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(54) Title: ANTIBODY AGAINST ALPHA-11 INTEGRIN AND ITS USE

(57) Abstract: In a first aspect, the present invention relates to an antibody directed against the alpha-11 integrin subunit, in particular, said antibody is an antibody binding the same epitope as the antibody produced by the hybridoma deposited as DSM ACC3320. Further, the present invention relates to a method for detecting the alpha-11 integrin subunit in a sample. In particular, the antibody according to the present is an anti-body suitable for use in samples being cryopreserved and cryosectioned or formaldehyde fixed and paraffin embedded. The present invention relates further to a kit for detecting the alpha-11 integrin subunit containing the antibody according to the present invention. Finally, the present invention provides the antibody in a humanized form. The antibody, in particular, in its humanized forms are useful for antibody drug conjugates.



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Antibody against alpha-11 integrin and its use

In a first aspect, the present invention relates to an antibody directed against the al-
5 pha-11 integrin subunit, in particular, said antibody is an antibody binding the same
epitope as the antibody produced by the hybridoma deposited as DSM ACC3320.
Further, the present invention relates to a method for detecting the alpha-11 integrin
subunit in a sample. In particular, the antibody according to the present is an anti-
body suitable for use in tissue samples being cryopreserved and cryosectioned or
10 formaldehyde fixed and paraffin embedded. The present invention relates further to a
kit for detecting the alpha-11 integrin subunit containing the antibody according to the
present invention. Finally, the present invention provides the antibody in a humanized
form. The antibody, in particular, in its humanized forms are useful for antibody drug
conjugates.

15

Prior art

Integrins are heterodimeric cell surface receptors composed of non-covalently asso-
ciated alpha and beta subunits, which act as cell surface links to extracellular matrix
(ECM) and to other cells in dynamic cell-cell linkages.

20

Integrin subunits are composed of different domains with different functions.
The extracellular domain of collagen-binding alpha integrin chain contain an inserted
alpha I domain, which is responsible for collagen-binding without direct involvement
of the beta subunit. Whereas different integrin alpha chains display conserved re-
gions including their cytoplasmic tail, the cytoplasmic tails of integrin alpha chains
25 show little sequence similarity except for the vary proximal membrane sequence
GFFXR (Seq.ID. NO 11). Of note, alpha-11 integrin lacks this conserved sequence
but contains the GFFRS sequence (Seq.ID. NO 12). This conserved sequence is in-
tegral for the activity of the integrin. Namely, mutations of this sequence in general
result in integrins being constitutively active.

Integrins are involved in various functions including various signal transduction pathways mediating cellular signals. These signal transduction pathways include the regulation of the cell cycle, the organization of the intracellular cytoskeleton, the movement of new receptors to the cell membrane, etc. The presence of integrins allows rapid and flexible responses to events at the surface. The integrin alpha-11 is a polypeptide that is encoded in humans by the ITGA11 gene. It is described that the integrin alpha-11 binds collagen when present as a heterodimer with beta-1 integrin. Alpha-11 beta-1 integrin represents a collagen receptor, which is the latest identified member of the integrin family. This integrin heterodimer has distinct functions *in vivo* from other collagen-binding integrins. It was demonstrated in mice that collagen-binding integrins are dispensable for normal development, but suggest important roles in tissue remodeling events occurring in wound healing, fibrosis and tumor-stroma interactions.

For example, it is described that non-small lung cancers (NSCLC) express alpha-11 integrin subunit in the activated stroma, when it has the potential to be a biomarker for activated cancer associated fibroblasts [1-3].

The importance of the tumor microenvironment for tumor growth and tumor spread is increasingly being recognized. The major cell types in the tumor stroma of solid tumors include cancer-associated fibroblasts (CAFs) of varying origin, endothelial cells, pericytes, mesenchymal stem cells, tumor stem cells and immune cells. The ECM in addition to serving as a structural scaffold serves as a reservoir of growth factors and cytokines, which take part in bidirectional communication that occurs between the stroma and the tumor cells. For example, CAFs produce collagen cross-linking enzymes of the LOX family, which can increase stiffness of ECM, affecting the growth and invasion of tumor cells. CAFs thus constitute a group of fibroblastic cells of different origin, some of which share characteristics with myofibroblasts in granulation tissue during wound healing and tissue fibrosis. The mesenchymal derived CAF population seems to represent a heterogeneous cell mixture compared to resident tissue fibroblasts in typical resting tissue. For contractile activated myofibroblasts and CAFs the protein alpha smooth muscle actin (alpha SMA) has been suggested as a suitable marker. However, careful analysis of collagen producing activated fibroblast in heart, lung and kidney fibrosis has revealed that only a fraction of these fibroblasts expressed alpha SMA suggesting that alpha SMA is an inconsistent marker of activated collagen producing cells [4]. There is thus a need for better cell type specific

markers to be able to understand the dynamics of different CAF population in the tumor stroma. As noted, NSCLC stroma express integrin alpha-11 and, in addition, recent studies suggest that the stroma expression of alpha-11 in NSCLC correlates with LOXL1 expression and tumor stiffness. In fibrosis models importance of integrins have been demonstrated. Although expression of alpha-11 integrin in NSCLC in the activated stroma has been described, and regulation of cancer stroma stiffness of said integrin as well as promoting tumorigenicity and metastases, is disclosed, a suitable marker to discriminate between different subclasses of fibroblasts is lacking. This is particularly true for typical tissue samples of tumor patients and the fibrotic tissues in various types of fibrosis. These samples obtained as biopsies or obtained from the tumor during surgery, are acetone fixed or formaldehyde fixed before embedding the sample into a suitable embedding medium. The most often used medium for embedding samples is paraffin. However, markers suitable for specific labeling of different fibroblasts subclasses in paraffin embedded samples are rare. In particular, for alpha-11 integrin, no suitable markers are described in the literature so far.

WO 2008/075045 A1 and WO 2008/075038 A1 identify binding agents to the integrin alpha-11 subunit. Therein, the antibody A03 representing a scFv molecule is described. However, this molecule is not available and, thus, cannot be used for comparison. However, according to the knowledge of the present inventors, this antibody is not suitable for staining of paraffin embedded sections.

Further, WO 2016/133449 A1 disclose a detection and treatment of malignant tumors and CNS using an antibody specific malignant tumor and integrin alpha-10 subunit.

25 Brief description of the present invention

Hence, an object of the present invention is the provision of a biomarker suitable for determining alpha-11 integrin subunit in samples, in particular, tissue samples, like tissue samples obtained from cancer and fibrosis patients.

