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(71) Applicant: BLACKHAWK GENOMICS LLC [US/US];
3415 56th Street, Lubbock, Texas 79413 (US).

(72) Inventors: TATUM, Owatha; 3415 56th Street, Lubbock,
Texas 79413 (US). LEE, Ji; 1111 Hunt Creek Lane, Sparks,
Maryland 21152 (US).

(74) Agent: ADELSON, Charles N.; Wilson Sonsini Goodrich
& Rosati, 650 Page Mill Road, Palo Alto, California 94304
(US).

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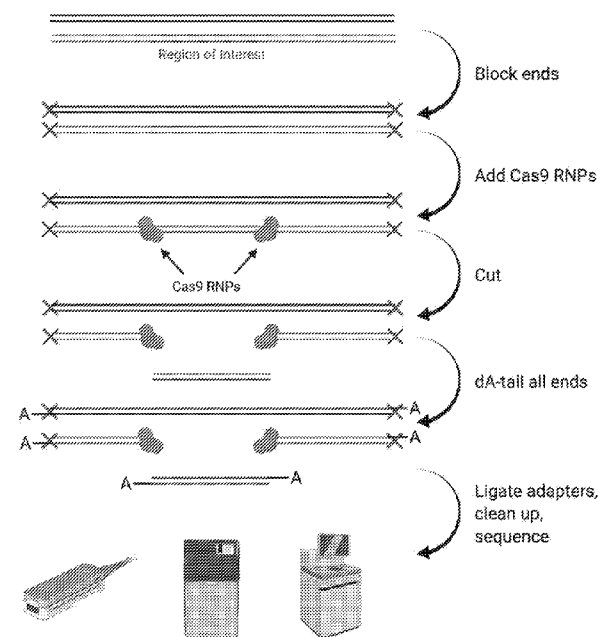


FIG. 2

(57) Abstract: Provided herein are systems and methods for detecting
genomic structural variants using a non-application gene-editing sample
preparation followed by long-read sequencing.

IDENTIFICATION OF GENOMIC STRUCTURAL VARIANTS USING LONG-READ SEQUENCING

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application claims the benefit of U.S. Provisional Application No. 62/913,886 filed October 11, 2019; and of U.S. Provisional Application No. 62/981,146, filed February 25, 2020, each of which is incorporated herein by reference.

BACKGROUND OF THE INVENTION

[0002] A genetic abnormality or genomic variation in the genetic makeup of an individual can cause a genetic disease or disorder in the individual. The genetic abnormality or genomic variation can range for a discrete mutation in a single base (e.g. single nucleotide variant) to a chromosomal abnormality or structural variant (SV) (e.g. copy number variant, segmental inversions, etc.) comprising the rearrangement, addition or deletion of one or more genes. Currently, more than 100,000 genetic variants have been classified as disease-causing in public databases. For example, sickle cell disease is caused by a single nucleotide mutation in the beta-globin gene; Fragile X syndrome is caused by tandem duplication of the CGG trinucleotide repeated over 200 times; and Down Syndrome is commonly caused by complete duplication of chromosome 21. Short-read sequencing technologies can identify small genomic variations such as single nucleotide variants, insertions and deletions, with high accuracy. However, these technologies are unable to identify structural variants larger than a few hundred base pairs with good accuracy. Several methods have emerged to try to detect structural variants; but they all have their limitations. For example, microscopy using fluorescent probes is low-throughput, is quite expensive, and has low resolution. Quantitative PCR (qPCR) and microarray assays are high-throughput and inexpensive but cannot identify unknown structural variants. Short-read sequencers, which are high-throughput and inexpensive, have difficulty resolving SVs and frequently are coupled with another technology, such as optical mapping or linked read sequencing, to identify SVs accurately. Whole genome sequencing using long-read sequencers can be used to detect large structural variants; however, whole genome sequencing is expensive, and some long-read sequencers have difficulty resolving very large structural variants. As such, there is an immediate need, especially in the clinical setting, for a fast, high-throughput yet cost-effective method to identify genomic structural variants, in particular, de novo structural variants.

SUMMARY OF THE INVENTION

[0003] In one aspect, provided herein is a method for identifying a set of guide RNAs (gRNAs) that are hybridizable to a genomic region of interest in a genome comprising designing a

plurality of gRNAs, wherein at least one gRNA is hybridizable to a target site within the genomic region of interest and is configured to produce a genomic variant that comprises at least 1000 bp; and said plurality of gRNAs comprises a plurality of CRISPR RNAs (crRNAs), wherein said plurality of crRNAs comprises a GC of at least about 20% to about 80%.

[0004] In another aspect, provided herein is a method for identifying a set of guide RNAs (gRNAs) that are hybridizable to a genomic region of interest in a genome comprising designing a plurality of gRNAs, wherein at least one gRNA is hybridizable to a target site within the genomic region of interest and is configured to produce a genomic variant that comprises at least 1000 bp; and said plurality of gRNAs comprises a plurality of CRISPR RNAs (crRNAs), wherein said plurality of crRNAs comprises a self-complementarity score of zero.

[0005] In another aspect, provided herein is a method for identifying a set of guide RNAs (gRNAs) that are hybridizable to a genomic region of interest in a genome comprising designing a plurality of gRNAs, wherein at least one gRNA is hybridizable to a target site within the genomic region of interest and is configured to produce a genomic variant that comprises at least 1000 bp; and said plurality of gRNAs comprises a plurality of CRISPR RNAs (crRNAs), wherein said plurality of crRNAs comprises an efficiency score of about 0.2.

[0006] In another aspect, provided herein is a method for identifying a set of guide RNAs (gRNAs) that are hybridizable to a genomic region of interest in a genome comprising designing a plurality of gRNAs, wherein at least one gRNA is hybridizable to a target site within the genomic region of interest and is configured to produce a genomic variant that comprises at least 1000 bp; and said plurality of gRNAs comprises a plurality of CRISPR RNAs (crRNAs), wherein said plurality of crRNAs comprises a mismatch profile of $MM0 < 2$, $MM1 < 3$, $MM2 < 3$, and $MM3 < 21$.

[0007] In some embodiments, the plurality of crRNAs comprises a mismatch profile of $MM3 < 5$.

[0008] In another aspect, provided herein is a method of detecting a genomic variant in a sample, the method comprising enriching said sample for a genomic region of interest comprising said genomic variant using a gene-editing based approach; and sequencing said enriched sample comprising said genomic region of interest using long-read sequencing.

[0009] In some embodiments, said genomic variant comprises a structural variant. In some cases, said genomic variant comprises at least 50 bp. In some embodiments, said genomic variant comprises a structural variant. In some cases, said genomic variant comprises at least 1000 bp.

[0010] In some embodiments, said gene-editing based approach comprises use of a clustered regularly interspersed short palindromic repeats (CRISPR)-Cas system. In some cases, said CRISPR-Cas system comprises Cas9.

[0011] In some embodiments, step (a) of enriching of said sample further comprises amplification of said genomic region of interest. In some embodiments, step (a) of enriching said sample does not require amplification of said genomic region of interest. In some embodiments, step (a) of enriching of said sample further comprises coupling a sequence of dAMPs to said genomic variant. In some embodiments, step (a) of enriching of said sample further comprises coupling a plurality of barcode molecules to said genomic variant. In some embodiments, step (a) of enriching of said sample further comprises coupling said genomic variant to a magnetic bead.

[0012] In some embodiments, said long-read sequencing comprises nanopore sequencing. In some embodiments, said long-read sequencing comprises single molecule, real-time (SMRT) sequencing.

[0013] In some embodiments, said CRISPR-Cas system further comprises a crRNA comprising a sequence of Tables 1-117.

[0014] In some embodiments, said genomic region of interest comprises two or more repeat regions. In some embodiments, said genomic region of interest comprises a GC content of greater than 30%.

[0015] In some embodiments, said sample comprises at least 10 genomic regions of interest.

[0016] In some embodiments, said genomic variant is associated with a disorder. In some cases, the disorder is selected from the group consisting of acute lymphoblastic leukemia (ALL), alpha-thalassemia, ataxia-telangiectasia (AT), autosomal recessive deafness 16, autosomal recessive deafness 22, beta-thalassemia, breast cancer, Canavan disease, cancer, celiac disease, chronic myeloid leukemia (CML), cystic fibrosis, cystinosis, deafness infertility syndrome (DIS), Duchenne muscular dystrophy, Ehlers-Danlos syndrome type III and IV, Ellis-van Creveld syndrome, Fabry disease, familial adenomatous polyposis (FAP), familial cutaneous melanoma, Fragile X, gastric cancer (including hereditary diffuse gastric cancer), Gaucher disease, hereditary predisposition to develop cancer, Huntington disease, hypophosphatasia (HPP), incontinentia pigmenti, Krabbe disease, Leber congenital amaurosis (LCA), Loeys-Dietz syndrome, Long QT syndrome, Lynch syndrome, Marfan syndrome, mental disorder, medium-chain acyl-coenzyme A dehydrogenase (MCAD) deficiency, MUTYH-associated polyposis, neuroblastoma, neuronal ceroid-lipofuscinoses (NCLs), Niemann-Pick Type C disease, pancreatic cancer syndromes, papillary renal carcinoma, Parkinson disease, phenylketonuria, Pompe disease, propionic acidemia, rheumatoid arthritis, solid tumors, spinal muscular atrophy, spinocerebellar ataxia, susceptibility to breast cancer, Tay-Sachs disease, very long-chain acyl-coenzyme A dehydrogenase deficiency, Von Hippel-Lindau syndrome, Wilms tumor, Wilson

disease, Wolfram syndrome type 1, X-linked creatine deficiency syndrome, X-linked hemophilia A, X-linked retinitis pigmentosa.

[0017] Provided herein is a method of designing a probe to target a genomic region of interest, the method comprising designing a plurality of nucleic acid probe options to target said genomic region of interest; selecting a first set of candidates from said plurality of nucleic acid probe options with a GC content of at least 20%; selecting a second set of candidates from said first set of candidates with a self-complementarity score of zero or a complementarity score of 1; selecting a third set of candidates from said second set of candidates with an efficiency greater than 0.2; and selecting a fourth set of candidates from said third set of candidates with a mismatch profile of MM0=0 or MM0=1, MM1=0 or MM1=1 or MM1=2, MM2=0 or MM2=1 or MM2=2, and MM3 <21, wherein said fourth set of candidates comprises said probe to target a genomic region of interest, wherein said fourth set of candidates comprises said probe to target a genomic region of interest.

[0018] In some embodiments, the fourth set of candidates comprises a mismatch profile of MM3<5. In some embodiments, said designing comprises using CHOPCHOP.

[0019] In some embodiments, said first set of candidates have a GC content of about 40% to about 80%.

[0020] In some embodiments, said nucleic acid probe of interest comprises a crRNA. In some embodiments, the probability of said crRNA cutting said genomic region of interest is greater than or equal to 80%. In some embodiments, the method further comprises estimating on-target value of said crRNA. In some embodiments, the method further comprises estimating off-target value of said crRNA.

[0021] In another aspect, provided herein is a kit comprising a set of guide RNAs (gRNAs) that are hybridizable to a genomic region of interest in a genome comprising designing a plurality of gRNAs, wherein at least one gRNA is hybridizable to a target site within the genomic region of interest and is configured to produce a genomic variant that comprises at least 1000 bp; and said plurality of gRNAs comprises a plurality of CRISPR RNAs (crRNAs), wherein said plurality of crRNAs comprises a GC of at least about 40% to about 80%.

[0022] In another aspect, provided herein is a kit comprising a set of guide RNAs (gRNAs) that are hybridizable to a genomic region of interest in a genome comprising designing a plurality of gRNAs, wherein at least one gRNA is hybridizable to a target site within the genomic region of interest and is configured to produce a genomic variant that comprises at least 1000 bp; and said plurality of gRNAs comprises a plurality of CRISPR RNAs (crRNAs), wherein said plurality of crRNAs comprises a self-complementarity score of zero.

[0023] In another aspect, provided herein is a kit comprising a set of guide RNAs (gRNAs) that are hybridizable to a genomic region of interest in a genome comprising designing a plurality of gRNAs, wherein at least one gRNA is hybridizable to a target site within the genomic region of interest and is configured to produce a genomic variant that comprises at least 1000 bp; and said plurality of gRNAs comprises a plurality of CRISPR RNAs (crRNAs), wherein said plurality of crRNAs comprises an efficiency score of about 0.2.

[0024] In another aspect, provided herein is a kit comprising a set of guide RNAs (gRNAs) that are hybridizable to a genomic region of interest in a genome comprising designing a plurality of gRNAs, wherein at least one gRNA is hybridizable to a target site within the genomic region of interest and is configured to produce a genomic variant that comprises at least 1000 bp; and said plurality of gRNAs comprises a plurality of CRISPR RNAs (crRNAs), wherein said plurality of crRNAs comprises a mismatch profile of MM0=0 or MM0=1, MM1=0 or MM1=1 or MM1 = 2, MM2=0 or MM2 = 1 or MM2=2, and MM3 <21.

INCORPORATION BY REFERENCE

[0025] All publications, patents, and patent applications mentioned in this specification are herein incorporated by reference to the same extent as if each individual publication, patent, or patent application was specifically and individually indicated to be incorporated by reference. To the extent publications and patents or patent applications incorporated by reference contradict the disclosure contained in the specification, the specification is intended to supersede and/or take precedence over any such contradictory material.

BRIEF DESCRIPTION OF THE DRAWINGS

[0026] The novel features of the invention are set forth with particularity in the appended claims. A better understanding of the features and advantages of the present invention will be obtained by reference to the following detailed description that sets forth illustrative embodiments, in which the principles of the invention are utilized, and the accompanying drawings (also “Figure” and “FIG.” herein), of which:

[0027] FIG. 1 provides exemplary genomic abnormalities and variants.

[0028] FIG. 2 provides an exemplary target enrichment sample preparation approach, in accordance with the embodiments provided herein.

[0029] FIG. 3 provides an exemplary design approach for crRNA probes, in accordance with the embodiments provided herein.

[0030] FIGS. 4A and 4B provide exemplary coverage of a crRNA probe embodiment, in accordance with the embodiments provided herein.

[0031] FIG. 5 provides an exemplary computer control system that is programmed to implement the methods provided, in accordance with the embodiments provided herein.

[0032] FIG. 6 provides an exemplary design approach for crRNA probes, in accordance with the embodiments provided herein.

DETAILED DESCRIPTION OF THE INVENTION

[0033] While various embodiments of the invention have been shown and described herein, it will be obvious to those skilled in the art that such embodiments are provided by way of example only. Numerous variations, changes, and substitutions may occur to those skilled in the art without departing from the invention. It should be understood that various alternatives to the embodiments of the invention described herein may be employed.

[0034] Where values are described as ranges, it will be understood that such disclosure includes the disclosure of all possible sub-ranges within such ranges, as well as specific numerical values that fall within such ranges irrespective of whether a specific numerical value or specific sub-range is expressly stated.

[0035] The terms “a,” “an,” and “the,” as used herein, generally refers to singular and plural references unless the context clearly dictates otherwise.

[0036] The term “subject,” as used herein, generally refers to an animal, such as a mammal (e.g., human) or avian (e.g., bird), or other organism, such as plant. For example, the subject can be a vertebrate, a mammal, a rodent (e.g., a mouse), a primate, a simian, or a human. Animals may include, but are not limited to, farm animals, sport animals, and pets. A subject may be a healthy or asymptomatic individual, an individual that has or is suspected of having a disease (e.g., a genetic disorder) or a pre-disposition to a disease, and/or an individual that is in need of therapy or suspected of needing therapy. A subject can be a patient.

[0037] The term “genome,” as used herein, generally refers to genomic information from a subject, which may be, for example, at least a portion or an entirety of a subject’s hereditary information. A genome can be encoded either in DNA or in RNA. A genome can include the sequence of all chromosomes together in an organism. For example, the human genome ordinarily has a total of 46 chromosomes. The sequence of all these together may constitute a human genome.

[0038] The term “sequencing,” as used herein, generally refers to methods and technologies for determining the sequence of nucleotide bases in one or more polynucleotides. The polynucleotides can be, for example, nucleic acid molecules such as deoxyribonucleic acid (DNA) or ribonucleic acid (RNA), including variants or derivatives thereof (e.g., single stranded DNA). Sequencing can be performed by various systems currently available, such as, without

limitation, sequencing system by Illumina®, Pacific Biosciences (PacBio®), Oxford Nanopore®, Life Technologies (Ion Torrent®), Roche®, Genapsys®, and MGI Tech®. Sequencing may be performed without using nucleic acid amplification. Alternatively, or in addition, sequencing may be performed using nucleic acid amplification, polymerase chain reaction (PCR) (e.g. digital PCR, quantitative PCR, or real time PCR), or isothermal amplification. Such systems may provide a plurality of raw genetic data corresponding to the genetic information of a subject (e.g., human), as generated by the systems from a sample provided by the subject. In some examples, such systems provide sequencing reads (also “reads” herein). A read may include a string of nucleic acid bases corresponding to a sequence of a nucleic acid molecule that has been sequenced. In some situations, systems and methods provided herein may be used with proteomic information.

[0039] The term “sample,” as used herein, generally refers to a biological sample of a subject. The biological sample may comprise any number of macromolecules, for example, cellular macromolecules. The sample may be a cell sample. The sample may be a cell line or cell culture sample. The sample can include one or more cells. The sample may include one or more microbes. The biological sample may be a nucleic acid sample or protein sample. The biological sample may also be a carbohydrate sample or a lipid sample. The biological sample may be derived from another sample. The sample may be a tissue sample, such as a biopsy, core biopsy, needle aspirate, or fine needle aspirate. The sample may be a fluid sample, such as blood sample, urine sample, or saliva sample. The sample may be a skin sample. The sample may be a cheek swab. The sample may be a plasma or serum sample. The sample may include cells or may be cell-free. A cell-free sample may include extracellular polynucleotides. Extracellular polynucleotides may be isolated from a bodily sample that may be selected from the group consisting of blood, plasma, serum, urine, saliva, mucosal excretions, sputum, stool and tears.

[0040] The term “short read,” as used herein, generally refers to a read length of a DNA or RNA polynucleotide of about 100 to about 600 bp.

[0041] The term “long read,” as used herein, generally refers to a read length of a DNA or RNA polynucleotide of greater than 1 Kbp.

[0042] The term “ribonucleoprotein (RNP),” as used herein is a ribonucleoprotein is a ribonucleic acid (RNA)-protein complex.

[0043] The term “CRISPR-Cas system,” as used herein, generally refers the clustered regularly short palindromic repeats (CRISPR system) which comprises an array of two types of DNA sequences: (i) repetitive, flanking DNA sequences; and (ii) spacer sequences that are endogenously derived from a virus, and can be used to target DNA or RNA sequences for

cleaving using the CRISPR-associated (Cas) enzyme (ribonucleoprotein) complex that are used to cleave the CRISPR sites that are complementary to those in spacer regions.

[0044] The term “barcoding,” as used herein, is the ligation of known, unique sequences to target DNA molecules, between the adapter and the ROI in order for the target sequence recognition in the downstream analysis, i.e. post-base calling.

[0045] The term “multiplexing,” as used herein, is the running of multiple samples in a single flow cell, identifying each sample’s DNA molecules through unique ‘barcode’ molecules that have been attached to the DNA ends. The decoded sequences of a sample’s DNA will be identified downstream once the sequences have been basecalled.

[0046] The term “crRNA,” as used herein, are the RNA sequences that recognize the target site. Together with the tracrRNA, this forms a single guide RNA (sgRNA) and when several are used together, gRNA.

[0047] The term “tracrRNA,” as used herein, refers to trans-activating-crRNA specific to Type II Cas/CRISPR system. It is used to process the pre-crRNA along with an RNase III. The tracrRNA provides structural support to the ribonucleoprotein and anneals to the pre-crRNA for processing via the internal endonuclease activity of the Cas protein. Non-limiting examples of Cas enzymes can include Cas1, Cas1B, Cas2, Cas3, Cas4, Cas5, Cas6, Cas7, Cas8, Cas9 (also known as Csn1 or Csx12), Cas10, Csy1, Csy2, Csy3, Cse1, Cse2, Csc1, Csc2, Csa5, Csn2, Csm2, Csm3, Csm4, Csm5, Csm6, Cmr1, Cmr3, Cmr4, Cmr5, Cmr6, Csb1, Csb2, Csb3, Csx17, Csx14, Csx10, Csx16, CsaX, Csx3, Csx1, Csx1S, Csf1, Csf2, CsO, Csf4, Cpf1, c2c1, c2c3, Cas9HiFi, homologues thereof, or modified versions thereof. In some cases, a catalytically dead Cas protein can be used, for example a dCas9. An unmodified CRISPR enzyme can have DNA cleavage activity, such as Cas9.

[0048] The term “protospacer,” as used herein, refers to a sequence acquired from a pathogenic organism’s DNA molecule. The sequence is converted into DNA and forms the gene of the crRNA, which, along with the PAM in the substrate sequence, directs the Cas-crRNA-tracrRNA ternary complex to cleave target molecule.

[0049] The term “protospacer adjacent motif (PAM),” as used herein, refers to pathogenic sequences from host sequences i.e. the crRNA gene. It is adjacent to the 3’ end of the protospacer and facilitates the pathogen’s sequence to be cut by the Cas-crRNA-tracrRNA ternary complex. In Type II CRISPR systems, NGG (where N is any nucleotide as per FASTA conventions), defines the sequences that will be cleaved. If a PAM is 3’ of crRNA sequence in the DNA, the crRNA-tracrRNA-Cas9 ternary complex will not cleave the host DNA/genome sequence.

[0050] The term “untranslated region (UTR),” as used herein, refers to the untranslated region (UTR) of a mRNA transcript and is present both at the 5’ and 3’ ends of the protein coding region. It is not translated via the protein synthesis process by the ribosome.

[0051] The human genome has many different types of genomic variations that range in size and type. FIG. 1 shows examples of genomic variations. A single nucleotide variant is a substitution of a single nucleotide at a specific position in the genome. A deletion is a loss of one or more nucleotides in the genome, ranging from a single base to an entire chromosome. In contrast, an insertion is the addition of one or more nucleotides to the genome. A tandem repeat consists of two or more adjacent copies of a sequence of at least two nucleotides in length. A tandem duplication occurs when a nucleotide sequence, which itself can contain a repeated sequence, is copied into two adjacent copies. Interspersed duplication differs from tandem duplication or repeat in that the repeated sequence is dispersed throughout the genome and is nonadjacent to the original copy. Inversion is a chromosome rearrangement in which a segment of a gene, structural element or chromosome is reversed end to end. Translocation is the unusual rearrangement of chromosomes. Copy number variants is a type of structural repetition in which one or more parts of the genome are repeated.

[0052] The types of genomic variants can be categorized based on the number of nucleotides involved. Single nucleotide variants (SNVs) affect a single nucleotide or base pair. Small insertions and deletions, commonly called indels, are shorter than 50 nucleotides in length. Structural variants are changes in the structure of chromosome and generally affect 50 or more nucleotides. The typical human genome has about 8 million bases that differ from a reference due to SNVs and indels. The typical human genome has about 20,000 structural variants that differ from the reference and affects about 10 million bases.

[0053] In one aspect, provided herein are systems and methods to detect one or more genomic variants in a sample, comprising (a) preparing a sample for sequencing using a non-amplification-based, gene-editing based approach, and (b) long-read sequencing, as described herein elsewhere.

[0054] In another aspect, provided herein are systems and methods for conducting a diagnostic assay for a genetic disorder, comprising (a) preparing a sample for sequencing using a non-amplification-based, gene-editing based approach, and (b) long-read sequencing, as described herein elsewhere.

[0055] In some cases, the one or more genomic variants comprise one or more structural variants. In some cases, the one or more genomic variants comprise at least one structural variant. In some cases, the structural variant is about 30 bp to about 1,000 bp. In some cases, the

structural variant is about 30 bp to about 50 bp, about 30 bp to about 100 bp, about 30 bp to about 500 bp, about 30 bp to about 750 bp, about 30 bp to about 1,000 bp, about 50 bp to about 100 bp, about 50 bp to about 500 bp, about 50 bp to about 750 bp, about 50 bp to about 1,000 bp, about 100 bp to about 500 bp, about 100 bp to about 750 bp, about 100 bp to about 1,000 bp, about 500 bp to about 750 bp, about 500 bp to about 1,000 bp, or about 750 bp to about 1,000 bp. In some cases, the structural variant is about 30 bp, about 50 bp, about 100 bp, about 500 bp, about 750 bp, or about 1,000 bp. In some cases, the structural variant is at least about 30 bp, about 50 bp, about 100 bp, about 500 bp, or about 750 bp. In some cases, the structural variant is at most about 50 bp, about 100 bp, about 500 bp, about 750 bp, or about 1,000 bp. In some cases, the structural variant is about 1 Kbp to about 1,000 Kbp. In some cases, the structural variant is about 1 Kbp to about 50 Kbp, about 1 Kbp to about 100 Kbp, about 1 Kbp to about 250 Kbp, about 1 Kbp to about 500 Kbp, about 1 Kbp to about 750 Kbp, about 1 Kbp to about 1,000 Kbp, about 50 Kbp to about 100 Kbp, about 50 Kbp to about 250 Kbp, about 50 Kbp to about 500 Kbp, about 50 Kbp to about 750 Kbp, about 50 Kbp to about 1,000 Kbp, about 100 Kbp to about 250 Kbp, about 100 Kbp to about 500 Kbp, about 100 Kbp to about 750 Kbp, about 100 Kbp to about 1,000 Kbp, about 250 Kbp to about 500 Kbp, about 250 Kbp to about 750 Kbp, about 250 Kbp to about 1,000 Kbp, about 500 Kbp to about 750 Kbp, about 500 Kbp to about 1,000 Kbp, or about 750 Kbp to about 1,000 Kbp. In some cases, the structural variant is about 1 Kbp, about 50 Kbp, about 100 Kbp, about 250 Kbp, about 500 Kbp, about 750 Kbp, or about 1,000 Kbp. In some cases, the structural variant is at least about 1 Kbp, about 50 Kbp, about 100 Kbp, about 250 Kbp, about 500 Kbp, or about 750 Kbp. In some cases, the structural variant is at most about 50 Kbp, about 100 Kbp, about 250 Kbp, about 500 Kbp, about 750 Kbp, or about 1,000 Kbp. In some cases, the structural variant is about 1 Mbp to about 10 Mbp. In some cases, the structural variant is at least about 1 Mbp. In some cases, the structural variant is at most about 10 Mbp. In some cases, the structural variant is about 1 Mbp to about 2 Mbp, about 1 Mbp to about 3 Mbp, about 1 Mbp to about 4 Mbp, about 1 Mbp to about 5 Mbp, about 1 Mbp to about 6 Mbp, about 1 Mbp to about 7 Mbp, about 1 Mbp to about 8 Mbp, about 1 Mbp to about 9 Mbp, about 1 Mbp to about 10 Mbp, about 2 Mbp to about 3 Mbp, about 2 Mbp to about 4 Mbp, about 2 Mbp to about 5 Mbp, about 2 Mbp to about 6 Mbp, about 2 Mbp to about 7 Mbp, about 2 Mbp to about 8 Mbp, about 2 Mbp to about 9 Mbp, about 2 Mbp to about 10 Mbp, about 3 Mbp to about 4 Mbp, about 3 Mbp to about 5 Mbp, about 3 Mbp to about 6 Mbp, about 3 Mbp to about 7 Mbp, about 3 Mbp to about 8 Mbp, about 3 Mbp to about 9 Mbp, about 3 Mbp to about 10 Mbp, about 4 Mbp to about 5 Mbp, about 4 Mbp to about 6 Mbp, about 4 Mbp to about 7 Mbp, about 4 Mbp to about 8 Mbp, about 4 Mbp to about 9 Mbp, about 4 Mbp to about 10 Mbp, about 5 Mbp to about 6 Mbp,

about 5 Mbp to about 7 Mbp, about 5 Mbp to about 8 Mbp, about 5 Mbp to about 9 Mbp, about 5 Mbp to about 10 Mbp, about 6 Mbp to about 7 Mbp, about 6 Mbp to about 8 Mbp, about 6 Mbp to about 9 Mbp, about 6 Mbp to about 10 Mbp, about 7 Mbp to about 8 Mbp, about 7 Mbp to about 9 Mbp, about 7 Mbp to about 10 Mbp, about 8 Mbp to about 9 Mbp, about 8 Mbp to about 10 Mbp, or about 9 Mbp to about 10 Mbp. In some cases, the structural variant is about 1 Mbp, about 2 Mbp, about 3 Mbp, about 4 Mbp, about 5 Mbp, about 6 Mbp, about 7 Mbp, about 8 Mbp, about 9 Mbp, or about 10 Mbp.

[0056] As described elsewhere, the one or more target genomic variants may comprise one or more structural variants. In some cases, the one or more target genomic variants may comprise at least one structural variant. In some cases, the sample comprises about 1 target genomic variant to about 100 target genomic variants.

[0057] In some embodiments, the sample comprises RNA transcripts. In some embodiments, the sample comprises genomic DNA (gDNA). In some embodiments, the sample comprises gDNA and RNA transcripts.

