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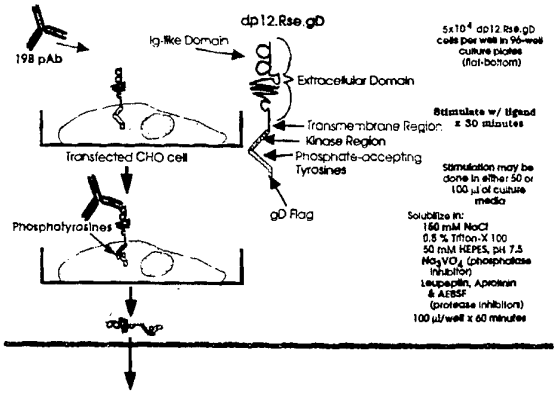
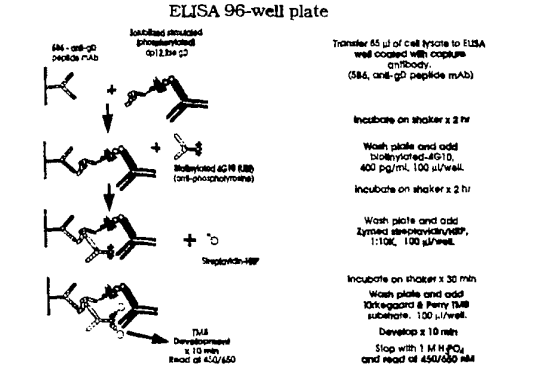
(54) Title: KINASE RECEPTOR ACTIVATION ASSAY

(57) Abstract

An assay for measuring activation (i.e., autophosphorylation) of a tyrosine kinase receptor of interest is disclosed. a) A first solid phase is coated with a substantially homogeneous population of cells so that the cells adhere to the first solid phase. The cells have either an endogenous tyrosine kinase receptor or have been transformed with DNA encoding a receptor or "receptor construct" and the DNA has been expressed so that the receptor or receptor construct is presented in the cell membranes of the cells. b) A ligand is then added to the solid phase having the adhering cells, such that the tyrosine kinase receptor is exposed to the ligand. c) Following exposure to the ligand, the adherent cells are solubilized, thereby releasing cell lysate. d) A second solid phase is coated with a capture agent which binds specifically to the tyrosine kinase receptor, or, in the case of a receptor construc, to the flag polypeptide. e) The cell lysate obtained in step c) is added to the wells containing the adhering capture agent so as to capture the receptor or receptor construct to the wells. f) A washing step is then carried out, so as to remove unbound cell lysate, leaving the captured receptor or receptor construct. g) The captured receptor or receptor construct is exposed to a labelled anti-phosphotyrosine antibody which identifies phosphorylated residues in the tyrosine kinase receptor. h) Binding of the anti-phosphotyrosine antibody to the captured receptor or receptor construct is measured.





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KINASE RECEPTOR ACTIVATION ASSAY

BACKGROUND OF THE INVENTION

Field of the Invention

This invention relates to a kinase receptor activation (KIRA) assay. In particular, the invention relates to an assay for measuring autophosphorylation of the kinase domain of a receptor protein tyrosine kinase (rPTK) using a kinase receptor activation, enzyme-linked immunosorbent assay (KIRA ELISA).

Description of Related Art

One mechanism for signal transduction in animals involves protein phosphorylation. Protein phosphorylation involves the action of protein kinase, an enzyme that transfers a phosphate group from a phosphate donor onto an acceptor amino acid in a substrate protein. Protein phosphatases provide a means for reversing the signal when the stimulus is removed.

Protein kinases have multiple substrates, and classification of the protein kinases is based on the acceptor amino acid specificity. The two most well characterized protein kinases are the protein kinases with a protein alcohol group as acceptor called protein serine/threonine kinases and the protein kinases with a protein phenolic group as acceptor called protein tyrosine kinases (Hunter, Methods in Enzymology **200**:3-9 [1991]).

The most well known type of signal-transducing protein kinases are growth factor receptor protein tyrosine kinases (rPTKs). rPTKs usually comprise a large, glycosylated, extracellular ligand binding domain (ECD) and an intracellular domain (ICD) which contains a tyrosine kinase catalytic domain. A single hydrophobic transmembrane (TM) domain connects the ECD and ICD. Examples of rPTKs include the insulin receptor, epidermal growth factor receptor (EGF-R), platelet-derived growth factor receptor (PDGF-R), insulin-like growth factor 1 receptor (IGF-1-R), and the HER2 receptor, to name a few. See, for example, Ullrich and Schlessinger Cell **61**:203-212 (1990) and Fantl *et al.*, Annu. Rev. Biochem. **62**:453-481 (1993). rPTKs can phosphorylate exogenous protein substrates and intrinsic tyrosine residues via their catalytic tyrosine kinase domain. The intrinsic tyrosine residues normally reside in the ICD of the rPTK (see Figure 1 herein). Activation of the intracellular kinase domain of rPTKs appears to be mediated by receptor oligomerization which results from the conformational alteration of the ECD upon ligand binding thereto. See

Ullrich and Schlessinger, *supra*.

Serine-threonine kinases have also been disclosed in the literature. While most of the known protein serine-threonine kinases are cytoplasmic proteins, a family of mammalian transmembrane receptors with serine-threonine kinase domains has recently been found. Members of this receptor family have been described as binding TGF-β's and activin. For reviews of serine-threonine kinases, see Sale, G., Biochem. Soc. Transactions 20: 664-670 (1992); ten Dijke *et al.*, Prog. in Growth Factor Res. 5: 55-72 (1994); and Mathews, L., Endoc. Rev. 15(3): 310-325 (1994).

Various assays have been developed which measure tyrosine kinase activity. Some of these assays measure the ability of a tyrosine kinase enzyme to phosphorylate a synthetic substrate polypeptide. For example, an assay has been developed which measures growth factor-stimulated tyrosine kinase activity by measuring the ability of the kinase to catalyze the transfer of the γ-phosphate of ATP to a suitable acceptor substrate. See Pike, L., Methods of Enzymology **146**:353-362 (1987) and Hunter, Journal of Biological Chemistry **257(9)**:4843-4848 (1982), for example. In this assay, the use of [γ-^{32}P]ATP permits the radioactive labeling of the phosphorylated substrate, which is a synthetic tyrosine-containing peptide. Others have described protein kinase assays wherein incorporation of ^{32}P into a tyrosine kinase receptor, such as the EGF receptor (see Donato *et al.*, Cell Growth Differ. **3**:259-268 [1992]), insulin receptor (see Kasuga *et al.*, Journal of Biological Chemistry **257(17)**:9891-9884 [1982] and Kasuga *et al.*, Methods in Enzymology **109**:609-621 [1985]), and liver growth hormone receptor (see Wang *et al.*, Journal of Biological Chemistry **267(24)**:17390-17396 [1992]), is measured.

The discovery of anti-phosphotyrosine antibodies has provided a non-radioactive, alternative means for measuring phosphorylation of tyrosine residues. For example, White and Backer (Methods in Enzymology **201**:65-67 [1991]) mention polyclonal antibodies which selectively bind to phosphotyrosine and are considered to be useful for studying rPTKs. An anti-phosphotyrosine monoclonal antibody was used in one of the assays referred to in Madden *et al.* (Anal Biochem **199**:210-215 [1991]), which measured phosphatase activity toward the insulin receptor. Anti-phosphotyrosine antibodies were also used by Cleaveland *et al.*, in their protein tyrosine kinase ELISA assay. See Cleaveland *et al.*, Analytical Biochemistry **190**:249-253 (1990). The method of Cleaveland *et al.* utilizes purified high-activity oncogene tyrosine kinases, v-src and v-fps, and

measures the ability of these tyrosine kinases to phosphorylate synthetic polymeric substrates which are coated on an ELISA microtiter plate. The phosphotyrosine produced by src-induced phosphorylation of the polymeric substrate is then quantitated by addition of an anti-phosphotyrosine antibody, the presence of which is detected using a second rabbit anti-mouse antibody which is linked to a reporter enzyme, horseradish peroxidase (HRPO). A similar ELISA assay has been developed by Lazaro *et al.*, which is used for detection of a protein tyrosine kinase. See Lazaro *et al.*, Analytical Biochemistry **192**:257-261 (1991). Like the assay of Cleaveland *et al.*, this assay also measures the ability of a protein tyrosine kinase to phosphorylate a synthetic substrate which is bound to microELISA wells.

A direct way to assess specific activation of rPTKs is by analysis of receptor autophosphorylation. See, *e.g.*, Hunter and Cooper Ann Rev Biochem **54**:897-930 (1985) and Ullrich and Schlessinger, Cell **61**:203-212 (1990). Using this direct approach, Knutson and Buck disclose assays for measuring autophosphorylation of the insulin receptor under *in situ* or *in vitro* conditions (Archives of Biochemistry and Biophysics **285(2)**:197-204 [1991]). In the *in situ* assay, monolayer cultures of embryonic mouse 3T3-C2 fibroblasts (having the endogenous insulin receptor) are incubated with insulin in large cell culture dishes. Following incubation, the insulin receptor is extracted from the membranes. To achieve extraction of the insulin receptor, the cell monolayers are scraped into a buffer containing protease inhibitors and the cells are then disrupted in a homogenizer. The cellular homogenate is subsequently subjected to centrifugation for 60 min., and the pellet which forms is extracted into buffer containing detergent. Following a further centrifugation step, the supernatant (containing the insulin receptor) is incubated with an anti-insulin receptor antibody. Then, the receptor-antibody complex is incubated with protein A-agarose and unoccupied protein A sites are blocked with normal rabbit IgG. The agarose beads are then centrifuged, the supernatants aspirated and the pellets are re-suspended in buffer containing the radiolabelled anti-phosphotyrosine antibody. The amount of bound iodinated anti-phosphotyrosine antibody is consequently measured.

Klein and his colleagues discuss an assay for measuring insulin activation of the insulin receptor (Klein *et al.*, Diabetes **42**:883-890 [1993]). In this assay, aliquots of a heterogeneous population of mononuclear blood cells (including T cells, B cells, macrophages etc) having the insulin receptor are exposed to insulin in centrifuge tubes.

The cells are then lysed in detergent using a motordriven homogenizer and the lysates are concentrated two- to fourfold using vacuum centrifugation. Sometimes, the insulin receptor is also partially purified using wheat germ agglutin agarose. The supernatants which form following centrifugation, are then transferred to anti-insulin receptor-coated microtiter plates. Insulin (8.7nM) as well as kinase and phosphatase inhibitors are present during receptor immobilization in order to optimize the percentage of receptors captured to the microtiter plates. Activation of the insulin receptor is then measured by transphosphorylation of the substrate Poly-Glu,Tyr with ^{32}P labeled ATP. The supernatants are then spotted onto absorbent paper and the paper is washed with cold TCA to remove unbound ^{32}P-ATP. Remaining ^{32}P-labeled Poly-Glu,Tyr on the washed absorbent paper is subsequently counted by scintillation counting.

Hagino *et al.* were also interested in studying the insulin receptor in patients (Hagino *et al.*, Diabetes **43**:274-280 [1994]). As a first step in the assay, Hagino *et al.* stimulate a primary cell suspension, which is not particularly homogeneous with respect to cell type. In particular, heparinized blood (1ml washed twice with medium and resuspended in 1 ml of medium containing bovine serum albumin, BSA) is exposed to varying concentrations of insulin. The autophosphorylation reaction is stopped, the cells centrifuged for 30 min, the supernatant is discarded and the erythrocyte ghosts thus obtained are resuspended in buffer and centrifuged again. The pellet thereby obtained is adjusted to 500μl and solubilized in detergent. The solubilized materials are then centrifuged and the resulting supernatant is subjected to sandwich ELISA (using anti-insulin receptor antibodies to capture the insulin receptor) to determine the extent of insulin receptor autophosphorylation.

King *et al.* in Life Sciences 53: 1465-1472 (1993) describe a colorimetric assay for examining inhibitors of the epidermal growth factor (EGF) receptor-associated tyrosine kinase in human intact epidermal A431 cells.

Several others have used an enzyme-conjugated form of the anti-phosphotyrosine antibody in Western blot analyses which measure receptor autophosphorylation. Briefly, Western blotting generally involves electrophoresing activated rPTK on polyacrylamide gel. The rPTK is then transferred to nitrocellulose and immunoblotted with the anti-phosphotyrosine antibody which is labelled to enable detection. See, for example, Wang, Molecular and Cellular Biology **5(12)**:3640-3643 (1985);

Glenney *et al.*, Journal of Immunological Methods **109**:277-285 (1988); Kamps, Methods in Enzymology **201**:101-110 (1991); Kozma *et al.*, Methods in Enzymology **201**:28-43 (1991); Holmes *et al.*, Science **256**:1205-10 (1992); and Corfas *et al.*, PNAS, USA **90**:1624-1628 (1993). However, with Western blot analysis, accurate quantitation can be very tedious. Furthermore, this technique tends to be time- consuming and generally does not allow high sample throughput.

It is an object of the instant invention to provide a sensitive, reliable assay that measures receptor protein tyrosine kinase (rPTK) autophosphorylation. The assay is desirably useful for qualitatively and quantitatively measuring kinase activation as well as facilitating identification and characterization of potential agonists and antagonists for a selected rPTK. It is a further object of the invention to provide an assay which enables ligand-receptor interactions to be studied for any selected rPTK.

This assay must have a capacity for high throughput, that is, the ability to reliably evaluate large numbers of samples in a relatively short period of time (*e.g.*, in one day). The assay ideally does not use radioactive materials and is also amenable to automation.

It is a further object, in at least one embodiment of the invention, to provide a generic assay which enables a rPTK of interest to be studied, regardless of whether or not a receptor-specific capture agent having the desired characteristics is available. Furthermore, it is an object of the invention to provide an assay which substantially represents the activity of the tyrosine kinase receptor *in situ*. This is desirable insofar as it reduces the possibility that altered interactions between the receptor and the ligand may occur as a consequence of the receptor not being membrane-bound. Furthermore, if the receptor is a multimeric complex, this assay enables the correctly assembled receptor to be studied. It is an additional object to provide a method for measuring serine-threonine kinase phosphorylation, phosphorylation of intracellular kinases and phosphatase activity.

These and other objects will be apparent to the ordinary artisan upon consideration of the specification as a whole.

SUMMARY OF THE INVENTION

Accordingly, the invention provides an assay for measuring activation (*i.e.*, autophosphorylation) of a tyrosine kinase receptor of interest.

The assay can be divided into two major stages, each of which is generally performed in separate assay plates. The first stage of the assay involves activating the receptor and is termed the kinase receptor activation (KIRA) stage of the assay. The second stage of the assay involves measuring receptor activation. Conveniently, this is achieved using an enzyme-linked immunosorbent assay (ELISA) to measure receptor activation.

The KIRA stage of the assay involves activating a tyrosine kinase receptor which is located in the cell membrane of an eukaryotic cell such that the extracellular domain of the receptor faces the external milieu of the cell, the transmembrane domain is located in the cell membrane and the kinase domain is located intracellularly. This stage of the overall assay involves steps (a) to (c) below:

(a) The first solid phase (*e.g.*, a well of a first assay plate) is coated with a substantially homogeneous population of cells (usually a mammalian cell line) so that the cells adhere to the solid phase. Often, the cells are adherent and thereby adhere naturally to the first solid phase. In one embodiment of the invention, the cells have an endogenous tyrosine kinase receptor presented in the cell membrane as discussed above. In an alternative embodiment, the cells have been transformed with DNA encoding a tyrosine kinase receptor or a "receptor construct" defined further below, which DNA is expressed by the cells such that the receptor or receptor construct is suitably positioned in the cell membranes thereof.

The receptor construct comprises a fusion of a kinase receptor and a flag polypeptide. The flag polypeptide is recognized by the capture agent, often a capture antibody, in the ELISA part of the assay. Use of a receptor construct as disclosed herein is particularly advantageous since it provides a "generic" assay wherein autophosphorylation of any tyrosine kinase receptor can be measured, regardless of whether or not a receptor-specific capture agent having the required characteristics is available. Often, the tyrosine kinase receptor is a fusion protein comprising the ECD of a selected tyrosine kinase and the catalytic ICD (and possibly the transmembrane domain) of another well characterized tyrosine kinase (*e.g.*, the Rse receptor).

(b) An analyte is then added to the wells having the adhering cells, such that the tyrosine kinase receptor is exposed to (or contacted with) the analyte. This assay enables identification of agonist and antagonist ligands for the tyrosine kinase receptor of interest. In order to detect

the presence of an antagonist ligand which blocks binding and/or activation of the receptor by an agonist ligand, the adhering cells are exposed to the suspected antagonist ligand first and then to the agonist ligand (or to a mixture of the agonist and antagonist) so that competitive inhibition of receptor binding and activation can be measured. Also, the assay can identify an antagonist which binds to the agonist ligand and thereby reduces or eliminates its ability to bind to, and activate, the rPTK. To detect such an antagonist, the suspected antagonist and the agonist for the rPTK are incubated together and the adhering cells are then exposed to this mixture of ligands.

(c) Following exposure to the analyte, the adhering cells are solubilized using a lysis buffer (which has a solubilizing detergent therein) and gentle agitation, thereby releasing cell lysate which can be subjected to the ELISA part of the assay directly, without the need for concentration or clarification of the cell lysate. Thus, this assay provides a significant improvement over assays described by Knutson and Buck, *supra*, Klein *et al.*, *supra*, and Hagino *et al.*, *supra*, insofar as it is surprisingly unnecessary to concentrate the cell lysate prior to the ELISA. Furthermore, unlike the other assays, in the instant assay the cells can be lysed in lysis buffer using gentle agitation without the need for homogenizing, centrifuging or clarifying the cells. The cell lysate thus prepared is then ready to be subjected to the ELISA stage of the assay. It has been discovered that, surprisingly, the first assay plate can be stored at freezing temperatures (*i.e.*, at about -20° to -70°C) for significant periods of time (at least 6 months) before the ELISA stage of the assay. This is a significant finding insofar as the KIRA and ELISA stages of the assay can be performed on separate days.

The ELISA component of the assay comprises steps (d) to (h), described below.

(d) As a first step, the second solid phase (usually a well of an ELISA microtiter plate) is coated with a capture agent (often a capture antibody) which binds specifically to the tyrosine kinase receptor, or, in the case of a receptor construct, to the flag polypeptide. Coating of the second solid phase is carried out so that the capture agent adheres to the second solid phase. The capture agent is generally a monoclonal antibody, but, as is described in the examples herein, polyclonal antibodies may also be used.

(e) The cell lysate obtained in step (c) of the above-mentioned KIRA stage of the assay is exposed to, or contacted with, the adhering capture agent so that the receptor or receptor construct adheres to (or is captured in) the second solid phase. Unlike the assay of Klein *et al.*, the instant assay does not require the ligand for the receptor as well as kinase inhibitors to be present to achieve suitable immobilization of the receptor or receptor construct to the second solid phase.

(f) A washing step is then carried out, so as to remove unbound cell lysate, leaving the captured receptor or receptor construct.

(g) The adhering or captured receptor or receptor construct is then exposed to, or contacted with, an anti-phosphotyrosine antibody which identifies phosphorylated tyrosine residues in the tyrosine kinase receptor. In the preferred embodiment, the anti-phosphotyrosine antibody is conjugated (directly or indirectly) to an enzyme which catalyses a color change of a non-radioactive color reagent. Accordingly, phosphorylation of the receptor can be measured by a subsequent color change of the reagent. The enzyme can be bound to the anti-phosphotyrosine antibody directly, or a conjugating molecule (*e.g.*, biotin) can be conjugated to the anti-phosphotyrosine antibody and the enzyme can be subsequently bound to the anti-phosphotyrosine antibody via the conjugating molecule.

(h) Finally, binding of the anti-phosphotyrosine antibody to the captured receptor or receptor construct is measured, *e.g.*, by a color change in the color reagent.

The invention also pertains to a Rse.flag reagent which is particularly useful for use in the KIRA ELISA assay. The Rse.flag reagent is a polypeptide comprising a fusion of a flag polypeptide (usually the gD flag described herein) to the carboxyl terminus of the intracellular domain of the Rse rPTK. Generally, the transmembrane domain of Rse and the extracellular domain of another rPTK of interest are also present in the fusion polypeptide reagent. The nucleic acid encoding this reagent and a cell transformed therewith are also claimed.

In yet a further aspect, the invention relates to a kit which can be used in the KIRA ELISA disclosed above which comprises an anti-flag polypeptide capture agent (*e.g.* a capture antibody) which is usually bound to the second solid phase as described herein. Thus, the kit generally provides an ELISA microtiter plate having an anti-flag polypeptide capture antibody adhering to a well thereof. Optionally, the kit also provides an anti-phosphotyrosine antibody which is often labelled, or reagents for

labelling the anti-phosphotyrosine antibody are supplied with the kit. Sometimes, a homogeneous population of cells which have been transformed with a receptor construct as described herein are also provided with the kit. The kit also suitably includes instructions for carrying out the KIRA ELISA.

BRIEF DESCRIPTION OF THE DRAWINGS

Figures 1A-1C are diagrammatic representations of Rse.gD (Figure 1A), Receptor ECD/Rse.gD chimera (Figure 1B) and a CHO cell transformed with the Receptor ECD/Rse.gD chimera (Figure 1C).

Figures 2A and 2B depict an alignment of the amino acid sequence (SEQ ID NO: 1) and nucleotide sequence (SEQ ID NO: 2) of Rse.gD. The residues of the signal sequence are indicated with an (*), the transmembrane domain of Rse is boxed and the ECD and ICD of Rse are also delineated. The residues of the gD flag sequence are underlined.

Figure 3 is a flow diagram of an exemplary strategy for selecting a suitable capture agent for use in the assay.

Figure 4 is a flow diagram of an exemplary strategy for selecting a transformed cell suitable for use in the assay, where the cell has a receptor construct with an amino-terminal flag polypeptide located in the cell membrane thereof.

Figure 5 is a flow diagram of an exemplary strategy for selecting a transformed cell suitable for use in the assay, where the cell has a receptor construct with a carboxyl-terminal flag polypeptide located in the cell membrane thereof.

Figure 6 is a flow chart and cartoon illustrating the KIRA ELISA assay for the HER2 receptor described in Example 1.

Figure 7 depicts a $p185^{HER2}$/$HRG\beta1_{177-244}$ KIRA ELISA standard curve obtained using the assay described in Example 1. To obtain the standard curve, MCF-7 cells ($2x10^5$) were stimulated with 3000, 1000, 333, 111, 37, 12, 4, or 0 pM $HRG\beta1_{177-244}$, as determined by quantitative amino acid analysis (q.a.a.a.). Each calibrator concentration was run in triplicate. The values derived from 10 such standard curves were averaged (total n = 30) and are presented as mean $ABS_{450/650}$ ± sd vs. $HRG\beta1_{177-244}$ concentration.

Figure 8 depicts heregulin specificity of $p185^{HER2}$/HRG KIRA ELISA of Example 1. In the assay, MCF-7 cells ($2x10^5$) were stimulated with either $HRG\beta1_{177-244}$ (■) at 3000, 1000, 333, 111, 37, 12, 4 or 0 pM or IGF-1(▲), EGF (□), VEGF (●) or insulin (♦) at 30000, 10000, 3333, 1111, 370, 120, 40 or

0 pM. For all concentrations of ligands, n = 3 and data are presented as average $ABS_{450/650}$ ± sd vs. ligand concentration.

Figure 9 is a flow chart and cartoon illustrating the KIRA ELISA assay for the Rse receptor described in Example 2.

Figure 10 depicts a Rse KIRA ELISA standard curve obtained using the assay described in Example 2. To obtain the standard curve, CHO cells transformed with the Rse.gD construct were stimulated with 1:100, 1:200, 1:400, 1:800, 1:1600, 1:3200 or 0 diluted, anti-Rse agonist antibody. Each calibrator concentration was run in triplicate. The values are presented as mean $ABS_{450/650}$ ± sd vs. 1/dilution agonist antibody.

Figure 11 is a flow chart and cartoon illustrating the KIRA ELISA assay for the trk receptors (*i.e.*, trk A, trk B, and trk C) described in Example 3.

Figures 12A-12D depict an alignment of the amino acid acid sequence (SEQ ID NO: 3) and nucleotide sequence (SEQ ID NO: 4) of gD.trk A used in the assay described in Example 3. The residues of the signal sequence are indicated with an (*), the residues of the gD flag sequence are underlined, the residues of the transmembrane domain of trk A are in bold and the ECD and ICD thereof are also delineated.

Figures 13A-13D depict an alignment of the amino acid sequence (SEQ ID NO: 5) and nucleotide sequence (SEQ ID NO: 6) of gD.trk B used in the assay described in Example 3. The residues of the signal sequence are indicated with an (*), the residues of the gD flag sequence are underlined, the residues of the transmembrane domain of trk B are in bold and the ECD and ICD thereof are also delineated.

Figures 14A-14D depict an alignment of the amino acid sequence (SEQ ID NO: 7) and nucleotide sequence (SEQ ID NO: 8) of gD.trk C used in the assay described in Example 3. The residues of the signal sequence are indicated with an (*), the residues of the gD flag sequence are underlined, the residues of the transmembrane domain of trk C are in bold and the ECD and ICD thereof are also delineated.

Figures 15A-15C depict standard curves for trk A, B and C, respectively, which were obtained using the assay described in Example 3. To obtain the standard curves, CHO cells transformed with the gD.trk constructs were stimulated with 3000, 1000, 333, 111, 37, 12, 4 or 0 pM of ligand, *i.e.* nerve growth factor (NGF, ■), neurotrophin 3 (NT3, ●) or neurotrophin 5 (NT5, ▲). The values are presented as mean $ABS_{450/650}$ ± sd vs. ligand concentration.

Figures 16A-16L depict the nucleotide sequence (SEQ ID NO: 9) of the pSVI17.ID.LL expression vector used for expression of Rse.gD in Example 2.

Figure 17 is a diagrammatic representation of the *MPL*/Rse.gD chimeric receptor described in Example 4.

Figure 18 is a flow chart and cartoon illustrating the KIRA ELISA for the *MPL*/Rse.gD chimeric receptor described in Example 4.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

I. Abbreviations and Definitions

"rPTK" means a receptor protein tyrosine kinase.

"ECD", "TM domain" and "ICD" refer to the extracellular domain, transmembrane domain and intracellular domain of a rPTK, respectively.

"Kinase Receptor Activation" or "KIRA" when used throughout this application refers to the first stage of the instantly claimed assay wherein a cell-bound rPTK is exposed to a potential agonist/antagonist ligand which may (or may not) induce phosphorylation of tyrosine residues in the intracellular domain of the rPTK. The KIRA is generally carried out in the "first assay plate" as defined herein.

"Enzyme-Linked Immunosorbent Assay" or "ELISA" refers to the second stage of the instantly claimed assay and involves measuring tyrosine phosphorylation of the rPTK. The ELISA is normally carried out in the "second assay plate" as disclosed in this application. The ELISA is a "sandwich ELISA" insofar as it involves capturing the rPTK or receptor construct to the second solid phase (usually the well of an ELISA microtiter plate). ELISA assays generally involve the preparation of enzyme-antibody conjugates. The conjugated enzyme cleaves a substrate to generate a colored reaction product that can be detected spectrophotometrically. In this assay, the absorbance of the colored solution in individual microtiter wells is proportional to the amount of phosphotyrosines. A review of ELISA is found in Current Protocols in Molecular Biology, Vol. 2, chapter 11 (1991). While the term "ELISA" is used to describe the second stage of the instant assay, it is only a preferred embodiment of the invention, since, as disclosed herein, techniques other than enzymatic detection are available for measuring binding of the anti-phosphotyrosine antibody to the activated receptor.

The terms "receptor", "kinase receptor", "tyrosine kinase", "tyrosine kinase receptor", "receptor protein tyrosine kinase" and "rPTK" are used

interchangeably herein and refer to a protein having at least one phosphate accepting phenolic group. The protein is usually a receptor insofar as it has a ligand-binding ECD, TM domain and ICD. The ICD usually comprises a catalytic kinase domain and has one or more phosphate accepting tyrosine residues. See Figures 1A and 1B, for example. Examples of tyrosine kinase receptors include the insulin receptor, insulin related receptor, epidermal growth factor receptor (EGF-R), platelet-derived growth factor receptors A and B (PDGF-R-A and PDGF-R-B), insulin-like growth factor 1 receptor (IGF-1-R), macrophage colony-stimulating factor receptor (M-CSF-R), HER2/*neu*/c-*erb*B-2 receptor, HER3/c-*erb*B-3 receptor, *Xmrk* receptor, IRR receptor, fibroblast growth factor (FGF) receptors *bek* and *flg*, c-*kit* receptor, Flk/kDR receptor, Rse receptor, the *Eph*, *Elk*, *Eck*, *Eek*, *Erk*, *Cek4/Mek4/HEK and Cek5* receptors, Axl receptor, hepatocyte growth factor receptor (HGF-R), Flt1 VEGF receptor, SAL-S1 receptor, HpTK 5 receptor, trkA receptor, trkB receptor, and trkC receptor. See, for example, Ullrich and Schlessinger Cell **81**:203-212 (1990); Fantl *et al.*, Annu. Rev. Biochem. **62**:453-481 (1993); Mark *et al.*, Journal of Biological Chemistry **269(14)**:10720-10728 (1994); and WO 93/15201.

The terms mentioned above encompass chimeric "receptor" molecules which comprise at least the extracellular domain of a selected tyrosine kinase and the intracellular domain, and optionally, the transmembrane domain of another tyrosine kinase. Of course, the tyrosine kinase of interest can provide the transmembrane domain and/or intracellular domain. The terms also encompass amino acid sequence variants and covalent derivatives of the various rPTKs provided they still display tyrosine kinase phosphorylation activity in the KIRA ELISA. Therefore, the variants will general have conservative amino acid alterations. The individual domains of the tyrosine kinase can be delineated based on sequence homology to known tyrosine kinases and hydrophobicity plots. For example, the hydrophobic transmembrane domain can be readily determined and the ECD and ICD are usually amino-terminal and carboxyl terminal to the transmembrane domain, respectively. Conveniently, the transmembrane domain and ICD of the Rse receptor can be fused to the ECD of a tyrosine kinase of interest, thereby forming a chimeric receptor which is encompassed by the terms denoting a receptor as mentioned above.

In the preferred embodiment, the rPTK is selected from the group consisting of HER2 receptor (Ullrich and Schlessinger, *supra*), Rse receptor

(Mark *et al.*, *supra* and SEQ ID NO: 1), trk A receptor (SEQ ID NO: 3), trk B receptor (SEQ ID NO: 5) and trk C receptor (SEQ ID NO: 7).

By "autophosphorylation" is meant activation of the catalytic kinase domain of the rPTK, whereby at least one intrinsic tyrosine residue is phosphorylated. Generally, autophosphorylation will result when an agonist molecule binds to the extracellular domain of the kinase receptor. Without being limited to any particular mechanism of action, it is thought that binding of the agonist molecule may result in oligomerization of the kinase receptor which causes activation of the catalytic kinase domain.

By "solid phase" is meant a non-aqueous matrix to which the cells (in the KIRA stage of the assay) or the capture agent (in the ELISA stage of the assay) can adhere. Usually, the solid phase comprises the well of an assay plate but the invention is by no means limited to this embodiment. For example, the solid phase can comprise a discontinuous solid phase of discrete particles. The particles can be porous and formed from a number of different materials, *e.g.*, polysaccharides (*e.g.* agarose), polyacrylamides, polystyrene, polyvinyl alcohol, silicones and glasses. For examples of suitable particulate solid phases, see U.S. Patent No. 4,275,149.

By "well" is meant a recess or holding space in which an aqueous sample can be placed. The well is provided in an "assay plate". The invention usually employs a "first assay plate" which is formed from a material (*e.g.* polystyrene) which optimizes adherence of cells (having the receptor or receptor construct) thereto. Generally, the individual wells of the first assay plate will have a high surface area to volume ratio and therefore a suitable shape is a flat bottom well (where the cells are adherent). The "second assay plate" is generally formed from a material (*e.g.* polystyrene) which optimizes adherence of the capture agent thereto. The second assay plate may have the same general construction and/or characteristics as the first assay plate. However, separate plates are used for the KIRA stage of the assay and the ELISA stage of the assay.

In the preferred embodiment of the invention, both the first assay plate and the second assay plate are "microtiter" plates. The term "microtiter" plate when used herein refers to an assay plate having between about 30 to 200 individual wells, usually 96 wells. Often, the individual wells of the microtiter plate will hold a maximum volume of about 250μl. Conveniently, the first assay plate is a 96 well polystyrene or plastic, cell culture microtiter plate (such as that

sold by Becton Dickinson Labware, Lincoln Park, NJ), which allows for automation. Often, about 50μl to 300μl, more preferably 100μl to 200μl, of an aqueous sample comprising cell culture media with the cells suspended therein will be added to each well of the first assay plate in the KIRA stage of the assay. It is desirable to seed between about 1×10^4 to 3×10^5 cells per well. More preferably, 5×10^4 to 1×10^5 cells per well are seeded. Usually, the second assay plate will comprise a polystyrene microtiter ELISA plate such as that sold by Nunc Maxisorp, Inter Med, Denmark.

The term "homogeneous population of cells" refers to a substantially homogeneous population of cells wherein at least about 80%, and preferably about 90%, of the cells in the population are of the same cell type. Therefore, it is convenient to use a cell line. The cell line is a eukaryotic cell line, normally an animal cell line and desirably a mammalian cell line.

The cells have, or are transformed to produce, the selected receptor or a receptor construct. For example, where the kinase receptor is known to be present in a certain cell line (*e.g.*, the HER2 receptor in the MCF-7 cell line) no transformation step is required. Conversely, it may be necessary to transform a cell with a nucleic acid encoding the receptor, where the cell does not make the receptor, or does not have suitable numbers of the receptor in the cell membrane thereof. Accordingly, the cell is transformed with a nucleic acid encoding the receptor (or receptor construct) and the nucleic acid is expressed so that the ECD of the receptor faces the external milieu of the cell, the transmembrane domain is located in the cell membrane and the kinase domain is located intracellularly.

Where the assay relies on activating the endogenous rPTK, a cell line is selected which is known to produce the rPTK of interest, provided sufficient levels of the rPTK are present in the cell membrane thereof to enable detection. As a general proposition, a minimum number of about 1×10^4 receptors/cell is required. For example, the MCF-7 cell line (ATCC-HTB 22) which produces the HER2 receptor was shown to be useful in the assay. There are 5×10^4 HER2 receptors/MCF-7 cell. Examples of other cell lines and their respective rPTKs include, embryonic mouse 3T3-C2 fibroblast cell line and the insulin receptor, and Hep 3B (ATCC # HB 8064) cell line and the Rse receptor. However, the degree of expression of the rPTK nucleic acid in the cell line is not so high that it results in constitutive

phosphorylation of the rPTK. For example, the SK-BR-3 cell line (ATCC HTB30), which has 3 x 10^6 HER2 receptors/cell, was found to be unsuitable for use in the assay disclosed herein. Therefore, it may be useful to use a cell line which has less than about 3 x 10^6 receptors/cell, depending on the type of receptor. The number of receptors/cell can be measured using Scatchard analysis, for example (Scatchard, Ann. NY Acad. Sci. 51:660-672 [1949]; and Goodwin *et al.*, Cell 73:447-456 [1993]). However, selection of a cell line having a suitable number of receptors/cell is possible using the techniques described herein.

The term "adherent" when used herein to describe the cell, refers to a cell which naturally adheres to the first solid phase (often the well of the first assay plate), thereby forming a fairly uniform coating of the cells on the inside surface of the well. The uniform coating of cells generally forms following incubation of the cells in the wells of the first assay plate for about 8-16 hours. After incubation, non-adhering cells and cell culture medium are decanted off the first assay plate. Incubation is usually carried out at a temperature which is optimal for cell growth, *i.e*, about 37°C. Examples of adherent cell lines include CHO cells (Urlaub and Chasin, Proc. Natl. Acad. Sci. USA 77:4216 [1980]), MCF-7 cells (ATCC HB 22), 293 cells (Graham *et al.*, J. Gen Virol. 36:59 [1977]), Swiss albino 3T3 fibroblast cell line (ATCC No. CCL 92) and U937 macrophage cell line (ATCC No. CRL 1593).

A "flag polypeptide" comprises a short polypeptide which has enough residues to provide an epitope (preferably a linear epitope) against which a "capture agent" thereagainst can be made, yet is short enough such that it does not interfere with activity of the rPTK. The flag polypeptide is also sufficiently unique so that the capture agent thereagainst does not bind to other reagents in the assay. Selection of a "unique" flag polypeptide sequence can be accomplished by comparing the sequence of a proposed flag polypeptide against other known sequences in Genbank or EMBL, for example. Suitable flag polypeptides generally have at least 6 amino acid residues and usually between about 8-80 amino acid residues (preferably between about 9-30 amino acid residues).

By "receptor construct" is meant a polypeptide which comprises a fusion of a kinase receptor and a flag polypeptide as defined above. The flag polypeptide is provided at a location in the receptor construct such that: a) the flag polypeptide does not interfere with ligand binding to the receptor; b) the flag polypeptide does not interfere with

autophosphorylation of the receptor and c) the flag polypeptide is presented in a suitable configuration so that it can bind to the capture agent in the ELISA stage of the assay. Often, the polypeptide flag will be present at the N-terminus of the receptor construct. See, for example, Example 3 which refers to the gD.trk constructs. Alternatively, the flag polypeptide may be present at the C-terminus of the receptor construct. See, for example, Example 2 which refers to the Rse.gD construct. See also Figures 1A-1C. The Rse construct disclosed herein is particularly useful, since the ICD (and optionally the transmembrane domain) thereof can be fused to the ECD of a kinase receptor of interest, thereby obviating the need to establish where the flag polypeptide should be located with respect to the kinase receptor of interest.

"Rse.gD" refers to a receptor construct which is the Rse receptor protein tyrosine kinase with the Herpes Simplex virus glycoprotein D (gD) flag polypeptide fused to the COOH-terminus thereof.

"Rse.flag reagent" refers to a polypeptide which comprises the ICD of the Rse receptor fused at its COOH-terminus to a flag polypeptide (normally the gD flag polypeptide). Sometimes, the TM domain of Rse and the ECD of a rPTK of interest will also be present in the Rse.gD. reagent. "Receptor ECD/Rse.gD Chimera" refers to a fusion of the ECD of a rPTK of interest to the TM and ICD domains of Rse which are fused COOH-terminally to the gD flag polypeptide.

"gD.trkA", "gD.trkB" and "gD.trkC" refer to each of the trk receptors (A-C) having the gD flag polypeptide fused to the amino-termini thereof.

By "capture agent" is meant a compound or agent which is able to adhere to the second solid phase, as herein defined, and which is selective for a rPTK or receptor construct. Thus, the capture agent captures the receptor or receptor construct to the wells of the second assay plate. Usually, the capture agent binds selectively to the flag polypeptide which has been fused to the receptor of interest. Binding of the capture agent is not affected by the presence or absence of ligand bound to the receptor and does not induce receptor activation upon capture. Furthermore, the capture agent does not sterically block access to the phosphorylated tyrosine(s) by the anti-phosphotyrosine antibody. Means for selecting suitable capture agents are described herein. Generally, the capture agent will comprise an antibody (*e.g.*, an affinity purified polyclonal antibody or a monoclonal antibody), but other selective agents, such as streptavidin which binds selectively to the "*strep*-tag" polypeptide can also be used

(see Schmidt *et al.*, Protein Engineering 6(1):109-122 [1993]). Streptavidin can be purchased commercially from Zymed Laboratories, S. San Francisco, CA, for example. Alternatively, the capture agent can comprise protein A (which binds specifically to immunoglobulins). In this embodiment of the invention, the activated receptor or receptor-construct present in the cell lysate is incubated with an antibody which binds specifically thereto, thereby forming a receptor-antibody complex. This complex can be captured by protein A by virtue of its specific binding to the antibody present in the complex. Protein A can be purchased commercially from Pharmacia Biotech, Inc., Piscataway, New Jersey, for example. A strategy for selecting a suitable capture agent is depicted in Figure 3 and will be described in more detail later herein.

In the most preferred embodiment, the capture agent is a monoclonal antibody which binds specifically to a flag polypeptide (which is present in the receptor construct). Examples of suitable flag polypeptides and their respective capture antibodies include the flu HA flag and its antibody 12CA5, (Field *et al.*, Mol. Cell. Biol. 8:2159-2165 [1988]); the c-myc flag and the 8F9, 3C7, 6E10, G4, B7 and 9E10 antibodies thereto (Evan *et al.*, Molecular and Cellular Biology **5(12)**:3610-3616 [1985]); as well as the Herpes Simplex virus glycoprotein D (gD) flag and the 5B6 antibody thereto (Paborsky *et al.*, Protein Engineering **3(6)**:547-553 [1990] and Mark *et al.*, Journal of Biological Chemistry **269(14)**:10720-10728 [1994]). Other flag polypeptides have been disclosed. Examples include the Flag-peptide (Hopp *et al.*, BioTechnology 6:1204-1210 [1988]); the KT3 epitope peptide (Martin *et al.*, Science **255**:192-194 [1992]); an α-tubulin epitope peptide (Skinner *et al.*, J. Biol. Chem **266**:15163-15166 [1991]); and the T7 gene 10 protein peptide tag (Lutz-Freyermuth *et al.*, Proc. Natl. Acad. Sci. USA **87**:6393-6397 [1990]). Once the flag polypeptide has been selected as discussed above, a capture antibody thereto can be generated using the techniques disclosed herein.

