



(51) International Patent Classification:

G01N 33/574 (2006.01)

(21) International Application Number:

PCT/US2014/022037

(22) International Filing Date:

7 March 2014 (07.03.2014)

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data:

61/775,316

8 March 2013 (08.03.2013)

US

(71) Applicant: MAYO FOUNDATION FOR MEDICAL EDUCATION AND RESEARCH [US/US]; 200 First Street S.W., Rochester, Minnesota 55905 (US).

(72) Inventors; and

(71) Applicants : KOSARI, Farhad [US/US]; P.O. Box 692, Ellsworth, Wisconsin 54011-0692 (US). VASMATZIS, George [GR/US]; 945 Cedar Point Lane S.E., Oronoco, Minnesota 55960 (US). AUBRY, Marie-Christine [CA/US]; 25260 525th Street, Pine Island, Minnesota 55963 (US). IDA, Cristiane M. [BR/US]; 508 4th Street S.W., Apt. 201, Rochester, Minnesota 55902 (US).

(74) Agent: FINN III, J. Patrick; Fish & Richardson P.C., P.O. Box 1022, Minneapolis, Minnesota 55440-1022 (US).

(81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BN, BR, BW, BY, BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IR, IS, JP, KE, KG, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LT, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PA, PE, PG, PH, PL, PT, QA, RO, RS, RU, RW, SA, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

(84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LR, LS, MW, MZ, NA, RW, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, RU, TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, KM, ML, MR, NE, SN, TD, TG).

[Continued on next page]

(54) Title: METHODS AND MATERIALS FOR IDENTIFYING AND TREATING MAMMALS HAVING LUNG ADENOCARCINOMA CHARACTERIZED BY NEUROENDOCRINE DIFFERENTIATION

(57) Abstract: This document provides methods and materials involved in identifying mammals having lung adenocarcinoma characterized by neuroendocrine differentiation as well as methods and materials involved in treating mammals having lung adenocarcinoma characterized by neuroendocrine differentiation. For example, methods and materials for using ASCL1 and RET expression levels to identify lung cancer patients having lung adenocarcinoma characterized by neuroendocrine differentiation are provided.

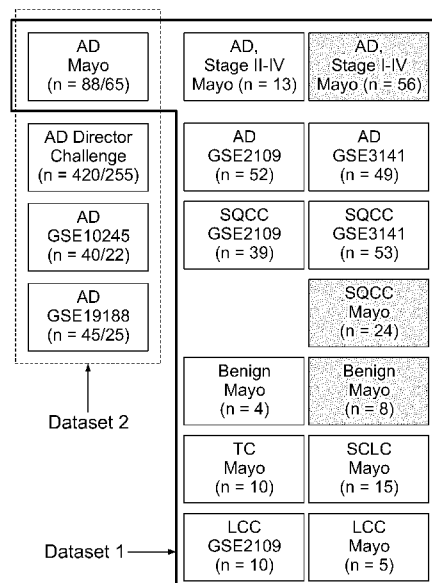


FIG. 1



Published:

— with international search report (Art. 21(3))

— with sequence listing part of description (Rule 5.2(a))

**METHODS AND MATERIALS FOR IDENTIFYING AND TREATING
MAMMALS HAVING LUNG ADENOCARCINOMA CHARACTERIZED BY
NEUROENDOCRINE DIFFERENTIATION**

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims the benefit of U.S. Provisional Application Serial No. 61/775,316, filed March 8, 2013. The disclosure of the prior application is considered part of (and is incorporated by reference in) the disclosure of this application.

BACKGROUND

1. Technical Field

This document relates to methods and materials involved in identifying mammals having lung adenocarcinoma characterized by neuroendocrine differentiation as well as methods and materials involved in treating mammals having lung adenocarcinoma characterized by neuroendocrine differentiation. For example, this document provides methods and materials for using achaete-scute homolog 1 (ASCL1) and RET expression levels to identify lung cancer patients having lung adenocarcinoma characterized by neuroendocrine differentiation.

2. Background Information

The clinical significance of neuroendocrine (NE) differentiation in lung adenocarcinoma, and the most appropriate biomarkers for this assessment, has been long debated. In the absence of a gold standard, investigators have most commonly used immunohistochemistry (IHC) of one or a combination of neuroendocrine markers, such as chromogranin (CHGA), synaptophysin (SYP), neuron-specific enolase (NSE), or neural cell adhesion molecule (CD56/NCAM) to assess the role of neuroendocrine differentiation in lung cancer survival.

SUMMARY

This document provides methods and materials involved in identifying mammals having lung adenocarcinoma characterized by neuroendocrine differentiation as well as

methods and materials involved in treating mammals having lung adenocarcinoma characterized by neuroendocrine differentiation. For example, this document provides methods and materials for using ASCL1 and RET expression levels to identify lung cancer patients having lung adenocarcinoma characterized by neuroendocrine differentiation. As described herein, the presence of an elevated level of ASCL1 expression and an elevated level of RET within a lung cancer sample can indicate that a mammal (e.g., a human) has lung adenocarcinoma characterized by neuroendocrine differentiation. In some cases, the absence of an elevated level of ASCL1 expression and an elevated level of RET within a lung cancer sample can indicate that a mammal (e.g., a human) does not have lung adenocarcinoma characterized by neuroendocrine differentiation.

Having the ability to identify mammals as having lung adenocarcinoma characterized by neuroendocrine differentiation as described herein can allow those lung cancer patients to be properly identified and treated in an effective and reliable manner. For example, the lung cancer treatments provided herein can be used to treat lung cancer patients identified as having lung adenocarcinoma characterized by neuroendocrine differentiation.

In general, one aspect of this document features a method for identifying a mammal as having lung adenocarcinoma characterized by neuroendocrine differentiation. The method comprises, or consist essentially of, determining whether or not cancer cells from the mammal contain an elevated level of ASCL1 expression and an elevated level of RET expression, wherein the presence of the elevated level of ASCL1 expression and the presence of the elevated level of RET expression indicates that the mammal has lung adenocarcinoma characterized by neuroendocrine differentiation, and wherein the absence of the elevated level of ASCL1 expression and the absence of the elevated level of RET expression indicates that the mammal does not have lung adenocarcinoma characterized by neuroendocrine differentiation. The mammal can be a human. The elevated level can be determined using PCR. The elevated level can be determined using immunohistochemistry.

In another aspect, this document features a method for identifying a mammal as having lung adenocarcinoma characterized by neuroendocrine differentiation. The

method comprises, or consists essentially of, (a) determining whether or not a lung cancer cells from the mammal contain an elevated level of ASCL1 expression and an elevated level of RET expression, (b) classifying the mammal as having lung adenocarcinoma characterized by neuroendocrine differentiation if the sample contains the elevated level of ASCL1 expression and the elevated level of RET expression, and (c) classifying the mammal as not having lung adenocarcinoma characterized by neuroendocrine differentiation if the sample lacks the elevated level of ASCL1 expression and the elevated level of RET expression. The mammal can be a human. The elevated level can be determined using PCR. The elevated level can be determined using immunohistochemistry.

In another aspect, this document features a method for identifying a mammal as having lung adenocarcinoma characterized by neuroendocrine differentiation, wherein the method comprises, or consists essentially of, (a) detecting the presence of an elevated level of ASCL1 expression and an elevated level of RET expression in lung cancer cells from the mammal, and (b) classifying the mammal as having lung adenocarcinoma characterized by neuroendocrine differentiation based at least in part on the presence of the elevated level of ASCL1 expression and the elevated level of RET expression. The mammal can be a human. The elevated level can be detecting using PCR. The elevated level can be detecting using immunohistochemistry.

In another aspect, this document features a method for treating lung cancer, wherein the method comprises, or consists essentially of, (a) detecting the presence of an elevated level of ASCL1 expression and an elevated level of RET expression in lung cancer cells from a mammal, and (b) administering a molecule to the mammal under conditions wherein the number of lung cancer cells within the mammal is reduced, wherein the molecule is selected from the group consisting of sunitinib, vandetanib, riluzole, alteplase, anistreplase, tenecteplase, sucalfate, dasatinib, pazopanib, tivozanib, OSI-930, telatinib, tandutinib, imatinib, sorafenib, levodopa, carbidopa, entacapone orion, L-dopa, ABT-089, mecamlamine, and succinylcholine.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention pertains. Although methods and materials similar or equivalent to those

described herein can be used to practice the invention, suitable methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

The details of one or more embodiments of the invention are set forth in the accompanying drawings and the description below. Other features, objects, and advantages of the invention will be apparent from the description and drawings, and from the claims.

DESCRIPTION OF THE DRAWINGS

Figure 1 is a schematic showing the composition of Datasets 1 and 2. Boxes with dotted shadings in Dataset 1 denote samples collected by laser capture microdissection (LCM). Numbers in parenthesis in Dataset 2 are all stages and stage 1 only sample sizes, respectively. Dataset 2 was used in survival analysis by RET. All samples in this dataset were collected in bulk.

Figure 2 is a graph plotting the threshold selection for ASCL1 positive and negative status. With minor exceptions, signal intensities at or above 8 produced a positive staining on the IHC. There was a strong Pearson's correlation ($\rho = 0.89$) between the Labeling index (LI) and microarray signal intensities (Log_2).

Figure 3 contains photographs and a heat graph of an immunohistochemical analysis. (A-E) AD characterized by ASCL1 mRNA expression (ASCL1^+ AD): (A) Adenocarcinoma with an acinar pattern. H&E. 400x; (B) ASCL1 protein expression (nuclear pattern, 70%, 2+). 400x; (C) CHGA (cytoplasmic pattern, 5%, 3+). 400x; (D) SYP (cytoplasmic pattern, 5%, 2+) 400x; and (E) CD56/NCAM (membranous pattern, 5%, 2+) 400x. (F-J) SCLC: (F) High-grade tumor characterized by extensive areas of necrosis and cells with high nuclear to cytoplasmic ratio, delicate nuclear chromatin and inconspicuous nucleoli. H&E 400x; (G) ASCL1 protein expression (nuclear pattern, 95%, 3+) 400x; (H) CHGA (cytoplasmic pattern, 100%, 2+) 400x; (I) SYP (cytoplasmic pattern, 100%, 1+) 400x; and (J) CD56/NCAM (membranous pattern, 100%, 2+) 400x. (K-O) LCNEC: (K) Poorly differentiated tumor with high mitotic activity (>10 mitotic

figures/ 2mm²) and organoid nesting composed by cells with vesicular nuclei, evident nucleoli and moderate amount of cytoplasm. H&E, 400x; (L) ASCL1 protein expression (nuclear pattern, 95%, 3+) 400x; (M) CHGA (cytoplasmic pattern, 90%, 3+) 400x; (N) SYP (cytoplasmic pattern, 95%, 2+) 400x; and (O) CD56/NCAM (membranous pattern, 90%, 3+) 400x. (P) Heat map of IHC protein expression of ASCL1, CHGA, SYP, and CD56/NCAM.

Figure 4 is a heat map of the microarray Log₂ expression values using known NE genes and RET. The map also includes SQCC and AD specific genes (DSG3 and NKX2.1, respectively). ASCL1 expression is much more frequent in AD than in SQCC.

Each gene is represented by the most variable probeset with the highest standard deviation.

Figure 5 contains graphs demonstrating that NE differentiation in never smoker AD is rare. In contrast with typical carcinoid (CT), no subset of AD has a distinguishably higher expression of the NE markers than non-neoplastic lung. Similar observations can be made about SQCC, but the number of samples is small.

Figure 6 is a graph of a KM plot of stage I AD in Dataset 2 based on ASCL1 status. The drop off for the ASCL1⁺ tumors is sharper than the ASCL1⁻ tumors, suggesting nonproportional hazard.

Figures 7A and 7B are graphs demonstrating the over representation of tumors expressing RET (211421_s_at probe set) in ASCL1⁺ compared with ASCL1⁻ in (A) stage I ADs (Dataset 2), and in (B) all lung cancers (Dataset 1). This over-representation was consistent in all datasets. Few samples (shown in black circles) expressed RET while ASCL1 was below noise level (arbitrary Log₂ signal intensity of 3.5). Data points correspond to the samples from the Mayo Clinic unless stated in the figure legends.

Figures 8A and 8B are graphs plotting overall survival in ASCL1⁺ stage I (A) and all (B) AD as a function of the RET mRNA expression level by microarrays.

Figure 9 contains photographs and graphs of RET protein expression by IHC. RET staining in fatal adenocarcinoma was typically much less intense in ASCL1⁻ (A) than in ASCL1⁺ (B) tumors. (C) Co-IHC of ASCL1 (nuclear brown staining) and RET (cytoplasmic red staining) identified areas with overlapping expression of the two proteins. (D) KM plot of 14 ASCL1⁺ AD samples indicate a significant association with

OS ($p = 0.05$). (E) When samples were not stratified by the ASCL1 expression, RET IHC was not significant in predicting OS (overall survival).

Figure 10 is a Kaplan Meir plot of the overall survival (OS) in ASCL1⁺ stage I AD in patients with low ($n = 38$) and high ($n = 15$) expression levels of RET. A significant association with the overall survival was identified ($p = 0.007$).

Figure 11 is a plot of ASCL1 and RET expression versus promoter methylation of the ASCL1 promoter in the GSE32867 dataset. This data indicates that promoter hypomethylation increases expression levels of the ASCL1 transcript. Interestingly, high expression level of ASCL1 is significantly associated with high level of RET. In the plot, samples with high transcript levels of RET are indicated by circles around the solid circles. RET and ASCL1 expression levels are based on the Illumina ILMN_1655610 and ILMN_1701653 probesets, respectively. ASCL1 promoter methylation is assessed by the cg20053158 probeset. High RET expression is marked by circles.

Figure 12 is a Kaplan Meir plot of the ASCL1⁺ AD patients overall survival based on the levels of the RET protein by IHC. Samples are from Mayo patients and were used in the discovery step by microarrays. A significant association between RET IHC and the overall survival was identified ($p = 0.038$).

Figure 13 contains three graphs of an *in vitro* analysis that confirms that ASCL1 regulates RET expression.

Figure 14 contains photographs of cells from a filling the gap scratch assay (see, e.g., Liang *et al.*, *Nature Protocols*, 2:329-333 (2007)). Wild-type HCC1833 cells and HCC1833 cells transfected with a control vector filled most of the gap by day 3, while HCC1833 cells transfected with an ASCL1 knock down vector (ASCL1-sh2 cells) did not fill as much of the gap, indicating that wild-type HCC1833 cells and cells transfected with empty vector (VEC) have a higher migration capacity than cells transfected with ASCL1 shRNA (ASCL1-sh2).

Figure 15 contains a bar graph plotting the clonogenic number of cells that do not express ASCL1 (A549 cells) and cells that express ASCL1 (A549-ASCL1 cells) after treatment with the indicated amount of cisplatin (e.g., 0 μ M, 3 μ M, or 9 μ M) for 14 days. Figure 15 also contains a line graph plotting the viability of A549 cells and A549-ASCL1 cells 72 hours after treatment with the indicated amount of cisplatin. These results

demonstrate that adenocarcinomas that express ASCL1 appear to be more resistant to treatment by cisplatin.

Figure 16 is a line graph plotting the viability of A549 cells and A549-ASCL1 cells 72 hours after treatment with the indicated amount of sunitinib (marketed as Sutent by Pfizer, and previously known as SU11248). The vertical line is drawn at about 3 μ M. These results demonstrate that adenocarcinomas that express ASCL1/RET are more susceptible to treatment by sunitinib.

DETAILED DESCRIPTION

This document provides methods and materials related to identifying mammals having lung adenocarcinoma characterized by neuroendocrine differentiation. For example, this document provides methods and materials for identifying mammals (e.g., humans) as having lung adenocarcinoma characterized by neuroendocrine differentiation by determining whether or not a lung cancer sample (e.g., lung tissue biopsy) from the mammal contains cancer cells having an elevated level of ASCL1 expression and/or an elevated level of RET expression. As described herein, if a mammal contains lung cancer cells with an elevated level of ASCL1 expression and/or an elevated level of RET expression, then that mammal can be classified as having lung adenocarcinoma characterized by neuroendocrine differentiation. If a mammal contains a lung cancer cells that lack an elevated level of ASCL1 expression and lack an elevated level of RET expression, then that mammal can be classified as not having lung adenocarcinoma characterized by neuroendocrine differentiation.

The term “elevated level” as used herein with respect to a level of expression (e.g., ASCL1 and/or RET expression) refers to any level that is greater than a reference level for that molecule (e.g., a reference level of ASCL1 and/or RET expression). The term “reference level” as used herein with respect to a particular molecule (e.g., a reference level of ASCL1 and/or RET expression) refers to the level of expression that is typically observed with normal healthy lung cells or lung adenocarcinoma characterized by a lack of neuroendocrine differentiation from mammals (e.g., humans). For example, a reference level of ASCL1 expression can be the average level of ASCL1 expression that is present in lung cells obtained from a random sampling of 50 humans free of lung

cancer. In some cases, an elevated level of expression (e.g., ASCL1 and/or RET expression) can be a level that is at least 10, 25, or 50 percent greater than a reference level for that molecule (e.g., a reference level of ASCL1 and/or RET expression). In some cases, an elevated level of ASCL1 expression or RET expression can be a
5 detectable level (e.g., an expression level detectable by immunocytochemistry). It will be appreciated that levels from comparable samples are used when determining whether or not a particular level is an elevated level.

As described herein, the level of ASCL1 and/or RET expression within lung cancer cells can be used to determine whether or not a particular mammal has lung
10 adenocarcinoma characterized by neuroendocrine differentiation. Any appropriate lung cancer sample can be used as described herein to identify mammals having lung adenocarcinoma characterized by neuroendocrine differentiation. For example, lung cancer tissue samples, lung cancer cell samples, and lung cancer needle biopsy specimen can be used to determine whether or not a mammal has lung adenocarcinoma
15 characterized by neuroendocrine differentiation.

In addition, any appropriate method can be used to obtain lung cancer cells. For example, a lung cancer sample can be obtained by a tissue biopsy or following a surgical resection. Once obtained, a sample can be processed prior to measuring a level of expression. For example, a lung cancer sample can be processed to extract RNA from
20 the sample. Once obtained, the RNA can be evaluated to determine the level of an mRNA of interest. In some embodiments, nucleic acids present within a sample can be amplified (e.g., linearly amplified) prior to determining the level of expression (e.g., using array technology). In another example, a lung cancer sample can be frozen, and sections of the frozen tissue sample can be prepared on glass slides. The frozen tissue
25 sections can be stored (e.g., at -80°C) prior to analysis, or they can be analyzed immediately (e.g., by immunohistochemistry with an antibody specific for a particular polypeptide of interest).