[0058] In some embodiments, the sample comprises one or more target genomic variants. In some cases, the sample comprises about 1 target genomic variant to about 2 target genomic variants, about 1 target genomic variant to about 4 target genomic variants, about 1 target genomic variant to about 6 target genomic variants, about 1 target genomic variant to about 8 target genomic variants, about 1 target genomic variant to about 10 target genomic variants, about 1 target genomic variant to about 20 target genomic variants, about 1 target genomic variant to about 30 target genomic variants, about 1 target genomic variant to about 40 target genomic variants, about 1 target genomic variant to about 50 target genomic variants, about 1 target genomic variant to about 75 target genomic variants, about 1 target genomic variant to about 100 target genomic variants, about 2 target genomic variants to about 4 target genomic variants, about 2 target genomic variants to about 6 target genomic variants, about 2 target genomic variants to about 8 target genomic variants, about 2 target genomic variants to about 10 target genomic variants, about 2 target genomic variants to about 20 target genomic variants, about 2 target genomic variants to about 30 target genomic variants, about 2 target genomic variants to about 40 target genomic variants, about 2 target genomic variants to about 50 target genomic variants, about 2 target genomic variants to about 75 target genomic variants, about 2 target genomic variants to about 100 target genomic variants, about 4 target genomic variants to about 6 target genomic variants, about 4 target genomic variants to about 8 target genomic variants, about 4 target genomic variants to about 10 target genomic variants, about 4 target genomic variants to about 20 target genomic variants, about 4 target genomic variants to about

30 target genomic variants, about 4 target genomic variants to about 40 target genomic variants, about 4 target genomic variants to about 50 target genomic variants, about 4 target genomic variants to about 75 target genomic variants, about 4 target genomic variants to about 100 target genomic variants, about 6 target genomic variants to about 8 target genomic variants, about 6 target genomic variants to about 10 target genomic variants, about 6 target genomic variants to about 20 target genomic variants, about 6 target genomic variants to about 30 target genomic variants, about 6 target genomic variants to about 40 target genomic variants, about 6 target genomic variants to about 50 target genomic variants, about 6 target genomic variants to about 75 target genomic variants, about 6 target genomic variants to about 100 target genomic variants, about 8 target genomic variants to about 10 target genomic variants, about 8 target genomic variants to about 20 target genomic variants, about 8 target genomic variants to about 30 target genomic variants, about 8 target genomic variants to about 40 target genomic variants, about 8 target genomic variants to about 50 target genomic variants, about 8 target genomic variants to about 75 target genomic variants, about 8 target genomic variants to about 100 target genomic variants, about 10 target genomic variants to about 20 target genomic variants, about 10 target genomic variants to about 30 target genomic variants, about 10 target genomic variants to about 40 target genomic variants, about 10 target genomic variants to about 50 target genomic variants, about 10 target genomic variants to about 75 target genomic variants, about 10 target genomic variants to about 100 target genomic variants, about 20 target genomic variants to about 30 target genomic variants, about 20 target genomic variants to about 40 target genomic variants, about 20 target genomic variants to about 50 target genomic variants, about 20 target genomic variants to about 75 target genomic variants, about 20 target genomic variants to about 100 target genomic variants, about 30 target genomic variants to about 40 target genomic variants, about 30 target genomic variants to about 50 target genomic variants, about 30 target genomic variants to about 75 target genomic variants, about 30 target genomic variants to about 100 target genomic variants, about 40 target genomic variants to about 50 target genomic variants, about 40 target genomic variants to about 75 target genomic variants, about 40 target genomic variants to about 100 target genomic variants, about 50 target genomic variants to about 75 target genomic variants, about 50 target genomic variants to about 100 target genomic variants, or about 75 target genomic variants to about 100 target genomic variants. In some cases, the sample comprises about 1 target genomic variant, about 2 target genomic variants, about 4 target genomic variants, about 6 target genomic variants, about 8 target genomic variants, about 10 target genomic variants, about 20 target genomic variants, about 30 target genomic variants, about 40 target genomic variants, about 50 target genomic variants, about 75 target genomic

variants, or about 100 target genomic variants. In some cases, the sample comprises at least about 1 target genomic variant, about 2 target genomic variants, about 4 target genomic variants, about 6 target genomic variants, about 8 target genomic variants, about 10 target genomic variants, about 20 target genomic variants, about 30 target genomic variants, about 40 target genomic variants, about 50 target genomic variants, or about 75 target genomic variants. In some cases, the sample comprises at most about 2 target genomic variants, about 4 target genomic variants, about 6 target genomic variants, about 8 target genomic variants, about 10 target genomic variants, about 20 target genomic variants, about 30 target genomic variants, about 40 target genomic variants, about 50 target genomic variants, about 75 target genomic variants, or about 100 target genomic variants.

TARGET ENRICHMENT SAMPLE PREPARATION FOR SEQUENCING

[0059] In some aspect, the target enrichment sample preparation approach describe herein may comprise one or more genome editing technologies. In some cases, the genome editing technology is an endonuclease-based genome editing technology. In some cases, the endonuclease-based genome editing technology comprises zinc-finger nucleases (ZFNs), homing nucleases, transcription activator-like effector nucleases (TALENs), and/or clustered regularly interspersed short palindromic repeats (CRISPR)-Cas systems. In some cases, the target enrichment sample preparation approach may further comprise DNA amplification. In some cases, the target enrichment sample preparation approach may not comprise DNA amplification.

[0060] In some embodiments, the target enrichment sample preparation approach comprises preparing a sample for sequencing using a non-amplification-based, gene-editing based approach. In some case, the sample preparation comprises Cas-mediated PCR-free enrichment of said sample as shown in FIG. 2. Cas-mediated PCR-free enrichment of said sample may comprise extracting genomic DNA (gDNA) from said sample; dephosphorylating 5' ends of the DNA to reduce ligation of sequencing adapters to non-target strands; adding Cas9 ribonucleoproteins (RNPs) comprising bound crRNA and tracrRNA to the gDNA to bind and cleave the region of interest (ROI); cleaving of gDNA by Cas9 to reveal blunt ends with ligatable 5' phosphates; dA-tailing of gDNA in said sample to prepare blunt ends for sequencing adapter ligation; and ligating sequencing adapters to the Cas9 cut sides, wherein the Cas9 cut sides are 3'dA-tailed and 5' phosphorylated.

[0061] In some embodiments, a two RNP (ribonucleoprotein complex comprising Cas9-crRNA-tracrRNA) complexes, designed to excise a ROI, bind to sequences on the (+) and (-) strands, upstream and downstream of the ROI, respectively. The crRNAs confer specificity and 'program' the RNPs to bind to the specific sequences. Background DNA has been

dephosphorylated (i.e. carries 5'-hydroxyl groups). Upon RNP binding, the duplex DNA is locally melted. crRNA hybridizes to the non-target DNA strand, which is complementary to the crRNA. Cas9 cleaves both of the DNA strands within the target site, 3 bp upstream of the PAM. Cleavage by Cas9 reveals 5' phosphates at each end of the ROI. Existing ends of the same molecule, which carry 5' hydroxyl groups, are considered non-target. The PAM-distal side is protected from ligation by Cas9 and/or the bound crRNA, whereas the PAM-proximal side is released for each RNP targeting the ROI. Because the RNPs here target the (+) strand and the (-) strand upstream and downstream of the ROI, the ROI is excised and both ends of the ROI are freed for dA-tailing and adapter ligation. Adapter ligation to the ROI results in directionality of the expected reads.

[0062] In some cases, an alternative to Cas9 may be used in the CRISPR-Cas system, wherein the alternative to Cas9 may be Cas3, Cas4, Cas5, Cas8a, Cas8b, Cas8c, Cas10, Cas10d, Cas13a, Cas13b, Cas13c, Cse1, Cse2, Csy1, Csy2, Csy3, Csm2, Cmr5, Csx10, Csx10, Csf1, Csn2, Cpf1, C2c1, or C2c3.

[0063] In some cases, the target enrichment sample preparation comprises preparing a sample for sequencing using the PacBio® sequencing system. In such cases, genomic DNA (gDNA) is dephosphorylated and then subjected to Cas-mediated PCR-free enrichment as described herein. Following the cleavage reaction, SMRTbell® adapters are ligated to the blunt template ends, forming SMRTbell® templates. In the final step, unligated DNA is eliminated by exonuclease digestion and then prepared for sequencing by annealing to the Sequencing Primers and binding to the polymerase.

[0064] In some cases, the target enrichment sample preparation comprises preparing a sample for sequencing using Illumina® sequencing system. In such embodiments, gDNA is dephosphorylated and then filled in using biotinylated nucleotides. The gDNA is then subjected to Cas-mediated PCR-free enrichment as described herein. After the cleavage reaction, non-target gDNA is removed using streptavidin beads. The target gDNA is then fragmented to the appropriate size, end-repaired, and dA-tailed. Illumina® adapters are ligated to the end-repaired, dA-tailed target gDNA, and is then ready for sequencing.

crRNA PROBES

[0065] In one aspect, provided herein are systems and methods to design crRNAs for use with the systems and methods described herein. In some embodiments as shown in Fig. 3 and Fig. 6, preliminary crRNA probes are designed using available guide RNA (gRNA) tools. Exemplary gRNA design tools include CHOPCHOP program, based on ONT recommended design options,

and Broad Institute sgRNA Designer. In some cases, the preliminary crRNA probes are designed from Benchling probe design tool and/or CRISPOR probe design tool.

[0066] The preliminary crRNA probes are filtered using one or more approaches as shown in Fig. 3 and Fig. 6. One filter approach is to retain preliminary crRNA probes with a GC content between about 40% and about 80%. If no candidates are obtained, the lower limit of the range is lowered to a GC content between about 20% and about 80%. Another filter approach is to retain preliminary crRNA probes with a self-complementarity score of zero. If no candidates are obtained, the self-complementarity score is increased to 1. Another filter approach is to retain preliminary crRNA probes with an efficiency score greater than 0.3. If no candidates are obtained, the efficiency score is lowered to greater than 0.2. Another filter approach is to retain preliminary crRNA probes with the following mismatches: MM0=0, MM1=0, MM2=0, and MM3<5. If no candidates are obtained, the stringency of the mismatches is decreased in the following order: MM0=1, MM1=1, MM2 < 2 and MM3 < 10, until candidates are produced. In another embodiment, the stringency of the mismatches is decreased in the following order: MM0 < 1, MM1 < 2, MM2 < 2 and MM3 < 21, until candidates are produced. In some cases, candidates are further filtered by retaining candidates without any single nucleotide polymorphisms (SNPs). In some cases, ambiguous bases are introduced at any position to increase on-target performance.

[0067] In some cases, two or more of the approaches are used. In some cases, three or more of the approaches are used. In some cases, four approaches are used. In some cases, the following approaches are used in the following order: GC content, self-complementarity score, efficiency score and mismatches. After filtering the preliminary crRNA probes using one or more of the filter approaches, the on-target and off-target performance of candidate crRNA probes are confirmed using a guide RNA check tool. Examples of guide RNA check tools include IDT CRISPR-Cas9 gRNA checker, Cas-OFFinder, Dharmacon's CRISPR specificity analysis tool, Synthego's CRISPR specificity analysis tool, or a combination thereof.

[0068] Candidate crRNA probes obtained using the methods provided herein are more likely to cut the target genomic region of interest than crRNA probes obtained using other methods. In some cases, the probability that a candidate crRNA probe will cut a target is about 60 % to about 99.9 %. In some cases, the probability that a candidate crRNA probe will cut a target is at least about 60 %. In some cases, the probability that a candidate crRNA probe will cut a target is at most about 99.9 %. In some cases, the probability that a candidate crRNA probe will cut a target is about 60 % to about 65 %, about 60 % to about 70 %, about 60 % to about 75 %, about 60 % to about 80 %, about 60 % to about 85 %, about 60 % to about 90 %, about 60 % to about 95 %, about 60 % to about 99.9 %, or a combination thereof.

about 60 % to about 99.9 %, about 65 % to about 70 %, about 65 % to about 75 %, about 65 % to about 80 %, about 65 % to about 85 %, about 65 % to about 90 %, about 65 % to about 95 %, about 65 % to about 99.9 %, about 70 % to about 75 %, about 70 % to about 80 %, about 70 % to about 85 %, about 70 % to about 90 %, about 70 % to about 95 %, about 70 % to about 99.9 %, about 75 % to about 80 %, about 75 % to about 85 %, about 75 % to about 90 %, about 75 % to about 95 %, about 75 % to about 99.9 %, about 80 % to about 85 %, about 80 % to about 90 %, about 80 % to about 95 %, about 80 % to about 99.9 %, about 85 % to about 90 %, about 85 % to about 95 %, about 85 % to about 99.9 %, about 90 % to about 95 %, about 90 % to about 99.9 %, or about 95 % to about 99.9 %. In some cases, the probability that a candidate crRNA probe will cut a target is about 60 %, about 65 %, about 70 %, about 75 %, about 80 %, about 85 %, about 90 %, about 95 %, or about 99.9 %.

[0069] Provided herein are exemplary target genomic sequences (*e.g.*, a protospacer) to which crRNA probes may be hybridizable for use with the systems and methods described herein. A guide RNA can target a nucleic acid sequence of or of about 20 nucleotides. A target nucleic acid can be less than or less than about 20 nucleotides. A target nucleic acid can be at least or at least about 5, 10, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 30 or more nucleotides. A target nucleic acid can be at most or at most about 5, 10, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 30 or more nucleotides in length. A target nucleic acid sequence can be or can be about 20 bases immediately 5' of the first nucleotide of the PAM. A guide RNA can target the nucleic acid sequence. A guiding polynucleic acid, such as a guide RNA, can bind to a genomic sequence with at least or at least about 50%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, or up to about 100% sequence identity and/or sequence similarity to any of the sequences of the tables below.

[0070] Table 1: Target sequences for ABCD1 gene

SEQ ID NOS	Target sequence
1	TCAGCAACAACGTGACCCAGTGG
2	GTCATGACGAAGCAGAACCCTGG
3	TCATGACGAAGCAGAACCCTGGG
4	GTTCTGTTGCAAAACCCACAAGG
5	TTGGAGGCCATTAGTTAGTGCGG
6	GAGGCCATTAGTTAGTGCGGAGG
7	AGGCCATTAGTTAGTGCGGAGGG
8	CAGGTCTCCTGATTTACCTCGGG

9	CTTGCCCCATCTCGCATACCCGG
10	TGAGGGGTAAACCACCTGTGCCGG
11	CCAGAAACCCGAGGTAAATCAGG
12	AAGTGTTACAAAGGGTCTCCAGG
13	CGGGTATGCGAGATGGGGCAAGG
14	TCATGGGGCCCCTGCGCGCAGGG
15	CGCAGGGCCACATATGCTCAGGG
16	GCGCAGGGCCACATATGCTCAGG
17	TTACCCCTCACCGCTCGCAGCGG
18	CTGAGGTAAGCTAAAGACCACGG
19	TCCAGGTAGACAGCTGTTCAAGG
20	GGGGCACTAAAGTGTTACAAAGG
21	CGGGGTTTATGATCAAGCATGGG
22	ACGGGGTTTATGATCAAGCATGG

[0071] Table 2: Target sequences for ACADM gene

SEQ ID NOS	Target sequence
23	AAGGGGTTACAATAGGCATATGG
24	CGTGTTGTAATCATCATAGAAGG
25	TGTATGGAGGGATTGAACACAGG
26	GATTACAACACGTGACCTCAGGG
27	CTTAATCACATGGTCCTCGGGGG
28	GTTCAATCCCTCCATACAAGTGG
29	ACTTAATCACATGGTCCTCGGGG
30	TGATTACAACACGTGACCTCAGG
31	AACTTAATCACATGGTCCTCGGG
32	TAGAATGAGGCCCAGCAACCAGG
33	ACAGTCATTTATTGCTACTAGGG
34	GAGGGATGCCAAAATCTATCAGG
35	GATAGGACTCTAATCTCACAGGG

[0072] Table 3: Target sequences for ACADVL gene

SEQ ID NOS	Target sequence
36	GGGTCTTGCCAAACGGCCAG

37	GGGGTCTTGCCAAACGGCCA
38	AGCACACCCCGATTCTCAGG
39	CACACCCCGATTCTCAGGAG
40	GGGAGCACACCCCGATTCTC
41	GAGGCCCGCAAGTATGCCAG
42	TCCCGCACTAGGTCCTGCAC
43	GACGTCCACCCATGTGCTGC

[0073] Table 4: Target sequences for AFF2 gene

SEQ ID NOS	Target sequence
44	GTGATACCATGTATGCCACGTGG
45	GATGCTAAGGTACACCACGAGG
46	CTAACGAAAGACACCAACTG
47	CCTTCCCTAAGTGAACCAAGGGG
48	TCGTATCTAACACTCCCCTGGGG
49	CTAGTATTATCGATACCCAGAGG
50	GTAGGTTTCATACCACAATGTGG
51	GGTCTCTATCAAGTTCAAGGTGG
52	TCTGCCACTAAAGCAACCAGCGG
53	GTTCTCATGATCTCGCAGAGGGG
54	TGAGCACAACCTCAACTGGGGGG
55	GTCATAATCACAGTACATCGTGG
56	ACCATACACCAAATGCCACGTGG
57	GTGCTACTGCCACCTCACGGTGG
58	TTACGCCAGCACAAAACGTGGG
59	TGCCTGCGATAATTACAGAGTGG
60	GAACTGTAAATATAGATACGTGG
61	GAACTCATATGCAAACCTCGTGG
62	TCTAACTAAGGATCAGCACAGGG
63	CGACACATTGGATGAAACGTGGG
64	TATCAAAAATACCCACAGCGGGG
65	TGTTACCTGGAGTACTACGATGG

66	TAGATGTCCACCATACTCAGAGG
67	CAACTTGTTCACTATGACGAGGG
68	CAAGTTCCTCAGATCGCTGAGG
69	ACCACAGTCCCTAATTACCGTGG
70	AGTGGCTTGGTACAACAATGAGG
71	GCTGGAGTATATAATCCCCGGGG
72	ATATGTGATATACTACCCTGTGG
73	TGGGCGTAAGAACTAATTGAGG
74	GCTGGTGACGAGGTTAGACGTGG
75	CGTCTTCGCAGTAATTCTGGAGG
76	CCTTACAATGTATGTCCCAGTGG
77	GATAATGCTCATATGTGACAGGG
78	TGGGACCTTTGCAATACACAGGG
79	CTTTTAGTGATACTTCCACGTGG
80	ACCATACTGTCACAACAAGTGGG
81	GGCTATGTAGATACCTGTGGAGG
82	GATCACTAATCGCCCACCCAGGG
83	GAAAGTCCTTAAAGACCCCGTGG
84	GAGCATTGTTAGTGAAGTGGGGG
85	ACTGACATAAATGCCGTGGGTGG
86	GGTATTATAGTTCATCAGGGTGG
87	CTCTATTGTGACATGCAAGGTGG
88	TTTTCATCGAGTGTGCGTCGTGG
89	ATAGCGAAATCTATCTCACGAGG
90	GGCACTTGACATTTAACGTGGG
91	ATGAAAACTCAGCTTACGGTGG
92	GGTACCAAGATCTAAAATGGGGG
93	ATCGTTTATATAATACCCAGAGG
94	CCACCTGAAGAAATACCGGGGG
95	TATGTGATATACTACCCTGTGGG
96	GCTTGTTATAAATTACACCAAGG

97	GGATTCACCTCTGGTAAAGCAGGG
98	TGTGCGCAGATTCTCCGATGAGG
99	GGGAGTTACATACCCGTACAGGG
100	CTCAAGATACCCCATGACAGAGG
101	GATAAGAAGCAATCCACTGGGGG
102	TACGTAGAGAGTAAGTCCAGAGG
103	GAGACAACGTTACAAAAGCGCGG
104	ATATCTTATCTCCTACACCAAGG
105	GCTCCCCTGATGGTAAGACGTGG
106	CAGTTCCCAGCAAATAAACGAGG
107	TGACCACGTCTTACCATCAGGGG
108	ACGGGCAACTGAGGTAATGGGGG
109	TCTGAACACTAATGTCACAAGGG
110	TAAGTTTGACAGCTAACCAGTGG
111	AATAGTCGGTTTCTTCAATGTGG
112	GACACACTTAAATAAGCACGTGG
113	TAGGCACTTGTCAGAAACGAGG
114	CACGCAATGAGATAATTCTG
115	TCATAGGAAATGGCTCGCTGCGG
116	GGTATAATAGGCCAGTCTTGCGG
117	GGTGCTTATCCACTATTCAGGGG
118	ATAACTCAGTAGATCAACCTGGG
119	GTCCTTGAGAAACCCTTCGGAGG
120	ATGCTCTATTGTGACATGCAAGG
121	CCGTTAGGCTTCATAAACCATGG
122	ATGTTTCTGATGAAGTGCCGTGG
123	CAGGGTATTATAGTTCATCAGGG
124	TATCTCAAGAAGGTAACCTCGGGG
125	AGGTTCTCATGATCTCGCAGAGG
126	GGAATGAACACCATAACCGAAGG
127	TCTAACCTAAATGGTCTGTGTGG

128	ACCTGTCCTATGGAGTATGGTGG
129	TCACACCCTTGTTAGACCAGTGG
130	CATAATCCTCTACTACGATG
131	CTTCTCTATAAATCTCAGGGAGG
132	TTACCAAAAAGGCTTATCCGTGG
133	AAATGCGGGGATAATTCCAAAGG
134	GGAGTTATAACAATTGTCCG
135	GAGTTGTCAAACCATTATCGTGG
136	TAGTGCCATCGCTTTAAGGGTGG
137	GCTAAAACACTGGTAACTCAGGG
138	GATGTGCTACTTCTACCTGAAGG
139	TATGAATAACAATGGTACGAAGG
140	ATTCATAGAGTGGTTGTCAGGGG
141	ATGATGATAGAACCTTAAGCCGG
142	GCTTGACATTAGATAGACCATGG
143	TCTGGAAGTTGGAAGTCCGTAGG

[0074] Table 5: Target sequences for ALPL gene

SEQ ID NOS	Target sequence
144	TGATACCATCTTAAGTCTCCTGG
145	CACTTAGGTGATTAAGGGCTTGG
146	CAACATGCACGTACTAGGCATGG
147	CTGCCCAAGGCTTAGCTAGGTGG
148	AGGCCACCTAGCTAAGCCTTGGG
149	AATTTCCCCATTGTGCGTCTTGG
150	TTACAACCCTTTGACCACCAGG
151	TAGGTCCCTCTGCTAAACAGGGG
152	GTACCTGCAAGTCCTGTCACAGG
153	GCATGGAACCTGGTGGTCAAAGG

[0075] Table 6: Target sequences for APC gene

SEQ ID NOS	Target sequence
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154	TATATCAGGCATTGTAACAC
155	TGTGTATGGCCCCACAAAGA
156	ATCTTTGTGGGGCCATACAC
157	TCGTTATAACACCAGTTCTG
158	CGTTATAACACCAGTTCTGT
159	AGATCTAGTTAGTTCTACAA
160	GTACAATCCAATGACATCTG
161	AGATGTCATTGGATTGTACC
162	CCAGTCATGTTTGATATACT
163	GTCTCCTGCACTACAAGACT
164	AAGTTCACAATACTGGTT
165	CTCTTGGAGGTTGTAACTC
166	CCTAGTATATCAAACATGAC
167	ATTAGGGTTTAGTGTACTAA
168	TTAGTCCTCTACCTTACTGG
169	GCCTATTTTGTGATTGCCAA
170	CATGAGAATAACGACCTCAA
171	GGTGACCCCCAGTAAGGTAG
172	CGACCTCAAAGGATATCATG
173	TAAGTGTCATCAACTAGGG
174	CCATGCAGTTAAGAGGTACC
175	AACCTTTGCTTACATGCCTA
176	GGTGGTACTTACCCTTCAT
177	TTGTTAATAGAGCTTACTAC
178	TGTCAACTATATTACCCTAT
179	CATCCAGGCTTATAATCTCC
180	TTTGCATGGATGCACCATAT
181	TGCACCATATAGGTTCCATG
182	TTTACGATCAATGTCCATTT
183	TTGGCCTCATGGAACCTATA
184	GTAGATTAGTTAAGTGGCTC

185	AATGAGGGGGATTAGCCACA
186	CAGGTAGTATATTAGTCACC
187	CTGGTGACTAATATACTACC
188	AAGACATCCAGACTGTCGCA
189	AATGCTTGGTACTCATGATA
190	TCTAAACTCATTTGGCCAC
191	GAATGCGTATCTAACAAGGG
192	ATGCGTATCTAACAAGGGTG
193	AATGCGTATCTAACAAGGGT
194	CTTAACCTAGACCTGGGATT
195	TTGTAGATACGCATTCATC
196	CTCATTTGTAGCTATCAAGC
197	ATGGGTCATCTAATTAGAGT
198	TAGCTACAAATGAGGACCAC
199	ACCAGTGAGGGACGGGCAAT
200	TGGTTGGCACTCTTACTTAC
201	TGGTACAAATAGCCAAGGTC
202	CTCTAGGTCAGATACAACCTC

[0076] Table 7: Target sequences for ASPA gene

SEQ ID NOS	Target sequence
203	CTATGTAAGTTCACATGATGTGG
204	AACCTGGCGTTACTAGTACATGG
205	CACTAACTACAGTTCTGAGTAGG
206	TTGGATCTGCCTTCTCAACCAGG
207	AAATTCTGAGTCCGTAATCCAGG
208	TTAGCTAAGTGACAGGTCTCAGG
209	ACTAAGTTCGCAGTCTCACATGG

[0077] Table 8: Target sequences for ATM gene

SEQ ID NOS	Target sequence
210	AATTGCGAGGACAACCTGTCT

211	GAATTGCGAGGACAACTGTC
212	GATCACAACCTGGGTAAGGGT
213	ATCACAACCTGGGTAAGGGTA
214	CAGGTCCAATCTTCCTATGA
215	CTTCATAGGAAGATTGGACC
216	GATTCTGTGAGATTGAATCG
217	GTAAACTGTCAGGTCACTT
218	CATCGTCAAGGAGTTGACAG
219	GCCATGATGAGTTGGTCCAA
220	AAAGGCTAGTATAAGCCCAA
221	ATGCATAAGTAGCTCCTAGA
222	AGTGATACTCTAGGGCAAAC
223	GATGCATAAGTAGCTCCTAG
224	GGGCAATACTCTCTTGGTAT
225	GCCTTTGGACCAACTCATCA
226	CCCTAGAGTATCACTTGTTA
227	GGAACCTTATTGGCTGGAAC
228	TAGTTAGGAACCTTATTGGC
229	ACCGAATTCACCTCTTGAA
230	AGGAGTGAATTCGGTAGCCA
231	GTTCTAATTAGGGACTCACC
232	GACCCAACCTTGCTACTCGCT
233	TTCAGGTTGAGTGGATAGTC
234	GCTCTACCTCCACATACACT
235	GGCTCTACCTCCACATACAC
236	CGAGTAGCAAGTTGGGTCCT
237	TGGCCTAGCGAGTAGCAAGT
238	GCGTAACACCCACATATTTA
239	TCCCATAGGCATAACCTAA
240	GGACTCAACTAATTGGTGTT
241	ATATGTGGGTGTTACGCAAA

242	CCAAATCCCTAACAGAGTTA
243	CCTTA ACTCTGTTAGGGATT
244	GCTTCAAGCTGACTTTAACC
245	CAGGTGATTTCTCCATCCCG
246	TAGATTTAGTGACCACGGGA
247	CACGGGATGGAGAAATCACC
248	TCAGCTTGAAGCTCTCGTGA
249	GTGTTTAGATTTAGTGACCA
250	CTATAATCTAGTAGGATCAC
251	ATCTAGTAGGATCACAGGAT
252	CTGTGATCCTACTAGATTAT
253	CATGCTTGAAGGCTCATTAT
254	ACAAGTGGACAAGTCAGATC
255	GCCAAGCTGTTTCTATCCAA
256	TACACGATTCTGACATCAA
257	GCTACTTATGTGTAGAGCAC
258	CTTTGCAGTTACCATAGGAG
259	TAGAGTATCTAACCCAACGT
260	GGGCATGTAGAATACTTATT
261	GTGAATTTATATACCTACGT
262	ATCTTAATGAACCACTCATA
263	AGACAGTCACGGATATTATA
264	TGGAGTACAACCCATATGAG
265	AAAGATGCCTCGGTTTCATAA
266	CGGTTTCATAAAGGTGCACAC
267	GCCCCACCCTTATTGACCAC
268	AAGATGAGTAACAGTCCATC
269	CATTAAGCCTGTGGTCAATA
270	GGCATTAAACCATTAAAGCCTG
271	AAGCCTGTGGTCAATAAGGG
272	AATAGGTCCCAATAATACGT

273	GTGTGCACCTTTATGAACCG
274	GGAGGTTGGTTGCACACCAC
275	ACTCACCATTAGTAGTATAC

[0078] Table 9: Target sequences for ATM gene

SEQ ID NOS	Target sequence
276	AATGGGATCCCTTCCTAAGG
277	GTACCAAGACGTGGATATGG
278	GGTACCAAGACGTGGATATG
279	GATGTGTAGGTACCAAGACG
280	CTGGAACCTATGATCAGGCA
281	TAGGTACCAAGACGTGGATA
282	AGGTACCAAGACGTGGATAT
283	TGTCTCTGGAACCTATGATC
284	TGTCACAAGAGGTGCTTACA
285	ATGACTCTGAACTGCCCACC

[0079] Table 10: Target sequences for ATXN1 gene

SEQ ID NOS	Target sequence
286	GGCAACTCAGATACTCACGTGGG
287	CCCCAATAGAGATTGCCCTGTGG
288	ACATCAGAACATGAGCACCGGGG
289	CAGGTGAGCGTACTGCACGGGGG
290	CGGGTCAAACCCCATCACAGTGG
291	TAAGTTGTCGTTGATCACAGGGG
292	GAAACAGGTATGATGCATGGGGG
293	GTCCACTTTATAAATCCCAGAGG
294	CTAAAACTTCTCATGCAAGGCGG
295	TTGCGATCAGAAACACAAGGAGG
296	TATAAGTGTTAAGGGCACCGGGG
297	AAGGTTACTCGGGTTCACAGAGG

298	CGAGACCTGACCATACTGTGGGG
299	GTAGTTCGAACACCCAACCAGGG
300	CAGAGTTTCGTACAGCAGCGTGG
301	TTGTAAACCAAGCTCCACCGAGG
302	TGTCACTTTAGACCAACCCGAGG
303	CCTGATCCAGTAAGTCACGGAGG
304	TTTCTGATGAGAGATCGCGGGGG
305	GTGTTTATGAACTCGCCAGGAGG
306	GTAGAAGATAGAATTCATTGGGG
307	AGTCTCAGCACATGACAACGTGG
308	GGTATACGTTCCAACCTCAGAGG
309	TGTATCACTACAGTTAAACGGGG
310	TGTCGGTAAATATTGCAAAGTGG
311	CAACCCACATATCAAACCGTGG
312	GGACTGGTGGAACAACCGGGAGG
313	TGATCGCTGTAAGACCAAAGAGG
314	ACCACGTTGCAATATCTGGGAGG
315	GCAATGTGATTCTACACCCGGGG
316	AATGATTTGTCACTTTACCGAGG
317	ACATATACCTTACCCAGCGAGG
318	TGTAGTAGAGCACACCAAGGGGG
319	TGGCCGGTTCCTATTCCATGGGG
320	GTGAACGCACCTGATCCATGAGG
321	GATCAATTCCAGGAGTTACGGGG
322	TGGTACTCTTGAGGTAAACGGGG
323	GGGCGATGAGGTAATTTGAGCGG
324	TCCAGAGATAAACTCCTCGGGGG
325	ACTAAGATTCATCTACCACGTGG
326	CATCTGGGTAGAGTACGTGGTGG
327	GCTCATTGTATCAACCAGTGTGG
328	GTACACTTTAAGATGCCACATGG