The present inventors aim in providing an antibody suitable for determining the alpha-11 integrin subunit in fixed samples, in particular, in acetone fixed or formaldehyde fixed samples, e.g. in samples being cryopreserved or paraffin embedded.

In a first aspect, the present invention provides an antibody that binds to the same epitope of the alpha-11 integrin subunit as bound by 210 F4B6A4 produced by the hybridoma deposited as DSM ACC3320.

In an embodiment, the antibody is the specific antibody produced by the hybridoma deposited as DSM ACC3320.

In an embodiment, the antibody comprise at least one of SEQ ID No. 1 to SEQ ID No. 6, in an embodiment, all sequences of SEQ ID No. 1, SEQ ID No. 2, SEQ ID No. 3, SEQ ID No. 4, SEQ ID No. 5 and SEQ ID No. 6 are present.

In a further aspect, the use of the antibody according to the present invention as a marker for the expression of the alpha-11 integrin subunit, in particular in tissue.

In a further embodiment, the present invention relates to a method for determining the alpha-11 integrin subunit expression in a sample containing cells and/or tissue. The method includes the step of incubating the sample with an antibody according to the present invention and determining binding of said antibody specifically to the alpha-11 integrin subunit by suitable detection means including immunohistological and immunocytochemical detection. The method is particularly suitable for detecting the alpha-11 integrin subunit in cryopreserved or paraffin embedded samples, e.g. tissue samples from cancer and fibrosis patients.

In a further aspect, the present invention provides a kit for detecting alpha-11 integrin subunit in a sample comprising the antibody according to the present invention and detection means for detecting binding specifically of the antibodies according to the present invention to the alpha-11 integrin subunit present in said sample.

Finally, the present invention provides molecules being antibody drug conjugates which are suitable for pharmaceutical purposes. In particular, said antibody drug conjugates contain antibodies being humanized by known methods.

Brief description of the drawings

Figure 1. Immunological characterization of 210F4B6A4 mAb

A-B. C2C12 mouse myoblast cells expressing human integrin α 11-EGFP or WT (wildtype) C2C12 cells were stained with 210F4B6A4 mAb for 1 hour at 4°C. Cells were then washed and stained with goat anti-mouse R-Phycoerythrin IgG. Samples were analysed by flow cytometry using Intellicyt iQue and fluorescence intensity was measured.

C. Immunoprecipitation (IP) of integrin α 11 β 1 with the 210F4B6A4 mAb and the polyclonal α 11 antibody (α 11 pAb)[5]. Protein bands of integrin chain α 11 at the expected size after immunoprecipitation using α 11 antibodies followed by Western blotting and detection using the polyclonal α 11 antibody.

D. Western-blotting (WB) using the $\alpha 11$ 210F4B6A4 mAb on C2C12-hu $\alpha 11$ -EGFP lysate, shown is the protein band of expected size of $\alpha 11$ -EGFP (180 kDa).

E. C2C12 mouse myoblast cells expressing human integrin $\alpha 11$ were allowed to attach on collagen I-coated coverslips for 2 hours, cell were then fixed with methanol and incubated with with 210F4B6A4 mAb or control IgG. Bound antibody was detected using goat anti-mouse Alexa Fluor 594. Integrin $\alpha 11$ staining in cell focal adhesions is indicated by white arrowheads.

Figure 2 shows integrin alpha-11 expression in human breast carcinomas by immunohistochemistry with formalin fixed, paraffin embedded sections of invasive human breast carcinoma stained with a monoclonal mouse anti-human integrin alpha-11 antibody clone 210 F4B6A4.

Detailed description of the present invention

The present invention aims to provide new antibodies suitable for specifically binding the alpha-11 integrin subunit. Namely, the present invention provides antibodies that's bind to the same epitope of the alpha-11 integrin subunit as bound by 210F4B6A4 antibody produced by the hybridoma deposited as DSM ACC3320. The antibodies according to the present invention are suitable for immune detection of the antigen, namely, the alpha-11 integrin subunit, in samples, in particular, in cryo-preserved or paraffin embedded samples.

As used herein, the term "comprise" or "comprising" as well as "contain" or "containing" include the embodiment of "consist" and "consisting of".

Further, the term "antibody" refers to a polypeptide encoded by an immunoglobulin gene or functional fragments thereof that specifically binds and recognizes an antigen. The recognized immunoglobulin genes include the kappa, lambda, alpha, gamma, delta, epsilon and mu constant region genes, as well as the myriad immunoglobulin variable region genes. Light chains are classified as either kappa or lambda. Heavy chains are classified as gamma, mu, alpha, delta or epsilon, which in turn define the immunoglobulin classes, IgG, IgM, IgA, IgD and IgE, respectively.

Unless otherwise indicated, the term "antibody" includes the embodiments of an antigen-binding fragment thereof with binding specificity for an integrin alpha-11 subunit, or a heterodimer thereof, or a variant, fusion or derivative of said antibody on antigen-binding fragment, a fusion of said variant or derivative thereof, which retains

the binding specificity for integrin alpha-11 subunit or a heterodimer thereof. For example, the antibody may be a monoclonal antibody.

By “antigen-binding fragment” a functional fragment of an antibody that is capable of binding to an integrin alpha-11 subunit, a heterodimer thereof, as defined
5 herein is meant.

Exemplary antigen-binding fragments of the invention may be selected from the group consisting of Fv fragments, like single chain Fv (scFv) and disulfide-bonded Fv, and Fab-like fragments, like Fab fragments, Fab' fragments and F(ab)₂ fragments. In one embodiment, the antigen binding fragment is a scFv.

10 The immunoglobulin (antibody) structure unit comprises typically a tetramer composed of two identical pairs of polypeptide chains, each pair having one light and one heavy chain. The N-terminus of each chain defines a variable region of about 100 to 110 or more amino acids primarily responsible for antigen recognition. Thus, the terms “variable heavy chain”, V_H, or V_H, refer to the variable region of an immunoglobulin heavy chain including an Fv, scFv, dsFv or Fab, while the terms “variable
15 light chain”, “V_L” or “vL” refers to the variable region of the immunoglobulin light chain, including of an Fv, scFv, dsFv or Fab.

Examples of antibody functional fragments include but are not limited to complete antibody molecules, antibody fragments, such as Fv, scFv, complementarity determining regions (CDRs), V_L (light chain variable region), V_H (heavy chain variable
20 region), Fab, F(ab)₂' and any combination of those or any other functional portion of an immunoglobulin peptide capable of binding to target antigen. The respective antibody and antibody fragments can be obtained by variety of methods known to the skilled person, namely, either by digestion of an intact antibody with an enzyme or by
25 de novo synthesis. The de novo synthesis can either be chemically or by using recombinant DNA technology. Unless otherwise indicated, the term antibody includes antibody fragments obtained by the methods known in the art also including phage display libraries. The term “antibody” also includes bivalent or bispecific molecules, diabodies, triabodies and tetrabodies etc.

