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FR-A1- 2 509 990
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DAE SIK JANG ET AL: "Constituents of Asparagus officinalis Evaluated for Inhibitory Activity against Cyclooxygenase-2", JOURNAL OF AGRICULTURAL AND FOOD CHEMISTRY, AMERICAN CHEMICAL SOCIETY, US, vol. 52, 2004, pages 2218-2222, XP008156190, ISSN: 0021-8561, DOI: 10.1021/JF0305229
XIONG Y; DENG K Z; GUO Y Q; GAO W Y; ZHANG T J: "New Chemical Constituents from the Rhizomes of Sparganium stoloniferum", ARCHIVES OF PHARMACAL RESEARCH, vol. 32, no. 5, May 2009 (2009-05), pages 717-720, XP002739585, DOI: 10.1007/s12272-009-1510-9
KOSHINO H ET AL: "Three phenolic acid derivatives from stromata of Epichloe typhina on Phleum pratense", PHYTOCHEMISTRY, PERGAMON PRESS, GB, vol. 27, no. 5, 1988, pages 1333-1338, XP026772840, ISSN: 0031-9422, DOI: 10.1016/0031-9422(88)80188-2 [retrieved on 1988-01-01]
COOPER R ET AL: "New phenolic diglycerides from Aegilops ovata", PHYTOCHEMISTRY, PERGAMON PRESS, GB, vol. 17, no. 9, 1978, pages 1673-1675, XP026617308, ISSN: 0031-9422, DOI: 10.1016/S0031-9422(00)94673-9 [retrieved on 1978-01-01]
SHIMOMURA H ET AL: "Phenolic glycerides from Lilium auratum", PHYTOCHEMISTRY, PERGAMON PRESS, GB, vol. 26, no. 3, 1987, pages 844-845, XP026659871, ISSN: 0031-9422, DOI: 10.1016/S0031-9422(00)84801-3 [retrieved on 1987-01-01]
ORI K ET AL: "Jatropha derivatives and steroidal saponins from the bulbs of Lilium hansonii",

Fortsættes ...

PHYTOCHEMISTRY, PERGAMON PRESS, GB, vol. 31, no. 8, August 1992 (1992-08), pages 2767-2775, XP026633207, ISSN: 0031-9422, DOI: 10.1016/0031-9422(92)83627-B [retrieved on 1992-08-01]

JANG, D. S. ET AL.: 'Constituents of Asparagus officinalis Evaluated for Inhibitory Activity against Cyclooxygenase-2' J. AGRIC. FOOD CHEM. vol. 52, 2004, pages 2218 - 2222, XP008156190

LI, F. ET AL.: 'Study on the Constituents of Mexican Propolis and Their Cytotoxic Activity against PANC-1 Human Pancreatic Cancer Cells' J. NAT. PROD. vol. 73, 22 March 2010, pages 623 - 627, XP008156191

BANSKOTA, A. H. ET AL.: 'Antiproliferative activity of the Netherlands propolis and its active principles in cancer cell lines' J. ETHNOPHARMACOLOGY vol. 80, 2002, pages 67 - 73, XP008156193

DESCRIPTION

FIELD OF INVENTION

[0001] The present invention relates to therapeutic agents for use in treating cancer. More particularly, the present invention relates to propane diyl dicinnamate derivatives for treating cancer and methods for their preparation.

BACKGROUND OF INVENTION

[0002] According to the World health Organization, malignant tumours were responsible for 12 per cent of the nearly 56 million deaths worldwide from all causes in the year 2000. In particular, 5.3 million men and 4.7 million women developed a malignant tumour and altogether 6.2 million died from the disease. The report also reveals that cancer has emerged as a major public health problem in developing countries, matching its effect in industrialized nations.

[0003] Although substantial progress has been made in the past two decades, there remain many cancers for which conventional therapy are either partially or totally ineffective. The main common problem with these therapies is unacceptable toxicity.

[0004] Thus, novel compounds, agents or methods are needed either to prevent the development of cancer, or, in the case where neoplasia has already developed, to render the host organism cancer-free or to reduce its neoplastic burden to a level compatible with life or at least to facilitate the use of concomitant therapies.

[0005] In its essence, neoplasia, including cancer, can be regarded as the inappropriate accumulation of cells, in violation of the delicate balance between cell renewal and cell death. For neoplasia to develop, either cell renewal must be increased or cell death decreased or both. A corollary to this relationship is that an agent that affects these processes favorably for the host organism (and, consequently, unfavorably for the neoplasm), is a potential antineoplastic drug.

[0006] Several antineoplastic agents have been isolated and identified from natural sources. For example, Curcumin (diferuloylmethane), a polyphenol derived from the plant *Curcuma longa*, commonly called turmeric has been extensively studied over the last 50 years and these these series of studies have shown that Curcumin suppresses the proliferation of a wide variety of tumor cells, including breast carcinoma, colon carcinoma, acute myelogenous leukemia, basal cell carcinoma, melanoma and prostate carcinoma.(1-7) Despite the remarkable pharmacological safety profile of this compound, its effectiveness as a plausible anticancer agent has been limited by poor systemic absorption and extensive metabolism with administered doses.

[0007] A specific component of the honeybee hive product propolis, identified as caffeic acid phenyl ester (CAPE), has been shown to selectively inhibit the growth of viral-transformed and oncogene-transformed rodent cells as well as human tumor cells, including glioblastoma multiforme (GBM-18), colon adenocarcinoma (HT-29), and melanoma (HO-1) cells. These studies also show that CAPE and several additional caffeic acid esters inhibit azoxymethane-induced colonie preneoplastic lesions and ornithine decarboxylase, tyrosine protein kinase, and lipoxygenase activities associated with colon carcinogenesis.

(8-13)

[0008] Although derived from different natural sources, CAPE and the Curcuminoids possess structural similarity that may likely explain their anticancer properties or at least partially explain their safety and selectivity profile. The significance of this structural similarity with regard to their anticancer properties is currently unknown.

[0009] An approach for developing new antineoplastic agents is to synthesize novel chemical compounds that are selective for cancer, stable in the biological milieu, maintain potency against cancer and exhibit low toxicity overall.

[0010] Banskota et al. (Journal of Ethnopharmacology (2002), 80(1), 67-73) isolated the cinnamic acid derivatives benzyl caffeate, phenethyl caffeate and cinnamyl caffeate, as well as the dicinnamate compounds (2E,2'E)-2-acetoxypropane-1,3-diyl bis(3-(4-hydroxyphenyl)acrylate) and (E)-2-acetoxy-3-(((E)-3-(4-hydroxy-3-methoxyphenyl)acryloyl)oxy)propyl 3-(4-hydroxyphenyl)acrylate from a MeOH extract of Netherlands *propolis*. Benzyl caffeate, phenethyl caffeate and cinnamyl caffeate displayed EC₅₀ values of 2.03 μM, 3.16 μM and 1.92 μM, respectively, against B16-BL6. The dicinnamate compounds (2E,2'E)-2-acetoxypropane-1,3-diyl bis(3-(4-hydroxyphenyl)acrylate) and (E)-2-acetoxy-3-(((E)-3-(4-hydroxy-3-methoxyphenyl)acryloyl)oxy)propyl 3-(4-hydroxyphenyl)acrylate displayed moderate EC₅₀ values of 81.9 μM and 66 μM, respectively, against the same cell line.

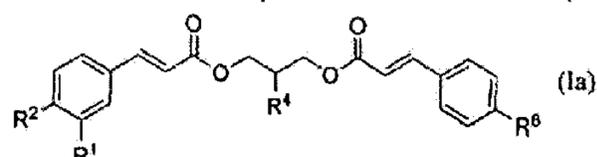
D1 = Dae Sik Jang et al. "Constituents of asparagus officinalis evaluated for inhibitory activity against cyclooxygenase-2", J. of agricultural and food chemistry, vol. 52, 2004, pages 2218-2222, discloses compounds 8 and 9, having a distinct pattern of substitution, wherein each of the terminal phenyl groups carries an OH-group in para position. The said compounds 8 and 9 are described in the context of their use in the medicinal field as cancer chemopreventive agents". However, their inhibitory activities against cyclooxygenase-2 have been described as low (30% or 0%) or ND.

D2 = XIONG Y; DENG K Z; GUO Y Q; ZHANG T J: "New chemical constituents from the rhizomes of sparganium stoloniferum" Archives of Pharmacal Research, vol. 32, no. 5, 2009, pages 717-720 describes the compound (2) = O-cis-Feruloyl-3-O-trans-p-coumaroylglycerol as having weak cytotoxic activity against the growth of LA795 tumor cell line.

SUMMARY OF INVENTION

[0011] The present invention relates to therapeutic agents for use in treating cancer. More particularly, the present invention relates to propane diyl dicinnamate derivatives for treating cancer and methods for their preparation.

[0012] In one aspect, the present invention provides a compound for use in the treatment of cancer, wherein the compound is of formula (Ia)



wherein:

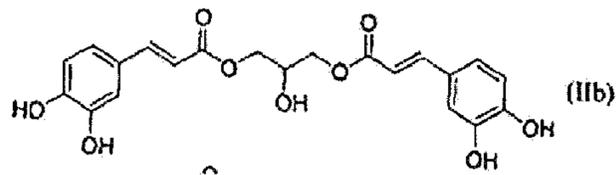
R¹ and R² are independently OH, alkoxy or alkylcarbonyloxy,

R⁴ is OH, and

R⁶ is H

or a pharmaceutically acceptable salt thereof.

[0013] In another aspect, the present invention provides a compound of formula (IIb) for use in the treatment of cancer.



BRIEF DESCRIPTION OF THE DRAWINGS

[0014] These and other features of the invention will become more apparent from the following description in which reference is made to the appended drawings wherein:

FIGS 1* and 9*-11* illustrate examples of synthetic schemes for producing diester compounds.

FIG. 2* illustrates examples of synthetic schemes for producing diester compounds.

FIG. 3 illustrates examples of methods of producing glycidyl cinnamate derivatives.

FIG. 4* illustrates examples of synthetic schemes for producing diester compounds having symmetrical ends.

FIGS. 5 and 6 illustrate examples of synthetic schemes of forming tri-substituted derivatives of intermediates for forming the compounds of the present invention.

FIGS. 7 and 8 illustrate examples of synthetic schemes of forming di-substituted derivatives of intermediates for forming the compounds of the present invention.

FIG. 12 shows micrographs of Human Colon Carcinoma HCT116 cells over time following treatment with no compound (control).

FIG. 13 shows micrographs of Human Colon Carcinoma HCT116 cells over time following treatment with 30 μ M Compound (IIb).

FIG. 14 shows micrographs of Human Colon Carcinoma HCT116 cells over time following treatment with 30 μ M Compound (Ie)

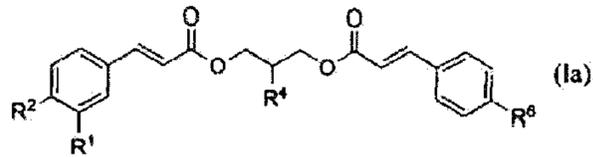
FIG. 15 shows micrographs of Human Colon Carcinoma HCT116 cells over time following treatment with 30 μ M Compound (Id)

* All Figures marked by an asterix do not form part of the invention, but they are left in the description for better understanding the invention.

DETAILED DESCRIPTION OF INVENTION

[0015] The present invention relates to therapeutic agents for use in treating cancer. More particularly, the present invention relates to propane diyl dicinnamate derivatives for treating cancer and methods for their preparation.

[0016] The present invention provides a compound of formula (Ia):



wherein:

R¹ and R² are independently OH, alkoxy or alkylcarbonyloxy,

R⁴ is OH and

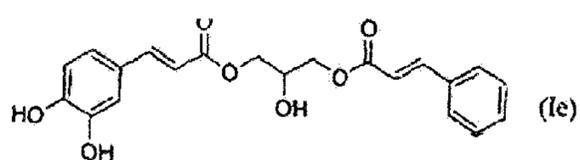
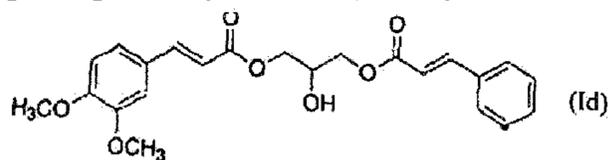
R⁶ is H,

or a pharmaceutically acceptable salt thereof.

[0017] In an example of the above-defined compound of formula (Ia) both R¹ and R² are OH, alkoxy or alkylcarbonyloxy.

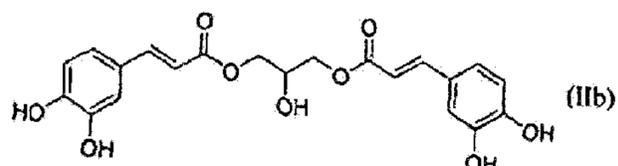
[0018] In other examples, the present invention relates to the compounds ²a of formula (Ia) described above, wherein R¹ and R² are both OH or both alkoxy.

[0019] More particularly, the present invention relates to a compound of the formula (Id), (Ie).



In another aspect, the present invention provides a pharmaceutical composition for use in the treatment of cancer comprising at least one compound as used herein before.

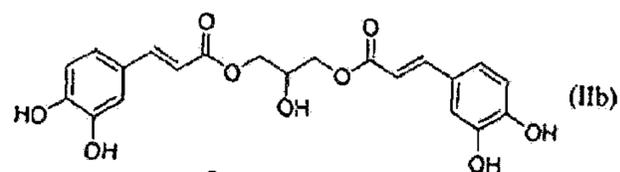
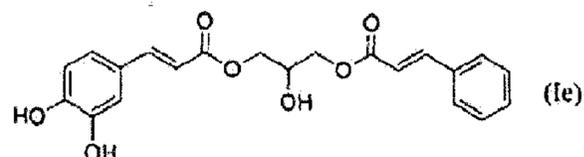
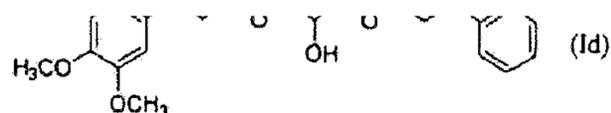
[0020] In still another aspect, the present invention provides a compound compound for use in the treatment of cancer, such as lung cancer, breast cancer, colon cancer, prostate cancer, ovarian cancer, skin cancer or leukemia wherein the compound is of formula (IIb):



or a pharmaceutically acceptable salt thereof.

[0021] More particularly, the present invention relates to the uses defined above, wherein the compound administered to the subject is of the formula (Id), (Ie), (IIb), or a combination thereof:





[0022] The present invention also relates to a pharmaceutical composition comprising the compound defined above, and a pharmaceutically acceptable diluent or carrier.

[0023] As used herein, the term "alkyl" refers to a straight- or branched-chain hydrocarbon radical of the general formula C_nH_{2n+1} , such as an alkyl group having 1 to 10, 1 to 9, 1 to 8, 1 to 7, 1 to 6, 1 to 5, 1 to 4, 1 to 3 or 1 to 2 carbon atoms. Examples of alkyl groups include, but are not limited to, methyl, ethyl, propyl, isopropyl, and t-butyl.

[0024] The term "pharmaceutically acceptable salt" refers to a salt that retains the biological effectiveness and properties of a corresponding free base or free acid, and which is not biologically or otherwise undesirable. The salt may be prepared from addition of an inorganic base or an organic base to a free acid. Salts derived from an inorganic base include, but are not limited to, the sodium, potassium, lithium, ammonium, calcium, magnesium salts and the like. Salts derived from organic bases include, but are not limited to salts of primary, secondary, and tertiary amines, substituted amines including naturally occurring substituted amines, cyclic amines and basic ion exchange resins, such as isopropylamine, trimethylamine, diethylamine, triethylamine, tripropylamine, ethanolamine, lysine, arginine, N-ethylpiperidine, piperidine, or a polyamine resin.

[0025] The compounds of the present invention or their corresponding pharmaceutically acceptable salts can be used in the form of pharmaceutical compositions for enteral, parenteral or topical administration. They can be administered, for example, perorally, e.g. in the form of tablets, coated tablets, dragees, hard and soft gelatine capsules, solutions, emulsions or suspensions, rectally, e.g. in the form of suppositories, parenterally, e.g. in the form of injection solutions or suspensions or infusion solutions, or topically, e.g. in the form of ointments, creams or oils.

[0026] Suitable carrier materials are not only inorganic carrier materials, but also organic carrier materials. Thus, for example, lactose, corn starch or derivatives thereof, talc, stearic acid or its salts can be used as carrier materials for tablets, coated tablets, dragees and hard gelatine capsules. Suitable carrier materials for soft gelatine capsules are, for example, vegetable oils, waxes, fats and semi-solid and liquid polyols (depending on the nature of the active ingredient no carriers might, however, be required in the case of soft gelatine capsules). Suitable carrier materials for the production of solutions and syrups are, for example, water, polyols, sucrose, invert sugar and the like. Suitable carrier materials for injection solutions are, for example, water, alcohols, polyols, glycerol and vegetable oils. Suitable carrier materials for suppositories are, for example, natural or hardened oils, waxes, fats and semiliquid or liquid polyols. Suitable carrier materials for topical preparations are glycerides, semi-synthetic and synthetic glycerides, hydrogenated oils, liquid waxes, liquid paraffins, liquid fatty alcohols, sterols,

polyethylene glycols and cellulose derivatives.

[0027] The propane diyl dicinnamate derivatives represented by the general formulas (Ia), (Ib) defined above are useful for treatment, prophylaxis and prevention of cancers and other proliferative diseases including, but not limited to, tumors, inflammation and human immunodeficiency (HIV). The anticancer activity exhibited by the compounds of the present invention may be through cytotoxicity, antiproliferation, cell cycle kinase inhibition or through cell differentiation.

[0028] The present invention also relates to processes for the preparation of compounds defined by the general formulas (Ia), (Ib) or mixtures thereof, their stereoisomers, their polymorphs, their pharmaceutical acceptable salts, and their pharmaceutically acceptable solvates.

In the following text passages, paragraphs marked with an asterix do not belong to the invention, however, they are left in the description for better understanding the invention.

[0029] * The compounds of general formula (Ib) and (Ib') can be formed according to the synthetic pathways outlined in Figures 1 and 9-11. Referring specifically to Figure 1, 1,3 glycerol diester 5 (formula (II), $R^4=OH$) can be formed by allowing cinnamic acid derivative 1 or 2 to react with the corresponding glycidyl cinnamate derivative 3 or 4 in the presence of a phase transfer catalyst (PTC). Diester 5 can be further alkylated or acylated to produce derivatized diester 6.

[0030] * Compounds of formula (II), where R^4 is hydrogen, can be formed by crosslinking of cinnamic acid derivatives 1 and 2 with crosslinking agent 7 to produce the diester compound 8. Subsequent partial or complete acylation of diester 8 with an acylating agent, such as $RC(O)L$, affords acylated diester derivative 17.

[0031] * As illustrated in Figure 2, compounds of formula (II') can be produced by esterification of cinnamic acid derivative 9 or 10 with the corresponding glycidyl cinnamate derivative 3 or 4 in the presence of a phase transfer catalyst (PTC) to afford 1,2 glycerol diester derivative 13, which can be further alkylated or acylated to produce diester 14.

[0032] * Compounds of formula (II'), where R^4 is hydrogen, can be formed by crosslinking of cinnamic acid derivatives 1 and 2 with crosslinking agent 15 to produce the diester compound 16. Subsequent partial or complete acylation of diester 8 with an acylating agent, such as $RC(O)L$, affords acylated diester derivative 18.

[0033] * Examples of phase transfer catalysts that can be used in forming the diester compounds 5 and 13 include tetrabutylammonium bromide or tetrabutylammonium chloride. The illustrated reactions for forming diesters 5, 6, 8, 13, 14 and 16-18 are conducted in a suitable inert solvent, such as dioxane, and carried out at a reaction temperature of 60-120°C or 100-105°C.

[0034] * Examples of the leaving groups that can be used in compounds 7 and 15 include, without limitation, a halogen, such as Cl, Br or iodo, methanesulfonyl, phenylsulfonyloxy, p-tosyloxy, methanesulfonyloxy, acetoxy or benzoyloxy.

[0035] * Intermediate glycidyl cinnamate compounds 3 and 4 can be produced by a substitution reaction involving the corresponding cinnamate derivatives 1 or 2 and an excess molar amount of epichlorohydrin or other suitable epihalohydrin (Figure 3). The reaction is conducted in a suitable inert solvent, such as dioxane, and optionally in the presence of a catalyst at a reaction temperature of about 50-100°C or 70-

95°C, for approximately 12 to 18 hours.

[0036] When the substituents R¹, R² and R³ are the same as the substituents R⁷, R⁶ and R⁵, respectively, compounds of the general formula (II) can be prepared in a reaction involving a cinnamyl halide derivative 9 and stoichiometric amounts of glycerol in the presence of a tertiary amine, such as pyridine, to produce the symmetrical 1,3 glycerol diester 5, which can be further alkylated or esterified to afford the protected compound 6 (Figure 4).

[0037] * To prepare compounds of general formula (II'), where the variable R⁴ is H, and the variables R¹, R² and R³ are equivalent to R⁷, R⁶ and R⁵, respectively, the cinnamyl halide derivative 9 can be reacted with methylglycol in the presence of a catalytic amount of pyridine to produce the diester compound 16.

[0038] * The cinnamyl halide derivative 9 or 10 can be prepared by reacting the corresponding derivative of cinnamic acid with a thionyl halide, such as thionyl chloride, in a suitable solvent, such as dioxane, at room temperature for approximately 15-20 minutes.

[0039] * The reactions for forming diester derivatives 5, 8, 13 and 16 are typically carried out in the presence of pyridine and allowed to react for approximately 20-30 minutes. Following the completion of the reactions, the reaction mixture is neutralized with sodium bicarbonate or other suitable neutralizing agent. The reaction mixture is then filtered and the final compound is isolated by drying the organic layer under reduced pressure followed by crystallization from ethyl acetate or a mixture of ethyl acetate and cyclohexane.

[0040] * Figures 5 and 6 illustrate examples of synthetic schemes for producing different tri-substituted derivatives of cinnamic acids 1 and 2. Each of these schemes initially involves the esterification of gallic acid 20 to produce methyl gallate 21, which is selectively benzylated at the para position to produce protected derivative 22. Alkylation of derivative 22 with two equivalents of an alkyl halide RX affords dialkylated ester 23, which is subsequently subjected to hydrogenolysis to produce phenolic derivative 24. Reduction of phenol 24 with lithium aluminum hydride (LAH) following by partial oxidation of the resulting alkyl alcohol with pyridinium chlorochromate (PCC) results in the formation of aldehyde 25, which is allowed to react with malonic acid and piperidine in pyridine to produce cinnamic acid derivative 26.

[0041] Alternatively, methyl gallate 21 can be directly reduced and the resulting methylene alcohol partially oxidized with PCC to afford the aldehyde 27, which is subsequently allowed to react with malonic acid and piperidine in pyridine to produce cinnamic acid derivative 26.

[0042] In another example, alkylation of derivative 22 with one equivalent of an alkyl halide RX affords monalkylated ester 29, which is subsequently subjected to hydrogenolysis to remove the benzyl protecting group. The resulting phenol is then reduced with LAH to produce an methylene alcohol that is partially oxidized with PCC to form aldehyde 30, which is allowed to react with malonic acid and piperidine in pyridine to produce cinnamic acid derivative 31.

[0043] * Figure 6 illustrates examples of synthetic schemes for producing mono, di- and trialkylated derivatives of trihydroxylated cinnamic acid (34, 37 and 40). The synthetic schemes involve the initial reaction of methyl gallate 21 with 1, 2 or 3 eq. of an alkyl halide RX in the presence of tetrabutylammonium iodide and K₂CO₃ to produce the mono-alkylated ester 32, the di-alkylated ester 35 and the tri-alkylated ester 38, respectively. These esters are then subsequently reduced with LAH to

produce respective alcohols that are partially oxidized with PCC to form aldehydes 33, 36 and 39, respectively, which are subsequently allowed to react with malonic acid and piperidine in pyridine to produce the alkylated cinnamic acid derivatives 34, 37 and 40, respectively.

[0044] * Figure 7 illustrates examples of synthetic schemes for producing mono and dialkylated derivatives of dihydroxylated cinnamic acid (44, 46 and 50). Caffeic acid 41 is first esterified in MeOH to produce caffeic acid methyl ester 42, which is then reacted with 1 or 2 eq. of an alkylating agent RX in the presence of TBAI and K_2CO_3 to produce the mono-alkylated intermediate 43 or the dialkylated intermediate 45. The esters 43 and 45 are then hydrolyzed under basic conditions to produce the corresponding acids 44 and 46, respectively.

[0045] To produce a derivative of caffeic acid having an alkoxy group in the 3-position of the benzene ring, the ester 42 is first protected with 1 eq. of a benzoyl halide to afford protected derivative 47 having a benzoyloxy group at the para-position of the phenyl ring, which is subsequently reacted with 1 eq. of an alkyl halide to produce mono-alkylated derivative 48. Deprotection of derivative 48 results in the production of ester 49 having a free hydroxyl group at the para-position of the phenyl ring and an alkoxy group at the 3-position of the phenyl ring. Finally, hydrolysis of the ester 49 under basic conditions produces carboxylic acid 50.

[0046] * Figure 8 illustrates examples of synthetic schemes for producing alkylated derivatives of 3,5-dihydroxycinnamic acid. In the illustrated synthetic schemes, 3,5-dihydroxycinnamic acid 51 is first esterified in MeOH to produce 3,5-dihydroxycinnamic acid methyl ester 52, which is then reacted with 1 or 2 eq. of an alkylating agent RX in the presence of TBAI and K_2CO_3 to produce the mono-alkylated intermediate 53 or the dialkylated intermediate 55. The esters 53 and 55 are then hydrolyzed under basic conditions to produce the corresponding acids 54 and 56, respectively.

[0047] Similarly, the monoalkylated derivative of m- or p-coumaric acid can be produced by esterifying coumaric acid in MeOH to produce coumaric acid methyl ester, which is then reacted with an alkylating agent to produce a mono-alkylated intermediate. This intermediate is subsequently hydrolyzed under basic conditions to produce the corresponding alkylated derivative of m- or p-coumaric acid.

[0048] * Figures 9-11 illustrate examples of synthetic schemes for forming different protected derivatives of the diesters 5 and 13. Referring to Figure 9, there is shown an example of a synthetic scheme for forming diester derivatives having acylated phenolic residues, which involves first protecting the secondary hydroxyl group of diester 5 with a tertbutyldimethylsilyl (TBS) group to form the TBS-ether 60 and then acylating the free phenolic hydroxyl groups with an acylating agent, such as an acyl halide, to form the acylated, protected derivative 61. The TBS group is then removed with tetrabutyl ammonium fluoride (TBAF) to produce the alcohol 62, which can then be further alkylated to afford the acylated, alkyl ether 63.

[0049] * In another example illustrated in Figure 10, the phenolic hydroxyl groups of TBS ether 60 are partially acylated to form the TBS ether 61a having one or more free phenolic hydroxyl groups. Following deprotection of the TBS ether 61a with TBAF, the phenolic hydroxyl groups of the resulting alcohol 62a are selectively protected with trityl-Cl to form the tritylated derivative 62b having a free secondary hydroxyl group. The alcohol 62b can then be further alkylated to afford the partially acylated alkyl ether 63a.

[0050] * In further examples illustrated in Figure 11, the free phenolic hydroxyl groups and the secondary

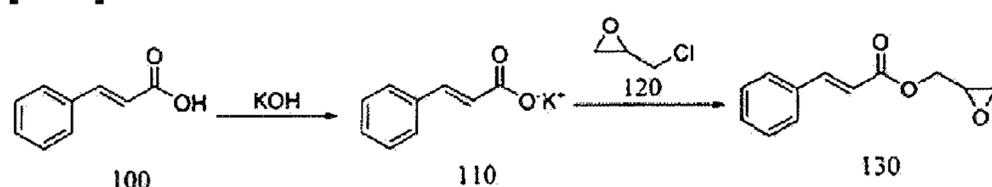
alcohol group of diester 5 can be acylated using an excess of an acylating agent, such as an acyl halide. Alternatively, the phenolic hydroxyl groups of diester 5 may be selectively protected with trityl-Cl to form the tritylated derivative 70 having a free secondary hydroxyl group. Subsequent alkylation or acylation of the alcohol 70 followed by deprotection of the phenolic hydroxyl groups results in alkylated or acylated derivative 72.

[0051] The following examples are given to illustrate the invention:

Examples

Example 1: Preparation of Glycidyl Cinnamate (130)

[0052]



[0053] To 5.66 g (0.10 mol) of potassium hydroxide dissolved in 50 mL of deionized water and heated to 50°-60°C was added 15 g (0.10 mol) of cinnamic acid (100) with stirring. The resulting reaction slurry was dried in a vacuum oven at 40-50°C to yield potassium cinnamate (110).

Alternative method for producing potassium cinnamate (110)

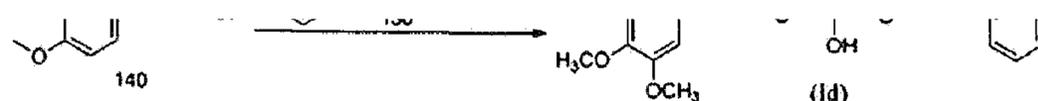
[0054] To a solution of cinnamic acid (100, 15 g, 0.10 mol) in THF (150 mL) at about 30°C was added 5.66 g (0.10 mol) of a powder of freshly ground potassium hydroxide pellets to produce a white precipitate of potassium cinnamate (110). The precipitate was filtered and dried in a vacuum oven at 40-50°C.

[0055] Potassium cinnamate (110; 5.3 g; 0.028 mol) and a catalytic amount (0.85 g; 2.7 mmol) of tetrabutylammonium bromide were added to 50 g (0.54 mol) of epichlorohydrin (120) in a reaction vessel equipped with a mechanical stirrer and a reflux assembly and heated to 95°-105°C to form a mixture that was allowed to react for 60 minutes. The mixture was then cooled to room temperature, diluted with 55 mL chloroform, and filtered to remove solid precipitates. The organic filtrate was washed sequentially with 50 mL of 5% NaHCO₃ and deionized water and the resulting organic layer was distilled under reduced pressure at 30°-40°C to yield glycidyl cinnamate (130).

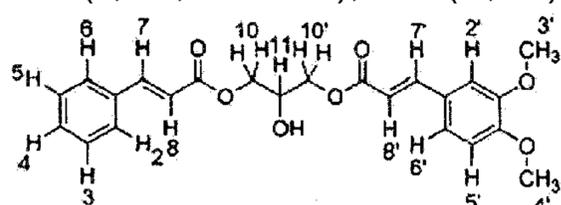
Example 2: Preparation of (E)-3-(cinnamoyloxy)-2-hydroxypropyl 3-(3, 4-dimethoxyphenyl) acrylate (1d)

[0056]





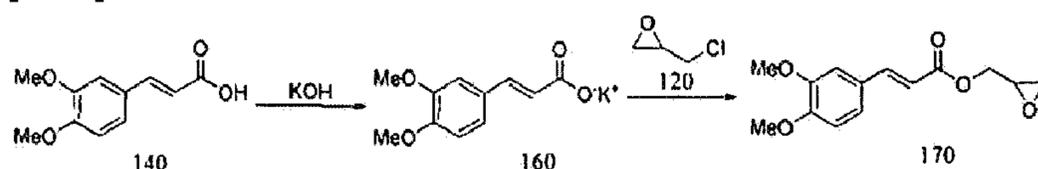
[0057] 3,4-dimethoxy cinnamic acid (140; 3.82 g; 18.37 mol) was dissolved in 60 mL of dioxane and heated under reflux (60°-70°C). A catalytic amount (22.5 mg; 0.7 mmol) of tetrabutylammonium bromide and 1.5 g (7.35 mmol) of glycidyl cinnamate (130) were consecutively added to the solution of 3,4-dimethoxy cinnamic acid (140). The resulting mixture was heated to 100°-105°C with continuous stirring for 15-16 h and then distilled under reduced pressure to yield compound (Id). ¹H-NMR (CD₃OD) δ 3.81 (s, 3H), 3.83 (s, 3H), 4.14 (m, 1H), 4.27 (d, 4H, J = 8 Hz), 6.42 (d, 1H, J = 16 Hz), 7.64 (d, 1H, J = 16 Hz), 7.15 (dd, 1H, J = 8 Hz, 2 Hz), 6.92 (d, 1H, J = 8 Hz), 7.18 (d, 1H, J = 2 Hz), 6.53 (d, 1H, J = 16 Hz), 7.70 (d, 1H, J = 16 Hz), 7.35 (m, 3H), 7.56 (m, 2H)



Proton	Chemical Shift
2,6	7.56
3,5	7.35
4	7.35
7	7.70
8	6.53
2'	7.18
5'	6.92
6'	7.15
7'	7.64
8'	6.42
10, 10'	4.27
11	4.14
OCH3	3.83
OCH3	3.81

Example 3: Preparation of (E)-oxiran-2-ylmethyl 3-(3,4-dimethoxyphenyl) (170)

[0058]



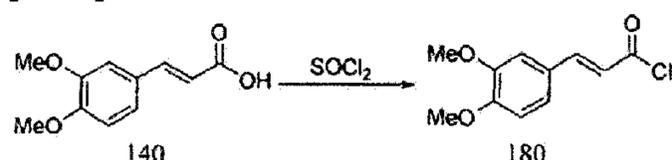
[0059] To 5.0 g (0.24 mol) of potassium hydroxide dissolved in 30 mL of deionized water and heated to 50°-60°C was added 1.34 g (0.024 mol) of 3,4-dimethoxy cinnamic acid (140) with stirring. The resulting

reaction slurry was dried in a vacuum oven at 40-50°C to yield potassium 3,4-dimethoxy cinnamate (160).

[0060] Potassium 3,4-dimethoxy cinnamate (160; 11.83 g; 0.048 mol) and 0.772 g (0.0024 mol) of tetrabutylammonium bromide were added to 55.32 g (0.60 mol) of epichlorohydrin (120) in a reaction vessel equipped with a mechanical stirrer and a reflux assembly to form a mixture that was allowed to react for 60 minutes and heated at 80°-90°C. The mixture was then cooled to room temperature, filtered to remove solid precipitate and dried under vacuum to yield Compound 170.

Example 4: Preparation of (E)-3-(3,4-dimethoxyphenyl)acryloyl chloride (180)

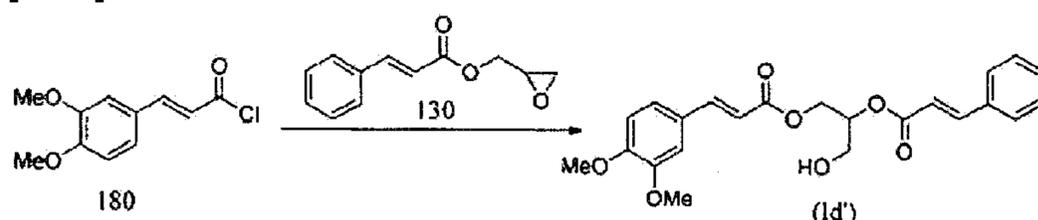
[0061]



[0062] To 5 mL of dried thionyl chloride (SOCl₂) contained in a reaction vessel was slowly added 4 g (19 mmol) of 3,4-dimethoxy cinnamic acid (140) and 50 μl of dried N,N'-dimethylformamide. The contents of the reaction vessel were mixed continuously at room temperature for 10-15 minutes, and the resulting product was dried in vacuo at 30-40°C to yield (E)-3-(3,4-dimethoxyphenyl)acryloyl chloride (180).

Example 5: Preparation of (E)-2-(cinnamoyloxy)-3-hydroxypropyl 3-(3,4-dimethoxyphenyl)acrylate (Id')

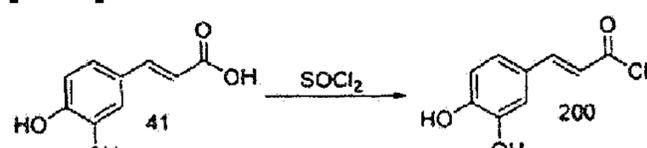
[0063]



[0064] To 3,4-dimethoxy cinnamoyl chloride (180; 2.5 g; 11 mmol) dissolved in 20 mL of dioxane was added a catalytic amount (250 mg) of tetrabutylammonium bromide and 1.33 g (6.5 mmol) of glycidyl cinnamate (130). The resulting mixture was heated under reflux (90°-95°C) with continuous stirring for 16-17 h and then distilled under reduced pressure to yield compound (Id').

Example 6: Preparation of Caffeoyl Chloride (200)

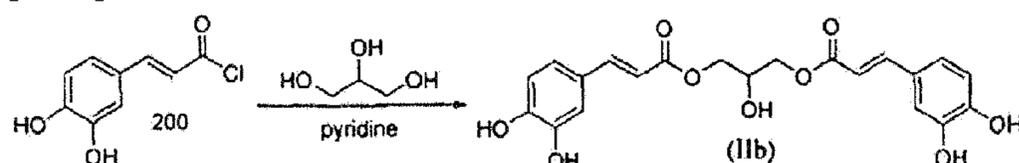
[0065]



[0066] 500 mg (2.78 mmol) of caffeic acid (190) was dissolved in 12 mL of dried dioxane at room temperature and then 600 μ E of SOCl_2 (thionyl chloride) was slowly added with mixing over approximately 20 minutes to produce a reaction mixture containing caffeoyl chloride (200).

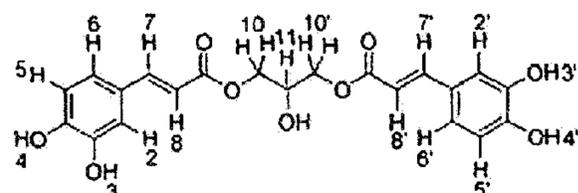
Example 7: Preparation of (2E, 2'E)-2-hydroxypropane-1,3-diyl bis (3-(3, 4-dihydroxyphenyl) acrylate (IIb))

[0067]



[0068] To the crude reaction mixture of Example 6 was added 248 mg (1.39 mmol) of anhydrous glycerol and a catalytic amount (300 μ L) of pyridine. The mixture was stirred for 30-40 minutes, and then approximately 1.0 g of NaHCO_3 and 10 mL of a methanol-ethyl acetate (50:50) mixture was added. After about 30 minutes of further stirring at room temperature, the reaction mixture was filtered and the solvent was removed. The reaction mixture was then dried under reduced pressure at approximately 60°C. The dried material was dissolved in 50 mL ethyl acetate, and washed three times with 100 mL ammonium formate, pH 3.7. The washed organic layer was dried over Na_2SO_4 and then under reduced pressure to obtain compound (IIb).

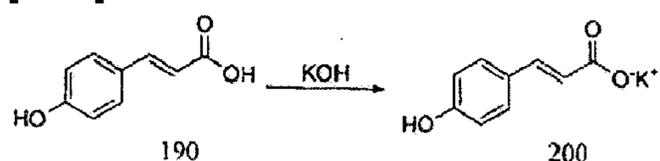
Compound (IIb) was then recrystallized from a mixture of ethyl acetate and cyclohexane (1 :3). $^1\text{H-NMR}$ (CD_3OD) δ 4.15 (m, 1H), 4.27 (m, 4H), 6.32 (d, 2H, $d = 16$ Hz), 7.60 (d, 2H, $J = 16$ Hz), 6.95 (dd, 2H, $J = 8, 2$ Hz), 6.77 (d, 2H, $d = 8$ Hz), 7.05 (d, 2H, $J = 2$ Hz)



Proton	Chemical Shift
2, 2'	7.05
5, 5'	6.77
6, 6'	6.95
7, 7'	7.60
8, 8'	6.32
10, 10'	4.27
11	4.15

Example 8*: Preparation of potassium 3-(4-hydroxyphenyl)acrylate (200)

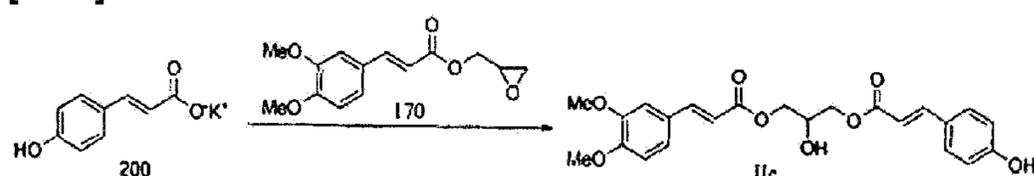
[0069]



[0070] To 15 mL of a 0.002 mM aqueous solution of potassium hydroxide was slowly added 5 g coumaric acid 190 (30 mmol) with mixing at 60-70°C. The resulting potassium coumarate (200) was dried under vacuum at 30-40°C.

Example 9*: Preparation of (E)-3-((E)-3-(3,4-dimethoxyphenyl)acryloyloxy)-2-hydroxypropyl 3-(4-hydroxyphenyl)acrylate (Ile)

[0071]

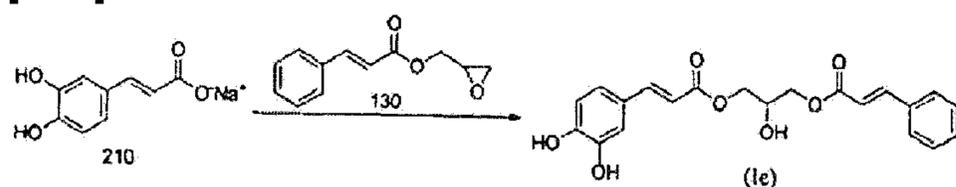


[0072] * forming not part of the invention, but left in the description for better understanding the invention.

[0073] To a solution of 1.5 g (5.68 mmol) of oxiran-2-ylmethyl 3-(3,4-dimethoxyphenyl)acrylate (170) dissolved in 30 mL of dried dioxane was added 2.29 g (1 1.36 mmol) of potassium coumarate (200) dissolved in 20 mL dioxane and 100 mg of tetrabutylammonium bromide, and the resulting mixture was heated to a temperature of 105-110°C for 16-18 hours to produce diester (Ile).

Example 10. Preparation of (E)-3-(cinnamoyloxy)-2-hydroxypropyl 3-(3,4-dihydroxyphenyl)acrylate (Ie)

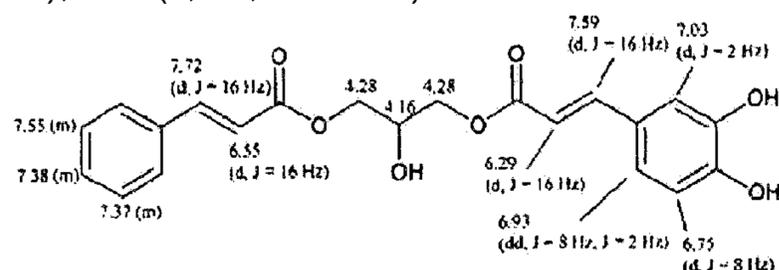
[0074]



[0075] Caffeic acid (41 ; 18.02 g; 0.10 mol) was dissolved in THF (180 mL) with stirring at about 30°C to yield a 10% solution. A powder of freshly ground sodium hydroxide pellets (4.00 g; 0.10 mol) was added to the caffeic acid solution and a solid precipitate of sodium caffeate (210) was formed, which was filtered and dried under reduced pressure.

[0076] To a solution of sodium caffeate (210; 7.35 mmol) in DMSO (20 mL) at 60°-70°C was added glycidyl cinnamate (130; 7.35 mmol). The resulting mixture was heated to 60°-75°C with continuous stirring for 22-24 h and then distilled under reduced pressure to yield compound (Ie). ¹H-NMR (CD₃OD)

8 4.16 (s, 1H), 2.28 (s, 4H), 6.29 (d, 1H, J = 16 Hz), 6.65 (d, 1H, J = 16 Hz), 6.75 (d, 1H, J = 8 Hz), 6.93 (1H, dd, J = 8, 2 Hz), 7.03 (d, 1H, J = 2 Hz), 7.37 (m, 1H), 7.38 (m, 1H), 7.55 (m), 7.59 (d, 1H, J = 16 Hz), 7.72 (d, 1H, J = 16 Hz).



Anti-cancer activity

[0077] The compounds prepared in the present invention exhibited good in vitro anticancer activity towards human B-16 melanoma cell lines. The cell lines were maintained in EMEM medium supplemented with 10% FCS, 0.1% sodium bicarbonate and 12mM glutamine. In a typical procedure, 1×10^4 cells were seeded into each of 96 well plate in 90 volume of medium. The plates were incubated for 24 h in the presence of CO₂ to allow for cell attachment. After 24 h, test compounds were evaluated at five 10 fold dilutions from 1 :10, 1 :100, 1 :1000, 1 :10000 and 1 :100000. To each test well 100 μ L of test compound solution was added and the medium with vehicle was added to control wells and the plates were further incubated. After 24 h of incubation, 10 μ L of [3H] - thymidine was added to each well to obtain a 1 μ Ci concentration per well and incubated for a further 24h. The plates were terminated, cells harvested and read by microbeta plate reader. The results are shown in Table 1.

Table 1. Values of EC₅₀ of compounds of the present invention in the B-16 melanoma cell line, μ M

Compound	EC ₅₀ (μ M)
 (Ia)	1.08
 (IIb)	6.12

MTT (cell proliferation and viability) assay for anticancer compounds of the present invention

[0078] Normal human fibroblast cell lines (GM9503 and GM8399) and human carcinoma cell lines MCF-7, A549, HCT116, SKOV-3 and PC-3 were maintained in DMEM (Dulbecco's Modified Essential Medium) supplemented with 10% FBS (Fetal Bovine Serum). In addition, murine leukemia MDAY-D2 cells were maintained in RPMI 1640 media supplemented with 5% FBS. All media were supplemented with 100 units/mL of streptomycin and 100 μ g/mL of penicillin (all from Hyclone, Logan, UT). Cells were incubated in a humidified air atmosphere containing 5% CO₂ at 37°C.

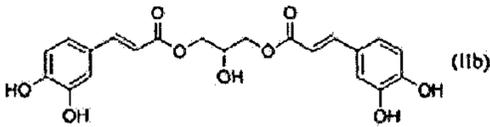
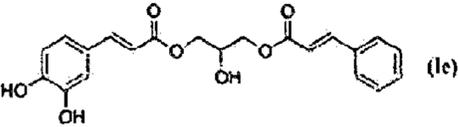
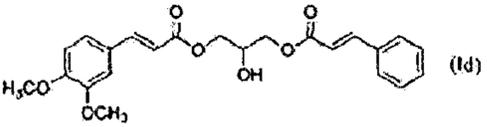
[0079] Cell growth and viability were measured using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) reduction assay. MTT reagent (Sigma, St. Louis, MO) was prepared at 5

mg/ml in ddH₂O (5X solution), filter sterilized and stored in the dark at - 20°C.

[0080] To determine the optimal number of cells to seed per well, preliminary experiments were conducted examining increasing cell densities in 96 well plates. Cell were seeded at various densities and examined under the microscope after 96 h. Based on these experiments and growth curve analyses, optimal cell densities were selected for each cell line. In brief, exponentially growing cells were harvested and 100 µL of cell suspension containing approximately 2000 cells was plated in 96-well microtiter plates. After 24 hours of incubation to allow for cell attachment, cells were treated with varying concentrations of test samples in medium (60 µL/well) supplemented with 5% FBS and incubated for 72 h at 37°C under 5% CO₂. Three hours after the addition of MTT, the amount of formazan formed was measured spectrophotometrically at 570 nm with a Spectramax Plus 384 plate reader.

[0081] To assess the effects of the novel compounds on growth and viability, GM9503 (normal human fibroblast cells; 2.08×10^3 cells/well), GM8399 (normal human fibroblast cells; 2.04×10^3 cells/well), A549 (human lung carcinoma cells; 2.05×10^3 cells/well), HCT116 (human colon carcinoma cells; 2.19×10^3 cells/well), MCF-7 (human breast carcinoma cells; 2.19×10^3 cells/well), SKOV-3 (human ovarian carcinoma cells; 2.10×10^3 cells/well), PC-3 (human prostate carcinoma cells; 2.08×10^3 cells/well) and MDAY-D2 (murine leukemia cells; 2.5×10^3 cells/well) were seeded in 96 well plates in supplemented medium. Twenty-four hours post-seeding, cells were treated with either DMEM supplemented with 5% FBS (as a control) or with compounds (IIb), (Ie) and (Id) at increasing concentrations. After incubation for 72 h, cell growth and viability were measured. Each compound was tested in triplicate and the data shown in Tables 2-4 represent the mean of viable cells compared to medium control.

Table 2. *In-vitro* inhibition of carcinoma cells, comparative EC₅₀ (µM)

Compound	Human (EC ₅₀) µM				Murine (EC ₅₀) µM
	MCF-7 (Breast Carcinoma)	A549 (Lung Carcinoma)	HCT116 (Colon Carcinoma)	GM9503 (Normal Skin Fibroblast)	MDAY-D2 (Leukemia)
 (IIb)	1.0	2.17	NE*	57.9	3.4
 (Ie)	25.9	0.64	59.2	NE*	1.5
 (Id)	72.2	0.59	102.5	NE*	5.0
DMSO	NE*	NF*	NE*	20.3	43.6

*NE (no significant inhibitory effect)

Table 3. *In-vitro* inhibition of carcinoma cells, comparative IC₅₀ (µM)

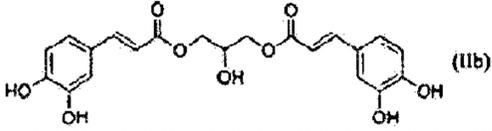
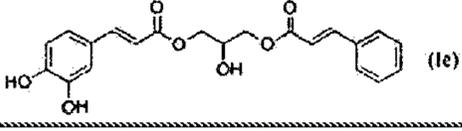
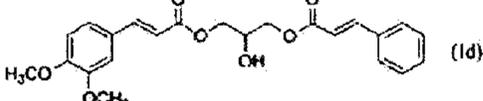
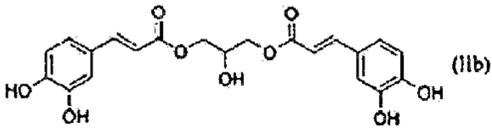
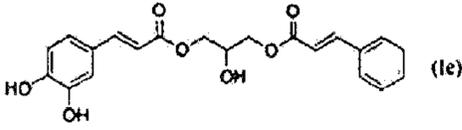
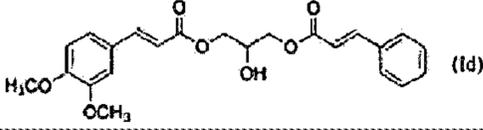
Compound	Anticancer Compound Inhibition Concentration (IC ₅₀) μM in human carcinoma cell lines				
	A549 (Lung)	MCF-7 (Breast)	HCT116 (Colon)	PC-3 (Prostate)	SKOV-3 (Ovarian)
paclitaxel	0.69	11.7	0.59	1.32	0.75
 (IIb)	21.04	74.46	26.98	6.76	15.78
 (Ie)	33.2	33.02	27.88	9.04	16.52
 (Id)	18.18	85.15	11.49	9.07	22.01

Table 4. Comparative Toxicity of compounds of the present invention in normal human fibroblast cells *in-vitro* (IC₅₀, μM)

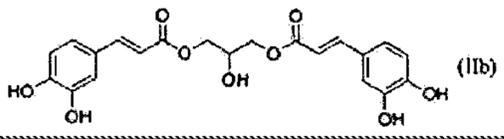
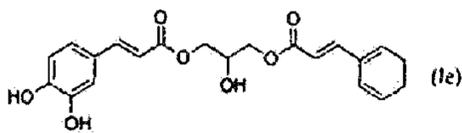
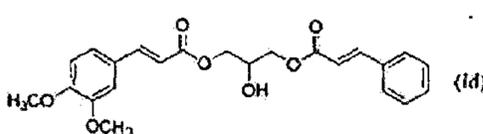
Compound	Inhibition Concentration (IC ₅₀) μM	
	GM9503 (Normal human fibroblast)	GM8399 (nonnal human fibroblast)
paclitaxel	0.7	2.1
caffeic Acid	22.0	24.0
 (IIb)	71.2	38
 (Ie)	37.0	23
 (Id)	60.0	16

[0082] As shown above in Table 4, in normal human fibroblast cell lines, paclitaxel and caffeic acid are more toxic than compounds (IIb), (Ie) and (Id).

[0083] The therapeutic index for an anticancer compound compares the toxicity of the compound in a normal cell with an anti-proliferative effect in carcinoma cells. The index is a measure of the safety of the anticancer compound. The ratio of an IC₅₀ value in a normal human cell (e.g. a human fibroblast GM9503 cell) to an IC₅₀ value in a human carcinoma cell line for a particular anticancer compound provides comparative information on the safety and selectivity of the anticancer compound. The values of the therapeutic index shown in Table 5 demonstrate that the anticancer compounds (Ib), (Ie), and (Id) possess a larger therapeutic index and are therefore less toxic in comparison to paclitaxel (an approved

anticancer drug). These results are significant since a successful anticancer therapy must not only demonstrate antitumor cytotoxicity but also show tolerable toxicity with respect to normal healthy cells.

Table 5. Values of therapeutic (safety) index* of anticancer compounds of the present invention

Compound	A549 (Lung)	MCF-7 (Breast)	HCT116 (Colon)	PC-3 (Prostate)	SKOV-3 (Ovarian)
Taxol (Control)	1.01	0.06	1.18	0.53	0.93
 (IIb)	3.38	0.95	2.63	10.52	4.51
 (Ie)	1.12	1.13	1.34	4.14	2.26
 (Id)	3.31	0.70	5.25	6.64	2.74
*Therapeutic index = toxicity (fibroblast)/ anticancer inhibition = $IC_{50}(GM9503)/IC_{50}$ (carcinoma cell line)					

Mechanism of action

[0084] Human colon carcinoma cell line HCT116 was treated with compounds (IIb), (Ie) and (Id) and observed over a time period to determine the mechanism of cell death. The experiment was conducted as follows:

[0085] HCT116 cells were seeded and treated with each compound. At each time point, the cells were imaged using a neutral microscope at 63X magnification prior to fixing. The time points were 2, 4, 6, 8, 12, and 16 hours.

[0086] Cells treated with the compounds (IIb), (Ie) and (Id) exhibited the characteristic membrane blebbing morphology commonly seen during apoptosis (see Figures 12-15). Compound (IIb) appeared to be the most aggressive, followed by compound (Ie) and compound (Id). At 4 hr, a few cells in each well displayed membrane blebbing and a majority of the initial cellular population appeared to be undergoing apoptosis at the 6 hr and 8 hr time-points. At the 10 and 12 hr time-points, significantly fewer live cells were observed compared to the 2 and 4 hr time points, however, the cellular population appeared to slightly increase at 16 hrs. This may potentially be caused by a population doubling at this time-point.

References

[0087]

1. (1) Jiang MC, Yang-Yen HF, Yen JJ and Lin JK: Curcumin induces apoptosis in immortalized N1H

- 3T3 and malignant cancer cell lines. *Nutr Cancer* 26: 111-20, 1996.
2. (2) Kuo ML, Huang TS and Lin JK: Curcumin, an antioxidant and anti-tumor promoter, induces apoptosis in human leukemia cells. *Biochim Biophys Acta* 1317: 95-100, 1996.
 3. (3) Mehta K, Pantazis P, McQueen T and Aggarwal BB: Antiproliferative effect of curcumin (diferuloylmethane) against human breast tumor cell lines. *Anticancer Drugs* 8: 470-81, 1997.
 4. (4) Samaha HS, Kelloff GJ, Steele V, Rao CV and Reddy BS: Modulation of apoptosis by sulindac, curcumin, phenylethyl-3-methylcaffeate, and 6-phenylhexyl isothiocyanate: apoptotic index as a biomarker in colon cancer chemoprevention and promotion. *Cancer Res* 57: 1301-5, 1997.
 5. (5) Simon A, Allais DP, Duroux JL, Basly JP, Durand-Fontanier S and Delage C: Inhibitory effect of curcuminoids on MCF-7 cell proliferation and structure- activity relationships. *Cancer Lett* 129:111-6, 1998.
 6. (6) Jee SH, Shen SC, Tseng CR, Chiu HC and Kuo ML: Curcumin induces a p53-dependent apoptosis in human basal cell carcinoma cells. *J Invest Dermatol* 111: 656-61, 1998.
 7. (7) Mukhopadhyay A, Bueso-Ramos C, Chatterjee D, Pantazis P and Aggarwal BB: Curcumin downregulates cell survival mechanisms in human prostate cancer cell lines. *Oncogene* 20: 7597-609, 2001.
 8. (8) Grunberger, D., Banerjee, R., Eisinger, K., Oltz, E. H., Efros, L., Caldwell, M., Estevez, V., and Nakanishi, K. Preferential cytotoxicity on tumor cells by caffeic acid phenethyl ester isolated from propolis. *Experientia*. 44: 230-232, 1988.
 9. (9) Su, Z-Z., Grunberger, D., and Fisher. P. B. Suppression of adenovirus type 5 E1A-mediated transformation and expression of the transformed phenotype by caffeic acid phenethyl ester (CAPE). *Mol. Carcinogenesis*. 4: 231-242, 1991.
 10. (10) Guarini. L., Su. Z-z., Zucker, S., Lin, J., Grunberger, D.. and Fisher. P. B. Growth inhibition and modulation of antigenic phenotype in human melanoma and glioblastoma multiforme cells by caffeic acid phenethyl ester (CAPE). *Cell. Mol. Biol.*, 38:513-527, 1992.
 11. (11) Rao, C. V., Desai, D., Kaul, B., Amin, S., and Reddy, B. S. Effect of caffeic acid esters on carcinogen-induced mutagenicity and human colon adenocarcinoma cell growth. *Chem.-Biol. Interact.*, 84: 277-290, 1992.
 12. (12) Rao, C. V., Desai, D., Simi, B., Kulkarni, N., Amin, S., and Reddy, B. S. Inhibitory effect of caffeic acid esters on azoxymethane-induced biochemical changes and aberrant crypt foci formation in rat colon. *Cancer Res.* 53: 4182-4188. 1993.
 13. (13) Frankel. K., Wei, H., Bhimani, R., Zadunaisky, J. A., Ferrara, T., Huang, M. T., Conney, A. H., and Grunberger, D. Inhibition of tumor promoter-mediated processes in mouse skin and bovine lens by caffeic acid phenethyl ester. *Cancer Res.*, 53:1255-1261, 1993.

REFERENCES CITED IN THE DESCRIPTION

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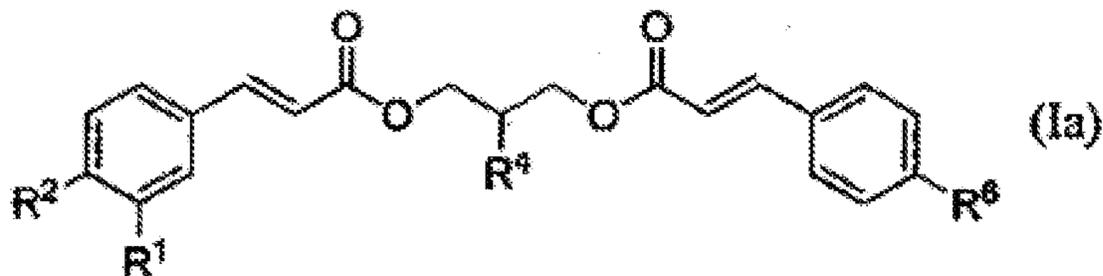
Non-patent literature cited in the description

- **BANSKOTA et al.**Journal of Ethnopharmacology, 2002, vol. 80, 167-73 [0010]
- **DAE SIK JANG et al.**Constituents of asparagus officinalis evaluated for inhibitory activity against cyclooxygenase-2J. of agricultural and food chemistry, 2004, vol. 52, 2218-2222 [0010]
- **XIONG YDENG K ZGUO Y QZHANG T J**New chemical constituents from the rhizomes of sparganium stoloniferumArchives of Pharmacal Research, 2009, vol. 32, 5717-720 [0010]
- **JIANG MCYANG-YEN HFYEN JLIN JK**Curcumin induces apoptosis in immortalized N1H 3T3 and malignant cancer cell linesNutr Cancer, 1996, vol. 26, 111-20 [0087]
- **KUO MLHUANG TSLIN JK**Curcumin, an antioxidant and anti-tumor promoter, induces apoptosis in human leukemia cellsBiochim Biophys Acta, 1996, vol. 1317, 95-100 [0087]
- **MEHTA KPANTAZIS PMCQUEEN TAGGARWAL BB**Antiproliferative effect of curcumin (diferuloylmethane) against human breast tumor cell linesAnticancer Drugs, 1997, vol. 8, 470-81 [0087]
- **SAMAHA HSKELLOFF GJSTEELE VRAO CVREDDY BS**Modulation of apoptosis by sulindac, curcumin, phenylethyl-3-methylcaffeate, and 6-phenylhexyl isothiocyanate: apoptotic index as a biomarker in colon cancer chemoprevention and promotionCancer Res, 1997, vol. 57, 1301-5 [0087]
- **SIMON AALLAIS DPDUROUX JLBASLY JPDURAND-FONTANIER SDELAGE C**Inhibitory effect of curcuminoids on MCF-7 cell proliferation and structure- activity relationshipsCancer Lett, 1998, vol. 129, 111-6 [0087]
- **JEE SHSHEN SCTSENG CRCHIU HCKUO ML**Curcumin induces a p53-dependent apoptosis in human basal cell carcinoma cellsJ Invest Dermatol, 1998, vol. 111, 656-61 [0087]
- **MUKHOPADHYAY ABUESO-RAMOS CCHATTERJEE DPANTAZIS PAGGARWAL BB**Curcumin downregulates cell survival mechanisms in human prostate cancer cell linesOncogene, 2001, vol. 20, 7597-609 [0087]
- **GRUNBERGER, D.BANERJEE, R.EISINGER, K.OLTZ. E. H.EFROS, L.CALDWELL, M.ESTEVEZ, V.NAKANISHI, K.**Preferential cytotoxicity on tumor cells by caffeic acid phenethyl ester isolated from propolisExperientia, 1988, vol. 44, 230-232 [0087]
- **SU, Z-Z.GRUNBERGER, D.FISHER. P. B.**Suppression of adenovirus type 5 E1A-mediated transformation and expression of the transformed phenotype by caffeic acid phenethyl ester (CAPE)Mol. Carcinogenesis, 1991, vol. 4, 231-242 [0087]
- **GUARINI. L.SU. Z-Z.ZUCKER, S.LIN, J.GRUNBERGER, D.FISHER. P. B.**Growth inhibition and modulation of antigenic phenotype in human melanoma and glioblastoma multiforme cells by caffeic acid phenethyl ester (CAPE)Cell. Mol. Biol., 1992, vol. 38, 513-527 [0087]
- **RAO, C. V.DESAI, D.KAUL, B.AMIN, S.REDDY, B. S.**Effect of caffeic acid esters on carcinogen-induced mutagenicity and human colon adenocarcinoma cell growthChem.-Biol. Interact., 1992, vol. 84, 277-290 [0087]
- **RAO, C. V.DESAI, D.SIMI, B.KULKARNI, N.AMIN, S.REDDY, B. S.**Inhibitory effect of caffeic acid esters on azoxymethane-induced biochemical changes and aberrant crypt foci formation in rat colonCancer Res., 1993, vol. 53, 4182-4188 [0087]
- **FRANKEL. K.WEI, H.BHIMANI, R.ZADUNAISKY, J. A.FERRARA, T.HUANG, M. T.CONNEY, A. H.GRUNBERGER, D.**Inhibition of tumor promoter-mediated processes in mouse skin and bovine lens by caffeic acid phenethyl esterCancer Res., 1993, vol. 53, 1255-1261 [0087]

PATENTKRAV

1. Forbindelse til anvendelse i behandlingen af cancer, hvor forbindelsen har formelen

(Ia):



5

hvor:

R^1 og R^2 uafhængigt er OH, alkoxy eller alkylcarbonyloxy,

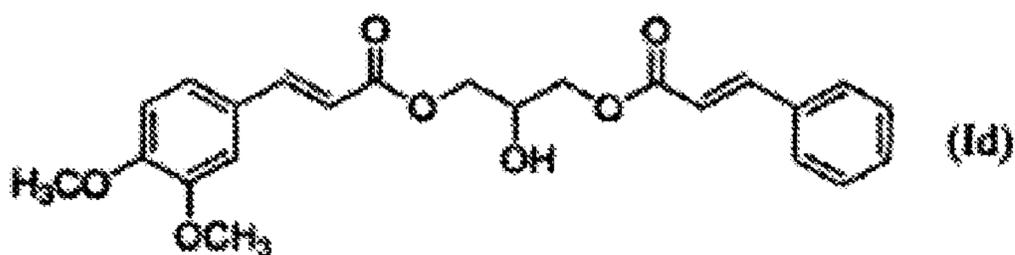
R^3 er OH, og

10

R^6 er H,

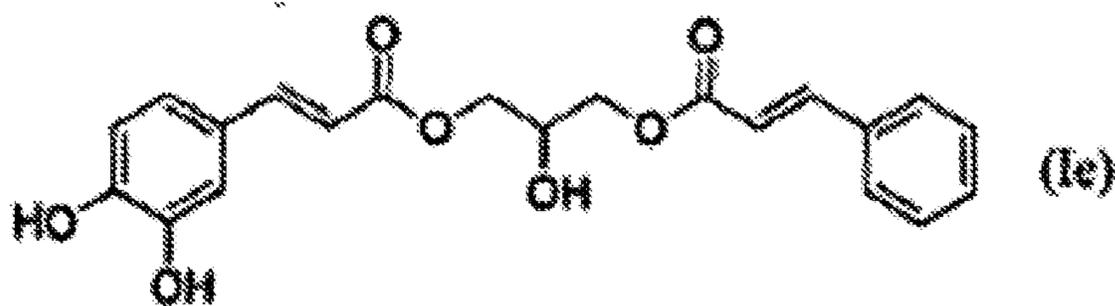
eller et farmaceutisk acceptabelt salt deraf.

2. Forbindelse ifølge krav 1 til anvendelse i behandlingen af cancer, hvor forbindelsen er mindst én forbindelse med formelen:



15

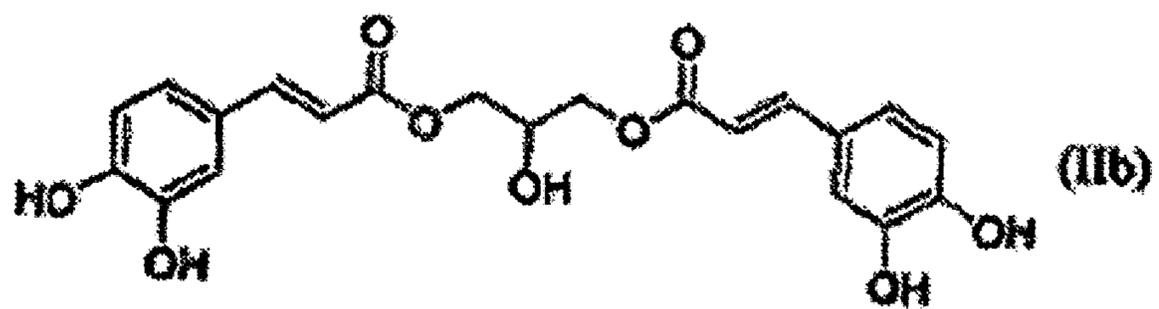
og



3. Farmaceutisk sammensætning til anvendelse i behandlingen af cancer omfattende mindst én forbindelse som anvendt i et hvilket som helst af kravene 1 til 2.

