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(54) **Title:** ANAPLASTIC THYROID CANCERS HARBOR NOVEL ONCOGENIC MUTATIONS OF THE ALK GENE

(57) **Abstract:** The present invention relates to diagnostic, therapeutic and prognostic methods for thyroid cancers. In one embodiment, the present invention provides a method for treating a anaplastic thyroid cancer (ATC) patient comprising the step of administering to the patient an effective amount of an anaplastic lymphoma kinase (ALK) inhibitor. In another embodiment, a method for diagnosing ATC in patient comprises the step of performing an assay on a biological sample from the patient to identify the presence or absence of a C3592T and/or a G3602A mutation in exon 23 of the ALK gene according to SEQ ID NO:6, wherein the presence of either of both of the mutations correlates with a diagnosis of ATC in the patient.



CROSS-REFERENCE TO RELATED APPLICATIONS

STATEMENT OF GOVERNMENTAL INTEREST

10 FIELD OF THE INVENTION

INCORPORATION-BY-REFERENCE OF MATERIAL SUBMITTED
ELECTRONICALLY

BACKGROUND OF THE INVENTION

Recently, ALK mutations were found in 6% to 14% of sporadic neuroblastomas (10-14). ALK mutations were also reported in familial neuroblastomas (13, 14). Moreover, genetic amplification of the ALK gene could also occur in neuroblastomas or cell lines

derived from this tumor (10, 11, 13, 15). Except for occasional mutations in the juxtamembrane domain, most ALK mutations identified so far are within the tyrosine kinase domain of ALK. ALK mutations and/or copy gain were found particularly in advanced and metastatic neuroblastomas, and patients with ALK mutations had a worse prognosis (11, 12, 14). Several common ALK mutations were shown to be functional. For example, siRNA-mediated knockdown of the ALK expression in cell lines harboring ALK mutants F1174L or R1275Q caused cell apoptosis and suppression of cell proliferation (12-14). The F1174L and another mutant ALK, K1062M, were shown to display increased tyrosine kinase activity and promote cell focus formation, cell transformation, and xenograft tumorigenicity in nude mice (10). The oncogenicity of ALK F1174L and R1275Q was also shown in another study (12). Genetic copy gain of the ALK is also functionally important, as suggested by the demonstration that inhibition of ALK in neuroblastoma cell lines harboring ALK copy gain induced cell apoptosis through reduced signaling of the PI3K/Akt and MAP kinase pathways (15).

Mutations of the ALK gene have not been reported in human cancers other than neuroblastomas. As described herein, the present inventors investigated the mutation status of the ALK gene in various thyroid cancers, including well-differentiated papillary thyroid cancer (PTC) and follicular thyroid cancer (FTC) and undifferentiated anaplastic thyroid cancer (ATC). Prompted by the finding of ALK mutations in ATC, a rapidly aggressive and deadly human cancer (16), the present inventors also examined melanoma and colon carcinoma for ALK mutation. Indeed, identifying mutations in human cancers is highly desirable because it can lead to the development of new therapeutics that target such fusion or mutant proteins, and to new diagnostics for identifying patients that have such gene mutations.

SUMMARY OF THE INVENTION

The present invention is based, at least in part, on the discovery of ALK gene mutations in thyroid cancer that may rationalize clinical evaluation of ALK inhibitors in this setting. In undifferentiated anaplastic thyroid cancer (ATC), the present inventors identified two novel point mutations in exon 23 of the ALK gene, C3592T and G3602A, in exon 23 of the *ALK* gene with a prevalence of 11.11%, but found no mutations in the matched normal tissues or in well-differentiated thyroid cancers. These two mutations, resulting in the L1198F and G1201E amino acid changes, respectively, both reside within the ALK tyrosine kinase domain where they dramatically increased tyrosine kinase activities. Similarly, these mutations heightened the ability of ALK to activate the PI3K/Akt and MAP kinase pathways

in established mouse cells. Further investigation demonstrated that these two ALK mutants strongly promoted cell focus formation, anchorage-independent growth, and cell invasion. Similar oncogenic properties were observed in the neuroblastoma-associated ALK mutants K1062M and F1174L, but not in wild-type ALK. Overall, the results reveal two novel gain-
5 of-function mutations of ALK in certain ATCs and they suggest efforts to clinically evaluate the use of ALK kinase inhibitors to treat patients who harbor ATCs with these mutations.

Furthermore, although these two novel mutations are found in thyroid cancer, they likely are also present in other human cancers and can therefore make ALK an effective therapeutic target also in those non-thyroid cancers harboring them. Indeed, the molecular
10 testing of these two novel ALK mutations in thyroid cancer as well as in other human cancers will be helpful in guiding their targeted treatments. These cancers may particularly include brain tumor, lymphoma, lung cancer, gastric cancer, pancreatic cancer, liver cancer, colon cancer, melanoma, breast cancer, ovarian cancer, cervical cancer, prostate cancer, testicle cancer, bone cancer, head and neck cancer, laryngeal cancer, leukemia, myofibroblastic
15 tumors, and other cancers.

Accordingly, in one aspect, the present invention provides methods for treating thyroid cancer. In one embodiment, a method for treating an anaplastic thyroid cancer (ATC) patient comprises the step of administering to the patient an effective amount of an anaplastic lymphoma kinase (ALK) inhibitor. In a specific embodiment, the ALK inhibitor is crizotinib.
20 In another embodiment, the patient has a C3592T mutation in exon 23 of the ALK gene. In yet another embodiment, the patient has a G3602A mutation in exon 23 of the ALK gene. The can further comprise the step of determining whether the patient has a C3592T and/or a G3602A mutation in exon 23 of the ALK gene prior to the administering step.

In another embodiment, a method for treating anaplastic thyroid cancer (ATC) in a
25 thyroid cancer patient comprises the steps of (a) determining whether the thyroid cancer patient has a C3592T and/or a G3602A mutation in exon 23 of the anaplastic lymphoma kinase (ALK) gene; and (b) treating the patient with an ALK inhibitor therapy if the patient has either or both of the mutations. Alternatively, a method for treating anaplastic thyroid cancer (ATC) in a thyroid cancer patient comprises the step of treating the patient with an
30 ALK inhibitor therapy if the thyroid cancer patient has a C3592T and/or a G3602A mutation in exon 23 of the ALK gene. In such embodiments, the ALK inhibitor is crizotinib.

In another aspect, the present invention provides methods for treating cancer patients that harbor the mutations described herein. For example, a method for treating a cancer patient comprises the step of administering to the patient an effective amount of an ALK

inhibitor, wherein the cancer patient has a C3592T and/or a G3602A mutation in exon 23 of the ALK gene. In another embodiment, a method for treating a cancer patient comprises the steps of (a) determining whether the cancer patient has a C3592T mutation in exon 23 of the anaplastic lymphoma kinase (ALK) gene; and (b) treating the patient with an ALK inhibitor therapy if the patient has either or both of the mutations. In such embodiments, the ALK inhibitor is crizotinib. In other specific embodiments, the cancer is a neuroblastoma.

In another specific embodiment, a method for treating a neuroblastoma cancer patient comprises the step of administering to the patient an effective amount of an ALK inhibitor, wherein the patient has a C3592T and/or a G3602A mutation in exon 23 of the ALK gene. In a more specific embodiment, the ALK inhibitor is crizotinib.

In additional embodiments, the methods described herein can further comprise the step of administering an inhibitor of the PI3K/Akt pathway. In other embodiments, the methods described herein can further comprise the step of administering an inhibitor of the MAP kinase pathway. In a specific embodiment, a method for treating a cancer patient comprises the step of administering to the patient an inhibitor of a protein or pathway selected from the group consisting of ALK, the PI3K/Akt pathway and the MAP kinase pathway, wherein the patient has a C3592T and/or a G3602A mutation in exon 23 of the ALK gene. In a more specific embodiment, the ALK inhibitor is crizotinib. In another embodiment, the patient has ATC. In yet another embodiment, the patient has a neuroblastoma.

In another aspect, the present invention provides diagnostic and prognostic methods relating to the mutations described herein. In one embodiment, a method for diagnosing ATC in patient comprises the step of performing an assay on a biological sample from the patient to identify the presence or absence of a C3592T and/or a G3602A mutation in exon 23 of the ALK gene according to SEQ ID NO:6, wherein the presence of either of both of the mutations correlates with a diagnosis of ATC in the patient. In another embodiment, a method for determining a patient's risk of developing ATC comprises the step of performing an assay on a biological sample from the patient to identify the presence or absence of a C3592T and/or a G3602A mutation in exon 23 of the ALK gene according to SEQ ID NO:6, wherein the presence of either of both of the mutations correlates with a prognosis that the patient has a higher risk of ATC than a patient without the mutations, and wherein the absence of the mutations correlates with a prognosis that the patient has a lower risk of ATC than a patient with either or both of the mutations.

In a more specific embodiment, a method for detecting ATC in a patient comprises the step of determining the presence of a C3592T and/or a G3602A mutation in exon 23 of the ALK gene according to SEQ ID NO:6 in a blood sample of a patient, wherein the presence of the mutation indicates ATC in the patient.

5 In yet another embodiment, a method for distinguishing ATC from non-ATC samples comprises the step of determining the presence of a C3592T and/or a G3602A mutation in exon 23 of the ALK gene according to SEQ ID NO:6 in thyroid sample of a patient, wherein the presence of either or both of the mutations indicates ATC and absence of either or both of the mutations indicates non-ATC. In a specific embodiment, the thyroid sample is a fine
10 needle aspirate (FNA). In another embodiment, the thyroid sample is a tissue sample. In yet another embodiment, the thyroid sample is a cytological sample. The mutations can be detected using methods and kits known to those of ordinary skill in the art. For example, as described below, genomic DNA can be isolated from a sample and then exon 23 can be PCR amplified and sequenced. The method may further comprise providing a diagnosis based on
15 the presence or absence of either or both of the mutations.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1A is a sequencing electropherogram of the ALK gene. Left, the sequencing results of the matched normal tissues of the 2 ATC cases, showing the wild-type ALK gene. Middle and right, the sequencing results of 2 ATC tumors. Top, the sequencing results of
20 sense and antisense strands of a region of exon 23 of the ALK gene in an ATC showing the heterozygous C>T mutation at nucleotide position 3,592 in codon 1,198, resulting in the L1198F amino acid change of ALK. Bottom, the sequencing results of sense and antisense strands of a region of exon 23 of the ALK gene in another ATC showing the heterozygous G>A mutation at nucleotide position 3,602 in codon 1,201, resulting in the G1201E amino
25 acid change. Arrows indicate the mutated nucleotides. Nucleotide numbers refer to the position within the coding sequence of the ALK gene, where position 1 corresponds to the first position of the translation initiation codon. All samples were sequenced in 2 repeated experiments with independent PCR by sense and antisense primers. FIG. 1B is a schematic diagram of the ALK. Shown are the relative positions of the novel somatic ALK mutations
30 L1198F and G1201E and the previously characterized mutations K1062M and F1174L from neuroblastoma. L1198F and G1201E are located in the tyrosine kinase domain of the ALK. FIG. 1C is an amino acid sequence alignment of the ALK proteins from 6 species. Shown are the L1198 and G1201 residues that are evolutionarily completely conserved among these

different species. Numbers indicate amino acid or codon positions. Amino acid sequences are numbered with the initiation codon (methionine) of each protein defined as number 1.

FIG. 2 shows the increased tyrosine kinase activities of ALK mutants L1198F and G1201E and their activation of the PI3K/Akt and MAP kinase pathways. FIG. 2A shows the results of an in vitro assay of tyrosine kinase activities of ALK mutants. NIH3T3 cells stably expressing Flag-tagged vector, wild-type ALK (wt-ALK), and each of ALK mutants as indicated were lysed. The cell lysates were assayed for tyrosine kinase activity as described in the Materials and Methods. The enzymatic activities were expressed as measured optical density value x 20. Results represent mean $\pm$ SD of 3 independent experiments. FIG. 2B shows the activation of the PI3K/Akt and MAP kinase pathways. This is reflected by increased phosphorylation of Akt (p-Akt) and phosphorylation of ERK (p-ERK), respectively. NIH3T3 cells stably transfected with the indicated vector constructs, as described in FIG. 1A cell lysate proteins, were subjected to Western blot analyses for the indicated proteins by using appropriate antibodies as described in Materials and Methods. Successful protein expression of Flag-tagged wild-type ALK and each of the ALK mutants is shown in the top row of FIG. 1B. The key molecules of the 2 pathways are shown in the subsequent rows. Total Akt, ERK, and b-actin were used for quality control of loading proteins.