30 The term “humanized” antibody is an antibody that retains the reactivity of a non-human antibody while being less immunogenic in humans. This can be achieved by methods along known in the art by retaining the non-human CDR regions and replacing the remaining parts of the antibodies with the human counterparts, a technic well known to the skilled person.

“Single chain Fv (scFv)” or “single chain antibodies” refers to a protein wherein the V_H and the V_L regions of a scFv antibody comprise a single chain which is folded to create an antigen binding site similar to that found in two chain antibodies. Methods for producing the same are described in the art. Single chain Fv antibodies optionally include a peptide linker of no more than 50 amino acids in length.

The phrase “specifically (or selectively) binds to an antibody” when referring to a protein or peptide, refers to a binding reaction which is determinative of the presence of the protein in the presence of a heterogeneous population of proteins and other biologics. That is, in the designated immunoassay conditions typically present in said immunoassays, the specific antibodies bind to a particular protein, in the present case, the antibody according to the present invention to the alpha-11 integrin subunit and do not bind in a significant amount to other proteins present in the sample. Typically, a specific or selective reaction will be at least twice the background signal or noise and more typically more than 10 to 100 times over the background.

The antibody which is in a one embodiment a monoclonal antibody according to the present invention can be obtained from any animal or the human being, whereby the monoclonal antibody from a mouse are preferred. Further, the monoclonal antibody may be altered biochemically, by genetic manipulation, or it may be synthetic, with the antibody possibly lacking portions completely or in parts, said portions being necessary for the recognition of alpha-11 integrin subunit and being substituted by others imparting further advantageous properties to the antibody.

A hybridoma cell line producing a preferred antibody of the present invention, namely, the monoclonal mouse antibody F210F4B6A4, was deposited at the Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH (DSMZ) in Braunschweig under the number DSM ACC3320 on March 21, 2017.

The antibody F210F4B6A4 deposited with the depositing number DSM ACC3320 include the sequences of SEQ ID Nos. 1 to 6:

DDYIH (Seq. ID No. 1);
WIDPENGDTHEYASNFQG (Seq. ID No. 2);
EYAY (Seq. ID No. 3);
RSSQSLLYSNGNTYLQ (Seq. ID No. 4);
RASNRFS (Seq. ID No. 5);
SQSTHVPWT (Seq. ID No. 6).

The term "epitope" also known as "antigenic determinant" is a part of an antigen that is recognized by the immune system in the present context by antibodies. The epitopes according to the present invention may be conformational epitopes and linear epitopes, based on their structure and interaction with the paratope, namely the
5 part of the antibody that binds to the epitope.

The skilled person is well aware of suitable methods to determine whether an antibody binds to the same epitope of the alpha-11 integrin subunit as bound by 210 F4B6A4 produced by the hybridoma deposited as DSM ACC3320. For example, appropriate tests may include experiments were competitive binding assays of the anti-
10 body produced by the hybridoma identified herein and the antibody in question are conducted.

An agent that specifically competes for binding reduces the specific binding of an antibody to a polypeptide. That is, competitive binding assays may be conducted to determine appropriate antibodies using the 210F4B6A4 antibody produced by the
15 hybridoma cell line disclosed herewith.

The term "sample" is intended to cover all types of samples suitable for the purpose of the invention. Examples of such samples are serum, sputum, urine, liquor, tissue, and biopsies. In particular, the sample may be a blood sample or a tissue sample. In an embodiment, the tissue sample is a paraffin embedded tissue sample,
20 a cryopreserved sample or a cell sample.

In an embodiment of the present invention, the antibody is the antibody 210F4B6A4 produced by the hybridoma deposited as DSM ACC3320.

In another embodiment, the antibody, which is typically a monoclonal antibody, is an antibody comprising the heavy and light chain complementary determining regions (CDR) of the antibody according to the present invention, in particular, wherein
25 the CDR are the CDR of the monoclonal antibody produced by the hybridoma deposited as DSM ACC3320.

The CDR of the monoclonal antibody deposited as DSM ACC3320 are as follows: heavy chain: CDR1 is SEQ ID No. 1; CDR2 is SED ID No. 2; CDR3 is SED ID
30 No. 3; the light chain CDRs are as follows: CDR1 is SEQ ID No. 4; CDR2 is SEQ ID No. 5; CDR3 is SEQ ID No. 6.

The CDR sequences including the framework region are shown in SEQ ID No. 7 of the heavy chain and SEQ ID No. 8 of the light chain of the deposited antibody, respectively.

In an embodiment of the present invention, the antibody is comprising a heavy chain variable region comprising the following CDR's:

- a) Seq. ID No. 1;
- b) Seq. ID No. 2; and
- 5 c) Seq. ID No. 3.

In a further embodiment, the heavy chain variable region comprises or consist of the amino acid sequence of Seq. ID No. 7.

Further, in another embodiment of the present invention, the antibody comprise a light chain variable region comprising the following CDR's:

- 10 a) Seq. ID No. 4;
- b) Seq. ID No. 5; and
- c) Seq. ID No. 6.

Further, the antibody according to another embodiment is an antibody wherein the light chain variable region comprises or consist of the amino acid sequence of
15 Seq. ID No. 8.

Moreover, another embodiment of the present invention is an antibody comprising the heavy chain variable region as defined above and a light chain variable region as defined above. The coding nucleic acid sequence of the heavy chain of Seq. ID. No. 7 and of the light chain of Seq. ID. No. 8 including the leader sequences are
20 shown in SEQ ID No. 9 (heavy chain) and SEQ ID No. 10 (light chain), respectively.

In an another embodiment, the present invention relates to antibodies, in particular, monoclonal antibodies being selected from the group consisting of scFv, Fab, (Fab')₂.

In an embodiment, the antibody according to the present invention, in particular,
25 monoclonal antibodies like the 210F4B6A4 monoclonal antibody or derivatives thereof are useful as a marker of the expression of the alpha-11 integrin subunit in a sample. As noted, the sample may be a body fluid or a tissue sample. In an embodiment of the present invention, the detection of the antibody binding specifically to the alpha-11 subunit according to the present invention is by an immunohistochemical or
30 immunocytochemical method.

The antibodies according to the present invention are excellently suitable in the application of diagnostic problems, e. g. determined by immunohistochemistry with quantitative expression parameters. Alternatively, said antibodies may be used in Western Blot or immunoprecipitation analysis.