20

4. Forbindelse med formelen (IIb) til anvendelse i behandlingen af cancer:



DRAWINGS

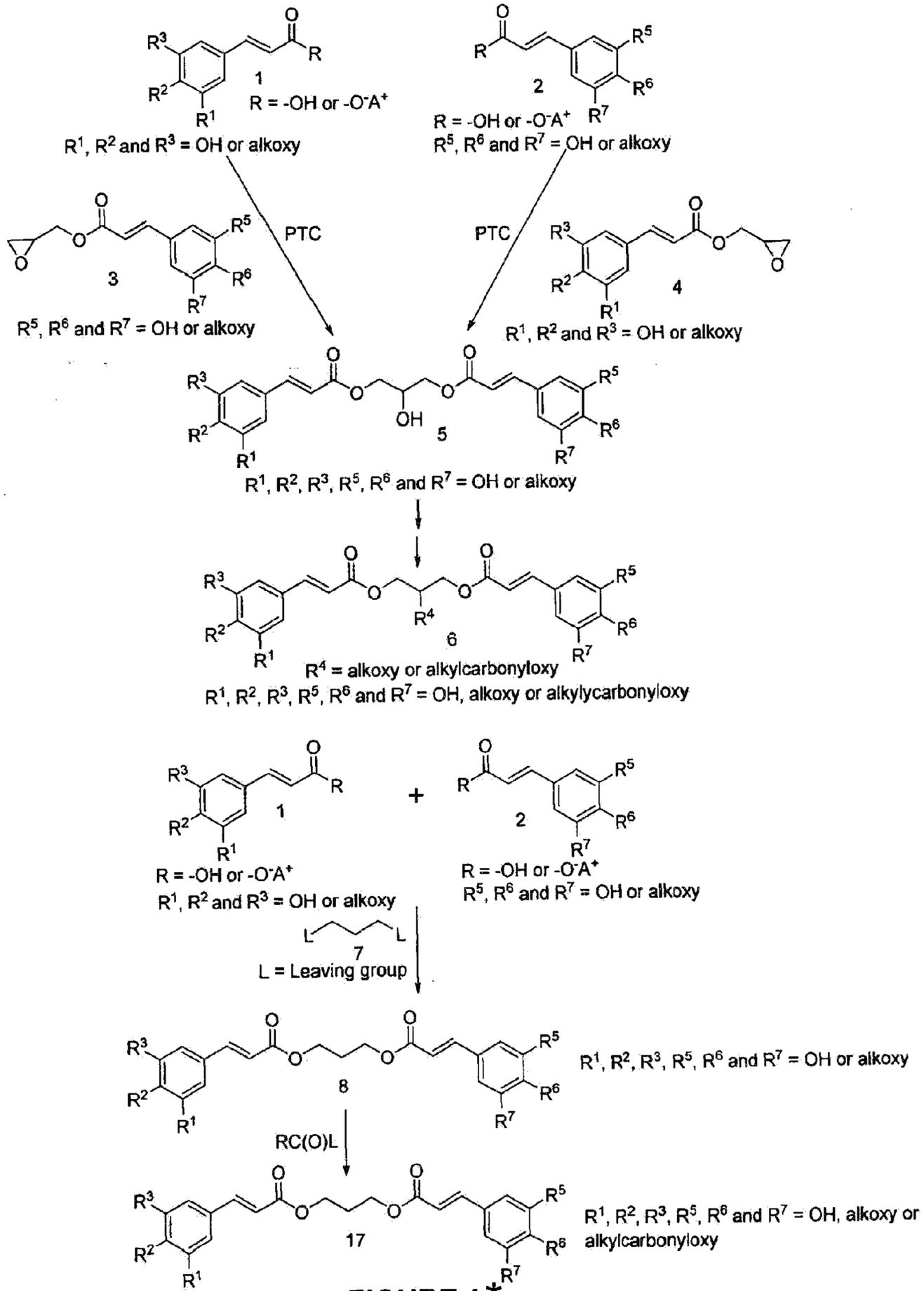


FIGURE 1*

* not according to the invention

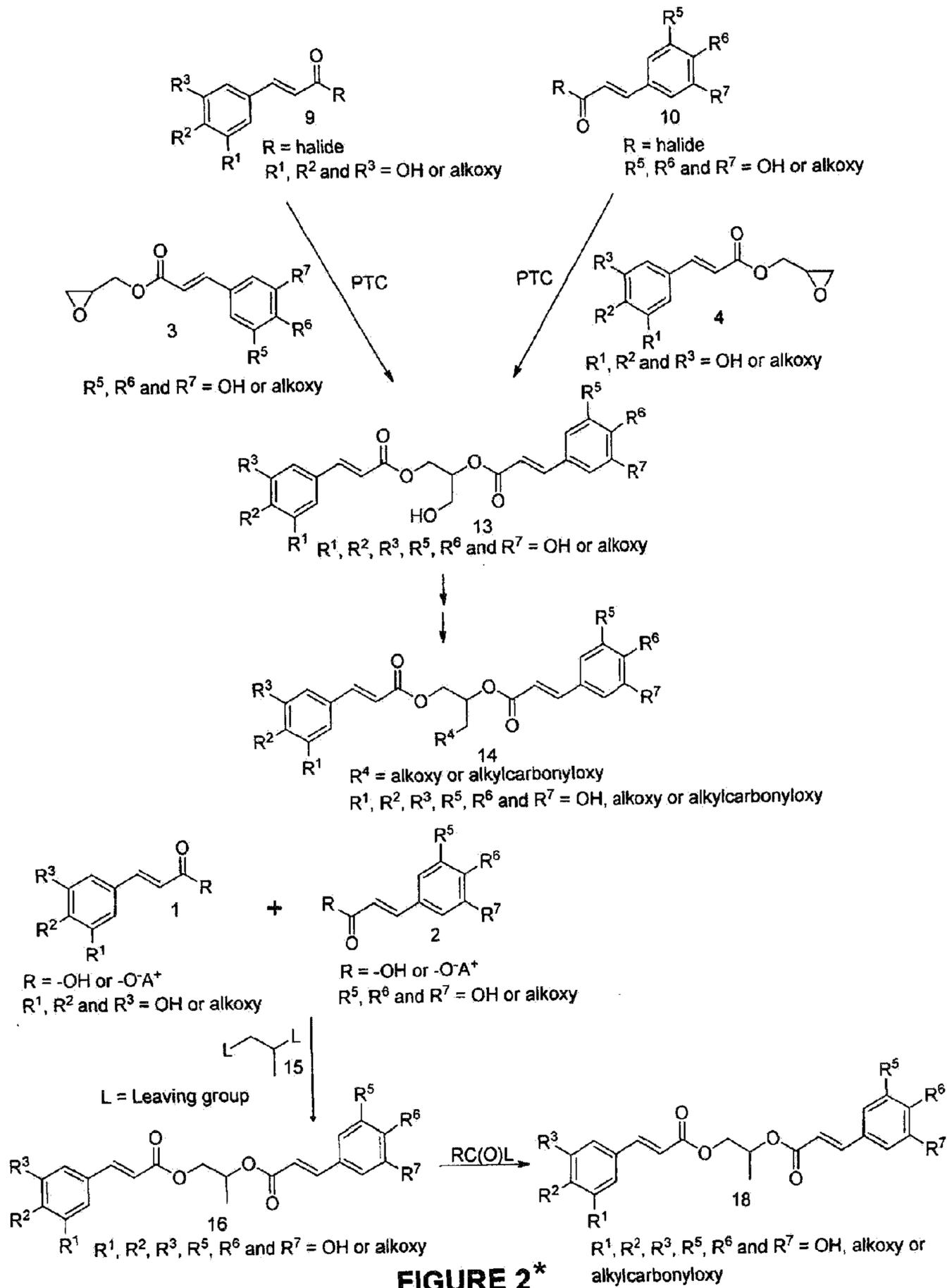


FIGURE 2*

* not according to the invention

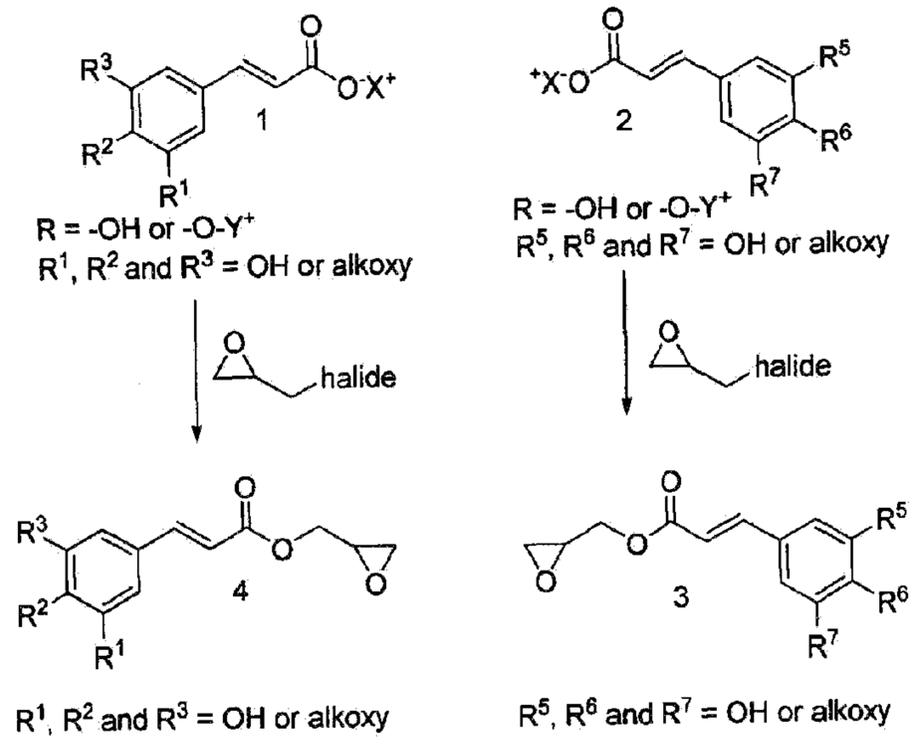


FIGURE 3

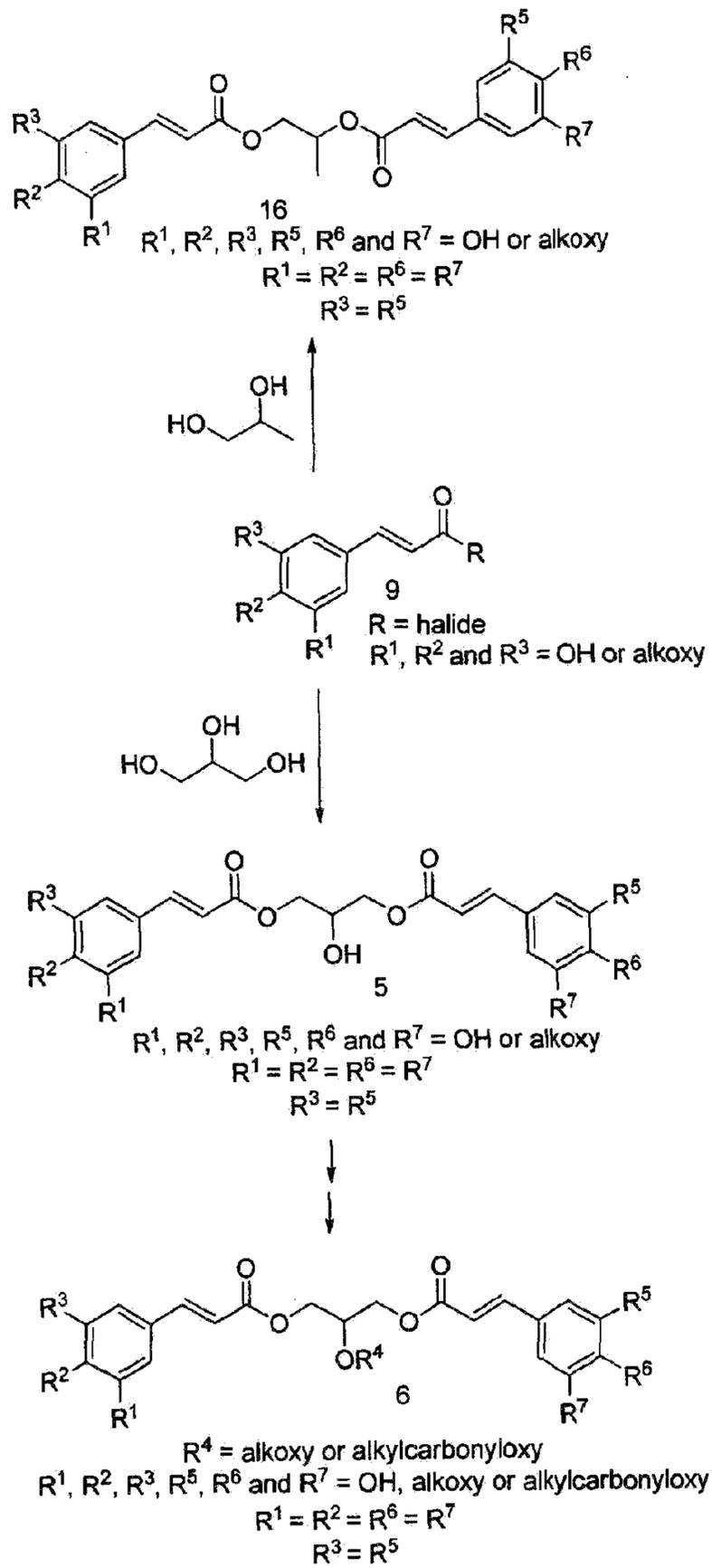


FIGURE 4 *

* not according to the invention

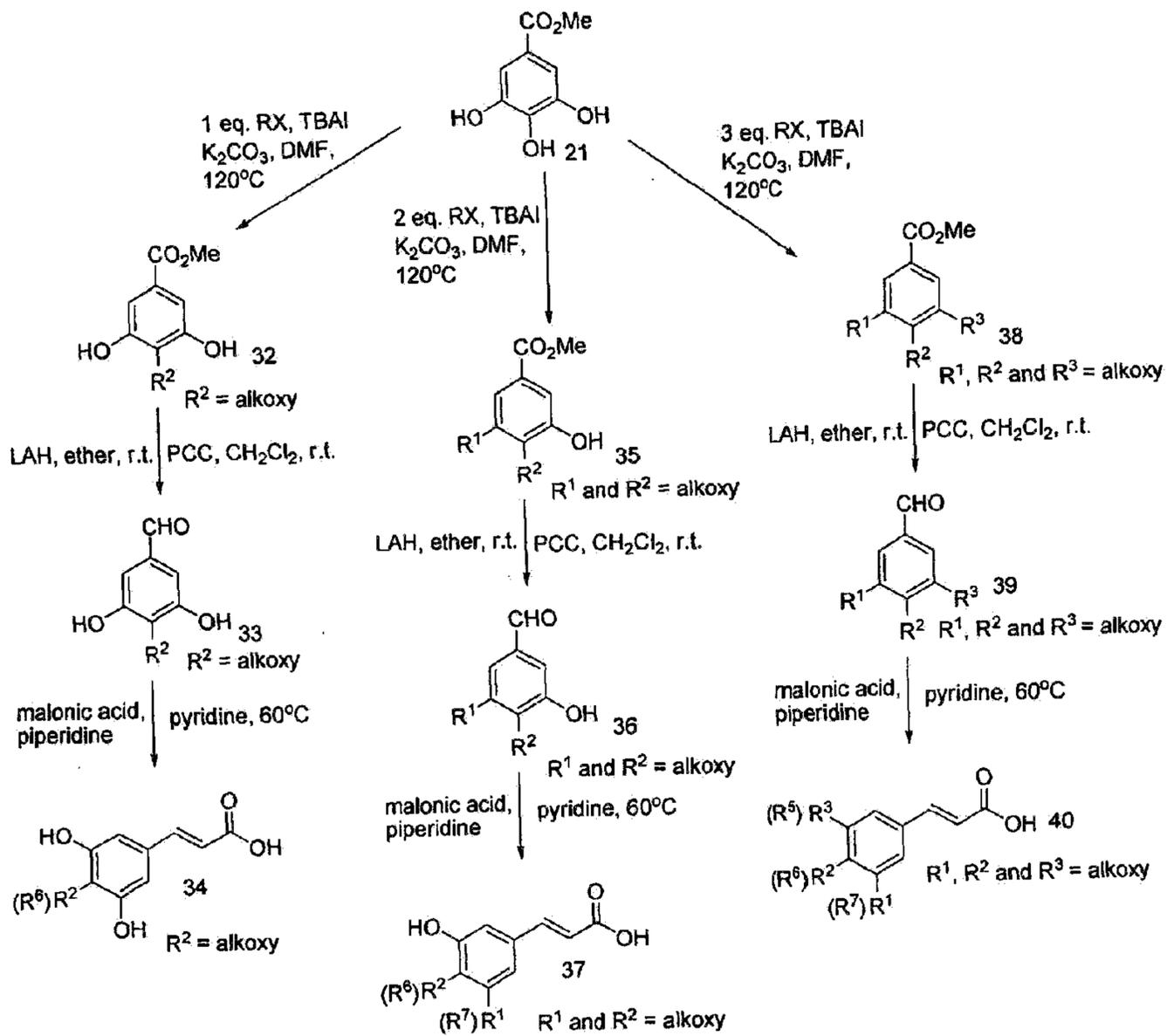


FIGURE 6

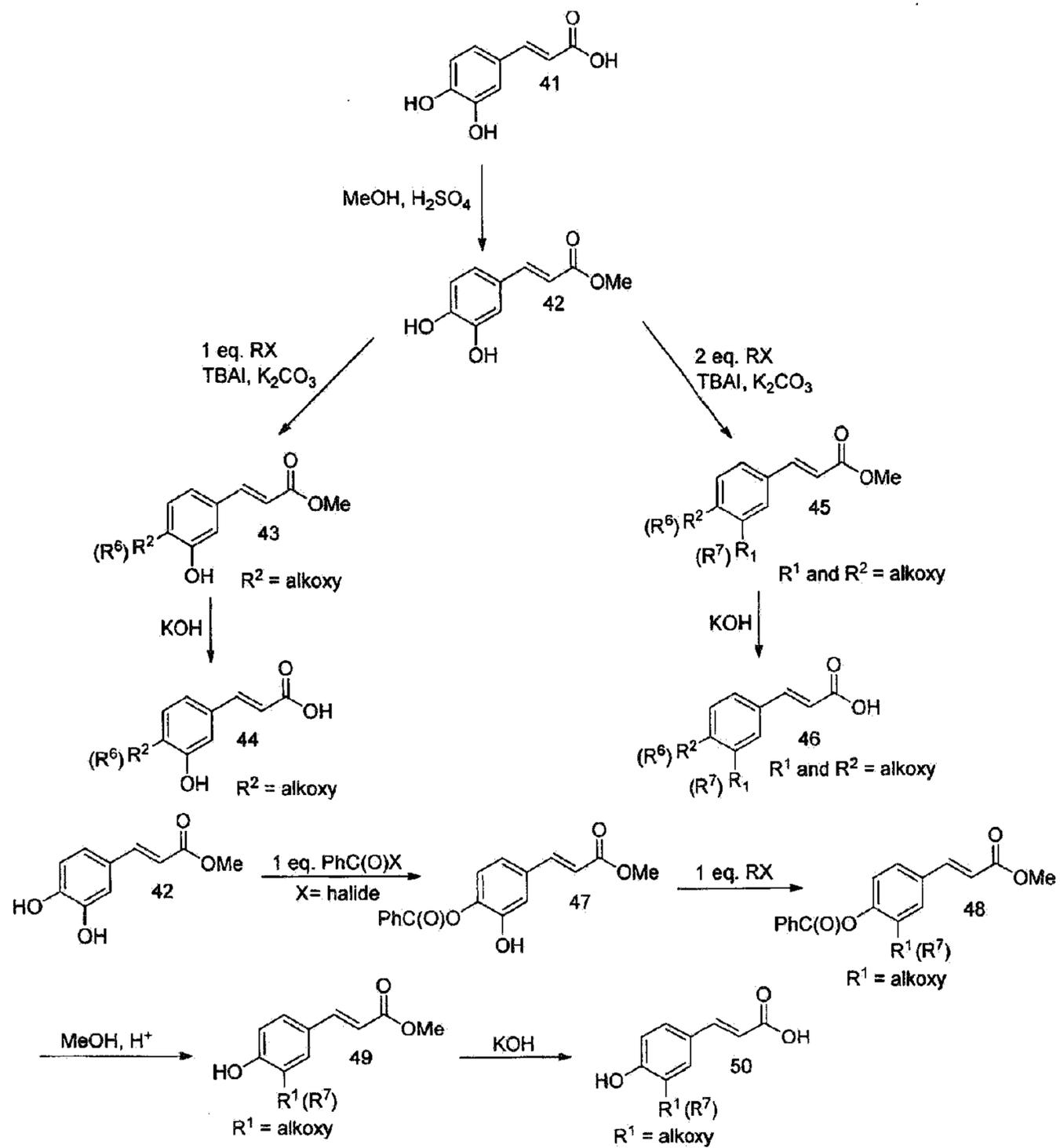


FIGURE 7

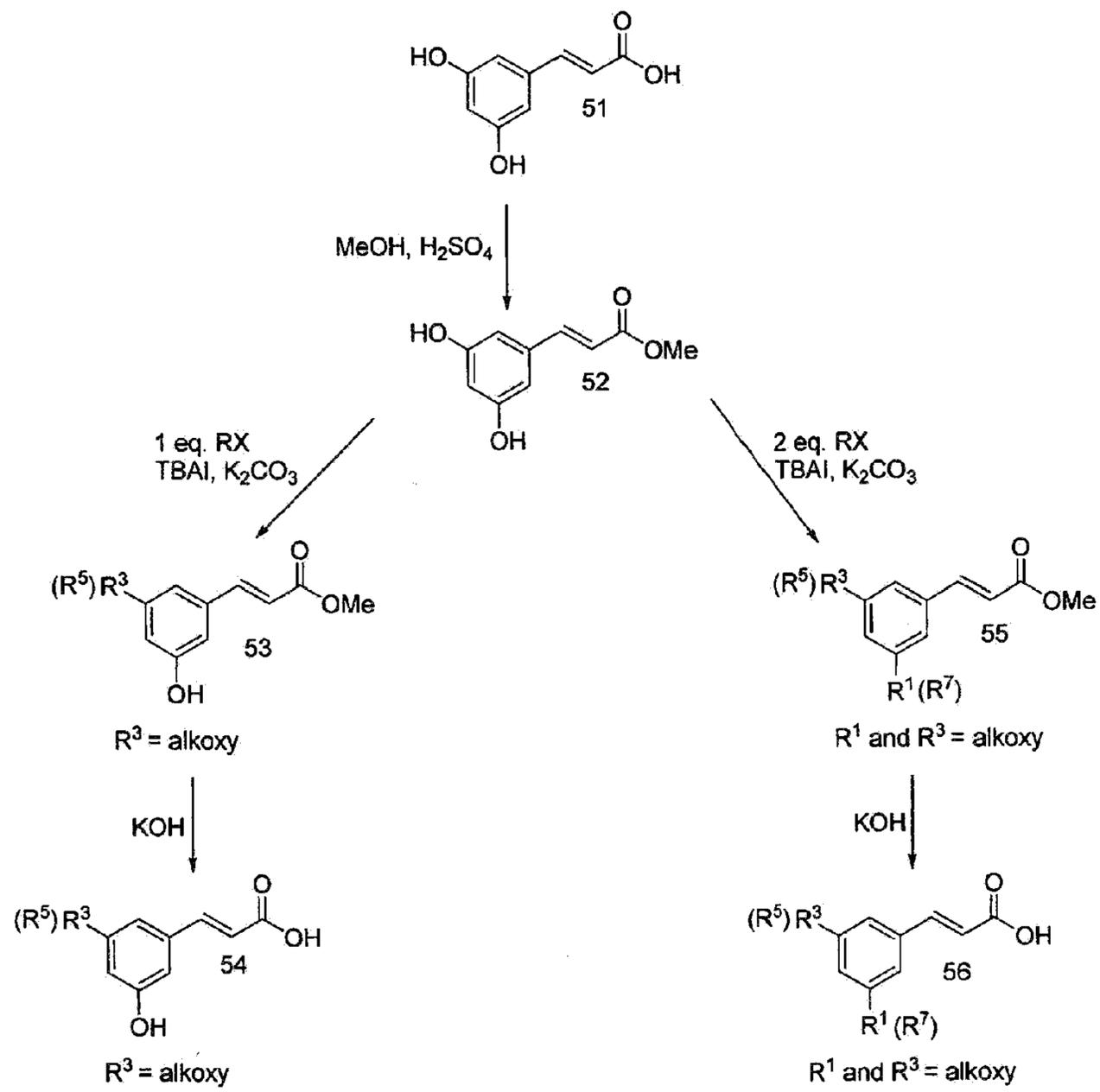


FIGURE 8

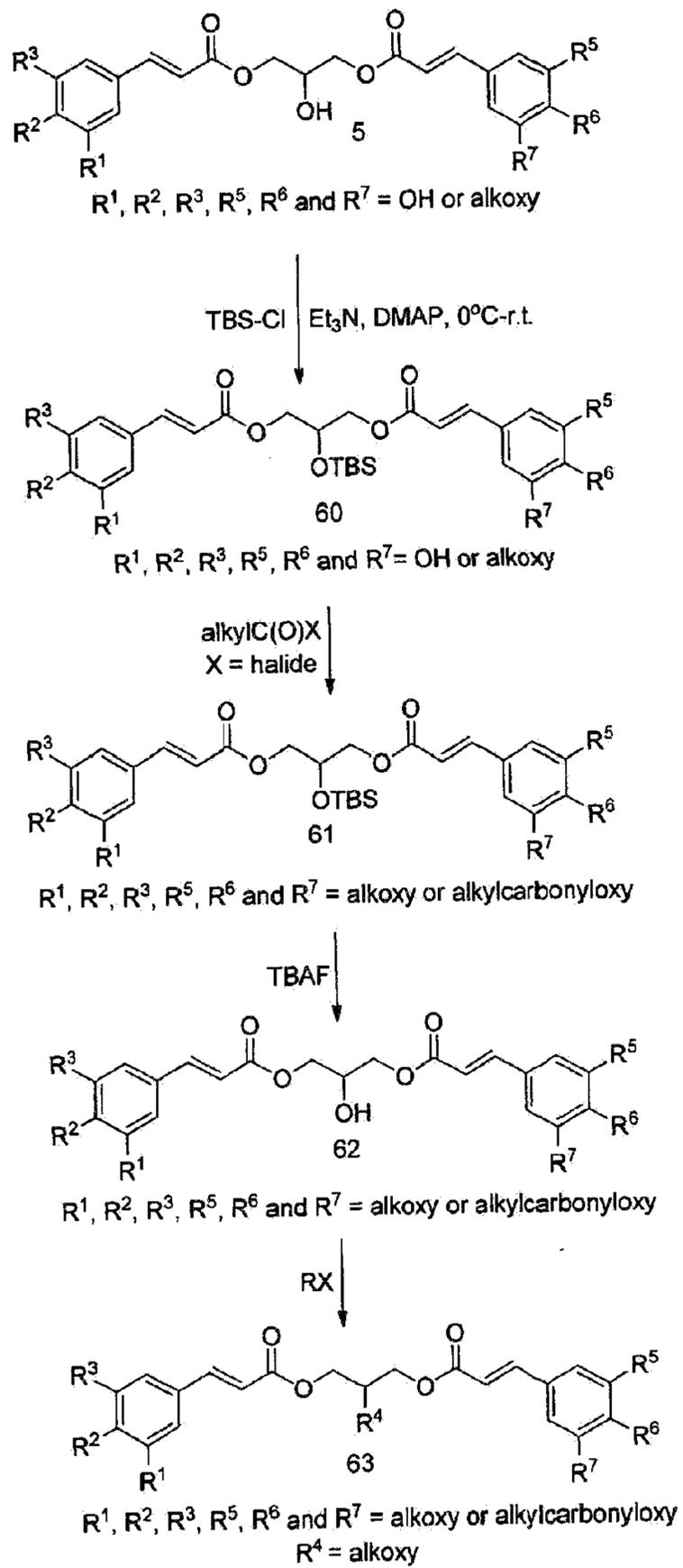


FIGURE 9*

* not according to the invention

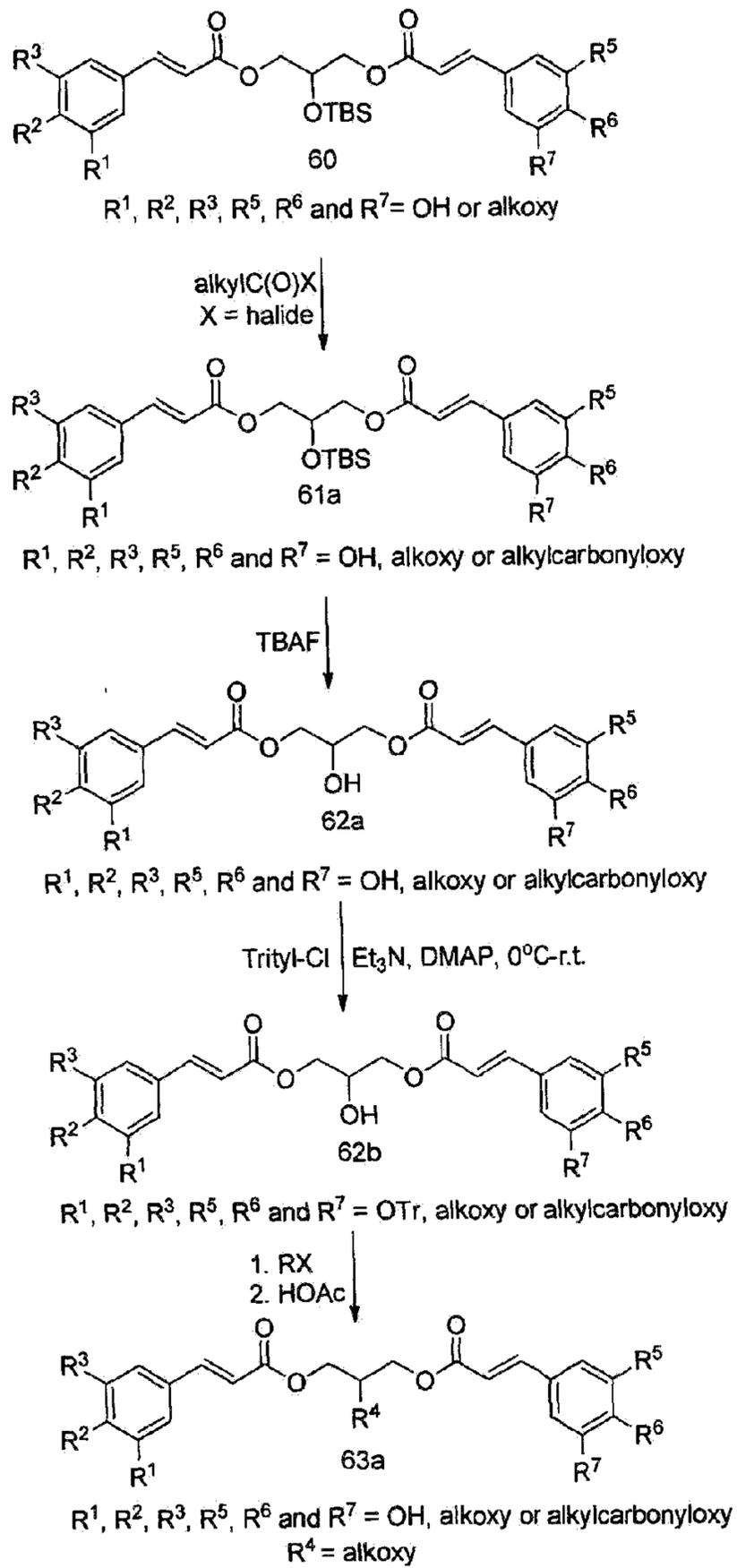


FIGURE 10*

* not according to the invention

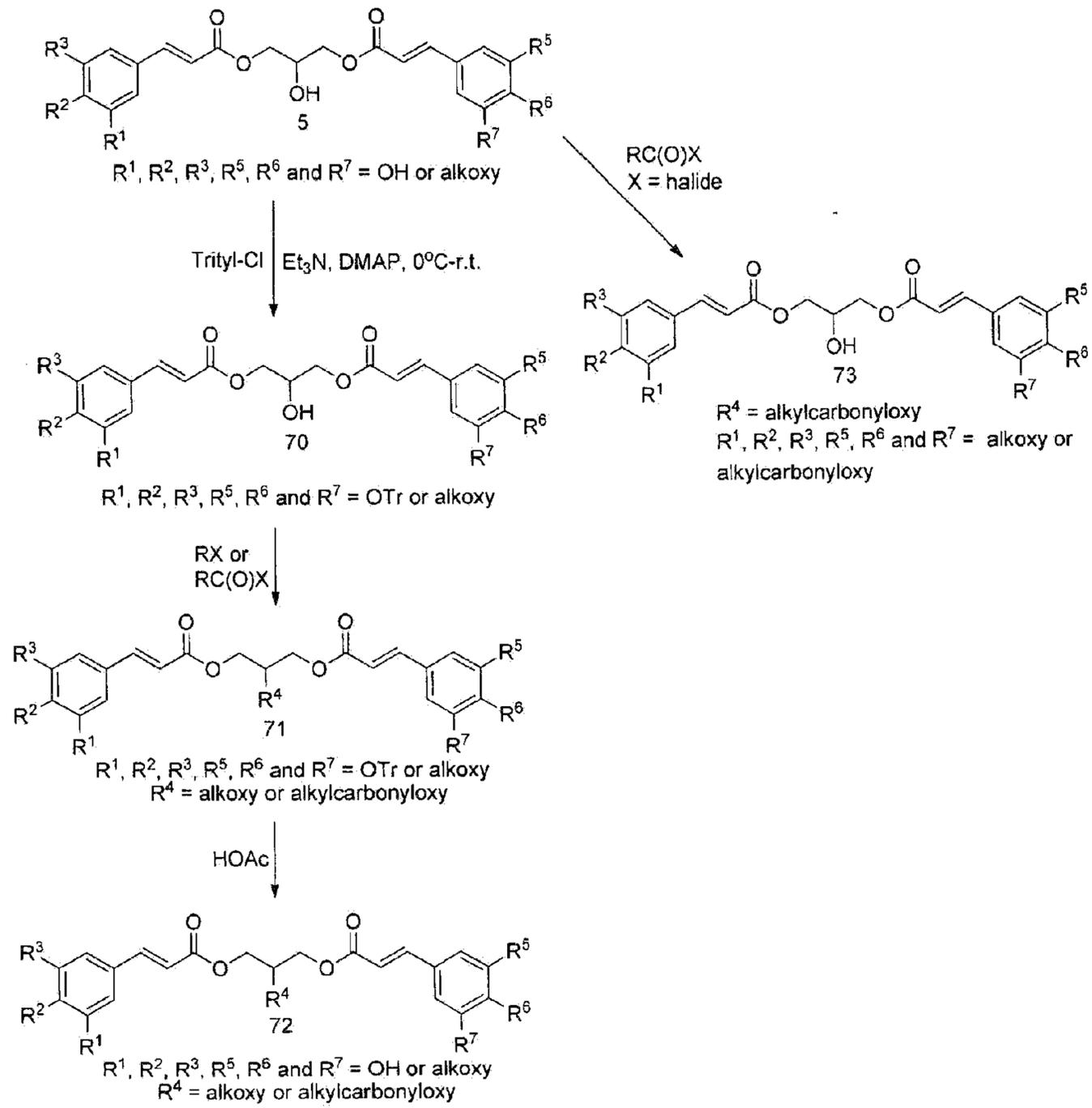


FIGURE 11*

* not according to the invention

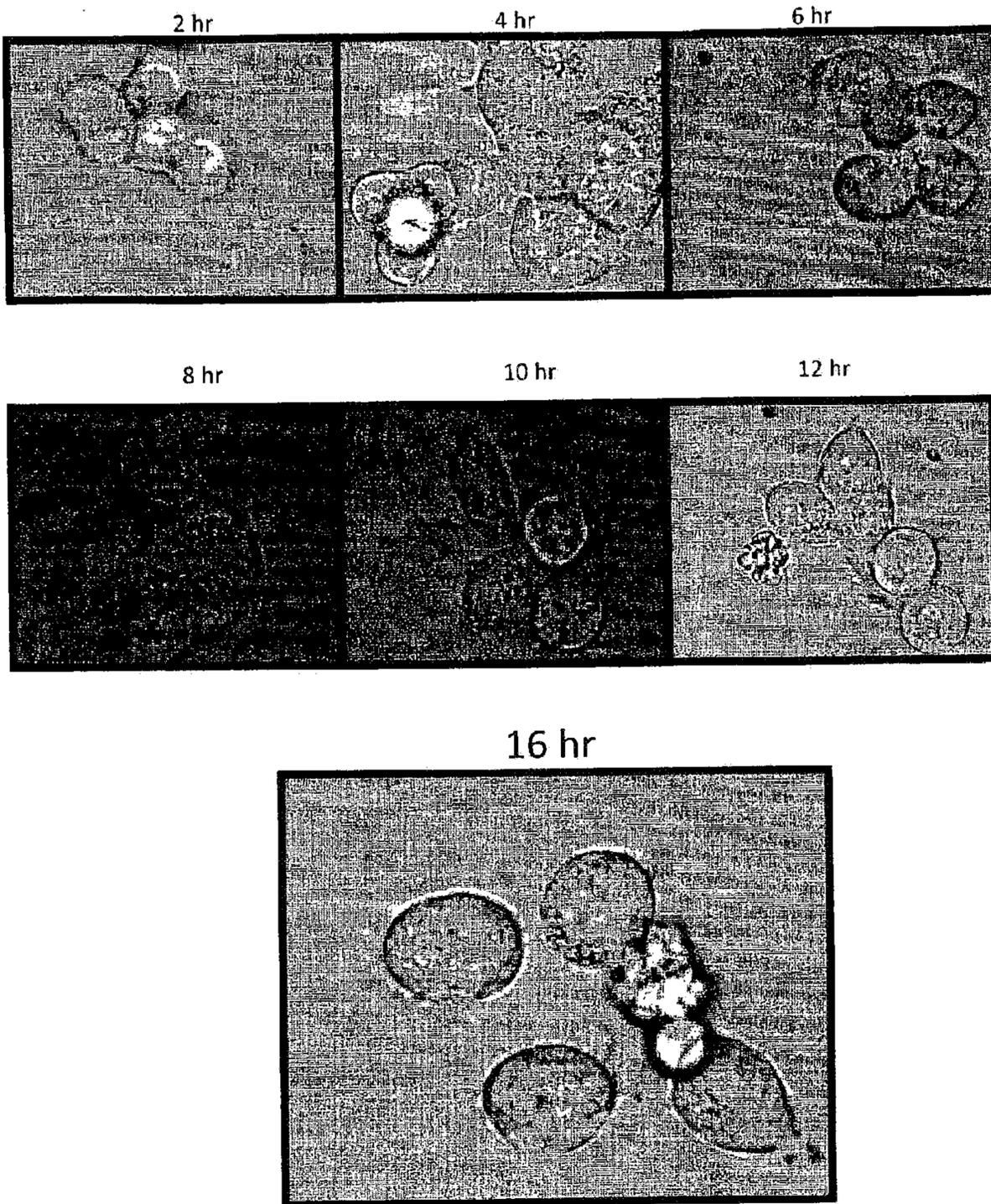


FIGURE 12

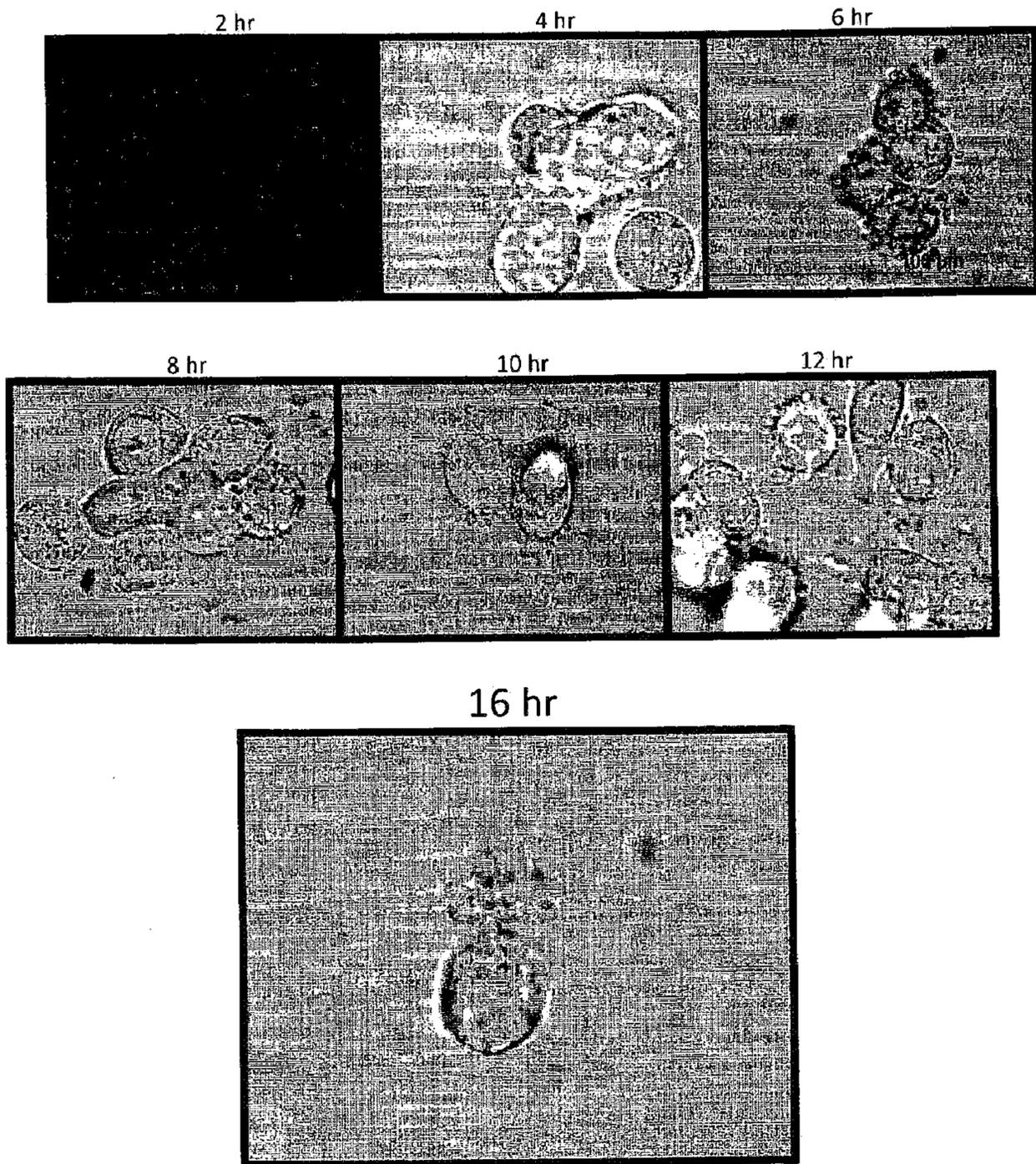


FIGURE 13

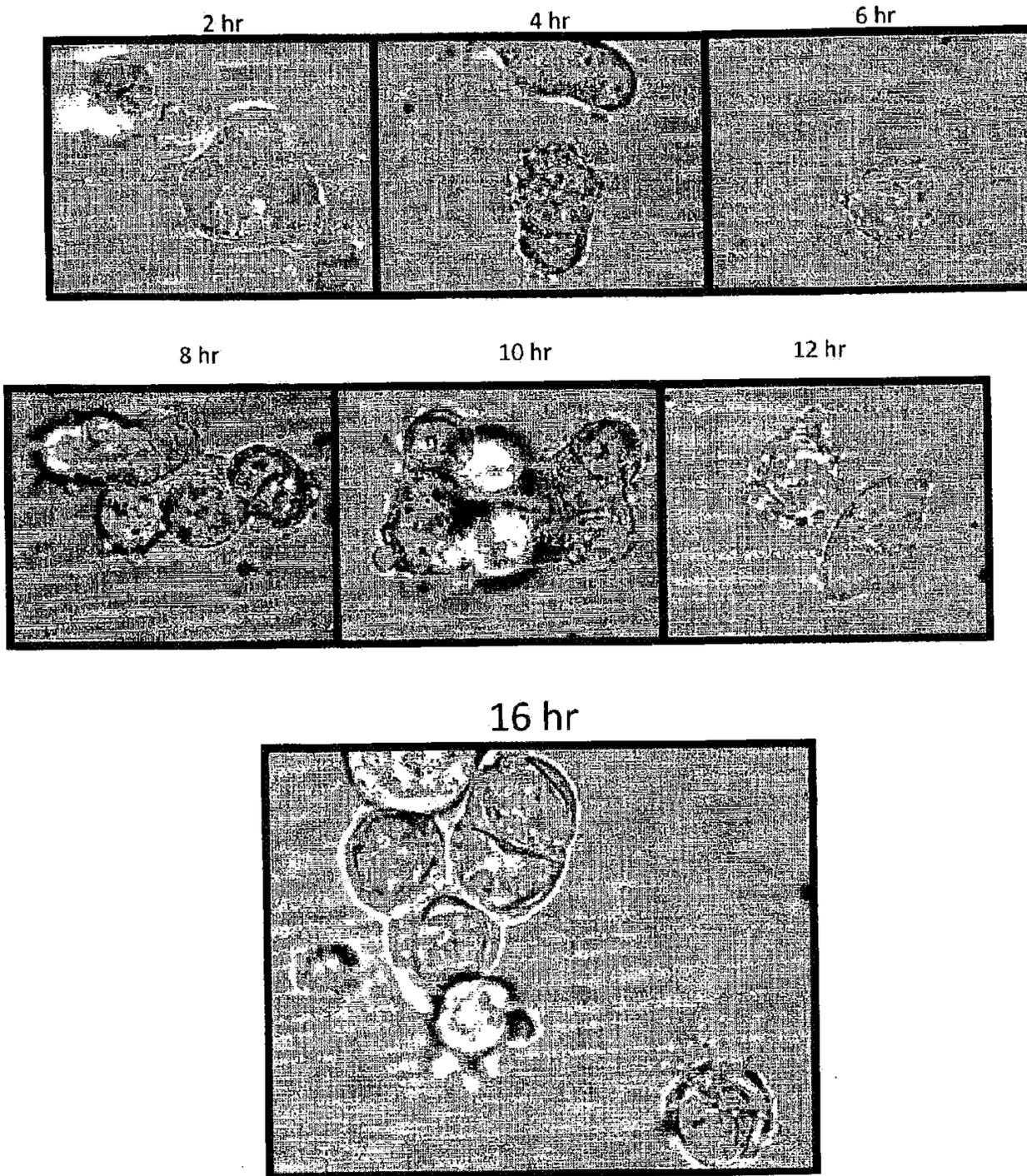


FIGURE 14

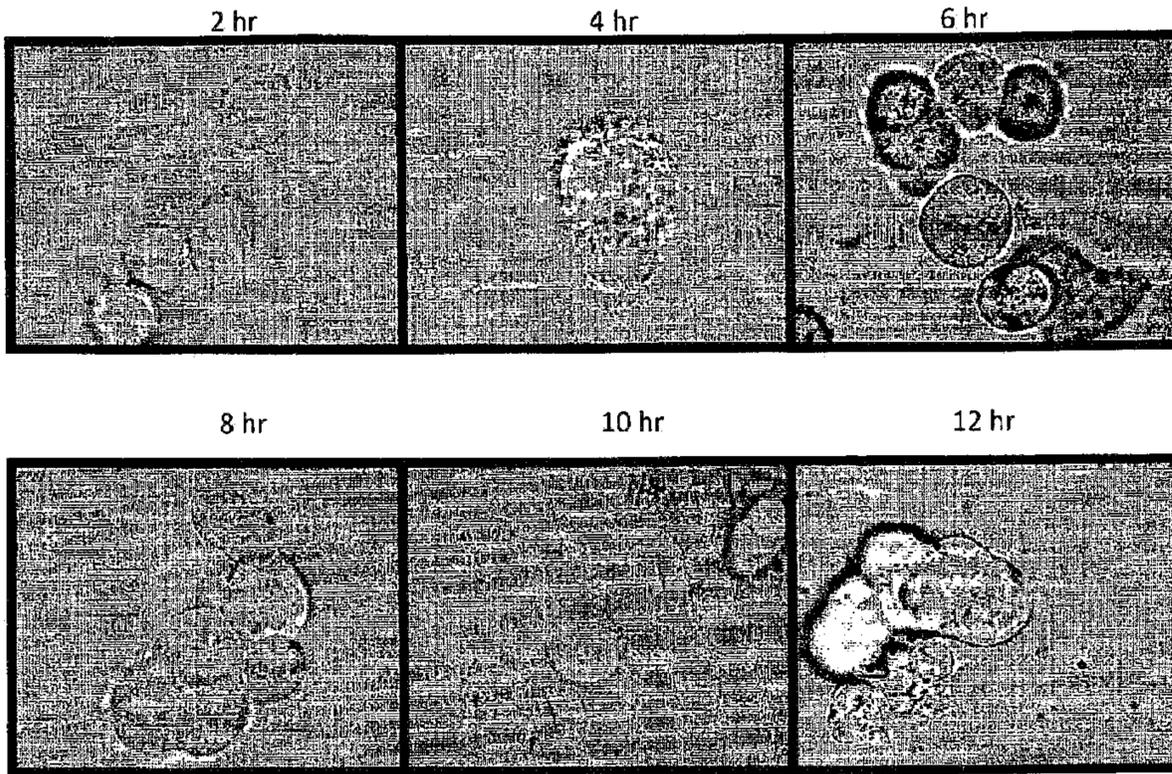


FIGURE 15