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(54) Title: METHOD FOR TREATING DISEASES USING HSP90-INHIBITING AGENTS IN COMBINATION WITH ENZYME INHIBITORS

(57) Abstract: The present invention provides a method for treating cancer. The method involves the administration of an HSP90 inhibitor and an enzyme inhibitor, where the combined administration provides a synergistic effect. In one aspect of the invention, a method of treating cancer is provided where a subject is treated with a dose of an HSP90 inhibitor in one step and a dose of an enzyme inhibitor in another step. In another aspect of the invention, a method of treating cancer is provided where a subject is first treated with a dose of an HSP90 inhibitor and subsequently treated with a dose of an enzyme inhibitor. In another aspect of the invention, a method of treating cancer is provided where a subject is first treated with a dose of an enzyme inhibitor and subsequently treated with a dose of an HSP90 inhibitor.

## METHOD FOR TREATING DISEASES USING HSP90-INHIBITING AGENTS IN COMBINATION WITH ENZYME INHIBITORS

### CROSS REFERENCE TO RELATED U.S. PATENT APPLICATIONS

5 [0001] The present application claims the benefit of Provisional Patent Application No. 60/474,906, which was filed May 30, 2003, under 35 U.S.C. § 119(e). The provisional application is hereby incorporated-by-reference into this application for all purposes.

### 10 BACKGROUND OF THE INVENTION

#### *Field of the Invention*

[0002] This invention relates to methods for treating cancer in which an inhibitor of Heat Shock Protein 90 (“HSP90”) is combined with an enzyme inhibitor. More particularly, this invention relates to combinations of the HSP90 inhibitor geldanamycin 15 and its derivatives, especially 17-alkylamino-17-desmethoxygeldanamycin (“17-AAG”) and 17-(2-dimethylaminoethyl)amino-17-desmethoxygeldanamycin (“17-DMAG”), with an enzyme inhibitor (*e.g.*, SAHA and Iressa).

#### *References*

- 20 [0003] Agnew *et al.*, “Clinical pharmacokinetics of 17-(allylamino)-17-demethoxy-geldanamycin and the active metabolite 17-(amino)-17-demethoxygeldanamycin given as a one-hour infusion daily for 5 days.” AACR, 2002.
- [0004] An *et al.*, “Depletion of p185erbB2, Raf-1 and mutant p53 proteins by geldanamycin derivatives correlates with antiproliferative activity.” *Cancer Chemother. Pharmacol.* 40:60-64, 1997.
- 25 [0005] Bagatell *et al.*, “Induction of a heat shock factor 1-dependent stress response alters the cytotoxic activity of hsp90-binding agents.” *Clin. Cancer Res.* 6:3312-3318, 2000.
- [0006] Bagatell *et al.*, “Destabilization of steroid receptors by heat shock protein 90-binding drugs: a ligand-independent approach to hormonal therapy of breast cancer.” *Clin. Cancer Res.* 7:2076-2084, 2001.

- [0007] Banerji *et al.*, "A pharmacokinetically (PK)-pharmacodynamically (PD) driven Phase I trial of the HSP90 molecular chaperone inhibitor 17-allylamino-17-demethoxygeldanamycin (17-AAG)." AACR, 2002.
- 5 [0008] Barent *et al.*, "Analysis of FKBP51/FKBP52 chimeras and mutants for Hsp90 binding and association with progesterone receptor complexes." *Mol. Endocrinol.* 12:342-354, 1998.
- [0009] Bilodeau *et al.*, "Tyrosine kinase inhibitors." U.S. Patent No. 6,245,759 issued June 12, 2001.
- 10 [0010] Citri *et al.*, "Drug-induced ubiquitylation and degradation of ErbB receptor tyrosine dinases: implications for cancer chemotherapy." *EMBO Journal* 21:2407-2417, 2002.
- [0011] Egorin *et al.*, "Metabolism of 17-(allylamino)-17-demethoxygeldanamycin (NSC 330507) by murine and human hepatic preparations." *Cancer Res.* 58:2385-2396, 1998.
- 15 [0012] Fraley *et al.*, "Tyrosine kinase inhibitors." U.S. Patent No. 6,306,874 issued October 23, 2001.
- [0013] Fraley *et al.*, "Tyrosine kinase inhibitors." U.S. Patent No. 6,313,138 issued November 6, 2001.
- 20 [0014] Gaidigk *et al.*, "NAD(P)H:quinone oxidoreductase: polymorphisms and allele frequencies n Caucasian, Chinese and Canadian Native Indian and Inuit populations." *Pharmacogenetics* 8:305-313, 1998.
- [0015] Gelmon *et al.*, "Anticancer agents targeting signaling molecules and cancer cell environment: challenges for drug development?" *J. Natl. Cancer Inst.* 91:1281-1287, 1999.
- 25 [0016] Goetz *et al.*, "The Hsp90 chaperone complex as a novel target for cancer therapy." *Ann. Oncol.* 14:1169-1176, 2003.
- [0017] Goh *et al.*, "Explaining interindividual variability of docetaxel pharmacokinetics and pharmacodynamics in Asians through phenotyping and genotyping strategies." *J. Clin. Oncol.* 20:3683-3690, 2002.
- 30 [0018] Grenert *et al.*, "The amino-terminal domain of heat shock protein 90 (hsp90) that binds geldanamycin is an ATP/ADP switch domain that regulates hsp90 conformation." *J. Biol. Chem.* 272:23843-23850, 1997.

- [0019] Johnson and Toft, "Binding of p23 and hsp90 during assembly with the progesterone receptor." *Mol. Endocrinol.* **9**:670-678, 1995.
- [0020] Hartl and Hayer-Hartl, "Molecular chaperones in the cytosol: from nascent chain to folded protein." *Science* **195**:1852-1858, 2002.
- 5 [0021] Hegde *et al.*, "Short circuiting stress protein expression via a tyrosine kinase inhibitor, herbimycin A." *J. Cell Physiol.* **165**:186-200, 1995.
- [0022] Hustert *et al.*, "The genetic determinants of the CYP3A5 polymorphism." *Pharmacogenetics* **11**:773-779, 2001.
- [0023] Kelland *et al.*, "DT-Diaphorase expression and tumor cell sensitivity to 17-allylamino, 17-demethoxygeldanamycin, an inhibitor of heat shock protein 90." *J. Natl. Cancer Inst.* **91**:1940-1949, 1999.
- [0024] Kuehl *et al.*, "Sequence diversity in CYP3A promoters and characterization of the genetic basis of polymorphic CYP3A5 expression." *Nat. Genet.* **27**:383-391, 2001.
- 10 [0025] Lawson *et al.*, "Geldanamycin, an hsp90/GRP94-binding drug, induces increased transcription of endoplasmic reticulum (ER) chaperones via the ER stress pathway." *J. Cell Physiol.* **174**:170-178, 1998.
- [0026] Lin *et al.*, "Co-regulation of CYP3A4 and CYP3A5 and contribution to hepatic and intestinal midazolam metabolism." *Mol. Pharmacol.* **62**:162-172, 2002.
- 15 [0027] Morimoto *et al.*, "The heat-shock response: regulation and function of heat-shock proteins and molecular chaperones." *Essays Biochem.* **32**:17-29, 1997.
- [0028] Munster *et al.*, "Phase I trial of 17-(allylamino)-17-demethoxygeldanamycin (17-AAG) in patients with advanced solid malignancies." *Proc. Am. Soc. Clin. Oncol.*, 83a, 2001.
- 20 [0029] Munster *et al.*, "Modulation of Hsp90 function by ansamycins sensitizes breast cancer cells to chemotherapy-induced apoptosis in an RB- and schedule-dependent manner." *Clin. Cancer Res.* **7**:2228-2236, 2001.
- [0030] Murakami *et al.*, "Induction of hsp 72/73 by herbimycin A, an inhibitor of transformation by tyrosine kinase oncogenes." *Exp. Cell Res.* **195**:338-344, 1991.
- 25 [0031] Pratt and Toft, "Steroid receptor interactions with heat shock protein and immunophilin chaperones." *Endocr. Rev.* **18**:306-60, 1997.
- [0032] Prodromou *et al.*, "Identification and structural characterization of the ATP/ADP-binding site in the Hsp90 molecular chaperone." *Cell* **90**:65-75, 1997.

[0033] Richter and Buchner, "Hsp90: chaperoning signal transduction." *J. Cell. Physiol.* **188**:281-290, **2001**.

[0034] Rosvold *et al.*, "Identification of an NAD(P)H:quinone oxidoreductase polymorphism and its association with lung cancer and smoking." *Pharmacogenetics* **5**:199-206, **1995**.

[0035] Schneider *et al.*, "Pharmacologic shifting of a balance between protein refolding and degradation mediated by Hsp90." *Proc. Natl. Acad. Sci. USA* **93**:14536-14541, **1996**.

[0036] Schnur *et al.*, "erbB-2 oncogene inhibition by geldanamycin derivatives: synthesis, mechanism of action, and structure-activity relationships." *J. Med. Chem.* **38**:3813-20, **1995**.

[0037] Schnur *et al.*, "Inhibition of the oncogene product p185erbB-2 in vitro and in vivo by geldanamycin and dihydrogeldanamycin derivatives." *J. Med. Chem.* **38**:3806-3812, **1995**.

[0038] Smith *et al.*, "Progesterone receptor structure and function altered by geldanamycin, an hsp90-binding agent." *Mol. Cell Biol.* **15**:6804-6812, **1995**.

[0039] Smith *et al.*, "Identification of a 60-kilodalton stress-related protein, p60, which interacts with hsp90 and hsp70." *Mol. Cell Biol.* **13**:869-876, **1993**.

[0040] Stebbins *et al.*, "Crystal structure of an Hsp90-geldanamycin complex: targeting of a protein chaperone by an antitumor agent." *Cell* **89**:239-250, **1997**.

[0041] Traver *et al.*, "NAD(P)H:quinone oxidoreductase gene expression in human colon carcinoma cells: characterization of a mutation which modulates DT-diaphorase activity and mitomycin sensitivity." *Cancer Res.* **52**:797-802, **1992**.

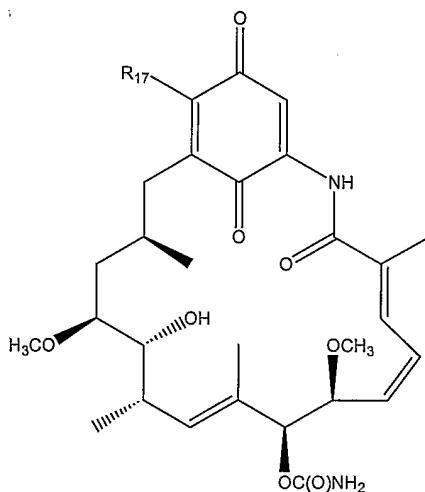
[0042] Whitesell *et al.*, "Inhibition of heat shock protein HSP90-pp60v-src heteroprotein complex formation by benzoquinone ansamycins: essential role for stress proteins in oncogenic transformation." *Proc. Natl. Acad. Sci. USA* **91**:8324-8328, **1994**.

[0043] Young *et al.*, "Hsp90: a specialized but essential protein-folding tool." *J. Cell Biol.* **154**:267-273, **2001**.

[0044] Zou *et al.*, "Repression of heat shock transcription factor HSF1 activation by HSP90 (HSP90 complex) that forms a stress-sensitive complex with HSF1." *Cell* **94**:471-480, **1998**.

*Discussion*

- [0045] Geldanamycin (figure below,  $R_{17} = -OCH_3$ ) is a benzoquinone ansamycin polyketide isolated from *Streptomyces geldanus*. Although originally discovered by screening microbial extracts for antibacterial and antiviral activity, geldanamycin was 5 later found to be cytotoxic to certain tumor cells *in vitro* and to reverse the morphology of cells transformed by the Rous sarcoma virus to a normal state.



Geldanamycin's nanomolar potency and apparent specificity for aberrant protein kinase 10 dependent tumor cells, as well as the discovery that its primary target in mammalian cells is the ubiquitous Hsp90 protein chaperone, has stimulated interest in the development of this compound as an anti-cancer drug. However, the association of unacceptable hepatotoxicity with the administration of geldanamycin led to its withdrawal from Phase I clinical trials.

[0046] More recently, attention has focused on 17-amino derivatives of 15 geldanamycin, in particular 17-(allylamino)-17-desmethoxygeldanamycin ("17-AAG",  $R_{17} = -NCH_2CH=CH_2$ ). This compound has reduced hepatotoxicity while maintaining useful Hsp90 binding. Certain other 17-amino derivatives of geldanamycin, 11-oxogeldanamycin, and 5,6-dihydrogeldanamycin, are disclosed in U.S. patents 4,261,989, 5,387,584 and 5,932,566, each of which is incorporated herein by reference. Treatment 20 of cancer cells with geldanamycin or 17-AAG causes a retinoblastoma protein-dependent G1 block, mediated by down-regulation of the induction pathways for cyclin D-cyclin dependent cdk4 and cdk6 protein kinase activity. Cell cycle arrest is followed by

differentiation and apoptosis. G1 progression is unaffected by geldanamycin or 17-AAG in cells with mutated retinoblastoma protein; these cells undergo cell cycle arrest after mitosis, again followed by apoptosis.

[0047] The above-described mechanism of geldanamycin and 17-AAG appears to be 5 a common mode of action among the benzoquinone ansamycins that further includes binding to Hsp90 and subsequent degradation of Hsp90-associated client proteins. Among the most sensitive client protein targets of the benzoquinone ansamycins are the Her kinases (also known as ErbB), Raf, Met tyrosine kinase, and the steroid receptors. Hsp90 is also involved in the cellular response to stress, including heat, radiation, and 10 toxins. Certain benzoquinone ansamycins, such as 17-AAG, have thus been studied to determine their interaction with cytotoxins that do not target Hsp90 client proteins.

[0048] U.S. Patents 6,245,759, 6,306,874 and 6,313,138, each of which is 15 incorporated herein by reference, disclose compositions comprising certain tyrosine kinase inhibitors together with 17-AAG and methods for treating cancer with such compositions. Münster, et al., "Modulation of Hsp90 function by ansamycins sensitizes breast cancer cells to chemotherapy-induced apoptosis in an RB- and schedule-dependent manner," Clinical Cancer Research (2001) 7:2228-2236, discloses that 17-AAG sensitizes cells in culture to the cytotoxic effects of Paclitaxel and doxorubicin. The Münster reference further discloses that the sensitization towards paclitaxel by 17-AAG is 20 schedule-dependent in retinoblastoma protein-producing cells due to the action of these two drugs at different stages of the cell cycle: treatment of cells with a combination of paclitaxel and 17-AAG is reported to give synergistic apoptosis, while pretreatment of cells with 17-AAG followed by treatment with paclitaxel is reported to result in abrogation of apoptosis. Treatment of cells with paclitaxel followed by treatment with 25 17-AAG 4 hours later is reported to show a synergistic effect similar to coincident treatment.

[0049] Citri, et al., "Drug-induced ubiquitylation and degradation of ErbB receptor tyrosine kinases: implications for cancer chemotherapy," EMBO Journal (2002) 30 21:2407-2417, discloses an additive effect upon co-administration of geldanamycin and an irreversible protein kinase inhibitor, CI-1033, on growth of ErbB2-expressing cancer cells in vitro. In contrast, an antagonistic effect of CI-1033 and anti-ErbB2 antibody, Herceptin is disclosed.

[0050] Thus, while there has been a great deal of research interest in the benzoquinone ansamycins, particularly geldanamycin and 17-AAG, there remains a need for effective therapeutic regimens to treat cancer or other disease conditions characterized by undesired cellular hyperproliferation using such compounds, whether alone or in 5 combination with other agents.

#### SUMMARY OF THE INVENTION

[0051] The present invention provides a method for treating cancer. The method involves the administration of an HSP90 inhibitor and an enzyme inhibitor, where the 10 combined administration provides a synergistic effect.

[0052] In one aspect of the invention, a method of treating cancer is provided where a subject is treated with a dose of an HSP90 inhibitor in one step and a dose of an enzyme inhibitor in another step.

[0053] In another aspect of the invention, a method of treating cancer is provided 15 where a subject is first treated with a dose of an HSP90 inhibitor and subsequently treated with a dose of an enzyme inhibitor.

[0054] In another aspect of the invention, a method of treating cancer is provided where a subject is first treated with a dose of an enzyme inhibitor and subsequently treated with a dose of an HSP90 inhibitor.

[0055] In another aspect of the invention, a method of treating cancer is provided 20 where a subject is first treated with a dose of an enzyme inhibitor (e.g., SAHA or Iressa). After waiting for a period of time sufficient to allow development of a substantially efficacious response of the enzyme inhibitor, a formulation comprising a synergistic dose of a benzoquinone ansamycin together with a second sub-toxic dose of the enzyme inhibitor is administered.

[0056] In another aspect of the invention, a method of treating cancer is provided 30 where a subject is treated first with a dose of a benzoquinone ansamycin, and second, a dose of an enzyme inhibitor. After waiting for a period of time sufficient to allow development of a substantially efficacious response of the enzyme inhibitor, a formulation comprising a synergistic dose of a benzoquinone ansamycin together with a second sub-toxic dose of the enzyme inhibitor drug is administered.

[0057] In another aspect of the invention, a method for treating cancer is provided where a subject is treated with a dose of an HSP90 inhibitor in one step and a dose of an

enzyme inhibitor in another step, and where a side effect profile for the combined, administered drugs is substantially better than for the enzyme inhibitor alone.

[0058] In another aspect of the invention, a method for treating breast or colorectal cancer is provided where a subject is treated with a dose of an HSP90 inhibitor in one step and a dose of an enzyme inhibitor in another step. The HSP90 inhibitor for this aspect is typically 17-AAG, while the enzyme inhibitor is usually SAHA or Iressa. For the treatment of colorectal cancer or breast cancer, the enzyme inhibitor is typically administered before the 17-AAG.

10 Definitions

[0059] “Enzyme inhibitor” refers to a drug that stops, prevents or reduces the activity of an enzyme or a prodrug thereof. Enzyme inhibitors include, without limitation, histone deacetylation inhibitors (*e.g.*, SAHA) and tyrosine kinase inhibitors (*e.g.*, Iressa).

[0060] “HSP90 inhibitor” refers to a compound that inhibits the activity of heat shock

15 protein 90, which is a cellular protein responsible for chaperoning multiple client proteins necessary for cell signaling, proliferation and survival. One class of HSP90 inhibitors is the benzoquinone ansamycins. Examples of such compounds include, without limitation, geldanamycin and geldanamycin derivatives (*e.g.*, 17-alkylamino-17-desmethoxy-

geldanamycin (“17-AAG”) and 17-(2-dimethylaminoethyl)amino-17-desmethoxy-

20 geldanamycin (“17-DMAG”). *See* Sasaki *et al.*, US 4,261,989 (1981) for synthesis of 17-

AAG and Snader *et al.*, US 2004/0053909 A1 (2004) for synthesis of 17-DMAG. In

addition to 17-AAG and 17-DMAG, other preferred geldanamycin derivatives are 11-*O*-

25 methyl-17-(2-(1-azetidinyl)ethyl)amino-17-demethoxygeldanamycin (A), 11-*O*-methyl-

17-(2-dimethylaminoethyl)amino-17-demethoxygeldanamycin (B), and 11-*O*-methyl-17-

25 (2-(1-pyrrolidinyl)ethyl)amino-17-demethoxygeldanamycin (C), whose synthesis is

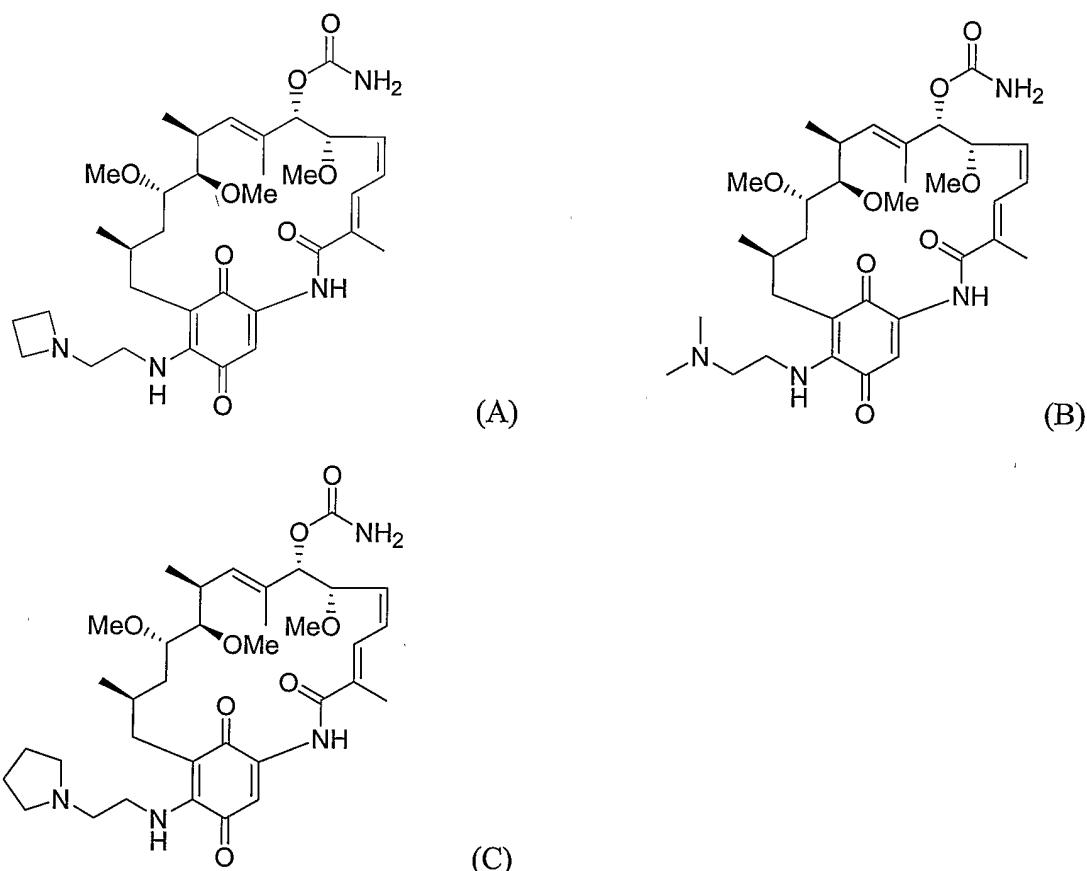
described in the co-pending commonly US patent application of Tian *et al.*, serial no.

10/825,788, filed Apr. 16, 2004, and in Tian *et al.*, PCT application no. PCT/US04/11638,

filed Apr. 16, 2004; the disclosures of which are incorporated herein by reference.

Additional preferred geldanamycin derivatives are described in Santi *et al.*, US

30 2003/0114450 A1 (2003), also incorporated by reference.



5 [0061] “MTD” refers to maximum tolerated dose. The MTD for a compound is determined using methods and materials known in the medical and pharmacological arts, for example through dose-escalation experiments. One or more patients is first treated with a low dose of the compound, typically about 10% of the dose anticipated to be therapeutic based on results of in vitro cell culture experiments. The patients are  
10 observed for a period of time to determine the occurrence of toxicity. Toxicity is typically evidenced as the observation of one or more of the following symptoms: vomiting, diarrhea, peripheral neuropathy, ataxia, neutropenia, or elevation of liver enzymes. If no toxicity is observed, the dose is increased about 2-fold, and the patients are again observed for evidence of toxicity. This cycle is repeated until a dose producing  
15 evidence of toxicity is reached. The dose immediately preceding the onset of unacceptable toxicity is taken as the MTD.

[0062] "Side effects" refer to a number of toxicities typically seen upon treatment of a subject with an antineoplastic drug. Such toxicities include, without limitation, anemia, anorexia, bilirubin effects, dehydration, dermatology effects, diarrhea, dizziness, dyspnea, edema, fatigue, headache, hematemesis, hypokalemia, hypoxia, musculoskeletal effects,

myalgia, nausea, neuro-sensory effects, pain, rash, serum glutamic oxaloacetic transaminase effects, serum glutamic pyruvic transaminase effects, stomatitis, sweating, taste effects, thrombocytopenia, voice change, and vomiting.

5 [0063] “Side effect grading” refers to National Cancer Institute common toxicity criteria (NCI CTC, Version 2). Grading runs from 1 to 4, with a grade of 4 representing the most serious toxicities.

#### Combination Therapy

10 [0064] The present invention provides a method for treating cancer. The method involves the administration of an HSP90 inhibitor and an enzyme inhibitor, where the combined administration provides a synergistic effect.

15 [0065] Suitable HSP90 inhibitors used in the present invention include benzoquinone ansamycins. Typically, the benzoquinone ansamycin is geldanamycin or a geldanamycin derivative. Preferably, the benzoquinone ansamycin is a geldanamycin derivative selected from a group consisting of 17-alkylamino-17-desmethoxy-geldanamycin (“17-AAG”) and 17-(2-dimethylaminoethyl)amino-17-desmethoxy-geldanamycin (“17-DMAG”).

20 [0066] Enzyme inhibitors employed in the present method include, without limitation, histone deacetylation inhibitors (*e.g.*, SAHA) and tyrosine kinase inhibitors (*e.g.*, Iressa).

25 [0067] The dose of enzyme inhibitor used as a partner in combination therapy with an HSP90 inhibitor (*e.g.*, benzoquinone ansamycin) is determined based on the maximum tolerated dose observed when the enzyme inhibitor is used as the sole therapeutic agent. In one embodiment of the invention, the dose of enzyme inhibitor when used in combination therapy with a benzoquinone ansamycin is the MTD. In other embodiments of the invention, the dose of enzyme inhibitor when used in combination therapy with a benzoquinone ansamycin is between about 1% of the MTD and the MTD, between about 5% of the MTD and the MTD, between about 5% of the MTD and 75% of the MTD, or between about 25% of the MTD and 75% of the MTD.

30 [0068] Use of the benzoquinone ansamycin allows for use of a lower therapeutic dose of an enzyme inhibitor, thus significantly widening the therapeutic window for treatment. In one embodiment, the therapeutic dose of enzyme inhibitor is lowered by at least about 10%. In other embodiments the therapeutic dose is lowered from about 10 % to 20%, from about 20% to 50%, from about 50% to 200%, or from about 100% to 1,000%.

[0069] For the treatment of a variety of carcinomas, the typical oral dose of various enzyme inhibitors is as follows: SAHA—up to 500 mg/day; Iressa—250 mg/day.

[0070] The synergistic dose of the benzoquinone ansamycin used in combination therapy is determined based on the maximum tolerated dose observed when the

5 benzoquinone ansamycin is used as the sole therapeutic agent. Clinical trials have determined an MTD for 17-AAG of about 40 mg/m<sup>2</sup> utilizing a daily x 5 schedule, and MTD of about 220 mg/m<sup>2</sup> utilizing a twice-weekly regimen, and an MTD of about 308 mg/m<sup>2</sup> utilizing a once-weekly regimen. In one embodiment of the invention, the dose of the benzoquinone ansamycin when used in combination therapy is the MTD. In other 10 embodiments of the invention, the dose of the benzoquinone ansamycin when used in combination therapy is between about 1% of the MTD and the MTD, between about 5% of the MTD and the MTD, between about 5% of the MTD and 75% of the MTD, or between about 25% of the MTD and 75% of the MTD.

15 [0071] Where the benzoquinone ansamycin is 17-AAG, and the administration of compound is weekly, its therapeutic dose is typically between 50 mg/m<sup>2</sup> and 450 mg/m<sup>2</sup>. Preferably, the dose is between 150 mg/m<sup>2</sup> and 350 mg/m<sup>2</sup>, and about 308 mg/m<sup>2</sup> is especially preferred. Where the administration of compound is biweekly (*i.e.*, twice per week), the therapeutic dose of 17-AAG is typically between 50 mg/m<sup>2</sup> and 250 mg/m<sup>2</sup>. Preferably, the dose is between 150 mg/m<sup>2</sup> and 250 mg/m<sup>2</sup>, and about 220 mg/m<sup>2</sup> is 20 especially preferred.

[0072] Where the present method involves the administration of 17-AAG and SAHA, a dosage regimen involving one or more administration of the combination per week is typical. Oftentimes, the combination is administered 2, 3, 4, 5, 6, or 7 times per week. Tables 1 and 2 below show a number of SAHA/17-AAG dosage combinations (*i.e.*, dosage combinations 0001 to 0080).

**Table 1: SAHA/17-AAG dosage combinations.**

	30-100 mg/m <sup>2</sup> 17-AAG	100-150 mg/m <sup>2</sup> 17-AAG	150-200 mg/m <sup>2</sup> 17-AAG	200-250 mg/m <sup>2</sup> 17-AAG
<b>0-50 mg/day SAHA</b>	0001	0002	0003	0004
<b>50-100 mg/day SAHA</b>	0005	0006	0007	0008
<b>100-150 mg/day SAHA</b>	0009	0010	0011	0012
<b>150-200 mg/day SAHA</b>	0013	0014	0015	0016
<b>200-250 mg/day SAHA</b>	0017	0018	0019	0020
<b>250-300 mg/day SAHA</b>	0021	0022	0023	0024
<b>300-350 mg/day SAHA</b>	0025	0026	0027	0028
<b>350-400 mg/day SAHA</b>	0029	0030	0031	0032
<b>400-450 mg/day SAHA</b>	0033	0034	0035	0036
<b>450-500 mg/day SAHA</b>	0037	0038	0039	0040

**Table 2: SAHA/17-AAG dosage combinations continued.**

	250-300 mg/m <sup>2</sup> 17-AAG	300-350 mg/m <sup>2</sup> 17-AAG	350-400 mg/m <sup>2</sup> 17-AAG	400-450 mg/m <sup>2</sup> 17-AAG
<b>0-50 mg/day SAHA</b>	0041	0042	0043	0044
<b>50-100 mg/day SAHA</b>	0045	0046	0047	0048
<b>100-150 mg/day SAHA</b>	0049	0050	0051	0052
<b>150-200 mg/day SAHA</b>	0053	0054	0055	0056
<b>200-250 mg/day SAHA</b>	0057	0058	0059	0060
<b>250-300 mg/day SAHA</b>	0061	0062	0063	0064
<b>300-350 mg/day SAHA</b>	0065	0066	0067	0068
<b>350-400 mg/day SAHA</b>	0069	0070	0071	0072
<b>400-450 mg/day SAHA</b>	0073	0074	0075	0076
<b>450-500 mg/day SAHA</b>	0077	0078	0079	0080

[0073] Where the present method involves the administration of 17-AAG and Iressa, a dosage regimen involving one or more administration of the combination per week is typical. Oftentimes, the combination is administered 2, 3, 4, 5, 6, or 7 times per week. Tables 3 and 4 below show a number of Iressa/17-AAG dosage combinations (*i.e.*, dosage combinations 0081 to 0160).

**Table 3: Iressa/17-AAG dosage combinations.**

	30-100 mg/m <sup>2</sup> 17-AAG	100-150 mg/m <sup>2</sup> 17-AAG	150-200 mg/m <sup>2</sup> 17-AAG	200-250 mg/m <sup>2</sup> 17-AAG
<b>0-25 mg/day Iressa</b>	0081	0082	0083	0084
<b>25-50 mg/day Iressa</b>	0085	0086	0087	0088
<b>50-75 mg/day Iressa</b>	0089	0090	0091	0092
<b>75-100 mg/day Iressa</b>	0093	0094	0095	0096
<b>100-125 mg/day Iressa</b>	0097	0098	0099	0100
<b>125-150 mg/day Iressa</b>	0101	0102	0103	0104
<b>150-175 mg/day Iressa</b>	0105	0106	0107	0108
<b>175-200 mg/day Iressa</b>	0109	0110	0111	0112
<b>200-225 mg/day Iressa</b>	0113	0114	0115	0116
<b>225-250 mg/day Iressa</b>	0117	0118	0119	0120

**Table 4: Iressa/17-AAG dosage combinations continued.**

	250-300 mg/m <sup>2</sup> 17-AAG	300-350 mg/m <sup>2</sup> 17-AAG	350-400 mg/m <sup>2</sup> 17-AAG	400-450 mg/m <sup>2</sup> 17-AAG
<b>0-25 mg/day Iressa</b>	0121	0122	0123	0124
<b>25-50 mg/day Iressa</b>	0125	0126	0127	0128
<b>50-75 mg/day Iressa</b>	0129	0130	0131	0132
<b>75-100 mg/day Iressa</b>	0133	0134	0135	0136
<b>100-125 mg/day Iressa</b>	0137	0138	0139	0140
<b>125-150 mg/day Iressa</b>	0141	0142	0143	0144
<b>150-175 mg/day Iressa</b>	0145	0146	0147	0148
<b>175-200 mg/day Iressa</b>	0149	0150	0151	0152
<b>200-225 mg/day Iressa</b>	0153	0154	0155	0156
<b>225-250 mg/day Iressa</b>	0157	0158	0159	0160

[0074] The method of the present invention may be carried out in at least two basic ways. A subject may first be treated with a dose on an HSP90 inhibitor and subsequently be treated with a dose of an enzyme inhibitor. Alternatively, the subject may first be treated with a dose of an enzyme inhibitor and subsequently be treated with a dose of an HSP90 inhibitor. The appropriate dosing regimen depends on the particular enzyme inhibitor employed.