For example, the antibody according to the present invention is particularly useful for determining the alpha-11 integrin subunit in samples whereby said samples were fixed before by known methods, for example which wherein the samples are acetone-fixed or formaldehyde-fixed before. That is, the use is particularly for staining
5 alpha-11 integrin subunit in tissues, preferably on cryopreserved or routinely fixed and paraffin embedded tissues. For this the antibodies according to the present inventions may be labelled appropriately, as described above, or employed in combination with label antibodies direct against them or other reagents.

That is, the antibodies according to the present invention can be used diagnostically to monitor protein levels of the alpha-11 integrin subunit in tissue as part of clinical
10 examination or clinical testing procedures, e.g. to determine the efficacy of a given treatment regimen or determining cancer as discussed below. Detection can be facilitated by coupling, e.g. physically linking, the antibody to a detectable substance. Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials and radioactive
15 materials. Examples of suitable enzymes include horseradish peroxidase; alkaline phosphatase (-galactosidase, or acetylcholinesterase); examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include fluorescein, fluorescein isothiocyanate, rhodamine, Cy-dyes, alexa fluor-dyes or brilliant violet dyes. Suitable radioactive materials include ^{125}I , ^{131}I , ^{35}S or ^3H while luminescent and bioluminescent materials include luminol as well as luciferase, luciferin and aequorin.

As noted above, detection may be also in combination with labelled secondary antibodies directed against the antibody according to the present invention. The
25 skilled person is well known of suitable secondary antibodies which means that these antibodies are directed against the species from which the antibody according to the present invention is derived. Suitable examples in case of mouse monoclonal antibodies include gold anti mouse antibodies labelled with enzymes or dyes etc. which are commercially available.

30 The antibody are useful for detecting the alpha-11 integrin subunit expression by cultured cells or in cryopreserved tissue or tissues present in paraffin embedded samples. The present antibodies allow to detect routinely and specifically the alpha-11 integrin subunit in cryosections or paraffin embedded samples.

In another embodiment, the present invention relates to the use of an antibody according to the present invention for detecting the alpha-11 integrin subunit expression wherein the samples as sampled being formaldehyde fixed (formalin) and paraffin embedded samples, in particular, tissue samples.

5 In a further embodiment, the antibody according to the present invention, i.e. the monoclonal antibody according to the present invention, are humanized antibodies. That is, the antibody is a humanized antibody, i.e. an antibody that retains the reactivity of a non-human antibody while being less immunogenic in humans. This can be achieved for instance by retaining the non-human CDR regions and replacing the
10 remaining parts of the antibody with the human counterparts. This is a well established procedure known in the art. In cases where the transfer of the CDR to a human framework leads to a loss of specificity for humanized antibody, back mutations can be introduced to the framework regions of the human portion of the antibodies. These methods are described in the art accordingly.

15 The antibody according to the present invention, in particular, the humanized antibodies, may be also useful as active agents in pharmaceuticals. That is, these active agents may be used to treat diseases, disorders or conditions where the alpha-11 integrin subunit expression is altered or when alteration of the alpha-11 integrin subunit expression is beneficial to the subject receiving said treatment. These dis-
20 eases, disorders and conditions include cancer, fibrosis, wound healing and degenerative inflammatory and non-inflammatory diseases such as rheumatoid arthritis and osteoarthritis.

Another embodiment of the present invention relates to a method for determining the alpha-11 integrin subunit expression in a sample whereby said sample com-
25 prises cells and tissue, including the step of incubating the sample with the antibody according to the present invention and determining specific binding of said antibody by suitable means including immunohistological and immunohistochemical detection. The detection means are described above including label and marker. Said label and marker may be present with the antibody itself or may be present on secondary anti-
30 bodies or secondary detection compounds containing a label or marker for detection accordingly.

In an embodiment of the method according to the present invention, the sample are tissue sections, in particular, human tissue sections. Further, in an embodiment of the present invention, the tissue sections are acetone, methanol or aldehyde fixed

tissue sections, in particular, said tissue sections, or samples in general, are cryosections or paraffin embedded samples including paraffin embedded tissue sections or paraffin embedded cell sections.

The method according to the present invention may comprise further suitable steps for embedding the tissue and/or cells including paraffin embedding steps. The skilled person is well aware of the steps required for embedding the samples accordingly. In addition, the method according to the present invention may further comprise the necessary steps to treat the paraffin embedded samples to allow detection of antigens by antibodies accordingly.

Moreover, this method according to the present invention for detecting alpha-11 integrin subunit contain further steps of detection including immunohistochemical and immunohistological detection steps, like binding of secondary antibodies and staining of the sections when using enzyme based detection systems or microscopy etc. when using fluorescence based or chromophore based systems.

The use of a method according to the present invention are useful for detecting various types of cancer. In particular, the antibodies according to the present invention as well as the method according to the present invention allow to identify fibroblastoid expression in keratin-negative, vimentin-positive, non-vessel associated stroma of invasive breast carcinoma, ovary adenocarcinoma, skin carcinoma and pancreas adenocarcinoma. In general, the antibodies according to the present invention may serve as a biomarker for CAFs in carcinoma-type tumor characterized by a desmoplastic stroma.

In an embodiment, the method according to the present invention is a method wherein the sample is a tissue section (form)aldehyde fixed and paraffin embedded tissue section.

In a further aspect, the present invention relates to a kit for detecting alpha-11 integrin subunit in a sample comprising the antibody according to the present invention. That is, the test kit also named diagnostic kit according to the present invention, in particular for use in the method according to the present invention, is an immunological kit for use in detecting the antibodies according to the present invention bound to the alpha-11 integrin subunit. This kit can generally comprise the antibody according to the present invention. Further, the kits will comprise in suitable container(s), one or more detecting agents, in particular, when the antibodies according to the present invention do not contain a marker or label. If required, the kit further comprises

substrate and further means for allowing reaction with an enzyme used as a label for the detecting agent which may be an antibody. The immunodetection agents of the kit can include detectable labels that are associated with or linked to the given detecting agent, typically, a detecting antibody being a secondary antibody. Detectable labels and markers include the labels or markers identified above. Optionally, the kits further comprise positive and negative controls accordingly. The term “determining” or “analyzing” as used herein refers to assessing the presence, absence, level or amount of the specific binding of the antibody to the alpha-11 subunit integrin within the sample.

10 Moreover, a method for identifying tissue or cells associated with a cancer condition or fibrosis is provided, the method comprising determining the amount of alpha-11 integrin subunit expression in a sample of cells or tissue to be tested using an antibody according to the present invention and comparing it to the amount of alpha-11 integrin subunit protein to a positive and/or negative control. In an embodiment, the method is a method wherein the cancer condition is a solid tumor including breast cancer. In another embodiment, the method is a method wherein the condition is fibrosis.