FIG. 3 shows the focus-formation and anchorage-independent growth of cells promoted by ALK mutants. FIG. 3A shows the cell focus-forming activities of ALK mutants. Shown are images of adherent growth of NIH3T3 cells transfected with Flag-tagged vector, wild-type ALK (wt-ALK), and each of the ALK mutants indicated. Cells were cultured in regular medium with 10% fetal calf serum under standard conditions. Images of cell foci were photographed with 10x magnification after appropriate culture of cells as described in the Materials and Methods. FIG. 3B shows the number of cell foci formed with the indicated transfections. The number of transformed foci was counted 14 days after cell transfection. Results represent mean $\pm$ SD of 3 independent experiments. FIG. 3C shows the anchorage-independent cell growth of ALK mutants on soft agar. NIH3T3 cells stably transfected with Flag-tagged vector, wild-type ALK, and each of the ALK mutants indicated were seeded in soft agar, and colonies formed 4 weeks later were photographed with 40x magnification. FIG. 3D shows the analyses of the number of colonies. The number of cell colonies corresponding to C that were greater than 0.1 mm in diameter was counted. Results represent mean $\pm$ SD of 3 independent experiments.

FIG. 4 shows cell invasion promoted by ALK mutants. FIG. 4A shows the results from the in vitro invasion assay of NIH3T3 cells with various transfections. Cells transfected with Flag-tagged vector, wild-type ALK (wt-ALK), and each construct of the indicated ALK mutants. Cell invasion assay was conducted as described in Materials and Methods. Shown are the cells that invaded on the Matrigel matrix- coated polyethylene terephthalate (PET) membrane after removal of the noninvasive cells. FIG. 4B shows the number of invasive cells with the indicated transfections. Results of each column represent the mean $\pm$ SD of the numbers of invasive cells from 3 independent experiments.

DETAILED DESCRIPTION OF THE INVENTION

It is understood that the present invention is not limited to the particular methods and components, etc., described herein, as these may vary. It is also to be understood that the terminology used herein is used for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention. It must be noted that as used herein and in the appended claims, the singular forms “a,” “an,” and “the” include the plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to a “protein” is a reference to one or more proteins, and includes equivalents thereof known to those skilled in the art and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Specific methods, devices, and materials are described, although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention.

All publications cited herein are hereby incorporated by reference including all journal articles, books, manuals, published patent applications, and issued patents. In addition, the meaning of certain terms and phrases employed in the specification, examples, and appended claims are provided. The definitions are not meant to be limiting in nature and serve to provide a clearer understanding of certain aspects of the present invention.

I. Definitions

The term “ALK inhibitor” refers to an agent that inhibits the synthesis or biological activity of ALK. ALK inhibitors are known in the art and are described herein. The term also includes agents that have activity in addition to ALK inhibitory activity.

As used herein, the term “effective,” means adequate to accomplish a desired, expected, or intended result. More particularly, a “therapeutically effective amount” as provided herein refers to an amount of an ALK inhibitor of the present invention, either alone

or in combination with another therapeutic agent, necessary to provide the desired therapeutic effect, e.g., an amount that is effective to prevent, alleviate, or ameliorate symptoms of disease or prolong the survival of the subject being treated. In a specific embodiment, the term “therapeutically effective amount” as provided herein refers to an amount of an ALK inhibitor, necessary to provide the desired therapeutic effect, e.g., an amount that is effective to prevent, alleviate, or ameliorate symptoms of disease or prolong the survival of the subject being treated. As would be appreciated by one of ordinary skill in the art, the exact amount required will vary from subject to subject, depending on age, general condition of the subject, the severity of the condition being treated, the particular compound and/or composition administered, and the like. An appropriate “therapeutically effective amount” in any individual case can be determined by one of ordinary skill in the art by reference to the pertinent texts and literature and/or by using routine experimentation.

As used herein, the terms “treatment,” “treating,” and the like, refer to obtaining a desired pharmacologic and/or physiologic effect. The effect may be prophylactic in terms of completely or partially preventing a disease or symptom thereof and/or may be therapeutic in terms of a partial or complete cure for a disease and/or adverse affect attributable to the disease. “Treatment,” as used herein, covers any treatment of a disease in a subject, particularly in a human, and includes: (a) preventing the disease from occurring in a subject which may be predisposed to the disease but has not yet been diagnosed as having it; (b) inhibiting the disease, i.e., arresting its development; and (c) relieving the disease, e.g., causing regression of the disease, e.g., to completely or partially remove symptoms of the disease.

As used herein, the term “antibody” is used in reference to any immunoglobulin molecule that reacts with a specific antigen. It is intended that the term encompass any immunoglobulin (e.g., IgG, IgM, IgA, IgE, IgD, etc.) obtained from any source (e.g., humans, rodents, non-human primates, caprines, bovines, equines, ovines, etc.). Specific types/examples of antibodies include polyclonal, monoclonal, humanized, chimeric, human, or otherwise-human-suitable antibodies. “Antibodies” also includes any fragment or derivative of any of the herein described antibodies. In specific embodiments, antibodies may be raised against ALK and used as ALK inhibitors.

The terms “specifically binds to,” “specific for,” and related grammatical variants refer to that binding which occurs between such paired species as antibody/antigen, enzyme/substrate, receptor/agonist, and lectin/carbohydrate which may be mediated by covalent or non-covalent interactions or a combination of covalent and non-covalent

interactions. When the interaction of the two species produces a non-covalently bound complex, the binding which occurs is typically electrostatic, hydrogen-bonding, or the result of lipophilic interactions. Accordingly, "specific binding" occurs between a paired species where there is interaction between the two which produces a bound complex having the characteristics of an antibody/antigen or enzyme/substrate interaction. In particular, the specific binding is characterized by the binding of one member of a pair to a particular species and to no other species within the family of compounds to which the corresponding member of the binding member belongs. Thus, for example, an antibody typically binds to a single epitope and to no other epitope within the family of proteins. In some embodiments, specific binding between an antigen and an antibody will have a binding affinity of at least 10^{-6} M. In other embodiments, the antigen and antibody will bind with affinities of at least 10^{-7} M, 10^{-8} M to 10^{-9} M, 10^{-10} M, 10^{-11} M, or 10^{-12} M.

Optional" or "optionally" means that the subsequently described event or circumstance can or cannot occur, and that the description includes instances where the event or circumstance occurs and instances where it does not.

As used herein, a "subject" or "patient" means an individual and can include domesticated animals, (e.g., cats, dogs, etc.); livestock (e.g., cattle, horses, pigs, sheep, goats, etc.), laboratory animals (e.g., mouse, rabbit, rat, guinea pig, etc.) and birds. In one aspect, the subject is a mammal such as a primate or a human. In particular, the term also includes mammals diagnosed with cancer.

The terms "sample," "patient sample," "biological sample," and the like, encompass a variety of sample types obtained from a patient, individual, or subject and can be used in a diagnostic or monitoring assay. The patient sample may be obtained from a healthy subject, a diseased patient or a patient having associated symptoms of cancer (e.g., anaplastic thyroid cancer). Moreover, a sample obtained from a patient can be divided and only a portion may be used for diagnosis. Further, the sample, or a portion thereof, can be stored under conditions to maintain sample for later analysis. The definition specifically encompasses blood and other liquid samples of biological origin (including, but not limited to, peripheral blood, serum, plasma, cerebrospinal fluid, urine, saliva, stool and synovial fluid), solid tissue samples such as a biopsy specimen or tissue cultures or cells derived therefrom and the progeny thereof. In a specific embodiment, a sample comprises a blood sample. In another embodiment, a sample comprises a plasma sample. In yet another embodiment, a serum sample is used.

The definition of "sample" also includes samples that have been manipulated in any way after their procurement, such as by centrifugation, filtration, precipitation, dialysis, chromatography, treatment with reagents, washed, or enriched for certain cell populations. The terms further encompass a clinical sample, and also include cells in culture, cell
5 supernatants, tissue samples, organs, and the like. Samples may also comprise fresh-frozen and/or formalin-fixed, paraffin-embedded tissue blocks, such as blocks prepared from clinical or pathological biopsies, prepared for pathological analysis or study by immunohistochemistry.

The term "inhibitor" includes any type of molecule or agent that directly or indirectly
10 inhibits the expression or activity of a target gene or protein. An inhibitor can be any type of compound, such as a small molecule, antibody or antisense compound. In certain embodiments, the target gene or protein is ALK. In other embodiments, the target gene or protein is a member of the PI3K/Akt pathway. The PI3K/Akt pathway is a signaling pathway involved in a number of cellular processes, such as cell growth, proliferation, differentiation,
15 motility, survival, intracellular trafficking, metabolism and angiogenesis. In the context of the present disclosure, members of the PI3K/Akt pathway include, but are not limited to, PIK3CA, PIK3D, PIK3B, Ras, and PTEN. Inhibitors of the PI3K/Akt pathway include, for example, Akt inhibitors, such as perifosine and triciribine; mTOR inhibitors, such as temsirolimus, everolimus; receptor tyrosine kinase inhibitors, e.g., motesanib, axitinib,
20 sunitinib; and inhibitors of other signaling pathways, such as the NF-kappa pathway.

In some embodiments, the target gene or protein is a member of the MAP kinase pathway. Members of the MAP kinase pathway include, for example, Grb-2, mSOS, Rsk1, BRAF, RET/PTC, Ras, MEK, and ERK. Inhibitors of the MAP kinase pathways include, for example, MEK inhibitors, such as CI-1040, PD0325901, AZD6244, RDEA119, RDEA436;
25 Raf inhibitors, such as PLX4720, BAY 43-9006 (sorafenib). When the two pathways are to be inhibited, two distinct inhibitors of two distinct proteins can be used. Multiple inhibitors of each pathway may also be used, if desired. Certain inhibitors such as Ras inhibitors may act on both pathways and these may also be used in the invention.

II. Anaplastic Lymphoma Kinase Inhibitors

In particular embodiments, the present invention utilizes ALK inhibitors. ALK
30 inhibitors are known in the art and include PF-2341066 or Crizotinib and related analogs. *See* WO2006021886 and WO2004076412 (Pfizer), WO2007130468 (Cephalon), WO2009154769 (Xcovery), WO201056311 and WO2010068292 (Ariad), and WO201059771 (OSI Pharmaceuticals). In other embodiments, ALK inhibitors may comprise

the 2,4-diaminopyrimidine pharmacophore kinase inhibitor motif. *See* WO2004080980 (e.g., NVP-TAE226 and NVP-TAE 684), WO2005026130, WO2005016894, WO2005026158, WO2006021357, WO2008073687, WO209158431, WO2009032668, and WO 201002655 (Novartis). *See also* WO2008051547 (Cephalon) and WO2009143389 (Ariad). Other ALK
5 inhibitors are based on bicyclic diaminopyrimidine scaffolds. *See* WO2009032703, WO2009126514, and WO2009126515 (Novartis); WO2009020990 and WO2010045451 (e.g., GSK1838705A, GSK), WO2009008371 (Astellas), WO2009143389 (Ariad), WO2009132202 (Incyte).

A pyridone-based scaffold is also useful for ALK inhibition. *See* WO2008021369
10 and WO2009117097 (Chembridge/St. Jude's Research Hospital), and Bristol-Myers Squib (BMS-536294 and BMS-695735). ALK inhibitors may also comprise triazine pyrazole compounds. *See* WO2009143389 (AstraZeneca) and WO2009015254 (Bristol-Myers Squib). In other embodiments, pyrazoloisoquinoline inhibitors may be used. *See* WO2005009389 (Exelixis).

15 ALK Inhibitors may also include amidothiazoles and amidoimidazoles (WO2005097765 (Exelixis) and WO2009121535 (Universita Degli Studi di Milano, Universite de Geneve, and Universite Claude Bernard Lyon)); α -carboline ALK inhibitors (WO2010025872 (Universita Degli Studi di Milano, Universite de Geneve, and Universite Claude Bernard Lyon); and amidoindazole ALK inhibitors (WO2008074749, WO200913126,
20 WO2009138440 and WO201069966 (Nerviano Medical Sciences)).

III. Antibodies to ALK

The present invention contemplates the use of antibodies specific for ALK in the treatment of cancer including ATC. Such antibodies can inhibit ALK kinase activity or inhibits its ability to bind to fusion proteins. *See* WO2007124610 and WO2008131575
25 (Esbatech), WO2007059300 (Medimmune), U.S. Patent Publication No. 20110159008 (Delenex Therapeutics AG).