Any appropriate methods can be used to determine the level of ASCL1 and/or RET expression within lung cancer cells. For example, quantitative real time PCR, *in situ* hybridization, or microarray technology can be used to determine whether or not a
30 particular sample contains an elevated level of mRNA expression for a particular nucleic

acid or lacks an elevated level of mRNA expression for a particular nucleic acid. In some cases, the level of expression can be determined using polypeptide detection methods such as immunochemistry techniques. For example, antibodies specific for ASCL1 and/or RET polypeptides can be used to determine the polypeptide level in a sample. In
5 some cases, polypeptide-based techniques such as ELISAs and immunocytochemistry techniques can be used to determine whether or not a particular sample contains an elevated level of polypeptide expression for a particular nucleic acid or lacks an elevated level of polypeptide expression for a particular nucleic acid.

Examples of a human ASCL1 nucleic acid can have the sequence set forth in
10 GenBank[®] Accession No. NM_004316 (GI No. 190343011), and a human ASCL1 polypeptide can have the sequence set forth in GenBank[®] Accession No. NP_004307 (GI No. 55743094). Examples of a human RET nucleic acid can have the sequence set forth in GenBank[®] Accession No. NM_020630 (GI No. 126273513) or NM_020975 (GI No. 126273511), and a human RET polypeptide can have the sequence set forth in GenBank[®]
15 Accession No. NP_065681 (GI No. 10862701) or NP_066124 (GI No. 10862703).

Once the level of ASCL1 and/or RET expression within lung cancer cells from a mammal is determined, the level(s) can be compared to reference level(s) and used to classify the mammal as having or lacking lung adenocarcinoma characterized by neuroendocrine differentiation as described herein.

20 This document also provides methods and materials to assist medical or research professionals in identifying a mammal as having lung adenocarcinoma characterized by neuroendocrine differentiation. Medical professionals can be, for example, doctors, nurses, medical laboratory technologists, and pharmacists. Research professionals can be, for example, principle investigators, research technicians, postdoctoral trainees, and
25 graduate students. A professional can be assisted by (a) determining the level of ASCL1 and/or RET expression within lung cancer cells, and (b) communicating information about that the level(s) to that professional.

Any method can be used to communicate information to another person (e.g., a professional). For example, information can be given directly or indirectly to a
30 professional. In addition, any type of communication can be used to communicate the information. For example, mail, e-mail, telephone, and face-to-face interactions can be

used. The information also can be communicated to a professional by making that information electronically available to the professional. For example, the information can be communicated to a professional by placing the information on a computer database such that the professional can access the information. In addition, the
 5 information can be communicated to a hospital, clinic, or research facility serving as an agent for the professional.

This document also provides methods and materials for treating lung adenocarcinoma characterized by neuroendocrine differentiation. For example, one or more molecules listed in Table 1, 2, or 3 can be administered to a mammal (e.g., a
 10 human) having lung adenocarcinoma characterized by neuroendocrine differentiation under conditions wherein the presence or progression of the lung adenocarcinoma characterized by neuroendocrine differentiation is reduced. For example, a molecule listed in Table 1 such as tedisamil can be administered to a human having lung adenocarcinoma characterized by neuroendocrine differentiation such that the number of
 15 lung adenocarcinoma cells within the human is reduced. In some cases, one or more of molecules listed in Table 2A or 2B can be administered in combination with one or more of molecules listed in Table 1 to treat lung adenocarcinoma characterized by neuroendocrine differentiation. For example, tedisamil can be administered in combination with riluzole to a human having lung adenocarcinoma characterized by
 20 neuroendocrine differentiation.

Table 1. Molecules for treating lung adenocarcinoma characterized by neuroendocrine differentiation.

Gene	Molecule	Dosage Range (mg/kg)
ADRA2A	Paliperidone	6-12 mg daily oral, 117 monthly if injectable but can vary 39-234 mg
FGB	sucralfate	1 g (e.g., 10 mL/2 teaspoonfuls) four times per day
TUBB2B	Brentixumab vedotin	1.8 mg/kg administered as an intravenous infusion over 30 minutes every 3 weeks
TUBB2B	cabazitaxel	20 to 25 mg/m ² administered as a one-hour intravenous infusion every three weeks

KCNMB4	tedisamil	0.5-4 mg/kg, i.v.
--------	-----------	-------------------

Table 2A. Molecules that can be used in combination with one or more molecules listed in Table 1 for treating lung adenocarcinoma characterized by neuroendocrine differentiation.

Gene	Molecule	Dosage Range (mg/kg)
RET	sunitinib	30 mg/kg
RET	vandetanib	60 mg/kg
SCN3A	riluzole	40-60 mg twice daily
FGA	Alteplase	0.9 mg/kg, and a total not exceeding 90 mg
FGA	Anistreplase	IV 30 units over 2 to 5 min into IV line or vein
FGA	Tenecteplase	less than 60 kg: 30 mg IV bolus administered over 5 seconds. 60 to less than 70 kg: 35 mg IV bolus administered over 5 seconds 70 to less than 80 kg: 40 mg IV bolus administered over 5 seconds 80 to less than 90 kg: 45 mg IV bolus administered over 5 seconds 90 kg or greater: 50 mg IV bolus administered over 5 seconds)
FGA	Sucralfate	1 g (10 mL/2 teaspoonfuls) four times per day
KIT	Dasatinib	100-180 mg once daily
KIT	sunitinib	30 mg/kg
KIT	pazopanib	400-800 mg orally once daily
KIT	tivozanib	1-2 mg daily
KIT	OSI-930	500 mg twice a day
KIT	Telatinib	20 mg once daily to 1,500 mg twice daily
KIT	tandutinib	50 mg to 700 mg twice daily
KIT	imatinib	400-800 mg a day

KIT	sorafenib	400-800 mg a day
DDC	Levodopa/Carbidopa/Entacapone Orion	200 mg/50 mg/200 mg dose is 7 tablets per day (maximum dose a day)
DDC	carbidopa/levodopa	1 tablet of carbidopa 25 mg/levodopa 100 mg orally 3 times a day, or 1 tablet of 10 mg carbidopa/100 mg levodopa orally 3 to 4 times a day. The dose may be increased by 1 tablet orally every 1 to 2 days to a dose of 8 tablets/day (2 tablets orally 4 times a day)
DDC	Carbidopa	70-100 mg a day
DDC	L-Dopa	100-500 mg
CHRNA9	ABT-089	1-50 mg
CHRNA9	mecamylamine	2-25 mg
CHRNA9	succinylcholine	0.3-2.0 mg/kg

Table 2B. Molecules that can be used to reduce or inhibit the activity of polypeptides encoded by the listed genes.

Gene	Molecule
KCNMB4	tedisamil
RET	sunitinib, vandetanib
SCN3A	riluzole
ADRA2A	paliperidone, risperidone, antazoline/naphazoline, acetaminophen/clemastine/pseudoephedrine, articaine/epinephrine, bupivacaine/epinephrine, caffeine/ergotamine, acetaminophen/dexbrompheniramine/pseudoephedrine, dapiprazole, dexbrompheniramine/pseudoephedrine, chlorpheniramine/ibuprofen/pseudoephedrine, dipivefrin,

	cetirizine/pseudoephedrine, asenapine, epinephrine/prilocaine, epinephrine/lidocaine, PYM- 50018, V2006, lurasidone, paliperidone palmitate, fexofenadine/pseudoephedrine, guaifenesin/phenylpropanolamine, oxymetazoline, prazosin, phenylpropanolamine, ephedrine, tolazoline, guanfacine, guanabenz, guanethidine, phenoxybenzamine, dexmedetomidine, UK 14304, clonidine, dexefaroxan, quinidine, polythiazide/prazosin, chlorothiazide/methyldopa, chlorthalidone/clonidine, propafenone, guanadrel, hydrochlorothiazide/methyldopa, metaraminol, tizanidine, quetiapine, D-pseudoephedrine, apraclonidine, venlafaxine, phentolamine, labetalol, mephentermine, propylhexedrine, yohimbine, dihydroergotamine, ergotamine, norepinephrine, alpha- methyl dopa, epinephrine, dopamine, chlorpheniramine/phenylpropanolamine, desloratadine/pseudoephedrine, acrivastine/pseudoephedrine, carbinoxamine/pseudoephedrine, brompheniramine/codeine/phenylpropanolamine, pseudoephedrine/triprolidine, codeine/pseudoephedrine/triprolidine, carbetapentane/chlorpheniramine/ephedrine/phenylephrine, brompheniramine/dextromethorphan/pseudoephedrine, chlorpheniramine/hydrocodone/pseudoephedrine, azatadine/pseudoephedrine, naphazoline, carbinoxamine/dextromethorphan/pseudoephedrine
FGA	F2
FGB	F2
KIT	dasatinib, sunitinib, pazopanib, tivozanib, OSI-930,

	telatinib, tandutinib, imatinib, sorafenib
DDC	carbidopa/entacapone/levodopa, carbidopa/levodopa, S(-)-carbidopa, L-dopa
TUBB2B	brentuximab vedotin, cabazitaxel
CHRNA9	ABT-089, isoflurane, mecamylamine, succinylcholine, rocuronium, doxacurium, amobarbital, mivacurium, pipecuronium, rapacurium, metocurine, atracurium, cisatracurium, acetylcholine, nicotine, D-tubocurarine, arecoline, enflurane, pancuronium, vecuronium

The invention will be further described in the following examples, which do not limit the scope of the invention described in the claims.

5

EXAMPLES

Example 1 – Elevated ASCL1 and RET expression can be used to identify patients with lung adenocarcinoma characterized by neuroendocrine differentiation

Patient sample population

Using the Mayo Clinic frozen tumor bank, lung specimens resected from 303 patients between 1997 and 2007 were selected. Neoadjuvant therapy was not given to any patient included in this study. Formalin-fixed paraffin-embedded H&E sections from the corresponding surgical specimens were reviewed, and the diagnoses were confirmed according to the 2004 World Health Organization classification of tumors.

Bronchioloalveolar carcinoma variant of lung adenocarcinoma (AD) was excluded; hence, all ADs analyzed were clearly and predominantly invasive tumors. Never-smokers (NS) were characterized by less than 100 cigarettes per lifetime. Samples exclusively from NS (n = 130) were analyzed on the Illumina platform, and samples from former and current smokers (S) patients (n = 186) and NS (n = 18) were analyzed on the Affymetrix platform. Table 3 describes the clinicopathologic features of the samples.

20

Table 3. Clinicopathologic features of samples used.

	Samples arrayed on the Affymetrix platform	Samples arrayed on the DASL platform
Age at diagnosis (median, range)	69 yrs, 31-93 yrs	67 yrs, 17-91 yrs
Sex		
Male	94	24
Female	92	82
Smoking status		
Smoker	166	0
Never-smoker	18	106
Not Available	2	0
Histological analysis		
Adenocarcinoma (AD)	132	70
Adeno-squamous	0	6
Squamous cell carcinoma (SQCC)	24	2
Small cell carcinoma (SCLC)	15	0
Large cell carcinoma (LCC)	5	0
Typical carcinoid (TC)	10	24
Atypical carcinoid (AC)	0	4
Non-neoplastic lung tissue (N)	12	118

Immunohistochemical (IHC) Analysis

IHC procedures for ASCL1, CHGA, SYP, CD56/NCAM, and RET were as follows. A representative formalin fixed paraffin embedded (FFPE) block from a subset of gene expression profiled lung tumors of smokers (S), consisting of adenocarcinoma (AD) (n=83), small cell lung carcinoma (SCLC) (n=12), large cell carcinoma (LCC) (n=4), and large cell neuroendocrine carcinoma (LCNEC) (n=2), was selected. The analysis was limited to S as NE differentiation was significantly more prevalent in this group of tumors. IHC studies using antibodies directed against ASCL1/MASH1 (monoclonal, clone 24B72D11.1, 1:50 dilution, BD/Pharmingen, San Diego, CA), CHGA (monoclonal, clone LK2H10, 1:500 dilution, Chemicon/Millipore, Billerica, MA), SYP (monoclonal, clone SY38, 1:40 dilution, ICN, Irvine, CA), and CD56/NCAM (monoclonal clone 123C3, 1:25 dilution, Monosan, Uden, the Netherlands) were performed. The IHC stains were detected by the Dako Advance polymer-based detection

system (Dako, Carpinteria, CA, U.S.) using the Dako Autostainer. For each IHC assay, a positive control and negative control were performed. Immunostained slides were reviewed and scored by two pathologists, who were blinded to the corresponding microarray data. A consensus score was achieved for all cases. Cases were considered immunoreactive when exhibiting 5% or more tumor cells showing a nuclear staining pattern for ASCL1, a clear granular cytoplasmic staining pattern for CHGA and SYP, and a distinct membranous staining pattern for CD56/NCAM.

Similarly, twenty nine AD samples (14 ASCL1⁺ and 15 ASCL1⁻) with microarray expression data were selected for RET IHC using 1:500 dilutions of Epitomics 3454-1 rabbit monoclonal antibody. An ASCL1/RET co-IHC was developed by DAB staining for ASCL1 first (1:100 dilution monoclonal, clone 24B72D11.1, BD/Pharmingen, San Diego, CA) and then Fast Red staining (1:500 dilutions of Epitomics 3454-1 rabbit monoclonal antibody) for RET.

Immunoreactivity was semi-quantitatively scored based on a) the percentage of positive tumor cells (Labeling index, LI), ranging from 0 to 100%, in increments of 5%; and b) the intensity of staining, graded as: weak +1, moderate +2, and strong +3. For a comparative analysis of NE markers (ASCL1, CHGA, SYP, and CD56/NCAM), the Log₂ of the product of the percentage of positive tumor cells (Labeling index, LI) multiplied by the intensity of staining was determined for each IHC NE marker and used to generate a heat map of the IHC NE markers using 'heatmap' function in the open source package R version 2.12.2 (World Wide Web at "r-project.org/"). RET IHC frequently had areas with different intensity of stains. In each case, RET IHC score was computed as the summation of Log₂ (LI) x intensity for each stained area.

Preparation of samples for expression profiling on the Affymetrix platform

Lung tumor cells and non-neoplastic cells were collected by either laser capture microdissection (LCM=86) or macrodissection (M=112) to assure high tumor content (>80%) as described elsewhere (Klee *et al.*, *BMC Med. Genomics*, 2:13 (2009)). Total RNA from samples collected by LCM was isolated using the Micropure kit (Qiagen Corp, Valencia, CA) as described elsewhere (Savci-Heijink *et al.*, *Am. J. Pathol.*, 174(5):1629-37 (2009)). Briefly, RNA quality and quantity were controlled by the

Agilent bioanalyzer and the Ribogreen assay or by a quantitative PCR assay based on the ratio of concentration of 3' to middle transcript of β -actin. Total RNA (10 ng) from these LCM- collected samples were labeled in a two round linear amplification/labeling process according to the Small Sample Preparation protocol (Affymetrix Corp, Santa Clara, CA). Affymetrix arrays were scanned according to the manufacturer's protocol. Total RNA from samples obtained by macrodissection was isolated using the RNeasy kit (Qiagen). The quality and quantity of RNA samples were controlled by the Agilent bioanalyzer and a NanoDrop spectrophotometer. Total RNA (1.2 μ g) was labeled according to the standard Affymetrix protocol. Labeled cRNA was hybridized to U133PLUS2 chipset.

Preparation of samples for expression profiling on the Illumina platform

RNA from macrodissected samples were purified by the RNeasy kit (Qiagen) and analyzed by the Agilent bioanalyzer and a NanoDrop spectrophotometer. For the WG-DASL assay (Illumina, San Diego, CA), total RNA (100 ng) was reverse transcribed with biotinylated primers. The resulting cDNA was annealed to chimeric query oligonucleotides, which contained a gene-specific region and a universal primer sequence for PCR amplification, and then bound to streptavidin-conjugated paramagnetic particles. The gene-specific oligonucleotides were extended by second-strand cDNA synthesis and then ligated. Subsequently, the products were sequestered by magnetic separation, washed to remove unbound molecules, and then amplified by PCR with fluorophore-labeled universal primers. The resulting PCR products were purified, applied to HumanRef-8 v3 beadchips, and then hybridized for 16 hours at 58° C. The beadchips were washed and scanned in a BeadArray Reader using BeadScan v3 software (Illumina).

Microarray data analysis

Normalized expression values from WG-DASL experiments were generated by the Bead Studio software (Illumina). Affymetrix intensity files (.CEL files) were processed and normalized by the 'germa' package in R. All subsequent analyses of DASL and Affymetrix data were carried out in R. Other than data generated at Mayo, expression analysis included various publically available Affymetrix datasets. Two

major datasets which were a compendium of smaller datasets and frequently used in this study were named Dataset 1 and Dataset 2. Compositions of these two sets are shown in Figure 1. AD and LCC samples with high expression of either DSG3 or KRT5 (squamous differentiation markers) were excluded. Similarly, squamous cell carcinoma (SQCC) samples with low expression of DSG3 and KRT5 were excluded. Pearson correlation coefficients between various NE markers were calculated by 'cor.test,' and a heatmap of all samples in Dataset 1 was generated by the 'heatmap' function.

Differentially expressed transcripts between ASCL1⁺ and ASCL1⁻ tumors

Dataset 2 (Figure 1) was used to examine the expression differences between ASCL1⁺ and ASCL1⁻ tumors in stage I AD. All files had follow up information. Array files (n = 593) with more than 22,000 common Affymetrix probesets were included in this dataset. The microarray signal intensity (.CEL) files were normalized and processed by the "gcrma" package in R. Threshold for ASCL1 status (+ or -) was chosen as before using 209988_s_at probeset at Log₂ intensity of 8. To identify most differentially expressed genes in ASCL1⁺ versus ASCL1⁻ tumors, probesets were ranked by signal to noise ratio calculated as $SNR = (\mu_{ASCL1+} - \mu_{ASCL1-}) / (\sigma_{ASCL1+} + \sigma_{ASCL1-})$ where μ 's were mean expression values and σ 's were maximum of $0.2 \times \mu$ and standard deviation (Golub *et al.*, *Science*, 286(5439):531-7 (1999)). SNR values greater than and less than zero potentially indicate over and under expression in ASCL1⁺ compared with ASCL1⁻ tumors, respectively. It was also required in this example that the average expression in samples over-expressing a gene had greater than 3.5 Log₂ intensities. Log₂ expression intensities for the gcrma normalized data ranged from 2 to 15. Based on experience with quantitative RT-PCR, gene expression intensities below 3.5 were not reliable and frequently not detected. Significant figures for over-expression in ASCL1⁺ compared with ASCL1⁻ tumors were calculated by t-test and then corrected for multiple comparison correction using the 'qvalue' package in R (Storey *et al.*, *Proc. Natl. Acad. Sci. USA*, 100(16):9440-5 (2003)).