In a further aspect, an epitope defined by an antibody according to the present invention wherein the antibody comprises the following amino acid sequences:

20 DDYIH (Seq. ID No. 1);
 WIDPENGDEYASNFGG (Seq. ID No. 2);
 EYAY (Seq. ID No. 3);
 RSSQSLLYSNGNTYLQ (Seq. ID No. 4);
 RASNRFSS (Seq. ID No. 5);
25 SQSTHVPWT (Seq. ID No. 6)
 Is provided.

In an embodiment of the present invention, the presence of the antibodies can be determined using secondary antibodies from different species in the sample and the antibody according to the present invention. In a further embodiment, the kit contains further marker molecules allowing detection of the antibodies bound to the alpha-11 subunit present in the sample by immunohistochemical means including immunofluorescence and immunohistochemistry.

It is noted that as used herein, each of the embodiments described may be in combination with another embodiment of the same matter. That is, the features of

each of the embodiments may be combined to provide an antibody, method or use having some or all of the features mentioned in the specific embodiments, accordingly.

5 Examples

The following examples have been included to illustrate modes of the present disclosed subject matter. In light of the present disclosure and the general level of the skilled in the art, those of skilled in the art will appreciate that the following examples are intended to be exemplary only and that numerous changes and modifications can
10 be applied without departing from the scope of the present disclosed subject matter.

Material and Methods

Cell lines - C2C12 cells stably expressing human $\alpha 11$ integrin or human $\alpha 2$ integrin subunits (C2C12- $\alpha 11$ and C2C12- $\alpha 2$, respectively [6]) were cultured in DMEM me-
15 dium and 10% fetal bovine serum (FBS; Gibco) supplemented with antibiotics.

Generation of Mab specific to integrin $\alpha 11$ chain

Mabs were produced using established procedures. NT-HRM mice (nanoTools Antikoerpertechnik, Germany) were immunized with human $\alpha 11\beta 1$ integrin (R&D
20 Systems), boosted twice and cell fusion was performed on day 68. Luminex beads coated with $\alpha 11\beta 1$ integrin were used to screen $\alpha 11$ binders. Supernatants from positive clones were tested in FACS using C2C12- $\alpha 11$ cells as positive control and wild-type C2C12 (do not express human integrins), C2C12 - $\alpha 2$ cells and A431 cells (do express human integrin $\beta 1$, but not $\alpha 11$) as negative controls. Positive clones were
25 further characterized and finally subcloned by limited dilution.

Immunostaining

Cells were washed in PBS and fixed with ice cold methanol 10 min at -20C. Cells were blocked with 5% BSA/PBS containing 0.1% Triton X-100 for 1 hour at RT. Next,
30 cover slips were incubated with primary antibody, 210F4B6A4, in 5% BSA/PBS with 0.1% Triton X-100 for 1 hour at 37°C. Cells were then washed with 0.05% Tween-20/PBS and incubated with Alexa fluor® 594 goat anti-mouse IgG (1:400, Jackson ImmunoResearch) for 1 hour at RT. Later, coverslips were incubated with DAPI (0.25

µg/ml, Invitrogen) and mounted with ProLong Diamond Antifade mounting medium (Thermo Scientific). Cells were visualized under a Zeiss AxioScope fluorescence microscope and pictures were acquired with a digital AxioCam MRm camera.

5 Immunohistochemical staining of formaldehyde fixed and paraffin embedded tissue was conducted by known methods. Immunohistochemistry was performed on formalin fixed, paraffin embedded (FFPE) sections of invasive human breast carcinomas with the monoclonal antibody according to the present invention and as deposited.

10 The immunohistochemistry protocol was a protocol as described in the art with respect to immunohistochemical staining of FFPE sections.

In figure 2 the staining is shown whereby integrin alpha-11 was found to be expressed in spindle shaped cells in a breast tumor stroma. Figure 2 is a pictures with 400x.

Flow cytometry

15 Cells were harvested in PBS (without Ca^{2+} and Mg^{2+}) and accutase® (Biochrom) and washed in PBS (without Ca^{2+} and Mg^{2+}). 40µl of integrin α11 antibody supernatants (diluted 1:8 with Fluorescence activated cell sorting (FACS) buffer (PBS/0.2 mM Ca^{2+} , 0.1% BSA and 0.1% Poloxamer 188)) were mixed with 10^5 cells and incubated 1 hour at 4°C. Cells were then washed once with FACS buffer and incubated 1 hour
20 at 4°C with 50µl of PE labelled goat anti-mouse (diluted 1:400 in FACS buffer). Cells were washed once with FACS buffer and the samples analyzed by flow cytometry using Intellicyt iQue.

Immunoprecipitation

25 Subconfluent C2C12-α11 cells were lysed using ice-cold solubilization buffer (1% Triton X-100, 0.15 M NaCl, 10 mM Tris-HCl pH 7.4, 1 mM MgCl_2 , 1 mM CaCl_2) containing a protease inhibitor cocktail (complete mini EDTA-free, Roche Diagnostics). Cell lysates were centrifuged for 10 min at 4°C at 13000 rpm. The supernatant was pre-cleared by incubation with 100 µg/ml of preimmune IgG and protein A-Sepharose CL
30 4B (GE Healthcare) at 4°C overnight. In the next step, samples were incubated with 100 µg/ml of rabbit polyclonal anti-human α11 antibody [5] or mouse Mab 210 F4B6A4 hybridoma supernatant. The precipitates were washed three times with high salt buffer

(1% Triton X-100, 0.5 M NaCl, 10 mM Tris-HCl pH 7.4, 1 mM MgCl₂, 1 mM CaCl₂) and three times in a physiological salt buffer (0.1% Triton X-100, 0.15 M NaCl, 10 mM Tris-HCl pH 7.4, 1 mM MgCl₂, 1 mM CaCl₂) before solubilization in sodium dodecyl sulphate (SDS) -sample buffer with reducing agents. Proteins were separated by 6% SDS - poly
5 acrylamide gel electrophoresis (PAGE) and proteins were then transferred using iBlot® system. Membranes were blocked with 5% non-fat dry milk (Marvel, UK) in Tris-buffered saline containing 0.1% Tween20 (TBS-T), incubated with primary rabbit polyclonal anti-human α 11 antibody or mouse mab 203E3 anti-human integrin α 11 supernatant and to β -actin, overnight at 4°C. Following incubations, membranes were washed
10 in TBS-T three times for 10 min, and incubated with goat anti-mouse or goat anti-rabbit HRP conjugated secondary antibodies for 1 h at room temperature. Membranes were developed using ECL™ western blotting systems kit (GE Healthcare), and photographed using the ChemiDoc XRS device and the Quantity One 1-D Analysis Software (Bio-Rad).