The phrases "binding specificity," "binding specifically to," "specific binding" or otherwise any reference to an antibody to ALK, refers to a binding reaction that is determinative of the presence of the corresponding ALK antigen to the antibody in a
30 heterogeneous population of antigens and other biologics. The parameters required to achieve such specificity can be determined routinely, using conventional methods in the art including, but not limited to, competitive binding studies. The binding affinity of an antibody can also be readily determined, for example, by Scatchard analysis (Scatchard, Ann. NY Acad. Sci. 51: 660-672, 1949). In some embodiments, the immunoglobulins of the present

invention bind to ALK at least about 5, at least about 10, at least about 100, at least about 10^3 , at least about 10^4 , at least 10^5 , and at least 10^6 fold higher than to other proteins.

Various procedures known in the art may be used for the production of antibodies to ALK or any subunit thereof, or a fragment, derivative, homolog or analog of the protein.

5 Antibodies of the present invention include, but are not limited to, synthetic antibodies, polyclonal antibodies, monoclonal antibodies, recombinantly produced antibodies, intrabodies, multispecific antibodies (including bi-specific antibodies), human antibodies, humanized antibodies, chimeric antibodies, synthetic antibodies, single-chain Fvs (scFv) (including bi-specific scFvs), single chain antibodies Fab fragments, F(ab') fragments,
10 disulfide-linked Fvs (sdFv), and anti-idiotypic (anti-Id) antibodies, and epitope-binding fragments of any of the above. In particular, antibodies of the present invention include immunoglobulin molecules and immunologically active portions of immunoglobulin molecules, e.g., molecules that contain an antigen binding site that immunospecifically binds to an antigen (e.g., one or more complementarity determining regions (CDRs) of an
15 antibody).

Another embodiment for the preparation of antibodies according to the invention is the use of peptide mimetics. Mimetics are peptide-containing molecules that mimic elements of protein secondary structure. See, for example, Johnson et al., "Peptide Turn Mimetics" in BIOTECHNOLOGY AND PHARMACY, Pezzuto et al., Eds., Chapman and Hall, New
20 York (1993). The underlying rationale behind the use of peptide mimetics in rational design is that the peptide backbone of proteins exists chiefly to orient amino acid side chains in such a way as to facilitate molecular interactions, such as those of antibody and antigen. A peptide mimetic is expected to permit molecular interactions similar to the natural molecule. These principles may be used to engineer second generation molecules having many of the natural
25 properties of the targeting antibodies disclosed herein, but with altered and even improved characteristics. More specifically, under this rational design approach, peptide mapping may be used to determine "active" antigen recognition residues, and along with molecular modeling and molecular dynamics trajectory analysis, peptide mimic of the antibodies containing antigen contact residues from multiple CDRs may be prepared.

30 In some embodiments, an antibody specifically binds an epitope of the ALK protein. It is to be understood that the peptide regions may not necessarily precisely map one epitope, but may also contain an ALK sequence that is not immunogenic. Methods of predicting other potential epitopes to which an immunoglobulin of the invention can bind are well-known to those of skill in the art and include, without limitation, Kyte-Doolittle Analysis (Kyte, J. and

Dolittle, R. F., 157 J. MOL. BIOL. 105-32 (1982)); Hopp and Woods Analysis (Hopp, T. P. and Woods, K. R., 78 PROC. NATL. ACAD. SCI. USA 3824-28 (1981); Hopp, T. J. and Woods, K. R., 20 MOL. IMMUNOL. 483-89 (1983); Hopp, T. J., 88 J. IMMUNOL. METHODS 1-18 (1986)); Jameson-Wolf Analysis (Jameson, B. A. and Wolf, H., 4 COMPUT. APPL. BIOSCI. 181-86 (1988)); and Emini Analysis (Emeni et al., 140 VIROLOGY 13-20 (1985)).

Amino acid sequence variants of the antibodies of the present invention may be prepared by introducing appropriate nucleotide changes into the polynucleotide that encodes the antibody or by peptide synthesis. Such modifications include, for example, deletions from, and/or insertions into and/or substitutions of, residues within the amino acid sequences of the antibody. Any combination of deletions, insertions, and substitutions may be made to arrive at the final construct.

Amino acid sequence insertions include amino-terminal and/or carboxyl-terminal fusions ranging in length from one residue to polypeptides containing a hundred or more residues, as well as intrasequence insertions of single or multiple amino acid residues.

Examples of terminal insertions include an antibody with an N-terminal methionyl residue or the antibody fused to a cytotoxic polypeptide. Other insertional variants of the antibody molecule include the fusion to the N- or C-terminus of the antibody of a polypeptide that increases the serum half-life of the antibody.

Another type of antibody variant is an amino acid substitution variant. These variants have at least one amino acid residue in the antibody molecule replaced by a different residue. For example, the sites of greatest interest for substitutional mutagenesis of antibodies include the hypervariable regions, but framework region (FR) alterations are also contemplated.

A useful method for the identification of certain residues or regions of the ALK antibodies that are preferred locations for substitution, i.e., mutagenesis, is alanine scanning mutagenesis. See Cunningham & Wells, 244 SCIENCE 1081-85 (1989). Briefly, a residue or group of target residues are identified (e.g., charged residues such as arg, asp, his, lys, and glu) and replaced by a neutral or negatively charged amino acid (most preferably alanine or polyalanine) to affect the interaction of the amino acids with antigen. The amino acid locations demonstrating functional sensitivity to the substitutions are refined by introducing further or other variants at, or for, the sites of substitution. Thus, while the site for introducing an amino acid sequence variation is predetermined, the nature of the mutation per se need not be predetermined. For example, to analyze the performance of a mutation at a given site, alanine scanning or random mutagenesis may be conducted at the target codon or region and the expressed antibody variants screened for the desired activity.

Substantial modifications in the biological properties of the antibody can be accomplished by selecting substitutions that differ significantly in their effect on, maintaining (i) the structure of the polypeptide backbone in the area of the substitution, for example, as a sheet or helical conformation, (ii) the charge or hydrophobicity of the molecule at the target site, or (iii) the bulk of the side chain. Naturally occurring residues are divided into groups based on common side-chain properties:

- (1) hydrophobic: norleucine, met, ala, val, leu, ile;
- (2) neutral hydrophilic: cys, ser, thr;
- (3) acidic: asp, glu;
- (4) basic: asn, gln, his, lys, arg;
- (5) residues that influence chain orientation: gly, pro; and
- (6) aromatic: trp, tyr, phe.

Non-conservative substitutions will entail exchanging a member of one of these classes for another class. Conservative substitutions involve exchanging of amino acids within the same class.

Any cysteine residue not involved in maintaining the proper conformation of the antibody also may be substituted, generally with serine, to improve the oxidative stability of the molecule and prevent aberrant crosslinking. Conversely, cysteine bond(s) may be added to the antibody to improve its stability, particularly where the antibody is an immunoglobulin fragment such as an Fv fragment.

Another type of substitutional variant involves substituting one or more hypervariable region residues of a parent antibody. Generally, the resulting variant(s), i.e., functional equivalents as defined above, selected for further development will have improved biological properties relative to the parent antibody from which they are generated. A convenient way for generating such substitutional variants is by affinity maturation using phage display. Briefly, several hypervariable region sites (e.g., 6-7 sites) are mutated to generate all possible amino substitutions at each site. The antibody variants thus generated are displayed in a monovalent fashion from filamentous phage particles as fusions to the gene III product of M13 packaged within each particle. The phage-displayed variants are then screened for their biological activity (e.g., binding affinity) as herein disclosed.

In order to identify candidate hypervariable region sites for modification, alanine-scanning mutagenesis may be performed to identify hypervariable region residues contributing significantly to antigen binding. Alternatively, or additionally, it may be beneficial to analyze a crystal structure of the antibody-antigen complex to identify contact

points between the antibody and antigen. Such contact residues and neighboring residues are candidates for substitution according to the techniques elaborated herein. Once generated, the panel of variants is subjected to screening as described herein and antibodies with superior properties in one or more relevant assays may be selected for further development.

5 It may be desirable to modify the antibodies of the present invention, i.e., create functional equivalents, with respect to effector function, e.g., so as to enhance antigen-dependent cell-mediated cytotoxicity (ADCC) and/or complement dependent cytotoxicity (CDC) of the antibody. This may be achieved by introducing one or more amino acid substitutions in an Fc region of an antibody. Alternatively or additionally, cysteine residue(s)
10 may be introduced in the Fc region, thereby allowing interchain disulfide bond formation in this region. The homodimeric antibody thus generated may have improved internalization capability and/or increased complement-mediated cell killing and antibody-dependent cellular cytotoxicity (ADCC). Caron et al., 176 J. EXP MED. 1191-95 (1992); Shopes, 148 J. IMMUNOL. 2918-22 (1992). Homodimeric antibodies with enhanced anti-tumor activity may
15 also be prepared using heterobifunctional cross-linkers as described in Wolff et al., 53 CANCER RESEARCH 2560-65 (1993). Alternatively, an antibody can be engineered which has dual Fc regions and may thereby have enhanced complement lysis and ADCC capabilities. Stevenson et al., 3 ANTI-CANCER DRUG DESIGN 219-30 (1989).

To increase the serum half life of an antibody, one may incorporate a salvage receptor
20 binding epitope into the antibody (especially an immunoglobulin fragment) as described in, for example, U.S. Pat. No. 5,739,277. As used herein, the term "salvage receptor binding epitope" refers to an epitope of the Fc region of an IgG molecule (e.g., IgG1, IgG2, IgG3, or IgG4) that is responsible for increasing the in vivo serum half-life of the IgG molecule.

Polynucleotide molecules encoding amino acid sequence variants of the antibody are
25 prepared by a variety of methods known in the art. These methods include, but are not limited to, isolation from a natural source (in the case of naturally occurring amino acid sequence variants) or preparation by oligonucleotide-mediated (or site directed) mutagenesis, PCR mutagenesis, and cassette mutagenesis of an earlier prepared variant or a non-variant version of the anti-ALK antibodies of the present invention.

30 IV. Other ALK Inhibitors

A. RNA Interference Compositions for Targeting ALK mRNAs

In one aspect of the present invention, the expression of ALK may be inhibited by the use of RNA interference techniques (RNAi). RNAi is a remarkably efficient process whereby double-stranded RNA (dsRNA) induces the sequence-specific degradation of

homologous mRNA in animals and plant cells. *See* Hutvagner and Zamore, 12 CURR. OPIN. GENET. DEV. 225-32 (2002); Hammond et al., 2 NATURE REV. GEN. 110-19 (2001); Sharp, 15 GENES DEV. 485-90 (2001). RNAi can be triggered, for example, by nucleotide (nt) duplexes of small interfering RNA (siRNA) (Chiu et al., 10 MOL. CELL. 549-61 (2002); Elbashir et al., 5 411 Nature 494-98 (2001)), micro-RNAs (miRNA), functional small-hairpin RNA (shRNA), or other dsRNAs which are expressed in-vivo using DNA templates with RNA polymerase III promoters. *See, e.g.,* Zeng et al., 9 MOL. CELL. 1327-33 (2002); Paddison et al., 16 GENES DEV. 948-58 (2002); Lee et al., 20 NATURE BIOTECHNOL. 500-05 (2002); Paul et al., 20 NATURE BIOTECHNOL. 505-08 (2002); Tuschl, 20 NATURE BIOTECHNOL. 440-48 (2002); Yu 10 et al., 99(9) PROC. NATL. ACAD. SCI. USA, 6047-52 (2002); McManus et al., 8 RNA 842-50 (2002); Sui et al., 99(6) PROC. NATL. ACAD. SCI. USA 5515-20 (2002).

1. Small Interfering RNA

In particular embodiments, the present invention features “small interfering RNA molecules” (“siRNA molecules” or “siRNA”), methods of making siRNA molecules and 15 methods for using siRNA molecules (e.g., research and/or therapeutic methods). The siRNAs of this invention encompass any siRNAs that can modulate the selective degradation of ALK mRNA.

In a specific embodiment, the siRNA of the present invention may comprise double-stranded small interfering RNA molecules (ds-siRNA). A ds-siRNA molecule of the present 20 invention may be a duplex made up of a sense strand and a complementary antisense strand, the antisense strand being sufficiently complementary to a target ALK mRNA to mediate RNAi. The siRNA molecule may comprise about 10 to about 50 or more nucleotides. More specifically, the siRNA molecule may comprise about 16 to about 30, e.g., 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, or 30 nucleotides in each strand. The strands may be 25 aligned such that there are at least 1, 2, or 3 bases at the end of the strands which do not align (e.g., for which no complementary bases occur in the opposing strand) such that an overhang of 1, 2 or 3 residues occurs at one or both ends of the duplex when strands are annealed.