Survival analysis

Given that only 15-20% of AD expresses ASCL1, any one dataset by itself did not provide sufficient samples for statistical analysis. Therefore, the Mayo dataset (n = 132) was combined with four other lung AD microarray datasets that had follow up information available. These included the Director's Challenge dataset (Shedden *et al.*, *Nat. Med.*, 14(8):822-7 (2008)) (n = 420), Bhattacharjee dataset (Bhattacharjee *et al.*, *Proc. Natl. Acad. Sci. USA*, 98(24):13790-5 (2001)) (n = 139), Kune dataset (GEO dataset GSE10245, n = 40), and Hou dataset (GEO dataset GSE19188, n = 45). With the exception of Bhattacharjee dataset, all other array files had common probesets for ASCL1, and the most variable probeset in all sets (209988_s_at) was chosen to determine the expression levels of ASCL1. Based on the IHC data, expression levels above signal intensity 8 (Log₂) were chosen as the threshold for ASCL1⁺ and ASCL1⁻ status (Figure 2). The range of microarray Log₂ signal intensities for this probeset was 2-15. Therefore, signal intensity of 8 or higher corresponded to a moderate to high expression level in the 85th percentile and higher. The ASCL1 status in the Bhattacharjee dataset was determined by inspecting ASCL1 expression histograms and selecting thresholds breakpoints at the 85 percentile. Survival analyses used the "survival" package in R ([http at "http://cran.at.r-project.org"](http://cran.at.r-project.org)) and included time to progression and overall survival stratified by stage. In the combined dataset, differences in survival times between ASCL1⁻ and ASCL1⁺ tumors for stage I patients who died was assessed by a group t-test. A subset of 11 AD in the Director's Challenge data with high expression of CHGA, SCG2, CHGB, NCAM1 (CD56), or SYP were identified as large cell NE carcinomas (LCNEC) after histologic review (Bryant *et al.*, *PLoS One*, 5(7):e11712 (2010)). In light of this finding, the cases of AD in this study were re-reviewed, and the diagnosis confirmed by morphology. Furthermore, in the Director's Challenge data, 19 cases were identified with high expression of at least one of these NE markers which could have represented LCNEC and thus NSCLC with poorer prognosis. A group t-test was repeated after excluding these samples, but did not alter the overall results and conclusions. In the analysis of overall survival, the proportional hazard assumption for stage I tumors was tested by the "cox.zph" routine in the "survival" package. A small p-value indicated non-

proportional hazard. Proportional hazard assumption was tested after censoring follow up times at five and more years.

Associations between RET expression and survival in ASCL1⁺ tumors

5 By Cox proportional hazards regression analysis in R (coxph), two probesets corresponding to the RET oncogene (215771_x_at, 205879_x_at) had significant associations with overall survival in stage I AD after the follow up data at 5 years was censored. To visualize this association by a Kaplan Meir (KM) plot, varying the threshold for “low” and “high” expression levels of RET (215771_x_at) was examined. 10 Values in 3.0 to 6.5 were significant with p values ranging 0.0005 to 0.029. Excluding AD samples where an alternative diagnosis of LCNEC was possible did not appreciably change these results (Bryant *et al.*, *PLoS One*, 5(7):e11712 (2010)). The reported KM plot used a threshold of 3.5, as signal intensities below this threshold are usually not detected by RT-PCR. If the data was not censored at 5 years, p-values ranged from 15 0.00053 to 0.037 as the threshold changed from 3.0 to 6.5. Same probeset and threshold was used in the KM plot of all AD stages. Also, a KM plot for RET stains was generated by using the mean of all RET IHC scores as the threshold for selecting “low” and “high” levels.

20 *Gene set analysis*

To find gene sets enriched in ASCL1⁺ tumors compared with ASCL1⁻ tumors, probesets (13166) with SNR greater or less than zero were used in the GSA package in R and using Molecular Signatures Database (MSigDB) version 3.0. The analysis used 500 permutations and an FDR default value of 0.05. For robustness, 20 iterations were 25 performed, and gene sets identified in at least 16 iterations (80%) were reported. To find gene sets associated with aggressive behavior in ASCL1⁺ tumors, these tumors were divided into aggressive and non-aggressive groups. Aggressive tumors were from patients who died in less than 3.5 years after surgery (n = 21) and non-aggressive tumors were from patients who survived 6 or more years after surgery (n = 20). Probesets 30 (13126) with SNR greater or less than zero in comparisons of aggressive versus non-

aggressive tumors were used in the GSA program with the same selection criteria as above.

Comparative IHC analysis of NE markers (ASCL1, CHGA, SYP and CD56/NCAM)

- 5 Immunostaining quality of ASCL1, CHGA, SYP, and CD56/NCAM was comparable, and all slides were interpretable. Scattered immunoreactive bronchiolar basal-located NE cells were considered as positive internal controls for the IHC reaction. Labeling indices (LIs) and immunoreactivity for ASCL1, CHGA, SYP, and CD56/NCAM for AD, SCLC, LCNEC and LCC are shown in Table 4.

10

Table 4. Detailed results of the immunohistochemical study for ASCL1 and other NE markers in all lung cancer subtypes.

SUBTYPE		AD	SCLC	LCNEC	LCC
No. of patients		83	12	2	4
ASCL1	LI mean +/- SD	54.4 +/- 29	84.6 +/- 25.6	95 +/- 0	0
	Range	5-95	5-95	95	0
	Immunoreactive cases	15/83 (18%)	12/12 (100%)	2/2 (100%)	0/4 (0%)
CHGA	LI mean +/- SD	42.5 +/- 40.2	74.6 +/- 31.4	55 +/- 49.5	0
	Range	5-85	50-100	20-90	0
	Immunoreactive cases	4/83 (5%)	11/12 (92%)	2/2 (100%)	0/4 (0%)
SYP	LI mean +/- SD	31.3 +/- 32.8	90.4 +/- 20.5	95 +/- 0	0
	Range	5-100	20-100	95	0
	Immunoreactive cases	20/83 (24%)	12/12 (100%)	2/2 (100%)	0/4 (0%)
CD56/NCAM	LI mean +/- SD	26 +/- 29	92.9 +/- 13.9	92.5 +/- 3.5	0
	Range	5-75	50-100	90-95	0
	Immunoreactive cases	5/83 (6%)	12/12 (100%)	2/2 (100%)	0/4 (0%)

- 15 The pattern of ASCL1 immunoreactivity varied according to tumor histological subtype. In AD showing ASCL1 immunoreactivity (ASCL1+AD), ASCL1⁺ cells were focal and admixed with ASCL1⁻ cells (Figures 3A and 3B), resulting in low to moderate LIs (Table 4). In SCLC (Figures 3F and 3G) and LCNEC examples (Figures 3K and 3L), ASCL1 immunostaining was diffuse, resulting in moderate to high LIs (Table 4). In addition, ASCL1 immunoreactivity in AD was more frequent than for CHGA (Figure

3C) and CD56/NCAM (Figure 3D); however, SYP (Figure 3E) was the most common IHC NE marker expressed in this group as shown in Table 5 and illustrated in Figure 3P. One ASCL1⁺ AD was not immunoreactive for any of the other IHC NE markers, whereas six ADs exhibiting reactivity for at least one of the other NE markers were ASCL1⁻ AD (Table 6). Among SCLC (Figures 3H, 3I and 3J) and LCNEC (Figures 3M, 3N and 3O), all cases were immunoreactive for ASCL1 as well as for most other IHC NE markers (CHGA, SYP, and CD56/NCAM); whereas the remaining 4 LCC examples were not immunoreactive for any IHC NE marker, including ASCL1 (Figure 3P).

10 Table 5. Detailed results of correlation between ASCL1 and other NE markers in all lung cancer subtypes

SUBTYPE		CHGA ⁺	CHGA ⁻	SYP ⁺	SYP ⁻	CD56/ NCAM ⁺	CD56/ NCAM ⁻
AD (n = 83)	No	4	79	20	63	5	78
	ASCL1⁺ (n=15)	3	12	14	1	4	11
	ASCL1⁻ (n=68)	1	67	6	62	1	67
SCLC (n = 12)	No	11	1	12	0	12	0
	ASCL1⁺ (n=12)	11	1	12	0	12	0
	ASCL1⁻ (n=0)	0	0	0	0	0	0
LCNEC (n = 2)	No	2	0	2	0	2	0
	ASCL1⁺ (n=2)	2	0	2	0	2	0
	ASCL1⁻ (n=0)	0	0	0	0	0	0
LCC (n = 4)	No	0	4	0	4	0	4
	ASCL1⁺ (n=0)	0	0	0	0	0	0
	ASCL1⁻ (n=4)	0	4	0	4	0	4

Table 6. Correlation of ASCL1 with other NE markers in AD

AD SUBTYPE	CHGA/SYP/CD56* ⁺	CHGA/SYP/CD56* ⁻
ASCL1⁺ AD (n=15)	14 (94%)	1 (6%)
ASCL1⁻ AD (n=68)	6 (9%)	62 (91%)

ASCL1 mRNA expression is more prevalent in AD than in SQCC

The expression of ASCL1 and other known NE markers in Dataset 1 (Figure 1) consisting of AD (n = 232), SQCC (n = 100), SCLC (n = 15), adjacent non-neoplastic lung (N, n = 12), and LCC and LCNEC (n = 9) is shown as a heatmap in Figure 4. The heatmap also includes the SQCC and AD differentiation genes DSG3 and NKX2.1/TTF1, respectively, and RET. ASCL1 had the highest correlation with calcitonin (CPRG/CALCA) (correlation coefficient = 0.65). However, in general, there was not a high correlation between the mRNA expression levels of the NE markers (Table 7). Most NE markers had similar frequency of expression in the AD and SQCC samples. In contrast, ASCL1 was much more specific to AD than to SQCC. For a quantitative analysis, a threshold was selected for ASCL1 that corresponded to a positive IHC stain (Figure 2). Microarray signal intensity levels above this threshold had excellent correlation with the IHC staining (correlation coefficient = 0.89, Figure 2). In AD, 44 of 232 cases (19.0%) were ASCL1⁺. On the other hand, in 100 SQCC only 1 of 100 cases (1.0%) was ASCL1⁺. Of the nine LCC, two had strong expression of ASCL1 and all other NE markers and were classified as LCNEC. Importantly, ASCL1 also was highly prevalent in other NE lung tumors, including SCLC and carcinoid tumors (CT). Six of 10 (60%) and 14 of 15 (93%) CT and SCLC, respectively, were ASCL1⁺.

Table 7. Correlation between any two NE markers in AD and SQCC that express either or both markers.

	CHGA	CHGB	SCG2	INSM1	PCSK1	SYP	NCAM1	ASCL1	CALCA
CHGA	1								
CHGB	0.219	1							
SCG2	0.395	0.207	1						
INSM1	NS*	0.259	0.439	1					
PCSK1	NS*	0.3	0.33	0.308	1				
SYP	NS*	NS*	NS*	NS*	NS*	1			
NCAM1	NS*	NS*	NS*	NS*	NS*	NS*	1		
ASCL1	NS*	NS*	NS*	NS*	0.497	NS*	NS*	1	
CALCA	NS*	NS*	0.177	NS*	0.593	NS*	NS*	0.65	1

*NS: Pearson correlation p-value > 0.05

Neuroendocrine differentiation is rare in non-smoker adenocarcinomas

Expression levels of known NE markers were examined in the Mayo Clinic lung cancer samples from NS, which included 75 AD, 32 CT, 8 adenosquamous carcinomas

and SQCC, and 125 adjacent non-neoplastic (N) samples. Compared with N, all NE markers were over-expressed in a majority of CT as expected (Figure 5). In contrast, the expression levels of NE markers in AD were within the range of N. A subset of AD was not identified with marked over-expression of any of the NE markers. These data suggest that NE differentiation in lung AD is largely restricted to smokers.

Survival analysis of lung AD in relation to ASCL1 mRNA expression

Given that ASCL1 is expressed in about 20% of AD, to obtain sufficient statistical power for survival analysis, the Mayo Clinic AD microarray data was combined with four publicly available AD datasets for which outcome data was available. An association was not identified between the ASCL1 expression status and survival or time to progression in stage I tumors nor in combined stage II-IV tumors ($p \geq 0.28$). However, the Kaplan Meir (KM) survival curves for ASCL1⁺ and ASCL1⁻ stage I tumors had different drop off profiles (Figure 6). ASCL1⁺ patients who died had significantly shorter survival times causing a sharp drop in the survival curve. Upon further investigation, this pattern was found to be consistent in all five data sets (Table 8). In the combined datasets, ASCL1⁺ patients who died had statistically shorter survival times than ASCL1⁻ patients who died ($p < 5 \times 10^{-6}$). This trend did not change after excluding samples suspected of LCNEC in the Director's Challenge dataset. A statistical test (cox.zph) indicated non-proportional hazards in ASCL1⁻ / ASCL1⁺ tumors when censoring times of 6.5 or more years ($p < 0.05$) were used. These observations suggested that ASCL1⁺ status might reflect a different underlying biology for these tumors. The roles of traditional prognostic markers such as age, gender, tumor grade, race, smoking status (former or current), tumor "T" stage, and tumor grade were assessed by cox analysis. The only significant parameters were age ($p = 10^{-6}$) and gender ($p = 0.045$). When stage I AD were stratified by ASCL1 status, differences in age and gender between the ASCL1⁺ and ASCL1⁻ patients were not identified (group t-test and chi-square $p \geq 0.09$).

Table 8. Survival times of patients with fatal stage I AD according to the ASCL1 status.

Dataset	ASCL1 ⁺			ASCL1 ⁻		
	n	median	mean	n	median	mean
Mayo Clinic	5	16.0	17.6	41	25.3	33.5
Director Challenge	14	31.9	29.4	93	45.8	50.3
Bhattacharjee <i>et al.</i>	4	20.3	20.0	27	25.5	31.5
Kune <i>et al.</i>	3	21.9	23.3	7	31.2	26.5
Hou <i>et al.</i>	1	4.9	4.9	11	24.2	38.4
All sets	27	23.6	24.3	179	38.2	41.9

RET mRNA expression in ASCL1⁺ AD is predictive of overall survival

To gain further insight into the biology of ASCL1⁺ tumors, gene expression data for ASCL1⁺ and ASCL1⁻ tumors were compared. Gene expression analysis used Dataset 2 (Figure 1), which was a compendium of four sets of microarray data with follow up information and more than 22,000 common Affymetrix probesets from 593 AD including 367 stage I AD. Genes (probesets) over-expressed in ASCL1⁺ compared with ASCL1⁻ tumors were identified by signal to noise ratio (SNR). The top 12 genes (16 probesets) following ASCL1 are listed in Table 9. All genes in the list were significantly over-expressed in ASCL1⁺ tumors after correcting for multiple comparisons (q-value < 10⁻⁶) (Storey and Tibshirani, *Proc. Natl. Acad. Sci. USA*, 100(16):9440-5 (2003)). RET was the fourth most over-expressed gene after ASCL1 followed by CALCA and C1orf95 (Table 9). Figure 7A illustrates the expression of RET in ASCL1⁻ and ASCL1⁺ stage I AD. The over-expression of RET in ASCL1⁺ tumors was consistent in all four datasets (Figure 7A). RET expression was more consistent with ASCL1 than other NE markers (Figure 4). Depending on the microarray signal threshold for calling a transcript present (Log₂ signal intensity of 3.5 or 4.5), 91 to 95% of samples that expressed RET also expressed ASCL1. In contrast, only 0% to 55% of samples that expressed RET also expressed CHGA, CHGB, SCG2, SYP, INSM1, PCSK1, or NCAM1. Also noted was a small portion of samples with high levels of RET in the absence of ASCL1 (black circles in Figure 7A), indicating that in rare cases RET is expressed independent of ASCL1.

Table 9. Probesets (20) with highest signal to noise ratio (SNR) in ASCL1⁺ compared with ASCL1⁻ tumors in Dataset 2.

Affy Probeset	Symbol	SNR	q-value	Drug
209988_s_at	ASCL1	2.78	1.4E-39	
209987_s_at	ASCL1	2.70	7.5E-27	
213768_s_at	ASCL1	1.51	4.5E-15	
217561_at	CALCA	1.40	1.2E-15	
210728_s_at	CALCA	1.33	4.5E-15	
210727_at	CALCA	1.32	5.0E-15	
217495_x_at	CALCA	1.16	4.4E-11	
209985_s_at	ASCL1	1.15	7.0E-11	
213925_at	C1orf95	1.09	9.1E-13	
211421_s_at	RET	1.06	1.4E-10	sunitinib, vandetanib
205549_at	PCP4	1.05	6.5E-20	
220782_x_at	KLK12	1.03	3.2E-11	
209617_s_at	CTNND2	0.97	3.2E-11	
214023_x_at	TUBB2B	0.93	1.2E-15	brentuximab vedotin, cabazitaxel
205305_at	FGL1	0.91	2.7E-13	
204623_at	TFF3	0.85	4.8E-18	
214058_at	MYCL1	0.85	7.0E-11	
205879_x_at	RET	0.82	5.6E-08	sunitinib, vandetanib
210432_s_at	SCN3A	0.80	3.1E-07	riluzole
209228_x_at	TUSC3	0.77	1.3E-13	

RET expression coinciding with *ASCL1* was not limited to stage I AD

5 A similar ASCL1/RET co-expression was observed in all stages of AD and other lung cancer subtypes. Figure 7B illustrates the expression of RET in Dataset 1. In SQCC where ASCL1 was largely absent, RET was also rarely expressed. In SCLC, CT, and LCC, RET expression was largely restricted to ASCL1⁺ tumors.

10 As in Figure 7A, RET mRNA was detectable in a limited number of lung cancers that did not express ASCL1 (black circles in Figure 7B).

Two probesets corresponding to RET were significant in predicting the overall survival (OS) in stage I ASCL1⁺ tumors by cox analysis (p values of 0.029 and 0.006). High expression of RET was associated with shorter survival. In contrast, an association was not identified between the OS and RET expression level in ASCL1⁻ tumors. For
15 illustration, a threshold for 'low' and 'high' expression of RET in a Kaplan Meir (KM) plot was selected as shown in Figure 8A. The results did not appreciably change after excluding samples where an alternative diagnosis of LCNEC was possible. Using the

same threshold, RET mRNA also was significant in predicting OS in all AD (Figure 8B).

RET protein expression analysis by IHC

A select set of Mayo AD samples with expression data by the microarrays were
 5 immunostained for RET. RET protein expression by IHC was more prevalent than
 expected from the microarrays, perhaps due to the sensitivity of antibody to multiple
 variants of RET. A blush staining was observed in some ASCL1⁻ cases with RET mRNA
 expression below detection levels by microarrays (Figure 9A). However, ASCL1⁺ cases
 from fatal tumors often had intensely stained areas (Figure 9B). There was a significant
 10 correlation between RET IHC scores and microarray signal intensity by the RET probeset
 used in OS analysis ($\rho = 0.43$, $p < 0.02$). An ASCL1/RET co-IHC assay was
 developed, and overlapping tumor areas with positive staining for both proteins were
 frequently found (Figure 9C). However, because of the discrepancies in sensitivity and
 specificity of RET and ASCL1 antibodies or because of ASCL1 independent activation
 15 of RET, areas with positive RET staining without ASCL1 expression also were
 encountered.