The term "analyte" refers to a compound or composition to be studied, usually to investigate its ability to activate (or prevent activation of) the tyrosine kinase receptor of interest. The analyte can comprise a bodily fluid (such as plasma or amniotic fluid) or a composition known to contain, or suspected of containing, a ligand for the tyrosine kinase receptor. The analyte can also comprise a cell which has a ligand to the rPTK of interest.

"Ligand" when used herein refers to a molecule which is able to bind to the ECD of the tyrosine kinase of interest or to a known agonist for the tyrosine kinase of interest. The ligand will usually be an agonist or antagonist for the tyrosine kinase.

By "agonist" is meant a molecule which is able activate the intracellular kinase domain of the tyrosine kinase upon binding to the ECD. Often, the agonist will comprise a growth factor (*i.e.*, a polypeptide that is able to stimulate cell division). Exemplary growth factors include heregulin (HRG), insulin, insulin-like growth factors I and II (IGF-I and IGF-II), epidermal growth factor (EGF), interleukins (*e.g.*, IL-8), macrophage colony-stimulating factor (M-CSF), erythropoietin (EPO), platelet-derived growth factor (PDGF), fibroblast growth factor (FGF), transforming growth factors alpha and beta (TGF-α and TGF-β), hepatocyte growth factor (HGF), and nerve growth factor (NGF). Alternatively, the agonist can be an antibody against the rPTK (see, *e.g.*, Yarden, Proc. Natl. Acad. Sci., USA **87**:2569-2573 [1990]). However, other non-protein agonists such as small organic molecules are also encompassed by the invention.

By "antagonist" is meant a molecule which blocks agonist action. Usually, the antagonist will either: (a) bind to the rPTK and thereby block binding and/or activation of the rPTK by an agonist thereto (the antagonist may bind to the ECD of the rPTK, but this is not necessarily the case) or (b) bind to the agonist and thus prevent activation of the rPTK by the agonist. This assay facilitates the detection of both types of antagonist. The antagonist may, for example, comprise a peptide fragment comprising the receptor binding domain of the endogenous agonist ligand for the receptor. The antagonist may also be an antibody which is directed against the ECD of the rPTK, or against a known agonist for the rPTK. However, other non-protein molecules are also encompassed by this term.

The term "antibody" is used in the broadest sense and specifically covers monoclonal antibodies and antibody compositions with polyepitopic specificity (*i.e.* polyclonal antibodies). The polyclonal antibodies are preferably "affinity purified" antibodies. The term "affinity purified" means that the antibodies have been purified using the antigen (*e.g.* the rPTK or fragment thereof or the flag polypeptide) to selectively purify the polyclonal antibodies. Affinity purification can be achieved by immobilizing the antigen on an affinity column (*e.g.* an agarose column) and passing the polyclonal antibodies through the column. The affinity purified antibodies can be subsequently eluted from the column by changing

the elution conditions or by adding a chaotropic agent, for example. For a review of affinity purification techniques with respect to antibodies, see Current Protocols in Immunology, Ed. Coligen *et al.*, Wiley publishers, Vols. 1 and 2, for example.

The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, *i.e.*, the individual antibodies comprising the population are identical except for possible naturally-occurring mutations that may be present in minor amounts. Monoclonal antibodies are highly specific, being directed against a single antigenic site. Furthermore, in contrast to conventional (polyclonal) antibody preparations which typically include different antibodies directed against different determinants (epitopes), each monoclonal antibody is directed against a single determinant on the antigen.

The monoclonal antibodies herein include hybrid and recombinant antibodies produced by splicing a variable (including hypervariable) domain of a selected antibody with a constant domain (*e.g.* "humanized" antibodies), or a light chain with a heavy chain, or a chain from one species with a chain from another species, or fusions with heterologous proteins, regardless of species of origin or immunoglobulin class or subclass designation, as well as antibody fragments (*e.g.*, Fab, $F(ab')_2$, and Fv), so long as they exhibit the desired biological activity. [See, *e.g.* U.S. Patent No. 4,816,567 and Mage & Lamoyi, in Monoclonal Antibody Production Techniques and Applications, pp.79-97 (Marcel Dekker, Inc., New York (1987)].

Thus, the modifier "monoclonal" indicates the character of the antibody as being obtained from a substantially homogeneous population of antibodies, and is not to be construed as requiring production of the antibody by any particular method. For example, the monoclonal antibodies to be used in accordance with the present invention may be made by the hybridoma method first described by Kohler & Milstein, Nature, **256**: 495 (1975), or may be made by recombinant DNA methods (U.S. Patent No. 4,816,567). The "monoclonal antibodies" may can also be isolated from phage antibody libraries using the techniques described in Clackson *et al.*, Nature, **352**: 624-628 (1991) and Marks *et al.*, J. Mol. Biol., **222**: 581-597 (1991), for example.

The term "anti-phosphotyrosine antibody" refers to a molecule, usually an antibody, which binds selectively to phosphorylated tyrosine

residues in the kinase domain of a rPTK. The antibody can be polyclonal, but is desirably a monoclonal antibody. Anti-phosphotyrosine polyclonal antibodies can be made using the techniques disclosed in White and Backer, Methods in Enzymology **201**:65-67 [1991] and monoclonal anti-phosphotyrosine antibodies can be obtained commercially from Upstate Biologicals, Inc. (UBI, Lake Placid, NY), for example.

The word "label" when used herein refers to a detectable compound or composition which is conjugated directly or indirectly with a molecule (such as the anti-phosphotyrosine antibody). The label may be detectable by itself (*e.g.* radioisotope labels or fluorescent labels) or, in the case of an enzymatic label, may catalyze a chemical alteration of a substrate compound or composition which is detectable. The preferred label is an enzymatic one which catalyzes a color change of a non-radioactive color reagent.

By "washing" is meant exposing the solid phase to an aqueous solution (usually a buffer or cell culture media) in such a way that unbound material (*e.g.*, non-adhering cells, non-adhering capture agent, unbound ligand, receptor, receptor construct, cell lysate, or anti-phosphotyrosine antibody) is removed therefrom. To reduce background noise, it is convenient to include a detergent (*e.g.* Triton X) in the washing solution. Usually, the aqueous washing solution is decanted from the wells of the assay plate following washing. Conveniently, washing can be achieved using an automated washing device. Sometimes, several washing steps (*e.g.*, between about 1 to 10 washing steps) may be required.

By "block buffer" is meant an aqueous, pH buffered solution containing at least one blocking compound which is able to bind to exposed surfaces of the second solid phase which are not coated with capture agent. The blocking compound is normally a protein such as bovine serum albumin (BSA), gelatin, casein or milk powder and does not cross-react with any of the reagents in the assay (*e.g.*, the anti-phosphotyrosine antibodies and detection reagents). The block buffer is generally provided at a pH between about 7 to 7.5 and suitable buffering agents include phosphate and TRIS.

By "lysis buffer" is meant an aqueous, pH buffered solution comprising a solubilizing detergent, one or more protease inhibitors and at least one phosphatase inhibitor (such as sodium orthovanadate). The term "solubilizing detergent" refers to a water miscible, non-ionic detergent which lyses cell membranes of eukaryotic cells but does not

denature or activate the receptor or receptor construct. Examples of suitable non-ionic detergents include Triton-X 100, Tween 20, CHAPS and Nonidet P-40 (NP40) available from Calbiochem, La Jolla, California, for example. Many other non-ionic detergents are available in the art. Examples of suitable protease inhibitors include phenylmethylsulfonyl fluoride (PMSF), leupeptin, pepstatin, aprotinin, 4-(2-aminoethyl)-benzenesulfonyl fluoride hydrochloride-bestatin, chymostatin and benzamidine. Preservatives (*e.g.*, thimerosal) and one or more compounds which maintain the isotonicity of the solution (*e.g.*, sodium chloride [NaCl] or sucrose) and a buffer (*e.g.*, Tris or PBS) are usually also present. Generally, the pH of the lysis buffer is in the range about 7 to 7.5.

Usually, following addition of the lysis buffer to the first assay plate, the first assay plate is "gently agitated" and this expression refers to the act of physically shaking the first assay plate (normally using a circular motion) at a substantially low velocity. Gentle agitation does not involve mechanically disrupting the cells (*e.g.* by homogenizing or centrifuging the cells). Exemplary shaking velocities are in the order of 200 to 500 rpm, preferably 300 to 400 rpm in a Bellco orbital shaker, for example.

II. Modes for Practicing the Invention

1. Kinase Receptor Activation - KIRA

The first stage of the assay involves phosphorylation of the kinase domain of a kinase receptor, wherein the receptor is present in the cell membrane of a eukaryotic cell. The receptor may be an endogenous receptor or nucleic acid encoding the receptor may be transformed into the cell. In one embodiment of the invention, nucleic acid encoding a receptor construct is transformed into the cell. Exemplary techniques for transforming the cell with either the receptor or the receptor construct nucleic acid follow.

A. Transformation of the cells

The instant invention provides a substantial improvement over soluble kinase receptor assays insofar as it is considered to more accurately reflect the activity of the receptor *in situ*. It has been discovered that it is possible to transform eukaryotic cells with a receptor construct (comprising the kinase receptor and either an amino- or carboxyl-terminal flag polypeptide) so that the receptor construct assembles itself appropriately in the cell membrane and still retains tyrosine kinase

activity which can be detected in the ELISA stage of the assay. This provides a generic assay for measuring tyrosine kinase activity of any tyrosine kinase of interest.

If a suitable capture agent as described herein is available for a selected rPTK, cells can be transformed with the nucleic acid encoding the receptor alone, without the flag polypeptide. Alternatively, if cells are available which produce the receptor (*e.g.*, MCF-7 cells which produce the HER2 receptor), it is not necessary to transform the cells for use in the assay.

In order to transform the cells with the nucleic acid encoding the rPTK or receptor construct, nucleic acid encoding the rPTK and, optionally, the flag polypeptide, is isolated. This can be achieved by screening a cDNA or genomic library known to contain the DNA encoding the rPTK or flag polypeptide of interest with a selected labelled probe (*e.g.*, an antibody or oligonucleotide- probe) for the rPTK or flag polypeptide, using standard procedures as described in chapters 10-12 of Sambrook *et al.*, Molecular Cloning: A Laboratory Manual (New York: Cold Spring Harbor Laboratory Press, 1989), for example. Alternatively, the nucleic acid encoding the flag polypeptide can be made synthetically using an oligo-synthesizing machine (Applied Biosystems, CA). An alternative means to isolate the nucleic acid encoding the rPTK or flag polypeptide is to use PCR methodology as described in section 14 of Sambrook *et al.*, *supra*. Isolation of only the ECD of the rPTK of interest is required, since this nucleic acid can be fused to the nucleic acid encoding the TM and ICD of the Rse-flag polypeptide construct disclosed herein. See Figures 1A - 1C and SEQ ID NOS: 1 and 2. If necessary however, conventional primer extension procedures as described in section 7.79 of Sambrook *et al.*, *supra*, can be used to detect precursors and processing intermediates of mRNA that may not have been reverse-transcribed into cDNA.

A preferred method of practicing this invention is to use carefully selected oligonucleotide sequences to screen cDNA libraries from various tissues, preferably mammalian cell lines having the rPTK of interest. The oligonucleotide sequences selected as probes should be of sufficient length and sufficiently unambiguous that false positives are minimized.

The oligonucleotide must be labeled such that it can be detected upon hybridization to DNA in the library being screened. The preferred method of labeling is to use ^{32}P- labeled ATP with polynucleotide kinase, as is well known in the art, to radiolabel the oligonucleotide. However, other

methods may be used to label the oligonucleotide, including, but not limited to, biotinylation or enzyme labeling.

In order to provide nucleic acid encoding a receptor construct, nucleic acid encoding the rPTK is fused at its 3' end to nucleic acid encoding the N-terminus of the flag polypeptide. Alternatively, the nucleic acid encoding the rPTK will be fused at its 5' end to nucleic acid encoding the carboxyl terminus of the flag polypeptide. Thus, the flag polypeptide is provided at either the carboxyl- or amino- terminus of the receptor construct. Examples of suitable flag polypeptides are provided above. Selection of other suitable flag polypeptides is possible using the techniques described herein.

In order to generate fusions between the Rse.flag reagent and a rPTK of interest, the nucleic acid encoding the ECD of the rPTK of interest is fused at its 3' end to the nucleic acid encoding the amino terminus of the Rse.flag reagent.

The nucleic acid (*e.g.*, cDNA or genomic DNA) encoding the rPTK or receptor construct is then inserted into a replicable vector for further cloning (amplification of the DNA) or for expression. Many vectors are available to the skilled practitioner but must be compatible with the cell which is to be used in the assay. The vector will have vector components the presence of which will depend on various factors. Such components include, for example, a signal sequence, an origin of replication, one or more marker genes, an enhancer element, a promoter, and a transcription termination sequence. Selection of these vector components shall be described below.

Incorporation of a signal sequence into the expression vector is required since the rPTK or receptor construct must be transported to the cell membrane where it is positioned such that the ECD faces the external milieu of the cell. Therefore, a signal sequence suitable for positioning the rPTK or receptor construct in such a manner is used. The signal sequence is generally a component of the vector, or it may be a part of the rPTK or receptor construct DNA that is inserted into the vector. If a heterologous signal sequence is used, it is from those that are recognized and processed (*i.e.*, cleaved by a signal peptidase) by the host cell.

For yeast secretion the native signal sequence may be substituted by, *e.g.*, the yeast invertase leader, alpha factor leader (including *Saccharomyces* and *Kluyveromyces* α-factor leaders, the latter described in U.S. Patent No. 5,010,182 issued 23 April 1991), or acid phosphatase

leader, the *C. albicans* glucoamylase leader (EP 362,179 published 4 April 1990), or the signal described in WO 90/13646 published 15 November 1990. In mammalian cells expression of the DNA encoding the native signal sequence (*e.g.*, the rPTK pre-sequence that normally directs secretion of rPTK from mammalian cells *in vivo*) is satisfactory, although other mammalian signal sequences may be suitable, such as signal sequences from other animal rPTKs, and signal sequences from secreted polypeptides of the same or related species, as well as viral secretory leaders, for example, the herpes simplex gD signal.

The DNA for such precursor region is ligated in reading frame to DNA encoding the rPTK or receptor construct.

Both expression and cloning vectors contain a nucleic acid sequence that enables the vector to replicate in one or more selected host cells. Generally, in cloning vectors this sequence is one that enables the vector to replicate independently of the host chromosomal DNA, and includes origins of replication or autonomously replicating sequences. The 2μ plasmid origin is suitable for yeast, and various viral origins (SV40, polyoma, adenovirus, VSV or BPV) are useful for cloning vectors in mammalian cells. Generally, the origin of replication component is not needed for mammalian expression vectors (the SV40 origin may typically be used only because it contains the early promoter).

Most expression vectors are "shuttle" vectors, *i.e.*, they are capable of replication in at least one class of organisms but can be transformed into another organism for expression. For example, a vector is cloned in *E. coli* and then the same vector is transformed into yeast or mammalian cells for expression even though it is not capable of replicating independently of the host cell chromosome.

DNA may also be amplified by insertion into the host genome. This is readily accomplished using *Bacillus* species as hosts, for example, by including in the vector a DNA sequence that is complementary to a sequence found in *Bacillus* genomic DNA. Transformation of *Bacillus* with this vector results in homologous recombination with the genome and insertion of rPTK or receptor construct DNA. However, the recovery of genomic DNA encoding the rPTK or receptor construct is more complex than that of an exogenously replicated vector because restriction enzyme digestion is required to excise the rPTK or receptor construct DNA.

Expression and cloning vectors usually contain a selection gene, also termed a selectable marker. This gene encodes a protein necessary for the

survival or growth of transformed host cells grown in a selective culture medium. Host cells not transformed with the vector containing the selection gene will not survive in the culture medium. Typical selection genes encode proteins that (a) confer resistance to antibiotics or other toxins, *e.g.*, ampicillin, neomycin, methotrexate, or tetracycline, (b) complement auxotrophic deficiencies, or (c) supply critical nutrients not available from complex media.

One example of a selection scheme utilizes a drug to arrest growth of a host cell. Those cells that are successfully transformed with a heterologous gene express the DNA encoding a protein conferring drug resistance and thus survive the selection regimen. Examples of such dominant selection use the drugs neomycin (Southern *et al.*, J. Molec. Appl. Genet. 1:327 [1982]), mycophenolic acid (Mulligan *et al.*, Science 209:1422 [1980]) or hygromycin (Sugden *et al.*, Mol. Cell. Biol. 5:410-413 [1985]). The three examples given above employ bacterial genes under eukaryotic control to convey resistance to the appropriate drug G418 or neomycin (geneticin), xgpt (mycophenolic acid), or hygromycin, respectively.

Another example of suitable selectable markers for mammalian cells are those that enable the identification of cells competent to take up the rPTK or receptor construct nucleic acid, such as DHFR or thymidine kinase. The mammalian cell transformants are placed under selection pressure that only the transformants are uniquely adapted to survive by virtue of having taken up the marker. Selection pressure is imposed by culturing the transformants under conditions in which the concentration of selection agent in the medium is successively changed, thereby leading to amplification of both the selection gene and the DNA that encodes the rPTK or receptor construct. Amplification is the process by which genes in greater demand for the production of a protein critical for growth are reiterated in tandem within the chromosomes of successive generations of recombinant cells. Increased quantities of rPTK or receptor construct are synthesized from the amplified DNA. Other examples of amplifiable genes include metallothionein-I and -II, preferably primate metallothionein genes, adenosine deaminase, ornithine decarboxylase, *etc.*

For example, cells transformed with the DHFR selection gene are first identified by culturing all of the transformants in a culture medium that contains methotrexate (Mtx), a competitive antagonist of DHFR. An appropriate host cell when wild-type DHFR is employed is the Chinese hamster ovary (CHO) cell line deficient in DHFR activity, prepared and

propagated as described by Urlaub and Chasin, Proc. Natl. Acad. Sci. USA 77:4216 (1980). The transformed cells are then exposed to increased levels of methotrexate. This leads to the synthesis of multiple copies of the DHFR gene, and, concomitantly, multiple copies of other DNA comprising the expression vectors, such as the DNA encoding the rPTK or receptor construct. This amplification technique can be used with any otherwise suitable host, *e.g.*, ATCC No. CCL61 CHO-K1, notwithstanding the presence of endogenous DHFR if, for example, a mutant DHFR gene that is highly resistant to Mtx is employed (EP 117,060).

Alternatively, host cells (particularly wild-type hosts that contain endogenous DHFR) transformed or co-transformed with DNA sequences encoding the rPTK or receptor construct, wild-type DHFR protein, and another selectable marker such as aminoglycoside 3' phosphotransferase (APH) can be selected by cell growth in medium containing a selection agent for the selectable marker such as an aminoglycosidic antibiotic, *e.g.*, kanamycin, neomycin, or G418. See U.S. Patent No. 4,965,199.

A suitable selection gene for use in yeast is the *trp1* gene present in the yeast plasmid YRp7 (Stinchcomb *et al.*, Nature **282**:39 [1979]; Kingsman *et al.*, Gene **7**:141 [1979]; or Tschemper *et al.*, Gene **10**:157 [1980]). The *trp1* gene provides a selection marker for a mutant strain of yeast lacking the ability to grow in tryptophan, for example, ATCC No. 44076 or PEP4-1 (Jones, Genetics **85**:12 [1977]). The presence of the trp1 lesion in the yeast host cell genome then provides an effective environment for detecting transformation by growth in the absence of tryptophan. Similarly, *Leu2*-deficient yeast strains (ATCC 20,622 or 38,626) are complemented by known plasmids bearing the *Leu2* gene.

In addition, vectors derived from the 1.6 μm circular plasmid pKD1 can be used for transformation of *Kluyveromyces* yeasts. Bianchi *et al.*, Curr. Genet. **12**:185 (1987). More recently, an expression system for large-scale production of recombinant calf chymosin was reported for *K. lactis*. Van den Berg, Bio/Technology **8**:135 (1990). Stable multi-copy expression vectors for secretion of mature recombinant human serum albumin by industrial strains of *Kluyveromyces* have also been disclosed. Fleer *et al.*, Bio/Technology **9**:968-975 (1991).

Expression and cloning vectors usually contain a promoter that is recognized by the host organism and is operably linked to the rPTK or receptor construct nucleic acid. Promoters are untranslated sequences located upstream (5') to the start codon of a structural gene (generally

within about 100 to 1000 bp) that control the transcription and translation of particular nucleic acid sequence, such as the rPTK nucleic acid sequence, to which they are operably linked. Such promoters typically fall into two classes, inducible and constitutive. Inducible promoters are promoters that initiate increased levels of transcription from DNA under their control in response to some change in culture conditions, *e.g.*, the presence or absence of a nutrient or a change in temperature. At this time a large number of promoters recognized by a variety of potential host cells are well known. These promoters are operably linked to rPTK or receptor construct-encoding DNA by removing the promoter from the source DNA by restriction enzyme digestion and inserting the isolated promoter sequence into the vector. Both the native rPTK promoter sequence and many heterologous promoters may be used to direct amplification and/or expression of the rPTK or receptor construct DNA. The promoter will be one which results in the accumulation of suitable numbers of receptor or receptor construct in the cell membrane of the transformed cell (*i.e.* so that autophosphorylation of the receptor is detectable in the ELISA but constitutive phosphorylation does not occur). Selection of a suitable promoter to achieve this is possible following the guidelines herein for selecting cells for use in the KIRA ELISA.

Virtually all eukaryotic genes have an AT-rich region located approximately 25 to 30 bases upstream from the site where transcription is initiated. Another sequence found 70 to 80 bases upstream from the start of transcription of many genes is a CXCAAT region where X may be any nucleotide. At the 3' end of most eukaryotic genes is an AATAAA sequence that may be the signal for addition of the poly A tail to the 3' end of the coding sequence. All of these sequences are suitably inserted into eukaryotic expression vectors.

Examples of suitable promoting sequences for use with yeast hosts include the promoters for 3-phosphoglycerate kinase (Hitzeman *et al.*, J. Biol. Chem. **255**:2073 [1980]) or other glycolytic enzymes (Hess *et al.*, J. Adv. Enzyme Reg., 7:149 [1968]; and Holland, Biochemistry 17:4900 [1978]), such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase.

Other yeast promoters, which are inducible promoters having the additional advantage of transcription controlled by growth conditions, are

the promoter regions for alcohol dehydrogenase 2, isocytochrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Suitable vectors and promoters for use in yeast expression are further described in Hitzeman *et al.*, EP 73,657A. Yeast enhancers also are advantageously used with yeast promoters.

rPTK or receptor construct transcription from vectors in mammalian host cells is controlled, for example, by promoters obtained from the genomes of viruses such as polyoma virus, fowlpox virus (UK 2,211,504 published 5 July 1989), adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis-B virus and most preferably Simian Virus 40 (SV40), from heterologous mammalian promoters, *e.g.*, the actin promoter or an immunoglobulin promoter, from heat-shock promoters, and from the promoter normally associated with the rPTK or receptor construct sequence, provided such promoters are compatible with the host cell systems.

The early and late promoters of the SV40 virus are conveniently obtained as an SV40 restriction fragment that also contains the SV40 viral origin of replication. Fiers *et al.*, Nature **273**:113 (1978); Mulligan and Berg, Science **209**:1422-1427 (1980); Pavlakis *et al.*, Proc. Natl. Acad. Sci. USA **78**:7398-7402 (1981). The immediate early promoter of the human cytomegalovirus is conveniently obtained as a HindIII E restriction fragment. Greenaway *et al.*, Gene **18**:355-360 (1982). A system for expressing DNA in mammalian hosts using the bovine papilloma virus as a vector is disclosed in U.S. 4,419,446. A modification of this system is described in U.S. 4,601,978. See also Gray *et al.*, Nature **295**:503-508 (1982) on expressing cDNA encoding immune interferon in monkey cells; Reyes *et al.*, Nature **297**:598-601 (1982) on expression of human β-interferon cDNA in mouse cells under the control of a thymidine kinase promoter from herpes simplex virus; Canaani and Berg, Proc. Natl. Acad. Sci. USA **79**:5166-5170 (1982) on expression of the human interferon β1 gene in cultured mouse and rabbit cells; and Gorman *et al.*, Proc. Natl. Acad. Sci. USA **79**:6777-6781 (1982) on expression of bacterial CAT sequences in CV-1 monkey kidney cells, chicken embryo fibroblasts, Chinese hamster ovary cells, HeLa cells, and mouse NIH-3T3 cells using the Rous sarcoma virus long terminal repeat as a promoter.

Transcription of DNA encoding the rPTK or receptor construct by higher eukaryotes may be increased, if increased numbers of the rPTK or receptor construct per cell are required to facilitate detection in the ELISA stage of the assay. This may be achieved by inserting an enhancer sequence into the vector. Enhancers are cis-acting elements of DNA, usually about from 10 to 300 bp, that act on a promoter to increase its transcription. Enhancers are relatively orientation and position independent, having been found 5' (Laimins *et al.*, Proc. Natl. Acad. Sci. USA **78**:993 [1981]) and 3' (Lusky *et al.*, Mol. Cell Bio. **3**:1108 [1983]) to the transcription unit, within an intron (Banerji *et al.*, Cell **33**:729 [1983]), as well as within the coding sequence itself (Osborne *et al.*, Mol. Cell Bio. **4**:1293 [1984]). Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin, α-fetoprotein, and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. See also Yaniv, Nature **297**:17-18 (1982) on enhancing elements for activation of eukaryotic promoters. The enhancer may be spliced into the vector at a position 5' or 3' to the rPTK or receptor construct-encoding sequence, but is preferably located at a site 5' from the promoter.

Expression vectors used in eukaryotic host cells will also contain sequences necessary for the termination of transcription and for stabilizing the mRNA. Such sequences are commonly available from the 5' and, occasionally 3', untranslated regions of eukaryotic or viral DNAs or cDNAs. These regions contain nucleotide segments transcribed as polyadenylated fragments in the untranslated portion of the mRNA encoding the rPTK or receptor construct.

Construction of suitable vectors containing one or more of the above listed components employs standard ligation techniques. Isolated plasmids or DNA fragments are cleaved, tailored, and religated in the form desired to generate the plasmids required.

For analysis to confirm correct sequences in plasmids constructed, the ligation mixtures are used to transform *E. coli* K12 strain 294 (ATCC 31,446) and successful transformants selected by ampicillin or tetracycline resistance where appropriate. Plasmids from the transformants are prepared, analyzed by restriction endonuclease digestion, and/or sequenced

by the method of Messing *et al.*, Nucleic Acids Res. 9:309 (1981) or by the method of Maxam *et al.*, Methods in Enzymology 65:499 (1980).

Other methods, vectors, and host cells suitable for adaptation to the synthesis of the rPTK or receptor construct in recombinant vertebrate cell culture are described in Gething *et al.*, Nature 293:620-625 (1981); Mantei *et al.*, Nature 281:40-46 (1979); Levinson *et al.*; EP 117,060; and EP 117,058. A particularly useful plasmid for mammalian cell culture expression of rPTK or receptor construct DNA is pRK5 (EP 307,247) or pSVI6B (PCT pub. no. WO 91/08291 published 13 June 1991).

Examples of suitable eukaryotic cell lines for transformation include *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe* (Beach and Nurse, Nature 290:140 [1981]; EP 139,383 published 2 May 1985); *Kluyveromyces* hosts (U.S. 4,943,529; Fleer *et al.*, Bio/Technology 9:968-975 [1991]) and *Aspergillus* hosts such as *A. nidulans* (Ballance *et al.*, Biochem. Biophys. Res. Commun. 112:284-289 [1983]; Tilburn *et al.*, Gene 26:205-221 [1983]; Yelton *et al.*, Proc. Natl. Acad. Sci. USA 81:1470-1474 [1984]) and *A. niger* (Kelly and Hynes, EMBO J. 4:475-479 [1985]), among lower eukaryotic host microorganisms.

Examples of useful animal host cell lines for transformation include monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, Graham *et al.*, J. Gen Virol. 36:59 [1977]); baby hamster kidney cells (BHK, ATCC CCL 10); Chinese hamster ovary cells/-DHFR (CHO, Urlaub and Chasin, Proc. Natl. Acad. Sci. USA 77:4216 [1980]); mouse sertoli cells (TM4, Mather, Biol. Reprod. 23:243-251 [1980]); monkey kidney cells (CV1 ATCC CCL 70); African green monkey kidney cells (VERO-76, ATCC CRL-1587); human cervical carcinoma cells (HELA, ATCC CCL 2); canine kidney cells (MDCK, ATCC CCL 34); buffalo rat liver cells (BRL 3A, ATCC CRL 1442); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); mouse mammary tumor (MMT 060562, ATCC CCL51); TRI cells (Mather *et al.*, Annals N.Y. Acad. Sci. 383:44-68 [1982]); MRC 5 cells; FS4 cells; and a human hepatoma line (Hep G2).

Host cells are transformed with the above-described expression or cloning vectors of this invention and cultured in conventional nutrient media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences. Transformation means introducing DNA into an organism so that the DNA is replicable, either as an extrachromosomal element or as a chromosomal

integrant. Depending on the host cell used, transformation is done using standard techniques appropriate to such cells. Successful transformation is generally recognized when any indication of the operation of this vector occurs within the host cell.

For mammalian cells, the calcium phosphate precipitation method of Graham and Van der Eb, Virology **52**:456-457 (1978) is preferred. General aspects of mammalian cell host system transformations have been described by Axel in U.S. 4,399,216 issued 16 August 1983. Transformations into yeast are typically carried out according to the method of Van Solingen *et al.*, J. Bact. **130**:946 (1977) and Hsiao *et al.*, Proc. Natl. Acad. Sci. (USA) **76**:3829 (1979). However, other methods for introducing DNA into cells, such as by nuclear microinjection, electroporation, bacterial protoplast fusion with intact cells, or polycations, *e.g.*, polybrene, polyornithine, *etc.*, may also be used. For various techniques for transforming mammalian cells, see Keown *et al.*, Methods in Enzymology (1989), Keown *et al.*, Methods in Enzymology **185**:527-537 (1990), and Mansour *et al.*, Nature **336**:348-352 (1988).

The mammalian host cells used to produce the rPTK or receptor construct may be cultured in a variety of media. Commercially available media such as Ham's F10 (Sigma), Minimal Essential Medium ([MEM], Sigma), RPMI-1640 (Sigma), and Dulbecco's Modified Eagle's Medium ([DMEM], Sigma) are suitable for culturing the host cells. In addition, any of the media described in Ham and Wallace, Meth. Enz. **58**:44 (1979), Barnes and Sato, Anal. Biochem. **102**:255 (1980), U.S. 4,767,704; 4,657,866; 4,927,762; or 4,560,655; WO 90/03430; WO 87/00195; U.S. Patent Re. 30,985; or U.S. Patent No. 5,122,469, the disclosures of each of which are incorporated herein by reference, may be used as culture media for the host cells. Any of these media may be supplemented as necessary with hormones and/or other growth factors (such as insulin, transferrin, or epidermal growth factor), salts (such as sodium chloride, calcium, magnesium, and phosphate), buffers (such as HEPES), nucleosides (such as adenosine and thymidine), antibiotics (such as Gentamycin™ drug), trace elements (defined as inorganic compounds usually present at final concentrations in the micromolar range), and glucose or an equivalent energy source. Any other necessary supplements may also be included at appropriate concentrations that would be known to those skilled in the art. The culture conditions, such as temperature, pH, and the like, are those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan.

In general, principles, protocols, and practical techniques for maximizing the productivity of mammalian cell cultures can be found in Mammalian Cell Biotechnology: a Practical Approach, M. Butler, ed., IRL Press, 1991.

Gene amplification and/or expression may be measured in a sample directly, for example, by conventional Southern blotting, Northern blotting to quantitate the transcription of mRNA (Thomas, Proc. Natl. Acad. Sci. USA 77:5201-5205 [1980]), dot blotting (DNA analysis), or *in situ* hybridization, using an appropriately labeled probe, based on the sequences provided herein. Various labels may be employed, most commonly radioisotopes, particularly ^{32}P. However, other techniques may also be employed, such as using biotin-modified nucleotides for introduction into a polynucleotide. The biotin then serves as the site for binding to avidin or antibodies, which may be labeled with a wide variety of labels, such as radionuclides, fluorophores, enzymes, or the like. Alternatively, antibodies may be employed that can recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. The antibodies in turn may be labeled and the assay may be carried out where the duplex is bound to a surface, so that upon the formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Gene expression, alternatively, may be measured by immunological methods, such as immunohistochemical staining to quantitate directly the expression of gene product.

B. Selecting cells for use in the assay

As mentioned above, the cells to be subjected to the assay can be (a) cells having an endogenous receptor, (b) cells which have been transformed with a rPTK, or (c) cells transformed with a receptor construct. The suitability of the cells for use in the assay is investigated.

Cells having the endogenous rPTK can be subjected to a test-run KIRA ELISA using a known ligand to the PTK (*e.g.* an agonist antibody) and a control (*e.g.* the diluent for the agonist antibody). A range of ligand concentrations such as those used herein (see Examples 1, 2 and 3) will be used to determine whether sufficient numbers of the receptor are present in the cells being tested. In order to discover whether a cell line is unsuitable because the receptor is constitutively phosphorylated, the cell line can be subjected to the KIRA ELISA disclosed herein, wherein it is exposed to both positive and negative controls (*e.g.* a known agonist ligand

in cell culture media as described herein as a positive control and the cell culture media without the agonist ligand as the negative control). If phosphorylation of the receptor is detected for both positive and negative controls, this may be indicative that constitutive phosphorylation of the receptor is occurring. However, it is possible that a constituent of the serum in the cell culture media is activating the receptor. Thus, the cells can be "starved" in serum-free media for about 2-12 hours (depending on cell survival) and then the assay is repeated using the positive and negative controls. If activation is detected for both controls, the cell line may be considered unsuitable and another cell line can be tested.

If the cell line is transformed with the receptor (without the flag polypeptide) a strategy similar to that depicted in Figure 4 can be used to discover whether or not the cell line is suitable for use in the assay. As a first step, successful transformation and expression of the nucleic acid encoding the rPTK is determined (see Figure 4, step b). In order to identify whether the ECD of the rPTK is present on the surface of the cells, flow cytometric analysis can be performed using an antibody to the ECD of the receptor. The antibody can be made using the techniques for generating antibodies discussed herein. Flow cytometric analysis can be carried out using the techniques described in Current Protocols in Immunology, Ed. Coligen *et al.*, Wiley publishers, Vols. 1 and 2, for example. Briefly, flow cytometric analysis involves incubating intact cells (having the receptor) with antibodies to the ECD thereof, followed by washing. The antibody-bound cells are then incubated with species specific anti-antibody antibodies conjugated to a fluorochrome. Following washing, the labeled cells are analyzed by fluorescence-activated flow cytometry to detect whether the ECD is present on the surface of the cells.

In the following step, *i.e.* Figure 4, step (c), the ability of the cell-bound receptor to be activated is tested. In order to determine this, the transformed cells are exposed to a known agonist to the receptor (*e.g.* the endogenous ligand or an agonist antibody for the receptor). Following exposure, the cells are lysed in a suitable buffer (*e.g.* sodium dodecylbenzenesulfonate in phosphate buffered saline; SDS in PBS) and subjected to Western blotting with anti-phosphotyrosine antibodies as described in Wang, Molecular and Cellular Biology **5(12)**:3640-3643 (1985); Glenney *et al.*, Journal of Immunological Methods **109**:277-285 (1988); Kamps, Methods in Enzymology **201**:101-110 (1991); Kozma *et al.*, Methods in

Enzymology 201:28-43 (1991); Holmes *et al.*, Science 256:1205-10 (1992); or Corfas *et al.*, PNAS, USA 90:1624-1628 (1993), for example.

Assuming the Western blotting step indicates that the rPTK can be activated, a KIRA ELISA test run can be performed, see Figure 4 step (d), to further establish whether or not the transformed cell line can be used in the assay.

In the preferred embodiment of the invention, the KIRA ELISA is a "generic" assay insofar as any rPTK of interest can be studied regardless of the availability of receptor-specific reagents (*i.e.*, capture agent). This embodiment employs a receptor construct having a flag polypeptide at either the amino or carboxyl terminus of the receptor.

If the flag polypeptide is provided at the NH_2-terminus (see, *e.g.*, the gD.trk A, B and C receptor constructs disclosed in Example 3), the procedure for selecting a transformed cell line for use in the assay summarized in Figure 4 can be performed. In this embodiment, the cells are transformed with the flag polypeptide-receptor construct as described earlier herein. See step (a). In step (b), successful transformation of the receptor and flag polypeptide (*i.e.* the receptor construct) is confirmed. In order to study this, two-dimensional flow cytometric analysis can be performed using antibodies to both the flag polypeptide and the ECD of the receptor. Techniques for two-dimensional flow cytometric analysis are disclosed in Current Protocols in Immunology, *supra*. Assuming successful transformation of the receptor construct is demonstrated, steps (c) and (d) of Figure 4 are then performed. See the discussion above, for an explanation of steps (c) to (d) of Figure 4.

A technique for identification of cells which have been successfully transformed with the receptor construct having a C-terminal flag polypeptide and which cells are also suitable for use in the assay is illustrated in Figure 5. Following cell transformation [step (a)], successful transformation of the receptor is determined by flow cytometric analysis using an antibody directed against the ECD of the receptor of interest, for example. Flow cytometric analysis can be performed substantially as described above. This forms step (b) of the procedure outlined in Figure 5.

Following step (b), successful transformation of the entire receptor construct (including the COOH-terminal flag polypeptide) is analyzed in step (c). This can be achieved by lysing the cells (using techniques for lysing cells disclosed herein) and immunoprecipitating the membrane extract

with an antibody against the receptor of interest. This immunoprecipitated membrane extract is then subjected to Western blot analysis with antibodies specific for the flag polypeptide. Alternatively, rPTK-specific ELISA analysis of anti-flag polypeptide captured membrane lysate can be carried out. Briefly, this involves coating ELISA wells with appropriate flag specific capture agent. The wells are blocked, washed, and the lysate is then incubated in the wells. Unbound receptor construct is removed by washing. The wells are then reacted with receptor-specific antibody or antibodies, either directly or indirectly conjugated to HRPO. The wells are washed and the HRPO is then exposed to the chromogenic substrate (*e.g.*, TMB).

Steps (d) and (e), *i.e.*, detecting receptor activation and KIRA ELISA test run, are essentially the same as those steps described above.

Once useful cells are identified, they are subjected to the KIRA stage of the instantly claimed assay.

C. Coating the first solid phase with the cells

The first solid phase (*e.g.* a well of a first assay plate) is coated with cells having the endogenous receptor or cells which have been transformed pursuant to the preceding sections.

Preferably, an adherent cell line is chosen, so that the cells naturally adhere to the first solid phase. However, use of an adherent cell line is not essential. For example, non-adherent cells (*e.g.* red blood cells) can be added to round bottomed wells of an assay plate such as that sold by Becton Dickinson Labware, Lincoln Park, New Jersey, for example. The assay plate is then placed in a plate carrier and centrifuged so as to create a pellet of cells adhering to the base of the wells. The cell culture supernatants are removed using a pipette. Thus, use of an adherent cell is clearly advantageous over non-adherent cells since it reduces variability in the assay (*i.e*, the cells in the pellet of the round bottom wells may be taken up with the supernatant when the alternative method is used).

The cells to be added to the wells of the first assay plate may be maintained in tissue culture flasks and utilized when cells densities of about 70-90% confluency are achieved. Then, generally between about 1×10^4 to 3×10^5 (and preferably 5×10^4 to 1×10^5) cells are seeded per flat-bottom well, using a pipette, for example. It has been found that, contrary to expectations, addition of cell concentrations mentioned above is sufficient to enable activation of the rPTK to be measured in the ELISA

stage of the assay, without the need to concentrate or clarify the cells or cell lysate prior thereto. Often, the cells are diluted in culture medium prior to seeding them in the wells of the microtiter plate to achieve the desired cell densities.

Usually, the cells are cultured in the microtiter plates for a sufficient period of time to optimize adherence to the wells thereof, but not too long such that the cells begin to deteriorate. Thus, incubation for about 8 to 16 hours at a temperature which is the physiological optimum for the cells (usually about 37°C) is preferred. Suitable media for culturing the cells are described in Section 1A above. Culturing in 5% CO_2 is recommended.

Following incubation overnight, the well supernatants are decanted and excess supernatant may be further removed by lightly tamping the microtiter plates with an absorbent substrate, *e.g.*, a paper towel, but a sponge works equally well. Thus, a substantially homogeneous layer of adhering cells remains on the internal surfaces of the individual wells of the microtiter plate. These adhering cells are then exposed to the analyte.

D. Preparation and addition of the analyte

As mentioned above, the analyte may comprise an agonist ligand (or suspected agonist) or an antagonist (or suspected antagonist) for the rPTK of interest. The ligand may be an endogenous polypeptide, or a synthetic molecule, such as an inorganic or organic molecule. Usually, the ligand is a polypeptide. This assay is useful for screening molecules which activate (or antagonize activation) of the tyrosine kinase receptor of interest. Thus, the assay can be used for developing therapeutically effective molecules.

Where the ligand is an agonist, the molecule can comprise the native growth factor *e.g.*, heregulin (HRG), insulin, insulin-like growth factors I and II (IGF-I and IGF-II), epidermal growth factor (EGF), interleukins (*e.g.*, IL-8), macrophage colony-stimulating factor (M-CSF), erythropoietin (EPO), platelet-derived growth factor (PDGF), transforming growth factors alpha and beta (TGF-α and TGF-β), hepatocyte growth factor (HGF), fibroblast growth factor (FGF) and nerve growth factor (NGF). Many of these growth factors are available commercially. Alternatively, the growth factor can be made by peptide synthesis or recombinant techniques which are described herein. Synthetic small molecule agonists can similarly be

generated by those skilled in the art using conventional chemical synthesis techniques.