In an alternative embodiment, the siRNA of the present invention may comprise single-stranded small interfering RNA molecules (ss-siRNA). Similar to the ds-siRNA 30 molecules, the ss-siRNA molecule may comprise about 10 to about 50 or more nucleotides. More specifically, the ss-siRNA molecule may comprise about 15 to about 45 or more nucleotides. Alternatively, the ss-siRNA molecule may comprise about 19 to about 40 nucleotides. The ss-siRNA molecules of the present invention comprise a sequence that is “sufficiently complementary” to a target mRNA sequence to direct target-specific RNA

interference (RNAi), as defined herein, e.g., the ss-siRNA has a sequence sufficient to trigger the destruction of the target mRNA by the RNAi machinery or process. In one embodiment, the ss-siRNA molecule can be designed such that every residue is complementary to a residue in the target molecule. Alternatively, substitutions can be made within the molecule to
5 increase stability and/or enhance processing activity of the molecule. Substitutions can be made within the strand or can be made to residues at the ends of the strand. In a specific embodiment, the 5'-terminus may be phosphorylated (e.g., comprises a phosphate, diphosphate, or triphosphate group). In another embodiment, the 3' end of an siRNA may be a hydroxyl group in order to facilitate RNAi, as there is no requirement for a 3' hydroxyl
10 group when the active agent is a ss-siRNA molecule. In other instances, the 3' end (e.g., C3 of the 3' sugar) of ss-siRNA molecule may lack a hydroxyl group (e.g., ss-siRNA molecules lacking a 3' hydroxyl or C3 hydroxyl on the 3' sugar (e.g., ribose or deoxyribose)).

In another aspect, the siRNA molecules of the present invention may be modified to improve stability under in vitro and/or in vivo conditions, including, for example, in serum
15 and in growth medium for cell cultures. In order to enhance the stability, the 3'-residues may be stabilized against degradation, e.g., they may be selected such that they consist of purine nucleotides, particularly adenosine or guanosine nucleotides. Alternatively, substitution of pyrimidine nucleotides by modified analogues, e.g., substitution of uridine by 2'-deoxythymidine is tolerated and does not affect the efficiency of RNA interference. For
20 example, the absence of a 2' hydroxyl may significantly enhance the nuclease resistance of the siRNAs in tissue culture medium.

Furthermore, the siRNAs of the present invention may include modifications to the sugar-phosphate backbone or nucleosides. These modifications can be tailored to promote selective genetic inhibition, while avoiding a general panic response reported to be generated
25 by siRNA in some cells. In addition, modifications can be introduced in the bases to protect siRNAs from the action of one or more endogenous enzymes.

In an embodiment of the present invention, the siRNA molecule may contain at least one modified nucleotide analogue. The nucleotide analogues may be located at positions where the target-specific activity, e.g., the RNAi mediating activity is not substantially
30 effected, e.g., in a region at the 5'-end and/or the 3'-end of the RNA molecule. Particularly, the ends may be stabilized by incorporating modified nucleotide analogues. Examples of nucleotide analogues include sugar- and/or backbone-modified ribonucleotides (e.g., include modifications to the phosphate-sugar backbone). For example, the phosphodiester linkages of natural RNA may be modified to include at least one of a nitrogen or sulfur heteroatom. In

backbone-modified ribonucleotides, the phosphoester group connecting to adjacent ribonucleotides may be replaced by a modified group, e.g., a phosphothioate group. In sugar-modified ribonucleotides, the 2' OH-group may be replaced by a group selected from H, OR, R, halo, SH, SR, NH₂, NHR, NR₂ or ON, wherein R is C₁-C₆ alkyl, alkenyl or alkynyl and halo is F, Cl, Br or I.

Nucleobase-modified ribonucleotides may also be utilized, e.g., ribonucleotides containing at least one non-naturally occurring nucleobase instead of a naturally occurring nucleobase. Bases may be modified to block the activity of adenosine deaminase. Exemplary modified nucleobases include, but are not limited to, uridine and/or cytidine modified at the 5-position, e.g., 5-(2-amino)propyl uridine, 5-bromo uridine; adenosine and/or guanosines modified at the 8 position, e.g., 8-bromo guanosine; deaza nucleotides, e.g., 7-deaza-adenosine; O- and N-alkylated nucleotides, e.g., N6-methyl adenosine are suitable. It should be noted that the above modifications may be combined.

Derivatives of siRNAs may also be utilized herein. For example, cross-linking can be employed to alter the pharmacokinetics of the composition, e.g., to increase half-life in the body. Thus, the present invention includes siRNA derivatives that include siRNA having two complementary strands of nucleic acid, such that the two strands are crosslinked. The present invention also includes siRNA derivatives having a non-nucleic acid moiety conjugated to its 3' terminus (e.g., a peptide), organic compositions (e.g., a dye), or the like. Modifying siRNA derivatives in this way may improve cellular uptake or enhance cellular targeting activities of the resulting siRNA derivative as compared to the corresponding siRNA, are useful for tracing the siRNA derivative in the cell, or improve the stability of the siRNA derivative compared to the corresponding siRNA.

The siRNAs of the present invention can be enzymatically produced or totally or partially synthesized. Moreover, the siRNAs can be synthesized in vivo or in vitro. For siRNAs that are biologically synthesized, an endogenous or a cloned exogenous RNA polymerase may be used for transcription in vivo, and a cloned RNA polymerase can be used in vitro. siRNAs that are chemically or enzymatically synthesized are preferably purified prior to the introduction into the cell.

Although one hundred percent (100%) sequence identity between the siRNA and the target region is preferred in particular embodiments, it is not required to practice the invention. siRNA molecules that contain some degree of modification in the sequence can also be adequately used for the purpose of this invention. Such modifications may include,

but are not limited to, mutations, deletions or insertions, whether spontaneously occurring or intentionally introduced.

Moreover, not all positions of a siRNA contribute equally to target recognition. In certain embodiments, for example, mismatches in the center of the siRNA may be critical and could essentially abolish target RNA cleavage. In other embodiments, the 3' nucleotides of the siRNA do not contribute significantly to specificity of the target recognition. In particular, residues 3' of the siRNA sequence which is complementary to the target RNA (e.g., the guide sequence) may not critical for target RNA cleavage.

Sequence identity may be determined by sequence comparison and alignment algorithms known to those of ordinary skill in the art. To determine the percent identity of two nucleic acid sequences (or of two amino acid sequences), the sequences are aligned for optimal comparison purposes (e.g., gaps can be introduced in the first sequence or second sequence for optimal alignment). The nucleotides (or amino acid residues) at corresponding nucleotide (or amino acid) positions are then compared. When a position in the first sequence is occupied by the same residue as the corresponding position in the second sequence, then the molecules are identical at that position. The percent identity between the two sequences is a function of the number of identical positions shared by the sequences (e.g., % homology = # of identical positions/total # of positions x 100), optionally penalizing the score for the number of gaps introduced and/or length of gaps introduced.

The comparison of sequences and determination of percent identity between two sequences can be accomplished using a mathematical algorithm. In one embodiment, the alignment generated over a certain portion of the sequence aligned having sufficient identity but not over portions having low degree of identity (e.g., a local alignment). A non-limiting example of a local alignment algorithm utilized for the comparison of sequences is the algorithm of Karlin and Altschul, 87 PROC. NATL. ACAD. SCI. USA 2264-68 (1990), and as modified as in Karlin and Altschul 90 PROC. NATL. ACAD. SCI. USA 5873-77 (1993). Such an algorithm is incorporated into the BLAST programs (version 2.0) of Altschul, et al., 215 J. MOL. BIOL. 403-10 (1990).

In another embodiment, the alignment may optimized by introducing appropriate gaps and determining percent identity over the length of the aligned sequences (e.g., a gapped alignment). To obtain gapped alignments for comparison purposes, Gapped BLAST can be utilized as described in Altschul et al., 25(17) NUCLEIC ACIDS RES. 3389-3402 (1997). In another embodiment, the alignment may be optimized by introducing appropriate gaps and determining percent identity over the entire length of the sequences aligned (e.g., a global

alignment). A non-limiting example of a mathematical algorithm utilized for the global comparison of sequences is the algorithm of Myers and Miller, CABIOS (1989). Such an algorithm is incorporated into the ALIGN program (version 2.0) which is part of the GCG sequence alignment software package. When utilizing the ALIGN program for comparing amino acid sequences, a PAM120 weight residue table, a gap length penalty of 12, and a gap penalty of 4 can be used.

In particular embodiments, greater than 90% sequence identity, e.g., 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or even 100% sequence identity, between the siRNA and the portion of the target gene may be used. Alternatively, the siRNA may be defined functionally as a nucleotide sequence (or oligonucleotide sequence) that is capable of hybridizing with a portion of the target gene transcript (e.g., 400 mM NaCl, 40 mM PIPES pH 6.4, 1 mM EDTA, 50°C or 70°C hybridization for 12-16 hours; followed by washing). Additional hybridization conditions include, but are not limited to, hybridization at 70°C in 1xSSC or 50°C in 1xSSC, 50% formamide followed by washing at 70°C in 0.3xSSC or hybridization at 70°C in 4xSSC or 50°C in 4xSSC, 50% formamide followed by washing at 67°C in 1xSSC. The hybridization temperature for hybrids anticipated to be less than 50 base pairs in length can be about 5-10°C less than the melting temperature (T_m) of the hybrid, where T_m is determined according to the following equations. For hybrids less than 18 base pairs in length, $T_m(^{\circ}\text{C}) = 2(\# \text{ of A+T bases}) + 4(\# \text{ of G+C bases})$. For hybrids between 18 and 49 base pairs in length, $T_m(^{\circ}\text{C}) = 81.5 + 16.6(\log_{10}[\text{Na}^+]) + 0.41(\% \text{ G+C}) - (600/N)$, where N is the number of bases in the hybrid, and $[\text{Na}^+]$ is the concentration of sodium ions in the hybridization buffer ($[\text{Na}^+]$ for 1xSSC=0.165 M). Additional examples of stringency conditions for polynucleotide hybridization are provided in Sambrook, J., E. F. Fritsch, and T. Maniatis, 1989, *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., chapters 9 and 11, and *Current Protocols in Molecular Biology*, 1995, F. M. Ausubel et al., eds., John Wiley & Sons, Inc., sections 2.10 and 6.3-6.4, incorporated herein by reference. The length of the identical nucleotide sequences may be at least about 10, 12, 15, 17, 20, 22, 25, 27, 30, 32, 35, 37, 40, 42, 45, 47 50 or more bases.

2. Other Compositions for Targeting ALK DNA or mRNA

Antisense molecules can act in various stages of transcription, splicing and translation to block the expression of a target gene. Without being limited by theory, antisense molecules can inhibit the expression of a target gene by inhibiting transcription initiation by forming a triple strand, inhibiting transcription initiation by forming a hybrid at an RNA polymerase binding site, impeding transcription by hybridizing with an RNA molecule being

synthesized, repressing splicing by hybridizing at the junction of an exon and an intron or at the spliceosome formation site, blocking the translocation of an mRNA from nucleus to cytoplasm by hybridization, repressing translation by hybridizing at the translation initiation factor binding site or ribosome binding site, inhibiting peptide chain elongation by hybridizing with the coding region or polysome binding site of an mRNA, or repressing gene expression by hybridizing at the sites of interaction between nucleic acids and proteins. An example of an antisense oligonucleotide of the present invention is a cDNA that, when introduced into a cell, transcribes into an RNA molecule having a sequence complementary to at least part of the ALK mRNA.

Furthermore, antisense oligonucleotides of the present invention include oligonucleotides having modified sugar-phosphodiester backbones or other sugar linkages, which can provide stability against endonuclease attacks. The present invention also encompasses antisense oligonucleotides that are covalently attached to an organic or other moiety that increase their affinity for a target nucleic acid sequence. For example, intercalating agents, alkylating agents, and metal complexes can be also attached to the antisense oligonucleotides of the present invention to modify their binding specificities.

The present invention also provides ribozymes as a tool to inhibit ALK expression. Ribozymes are enzymatic RNA molecules capable of catalyzing the specific cleavage of RNA. The characteristics of ribozymes are well-known in the art. *See, e.g.,* Rossi, 4 CURRENT BIOLOGY 469-71 (1994). Without being limited by theory, the mechanism of ribozyme action involves sequence specific hybridization of the ribozyme molecule to complementary target RNA, followed by an endonucleolytic cleavage. In particular embodiments, the ribozyme molecules include one or more sequences complementary to the target gene mRNA, and include the well known catalytic sequence responsible for mRNA cleavage. *See* U.S. Patent No. 5,093,246. Using the known sequence of the target ALK mRNA, a restriction enzyme-like ribozyme can be prepared using standard techniques.

The expression of the ALK gene can also be inhibited by using triple helix formation. Nucleic acid molecules to be used in triple helix formation for the inhibition of transcription can be single stranded and composed of deoxynucleotides. The base composition of these oligonucleotides must be designed to promote triple helix formation via Hoogsteen base pairing rules, which generally require sizeable stretches of either purines or pyrimidines to be present on one strand of a duplex. Nucleotide sequences may be pyrimidine-based, which will result in TAT and CGC⁺ triplets across the three associated strands of the resulting triple helix. The pyrimidine-rich molecules provide base complementarity to a purine-rich region of

a single strand of the duplex in a parallel orientation to that strand. In addition, nucleic acid molecules that are purine-rich, e.g., containing a stretch of G residues, may be chosen. These molecules will form a triple helix with a DNA duplex that is rich in GC pairs, in which the majority of the purine residues are located on a single strand of the targeted duplex, resulting in GGC triplets across the three strands in the triplex.

Alternatively, the potential sequences that can be targeted for triple helix formation may be increased by creating a so-called "switchback" nucleic acid molecule. Switchback molecules are synthesized in an alternating 5'-3', 3'-5' manner, such that they base pair first with one strand of a duplex and then the other, eliminating the necessity for a sizeable stretch of either purines or pyrimidines to be present on one strand of a duplex.

It would be readily apparent to one of ordinary skill in the art that other methods of gene expression inhibition that selectively target ALK DNA or mRNA can also be used in connection with this invention without departing from the spirit of the invention. In a specific embodiment, using techniques known to those of ordinary skill in the art, the present invention contemplates affecting the promoter region of ALK to effectively switch off transcription.

3. Design and Production of the RNAi Compositions

One or more of the following guidelines may be used in designing the sequence of siRNA and other nucleic acids designed to bind to a target mRNA, e.g., shRNA, stRNA, antisense oligonucleotides, ribozymes, and the like, that are advantageously used in accordance with the present invention.

Beginning with the AUG start codon of the ALK gene, each AA dinucleotide sequence and the 3' adjacent 16 or more nucleotides are potential siRNA targets. In a specific embodiment, the siRNA is specific for a target region that differs by at least one base pair between the wild type and mutant allele or between splice variants. In dsRNAi, the first strand is complementary to this sequence, and the other strand identical or substantially identical to the first strand. siRNAs with lower G/C content (35-55%) may be more active than those with G/C content higher than 55%. Thus in one embodiment, the invention includes nucleic acid molecules having 35-55% G/C content. In addition, the strands of the siRNA can be paired in such a way as to have a 3' overhang of 1 to 4, e.g., 2, nucleotides. Thus in another embodiment, the nucleic acid molecules may have a 3' overhang of 2 nucleotides, such as TT. The overhanging nucleotides may be either RNA or DNA. In one embodiment, it may be desirable to choose a target region wherein the mismatch is a purine:purine mismatch.

Using any method known in the art, compare the potential targets to the appropriate genome database (human, mouse, rat, etc.) and eliminate from consideration any target sequences with significant homology to other coding sequences. One such method for such sequence homology searches is known as BLAST, which is available at National Center for
5 Biotechnology Information website (<http://www.ncbi.nih.gov>). Select one or more sequences that meet the criteria for evaluation.

Another method includes selecting in the sequence of the target mRNA, a region located from about 50 to about 100 nt 3' from the start codon. In this region, search for the following sequences: AA(N₁₉)TT or AA(N₂₁), where N=any nucleotide. The GC content of
10 the selected sequence should be from about 30% to about 70%, preferably about 50%. To maximize the specificity of the RNAi, it may be desirable to use the selected sequence in a search for related sequences in the genome of interest; sequences absent from other genes are preferred. The secondary structure of the target mRNA may be determined or predicted, and it may be preferable to select a region of the mRNA that has little or no secondary structure,
15 but it should be noted that secondary structure seems to have little impact on RNAi. When possible, sequences that bind transcription and/or translation factors should be avoided, as they might competitively inhibit the binding of a siRNA, sbRNA or stRNA (as well as other antisense oligonucleotides) to the mRNA. Further general information about the design and use of siRNA may be found in "The siRNA User Guide," available at The Max-Planck-
20 Institut für Biophysikalische Chemie website (<http://www.mpibpc.mpg.de>).

Negative control siRNAs should have the same nucleotide composition as the selected siRNA, but without significant sequence complementarity to the appropriate genome. Such negative controls may be designed by randomly scrambling the nucleotide sequence of the selected siRNA; a homology search can be performed to ensure that the negative control
25 lacks homology to any other gene in the appropriate genome.

4. Delivery of ALK RNA Targeting Compositions

Delivery of the compositions of the present invention (e.g., siRNAs, antisense oligonucleotides, or other compositions described herein) into a patient can either be direct, e.g., the patient is directly exposed to the compositions of the present invention or compound-
30 carrying vector, or indirect, e.g., cells are first transformed with the compositions of this invention in vitro, then transplanted into the patient for cell replacement therapy. These two approaches are known as in vivo and ex vivo therapy, respectively.

In the case of in vivo therapy, the compositions of the present invention are directly administered in vivo, where they are expressed to produce the encoded product. This can be

accomplished by any of numerous methods known in the art, e.g., by constructing them as part of an appropriate nucleic acid expression vector and administering them so that they become intracellular, by infection using a defective or attenuated retroviral or other viral vector, by direct injection of naked DNA, by coating with lipids or cell-surface receptors or
5 transfecting agents, encapsulation in liposomes, nanoparticles, microparticles, or microcapsules, by administering them in linkage to a peptide which is known to enter the cell or nucleus, or by administering them in linkage to a ligand subject to receptor-mediated endocytosis which can be used to target cell types specifically expressing the receptors. Further, the compositions of the present invention can be targeted in vivo for cell specific
10 uptake and expression, by targeting a specific receptor. *See, e.g.*, WO93/14188, WO 93/20221, WO 92/22635, WO92/20316, and WO 92/06180.

Ex vivo therapy involves transferring the compositions of the present invention to cells in tissue culture by methods well-known in the art such as electroporation, transfection, lipofection, microinjection, calcium phosphate mediated transfection, cell fusion,
15 chromosome-mediated gene transfer, microcell-mediated gene transfer, spheroplast fusion, and infection with a viral vector containing the nucleic acid sequences. These techniques should provide for the stable transfer of the compositions of this invention to the cell, so that they are expressible by the cell and preferably heritable and expressible by its cell progeny. In particular embodiments, the method of transfer includes the transfer of a selectable marker
20 to the cells. The cells are then placed under selection to isolate those cells that have taken up and are expressing the transferred compositions. The resulting recombinant cells can be delivered to a patient by various methods known in the art. Examples of the delivery methods include, but are not limited to, subcutaneous injection, skin graft, and intravenous injection.

25 B. ALK Vaccines

Not only are ALK fusion proteins required for the development of anaplastic large cell lymphoma (ALCL), they are also antigenic: patients with ALCL mount ALK specific T- and B-cell responses. These two characteristics make ALK a unique target for vaccine therapy. Inghirami and co-workers vaccinated BALB/c mice with a DNA plasmid which
30 coded for a large portion of the intracytoplasmic domain of ALK. Mice were then treated with syngeneic ALK positive lymphoma cells. Untreated mice developed lymphoma, while ALK-vaccinated mice did not. In another study, mice were pretreated with a low tumor load of lymphoma cells. Administration of the ALK vaccine provided protection to these mice as well. Combination therapy with chemotherapeutics provided an additional benefit. *See*

Chiarle et al., 14(6) NAT. MED. 676-80 (2009); EP2042191; Mastini et al., 87(7) J. MOL. MED. 669-77 (2009); and WO2004096142.

Without further elaboration, it is believed that one skilled in the art, using the preceding description, can utilize the present invention to the fullest extent. The following
5 examples are illustrative only, and not limiting of the remainder of the disclosure in any way whatsoever.

EXAMPLES

The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how the compounds, compositions, articles,
10 devices, and/or methods described and claimed herein are made and evaluated, and are intended to be purely illustrative and are not intended to limit the scope of what the inventors regard as their invention. Efforts have been made to ensure accuracy with respect to numbers (e.g., amounts, temperature, etc.) but some errors and deviations should be accounted for herein. Unless indicated otherwise, parts are parts by weight, temperature is in degrees
15 Celsius or is at ambient temperature, and pressure is at or near atmospheric. There are numerous variations and combinations of reaction conditions, e.g., component concentrations, desired solvents, solvent mixtures, temperatures, pressures and other reaction ranges and conditions that can be used to optimize the product purity and yield obtained from the described process. Only reasonable and routine experimentation will be required to
20 optimize such process conditions.

Materials and Methods

Tumor Samples, Cell Lines, and DNA Isolation. A large number of human tumor samples, as indicated in the Results below, and human thyroid cancer cell lines K1, BCPAP, K5, FTC133, OCUT-1, OCUT-2, FB-1, SW1736, HTh7, HTh74, KAT18, and C643 (with
25 the later 8 cell lines being derived from ATC), melanoma cell lines M14, A375, and UACC62, and colon carcinoma cell lines T84, RKO, and HT-29 were used for mutational analysis of the ALK gene. The original sources of the cell lines used in this study were as follows: K1 and K5 from Dr. David Wynford-Thomas (University of Wales College of Medicine, Cardiff, UK); BCPAP from Dr. Massimo Santoro (University of Federico II,
30 Naples, Italy); FTC133 from Dr. Georg Brabant (University of Manchester, Manchester, UK); OCUT-1 and OCUT-2 from Dr. Naoyoshi Onoda (Osaka City University Graduate School of Medicine, Osaka, Japan); KAT18 from Dr. Kenneth B. Ain (University of Kentucky Medical Center, Lexington, KY); FB-1 from Dr. Riccardo Giannini (Department of Surgery, University of Pisa, Pisa, Italy); SW1736, HTh7, HTh74, and C643 from Dr. N. E.

Heldin (University of Uppsala, Uppsala, Sweden). These cell lines have recently been tested and authenticated to be distinct thyroid cancer cell lines (17). The melanoma cell lines UACC62 and M14 were obtained from the cell bank of National Cancer Institute (Bethesda, MD), and the melanoma cell line A375 and the colon cancer cell lines T84, RKO, and HT-29 were obtained from American Type Culture Collection (ATCC). Use of tumor samples was Institutional Review Board approved as reported previously (18). Except for FTC133 cells cultured in DMEM/ Ham's F-12 medium, all tumor cell lines were cultured in RPMI-1640 medium, supplemented with 10% FBS, 2 mmol/L L-glutamine, streptomycin (100 mg/mL), and penicillin (100 U/ mL). Genomic DNA (from cell lines and tumors) was isolated by phenol-chloroform extraction, using MaXtract high-density gel tubes (Qiagen) as described previously (19).

PCR Amplification and Sequencing. PCR amplification of exons 23, 24, and 25 of the ALK gene was conducted using the primers and conditions as described previously (10) (ALK ex 23: Forward primer AAGATTTGCCAGACTCAGC (SEQ ID NO:8); Reverse primer TGTCTTGGCACAACAACACTG (SEQ ID NO:9)). The amplified PCR products were directly sequenced using a BigDye terminator v3.1 cycle sequencing ready reaction kit (Applied Biosystems) and an ABI PRISM 3730 automated next generation genetic analyzer (Applied Biosystems). GenBank accession number for ALK is NM_004304.3 (SEQ ID NO:6).

Multiple Amino Acid Sequence Alignment. Original amino acid sequences of ALK of various species were obtained from NCBI database (<http://www.ncbi.nlm.nih.gov/protein/>) as follows: H_sapiens (NP_004295.2) (SEQ ID NO:7), C_lupus (XP_540136.2), B_taurus (XP_616782.3), M_musculus (NP_031465.2), G_gallus (XP_419364.2), and D_rerio (XP_691964.2). These amino acid sequences were compared using a computer-based multiple sequence alignment program (<http://pir.georgetown.edu/pirwww/search/multialn.shtml>).

Expression Vectors and Site-Directed Mutagenesis. The expression vector pcDNA3 carrying wild-type ALK and mutant ALKs (K1062M and F1174L) are a kind gift from Profs. Yasubide Hayashi and Seishi Ogawa at The University of Tokyo, Tokyo, Japan (10). The expression vector pcDNA3 carrying wild-type ALK was used to generate the novel ALK mutants L1198F and G1201E discovered in the present study with a Quick Change XL II Site-Directed mutagenesis kit (Stratagene) according to the instructions of the manufacturer. The primers were designed using a template-specific mutagenic primer design program. The primer sequences are as follows: for L1198F: sense, ALK-C3592T_F 5'-

CATCCTGCTGGAGTTCATGGCGGGGGG- 3' (SEQ ID NO:1); antisense, ALK-C3-592T_R 5'-CCCCCGCCATGAACTCCAGCAGGATG-3' (SEQ ID NO:2). For G1201E: sense, ALK-G3602A_F 5'-GAGCTCATGGCGGAGGGAGACCTCAAG- 3' (SEQ ID NO:3); and antisense, ALK-G3602A_R 5'-CTTGAGGTCTCCCTCCGCCATGAGCTC- 3' (SEQ ID NO:4). The mutations were confirmed in the vectors by sequencing with the primer ALKVEC__F 5'-TCTCGCTGTGGTGACCTCTG-3' (SEQ ID NO:5). Plasmid DNAs for the transfection experiments were purified using a mini prep kit (catalogue no. K2100-11; Invitrogen).

Cell Culture, Transfection, and Pooled Stable Expression. NIH3T3 cells (ATCC) were grown in DMEM and supplemented with fetal calf serum and plated (8.0×10^5 cells/well) on 6-well plates. Twenty-four hours later, cells were transfected using the Lipofectamine 2000 Transfection Reagent (Invitrogen) with equal amount of empty vector or vector containing wild-type ALK or mutant ALK DNA per manufacturer's instructions. Cells were selected using 800 mg/ mL G418 (Stratagene) 48 hours after transfection. Medium was changed after every 3 days. After 14 days of selection, stably formed clones were pooled and integration of the plasmid was checked by PCR and expression of the ALK protein was confirmed by Western blotting. Stably transfected pooled clones were used for functional studies.

ALK Kinase Assay. The ALK tyrosine kinase assay was a non-radioactive solid phase ELISA, which was conducted as described previously (10) using the Universal Tyrosine Kinase Assay Kit (catalogue no. MK410; Takara Bio Inc.). Briefly, cells stably transfected with empty vector, wild-type ALK, the 2 novel ALK mutants (L1198F and G1201E), and the 2 positive ALK mutant controls (K1062M and F1174L) were lysed and lysates were centrifuged at $12,000 \times g$ for 10 minutes at 4°C. The supernatants were collected, and protein concentration was measured using a DC protein assay kit (Bio-Rad Laboratories). For each enzymatic reaction, an equal volume of cell lysate containing the same amount of protein was added to the kinase reaction buffer and the mixture was transferred to the microplate supplemented with the kit containing the substrate [immobilized peptide poly(Glu-Tyr)]. The kinase reaction was initiated by adding 40 mmol/L ATP-2Na solution, followed by incubation for 20 minutes at room temperature.

Western Blotting. Western blotting was done as described previously (20). Briefly, 15 mg of cell lysate proteins prepared for ALK kinase assay was separated on SDS-PAGE and transferred to polyvinylidene difluoride membrane (Millipore Co.). After transfer, the membrane was blocked with 5% skim-milk/PBS containing 0.1% Tween 20 (PBST) for 1

hour at room temperature and the membrane was sliced on the basis of the molecular weight and incubated with primary antibodies. Membranes were incubated overnight at 4°C with anti-Flag (catalogue no. 2368; Cell Signaling Technology), anti-phospho-Akt (catalogue no. sc-7985; Santa Cruz Biotechnology), or anti-phospho-ERK (catalogue no. sc-7383; Santa Cruz Biotechnology) primary antibodies. Membranes were incubated with anti-Akt (catalogue no. sc-8312), anti-ERK (catalogue no. sc-94), or anti-actin (catalogue no. sc-1616; Santa Cruz Biotechnology) primary antibodies for 1 hour at room temperature. After washing 4 times with PBST, blots were incubated with respective horseradish peroxidase-conjugated secondary antibodies (catalogue nos. sc-2004 and sc-2005; Santa Cruz Biotechnology) for 1 hour at room temperature. After washing with PBST, protein bands on the membrane were detected with enhanced chemiluminescence reaction and exposure to X-ray films.

Cell Focus Formation Assay. Cell focus formation assay was conducted as described previously (21). Briefly, NIH3T3 cells were transfected with equal amount of empty vector, wild-type, or each of mutant ALK expression vectors, using the Lipofectamine 2000 transfection reagent following the manufacturer's instructions (Invitrogen Life Technologies). Twenty-four hours after the transfection, cells were selected using G418 (800 mg/mL) for 7 days. Selected cells were trypsinized, pooled, and plated (5×10^5 cells) on 6-well plates. Medium was changed every 3 to 4 days. After 14 days, the number of morphologically transformed foci was counted and photographed (Zeiss Axiovert 200M).

Soft Agar Colony Formation Assay. Soft agar colony formation assay was conducted as previously described (10). Briefly, NIH3T3 cells stably expressing vector, wild-type, or mutant ALKs were seeded (1.0×10^4 cells) on 6-well plates (Costar; Corning) in 0.3% agar (catalogue no. 214010; BD Biosciences) over a bottom layer of 0.6% agar. After 4 weeks, the colonies of greater than 0.1 mm diameter were counted and photographed (Zeiss Axiovert 200M; Carl Zeiss).

Invasion Assay. Cell invasion assay was conducted as described previously (20). Briefly, the assay was conducted using Matrigel invasion chambers consisting of BD Falcon cell culture inserts containing a polyethylene terephthalate (PET) membrane with 8-mm pores coated with Matrigel matrix (BD BioCoat Matrigel Invasion Chamber; BD Biosciences). Cells expressing the vector, wild-type, or mutant ALK were completely serum starved for 7 to 8 hours and then collected and resuspended (5×10^4 cells) in 500 mL of serum-free DMEM with 0.1% bovine serum albumin. Culture inserts were placed in the wells of a BD Falcon 24-well multiwell companion plate, and 750 mL of DMEM containing 1% serum was

added to the lower compartment of each well. Cell suspensions were added to each culture inserts. After a 22-hour incubation at 37°C with 5% CO₂, the noninvading cells on top of the Matrigel were removed using cotton swab and invaded cells on the lower side of the membrane were fixed with 70% ethanol and stained with Coomassie Brilliant Blue. Invading
5 cells were counted and photographed under a microscope with 10x magnification (Nikon Eclipse ME-600-DS-5M-L1). All the data of assays presented represent at least 2 similar experiments.

Results

Example 1: Identification of Novel Somatic ALK Mutations in ATC. The tyrosine
10 kinase domain exons 23 and 25 of the ALK gene were analyzed for mutation in 12 thyroid cancer cell lines (including 8 ATC cell lines), 36 PTCs (including 12 cases of each conventional, follicular variant, and tall cell PTC), 20 FTCs, and 18 ATC tumor samples. These exons were chosen for analysis because they contained the hot spots for ALK mutations found in neuroblastomas. No mutation was found in the thyroid cancer cell lines
15 as well as in the PTC and FTC tumor samples. However, 2 novel missense heterozygous ALK point mutations were found in ATC tumor samples. As shown in FIG. 1A, a mutation found in a case of ATC represented a C>T transition in nucleotide position 3,592. This mutation changed codon 1,198 from CTC to TTC, resulting in the amino acid change from lysine to phenylalanine (L1198F) of ALK. As shown also in FIG. 1A, the other mutation
20 found in another case of ATC represented a G>A transition in nucleotide position 3,602. This mutation changed codon 1,201 from GGG to GAG, resulting in the amino acid change of glycine to glutamic acid (G1201E) of ALK. Reverse sequencing using antisense primers confirmed these 2 mutations (Fig. 1A). The matched normal tissues in these 2 cases showed the wild-type ALK gene (Fig. 1A), suggesting that the novel ALK mutations identified in the
25 ATC samples were somatic mutations. These mutations were not found in COSMIC Catalog Of Somatic Mutations In Cancer, a database of Sanger Institute, UK (<http://www.sanger.ac.uk/genetics/CGP/cosmic/>), and in the literature. The databases of single-nucleotide polymorphism were also checked, including the major Web sites of Ensembl (<http://uswest.ensembl.org>) and NCBI
30 (<http://www.ncbi.nlm.nih.gov/projects/SNP/>), and did not find germ line variations that represented these ALK mutations. Therefore, this is the first report of these somatic mutations of the ALK gene in human cancers. Exon 24 of the ALK gene was also examined in the ATC samples but no mutations were found. In these ATC tumors, ALK mutations that

had been previously reported in neuroblastomas were not found. The prevalence of ALK mutations in ATC in the present study was 11.11% (2 of 18).

Exons 23 and 25 of the ALK gene were also analyzed in 3 melanoma cell lines, 3 colon carcinoma cell lines, 44 melanomas, and 47 colon carcinoma samples. The ALK mutation was not found in any of these samples, suggesting that ALK mutation is not a common event in these cancers. A few silent mutations were found in these cancers (data not shown).

Example 2: Increased Tyrosine Kinase Activities of Novel Somatic ALK Mutants L1198F and G1201E and Their Activation of the PI3K/Akt and MAP Kinase Pathways. As

shown in FIG. 1B, the somatically mutated amino acid residues L1198F and G1201E are located within the tyrosine kinase domain (amino acids 1,057-1,383) of the ALK protein. Moreover, as shown in FIG. 1C, alignment comparison of amino acid sequences of ALK proteins from 6 different species revealed that the L1198 and G1201 residues were evolutionarily conserved residues of the ALK proteins among various species. These novel somatic ALK mutations likely altered the tyrosine kinase activity of ALK. To test this idea, in vitro mutagenesis was conducted to generate these 2 mutations and tested their tyrosine kinase activities. To this end, NIH3T3 cells stably expressing vector, wild-type, and each mutant ALK were lysed and assayed for in vitro tyrosine kinase activities. As shown in FIG. 2A, the novel ALK mutants L1198F and G1201E displayed dramatically increased tyrosine kinase activities compared with the wildtype ALK. The neuroblastoma-associated ALK mutants K1062M and F1174L, as positive controls in the assay, also displayed high activities as expected (Fig. 2A). Western blot analysis confirmed the corresponding protein expression of the expression vector constructs (Fig. 2B, top). These results showed that the 2 novel ALK mutations found in ATC are gain of- function mutations.

As oncogenic fusion ALK proteins were previously shown to activate the PI3K/Akt and MAP kinase pathways (8, 9), whether the novel ALK mutants discovered in this study had any impact on the signaling of these 2 pathways was tested next. As shown in FIG. 2B, compared with wild-type ALK, phosphorylation of both Akt and ERK was elevated in NIH3T3 cells expressing the mutant L1198F or G1201E, similar to ALK mutants K1062M and F1174L tested here as positive controls. This was consistent with high immunostaining scores for the phosphorylation levels of Akt and ERK in the 2 cases of ATC harboring the ALK mutations, which were revealed in a previous study of ours (19). Thus, the role of the 2 novel ALK mutations in the tumorigenesis of ATC likely involves aberrant activation of the PI3K/Akt and MAP kinase pathways.

Example 3: Novel Somatic ALK Mutants L1198F and G1201E Promoted Cell

Transformation and Invasion. Given the known cell-transforming abilities of fusion ALK proteins and neuroblastoma-associated ALK mutants (10), the transforming abilities of the 2 novel ALK mutants L1198F and G1201E was examined to functionally test their oncogenic potential. To this end, NIH3T3 cells were transfected with empty vector, wild-type ALK, and various ALK mutants and examined their ability to form cell focus and anchorage-independent colonies. As shown in FIG. 3A, cells expressing ALK mutants L1198F and G1201E lost cell contact-mediated growth inhibition and grew foci of multilayers of cells whereas control cells (vector and wild-type ALK) exhibited contact inhibition and grew in monolayer of cells. Moreover, ALK mutants L1198F and G1201E were also associated with a much larger number of foci of multilayers of cells than the wild-type ALK (Fig. 3B). As a positive control, the previously well-characterized neuroblastoma-associated ALK mutants K1062M and F1174L showed similar cell focus-forming abilities (Fig. 3A and B).