15 It is demonstrated herein, that the antibody according to the present invention is suitable in staining specifically alpha 11 integrin subunit in tissues and cells. In particular, the antibody allow the bind and stain specifically the alpha 11 subunit in tissue, either cryopreserved or paraffin embedded. In particular, in the tumor context, alpha 11 has the potential to serve as a biomarker for tumor tissue characterized by a desmo-
20 plastic stroma.

Antibody sequences of the deposited clone

Total RNA was isolated from frozen hybridoma cell lysates following the technical manual of TRIzol® Reagent. Total RNA was then reversed transcribed into cDNA using isotype specific anti-sense primers or universal primers following the
25 technical manual of PrimeScript™ first strand cDNA synthesis kit. The antibody fragments of V_H and V_L were amplified according to standard operation procedure (SOP) of rapid amplification of sDNA ends (RACE) of GgenScript. Amplified antibody fragments were cloned into a standard cloning vector separately. Colony PCR was performed to screen for clones with inserts of correct sizes. No less than five colonies
30 with inserts of correct sizes were sequenced for each fragment. The sequences of different clones were aligned and the consensus sequence of the clones were provided.

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PCT

Ausdruck (Original in elektronischem Format)
 (Dieses Blatt zählt nicht als Blatt der internationalen Anmeldung und ist nicht Teil derselben)

0-1	Formular PCT/RO/134 Angaben zu einem hinterlegten Mikroorganismus und/oder anderem hinterlegten biologischen Material	
0-1-1	Erstellt mit	CMS Online Filing Version CMS 1.15 MT/FOP 20020701/0.20.5.20
0-2	Internationales Aktenzeichen	
0-3	Aktenzeichen des Anmelders oder Anwalts	4200 29 PCT 1
1	Die nachstehenden Angaben betreffen den Mikroorganismus und/ oder anderes biologisches Material, der/das in der Beschreibung genannt ist	
1-1	Seite	8
1-2	Zeile	15-18
1-3	Angaben betr. Hinterlegung	
1-3-1	Name der Hinterlegungsstelle	DSMZ Leibniz-Institut DSMZ - Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH (DSMZ)
1-3-2	Anschrift der Hinterlegungsstelle	Inhoffenstr. 7B 38124 Braunschweig, Germany
1-3-3	Datum der Hinterlegung	03. April 2017 (03.04.2017)
1-3-4	Eingangsnummer	DSMZDSM ACC3320
1-4	Weitere Angaben	
1-5	Bestimmungsstaaten, für die besondere Angaben gemacht werden	Alle Bestimmungsstaaten
1-6	Gesondert eingereichte Angaben Diese Angaben werden dem Internationalen Büro später übermittelt	depositors statement of authorization

VOM ANMELDEAMT AUSZUFÜLLEN

0-4	Dieses Formular ist mit der internationalen Anmeldung eingegangen (ja oder nein)	ja
0-4-1	Bevollmächtigter Bediensteter	Benzler, Annemarie

VOM INTERNATIONALEN BÜRO AUSZUFÜLLEN

0-5	Dieses Formular ist an folgendem Datum beim internationalen Büro eingegangen	
0-5-1	Bevollmächtigter Bediensteter	

Claims

1. An antibody that binds to the same epitope of the alpha-11 integrin subunit as bound by 210 F4B6A4 produced by the hybridoma deposited as DSM ACC3320.
5
2. The antibody of claim 2 wherein the antibody is 210 F4B6A4 produced by the hybridoma deposited as DSM ACC3320.
3. The antibody of claim 1 or 2 comprising the heavy and light chain complementarity determining regions (CDR) of the antibody as defined in claim 1 or 2, in particular,
10 wherein the CDR are CDR of the antibody produced by the hybridoma deposited as DSM ACC3320.
4. The antibody according to any one of the preceding claims wherein the antibody
15 comprises at least one the following amino acid sequences:
DDYIH (Seq. ID No. 1);
WIDPENGDTHEYASNFG (Seq. ID No. 2);
EYAY (Seq. ID No. 3);
RSSQSLLYSNGNTYLQ (Seq. ID No. 4);
20 RASNRFS (Seq. ID No. 5);
SQSTHVPWT (Seq. ID No. 6).
5. The antibody according to any one of the preceding claims wherein the antibody
is selected from the group consisting of scFv, a Fab, and a (Fab')₂.
25
6. The antibody according to any one of the preceding claims comprising a heavy chain variable region comprising the following CDR's:
a) Seq. ID No. 1;
b) Seq. ID No. 2; and
30 c) Seq. ID No. 3.
7. The antibody according to claim 6 wherein the heavy chain variable region comprises or consist of the amino acid sequence of Seq. ID No. 7.

8. The antibody according to any one of the preceding claims comprising a light chain variable region comprising the following CDR's:
- a) Seq. ID No. 4;
 - b) Seq. ID No. 5; and
 - 5 c) Seq. ID No. 6.
9. The antibody according to claim 8 wherein the light chain variable region comprises or consist of the amino acid sequence of Seq. ID No. 8.
- 10 10. The antibody according to any one of the preceding claims comprising the heavy chain variable region as defined in claim 7 or 8 and a light chain variable region as defined in claim 8 or 9.
11. The use of an antibody according to any one of the preceding claims as a
15 marker for the expression of the alpha-11 integrin subunit.
12. The use of an antibody according to claim 11 for detecting the alpha-11 integrin subunit expression in a sample by an immunohistochemical or immunocytochemical method.
- 20 13. The use of an antibody according to claim 11 or 12 wherein the sample is a sample which was fixed before, preferably which was acetone-, methanol- fixed or formaldehyde fixed before.
- 25 14. The use of antibody according to claim 13 wherein the cells or tissue present in the sample are cryopreserved and cryosectioned, or paraffin embedded samples.
15. The use according to any one of claims 11 to 14 wherein the sample is a sample being (form)aldehyde fixed and paraffin embedded samples in particular, tissue
30 samples.
16. A method for determining the alpha-11 integrin subunit expression in a sample comprising cells and/or tissue, including the step of incubating the sample with an an-

tibody according to any one of claims 1 to 10 and determining binding of said antibody by suitable means including immunohistological and immunocytochemical detection.

- 5 17. The method according to claim 16 wherein the sample are tissue sections, in particular, human tissue sections.
18. The method according to claim 17 wherein the tissue sections are acetone-methanol- or (form)aldehyde fixed tissue sections, in particular, cryosections or paraffin
10 fin embedded tissue sections.
19. The method according to any one of claims 16 to 18 wherein the sample is a tissue section being (form)aldehyde fixed and paraffin embedded tissue section.
- 15 20. A kit for detecting alpha-11 integrin subunit in a sample comprising an antibody as defined in any one of claims 1 to 10
21. The kit according to claim 20 further comprising means for determining binding of the antibodies.
20
22. The kit according to claim 21 wherein the means for determining the presence of the antibodies include secondary antibodies from different species than the sample and the antibody as defined in any one of claims 1 to 4.
- 25 23. The kit according to any one of claims 20 to 22 further containing marker molecules allowing detection of the antibodies bound to the alpha-11 integrin subunit present in the sample by immunohistochemical means including immunofluorescence.
- 30 24. A method for identifying tissue or cells associated with a cancer condition or fibrosis, the method comprising determining the amount of alpha-11 integrin subunit expression in a sample of cells or tissue to be tested using an antibody as defined in any one of claims 1 to 10 and comparing it to the amount of alpha-11 integrin subunit protein to a positive and/or negative control.

25. The method according to claim 24 wherein the cancer condition is a solid tumor including breast cancer.

26. The method according to claim 24 wherein the condition is fibrosis.

5

27. An epitope defined by an antibody wherein the antibody comprises the following amino acid sequences:

DDYIH (Seq. ID No. 1);

WIDPENGDTHEYASNFQG (Seq. ID No. 2);

10 EYAY (Seq. ID No. 3);

RSSQSLLYSNGNTYLQ (Seq. ID No. 4);

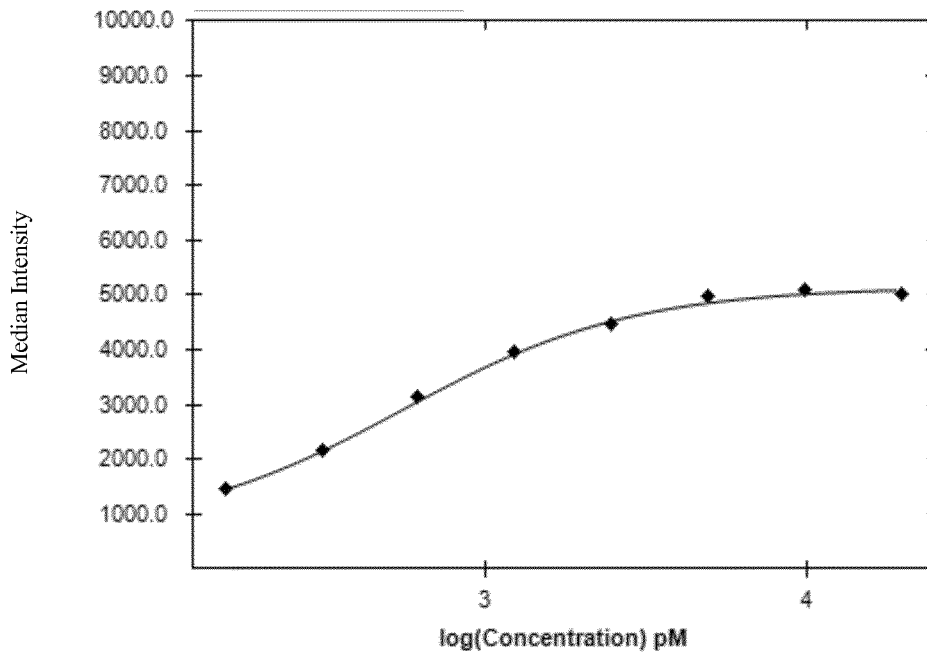
RASNRFS (Seq. ID No. 5);

SQSTHVPWT (Seq. ID No. 6).