329	CTGTGCGATTGCCCACAAGGAGG
330	TTAGAAAGCACGTCCCAACGTGG
331	AGGCTATTATCTCATAACCGGGG
332	GTATCACTACAGTTAAACGGGGG
333	TTGACCGCCAAAACCAACCAGGG
334	GCTCATCGTAACTAAACCAGTGG
335	GCCGCAAACCAAGACATGTGAGG
336	TCCACATTCACTATTCCGTGTGG
337	ATCCGTAATAGATTGCTGAGAGG
338	GCGCAGCACTGGAACCACGTAGG
339	ATTAAAGTAGACCCCCCAAGGG
340	ACGTCCTCTGATGAAAAGGGCGG
341	ACCTCCCTCTTGACAAACGGGGG
342	GCATCCAGATGCGACCCCCGAGG
343	TACACACAGCAGAGATCACGGGG
344	TATATCCAGGAGTTTGTAGGGGG
345	GTGACATTGTGATACCCCAAAGG
346	ATCTCCGGGGTATAAGACATAGG
347	GAAACTCACAAATGGCTACGAGG
348	ACTATTCCGTGTGGTGACAGGGG
349	CTTCACTCTAATGAGATACGTGG
350	CGGTGCATAGACACTTAAGGTGG
351	GAGCGTGACAGGAACCCCAAGGG
352	AGTACATGCATCATCCCAAGAGG
353	GTGCACAGGTCGTCTCTCAGGGG
354	AAGTTCTACAATGACACAGGTGG
355	ATACGTGGAACAAATTACTGGGG
356	ATTGAGGAGACACCTACCAAGTGG
357	TCTGTGACCCTAATCTACGTGGG
358	TTAGGAAGTTCGATCCAACAGGG
359	TCAGGAGGAGAATGGTCGCGGGG

360	GTTTAAGATGATGTAATCGAGGG
361	CAGGATACATAACCCACAGGAGG
362	CTGTATACCACGAGACATGGAGG
363	GATACTTTTGGTAAACAACGTGG
364	ACTAGTTAGGATAGATGACAAGG
365	GAACACCGTAATGATGGCAGTGG
366	GCGTATCAGCACCACCCCGTGG
367	GGTTCTAAGCTCAACTCCAGAGG
368	CGTCAAGTATGAAACCGGTGGGG
369	GCCACGACCAGATATCAGCTGGG
370	ATGATGGCACCCGAAGACAGTGG
371	GGGTGAAACAATTCCTTACGGGG
372	TCGCAACTTCAGCATAACAACGG
373	CATAGTATGTCAAACCTCACGAGG
374	TCATCATCTTTTTGTCAACGGGG
375	CTAAGATTCATCTACCACGTGGG
376	GAATGGCGGTGATAAGCTAGAGG
377	ATTCGTTTGAGGGTGTGGGAGG
378	AACCTACTTCCTACCCAAGGAGG
379	TCTAAGACGTTGCAGCAGTGGGG
380	GCTACTACCATATGCCCCGAGGGG
381	GAAACTCTATGATACCCCAAAGG
382	CAGGTCATATAACAACATCCGTGG
383	GCTATTTGAAAATCCCACGTAGG
384	AAGTGACGTTTTCCTCCTGGAGG
385	GTAATTCCCAAGACGCATGGTGG

[0080] Table 11: Target sequences for ATXN2 gene

SEQ ID NOS	Target sequence
386	GGTAACTCCACAATTCTACGAGG
387	TATGTGGTTCTGTACTTACGTGG

388	ACGATGATCTGGTATCCTGGTGG
389	GATACGCACAAACCTAAGTGAGG
390	ACCGTGGGTAAAGTCCTCTGGGG
391	TACCCCTTTAACCGGCACGGGGG
392	GTCCAAGATAATGACCTGAGAGG
393	AGGCCAGGGATCTATCATCAGGG
394	TGATGACCACGTTCCCCCGAGG
395	TGTATCCATCTTCACGAGGGTGG
396	AAGTTGTATAGGCACTGACTTGG
397	GAACTTGGTACAGAGACGCTGGG
398	GTAAGTATGAGGATCTCGAATGG
399	TGACGATCAGTTCAGCATCAGGG
400	CAGATCACGTGTATTTGAGAGGG
401	TGTTAAATAGTGCGCCAGTGAGG
402	GATGACCACGTTCCCCCGAGGG
403	GTGCCGCCAGAGCTTACCAAGGG
404	CCAGATCACGTGTATTTGAGAGG
405	GGAATCATCAGGGTCTGTCGGGG
406	ATTGTTTTGAGATCGTGCCCAGG
407	TGCCCAGTACAAAGCTCGAGTGG
408	AGTTTAGGCCCAAAGCTCCACGG
409	GGGACTGAATATGCGTGCAAAGG
410	TATAGTTACTTATCAACTGGAGG
411	ATTGCGTGGAGTAAGCTGGTGGG
412	GTAGTGTTGTACGATATCATGGG
413	AATCAAACAGCGTGTAACAGAGG
414	ACGGGTAGACATAATAGTTGGGG
415	AATGTGCGAACTTTAGACCTTGG
416	ATATTCAACGATTCCAAGGTCGG
417	AACCCCTCCCAACACGCGTGGGG
418	TTTAGGTGTGAACGTTGGAGGGG

419	CGTCTGTGGAAACCCCGAGTCGG
420	CATATGTTTTAGTGGTATCGGGG
421	TTGCGTGGAGTAAGCTGGTGGGG
422	TCACAGCTCATATGACGTAAAGG
423	GTAAGGTAGATTCTTCACGTTGG
424	CGTGGAGTAAGCTGGTGGGGTGG
425	TATCGCATCGTCAGAACACATGG
426	TCTGACCCAGAATTTGACGATGG
427	AACTTGCGAGTATATTAACAAGG
428	ACCGTGAGTTTATTCTCCAGGG
429	AGACAGAATTCACCGCGTATGGG
430	TCACAGGATTATATGTACCCTGG
431	AAGCATCCTAAGTGGTGTGTGGG
432	TAAATACCGGTAAACTTGCAAGG
433	CCACGTAAATGGTGTGCAGAGGG
434	TGGGGTGGGTTGGTATACGCCGG
435	GATATGACGTCTTCATGCCAGGG
436	GAACCCCTCCCAACACGCGTGGG
437	ATCTCGAGTGATTGAATCTGAGG
438	AGACGCATGCGATGTATGGTAGG
439	ACAGGCCCCGGTAGTCACTTCGG
440	TACTGAACCGCAATAAACAAAGG
441	ACATCATGTGCGTAACATTGTGG
442	CTAAGTATATCATATTGACCAGG
443	TAGGGTCTTAAGCACCAAAAGG
444	AGAACCCCTCCCAACACGCGTGG
445	CGTAATAAAAAGTTACCGCAAGG
446	AGTGCCGCCAGAGCTTACCAAGG
447	GTCGGCTCTGTCTCTACCGAGGG
448	GCAAGAGTAAACTTCCATAGAGG
449	TGTGGAACATCGGTGGGTGAGGG

450	AGGCCTGTCAAACCTTCGTAAAGG
451	GCAATGGAGCAGGTCGTCAATGG
452	TTTAGGGGAAGTTGTGCTAAGG
453	GGATCTATCATCAGGGCCGAAGG
454	GCGTCTGAACCAAAGATGTACGG
455	GTTACGCACATGATGTATAGAGG
456	CTACCCCTTTAACCGGCACGGGG
457	TATTGCGTGGAGTAAGCTGGTGG
458	GTCGGCAATATAAGTGAACGTGG
459	CTAACCTATAACCTCAGCATAGG
460	ATCCGGTCATATAATCATCTAGG
461	AGTGATCGTTTCCCAAGTAGGG
462	GTAGGCGCTCCAGTGGCTCGGGG
463	GTTGATGACCCACCATAGATGGG
464	GTTAGGGGGATGGCCGATGTTGG
465	TATACAACCGTTCCTCTCAAAGG
466	ACGATTAACCTCTAACTGCCAGG
467	GAAAACCTAACAACCAAGCTTGG
468	TGATGGTGCTGCAAAGCGACAGG
469	G TTCACACCTAAACCGGGAGTGG
470	GATGAAACTGTTCCACCGGCCGG
471	TCGAGTCAAACCCAGTTAGCCGG
472	GATAACCTATAGTCAGGGCATGG
473	TACGCCGGCTGAACGTGAGAAGG
474	GACACGCAAAGTCAGCTACATGG
475	GTGATTTGAGGATGTCGCTGGG
476	ATACCACCTGTGTAACTGCAGG
477	GTGTTCAAGTAACACGTTGCAAGG
478	GTCCTATTCTCATTAACTACGG
479	TGCTTCACTACTTGATCTGAGGG
480	TGGTATGCCCCCTATGGATCAAGG

481	CTGTAGTGCACTTTGAGCGAGGG
482	ATACACGTGATCTGGCCCTAAGG
483	TACTTATTGACCTACTAAGCTGG
484	CGTTTAGGCATAGTAGAGACAGG
485	CAACAGGTAGGGGTCATAGAAGG

[0081] Table 12: Target sequences for BRCA1 gene

SEQ ID NOS	Target sequence
486	TCAGGTAGCACTCTTAACCTGGG
487	CCTGTCACCTGTCTATGGGTCGG
488	ACATAGACCCCTCTGTTGATGGG
489	GTAGTCAGACTAGTTCAATGAGG
490	GTAGTTGACCTGCACTCTACAGG
491	GTTTGATGTTTATCCAGACTTGG
492	TACTCCACTATGTAAGACAAAGG
493	GCTTTAACTTGTTAGATGCAAGG
494	AGTGCTAGATACTTTCACACAGG
495	TGTAATTTGGATTCCCGTCTCGG
496	ACGTCATATTTAAGGCATTCAGG
497	GCTAAGATCTGAACCCGAGACGG
498	GTATCTGAAGAACCGTTACCCGG
499	TTCCAAATATCCATACCTGCTGG
500	GTATCTTTACCATCTACCTCTGG
501	CTTTCAGGCAATCACTCCATTGG
502	GTCAACCCTGACATATTGGCAGG
503	ATATGTCAACCCTGACATATTGG
504	GCTGCAGATTAGACTACAAGTGG
505	CCATCGCCACCAATTGTGAAAGG
506	GTCAGGGTTGACATATAACATGG
507	TTATGCTAAGTAACTACCTATGG
508	CACGTAGAGGTTAGATGTGATGG

509	AACTACTCACGAGTACCACGTGG
510	TTTATGTAATGGCCACGTAGAGG
511	GTATTTGGCCACTTACCTGCTGG
512	TACTCACGAGTACCACGTGGTGG
513	TATTTGGCCACTTACCTGCTGGG
514	GGTTGCCAAAAAGTCCAGTGGGG
515	CATCACATCTAACCTCTACGTGG
516	TGGTTGCCAAAAAGTCCAGTGGG
517	GGTAAGTGGCCAAATACATTAGG
518	GTTGCCAAAAAGTCCAGTGGGGG
519	GCAATGCCATTGCCACCACGTGG
520	ATTACTGGTGGACTTACTTCTGG
521	AAGTAAGTCCACCAGTAATTAGG
522	GCTTTGCTACAATCCAATTCTGG
523	GGTACTTGAAGCATCTATATCGG
524	TAGAAAGTAGCCCAGCGCAATGG
525	AAGGTACTTATGTACAGTGGAGG
526	GGTTATATTGGATCCAGAATTGG
527	GTAGAGTGGTTAGCCCAAGGTGG
528	ATGTTGGTACAAGTTATCTCAGG
529	GAGATAACTTGTACCAACATTGG
530	AACTACGAGTGCGCAGACATGGG
531	CTCGGTCCCTCAGAACACGAAGG
532	AAAACGACCACCCCATTGACTGG

[0082] Table 13: Target sequences for BRCA2 gene

SEQ ID NOS	Target sequence
533	CACGTAAGCACTCTCCCACC
534	GTACTTTACCATCATGCAAG
535	AGGCCCCTGATTTACTCT
536	GTCAGTGGTTAAACTAAGG

537	TCACTGGTTAAACTAAGGT
538	GTCACAAATTTGTCTGTCAC
539	CCTTAGTACTACTCACAAGG
540	CTTCCTTAGTACTACTCACA
541	CCACCTTGTGAGTAGTACTA
542	GGTGTCTCTCTGTAATACAT
543	CGGTGTCTCTCTGTAATACA
544	GCAAATAACACCTCCCATGA
545	CTCTCAAAGATGGCACGTAC
546	ATATACTACCCATTAATGGC
547	AATCCAGTTCATTAAACCCC
548	ACACTTTGGGTAGATATCC
549	AACTCCCCTCATCTACTTG
550	ACTTCCCCTCATCTACTTGA
551	GGGACCCTCAAGTAGATGAG
552	TTGGGACCCTCAAGTAGATG
553	TGGGACCCTCAAGTAGATGA
554	AGGATTATCAAGTACACTCC
555	GATGTGCCAGACGAGTGTGG
556	TTGATAATCCTCTACCCTAA
557	CTTGATAATCCTCTACCCTA
558	TAATCCTCCACCCACACATA
559	TAGAGCCTGCCCATATGTGT
560	CAGCAAGCTGTCATATGATC
561	TCTTGGAACAGACGTGAGGT
562	TAATTCTTGGAACAGACGTG
563	AATTAGGCACCCCAGGATAT
564	TGCAAGAAATTAGGCACCCC

[0083] Table 14: Target sequences for C9orf72 gene

SEQ ID NOS	Target sequence
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565	ACACTCCGATGATTATCCACTGG
566	TCTAACTCATCGGGGTCAAGTGG
567	TAGACAAGATCCCTATCCCATGG
568	TACTCTCAACTAAAAGTTAGAGG
569	CAGTAACAGCAAGGTGAGTCAGG
570	ACATGCAATGAGGTAGTGACTGG
571	TGTTACTGACGTGGTATCACCGG
572	GTTACTGACGTGGTATCACCGGG
573	CGTGGTATCACCGGGAATCATGG
574	GAATGTCCGCGCTCCACAGATGG

[0084] Table 15: Target sequences for CATSPER2 gene

SEQ ID NOS	Target sequence
575	GAGACTTCCGGTCCAGAAAC
576	ATTGCCAGGTGAGCTTGACT
577	CATAACTCTCATGTCAGATG
578	TGTCAGATGTGGGCCAAACT
579	GGGATGTCTAGTCTGTAGAC

[0085] Table 16: Target sequences for CDH1 gene

SEQ ID NOS	Target sequence
580	TAGGTTTGGGTGAACTCTAA
581	CCCATTTACAATCAACCTTA
582	GTCCAATCTGCCGTAACTC
583	TCCAATCTGCCGTAACTCA
584	GCCTGGTGCTAACACACAAC
585	GGACGGAACATACATGCCAA
586	GACGGAACATACATGCCAAT
587	ACATGAATAAATCGCTATCT
588	CCCTTAAGGTTGATTGTAAA
589	ACCCCAGGTACATGAGTCAA

[0086] Table 17: Target sequences for CDK4 gene

SEQ ID NOS	Target sequence
590	AATCTCTAGGGTACTTCCGG
591	CTAACCTTTGGGAGTGCCTA
592	CAAGTCCTCTTGTATGGCCT
593	GATTAAGGGGGTTTGTCTGA
594	TCTAACCTTTGGGAGTGCCT
595	GGGAATGAACTGAAGGCCGT
596	AAGAGCTGTGCAAGTGTCGG
597	TGCAAGTGTCGGAGGTGTGA
598	AAGTTGACTAGGTGTGTGTC
599	GTGCAAGTGTCGGAGGTGTG
600	TAAATGACGCAGGTGTACCA
601	GGTGAGTGGTTAAATGACGC

[0087] Table 18: Target sequences for CDKN2A gene

SEQ ID NOS	Target sequence
602	CAGTTAGGAAGGTTGTATCGCGG
603	CAAGATATACTGGGTCTACAAGG
604	GCTAATTGAGAGGTACCCGAGG
605	GGTGATTTGATTCTCGGTGGGG
606	AGGTGATTTGATTCTCGGTGGG
607	CAGGTGATTTGATTCTCGGTGG
608	GTACAGGTGATTTGATTCTCGG
609	AGCCGTTTTACACGCAGGAGGGG
610	CAGCCGTTTTACACGCAGGAGGG
611	GGCTTAACACAGCTGTACCTGGG

[0088] Table 19: Target sequences for CDKN2B gene

SEQ ID NOS	Target sequence
612	ATCTGAGTTATGTGCAACATTGG
613	TATCAGACGCTGCTTGTGAGGGG

614	TGCGGCAATTGACAGCATAGGGG
615	TTGCGGCAATTGACAGCATAGGG
616	CTAGTAAGCGCGAATGCCCCGG
617	GCTCAAATAAAGCGCCGCCGGG
618	CTCAAATAAAGCGCCGCCGGGG
619	TCGCTTCATGGTGAGTGTCGAGG
620	CGCTTCATGGTGAGTGTCGAGGG
621	GGCTTTCCGCCGCTCCCCGTTGG

[0089] Table 20: Target sequences for CTFR gene

SEQ ID NOS	Target sequence
622	GTATGCTTTTGGCCACGGAA
623	GACCCTTGCCTTAGATGTGT
624	TTGCCGACACATCTAAGGCA
625	CTTTATTGCCGACACATCTA
626	GTGTCGGCAATAAAGTAATC
627	TGCCGACACATCTAAGGCAA
628	CACAATAAGGCCAAACAAGT
629	GGTCACACTATGCCACAATA
630	ATGTAGAGTGCCCACTTGTT
631	TCCAAGCACCTAGACTGTA
632	GATAGAATAGAGCACACCAT
633	AAAGTGATGGCACACCCACC
634	TGGGATAGTATGCACCAGGT
635	ATACTGGGATAGTATGCACC
636	CTGAAGACCTTGCATGATCA
637	AAACACGCTTTCCCCTCAA
638	GGATAATTAATACGCCATGA
639	GGAAGTAGCAGCACCTTTGA
640	CAACTCCTACTGATAACCAA
641	ATTGGTGAGTAAAGGATCCT

642	TATTGGTGAGTAAAGGATCC
643	CATGGAGCTGTTACCATTCA
644	GACTATGTCCTCTTCGGTTG
645	CAGTACTCTATTGTCCCTAG
646	CCATTGTAGGCCAATAAGTG
647	GGAGGGTTGTCCAACCACTA
648	AATCACGATCTCTAAACTGG
649	ATCACGATCTCTAAACTGGA
650	CGGGTGTAGAGATCAAATAA
651	GTAGAGATCAAATAAGGGGC
652	TGGAGGGTTGTCCAACCACT
653	TGCCTTAATCCAACATTGGA
654	AATGTGCCTTAATCCAACAT
655	TTGGATTAAGGCACATTAGT
656	TGCCAGGTAAAGTTGTTCTT
657	GCCAGGTAAAGTTGTTCTTA
658	ACCCTAAGAACAACCTAACC
659	TGTATTAGCAAGTGGACTCC
660	GGGTCAATTGTATTAGCAAG
661	ACACTAACACCTACCCTACC
662	AGATCCTGAACTGTCTAGCC
663	CAGCCTACAAGTTCTTTGAC
664	CTGAGCTAGAGGTACCCTTA
665	GTAATTTAGATCTTAGGACC
666	AGACTTACTTACCAGGGAGC
667	CATGTACATTGGACCCTAAC
668	GGACCCTAACAGGAGTTCCA
669	TGTCTTAGATGATTCTAGTC
670	GAGTTTGGGGGCACACGAAA
671	GCTATTACTAAAGGTTTCTC
672	ATGGCCTTCAAAGTTGGCTC

673	CCCTTGAATAAGAGATATCC
674	ATGGCCTACACGACCCTACA
675	GGTCGTGTAGGCCATCTTAA
676	GTAGACAGCACGATGATTTC

[0090] Table 21: Target sequences for CHEK2 gene

SEQ ID NOS	Target sequence
677	AACGCACTGAGCTGTGTAGGAGG
678	TATTACTTGCAAGCTGAAACAGG
679	GTCATATGGGGAACCTCTGTTGG
680	ACCTCGACGTGTCTCTCGCCCGG
681	TGTTGACACAATACTTCAGCAGG
682	TTGGCAAATCGTATCTATGCAGG
683	CAACGTATGTATGTAGTAGTGGG
684	TCAACGTATGTATGTAGTAGTGG
685	GCAGATGTTCTAAGCTCTTGTGG
686	AAGGTGACCCTTATTAAAGTAGG

[0091] Table 22: Target sequences for CLN3 gene

SEQ ID NOS	Target sequence
687	GGGCATCGATTAGGGGTACGAGG
688	GCCAGAAGGGGCATCGATTAGGG
689	TGGGCGCCCCCATCAGCTCAGG
690	CTTTCTCGTGCGGTTTCCCAGG
691	GCTGTGAGGAGCTTCTCGAGAGG
692	AGTCCGACGAAAAGAGGGCCGGG
693	GAGTCCGACGAAAAGAGGGCCGG
694	GCATCATGCCAGGGTGCGCGAGG
695	GTTATCCCCGCCAGTTCTGAGG
696	CTCCGCTTCTCTTCGGGTAAGGG

[0092] Table 23: Target sequences for CLN6 gene

SEQ ID NOS	Target sequence
697	CATGCACTCCTCAACTGTCGTGG
698	ACCCTTGAGATACGATCTACTGG
699	TTTGGTACGACCTGGATGAAGGG
700	TTTTGGTACGACCTGGATGAAGG
701	AGGAAGTTTTTGGTACGACCTGG
702	GAGTGAGCGGTCATCTTGGAGGG
703	GGTACACACACCTCGTCACTCGG
704	TTCCCGCGTTCCAGCGACCCGGG
705	CTTCCCGCGTTCCAGCGACCCGG
706	GAGCGCCCGCCCGAAGTTTGGGG

[0093] Table 24: Target sequences for CNBP gene

SEQ ID NOS	Target sequence
707	TTAATAGGGAGGGTAGTTCCAGG
708	TGGGGTGTCGATGATTCAAAGG
709	GTTGGCAGGTATTGCTCAACTGG
710	GCTTCAGAAGCAAATACGAGAGG
711	AGTACTTGATTAGATTGGATTGG
712	CAGTGCAGTATACGTACATCAGG
713	ACCACCTGATTCACTGCGATAGG
714	GTTCCACATGTAAACCATATGG
715	ATGCGGGTCTTCGGCGCCACGG
716	AGCTGGGTCGCCGAGCATGCGGG

[0094] Table 25: Target sequences for COL3A1 gene

SEQ ID NOS	Target sequence
717	TTGGTGTGAAGAGTAATAACAGG
718	TCTATACTGCAGGTAAAGCAAGG
719	CTCTCAACTATGATACTTACAGG
720	GACTACCATTAATCCCAGGAGGG
721	GGACTACCATTAATCCCAGGAGG

722	AAACTTACGCGTTCACCACGTGG
723	TATTATGTCATCGCAGAGAACGG
724	TACCGTAATTGTTATACCTGAGG
725	GAAACATTTGTACGTACAGCTGG
726	GCCAGTTTTAGGTAACAATGAGG

[0095] Table 26: Target sequences for CRB1 gene

SEQ ID NOS	Target sequence
727	CCTGGAAACGAACAGCACCAAGG
728	CAGTAACCTAACTTACAGGGTGG
729	GA CTCAAAGATAGTGCCGGGAGG
730	TCATCAGTAGAGATCCTGGGAGG
731	GATAAGCTCTGGTAACAGGGTGG
732	GACATGTGTATTCTATACGGTGG
733	ACCTCCAGCATAACACAAGGAGG
734	TAGGGGCTAAAACCGACATGCGG
735	TGAATTGCAGGAACTCATCGCGG
736	CTATAAGTAGAACGTCTGGGAGG

[0096] Table 27: Target sequences for CRX gene

SEQ ID NOS	Target sequence
737	ACACATCTGTGGAGGGTCTTGGG
738	GGCGTAGGTCATGGCATAGGGGG
739	GGGGCGTAGGTCATGGCATAGGG
740	CGGGGCGTAGGTCATGGCATAGG
741	ATCCCGGGATCTAAACTGCAGGG
742	CATCCCGGGATCTAAACTGCAGG
743	GCGGTCACAATCGTGCCAGACGG
744	GAGCTCGTGGTGTACTTCAGCGG
745	CTTACCAGTTACTCACCATGGGG
746	CACTTACCAGTTACTCACCATGG

[0097] Table 28: Target sequences for CTNS gene

SEQ ID NOS	Target sequence
747	GATGCCACGTACAGTTACCGAGG
748	GGTCTTAGAAAACCATCGTGGGG
749	TTATGCGCTTCCTTACACGAGGG
750	GAATCACAGGAGATCGCTAGCGG
751	ACGATCAGTCTCCAGCATGTGGG
752	ATTGGTTACTTACTTCATCGGGG
753	AGCGCAGAGGAGATTCACGATGG
754	AGAATGCTCACCCACGCAGGAGG
755	CTTGACGCCGCAATCCTCCAGGG
756	GCCGATGTTGAATACACTGTAGG

[0098] Table 29: Target sequences for CYP11A1 gene

SEQ ID NOS	Target sequence
757	AATCTCTGATAGTATAAGATAGG
758	CTCATCATTAAACGTCACCTACGG
759	GGGGGGGTCTCCCTACAGTAAGG
760	CATAAGTGTGGTGGTATCATGGG
761	GACTGTAACGCTTGTGCGATAGG
762	GTACCGAGGGTCAAAGATGGTGG
763	GCAATCAGGAAACCTCGTGTAGG
764	TGTCCAAACAAGTAACTACCAGG
765	CAAGTAAGCTCAGTGATCCAAGG
766	ACACCGATCTTTATCCCCCTGGG
767	GACACCGATCTTTATCCCCCTGG
768	TCTCGCTATTGAAACATTGTTGG
769	TACTCTTATACCCCAAAGTGAGG
770	GAGGGTCCAGATCAATCCATTGG
771	TGAACATTGACCTCCATTACGG
772	GCAAGAGGCATAATGTGGGCAGG

773	CAATTCTGAATCATGACAACAGG
774	GATTCAATGGGTAATCCCCTTGG
775	TTAGTTATACTCTACACATAAGG
776	GTCCTGAGTACTTGGATACTTGG
777	GTA CTGCCCTTCTTTGGAACGGG

[0099] Table 30: Target sequences for CYP2C19 gene

SEQ ID NOS	Target sequence
778	GGAGAACTATTAGTCATTGCTGG
779	TAGTAGGCTATATTAAATAGAGG
780	GTTAAGGGTCATCACTTTCAGGG
781	CTAACGTTTAAATCTTTGGCCGG
782	AGTATTGTAATCTATATGGGAGG
783	GTCCCCTCAATATTAGTATTTGG
784	GGGGCGCACGCATGTGTGACAGG
785	TTGTTCTGGCTACTCTTAAGTGG
786	GTA CTAAATCAGTGACCTCAGGG
787	CTTATGTCAAGGGAATCCACTGG
788	AACTCCTCACTCACCTCTATAGG
789	TTGCTAAAATGCCCACAATCAGG
790	ACTGTTCCGGTGAATCATAGGAGG
791	AGAACTGTTCCGGTGAATCATAGG
792	CTACATATACTGCAGTATTGAGG
793	TGAATATCCCAATATAGATCAGG
794	GCGAATATAATACGTTTTTGTGG
795	CTTTAGTCTGGTGGCCACATTGG
796	AATAGACCTGCTGAATATGTTGG
797	AATGGCCCTATCACACCCCTAGG
798	ACGAGGAGTATGTACAAGGGAGG
799	CAGGTTTGT CATCGTACCCCAGG
800	TTATCCGATTTTACAGTGTGTGG

801	AAAGGAGCACCGGGCTGTATGGG
802	TAGTTACACCCCCATTGGAAGGG
803	ATAGTTACACCCCCATTGGAAGG
804	CTTTATAGTTACACCCCCATTGG
805	GGCTCCACTCCTCAATCTTAGGG
806	GGCTACTCACCTTCAACTGGGG

[00100] Table 31: Target sequences for CYP2D6 gene

SEQ ID NOS	Target sequence
807	TCCGGTGTCTGAAGTGGGGGGCGG
808	GAATCCGGTGTCTGAAGTGGGGGG
809	CGGCCCCGAAACCCAGGATCTGGG
810	GACGAGATCTCCAAATGCCCAGG
811	CCCTCTACAGGTGGATTGTATGG
812	GCCATACAATCCACCTGTAGAGG
813	CGGGGTTGATAAGTCCGCTGGGG
814	GGGGTTGATAAGTCCGCTGGGGG
815	ATAAGTCCGCTGGGGGTGACGGG
816	GGCACAGGATTCACTTATTGAGG
817	GCAGTCCGGTGGAGTGCTGTCTGG
818	CTTTCCGACATACACGCAATGGG
819	AATTGTTCCAATCTGCTCTTGGG
820	CGGCTGGAACCTGCTGATCTCGG
821	GGGCGATAATGTGGCAACTCCGG
822	GAAGCGAAGTCTTTGCCGAGTGG
823	AAGCGAAGTCTTTGCCGAGTGGG
824	CCGGGCGTGGCTTCAGTGCTCGG
825	ACCTCCGGTTGCTTCCTGAGGGG
826	GTCAAGACAAGTTCTCACAGAGG
827	AAGGCGAGGTCGTTAAAGAAAGG
828	AGGCGAGGTCGTTAAAGAAAGGG

