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NANOVECTORS FOR PENETRATING BRAIN TUMOR TISSUES TO CONDUCT GENE THERAPY

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This U.S. Non Provisional Application claims priority to U.S. Provisional application, Serial No. 62/072,125, filed October 29, 2014, which is hereby incorporated in its entirety by reference.

STATEMENT REGARDING SPONSORED RESEARCH

[0002] Not applicable.

TECHNICAL FIELD

[0003] This disclosure relates to methods of selectively targeting cells with a therapeutic agent. More specifically, it relates to methods of targeting cells, tumors, and solid tumors using nanospears comprising magnetized carbon nanotubes and therapeutic agents. Even more specifically this disclosure relates to methods of using a non-viral gene vector to treat a solid tumor (such as glioblastoma), wherein a nanospear is employed to deliver the vector directly to the cellular target.

BACKGROUND

[0004] Glioblastoma (GBM) is an incurable form of brain cancer with poor prognosis. Current treatments of GBM have a low success rate due to a number of reasons including: the non-specific cell toxicity of current treatments, radio exposure of healthy cells, insufficient transport of drugs across blood-brain-barrier, heterogenic GBM of the tumor, and the highly infiltrative nature of GBM cells. Current drug delivery approaches typically leave drug molecules to passively diffuse to a target after their release into the subject, therefore resulting in sub-optimal delivery to deep and solid tissues, such as is typical in GBM deep tissue tumors. Viral vectors (that are used to deliver genetic material into cells) display high delivery efficiency but exhibit no target selectivity in their infection and may trigger immunogenic responses and oncogenesis concerns. Therefore a method of selectively and efficiently targeting deep or solid tumors, such as but not limited to GBM, is an unmet need in the art.

BRIEF SUMMARY

[0005] In an effort to address such unmet needs as described above, disclosed herein, is a non-viral gene therapy vector. In some embodiments the non-viral gene therapy vector may function with the efficiency of viral infection. In some embodiments herein disclosed is a therapeutic composition comprising: a nanospear, wherein the

nanospear comprises: a carbon nanotube (CNT); a magnetic particle; and a therapeutic agent, wherein the nanospear comprises a polymer coating, wherein the coating comprises said therapeutic agent, in some embodiments the therapeutic agent is selected from the group comprising: Temozolomide, BCNU, Irinotecan, Carboplatin, Cisplatin cpt-11, Taxol, Methotrexate, a non-viral gene vector, or combinations thereof, in other embodiments the therapeutic agent is a non-viral gene vector that comprises a transgene plasmid, wherein the plasmid comprises miRNA-124 target sites that suppress the mRNA of the anti-neural function protein SCP1 (small C-terminal domain phosphatase 1) In some embodiments, the therapeutic composition comprises a non-viral gene vector which comprises a transgene plasmid, wherein said plasmid further comprises miRNA-124 target sites.

[0006] In another embodiment, a method of selectively targeting a tumor with a therapeutic agent is described, the method comprises (a) targeting the tumor with a nanospear; wherein the nanospear is coated with a polymer and wherein the polymer encapsulates a therapeutic agent, the nanospear further comprises a magnetic particle and a block chain linker, wherein the block chain linker bonds the therapeutic agent to the polymer; (b) sensing the presence of the tumor; wherein sensing is by biorecognition of enzymatic activity associated with tumorigenesis; c) enzymatically degrading the block chain linker; wherein the tumor comprises an enzyme that degrades the block chain linker; and (c) releasing the therapeutic agent from the nanospear, wherein the agent is selective for the tumor. In another embodiment the tumor is comprised of tumor tissue; blood vessels that surround the tumor tissue; and tumor cells. In some further embodiments, the nanospears concentrate within the tumor; and in a still further embodiment the nanospears after step (a) further penetrate the tumor. In another embodiment of selectively targeting a tumor, the enzyme in step (c) comprises a matrix metalloproteinase 2, a metalloproteinase 9, or a combination thereof; in some embodiments the therapeutic agent is selected from one of more chemotherapy drugs to treat heterogenous tumor cells. In a further embodiment the nanospear penetrates at least a first layer of tumor cells.

[0007] In some embodiments of the method of selectively targeting a tumor with a therapeutic agent, the therapeutic agent is: a drug molecule; a non-viral gene therapy vector, a molecule that reduces the growth of said tumor; a molecule that induced apoptosis, an imaging agent, a molecule that inhibits growth of said tumor; or a combination thereof. In other embodiments the tumor is a solid tissue tumor, and in a

further embodiment, the tumor is glioblastoma (GBM). In some embodiments of the method of selectively targeting a tumor with a therapeutic agent the targeting comprises subjecting the nanospear is a magnetic force, in a further embodiment the force is by produced by a Halbach magnet.

[0008] In one embodiment disclosed herein, is a therapeutic method of treating a subject, wherein the subject comprises a tumor, and the method comprises administering to the subject a nanospear, subjecting the nanospear to a magnetic force, wherein the magnetic force guides the nanospear, localizing the nanospear at the tumor, penetrating the tumor, and releasing a therapeutic agent from the nanospear. In some embodiments, administering is by intravenous injection or subcutaneous injection.

[0009] In one embodiment disclosed herein, is a method of selectively targeting a cell with a therapeutic agent, the method comprising: targeting a cell with a nanospear, puncturing the cell with the nanospear; releasing a therapeutic agent from the nanospear, wherein the therapeutic agent enters the cell and thereby effecting the growth of the cell. In some embodiments described herein, the cell comprises: a multilayer cell culture, a 3D neuron cultures, a spheroid, a GBM tumor tissue, or combinations thereof. In some embodiments, effecting the growth of a cell comprises at least one of inducing cell growth stasis or inhibition, inhibition of molecular pathways, cellular mechanisms, or by cell death/apoptosis.

