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(54) **Title**: RELAXIN FUSION POLYPEPTIDES AND USES THEREOF

(57) **Abstract**: The present invention provides Relaxin fusion polypeptides A-L-B with a non-wild type array of the Relaxin A-chain and Relaxin B-chain, wherein the A-and B-chains are connected by a linker peptide. The invention further provides Relaxin fusion polypeptides with extended half-life. Furthermore, the invention provides nucleic acid sequences encoding the foregoing fusion polypeptides, vectors containing the same, pharmaceutical compositions and medical use of such fusion polypeptides.

Relaxin fusion polypeptides and uses thereof

The present invention provides Relaxin fusion polypeptides A-L-B with a non-wild type array of the Relaxin A-chain and Relaxin B-chain, wherein the A- and B-chains are connected by a linker peptide. The invention further provides Relaxin fusion polypeptides with extended half-life. Furthermore, the invention provides nucleic acid sequences encoding the foregoing fusion polypeptides, vectors containing the same, pharmaceutical compositions and medical use of such fusion polypeptides.

Background of the invention

Relaxin 2 (H2 relaxin, RLN2) as a member of the insulin superfamily is a 2-chain peptide exhibiting, on the genetic level, the typical B - C - A chain prohormone structure, arranged from N- to C-terminus. Other members of this superfamily, encoded by 7 genes in human, are the relaxin genes RLN 1, RLN3, and the insulin-like peptide genes INSL3, INSL4, INSL5, and INSL6. The overall sequence homology between members of this family is low; nevertheless, phylogenetic analysis indicates that these genes have evolved from the RLN3 ancestral gene (Hsu, S. Y. (2003); Wilkinson, T. N. et al. (2005)). The mature protein has a molecular weight of approximately 6000 Da and is the product of an enzymatic cleavage of the prohormone catalyzed by the Prohormone-Convertase 1 (PC1) and 2 (PC2) (Hudson P. et al. (1983)). The resulting A- and B-chains are joined by two intermolecular cysteine bridges; the A-chain exhibits an additional intramolecular disulfide bond.

Relaxin initiates pleiotropic effects through multiple pathways on a variety of cell types. It confers its activity by binding to the class I (rhodopsin like) G-protein–coupled receptor termed LGR7 (leucine-rich G protein-coupled receptor 7) also named RXFP1 (relaxin family peptide 1 receptor), and with significantly lower affinity to LRG8/RXFP2 (relaxin family peptide 2 receptor) (Kong RC et al. (2010) Mol Cell Endocrinol. 320:1-15). Within the Relaxin molecule, an amino acid motif in the B-chain (Arg-X-X-X-Arg-X-X-Ile/Val-X) (Schwabe and Büllesbach (2007) Adv Exp Med Biol. 612:14-25 and Büllesbach and Schwabe J Biol Chem. 2000 Nov 10;275(45):35276-80) is conserved in all of the Relaxin peptides and is crucial for the interaction of these peptides with the corresponding receptor. Binding of Relaxin to LGR7/RXFP1 leads to activation of adenylate cyclase and to an increase of the second messenger molecule cAMP. Via this mechanism, Relaxin 2 for example mediates the release of atrial natriuretic peptide in rat hearts (Toth, M. et al. (1996)). A positive inotropic effect of Relaxin 2 on rat atrial myocytes has also been shown (Piedras-Renteria, E. S. et al. (1997)). Other signal transduction molecules which are activated by the Relaxin/LGR7 complex are the phosphoinositide-3 kinase, tyrosine kinases, and phosphodiesterases (Bartsch, O. et al. (2001), Bartsch, O. et al. (2004)). Additional signal transduction pathways activated by this system

include the nitric oxide (NO) pathway leading to increased levels of cyclic GMP in rat and guinea-pig hearts (Bani-Sacchi, T. et al. (1995)).

Relaxin acts as a pleiotropic hormone (Dschietzig T. et al. (2006)) possessing biological activity on organs such as lung, kidney, brain, and heart. A strong antifibrotic and vasodilator activity of Relaxin is most notably responsible for the positive effects obtained with this peptide in various animal disease models as well as in clinical studies (McGuane J.T. et al. (2005)). RLN2 has multiple beneficial actions in the cardiovascular system under pathological conditions. It maintains tissue homeostasis and protects the injured myocardium during various pathophysiological processes. It exhibits prominent vasodilatory effects, e.g. affecting flow and vasodilation in rodent coronary arteries (Nistri, S. et al. (2003)) and in the vascular beds of other organs. In spontaneously hypertensive rats RLN2 lowered blood pressure, an effect mediated by increased NO production.

A cardioprotective activity of Relaxin 2 has been evaluated in different animal models such as guinea pig, rat and pig (Perna A.M. et al. (2005), Bani, D. et al. (1998)). RLN2 ameliorates myocardial injury, inflammatory cell infiltration and subsequent fibrosis, thereby alleviating severe ventricular dysfunction (Zhang J. et al. (2005)).

Relaxin 2 exhibits strong antifibrotic activity. In injured tissues, fibroblast activation and proliferation causes increased collagen production and interstitial fibrosis. Fibrosis in the heart is increased by biomechanical overload, and influences ventricular dysfunction, remodeling, and arrhythmogenesis. In animal models, continuous infusion of Relaxin 2 inhibits or even reverses cardiac dysfunction caused by cardiomyopathy, hypertension, isoprenaline-induced cardiac toxicity, diabetic cardiomyopathy and myocardial infarction. This inhibition of fibrogenesis or reversal of established fibrosis can reduce ventricular stiffening and improve diastolic function. Notably, although Relaxin 2 reduces aberrant collagen accumulation, it does not affect basal collagen content in healthy tissues, highlighting its safety for therapeutic use.

Relaxin 2 has been tested in several clinical studies as a pleiotropic vasodilator for the treatment of patients with acute heart failure with very promising outcome. In these studies, Relaxin 2 was associated with favourable relief of dyspnoea and other clinical outcomes (Teerlink J.R. et al. (2009), Metra M. et al. (2010))

Due to the limited in-vivo half life of Relaxin, treatment of patients has to be repeated every 14 to 21 days, whereby compound administration has to be performed as a continuous infusion for at least 48 hours.

Furthermore, Relaxin 2 may also be useful in the treatment of diseases such as pancreatitis, inflammation-related diseases like rheumatoid arthritis, and cancer (Cosen-Binker L.I. et al. (2006) Santora K. Et al. (2007)) or scleroderma, pulmonary, renal, and hepatic fibrosis (Bennett RG. (2009)). Relaxin 2 reduces xenograft tumour growth of human MDA-MB-231 breast cancer cells (Radestock Y, Hoang-Vu C, Hombach-Klonisch S. (2008) Breast Cancer Res. 10:R71).

The synthesis of Relaxin 2 by chemical methods is difficult. Due to the low solubility of the B-chain and the requirement for the laborious, specific introduction of cysteine bridges between A and B-chains, yields of active peptide obtained by these methods are extremely low (Barlos K.K. et al. (2010)). Alternatively, recombinant expression of Relaxin 2 can be performed. To allow efficient cleavage of the prepro-peptide during post-translational modifications and the secretion of mature and biological active peptides, expression host cells are routinely co-transfected with expression constructs encoding the Prohormone-Convertase 1 and/or 2 (Park J.I. et al. (2008)). Nevertheless, the endoproteolytic processing efficiency of prepro-peptides in heterologous cells often limits the production of bioactive molecules significantly (Shaw J.A. et al. (2002)).

Therefore, it would be of great advantage to generate a Relaxin molecule which independent of endoproteolytic processing mediated by specific proteases exhibits full biological activity and can be produced in significant yields using heterologous expression systems.

For human Insulin, single-chain variants have been generated in which an uncleavable polypeptide connects the insulin B-chain with the insulin A-chain (Rajpal G. et al. (2009)). For these variants, endoproteolytic processing is dispensable.

Surprisingly, we identified a Relaxin variant in which the orientation of the two active chains, designated as A chain and B chain, are exchanged and the cleavable C chain is substituted by linker peptide. As shown in Figure 1, instead of the genetically determined orientation of the single chains encoding Relaxin, namely B chain – C chain – A chain, the orientation of the chains of the modified molecule is: A chain –linker peptide – B chain. The resulting molecule exhibits full biological activity, independent of any endoproteolytic processing. This new single-chain Relaxin variant provided by the invention thus solves the problem of low expression yields or the requirement of co-transfection with a processing protease.

The half-life of intravenously administrated Relaxin 2 in humans is less than 10 minutes (Dschietzig T. et al. (2009)). As a consequence, in clinical trials Relaxin 2 has to be administered continuously over 48h. Therefore, the improvement of the biological half life of Relaxin could be of great advantage.

Improving biological half life can either be performed by chemical modification such as PEGylation or HESylation of the polypeptide of interest, introduction of additional, non-natural N-glycosylation sites or by genetically fusing this polypeptide with other molecules such as the immunoglobulin Fc fragment of antibodies, transferrin, albumin, binding modules that bind in-vivo to other molecules mediating longer half-life, or other proteins, respectively. This invention provides single-chain Relaxin variants fused to the Fc part of antibodies with improved half-life. Surprisingly, these variants show biological activity in the range of the wild-type Relaxin.

Summary of the invention

The invention concerns fusion polypeptides, hereafter also referred to as single chain Relaxin (scRelaxin).

Current standard of Relaxin 2 production is the chemical synthesis of this molecule, which is a complex and expensive procedure. Due to the fact that Relaxin undergoes posttranslational modifications, especially the cleavage of the prepro-protein by the Prohormon Convertase 1 and Prohormone Convertase 2, choice of an adequate expression system is mandatory for recombinant expression. Endoproteolytic processing of proteins belonging to the insulin superfamily often limits the production of bioactive molecules from heterologous cells. To avoid the endoproteolytic processing of Relaxin, the fusion polypeptides of the invention are molecules in which the genetically encoded orientation of the two active chains of Relaxin, designated as A chain and B chain, is reversed wherein the A chain and B chain are connected by a linker peptide. In detail, instead of the genetically determined orientation of the individual DNA segments encoding Relaxin domains, namely, B chain - C chain - A chain, the orientation the DNA segments in the Relaxin variants provided by this invention is: A chain – peptide linker – B chain. This results in a single chain Relaxin wherein the carboxy-terminus of Relaxin A chain is fused to the amino-terminus of the linker polypeptide L, which carboxy-terminus is fused to the amino-terminus of the Relaxin B chain, designated A-L-B (see Figure 1 for an illustration). The resulting molecule exhibits biological activity similar to the wild-type Relaxin, but its expression is independent of endo-proteolytic processing.

One embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein A comprises a Relaxin A chain polypeptide or a functional variant thereof, B comprises a Relaxin B chain polypeptide or a functional variant thereof and L is a linker polypeptide.

In a further embodiment the Relaxin A chain polypeptide of A-L-B comprises a Relaxin 2 A chain polypeptide or a functional variant thereof and the Relaxin B chain polypeptide comprises a Relaxin 2 B chain polypeptide or a functional variant thereof.

In a preferred embodiment the Relaxin A chain polypeptide of A-L-B comprises a human minimal Relaxin 2 A chain polypeptide (SEQ ID NO: 118) or a functional variant thereof, or comprises a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof.

In a preferred embodiment the Relaxin B chain polypeptide of A-L-B comprises a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof.

In a more preferred embodiment the Relaxin A chain polypeptide of A-L-B comprises a human minimal Relaxin 2 A chain polypeptide (SEQ ID NO: 118) or a functional variant thereof, or comprises a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof and the Relaxin B chain polypeptide comprises a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof.

In an even more preferred embodiment the Relaxin A chain polypeptide of A-L-B is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof and the Relaxin B chain polypeptide is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof.

In one embodiment the linker polypeptide L of the aforementioned fusion polypeptides A-L-B consists of a polypeptide which is 6-14 amino acid residues in length. Further preferred are polypeptide linkers L which are 7-13 amino acid residues in length. Further preferred are polypeptide linkers L which are 8-12 amino acid residues in length. Even more preferred are polypeptide linkers L which are 7-11 or 9-11 amino acid residues in length. Even more preferred are polypeptide linkers L which are 9 amino acid residues in length. In a further preferred embodiment, the integer of the length of the polypeptide linker L is selected from the group consisting of the integers 6, 7, 8, 9, 10, 11, 12, 13 and 14.

The linker peptide L can be composed of any amino acid. In a preferred embodiment the linker polypeptide L comprises at least one Gly, Ser, Arg, Leu, Cys, Ala, Leu and/or Lys residue. In a more preferred embodiment the linker polypeptide L comprises Gly and Ser residues. A further preferred embodiment is a linker L which comprises Gly and Ser residues and has a ratio of Gly to Ser of at least 3 to 1.

In a further embodiment the aforementioned linker L comprises at least one attachment site for covalent coupling of a half-life extending moiety. In an embodiment of the invention the aforementioned attachment site is a Lys or a Cys residue.

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof, and

L is a linker polypeptide which is 6-14, 7-13, 8-12, 7-11, 9-11, or 9 amino acid residues in length. The linker peptide L can be composed of any amino acid. In a preferred embodiment the linker polypeptide L comprises at least one Gly, Ser, Arg, Leu, Cys, Ala, Leu and/or Lys residue. In a more preferred embodiment the linker polypeptide L comprises Gly and Ser residues. A further preferred embodiment is a linker L which comprises Gly and Ser residues and has a ratio of Gly to Ser of at least 3 to 1. In a further embodiment the aforementioned linker L comprises at least one attachment site for covalent coupling of a non-proteinaceous polymer half-life extending moiety. In an embodiment of the invention the aforementioned attachment site is a Lys or a Cys residue.

A preferred embodiment of the invention is a fusion polypeptide A-L-B further comprising a half-life extending moiety.

In a further embodiment the aforementioned fusion polypeptides have Relaxin activity. In a further preferred embodiment the Relaxin activity is activation of the relaxin receptor LGR7. In an even further preferred embodiment, the activation of the relaxin receptor LGR7 is determined by a method disclosed in experimental methods.

In another aspect, the invention provides a polynucleotide encoding an aforementioned fusion polypeptide. Such a polynucleotide may further comprise a coding sequence for a signal peptide allowing secretion of the fusion polypeptide. Vectors containing polynucleotides for such fusion polypeptides are included as well. Suitable vectors are for example expression vectors. A further embodiment of the invention is a host cell comprising a polynucleotide, a vector, or expression vector encoding the aforementioned fusion polypeptides. The host cell of the invention can be an eukaryotic cell or a prokaryotic cell. An eukaryotic cell can be a mammalian cell or a yeast or insect cell, preferably a mammalian cell. A prokaryotic cell can be for example an E. coli cell.

In another embodiment the invention provides pharmaceutical compositions comprising the aforementioned fusion polypeptides. The composition may be formulated for intravenous, intraperitoneal or subcutaneous administration.

Another embodiment of the invention provides a pharmaceutical composition or a fusion polypeptide as medicament. A further embodiment is the use of a pharmaceutical composition

or a fusion polypeptide in the treatment of cardiovascular diseases, pancreatitis, inflammation, cancer, scleroderma, pulmonary, renal, and hepatic fibrosis.

Brief description of the drawings

Fig. 1 Schematic representation of the genetic organization of domains of the wildtype Relaxin and single chain Relaxin as well as their corresponding polypeptides.

Fig. 2 Schematic representation of single chain Relaxin variants.

Fig. 3 Schematic representation of domain organisation of single chain Relaxin fusion protein variants as well as single chain Relaxin variants designed for PEGylation.

Fig. 4a-e Activity in a functional assay of scR 3, scR 4, and scR 5 (Fig. 4a), scR 7, scR 8, scR9, and scR10 (Fig. 4b), scR11 and scE12 (Fig. 4c), human Relaxin 3, scR14, and scR15 (Fig. 4d) and scR17 (Fig. 4e) using the CHO-CRE-LGR7 cell line. As control, hRelaxin 2 (R&D Systems, catalogue number 6586-RN-025) was used. Data are expressed as Relative Light Units, representing the activity of single chain Relaxin variants and Relaxin 2 induced luciferase expression. Symbols represent means, error bars represent S.E.M.

Fig. 5 Activity in a functional assay of scR-Fc 1 by using the CHO-CRE-LGR7 cell line. As control, hRelaxin 2 (R&D Systems, catalogue number 6586-RN-025) was used. Data are expressed as Relative Light Units, representing the activity of scR-Fc 1 and hRelaxin 2 induced luciferase expression. Symbols represent means, error bars represent S.E.M.

Fig. 6 Activity in a functional assay of scR-Fc 5, scR-Fc 6, and scR-Fc 7 using the CHO-CRE-LGR7 cell line. hRelaxin 2 (R&D Systems, catalogue number 6586-RN-025). was used as control. Data are expressed as Relative Light Units, representing the activity of the scR-Fc variants and hRelaxin 2 induced luciferase expression. Symbols represent means, error bars represent S.E.M.

Fig. 7 Activity in a functional assay of scR-Fc 11, scR-Fc 12, and scR-Fc 13 using the CHO-CRE-LGR7 cell line. As control, hRelaxin 2 (R&D Systems, catalogue number 6586-RN-025) was used. Data are expressed as Relative Light Units, representing the activity of scR-Fc variants and hRelaxin 2 induced luciferase expression. Symbols represent means, error bars represent S.E.M.

Fig. 8: Activity in a functional assay of scR-Var 3, scR-Var 4, scR-Var 5, and scR-Var 6 using the CHO-CRE-LGR7 cell line. As control, hRelaxin 2 (R&D Systems, catalogue number 6586-RN-025) was used. Data are expressed as Relative Light Units, representing the activity of scR-Fc variants and hRelaxin 2 induced luciferase expression. Symbols represent means, error bars represent S.E.M.

Fig. 9 In vivo half-life analysis of intravenously administrated hRelaxin 2 or scR-Fc 13. Eight weeks old male Wistar rats (three animals per group) were given a single application of human Relaxin 2 and scR-Fc 13, respectively (0,24 mg/kg). Blood samples were collected at the indicated time points after application and serum levels of each protein were measured by using a quantification ELISA.

Fig. 10: Activity of Relaxin 2 and Relaxin variants in blood samples

Relaxin activity in blood samples obtained from scR-Fc 13 treated rats by using the CHO-CRE-LGR7 cell line was determined. Blood samples collected 3, 5, and 7 days after intravenous administration of scR-Fc 13 were incubated on the CHO-CRE-LGR7 cell line and Relative Lights Units were determined. Calibration curves were determined using hRelaxin 2 (R&D Systems, catalogue number 6586-RN-025) and purified scR-Fc 13. The EC50 within the dose response curve is marked by an X. Data are expressed as Relative Light Units, representing the activity of scR-Fc variants and hRelaxin 2 induced luciferase expression. Symbols represent means, error bars represent S.E.M.

Fig. 11: Influence of hRelaxin 2 and scR-Fc 13 on heart rate, coronary flow and contractility in the isolated perfused rat heart model.

At a concentration of 1 nM, application of hRelaxin 2 leads to an increase of heart rate and coronary flow and exhibits a negative inotrophic activity (Fig. 11 a – d). Comparable effects were obtained with scR-Fc 13, although at a ten fold higher concentration (Fig. 11 e – h).

Detailed description of the invention

Definitions:

The term "amino acid residue" is intended to indicate an amino acid residue contained in the group consisting of alanine (Ala or A), cysteine (Cys or C), aspartic acid (Asp or D), glutamic acid (Glu or E), phenylalanine (Phe or F), glycine (Gly or G), histidine (His or H), isoleucine (Ile or I), lysine (Lys or K), leucine (Leu or L), methionine (Met or M), asparagine (Asn or N), proline (Pro or P), glutamine (Gln or Q), arginine (Arg or R), serine (Ser or S), threonine (Thr or T), valine (Val or V), tryptophan (Trp or W), and tyrosine (Tyr or Y) residues.

The term "activity of Relaxin" or "Relaxin Acitvity" is defined by the ability of Relaxin or variants thereof to the activation of the stimulatory G-protein Gs, thus the subsequent generation of the second messenger cyclic AMP, and/or the stimulation of PI3-kinase. Relaxin or variants thereof bind to LGR7 leading to the intracellular activation of the stimulatory G-protein Gs, resulting in the subsequent generation of the second messenger cyclic AMP (cAMP). However, cAMP generation is a time-dependent biphasic response. After an initial short Gs-adenylate cyclase-mediated cAMP response the receptor signal is switching to an inhibitory G protein activation and by this to PI3-kinase–mediated response. (Halls M.L., Bathgate R.A., Summers, R.J. (2005) Signal Switching after Stimulation of LGR7 Receptors by Human Relaxin 2. Ann. N.Y. Acad. Sci. 1041:288–291).

The term "half-life extending moiety" refers to a pharmaceutically acceptable moiety, domain, or "vehicle" covalently linked ("conjugated") to the Relaxin fusion polypeptide directly or via a linker, that prevents or mitigates in vivo proteolytic degradation or other activity-diminishing chemical modification of the Relaxin fusion polypeptide , increases half-life or other pharmacokinetic properties such as but not limited to increasing the rate of absorption, reduces toxicity, improves solubility, increases biological activity and/or target selectivity of the Relaxin fusion polypeptide, increases manufacturability, and/or reduces immunogenicity of the Relaxin fusion polypeptide, compared to an unconjugated form of the Relaxin fusion polypeptide. The term "half-life extending moiety" includes non-proteinaceous, half-life extending moieties, such as PEG or HES, and proteinaceous half-life extending moieties, such as serum albumin, transferrin or Fc domain.

"Polypeptide", peptide" and "protein" are used interchangeably herein and include a molecular chain of two or more amino acids linked through peptide bonds. The terms do not refer to a specific length of the product. The terms include post-translational modifications of the polypeptide, for example, glycosylations, acetylations, phosphorylations and the like. In addition, protein fragments, analogs, mutated or variant proteins, fusion proteins and the like are included within the meaning of polypeptide. The terms also include molecules in which one or more amino acid analogs or non-canonical or unnatural amino acids are included as can be synthesized, or expressed recombinantly using known protein engineering techniques. In addition, inventive fusion proteins can be derivatized as described herein by well-known organic chemistry techniques.

The term "functional variant" refers to a variant polypeptide which at least retains some of its natural biological activity. In case of the Relaxin 2 variants according to the invention, a functional variant is a variant which shows at least some of its natural activity, such as the

activation of the relaxin receptor LGR7. The activation of the relaxin receptor LGR7 can be determined by a method disclosed in experimental methods.

The terms “fragment,” “variant,” “derivative,” and “analog” when referring to polypeptides of the present invention include any polypeptides that retain at least some of the receptor binding properties of the corresponding wild-type Relaxin polypeptide. Fragments of polypeptides of the present invention include proteolytic fragments, as well as deletion fragments, and also polypeptides with altered amino acid sequences due to amino acid substitutions, deletions, or insertions. Variants may occur naturally or be non-naturally occurring. Non-naturally occurring variants may be produced using art-known mutagenesis techniques. Variant polypeptides may comprise conservative or non-conservative amino acid substitutions, deletions, or additions. Variant polypeptides may also be referred to herein as “polypeptide analogs.” As used herein a “derivative” of a polypeptide refers to a subject polypeptide having one or more residues chemically derivatized by reaction of a functional side group. Also included as “derivatives” are those peptides that contain one or more naturally occurring amino acid derivatives of the twenty standard amino acids. For example, 4-hydroxyproline may be substituted for proline; 5-hydroxylysine may be substituted for lysine; 3-methylhistidine may be substituted for histidine; homoserine may be substituted for serine; and ornithine may be substituted for lysine.

The term “fusion protein” indicates that the protein includes polypeptide components derived from more than one parental protein or polypeptide and/or that the fusion protein includes protein domains derived from one or more parental protein or polypeptide which are not arrayed in their wild type orientation. Typically, a fusion protein is expressed from a fusion gene in which a nucleotide sequence encoding a polypeptide sequence from one protein is appended in frame with, and optionally separated by a linker or stretcher from, a nucleotide sequence encoding a polypeptide sequence from a different protein. The fusion gene can then be expressed by a recombinant host cell as a single protein.

The term "nucleotide sequence" or “polynucleotide “is intended to indicate a consecutive stretch of two or more nucleotide molecules. The nucleotide sequence may be of genomic, cDNA, RNA, semisynthetic, synthetic origin, or any combinations thereof.

The term “EC_{50}” (half maximal effective concentration) refers to the effective concentration of a therapeutic compound which induces a response halfway between the baseline and maximum after some specified exposure time.

The term "immunogenicity" as used in connection with a given substance is intended to indicate the ability of the substance to induce a response from the immune system. The immune response may be a cell or antibody mediated response (see, e.g., Roitt: Essential Immunology

(8th Edition, Black-well) for further definition of immunogenicity). Normally, reduced antibody reactivity will be an indication of reduced immunogenicity. The reduced immunogenicity may be determined by use of any suitable method known in the art, e.g. in vivo or in vitro.

The term "polymerase chain reaction" or "PCR" generally refers to a method for amplification of a desired nucleotide sequence in vitro, as described, for example, in US Pat. No. US 4,683,195 and US 4,683,195. In general, the PCR method involves repeated cycles of primer extension synthesis, using oligonucleotide primers capable of hybridising preferentially to a template nucleic acid.

The term "vector" refers to a plasmid or other nucleotide sequences that are capable of replicating within a host cell or being integrated into the host cell genome, and as such, are useful for performing different functions in conjunction with compatible host cells (a vector-host system): to facilitate the cloning of the nucleotide sequence, i.e. to produce usable quantities of the sequence, to direct the expression of the gene product encoded by the sequence and to integrate the nucleotide sequence into the genome of the host cell. The vector will contain different components depending upon the function it is to perform.

"Cell", "host cell", "cell line" and "cell culture" are used interchangeably herein and all such terms should be understood to include progeny resulting from growth or culturing of a cell.

The term "functional in vivo half-life" is used in its normal meaning, i.e. the time at which 50% of the biological activity of the polypeptide is still present in the body/target organ, or the time at which the activity of the polypeptide is 50% of the initial value.

As an alternative to determining functional in vivo half-life, "serum half-life" may be determined, i.e. the time at which 50% of the polypeptide circulates in the plasma or bloodstream prior to being cleared. Determination of serum half-life is often more simple than determining the functional in vivo half-life and the magnitude of serum half-life is usually a good indication of the magnitude of functional in vivo half-life. Alternatively terms to serum half-life include "plasma half-life", "circulating half-life", "serum clearance", "plasma clearance" and "clearance half-life". The polypeptide is cleared by the action of one or more of the reticuloendothelial systems (RES), kidney, spleen or liver, by tissue factor, SEC receptor or other receptor mediated elimination, or by specific or unspecific proteolysis. Normally, clearance depends on size (relative to the cutoff for glomerular filtration), charge, attached carbohydrate chains, and the presence of cellular receptors for the protein. The functionality to be retained is normally selected from receptor binding or receptor activation. The functional in vivo half-life and the serum half-life may be determined by any suitable method known in the art and may for example generally involve the steps of suitably administering to a mammalian a

suitable dose of the amino acid sequence or compound to be treated; collecting blood samples or other samples from said mammalian at regular intervals; determining the level or concentration of the amino acid sequence or compound of the invention in said blood sample; and calculating, from (a plot of) the data thus obtained, the time until the level or concentration of the amino acid sequence or compound of the invention has been reduced by 50% compared to the initial level upon dosing. Reference is for example made to the standard handbooks, such as Kenneth, A et al: Chemical Stability of Pharmaceuticals: A Handbook for Pharmacists and in Peters et al, Pharmacokinete analysis: A Practical Approach (1996). Reference is also made to "Pharmacokinetics", M Gibaldi & D Perron, published by Marcel Dekker, 2nd Rev. edition (1982).

"Glycosylation" is a chemical modification wherein sugar moieties are added to the polypeptide at specific sites. Glycosylation of polypeptides is typically either N-linked or O-linked. N-linked refers to the attachment of a carbohydrate moiety to the side chain of an asparagine residue. The tripeptide sequences Asn-X-Ser and Asn-X-Thr ("N-X-S/T"), where X is any amino acid except proline, are the recognition sequences for enzymatic attachment of the carbohydrate moiety to the asparagine side chain. Thus, the presence of either of these tripeptide sequences (or motifs) in a polypeptide creates a potential N-linked glycosylation site. O-linked refers to the attachment of a carbohydrate moiety to the hydroxyl-group oxygen of serine and threonine.

An "isolated" fusion polypeptide is one that has been identified and separated from a component of the cell that expressed it. Contaminant components of the cell are materials that would interfere with diagnostic or therapeutic uses of the fusion polypeptide, and may include enzymes, hormones, and other proteinaceous or nonproteinaceous solutes. In preferred embodiments, the fusion polypeptide is purified (1) to greater than 95% by weight of fusion polypeptide as determined e.g. by the Lowry method, UV-Vis spectroscopy or by by SDS-Capillary Gel electrophoresis (for example on a Caliper LabChip GXII, GX 90 or Biorad Bioanalyzer device), and in further preferred embodiments more than 99% by weight, (2) to a degree sufficient to obtain at least 15 residues of N-terminal or internal amino acid sequence, or (3) to homogeneity by SDS-PAGE under reducing or non-reducing conditions using Coomassie blue or, preferably, silver stain. Ordinarily, however, isolated fusion polypeptides will be prepared by at least one purification step.

Overview

The application provides an A-L-B fusion polypeptide, also used terms herein are single chain Relaxin abbreviated as scRelaxin or scR, wherein "A" is a Relaxin A chain, "B" is a Relaxin B chain and "L" is a linker polypeptide. The present application describes an improved Relaxin molecule, wherein the C-terminus of an A chain is linked via a polypeptide linker to the N-

terminus of a B chain allowing the fusion polypeptide being expressed as a functional scRelaxin. The application relates, in part, on the surprising discovery that the A-L-B fusion polypeptides can be functionally expressed without the need for endoproteolytic prohormone processing as known for wildtype Relaxin.

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Single Chain Versions of Relaxin

Relaxin A and B Domains:

One embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein A comprises a Relaxin A chain polypeptide or a functional variant thereof, B comprises a Relaxin B chain polypeptide or a functional variant thereof and L is a linker polypeptide.

A further embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein A comprises a Relaxin A chain polypeptide or a functional variant thereof, B comprises a Relaxin B chain polypeptide or a functional variant thereof and L is a linker polypeptide, wherein Relaxin is selected from the group of Relaxins consisting of Relaxin 1, Relaxin 2, Relaxin 3, INSL3, INSL4, INSL5, and INSL6. In a further preferred embodiment the Relaxin is Relaxin 2 or Relaxin 3. In a further embodiment the aforementioned Relaxins are human Relaxins.

In a further embodiment the Relaxin A chain polypeptide of A-L-B comprises a Relaxin 2 A chain polypeptide or a functional variant thereof. In a further embodiment the Relaxin B chain polypeptide of A-L-B comprises a Relaxin 2 B chain polypeptide or a functional variant thereof.

In a further embodiment the Relaxin A chain polypeptide of A-L-B comprises a Relaxin 2 A chain polypeptide or a functional variant thereof and the Relaxin B chain polypeptide comprises a Relaxin 2 B chain polypeptide or a functional variant thereof.

In a preferred embodiment the Relaxin A chain polypeptide of A-L-B comprises a human minimal Relaxin 2 A chain polypeptide (SEQ ID NO: 118) or a functional variant thereof, or comprises a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof. In a preferred embodiment the Relaxin B chain polypeptide of A-L-B comprises a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof.

In a more preferred embodiment the Relaxin A chain polypeptide of A-L-B comprises a human minimal Relaxin 2 A chain polypeptide (SEQ ID NO: 118) or a functional variant thereof, or comprises a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof and the Relaxin B chain polypeptide comprises a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof.

In a further embodiment the Relaxin A chain polypeptide of A-L-B comprises a Relaxin 3 A chain polypeptide or a functional variant thereof. In a further embodiment the Relaxin B chain polypeptide of A-L-B comprises a Relaxin 3 B chain polypeptide or a functional variant thereof.

In a further embodiment the Relaxin A chain polypeptide of A-L-B comprises a human Relaxin 3 A chain polypeptide (SEQ ID NO:124) or a functional variant thereof. In a further

embodiment the Relaxin B chain polypeptide of A-L-B comprises a human Relaxin 3 B chain polypeptide (SEQ ID NO: 125) or a functional variant thereof. In a preferred embodiment the Relaxin A chain polypeptide of A-L-B comprises a human Relaxin 3 A chain polypeptide (SEQ ID NO: 124) or a functional variant thereof and the Relaxin B chain polypeptide comprises a human Relaxin 3 B chain polypeptide (SEQ ID NO: 125) or a functional variant thereof.

In a preferred embodiment of the aforementioned fusion polypeptides A-L-B a functional variant of the Relaxin A or B chain has 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions, insertions and/or deletions compared to the wild type Relaxin A and B chain, respectively. Further preferred is an aforementioned Relaxin 2 B variant that further comprises the conserved motif Arg-X-X-X-Arg-X-X-Ile/Val-X.

Relaxin A and B chain variants are known in the art. The well characterized binding site geometry of Relaxin provides the skilled person with guidance to design Relaxin A and B chain variants, see for example Büllesbach and Schwabe J Biol Chem. 2000 Nov 10; 275(45):35276-80 for variations of the Relaxin B chain and Hossain et al. J Biol Chem. 2008 Jun 20; 283(25):17287-97 for variations of the Relaxin A chain and the "minimal" Relaxin A chain. For example, for the conserved Relaxin 2 B motif (Arg-X-X-X-Arg-X-X-Ile/Val-X) X represents amino acids which are able to form a helical structure example to select appropriate amino acids X in the conserved motif as the three defined amino acids form a triangular contact region on the surface of the Relaxin B chain (Büllesbach and Schwabe J Biol Chem. 2000 Nov 10; 275(45)).

In an even more preferred embodiment the Relaxin A chain polypeptide of A-L-B is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof and the Relaxin B chain polypeptide is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof. In an even more preferred embodiment, the functional variant of human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) is a functional variant having 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions, deletions and/or insertions compared to SEQ ID NO: 117. Further preferred is a functional variant of human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) wherein the functional variant has 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions, deletions and/or insertions compared to SEQ ID NO: 119. Even further preferred is an aforementioned human Relaxin 2 B variant that further comprises the conserved motif Arg-X-X-X-Arg-X-X-Ile/Val-X.

In an even more preferred embodiment the Relaxin A chain polypeptide of A-L-B is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof having 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid exchanges compared to SEQ ID NO: 117 and the Relaxin B chain polypeptide is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional

variant thereof having 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid exchanges compared to SEQ ID NO:119 and comprising the conserved motif Arg-X-X-X-Arg-X-X-Ile/Val-X.

The person skilled in the art knows how to obtain functional variants. Examples of functional variants are disclosed for the Relaxin A chain in Hossain et al J Biol Chem. 2008 Jun 20; 283(25):17287-97 or in US Pat. publication No. US2011/0130332 and for the Relaxin B chain in Schwabe and Büllesbach (2007) Adv Exp Med Biol. 612:14-25 and Büllesbach and Schwabe J Biol Chem. 2000 Nov 10;275(45):35276-80).

Linker L:

In one embodiment the linker polypeptide L of the aforementioned fusion polypeptides A-L-B consists of a polypeptide which is 6-14 amino acid residues in length. Further preferred are polypeptide linkers L which are 7-13 amino acid residues in length. Further preferred are polypeptide linkers L which are 8-12 amino acid residues in length. Even more preferred are polypeptide linkers L which are 7-11, or 9-11 amino acid residues in length. Even more preferred are polypeptide linkers L which are 9 amino acid residues in length. In a further preferred embodiment, the integer of the length of the polypeptide linker L is selected from the group consisting of the integers 6, 7, 8, 9, 10, 11, 12, 13 and 14.

The amino acid composition of the linker can vary, although a linker exhibiting a low immunogenicity score is preferred. Examples of linkers are well known to those skilled in the art and comprise sequences such as (GGGS)n, (GGSG)n, where n are integers. The linker peptide L can be composed of any amino acid. In a preferred embodiment the linker polypeptide L comprises at least one Gly, Ser, Arg, Cys, Leu and/or Lys residue. In a more preferred embodiment the linker polypeptide L comprises Gly and Ser residues. In a further preferred embodiment the linker peptide L is a glycine-rich linker such as for example peptides comprising the sequence $[GGGGS]_n$ as disclosed in U.S. Patent No. 7,271,149. In other embodiments, a serine-rich linker peptide L is used, as described for example in U.S. Patent No. 5,525,491.

A further preferred embodiment is a linker L which comprises Gly and Ser residues and has a ratio of Gly to Ser of at least 3 to 1.

In a further embodiment the aforementioned linker L comprises at least one attachment site for covalent coupling of a non-proteinaceous polymer half-life extending moiety. In an embodiment of the invention the aforementioned attachment site is a Lys or a Cys residue.

Examples of such linkers are [GlyGlyGlySerGlyGly] (SEQ ID NO: 137),

[GlyGlyGlySerGlyGlyGly] (SEQ ID NO: 138),

[GlyGlyGlySerGlyGlyGlySerGly] (SEQ ID NO: 139),

[GlyGlyGlySerGlyGlyGlySerGlyGlyGlySer] (SEQ ID NO: 140),

[GlyGlyGlySerGlyCysGlyGlySerGly] (SEQ ID NO: 141),

[GlyGlyGlySerGlyGlyGlySerGlyGlyGlySerGlyGly] (SEQ ID NO: 143),

[LysArgSerLeuSerArgLysLysArg] (SEQ ID NO: 144),

[GlyGlyGlySerGlyLysGlyGlySerGly] (SEQ ID NO: 142),

[GlyGlyGlySerGlyGlyGlySerGlyGlyGlySerGly] (SEQ ID NO: 145), and

[GlyGlyGlySerGlyGlyGlySerGlyGlyGly] (SEQ ID NO: 146).

It is contemplated that the optimal linker length and amino acid composition can be determined by routine methods known in the art.

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof having 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions compared to SEQ ID NO: 117,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof having 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions compared to SEQ ID NO:119 and comprising the conserved motif Arg-X-X-X-Arg-X-X-Ile/Val-X, and

L is a linker polypeptide which is 6-14, 7-13, 8-12, 7-11, 9-11, or 9 amino acid residues in length.

In a preferred embodiment the linker polypeptide L of the aforementioned fusion polypeptide A-L-B is 7-11, or 9-11 amino acid residues in length. Even more preferred are polypeptide linkers L which are 9 amino acid residues in length. In a further preferred embodiment, the integer of the length of the polypeptide linker L is selected from the group consisting of the integers 6, 7, 8, 9, 10, 11, 12, 13 and 14. The linker polypeptide L can be composed of any amino acid. In a preferred embodiment the linker polypeptide L is a flexible linker.

In a preferred embodiment the linker polypeptide L comprises at least one Gly, Ser, Arg, Leu, Cys, and/or Lys residue. In a further preferred embodiment the linker polypeptide L is consists of amino acid residues selected from the group of amino acids consisting of Gly, Ser, Arg, Leu, Cys, and Lys residues.

In a more preferred embodiment the linker polypeptide L comprises Gly and Ser residues. In a further preferred embodiment the linker peptide L is a glycine-rich linker such as peptides comprising the sequence $[GGGGS]_n$ as disclosed in U.S. Patent No. 7,271,149. In other embodiments, a serine-rich linker peptide L is used, as described in U.S. Patent No. 5,525,491.

A further preferred embodiment is a linker polypeptide L which comprises Gly and Ser residues and has a ratio of Gly to Ser of at least 2 to 1.

A further preferred embodiment is a linker polypeptide L which comprises Gly and Ser residues and has a ratio of Gly to Ser of at least 3 to 1.

A further preferred embodiment is a linker polypeptide L which comprises Gly and Ser residues and has a ratio of Gly to Ser of at least 1 to 2.

A further preferred embodiment is a linker polypeptide L which comprises Gly and Ser residues and has a ratio of Gly to Ser of at least 1 to 3.

A further preferred embodiment is a linker polypeptide L with the aforementioned preferred length, wherein all but 4 amino acid residues of the linker L consist of Gly and/or Ser residues and the remaining 4 amino acid residues are selected from the group of natural amino acids.

A further preferred embodiment is a linker polypeptide L with the aforementioned preferred length, wherein all but 3 amino acid residues of the linker L consist of Gly and/or Ser residues and the remaining 3 amino acid residues are selected from the group of natural amino acids.

A further preferred embodiment is a linker polypeptide L with the aforementioned preferred length, wherein all but 2 amino acids residues of the linker L consist of Gly and/or Ser residues and the remaining 2 amino acid residues are selected from the group of natural amino acids.

A further preferred embodiment is a linker polypeptide L with the aforementioned preferred length, wherein all but 1 amino acid residues of the linker L consist of Gly and/or Ser residues and the remaining amino acid residue is selected from the group of natural amino acids.

In a further preferred embodiment the aforementioned group of natural amino acids excludes the amino acid prolin.

A further preferred embodiment is a linker polypeptide L with the aforementioned preferred length, wherein all but 1 amino acid residues of the linker L consist of Gly and/or Ser and the remaining amino acid is selected from the group of Cys and Lys.

In a further preferred embodiment the linker polypeptide L consists of amino acid residues selected from the group of amino acid residues consisting of Gly and Ser residues.

In a further preferred embodiment the linker L consists of amino acid residues selected from the group of amino acids consisting of Gly and Ser residues wherein the ratio of Gly to Ser is at least 2 to 1.

In a further preferred embodiment the linker L consists of amino acid residues selected from the group of amino acids consisting of Gly and Ser residues wherein the ratio of Gly to Ser is at least 3 to 1.

In a further preferred embodiment the linker L consists of amino acid residues selected from the group of amino acids consisting of Gly and Ser residues wherein the ratio of Gly to Ser is at least 1 to 2.

In a further preferred embodiment the linker L consists of amino acid residues selected from the group of amino acids consisting of Gly and Ser residues wherein the ratio of Gly to Ser is at least 1 to 3.

In a further embodiment the aforementioned linker L comprises at least one attachment site for covalent coupling of a nonproteinaceous polymer half-life extending moiety. In an embodiment of the invention the aforementioned attachment site is a Lys or a Cys residue.

Preferred linker polypeptides L are selected from the group of linker polypeptides consisting of [GlyGlyGlySerGlyGly] (SEQ ID NO: 137),

[GlyGlyGlySerGlyGlyGly] (SEQ ID NO: 138),

[GlyGlyGlySerGlyGlyGlySerGly] (SEQ ID NO: 139),

[GlyGlyGlySerGlyGlyGlySerGlyGlyGlySer] (SEQ ID NO: 140),

[GlyGlyGlySerGlyCysGlyGlySerGly] (SEQ ID NO: 141),

[GlyGlyGlySerGlyGlyGlySerGlyGlyGlySerGlyGly] (SEQ ID NO: 143),

[LysArgSerLeuSerArgLysLysArg] (SEQ ID NO: 144),

[GlyGlyGlySerGlyLysGlyGlySerGly] (SEQ ID NO: 142),

[GlyGlyGlySerGlyGlyGlySerGlyGlyGlySerGly] (SEQ ID NO: 145), and

[GlyGlyGlySerGlyGlyGlySerGlyGlyGly] (SEQ ID NO: 146).

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof, and

L is a linker polypeptide, which is 7, 8, 9 or 10 amino acids in length.

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof having 1, 2, 3 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions compared to SEQ ID NO: 117,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof having 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions compared to SEQ ID NO:119 and comprising the conserved motif Arg-X-X-X-Arg-X-X-Ile/Val-X, and

L is a linker polypeptide, which is 7, 8, 9 or 10 amino acids in length.

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117),

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119), and

L is a linker polypeptide, which is 7, 8, 9 or 10 amino acids in length.

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof, and

L is a linker polypeptide, which is 9 amino acids in length.

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof having 1, 2, 3 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions compared to SEQ ID NO: 117,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof having 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions compared to SEQ ID NO:119 and comprising the conserved motif Arg-X-X-X-Arg-X-X-Ile/Val-X, and

L is a linker polypeptide, which is 9 amino acids in length.

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117),

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119), and

L is a linker polypeptide, which is 9 amino acids in length.

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof, and

L is a linker polypeptide, which is 7, 8, 9 or 10 amino acids in length and which comprises Glycin and Serin residues in a ratio of at least 3:1.

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof having 1, 2, 3 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions compared to SEQ ID NO: 117,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof having 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions compared to SEQ ID NO:119 and comprising the conserved motif Arg-X-X-X-Arg-X-X-Ile/Val-X, and

L is a linker polypeptide, which is 7, 8, 9 or 10 amino acids in length and which comprises Glycin and Serin residues in a ratio of at least 3:1.

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117),

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119), and

L is a linker polypeptide, which is 7, 8, 9 or 10 amino acids in length and which comprises Glycin and Serin residues in a ratio of at least 3:1.

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof, and

L is a linker polypeptide, which is 9 amino acids in length and which comprises Glycin and Serin residues in a ratio of at least 3:1.

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof having 1, 2, 3 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions compared to SEQ ID NO: 117,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof having 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions compared to SEQ ID NO:119 and comprising the conserved motif Arg-X-X-X-Arg-X-X-Ile/Val-X, and

L is a linker polypeptide, which is 9 amino acids in length and which comprises Glycin and Serin residues in a ratio of at least 3:1.

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117),

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119), and

L is a linker polypeptide, which is 9 amino acids in length and which comprises Glycin and Serin residues in a ratio of at least 3:1.

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof, and

L is a linker polypeptide, which has the sequence GlyGlyGlySerGlyGlyGlySerGly (SEQ ID NO: 139).

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof having 1, 2, 3 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions compared to SEQ ID NO: 117,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof having 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions compared to SEQ ID NO:119 and comprising the conserved motif Arg-X-X-X-Arg-X-X-Ile/Val-X, and

L is a linker polypeptide, which has the sequence GlyGlyGlySerGlyGlyGlySerGly (SEQ ID NO: 139).

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117),

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119), and

L is a linker polypeptide, which has the sequence GlyGlyGlySerGlyGlyGlySerGly (SEQ ID NO: 139).

A more preferred embodiment of the invention is a fusion polypeptide comprising the sequence of scR4 (SEQ ID NO: 4).

A more preferred embodiment of the invention is a fusion polypeptide comprising the sequence of scR4 w/o Tag (SEQ ID NO: 45).

A preferred embodiment of the invention is a fusion polypeptide comprising A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof having 1, 2, 3 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions compared to SEQ ID NO: 117,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof having 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid substitutions compared to SEQ ID NO:119 and comprising the conserved motif Arg-X-X-X-Arg-X-X-Ile/Val-X, and

L is a linker polypeptide, selected from the group of linker peptides consisting of linkers having the amino acid sequence of SEQ ID NO: 137 – 146.

The linker length can be between 6 and 14 of amino acids while longer linker peptides that themselves mediate additional functions are conceivable.

In a further embodiment the aforementioned fusion polypeptides A-L-B have Relaxin activity. In a further preferred embodiment the Relaxin activity is activation of the relaxin receptor LGR7. Methods for determining Relaxin activity are known in the art or are provided herein. In an even further preferred embodiment, the activation of the relaxin receptor LGR7 is determined by a method disclosed in experimental methods herein. In an even further preferred embodiment, the determination of the activation of the Relaxin receptor LGR7 is determining an EC_{50} value. In an even more preferred embodiment the aforementioned Relaxin activity is less than 10^5 fold, 10^4 fold, 10^3 fold, 100 fold, 75 fold, 50 fold, 25 fold or 10 fold lower compared to the corresponding wild type Relaxin effective concentration inducing a half maximal activity. For example, the corresponding wild type Relaxin for a fusion polypeptide A-L-B based on human Relaxin 2 is the human Relaxin 2 protein.

Improvement of the biological half life of single chain Relaxin variants

The improvement of the half-life of a fusion polypeptide of the invention can be achieved by adding a half-life extending moiety.

In an embodiment of the invention the aforementioned fusion polypeptide A-L-B further comprise at least one half-life extending moiety. In one embodiment the half-life extending moieties are proteinaceous or non-proteinaceous polymers.

Half-life extension via non-proteinaceous polymer half-life extending moieties:

Improving the biological half-life of a fusion polypeptide A-L-B can be achieved by a non-proteinaceous polymer half-life extending moiety which is covalently coupled to a stretcher polypeptide comprising an attachment site for a non-proteinaceous polymer half-life extending moiety fused to the N- and/or C-terminus of A-L-B. Methods attaching such moieties are known in the art.

Non-proteinaceous polymer half-life extending moieties can be covalently coupled to an attachment site of the fusion polypeptide A-L-B. An attachment site can be either within A, L or B or added by a polypeptide comprising such attachment site recombinantly fused to the N-terminus and/or C-terminus of to the aforementioned fusion polypeptides A-L-B. Preferred is a coupling via the linker polypeptide L, or N- and/or C-terminally to the fusion polypeptide A-L-B fused stretcher comprising an attachment site. An attachment site can be an attachment amino acid, for example Cys or Lys, or a sugar moiety of a carbohydrate.

The non-proteinaceous polymer molecule to be coupled to the variant polypeptide may be any suitable polymer molecule, such as a natural or synthetic homo-polymer or hetero-polymer, typically with a molecular weight in the range of about 300-100,000 Da, such as about 500-20,000 Da, more preferably in the range of about 500-15,000 Da, even more preferably in the range of about 2-12 kDa, such as in the range of about 3-10 kDa. When the term "about" is used herein in connection with a certain molecular weight, the word "about" indicates an approximate average molecular weight and reflects the fact that there will normally be a certain molecular weight distribution in a given polymer preparation. Examples of homo-polymers include a polyol (i.e. poly-OH), a polyamine (i.e. poly-NH2) and a polycarboxylic acid (i.e. poly-COOH). A hetero-polymer is a polymer comprising different coupling groups, such as a hydroxyl group and an amine group.

Examples of suitable polymer molecules include polymer molecules selected from the group consisting of polyalkylene oxide (PAO), including polyalkylene glycol (PAG), such as polyethylene glycol (PEG) and polypropylene glycol (PPG), branched PEGs, hydroxyalkyl starch (HAS), such as hydroxyethyl starch (HES), polysialic acid (PSA), poly-vinyl alcohol (PVA), poly-carboxylate, poly-(vinylpyrolidone), polyethylene-co-maleic acid anhydride, polystyrene-co-maleic acid anhydride, dextran, including carboxymethyl-dextran, or any other biopolymer suitable for reducing immunogenicity and/or increasing functional in vivo half-life and/or serum half-life. Another example of a polymer molecule is human albumin or another abundant plasma protein. Generally, polyalkylene glycol-derived polymers are biocompatible, non-toxic, non-antigenic, non-immunogenic, have various water solubility properties, and are easily excreted from living organisms.

PEG is the preferred polymer molecule, since it has only few reactive groups capable of cross-linking compared to, e.g., polysaccharides such as dextran. In particular, monofunctional PEG, e.g. methoxypolyethylene glycol (mPEG), is of interest since its coupling chemistry is relatively simple (only one reactive group is available for conjugating with attachment groups on the polypeptide). Consequently, as the risk of cross-linking is eliminated, the resulting conjugated fusion polypeptides of the invention are more homogeneous and the reaction of the polymer molecules with the variant polypeptide is easier to control.

To effect covalent attachment of the polymer molecule(s) to the fusion polypeptides of the invention, the hydroxyl end groups of the polymer molecule must be provided in activated form, i.e. with reactive functional groups (examples of which include primary amino groups, hydrazide (HZ), thiol, succinate (SUC), succinimidyl succinate (SS), succinimidyl succinamide (SSA), succinimidyl propionate (SPA), succinimidyl butyrate (SBA), succinimidy carboxymethylate (SCM), benzotriazole carbonate (BTC), N-hydroxysuccinmide (NHS),

aldehyde, nitrophenylcarbonate (NPC), and tresylate (TRES)). Suitable activated polymer molecules are commercially available, e.g. from Shearwater Polymers, Inc., Huntsville, AL, USA, or from PolyMASC Pharmaceuticals plc, UK.

Alternatively, the polymer molecules can be activated by conventional methods known in the art, e.g. as disclosed in WO 90/13540. Specific examples of activated linear or branched polymer molecules for use in the present invention are described in the Shearwater Polymers, Inc. 1997 and 2000 Catalogs (Functionalized Biocompatible Polymers for Research and pharmaceuticals, Polyethylene Glycol and Derivatives, incorporated herein by reference). Specific examples of activated PEG polymers include the following linear PEGs: NHS-PEG (e.g. SPA-PEG, SSPA-PEG, SBA-PEG, SS-PEG, SSA-PEG, SC-PEG, SG-PEG, and SCM-PEG), and NOR-PEG, BTC-PEG, EPOXPEG, NCO-PEG, NPC-PEG, CDI-PEG, ALD-PEG, TRES-PEG, VS-PEG, IODO-PEG, and MAL-PEG, and branched PEGs such as PEG2-NHS and those disclosed in US 5,932,462 and US 5,643,575, both of which are incorporated herein by reference. Furthermore, the following publications disclose useful polymer molecules and/or PEGylation chemistries: US 5,824,778, US 5,476,653, WO 97/32607, EP 229,108, EP 402,378, US 4,902,502, US 5,281,698, US 5,122,614, US 5,219,564, WO 92/16555, WO 94/04193, WO 94/14758, WO 94/17039, WO 94/18247, WO 94/28024, WO 95/00162, WO 95/11924, WO95/13090, WO 95/33490, WO 96/00080, WO 97/18832, WO 98/41562, WO 98/48837, WO 99/32134, WO 99/32139, WO 99/32140, WO 96/40791, WO 98/32466, WO 95/06058, EP 439 508, WO 97/03106, WO 96/21469, WO 95/13312, EP 921 131, US 5,736,625, WO 98/05363, EP 809 996, US 5,629,384, WO 96/41813, WO 96/07670, US 5,473,034, US 5,516,673, EP 605 963, US 5,382,657, EP 510 356, EP 400 472, EP 183 503 and EP 154 316.

Specific examples of activated PEG polymers particularly preferred for coupling to cysteine residues, include the following linear PEGs: vinylsulfone-PEG (VS-PEG), preferably vinylsulfone-mPEG (VS-mPEG); maleimide-PEG (MAL-PEG), preferably maleimide-mPEG (MAL-mPEG) and orthopyridyl-disulfide-PEG (OPSS-PEG), preferably orthopyridyl-disulfide-mPEG (OPSS-mPEG). Typically, such PEG or mPEG polymers will have a size of about 5 kDa, about 10 kD, about 12 kDa or about 20 kDa.

The conjugation of the fusion polypeptides of the invention and the activated polymer molecules is conducted by use of any conventional method, e.g. as described in the following references (which also describe suitable methods for activation of polymer molecules): Harris and Zalipsky, eds., Poly(ethylene glycol) Chemistry and Biological Applications, AZC Washington; R.F. Taylor, (1991), "Protein immobilisation. Fundamental and applications", Marcel Dekker, N.Y.; S.S. Wong, (1992), "Chemistry of Protein Conjugation and

Crosslinking", CRC Press, Boca Raton; G.T. Hermanson et al., (1993), "Immobilized Affinity Ligand Techniques", Academic Press, N.Y.).

The skilled person will be aware that the activation method and/or conjugation chemistry to be used depends on the attachment group(s) of the fusion polypeptide (examples of which are given further above), as well as the functional groups of the polymer (e.g. being amine, hydroxyl, carboxyl, aldehyde, sulfydryl, succinimidyl, maleimide, vinysulfone or haloacetate). The PEGylation may be directed towards conjugation to all available attachment groups on the fusion polypeptide (i.e. such attachment groups that are exposed at the surface of the polypeptide) or may be directed towards one or more specific attachment groups, e.g. the N-terminal amino group as described in US 5,985,265 or to cysteine residues. Furthermore, the conjugation may be achieved in one step or in a stepwise manner (e.g. as described in WO 99/55377).

For PEGylation to cysteine residues (see above) the fusion polypeptide is usually treated with a reducing agent, such as dithiothreitol (DDT) prior to PEGylation. The reducing agent is subsequently removed by any conventional method, such as by desalting. Conjugation of PEG to a cysteine residue typically takes place in a suitable buffer at pH 6-9 at temperatures varying from 4°C to 25°C for periods up to 16 hours.

It will be understood that the PEGylation is designed so as to produce the optimal molecule with respect to the number of PEG molecules attached, the size and form of such molecules (e.g. whether they are linear or branched), and the attachment site(s) in the fusion polypeptide. The molecular weight of the polymer to be used may e.g. be chosen on the basis of the desired effect to be achieved.

In connection with conjugation to only a single attachment group on the protein (e.g. the N-terminal amino group), it may be advantageous that the polymer molecule, which may be linear or branched, has a high molecular weight, preferably about 10-25 kDa, such as about 15-25 kDa, e.g. about 20 kDa.

Normally, the polymer conjugation is performed under conditions aimed at reacting as many of the available polymer attachment groups with polymer molecules. This is achieved by means of a suitable molar excess of the polymer relative to the polypeptide. Typically, the molar ratios of activated polymer molecules to polypeptide are up to about 1000-1, such as up to about 200-1, or up to about 100-1. In some cases the ratio may be somewhat lower, however, such as up to about 50-1, 10-1, 5-1, 2-1 or 1-1 in order to obtain optimal reaction.

It is also contemplated according to the invention to couple the polymer molecules to the polypeptide through a linker. Suitable linkers are well known to the skilled person. A preferred example is cyanuric chloride (Abuchowski et al., (1977), J. Biol. Chem., 252, 3578-3581; US 4,179,337; Shafer et al., (1986), J. Polym. Sci. Polym. Chem. Ed., 24, 375-378).

Subsequent to the conjugation, residual activated polymer molecules are blocked according to methods known in the art, e.g. by addition of primary amine to the reaction mixture, and the resulting inactivated polymer molecules are removed by a suitable method.

It will be understood that depending on the circumstances, e.g. the amino acid sequence of the fusion polypeptide, the nature of the activated PEG compound being used and the specific PEGylation conditions, including the molar ratio of PEG to polypeptide, varying degrees of PEGylation may be obtained, with a higher degree of PEGylation generally being obtained with a higher ratio of PEG to fusion polypeptide. The PEGylated fusion polypeptides resulting from any given PEGylation process will, however, normally comprise a stochastic distribution of conjugated fusion polypeptide having slightly different degrees of PEGylation.

For improvement of the biological half life of Relaxin or of fusion polypeptides of the invention, chemical modification such as PEGylation, or HESylation are applicable.

HAS and HES non-proteinaceous polymers, as well as methods of producing HAS or HES conjugates are disclosed for example in WO02/080979, WO03/070772, WO057092391 and WO057092390.

Polysialytion is another technology, which uses the natural polymer polysialic acid (PSA) to prolong the half-life and improve the stability of therapeutic peptides and proteins. PSA is a polymer of sialic acid (a sugar). When used for protein and therapeutic peptide drug delivery, polysialic acid provides a protective microenvironment on conjugation. This increases the active life of the therapeutic protein in the circulation and prevents it from being recognized by the immune system. The PSA polymer is naturally found in the human body. It was adopted by certain bacteria which evolved over millions of years to coat their walls with it. These naturally potysialylated bacteria were then able, by virtue of molecular mimicry, to foil the body's defence system. PSA, nature's ultimate stealth technology, can be easily produced from such bacteria in large quantities and with predetermined physical characteristics. Bacterial PSA is completely non-immunogenic, even when coupled to proteins, as it is chemically identical to PSA in the human body.

Half-life extension via proteinaceous half-life extending moieties:

A further possibility improving the half-life of a fusion polypeptide A-L-B is a fusion with a proteinaceous half-life extending moiety, such as the immunoglobulin Fc fragment of antibodies, transferrin, transferrin receptor or at least the transferrin-binding portion thereof, serum albumin, or variants thereof or binding modules that bind in-vivo to other molecules mediating longer half-life, e.g. serum albumin binding protein is a commonly used method.

The scRelaxin polypeptides described above can be fused directly or via a peptide linker to the Fc portion of an immunoglobulin. "Immunoglobulins" are molecules containing polypeptide chains held together by disulfide bonds, typically having two light chains and two heavy chains. In each chain, one domain (V) has a variable amino acid sequence depending on the antibody specificity of the molecule. The other domains (C) have a rather constant sequence common to molecules of the same class.

As used herein, the "Fc" portion of an immunoglobulin has the meaning commonly given to the term in the field of immunology. Specifically, this term refers to an antibody fragment that is obtained by removing the two antigen binding regions (the Fab fragments) from the antibody. One way to remove the Fab fragments is to digest the immunoglobulin with papain protease. Thus, the Fc portion is formed from approximately equal sized fragments of the constant region from both heavy chains, which associate through non-covalent interactions and disulfide bonds. The Fc portion can include the hinge regions and extend through the CH2 and CH3 domains to the C-terminus of the antibody. Representative hinge regions for human and mouse immunoglobulins can be found in Antibody Engineering, A Practical Guide, Borrebaeck, C.A.K., ed., W.H. Freeman and Co., 1992.

There are five types of human immunoglobulin Fc regions with different effector and pharmacokinetic properties: IgG, IgA, IgM, IgD, and IgE. IgG is the most abundant immunoglobulin in serum. IgG also has the longest half-life in serum of any immunoglobulin (23 days). Unlike other immunoglobulins, IgG is efficiently recirculated following binding to an Fc receptor. There are four IgG subclasses G1, G2, G3, and G4, each of which have different effect or functions. These effector functions are generally mediated through interaction with the Fc receptor (FcγR) or by binding C1q and fixing complement. Binding to FcγR can lead to antibody dependent cell mediated cytolysis, whereas binding to complement factors can lead to complement mediated cell lysis. In designing heterologous Fc fusion proteins wherein the Fc portion is being utilized solely for its ability to extend half-life, it is important to minimize any effector function. All IgG subclasses are capable of binding to Fc receptors (CD16, CD32, CD64) with G1 and G3 being more effective than G2 and G4. The Fc receptor binding region of IgG is formed by residues located in both the hinge and the carboxy terminal regions of the CH2 domain.

Depending on the desired in vivo effect, the heterologous fusion proteins of the present invention may contain any of the isotypes described above or may contain mutated Fc regions wherein the complement and/or Fc receptor binding functions have been altered. Thus, the heterologous fusion proteins of the present invention may contain the entire Fc portion of an immunoglobulin, fragments of the Fc portion of an immunoglobulin, or analogs thereof fused to a scRelaxin compound.

Regardless of the final structure of the fusion protein, the Fc or Fc-like region must serve to prolong the in vivo plasma half-life of the scRelaxin compound fused at the C-terminus or N-terminus. Preferrably, the fused scRelaxin compound retains some biological activity. Biological activity can be determined by in vitro and in vivo methods known in the art.

It is preferable that the Fc region used for the heterologous fusion proteins of the present invention be derived from an IgG1 or an IgG2 Fc region.

Generally, the Fc region used for the heterologous fusion proteins of the present invention can be derived from any species including but not limited to human, rat, mouse and pig. Preferably, the Fc region used for the present invention is derived from human or rat. However, most preferred are human Fc regions and fragments and variants thereof to reduce the risk of the fusion protein being immunogenic in humans. A "native sequence Fc region" comprises an amino acid sequence identical to the amino acid sequence of an Fc region found in nature. A "variant Fc region" comprises an amino acid sequence which differs from that of a native sequence Fc region by virtue of at least one amino acid modification. Preferably, the variant Fc region has at least one amino acid substitution compared to a native sequence Fc region or to the Fc region of a parent polypeptide, e.g., from about one to about ten amino acid substitutions, and preferably from about one to about five amino acid substitutions in a native sequence Fc region or in the Fc region of the parent polypeptide. The variant Fc region herein will preferably possess at least about 80% sequence identity with a native sequence Fc region and/or with an Fc region of a parent polypeptide, and most preferably at least about 90% sequence identity therewith, more preferably at least about 95% sequence identity therewith.

The scRelaxin compounds described above can be fused directly or via a peptide stretcher to albumin or an analog, fragment, or derivative thereof. Generally the albumin proteins making up part of the fusion proteins of the present invention can be derived from albumin cloned from any species. However, human albumin and fragments and analogs thereof are preferred to reduce the risk of the fusion protein being immunogenic in humans. Human serum albumin (HSA) consists of a single non-glycosylated polypeptide chain of 585 amino acids with a formula molecular weight of 66,500. The amino acid sequence of HSA (SEQ ID NO:123) has been described e.g. in Meloun, et al. (1975); Behrens, et al. (1975); Lawn, et al. (1981) and

Minghetti, et al. (1986). A variety of polymorphic variants as well as analogs and fragments of albumin have been described (see Weitkamp, et al. (1973)). For example, in EP0322094 and EP0399666 various fragments of human serum albumin are disclosed. It is understood that the heterologous fusion proteins of the present invention include scRelaxin compounds that are coupled to any albumin protein including fragments, analogs, and derivatives wherein such fusion protein is biologically active and has a longer plasma half-life than the scRelaxin compound alone. Thus, the albumin portion of the fusion protein need not necessarily have a plasma half-life equal to that of native human albumin. Fragments, analogs, and derivatives are known or can be generated that have longer half-lives or have half-lives intermediate to that of native human albumin and the scRelaxin compound of interest. The techniques are well-known in the art, see, e.g., WO 93/15199, WO 93/15200, WO 01/77137 and EP0413622.

In an embodiment of the invention the proteinaceous half-life extending moiety has low immunogenicity, is human or humanized. In a preferred embodiment the proteinaceous half-life extending moiety is human, such as human transferrin (SEQ ID NO: 122), human serum albumin (SEQ ID NO: 123), or human IgG1 Fc (SEQ ID NO: 120).

Additionally, other proteins, protein domains or peptides improving the biological half life can also be used as fusion partners.

Half-life extension via fusion to human serum albumin is disclosed for example in WO93/15199. Albumin binding as a general strategy for improving the pharmacokinetics of proteins is described for example in Dennis et al., The Journal of Biological Chemistry, Vol. 277, No 38, Issue of September 20, pp. 35035-35043. Half-life extension via fusion to human serum albumin binding proteins is disclosed for example in US20100104588. Half-life extension via fusion to human serum albumin or IgG-Fc binding proteins is disclosed for example in WO01/45746. A further example of half-life extension via fusion to human serum albumin binding peptides is disclosed in WO2010/054699.

Half-life extension via fusion to an Fc domain is disclosed for example in WO2001/058957.

The biological activity determines the preferred orientation of the protein of interest to its fusion partner. C-terminal as well as N-terminal orientations of fusion partners are included. In addition, for improvement of the biological half life or other functions, fusion partners may be modified by phosphorylation, sulfation, acrylation, glycosylation, deglycosylation, methylation, farnesylation, acetylation, amidation or others.

Proteinaceous half-life extending moieties are recombinantly fused to the N-terminus and/or C-terminus of the aforementioned fusion polypeptides A-L-B. The fusion can be with or without

an additional stretcher polypeptide. Examples of proteinaceous half-life extending moieties are transferrin, transferrin receptor or at least the transferrin-binding portion thereof, serum albumin, serum albumin binding proteins, Immunglobulins, and the Fc domain of an immunoglobulin. Preferred are human proteinaceous half-life extending moieties, e.g human transferrin, human transferrin receptor or at least the transferrin-binding portion thereof, human serum albumin, human immunoglobulin or human Fc domains. Fusion partners are linked either directly or by a stretch of amino acids, also termed stretcher. The fusion junction is defined as the position between the last C-terminal amino acid of the first protein or peptide and the first N-terminal amino acid of the second protein or peptide in a fusion protein. Accordingly, a fusion junction or stretcher includes any amino acid between the last amino acid the N-terminal fusion partner and the first amino acid of the C-terminal fusion partner.

Stretcher Units:

Such stretchers are known in the art and are 1 to about 100 amino acids in length, are 1 to about 50 amino acids in length, are 1 to about 25 amino acids in length, are 1 to about 15 amino acids in length, are 1 to 10 amino acids in length, are 4 to 25 amino acids in length, are 4 to 20 amino acids in length, are 4 to 15 amino acids in length, or are 4 to 10 amino acids in length.

The amino acid composition of stretcher sequences is variable, although a stretcher exhibiting a low immunogenicity score is preferred. In an embodiment of the invention a strecher polypeptide connecting a fusion polypeptide A-L-B with a proteinaceous half-life extending moiety can be composed of any amino acid. As shown for example the stretcher polypeptide employed in scR-Fc1 is composed of charged and bulky amino acids (e.g. Glu, Arg or Asp) whereas the stretcher polypeptide in scR-Fc2 is composed of uncharged amino acids (e.g. Gly and Ser).

In a preferred embodiment the stretcher polypeptide comprises at least one Gly, Ser, Ile, Glu, Arg, Met, and/or Asp residue. In a more preferred embodiment the stretcher polypeptide comprises Gly and Ser residues. In a further preferred embodiment the stretcher peptide is a glycine-rich linker such as peptides comprising the sequence $[GGGGS]_n$ as disclosed in U.S. Patent No. 7,271,149. In other embodiments, a serine-rich strecher polypeptide is used, as described in U.S. Patent No. 5,525,491. A further preferred embodiment is a stretcher polypeptide which comprises Gly and Ser residues and has a ratio of Gly to Ser of at least 3 to 1. Further preferred are stretcher polypeptides having a Prolin residue at the C- and/or N-terminal end.

Preferred stretcher peptides are [GlyGlySerPro] (SEQ ID NO: 148),

[GlyGlySerGlyGlySerPro] (SEQ ID NO: 149), and

[GlyGlySerGlyGlySerGlyGlySerPro] (SEQ ID NO: 150).

Such fusion polypeptides with improved half-life can be represented by fusion polypeptide comprising the sequence $(R1)_m$-$(S1)_n$-A-L-B-$(S2)_o$-$(R2)_p$.

A further embodiment of the invention is a fusion polypeptide comprising

$(R1)_m$-$(S1)_n$-A-L-B-$(S2)_o$-$(R2)_p$, wherein

A, L and B have the definitions as disclosed above,

R1 and R2 are proteinaceous half-life extending moieties,

S1 and S2 are stretcher peptides as defined above,

and wherein m, n, o, and p independently have the integer 0 or 1 , provided that at least one of m, n, o, and p are 1. For example, $(S1)_{n=0}$ means that no linker S1 is present in the fusion polypeptide.

In a further embodiment n has the integer 1 if m has the integer 1. In a further embodiment o has the integer 1 if p has the integer 1.

In a preferred embodiment n and m are 0 and o and p are 1. In a further preferred embodiment n and m are 1 and o and p are 0.

A further embodiment of the invention is a fusion polypeptide comprising

$(R1)_{m=1}$-$(S1)_{n=0}$-A-L-B-$(S2)_{o=0}$-$(R2)_{p=0}$.

A further embodiment of the invention is a fusion polypeptide comprising

$(R1)_{m=0}$-$(S1)_{n=0}$-A-L-B-$(S2)_{o=0}$-$(R2)_{p=1}$.

In a preferred embodiment the proteinaceous half-life extending moiety is selected form the group consisting of serum albumin, transferrin, Fc domain, IgG1 Fc domain, and serum albumin binding protein.

In a further embodiment the aforementioned fusion polypeptides further comprising at least one half-life extending moiety have an extended half-life compared to the corresponding wild type Relaxin, wherein the half-life extension is at least 5, 10, 20, 50, 100 or 500-fold. Preferably, the half-life is determined as serum half-life, meaning detection of the fusion protein in serum or

whole blood, for example by using a commercially available quantification ELISA assay (e.g. R&D Systems, Human Relaxin-2 Quantikine ELISA kit, catalogue number DRL200). The half-life is preferably a human blood half-life. Preferably, the half-life is determined as functional in vivo half-life, meaning the activity of fusion polypeptide in serum or blood samples is determined. Assays to determine the activity of a fusion polypeptide A-L-B of the invention are known in the art and are described herein.

A preferred embodiment of the invention is a fusion polypeptide comprising

$(R1)_m$-$(S1)_n$-A-L-B-$(S2)_o$-$(R2)_p$, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof,

L is a linker polypeptide, which is 9 amino acids in length,

R1 and R2 are half-life extending moieties, preferably proteinaceous half-life extending moieties,

S1 and S2 are stretcher peptides as defined above,

and wherein m, n, o, and p independently have the integer 0 or 1, provided that at least one of m, n, o, and p are 1, preferably at least m or p is 1, more preferably m and n are 0 and o and p are 1, and most preferably m and n are 1 and o and p are 0.

A preferred embodiment of the invention is a fusion polypeptide comprising

$(R1)_m$-$(S1)_n$-A-L-B-$(S2)_o$-$(R2)_p$, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117),

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119),

L is a linker polypeptide, which is 9 amino acids in length,

R1 and R2 are half-life extending moieties, preferably proteinaceous half-life extending moieties,

S1 and S2 are stretcher peptides as defined above,

and wherein m, n, o, and p independently have the integer 0 or 1, provided that at least one of m, n, o, and p are 1, preferably at least m or p is 1, more preferably m and n are 0 and o and p are 1, and most preferably m and n are 1 and o and p are 0.

A preferred embodiment of the invention is a fusion polypeptide comprising

$(R1)_m$-$(S1)_n$-A-L-B-$(S2)_o$-$(R2)_p$, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof,

L is a linker polypeptide, which has the sequence GlyGlyGlySerGlyGlyGlySerGly (SEQ ID NO: 139),

R1 and R2 are half-life extending moieties, preferably proteinaceous half-life extending moieties,

S1 and S2 are stretcher peptides as defined above,

and wherein m, n, o, and p independently have the integer 0 or 1, provided that at least one of m, n, o, and p are 1, preferably at least m or p is 1, more preferably m and n are 0 and o and p are 1, and most preferably m and n are 1 and o and p are 0.

A preferred embodiment of the invention is a fusion polypeptide comprising

$(R1)_m$-$(S1)_n$-A-L-B-$(S2)_o$-$(R2)_p$, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117),

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119),

L is a linker polypeptide, which has the sequence GlyGlyGlySerGlyGlyGlySerGly (SEQ ID NO: 139),

R1 and R2 are half-life extending moieties, preferably proteinaceous half-life extending moieties,

S1 and S2 are stretcher peptides as defined above,

and wherein m, n, o, and p independently have the integer 0 or 1, provided that at least one of m, n, o, and p are 1, preferably at least m or p is 1, more preferably m and n are 0 and o and p are 1, and most preferably m and n are 1 and o and p are 0.

A preferred embodiment of the invention is a fusion polypeptide comprising

(R1)-(S1)-A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof,

L is a linker polypeptide, which has the sequence GlyGlyGlySerGlyGlyGlySerGly (SEQ ID NO: 139),

R1 is a half-life extending moiety, preferably a proteinaceous half-life extending moiety, and

S1 is a stretcher peptide as defined above.

A preferred embodiment of the invention is a fusion polypeptide comprising

(R1)-(S1)-A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117),

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119),

L is a linker polypeptide, which has the sequence GlyGlyGlySerGlyGlyGlySerGly (SEQ ID NO: 139),

R1 is a half-life extending moiety, preferably a proteinaceous half-life extending moiety, and

S1 is a stretcher peptide as defined above.

A preferred embodiment of the invention is a fusion polypeptide comprising

(R1)-(S1)-A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof,

L is a linker polypeptide, which has the sequence GlyGlyGlySerGlyGlyGlySerGly (SEQ ID NO: 139),

R1 is a proteinaceous half-life extending moiety,

S1 is a stretcher peptide being 4 – 10 amino acids in length, preferably selected from the group consisting of GlyGlySerPro (SEQ ID NO: 148),

GlyGlySerGlyGlySerPro (SEQ ID NO: 149), and

GlyGlySerGlyGlySerGlyGlySerPro (SEQ ID NO: 150).

A preferred embodiment of the invention is a fusion polypeptide comprising

(R1)-(S1)-A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117),

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119),

L is a linker polypeptide, which has the sequence GlyGlyGlySerGlyGlyGlySerGly (SEQ ID NO: 139),

R1 is a proteinaceous half-life extending moiety,

S1 is a stretcher peptide being 4 – 10 amino acids in length, preferably selected from the group consisting of GlyGlySerPro (SEQ ID NO: 148),

GlyGlySerGlyGlySerPro (SEQ ID NO: 149), and

GlyGlySerGlyGlySerGlyGlySerPro (SEQ ID NO: 150).

A preferred embodiment of the invention is a fusion polypeptide comprising

(R1)-(S1)-A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof,

L is a linker polypeptide, which has the sequence GlyGlyGlySerGlyGlyGlySerGly (SEQ ID NO: 139),

R1 is a proteinaceous half-life extending moiety,

S1 is a stretcher peptide being 10 amino acids in length.

A preferred embodiment of the invention is a fusion polypeptide comprising

(R1)-(S1)-A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117),

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119),

L is a linker polypeptide, which has the sequence GlyGlyGlySerGlyGlyGlySerGly (SEQ ID NO: 139),

R1 is a proteinaceous half-life extending moiety,

S1 is a stretcher peptide being 10 amino acids in length.

A preferred embodiment of the invention is a fusion polypeptide comprising

(R1)-(S1)-A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof,

L is a linker polypeptide, which has the sequence GlyGlyGlySerGlyGlyGlySerGly (SEQ ID NO: 139),

R1 is a proteinaceous half-life extending moiety,

S1 is a stretcher peptide consisting of GlyGlySerGlyGlySerGlyGlySerPro (SEQ ID NO: 150).

A preferred embodiment of the invention is a fusion polypeptide comprising

(R1)-(S1)-A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117),

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119),

L is a linker polypeptide, which has the sequence GlyGlyGlySerGlyGlyGlySerGly (SEQ ID NO: 139),

R1 is a proteinaceous half-life extending moiety,

S1 is a stretcher peptide consisting of GlyGlySerGlyGlySerGlyGlySerPro (SEQ ID NO: 150).

A preferred embodiment of the invention is a fusion polypeptide comprising

(R1)-(S1)-A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117) or a functional variant thereof,

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119) or a functional variant thereof,

L is a linker polypeptide, which has the sequence GlyGlyGlySerGlyGlyGlySerGly (SEQ ID NO: 139),

R1 is a Fc domain of an antibody, preferably a human IgG1 or IgG2 Fc domain,

S1 is a stretcher peptide consisting of GlyGlySerGlyGlySerGlyGlySerPro (SEQ ID NO: 150).

A preferred embodiment of the invention is a fusion polypeptide comprising

(R1)-(S1)-A-L-B, wherein

A is a human Relaxin 2 A chain polypeptide (SEQ ID NO: 117),

B is a human Relaxin 2 B chain polypeptide (SEQ ID NO: 119),

L is a linker polypeptide, which has the sequence GlyGlyGlySerGlyGlyGlySerGly (SEQ ID NO: 139),

R1 is a Fc domain of an antibody, preferably a human IgG1 or IgG2 Fc domain,

S1 is a stretcher peptide consisting of GlyGlySerGlyGlySerGlyGlySerPro (SEQ ID NO: 150).

A further preferred embodiment of the invention is a fusion polypeptide comprising a polypeptide as set forth in table 3.

A further preferred embodiment of the invention are fusion polypeptides as set forth in table 3.

Table 3:

Construct	SEQ ID NO
scR3	SEQ ID NO:3
scR4	SEQ ID NO:4
scR5	SEQ ID NO:5
scR7	SEQ ID NO:7
scR8	SEQ ID NO:8

Construct	SEQ ID NO
scR9	SEQ ID NO:9
scR10	SEQ ID NO:10
scR11	SEQ ID NO:11
scR12	SEQ ID NO:12
scR13	SEQ ID NO:13
scR14	SEQ ID NO:14
scR15	SEQ ID NO:15
scR-Fc 1	SEQ ID NO:16
scR-Fc 2	SEQ ID NO:17
scR-Fc 3	SEQ ID NO:18
scR-Fc 4	SEQ ID NO:19
scR-Fc 5	SEQ ID NO:20
scR-Fc 6	SEQ ID NO:21
scR-Fc 7	SEQ ID NO:22
scR-Fc 8	SEQ ID NO:23
scR-Fc 9	SEQ ID NO:24
scR-Fc 10	SEQ ID NO:25
scR-Fc 11	SEQ ID NO:26
scR-Fc 12	SEQ ID NO:27
scR-Fc 13	SEQ ID NO:28
scR-Fc 14	SEQ ID NO:29

Construct	SEQ ID NO
scR-Fc 15	SEQ ID NO:30
scR-Fc 16	SEQ ID NO:31
scR-Fc 17	SEQ ID NO:32
scR-Fc 18	SEQ ID NO:33
scR-Var1	SEQ ID NO:34
scR-Var2	SEQ ID NO:35
scR-Var3	SEQ ID NO:36
scR-Var4	SEQ ID NO:37
scR-Var5	SEQ ID NO:38
scR-Var6	SEQ ID NO:39
scR-Var7	SEQ ID NO:40
scR-Var8	SEQ ID NO:41
scR3 w/o Tag	SEQ ID NO:44
scR4 w/o Tag	SEQ ID NO:45
scR5 w/o Tag	SEQ ID NO:46
scR6 w/o Tag	SEQ ID NO:47
scR7 w/o Tag	SEQ ID NO:48
scR8 w/o Tag	SEQ ID NO:49
scR9 w/o Tag	SEQ ID NO:50
scR10 w/o Tag	SEQ ID NO:51
scR-Fc 1 w/o Tag	SEQ ID NO:52

Construct	SEQ ID NO
scR-Fc 8 w/o Tag	SEQ ID NO:53
scR-Fc 9 w/o Tag	SEQ ID NO:54
scR-Fc 10 w/o Tag	SEQ ID NO:55
scR-Fc 11 w/o Tag	SEQ ID NO:56
scR-Fc 12 w/o Tag	SEQ ID NO:57
scR-Fc 13 w/o Tag	SEQ ID NO:58
scR17	SEQ ID NO:153
scR19	SEQ ID NO:155

In a further embodiment the aforementioned fusion polypeptides A-L-B further comprising a half-life extending moiety have Relaxin activity. In a further preferred embodiment the Relaxin activity is activation of the relaxin receptor LGR7. Methods for determining Relaxin activity are known in the art or are provided herein. In an even further preferred embodiment, the activation of the relaxin receptor LGR7 is determined by a method disclosed in experimental methods herein. In an even further preferred embodiment, the determination of the activation of the relaxin receptor LGR7 is determining an EC_{50} value. In an even more preferred embodiment the aforementioned Relaxin activity is less than 10^5 fold, 10^4 fold, 10^3 fold, 100 fold, 75 fold, 50 fold, 25 fold or 10 fold lower compared to the corresponding wild type Relaxin activity. For example, the corresponding wild type Relaxin for a fusion polypeptide A-L-B based on human Relaxin 2 is the human Relaxin 2 protein.

Cloning, vector systems, expression, hosts, and purification

The invention also provides for a vector which comprises an isolated nucleic acid molecule encoding a fusion polypeptide of the invention. This vector system is operatively linked to an expression sequence capable of directing its expression in a host cell.

A suitable host cell may be selected from the group consisting of bacterial cells (such as E. coli), yeast cells (such as Saccharomyces cerevisiae), fungal cells, plant cells, insect cells and animals cells. Animal cells include, but are not limited to, HEK293 cells, CHO cells, COS cells,

BHK cells, HeLa cells and various primary mammalian cells. Derivatives of mammalian cells such as HEK293T cells are also applicable.

DNA molecules of the invention

The present invention also relates to the DNA molecules that encode a fusion protein of the invention. These sequences include, but are not limited to, those DNA molecules set forth in table 4.

Table 4:

Construct	SEQ ID NO
scR1	SEQ ID NO:59
scR2	SEQ ID NO:60
scR3	SEQ ID NO:61
scR4	SEQ ID NO:62
scR5	SEQ ID NO:63
scR6	SEQ ID NO:64
scR7	SEQ ID NO:65
scR8	SEQ ID NO:66
scR9	SEQ ID NO:67
scR10	SEQ ID NO:68
scR11	SEQ ID NO:69
scR12	SEQ ID NO:70
scR13	SEQ ID NO:71
scR14	SEQ ID NO:72
scR15	SEQ ID NO:73

Construct	SEQ ID NO
scR-Fc 1	SEQ ID NO:74
scR-Fc 2	SEQ ID NO:75
scR-Fc 3	SEQ ID NO:76
scR-Fc 4	SEQ ID NO:77
scR-Fc 5	SEQ ID NO:78
scR-Fc 6	SEQ ID NO:79
scR-Fc 7	SEQ ID NO:80
scR-Fc 8	SEQ ID NO:81
scR-Fc 9	SEQ ID NO:82
scR-Fc 10	SEQ ID NO:83
scR-Fc 11	SEQ ID NO:84
scR-Fc 12	SEQ ID NO:85
scR-Fc 13	SEQ ID NO:86
scR-Fc 14	SEQ ID NO:87
scR-Fc 15	SEQ ID NO:88
scR-Fc 16	SEQ ID NO:89
scR-Fc 17	SEQ ID NO:90
scR-Fc 18	SEQ ID NO:91
scR-Var1	SEQ ID NO:92
scR-Var2	SEQ ID NO:93
scR-Var3	SEQ ID NO:94

Construct	SEQ ID NO
scR-Var4	SEQ ID NO:95
scR-Var5	SEQ ID NO:96
scR-Var6	SEQ ID NO:97
scR-Var7	SEQ ID NO:98
scR-Var8	SEQ ID NO:99
scR3 w/o Tag	SEQ ID NO:102
scR4 w/o Tag	SEQ ID NO:103
scR5 w/o Tag	SEQ ID NO:104
scR6 w/o Tag	SEQ ID NO:105
scR7 w/o Tag	SEQ ID NO:106
scR8 w/o Tag	SEQ ID NO:107
scR9 w/o Tag	SEQ ID NO:108
scR10 w/o Tag	SEQ ID NO:109
scR-Fc 1 w/o Tag	SEQ ID NO:110
scR-Fc 8 w/o Tag	SEQ ID NO:111
scR-Fc 9 w/o Tag	SEQ ID NO:112
scR-Fc 10 w/o Tag	SEQ ID NO:113
scR-Fc 11 w/o Tag	SEQ ID NO:114
scR-Fc 12 w/o Tag	SEQ ID NO:115
scR-Fc 13 w/o Tag	SEQ ID NO:116
scR17	SEQ ID NO:158

Construct	SEQ ID NO
scR19	SEQ ID NO:160

DNA molecules of the invention are not limited to the sequences disclosed herein, but also include variants thereof. DNA variants within the invention may be described by reference to their physical properties in hybridization. The skilled worker will recognize that DNA can be used to identify its complement and, since DNA is double stranded, its equivalent or homolog, using nucleic acid hybridization techniques. It also will be recognized that hybridization can occur with less than 100% complementarity. However, given appropriate choice of conditions, hybridization techniques can be used to differentiate among DNA sequences based on their structural relatedness to a particular probe. For guidance regarding such conditions see, Sambrook et al., 1989 supra and Ausubel et al., 1995 (Ausubel, F. M., Brent, R., Kingston, R. E., Moore, D. D., Sedman, J. G., Smith, J. A., & Struhl, K. eds. (1995). Current Protocols in Molecular Biology. New York: John Wiley and Sons).

Structural similarity between two polynucleotide sequences can be expressed as a function of "stringency" of the conditions under which the two sequences will hybridize with one another. As used herein, the term "stringency" refers to the extent that the conditions disfavor hybridization. Stringent conditions strongly disfavor hybridization, and only the most structurally related molecules will hybridize to one another under such conditions. Conversely, non-stringent conditions favor hybridization of molecules displaying a lesser degree of structural relatedness. Hybridization stringency, therefore, directly correlates with the structural relationships of two nucleic acid sequences. The following relationships are useful in correlating hybridization and relatedness (where T_m is the melting temperature of a nucleic acid duplex):

a. $T_m = 69.3 + 0.41(G+C)\%$

b. The T_m of a duplex DNA decreases by 1°C with every increase of 1% in the number of mismatched base pairs.

c. $(T_m)_{\mu 2} - (T_m)_{\mu 1} = 18.5 \log_{10} \mu 2/\mu 1$

where $\mu 1$ and $\mu 2$ are the ionic strengths of two solutions.

Hybridization stringency is a function of many factors, including overall DNA concentration, ionic strength, temperature, probe size and the presence of agents which disrupt hydrogen bonding. Factors promoting hybridization include high DNA concentrations, high ionic

strengths, low temperatures, longer probe size and the absence of agents that disrupt hydrogen bonding. Hybridization typically is performed in two phases: the "binding" phase and the "washing" phase.

First, in the binding phase, the probe is bound to the target under conditions favoring hybridization. Stringency is usually controlled at this stage by altering the temperature. For high stringency, the temperature is usually between 65°C and 70°C, unless short (< 20 nt) oligonucleotide probes are used. A representative hybridization solution comprises 6X SSC, 0.5% SDS, 5X Denhardt's solution and 100 µg of nonspecific carrier DNA. See Ausubel et al., section 2.9, supplement 27 (1994). Of course, many different, yet functionally equivalent, buffer conditions are known. Where the degree of relatedness is lower, a lower temperature may be chosen. Low stringency binding temperatures are between about 25°C and 40°C. Medium stringency is between at least about 40°C to less than about 65°C. High stringency is at least about 65°C.

Second, the excess probe is removed by washing. It is at this phase that more stringent conditions usually are applied. Hence, it is this "washing" stage that is most important in determining relatedness via hybridization. Washing solutions typically contain lower salt concentrations. One exemplary medium stringency solution contains 2X SSC and 0.1% SDS. A high stringency wash solution contains the equivalent (in ionic strength) of less than about 0.2X SSC, with a preferred stringent solution containing about O.1X SSC. The temperatures associated with various stringencies are the same as discussed above for "binding." The washing solution also typically is replaced a number of times during washing. For example, typical high stringency washing conditions comprise washing twice for 30 minutes at 55° C. and three times for 15 minutes at 60° C.

An embodiment of the invention is an isolated nucleic acid sequence that encodes a fusion polypeptide of the invention.

Recombinant DNA constructs and expression

The present invention further provides recombinant DNA constructs comprising one or more of the nucleotide sequences of the present invention. The recombinant constructs of the present invention are used in connection with a vector, such as a plasmid, phagemid, phage or viral vector, into which a DNA molecule encoding a fusion polypeptide of the invention is inserted.

A fusion polypeptide as provided herein can be prepared by recombinant expression of nucleic acid sequences encoding a fusion polypeptide in a host cell. To express a fusion polypeptide recombinantly, a host cell can be transfected with a recombinant expression vectors carrying

DNA fragments encoding a fusion polypeptide such that the fusion polypeptide is expressed in the host cell. Standard recombinant DNA methodologies are used to prepare and/or obtain nucleic acids encoding a fusion polypeptide, incorporate these nucleic acids into recombinant expression vectors and introduce the vectors into host cells, such as those described in Sambrook, Fritsch and Maniatis (eds.), Molecular Cloning; A Laboratory Manual, Second Edition, Cold Spring Harbor, N.Y., (1989), Ausubel, F. M. et al. (eds.) Current Protocols in Molecular Biology, Greene Publishing Associates, (1989) and in U.S. Pat. No. 4,816,397 by Boss et al.

To express the fusion polypeptide standard recombinant DNA expression methods can be used (see, for example, Goeddel; Gene Expression Technology. Methods in Enzymology 185, Academic Press, San Diego, Calif. (1990)). For example, DNA encoding the desired polypeptide can be inserted into an expression vector which is then transfected into a suitable host cell. Suitable host cells are prokaryotic and eukaryotic cells. Examples for prokaryotic host cells are e.g. bacteria, examples for eukaryotic host cells are yeast, insect or mammalian cells. It is understood that the design of the expression vector, including the selection of regulatory sequences is affected by factors such as the choice of the host cell, the level of expression of protein desired and whether expression is constitutive or inducible.

Bacterial Expression

Useful expression vectors for bacterial use are constructed by inserting a structural DNA sequence encoding a desired protein together with suitable translation initiation and termination signals in operable reading phase with a functional promoter. The vector will comprise one or more phenotypic selectable markers and an origin of replication to ensure maintenance of the vector and, if desirable, to provide amplification within the host. Suitable prokaryotic hosts for transformation include E. coli, Bacillus subtilis, Salmonella typhimurium and various species within the genera Pseudomonas, Streptomyces, and Staphylococcus.

Bacterial vectors may be, for example, bacteriophage-, plasmid- or phagemid-based. These vectors can contain a selectable marker and bacterial origin of replication derived from commercially available plasmids typically containing elements of the well known cloning vector pBR322 (ATCC 37017). Following transformation of a suitable host strain and growth of the host strain to an appropriate cell density, the selected promoter is de-repressed/induced by appropriate means (e.g., temperature shift or chemical induction) and cells are cultured for an additional period. Cells are typically harvested by centrifugation, disrupted by physical or chemical means, and the resulting crude extract retained for further purification.

In bacterial systems, a number of expression vectors may be advantageously selected depending upon the use intended for the protein being expressed. For example, when a large quantity of such a protein is to be produced vectors which direct the expression of high levels of fusion polypeptide products that are readily purified may be desirable. Fusion polypeptide of the present invention include purified products, products of chemical synthetic procedures, and products produced by recombinant techniques from a prokaryotic host, including, for example, E. coli, Bacillus subtilis, Salmonella typhimurium and various species within the genera Pseudomonas, Streptomyces, and Staphylococcus, preferably, from E. coli cells.

Eukaryotic Expression

Eukaryotic cells can be used to express the polypeptides of the invention. Systems for expression of proteins are known in the art. Such systems include e.g. include the eukaryotic cell, growth media, and corresponding expression vectors. Common eukaryotic cells for expression are e.g. a mammalian cell, a yeast cell, a plant cell, or an insect cell.

Mammalian Expression and Purification

Preferred regulatory sequences for mammalian host cell expression include viral elements that direct high levels of protein expression in mammalian cells, such as promoters and/or enhancers derived from cytomegalovirus (CMV) (such as the CMV promoter/enhancer), Simian Virus 40 (SV40) (such as the SV40 promoter/enhancer), adenovirus, (e.g., the adenovirus major late promoter (AdMLP)) and polyoma. For further description of viral regulatory elements, and sequences thereof, see e.g., U.S. 5,168,062 by Stinski, U.S. 4,510,245 by Bell et al. and U.S. 4,968,615 by Schaffner et al. The recombinant expression vectors can also include origins of replication and selectable markers (see e.g., U.S. 4,399,216, 4,634,665 and U.S. 5,179,017, by Axel et al.). Suitable selectable markers include genes that confer resistance to drugs such as G418, hygromycin or methotrexate, on a host cell into which the vector has been introduced. For example, the dihydrofolate reductase (DHFR) gene confers resistance to methotrexate and the neo gene confers resistance to G418.

Transfection of the expression vector into a host cell can be carried out using standard techniques such as electroporation, calcium-phosphate precipitation, and DEAE-dextran, lipofection or polycation-mediated transfection.

Suitable mammalian host cells for expressing the fusion polypeptides provided herein include Chinese Hamster Ovary (CHO cells) (including dhfr- CHO cells, described in Urlaub and Chasin, (1980) Proc. Natl. Acad. Sci. USA 77:4216-4220, used with a DHFR selectable marker, e.g., as described in R. J. Kaufman and P. A. Sharp (1982) Mol. Biol. 159:601-621, NSO

myeloma cells, COS cells and SP2 cells. In some embodiments, the expression vector is designed such that the expressed protein is secreted into the culture medium in which the host cells are grown. Transient transfection/epression of antibodies can for example be achieved following the protocols by Durocher et al (2002) Nucl.Acids Res. Vol 30 e9. Stable transfection/expression of antibodies can for example be achieved following the protocols of the UCOE system (T. Benton et al. (2002) Cytotechnology 38: 43–46).

The fusion polypeptide can be recovered from the culture medium using standard protein purification methods.

A fusion polypeptide of the invention can be recovered and purified from recombinant cell cultures by well-known methods including, but not limited to ammonium sulfate or ethanol precipitation, acid extraction, Protein A chromatography, Protein G chromatography, anion or cation exchange chromatography, phospho-cellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography. High performance liquid chromatography ("HPLC") can also be employed for purification. See, e.g., Colligan, Current Protocols in Immunology, or Current Protocols in Protein Science, John Wiley & Sons, NY, N.Y., (1997-2001), e.g., Chapters 1, 4, 6, 8, 9, 10, each entirely incorporated herein by reference.

Fusion polypeptides of the invention include purified or isolated products, products of chemical synthetic procedures, and products produced by recombinant techniques from a eukaryotic host, including, for example, yeast (for example Pichia), higher plant, insect and mammalian cells, preferably from mammalian cells. Depending upon the host employed in a recombinant production procedure, the fusion polypeptide of the present invention can be glycosylated or can be non-glycosylated, with glycosylated preferred. Such methods are described in many standard laboratory manuals, such as Sambrook, supra, Sections 17.37-17.42; Ausubel, supra, Chapters 10, 12, 13, 16, 18 and 20.

Therapeutic Use

An embodiment of the invention is the use of a pharmaceutical composition or a fusion polypeptide of the invention in the treatment of cardiovascular diseases, kidney diseases, pancreatitis, inflammation, cancer, scleroderma, pulmonary, renal, and hepatic fibrosis.

Cardiovascular Diseases

Disorders of the cardiovascular system, or cardiovascular disorders, mean in the context of the present invention for example the following disorders: hypertension (high blood pressure), peripheral and cardiac vascular disorders, coronary heart disease, stable and unstable angina

pectoris, myocardial insufficiency, persistent ischemic dysfunction ("hibernating myocardium"), temporary postischemic dysfunction ("stunned myocardium"), heart failure, disturbances of peripheral blood flow, acute coronary syndrome, heart failure and myocardial infarction.

In the context of the present invention, the term heart failure includes both acute and chronic manifestations of heart failure, as well as more specific or related types of disease, such as acute decompensated heart failure, right heart failure, left heart failure, global failure, ischemic cardiomyopathy, dilated cardiomyopathy, congenital heart defects, heart valve defects, heart failure associated with heart valve defects, mitral stenosis, mitral insufficiency, aortic stenosis, aortic insufficiency, tricuspid stenosis, tricuspid insufficiency, pulmonary stenosis, pulmonary valve insufficiency, combined heart valve defects, myocardial inflammation (myocarditis), chronic myocarditis, acute myocarditis, viral myocarditis, diabetic heart failure, alcoholic cardiomyopathy, cardiac storage disorders, and diastolic and systolic heart failure and acute phases of worsening heart failure.

The compounds according to the invention are further also suitable for reducing the area of myocardium affected by an infarction, and for the prophylaxis of secondary infarctions.

The compounds according to the invention are furthermore suitable for the prophylaxis and/or treatment of thromboembolic disorders, reperfusion damage following ischemia, micro- and macrovascular lesions (vasculitis), arterial and venous thromboses, edemas, ischemias such as myocardial infarction, stroke and transient ischemic attacks, for cardio protection in connection with coronary artery bypass operations (CABG), primary percutaneous transluminal coronary angioplasties (PTCAs), PTCAs after thrombolysis, rescue PTCA, heart transplants and open-heart operations, and for organ protection in connection with transplants, bypass operations, catheter examinations and other surgical procedures.

Other areas of indication are, for example, the prevention and/or treatment of respiratory disorders, such as, for example, chronic obstructive pulmonary disease (chronic bronchitis, COPD), asthma, pulmonary emphysema, bronchiectases, cystic fibrosis (mucoviscidosis) and pulmonary hypertension, in particular pulmonary arterial hypertension.

Kidney disease

The present invention relates to the use of a fusion polypeptide of the invention as a medicament for the prophylaxis and/or treatment of kidney diseases, especially of acute and chronic kidney diseases and acute and chronic renal insufficiencies, as well as acute and chronic renal failure, including acute and chronic stages of renal failure with and without the requirement of dialysis, as well as the underlying or related kidney diseases such as renal

hypoperfusion, dialysis induced hypotension, glomerulopathies, glomerular and tubular proteinuria, renal edema, hematuria, primary, secondary, as well as acute and chronic glomerulonephritis, membranous and membranoproliferative glomerulonephritis, Alport-Syndrome, glomerulosclerosis, interstistial tubular diseases, nephropathic diseases, such as primary and inborn kidney diseases, renal inflammation, immunological renal diseases like renal transplant rejection, immune complex induced renal diseases, as well as intoxication induced nephropathic diseases, diabetic and non-diabetic renal diseases, pyelonephritis, cystic kidneys, nephrosclerosis, hypertensive nephrosclerosis, nephrotic syndrome, that are characterized and diagnostically associated with an abnormal reduction in creatinine clearance and/or water excretion, abnormal increased blood concentrations of urea, nitrogen, potassium and/or creatinine, alteration in the activity of renal enzymes, such as glutamylsynthetase, urine osmolarity and urine volume, increased microalbuminuria, macroalbuminuria, glomerular and arteriolar lesions, tubular dilation, hyperphosphatemia and /or the requirement of dialysis.

In addition, a fusion polypeptide of the invention can be used as a medicament for the prophylaxis and/or treatment of renal carcinomas, after incomplete resection of the kidney, dehydration after overuse of diuretics, uncontrolled blood pressure increase with malignant hypertension, urinary tract obstruction and infection, amyloidosis, as well as systemic diseases associated with glomerular damage, such as Lupus erythematodes, and rheumatic immunological systemic diseases, as well as renal artery stenosis, renal artery thrombosis, renal vein thrombosis, analgetics induced nephropathy and renal tubular acidosis.

In addition, a fusion polypeptide of the invention can be used as a medicament for the prophylaxis and/or treatment of contrast medium induced and drug induced acute and chronic interstitial kidney diseases, metabolic syndrome and dyslipemia.

In addition, the present invention includes the use of a fusion polypeptide of the invention as a medicament for the prophylaxis and/or treatment of aftereffects associated with acute and/or chronic kidney diseases, such as pulmonary edema, heart failure, uremia, anemia, electrolyte disturbances (e.g. hyperkalemia, hyponatremia), as well as bony and carbohydrate metabolism.

Lung Diseases

Furthermore, the fusion polypeptides according to the invention are also suitable for the treatment and/or prophylaxis of lung diseases especially of asthmatic disorders, pulmonary arterial hypertension (PAH) and other forms of pulmonary hypertension (PH) including left-heart disease, HIV, sickle cell anaemia, thromboembolisms (CTEPH), sarkoidosis, COPD or pulmonary fibrosis-associated pulmonary hypertension, chronic-obstructive pulmonary disease (COPD), acute respiratory distress syndrome (ARDS), acute lung injury (ALI), alpha-1-

antitrypsin deficiency (AATD), pulmonary fibrosis, pulmonary emphysema (for example pulmonary emphysema induced by cigarette smoke) and cystic fibrosis (CF).

Fibrotic Disorders

The fusion polypeptides according to the invention are furthermore suitable for the treatment and/or prophylaxis of fibrotic disorders of the internal organs such as, for example, the lung, the heart, the kidney, the bone marrow and in particular the liver, and also dermatological fibroses and fibrotic eye disorders. In the context of the present invention, the term fibrotic disorders includes in particular the following terms: hepatic fibrosis, cirrhosis of the liver, pulmonary fibrosis, endomyocardial fibrosis, nephropathy, glomerulonephritis, interstitial renal fibrosis, fibrotic damage resulting from diabetes, bone marrow fibrosis and similar fibrotic disorders, scleroderma, morphea, keloids, hypertrophic scarring (also following surgical procedures), naevi, diabetic retinopathy, proliferative vitreoretinopathy and disorders of the connective tissue (for example sarcoidosis).

Cancer

Cancer is disease in which a group of cells display uncontrolled growth. Cancers are usually classified in carcinomas which is a cancer derived from epithelial cells (This group includes many of the most common cancers, including those of the breast, prostate, lung and colon.); sarcomas, which are derived from connective tissue, or mesenchymal cells; lymphoma and leukemia, derived from hematopoietic cells; germ cell tumor, which is derived from pluripotent; and blastomas, which is a cancer derived from immature "precursor" or embryonic tissue.

The present invention furthermore provides the use of a fusion polypeptide of the invention for preparing a medicament for the treatment and/or prevention of disorders, in particular the disorders mentioned above.

The present invention furthermore provides a method for the treatment and/or prevention of disorders, in particular the disorders mentioned above, using an effective amount of at least one fusion polypeptide of the invention.

The present invention furthermore provides a fusion polypeptide of the invention for use in a method for the treatment and/or prophylaxis of coronary heart disease, acute coronary syndrome, heart failure, and myocardial infarction.

Pharmaceutical Compositions and Administration

The present invention also provides for pharmaceutical compositions comprising a single chain Relaxin fusion protein in a pharmacologically acceptable vehicle. The single chain Relaxin

fusion protein may be administrated systemically or locally. Any appropriate mode of administration known in the art may be used including, but not limited to, intravenous, intraperitoneal, intraarterial, intranasal, by inhalation, oral, subcutaneous administration, by local injection or in form of a surgical implant.

The present invention also relates to pharmaceutical compositions which may comprise inventive fusion polypeptides, alone or in combination with at least one other agent, such as stabilizing compound, which may be administered in any sterile, biocompatible pharmaceutical carrier, including, but not limited to, saline, buffered saline, dextrose, and water. Any of these molecules can be administered to a patient alone, or in combination with other agents, drugs or hormones, in pharmaceutical compositions where it is mixed with excipient(s) or pharmaceutically acceptable carriers. In one embodiment of the present invention, the pharmaceutically acceptable carrier is pharmaceutically inert.

The present invention also relates to the administration of pharmaceutical compositions. Such administration is accomplished orally or parenterally. Methods of parenteral delivery include topical, intra-arterial, intramuscular, subcutaneous, intramedullary, intrathecal, intraventricular, intravenous, intraperitoneal, or intranasal administration. In addition to the active ingredients, these pharmaceutical compositions may contain suitable pharmaceutically acceptable carriers comprising excipients and auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically. Further details on techniques for formulation and administration may be found in the latest edition of Remington's Pharmaceutical Sciences (Ed. Maack Publishing Co, Easton, Pa.).

Pharmaceutical compositions for oral administration can be formulated using pharmaceutically acceptable carriers well known in the art in dosages suitable for oral administration. Such carriers enable the pharmaceutical compositions to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions and the like, for ingestion by the patient.

Pharmaceutical formulations for parenteral administration include aqueous solutions of active compounds. For injection, the pharmaceutical compositions of the invention may be formulated in aqueous solutions, preferably in physiologically compatible buffers such as Hank's solution, Ringer's solution, or physiologically buffered saline. Aqueous injection suspensions may contain substances that increase viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran. Additionally, suspensions of the active compounds may be prepared as appropriate oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils such as sesame oil, or synthetic fatty acid esters, such as ethyl oleate or triglycerides, or liposomes. Optionally, the suspension may also contain suitable stabilizers or

agents which increase the solubility of the compounds to allow for the preparation of highly concentrated solutions.

A fusion polypeptide according to the invention can be used alone or, if required, in combination with other active compounds. The present invention furthermore provides medicaments comprising at least one fusion polypeptide according to the invention and one or more further active ingredients, in particular for the treatment and/or prevention of the disorders mentioned above.

Suitable active ingredients for combination are, by way of example and by way of preference: active ingredients which modulate lipid metabolism, antidiabetics, hypotensive agents, perfusion-enhancing and/or antithrombotic agents, antioxidants, chemokine receptor antagonists, p38-kinase inhibitors, NPY agonists, orexin agonists, anorectics, PAF-AH inhibitors, antiphlogistics (COX inhibitors, LTB_4-receptor antagonists), analgesics for example aspirin, antidepressants and other psychopharmaceuticals.

The present invention relates in particular to combinations of at least one of the fusion polypeptides according to the invention with at least one lipid metabolism-altering active ingredient, antidiabetic, blood pressure reducing active ingredient and/or agent having antithrombotic effects.

The fusion polypeptides according to the invention can preferably be combined with one or more

- lipid metabolism-modulating active ingredients, by way of example and by way of preference from the group of the HMG-CoA reductase inhibitors, inhibitors of HMG-CoA reductase expression, squalene synthesis inhibitors, ACAT inhibitors, LDL receptor inductors, cholesterol absorption inhibitors, polymeric bile acid adsorbers, bile acid reabsorption inhibitors, MTP inhibitors, lipase inhibitors, LpL activators, fibrates, niacin, CETP inhibitors, PPAR-α, PPAR-γ and/or PPAR-δ agonists, RXR modulators, FXR modulators, LXR modulators, thyroid hormones and/or thyroid mimetics, ATP citrate lyase inhibitors, Lp(a) antagonists, cannabinoid receptor 1 antagonists, leptin receptor agonists, bombesin receptor agonists, histamine receptor agonists and the antioxidants/radical scavengers;

- antidiabetics mentioned in the Rote Liste 2004/II, chapter 12, and also, by way of example and by way of preference, those from the group of the sulfonylureas, biguanides, meglitinide derivatives, glucosidase inhibitors, inhibitors of dipeptidyl-peptidase IV (DPP-IV inhibitors), oxadiazolidinones, thiazolidinediones, GLP 1 receptor agonists, glucagon antagonists, insulin sensitizers, CCK 1 receptor agonists, leptin receptor agonists, inhibitors of liver enzymes

involved in the stimulation of gluconeogenesis and/or glycogenolysis, modulators of glucose uptake and also potassium channel openers, such as, for example, those disclosed in WO 97/26265 and WO 99/03861;

- hypotensive active ingredients, by way of example and by way of preference from the group of the calcium antagonists, angiotensin AII antagonists, ACE inhibitors, renin inhibitors, beta-receptor blockers, alpha-receptor blockers, aldosterone antagonists, mineralocorticoid receptor antagonists, ECE inhibitors, ACE/NEP inhibitors and the vasopeptidase inhibitors; and/or

- antithrombotic agents, by way of example and by way of preference from the group of the platelet aggregation inhibitors or the anticoagulants;

- diuretics;

- vasopressin receptor antagonists;

- organic nitrates and NO donors;

- compounds with positive inotropic activity;

- compounds which inhibit the degradation of cyclic guanosine monophosphate (cGMP) and/or cyclic adenosine monophosphat (cAMP), such as, for example, inhibitors of phosphodiesterases (PDE) 1, 2, 3, 4 and/or 5, in particular PDE 5 inhibitors, such as sildenafil, vardenafil and tadalafil, and also PDE 3 inhibitors, such as milrinone;

- natriuretic peptides, such as, for example, "atrial natriuretic peptide" (ANP, anaritide), "B-type natriuretic peptide" or "brain natriuretic peptide" (BNP, nesiritide), "C-type natriuretic peptide" (CNP) and also urodilatin;

- agonists of the prostacyclin receptor (IP receptor), such as, by way of example, iloprost, beraprost, cicaprost;

- inhibitors of the I_f (funny channel) channel, such as, by way of example, ivabradine;

- calcium sensitizers, such as, by way of example and by way of preference, levosimendan;

- potassium supplements;

- NO-independent, but heme-dependent stimulators of guanylate cyclase, such as, in particular, the compounds described in WO 00/06568, WO 00/06569, WO 02/42301 and WO 03/095451;

- NO- and heme-independent activators of guanylate cyclase, such as, in particular, the

compounds described in WO 01/19355, WO 01/19776, WO 01/19778, WO 01/19780, WO 02/070462 and WO 02/070510;

- inhibitors of human neutrophil elastase (HNE), such as, for example, sivelestat and DX-890 (Reltran);

- compounds which inhibit the signal transduction cascade, such as, for example, tyrosine-kinase inhibitors, in particular sorafenib, imatinib, gefitinib and erlotinib; and/or

- compounds which modulate the energy metabolism of the heart, such as, for example, etomoxir, dichloroacetate, ranolazine and trimetazidine.

Lipid metabolism-modifying active ingredients are to be understood as meaning, preferably, compounds from the group of the HMG-CoA reductase inhibitors, squalene synthesis inhibitors, ACAT inhibitors, cholesterol absorption inhibitors, MTP inhibitors, lipase inhibitors, thyroid hormones and/or thyroid mimetics, niacin receptor agonists, CETP inhibitors, PPAR-α agonists PPAR-γ agonists, PPAR-δ agonists, polymeric bile acid adsorbers, bile acid reabsorption inhibitors, antioxidants/radical scavengers and also the cannabinoid receptor 1 antagonists.

In a preferred embodiment of the invention, a fusion polypeptide according to the invention is administered in combination with an HMG-CoA reductase inhibitor from the class of the statins, such as, by way of example and by way of preference, lovastatin, simvastatin, pravastatin, fluvastatin, atorvastatin, rosuvastatin or pitavastatin.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a squalene synthesis inhibitor, such as, by way of example and by way of preference, BMS-188494 or TAK-475.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with an ACAT inhibitor, such as, by way of example and by way of preference, avasimibe, melinamide, pactimibe, eflucimibe or SMP-797.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a cholesterol absorption inhibitor, such as, by way of example and by way of preference, ezetimibe, tiqueside or pamaqueside.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with an MTP inhibitor, such as, by way of example and by way of preference, implitapide, BMS-201038, R-103757 or JTT-130.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a lipase inhibitor, such as, by way of example and by way of preference, orlistat.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a thyroid hormone and/or thyroid mimetic, such as, by way of example and by way of preference, D-thyroxine or 3,5,3'-triiodothyronine (T3).

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with an agonist of the niacin receptor, such as, by way of example and by way of preference, niacin, acipimox, acifran or radecol.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a CETP inhibitor, such as, by way of example and by way of preference, dalcetrapib, BAY 60-5521, anacetrapib or CETP vaccine (CETi-1).

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a PPAR-γ agonist, for example from the class of the thiazolidinediones, such as, by way of example and by way of preference, pioglitazone or rosiglitazone.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a PPAR-δ agonist, such as, by way of example and by way of preference, GW-501516 or BAY 68-5042.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a polymeric bile acid adsorber, such as, by way of example and by way of preference, cholestyramine, colestipol, colesolvam, CholestaGel or colestimide.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a bile acid reabsorption inhibitor, such as, by way of example and by way of preference, ASBT (= IBAT) inhibitors, such as, for example, AZD-7806, S-8921, AK-105, BARI-1741, SC-435 or SC-635.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with an antioxidant/radical scavenger, such as, by way of example and by way of preference, probucol, AGI-1067, BO-653 or AEOL-10150.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a cannabinoid receptor 1 antagonist, such as, by way of example and by way of preference, rimonabant or SR-147778.

Antidiabetics are to be understood as meaning, preferably, insulin and insulin derivatives, and also orally effective hypoglycemic active ingredients. Here, insulin and insulin derivatives include both insulins of animal, human or biotechnological origin and also mixtures thereof. The orally effective hypoglycemic active ingredients preferably include sulfonylureas, biguanides, meglitinide derivatives, glucosidase inhibitors and PPAR-gamma agonists.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with insulin.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a sulfonylurea, such as, by way of example and by way of preference, tolbutamide, glibenclamide, glimepiride, glipizide or gliclazide.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a biguanide, such as, by way of example and by way of preference, metformin.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a meglitinide derivative, such as, by way of example and by way of preference, repaglinide or nateglinide.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a glucosidase inhibitor, such as, by way of example and by way of preference, miglitol or acarbose.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a DPP-IV inhibitor, such as, by way of example and by way of preference, sitagliptin and vildagliptin.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a PPAR-gamma agonist, for example from the class of the thiazolinediones, such as, by way of example and by way of preference, pioglitazone or rosiglitazone.

The hypotensive agents are preferably understood as meaning compounds from the group of the calcium antagonists, angiotensin AII antagonists, ACE inhibitors, beta-receptor blockers, alpha-receptor blockers and diuretics.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a calcium antagonist, such as, by way of example and by way of preference, nifedipine, amlodipine, verapamil or diltiazem.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with an angiotensin AII antagonist, such as, by way of example and by way of preference, losartan, valsartan, candesartan, embusartan, olmesartan or telmisartan.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with an ACE inhibitor, such as, by way of example and by way of preference, enalapril, captopril, lisinopril, ramipril, delapril, fosinopril, quinopril, perindopril or trandopril.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a beta-receptor blocker, such as, by way of example and by way of preference, propranolol, atenolol, timolol, pindolol, alprenolol, oxprenolol, penbutolol, bupranolol, metipranolol, nadolol, mepindolol, carazalol, sotalol, metoprolol, betaxolol, celiprolol, bisoprolol, carteolol, esmolol, labetalol, carvedilol, adaprolol, landiolol, nebivolol, epanolol or bucindolol.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with an alpha-receptor blocker, such as, by way of example and by way of preference, prazosin.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a diuretic, such as, by way of example and by way of preference, furosemide, bumetanide, torsemide, bendroflumethiazide, chlorothiazide, hydrochlorothiazide, hydroflumethiazide, methyclothiazide, polythiazide, trichloromethiazide, chlorothalidone, indapamide, metolazone, quinethazone, acetazolamide, dichlorophenamide, methazolamide, glycerol, isosorbide, mannitol, amiloride or triamteren.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with an aldosterone or mineralocorticoid receptor antagonist, such as, by way of example and by way of preference, spironolactone or eplerenone.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a vasopressin receptor antagonist, such as, by way of example and by way of preference, conivaptan, tolvaptan, lixivaptan or SR-121463.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with an organic nitrate or NO donor, such as, by way of example and by way of preference, sodium nitroprusside, nitroglycerol, isosorbide mononitrate, isosorbide dinitrate, molsidomin or SIN-1, or in combination with inhalative NO.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a positive-inotropic compound, such as, by way of example and by way of preference, cardiac glycosides (digoxin), beta-adrenergic and dopaminergic agonists, such as isoproterenol, adrenaline, noradrenaline, dopamine or dobutamine.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with antisympathotonics, such as reserpine, clonidine or alpha-methyldopa, or in combination with potassium channel agonists, such as minoxidil, diazoxide, dihydralazine or hydralazine, or with substances which release nitrogen oxide, such as glycerol nitrate or sodium nitroprusside.

Antithrombotics are to be understood as meaning, preferably, compounds from the group of the platelet aggregation inhibitors or the anticoagulants.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a platelet aggregation inhibitor, such as, by way of example and by way of preference, aspirin, clopidogrel, ticlopidine or dipyridamol.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a thrombin inhibitor, such as, by way of example and by way of preference, ximelagatran, melagatran, dabigatran, bivalirudin or clexane.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a GPIIb/IIIa antagonist, such as, by way of example and by way of preference, tirofiban or abciximab.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a factor Xa inhibitor, such as, by way of example and by way of preference, rivaroxaban (BAY 59-7939), DU-176b, apixaban, otamixaban, fidexaban, razaxaban, fondaparinux, idraparinux, PMD-3112, YM-150, KFA-1982, EMD-503982, MCM-17, MLN-1021, DX 9065a, DPC 906, JTV 803, SSR-126512 or SSR-128428.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with heparin or a low molecular weight (LMW) heparin derivative.

In a preferred embodiment of the invention, the fusion polypeptides according to the invention are administered in combination with a vitamin K antagonist, such as, by way of example and by way of preference, coumarin.

In the context of the present invention, particular preference is given to combinations comprising at least one of the fusion polypeptides according to the invention and also one or more further active ingredients selected from the group consisting of HMG-CoA reductase inhibitors (statins), diuretics, beta-receptor blockers, organic nitrates and NO donors, ACE inhibitors, angiotensin AII antagonists, aldosterone and mineralocorticoid receptor antagonists, vasopressin receptor antagonists, platelet aggregation inhibitors and anticoagulants, and also their use for the treatment and/or prevention of the disorders mentioned above.

The present invention furthermore provides medicaments comprising at least one fusion polypeptides according to the invention, usually together with one or more inert nontoxic pharmaceutically suitable auxiliaries, and also their use for the purposes mentioned above.

Therapeutically Effective Dose

Pharmaceutical compositions suitable for use in the present invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose, e.g. heart failure. The determination of an effective dose is well within the capability of those skilled in the art.

For any compound, the therapeutically effective dose can be estimated initially either in in vitro assays, e.g. LGR7 receptor activation, ex vivo in isolated perfused rat hearts, or in animal models, usually mice, rabbits, dogs, or pigs. The animal model is also used to achieve a desirable concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of fusion polypeptide that ameliorates the symptoms or condition. Therapeutic efficacy and toxicity of such compounds can be determined by standard pharmaceutical procedures in vitro or experimental animals, e.g., ED50 (the dose therapeutically effective in 50% of the population) and LD50 (the dose lethal to 50% of the population). The dose ratio between therapeutic and toxic effects is the therapeutic index, and it can be expressed as the ratio, ED50/LD50. Pharmaceutical compositions that exhibit large therapeutic indices are preferred. The data obtained from in vitro assays and animal studies are

used in formulating a range of dosage for human use. The dosage of such compounds lies preferably within a range of circulating concentrations what include the ED50 with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, sensitivity of the patient, and the route of administration.

Normal dosage amounts may vary from 0.1 to 100,000 milligrams total dose, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature. See U.S. Pat. No. 4,657,760; 5,206,344; or 5,225,212. Those skilled in the art will employ different formulations for polynucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

The present invention is further described by the following examples. The examples are provided solely to illustrate the invention by reference to specific embodiments. These exemplifications, while illustrating certain specific aspects of the invention, do not portray the limitations or circumscribe the scope of the disclosed invention.

All examples were carried out using standard techniques, which are well known and routine to those of skill in the art, except where otherwise described in detail. Routine molecular biology techniques of the following examples can be carried out as described in standard laboratory manuals, such as Sambrook et al., Molecular Cloning: A Laboratory Manual, 2nd Ed.; Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1989.

Further preferred embodiments are:

1. A fusion polypeptide having Relaxin activity comprising A-L-B,

wherein

B comprises a Relaxin B chain polypeptide or a functional variant thereof,

A comprises a Relaxin A chain poylpeptide or a functional variant thereof, and

L is a linker polypeptide.

2. A fusion polypeptide according to count 1, wherein

B is a Relaxin B chain polypeptide or a functional variant thereof,

A is a Relaxin A chain poylpeptide or a functional variant thereof, and

L is a linker polypeptide.

3. A fusion polypeptide according to count 1 or 2, wherein the Relaxin B chain is a Relaxin 2B or a Relaxin 3B chain.

4. A fusion polypeptide according to anyone of the foregoing counts, wherein the Relaxin A chain is a Relaxin 2A or a Relaxin 3A chain.

5. A fusion polypeptide according to anyone of the foregoing counts, wherein the Relaxin A chain is a Relaxin 2A chain.

6. A fusion polypeptide according to anyone of the foregoing counts, wherein the Relaxin A chain is a Relaxin 3A chain.

7. A fusion polypeptide according to anyone of the foregoing counts, wherein the Relaxin A chain is a Relaxin 2A chain and the Relaxin B chain is a Relaxin 2B chain.

8. A fusion polypeptide according to anyone of the foregoing counts, wherein the Relaxin A and B chains are human Relaxin A and B chains.

9. A fusion polypeptide according to anyone of the foregoing counts, wherein the fusion polypeptide further comprises at least one half-life extending moiety.

10. A fusion polypeptide according to count 9, wherein the half-life extending moiety is a non-proteinaceous or a proteinaceous half-life extending moiety.

11. A fusion polypeptide according to count 9 or 10, wherein the polypeptide has the formula

(R1)m – (S1)n – A – L –B – (S2)o – (R2)p,

wherein

R1 and R2 are proteinaceous half-life extending moieties,

S1 and S2 are stretcher peptides,

and wherein m, n, o and p are independently the number 0 or 1, provided that at least one of m, n, o, and p are 1.

12. A fusion polypeptide according to count 11, wherein m and n are 0 and o and p are 1.

13. A fusion polypeptide according to count 11, wherein m and n are 1 and o and p are 0.

14. A fusion polypeptide according to count 11, wherein m is 1 and n, o and p are 0.

15. A fusion polypeptide according to count 11, wherein m, n and o are 0 and p is 1.

16. A fusion polypeptide according to any one of counts 11 to 15, wherein R1 and R2 are proteinaceous half-life extending moieties comprised in a group of proteinaceous half-life extending moieties consisting of immunoglobulin Fc domain, serum albumin, transferrin and serum albumin binding protein.

17. A fusion polypeptide according to any one of counts 10 to 16, wherein the proteinaceous half-life extending moiety is an IgG1 Fc domain.

18. A fusion polypeptide according to any one of counts 10 to 17, wherein the proteinaceous half-life extending moiety is human.

19. A fusion polypeptide according to count 10, wherein the non-proteinaceous half-life extending moiety is PEG or HES.

20. A fusion polypeptide according to anyone of counts 11 – 19, wherein the stretcher polypeptides S1 and S2 are 1 – 25 amino acids in length.

21. A fusion polypeptide according to anyone of counts 11 – 20, wherein the stretcher polypeptides S1 and S2 are 4 – 10 amino acids in length, preferably 10 amino acids in length

22. A fusion polypeptide according to count 21, wherein the stretcher polypeptide S1 and S2 is comprised in the group of stretcher polypeptides consisting of polypeptides as set forth in SEQ ID NO: 148, SEQ ID NO: 149 , and SEQ ID NOs: 150.

23. A fusion polypeptide according to anyone of the foregoing counts, wherein the linker polypeptide L is 6 – 14 amino acids in length.

24. A fusion polypeptide according to anyone of the foregoing counts, wherein the linker polypeptide L is 7 – 11 amino acids in length.

25. A fusion polypeptide according to anyone of the foregoing counts, wherein the linker polypeptide L is 8, 9, or 10 amino acids in length.

26. A fusion polypeptide according to anyone of the foregoing counts, wherein the linker polypeptide L is 9 amino acids in length.

27. A fusion polypeptide according to anyone of the foregoing counts, wherein the linker polypeptide L is comprised in a group of linkers consisting of linkers having 6, 7, 8, 9, 10, 11, 12, 13 and 14 amino acids in length.

28. A fusion polypeptide according to anyone of the foregoing counts, wherein in the linker polypeptide L all but 4 amino acid residues of the linker L consist of Gly and/or Ser residues and the remaining 4 amino acid residues are selected from the group of natural amino acids.

29. A fusion polypeptide according to anyone of the foregoing counts, wherein the linker polypeptide L comprises at least one Gly, Ser, Arg, Cys, Leu and/or Lys residue.

30. A fusion polypeptide according to anyone of the foregoing counts, wherein the linker polypeptide L comprises Gly and Ser residues.

31. A fusion polypeptide according to anyone of the foregoing counts, wherein the linker polypeptide L consists of Gly and Ser residues.

32. A fusion polypeptide according to anyone of the foregoing counts, wherein the linker polypeptide L comprises Gly and Ser residues and has a Gly to Ser ratio of at least 3 to 1.

33. A fusion polypeptide according to anyone of the foregoing counts, wherein the linker polypeptide L is comprised in the group of linker polypeptides consisting of polypeptides as set forth in SEQ ID NO: 137, SEQ ID NO: 138, SEQ ID NO: 139, SEQ ID NO: 140, SEQ ID NO: 141, SEQ ID NO: 142, SEQ ID NO: 143, SEQ ID NO: 145 and SEQ ID NO: 146.

34. A fusion polypeptide according to anyone of the foregoing counts, wherein the Relaxin A chain is human Relaxin 2 A chain (SEQ ID NO: 117) and the Relaxin B chain is human Relaxin 2 B chain (SEQ ID NO: 119).

35. A fusion polypeptide according to anyone of the foregoing counts, wherein A is the human Relaxin 2 A chain (SEQ ID NO: 117) and B is the human Relaxin 2 B chain (SEQ ID NO: 119).

36. A fusion polypeptide according to anyone of the foregoing counts comprising a polypeptide as depicted in table 3.

37. A fusion polypeptide according to anyone of the foregoing counts, wherein A-L-B is selected from the group of A-L-B polypeptides consisting of scR3, scR4, scR5, scR3 w/o Tag, scR4 w/o Tag, scR5 w/o Tag, scR-Fc5, scR-Fc6 and scR-Fc7.

38. A fusion polypeptide as depicted in table 3.

39. A fusion polypeptide selected from the group consisting of scR3, scR4, scR5, scR3 w/o Tag, scR4 w/o Tag, scR5 w/o Tag, scR-Fc5, scR-Fc6 and scR-Fc7.

40. A polynucleotide encoding a fusion polypeptide according to anyone of the foregoing counts.

41. A vector comprising a polynucleotide according to count 40.

42. A host cell comprising a vector according to count 41 or a polynucleotide according to count 40.

43. A host cell according to count 42, wherein the host cell is a eukaryotic or prokaryotic cell.

44. A host cell according to count 42 or 43, wherein the eukaryotic host cell is a mammalian, yeast, insect or plant cell.

45. A host cell according to count 44, wherein the mammalian host cell is a CHO cell.

46. A host cell according to count 43, wherein the prokaryotic host cell is a bacterial cell, preferably an E.coli cell.

47. A method of producing a polypeptide according to anyone of counts 1 – 39 comprising the steps of cultivating a host cell of counts 42 - 46 and isolating the polypeptide.

48. A pharmaceutical composition comprising a fusion polypeptide according to anyone of counts 1 – 39.

49. A pharmaceutical composition according to count 48 or a fusion polypeptide according to anyone of counts 1 – 39 as medicament.

50. A pharmaceutical composition according to count 48 and 49 or a fusion polypeptide according to anyone of counts 1 – 39 as medicament for the treatment of cardiovascular disease, lung disease, fibrotic disorder or kidney disease.

51. A method of treating a cardiovascular disease, lung disease, fibrotic disorder or kidney disease comprising the administration of a therapeutically effective dose of a pharmaceutical composition according to count 48 and 49 or a fusion polypeptide according to anyone of counts 1 – 39.

52. A treatment according to counts 50 and 51, wherein the cardiovascular disease is coronary heart disease, acute coronary syndrome, heart failure, and myocardial infarction.

Examples

Experimental protocols

Construction of Relaxin variants:

The cDNA sequences of the Relaxin variants were generated by chemical gene synthesis. The synthesized genes were subcloned into the mammalian expression vector pCEP4 (Invitrogen, catalogue number V044-50). As signal leader sequence for correct secretion of the resulting protein, either the leader sequence of the LDL receptor-related protein (LRP, amino acid composition MLTPPLLLLLPLLSALVAA) or of CD33 (amino acid composition MPLLLLLPLLWAGALA) were used. For subcloning of the synthesized constructs the restriction enzymes HindIII and BamH1 were used according to manufactures' instruction.

Expression of Relaxin variants:

For small scale expression (up to 2 milliliter culture volume) HEK293 (ATCC, catalogue number CRL-1573) cells were transiently transfected using Lipofectamine2000 Transfection Reagent (Invitrogen, catalogue number 11668-019) according to manufactures' Instructions. Cells were cultivated in D-Mem F12 (Gibco, #31330), 1% Penicillin-Streptomycin (Gibco, #15140) and 10% fetal calf serum (FCS, Gibco, #11058) in a humified incubator at 5% carbon dioxide at 37°C.

Three to five days following transfection, conditioned medium of the transfected cells were tested for activity using the stably transfected CHO-CRE-GR7 cell line.

For large scale expression (10 milliliter culture volume and more) the constructs were transiently expressed in mammalian cell cells as described in Tom et al., 2007. Briefly, the expression plasmid transfected into HEK293-6E cells and incubated in Fernbach–Flasks or Wave-Bags. Expression was at 37°C for 5 to 6 days in F17 Medium (Invitrogen). 5 g/l Tryptone TN1 (Organotechnie), 1% Ultra-Low IgG FCS (Invitrogen) and 0.5 mM Valproic acid (Sigma) were supplemented after transfection.

Purification of Relaxin variants:

Relaxin Fc-Fusion constructs were purified from mammalian cell culture supernatants. First supernatants were clarified from cell debris by centrifugation. Proteins were purified by Protein A (MabSelect Sure, GE Healthcare) affinity chromatography followed by size exclusion chromatography (SEC). Therefore the supernatant was applied to a Protein A column previously equilibrated in PBS pH 7.4 (Sigma /Aldrich), contaminants were removed with 10 column volumes of PBS pH 7.4 + 500 mM NaCl. Relaxin Fc Fusion constructs were eluted with 50 mM Na-acetate pH 3.5 + 500 mM NaCl and further purified by SEC on a Superdex 200 column in PBS pH 7.4.

For purification of c-Myc tagged proteins or polypeptides, the c-Myc tagged Protein Mild Purification Gel is used (Biozol Diagnostic, Protein Mild Purification Gel, catalogue number 3306) according to the manufactures instructions.

For purification of His tagged proteins or polypeptides, Ni-NTA spin columns are used (Qiagen, Ni-NTA Spin Kit, catalogue number 31314) according to the manufactures instructions.

Quantification of expressed Relaxin variants:

For quantification of secreted and purified recombinant Relaxin variants, the commercially available quantification ELISA (R&D Systems, Human Relaxin-2 Quantikine ELISA Kit, catalogue number DRL200) was used according to the manufactures' instructions.

In addition for some constructs proteins were quantified by using FC-ELISA. For the Fc ELISA, 96 well microtitter plates (Nunc, Maxi Sorp black, catalogue number 460918) were coated with an anti-Fc antibody (SigmaAldrich, catalogue number A2136) over night at 4°C and a concentration of 5 µg per milliliter. Plates were washed once by using 50 microliter per well of a buffer consisting of PBS and 0,05% Tween 20 (SigmaAldrich, catalogue number 63158) buffer. Thirty microliter of a blocking buffer (Candor Bioscience, catalogue number 113500) was added and the plate incubated for 1 hour at 37°C. Plates were washed 3 times using 50 microliter per well of the PBS/0,05% Tween 20 buffer. Samples were added and the plates incubated were for 1 hour at 37°C. If necessary, samples have to be diluted by using the above mentioned blocking buffer. After incubation, plates were washed 3 times using 50 microliter per well of the PBS/0,05% Tween 20 buffer.

For detection 30 microliter of a Anti-h-Fc-POD (SigmaAldrich, catalogue number A0170) diluted 1:10000 in 10% blocking buffer was added and incubated for 1 hour at 37°C. After incubation, plates were washed 3 times using 50 microliter per well of the PBS/0,05% Tween 20 buffer. Thirty microliter of BM Blue Substrate POD (Roche Diagnostics, catalogue number 11484281001) was added and after five minutes of incubation, the reaction was stopped by the addition of a 1 molar acid sulfur solution. Absorption was measured using the Tecan Infinite 500 reader, absorbance mode, extinction 450nm, emission 690nm.

For determination of the concentration of Myc tagged proteins the Human c-Myc ELISA kit (EIAab & USCNLIFE, Wuhan EIAab Science Co.,Ltd, catalogue number E1290h) was used according to the manufactures instruction.

For determination of the concentration of His tagged proteins a His-Tag Protein ELISA Kit (BIOCAT GmbH, catalogue number AKR-130) was used according to the manufactures instruction.

For determination of the concentration of HA (hemagglutinin) tagged proteins a Human hemagglutinin,HA ELISA Kit (Hölzel Diagnostika, catalogue number CSB-E09360h) was used according to the manufactures instruction.

Activity testing:

CHO K1 cells (ATCC, catalogue number CCL-61) were stably transfected with the cyclic AMP responsive element (CRE) Luciferase reporter gene construct (Biomyx Technology, pHTS-CRE, catalogue number P2100) resulting in a CHO-CRE-Luciferase cell line.

This cell line was subsequently stably transfected with the human LGR7/RXFP1 receptor (accession numbers NM_021634.2), cloned as 2271 base pair long DNA fragment into the mammalian expression vector pcDNA3.1(-) (Invitrogen, catalogue number V79520), resulting in a CHO-CRE-LGR7 cell line. This cell line was cultivated in D-Mem F12 (Gibco, #31330) 2 mM Glutamax (Gibco, #35050), 100 nM Pyruvat (Gibco, # 11360-070), 20 mM Hepes (Gibco, # 15630), 1% Penicillin-Streptomycin (Gibco, #15140) and 10% fetal calf serum (FCS, Gibco, #11058).

For stimulation, medium was exchanged by OptiMem (Gibco, #11058) + 1% FCS containing different concentrations of the recombinantly expressed Relaxin variant proteins (usually starting at a concentration of 100 nM, followed by 1:2 dilutions). As positive control, commercially available recombinant expressed human Relaxin 2 (Genbank Accession number NP_604390.1) was used (R&D Systems, catalogue number 6586-RN-025). Subsequently, cells were incubated for 6 hours in a humified incubator at 5% carbon dioxide at 37°C. After 6 hours cells were tested for Luciferase activity using a Luciferase Assay System (Promega, # E1500) and using the Tecan Infinite 500 reader, luminescence mode, 1000 milliseconds integration time, measurement time 30 seconds.

Relative luminescence units were used to determine EC50 values of the different molecules by using the computer program Graph Pad Prism Version 5.

For alternative activity testing of Relaxin as well as of fusion polypeptides of the invention, cell lines (e.g. THP1, ATCC catalogue number TIB-202) or primary cells (e.g. Celprogen Inc., Human Cardiomyocyte Cell Culture, catalogue number 36044-15) with endogenous expression of the LGR7 receptor are used. These cells are cultivated according to the manufactures instruction.

Methods for the detection of Relaxin or Relaxin variants induced generation of cAMP are known in the art. For example, such measurement is performed using a cAMP ELISA (e.g. IBL

International GmbH, cAMP ELISA, catalogue number CM 581001) according to the manufactures instruction.

Methods for the detection of Relaxin or Relaxin variants induced activation of PI3 kinase are known in the art. For example, such measurement is performed using a PI3-Kinase HTRF Assay according to the manufactures instruction (e.g. Millipore, PI3-Kinase HTRF Assay, catalogue number 33-016).

PEGylation

For PEGylation to cysteine residues the fusion polypeptide is usually treated with a reducing agent, such as dithiothreitol (DDT) prior to PEGylation. The reducing agent is subsequently removed by any conventional method, such as by desalting. Conjugation of PEG to a cysteine residue typically takes place in a suitable buffer at pH 6-9 at temperatures varying from 4°C to 25°C for periods up to 16 hours.

It will be understood that the PEGylation is designed so as to produce the optimal molecule with respect to the number of PEG molecules attached, the size and form of such molecules (e.g. whether they are linear or branched), and the attachment site(s) in the fusion polypeptide. The molecular weight of the polymer to be used may e.g. be chosen on the basis of the desired effect to be achieved.

Immunogenicity testing

Immunogenicity testing is performed by using the computer program NetMHCIIpan (Center for Biological Sequence Analysis; Department of Systems Biology; Technical University of Denmark) which calculates the potential binding affinity of proteins or peptides to MHCII complex. The higher the calculated binding affinity the higher is the risk to induce antibodies directed against the protein or polypeptide of interest.

In vitro determination of mapping T cell epitopes is performed according to the protocol published by Reijonen and Kwok (Reijonen H., Kwok WW. (2003) Use of HLA class II tetramers in tracking antigen-specific T cells and mapping T-cell epitopes. Methods 29:282-288).

Constructs of single chain Relaxin variants

Determination of the optimal linker length of single chain Relaxin variants

Single chain Relaxin variants with different linker length connecting the A and B chain were generated as described above. As depicted in the sequences, for alternative determination of

protein expression, in some constructs a Myc Tag (amino acid sequence EQKLISEEDL) was added to the N terminal end of the A chain either with or without a hemagglutinin tag (amino acid sequence YPYDVPDYA) as well as a 6 Histidine tag (amino acid sequence HHHHHH) was added at the C terminal end of the B chain.

Example 1: scR1

In scR1 composition of the linker sequence connecting the A chain and B chain of human Relaxin 2 is three amino in acids length and consist of the polypeptide with the sequence GlyGlyGly. For alternative determination of protein expression, a Myc tag was added at the N terminal end of the A chain and a hemagglutinin tag and a 6 Histidine tag was added at the C terminal end of the B chain.

Example 2: scR2

In scR2 composition of the linker sequence connecting the A chain and B chain of human Relaxin 2 is five amino acids in length and consist of the polypeptide with the sequence GlyGlyGlySerGly. For alternative determination of protein expression, a Myc tag was added at the N terminal end of the A chain and a hemagglutinin tag and a 6 Histidine tag was added at the C terminal end of the B chain.

Example 3: scR3

In scR3 composition of the linker sequence connecting the A chain and B chain of human Relaxin 2 is seven amino acids in length and consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGly. For alternative determination of protein expression, a Myc tag was added at the N terminal end of the A chain and a hemagglutinin tag and a 6 Histidine tag was added at the C terminal end of the B chain.

Example 4: scR4

In scR4 composition of the linker sequence connecting the A chain and B chain of human Relaxin 2 is nine amino acids in length and consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGly. For alternative determination of protein expression, a Myc tag was added at the N terminal end of the A chain and a hemagglutinin tag and a 6 Histidine tag was added at the C terminal end of the B chain.

Example 5: scR5

In scR5 composition of the linker sequence connecting the A chain and B chain of human Relaxin 2 is eleven amino acids in length and consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGlyGlyGly. For alternative determination of protein expression, a Myc tag was added at the N terminal end of the A chain and a hemagglutinin tag and a 6 Histidine tag was added at the C terminal end of the B chain.

Example 6: scR6

In scR6 composition of the linker sequence connecting the A chain and B chain of human Relaxin 2 is fifteen amino acids in length and consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGlyGlyGlySerGlyGlyGly. For alternative determination of protein expression, a Myc tag was added at the N terminal end of the A chain and a hemagglutinin tag and a 6 Histidine tag was added at the C terminal end of the B chain.

Example 7: scR7

In scR7 composition of the linker sequence connecting the A chain and B chain of human Relaxin 2 is six amino acids in length and consist of the polypeptide with the sequence GlyGlyGlySerGlyGly. For alternative determination of protein expression, a Myc tag is added at the N terminal end of the A chain. Activity is measured according to the protocol as described above.

Example 8: scR8

In scR8 composition of the linker sequence connecting the A chain and B of human Relaxin 2 chain is twelve amino acids in length and consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGlyGlyGlySer. For alternative determination of protein expression, a Myc tag is added at the N terminal end of the A chain. Activity is measured according to the protocol described above.

Example 9: scR9

In scR9 composition of the linker sequence connecting the A chain and B chain of human Relaxin 2 is be thirteen amino acids in length and consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGlyGlyGlySerGly. For alternative determination of protein expression, a Myc tag is added at the N terminal end of the A chain. Activity is measured according to the protocol described above.

Example 10: scR10

In scR10 composition of the linker sequence connecting the A chain and B chain of human Relaxin 2 is fourteen amino acids in length and consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGlyGlyGlySerGlyGly. For alternative determination of protein expression, a Myc tag is added at the N terminal end of the A chain. Activity will be measured according to the protocol described above.

Example 11: scR11

In scR11 composition of the linker sequence connecting the A chain and B of human Relaxin 2 chain is ten amino acids in length and consist of the polypeptide with the sequence GlyGlyGlySerGlyCysGlyGlySerGly. For activity testing of the non-PEGylated fusion polypeptide non-purified protein was used.

To improve the biological half life of this construct, PEGylation of the Cysteine within the linker connecting the A chain and B chain following the protocol as described above is performed. Activity of the PEGylated variant is measured according to the protocol described above.

Example 12: scR12

In scR12 composition of the linker sequence connecting the A chain and B chain of human Relaxin 2 is ten amino acids in length and consist of the polypeptide with the sequence GlyGlyGlySerGlyLysGlyGlySerGly. For activity testing of the non-PEGylated fusion polypeptide non-purified protein was used.

To improve the biological half life of this construct, PEGylation of the Lysine within the linker connecting the A chain and B chain following the protocol as described above could be an option. Activity of the PEGylated variant is measured according to the protocol described above.

Example 13: scR13

In scR13 composition of the linker sequence connecting the C terminal end of the A chain and the N terminal end of the B chain of human Relaxin 2 is nine amino acids long and consists of the polypeptide with the sequence LysArgSerLeuSerArgLysLysArg. For activity testing non-purified fusion polypeptide was used.

Example 14: scR14

In scR14 composition of the linker sequence connecting the C terminal end of the A chain and N terminal end of the B chain of human Relaxin 3 (accession number NP_543140.1) is nine

amino in acids length and will consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGly. Activity is measured according to the protocol described above. For activity testing non-purified fusion polypeptide was used.

Example 15: scR15

In scR15 composition of the linker sequence connecting the C terminal end of the A chain and N terminal end of the B chain of human Relaxin 3 (accession number NP_543140.1) is nine amino in acids length and will consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGly. For alternative determination of protein expression, a Myc tag is added at the N terminal end of the A chain. Activity is measured according to the protocol described above.

Example 16: scR16

In scR16 composition of the linker sequence connecting the C-terminus of the B chain and the N-terminus of the A chain of human Relaxin 2 is nine amino in acids length and will consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGly. For alternative determination of protein expression, a Myc tag is added at the N terminal end of the A chain. Activity is measured according to the protocol described above.

Example 17: scR17

In scR17 composition of the linker sequence connecting the C-terminus of the A chain of human Relaxin 3 (accession number NP_543140.1)and the N-terminus of the B chain of human Relaxin 2 (accession number NP_604390.1) is nine amino in acids length and will consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGly. For alternative determination of protein expression, a Myc tag is added at the N terminal end of the A chain. Activity is measured according to the protocol described above.

Example 18: scR18

In scR18 composition of the linker sequence connecting the C-terminus of the B chain of human Relaxin 2 (accession number NP_604390.1) and the N-terminus of the A chain of human Relaxin 3 (accession number NP_543140.1) is nine amino in acids length and will consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGly. For alternative determination of protein expression, a Myc tag is added at the N terminal end of the A chain. Activity is measured according to the protocol described above.

Example 19: scR19

In scR19 composition of the linker sequence connecting the C-terminus of the A chain of human Relaxin 2 (accession number NP_604390.1) and the N-terminus of the B chain of human Relaxin 3 (accession number NP_543140.1) is nine amino in acids length and will consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGly. For alternative determination of protein expression, a Myc tag is added at the N terminal end of the A chain. Activity is measured according to the protocol described above.

Example 20: scR20

In scR20 composition of the linker sequence connecting the C-terminus of the B chain of human Relaxin 3 (accession number NP_543140.1) and the N-terminus of the A chain of human Relaxin 2 (accession number NP_604390.1) is nine amino in acids length and will consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGly. For alternative determination of protein expression, a Myc tag is added at the N terminal end of the A chain. Activity is measured according to the protocol described above.

A graphical representation of all single chain Relaxin variants is given in Fig. 2.

Table 1 summarizes the results regarding the expression as well as the biological activity of various scR constructs. Whereas single chain Relaxin variants having a linker length of three, five, and fifteen amino acids do not show any detectable biological activity in the assay described above, surprisingly the tested linker lengths of six, seven, nine, ten, eleven, twelve, thirteen, and fourteen amino acids lead to single chain variants exhibiting biological activity comparable to human Relaxin 2.

Although the length of the linker connecting the C-terminus of the A chain with the N-terminus of the B chain is important for the generation of a biological active molecule, the composition of the amino acids of the linker is variable. Examples are scR11 to scR13. Thereby, scR11 and scR12 exhibit an additional amino acid in the linker sequence (C in the linker of scR11 and K in the linker of scR12) or in case of the construct scR13, which exhibits a linker sequence which does not show any homology to the linker sequences mentioned above.

Generation of single chain Relaxin variants is not limited to Relaxin 2. Constructs scR14 and 15 are single chain variants of Relaxin 3. Although the overall sequence homology between Relaxin 2 and Relaxin 3 is low, the genomic organization of these two genes as memebers of the insulin superfamily is identical. Like Relaxin 2, Relaxin 3 consists of the classical B chain - C chain – A chain structure. Like for Relaxin 2, the C chain is cleaved off from the Relaxin 3 propeptide by Prohormone convertase I and II and the B and A chain are connected via disulfide bridges and by this the active molecule is formed. Constructs scR14 and scR15 are single chain

variants of Relaxin 3, exhibiting the same linker molecule connecting the C-terminus of the A chain with the N-terminus of the B chain as for example already shown for Relaxin 2 with the construct scR4. scR14 and scR15 exhibit detectable biological activity.

scR16, scR17, scR18, scR19, and scR20 are chimeras between the A chain of Relaxin 3 and the B chain of Relaxin 2 and vice versa. Thereby, for activation of the LGR7 receptor it is mandatory that the B-chain of the Relaxin 2 and Relaxin 3, respectively, are located in the C-terminal part of a Relaxin 3/Relaxin 2 chimera.

Clone	Expression	EC_{50} (M)*
hRelaxin 2		2,60E-11
hRelaxin 3		2,30E-09
scR1	detectable	not detectable
scR2	detectable	not detectable
scR3	detectable	7,70E-11
scR4	detectable	3,40E-11
scR5	detectable	3,70E-11
scR6	detectable	not detectable
scR7	detectable	5,30E-08
scR8	detectable	2,40E-08
scR9	detectable	1,10E-07
scR10	detectable	4,40E-08
scR11	detectable	2,50E-08
scR12	detectable	3,60E-08
scR13	detectable	active (EC_{50} n.d.)
scR14	detectable	5,90E-10
scR15	detectable	6,20E-10
scR16	detectable	not detectable
scR17	detectable	1,30E-08
scR18	detectable	not detectable
scR19	detectable	active (EC_{50} n.d.)
scR20	detectable	not detectable

* values are examples of three to five independent experiments.

Dose response curves and the corresponding EC_{50} values comparing the activity of hRelaxin 2, scR3, scR4, and scR5 are shown in Figure 4a, for hRelaxin 2, scR7, scR8, scR9, and scR10 are shown in Figure 4b, for hRelaxin 2, scR11 and scR12 are shown in Figure 4c, for hRelaxin 2,

hRelaxin 3, scR14 and scR15 are shown in Figure 4d, and for hRelaxin 3 and scR17 are shown in Figure 4e.

Conclusion: This shows that a linker length of more than five amino acids and less than fifteen amino acids are required for biological activity of single chain Relaxin variants wherein the C terminus of the A chain is connected via such linkers to the N terminus of the B chain. Furthermore, the generation of single chain Relaxin of the invention is not limited to Relaxin 2.

Binding of Relaxin 2 to its corresponding receptor LGR7 is a two-step process. In a first step, the A chain of human Relaxin 2 binds to the N terminal ectodomain of the receptor. In a second step, this bound ectodomain undergoes a conformational change and secondary interactions between the B chain of Relaxin and the transmembrane domain of LGR7 mediates receptor signaling. This second step is the most relevant in the activation of the ligand – receptor complex. Therefore, due to the fact that the variant scR17 contains the A chain of human Relaxin 3 instead of human Relaxin 2, leads to a construct with reduced activity. A further reduction in the activity is observed with the variant scR19, which contains the B chain of human Relaxin 3 instead of the B chain of human Relaxin 2. Binding to the ectodomain occurs via the A chain of the human Relaxin 2, but the B chain of the human Relaxin 3 is suboptimal for activating LGR7. The corresponding receptor for Relaxin 3 is LGR8. Therefore, it is very likely, that by using the scR19 as ligand and LGR8 as the corresponding receptor, signal intensity were much higher. This is also a mean to modulate the activity of an fusion polypeptide of the invention.

The non-purification of scR13 is an explanation of the lower activity as possible impurities in the sample leads to false determination of the concentration or could have an negative impact on the accuracy the cell based Luciferase assay.

In conclusion this shows that useful linker sequences are not restricted to Glycine/Serine rich sequences as other linker sequences (within the inventive length) also lead to fully active single chain Relaxins.

Construction of single chain Relaxin fusion proteins with improved biological half life.

In order to improve the biological half life of single chain Relaxin variants, constructs were designed where the Fc moiety of immunoglobulin molecules were added at the N terminal or C terminal end of the single chain Relaxin variants.

Thereby, single chain Relaxin variants were directly fused to the Fc part of an immunoglobulin or linked by a polypeptide of different length and amino acid compositions.

Another option to improve the biological half life of polypeptides are fusions with polypeptides like Transferrin (accession number P02787) or Albumin (accession number P02768) (SR Schmid (2009)).

PEGylation is a commonly used method to improve the biological half life of polypeptides. Hereby polyethylene glycol polymer chains are added covalently attached to a polypeptide. Thereby a reactive derivative of PEG is incubated with the target polypeptide. Preferred amino acids reacting with PEG are Cysteins and Lysins.

Pasut and Veronese (2009))

Generation of a Relaxin fusion protein - Relaxin-Fc

To improve the biological half life the Fc part of the human IgG1 was combined with human Relaxin 2 by chemically based gene synthesis. The carboxy-terminal part of human Relaxin 2 (according to its genomic organization arranged as follows: B chain – C chain – A chain) was fused to N terminal end of the human IgG1 Fc moiety, whereby these two parts of the fusion protein were connected by a 6 amino acids long linker sequence consisting of a polypeptide with the sequence IleGluGlyArgMetAsp encoding the coagulation factor Xa cleavage site. However, Relaxin Fc showed no activity determined by a CHO-CRE-LGR7 cell line.

Example 16: scR-Fc 1

In scR-Fc 1 composition of the linker sequence connecting the C terminal end of scR 4 with the N terminal end of the human IgG1 Fc moiety is 6 amino acids long and consists of the polypeptide with the sequence IleGluGlyArgMetAsp encoding the coagulation factor Xa cleavage site. This polypeptide and Fc moiety replaces the hemagglutinin tag and 6 Histidine tag in scR 4. For alternative determination of protein expression, a Myc tag was added at the N terminal end of the A chain.

Example 17: scR-Fc 2

In scR-Fc 2 composition of the linker sequence connecting the C terminal end of the single chain Relaxin scR4 with the N terminal end of the human IgG1 Fc moiety is 4 amino acids long and consists of the polypeptide with the sequence GlyGlySerPro. In contrast to scR-Fc 1, this construct has no Myc tag at the N terminal end of the A chain.

Example 18: scR-Fc 3

In scR-Fc 3 composition of the linker sequence connecting the C terminal end of the single chain Relaxin scR4 with the N terminal end of the human IgG1 Fc moiety is 7 amino acids long and consists of the polypeptide with the sequence GlyGlySerGlyGlySerPro. In contrast to scR-Fc 1, this construct has no Myc tag at the N terminal end of the A chain.

Example 19: scR-FC 4

In scR-Fc 4 composition of the linker sequence connecting the C terminal end of the single chain Relaxin scR4 with the N terminal end of the human IgG1 Fc moiety is 10 amino acids long and consists of the polypeptide with the sequence GlyGlySerGlyGlySerGlyGlySerPro. In contrast to scR-Fc 1, this construct has no Myc tag at the N terminal end of the A chain.

Example 20: scR-Fc 5

In scR-Fc 5 composition of the linker sequence connecting the N terminal end of the single chain Relaxin scR4 with the C terminal end of the human IgG1 Fc moiety is 4 amino acids long and consists of the polypeptide with the sequence GlyGlySerPro. The Fc moiety replaces the Myc tag at the N terminal end of the A chain. This construct has no hemagglutinin tag and/or 6 Histidine tag at its C terminal end.

Example 21: scR-Fc 6

In scR-Fc 6 composition of the linker sequence connecting the N terminal end of the single chain Relaxin scR4 with the C terminal end of the human IgG1 Fc moiety is 7 amino acids long and consists of the polypeptide with the sequence GlyGlySerGlyGlySerPro. The Fc moiety replaces the Myc tag at the N terminal end of the A chain. This construct has no hemagglutinin tag and/or 6 Histidine tag at its C terminal end.

Example 22: scR-Fc 7

In scR-Fc 7 composition of the linker sequence connecting the N terminal end of the single chain Relaxin scR4 with the C terminal end of the human IgG1 Fc moiety is 10 amino acids long and consists of the polypeptide with the sequence GlyGlySerGlyGlySerGlyGlySerPro. The

Fc moiety replaces the Myc tag at the N terminal end of the A chain. This construct has no hemagglutinin tag and/or 6 Histidine tag at its C terminal end.

Example 23: scR-FC 8

In scR-Fc 8 composition of the linker sequence connecting the C terminal end of the single chain Relaxin scR4 with the N terminal end of the rat IgG2b Fc moiety is 4 amino acids long and consists of the polypeptide with the sequence GlyGlySerPro. Additionally a 6 Histidine tag is added at the C terminal end of the Fc part. In contrast to scR4, this construct has no Myc tag the N terminal end of the A chain. The rat IgG2b Fc moiety replaces the hemagglutinin tag and 6 Histidine tag.

Example 24: scR-Fc 9

In scR-Fc 9 composition of the linker sequence connecting the C terminal end of the single chain Relaxin scR4 with the N terminal end of the rat IgG2b Fc moiety is 7 amino acids long and consists of the polypeptide with the sequence GlyGlySerGlyGlySerPro. Additionally a 6 Histidine tag is added at the C terminal end of the Fc part. In contrast to scR4, this construct has no Myc tag the N terminal end of the A chain. The rat IgG2b Fc moiety replaces the hemagglutinin tag and 6 Histidine tag.

Example 25: scR-Fc 10

In scR-Fc 10 composition of the linker sequence connecting the C terminal end of the single chain Relaxin scR4 with the N terminal end of the rat IgG2b Fc moiety is 10 amino acids long and consists of the polypeptide with the sequence GlyGlySerGlyGlySerGlyGlySerPro. Additionally a 6 Histidine tag is added at the C terminal end of the Fc part. In contrast to scR4, this construct has no Myc tag at the N terminal end of the A chain. The rat IgG2b Fc moiety replaces the hemagglutinin tag and 6 Histidine tag.

Example 26: scR-Fc 11

In scR-Fc 11 composition of the linker sequence connecting the N terminal end of the single chain Relaxin scR4 with the C terminal end of the rat IgG2b Fc moiety is 4 amino acids long and consists of the polypeptide with the sequence GlyGlySerPro. Additionally a 6 Histidine tag is added at the N terminal end of the Fc part. The rat IgG2b Fc moiety replaces the Myc tag. Additionally this construct has no hemagglutinin tag and/or 6 Histidine tag at its C terminal end.

Example 27: scR-Fc 12

In scR-Fc 11 composition of the linker sequence connecting the N terminal end of the single chain Relaxin scR1 with the C terminal end of the rat IgG2b Fc moiety is 7 amino acids long and consists of the polypeptide with the sequence GlyGlySerGlyGlySerPro. Additionally a 6 Histidine tag is added at the N terminal end of the Fc part. The rat IgG2b Fc moiety replaces the Myc tag. Additionally this construct has no hemagglutinin tag and/or 6 Histidine tag at its C terminal end.

Example 28: scR-Fc 13

In scR-Fc 11 composition of the linker sequence connecting the N terminal end of the single chain Relaxin scR4 with the C terminal end of the rat IgG2b Fc moiety is 10 amino acids long and consists of the polypeptide with the sequence GlyGlySerGlyGlySerGlyGlySerPro. Additionally a 6 Histidine tag is added at the N terminal end of the Fc part. The rat IgG2b Fc moiety replaces the Myc tag. Additionally this construct has no hemagglutinin tag and/or 6 Histidine tag at its C terminal end.

Example 29: scR-Fc 14

In order to analyze the influence of a linker sequence connecting single chain Relaxin variants and Fc moieties, in scR-Fc 14 the C terminal end of sequence scR4 was directly fused to the Fc part of the human IgG1. This Fc moiety replaces the hemagglutinin tag and 6 Histidine tag in scR4. This construct has no Myc tag at the N terminal end of the A chain.

Example 30: scR-Fc 15

In scR-Fc 15 composition of the linker sequence connecting the C terminal end of the single chain Relaxin scR4 with the N terminal end of the human IgG1 Fc moiety is 6 amino acids long and consists of the polypeptide with the sequence GlySerGlySerGlySer. The human IgG1 Fc moiety replaces the hemagglutinin tag and 6 Histidine tag. This construct has no Myc tag at the N terminal end of the A chain.

Example 31: scR-Fc 16

scR-Fc 16 was designed to analyze the influence of disulfide bridges within the Fc moiety on protein expression and fusion protein activity. For this, the Cysteine residue at position 86 within the Fc part of the human IgG1 in scR-Fc 15 was replaced by Alanin.

Example 32: scR-Fc 17

In scR-Fc 17 composition of the linker sequence connecting the C terminal end of the single chain Relaxin scR4 with the N terminal end of the rat IgG2b Fc moiety is 6 amino acids long

and consists of the polypeptide with the sequence GlySerGlySerGlySer. The rat IgG2b Fc moiety replaces the hemagglutinin tag and 6 Histidine tag. This construct has no Myc tag at the N terminal end of the A chain.

Example 33: scR-Fc 18

In scR-Fc 18 composition of the linker sequence connecting the C terminal end of the single chain Relaxin scR4 with the N terminal end of the human IgG1 Fc moiety is 21 amino acids long and consists of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGlyThrLysValThrValSerSerGluSerLysTyrGly. The human IgG1 Fc moiety replaces the hemagglutinin tag and 6 Histidine tag. This construct has no Myc tag at the N terminal end of the A chain.

Example 34: scR-Var 1

In scR-Var1 composition of the linker sequence connecting the A chain and B chain of the human Relaxin 2 is of nine amino acids in length and consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGly. Additionally a polypeptide of six amino acids in length and with the sequence GlyGlySerGlyCysGly was added at the C terminal end of the B chain. For activity testing of the non-PEGylated fusion polypeptide non-purified protein was used.

To improve the biological half life of this construct, PEGylation of the Cysteine within the stretcher polypeptide fused at the C terminal end of the B chain is performed following the protocol as described above. Activity of the PEGylated variant is measured according to the protocol described above.

Example 35: scR-Var 2

In scR-Var2 composition of the linker sequence connecting the A chain and B chain of the human Relaxin 2 is of nine amino acids length and consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGly. Additionally a polypeptide of six amino acids in length and with the sequence GlyCysGlySerGlyGly was added at the N terminal end of the A chain. For activity testing of the non-PEGylated fusion polypeptide non-purified protein was used.

To improve the biological half life of this construct, PEGylation of the Cysteine within the stretcher polypeptide fused at the N terminal end of the A chain is performed following the protocol as described above. Activity of the PEGylated variant is measured according to the protocol described above.

Example 36: scR-Var3

In scR-Var3 composition of the linker sequence connecting the C terminal end of the A chain and the N terminal end of the B chain of of the human Relaxin 2 is of nine amino acids in length and consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGly. At the N terminal end of the A chain a polypeptide with the sequence IleGluGlyArgMetAsp encoding the coagulation factor Xa cleavage site connects this variant with the C terminal end of the human Transferrin protein (accession number NP_001054.1). Activity is measured according to the protocol described above.

Example 37: scR-Var4

In scR-Var4 wild type proRelaxin 2 (genomic organization) is fused to Transferrin. For this, at the N terminal end of the B chain is a polypeptide with the sequence IleGluGlyArgMetAsp encoding the coagulation factor Xa cleavage site connects this variant with the C terminal end of the human Transferrin protein (accession number NP_001054.1). Activity is measured according to the protocol described above.

Example 38: scR-Var5

In scR-Var5 composition of the linker sequence connecting the C terminal end of the A chain and the N terminal end of the B chain of of human Relaxin 2 is of nine amino acids length and consist of the polypeptide with the sequence GlyGlyGlySerGlyGlyGlySerGly. At the N terminal end of the A chain a polypeptide with the sequence IleGluGlyArgMetAsp encoding the coagulation factor Xa cleavage site connects this variant with the C terminal end of the human Albumin protein (accession number NP_000468.1). Activity is measured according to the protocol described above.

Example 39: scR-Var6

In scR-Var6, a polypeptide with the sequence IleGluGlyArgMetAsp encoding the coagulation factor Xa cleavage site located at the N terminal end of the B chain connects this variant with the C terminal end of the human Albumin protein (accession number NP_000468.1). Activity is measured according to the protocol described above.

Example 40: scR-Var7

In scR-Var7 composition of the linker sequence connecting the C terminal end of the A chain of human Relaxin 2 and the N terminal end of the B chain of human Relaxin 2 is nine amino acids long and consists of the polypeptide with the sequence LysArgSerLeuSerArgLysLysArg,

A linker sequence connecting the C terminal end of the B chain with the N terminal end of the human IgG1 Fc moiety is 6 amino acids long and consists of the polypeptide with the sequence IleGluGlyArgMetAsp encoding the coagulation factor Xa cleavage site.

Example 41: scR-Var8

In scR-Var8 composition of the linker sequence connecting the C terminal end of the A chain and the N terminal end of the B chain is nine amino acids long and consists of the polypeptide with the sequence LysArgSerLeuSerArgLysLysArg.

A linker sequence connecting the N terminal end of the A chain with the C terminal end of the human IgG1 Fc moiety is 6 amino acids long and consists of the polypeptide with the sequence IleGluGlyArgMetAsp encoding the coagulation factor Xa cleavage site.

A graphical representation of all single chain Relaxin fusion proteins as well as the variants designed for PEGylation is given in Fig. 3.

Table 2 summarizes the results for expression as well as biological activity of various scR fusion protein constructs.

Clone	Expression	EC_{50} (M)*
Relaxin		3,50E-11
Relaxin Fc	detectable	not detectable
scR-Fc 1	detectable	1,30E-08
scR-Fc 2	detectable	3,30E-09
scR-Fc 3	detectable	2,40E-09
scR-Fc 4	detectable	3,10E-09
scR-Fc 5	detectable	1,30E-10
scR-Fc 6	detectable	4,20E-10
scR-Fc 7	detectable	7,40E-10
scR-Fc 8	detectable	7,20E-09
scR-Fc 9	detectable	9,90E-09
scR-Fc 10	detectable	4,80E-09
scR-Fc 11	detectable	1,20E-09
scR-Fc 12	detectable	9,50E-10
scR-Fc 13	detectable	8,90E-10
scR-Fc 14	detectable	3,90E-07
scR-Fc 15	detectable	3,40E-09
scR-Fc 16	detectable	2,50E-09
scR-Fc 17	detectable	2,50E-09
scR-Fc 18	detectable	active (EC_{50} n.d.)
scR-Var1	detectable	1,10E-07
scR-Var2	detectable	4,20E-08
scR-Var3	detectable	1,00E-09
scR-Var4	detectable	1,30E-10
scR-Var5	detectable	5,50E-09
scR-Var6	detectable	8,30E-09
scR-Var7	detectable	active (EC_{50} n.d.)
scR-Var8	detectable	active (EC_{50} n.d.)

* values are examples of three to five independent experiments.

For all variants listed, expression could be determined by using the Human Relaxin-2 Quantikine ELISA Kit and activity could be measured by using the CHO-CRE-LGR7 cell line. Exemplarily, dose response curves for scR-Fc 1, scR-Fc 5 to scR-Fc 7, scR-Fc 11 to scR-Fc 13, and scR-Var3 to scR-Var6 are shown in Figure 5, Figure 6, Figure 7, and Figure 8, respectively.

The human wildtype Relaxin 2 molecule with its orientation B chain – C chain – A chain fused to the Fc moiety of the human IgG molecule does not show any detectable activity. Possible explanation for the non-activity of this molecule could be an incomplete processing of the C chain. In contrast, in all fusion constructs containing the single chain human Relaxin 2, a

significant activity can be detected. As shown above, the single chain Relaxin exhibits activity comparable to the human wildtype Relaxin 2, although no proteolytic processing takes place.

For the single chain Relaxin 2 fusion constructs, the orientation of the Fc moiety seems to have a significant impact on the activity of these molecules. Constructs carrying the Fc part at the C terminal end of the B chain (e.g. scR-Fc 1 to scR-Fc 4 and scR-Fc 13 to scR-Fc 18) exhibit a slightly lower activity than constructs carrying the Fc moiety at the N terminal end of the A chain (e.g. scR-Fc 5 to scR-Fc 6 and scR-Fc 11 to scR-Fc 12). As mentioned above, after binding of the A chain to the ectodomain of the corresponding receptor LGR7, a conformational change within the receptor molecule brings the B chain in contact with the extracellular loops of the transmembrane domains. The second step than leads to the activation of the receptor. Therefore, the Fc moiety coupled to the B chain could inhibit the optimal binding of the B chain and by this inhibits the full activation of the receptor.

Analysis of the in vivo plasma stability of Fc-single chain Relaxin

scR-Fc 13 and hRelaxin2 were administrated intravenously in 8 weeks old, male Wistar rats at concentrations of 240 µg/kg. At time points 0 hour, 1 hour, 3 days, 5 days, and 7 days after compound administration, blood samples were taken and the concentrations of the Fc-single chain Relaxin and non-modified hRelaxin2 were determined using the commercially available quantification ELISA (R&D Systems, Human Relaxin-2 Quantikine ELISA Kit, catalogue number DRL200).

As shown in Fig. 9, three days after application, non-modified hRelaxin2 was undetectable whereas for scR-Fc13 even 7 days after intravenous administration significant concentrations were detected, that were even above the EC50 value obtained for the CHO-LGR7 based activity test.

Determination of Fc-single chain relaxin activity isolated from plasma.

In order to determine whether scR-Fc 13 still exhibits activity after 3, 5, and 7 days after intravenous administration, plasma samples were tested on the CHO-CRE-LGR7 cell line. As shown in Figure 10, for all three samples activity could be determined and for all three samples, activity values are similar to the EC_{50} value obtained with the purified scR-Fc 13 variant.

Isolated perfused rat heart

Male Wistar rats (200-250 g) were anesthetized using Narcoren (100 mg/kg i.p.). The heart was rapidly excised and connected to a Langendorff perfusion system (FMI GmbH, Seeheim-Ober Beerbach, Germany). The heart was perfused at a constant rate of 10 ml/min with Krebs-Henseleit bicarbonate buffer solution equilibrated with 95% O_2 - 5% CO_2. The perfusion solution contained (in mmol/l): NaCl 118; KCl 3; $NaHCO_3$ 22; KH_2PO_4 1,2; $MgSO_4$ 1,2; $CaCl_2$ 1,8; Glucose 10; Na-Pyruvat 2. A pressure transducer registered the perfusion pressure in the perfusion system. The left ventricular pressure (LVP) was measured using a second pressure transducer connected to a water-filled balloon which was inserted into the left ventricle via the left atrium. The end diastolic pressure was initially set to 8 mm Hg by adjusting the volume of the balloon. The hearts were spontaneously beating. The signals from the pressure transducer were amplified, registered and used for the calculation of the heart frequency and +dp/dt by a personal computer.

As shown in Figure 11, perfusion of human Relaxin 2 (Fig. 11 a – d) as well as scR-Fc 13 (Fig. 11 e – h) are leading to a significant increase in heart rate and coronary flow and to a decrease in the left ventricular diastolic pressure and the left ventricular pressure (+dp/dtmax). Thereby, hRelaxin 2 is ten fold more potent than scR-Fc 13, reflecting the differences in the EC50 values

for sc Relaxin fusion protein variants and of Relaxin 2 determined with the CHO-CRE-LGR7 cell line.

Table 5:

List of constructs and corresponding SEQ ID NOs.

Construct	type	SEQ ID NO
scR1	PRT	SEQ ID NO:1
scR2	PRT	SEQ ID NO:2
scR3	PRT	SEQ ID NO:3
scR4	PRT	SEQ ID NO:4
scR5	PRT	SEQ ID NO:5
scR6	PRT	SEQ ID NO:6
scR7	PRT	SEQ ID NO:7
scR8	PRT	SEQ ID NO:8
scR9	PRT	SEQ ID NO:9
scR10	PRT	SEQ ID NO:10
scR11	PRT	SEQ ID NO:11
scR12	PRT	SEQ ID NO:12
scR13	PRT	SEQ ID NO:13
scR14	PRT	SEQ ID NO:14
scR15	PRT	SEQ ID NO:15
scR-Fc 1	PRT	SEQ ID NO:16
scR-Fc 2	PRT	SEQ ID NO:17

Construct	type	SEQ ID NO
scR-Fc 3	PRT	SEQ ID NO:18
scR-Fc 4	PRT	SEQ ID NO:19
scR-Fc 5	PRT	SEQ ID NO:20
scR-Fc 6	PRT	SEQ ID NO:21
scR-Fc 7	PRT	SEQ ID NO:22
scR-Fc 8	PRT	SEQ ID NO:23
scR-Fc 9	PRT	SEQ ID NO:24
scR-Fc 10	PRT	SEQ ID NO:25
scR-Fc 11	PRT	SEQ ID NO:26
scR-Fc 12	PRT	SEQ ID NO:27
scR-Fc 13	PRT	SEQ ID NO:28
scR-Fc 14	PRT	SEQ ID NO:29
scR-Fc 15	PRT	SEQ ID NO:30
scR-Fc 16	PRT	SEQ ID NO:31
scR-Fc 17	PRT	SEQ ID NO:32
scR-Fc 18	PRT	SEQ ID NO:33
scR-Var1	PRT	SEQ ID NO:34
scR-Var2	PRT	SEQ ID NO:35
scR-Var3	PRT	SEQ ID NO:36
scR-Var4	PRT	SEQ ID NO:37
scR-Var5	PRT	SEQ ID NO:38

Construct	type	SEQ ID NO
scR-Var6	PRT	SEQ ID NO:39
scR-Var7	PRT	SEQ ID NO:40
scR-Var8	PRT	SEQ ID NO:41
scR1 w/o Tag	PRT	SEQ ID NO:42
scR2 w/o Tag	PRT	SEQ ID NO:43
scR3 w/o Tag	PRT	SEQ ID NO:44
scR4 w/o Tag	PRT	SEQ ID NO:45
scR5 w/o Tag	PRT	SEQ ID NO:46
scR6 w/o Tag	PRT	SEQ ID NO:47
scR7 w/o Tag	PRT	SEQ ID NO:48
scR8 w/o Tag	PRT	SEQ ID NO:49
scR9 w/o Tag	PRT	SEQ ID NO:50
scR10 w/o Tag	PRT	SEQ ID NO:51
scR-Fc 1 w/o Tag	PRT	SEQ ID NO:52
scR-Fc 8 w/o Tag	PRT	SEQ ID NO:53
scR-Fc 9 w/o Tag	PRT	SEQ ID NO:54
scR-Fc 10 w/o Tag	PRT	SEQ ID NO:55
scR-Fc 11 w/o Tag	PRT	SEQ ID NO:56
scR-Fc 12 w/o Tag	PRT	SEQ ID NO:57
scR-Fc 13 w/o Tag	PRT	SEQ ID NO:58
scR1	DNA	SEQ ID NO:59

Construct	type	SEQ ID NO
scR2	DNA	SEQ ID NO:60
scR3	DNA	SEQ ID NO:61
scR4	DNA	SEQ ID NO:62
scR5	DNA	SEQ ID NO:63
scR6	DNA	SEQ ID NO:64
scR7	DNA	SEQ ID NO:65
scR8	DNA	SEQ ID NO:66
scR9	DNA	SEQ ID NO:67
scR10	DNA	SEQ ID NO:68
scR11	DNA	SEQ ID NO:69
scR12	DNA	SEQ ID NO:70
scR13	DNA	SEQ ID NO:71
scR14	DNA	SEQ ID NO:72
scR15	DNA	SEQ ID NO:73
scR-Fc 1	DNA	SEQ ID NO:74
scR-Fc 2	DNA	SEQ ID NO:75
scR-Fc 3	DNA	SEQ ID NO:76
scR-Fc 4	DNA	SEQ ID NO:77
scR-Fc 5	DNA	SEQ ID NO:78
scR-Fc 6	DNA	SEQ ID NO:79
scR-Fc 7	DNA	SEQ ID NO:80

Construct	type	SEQ ID NO
scR-Fc 8	DNA	SEQ ID NO:81
scR-Fc 9	DNA	SEQ ID NO:82
scR-Fc 10	DNA	SEQ ID NO:83
scR-Fc 11	DNA	SEQ ID NO:84
scR-Fc 12	DNA	SEQ ID NO:85
scR-Fc 13	DNA	SEQ ID NO:86
scR-Fc 14	DNA	SEQ ID NO:87
scR-Fc 15	DNA	SEQ ID NO:88
scR-Fc 16	DNA	SEQ ID NO:89
scR-Fc 17	DNA	SEQ ID NO:90
scR-Fc 18	DNA	SEQ ID NO:91
scR-Var1	DNA	SEQ ID NO:92
scR-Var2	DNA	SEQ ID NO:93
scR-Var3	DNA	SEQ ID NO:94
scR-Var4	DNA	SEQ ID NO:95
scR-Var5	DNA	SEQ ID NO:96
scR-Var6	DNA	SEQ ID NO:97
scR-Var7	DNA	SEQ ID NO:98
scR-Var8	DNA	SEQ ID NO:99
scR1 w/o Tag	DNA	SEQ ID NO:100
scR2 w/o Tag	DNA	SEQ ID NO:101

Construct	type	SEQ ID NO
scR3 w/o Tag	DNA	SEQ ID NO:102
scR4 w/o Tag	DNA	SEQ ID NO:103
scR5 w/o Tag	DNA	SEQ ID NO:104
scR6 w/o Tag	DNA	SEQ ID NO:105
scR7 w/o Tag	DNA	SEQ ID NO:106
scR8 w/o Tag	DNA	SEQ ID NO:107
scR9 w/o Tag	DNA	SEQ ID NO:108
scR10 w/o Tag	DNA	SEQ ID NO:109
scR-Fc 1 w/o Tag	DNA	SEQ ID NO:110
scR-Fc 8 w/o Tag	DNA	SEQ ID NO:111
scR-Fc 9 w/o Tag	DNA	SEQ ID NO:112
scR-Fc 10 w/o Tag	DNA	SEQ ID NO:113
scR-Fc 11 w/o Tag	DNA	SEQ ID NO:114
scR-Fc 12 w/o Tag	DNA	SEQ ID NO:115
scR-Fc 13 w/o Tag	DNA	SEQ ID NO:116
RLN2 A chain	PRT	SEQ ID NO:117
RLN2 minimal A chain	PRT	SEQ ID NO:118
RLN2 B chain	PRT	SEQ ID NO:119
Fc IgG1 human	PRT	SEQ ID NO:120
Fc IgG2b rat	PRT	SEQ ID NO:121
Transferrin	PRT	SEQ ID NO:122

Construct	type	SEQ ID NO
Albumin	PRT	SEQ ID NO:123
RLN3 A chain	PRT	SEQ ID NO:124
RLN3 B chain	PRT	SEQ ID NO:125
RLN3 minimal A chain	PRT	SEQ ID NO:126
RLN2 A chain	DNA	SEQ ID NO:127
RLN2 minimal A chain	DNA	SEQ ID NO:128
RLN2 B chain	DNA	SEQ ID NO:129
Fc IgG1 human	DNA	SEQ ID NO:130
Fc IgG2b rat	DNA	SEQ ID NO:131
Transferrin	DNA	SEQ ID NO:132
Albumin	DNA	SEQ ID NO:133
RLN3 A chain	DNA	SEQ ID NO:134
RLN3 B chain	DNA	SEQ ID NO:135
RLN3 minimal A chain	DNA	SEQ ID NO:136
linker 1	PRT	SEQ ID NO:137
linker 2	PRT	SEQ ID NO:138
linker 3	PRT	SEQ ID NO:139
linker 4	PRT	SEQ ID NO:140
linker 5	PRT	SEQ ID NO:141
linker 6	PRT	SEQ ID NO:142
linker 7	PRT	SEQ ID NO:143

Construct	type	SEQ ID NO
linker 8	PRT	SEQ ID NO:144
linker 9	PRT	SEQ ID NO:145
linker 10	PRT	SEQ ID NO:146
stretcher 1	PRT	SEQ ID NO:147
stretcher 2	PRT	SEQ ID NO:148
stretcher 3	PRT	SEQ ID NO:149
stretcher 4	PRT	SEQ ID NO:150
stretcher 5	PRT	SEQ ID NO:151
scR16	PRT	SEQ ID NO:152
scR17	PRT	SEQ ID NO:153
scR18	PRT	SEQ ID NO:154
scR19	PRT	SEQ ID NO:155
scR20	PRT	SEQ ID NO:156
scR16	DNA	SEQ ID NO:157
scR17	DNA	SEQ ID NO:158
scR18	DNA	SEQ ID NO:159
scR19	DNA	SEQ ID NO:160
scR20	DNA	SEQ ID NO:161

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WO 02/070510

Claims:

1. A fusion polypeptide having Relaxin activity comprising A-L-B,

wherein

B comprises a Relaxin B chain polypeptide or a functional variant thereof,

A comprises a Relaxin A chain poylpeptide or a functional variant thereof, and

L is a linker polypeptide.

2. A fusion polypeptide according to claim 1, wherein the fusion polypeptide further comprises at least one half-life extending moiety.

3. A fusion polypeptide according to claim 2, wherein half-life extending moiety is an IgG1 Fc domain, PEG or HES.

4. A fusion polypeptide according to anyone of the foregoing claims, wherein the linker polypeptide L is 6 – 14 amino acids in length.

5. A fusion polypeptide according to anyone of the foregoing claims, wherein the Relaxin A chain is human Relaxin 2 A chain (SEQ ID NO: 117) and the Relaxin B chain is human Relaxin 2 B chain (SEQ ID NO: 119).

6. A fusion polypeptide comprising a polypeptide as depicted in table 3.

7. A fusion polypeptide according to anyone of claims 1 to 6, wherein A-L-B is selected from the group of A-L-B polypeptides consisting of scR3, scR4, scR5, scR3 w/o Tag, scR4 w/o Tag, scR5 w/o Tag, scR-Fc5, scR-Fc6 and scR-Fc7.

8. A polynucleotide encoding a fusion polypeptide according to anyone of the foregoing claims.

9. A vector comprising a polynucleotide according to claim 8.

10. A host cell comprising a vector according to claim 9 or a polynucleotide according to claim 8.

11. A method of producing a polypeptide according to anyone of claims 1 – 7 comprising the steps of cultivating a host cell of claim 10 and isolating the polypeptide.

12. A pharmaceutical composition comprising a fusion polypeptide according to anyone of claims 1 – 7.

13. A pharmaceutical composition according to claim 12 or a fusion polypeptide according to anyone of claims 1 – 7 as medicament.

14. A pharmaceutical composition according to claim 12 and 13 or a fusion polypeptide according to anyone of claims 1 – 7 as medicament for the treatment of cardiovascular disease, lung disease, fibrotic disorder or kidney disease.

15. A method of treating a cardiovascular disease, lung disease, fibrotic disorder or kidney disease comprising the administration of a therapeutically effective dose of a pharmaceutical composition according to claim 12 and 13 or a fusion polypeptide according to anyone of claims 1 – 7.

16. A treatment according to claims 14 and 15, wherein the cardiovascular disease is coronary heart disease, acute coronary syndrome, heart failure, and myocardial infarction.

Figure 1

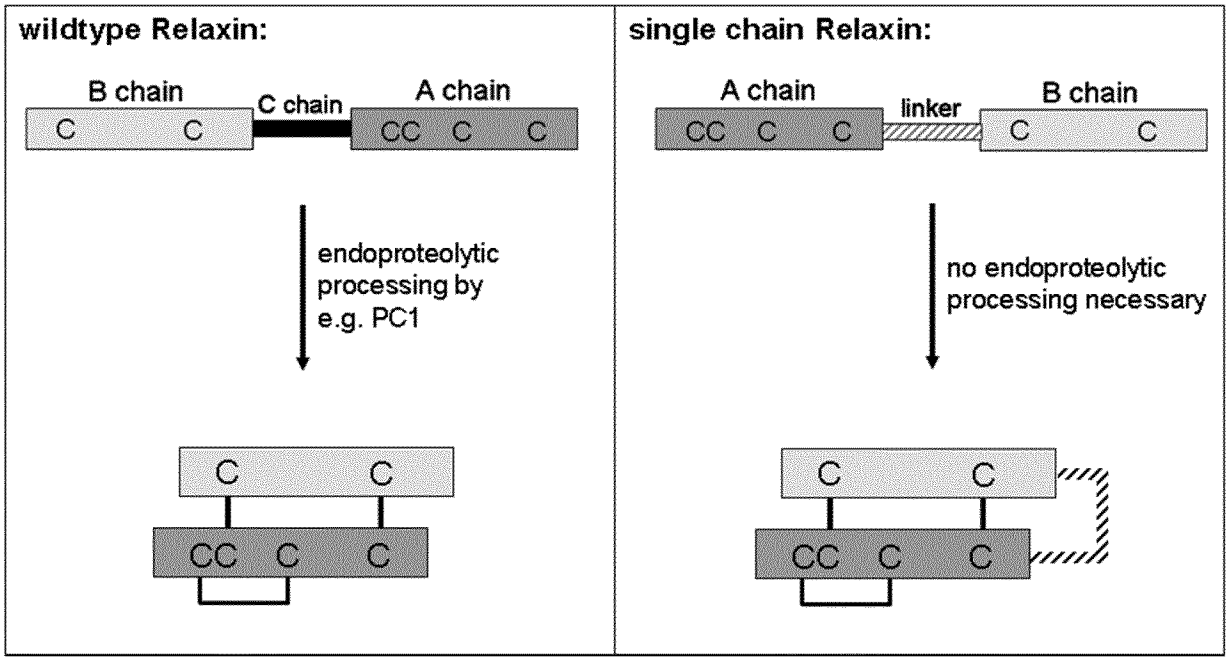

wildtype Relaxin:
B chain
C chain
A chain
C C
CC C C
endoproteolytic processing by e.g. PC1
C C
CC C C
single chain Relaxin:
A chain
linker
B chain
CC C C
C C
no endoproteolytic processing necessary
C C
CC C C

Figure 2

Clone	Construct
hRelaxin 2	B C A
scR1	Myc A 3aaG B FXa HA His
scR2	Myc A 5aaGS B FXa HA His
scR3	Myc A 7aaGS B FXa HA His
scR4	Myc A 9aaGS B FXa HA His
scR5	Myc A 11aaGS B FXa HA His
scR6	Myc A 15aaGS B FXa HA His
scR7	Myc A 6aaGS B
scR8	Myc A 12aaGS B
scR9	Myc A 13aaGS B
scR10	Myc A 14aaGS B
scR11	A 9aaGS + C B
scR12	A 9aaGS + K B
scR13	A short linker B
scR14	A (RLN3) 9aaGS B (RLN3)
scR15	Myc A (RLN3) 9aaGS B (RLN3)
scR16	Myc B (RLN2) 9aaGS A (RLN2)
scR17	Myc A (RLN3) 9aaGS B (RLN2)
scR18	Myc B (RLN2) 9aaGS A (RLN3)
scR19	Myc A (RLN2) 9aaGS B (RLN3)
scR20	Myc B (RLN3) 9aaGS A (RLN2)

Figure 3

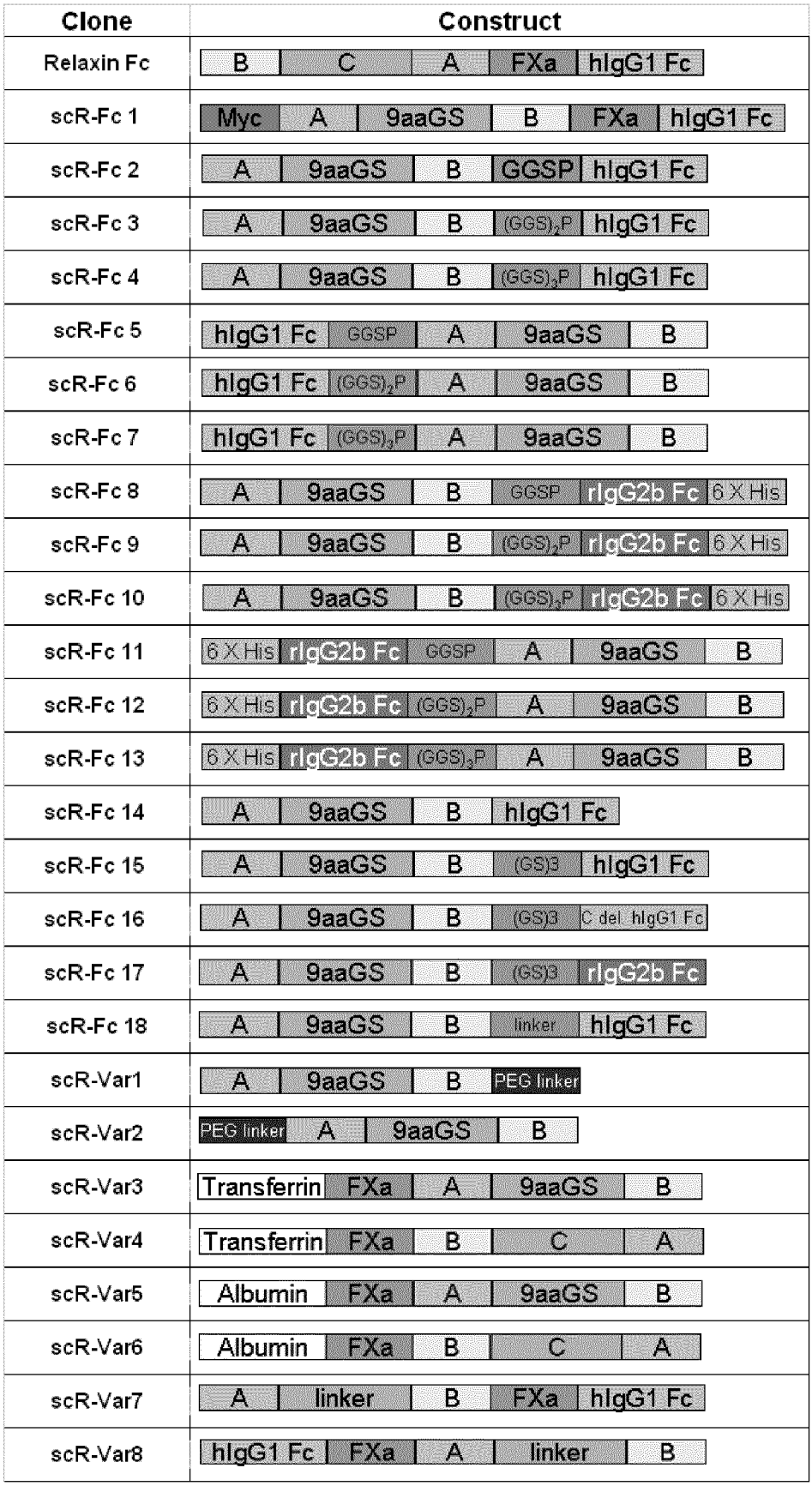
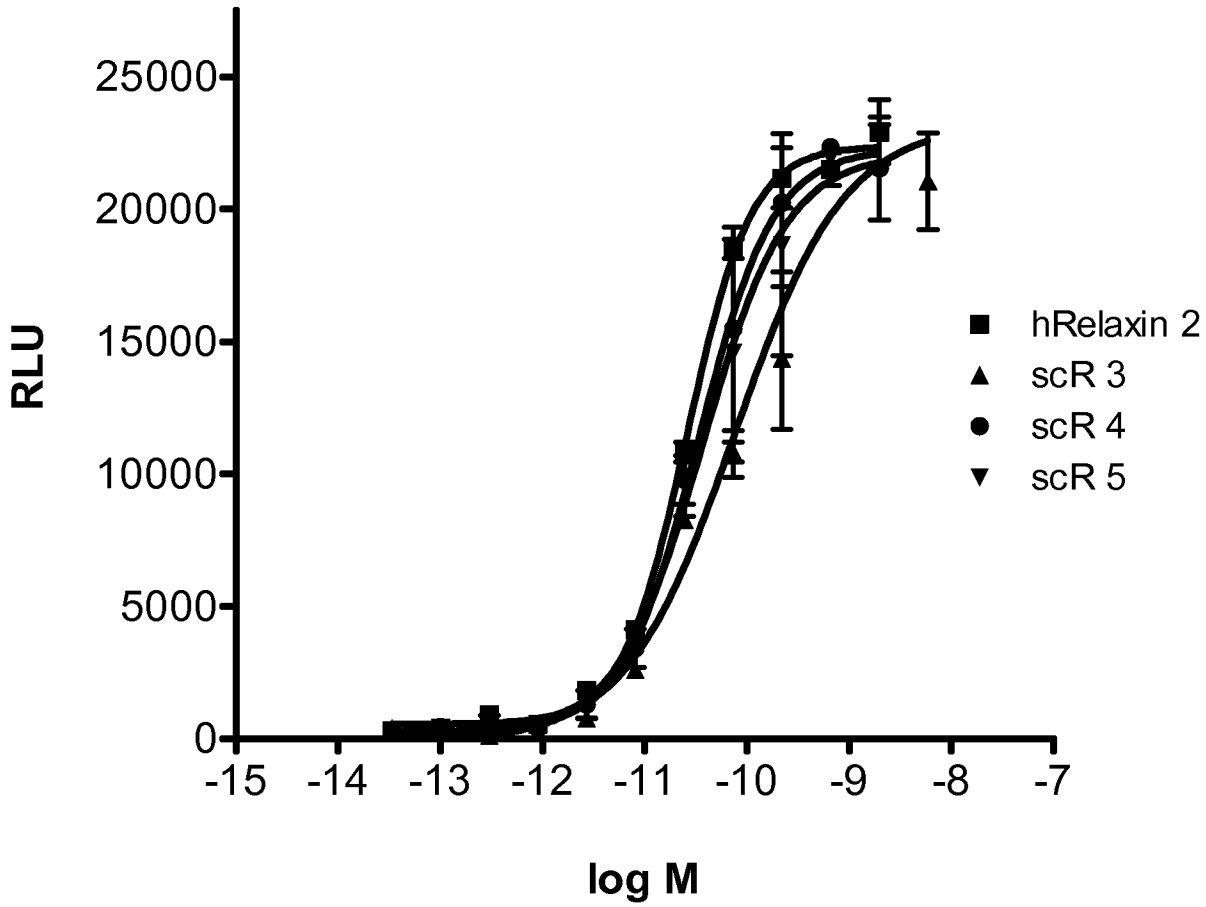
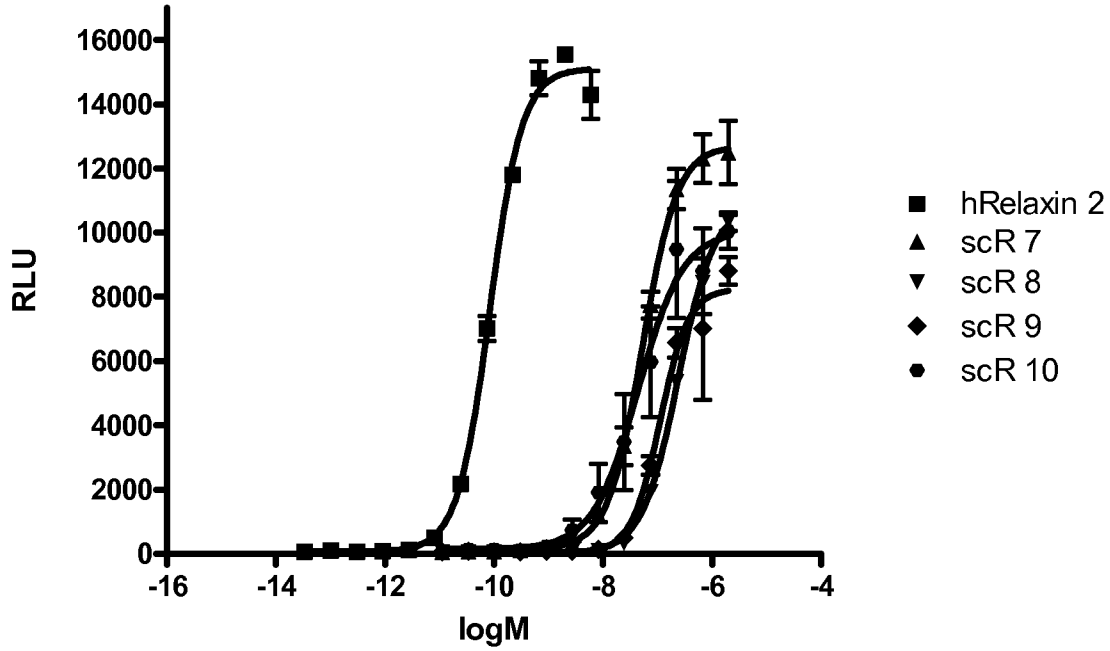

Clone
Construct
Relaxin Fc
B C A FXa hIgG1 Fc
scR-Fc 1
Myc A 9aaGS B FXa hIgG1 Fc
scR-Fc 2
A 9aaGS B GGSP hIgG1 Fc
scR-Fc 3
A 9aaGS B (GGS)2P hIgG1 Fc
scR-Fc 4
A 9aaGS B (GGS)3P hIgG1 Fc
scR-Fc 5
hIgG1 Fc GGSP A 9aaGS B
scR-Fc 6
hIgG1 Fc (GGS)2P A 9aaGS B
scR-Fc 7
hIgG1 Fc (GGS)3P A 9aaGS B
scR-Fc 8
A 9aaGS B GGSP rIgG2b Fc 6 X His
scR-Fc 9
A 9aaGS B (GGS)2P rIgG2b Fc 6 X His
scR-Fc 10
A 9aaGS B (GGS)3P rIgG2b Fc 6 X His
scR-Fc 11
6 X His rIgG2b Fc GGSP A 9aaGS B
scR-Fc 12
6 X His rIgG2b Fc (GGS)2P A 9aaGS B
scR-Fc 13
6 X His rIgG2b Fc (GGS)3P A 9aaGS B
scR-Fc 14
A 9aaGS B hIgG1 Fc
scR-Fc 15
A 9aaGS B (GS)3 hIgG1 Fc
scR-Fc 16
A 9aaGS B (GS)3 C del hIgG1 Fc
scR-Fc 17
A 9aaGS B (GS)3 rIgG2b Fc
scR-Fc 18
A 9aaGS B linker hIgG1 Fc
scR-Var1
A 9aaGS B PEG linker
scR-Var2
PEG linker A 9aaGS B
scR-Var3
Transferrin FXa A 9aaGS B
scR-Var4
Transferrin FXa B C A
scR-Var5
Albumin FXa A 9aaGS B
scR-Var6
Albumin FXa B C A
scR-Var7
A linker B FXa hIgG1 Fc
scR-Var8
hIgG1 Fc FXa A linker B

Figure 4a



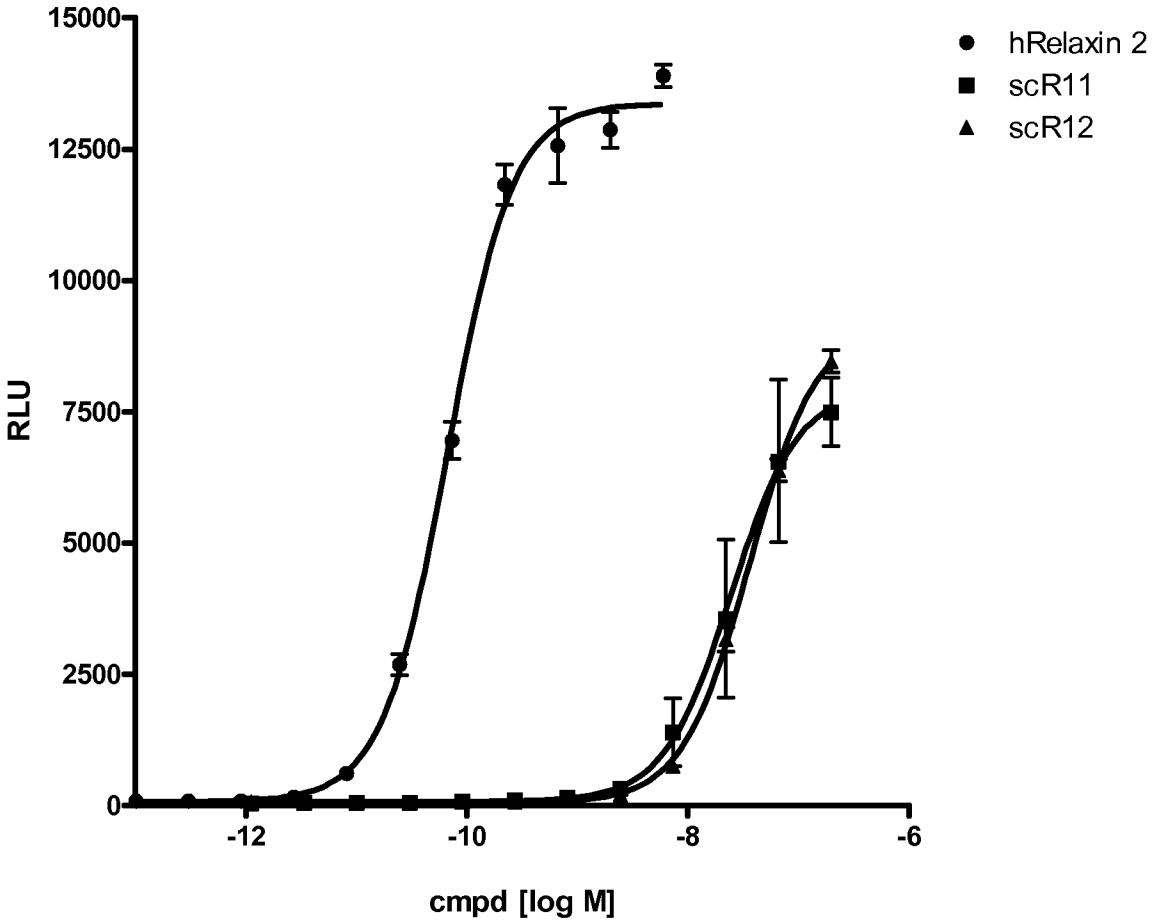
	hRelaxin 2	scR 3	scR 4	scR 5
EC50	2.630e-011	7.769e-011	3.390e-011	3.721e-011

Figure 4b



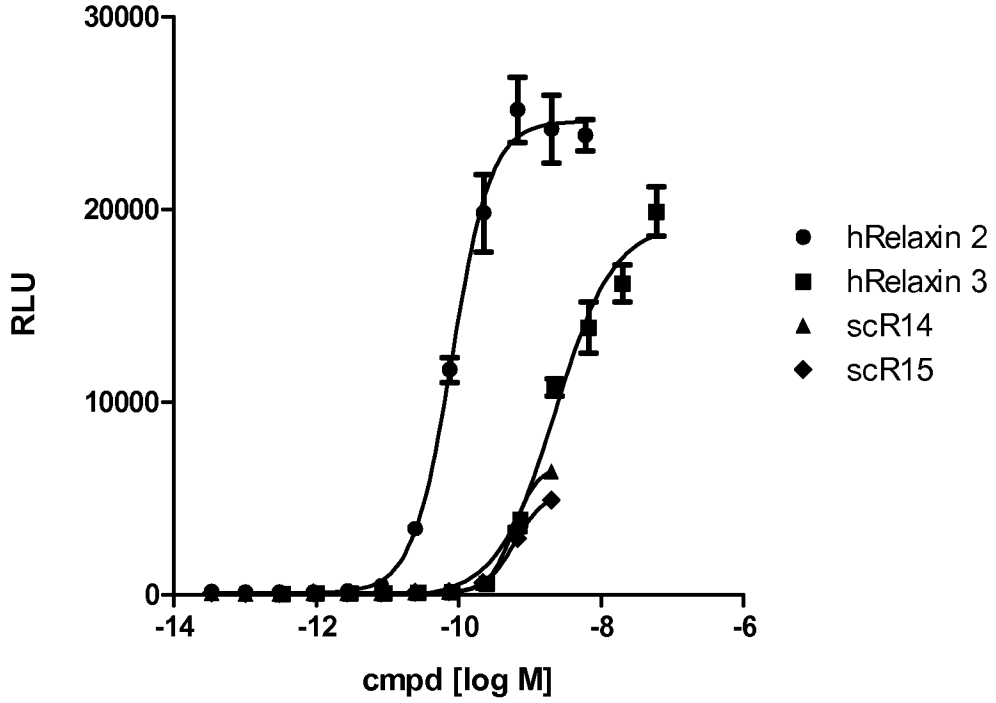
	hRelaxin 2	scR 7	scR 8	scR 9	scR 10
EC50	8.446e-011	5.293e-008	2.382e-007	1.114e-007	4.422e-008

Figure 4c



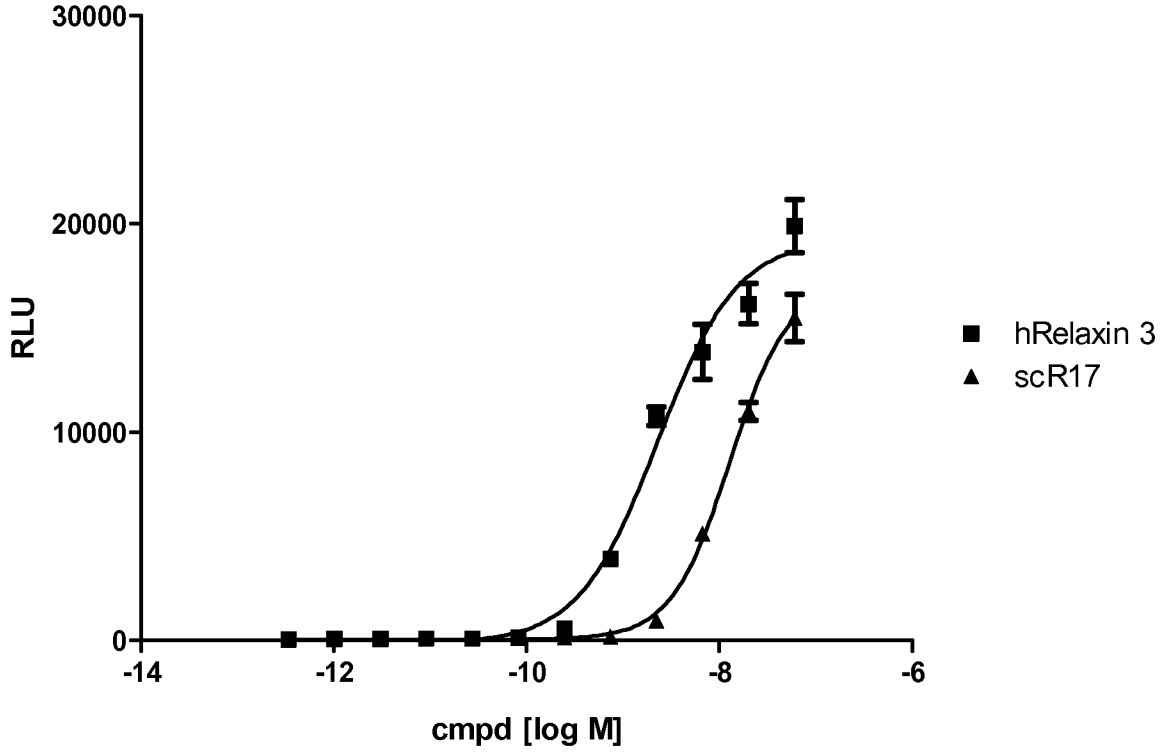
	hRelaxin 2	scR11	scR12
EC50	6.746e-011	2.469e-008	3.640e-008

Figure 4d



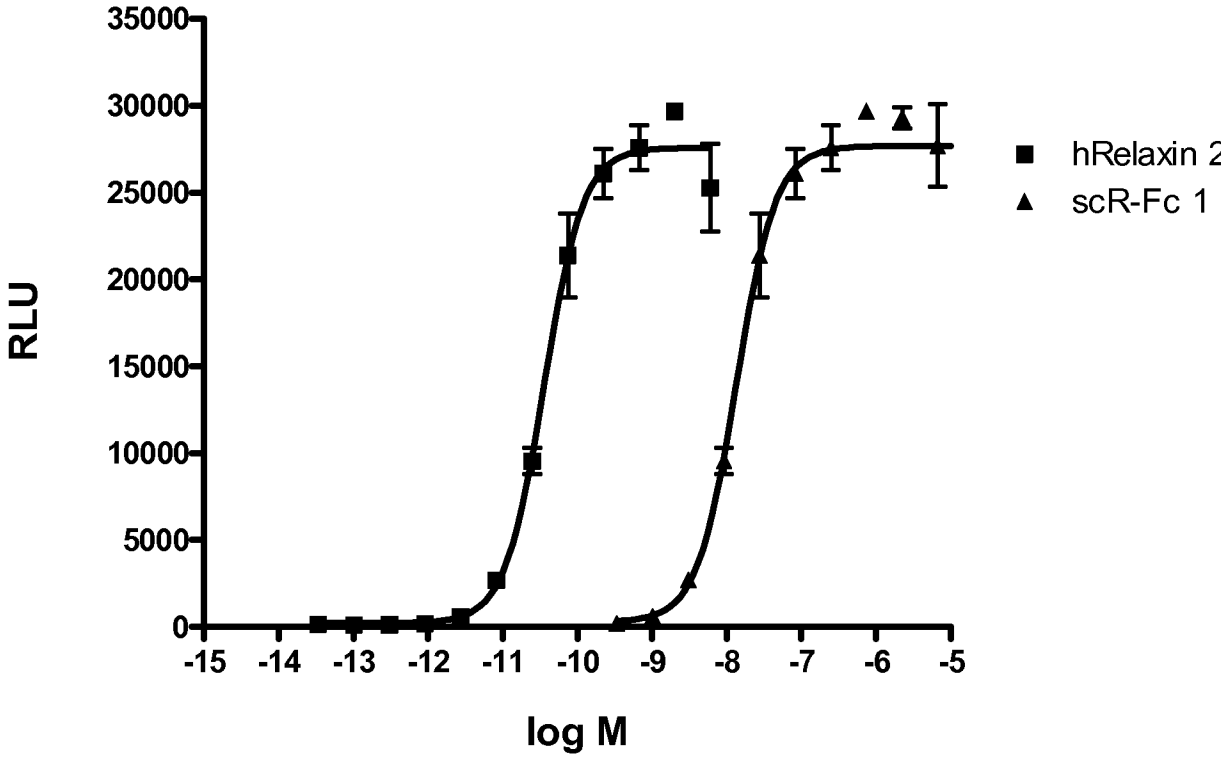
	hRelaxin 2	hRelaxin 3	scR14	scR15
EC50	8.104e-011	2.297e-009	5.891e-010	6.185e-010

Figure 4e



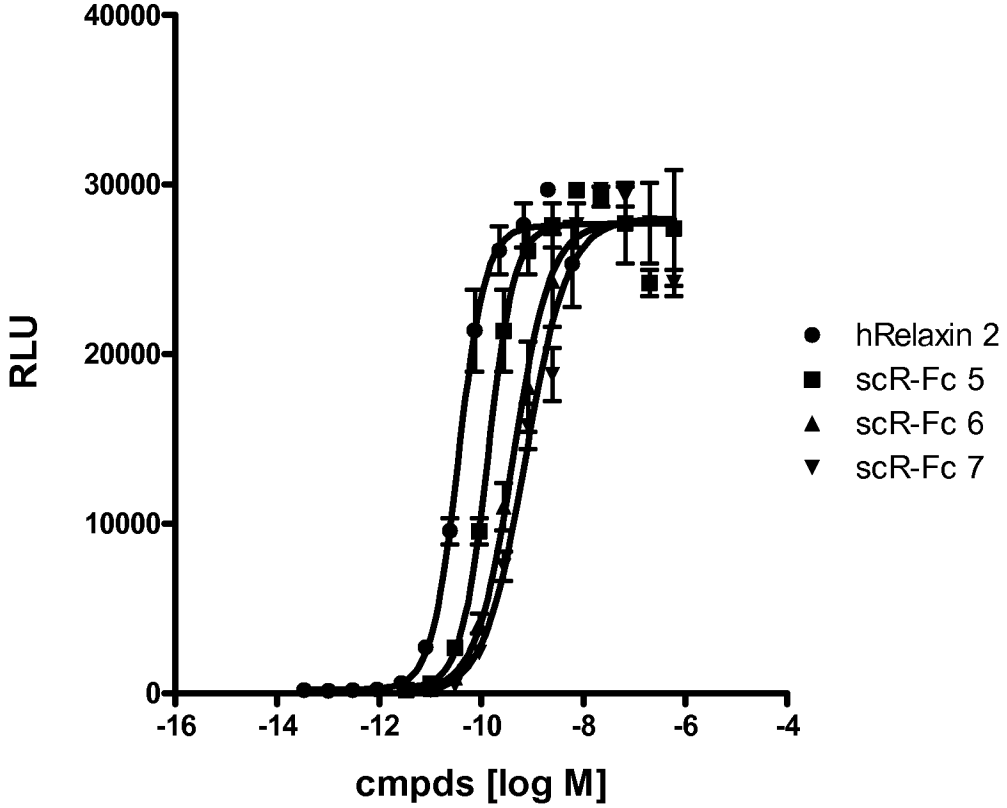
	hRelaxin 3	scR17
EC50	2.297e-009	1.286e-008

Figure 5



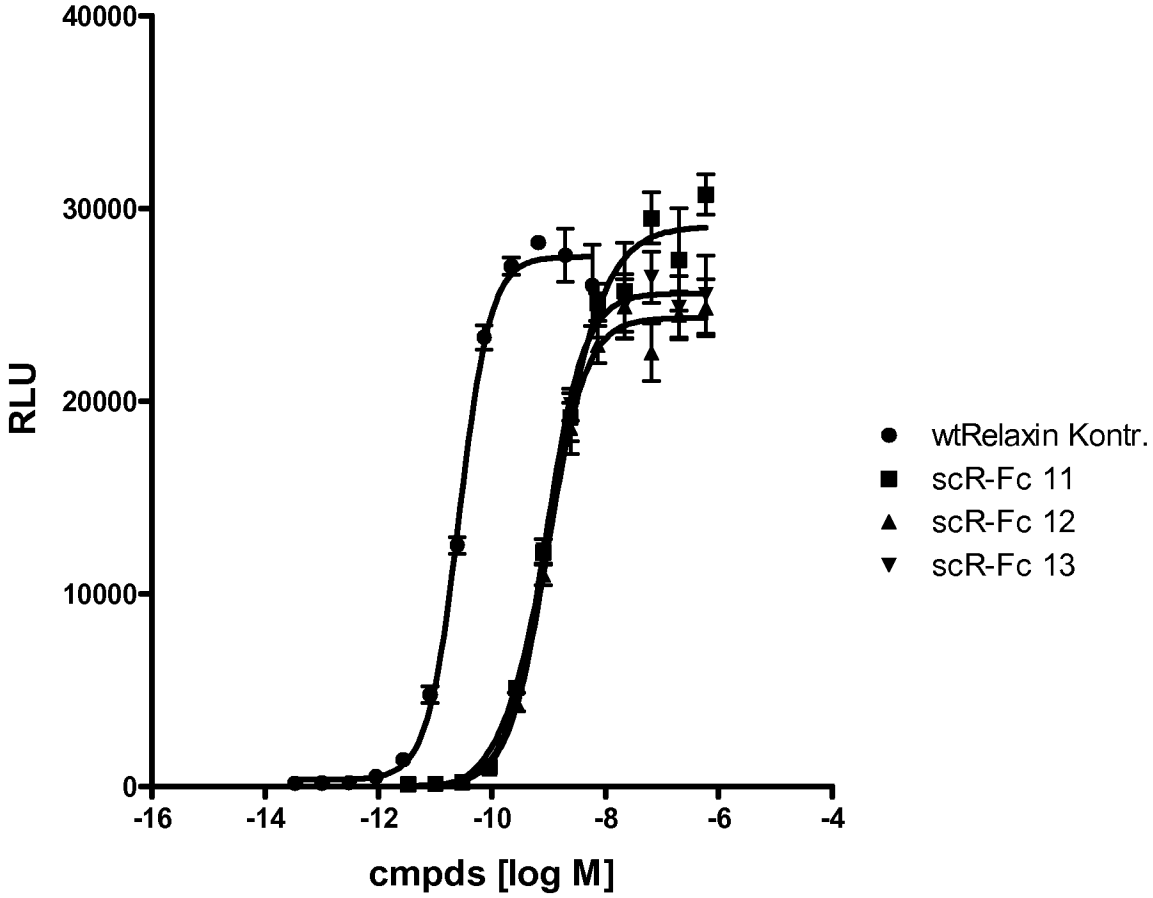
	hRelaxin 2	scR-Fc 1
EC50	3.596e-011	1.345e-008

Figure 6



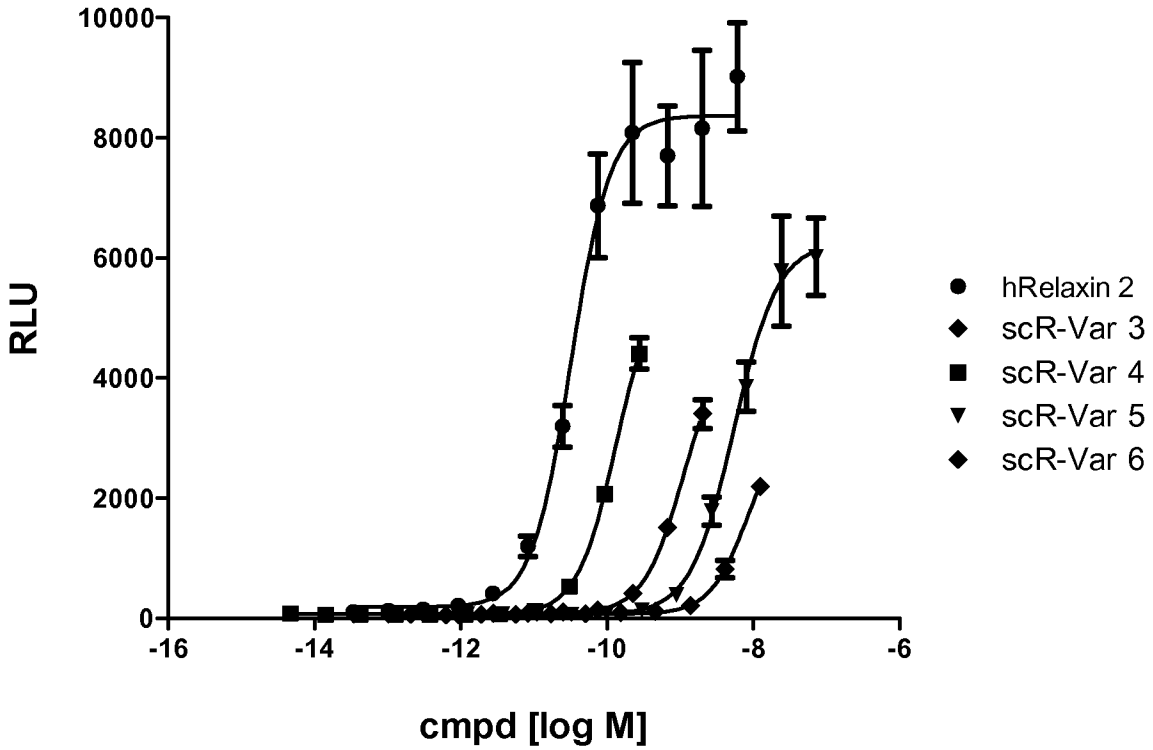
	hRelaxin 2	scR-Fc 5	scR-Fc 6	scR-Fc 7
EC50	3.595e-011	1.346e-010	4.232e-010	7.411e-010

Figure 7



	wtRelaxin Kontr.	scR-Fc 11	scR-Fc 12	scR-Fc 13
EC50	2.664e-011	1.204e-009	9.530e-010	8.953e-010

Figure 8



	hRelaxin 2	scR-Var 3	scR-Var 4	scR-Var 5	scR-Var 6
EC50	3.243e-011	1.119e-009	1.305e-010	5.508e-009	8.368e-009

Figure 9

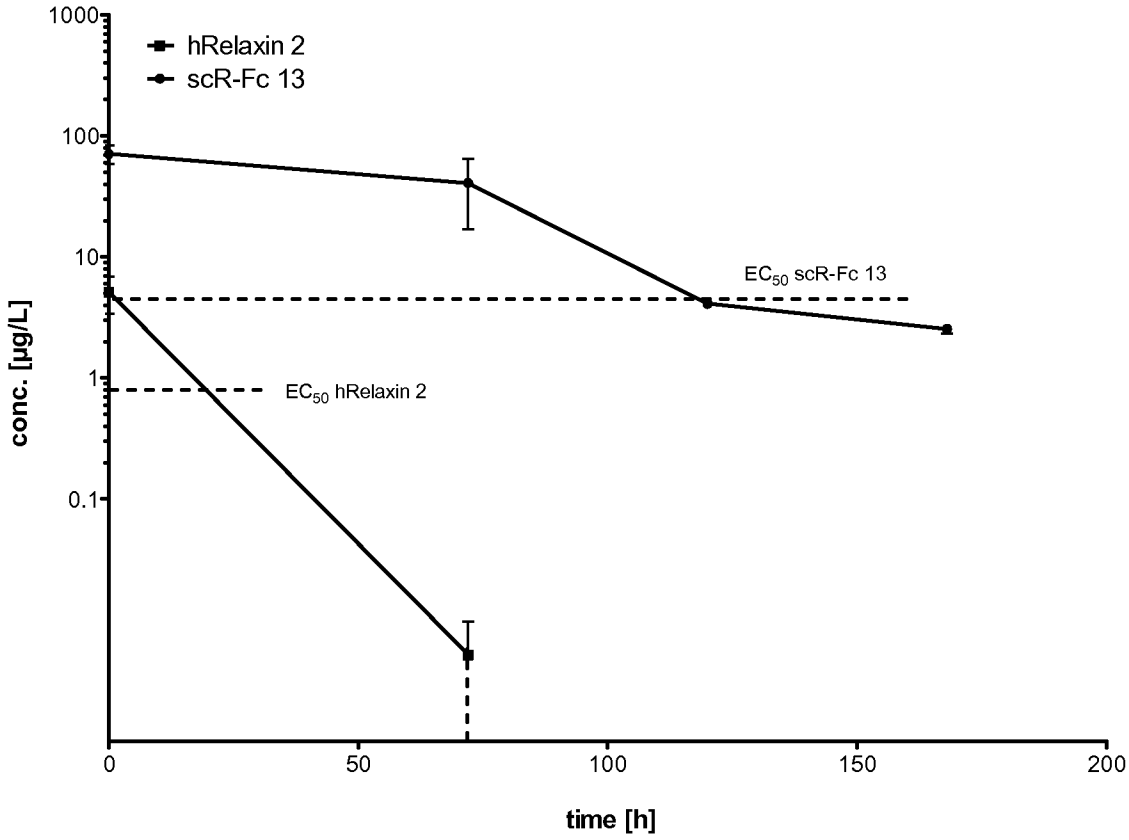
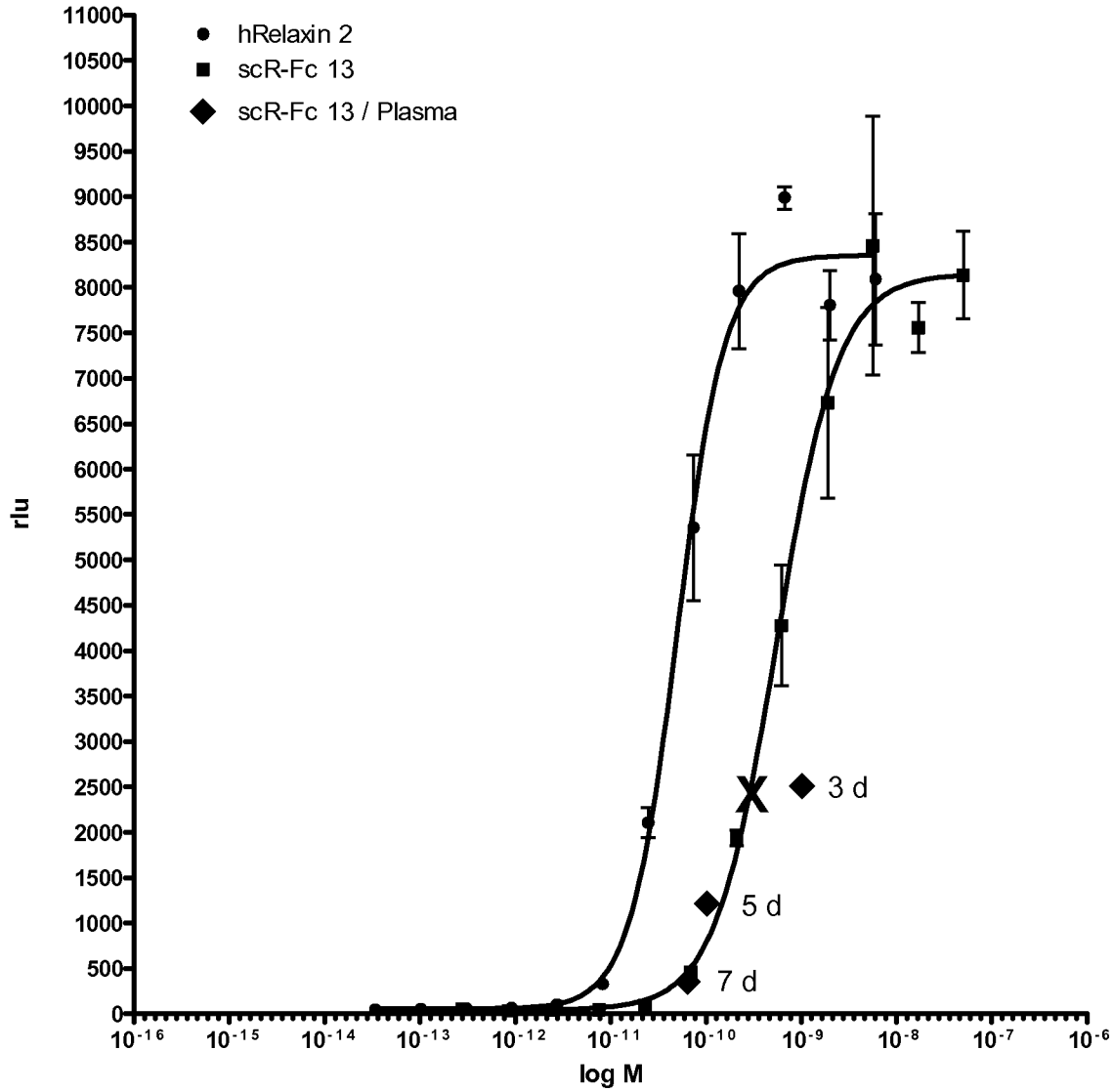

1000
100
10
1
0.1
conc. [µg/L]
hRelaxin 2
scR-Fc 13
EC50 scR-Fc 13
EC50 hRelaxin 2
0
50
100
150
200
time [h]

Figure 10



	hRelaxin 2	scR-Fc 13
EC50	5.010e-011	5.534e-010

Figure 11 a and b

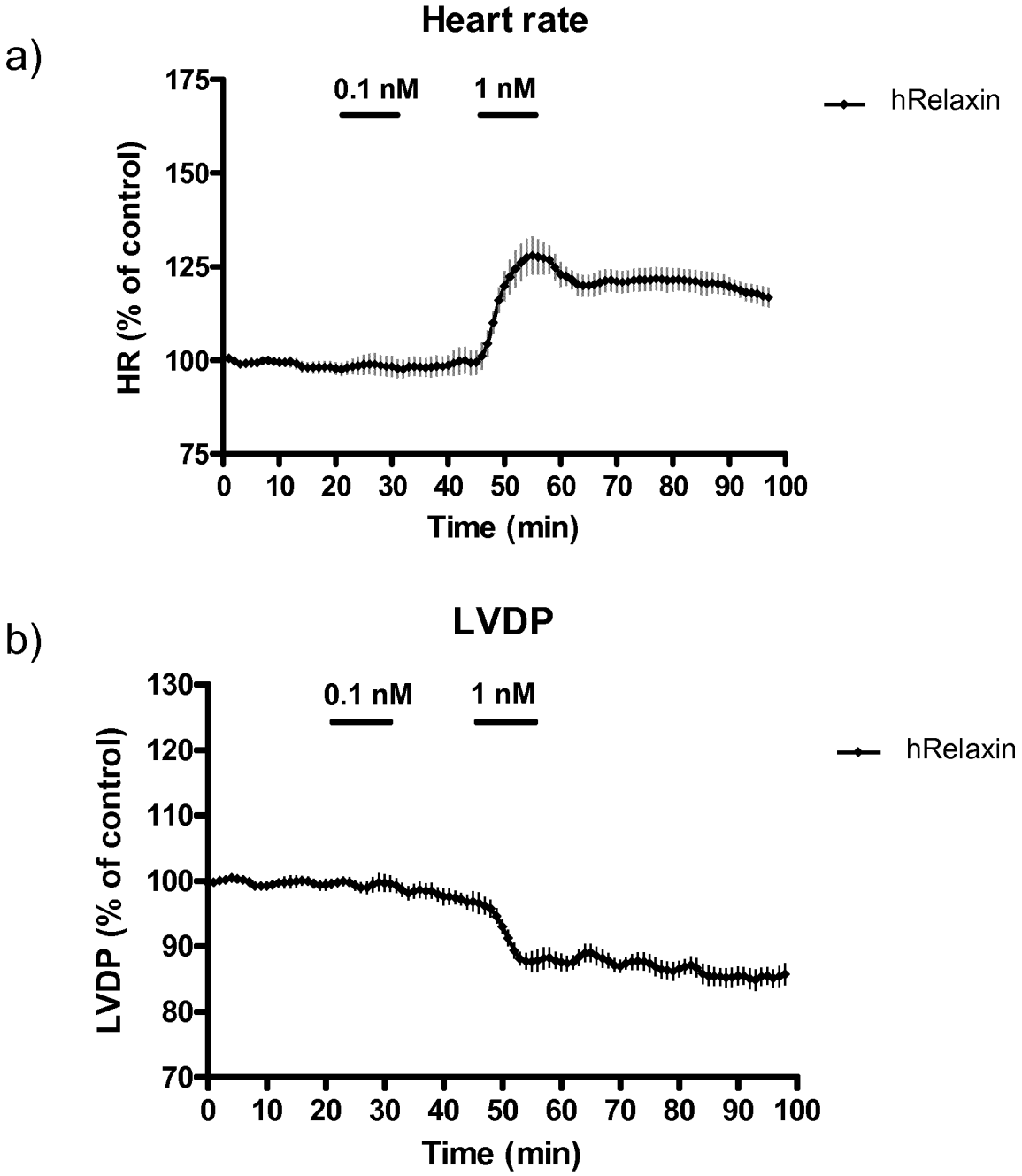


Figure 11 c and d

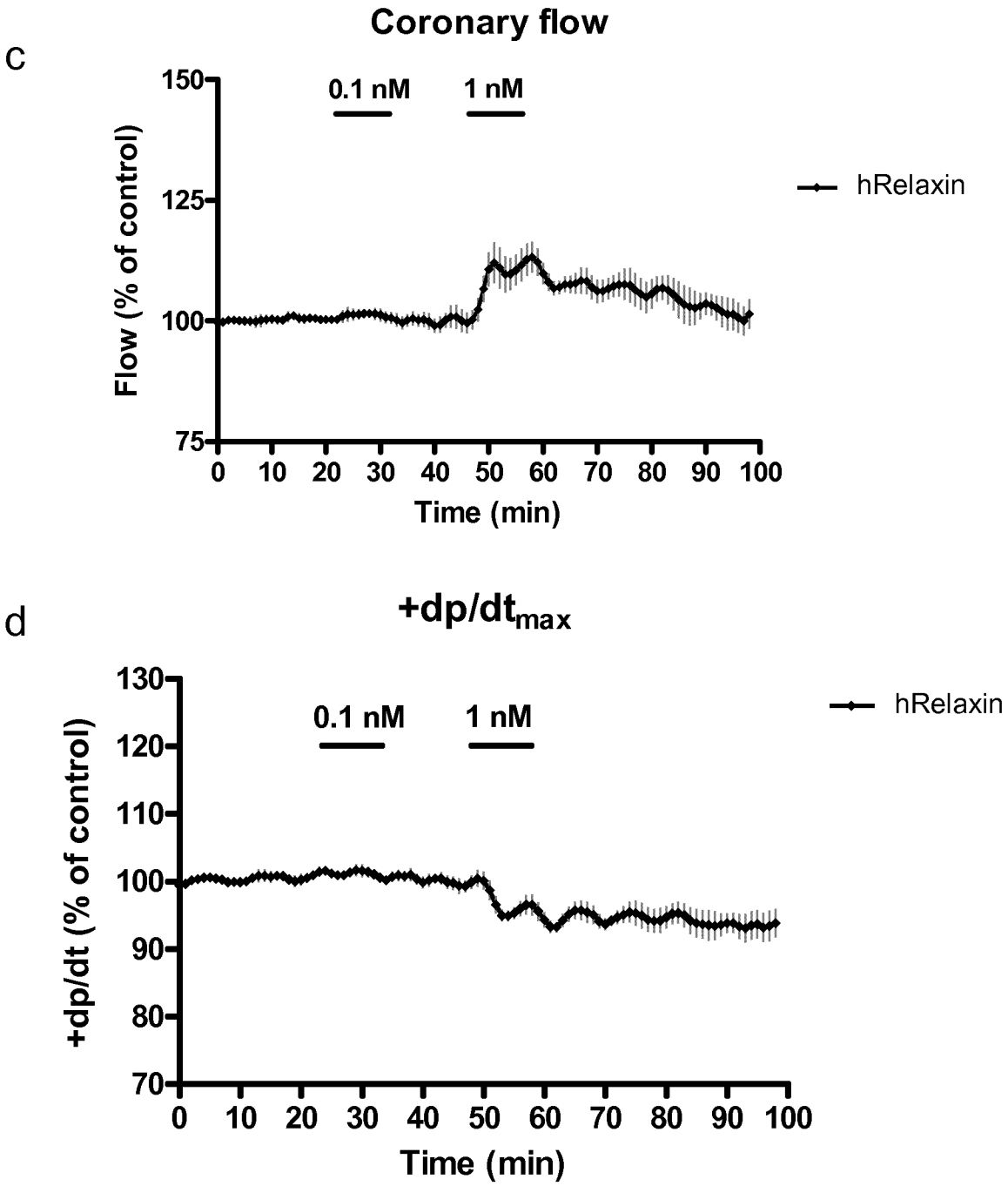


Figure 11 e and f

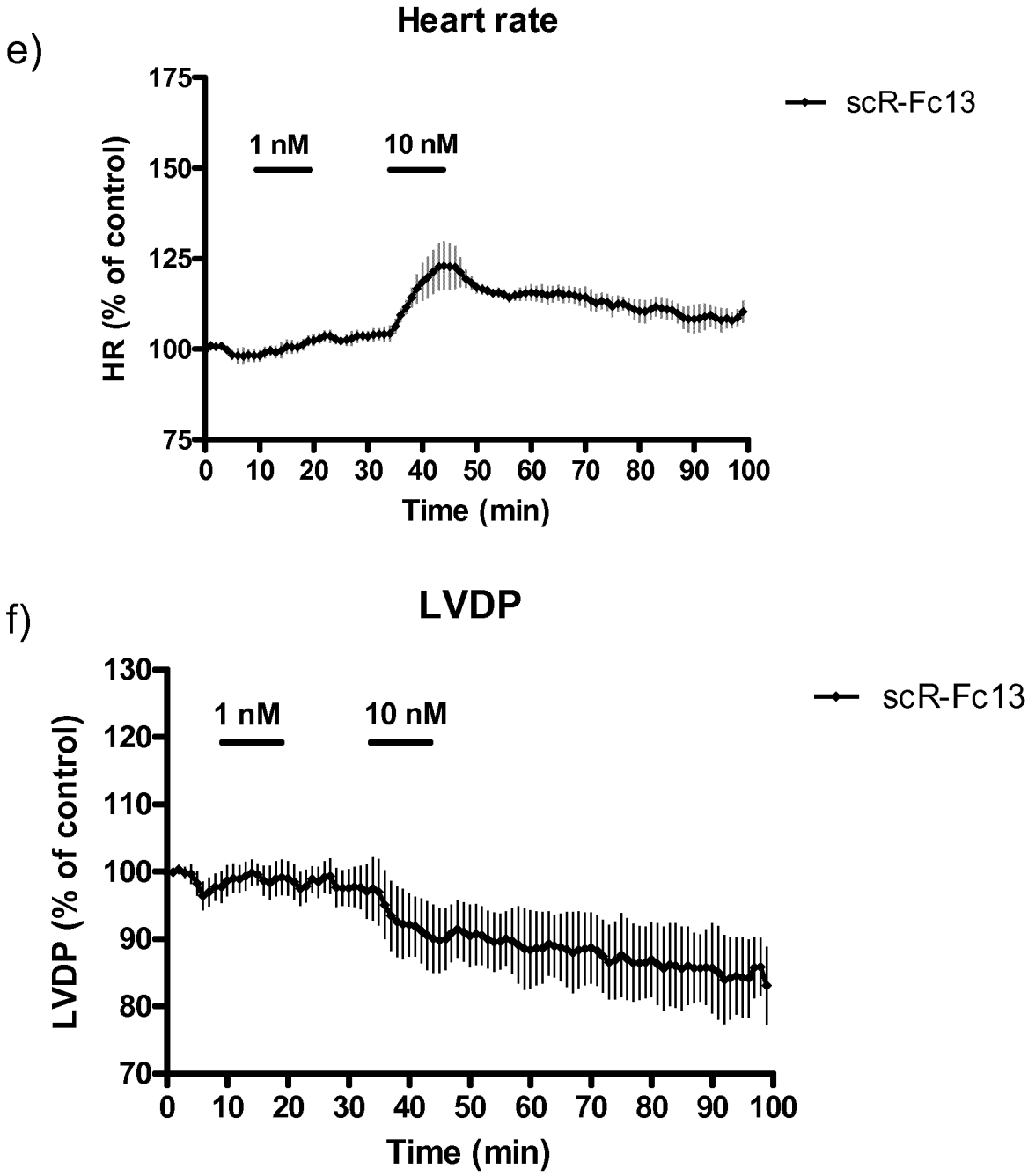
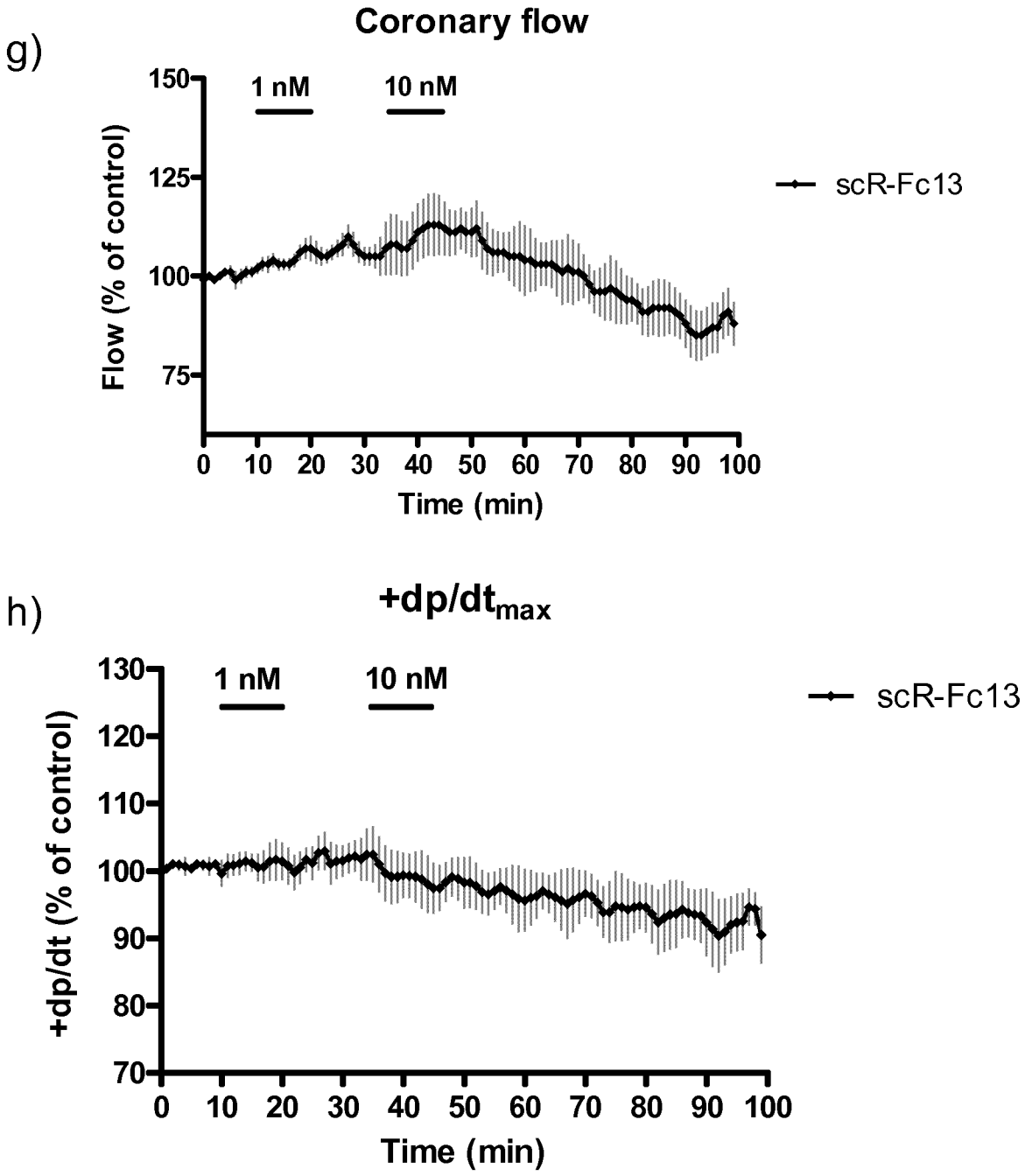


Figure 11 g and h



INTERNATIONAL SEARCH REPORT

International application No
PCT/EP2012/062665

A. CLASSIFICATION OF SUBJECT MATTER
INV. C07K14/64
ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
EPO-Internal, WPI Data, BIOSIS, CHEM ABS Data, Sequence Search, EMBASE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 01/58468 A1 (CONNETICS CORP [US]; UNIV PITTSBURGH [US]; UNIV NEW JERSEY MED [US]) 16 August 2001 (2001-08-16) page 7, lines 4-11,28-34; claims	1-3,5, 7-16
X	US 2011/130332 A1 (PARK JAE-II [US] ET AL) 2 June 2011 (2011-06-02) cited in the application	1-3,7-16
Y	paragraphs [0015], [0044] - [0046]; claims 14,15,20-22; figure 4A	6
X	WO 90/13659 A1 (GENENTECH INC [US]) 15 November 1990 (1990-11-15) abstract; example 5	1,2,7-11

-/--

[X] Further documents are listed in the continuation of Box C.

[X] See patent family annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

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"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

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"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
17 August 2012	28/08/2012

Name and mailing address of the ISA/	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Fax: (+31-70) 340-3016	Pilat, Daniel

1

Form PCT/ISA/210 (second sheet) (April 2005)

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 97/16549 A2 (REGENERON PHARMA [US]; DAVIS SAMUEL [US]) 9 May 1997 (1997-05-09) page 8, line 2 - line 4 page 14, line 24 - line 29; figure 2	1-3,7-12
Y	SCHMIDT SR: "Fusion-proteins as biopharmaceuticals--applications and challenges", CURRENT OPINION IN DRUG DISCOVERY AND DEVELOPMENT, CURRENT DRUGS, LONDON, GB, vol. 12, no. 2, 1 March 2009 (2009-03-01), pages 284-295, XP009132026, ISSN: 1367-6733 abstract	6
X,P	US 2011/243942 A1 (WANG TONY [US]) 6 October 2011 (2011-10-06) the whole document	1-16
X,P	US 2012/046229 A1 (KRAYNOV VADIM [US] ET AL) 23 February 2012 (2012-02-23) the whole document	1-16

INTERNATIONAL SEARCH REPORT

International application No.
PCT/EP2012/062665

Box No. I Nucleotide and/or amino acid sequence(s) (Continuation of item 1.c of the first sheet)

1. With regard to any nucleotide and/or amino acid sequence disclosed in the international application and necessary to the claimed invention, the international search was carried out on the basis of:

   a. (means)

   [ ] on paper

   [X] in electronic form

   b. (time)

   [X] in the international application as filed

   [ ] together with the international application in electronic form

   [ ] subsequently to this Authority for the purpose of search

2. [ ] In addition, in the case that more than one version or copy of a sequence listing and/or table relating thereto has been filed or furnished, the required statements that the information in the subsequent or additional copies is identical to that in the application as filed or does not go beyond the application as filed, as appropriate, were furnished.

3. Additional comments:

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No
PCT/EP2012/062665

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
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