RET protein level by IHC was predictive of OS in the Mayo AD samples, which
 also were positive for ASCL1 by IHC (log-rank test p value = 0.05, Figure 9D). When
 cases were not stratified by the ASCL1 expression, RET IHC was not predictive of OS
 20 (Figure 9E). In this situation, median survival times of tumors with 'low' levels of RET
 was slightly less than tumors with 'high' levels of RET, but this difference was
 statistically insignificant. These results indicate that RET protein is predictive of OS
 survival only in the context of ASCL1 expression.

Gene Set Analysis of ASCL1⁺ tumors

To gain further insight in the biology of ASCL1⁺ tumors, gene set enrichment
 analysis was performed by the GSA program and MSigDB version 3 with close to 7000
 gene sets. The results are shown in Table 10. Notably, positively and negatively
 associated gene sets included OSADA_ASCL1_TARGETS_UP and _DN, respectively.
 30 These sets contained genes that were up and down regulated by ASCL1 in a study of
 ASCL1-transduced A549 lung AD cells (Osada *et al.*, *Cancer Res.*, 68(6):1647-55

(2008)). Importantly, RET was among the target genes up regulated by ASCL1 in the OSADA_ASCL1_TARGETS_UP set corroborating the observations in patient data. In the module corresponding to human chromosome and cytogenetic bands, 12q22 and 8p22 were enriched. Ten of 37 genes (including ASCL1) on chr12q22 and twelve of 41 genes on chr 8p22 were significantly over-expressed in ASCL1⁺ tumors. The high concentration of over-expressed genes in these regions suggested potential copy number changes.

Table 10. Gene sets positively and negatively associated with ASCL1⁺ compared with ASCL1⁻ stage I tumors.

Positive Associations		
Pathway Names	Frequency	Score
TATCTGG,MIR-488	100	0.76
BONE_REMODELING	90	1.2
TISSUE_REMODELING	85	1.16
module_382	85	2.46
chr12q22	85	1.64
OSADA_ASCL1_TARGETS_UP	80	1.16
chr8p22	80	1.72
HANN_RESISTANCE_TO_BCL2_INHIBITOR_DN	80	0.95
Negative Associations		
Pathway Names	Frequency	Score
CHARAFE_BREAST_CANCER_LUMINAL_VS_BASAL_DN	100	-0.85
HUANG_DASATINIB_RESISTANCE_UP	100	-1.34
BOYLAN_MULTIPLE_MYELOMA_D_DN	100	-0.92
MARSON_FOXP3_TARGETS_UP	100	-1.06
module_543	100	-2.66
HUMMEL_BURKITT'S_LYMPHOMA_DN	95	-1.48
WANG_BARRETT'S_ESOPHAGUS_AND_ESOPHAGUS_CA	95	-1.34
LEE_EARLY_T_LYMPHOCYTE_DN	95	-1.49
KEGG_VIRAL_MYOCARDITIS	95	-1.55
module_411	95	-0.68
FUJII_YBX1_TARGETS_UP	90	-1.51
GNF2_RAP1B	90	-1.41
module_223	90	-0.96
KEGG_CELL_ADHESION_MOLECULES_CAMS	90	-1.18
BIOCARTA_TH1TH2_PATHWAY	90	-2.42

module_341	90	-0.65
WU_CELL_MIGRATION	85	-0.74
OSADA_ASCL1_TARGETS_DN	80	-1.29
KIM_LRRC3B_TARGETS	80	-1.94
CASTELLANO_NRAS_TARGETS_UP	80	-0.86

mRNA correlates of aggressive behavior in stage I ASCL1⁺ AD also were examined. Tumors from patients who died in less than 3.5 years following surgery (n=21) and from patients who survived more than 6 years following surgery (n = 20) were designated as aggressive and non-aggressive tumors, respectively. When probesets were ranked by SNR in aggressive versus non-aggressive tumors, two probesets for RET were among the list of top 10 probesets. GSA analysis in these tumors identified six gene sets (Table 11). Most notably, KANG-CISPLATIN-RESISTANCE-UP was positively associated with aggressive tumors. This set included genes that were up-regulated in gastric cancer cell lines resistant to cisplatin (Kang *et al.*, *Clin. Cancer Res.*, 10(1 Pt 1):272-84 (2004)).

Table 11. Gene sets positively and negatively associated with aggressive behavior in ASCL1⁺ stage I AD.

Positive Associations		
Pathway Names	Frequency	Score
CHIANG_LIVER_CANCER_SUBCLASS_POLYSOMY7_UP	100	1.39
V\$AP1_Q4	85	0.35
module_94	85	0.4
KANG_CISPLATIN_RESISTANCE_UP	80	0.97
ENZYME_INHIBITOR_ACTIVITY	80	0.53
Negative Associations		
Pathway Names	Frequency	Score
V\$MAX_01	85	-0.3

In summary, the results provided herein demonstrate that lung cancer patients can be examined for the presence of lung cancer cells expressing ASCL1 (e.g., an elevated level ASCL1) and RET (e.g., an elevated level RET). If the presence of lung cancer cells expressing ASCL1 and RET is detected in a particular lung cancer patient, then that lung cancer patient can be classified as having lung adenocarcinoma characterized by

neuroendocrine differentiation and/or as having a poor survival prognosis. In some cases, lung cancer patients classified as having lung adenocarcinoma characterized by neuroendocrine differentiation can be treated as described herein.

5 Example 2 – Genes over expressed in lung adenocarcinoma
 expressing ASCL1 and RET

The genes listed in Table 12 were found to be overexpressed in lung adenocarcinoma samples that express ASCL1 and RET. Additional information about each of these ten genes is provided in Table 13. Possible drugs for treating lung
 10 adenocarcinoma characterized by neuroendocrine differentiation are listed in Table 14, Table 1, Table 2A, or Table 2B.

Table 12.

Symbol	Entrez Gene Name
KCNMB4	potassium large conductance calcium-activated channel, subfamily M, beta member 4
RET	ret proto-oncogene
SCN3A	sodium channel, voltage-gated, type III, alpha subunit
ADRA2A	adrenoceptor alpha 2A
FGA	fibrinogen alpha chain
FGB	fibrinogen beta chain
KIT	v-kit Hardy-Zuckerman 4 feline sarcoma viral oncogene homolog
DDC	dopa decarboxylase (aromatic L-amino acid decarboxylase)
TUBB2B	tubulin, beta 2B class IIb
CHRNA9	cholinergic receptor, nicotinic, alpha 9 (neuronal)

15 Table 13.

Symbol	Affymetrix	Location	Type(s)	Entrez Gene ID for Human	Entrez Gene ID for Mouse	Entrez Gene ID for Rat
KCNMB4	219287_at	Plasma Membrane	ion channel	27345	58802	66016
RET	205879_x_at	Plasma Membrane	kinase	5979	19713	24716
SCN3A	210432_s_at	Plasma Membrane	ion channel	6328	20269	497770
ADRA2A	209869 at	Plasma	G-protein	150	11551	25083

		Membrane	coupled receptor			
FGA	205649_s_at	Extracellular Space	other	2243	14161	361969
FGB	204988_at	Extracellular Space	other	2244	110135	24366
KIT	205051_s_at	Plasma Membrane	kinase	3815	16590	64030
DDC	205311_at	Cytoplasm	enzyme	1644	13195	24311
TUBB2B	214023_x_at	Cytoplasm	other	347733	73710	291081
CHRNA9	221107_at	Plasma Membrane	transmembrane receptor	55584	231252	65024

Table 14.

Symbol	Drug(s) for treating lung adenocarcinoma characterized by neuroendocrine differentiation
KCNMB4	tedisamil
RET	sunitinib, vandetanib
SCN3A	riluzole
ADRA2A	paliperidone, risperidone, antazoline/naphazoline, acetaminophen/clemastine/pseudoephedrine, articaïne/epinephrine, bupivacaine/epinephrine, caffeine/ergotamine, acetaminophen/dexbrompheniramine/pseudoephedrine, dapiprazole, dexbrompheniramine/pseudoephedrine, chlorpheniramine/ibuprofen/pseudoephedrine, dipivefrin, cetirizine/pseudoephedrine, asenapine, epinephrine/prilocaine, epinephrine/lidocaine, PYM-50018, V2006, lurasidone, paliperidone palmitate, fexofenadine/pseudoephedrine, guaifenesin/phenylpropanolamine, oxymetazoline, prazosin, phenylpropanolamine, ephedrine, tolazoline, guanfacine, guanabenz, guanethidine, phenoxybenzamine, dexmedetomidine, UK 14304, clonidine, dexefaroxan, quinidine, polythiazide/prazosin, chlorothiazide/methyldopa, chlorthalidone/clonidine, propafenone, guanadrel, hydrochlorothiazide/methyldopa, metaraminol, tizanidine, quetiapine, D-pseudoephedrine, apraclonidine, venlafaxine, phentolamine, labetalol, mephentermine, propylhexedrine, yohimbine, dihydroergotamine, ergotamine, norepinephrine, alpha-methyl dopa, epinephrine, dopamine, chlorpheniramine/phenylpropanolamine, desloratadine/pseudoephedrine, acrivastine/pseudoephedrine, carbinoxamine/pseudoephedrine, brompheniramine/codeine/phenylpropanolamine, pseudoephedrine/triprolidine, codeine/pseudoephedrine/triprolidine, carbetapentane/chlorpheniramine/ephedrine/phenylephrine, brompheniramine/dextromethorphan/pseudoephedrine, chlorpheniramine/hydrocodone/pseudoephedrine, azatadine/pseudoephedrine, naphazoline,

	carbinoxamine/dextromethorphan/pseudoephedrine
FGA	F2
FGB	F2
KIT	dasatinib, sunitinib, pazopanib, tivozanib, OSI-930, telatinib, tandutinib, imatinib, sorafenib
DDC	carbidopa/entacapone/levodopa, carbidopa/levodopa, S(-)-carbidopa, L-dopa
TUBB2B	brentuximab vedotin, cabazitaxel
CHRNA9	ABT-089, isoflurane, mecamylamine, succinylcholine, rocuronium, doxacurium, amobarbital, mivacurium, pipecuronium, rapacuronium, metocurine, atracurium, cisatracurium, acetylcholine, nicotine, D-tubocurarine, arecoline, enflurane, pancuronium, vecuronium

Example 3 – Methods for confirming effectiveness of drugs for treating lung adenocarcinoma characterized by neuroendocrine differentiation

Two cell lines are used to confirm the effectiveness of drugs for treating lung adenocarcinoma characterized by neuroendocrine differentiation. The first is the HCC1833 cell line, which was derived from lung AD and has high expression levels of ASCL1 and RET. The second is the A549 cell line which has low endogenous expression of ASCL1 and is stably transfected with ASCL1 (A549-As⁺). A549-As⁺ captures salient features of ASCL1⁺ lung AD from patients, including increased expression of RET. HCC1833 are stably transfected with ASCL1 siRNA to knock down ASCL1 and produce a HCC1833-AsKD cell line. Lowering ASCL1 expression leads to low levels of RET expression.

Selected drugs such as sunitinib, sorafenib, or others listed in Table 14 are incubated with A549 and A549-As⁺ cells *in vitro* and HCC1833 and HCC1833-AsKD cells *in vitro*. The cells are treated in culture at various concentrations (e.g., 10 to 100 nM or 2 to 10 μ M concentrations). The treated cell lines are examined for sensitivity to the selected drugs. Cell viability and apoptosis are assessed using standard assays to compare sensitivity of A549, A549-As⁺, HCC1833, and HCC1833-AsKD cells to the selected drugs.

In vivo methods are performed as follows. HCC1833 or A549-As⁺ cells are transplanted into Nude mice subcutaneously or by IP injections. Tumors are allowed to grow, and the animals are treated to receive daily treatments of a selected drug (e.g., sunitinib and/or sorafenib) given by oral administration at a particular dose (e.g., 30

mg/kg or 60 mg/kg). Tumor growth is evaluated twice-weekly by measurement of tumor volume, and histology of the tumors is assessed at the end of the treatment or after mice become moribund.

The HCC1833 adenocarcinoma cell line expressed high endogenous levels of ASCL1 and RET (Figure 13, left). Transfection with sh1 (a small interfering RNA construct that includes: GCCAACAAGAAGATGAGTAAG (SEQ ID NO:1)) or sh2 (a small interfering RNA construct that includes: CAACCGCGTCAAGTTGGTCAA (SEQ ID NO:2)) reduced ASCL1 expression as well as RET expression (Figure 13, left). These results suggest that ASCL1 is an upstream regulator of RET. In addition, STAT3 expression (in JAK/STAT3 pathway) went down. Knocking down ASCL1 expression also caused reduced cell proliferation (Figure 13, right). Also, HCC1833 cells with reduced ASCL1 expression (ASCL1-sh2 cells) exhibited a much slower ability to filling the gap in a scratch assay (Figure 14).

A549 cells expressed little ASCL1 and RET, while A549 cells transfected with ASCL1 lentivirus (A549-ASCL1 cells or A549-As⁺ cells) exhibited much more ASCL1 expression (Figure 13, center). Importantly, A549-As⁺ cells also expressed high levels of RET, again suggesting that ASCL1 is an up-stream regulator of RET. Also, the A549-As⁺ cells exhibited increased resistance to cisplatin-induced cytotoxicity (Figure 15). Furthermore, after cisplatin treatment, the remaining clonogenic potential in ASCL1 over-expressing cells was higher than in cells that did not express ASCL1. The effects of sunitinib, a tyrosine kinase inhibitor, also were examined. A549-As⁺ cells were more susceptible to sunitinib than the wild type A549 cells (Figure 16).

Example 4 – Treating lung adenocarcinoma characterized by neuroendocrine differentiation with brentuximab vedotin

A patient is identified as having lung adenocarcinoma characterized by neuroendocrine differentiation and is administered Brentuximab vedotin at a dose that is between 1.5 and 2.0 mg/kg (1.8 mg/kg) via intravenous infusion over 30 minutes every 3 weeks.

Example 5 – Treating lung adenocarcinoma characterized by neuroendocrine differentiation with sucralfate

5 A patient is identified as having lung adenocarcinoma characterized by neuroendocrine differentiation and is administered sucralfate at a dose of about 1 g (10 mL/2 teaspoonfuls) four times per day.

Example 6 – Treating lung adenocarcinoma characterized by neuroendocrine differentiation with paliperidone

10 A patient is identified as having lung adenocarcinoma characterized by neuroendocrine differentiation and is administered paliperidone at a dose of 6 to 12 mg daily orally.

OTHER EMBODIMENTS

15 It is to be understood that while the invention has been described in conjunction with the detailed description thereof, the foregoing description is intended to illustrate and not limit the scope of the invention, which is defined by the scope of the appended claims. Other aspects, advantages, and modifications are within the scope of the following claims.

WHAT IS CLAIMED IS:

1. A method for identifying a mammal as having lung adenocarcinoma characterized by neuroendocrine differentiation, wherein said method comprises determining whether
5 or not cancer cells from said mammal contain an elevated level of ASCL1 expression and an elevated level of RET expression, wherein the presence of said elevated level of ASCL1 expression and the presence of said elevated level of RET expression indicates that said mammal has lung adenocarcinoma characterized by neuroendocrine
10 differentiation, and wherein the absence of said elevated level of ASCL1 expression and the absence of said elevated level of RET expression indicates that said mammal does not have lung adenocarcinoma characterized by neuroendocrine differentiation.
2. The method of claim 1, wherein said mammal is a human.
- 15 3. The method of claim 1, wherein said elevated level is determined using PCR.
4. The method of claim 1, wherein said elevated level is determined using immunohistochemistry.
- 20 5. A method for identifying a mammal as having lung adenocarcinoma characterized by neuroendocrine differentiation, wherein said method comprises:
 - (a) determining whether or not a lung cancer cells from said mammal contain an elevated level of ASCL1 expression and an elevated level of RET expression,
 - (b) classifying said mammal as having lung adenocarcinoma characterized by
25 neuroendocrine differentiation if said sample contains said elevated level of ASCL1 expression and said elevated level of RET expression, and
 - (c) classifying said mammal as not having lung adenocarcinoma characterized by neuroendocrine differentiation if said sample lacks said elevated level of ASCL1
30 expression and said elevated level of RET expression.
6. The method of claim 5, wherein said mammal is a human.

7. The method of claim 5, wherein said elevated level is determined using PCR.

8. The method of claim 5, wherein said elevated level is determined using
5 immunohistochemistry.

9. A method for identifying a mammal as having lung adenocarcinoma characterized by neuroendocrine differentiation, wherein said method comprises:

(a) detecting the presence of an elevated level of ASCL1 expression and an
10 elevated level of RET expression in lung cancer cells from said mammal, and

(b) classifying said mammal as having lung adenocarcinoma characterized by neuroendocrine differentiation based at least in part on said presence of said elevated level of ASCL1 expression and said elevated level of RET expression.

15 10. The method of claim 9, wherein said mammal is a human.

11. The method of claim 9, wherein said elevated level is detecting using PCR.

12. The method of claim 9, wherein said elevated level is detecting using
20 immunohistochemistry.

13. A method for treating lung cancer, wherein said method comprises:

(a) detecting the presence of an elevated level of ASCL1 expression and an elevated level of RET expression in lung cancer cells from a mammal, and

25 (b) administering a molecule to said mammal under conditions wherein the number of lung cancer cells within said mammal is reduced, wherein said molecule is selected from the group consisting of sunitinib, vandetanib, riluzole, alteplase, anistreplase, tenecteplase, sucralfate, dasatinib, pazopanib, tivozanib, OSI-930, telatinib, tandutinib, imatinib, sorafenib, levodopa, carbidopa, entacapone orion, L-dopa, ABT-
30 089, mecamlamine, and succinylcholine.