Anchorage-independent growth on soft agar also reflects a transformation property of cells. As shown in FIG. 3C, like the positive control ALK mutants K1062M and F1174L, the novel ALK mutants L1198F and G1201E induced the formation of much larger cell colonies on soft agar than that by vector and wild-type ALK. The number of large cell colonies induced by these mutants was also much bigger than that of the vector and the wild-type ALK (Fig. 3D).

An earlier study showed cell invasion-promoting properties of various fusion ALK proteins (22). Such properties of the ALK mutants L1198F and G1201E were tested in the present studies. As shown in FIG. 4A, like ALK K1062M and F1174L, NIH3T3 cells transfected with the 2 novel ALK mutants were much more invasive on Matrigel matrix-coated membranes than vector or wild-type ALK-transfected cells. The number of invading cells was much bigger with the ALK mutants than with the vector and wild-type ALK (Fig. 4B).

Discussion

For the first time the ALK gene was analyzed for mutations in tumors other than neuroblastomas and 2 somatic novel missense ALK point mutations, C3592T and G3602A, were identified in ATC. The 2 mutations caused amino acid change from lysine to phenylalanine (L1198F) and from glycine to glutamic acid (G1201E) of ALK, respectively. They are both in exon 23 and only 3 codons apart, with a relatively high ALK mutation prevalence of 11% in ATC, similar to that in sporadic neuroblastomas (10-12, 14). The novel mutations L1198F and G1201E are located in the tyrosine kinase domain of the ALK. More

specifically, they are in the hinge region of this domain. It has been shown that ALK binds ADP and staurosporine at the hinge region of the interlobe cleft, suggesting important specific functions of this region (23). Therefore, it may be expected that amino acid changes in this region may result in conformational change of the ALK protein, with significant functional consequences to the tyrosine kinase activity of ALK. Indeed, both mutations were shown to be gain-of-function mutations, conferring that ALK dramatically increased tyrosine kinase activities. Like several previously characterized ALK mutants in neuroblastomas, the 2 novel ALK mutants found in ATC strongly promoted cell focus formation, anchorage-independent growth, and cell invasion, effectively showing their oncogenic functions. The present inventors thus report ALK mutations in ATC with a comparable prevalence and oncogenic power with those in neuroblastomas. Expression of ALK seems to be restricted to tissues originating from neuroectodermal developmental origin (24). Thyroid cancer cells were specifically shown to express ALK (24). Thus, the mutant ALK gene can be expected to be expressed in thyroid cells, making such mutations relevant for thyroid tumorigenesis. ATC is the most aggressive type of thyroid cancer; in fact, it is one of the most rapidly developing lethal human cancers (16). With the current available treatments, patients with ATC usually die within 5 to 6 months after initial diagnosis. The finding of the ALK mutations in ATC, but not in generally indolent differentiated PTC and FTC, suggests that activating genetic alterations of the ALK gene play a role in the aggressiveness of a subgroup of ATC. This is interestingly consistent with previous observations that ALK mutations were also associated with advanced disease stages and poor outcomes of neuroblastoma (11, 12, 14).

The molecular signaling pathways mediating the oncogenic role of ALK mutations in ATC involved the PI3K/Akt and MAP kinase pathways, as suggested by the present inventors' demonstration of the coupling of the 2 novel ALK mutants to increased phosphorylation of Akt and ERK in both ALK-transfected cell lines and ATC tumor tissues. This is not surprising, as tyrosine kinases, particularly RTK, typically activate these 2 pathways. Interestingly, in neuroblastoma, ALK mutations were also shown to be coupled to PI3K/Akt and MAP kinase signaling (8, 9). Thus, it seems that coupling to these pathways is a common mechanism involved in the oncogenic role of genetic alterations of the ALK gene in human cancer. The PI3K/Akt and MAP kinase pathways play a fundamental role in thyroid tumorigenesis (25, 26). Dual activation of the 2 pathways driven by genetic alterations is a fundamental mechanism for the pathogenesis of ATC (19, 27). ALK

mutations discovered in the present study add a novel set of genetic alterations that contribute to this mechanism.

Development of inhibitors targeting ALK is an exciting current research area as a novel therapeutic strategy for neuroblastomas harboring ALK mutations, ALCL NPM1-ALK, NSCLC harboring EML4-ALK, and IMT harboring TMP3/4- ALK and RANBP2-ALK fusion genes (1, 2). It has been recently reported that most of the patients with NSCLC harboring EML4-ALK responded effectively to the treatment with an anti-ALK agent, crizotinib (28), which has also been recently reported to be effective in treating a patient with IMT harboring RANBP2-ALK (29). In future studies, it will be interesting to examine the effects of this ALK inhibitor on the growth of ALK mutants in in vitro cell line assay and in vivo animal tumor studies. This will not only test the dependence of cells or tumors on these ALK mutants but also test the therapeutic potentials of targeting these novel ALK mutants in human cancers, particularly ATC. It is therefore highly attractive to speculate that the subset of patients with ATC harboring ALK mutations may be similarly and effectively treated with this and other ALK inhibitors.

In summary, in the present study, the present inventors report novel ALK mutations in ATC with a comparable prevalence and oncogenic power to those in neuroblastomas. This is the first report on ALK mutations in non-neuroblastoma human cancers. These are gain-of-function mutations that cause dual activation of the PI3K/Akt and MAP kinase pathways in ATC. They may represent a novel therapeutic target in ATC for the recently developed ALK inhibitors.

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That which is claimed is:

1. A method for treating a anaplastic thyroid cancer (ATC) patient comprising the step of administering to the patient an effective amount of an anaplastic lymphoma kinase
5 (ALK) inhibitor.
2. The method of claim 1, wherein the ALK inhibitor is crizotinib.
3. The method of claim 1, wherein the patient has a C3592T mutation in exon 23 of the
10 ALK gene.
4. The method of claim 1, wherein the patient has a G3602A mutation in exon 23 of the ALK gene.
- 15 5. The method of claim 1, further comprising the step of determining whether the patient has a C3592T and/or a G3602A mutation in exon 23 of the ALK gene prior to the administering step.
6. A method for treating ATC in a thyroid cancer patient comprising the steps of:
20 a. determining whether the thyroid cancer patient has a C3592T and/or a G3602A mutation in exon 23 of the anaplastic lymphoma kinase (ALK) gene; and
b. treating the patient with an ALK inhibitor therapy if the patient has either or both of the mutations.
- 25 7. A method for treating anaplastic thyroid cancer (ATC) in a thyroid cancer patient comprising the step of treating the patient with an ALK inhibitor therapy if the thyroid cancer patient has a C3592T and/or a G3602A mutation in exon 23 of the ALK gene.
8. The method of claims 6 or 7, wherein the ALK inhibitor is crizotinib.
30
9. A method for treating a cancer patient comprising the step of administering to the patient an effective amount of an ALK inhibitor, wherein the cancer patient has a C3592T and/or a G3602A mutation in exon 23 of the ALK gene.

10. A method for treating a cancer patient comprising the steps of:
- a. determining whether the cancer patient has a C3592T mutation in exon 23 of the anaplastic lymphoma kinase (ALK) gene; and
 - b. treating the patient with an ALK inhibitor therapy if the patient has either or
5 both of the mutations.
11. The method of claim 9 or 10, wherein the ALK inhibitor is crizotinib.
12. The method of claim 9 or 10, wherein the cancer is a neuroblastoma.
- 10 13. A method for treating a neuroblastoma cancer patient comprising the step of administering to the patient an effective amount of an ALK inhibitor, wherein the patient has a C3592T and/or a G3602A mutation in exon 23 of the ALK gene.
- 15 14. The method of claim 13, wherein the ALK inhibitor is crizotinib.
15. The method of any of claims 1-14, further comprising the step of administering an inhibitor of the PI3K/Akt pathway.
- 20 16. The method of any of claims 1-15, further comprising the step of administering an inhibitor of the MAP kinase pathway.
17. A method for treating a cancer patient comprising the step of administering to the patient an inhibitor of a protein or pathway selected from the group consisting of ALK, the
25 PI3K/Akt pathway and the MAP kinase pathway, wherein the patient has a C3592T and/or a G3602A mutation in exon 23 of the ALK gene.
18. The method of claim 17, wherein the ALK inhibitor is crizotinib.
- 30 19. The method of claim 17, wherein the patient has ATC.
20. The method of claim 17, wherein the patient has a neuroblastoma.
21. A method for diagnosing ATC in patient comprising the step of performing an assay
35 on a biological sample from the patient to identify the presence or absence of a C3592T

and/or a G3602A mutation in exon 23 of the ALK gene according to SEQ ID NO:6, wherein the presence of either of both of the mutations correlates with a diagnosis of ATC in the patient.

- 5 22. A method for determining a patient's risk of developing ATC comprising the step of performing an assay on a biological sample from the patient to identify the presence or absence of a C3592T and/or a G3602A mutation in exon 23 of the ALK gene according to SEQ ID NO:6, wherein the presence of either of both of the mutations correlates with a prognosis that the patient has a higher risk of ATC than a patient without the mutations, and
10 wherein the absence of the mutations correlates with a prognosis that the patient has a lower risk of ATC than a patient with either or both of the mutations.

23. A method for detecting ATC in a patient comprising the step of determining the presence of a C3592T and/or a G3602A mutation in exon 23 of the ALK gene according to
15 SEQ ID NO:6 in a blood sample of a patient, wherein the presence of the mutation indicates ATC in the patient.

24. A method for distinguishing ATC from non-ATC samples comprising the step of determining the presence of a C3592T and/or a G3602A mutation in exon 23 of the ALK
20 gene according to SEQ ID NO:6 in thyroid sample of a patient, wherein the presence of either or both of the mutations indicates ATC and absence of either or both of the mutations indicates non-ATC.

- 25 25. The method of claim 24, wherein the thyroid sample is a fine needle aspirate (FNA).

26. The method of claim 24, wherein the thyroid sample is a tissue sample.

27. The method of claim 24, wherein the thyroid sample is a cytological sample.

- 30 28. The method of claim 24, further comprising providing a diagnosis based on the presence or absence of either or both of the mutations.

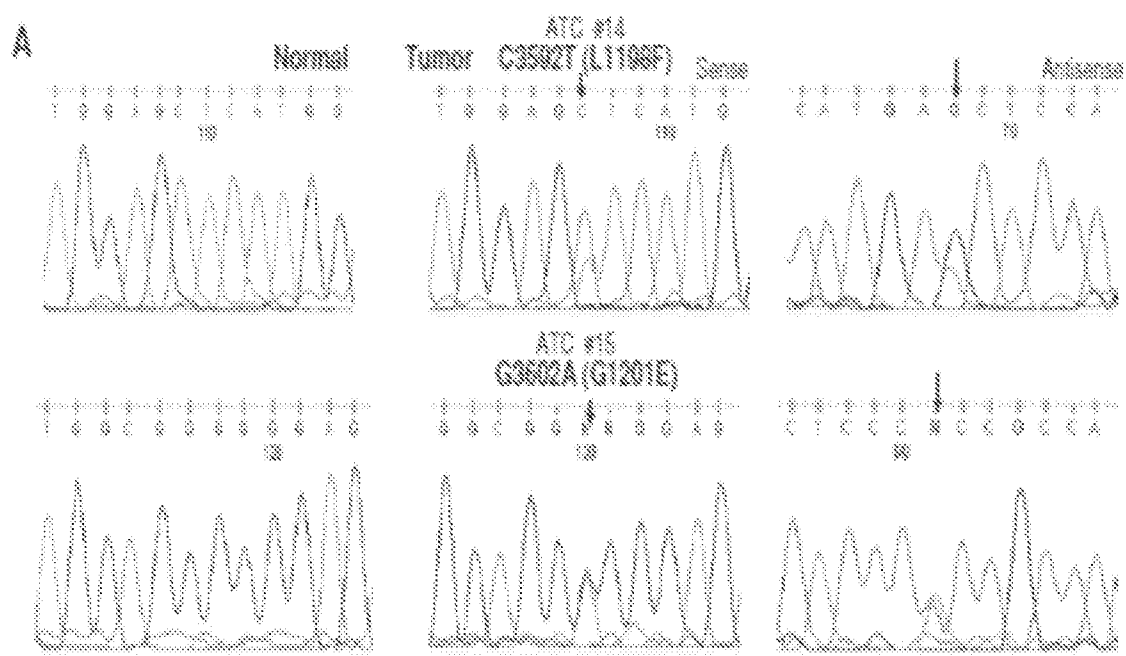
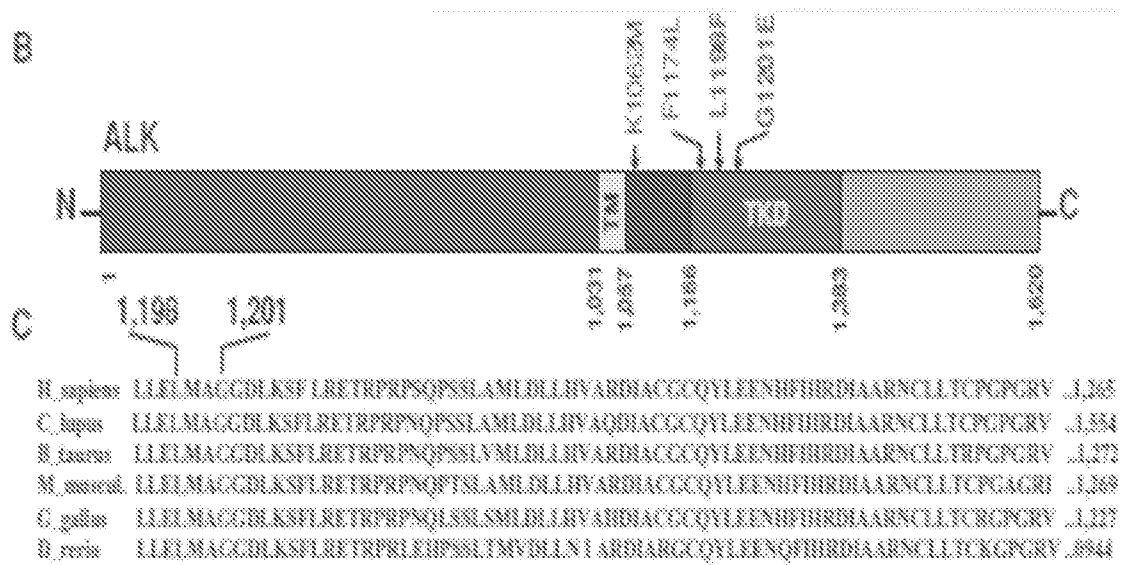