[0010] The foregoing has broadly outlined certain features exemplary embodiments of the present disclosure in order that the detailed description that follows may be better understood. It should be appreciated by those skilled in the art that the conception and the specific embodiments disclosed may be readily utilized as a basis for modifying or designing other methods and structures for carrying out the same purposes of the disclosure as claimed below. It should also be realized by those skilled in the art that such equivalent constructions do not depart from the spirit and scope of the disclosure as set forth in the appended claims.

BRIEF DESCRIPTION OF THE DRAWINGS

[0011] For a more complete understanding of the present disclosure, reference will now be made to the accompanying figures:

[0012] Fig. 1 (A-G) illustrates an embodiment of surface modification and characterization of CNTs as described herein. Fig.1A shows a schematic illustration of the surface modification of CNTs: wherein Ni-coat CNTs array by e-beam

evaporation of Ni on an aligned CNTs array, and poly-L-tyrosine coating by electropolymerization. Fig. 1B depicts recording of cyclic voltammetry (CV) for electropolymerization of L-tyrosine on CNTs, with CNTs and Ag/AgCl as the working and reference electrodes, respectively. Fig. 1C shows deposition charge (Q) by integration of each cycle of CV versus the cycles. (D) SEM image of Ni-coated CNTs. Fig. 1E shows TEM images of Ni-coated CNTs with surface modified by poly-L-tyrosine coating, as indicated by the red arrow; inset: a low magnification image. Fig. 1F shows magnetization measurement of Ni-coated CNTs. Fig. 1G shows aqueous suspension of the magnetized CNTs.

DETAILED DESCRIPTION OF THE DISCLOSURE

[0013] The following discussion is directed to various exemplary embodiments of the disclosure. However, the embodiments disclosed should not be interpreted, or otherwise used, as limiting the scope of the disclosure, including the claims. In addition, one skilled in the art will understand that the following description has broad application, and the discussion of any embodiment is meant only to be exemplary of that embodiment, and that the scope of this disclosure, including the claims, is not limited to that embodiment.

[0014] In the following discussion and in the claims, the terms “including” and “comprising” are used in an open-ended fashion, and thus should be interpreted to mean “including, but not limited to....” Also, the term “couple” or “couples” is intended to mean either an indirect or direct connection. Thus, if a first device couples to a second device, that connection may be through a direct engagement between the two devices, or through an indirect connection via other intermediate devices and connections. As used herein, the term “about,” when used in conjunction with a percentage or other numerical amount, means plus or minus 10% of that percentage or other numerical amount. For example, the term “about 80%,” would encompass 80% plus or minus 8%.

[0015] In some embodiments described herein, a nanospear methodology is disclosed, wherein a gene-bearing nanospear structure may be injected into a target, wherein the target may be isolated cells *in vitro*, or in a subject (or patient) and *in vivo*. The administration of such nanospears may therefore be intravenous or subcutaneous. The nanospears comprise magnetized carbon nanotubes that are coated with a biocompatible polymer that may be linked or chemically bonded or attached to a therapeutic agent. The nanospear may be guided by a magnetic source

to a cellular target such as GBM, where the specific localized tumor environment may induce cleavage of the linker between the polymer and the therapeutic agent, thereby delivering the agent directly and specifically to the localized target cells that comprise for example a GBM tumor.

[0016] In some embodiments the nanospears will be concentrated at the blood vessels surrounding the tumor tissue, and will further spear, and may penetrate into the cells of the tumor tissue itself. Such spears, in some embodiments therefore penetrate the cells of the tumor tissue layer by layer. In some embodiments Gene therapy molecules are disseminated into such cells, in some embodiments the trajectory of the nanospear and penetration of the cells within that trajectory facilitates the dissemination of the gene therapy molecules into the cells.

EXAMPLES:

[0017] Production of nanospears: In some embodiments the nanospears disclosed herein comprise: biocompatible and biodegradable materials (coating polymers), iron oxide magnetic nanoparticles, carbon nanotubes and therapeutic agents. Therapeutic agents delivered by the nanospears include but are not limited to chemotherapy drugs, for example those selective for GBM, such as Temozolomide, BCNU, Irinotecan, Carboplatin, Cisplatin cpt-11, Taxol, Methotrexate. The drugs (therapeutic agents) are entrapped in a polymer coating outside the magnetic bmCNT. (carbon-nanotube). In some embodiments the coating on the carbon nanotube may be produced by electropolymerization, or self-assembly by static electrical charge, or by micelle interaction. Examples of the polymer coating include (but are not limited to) polyphenol, polytyrosine, polyaniline, and polypyrrole. In some embodiments, the polymer used for drug encapsulation of the nanospear may be designed to response to a specific environmental change that is related only to cancer, so that the drug may be released only around the cancer target. For example, in some embodiments, a block polymer that is sensitive to the cancer related enzymatic activity maybe incorporated in the nanostructures to render such a feature of targeted release. In some embodiments a chemical linker in the form of a block chain linker links the polymer coating to the therapeutic agent, when the nanospear comes into contact with matrix metalloproteinases found in the vicinity of GMB tumors, matrix metalloproteinase 2, a metalloproteinase 9, or a combination thereof, digest the linker and release the therapeutic agent, thereby allowing delivery of the agent to the target tumor cell. Examples of block chain linkers include but are not limited to

KRGPQGIWGQDRCGR (Seq.1), KRGPQGIAGQDRCGR (Seq.2), KRGDQGIAGFDRCGR (Seq. 3) and GPQGIFGQ (Seq.4).