15

1/3

C2C12- α 11; 210F4B6A4 maAb



C2C12 WT; 210F4B6A4 mAb

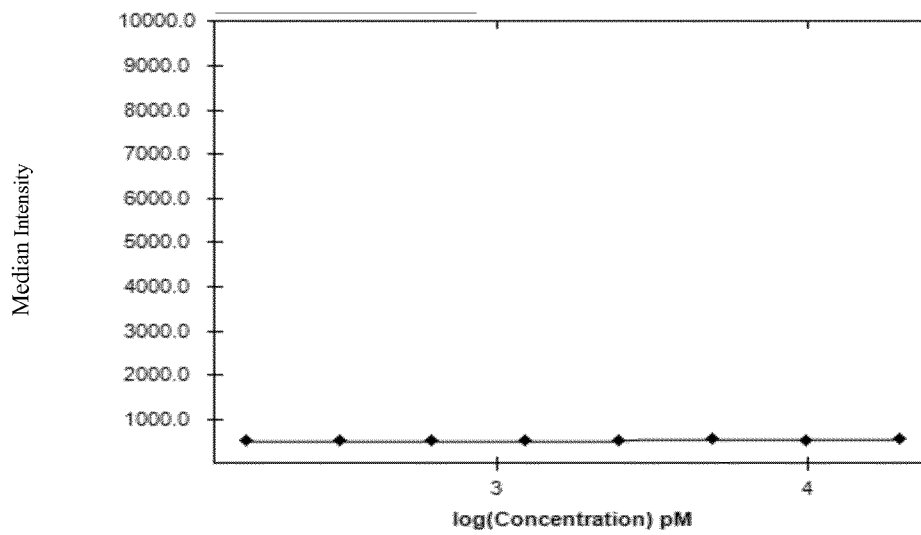


Figure 1 A,B

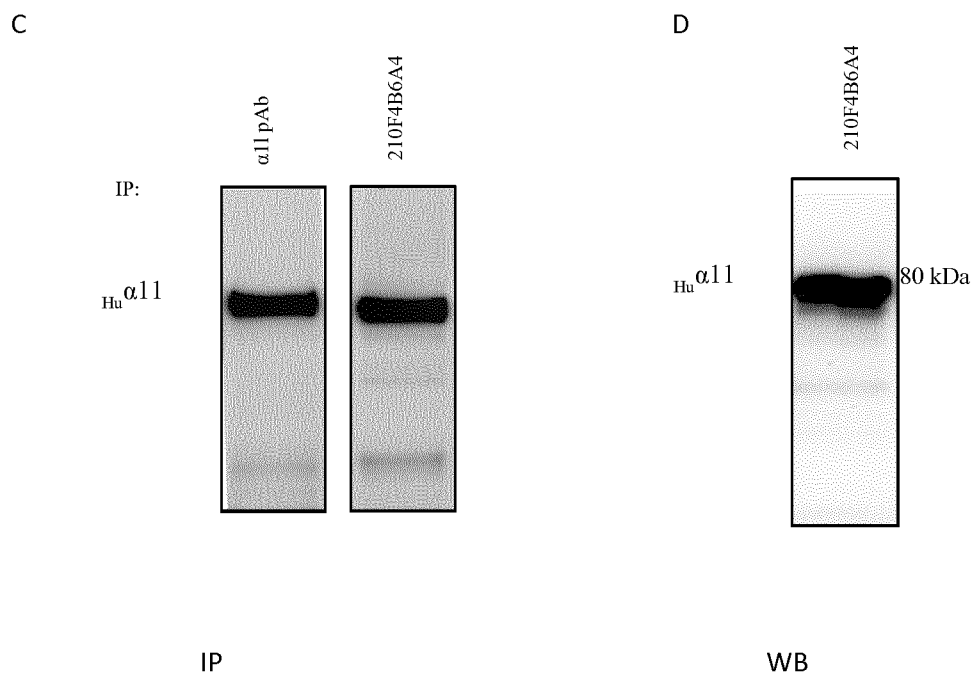


Figure 1 C, D

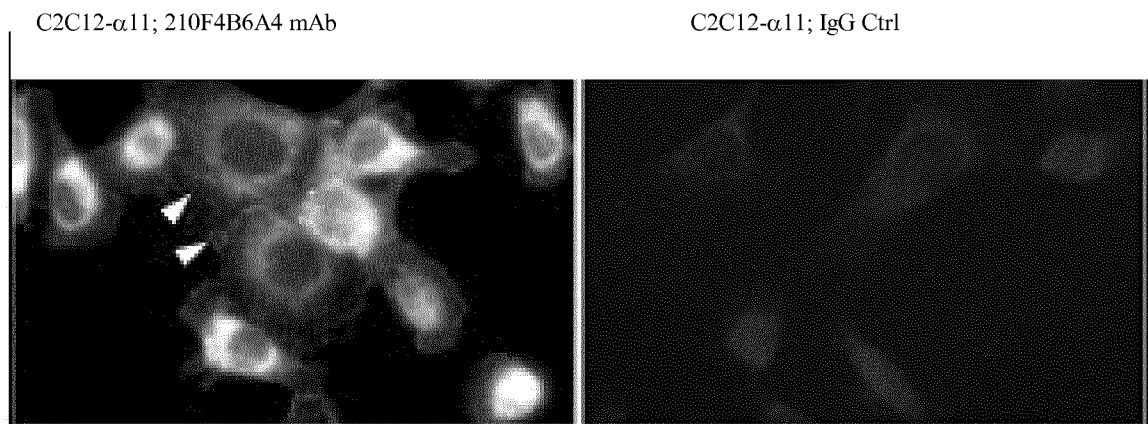


Figure 1E

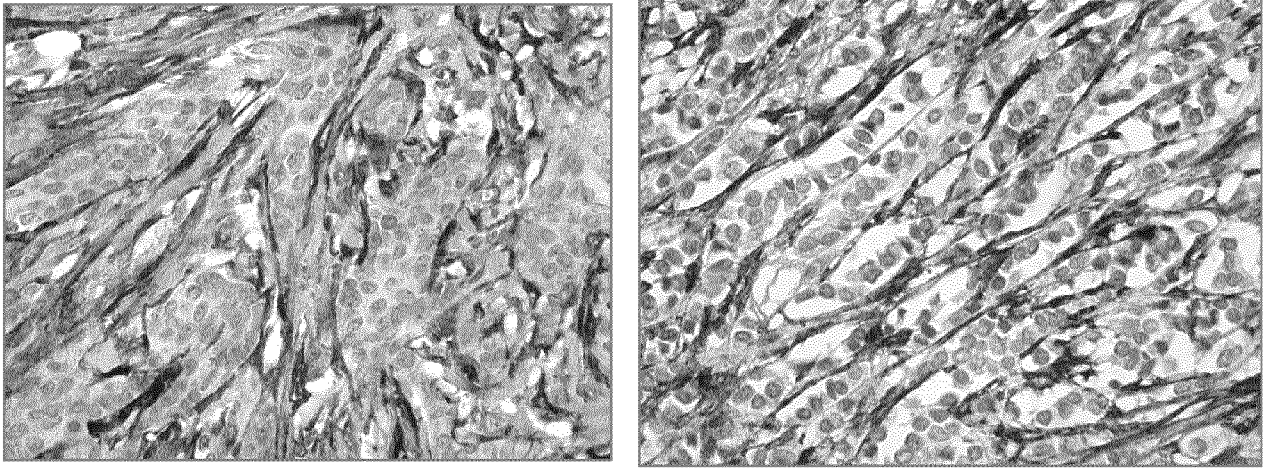


Figure 2

INTERNATIONAL SEARCH REPORT

International application No
PCT/EP2019/051716

A. CLASSIFICATION OF SUBJECT MATTER
INV. C07K16/28
ADD.
According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED
Minimum documentation searched (classification system followed by classification symbols)
C07K G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
EPO-Internal, BIOSIS, EMBASE, WPI Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 2016/133449 A1 (XINTELA AB [SE]) 25 August 2016 (2016-08-25) claims 1-57	1-27
X	WO 2008/075045 A1 (CARTELA R & D AB [SE]; LUNDGREN-AAKERLUND EVY [SE]) 26 June 2008 (2008-06-26) claims 1-109	1-23,27
Y		24-26
X	WO 2008/075038 A1 (BIOINVENT INT AB [SE]; FRENDEUS BJOERN [SE]; CARLSSON ROLAND [SE]; JAN) 26 June 2008 (2008-06-26)	1-23,27
Y	page 7, paragraph 3; page 46, paragraph 5; page 37, paragraph 1; claims 1-47	24-26
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Further documents are listed in the continuation of Box C.

See patent family annex.

* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
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- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
- "&" document member of the same patent family

Date of the actual completion of the international search 3 July 2019	Date of mailing of the international search report 11/07/2019
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Name and mailing address of the ISA/ European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Fax: (+31-70) 340-3016	Authorized officer Klee, Barbara
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INTERNATIONAL SEARCH REPORT

International application No
PCT/EP2019/051716

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	WO 2017/069627 A1 (UNIV TWENTE [NL]; PRAKASH JAI [NL]) 27 April 2017 (2017-04-27) page 2, paragraph 2 - page 5, paragraph 3; claims 1,17-24 page 1, paragraph 4 page 41, paragraph 3 -----	24-26

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No

PCT/EP2019/051716

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