Where the ligand is present in a biological fluid, the analyte can be prepared using techniques which are well known in the art. Body fluid such as blood or amniotic fluid may be used directly, however concentration may be required. If the analyte to be tested comprises a particular tissue, the cells thereof can be grown in cell culture and the supernatant can be tested for secreted ligand.

Often, the ligand is diluted in an aqueous diluent (such as cell culture media) so that a standard curve can be generated. However, the ligand may be present in a cell or a cell component (*e.g.*, the cell membrane). In particular, it has been found that the assay can be used to detect the presence of a ligand in the cell membrane of a selected cell line. This is clearly useful for discovering a novel endogenous ligand for a known rPTK.

The ligand composition is added to each well which contains the adhering cells using a pipette, for example. At least one control well (*e.g.* to which the aqueous diluent for the ligand is added) is included in the assay.

The adhering cells are usually stimulated for a sufficient period of time to optimize the signal, but not too long such that the signal decreases as a consequence of dephosphorylation of the rPTK by endogenous phosphatases. A suitable stimulation period is between about 10 to 60 minutes, preferably about 30 minutes at a physiologically optimal temperature for the cells (usually about 37°C).

Following activation, well supernatants are decanted and the plates can then be lightly tamped with an absorbent substrate to remove excess supernatant.

The assay can be used to detect antagonist ligands for the rPTK of interest. Antagonists generally fall into two categories (a) ones which bind to the rPTK and thereby block binding and/or activation of the rPTK by an agonist thereto (the antagonist may bind to the ECD, but this is not necessarily the case) and (b) those which bind to the agonist and thus prevent activation of the rPTK by the agonist.

In order to detect antagonist molecules from category (a) above, the cells are exposed to the suspected antagonist ligand substantially as mentioned above. Following exposure to the antagonist, the well supernatants are decanted and the plates are lightly tamped. Then, a known

agonist (*e.g.*, the endogenous growth factor) is added to the washed cells essentially as discussed in the preceding paragraphs, following which, the well supernatants are decanted and plates are lightly tamped. Alternatively, a composition comprising both the antagonist and agonist can be added to the adhering cells substantially as discussed above. Ability of the suspected antagonist to block binding and/or activation of the rPTK can subsequently be measured by ELISA as discussed below.

To detect antagonist molecules from category (b) above, a known agonist is pre-incubated with the suspected antagonist prior to the KIRA stage of the assay. This incubation is carried out for a sufficient period of time to enable a complex of the antagonist-agonist to form; from 30 min. to 12 hours, for example. This complex is then subjected to the assay with the non-complexed agonist and antagonist used as controls.

Following exposure to the agonist (and optionally the antagonist) ligand, the cells are lysed, as discussed below.

E. Solubilizing the cells

In this step of the assay, the cells are lysed so as to solubilize the rPTK such that it remains activated (*i.e.*, the tyrosine residues remain phosphorylated) for the ELISA stage of the assay. Thus, the cells are lysed using a lysis buffer as described above which serves to solubilize the rPTK or receptor construct, yet does not dephosphorylate or denature the rPTK.

Where microtiter plates are used as mentioned above, about 75 to 200μl of lysis buffer is added to each well. The plates can then be agitated gently using a plate shaker (*e.g.*, such as that sold by Bellco Instruments, Vineland, NJ) for about 1 to 2 hours. Shaking can be carried out at room temperature.

2. Enzyme-Linked Immunosorbent Assay - ELISA

The second stage of the assay involves a sandwich ELISA performed in the second assay plate. In order to carry out the ELISA, a capture agent is prepared.

A. Preparation of the capture agent

As mentioned above, the capture agent often comprises a polyclonal antibody (usually an affinity purified polyclonal antibody) or monoclonal antibody. Other capture agents are envisaged and are discussed in the definitions section above. The capture agent either binds specifically to the kinase receptor, or to the flag polypeptide (*i.e.* the antigen).

Polyclonal antibodies to the antigen (either the receptor or the flag polypeptide) generally are raised in animals by multiple subcutaneous (sc)

or intraperitoneal (ip) injections of the antigen or an antigenic fragment thereof (often the ECD of the rPTK) and an adjuvant. It may be useful to conjugate the antigen or a fragment containing the target amino acid sequence to a protein that is immunogenic in the species to be immunized (*e.g.*, keyhole limpet hemocyanin), using a bifunctional or derivatizing agent.

The route and schedule for administration of immunogen to the host animal or cultured antibody-producing cells therefrom are generally in keeping with established and conventional techniques for antibody stimulation and production. While mice are frequently employed as the test model, it is contemplated that any mammalian subject including human subjects or antibody-producing cells obtained therefrom can be manipulated according to the processes of this invention to serve as the basis for production of mammalian, including human, hybrid cell lines.

Animals are typically immunized against the immunogenic conjugates or derivatives by combining 1 mg or 1 μg of conjugate (for rabbits or mice, respectively) with 3 volumes of Freund's complete adjuvant and injecting the solution intradermally at multiple sites. One month later the animals are boosted with 1/5 to 1/10 the original amount of conjugate in Freund's complete adjuvant (or other suitable adjuvant) by subcutaneous injection at multiple sites. 7 to 14 days later animals are bled and the serum is assayed for anti-antigen titer. Animals are boosted until the titer plateaus. Preferably, the animal is boosted with the conjugate of the same antigen, but conjugated to a different protein and/or through a different cross-linking agent. Conjugates also can be made in recombinant cell culture as protein fusions. Also, aggregating agents such as alum are used to enhance the immune response.

After immunization, monoclonal antibodies can be prepared by recovering immune cells (typically spleen cells or lymphocytes from lymph node tissue) from immunized animals and immortalizing the cells in conventional fashion, *e.g.*, by fusion with myeloma cells or by Epstein-Barr (EB)-virus transformation and screening for clones producing the desired antibody. The hybridoma technique described originally by Kohler and Milstein, Eur. J. Immunol. 6:511 (1976), and also described by Hammerling *et al.*, In: Monoclonal Antibodies and T-Cell Hybridomas, Elsevier, N.Y., pp. 563-681 (1981) has been widely applied to produce hybrid cell lines that secrete high levels of monoclonal antibodies against many specific antigens.

It is possible to fuse cells of one species with another. However, it is preferable that the source of the immunized antibody producing cells and the myeloma be from the same species.

The hybrid cell lines can be maintained in culture in cell culture media. The cell lines of this invention can be selected and/or maintained in a composition comprising the continuous cell line in hypoxanthine-aminopterin-thymidine (HAT) medium. In fact, once the hybridoma cell line is established, it can be maintained on a variety of nutritionally adequate media. Moreover, the hybrid cell lines can be stored and preserved in any number of conventional ways, including freezing and storage under liquid nitrogen. Frozen cell lines can be revived and cultured indefinitely with resumed synthesis and secretion of monoclonal antibody.

The secreted antibody is recovered from tissue culture supernatant by conventional methods such as precipitation, ion exchange chromatography, affinity chromatography, or the like. The antibodies described herein are also recovered from hybridoma cell cultures by conventional methods for purification of IgG or IgM, as the case may be, that heretofore have been used to purify these immunoglobulins from pooled plasma, *e.g.*, ethanol or polyethylene glycol precipitation procedures. The purified antibodies are then sterile filtered. Where the antibody is a polyclonal antibody, it is generally affinity purified using an affinity column generated from the antigen of interest so as to provide a substantially specific capture antibody. Affinity chromatography is usually preceded by other purification techniques, such as liquid chromatography.

In a further embodiment, antibodies or antibody fragments can be isolated from antibody phage libraries generated via the techniques described in McCafferty *et al.*, Nature, **348**:552-554 (1990), using the flag polypeptide, rPTK, or a fragment thereof, to select for a suitable antibody or antibody fragment. Clackson *et al.*, Nature, **352**:624-628 (1991) and Marks *et al.*, J. Mol. Biol., **222**:581-597 (1991) describe the isolation of murine and human antibodies, respectively, using phage libraries. Subsequent publications describe the production of high affinity (nM range) human antibodies by chain shuffling (Mark *et al.*, Bio/Technol. **10**:779-783 [1992]), as well as combinatorial infection and *in vivo* recombination as a strategy for constructing very large phage libraries (Waterhouse *et al.*, Nuc. Acids Res., **21**:2265-2266 [1993]). Thus, these techniques are viable alternatives to traditional monoclonal antibody hybridoma techniques for

isolation of "monoclonal" antibodies which are encompassed by the present invention.

DNA encoding the monoclonal antibodies of the invention is readily isolated and sequenced using conventional procedures (*e.g.*, by using oligonucleotide probes that are capable of binding specifically to genes encoding the heavy and light chains of murine antibodies). The hybridoma cells of the invention serve as a preferred source of such DNA. Once isolated, the DNA may be placed into expression vectors, which are then transfected into host cells such as simian COS cells, Chinese hamster ovary (CHO) cells, or myeloma cells that do not otherwise produce immunoglobulin protein, to obtain the synthesis of monoclonal antibodies in the recombinant host cells. The DNA also may be modified, for example, by substituting the coding sequence for human heavy and light chain constant domains in place of the homologous murine sequences, Morrison *et al.*, Proc. Nat. Acad. Sci. **81**, 6851 (1984), or by covalently joining to the immunoglobulin coding sequence all or part of the coding sequence for a non-immunoglobulin polypeptide. In that manner, "chimeric" or "hybrid" antibodies are prepared that have the binding specificity of an anti-rPTK or anti-flag polypeptide monoclonal antibody herein. Thus, the antibody may be made by recombinant DNA methods (Cabilly *et al.*, U.S. Pat. No. 4,816,567).

Binding of the capture agent is not affected by the presence or absence of a ligand bound to the receptor and the capture agent does not sterically block access to the phosphorylated tyrosine(s) by the anti-phosphotyrosine antibody. Furthermore, the capture agent does not, of course, activate the receptor of interest. In order to screen for an antibody having these characteristics, the procedure outlined in Figure 3 can be carried out.

First, once the capture agent (*e.g.* an antibody or streptavidin) has been chosen, binding to either the receptor or the flag polypeptide (where a receptor construct is to be used in the assay) is confirmed. This can be determined by flow cytometric analysis, immuno-precipitation or antigen-coat ELISA, for example. Flow cytometric analysis has been described above. Immunoprecipitation usually involves lysing the cells (having the receptor or receptor construct) in non-ionic detergent (*e.g.* 0.5% Triton X-100) in a suitable buffer (*e.g.* PBS) and the cell lysates thus obtained are then incubated with the potential anti-receptor or anti-flag polypeptide capture agent. The immune complexes are precipitated with

either (a) anti-capture agent antibodies in the presence of polyethylene glycol (PEG) which enhances precipitation of the immune complex or with (b) insoluble (*e.g.* agarose bound) protein A or protein G. The immunoprecipitated material is then analyzed by polyacrylamide gel electrophoresis (PAGE). For antigen-coat ELISA, ELISA wells are coated overnight with either the purified receptor, purified flag polypeptide or purified receptor construct. The coated wells are then exposed to the potential capture agent and screened with HRPO-conjugated species specific anti-capture agent antibody.

The ability of the capture agent to bind to the receptor or flag polypeptide in the presence of a ligand to the receptor is also confirmed. This can be analyzed by incubating the receptor or receptor construct with a known ligand for the receptor (*e.g.* the endogenous growth factor or an agonist antibody thereto). Flow cytometric analysis, immunoprecipitation or antigen-coat ELISA can then be performed substantially as described above to investigate binding of the capture agent.

Assuming the capture agent is suitable as determined by the preceding two steps, it is then shown that the capture agent does not induce receptor activation (*i.e.* autophosphorylation) either before or after cell lysis. Thus, the cell-bound receptor or receptor construct is exposed to either the potential capture agent or a negative control (*e.g.* a control antibody which does not activate the receptor). Following cell lysis, the receptor or receptor construct can be subjected to Western blot analysis using labeled anti-phosphotyrosine antibodies. See, *e.g.*, Glenney *et al.*, Journal of Immunological Methods **109**:277-285 (1988); Kamps, Methods in Enzymology **201**:101-110 (1991); Kozma *et al.*, Methods in Enzymology **201**:28-43 (1991); or Holmes *et al.*, Science **256**:1205-10 (1992). To establish whether the capture agent induces receptor activation following cell lysis, a trial run of the KIRA ELISA (with both the capture agent and a negative control as discussed above) can be performed.

Finally, the ability of an anti-phosphotyrosine antibody (*e.g.* biotinylated anti-phosphotyrosine antibody) to bind the activated receptor in the presence of the potential capture agent is confirmed by a trial run in the KIRA ELISA disclosed herein.

Assuming the capture agent meets all the criteria specified above, it has good potential for use in the KIRA ELISA.

Once a suitable capture agent has been prepared, the second solid phase is coated therewith. Between about 0.1 to 10 μg/ml of capture agent

can be added to each well of the second assay plate using a pipette, for example. The capture agent is often provided in a buffer at a high pH (*e.g.*, between about 7.5 to 9.6) so that it has an increased overall charge and therefore displays enhanced binding to the second assay plate. Usually, the capture agent will be incubated in the wells for between about 8 to 72 hours to enable a sufficient coating of the capture agent to form on the inside walls of the wells. This incubation is generally carried out at low temperatures (*e.g.*, between about 3-8°C) to avoid or reduce degradation of the capture agent.

Following incubation, the wells of the plate are decanted and tamped lightly with an absorbent substrate. Non-specific binding is then blocked. In order to achieve this, a block buffer, is added to the wells. For example, a block buffer containing bovine serum albumin (BSA) such as that sold by Intergen Company, Purchase, NY, is suitable. It has been found that addition of between about 100 to 200μl of block buffer to each well followed by gentle agitation at room temperature for between about 1-2 hours is sufficient to block non-specific binding. It is also possible to add the block buffer directly to the cell lysate obtained in the previous step rather than to the second assay plate.

Following this, the capture agent-coated plates are washed several times (usually between about 3-8 times) with a wash buffer. The wash buffer can comprise phosphate buffered saline (PBS) at pH 7.0 to 7.5, for example. However, other wash buffers are available which can also be used. Conveniently, an automated plate washer, such as the ScanWasher 300 (Skatron Instruments, Inc., Sterling, VA) can be used for this, and other, washing steps of the assay.

B. Measuring tyrosine phosphorylation

The activated, solubilized rPTK (or receptor construct) is then added to the wells having the capture agent adhering thereto. As a general proposition, about 80% of cell lysate obtained as mentioned under Section 1E above can be added to each well (*i.e.*, about 60 to 160 μl depending on the original volume of the wells). The lysate is incubated with the capture agent for an adequate period of time to enable the rPTK to be captured in the wells, *e.g.*, from 1 to 3 hours. Incubation can be carried out at room temperature.

Unbound cell lysate is then removed by washing with wash buffer. Following this washing step, an amount of the anti-phosphotyrosine antibody which is equal to, or less than, the amount of block buffer added

previously, is added to each well. For example, about 50 to 200μl of an anti-phosphotyrosine antibody preparation having between about 0.3 to 0.5 μg/ml of antibody in a suitable buffer (*e.g.*, PBS with a detergent such as those included in the lysis buffer) is added to the well. This is followed by a washing step to remove unbound anti-phosphotyrosine antibody.

Tyrosine phosphorylation is then quantified by the amount of anti-phosphotyrosine antibody binding to the second solid phase. Many systems for detecting the presence of an antibody are available to those skilled in the art. Some examples follow.

Generally, the anti-phosphotyrosine antibody will be labelled either directly or indirectly with a detectable label. Numerous labels are available which can be generally grouped into the following categories:

(a) Radioisotopes, such as ^{35}S, ^{14}C, ^{125}I, ^{3}H, and ^{131}I. The antibody can be labeled with the radioisotope using the techniques described in Current Protocols in Immunology, *supra*, for example and radioactivity can be measured using scintillation counting.

(b) Fluorescent labels such as rare earth chelates (europium chelates) or fluorescein and its derivatives, rhodamine and its derivatives, dansyl, Lissamine, phycoerythrin and Texas Red are available. The fluorescent labels can be conjugated to the antibody using the techniques disclosed in Current Protocols in Immunology, *supra*, for example. Fluorescence can be quantified using a fluorimeter (Dynatech).

(c) Various enzyme-substrate labels are available and U.S. Patent No. 4,275,149 provides a review of some of these. The enzyme generally catalyses a chemical alteration of the chromogenic substrate which can be measured using various techniques. For example, the enzyme may catalyze a color change in a substrate, which can be measured spectrophotometrically. Alternatively, the enzyme may alter the fluorescence or chemiluminescence of the substrate. Techniques for quantifying a change in fluorescence are described above. The chemiluminescent substrate becomes electronically excited by a chemical reaction and may then emit light which can be measured (using a Dynatech ML3000 chemiluminometer, for example) or donates energy to a fluorescent acceptor. Examples of enzymatic labels include luciferases (*e.g.*, firefly luciferase and bacterial luciferase; U.S. Patent No. 4,737,456), luciferin, 2,3-dihydrophthalazinediones, malate dehydrogenase, urease, peroxidase such as horseradish peroxidase (HRPO), alkaline phosphatase, β-galactosidase, glucoamylase, lysozyme, saccharide oxidases (*e.g.*, glucose oxidase, galactose oxidase, and glucose-6-phosphate

dehydrogenase), heterocyclic oxidases (such as uricase and xanthine oxidase), lactoperoxidase, microperoxidase, and the like. Techniques for conjugating enzymes to antibodies are described in O'Sullivan *et al.*, Methods for the Preparation of Enzyme-Antibody Conjugates for use in Enzyme Immunoassay, in Methods in Enzym. (ed J. Langone & H. Van Vunakis), Academic press, New York, 73: 147-166 (1981) and Current Protocols in Immunology, *supra*.

Examples of enzyme-substrate combinations include, for example:

(i) Horseradish peroxidase (HRPO) with hydrogen peroxidase as a substrate, wherein the hydrogen peroxidase oxidizes a dye precursor (*e.g.* orthophenylene diamine [OPD] or 3,3',5,5'-tetramethyl benzidine hydrochloride [TMB]).

(ii) alkaline phosphatase (AP) with para-Nitrophenyl phosphate as chromogenic substrate.

(iii) β-D-galactosidase (β-D-Gal) with a chromogenic substrate (*e.g.* p-nitrophenyl-β-D-galactosidase) or fluorogenic substrate 4-methylumbelliferyl-β-D-galactosidase.

Numerous other enzyme-substrate combinations are available to those skilled in the art. For a general review of these, see U.S. Patent Nos. 4,275,149 and 4,318,980.

Sometimes, the label is indirectly conjugated with the antibody. The skilled artisan will be aware of various techniques for achieving this. For example, the antibody can be conjugated with biotin and any of the three broad categories of labels mentioned above can be conjugated with avidin, or *vice versa*. Biotin binds selectively to avidin and thus, the label can be conjugated with the antibody in this indirect manner. See, Current Protocols in Immunology, *supra*, for a review of techniques involving biotin-avidin conjugation. Alternatively, to achieve indirect conjugation of the label with the antibody, the antibody is conjugated with a small hapten (*e.g.* digoxin) and one of the different types of labels mentioned above is conjugated with an anti-hapten antibody (*e.g.* anti-digoxin antibody). Thus, indirect conjugation of the label with the antibody can be achieved.

In another embodiment of the invention, the anti-phosphotyrosine antibody need not be labeled, and the presence thereof can be detected using a labeled anti-antiphosphotyrosine antibody (*e.g.* anti-mouse anti-phosphotyrosine antibody conjugated with HRPO).

In the preferred embodiment, the anti-phosphotyrosine antibody is labeled with an enzymatic label which catalyzes a color change of a substrate (such as tetramethyl benzimidine [TMB], or orthaphenylene diamine [OPD]). Thus, the use of radioactive materials is avoided. A color change of the reagent can be determined spectrophotometrically at a suitable wavelength (*e.g.* 450nm for TMB and 490nm for OPD, with a reference wavelength of 650 nm).

3. Intracellular Kinase Activity

The assay described herein is also useful for measuring phosphorylation and/or activation of intracellular kinases (*e.g.* cytoplasmic tyrosine kinases and/or cytoplasmic serine-threonine kinases). Phosphorylation of these molecules can occur as a consequence of trans-phosphorylation of the intracellular kinase by a kinase receptor or "receptor complex" (which comprises one or more kinase receptors residing in a cell membrane). Examples of intracellular tyrosine kinases include insulin receptor substrate I (IRS-1), Shc, Ras and GRB2, for example. Antibodies to human Shc, human Ras and GRB2 can be obtained commercially from UBI, NY, which can be used as capture agents for these tyrosine kinases. Examples of intracellular serine-threonine kinases include MEK and MAPK.

In order to measure phosphorylation of these kinases, the procedure is essentially as described above except that a chimera of the intracellular kinase and the flag polypeptide is normally formed (*i.e.* a "kinase construct"). Alternatively, the cell has an endogenous intracellular kinase or is transformed with nucleic acid encoding an intracellular kinase of interest. Generally, a eukaryotic cell will be transformed with nucleic acid encoding a kinase construct. Upon expression of the nucleic acid, the kinase or kinase construct will reside intracellularly (*i.e.* in the cytoplasm). The cells comprising the kinase or kinase construct are subjected to the KIRA as discussed above. Exposure to the agonist may result in trans-phosphorylation of the intracellular kinase which can be quantified in the ELISA as elaborated above. The capture agent in the ELISA binds to either the intracellular kinase or to the flag polypeptide.

4. Serine-Threonine Kinase Activity

This assay is further useful for measuring phosphorylation and/or activation of serine-threonine kinases. The term "serine-threonine kinase" refers to a kinase which phosphorylates a substrate which has at least one

phosphate accepting alcohol group. The serine-threonine kinase is usually a "receptor" insofar as it has a ligand-binding ECD, TM domain and ICD. The ICD usually comprises a catalytic kinase domain and generally has one or more phosphate accepting serine and/or threonine residues. Examples of intracellular serine-threonine kinases include MEK and MAPK. See section 3 above for a discussion as to measuring phosphorylation of intracellular serine-threonine kinases. Examples of serine-threonine kinase receptors include *daf*-1, activin type II receptor (ActR-II), activin type IIB receptor (ActR-IIB), TGF-β type II receptor (TβR-II), activin receptor-like kinase (ALK) -1, -2, -3, -4 and TGF-β type I receptor (TβR-1)/ALK-5. See ten Dijke *et al.*, *supra*. The serine-threonine kinase assay is essentially the same as described above for tyrosine kinases, except that phosphorylation is quantified using anti-phosphoserine and/or anti-phosphothreonine antibodies. Anti-phosphoserine and anti-phosphothreonine monoclonal antibodies can be purchased from Sigma Immuno Chemicals, St Louis, MO, for example.

5. Phosphatase Activity

Phosphatase activity can similarly be measured using the assay described herein. Phosphatase enzymes are able to dephosphorylate phosphorylated tyrosine, serine and/or threonine residues (*i.e.* liberate inorganic phosphate from phosphoric esters of such amino acid residues). Generally the phosphatase enzyme is specific for either tyrosine residues or serine-threonine residues but sometimes can dephosphorylate tyrosine, serine and threonine residues. Sometimes "endogenous" phosphatase activity is measured and this refers to the activity of phosphatase enzyme(s) which exist in nature in a selected cell.

In order to quantify endogenous phosphatase activity, cells possessing at least one phosphatase are stimulated in the presence and absence of one or more phosphatase inhibitors. Examples of protein tyrosine phosphatase (PTPase) inhibitors include sodium orthovanadate and sodium molybdate (Sigma Chemical Co., St. Louis, MO). ICN Biochemicals supply okadaic acid which is a serine-threonine phosphatase inhibitor. As a general proposition, between about 1-10μM phosphatase inhibitor can be added to each well of the assay plate. In all other respects, the assay is performed essentially as discussed above. Thus, the ability of endogenous phosphatases to dephosphorylate a kinase in the selected cell can be quantified.

In the preferred embodiment, a phosphatase enzyme of interest can be studied. Examples of protein tyrosine phosphatases (PTPases) include PTP1B, PTPMEG, PTP1c, Yop51, VH1, cdc25, CD45, HLAR, PTP18, HPTPα and DPTP10D. See Zhang and Dixon, Adv. Enzym. 68: 1-36 (1994). Examples of protein serine-threonine phosphatases include PP1, PP2A, PP2B and PP2C. See Meth. Enzym., ed Hunter & Sefton, Academic press, New York, 201:389-398 (1991). These proteins can be purchased commercially or made using the recombinant techniques described herein. To measure phosphatase activity, the KIRA ELISA can be performed essentially as described above with the following modifications. Following capture of the kinase or kinase construct (*e.g.* receptor construct) to the second solid phase and the washing step (to remove unbound cell lysate), the phosphatase of interest is added to the wells of the second assay plate and incubated with the adhering kinase or kinase construct. For example, between about 50-200 μl of the phosphatase in a suitable dilution buffer (see Meth. Enzym., ed Hunter & Sefton, Academic press, New York, 201:416-440 [1991]) can be added to each well. This is generally followed by gentle agitation at room temperature (or 37°C) for between about 30 min to 2 hours to allow the phosphatase to dephosphorylate the kinase. Following washing to remove the phosphatase, the decreased degree of phosphorylation of the kinase relative to the control (*i.e.* no phosphatase added) is quantified by ELISA as described earlier herein.

6. Kits

As a matter of convenience, the reagents can be provided in a kit, *i.e.*, a packaged combination of reagents, for combination with the analyte in assaying the ability of the analyte to activate or prevent activation of a rPTK of interest. The components of the kit will be provided in predetermined ratios. Thus, a kit will comprise the specific second solid phase for the assay as well as the anti-flag polypeptide capture agent either packaged separately or captured to the second solid phase (*e.g.* a microtiter plate). Usually, other reagents, such as the anti-phosphotyrosine antibody labelled directly or indirectly with an enzymatic label will also be provided in the kit. Where the detectable label is an enzyme, the kit will include substrates and cofactors required by the enzyme (*e.g.* a substrate precursor which provides the detectable chromophore or fluorophore). In addition, other additives may be included such as stabilizers, buffers (*e.g.* a block buffer and a lysis buffer) and the like. Conveniently, the kit can also supply the homogeneous population

of cells which have been transformed with the receptor construct. The relative amounts of the various reagents may be varied widely to provide for concentrations in solution of the reagents which substantially optimize the sensitivity of the assay. Particularly, the reagents may be provided as dry powders, usually lyophilized, including excipients which on dissolution will provide a reagent solution having the appropriate concentration. The kit also suitably includes instructions for carrying out the KIRA ELISA.

7. Uses for the Assay

This application provides two assays which are useful for reliable, sensitive and quantitative detection of kinase activation. The first assay can be used where a kinase receptor-specific capture antibody having the desired characteristics herein described is available or has been prepared. The second assay is a generic assay which enables activation of any kinase receptor to be measured via the use of a flag polypeptide and a capture agent which binds specificity thereto.

These assays are useful for identifying novel agonists/antagonists for a selected kinase receptor. Also, the assay provides a means for studying ligand-receptor interactions (*i.e.*, mechanism studies). Also the presence of an endogenous receptor in a selected cell line can be quantified using the assay. The assays are further useful for identifying the presence of a ligand for a selected kinase receptor in a biological sample and, *e.g.*, establishing whether a growth factor has been isolated following a purification procedure. It is desirable to have an assay for measuring the ability of these growth factors to activate their respective receptors.

The assay also has clinical applications for detecting the presence of a ligand for a selected rPTK (*e.g.* the insulin receptor) in a biological sample taken from a human and thus patients having elevated or depressed levels of the ligand can be identified. This is particularly desirable where elevated or depressed levels of the ligand cause a pathological condition. Accordingly, candidates for administration of the selected ligand (*e.g.* insulin) can be identified through this diagnostic method. It is possible, using the assay disclosed herein, to assay the pK of agonists or antagonists administered to a patient. This assay also facilitates the detection of shed receptor in a biological sample.

The assay is also useful to quantify phosphatase activity of endogenous phosphatases or, in the preferred embodiment, a phosphatase of

interest. This can be used for screening phosphatase inhibitors, for example.

Below are examples of specific embodiments for carrying out the present invention. The examples are offered for illustrative purposes only, and are not intended to limit the scope of the present invention in any way.

All publications, patents and patent applications cited herein, whether *supra* or *infra*, are hereby incorporated by reference in their entirety.

EXAMPLE 1

KIRA ELISA of the HER2 Receptor

The assay system described in this example was developed to measure the extent of autophosphorylation as a result of the interactions between the HER2 receptor and its specific activator, heregulin (HRG). The overexpression of $p185^{HER2}$ has been correlated with poor clinical outcome in a number of epithelial-derived cancers. Heregulin and its rodent homologue, *neu* differentiation factor (NDF), were originally purified based on their ability to stimulate the autophosphorylation of a 185 kDa protein in the breast carcinoma cell lines MCF-7 and MDA-453, respectively. In this embodiment of the invention, the cell line expressing the tyrosine kinase receptor DNA (either endogenous or transformed) is adherent and there is an antibody (*e.g.* monoclonal or affinity purified polyclonal) capable of specifically binding the receptor such that it neither stimulates autophosphorylation in the absence of ligand nor suffers impaired binding due to the presence of bound ligand. Standard curve preparations and many samples may easily be run simultaneously in replicate and at several dilutions using this assay, readily allowing quantitation of ligand activity in a large number of unknown samples.

(i) Capture agent preparation

Polyclonal anti-HER2 antibody was isolated from pooled immune sera from New Zealand White rabbits immunized with the extracellular domain of the HER2 molecule (Fendly *et al.*, Journal of Biological Response Modifiers 9:449-455 [1990]). The rHER2 ECD specific antibodies were affinity purified using an FPLC (Pharmacia Biotech, Inc, Piscataway, NJ) with an affinity column generated from rHER2 ECD conjugated to Avidgel F (Bioprobe International, Inc, Tustin, CA). The resulting purified antibody stock was

0.829 mg/ml in phosphate buffered saline (PBS), pH 7.4, and was stored as 0.5 ml aliquots at -20°C.

(ii) Anti-phosphotyrosine antibody preparation

Monoclonal anti-phosphotyrosine, clone 4G10, was purchased from Upstate Biologicals, Inc (UBI, Lake Placid, NY) and biotinylated using long-arm biotin-N-hydroxysuccinamide (Biotin-X-NHS, Research Organics, Cleveland, OH).

(iii) Ligand

The recombinant truncated form of β1heregulin (MW= 7.88 Kd) corresponding to residues 177-244 ($HRG\beta1_{177-244}$) was produced in *E. coli* and purified to homogeneity as described in Holmes *et al.*, Science, **256**: 1205-1210 (1992) and was stored at 4°C as an 89.7 μM stock solution in 50 mM Tris/HCl, pH 7.5.

(iv) Adherent Cells

MCF-7 (ATCC-HTB 22), an adherent cell line isolated from a human breast adenocarcinoma, was obtained from American Type Culture Collection (ATCC, Rockville, MD). MCF-7 cells have been shown to produce measurable levels of surface $p185^{HER2}$ by both FACS and ELISA analysis. The cells were maintained in 150 cm^2 tissue culture flasks (Corning Inc, Corning, NY) and utilized when at cell densities of 60 % to 75 % confluency. For the assay, 2 x 10^5 cells were seeded per well in flat-bottom microtiter plates (Falcon 3072, Becton Dickinson Labware, Lincoln Park, NJ) cultured overnight at 37°C in 5 % CO_2. Cells were grown in F12/DMEM 50:50 Gibco as a custom formulation (Gibco/BRL, Life Technologies, Grand Island, NY). The medium was supplemented with 10 % FBS (HyClone, Logan, Utah), 25 mM HEPES (Gibco) and 2 mM L-glutamine (Gibco).

(v) KIRA ELISA

MCF-7 cells (2 x 10^5) in 100 μl media were added to each well in a flat-bottom-96 well culture plate and cultured overnight at 37°C in 5% CO_2. The following morning the well supernatants were decanted, and the plates were lightly tamped on a paper towel. 50 μl of media containing either experimental samples or the recombinant $HRG\beta1_{177-244}$ standards (3000, 1000, 333, 111, 37, 12, 4, and 0 pM) was then added to each well. The cells were stimulated at 37°C for 30 min., the well supernatants were decanted, and the plates were once again lightly tamped on a paper towel. To lyse the cells and solubilize the receptors, 100 μl of lysis buffer was added to each well. Lysis buffer consisted of 150 mM NaCl containing 50 mM HEPES (Gibco), 0.5 % Triton-X 100 (Gibco), 0.01 % thimerosal, 30 KIU/ml aprotinin

(ICN Biochemicals, Aurora, OH), 1mM 4-(2-aminoethyl)-benzenesulfonyl fluoride hydrochloride (AEBSF; ICN Biochemicals), 50 μM leupeptin (ICN Biochemicals), and 2 mM sodium orthovanadate (Na_3VO_4, Sigma Chemical Co, St. Louis, MO), pH 7.5. The plate was then agitated gently on a plate shaker (Bellco Instruments, Vineland, NJ) for 60 min. at room temperature.

While the cells were being solubilized, an ELISA microtiter plate (Nunc Maxisorp, Inter Med, Denmark) coated overnight at 4°C with the affinity-purified polyclonal anti-HER2 ECD (1.0 μg/ml in 50 mM carbonate buffer, pH 9.6, 100 μl/well) was decanted, tamped on a paper towel and blocked with 150 μl/well of Block Buffer [PBS containing 0.5 % BSA (Intergen Company, Purchase, NY) and 0.01 % thimerosal] for 60 min. at room temperature with gentle agitation. After 60 minutes, the anti-HER2 ECD coated plate was washed 6 times with wash buffer (PBS containing 0.05 % Tween-20 and 0.01 % thimerosal) using an automated plate washer (ScanWasher 300, Skatron Instruments, Inc, Sterling, VA).

The lysate containing solubilized $p185^{HER2}$ from the cell-culture microtiter well was transferred (85 μl/well) to anti-rHER2 ECD coated and blocked ELISA well and was incubated for 2 h at room temperature with gentle agitation. The unbound receptor was removed by washing with wash buffer and 100 μl of biotinylated 4G10 (anti-phosphotyrosine) diluted 1:2000 in dilution buffer (PBS containing 0.5 % BSA, 0.05 % Tween-20, 5 mM EDTA, and 0.01 % thimerosal), *i.e.* 400pg/ml, was added to each well. After incubation for 2 h at room temperature the plate was washed and 100 μl of HRPO-conjugated streptavidin (Zymed Laboratories, S. San Francisco, CA) diluted 1:10000 in dilution buffer was added to each well. The plate was incubated for 30 minutes at room temperature with gentle agitation. The free avidin-conjugate was washed away and 100 μl freshly prepared substrate solution (tetramethyl benzidine [TMB]; 2-component substrate kit; Kirkegaard and Perry, Gaithersburg, MD) was added to each well. The reaction was allowed to proceed for 10 minutes, after which the color development was stopped by the addition of 100 μl/well 1.0 M H_3PO_4. The absorbance at 450 nm was read with a reference wavelength of 650 nm ($ABS_{450/650}$), using a *vmax* plate reader (Molecular Devices, Palo Alto, CA) controlled with a Macintosh Centris 650 (Apple Computers, Cupertino, CA) and DeltaSoft software (BioMetallics, Inc, Princeton, NJ).

The standard curve shown in Figure 7 was generated by stimulating MCF-7 cells with 3000, 1000, 333, 111, 37, 12, 4, or 0 pM $HRG\beta1_{177-244}$ and presented as pM $HRG\beta1_{177-244}$ vs. mean $ABS_{450/650}$ ± sd using the DeltaSoft

program. Sample concentrations were obtained by interpolation of their absorbance on the standard curve and are expressed in terms of pM $HRG\beta1_{177-244}$ activity.

When the data were fitted to a 4-parameter nonlinear least squares equation, they resulted in a correlation coefficient of 0.9998. For the data shown in Figure 7, the EC_{50} of receptor activation by $HRG\beta1_{177-244}$ was 373 pM. To demonstrate the highly reproducible nature of the $p185^{HER2}$ KIRA ELISA, seven standard curves were generated over the period of one month and the EC_{50}'s are averaged. This gives an $EC_{50}ave$ for $HRG\beta1_{177-244}$ of 360 ± 40 pM (average ± SD).

(vi) Intra- and inter-assay precision and assay specificity

The intra-assay variability was determined by performing the $p185^{HER2}$ KIRA ELISA on three separate days. For each test, the standard curve is run in triplicate. Controls with $HRG\beta1_{177-244}$ corresponding to high (1000 pM), mid (200 pM) and low (40 pM) were assayed in 24 replicates. The $ABS_{450/650}$ of the individual test samples were converted to pM $HRG\beta1_{177-244}$ activity and the 24 converted values for each test concentration were averaged. The data are expressed as *averaged value* and *% coefficient of variation (%cv;* [(intra-assay standard deviation/intra-assay averaged calculated value) x 100]. See Table 1A below.

Table 1

Intra- and Inter-assay Variation

A. Intra-assay Precision (n-24 per test)

	High Value[a]		Mid Value		Low Value	
	Average Value (pM)	% cv[b]	Average Value (pM)	% cv	Average Value (pM)	% cv
Test #1	1256	19.5%	209	10.8%	33	12.3%
Test #2	1078	10.0%	196	5.1%	38	7.5%
Test #3	999	14.3%	196	6.3%	35	11.3%

B. Inter-assay Precision (n=3)

Average Value (pM)	%cv[c]	Average Value (pM)	% cv	Average Value (pM)	% cv
1100	4.3%	200	6.3%	34	9.0%

[a]Expected high value: 1000 pM; mid value: 200 pM; low value: 40 pM
[b]Intra-assay % cv determined as intra-assay sd/intra-assay average x 100
[c]Inter-assay % cv determined as inter-assay sd/inter-assay average x 100

The intra-assay variability of the KIRA ELISA was within acceptable limits despite the fact that the assay actually consists of both bioassay and ELISA components. The coefficients of variance (%) for the highest values were under 20% and for the mid and low values were at or under 10%.

The inter-assay variability was determined by averaging the values from upper-most three adjacent wells (of the 24 wells run) for a given sample concentration from each run. The three separate averages for each test concentration were then averaged. The data were expressed as *averaged value* and *%cv* [(inter-assay standard deviation/inter-assay averaged value) x 100]. See Table 1B. above. The inter-assay variability of the KIRA ELISA was within acceptable limits.

In order to confirm the specificity of the assay, MCF-7 cells were stimulated with either $HRG\beta1_{177-244}$ at 3000, 1000, 333, 111, 37, 12, 4 or 0 pM or insulin-like growth factor-1 (IGF-1), epidermal growth factor (EGF), vascular endothelial growth factor (VEGF), or insulin at 30000, 10000, 3333, 1111, 370, 120, 40 or 0 pM. The $p185^{HER2}$ KIRA ELISA was then performed as described above. The results are depicted in Figure 8.

The $p185^{HER2}$ KIRA ELISA was clearly specific for heregulin. While $HRG\beta1_{177-244}$ induced normal receptor stimulation and autophosphorylation, the closely related EGF gives only a slight stimulation ($OD_{450/650}$ = 0.239) at the highest concentration tested (100 nM). Since EGF-R is produced in MCF-7 cells, this signal is likely due to EGF receptor transphosphorylation of $p185^{HER2}$. Neither insulin-like growth factor-1 (IGF-1), vascular endothelial growth factor (VEGF) nor insulin have any detectable effect on the MCF-7 $p185^{HER2}$ KIRA ELISA, the latter despite the fact that MCF-7 cells produce active insulin receptors.

The results presented in this example demonstrate that the KIRA ELISA is a useful method for assaying ligand activation of a kinase receptor, *e.g.*, heregulin activation of the $p185^{HER2}$ receptor. Levels of receptor activation in terms of tyrosine phosphorylation are easily quantified and an EC_{50} for a given ligand is readily determined. One potential use for this

assay would be to screen compounds for receptor agonist or antagonist activities. The potential throughput for this assay greatly surpasses that of Western blot analysis. Since the cell-culture portion of the assay is conducted in 96-well plates, many samples may be run in replicate at different dilutions at one time in a one-day assay.

EXAMPLE 2

KIRA ELISA of the Rse Receptor

Mark *et al.*, Journal of Biological Chemistry **269(14)**:10720-10728 (1994) describe isolation of the Rse receptor protein tyrosine kinase from human and murine tissues. This Rse receptor with a carboxyl-terminal flag polypeptide (*i.e.* Rse.gD) was subjected to the KIRA ELISA described herein. The experimental procedure is outlined below.

(i) Capture agent preparation

Monoclonal anti-gD (clone 5B6) was produced against a peptide from Herpes simplex virus glycoprotein D (Paborsky *et al.*, Protein Engineering **3(6)**:547-553 [1990]). The purified stock preparation was adjusted to 3.0mg/ml in phosphate buffered saline (PBS), pH 7.4 and 1.0ml aliquots were stored at -20°C.

(ii) Anti-phosphotyrosine antibody preparation

Monoclonal anti-phosphotyrosine, clone 4G10, was purchased from Upstate Biologicals, Inc (UBI, Lake Placid, NY) and biotinylated using long-arm biotin-N-hydroxysuccinamide (Biotin-X-NHS, Research Organics, Cleveland, OH).

(iii) Ligand

Since the endogenous ligand for the Rse receptor was not available, an agonist antibody for the Rse receptor was prepared which forms the ligand for the KIRA ELISA described in this Example. To generate the agonist antibody, a Rse.IgG chimera was generated. Briefly, the coding sequence of the ECD of Rse was fused to that of the human IgG-γ1 heavy chain in a multi-step process. PCR was used to generate a fragment with a unique BstEII site 3' to the coding sequences of the Rse amino acid 428. The PCR product was joined to the human IgG-γ_1 heavy chain cDNA through a unique BstEII site in that construct (Mark *et al.*, J. Cell. Biol., **267**: 26166-26171 [1992]). The resulting construct (termed pRK.bpTK3.IgG.fusion) contained the coding sequences for amino acids 375-428 of Rse joined to those encoding human IgG-γ_1 heavy chain. The remaining portion of the Rse

ECD (amino acids 1-374) was then added by linkage through the Bam HI site in pRK.bpTK3.IgG.fusion to yield pRK.Rse.IgG.

To generate stable cell populations expressing Rse.IgG, the cDNA encoding Rse.IgG was subcloned into the episomal CMV-driven expression plasmid pCIS.EBON, a pRK5 derivative disclosed in Cachianes *et al.*, Bio. Techniques, **15**: 225-259 (1993). Human fetal kidney 293 cells (obtained from ATCC, 12301 Parklawn Drive, Rockville, MD, USA) were transfected by the calcium phosphate technique. Cell monolayers were incubated for four hours in the presence of the DNA precipitate, glycerol shocked, and cultured in F12:DMEM (1:1) containing 2mM glutamine, 10% fetal bovine serum, penicillin and streptomycin. After 48 hours, populations were replated in media containing G418 to select for a stable population of cells. Conditioned media was collected from cells expressing Rse.IgG nucleic acid that have been cultured in serum-free media for 72 hours in the absence of G418.

Rse.IgG was purified by affinity chromatography on a protein A column using procedures as described by Chamow, S.M., *et al.*, Biochemistry, **29**:9885-9891 (1990) with the following minor modifications. Conditioned media collected from cells expressing the Rse.IgG was adjusted to 0.1 M citrate pH 6.0 and loaded directly onto a protein A column (Repligen). The column was washed with 0.1 M citrate, pH 6.0, and was eluted with 3 M $MgCl_2$ with 10% glycerol. Fractions were pooled and desalted on a PD-10 column, dialyzed and concentrated against PBS. Protein concentrations were determined by an ELISA against human IgG (Fc). The protein was analyzed for purity by Coomassie staining of PAGE gels.

Polyclonal antibodies were generated in New Zealand white rabbits against the Rse.IgG formed as described above. 4μg of Rse.IgG in 100μL PBS was emulsified with 100μL Freund's adjuvant (complete adjuvant for the primary injection and incomplete adjuvant for all boosts). For the primary immunization and the first boost, the protein was injected directly into the popliteal lymph nodes (Sigel *et al.*, Methods Enzymol., **93**, 3-12 [1983]). For subsequent boosts, the protein was injected into subcutaneous and intramuscular sites. 1.3 μg protein/kg body weight was injected every 3 weeks with bleeds taken 1 and 2 weeks following each boost. The polyclonal antisera generated was then precipitated in 50% ammonium sulphate.

The resultant, purified polyclonal antisera is called "19B" herein. To confirm the ability of the 19B antisera to induce autophosphorylation of the Rse receptor, serum starved 3T3.gD.R11 cells (transformed with

nucleic acid encoding the Rse receptor with an amino terminal gD flag polypeptide [*i.e.* gD.Rse] using the techniques described in Mark *et al.*, Journal of Biological Chemistry **269(14)**:10720-10728 [1994]) or NIH3T3 cells were exposed to pre-immune serum or 19B polyclonal antisera at a 1:200 dilution for 10 minutes. The gD.Rse protein was immunoprecipitated from extracts using the anti-gD monoclonal antibody 5B6. Proteins were fractionated on 7% SDS-PAGE under reducing conditions and transferred to nitrocellulose. Phosphorylation of Rse was detected with labelled anti-phosphotyrosine antibody. Treatment of the 3T3.gD.R11 cells with 19B antisera stimulated the phosphorylation of the 140kD gD.Rse protein. This increase was not observed in cells treated with pre-immune sera.

The purified 19B polyclonal antisera was stored at 4°C as an 2.8mg/ml stock solution in PBS, pH 7.5.

(iv) Preparation of Rse.gD nucleic acid

Synthetic double stranded oligonucleotides were used to reconstitute the coding sequence for the C-terminal 10 amino acids (880 - 890) of human Rse and add an additional 21 amino acids containing an epitope for the antibody 5B6 and a stop codon. The final sequence of the synthetic portion of the fusion gene was:

coding strand:

5'-TGCAGCAAGGGCTACTGCCACACTCGAGCTGCGCAGATGCTAGCCTCAAGATGGCT G ATCCAAATCGATTCCGCGGCAAAGATCTTCCGGTCCTGTAGAAGCT-3' (SEQ ID NO: 10)

noncoding (anti-sense) strand:

5'-AGCTTCTACAGGACCGGAAGATCTTTGCCGCGGAATCGATTTGGATCAGCCATCTT G AGGCTAGCATCTGCGCAGCTCGAGTGTGGCAGTAGCCCTTGCTGCA-3'(SEQ ID NO: 11).

The synthetic DNA was ligated with the cDNA encoding amino acids 1-880 of human Rse at the PstI site beginning at nucleotide 2644 of the published human Rse cDNA sequence (Mark *et al.*, Journal of Biological Chemistry **269(14)**:10720-10728 [1994]) and HindIII sites in the polylinker of the expression vector pSVI7.ID.LL (See Figure 16; SEQ ID NO: 9) to create the expression plasmid pSV.ID.Rse.gD. Briefly, the expression plasmid comprises a dicistronic primary transcript which contains sequence encoding DHFR bounded by 5' splice donor and 3' splice acceptor intron splice sites, followed by sequence that encodes the Rse.gD. The full length (non-spliced) message contains DHFR as the first open reading frame and therefore generates DHFR protein to allow selection of stable transformants.

(v) Cell transformation

dp12.CHO cells (EP 307,247 published 15 March 1989) were electroporated with 20 μgs of pSV.ID.Rse.gD which had been linearized at a unique NotI site in the plasmid backbone. The DNA was ethanol precipitated after phenol/chloroform extraction and was resuspended in 20μl 1/10 Tris EDTA. Then, 10μg of DNA was incubated with 10^7 CHO.dp12 cells in 1 ml of PBS on ice for 10 min. before electroporation at 400 volts and 330μf. Cells were returned to ice for 10 min. before being plated into non-selective medium. After 24 hours cells were fed nucleoside-free medium to select for stable DHFR+ clones.

(vi) Selection of transformed cells for use in the KIRA ELISA

To identify a cell line that expresses Rse.gD nucleic acid, candidate clones were screened by fluorescence activated cell sorting (FACS) analysis using the polyclonal antiserum 19B generated as described above, which recognizes epitopes in the extracellular domain of Rse. See Figure 5, step (b).

To confirm that clones that scored positive in the FACS assay express full-length Rse.gD nucleic acid, cell lysates were prepared (Lokker *et al.*, EMBO J, **11**:2503-2510 [1992]) and solubilized Rse.gD was immunoprecipitated with the 19B antisera. The immunoprecipitated proteins were fractionated under reducing conditions using 7% PAGE, blotted onto nitrocellulose and then probed with the anti-gD 5B6 antibody which was detected with a horseradish peroxidase conjugated anti-mouse IgG antibody. See Figure 5, step (c). The ability of Rse.gD in cell clones to be activated to undergo autophosphorylation in response to the 19B agonistic antibody was determined. Briefly, serum starved dp.CHO cells transformed with Rse.gD nucleic acid as described above were exposed to pre-immune or 19B antisera at a 1:200 dilution for 10 min. The Rse.gD protein was immunoprecipitated from extracts using the anti-gD 5B6 monoclonal antibody. Proteins were fractionated on 7% SDS-PAGE under reducing conditions and transferred to nitrocellulose. Phosphorylation of Rse was detected with labelled antiphosphotyrosine antibody. See Figure 5, step (d).

(vii) Media

Cells were grown in F12/DMEM 50:50 (Gibco/BRL, Life Technologies, Grand Island, NY). The media was supplemented with 10% diafiltered FBS (HyClone, Logan, Utah), 25mM HEPES and 2mM L-glutamine.

(viii) KIRA ELISA

Rse.gD transformed dp12.CHO cells (EP 307,247 published 15 March 1989) were seeded ($5x10^4$ per well) in the wells of a flat-bottom-96 well culture plate in 100 µl media and cultured overnight at 37°C in 5% CO_2. The following morning the well supernatants were decanted, and the plates were lightly tamped on a paper towel. 100µl of media containing either experimental samples or 1:100, 1:200, 1:400, 1:800, 1:1600, 1:3200 or 0 diluted, anti-Rse agonist polyclonal antibody (19B pAb) was then added to each well. The cells were stimulated at 37°C for 30 min., the well supernatants were decanted, and the plates were once again lightly tamped on a paper towel. To lyse the cells and solubilize the receptors, 100 µl of lysis buffer was added to each well. Lysis buffer consisted of 150 mM NaCl containing 50 mM HEPES (Gibco), 0.5 % Triton-X 100 (Gibco), 0.01 % thimerosal, 30 KIU/ml aprotinin (ICN Biochemicals, Aurora, OH), 1mM 4-(2-aminoethyl)-benzenesulfonyl fluoride hydrochloride (AEBSF; ICN Biochemicals), 50 µM leupeptin (ICN Biochemicals), and 2 mM sodium orthovanadate (Na_3VO_4; Sigma Chemical Co, St. Louis, MO), pH 7.5. The plate was then agitated gently on a plate shaker (Bellco Instruments, Vineland, NJ) for 60 min. at room temperature.

While the cells were being solubilized, an ELISA microtiter plate (Nunc Maxisorp, Inter Med, Denmark) coated overnight at 4°C with the 5B6 monoclonal anti-gD antibody (0.5 µg/ml in 50 mM carbonate buffer, pH 9.6, 100 µl/well) was decanted, tamped on a paper towel and blocked with 150 µl/well of Block Buffer [PBS containing 0.5 % BSA (Intergen Company, Purchase, NY) and 0.01 % thimerosal] for 60 min. at room temperature with gentle agitation. After 60 minutes, the anti-gD 5B6 coated plate was washed 6 times with wash buffer (PBS containing 0.05 % Tween-20 and 0.01 % thimerosal) using an automated plate washer (ScanWasher 300, Skatron Instruments, Inc, Sterling, VA).

The lysate containing solubilized Rse.gD from the cell-culture microtiter well was transferred (85 µl/well) to anti-gD 5B6 coated and blocked ELISA well and was incubated for 2 h at room temperature with gentle agitation. The unbound Rse.gD was removed by washing with wash buffer and 100 µl of biotinylated 4G10 (anti-phosphotyrosine) diluted 1:2000 in dilution buffer (PBS containing 0.5 % BSA, 0.05 % Tween-20, 5 mM EDTA, and 0.01 % thimerosal), *i.e.* 400 pg/ml was added to each well. After incubation for 2 h at room temperature the plate was washed and 100 µl of HRPO-conjugated streptavidin (Zymed Laboratories, S. San Francisco, CA)

diluted 1:10000 in dilution buffer was added to each well. The plate was incubated for 30 minutes at room temperature with gentle agitation. The free avidin-conjugate was washed away and 100 µl freshly prepared substrate solution (tetramethyl benzidine [TMB]; 2-component substrate kit; Kirkegaard and Perry, Gaithersburg, MD) was added to each well. The reaction was allowed to proceed for 10 minutes, after which the color development was stopped by the addition of 100 µl/well 1.0 M H_3PO_4. The absorbance at 450 nm was read with a reference wavelength of 650 nm ($ABS_{450/650}$), using a *vmax* plate reader (Molecular Devices, Palo Alto, CA) controlled with a Macintosh Centris 650 (Apple Computers, Cupertino, CA) and DeltaSoft software (BioMetallics, Inc, Princeton, NJ).

The standard curve shown in Figure 10 was generated by stimulating Rse.gD transformed CHO cells with 1:100, 1:200, 1:400, 1:800, 1:1600, 1:3200 or 0 diluted, anti-Rse agonist antibody (19B) and presented as 1/dilution anti-Rse agonist antibody (19B) vs. mean $ABS_{450/650}$ ± sd using the DeltaSoft program.

The results presented in this example demonstrate that the KIRA ELISA is a useful method for assaying ligand activation of a receptor construct having a carboxyl terminal flag polypeptide, *e.g.*, activation of Rse.gD. Levels of receptor activation in terms of tyrosine phosphorylation are easily quantified and an EC_{50} for a given ligand (*e.g.* an agonist antibody for the receptor) is readily determined.

EXAMPLE 3

KIRA ELISA of the trk A, B and C Receptors

Neurotrophins belong to a family of small, basic proteins which play a crucial role in the development and maintenance of the nervous system. The first identified and probably best understood member of this family is nerve growth factor (NGF). See U.S. Patent No. 5,169,762, issued December 8, 1992. Recently, sequentially related but distinct polypeptides with similar functions to NGF have been identified. For example, brain-derived neurotrophic factor (BDNF), now also referred to as neurotrophin-2 (NT2), was cloned and sequenced by Leibrock *et al.* (Nature, **341**: 149-152 [1989]). Several groups identified a neurotrophic factor originally called neuronal factor (NF), and now referred to as neurotrophin-3 (NT3). (Ernfors *et al.*, Proc. Natl. Acad. Sci. USA, **87**: 5454-5458 [1990]; Höhn *et al.*, Nature, **344**: 339 [1990]; Maisonpierre *et al.*, Science, **247**: 1446 [1990]; Rosenthal *et al.*, Neuron, **4**: 767 [1990]; Jones and Reichardt, Proc. Natl. Acad. Sci.

USA, 87: 8060-8064 [1990]; Kaisho *et al.*, FEBS Lett., 266: 187 [1990]). Neurotrophins-4 and -5 (NT4 and NT5) have been recently added to the family (Hallbook *et al.*, Neuron, 6: 845-858 [1991]; Berkmeier *et al.*, Neuron, 7: 857-866 [1991]; Ip *et al.*, Proc. Natl. Acad. Sci. USA, 89: 3060-3064 [1992]).

Neurotrophins, similarly to other polypeptide growth factors, affect their target cells through interactions with cell surface rPTKs (called Trk receptors). The first member of the trk receptor family, trkA, was initially identified as the result of an oncogenic transformation caused by the translocation of tropomyosin sequences onto its catalytic domain. Later work identified trkA as a signal transducing receptor for NGF. Subsequently, two other related receptors, mouse and rat trkB (Klein *et al.*, EMBO J., 8: 3701-3709 [1989]; Middlemas *et al.*, Mol. Cell. Biol., 11: 143-153 [1991]; EP 455,460 published 6 November 1991) and porcine, mouse and rat trkC (Lamballe *et al.*, Cell, 66: 967-979 [1991]; EP 522,530 published 13 January 1993), were identified as members of the trk receptor family. The structures of the trk receptors are quite similar, but alternate splicing increases the complexity of the family by giving rise to two known forms of trkA, three known forms of trkB (two without functional tyrosine kinase domains) and at least four forms of trkC (several without functional tyrosine kinase domain, and two with small inserts in the tyrosine kinase domain). Human trk A, B and C receptor sequences are disclosed in U.S. Patent application Serial No. 08/215,139, filed March 18, 1994, specifically incorporated herein by reference.

The following KIRA ELISA was performed using trk A, B and C receptor constructs having amino-terminal flag polypeptides.

(i) Capture agent preparation

Monoclonal anti-gD (clone 5B6) was produced against a peptide from Herpes simplex virus glycoprotein D as discussed above in Example 2. The purified stock preparation was adjusted to 3.0mg/ml in phosphate buffered saline (PBS), pH 7.4 and 1.0ml aliquots were stored at -20°C.

(ii) Anti-phosphotyrosine antibody preparation

Monoclonal anti-phosphotyrosine, clone 4G10, was purchased from Upstate Biologicals, Inc (UBI, Lake Placid, NY) and biotinylated using long-arm biotin-N-hydroxysuccinamide (Biotin-X-NHS, Research Organics, Cleveland, OH).

(iii) Ligands

Nerve growth factor (NGF), neurotrophin 3 (NT3), and neurotrophin 5 (NT5) were prepared by recombinant techniques using the sequence data provided for each of these proteins in the above-mentioned references. The purified NGF, NT3 and NT5 were stored at 4°C as stock solutions (180μM, 8.8μM and 26.9μM, respectively) in PBS, pH 7.5.

(iv) Preparation of gD.trk nucleic acid

In order to express the various trk receptors with gD flags (*i.e.* gD.trk constructs), DNA constructs were made which encoded the signal and epitope of gD (see Paborsky *et al.*, *supra*) fused to the amino terminus of the various trk receptors. These were made by inserting the trk receptor and gD sequences into pRK5 or pRK7 (Suva *et al.*, Science, **237**: 893-896 [1987]) using standard molecular biology techniques, to generate the constructs shown in Figures 12-14. In addition to the gD.trk constructs, constructs were also made to express gD tagged trk.IgG fusion proteins (*i.e.*, gD.trk.IgG). DNA constructs encoding the chimeras of trk extracellular domain and IgG-1 Fc domains were made with the Fc region clones of human IgG-1 (Ashkenazi *et al.*, Immunoadhesins Intern. Rev. Immunol., **10**: 219-227 [1993]). More specifically, the source of the IgG-1 encoding sequence was the CD4-IgG-1 expression plasmid $pRKCD4_2Fc_1$ (Capon *et al.*, Nature, **334**: 525 [1989]; Byrn *et al.*, Nature, **344**: 667 [1990]) containing a cDNA sequence encoding a hybrid polypeptide consisting of residues 1-180 of the mature human CD4 protein fused to human IgG-1 sequences beginning at aspartic acid 216 (taking amino acid 114 as the first residue of the heavy chain constant region; Kabat *et al.*, Sequences of Proteins of Immunological Interest 4th ed. [1987]), which is the first residue of the IgG-1 hinge after the cysteine residue involved in heavy-light chain bonding, and ending with residues 441 to include the CH2 and CH3 Fc domains of IgG-1. The CD4-encoding sequence was deleted from the expression plasmid $pRKCD4_2Fc_1$ and the vector was fused to DNA encoding the trk receptors, with the splice between aspartate 216 of the IgG-1 and valine 402 of trkA, threonine 422 of trkB, or threonine 413 of trkC. The gD tag was added to the amino terminus of each trk.IgG in the same way as for the gD.trk constructs.

(v) Cell transformation

Human embryonic kidney 293 cells (obtained from ATCC, Rockville, MD) were transiently transfected with the nucleic acid encoding gD.trk.IgG using a calcium phosphate protocol (Gorman, DNA Cloning: A Practical Approach [Glover, D., ed.] Vol II: 143-190, IRL Press, Washington DC).

After twelve hours, the transformed cells were rinsed three times with serum free F12/DMEM 50:50 media (Gibco) and then serum free media was added for a 48 hour collection.

Cell lines stably expressing each of the gD.trk constructs were made by co-transfecting dp12.CHO cells (EP 307,247 published 15 March 1989) with the pRK plasmids encoding the gD tagged trk receptors and a plasmid encoding DHFR, again using calcium phosphate mediated transfection.

The media mentioned above (having the gD.trk.IgG) was used without further purification in binding assays to assess the effects of the presence of the gD flag polypeptide on neurotrophin binding to the gD.trk.IgG polypeptides. DNA encoding untagged trk.IgG polypeptide was run in parallel as a control. trk.IgG and gD tagged trk.IgG containing cell supernatants were prepared as described and used in competitive displacement assays with the appropriate iodinated neurotrophin. NGF is used as ligand for trkA, NT5 is used as ligand for trkB, and NT3 is used as a ligand for trkC. A summary of the results obtained is shown in the following table.

TABLE 1

Binding of Neurotrophins to trk.IgG

	IC50 without gD	IC50 with gD
trkA	68.4+/-11.9 pM	68.8+/-3.0 pM
trkB	31.1+/-15.6 pM	12.1+/-18 pM
trkC	31.1+/-1.1 pM	30.2+/-0.7 pM

(vi) Selection of transformed cells for use in the KIRA ELISA

It was apparent from the preceding experiment that there was no observable change in the affinity of interaction of neurotrophins with their receptor due to the presence of the gD flag polypeptide on the amino terminus. Based on this result, cells were transformed with the gD.trk constructs for use in the KIRA ELISA using the techniques described in the previous section.

After two days, dp12.CHO cells (EP 307,247 published 15 March 1989) transformed with gD.trk constructs were selected for by growth in media without GHT, and after two weeks, growing cells were sorted by FACS analysis using the 5B6 monoclonal to select cells expressing the gD flag polypeptide on their surface. gD positive cells were cloned by plating at limiting dilution and resultant colonies were then rescreened by FACS

analysis (using the anti-gD 5B6 monoclonal antibody), neurotrophin binding (as discussed above), tyrosine phosphorylation indicated by Western blot using an anti-phosphotyrosine antibody, gD expression by Western blot using th anti-gD 5B6 antibody, and immunocytochemistry using the 5B6 antibody. Clones which were positive were then recloned by limiting dilution and were subjected to the KIRA ELISA as described below.

(vii) Media

Cells were grown in F12/DMEM 50:50 (Gibco/BRL, Life Technologies, Grand Island, NY). The media was supplemented with 10% diafiltered FBS (HyClone, Logan, Utah), 25mM HEPES and 2mM L-glutamine.

(viii) KIRA ELISA

gD.trk transformed dp12.CHO cells (EP 307,247 published 15 March 1989) were seeded (5 x 10^4 per well) in a flat-bottom-96 well culture plate in 100 μl media and cultured overnight at 37°C in 5% CO_2. The following morning the well supernatants were decanted, and the plates were lightly tamped on a paper towel. 100μl of media containing either experimental samples or the recombinant purified NGF, NT3, or NT5 standards (3000, 1000, 333, 111, 37, 12, 4, and 0 pM) was then added to each well. The cells were stimulated at 37°C for 30 min., the well supernatants were decanted, and the plates were once again lightly tamped on a paper towel. To lyse the cells and solubilize the receptors, 100 μl of lysis buffer was added to each well. Lysis buffer consisted of 150 mM NaCl containing 50 mM HEPES (Gibco), 0.5 % Triton-X 100 (Gibco), 0.01 % thimerosal, 30 KIU/ml aprotinin (ICN Biochemicals, Aurora, OH), 1mM 4-(2-aminoethyl)-benzenesulfonyl fluoride hydrochloride (AEBSF; ICN Biochemicals), 50 μM leupeptin (ICN Biochemicals), and 2 mM sodium orthovanadate (Na_3VO_4; Sigma Chemical Co, St. Louis, MO). pH 7.5. The plate was then agitated gently on a plate shaker (Bellco Instruments, Vineland, NJ) for 60 min. at room temperature.

While the cells were being solubilized, an ELISA microtiter plate (Nunc Maxisorp, Inter Med, Denmark) coated overnight at 4°C with the 5B6 monoclonal anti-gD antibody (0.5 μg/ml in 50 mM carbonate buffer, pH 9.6, 100 μl/well) was decanted, tamped on a paper towel and blocked with 150 μl/well of Block Buffer [PBS containing 0.5 % BSA (Intergen Company, Purchase, NY) and 0.01 % thimerosal] for 60 min. at room temperature with gentle agitation. After 60 minutes, the anti-gD 5B6 coated plate was washed 6 times with wash buffer (PBS containing 0.05 % Tween-20 and 0.01 % thimerosal) using an automated plate washer (ScanWasher 300, Skatron Instruments, Inc, Sterling, VA).

The lysate containing solubilized gD.trk from the cell-culture microtiter well was transferred (85 μl/well) to anti-gD 5B6 coated and blocked ELISA well and was incubated for 2 h at room temperature with gentle agitation. The unbound gD.trk was removed by washing with wash buffer and 100 μl of biotinylated 4G10 (anti-phosphotyrosine) diluted 1:2000 in dilution buffer (PBS containing 0.5 % BSA, 0.05 % Tween-20, 5 mM EDTA, and 0.01 % thimerosal), *i.e.*, 400pg/ml, was added to each well. After incubation for 2 h at room temperature the plate was washed and 100 μl of HRPO-conjugated streptavidin (Zymed Laboratories, S. San Francisco, CA) diluted 1:10000 in dilution buffer was added to each well. The plate was incubated for 30 minutes at room temperature with gentle agitation. The free avidin-conjugate was washed away and 100 μl freshly prepared substrate solution (tetramethyl benzidine; 2-component substrate kit; Kirkegaard and Perry, Gaithersburg, MD) was added to each well. The reaction was allowed to proceed for 10 minutes, after which the color development was stopped by the addition of 100 μl/well 1.0 M H_3PO_4. The absorbance at 450 nm was read with a reference wavelength of 650 nm ($ABS_{450/650}$), using a *vmax* plate reader (Molecular Devices, Palo Alto, CA) controlled with a Macintosh Centris 650 (Apple Computers, Cupertino, CA) and DeltaSoft software (BioMetallics, Inc, Princeton, NJ).

The standard curves shown in Figures 15A-15C were generated by stimulating gD.trk transformed CHO cells with 3000, 1000, 333, 111, 37, 12, 4, and 0 pM NGF, NT3 or NT5 and were presented as pM neurotrophin vs. mean $ABS_{450/650}$ ± sd using the DeltaSoft program. Sample concentrations were obtained by interpolation of their absorbance on the standard curve and are expressed in terms of pM neurotrophin activity.

The results presented in this example demonstrate that the KIRA ELISA is a useful method for assaying ligand activation of a receptor construct having an amino terminal flag polypeptide, *e.g.*, activation of gD.trk receptor constructs. Levels of receptor activation in terms of tyrosine phosphorylation are easily quantified and an EC_{50} for a given ligand is readily determined.

EXAMPLE 4

KIRA ELISA of the *MPL*/Rse Chimeric Receptor

The human *MPL* receptor has been disclosed by Vigon *et al.*, PNAS, USA 89:5640-5644 (1992). A chimeric receptor comprising the ECD of the *MPL* receptor and the TM and ICD of Rse (Mark *et al.*, *supra*) with a carboxyl-

terminal flag polypeptide (*i.e.* Rse.gD; see Example 2) was subjected to the KIRA ELISA described herein. The experimental procedure is outlined below. See also Figs. 16 and 17.

(i) Capture agent preparation

Monoclonal anti-gD (clone 5B6) was produced against a peptide from Herpes simplex virus glycoprotein D (Paborsky *et al.*, Protein Engineering 3(6):547-553 [1990]). The purified stock preparation was adjusted to 3.0mg/ml in phosphate buffered saline (PBS), pH 7.4 and 1.0ml aliquots were stored at -20°C.

(ii) Anti-phosphotyrosine antibody preparation

Monoclonal anti-phosphotyrosine, clone 4G10, was purchased from UBI (Lake Placid, NY) and biotinylated using long-arm biotin-N-hydroxysuccinamide (Biotin-X-NHS, Research Organics, Cleveland, OH).

(iii) Ligand

The *MPL* ligand [de Sauvage *et al.*, Nature 369: 533-538 (1994)] was prepared by recombinant techniques. The purified *MPL* ligand was stored at 4°C as a stock solution.

(iv) Preparation of *MPL*/Rse.gD nucleic acid

The expression plasmid pSV.ID.Rse.gD produced as described in Example 2 above was modified to produce plasmid pSV.ID.M.tmRd6 which contained the coding sequences of the ECD of human *MPL* (amino acids 1-491) fused to the transmembrane domain and intracellular domain of Rse.gD (amino acids 429-911). Synthetic oligonucleotides were used to join the coding sequence of a portion of the extracellular domain of human *MPL* to a portion of the Rse coding sequence in a two step PCR cloning reaction as described by Mark *et al.* in J. Biol. Chem. 267: 26166-26171 (1992). Primers used for the first PCR reaction were M1 (5'-TCTCGCTACCGTTTACAG - SEQ ID NO:12) and M2 (5'-CAGGTACCCACCAGGCGGTCTCGGT - SEQ ID NO: 13) with a *MPL* cDNA template and R1 (5'-GGGCCATGACACTGTCAA - SEQ ID NO: 14) and R2 (5'-GACCGCCACCGAGACCGCCTGGTGGGTACCTGTGGTCCTT - SEQ ID NO: 15) with a Rse cDNA template. The PvuII-SmaI portion of this fusion junction was used for the construction of the full-length chimeric receptor.

(v) Cell transformation

dp12.CHO cells (EP 307,247 published 15 March 1989) were electroporated with pSV.ID.M.tmRd6 which had been linearized at a unique NotI site in the plasmid backbone. The DNA was ethanol precipitated after phenol/chloroform extraction and was resuspended in 20μl 1/10 Tris EDTA. Then, 10μg of DNA was incubated with 10^7 CHO.dp12 cells in 1 ml of PBS on

ice for 10 min. before electroporation at 400 volts and 330μf. Cells were returned to ice for 10 min. before being plated into non-selective medium. After 24 hours cells were fed nucleoside-free medium to select for stable DHFR+ clones.

(vi) Selection of transformed cells for use in the KIRA ELISA

Clones expressing *MPL*/Rse.gD were identified by western-blotting of whole cell lysates post-fractionation by SDS-PAGE using the antibody 5B6 which detects the gD epitope tag.

(vii) Media

Cells were grown in F12/DMEM 50:50 (Gibco/BRL, Life Technologies, Grand Island, NY). The media was supplemented with 10% diafiltered FBS (HyClone, Logan, Utah), 25mM HEPES and 2mM L-glutamine.

(viii) KIRA ELISA

MPL/Rse.gD transformed dp12.CHO cells were seeded ($3x10^4$ per well) in the wells of a flat-bottom-96 well culture plate in 100 μl media and cultured overnight at 37°C in 5% CO_2. The following morning the well supernatants were decanted, and the plates were lightly tamped on a paper towel. 50μl of media containing either experimental samples or 200, 50, 12.5, 3.12, 0.78, 0.19, 0.048 or 0 ng/ml *MPL* ligand was then added to each well. The cells were stimulated at 37°C for 30 min., the well supernatants were decanted, and the plates were once again lightly tamped on a paper towel. To lyse the cells and solubilize the chimeric receptors, 100 μl of lysis buffer was added to each well. Lysis buffer consisted of 150 mM NaCl containing 50 mM HEPES (Gibco), 0.5 % Triton-X 100 (Gibco), 0.01 % thimerosal, 30 KIU/ml aprotinin (ICN Biochemicals, Aurora, OH), 1mM 4-(2-aminoethyl)-benzenesulfonyl fluoride hydrochloride (AEBSF; ICN Biochemicals), 50 μM leupeptin (ICN Biochemicals), and 2 mM sodium orthovanadate (Na_3VO_4; Sigma Chemical Co, St. Louis, MO), pH 7.5. The plate was then agitated gently on a plate shaker (Bellco Instruments, Vineland, NJ) for 60 min. at room temperature.

While the cells were being solubilized, an ELISA microtiter plate (Nunc Maxisorp, Inter Med, Denmark) coated overnight at 4°C with the 5B6 monoclonal anti-gD antibody (5.0 μg/ml in 50 mM carbonate buffer, pH 9.6, 100 μl/well) was decanted, tamped on a paper towel and blocked with 150 μl/well of Block Buffer [PBS containing 0.5 % BSA (Intergen Company, Purchase, NY) and 0.01 % thimerosal] for 60 min. at room temperature with gentle agitation. After 60 minutes, the anti-gD 5B6 coated plate was washed 6 times with wash buffer (PBS containing 0.05 % Tween-20 and 0.01 %

thimerosal) using an automated plate washer (ScanWasher 300, Skatron Instruments, Inc, Sterling, VA).

The lysate containing solubilized *MPL*/Rse.gD from the cell-culture microtiter well was transferred (85 μl/well) to anti-gD 5B6 coated and blocked ELISA well and was incubated for 2 h at room temperature with gentle agitation. The unbound *MPL*/Rse.gD was removed by washing with wash buffer and 100 μl of biotinylated 4G10 (anti-phosphotyrosine) diluted 1:18000 in dilution buffer (PBS containing 0.5 % BSA, 0.05 % Tween-20, 5 mM EDTA, and 0.01 % thimerosal), *i.e.* 56 ng/ml was added to each well. After incubation for 2 h at room temperature the plate was washed and 100 μl of HRPO-conjugated streptavidin (Zymed Laboratories, S. San Francisco, CA) diluted 1:60000 in dilution buffer was added to each well. The plate was incubated for 30 minutes at room temperature with gentle agitation. The free avidin-conjugate was washed away and 100 μl freshly prepared substrate solution (tetramethyl benzidine [TMB]; 2-component substrate kit; Kirkegaard and Perry, Gaithersburg, MD) was added to each well. The reaction was allowed to proceed for 10 minutes, after which the color development was stopped by the addition of 100 μl/well 1.0 M H_3PO_4. The absorbance at 450 nm was read with a reference wavelength of 650 nm ($ABS_{450/650}$), using a *vmax* plate reader (Molecular Devices, Palo Alto, CA) controlled with a Macintosh Centris 650 (Apple Computers, Cupertino, CA) and DeltaSoft software (BioMetallics, Inc, Princeton, NJ).

The results demonstrated that *MPL* ligand was able to activate the *MPL*/Rse.gD chimeric receptor in a concentration-dependent and ligand-specific manner.

CLAIMS

1. A method for measuring autophosphorylation of a tyrosine kinase receptor comprising the steps of:

(a) coating a first solid phase with a homogeneous population of eukaryotic cells so that the cells adhere to the first solid phase, wherein, positioned in their membranes, the cells have a receptor construct comprising a flag polypeptide and the tyrosine kinase receptor;

(b) exposing the adhering cells to an analyte;

(c) solubilizing the adhering cells, thereby releasing cell lysate therefrom;

(d) coating a second solid phase with a capture agent which binds specifically to the flag polypeptide so that the capture agent adheres to the second solid phase;

(e) exposing the adhering capture agent to the cell lysate obtained in step (c) so that the receptor construct adheres to the second solid phase;

(f) washing the second solid phase so as to remove unbound cell lysate;

(g) exposing the adhering receptor construct to an anti-phosphotyrosine antibody which identifies phosphorylated tyrosine residues in the tyrosine kinase receptor; and

(h) measuring binding of the anti-phosphotyrosine antibody to the adhering receptor construct.

2. The method of claim 1 wherein the cells are transformed with nucleic acid encoding the receptor construct prior to step (a).

3. The method of claim 1 wherein the cells comprise a mammalian cell line.

4. The method of claim 1 wherein the cells are adherent.

5. The method of claim 1 wherein the capture agent comprises a capture antibody.

6. The method of claim 1 wherein the first solid phase comprises a well of a first assay plate.

7. The method of claim 6 wherein the first assay plate is a microtiter plate.

8. The method of claim 6 wherein between about 1×10^4 to 3×10^5 cells are added to the well in step (a).

9. The method of claim 1 wherein the second solid phase comprises a well of a second assay plate.

10. The method of claim 1 wherein the cell lysate is not concentrated or clarified prior to step (e).

11. The method of claim 6 wherein step (c) comprises adding a lysis buffer to the well of the first assay plate and gently agitating the first assay plate.

12. The method of claim 11 wherein the lysis buffer comprises a solubilizing detergent.

13. The method of claim 1 wherein the anti-phosphotyrosine antibody is labelled.

14. The method of claim 13 wherein the label comprises an enzyme which is exposed to a color reagent and the color change of the color reagent is determined in step (h).

15. The method of claim 1 wherein the flag polypeptide is fused to the amino terminus of the tyrosine kinase receptor.

16. The method of claim 15 wherein the tyrosine kinase receptor is a trk A receptor, trk B receptor or trk C receptor.

17. The method of claim 1 wherein the flag polypeptide is fused to the carboxyl terminus of the tyrosine kinase receptor.

18. The method of claim 17 wherein the tyrosine kinase receptor is the Rse receptor.

19. The method of claim 17 wherein the receptor construct comprises the extracellular domain of a receptor of interest and the intracellular domain of the Rse receptor.

20. The method of claim 19 wherein the receptor of interest is the *MPL* receptor.

21. The method of claim 20 wherein the receptor construct further comprises the transmembrane domain of the Rse receptor and the flag polypeptide comprises the gD polypeptide.

22. The method of claim 1 wherein the analyte comprises an agonist for the tyrosine kinase receptor.

23. The method of claim 1 wherein the analyte comprises an antagonist for the tyrosine kinase receptor.

24. The method of claim 23 wherein the antagonist competitively inhibits binding or activation of the tyrosine kinase receptor by an agonist thereto and step (b) is followed by a step wherein the adhering cells are exposed to the agonist.

25. The method of claim 1 wherein the analyte is a composition which comprises an antagonist and an agonist for the receptor and the assay measures the ability of the antagonist to bind to the agonist and thereby reduce activation of the tyrosine kinase receptor by the agonist.

26. The method of claim 1 wherein a block buffer is added to the second solid phase following step (d).

27. A method for measuring autophosphorylation of a tyrosine kinase receptor comprising the steps of:

(a) coating a well of a first assay plate with a homogeneous population of adherent cells so that the cells adhere to the

well, wherein the cells have a tyrosine kinase receptor positioned in the cell membranes thereof;

(b) exposing the adhering cells to an analyte;

(c) solubilizing the adhering cells thereby releasing cell lysate therefrom;

(d) coating a well of a second assay plate with a capture agent which binds specifically to the tyrosine kinase receptor so that the capture agent adheres to the well;

(e) exposing the cell lysate obtained in step (c) to the adhering capture agent so that the tyrosine kinase receptor adheres to the well;

(f) washing the well so as to remove unbound cell lysate;

(g) exposing the adhering tyrosine kinase receptor to an anti-phosphotyrosine antibody which binds selectively to phosphorylated tyrosine residues in the tyrosine kinase receptor;

(h) measuring binding of the anti-phosphotyrosine antibody to the adhering tyrosine kinase receptor.

28. The method of claim 27 wherein the tyrosine kinase receptor comprises the HER2 receptor.

29. A polypeptide comprising a flag polypeptide fused to the carboxyl terminus of the intracellular domain of the Rse receptor.

30. The polypeptide of claim 29 further comprising the transmembrane domain of the Rse receptor.

31. The polypeptide of claim 30 further comprising the extracellular domain of a receptor protein tyrosine kinase, other than the Rse receptor.

32. The polypeptide of claim 29 wherein the flag polypeptide comprises the gD flag.

33. A kit comprising a solid phase coated with a capture agent which binds specifically to a flag polypeptide.

34. The kit of claim 33 wherein the solid phase comprises a well of a microtiter plate.

35. The kit of claim 33 further comprising a labeled anti-phosphotyrosine antibody.

36. The kit of claim 35 wherein the label comprises an enzyme.

37. The kit of claim 33 further comprising a cell transformed with a nucleic acid encoding a receptor construct.

38. An assay for measuring phosphorylation of a kinase comprising the steps of:

(a) coating a first solid phase with a homogeneous population of eukaryotic cells so that the cells adhere to the first solid phase, wherein the cells comprise a kinase construct comprising a flag polypeptide and the kinase;

(b) exposing the adhering cells to an analyte;

(c) solubilizing the adhering cells, thereby releasing cell lysate therefrom;

(d) coating a second solid phase with a capture agent which binds specifically to the flag polypeptide so that the capture agent adheres to the second solid phase;

(e) exposing the adhering capture agent to the cell lysate obtained in step (c) so that the kinase construct adheres to the second solid phase;

(f) washing the second solid phase so as to remove unbound cell lysate;

(g) exposing the adhering kinase construct to an antibody which identifies phosphorylated residues in the kinase construct; and

(h) measuring binding of the antibody to the adhering kinase construct.

39. The assay of claim 38 wherein the kinase is a receptor.

40. The assay of claim 38 wherein the kinase is a serine-threonine kinase.

41. The assay of claim 38 which measures phosphatase activity.

42. The assay of claim 41 wherein the cells further comprise a phosphatase and the assay further comprises the step of exposing the eukaryotic cells to a phosphatase inhibitor prior to step (c).

43. The assay of claim 41 which further comprises the steps in between steps (f) and (g) of exposing the adhering kinase construct to a phosphatase and then washing the second solid phase so as to remove unbound phosphatase.