829	CAGCCTCGTCACCTCACCACAGG
830	ACGTACCCCTGTCTCAAATGCGG
831	GCCTGGCCGCATTTGAGACAGGG
832	TCGAAATCTCTGACGTGGATAGG

[00101] Table 32: Target sequences for CYP11B1 gene

SEQ ID NOS	Target sequence
833	CGCGTGTTCTCTACTCTCTGGG
834	GCGCGTGTTCTCTACTCTCTGG
835	ACAGTACACCAGCATCGTGGCGG
836	TCAACAGTACACCAGCATCGTGG
837	CATGACGTGATCCCTCTCGAAGG
838	CAAGGCTCTTGGATAAGATAAGG
839	GCGTACCAGATGACGAGAGTGGG
840	AGCGTACCAGATGACGAGAGTGG
841	GCTGCTAAACCGGGTCAGGTGGG
842	ATGGTGAGGAGCGTACCATCTGG
843	GAGCCGGTACTGGGAGAACCTGG
844	AACACGCGCACCAATGTCTGCGG
845	GACCCCCGAGCTGCGACACTGG
846	ACGATGCTGGTGTACTGTTGAGG
847	TGACCCACAGGGCTTATCAGTGG
848	CAATGCAGGCACACCCCATTTGG
849	ACGCCGGGGCATGGCTTCAAAGG
850	CTTCGAGAGGGATCACGTCATGG
851	TTCGAGAGGGATCACGTCATGGG
852	GGGGGTGCATGAGCGTAGACAGG
853	GGATTATTCATCTCCTTGCAAGG
854	CTTAGAGATTTTCAAGTCCGTGG
855	TCATGCCCACTCTCGTCATCTGG
856	ACTCTCGTCATCTGGTACGCTGG

857	ATCACCAAATGTATTAGTGCAGG
858	GCACCGTTCCCCCTTGATACTGG
859	CTGTCAGTTCGAGGTGAATCTGG
860	AGTAGTGCATTCTGAACTGAGGG
861	GGTAAAAGGCTCTTTGGGGGAGG
862	GGTTATTAAGGATTGCCACAAGG
863	ACCGGTGAGTCATTCCAGTCTGG
864	CCGGTGAGTCATTCCAGTCTGGG
865	TGGTATATATGAGTGCTGTAGGG
866	GGCTGGGTACACTCTCAAAGTGG
867	ATCCGGCCGGCCAGAGTTCAGG
868	GTATGGCCACACGAGGAGCCTGG
869	TCGGGAGTTCATTTGTGCTGGG
870	CGGGAGTTCATTTGTGCTGGGG
871	GCAGAGACGTGATTAGTTGATGG

[00102] Table 33: Target sequences for CYP11B2 gene

SEQ ID NOS	Target sequence
872	TGTGAGAACCCGCCCTGAAGAGG
873	AACCGCCCTCAAACTACACAGG
874	CTTGTTGTAAGCGGCGAGTTGGG
875	TCTTGTTGTAAGCGGCGAGTTGG
876	ACGTATCGAGATTCCTCACATGG
877	CAGAAAAGCTCGTCTATGTCAGG
878	GGCTCTATGAATCTGAACTACGG
879	ACCTCTTCACTGCGTCAGCACGG
880	TGCGGCCAGACCTATGGGCAGGG
881	GTGCGGCCAGACCTATGGGCAGG
882	GGGGGGTGCGGCCAGACCTATGG
883	CCTTGGGCGACAGCACATCTGGG
884	ATCCCCACCTAAACACTGTCGGG

885	CCCCACCTAAACACTGTCGGGGG
886	CAGTGCAGACGCGACCCACAGG
887	ACAGTAACCGCACCCCGCCTGG
888	CCCAACCCGTGAACATTACAAGG
889	CCAACCCGTGAACATTACAAGGG
890	CTCACATGTGGCACGCTACACGG
891	GTACTTCCCGAATTACCAAATGG
892	CTGAGTTGAGGGCCGTTCTCAGG
893	ACCAGGCACCGAACCTTGCAGGG
894	AATCCCGAGATCTGCTCCGCTGG
895	AAGGCACTCACTCCAAGTTGAGG
896	AAGTTGAGGGGGGCGGCACCTGG
897	CAGAAAGGGCCGACCGCGGTGGG
898	CACCCCTCCGCATTCTCGTCAGG
899	ACCCCTCCGCATTCTCGTCAGGG
900	CAGAGCTTGCCGGCTAACTTGGG
901	AGAGCTTGCCGGCTAACTTGGGG
902	CGTTTCAGCGGGTGATTGCTCGG
903	TTTCAGCGGGTGATTGCTCGGGG
904	CCCGAGTCAAGTCCTTCCAACGG
905	CAACCAACATCCGCCCCGACAGG
906	TCCCGCTGTCATGTCAGAGCTGG
907	GCAACTGTCTTCGAATAGGCTGG
908	CAATAACATTGGCCAACCTCTGG
909	ATGGATCATACTGTTGTTCCAGG
910	ATTAACCATGGATTGTACCATGG
911	TTATGACCAAAAGGCCCCCATGG
912	TAACCACGCAACTTAGGCTCAGG
913	AGCTACACTAAGGCATGAACTGG
914	ACTTAGATGAAGGTGTTCTGGGGG
915	TCCCGTGCCGAAGAGACACCTGG

916	GCCCAAGGCAGGTTACGTAGGG
917	CATGGCTCCGTATCAACCAGAGG
918	TTACCAAAGTGTGACCTCGATGG
919	TAATCGCTCTGAAAGTGAGGAGG
920	TCTCCATGTATGAGCACTCCCGG
921	ACGCCGACCTCAACCAACCAAGG
922	CATTGCGACCCAGCGAGTAGAGG
923	GAGGTTACCGGTATGGAGCCAGG
924	CCTGTACCAATGTCTGCGGACGG

[00103] Table 34: Target sequences for DMD gene

SEQ ID NOS	Target sequence
925	CACTCATGCATCCTCTTAGATGG
926	GTGGTGGTTGACTATGGTAAGGG
927	AAATCCGAATCCCCAGGCCAGGG
928	GTGGCCAAATCCGAATCCCCAGG
929	TGAAACTCGCATTCTATAAGGAGG
930	GCATTCTATAAGGAGGCACACAGG
931	GTATATAGCAGTGCATGCCAGGG
932	ATTGTAATAGAGGAGGCCATAGG
933	TCTTGCCATATATGATCCTATGG
934	GTCCTCAGGAATACTGCCATTGG
935	AAGTACCATCTACACAGATCAGG
936	GCATCGAATCTCAAGAAATATGG
937	GATCTCAACATAACGTCTTCCGG
938	GGAAATGTAGTGAAGATCGGGGG
939	GGGAAATGTAGTGAAGATCGGGG
940	TGGCTAGCTTTCCCTACCAAAGG
941	GATCAAGTGCTTAATAGAGGTGG
942	TTGGTAGGGAAAGCTAGCCAGGG
943	AGGGTAAACAGGAACGCTTCAGG

944	ATGGATGGCCCTGAAGTCACAGG
945	GTATGGTGGGTCCTAAACAATGG
946	TATGGTGGGTCCTAAACAATGGG
947	CATACCTGCTACACGTATATAGG
948	GTCCAAATTGCTCTTTGAGCTGG
949	AATTCCTATATACGTGTAGCAGG
950	GGGATCTCAAGGATGTAGGCAGG
951	ATAGATTTTCATGACGTACTAAGG
952	CCATAAGATTGCCTCAACTCAGG
953	GCCTCAACTCAGGTGTACCTCGG
954	CTGAGTTGAGGCAATCTTATGGG
955	TGAGTTGAGGCAATCTTATGGGG
956	ACGTCATGAAATCTATATAGTGG
957	CATTCCACTCAGGTACCTAAAGG
958	GATCTATCTGGCTTTCAATCAGG
959	TTTAAGGGGATCATTGCCACTGG
960	GTGCATACATACAAGTTCTATGG
961	TAGTAGGTGCTGGTATCACAAGG
962	GCTGTGGATTAGGCCTAGATTGG
963	CACGTCTTCTGACAATGAGATGG
964	ACGTCTTCTGACAATGAGATGGG
965	CGTGTTAAATATCCCTGTGTTGG
966	TAACACGTTGATTGCTGTTAAGG
967	TACTGCAAACCAGCCAACACAGG
968	ACTTGAATTGGAGCAATGCCTGG
969	GTTAAGGCTAAGATGTAGTTAGG
970	ATCTGGCTTTAGAGCTGAATGGG
971	TAACCACCACTCCTTCGTCACGG
972	GCACTATTTTGGTGGAATGCTGG
973	TAGTGGGATCACATCCCTGTGGG
974	TGGAAATTAGCCCGGTGGCATGG

975	TTACATGGAAATTAGCCCGGTGG
976	GAGATGGCATTACCCTTAGATGG
977	GAATGTTCTTGGAGAAGCGTTGG
978	AATGTTCTTGGAGAAGCGTTGGG
979	TCGGTGAGGTGAAAGATTAAAGG
980	GTGCATTTTAGAAATCGGTGAGG
981	ATGATACCCTTAAGGTACTTGGG
982	TCATAGACCCAAGTACCTTAAGG
983	CATAGACCCAAGTACCTTAAGGG
984	AACTATAGGTCCCACCCAACAGG
985	AGTTTGATGTGCTTTTCGAAAGG
986	ATGTGCTTTTCGAAAGGTTATGG
987	GTCCTCAGAGCCTATGCCAGGG
988	TTAGGCCTCTTTCGGAGAGAAGG
989	AACAGTTTGTGTCGGTATAGAGG
990	AGATTTCAAGGAGCCTAATAGAGG
991	GGCTATATTGTTGTCACAGCAGG
992	GAAGACCCAATCTTGACACCAGG
993	CTATACTGTGCCCTAAGATGAGG
994	CTGACCCTGGTGTCAAGATTGGG

[00104] Table 35: Target sequences for DMPK gene

SEQ ID NOS	Target sequence
995	GGGCACTCAGTCTTCCAACGGGG
996	CTGGTCATGGAGTATTACGTGGG
997	GATGGCGCGCTTCTACCTGGCGG
998	CGTCATTGGCTGCTTCCTAGCGG
999	GCGGTTGATCGACAAGACCAAGG
1000	TGGGCAGACGCCCTTCTACGCGG
1001	CAACTCCCCGAGTGGCACAGTGG
1002	ATAAATACCGAGGAATGTCGGGG

1003	GAAGTAACCTCGTCTCTCCGTGG
1004	AGTCCCCACGTATATGGCAGGG
1005	CGAAGTTCTGGTTGTCCGTGCGG
1006	GACATTCTACATGAGAACGTGGG
1007	CCTTCTTATGAAACCCTTGGGGG
1008	CCCCTCTTCTCGACGCTCGGTGG
1009	GCCTGACGTAGTAAAGATCGGGG
1010	GGAGAGCGGTACCACTTGTGGGG
1011	GCTCCCGTTCACCAGGATGGAGG
1012	GTCTCAGTGCATCCAAAACGTGG
1013	AACCGCATCGTGAAGCAGGACGG
1014	TTGCGAACCAACGATAGGTGGGG
1015	GTGGGGTTCGCACTCTTACGAGG
1016	CAGCGTGCCCCCCTTTACACCGG
1017	GGACATTCTACATGAGAACGTGG
1018	GTCCTTCACCGAGGGCCGCGTGG
1019	TACATGGGAAGGTGGATCCGTGG
1020	TGCGAACCAACGATAGGTGGGGG
1021	CCAGGCCGTTGATGATGACGGGG
1022	GGGCCACACCCGTCACGATGGGG
1023	CAAATGCGCAGCTAAGCGGGTGG
1024	CCGGCCCACAACGCAAACCGCGG
1025	AAGAGGCATAGGGCGCGTGGAGG
1026	TCTAAAGTCGCAAAGACGTAGGG
1027	CCCAATAGAGGCTAAAACGGTGG
1028	GAAGCTCCCGTTCACCAGGATGG
1029	GTCATGGAGTATTACGTGGGCGG
1030	CACTTAGTCCCCGCGCCCCGCGG
1031	AGGTTACGTTTCACAACAAAGG
1032	TCGAGCTTGCGTCCCAGGAGCGG
1033	GTCAACCTCACCCCCTGCGGTGG

1034	AAATATCCAAACCGCCGAAGCGG
1035	TAGGGTTCAGGGAGCGCGGGCGG
1036	ATGAAATGCGGGGTGTCGGAAGG
1037	GGCGCTTCTCGTCCGGCGTGGGG
1038	AAGATCCGCCCTCCTGCCGTGGG
1039	AAGCCTGACGTAGTAAAGATCGG
1040	AGCAAATTTCCCGAGTAAGCAGG
1041	CGGCCGGCCGCAGAGAGAAAGTGG
1042	GCGAGGTCAACACCCGGCATGGG
1043	TGCGTCTTCAGCACCAATGTCGG
1044	GCAGCGGTTCAAGTCAAGCTGG

[00105] Table 36: Target sequences for EGFR gene

SEQ ID NOS	Target sequence
1045	GGGCACTCAGTCTTCCAACGGGG
1046	CTGGTCATGGAGTATTACGTGGG
1047	GATGGCGCGCTTCTACCTGGCGG
1048	CGTCATTGGCTGCTTCCTAGCGG
1049	GCGGTTGATCGACAAGACCAAGG
1050	TGGGCAGACGCCCTTCTACGCGG
1051	CAACTCCCCGAGTGGCACAGTGG
1052	ATAAATACCGAGGAATGTCGGGG
1053	GAAGTAACCTCGTCTCTCCGTGG
1054	AGTCCCCCACGTATATGGCAGGG

[00106] Table 37: Target sequences for EPCAM gene

SEQ ID NOS	Target sequence
1055	AGCAAATGATTCAACACCGGGGG
1056	AGGCTTTATATATGCCCTCTGG
1057	GAGGTCTCTAAATCTATCAAAGG
1058	AACGGCAGCAGCGAACCATTGG

1059	TCTAGCTGCCATCCCACTGAGGG
1060	TTAGGGTACTTGGGATACGAAGG
1061	CAGTCCCCCTCGCTACCCATTGG
1062	AAGATGAAGTTCTCCCGATTAGG
1063	AAAGATCCCTAACGCCGCCATGG
1064	CGCCATGGAGACGAAGCACCTGG
1065	CAACGAGCACCAGCGGCCAGAGG
1066	GCGAGCGAGCACCTTCGACGCGG
1067	CGAGCACCTTCGACGCGGTCCGG
1068	GAGCACCTTCGACGCGGTCCGGG
1069	AGCACCTTCGACGCGGTCCGGGG
1070	CCCCGCAGGTCCTCGCGTTCGGG
1071	G TTCGGGCTTCTGCTTGCCGCGG
1072	GCTTCTGCTTGCCGCGGCGACGG
1073	GCCCTCCGCGCGGTAGGAAACGG
1074	GTTTCCTGCGGCCACCGAACCGG
1075	CCCTGGCGCACCCACGTCCTCGG
1076	GCGCACCCACGTCCTCGGTTCGG
1077	GCACCCACGTCCTCGGTTCGGGG
1078	CCCACGTCCTCGGTTCGGGGTGG
1079	GGCCGCTATGCACCTGCGCGCGG
1080	GCTATGCACCTGCGCGCGGCAGG
1081	TATAATATTGCCCCAGCAGGTGG
1082	ATAATATTGCCCCAGCAGGTGGG
1083	TGTGTAATACTGATGTTCCAGG
1084	GATCACAACGCGTTATCAACTGG
1085	ACAGTAGTAGGAAAGGCGTTGGG
1086	TGTTGATACAAGCTGTGCACAGG
1087	ATATTCTTGCGTGAGTTCCATGG
1088	CCATTCTGTAGTAGGTCATCTGG

[00107] Table 38: Target sequences for ERG gene

SEQ ID NOS	Target sequence
1089	CGGCACTGAATACATCCCAGAGG
1090	GTATTACATTGAGAACCATGTGG
1091	GGAATCTGACGATATCCCTGTGG
1092	AAAGCTGGTTCGATGCAGTGGGG
1093	ATCAGAGTCTACTTACAGCGAGG
1094	ATAACGTGATCACAGCGTGGCGG
1095	TTAATAACGTGATCACAGCGTGG
1096	GCTCACGAACACCATCACATGGG
1097	GATGCACAGAACACGCACAAGGG
1098	CGGGGCACAGGAGTACACCAAGG

[00108] Table 39: Target sequences for EVC gene

SEQ ID NOS	Target sequence
1099	TAGGTGGAAGATCTGAACCAGGG
1100	CCACCACACTCTCAATACGGAGG
1101	ATGCCTGAATAAACCCACCGGGG
1102	GCGATGCCCTGTGAGCAACACGG
1103	ATTTGAGAGATCCATCCGTGTGG
1104	GTGTCATCCCAATAACAGCGGGG
1105	TGTGGCTTAGATACCCTGGTAGG
1106	GCGCCCAAACCGAATCAGAGCGG
1107	GTGATGTGAGATCGTCAGGGAGG
1108	AGAGCGAAACCAGAGCTCGGTGG
1109	ATAATACAAGCATACCATGGAGG

[00109] Table 40: Target sequences for EVC2 gene

SEQ ID NOS	Target sequence
1110	GTATAGAAGACGAACCCCAGAGG
1111	ACCTACAATGTACCGCACAGTGG
1112	CGTAAGTGAACCCACCACAGGGG

1113	GCCGAAGCGTTAGTGCACAGTGG
1114	GGCGTAATCAGCAAACAGCGGGG
1115	ACAGGCTATATAGTCCAGAGGGG
1116	CCACCACACTCTCAATACGGAGG
1117	ATTGCGAAAGAATGGCCCAGAGG
1118	TAATATCTTTGAGTGCTACGGGG
1119	GCGCCCAAACCGAATCAGAGCGG

[00110] Table 41: Target sequences for F8 gene

SEQ ID NOS	Target sequence
1120	ACTGTAGTAAGAACACAACGTGG
1121	GTACACAGAATGACGCCACGAGG
1122	GTTGTGGGAGTGGAACACTACGTGG
1123	TTATGGGCAGACAACCACACAGG
1124	CCGATCTGAGATACCCATGAAGG
1125	TACGATGGTAGACACAAAGGAGG
1126	GGACACACCCCACTAAACGATGG
1127	TGTATCGAGCAATAATTGGAGGG
1128	TGTATGCACTACTTCTGGAGGGG
1129	TGTTACGATGGTAGACACAAAGG

[00111] Table 42: Target sequences for FBN1 gene

SEQ ID NOS	Target sequence
1130	GAAGTCCAAGTACTACACAGTGG
1131	ATAACAGAGTGATACCCACGAGG
1132	CATATGTTTAGTCCACATGGGGG
1133	ACACTCGTCATTACAGCACCAGGG
1134	GGTACATACAAACACCTCTGGGG
1135	TGCTCATACGAAGACAACCGAGG
1136	GAGTGTATCAGATCACCTAGAGG
1137	TTTCTCCTTACCGATACACGCGG

1138	TACCAATACACTCCCCACGGAGG
1139	ACATACCATCAGGTTCCGTGGGG

[00112] Table 43: Target sequences for FGFR1 gene

SEQ ID NOS	Target sequence
1140	CATGTGTTAACAGTGCATTGCGG
1141	GAACACGCTTGATACACATGTGG
1142	TACTGATCCAACATACAGGGTGG
1143	CTAAATTACAGTGACGAGGTGGG
1144	TGAGGAATGATCCCATTGCGGGG
1145	GTTGCCCCGCAACAAAACAGTGG
1146	GCACTGTCAAGGCTACGTGGGGG
1147	GTGAGGAATGATCCCATTGCGGG
1148	CCTCGACGTCCATCCAAGTGGG
1149	ATGAGTCCAGAAGTTGCGGGGGG

[00113] Table 44: Target sequences for FGFR2 gene

SEQ ID NOS	Target sequence
1150	TGACCAAACGTATCCCCCTGCGG
1151	GTGCGTTGCTTGATCAATGGGG
1152	CAACTGTTACCTCCCACCCGGGG
1153	AACCAGTGCACTAAACACGTGGG
1154	TCCAGGAGTACTATCCACCTGGG
1155	AGACCAATGAGATTCCACGTGGG
1156	GTTGCGTTGACGTAATGACAGGG
1157	ACTTTAAAGTCCCCGCCATGTGG
1158	ATGACGTTAACACCCAGCAGAGG
1159	GAGGCCCTTAGAGCGTTCCGAGG

[00114] Table 45: Target sequences for FGFR3 gene

SEQ ID NOS	Target sequence
1160	ATCGTGAACGTATTGCCAAGTGG

1161	GAATTGCCGCTCACACCACAGGG
1162	GAGATCGCATGGCTCCCAGGGGG
1163	TTTCCGTCATGACCGCCGTGTGG
1164	AGAAGCTCCGTACCCCCGGGAGG
1165	CATCGTGGCACAGACATGGGGGG
1166	GACCCCAAGGTACAGATCGAGG
1167	GTTAGAATATACCTCGTGTGAGG
1168	GTGCGTAGTGGGCAGAACGGCGG
1169	CGTGCAGGTGAGGGTCATCGTGG

[00115] Table 46: Target sequences for FMR1 gene

SEQ ID NOS	Target sequence
1170	TACACTAACCATCATAGTAG
1171	GGCATACTCGGTAGCAAAGTAGG
1172	AACAATCTGCTATCAGTAAC
1173	CTGGGTTTGAGCACATCAAT
1174	GTATGTTTGCAATACAACACTGG
1175	AAACTGCTGGAGTACCCCAA
1176	AAGAGGACTATAACGGCAAG
1177	GCTTAAATTAGAGTGGCCCTTGG
1178	GATTGGATATGTCTCATTGCCGG
1179	CTTAAATTAGAGTGGCCCTTGGG
1180	TGCCAGACTTGGAGTGCCAAAGG
1181	CAACTATTCTAATGGCACTTAGG
1182	GACTGCATCAACTATTCTAA
1183	TAATGGCACTTAGGTGCTGAGGG

[00116] Table 47: Target sequences for FXN gene

SEQ ID NOS	Target sequence
1184	GTAATCCAGATACACCCAAGAGG
1185	GGCCTAAAGTAAGACACCAGGGG

1186	CTGCTGTAAACCCATACCGGCGG
1187	ACTTAGGGCAAGGTTACACAGGG
1188	TATCAGAGTATAGGGCCAAGGGG
1189	TACCCTGAGAGGATCGCATGTGG
1190	TATCTGACCCAGTTACGCCACGG
1191	TTTCAGAGTTCGAACCAACGTGG
1192	GGGCCTAAAGTAAGACACCAGGG
1193	GAAGTCAAGAGGTACCCCAAAGG

[00117] Table 48: Target sequences for G6PD gene

SEQ ID NOS	Target sequence
1194	GGTTCTGCATCACGTCCCTGGGG
1195	GCCGTGAGTTGATGTGACATGGG
1196	GGGGATTCGGGAGCACTACGCGG
1197	CAATGACAATATGCGTGGAGCGG
1198	AAAAAACCCGGTAAATTGCGGGG
1199	GCTTTTGAAACGAGGGCCCAGGG
1200	ATAATGGGAGAGGATTGCGAGGG
1201	GCTTCATCTCAAATTACACGTGG
1202	CTCGGTAATGATAAGCACGCCGG
1203	AGTAGGCGCCCAGAGCTGAAGGG

[00118] Table 49: Target sequences for GAA gene

SEQ ID NOS	Target sequence
1204	AATAGCAACGAGACCTGAGGGGG
1205	AGTTGGCATCAGTTCCAACGAGG
1206	CCGCAGGCTGAACACGACGGTGG
1207	ACCGTCCCCACTCTACAGCGTGG
1208	CTTAACGCACGCCAGAAACGCGG
1209	TCCAGCTAACAGGCGCTACGAGG
1210	TTGATTATATTCTCTCACGTGGG

1211	CTCCTTGATAACCTACACTGCGG
1212	GGTCGTACCATGTGCCCAAGGGG
1213	GCGGTCATTATAAATCTGCGTGG
1214	AACGCGGTGCTGCTTCAACACGG

[00119] **Table 50: Target sequences for GALC gene**

SEQ ID NOS	Target sequence
1215	TACTATACACCCACAATTAAGGG
1216	ACGCCGCTTTCATGATGTTCTGG
1217	ATGGGGCGCTGTTTTCTATCAGG
1218	TACTACTCAAACCACTCCTAAGG
1219	AATACGAATGCTGGTCTGTCTGG
1220	ATGTATGGCCCACTACTTAGTGG
1221	GTCTTGGAAGTATAACGTAATGG
1222	CCTCCCTGGTTAGAGAATCAGGG
1223	TACAGAGTATATGGGTCTTGTGG

[00120] **Table 51: Target sequences for GALT gene**

SEQ ID NOS	Target sequence
1224	GAATGAGCTCAATACCCCCGAGG
1225	AGGCAGACCTTATCACCTGGGG
1226	GCTTGTATCAACATTCCCCAAGG
1227	GATCCGCTGGAAAATCTGCAGGG
1228	GAAGTCGTTGTCAAACAGGAAGG
1229	TGGGGATTACCTACCGACAAGG
1230	ATGTCTGCCAGCGTGAGAGTGGG
1231	GTATAAGCGCTCGTGACAGAGGG
1232	CTAGGCAGACCTTATCACCTGG
1233	GACAATTCATAAGAACCCTGGG

[00121] **Table 52: Target sequences for GATA6 gene**

SEQ ID NOS	Target sequence
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1234	GCCGAAATAAATCAACCCTGGGG
1235	TTTTTCCGCGAAGTGACGGGGG
1236	GCCAATATAGGAGAACGCGGCGG
1237	ACCCGAGTTAAAGTTCCCAAAGG
1238	CCGGGGGAGACACTTTAGGGCGG
1239	TACTCCAAACAGTCCTACCCCGG
1240	CTTTTTATTACCAGCAGCGCGG
1241	CTTATTGATCTCCACGCCCGGGG
1242	TCGAATCGCGAATAGTGGTGTGG
1243	TCGCGAATAGTGGTGTGGCGCGG

[00122] Table 53: Target sequences for GBA Gene Cluster

SEQ ID NOS	Target sequence
1244	AAGCCATGGACGTTAGTAGT
1245	TAGAAAAGAGGGCTTACGGT
1246	AGCCATGGACGTTAGTAGTA
1247	AGGGCTTACGGTGGGCAATG
1248	AAAGATGGTACTTAAAGCCA
1249	GTAGAAAAGAGGGCTTACGG
1250	ACCAGATATGCTGAGTTGGA
1251	AGTTGGATGGCGCTCAAGAG
1252	TCCAACCAGATATGCTGAGT
1253	CGCTCAAGAGAGGTCAAGGC
1254	ACCTCCACTCTTTCTATAGG
1255	CCTGCTGAACTGCTTAACAT
1256	AAACTTTCCAGTGACCACAG
1257	TTGAATTTGTCCCTTTGAAA
1258	CTGCATCTCACTTGACCTCG
1259	CTGAACTGCTTAACATTGGA
1260	GCTGAACTGCTTAACATTGG
1261	CTCCTCCTTTTCACAGCAAT

1262	GCTTAACATTGGAGGGCCCC
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[00123] Table 54: Target sequences for GCH1 gene

SEQ ID NOS	Target sequence
1263	CGCGATAGATCCTGTGGTATTGG
1264	GGGGTTACTTCGTACTATAATGG
1265	TAGTCTAAAGTCAACTTGATTGG
1266	CTACTAAGCATTAAAGACAACAGG
1267	ATGGCGATTGAGCTGGGCGCAGG
1268	CACTACACCACTTTTATTGGAGG
1269	ATTGATGAGGTGAGGAGCCGGG
1270	GTTTGGCTAAATGTTGCGACTGG
1271	GAACTTGGCCAATCAATCTTCGG
1272	G TTCAGGTGCGTGGAAGCTATGG
1273	ACTAACTGGAAGTTTTGCCCTGG
1274	TGGCGATTGAGCTGGGCGCAGGG
1275	CACCATTATGACGTTACTAAAGG
1276	TCTGTGCTCGTTCAGGTGCGTGG
1277	AGTGCATTTTCACAGATCGTTGG
1278	TGTAAGGCGCTCCTGAACTGTGG
1279	ATACGCTTTGGTTAAAACGTTGG
1280	GGTCCCTGATAGAACCAGAATGG
1281	AGGCAACGCGATAGATCCTGTGG
1282	GTTACCAAGCACCTCCATGGAGG

[00124] Table 55: Target sequences for GJB2 gene

SEQ ID NOS	Target sequence
1283	GCACTGATGGAACCGTCCTGAGG
1284	CCAAGTACAGGAGAACCGTGAGG
1285	GGCTACGTGATATTGCATGTAGG
1286	ATTTAGAGCATTCTTCCCGGCGG

1287	ACGCTGCAGACGATCCTGGGGGG
1288	TTGTCAAAGACCAACCCGTGGGG
1289	GACATAGAAGACGTACATGAAGG
1290	GTTCGCGAAGAGGTGGTGTGCGG
1291	GTCTTCTATGTCATGTACGACGG
1292	GCTCACAGGAGATTATCCACTGG

[00125] Table 56: Target sequences for GJB6 gene

SEQ ID NOS	Target sequence
1293	ACCCACTCATCATACCACGAGGG
1294	ACACGCAGCAAATGAAACGGGGG
1295	ACCGAGTCTTGGAATCACAATGG
1296	GTACCAATCTATAAAAACCAAGG
1297	TATCTCTTGACACTTGCAGGGG
1298	TATGGCATAAAGTCTACTTGAGG
1299	AAACCAGCGCAATGGATTGGGGG
1300	ACGCTGCACACTTTCATCGGGGG
1301	ACCCTCGTGGTATGATGAGTGGG
1302	TCGCAGAAGGATAGACCCAATGG

[00126] Table 57: Target sequences for GLA gene

SEQ ID NOS	Target sequence
1303	ACCGAGATCTCACATGACGTAGG
1304	ACGGCCATAAACTACACTGAGG
1305	ACGAAACGTTGAAAGCTGCGGGG
1306	ATAGCCATGAGCTTTCGAGGGGG
1307	GCCACACATACTGTACCACAGGG
1308	AGTGGGTTCGAACTTCAGCTCGG
1309	TCAATAAGGAGGGTATAAGGGGG
1310	CGATGGCAGAGTTACCGGTGAGG
1311	ACTGCGATGGTATAAGAGCGAGG