FIG. 1A



FIGS. 1B and 1C

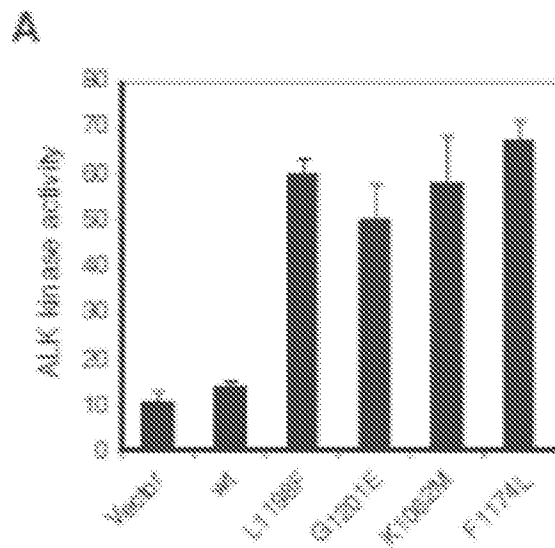


FIG. 2A

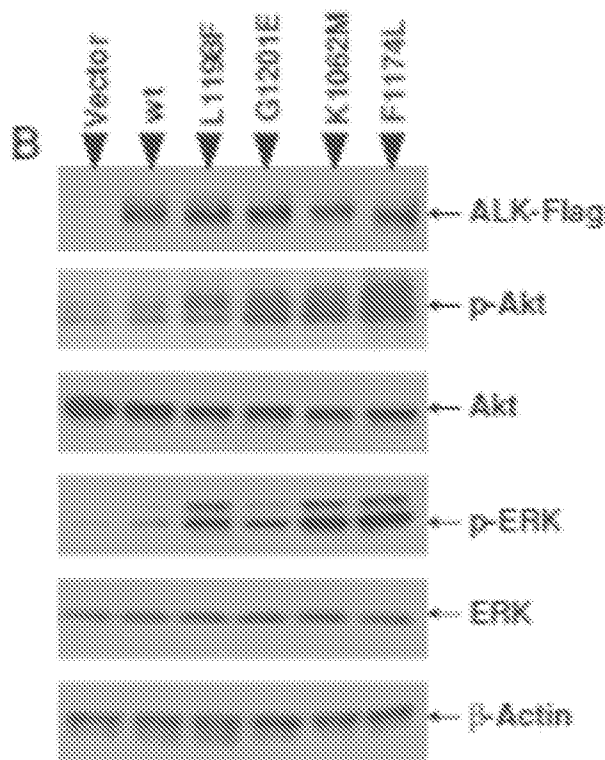


FIG. 2B

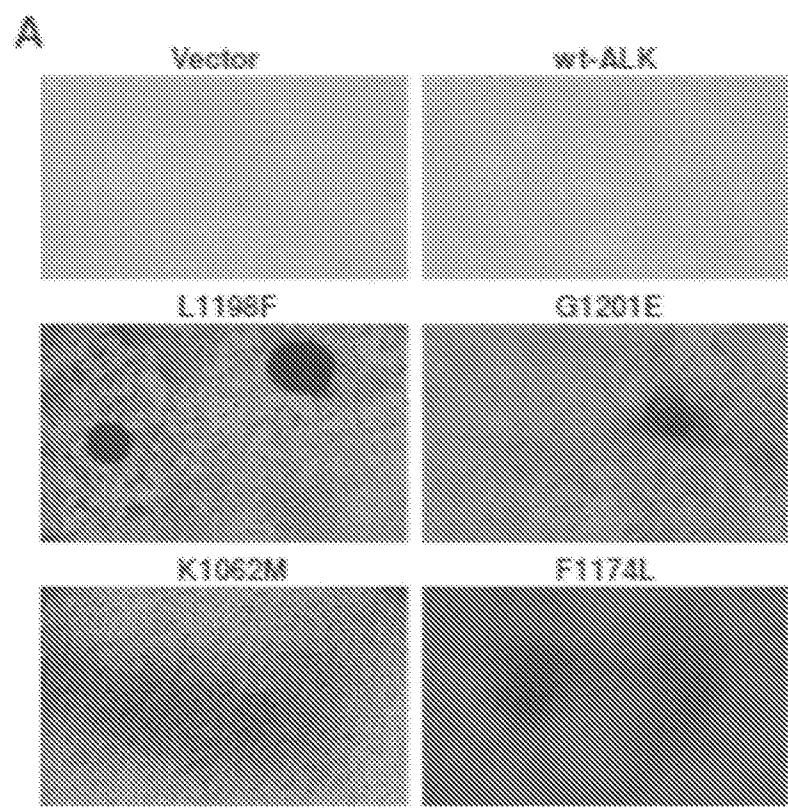


FIG. 3A

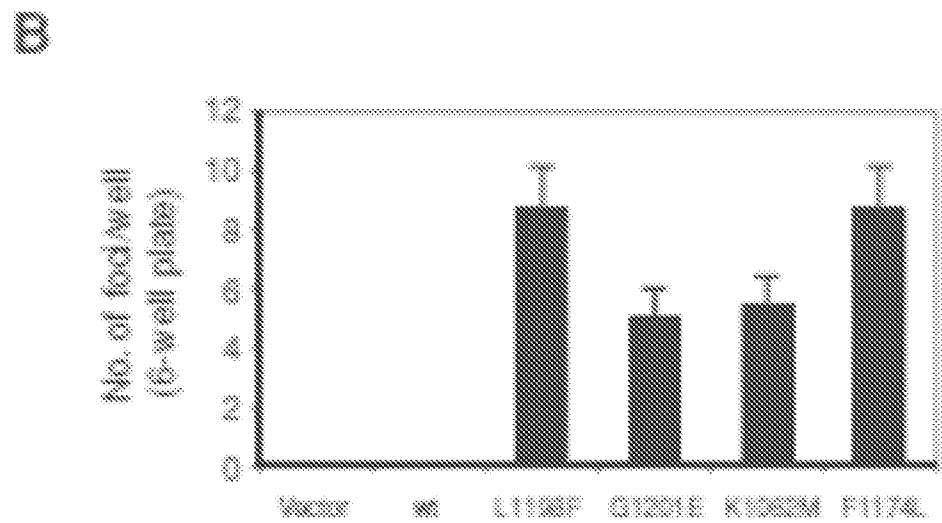


FIG. 3B

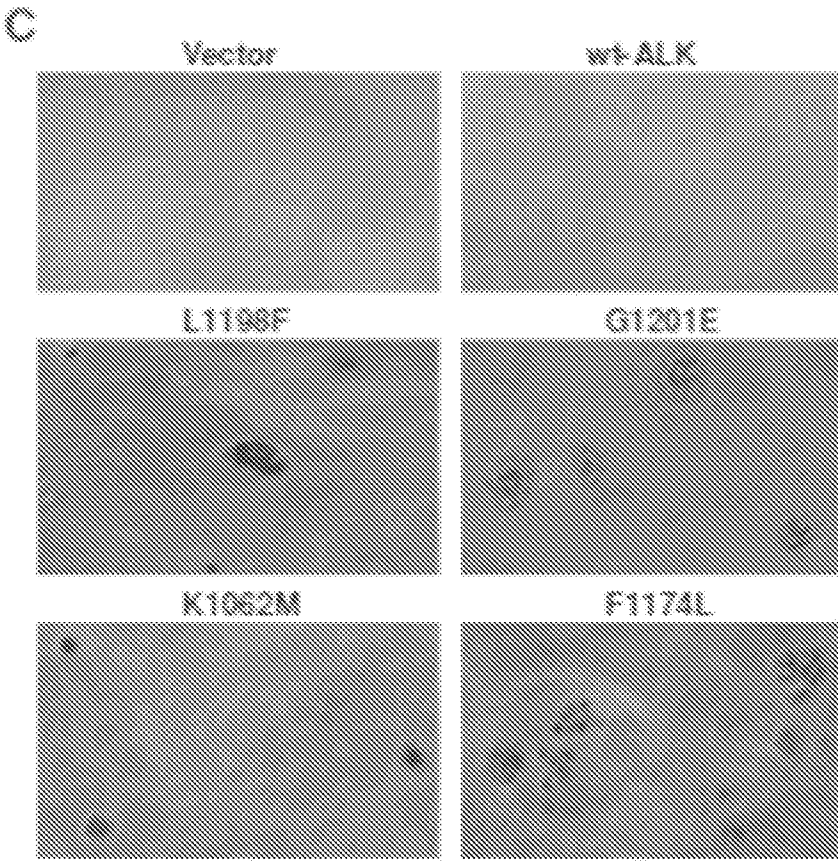


FIG. 4A

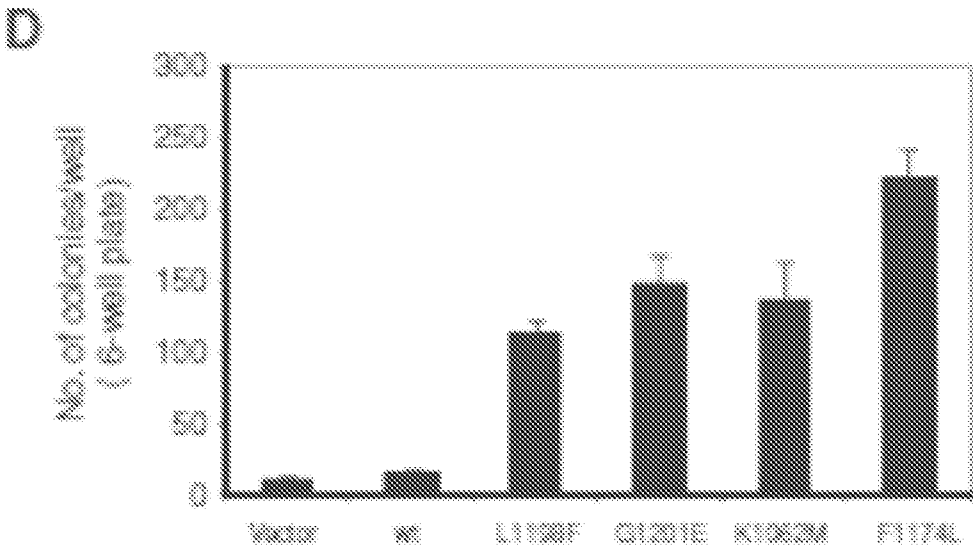


FIG. 4B
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