[0018] In some embodiments, nanospears comprise carbon nanotubes (CNTs), a magnetic metal and an outer polymeric layer. In an embodiment, the magnetic metal may comprise magnetic particles and a magnetic metal layer. In an embodiment, the magnetic metal may comprise nickel, Iron (Fe), Iron oxide, superparamagnetic materials, and the like, or combinations thereof.

[0019] In an embodiment, the CNTs have a rod shape or cylindrical geometry. In such embodiment, the CNTs may be characterized by having two ends, which correspond to the ends of the rod or cylinder. Further, in such an embodiment, the magnetic metal may coat only one end of the nanospears. Coating only one end of the CNTs with a magnetic material such as, magnetic metal) ensures that the resulting nanospears could be oriented in the magnetic field, and could consequently be "speared" in the desired direction. For purposes of the disclosure herein, the terms "spear" or "spearing," and "nanospear" or "nanospearing," may be used interchangeably and all these related terms refer to a directed movement of a magnetized nanostructure (MNS) within and/or through a bioentity (such as, a single cell, distinct cell layers, a clump of cells, a piece of live tissue, etc.). Non-limiting examples of MNS include nanospear, nanotube, nanoparticle, nanorod, nanowire, nanohorn, nanostar, nanovesicle, nanocapsule that may comprise inorganic, organic, polymeric, metallic, non-metallic, oxide, alloy, or composite materials, and the like, or combinations thereof.

[0020] In some embodiments, the nanospears may be characterized by a nanospear length of from about 0.5 mm to about 5 mm, alternatively from about 1 mm to about 3 mm, or alternatively from about 1 mm to about 2 mm.

[0021] In an embodiment, the nanospears may be characterized by a nanospear diameter of from about 50 nm to about 300 nm, alternatively from about 75 nm to about 200 nm, or alternatively from about 75 nm to about 125 nm.

[0022] In an embodiment, a method of preparing nanospears may comprise growing carbon nanotubes; coating the carbon nanotubes with a magnetic metal to yield nanospears, wherein the magnetic metal may comprise nickel; and coating the nanospears with an outer polymeric layer, wherein the outer polymeric layer may be hydrophilic and biocompatible.

[0023] In some embodiments, the CNTs may be grown by using any suitable methodology (such as for example those disclosed in U.S. Provisional Patent application 62/032,996 incorporated herein in its entirety). In an embodiment, the CNTs may be grown by using a plasma-enhanced chemical vapor deposition system, as described in more detail in science 1998, 282(5391):1105-1107 (27), which is also incorporated by reference herein in its entirety. The growth of the CNTs may result in straight-aligned CNTs with magnetic nickel (Ni) particles enclosed at the tips (or in further embodiments as described herein with Iron, or Iron oxide enclosed at the tips) which make the CNTs magnetically drivable. In an embodiment, a layer of a magnetic metal (such as, Ni or Fe) may be deposited along the surface of individual CNTs by using any suitable methodology, such as for example e-beam evaporation. In an embodiment, the layer of magnetic metal may enhance the magnetization, thereby leading to an enhanced magnetic force, wherein such magnetic force may be required for cell penetration. The magnetic metal may exacerbate toxicity and hydrophobicity of the nanospears for biological applications.

[0024] In an embodiment, the magnetic metal layer may be characterized by a magnetic metal layer thickness of from about 5 nm to about 50 nm, alternatively from about 10 nm to about 30 nm, or alternatively from about 15 nm to about 25 nm.

[0025] In an embodiment, the nanospears (such as, nanospears array) may be further coated with the outer polymeric layer by using any suitable methodology, such as for example electropolymerization, thereby reducing the toxicity of metal (such as, Ni)-coated CNTs. In such embodiment, the outer polymeric layer may comprise poly-L-tyrosine. In an embodiment, the outer polymeric layer may be hydrophilic, thereby rendering the nanospears hydrophilic. In an embodiment, the outer polymeric layer may be biocompatible, thereby rendering the nanospears biocompatible.

[0026] In an embodiment, the outer polymeric layer may be characterized by an outer polymeric layer thickness of from about 1 nm to about 50 nm, alternatively from about 2 nm to about 25 nm, or alternatively from about 5 nm to about 15 nm. In an embodiment, the nanospears (such as, nanospears array) may be connected to an electrochemistry system to conduct electropolymerization of a monomer (such as, L-tyrosine) on the surfaces of the nanospears, as illustrated in figure 1. It has been previously shown that electropolymerization of L-tyrosine may be a feasible way to create a hydrophilic and biocompatible film that is suitable in diverse biological

applications, as described in more detail in *biomacromolecules* 2005, 6(3):1698-1706 and *anal biochem* 2009, 384(1):86-95 (28, 29), each of which is incorporated by reference herein in its entirety. In an embodiment, electropolymerization of γ -tyrosine into poly- γ -tyrosine may comprise cyclic voltammetry.