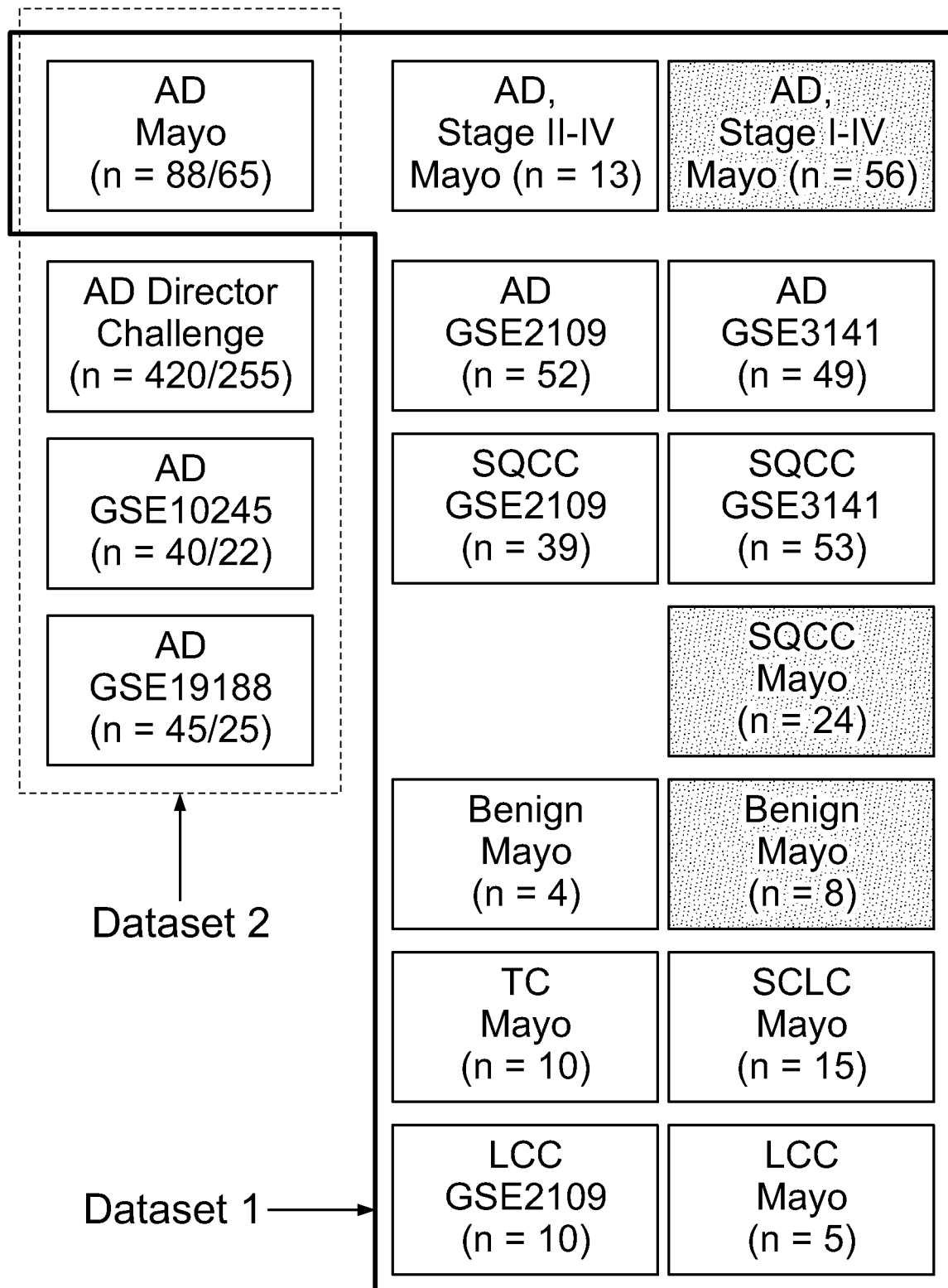


FIG. 1

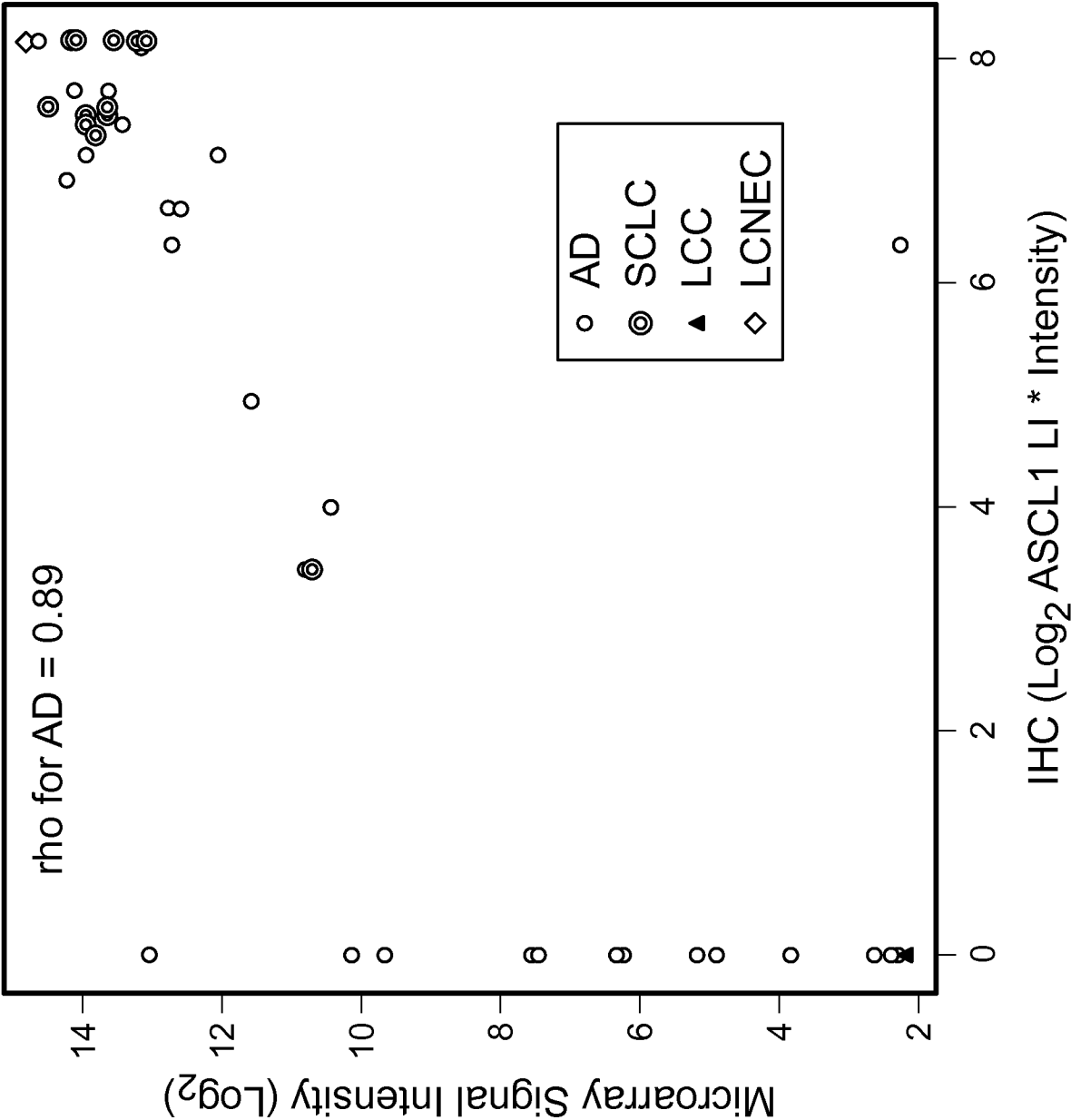


FIG. 2

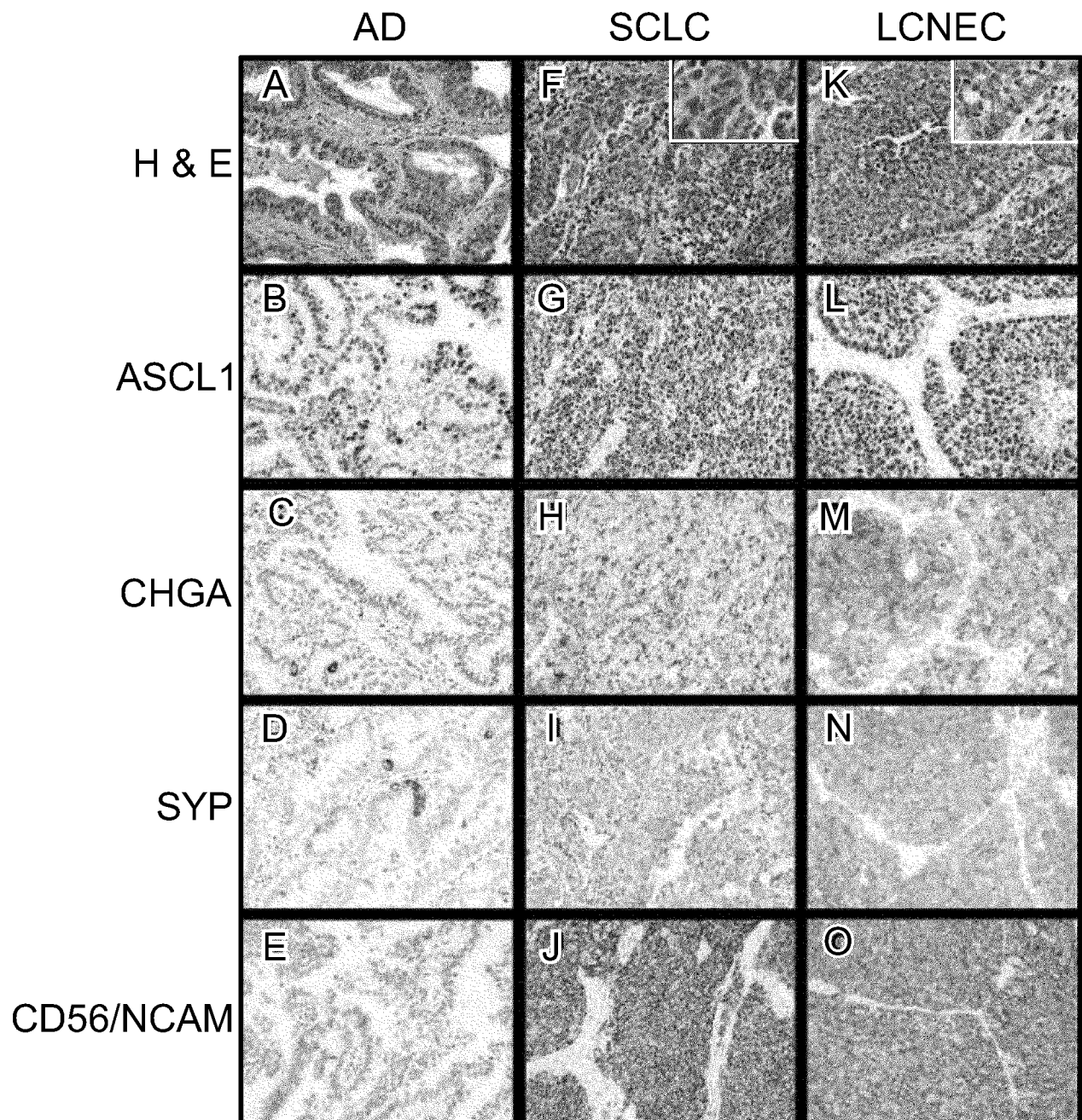


FIG. 3

4/20

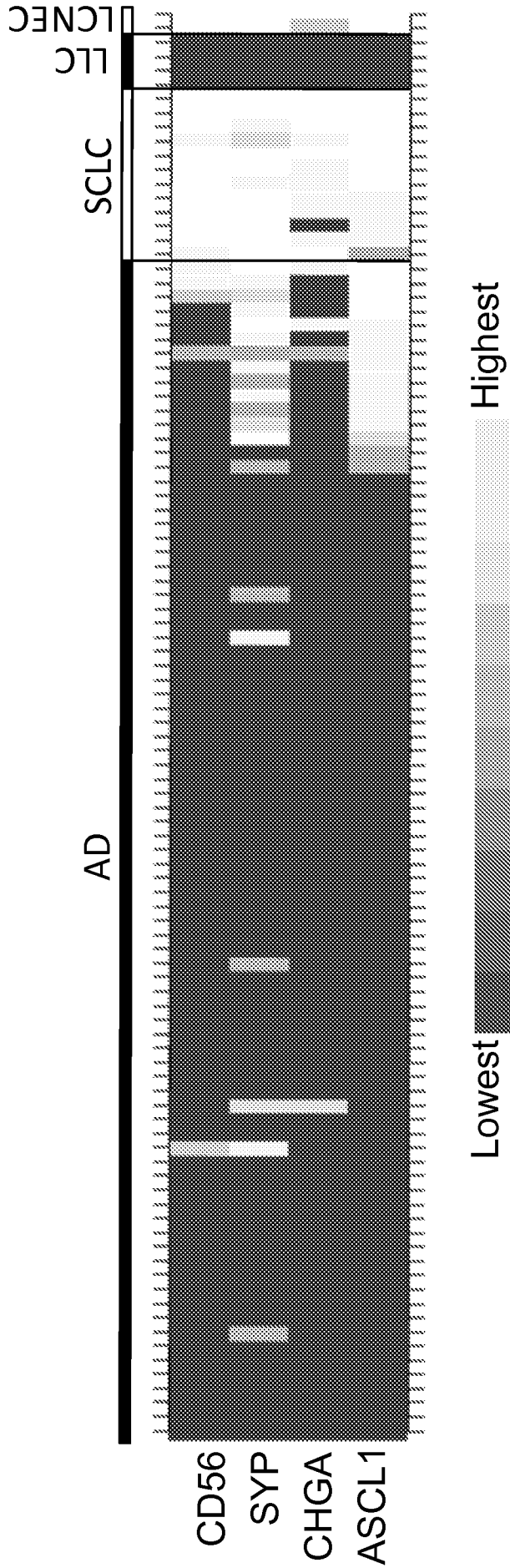


FIG. 3P

5/20

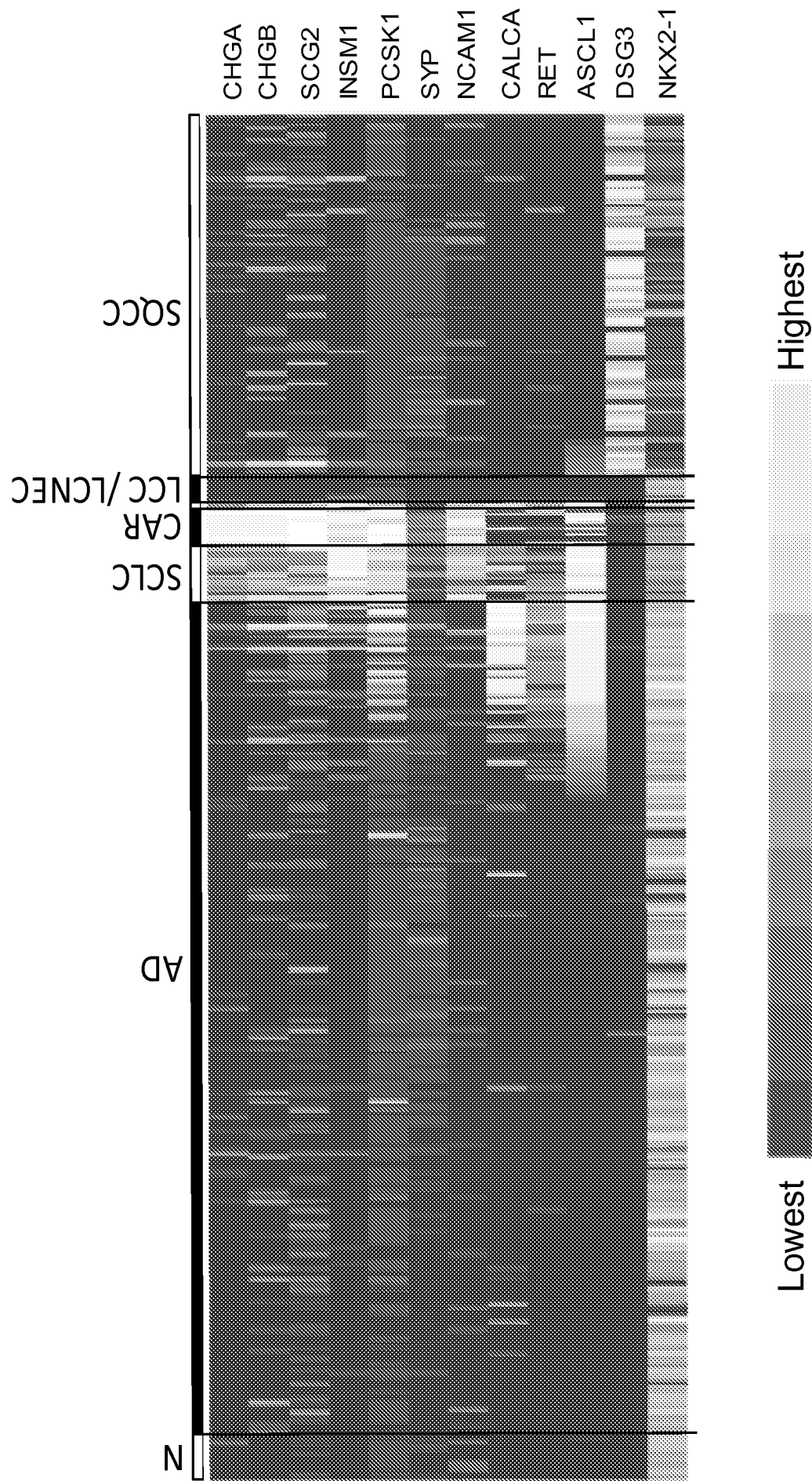


FIG. 4

6/20

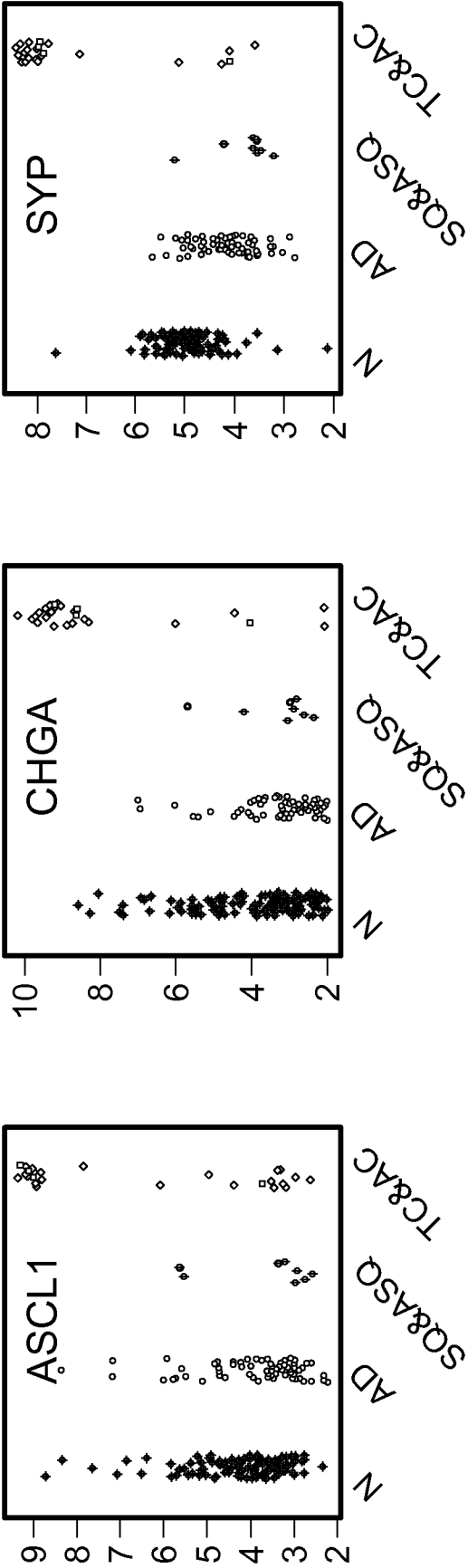
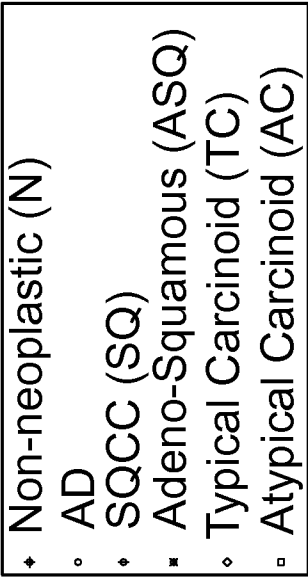


