

FORM 1

623131

SPRUSON & FERGUSON

COMMONWEALTH OF AUSTRALIA
PATENTS ACT 1952
APPLICATION FOR A STANDARD PATENT

F Hoffmann-La Roche & Co Aktiengesellschaft, of Grenzacherstrasse 124-184,
4002 Basle, SWITZERLAND, hereby apply for the grant of a standard patent for
an invention entitled:

Tyr-Peptide Analogs

which is described in the accompanying complete specification.

Details of basic application(s):-

<u>Basic Applic. No:</u>	<u>Country:</u>	<u>Application Date:</u>
185228	US	25 April 1988
311872	US	21 February 1989

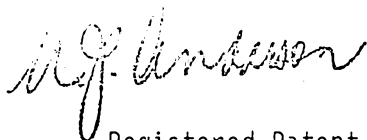
The address for service is:-

Spruson & Ferguson
Patent Attorneys
Level 33 St Martins Tower
31 Market Street
Sydney New South Wales Australia

DATED this TWENTIETH day of APRIL 1989

F Hoffmann-La Roche & Co Aktiengesellschaft

By:



Registered Patent Attorney

TO: THE COMMISSIONER OF PATENTS
OUR REF: 93154
S&F CODE: 55541

REPRINT OF RECEIPT

3006999 24/04/89

5845/2

COMMONWEALTH OF AUSTRALIA

THE PATENTS ACT 1952

DECLARATION IN SUPPORT OF A
CONVENTION APPLICATION FOR A PATENTIn support of the Convention Application made for a
patent for an invention entitled:AUSTRALIA
CONVENTION
STANDARD
& PETTY PATENT
DECLARATION~~4105/116~~

Title of Invention

Tyr-Peptide Analogs

Full name(s) and
address(es) of
Declarant(s)

I Roland Borer
of 10 Stockackerstrasse, CH-4153 Reinach, Switzerland

do solemnly and sincerely declare as follows:-

Full name(s) of
Applicant(s)

1. I am authorised by
F. HOFFMANN-LA ROCHE & CO. Aktiengesellschaft,
(of 124-184 Grenzacherstrasse, Basle, Switzerland,)

the applicant(s) for the patent to make this declaration on
its/their behalf.

2. The basic application(s) as defined by Section 141 of the
Act was/were made

Basic Country(ies)

in the United States of America

Priority Date(s)

on April 25, 1988 and on February 21, 1989

Basic Applicant(s)

by ☐ F. HOFFMANN-LA ROCHE & CO., Aktiengesellschaft☒ the inventor(s) cited in paragraph 3.

3.

Full name(s) and
address(es) of
inventor(s)

- 1) Waleed Danho
118 Bunker Hill Road, Wayne, N.J. 07470, U.S.A.
2) Jefferson Wright Tilley
19 Evergreen Drive, North Caldwell, N.J. 07006, U.S.A.
3) Joseph Triscari
22 Evergreen Drive, North Caldwell, N.J. 07006, U.S.A.
4) Rolf Wagner
33 Madelyn Avenue, West Milford, N.J. 07480, U.S.A.

(respectively)-

is/are the actual inventor(s) of the invention and the facts upon
which the applicant(s) is/are entitled to make the application are
as follows:

- (X) the inventor(s) have assigned the invention to
Hoffmann-La Roche Inc., Nutley, USA,
who have re-assigned all their rights for Australia
to the Applicant.
() the Applicant is the assignee of the invention from
the inventor(s).

4. The basic application(s) referred to in paragraph 2 of this
Declaration was/were the first application(s) made in a Convention
country in respect of the invention(s) the subject of the application.

To:

Declared at Basle this 31st day of March 1989

The Commissioner of Patents,
COMMONWEALTH OF AUSTRALIASignature of Declarant(s)
Roland Borer

(12) PATENT ABRIDGMENT (11) Document No. AU-B-33359/89
(19) AUSTRALIAN PATENT OFFICE (10) Acceptance No. 623131

(54) Title
TYR-SULPHATE/PHOSPHATE CONTAINING PEPTIDES

International Patent Classification(s)
(51)⁴ C07K 007/06 A61K 037/24 C07C 057/34 C07C 057/42
C07C 057/58 C07C 057/60 C07C 069/65 C07C 101/20
C07C 101/28 C07C 101/34 C07C 103/46 C07C 103/84
C07C 125/00 C07D 209/48 C07D 257/04 C07D 403/10
C07C 069/612 C07C 069/618 C07C 125/065 C07D 207/404
C07D 207/408

(21) Application No. : 33359/89

(22) Application Date : 24.04.89

(30) Priority Data

(31) Number	(32) Date	(33) Country
185228	25.04.88	US UNITED STATES OF AMERICA
311872	21.02.89	US UNITED STATES OF AMERICA

(43) Publication Date : 26.10.89

(44) Publication Date of Accepted Application : 07.05.92

(71) Applicant(s)
F HOFFMANN-LA ROCHE & CO AKTIENGESELLSCHAFT

(72) Inventor(s)
WALEED DANHO; JEFFERSON WRIGHT TILLEY; JOSEPH TRISCARI; ROLF WAGNER

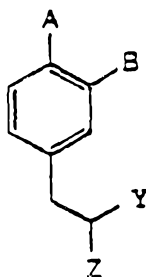
(74) Attorney or Agent
SPRUSON & FERGUSON, GPO Box 3898, SYDNEY NSW 2001

(56) Prior Art Documents
AU 49430/90 C07K 7/06
DE 2917603
GB 1309464

(57) ALSO INCLUDES INTERMEDIATES, AND THEIR USE AS APPETITE SUPPRESSANTS.

CLAIM

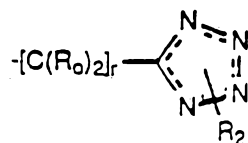
1. An analog of a tyrosine sulfate- or tyrosine phosphate residue(s) containing peptide wherein one or more of such residue(s) is/are replaced by (a) residue(s) of the formula



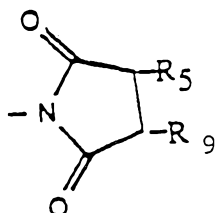
wherein one of A or B is selected from the group consisting of: $-[C(R_O)_2]_s-CO_2R_1$ or,

(11) AU-B-33359/89
(10) 623131

-2-



and the other of A or B is H;
Z is H, -NH_2 , -NH- , lower alkyl, -NHCOR_3 , $\text{-NHCO}_2\text{R}_4$,
or



R_0 is H or F;

R_1 is H, substituted or unsubstituted lower alkyl with the substituents selected from the group consisting of hydroxyl, halogen, or aryl;

R_2 is H, lower alkenyl, lower alkyl or lower alkyl substituted by 1 to 3 aryl groups;

R_3 is H, lower alkyl, alkyl substituted by one or two aryl groups or aryl;

R_4 is lower alkyl or alkyl substituted by one or two aryl groups;

R_5 and each independently H, lower alkyl,

(11) AU-B-33359/89
(10) 623131

-3-

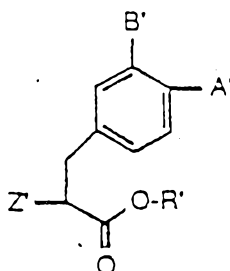
R_9 are or taken together and including the carbon atoms to which they are attached may form a six numbered ring which may be aromatic;

s and r are 0, 1 or 2;

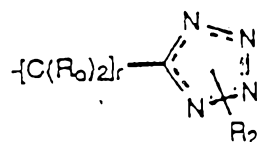
and Y is $-\text{COOH}$, alkoxycarbonyl, $-\text{CONH}_2$ or $-\text{CO}-$;

with the proviso that if Z is other than $-\text{NH}-$ then Y is $-\text{CO}-$; or if Y is other than $-\text{CO}-$ then Z is $-\text{NH}-$; and pharmaceutically acceptable salts thereof.

62. A compound when used for the preparation of an analog of a tyrosine sulphate- or tyrosine phosphate residue(s) containing peptide as defined in claim 1, said compound having the formula:



wherein one of A' or B' , is selected from $-\text{[C(R}_0\text{)]}_2\text{]-CO}_2\text{R}_1'$, or



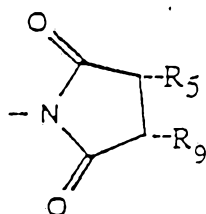
and the other of A' or B' is H wherein R_0 is H or F, R_1' is H, substituted or unsubstituted lower alkyl with the substituents selected from hydroxyl, halogen, or aryl, R_2 is H, lower alkenyl, lower alkyl or lower alkyl substituted by 1 to 3 aryl groups,

r is 0-2

s is 1-2

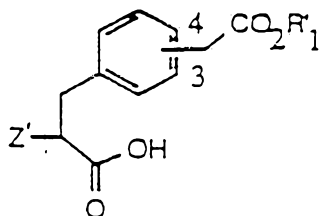
R' is H, lower alkyl or lower alkyl substituted by 1 to 3 aryl groups,

Z' is H, lower alkyl, $-\text{NH}_2$, $-\text{NHCOR}_3$, $-\text{KHCO}_2\text{R}_4$, or



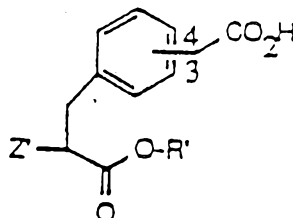
and R_3 and R_4 are H, lower alkyl or lower alkyl substituted by one, two or three aryl groups; and R_5 and R_9 are each independently H, lower alkyl, or taken together and including the carbon atoms to which they are attached may form a six membered ring which may be aromatic.

63. A compound when used for the preparation of an analog of a tyrosine sulphate- or tyrosine phosphate residue(s) containing peptide as defined in claim 1, said compound having the formula:



wherein Z' and R'_1 are as defined in claim 62.

64. A compound when used for the preparation of an analog of a tyrosine sulphate- or tyrosine phosphate residue(s) containing peptide as defined in claim 1, said compound having the formula:

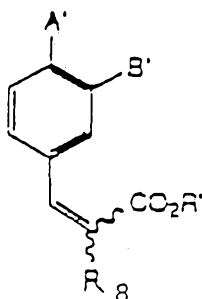


wherein R' and Z' are as defined in claim 62.

66. A compound when used for the preparation of an analog of a tyrosine sulphate- or tyrosine phosphate residue(s) containing peptide as defined in claim 1, said compound having the formula:

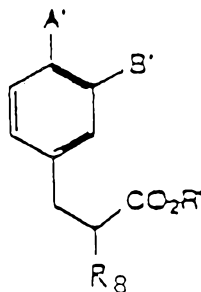
(11) AU-B-33359/89
(10) 623131

-5-



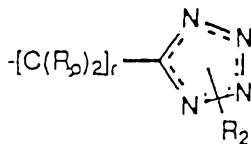
wherein A', B', and R' are as defined in claim 62 and R₈ is H or lower alkyl.

70. A compound when used for the preparation of an analog of a tyrosine sulphate- or tyrosine phosphate residue(s) containing peptide as defined in claim 1, said compound having the formula:



wherein A', B', and R' are as defined in the second embodiment and R₈ is H or lower alkyl.

71. The compound of claim 70 wherein A' is



and B' is H and R₀, r, and R₂ are as defined in claim 62.

623131
FORM 10

S & F Ref: 93154

COMMONWEALTH OF AUSTRALIA

PATENTS ACT 1952

COMPLETE SPECIFICATION

(ORIGINAL)

FOR OFFICE USE:

Class Int Class

Complete Specification Lodged:
Accepted:
Published:

Priority:

Related Art:

Name and Address
of Applicant: F Hoffmann-La Roche & Co Aktiengesellschaft
Grenzacherstrasse 124-184
4002 Basle
SWITZERLAND

Address for Service: Spruson & Ferguson, Patent Attorneys
Level 33 St Martins Tower, 31 Market Street
Sydney, New South Wales, 2000, Australia

Complete Specification for the invention entitled:

Tyr-Peptide Analogs

The following statement is a full description of this invention, including the best method of performing it known to me/us

ABSTRACT OF DISCLOSURE

5 Analog of tyrosine sulfate- or tyrosine phosphate
 residue(s) containing peptides, the novel intermediate
 compounds used in the preparation of these analogs, as well
 as a method for suppressing appetite in subjects by
 administering to the subject an effective amount of CCK
 analog wherein one or more of any tyrosine sulfate
 residue(s) present is/are replaced by a residue of the
10 invention.

15

20

25

30

35

- 1A -

Peptides are ubiquitous biological molecules and have, in recent years, become the subject of extensive research and investigation. For example, the possibilities for utilizing natural biological substances, such as peptides, as therapeutics for various diseases is being aggressively explored.

Elucidation of the amino acid sequences of such peptides such as Growth Hormone, Growth Hormone Releasing Factor, or Cholecystokinin (CCK) has lead to advancements in the understanding of how these molecules work in treating various disorders. However, peptides suffer from rapid degradation upon exposure to the internal milieu often resulting in low bioavailability. It has been recently discovered that in many instances if the amino acid constituency of many naturally occurring peptides is altered by single or multiple amino acid replacements at different sites, the analogs of the natural peptide may degrade less rapidly and hence exhibit greater bioavailability and efficacy.

For example, CCK is a family of peptide hormones which vary in length up to 58 amino acids. The amino acid sequence of CCK first discovered had a length of 33 amino acids. CCK as well as fragments thereof, such as CCK-8 and CCK-7, have been shown to have satiety-inducing effects when administered peripherally to animals. CCK-8 has the amino acid sequence:

26 27 28 29 30 31 32 33
Asp-Tyr-(SO₃H)-Met-Gly-Trp-Met-Asp-Phe-NH₂.

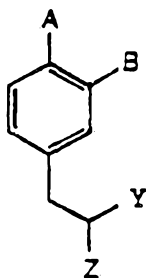
CCK-7 corresponds to the amino acid sequence 27-33 shown above.

While CCK analogs are known to have satiety inducing effects, they exhibit low bioavailability and are poorly absorbed. Tyrosine sulfate containing peptides are well known to suffer loss of the sulfate moiety upon storage, particularly when stored in solution. This has led to the synthesis of various CCK analogs wherein the attempt to improve properties such as stability and bioavailability has been made. A multitude of CCK analogs with various amino acid replacements resulted in compounds with altered properties which enhance their potential usefulness in human therapeutics.

The instant invention comprises analogs of tyrosine sulfate- or tyrosine phosphate residue(s) containing peptides. Examples of tyrosine sulfate residue(s) containing peptides include peptides such as gastrin, cholecystokinin, platelet factor 4, or hirudin. Many tyrosine phosphate residue(s) containing peptides are also known such as human insulin receptor, progesterone receptor, or Lipocortin-I. All of these peptides may yield analogs according to the instant invention.

For example, a tyrosine sulfate residue occupies position 27 of the amino acid sequence of CCK or the corresponding position in its shorter analogs such as CCK-7 and CCK-8. When this tyrosine sulfate residue is replaced by a residue of the invention, the result is a CCK analog which may exhibit greater bioavailability, and hence may lend itself more readily to absorption following oral administration.

The instant invention comprises analogs of tyrosine sulfate- or tyrosine phosphate residue(s) containing peptides wherein one or more of such residue(s) is/are replaced by (a) residue(s) of the formula:



I

5

wherein A, B, Z, and Y are as described hereinafter,
10 and pharmaceutically acceptable salts thereof.

The instant invention also comprises the novel intermediate compounds.

15

The instant invention also comprises a method of suppressing appetite in subjects by administering an appetite suppressing effective amount of a CCK analog wherein tyrosine sulfate residue(s) is are replaced by residue(s) of the invention.

20

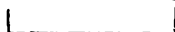
The following symbols and terminology as utilized in this specification are defined as follows:

1. cyclic peptide

25

means a peptide where the omega carboxy terminus of one amino di-acid in the peptide chain is attached to the omega amino terminus of another di-amino acid in the peptide chain via the formation of an amide bond. The bonding between the two amino acids in the chain illustrated by the designation

30



35

results in a ring structure.

2. lower alkyl means straight or branched chain saturated hydrocarbon residues containing from 1 to 7 carbon atoms.
- 5
3. aryl means substituted or unsubstituted phenyl or naphthyl wherein the substituents are one or more halogen, lower alkyl, lower alkoxy or nitro.
- 10
4. lower alkoxy means a lower alkyl oxy group such as methoxy, ethoxy, propoxy, etc.
- 15
5. lower alkenyl means a straight or branched chain unsaturated hydrocarbon residue containing from 2 to 7 carbon atoms.
- 20
6. Phe(3-COOH) means the 3 position of the phenyl on phenylalanine is substituted with COOH.
- 25
7. Phe(4-COOH) means the 4 position on the phenyl of phenylalanine is substituted with COOH.
- 30
8. Phe(4-CH₂COOH) means the 4 position on the phenyl of phenylalanine is substituted with CH₂COOH.

- 5 9. Phe(4-CH₂COOC₂H₅) means the 4 position on the phenyl of phenylalanine is substituted with CH₂COOC₂H₅.
- 10 10. Phe(4-CH₂CH₂COOH) means the 4 position on the phenyl of phenylalanine is substituted with CH₂CH₂COOH.
11. Phe(4-CF₂COOH) means the 4 position on the phenyl of phenylalanine is substituted with CF₂COOH.
- 15 12. Phe(4-tetrazole) means the 4 position on the phenyl of phenylalanine is substituted with a 5-tetrazolyl group.
- 20 13. Phe(4-CH₂-tetrazole) means that 4 position on the phenyl of phenylalanine is substituted with a CH₂-[tetrazolyl] group.
- 25 14. peptide means a linear or cyclic peptide.
15. Ac means acetyl.

30

35

16. (D,L), (D), or (L)

preceding the amino acid designation means that this amino acid exists in that specific isomeric form. i.e. (D,L)Phe means that the amino acid phenylalanine exists as a racemic mixture; (D)Phe means that the amino acid Phenylalanine exists as the R stereoisomer; (L)Phe means that the amino acid Phe exists as the L stereoisomer or implied S configuration. In case that there is no designation preceeding the amino acid the L isomer is implied (S configuration). All amino acids are represented by their commonly understood three letter designations.

17. Compounds

where this term is used it includes enantiomers and racemates of the compounds.

18. Tyrosine Sulfate

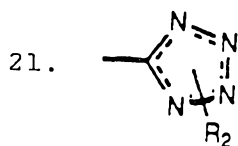
means the amino acid tyrosine wherein a sulfate ester is present on the 4-position of the aromatic ring.

19. Tyrosine Phosphate

means the amino acid tyrosine wherein a phosphate ester is present on the 4-position of the aromatic ring.

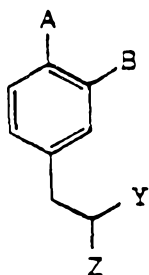
20. Desamino

means an amino acid which lacks the alpha-amino functional group. For example, in the context of the instant invention "Desamino Phe" means that the phenylalanine lacks the alpha amino functional group.



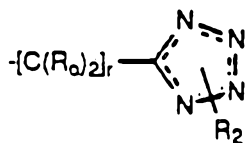
means a tetrazole residue with a functional group R_2 attached to a nitrogen in position 1 or 2.

The instant invention comprises analogs of tyrosine sulfate- or tyrosine phosphate residue(s) containing peptides wherein one or more of such residue(s) is/are replaced by residue(s) of the formula



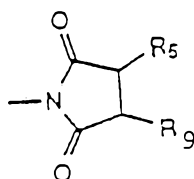
I

wherein one of A or B is selected from the group consisting of $-[C(R_0)_2]-CO_2R_1$, or



and the other of A or B is H;

Z is H, $-\text{NH}_2$, $-\text{NH}-$, lower alkyl, $-\text{NHCOR}_3$, $-\text{NHCO}_2\text{R}_4$ or



R_0 is H or F;

R_1 is H, substituted or unsubstituted lower alkyl with the substituents selected from the group consisting of hydroxyl, halogen, or aryl;

R_2 is H, lower alkenyl, lower alkyl or lower alkyl substituted by 1 to 3 aryl groups;

R_3 is H, lower alkyl, alkyl substituted by one or two aryl groups or aryl;

R_4 is lower alkyl or alkyl substituted by one or two aryl groups;

R_5 and R_9 are each independently H, lower alkyl, or taken together and including the carbon atoms to which they are attached may form a six membered ring which may be aromatic;

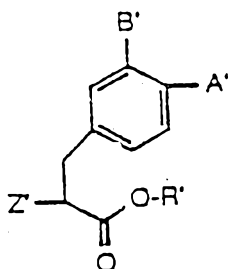
s and r are 0, 1 or 2;

and Y is $-\text{COOH}$, alkoxycarbonyl, $-\text{CONH}_2$ or $-\text{CO}-$;

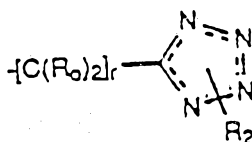
with the proviso that if Z is other than $-\text{NH}-$ then Y is $-\text{CO}-$; or if Y is other than $-\text{CO}-$ then Z is $-\text{NH}-$;

and pharmaceutically acceptable salts thereof.

According to a second embodiment of this invention, there is provided a compound when used for the preparation of an analog of a tyrosine sulphate- or tyrosine phosphate residue(s) containing peptide according to the first embodiment, said compound having the formula:



wherein one of A' or B' , is selected from $-\text{[C(R}_0\text{)}_2\text{]}_s-\text{CO}_2\text{R}_1'$, or



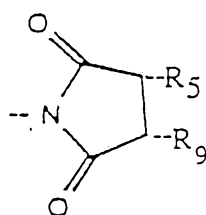
and the other of A' or B' is H wherein R_0 is H or F, R_1' is H, substituted or unsubstituted lower alkyl with the substituents selected from hydroxyl, halogen, or aryl, R_2 is H, lower alkenyl, lower alkyl or lower alkyl substituted by 1 to 3 aryl groups,

5 r is 0-2

s is 1-2

R' is H, lower alkyl or lower alkyl substituted by 1 to 3 aryl groups,

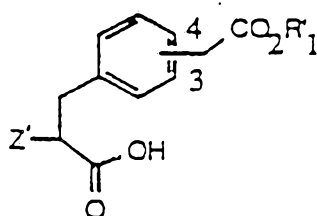
Z' is H, lower alkyl, $-NH_2$, $-NHCOR_3$, $-NHCO_2R_4$, or



10 and R_3 and R_4 are H, lower alkyl or lower alkyl substituted by one, two or three aryl groups; and R_5 and R_9 are each independently H, lower alkyl, or taken together and including the carbon atoms to which they are attached may form a six membered ring which may be aromatic.

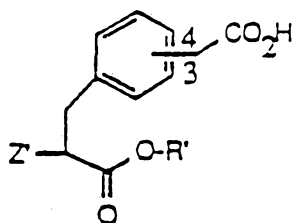
According to a third embodiment of this invention, there is

15 provided a compound when used for the preparation of an analog of a tyrosine sulphate- or tyrosine phosphate residue(s) containing peptide according to the first embodiment, said compound having the formula:



wherein Z' and R_1' are as defined in the second embodiment.

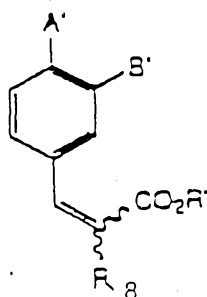
20 According to a fourth embodiment of this invention, there is provided a compound when used for the preparation of an analog of a tyrosine sulphate- or tyrosine phosphate residue(s) containing peptide according to the first embodiment, said compound having the formula:



wherein R' and Z' are as defined in the second embodiment.

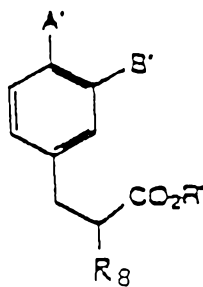
According to a fifth embodiment of this invention, there is provided a compound when used for the preparation of an analog of a tyrosine sulphate- or tyrosine phosphate residue(s) containing peptide

5 according to the first embodiment, said compound having the formula:



wherein A', B', and R' are as defined in the second embodiment and R₈ is H or lower alkyl.

According to a sixth embodiment of this invention, there is provided a compound when used for the preparation of an analog of a tyrosine sulphate- or tyrosine phosphate residue(s) containing peptide according to the first embodiment, said compound having the formula:



wherein A', B', and R' are as defined in the second embodiment and R₈ is H or lower alkyl.

According to a seventh embodiment of this invention, there is provided a process for the manufacture of a compound according to the first embodiment, which process comprises treating the resin bound peptide with appropriate cleavage reagent(s) and if desired, converting such a peptide obtained into a pharmaceutically acceptable salt.



According to a eighth embodiment of this invention, there is provided a pharmaceutical composition containing a compound according to the first embodiment and a non-toxic, inert, therapeutically acceptable carrier material.

- 5 According to a ninth embodiment of this invention, there is provided a pharmaceutical composition for suppressing appetite containing a compound according to the first embodiment and a therapeutically inert carrier material.

- 10 According to a tenth embodiment of this invention, there is provided the use of a compound according to the first embodiment as a therapeutically active substance.

According to an eleventh embodiment of this invention, there is provided the use of a compound according to the first embodiment in the suppression of appetite.

- 15 According to a twelfth embodiment of this invention, there is provided a method for suppressing appetite in subjects by administering an appetite suppressing effective amount of an analog of CCK wherein the tyrosine sulfate residue is replaced by a residue of Formula I as defined in the first embodiment.

- 20 A preferred embodiment of this invention is analogs of tyrosine sulfate residue(s) containing peptides wherein one or more tyrosine sulfate residue(s) are/is replaced by residue(s) of formula I.

- 25 A further preferred embodiment of the present invention is analogs of CCK wherein the tyrosine sulfate residue(s) is/are replaced by (a) residue(s) of formula I wherein Z is H, NHCOR_3 , or NHCO_2R_4 and R_3 is lower alkyl, R_4 is lower alkyl optionally substituted with one to three aryl groups and such analogs wherein R_3 is CH_3 .



Another preferred embodiment of the present invention are analogs of CCK wherein the tyrosine sulfate residue(s) is/are replaced by a residue of formula I, wherein Z in formula I is H or NHCOR_3 with R_3 = lower alkyl or H.

5

Further preferred are analogs wherein one of A or B in formula I is $-\text{[C(R}_0\text{)]}_s\text{-CO}_2\text{R}_1$ and the other of A or B is H.

10

Further preferred are analogs wherein A in formula I is $-\text{[C(R}_0\text{)]}_s\text{CO}_2\text{R}_1$ and B in formula I is H.

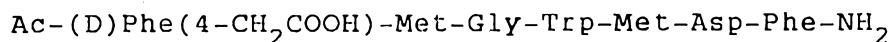
Also preferred are analogs wherein s in formula I is 0 and R_1 is H.

15

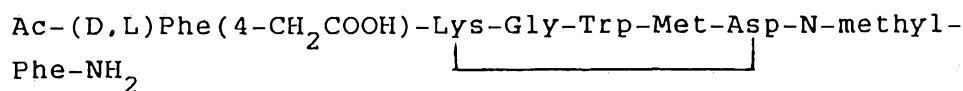
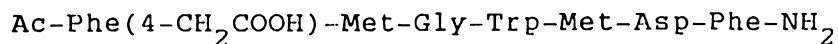
More preferred are analogs of CCK wherein tyrosine sulfate residue(s) is/are replaced by residue(s) of formula I wherein s is 1, R_0 is H or F, and R_1 is H.

20

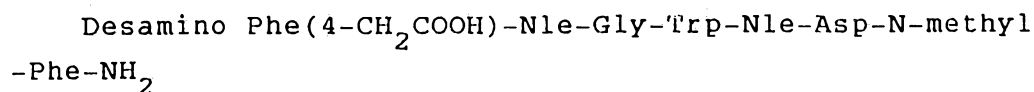
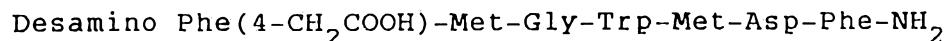
Most preferred are analogs wherein R_0 in formula I is H, yielding the preferred analogs of the formula:



25

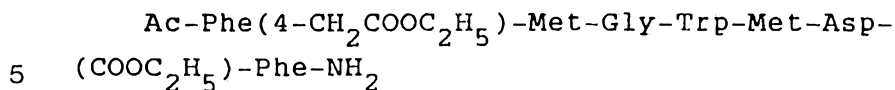
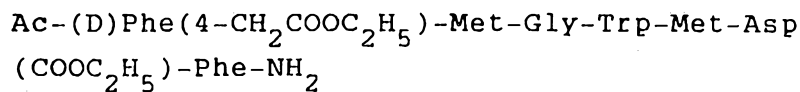


30



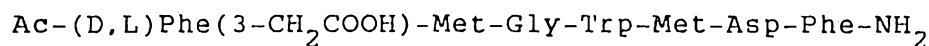
35

Also preferred are analogs wherein s in formula I is 1, R_0 in formula I is H, and R_1 in formula I is $-\text{CH}_2\text{CH}_3$ yielding preferred analogs of the formula:



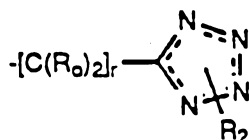
Also preferred are analogs wherein B in formula I is $-\text{[C(R}_0\text{)}_2\text{]}_s\text{CO}_2\text{R}_1$ and A in Formula I is H.