Nanospear Targets:

[0027] *In vitro* targets may be prepared for analysis of the therapeutic effects and compositions of embodiments of the nanospears herein provided. For example, brain tumor cells may be enzymatically digested and dispersed in a petri dish and maintained in a CO₂ incubator at 37°C, under controlled CO₂ concentration and saturated humidity. The mono-dispersed cells may be re-digested and re-suspended, and transferred to 3D culture hydrogel. Some embodiments herein described use Mebiol Gel (Cosmo Bio Co., Ltd), which is liquidized poly(N-isopropylacrylamide) and poly(ethylene glycol) hydrogel in a cell culture medium on ice; the cells are then mixed with the hydrogel at low temperature (2-10°C); (3) the hydrogel was warmed to 37°C to solidify the hydrogel and maintain the cell in 3D scaffold. In some embodiments the resultant 3D culture environment provides conditions for cell proliferation, cell communication, gas and mass exchange, and maintains the specific location of the cells. The hydrogel may be kept in a cell culture plate or petri dish, wherein the cancer may be produced in days. Epithelial cells may be cultured as a non-cancer cell control to evaluate the selectivity of the targeted release. Further, in some embodiments described herein the environmental selectivity and stability of the polymeric nanospear may be measured in the artificial environment provided by buffer, or in the cultured normal and cancer cells. In some embodiments, the drug-bearing bmCNT, i.e. nanospears described herein were suspended in culture medium and applied to the hydrogel containing the spheroids. A magnet (Halbach) may be placed under the container (comprising the spheroids) to pull the nanospears into the hydrogel and the spheroid, wherein drug molecule may be released directly into the target cell (such as the solid tumor *in vivo*). Fluorescent molecules may be used instead of therapeutic agents to further analyze the progress of nanospear motion in the hydrogel visualized with a confocal microscope. In some embodiments the pharmacokinetics of the targeted release of the therapeutic agents from the nanospears may be evaluated by measuring fluorescence leakage of fluorescent surrogate under artificial environment with TIRF microscopy. The drugs molecules may be loaded into the nanospears at different

molar concentrations such as but not limited to: 1 μ M, 10 μ M, 100 μ M, 1 mM, 10 mM, 20 mM, 40 mM, and the cells may be maintained in the hydrogel for a selected time periods without disturbance for 0.5, 1, 2, 4, 8, 16, 24, 48, 72 hours for example.

[0028] The cells may be harvest into the liquid medium by cooling to 2-10°C. With this hydrogel feature, the dose response and time lapse of response may be obtained by propidium iodide staining. In further embodiments the nanospear may be electrospun, comprise nanofibers or nanobeads and may be produced with the polymers, drugs and magnetic particles described herein.

[0029] In some embodiments, nanospears may be administered in vivo or in vitro to selectively conduct GBM suppression by nanospear gene vector. The transgene plasmid may comprise miRNA-124 target sites, so that transgene expression is prevented in neuron, rather in glia. In some embodiments a mixed culture of neuron and glial in the form of cell layers, spheroids may be produced, thereby characterizing the vector dosage based on the viability of glial cells and neurons. In some embodiments a xenograft animal model may be utilized along with I.V. administration to assess the shrinkage of tumor size. Some embodiments of the method herein described showed high efficient plasmid transfection in cultured primary cortical neurons, and in further embodiments biocompatible nanospears thoroughly penetrated mammalian cell bodies

[0030] While exemplary embodiments of the disclosure have been shown and described, modifications thereof may be made by one skilled in the art without departing from the spirit and teachings of those embodiments. The embodiments described herein are exemplary only, and are not intended to be limiting. Many variations and modifications of the disclosed embodiments are possible and are within the scope of the claimed disclosure. Where numerical ranges or limitations are expressly stated, such express ranges or limitations should be understood to include iterative ranges or limitations of like magnitude falling within the expressly stated ranges or limitations (such as., from about 1 to about 10 includes, 2, 3, 4, etc.; greater than 0.10 includes 0.11, 0.12, 0.13, etc.). For example, whenever a numerical range with a lower limit, R_l , and an upper limit, R_u , is disclosed, any number falling within the range is specifically disclosed. In particular, the following numbers within the range are specifically disclosed: $R = R_l + k * (R_u - R_l)$, wherein k is a variable ranging from 1 percent to 100 percent with a 1 percent increment, i.e., k is 1 percent, 2 percent, 3 percent, 4 percent, 5 percent, 50 percent, 51 percent, 52

percent, 95 percent, 96 percent, 97 percent, 98 percent, 99 percent, or 100 percent. Moreover, any numerical range defined by two R numbers as defined in the above is also specifically disclosed. Use of the term "optionally" with respect to any element of a claim is intended to mean that the subject element is required, or alternatively, is not required. Both alternatives are intended to be within the scope of the claim. Use of broader terms such as comprises, includes, having, etc. should be understood to provide support for narrower terms such as consisting of, consisting essentially of, comprised substantially of, etc.

[0031] Accordingly, the scope of protection is not limited by the description set out above but is only limited by the claims which follow, that scope including all equivalents of the subject matter of the claims. Each and every claim is incorporated into the specification as an embodiment of the present disclosure. Thus, the claims are a further description and are an addition to the embodiments of the present disclosure. The disclosures of all patents, patent applications, and publications cited herein are hereby incorporated by reference, to the extent that they provide exemplary, procedural or other details supplementary to those set forth herein.

CLAIMS

What is claimed is:

1. A therapeutic composition comprising:
 - a nanospear, wherein said nanospear comprises:
 - a carbon nanotube (CNT);
 - a magnetic particle; and
 - a therapeutic agent, wherein said CNT comprises a polymer coating, and wherein said coating comprises said therapeutic agent.
2. The composition of claim 1, wherein said therapeutic agent is selected from the group comprising: Temozolomide, BCNU, Irinotecan, Carboplatin, Cisplatin cpt-11, Taxol, Methotrexate, a non-viral gene vector, or combinations thereof.
3. The composition of claim 2, wherein said non-viral gene vector comprises a transgene plasmid, wherein said plasmid comprises miRNA-124 target sites.
4. A method of selectively targeting a tumor with a therapeutic agent, the method comprising:
 - (a) targeting said tumor with a nanospear; wherein said nanospear comprises: a carbon nanotube coated with a polymer, wherein said polymer encapsulates a therapeutic agent, and wherein said therapeutic agent and said polymer are linked by a block chain linker, and wherein said nanospear comprises a magnetic particle;
 - (b) sensing the presence of the tumor;
 - (c) enzymatically degrading said block chain linker; wherein said tumor comprises an enzyme that degrades said polymer; and
 - (d) releasing said therapeutic agent from said nanospear, wherein said agent is selective for said tumor.
5. The method of claim 4 wherein said tumor is comprised tumor tissue; blood vessels that surround said tumor tissue; and tumor cells.