1312	TTAAGGAATAGAGCGGTGCAGGG
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[00127] Table 58: Target sequences for HBA Gene Cluster

SEQ ID NOS	Target sequence
1313	AGAGTTTCACTGCATTAGCG
1314	TCCCGAGTAGCTGAGTAGCT
1315	ACATCTACAACACTGCCCAC
1316	CTGCCATAGGTGTTTACCAA
1317	GGGAAGGACATCACAAACGC
1318	ACAGTTGATACTGTACCCAC
1319	GGAGAAGGGACCTTCTAGCC
1320	GCCTGATCTTGACAGCCCCA
1321	CCAGCCTCAGGGGAGCTGAG
1322	CTCTCCAGTCGCAATGGGAC
1323	GTTTACCAAGGGTGATTCAT
1324	TGTTTACCAAGGGTGATTCA
1325	CTGCCATAGGTGTTTACCAA
1326	CTCTCCTCTCCAGTCGCAAT
1327	TTCCTATCAGTTGAGGGCCA
1328	AACCCTCCCTCTGATACCCC
1329	TGAGCATTCTGGGGTGACCT
1330	GTCTGGTGTGTGAGCATTCT
1331	AAGATATTCCTATCAGTTGA
1332	TCTGGTGTGTGAGCATTCTG
1333	GTGAGCATTCTGGGGTGACC
1334	TGTCTGGTGTGTGAGCATTC
1335	ATTCCTATCAGTTGAGGGCC

[00128] Table 59: Target sequences for HBB gene

SEQ ID NOS	Target sequence
1336	TTGGTTCTTCTATGGCTATCTGG

1337	CGGTTTGTTTCTATGGGTTCTGG
1338	GTAGACCTTATGATCTTGATAGG
1339	TACCTGTCTCAACCCTCATCAGG
1340	TTGTCTCTCCACATGGGTATGGG
1341	TAGACCTTATGATCTTGATAGGG
1342	AACCATCTCGCCGTAAACATGG
1343	ATATCCCCCAGTTTAGTAGTTGG
1344	TCACACTAAGTAACTACCATTGG
1345	CCTAATTGTGTAATCGATTGTGG
1346	GATTACTGGTGGTCTACCCTTGG
1347	TTACCTCTATAATCATACATAGG
1348	CTTTCCTTACTAAACCGACATGG
1349	GGAGTAGATTGGCCAACCCTAGG
1350	GGCCAAGAGATATATCTTAGAGG
1351	GCGAGCTTAGTGATACTTGTGGG
1352	TGGTTATCAGGAAACAGTCCAGG
1353	CGTAAATACACTTGCAAAGGAGG
1354	GGGTTGGCCAATCTACTCCCAGG
1355	GAGTAGATTGGCCAACCCTAGGG
1356	ATCTCGCCGTAAACATGGAAGG
1357	GCTGGCCCGCAACTTTGGCAAGG
1358	TGTATGATTATAGAGGTAAGAGG
1359	ACCGACATGGGTTTCCAGGTAGG
1360	AGCGAGCTTAGTGATACTTGTGG
1361	CTATCTTACTTACACATGAGTGG
1362	ACTATCAATGGGGTAATCAGTGG
1363	ACCACCAGTAATCTGAGGGTAGG
1364	GCATTTATGAGGTCAGCGTAGGG
1365	GACGAATGATTGCATCAGTGTGG
1366	AAGTCCAACCTACTAACTGGGGG
1367	TAAGTCCAACCTACTAACTGGGG

[00129] Table 60: Target sequences for HEXA gene

SEQ ID NOS	Target sequence
1368	TGGTTGACCCACCTACAGGAGG
1369	ATTTACCACAGGCCCGCGTGCGG
1370	AGGAGGTCATTGAATACGCACGG
1371	TCCTTCTACATCCAGACGTGAGG
1372	TAGAAGGAAATGTCTCGTCGTGG
1373	AACCTGACCAATCTCCTTAGGGG
1374	GTCTGTATTTGGTGTCCGAGAGG
1375	CATGAGCTTTAAGTACGTAATGG
1376	GTAACATGAAAGTTATGACCAGG
1377	ATTACCCAGAAGCTGTAGGAGG

[00130] Table 61: Target sequences for HLA-A gene

SEQ ID NOS	Target sequence
1378	TCCCTTGTCGGTTGTGTGAGCGG
1379	CTCACCTTTACAAGCTGTGAGGG

[00131] Table 62: Target sequences for HLA-C gene

SEQ ID NOS	Target sequence
1380	AGGCTGAAAACACACATCCCGG
1381	GTAAGCGATGACACTCTGAACGG
1382	CGAGGCTGATGCAGACATGTGGG
1383	CTATATGTGGAGGTGGCATCTGG
1384	CATGTGGGATCCTGGTGTCTGG
1385	TTGGAGTGGCATTGTGTGCTTGG
1386	GATGCAGACATGTGGGATCCTGG
1387	TAAGAGGTCACACCACATAAAGG
1388	CACGGATGTACTCACCAGTTGGG
1389	ACCGCACAGCAGGTCACTAGTGG
1390	GCACGTCTGTTTATAGGCTCTGG

[00132] Table 63: Target sequences for HTT gene

SEQ ID NOS	Target sequence
1391	ATATACAGTACGTTAATACGTGG
1392	TAATTGCCGAGGGATGAATGAGG
1393	TTATTCCAACCCATCCAGGGAGG
1394	TTTTGCAGTGATACGTCTGGGGG
1395	TGTAATCGTTGATATACGTGAGG
1396	AGTAAAGTGGTGAACCTACGTGG
1397	CCTGTCCTGAATTCACCGAGGGG
1398	CTTAGAAATCTTTCACCGAGGGG
1399	AGTAGTGGTATTCCAGATGGGGG
1400	TGTATCGTCACACGTTCTGTGGG

[00133] Table 64: Target sequences for IKBKG gene

SEQ ID NOS	Target sequence
1401	AAGACGAGGAGGGTTAAACGAGG
1402	CGAGTCACTTACAAACAAAGTGG
1403	CGGGGGCTCATGAGTCACCGGGG
1404	GCCGTGAGTTGATGTGACATGGG
1405	GGGGATTCGGGAGCACTACGCGG
1406	AAGTGTACGACCGTTTCCGGGGG
1407	CCTAAGTGTCCACCCCATCGTGG
1408	AACCGAGTAAAATCCTTGTGGGG
1409	GCTTTTGAAACGAGGGCCCAGGG
1410	ATAATGGGAGAGGATTGCGAGGG

[00134] Table 65: Target sequences for IKZF1 gene

SEQ ID NOS	Target sequence
1411	GGGTGTCGTAAACAAAACAGAGG
1412	GGTTTAGAGAGACGTACCAGCGG
1413	TGACTTGAGCGTCAAACCTGCGG

1414	CTACGCAAACTCAGCACAAAGG
1415	GGTTAACGAAGAATTCATCAAGG
1416	ATTGAACCCCGATATCAGTGAGG
1417	TGGCACTCACCACCAACCGAGG
1418	GCGTCACCCCAAAGTTTGCAGGGG
1419	GTAGTGCTAAAGGATTTCTGTGG
1420	CGGAAGCATAAACACTCTGGTGG

[00135] Table 66: Target sequences for JAK2 gene

SEQ ID NOS	Target sequence
1421	GTGCACTTACTCACATCACATGG
1422	GCGCCATCTCACACTTACTGAGG
1423	TTGCTTATACTTTCCCTACGTGG
1424	TGTGTAACGTATGTACAGACTGG
1425	AGACAGTTGAGCGTATATTGTGG
1426	TCGATAACTTATAAATCTGAGGG
1427	GCCCGGTCTCCTGCCATTCGGGG
1428	GGCAGCACAATAATTGGTAGGGG
1429	GGTTTGCTTTTCAGTGACGGAGG
1430	GGGGCAGCACAATAATTGGTAGG

[00136] Table 67: Target sequences for KCNH2 gene

SEQ ID NOS	Target sequence
1431	GGGGTATAAAGTCTCCACGGGGG
1432	CCCTCCACTGAAAAACGACGGGG
1433	GGCTCCATCGAGATCCTGCGGGG
1434	TCGCCCCGGGATACCTGACAGGGG
1435	CCGATGCGTGAGTCCATGTGTGG
1436	AGTCAACAAACCCACCTCCGAGG
1437	GTCAACAAACCCACCTCCGAGGG
1438	ACTGGCACATTTGCTGACGTGGG

1439	CTCTAACTCCGTACTGCCGGGGG
1440	ACCATCGTGACATGGTTTGGGGG

[00137] Table 68: Target sequences for KCNQ1 gene

SEQ ID NOS	Target sequence
1441	GTGCTGTAGATGGAGACGCGCGG
1442	AGTTATCTTACTGCACCCAAGGG
1443	CGGGATAGATGACACGAGCGGGG
1444	GCTCGAGGAAGTTGTAGACGCGG
1445	TTTGGCTCCACACCTCCGGGAGG
1446	GGGTGCGTGTTAATCAACAATGG
1447	ACGAGCGGGGCTAAGCAGGTGGG
1448	CAGGCGGGGTAAATGCACACTGG
1449	CGGTCTTTATGAGCATGCGGGGG
1450	GAACCTTTGCATATAACGTGCGG

[00138] Table 69: Target sequences for KLF5 gene

SEQ ID NOS	Target sequence
1451	GAAGTTGTGTACAAACTGCGCGG
1452	ACCCGTACCTACATAAGACGGGG
1453	CCCCAAGGTTTCATACCCGGTGG
1454	TTTACTCTCAGCGAAACGCGGGG
1455	GTTTCGCTGAGAGTAAATGGGGG
1456	TGCGTCGTTTCTCAAATCGGGG
1457	GTCAAGTGTCAGTAGTCGCGGGG
1458	TTAAGGTCTCGTGCATTACGTGG
1459	TGGTACTGATAACTTCACATTGG
1460	AATGGTACAGCACTACTAAGCGG

[00139] Table 70: Target sequences for KRAS gene

SEQ ID NOS	Target sequence
1461	GATTAGGTCAAATCCCTTTATGG

1462	AATACGCATCGTGTTATCTCTGG
1463	GCTTACTATTCAACTCTAACAGG
1464	AACTTTTTCGTTCCACGTACTGG
1465	CCTACTGTCGCTAATGGATTGGG
1466	TCCTACTGTCGCTAATGGATTGG
1467	TAGTTACTACTCAGTTGAACAGG
1468	TATACTTACGTAAATCCATTGG
1469	GCAATGTCATGAGTGAATACTGG
1470	CACCTATCCTACCCACGAATTGG
1471	ATACGCATCGTGTTATCTCTGGG
1472	ACTTTTTCGTTCCACGTACTGGG
1473	ACGCACCCTGAAATTGGAAGTGG
1474	TGCCAATTCGTGGGTAGGATAGG
1475	CGCCGAATGGTGACAGCAAGAGG
1476	GGGCAATGTTCATGAGTGCTGGG
1477	AAGGCTGCCAATTCGTGGGTAGG
1478	ATGACTTAGGTTTGCCAATGTGG
1479	ATCTCTGGGTCGTATACCAAAGG
1480	AGTATTCCATATCCATTTCGGGG
1481	GTTTAAAGTGACCCCAACACAGG
1482	AGTAGAGTGTGTGCGCCGAATGG
1483	GCTTTTATAGATCTGTATACGTGG
1484	TATAACTATATCCAGTACGTGG

[00140] Table 71: Target sequences for LCA5 gene

SEQ ID NOS	Target sequence
1485	GGTCACTGGGAAACTTATAAGG
1486	GAATAACTTCAGACCGAGTTTGG
1487	CAATGAGCAGGTGCAGTATATGG
1488	TACGGTAGTTTGATGTGATATGG
1489	TCCCCTAATGAGTTCGCATTTGG

1490	TATCGTCTGCATGTTTTAATCGG
1491	AGAACTCCATGTCGTAAAACAGG
1492	GCGAACTCATTAGGGGAGGCTGG
1493	ACCCCTGGCCCTATCCATAAAGG
1494	ACGGGTTCAGTGACATAAGAAGG
1495	CGAGTAGTACTTTAGAATAGTGG
1496	CTCTATGGAATACCTCCGTATGG
1497	CCGATACGTTGTTTTCTTG GGG
1498	TTGATGACCTTGGATCATGCTGG
1499	GAAAACGTTAGTTACTGTACAGG
1500	TATTCTAAAGTACTACTCGTTGG
1501	CGGGGATTCCCTAACTACCATGG
1502	TATGGCAAGTTTAACTGCACTGG
1503	GTAGCCCTCTTGTGTATGGTTGG
1504	TTCTAAAGTACTACTCGTTGGGG
1505	GCGTGGAGAGTAAACCAGACAGG
1506	AGGTCTGTTCTATACGAAGTGGG
1507	ATGCAACCCAAAAGTTCCGTGGG
1508	TATGCAACCCAAAAGTTCCGTGG
1509	TGCTTTACACATTATGACCGGGG
1510	CAGGTCTGTTCTATACGAAGTGG
1511	TGATGACCTTGGATCATGCTGGG
1512	AAAACGTTAGTTACTGTACAGGG
1513	AAATGCGAACTCATTAGGGGAGG
1514	GGTTACCCATGAGATTACACAGG
1515	ACTCCATGTCGTAAAACAGGAGG

[00141] Table 72: Target sequences for LRRK2 gene

SEQ ID NOS	Target sequence
1516	CAACGTCTGTTCACTTACGTGG
1517	CGTCTGTTCACTTACGTGGAGG

1518	CTGTTTCAGCTTACGTGGAGGAGG
1519	TCATCCGTTCTTATACAATCTGG
1520	CTATAGAACTATACTTGACATGG
1521	TTCATCTCCGGTTTGAAATCAGG
1522	ACATGCCAATTGTCTAAATAAGG
1523	GGTACAATGCAAAGCTTAATGGG
1524	GCTTGATACACCCAGATATAAGG
1525	GGCTCCCCGCTTTCATCCTAGGG
1526	GCTCCCCGCTTTCATCCTAGGGG
1527	ACCCAATATCCAGGTTGAGTAGG
1528	CCAGTTCCCAGACCTTCCGTGGG
1529	TTCTGCGCGGCCCGTCGCCTCGG
1530	GGCCCCTGAGCTCGTTTTTGGGG
1531	GGCCCCAAAAACGAGCTCAGGGG
1532	TCCTCATAAACAGGCGGGCGTGG
1533	GTGTTACGTA CTCCGAGCGCGG
1534	TTTCAAGTGATTACCGCGCTCGG
1535	GAGTCCAAGACGATCAACAGAGG
1536	GAGAGTCGCGAGTGTGCAGCAGG
1537	CGCGAGTGTGCAGCAGGTAAAGG
1538	ACAAGGTATACTACAATAAAGG
1539	GGGTAGGCGTTTTGGTCTGCAGG
1540	AGTGCTATACTTGACAACCCAGG
1541	GTGCTATACTTGACAACCCAGGG

[00142] Table 73: Target sequences for MDM4 gene

SEQ ID NOS	Target sequence
1542	GGGATATTATCGTTAAATATAGG
1543	ACTCCAACAAC TACTCATTGGG
1544	ATAGGGCCAGTTAGGGAGCGTGG
1545	AGTTAGGGAGCGTGGTTCATTGG

1546	AGATAGGGAATACAAGCGGTTGG
1547	GATAGGGAATACAAGCGGTTGGG
1548	TATAGGAACCTTAAGTCAGCGGG
1549	ATAGGAACCTTAAGTCAGCGGGG
1550	GGTGCATCCGTTACTATTATGGG
1551	CTCGTGTGAGGCCGTGTGGGAGG
1552	CGGCCGTACCGCCAGTTGTGCGG
1553	GGCCGTACCGCCAGTTGTGCGGG
1554	GTGAAGTAACTTTGGCCAACAGG
1555	AAACCTAAAGTCGACGTAGTTGG
1556	CACGTCAATGTCATTCTACCCGG
1557	ACCTACAGACAGTATCGAGATGG
1558	CCTACAGACAGTATCGAGATGGG
1559	CTACAGACAGTATCGAGATGGGG
1560	CGGGTGTTGCTTTTAACTGTGG
1561	GTAACCTGCAGTTAGTAGGTAGG

[00143] Table 74: Target sequences for MET gene

SEQ ID NOS	Target sequence
1562	ATAACTGTTTGATAAGACCGTGG
1563	CCATAACATTCTCCTAACAGTGG
1564	CAATTTTTTGACAACCTACGAGG
1565	AACTCTTCATCAGCTAACCAAGG
1566	AGGTCGTTTTGGTATCAGAAAGG
1567	CAAATCTCTCTAAACCCGGGTGG
1568	CAAAGCTCGCGCCCTTCCCGGGG
1569	TGTCAGTTCCTATTGGCACGTGG
1570	CTATTATGTAGATCTGCAGAAGG
1571	GGTAGAGTATCATATGTGCTAGG

[00144] Table 75: Target sequences for MLH1 gene

SEQ ID NOS	Target sequence
1572	TCTTGACTACAAAGCCTTA
1573	CAGTTTGGACGGCTGGTACT
1574	TTGTGATCAGTTTGGACGGC
1575	AGTTGTGGCAACCCGAAACA
1576	CAGTTGTGGCAACCCGAAAC
1577	ACCGGGCTCCATTTAGTTG
1578	CTCACAAGGTCATCCCAACC
1579	TCTCACAAGGTCATCCCAAC
1580	TGGCAACCCGAAACAGGGCT
1581	TTAATTGTGATCAGTTTGGA
1582	TACCTATAAGAATACTCATC
1583	ACCTTAAACAAGGCCAGACG
1584	TGGGTAGAAAGATATCCAAC
1585	CTAGATAGGACTATATTTAC
1586	GTAGCCATTAAACCTAGAT
1587	ACTCATCAGGACCTTAAACA
1588	TAGATAGGACTATATTTACT
1589	TCAGGAGTTCAAGACCAGCC
1590	GTCCTGATGAGTATTCTTAT
1591	CAGTAAATATAGTCCTATCT
1592	CTTGGCCTTGCAAAGTGCTG
1593	ACCACGTCTGGCCTTGTTTA
1594	ATGGTGATTTTACATGCAG
1595	TGCAGAGGGGAGCAACTATG
1596	TGGTGATTTTACATGCAGA
1597	CAACACGAATCTAGTCTTTA
1598	ATCCATATACCTCCCATATA
1599	TATAAAGTCCTGAGACCGCT
1600	AGACCGCTAGGAATCTATGA
1601	TGATTCACGCCACAGAATCT

1602	CACAAAGCCTGGAATATGAG
1603	AACACGAATCTAGTCTTTAA
1604	CGAATCTAGTCTTTAAGGGC
1605	CTAACTTCTAGCACAAAGCC
1606	GAGACCCTTCCATATATGGG
1607	TACTGAGACCCTTCCATATA
1608	GTGAATCATGTGTTCTTTCA
1609	GGGGAAAAGTGCTTGCATTA
1610	TTTCCATCATAGATTCTAG
1611	ACTGAGACCCTTCCATATAT
1612	TTAATTGCCTCTCATATTCC
1613	CAGGCTTTGTGCTAGAAGTT
1614	AGGCTTTGTGCTAGAAGTTA
1615	CCAACCCCTTGGACCTCAAC
1616	ATGATCTCTGGCCAACCCCT
1617	CAACCCCTTGGACCTCAACT
1618	CCATTCTGATATTGCAACCA
1619	TACTCAACTATTAGTGAATG
1620	CATACTCAACTATTAGTGAA
1621	GAGCAAATGGCAATCACTCT
1622	TCCATTCTGATATTGCAACC
1623	ATACTCAACTATTAGTGAAT
1624	CACTTCTCCCAAATCACTGT
1625	AGAGCAAATGGCAATCACTC
1626	CTGAGAGGTTCTTCTCCCA
1627	TCCATCTTCATTTCACTT
1628	TATGTAGATTTGCTAGGACC
1629	GTTAGGGCAAGTGGCGGTGA
1630	TCTGTCCCAGTTGAGGTCCA
1631	CCAGTTGAGGTCCAAGGGGT
1632	CTGTCCCAGTTGAGGTCCAA

1633	CAAATAGAGAGGTTTTCATC
1634	CTACAGTGATTTGGGAGAAG
1635	TGTCCCAGTTGAGGTCCAAG
1636	AAGTGGCGGTGATGGAGTTG
1637	AAGGGGTTGGCCAGAGATCA
1638	GTTGAGTATGTAGATTTGCT
1639	CATTGCTCTGTCCCAGTTG
1640	GGATCTGTTAGGGCAAGTGG
1641	GTCTGAGTATGGATCTGTTA
1642	CCCTGGTTGCAATATCAGAA
1643	GTTGCAATATCAGAATGGAT
1644	TATGGATCTGTTAGGGCAAG
1645	GGTCTGAGTATGGATCTGTT
1646	TGGTGAGGGACTTGAGACAC
1647	GATGGAAGCCTACAGTGATT
1648	CATGGTGTTCTTTAAGGCAG
1649	TGAAATTAAGTGTGGATATC
1650	CTGGAGGCAAAAAACGTTAA
1651	TCACTTCCTACTTCTGAGCT
1652	ACCCTGGCTTTCTGCTGAAC
1653	CCATGGTGTTCTTTAAGGCA
1654	TCTGGAGGCAAAAAACGTTA
1655	ACCATGGTGTTCTTTAAGGC
1656	TGTCACCTAGTGACAAACCA
1657	GGCCAGTTCAGCAGAAAGCC
1658	TGCTCTTTGGTGAACAGTCC
1659	GCAAAGGCACTGGCATAACAG
1660	TATATCCATGGTTTGTCACT
1661	GCTCTTTGGTGAACAGTCCT
1662	GGTCTGAATGTATATATCCA
1663	AAGGGTGTCTTGATCATCTC

1664	ATGGTTTGTCACTAGGTGAC
1665	AATAATCAAAAGTAGACCTA
1666	GAGAAGTAATCCCTGAAACA
1667	ATGTTCTGTCCCTACCTGTC
1668	TTCCCAACGTCTTCAACCAG
1669	GATTCCCAACGTCTTCAACC
1670	ATTCCCAACGTCTTCAACCA
1671	TGCTTGAGATACAACCAGTT
1672	CTGGCCAGCCTCTAACAGAC
1673	TCTTCAACCAGGGGTCTGAC
1674	TAGAGCACTAAGACCAAGTC
1675	TGTGAATGGTTTTCCAGTAA
1676	GAATAAGTCAGCTACTCAAT
1677	GAAGTCTTTAAGCAAGTCTA
1678	TGAATGGTTTTCCAGTAAGG
1679	GTTTAAGGGAATGACCTCCA
1680	CGGGTTCAGAGTTCAATATC
1681	GTGAATGGTTTTCCAGTAAG
1682	AGTCTTTAAGCAAGTCTATG
1683	CACCAAATGCAGACATAGA
1684	ATGTGAATGGTTTTCCAGTA
1685	CAGCTGTTAATAAATGTGAA
1686	GGGGAAAATCTAGTACTAA
1687	TCACCCAGGCTGGAGTGCAG
1688	TCTTCTGCCTCAGCCTCCCG
1689	TTTCGCTCAGTCACCCAGGC
1690	GGAGTGCAGTGGCACGATCT
1691	AATAAACTAGACTTTAAAA

[00145] Table 76: Target sequences for MSH2 gene

SEQ ID NOS	Target sequence
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1692	AAGACCCATTATGTGTGGGC
1693	GGTATTTCAACGTTTGGCCT
1694	GTCTGTGGTATTTCAACGTT
1695	ACACTCAAGCTATAGGTCAT
1696	GGTAAACTAACAATCGAAGG
1697	TAATTTAACGACCCACTACT
1698	TGAGTCATCTGTAATGCCTA
1699	GCAAGGTGTGACCCAGTAGT
1700	TGCAAGGTGTGACCCAGTAG
1701	TTAGGGAGTTCCTAATGACC
1702	TAGGGAGTTCCTAATGACCA
1703	GGTCATTAGGAACTCCCTAA
1704	GTTGAATTTTAGGTGTACCC
1705	TTAGGTGTACCCTGGTCATT
1706	GCCATGGCAATTTGTTCCCG
1707	TCCACGGGAACAAATTGCCA
1708	TACCTACAGTATACTTACCT
1709	TGCCTAGGTAAGTATACTGT
1710	CCAAGACATTAGTACGTTGT
1711	GTTACAGTAGGACACATAAC
1712	CTACAACGTAATAATGTCTT
1713	CCTACAACGTAATAATGTCT
1714	GATTCCACTTGGATATACGT
1715	CACTTGGATATACGTTGGAG
1716	CGTTGGAGTGGAATTGTCTG

[00146] Table 77: Target sequences for MSH6 gene

SEQ ID NOS	Target sequence
1717	TAGTTCAACCTAGTATAAGG
1718	ACATAGTTCAACCTAGTATA
1719	GGGTGGTTGTAAACCAGACA

1720	GTAAACCAGACAAGGCCACC
1721	GTTTACAACCAACCCCTTTGA
1722	TTGTATAGGTGCTACTAATT
1723	TCGAGCCTTTTCATGGTCAA
1724	GGACTTATTACTCCCAAAGC
1725	CGTGTTTAAGACTGTAAGT
1726	ATCCCATGCATGATTCTAC

[00147] **Table 78: Target sequences for MUTYH gene**

SEQ ID NOS	Target sequence
1727	CAACTCCGGACGATCAGCCC
1728	TGAGCCGGACTCCCCAACTC
1729	TAAACCGAACTTTGGCCAGA
1730	TCACAGGTATTGTGTACCTC
1731	GCTGAACTCAAGAAGCCGCA
1732	ATTCCTCACCATTTCGGA

[00148] **Table 79: Target sequences for MYC gene**

SEQ ID NOS	Target sequence
1733	CTTCGGGGAGACAACGACGGCGG
1734	GCCGTATTTCTACTGCGACGAGG
1735	ACCCCTCCATAAATACAAGGGGG
1736	TCCGTATTGAGTGCGAAGGGAGG
1737	TAAGTGATCAGACACCGTCAGGG
1738	GCGCGCGTAGTTAATTCATGCGG
1739	GGCGGGTTGGAATCGCCGCGGGG
1740	TGCGTAGTTGTGCTGATGTGTGG
1741	GTCAAACAGTACTGCTACGGAGG
1742	CGAGGGGTCGATGCACTCTGAGG

[00149] **Table 80: Target sequences for MYCN gene**

SEQ ID NOS	Target sequence
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1743	AAGCGAGTTAAACAACCCTGTGG
1744	ACAACACGCAGTCAAAGCGGGGG
1745	ACATACGAGCACTAACAAAGGGG
1746	GCTCCCCAACTGGTACAACGAGG
1747	TCGCACACCCTTGAGATACGAGG
1748	CTCCCCAACTGGTACAACGAGGG
1749	AGAAATCGACGTGGTCACTGTGG
1750	CTTTCTGCTCAGTCTCCGCGAGG
1751	TCCATGACAGCGCTAACGTTGG
1752	TCACCAACCTCGTATCTCAAGGG

[00150] Table 81: Target sequences for MYH11 gene

SEQ ID NOS	Target sequence
1753	TTGTGTTGCACTAACCCAAGCGG
1754	TTATACGTGTTAATCCAAGGTGG
1755	GATGCTCAAATTCAGCGCAGAGG
1756	GATTCCTACTTCCTACAAGCGG
1757	TGTTGCACTAACCCAAGCGGAGG
1758	TCACTCAAGATGATTCCCGAGG
1759	GTGGGATTTCACGCACCATGG
1760	AACTTGAGACCTTACACGTGG
1761	TGTGACGAAGAGAGCTGTGTGGG
1762	CCTTGTCAAAGACGTGAACGTGG

[00151] Table 82: Target sequences for NPC1 gene

SEQ ID NOS	Target sequence
1763	GTGGTAGGTCATGAAGTACGTGG
1764	CGTCCGTTCTGTCCACGATGTGG
1765	TGCCGAGCAGAGTTATGCGATGG
1766	AGCGAAACCAGCGTTTGCGAGGG
1767	TTATGCTCTGGAACCTACCGAGG

1768	GAGAAATATTAATCCGTGAGTGG
1769	CCTGTAAGGAAATACTCGGTAGG
1770	GTACAGTAAGATTGGTGTGATGG
1771	CCAACCGCACATCACACGCTGGG
1772	GGGTTATCCGAAAGGAACATGGG

[00152] Table 83: Target sequences for NPC2 gene

SEQ ID NOS	Target sequence
1773	AGCTGCCAGGAAACGCATCGCGG
1774	AACCCCGACGACAGGCAAGGAGG
1775	CAGATGCACCGAACTCAATGAGG
1776	TACCACTTAACACTGAACAGAGG
1777	TGCGCGGTCGGGTTTCATGGAGG
1778	GGCTTTTGAAATCACCGAAGGG
1779	ATCAACCCCGACGACAGGCAAGG
1780	GGGTTCCCTAAATCTTAAGGAGG
1781	TAGTCGGTAGAAAGTCAGGCCGG
1782	CGGTCACAAGACAAACCTGTCGG

[00153] Table 84: Target sequences for OTOA gene

SEQ ID NOS	Target sequence
1783	AAGTTGGCAATTCAGTAGAGGG
1784	AACTGGGTATCCCTGATATGAGG
1785	GTACCCATTGGTGTATCTTAGG
1786	GTACAAAGTCCTAACACCCCTGG
1787	TCAACTGAAGCTCCACGTGTGG
1788	AGCACAAGCGTGTTGATAGGTGG
1789	AGCGTGTTGATAGGTGGCAAAGG
1790	CAGTCATGATACTACCCACAAGG
1791	AATGGGGGAATCGGGCTGGCTGG
1792	CCTAAAAGGGGATGTGCGCCCGG

1793	TTAAATGTTGGCGGCTAATGAGG
1794	TCTCCCAACACCCCAAATACAGG
1795	CTCATACGACACAGTGATGCTGG
1796	ATGTATCAGCTACCCTAATCAGG
1797	CGTGATTGAGAACAGGTGACTGG
1798	TACTATGATCGATAAGAAATAGG
1799	CTATGATCGATAAGAAATAGGGG
1800	TCGATAAGAAATAGGGGTCTTGG
1801	CGATAAGAAATAGGGGTCTTGGG