FIG. 1 A

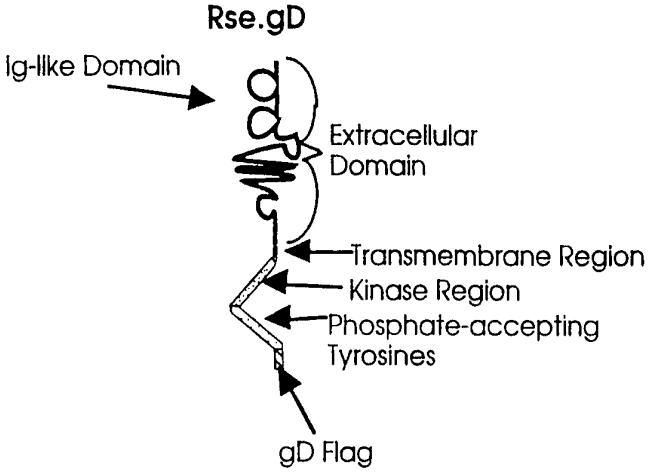


FIG. 1 B

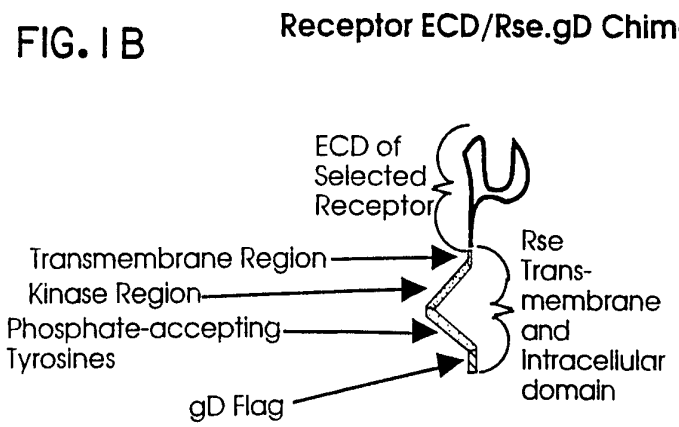
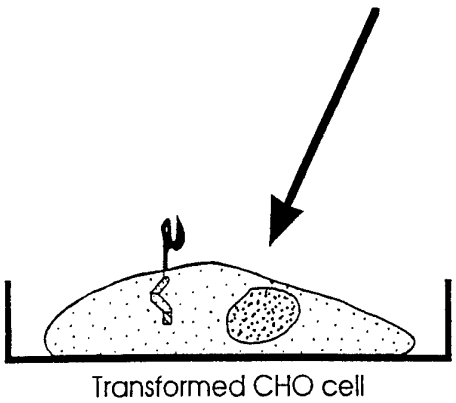


FIG. 1 C



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     signal sequence
     * * * *  * * *  * * *  * * * *  * * *  * * *  * * * *  * * *  * * *  * * * *
  1  M  A  L  R   R  S  M   G  R  P   G  L  P  P   L  P  L   P  P  P   P  R  L  G   L  L  L   A  A  L   A  S  L  L
  1  ATGGCGCTGA GGCGGAGCAT GGGGCGGCCG GGGCTCCCGC CGCTGCCGCT GCCGCCGCCA CCGCGGCTCG GGCTGCTGCT GGCGGCTCTG GCTTCTCTGC
     * * *  * * *  extracellular domain
 35   L  P  E   S  A  A   A  G  L  K   L  M  G   A  P  V   K  L  T  V   S  Q  G   Q  P  V   K  L  N  C   S  V  E
101  TGCTCCCGGA GTCCGCCGCC GCAGGTCTGA AGCTCATGGG AGCCCCGGTG AAGCTGACAG TGTCTCAGGG GCAGCCGGTG AAGCTCAACT GCAGTGTGGA

 68  G  M  E   E  P  D  I   Q  W  V   K  D  G   A  V  V  Q   N  L  D   Q  L  Y   I  P  V  S   E  Q  H   W  I  G
201  GGGGATGGAG GAGCCTGACA TCCAGTGGGT GAAGGATGGG GCTGTGGTCC AGAACTTGGA CCAGTTGTAC ATCCCAGTCA GCGAGCAGCA CTGGATCGGC

101  F  L  S  L   K  S  V   E  R  S   D  A  G  R   Y  W  C   Q  V  E   D  G  G  E   T  E  I   S  Q  P   V  W  L  T
301  TTCCTCAGCC TGAAGTCAGT GGAGCGCTCT GACGCCGGCC GGTACTGGTG CCAGGTGGAG GATGGGGGTG AAACCGAGAT CTCCCAGCCA GTGTGGCTCA

135   V  E  G   V  P  F   F  T  V  E   P  K  D   L  A  V   P  P  N  A   P  F  Q   L  S  C   E  A  V  G   P  P  E
401  CGGTAGAAGG TGTGCCATTT TTCACAGTGG AGCCAAAAGA TCTGGCAGTG CCACCCAATG CCCCTTTCCA ACTGTCTTGT GAGGCTGTGG GTCCCCCTGA

168  P  V  T   I  V  W  W   R  G  T   T  K  I   G  G  P  A   P  S  P   S  V  L   N  V  T  G   V  T  Q   S  T  M
501  ACCTGTTACC ATTGTCTGGT GGAGAGGAAC TACGAAGATC GGGGGACCCG CTCCCTCTCC ATCTGTTTTA AATGTAACAG GGGTGACCCA GAGCACCATG

201  F  S  C  E   A  H  N   L  K  G   L  A  S  S   R  T  A   T  V  H   L  Q  A  L   P  A  A   P  F  N   I  T  V  T
601  TTTTCCTGTG AAGCTCACAA CCTAAAAGGC CTGGCCTCTT CTCGCACAGC CACTGTTCAC CTTCAAGCAC TGCCTGCAGC CCCCTTCAAC ATCACCGTGA

235   K  L  S   S  S  N   A  S  V  A   W  M  P   G  A  D   G  R  A  L   L  Q  S   C  T  V   Q  V  T  Q   A  P  G
701  CAAAGCTTTC CAGCAGCAAC GCTAGTGTGG CCTGGATGCC AGGTGCTGAT GGCCGAGCTC TGCTACAGTC CTGTACAGTT CAGGTGACAC AGGCCCCAGG

268  G  W  E   V  L  A  V   V  V  P   V  P  P   F  T  C  L   L  R  D   L  V  P   A  T  N  Y   S  L  R   V  R  C
801  AGGCTGGGAA GTCCTGGCTG TTGTGGTCCC TGTGCCCCCC TTTACCTGCC TGCTCCGGGA CCTGGTGCCT GCCACCAACT ACAGCCTCAG GGTGCGCTGT

301  A  N  A  L   G  P  S   P  Y  A   D  W  V  P   F  Q  T   K  G  L   A  P  A  S   A  P  Q   N  L  H   A  I  R  T
901  GCCAATGCCT TGGGGCCCTC TCCCTATGCT GACTGGGTGC CCTTTCAGAC CAAGGGTCTA GCCCCAGCCA GCGCTCCCCA AAACCTCCAT GCCATCCGCA

335   D  S  G   L  I  L   E  W  E  E   V  I  P   E  A  P   L  E  G  P   L  G  P   Y  K  L   S  W  V  Q   D  N  G
1001 CAGATTCAGG CCTCATCTTG GAGTGGGAAG AAGTGATCCC CGAGGCCCCT TTGGAAGGCC CCCTGGGACC CTACAAACTG TCCTGGGTTC AAGACAATGG
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FIG.2A

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 368  T  Q  D   E  L  T  V   E  G  T   R  A  N   L  T  G  W   D  P  Q   K  D  L   I  V  R  V   C  V  S   N  A  V
1101 AACCCAGGAT GAGCTGACAG TGGAGGGGAC CAGGGCCAAT TTGACAGGCT GGGATCCCCA AAAGGACCTG ATCGTACGTG TGTGCGTCTC CAATGCAGTT
                                                                                                  transmembrane domain
 401 G  C  G  P   W  S  Q   P  L  V   V  S  S  H   D  R  A   G  Q  Q   G  P  P  H   S  R  T   S  W  V   P  V  V  L
1201 GGCTGTGGAC CCTGGAGTCA GCCACTGGTG GTCTCTTCTC ATGACCGTGC AGGCCAGCAG GGCCCTCCTC ACAGCCGCAC ATCCTGGGTA CCTGTGGTCC
                                                                  intracellular domain
 435  G  V  L   T  A  L   V  T  A  A   A  L  A   L  I  L   L  R  K  R   R  K  E   T  R  F   G  Q  A  F   D  S  V
1301 TTGGTGTGCT AACGGCCCTG GTGACGGCTG CTGCCCTGGC CCTCATCCTG CTTCGAAAGA GACGGAAAGA GACGCGGTTT GGGCAAGCCT TTGACAGTGT

 468  M  A  R   G  E  P  A   V  H  F   R  A  A   R  S  F  N   R  E  R   P  E  R   I  E  A  T   L  D  S   L  G  I
1401 CATGGCCCGG GGAGAGCCAG CCGTTCACTT CCGGGCAGCC CGGTCCTTCA ATCGAGAAAG GCCCGAGCGC ATCGAGGCCA CATTGGACAG CTTGGGCATC

 501 S  D  E  L   K  E  K   L  E  D   V  L  I  P   E  Q  Q   F  T  L   G  R  M  L   G  K  G   E  F  G   S  V  R  E
1501 AGCGATGAAC TAAAGGAAAA ACTGGAGGAT GTGCTCATCC CAGAGCAGCA GTTCACCCTG GGCCGGATGT TGGGCAAAGG AGAGTTTGGT TCAGTGCGGG

 535  A  Q  L   K  Q  E   D  G  S  F   V  K  V   A  V  K   M  L  K  A   D  I  I   A  S  S   D  I  E  E   F  L  R
1601 AGGCCCAGCT GAAGCAAGAG GATGGCTCCT TTGTGAAAGT GGCTGTGAAG ATGCTGAAAG CTGACATCAT TGCCTCAAGC GACATTGAAG AGTTCCTCAG

 568  E  A  A   C  M  K  E   F  D  H   P  H  V   A  K  L  V   G  V  S   L  R  S   R  A  K  G   R  L  P   I  P  M
1701 GGAAGCAGCT TGCATGAAGG AGTTTGACCA TCCACACGTG GCCAAACTTG TTGGGGTAAG CCTCCGGAGC AGGGCTAAAG GCCGTCTCCC CATCCCCATG

 601 V  I  L  P   F  M  K   H  G  D   L  H  A  F   L  L  A   S  R  I   G  E  N  P   F  N  L   P  L  Q   T  L  I  R
1801 GTCATCTTGC CCTTCATGAA GCATGGGGAC CTGCATGCCT TCCTGCTCGC CTCCCGGATT GGGGAGAACC CCTTTAACCT ACCCCTCCAG ACCCTGATCC

 635  F  M  V   D  I  A   C  G  M  E   Y  L  S   S  R  N   F  I  H  R   D  L  A   A  R  N   C  M  L  A   E  D  M
1901 GGTTCATGGT GGACATTGCC TGCGGCATGG AGTACCTGAG CTCTCGGAAC TTCATCCACC GAGACCTGGC TGCTCGGAAT TGCATGCTGG CAGAGGACAT

 668  T  V  C   V  A  D  F   G  L  S   R  K  I   Y  S  G  D   Y  Y  R   Q  G  C   A  S  K  L   P  V  K   W  L  A
2001 GACAGTGTGT GTGGCTGACT TCGGACTCTC CCGGAAGATC TACAGTGGGG ACTACTATCG TCAAGGCTGT GCCTCCAAAC TGCCTGTCAA GTGGCTGGCC
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FIG.2B

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 701 L  E  S  L   A  D  N   L  Y  T   V  Q  S  D   V  W  A   F  G  V   T  M  W  E   I  M  T   R  G  Q   T  P  Y  A
2101 CTGGAGAGCC TGGCCGACAA CCTGTATACT GTGCAGAGTG ACGTGTGGGC GTTCGGGGTG ACCATGTGGG AGATCATGAC ACGTGGGCAG ACGCCATATG

 735  G  I  E   N  A  E   I  Y  N  Y   L  I  G   G  N  R   L  K  Q  P   P  E  C   M  E  D   V  Y  D  L   M  Y  Q
2201 CTGGCATCGA AAACGCTGAG ATTTACAACT ACCTCATTGG CGGGAACCGC CTGAAACAGC CTCCGGAGTG TATGGAGGAC GTGTATGATC TCATGTACCA

 768  C  W  S   A  D  P  K   Q  R  P   S  F  T   C  L  R  M   E  L  E   N  I  L   G  Q  L  S   V  L  S   A  S  Q
2301 GTGCTGGAGT GCTGACCCCA AGCAGCGCCC GAGCTTTACT TGTCTGCGAA TGGAACTGGA GAACATCTTG GGCCAGCTGT CTGTGCTATC TGCCAGCCAG

 801 D  P  L  Y   I  N  I   E  R  A   E  E  P  T   A  G  G   S  L  E   L  P  G  R   D  Q  P   Y  S  G   A  G  D  G
2401 GACCCCTTAT ACATCAACAT CGAGAGAGCT GAGGAGCCCA CTGCGGGAGG CAGCCTGGAG CTACCTGGCA GGGATCAGCC CTACAGTGGG GCTGGGGATG

 835  S  G  M   G  A  V   G  G  T  P   S  D  C   R  Y  I   L  T  P  G   G  L  A   E  Q  P   G  Q  A  E   H  Q  P
2501 GCAGTGGCAT GGGGGCAGTG GGTGGCACTC CCAGTGACTG TCGGTACATA CTCACCCCCG GAGGGCTGGC TGAGCAGCCA GGGCAGGCAG AGCACCAGCC
                                                                                 gD flag polypeptide
 868  E  S  P   L  N  E  T   Q  R  L   L  L  L   Q  Q  G  L   L  P  H   S  S  C   A  D  A  S   L  K  M   A  D  P
2601 AGAGAGTCCC CTCAATGAGA CACAGAGGCT TTTGCTGCTG CAGCAAGGGC TACTGCCACA CTCGAGCTGC GCAGATGCTA GCCTCAAGAT GGCTGATCCA

 901 N  R  F  R   G  K  D   L  P  V   L  O
2701 AATCGATTCC GCGGCAAAGA TCTTCCGGTC CTGTAGAAGC TT
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FIG.2C

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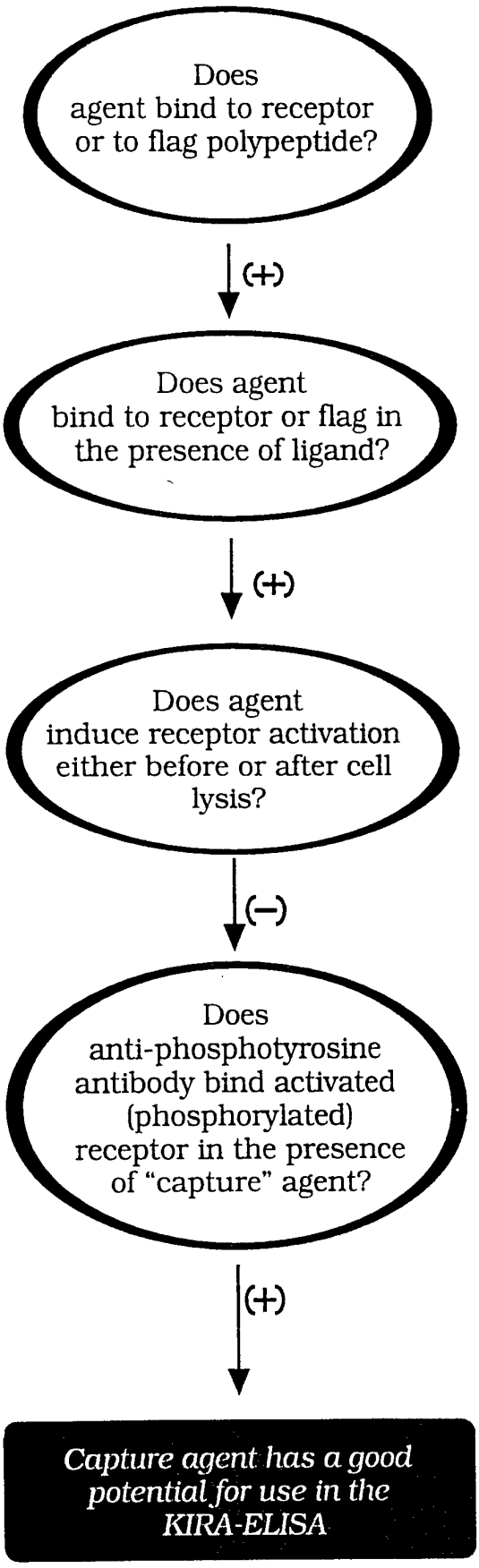
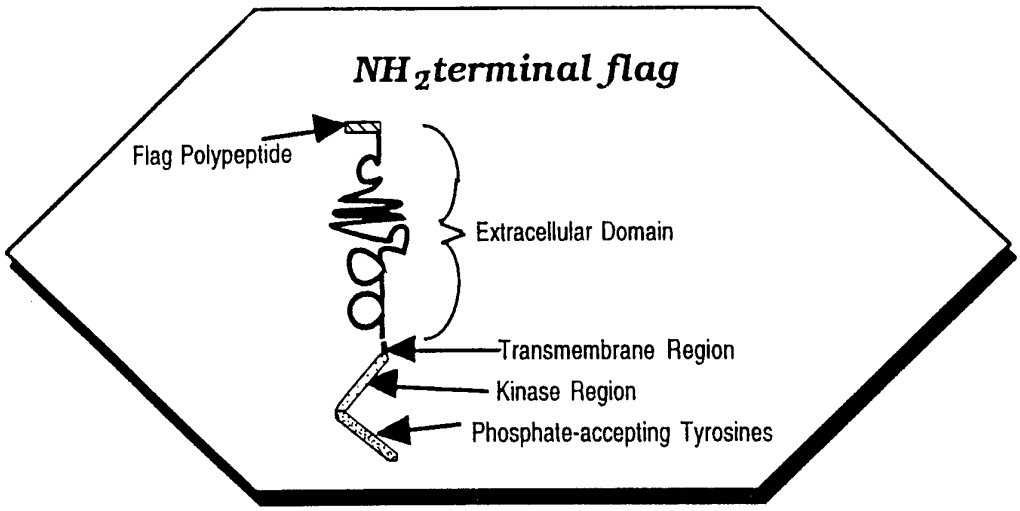

Does agent bind to receptor or to flag polypeptide?
(+)
Does agent bind to receptor or flag in the presence of ligand?
(+)
Does agent induce receptor activation either before or after cell lysis?
(−)
Does anti-phosphotyrosine antibody bind activated (phosphorylated) receptor in the presence of "capture" agent?
(+)
Capture agent has a good potential for use in the KIRA-ELISA

FIG.3


NH2terminal flag
Flag Polypeptide
Extracellular Domain
Transmembrane Region
Kinase Region
Phosphate-accepting Tyrosines

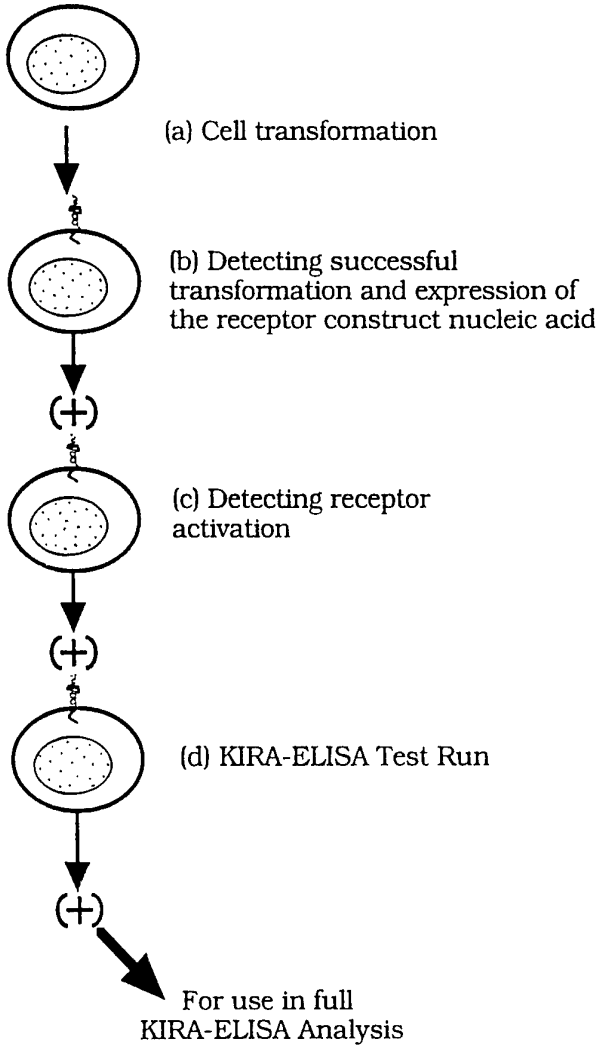
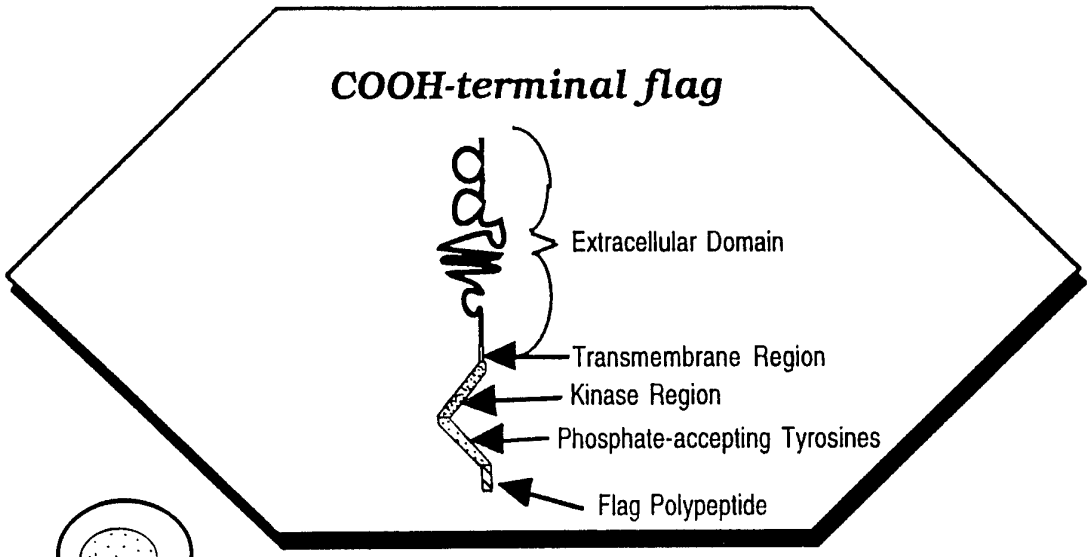

(a) Cell transformation
(b) Detecting successful transformation and expression of the receptor construct nucleic acid
(+)
(c) Detecting receptor activation
(+)
(d) KIRA-ELISA Test Run
(+)
For use in full KIRA-ELISA Analysis

FIG.4


COOH-terminal flag
Extracellular Domain
Transmembrane Region
Kinase Region
Phosphate-accepting Tyrosines
Flag Polypeptide

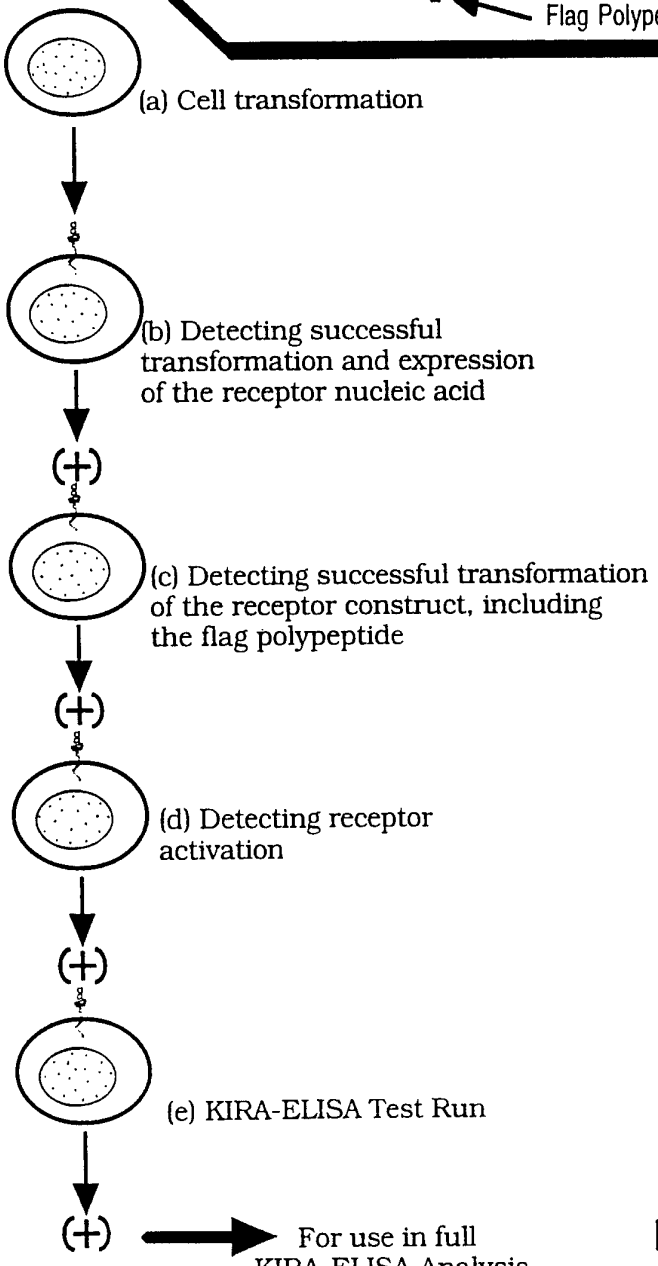

(a) Cell transformation
(b) Detecting successful transformation and expression of the receptor nucleic acid
(+)
(c) Detecting successful transformation of the receptor construct, including the flag polypeptide
(+)
(d) Detecting receptor activation
(+)
(e) KIRA-ELISA Test Run
(+)
For use in full KIRA-ELISA Analysis

FIG.5

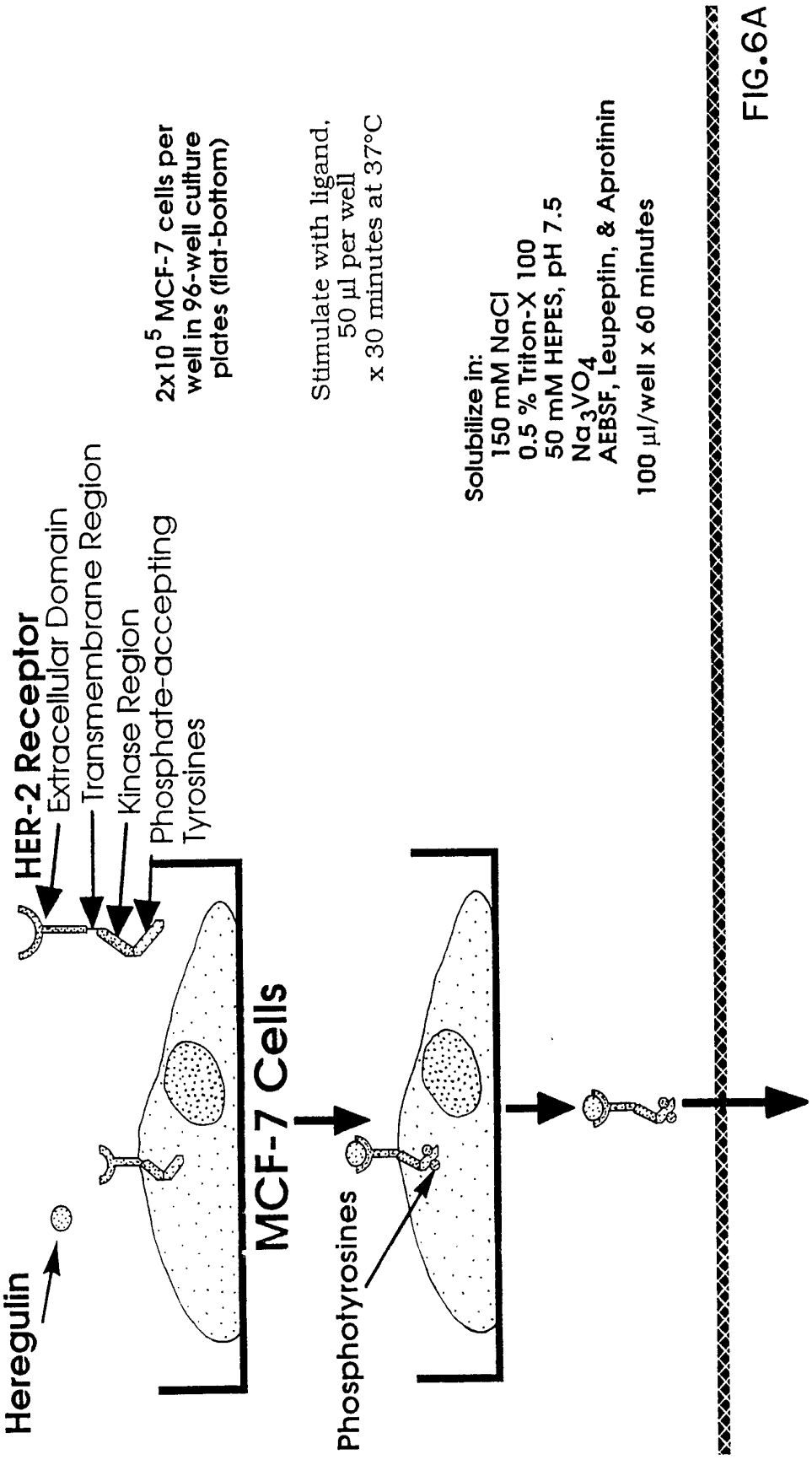

HER-2 Receptor
Extracellular Domain
Transmembrane Region
Kinase Region
Phosphate-accepting Tyrosines
2x10^5 MCF-7 cells per well in 96-well culture plates (flat-bottom)
Stimulate with ligand, 50 µl per well x 30 minutes at 37°C
Solubilize in:
150 mM NaCl
0.5 % Triton-X 100
50 mM HEPES, pH 7.5
Na3VO4
AEBSF, Leupeptin, & Aprotinin
100 µl/well x 60 minutes
Heregulin
MCF-7 Cells
Phosphotyrosines

FIG.6A

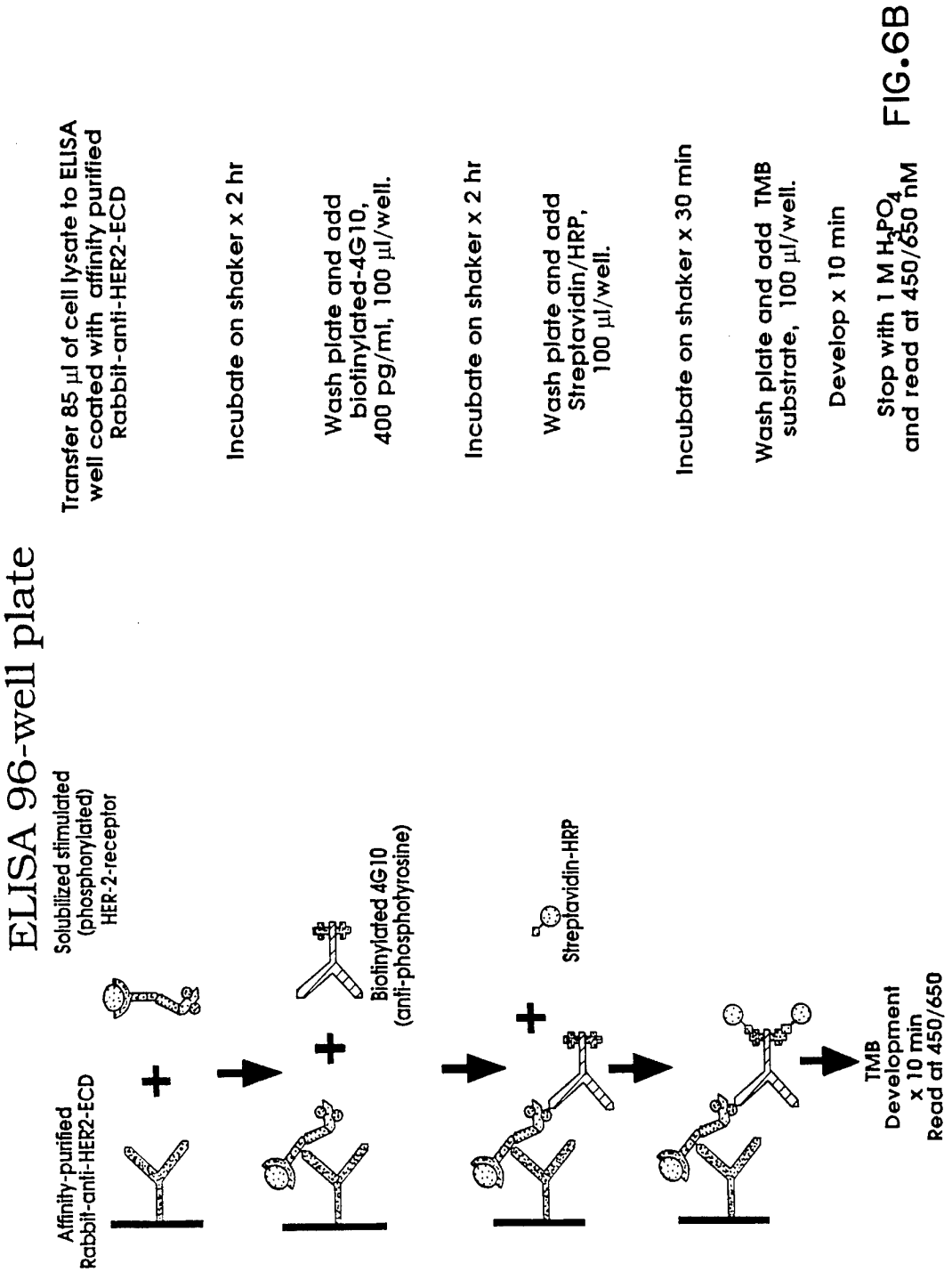

ELISA 96-well plate
Affinity-purified Rabbit-anti-HER2-ECD
Solubilized stimulated (phosphorylated) HER-2-receptor
Biotinylated 4G10 (anti-phosphohyrosine)
Streptavidin-HRP
TMB Development x 10 min Read at 450/650
Transfer 85 µl of cell lysate to ELISA well coated with affinity purified Rabbit-anti-HER2-ECD
Incubate on shaker x 2 hr
Wash plate and add biotinylated-4G10, 400 pg/ml, 100 µl/well.
Incubate on shaker x 2 hr
Wash plate and add Streptavidin/HRP, 100 µl/well.
Incubate on shaker x 30 min
Wash plate and add TMB substrate, 100 µl/well.
Develop x 10 min
Stop with 1 M H_3PO_4 and read at 450/650 nM

FIG.6B

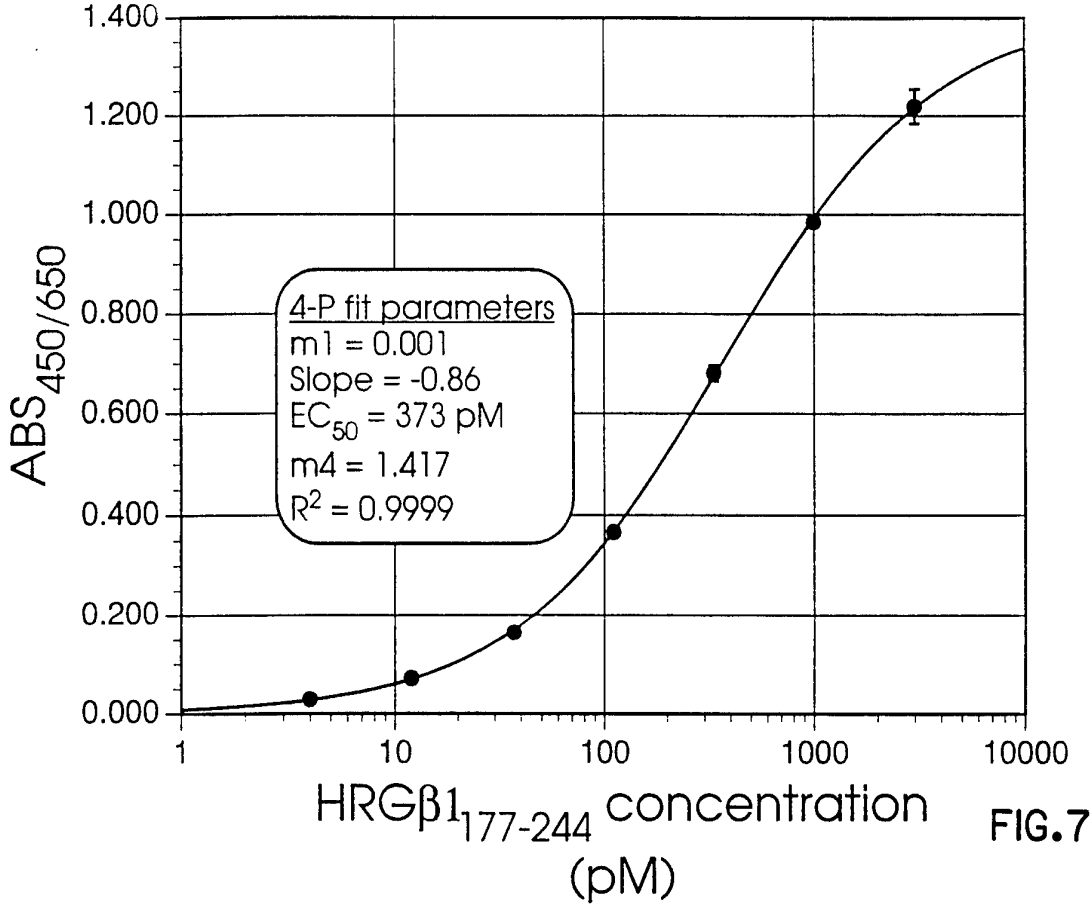

1.400
1.200
1.000
0.800
0.600
0.400
0.200
0.000
ABS450/650
4-P fit parameters
m1 = 0.001
Slope = -0.86
EC50 = 373 pM
m4 = 1.417
R2 = 0.9999
1
10
100
1000
10000
HRGβ1177-244 concentration
(pM)

FIG.7

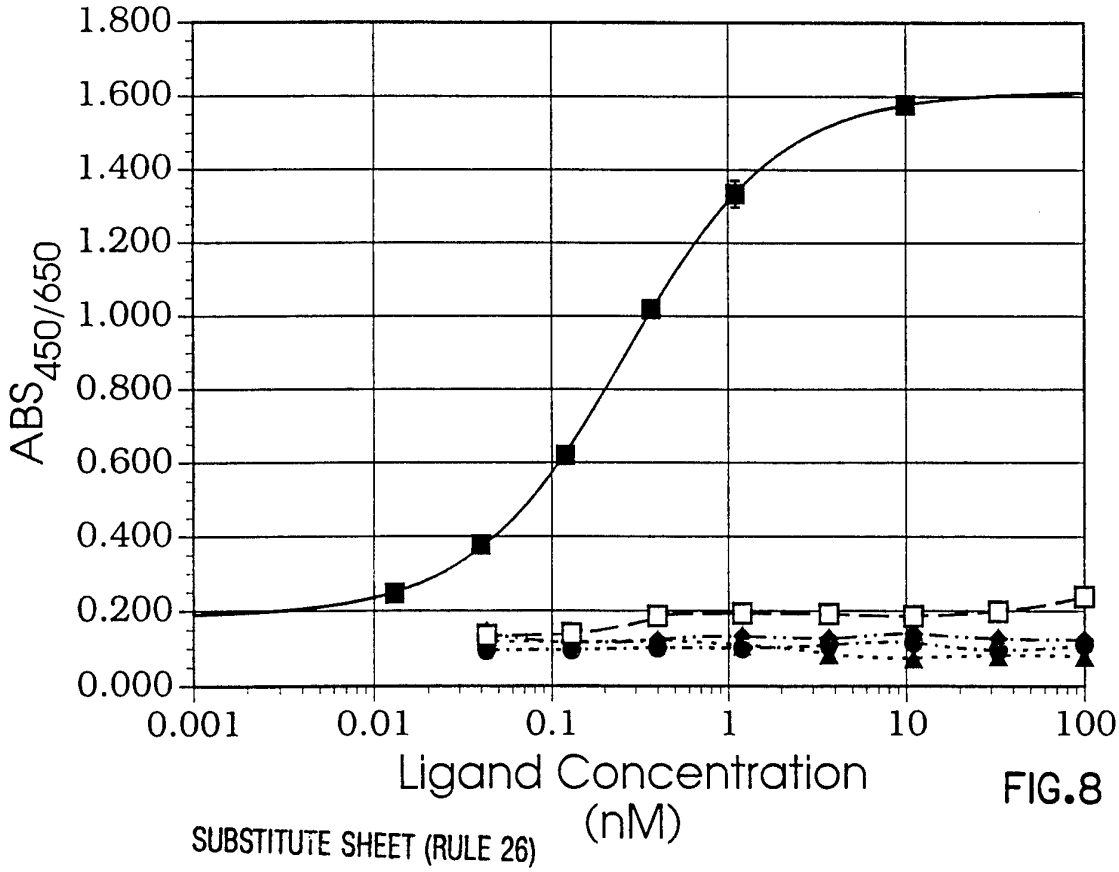

1.800
1.600
1.400
1.200
1.000
0.800
0.600
0.400
0.200
0.000
ABS450/650
0.001
0.01
0.1
1
10
100
Ligand Concentration
(nM)