6. The method of claim 4, wherein said nanospears concentrate within said tumor.
7. The method of claim 4, wherein said nanospears after step (a) further penetrate said tumor.
8. The method of claim 4, wherein said enzyme in step (c) comprises a matrix metalloproteinase 2, a metalloproteinase 9, or a combination thereof.
9. The method of claim 4, wherein said therapeutic agent is selected from one of more chemotherapy drugs that target heterogenous tumor cells.
10. The method of claim 5, wherein said nanospear penetrates at least a first layer of tumor cells.
11. The method of claim 4, wherein said therapeutic agent is a drug molecule; a non-viral gene therapy vector, a molecule that reduces the growth of said tumor; a molecule that induced apoptosis, an imaging agent, a molecule that inhibits growth of said tumor; or a combination thereof.
12. The method of claim 4, wherein said tumor is a solid tissue tumor.
13. The method of claim 12, wherein said tumor is glioblastoma (GBM).
14. The method of claim 4, wherein said targeting comprises subjecting said nanospear to a magnetic force.
15. The method of claim 14, wherein said force is produced by a Halbach magnet.
16. A therapeutic method of treating a subject, wherein the subject comprises a tumor, said method comprising:
administering to said subject a nanospear;

subjecting the nanospear to a magnetic force, wherein said magnetic force guides said nanospear;

localizing said nanospear at said tumor;

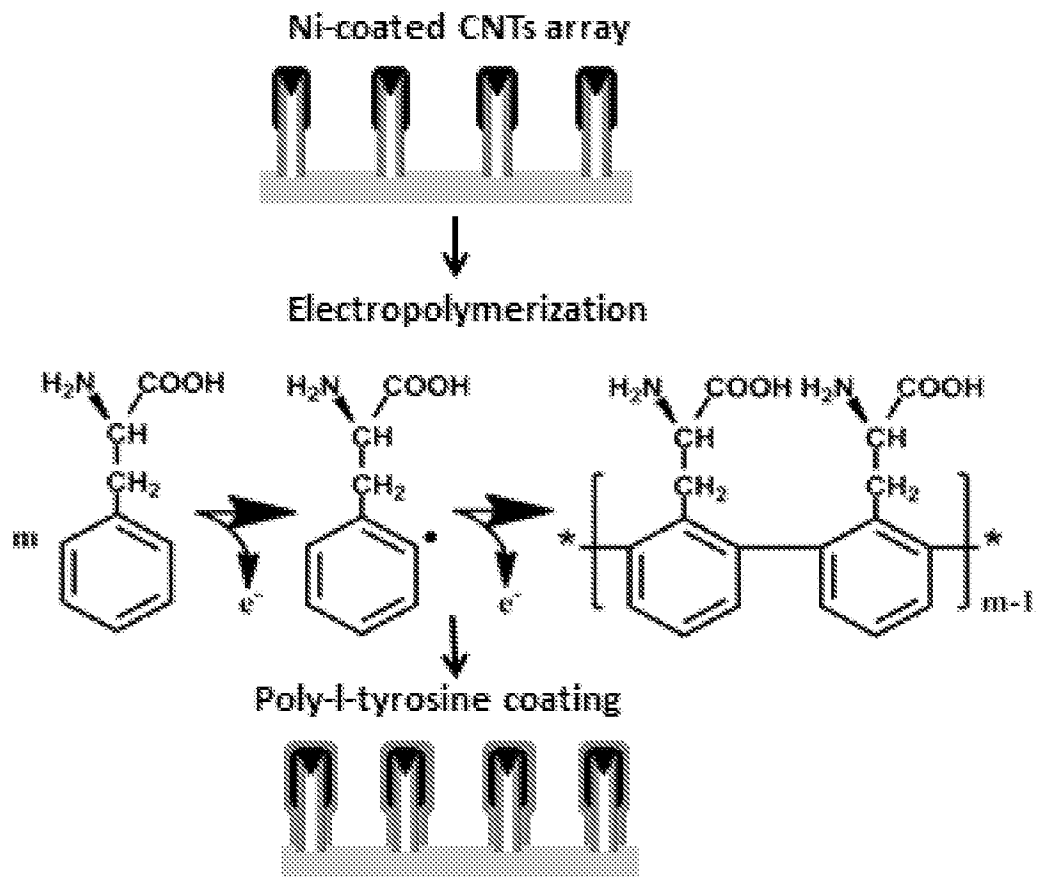
penetrating said tumor; and

releasing a therapeutic agent from said nanospear.