[0075] In another aspect of the invention, a subject is first treated with a dose of an enzyme inhibitor (e.g., SAHA or Iressa). After waiting for a period of time sufficient to allow development of a substantially efficacious response of the enzyme inhibitor, a formulation comprising a synergistic dose of a benzoquinone ansamycin together with a second sub-toxic dose of the enzyme inhibitor is administered. In general, the appropriate period of time sufficient to allow development of a substantially efficacious response to the enzyme inhibitor will depend upon the pharmacokinetics of the enzyme inhibitor, and will have been determined during clinical trials of therapy using the enzyme inhibitor alone. In one embodiment of the invention, the period of time sufficient to allow development of a substantially efficacious response to the enzyme inhibitor is between about 1 hour and 96 hours. In another aspect of the invention, the period of time sufficient to allow development of a substantially efficacious response to the enzyme

inhibitor is between about 2 hours and 48 hours. In another embodiment of the invention, the period of time sufficient to allow development of a substantially efficacious response to the enzyme inhibitor is between about 4 hours and 24 hours.

- 5 [0076] In another aspect of the invention, a subject is treated first with one of the above-described benzoquinone ansamycins, and second, a dose of an enzyme inhibitor, such as, but not limited to, SAHA and Iressa. After waiting for a period of time sufficient to allow development of a substantially efficacious response of the enzyme inhibitor, a formulation comprising a synergistic dose of a benzoquinone ansamycin together with a second sub-toxic dose of the enzyme inhibitor is administered. In general, the appropriate 10 period of time sufficient to allow development of a substantially efficacious response to the enzyme inhibitor will depend upon the pharmacokinetics of the enzyme inhibitor, and will have been determined during clinical trials of therapy using the enzyme inhibitor alone. In one embodiment of the invention, the period of time sufficient to allow development of a substantially efficacious response to the enzyme inhibitor is between 15 about 1 hour and 96 hours. In another aspect of the invention, the period of time sufficient to allow development of a substantially efficacious response to the enzyme inhibitor is between about 2 hours and 48 hours. In another embodiment of the invention, the period of time sufficient to allow development of a substantially efficacious response to the enzyme inhibitor is between about 4 hours and 24 hours.
- 20 [0077] As noted above, the combination of an HSP90 inhibitor and an enzyme inhibitor allows for the use of a lower therapeutic dose of the enzyme inhibitor for the treatment of cancer. That a lower dose of enzyme inhibitor is used oftentimes lessens the side effects observed in a subject. The lessened side effects can be measured both in terms of incidence and severity. Severity measures are provided through a grading 25 process delineated by the National Cancer Institute (common toxicity criteria NCI CTC, Version 2). For instance, the incidence of side effects are typically reduced 10%. Oftentimes, the incidence is reduced 20%, 30%, 40% or 50%. Furthermore, the incidence of grade 3 or 4 toxicities for more common side effects associated with enzyme inhibitor administration (*e.g.*, anemia, anorexia, diarrhea, fatigue, nausea and vomiting) is oftentimes reduced 10%, 20%, 30%, 40% or 50%.

30 [0078] Formulations used in the present invention may be in any suitable form, such as a solid, semisolid, or liquid form. See *Pharmaceutical Dosage Forms and Drug Delivery Systems*, 5<sup>th</sup> edition, Lippincott Williams & Wilkins (1991), incorporated herein

by reference. In general the pharmaceutical preparation will contain one or more of the compounds of the present invention as an active ingredient in admixture with an organic or inorganic carrier or excipient suitable for external, enteral, or parenteral application.

The active ingredient may be compounded, for example, with the usual non-toxic,

5 pharmaceutically acceptable carriers for tablets, pellets, capsules, suppositories, pessaries, solutions, emulsions, suspensions, and any other form suitable for use. The carriers that can be used include water, glucose, lactose, gum acacia, gelatin, mannitol, starch paste,

magnesium trisilicate, talc, corn starch, keratin, colloidal silica, potato starch, urea, and other carriers suitable for use in manufacturing preparations in solid, semi-solid, or

10 liquefied form. In addition, auxiliary stabilizing, thickening, and coloring agents and perfumes may be used. Where applicable, the compounds useful in the methods of the invention may be formulated as microcapsules and nanoparticles. General protocols are

described, for example, by *Microcapsules and Nanoparticles in Medicine and Pharmacy* by Max Donbrow, ed., CRC Press (1992) and by U.S. Patent Nos. 5,510,118, 5,534,270

15 and 5,662,883 which are all incorporated herein by reference. By increasing the ratio of surface area to volume, these formulations allow for the oral delivery of compounds that would not otherwise be amenable to oral delivery. The compounds useful in the methods of the invention may also be formulated using other methods that have been previously used for low solubility drugs. For example, the compounds may form emulsions with

20 vitamin E or a PEGylated derivative thereof as described by PCT publications WO 98/30205 and WO 00/71163, each of which is incorporated herein by reference.

Typically, the compound useful in the methods of the invention is dissolved in an aqueous solution containing ethanol (preferably less than 1% w/v). Vitamin E or a

PEGylated-vitamin E is added. The ethanol is then removed to form a pre-emulsion that 25 can be formulated for intravenous or oral routes of administration. Another method involves encapsulating the compounds useful in the methods of the invention in liposomes. Methods for forming liposomes as drug delivery vehicles are well known in the art. Suitable protocols include those described by U.S. Patent Nos. 5,683,715,

5,415,869, and 5,424,073 which are incorporated herein by reference relating to another

30 relatively low solubility cancer drug paclitaxel and by PCT Publication WO 01/10412 which is incorporated herein by reference relating to epothilone B. Of the various lipids that may be used, particularly preferred lipids for making encapsulated liposomes include

phosphatidylcholine and polyethyleneglycol-derivatized distearyl phosphatidyl-ethanoloamine.

**[0079]** Yet another method involves formulating the compounds useful in the methods of the invention using polymers such as biopolymers or biocompatible (synthetic or naturally occurring) polymers. Biocompatible polymers can be categorized as biodegradable and non-biodegradable. Biodegradable polymers degrade in vivo as a function of chemical composition, method of manufacture, and implant structure.

Illustrative examples of synthetic polymers include polyanhydrides, polyhydroxyacids such as polylactic acid, polyglycolic acids and copolymers thereof, polyesters, polyamides, 10 polyorthoesters and some polyphosphazenes. Illustrative examples of naturally occurring polymers include proteins and polysaccharides such as collagen, hyaluronic acid, albumin, and gelatin.

**[0080]** Another method involves conjugating the compounds useful in the methods of the invention to a polymer that enhances aqueous solubility. Examples of suitable 15 polymers include polyethylene glycol, poly-(d-glutamic acid), poly-(l-glutamic acid), poly-(l-glutamic acid), poly-(d-aspartic acid), poly-(l-aspartic acid) and copolymers thereof. Polyglutamic acids having molecular weights between about 5,000 to about 100,000 are preferred, with molecular weights between about 20,000 and 80,000 being more preferred and with molecular weights between about 30,000 and 60,000 being 20 most preferred. The polymer is conjugated via an ester linkage to one or more hydroxyls of an inventive geldanamycin using a protocol as essentially described by U.S. Patent No. 5,977,163 which is incorporated herein by reference.

**[0081]** In another method, the compounds useful in the methods of the invention are conjugated to a monoclonal antibody. This method allows the targeting of the inventive 25 compounds to specific targets. General protocols for the design and use of conjugated antibodies are described in *Monoclonal Antibody-Based Therapy of Cancer* by Michael L. Grossbard, ED. (1998), which is incorporated herein by reference.

**[0082]** The amount of active ingredient that may be combined with the carrier materials to produce a single dosage form will vary depending upon the subject treated 30 and the particular mode of administration. For example, a formulation for intravenous use comprises an amount of the inventive compound ranging from about 1 mg/mL to about 25 mg/mL, preferably from about 5 mg/mL, and more preferably about 10 mg/mL.

Intravenous formulations are typically diluted between about 2 fold and about 30 fold with normal saline or 5% dextrose solution prior to use.

**[0083]** Preferably, 17-AAG is formulated as a pharmaceutical solution formulation comprising 17-AAG in an concentration of up to 15 mg/mL dissolved in a vehicle

5 comprising (i) a first component that is ethanol, in an amount of between about 40 and about 60 volume %; (ii) a second component that is a polyethoxylated castor oil, in an amount of between about 15 to about 50 volume %; and (iii) a third component that is selected from the group consisting of propylene glycol, PEG 300, PEG 400, glycerol, and combinations thereof, in an amount of between about 0 and about 35 volume %. The 10 aforesaid percentages are volume/volume percentages based on the combined volumes of the first, second, and third components. The lower limit of about 0 volume % for the third component means that it is an optional component; that is, it may be absent. The pharmaceutical solution formulation is then diluted into water to prepare a diluted formulation containing up to 3 mg/mL 17-AAG, for intravenous formulation.

15 **[0084]** Preferably, the second component is Cremophor EL and the third component is propylene glycol. In an especially preferred formulation, the percentages of the first, second, and third components are 50%, 20-30%, and 20-30%, respectively.

20 **[0085]** Other formulations designed for 17-AAG are described in Tabibi et al., US 6,682,758 B1 (2004) and Ulm et al., WO 03/086381 A1 (2003); the disclosures of which are incorporated herein by reference.

25 **[0086]** The method of the present invention is used for the treatment of cancer. In one embodiment, the methods of the present invention are used to treat cancers of the head and neck, which include, but are not limited to, tumors of the nasal cavity, paranasal sinuses, nasopharynx, oral cavity, oropharynx, larynx, hypopharynx, salivary glands, and paragangliomas. In another embodiment, the compounds of the present invention are used to treat cancers of the liver and biliary tree, particularly hepatocellular carcinoma. In another embodiment, the compounds of the present invention are used to treat intestinal cancers, particularly colorectal cancer. In another embodiment, the compounds of the present invention are used to treat ovarian cancer. In another embodiment, the 30 compounds of the present invention are used to treat small cell and non-small cell lung cancer. In another embodiment, the compounds of the present invention are used to treat breast cancer. In another embodiment, the compounds of the present invention are used to treat sarcomas, including fibrosarcoma, malignant fibrous histiocytoma, embryonal

rhabdomysocarcoman, leiomyosarcoma, neuro-fibrosarcoma, osteosarcoma, synovial sarcoma, liposarcoma, and alveolar soft part sarcoma. In another embodiment, the compounds of the present invention are used to treat neoplasms of the central nervous systems, particularly brain cancer. In another embodiment, the compounds of the present invention are used to treat lymphomas which include Hodgkin's lymphoma, lymphoplasmacytoid lymphoma, follicular lymphoma, mucosa-associated lymphoid tissue lymphoma, mantle cell lymphoma, B-lineage large cell lymphoma, Burkitt's lymphoma, and T-cell anaplastic large cell lymphoma.

10                   Examples

[0087] The following Examples are provided to illustrate certain aspects of the present invention and to aid those of skill in the art in practicing the invention.

15                   Materials and Methods

15                   Cell line and reagents

[0088] Human colon adenocarcinoma cell line, DLD-1, and human breast adenocarcinoma cell line, SKBr-3, were obtained from American Type Culture Collection (manassas, VA). DLD-1 cells were maintained in RPMI 1640 medium supplemented with 10% fetal bovine serum, and SKBr-3 cells were cultured in McCoy's 5a medium supplemented with 10 % fetal bovine serum. 17-DMAG and 17-AAG were obtained using published procedures. Other cytotoxic agents were purchased commercially from suppliers such as Sigma Chemical Co. (St. Louis, MO) and Sequoia Research Products (Oxford, UK).

25                   Cell viability assay and combination effect analysis

[0089] Cells were seeded in duplicate in 96-well microtiter plates at a density of 5,000 cells per well and allowed to attach overnight. Cells were treated with 17-AAG or 17-DMAG and the corresponding enzyme inhibitor at varying concentrations, ranging from 0.5 picomolar ("pM") to 50 micromolar ("μM"), for 3 days. Cell viability was determined using the MTS assay (Promega). For the drug combination assay, cells were seeded in duplicate in 96-well plates (5,000 cells/well). After an overnight incubation, cells were treated with drug alone or a combination and the IC<sub>50</sub> value (the concentration of drug required to inhibit cell growth by 50%) was determined. Based on the IC<sub>50</sub> values

of each individual drug, combined drug treatment was designed at constant ratios of two drugs, *i.e.*, equivalent to the ratio of their IC<sub>50</sub>. Two treatment schedules were used: In one schedule, the cells were exposed to 24 hours of 17-AAG or 17-DMAG. The drug was then added to the cells and incubated for 48 hours. In another schedule, cells were 5 exposed to the drug alone for 24 hours followed by addition of 17-AAG or 17-DMAG for 48 hours. Cell viability was determined by the MTS assay.

[0090] Synergism, additivity or antagonism was determined by median effect analysis using the combination index (CI) calculated using Calcusyn (Biosoft, Cambridge, UK). The combination index is defined as follows:

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$$CI = [D]_1/[D_x]_1 + [D]_2/[D_x]_2$$

The quantities [D]<sub>1</sub> and [D]<sub>2</sub> represent the concentrations of the first and second drug, respectively, that in combination provide a response of x % in the assay. The quantities 15 [D<sub>x</sub>]<sub>1</sub> and [D<sub>x</sub>]<sub>2</sub> represent the concentrations of the first and second drug, respectively, that when used alone provide a response of x % in the assay. Values of CI < 1, CI = 1, and CI > 1 indicated drug-drug synergism, additivity, and antagonism respectively (Chou and Talalay 1984). The “enhancing” effect of two drugs can also be determined.

20

## Results

### *17-AAG combination in DLD-1 cells*

[0091] The following table provides CI values for combinations of 17-AAG and the enzyme inhibitor Iressa in a DLD-1 cell assay. “Pre-administration” refers to the administration of 17-AAG to the cells before the administration of enzyme inhibitor; 25 “post-administration” refers to the administration of 17-AAG to the cells after the administration of enzyme inhibitor.

**Table 5: CI values for combinations in DLD-1 cells (human colorectal cancer cells).**

Enzyme Inhibitor	17-AAG Pre-Administration	17-AAG Post-Administration
Iressa	0.76 ± 0.37	0.41 ± 0.01

*17-AAG combination in SKSBr-3 cells*

[0092] The following table provides CI values for combinations of 17-AAG and the enzyme inhibitors SAHA, Trichostatin A, and Iressa in an SKBr-3 cell assay.

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**Table 6: CI values for combinations in SKBr cells (human breast cancer cells).**

Enzyme Inhibitor	17-AAG Pre-Administration	17-AAG Post-Administration
SAHA	1.05 ± 0.03	0.15 ± 0.12
Trichostatin A	1.16 ± 0.02	0.91 ± 0.007
Iressa	0.77 ± 0.29	0.67 ± 0.02

*Additional Observations*

[0093] Additional analysis indicated that both 17-AAG and 17-DMAG reduced the expression of ErbB2 protein in SKBr3 and glioma cells. This observation, taken in combination with the results reported above, indicates that combinations of 17-AAG or 17-DMAG with any of the enzyme inhibitors above that are known to be useful to treat diseases characterized by elevated ErbB2 protein expression (*i.e.*, levels of expressions of ErbB2 protein greater than those found in healthy cells).

Claims

1. A method for treating breast cancer in a patient, wherein the method comprises administering an HSP90 inhibitor and an enzyme inhibitor to the patient.

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2. The method of claim 1, wherein the HSP90 inhibitor is administered to the patient before the enzyme inhibitor.

10 3. The method of claim 1, wherein the HSP90 inhibitor is administered to the patient after the enzyme inhibitor.

4. The method of claim 2, wherein the HSP90 inhibitor is geldanamycin or a geldanamycin derivative.

15 5. The method of claim 3, wherein the HSP90 inhibitor is geldanamycin or a geldanamycin derivative.

6. The method of claim 4, wherein the HSP90 inhibitor is a geldanamycin derivative, and wherein the derivative is 17-AAG.

20

7. The method of claim 5, wherein the HSP 90 inhibitor is a geldanamycin derivative, and wherein the derivative is 17-AAG.

8. The method of claim 6, wherein the enzyme inhibitor is SAHA or Iressa.

25

9. A method for treating colorectal cancer in a patient, wherein the method comprises administering an HSP90 inhibitor and an enzyme inhibitor to the patient.

30 10. The method of claim 9, wherein the HSP90 inhibitor is administered to the patient after the enzyme inhibitor.

11. The method of claim 10, wherein the HSP90 inhibitor is administered to the patient before the enzyme inhibitor.

12. The method of claim 10, wherein the HSP90 inhibitor is geldanamycin or a geldanamycin derivative.

5 13. The method of claim 11, wherein the HSP90 inhibitor is geldanamycin or a geldanamycin derivative.

14. The method of claim 12, wherein the HSP90 inhibitor is a geldanamycin derivative, and wherein the derivative is 17-AAG.

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15. The method of claim 13, wherein the HSP90 inhibitor is a geldanamycin derivative, and wherein the derivative is 17-AAG.

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17. The method of claim 1, wherein the enzyme inhibitor is a histone deacetylase inhibitor.

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18. The method of claim 1, wherein the enzyme inhibitor is a tyrosine kinase inhibitor.

19. The method of claim 9, wherein the enzyme inhibitor is a histone deacetylase inhibitor.

25

20. The method of claim 9, wherein the enzyme inhibitor is a tyrosine kinase inhibitor.

30

21. The method of claim 1, wherein the HSP90 inhibitor is 17-AAG, and wherein the administration of 17-AAG and the enzyme inhibitor is performed once per week.

22. The method of claim 1, wherein the HSP90 inhibitor is 17-AAG, and wherein the administration of 17-AAG and the enzyme inhibitor is performed twice per week.

5 23. The method of claim 9, wherein the HSP90 inhibitor is 17-AAG, and wherein the administration of 17-AAG and the enzyme inhibitor is performed once per week.

10 24. The method of claim 9, wherein the HSP90 inhibitor is 17-AAG, and wherein the administration of 17-AAG and the enzyme inhibitor is performed twice per week.

25. The method of claim 21, wherein the therapeutic dose of 17-AAG is between 50 mg/m<sup>2</sup> and 450 mg/m<sup>2</sup>.

15 26. The method of claim 22, wherein the therapeutic dose of 17-AAG is between 50 mg/m<sup>2</sup> and 250 mg/m<sup>2</sup>.

20 27. The method of claim 23, wherein the therapeutic dose of 17-AAG is between 50 mg/m<sup>2</sup> and 450 mg/m<sup>2</sup>.

28. The method of claim 24, wherein the therapeutic dose of 17-AAG is between 50 mg/m<sup>2</sup> and 250 mg/m<sup>2</sup>.