10 More preferred are analogs wherein s in formula I is 1 and R_1 in Formula I is H:



15 Also preferred are analogs wherein s in formula I is 2 and R_0 is H and R_1 is H.

Also preferred are analogs of CCK wherein the tyrosine sulfate residue(s) is/are replaced by residue(s) of formula
20 I wherein one of A or B in Formula I is



25 and the other of A or B in Formula I is H.

Further preferred are analogs wherein A in formula I is



and B in formula I is H.

35 Most preferred are analogs wherein r in formula I is 0 and R_2 in formula I is H yielding the preferred analogs of the formula:

Ac-Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂

Desamino-Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂

5 Desamino-Phe(4-tetrazole)-Lys-Gly-Trp-Met-Asp-N-methyl-
Phe-NH₂

Ac-(D,L)Phe(4-tetrazole)-Lys-Gly-Trp-Met-Asp-N-methyl-
Phe-NH₂

10

Also preferred are analogs containing the residue of
formula I wherein r in formula I is 1 and R₂ in formula I
is H and Ro is H yielding preferred analogs of the formula:

15 Ac-(D)Phe(4-CH₂-tetrazole)-Met-Gly-Trp-Met-Asp-
Phe-NH₂

Ac-Phe(4-CH₂-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂

20 Ac-(D)Phe(4-CH₂-tetrazole)-Nle-Gly-Trp-Nle-Asp-N-methyl-
Phe-NH₂

Ac-Phe(4-CH₂-tetrazole)-Nle-Gly-Trp-Nle-Asp-N-methyl-Phe
-NH₂

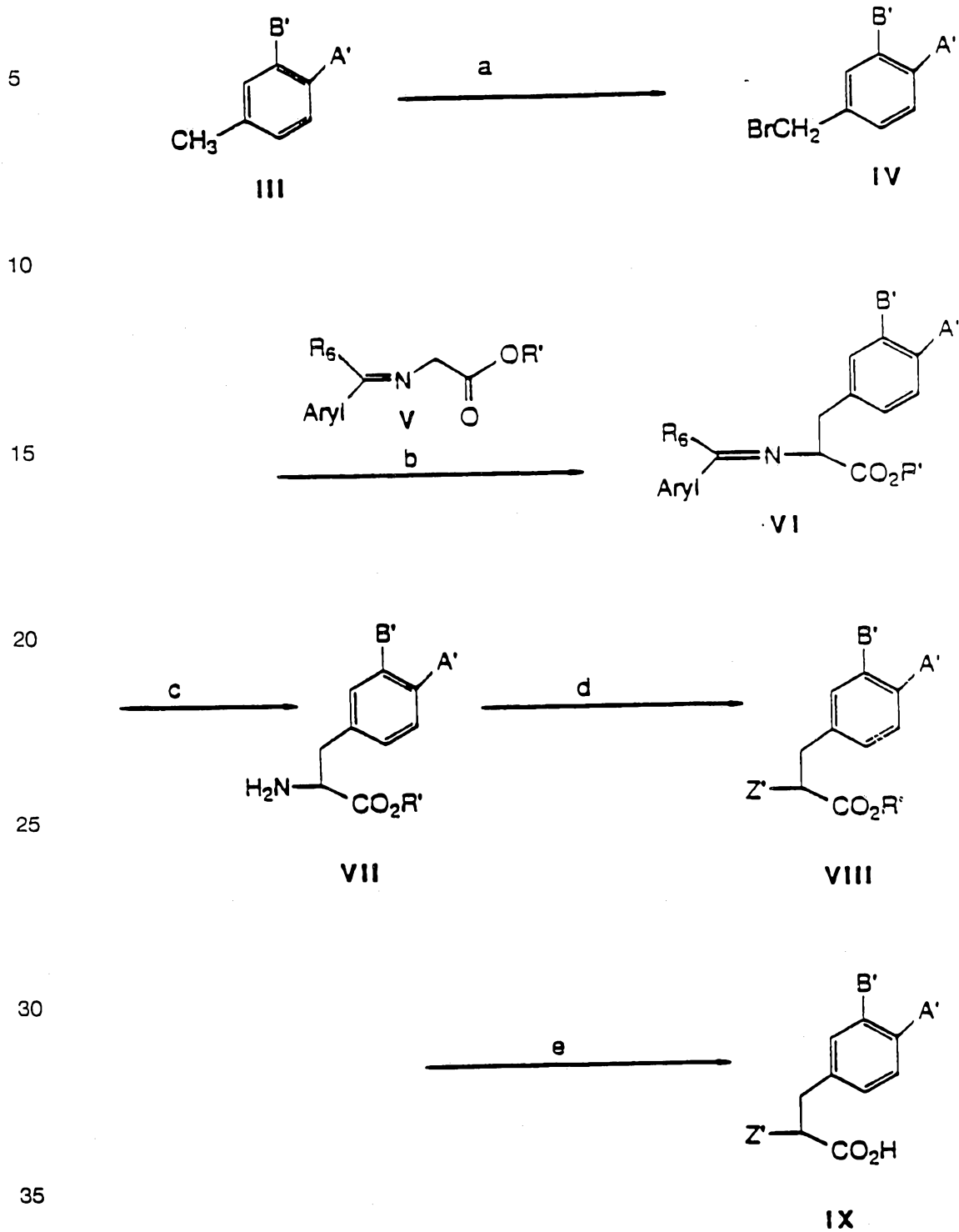
25

Desamino-Phe(4-CH₂-tetrazole)-Nle-Gly-Trp-Nle-Asp-N-
methyl-Phe-NH₂

The residues incorporated into the analogs of tyrosine
30 sulfate- or tyrosine phosphate residue(s) containing
peptides according to the invention may be synthesized as
follows:

35

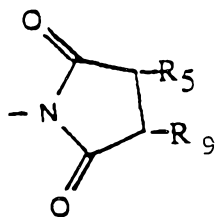
Scheme I



In Reaction Scheme I A' and B' are identical with A and B in formula I with the proviso that R₁ and R₂ are not H.

R' may be a substituted or unsubstituted lower alkyl with the substituents selected from the group consisting of hydroxyl, halogen, or aryl; or any suitable protecting group chosen so as to be easily removed selectively without affecting R₁, R₂ or Z' in step (e).

R₆ may be H or aryl, and Z' is -NH₂, -NHCOR₃, -NHCO₂R₄, or



In Reaction Scheme I the formula III compound is reacted in step (a) with a halogenating agent dissolved in a nonpolar inert organic solvent which contains a catalytic quantity of a radical initiator at a temperature ranging from 60°C to 80°C until most of the formula III compound is consumed. A suitable halogenating agent is N-bromo-succinimide, a suitable nonpolar inert organic solvent is carbon tetrachloride, and a suitable catalytic radical initiator is azobisisobutyronitrile or benzoyl peroxide. The resulting formula IV compound can then be isolated by conventional methods such as chromatography or distillation. In step (b) the formula IV compound is reacted at about room temperature with a formula V compound wherein R₆ is aryl, in a biphasic mixture of, for example, methylene chloride and water which contains an inorganic base, such as alkali metal hydroxide and a phase transfer catalyst, such as a tetraalkylammonium salt [see the procedure described in O'Donnell, et al., Tetrahedron Letters, 2641 (1978)]. The resulting formula VI compound can be isolated by conventional methods, such as chromatography or can be used directly in step (c).

Alternatively, the formula V compound wherein R_6 is H, or aryl, can be deprotonated below room temperature with a suitable strong base, such as lithium diisopropylamide or potassium hexamethyldisilazide, in an inert ethereal solvent, such as diethylether, tetrahydrofuran or dimethoxyethane to which is added a metal coordinating agent such as hexamethylphosphoric triamide. A solution of a formula IV compound in an ethereal solvent such as diethyl ether, tetrahydrofuran or dimethoxyethane, can then be added to the mixture preferably at a reaction temperature of about -78°C . After complete reaction and aqueous workup, the resulting formula VI compound may be isolated by conventional methods or may be used directly in step (c).

In step (c) the compound of formula VI is treated with at least one equivalent of a strong acid dissolved in a suitable organic solvent which contains excess water. Suitable acids are toluenesulfonic acid or hydrochloric acid, and tetrahydrofuran, ether or acetonitrile are suitable solvents. After complete reaction, the resultant compound of formula VII can be isolated as its salt by conventional methods such recrystallization or may be used directly in step (d), wherein the formula VII compound is treated with an acylating agent, such as acetic anhydride or t-butylpyrocarbonate, in the presence of a base which may be a tertiary amine such as triethyl amine; at a temperature ranging from 0°C to room temperature. The choice of solvent is not critical for this reaction and is normally based on considerations of solubility and reactivity of the reaction components. Dichloromethane is useful for laboratory scale preparations. Isolation of the resulting formula VIII compound can be accomplished using conventional methods.

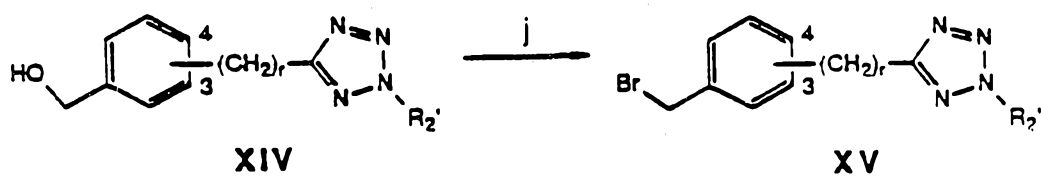
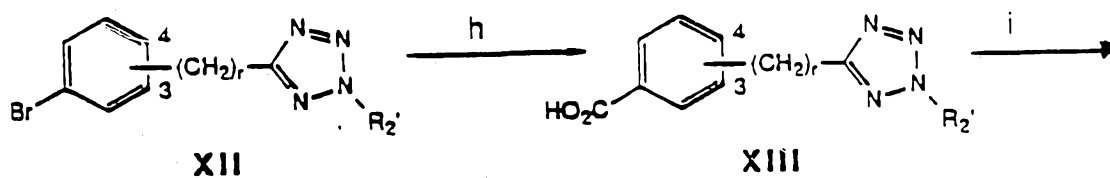
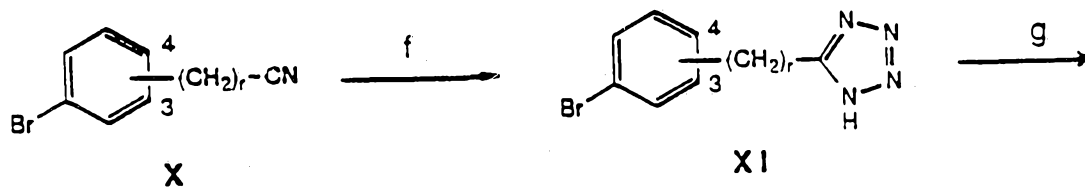
In step (e), the conversion of the formula VIII compound to a formula IX compound is carried out by selective ester hydrolysis. Appropriate conditions for this hydrolysis depend on the particular choice of R' , R_1 , R_2 and Z' in the compound of formula VIII. Optimally, these groups are chosen so as to facilitate the selective conversion of R' to hydrogen without affecting R_1 , R_2 or Z' using methods known in the art. For example, where R' is a mono- or diarylmethyl group it can be removed through treatment with hydrogen gas at atmospheric pressure in a polar organic solvent such as ethanol in the presence of of a noble metal catalyst such as 10% Pd/C. These reaction conditions may be controlled so as not to affect R_1 , or R_2 when these groups are lower alkyl, mono-, di-, or trihaloalkyl, or trialkylsilylalkyl and will not affect Z' when R_3 and R_4 are appropriately selected, as for example lower alkyl. The resulting formula IX compound can be isolated by conventional methods.

Alternatively, when R' is a small alkyl group such as methyl and R_1 is a bulky alkyl group such as a tertiary butyl group; selective base hydrolysis may be effected by treatment of the formula VIII compound with an alkali metal hydroxide, in the presence of a polar solvent until hydrolysis of R' is complete. When done carefully these reaction conditions will not affect R_1 , R_2 or Z' . Isolation of the formula IX compound can then be accomplished using conventional methods.

30

35

Scheme II



In Reaction Scheme II, r and R'_2 are as defined in formula I for r and R_2 respectively except that R'_2 should not be H, and the side chain may be attached to either position 3 or 4 on the aromatic ring as indicated.

- 5 In step (f) the formula X compound is treated with sodium azide and ammonium chloride in a polar aprotic solvent such as dimethylformamide at a temperature of about 90-100°C. The resultant formula XI compound can then be isolated by conventional techniques. In step (g) the tetrazole of
- 10 formula XI is treated with an excess of an alcohol capable of forming a stabilized carbonium ion in the presence of a strong acid (such as trifluoroacetic acid), at about room temperature. The major product from this process will be a formula XII compound, however, the regioisomer resulting
- 15 from alkylation at N-1 of the tetrazole will also be formed in varying amounts depending on the selection of r and the particular reaction conditions. Formula XII compounds and their regioisomers may be separated but this is generally not necessary as both isomers will yield the same product
- 20 when R'_2 is converted to hydrogen. Alternatively, a primary or secondary alcohol can be reacted with a formula XI compound in the presence of a dialkyl azodicarboxylate and a phosphine in an inert solvent according to the procedure described by O. Mitsunobu, Synthesis, (1981) 1; to
- 25 give a formula XII compound, wherein R'_2 is derived from a primary or secondary alcohol.

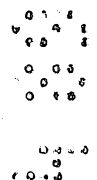
- In step (h) the formula XII compound is carbonylated according to the method of Schoenberg, et al. J. Org. Chem.,
- 30 39, 3318 (1974). Thus a compound of formula XII is maintained under an atmosphere of carbon monoxide ranging from 100-200 psi in the presence of a tertiary amine base in a polar solvent system that contains a trialkyl or triaryl phosphine and a source of catalytic palladium zero. This
- 35 mixture is maintained at about 100° C from 1 to 3 days. The resulting formula XIII compound may be isolated by conventional techniques.

In step (i) the formula XIII compound is treated with a metal hydride reducing agent in an inert solvent at room temperature. The resulting compound of formula XIV
5 compound may be isolated by conventional techniques. In step (j) the formula XIV alcohol is treated with triphenylphosphine and carbon tetrabromide in an inert solvent at or below room temperature, preferably at about 0°C. The resulting formula XV compound can be isolated by
10 conventional means.

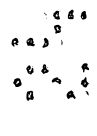


15

20



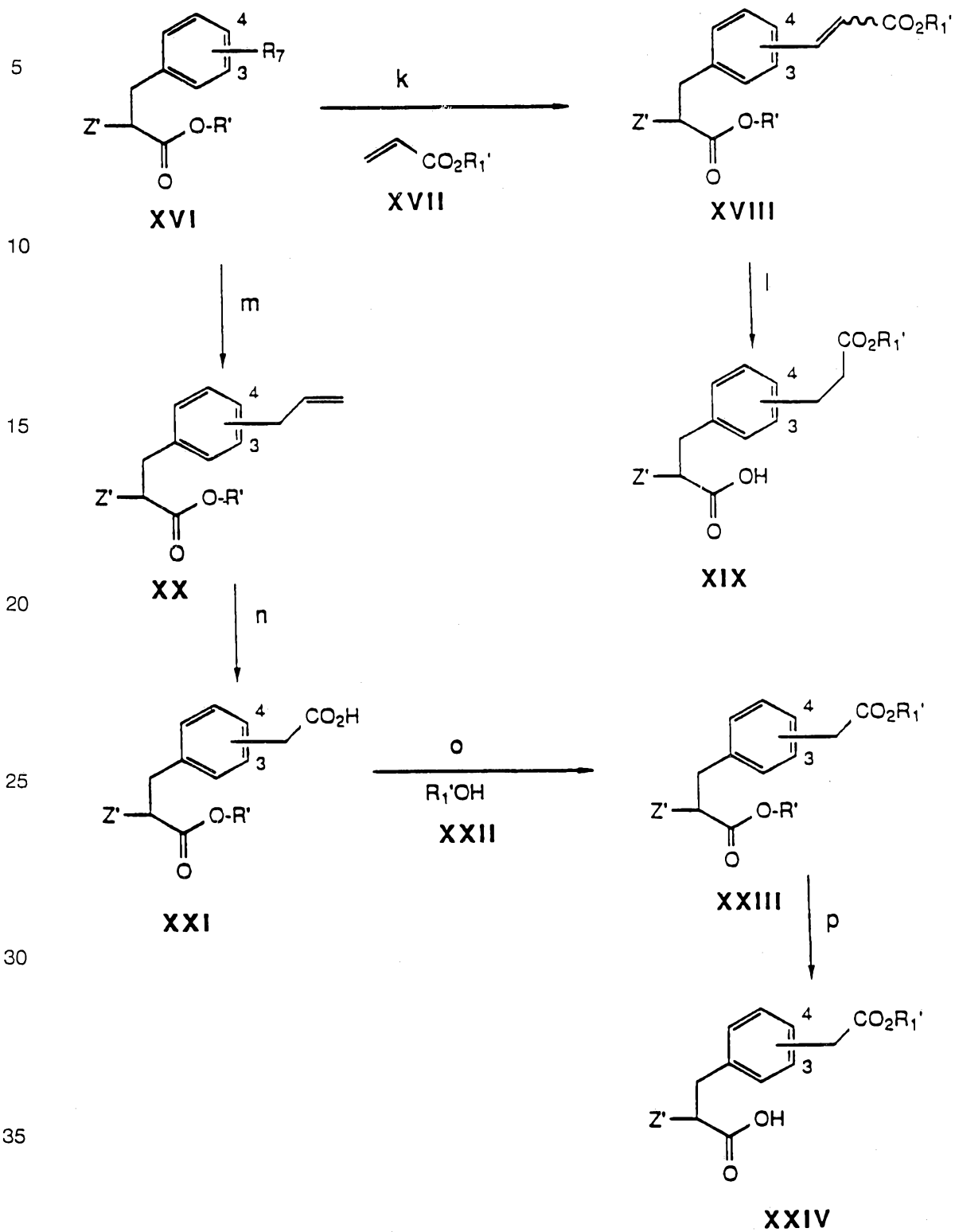
25



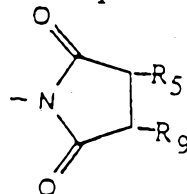
30

35

Scheme III



In Reaction Scheme III, R' is as previously defined, Z' is H, lower alkyl, -NHCOR_3 , $\text{-NHCO}_2\text{R}_4$, or



5

and R₇ is bromide, iodide, or a perfluoroalkylsulfonate ester which may be attached to the phenyl at either position 3 or 4 as set forth in formula XVI, and R'₁ is the same as R₁ in formula I with the proviso that R'₁ is other than

10

H. In step (k) the formula XVI compound is reacted with an acrylate of formula XVII in the presence of a source of palladium zero and a tertiary amine base. When R₇ is a perfluoroalkylsulfonate ester, this procedure is carried out in a polar aprotic solvent at a reaction temperature of

15

70-100°C as described by Chen and Yang, Tetrahedron Letters, 27, 1171 (1986). When R₇ is bromide or iodide, the reaction can be carried out as described by Heck, Organic Reactions, 27, (1982) 345. The side chain of the resulting formula XVIII compound becomes attached to the site of the departing R₇. The formula XVIII compounds may be isolated by conventional means.

20

Step (l) can be carried out in one or two steps depending on the particular choice of R'₁ and R'.

25

When R' is mono- or diarylmethyl and R'₁ is lower alkyl, halosubstituted alkyl, or hydroxy substituted alkyl, and Z' does not contain a mono-, di- or triarylmethoxy group, catalytic hydrogenation over a noble metal catalyst, in a polar solvent at a hydrogen pressure of from one to three atmospheres results in the simultaneous reduction of the double bond and hydrogenolysis of the R' group yielding a formula XIX compound after a suitable isolation procedure. Alternatively, when R' is methyl or ethyl and R'₁ is a bulky substituent such as, tertiary butyl,

30

35

catalytic hydrogenolysis may be carried out as above, and a

separate selective hydrolysis step may be carried out as described above for step (e) in Scheme I to yield the formula XIX compound.

5 In step (m) the formula XVI compound where R_7 is perfluoroalkylsulfonate ester is treated with an allyl trialkylstannane, a source of palladium zero, and an excess of lithium chloride in a polar aprotic solvent at about 90°C. The resulting formula XX compound can be isolated by
10 conventional techniques. In step (n) the formula XX compound is reacted with a suitable oxidizing agent and a catalytic amount of ruthenium trichloride in a two phase system as described by Carlsen et al., J. Organic Chemistry, 46, (1981), 3936. Depending on the particular choice of
15 reaction conditions this process may lead either directly to a compound of formula XXI or to an intermediate aldehyde which may be oxidized to a formula XXI compound according to the procedure described by Bal et al., Tetrahedron, 37, 2091 (1981). The resulting formula XXI compounds can be isolated
20 by conventional techniques.

In step (o), an acid of formula XXI is converted to an ester of formula XXIII. The choice of R_1 should be such that it will survive the reaction conditions required
25 to generate an acid of formula XXIV and will thus depend on the selection of R' in the formula XXI compound. The formula XXI compound can be reacted with an alcohol of formula XXII in the presence of a coupling reagent and a catalyst in an inert solvent. Alternatively the formula XXI
30 compound may be reacted with a dimethylformamide dialkoxyacetal in a inert solvent at a reaction temperature ranging from 60 to 80°C. The resulting formula XXIII compound can be isolated by convention means.

35 In step (p) the formula XXIII compound is converted to the formula XXIV compound as described in step (e) of scheme I.

Scheme IV

5

10

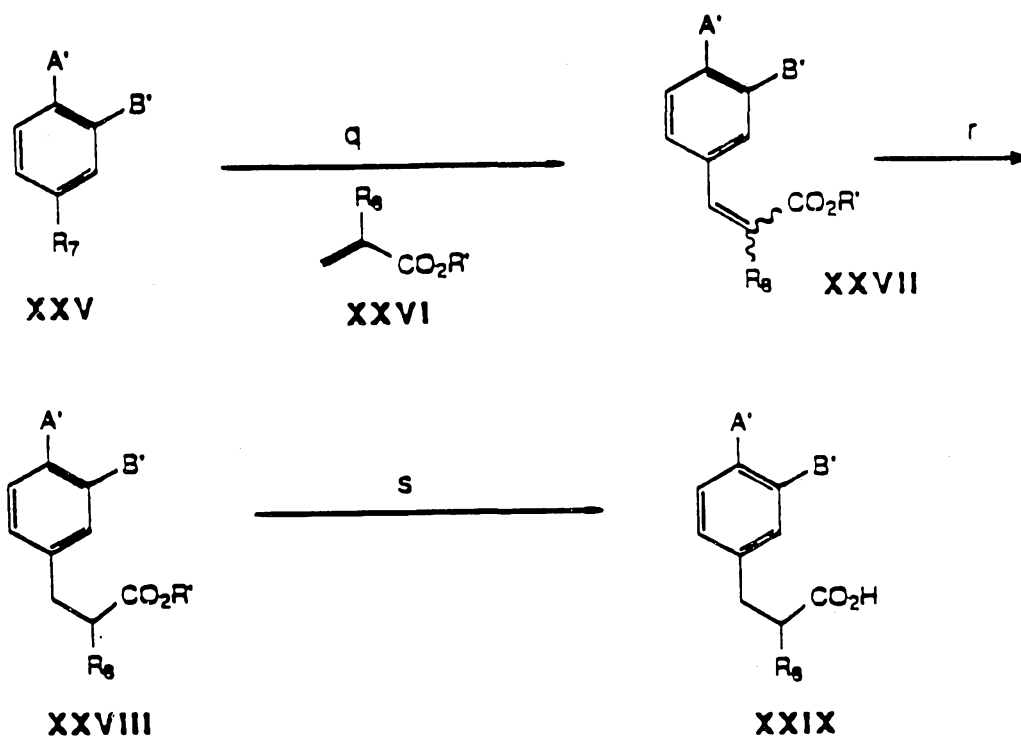
15

20

25

30

35



In reaction scheme IV, A', B', R₇ and R' are as previously defined and R₈ is hydrogen or lower alkyl.

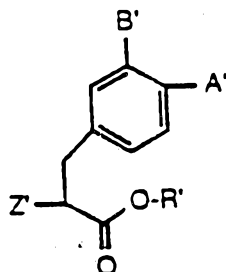
The compound of formula XXV can be prepared by known methods. In Step q the compound of formula XXV is treated with a compound of formula XXVI under the general conditions described for step k in scheme III to give a compound of formula XXVII which can be isolated by conventional means. In step (r) formula XXVII compound is reduced to a formula XXVIII compound. When A' and B' contain groups resistant to catalytic hydrogenation, this process is conveniently carried out by hydrogenation over a noble metal catalyst (for example palladium/carbon) in a suitable solvent until reduction is complete. The resulting compound of formula XXVIII can be isolated by conventional techniques such as recrystallization. When R' is susceptible to hydrogenolysis, the conditions of step (r) may lead directly to a compound of formula XXIX. Otherwise, a selective hydrolysis may be carried out using the conditions described for step (e) in reaction scheme I to give a compound of formula XXIX.

The instant invention also comprises all the novel intermediate compounds which are used to prepare the compounds of the invention.

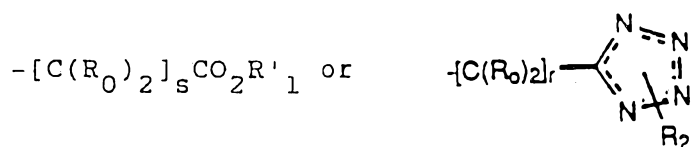
These novel intermediates are compounds represented by formulas VII, VIII, and IX in scheme I and formulas XIX, XXI, XXIII, and XXIV in scheme III and formula XXIX in scheme IV ; wherein the substituents are as defined in schemes I-IV except as noted.

Further novel intermediates are the following compounds:

(a) a compound of the formula:



wherein one of A' or B' is selected from the group consisting of



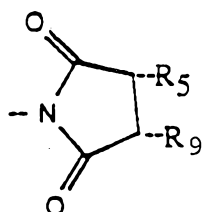
and the other of A' or B' is H wherein R₀ is H or F, R'₁ is H, substituted or unsubstituted lower alkyl with the substituents selected from the group consisting of hydroxyl, halogen, or aryl, R₂ is H, lower alkenyl, lower alkyl or lower alkyl substituted by 1 to 3 aryl groups,

r is 0-2

s is 1-2

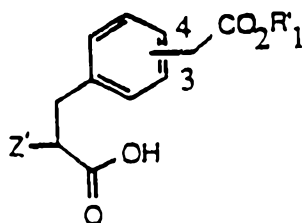
R' is H, lower alkyl or lower alkyl substituted by 1 to 3 aryl groups.

Z' is H, lower alkyl, -NH₂, -NHCOR₃, -NHCO₂R₄.



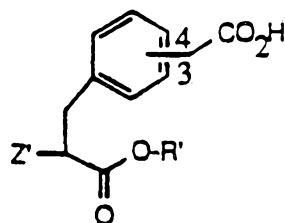
and R_3 and R_4 are H, substituted or unsubstituted lower alkyl with the substituents selected from one, two, or three aryl groups; and R_5 is each independently H, lower alkyl, or taken together may form a six membered ring which may be aromatic, or

(b) a compound of the formula



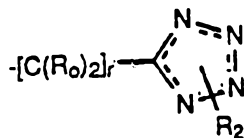
wherein Z' and R'_1 are as defined under (a), or

(c) a compound of the formula



wherein R' and Z' are as defined under (a), or

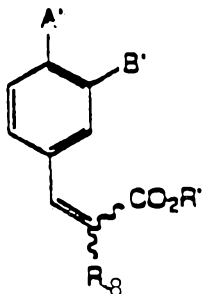
(d) a compound as defined under (a) wherein one of A' or B' is



and the other of A' or B' is H.