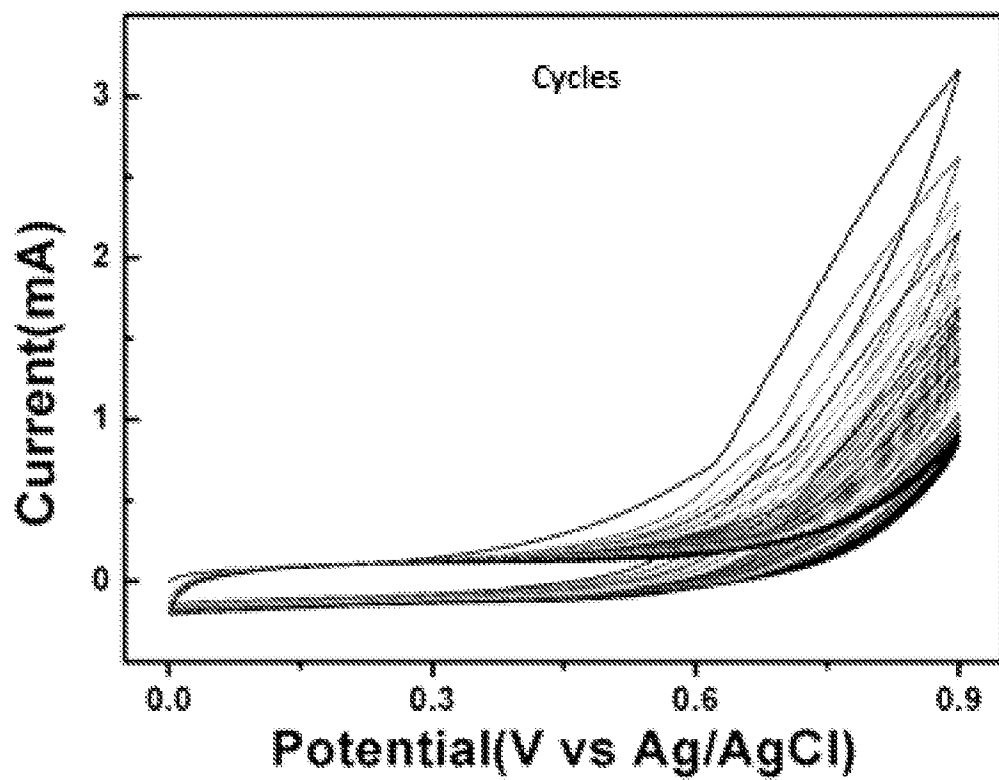
17. The method of claim 16, wherein administering is by intravenous injection or subcutaneous injection.
18. A method of selectively targeting a cell with a therapeutic agent, said method comprising:
 - targeting a cell with a nanospear,
 - puncturing said cell with said nanospear;
 - releasing a therapeutic agent from said nanospear, wherein said therapeutic agent effects cell growth.
19. The method of claim 4, wherein said cell comprises: a multilayer cell culture, a 3D neuron cultures, a spheroid, a GBM tumor tissue, or combinations thereof.
20. The method of claim 18, wherein effects cell growth is by at least one of: inducing cell stasis, cell death, or inhibition of cellular mechanisms.