FIG. 5

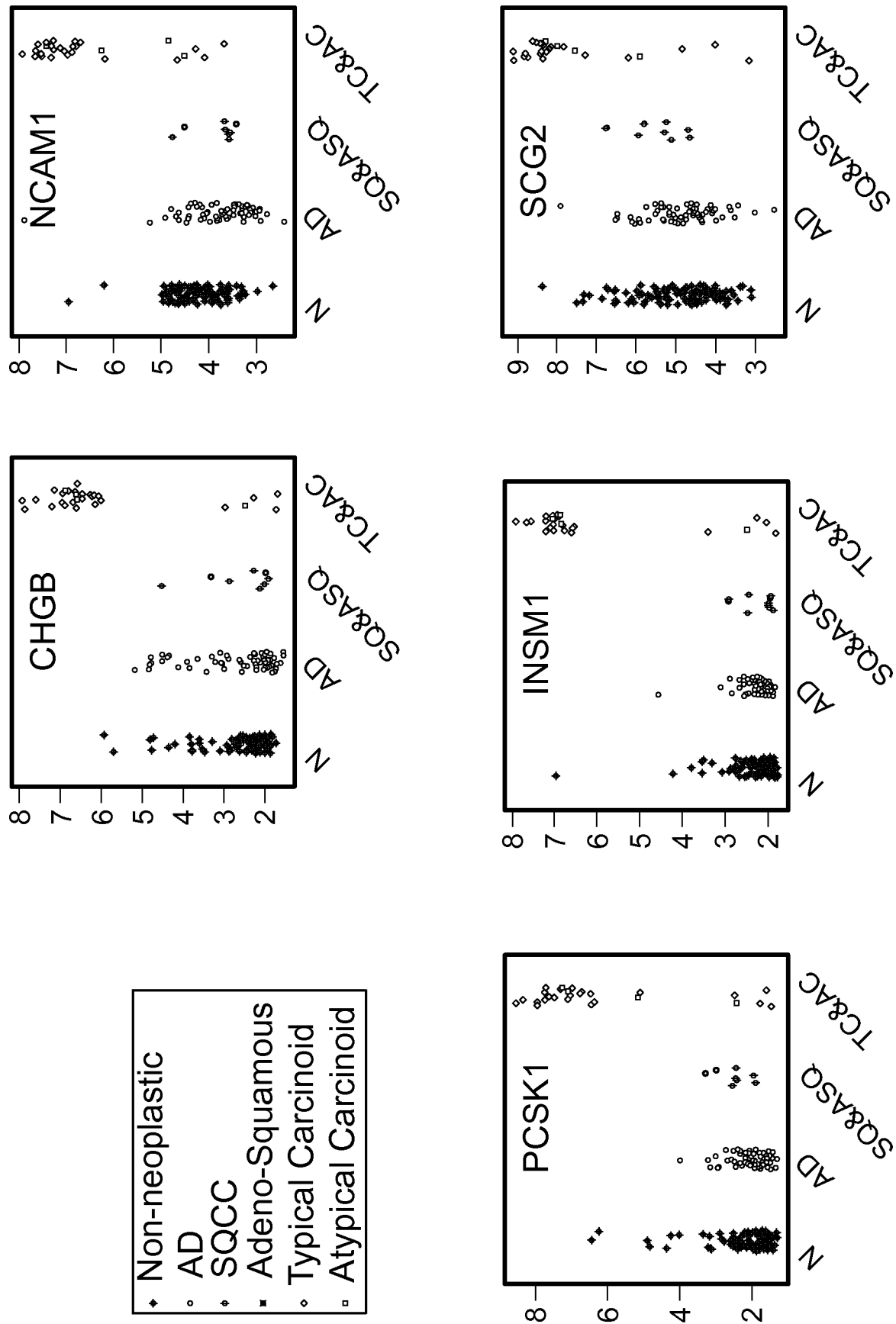


FIG. 5 (CONTINUED)