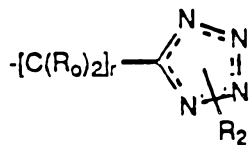
wherein Z' , R_2 , and R' are as defined under (a), or

(e) a compound of the formula:



wherein A', B', and R' are as defined under (a) and R₈ is H or lower alkyl, or

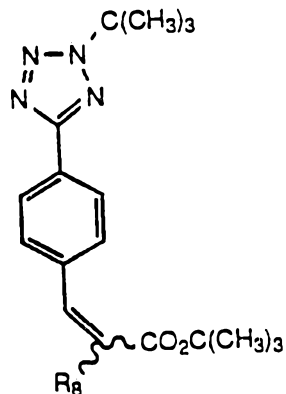
(f) a compound as defined under (e) wherein A' is



and B' is H, and R₀, r, and R₂ are as defined under (a),

(g) a compound as defined under (f) wherein r=0, or

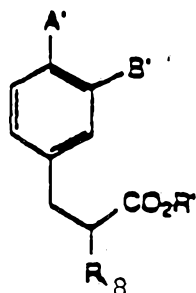
(h) a compound as defined under (e) wherein R' is t-butyl and R₈ is H said compound having the formula:



, or

(i) a compound of the formula:

5

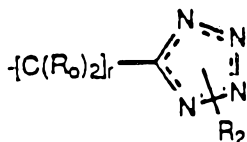


10

wherein A', B', and R' are as defined under (a) and R₈ is H or lower alkyl, or

(k) a compound as defined under (i) wherein A' is

15



20

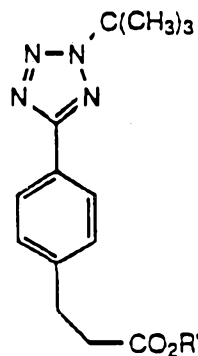
and B' is H and R₀, r, and R₂ are as defined under (a), or

(l) a compound as defined under (k) wherein r=0, or

25

(m) a compound as defined under (l) wherein R' is lower alkyl and R₂ is t-butyl said compound having the formula:

30

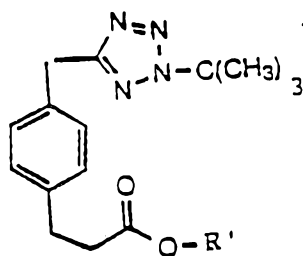


35

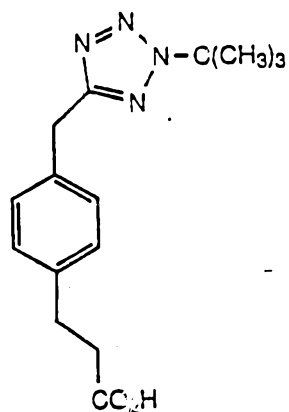
(n) a compound as defined under (i) wherein R' is H and R₂ is t-butyl said compound having the formula:

(o) a compound as defined under (k) wherein $r=1$ and R_0 is H, or

(p) a compound as defined under (o) wherein R' is lower alkyl and R₂ is t-butyl said compound having the formula:



(q) a compound as defined under p wherein R' is H and R₂ is t-butyl said compound having the formula:



The residues of formula I may be incorporated into peptides in a protected form, that is R_1 or R_2 may be substituted alkyl such as tertiary butyl and Z may be hydrogen, $NHCR_3$, or $NHCOR_4$ as appropriate. Such a residue is then incorporated into the growing peptide chain according to methods known in the art. The protecting groups are then removed according to suitable known methods.

The instant invention also comprises a method for suppressing appetite in subjects by administering to the subject an appetite suppressing effective amount of an analog of CCK wherein the tyrosine sulfate residue(s) is/are replaced by residue(s) of formula I or of an pharmaceutically acceptable salt thereof.

Particularly preferred is a method for suppressing appetite in subjects by administering to a subject an appetite suppressing effective amount of the following CCK analogs wherein the tyrosine sulfate residue has been replaced by a residue of formula I :

Ac-(D)Phe-(4- CH_2COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂
Ac-Phe(4- CH_2COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂
Ac-(D)Phe(4- $CH_2COOC_2H_5$)-Met-Gly-Trp-Met-Asp
($COOC_2H_5$)-Phe-NH₂
Ac-Phe(4- $CH_2COOC_2H_5$)-Met-Gly-Trp-Met-Asp
($COOC_2H_5$)-Phe-NH₂
Ac-(D,L)Phe(4- CF_2COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂
Ac-(D)Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂
Ac-Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂
Ac-(D,L)Phe(4- CH_2COOH)Lys-Gly-Trp-Met-Asp-N-methyl-Phe-NH₂
Ac-(D,L)Phe(4-tetrazole)-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-NH₂
Ac(D,L)Phe(3- CH_2COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂

Desamino Phe(4-CH₂COOH)-Nle-Gly-Trp-Nle-Asp-N-methyl-
Phe-NH₂
Desamino Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂
Desamino Phe(4-tetrazole)-Lys-Gly-Trp-Met-Asp-N-methyl-
5 Phe-NH₂
Ac-(D)Phe(4-CH₂-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂
Ac-Phe(4-CH₂-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂
Ac-(D)Phe(4-CH₂-tetrazole)-Nle-Gly-Trp-Nle-Asp-N-methyl-
Phe-NH₂
10 Ac-Phe(4-CH₂-tetrazole)-Nle-Gly-Trp-Nle-Asp-N-methyl-
Phe-NH₂
Desamino-Phe(4-CH₂-tetrazole)-Nle-Gly-Trp-Nle-Asp-N-
methyl-Phe-NH₂

15 or of their pharmaceutically acceptable salts.

Most preferred is a method for suppressing appetite in
subjects by administering to the subject an appetite
suppressing effective amount of the following CCK analogs
20 wherein the tyrosine sulfate residue has been replaced by a
residue of formula I :

Ac-(D)Phe(4-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂
25 Ac-Phe(4-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂
Ac-(D)Phe(4-CH₂COOHC₂H₅)-Met-Gly-Trp-Met-Asp-
(COOC₂H₅)-Phe-NH₂
30 Ac-Phe(4-CH₂COOC₂H₅)-Met-Gly-Trp-Met-Asp
(COOC₂H₅)-Phe-NH₂
Ac-Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂
35 Ac-(D,L)Phe(4-CH₂COOH)-Lys-Gly-Trp-Met-Asp-N-methyl-
Phe-NH₂

Ac-(D,L)Phe(4-tetrazole)-Lys-Gly-Trp-Met-Asp-N-methyl-
Phe-NH₂

5 Desamino-Phe(4-tetrazole)Lys-Gly-Trp-Met-Asp-N-methyl-
Phe-NH₂

Ac-(D,L)Phe(3-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂

10 Desamino-Phe(4-CH₂tetrazole)-Met-Gly-Trp-Met-Asp-
Phe-NH₂

Desamino-Phe(4-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂

15 Desamino-Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂
or of their pharmaceutically acceptable salts.

The invention is further directed to compositions for
suppressing appetite in subjects comprising an appetite
suppressing effective amount of the peptides of the
20 invention, or of their pharmaceutically acceptable salts.
Such compositions may be prepared in a conventional way and
may contain conventional carriers and/or diluents.

An "appetite suppressing effective amount" as used
25 herein refers to the amount of the peptide or its
pharmaceutically acceptable salt (on a weight basis) per kg
of body weight of the subject which must be administered to
suppress appetite. It is within the skill of the art to
calculate such amounts considering the method of
30 administration, the particular subject and the weight of the
subject. See Morley, J.E. "Minireview. The Ascent of
Cholecystokinin (CCK) from Gut to Brain" Life Sciences, 30,
(1982) 479.

35 The analogs may be administered as water soluble salts,
generally as salts of alkaline metals such as sodium or
potassium salts, as alkylamine salts, preferably

diethylamine salts or as acid addition salts. The analogs of the invention can be converted to the pharmaceutically acceptable salts by known methods.

5 The analogs of the invention may be administered to the subject by any suitable route including, nasal, sublingual, buccal, intraperitoneal, or parenteral including intravenous, intramuscular, or transdermal.

10 If the analogs of the invention are administered intranasally such vehicles of administration may include foams, creams, inhalants, etc. The effective appetite suppressing amount of the analog as the active ingredient is dissolved in pharmaceutically acceptable foams or inhalant
15 compositions suitable for intranasal administration, which compositions are known to those skilled in the art.

Where the peptides of the invention are administered parenterally, or intraperitoneally the appropriate amount of
20 the analog as the active ingredient is dissolved in sterile injectable solutions or suspensions. These types of solutions are well known to skilled artisans and comprise for example saline solutions, etc.

25 The present invention is also directed to the use of the peptides of the invention and their pharmaceutically acceptable salts as therapeutically active substances, especially as appetite suppressing substances.

30 The peptides of the invention may be prepared using the method of solid phase synthesis as generally described by Merrifield, J. Am. Chem. Soc., 85, 2149 (1963), although other equivalent methods of chemical synthesis known in the art may also be used. Solid-phase synthesis is commenced
35 from the C-terminal end of the peptide to be synthesized by coupling a protected α -amino acid by an amide bond to a suitable resin, e.g., benzhydrylamine (BHA), methylbenz-

hydramine (MBHA) or 4-(oxymethyl)-phenylacetamidomethyl (PAM) or 5-[(2' or 4')-aminomethyl-3',5'-dimethoxyphenoxy]-valerate (PAL). BHA, MBHA, PAM and PAL resin supports are commercially available.

5

All solvents used in the peptide preparations described herein, e.g. methylene chloride (CH_2Cl_2), 2-propanol, dimethylformamide (DMF), and methanol, were Burdick and Jackson "distilled in glass" grade and used without
10 additional distillation. Trifluoroacetic acid (TFA), diisopropylethylamine (DIPEA), piperidine (PIP), dicyclohexylcarbodiimide (DCC), 1-hydroxybenzotriazole (HOBT), and [benzotriazole-1-yl-oxy-tris(dimethyl) phosphonium hexafluorophosphate] (BOP) were purchased from
15 Chemical Dynamics Corp. and were "sequential" grade purity. 1,2-ethanedithiol (EDT) was purchased from Sigma Chemical Co. and used without further purification. All protected amino acids were of the L-configuration unless otherwise indicated and were obtained from Bachem.

20

In solid phase synthesis, the reactive side chain groups of the various amino acid moieties are typically protected with suitable protecting groups which will prevent a chemical reaction from occurring at that site until the
25 protecting group is ultimately removed. While specific protecting groups are disclosed in regard to the solid phase synthesis aspect, it should be noted that each amino acid can be protected by any protective group conventionally used for the respective amino acids in solution phase synthesis.
30 Purity of the protected amino acids was confirmed by thin layer chromatography (TLC), elemental analysis, IR, MS, NMR and optical rotation.

The following instrumentation was utilized. TLC was
35 performed on glass backed precoated silica gel 60 F254 plates purchased from Merck using appropriate solvent systems. Detection of spots was by UV fluorescence

quenching (254 nm absorption), iodine staining, or ninhydrin spray for primary and secondary amines.

For amino acid analyses, peptides were hydrolyzed in 6N HCl containing phenol at 115° C for 24 hours in evacuated Reacti-Therm hydrolysis tubes (Wheaton Scientific Company, Milville, N.J. 08332, USA). Analyses were performed on a Beckman 121M amino acid analyzer.

High pressure liquid chromatography (HPLC) was conducted on a laboratory data control (LDC) apparatus consisting of a Constametric I pump, a Constametric III pump, a Gradient Master solvent programmer and mixer, and a Spectromonitor III variable wavelength UV detector. Analytical HPLC chromatography was performed with Waters Micro Bondapack C₁₈ on reversed phase columns (0.4 x 25 cm). Preparative HPLC separations were run on (2.5 x 50 cm) Partisil M20 10/50 ODS-3 columns, or on (2.3 x 30) cm micro Bondapack C₁₈ column; in both cases, a pre-column of Whatman Co:Pell ODS pellicular packing was used. The peptides were assembled in a stepwise manner on a solid support using a Vega 1000 peptide synthesizer. The 1000 peptide synthesizer was controlled by an Apple IIe microprocessor with manual operations at steps 16 and 20 for the Boc-protocol and 7 and 10 for the Fmoc-protocol.

Boc-Phe (6.0g, 23 mmol) was coupled to the BHA resin (25g) using DCC (4.6g, 22 mmol) and HOBt (4.5g, 33 mmol) at 0°C. Loading was determined by amino acid analysis to be 0.31 mmol/g resin. Any unreacted amino groups were capped by treatment with 6 equivalents each of acetic anhydride and pyridine.

Boc-N-methyl-Phe (1.35g, 5 mmol) was coupled to the BHA resin (8g) using DCC (1.1g, 5 mmol) and HOBt (1g, 8 mmol) at 0°C. Loading was determined by amino acid analysis to be 0.30 mmol/g. Any unreacted amino groups were capped by

treatment with 6 equivalents each of acetic anhydride and pyridine.

Boc-Phe-Pam resin was purchased from Vega
 5 Biotechnologies (Tucson, Arizona, USA). The loading was
 0.32 mmol/g. The PAL-linker was purchased from Biosearch
 (San Rafael, California, USA). The initial synthesis was
 started with the Boc-amino acid resin and portions of
 peptide resin were removed at various points of the
 10 synthetic cycle for separate analog preparations. The
 protocol for a typical Boc-synthetic cycle was as follows:

	<u>Step</u>	<u>Reagent</u>	<u>Time</u>
	1	1% EDT/CH ₂ Cl ₂	1 x 30 sec.
	2	50% TFA/CH ₂ Cl ₂ 1% EDT	1 x 1 min.
15	3	Repeat Step 1	
	4	50% TFA/CH ₂ Cl ₂ 1% EDT	1 x 15 min.
	5	CH ₂ Cl ₂	1 x 30 sec.
	6	Methanol	1 x 30 sec.
	7-8	Repeat steps 5 and 6	
20	9	CH ₂ Cl ₂	2 x 30 sec.
	10	8% DIPEA	2 x 2 min.
	11-15	Repeat step 5-9	
	16	3 equiv. Boc-AA, DCC, HOBt	1 x 60 min.
	17	1% DIPEA	1 x 30 min.
25	18-19	Repeat steps 6 and 9	
	20-21	Repeat steps 16 and 17 if Kaiser test is positive	
	22	Methanol	1 x 30 sec.
	23-24	Repeat steps 5 and 6	
30	25	CH ₂ Cl ₂	1 x 30 sec.
	26	Methanol	2 x 30 sec.
	27	CH ₂ Cl ₂	3 x 30 sec.

The protocol for a typical Fmoc-synthetic cycle was as follows

	<u>Step</u>	<u>Reagent</u>	<u>Time</u>
	1	20% piperidine/DMF	1 x 5 min.
5	2	20% piperidine/DMF	1 x 5 min.
	3	DMF	2 x 1 min.
	4	CH ₂ Cl ₂	2 x 1 min.
	5	2-propanol	2 x 1 min.
	6	CH ₂ Cl ₂ /DMF	2 x 1 min.
10	7	3 equiv. Fmoc-AA, DCC, HOBT	1 x 60 min.
	8	CH ₂ Cl ₂	2 x 1 min.
	9	DMF	2 x 1 min.
	10-12	Repeat steps 7, 8, 9 if Kaiser test is positive	
15	13	CH ₂ Cl ₂	2 x 1 min.
	14	DMF	2 x 1 min.
	15	2-propanol	2 x 1 min.
	16	DMF	2 x 1 min.

20 Solvents for all washings and couplings were measured to volumes of 10-20 ml/g resin. Couplings were performed using the DCC/HOBT procedure. Coupling reactions were monitored by the Kaiser ninhydrin test to determine whether coupling was complete at step 19 by the Boc-synthetic protocol or at step 25 9 by the Fmoc-synthetic protocol as set forth by Kaiser et al., Analytical Biochemistry 34, 595-598 (1970).

The fully assembled peptide-resins were dried under high vacuum overnight and cleaved with appropriate cleavage 30 reagents. For the Boc-synthesis the modified procedures of Tam et al. [Tetrahedron Letter, 23, 4435-4438 (1982)] were used. In brief: The peptide-resin was treated in a teflon HF apparatus (Peninsula) with HF, dimethylsulfide and p-cresol (5:13:2) for 1h at 0°C. After evaporation to a low 35 volume fresh anhydrous HF was distilled into the reaction vessel (18 ml) for a second treatment for 1.5 h at 0°C. After thorough evaporation, the dry resin was washed with 3

volumes each of Et₂O and EtOAc, then triturated with 4 x 15 ml of 30% acetic acid and filtered. Lyophilization of the aqueous filtrate yielded the crude peptide.

- 5 For the Fmoc-synthesis the procedure of Mitchell et al. [J. Org. Chem., 43, 2845, (1978)] was used. In brief: The peptide-resin was placed into a pressure bottle, suspended in methanol, saturated with NH₃ at -20°C and sealed. The suspension was stirred at room temperature for 2-3 days.
- 10 After venting the excess NH₃, the PAM-resin was filtered off and washed with methanol. The filtrate was evaporated to dryness to give the crude peptide.

- 15 Preparative purification was carried out directly with the crude peptide by HPLC on a (2.3 x 30) cm micro Bondapack C₁₈ or (2.5 x 50) cm Whatman ODS-3 column. The peptides were applied in a minimum volume of 50% AcOH, and eluted with a slow gradient (4 hr) of 5-65%, 0.022% TFA/CH₃CN, at a flow rate of 8.0 ml/min. Fractions were collected at 3
- 20 minute intervals and cuts were made after inspection by analytical HPLC. Fractions, judged to be greater than 97% pure, were pooled and lyophilized.

- 25 Purity of the individual peptides was checked by HPLC and determined to be 99% in all cases. Amino acid analyses of the individual peptides were performed and the expected values were obtained in each case. U.V., N.M.R. and M.S. were also performed on the analogs confirming the chemical integrity of the peptides.

- 30 The separation of peptides containing (D,L) amino acids into their optically pure enantiomers was achieved by preparative HPLC on (2.3 x 30) cm micro Bondapack C₁₈ column from E.S. Industries using CH₃CN/0.01M NH₄OAc.

- 35 The chirality of the amino acids were determined by the method of Bayer et al. [H. Frank, W. Woiwode, G. Nicholson

and E. Bayer, Liebig Ann. Chem. 354, (1981)] using glass capillary gas chromatography.

5 The present invention will be further described in connection with the following examples which are set forth for the purposes of illustration only.

For the preparation of the compounds described in Examples 1-36, melting points were taken on a Buchi 510
10 melting point apparatus and are uncorrected. Preparative high pressure liquid chromatography (HPLC) was performed on silica gel Pre-Pak 500 cartridges using a Waters Associates Prep LC 500A. Dry dichloromethane was distilled from P_2O_5 . DMF was dried over Linde 3A molecular sieves and
15 triethylamine was distilled from calcium hydride. Concentration refers to evaporation under aspirator vacuum using a Buchi rotary evaporator.

20

25

30

35

EXAMPLE 1

Preparation of (S)-4-[[[(trifluoromethyl)sulfonyl]oxy]-
alpha-[[[(1,1-dimethylethoxy)carbonyl]amino]benzenepropanoic
5 acid methyl ester

A solution of 19.23 g (0.065 mol) of (S)-4-hydroxy-
alpha-[[[(1,1-dimethylethoxy)carbonyl]amino]benzenepropanoic
acid methyl ester and 24.0 g (0.067 mol) of N-phenyl-N-
10 trifluoromethylsulfonyl-1,1,1-trifluoromethane sulfonamide
in 175 ml of dry dichloromethane was cooled in an ice bath
and 9.7 ml (0.70 mol) of triethylamine was added over three
minutes. The resulting mixture was held at 0°C for 1 hour
and allowed to warm to room temperature over 1 hour. The
15 reaction mixture was diluted with 500 ml of ether and washed
successively with water (1 X 100 ml), 1 N sodium hydroxide
solution (2 X 100 ml), water (1 X 100 ml), and saturated
sodium chloride solution (1 X 100 ml). The organic phase
was dried over magnesium sulfate and concentrated to an oil
20 which was purified by preparative liquid chromatography
using silica gel cartridges on a Waters Prep 500
chromatograph, eluting with 20% ethyl acetate-hexane. The
pure fractions were combined and evaporated to give 26.78 g
(96%) of (S)-4-[[[(trifluoromethyl)sulfonyl]oxy]-alpha-
25 [[[(1,1-dimethylethoxy)carbonyl]amino]benzenepropanoic acid
methyl ester as a colorless oil which crystallized on
standing, melting point (mp) 48-49°C.

EXAMPLE 2

30

Preparation of (S)-alpha-[[[(1,1-dimethylethoxy)carbonyl]-
amino]-4-(2-propenyl)benzenepropanoic acid methyl ester

35

Argon was passed through a solution of 7.0 g (0.0164
mol) of (S)-4-[[[(trifluoromethyl)sulfonyl]oxy]-alpha-[[[(1,1-
dimethylethoxy)carbonyl]amino]benzenepropanoic acid methyl
ester, 5.7 g (0.0165 mol) of allyltributyl tin, and 1.42 g

(0.04 mol) of lithium chloride in 50 ml of dimethylformamide for 10 minutes and 210 mg (0.0003 mol) of bis(triphenylphosphine)palladium dichloride was added. The bath temperature was raised to 90-95°C for 40 minutes and the mixture was allowed to cool. The mixture was diluted with ether and washed with water and saturated sodium chloride solution and dried over magnesium sulfate. The residue obtained after filtration and evaporation was purified by preparative liquid chromatography using silica gel cartridges on a Waters Prep 500 chromatograph, eluting with 10% ethyl acetate-hexane to give 4.83 g (92%) of (S)-alpha-[[[(1,1-dimethylethoxy)carbonyl]amino]-4-(2-propenyl)benzenepropanoic acid methyl ester, mp 56-59°C.

Example 3

Preparation of (S)-4-carboxymethyl-alpha-[[[(1,1-dimethylethoxy)carbonyl]amino]benzenepropanoic acid methyl ester

To a solution of 4.00 g (0.0125 mol) of (S)-alpha-[[[(1,1-dimethylethoxy)carbonyl]amino]-4-(2-propenyl)-benzenepropanoic acid methyl ester in 80 ml of carbon tetrachloride and 80 ml of acetonitrile was added a solution of 8.00 g (0.037 mol) of sodium metaperiodate in 200 ml of water. The two phase mixture was stirred mechanically and 0.2 g (0.0001 mol) of ruthenium trichloride hydrate was added. The resulting dark mixture was stirred at room temperature for 1 hour and was diluted with 500 ml of dichloromethane. The layers were separated and the organic layer was washed with water and was dried over magnesium sulfate. Filtration and evaporation gave 3.84 g of a dark oil. This oil was dissolved in 120 ml of t-butyl alcohol and 40 ml of 2-methyl-2-butene and a solution of 12.6 g of sodium chlorite and 12.6 g of sodium dihydrogen phosphate in 80 ml of water was added. The resulting mixture was stirred mechanically for 2 hours. The mixture was diluted with

500 ml of ether, the layers were separated, the organic phase was washed with 100 ml portions of water, 10% sodium thiosulfate and saturated sodium chloride solution and was dried over magnesium sulfate. The residue obtained after filtration and concentration was chromatographed over 150 g of silica gel, eluting with 40:59:1 ethyl acetate-hexane-acetic acid. The earlier fractions contained 0.631 g of a mixture from which (S)-4-carboxy-alpha-[[[(1,1-dimethylethoxy)carbonyl]amino]benzenepropanoic acid methyl ester was obtained by crystallization from ether-hexane, mp 101-104°C, $[\alpha]_D +4.00$ (ethanol). The later fractions were combined, diluted with toluene and evaporated to remove traces of acetic acid and were evacuated under high vacuum for 72 hours to give 3.078 g of (S)-4-carboxymethyl-alpha-[[[(1,1-dimethylethoxy)carbonyl]amino]benzenepropanoic acid methyl ester as a colorless oil, dicyclohexylamine salt, mp 141-143°C, $[\alpha]_D +7.25$ (ethanol).

EXAMPLE 4

Preparation of (S)-alpha-[[[(1,1-dimethylethoxy)-carbonyl]amino]-4-[2-(1,1-dimethylethoxy)-2-oxoethyl]benzene-propanoic acid methyl ester

a. A solution of 2.85 g of (S)-4-carboxymethyl)alpha-[[[(1,1-dimethylethoxy)carbonyl]amino]benzenepropanoic acid methyl ester in 12 ml of t-butyl alcohol was treated with 2.06 g of dicyclohexylcarbodiimide followed by 0.13 g of 4-pyrrolidinopyridine. A white precipitate formed after a few minutes and the reaction mixture was allowed to stir for 18 hours. The mixture was filtered, and the solid washed with ether. The ether extracts and the filtrate were combined and washed with 1N hydrochloric acid, water, and saturated sodium bicarbonate solution and were dried over magnesium sulfate. The residue obtained after filtration and evaporation was chromatographed over 150 g of silica gel, eluting with 20% ethyl acetate-hexane to afford 1.613 g

of (S)-alpha-[[[(1,1-dimethylethoxy)carbonyl]amino]-4-[2-(1,1-dimethylethoxy)-2-oxoethyl]benzenepropanoic acid methyl ester as a colorless oil, $[\alpha]_D +6.07$ (ethanol).

5 b. A solution of 3.50 g of (S)-4-carboxymethyl)-alpha-
[[[(1,1-dimethylethoxy)carbonyl]amino]benzenepropanoic acid
methyl ester in 40 ml of dry toluene and 10 ml of
dimethylformamide di-t-butyl acetal was heated to a bath
temperature of 80°C for 4 hours. After cooling, the mixture
10 was diluted with 100 ml of ether and was washed with water
and saturated sodium chloride solution and was dried over
magnesium sulfate. The residue obtained after filtration
and evaporation was chromatographed over 150 g of silica
gel, eluting with 20% ethyl acetate-hexane to give 2.663 g
15 of (S)-alpha-[[[(1,1-dimethylethoxy)carbonyl]amino]
-4-[2-(1,1-dimethylethoxy)-2-oxoethyl]benzenepropanoic acid
methyl ester.

EXAMPLE 5

20 Preparation of (S)-alpha-[[[(1,1-dimethylethoxy)carbonyl]
amino]-4-[2-(1,1-dimethylethoxy)-2-oxoethyl]-
benzenepropanoic acid

25 A solution of 1.471 g of (S)-alpha-[[[(1,1-dimethyl-
ethoxy)carbonyl]amino]-4-[2-(1,1-dimethylethoxy)-2-oxoethyl]-
benzenepropanoic acid methyl ester in 25 ml of methanol and
5 ml of 1 N sodium hydroxide solution was stirred at room
temperature for 2 hours. The mixture was acidified with a
30 slight excess of hydrochloric acid, was diluted with 100 ml
of ether, and was washed with water and saturated sodium
chloride solution. The residue obtained after filtration
and evaporation was chromatographed over 100 g of silica
gel, eluting with 40:59:0.5 ethyl acetate-hexane-acetic
35 acid. The product containing fractions were combined,
evaporated, diluted with toluene and evaporated finally
under high vacuum to give 1.105 g of (S)-alpha-[[[(1,1-

-dimethylethoxy)carbonyl]amino]-4-[2-(1,1-dimethylethoxy)-
-2-oxoethyl]benzenepropanoic acid $[\alpha]_D + 21.41$ (ethanol).

EXAMPLE 6

5

Preparation of 5-(4-methylphenyl)-2-(1,1-dimethylethyl)-
2H-tetrazole

10

15

20

A solution of 5-(4-methylphenyl)-2H-tetrazole [Finnegan,
W.G. et al., J. Am. Chem. Soc. 80, 3908, (1958)] (19.9 g,
0.12 mol), t-butanol (19.5 g, 0.26 mol) and concentrated
sulfuric acid (5.84 g, 0.06 mol) in trifluoroacetic acid
(122 ml) was stirred at room temperature (3 h) and then
diluted with ethyl acetate (250 ml). The mixture was washed
sequentially with water (2 x 50 ml), 10% aqueous sodium
hydroxide until washings were basic, water (2 x 30 ml), and
then dried over sodium sulfate (Na_2SO_4) and filtered.
Following removal of the solvents in vacuo, the resultant
oil was purified by HPLC on silica gel eluting with 20:1
mixture of hexanes-ethyl acetate to provide 17.5 g of
5-(4-methylphenyl)-2-(1,1-dimethylethyl)-2H-tetrazole as a
clear oil, bp. 120-122°C (0.3 mm Hg).

EXAMPLE 7

25

Preparation of benzyl bromide derivatives

30

35

N-Bromosuccinimide (NBS) (35.8 g, 0.20 mol) was added in
several portions to a solution of 1,1-dimethylethyl
4-methylbenzoate (prepared by the reaction of the respective
acid chloride with t-butanol.) (38.0g, 0.20 mol) and benzoyl
peroxide (0.36 g, 1.5 mmol) in carbon tetrachloride
(1200 ml) at vigorous reflux. Another portion of benzoyl
peroxide (0.36 g, 1.5 mmol) was added just before the final
addition of NBS. After the foaming subsided, the reaction
mixture was cooled to room temperature, washed with water
(2 x 60 ml) and dried over magnesium sulfate. Filtration

and concentration provided an oil that was purified by HPLC on silica gel eluting with a mixture of hexanes and ethyl acetate to give 1,1-dimethylethyl 4-bromomethylbenzoate (21.4 g) in 40% yield.

5

Following the above procedure, the analogs listed below were prepared.

10 a. From 1,1-dimethylethyl 3-methylbenzoate (40.0 g, 0.21 mol) there was obtained 1,1-dimethylethyl 3-bromomethylbenzoate (23.0 g, 0.09 mol) in 41% yield.

15 b. From 1,1-dimethylethyl 2-(4-methylphenyl)acetate (25.0 g, 0.12 mol) there was obtained 1,1-dimethylethyl 2-(4-bromomethylphenyl)acetate (10.9 g, 0.04 mol) in 32% yield.

20 c. From 5-(4-methylphenyl)-2-(1,1-dimethylethyl)-2H-tetrazole (18 g, 0.08 mol) there was obtained 5-(4-bromomethylphenyl)-2-(1,1-dimethylethyl)-2H-tetrazole (17.4 g, 0.06 mol), in 71% yield.

25 d. From 1,1-dimethylethyl 2-(4-methylphenyl)-2,2-difluoroacetate (8.0 g, 0.03 mol) there was obtained 1,1-dimethylethyl 2-(4-bromomethylphenyl)-2,2-difluoroacetate (7.7 g, 0.024 mol) in 73% yield.

EXAMPLE 8

30 Preparation of 5-[(4-bromophenyl)methyl]-2H-tetrazole

4-Bromophenylacetonitrile (30.0 g, 0.15 mol), sodium azide (10.9 g, 0.17 mol) and ammonium chloride (8.9 g, 0.17 mol) were heated in DMF (300 ml) at 90°C for 2 days.
35 After concentration, water (200 ml) was added to the residue, the mixture was basified with 1 M NaOH (170 ml) and washed with ether (2 x 100 ml). Acidification of the

aqueous layer with 1N HCl and collection of the precipitate by suction filtration produced the crude product. This was purified by recrystallization from ethanol to provide 5-[(4-bromophenyl)methyl]-2H-tetrazole (17.2 g, 0.07 mol) in 44% yield, mp. 173-175°C.

EXAMPLE 9

Preparation of 5-[(4-bromophenyl)methyl]-2-(1,1-dimethylethyl)-2H-tetrazole

A mixture of 5-[(4-bromophenyl)methyl]-2H-tetrazole (18.5 g, 77 mmol), t-butanol (11.4 g, 150 mmol), trifluoroacetic acid (76 ml, 1.0 mol) and concentrated sulfuric acid (3.8 g, 39 mmol) was stirred for 24 h and then partitioned between ethyl acetate (250 ml) and water (100 ml). The organic layer was washed with water (4 x 100 ml) and 1 M NaOH (2 x 100 ml) and was dried over sodium sulfate. Filtration and removal of the solvent in vacuo produced a solid that was recrystallized from hexane to give 5-[(4-bromophenyl)methyl]-2-(1,1-dimethylethyl)-2H-tetrazole. (12.3 g, 47 mmol) in 54% yield, mp. 69-70°C.

EXAMPLE 10

Preparation of 4-[[2-(1,1-dimethylethyl)-2H-tetrazol-5-yl]methyl]benzoic acid

To a deoxygenated mixture of 5-[(4-bromophenyl)methyl]-2-(1,1-dimethylethyl)-2H-tetrazole (12.0 g, 41 mmol), triphenylphosphine (11.0 g, 42 mmol), tributylamine (8.4 g, 45 mmol), t-butanol (70 ml) and water (45 ml) was added bis(triphenylphosphine)palladium dichloride (1.0 g, 13 mmol). This mixture was placed in a glass lined autoclave and maintained under carbon monoxide (200 psi) at 100°C for 2 days. The reaction mixture was diluted with dichloromethane (500 ml), washed with water (3 x 100 ml) and

dried over magnesium sulfate. Filtration and concentration produced the crude product mixture. This was purified by flash chromatography on silica gel using a 4:1 mixture of hexane and ethyl acetate to remove the less polar impurities and 5% acetic acid in hexane-ethyl acetate to elute 4-[[2-(1,1-dimethylethyl)-2H-tetrazole-5-yl]methyl]benzoic acid (4.2 g, 16 mmol) in 49% yield, mp. 139-140°C.

EXAMPLE 11

Preparation of 4-[[2-(1,1-dimethylethyl)-2H-tetrazol-5-yl]-methyl]benzyl alcohol

Diisobutylaluminum hydride (48 ml of a 0.8 M solution in toluene, 38 mmol) was added dropwise to a mixture of 4-[[2-(1,1-dimethylethyl)-2H-tetrazole-5-yl]methyl]benzoic acid (2.5 g, 9.6 mmol) in toluene (250 ml) at 0°C and the mixture was then warmed to room temperature. After stirring overnight, the reaction was quenched at 0°C by careful addition of a mixture of water and THF (8 ml, 5:3), 10% NaOH (5 ml) and a final portion of water (15 ml). After the addition of ether (200 ml) and stirring at room temperature for 2 hours, the white precipitate was removed by filtration and the filtrate was concentrated. The residue was purified by HPLC on silica gel eluting with a 7:3 mixture of hexane-ethyl acetate to give 4-[[2-(1,1-dimethylethyl)-2-tetrazol-5-yl]methyl]benzylalcohol (1.6 g, 6.7 mmol) in 70% yield, mp. 65-66°C.

EXAMPLE 12

Preparation of 5-[[4-(Bromomethyl)phenyl]methyl]-2-(1,1-dimethylethyl)-2H-tetrazole

Triphenylphosphine (6.5 g, 25 mmol) was added to a solution of 4-[[2-(1,1-dimethylethyl)-2H-tetrazol-5-yl]-methyl]benzenemethanol (3.0 g, 12.4 mmol) and carbon

tetrabromide (8.3 g, 25 mmol) in diethyl ether (150 ml). After 3 hours the reaction mixture was concentrated and the residue purified by chromatography on silica gel eluting with a 9:1 mixture of hexane-ethyl acetate to give

5 5-[[4-(bromomethyl)phenyl]methyl]-2-(1,1-dimethylethyl)-2H-tetrazole (2.4 g, 7.8 mmol) in 63% yield, mp. 73-74°C.

EXAMPLE 13

10 Preparation of benzyl phenylalaninate-benzophenone imine derivatives

Tetrabutyl ammonium sulfate (3.0 g, 9 mmol) was added to a biphasic mixture consisting of 10% aqueous sodium
15 hydroxide (75 ml), 2-[(diphenylmethylene)amino]acetic acid benzyl ester (3.05 g, 9.3 mmol) and 1,1-dimethylethyl 4-bromomethylbenzoate (3.0 g, 9 mmol) in dichloromethane (58 ml). After several hours of vigorous stirring at room temperature, the reaction mixture was diluted with ether
20 (300 ml) and the layers were separated. The organic layer was washed with water (3 x 30 ml), dried (K_2CO_3) and filtered. Concentration provided an oil that was purified by HPLC on silica gel (the columns were pre-treated with 10% triethylamine/hexanes, ethyl acetate and finally hexanes)
25 eluting with a mixture of hexanes-ethyl acetate to give rac.-2-[(diphenylmethylene)amino]-3-[4-[(1,1-dimethylethoxy)carbonyl]phenyl]propanoic acid benzyl ester, (1.7 g, 3 mmol) in 36% yield.

30 Following the above procedure, the analogs listed below were prepared.

a. From 1,1-dimethylethyl 3-bromomethylbenzoate (10.0 g, 37 mmol) there was obtained rac.-2-[(diphenylmethylene) amino]-3-[3-[(1,1-dimethylethoxy)carbonyl]phenyl]propanoic
35 acid benzyl ester, (9.4 g, 18 mmol) in 49% yield.

b. From 1,1-dimethylethyl 4-bromomethylphenylacetate (3.1 g, 11 mmol) there was obtained rac.-2-[(diphenylmethylene)amino]-3-[4-[2-(1,1-dimethylethoxy)-2-oxoethyl]phenyl]propanoic acid benzyl ester, (2.9 g, 5 mmol) in 50% yield.

c. From 5-(4-bromomethylphenyl)-2-(1,1-dimethylethyl)-2H-tetrazole (16.0 g, 54 mmol) there was obtained rac.-2-[(diphenylmethylene)amino]-3-[4-[5-[(2-(1,1-dimethylethyl)-2H-tetrazoyl)]phenyl]propanoic acid benzyl ester, (10.9 g, 32 mmol) in 59% yield.

d. From 5-(4-bromomethylphenylmethyl)-2-(1,1-dimethylethyl)-2H-tetrazole (2.0 g, 6.5 mmol) there was obtained rac.-2-[(diphenylmethylene)amino]-3-[4-[5-[2-(1,1-dimethylethyl)-2H-tetrazoyl]]methyl]phenyl] propanoic acid benzyl ester, (1.9 g, 3.3 mmol) in 50% yield.

EXAMPLE 14

Preparation of rac.-2-[(diphenylmethylene)amino]-3-[4-[1,1-difluoro-[2-(1,1-dimethylethoxy)-2-oxoethyl]phenyl]propanoic acid benzyl ester

Lithium diisopropyl amide (8.8 mmol) was generated in 25 ml of THF at 0°C and chilled to -78°C. A solution of 2-[(diphenylmethylene)amino]acetic acid benzyl ester (2.9 g, 8.8 mmol) in THF (5 ml) was added over 5 minutes followed by the dropwise addition of hexamethylphosphoramide (1.6 g, 8.8 mmol) and the mixture was stirred for 15 min. A solution of 1,1-dimethylethyl 2-(4-bromomethylphenyl)-2,2-difluoroacetate (2.7 g, 8.0 mmol) in 5 ml THF was added slowly and the reaction mixture was warmed over the course of 2 hours to room temperature. The mixture was partitioned between saturated aqueous NH_4Cl (50 ml) and ether (30 ml). The aqueous layer was extracted with additional ether (2 x 30 ml), and the combined organic layers were washed

with brine (20 ml), and dried (MgSO_4). Filtration and concentration yielded an oil that was purified by flash chromatography on silica gel (pretreated with 10% triethylamine in hexanes) eluting with a 9:1 mixture of hexane-ethyl acetate. This provided
5 rac.-2-[(diphenylmethylene)amino]-3-[4-[1,1-difluoro-[2-(1,1-dimethylethoxy)-2-oxoethyl]phenyl] propanoic acid benzyl ester (3.2 g, 5.6 mmol), as a colorless oil in 71% yield.

10

EXAMPLE 15

Preparation of benzyl phenylalaninate p-toluenesulfonic acid salt derivatives

15

A solution of rac.-2-[(diphenylmethylene)amino]-3-[4-[(1,1-dimethylethoxy)carbonyl]phenyl]propanoic acid benzyl ester (11.5 g, 22 mmol) and p-toluenesulfonic acid monohydrate (4.18 g, 22 mmol) in a 10/1 mixture of
20 acetonitrile and water (640 ml) was stirred for 2 h at room temperature. Concentration provided a crude solid product which was recrystallized from ethanol-ether to give rac.-2-amino-3-[4-[(1,1-dimethylethoxy)carbonyl]phenyl]propanoic acid benzyl ester p-toluenesulfonic acid salt, (8.1 g, 15
25 mmol) in 67% yield, mp. 154-157°C.

Following the previous procedure, the analogs listed below were prepared.

30

a. From rac.-2-[(diphenylmethylene)amino]-3-[3-[(1,1-dimethylethoxy)carbonyl]phenyl]propanoic acid benzyl ester (9.4 g, 18 mmol) there was obtained rac.-2-amino-3-[3-[(1,1-dimethylethoxy)carbonyl]phenyl]propanoic acid benzyl ester p-toluenesulfonic acid salt which was carried on to the next
35 step without purification.

b. From rac.-2-[(diphenylmethylene)amino]-3-[4-[2-(1,1-dimethylethoxy)-2-oxoethyl]phenyl]propanoic acid benzyl ester (6.9 g, 13 mmol) there was obtained rac.-2-amino-3-[4-[2-(1,1-dimethylethoxy)-2-oxoethyl]phenyl]propanoic acid benzyl ester p-toluenesulfonic acid salt, (6.1 g, 11 mmol) in 87% yield, mp. 139-141°C.

c. From rac.-2-[(diphenylmethylene)amino]-3-[4-[5-[2-(1,1-dimethylethyl)-2H-tetrazoyl]phenyl]propanoic acid benzyl ester (8.0 g, 15 mmol) there was obtained rac.-2-amino-3-[4-[5-[2-(1,1-dimethylethyl)-2H-tetrazoyl]]phenyl]propanoic acid benzyl ester p-toluenesulfonic acid salt, (6.6 g, 12 mmol) in 81% yield, mp. 204-205°C.

d. From rac.-2-[(diphenylmethylene)amino]-3-[4-[1,1-difluoro-2-(1,1-dimethylethoxy)-2-oxoethyl]phenyl]propanoic acid benzyl ester (3.2 g, 5.6 mmol) there was obtained rac.-2-amino-3-[4-[1,1-difluoro-2-(1,1-dimethylethoxy)-2-oxoethyl]phenyl]propanoic acid benzyl ester p-toluenesulfonic acid salt, (2.3 g, 3.9 mmol) in 70% yield, mp. 145-149°C.

e. From rac.-2-[(diphenylmethylene)amino]-3-[4-[[5-[2-(1,1-dimethylethyl)-2H-tetrazoyl]methyl]phenyl]propanoic acid benzyl ester (4.4 g, 7.9 mmol) there was obtained rac.-2-amino-3-[4-[[5-[2-(1,1-dimethylethyl)-2H-tetrazoyl]methyl]phenyl]propanoic acid benzyl ester p-toluenesulfonic acid salt, (3.0 g, 5.3 mmol) in 67% yield, mp. 151-154°C.

EXAMPLE 16

Preparation of rac.-benzyl N-acetylphenylalaninate derivatives

Acetic anhydride (1.0 ml, 10.8 mmol) and triethylamine (2.4 g, 24 mmol) were added sequentially to a solution of

rac.-2-amino-3-[4-[(1,1-dimethylethoxy)carbonyl]phenyl]
propanoic acid benzyl ester p-toluenesulfonic acid salt
(5.0 g, 9 mmol) in dichloromethane (100 ml) at 0°C. After
2 h, the reaction mixture was diluted with dichloromethane
5 (250 ml) and washed with 1 N HCl (2 x 20 ml), saturated
aqueous sodium bicarbonate (2 x 20 ml) and dried
(Na₂SO₄). Filtration and concentration provided the
crude product which was purified by HPLC on silica gel
eluting with a mixture of hexanes and ethyl acetate to give
10 rac.- 2-acetamido-3-[4-[(1,1-dimethylethoxy)carbonyl]-
phenyl]propanoic acid benzyl ester (3.3 g, 8 mmol) in 91%
yield.

Following the above procedure, the analogs listed below
15 were prepared.

a. From rac.-2-amino-3-[3-[(1,1-dimethylethoxy)-
carbonyl]phenyl]propanoic acid benzyl ester
p-toluenesulfonic acid salt obtained as the crude product in
20 the hydrolysis of the corresponding diphenylmethyl imine,
there was obtained rac.-2-acetamido-3-[3-[(1,1-dimethyl-
ethoxy)carbonyl]phenyl]propanoic acid benzyl ester as an oil.

b. From rac.-2-amino-3-[4-[2-(1,1-dimethylethoxy)-2-
25 oxoethyl]phenyl]propanoic acid benzyl ester
p-toluenesulfonic acid salt (5.9 g, 11 mmol) there was
obtained rac.-2-acetamido-3-[4-[2-(1,1-dimethylethoxy)-2-
oxoethyl]]phenyl]propanoic acid benzyl ester, (3.1 g, 8.4
mmol) in 76% yield.

c. From rac.-2-amino-3-[4-[5-[2-(1,1-dimethylethyl)-
2H-tetrazoyl]]phenyl]propanoic acid benzyl ester
p-toluenesulfonic acid salt (2.0 g, 3.8 mmol) there was
obtained rac.-2-acetamido-3-[4-[5-[2-(1,1-dimethylethyl)-
30 2H-tetrazoyl]]phenyl]propanoic acid benzyl ester, (1.4 g,
3.3 mmol) in 87% yield.

d. From rac.-2-amino-3-[4-[1,1-difluoro-[2-(1,1-dimethylethoxy)-2-oxoethyl]]phenyl]propanoic acid benzyl ester p-toluenesulfonic acid salt (2.0 g, 3.4 mmol) there was obtained rac.-2-acetamido-3-[4-[1,1-difluoro-[2-(1,1-dimethylethoxy)-2-oxoethyl]]phenyl]propanoic acid benzyl ester, (1.2 g, 2.7 mmol) in 80% yield, mp. 98-100°C (ethyl acetate/hexanes).

e. From rac.-2-amino-3-[4-[[5-[2-(1,1-dimethylethyl)-H-tetrazoyl]]methyl]phenyl]propanoic acid benzyl ester p-toluenesulfonic acid salt (4.3 g, 7.4 mmol) there was obtained rac.-2-amino-3-[4-[[5-[2-(1,1-dimethylethyl)-2H-tetrazoyl]]methyl]phenyl]propanoic acid benzyl ester, (2.0 g, 4.6 mmol) in 63% yield, mp. 113-114.5°C (ethyl acetate/hexanes).

EXAMPLE 17

Preparation of N-acetylphenylalanine derivatives

A suspension of rac.-2-acetamido-3-[4-[(1,1-dimethylethoxy)carbonyl]phenyl]propanoic acid benzyl ester (3.3 g, 8 mmol) and 10 % Pd/C (320 mg) in ethanol (100 ml) was blanketed with hydrogen (1 atm) at room temperature. Upon the consumption of 1 equivalent of hydrogen, the mixture was filtered through celite. The celite pad was washed with ethanol (50 ml) and the filtrates were combined and concentrated. The resultant solid was purified by recrystallization from ethyl acetate/hexane to give rac.-2-acetamido-3-[4-[(1,1-dimethylethoxy)carbonyl]phenyl]propanoic acid, (2.3 g, 7.4 mmol) in 89% yield, mp. 187-188°C.

Following the above procedure, the analogs listed below were prepared.

a. From rac.-2-acetamido-3-[3-[(1,1-dimethylethoxy)-carbonyl]phenyl]propanoic acid benzyl ester (3.1 g, 7.8 mmol) there was obtained rac.-2-acetamido-3-[3-[(1,1-dimethylethoxy)carbonyl]phenyl]propanoic acid, (1.8 g, 5.8 mmol) in 75% yield, mp. 164-166°C (ethyl acetate/hexanes).

b. From rac.-2-acetamido-3-[4-[-2-(1,1-dimethylethoxy)-2-oxoethyl]phenyl]propanoic acid benzyl ester (2.6 g, 6.5 mmol) there was obtained rac.-2-acetamido-3-[4-[-2-(1,1-dimethylethoxy)-2-oxoethyl]phenyl]propanoic acid, (1.9 g, 5.9 mmol) in 91% yield, mp. 170-172°C (ethyl acetate/hexanes).

c. From rac.-2-acetamido-3-[4-[5-[2-(1,1-dimethylethyl)-2H-tetrazoyl]]phenyl]propanoic acid benzyl ester (3.2 g, 7.6 mmol) there was obtained rac.-2-acetamido-3-[4-[5-[(2-(1,1-dimethylethyl)-2H-tetrazoyl)]phenyl]propanoic acid, (2.2 g, 6.5 mmol) in 86% yield, mp. 206-207°C (ethyl acetate/hexanes).

d. From rac.-2-acetamido-3-[4-[1,1-difluoro-[2-(1,1-dimethylethoxy)-2-oxoethyl]phenyl]propanoic acid benzyl ester (1.1 g, 2.5 mmol) there was obtained rac.-2-acetamido-3-[4-[1,1-difluoro[2-(1,1-dimethylethoxy)-2-oxoethyl]phenyl]propanoic acid, (0.6 g, 1.7 mmol) in 69% yield, mp. 149-150.5°C (acetonitrile).

e. From rac.-2-acetamido-3-[4-[[5-[2-(1,1-dimethylethyl)-2H-tetrazoyl]]methyl]phenyl]propanoic acid benzyl ester (2.0 g, 4.6 mmol) there was obtained rac.-2-acetamido-3-[4-[[5-[2-(1,1-dimethylethyl)-2H-tetrazoyl]]methyl]phenyl]propanoic acid, (1.2 g, 3.5 mmol) in 76% yield, mp. 193-194.5°C (acetonitrile).

EXAMPLE 18

Preparation of (S)-2-[[[(1,1-dimethylethoxy)carbonyl]-
amino]-3-[4-[[[(trifluoromethyl)sulfonyl]oxy]phenyl]-
5 propanoic acid diphenylmethyl ester

Triethylamine (4.0 g, 39 mmol) was added dropwise to a
suspension of (S)-2-[[[(1,1-dimethylethoxy)carbonyl]-
amino]-3-(4-hydroxyphenyl)propanoic acid diphenylmethyl
10 ester (17.0 g, 38 mmol) and N-phenyl-N-trifluorosulfonyl
-1,1,1-trifluoromethyl sulfonamide (13.6 g, 38 mmol) in
dichloromethane (170 ml) at 0°C. After 1 hour the reaction
mixture was warmed to room temperature and washed with 1 N
sodium hydroxide (2 x 25 ml), saturated aqueous sodium
15 bicarbonate (2 x 25 ml) and water (2 x 25 ml). The organic
layer was dried (K_2CO_3), filtered and concentrated.
Recrystallization of the solid from hexane gave
(S)-2-[[[(1,1-dimethylethoxy)carbonyl]amino]-3-[4-[[[(
trifluoromethyl)sulfonyl]oxy]phenyl]]propanoic acid
20 diphenylmethyl ester (13.7 g, 24 mmol) in 62% yield,
mp. 110-112°C, $[\alpha]_D -14.23^\circ$ (0.12% in ethanol).

EXAMPLE 19

25 Preparation of (S)-2-[[[(1,1-dimethylethoxy)carbonyl]-
amino]-3-[4-[3-(1,1-dimethylethoxy)3-oxo-1-propenyl]phenyl]
propanoic acid diphenylmethyl ester

Bis(triphenylphosphine) palladium dichloride (120 mg,
30 0.2 mmol) was added to a deoxygenated mixture of
(S)-2-[[[(1,1-dimethylethoxy)carbonyl]amino]-3-[4-
[[[(trifluoromethyl)sulfonyl]oxy]phenyl]]propanoic acid
diphenylmethyl ester (3.0 g, 5.2 mmol), t-butyl acrylate
(1.5 ml, 10.2 mmol) and triethylamine (4.0 ml, 30.0 mmol) in
35 DMF (50 ml) and the well-stirred suspension was then heated
at 90°C (24 h). The solvent was removed in vacuo and the
resulting residue was purified by flash chromatography on

silica gel eluting with a mixture of dichloromethane and hexanes (8:1) to give the (S)-2-[[[(1,1-dimethylethoxy)carbonyl]amino]-3-[4-[3-(1,1-dimethylethoxy)-3-oxo-1-propenyl]phenyl]propanoic acid diphenylmethyl ester (1.6 g, 2.8 mmol) in 54% yield, mp. 135-137°C, $[\alpha]_D -5.35^\circ$ (0.97% in ethanol).

EXAMPLE 20

Preparation of (S)-2-[[[(1,1-dimethylethoxy)carbonyl]amino]-3-[4-[3-(1,1-dimethylethoxy)-3-oxopropyl]phenyl]propanoic acid

(S)-2-[[[(1,1-Dimethylethoxy)carbonyl]amino]-3-[4-[3-(1,1-dimethylethoxy)-3-oxo-1-propenyl]phenyl]propanoic acid diphenylmethyl ester (1.6 g, 2.8 mmol) and 10 % Pd/C (300 mg) in t-butanol (20 ml) was blanketed with hydrogen (1 atm). To prevent the solvent from freezing, a warm water bath (40°C) was used to heat the reaction mixture. Upon the consumption of 2 equivalents of hydrogen, the mixture was filtered through celite. The celite pad was washed with ethanol (50 ml) and the filtrates combined and concentrated. The residue was purified by recrystallization from hexanes to give (S)-2-[[[(1,1-dimethylethoxy)carbonyl]amino]-3-[4-[3-(1,1-dimethylethoxy)-3-oxopropyl]phenyl]propanoic acid (1.0 g, 2.6 mmol) in 94% yield, mp. 88.5-90°C, $[\alpha]_D +21.2^\circ$ (0.90% in ethanol).

EXAMPLE 21

Preparation of benzyl 2-(4-bromophenyl)acetate

A mixture of 2-(4-bromophenyl)acetic acid (50 g, 0.23 mol), benzyl alcohol (26 ml, 0.25 mol) and p-toluenesulfonic acid (0.3 g) in 200 ml toluene were heated at reflux with the azeotropic removal of water (15 h). The yellow solution was washed with water (250 ml), saturated aqueous sodium

bicarbonate (250 ml) and then dried (MgSO_4).
Concentration provided a light yellow solid that was
recrystallized from cold hexanes to give benzyl
2-(4-bromophenyl)acetate (48.5 g, 0.16 mol) in 69% yield,
mp. 47.5-48°C.

EXAMPLE 22

Preparation of benzyl (E)-2-[4-[3-(1,1-dimethylethoxy)-3- 10 oxo-1-propenyl]phenyl]acetate

A solution of benzyl 2-(4-bromophenyl)acetate (33.5 g,
0.11 mol), t-butyl acrylate (31.6 ml, 0.22 mol) and
triethylamine (85 ml, 0.64 mol) in DMF (1000 ml) was
15 deoxygenated with argon, whereupon bis(triphenylphosphine)-
palladium dichloride (5.1 g, 8.3 mmol) was added. After
heating at 75°C for 12 hours the solvent was removed
in vacuo providing a brown residue that was purified by
chromatography on silica gel using a mixture of hexanes and
20 ethyl acetate (9:1) to give benzyl (E)-2-[4-[3-(1,1-dime-
thylethoxy)-3-oxo-1-propenyl]phenyl]acetate as a pale yellow
oil (23.3 g, 0.07 mol) in 60% yield.

EXAMPLE 23

Preparation of 1,1-dimethylethyl 4-(carboxymethyl)- 25 phenylpropanoate

A mixture of benzyl (E)-2-[4-[3-(1,1-dimethylethoxy)-3-
30 oxo-1-propenyl]phenyl]acetate (11.6 g, 33.0 mmol) and 10%
Pd/C (0.5 g) in ethanol (120 ml) was stirred under H_2 for
3 hours at 1 atm. Filtration of the mixture through a pad
of Celite, followed by concentration gave a colorless oil
that was recrystallized from a mixture of ethyl acetate and
35 hexanes to give 1,1-dimethylethyl-4-(carboxymethyl)phenyl
propanoate (7.5 g, 28.4 mmol) in 86% yield, mp. 55.5-56.5°C.

EXAMPLE 24

Preparation of 1,1-dimethylethyl 3-[4-(2-benzyloxy-2-oxo-ethyl)phenyl]propanoate

5

To a stirred suspension of 1,1-dimethylethyl 4-(carboxymethyl)phenylpropanoate (5.3 g, 20.0 mmol) and potassium carbonate (2.8 g, 20.0 mmol) in DMF (30 ml) was added benzyl bromide (3.42 g, 20 mmol). After stirring for 10 24 h, the solvent was removed in vacuo and the residue was extracted with hexanes (100 ml). Filtration and removal of the solvent under reduced pressure gave 1,1-dimethylethyl 3-[4-(2-benzyloxy-2-oxo-ethyl)phenyl]propanoate (6.5 g, 18.4 mmol) as a colorless liquid in 92% yield.

15

EXAMPLE 25

Preparation of 3-[4-(2-benzyloxy-2-oxoethyl)phenyl]propanoic acid

20

1,1-dimethylethyl 3-[4-(2-benzyloxy-2-oxoethyl)phenyl]propanoate (3.0 g, 8.5 mmol) was stirred in formic acid (10 ml) for 8 hours. Removal of the solvent in vacuo provided a colorless oil that was shaken vigorously with a 25 mixture of ethyl acetate and hexanes (1:3, 20 ml) until it solidified. The mixture was cooled in ice water and the product, collected by suction filtration to give 3-[4-(2-benzyloxy-2-oxoethyl)phenyl]propanoic acid, as a white solid (2.2 g, 7.4 mmol) in 87% yield, mp. 62-64.5°C.

30

35

EXAMPLE 26

Preparation of N-Hydroxysuccinyl
3-(4-carboxymethyl)phenylpropanoate

5

3-[4-(2-benzyloxy-2-oxoethyl)phenyl]propanoic acid
(1.10 g; 3.68 mmol) N-hydroxy succinimide (0.51 g; 4.43
mmol); and 3.86 ml of 1M dicyclohexylcarbodiimide in
CH₂Cl₂ were dissolved in CH₂Cl₂ (50 ml). The
10 mixture was stirred at room temperature for 22 hours.
1.0 ml of acetic acid was added and the mixture stirred for
3 hours after which the dicyclohexylurea was removed by
filtration. The solvent was removed in vacuo and the
residue was redissolved in DMF (40 ml.). After standing
15 approx. 30 min. additional dicyclohexylurea was removed by
filtration. The volume was adjusted to 100 ml with
additional DMF and the solution was transferred to a
pressure bottle. After flushing with N₂, 10% Pd/C (1.0 g)
was added and the bottle was placed on a Parr hydrogenation
20 apparatus. After flushing the system 3 times with 20 psi
H₂ the mixture was shaken under 50 psi H₂ for approx. 1
1/2 hour (or until H₂ up take ceased.) The pressure bottle
was then flushed 3x 20 psi N₂ before removing from the
parr apparatus. The catalyst was removed by filtration
25 through celite and MgSO₄. The final volume was adjusted
to 140 ml with DMF. 35 ml (0.92 mmol) of the solution of
N-hydroxysuccinyl 3-(4-carboxymethyl)phenylpropanoate was
used per coupling.

30

EXAMPLE 27

Preparation of rac.-2-amino-3-hydroxyphenyl)propanoic acid
benzyl ester

35

A 500 ml round bottom flask fitted with a Dean-Stark
water separator was charged with a suspension of 10.0 g
(0.055 mol) of rac.-3-Hydroxyphenylalanine and 11.5 g (0.066

mol) of p-toluenesulfonic acid monohydrate in 60 ml of benzyl alcohol and 250 ml of toluene. The resulting mixture was heated to reflux for 4 hours as about 2 ml of water was collected in the trap. The mixture was allowed to cool, was
5 diluted with ether and was extracted repeatedly with 1 N hydrochloric acid. The combined extracts were neutralized with excess solid sodium bicarbonate to precipitate 8.72 g of rac.-2-amino-3-(3-hydroxyphenyl)propanoic acid benzyl ester.

10

EXAMPLE 28

Preparation of rac.-2-acetamido-3-(3-hydroxyphenyl)propanoic acid benzy ester

15

A suspension of 8.72 g (0.032 mol) of rac.-2-amino-3-(3-hydroxyphenyl)propanoic acid benzyl ester in 400 ml of ice cold dichloromethane was treated dropwise with 1.9 ml (0.020 mol) of acetic anhydride. Upon completion of the addition,
20 a solution of 3.70 g of sodium carbonate in 30 ml of water was added simultaneously with an additional 1.9 ml of acetic anhydride. After 1 hour, the layers were separated and the organic layer was washed with water. The combined aqueous layers were extracted with dichloromethane and the combined
25 extracts were washed with brine and dried over magnesium sulfate. Filtration and concentration afforded a residue which was chromatographed on a Waters Prep 500 liquid chromatograph fitted with two silica gel cartridges, eluting with 20% ethyl acetate-hexane to give 8.49 g (84%) of
30 rac.-2-acetamido-3-(3-hydroxyphenyl)propanoic acid benzyl ester as a thick oil.

35

EXAMPLE 29

Preparation of rac.-2-acetamido-3-[3-[(trifluoromethyl-sulfonyl)oxy]phenyl]propanoic acid benzyl ester

5

A solution of 8.30 g (0.026 mol) of rac.-2-acetamido-3-(3-hydroxyphenyl)propanoic acid benzyl ester and 10.0 g (0.028 mol) of N-phenyl-N-trifluoromethyl-1,1,1-trifluoromethyl sulfonyl sulfonamide in 110 ml of freshly distilled dichloromethane was cooled in an ice bath and 4.10 ml (0.029 mol) of triethylamine was added dropwise. The resulting mixture was stirred 1 hour at 0°C and was allowed to warm to room temperature over 2 hours. The mixture was diluted with 250 ml of ethyl acetate and was washed with successive 50 ml portions of water, 1 N sodium hydroxide, 1 N hydrochloric acid, water, and brine and was dried over magnesium sulfate. Filtration and concentration afforded an oil which was purified by preparative chromatography on a Waters Prep 500 liquid chromatograph fitted with two Prep-Pak silica gel cartridges, eluting with 40% ethyl acetate-hexane to give 10.02 g (85%) of rac.-2-acetamido-3-[3-[(trifluoromethylsulfonyl)oxy]phenyl]propanoic acid benzyl ester.

25

EXAMPLE 30

Preparation of rac.-2-acetamido-3-[3-(2-propenyl)phenyl]propanoic acid benzyl ester

30

Argon was passed through a solution of 9.89 g (0.022 mol) of rac.-2-acetamido-3-[3-(3-trifluoromethylsulfonyl)oxy]phenyl]propanoic acid benzyl ester, 2.8 g (0.066 mol) of lithium chloride, and 7.00 ml (0.0225 mol) of allyltributyltin in 50 ml of dimethylformamide for ten minutes and 0.20 g (0.00028 mol) of bis(triphenylphosphine)-palladium dichloride was added. The bath temperature was raised to 95-100°C for 2 hours at which time a black

35

precipitate was observed to form. The mixture was cooled, diluted with 250 ml of ether, was washed with 3 x 50 ml of water and 1 x 50 ml of brine and was dried over magnesium sulfate. Filtration and concentration afforded an oil which
5 was purified by preparative chromatography on a Waters Prep 500 liquid chromatograph fitted with two silica gel cartridges and eluting with 33% ethyl acetate-hexane to give 6.70 g (85%) of rac.-2-acetamido-3-[3-(2-propenyl)phenyl] propanoic acid benzyl ester as a white solid, mp 55-56.5°C.
10 A portion was recrystallized from ether-hexane resulting in a mp 57-58°C.

EXAMPLE 31

15 Preparation of rac.-2-acetamido-3-[3-(carboxymethyl)phenyl]propanoic acid benzyl ester

Solutions of 6.25 g (0.0185 mol) of rac.-2-acetamido-3-[3-(2-propenyl)phenyl]propanoic acid benzyl ester in 120 ml each of acetonitrile and carbon tetrachloride and 11.9 g
20 (0.0555 mol) of sodium metaperiodate in 240 ml of water were combined and stirred mechanically as 0.25 g (0.0013 mol) of ruthenium chloride hydrate was added to the mixture. The mixture darkened immediately and after 1 hour, was diluted
25 with 300 ml of dichloromethane. The layers were separated and the organic layer was washed with 100 ml of water. The combined aqueous layers were extracted with 200 ml of ether and the combined extracts were dried over magnesium sulfate, filtered and concentrated.

30 The residue was dissolved in 180 ml of tert-butyl alcohol and 60 ml of 2-methyl-2-butene and a solution of 19.0 g (0.21 mol) of sodium chlorite and 19.0 g (0.137 mol) of monobasic sodium phosphate in 130 ml of water was added
35 all at once. The mixture was stirred at 0°C for 2 hours, was diluted with ether and the layers were separated. The organic layer was washed with 10% sodium thiosulfate and

brine and was dried over magnesium sulfate. Filtration and evaporation gave a oily residue which was chromatographed over 200 g of silica gel eluting with 60:40:1 ethyl acetate-hexane-acetic acid to give 4.64 g of a white solid, mp 129-139°C. Recrystallization from dichloromethane-hexane gave 4.14 g (63%) of rac.-2-acetamido-3-[3-(carboxymethyl)phenyl]propanoic acid benzyl ester, mp 130-131°C.

10 EXAMPLE 32

Preparation of rac.-2-acetamido-3-[3-[2-(1,1-dimethylethoxy)-2-oxoethyl]benzene]propanoic acid benzyl ester

15 A suspension of 4.00 g (0.0113 mol) of rac.-2-acetamido-3-[3-(carboxymethyl)benzene]propanoic acid benzyl ester in 40 ml of toluene and 15 ml of dimethylformamide di-tert-butyl acetal was heated to a bath temperature of 55°C. After 30 min, a clear solution formed and after 3 hours, the mixture was cooled, diluted with 50 ml of ether and washed with 3 x 25 ml of water, 1 x 25 ml of brine and was dried over magnesium sulfate. Filtration and concentration gave a white solid which was recrystallized from ether-hexane to afford 3.90 g (84%) of rac.-2-acetamido-3-[3-[2-(1,1-dimethylethoxy)-2-oxoethyl]benzene]propanoic acid benzyl ester, mp 80-83°C.

EXAMPLE 33

30 Preparation of rac.-2-acetamido-3-[3-[2-(1,1-dimethylethoxy)-2-oxoethyl]benzene]propanoic acid

A suspension of 3.30 g (0.00802 mol) of rac.-2-acetamido-3-[3-[2-(1,1-dimethylethoxy)-2-oxoethyl]propanoic acid benzyl ester in 50 ml of ethanol was hydrogenated over 200 mg of 10% palladium on carbon for 3 hours at room temperature under 1 atmosphere of hydrogen. Hydrogen uptake

amounted to 220 ml at which time the mixture was filtered and concentrated to afford 2.54 g of a white solid, mp 167-170°C. Recrystallization from ethanol-hexane afforded 1.77 g (69%) of rac.-2-acetamido-3-[3-[2-(1,1-dimethylethoxy)-2-oxoethyl]benzene propanoic acid, mp 169-172°C.

EXAMPLE 34

10 Preparation of 1,1-dimethylethyl 4-[[2-(1,1-dimethylethyl)-2H-tetrazol-5-yl]methyl]benzenepropenoate

A mixture of Pd(OAc)₂ (90 mg, 0.42 mmol) and 1,1'-bis(diphenylphosphino)ferrocene (300 mg, 0.56 mmol) in 15 DMF (40 ml) was heated at 90°C for 15 minutes while argon was being passed through the mixture. After cooling the burgundy solution to ambient temperature, 5-[(4-bromophenyl)methyl]-2-(1,1-dimethylethyl)-2H-tetrazole (4.0 g, 14 mmol), t-butyl acrylate (3.1 g, 23 mmol) and triethylamine (2.8 ml, 20 mmol) were added to the mixture. The reaction was heated at 90°C overnight and then concentrated. After filtering the dark mixture through a short column of silica gel using a mixture of hexanes and ethyl acetate (4:1), the fractions containing product were pooled and concentrated. Final 25 purification was achieved by chromatography on a Waters Prep 500 liquid chromatograph fitted with two silica gel cartridges eluting with a mixture of hexanes and ethyl acetate (9:1) to give pure 1,1-dimethylethyl 4-[[2-(1,1-dimethylethyl)-2H-tetrazol-5-yl]methyl]benzene 30 propenoate (3.9 g, 11.3 mmol) in 81% yield. The analytical sample was recrystallized from ethyl acetate and hexanes, mp. 105-106°C.

EXAMPLE 35

Preparation of 1,1-dimethylethyl 4-[[2-(1,1-dimethylethyl)-2H-tetrazol-5-yl]methyl]benzene propanoate

5
1,1-dimethylethyl 4-[[2-(1,1-dimethylethyl)-2H-tetrazol-5-yl]methyl]benzene propanoate (3.4 g, 9.9 mmol) in ethanol (30 ml) containing 10% Pd/C (200 mg) was stirred under a blanket of hydrogen (1 atm). After hydrogen uptake
10 ceased, the mixture was filtered through a pad of Celite followed by additional ethanol (30 ml). The filtrate was concentrated and then purified by chromatography on a Waters Prep 500 liquid chromatograph fitted with two silica gel
15 cartridges eluting with a mixture of hexanes and ethyl acetate (4:1) followed by recrystallization from hexanes to give pure 1,1-dimethylethyl 4-[[2-(1,1-dimethylethyl)-2H-tetrazol-5-yl]methyl]benzenepropanoate (3.0 g, 8.6 mmol) in 87% yield, mp. 54-55°C.

20 EXAMPLE 36

Preparation of 4-[[2-(1,1-dimethylethyl)-2H-tetrazol-5-yl]methyl]benzene propanoic acid

25 1,1-dimethylethyl 4-[[2-(1,1-dimethylethyl)-2H-tetrazol-5-yl]methyl]benzenepropanoate (3.0 g, 8.7 mmol) was dissolved in trifluoroacetic acid (33 ml) and stirred at room temperature under argon for 12 hours. The mixture was concentrated and then dissolved in water (50 ml). After
30 adding enough sat. NaHCO₃ to raise the pH to 8, the solution was washed with diethyl ether (2 x 25 ml). The aqueous layer was acidified to pH 3 with 1N HCl and the product collected by suction filtration. Recrystallization from hexanes gave 4-[[2-(1,1-dimethylethyl)-2H-tetra-
35 zol-5-yl]methyl]benzene-propanoic acid (1.5 g, 5.7 mmol) in 66% yield, mp. 139-141 °C.

EXAMPLE 37

Preparation of 5-(4-bromophenyl)-2H-tetrazole

5 4-Bromobenzonitrile (20 g, 110 mmol), sodium azide
 (7.9 g, 121 mmol) and NH_4Cl (6.5 g, 121 mmol) in DMF
 (340 ml) were heated at 90°C under argon. After 2 days the
 reaction mixture was concentrated and then diluted with
 water (300 ml). After adding enough 1M NaOH to render the
10 mixture basic it was washed with ether (4 x 25 ml). The
 aqueous portion was acidified to pH 3 with 1N HCl, the
 precipitated product was collected by suction filtration and
 then washed with water. The crude product was
 recrystallized from ethanol to give crystalline
15 5-(4-bromophenyl)-2H-tetrazole (19.2 g, 83.6 mmol) in 76%
 yield, mp. 271-273°C.

EXAMPLE 38

20 Preparation of 5-(4-bromophenyl)-2-(1,1-dimethylethyl)-
 2H-tetrazole

 5-(4-Bromophenyl)-2H-tetrazole (19.0 g, 81.9 mmol),
 t-butanol (12.1 g, 164 mmol), trifluoroacetic acid (80 ml)
25 and concentrated sulfuric acid (4.6 g, 41 mmol) were stirred
 at ambient temperature for 24 hours. The mixture was
 concentrated and dissolved in ethyl acetate (200 ml). After
 washing with water (3 x 25 ml), 1M NaOH (3 x 25 ml) and
 brine (25 ml) the organic layer was dried (Na_2SO_4),
30 filtered and concentrated. The oil was purified by
 chromatography on a Waters Prep 500 liquid chromatograph
 fitted with two silica gel cartridges eluting with a mixture
 of hexanes and ethyl acetate (9:1) to provide
 5-(4-bromophenyl)-2-(1,1-dimethylethyl)-2H-tetrazole (17.1
35 g, 58.1 mmol) as a yellow oil in 71% yield.

EXAMPLE 39

Preparation of 1,1-dimethylethyl 4-[2-(1,1-dimethylethyl)-2H-tetrazol-5-yl]benzenepropenoate

5

A solution of 5-(4-bromophenyl)-2-(1,1-dimethylethyl)-2H-tetrazole (3.0 g, 10.2 mmol), t-butyl acrylate (2.9 ml, 20.4 mmol), and triethylamine (7.9 ml, 59 mmol) in DMF (93 ml) was deoxygenated with argon, whereupon
10 bis(triphenylphosphine)palladium dichloride (0.5 g, 0.8 mmol) was added. After heating at 75°C for 12 hours the mixture was concentrated under reduced pressure and the brown residue was filtered through a short column of silica gel eluting with a mixture of hexanes and ethyl acetate
15 (9:1). The eluent was concentrated and then purified by chromatography on a Waters Prep 500 liquid chromatograph fitted with two silica gel cartridges eluting with a mixture of hexanes and ethyl acetate (9:1). The fractions containing product were pooled, concentrated and the residue
20 was recrystallized from hexanes to give 1,1-dimethylethyl 4-[2-(1,1-dimethylethyl)-2H-tetrazol-5-yl]benzene propenoate (1.6 g, 5.6 mmol) in 55% yield, mp. 118.5-120°C.

EXAMPLE 40

25

Preparation of 5-[4-(3-(1,1-dimethylethoxy)-3-oxopropyl)phenyl]-2-(1,1-dimethylethyl)-2H-tetrazole

A suspension of 1,1-dimethylethyl
30 4-[2-(1,1-dimethylethyl)-2H-tetrazol-5-yl]benzene propenoate (1.6 g, 5.6 mmol) and 100 mg of 10% Pd/C in ethanol (17 ml) was stirred under a blanket of hydrogen (1 atm). After hydrogen uptake ceased, the mixture was filtered through a pad of Celite washing with ethanol (30 ml). The filtrate
35 was concentrated to give 5-[4-(3-(1,1-dimethylethoxy)-3-oxopropyl)phenyl]-2-(1,1-dimethylethyl)-2H-tetrazole (1.2 g, 4.4 mmol) in 78% yield.

EXAMPLE 41

Preparation of 4-[2-(1,1-dimethylethyl)-2H-tetrazol-5-yl]-
benzenepropanoic acid

5 5-[4-(3-(1,1-dimethylethoxy)-3-oxopropyl)phenyl]-2-
 (1,1-dimethylethyl)-2H-tetrazole (1.0 g, 3.7 mmol) was
 dissolved in trifluoroacetic acid (14 ml) and stirred at
 room temperature under argon for 12 hours. The mixture was
10 concentrated and then dissolved in water (35 ml). After
 adding enough sat. NaHCO_3 to raise the pH to 8, the
 solution was washed with diethyl ether (2 x 10 ml). The
 aqueous layer was acidified to pH 3 with 1N HCl and the
 product was collected by suction filtration to give
15 4-[2-(1,1-dimethylethyl)-2H-tetrazole-5-yl]benzene-
 propanoic acid (0.7 g, 3.1 mmol) in 84% yield, mp. 88-90°C

EXAMPLE 42

20 Preparation of Ac-(D,L)Phe(3-COOH)-Met-Gly-Trp-Met-Asp-
 Phe-NH₂

 6 g of Boc-Phe-PAM resin (substitution 0.36 mmol/g) was
 suspended and shaken in TFA/ CH_2Cl_2 (1:1 by volume, 3 x
25 50 ml) 10 min. each time at room temperature to remove the
 Boc-group. The product was isolated by filtration and
 washed (3 x 50 ml each) with CH_2Cl_2 , 8% DIEA in
 CH_2Cl_2 and CH_2Cl_2 to give the free base of
 Phe-PAM-resin. This was subjected to sequential solid phase
30 synthesis using the Fmoc-protocol. All couplings were
 performed using the DCC/HOBt procedure. At step 7 the
 Fmoc-amino acids, DCC, and HOBt were added with the
 corresponding reaction times as follows: Fmoc-Asp(OtBu)-OH
 (2.46 g, 6 mmol), DCC (1.24 g, 6 mmol) and HOBt (1.2 g,
35 9 mmol) were dissolved in 50 ml of 1:1 by volume
 DMF/ CH_2Cl_2 , and allowed to couple for 60 min at room
 temperature. Fmoc-Met-OH (2.2 g, 6 mmol), DCC (1.24 g,

6 mmol) and HOBt (1.2 g, 9 mmol) were dissolved in 50 ml of 1:1 by volume DMF/CH₂Cl₂, and allowed to couple for 60 min at room temperature. Fmoc-Trp-OH (2.6 g, 6 mmol), DCC (1.24 g, 6 mmol) and HOBt (1.2 g, 9 mmol) were dissolved in 50 ml of 1:1 by volume DMF/CH₂Cl₂, and allowed to couple for 60 min at room temperature. Fmoc-Gly-OH (1.8 g, 6 mmol), DCC (1.24 g, 6 mmol) and HOBt (1.2 g, 9 mmol) were dissolved in 50 ml of 1:1 by volume DMF/CH₂Cl₂, and allowed to couple for 60 min. Fmoc-Met-OH (2.2 g, 6 mmol), DCC (1.24 g, 6 mmol) and HOBt (1.2 g, 9 mmol) were dissolved in 50 ml of 1:1 by volume DMF/CH₂Cl₂, and allowed to couple for 60 min at room temperature. At this point the peptide-resin was dried under high vacuum to provide 7.98 g of Fmoc-Met-Gly-Trp-Met-Asp(OtBu)-Phe-PAM resin.

15 A portion (1.5 g, 0.4 mmol) of Fmoc-Met-Gly-Trp-Met-Asp-(OtBu)-Phe-PAM resin was deprotected with 20% piperidine/DMF (step 1-6) using the Fmoc protocol and coupled to the compound of Example 17(a), rac.-2-acetamido-3-[3-[1,1-dimethyl ethoxy)carbonyl]phenyl]propanoic acid [Ac-(D,L)Phe (3-CO₂OtBu)] (400 mg, 1.5 mmol), using DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) which were dissolved in 50 ml of DMF/CH₂Cl₂ (1:1) by volume and allowed to react for 60 min at room temperature, then washed (step 8-16), and dried to yield Ac-(D,L)Phe(3-CO₂OtBu)-Met-Gly-Trp-Met-Asp(OtBu)-Phe-PAM-resin. This peptidyl-resin was suspended and shaken in 50% TFA/CH₂Cl₂ with 1% EDT (2 x 50 ml) 10 min each time at room temperature to remove the OtBu groups. The peptidyl-PAM resin was then isolated by filtration, washed (3 x 50 ml each) with CH₂Cl₂, DMF and methanol and placed in a pressure bottle, suspended in 100 ml methanol, saturated with NH₃ at -20°C and sealed. The suspension was stirred at room temperature for 3 days. After venting the excess NH₃, the PAM-resin was filtered off and washed with methanol. The combined filtrates were evaporated to dryness to give 1.00 g of crude peptide.

100 mg of the crude peptide was purified by preparative HPLC on a (2.3 x 30) cm micro Bondapack C₁₈ column. The peptide was eluted with a linear gradient (4h) of 5% to 65%, 0.022% TFA/CH₃CN at a flow rate of 8 ml/min, detection at 280 nm. The main peak was collected and lyophilized to yield 25 mg (61%) of Ac-(D,L)Phe(3-COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂. This material was homogeneous by HPLC and gave the correct amino acid analysis and MS. Amino acid analysis: Asp, 1.00(1); Gly, 1.00(1), Met, 1.95(2); Phe, 1.01(1); Trp, 0.80(1); (D,L)Phe(3-COOH), n.d. Empirical Formula: C₄₈H₅₉N₉O₁₂S₂. M.W. 1018.18.

EXAMPLE 43

Preparation of Ac-(D,L)Phe(4-COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂

1.76 g (0.47 mmol) of Fmoc-Met-Gly-Trp-Met-Asp(OtBu)-Phe-PAM-resin obtained from Example 42 was deprotected with 20% piperidine/DMF (step 1-6) using the Fmoc protocol and coupled to the compound of Example 17, rac.-2-acetamido-3-[4-[(1,1-dimethylethoxy)carbonyl]phenyl]propanoic acid [Ac-(D,L)Phe(4-CO₂OtBu)] (700 mg, 1.5 mmol) using DCC (310 mg, 1.5 mmol) and HOBT (270 mg, 2 mmol) which were dissolved in 50 ml of DMF/CH₂Cl₂ (1:1) by volume and allowed to react for 60 min at room temperature, then washed (step 8-16) and dried to yield Ac-(D,L)Phe(4-CO₂OtBu)-Met-Gly-Trp-Met-Asp(OtBu)-Phe-PAM-resin. This peptidyl-resin was suspended and shaken in 50% TFA/CH₂Cl₂ with 1% EDT (2 x 50 ml) 10 min each time at room temperature to remove the OtBu groups. The peptidyl-resin was then isolated by filtration, washed (3 x 50 ml each) with CH₂Cl₂, DMF and methanol and placed in a pressure bottle suspended in 100 ml of methanol, saturated with NH₃ at -20°C and sealed. The suspension was stirred at room temperature for 3 days.

After venting the excess NH_3 , the PAM-resin was filtered off and washed with methanol. The combined filtrates were evaporated to dryness to yield 541 mg of crude peptide.

- 5 80 mg of the crude peptide was purified by preparative HPLC on a (2.3 x 30) cm micro Bondapack C_{18} column. The peptide was eluted with a linear gradient (4h) of 5% to 65%, 0.022% TFA/ CH_3CN at a flow rate of 8 ml/min, detection at 280 nm. The main peak was collected and lyophilized to
- 10 yield 23 mg (33 %) of Ac-(D,L)Phe(4-COOH)-Met-Gly-Trp-Met-Asp-Phe- NH_2 . This material was homogeneous by HPLC and gave the correct amino acid analysis and MS. Amino acid analysis: Asp, 1.00(1); Gly, 0.98(1); Met, 2.00(2); Phe, 1.00(1); Trp, 0.76(1); (D,L)Phe(4-COOH), n.d. Empirical
- 15 Formula: $\text{C}_{48}\text{H}_{59}\text{N}_9\text{O}_{12}\text{S}_2$. M.W. 1018.18.

EXAMPLE 44

Preparation of Ac-(D)Phe(4-COOH)-Met-Gly-Trp-Met-Asp- 20 Phe- NH_2

- 4 mg of Ac-(D,L)Phe-(4-COOH)-Met-Gly-Trp-Met-Asp-Phe- NH_2 were dissolved in 0.5 ml of 0.2N NH_4OH and applied to (1.25 x 30) cm micro Bondapack C_{18} column from E.S.
- 25 Industries. The column was previously equilibrated with 10% $\text{CH}_3\text{CN}/0.01\text{M NH}_4\text{OAc}$ and eluted with a linear gradient of 10-40% CH_3CN in 0.01M NH_4OAc for 120 min at 5 ml/min. The 280 nm absorption of the column effluent was monitored. Two peaks were detected eluting at 78 min at 84 min.
- 30 Fractions containing these peaks were pooled and lyophilized to 1.5 mg of white powder. Analysis by Glass Capillary Gas Chromatography shows that the compound at 78 min retention time contains the (D)Phe(4-COOH) enantiomer. The purity of this compound was also determined by analytical HPLC, amino
- 35 acid analysis, UV and IR.

EXAMPLE 45

Preparation of Ac-Phe(4-COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂

5

From Example 44, the compound eluting at 84 min retention time was analyzed by Glass Capillary Gas Chromatography and showed to contain the (L)Phe(4-COOH) enantiomer. The purity of this compound was also determined by analytical HPLC, amino acid analysis, UV and IR.

10

EXAMPLE 46

Preparation of Ac-(D,L)Phe(4-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂

15

1.00 g (0.27 mmol) of Fmoc-Met-Gly-Trp-Met-Asp(OtBu)-Phe-PAM-resin obtained from Example 42 was deprotected with 20% piperidine/DMF (step 1-6) using the Fmoc protocol and coupled to the compound of Example 17(b,) rac.-2-acetamido-3-[4-[1,1-dimethylethoxy)-2-oxoethyl]carbonyl]phenyl] p opanoic acid [Ac-(D,L)Phe(4-CH₂COOtBu)] (280 mg, 0.8 mmol) using the DCC (165 mg, 0.8 mmol) and HOBt (200 mg, 1.5 mmol) which were dissolved in 40 ml DMF/CH₂Cl₂ (1:1) by volume and allowed to react for 60 min at room temperature, then washed (step 8-16) and dried to yield Ac-(D,L)Phe(4-CH₂COOtBu)-Met-Gly-Trp-Met-Asp(OtBu)-Phe-PAM-resin. This peptidyl-resin was suspended and shaken in 50% TFA/CH₂Cl₂ with 1% EDT (2 x 50 ml) 10 min each time at room temperature to remove the OtBu groups. The peptidyl-resin was then isolated by filtration, washed (3 x 50 ml each) with CH₂Cl₂, DMF and methanol, and placed in a pressure bottle, suspended in 100 ml of methanol, saturated with NH₃ at -20°C and sealed. The suspension was stirred at room temperature for 3 days. After venting the excess NH₃, the PAM-resin was filtered off and washed with methanol. The combined filtrates were

30

35

evaporated to dryness to yield 392 mg of crude peptide.
80 mg of crude peptide was purified by preparative HPLC on a
(2.3 x 30) cm micro Bondapack C₁₈ column. The peptide was
eluted with a linear gradient (4h) of 5% to 65% 0.022%
5 TFA/CH₃CN at a flow rate of 8 ml/min, detection at
280 nm. The main peak was collected and lyophilized to
yield 13 mg (24 %) of Ac-(D,L)Phe(4-CH₂COOH)-Met-Gly-Trp-
Met-Asp-Phe-NH₂. This material was homogeneous by HPLC and
gave the correct amino acid analysis and MS. Amino acid
10 analysis: Asp, 1.02(1); Gly, 1.00(1); Met, 1.96(2); Phe,
1.00(1); Trp, 0.75(1); Phe(4-CH₂COOH), 1.00(1). Empirical
Formula: C₄₉H₆₁N₉O₁₂S₂. M.W. 1032.20.

EXAMPLE 47

15 Preparation of Ac-(D)Phe(4-CH₂COOH)-Met-Gly-Trp-
Met-Asp-Phe-NH₂

2 mg of Ac-(D,L)Phe(4-CH₂COOH)-Met-Gly-Trp-Met-Asp-
20 Phe-NH₂ were dissolved in 0.5 ml of 0.2N NH₄OH and
applied to (2.3 x 30) cm micro Bondapack C₁₈ column from
E.S. Industries. The column was previously equilibrated
with 2% CH₃CN/0.01M NH₄OAc and eluted with a linear
gradient of 2-20% CH₃CN in 0.01M NH₄OAc over 5 min at
25 8 ml/min, then held at 20% CH₃CN for 120 min. The 280 nm
absorption of the column effluent was monitored. Two peaks
were detected eluting at 40 min and 46 min. Fractions
containing these peaks were pooled and lyophilized to 0.8 mg
of white powder. Analysis by Glass Capillary Gas
30 Chromatography shows that the compound eluting at 40 min
retention time contains the (D)Phe(4-CH₂COOH) enantiomer.
The purity of this compound was also confirmed by analytical
HPLC, amino acid analysis, UV and IR.

EXAMPLE 48

Preparation of Ac-Phe(4-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂

5

From Example 47, the compound eluting at 46 min retention time was analyzed by Glass Capillary Gas Chromatography and showed to contain the (L)Phe(4-CH₂COOH) enantiomer. The purity of this compound was also determined by analytical HPLC, amino acid analysis, UV and IR.

10

EXAMPLE 49

Preparation of Ac-(D)Phe(4-CH₂COOC₂H₅)-Met-Gly-Trp-Met-Asp(COOC₂H₅)-Phe-NH₂

15

15 mg of Ac-(D,L)Phe(4-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂ (0.015 mmol) were dissolved in 4 ml of 2:1 by volume ethanol/DMF. HOBT (6 mg, 0.04 mmol) was added and the mixture was cooled to 0°C. DCC (9 mg, 0.04 mmol) was then added and the reaction mixture was stirred for 1 hr at 0°C and allowed to stand at 5°C for 24 hours and at room temperature for 15 hours. The reaction mixture was then evaporated in vacuo. The residue was taken off in 1 ml of DMF and applied into a (2.3 x 30) cm micro Bondapack C₁₈ column. The column was previously equilibrated with 5% CH₃CN/0.022% TFA and eluted with a linear gradient of 5-65% CH₃CN in 0.022% TFA over 240 min at 8 ml/min. The 280 nm absorption of the column effluent was monitored. Two peaks were detected eluting at 168 min and 171 min. Fractions containing these peaks were pooled and lyophilized to yield 4 mg (49%) of white powder. Analysis by MS, NMR and Glass Capillary Gas Chromatography shows that the compound eluting at 168 min retention time is:

Ac-(D)Phe(4-CH₂COOC₂H₅)-Met-Gly-Trp-Met-Asp(COOC₂H₅)-Phe-NH₂. Empirical Formula: C₅₃H₆₉N₉O₁₂S₂.
M.W. 1088.31.

EXAMPLE 50

Preparation of Ac-Phe(4-CH₂COOC₂H₅)-Met-Gly-Trp-Met-Asp-(COOC₂H₅)-Phe-NH₂

5

From Example 49, the compound eluting at 171 min retention time was analyzed by MS, NMR and Glass Capillary Gas Chromatography and showed to be

10 Ac-Phe(4-CH₂COOC₂H₅)-Met-Gly-Trp-Met-Asp(COOC₂H₅)-Phe-NH₂.

EXAMPLE 51

15 Preparation of Ac-Phe(4-CH₂CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂

1.76 g (0.47 mmol) of Fmoc-Met-Gly-Trp-Met-Asp(OtBu)-Phe-PAM-resin obtained from Example 42 was deprotected with 20% piperidine/DMF (step 1-6) using the Fmoc protocol and
20 coupled to the compound of Example 20 (S)-2-[[[(1,1-dimethylethoxy)carbonyl]amino]-3-[4-[3-(1,1-dimethylethoxy)3-oxo-1-propenyl]phenyl]propanoic acid
[Boc-Phe(4-CH₂CH₂COOtBu)] (550 mg, 1.5 mmol) using DCC (310 mg, 1.5 mmol) and HOBT (270 mg, 2 mmol) which were
25 dissolved in 50 ml of DMF/CH₂Cl₂ (1:1) by volume and allowed to react for 60 min at room temperature then washed (step 8-16) and dried to yield Boc-Phe(4-CH₂CH₂COOtBu)-Met-Gly-Trp-Met-Asp(OtBu)-Phe-PAM-resin. This
peptidyl-resin was suspended and shaken in 50%
30 TFA/CH₂Cl₂ with 1% EDT (2 x 50 ml) 10 min each time at room temperature to remove the Boc and OtBu groups. The
peptidyl-resin was then isolated by filtration, washed (3 x 50 ml each) with CH₂Cl₂, DMF and methanol, and then
acetylated using 3 equivalents of acetic acid and BOP
35 reagent in the presence of 1.5% DIEA according to the same protocol as used for the Boc-amino acid residues. The
acetylated peptidyl-resin was then placed in a pressure

bottle, suspended in 75 ml of methanol saturated with NH_3 at -20°C and sealed. The suspension was stirred at room temperature for 3 days. After venting the excess NH_3 , the PAM resin was filtered off and washed with methanol and 90% acetic acid. The combined filtrates were evaporated and the residue lyophilized to yield 831 mg of crude peptide. 100 mg crude peptide was purified by preparative HPLC on a (2.3 x 30) cm micro Bondapak C_{18} column. The peptide was eluted with a linear gradient (4h) of 5% to 65% 0.022% TFA/ CH_3CN at a flow rate of 8 ml/min, detection at 280 nm. The main peak was collected and lyophilized to yield 17mg (29%) of Ac-Phe(4- $\text{CH}_2\text{CH}_2\text{COOH}$)-Met-Gly-Trp-Met-Asp-Phe- NH_2 . This material was homogeneous by HPLC and gave the correct amino acid analysis and MS. Amino acid analysis: Asp, 1.00(1); Gly, 1.01(1); Met, 1.95(2); Phe, 1.00(1); Phe($\text{CH}_2\text{CH}_2\text{COOH}$), 0.97(1); Trp, 0.70(1). Empirical Formula: $\text{C}_{50}\text{H}_{63}\text{N}_9\text{O}_{12}\text{S}_2$. M.W. 1046.22.

EXAMPLE 52

Preparation of Ac-(D,L)Phe(4- CF_2COOH)-Met-Gly-Trp-Met-Asp-Phe- NH_2

1.00 g (0.27 mmol) of Fmoc-Met-Gly-Trp-Met-Asp(OtBu)-Phe-PAM-resin obtained from Example 42 was deprotected with 20% piperidine/DMF (step 1-6) using the Fmoc protocol and coupled to the compound of Example 17(d), rac.-2-acetamido-3-[4-[1,1-difluoro[2-(1,1-dimethylethoxy)-2-oxoethyl]phenyl]propanoic acid [Ac-(D,L)Phe(4- CF_2COOtBu)] (310 mg, 0.8 mmol) using the DCC (165 mg, 0.8 mmol) and HOBt (200 mg, 1.5 mmol) which were dissolved in 40 ml DMF/ CH_2Cl_2 (1:1) by volume and allowed to react for 60 min at room temperature then washed (step 8-16) and dried to yield Ac-(D,L)Phe(4- CF_2COOtBu)-Met-Gly-Trp-Met-Asp(OtBu)-Phe-PAM-resin. This peptidyl-resin was suspended and shaken in 50% TFA/ CH_2Cl_2 with 1% EDT (2 x 50 ml) 10 min each time at room temperature to remove the OtBu groups. The

peptidyl-resin was then isolated by filtration, washed (3 x 50 ml each) with CH_2Cl_2 , DMF and methanol and placed in a pressure bottle suspended in 100 ml of methanol, saturated with NH_3 at -20°C and sealed. The suspension
5 was stirred at room temperature for 3 days. After venting the excess NH_3 , the PAM-resin was filtered off and washed with methanol. The combined filtrates were evaporated to dryness to yield 388 mg of crude peptide. 70 mg of the crude peptide was purified by preparative HPLC on a (2.3 x 30) cm
10 micro Bondapack C_{18} column. The peptide was eluted with a linear gradient (4h) of 5 to 65% 0.022% TFA/ CH_3CN at a flow rate of 8 ml/min, detection at 280 nm. The main peak was collected and lyophilized to yield 14 mg (27 %) of Ac-(D,L)Phe(CF_2COOH)-Met-Gly-Trp-Met-Asp-Phe- NH_2 . This
15 material was homogeneous by HPLC and gave the correct amino acid analysis and MS. Amino acid analysis: Asp, 1.00(1); Gly, 1.00(1); Met, 2.04(2); Phe, 1.00(1); Trp, 0.70(1); Phe ($4\text{CF}_2\text{COOH}$) n.d. Empirical Formula:
 $\text{C}_{49}\text{H}_{59}\text{N}_9\text{O}_{12}\text{F}_2\text{S}_2$. M.W. 1068.20.

20

EXAMPLE 53

Preparation of Ac-(D,L)Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe- NH_2

25

Boc-Phe (2.6 g, 10 mmol) and HOBt (2.0 g, 15 mmol) were dissolved in a mixture of 20 ml CH_2Cl_2 and 20 ml DMF chilled to 0°C and with stirring (2.06 g, 10 mmol) DCC was added and the mixture was stirred for 60 minutes at 0°C .
30 Separately 10 g of benzhydrylamine copolysterene 1% divinylbenzene cross-linked resin (0.56 mmole N/g) was washed with 10% diisopropylethylamine in methylene chloride for 30 min, filtered and washed with methylene chloride dimethylformamide and methylene chloride. The chilled
35 mixture above was added to the resin and stirred for 24 hours at room temperature. The resin was filtered and washed with methylene chloride, dimethylformamide,

isopropanol, methylene chloride, dimethylformamide, isopropanol, methylene chloride and dried under high vacuum. Amino acid analysis showed the resin to contain 0.32 mmoles of phenylalanine per gram of resin. Unreacted amino groups were capped by shaking the resin with 5 ml of acetic anhydride and 5 ml diisopropylethylamine in methylene chloride for 60 minutes. The resin was filtered and washed with methylene chloride, isopropanol, dimethylformamide and methylene chloride. 1.5 g (0.48 mmol) Boc-Phe-BHA resin was subjected to sequential solid phase synthesis using the Boc protocol. All couplings were performed using the DCC/HOBt procedure. At step 16 the Boc-amino acid, DCC and HOBt were added with the corresponding reaction times as follows:

Boc-Asp(OBzl)-OH (485 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by volume DMF/CH₂Cl₂, and allowed to couple for 60 min at room temperature. Boc-Met-OH (380 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by volume DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. Boc-Trp(For)-OH (500 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of DMF, and allowed to couple for 60 min at room temperature. Boc-Gly-OH (270 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by volume DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. Boc-Met-OH (380 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by volume DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. The compound of Example 17(c), rac.-2-acetamido-3-[4-[5-(2-(1,1-dimethylethyl)-2H-tetrazoyl]]phenyl]propanoic acid [Ac-(D,L)Phe(4-tetrazole-tBu)] (500 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by volume DMF/CH₂Cl₂, and allowed to couple for 60 min at room temperature. At this point the peptidyl-resin was dried

under high vacuum to provide 2.25 g of Ac-(D,L)Phe(4-tetrazole-tBu)-Met-Gly-Trp(For)-Met-Asp(OBzl)-Phe-BHA-resin.

2.25 g of the resin was cleaved by treatment with 5 ml of HF containing 2.0 ml of anisole, 1.0 ml of EDT and 15 ml of dimethylsulfide for 1 hour at 0°C. After evaporation to a low volume, fresh anhydrous HF (20 ml) was distilled into the reaction vessel for a second treatment for 2h at 0°C. After thorough evaporation, the resin was washed with 2 volumes of ethylacetate then triturated with 4 x 15 ml of 30% acetic acid, filtered and lyophilized to yield 415 mg of crude peptide.

100 mg of the crude peptide was purified by preparative HPLC on a (2.3 x 30) cm micro Bondapack C₁₈ column. The peptide was eluted with a linear gradient (4h) of 5 to 65% 0.022% TFA/CH₃CN of a flow rate of 8 ml/min, detection at 280 nm. The main peak was collected and lyophilized to yield 8 mg (6.6%) of Ac-(D,L)Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂. This material was homogeneous by HPLC and gave the correct amino acid analysis and MS. Amino acid analysis: Asp, 0.98(1); Gly, 1.00(1); Met, 1.92(2); Phe, 1.00(1); (D,L)Phe(4-tetrazole), 0.96(1); Trp, n.d. Empirical Formula: C₄₈H₅₉N₁₃O₁₀S₂. M.W. 1042.19.

EXAMPLE 54

Preparation of Ac-(D)Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂

2 mg of Ac-(D,L)Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂ from example 53 were dissolved in 0.5 ml of 0.2N NH₄OH and applied to (2.3 x 30) cm micro Bondapack C₁₈ column from E.S. Industries. The column was previously equilibrated with 1% CH₃CN/0.01M NH₄OAc and eluted with a step gradient of 1-20% CH₃CN/0.01M NH₄OAc over 5 min at 8 ml/min, then 20-30% CH₃CN/0.01M NH₄OAc over 120 min

at 8 ml/min. The 280 nm absorption of the column effluent was monitored. Two peaks were detected eluting at 35 min and 39 min. Fractions containing these peaks were pooled and lyophilized to 1 mg of white powder. Analysis by Glass
5 Capillary Gas Chromatography shows that the compound eluting at 35 min retention time contains the (D)Phe(4-tetrazole) enantiomer. The purity of this compound was also confirmed by analytical HPLC, amino acid analysis, UV and IR.

10 EXAMPLE 55

Preparation of Ac-Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂

15 From Example 54, the compound eluting at 39 min retention time was analyzed by Glass Capillary Gas Chromatography and shown to contain the (L)Phe(4-tetrazole) enantiomer. The purity of this compound was also determined by analytical HPLC, amino acid analysis, UV and IR.

20 EXAMPLE 56

Preparation of Ac-(D,L)Phe(4-CH₂COOH)-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-NH₂

25 Boc-N-methyl-Phe (5 g, 17.8 mmol) and HOBt (34 g, 25 mmol) were dissolved in a mixture of 20 ml CH₂Cl₂ and 20 ml DMF chilled to 0°C and with stirring (4.12 g, 20 mmol) DCC was added and the mixture was stirred for 60 minutes at
30 0°C. Separately 10 g of benzhydrylamine copolysterene 1% divinylbenzene cross-linked resin (0.39 mmol N/g) was washed with 10% diisopropylethylamine in methylene chloride for 30 min, filtered and washed with methylene chloride dimethylformamide and methylene chloride. The chilled
35 mixture above was added to the resin and stirred for 24 hours at room temperature. The resin was filtered and washed with methylene chloride, dimethylformamide,

isopropanol, methylene chloride, and dried under high vacuum. Amino acid analysis showed the resin to contain 0.30 mmoles of N-methylphenylalanine per gram of resin. Unreacted amino groups were capped by shaking the resin with 5 ml of acetic anhydride and 5 ml diisopropylethylamine in methylene chloride for 60 min. The resin was filtered and washed with methylene chloride, isopropanol, dimethylformamide and methylene chloride.

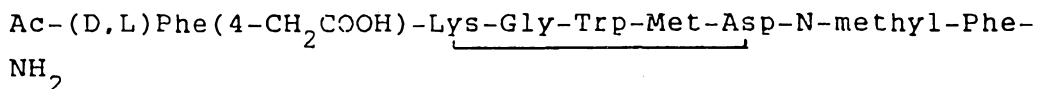
4 g (1.2 mmol) Boc-N-methyl-Phe-BHA resin was suspended and shaken in TFA/CH₂Cl₂ (1:1) by volume (3 x 40 ml) 10 min, each time at room temperature to remove the Boc group. The product was isolated by filtration and washed (3 x 50 ml each) with CH₂Cl₂, 8% DIEA in CH₂Cl₂ and CH₂Cl₂ to give the free base of N-methyl-Phe-BHA resin. This was subjected to sequential solid phase peptide synthesis using the DCC/HOBt procedure. At step 7 the Fmoc amino acid, DCC and HOBt were added with the corresponding reaction times as follows: Fmoc-Asp(OtBu)-OH (1.64 g, 4 mmol), DCC (825 mg, 4 mmol) and HOBt (810 mg, 6 mmol) were dissolved in 40 ml of 1:1 by volume DMF/CH₂Cl₂, and allowed to couple for 60 min at room temperature. Fmoc-Met-OH (1.5 g, 4 mmol), DCC (825 mg, 1 mmol) and HOBt (810 mg, 6 mmol) were dissolved in 40 ml of 1:1 by volume DMF/CH₂Cl₂, and allowed to couple for 60 min. at room temperature. Fmoc-Trp-OH (1.7 g, 4 mmol), DCC (825 mg, 4 mmol) and HOBt (810 mg, 6 mmol) were dissolved in 40 ml of 1:1 by volume DMF/CH₂Cl₂, and allowed to couple for 60 min at room temperature. Fmoc-Gly-OH (1.2 g, 4 mmol), DCC (825 mg, 4 mmol) and HOBt (810 mg, 6 mmol) were dissolved in 40 ml of 1:1 by volume DMF/CH₂Cl₂, and allowed to couple for 60 min at room temperature. Fmoc-Lys(Boc) (1.9 g, 4 mmol), DCC (825 mg, 4 mmol) and HOBt (810 mg, 6 mmol) were dissolved in 40 ml of 1:1 by volume DMF/CH₂Cl₂, and allowed to couple for 60 min. At this point the peptide-resin was dried under high vacuum to provide 5.2 g of Fmoc-Lys(Boc)-Gly-Trp-Met-Asp(OtBu)-N-methyl-Phe-BHA-resin. This peptidyl-resin was

suspended and shaken in 50% TFA/CH₂Cl₂ with 1% EDT (2 x 50 ml) 10 min each time at room temperature to remove the OtBu and the Boc groups. The peptidyl-BHA resin was then neutralized with 8% DIPEA in CH₂Cl₂, washed (3 x 50 ml each) with CH₂Cl₂, DMF and methanol, and isolated by filtration to yield Fmoc-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-BHA resin. The cyclization on the resin between the ε-amino group of Lys and the β-carboxyl group of Asp was achieved using the BOP reagent (2.0 g, 4 mmol) in DMF (50 ml) containing 1.5% DIEA (v/v) for 48h. A negative Kaiser ninhydrin test was observed and the peptide-resin was washed and dried to yield:

Fmoc-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-BHA-resin.

1.1 g (0.33 mmol) of Fmoc-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-BHA-resin was deprotected with 20% piperidine/DMF (step 1-6) using the Fmoc protocol and coupled to the compound of Example 17(b), rac.-2-acetamido-3-[4-[(1,1-dimethylethoxy)-2-oxoethyl]phenyl]propanoic acid [Ac-(D,L)Phe(4-CH₂COOtBu)] (350 mg, 1 mmol) using DCC (210 mg, 1 mmol) and HOBT (202 mg, 15 mmol) which were dissolved in 40 ml DMF/CH₂Cl₂ (1:1) by volume and allowed to react for 60 min at room temperature, then washed (step 8-16) and dried to yield 1.2 g Ac-(D,L)Phe(4-CH₂COOtBu)-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-BHA-resin. This peptidyl-resin was suspended and shaken in 50% TFA/CH₂Cl₂ with 1% EDT (2 x 20 ml) 10 min each time at room temperature to remove the OtBu group, washed (3 x 50 ml each) with CH₂Cl₂ and dried in vacuo. The peptide was cleaved from the resin by treatment with 15 ml of HF containing 1.0 ml of anisole, 0.5 ml of DTE for 2 hours at 0°C. After thorough evaporation, the resin was washed with 2 volumes of ethylacetate, then triturated with 4 x 15 ml of 30% acetic acid, filtered and lyophilized to yield 200 mg of crude peptide.

100 mg of the crude peptide was purified by preparative HPLC on a (2.3 x 30) cm micro Bondapack C₁₈ column. The peptide was eluted with a linear gradient (4h) of 5 to 65% 0.022% TFA/CH₃CN of a flow rate of 8 ml/min, detection at 280 nm. The main peak was collected and lyophilized to yield 12 mg (7%) of:



10

This material was homogeneous by HPLC and gave the correct amino acid analysis and MS. Amino acid analysis: Asp, 1.08(1); Gly, 1.00(1); Met, 1.00(1); Lys, 0.95(1); Phe(4-CH₂COOH), 0.92(1); N-methyl-Phe (n.d.); Trp, 0.70(1). Empirical Formula: C₅₁H₆₄N₁₀O₁₁S₁. M.W. 1025.18.

EXAMPLE 57

20 Preparation of Ac-(D,L)Phe(4-tetrazole)-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-NH₂

1.1 g (0.33 mmol) of Fmoc-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-BHA-resin obtained from Example 56 was deprotected with 20% piperidine/DMF (step 1-6) using the Fmoc protocol and coupled to the compound of Example 17(c), rac.-2-acetamido-3-[4-[5-2-(1,1-dimethylethyl)-2H-tetrazoyl]]-phenyl]propanoic acid [Ac-(D,L)Phe(4-tetrazole-tBu)] (500 mg, 1.5 mmol), using DCC (310 mg, 1.5 mmol), and HOBT (270 mg, 2 mmol) which were dissolved in 40 ml DMF/CH₂Cl₂ (1:1) by volume and allowed to react for 60 min at room temperature, then washed (step 8-16) and dried to yield 1.2 g of:

35 Ac-(D,L)Phe(4-tetrazole-tBu)-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-BHA resin which was cleaved by treatment with HF (15 ml) containing 1.0 ml anisole, 0.5 ml DTE, and 0.5 ml

dimethylsulfide for 2h at 0°C. After thorough evaporation, the resin was washed with 2 volumes of ethylacetate, then triturated with 4 x 15 ml of 30% acetic acid, filtered and lyophilized to yield 123 mg of crude peptide.

5

123 mg of the crude peptide was purified by preparative HPLC on a (2.3 x 30) cm micro Bondapack C₁₈ column. The peptide was eluted with a linear gradient (4h) of 5 to 65% 0.022% TFA/CH₃CN at a flow rate of 8 ml/min, detection at 280 nm. The main peak was collected and lyophilized to yield 20 mg (6%) of:

Ac-(D,L)Phe(4-tetrazole)-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-NH₂.

15

This material was homogeneous by HPLC and gave the correct amino acid analysis and MS. Amino acid analysis: Asp, 1.00(1); Gly, 1.00(1); Met, 1.03(1); Lys, 1.01(1); Trp, 0.6(1); Phe(4-tetrazole); 0.97(1); N-methyl-Phe, n.d. Empirical Formula: C₅₀H₆₂N₁₄O₉S. M.W. 1035.90.

20

EXAMPLE 58

Preparation of Ac-(D,L)Phe-(3-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂

25

1.00 g (0.87 mmol) of Fmoc-Met-Gly-Trp-Met-Asp(OtBu)-Phe-PAM-resin obtained from Example 42 was deprotected with 20% piperidine/DMF (step 1-6) using the Fmoc protocol and coupled to the compound of Example 33, rac.-N-acetyl-3-[2-(1,1-dimethylethoxy)-2-oxoethyl]phenylalanine [Ac-(D,L)Phe(3-CH₂CO₂tBu)] (280 mg 0.8 mmol) using DCC (165 mg. 0.8 mmol) and HOBt (200 mg. 1.5 mmol) which were dissolved in 40 ml DMF/CH₂Cl₂ (1:1) by volume and allowed to react for 60 min at room temperature. Then washed (step 8-16) and dried to yield Ac-(D,L)Phe(3-CH₂COOtBu)-Met-Gly-Trp-Met-Asp(OtBu)-Phe-PAM resin. This peptidyl-resin was suspended and shaken

35

in 50% TFA/CH₂Cl₂ with 1% EDT (2 x 50 ml) 10 min each time at room temperature to remove the OtBu groups. The peptidyl-resin was then isolated by filtration, washed (3 x 50 ml each) with CH₂Cl₂, DMF and methanol, and placed in a pressure bottle suspended in 100 ml of methanol, saturated with NH₃ at 20° and sealed. The suspension was stirred at room temperature for 3 days. After venting the excess NH₃, the PAM-resin was filtered off and washed with methanol. The combined filtrates were evaporated to dryness to yield 456 mg of crude peptide. 80 mg of the crude peptide was purified by preparative HPLC on a (2.3x30) cm micro Bondapack C₁₈ column. The peptide was eluted with a linear gradient (4h) of 5 to 65% 0.022% TFA/CH₃CN of a flow rate at 8 ml/min. detection at 280 nm. The main peak was collected and lyophilized to yield 14 mg (25%) of Ac-(D,L)Phe(3-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe- NH₂. This material was homogeneous by HPLC and gave the correct amino acid analysis and MS. Amino acid analysis: Asp 1.00 (1); Gly 0.80 (1); Met 2.08 (2); Phe 1.00 (1); Trp 0.70 (1); Phe(3-CH₂COOH) 1.00 (1); Empirical Formula C₄₉H₆₁N₉O₁₂S₂, M.W. 1032.20.

EXAMPLE 59

Preparation of Desamino Phe(4-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂

1.00 g (0.87 mmol) of Fmoc-Met-Gly-Trp-Met-Asp-(OtBu)-Phe-PAM resin obtained from example 42 was deprotected with 20% piperidine/DMF (Step 1-6) using the Fmoc protocol and coupled to the compound of Example 26, N-hydroxysuccinyl 3-(4-carboxymethyl)phenylpropionate, (244 mg. 0.8 mmol) which was dissolved in 20 ml of DMF/CH₂Cl₂ (1:1) by volume and allowed to react for 24 hours at room temperature, then washed (step 8-16) and dried to yield Desamino Phe(4-CH₂COOH)-Met-Gly-Trp-Met-Asp (OtBu)-Phe-PAM resin. This peptidyl-resin was suspended and shaken in 50%

TFA/CH₂Cl₂ with 1% EDT (2 x 50 ml) 10 min each time at room temperature to remove the OtBu group. The peptidyl-resin was then isolated by filtration, washed (3 x 50 ml each) with CH₂Cl₂, DMF and methanol, and placed in a pressure bottle, suspended in 100 ml of methanol, saturated with NH₃ at -20° C and sealed. The suspension was stirred at room temperature for 3 days. After venting the excess NH₃, the PAM-resin was filtered off and washed with methanol. The combined filtrates were evaporated to dryness to yield 733 mg of crude peptide. 80 mg of the crude peptide was purified by preparation. HPLC on a (2.3 x 30 cm) micro Bondapack C₁₈ column. The peptide was eluted with a linear gradient (4 h) of 5 to 65% 0.022%TFA/CH₃CN at a flow rate of 8 ml/min. and detection at 280 nm. The main peak was collected and lyophilized to yield 8 mg (38%) of Desamino Phe(4-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂. This material was homogeneous by HPLC and gave the correct amino acid analysis and MS. Amino acid analysis: Asp 1.00 (1); Gly 1.00 (1); Met 2.00(2); Phe 1.00 (1); Trp 0.71 (1); Empirical formula C₄₇H₅₈N₈O₁₁S₂; MW 975.2.

EXAMPLE 60

Preparation of Ac-(D,L)Phe(4-CH₂tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂

1.5 g (0.48 mmol) Boc-Met-Gly-Trp(For)-Met-Asp(OBzl)-Phe-BHA resin obtained from example 53 was deprotected with 50% TFA/CH₂Cl₂ with 1% EDT (step 1-15) using the Boc protocol and coupled to the compound of Example 17(e), rac.-2-amino-3-[4-[[5-[2-(1,1-dimethylethyl)-2H-tetrazoyl]]-methyl] phenyl]propanoic acid, [Ac-(D,L)Phe(4-CH₂-tetrazole-tBu] (525 mg, 1.5 mmol). DCC (310 mg, 1.5 mmol) and HOBt (270mg, 2 mmol) which were dissolved in 50 ml DMF/CH₂Cl₂ (1:1) by volume and allowed to react for 60 min at room temperature. At this point, the peptidyl-resin was dried under high vacuum to

provide 1.7 g of Ac-(D,L)Phe(4-CH₂tetrazole-tBu)-Met-Gly-Trp(For)Met-Asp(OBZl)-Phe-BHA-resin. 1.7 g of the resin was cleaved by treatment with 5 ml of HF containing 2.0 ml of anisole, 1.0 ml of EDT and 15 ml of dimethylsulfide for 1
5 hour at 0°C. After evaporation to a low volume, fresh anhydrous HF (20 ml) was distilled into the reaction vessel for a second treatment for 2 hours at 0 C. After thorough evaporation, the resin was washed with 2 volumes of ethylacetate then triturated with 4 x 15 ml of 30% acetic
10 acid, filtered and lyophilized to yield 456 mg of crude peptide. 80 mg of the crude peptide was purified by preparative HPLC as a (2.3 x 30) cm micro Bondapak C₁₈ column. The peptide was eluted with a linear gradient (4 h) of 5 to 65%. 0.022% TFA/CH₃CN of a flow rate of 8 ml/min.
15 detection of 280 nm. The main peak was collected and lyophilized to yield 18 mg (20%) of Ac-(D,L)Phe(4-CH₂-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂. This material was homogeneous by HPLC and gave the correct amino acid analysis and MS. Amino acid analysis Asp 1.00 (1); Gly 1.06(1); Met
20 1.80(2)Phe 1.00(1); Phe(4-CH₂tetrazole) 0.95 (1); Trp 0.70 (1); Empirical Formula: C₄₉H₆₁N₁₃O₁₀S₂ MW 1056.30.

EXAMPLE 61

25 Preparation of Ac-(D)Phe(4-CH₂-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂

2 mg of Ac-(D,L)-Phe(4-CH₂-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂ obtained from Example 60 were dissolved in
30 0.5 ml of 0.2N NH₄OH and applied to (2.3 x 30 cm) micro Bondapak C-18 column from E.S. Industries. The column was previously equilibrated with 1% CH₃CN/0.01M NH₄OAc and eluted with a step gradient of 1-20% CH₃CN/0.01M NH₄OAc
35 over 5 min at 8 ml/min, then 20-30% CH₃CN/0.01M NH₄OAc over 120 min at 8 ml/min. The 280 nm absorption of the column effluent was monitored. Two peaks were detected eluting at 51 min and 55 min. Fractions containing these

peaks were pooled and lyophilized to 1 mg of white powder. Analysis by Glass Capillary Gas Chromatography shows that the compound eluting at 51 min retention time contains the (D)-Phe(CH₂-tetrazole) enantiomer. The purity of this compound was also confirmed by analytical HPLC, amino acid analysis, UV and IR.

EXAMPLE 62

10 Preparation of Ac-Phe(4-CH₂-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂

From Example 61, the compound eluting at 55 min retention time was analyzed by Glass Capillary Gas Chromatography and shown to contain the (L)-Phe(4-CH₂-tetrazole) enantiomer. The purity of this compound was also determined by analytical HPLC, amino acid analysis, UV and IR.

20 EXAMPLE 63

Preparation of Ac-(D,L)Phe(4-CH₂-tetrazole)-Nle-Gly-Trp-Nle-Asp-N-methyl-Phe-NH₂

25 1 g (0.37 mmol) Boc-N-methyl-Phe-BHA resin was suspended and shaken in TFA/CH₂Cl₂ (1:1) by volume (3 x 40 ml) 10 min each time at room temperature to remove the Boc-group. The product was isolated by filtration and washed (3 x 50 ml each) with CH₂Cl₂, 8% DIEA in CH₂Cl₂ and CH₂Cl₂ to give the free base of N-methyl-Phe-BHA resin. This was subjected to sequential solid phase peptide synthesis using the DCC/HOBt procedure. At Step 16 the Boc-amino acid, DCC and HOBt were added with the corresponding reaction times as follows: Boc-Asp(OBzl)-OH (485 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml 35 1:1 by volume DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. Boc-Nle-OH (350 mg, 1.5 mmol), DCC

(310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by volume DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. Boc-Trp(For)-OH (500 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of DMF, and allowed to couple for 60 min at room temperature. Boc-Gly-OH (270 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by volume DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. Boc-Nle-OH (350 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by volume DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. The compound of Example (17e) rac.-2-acetamido-3-[4-[[5-[2-(1,1-dimethylethyl)-2H-methyl]-tetrazoyl]]phenyl]propanoic acid (530 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 30 ml of 1:1 by volume DMF (CH₂Cl₂) and allowed to couple for 60 min at room temperature. At this point the peptidyl-resin was dried under high vacuum to provide 1.7 g of Ac-(D,L)-Phe(4-CH₂-tetrazole-tBu)-Nle-Gly-Trp(For)-Nle-Asp(OBzl)-N-methyl-Phe-BHA resin. 1.7 g of the resin was cleaved by treatment with 5 ml of HF containing 2.0 ml of anisole, 1.0 ml of EDT and 15 ml of dimethylsulfide for 1 h at 0°C. After evaporation to a low volume, fresh anhydrous HF (20 ml) was distilled into the reaction vessel for a second treatment for 2 h at 0°C. After thorough evaporation, the resin was washed with 2 volumes of ethylacetate, then triturated with 4 x 15 ml of 30% acetic acid, filtered and lyophilized to yield 530 mg of crude peptide.

100 mg of the crude peptide was purified by preparative HPLC on a (2.3 x 30 cm) micro Bondapak C-18 column. The peptide was eluted with a linear gradient (4 h) of 5 to 65% 0.022% TFA/CH₃CN of a flow rate of 8 ml/min, detection at 290 nm. The main peak was collected and lyophilized to yield 10 mg (13.8%) of Ac-(D,L)-Phe(4-CH₂-tetrazole)-Nle-

Gly-Trp-Nle-Asp-N-methyl-Phe-NH₂. This material was homogeneous by HPLC and gave the correct amino acid analysis and MS. Amino acid analysis: Asp, 0.98 (1); Gly, 0.96 (1); Nle, 2.00 (2); N-methyl-Phe, n.d.; Trp, n.d.; Phe(4-CH₂-tetrazole), 0.90 (1): Empirical formula
5 C₅₂H₆₇N₁₃O₁₀. M.W. 1034.12.

EXAMPLE 64

10 Preparation of Ac-(D)Phe(4-CH₂-tetrazole)-
Nle-Gly-Trp-Nle- Asp-N-methyl-Phe-NH₂

2 mg of Ac-(D,L)-Phe(4-CH₂-tetrazole)-Nle-Gly-Trp-Nle-Asp-N-methyl-Phe-NH₂ from example 63 were dissolved in 0.5
15 ml of 0.2N NH₄OH and applied to (2.3 x 30 cm) micro Bondapak C-18 column from E.S. Industries. The column was previously equilibrated with 1% CH₃CN/0.01M NH₄OAc and eluted with a step gradient of 2-25% CH₃CN/0.01M NH₄OAc over 5 min at 8 ml/min, then 25-40% CH₃CN/0.01M NH₄OAc
20 over 120 min at 8 ml/min. The 280 nm absorption of the column effluent was monitored. Two peaks were detected at 41 min and 44 min. Fractions containing these peaks were pooled and lyophilized to 1 mg of white powder. Analysis by Glass Capillary Gas Chromatography shows that the compound
25 eluting at 41 min retention time contains the (D)Phe(4-CH₂-tetrazole) enantiomer. The purity of this compound was also confirmed by analytical HPLC, amino acid analysis, UV and IR.

30 EXAMPLE 65

Preparation of Ac-Phe(4-CH₂-tetrazole)-Nle-Gly-Trp-
Nle-Asp- N-methyl-Phe-NH₂

35 From Example 64, the compound eluting at 44 min retention time was analyzed by Glass Capillary Gas Chromatography and shown to contain the (L)Phe(4-CH₂-

tetrazole) enantiomer. The purity of this compound was also determined by analytical HPLC, amino acid analysis, UV and IR.

5

EXAMPLE 66

Desamino-Phe(4-CH₂-tetrazole)-Nle-Gly-Trp-Nle-Asp-N-methyl-Phe-NH₂

10 1.2 g (0.37 mmol) Boc-Nle-Gly-Trp(For)-Nle-Asp(OBzl)-N-methyl-Phe-BHA-resin obtained from Example 63 was deprotected with TFA/CH₂Cl₂ (1:1) by volume (Steps 1-15) using the Boc-protocol and coupled to the compound of Example 36, 4-[[2-(1,1-dimethylethyl)-2H-tetrazol
15 -5-yl]methly] benzenepropanoic acid (500 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) which were dissolved in 50 ml of DMF/CH₂Cl₂ (1:1) by volume and allowed to react for 60 min at room temperature, then washed (Steps 17-27) and dried to yield 1.4 g Desamino-Phe(4-CH₂-
20 tetrazole-tBu)-Nle-Gly-Trp(For)-Nle-Asp(OBzl)-N-methyl-Phe-BHA-resin. 1.4 g of the resin was cleaved by treatment with HF using the same condition as described in Example 63 to yield 400 mg of crude peptide. 100 mg of the crude peptide was purified by preparative HPLC on a (2.3 x 30 cm) micro
25 Bondapak C-18 column. The peptide was eluted with a linear gradient (4 h) of 5 to 65% 0.022% TFA/CH₃CN of a flow rate of 8 ml/min, detection at 280 nm. The main peak was collected and lyophilized to yield 16 mg (17.7%) of Desamino-Phe(4-CH₂-tetrazole)-Nle-Gly-Trp-Nle-Asp-Phe-
30 NH₂. This material was homogeneous by HPLC and gave the correct amino acid analysis and MS. Amino acid analysis: Asp, 1.01 (1); Gly, 1.00 (1); Nle, 1.90 (2); Trp, 0.80 (1); N-methyl-Phe, n.d. Empirical formula: C₅₀H₆₄N₁₂O₉.
M.W. 977.15.

35

EXAMPLE 67

Desamino-Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂

5 1.4 g (0.48 mmol) of Boc-Met-Gly-Trp(For)-Met-Asp(OBzl)-Phe-BHA-resin obtained from Example 53 was deprotected with TFA/CH₂Cl₂ (1:1) by volume (Steps 1-15) using the Boc-protocol and coupled to the compound of Example 41
10 4-[2-(1,1-dimethylethyl)-2H-tetrazol-5-yl]-benzenepropanoic acid (475 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBT (270 mg, 2 mmol) which were dissolved in 50 ml of DMF/CH₂Cl₂ (1:1) by volume and allowed to react for 60 min at room temperature, then washed (Steps 17-27) and dried to yield 1.52 g of Desamino-Phe(4-tetrazole-tBu)-Met-
15 Gly-Trp(For)-Met-Asp(OBzl)-Phe-BHA-resin. 1.52 g of the resin was cleaved by treatment with HF using the same conditions as described in Example 63 to yield 340 mg of crude peptide. 100 mg of the crude peptide was purified by preparative HPLC on a (2.3 x 30 cm) micro Bondapak C-18
20 column. The peptide was eluted with a linear gradient (4 h) of 5 to 65% 0.022% TFA/CH₃CN of a flow rate of 8 ml/min, detection at 280 nm. The main peak was collected and lyophilized to yield 15 mg (10.8%) of Desamino-Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂. This material was
25 homogeneous by HPLC and gave the correct amino acid analysis and MS. Amino acid analysis: Asp, 1.07 (1); Gly, 0.97 (1); Met, 2.00 (2); Phe, 1.10 (1); Trp, n.d. Empirical formula C₄₆H₅₆N₁₂O₉S₂. M.W. 985.16.

30

EXAMPLE 68

Preparation of Desamino-Phe(4-tetrazole)-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-NH₂

35 1.1 g (0.33 mmol) of Fmoc-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-BHA-resin obtained from Example 56 was deprotected with 20% piperidine/DMF (Steps 1-6) using the

Fmoc protocol and coupled to the compound of Example 41
4-[2-(1,1-dimethylethyl)-2H-tetrazol-5-yl]-benzenepropanoic
acid tetrazole (475 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol)
and HOBt (270 mg, 2 mmol) which were dissolved in 50 ml of
5 DMF/CH₂Cl₂ (1:1) by volume and allowed to react for 60
min at room temperature, then washed (Steps 8-16) and dried
to yield 1.3 g of Desamino-Phe(4-tetrazole-tBu)-Lys-Gly-Trp-
Met-Asp-N-methyl-Phe-BHA-resin. 1.3 g of the resin was
cleaved by treatment with HF using the same conditions as
10 described in Example 68 to yield 439 mg of crude peptide.
150 mg of the crude peptide was purified by preparative HPLC
on a (2.3 x 30 cm) micro Bondapak C-18 column. The peptide
was eluted with a linear gradient (4 h) of 5 to 65% 0.022%
TFA/CH₃CN of a flow rate of 8 ml/min, detection at 280
15 nm. The main peak was collected and lyophilized to yield 15
mg (13.7%) of Desamino-Phe-(4-tetrazole)-Lys-Gly-Trp-
Met-Asp-N-methyl-Phe-NH₂. This material was homogeneous
by HPLC and gave the correct amino acid analysis and MS.
Amino acid analysis: Asp, 0.60 (1); Gly, 0.92 (1); Met, 1.00
20 (1); Lys, 0.60 (1); Trp, 0.90 (1); N-methyl-Phe, n.d.
Empirical formula C₄₈H₅₉N₁₃O₈S. M.W. 978.17.

EXAMPLE 69

25 Preparation of Desamino-Phe(4-CH₂-COOH)-Met-Gly-Trp-Met- Asp-N-methyl-Phe-NH₂

5 g of Fmoc-PAL-resin (substitution 0.38 mmol/g) was
suspended and shaken in 20% piperidine/DMF (Steps 1-6) using
30 the Fmoc protocol and coupled to Fmoc-N-methyl-Phe (2.4 g, 6
mmol) using DCC (1.25 g, 6 mmol) and HOBt (1.2 g, 9 mmol)
which were dissolved in 100 ml DMF/CH₂Cl₂ (2:1) by
volume and allowed to react for 12 h at room temperature,
then washed (Steps 8-16) and dried to yield 5.5 g of Fmoc-N-
35 methyl-Phe-PAL-resin. The substitution was determined by the
Gisin method to be 0.34 mmol/g. A portion, 1 g (0.34 mmol)
of the Fmoc-N-methyl-Phe-PAL-resin was subjected to

sequential solid phase synthesis using the Fmoc-protocol. All couplings except the last residue were performed using DCC/HOBt procedure. At Step 7 the Fmoc-amino acids, DCC and HOBt were added with the corresponding reaction times as follows: Fmoc-Asp(OtBu)-OH (615 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol), were dissolved in 20 ml of 1:1 by volume DMF/CH₂Cl₂, and allowed to couple for 60 min at room temperature. Fmoc-Met-OH (550 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by volume DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. Fmoc-Trp-OH (650 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by volume DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. Fmoc-Gly-OH (450 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by volume DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. Fmoc-Met-OH (550 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by volume DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. At this point the Fmoc-Met-Gly-Trp-Met-Asp(OtBu)-N-methyl-Phe-PAL-resin was suspended and shaken in 20% piperidine/DMF (Steps 1-6) using the Fmoc-protocol and coupled to the compound of Example 26 N-hydroxysuccinyl 3-(4-carboxymethyl)phenylpropanoate (610 mg, 2 mmol) which was dissolved in 20 ml DMF/CH₂Cl₂ (1:1) by volume and allowed to react for 6 h at room temperature, then washed (Steps 8-16) and dried to yield 1.40 g of Desamino-Phe(4-CH₂COOH)-Met-Gly-Trp-Met-Asp(OtBu)-N-methyl-Phe-PAL resin. This peptidyl-resin was suspended and shaken in 50 ml of TFA/EDT/CH₂Cl₂ (14/1/5) by volume for 1 hour at room temperature, then the PAL resin was filtered off and washed with 20 ml of TFA/CH₂Cl₂ (1:1) by volume. The combined filtrates were evaporated to dryness, precipitated with ether, filtered off and dried to yield 268 mg of crude peptide. 135 mg of the crude peptide

was purified by preparative HPLC on a (2.3 x 30 cm) micro Bondapak C-18 column. The peptide was eluted with a linear gradient of 5 to 65% of 0.022% TFA/CH₃CN at a flow rate of 8 ml/min, detection at 280 nm. The main peak was collected
5 and lyophilized to yield 12 mg (7.2%) of Desamino-Phe(4-CH₂COOH)-Met-Gly-Trp-Met-Asp-N-methyl-Phe-NH₂. This material was homogeneous by HPLC and gave the correct amino acid analysis and MS. Amino acid analysis: Asp, 1.00 (1); Gly, 1.05 (1); Met, 1.80 (2); N-methyl-Phe, n.d.; Trp, n.d.
10 Empirical formula: C₄₈H₆₀N₈O₁₁S₂. M.W. 989.15.

EXAMPLE 70

Preparation of Desamino-Phe(4-CH₂COOH)-Nle-Gly-Trp-Nle-Asp-N-methyl-Phe-NH₂ 15

1 g (0.34 mmol) of Fmoc-N-methyl-Phe-PAL-resin obtained from Example 69 was subjected to sequential solid phase synthesis using the Fmoc-protocol. All couplings except the
20 last residue were performed using the DCC/HOBt procedure. At Step 7 the Fmoc-amino acid, DCC and HOBt were added with the corresponding reaction times as follows:
Fmoc-Asp(OtBu)-OH (615 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by
25 volume DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. Fmoc-Nle-OH (520 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in
20 ml of 1:1 by volume DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. Fmoc-Trp-OH (650 mg, 1.5
30 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by volume DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature.
Fmoc-Gly-OH (450 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by
35 volume DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. Fmoc-Nle-OH (520 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 1.5 mmol) were dissolved in

20 ml of 1:1 by volume of DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. At this point the Fmoc-Nle-Gly-Trp-Nle-Asp(OtBu)-N-methyl-Phe-PAL-resin was suspended and shaken in 20% piperidine/DMF (Steps 1-6) using the Fmoc protocol and coupled to the compound of Example 26 N-hydroxysuccinyl 3-(4-carboxymethyl)phenylpropionate (610 mg, 2 mmol) which was dissolved in 20 ml DMF/CH₂Cl₂ (1:1) by volume and allowed to react for 6 h at room temperature, then washed (Steps 8-16) and dried to yield 1.45 g of Desamino-Phe(4-CH₂COOH)-Nle-Gly-Trp-Nle-Asp-(OtBu)-N-methyl-Phe-PAL-resin. This peptidyl-resin was suspended and shaken in 50 ml of TFA/EDT/CH₂Cl₂ (14/1/5) by volume for 1 h at room temperature, then the PAL resin was filtered off and washed with 20 ml TFA (CH₂Cl₂ (1:1) by volume. The combined filtrates were evaporated to dryness, precipitated with ether, filtered off and dried to yield 240 mg of crude peptide.

120 mg of the crude peptide was purified by preparative HPLC on a (2.3 x 30 cm) micro Bondapak C-18 column. The peptide was eluted with a linear gradient of 5 to 65% of 0.022% TFA/CH₃CN at a flow rate of 8 ml/min, detection at 280 nm. The main peak was collected and lyophilized to yield 10 mg (6.2%) of Desamino-Phe(4-CH₂COOH)-Nle-Gly-Trp-Nle-Asp-N-methyl-Phe-NH₂. This material was homogeneous by HPLC and gave the correct amino acid analysis and MS. Amino acid analysis: Asp, 0.96 (1); Gly, 0.93 (1); Nle, 2.00 (2); N-methyl-Phe, n.d.; Empirical formula: C₅₀H₆₄N₈O₁₁; M.W. 953.11.

EXAMPLE 71

Preparation of Desamino-Phe(4-CH₂COOH)-Nle-(D)Ala-Trp-Nle-Asp-N-methyl-Phe-NH₂

1.25 g (0.43 mmol) of Fmoc-Trp-Nle-Asp(OtBu)-N-methyl-Phe-PAL-resin obtained from Example 70 was subjected to

sequential solid phase synthesis using the Fmoc protocol. All couplings except the last residue were performed using the DCC/HOBt procedure. At Step 7 the Fmoc-amino acid, DCC and HOBt were added with the corresponding reaction times as follows: Fmoc(D)-Ala-OH (480 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by volume DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. Fmoc-Nle-OH (520 mg, 1.5 mmol), DCC (310 mg, 1.5 mmol) and HOBt (270 mg, 2 mmol) were dissolved in 20 ml of 1:1 by volume of DMF/CH₂Cl₂ and allowed to couple for 60 min at room temperature. At this point the Fmoc-Nle-(D)Ala-Trp-Nle-Asp(OtBu)-N-methyl-Phe-PAL-resin was suspended and shaken in 20% piperidine/DMF (Steps 1-6) using the Fmoc protocol and coupled to the compound of Example 26 N-hydroxysuccinyl 3-(4-carboxymethyl)phenylpropanoate (610 mg, 2 mmol) which was dissolved in 20 ml DMF/CH₂Cl₂ (1:1) by volume and allowed to react for 6 h at room temperature, then washed (Steps 6-16) and dried to yield 1.5 g of Desamino-Phe(4-CH₂COOH)-Nle-(D)-Ala-Trp-Nle-Asp(OtBu)-N-methyl-Phe-PAL-resin. This peptidyl-resin was suspended and shaken in 50 ml of TFA/EDT/CH₂Cl₂ (14/1/5) by volume for 1h at room temperature, then the resin was filtered off and washed with 20 ml TFA/CH₂Cl₂ (1:1) by volume. The combined filtrates were evaporated to dryness, precipitated with ether, filtered off and dried to yield 300 mg of crude peptide.

150 mg of the crude peptide was purified by preparative HPLC on a (2.3 x 30 cm) micro Bondapak C-18 column. The peptide was eluted with a linear gradient of 5 to 65% of 0.022% TFA/CH₃CN at a flow rate of 8 ml/min, detection at 280 nm. The main peak was collected and lyophilized to yield 20 mg (12.1%) of Desamino-Phe(4-CH₂COOH)-Nle-(D)Ala-Trp-Nle-Asp-N-methyl-Phe-NH₂. This material was homogeneous by HPLC and gave the correct amino acid analysis and MS. Amino acid analysis: Asp, 0.95 (1); Ala, 1.03 (1);

Nle, 2.02 (2); N-methyl-Phe, n.d.; Trp, n.d.; Empirical
formula: $C_{51}H_{66}N_8O_{11}$; M.W. 967.13.

EXAMPLE 72

5

In Vitro Receptor Binding Assay

Frozen bovine striatum (approx. 5 g) or fresh rat
pancreas (approx. 5 g) cleaned of fat and extraneous tissue
10 were homogenized in HEPES buffer #1 (10 mM HEPES + 130 mM
NaCl + 5 mM $MgCl_2$, pH 7.4) using 35 parts buffer per 1
part tissue on a wet weight/volume basis (approx. 175 ml).
The tissue was homogenized 2 x for approx. 15 sec. at 0°C
using a Polytron homogenizer at a setting of 6. The tissue
15 was isolated by centrifugation at 48,000 x g for 10 min at
0°C. The resulting tissue pellet was resuspended in HEPES
buffer #2 (10 mM HEPES + 130 mM NaCl + 5 mM $MgCl_2$ + 1 mg/L
phenylmethanesulfonyl fluoride (PMSF) + 200 mg/L
Bacitracin): 1 part striatal tissue (original wet weight)
20 per 80 parts buffer and 1 part pancreas tissue (original wet
weight) per 500 to 1000 parts buffer.

Incubation was initiated by combining various
concentrations of native CCK-8 or peptides of the invention
25 with 3H -CCK-8- (SO_3H) (final conc. = 0.15 nM) and tissue
homogenate (striatum approximately 0.26 mg protein in 2 ml
final volume; pancreas approximately 0.100 mg protein in
1 ml final volume). Samples were incubated for 30 min at
25°C and the incubation terminated by pouring the mixture
30 onto a pre-wetted Whatman GF/B filter on a Sandbeck Vacuum
Filtration Manifold. The incubation tubes were washed with
2 x 3 ml of ice-cold HEPES Buffer #2 and the wash filtered
through the GF/B filter. The filter was air dried for 10 min
and then placed in a scintillation vial with 12 ml of
35 DuPont/NEN Aquasol scintillation cocktail. The vials were
shaken overnight and then counted using a liquid
scintillation spectrometer. Non-specific binding was

determined in the presence of 1 micromolar native CCK-8 and subtracted from all samples to determine specific binding. The concentration of peptide necessary to inhibit 50% of total specific ^3H -CCK-8-(SO_3H) binding (IC_{50} value) was determined by log-probit analysis. The results are summarized in Table I.

EXAMPLE 73

Two-Meal Feeding Assay

Male Sprague-Dawley (CD) rats weighting 180-200 grams (Charles River Breeding Laboratories) were acclimated to a 12h light/dark cycle (6 a.m. to 6 p.m.) in a room kept at 22°C. They were subsequently fasted for two days, weighed, placed in individual cages, and a four-day period of meal training was begun. During this time the rats were given ground laboratory chow (Purina Lab Chow) in jars for one hour from 9:00 a.m. until 10:00 a.m., the jars were removed from 10:00 a.m. to 12:00 p.m., and placed back in the cages from 12:00 until 1:00 p.m. Under this '1-2-1' meal feeding regime, most rats learn to eat approximately as much per day during the two hours they have access to food as rats which have food ad libitum over the entire 24-hour day. On the fourth day, the rats were weighted again, and any which lost more than five grams body weight were excluded from the test. The animals were then distributed into experimental (n = 5 to 6) and control groups (n = 6-12), but not matched for body weight.

Peptides of the invention were suspended either in saline, if soluble, or in 0.5% DMSO/saline, if insoluble, at concentrations of 0 to 320 $\mu\text{g/ml/kg}$ body weight and were administered intraperitoneally 15 min before the first meal on day 5 of meal feeding. The rats were then given their meals as they had been during the previous four days, and the food cups were weighed both before and after each meal

to determine food consumption. Food intake was expressed as
a mean and standard error of the mean in percent of control
values for the various groups. The treated groups were
compared to the control groups by t-test analysis. The
5 results are summarized in Table 1.

While the invention has been described in connection
with the preferred embodiment, it is not intended to limit
the scope of the invention to the particular form set forth,
10 but, on the contrary, it is intended to cover such
alternatives, modifications, and equivalents as may be
included within the spirit and scope of the appended claims.

15

20

25

30

35

TABLE I (page 1)

Peptides	Ex. No.	Bovine Striatum (nM)	Rat Pancreas (nM)	Food Intake		
				Dose Microgram/kg	1st Meal % of Control	2nd Meal
Asp-Tyr(SO ₃ H)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	CK-8	1-3.2	1-4.6	32	27±17***	149±6***
Ac-(D,L)Phe(3-COOH)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	42	32	128	32 320	125±12 98±14	98±9 110±12
Ac-(D,L)Phe(4-COOH)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	43	34	62	320 1060	43±11*** 41±11***	118±13 113±8
Ac-(D)Phe(4-COOH)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	44	68	210	32 320	89±6 85±4	123±9 146±6*
Ac-Phe(4-COOH)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	45	62	30	32 320	79±12 12±6***	140±6* 165±8**
Ac-(D,L)Phe(4-CH ₂ COOH)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	46	47	3.6	3 16 32 320	97±15 26±5*** 12±4*** 1±1***	98±6 157±18** 133±10** 142±6**
Ac-(D)Phe(4-CH ₂ COOH)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	47	145	21	3 16 320	95±4 53±5*** 31±19**	100±10 172±6** 141±12*

35

30

25

20

15

10

5

TABLE I (page 2)

Peptides	Ex. No.	Bovine Striatum (nM)	Rat Pancreas (nM)	Dose Microgram/kg	Food Intake % of Control	
					1st Meal	2nd Meal
Ac-Phe(4-CH ₂ COOH)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	48	54	2.6	3 16 320	76+6* 38+5*** 1+0***	139+16 179+20* 116+14
Ac-(D)Phe(4-CH ₂ COOC ₂ H ₅)-Met-Gly-Trp-Met-Asp (COOC ₂ H ₅)-Phe-NH ₂	49	10000	1000	1 3 32 320	93+3 71+8** 12+6*** 1+0***	98+15 165+14** 228+16*** 109+5
Ac-Phe(4-CH ₂ COCH ₃)-Met-Gly-Trp-Met-Asp(COOC ₂ H ₅) -Phe-NH ₂	50	10000	3600	3 32 96 320	97+3 66+6* 50+14** 16+6***	112+8 138+5* 169+22** 165+8***
Ac-Phe(4-CH ₂ CH ₂ COOH)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	51	620	390	320	103+5	114+7
Ac-(D,L)Phe(4-CF ₂ COOH)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	52	66	81	32 100	54+6*** 10+2***	186+3*** 201+3***

TABLE I (page 3)

Peptides	Ex. No.	Bovine Striatum (nM)	Rat Pancreas (nM)	Dose Microgram/kg	Food Intake % of Control	
					1st Meal	2nd Meal
Ac-(D,L)Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	53	1.6	22	16 32 320	49±11** 28±9*** 14±5***	144±8** 146±12** 131±8*
Ac-(D)Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	54	50	120	3 16	86±16 89±6	140±25 145±12
Ac-Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	55	0.51	3.0	3 16	70±10 32±10***	156±8* 191±10**
Ac-(D,L)Phe(4-CH ₂ COOH)-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-NH ₂	56	10000	4650	32 320	89±7 80±3**	98±9 121±2
Ac-(D,L)Phe(4-tetrazole)-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-NH ₂	57	580	1100	32 320	75±6** 36±3***	160±14** 148±6*
Ac-(D,L)Phe(3-CH ₂ COOH)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	58	53	23	32 320	40±9*** 0±0***	169±11*** 163±13***
Des amino Phe(4-CH ₂ COOH)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	59	22	57	3 32	59±5** 10±4***	197±21** 121±14

TABLE I (page 4)

Peptides	Ex. No.	Bovine Striatum (nM)	Rat Pancreas (nM)	Dose Microgram/kg	Food Intake	
					1st Meal % of Control	2nd Meal
Ac-(D,L)Phe(4-CH ₂ -tetrazole)-Met-Gly-Trp-Met-Asp- Phe-NH ₂	60	340	730	32	63±9**	140±15
				320	124±4***	165±19

TABLE I (page 5)

Peptides	Ex. No.	Bovine Striatum (nM)	Rat Pancreas (nM)	Food Intake		
				Dose Microgram/kg	1st Meal % of Control	2nd Meal % of Control
Ac-(D)Phe(4-CH ₂ -tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	61	62	380	32 100 320	80+6* 58+6*** 47+13***	119+28 159+9* 170+21*
Ac-Phe(4-CH ₂ -tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	62	18	67	3 10 32 100	85+6* 65+4*** 25+4*** 16+3***	118+9 129+9*** 170+12** 179+14**
Ac-(D,L)Phe(4-CH ₂ -tetrazole)-Nle-Gly-Trp-Nle-Asp-N-methyl-Phe-NH ₂	63	260	640	32 320	53+10** 19+5***	171+3*** 141+17
Ac-(D)Phe(4-CH ₂ -tetrazole)-Nle-Gly-Trp-Nle-Asp-N-methyl-Phe-NH ₂	64	160	800	10 32 100 320	112+10 79+5* 56+6*** 21+6***	102+9 148+12* 173+5*** 207+20***



35 30 25 20 15 10 5

TABLE I (page 6)

Peptides	Ex. No.	Bovine Striatum (nM)	Rat Pancreas (nM)	Dose Microgram/kg	Food Intake	
					1st Meal % of Control	2nd Meal
Ac-Phe(4-CH ₂ -tetrazole)-Nle-Gly-Trp-Nle-Asp-N-Methyl-Phe-NH ₂	65	69	260	10 100	86+11 30+4***	133+11* 164+9***
Des amino Phe(4-CH ₂ -tetrazole)-Nle-Gly-Trp-Nle-Asp-N-Methyl-Phe-NH ₂	66	160	710	3 32 100 320	94+9 68+6* 56+5** 34+5***	109+16 179+16** 205+7*** 177+14***
Des amino Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH ₂	67	5.1	35	3 32	99+9 26+4***	116+6 204+9***
Des amino Phe(4-tetrazole)-Lys-Gly-Trp-Met-Asp-N-Methyl-Phe-NH ₂	68	1400	2900	32 320	77+8* 24+3***	165+18** 177+14***

TABLE I (page 7)

Peptides	Ex. No.	Bovine Striatum (nM)	Rat Pancreas (nM)	Dose Microgram/kg	1st Meal % of Control	Food Intake 2nd Meal
Des amino-Phe(4-CH ₂ -COOH)-Met-Gly-Trp-Met-Asp-N-Methyl-Phe-NH ₂	69	3.2	1.8	1 3 10 32	99+7 99+9 53+11* 16+5***	106+8 127+10 163+10*** 157+13*
Des amino-Phe(4-CH ₂ -COOH)-Nle-Gly-Trp-Nle-Asp-N-Methyl-Phe-NH ₂	70	200	330	3 10 32 320	78+6* 65+7** 27+6*** 9+4***	151+16* 165+7*** 140+12** 113+9
Des amino-Phe(4-CH ₂ -COOH)-Nle(D)Ala-Trp-Nle-Asp-N-Methyl-Phe-NH ₂	71	1400	1800	36 320	93+9 105+4	109+6* 89+12*

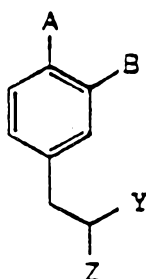
Values significantly different than their respective controls.

*** $p \leq 0.001$, ** $p \leq 0.01$, * $p \leq 0.05$

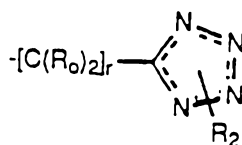
CLAIMS:

The claims defining the invention are as follows:

1. An analog of a tyrosine sulfate- or tyrosine phosphate residue(s) containing peptide wherein one or more of such residue(s) is/are replaced by (a) residue(s) of the formula

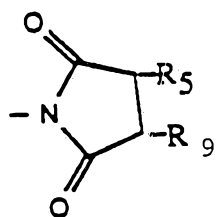


wherein one of A or B is selected from the group consisting of: $-[C(R_O)_2]_S-CO_2R_1$ or,



and the other of A or B is H;

Z is H, $-NH_2$, $-NH-$, lower alkyl, $-NHCOR_3$, $-NHCO_2R_4$, or



R_0 is H or F;

5 R_1 is H, substituted or unsubstituted lower alkyl with the substituents selected from the group consisting of hydroxyl, halogen, or aryl;

10 R_2 is H, lower alkenyl, lower alkyl or lower alkyl substituted by 1 to 3 aryl groups;

R_3 is H, lower alkyl, alkyl substituted by one or two aryl groups or aryl;

15 R_4 is lower alkyl or alkyl substituted by one or two aryl groups;

20 R_5 and R_9 are each independently H, lower alkyl, or taken together and including the carbon atoms to which they are attached may form a six numbered ring which may be aromatic;

s and r are 0, 1 or 2;

25 and Y is $-\text{COOH}$, alkoxycarbonyl, $-\text{CONH}_2$ or $-\text{CO}-$;

with the proviso that if Z is other than $-\text{NH}-$ then Y is $-\text{CO}-$; or if Y is other than $-\text{CO}-$ then Z is $-\text{NH}-$; and
30 pharmaceutically acceptable salts thereof.

2. A compound as claimed in claim 1 which is an analog of a tyrosine sulfate residue(s) containing peptide.

3. A compound as claimed in claim 2 which is an analog
5 of cholecystokinin (CCK).

4. The compound of claim 3 wherein the tyrosine sulfate residue(s) of CCK is/are replaced by a residue of Formula I wherein Z is H, NHCOR_3 , or NHCO_2R_4 and R_3
10 is lower alkyl, R_4 is lower alkyl optionally substituted with one to three aryl groups.

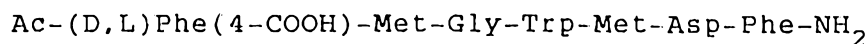
5. The compound of claim 4 wherein R_3 is CH_3

15 6. The compound of claim 5 wherein one of A or B is $-\text{[C(R}_O)_2]_s-\text{CO}_2\text{R}_1$ and the other of A or B is hydrogen.

7. The analog of claim 6 wherein A is
20 $-\text{[C(R}_O)_2]_s-\text{CO}_2\text{R}_1$ and B is H.

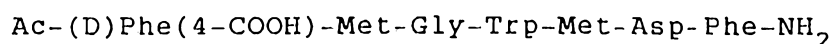
8. The compound of claim 7 wherein s is 0 and R_1 is H.

25 9. The compound of claim 8 having the formula:



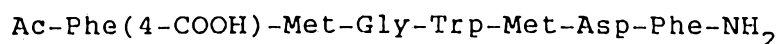
10. The compound of claim 8 having the formula:

30



11. The compound of claim 8 having the formula:

35



12. The compound of claim 7 wherein s is 1.

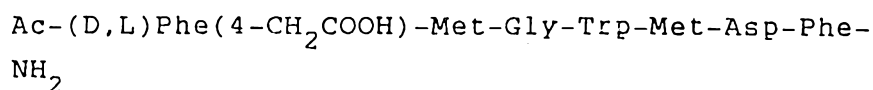
13. The compound of claim 12 wherein R_0 is H or F.

5 14. The compound of claim 13 wherein R_1 is H.

15. The compound of claim 14 wherein R_0 is H.

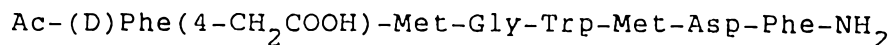
16. The compound of claim 15 having the formula:

10



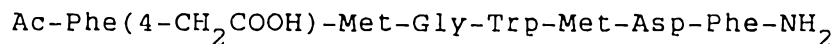
17. The compound of claim 15 having the formula:

15



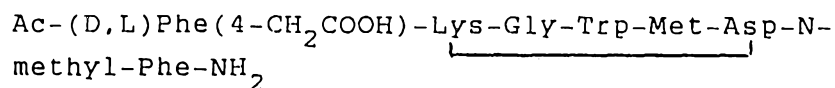
18. The compound of claim 15 having the formula:

20



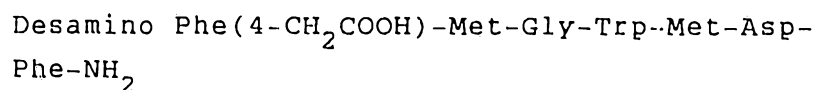
19. The compound of claim 15 having the formula:

25



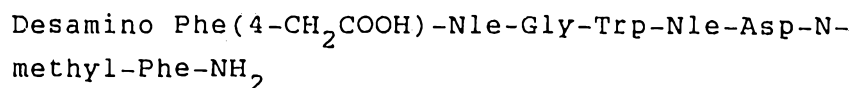
20. The compound of claim 15 having the formula:

30

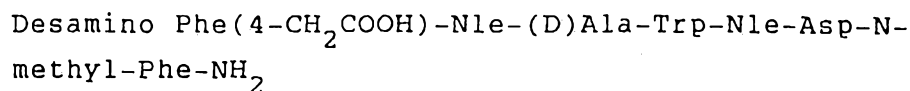


21. The compound of claim 15 having the formula:

35



22. The compound of claim 15 having the formula:

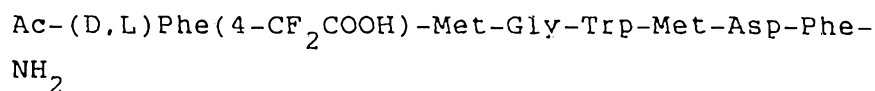


5

23. The compound of claim 13 wherein R₀ is F.

24. The compound of claim 23 having the formula:

10



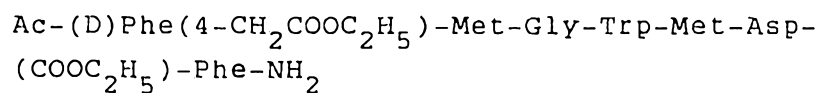
25. The compound of claim 13 wherein R₀ is H.

15

26. The compound of claim 25 wherein R₁ is CH₂CH₃.

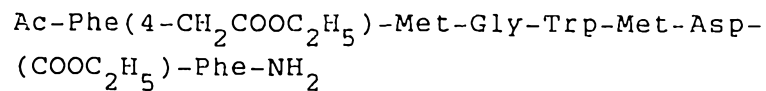
27. The compound of claim 26 having the formula:

20



28. The compound of claim 26 having the formula:

25



29. The compound of claim 7 wherein S is 2.

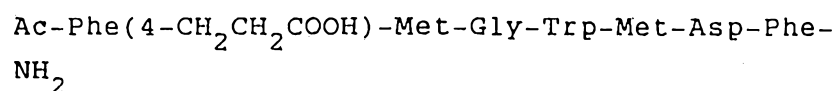
30. The compound of claim 29 wherein R₀ is H.

30

31. The compound of claim 30 wherein R₁ is H.

32. The compound of claim 31 having the formula:

35



33. The compound of claim 6 wherein B is $-[C(R_O)_2]_s-CO_2R_1$ and A is H.

34. The compound of claim 33 wherein s is O.

5

35. The compound of claim 34 wherein R_1 is H.

36. The compound of claim 35 having the formula:

10

Ac-(D,L)Phe(3-COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂

37. The compound of claim 33 wherein S is 1.

38. The compound of claim 37 wherein R_0 is H.

15

39. The compound of claim 38 wherein R_1 is H.

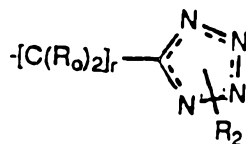
40. The compound of claim 39 having the formula:

20

Ac-(D,L)Phe(3-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂

41. The compound of claim 5 wherein one of A or B is

25

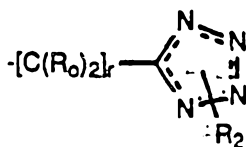


and the other of A or B is hydrogen.

30

35

42. The compound of claim 41 wherein A is

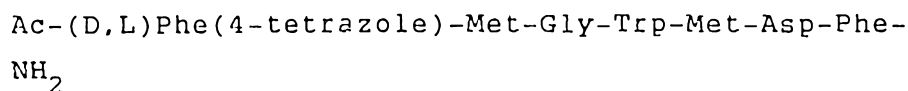


and B is H.

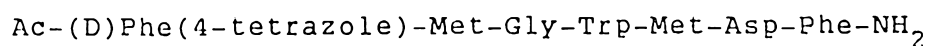
43. The compound of claim 42 wherein r is 0.

44. The compound of claim 43 wherein R_2 is H.

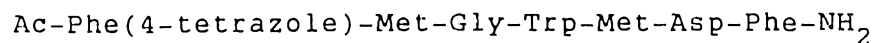
45. The compound of claim 44 having the formula:



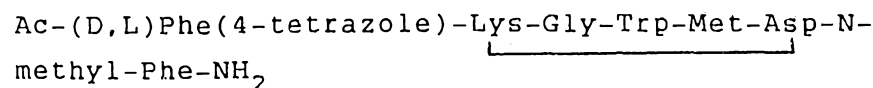
46. The compound of claim 44 having the formula:



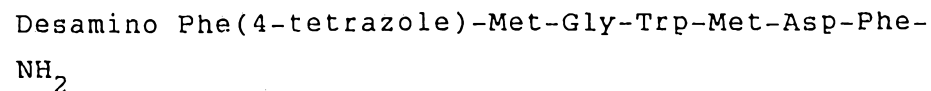
47. The compound of claim 44 having the formula:



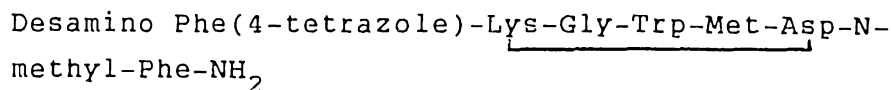
48. The compound of claim 44 having the formula:



49. The compound of claim 44 having the formula:



50. The compound of claim 44 having the formula:



5

51. The compound of claim 42 wherein r is 1.

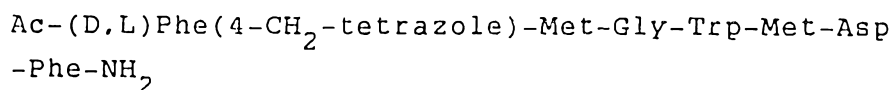
52. The compound of claim 51 wherein R₀ is H.

10

53. The compound of claim 52 wherein R₂ is H.

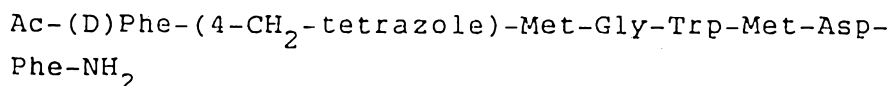
54. The compound of claim 53 having the formula:

15



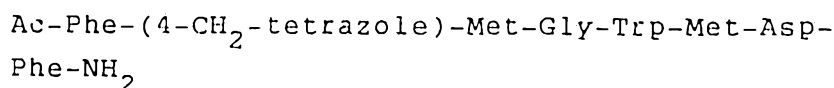
55. The compound of claim 53 having the formula:

20



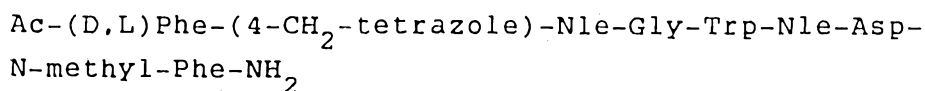
56. The compound of claim 53 having the formula:

25



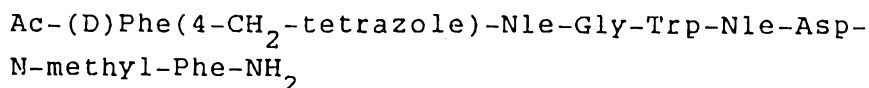
57. The compound of claim 53 having the formula:

30

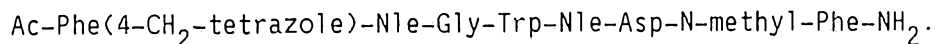


58. The compound of claim 53 having the formula:

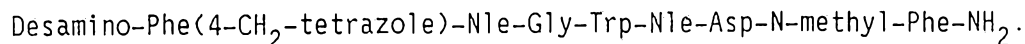
35



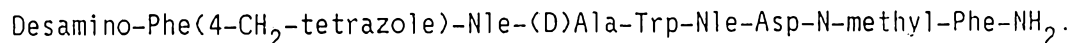
59. The compound of claim 53 having the formula:



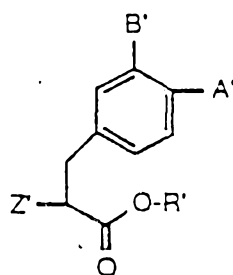
60. The compound of claim 53 having the formula:



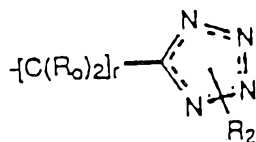
5 61. The compound of claim 53 having the formula:



62. A compound when used for the preparation of an analog of a tyrosine sulphate- or tyrosine phosphate residue(s) containing peptide as defined in claim 1, said compound having the formula:



wherein one of A' or, B', is selected from $-[C(R_0)_2]_s-CO_2R_1'$, or



and the other of A' or B' is H wherein R₀ is H or F, R₁' is H,

substituted or unsubstituted lower alkyl with the substituents selected

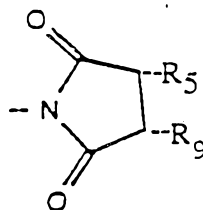
15 from hydroxyl, halogen, or aryl, R₂ is H, lower alkenyl, lower alkyl or lower alkyl substituted by 1 to 3 aryl groups,

r is 0-2

s is 1-2

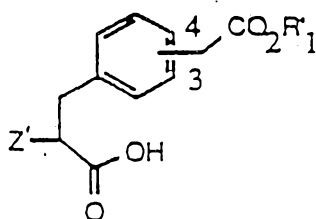
R' is H, lower alkyl or lower alkyl substituted by 1 to 3 aryl groups,

20 Z' is H, lower alkyl, -NH₂, -NHCOR₃, -NHCO₂R₄, or



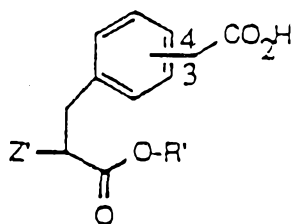
and R_3 and R_4 are H, lower alkyl or lower alkyl substituted by one, two or three aryl groups; and R_5 and R_9 are each independently H, lower alkyl, or taken together and including the carbon atoms to which they are attached may form a six membered ring which may be aromatic.

- 5 63. A compound when used for the preparation of an analog of a tyrosine sulphate- or tyrosine phosphate residue(s) containing peptide as defined in claim 1, said compound having the formula:



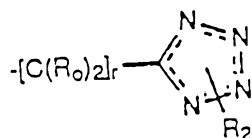
wherein Z' and R_1' are as defined in claim 62.

- 10 64. A compound when used for the preparation of an analog of a tyrosine sulphate- or tyrosine phosphate residue(s) containing peptide as defined in claim 1, said compound having the formula:



wherein R' and Z' are as defined in claim 62.

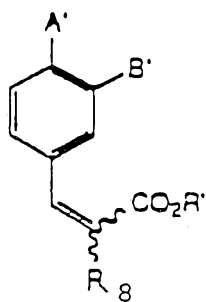
- 15 65. The compound of claim 62 wherein one of A' or B' is



and the other of A' or B' is H.

- 20 66. A compound when used for the preparation of an analog of a tyrosine sulphate- or tyrosine phosphate residue(s) containing peptide as defined in claim 1, said compound having the formula:



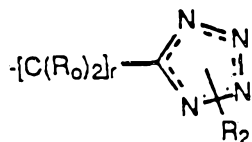


wherein A', B', and R' are as defined in claim 62 and R₈ is H or lower alkyl.



67. The compound of claim 66 wherein A' is

5

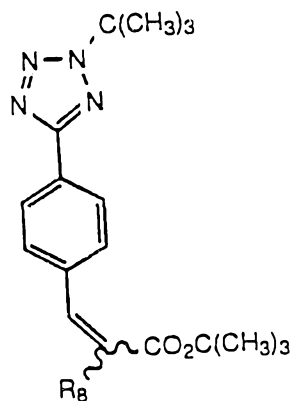


and B' is H, and R₀, r, and R₂ are as defined in claim
10 62.

68. The compound of claim 67 wherein r=0.

69. The compound of claim 68 wherein R' is t-butyl and
15 R₈ is H said compound having the formula:

20

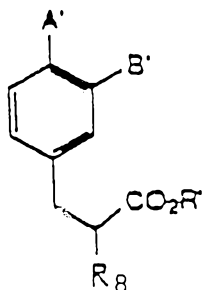


25

30

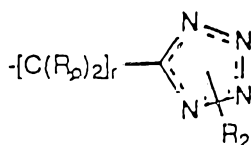
35

70. A compound when used for the preparation of an analog of a tyrosine sulphate- or tyrosine phosphate residue(s) containing peptide as defined in claim 1, said compound having the formula:



5 wherein A', B', and R' are as defined in the second embodiment and R₈ is H or lower alkyl.

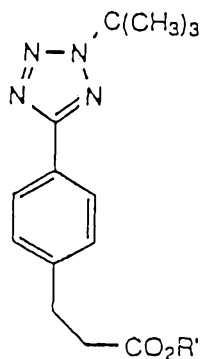
71. The compound of claim 70 wherein A' is



and B' is H and R₀, r, and R₂ are as defined in claim 62.

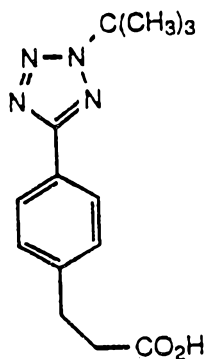
10 72. The compound of claim 71 wherein r=0.

73. The compound of claim 72 wherein R' is lower alkyl and R₂ is t-butyl said compound having the formula:



74. The compound of claim 70 wherein R' is H and R₂ is t-butyl said compound having the formula:

5



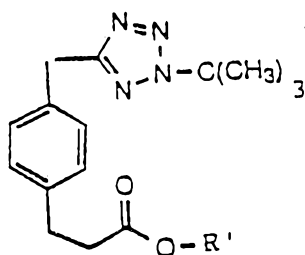
10

75. The compound of claim 71 wherein r=1 and R₀ is H.

15

76. The compound of claim 75 wherein R' is lower alkyl and R₂ is t-butyl said compound having the formula:

20

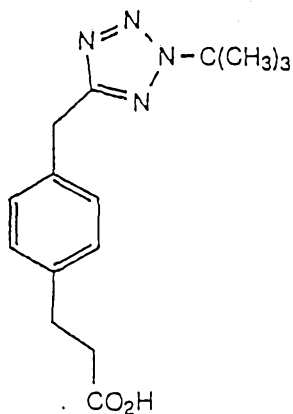


25

30

35

77. The compound of claim 76 wherein R' is H and R₂ is t-butyl said compound having the formula:



78. A process for the manufacture of a compound according to any one of claims 1 to 61, which process comprises treating the resin bound peptide with appropriate cleavage reagent(s) and if desired, converting such a peptide obtained into a pharmaceutically acceptable salt.

79. A pharmaceutical composition containing a compound according to any one of claims 1 to 61 and a non-toxic, inert, therapeutically acceptable carrier material.

80. A pharmaceutical composition for suppressing appetite containing a compound according to any one of claims 1 to 61 and a therapeutically inert carrier material.

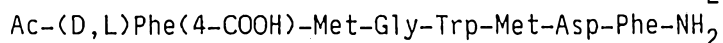
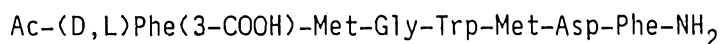
81. The use of a compound according to any one of claims 1 to 61 as a therapeutically active substance.

82. The use of a compound according to any one of claims 1 to 61 in the suppression of appetite.

83. Compounds as claimed in any one of claims 1 to 61 whenever prepared according to a process of claim 78.

84. A method for suppressing appetite in subjects by administering an appetite suppressing effective amount of an analog of CCK wherein the tyrosine sulfate residue is replaced by a residue of Formula I as defined in claim 1.

85. The method of claim 84 wherein the CCK analog is a compound selected from:



- Ac-(D)Phe(4-COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂
 Ac-Phe(4-COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂
 Ac-(D,L)Phe(4-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂
 Ac-(D)Phe(4-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂
 5 Ac-Phe(4-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂
 Ac-(D)Phe(4-CH₂COOC₂H₅)-Met-Gly-Trp-Met-Asp(COOC₂H₅)-Phe-NH₂
 Ac-Phe(4-CH₂COOC₂H₅)-Met-Gly-Trp-Met-Asp(COOC₂H₅)-Phe-NH₂
 Ac-Phe(4-CH₂CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂
 Ac-(D,L)Phe(4-CF₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂
 10 Ac-(D,L)Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂
 Ac-(D)Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂
 Ac-Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂
 Desamino Phe(4-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂
 Ac-(D,L)Phe(3-CH₂COOH)-Met-Gly-Trp-Met-Asp-Phe-NH₂
 15 Ac-(D,L)Phe(4-CH₂-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂
 Ac-(D,L)Phe(4-CH₂COOH)-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-NH₂
 Ac-(D,L)Phe(4-tetrazole)-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-NH₂
 Desamino Phe(4-CH₂COOH)-Nle-Gly-Trp-Nle-Asp-N-methyl-Phe-NH₂
 Desamino Phe(4-CH₂COOH)-Nle-(D)Ala-Trp-Nle-Asp-N-methyl-Phe-NH₂
 20 Desamino Phe(4-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂
 Desamino Phe(4-tetrazole)-Lys-Gly-Trp-Met-Asp-N-methyl-Phe-NH₂
 Ac-(D)Phe(4-CH₂-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂
 Ac-Phe(4-CH₂-tetrazole)-Met-Gly-Trp-Met-Asp-Phe-NH₂
 Ac-(D,L)Phe(4-CH₂-tetrazole)-Nle-Gly-Trp-Nle-Asp-N-methyl-Phe-NH₂
 25 Ac-Phe(4-CH₂-tetrazole)-Nle-Gly-Trp-Nle-Asp-N-methyl-Phe-NH₂
 Desamino-Phe(4-CH₂-tetrazole)-Nle-Gly-Trp-Nle-Asp-N-methyl-Phe-NH₂
 86. The method of claim 84 or claim 85 wherein the analog is administered intranasally.
 87. The method of claim 84 or claim 85 wherein the analog is
 30 administered parenterally.
 88. An analog of CCK wherein the tyrosine sulfate residue is replaced by a residue of Formula I as defined in claim 1, which analog is substantially as hereinbefore described with reference to Example 69.



89. A process for the manufacture of an analog of CCK wherein the tyrosine sulfate residue is replaced by a residue of Formula I as defined in claim 1, which process is substantially as hereinbefore described with reference to any one of Examples 42 to 71.

5 90. The CCK analogue product when produced by the process of claim 89.

91. A pharmaceutical composition for appetite suppression in a subject, which composition comprises an amount effective as an appetite suppressant in said subject of the CCK analog product of claim 90
10 together with a pharmaceutically acceptable carrier, diluent, adjuvant and/or excipient.

92. A method of appetite suppression in a subject, which method comprises administering to a subject in need of appetite suppression, an amount effective as an appetite suppressant in said subject of the CCK
15 analog product of claim 90, or the composition of claim 91.

DATED this TWELFTH day of AUGUST 1991
F. Hoffmann-La Roche & Co. Aktiengesellschaft

Patent Attorneys for the Applicant
SPRUSON & FERGUSON