[00154] Table 85: Target sequences for PAH gene

SEQ ID NOS	Target sequence
1802	GCAGCTTATAGGTTACCAGAGG
1803	CTGTGATGTAGAAGGAATCGGGG
1804	TCCGTTTTGATATGCAACCTGGG
1805	ATCCGTTTTGATATGCAACCTGG
1806	TAAGTAATTTACACCTTACGAGG
1807	GCTACGACCCATACACCCAAAGG
1808	TATTATGGCCCTTGTGACCATGG
1809	TGATTTACCCCTACCCTACTAGG
1810	TCATTTTAGGCCACACCAAGTGG
1811	AAGTATTACAGACGCACTGGTGG
1812	ACTTGGTGGTTGCGTTGAACAGG
1813	CTTGGTGGTTGCGTTGAACAGGG
1814	AACTCTCTGCCACGTAATAGAGG
1815	ACTCCGTGACAGTGTAATTTTGG
1816	CGTGACAGTGTAATTTTGGATGG
1817	AGCTCATTAGGCACAACAGTGGG
1818	TCAGTACTGGCAACAAATGTGGG
1819	GTTCTACTCCAATATATGGCAGG
1820	ATATGGCAGGGTGGGTCTTAGGG

1821	AGGGTGCATACACACTTTACTGG
1822	CCCAGCTGGCATATATAAGCAGG
1823	TAACACCCCATCAGTGGATCAGG
1824	TCGATTACTGAGAAACCGAGTGG
1825	ACCTCAATCCTTTGGGTGTATGG

[00155] Table 86: Target sequences for PCCA gene

SEQ ID NOS	Target sequence
1826	GTTACCTAATGAGACCATGGGGG
1827	CTTATCGACATGGAAGTGAGTGG
1828	TTTGTGTCCAATTCAGCGTGCGG
1829	AGTACCACATCGAACTGGAAAGG
1830	CGAGTCTGTCGTTAATTCTGGGG
1831	TGTTCTCGCGGGGATCCTGCGG
1832	GAAGTTCATATCACTCTAGGGG
1833	TCCCCTTCCGCAAGTTAGGGGG
1834	CGTTGCAGCTGTTCTCGCGGGG
1835	GCCAGTAGTTGTACTAACAAGGG

[00156] Table 87: Target sequences for PCCB gene

SEQ ID NOS	Target sequence
1836	CGTGCCCCATGAAAGAGTGATGG
1837	ACATGCGTACTCAGGTGCGCCGG
1838	TGTGCGCGTGCAGGAACTTGTGG
1839	GTA CT CAGGTGCGCCGGTAGGGG
1840	GCGACCTATCACTGCGTGCCCGG
1841	AACGCATCGAAAACAAGCGCCGG
1842	CGCATTTGACAAGGGTCCAAAGG
1843	AAGGTCAAGAGTACCCATTACGG
1844	GCGTACTCAGGTGCGCCGGTAGG
1845	GTCACAGATACCAGGATACTGGG

1846	CGGCACAGCAAAAATGGCGGCGG
1847	GTAATTGCATTGAGATCAACGGG
1848	TCCGTAGATTTTCCCAGAAGAGG
1849	TTGCCAGTGTGTCCGTGACTGG
1850	TGAATGACCCTGTGTTATCCAGG
1851	TGGTATTAAAGGGCAATTACTGG
1852	GGCTACTCTCGATGTTTGGCTGG
1853	CGTACTCAGGTGCGCCGGTAGGG
1854	TACTTCTAACCTACTCTGTTAGG
1855	TGGAGCATAGTGGTATTAAAGGG
1856	CGCAGGCTACTCTCGATGTTTGG
1857	CGGGGCAAGGCTCAGCGTTCTGG

[00157] Table 88: Target sequences for PHEX gene

SEQ ID NOS	Target sequence
1858	TGGGTGTAAGTGGCTTCGAGTGG
1859	ATCGGTTGAAAGATTCTCCGCGG
1860	TATCTTGCATATGTTTCCGAGGG
1861	CCTGTCGGTAAGTGATGGGTAGG
1862	AGGGTCGTCGTCTCTTCAAGGGG
1863	TATATCGTTAGTGAAAGGCCTGG
1864	CTAAACCATCCATACAGATACGG
1865	AATTCCTGTCGGTAAGTGATGGG
1866	TTTATCTAACGATGAGCAGAAGG
1867	TTTCCGTGTTACTTTAAGTGTGG

[00158] Table 89: Target sequences for PIK3CA gene

SEQ ID NOS	Target sequence
1868	AGCAAGCACATCCACAGCGTAGG
1869	GTAAAGGGAGCGCAACAAGAGGG
1870	AACTGTACATAAACTTCGGGCGG

1871	CCCCGAGCGTGAGTAGAGCGCGG
1872	GTAAACACCAGACGTTTCAGCCGG
1873	AAGGTATAGGTACTCAGGAGAGG
1874	GGGTGTCATGTATAATACAGAGG
1875	GTGTCATGCATTCAAGTACCAGG
1876	CGATCACGAATCAGAAAACACGG
1877	CGAGTATTATGAGATTACCTGGG

[00159] Table 90: Target sequences for PKD1 gene

SEQ ID NOS	Target sequence
1878	GCTGCCGTCAGAAATCCCCGCGG
1879	CGGCAGAAAGTAATACTGAGCGG
1880	GACCGGGCATATCAGCATGGTGG
1881	ACGCAAACTCACGCCCGGGGGG
1882	CGGCGGTGTTAAGAGGGCAAAGG
1883	CCGATATCTACCCCTCCAAGTGG
1884	CACGCAAACTCACGCCCGGGGGG
1885	CCGAAGCACTGTCCGAGCAAGGG
1886	GGCAGCGAAGACACGTTGAGGGG
1887	GGGCGTACCGAGGTGAGCAGAGG

[00160] Table 91: Target sequences for PLP1 gene

SEQ ID NOS	Target sequence
1888	ACTTAAATCTAAATGCACCGGGG
1889	GTGCACACTATGAGGAATCGGGG
1890	CAATGGTGCTCATTTTCATGGGGG
1891	CGAATTGATTCATTAACCAGGGG
1892	GCACAGTTCGAGGTCCCAGAGGG
1893	GCACGATTGAGGATGCACATTGG
1894	TCCATAGATGACATACTGGAAGG
1895	GGTTATCCATGCTTTGAGTGAGG

1896	AACAAGGCTTCTTTGTCCGGGGG
1897	CGTAGAATCTGTGTAGACGAAGG

[00161] **Table 92: Target sequences for PMP22 gene**

SEQ ID NOS	Target sequence
1898	GCGCGTAAAGCTTCACACAGAGG
1899	CAGGATGTAGGCGAAACCGTAGG
1900	TGTCAGGAGCGAAATCATTGCGG
1901	TATAAATCCAGTATGCCGTGTGG
1902	CTTCTTTAAGGCTCAACACGAGG
1903	GCCAGGTTTTCCCAAAACGTGGG
1904	TCCGACCGTAAGAAAAATGTGGG
1905	ACACACAACAAAAGGTCGACGGG
1906	ACAGACAGCGTCCCCCACAAGG
1907	TGTCACACGATAAGGGAACCAGG

[00162] **Table 93: Target sequences for PMS2 gene**

SEQ ID NOS	Target sequence
1908	CTCCTGTGTCTACGGTGAGC
1909	ACTAGTAAAACTGGACCTT
1910	AAGGTCCAGTTTTTACTAGT
1911	TCTTTTGACGAGCATAGAT
1912	CTATGCTCGTCAAAAAGACG
1913	TCGTCAAAAAGACGTGGATG
1914	GTGGTGCATTGGTTGACTGT
1915	TTAGACTTCATTGACAAACC
1916	TGAGATATAAGCGTCCTACC
1917	GACGCTTATATCTCATGTCT
1918	AGGATCACTATTGCAGTTCA
1919	GGATCACTATTGCAGTTCAC
1920	ACAGTCAACCAATGCACCAC

1921	GAGACCCACCCCAGGGATAC
1922	AGGATGGTCAAAGTGCAACG
1923	CCAATAAAGAGAACGGGGAC
1924	GTCCTCAAGTTAGAGAAGTC

[00163] Table 94: Target sequences for PRSS1 gene

SEQ ID NOS	Target sequence
1925	TAGTAAGTTATGTGCTATATAGG
1926	GCCCTTCCCGCAAGGATGCTGG
1927	AACGCCCTGCAGGCTTGTTAAGG
1928	CGGGGTTGGCACATGACATATGG
1929	TGACCTTGCCCGACACTGACTGG
1930	CCATAAACTAATCGACAGTCAGG
1931	CACGGTTCCACGTGAGTACATGG
1932	GTATCTACAGTTGTTAGAGCAGG
1933	AGAGGCACGTCATCACCAACAGG
1934	TCTTCCTGTCGTATTGGGGGTGG
1935	GAGTCTTCCTGTCGTATTGGGGG
1936	GGCGTTGATTACTGCACGTGAGG
1937	AAGAGTCTTAGTGGCCCAGGTGG
1938	TAGGAGCTTAGTGCATCTGGAGG

[00164] Table 95: Target sequences for PTCH1 gene

SEQ ID NOS	Target sequence
1939	ATTTCAAAAGCGTCTCTGCGCGG
1940	TTGAAAGAGCACTAATGACGGGG
1941	GGAGGTCTATAATTACCAAGAGG
1942	CGAGGAGCTTCGGCACTACGAGG
1943	CCCATGTGACCAATTCGCTGTGG
1944	TAAGAGATGCCGTAGACACGAGG
1945	AGTGCCGAAGCTCCTCGCTGAGG

1946	GAAGCACGTACCCTAAACACTGG
1947	GTCCAATTATGCATCTCAAGGGG
1948	TATTACTGCTACCCAAGATGGGG

[00165] Table 96: Target sequences for PTEN gene

SEQ ID NOS	Target sequence
1949	GTAGTCCCGGAGTTAGGTAA
1950	CCAGGTTTAATTAGTAGTCC
1951	CCTATGGAAGAACGTATATG
1952	AGGTTAGACTAACCTTAAAT
1953	CCACATATACGTTCTTCCAT
1954	CATATACGTTCTTCCATAGG
1955	ATTTAAGTTGCCCCAACCAAC
1956	TAGCGAGAGCAAACTGTAG
1957	GGTTATAGCTACCAATACTC
1958	ATTGGTAGCTATAACCACTT
1959	TTGGTAGCTATAACCACTTT
1960	GGTATGAGTACTAATCTGGC
1961	GTATGAGTACTAATCTGGCT
1962	GGTGAAGTTATTGCAATCTA
1963	TTTTGGTATGAGTACTAATC
1964	TGGTGTGCTAGTTTTTACGT
1965	CCCGATTAATATTTAGCCAG
1966	CAATGGTTGGTACTAACAGG
1967	GAACAATGGTTGGTACTAAC
1968	ACGTGATATCTTTTGTAAAC
1969	AGTTTAAACCATAGACGCAA
1970	GGGAACATACTACCACTGTT
1971	CTTTGTAGGAGAGGTTTATC
1972	TCCTACAAAGAGCCTTGTTG
1973	CGGATACCATAGTGTTTCTT

1974	AGGGTTAGACTATCAGAACT
1975	GGGTTAGACTATCAGAACTG
1976	CCATTAAACTGAGTCACTTC
1977	CCTATTTCAACAACCCCTAC
1978	GGGATATTCCAACCTATGCA
1979	GATGAAATCGTAAGTCCTGT
1980	ATGAAATCGTAAGTCCTGTA
1981	CTATCACTCAATAACTCTTC
1982	GCCCTACCCACAACATAAAC
1983	AATTCATTTGTCATACGCTG
1984	TGGCACTTCTTAACCTCCTA
1985	GTAGTAGGTGTTTACTAAAC
1986	GCTCATATTACAACGTACAA

[00166] Table 97: Target sequences for REEP1 gene

SEQ ID NOS	Target sequence
1987	CATCTGGTCCAATCACCGTGAGG
1988	TCCCCATATAAGTCTCACAAGGG
1989	ATTGGCGTTTTCTGACGACGAGG
1990	GGCGTTTTCTGACGACGAGGAGG
1991	TGATCTGTGTATCCCATGGAAGG
1992	GTTGGCTCATCTCACTCACGTGG
1993	AGGCAGATTACTATAAAGGTGGG
1994	CACTTAACATCTAACACACCAGG
1995	CAGATGTTAATTAAGCTGGATGG
1996	GGTTTTAGAAGATTGCGAGTTGG

[00167] Table 98: Target sequences for RPGR gene

SEQ ID NOS	Target sequence
1997	TCGCTTGTCAGAGATCCCAGAGG
1998	ATATTGACCCTACGACAACAAGG

1999	AGGTTTCTCTCAGAACATCGTGG
2000	TGCCAACTCAGTAAACCGAAGGG
2001	AATGGCACCAAGTAACCAAGTGGG
2002	GGTAGCAACTAATAATGACCAGG
2003	GTTCTTAACGAGCAAACCAGAGG
2004	AATACAGGTATGATGCGTGATGG
2005	GGA CTCTATCAGCACGTATGCGG
2006	TAAACTAATTCGTACCAGAAAGG

[00168] Table 99: Target sequences for SBDS gene

SEQ ID NOS	Target sequence
2007	TGGCGAAAGTAAATACGCCAAGG
2008	GCTGTATCAAATGGTGACATGG
2009	GCGGTACCAGTGCGAATCATCGG
2010	CGGTACCAGTGCGAATCATCGGG
2011	ATCCTGGTGGTATCTTGTCGTGG
2012	TGCGAATCATCGGGCTATCCAGG
2013	CACTCGGTACGCCGCTAACGCGG

[00169] Table 100: Target sequences for SCN1A gene

SEQ ID NOS	Target sequence
2014	TCCCGATGCAACTCAGTTCATGG
2015	GACCCTAATAAAGTTAACCTGG
2016	AAACTTGTACCTATACTGTTGGG
2017	CATTTTGTACGCATCAATCTGG
2018	TGATATGTGTTGCATACCTCTGG
2019	ATGGTTGCCAAGTAATATCAGGG
2020	ACTCACTAAGCATAAGGTCTTGG
2021	TTCGCCACTCACTAAGCATAAGG
2022	GCTTAGTGAGTGGCGAAATTTGG
2023	ATCAGAAGTTATCCATTATAGG

2024	TGAACAATTGAATTGCTCCGTGG
2025	TTTATATAGTTCGAGTGTCTGGG
2026	GGTAGTATAAAAAGTCTGCAGGG
2027	GTCAGTCCATTGTACAAGGATGG
2028	CCAATTGGGAGCTCAAAGGTAGG
2029	TCCAGTGACATATCTACCCTAGG
2030	ACCTTCAATTCAGTTAGTGCAGG
2031	TGGGGGGTATGGCAACCACATGG
2032	TTTGTCACCCGGTCATAGGAAGG
2033	GTCACCCGGTCATAGGAAGGTGG

[00170] Table 101: Target sequences for SDHB gene

SEQ ID NOS	Target sequence
2034	GCGTCTCTGGGAAGAAACCGGGG
2035	TGAAATTTCCAGTCCCACGTGGG
2036	TTTGGTACAGGAACACACGTTGG
2037	GTGTGTATAATTAAGCACCCGGG
2038	GGCCGATCATGAAACTGGAAGGG
2039	CTCTCGGTGTGTGGTCATCGAGG
2040	GCCACTGCCAATCCTGACGGAGG
2041	GGTCCCCACAGGGTCAGTAAGGG
2042	CTTCCAATCCCGCGGCTGAGGGG
2043	CGAGTTAATCACATGACCATAGG

[00171] Table 102: Target sequences for SDHC gene

SEQ ID NOS	Target sequence
2044	TATTATCACTGGTCTCCCCGAGG
2045	ATACATTCACCACATCGCGGTGG
2046	TAGTATCTCACCTTGGAACGGGG
2047	GTGGTTCCATCAATATCCTGAGG
2048	AAACAACGCACTTCACAACGTGG

2049	ATTGTGGGGTCTAATCGAGGTGG
2050	ATTGTCTGACCAACGCTGGGGGG
2051	TGGTCCGCAAGGTCTTCTCGAGG
2052	GTGCGCTCCGTAGGGCTTCGGGG
2053	TTGATGGATATGTACGACAGTGG

[00172] Table 103: Target sequences for SDHD gene

SEQ ID NOS	Target sequence
2054	CTGAGCACTACCGGTCACCAGGG
2055	AAAACTCTGAATCGGTCGAGGGG
2056	TAGGTGGGTTAATAAGCTAGAGG
2057	GCGTTAGAACCATGTCCGAAGGG
2058	TAGCATTACTACAGTACCTGAGG
2059	TATTCCCAGCAGAACCACGAGGG
2060	GCTGGATCCAATAGTGACCTGGG
2061	GAGATTCCTTGAACATGCCAGGG
2062	GTTGAAAATCATTTAACCTGGG
2063	GCGATGGAGAGAACATACAATGG

[00173] Table 104: Target sequences for SHOX gene

SEQ ID NOS	Target sequence
2064	TTGGCAACGAAAACGTGTGGGG
2065	CCCAAGATCGTGCGTCCCCGGGG
2066	AATCAATAAACAGCGTCGGAGGG
2067	CGTCAATCAATAAACAGCGTCGG
2068	AACCCCTGCGCTACCCGCGGGG
2069	TTGCAGCTCCCGTCTCGCCAGGG
2070	ATTGGCAACGAAAACGTGTGGG
2071	CGGCGCATCTTCCTCCCCGGCGG
2072	GCTTTTCTCCGAGGCCGAGGGG
2073	CCAAATCACCTGGCCACACGGGG

[00174] Table 105: Target sequences for SLC6A4 gene

SEQ ID NOS	Target sequence
2074	GTAGTAAAAAGGGGCAAACGTGG
2075	CTACTGCACCCATAAATATGAGG
2076	TAAGGGGAGTTGCTTTACAGTGG
2077	GAACGTATTTGTGAACCGATAGG
2078	AGGAGCTCGTAGAATTGTCATGG
2079	TGAAAGTTCTGCCCCGAGAGGG
2080	CTTTCCAGCAACAGCACGAGCGG
2081	GTGCAGGCCACGAGACCCGAAGG
2082	CACGCTGCAAGGTAAGATGTTGG
2083	TGAGAGCGCTATAAAGGCAGCGG

[00175] Table 106: Target sequences for SLC6A5 gene

SEQ ID NOS	Target sequence
2084	CTTGCTTAACCTCCGCACTGCGG
2085	GTTTAGGCAGAAACACTCGTAGG
2086	GCTACCCCCATACAACCGAGTGG
2087	TACCTCTTCTGTACCCACCGAGG
2088	ATTCAGACCGAATGGCTGCGCGG
2089	CGGTCCGGTTGAGAAGATGTGGG
2090	ACGCGCGGCAGTCTCCACGCCGG
2091	CGAGTTGCTCTGGGTCCTAGAGG
2092	AGGCTTGAGTGCATAACCAGAGG
2093	GGGATCTGCGAAGAGCGGCGGGG

[00176] Table 107: Target sequences for SLC6A8 gene

SEQ ID NOS	Target sequence
2094	ACTCTCCAAGCACATTACAGGGG
2095	ATAGGTCTATGTGGTCCGGGTGG
2096	GGTCTGATCAGGTCTTGAAGGGG
2097	GATGAGGCGCTTCACCCCGTGG

2098	AGGCAAGCGAGTCCTCTACCCGG
2099	CGCGTCCAGGTCTCGCGCGGCGG
2100	GGTCATCCTGCAAACCTTCGGGG
2101	ACCGCAGCATTCTGGTCCGTAGG
2102	GCAGACAAACGAGGCGCCCAGGG
2103	CCGCAGCATTCTGGTCCGTAGGG

[00177] Table 108: Target sequences for SLC22A5 gene

SEQ ID NOS	Target sequence
2104	GCTAATCCCCAGTACCCAGGG
2105	TGGCTTGGGAACGCTTCACGAGG
2106	GCATCCAACCCCTAATCAGGAGG
2107	GGGCCATAGAGCATCGCCCAGGG
2108	GAGTTGTCAAGGGCGGTCAGTGG
2109	GTCCCTCTTATAAGATTAGGCGG
2110	GTATTATAGAAGGGTTTTCGGGG
2111	TGAGGTAAGGGATGTGCTCGGGG
2112	GTGGGTAAGTATCCCTGCAGTGG
2113	TTACATAGGGCGCACGACCAGGG

[00178] Table 109: Target sequences for SMAD4 gene

SEQ ID NOS	Target sequence
2114	CTTCGGGAAGAAACAGACGCTGG
2115	TCTTATAACCACCTACCACTAGG
2116	TAGTAGAATCATTACATGCGAGG
2117	GTCATACCAAAAGGCCACATTGG
2118	TGAGTGGCGAAGGCGTACGGTGG
2119	ATTCTCCCACGAGCTGCAAGCGG
2120	GTTTGAGGGAGTGGTCGCCGGGG
2121	GCAATTCAACCATGTGAGGGTGG
2122	TTAATGGGGTAAGCTAAGCCAGG

2123	TTTTGCACCGTAGTTTAAGGTGG
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[00179] Table 110: Target sequences for SMARCE1 gene

SEQ ID NOS	Target sequence
2124	GTAGTGCTATGGATTAAACGAGG
2125	GTGTAGGAATCATATCACCTGGG
2126	CAGAACCATGACGACCTTAGGGG
2127	AGCTATTGTCCCAGAATACGTGG
2128	GCATCGTTGCAAGAAGTGGGAGG
2129	AAGTAACTACTCTAACTATGGGG
2130	CAGTGAGGGCCATAGTTCGTTGG
2131	GTGCCTATACCACAAATCCCAGG
2132	GGTAGATTTAGGCATGGTGTAGG
2133	AGTCCTCTCCATATAGGCACAGG

[00180] Table 111: Target sequences for SMN1 gene

SEQ ID NOS	Target sequence
2134	CCGGGTGTAAGGGGGCCATTAGG
2135	TTCAAATAATGTCGGGGTGGTGG
2136	CTTCATATCACTGTACCTACTGG
2137	GCCGAGTTCCGGGTGTAAGGGGG
2138	ACACACTGGAGTTCGAGACGAGG
2139	GAAGGATGGCCAGCTCTTATTGG
2140	AAGGATGGCCAGCTCTTATTGGG
2141	TACATGAGTGGCTATCATACTGG
2142	GTTGTTGCGCAATAGATCTTCGG
2143	CATATCTTATACAGGTGACATGG
2144	TCATCTCGTTTTGATCAGTGGGG
2145	GGTGTAGATTAGTAATGAAGTGG
2146	GCATGGCAGCGCACTGTAAAGG
2147	GCAGTCCTGGTGGTCCGTTCTGG

2148	CACATCTATGATACGTGAATGGG
2149	TCATACACAATCTTGCTGTCTGG
2150	AAACCCGCGGGTGCGCAGCGTGG
2151	ACGAATCTGCCAAACTTAGTGG
2152	CTTCTCACGCTTTCTACGAGTGG
2153	GCGTTTGGAGCATATTGTGTAGG
2154	AGTTTCAAATAATGTCGGGGTGG
2155	CGCACGAAAAGTGGCCAGCACGG
2156	CGTGCTGGGCAGTTTTCGTGCGG
2157	TGCCGCACCCAGCTGTAACTGG
2158	CTATAGGGTAGAGTTGGATTGG
2159	CAGGAAACTTACCTGGTTAGAGG
2160	TTCCCTGGTCATATCTTGGTTGG
2161	ATCATCTCGTTTTGATCAGTGGG
2162	AAGTTGGTGTCTATGCCATAAGG
2163	ATATCTTATACAGGTGACATGGG
2164	GTGTAGATTAGTAATGAAGTGGG
2165	AGAGCTCAATTCATTAAGCGTGG
2166	ACATCGGTAGGCATATTTCAAGG
2167	GATTCGTGGTCATGAGTTGAAGG
2168	CGTCACTCTTAAGAAGGGACGGG
2169	GCTATGGCGATGAGCAGCGGCGG
2170	GAGCCCAAAGTCTCGAGGAAGG
2171	GATTCCGTGCTGTTCCGGCGCGG
2172	CCGCTATTCACAACAAATATGGG
2173	TCTACTCATGGTATGTGGATAGG
2174	TAGGCATTCCCAATAAGAGCTGG
2175	GATTGAAATGGGGCTCGATGTGG
2176	CAGAAGTAATGAAACCGTTGGGG
2177	CACGTTACTAAGAGCAACTCTGG
2178	AACCCGCGGGTGCGCAGCGTGGG

2179	GGCCGAGTTCGGGTGTAAGGGG
2180	GTTACTACAAGCGGTCCTCCCGG
2181	GTTTTCGTGCGGCTGTCTCGTGG
2182	CCCGCTATTCACAACAAATATGG
2183	GAAGCGTTATAGAAGATAACTGG
2184	CGTGAGCTTAGAGCATAGACTGG
2185	TAGGCCGAGTTCGGGTGTAAGG
2186	GAACTGCGATGTAAACATTAAGG
2187	TGTCTTTATATAGATCAAGCAGG
2188	CGATAGTTAGACAGAGTCCTCGG
2189	TCAGATAGATTCGATAACGGAGG
2190	CTTAAGGTTACATTCGCACTTGG
2191	ATAGCAATGTAGGGCCCCAACGG
2192	AATAAGGTATAAGCGGGCTCAGG
2193	AGGCCGAGTTCGGGTGTAAGGG
2194	CATCAAGTCGATCCGCTCACTGG
2195	CGATCCGCTCACTGGAGTTGTGG
2196	AGGTTACATTCGCACTTGGAAGG
2197	GGTTACATTCGCACTTGGAAGGG
2198	GTTGTCAGTTTGATCCACCGAGG

[00181] Table 112: Target sequences for SMN2 gene

SEQ ID NOS	Target sequence
2199	TCATCTCGTTTTGATCAGTGGGG
2200	GTTGTCAGTTTGATCCACCGAGG
2201	GATTCCGTGCTGTTCCGGCGCGG
2202	ATATCTTATACAGGTGACATGGG
2203	ACACACTGGAGTTCGAGACGAGG
2204	CAGAAGTAATGAAACCGTTGGGG
2205	CGTGCTGGGCAGTTTTCTGTGCGG
2206	ATAGCAATGTAGGGCCCCAACGG

2207	AGAGCTCAATTCATTAAGCGTGG
2208	TCAGATAGATTGATAACGGAGG
2209	CGCACGAAAACGCCCCAGCACGG
2210	AGTTTCAAATAATGTCGGGGTGG
2211	GTGTAGATTAGTAATGAAGTGGG
2212	GAGCCCAAACGCTCGAGGAAGG
2213	GCCGAGTTCGGGTGTAAGGGGG
2214	GGTTACATTCGCACTTGGAAGGG
2215	GGTGTAGATTAGTAATGAAGTGG
2216	GCGTTTGGAGCATATTGTGTAGG
2217	CGATAGTTAGACAGAGTCCTCGG
2218	GTTTTCGTGCGGCTGTCTCGTGG
2219	GATTGAAATGGGGCTCGATGTGG
2220	TGTCTTTATATAGATCAAGCAGG
2221	GCTATGGCGATGAGCAGCGGCGG
2222	CGATCCGCTCACTGGAGTTGTGG
2223	TAGGCATTCCCAATAAGAGCTGG
2224	GTTACTACAAGCGGTCCTCCCGG
2225	AGGTTACATTCGCACTTGGAAGG
2226	CATATCTTATACAGGTGACATGG
2227	CGTCACTCTTAAGAAGGGACGGG
2228	CATCAAGTCGATCCGCTCACTGG
2229	CTTCTCACGCTTTCTACGAGTGG
2230	CGTGAGCTTAGAGCATAGACTGG
2231	AAGTTGGTGTCTATGCCATAAGG
2232	CTTCATATCACTGTACCTACTGG
2233	GTTGTTGCGCAATAGATCTTCGG
2234	ATCATCTCGTTTTGATCAGTGGG
2235	GATTCGTGGTCATGAGTTGAAGG
2236	ACGAATCTGCCAAAACCTTAGTGG
2237	GGCCGAGTTCGGGTGTAAGGGG

2238	TACATGAGTGGCTATCATACTGG
2239	CTTAAGGTTACATTCGCACTTGG
2240	AAACCCGCGGGTGCGCAGCGTGG
2241	ACATCGGTAGGCATATTTCAAGG
2242	TCATACACAATCTTGCTGTCTGG
2243	AATAAGGTATAAGCGGGCTCAGG
2244	TTCAAATAATGTCGGGGTGGTGG
2245	TCTACTCATGGTATGTGGATAGG
2246	GAACTGCGATGTAAACATTAAGG
2247	CACATCTATGATACGTGAATGGG
2248	TTCCCTGGTCATATCTTGTTGG
2249	CAGGAACTTACCTGGTTAGAGG
2250	AGGCCGAGTTCCGGGTGTAAGGG
2251	CTATAGGGTAGAGTTGGATTTGG
2252	GAAGCGTTATAGAAGATAACTGG
2253	TAGGCCGAGTTCCGGGTGTAAGG
2254	CACGTTACTAAGAGCAACTCTGG
2255	CCGCTATTCACAACAAATATGGG
2256	TGCCGCACCCAGCTGTAAACTGG
2257	CCGGGTGTAAGGGGGCCATTAGG
2258	GCAGTCCTGGTGGTCCGTTCTGG
2259	AAGGATGGCCAGCTCTTATTGGG
2260	AACCCGCGGGTGCGCAGCGTGGG
2261	GCATGGCAGCGCACTGTAAAGG
2262	GAAGGATGGCCAGCTCTTATTGG
2263	CCCGCTATTCACAACAAATATGG