8/20

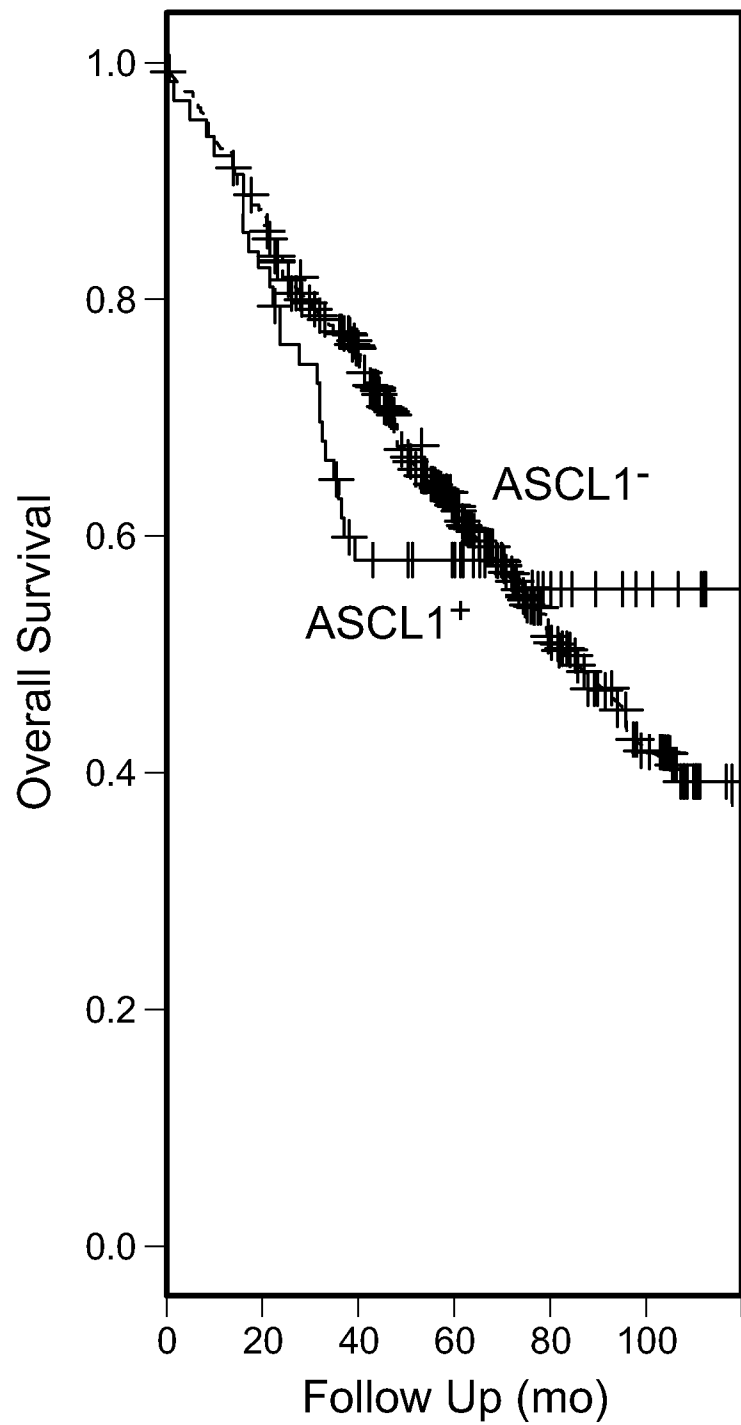


FIG. 6

9/20

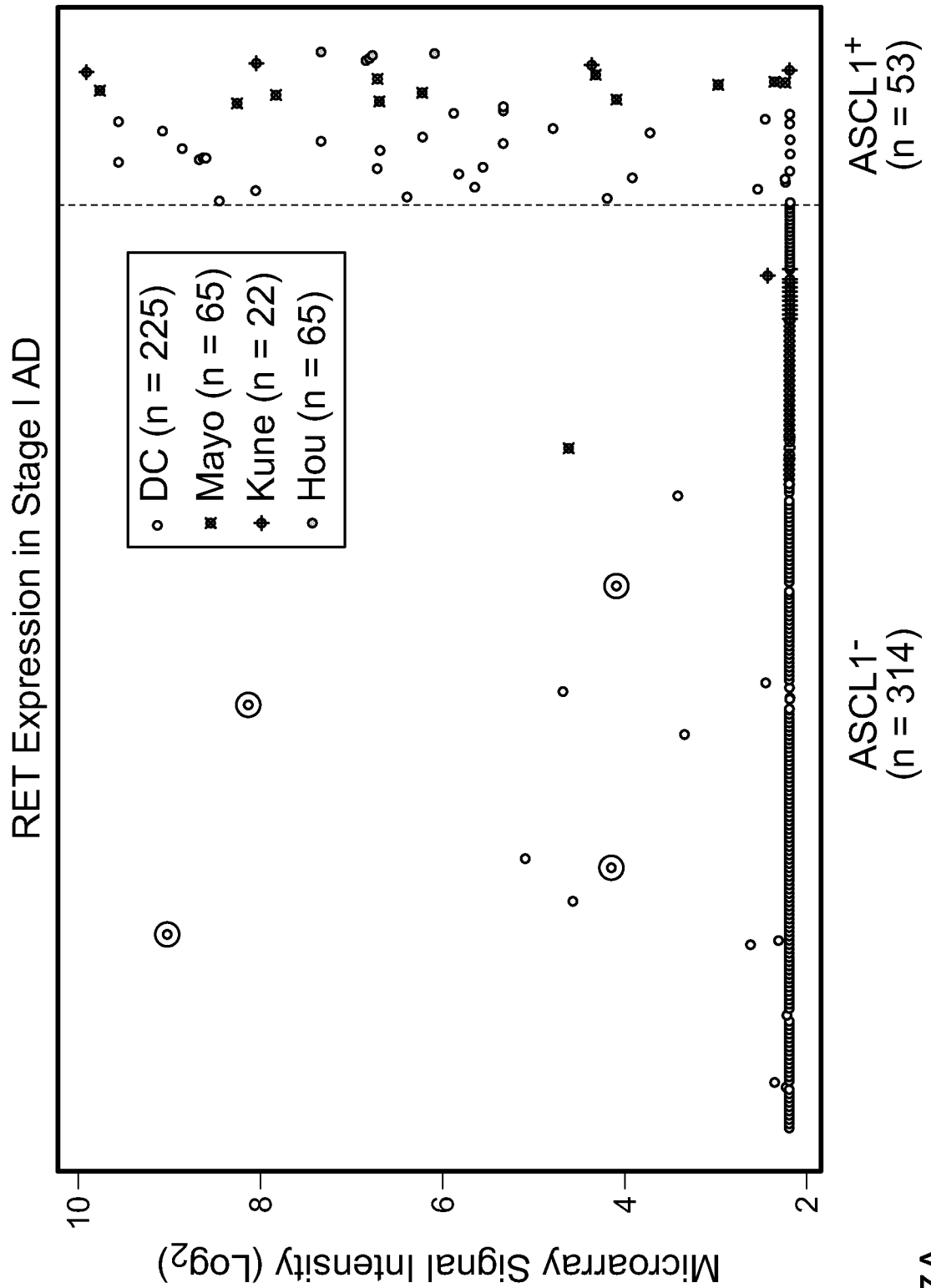


FIG. 7A

10/20

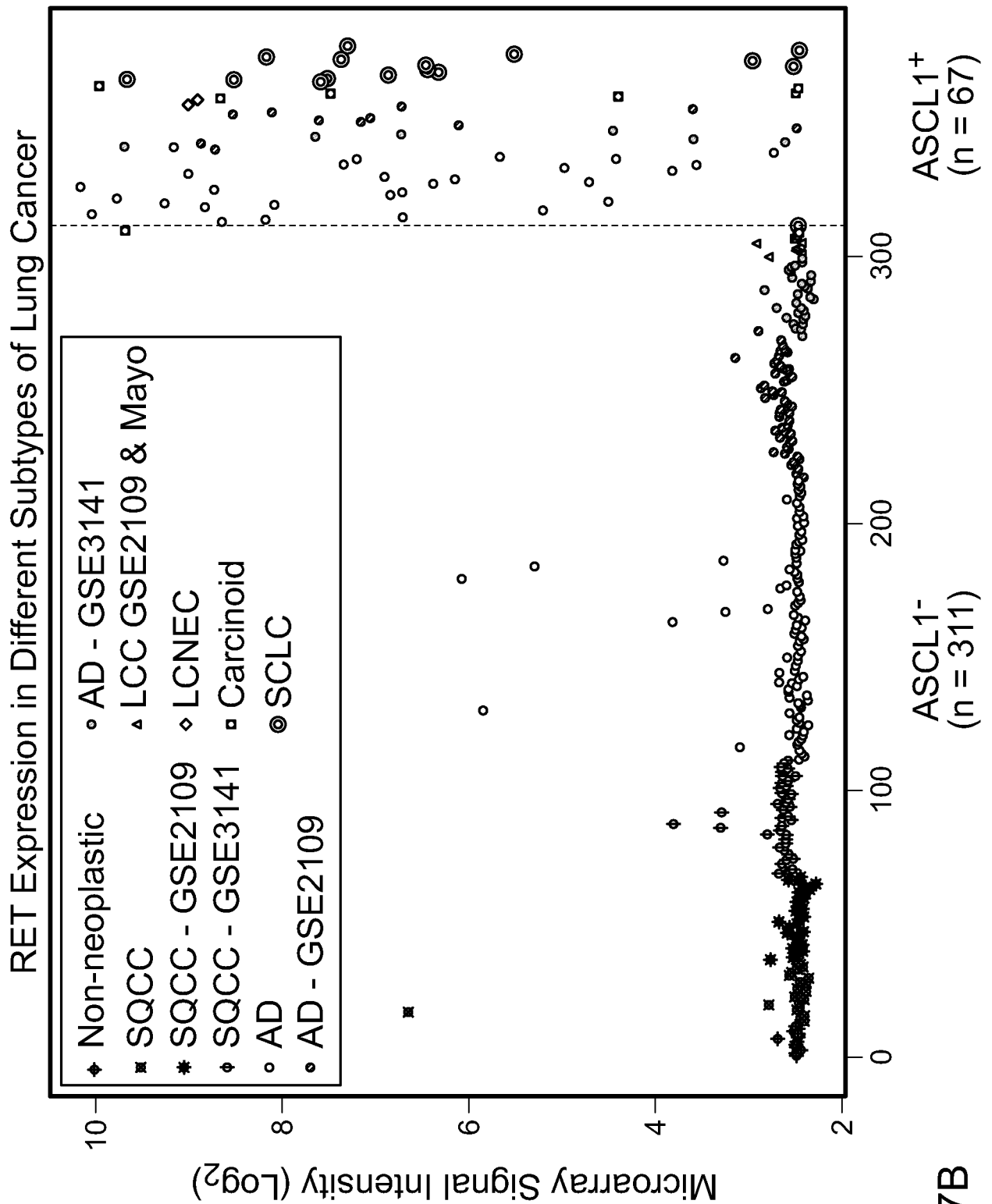


FIG. 7B

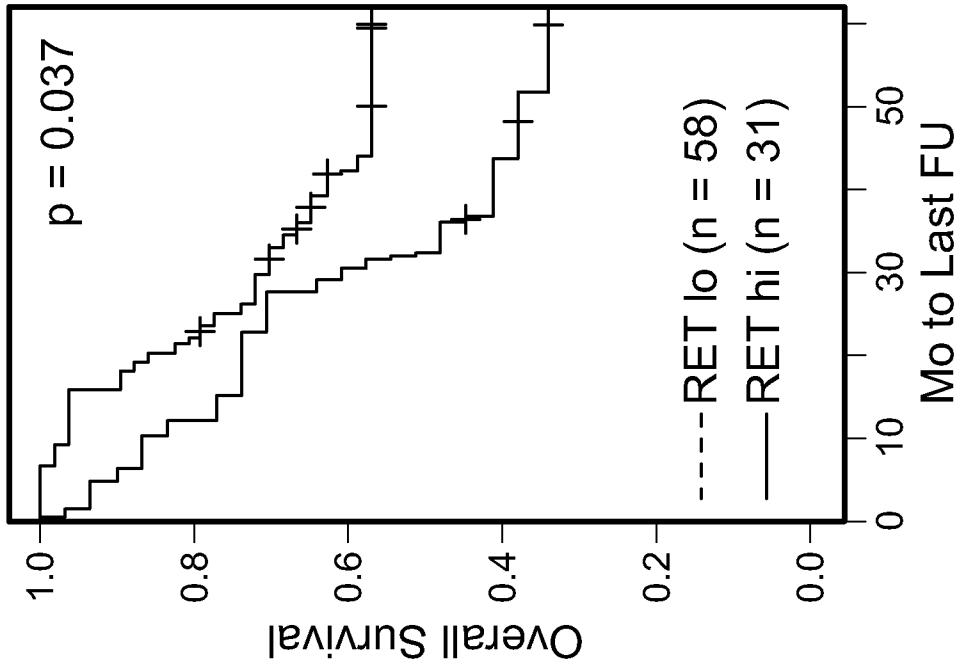


FIG. 8B

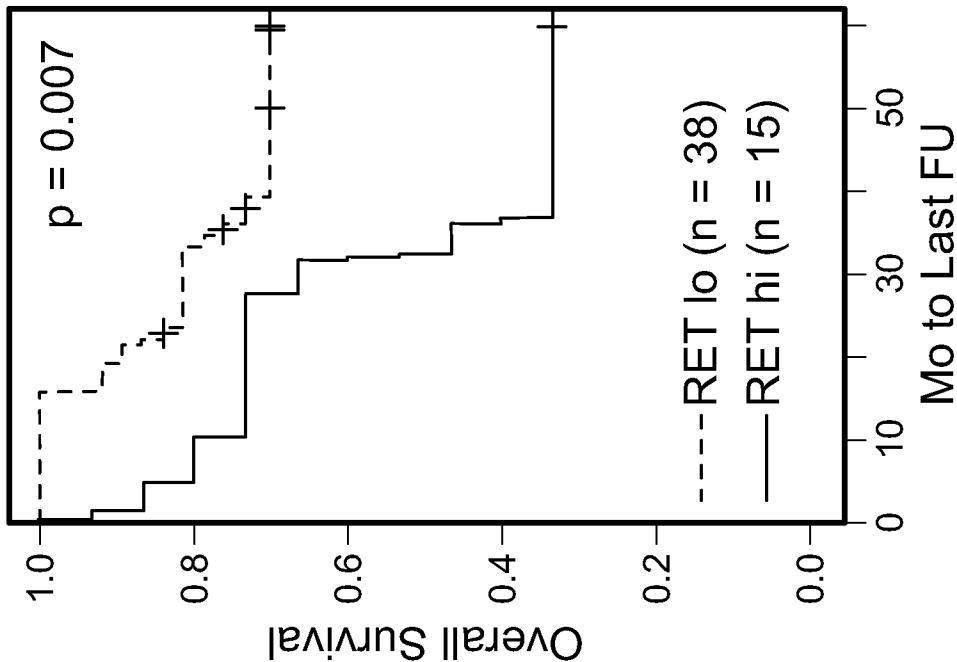


FIG. 8A

12/20

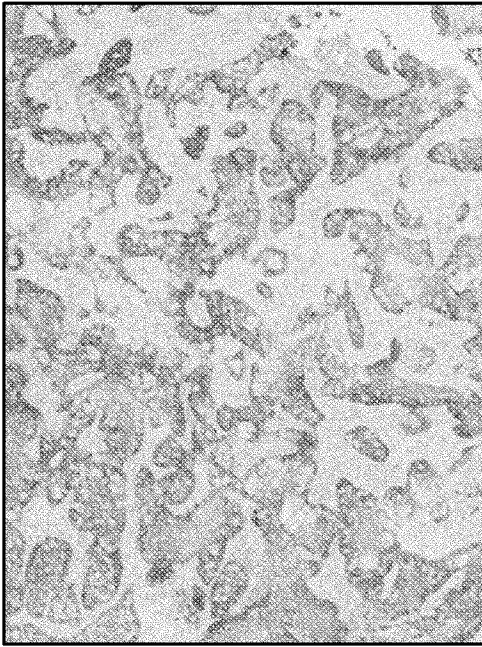


FIG. 9B

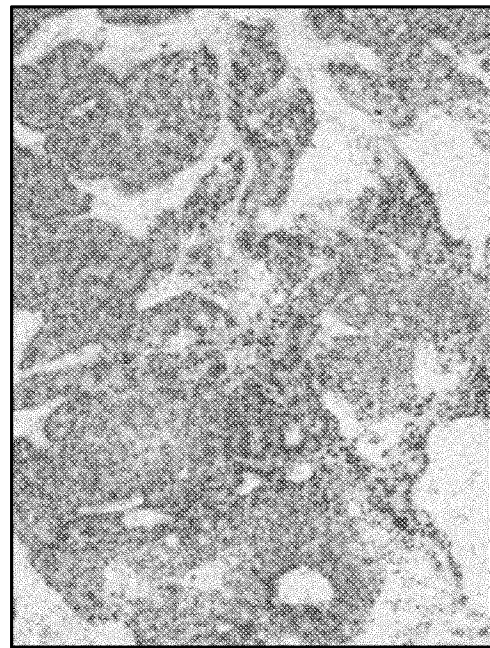


FIG. 9C

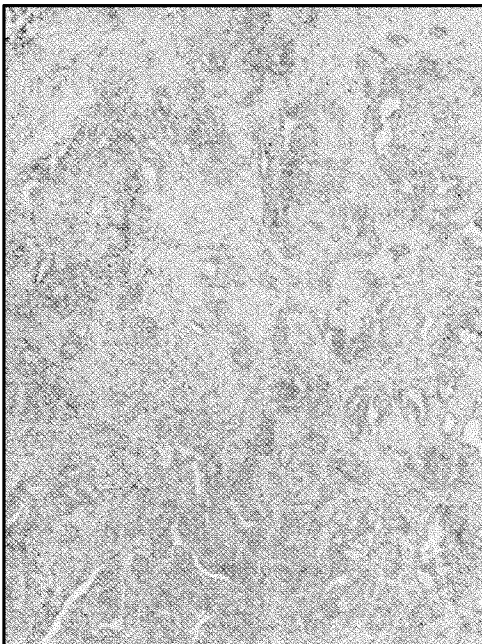


FIG. 9A

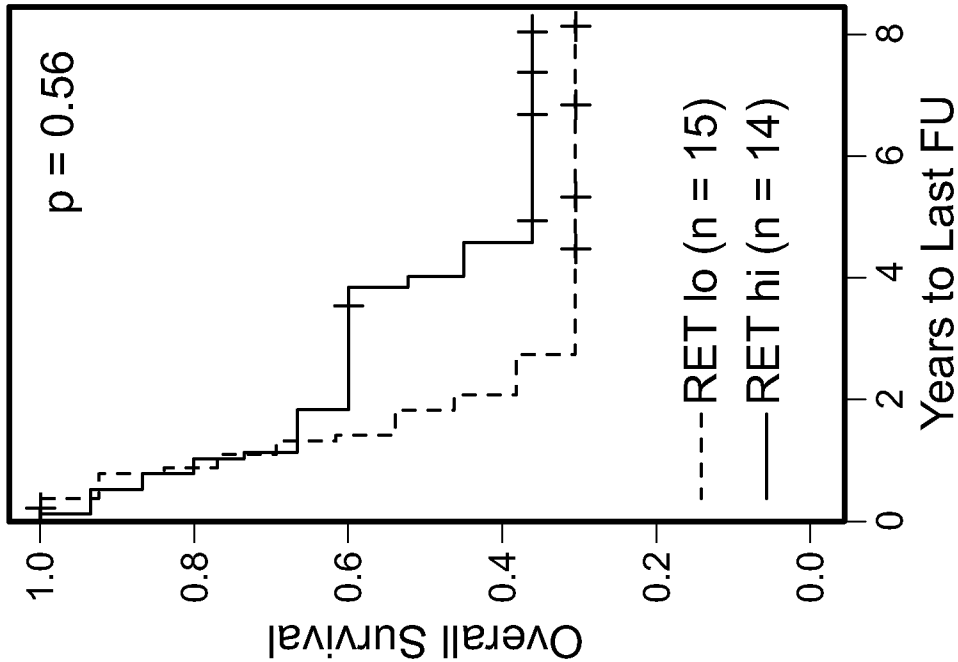


FIG. 9E

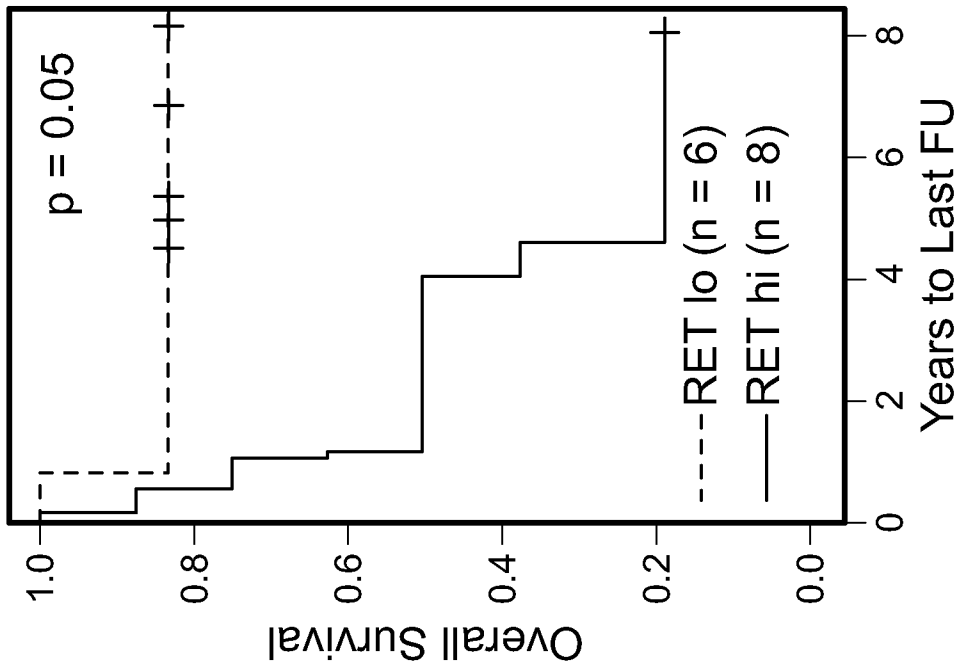


FIG. 9D

14/20

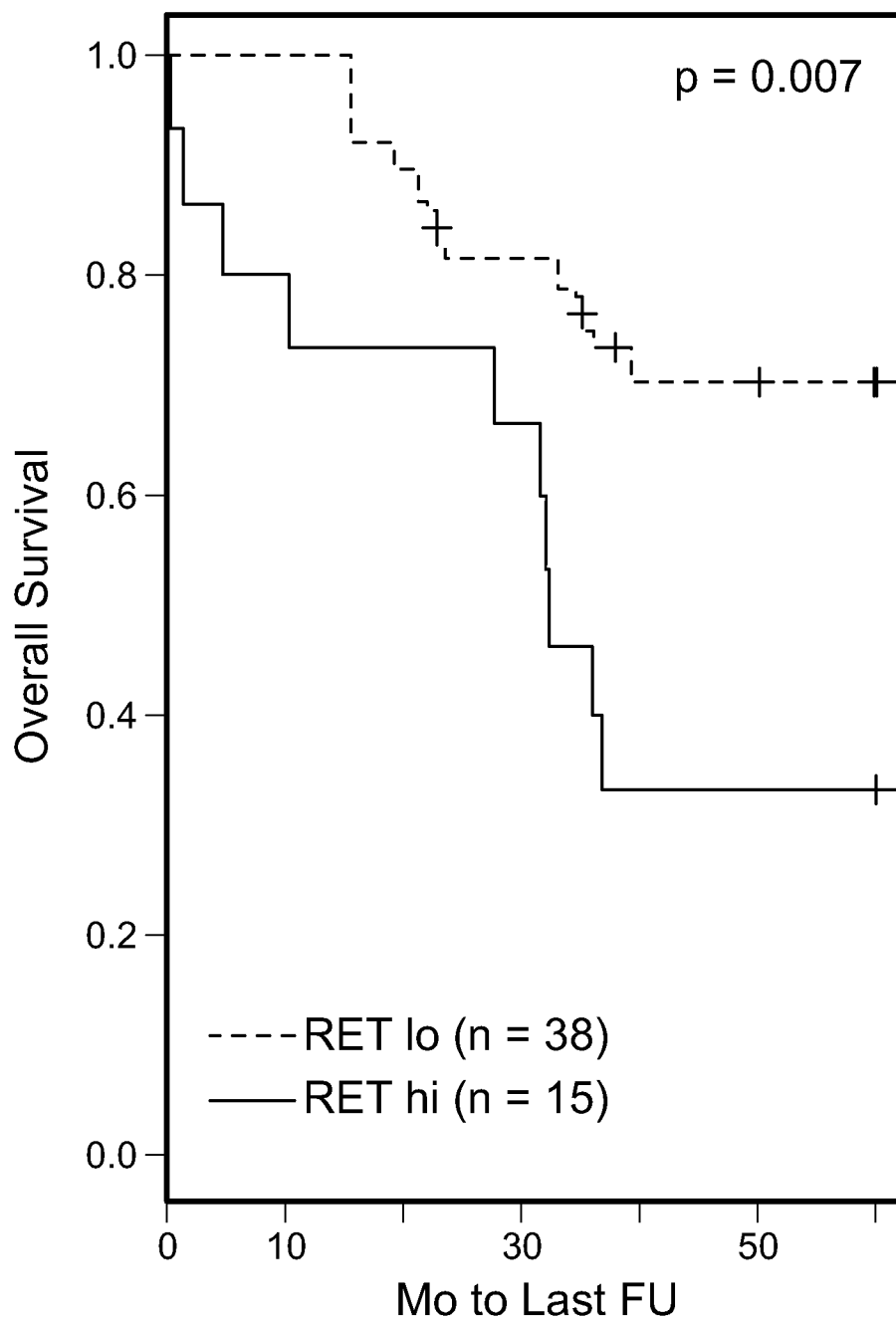
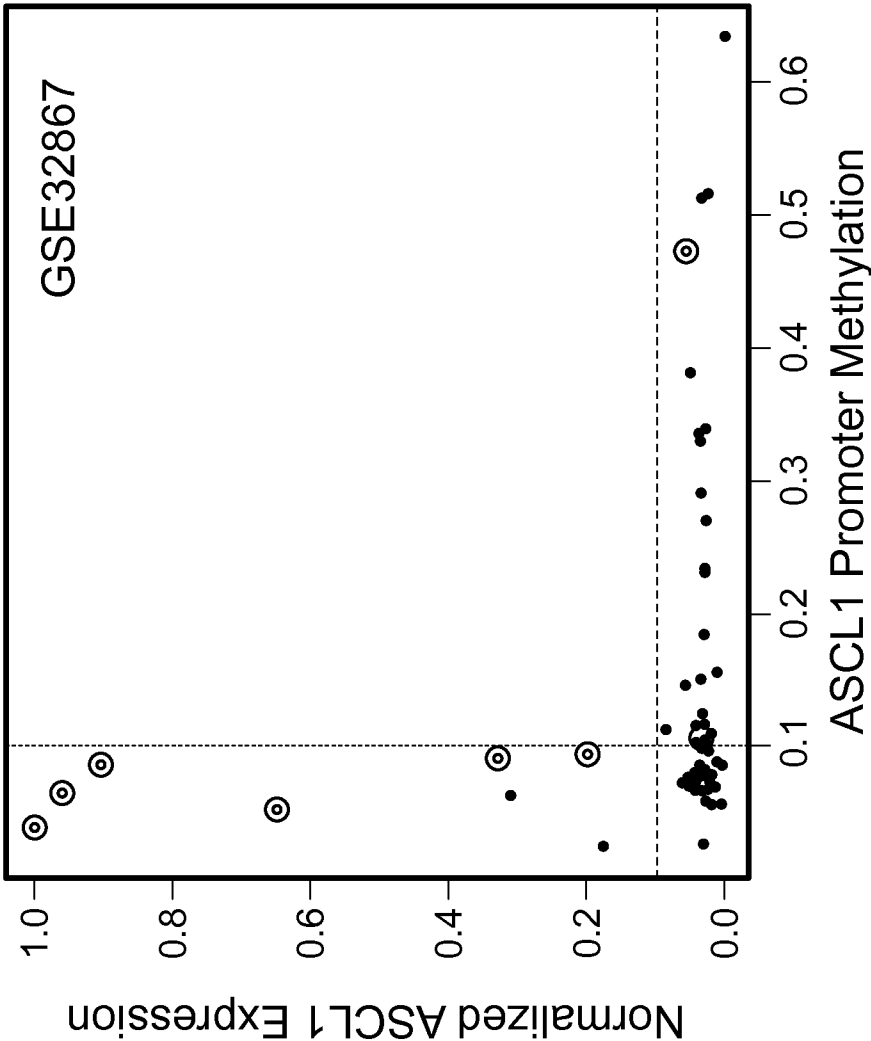


FIG. 10

ASCL1 and RET Expression According to the ASCL1 Promoter Methylation



Ret expression based on ILMN_1655610 probeset
Ascl1 expression based on the ILMN_1701653 probeset
Ascl1 promoter methylation based on cg20053158 probeset