FIG.8

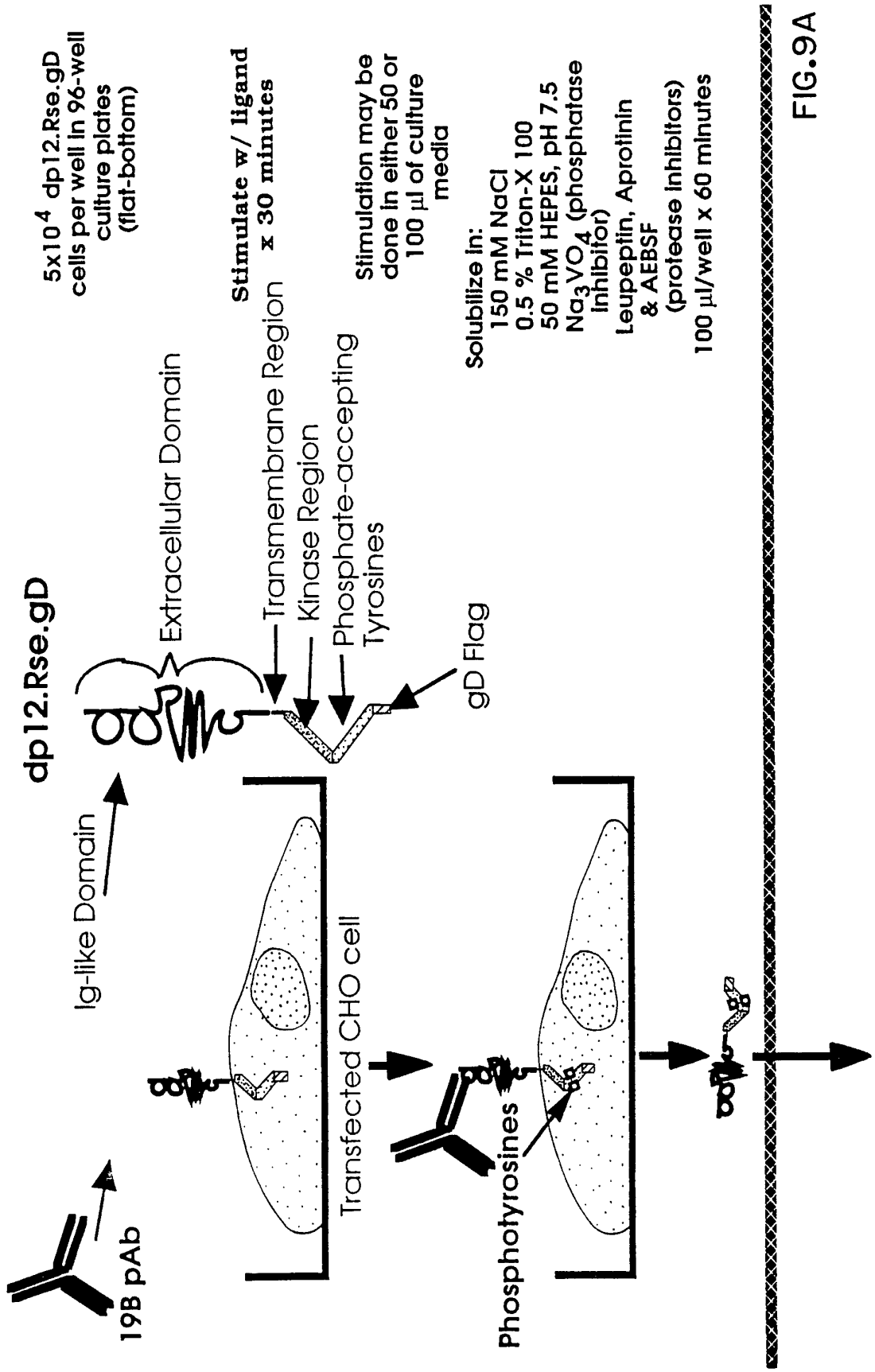

5x10^4 dp12.Rse.gD cells per well in 96-well culture plates (flat-bottom)
Stimulate w/ ligand x 30 minutes
Stimulation may be done in either 50 or 100 µl of culture media
Solubilize in:
150 mM NaCl
0.5 % Triton-X 100
50 mM HEPES, pH 7.5
Na3VO4 (phosphatase inhibitor)
Leupeptin, Aprotinin & AEBSF (protease inhibitors)
100 µl/well x 60 minutes
dp12.Rse.gD
Extracellular Domain
Transmembrane Region
Kinase Region
Phosphate-accepting Tyrosines
gD Flag
Ig-like Domain
19B pAb
Transfected CHO cell
Phosphotyrosines

FIG.9A

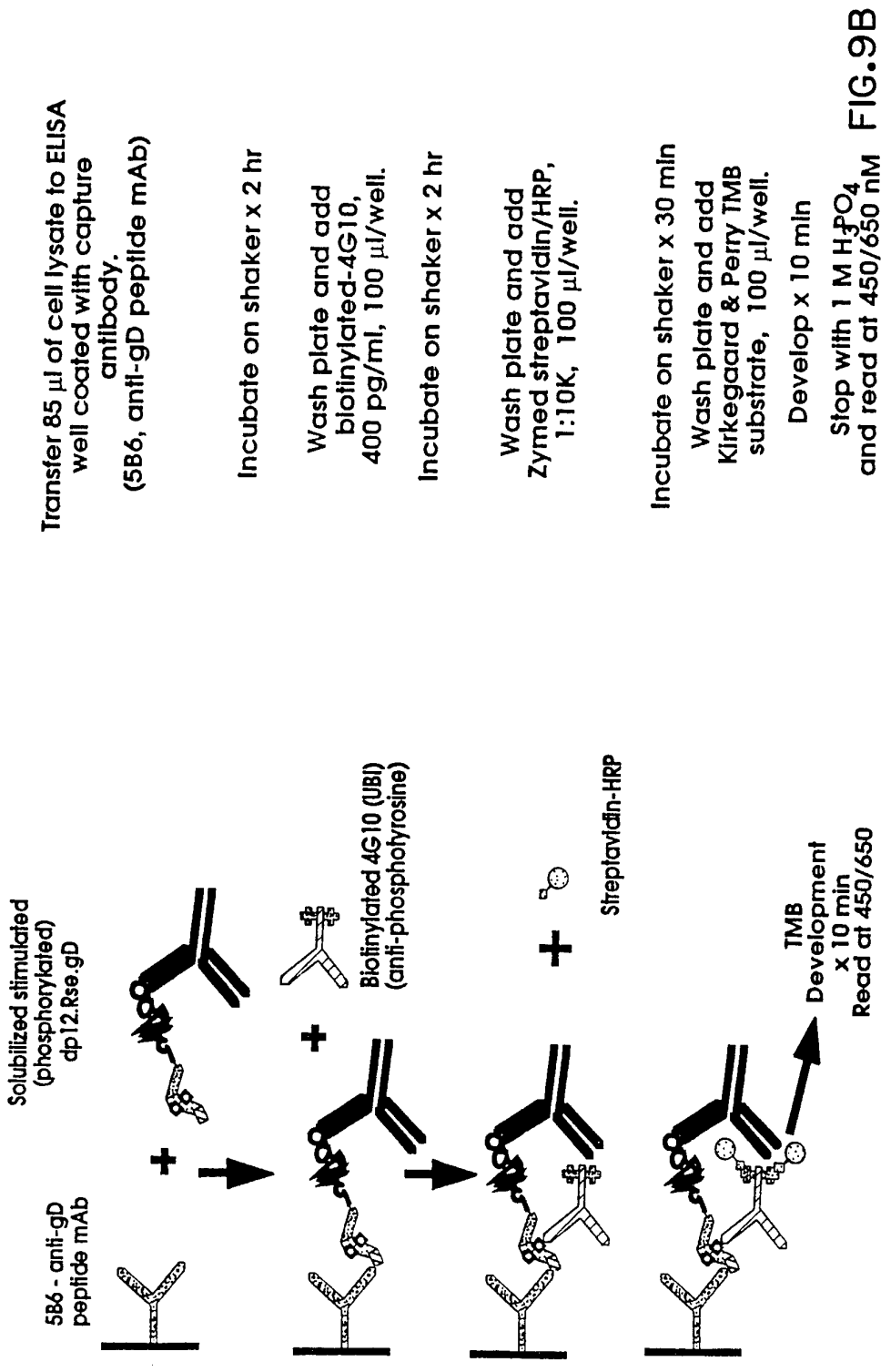

ELISA 96-well plate
5B6 - anti-gD peptide mAb
Solubilized stimulated (phosphorylated) dp12.Rse.gD
Biotinylated 4G10 (UBI) (anti-phosphotyrosine)
Streptavidin-HRP
TMB Development x 10 min Read at 450/650
Transfer 85 µl of cell lysate to ELISA well coated with capture antibody. (5B6, anti-gD peptide mAb)
Incubate on shaker x 2 hr
Wash plate and add biotinylated-4G10, 400 pg/ml, 100 µl/well.
Incubate on shaker x 2 hr
Wash plate and add Zymed streptavidin/HRP, 1:10K, 100 µl/well.
Incubate on shaker x 30 min
Wash plate and add Kirkegaard & Perry TMB substrate, 100 µl/well.
Develop x 10 min
Stop with 1 M H_3PO_4 and read at 450/650 nM

FIG.9B

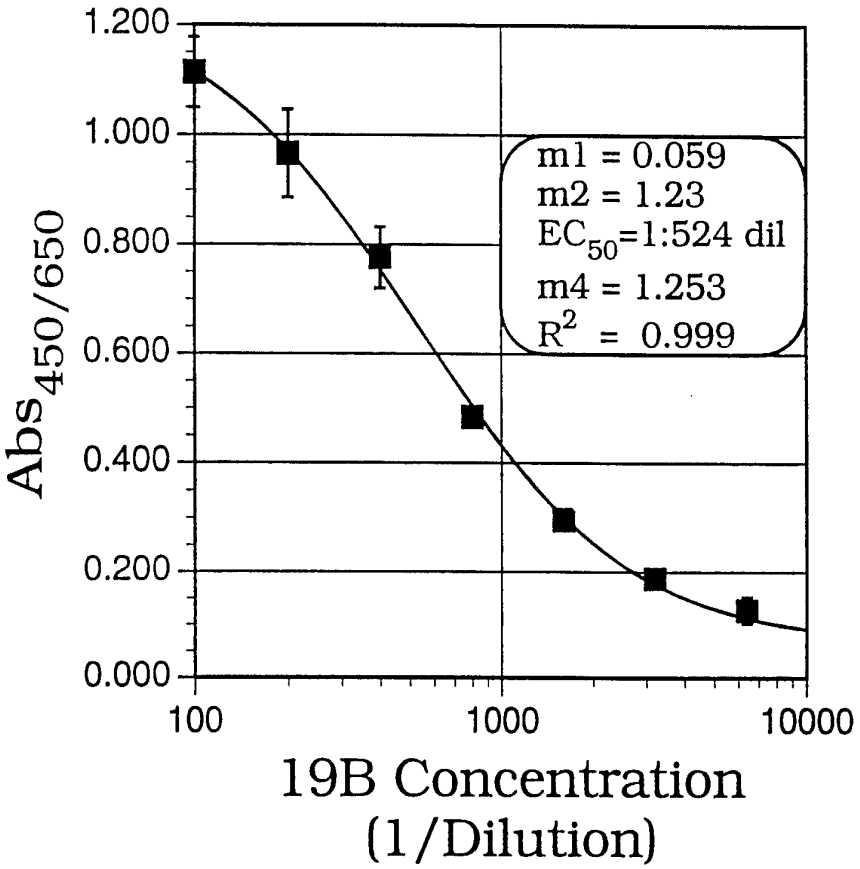

1.200
1.000
0.800
0.600
0.400
0.200
0.000
Abs450/650
100
1000
10000
m1 = 0.059
m2 = 1.23
EC50=1:524 dil
m4 = 1.253
R2 = 0.999
19B Concentration
(1/Dilution)

FIG. 10

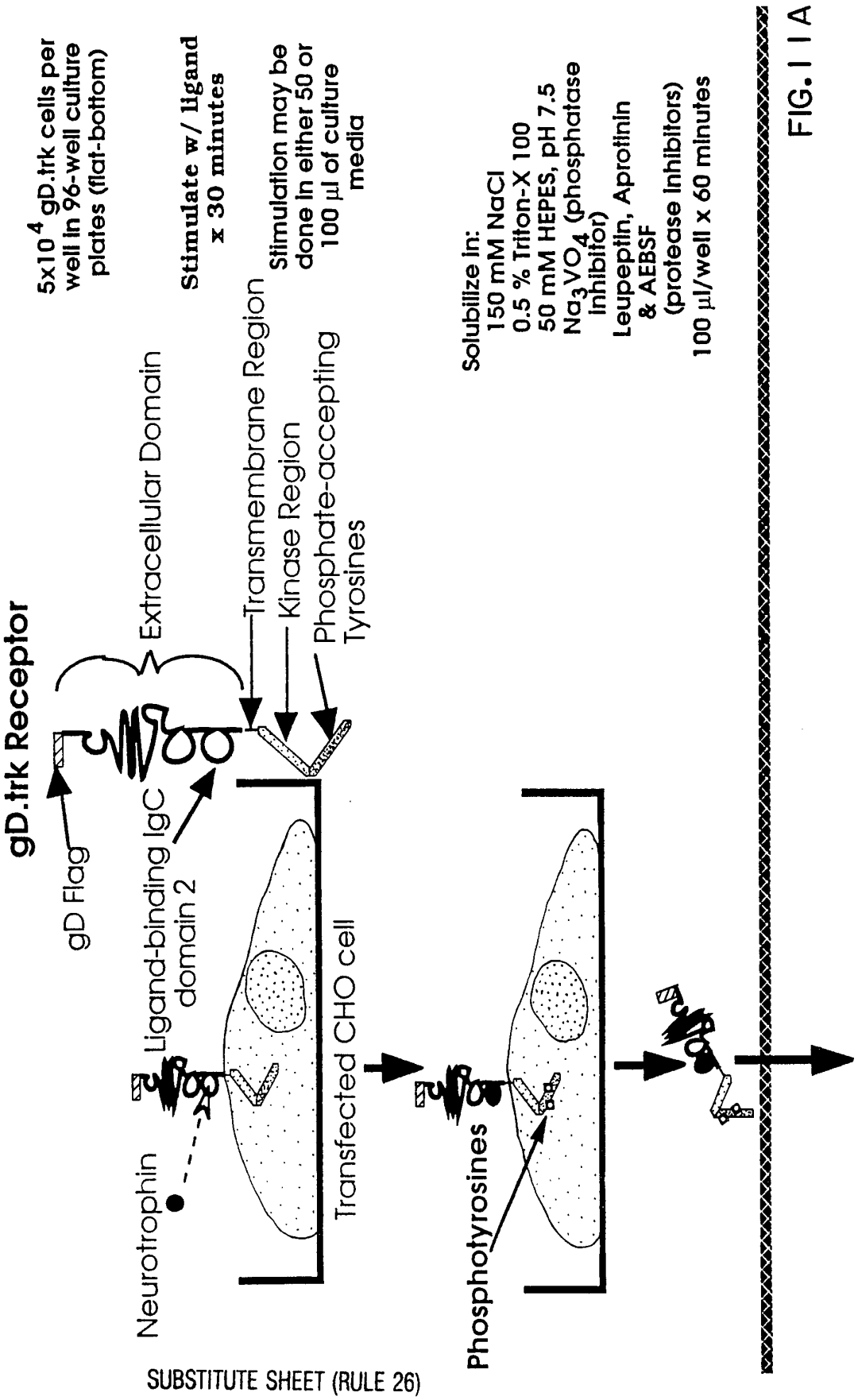

5x10^4 gD.trk cells per well in 96-well culture plates (flat-bottom)
Stimulate w/ ligand x 30 minutes
Stimulation may be done in either 50 or 100 µl of culture media
gD.trk Receptor
Extracellular Domain
Transmembrane Region
Kinase Region
Phosphate-accepting Tyrosines
gD Flag
Ligand-binding IgC domain 2
Neurotrophin
Transfected CHO cell
Solubilize in:
150 mM NaCl
0.5 % Triton-X 100
50 mM HEPES, pH 7.5
Na3VO4 (phosphatase inhibitor)
Leupeptin, Aprotinin & AEBSF
(protease inhibitors)
100 µl/well x 60 minutes
Phosphotyrosines

FIG. 11A

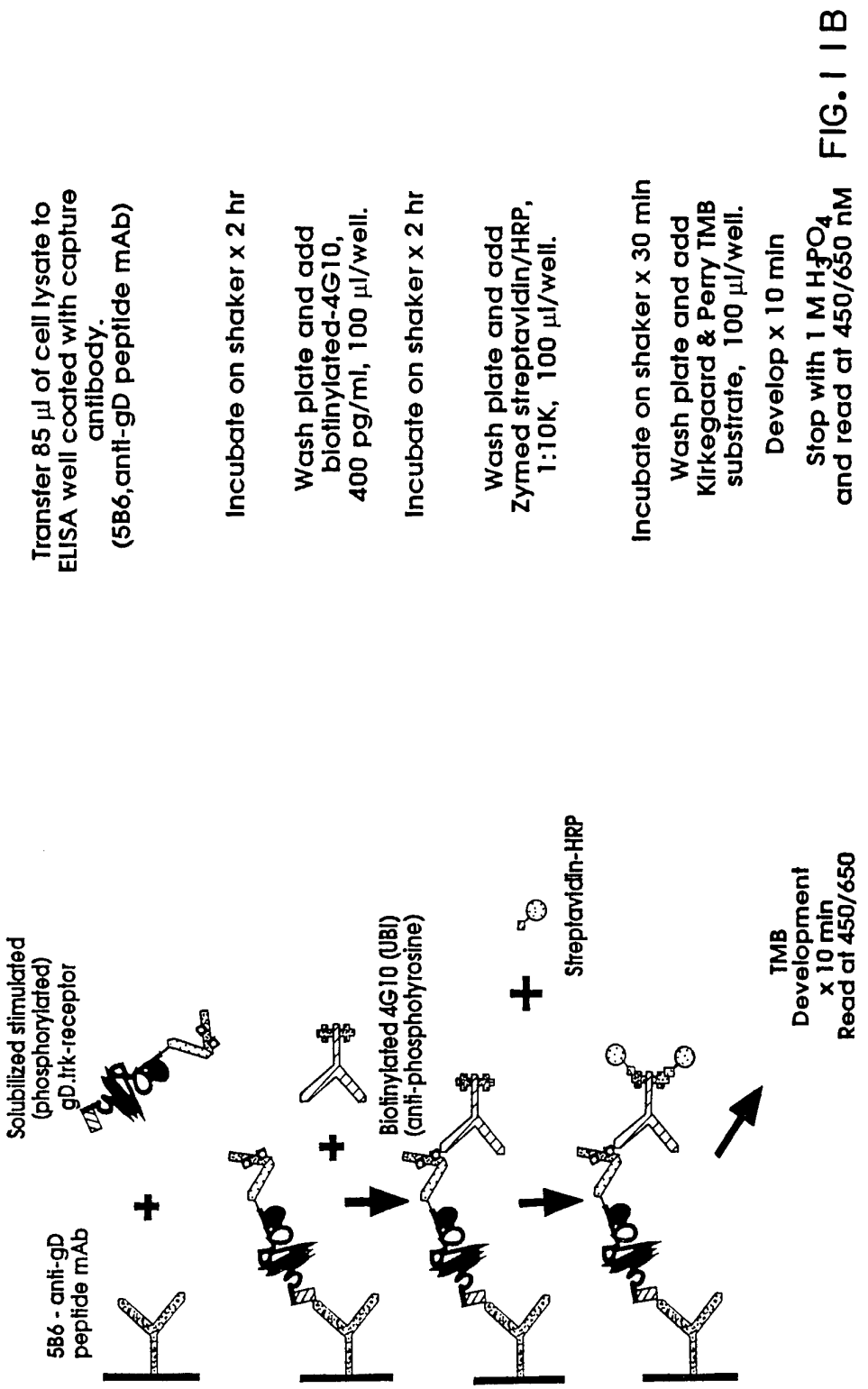

ELISA 96-well plate
Transfer 85 µl of cell lysate to ELISA well coated with capture antibody. (5B6,anti-gD peptide mAb)
Incubate on shaker x 2 hr
Wash plate and add biotinylated-4G10, 400 pg/ml, 100 µl/well.
Incubate on shaker x 2 hr
Wash plate and add Zymed streptavidin/HRP, 1:10K, 100 µl/well.
Incubate on shaker x 30 min
Wash plate and add Kirkegaard & Perry TMB substrate, 100 µl/well.
Develop x 10 min
Stop with 1 M H_3PO_4 and read at 450/650 nM
5B6 - anti-gD peptide mAb
Solubilized stimulated (phosphorylated) gD.trk-receptor
Biotinylated 4G10 (UBI) (anti-phosphotyrosine)
Streptavidin-HRP
TMB Development x 10 min Read at 450/650

FIG. 11B

SUBSTITUTE SHEET (RULE 26)

```
          ^sp6 RNA start
 841 TATAGAATAA CATCCACTTT GCCTTTCTCT CCACAGGTGT CCACTCCCAG GTCCAACTGC
     ATATCTTATT GTAGGTGAAA CGGAAAGAGA GGTGTCCACA GGTGAGGGTC CAGGTTGACG

       ^cloning linker
          ^R1 site mutated in
          ^begin gD from pchadII
 901 ACCTGAATTC CACTGCCTTC CACCAAGCTC TGCAGGATCC CAGAGTCAGG GGTCTGTATC
     TGGACTTAAG GTGACGGAAG GTGGTTCGAG ACGTCCTAGG GTCTCAGTCC CCAGACATAG

 961 TTCCTGCTGG TGGCTCCAGT TCAGGAACAG TAAACCCTGC TCCGAATATT GCCTCTCACA
     AAGGACGACC ACCGAGGTCA AGTCCTTGTC ATTTGGGACG AGGCTTATAA CGGAGAGTGT

1021 TCTCGTCAAT CTCCGCGAGG ACTGGGGACC CTGTGACAAG CTTCAGCGCG AACGACCAAC
     AGAGCAGTTA GAGGCGCTCC TGACCCCTGG GACACTGTTC GAAGTCGCGC TTGCTGGTTG

                                                          ^Start gD
   1                                                      M* G* G*
1081 TACCCCGATC ATCAGTTATC CTTAAGGTCT CTTTTGTGTG GTGCGTTCCG GTATGGGGGG
     ATGGGGCTAG TAGTCAATAG GAATTCCAGA GAAAACACAC CACGCAAGGC CATACCCCCC

   4  T* A* A*  R* L* G* A*  V* I* L*  F* V* V*  I* V* G* L*  H* G* V*
1141 GACTGCCGCC AGGTTGGGGG CCGTGATTTT GTTTGTCGTC ATAGTGGGCC TCCATGGGGT
     CTGACGGCGG TCCAACCCCC GGCACTAAAA CAAACAGCAG TATCACCCGG AGGTACCCCA

  24  R* G* K*  Y* A* L* A  D  A  S  L  K  M  A  D  P  N  R  F  R
1201 CCGCGGCAAA TATGCCTTGG CGGATGCCTC TCTCAAGATG GCCGACCCCA ATCGATTTCG
     GGCGCCGTTT ATACGGAACC GCCTACGGAG AGAGTTCTAC CGGCTGGGGT TAGCTAAAGC
```

FIG. 12A

SUBSTITUTE SHEET (RULE 26)

```
                                        ^Xho and GTA mutated in
                                                  ^begin mature trkA
  44  G  K  D   L  P  V  L   D  Q  L   L  E  V   A  A  P  C   P  D  A
1261  CGGCAAAGAC CTTCCGGTCC TGGACCAGCT GCTCGAGGTA GCCGCACCCT GCCCCGATGC
      GCCGTTTCTG GAAGGCCAGG ACCTGGTCGA CGAGCTCCAT CGGCGTGGGA CGGGGCTACG

  64  C  C  P   H  G  S  S   G  L  R   C  T  R   D  G  A  L   D  S  L
1321  CTGCTGCCCC CACGGCTCCT CGGGACTGCG ATGCACCCGG GATGGGGCCC TGGATAGCCT
      GACGACGGGG GTGCCGAGGA GCCCTGACGC TACGTGGGCC CTACCCCGGG ACCTATCGGA

  84  H  H  L   P  G  A  E   N  L  T   E  L  Y   I  E  N  Q   Q  H  L
1381  CCACCACCTG CCCGGCGCAG AGAACCTGAC TGAGCTCTAC ATCGAGAACC AGCAGCATCT
      GGTGGTGGAC GGGCCGCGTC TCTTGGACTG ACTCGAGATG TAGCTCTTGG TCGTCGTAGA

 104  Q  H  L   E  L  R  D   L  R  G   L  G  E   L  R  N  L   T  I  V
1441  GCAGCATCTG GAGCTCCGTG ATCTGAGGGG CCTGGGGGAG CTGAGAAACC TCACCATCGT
      CGTCGTAGAC CTCGAGGCAC TAGACTCCCC GGACCCCCTC GACTCTTTGG AGTGGTAGCA

 124  K  S  G   L  R  F  V   A  P  D   A  F  H   F  T  P  R   L  S  R
1501  GAAGAGTGGT CTCCGTTTCG TGGCGCCAGA TGCCTTCCAT TTCACTCCTC GGCTCAGTCG
      CTTCTCACCA GAGGCAAAGC ACCGCGGTCT ACGGAAGGTA AAGTGAGGAG CCGAGTCAGC

 144  L  N  L   S  F  N  A   L  E  S   L  S  W   K  T  V  Q   G  L  S
1561  CCTGAATCTC TCCTTCAACG CTCTGGAGTC TCTCTCCTGG AAAACTGTGC AGGGCCTCTC
      GGACTTAGAG AGGAAGTTGC GAGACCTCAG AGAGAGGACC TTTTGACACG TCCCGGAGAG

 164  L  Q  E   L  V  L  S   G  N  P   L  H  C   S  C  A  L   R  W  L
1621  CTTACAGGAA CTGGTCCTGT CGGGGAACCC TCTGCACTGT TCTTGTGCCC TGCGCTGGCT
      GAATGTCCTT GACCAGGACA GCCCCTTGGG AGACGTGACA AGAACACGGG ACGCGACCGA
```

FIG. 12B

SUBSTITUTE SHEET (RULE 26)

```
 184  Q  R  W   E  E  E  G   L  G  G   V  P  E   Q  K  L  Q   C  H  G
1681 ACAGCGCTGG GAGGAGGAGG GACTGGGCGG AGTGCCTGAA CAGAAGCTGC AGTGTCATGG
     TGTCGCGACC CTCCTCCTCC CTGACCCGCC TCACGGACTT GTCTTCGACG TCACAGTACC

 204  Q  G  P   L  A  H  M   P  N  A   S  C  G   V  P  T  L   K  V  Q
1741 GCAAGGGCCC CTGGCCCACA TGCCCAATGC CAGCTGTGGT GTGCCCACGC TGAAGGTCCA
     CGTTCCCGGG GACCGGGTGT ACGGGTTACG GTCGACACCA CACGGGTGCG ACTTCCAGGT

 224  V  P  N   A  S  V  D   V  G  D   D  V  L   L  R  C  Q   V  E  G
1801 GGTGCCCAAT GCCTCGGTGG ATGTGGGGGA CGACGTGCTG CTGCGGTGCC AGGTGGAGGG
     CCACGGGTTA CGGAGCCACC TACACCCCCT GCTGCACGAC GACGCCACGG TCCACCTCCC

 244  R  G  L   E  Q  A  G   W  I  L   T  E  L   E  Q  S  A   T  V  M
1861 GCGGGGCCTG GAGCAGGCCG GCTGGATCCT CACAGAGCTG GAGCAGTCAG CCACGGTGAT
     CGCCCCGGAC CTCGTCCGGC CGACCTAGGA GTGTCTCGAC CTCGTCAGTC GGTGCCACTA

 264  K  S  G   G  L  P  S   L  G  L   T  L  A   N  V  T  S   D  L  N
1921 GAAATCTGGG GGTCTGCCAT CCCTGGGGCT GACCCTGGCC AATGTCACCA GTGACCTCAA
     CTTTAGACCC CCAGACGGTA GGGACCCCGA CTGGGACCGG TTACAGTGGT CACTGGAGTT

 284  R  K  N   L  T  C  W   A  E  N   D  V  G   R  A  E  V   S  V  Q
1981 CAGGAAGAAC TTGACGTGCT GGGCAGAGAA CGATGTGGGC CGGGCAGAGG TCTCTGTTCA
     GTCCTTCTTG AACTGCACGA CCCGTCTCTT GCTACACCCG GCCCGTCTCC AGAGACAAGT

 304  V  N  V   S  F  P  A   S  V  Q   L  H  T   A  V  E  M   H  H  W
2041 GGTCAACGTC TCCTTCCCGG CCAGTGTGCA GCTGCACACG GCGGTGGAGA TGCACCACTG
     CCAGTTGCAG AGGAAGGGCC GGTCACACGT CGACGTGTGC CGCCACCTCT ACGTGGTGAC
```

FIG. 12C

```
 324  C  I  P  F  S  V  D  G  Q  P  A  P  S  L  R  W  L  F  N  G
2101 GTGCATCCCC TTCTCTGTGG ATGGGCAGCC GGCACCGTCT CTGCGCTGGC TCTTCAATGG
     CACGTAGGGG AAGAGACACC TACCCGTCGG CCGTGGCAGA GACGCGACCG AGAAGTTACC

 344  S  V  L  N  E  T  S  F  I  F  T  E  F  L  E  P  A  A  N  E
2161 CTCCGTGCTC AATGAGACCA GCTTCATCTT CACTGAGTTC CTGGAGCCGG CAGCCAATGA
     GAGGCACGAG TTACTCTGGT CGAAGTAGAA GTGACTCAAG GACCTCGGCC GTCGGTTACT

 364  T  V  R  H  G  C  L  R  L  N  Q  P  T  H  V  N  N  G  N  Y
2221 GACCGTGCGG CACGGGTGTC TGCGCCTCAA CCAGCCCACC CACGTCAACA ACGGCAACTA
     CTGGCACGCC GTGCCCACAG ACGCGGAGTT GGTCGGGTGG GTGCAGTTGT TGCCGTTGAT

 384  T  L  L  A  A  N  P  F  G  Q  A  S  A  S  I  M  A  A  F  M
2281 CACGCTGCTG GCTGCCAACC CCTTCGGCCA GGCCTCCGCC TCCATCATGG CTGCCTTCAT
     GTGCGACGAC CGACGGTTGG GGAAGCCGGT CCGGAGGCGG AGGTAGTACC GACGGAAGTA

 404  D  N  P  F  E  F  N  P  E  D  P  I  P  D  T  N  S  T  S  G
2341 GGACAACCCT TTCGAGTTCA ACCCCGAGGA CCCCATCCCT GACACTAACA GCACATCTGG
     CCTGTTGGGA AAGCTCAAGT TGGGGCTCCT GGGGTAGGGA CTGTGATTGT CGTGTAGACC

                          ^begin TM
 424  D  P  V  E  K  K  D  E  T  P  F  G  V  S  V  A  V  G  L  A
2401 AGACCCGGTG GAGAAGAAGG ACGAAACACC TTTTGGGGTC TCGGTGGCTG TGGGCCTGGC
     TCTGGGCCAC CTCTTCTTCC TGCTTTGTGG AAAACCCCAG AGCCACCGAC ACCCGGACCG

                                                ^end TM
 444  V  F  A  C  L  F  L  S  T  L  L  L  V  L  N  K  C  G  R  R
2461 CGTCTTTGCC TGCCTCTTCC TTTCTACGCT GCTCCTTGTG CTCAACAAAT GTGGACGGAG
     GCAGAAACGG ACGGAGAAGG AAAGATGCGA CGAGGAACAC GAGTTGTTTA CACCTGCCTC
```

FIG. 12D

```
 464  N  K  F     G  I  N  R     P  A  V     L  A  P     E  D  G  L     A  M  S
2521  AAACAAGTTT  GGGATCAACC  GCCCGGCTGT  GCTGGCTCCA  GAGGATGGGC  TGGCCATGTC
      TTTGTTCAAA  CCCTAGTTGG  CGGGCCGACA  CGACCGAGGT  CTCCTACCCG  ACCGGTACAG

 484  L  H  F     M  T  L  G     G  S  S     L  S  P     T  E  G  K     G  S  G
2581  CCTGCATTTC  ATGACATTGG  GTGGCAGCTC  CCTGTCCCCC  ACCGAGGGCA  AAGGCTCTGG
      GGACGTAAAG  TACTGTAACC  CACCGTCGAG  GGACAGGGGG  TGGCTCCCGT  TTCCGAGACC

 504  L  Q  G     H  I  I  E     N  P  Q     Y  F  S     D  A  C  V     H  H  I
2641  GCTCCAAGGC  CACATCATCG  AGAACCCACA  ATACTTCAGT  GATGCCTGTG  TTCACCACAT
      CGAGGTTCCG  GTGTAGTAGC  TCTTGGGTGT  TATGAAGTCA  CTACGGACAC  AAGTGGTGTA

 524  K  R  R     D  I  V  L     K  W  E     L  G  E     G  A  F  G     K  V  F
2701  CAAGCGCCGG  GACATCGTGC  TCAAGTGGGA  GCTGGGGGAG  GGCGCCTTTG  GGAAGGTCTT
      GTTCGCGGCC  CTGTAGCACG  AGTTCACCCT  CGACCCCCTC  CCGCGGAAAC  CCTTCCAGAA

 544  L  A  E     C  H  N  L     L  P  E     Q  D  K     M  L  V  A     V  K  A
2761  CCTTGCTGAG  TGCCACAACC  TCCTGCCTGA  GCAGGACAAG  ATGCTGGTGG  CTGTCAAGGC
      GGAACGACTC  ACGGTGTTGG  AGGACGGACT  CGTCCTGTTC  TACGACCACC  GACAGTTCCG

 564  L  K  E     A  S  E  S     A  R  Q     D  F  Q     R  E  A  E     L  L  T
2821  ACTGAAGGAG  GCGTCCGAGA  GTGCTCGGCA  GGACTTCCAA  CGTGAGGCTG  AGCTGCTCAC
      TGACTTCCTC  CGCAGGCTCT  CACGAGCCGT  CCTGAAGGTT  GCACTCCGAC  TCGACGAGTG

 584  M  L  Q     H  Q  H  I     V  R  F     F  G  V     C  T  E  G     R  P  L
2881  CATGCTGCAG  CACCAGCACA  TCGTGCGCTT  CTTCGGCGTC  TGCACCGAGG  GCCGCCCCCT
      GTACGACGTC  GTGGTCGTGT  AGCACGCGAA  GAAGCCGCAG  ACGTGGCTCC  CGGCGGGGGA
```

FIG. 12E

```
 604  L  M  V  F  E  Y  M  R  H  G  D  L  N  R  F  L  R  S  H  G
2941  GCTCATGGTC TTTGAGTATA TGCGGCACGG GGACCTCAAC CGCTTCCTCC GATCCCATGG
      CGAGTACCAG AAACTCATAT ACGCCGTGCC CCTGGAGTTG GCGAAGGAGG CTAGGGTACC

 624  P  D  A  K  L  L  A  G  G  E  D  V  A  P  G  P  L  G  L  G
3001  ACCTGATGCC AAGCTGCTGG CTGGTGGGGA GGATGTGGCT CCAGGCCCCC TGGGTCTGGG
      TGGACTACGG TTCGACGACC GACCACCCCT CCTACACCGA GGTCCGGGGG ACCCAGACCC

 644  Q  L  L  A  V  A  S  Q  V  A  A  G  M  V  Y  L  A  G  L  H
3061  GCAGCTGCTG GCCGTGGCTA GCCAGGTCGC TGCGGGGATG GTGTACCTGG CGGGTCTGCA
      CGTCGACGAC CGGCACCGAT CGGTCCAGCG ACGCCCCTAC CACATGGACC GCCCAGACGT

 664  F  V  H  R  D  L  A  T  R  N  C  L  V  G  Q  G  L  V  V  K
3121  TTTTGTGCAC CGGGACCTGG CCACACGCAA CTGTCTAGTG GGCCAGGGAC TGGTGGTCAA
      AAAACACGTG GCCCTGGACC GGTGTGCGTT GACAGATCAC CCGGTCCCTG ACCACCAGTT

 684  I  G  D  F  G  M  S  R  D  I  Y  S  T  D  Y  Y  R  V  G  G
3181  GATTGGTGAT TTTGGCATGA GCAGGGATAT CTACAGCACC GACTATTACC GTGTGGGAGG
      CTAACCACTA AAACCGTACT CGTCCCTATA GATGTCGTGG CTGATAATGG CACACCCTCC

 704  R  T  M  L  P  I  R  W  M  P  P  E  S  I  L  Y  R  K  F  T
3241  CCGCACCATG CTGCCCATTC GCTGGATGCC GCCCGAGAGC ATCCTGTACC GTAAGTTCAC
      GGCGTGGTAC GACGGGTAAG CGACCTACGG CGGGCTCTCG TAGGACATGG CATTCAAGTG

 724  T  E  S  D  V  W  S  F  G  V  V  L  W  E  I  F  T  Y  G  K
3301  CACCGAGAGC GACGTGTGGA GCTTCGGCGT GGTGCTCTGG GAGATCTTCA CCTACGGCAA
      GTGGCTCTCG CTGCACACCT CGAAGCCGCA CCACGAGACC CTCTAGAAGT GGATGCCGTT

 744  Q  P  W  Y  Q  L  S  N  T  E  A  I  D  C  I  T  Q  G  R  E
```

FIG. 12F

SUBSTITUTE SHEET (RULE 26)

```
3361 GCAGCCCTGG TACCAGCTCT CCAACACGGA GGCAATCGAC TGCATCACGC AGGGACGTGA
     CGTCGGGACC ATGGTCGAGA GGTTGTGCCT CCGTTAGCTG ACGTAGTGCG TCCCTGCACT

 764  L  E  R   P  R  A  C   P  P  E   V  Y  A   I  M  R  G   C  W  Q
3421 GTTGGAGCGG CCACGTGCCT GCCCACCAGA GGTCTACGCC ATCATGCGGG GCTGCTGGCA
     CAACCTCGCC GGTGCACGGA CGGGTGGTCT CCAGATGCGG TAGTACGCCC CGACGACCGT

 784  R  E  P   Q  Q  R  H   S  I  K   D  V  H   A  R  L  Q   A  L  A
3481 GCGGGAGCCC CAGCAACGCC ACAGCATCAA GGATGTGCAC GCCCGGCTGC AAGCCCTGGC
     CGCCCTCGGG GTCGTTGCGG TGTCGTAGTT CCTACACGTG CGGGCCGACG TTCGGGACCG

            R1 site added with cloning primer^
              R1 site removed with cut and fill^
 804  Q  A  P   P  V  Y  L   D  V  L   G  O
3541 CCAGGCACCT CCTGTCTACC TGGATGTCCT GGGCTAGAAT TAATTCAATC GATGGCCGCC
     GGTCCGTGGA GGACAGATGG ACCTACAGGA CCCGATCTTA ATTAAGTTAG CTACCGGCGG

            ^sv40 early poly A
3601 ATGGCCCAAC TTGTTTATTG CAGCTTATAA TGGTTACAAA TAAAGCAATA GCATCACAAA
     TACCGGGTTG AACAAATAAC GTCGAATATT ACCAATGTTT ATTTCGTTAT CGTAGTGTTT
```

FIG. 12G

```
      ^sp6 RNA start
841   TATAGAATAA CATCCACTTT GCCTTTCTCT CCACAGGTGT CCACTCCCAG GTCCAACTGC
      ATATCTTATT GTAGGTGAAA CGGAAAGAGA GGTGTCCACA GGTGAGGGTC CAGGTTGACG

        ^cloning linker    ^begin gD from pchadII
901   ACCTCGGTTC TATCGATTGA ATTCCACCTGC CTTCCACCAA GCTCTGCAGG ATCCCAGAGT
      TGGAGCCAAG ATAGCTAACT TAAGGTGACG GAAGGTGGTT CGAGACGTCC TAGGGTCTCA

961   CAGGGGTCTG TATCTTCCTG CTGGTGGCTC CAGTTCAGGA ACAGTAAACC CTGCTCCGAA
      GTCCCCAGAC ATAGAAGGAC GACCACCGAG GTCAAGTCCT TGTCATTTGG GACGAGGCTT

1021  TATTGCCTCT CACATCTCGT CAATCTCCGC GAGGACTGGG GACCCTGTGA CAAGCTTCAG
      ATAACGGAGA GTGTAGAGCA GTTAGAGGCG CTCCTGACCC CTGGGACACT GTTCGAAGTC

1081  CGCGAACGAC CAACTACCCC GATCATCAGT TATCCTTAAG GTCTCTTTTG TGTGGTGCGT
      GCGCTTGCTG GTTGATGGGG CTAGTAGTCA ATAGGAATTC CAGAGAAAAC ACACCACGCA

            ^Start gD
        1   M* G*  G* T* A*  A* R* L*  G* A* V* I*  L* F* V*  V* I* V*
1141  TCCGGTATGG GGGGGACTGC CGCCAGGTTG GGGGCCGTGA TTTTGTTTGT CGTCATAGTG
      AGGCCATACC CCCCCTGACG GCGGTCCAAC CCCCGGCACT AAAACAAACA GCAGTATCAC

       19  G* L* H* G*  V* R* G*  K* Y* A*  L* A  D  A  S  L  K  M  A  D
1201  GGCCTCCATG GGGTCCGCGG CAAATATGCC TTGGCCGGATG CCTCTCTCAA GATGGCCGAC
      CCGGAGGTAC CCCAGGCGCC GTTTATACGG AACCGCCTAC GGAGAGAGTT CTACCGGCTG
```