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Fig. 1A

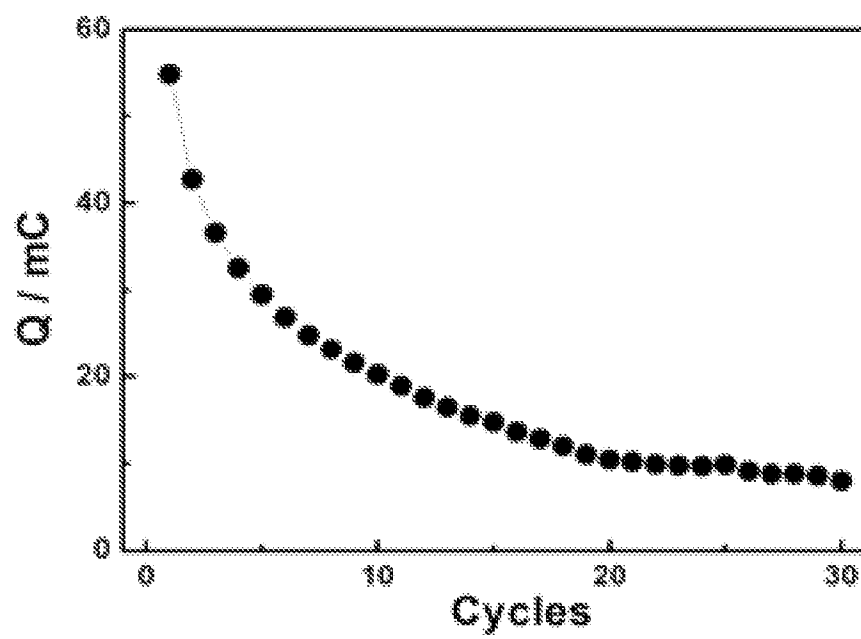


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Fig. 1B

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Fig. 1C



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Fig. 1D

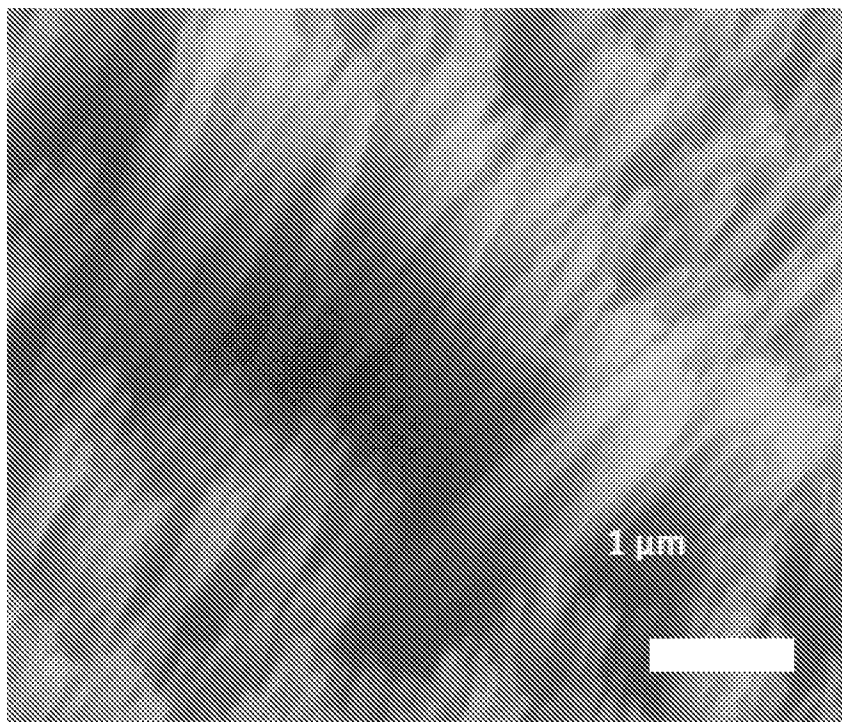


Fig. 1E

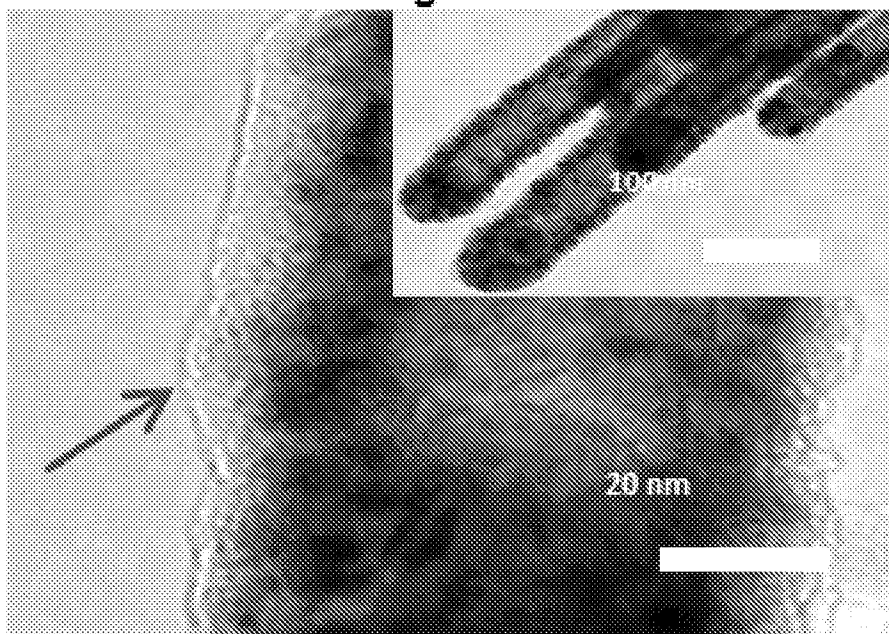
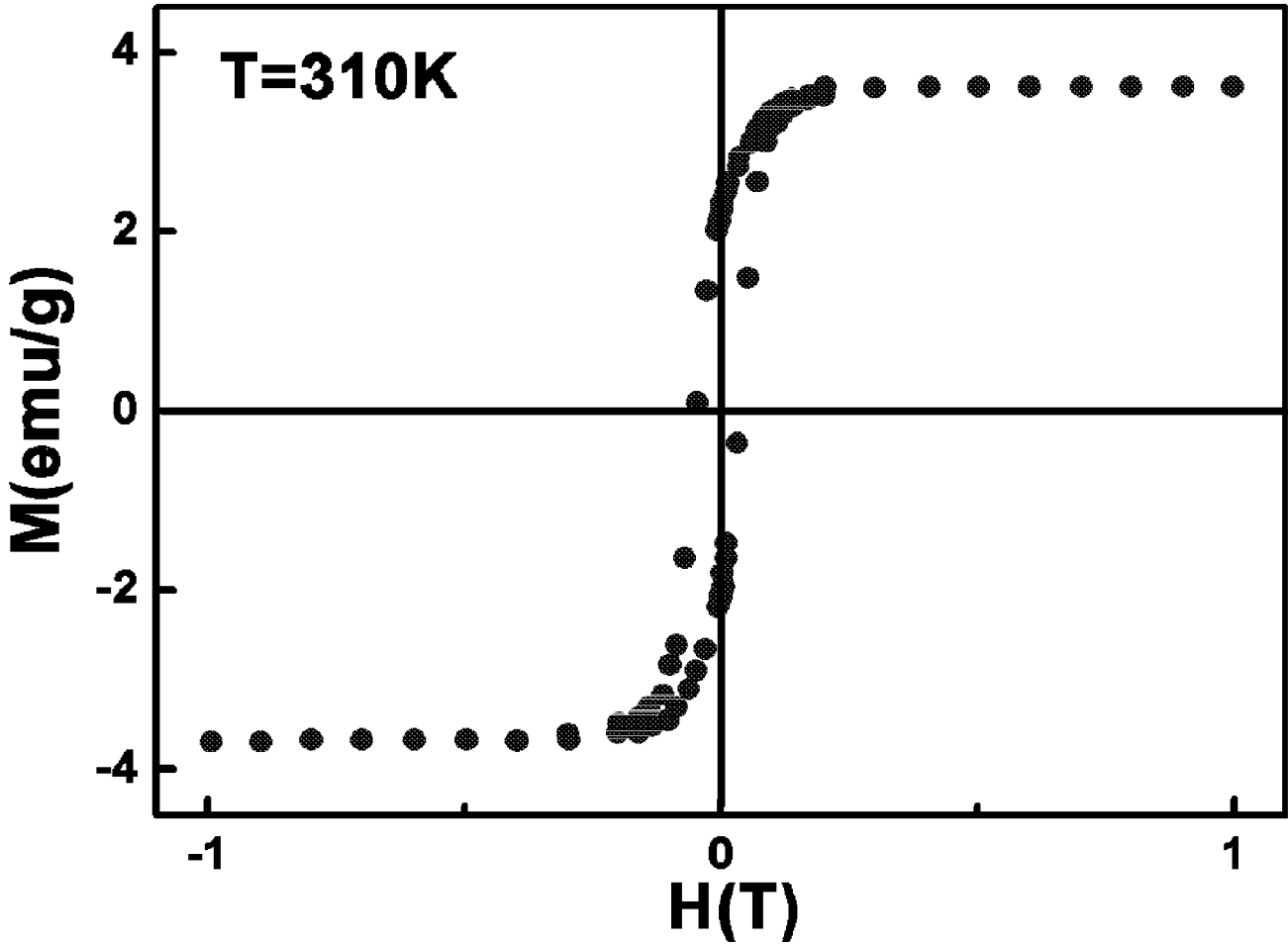
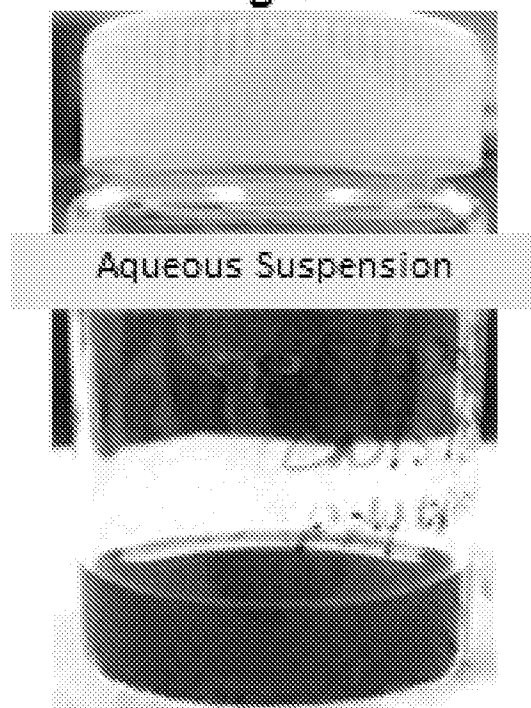


Fig. 1F



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Fig. 1G



A. CLASSIFICATION OF SUBJECT MATTER

A61K 9/16(2006.01)i, A61K 47/30(2006.01)i, A61K 31/4188(2006.01)i, A61K 31/4738(2006.01)i, A61K 31/4741(2006.01)i, A61K 31/337(2006.01)i, A61K 33/44(2006.01)i, A61K 48/00(2006.01)i, A61P 35/00(2006.01)i

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

A61K 9/16; A61K 47/30; A61K 31/4188; A61K 31/4738; A61K 31/4741; A61K 31/337; A61K 33/44; A61K 48/00; A61P 35/00

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Korean utility models and applications for utility models

Japanese utility models and applications for utility models

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

eKOMPASS(KIPO internal) & Keywords: drug delivery, gene delivery, nanospear, nanotube, magnetic, polymer coating, cancer, glioblastoma, cell penetration

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	LU, Y.-J. et al., 'Dual targeted delivery of doxorubicin to cancer cells using folate-conjugated magnetic multi-walled carbon nanotubes', Colloids and Surfaces B: Biointerfaces, 2012, Vol.89, pp.1-9 See abstract; pages 2, 3, 5, 7; and figure 1.	1-3
A	MOORE, T. L. et al., 'Multifunctional polymer-coated carbon nanotubes for safe drug delivery', Particle & Particle Systems Characterization, 2013, Vol.30, pp.365-373 See abstract; page 365.	1-3
A	CAI, D. et al., 'Nanospearing-biomolecule delivery and its biocompatibility', Nanomaterials for Application in Medicine and Biology, ISBN: 978-1-4020-6827-0, 2008, pp.81-92 See abstract.	1-3
A	MOORE, T. L. et al., 'Multilayered polymer-coated carbon nanotubes to deliver dasatinib', Molecular Pharmaceutics, Epub.2013, Vol.11, pp.276-282 See abstract; and page 277.	1-3



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

29 January 2016 (29.01.2016)

Date of mailing of the international search report

29 January 2016 (29.01.2016)

Name and mailing address of the ISA/KR

International Application Division

Korean Intellectual Property Office

189 Cheongsa-ro, Seo-gu, Daejeon, 35208, Republic of Korea

Facsimile No. +82-42-472-7140

Authorized officer

LEE, Jeong A

Telephone No. +82-42-481-8740



INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2015/057828

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>PEREZ-MARTINEZ, F. C. et al., 'Barriers to non-viral vector-mediated gene delivery in the nervous system', Pharmaceutical Research, 2011, Vol.28, pp.1843-1858 See abstract; and pages 1853-1854</p>	1-3

Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.: 4-20
because they relate to subject matter not required to be searched by this Authority, namely:
Claims 4-20 pertain to methods for treatment of the human body by surgery or therapy, and thus relate to a subject matter which this International Searching Authority is not required, under PCT Article 17(2)(a)(i) and PCT Rule 39.1(iv), to search.
2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box No. III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fees, this Authority did not invite payment of any additional fees.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee.
- ☐ The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation.
- ☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No.

PCT/US2015/057828Patent document
cited in search reportPublication
datePatent family
member(s)Publication
date

None