[00182] Table 113: Target sequences for STK11 gene

SEQ ID NOS	Target sequence
2264	GGACTCTTCTGTCAATTCG
2265	ACACCCAGGCCTATTTGTCG

2266	GGGCACAAACAGAGGCCTCG
2267	GCGAAAATCCTCTTTACCAT
2268	CAGATGCTGGAACCCCATAA
2269	TGCTTGGACCTATGGTAAAG
2270	TTGGCAGATGCTTGGACCTA
2271	GTAGGTCTTTACATCCCAGG
2272	GATACCTGGACGCTCCTAAG
2273	ATACCTGGACGCTCCTAAGG
2274	GTGATACCTGGACGCTCCTA
2275	TGATACCTGGACGCTCCTAA
2276	TCCACTCCTGGGACATGCCG
2277	GAGCGTCCAGGTATCACCCA
2278	GGAGCGTCCAGGTATCACCC
2279	AAGCCCAGGGCCACGTCGG
2280	CGGCTCCACGTCCACTGGG
2281	CACGTCGGTGGGATGGGAAT

[00183] Table 114: Target sequences for TGFBR1 gene

SEQ ID NOS	Target sequence
2282	TGGGTTTTTAGTGACACCTCAGG
2283	TTTCCAACCTGGATCGGGAAGGG
2284	TCACAACGATCAGGTAAATTAGG
2285	ACACTATCTTCACAACGATCAGG
2286	GTCATGGTTGCTGATGTTACAGG
2287	GTGTCAGCTTTACTATCTCCTGG
2288	CTGAAGTCCTAGCTTGTATCTGG
2289	TTTTATTCTAGGCCACCAAAGG
2290	GTAGTAGAAAGGTCCTAAACAGG
2291	GTAGGAGTCTAAACCAAATCAGG
2292	CATGTCTTAACCTTTCAGTCTGG
2293	TAAACCAAATCAGGTCCACCTGG

2294	GTCTTAACCTTTTCAGTCTGGAGG
2295	AGTTGCGTAGGTTTCACTCGTGG
2296	CCTTCCCCACTTATCACATCAGG
2297	AGTGACCTGATGTGATAAGTGGG
2298	TCAATAAGTCAGCTCCATGGTGG
2299	AGTGATACCTCTAACACATGGGG
2300	TAGGTTAAATTAGATTGTCGTGG
2301	ACTATGTTCTGATACTAAAGG
2302	AACACTGTAATAGGTCTCTCAGG
2303	GATGCTTCAGTGGTTACTCCAGG
2304	AAGAGTGTGCATTCTGTTCTGTGG
2305	AGTGTGCATTCTGTTCTGTGGTGG

[00184] Table 115: Target sequences for TGFBR2 gene

SEQ ID NOS	Target sequence
2306	CACCACTATCACTTCGTGATAGG
2307	TACCCCGTTTGCACATGAGAGGG
2308	TTCCATTGAGATCACAAGACAGG
2309	TTCCAACACCCATGCTATAATGG
2310	ACTACTTGTCATTATAGCATGG
2311	GTCCATTATAGCATGGGTGTTGG
2312	CATGGGTGTTGGAAGACTAGAGG
2313	ACAGTCCTAATCAAGCCCACTGG
2314	GGATTCCATAGCAAGTCTTCTGG
2315	TGAGATACAGGCCACATAACAGG
2316	TTGTTAGAAACCAAGCGCCTTGG
2317	TCCCAAATATGGTAGTACTCTGG
2318	TATACAACCTTATGCTGCTGAGGG
2319	ATAGAAATTCTTCTCCGTGCTGG
2320	AACCCAGACCTATAGTTAGTTGG
2321	TTTCCAACCTAATAGGTCTGG

2322	TCACTATTCTCACGTTTCTAAGG
2323	TTCCAACAACTATAGGTCTGGG
2324	CAATGCTAGTAAACATGCCTGGG
2325	TTGATAAATGGCCTGCAAGTTGG
2326	TCTCTGACAGTAGAATACCCAGG
2327	TCTAGTCAATTAAGTGGGAGG
2328	CGGGCACACTTAGAATAACGAGG
2329	CTTGCCATCCCCACGGACAGGG
2330	ACTGAGTGTTATCTAAGCTCAGG
2331	ACGGACAGGGAAGTCCATGCTGG
2332	GTTGTACTGAATTGTTACCTAGG
2333	ATGGAGTTCCTGTCCGTGGGGG
2334	AGCAACTTGACAATACACTAAGG
2335	ACGTGTCAGCTTCTATTCAAAGG
2336	GGGACAGCAATGGTATTCCTCGG
2337	GTGTTACTGTTCTACGAAAAGG
2338	ACGGGTAGTCTGAAAGGTGCTGG
2339	GAGGTCACGGGTAGTCTGAAAGG
2340	GTTACATGAGGTCTCATCCTAGG
2341	AGGTTGAAATACCCTGGTGCAGG

[00185] Table 116: Target sequences for VHL gene

SEQ ID NOS	Target sequence
2342	ACCATAGGTGGTACATAGTAGGG
2343	CACCATAGGTGGTACATAGTAGG
2344	TATTGAAGTGCAGTGAAGGCAGG
2345	TCAACACTTATCACCATAGGTGG
2346	TAGTAATTTACCTTGAAATGGG
2347	GGCCCCCTATGGACACCTCATGG
2348	ATTTACCTTGAAATGGGCTGGG
2349	CAGTACAAGGAACGAACAAGAGG

2350	CTCAGGCGATCTACTGACGTTGG
2351	GTATAAAAGCAGAAGTCAGCAGG
2352	CACCATGAGGTGTCCATAGGGGG
2353	TCAAGGTGAAATTACTACAGAGG
2354	TCTAGCCCATGCCCTCACTGTGG
2355	GCCAATGACTAGCAGAGCGTGGG
2356	ACTAGCAGAGCGTGGGACTGAGG

[00186] Table 117: Target sequences for WT1 gene

SEQ ID NOS	Target sequence
2357	AATCTTGTCTAACATTCCCGAGG
2358	GTTCCCAACTTACTCAACAAGGG
2359	TGGTATGGTTTCTCACCTTGGGG
2360	TTGATCGTCCTAACTGTACAGGG
2361	TGTAGCGAGGATCTACAGGGTGG
2362	GAATGCTACTAACACTGGTGGGG
2363	GTCCTGAGCTCATAATTCCGGTGG
2364	GTAGCGAGGATCTACAGGGTGGG
2365	TACTCCTTACAACTGCCCCGTAGG
2366	CTCCTTACAACTGCCCCGTAGGGG

SEQUENCING

[00187] Following target enrichment of the sample using the methods and systems described elsewhere, the highly fragmented gDNA samples can be sequenced to detect genomic variations. In some embodiments, short-read sequencing is used. In some embodiments, long-read sequencing. In some cases, the sample contains high fragmented RNA samples. In some case the sample contains full-length RNA transcripts.

[00188] In some embodiments, the long-read sequencing platform may be single molecule real time sequencing (SMRT) (e.g. Pacific Biosciences long-read sequencing technology), or a variation thereof. Single-molecule real-time sequencing (SMRT) is a parallelized single molecule DNA sequencing method. Single-molecule real-time sequencing utilizes a zero-mode waveguide (ZMW). A single DNA polymerase enzyme is affixed at the bottom of a ZMW with a single molecule of DNA as a template. The ZMW is a structure that creates an illuminated

observation volume that is small enough to observe only a single nucleotide of DNA being incorporated by DNA polymerase. Each of the four DNA bases is attached to one of four different fluorescent dyes. When a nucleotide is incorporated by the DNA polymerase, the fluorescent tag is cleaved off and diffuses out of the observation area of the ZMW where its fluorescence is no longer observable. A detector detects the fluorescent signal of the nucleotide incorporation, and the base call is made according to the corresponding fluorescence of the dye.

[00189] In other embodiments, the long-read sequencing platform may be nanopore sequencing (e.g. Oxford Nanopore long-read sequencing technology), or a variation thereof. Nanopore sequencing uses electrophoresis to transport an unknown sample through an orifice of about 10^{-9} meters in diameter. A nanopore system can contain an electrolytic solution; when a constant electric field is applied, an electric current can be observed in the system. The magnitude of the electric current density across a nanopore surface depends on the nanopore's dimensions and the composition of DNA or RNA molecule that is occupying the nanopore. Sequencing is made possible because, while traversing through the nanopore, samples cause characteristic changes in electric current density across nanopore surfaces. The total charge flowing through a nanopore channel is equal to the surface integral of electric current density flux across the nanopore unit normal surfaces between times t_1 and t_2 .

[00190] In some cases, long-read sequencing requires application of the sample. In other cases, long-read sequencing does not require application of the sample.

CLINICAL APPLICATIONS

[00191] The systems and methods described herein can be used in clinical settings to detect and diagnose genetic diseases or disorders. In some embodiments, the systems and methods described herein can be used in the detection, treatment and/or monitoring of hereditary breast cancer-related disorders by detecting genetic variations in relevant genes such as BRCA1, BRCA2, MLH1, MSH2, and STK11. In some embodiments, the systems and methods described herein can be used in the detection, treatment and/or monitoring of hereditary colon cancer-related disorders by detecting genetic variations in relevant genes such as MLH1, MSH2, EPCAM, SMAD4, and STK11. In some embodiments, the systems and methods described herein can be used in the detection, treatment and/or monitoring of hereditary neuroendocrine tumor disorders by detecting genetic variations in relevant genes such as SDHB, SHDC, SDHD, and VHL. In some embodiments, the systems and methods described herein can be used in the detection, treatment and/or monitoring of Cowden Syndrome by detecting genetic variations in relevant genes such as PTEN.

[00192] In some embodiments, the systems and methods described herein can be used can be used in the detection, treatment and/or monitoring of neuromuscular disorders such as Duchenne Muscular Dystrophy and Spinal Muscular Atrophy by detecting genetic variations in relevant genes such as DMD, SMN1, and SMN2.

[00193] In some embodiments, the systems and methods described herein can be used can be used in the detection, treatment and/or monitoring of Fragile X Syndrome by detecting genetic variations in relevant genes such as FMR1.

[00194] In some embodiments, the systems and methods described herein can be used can be used in the detection, treatment and/or monitoring of cardiovascular disorders such as aortic dysfunction and dilation, and cardiac ion channelopathies, by detecting genetic variations in relevant genes such as TGFBR1, TFRBR2, MYH11, COL3A1, KCNH2 and KCNQ1.

[00195] In some embodiments, the systems and methods described herein can be used can be used in the detection, treatment and/or monitoring of movement disorders such as Parkinson Disease, Hereditary Ataxia, and Dystonia 5, by detecting genetic variations in relevant genes such as SCNA, PARK2, PARK7, PINK1, SCA1 (ATXN1), SCA10 (ATXN10), SCA17 (TBP), SCA2 (ATXN2), SCA3 (MJD/ATXN3), SCA6 (CACNA1A), SCA7 (ATXN7), SCA8 (ATXN8OS) and GCH1.

[00196] In some embodiments, the systems and methods described herein can be used can be used in the detection, treatment and/or monitoring of renal disorders (e.g. Alport Syndrome and Polycystic Kidney Disease) by detecting genetic variations in relevant genes such as COL4A5, PKD1 and PKD2.

[00197] In some embodiments, the systems and methods described herein can be used can be used in the detection, treatment and/or monitoring of adrenal disorders (e.g. Congenital Adrenal Hyperplasia) by detecting genetic variations in relevant genes such as CYP21A2.

[00198] In some embodiments, the systems and methods described herein can be used can be used in the detection, treatment and/or monitoring of neurodevelopmental disorders (e.g. Rett Syndrome) by detecting genetic variations in relevant genes such as FOXG1, and MeCP2.

[00199] In some embodiments, the systems and methods described herein can be used can be used in the detection, treatment and/or monitoring of cerebrovascular disorders (e.g. Cerebral Cavernous Malformations) by detecting genetic variations in relevant genes such as KRIT1 and PDCD10.

[00200] In some embodiments, the systems and methods described herein can be used can be used in the detection, treatment and/or monitoring of neuro-oncology (e.g. Neurofibromatosis

Type 1 and Neurofibromatosis Type 2) by detecting genetic variations in relevant genes such as NF1 and NF2.

[00201] In some embodiments, the systems and methods described herein can be used can be used in the detection, treatment and/or monitoring of epilepsy (e.g. Unverricht-Lundborg disease) by detecting genetic variations in relevant genes such as CSTB.

[00202] In some embodiments, the systems and methods described herein can be used can be used in the detection, treatment and/or monitoring of peripheral neuropathy by detecting genetic variations in relevant genes such as GJB1 and PMP22.

[00203] Use systems and methods described herein with existing clinical sequencing methods. In some cases, a sample can be analyzed using short-read sequencing to detect SNVs and indels, and long-read sequencing to detect SVs.

KITS

[00204] In some embodiment, a kit is described herein. The kit may comprise a plurality of crRNA probes disclosed herein. Further, the kit may comprise a plurality of tracerRNA molecules. The kit may comprise reagents that can be used to performing dA tailing and adapter ligation. Moreover, the kit may comprise any buffer that can be used in performing needed experiments. The kit may comprise instructions for performing any experiments and procedures described herein.

COMPUTER SYSTEMS

[00205] The present disclosure provides computer systems that are programmed to implement methods of the disclosure. FIG. 5 shows an example computer system 501 that can be programmed or otherwise configured to, for example, process and/or analyze a metabolite, control addition of reagents to reaction mixtures, control partition generation, control of reagent addition to partitions, provide conditions sufficient to conduct reactions, obtain and process sequencing data, output sequencing results to a user, provide an interface for user input to control devices coupled to the computer processor. The computer system 501 can regulate various aspects of the present disclosure, such as, for example, regulating fluid flow, delivery of reagents, partition generation, modulate reactions conditions, etc. The computer system 501 can be an electronic device of a user or a computer system that is remotely located with respect to the electronic device. The electronic device can be a mobile electronic device.

[00206] The computer system 501 includes a central processing unit (CPU, also “processor” and “computer processor” herein) 505, which can be a single core or multi core processor, or a plurality of processors for parallel processing. The computer system 501 also includes memory or memory location 510 (e.g., random-access memory, read-only memory,

flash memory), electronic storage unit 515 (e.g., hard disk), communication interface 520 (e.g., network adapter) for communicating with one or more other systems, and peripheral devices 525, such as cache, other memory, data storage and/or electronic display adapters. The memory 510, storage unit 515, interface 520 and peripheral devices 525 are in communication with the CPU 505 through a communication bus (solid lines), such as a motherboard. The storage unit 515 can be a data storage unit (or data repository) for storing data. The computer system 501 can be operatively coupled to a computer network (“network”) 530 with the aid of the communication interface 520. The network 530 can be the Internet, an internet and/or extranet, or an intranet and/or extranet that is in communication with the Internet. The network 530 in some cases is a telecommunication and/or data network. The network 530 can include one or more computer servers, which can enable distributed computing, such as cloud computing. The network 530, in some cases with the aid of the computer system 501, can implement a peer-to-peer network, which may enable devices coupled to the computer system 501 to behave as a client or a server.

[00207] The CPU 505 can execute a sequence of machine-readable instructions, which can be embodied in a program or software. The instructions may be stored in a memory location, such as the memory 510. The instructions can be directed to the CPU 505, which can subsequently program or otherwise configure the CPU 505 to implement methods of the present disclosure. Examples of operations performed by the CPU 505 can include fetch, decode, execute, and writeback.

[00208] The CPU 505 can be part of a circuit, such as an integrated circuit. One or more other components of the system 501 can be included in the circuit. In some cases, the circuit is an application specific integrated circuit (ASIC).

[00209] The storage unit 515 can store files, such as drivers, libraries and saved programs. The storage unit 515 can store user data, e.g., user preferences and user programs. The computer system 501 in some cases can include one or more additional data storage units that are external to the computer system 501, such as located on a remote server that is in communication with the computer system 501 through an intranet or the Internet.

[00210] The computer system 501 can communicate with one or more remote computer systems through the network 530. For instance, the computer system 501 can communicate with a remote computer system of a user (e.g., operator). Examples of remote computer systems include personal computers (e.g., portable PC), slate or tablet PC's (e.g., Apple® iPad, Samsung® Galaxy Tab), telephones, Smart phones (e.g., Apple® iPhone, Android-enabled

device, Blackberry®), or personal digital assistants. The user can access the computer system 501 via the network 530.

[00211] Methods as described herein can be implemented by way of machine (e.g., computer processor) executable code stored on an electronic storage location of the computer system 501, such as, for example, on the memory 510 or electronic storage unit 515. The machine executable or machine readable code can be provided in the form of software. During use, the code can be executed by the processor 505. In some cases, the code can be retrieved from the storage unit 515 and stored on the memory 510 for ready access by the processor 505. In some situations, the electronic storage unit 515 can be precluded, and machine-executable instructions are stored on memory 510.

[00212] The code can be pre-compiled and configured for use with a machine having a processor adapted to execute the code, or can be compiled during runtime. The code can be supplied in a programming language that can be selected to enable the code to execute in a pre-compiled or as-compiled fashion.

[00213] Aspects of the systems and methods provided herein, such as the computer system 501, can be embodied in programming. Various aspects of the technology may be thought of as “products” or “articles of manufacture” typically in the form of machine (or processor) executable code and/or associated data that is carried on or embodied in a type of machine readable medium. Machine-executable code can be stored on an electronic storage unit, such as memory (e.g., read-only memory, random-access memory, flash memory) or a hard disk. “Storage” type media can include any or all of the tangible memory of the computers, processors or the like, or associated modules thereof, such as various semiconductor memories, tape drives, disk drives and the like, which may provide non-transitory storage at any time for the software programming. All or portions of the software may at times be communicated through the Internet or various other telecommunication networks. Such communications, for example, may enable loading of the software from one computer or processor into another, for example, from a management server or host computer into the computer platform of an application server. Thus, another type of media that may bear the software elements includes optical, electrical and electromagnetic waves, such as used across physical interfaces between local devices, through wired and optical landline networks and over various air-links. The physical elements that carry such waves, such as wired or wireless links, optical links or the like, also may be considered as media bearing the software. As used herein, unless restricted to non-transitory, tangible “storage” media, terms such as computer or machine “readable medium” refer to any medium that participates in providing instructions to a processor for execution.

[00214] Hence, a machine readable medium, such as computer-executable code, may take many forms, including but not limited to, a tangible storage medium, a carrier wave medium or physical transmission medium. Non-volatile storage media include, for example, optical or magnetic disks, such as any of the storage devices in any computer(s) or the like, such as may be used to implement the databases, etc. shown in the drawings. Volatile storage media include dynamic memory, such as main memory of such a computer platform. Tangible transmission media include coaxial cables; copper wire and fiber optics, including the wires that comprise a bus within a computer system. Carrier-wave transmission media may take the form of electric or electromagnetic signals, or acoustic or light waves such as those generated during radio frequency (RF) and infrared (IR) data communications. Common forms of computer-readable media therefore include for example: a floppy disk, a flexible disk, hard disk, magnetic tape, any other magnetic medium, a CD-ROM, DVD or DVD-ROM, any other optical medium, punch cards paper tape, any other physical storage medium with patterns of holes, a RAM, a ROM, a PROM and EPROM, a FLASH-EPROM, any other memory chip or cartridge, a carrier wave transporting data or instructions, cables or links transporting such a carrier wave, or any other medium from which a computer may read programming code and/or data. Many of these forms of computer readable media may be involved in carrying one or more sequences of one or more instructions to a processor for execution.

[00215] The computer system 501 can include or be in communication with an electronic display 535 that comprises a user interface (UI) 540 for providing, for example, monitoring of sample preparation, monitoring of reactions and/or reaction conditions, monitoring of sequencing, results of sequencing, and permitting user inputs for sample preparation, reactions, sequencing and/or sequencing analysis. Examples of UIs include, without limitation, a graphical user interface (GUI) and web-based user interface.

[00216] Methods and systems of the present disclosure can be implemented by way of one or more algorithms. An algorithm can be implemented by way of software upon execution by the central processing unit 505. The algorithm can, for example, implement sample preparation protocols, reaction protocols, sequencing protocols, data analysis protocols and system or device operation protocols.

[00217] Devices, systems, compositions and methods of the present disclosure may be used for various applications, such as, for example, processing a single analyte (e.g., RNA, DNA, or protein) or multiple analytes (e.g., DNA and RNA, DNA and protein, RNA and protein, or RNA, DNA and protein) from a cell. For example, a biological particle or analyte carrier (e.g., a cell or cell bead) is partitioned in a partition (e.g., droplet), and multiple analytes from the

biological particle or analyte carrier are processed for subsequent processing. The multiple analytes may be from the cell. This may enable, for example, simultaneous proteomic, transcriptomic and genomic analysis of the cell.

EXAMPLES

[00218] Example 1: Target enrichment protocol

[00219] An exemplary target enrichment protocol begins with preparing the Cas9 ribonucleoprotein complexes (RNPs). Prior to guide RNA assembly, all crRNAs are pooled into an equimolar mix, with a total concentration of 50-100 μ M. The crRNA mix and tracrRNA are then combined such that the tracrRNA concentration and the total crRNA concentration are both 5-10 μ M. The gRNA duplexes are formed by denaturation at 95 °C and then cooling to room temperature. Ribonucleoprotein complexes (RNPs) are constructed by combining the gRNA duplexes with Cas9 nucleases and then incubating at room temperature.

[00220] The next stage comprises dephosphorylating the genomic DNA. Between one to four genomic DNA samples can be pooled into the dephosphorylation reaction, for a total of 1-5 μ g of gDNA in each phosphorylation reaction. The input DNA is dephosphorylated using Calf Intestinal Phosphatase or Shrimp Alkaline Phosphatase.

[00221] The next stage comprises cleaving and dA-tailing target DNA. RNPs are added to the dephosphorylated gDNA along with dATP and Taq DNA polymerase. The sample is then incubated at 37 °C for Cas9 cleavage followed by 72 °C for dA-tailing. The reaction is then cleaned up using SPRI beads. Next is barcode ligation. Barcodes are ligated to the dA-tailed ends of the gDNA using ligase. The reaction is incubated at room temperature and then cleaned up using SPRI beads.

[00222] Next stage is sequencing adapter ligation and clean-up. All the barcoded DNA are pooled together at an equimolar amount. Sequencing adapters are ligated to the pool of barcoded DNA using ligase. The DNA is then cleaned up using SPRI beads, and then eluted in elution buffer.

[00223] The next stage is priming and loading the Flow Cell. Libraries were prepared for sequencing by adding the following to the eluate: Sequencing Buffer, Loading Beads, and Flush Tether. The sequencing libraries are then loaded onto the flow cell for sequencing.

[00224] Example 2: BRCA1 crRNA probe design

[00225] In the case of BRCA1, the CHOPCHOP design program yielded a total of 5567 possible crRNA probes along the entire length of the BRCA1 genomic locus. These crRNA sequences were then filtered using the filtering scheme described in **[0041]**, reducing the number to 233 crRNA probes. The crRNA sequences were then checked using a second design checker

tool, e.g. IDT CRISPR-Cas9 guide RNA design checker tool. The number of candidate crRNA probes was reduced to 86 probes. The final set of crRNA probes was chosen based upon the location of the target sites.

[00226] As shown in FIG. 4A, successful Cas9 cleavage and sequencing results in increased sequencing coverage of the target region with little or no sequencing coverage of non-target regions. In a sample with a known deletion of exons 15 and 16, a sharp drop in sequencing coverage is observed where the deletion occurs (FIG. 4B).

[00227] While preferred embodiments of the present invention have been shown and described herein, it will be obvious to those skilled in the art that such embodiments are provided by way of example only. It is not intended that the inventions be limited by the specific examples provided within the specification. While the invention has been described with reference to aforementioned specification, the descriptions and illustrations of the embodiments herein are not meant to be construed in a limiting sense. Numerous variations, changes, and substitutions will now occur to those skilled in the art without departing from the invention. Furthermore, it shall be understood that all aspects of the inventions are not limited to the specific depictions, configurations or relative proportions set forth herein which depend upon a variety of conditions and variables. It should be understood that various alternatives to the embodiments of the invention described herein may be employed in practicing the invention. It is therefore contemplated that the invention shall also cover any such alternatives, modifications, variations or equivalents. It is intended that the following claims define the scope of the invention and that methods and structures within the scope of these claims and their equivalents be covered thereby.

CLAIMS

WHAT IS CLAIMED IS:

1. A method for identifying a set of guide RNAs (gRNAs) that are hybridizable to a genomic region of interest in a genome comprising designing a plurality of gRNAs, wherein:
 - at least one gRNA is hybridizable to a target site within the genomic region of interest and is configured to produce a genomic variant that comprises at least 1000 bp; and
 - said plurality of gRNAs comprises a plurality of CRISPR RNAs (crRNAs), wherein said plurality of crRNAs comprises a GC of at least about 40% to about 80%.
2. A method for identifying a set of guide RNAs (gRNAs) that are hybridizable to a genomic region of interest in a genome comprising designing a plurality of gRNAs, wherein:
 - at least one gRNA is hybridizable to a target site within the genomic region of interest and is configured to produce a genomic variant that comprises at least 1000 bp; and
 - said plurality of gRNAs comprises a plurality of CRISPR RNAs (crRNAs), wherein said plurality of crRNAs comprises a self-complementarity score of zero.
3. A method for identifying a set of guide RNAs (gRNAs) that are hybridizable to a genomic region of interest in a genome comprising designing a plurality of gRNAs, wherein:
 - at least one gRNA is hybridizable to a target site within the genomic region of interest and is configured to produce a genomic variant that comprises at least 1000 bp; and
 - said plurality of gRNAs comprises a plurality of CRISPR RNAs (crRNAs), wherein said plurality of crRNAs comprises an efficiency score of about 0.2.
4. A method for identifying a set of guide RNAs (gRNAs) that are hybridizable to a genomic region of interest in a genome comprising designing a plurality of gRNAs, wherein:

at least one gRNA is hybridizable to a target site within the genomic region of interest and is configured to produce a genomic variant that comprises at least 1000 bp; and

said plurality of gRNAs comprises a plurality of CRISPR RNAs (crRNAs), wherein said plurality of crRNAs comprises a mismatch profile of MM0=0 or MM0=1, MM1=0 or MM1=1, MM2=0 or MM2=2, and MM3 <21.

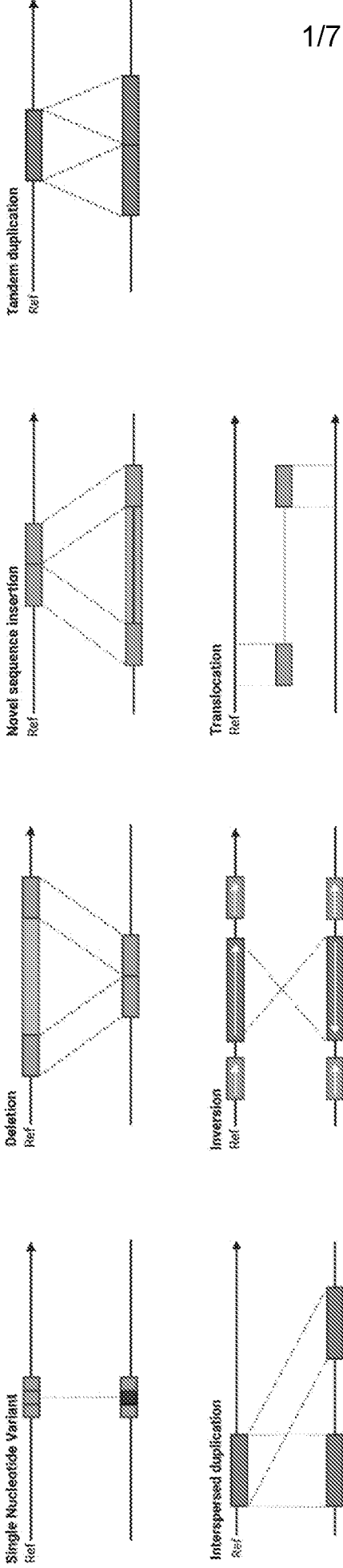
5. The method of claim 4, wherein said plurality of crRNAs comprises a mismatch profile of MM3<5.
6. A method of detecting a genomic variant in a sample, the method comprising:
 - (a) enriching said sample for a genomic region of interest comprising said genomic variant using a gene-editing based approach; and
 - (b) sequencing said enriched sample comprising said genomic region of interest using long-read sequencing.
7. The method of any of claims 1-6, wherein said genomic variant comprises a structural variant.
8. The method of claim 5, wherein said genomic variant comprises at least 50 bp.
9. The method of claim 5, wherein said genomic variant comprises at least 1000 bp.
10. The method of any of claims 1-5, wherein said gene-editing based approach comprises use of a clustered regularly interspersed short palindromic repeats (CRISPR)-Cas system.
11. The method of claim 9, wherein said CRISPR-Cas system comprises Cas 9.
12. The method of claim 5, wherein step (a) of enriching said sample further comprises an amplification of said genomic region of interest.
13. The method of claim 5, wherein step (a) of enriching said sample does not require an amplification of said genomic region of interest.
14. The method of claim 5, wherein step (a) of enriching said sample further comprises coupling a sequence of dAMPs to said genomic variant.
15. The method of claim 5, wherein step (a) of enriching said sample further comprises coupling a plurality of barcode molecules to said genomic variant.
16. The method of claim 5, wherein step (a) of enriching said sample further comprises coupling said genomic variant to a magnetic bead.
17. The method of any of claims 1-5, wherein said long-read sequencing comprises nanopore sequencing.

18. The method of any of claims 1-5, wherein said long-read sequencing comprises single molecule, real-time (SMRT) sequencing.
19. The method of any of claims 1-5, wherein said CRISPR-Cas system further comprises a crRNA that is hybridizable to a sequence listed in Tables 1-117.
20. The method of any of claims 1-5, wherein said genomic region of interest comprises two or more repeat regions.
21. The method of any of claims 1-5, wherein said sample comprises at least 10 genomic regions of interest.
22. The method of any of claims 1-5, wherein said genomic variant is associated with a disorder and drug response (pharmacogenomics).
23. The method of claim 22, wherein said disorder is selected from the group consisting of acute lymphoblastic leukemia (ALL), alpha-thalassemia, ataxia-telangiectasia (AT), autosomal recessive deafness 16, autosomal recessive deafness 22, beta-thalassemia, breast cancer, Canavan disease, cancer, celiac disease, chronic myeloid leukemia (CML), cystic fibrosis, cystinosis, deafness infertility syndrome (DIS), Duchenne muscular dystrophy, Ehlers-Danlos syndrome type III and IV, Ellis-van Creveld syndrome, Fabry disease, familial adenomatous polyposis (FAP), familial cutaneous melanoma, Fragile X, gastric cancer (including hereditary diffuse gastric cancer), Gaucher disease, hereditary predisposition to develop cancer, Huntington disease, hypophosphatasia (HPP), incontinentia pigmenti, Krabbe disease, Leber congenital amaurosis (LCA), Loeys-Dietz syndrome, Long QT syndrome, Lynch syndrome, Marfan syndrome, mental disorder, medium-chain acyl-coenzyme A dehydrogenase (MCAD) deficiency, MUTYH-associated polyposis, neuroblastoma, neuronal ceroid-lipofuscinoses (NCLs), Niemann-Pick Type C disease, pancreatic cancer syndromes, papillary renal carcinoma, Parkinson disease, phenylketonuria, Pompe disease, propionic acidemia, rheumatoid arthritis, solid tumors, spinal muscular atrophy, spinocerebellar ataxia, susceptibility to breast cancer, Tay-Sachs disease, very long-chain acyl-coenzyme A dehydrogenase deficiency, Von Hippel-Lindau syndrome, Wilms tumor, Wilson disease, Wolfram syndrome type 1, X-linked creatine deficiency syndrome, X-linked hemophilia A, and X-linked retinitis pigmentosa.
24. A method of designing a probe to target a genomic region of interest, the method comprising:

- (c) designing a plurality of nucleic acid probe options to target said genomic region of interest;
 - (d) selecting a first set of candidates from said plurality of nucleic acid probe options with a GC content of at least 20%;
 - (e) selecting a second set of candidates from said first set of candidates with a self-complementarity score of zero or a complementarity score of 1;
 - (f) selecting a third set of candidates from said second set of candidates with an efficiency greater than or equal to 0.2; and
 - (g) selecting a fourth set of candidates from said third set of candidates with a mismatch profile of MM0=0 or MM0=1, MM1=0 or MM1=1 or MM1=2, MM2=0 or MM2=1 or MM2=2, and MM3 <21, wherein said fourth set of candidates comprises said probe to target a genomic region of interest.
25. The method of claim 24, wherein fourth set of candidates comprises a mismatch profile of MM3<5.
26. The method of claim 24, wherein said designing comprises using CHOPCHOP.
27. The method of claim 24, wherein said first set of candidates have a GC content of about 40% to about 80%.
28. The method of claim 24, wherein said nucleic acid probe of interest comprises a crRNA.
29. The method of claim 27, wherein a probability of said crRNA cutting said genomic region of interest is greater than or equal to 80%.
30. The method of claim 21, further comprising estimating on-target value of said crRNA.
31. The method of claim 29, further comprising estimating off-target value of said crRNA.
32. A kit comprising a set of guide RNAs (gRNAs) that are hybridizable to a genomic region of interest in a genome comprising designing a plurality of gRNAs, wherein:
- at least one gRNA is hybridizable to a target site within the genomic region of interest and is configured to produce a genomic variant that comprises at least 1000 bp; and
 - said plurality of gRNAs comprises a plurality of CRISPR RNAs (crRNAs), wherein said plurality of crRNAs comprises a GC of at least about 40% to about 80%.

33. A kit comprising a set of guide RNAs (gRNAs) that are hybridizable to a genomic region of interest in a genome comprising designing a plurality of gRNAs, wherein:
- at least one gRNA is hybridizable to a target site within the genomic region of interest and is configured to produce a genomic variant that comprises at least 1000 bp; and
 - said plurality of gRNAs comprises a plurality of CRISPR RNAs (crRNAs), wherein said plurality of crRNAs comprises a self-complementarity score of zero.
34. A kit comprising a set of guide RNAs (gRNAs) that are hybridizable to a genomic region of interest in a genome comprising designing a plurality of gRNAs, wherein:
- at least one gRNA is hybridizable to a target site within the genomic region of interest and is configured to produce a genomic variant that comprises at least 1000 bp; and
 - said plurality of gRNAs comprises a plurality of CRISPR RNAs (crRNAs), wherein said plurality of crRNAs comprises an efficiency score of about 0.2.
35. A kit comprising a set of guide RNAs (gRNAs) that are hybridizable to a genomic region of interest in a genome comprising designing a plurality of gRNAs, wherein:
- at least one gRNA is hybridizable to a target site within the genomic region of interest and is configured to produce a genomic variant that comprises at least 1000 bp; and
 - said plurality of gRNAs comprises a plurality of CRISPR RNAs (crRNAs), wherein said plurality of crRNAs comprises a mismatch profile of MM0=0 or MM0=1, MM1=0 or MM1=1, MM2=0 or MM2=2, and MM3 <21.

Types of Variants



SNVs 1 bp	Indels < 50 bp	Structural Variants ≥ 50 bp
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FIG. 1

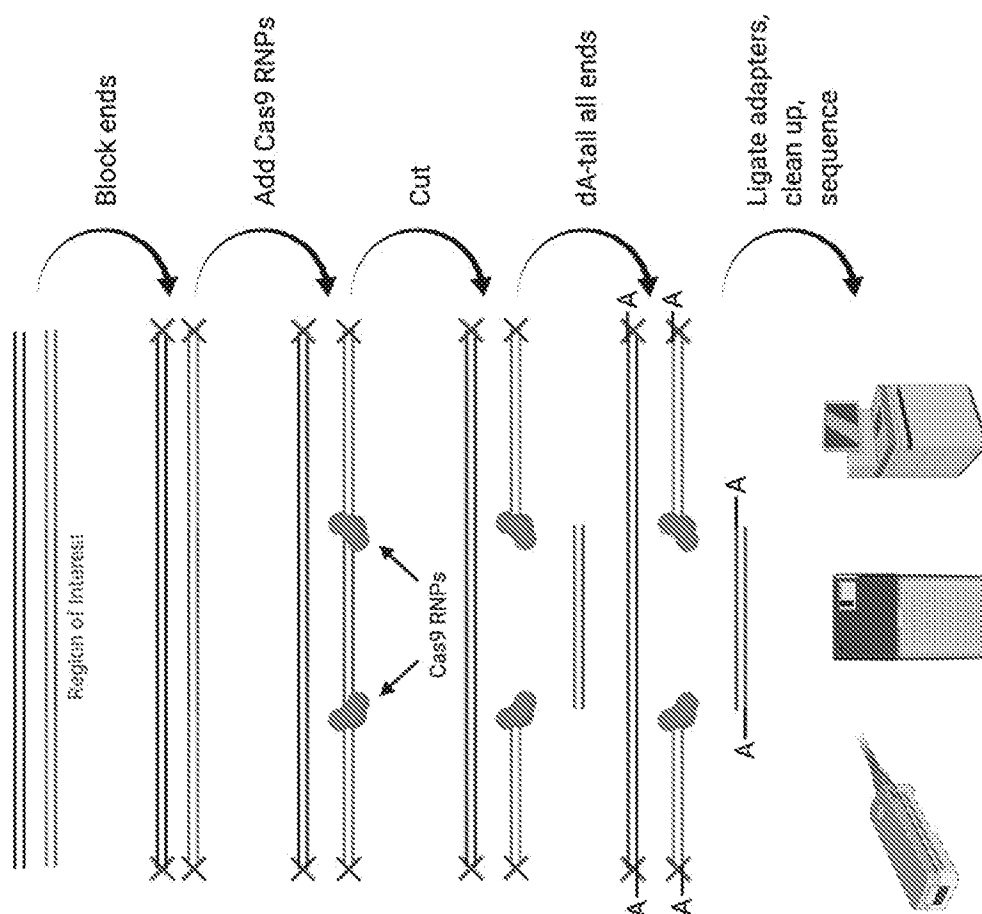


FIG. 2

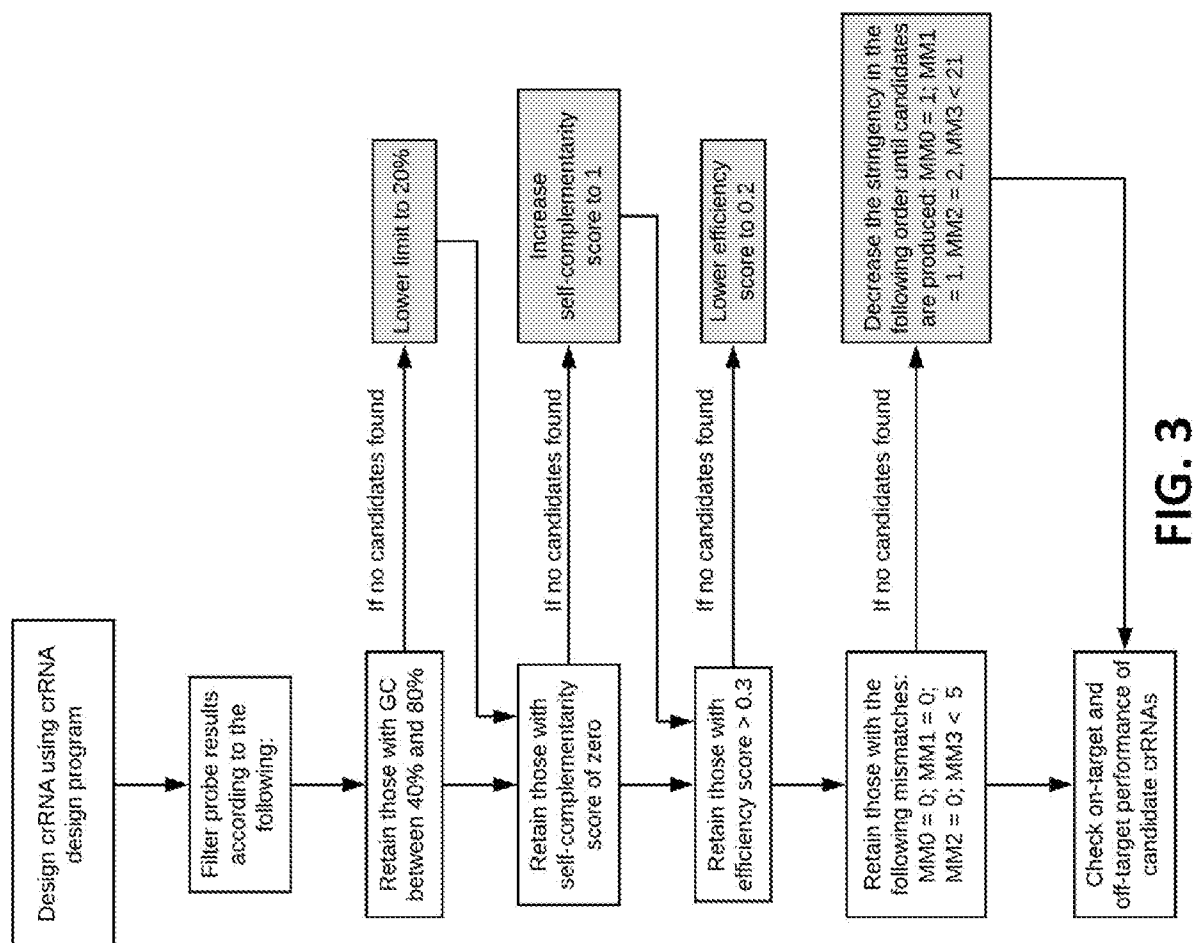


FIG. 3

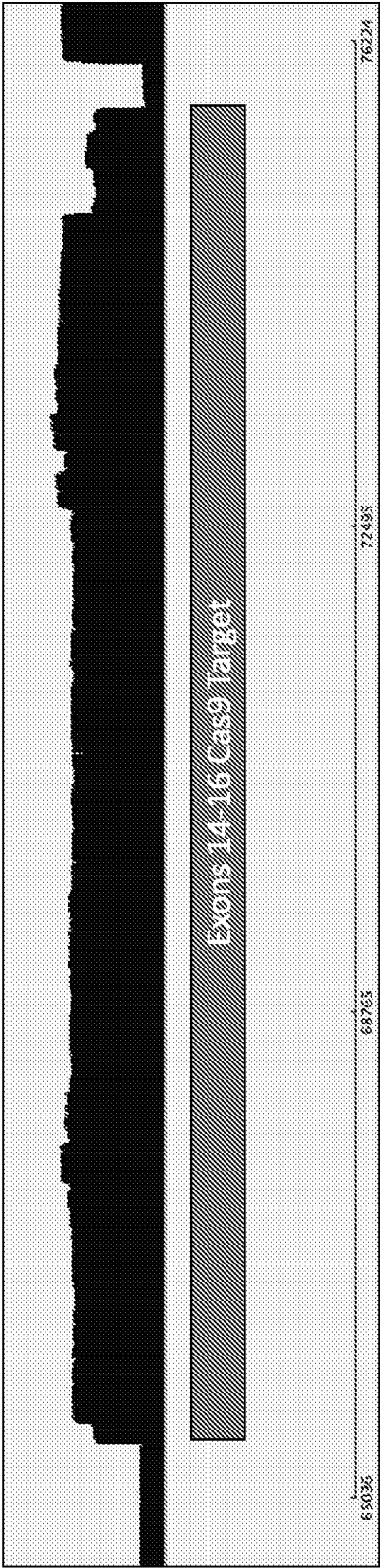
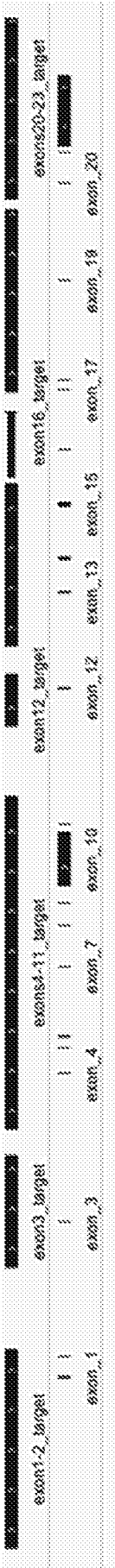


FIG. 4A

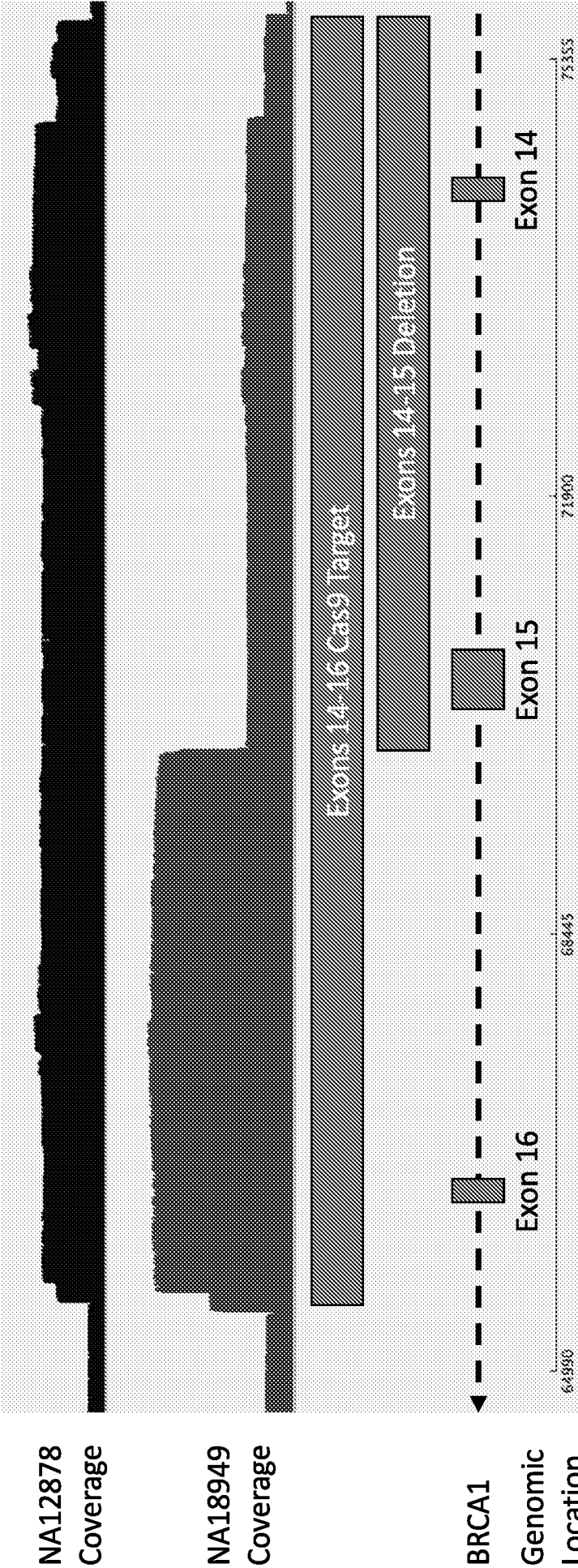


FIG. 4B

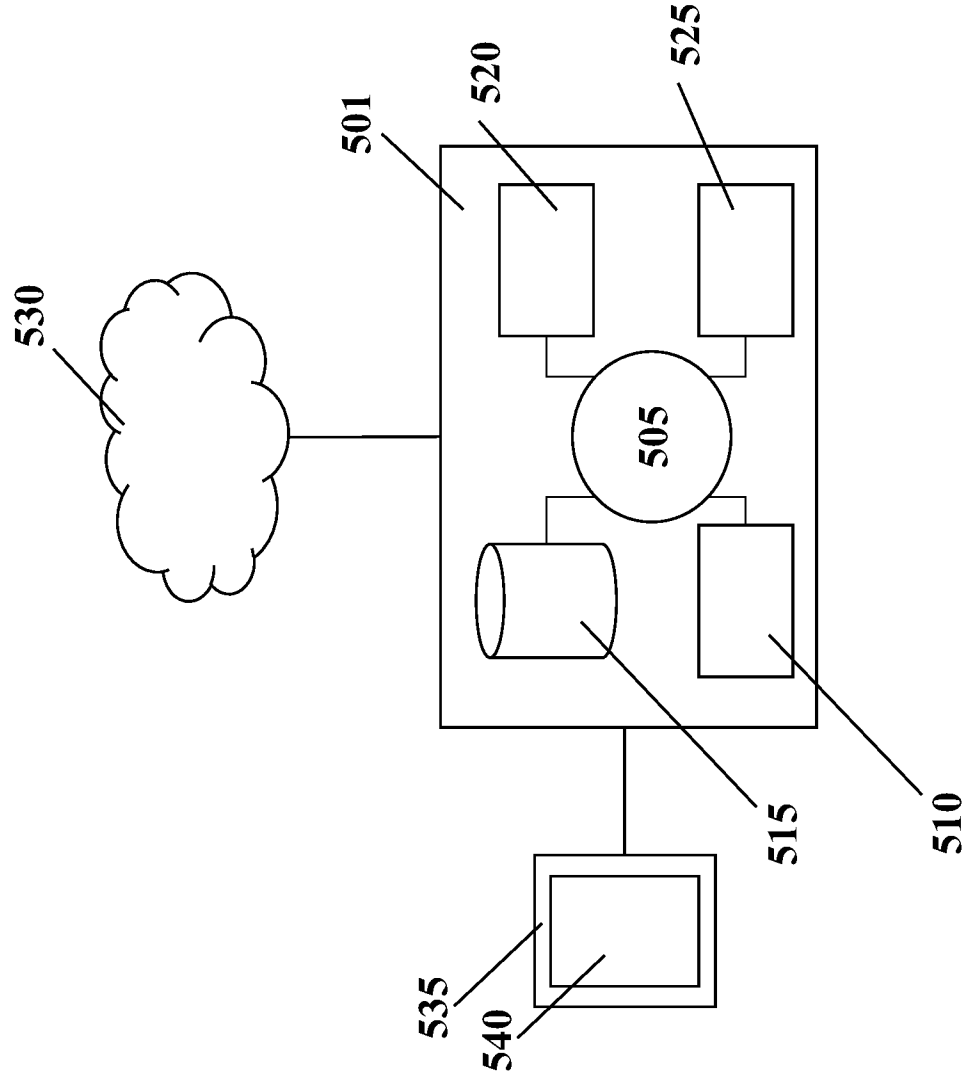


FIG. 5

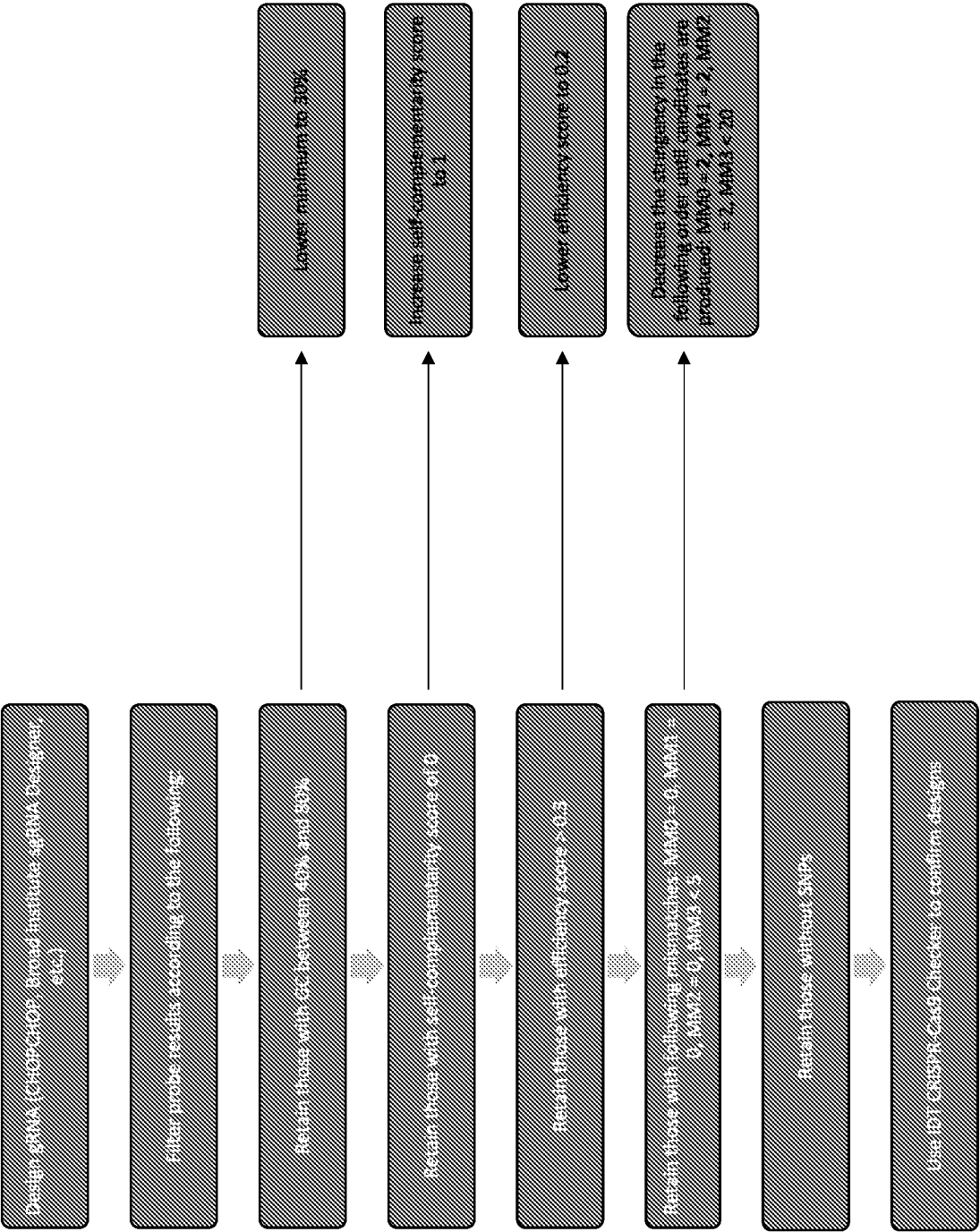


FIG. 6

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2020/055293

A. CLASSIFICATION OF SUBJECT MATTER

C12Q 1/6811 (2018.01) C12Q 1/6806 (2018.01) C12Q 1/6869 (2018.01)

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

PATENW, HCAPLUS, BIOSIS, EMBASE, MEDLINE: CRISPR, guide RNA, long read sequencing, Oxford nanopore sequencing, SMRT, PacBio and like terms; CPC/IPC: C12Q1/6806, C12Q1/6869.

PUBMED, ESPACENET, Internal Databases: Inventor and Applicant names.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
	Documents are listed in the continuation of Box C	



Further documents are listed in the continuation of Box C



See patent family annex

* Special categories of cited documents:	
"A" document defining the general state of the art which is not considered to be of particular relevance	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"D" document cited by the applicant in the international application	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier application or patent but published on or after the international filing date	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&" document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means	
"P" document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search
24 November 2020Date of mailing of the international search report
24 November 2020

Name and mailing address of the ISA/AU

AUSTRALIAN PATENT OFFICE
PO BOX 200, WODEN ACT 2606, AUSTRALIA
Email address: pct@ipaaustralia.gov.au

Authorised officer

Richard Filmer
AUSTRALIAN PATENT OFFICE
(ISO 9001 Quality Certified Service)
Telephone No. +61262832735

Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
the subject matter listed in Rule 39 on which, under Article 17(2)(a)(i), an international search is not required to be carried out, including
2. ☒ Claims Nos.: 2-5, 24-31 and 33-35
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
See Supplemental Box
3. ☐ Claims Nos:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a)

Box No. III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

See Supplemental Box for Details

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☒ As all searchable claims could be searched without effort justifying additional fees, this Authority did not invite payment of additional fees.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee.
- ☐ The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation.
- ☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT		International application No.
C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		PCT/US2020/055293
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	LIU, X. et al., "Sequence features associated with the cleavage efficiency of CRISPR/Cas9 system", Scientific Reports, 2016, vol. 6, no. 19675 Abstract; Results: Logistic Regression Analysis; Discussion	1, 32
X	CHOPCHOP: Instructions [retrieved from Internet on 13 November 2020] <URL: https://web.archive.org/web/20190811181724/https://chopchop.cbu.uib.no/instructions > published 11 August 2019 as per Wayback Machine 7. Interpreting the Results	1, 32
X	GILPATRICK, T. et al., "Targeted Nanopore Sequencing with Cas9 for studies of methylation, structural variants, and mutations", bioRxiv, 4 June 2019, doi: https://doi.org/10.1101/604173 Abstract; Methods, Discussion; Fig. 1A	6-17, 19-23
X	WO 2016/028887 A1 (PACIFIC BIOSCIENCES OF CALIFORNIA, INC.) 25 February 2016 [0006]-[0009], [0032], [0038], [0065], [0072], [0111], [0124]	6-23
A	LUBUN, K. et al., "CHOPCHOP v3: expanding the CRISPR web toolbox beyond genome editing", Nucleic Acids research, 20 May 2019, vol. 47, W171-W174. Improvements in the New Release	

Supplemental Box**Continuation of Box II**

The claims recite score and profile values that are so inadequately defined that the scope of the claims cannot be determined.

Continuation of: Box III

This International Application does not comply with the requirements of unity of invention because it does not relate to one invention or to a group of inventions so linked as to form a single general inventive concept.

This Authority has found that there are different inventions based on the following features that separate the claims into distinct groups:

- Invention 1 is defined by claims 1 and 32 (completely) and claims 24-31 (in part). The feature of a method for designing a plurality of gRNAs having crRNAs comprising a defined GC percentage is specific to this group of claims.
- Invention 2 is defined by claims 2 and 33 (completely) and claims 24-31 (in part). The feature of a method for designing a plurality of gRNAs having crRNAs comprising a certain self-complementarity score is specific to this group of claims.
- Invention 3 is defined by claims 3 and 34 (completely) and claims 24-31 (in part). The feature of a method for designing a plurality of gRNAs having crRNAs comprising a certain efficiency score is specific to this group of claims.
- Invention 4 is defined by claims 4, 5 and 35 (completely) and claims 24-31 (in part). The feature of a method for designing a plurality of gRNAs having crRNAs comprising a certain mismatch profile is specific to this group of claims.
- Invention 5 is defined by claims 6-23 (completely). A method for detecting a genomic variant in a sample comprising the step of enriching a sample for a genomic region using a gene-editing approach and a step of sequencing the genomic region using long read sequencing, is specific to this group of claims.

PCT Rule 13.2, first sentence, states that unity of invention is only fulfilled when there is a technical relationship among the claimed inventions involving one or more of the same or corresponding special technical features. PCT Rule 13.2, second sentence, defines a special technical feature as a feature which makes a contribution over the prior art.

When there is no special technical feature common to all the claimed inventions there is no unity of invention.

In the above groups of claims, the identified features may have the potential to make a contribution over the prior art but are not common to all the claimed inventions and therefore cannot provide the required technical relationship. The only feature common to all of the claimed inventions and which provides a technical relationship among them is a guide RNA. The only feature common to Inventions 1-4 is a method for identifying a guide RNA comprising certain properties. However this feature does not make a contribution over the prior art because it is disclosed in:

LUBUN, K. et al., "CHOPCHOP v3: expanding the CRISPR web toolbox beyond genome editing", Nucleic Acids Research, 20 May 2019, vol. 47, W171-W174.

This document discloses a method for designing a guide RNA capable of producing a genomic variant having a certain GC content, self-complementarity, or efficiency, and a method for designing guide RNAs suitable for enriching nucleic acids for sequencing with Oxford Nanopore Technologies (Improvements in the New Release).

Therefore in the light of this document these common features cannot be a special technical feature. Therefore there is no special technical feature common to all the claimed inventions and the requirements for unity of invention are consequently not satisfied *a posteriori*.

INTERNATIONAL SEARCH REPORT Information on patent family members		International application No. PCT/US2020/055293	
This Annex lists known patent family members relating to the patent documents cited in the above-mentioned international search report. The Australian Patent Office is in no way liable for these particulars which are merely given for the purpose of information.			
Patent Document/s Cited in Search Report		Patent Family Member/s	
Publication Number	Publication Date	Publication Number	Publication Date
WO 2016/028887 A125 February 2016 WO 2016028887 A125 Feb 2016 EP 3183367 A128 Jun 2017 EP 3183367 B126 Jun 2019 EP 3633047 A108 Apr 2020 US 2016208241 A121 Jul 2016 US 10435685 B208 Oct 2019 US 2020109396 A109 Apr 2020			
End of Annex			
Due to data integration issues this family listing may not include 10 digit Australian applications filed since May 2001. Form PCT/ISA/210 (Family Annex)(July 2019)			