FIG. 13A

xho and GTA mutated in^
start mature trkB^

```
  39 P  N  R  F   R  G  K   D  L  P   V  L  D  Q   L  L  E   V  C  P
1261 CCCAATCGAT TTCGCGGCAA AGACCTTCCG GTCCTGGACC AGCTGCTCGA GGTATGTCCC
     GGGTTAGCTA AAGCGCCGTT TCTGGAAGGC CAGGACCTGG TCGACGAGCT CCATACAGGG

  59 T  S  C  K   C  S  A   S  R  I   W  C  S  D   P  S  P   G  I  V
1321 ACGTCCTGCA AATGCAGTGC CTCTCGGATC TGGTGCAGCG ACCCTTCTCC TGGCATCGTG
     TGCAGGACGT TTACGTCACG GAGAGCCTAG ACCACGTCGC TGGGAAGAGG ACCGTAGCAC

  79 A  F  P  R   L  E  P   N  S  V   D  P  E  N   I  T  E   I  F  I
1381 GCATTTCCGA GATTGGAGCC TAACAGTGTA GATCCTGAGA ACATCACCGA AATTTTCATC
     CGTAAAGGCT CTAACCTCGG ATTGTCACAT CTAGGACTCT TGTAGTGGCT TTAAAAGTAG

  99 A  N  Q  K   R  L  E   I  I  N   E  D  D  V   E  A  Y   V  G  L
1441 GCAAACCAGA AAAGGTTAGA AATCATCAAC GAAGATGATG TTGAAGCTTA TGTGGGACTG
     CGTTTGGTCT TTTCCAATCT TTAGTAGTTG CTTCTACTAC AACTTCGAAT ACACCCTGAC

 119 R  N  L  T   I  V  D   S  G  L   K  F  V  A   H  K  A   F  L  K
1501 AGAAATCTGA CAATTGTGGA TTCTGGATTA AAATTTGTGG CTCATAAAGC ATTTCTGAAA
     TCTTTAGACT GTTAACACCT AAGACCTAAT TTTAAACACC GAGTATTTCG TAAAGACTTT

 139 N  S  N  L   Q  H  I   N  F  T   R  N  K  L   T  S  L   S  R  K
1561 AACAGCAACC TGCAGCACAT CAATTTTACC CGAAACAAAC TGACGAGTTT GTCTAGGAAA
     TTGTCGTTGG ACGTCGTGTA GTTAAAATGG GCTTTGTTTG ACTGCTCAAA CAGATCCTTT

 159 H  F  R  H   L  D  L   S  E  L   I  L  V  G   N  P  F   T  C  S
1621 CATTTCCGTC ACCTTGACTT GTCTGAACTG ATCCTGGTGG GCAATCCATT TACATGCTCC
     GTAAAGGCAG TGGAACTGAA CAGACTTGAC TAGGACCACC CGTTAGGTAA ATGTACGAGG
```

FIG. 13B

```
 179 C  D  I  M    W  I  K    T  L  Q    E  A  K  S    S  P  D    T  Q  D
1681 TGTGACATTA TGTGGATCAA GACTCTCCAA GAGGCTAAAT CCAGTCCAGA CACTCAGGAT
     ACACTGTAAT ACACCTAGTT CTGAGAGGTT CTCCGATTTA GGTCAGGTCT GTGAGTCCTA

 199 L  Y  C  L    N  E  S    S  K  N    I  P  L  A    N  L  Q    I  P  N
1741 TTGTACTGCC TGAATGAAAG CAGCAAGAAT ATTCCCCTGG CAAACCTGCA GATACCCAAT
     AACATGACGG ACTTACTTTC GTCGTTCTTA TAAGGGGACC GTTTGGACGT CTATGGGTTA

 219 C  G  L  P    S  A  N    L  A  A    P  N  L  T    V  E  E    G  K  S
1801 TGTGGTTTGC CATCTGCAAA TCTGGCCGCA CCTAACCTCA CTGTGGAGGA AGGAAAGTCT
     ACACCAAACG GTAGACGTTT AGACCGGCGT GGATTGGAGT GACACCTCCT TCCTTTCAGA

 239 I  T  L  S    C  S  V    A  G  D    P  V  P  N    M  Y  W    D  V  G
1861 ATCACATTAT CCTGTAGTGT GGCAGGTGAT CCGGTTCCTA ATATGTATTG GGATGTTGGT
     TAGTGTAATA GGACATCACA CCGTCCACTA GGCCAAGGAT TATACATAAC CCTACAACCA

 259 N  L  V  S    K  H  M    N  E  T    S  H  T  Q    G  S  L    R  I  T
1921 AACCTGGTTT CCAAACATAT GAATGAAACA AGCCACACAC AGGGCTCCTT AAGGATAACT
     TTGGACCAAA GGTTTGTATA CTTACTTTGT TCGGTGTGTG TCCCGAGGAA TTCCTATTGA

 279 N  I  S  S    D  D  S    G  K  Q    I  S  C  V    A  E  N    L  V  G
1981 AACATTTCAT CCGATGACAG TGGGAAGCAG ATCTCTTGTG TGGCGGAAAA TCTTGTAGGA
     TTGTAAAGTA GGCTACTGTC ACCCTTCGTC TAGAGAACAC ACCGCCTTTT AGAACATCCT

 299 E  D  Q  D    S  V  N    L  T  V    H  F  A  P    T  I  T    F  L  E
2041 GAAGATCAAG ATTCTGTCAA CCTCACTGTG CATTTTGCAC CAACTATCAC ATTTCTCGAA
     CTTCTAGTTC TAAGACAGTT GGAGTGACAC GTAAAACGTG GTTGATAGTG TAAAGAGCTT
```

FIG. 13C

SUBSTITUTE SHEET (RULE 26)

SUBSTITUTE SHEET (RULE 26)

```
 319 S  P  T  S    D  H  H    W  C  I    P  F  T  V    K  G  N    P  K  P
2101 TCTCCAACCT CAGACCACCA CTGGTGCATT CCATTCACTG TGAAAGGCAA CCCAAAACCA
     AGAGGTTGGA GTCTGGTGGT GACCACGTAA GGTAAGTGAC ACTTTCCGTT GGGTTTTGGT

 339 A  L  Q  W    F  Y  N    G  A  I    L  N  E  S    K  Y  I    C  T  K
2161 GCGCTTCAGT GGTTCTATAA CGGGGCAATA TTGAATGAGT CCAAATACAT CTGTACTAAA
     CGCGAAGTCA CCAAGATATT GCCCCGTTAT AACTTACTCA GGTTTATGTA GACATGATTT

 359 I  H  V  T    N  H  T    E  Y  H    G  C  L  Q    L  D  N    P  T  H
2221 ATACATGTTA CCAATCACAC GGAGTACCAC GGCTGCCTCC AGCTGGATAA TCCCACTCAC
     TATGTACAAT GGTTAGTGTG CCTCATGGTG CCGACGGAGG TCGACCTATT AGGGTGAGTG

 379 M  N  N  G    D  Y  T    L  I  A    K  N  E  Y    G  K  D    E  K  Q
2281 ATGAACAATG GGGACTACAC TCTAATAGCC AAGAATGAGT ATGGGAAGGA TGAGAAACAG
     TACTTGTTAC CCCTGATGTG AGATTATCGG TTCTTACTCA TACCCTTCCT ACTCTTTGTC

 399 I  S  A  H    F  M  G    W  P  G    I  D  D  G    A  N  P    N  Y  P
2341 ATTTCTGCTC ACTTCATGGG CTGGCCTGGA ATTGACGATG GTGCAAACCC AAATTATCCT
     TAAAGACGAG TGAAGTACCC GACCGGACCT TAACTGCTAC CACGTTTGGG TTTAATAGGA

 419 D  V  I  Y    E  D  Y    G  T  A    A  N  D  I    G  D  T    T  N  R
2401 GATGTAATTT ATGAAGATTA TGGAACTGCA GCGAATGACA TCGGGGACAC CACGAACAGA
     CTACATTAAA TACTTCTAAT ACCTTGACGT CGCTTACTGT AGCCCCTGTG GTGCTTGTCT

 439 S  N  E  I    P  S  T    D  V  T    D  K  T  G    R  E  H    L  S  V
2461 AGTAATGAAA TCCCTTCCAC AGACGTCACT GATAAAACCG GTCGGGAACA TCTCTCGGTC
     TCATTACTTT AGGGAAGGTG TCTGCAGTGA CTATTTTGGC CAGCCCTTGT AGAGAGCCAG

 459 Y  A  V  V    V  I  A    S  V  V    G  F  C  L    L  V  M    L  F  L
2521 TATGCTGTGG TGGTGATTGC GTCTGTGGTG GGATTTTGCC TTTTGGTAAT GCTGTTTCTG
     ATACGACACC ACCACTAACG CAGACACCAC CCTAAAACGG AAAACCATTA CGACAAAGAC
```

FIG. 13D

SUBSTITUTE SHEET (RULE 26)

```
 479 L  K  L  A    R  H  S    K  F  G    M  K  G  P    A  S  V    I  S  N
2581 CTTAAGTTGG CAAGACACTC CAAGTTTGGC ATGAAAGGCC CAGCCTCCGT TATCAGCAAT
     GAATTCAACC GTTCTGTGAG GTTCAAACCG TACTTTCCGG GTCGGAGGCA ATAGTCGTTA

 499 D  D  D  S    A  S  P    L  H  H    I  S  N  G    S  N  T    P  S  S
2641 GATGATGACT CTGCCAGCCC ACTCCATCAC ATCTCCAATG GGAGTAACAC TCCATCTTCT
     CTACTACTGA GACGGTCGGG TGAGGTAGTG TAGAGGTTAC CCTCATTGTG AGGTAGAAGA

 519 S  E  G  G    P  D  A    V  I  I    G  M  T  K    I  P  V    I  E  N
2701 TCGGAAGGTG GCCCAGATGC TGTCATTATT GGAATGACCA AGATCCCTGT CATTGAAAAT
     AGCCTTCCAC CGGGTCTACG ACAGTAATAA CCTTACTGGT TCTAGGGACA GTAACTTTTA

 539 P  Q  Y  F    G  I  T    N  S  Q    L  K  P  D    T  F  V    Q  H  I
2761 CCCCAGTACT TTGGCATCAC CAACAGTCAG CTCAAGCCAG ACACATTTGT TCAGCACATC
     GGGGTCATGA AACCGTAGTG GTTGTCAGTC GAGTTCGGTC TGTGTAAACA AGTCGTGTAG

 559 K  R  H  N    I  V  L    K  R  E    L  G  E  G    A  F  G    K  V  F
2821 AAGCGACATA ACATTGTTCT GAAAAGGGAG CTAGGCGAAG GAGCCTTTGG AAAAGTGTTC
     TTCGCTGTAT TGTAACAAGA CTTTTCCCTC GATCCGCTTC CTCGGAAACC TTTTCACAAG

 579 L  A  E  C    Y  N  L    C  P  E    Q  D  K  I    L  V  A    V  K  T
2881 CTAGCTGAAT GCTATAACCT CTGTCCTGAG CAGGACAAGA TCTTGGTGGC AGTGAAGACC
     GATCGACTTA CGATATTGGA GACAGGACTC GTCCTGTTCT AGAACCACCG TCACTTCTGG

 599 L  K  D  A    S  D  N    A  R  K    D  F  H  R    E  A  E    L  L  T
2941 CTGAAGGATG CCAGTGACAA TGCACGCAAG GACTTCCACC GTGAGGCCGA GCTCCTGACC
     GACTTCCTAC GGTCACTGTT ACGTGCGTTC CTGAAGGTGG CACTCCGGCT CGAGGACTGG
```

FIG. 13E

```
 619 N  L  Q  H    E  H  I    V  K  F    Y  G  V  C    V  E  G    D  P  L
3001 AACCTCCAGC ATGAGCACAT CGTCAAGTTC TATGGCGTCT GCGTGGAGGG CGACCCCCTC
     TTGGAGGTCG TACTCGTGTA GCAGTTCAAG ATACCGCAGA CGCACCTCCC GCTGGGGGAG

 639 I  M  V  F    E  Y  M    K  H  G    D  L  N  K    F  L  R    A  H  G
3061 ATCATGGTCT TTGAGTACAT GAAGCATGGG GACCTCAACA AGTTCCTCAG GGCACACGGC
     TAGTACCAGA AACTCATGTA CTTCGTACCC CTGGAGTTGT TCAAGGAGTC CCGTGTGCCG

 659 P  D  A  V    L  M  A    E  G  N    P  P  T  E    L  T  Q    S  Q  M
3121 CCTGATGCCG TGCTGATGGC TGAGGGCAAC CCGCCCACGG AACTGACGCA GTCGCAGATG
     GGACTACGGC ACGACTACCG ACTCCCGTTG GGCGGGTGCC TTGACTGCGT CAGCGTCTAC

 679 L  H  I  A    Q  Q  I    A  A  G    M  V  Y  L    A  S  Q    H  F  V
3181 CTGCATATAG CCCAGCAGAT CGCCGCGGGC ATGGTCTACC TGGCGTCCCA GCACTTCGTG
     GACGTATATC GGGTCGTCTA GCGGCGCCCG TACCAGATGG ACCGCAGGGT CGTGAAGCAC

 699 H  R  D  L    A  T  R    N  C  L    V  G  E  N    L  L  V    K  I  G
3241 CACCGCGATT TGGCCACCAG GAACTGCCTG GTCGGGGAGA ACTTGCTGGT GAAAATCGGG
     GTGGCGCTAA ACCGGTGGTC CTTGACGGAC CAGCCCCTCT TGAACGACCA CTTTTAGCCC

 719 D  F  G  M    S  R  D    V  Y  S    T  D  Y  Y    R  V  G    G  H  T
3301 GACTTTGGGA TGTCCCGGGA CGTGTACAGC ACTGACTACT ACAGGGTCGG TGGCCACACA
     CTGAAACCCT ACAGGGCCCT GCACATGTCG TGACTGATGA TGTCCCAGCC ACCGGTGTGT

 739 M  L  P  I    R  W  M    P  P  E    S  I  M  Y    R  K  F    T  T  E
3361 ATGCTGCCCA TTCGCTGGAT GCCTCCAGAG AGCATCATGT ACAGGAAATT CACGACGGAA
     TACGACGGGT AAGCGACCTA CGGAGGTCTC TCGTAGTACA TGTCCTTTAA GTGCTGCCTT

 759 S  D  V  W    S  L  G    V  V  L    W  E  I  F    T  Y  G    K  Q  P
3421 AGCGACGTCT GGAGCCTGGG GGTCGTGTTG TGGGAGATTT TCACCTATGG CAAACAGCCC
```

FIG. 13F

SUBSTITUTE SHEET (RULE 26)

```
     TCGCTGCAGA CCTCGGACCC CCAGCACAAC ACCCTCTAAA AGTGGATACC GTTTGTCGGG

 779 W  Y  Q  L   S  N  N   E  V  I   E  C  I  T   Q  G  R   V  L  Q
3481 TGGTACCAGC TGTCAAACAA TGAGGTGATA GAGTGTATCA CTCAGGGCCG AGTCCTGCAG
     ACCATGGTCG ACAGTTTGTT ACTCCACTAT CTCACATAGT GAGTCCCGGC TCAGGACGTC

 799 R  P  R  T   C  P  Q   E  V  Y   E  L  M  L   G  C  W   Q  R  E
3541 CGACCCCGCA CGTGCCCCCA GGAGGTGTAT GAGCTGATGC TGGGGTGCTG GCAGCGAGAG
     GCTGGGGCGT GCACGGGGGT CCTCCACATA CTCGACTACG ACCCCACGAC CGTCGCTCTC

 819 P  H  M  R   K  N  I   K  G  I   H  T  L  L   Q  N  L   A  K  A
3601 CCCCACATGA GGAAGAACAT CAAGGGCATC CATACCCTCC TTCAGAACTT GGCCAAGGCA
     GGGGTGTACT CCTTCTTGTA GTTCCCGTAG GTATGGGAGG AAGTCTTGAA CCGGTTCCGT

 839 S  P  V  Y   L  D  I   L  G  O
3661 TCTCCGGTCT ACCTGGACAT TCTAGGCTAG GGCCCTTTTC CCCAGACCGA TCCTTCCCAA
     AGAGGCCAGA TGGACCTGTA AGATCCGATC CCGGGAAAAG GGGTCTGGCT AGGAAGGGTT

3721 CGTACTCCTC AGACGGGCTG AGAGGATGAA CATCTTTTAA CTGCCGCTGG AGGCCACCAA
     GCATGAGGAG TCTGCCCGAC TCTCCTACTT GTAGAAAATT GACGGCGACC TCCGGTGGTT

                          half Xho half Sal site from subcloning^
3781 GCTGCTCTCC TTCACTCTGA CAGTATTAAC ATCAAAGACT CCGAGAAGCT CTCGACCTGC
     CGACGAGAGG AAGTGAGACT GTCATAATTG TAGTTTCTGA GGCTCTTCGA GAGCTGGACG

                                ^sv40 early poly A
3841 AGAAGCTTGG CCGCCATGGC CCAACTTGTT TATTGCAGCT TATAATGGTT ACAAATAAAG
     TCTTCGAACC GGCGGTACCG GGTTGAACAA ATAACGTCGA ATATTACCAA TGTTTATTTC
```

FIG. 13G

SUBSTITUTE SHEET (RULE 26)

```
          ^sp6 RNA start
  841 TATAGAATAA CATCCACTTT GCCTTTCTCT CCACAGGTGT CCACTCCCAG GTCCAACTGC
      ATATCTTATT GTAGGTGAAA CGGAAAGAGA GGTGTCCACA GGTGAGGGTC CAGGTTGACG

          ^cloning linker
             ^RI site mutated in
             ^gD from pchadII
  901 ACCTGAATTC CACTGCCTTC CACCAAGCTC TGCAGGATCC CAGAGTCAGG GGTCTGTATC
      TGGACTTAAG GTGACGGAAG GTGGTTCGAG ACGTCCTAGG GTCTCAGTCC CCAGACATAG

  961 TTCCTGCTGG TGGCTCCAGT TCAGGAACAG TAAACCCTGC TCCGAATATT GCCTCTCACA
      AAGGACGACC ACCGAGGTCA AGTCCTTGTC ATTTGGGACG AGGCTTATAA CGGAGAGTGT

 1021 TCTCGTCAAT CTCCGCGAGG ACTGGGGACC CTGTGACAAG CTTCAGCGCG AACGACCAAC
      AGAGCAGTTA GAGGCGCTCC TGACCCCTGG GACACTGTTC GAAGTCGCGC TTGCTGGTTG

                                                          ^Start
gD
    1                                                     M*  G*  G*
 1081 TACCCCGATC ATCAGTTATC CTTAAGGTCT CTTTTGTGTG GTGCGTTCCG GTATGGGGGG
      ATGGGGCTAG TAGTCAATAG GAATTCCAGA GAAAACACAC CACGCAAGGC CATACCCCCC

    4  T*  A*  A*   R*  L*  G*  A*   V*  I*  L*   F*  V*  V*   I*  V*  G*  L*   H*  G*  V*
 1141 GACTGCCGCC AGGTTGGGGG CCGTGATTTT GTTTGTCGTC ATAGTGGGCC TCCATGGGGT
      CTGACGGCGG TCCAACCCCC GGCACTAAAA CAAACAGCAG TATCACCCGG AGGTACCCCA

   24  R*  G*  K*   Y*  A*  L*  A   D   A   S   L   K   M   A   D   P   N   R   F   R
 1201 CCGCGGCAAA TATGCCTTGG CGGATGCCTC TCTCAAGATG GCCGACCCCA ATCGATTTCG
      GGCGCCGTTT ATACGGAACC GCCTACGGAG AGAGTTCTAC CGGCTGGGGT TAGCTAAAGC
```

FIG. 14A

```
                                                  ^Xho site and GTA mutated in
                                                                ^begin mature trkC
  44  G  K  D     L  P  V  L     D  Q  L     L  E  V     C  P  A  N     C  V  C
1261  CGGCAAAGAC  CTTCCGGTCC  TGGACCAGCT  GCTCGAGGTA  TGCCCTGCAA  ATTGTGTCTG
      GCCGTTTCTG  GAAGGCCAGG  ACCTGGTCGA  CGAGCTCCAT  ACGGGACGTT  TAACACAGAC

  64  S  K  T     E  I  N  C     R  R  P     D  D  G     N  L  F  P     L  L  E
1321  CAGCAAGACT  GAGATCAATT  GCCGGCGGCC  GGACGATGGG  AACCTCTTCC  CCCTCCTGGA
      GTCGTTCTGA  CTCTAGTTAA  CGGCCGCCGG  CCTGCTACCC  TTGGAGAAGG  GGGAGGACCT

  84  G  Q  D     S  G  N  S     N  G  N     A  N  I     N  I  T  D     I  S  R
1381  AGGGCAGGAT  TCAGGGAACA  GCAATGGGAA  CGCCAATATC  AACATCACGG  ACATCTCAAG
      TCCCGTCCTA  AGTCCCTTGT  CGTTACCCTT  GCGGTTATAG  TTGTAGTGCC  TGTAGAGTTC

 104  N  I  T     S  I  H  I     E  N  W     R  S  L     H  T  L  N     A  V  D
1441  GAATATCACT  TCCATACACA  TAGAGAACTG  GCGCAGTCTT  CACACGCTCA  ACGCCGTGGA
      CTTATAGTGA  AGGTATGTGT  ATCTCTTGAC  CGCGTCAGAA  GTGTGCGAGT  TGCGGCACCT

 124  M  E  L     Y  T  G  L     Q  K  L     T  I  K     N  S  G  L     R  S  I
1501  CATGGAGCTC  TACACCGGAC  TTCAAAAGCT  GACCATCAAG  AACTCAGGAC  TTCGGAGCAT
      GTACCTCGAG  ATGTGGCCTG  AAGTTTTCGA  CTGGTAGTTC  TTGAGTCCTG  AAGCCTCGTA

 144  Q  P  R     A  F  A  K     N  P  H     L  R  Y     I  N  L  S     S  N  R
1561  TCAGCCCAGA  GCCTTTGCCA  AGAACCCCCA  TTTGCGTTAT  ATAAACCTGT  CAAGTAACCG
      AGTCGGGTCT  CGGAAACGGT  TCTTGGGGGT  AAACGCAATA  TATTTGGACA  GTTCATTGGC

 164  L  T  T     L  S  W  Q     L  F  Q     T  L  S     L  R  E  L     Q  L  E
1621  GCTCACCACA  CTCTCGTGGC  AGCTCTTCCA  GACGCTGAGT  CTTCGGGAAT  TGCAGTTGGA
```

FIG. 14B

SUBSTITUTE SHEET (RULE 26)

```
      CGAGTGGTGT GAGAGCACCG TCGAGAAGGT CTGCGACTCA GAAGCCCTTA ACGTCAACCT

 184  Q  N  F   F  N  C  S   C  D  I   R  W  M   Q  L  W  Q   E  Q  G
1681  GCAGAACTTT TTCAACTGCA GCTGTGACAT CCGCTGGATG CAGCTCTGGC AGGAGCAGGG
      CGTCTTGAAA AAGTTGACGT CGACACTGTA GGCGACCTAC GTCGAGACCG TCCTCGTCCC

 204  E  A  K   L  N  S  Q   N  L  Y   C  I  N   A  D  G  S   Q  L  P
1741  GGAGGCCAAG CTCAACAGCC AGAACCTCTA CTGCATCAAT GCTGATGGCT CCCAGCTTCC
      CCTCCGGTTC GAGTTGTCGG TCTTGGAGAT GACGTAGTTA CGACTACCGA GGGTCGAAGG

 224  L  F  R   M  N  I  S   Q  C  D   L  P  E   I  S  V  S   H  V  N
1801  TCTCTTCCGC ATGAACATCA GTCAGTGTGA CCTTCCTGAG ATCAGCGTGA GCCACGTCAA
      AGAGAAGGCG TACTTGTAGT CAGTCACACT GGAAGGACTC TAGTCGCACT CGGTGCAGTT

 244  L  T  V   R  E  G  D   N  A  V   I  T  C   N  G  S  G   S  P  L
1861  CCTGACCGTA CGAGAGGGTG ACAATGCTGT TATCACTTGC AATGGCTCTG GATCACCCCT
      GGACTGGCAT GCTCTCCCAC TGTTACGACA ATAGTGAACG TTACCGAGAC CTAGTGGGGA

 264  P  D  V   D  W  I  V   T  G  L   Q  S  I   N  T  H  Q   T  N  L
1921  TCCTGATGTG GACTGGATAG TCACTGGGCT GCAGTCCATC AACACTCACC AGACCAATCT
      AGGACTACAC CTGACCTATC AGTGACCCGA CGTCAGGTAG TTGTGAGTGG TCTGGTTAGA

 284  N  W  T   N  V  H  A   I  N  L   T  L  V   N  V  T  S   E  D  N
1981  GAACTGGACC AATGTTCATG CCATCAACTT GACGCTGGTG AATGTGACGA GTGAGGACAA
      CTTGACCTGG TTACAAGTAC GGTAGTTGAA CTGCGACCAC TTACACTGCT CACTCCTGTT

 304  G  F  T   L  T  C  I   A  E  N   V  V  G   M  S  N  A   S  V  A
2041  TGGCTTCACC CTGACGTGCA TTGCAGAGAA CGTGGTGGGC ATGAGCAATG CCAGTGTTGC
      ACCGAAGTGG GACTGCACGT AACGTCTCTT GCACCACCCG TACTCGTTAC GGTCACAACG
```

FIG. 14C

SUBSTITUTE SHEET (RULE 26)

```
 324  L  T  V   Y  Y  P  P   R  V  V   S  L  E   E  P  E  L   R  L  E
2101  CCTCACTGTC TACTATCCCC CACGTGTGGT GAGCCTGGAG GAGCCTGAGC TGCGCCTGGA
      GGAGTGACAG ATGATAGGGG GTGCACACCA CTCGGACCTC CTCGGACTCG ACGCGGACCT

 344  H  C  I   E  F  V  V   R  G  N   P  P  P   T  L  H  W   L  H  N
2161  GCACTGCATC GAGTTTGTGG TGCGTGGCAA CCCCCCACCA ACGCTGCACT GGCTGCACAA
      CGTGACGTAG CTCAAACACC ACGCACCGTT GGGGGGTGGT TGCGACGTGA CCGACGTGTT

 364  G  Q  P   L  R  E  S   K  I  I   H  V  E   Y  Y  Q  E   G  E  I
2221  TGGGCAGCCT CTGCGGGAGT CCAAGATCAT CCATGTGGAA TACTACCAAG AGGGAGAGAT
      ACCCGTCGGA GACGCCCTCA GGTTCTAGTA GGTACACCTT ATGATGGTTC TCCCTCTCTA

 384  S  E  G   C  L  L  F   N  K  P   T  H  Y   N  N  G  N   Y  T  L
2281  TTCCGAGGGC TGCCTGCTCT TCAACAAGCC CACCCACTAC AACAATGGCA ACTATACCCT
      AAGGCTCCCG ACGGACGAGA AGTTGTTCGG GTGGGTGATG TTGTTACCGT TGATATGGGA

 404  I  A  K   N  P  L  G   T  A  N   Q  T  I   N  G  H  F   L  K  E
2341  CATTGCCAAA AACCCACTGG GCACAGCCAA CCAGACCATC AATGGCCACT TCCTCAAGGA
      GTAACGGTTT TTGGGTGACC CGTGTCGGTT GGTCTGGTAG TTACCGGTGA AGGAGTTCCT

                ^begin ecd insert          ^end ecd insert
 424  P  F  P   E  S  T  D   N  F  I   L  F  D   E  V  S  P   T  P  P
2401  GCCCTTTCCA GAGAGCACGG ATAACTTTAT CTTGTTTGAC GAAGTGAGTC CCACACCTCC
      CGGGAAAGGT CTCTCGTGCC TATTGAAATA GAACAAACTG CTTCACTCAG GGTGTGGAGG

                                        ^begin TM
 444  I  T  V   T  H  K  P   E  E  D   T  F  G   V  S  I  A   V  G  L
2461  TATCACTGTG ACCCACAAAC CAGAAGAAGA CACTTTTGGG GTATCCATAG CAGTTGGACT
```

FIG. 14D

SUBSTITUTE SHEET (RULE 26)

```
     ATAGTGACAC TGGGTGTTTG GTCTTCTTCT GTGAAAACCC CATAGGTATC GTCAACCTGA

                                                          ^end TM
 464  A  A  F  A  C  V  L  L  V  V  L  F  V  M  I  N  K  Y  G  R
2521 TGCTGCTTTT GCCTGTGTCC TGTTGGTGGT TCTCTTCGTC ATGATCAACA AATATGGTCG
     ACGACGAAAA CGGACACAGG ACAACCACCA AGAGAAGCAG TACTAGTTGT TTATACCAGC

 484  R  S  K  F  G  M  K  G  P  V  A  V  I  S  G  E  E  D  S  A
2581 ACGGTCCAAA TTTGGAATGA AGGGTCCCGT GGCTGTCATC AGTGGTGAGG AGGACTCAGC
     TGCCAGGTTT AAACCTTACT TCCCAGGGCA CCGACAGTAG TCACCACTCC TCCTGAGTCG

 504  S  P  L  H  H  I  N  H  G  I  T  T  P  S  S  L  D  A  G  P
2641 CAGCCCACTG CACCACATCA ACCACGGCAT CACCACGCCC TCGTCACTGG ATGCCGGGCC
     GTCGGGTGAC GTGGTGTAGT TGGTGCCGTA GTGGTGCGGG AGCAGTGACC TACGGCCCGG

 524  D  T  V  V  I  G  M  T  R  I  P  V  I  E  N  P  Q  Y  F  R
2701 CGACACTGTG GTCATTGGCA TGACTCGCAT CCCTGTCATT GAGAACCCCC AGTACTTCCG
     GCTGTGACAC CAGTAACCGT ACTGAGCGTA GGGACAGTAA CTCTTGGGGG TCATGAAGGC

 544  Q  G  H  N  C  H  K  P  D  T  Y  V  Q  H  I  K  R  R  D  I
2761 TCAGGGACAC AACTGCCACA AGCCGGACAC GTATGTGCAG CACATTAAGA GGAGAGACAT
     AGTCCCTGTG TTGACGGTGT TCGGCCTGTG CATACACGTC GTGTAATTCT CCTCTCTGTA

                    ^begin TK
 564  V  L  K  R  E  L  G  E  G  A  F  G  K  V  F  L  A  E  C  Y
2821 CGTGCTGAAG CGAGAACTGG GTGAGGGAGC CTTTGGAAAG GTCTTCCTGG CCGAGTGCTA
     GCACGACTTC GCTCTTGACC CACTCCCTCG GAAACCTTTC CAGAAGGACC GGCTCACGAT
```

FIG. 14E

SUBSTITUTE SHEET (RULE 26)

```
 584  N  L  S   P  T  K  D   K  M  L   V  A  V   K  A  L  K   D  P  T
2881 CAACCTCAGC CCGACCAAGG ACAAGATGCT TGTGGCTGTG AAGGCCCTGA AGGATCCCAC
     GTTGGAGTCG GGCTGGTTCC TGTTCTACGA ACACCGACAC TTCCGGGACT TCCTAGGGTG

 604  L  A  A   R  K  D  F   Q  R  E   A  E  L   L  T  N  L   Q  H  E
2941 CCTGGCTGCC CGGAAGGATT TCCAGAGGGA GGCCGAGCTG CTCACCAACC TGCAGCATGA
     GGACCGACGG GCCTTCCTAA AGGTCTCCCT CCGGCTCGAC GAGTGGTTGG ACGTCGTACT

 624  H  I  V   K  F  Y  G   V  C  G   D  G  D   P  L  I  M   V  F  E
3001 GCACATTGTC AAGTTCTATG GAGTGTGCGG CGATGGGGAC CCCCTCATCA TGGTCTTTGA
     CGTGTAACAG TTCAAGATAC CTCACACGCC GCTACCCCTG GGGGAGTAGT ACCAGAAACT

 644  Y  M  K   H  G  D  L   N  K  F   L  R  A   H  G  P  D   A  M  I
3061 ATACATGAAG CATGGAGACC TGAATAAGTT CCTCAGGGCC CATGGGCCAG ATGCAATGAT
     TATGTACTTC GTACCTCTGG ACTTATTCAA GGAGTCCCGG GTACCCGGTC TACGTTACTA

 664  L  V  D   G  Q  P  R   Q  A  K   G  E  L   G  L  S  Q   M  L  H
3121 CCTTGTGGAT GGACAGCCAC GCCAGGCCAA GGGTGAGCTG GGGCTCTCCC AAATGCTCCA
     GGAACACCTA CCTGTCGGTG CGGTCCGGTT CCCACTCGAC CCCGAGAGGG TTTACGAGGT

 684  I  A  S   Q  I  A  S   G  M  V   Y  L  A   S  Q  H  F   V  H  R
3181 CATTGCCAGT CAGATCGCCT CGGGTATGGT GTACCTGGCC TCCCAGCACT TTGTGCACCG
     GTAACGGTCA GTCTAGCGGA GCCCATACCA CATGGACCGG AGGGTCGTGA AACACGTGGC

 704  D  L  A   T  R  N  C   L  V  G   A  N  L   L  V  K  I   G  D  F
3241 AGACCTGGCC ACCAGGAACT GCCTGGTTGG AGCGAATCTG CTAGTGAAGA TTGGGGACTT
     TCTGGACCGG TGGTCCTTGA CGGACCAACC TCGCTTAGAC GATCACTTCT AACCCCTGAA

                                                ^TK insert site
 724  G  M  S   R  D  V  Y   S  T  D   Y  Y  R   V  G  G  H   T  M  L
```

FIG. 14F

```
3301 CGGCATGTCC AGAGATGTCT ACAGCACGGA TTATTACAGG GTGGGAGGAC ACACCATGCT
     GCCGTACAGG TCTCTACAGA TGTCGTGCCT AATAATGTCC CACCCTCCTG TGTGGTACGA

 744  P  I  R  W  M  P  P  E  S  I  M  Y  R  K  F  T  T  E  S  D
3361 CCCCATTCGC TGGATGCCTC CTGAAAGCAT CATGTACCGG AAGTTCACTA CAGAGAGTGA
     GGGGTAAGCG ACCTACGGAG GACTTTCGTA GTACATGGCC TTCAAGTGAT GTCTCTCACT

 764  V  W  S  F  G  V  I  L  W  E  I  F  T  Y  G  K  Q  P  W  F
3421 TGTATGGAGC TTCGGGGTGA TCCTCTGGGA GATCTTCACC TATGGAAAGC AGCCATGGTT
     ACATACCTCG AAGCCCCACT AGGAGACCCT CTAGAAGTGG ATACCTTTCG TCGGTACCAA

 784  Q  L  S  N  T  E  V  I  E  C  I  T  Q  G  R  V  L  E  R  P
3481 CCAACTCTCA AACACGGAGG TCATTGAGTG CATTACCCAA GGTCGTGTTT TGGAGCGGCC
     GGTTGAGAGT TTGTGCCTCC AGTAACTCAC GTAATGGGTT CCAGCACAAA ACCTCGCCGG

 804  R  V  C  P  K  E  V  Y  D  V  M  L  G  C  W  Q  R  E  P  Q
3541 CCGAGTCTGC CCCAAAGAGG TGTACGATGT CATGCTGGGG TGCTGGCAGA GGGAACCACA
     GGCTCAGACG GGGTTTCTCC ACATGCTACA GTACGACCCC ACGACCGTCT CCCTTGGTGT

 824  Q  R  L  N  I  K  E  I  Y  K  I  L  H  A  L  G  K  A  T  P
3601 GCAGCGGTTG AACATCAAGG AGATCTACAA AATCCTCCAT GCTTTGGGGA AGGCCACCCC
     CGTCGCCAAC TTGTAGTTCC TCTAGATGTT TTAGGAGGTA CGAAACCCCT TCCGGTGGGG

                          ^stop
                R1 site removed with cut and fill^
 844  I  Y  L  D  I  L  G  O
3661 AATCTACCTG GACATTCTTG GCTAGTGGTG GCTGGTGGTC ATGAATTAAT TCAATCGATG
     TTAGATGGAC CTGTAAGAAC CGATCACCAC CGACCACCAG TACTTAATTA AGTTAGCTAC

                   ^sv40 early poly A
3721 GCCGCCATGG CCCAACTTGT TTATTGCAGC TTATAATGGT TACAAATAAA GCAATAGCAT
     CGGCGGTACC GGGTTGAACA AATAACGTCG AATATTACCA ATGTTTATTT CGTTATCGTA
```

FIG. 14G

gD.TrkA

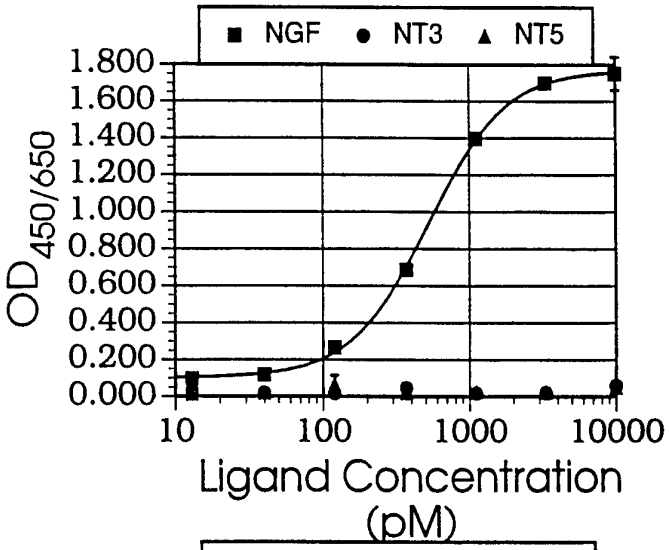


FIG. 15A

gD.TrkB

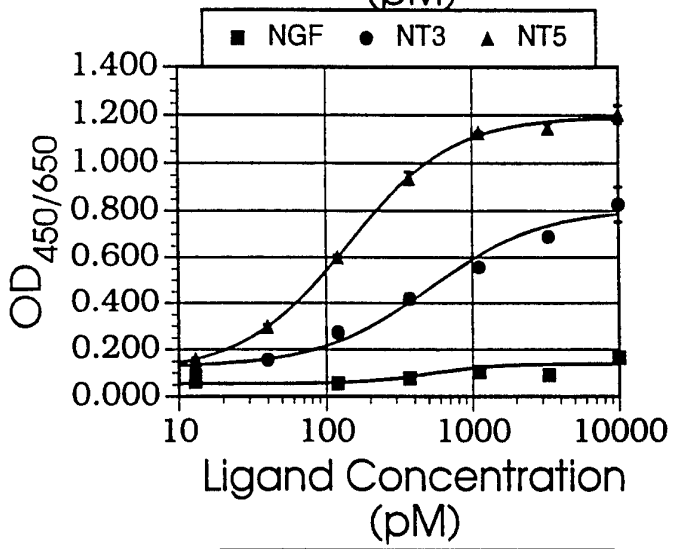


FIG. 15B

gD.TrkC

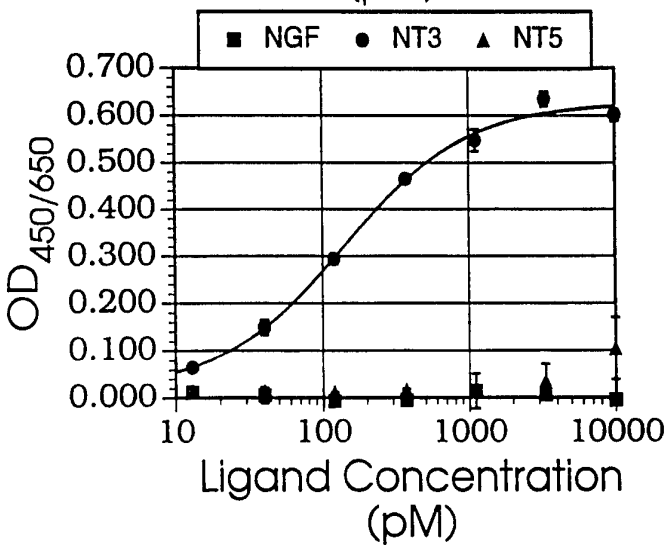


FIG. 15C

```
                                                
```

```
                    nlaIV
          scrFI                                sfaNI
               mvaI                                     ppul0I
          ecoRII                            nsiI/avaIII
          dsaV                             nlaIII
               bstNI                                 sphI
  [dcm+]       apyI[dcm+]                            nspI
              bsaJI                                  nspHI
141 GTGTGGAAAG TCCCCAGGCT CCCCAGCAGG CAGAAGTATG CAAAGCATGC ATCTCAATTA GTCAGCAACC
    CACACCTTTC AGGGGTCCGA GGGGTCGTCC GTCTTCATAC GTTTCGTACG TAGAGTTAAT CAGTCGTTGG

                                                                        nlaIII
                                                                       styI
                                                                       ncoI
                                  aciI                                bslI dsaI
         aciI          aciI  fokI              aciI bsrI  aciI         aciI bsaJI
211 ATAGTCCCGC CCCTAACTCC GCCCATCCCG CCCCTAACTC CGCCCAGTTC CGCCCATTCT CCGCCCCATG
    TATCAGGGCG GGGATTGAGG CGGGTAGGGC GGGGATTGAG GCGGGTCAAG GCGGGTAAGA GGCGGGGTAC

                                        fnu4HI
                                        bglI
                                       sfiI
                                       haeIII/palI
                                     mnlI   mnlI      ddeI
                                haeIII/palI bsaJI  mnlI   aluI
                              mnlI bsaJI aciI    haeIII/palI                 mnlI
281 GCTGACTAAT TTTTTTTATT TATGCAGAGG CCGAGGCCGC CTCGGCCTCT GAGCTATTCC AGAAGTAGTG
    CGACTGATTA AAAAAAATAA ATACGTCTCC GGCTCCGGCG GAGCCGGAGA CTCGATAAGG TCTTCATCAC
```

FIG. 16B

```
                                                
```

```
                                  haeIII/palI
                                 haeI
                               scrFI
                               mvaI     bsrBI
                               ecoRII
                               dsaV
                               bstNI    aciI                  rsaI
                     bsmAI     apyI[dcm+]        xmnI        csp6I
                     bsaI     bsaJI  mnlI  ddeI  asp700     scaI
541 GGGATTGGCA AGAACGGAGA CCTACCCTGG CCTCCGCTCA GGAACGAGTT CAAGTACTTC CAAAGAATGA
    CCCTAACCGT TCTTGCCTCT GGATGGGACC GGAGGCGAGT CCTTGCTCAA GTTCATGAAG GTTTCTTACT

                                                                scrFI
                                                                mvaI
                                                                ecoRII
             eco57I                                             dsaV
            mboII                 tfiI                          bstNI
           earI/ksp632I           hinfI                         apyI[dcm+]
          mnlI                  alwNI    hphI                  sexAI            ddeI
611 CCACAACCTC TTCAGTGGAA GGTAAACAGA ATCTGGTGAT TATGGGTAGG AAAACCTGGT TCTCCATTCC
    GGTGTTGGAG AAGTCACCTT CCATTTGTCT TAGACCACTA ATACCCATCC TTTTGGACCA AGAGGTAAGG

          tfiI       tru9I         tru9I
          hinfI      mseI          mseI         ddeI
       mboII taqI   ahaIII/draI   aseI/asnI/vspI                      bslI   mnlI
681 TGAGAAGAAT CGACCTTTAA AGGACAGAAT TAATATAGTT CTCAGTAGAG AACTCAAAGA ACCACCACGA
    ACTCTTCTTA GCTGGAAATT TCCTGTCTTA ATTATATCAA GAGTCATCTC TTGAGTTTCT TGGTGGTGCT
```

FIG. 16D

```
     sstI
     sacI
     hgiJII
     hgiAI/aspHI
     ecl136II
     bsp1286
     bsiHKAI
     bmyI                                  tru9I                 mspI
     banII                                aflII/bfrI             hpaII
      aluI           bstXI      fokI sfaNI mseI                 bsaWI
751 GGAGCTCATT TTCTTGCCAA AAGTTTGGAT GATGCCTTAA GACTTATTGA ACAACCGGAA TTGGCAAGTA
    CCTCGAGTAA AAGAACGGTT TTCAAACCTA CTACGGAATT CTGAATAACT TGTTGGCCTT AACCGTTCAT

                                                                        haeIII/palI
                                                                       haeI
                                                 scrFI          scrFI
                                                 mvaI                 mvaI
                                                 ecoRII         ecoRII
                                                 dsaV   tfiI    dsaV
                                                 bstNI    nlaIII     bstNI     ddeI
      accI nlaIII             mnlI               apyI[dcm+]   hinfI  apyI[dcm+]
821 AAGTAGACAT GGTTTGGATA GTCGGAGGCA GTTCTGTTTA CCAGGAAGCC ATGAATCAAC CAGGCCACCT
    TTCATCTGTA CCAAACCTAT CAGCCTCCGT CAAGACAAAT GGTCCTTCGG TACTTAGTTG GTCCGGTGGA
```

FIG. 16E

```
                                                
```

```
                                                
```

```
                    styI
                    bsaJI
              sau3AI                    fnu4HI
              mboI/ndeII[dam-]          aciI
              dpnI[dam+]               thaI
              dpnII[dam-]              fnuDII/mvnI tru9I
              alwI[dam-]               bstUI       mseI
             bstYI/xhoII               bsh1236I   aseI/asnI/vspI
1131 CTGGCTTTAG ATCCCCTTGG CTTCGTTAGA ACGCGGCTAC AATTAATACA TAACCTTATG TATCATACAC
     GACCGAAATC TAGGGGAACC GAAGCAATCT TGCGCCGATG TTAATTATGT ATTGGAATAC ATAGTATGTG

                                                                          sau96I
                                                                          avaII
                                                                          asuI
                                                                       scrFI
                                                                       mvaI
                                                                       ecoRII
                                                                       dsaV
                                                                       bstNI
                maeIII                                                 apyI[dcm+]
               hphI   scfI       fokI                           bslI  bsaJI
1201 ATACGATTTA GGTGACACTA TAGATAACAT CCACTTTGCC TTTCTCTCCA CAGGTGTCCA CTCCCAGGTC
     TATGCTAAAT CCACTGTGAT ATCTATTGTA GGTGAAACGG AAAGAGAGGT GTCCACAGGT GAGGGTCCAG
```

FIG. 16H

```
                                                
```

```
                                                
```

```
                                                
```

FIG. 16L

```
                                                                  nlaIV
                                                               scrFI
                                                               mvaI
                                                               ecoRII
                                                               dsaV
                                                               bstNI
          aluI                                                 apyI[dcm+]
         pvuII                                                bsaJI
         nspBII
1571 AACCAGCTGT GGAATGTGTG TCAGTTAGGG TGTGGAAAGT CCCCAGGCTC CCCAGCAGGC AGAAGTATGC
     TTGGTCGACA CCTTACACAC AGTCAATCCC ACACCTTTCA GGGGTCCGAG GGGTCGTCCG TCTTCATACG

                                                                  nlaIV
      sfaNI                  scrFI        scrFI
          ppu10I                   mvaI                  mvaI
    nsiI/avaIII              ecoRII       ecoRII
   nlaIII                    dsaV         dsaV
        sphI                       bstNI                 bstNI
        nspI                       apyI[dcm+]            apyI[dcm+]
        nspHI                     sexAI                 bsaJI
1641 AAAGCATGCA TCTCAATTAG TCAGCAACCA GGTGTGGAAA GTCCCCAGGC TCCCCAGCAG GCAGAAGTAT
     TTTCGTACGT AGAGTTAATC AGTCGTTGGT CCACACCTTT CAGGGGTCCG AGGGGTCGTC CGTCTTCATA

              sfaNI
            ppu10I
            nsiI/avaIII
           nlaIII
          sphI
          nspI                                                                aciI
          nspHI                                  aciI          aciI  fokI
1711 GCAAAGCATG CATCTCAATT AGTCAGCAAC CATAGTCCCG CCCCTAACTC CGCCCATCCC GCCCCTAACT
     CGTTTCGTAC GTAGAGTTAA TCAGTCGTTG GTATCAGGGC GGGGATTGAG GCGGGTAGGG CGGGGATTGA
```

```
                                    nlaIII
                                   styI
                                   ncoI
          bsrI                 bslI dsaI
      asiI       aciI          aciI bsaJI                                  mnlI
1781  CCGCCCAGTT CCGCCCATTC TCCGCCCCAT GGCTGACTAA TTTTTTTTAT TTATGCAGAG
      GGCGGGTCAA GGCGGGTAAG AGGCGGGGTA CCGACTGATT AAAAAAAATA AATACGTCTC

                                                                      styI
            fnu4HI                                                    bsaJI
            bglI                                                      blnI
           sfiI                                                       avrII
           haeIII/palI                                              haeIII/palI
         mnlI    mnlI     ddeI                                      stuI
    haeIII/palI  bsaJI mnlI   aluI                   mnlI            haeI
       bsaJI aciI    haeIII/palI                  mnlI             mnlI
1841  GCCGAGGCCG CCTCGGCCTC TGAGCTATTC CAGAAGTAGT GAGGAGGCTT TTTTGGAGGC
      CGGCTCCGGC GGAGCCGGAG ACTCGATAAG GTCTTCATCA CTCCTCCGAA AAAACCTCCG
```

FIG. 16M

```
                                                
```

```
                                                
```

```
                                                                  hinPI
                                                                 thaI
                                                                 fnuDII/mvnI
                                                                 bstUI scfI     hinPI
                                                                 bsh1236I       hhaI/cfoI
                                                            rsaI hhaI/cfoI  fnu4HI
        aciI          aciI   maeII                         csp6I   bslI    aciI
2171 TGTGCGGTAT TTCACACCGC ATACGTCAAA GCAACCATAG TACGCGCCCT GTAGCGGCGC
     ACACGCCATA AAGTGTGGCG TATGCAGTTT CGTTGGTATC ATGCGCGGGA CATCGCCGCG

            fnu4HI                  fnu4HI
           thaI                   hinPI
           fnuDII/mvnI            hhaI/cfoI                                  hinPI
           bctUI                 thaI                                        hhaI/cfoI
          hinPI aciI             fnuDII/mvnI                            rmaI
          hhaI/cfoI              bstUI                            hinPI   haeII
      tru9I aciI                 bsh1236I      aciI               hhaI/cfoI    bsrBI
      mseI bsh1236I           maeIII bbvI  maeIII               haeII maeI    aciI
2231 ATTAAGCGCG GCGGGTGTGG TGGTTACGCG CAGCGTGACC GCTACACTTG CCAGCGCCCT AGCGCCCGCT
     TAATTCGCGC CGCCCACACC ACCAATGCGC GTCGCACTGG CGATGTGAAC GGTCGCGGGA TCGCGGGCGA

                                                                                nlaIV
                                                  mspI                         hgiJII
                                                  hpaII                        bsp1286
                                                 naeI                          bmyI
                mboII                   maeII cfr10I              aluI         banII
2301 CCTTTCGCTT TCTTCCCTTC CTTTCTCGCC ACGTTCGCCG GCTTTCCCCG TCAAGCTCTA AATCGGGGGC
     GGAAAGCGAA AGAAGGGAAG GAAAGAGCGG TGCAAGCGGC CGAAAGGGGC AGTTCGAGAT TTAGCCCCCG
```

FIG. 16P

```
                                                
```

```
                                                    hgiAI/aspHI
       thaI                                         bsp1286
       fnuDII/mvnI              maeII               bsiHKAI
          apoI                 psp1406I             bmyI   ddeI
       bstUI    tru9I        tru9I                  apaLI/snoI  rsaI
       bsh1236I mseI   sspI mseI                    alw44I/snoI csp6I
2631 AACGCGAATT TTAACAAAAT ATTAACGTTT ACAATTTTAT GGTGCACTCT CAGTACAATC
     TTGCGCTTAA AATTGTTTTA TAATTGCAAA TGTTAAAATA CCACGTGAGA GTCATGTTAG

                                                        bsrI           hinPI
                                                     maeIII         fnu4HI
              fnu4HI     tru9I                      maeII        nlaIII hhaI/cfoI
           sfaNI aciI    mseI        aciI       bsaAI tth111I/aspI bbvI
2691 TGCTCTGATG  CCGCATAGTT AAGCCAACTC CGCTATCGCT ACGTGACTGG GTCATGGCTG CGCCCCGACA
     ACGAGACTAC  GGCGTATCAA TTCGGTTGAG GCGATAGCGA TGCACTGACC CAGTACCGAC GCGGGGCTGT

                         hinPI                          sfaNI
                         hhaI/cfoI                   mspI
                        thaI                         hpaII
                        fnuDII/mvnI                 scrFI
                        bstUI                       nciI
                 nspBII bsh1236I                    dsaV fokI                 maeIII
      aciI       aciI hgaI       drdI               cauII    aciI          aluI
2761 CCCGCCAACA CCCGCTGACG CGCCCTGACG GGCTTGTCTG CTCCCGGCAT CCGCTTACAG ACAAGCTGTG
     GGGCGGTTGT GGGCGACTGC GCGGGACTGC CCGAACAGAC GAGGGCCGTA GGCGAATGTC TGTTCGACAC
```

FIG. 16R

```
                                                
```

```
                                                                                  bsmAI
                                                                              rcaI
                                                                        bsrBI nlaIII
                                                                        aciI bspHI
      nlaIV
3001 CGGAACCCCT ATTTGTTTAT TTTTCTAAAT ACATTCAAAT ATGTATCCGC TCATGAGACA ATAACCCTGA
     GCCTTGGGGA TAAACAAATA AAAAGATTTA TGTAAGTTTA TACATAGGCG AGTACTCTGT TATTGGGACT

                                mboII
                   sspI         earI/ksp632I
3071 TAAATGCTTC AATAATATTG AAAAAGGAAG AGTATGAGTA TTCAACATTT CCGTGTCGCC CTTATTCCCT
     ATTTACGAAG TTATTATAAC TTTTTCCTTC TCATACTCAT AAGTTGTAAA GGCACAGCGG GAATAAGGGA

          fnu4HI
          aciI                          hphI            hphI          sfaNI
3141 TTTTTGCGGC ATTTTGCCTT CCTGTTTTTG CTCACCCAGA AACGCTGGTG AAAGTAAAAG
     AAAAACGCCG TAAAACGGAA GGACAAAAAC GAGTGGGTCT TTGCGACCAC TTTCATTTTC

                        hgiAI/aspHI
                        bsp1286                          sau3AI
             sau3AI     bsiHKAI                          mboI/ndeII[dam-]
             mboI/ndeII[dam-]                            dpnI[dam+]
             dpnI[dam+] bmyI                             dpnII[dam-]
             dpnII[dam-]                                bstYI/xhoII
          mboII[dam-]   apaLI/snoI          taqI        alwI[dam-]  aciI
        eco57I          alw44I/snoI   maeIII      bsrI          nspBII
3201 ATGCTGAAGA TCAGTTGGGT GCACGAGTGG GTTACATCGA ACTGGATCTC AACAGCGGTA
     TACGACTTCT AGTCAACCCA CGTGCTCACC CAATGTAGCT TGACCTAGAG TTGTCGCCAT
```

FIG. 16T

```
      sau3AI
      mboI/ndeII[dam-]                maeII
      dpnI[dam+]                     psp1406I          hgiAI/aspHI
      dpnII[dam-]                   xmnI               bsp1286  tru9I
      alwI[dam-]                    asp700             bsiHKAI  mseI
     bstYI/xhoII                mboII                  bmyI    ahaIII/draI
3261 AGATCCTTGA GAGTTTTCGC CCCGAAGAAC GTTTTCCAAT GATGAGCACT TTTAAAGTTC
     TCTAGGAACT CTCAAAAGCG GGGCTTCTTG CAAAAGGTTA CTACTCGTGA AAATTTCAAG

                                          scrFI
                 aciI                     nciI
                thaI                      mspI
                fnuDII/mvnI               hpaII
                bstUI                     dsaV
                bsh1236I             hinlI/acyI
              hinPI                  hgaI cauII                    aciI
              hhaI/cfoI              ahaII/bsaHI     bcgI  mcrI  fnu4HI
3321 TGCTATGTGG CGCGGTATTA TCCCGTGATG ACGCCGGGCA AGAGCAACTC GGTCGCCGCA
     ACGATACACC GCGCCATAAT AGGGCACTAC TGCGGCCCGT TCTCGTTGAG CCAGCGGCGT

                                    rsaI
                                    csp6I   bsrI
              ddeI                 scaI  hphI  maeIII      sfaNI     fokI
3381 TACACTATTC TCAGAATGAC TTGGTTGAGT ACTCACCAGT CACAGAAAAG CATCTTACGG
     ATGTGATAAG AGTCTTACTG AACCAACTCA TGAGTGGTCA GTGTCTTTTC GTAGAATGCC
```

FIG. 16U

```
                                                                                  haeIII/palI
                                                                                 eaeI
                                                                                 cfrI
                                        fnu4HI                                fnu4HI
          nlaIII                        bbvI          nlaIII                  aciI
3441 ATGGCATGAC AGTAAGAGAA TTATGCAGTG CTGCCATAAC CATGAGTGAT AACACTGCGG CCAACTTACT
     TACCGTACTG TCATTCTCTT AATACGTCAC GACGGTATTG GTACTCACTA TTGTGACGCC GGTTGAATGA

                        sau96I
                        avaII
                sau3AI  asuI                                                 nlaIII
                mboI/ndeII[dam-]                                          sau3AI maeIII
                dpnI[dam+]                                                mboI/ndeII[dam-]
                dpnII[dam-]                                               dpnI[dam+]
               pvuI/bspCI                                                 dpnII[dam-]
               mcrI   mnlI          aluI   aciI                 nlaIII alwI[dam-]
3511 TCTGACAACG ATCGGAGGAC CGAAGGAGCT AACCGCTTTT TTGCACAACA TGGGGGATCA TGTAACTCGC
     AGACTGTTGC TAGCCTCCTG GCTTCCTCGA TTGGCGAAAA AACGTGTTGT ACCCCCTAGT ACATTGAGCG

                       mspI
         sau3AI   nlaIV
         mboI/ndeII[dam-] aluI
         dpnI[dam+]   hpaII                                                      fnu4HI
         dpnII[dam-] bsaWI                                     maeIII     sfaNI   bbvI
3581 CTTGATCGTT GGGAACCGGA GCTGAATGAA GCCATACCAA ACGACGAGCG TGACACCACG ATGCCAGCAG
     GAACTAGCAA CCCTTGGCCT CGACTTACTT CGGTATGGTT TGCTGCTCGC ACTGTGGTGC TACGGTCGTC
```

FIG. 16V

```
                                                
```

FIG. 16X

```
                                                                  pleI
                                                                  hinfI
                                                      eam1105I
                                                      mnlI
                                                      bsrI haeIII/palI
                                                      nlaIV
                                                      asuI
                                                      sau96I
3841 ACTGGGGCCA GATGGTAAGC CCTCCCGTAT CGTAGTTATC TACACGACGG GGAGTCAGGC
     TGACCCCGGT CTACCATTCG GGAGGGCATA GCATCAATAG ATGTGCTGCC CCTCAGTCCG

                                                ddeI
                                          nlaIV sau3AI
                                    mboI/ndeII[dam-]
                                    hgiCI   dpnI[dam+]
                                    tru9I   mseI
                                    banI mnlI dpnII[dam-]
3901 AACTATGGAT GAACGAAATA GACAGATCGC TGAGATAGGT GCCTCACTGA TTAAGCATTG
     TTGATACCTA CTTGCTTTAT CTGTCTAGCG ACTCTATCCA CGGAGTGACT AATTCGTAAC

                                                tru9I
                                                mseI      tru9I
                                                ahaIII/draI mseI
3961 GTAACTGTCA GACCAAGTTT ACTCATATAT ACTTTAGATT GATTTAAAAC TTCATTTTTA
     CATTGACAGT CTGGTTCAAA TGAGTATATA TGAAATCTAA CTAAATTTTG AAGTAAAAAT

                                          rmaI      sau3AI
                                    sau3AI hphI   mboI/ndeII[dam-]
                                    mboI/ndeII[dam-]
                                    dpnI[dam+]    dpnI[dam+]
                                    dpnII[dam-]   dpnII[dam-]
                              tru9I bstYI/xhoII   alwI[dam-]      nlaIII      maeII
                              mseI  alwI[dam-]    bstYI/xhoII     rcaI        tru9I
                           ahaIII/draI maeI  mboII[dam-]        bspHI       mseI
4021 ATTTAAAAGG ATCTAGGTGA AGATCCTTTT TGATAATCTC ATGACCAAAA TCCCTTAACG TGAGTTTTCG
     TAAATTTTCC TAGATCCACT TCTAGGAAAA ACTATTAGAG TACTGGTTTT AGGGAATTGC ACTCAAAAGC
```

FIG. 16Y

```
                                                    sau3AI
                                                    mboI/ndeII[dam-]
                                                    dpnI[dam+]   sau3AI
                                                    dpnII[dam-] mboI/ndeII[dam-]
                                                   bstYI/xhoII  dpnI[dam+]
                                    sau3AI  alwI[dam-]   dpnII[dam-]
                                    mboI/ndeII[dam-]     alwI[dam-]
            hgaI                    dpnI[dam+] mboII[dam-]
         ddeI                       dpnII[dam-]         bstYI/xhoII
4091 TTCCACTGAG CGTCAGACCC CGTAGAAAAG ATCAAAGGAT CTTCTTGAGA TCCTTTTTTT
     AAGGTGACTC GCAGTCTGGG GCATCTTTTC TAGTTTCCTA GAAGAACTCT AGGAAAAAAA

        thaI
        fnuDII/mvnI
        bstUI
        bsh1236I
      hinPI       fnu4HI                                  aciI
      hhaI/cfoI   bbvI                          aciI    nspBII
4151 CTGCGCGTAA TCTGCTGCTT GCAAACAAAA AAACCACCGC TACCAGCGGT GGTTTGTTTG
     GACGCGCATT AGACGACGAA CGTTTGTTTT TTTGGTGGCG ATGGTCGCCA CCAAACAAAC

        sau3AI
        mboI/ndeII[dam-]
        dpnI[dam+]
        dpnII[dam-]
      alwI[dam-]
     mspI                                      bsrI              hinPI
     hpaII      aluI                         maeIII   eco57I     hhaI/cfoI
4211 CCGGATCAAG AGCTACCAAC TCTTTTTCCG AAGGTAACTG GCTTCAGCAG AGCGCAGATA CCAAATACTG
     GGCCTAGTTC TCGATGGTTG AGAAAAAGGC TTCCATTGAC CGAAGTCGTC TCGCGTCTAT GGTTTATGAC
```


SUBSTITUTE SHEET (RULE 26)

```
          rmaI                 haeIII/palI
          maeI      bslI      haeI                    scfI      aciI         mnlI
4281 TCCTTCTAGT GTAGCCGTAG TTAGGCCACC ACTTCAAGAA CTCTGTAGCA CCGCCTACAT ACCTCGCTCT
     AGGAAGATCA CATCGGCATC AATCCGGTGG TGAAGTTCTT GAGACATCGT GGCGGATGTA TGGAGCGAGA

                                                                scrFI
                                                                nciI
                                fnu4HI                          mspI
                      alwNI     bbvI                            hpaII
                     bsrI  fnu4HI                               dsaV    pleI
               maeIII     bbvI    bsrI                          cauII   hinfI
4351 GCTAATCCTG TTACCAGTGG CTGCTGCCAG TGGCGATAAG TCGTGTCTTA CCGGGTTGGA CTCAAGACGA
     CGATTAGGAC AATGGTCACC GACGACGGTC ACCGCTATTC AGCACAGAAT GGCCCAACCT GAGTTCTGCT

                                                        hgiAI/aspHI
                         nspBII                         bsp1286
           mspI         fnu4HI                          bsiHKAI
           hpaII        bbvI mcrI                       bmyI
          bsaWI       hinPI aciI                        apaLI/snoI
       maeIII         hhaI/cfoI                         alw44I/snoI   aluI
4421 TAGTTACCGG ATAAGGCGCA GCGGTCGGGC TGAACGGGGG GTTCGTGCAC ACAGCCCAGC TTGGAGCGAA
     ATCAATGGCC TATTCCGCGT CGCCAGCCCG ACTTGCCCCC CAAGCACGTG TGTCGGGTCG AACCTCGCTT

                                                         hinPI
                                                         hhaI/cfoI
                      ddeI      scfI                    haeII
4491 CGACCTACAC CGAACTGAGA TACCTACAGC GTGAGCATTG AGAAAGCGCC ACGCTTCCCG AAGGGAGAAA
     GCTGGATGTG GCTTGACTCT ATGGATGTCG CACTCGTAAC TCTTTCGCGG TGCGAAGGGC TTCCCTCTTT
```

FIG. 16Z-1

```
                                                                                scrFI
                                                                                mvaI
                                                                                ecoRII   mvaI
                    mspI                                                        dsaV     ecoRII
                    hpaII                                                       bstNI
                    bslI    fnu4HI                        hinPI mnlI            bsaJI
      aciI         bsaWI    aciI                          hhaI/cfoI   aluI apyI[dcm+]
4561 GGCGGACAGG TATCCGGTAA GCGGCAGGGT CGGAACAGGA GAGCGCACGA GGGAGCTTCC AGGGGGAAAC
     CCGCCTGTCC ATAGGCCATT CGCCGTCCCA GCCTTGTCCT CTCGCGTGCT CCCTCGAAGG TCCCCCTTTG

      scrFI


      dsaV
      bstNI                                                        taqI
      apyI[dcm+]                                 mnlI  drdI   hgaI            sfaNI
4631 GCCTGGTATC TTTATAGTCC TGTCGGGTTT CGCCACCTCT GACTTGAGCG TCGATTTTTG TGATGCTCGT
     CGGACCATAG AAATATCAGG ACAGCCCAAA GCGGTGGAGA CTGAACTCGC AGCTAAAAAC ACTACGAGCA

                                                  haeIII/palI
                                                fnu4HI
                                                aciI
                                               thaI bslI
                                               fnuDII/mvnI
                nlaIV                          bstUI
              aciI                             bsh1236I
4701 CAGGGGGGCG GAGCCTATGG AAAAACGCCA GCAACGCGGC
     GTCCCCCCGC CTCGGATACC TTTTTGCGGT CGTTGCGCCG
```

FIG. 16Z-2

```
                    haeIII/palI
                 scrFI
                 mvaI bslI
                 ecoRII
                 dsaV                    nlaIII
                 bstNI                        nspI
                 apyI[dcm+]     haeIII/palI   nspHI                          tfiI
            nlaIV  haeI         haeI          aflIII                         hinfI
4741 CTTTTTACGG TTCCTGGCCT TTTGCTGGCC TTTTGCTCAC ATGTTCTTTC CTGCGTTATC CCCTGATTCT
     GAAAAATGCC AAGGACCGGA AAACGACCGG AAAACGAGTG TACAAGAAAG GACGCAATAG GGGACTAAGA

                                                        fnu4HI
                                                        bbvI
                                                 bsrBI   aciI
                      aciI          aluI       aciI   fnu4HI       mcrI
4811 GTGGATAACC GTATTACCGC CTTTGAGTGA GCTGATACCG CTCGCCGCAG CCGAACGACC
     CACCTATTGG CATAATGGCG GAAACTCACT CGACTATGGC GAGCGGCGTC GGCTTGCTGG

                                                hinPI
        fnu4HI                                 haeII
        bbvI pleI                           sapI hhaI/cfoI
      hinPI  hinfI                          mboII                      mnlI
      hhaI/cfoI              mnlI  aciI  earI/ksp632I             aciI
4871 GAGCGCAGCG AGTCAGTGAG CGAGGAAGCG  GAAGAGCGCC CAATACGCAA ACCGCCTCTC
     CTCGCGTCGC TCAGTCACTC GCTCCTTCGC  CTTCTCGCGG GTTATGCGTT TGGCGGAGAG
```

FIG. 16Z-3

```
          thaI
          fnuDII/mvnI
          bstUI
          bsh1236I
         hinPI
         hhaI/cfoI
        thaI
        fnuDII/mvnI
        bstUI                tru9I  aluI
        bsh1236I haeIII/palI      pvuII
       bslI    eaeI  tfiI aseI/asnI/vspI
       aciI    cfrI  hinfI  mseI  nspBII                    bsrI     aciI
4931 CCCGCGCGTT GGCCGATTCA TTAATCCAGC TGGCACGACA GGTTTCCCGA CTGGAAAGCG
     GGGCGCGCAA CCGGCTAAGT AATTAGGTCG ACCGTGCTGT CCAAAGGGCT GACCTTTCGC

                                                            scrFI
                                                            mvaI
                                                            ecoRII
                                                            dsaV
                         tru9I                       nlaIV  bstNI
              hinPI      mseI      maeIII            hgiCI  apyI[dcm+]
              hhaI/cfoI aseI/asnI/vspI  mnlI         banI bsaJI
4991 GGCAGTGAGC GCAACGCAAT TAATGTGAGT TACCTCACTC ATTAGGCACC CCAGGCTTTA CACTTTATGC
     CCGTCACTCG CGTTGCGTTA ATTACACTCA ATGGAGTGAG TAATCCGTGG GGTCCGAAAT GTGAAATACG
```

FIG. 16Z-4

```
        mspI                                aciI
        hpaII                            bsrBI                                aluI      nlaIII
5061 TTCCGGCTCG TATGTTGTGT GGAATTGTGA GCGGATAACA ATTTCACACA GGAAACAGCT ATGACCATGA
     AAGGCCGAGC ATACAACACA CCTTAACACT CGCCTATTGT TAAAGTGTGT CCTTTGTCGA TACTGGTACT

            tru9I
            mseI
           aseI/asnI/vspI
         xmnI
         asp700
5131 TTACGAATTA A
     AATGCTTAAT T

>length: 5141
```

FIG. 16Z-5

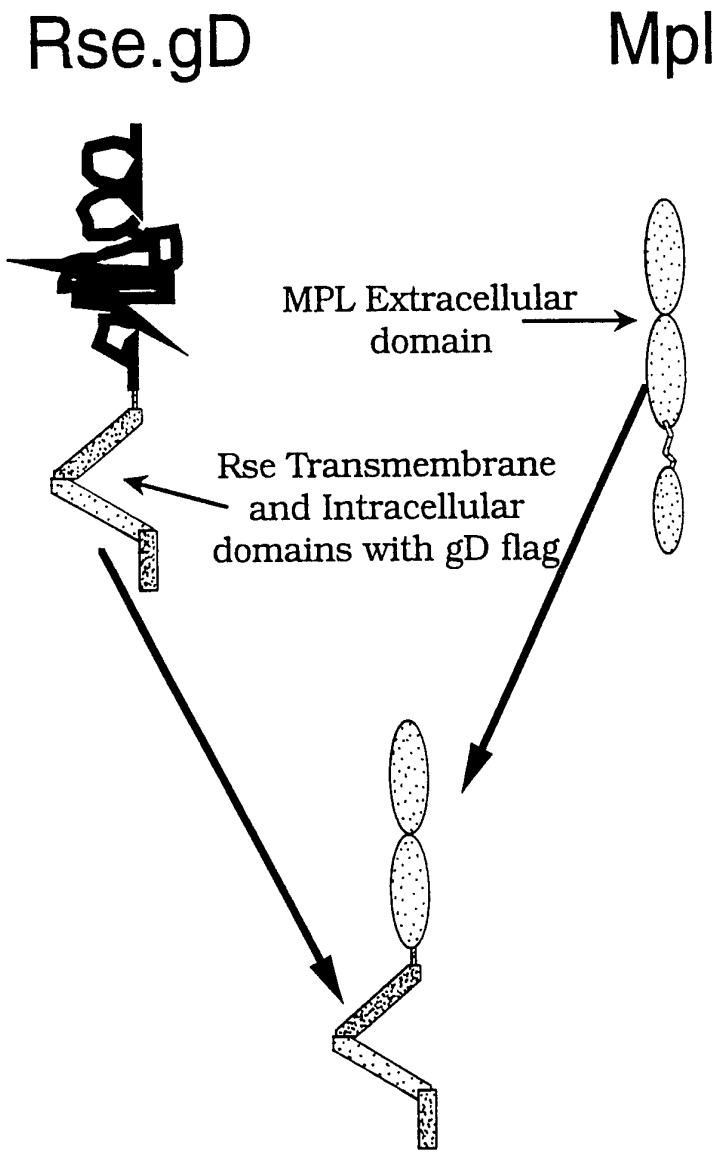

Rse.gD
Mpl
MPL Extracellular domain
Rse Transmembrane and Intracellular domains with gD flag

FIG. 17

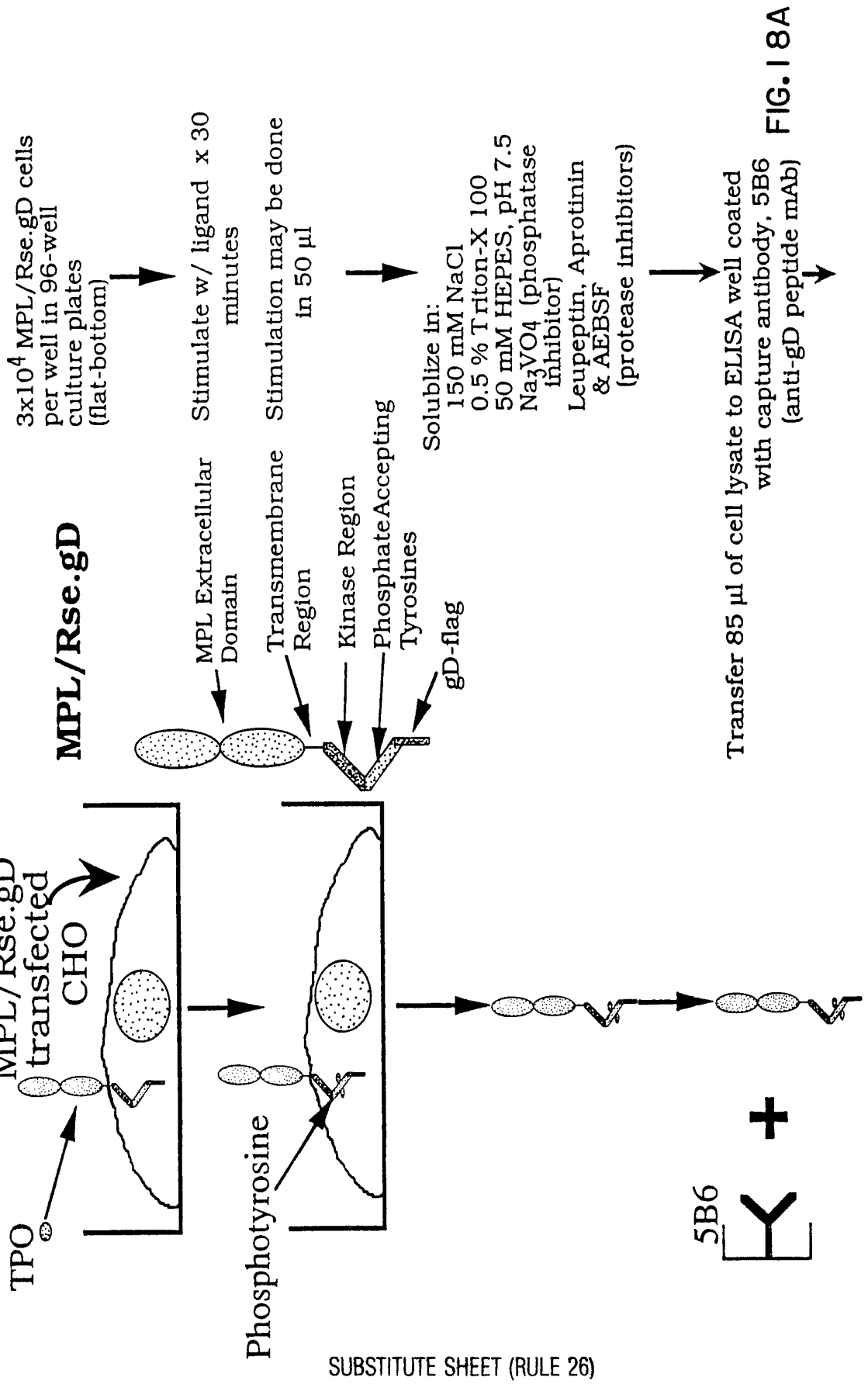

3x10^4 MPL/Rse.gD cells per well in 96-well culture plates (flat-bottom)
Stimulate w/ ligand x 30 minutes
Stimulation may be done in 50 µl
Solublize in:
150 mM NaCl
0.5 % Triton-X 100
50 mM HEPES, pH 7.5
Na3VO4 (phosphatase inhibitor)
Leupeptin, Aprotinin & AEBSF
(protease inhibitors)
Transfer 85 µl of cell lysate to ELISA well coated with capture antibody, 5B6 (anti-gD peptide mAb)
MPL/Rse.gD
MPL Extracellular Domain
Transmembrane Region
Kinase Region
PhosphateAccepting Tyrosines
gD-flag
MPL/Rse.gD transfected CHO
TPO
Phosphotyrosine
5B6

FIG. 18A

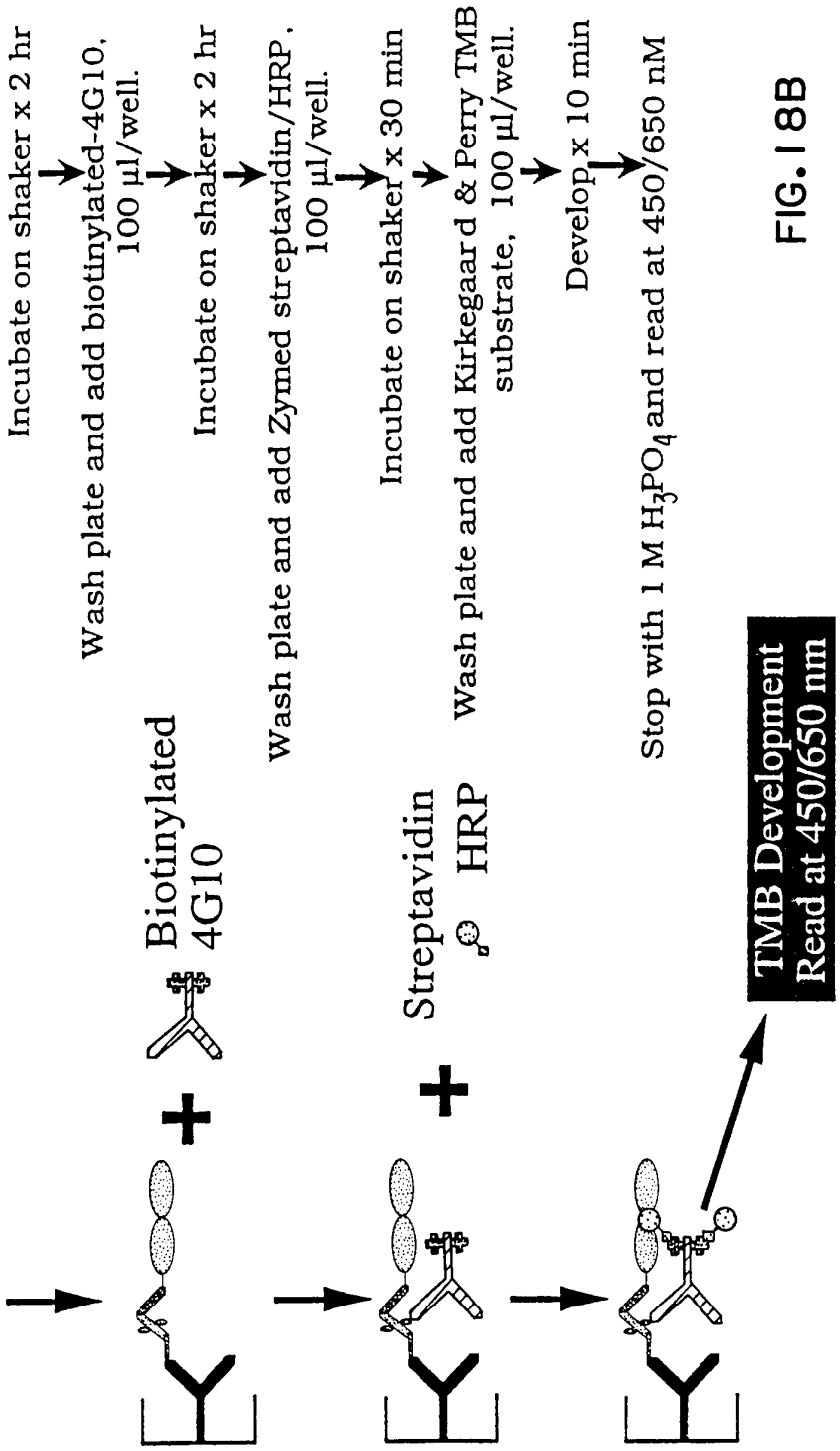

Incubate on shaker x 2 hr
Wash plate and add biotinylated-4G10, 100 µl/well.
Incubate on shaker x 2 hr
Wash plate and add Zymed streptavidin/HRP, 100 µl/well.
Incubate on shaker x 30 min
Wash plate and add Kirkegaard & Perry TMB substrate, 100 µl/well.
Develop x 10 min
Stop with 1 M H_3PO_4 and read at 450/650 nM
Biotinylated 4G10
Streptavidin HRP
TMB Development Read at 450/650 nm

FIG. 18B

INTERNATIONAL SEARCH REPORT

Internatic Application No
PCT/US 94/13329

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 G01N33/573 G01N33/566 G01N33/543 G01N33/532 G01N33/577
C07K14/705

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 G01N C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X,P	JOURNAL OF BIOLOGICAL CHEMISTRY, vol.269, no.14, 1 April 1994, WASHINGTON DC USA pages 10720 - 10728 M.R. MARK ET AL. 'RSE, a novel receptor-type tyrosine kinase with homology to Axl/Ufo, is expressed at high levels in the brain.' cited in the application see the whole document	29-32
T	see the whole document	1-43

-/--

[X] Further documents are listed in the continuation of box C.

[X] Patent family members are listed in annex.

° Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
3 April 1995	27.04.95

Name and mailing address of the ISA	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax (+31-70) 340-3016	Van Bohemen, C

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	ARCHIVES OF BIOCHEMISTRY AND BIOPHYSICS, vol.285, no.2, 1 March 1991, NEW YORK NY USA pages 197 - 204 V.P. KNUTSON ET AL. 'Comparison of insulin receptor tyrosine phosphorylation under in vitro and in situ conditions: assessment of specific protein tyrosine phosphorylation without the use of 32P-phosphate labeled substrates.' cited in the application see figure 1 ---	1-28, 33-43
Y	DIABETES, vol.42, 1 June 1993, WASHINGTON DC USA pages 883 - 890 H.H. KLEIN ET AL. W 'A microtiter well assay system to measure insulin activation of insulin receptor kinase in intact human mononuclear cells.' cited in the application see page 883, abstract see page 884, column 2, line 9 - line 44 ---	1-28, 33-43
Y	EP,A,0 244 221 (GENENTECH INCORPORATED) 4 November 1987 cited in the application see claim 1 -----	1-28, 33-43

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INTERNATIONAL SEARCH REPORT

Internatic Application No
PCT/US 94/13329

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
EP-A-0244221	04-11-87	US-A-	4859609	22-08-89
		CA-A-	1328419	12-04-94
		JP-A-	62272990	27-11-87
		US-A-	5030576	09-07-91