FIG. 11

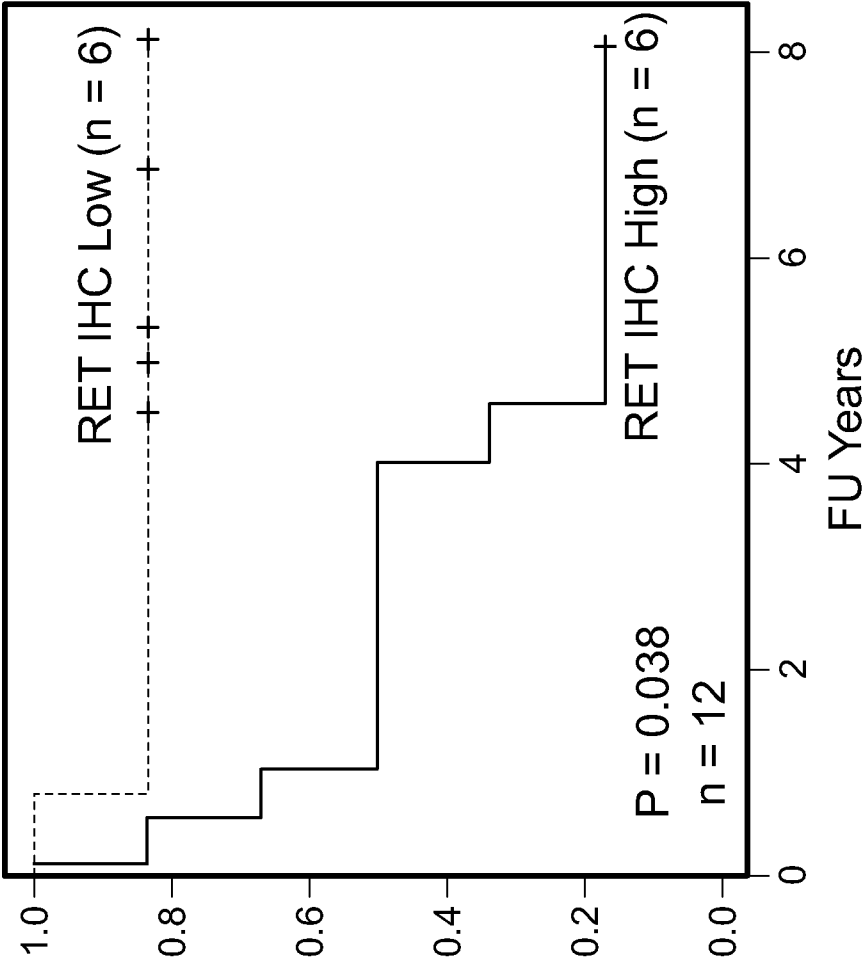


FIG. 12

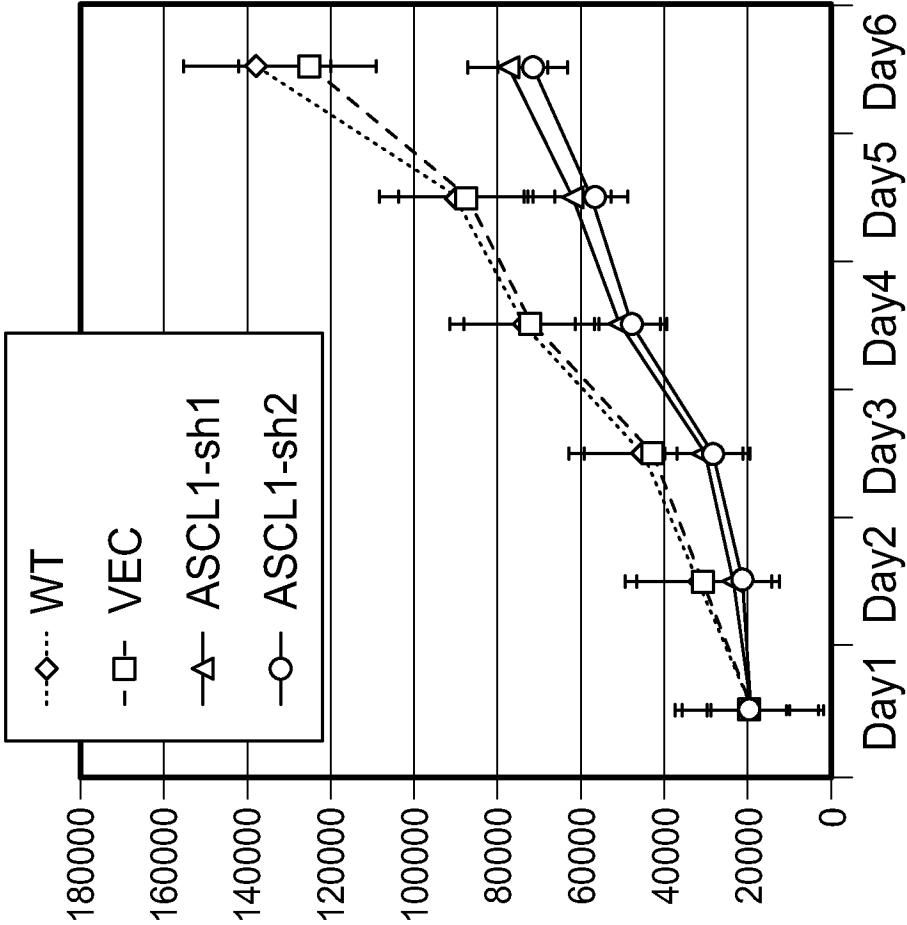
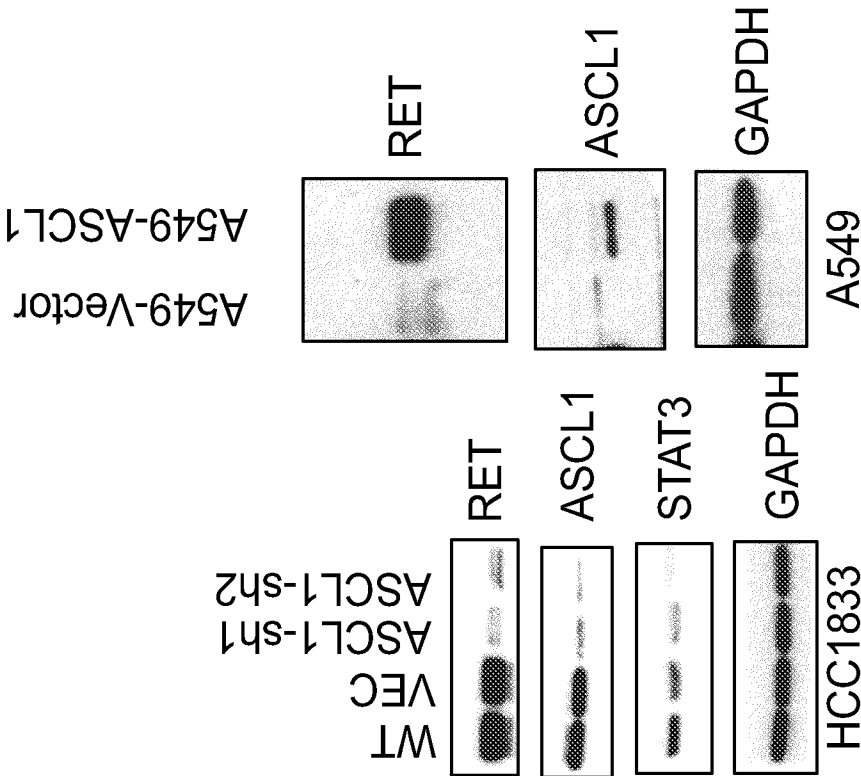


FIG. 13



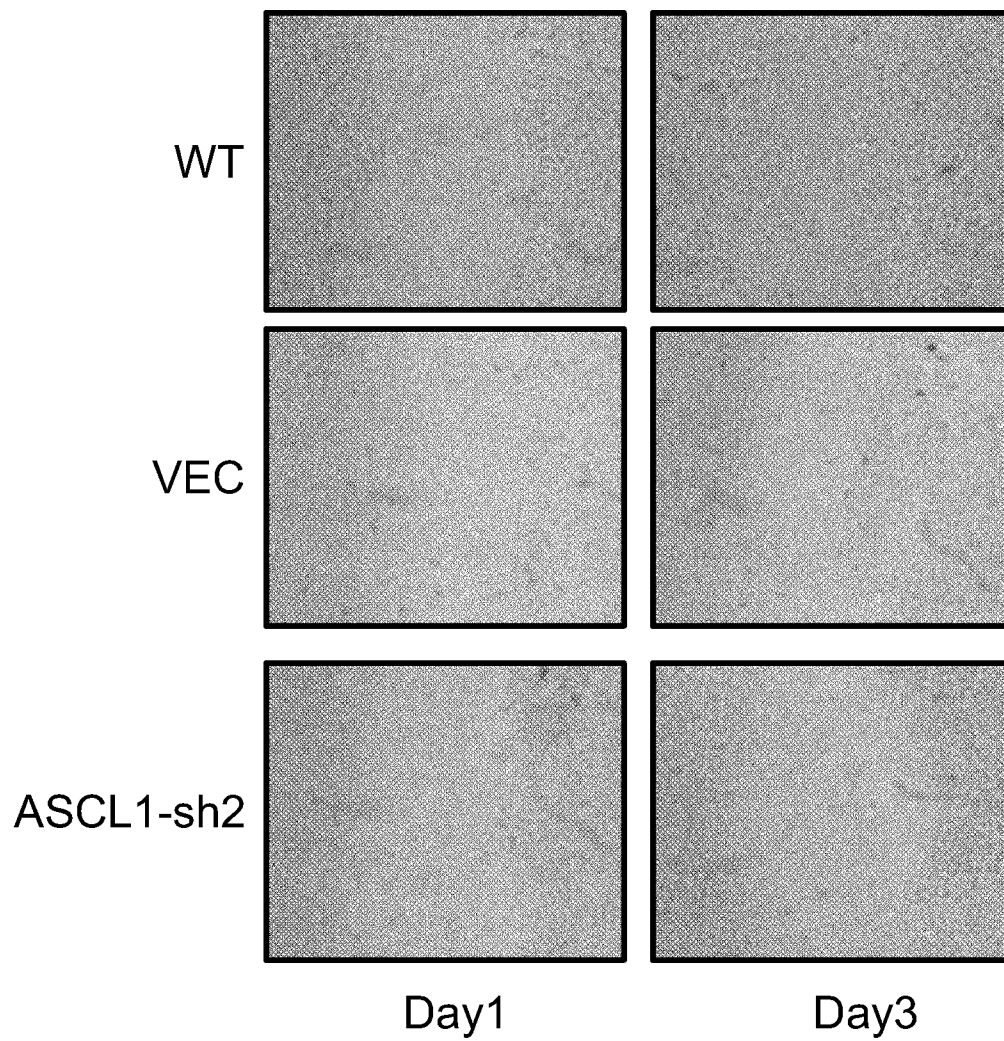


FIG. 14

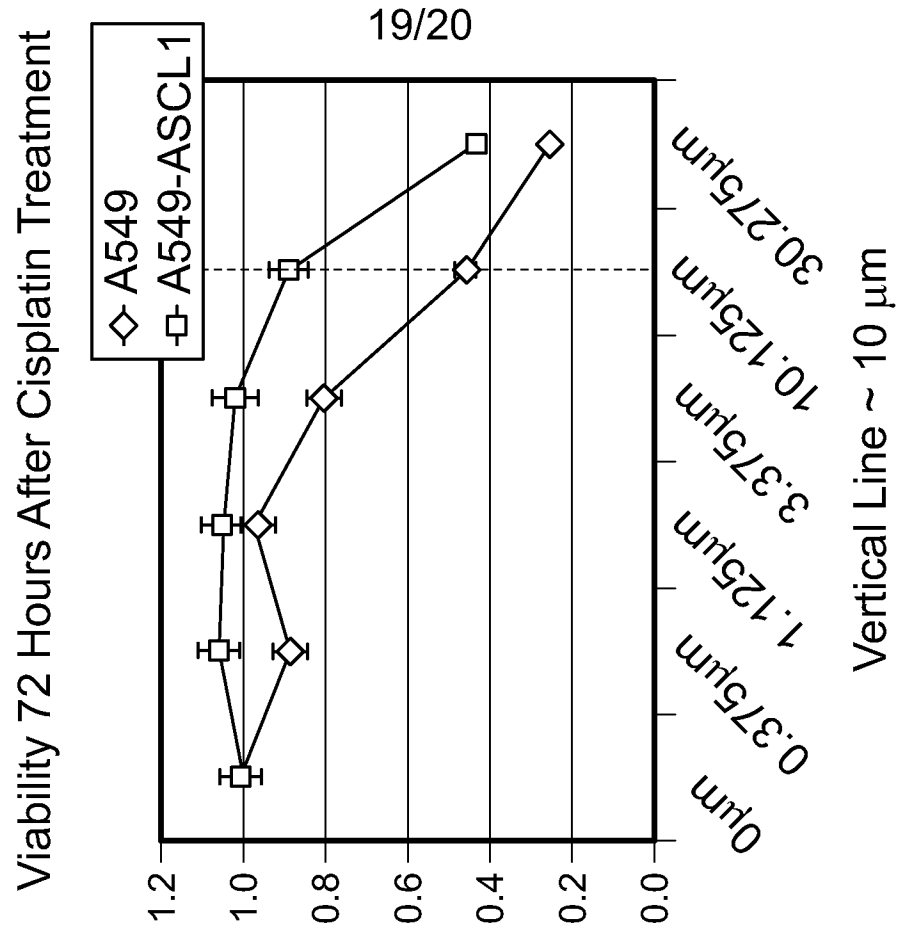
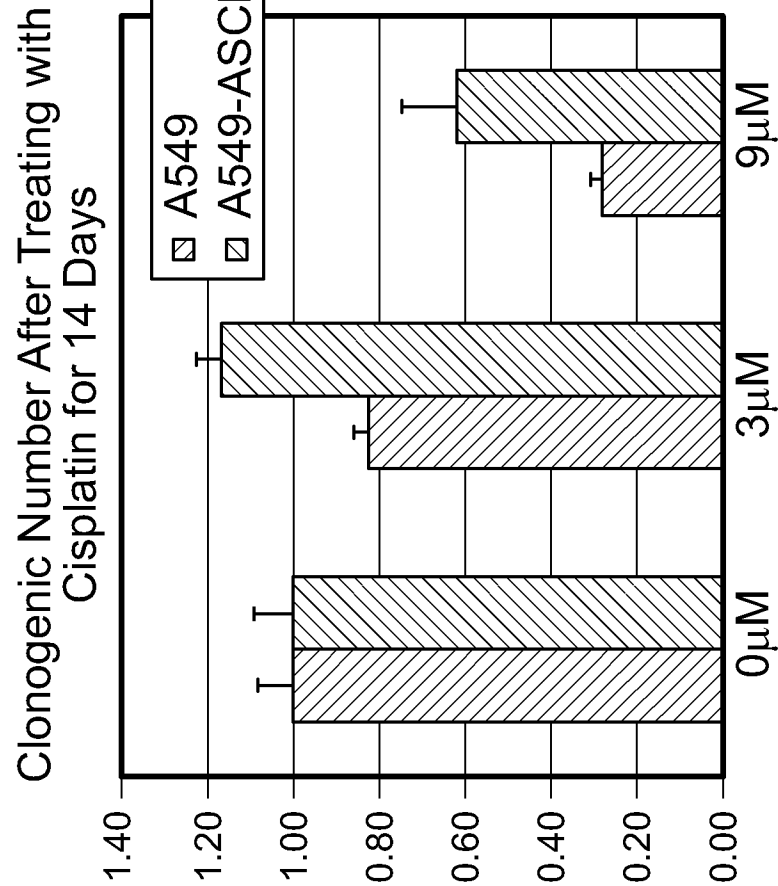


FIG. 15

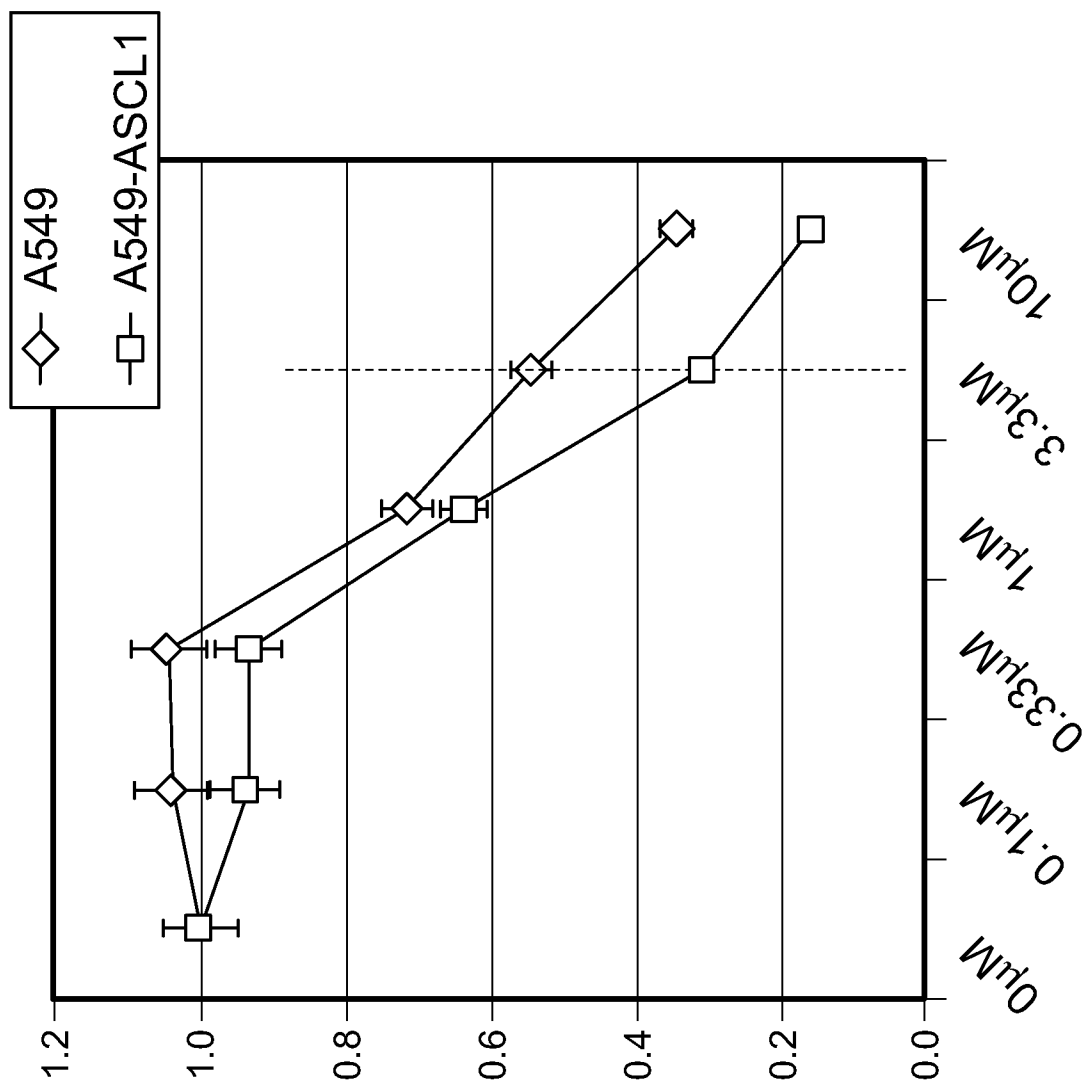


FIG. 16

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2014/022037

A. CLASSIFICATION OF SUBJECT MATTER

IPC(8) - G01N 33/574 (2014.01)

USPC - 435/7.23

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC(8) - C07K 14/435; C12Q 1/68; G01N 33/50, 33/53, 33/574 (2014.01)

USPC - 424/557; 435/6.1, 6.14, 7.23; 514/19.3; 536/24.5

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

CPC - C12Q 1/6886, 2600/00, 2600/118, 2600/158; G01N 33/57423 (2014.02)

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

PatBase, Google Patents, Google, PubMed

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	WO 2013/025952 A2 (CHAPMAN et al) 21 February 2013 (21.02.2013). entire document	1-13
Y	ENDOHO et al. 'Prognostic Model of Pulmonary Adenocarcinoma by Expression Profiling of Eight Genes As Determined by Quantitative Real-Time Reverse Transcriptase Polymerase Chain Reaction. J Clin Oncol. 22(5): 811-819. 01 March 2004. entire document	1-12
Y	ZAREBCZAN et al. 'Signaling Mechanisms in Neuroendocrine Tumors as Targets for Therapy.' Endocrinol Metab Clin North Am. 39(4): 801-810. December 2010. entire document	13
A	US 2011/0195072 A1 (BOULAY et al) 11 August 2011 (11.08.2011) entire document	1-13
A	KUNNIMALAIYAAN et al. 'Over Expression of the Notch1 Intracellular Domain (NICD) Inhibits Cellular Proliferation and Alters the Neuroendocrine Phenotype of Medullary Thyroid Cancer Cells.' J. Biol. Chem. 281: 1-23. 07 November 2006. entire document	1-13
A	WESTERMAN et al. 'Quantitative Reverse Transcription-Polymerase Chain Reaction Measurement of HASH1 (ASCL1), a Marker for Small Cell Lung Carcinomas with Neuroendocrine Features. Clin Cancer Res. 8: 1082-1086. 2002. entire document	1-13

☐ Further documents are listed in the continuation of Box C.


* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

13 May 2014

Date of mailing of the international search report

22 MAY 2014

Name and mailing address of the ISA/US

Mail Stop PCT, Attn: ISA/US, Commissioner for Patents
P.O. Box 1450, Alexandria, Virginia 22313-1450
Facsimile No. 571-273-3201

Authorized officer:

Blaine R. Copenheaver

PCT Helpdesk: 571-272-4300
PCT OSP: 571-272-7774

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2014/022037

Box No. 1 Nucleotide and/or amino acid sequence(s) (Continuation of item 1.c of the first sheet)

1. With regard to any nucleotide and/or amino acid sequence disclosed in the international application, the international search was carried out on the basis of a sequence listing filed or furnished:

a. (means)

☐

on paper

☒

in electronic form

b. (time)

☒

in the international application as filed

☐

together with the international application in electronic form

☐

subsequently to this Authority for the purposes of search

2. ☐ In addition, in the case that more than one version or copy of a sequence listing has been filed or furnished, the required statements that the information in the subsequent or additional copies is identical to that in the application as filed or does not go beyond the application as filed, as appropriate, were furnished.

3. Additional comments: