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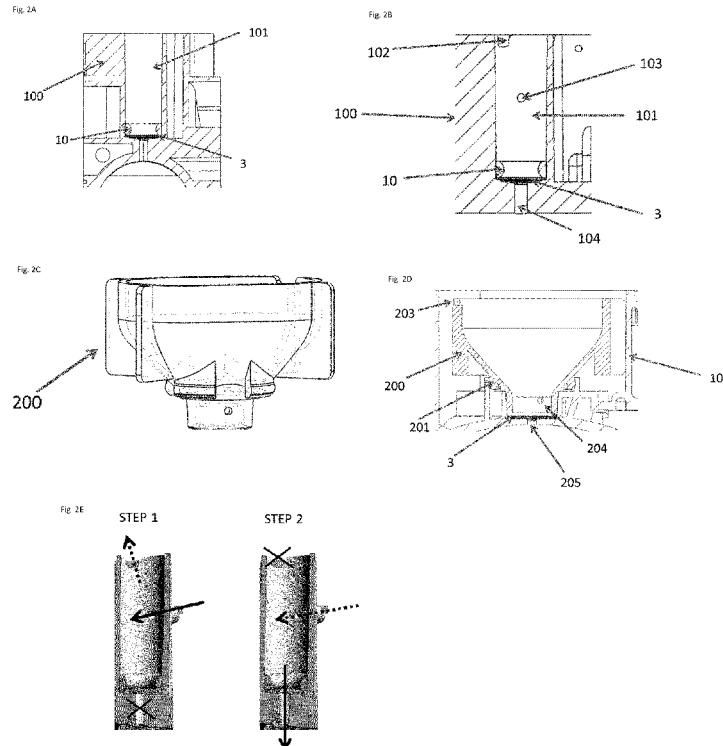
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(57) Abstract: A microfluidic device is disclosed having an enclosed chamber containing a filter for purifying biological or chemical analytes from a complex biological sample, said chamber housing a plurality of ports in addition to said filter, as follows: a first port enabling gas communication of the chamber with a vacuum generator, via a first flow path; a second port enabling liquid communication of the chamber with one or more reservoirs, via a second flow path; a third port enabling gas and liquid communication of the chamber with both one or more receiving containers and a vacuum generator, via a third flow path; and a filter located between the third port and both the first and second port, so that a fluid entering the chamber through the first and/or second port and exiting the chamber through the third port flows through the filter. The invention also relates to a method using the microfluidic device.

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NUCLEIC ACID PURIFICATION CARTRIDGE

Introduction

5 The original nucleic acid purification method based on the affinity of DNA and RNA for the silica surfaces (solid phase adsorption) was described by Boom et al. Nucleic acid attraction for silica surfaces is promoted by a high concentration of chaotropic salts (typically guanidine isothiocyanate or guanidine hydrochloride). The Boom method uses a chaotropic salt solution to denaturalize the biological sample and make it go through the filter using centrifugal forces to 10 promote DNA and RNA adsorption onto the silica surface. Once the nucleic acids are bound to the filter, one or more washes with ethanolic buffers are performed to get rid of the chaotropic salts and other biological impurities while keeping nucleic acids bound (chaotropic salts are disruptive for most nucleic acids in downstream applications). As a final step, after getting rid of ethanol (with a high speed spin), nucleic acids need to get rehydrated using an elution buffer (water or 15 low salt buffer). Rehydration promotes unbinding of the DNA and RNA from the silica surface and a final spin yields a solution where purified nucleic acids are resuspended.

Variations of this protocol have been described elsewhere, using either centrifugal force or 20 vacuum as a liquid flow driving force. However, all these methods are rather cumbersome and time-consuming, comprising several pipetting steps and the sequential application of different driving forces to control the flow of liquid, which usually produces a high variation of yield among 25 repeated purification processes.

For example, in a manual execution of the purification protocol by vacuum, the purification process consists of 5 main steps corresponding to the flow of 5 different liquids through the silica filter, i.e: sample mixture containing the nucleic acids, wash buffer 1 and wash buffer 2 to rinse 25 the filter and eliminate any amounts of contaminants, air to dry out the filter and eliminate any traces of volatile contaminants, and elution buffer to release the nucleic acids from the filter, so that the NAs are further available for downstream applications (such as qPCR amplification and detection). At the end of each step, the vacuum suction is maintained for one or two minutes even after the volume of liquid has flown through the filter In order to ensure that almost no 30 liquid is left within the filter crevices before the next liquid is pipetted.

The reproducibility of the yield of nucleic acids is dependent on the possibility to reproduce the contact time of the sample and buffers with the filter and the magnitude and distribution of liquid flow, which is dependent on the operator's skill.

There is a need for a purification device which provides reproducible purification results independent of the individual operator.

Object of the Invention

It is an object of the present invention to at least substantially satisfy the above need.

Summary

The present disclosure relates to a microfluidic device and a method for purifying biological or chemical analytes from a complex biological sample. Embodiments of the microfluidic device comprise a chamber wherein a filter is embedded, several reservoirs and valves. Embodiments of the device can be interfaced with external pumps that are operated by an automated instrument. Embodiments of the device and method herein thus provide an end-to-end automated implementation of the classical nucleic acid purification method.

In a first aspect, the present invention provides a microfluidic device comprising:

- (a) one or more reservoirs;
- (b) one or more receiving containers;
- (c) a purification cavity configured to be detachable from the microfluidic device, the purification cavity comprising:
 - (i) a first port enabling gas communication of the purification cavity with a vacuum generator, via a first flow path;
 - (ii) a second port enabling liquid communication of the purification cavity with the one or more reservoirs, via a second flow path; and
 - (iii) a third port enabling gas and liquid communication of the purification cavity with both the one or more receiving containers and a vacuum generator, via a third flow path; and
- (d) a filter configured to purify biological or chemical analytes from a complex biological sample and located over the third port such that fluid entering the purification cavity through the first and/or second port and exiting the purification cavity through the third port flows through the filter.

In a second aspect, the present invention provides a microfluidic device having an enclosed chamber containing a filter for purifying biological or chemical analytes from a complex biological sample, said chamber housing a plurality of ports in addition to said filter, as follows:

- a. a first port enabling gas communication of the chamber with a vacuum generator, via a first flow path;
- b. a second port enabling liquid communication of the chamber with one or more reservoirs, via a second flow path;
- c. a third port enabling gas and liquid communication of the chamber with both one or more receiving containers and a vacuum generator, via a third flow path; and
- d. a filter located between the third port and both the first and second port, so that a fluid entering the chamber through the first and/or second port and exiting the chamber through the third port flows through the filter.

There is disclosed herein a method of purifying a biological or chemical analyte from a complex biological sample, the method comprising the steps of: (a) allowing a liquid sample to enter the chamber through the second port by applying a negative pressure difference between the chamber and the first reservoir, while the valves within the flow paths are open for the first and second ports and closed for the third port; (b) allowing the sample to flow through the filter into a first receiving container by applying a negative pressure difference between said first receiving container and the chamber, while the valves within the flow paths are closed for the first port, vented to atmospheric pressure for the second port and open for the third port; and (c) eluting the analyte from the filter by applying a negative pressure difference between the chamber and one of the

receiving containers, while the valves within the flow paths are open for the first and second ports and closed for the third port.

Detailed Description

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Purification can in principle be based on any effect widely known in chromatography (e.g. displacement, affinity, cation exchange, anion exchange, size exclusion, reversed phase and normal phase) and its choice mainly depends on the analyte to be purified. Size exclusion is, however, less preferred than the other techniques, because a permanent binding cannot be 10 achieved in the case of the first technique. For the latter one may find conditions, under which the analyte to be purified is selectively bound to the medium, while ideally the other constituents of the sample pass through the medium without binding.

15 The microfluidic device of the present invention comprises an enclosed chamber containing a filter. A filter herein denotes a medium which differentially interacts with different constituents of a sample. In conventional chromatography such medium would usually be called a stationary phase. The differential interaction (also called partitioning) will cause differential retention times and thus a purification effect, if a sample is moved in a suitable buffer (in chromatography usually called mobile phase) through said medium.

20

25 The filter used in the herein disclosed device is one which is suited for purifying biological or chemical analytes from a complex biological sample. The analyte is the substance to be purified. A complex biological sample is a sample which comprises besides the analyte to the purified many different constituents of varying size and chemistry, such as proteins, nucleic acids, hormones, lipids, salts. A preferred sample is a cell lysate.

30 In a preferred embodiment the filter is made of or at least comprises silica. For example, the filter may be in the form of a silica membrane or a resin containing either silica beads or silica coated beads. Silica surfaces are useful for separating or purifying nucleic acids, in particular DNA. Silica is known to adsorb DNA molecules under certain salt and pH conditions and silica adsorption has become an important technique for purifying DNA.

In one embodiment of the invention the filter element is integrated in the purification cavity and fixated by a fixation ring. In a preferred embodiment the purification membrane filters are

inserted in a cavity, which is part of the body of the microfluidic device, and the membrane filter is held in position by a fixation ring that compresses it. (Figure 2 A and D)

In an alternative embodiment the purification cavity is a separate part which is assembled into the microfluidic device and eliminates the need for a fixation ring to keep the membrane filter in place. The purification cavity itself provides a clipping feature that fixes the cavity and the membrane filter in position, with the right compression of the membrane (Figure 2 B and C).

A separate cavity, which is hold in position, preferably by a clipping feature, has several advantages compared with cavities comprising a fixation ring:

A reproducible compression of the membrane filter is achieved by a clipping feature. This guarantees a reproducible compression, which generates a reproducible flow of liquids through the membrane filter and therefore a reproducible yield of purified nucleic acids or other purified analyte.

The correct positioning of the purification cavity into the microfluidic device is preferably guaranteed by its clipping feature, without needs to control the amount of compression, which is given by design. This facilitates manufacturing.

The purification cavity eliminates the need for a fixation ring, which results in reduced contamination of the sample. Fixation rings disrupt the fluidic path and gather remnants of liquids, which creates a contamination between buffers and results in a certain amount of contaminants in the final purified eluate, which can inhibit downstream analysis such as PCR. A detachable purification cavity creates a smooth transition on its walls which reduces the amount of contaminants that can be stuck to the wall, typically 5 to 10-fold

It is preferred that the analytes are nucleic acids. The term nucleic acid comprises mRNA (messenger RNA) in processed and unprocessed form, tRNA (transfer RNA), hn-RNA (heterogeneous nuclear RNA), rRNA (ribosomal RNA), LNA (locked nucleic acid), mtRNA

(mitochondrial RNA), nRNA (nuclear RNA), siRNA (short interfering RNA), snRNA (small nuclear RNA), snoRNA (small nucleolar RNA), scaRNA (Small Cajal Body specific RNA), microRNA, dsRNA (doubled-stranded RNA), ribozyme, riboswitch, viral RNA, dsDNA (double-stranded DNA), ssDNA (single-stranded DNA), plasmid DNA, cosmid DNA, chromosomal DNA, viral DNA, mtDNA
5 (mitochondrial DNA), nDNA (nuclear DNA), snDNA (small nuclear DNA) or the like or as well as all other conceivable nucleic acids.

The chamber houses a plurality of ports in addition to said filter, as follows: a first port enabling gas communication of the chamber with a vacuum generator, via a first flow path; a second port
10 enabling liquid communication of the chamber with one or more reservoirs, via a second flow path; a third port enabling gas and liquid communication of the chamber with both one or more receiving containers and a vacuum generator, via a third flow path.

The vacuum generator is located upstream of the chamber. The one or more reservoirs are also
15 located upstream of the chamber, but in another flow path than the vacuum generator. The one or more receiving containers are located downstream of the chamber. Further downstream of the receiving container(s) the vacuum generator of the third flow path is located.

The reservoir(s) usually comprise at least a reservoir containing the sample to be purified and
20 optionally one or more reservoirs comprising one or more washing buffers and/or an elution buffer and/or a regeneration buffer. The one or more containers usually comprise at least one container for receiving the analyte and optionally one or more containers for receiving other liquids, e.g. the flow through, washing buffer(s) and/or regeneration buffer(s).

25 A filter is located between the third port and both the first and second port, so that a fluid entering the chamber through the first and/or second port and exiting the chamber through the third port flows through the filter. Most conveniently the filter expands across the complete cross-section of the chamber. It is, however, not necessary that the medium fill up the full height of the chamber. Preferably, the filter is placed directly over the third port.

30

It is preferred that the device is a microfluidic cartridge. A cartridge means a consumable component which can be actuated by a larger unit through a suitable interface. Usually, the unit contains costly and/or durable elements or elements which are easy to clean, and a piece of software code to automate the control of the process. The unit may alternatively comprise

further elements for performing other processes upstream or downstream of the purification unit.

In one embodiment the device is disposable meaning that the device is designed for a single use after which it is disposed. In another embodiment the device is reusable which usually necessitates a regeneration of the device after each use.

The device may further comprise the valves ideally the vacuum generator is separate. The vacuum generator evacuates the chamber's pressure thereby generating a relative negative pressure.

Depending on the ports configuration (i.e. open or closed), a fluid is sucked from one of the reservoirs into the chamber and/or out of the chamber into one of the receiving containers. In a preferred embodiment the vacuum generator is a syringe pump or a diaphragm pump. In a further preferred embodiment, the vacuum can be applied to the first port and/or the third port with the same vacuum generator.

Known microfluidic devices do not include means for tracking the pressure in the system. The present invention preferably includes one or more pressure sensors. A Pressure sensor is preferably located within the third flow path upstream of the receiving containers. Another pressure sensor is preferably located within the first flow path downstream of the vacuum generator. The above pressure sensors may be used to determine the pressure drop caused by the filter which indicates the fluidic state of the filter. Thereby, one may determine (i) when a method step is completed thus minimizing time and buffers (e.g. when the filter is completely dry during a drying step; when the filter is sufficiently purged of liquid leftovers during the purging steps, which advantageously take place after the flow of each liquid and before the flow of the next one); (ii) if the liquids have flown completely through the filter, allowing the system to apply a 'just-in-time' increase in the suction pressure if there is an increased resistance to liquid flow due to the density and viscosity of the sample; (iii) if the filter is clogged; and (iv) the time required for each liquid to flow through the filter, which can be compared to a pre-determined threshold as a control for the purification method.

As described above, the herein disclosed device has three ports for communication with the chamber: a first port (gas outlet port), a second port (liquid inlet port) and a third port (liquid/gas outlet port). Each port can be individually opened, closed or vented to the atmosphere by means of a valve located within the respective flow path. Conveniently, multiport valves are used and, if

desired, two or three ports actuated with the same multiport valve. It is preferred that the dead volume enclosed by the third flow path between its corresponding valve and the filter is between 1uL and 10 mL. The controlled flow of liquids (including no-flow situation for a complete wetting of the filter) is achieved by applying a vacuum to the appropriate port and by opening and closing 5 the appropriate valves at each step. This confers a higher reproducibility to the device than the known devices independently of the biological sample type.

For example, in a conventional purification kit with a manual execution of the purification protocol by vacuum, the purification process consists of five main steps corresponding to the flow 10 of five different liquids through the filter, i.e. loading a sample mixture containing the nucleic acids, washing with wash buffer 1 and wash buffer 2 to rinse the filter and eliminate any amounts of contaminants, air drying the filter and eliminating any traces of volatile contaminants, and eluting to release the nucleic acids from the filter in order to be further available for downstream 15 applications (such as qPCR amplification and detection). At the end of each step, the vacuum suction is maintained for one or two minutes even after the volume of liquid has flown through the filter in order to ensure that almost no liquid is left within the filter crevices before the next liquid is pipetted, an operation that is referred to as 'purging'.

In order to achieve the same effect, the automated protocol comprises basically the same steps, 20 which are achieved by connecting each port to the appropriate reservoirs and applying the appropriate pressure differences at all times, by means of a pressure source (e.g. syringe or rotary pump), a set of valves, a set of microfluidic channels and a microcontroller using a software to automate all the steps.

25 The herein disclosed microfluidic device is particularly suited for being used in methods in which one or more analytes are to be separated from other constituents, i.e. in a purification method. Therefore, another object of the invention is a method of purifying a biological or chemical analyte from a complex biological sample using the microfluidic device described herein, the method comprising in this order the following steps: (a) allowing a liquid sample to enter the 30 chamber through the second port by applying a negative pressure difference between the chamber and the first reservoir, while the valves within the flow paths are open for the first and second ports and closed for the third port; (b) allowing the sample to flow through the filter into a first receiving container by applying a negative pressure difference between said first receiving container and the chamber, while the valves within the flow paths are closed for the first port,

vented to atmospheric pressure for the second port and open for the third port; and (c) eluting the analyte from the filter by applying a negative pressure difference between the chamber and one of the receiving containers, while the valves within the flow paths are open for the first and second ports and closed for the third port.

5

The pressure in step a may be generated by the vacuum generator located in the first flow path. The pressure in step b may be generated by the vacuum generator located in the third flow path.

The elution in step c may be carried out in detail as follows:

10

i) allowing an elution buffer contained in a third reservoir to enter the chamber through the second port by applying a negative pressure difference between the chamber and the third reservoir, while the valves within the flow paths are open for the first and second ports and closed for the third port. The pressure may be generated by the vacuum generator located in the first flow path; and

ii) allowing the elution buffer to be in contact with the filter for a predetermined time, while the valves within the flow paths are open for the first and second ports and closed for the third port. This step allows sufficient wetting of the filter in order to release the desired analyte; and

iii) allowing the elution buffer (containing the released analyte) to flow through the filter into a second receiving container by applying a negative pressure difference between said second receiving container and the chamber, while the valves within the flow paths are closed for the first port, vented to atmospheric pressure for the second port and open for the third port. The pressure may be generated by the vacuum generator of the third flow path.

25 Preferably, the method further comprises between steps a and b the step of allowing the sample to be in contact with the filter for a predetermined time, while the valves within the flow paths remain open for the first and second ports and closed for the third port.

The method may optionally comprise between steps b and c one or more of the following steps:

30 (i) cleaning and drying the filter for a predetermined time by applying a negative pressure between the third flow path and the chamber, said negative pressure difference being generated by the vacuum generator located in the third flow path, while the valves within the flow paths are closed for the first port, vented to atmospheric pressure for the second port and open for the third port; and/or

(ii) allowing a washing buffer located in a second reservoir to enter the chamber through the second port and to flow through the filter and into a receiving container by applying a negative pressure difference between said receiving reservoir and said second reservoir, while the valves within the fluidic paths are open for the second and third ports and closed for the first port. The pressure may be generated by the vacuum generator located in the third flow path; and/or

5 iii) allowing gas to flow through the filter for a predetermined time, by applying a negative pressure between the third flow path and the chamber, while the valves within the flow paths are closed for the first port, vented to atmospheric pressure for the second port and open for the third port. The gas displaces liquid and dries the filter. The pressure may be generated by the

10 vacuum generator located in the third flow path.

Preferably, the pressure difference is determined in order to determine when one or more valves are to be switched and, thus, the next method step is to be performed. A pressure drop across the filter indicates that valve(s) can be actuated to perform the next method step.

15

It is further preferred that the gas flow for drying the filter is applied only if the value of the first derivative of the pressure difference between the first and second pressure sensors is below a predefined threshold value.

20 Table 1 details one example of the method steps to be performed for a purification process by automatic means. Fluidic diagrams shown in the figures show the port/valve configuration for each of said steps.

Table 1. Detailed protocol for an example of a purification process.

SAMPLE	Step 1	FIGURE 3	Initial step_ Sample to purification filter
	Step 2	FIGURE 4	Sample to purification filter (plunger drive)
	Step 3	FIGURE 5	Pressure release
	Step 4	FIGURE 6	Plunger to home
	Step 5	FIGURE 7	Incubation of Sample
	Step 6	FIGURE 8	Sample from purification filter to waste/container nº13 (plunger drive)
	Step 7	FIGURE 5	Pressure release
	Step 8	FIGURE 6	Plunger to home
	Step 9	FIGURE 9	Purge
	Step 10	FIGURE 3	Wash 1 to purification filter
WASH 1 (valve nº7 → position 2; Reservoir nº16)	Step 11	FIGURE 4	Wash 1 to purification filter (plunger drive)
	Step 12	FIGURE 5	Pressure release
	Step 13	FIGURE 6	Plunger to home
	Step 14	FIGURE 7	Incubation of Wash Buffer 1
	Step 15	FIGURE 8	Wash 1 from purification filter to waste/container nº13

			(plunger drive)
Step 16	FIGURE 5	Pressure release	
Step 17	FIGURE 6	Plunger to home	
Step 18	FIGURE 9	Purge	
WASH 2 (valve n°7 → position 3; Reservoir n°17)	Step 19	FIGURE 3	Wash 2 to purification filter
	Step 20	FIGURE 4	Wash 2 to purification filter (plunger drive)
	Step 21	FIGURE 5	Pressure release
	Step 22	FIGURE 6	Plunger to home
	Step 23	FIGURE 7	Incubation of Wash Buffer 2
	Step 24	FIGURE 8	Wash 2 from purification filter to waste/container n°13 (plunger drive)
	Step 25	FIGURE 5	Pressure release
	Step 26	FIGURE 6	Plunger to home
	Step 27	FIGURE 9	Purge
DRYING	Step 28	FIGURE 10	Drying of the purification filter
ELUTION (valve n°7 → position 4; Reservoir n°18)	Step 29	FIGURE 3	Elution buffer to purification filter
	Step 30	FIGURE 4	Elution buffer to purification filter (plunger drive)
	Step 31	FIGURE 5	Pressure release
	Step 32	FIGURE 6	Plunger to home
ELUTION	Step 33	FIGURE 11	Incubation of Elution Buffer
	Step 34	FIGURE 12	Elution buffer from purification filter to container n°14 (plunger drive)

The invention has the advantage that whole automation can be achieved and no manual manipulation is needed for the purification process to be completed, thus achieving reproducible results. Further, the purification device/process can be linked to other upstream or downstream devices/processes (e.g., lysis, amplification and detection). In contrast with the prior art, liquids are moved into the chamber and onto the filter by means of a vacuum applied to the first port,

while the third port is closed. This prevents flow through the filter while allowing pre-wetting of the filter for a predetermined incubation time. Liquids are moved through the filter by means of a vacuum applied to the third port, after opening the valve of that port. Taken together, equal or higher nucleic acid yields are obtained as compared to known methods.

Figure captions

Preferred embodiments of the invention will be described hereinafter, by way of examples only, with reference to the accompanying drawings, wherein:

Fig. 1 shows a purification column (1) from a commercial kit for manual purification consisting of a plastic body (2) and a membrane filter (3) compressed and held in place by a fixation ring (10).

The plastic body comprises a liquid inlet (4) and a liquid outlet (5).

Fig. 2A shows part of a microfluidic device (100) with an integrated purification cavity (101), and a membrane filter (3) held in place by a fixation ring (10).

Figure 2B shows a rotated view for the microfluidic device (100) with an integrated purification cavity (101), and a membrane filter (3) held in place by a fixation ring (10). The gas port (102),

liquid inlet port (103) and outlet port (104) connected to the purification cavity (101) are also shown.

Fig. 2C shows a separate purification cavity (200).

Fig. 2D shows a purification cavity (200) assembled in a microfluidic device (100). The clipping feature (201) holds the purification cavity (200) in place and applies the right compression to the membrane filter (3). The gas port (203), liquid inlet port (204) and outlet port (205) are also shown.

Fig. 2E illustrates the port configuration and the flow direction during loading of liquid (STEP 1) and washing/elution (STEP 2). Solid arrow indicates liquid flow; dotted arrow indicates gas flow; X

10 indicates port closed by valve.

Figs. 3 to 11: Detailed fluidic diagrams showing the port/valve configuration for each of the steps detailed in table 1. It is to be understood that the device of the present invention may but does not necessarily comprise each of the elements shown in the figures. The description and/or claims

15 denote the essential elements. In addition to said elements one or more further optional elements may be independently chosen from each other. The optional elements are indicated in the following. 1: vacuum generator 1 (e.g. syringe pump); 2: vacuum generator 2 (optional, e.g. diaphragm pump); 3 - 7: valves (e.g. multiport valves); 8 and 9: pressure sensors (optional); 10: chamber; 11: fixation ring (optional); 12: filter; 13: waste receiving container (optional); 14: eluate receiving container; 15: sample reservoir; 16 - 17: reservoirs (optional); 18: elution buffer

20 reservoir.

CLAIMS

1. A microfluidic device comprising:
 - (a) one or more reservoirs upstream of a purification cavity;
 - (b) one or more waste receiving containers downstream of the purification cavity;
 - (c) the purification cavity defining an enclosed column with a tapered shape and hollow interior configured to be detachable from the microfluidic device, the purification cavity separate from and non-contiguous with any of the reservoirs or receiving containers, and comprising:
 - (i) a first port enabling gas communication of the purification cavity with a vacuum generator, via a first flow path;
 - (ii) a second port enabling liquid communication of the purification cavity with the one or more reservoirs, via a second flow path; and
 - (iii) a third port enabling gas and liquid communication of the purification cavity with both the one or more receiving containers and a vacuum generator, via a third flow path, wherein the first, second, and third ports are positioned on an exterior peripheral surface of the purification cavity; and
 - (d) a filter spanning a cross-section of the purification cavity orthogonal to the direction of liquid flow, configured to purify biological or chemical analytes from a complex biological sample and located adjacent to the third port such that fluid entering the purification cavity through the first and/or second port and exiting the purification cavity through the third port flows through the filter.
2. The microfluidic device of claim 1, wherein the microfluidic device is a microfluidic cartridge and is either disposable or reusable.
3. The microfluidic device of claims 1 or 2, wherein the analytes to be purified are nucleic acids.
4. The microfluidic device of any one of claims 1 to 3, wherein the filter comprises silica.
5. The microfluidic device of claim 4, wherein the filter comprises a silica membrane or a resin containing either silica beads or silica-coated beads.

6. The microfluidic device of any one of claims 1 to 5, wherein the purification cavity comprises a clipping feature configured to fix the position of the purification cavity in the microfluidic device.
7. The microfluidic device of any one of claims 1 to 6, wherein the vacuum generator is a syringe pump, a diaphragm pump or a combination of them.
8. The microfluidic device of any one of claims 1 to 7, wherein vacuum can be applied to the first port and/or third port with the same vacuum generator.
9. The microfluidic device of any one of claims 1 to 8, wherein a pressure sensor is located within the third flow path upstream of the receiving containers.
10. The microfluidic device of claim 9, wherein another pressure sensor is located within the first flow path between the purification cavity and the vacuum generator.
11. The microfluidic device of any one of claims 1 to 10, wherein the ports can be individually opened, closed or vented to the atmosphere by means of one or more multiport valves within their flow paths.
12. The microfluidic device of claim 11, wherein the dead volume enclosed by the third fluidic path between its corresponding valve and the filter is between 1 μ L and 10 mL.
13. A method of purifying a biological or chemical analyte from a complex biological sample using the microfluidic device according to any one of claims 1 to 12, the method comprising the steps of:
 - (a) allowing a liquid sample to enter the purification cavity through the second port by applying a negative pressure difference between the purification cavity and a first reservoir;
 - (b) allowing the sample to flow through the filter into a first receiving container by applying a negative pressure difference between said first receiving container and the purification cavity; and
 - (c) eluting the analyte from the filter by applying a negative pressure difference between the purification cavity and one of the receiving containers.

14. The method of claim 13, wherein the elution in step c is carried out as follows:
 - (i) allowing an elution buffer contained in a third reservoir to enter the purification cavity through the second port by applying a negative pressure difference between the purification cavity and the third reservoir;
 - (ii) allowing the elution buffer to be in contact with the filter for a predetermined incubation time; and
 - (iii) allowing the elution buffer to flow through the filter into a second receiving container by applying a negative pressure difference between said second receiving container and the purification cavity.
15. The method of claim 13 or 14 further comprising, between steps a and b, the step of allowing the sample to be in contact with the filter for a predetermined incubation time.
16. The method of any one of claims 13 to 15 further comprising, between steps b and c, one or more of the following steps:
 - (i) cleaning and drying the filter for a predetermined time by applying a negative pressure between the third flow path and the purification cavity; and/or
 - (ii) allowing a washing buffer located in a second reservoir to enter the purification cavity through the second port and to flow through the filter and into a receiving container by applying a negative pressure difference between said receiving reservoir and said second reservoir; and/or
 - (iii) allowing gas to flow through the filter for a predetermined time, by applying a negative pressure between the third flow path and the purification cavity.
17. The method of any one of claims 13 to 16, further comprising triggering one or more multiport valves within the first flow path, second flow path, and third flow path by a pressure difference.
18. A microfluidic device having an enclosed chamber containing a filter for purifying biological or chemical analytes from a complex biological sample, said chamber housing a plurality of ports in addition to the filter, as follows:
 - a. a first port enabling gas communication of the chamber with a vacuum generator, via a first flow path;

- b. a second port enabling liquid communication of the chamber with one or more reservoirs upstream of the chamber, via a second flow path;
- c. a third port enabling gas and liquid communication of the chamber with both one or more waste receiving containers downstream of the chamber and a vacuum generator, via a third flow path; wherein the first, second and third ports are positioned on an exterior peripheral surface of the chamber;
- d. the filter is located between the third port and both the first and second port, spanning a cross-section of the chamber orthogonal to the direction of liquid flow, adjacent to the third port, so that a fluid entering the chamber through the first and/or second port and exiting the chamber through the third port flows through the filter; and
- e. the chamber defines an enclosed column with a tapered shape and a hollow interior.

19. The device of claim 18, wherein the device is a microfluidic cartridge and either disposable or reusable.

20. The device of claims 18 or 19, wherein the analytes to be purified are nucleic acids.

21. The device of any one of claims 18 to 20, wherein the filter is silica, preferably a silica membrane or a resin containing either silica beads or silica coated beads.

22. The device of any one of claims 18 to 21, wherein the filter is integrated in a purification cavity and fixed by a fixation ring.

23. The device of any one of claims 18 to 21, wherein the purification cavity is a separate part, which is positioned in the device, thereby fixating said filter.

24. The device of any one of claims 18 to 23, wherein the vacuum generator is a syringe pump, a diaphragm pump or a combination of them.

25. The device of any one of claims 18 to 24, wherein vacuum can be applied to the first port and/or third port with the same vacuum generator.

26. The device of any one of claims 18 to 25, wherein a pressure sensor is located within the third flow path upstream of the receiving containers.

27. The device of claim 26, wherein another pressure sensor is located within the first flow path downstream of the vacuum generator.

28. The device of any one of claims 18 to 27, wherein the ports can be individually opened, closed or vented to the atmosphere by means of one or more multiport valves within their flow paths.

29. The device of claim 28, wherein the dead volume enclosed by the third fluidic path between its corresponding valve and the filter is between 1 uL and 10 mL.

30. A method of purifying a biological or chemical analyte from a complex biological sample using the microfluidic device according to claims 18 to 29, the method comprising the steps of:

- (a) allowing a liquid sample to enter the chamber through the second port by applying a negative pressure difference between the chamber and the first reservoir, while the valves within the flow paths are open for the first and second ports and closed for the third port;
- (b) allowing the sample to flow through the filter into a first receiving container by applying a negative pressure difference between said first receiving container and the chamber, while the valves within the flow paths are closed for the first port, vented to atmospheric pressure for the second port and open for the third port; and
- (c) eluting the analyte from the filter by applying a negative pressure difference between the chamber and one of the receiving containers, while the valves within the flow paths are open for the first and second ports and closed for the third port.

31. The method of claim 30, wherein the elution in step c is carried out as follows:

- i) allowing an elution buffer contained in a third reservoir to enter the chamber through the second port by applying a negative pressure difference between the chamber and the third reservoir, while the valves within the flow paths are open for the first and second ports and closed for the third port; and

- ii) allowing the elution buffer to be in contact with the filter for a predetermined incubation time, while the valves within the flow paths are open for the first and second ports and closed for the third port; and
- iii) allowing the elution buffer to flow through the filter into a second receiving container by applying a negative pressure difference between said second receiving container and the chamber, while the valves within the flow paths are closed for the first port, vented to atmospheric pressure for the second port and open for the third port.

32. The method of claim 30 or 31 further comprising between steps a and b the step of allowing the sample to be in contact with the filter for a predetermined incubation time, while the valves within the flow paths are open for the first and second ports and closed for the third port.

33. The method of any one of claims 30 to 32 further comprising between steps b and c one or more of the following steps:

- (i) cleaning and drying the filter for a predetermined time by applying a negative pressure between the third flow path and the chamber, while the valves within the flow paths are closed for the first port, vented to atmospheric pressure for the second port and open for the third port; and/or
- (ii) allowing a washing buffer located in a second reservoir to enter the chamber through the second port and to flow through the filter and into a receiving container by applying a negative pressure difference between said receiving reservoir and said second reservoir, while the valves within the flow paths are open for the second and third ports and closed for the first port; and/or
- (iii) allowing gas to flow through the filter for a predetermined time, by applying a negative pressure between the third flow path and the chamber, while the valves within the flow paths are closed for the first port, vented to atmospheric pressure for the second port and open for the third port.

34. The method of any one of claims 30 to 33, wherein one or more of the valves are triggered by the pressure difference.

STAT-Diagnostica & Innovation SL
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SPRUSON & FERGUSON

14 Jan 2021

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Figures

Fig. 1:

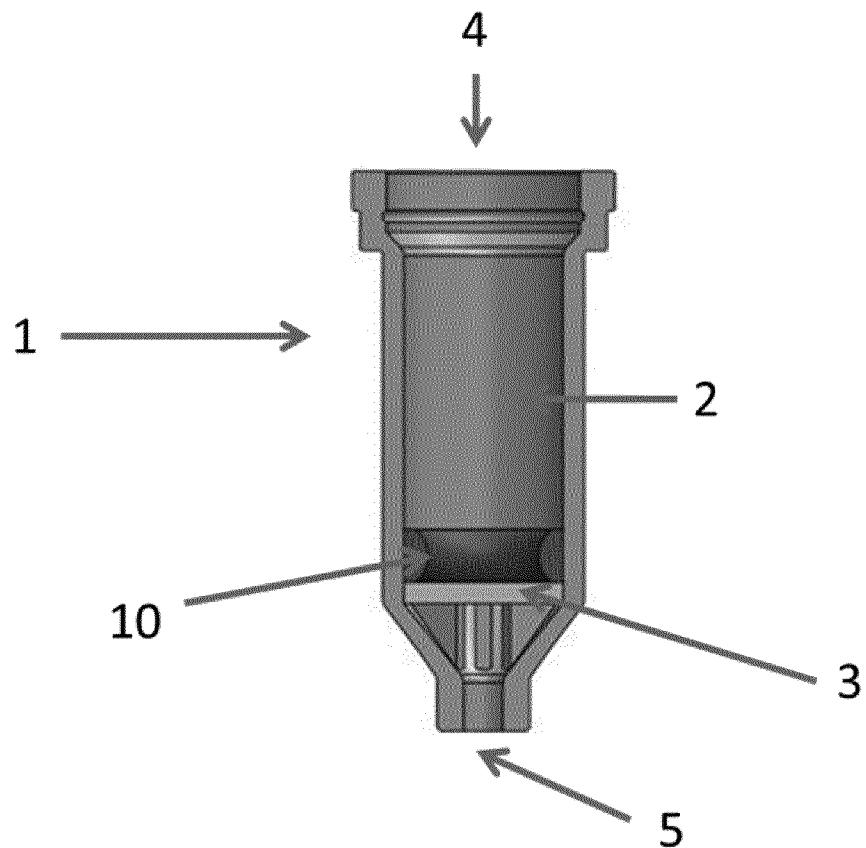


Fig. 2A

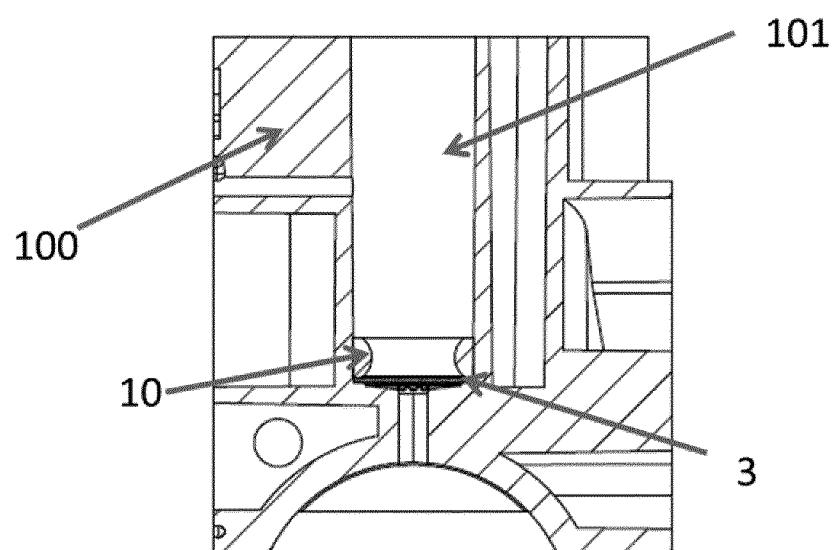


Fig. 2B

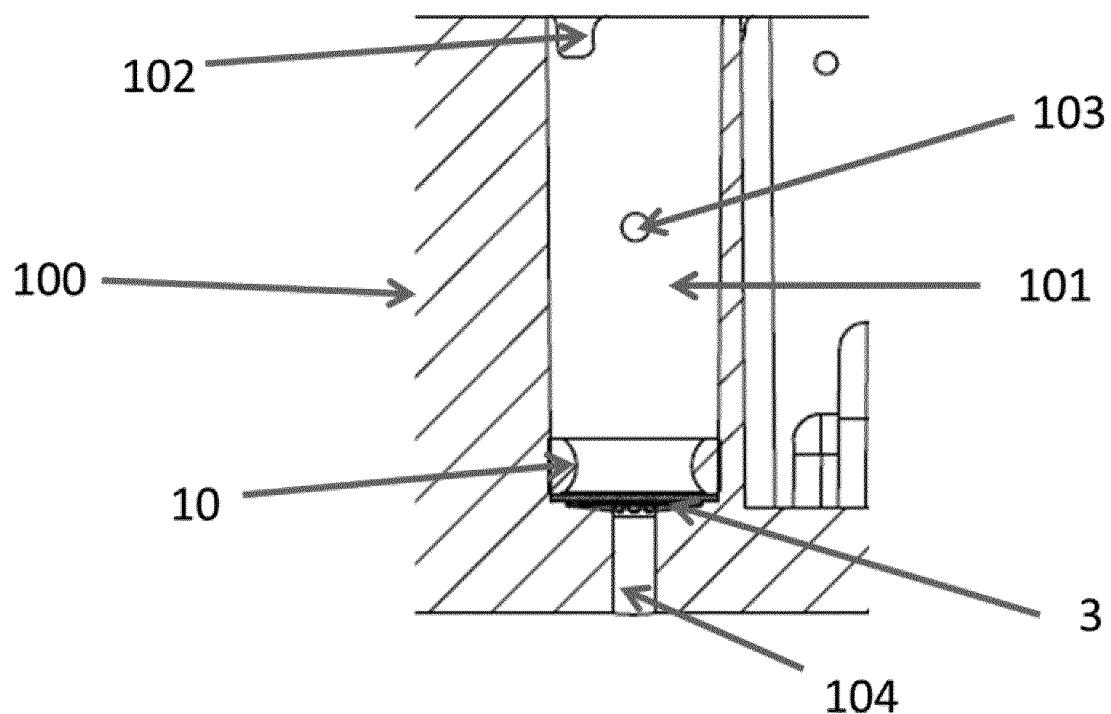


Fig. 2C:

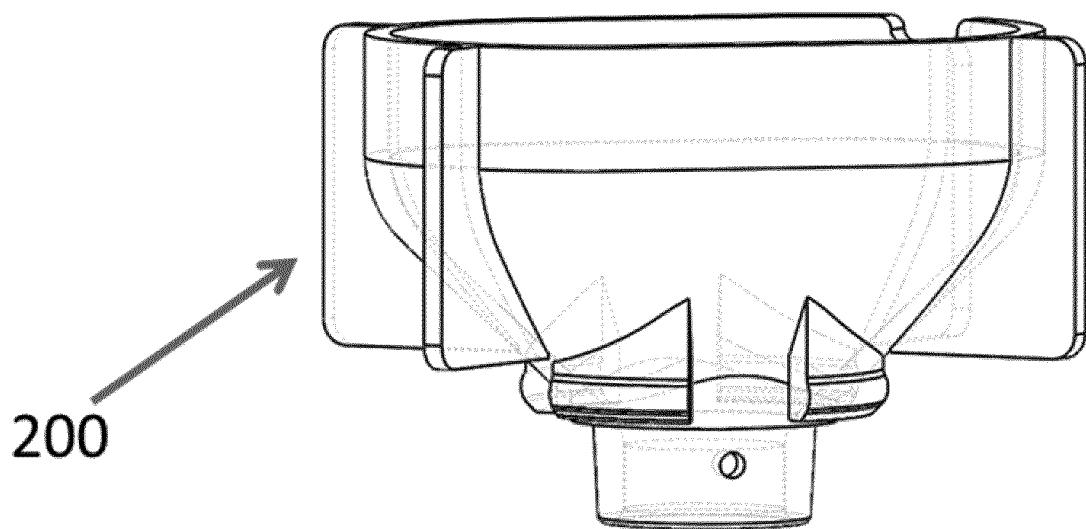


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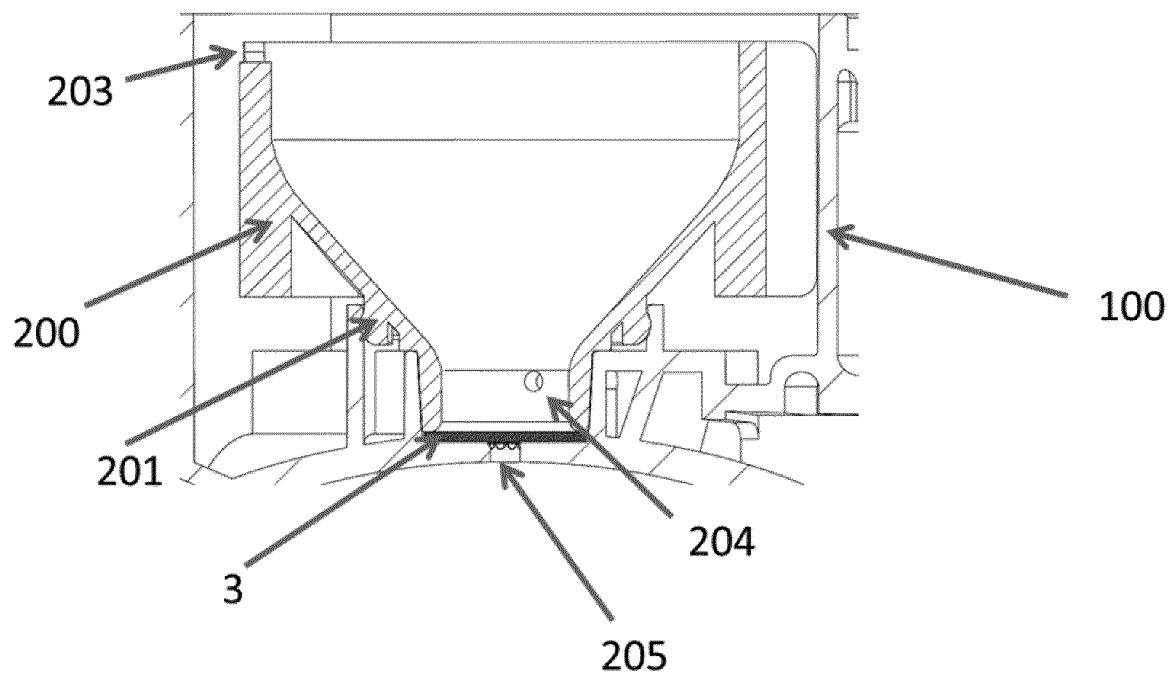


Fig. 2E

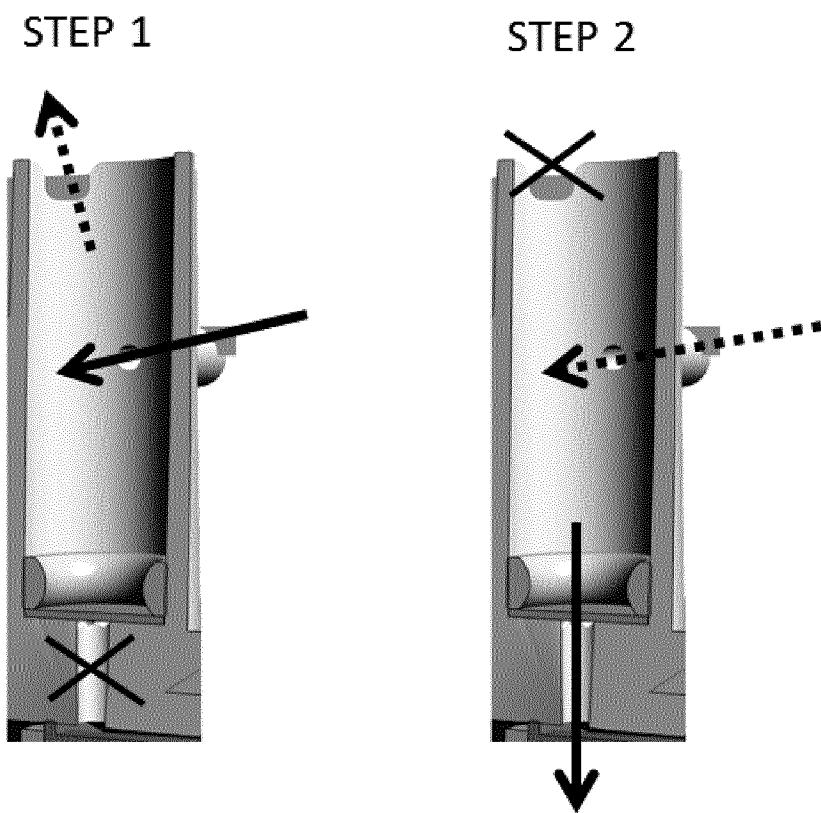


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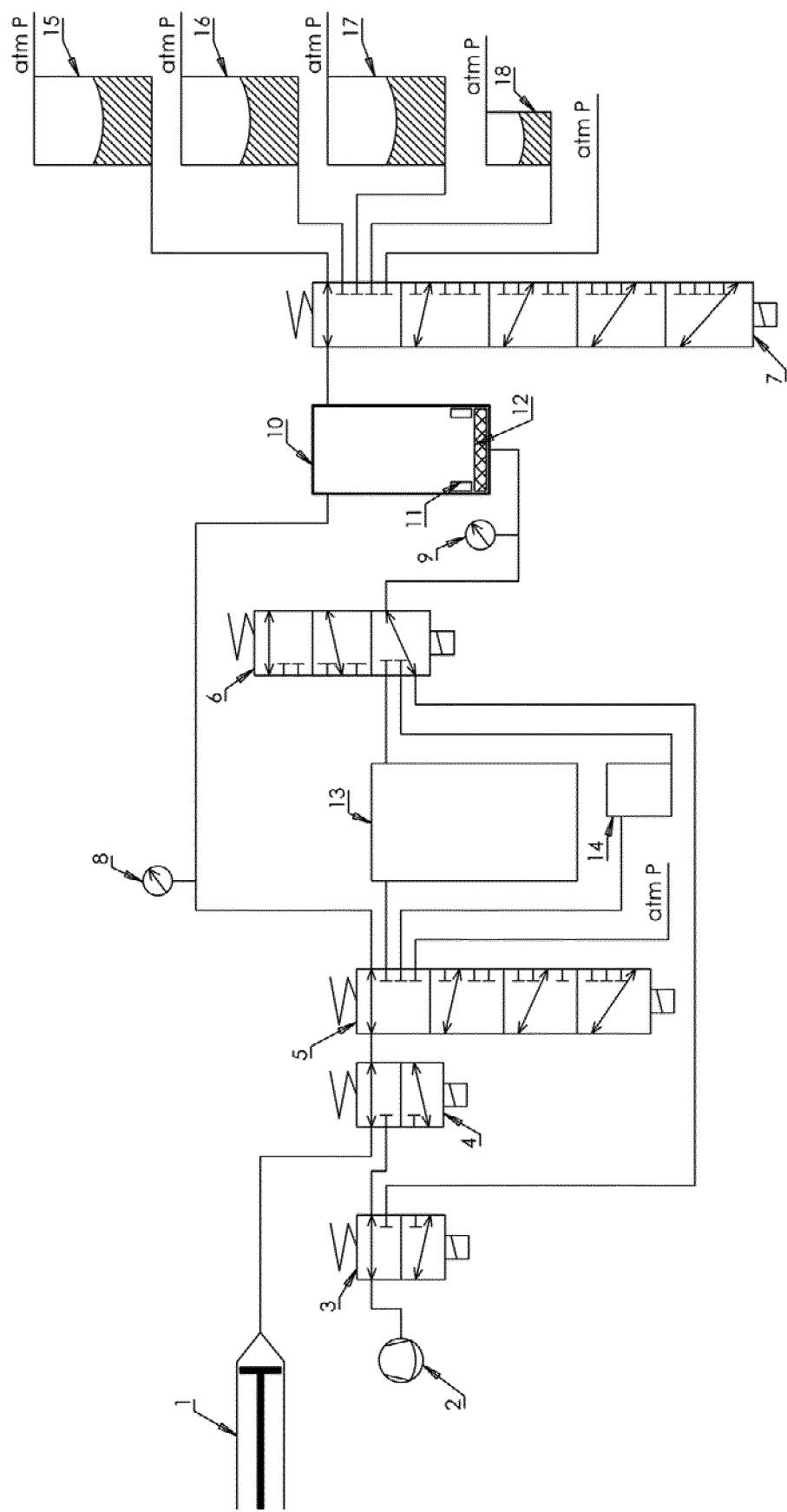


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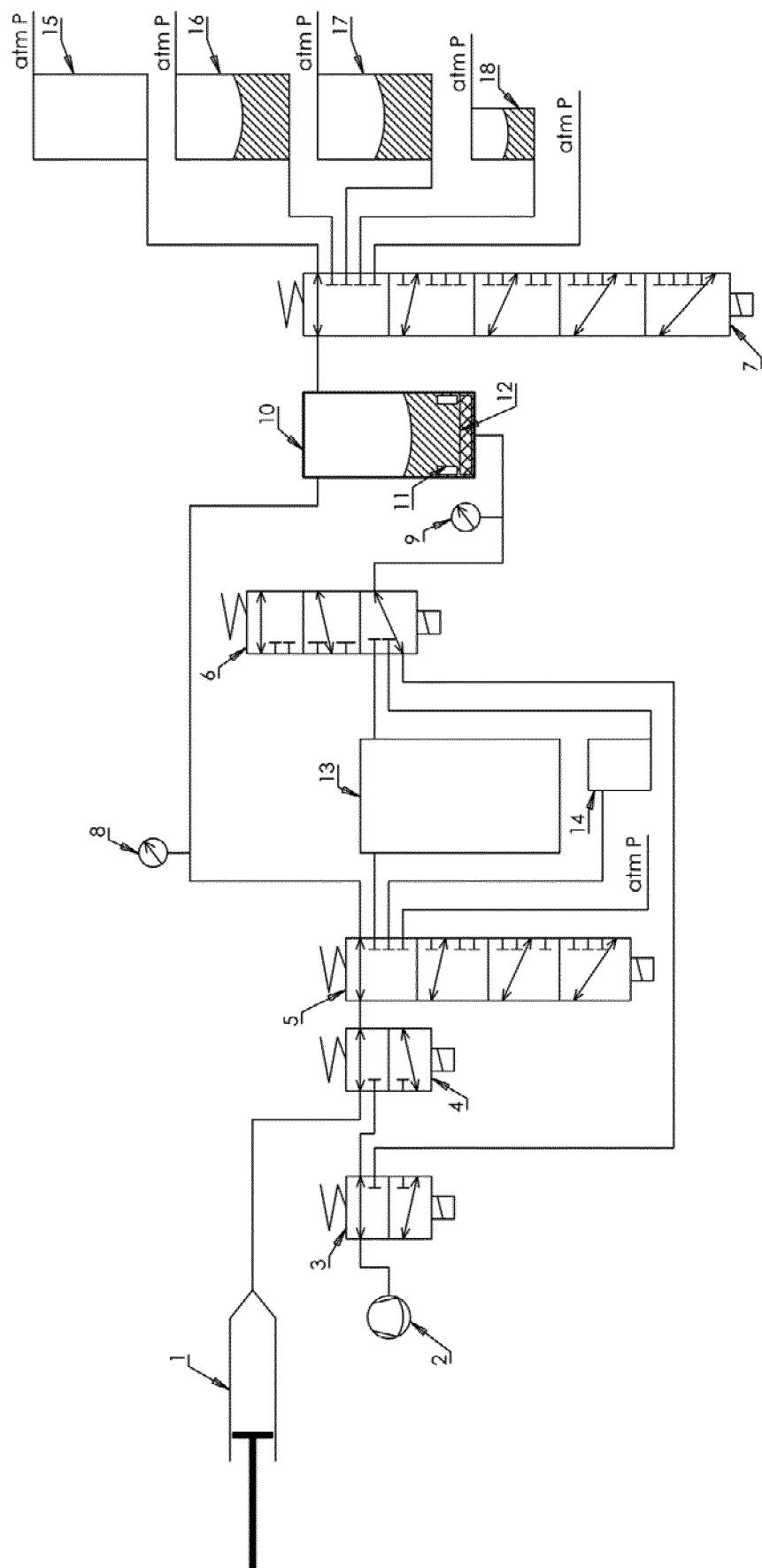


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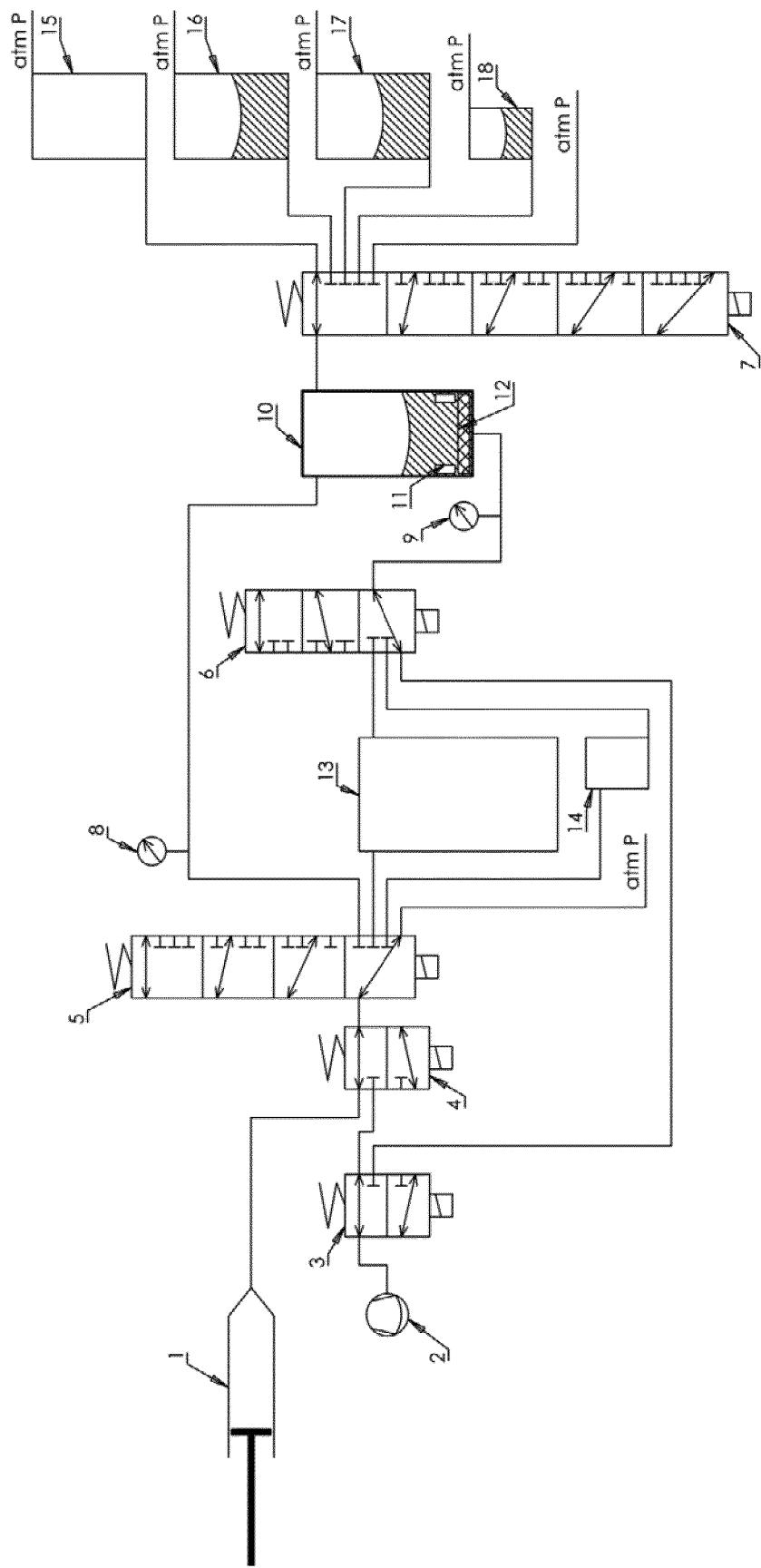


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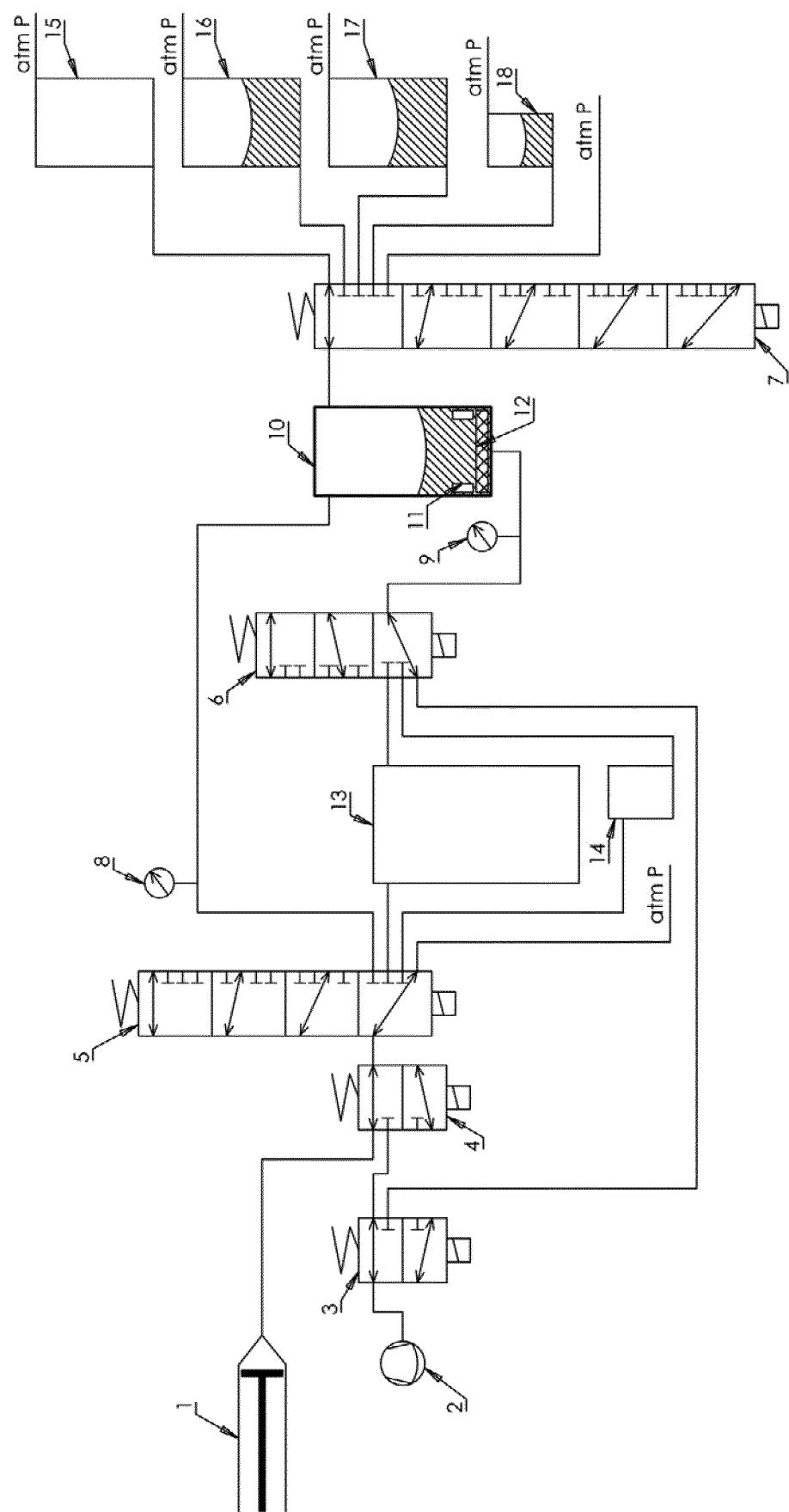


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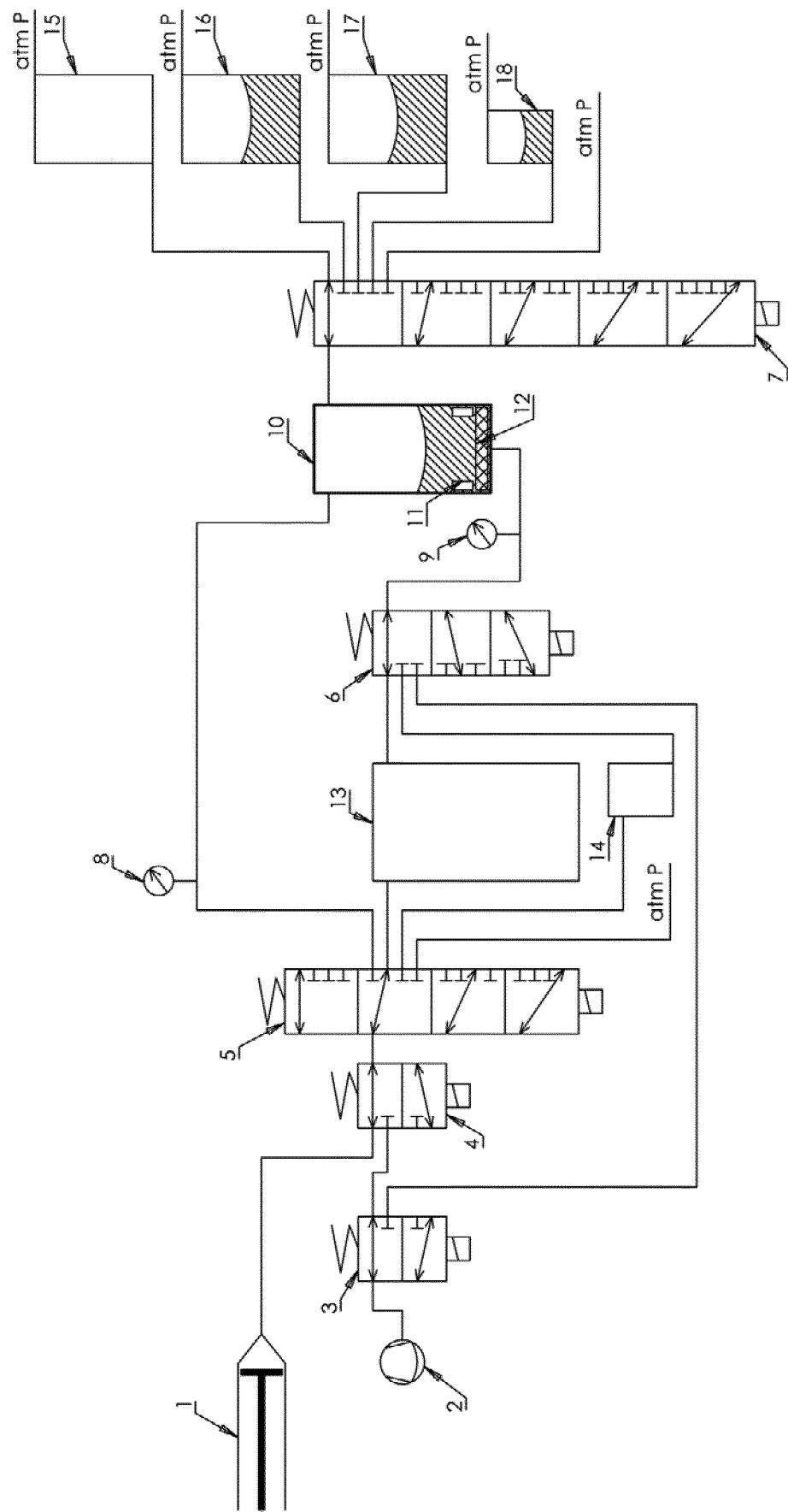


Figure 8:

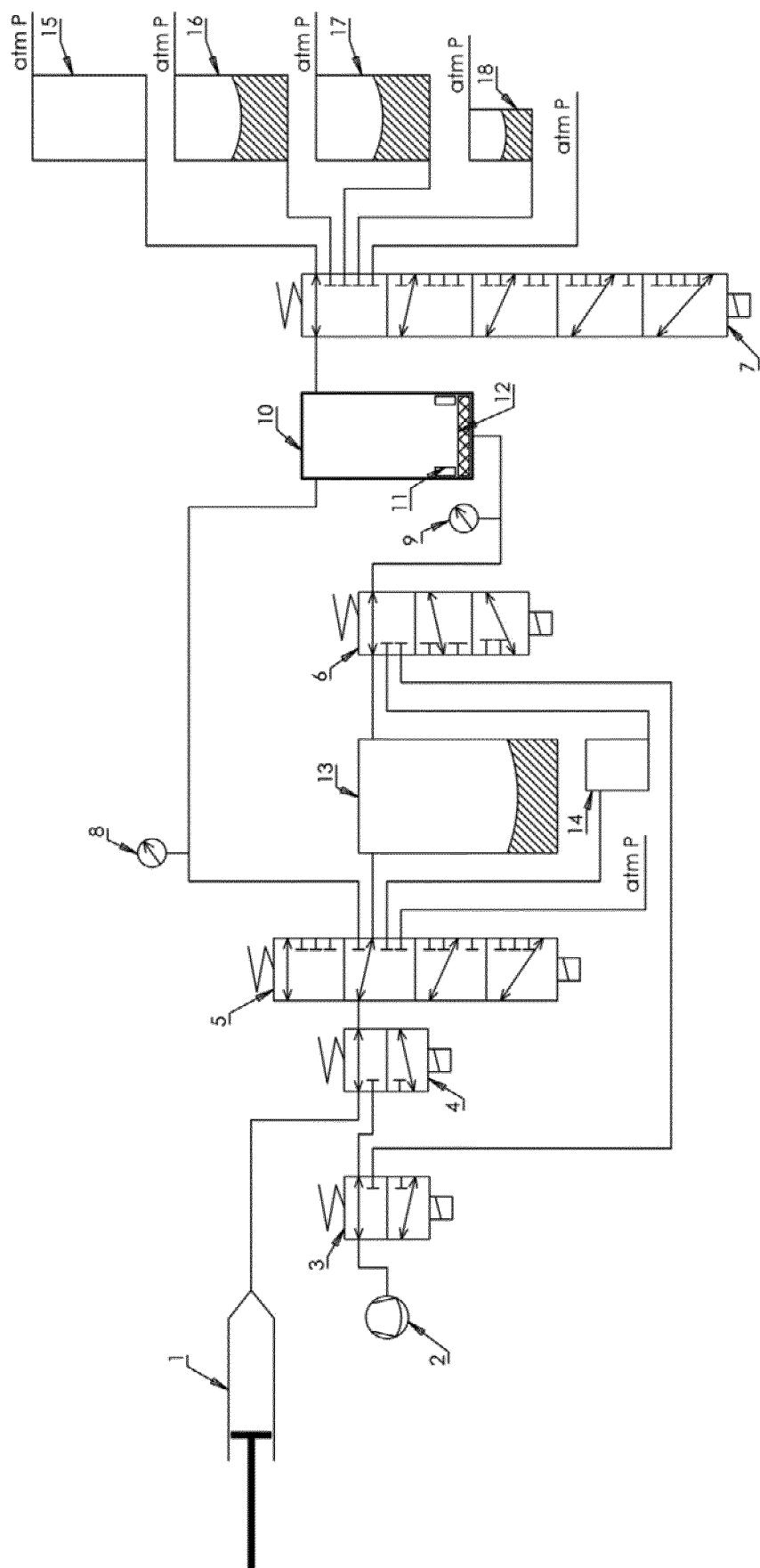


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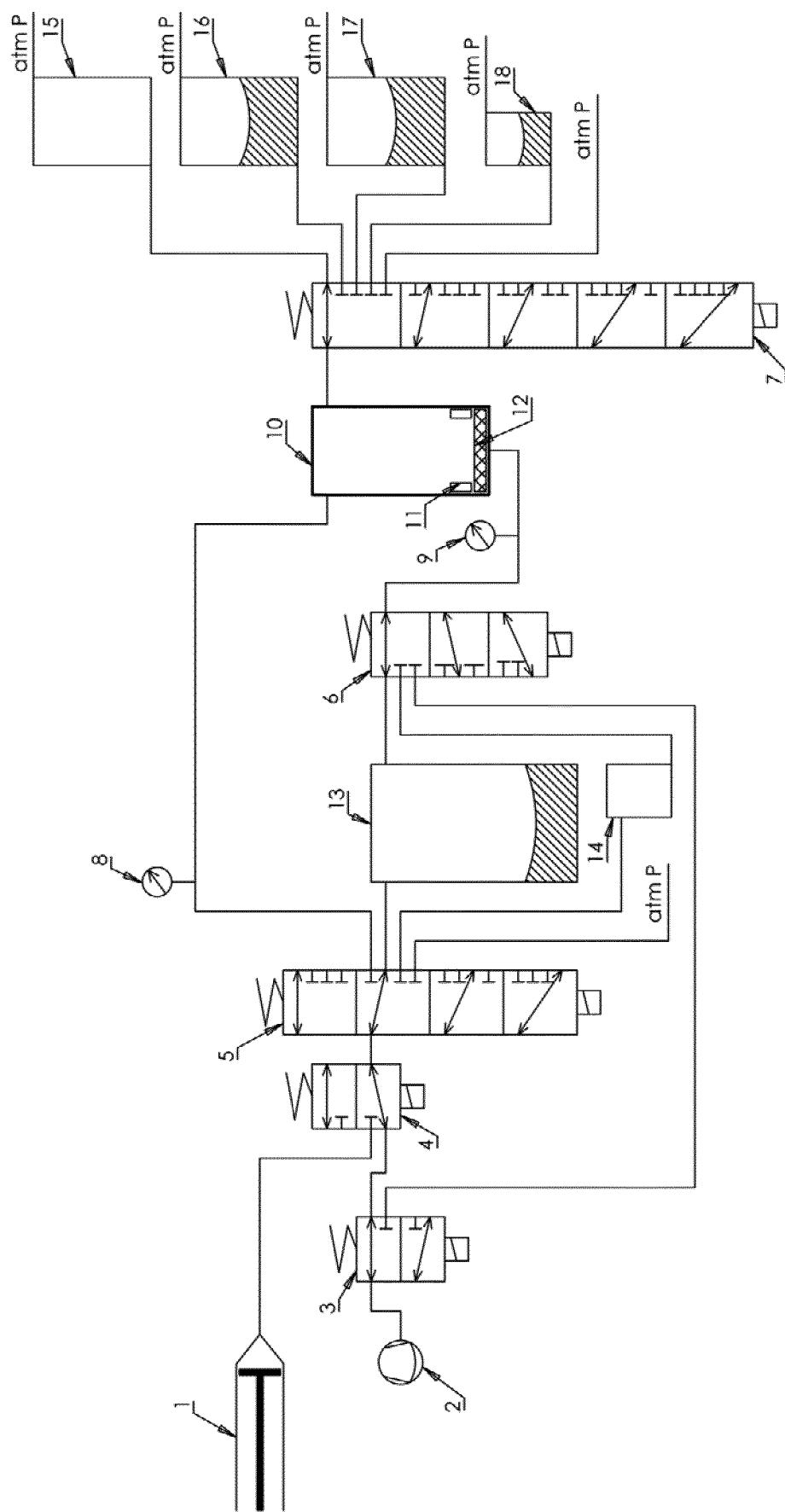


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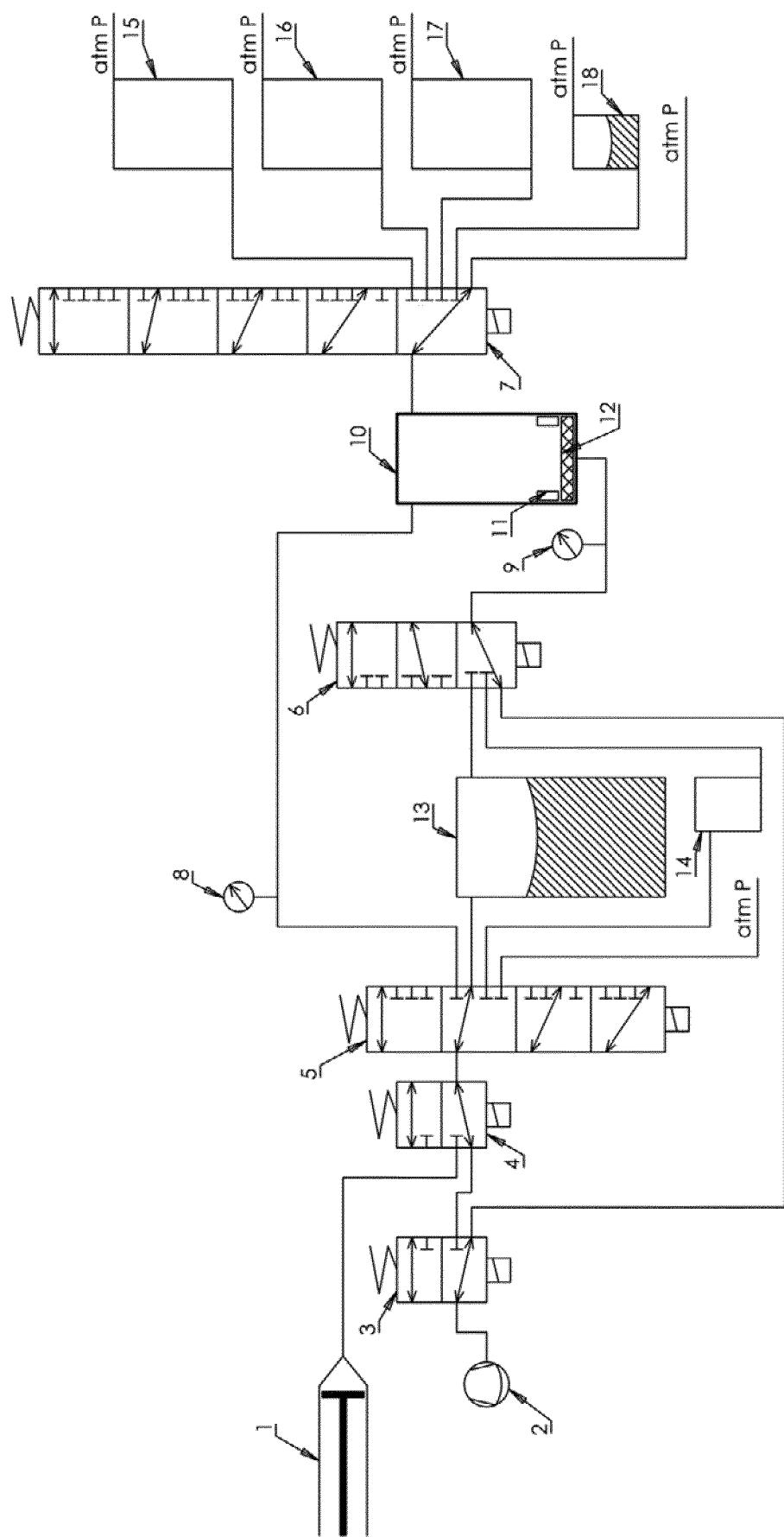


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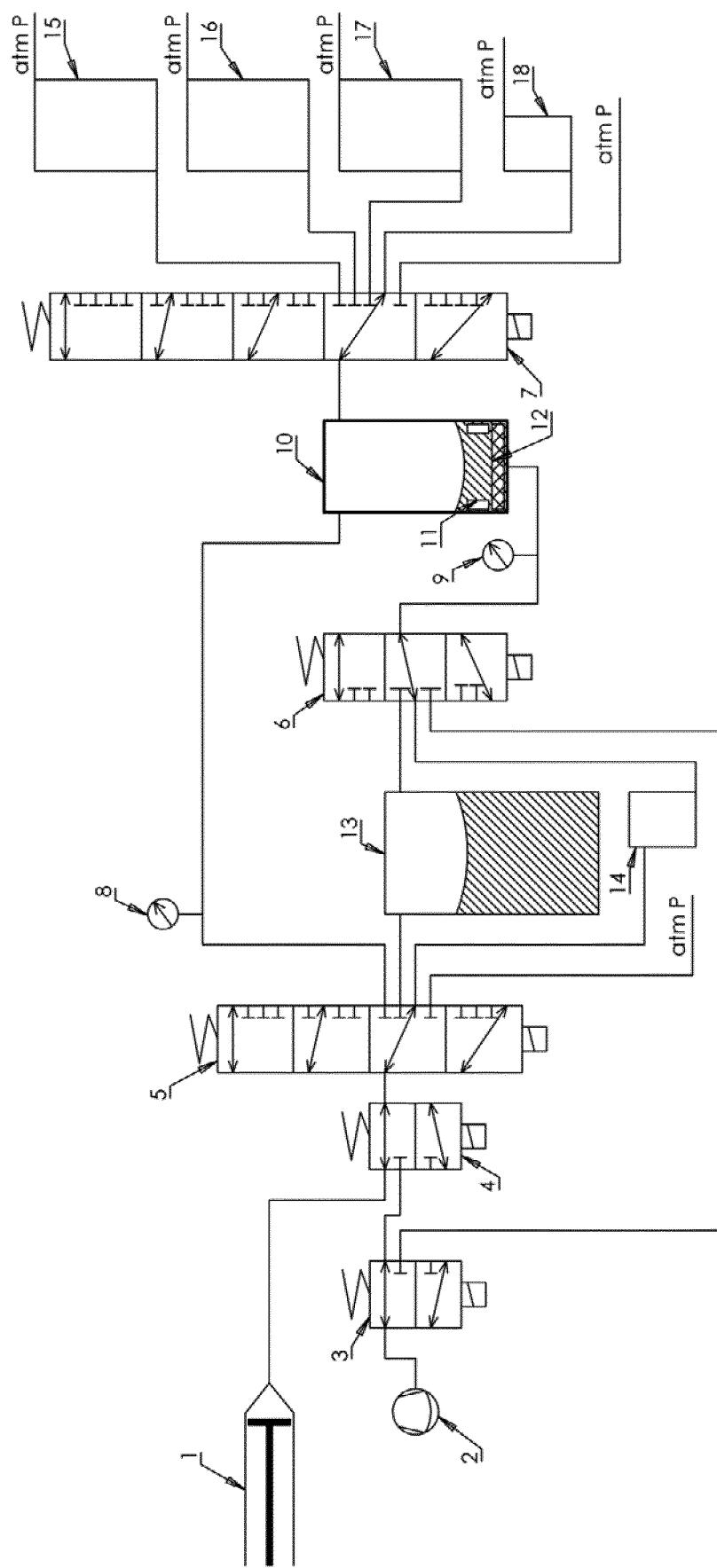


Figure 12:

