

(19) AUSTRALIAN PATENT OFFICE

(54) Title
Antisense modulation of PTP1B expression

(51)⁶ International Patent Classification(s)
C12N 15/09 (2006.01) 31/7125
A61K 31/7115 20060101ALI2005122
(2006.01) OBMJP **A61K**
A61K 31/712 48/00
(2006.01) 20060101ALI2005122
A61K 31/7125 OBMJP **A61P**
(2006.01) 3/04
A61K 48/00 (2006.01) 20060101ALI2005122
A61P 3/04 (2006.01) OBMJP **A61P**
A61P 3/10 (2006.01) 3/10
A61P 35/00 (2006.01) 20060101ALI2005122
C12N 15/11 (2006.01) OBMJP **A61P**
A61K 38/00 (2006.01) 35/00
C12N 15/09 20060101ALI2005122
20060101AFI2005122 OBMJP **C12N**
31/7115 15/11
20060101ALI2005122 8BMJP **A61K**
31/712 38/00
20060101ALI2005122 20060101ALN200510
OBMJP **A61K** 08BMJP
PCT/US02/15301

(21) Application No: 2002309819

(22) Application Date: 2002 .05 .13

(87) WIPO No: W002/092772

(30) Priority Data

(31) Number	(32) Date	(33) Country
09/854,883	2001 .05 .14	US

(43) Publication Date : 2002 .11 .25

(43) Publication Journal Date : 2003 .05 .01

(62) Divisional of:

2001278077

2001226246

(71) Applicant(s)

Isis Pharmaceuticals, Inc.

(72) Inventor(s)

MONIA, Brett, P.; WYATT, Jacqueline; COWSERT, Lex, M.; BUTLER, Madeline, M.; MCKAY, Robert; FREIER, Susan, M.

(74) Agent/Attorney

Davies Collison Cave, 255 Elizabeth Street, Sydney, NSW, 2000

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
21 November 2002 (21.11.2002)

PCT

(10) International Publication Number
WO 02/092772 A2

- (51) International Patent Classification⁷: C12N
- (21) International Application Number: PCT/US02/15301
- (22) International Filing Date: 13 May 2002 (13.05.2002)
- (25) Filing Language: English
- (26) Publication Language: English
- (30) Priority Data:
09/854,883 14 May 2001 (14.05.2001) US
- (71) Applicant (for all designated States except US): **ISIS PHARMACEUTICALS, INC.** [US/US]; Carlsbad Research Center, 2292 Faraday Avenue, Carlsbad, CA 92008 (US).
- (72) Inventors; and
- (75) Inventors/Applicants (for US only): **COWSERT, Lex, M.** [US/US]; 3008 Newshire Street, Carlsbad, CA 92008 (US). **WYATT, Jacqueline** [US/US]; 1065 Hymettus Avenue, Encinitas, CA 92024 (US). **FREIER, Susan, M.** [US/US]; 2946 Renault Street, San Diego, CA 92112 (US). **MONIA, Brett, P.** [US/US]; 7605 Nueva Castille Way, La Costa, CA 92009 (US). **BUTLER, Madeline, M.** [US/US]; 15951 Avenida Calma, Rancho Santa Fe, CA 92091 (US). **MCKAY, Robert** [US/US]; 277 Caminito Pescado #73, San Diego, CA 92116 (US).
- (74) Agents: **BAK, Mary, E.** et al.; Howson and Howson, Spring House Corporate Center, P.O. Box 457, Spring House, PA 19477 (US).
- (81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZM, ZW.
- (84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).
- Published:
— without international search report and to be republished upon receipt of that report
- For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

WO 02/092772 A2

(54) Title: ANTISENSE MODULATION OF PTP1B EXPRESSION

(57) Abstract: Compounds, compositions and methods are provided for modulating the expression of PTP1B. The compositions comprise antisense compounds, particularly antisense oligonucleotides, targeted to nucleic acids encoding PTP1B. Methods of using these compounds for modulation of PTP1B expression and for treatment of diseases associated with expression of PTP1B are provided.

ANTISENSE MODULATION OF PTP1B EXPRESSION

This application claims Convention priority from USSN 09/854,883 (14.05.2001) and International patent application No. PCT/US02/15301 (13.05.2002). This application is a divisional patent application of Australian patent application No. 2001226246 (14.01.2001), the Australian designation of International patent application No. PCT/US01/00109, and is a divisional patent application of Australian patent application No. 2001278077 (30.07.2001), the Australian designation of International patent application No. PCT/US01/23874).

BACKGROUND OF THE INVENTION

The process of phosphorylation, defined as the attachment of a phosphate moiety to a biological molecule through the action of enzymes called kinases, represents one course by which intracellular signals are propagated resulting finally in a cellular response. Within the cell, proteins can be phosphorylated on serine, threonine or tyrosine residues and the extent of phosphorylation is regulated by the opposing action of phosphatases, which remove the phosphate moieties. While the majority of protein phosphorylation within the cell is on serine and threonine residues, tyrosine phosphorylation is modulated to the greatest extent during oncogenic transformation and growth factor stimulation (Zhang, *Crit. Rev. Biochem. Mol. Biol.*, 1998, 33, 1-52).

Because phosphorylation is such a ubiquitous process within cells and because cellular phenotypes are largely influenced by the activity of these pathways, it is currently believed that a number of disease states and/or disorders are a result of either aberrant activation of,

or functional mutations in, kinases and phosphatases. Consequently, considerable attention has been devoted recently to the characterization of tyrosine kinases and tyrosine phosphatases.

- 5 PTP1B (also known as protein phosphatase 1B and PTPN1) is an endoplasmic reticulum (ER)-associated enzyme originally isolated as the major protein tyrosine phosphatase of the human placenta (Tonks *et al.*, *J. Biol. Chem.*, 1988, 263, 6731-6737; Tonks *et al.*, *J. Biol.*
- 10 *Chem.*, 1988, 263, 6722-6730).

An essential regulatory role in signaling mediated by the insulin receptor has been established for PTP1B. PTP1B interacts with and dephosphorylates the activated insulin receptor both in vitro and in intact cells
5 resulting in the downregulation of the signaling pathway (Goldstein et al., *Mol. Cell. Biochem.*, 1998, 182, 91-99; Seely et al., *Diabetes*, 1996, 45, 1379-1385). In addition, PTP1B modulates the mitogenic actions of insulin (Goldstein et al., *Mol. Cell. Biochem.*, 1998,
10 182, 91-99). In rat adipose cells overexpressing PTP1B, the translocation of the GLUT4 glucose transporter was inhibited, implicating PTP1B as a negative regulator of glucose transport as well (Chen et al., *J. Biol. Chem.*, 1997, 272, 8026-8031).

15 Mouse knockout models lacking the PTP1B gene also point toward the negative regulation of insulin signaling by PTP1B. Mice harboring a disrupted PTP1B gene showed increased insulin sensitivity, increased phosphorylation of the insulin receptor and when placed on a high-fat
20 diet, PTP1B -/- mice were resistant to weight gain and remained insulin sensitive (Elchebly et al., *Science*, 1999, 283, 1544-1548). These studies clearly establish PTP1B as a therapeutic target in the treatment of diabetes and obesity.

25 PTP1B, which is differentially regulated during the cell cycle (Schievella et al., *Cell. Growth Differ.*, 1993, 4, 239-246), is expressed in insulin sensitive tissues as two different isoforms that arise from alternate splicing of the pre-mRNA (Shifrin and Neel, *J.*
30 *Biol. Chem.*, 1993, 268, 25376-25384). It was recently demonstrated that the ratio of the alternatively spliced products is affected by growth factors such as insulin

and differs in various tissues examined (Sell and Reese, *Mol. Genet. Metab.*, **1999**, *66*, 189-192). In these studies it was also found that the levels of the variants correlated with the plasma insulin concentration and percentage body fat and may therefore be used as a biomarker for patients with chronic hyperinsulinemia or type 2 diabetes.

Liu and Chernoff have shown that PTP1B binds to and serves as a substrate for the epidermal growth factor receptor (EGFR) (Liu and Chernoff, *Biochem. J.*, **1997**, *327*, 139-145). Furthermore, in A431 human epidermoid carcinoma cells, PTP1B was found to be inactivated by the presence of H₂O₂ generated by the addition of EGF. These studies indicate that PTP1B can be negatively regulated by the oxidation state of the cell, which is often deregulated during tumorigenesis (Lee et al., *J. Biol. Chem.*, **1998**, *273*, 15366-15372).

Overexpression of PTP1B has been demonstrated in malignant ovarian cancers and this correlation was accompanied by a concomitant increase in the expression of the associated growth factor receptor (Wiener et al., *Am. J. Obstet. Gynecol.*, **1994**, *170*, 1177-1183).

PTP1B has been shown to suppress transformation in NIH3T3 cells induced by the neu oncogene (Brown-Shimer et al., *Cancer Res.*, **1992**, *52*, 478-482), as well as in rat 3Y1 fibroblasts induced by v-src, v-src, and v-ras (Liu et al., *Mol. Cell. Biol.*, **1998**, *18*, 250-259) and rat-1 fibroblasts induced by bcr-abl (LaMontagne et al., *Proc. Natl. Acad. Sci. U. S. A.*, **1998**, *95*, 14094-14099). It has also been shown that PTP1B promotes differentiation of K562 cells, a chronic myelogenous leukemia cell line, in a similar manner as does an inhibitor of the bcr-abl

oncoprotein. These studies describe the possible role of PTP1B in controlling the pathogenesis of chronic myeloid leukemia (LaMontagne et al., *Proc. Natl. Acad. Sci. U. S. A.*, **1998**, *95*, 14094-14099).

5 PTP1B negatively regulates integrin signaling by interacting with one or more adhesion-dependent signaling components and repressing integrin-mediated MAP kinase activation (Liu et al., *Curr. Biol.*, **1998**, *8*, 173-176). Other studies designed to study integrin signaling, using
10 a catalytically inactive form of PTP1B, have shown that PTP1B regulates cadherin-mediated cell adhesion (Balsamo et al., *J. Cell. Biol.*, **1998**, *143*, 523-532) as well as cell spreading, focal adhesion and stress fiber formation and tyrosine phosphorylation (Arregui et al., *J. Cell.*
15 *Biol.*, **1998**, *143*, 861-873).

Currently, therapeutic agents designed to inhibit the synthesis or action of PTP1B include small molecules (Ham et al., *Bioorg. Med. Chem. Lett.*, **1999**, *9*, 185-186; Skorey et al., *J. Biol. Chem.*, **1997**, *272*, 22472-22480;
20 Taing et al., *Biochemistry*, **1999**, *38*, 3793-3803; Taylor et al., *Bioorg. Med. Chem.*, **1998**, *6*, 1457-1468; Wang et al., *Bioorg. Med. Chem. Lett.*, **1998**, *8*, 345-350; Wang et al., *Biochem. Pharmacol.*, **1997**, *54*, 703-711; Yao et al., *Bioorg. Med. Chem.*, **1998**, *6*, 1799-1810) and peptides
25 (Chen et al., *Biochemistry*, **1999**, *38*, 384-389; Desmarais et al., *Arch. Biochem. Biophys.*, **1998**, *354*, 225-231; Roller et al., *Bioorg. Med. Chem. Lett.*, **1998**, *8*, 2149-2150). In addition, disclosed in the PCT publication WO 97/32595 are phosphopeptides and antibodies that inhibit
30 the association of PTP1B with the activated insulin receptor for the treatment of disorders associated with

insulin resistance. Antisense nucleotides against PTP1B are also generally disclosed in that document.

There remains a long felt need for additional agents capable of effectively inhibiting PTP1B function and antisense technology is emerging as an effective means for reducing the expression of specific gene products. This technology may therefore prove to be uniquely useful in a number of therapeutic, diagnostic, and research applications for the modulation of PTP1B expression.

SUMMARY OF THE INVENTION

The present invention provides compositions and methods for modulating the expression of PTP1B. In particular, the present invention provides compositions and methods for modulating the alternatively spliced form of PTP1B. This invention relates to antisense compounds, particularly antisense oligonucleotides, specifically hybridizable with and targeted to a nucleic acid encoding PTP1B. Such oligonucleotides have been shown to modulate the expression of PTP1B.

Pharmaceutical and other compositions comprising the antisense compounds of the invention are also provided. Further provided are methods of modulating the expression of PTP1B in cells or tissues comprising contacting said cells or tissues with one or more of the antisense compounds or compositions of the invention.

Further provided are methods of treating an animal, particularly a human, suspected of having or being prone to a disease or condition associated with expression of PTP1B by administering a therapeutically or prophylactically effective amount of one or more of the antisense compounds or compositions of the invention.

Other aspects and advantages of this invention are encompassed in the following detailed description of the invention.

5 DETAILED DESCRIPTION OF THE INVENTION

The present invention employs oligomeric antisense compounds, particularly oligonucleotides, for use in modulating the function of nucleic acid molecules encoding PTP1B, ultimately modulating the amount of PTP1B
10 produced. This is accomplished by providing antisense compounds which specifically hybridize with one or more nucleic acids encoding PTP1B.

As used herein, the terms "target nucleic acid" and "nucleic acid encoding PTP1B" encompass DNA encoding
15 PTP1B, RNA (including pre-mRNA and mRNA) transcribed from such DNA, and also cDNA derived from such RNA. The specific hybridization of an oligomeric compound with its target nucleic acid interferes with the normal function of the nucleic acid. This modulation of function of a
20 target nucleic acid by compounds which specifically hybridize to it is generally referred to as "antisense". The functions of DNA to be interfered with include replication and transcription. The functions of RNA to be interfered with include all vital functions such as,
25 for example, translocation of the RNA to the site of protein translation, translation of protein from the RNA, splicing of the RNA to yield one or more mRNA species, and catalytic activity which may be engaged in or facilitated by the RNA. The overall effect of such
30 interference with target nucleic acid function is modulation of the expression of PTP1B.

In the context of the present invention, "modulation" means either an increase (stimulation) or a

decrease (inhibition) in the expression of a gene. In the context of the present invention, inhibition is the preferred form of modulation of gene expression and mRNA is a preferred target.

5 It is preferred to target specific nucleic acids for antisense. "Targeting" an antisense compound to a particular nucleic acid, in the context of this invention, is a multistep process. The process usually begins with the identification of a nucleic acid sequence
10 whose function is to be modulated. This may be, for example, a cellular gene (or mRNA transcribed from the gene) whose expression is associated with a particular disorder or disease state, or a nucleic acid molecule from an infectious agent. In the present invention, the
15 target is a nucleic acid molecule encoding PTP1B. The targeting process also includes determination of a site or sites within this gene for the antisense interaction to occur such that the desired effect, e.g., detection or modulation of expression of the protein, will result.
20 Within the context of the present invention, a preferred intragenic site is the region encompassing the translation initiation or termination codon of the open reading frame (ORF) of the gene.

 Since, as is known in the art, the translation
25 initiation codon is typically 5'-AUG (in transcribed mRNA molecules; 5'-ATG in the corresponding DNA molecule), the translation initiation codon is also referred to as the "AUG codon," the "start codon" or the "AUG start codon". A minority of genes have a translation initiation codon
30 having the RNA sequence 5'-GUG, 5'-UUG or 5'-CUG, and 5'-AUA, 5'-ACG and 5'-CUG have been shown to function in vivo. Thus, the terms "translation initiation codon" and "start codon" can encompass many codon sequences, even

though the initiator amino acid in each instance is typically methionine (in eukaryotes) or formylmethionine (in prokaryotes). It is also known in the art that eukaryotic and prokaryotic genes may have two or more alternative start codons, any one of which may be preferentially utilized for translation initiation in a particular cell type or tissue, or under a particular set of conditions. In the context of the invention, "start codon" and "translation initiation codon" refer to the codon or codons that are used *in vivo* to initiate translation of an mRNA molecule transcribed from a gene encoding PTP1B, regardless of the sequence(s) of such codons.

It is also known in the art that a translation termination codon (or "stop codon") of a gene may have one of three sequences, i.e., 5'-UAA, 5'-UAG and 5'-UGA (the corresponding DNA sequences are 5'-TAA, 5'-TAG and 5'-TGA, respectively). The terms "start codon region" and "translation initiation codon region" refer to a portion of such an mRNA or gene that encompasses from about 25 to about 50 contiguous nucleotides in either direction (i.e., 5' or 3') from a translation initiation codon. Similarly, the terms "stop codon region" and "translation termination codon region" refer to a portion of such an mRNA or gene that encompasses from about 25 to about 50 contiguous nucleotides in either direction (i.e., 5' or 3') from a translation termination codon.

The open reading frame (ORF) or "coding region," which is known in the art to refer to the region between the translation initiation codon and the translation termination codon, is also a region which may be targeted effectively. Other target regions include the 5' untranslated region (5'UTR), known in the art to refer to

the portion of an mRNA in the 5' direction from the translation initiation codon, and thus including nucleotides between the 5' cap site and the translation initiation codon of an mRNA or corresponding nucleotides on the gene, and the 3' untranslated region (3'UTR), known in the art to refer to the portion of an mRNA in the 3' direction from the translation termination codon, and thus including nucleotides between the translation termination codon and 3' end of an mRNA or corresponding nucleotides on the gene. The 5' cap of an mRNA comprises an N7-methylated guanosine residue joined to the 5'-most residue of the mRNA via a 5'-5' triphosphate linkage. The 5' cap region of an mRNA is considered to include the 5' cap structure itself as well as the first 50 nucleotides adjacent to the cap. The 5' cap region may also be a preferred target region.

Although some eukaryotic mRNA transcripts are directly translated, many contain one or more regions, known as "introns," which are excised from a transcript before it is translated. The remaining (and therefore translated) regions are known as "exons" and are spliced together to form a continuous mRNA sequence. mRNA splice sites, i.e., intron-exon junctions, may also be preferred target regions, and are particularly useful in situations where aberrant splicing is implicated in disease, or where an overproduction of a particular mRNA splice product is implicated in disease. Aberrant fusion junctions due to rearrangements or deletions are also preferred targets. It has also been found that introns can also be effective, and therefore preferred, target regions for antisense compounds targeted, for example, to DNA or pre-mRNA.

Once one or more target sites have been identified, oligonucleotides are chosen which are sufficiently complementary to the target, i.e., hybridize sufficiently well and with sufficient specificity, to give the desired effect.

In the context of this invention, "hybridization" means hydrogen bonding, which may be Watson-Crick, Hoogsteen or reversed Hoogsteen hydrogen bonding, between complementary nucleoside or nucleotide bases. For example, adenine and thymine are complementary nucleobases which pair through the formation of hydrogen bonds. "Complementary," as used herein, refers to the capacity for precise pairing between two nucleotides. For example, if a nucleotide at a certain position of an oligonucleotide is capable of hydrogen bonding with a nucleotide at the same position of a DNA or RNA molecule, then the oligonucleotide and the DNA or RNA are considered to be complementary to each other at that position. The oligonucleotide and the DNA or RNA are complementary to each other when a sufficient number of corresponding positions in each molecule are occupied by nucleotides which can hydrogen bond with each other. Thus, "specifically hybridizable" and "complementary" are terms which are used to indicate a sufficient degree of complementarity or precise pairing such that stable and specific binding occurs between the oligonucleotide and the DNA or RNA target. It is understood in the art that the sequence of an antisense compound need not be 100% complementary to that of its target nucleic acid to be specifically hybridizable. An antisense compound is specifically hybridizable when binding of the compound to the target DNA or RNA molecule interferes with the normal function of the target DNA or RNA to cause a loss of

utility, and there is a sufficient degree of complementarity to avoid non-specific binding of the antisense compound to non-target sequences under conditions in which specific binding is desired, i.e.,
5 under physiological conditions in the case of *in vivo* assays or therapeutic treatment, and in the case of *in vitro* assays, under conditions in which the assays are performed.

Antisense compounds are commonly used as research
10 reagents and diagnostics. For example, antisense oligonucleotides, which are able to inhibit gene expression with exquisite specificity, are often used by those of ordinary skill to elucidate the function of particular genes. Antisense compounds are also used, for
15 example, to distinguish between functions of various members of a biological pathway. Antisense modulation has, therefore, been harnessed for research use.

The specificity and sensitivity of antisense is also harnessed by those of skill in the art for therapeutic
20 uses. Antisense oligonucleotides have been employed as therapeutic moieties in the treatment of disease states in animals and man. Antisense oligonucleotides have been safely and effectively administered to humans and numerous clinical trials are presently underway. It is
25 thus established that oligonucleotides can be useful therapeutic modalities that can be configured to be useful in treatment regimes for treatment of cells, tissues and animals, especially humans.

In the context of this invention, the term
30 "oligonucleotide" refers to an oligomer or polymer of ribonucleic acid (RNA) or deoxyribonucleic acid (DNA) or mimetics thereof. This term includes oligonucleotides composed of naturally-occurring nucleobases, sugars and

covalent internucleoside (backbone) linkages as well as oligonucleotides having non-naturally-occurring portions which function similarly. Such modified or substituted oligonucleotides are often preferred over native forms because of desirable properties such as, for example, enhanced cellular uptake, enhanced affinity for nucleic acid target and increased stability in the presence of nucleases.

While antisense oligonucleotides are a preferred form of antisense compound, the present invention comprehends other oligomeric antisense compounds, including but not limited to oligonucleotide mimetics such as are described below. The antisense compounds in accordance with this invention preferably comprise from about 8 to about 50 nucleobases (i.e. from about 8 to about 50 linked nucleosides). Particularly preferred antisense compounds are antisense oligonucleotides, even more preferably those comprising from about 12 to about 30 nucleobases. As is known in the art, a nucleoside is a base-sugar combination. The base portion of the nucleoside is normally a heterocyclic base. The two most common classes of such heterocyclic bases are the purines and the pyrimidines. Nucleotides are nucleosides that further include a phosphate group covalently linked to the sugar portion of the nucleoside. For those nucleosides that include a pentofuranosyl sugar, the phosphate group can be linked to either the 2', 3' or 5' hydroxyl moiety of the sugar. In forming oligonucleotides, the phosphate groups covalently link adjacent nucleosides to one another to form a linear polymeric compound. In turn the respective ends of this linear polymeric structure can be further joined to form a circular structure, however, open linear structures are

generally preferred. Within the oligonucleotide structure, the phosphate groups are commonly referred to as forming the internucleoside backbone of the oligonucleotide. The normal linkage or backbone of RNA and DNA is a 3' to 5' phosphodiester linkage.

Specific examples of preferred antisense compounds useful in this invention include oligonucleotides containing modified backbones or non-natural internucleoside linkages. As defined in this specification, oligonucleotides having modified backbones include those that retain a phosphorus atom in the backbone and those that do not have a phosphorus atom in the backbone. For the purposes of this specification, and as sometimes referenced in the art, modified oligonucleotides that do not have a phosphorus atom in their internucleoside backbone can also be considered to be oligonucleosides.

Preferred modified oligonucleotide backbones include, for example, phosphorothioates, chiral phosphorothioates, phosphorodithioates, phosphotriesters, aminoalkylphosphotriesters, methyl and other alkyl phosphonates including 3'-alkylene phosphonates and chiral phosphonates, phosphinates, phosphoramidates including 3'-amino phosphoramidate and aminoalkylphosphoramidates, thionophosphoramidates, thionoalkylphosphonates, thionoalkylphosphotriesters, and boranophosphates having normal 3'-5' linkages, 2'-5' linked analogs of these, and those having inverted polarity wherein the adjacent pairs of nucleoside units are linked 3'-5' to 5'-3' or 2'-5' to 5'-2'. Various salts, mixed salts and free acid forms are also included.

Representative United States patents that teach the preparation of the above phosphorus-containing linkages include, but are not limited to, U.S.: 3,687,808;

4,469,863; 4,476,301; 5,023,243; 5,177,196; 5,188,897;
5,264,423; 5,276,019; 5,278,302; 5,286,717; 5,321,131;
5,399,676; 5,405,939; 5,453,496; 5,455,233; 5,466,677;
5,476,925; 5,519,126; 5,536,821; 5,541,306; 5,550,111;
5 5,563,253; 5,571,799; 5,587,361; and [COMMENT][a]Page: 1

The patent No. 5,697,248 was found to be wrong. On 9-3-98 deleted it and changed the wording from "5,625,050; and 5,697,248" to " and 5,625,050" 5,625,050, certain of which are commonly owned with this application, and each of which is herein incorporated by reference.

Preferred modified oligonucleotide backbones that do not include a phosphorus atom therein have backbones that are formed by short chain alkyl or cycloalkyl internucleoside linkages, mixed heteroatom and alkyl or cycloalkyl internucleoside linkages, or one or more short chain heteroatomic or heterocyclic internucleoside linkages. These include those having morpholino linkages (formed in part from the sugar portion of a nucleoside); siloxane backbones; sulfide, sulfoxide and sulfone backbones; formacetyl and thioformacetyl backbones; methylene formacetyl and thioformacetyl backbones; alkene containing backbones; sulfamate backbones; methyleneimino and methylenehydrazino backbones; sulfonate and sulfonamide backbones; amide backbones; and others having mixed N, O, S and CH₂ component parts.

Representative United States patents that teach the preparation of the above oligonucleosides include, but are not limited to, U.S.: 5,034,506; 5,166,315; 5,185,444; 5,214,134; 5,216,141; 5,235,033; 5,264,562; 5,264,564; 5,405,938; 5,434,257; 5,466,677; 5,470,967; 5,489,677; 5,541,307; 5,561,225; 5,596,086; 5,602,240; 5,610,289; 5,602,240; 5,608,046; 5,610,289; 5,618,704; 5,623,070; 5,663,312; 5,633,360; 5,677,437; and

5,677,439, certain of which are commonly owned with this application, and each of which is herein incorporated by reference.

In other preferred oligonucleotide mimetics, both
5 the sugar and the internucleoside linkage, i.e., the backbone, of the nucleotide units are replaced with novel groups. The base units are maintained for hybridization with an appropriate nucleic acid target compound. One such oligomeric compound, an oligonucleotide mimetic that
10 has been shown to have excellent hybridization properties, is referred to as a peptide nucleic acid (PNA). In PNA compounds, the sugar-backbone of an oligonucleotide is replaced with an amide containing backbone, in particular an aminoethylglycine backbone.
15 The nucleobases are retained and are bound directly or indirectly to aza nitrogen atoms of the amide portion of the backbone. Representative United States patents that teach the preparation of PNA compounds include, but are not limited to, U.S.: 5,539,082; 5,714,331; and
20 5,719,262, each of which is herein incorporated by reference. Further teaching of PNA compounds can be found in Nielsen et al., *Science*, 1991, 254, 1497-1500.

Most preferred embodiments of the invention are oligonucleotides with phosphorothioate backbones and
25 oligonucleosides with heteroatom backbones, and in particular -CH₂-NH-O-CH₂-, -CH₂-N(CH₃)-O-CH₂- [known as a methylene (methylimino) or MMI backbone], -CH₂-O-N(CH₃)-CH₂-, -CH₂-N(CH₃)-N(CH₃)-CH₂- and -O-N(CH₃)-CH₂-CH₂- [wherein the native phosphodiester backbone is
30 represented as -O-P-O-CH₂-] of the above referenced U.S. patent 5,489,677, and the amide backbones of the above referenced U.S. patent 5,602,240. Also preferred are

oligonucleotides having morpholino backbone structures of the above-referenced U.S. patent 5,034,506.

Modified oligonucleotides may also contain one or more substituted sugar moieties. Preferred oligonucleotides comprise one of the following at the 2' position: OH; F; O-, S-, or N-alkyl; O-, S-, or N-alkenyl; O-, S- or N-alkynyl; or O-alkyl-O-alkyl, wherein the alkyl, alkenyl and alkynyl may be substituted or unsubstituted C₁ to C₁₀ alkyl or C₂ to C₁₀ alkenyl and alkynyl. Particularly preferred are O[(CH₂)_nO]_mCH₃, O(CH₂)_nOCH₃, O(CH₂)_nNH₂, O(CH₂)_nCH₃, O(CH₂)_nONH₂, and O(CH₂)_nON[(CH₂)_nCH₃]₂, where n and m are from 1 to about 10. Other preferred oligonucleotides comprise one of the following at the 2' position: C₁ to C₁₀ lower alkyl, substituted lower alkyl, alkaryl, aralkyl, O-alkaryl or O-aralkyl, SH, SCH₃, OCN, Cl, Br, CN, CF₃, OCF₃, SOCH₃, SO₂CH₃, ONO₂, NO₂, N₃, NH₂, heterocycloalkyl, heterocycloalkaryl, aminoalkylamino, polyalkylamino, substituted silyl, an RNA cleaving group, a reporter group, an intercalator, a group for improving the pharmacokinetic properties of an oligonucleotide, or a group for improving the pharmacodynamic properties of an oligonucleotide, and other substituents having similar properties. A preferred modification includes 2'-methoxyethoxy (2'-O-CH₂CH₂OCH₃, also known as 2'-O-(2-methoxyethyl) or 2'-MOE) (Martin et al., *Helv. Chim. Acta*, 1995, 78, 486-504) i.e., an alkoxyalkoxy group. A further preferred modification includes 2'-dimethylaminoethoxy, i.e., a O(CH₂)₂ON(CH₃)₂ group, also known as 2'-DMAOE, as described in examples hereinbelow, and 2'-dimethylaminoethoxyethoxy (also known in the art as 2'-O-dimethylaminoethoxyethyl or 2'-DMAEOE), i.e., 2'-O-CH₂-O-CH₂-N(CH₂)₂, also described in examples hereinbelow.

Other preferred modifications include 2'-methoxy (2'-O-CH₃), 2'-aminopropoxy (2'-OCH₂CH₂CH₂NH₂) and 2'-fluoro (2'-F). Similar modifications may also be made at other positions on the oligonucleotide, particularly the 3' position of the sugar on the 3' terminal nucleotide or in 2'-5' linked oligonucleotides and the 5' position of 5' terminal nucleotide. Oligonucleotides may also have sugar mimetics such as cyclobutyl moieties in place of the pentofuranosyl sugar. Representative United States patents that teach the preparation of such modified sugar structures include, but are not limited to, U.S.: 4,981,957; 5,118,800; 5,319,080; 5,359,044; 5,393,878; 5,446,137; 5,466,786; 5,514,785; 5,519,134; 5,567,811; 5,576,427; 5,591,722; 5,597,909; 5,610,300; 5,627,053; 5,639,873; 5,646,265; 5,658,873; 5,670,633; and 5,700,920, certain of which are commonly owned with the instant application, and each of which is herein incorporated by reference in its entirety.

Oligonucleotides may also include nucleobase (often referred to in the art simply as "base") modifications or substitutions. As used herein, "unmodified" or "natural" nucleobases include the purine bases adenine (A) and guanine (G), and the pyrimidine bases thymine (T), cytosine (C) and uracil (U). Modified nucleobases include other synthetic and natural nucleobases such as 5-methylcytosine (5-me-C), 5-hydroxymethyl cytosine, xanthine, hypoxanthine, 2-aminoadenine, 6-methyl and other alkyl derivatives of adenine and guanine, 2-propyl and other alkyl derivatives of adenine and guanine, 2-thiouracil, 2-thiothymine and 2-thiocytosine, 5-halouracil and cytosine, 5-propynyl uracil and cytosine, 6-azo uracil, cytosine and thymine, 5-uracil (pseudo-uracil), 4-thiouracil, 8-halo, 8-amino, 8-thiol, 8-

thioalkyl, 8-hydroxyl and other 8-substituted adenines and guanines, 5-halo particularly 5-bromo, 5-trifluoromethyl and other 5-substituted uracils and cytosines, 7-methylguanine and 7-methyladenine, 8-azaguanine and 8-azaadenine, 7-deazaguanine and 7-deazaadenine and 3-deazaguanine and 3-deazaadenine. Further nucleobases include those disclosed in United States Patent No. 3,687,808, those disclosed in *The Concise Encyclopedia Of Polymer Science And Engineering*, pages 858-859, Kroschwitz, J.I., ed. John Wiley & Sons, 1990, those disclosed by Englisch et al., *Angewandte Chemie*, International Edition, 1991, 30, 613, and those disclosed by Sanghvi, Y.S., Chapter 15, *Antisense Research and Applications*, pages 289-302, Crooke, S.T. and Lebleu, B., ed., CRC Press, 1993. Certain of these nucleobases are particularly useful for increasing the binding affinity of the oligomeric compounds of the invention. These include 5-substituted pyrimidines, 6-azapyrimidines and N-2, N-6 and O-6 substituted purines, including 2-aminopropyladenine, 5-propynyluracil and 5-propynylcytosine. 5-methylcytosine substitutions have been shown to increase nucleic acid duplex stability by 0.6-1.2 C (Sanghvi, Y.S., Crooke, S.T. and Lebleu, B., eds., *Antisense Research and Applications*, CRC Press, Boca Raton, 1993, pp. 276-278) and are presently preferred base substitutions, even more particularly when combined with 2'-O-methoxyethyl sugar modifications.

Representative United States patents that teach the preparation of certain of the above noted modified nucleobases as well as other modified nucleobases include, but are not limited to, the above noted U.S. 3,687,808, as well as U.S.: 4,845,205; 5,130,302; 5,134,066; 5,175,273; 5,367,066; 5,432,272; 5,457,187;

5,459,255; 5,484,908; 5,502,177; 5,525,711; 5,552,540;
5,587,469; 5,594,121, 5,596,091; 5,614,617; and
5,681,941, certain of which are commonly owned with the
instant application, and each of which is herein
5 incorporated by reference, and United States patent
5,750,692, which is commonly owned with the instant
application and also herein incorporated by reference.

Another modification of the oligonucleotides of the
invention involves chemically linking to the oligo-
10 nucleotide one or more moieties or conjugates which
enhance the activity, cellular distribution or cellular
uptake of the oligonucleotide. Such moieties include but
are not limited to lipid moieties such as a cholesterol
moiety (Letsinger et al., *Proc. Natl. Acad. Sci. USA*,
15 **1989**, *86*, 6553-6556), cholic acid (Manoharan et al.,
Bioorg. Med. Chem. Lett., **1994**, *4*, 1053-1060), a
thioether, e.g., hexyl-S-tritylthiol (Manoharan et al.,
Ann. N.Y. Acad. Sci., **1992**, *660*, 306-309; Manoharan et
al., *Bioorg. Med. Chem. Lett.*, **1993**, *3*, 2765-2770), a
20 thiocholesterol (Oberhauser et al., *Nucl. Acids Res.*,
1992, *20*, 533-538), an aliphatic chain, e.g., dodecandiol
or undecyl residues (Saison-Behmoaras et al., *EMBO J.*,
1991, *10*, 1111-1118; Kabanov et al., *FEBS Lett.*, **1990**,
259, 327-330; Svinarchuk et al., *Biochimie*, **1993**, *75*, 49-
25 54), a phospholipid, e.g., di-hexadecyl-rac-glycerol or
triethylammonium 1,2-di-O-hexadecyl-rac-glycero-3-H-
phosphonate (Manoharan et al., *Tetrahedron Lett.*, **1995**,
36, 3651-3654; Shea et al., *Nucl. Acids Res.*, **1990**, *18*,
3777-3783), a polyamine or a polyethylene glycol chain
30 (Manoharan et al., *Nucleosides & Nucleotides*, **1995**, *14*,
969-973), or adamantane acetic acid (Manoharan et al.,
Tetrahedron Lett., **1995**, *36*, 3651-3654), a palmityl

moiety (Mishra et al., *Biochim. Biophys. Acta*, **1995**,
1264, 229-237), or an octadecylamine or hexylamino-
carbonyl-oxycholesterol moiety (Crooke et al., *J.*
Pharmacol. Exp. Ther., **1996**, 277, 923-937.

5 Representative United States patents that teach the
preparation of such oligonucleotide conjugates include,
but are not limited to, U.S.: 4,828,979; 4,948,882;
5,218,105; 5,525,465; 5,541,313; 5,545,730; 5,552,538;
5,578,717; 5,580,731; 5,580,731; 5,591,584; 5,109,124;
10 5,118,802; 5,138,045; 5,414,077; 5,486,603; 5,512,439;
5,578,718; 5,608,046; 4,587,044; 4,605,735; 4,667,025;
4,762,779; 4,789,737; 4,824,941; 4,835,263; 4,876,335;
4,904,582; 4,958,013; 5,082,830; 5,112,963; 5,214,136;
5,082,830; 5,112,963; 5,214,136; 5,245,022; 5,254,469;
15 5,258,506; 5,262,536; 5,272,250; 5,292,873; 5,317,098;
5,371,241; 5,391,723; 5,416,203; 5,451,463; 5,510,475;
5,512,667; 5,514,785; 5,565,552; 5,567,810; 5,574,142;
5,585,481; 5,587,371; 5,595,726; 5,597,696; 5,599,923;
5,599,928 and 5,688,941, certain of which are commonly
20 owned with the instant application, and each of which is
herein incorporated by reference.

It is not necessary for all positions in a given
compound to be uniformly modified, and in fact more than
one of the aforementioned modifications may be
25 incorporated in a single compound or even at a single
nucleoside within an oligonucleotide. The present
invention also includes antisense compounds which are
chimeric compounds. "Chimeric" antisense compounds or
"chimeras," in the context of this invention, are
30 antisense compounds, particularly oligonucleotides, which
contain two or more chemically distinct regions, each
made up of at least one monomer unit, i.e., a nucleotide
in the case of an oligonucleotide compound. These

oligonucleotides typically contain at least one region wherein the oligonucleotide is modified so as to confer upon the oligonucleotide increased resistance to nuclease degradation, increased cellular uptake, and/or increased binding affinity for the target nucleic acid. An additional region of the oligonucleotide may serve as a substrate for enzymes capable of cleaving RNA:DNA or RNA:RNA hybrids. By way of example, RNase H is a cellular endonuclease which cleaves the RNA strand of an RNA:DNA duplex. Activation of RNase H, therefore, results in cleavage of the RNA target, thereby greatly enhancing the efficiency of oligonucleotide inhibition of gene expression. Consequently, comparable results can often be obtained with shorter oligonucleotides when chimeric oligonucleotides are used, compared to phosphorothioate deoxyoligonucleotides hybridizing to the same target region. Cleavage of the RNA target can be routinely detected by gel electrophoresis and, if necessary, associated nucleic acid hybridization techniques known in the art.

Chimeric antisense compounds of the invention may be formed as composite structures of two or more oligonucleotides, modified oligonucleotides, oligonucleosides and/or oligonucleotide mimetics as described above. Such compounds have also been referred to in the art as hybrids or gapmers. Representative United States patents that teach the preparation of such hybrid structures include, but are not limited to, U.S.: 5,013,830; 5,149,797; 5,220,007; 5,256,775; 5,366,878; 5,403,711; 5,491,133; 5,565,350; 5,623,065; 5,652,355; 5,652,356; and 5,700,922, certain of which are commonly owned with the instant application, and each of which is herein incorporated by reference in its entirety.

The antisense compounds used in accordance with this invention may be conveniently and routinely made through the well-known technique of solid phase synthesis. Equipment for such synthesis is sold by
5 several vendors including, for example, Applied Biosystems (Foster City, CA). Any other means for such synthesis known in the art may additionally or alternatively be employed. It is well known to use similar techniques to prepare oligonucleotides such as
10 the phosphorothioates and alkylated derivatives.

The antisense compounds of the invention are synthesized *in vitro* and do not include antisense compositions of biological origin, or genetic vector constructs designed to direct the *in vivo* synthesis of
15 antisense molecules. The compounds of the invention may also be admixed, encapsulated, conjugated or otherwise associated with other molecules, molecule structures or mixtures of compounds, as for example, liposomes, receptor targeted molecules, oral, rectal, topical or
20 other formulations, for assisting in uptake, distribution and/or absorption. Representative United States patents that teach the preparation of such uptake, distribution and/or absorption assisting formulations include, but are not limited to, U.S.: 5,108,921; 5,354,844; 5,416,016;
25 5,459,127; 5,521,291; 5,543,158; 5,547,932; 5,583,020; 5,591,721; 4,426,330; 4,534,899; 5,013,556; 5,108,921; 5,213,804; 5,227,170; 5,264,221; 5,356,633; 5,395,619; 5,416,016; 5,417,978; 5,462,854; 5,469,854; 5,512,295; 5,527,528; 5,534,259; 5,543,152; 5,556,948; 5,580,575;
30 and 5,595,756, each of which is herein incorporated by reference.

The antisense compounds of the invention encompass any pharmaceutically acceptable salts, esters, or salts

of such esters, or any other compound which, upon administration to an animal including a human, is capable of providing (directly or indirectly) the biologically active metabolite or residue thereof. Accordingly, for example, the disclosure is also drawn to prodrugs and pharmaceutically acceptable salts of the compounds of the invention, pharmaceutically acceptable salts of such prodrugs, and other bioequivalents.

The term "prodrug" indicates a therapeutic agent that is prepared in an inactive form that is converted to an active form (i.e., drug) within the body or cells thereof by the action of endogenous enzymes or other chemicals and/or conditions. In particular, prodrug versions of the oligonucleotides of the invention are prepared as SATE [(S-acetyl-2-thioethyl) phosphate] derivatives according to the methods disclosed in WO 93/24510 to Gosselin et al., published December 9, 1993 or in WO 94/26764 to Imbach et al.

The term "pharmaceutically acceptable salts" refers to physiologically and pharmaceutically acceptable salts of the compounds of the invention: i.e., salts that retain the desired biological activity of the parent compound and do not impart undesired toxicological effects thereto.

Pharmaceutically acceptable base addition salts are formed with metals or amines, such as alkali and alkaline earth metals or organic amines. Examples of metals used as cations are sodium, potassium, magnesium, calcium, and the like. Examples of suitable amines are N,N'-dibenzylethylenediamine, chloroprocaine, choline, diethanolamine, dicyclohexylamine, ethylenediamine, N-methylglucamine, and procaine (see, for example, Berge et al., "Pharmaceutical Salts," *J. of Pharma Sci.*, 1977, 66, 1-

19). The base addition salts of said acidic compounds are prepared by contacting the free acid form with a sufficient amount of the desired base to produce the salt in the conventional manner. The free acid form may be regenerated by contacting the salt form with an acid and isolating the free acid in the conventional manner. The free acid forms differ from their respective salt forms somewhat in certain physical properties such as solubility in polar solvents, but otherwise the salts are equivalent to their respective free acid for purposes of the present invention. As used herein, a "pharmaceutical addition salt" includes a pharmaceutically acceptable salt of an acid form of one of the components of the compositions of the invention. These include organic or inorganic acid salts of the amines. Preferred acid salts are the hydrochlorides, acetates, salicylates, nitrates and phosphates. Other suitable pharmaceutically acceptable salts are well known to those skilled in the art and include basic salts of a variety of inorganic and organic acids, such as, for example, with inorganic acids, such as for example hydrochloric acid, hydrobromic acid, sulfuric acid or phosphoric acid; with organic carboxylic, sulfonic, sulfo or phospho acids or N-substituted sulfamic acids, for example acetic acid, propionic acid, glycolic acid, succinic acid, maleic acid, hydroxymaleic acid, methylmaleic acid, fumaric acid, malic acid, tartaric acid, lactic acid, oxalic acid, gluconic acid, glucaric acid, glucuronic acid, citric acid, benzoic acid, cinnamic acid, mandelic acid, salicylic acid, 4-aminosalicylic acid, 2-phenoxybenzoic acid, 2-acetoxybenzoic acid, embonic acid, nicotinic acid or isonicotinic acid; and with amino acids, such as the 20 alpha-amino acids involved in the synthesis of

proteins in nature, for example glutamic acid or aspartic acid, and also with phenylacetic acid, methanesulfonic acid, ethanesulfonic acid, 2-hydroxyethanesulfonic acid, ethane-1,2-disulfonic acid, benzenesulfonic acid, 4-methylbenzenesulfonic acid, naphthalene-2-sulfonic acid, naphthalene-1,5-disulfonic acid, 2- or 3-phosphoglycerate, glucose-6-phosphate, N-cyclohexylsulfamic acid (with the formation of cyclamates), or with other acid organic compounds, such as ascorbic acid.

Pharmaceutically acceptable salts of compounds may also be prepared with a pharmaceutically acceptable cation. Suitable pharmaceutically acceptable cations are well known to those skilled in the art and include alkaline, alkaline earth, ammonium and quaternary ammonium cations. Carbonates or hydrogen carbonates are also possible.

For oligonucleotides, preferred examples of pharmaceutically acceptable salts include but are not limited to (a) salts formed with cations such as sodium, potassium, ammonium, magnesium, calcium, polyamines such as spermine and spermidine, etc.; (b) acid addition salts formed with inorganic acids, for example hydrochloric acid, hydrobromic acid, sulfuric acid, phosphoric acid, nitric acid and the like; (c) salts formed with organic acids such as, for example, acetic acid, oxalic acid, tartaric acid, succinic acid, maleic acid, fumaric acid, gluconic acid, citric acid, malic acid, ascorbic acid, benzoic acid, tannic acid, palmitic acid, alginic acid, polyglutamic acid, naphthalenesulfonic acid, methanesulfonic acid, p-toluenesulfonic acid, naphthalenedisulfonic acid, polygalacturonic acid, and the like; and (d) salts formed from elemental anions such as chlorine, bromine, and iodine.

The antisense compounds of the present invention can be utilized for diagnostics, therapeutics, prophylaxis and as research reagents and kits. For therapeutics, an animal, preferably a human, suspected of having a disease or disorder which can be treated by modulating the expression of PTP1B is treated by administering antisense compounds in accordance with this invention. The compounds of the invention can be utilized in pharmaceutical compositions by adding an effective amount of an antisense compound to a suitable pharmaceutically acceptable diluent or carrier. Use of the antisense compounds and methods of the invention may also be useful prophylactically, e.g., to prevent or delay infection, inflammation or tumor formation, for example.

The antisense compounds of the invention are useful for research and diagnostics, because these compounds hybridize to nucleic acids encoding PTP1B, enabling sandwich and other assays to easily be constructed to exploit this fact. Hybridization of the antisense oligonucleotides of the invention with a nucleic acid encoding PTP1B can be detected by means known in the art. Such means may include conjugation of an enzyme to the oligonucleotide, radiolabelling of the oligonucleotide or any other suitable detection means. Kits using such detection means for detecting the level of PTP1B in a sample may also be prepared.

The present invention also includes pharmaceutical compositions and formulations which include the antisense compounds of the invention. The pharmaceutical compositions of the present invention may be administered in a number of ways depending upon whether local or systemic treatment is desired and upon the area to be

treated. Administration may be topical (including ophthalmic and to mucous membranes including vaginal and rectal delivery), pulmonary, e.g., by inhalation or insufflation of powders or aerosols, including by nebulizer; intratracheal, intranasal, epidermal and transdermal), oral or parenteral. Parenteral administration includes intravenous, intraarterial, subcutaneous, intraperitoneal or intramuscular injection or infusion; or intracranial, e.g., intrathecal or intraventricular, administration. Oligonucleotides with at least one 2'-O-methoxyethyl modification are believed to be particularly useful for oral administration.

Pharmaceutical compositions and formulations for topical administration may include transdermal patches, ointments, lotions, creams, gels, drops, suppositories, sprays, liquids and powders. Conventional pharmaceutical carriers, aqueous, powder or oily bases, thickeners and the like may be necessary or desirable. Coated condoms, gloves and the like may also be useful.

Compositions and formulations for oral administration include powders or granules, suspensions or solutions in water or non-aqueous media, capsules, sachets or tablets. Thickeners, flavoring agents, diluents, emulsifiers, dispersing aids or binders may be desirable.

Compositions and formulations for parenteral, intrathecal or intraventricular administration may include sterile aqueous solutions which may also contain buffers, diluents and other suitable additives such as, but not limited to, penetration enhancers, carrier compounds and other pharmaceutically acceptable carriers or excipients.

Pharmaceutical compositions of the present invention include, but are not limited to, solutions, emulsions, and liposome-containing formulations. These compositions may be generated from a variety of components that include, but are not limited to, preformed liquids, self-emulsifying solids and self-emulsifying semisolids.

The pharmaceutical formulations of the present invention, which may conveniently be presented in unit dosage form, may be prepared according to conventional techniques well known in the pharmaceutical industry. Such techniques include the step of bringing into association the active ingredients with the pharmaceutical carrier(s) or excipient(s). In general the formulations are prepared by uniformly and intimately bringing into association the active ingredients with liquid carriers or finely divided solid carriers or both, and then, if necessary, shaping the product.

The compositions of the present invention may be formulated into any of many possible dosage forms such as, but not limited to, tablets, capsules, liquid syrups, soft gels, suppositories, and enemas. The compositions of the present invention may also be formulated as suspensions in aqueous, non-aqueous or mixed media. Aqueous suspensions may further contain substances which increase the viscosity of the suspension including, for example, sodium carboxymethylcellulose, sorbitol and/or dextran. The suspension may also contain stabilizers.

In one embodiment of the present invention the pharmaceutical compositions may be formulated and used as foams. Pharmaceutical foams include formulations such as, but not limited to, emulsions, microemulsions, creams, jellies and liposomes. While basically similar

in nature these formulations vary in the components and the consistency of the final product. The preparation of such compositions and formulations is generally known to those skilled in the pharmaceutical and formulation arts and may be applied to the formulation of the compositions of the present invention.

Emulsions: The compositions of the present invention may be prepared and formulated as emulsions. Emulsions are typically heterogenous systems of one liquid dispersed in another in the form of droplets usually exceeding 0.1 μ m in diameter (Idson, in *Pharmaceutical Dosage Forms*, Lieberman, Rieger and Banker (Eds.), 1988, Marcel Dekker, Inc., New York, N.Y., volume 1, p. 199; Rosoff, in *Pharmaceutical Dosage Forms*, Lieberman, Rieger and Banker (Eds.), 1988, Marcel Dekker, Inc., New York, N.Y., Volume 1, p. 245; Block in *Pharmaceutical Dosage Forms*, Lieberman, Rieger and Banker (Eds.), 1988, Marcel Dekker, Inc., New York, N.Y., volume 2, p. 335; Higuchi et al., in *Remington's Pharmaceutical Sciences*, Mack Publishing Co., Easton, PA, 1985, p. 301). Emulsions are often biphasic systems comprising of two immiscible liquid phases intimately mixed and dispersed with each other. In general, emulsions may be either water-in-oil (w/o) or of the oil-in-water (o/w) variety. When an aqueous phase is finely divided into and dispersed as minute droplets into a bulk oily phase the resulting composition is called a water-in-oil (w/o) emulsion. Alternatively, when an oily phase is finely divided into and dispersed as minute droplets into a bulk aqueous phase the resulting composition is called an oil-in-water (o/w) emulsion. Emulsions may contain additional components in addition to the dispersed phases and the active drug which may be present as a solution in

either the aqueous phase, oily phase or itself as a separate phase. Pharmaceutical excipients such as emulsifiers, stabilizers, dyes, and anti-oxidants may also be present in emulsions as needed. Pharmaceutical emulsions may also be multiple emulsions that are comprised of more than two phases such as, for example, in the case of oil-in-water-in-oil (o/w/o) and water-in-oil-in-water (w/o/w) emulsions. Such complex formulations often provide certain advantages that simple binary emulsions do not. Multiple emulsions in which individual oil droplets of an o/w emulsion enclose small water droplets constitute a w/o/w emulsion. Likewise a system of oil droplets enclosed in globules of water stabilized in an oily continuous provides an o/w/o emulsion.

Emulsions are characterized by little or no thermodynamic stability. Often, the dispersed or discontinuous phase of the emulsion is well dispersed into the external or continuous phase and maintained in this form through the means of emulsifiers or the viscosity of the formulation. Either of the phases of the emulsion may be a semisolid or a solid, as is the case of emulsion-style ointment bases and creams. Other means of stabilizing emulsions entail the use of emulsifiers that may be incorporated into either phase of the emulsion. Emulsifiers may broadly be classified into four categories: synthetic surfactants, naturally occurring emulsifiers, absorption bases, and finely dispersed solids (Idson, in *Pharmaceutical Dosage Forms*, Lieberman, Rieger and Banker (Eds.), 1988, Marcel Dekker, Inc., New York, N.Y., volume 1, p. 199).

Synthetic surfactants, also known as surface active agents, have found wide applicability in the formulation

of emulsions and have been reviewed in the literature
(Rieger, in *Pharmaceutical Dosage Forms*, Lieberman,
Rieger and Banker (Eds.), 1988, Marcel Dekker, Inc., New
York, N.Y., volume 1, p. 285; Idson, in *Pharmaceutical*
5 *Dosage Forms*, Lieberman, Rieger and Banker (Eds.), Marcel
Dekker, Inc., New York, N.Y., 1988, volume 1, p. 199).
Surfactants are typically amphiphilic and comprise a
hydrophilic and a hydrophobic portion. The ratio of the
hydrophilic to the hydrophobic nature of the surfactant
10 has been termed the hydrophile/lipophile balance (HLB)
and is a valuable tool in categorizing and selecting
surfactants in the preparation of formulations.
Surfactants may be classified into different classes
based on the nature of the hydrophilic group: nonionic,
15 anionic, cationic and amphoteric (Rieger, in
Pharmaceutical Dosage Forms, Lieberman, Rieger and Banker
(Eds.), 1988, Marcel Dekker, Inc., New York, N.Y.,
volume 1, p. 285).
Naturally occurring emulsifiers used in emulsion
20 formulations include lanolin, beeswax, phosphatides,
lecithin and acacia. Absorption bases possess
hydrophilic properties such that they can soak up water
to form w/o emulsions yet retain their semisolid
consistencies, such as anhydrous lanolin and hydrophilic
25 petrolatum. Finely divided solids have also been used as
good emulsifiers especially in combination with
surfactants and in viscous preparations. These include
polar inorganic solids, such as heavy metal hydroxides,
nonswelling clays such as bentonite, attapulgate,
30 hectorite, kaolin, montmorillonite, colloidal aluminum
silicate and colloidal magnesium aluminum silicate,
pigments and nonpolar solids such as carbon or glyceryl
tristearate.

A large variety of non-emulsifying materials are also included in emulsion formulations and contribute to the properties of emulsions. These include fats, oils, waxes, fatty acids, fatty alcohols, fatty esters, humectants, hydrophilic colloids, preservatives and antioxidants (Block, in *Pharmaceutical Dosage Forms*, Lieberman, Rieger and Banker (Eds.), 1988, Marcel Dekker, Inc., New York, N.Y., volume 1, p. 335; Idson, in *Pharmaceutical Dosage Forms*, Lieberman, Rieger and Banker (Eds.), 1988, Marcel Dekker, Inc., New York, N.Y., volume 1, p. 199).

Hydrophilic colloids or hydrocolloids include naturally occurring gums and synthetic polymers such as polysaccharides (for example, acacia, agar, alginic acid, carrageenan, guar gum, karaya gum, and tragacanth), cellulose derivatives (for example, carboxymethyl-cellulose and carboxypropylcellulose), and synthetic polymers (for example, carbomers, cellulose ethers, and carboxyvinyl polymers). These disperse or swell in water to form colloidal solutions that stabilize emulsions by forming strong interfacial films around the dispersed-phase droplets and by increasing the viscosity of the external phase.

Since emulsions often contain a number of ingredients such as carbohydrates, proteins, sterols and phosphatides that may readily support the growth of microbes, these formulations often incorporate preservatives. Commonly used preservatives included in emulsion formulations include methyl paraben, propyl paraben, quaternary ammonium salts, benzalkonium chloride, esters of p-hydroxybenzoic acid, and boric acid. Antioxidants are also commonly added to emulsion formulations to prevent deterioration of the formulation.

Antioxidants used may be free radical scavengers such as tocopherols, alkyl gallates, butylated hydroxyanisole, butylated hydroxytoluene, or reducing agents such as ascorbic acid and sodium metabisulfite, and antioxidant synergists such as citric acid, tartaric acid, and lecithin.

The application of emulsion formulations via dermatological, oral and parenteral routes and methods for their manufacture have been reviewed in the literature (Idson, in *Pharmaceutical Dosage Forms*, Lieberman, Rieger and Banker (Eds.), 1988, Marcel Dekker, Inc., New York, N.Y., volume 1, p. 199). Emulsion formulations for oral delivery have been very widely used because of reasons of ease of formulation, efficacy from an absorption and bioavailability standpoint. (Rosoff, in *Pharmaceutical Dosage Forms*, Lieberman, Rieger and Banker (Eds.), 1988, Marcel Dekker, Inc., New York, N.Y., volume 1, p. 245; Idson, in *Pharmaceutical Dosage Forms*, Lieberman, Rieger and Banker (Eds.), 1988, Marcel Dekker, Inc., New York, N.Y., volume 1, p. 199). Mineral-oil base laxatives, oil-soluble vitamins and high fat nutritive preparations are among the materials that have commonly been administered orally as o/w emulsions.

In one embodiment of the present invention, the compositions of oligonucleotides and nucleic acids are formulated as microemulsions. A microemulsion may be defined as a system of water, oil and amphiphile which is a single optically isotropic and thermodynamically stable liquid solution (Rosoff, in *Pharmaceutical Dosage Forms*, Lieberman, Rieger and Banker (Eds.), 1988, Marcel Dekker, Inc., New York, N.Y., volume 1, p. 245). Typically microemulsions are systems that are prepared by first dispersing an oil in an aqueous surfactant solution and

then adding a sufficient amount of a fourth component, generally an intermediate chain-length alcohol to form a transparent system. Therefore, microemulsions have also been described as thermodynamically stable, isotropically clear dispersions of two immiscible liquids that are stabilized by interfacial films of surface-active molecules (Leung and Shah, in: *Controlled Release of Drugs: Polymers and Aggregate Systems*, Rosoff, M., Ed., 1989, VCH Publishers, New York, pages 185-215). Microemulsions commonly are prepared via a combination of three to five components that include oil, water, surfactant, cosurfactant and electrolyte. Whether the microemulsion is of the water-in-oil (w/o) or an oil-in-water (o/w) type is dependent on the properties of the oil and surfactant used and on the structure and geometric packing of the polar heads and hydrocarbon tails of the surfactant molecules (Schott, in *Remington's Pharmaceutical Sciences*, Mack Publishing Co., Easton, PA, 1985, p. 271).

The phenomenological approach utilizing phase diagrams has been extensively studied and has yielded a comprehensive knowledge, to one skilled in the art, of how to formulate microemulsions (Rosoff, in *Pharmaceutical Dosage Forms*, Lieberman, Rieger and Banker (Eds.), 1988, Marcel Dekker, Inc., New York, N.Y., volume 1, p. 245; Block, in *Pharmaceutical Dosage Forms*, Lieberman, Rieger and Banker (Eds.), 1988, Marcel Dekker, Inc., New York, N.Y., volume 1, p. 335). Compared to conventional emulsions, microemulsions offer the advantage of solubilizing water-insoluble drugs in a formulation of thermodynamically stable droplets that are formed spontaneously.

Surfactants used in the preparation of microemulsions include, but are not limited to, ionic surfactants, non-ionic surfactants, Brij 96, polyoxyethylene oleyl ethers, polyglycerol fatty acid esters, tetraglycerol monolaurate (ML310), tetraglycerol monooleate (MO310), hexaglycerol monooleate (PO310), hexaglycerol pentaoleate (PO500), decaglycerol monocaprate (MCA750), decaglycerol monooleate (MO750), decaglycerol sequioleate (SO750), decaglycerol decaoleate (DAO750), alone or in combination with cosurfactants. The cosurfactant, usually a short-chain alcohol such as ethanol, 1-propanol, and 1-butanol, serves to increase the interfacial fluidity by penetrating into the surfactant film and consequently creating a disordered film because of the void space generated among surfactant molecules. Microemulsions may, however, be prepared without the use of cosurfactants and alcohol-free self-emulsifying microemulsion systems are known in the art. The aqueous phase may typically be, but is not limited to, water, an aqueous solution of the drug, glycerol, PEG300, PEG400, polyglycerols, propylene glycols, and derivatives of ethylene glycol. The oil phase may include, but is not limited to, materials such as Captex 300, Captex 355, Capmul MCM, fatty acid esters, medium chain (C8-C12) mono, di, and tri-glycerides, polyoxyethylated glyceryl fatty acid esters, fatty alcohols, polyglycolized glycerides, saturated polyglycolized C8-C10 glycerides, vegetable oils and silicone oil.

Microemulsions are particularly of interest from the standpoint of drug solubilization and the enhanced absorption of drugs. Lipid based microemulsions (both o/w and w/o) have been proposed to enhance the oral

bioavailability of drugs, including peptides
(Constantinides et al., *Pharmaceutical Research*, **1994**,
11, 1385-1390; Ritschel, *Meth. Find. Exp. Clin.*
Pharmacol., **1993**, 13, 205). Microemulsions afford
5 advantages of improved drug solubilization, protection of
drug from enzymatic hydrolysis, possible enhancement of
drug absorption due to surfactant-induced alterations in
membrane fluidity and permeability, ease of preparation,
ease of oral administration over solid dosage forms,
10 improved clinical potency, and decreased toxicity
(Constantinides et al., *Pharmaceutical Research*, **1994**,
11, 1385; Ho et al., *J. Pharm. Sci.*, **1996**, 85, 138-143).
Often microemulsions may form spontaneously when their
components are brought together at ambient temperature.
15 This may be particularly advantageous when formulating
thermolabile drugs, peptides or oligonucleotides.
Microemulsions have also been effective in the
transdermal delivery of active components in both
cosmetic and pharmaceutical applications. It is expected
20 that the microemulsion compositions and formulations of
the present invention will facilitate the increased
systemic absorption of oligonucleotides and nucleic acids
from the gastrointestinal tract, as well as improve the
local cellular uptake of oligonucleotides and nucleic
25 acids within the gastrointestinal tract, vagina, buccal
cavity and other areas of administration.
Microemulsions of the present invention may also
contain additional components and additives such as
sorbitan monostearate (Grill 3), Labrasol, and
30 penetration enhancers to improve the properties of the
formulation and to enhance the absorption of the
oligonucleotides and nucleic acids of the present
invention. Penetration enhancers used in the micro-

emulsions of the present invention may be classified as belonging to one of five broad categories: surfactants, fatty acids, bile salts, chelating agents, and non-chelating non-surfactants (Lee et al., *Critical Reviews in Therapeutic Drug Carrier Systems*, 1991, p. 92). Each of these classes has been discussed above.

Liposomes: There are many organized surfactant structures besides microemulsions that have been studied and used for the formulation of drugs. These include monolayers, micelles, bilayers and vesicles. Vesicles, such as liposomes, have attracted great interest because of their specificity and the duration of action they offer from the standpoint of drug delivery. As used in the present invention, the term "liposome" means a vesicle composed of amphiphilic lipids arranged in a spherical bilayer or bilayers.

Liposomes are unilamellar or multilamellar vesicles which have a membrane formed from a lipophilic material and an aqueous interior. The aqueous portion contains the composition to be delivered. Cationic liposomes possess the advantage of being able to fuse to the cell wall. Non-cationic liposomes, although not able to fuse as efficiently with the cell wall, are taken up by macrophages in vivo.

In order to cross intact mammalian skin, lipid vesicles must pass through a series of fine pores, each with a diameter less than 50 nm, under the influence of a suitable transdermal gradient. Therefore, it is desirable to use a liposome which is highly deformable and able to pass through such fine pores.

Further advantages of liposomes include; liposomes obtained from natural phospholipids are biocompatible and biodegradable; liposomes can incorporate a wide range of

water and lipid soluble drugs; liposomes can protect encapsulated drugs in their internal compartments from metabolism and degradation (Rosoff, in *Pharmaceutical Dosage Forms*, Lieberman, Rieger and Banker (Eds.), 1988, Marcel Dekker, Inc., New York, N.Y., volume 1, p. 245).
5 Important considerations in the preparation of liposome formulations are the lipid surface charge, vesicle size and the aqueous volume of the liposomes.

Liposomes are useful for the transfer and delivery
10 of active ingredients to the site of action. Because the liposomal membrane is structurally similar to biological membranes, when liposomes are applied to a tissue, the liposomes start to merge with the cellular membranes. As the merging of the liposome and cell progresses, the
15 liposomal contents are emptied into the cell where the active agent may act.

Liposomal formulations have been the focus of extensive investigation as the mode of delivery for many drugs. There is growing evidence that for topical
20 administration, liposomes present several advantages over other formulations. Such advantages include reduced side-effects related to high systemic absorption of the administered drug, increased accumulation of the administered drug at the desired target, and the ability
25 to administer a wide variety of drugs, both hydrophilic and hydrophobic, into the skin.

Several reports have detailed the ability of liposomes to deliver agents including high-molecular weight DNA into the skin. Compounds including
30 analgesics, antibodies, hormones and high-molecular weight DNAs have been administered to the skin. The majority of applications resulted in the targeting of the upper epidermis.

Liposomes fall into two broad classes. Cationic liposomes are positively charged liposomes which interact with the negatively charged DNA molecules to form a stable complex. The positively charged DNA/liposome complex binds to the negatively charged cell surface and is internalized in an endosome. Due to the acidic pH within the endosome, the liposomes are ruptured, releasing their contents into the cell cytoplasm (Wang et al., *Biochem. Biophys. Res. Commun.*, **1987**, 147, 980-985).

Liposomes which are pH-sensitive or negatively-charged, entrap DNA rather than complex with it. Since both the DNA and the lipid are similarly charged, repulsion rather than complex formation occurs. Nevertheless, some DNA is entrapped within the aqueous interior of these liposomes. pH-sensitive liposomes have been used to deliver DNA encoding the thymidine kinase gene to cell monolayers in culture. Expression of the exogenous gene was detected in the target cells (Zhou et al., *Journal of Controlled Release*, **1992**, 19, 269-274).

One major type of liposomal composition includes phospholipids other than naturally-derived phosphatidylcholine. Neutral liposome compositions, for example, can be formed from dimyristoyl phosphatidylcholine (DMPC) or dipalmitoyl phosphatidylcholine (DPPC). Anionic liposome compositions generally are formed from dimyristoyl phosphatidylglycerol, while anionic fusogenic liposomes are formed primarily from dioleoyl phosphatidylethanolamine (DOPE). Another type of liposomal composition is formed from phosphatidylcholine (PC) such as, for example, soybean PC, and egg PC. Another type is formed from mixtures of phospholipid and/or phosphatidylcholine and/or cholesterol.

Several studies have assessed the topical delivery of liposomal drug formulations to the skin. Application of liposomes containing interferon to guinea pig skin resulted in a reduction of skin herpes sores while delivery of interferon via other means (e.g. as a solution or as an emulsion) were ineffective (Weiner et al., *Journal of Drug Targeting*, 1992, 2, 405-410). Further, an additional study tested the efficacy of interferon administered as part of a liposomal formulation to the administration of interferon using an aqueous system, and concluded that the liposomal formulation was superior to aqueous administration (du Plessis et al., *Antiviral Research*, 1992, 18, 259-265).

Non-ionic liposomal systems have also been examined to determine their utility in the delivery of drugs to the skin, in particular systems comprising non-ionic surfactant and cholesterol. Non-ionic liposomal formulations comprising NovasomeJ I (glyceryl dilaurate/cholesterol/polyoxyethylene-10-stearyl ether) and NovasomeJ II (glyceryl distearate/cholesterol/ polyoxyethylene-10-stearyl ether) were used to deliver cyclosporin-A into the dermis of mouse skin. Results indicated that such non-ionic liposomal systems were effective in facilitating the deposition of cyclosporin-A into different layers of the skin (Hu et al. *S.T.P. Pharma. Sci.*, 1994, 4, 6, 466).

Liposomes also include "sterically stabilized" liposomes, a term which, as used herein, refers to liposomes comprising one or more specialized lipids that, when incorporated into liposomes, result in enhanced circulation lifetimes relative to liposomes lacking such specialized lipids. Examples of sterically stabilized liposomes are those in which part of the vesicle-forming

lipid portion of the liposome (A) comprises one or more glycolipids, such as monosialoganglioside G_{M1}, or (B) is derivatized with one or more hydrophilic polymers, such as a polyethylene glycol (PEG) moiety. While not wishing to be bound by any particular theory, it is thought in the art that, at least for sterically stabilized liposomes containing gangliosides, sphingomyelin, or PEG-derivatized lipids, the enhanced circulation half-life of these sterically stabilized liposomes derives from a reduced uptake into cells of the reticuloendothelial system (RES) (Allen et al., *FEBS Letters*, **1987**, 223, 42; Wu et al., *Cancer Research*, **1993**, 53, 3765). Various liposomes comprising one or more glycolipids are known in the art. Papahadjopoulos et al. (*Ann. N.Y. Acad. Sci.*, **1987**, 507, 64) reported the ability of monosialoganglioside G_{M1}, galactocerebroside sulfate and phosphatidyl-inositol to improve blood half-lives of liposomes. These findings were expounded upon by Gabizon et al. (*Proc. Natl. Acad. Sci. U.S.A.*, **1988**, 85, 6949). U.S. Patent No. 4,837,028 and WO 88/04924, both to Allen et al., disclose liposomes comprising (1) sphingomyelin and (2) the ganglioside G_{M1} or a galactocerebroside sulfate ester. U.S. Patent No. 5,543,152 (Webb et al.) discloses liposomes comprising sphingomyelin. Liposomes comprising 1,2-*sn*-dimyristoylphosphatidylcholine are disclosed in WO 97/13499 (Lim et al.).

Many liposomes comprising lipids derivatized with one or more hydrophilic polymers, and methods of preparation thereof, are known in the art. Sunamoto et al. (*Bull. Chem. Soc. Jpn.*, **1980**, 53, 2778) described liposomes comprising a nonionic detergent, 2C₁₂15G, that contains a PEG moiety. Illum et al. (*FEBS Lett.*, **1984**,

167, 79) noted that hydrophilic coating of polystyrene particles with polymeric glycols results in significantly enhanced blood half-lives. Synthetic phospholipids modified by the attachment of carboxylic groups of polyalkylene glycols (e.g., PEG) are described by Sears (U.S. Patent Nos. 4,426,330 and 4,534,899). Klivanov et al. (*FEBS Lett.*, 1990, 268, 235) described experiments demonstrating that liposomes comprising phosphatidylethanolamine (PE) derivatized with PEG or PEG stearate have significant increases in blood circulation half-lives. Blume et al. (*Biochimica et Biophysica Acta*, 1990, 1029, 91) extended such observations to other PEG-derivatized phospholipids, e.g., DSPE-PEG, formed from the combination of distearoylphosphatidylethanolamine (DSPE) and PEG. Liposomes having covalently bound PEG moieties on their external surface are described in European Patent No. EP 0 445 131 B1 and WO 90/04384 to Fisher. Liposome compositions containing 1-20 mole percent of PE derivatized with PEG, and methods of use thereof, are described by Woodle et al. (U.S. Patent Nos. 5,013,556 and 5,356,633) and Martin et al. (U.S. Patent No. 5,213,804 and European Patent No. EP 0 496 813 B1). Liposomes comprising a number of other lipid-polymer conjugates are disclosed in WO 91/05545 and U.S. Patent No. 5,225,212 (both to Martin et al.) and in WO 94/20073 (Zalipsky et al.) Liposomes comprising PEG-modified ceramide lipids are described in WO 96/10391 (Choi et al.). U.S. Patent Nos. 5,540,935 (Miyazaki et al.) and 5,556,948 (Tagawa et al.) describe PEG-containing liposomes that can be further derivatized with functional moieties on their surfaces.

A limited number of liposomes comprising nucleic acids are known in the art. WO 96/40062 to Thierry et

al. discloses methods for encapsulating high molecular weight nucleic acids in liposomes. U.S. Patent No. 5,264,221 to Tagawa et al. discloses protein-bonded liposomes and asserts that the contents of such liposomes may include an antisense RNA. U.S. Patent No. 5,665,710 to Rahman et al. describes certain methods of encapsulating oligodeoxynucleotides in liposomes. WO 97/04787 to Love et al. discloses liposomes comprising antisense oligonucleotides targeted to the raf gene.

Transfersomes are yet another type of liposomes, and are highly deformable lipid aggregates which are attractive candidates for drug delivery vehicles. Transfersomes may be described as lipid droplets which are so highly deformable that they are easily able to penetrate through pores which are smaller than the droplet. Transfersomes are adaptable to the environment in which they are used, e.g. they are self-optimizing (adaptive to the shape of pores in the skin), self-repairing, frequently reach their targets without fragmenting, and often self-loading. To make transfersomes it is possible to add surface edge-activators, usually surfactants, to a standard liposomal composition. Transfersomes have been used to deliver serum albumin to the skin. The transfersome-mediated delivery of serum albumin has been shown to be as effective as subcutaneous injection of a solution containing serum albumin.

Surfactants find wide application in formulations such as emulsions (including microemulsions) and liposomes. The most common way of classifying and ranking the properties of the many different types of surfactants, both natural and synthetic, is by the use of the hydrophile/lipophile balance (HLB). The nature of

the hydrophilic group (also known as the "head") provides the most useful means for categorizing the different surfactants used in formulations (Rieger, in *Pharmaceutical Dosage Forms*, Marcel Dekker, Inc., New York, NY, 1988, p. 285).

If the surfactant molecule is not ionized, it is classified as a nonionic surfactant. Nonionic surfactants find wide application in pharmaceutical and cosmetic products and are usable over a wide range of pH values. In general their HLB values range from 2 to about 18 depending on their structure. Nonionic surfactants include nonionic esters such as ethylene glycol esters, propylene glycol esters, glyceryl esters, polyglyceryl esters, sorbitan esters, sucrose esters, and ethoxylated esters. Nonionic alkanolamides and ethers such as fatty alcohol ethoxylates, propoxylated alcohols, and ethoxylated/propoxylated block polymers are also included in this class. The polyoxyethylene surfactants are the most popular members of the nonionic surfactant class.

If the surfactant molecule carries a negative charge when it is dissolved or dispersed in water, the surfactant is classified as anionic. Anionic surfactants include carboxylates such as soaps, acyl lactylates, acyl amides of amino acids, esters of sulfuric acid such as alkyl sulfates and ethoxylated alkyl sulfates, sulfonates such as alkyl benzene sulfonates, acyl isethionates, acyl taurates and sulfosuccinates, and phosphates. The most important members of the anionic surfactant class are the alkyl sulfates and the soaps.

If the surfactant molecule carries a positive charge when it is dissolved or dispersed in water, the surfactant is classified as cationic. Cationic

surfactants include quaternary ammonium salts and ethoxylated amines. The quaternary ammonium salts are the most used members of this class.

5 If the surfactant molecule has the ability to carry either a positive or negative charge, the surfactant is classified as amphoteric. Amphoteric surfactants include acrylic acid derivatives, substituted alkylamides, N-alkylbetaines and phosphatides.

10 The use of surfactants in drug products, formulations and in emulsions has been reviewed (Rieger, in *Pharmaceutical Dosage Forms*, Marcel Dekker, Inc., New York, NY, 1988, p. 285).

15 Penetration Enhancers: In one embodiment, the present invention employs various penetration enhancers to effect the efficient delivery of nucleic acids, particularly oligonucleotides, to the skin of animals. Most drugs are present in solution in both ionized and nonionized forms. However, usually only lipid soluble or lipophilic drugs readily cross cell membranes. It has
20 been discovered that even non-lipophilic drugs may cross cell membranes if the membrane to be crossed is treated with a penetration enhancer. In addition to aiding the diffusion of non-lipophilic drugs across cell membranes, penetration enhancers also enhance the permeability of
25 lipophilic drugs.

Penetration enhancers may be classified as belonging to one of five broad categories, i.e., surfactants, fatty acids, bile salts, chelating agents, and non-chelating non-surfactants (Lee et al., *Critical
30 Reviews in Therapeutic Drug Carrier Systems*, 1991, p.92). Each of the above mentioned classes of penetration enhancers are described below in greater detail.

Surfactants: In connection with the present invention, surfactants (or "surface-active agents") are chemical entities which, when dissolved in an aqueous solution, reduce the surface tension of the solution or the interfacial tension between the aqueous solution and another liquid, with the result that absorption of oligonucleotides through the mucosa is enhanced. In addition to bile salts and fatty acids, these penetration enhancers include, for example, sodium lauryl sulfate, polyoxyethylene-9-lauryl ether and polyoxyethylene-20-cetyl ether) (Lee et al., *Critical Reviews in Therapeutic Drug Carrier Systems*, 1991, p.92); and perfluorochemical emulsions, such as FC-43. Takahashi et al., *J. Pharm. Pharmacol.*, 1988, 40, 252).

Fatty acids: Various fatty acids and their derivatives which act as penetration enhancers include, for example, oleic acid, lauric acid, capric acid (n-decanoic acid), myristic acid, palmitic acid, stearic acid, linoleic acid, linolenic acid, dicaprate, tricaprate, monoolein (1-monooleoyl-rac-glycerol), dilaurin, caprylic acid, arachidonic acid, glycerol 1-monocaprate, 1-dodecylazacycloheptan-2-one, acylcarnitines, acylcholines, C₁₋₁₀ alkyl esters thereof (e.g., methyl, isopropyl and t-butyl), and mono- and diglycerides thereof (i.e., oleate, laurate, caprate, myristate, palmitate, stearate, linoleate, etc.) (Lee et al., *Critical Reviews in Therapeutic Drug Carrier Systems*, 1991, p.92; Muranishi, *Critical Reviews in Therapeutic Drug Carrier Systems*, 1990, 7, 1-33; El Hariri et al., *J. Pharm. Pharmacol.*, 1992, 44, 651-654).

Bile salts: The physiological role of bile includes the facilitation of dispersion and absorption of

lipids and fat-soluble vitamins (Brunton, Chapter 38 in:
 Goodman & Gilman's *The Pharmacological Basis of
 Therapeutics*, 9th Ed., Hardman et al. Eds., McGraw-Hill,
 New York, 1996, pp. 934-935). Various natural bile salts,
 5 and their synthetic derivatives, act as penetration
 enhancers. Thus the term "bile salts" includes any of
 the naturally occurring components of bile as well as any
 of their synthetic derivatives. The bile salts of the
 invention include, for example, cholic acid (or its
 10 pharmaceutically acceptable sodium salt, sodium cholate),
 dehydrocholic acid (sodium dehydrocholate), deoxycholic
 acid (sodium deoxycholate), glucolic acid (sodium
 glucolate), glycholic acid (sodium glycocholate),
 glycodeoxycholic acid (sodium glycodeoxycholate),
 15 taurocholic acid (sodium taurocholate), taurodeoxycholic
 acid (sodium taurodeoxycholate), chenodeoxycholic acid
 (sodium chenodeoxycholate), ursodeoxycholic acid (UDCA),
 sodium tauro-24,25-dihydro-fusidate (STDHF), sodium
 glycodihydrofusidate and polyoxyethylene-9-lauryl ether
 20 (POE) (Lee et al., *Critical Reviews in Therapeutic Drug
 Carrier Systems*, 1991, page 92; Swinyard, Chapter 39 In:
Remington's Pharmaceutical Sciences, 18th Ed., Gennaro,
 ed., Mack Publishing Co., Easton, PA, 1990, pages 782-
 783; Muranishi, *Critical Reviews in Therapeutic Drug
 Carrier Systems*, 1990, 7, 1-33; Yamamoto et al., *J.*
 25 *Pharm. Exp. Ther.*, 1992, 263, 25; Yamashita et al., *J.*
Pharm. Sci., 1990, 79, 579-583).

Chelating Agents: Chelating agents, as used in
 connection with the present invention, can be defined as
 30 compounds that remove metallic ions from solution by
 forming complexes therewith, with the result that
 absorption of oligonucleotides through the mucosa is

enhanced. With regards to their use as penetration enhancers in the present invention, chelating agents have the added advantage of also serving as DNase inhibitors, as most characterized DNA nucleases require a divalent metal ion for catalysis and are thus inhibited by

5 chelating agents (Jarrett, *J. Chromatogr.*, **1993**, 618, 315-339). Chelating agents of the invention include but are not limited to disodium ethylenediaminetetraacetate (EDTA), citric acid, salicylates (e.g., sodium

10 salicylate, 5-methoxysalicylate and homovanilate), *N*-acyl derivatives of collagen, laureth-9 and *N*-amino acyl derivatives of beta-diketones (enamines) (Lee et al., *Critical Reviews in Therapeutic Drug Carrier Systems*, **1991**, page 92; Muranishi, *Critical Reviews in Therapeutic*

15 *Drug Carrier Systems*, **1990**, 7, 1-33; Buur et al., *J. Control Rel.*, **1990**, 14, 43-51).

Non-chelating non-surfactants: As used herein, non-chelating non-surfactant penetration enhancing compounds can be defined as compounds that demonstrate

20 insignificant activity as chelating agents or as surfactants but that nonetheless enhance absorption of oligonucleotides through the alimentary mucosa (Muranishi, *Critical Reviews in Therapeutic Drug Carrier Systems*, **1990**, 7, 1-33). This class of penetration

25 enhancers include, for example, unsaturated cyclic ureas, 1-alkyl- and 1-alkenylazacyclo-alkanone derivatives (Lee et al., *Critical Reviews in Therapeutic Drug Carrier Systems*, **1991**, page 92); and non-steroidal anti-inflammatory agents such as diclofenac sodium,

30 indomethacin and phenylbutazone (Yamashita et al., *J. Pharm. Pharmacol.*, **1987**, 39, 621-626).

Agents that enhance uptake of oligonucleotides at the cellular level may also be added to the pharmaceutical and other compositions of the present invention. For example, cationic lipids, such as
5 lipofectin (Junichi et al, U.S. Patent No. 5,705,188), cationic glycerol derivatives, and polycationic molecules, such as polylysine (Lollo et al., PCT Application WO 97/30731), are also known to enhance the cellular uptake of oligonucleotides.

10 Other agents may be utilized to enhance the penetration of the administered nucleic acids, including glycols such as ethylene glycol and propylene glycol, pyrrols such as 2-pyrrol, azones, and terpenes such as limonene and menthone.

15 Carriers: Certain compositions of the present invention also incorporate carrier compounds in the formulation. As used herein, "carrier compound" or "carrier" can refer to a nucleic acid, or analog thereof, which is inert (*i.e.*, does not possess biological
20 activity *per se*) but is recognized as a nucleic acid by *in vivo* processes that reduce the bioavailability of a nucleic acid having biological activity by, for example, degrading the biologically active nucleic acid or promoting its removal from circulation. The
25 coadministration of a nucleic acid and a carrier compound, typically with an excess of the latter substance, can result in a substantial reduction of the amount of nucleic acid recovered in the liver, kidney or other extracirculatory reservoirs, presumably due to
30 competition between the carrier compound and the nucleic acid for a common receptor. For example, the recovery of a partially phosphorothioate oligonucleotide in hepatic tissue can be reduced when it is coadministered with

polyinosinic acid, dextran sulfate, polycytidic acid or
4-acetamido-4'-isothiocyano-stilbene-2,2'-disulfonic acid
(Miyao *et al.*, *Antisense Res. Dev.*, **1995**, 5, 115-121;
Takakura *et al.*, *Antisense & Nucl. Acid Drug Dev.*, **1996**,
5 6, 177-183).

Excipients: In contrast to a carrier compound, a
"pharmaceutical carrier" or "excipient" is a
pharmaceutically acceptable solvent, suspending agent or
any other pharmacologically inert vehicle for delivering
10 one or more nucleic acids to an animal. The excipient
may be liquid or solid and is selected, with the planned
manner of administration in mind, so as to provide for
the desired bulk, consistency, etc., when combined with a
nucleic acid and the other components of a given
15 pharmaceutical composition. Typical pharmaceutical
carriers include, but are not limited to, binding agents
(e.g., pregelatinized maize starch, polyvinylpyrrolidone
or hydroxypropyl methylcellulose, etc.); fillers (e.g.,
lactose and other sugars, microcrystalline cellulose,
20 pectin, gelatin, calcium sulfate, ethyl cellulose,
polyacrylates or calcium hydrogen phosphate, etc.);
lubricants (e.g., magnesium stearate, talc, silica,
colloidal silicon dioxide, stearic acid, metallic
stearates, hydrogenated vegetable oils, corn starch,
25 polyethylene glycols, sodium benzoate, sodium acetate,
etc.); disintegrants (e.g., starch, sodium starch
glycolate, etc.); and wetting agents (e.g., sodium lauryl
sulphate, etc.).

Pharmaceutically acceptable organic or inorganic
30 excipient suitable for non-parenteral administration
which do not deleteriously react with nucleic acids can
also be used to formulate the compositions of the present
invention. Suitable pharmaceutically acceptable carriers

include, but are not limited to, water, salt solutions, alcohols, polyethylene glycols, gelatin, lactose, amylose, magnesium stearate, talc, silicic acid, viscous paraffin, hydroxymethylcellulose, polyvinylpyrrolidone and the like.

Formulations for topical administration of nucleic acids may include sterile and non-sterile aqueous solutions, non-aqueous solutions in common solvents such as alcohols, or solutions of the nucleic acids in liquid or solid oil bases. The solutions may also contain buffers, diluents and other suitable additives. Pharmaceutically acceptable organic or inorganic excipients suitable for non-parenteral administration which do not deleteriously react with nucleic acids can be used.

Suitable pharmaceutically acceptable excipients include, but are not limited to, water, salt solutions, alcohol, polyethylene glycols, gelatin, lactose, amylose, magnesium stearate, talc, silicic acid, viscous paraffin, hydroxymethylcellulose, polyvinylpyrrolidone and the like.

Other Components: The compositions of the present invention may additionally contain other adjunct components conventionally found in pharmaceutical compositions, at their art-established usage levels. Thus, for example, the compositions may contain additional, compatible, pharmaceutically-active materials such as, for example, antipruritics, astringents, local anesthetics or anti-inflammatory agents, or may contain additional materials useful in physically formulating various dosage forms of the compositions of the present invention, such as dyes, flavoring agents, preservatives, antioxidants, opacifiers, thickening agents and

stabilizers. However, such materials, when added, should not unduly interfere with the biological activities of the components of the compositions of the present invention. The formulations can be sterilized and, if
5 desired, mixed with auxiliary agents, e.g., lubricants, preservatives, stabilizers, wetting agents, emulsifiers, salts for influencing osmotic pressure, buffers, colorings, flavorings and/or aromatic substances and the like which do not deleteriously interact with the nucleic
10 acid(s) of the formulation.

Aqueous suspensions may contain substances which increase the viscosity of the suspension including, for example, sodium carboxymethylcellulose, sorbitol and/or dextran. The suspension may also contain stabilizers.

15 Certain embodiments of the invention provide pharmaceutical compositions containing (a) one or more antisense compounds and (b) one or more other chemotherapeutic agents which function by a non-antisense mechanism. Examples of such chemotherapeutic agents
20 include, but are not limited to, anticancer drugs such as daunorubicin, dactinomycin, doxorubicin, bleomycin, mitomycin, nitrogen mustard, chlorambucil, melphalan, cyclophosphamide, 6-mercaptopurine, 6-thioguanine, cytarabine (CA), 5-fluorouracil (5-FU), floxuridine
25 (5-FUdR), methotrexate (MTX), colchicine, vincristine, vinblastine, etoposide, teniposide, cisplatin and diethylstilbestrol (DES). See, generally, *The Merck Manual of Diagnosis and Therapy*, 15th Ed., Berkow et al., eds., 1987, Rahway, N.J., pages 1206-1228). Anti-
30 inflammatory drugs, including but not limited to nonsteroidal anti-inflammatory drugs and corticosteroids, and antiviral drugs, including but not limited to ribivirin, vidarabine, acyclovir and ganciclovir, may

also be combined in compositions of the invention. See, generally, *The Merck Manual of Diagnosis and Therapy*, 15th Ed., Berkow et al., eds., 1987, Rahway, N.J., pages 2499-2506 and 46-49, respectively). Other non-antisense
5 chemotherapeutic agents are also within the scope of this invention. Two or more combined compounds may be used together or sequentially.

In another related embodiment, compositions of the invention may contain one or more antisense compounds,
10 particularly oligonucleotides, targeted to a first nucleic acid and one or more additional antisense compounds targeted to a second nucleic acid target. Numerous examples of antisense compounds are known in the art. Two or more combined compounds may be used together
15 or sequentially.

The formulation of therapeutic compositions and their subsequent administration is believed to be within the skill of those in the art. Dosing is dependent on severity and responsiveness of the disease state to be
20 treated, with the course of treatment lasting from several days to several months, or until a cure is effected or a diminution of the disease state is achieved. Optimal dosing schedules can be calculated from measurements of drug accumulation in the body of the
25 patient. Persons of ordinary skill can easily determine optimum dosages, dosing methodologies and repetition rates. Optimum dosages may vary depending on the relative potency of individual oligonucleotides, and can generally be estimated based on EC₅₀s found to be
30 effective in *in vitro* and *in vivo* animal models. In general, dosage is from 0.01 ug to 100 g per kg of body weight, and may be given once or more daily, weekly, monthly or yearly, or even once every 2 to 20 years.

Persons of ordinary skill in the art can easily estimate repetition rates for dosing based on measured residence times and concentrations of the drug in bodily fluids or tissues. Following successful treatment, it may be
5 desirable to have the patient undergo maintenance therapy to prevent the recurrence of the disease state, wherein the oligonucleotide is administered in maintenance doses, ranging from 0.01 μ g to 100 g per kg of body weight, once or more daily, to once every 20 years.

10 While the present invention has been described with specificity in accordance with certain of its preferred embodiments, the following examples serve only to illustrate the invention and are not intended to limit the same.

15 **EXAMPLES**

EXAMPLE 1: NUCLEOSIDE PHOSPHORAMIDITES FOR

OLIGONUCLEOTIDE SYNTHESIS DEOXY AND 2'-ALKOXY AMIDITES

2'-Deoxy and 2'-methoxy beta-cyanoethyl-diisopropyl
20 phosphoramidites were purchased from commercial sources (e.g. Chemgenes, Needham, MA or Glen Research, Inc., Sterling, VA). Other 2'-O-alkoxy substituted nucleoside amidites are prepared as described in U.S. Patent 5,506,351, herein incorporated by reference. For
25 oligonucleotides synthesized using 2'-alkoxy amidites, the standard cycle for unmodified oligonucleotides was utilized, except the wait step after pulse delivery of tetrazole and base was increased to 360 seconds.

Oligonucleotides containing 5-methyl-2'-
30 deoxycytidine (5-Me-C) nucleotides were synthesized according to published methods (Sanghvi, et al, *Nucleic Acids Research*, 1993, 21, 3197-3203) using commercially

available phosphoramidites (Glen Research, Sterling, VA, or ChemGenes, Needham, MA).

2'-FLUORO AMIDITES

(a) 2'-FLUORODEOXYADENOSINE AMIDITES

5 2'-fluoro oligonucleotides were synthesized as described previously (Kawasaki, *et al*, *J. Med. Chem.*, 1993, 36, 831-841) and United States patent 5,670,633, herein incorporated by reference. Briefly, the protected nucleoside N6-benzoyl-2'-deoxy-2'-fluoroadenosine was
10 synthesized utilizing commercially available 9-beta-D-arabinofuranosyladenine as starting material and by modifying literature procedures whereby the 2'-alpha-fluoro atom is introduced by a S_N2-displacement of a 2'-beta-trityl group. Thus N6-benzoyl-9-beta-D-
15 arabinofuranosyladenine was selectively protected in moderate yield as the 3',5'-ditetrahydropyranyl (THP) intermediate. Deprotection of the THP and N6-benzoyl groups was accomplished using standard methodologies and standard methods were used to obtain the 5'-
20 dimethoxytrityl-(DMT) and 5'-DMT-3'-phosphoramidite intermediates.

(b) 2'-FLUORODEOXYGUANOSINE

The synthesis of 2'-deoxy-2'-fluoroguanosine was accomplished using tetraisopropylidisiloxanyl (TPDS)
25 protected 9-beta-D-arabinofuranosylguanine as starting material, and conversion to the intermediate diisobutyrylarabinofuranosylguanosine. Deprotection of the TPDS group was followed by protection of the hydroxyl group with THP to give diisobutyryl di-THP protected
30 arabinofuranosylguanine. Selective O-deacylation and triflation was followed by treatment of the crude product with fluoride, then deprotection of the THP groups.

Standard methodologies were used to obtain the 5'-DMT- and 5'-DMT-3'-phosphoramidites.

(c) 2'-FLUOROURIDINE

Synthesis of 2'-deoxy-2'-fluorouridine was accomplished by the modification of a literature procedure in which 2,2'-anhydro-1-beta-D-arabinofuranosyluracil was treated with 70% hydrogen fluoride-pyridine. Standard procedures were used to obtain the 5'-DMT and 5'-DMT-3'phosphoramidites.

(d) 2'-FLUORODEOXYCYTIDINE

2'-deoxy-2'-fluorocytidine was synthesized via amination of 2'-deoxy-2'-fluorouridine, followed by selective protection to give N4-benzoyl-2'-deoxy-2'-fluorocytidine. Standard procedures were used to obtain the 5'-DMT and 5'-DMT-3'phosphoramidites.

2'-O-(2-METHOXYETHYL) MODIFIED AMIDITES

2'-O-Methoxyethyl-substituted nucleoside amidites are prepared as follows, or alternatively, as per the methods of Martin, P., *Helvetica Chimica Acta*, 1995, 78, 486-504.

(a) 2,2'-ANHYDRO[1-(BETA-D-ARABINOFURANOSYL)-5-METHYLURIDINE]

5-Methyluridine (ribosylthymine, commercially available through Yamasa, Choshi, Japan) (72.0 g, 0.279 M), diphenylcarbonate (90.0 g, 0.420 M) and sodium bicarbonate (2.0 g, 0.024 M) were added to DMF (300 mL). The mixture was heated to reflux, with stirring, allowing the evolved carbon dioxide gas to be released in a controlled manner. After 1 hour, the slightly darkened solution was concentrated under reduced pressure. The resulting syrup was poured into diethylether (2.5 L), with stirring. The product formed a gum. The ether was decanted and the residue was dissolved in a minimum

amount of methanol (ca. 400 mL). The solution was poured into fresh ether (2.5 L) to yield a stiff gum. The ether was decanted and the gum was dried in a vacuum oven (60 C at 1 mm Hg for 24 h) to give a solid that was crushed to a light tan powder (57 g, 85% crude yield). The NMR spectrum was consistent with the structure, contaminated with phenol as its sodium salt (ca. 5%). The material was used as is for further reactions (or it can be purified further by column chromatography using a gradient of methanol in ethyl acetate (10-25%) to give a white solid, mp 222-4 C).

(b) 2'-O-METHOXYETHYL-5-METHYLURIDINE

2,2'-Anhydro-5-methyluridine (195 g, 0.81 M), tris(2-methoxyethyl)borate (231 g, 0.98 M) and 2-methoxyethanol (1.2 L) were added to a 2 L stainless steel pressure vessel and placed in a pre-heated oil bath at 160 C. After heating for 48 hours at 155-16 C, the vessel was opened and the solution evaporated to dryness and triturated with MeOH (200 mL). The residue was suspended in hot acetone (1 L). The insoluble salts were filtered, washed with acetone (150 mL) and the filtrate evaporated. The residue (280 g) was dissolved in CH₃CN (600 mL) and evaporated. A silica gel column (3 kg) was packed in CH₂Cl₂/acetone/MeOH (20:5:3) containing 0.5% Et₃NH. The residue was dissolved in CH₂Cl₂ (250 mL) and adsorbed onto silica (150 g) prior to loading onto the column. The product was eluted with the packing solvent to give 160 g (63%) of product. Additional material was obtained by reworking impure fractions.

(c) 2'-O-METHOXYETHYL-5'-O-DIMETHOXYTRITYL-5-METHYLURIDINE

2'-O-Methoxyethyl-5-methyluridine (160 g, 0.506 M) was co-evaporated with pyridine (250 mL) and the dried

residue dissolved in pyridine (1.3 L). A first aliquot of dimethoxytrityl chloride (94.3 g, 0.278 M) was added and the mixture stirred at room temperature for one hour. A second aliquot of dimethoxytrityl chloride (94.3 g,
5 0.278 M) was added and the reaction stirred for an additional one hour. Methanol (170 mL) was then added to stop the reaction. HPLC showed the presence of approximately 70% product. The solvent was evaporated and triturated with CH₃CN (200 mL). The residue was
10 dissolved in CHCl₃ (1.5 L) and extracted with 2x500 mL of saturated NaHCO₃ and 2x500 mL of saturated NaCl. The organic phase was dried over Na₂SO₄, filtered and evaporated. 275 g of residue was obtained. The residue was purified on a 3.5 kg silica gel column, packed and
15 eluted with EtOAc/hexane/acetone (5:5:1) containing 0.5% Et₃NH. The pure fractions were evaporated to give 164 g of product. Approximately 20 g additional was obtained from the impure fractions to give a total yield of 183 g (57%).

20 (d) 3'-O-ACETYL-2'-O-METHOXYETHYL-5'-O-DIMETHOXYTRITYL-5-METHYLURIDINE
2'-O-Methoxyethyl-5'-O-dimethoxytrityl-5-methyluridine (106 g, 0.167 M), DMF/pyridine (750 mL of a 3:1 mixture prepared from 562 mL of DMF and 188 mL of
25 pyridine) and acetic anhydride (24.38 mL, 0.258 M) were combined and stirred at room temperature for 24 hours. The reaction was monitored by TLC by first quenching the TLC sample with the addition of MeOH. Upon completion of the reaction, as judged by TLC, MeOH (50 mL) was added
30 and the mixture evaporated at 35 C. The residue was dissolved in CHCl₃ (800 mL) and extracted with 2x200 mL of saturated sodium bicarbonate and 2x200 mL of saturated NaCl. The water layers were back extracted with 200 mL

of CHCl_3 . The combined organics were dried with sodium sulfate and evaporated to give 122 g of residue (approx. 90% product). The residue was purified on a 3.5 kg silica gel column and eluted using EtOAc/hexane(4:1). Pure product fractions were evaporated to yield 96 g (84%). An additional 1.5 g was recovered from later fractions.

(e) 3'-O-ACETYL-2'-O-METHOXYETHYL-5'-O-DIMETHOXYTRITYL-5-METHYL-4-TRIAZOLEURIDINE

A first solution was prepared by dissolving 3'-O-acetyl-2'-O-methoxyethyl-5'-O-dimethoxytrityl-5-methyluridine (96 g, 0.144 M) in CH_3CN (700 mL) and set aside. Triethylamine (189 mL, 1.44 M) was added to a solution of triazole (90 g, 1.3 M) in CH_3CN (1 L), cooled to -5 C and stirred for 0.5 h using an overhead stirrer. POCl_3 was added dropwise, over a 30 minute period, to the stirred solution maintained at 0-10 C, and the resulting mixture stirred for an additional 2 hours. The first solution was added dropwise, over a 45 minute period, to the latter solution. The resulting reaction mixture was stored overnight in a cold room. Salts were filtered from the reaction mixture and the solution was evaporated. The residue was dissolved in EtOAc (1 L) and the insoluble solids were removed by filtration. The filtrate was washed with 1x300 mL of NaHCO_3 and 2x300 mL of saturated NaCl, dried over sodium sulfate and evaporated. The residue was triturated with EtOAc to give the title compound.

(f) 2'-O-METHOXYETHYL-5'-O-DIMETHOXYTRITYL-5-METHYLCYTIDINE

A solution of 3'-O-acetyl-2'-O-methoxyethyl-5'-O-dimethoxytrityl-5-methyl-4-triazoleuridine (103 g, 0.141 M) in dioxane (500 mL) and NH_4OH (30 mL) was stirred at

room temperature for 2 hours. The dioxane solution was evaporated and the residue azeotroped with MeOH (2x200 mL). The residue was dissolved in MeOH (300 mL) and transferred to a 2 liter stainless steel pressure vessel.
5 MeOH (400 mL) saturated with NH₃ gas was added and the vessel heated to 100 C for 2 hours (TLC showed complete conversion). The vessel contents were evaporated to dryness and the residue was dissolved in EtOAc (500 mL) and washed once with saturated NaCl (200 mL). The
10 organics were dried over sodium sulfate and the solvent was evaporated to give 85 g (95%) of the title compound.

(g) N4-BENZOYL-2'-O-METHOXYETHYL-5'-O-

DIMETHOXYTRITYL-5-METHYLCYTIDINE

2'-O-Methoxyethyl-5'-O-dimethoxytrityl-5-methyl-
15 cytidine (85 g, 0.134 M) was dissolved in DMF (800 mL) and benzoic anhydride (37.2 g, 0.165 M) was added with stirring. After stirring for 3 hours, TLC showed the reaction to be approximately 95% complete. The solvent was evaporated and the residue azeotroped with MeOH (200
20 mL). The residue was dissolved in CHCl₃ (700 mL) and extracted with saturated NaHCO₃ (2x300 mL) and saturated NaCl (2x300 mL), dried over MgSO₄ and evaporated to give a residue (96 g). The residue was chromatographed on a 1.5 kg silica column using EtOAc/hexane (1:1) containing 0.5%
25 Et₃NH as the eluting solvent. The pure product fractions were evaporated to give 90 g (90%) of the title compound.

(h) N4-BENZOYL-2'-O-METHOXYETHYL-5'-O-

DIMETHOXYTRITYL-5-METHYLCYTIDINE-3'-AMIDITE

N4-Benzoyl-2'-O-methoxyethyl-5'-O-dimethoxytrityl-
30 5-methylcytidine (74 g, 0.10 M) was dissolved in CH₂Cl₂ (1 L). Tetrazole diisopropylamine (7.1 g) and 2-cyanoethoxy-tetra(isopropyl)phosphite (40.5 mL, 0.123 M) were added with stirring, under a nitrogen atmosphere. The

resulting mixture was stirred for 20 hours at room temperature (TLC showed the reaction to be 95% complete). The reaction mixture was extracted with saturated NaHCO₃ (1x300 mL) and saturated NaCl (3x300 mL). The aqueous washes were back-extracted with CH₂Cl₂ (300 mL), and the extracts were combined, dried over MgSO₄ and concentrated. The residue obtained was chromatographed on a 1.5 kg silica column using EtOAc/hexane (3:1) as the eluting solvent. The pure fractions were combined to give 90.6 g (87%) of the title compound.

2'-O-(AMINOXYETHYL) NUCLEOSIDE AMIDITES AND 2'-O-(DIMETHYLAMINOXYETHYL) NUCLEOSIDE AMIDITES

(a) 2'-(DIMETHYLAMINOXYETHOXY) NUCLEOSIDE AMIDITES

2'-(Dimethylaminoxyethoxy) nucleoside amidites [also known in the art as 2'-O-(dimethylaminoxyethyl) nucleoside amidites] are prepared as described in the following paragraphs[COMMENT] [1]Page: 1

T = cpds 102-108 from ISIS-2824. Adenosine, cytidine and guanosine nucleoside amidites are prepared similarly to the thymidine (5-methyluridine) except the exocyclic amines are protected with a benzoyl moiety in the case of adenosine and cytidine and with isobutyryl in the case of guanosine.

(b) 5'-O-TERT-BUTYLDIPHENYLSILYL-O²-2'-ANHYDRO-5-METHYLURIDINE

O²-2'-anhydro-5-methyluridine (Pro. Bio. Sint., Varese, Italy, 100.0 g, 0.416 mmol), dimethylamino-pyridine (0.66 g, 0.013 eq, 0.0054 mmol) were dissolved in dry pyridine (500 mL) at ambient temperature under an argon atmosphere and with mechanical stirring. *tert*-Butyldiphenylchlorosilane (125.8 g, 119.0 mL, 1.1 eq, 0.458 mmol) was added in one portion. The reaction was

stirred for 16 h at ambient temperature. TLC (Rf 0.22, ethyl acetate) indicated a complete reaction. The solution was concentrated under reduced pressure to a thick oil. This was partitioned between dichloromethane (1 L) and saturated sodium bicarbonate (2x1 L) and brine (1 L). The organic layer was dried over sodium sulfate and concentrated under reduced pressure to a thick oil. The oil was dissolved in a 1:1 mixture of ethyl acetate and ethyl ether (600 mL) and the solution was cooled to -10°C. The resulting crystalline product was collected by filtration, washed with ethyl ether (3x200 mL) and dried (40°C, 1mm Hg, 24 h) to 149g (74.8%) of white solid. TLC and NMR were consistent with pure product.

(c) 5'-O-TERT-BUTYLDIPHENYLSILYL-2'-O-(2-HYDROXYETHYL)-5-METHYLURIDINE

In a 2 L stainless steel, unstirred pressure reactor was added borane in tetrahydrofuran (1.0 M, 2.0 eq, 622 mL). In the fume hood and with manual stirring, ethylene glycol (350 mL, excess) was added cautiously at first until the evolution of hydrogen gas subsided. 5'-O-tert-Butyldiphenylsilyl-O²-2'-anhydro-5-methyluridine (149 g, 0.311 mol) and sodium bicarbonate (0.074 g, 0.003 eq) were added with manual stirring. The reactor was sealed and heated in an oil bath until an internal temperature of 160°C was reached and then maintained for 16 h (pressure < 100 psig). The reaction vessel was cooled to ambient and opened. TLC (Rf 0.67 for desired product and Rf 0.82 for ara-T side product, ethyl acetate) indicated about 70% conversion to the product. In order to avoid additional side product formation, the reaction was stopped, concentrated under reduced pressure (10 to 1 mm Hg) in a warm water bath (40-100°C) with the more extreme conditions used to remove the ethylene glycol.

[Alternatively, once the low boiling solvent is gone, the remaining solution can be partitioned between ethyl acetate and water. The product will be in the organic phase.] The residue was purified by column chromatography (2 kg silica gel, ethyl acetate-hexanes gradient 1:1 to 4:1). The appropriate fractions were combined, stripped and dried to product as a white crisp foam (84 g, 50%), contaminated starting material (17.4 g) and pure reusable starting material 20 g. The yield based on starting material less pure recovered starting material was 58%. TLC and NMR were consistent with 99% pure product.

(d) 2'-O-([2-PHTHALIMIDOXY)ETHYL]-5'-T-BUTYLDIPHENYLSILYL-5-METHYLURIDINE
5'-O-tert-Butyldiphenylsilyl-2'-O-(2-hydroxyethyl)-5-methyluridine (20 g, 36.98 mmol) was mixed with triphenylphosphine (11.63 g, 44.36 mmol) and N-hydroxyphthalimide (7.24 g, 44.36 mmol). It was then dried over P₂O₅ under high vacuum for two days at 40 C. The reaction mixture was flushed with argon and dry THF (369.8 mL, Aldrich, sure seal bottle) was added to get a clear solution. Diethyl-azodicarboxylate (6.98 mL, 44.36 mmol) was added dropwise to the reaction mixture. The rate of addition is maintained such that resulting deep red coloration is just discharged before adding the next drop. After the addition was complete, the reaction was stirred for 4 hrs. By that time TLC showed the completion of the reaction (ethylacetate:hexane, 60:40). The solvent was evaporated in vacuum. Residue obtained was placed on a flash column and eluted with ethyl acetate:hexane (60:40), to get 2'-O-([2-phthalimidoxy)ethyl]-5'-t-butyldiphenylsilyl-5-methyluridine as white foam (21.819 g, 86%).

(e) 5'-O-*TERT*-BUTYLDIPHENYLSILYL-2'-O-[(2-
 FORMADOXIMINOXY)ETHYL]-5-METHYLURIDINE
 2'-O-[(2-phthalimidoxy)ethyl]-5'-*t*-
 butyldiphenylsilyl-5-methyluridine (3.1 g, 4.5 mmol) was
 5 dissolved in dry CH₂Cl₂ (4.5 mL) and methylhydrazine (300
 mL, 4.64 mmol) was added dropwise at -10 C to 0 C. After
 1 h the mixture was filtered, the filtrate was washed
 with ice cold CH₂Cl₂ and the combined organic phase was
 washed with water, brine and dried over anhydrous Na₂SO₄.
 10 The solution was concentrated to get 2'-O-(aminooxyethyl)
 thymidine, which was then dissolved in MeOH (67.5 mL).
 To this formaldehyde (20% aqueous solution, w/w, 1.1 eq.)
 was added and the resulting mixture was stirred for 1 h.
 Solvent was removed under vacuum; residue chromatographed
 15 to get 5'-O-*tert*-butyldiphenylsilyl-2'-O-[(2-
 formadoximinooxy) ethyl]-5-methyluridine as white foam
 (1.95 g, 78%).

(f) 5'-O-*TERT*-BUTYLDIPHENYLSILYL-2'-O-[N,N-
 DIMETHYLAMINOXYETHYL]-5-METHYLURIDINE
 20 5'-O-*tert*-butyldiphenylsilyl-2'-O-[(2-
 formadoximinooxy)ethyl]-5-methyluridine (1.77 g, 3.12
 mmol) was dissolved in a solution of 1M pyridinium p-
 toluenesulfonate (PPTS) in dry MeOH (30.6 mL). Sodium
 cyanoborohydride (0.39 g, 6.13 mmol) was added to this
 25 solution at 10 C under inert atmosphere. The reaction
 mixture was stirred for 10 minutes at 10 C. After that
 the reaction vessel was removed from the ice bath and
 stirred at room temperature for 2 h, the reaction
 monitored by TLC (5% MeOH in CH₂Cl₂). Aqueous NaHCO₃
 30 solution (5%, 10 mL) was added and extracted with ethyl
 acetate (2x20 mL). Ethyl acetate phase was dried over
 anhydrous Na₂SO₄, evaporated to dryness. Residue was
 dissolved in a solution of 1M PPTS in MeOH (30.6 mL).

Formaldehyde (20% w/w, 30 mL, 3.37 mmol) was added and the reaction mixture was stirred at room temperature for 10 minutes. Reaction mixture cooled to 10 C in an ice bath, sodium cyanoborohydride (0.39 g, 6.13 mmol) was added and reaction mixture stirred at 10 C for 10 minutes. After 10 minutes, the reaction mixture was removed from the ice bath and stirred at room temperature for 2 hrs. To the reaction mixture 5% NaHCO₃ (25 mL) solution was added and extracted with ethyl acetate (2x25 mL). Ethyl acetate layer was dried over anhydrous Na₂SO₄ and evaporated to dryness. The residue obtained was purified by flash column chromatography and eluted with 5% MeOH in CH₂Cl₂ to get 5'-O-*tert*-butyldiphenylsilyl-2'-O-[N,N-dimethylaminooxyethyl]-5-methyluridine as a white foam (14.6 g, 80%).

(g) 2'-O-(DIMETHYLAMINOXYETHYL)-5-METHYLURIDINE
Triethylamine trihydrofluoride (3.91 mL, 24.0 mmol) was dissolved in dry THF and triethylamine (1.67 mL, 12 mmol, dry, kept over KOH). This mixture of triethylamine-2HF was then added to 5'-O-*tert*-butyldiphenylsilyl-2'-O-[N,N-dimethylaminooxyethyl]-5-methyluridine (1.40 g, 2.4 mmol) and stirred at room temperature for 24 hrs. Reaction was monitored by TLC (5% MeOH in CH₂Cl₂). Solvent was removed under vacuum and the residue placed on a flash column and eluted with 10% MeOH in CH₂Cl₂ to get 2'-O-(dimethylaminooxyethyl)-5-methyluridine (766 mg, 92.5%).

(h) 5'-O-DMT-2'-O-(DIMETHYLAMINOXYETHYL)-5-METHYLURIDINE
2'-O-(dimethylaminooxyethyl)-5-methyluridine (750 mg, 2.17 mmol) was dried over P₂O₅ under high vacuum overnight at 40 C. It was then co-evaporated with anhydrous pyridine (20 mL). The residue obtained

was dissolved in pyridine (11 mL) under argon atmosphere. 4-dimethylaminopyridine (26.5 mg, 2.60 mmol), 4,4'-dimethoxytrityl chloride (880 mg, 2.60 mmol) was added to the mixture and the reaction mixture was stirred at room temperature until all of the starting material disappeared. Pyridine was removed under vacuum and the residue chromatographed and eluted with 10% MeOH in CH₂Cl₂ (containing a few drops of pyridine) to get 5'-O-DMT-2'-O-(dimethylamino-oxyethyl)-5-methyluridine (1.13 g, 80%). (i) 5'-O-DMT-2'-O-(2-N,N-DIMETHYLAMINOXYETHYL)-5-METHYLURIDINE-3'-[(2-CYANOETHYL)-N,N-DIISO-PROPYLPHOSPHORAMIDITE] 5'-O-DMT-2'-O-(dimethylaminooxyethyl)-5-methyluridine (1.08 g, 1.67 mmol) was co-evaporated with toluene (20 mL). To the residue N,N-diisopropylamine tetrazonide (0.29 g, 1.67 mmol) was added and dried over P₂O₅ under high vacuum overnight at 40 C. Then the reaction mixture was dissolved in anhydrous acetonitrile (8.4 mL) and 2-cyanoethyl-N,N,Nⁱ,Nⁱ-tetraisopropylphosphoramidite (2.12 mL, 6.08 mmol) was added. The reaction mixture was stirred at ambient temperature for 4 hrs under inert atmosphere. The progress of the reaction was monitored by TLC (hexane:ethyl acetate 1:1). The solvent was evaporated, then the residue was dissolved in ethyl acetate (70 mL) and washed with 5% aqueous NaHCO₃ (40 mL). Ethyl acetate layer was dried over anhydrous Na₂SO₄ and concentrated. Residue obtained was chromatographed (ethyl acetate as eluent) to get 5'-O-DMT-2'-O-(2-N,N-dimethylaminooxyethyl)-5-methyluridine-3'-[(2-cyanoethyl)-N,N-diisopropylphosphoramidite] as a foam (1.04 g, 74.9%).
 2'-(AMINOXYETHOXY) NUCLEOSIDE AMIDITES

2'-(Aminooxyethoxy) nucleoside amidites [also known
in the art as 2'-O-(aminooxyethyl) nucleoside amidites]
are prepared as described in the following
paragraphs [COMMENT] [1] Page: 1

5 G= cpds 26-31 from ISIS 2824. Adenosine, cytidine and
thymidine nucleoside amidites are prepared similarly.

N2-ISOBUTYRYL-6-O-DIPHENYLCARBAMOYL-2'-O-(2-
ETHYLACETYL)-5'-O-(4,4'-DIMETHOXYTRITYL)GUANOSINE-
3'-[(2-CYANOETHYL)-N,N-DIISOPROPYLPHOSPHORAMIDITE]

10 The 2'-O-aminooxyethyl guanosine analog may be
obtained by selective 2'-O-alkylation of diaminopurine
riboside. Multigram quantities of diaminopurine riboside
may be purchased from Schering AG (Berlin) to provide 2'-
O-(2-ethylacetyl) diaminopurine riboside along with a
15 minor amount of the 3'-O-isomer. 2'-O-(2-ethylacetyl)
diaminopurine riboside may be resolved and converted to
2'-O-(2-ethylacetyl)guanosine by treatment with adenosine
deaminase. (McGee, D. P. C., Cook, P. D., Guinosso, C.
J., WO 94/02501 A1 940203.) Standard protection
20 procedures should afford 2'-O-(2-ethylacetyl)-5'-O-(4,4'-
dimethoxytrityl)guanosine and 2-N-isobutyryl-6-O-
diphenylcarbamoyl-2'-O-(2-ethylacetyl)-5'-O-(4,4'-
dimethoxytrityl)guanosine which may be reduced to provide
2-N-isobutyryl-6-O-diphenylcarbamoyl-2'-O-(2-
25 ethylacetyl)-5'-O-(4,4'-dimethoxytrityl)guanosine. As
before the hydroxyl group may be displaced by N-
hydroxyphthalimide via a Mitsunobu reaction, and the
protected nucleoside may phosphitylated as usual to yield
2-N-isobutyryl-6-O-diphenylcarbamoyl-2'-O-(2-
30 ethylacetyl)-5'-O-(4,4'-dimethoxytrityl)guanosine-3'-[(2-
cyanoethyl)-N,N-diisopropylphosphoramidite].

2'-DIMETHYLAMINOETHOXYETHOXY (2'-DMAEOE)
NUCLEOSIDE AMIDITES

2'-dimethylaminoethoxyethoxy nucleoside amidites
(also known in the art as 2'-O-dimethylaminoethoxyethyl,
i.e., 2'-O-CH₂-O-CH₂-N(CH₂)₂, or 2'-DMAEOE nucleoside
amidites) are prepared as follows. Other nucleoside
5 amidites are prepared similarly.

2'-O-[2(2-N,N-DIMETHYLAMINOETHOXY)ETHYL]-5-METHYL
URIDINE

2[2-(Dimethylamino)ethoxy]ethanol (Aldrich, 6.66 g,
50 mmol) is slowly added to a solution of borane in
10 tetrahydrofuran (1 M, 10 mL, 10 mmol) with stirring in a
100 mL bomb. Hydrogen gas evolves as the solid
dissolves. O²-,2'-anhydro-5-methyluridine (1.2 g, 5 mmol),
and sodium bicarbonate (2.5 mg) are added and the bomb is
sealed, placed in an oil bath and heated to 155 C for 26
15 hours. The bomb is cooled to room temperature and
opened. The crude solution is concentrated and the
residue partitioned between water (200 mL) and hexanes
(200 mL). The excess phenol is extracted into the hexane
layer. The aqueous layer is extracted with ethyl acetate
20 (3x200 mL) and the combined organic layers are washed
once with water, dried over anhydrous sodium sulfate and
concentrated. The residue is columned on silica gel
using methanol/methylene chloride 1:20 (which has 2%
triethylamine) as the eluent. As the column fractions
25 are concentrated a colorless solid forms which is
collected to give the title compound as a white solid.

5'-O-DIMETHOXYTRITYL-2'-O-[2(2-N,N-DIMETHYL-
AMINOETHOXY)ETHYL]-5-METHYL URIDINE

To 0.5 g (1.3 mmol) of 2'-O-[2(2-N,N-dimethylamino-
30 ethoxy)ethyl]-5-methyl uridine in anhydrous pyridine (8
mL), triethylamine (0.36 mL) and dimethoxytrityl chloride
(DMT-Cl, 0.87 g, 2 eq.) are added and stirred for 1 hour.
The reaction mixture is poured into water (200 mL) and

extracted with CH₂Cl₂ (2x200 mL). The combined CH₂Cl₂ layers are washed with saturated NaHCO₃ solution, followed by saturated NaCl solution and dried over anhydrous sodium sulfate. Evaporation of the solvent followed by silica gel chromatography using MeOH:CH₂Cl₂:Et₃N (20:1, v/v, with 1% triethylamine) gives the title compound.

5'-O-DIMETHOXYTRITYL-2'-O-[2(2-N,N-DIMETHYLAMINOETHOXY)ETHYL]-5-METHYL URIDINE-3'-O-(CYANOETHYL-N,N-DIISOPROPYL) PHOSPHORAMIDITE

Diisopropylaminotetrazolide (0.6 g) and 2-cyanoethoxy-N,N-diisopropyl phosphoramidite (1.1 mL, 2 eq.) are added to a solution of 5'-O-dimethoxytrityl-2'-O-[2(2-N,N-dimethylaminoethoxy)ethyl]-5-methyluridine (2.17 g, 3 mmol) dissolved in CH₂Cl₂ (20 mL) under an atmosphere of argon. The reaction mixture is stirred overnight and the solvent evaporated. The resulting residue is purified by silica gel flash column chromatography with ethyl acetate as the eluent to give the title compound.

EXAMPLE 2: OLIGONUCLEOTIDE SYNTHESIS

Unsubstituted and substituted phosphodiester (P=O) oligonucleotides are synthesized on an automated DNA synthesizer (Applied Biosystems model 380B) using standard phosphoramidite chemistry with oxidation by iodine.

Phosphorothioates (P=S) are synthesized as for the phosphodiester oligonucleotides except the standard oxidation bottle was replaced by 0.2 M solution of 3H-1,2-benzodithiole-3-one 1,1-dioxide in acetonitrile for the stepwise thiation of the phosphite linkages. The thiation wait step was increased to 68 sec and was followed by the capping step. After cleavage from the

CPG column and deblocking in concentrated ammonium hydroxide at 55 C (18 h), the oligonucleotides were purified by precipitating twice with 2.5 volumes of ethanol from a 0.5 M NaCl solution. Phosphinate
5 oligonucleotides are prepared as described in U.S. Patent 5,508,270, herein incorporated by reference.

Alkyl phosphonate oligonucleotides are prepared as described in U.S. Patent 4,469,863, herein incorporated by reference. 3'-Deoxy-3'-methylene phosphonate
10 oligonucleotides are prepared as described in U.S. Patents 5,610,289 or 5,625,050, herein incorporated by reference. Phosphoramidite oligonucleotides are prepared as described in U.S. Patent, 5,256,775 or U.S. Patent 5,366,878, herein incorporated by reference. Alkylphosphonothioate oligonucleotides are prepared as described
15 in published PCT applications PCT/US94/00902 and PCT/US93/06976 (published as WO 94/17093 and WO 94/02499, respectively), herein incorporated by reference. 3'-Deoxy-3'-amino phosphoramidate oligonucleotides are
20 prepared as described in U.S. Patent 5,476,925, herein incorporated by reference. Phosphotriester oligonucleotides are prepared as described in U.S. Patent 5,023,243, herein incorporated by reference. Borano phosphate oligonucleotides are prepared as described in U.S.
25 Patents 5,130,302 and 5,177,198, both herein incorporated by reference.

EXAMPLE 3: OLIGONUCLEOSIDE SYNTHESIS

Methylenemethylimino linked oligonucleosides, also
30 identified as MMI linked oligonucleosides, methylenedimethylhydrazo linked oligonucleosides, also identified as MDH linked oligonucleosides, and methylenecarbonylamino linked oligonucleosides, also identified as amide-3

linked oligonucleosides, and methyleneaminocarbonyl
linked oligonucleosides, also identified as amide-4
linked oligonucleosides, as well as mixed backbone
compounds having, for instance, alternating MMI and P=O
5 or P=S linkages are prepared as described in U.S. Patents
5,378,825, 5,386,023, 5,489,677, 5,602,240 and
5,610,289, all of which are herein incorporated by
reference.

Formacetal and thioformacetal linked oligonucleo-
10 sides are prepared as described in U.S. Patents 5,264,562
and 5,264,564, herein incorporated by reference. Ethylene
oxide linked oligonucleosides are prepared as described
in U.S. Patent 5,223,618, herein incorporated by
reference.

15
EXAMPLE 4: PNA SYNTHESIS

Peptide nucleic acids (PNAs) are prepared in
accordance with any of the various procedures referred to
in Peptide Nucleic Acids (PNA): Synthesis, Properties and
20 Potential Applications, *Bioorganic & Medicinal Chemistry*,
1996, 4, 5-23. They may also be prepared in accordance
with U.S. Patents 5,539,082, 5,700,922, and 5,719,262,
herein incorporated by reference.

25
EXAMPLE 5: SYNTHESIS OF CHIMERIC OLIGONUCLEOTIDES

Chimeric oligonucleotides, oligonucleosides or
mixed oligonucleotides/oligonucleosides of the invention
can be of several different types. These include a first
type wherein the "gap" segment of linked nucleosides is
30 positioned between 5' and 3' "wing" segments of linked
nucleosides and a second "open end" type wherein the
"gap" segment is located at either the 3' or the 5'
terminus of the oligomeric compound. Oligonucleotides of

the first type are also known in the art as "gapmers" or gapped oligonucleotides. Oligonucleotides of the second type are also known in the art as "hemimers" or "wingmers".

5 [2'-O-ME]--[2'-DEOXY]--[2'-O-ME] CHIMERIC
 PHOSPHOROTHIOATE OLIGONUCLEOTIDES
 Chimeric oligonucleotides having 2'-O-alkyl
phosphorothioate and 2'-deoxy phosphorothioate oligo-
nucleotide segments are synthesized using an Applied
10 Biosystems automated DNA synthesizer Model 380B, as
above. Oligonucleotides are synthesized using the
automated synthesizer and 2'-deoxy-5'-dimethoxytrityl-3'-
O-phosphoramidite for the DNA portion and 5'-dimethoxy-
trityl-2'-O-methyl-3'-O-phosphoramidite for 5' and 3'
15 wings. The standard synthesis cycle is modified by
increasing the wait step after the delivery of tetrazole
and base to 600 s repeated four times for RNA and twice
for 2'-O-methyl. The fully protected oligonucleotide is
cleaved from the support and the phosphate group is
20 deprotected in 3:1 ammonia/ethanol at room temperature
overnight then lyophilized to dryness. Treatment in
methanolic ammonia for 24 hrs at room temperature is then
done to deprotect all bases and sample was again
lyophilized to dryness. The pellet is resuspended in 1M
25 TBAF in THF for 24 hrs at room temperature to deprotect
the 2' positions. The reaction is then quenched with 1M
TEAA and the sample is then reduced to 1/2 volume by
rotovac before being desalted on a G25 size exclusion
column. The oligo recovered is then analyzed
30 spectrophotometrically for yield and for purity by
capillary electrophoresis and by mass spectrometry.

[2'-O-(2-METHOXYETHYL)]--[2'-DEOXY]--[2'-O-(METHOXYETHYL)] CHIMERIC PHOSPHOROTHIOATE OLIGONUCLEOTIDES

[2'-O-(2-methoxyethyl)]--[2'-deoxy]--[2'-O-(methoxyethyl)] chimeric phosphorothioate oligonucleotides were prepared as per the procedure above for the 2'-O-methyl chimeric oligonucleotide, with the substitution of 2'-O-(methoxyethyl) amidites for the 2'-O-methyl amidites.

[2'-O-(2-METHOXYETHYL)PHOSPHODIESTER]--[2'-DEOXY PHOSPHOROTHIOATE]--[2'-O-(2-METHOXYETHYL) PHOSPHODIESTER] CHIMERIC OLIGONUCLEOTIDES

[2'-O-(2-methoxyethyl phosphodiester)]--[2'-deoxy phosphorothioate]--[2'-O-(methoxyethyl) phosphodiester] chimeric oligonucleotides are prepared as per the above procedure for the 2'-O-methyl chimeric oligonucleotide with the substitution of 2'-O-(methoxyethyl) amidites for the 2'-O-methyl amidites, oxidization with iodine to generate the phosphodiester internucleotide linkages within the wing portions of the chimeric structures and sulfurization utilizing 3,4,5-benzodithiole-3-one 1,1-dioxide (Beaucage Reagent) to generate the phosphorothioate internucleotide linkages for the center gap.

Other chimeric oligonucleotides, chimeric oligonucleosides and mixed chimeric oligonucleotides/oligonucleosides are synthesized according to United States patent 5,623,065, herein incorporated by reference.

EXAMPLE 6: OLIGONUCLEOTIDE ISOLATION

After cleavage from the controlled pore glass column (Applied Biosystems) and deblocking in concentrated ammonium hydroxide at 55 C for 18 hours, the

oligonucleotides or oligonucleosides are purified by precipitation twice out of 0.5 M NaCl with 2.5 volumes ethanol. Synthesized oligonucleotides were analyzed by polyacrylamide gel electrophoresis on denaturing gels and
5 judged to be at least 85% full length material. The relative amounts of phosphorothioate and phosphodiester linkages obtained in synthesis were periodically checked by ³¹P nuclear magnetic resonance spectroscopy, and for some studies oligonucleotides were purified by HPLC, as
10 described by Chiang et al., *J. Biol. Chem.* **1991**, *266*, 18162-18171. Results obtained with HPLC-purified material were similar to those obtained with non-HPLC purified material.

15 **EXAMPLE 7: OLIGONUCLEOTIDE SYNTHESIS - 96 WELL PLATE FORMAT**

Oligonucleotides were synthesized via solid phase P(III) phosphoramidite chemistry on an automated synthesizer capable of assembling 96 sequences
20 simultaneously in a standard 96 well format. Phosphodiester internucleotide linkages were afforded by oxidation with aqueous iodine. Phosphorothioate internucleotide linkages were generated by sulfurization utilizing 3,4-dihydro-2H-benzothiole-3-one 1,1-dioxide
25 (Beaucage Reagent) in anhydrous acetonitrile. Standard base-protected beta-cyanoethyl-diisopropyl phosphoramidites were purchased from commercial vendors (e.g. PE-Applied Biosystems, Foster City, CA, or Pharmacia, Piscataway, NJ). Non-standard nucleosides are
30 synthesized as per known literature or patented methods. They are utilized as base protected beta-cyanoethyl-diisopropyl phosphoramidites.

Oligonucleotides were cleaved from support and deprotected with concentrated NH_4OH at elevated temperature (55-60 C) for 12-16 hours and the released product then dried in vacuo. The dried product was then re-suspended in sterile water to afford a master plate from which all analytical and test plate samples are then diluted utilizing robotic pipettors.

EXAMPLE 8: OLIGONUCLEOTIDE ANALYSIS - 96 WELL PLATE
FORMAT

The concentration of oligonucleotide in each well was assessed by dilution of samples and UV absorption spectroscopy. The full-length integrity of the individual products was evaluated by capillary electrophoresis (CE) in either the 96 well format (Beckman P/ACEJ MDQ) or, for individually prepared samples, on a commercial CE apparatus (e.g., Beckman P/ACEJ 5000, ABI 270). Base and backbone composition was confirmed by mass analysis of the compounds utilizing electrospray-mass spectroscopy. All assay test plates were diluted from the master plate using single and multi-channel robotic pipettors. Plates were judged to be acceptable if at least 85% of the compounds on the plate were at least 85% full length.

EXAMPLE 9: CELL CULTURE AND OLIGONUCLEOTIDE TREATMENT

The effect of antisense compounds on target nucleic acid expression can be tested in any of a variety of cell types provided that the target nucleic acid is present at measurable levels. This can be routinely determined using, for example, PCR or Northern blot analysis. The following 5 cell types are provided for illustrative purposes, but other cell types can be routinely used,

provided that the target is expressed in the cell type chosen. This can be readily determined by methods routine in the art, for example Northern blot analysis, Ribonuclease protection assays, or RT-PCR.

5 T-24 CELLS: The human transitional cell bladder carcinoma cell line T-24 was obtained from the American Type Culture Collection (ATCC) (Manassas, VA). T-24 cells were routinely cultured in complete McCoy's 5A basal media (Gibco/Life Technologies, Gaithersburg, MD) supplemented with 10% fetal calf serum (Gibco/Life Technologies, Gaithersburg, MD), penicillin 100 units per mL, and streptomycin 100 micrograms per mL (Gibco/Life Technologies, Gaithersburg, MD). Cells were routinely passaged by trypsinization and dilution when they reached 10
10 90% confluence. Cells were seeded into 96-well plates (Falcon-Primaria #3872) at a density of 7000 cells/well for use in RT-PCR analysis.

 For Northern blotting or other analysis, cells may be seeded onto 100 mm or other standard tissue culture plates and treated similarly, using appropriate volumes of medium and oligonucleotide.

20 A549 CELLS: The human lung carcinoma cell line A549 was obtained from the American Type Culture Collection (ATCC) (Manassas, VA). A549 cells were routinely cultured in DMEM basal media (Gibco/Life Technologies, Gaithersburg, MD) supplemented with 10% fetal calf serum (Gibco/Life Technologies, Gaithersburg, MD), penicillin 100 units per mL, and streptomycin 100 micrograms per mL (Gibco/Life Technologies, Gaithersburg, MD). Cells were routinely passaged by trypsinization and dilution when they reached 90% confluence.

30 NHDF CELLS: Human neonatal dermal fibroblast (NHDF) were obtained from the Clonetics Corporation

(Walkersville MD). NHDFs were routinely maintained in Fibroblast Growth Medium (Clonetics Corporation, Walkersville, MD) supplemented as recommended by the supplier. Cells were maintained for up to 10 passages as recommended by the supplier.

HEK CELLS: Human embryonic keratinocytes (HEK) were obtained from the Clonetics Corporation (Walkersville, MD). HEKs were routinely maintained in Keratinocyte Growth Medium (Clonetics Corporation, Walkersville MD) formulated as recommended by the supplier. Cells were routinely maintained for up to 10 passages as recommended by the supplier.

PC-12 CELLS: The rat neuronal cell line PC-12 was obtained from the American Type Culture Collection (Manassas, VA). PC-12 cells were routinely cultured in DMEM, high glucose (Gibco/Life Technologies, Gaithersburg, MD) supplemented with 10% horse serum + 5% fetal calf serum (Gibco/Life Technologies, Gaithersburg, MD). Cells were routinely passaged by trypsinization and dilution when they reached 90% confluence. Cells were seeded into 96-well plates (Falcon-Primaria #3872) at a density of 20000 cells/well for use in RT-PCR analysis.

For Northern blotting or other analysis, cells may be seeded onto 100 mm or other standard tissue culture plates and treated similarly, using appropriate volumes of medium and oligonucleotide.

TREATMENT WITH ANTISENSE COMPOUNDS:

When cells reached 80% confluency, they were treated with oligonucleotide. For cells grown in 96-well plates, wells were washed once with 200 μ L OPTI-MEMJ-1 reduced-serum medium (Gibco BRL) and then treated with 130 μ L of OPTI-MEMJ-1 containing 3.75 μ g/mL LIPOFECTINJ (Gibco BRL) and the desired concentration of

oligonucleotide. After 4-7 hours of treatment, the medium was replaced with fresh medium. Cells were harvested 16-24 hours after oligonucleotide treatment.

The concentration of oligonucleotide used varies from cell line to cell line. To determine the optimal oligonucleotide concentration for a particular cell line, the cells are treated with a positive control oligonucleotide at a range of concentrations. For human cells the positive control oligonucleotide is ISIS 13920, **TCCGTCATCGCTCCTCAGGG**, SEQ ID NO: 1, a 2'-O-methoxyethyl gapmer (2'-O-methoxyethyls shown in bold) with a phosphorothioate backbone which is targeted to human H-ras. For mouse or rat cells the positive control oligonucleotide is ISIS 15770, **ATGCATTCTGCCCCAAGGA**, SEQ ID NO: 2, a 2'-O-methoxyethyl gapmer (2'-O-methoxyethyls shown in bold) with a phosphorothioate backbone which is targeted to both mouse and rat c-raf. The concentration of positive control oligonucleotide that results in 80% inhibition of c-Ha-ras (for ISIS 13920) or c-raf (for ISIS 15770) mRNA is then utilized as the screening concentration for new oligonucleotides in subsequent experiments for that cell line. If 80% inhibition is not achieved, the lowest concentration of positive control oligonucleotide that results in 60% inhibition of H-ras or c-raf mRNA is then utilized as the oligonucleotide screening concentration in subsequent experiments for that cell line. If 60% inhibition is not achieved, that particular cell line is deemed as unsuitable for oligonucleotide transfection experiments.

EXAMPLE 10: ANALYSIS OF OLIGONUCLEOTIDE INHIBITION OF PTP1B EXPRESSION

Antisense modulation of PTP1B expression can be assayed in a variety of ways known in the art. For example, PTP1B mRNA levels can be quantitated by, e.g., Northern blot analysis, competitive polymerase chain reaction (PCR), or real-time PCR (RT-PCR). Real-time quantitative PCR is presently preferred. RNA analysis can be performed on total cellular RNA or poly(A)+ mRNA. Methods of RNA isolation are taught in, for example, Ausubel, F.M. et al., *Current Protocols in Molecular Biology*, Volume 1, pp. 4.1.1-4.2.9 and 4.5.1-4.5.3, John Wiley & Sons, Inc., 1993. Northern blot analysis is routine in the art and is taught in, for example, Ausubel, F.M. et al., *Current Protocols in Molecular Biology*, Volume 1, pp. 4.2.1-4.2.9, John Wiley & Sons, Inc., 1996. Real-time quantitative (PCR) can be conveniently accomplished using the commercially available ABI PRISMJ 7700 Sequence Detection System, available from PE-Applied Biosystems, Foster City, CA and used according to manufacturer's instructions. Prior to quantitative PCR analysis, primer-probe sets specific to the target gene being measured are evaluated for their ability to be "multiplexed" with a GAPDH amplification reaction. In multiplexing, both the target gene and the internal standard gene GAPDH are amplified concurrently in a single sample. In this analysis, mRNA isolated from untreated cells is serially diluted. Each dilution is amplified in the presence of primer-probe sets specific for GAPDH only, target gene only ("single-plexing"), or both (multiplexing). Following PCR amplification, standard curves of GAPDH and target mRNA signal as a function of dilution are generated from both the single-plexed and multiplexed samples. If both the slope and correlation coefficient of the GAPDH and target signals

generated from the multiplexed samples fall within 10% of their corresponding values generated from the single-plexed samples, the primer-probe set specific for that target is deemed as multiplexable. Other methods of PCR are also known in the art.

Protein levels of PTP1B can be quantitated in a variety of ways well known in the art, such as immunoprecipitation, Western blot analysis (immunoblotting), ELISA or fluorescence-activated cell sorting (FACS). Antibodies directed to PTP1B can be identified and obtained from a variety of sources, such as the MSRS catalog of antibodies (Aerie Corporation, Birmingham, MI), or can be prepared via conventional antibody generation methods. Methods for preparation of polyclonal antisera are taught in, for example, Ausubel, F.M. et al., *Current Protocols in Molecular Biology*, Volume 2, pp. 11.12.1-11.12.9, John Wiley & Sons, Inc., 1997. Preparation of monoclonal antibodies is taught in, for example, Ausubel, F.M. et al., *Current Protocols in Molecular Biology*, Volume 2, pp. 11.4.1-11.11.5, John Wiley & Sons, Inc., 1997.

Immunoprecipitation methods are standard in the art and can be found at, for example, Ausubel, F.M. et al., *Current Protocols in Molecular Biology*, Volume 2, pp. 10.16.1-10.16.11, John Wiley & Sons, Inc., 1998. Western blot (immunoblot) analysis is standard in the art and can be found at, for example, Ausubel, F.M. et al., *Current Protocols in Molecular Biology*, Volume 2, pp. 10.8.1-10.8.21, John Wiley & Sons, Inc., 1997. Enzyme-linked immunosorbent assays (ELISA) are standard in the art and can be found at, for example, Ausubel, F.M. et al.,

Current Protocols in Molecular Biology, Volume 2, pp.
11.2.1-11.2.22, John Wiley & Sons, Inc., 1991.

EXAMPLE 11: POLY(A)+ MRNA ISOLATION

5 Poly(A)+ mRNA was isolated according to Miura et
al., *Clin. Chem.*, 1996, 42, 1758-1764. Other methods
for poly(A)+ mRNA isolation are taught in, for example,
Ausubel, F.M. et al., *Current Protocols in Molecular*
10 *Biology*, Volume 1, pp. 4.5.1-4.5.3, John Wiley & Sons,
Inc., 1993. Briefly, for cells grown on 96-well plates,
growth medium was removed from the cells and each well
was washed with 200 μ L cold PBS. 60 μ L lysis buffer (10
mM Tris-HCl, pH 7.6, 1 mM EDTA, 0.5 M NaCl, 0.5% NP-40,
20 mM vanadyl-ribonucleoside complex) was added to each
15 well, the plate was gently agitated and then incubated at
room temperature for five minutes. 55 μ L of lysate was
transferred to Oligo d(T) coated 96-well plates (AGCT
Inc., Irvine CA). Plates were incubated for 60 minutes
at room temperature, washed 3 times with 200 μ L of wash
20 buffer (10 mM Tris-HCl pH 7.6, 1 mM EDTA, 0.3 M NaCl).
After the final wash, the plate was blotted on paper
towels to remove excess wash buffer and then air-dried
for 5 minutes. 60 μ L of elution buffer (5 mM Tris-HCl pH
7.6), preheated to 70 C was added to each well, the plate
25 was incubated on a 90 C hot plate for 5 minutes, and the
eluate was then transferred to a fresh 96-well plate.

Cells grown on 100 mm or other standard plates may
be treated similarly, using appropriate volumes of all
solutions.

30

EXAMPLE 12: TOTAL RNA ISOLATION

Total mRNA was isolated using an RNEASY 96J kit and
buffers purchased from Qiagen Inc. (Valencia, CA)

following the manufacturer's recommended procedures.
Briefly, for cells grown on 96-well plates, growth medium
was removed from the cells and each well was washed with
200 μ L cold PBS. 100 μ L Buffer RLT was added to each well
5 and the plate vigorously agitated for 20 seconds. 100 μ L
of 70% ethanol was then added to each well and the
contents mixed by pipetting three times up and down. The
samples were then transferred to the RNEASY 96J well
plate attached to a QIAVACJ manifold fitted with a waste
10 collection tray and attached to a vacuum source. Vacuum
was applied for 15 seconds. 1 mL of Buffer RW1 was added
to each well of the RNEASY 96J plate and the vacuum again
applied for 15 seconds. 1 mL of Buffer RPE was then
15 added to each well of the RNEASY 96J plate and the vacuum
applied for a period of 15 seconds. The Buffer RPE wash
was then repeated and the vacuum was applied for an
additional 10 minutes. The plate was then removed from
the QIAVACJ manifold and blotted dry on paper towels.
The plate was then re-attached to the QIAVACJ manifold
20 fitted with a collection tube rack containing 1.2 mL
collection tubes. RNA was then eluted by pipetting 60 μ L
water into each well, incubating 1 minute, and then
applying the vacuum for 30 seconds. The elution step was
repeated with an additional 60 μ L water.
25 The repetitive pipetting and elution steps may be
automated using a QIAGEN Bio-Robot 9604 (Qiagen, Inc.,
Valencia, CA). Essentially, after lysing of the cells on
the culture plate, the plate is transferred to the robot
deck where the pipetting, DNase treatment and elution
30 steps are carried out.

EXAMPLE 13: REAL-TIME QUANTITATIVE PCR ANALYSIS OF PTP1B
MRNA LEVELS

Quantitation of PTP1B mRNA levels was determined by real-time quantitative PCR using the ABI PRISMJ 7700 Sequence Detection System (PE-Applied Biosystems, Foster City, CA) according to manufacturer's instructions. This is a closed-tube, non-gel-based, fluorescence detection system which allows high-throughput quantitation of polymerase chain reaction (PCR) products in real-time. As opposed to standard PCR, in which amplification products are quantitated after the PCR is completed, products in real-time quantitative PCR are quantitated as they accumulate. This is accomplished by including in the PCR reaction an oligonucleotide probe that anneals specifically between the forward and reverse PCR primers, and contains two fluorescent dyes. A reporter dye (e.g., JOE, FAM, or VIC, obtained from either Operon Technologies Inc., Alameda, CA or PE-Applied Biosystems, Foster City, CA) is attached to the 5' end of the probe and a quencher dye (e.g., TAMRA, obtained from either Operon Technologies Inc., Alameda, CA or PE-Applied Biosystems, Foster City, CA) is attached to the 3' end of the probe. When the probe and dyes are intact, reporter dye emission is quenched by the proximity of the 3' quencher dye. During amplification, annealing of the probe to the target sequence creates a substrate that can be cleaved by the 5'-exonuclease activity of Taq polymerase. During the extension phase of the PCR amplification cycle, cleavage of the probe by Taq polymerase releases the reporter dye from the remainder of the probe (and hence from the quencher moiety) and a sequence-specific fluorescent signal is generated. With each cycle, additional reporter dye molecules are cleaved from their respective probes, and the fluorescence intensity is monitored at regular intervals by laser

optics built into the ABI PRISMJ 7700 Sequence Detection System. In each assay, a series of parallel reactions containing serial dilutions of mRNA from untreated control samples generates a standard curve that is used to quantitate the percent inhibition after antisense oligonucleotide treatment of test samples.

PCR reagents were obtained from PE-Applied Biosystems, Foster City, CA. RT-PCR reactions were carried out by adding 25 L PCR cocktail (1x TAQMANJ buffer A, 5.5 mM MgCl₂, 300 M each of dATP, dCTP and dGTP, 600 M of dUTP, 100 nM each of forward primer, reverse primer, and probe, 20 Units RNase inhibitor, 1.25 Units AMPLITAQ GOLDJ, and 12.5 Units MuLV reverse transcriptase) to 96 well plates containing 25 L poly(A) mRNA solution. The RT reaction was carried out by incubation for 30 minutes at 48 C. Following a 10 minute incubation at 95 C to activate the AMPLITAQ GOLDJ, 40 cycles of a two-step PCR protocol were carried out: 95 C for 15 seconds (denaturation) followed by 60 C for 1.5 minutes (annealing/extension).

Probes and primers to human PTP1B were designed to hybridize to a human PTP1B sequence, using published sequence information (GenBank accession number M31724, incorporated herein as SEQ ID NO:3). For human PTP1B the PCR primers were:

forward primer: GGAGTTCGAGCAGATCGACAA (SEQ ID NO: 4)
reverse primer: GGCCACTCTACATGGGAAGTC (SEQ ID NO: 5) and
the PCR probe was: FAM-AGCTGGGCGGCCATTTACCAGGAT-TAMRA (SEQ ID NO: 6) where FAM (PE-Applied Biosystems, Foster City, CA) is the fluorescent reporter dye) and TAMRA (PE-Applied Biosystems, Foster City, CA) is the quencher dye. For human GAPDH the PCR primers were:

forward primer: GAAGGTGAAGGTCGGAGTC (SEQ ID NO: 7)

reverse primer: GAAGATGGTGATGGGATTTTC (SEQ ID NO: 8) and
the PCR probe was: 5' JOE-CAAGCTTCCCGTTCTCAGCC- TAMRA 3'
(SEQ ID NO: 9) where JOE (PE-Applied Biosystems, Foster
City, CA) is the fluorescent reporter dye) and TAMRA (PE-
5 Applied Biosystems, Foster City, CA) is the quencher dye.

Probes and primers to rat PTP1B were designed to
hybridize to a rat PTP1B sequence, using published
sequence information (GenBank accession number M33962,
incorporated herein as SEQ ID NO:10). For rat PTP1B the
10 PCR primers were:

forward primer: CGAGGGTGCAAAGTTCATCAT (SEQ ID NO:11)
reverse primer: CCAGGTCTTCATGGGAAAGCT (SEQ ID NO: 12) and
the PCR probe was: FAM-CGACTCGTCAGTGCAGGATCAGTGGA-TAMRA
(SEQ ID NO: 13) where FAM (PE-Applied Biosystems, Foster
15 City, CA) is the fluorescent reporter dye) and TAMRA (PE-
Applied Biosystems, Foster City, CA) is the quencher dye.
For rat GAPDH the PCR primers were:

forward primer: TGTTCTAGAGACAGCCGCATCTT (SEQ ID NO: 14)
reverse primer: CACCGACCTTCACCATCTTGT (SEQ ID NO: 15) and
20 the PCR probe was: 5' JOE-TTGTGCAGTGCCAGCCTCGTCTCA- TAMRA
3' (SEQ ID NO: 16) where JOE (PE-Applied Biosystems,
Foster City, CA) is the fluorescent reporter dye) and
TAMRA (PE-Applied Biosystems, Foster City, CA) is the
quencher dye.

25
EXAMPLE 14: NORTHERN BLOT ANALYSIS OF PTP1B MRNA LEVELS

Eighteen hours after antisense treatment, cell
monolayers were washed twice with cold PBS and lysed in 1
mL RNAzolW (TEL-TEST "B" Inc., Friendswood, TX). Total
30 RNA was prepared following manufacturer's recommended
protocols. Twenty micrograms of total RNA was
fractionated by electrophoresis through 1.2% agarose gels
containing 1.1% formaldehyde using a MOPS buffer system

(AMRESCO, Inc. Solon, OH). RNA was transferred from the gel to HYBONDJ-N+ nylon membranes (Amersham Pharmacia Biotech, Piscataway, NJ) by overnight capillary transfer using a Northern/Southern Transfer buffer system (TEL-TEST "B" Inc., Friendswood, TX). RNA transfer was confirmed by UV visualization. Membranes were fixed by UV cross-linking using a STRATALINKERJ UV Crosslinker 2400 (Stratagene, Inc, La Jolla, CA) and then probed using QUICKHYBJ hybridization solution (Stratagene, La Jolla, CA) using manufacturer's recommendations for stringent conditions.

To detect human PTP1B, a human PTP1B specific probe was prepared by PCR using the forward primer GGAGTTCGAGCAGATCGACAA (SEQ ID NO: 4) and the reverse primer GGCCACTCTACATGGGAAGTC (SEQ ID NO: 5). To normalize for variations in loading and transfer efficiency membranes were stripped and probed for human glyceraldehyde-3-phosphate dehydrogenase (GAPDH) RNA (Clontech, Palo Alto, CA).

To detect rat PTP1B, a rat PTP1B specific probe was prepared by PCR using the forward primer CGAGGGTGCAAAGTTCATCAT (SEQ ID NO:11) and the reverse primer CCAGGTCTTCATGGGAAAGCT (SEQ ID NO: 12). To normalize for variations in loading and transfer efficiency membranes were stripped and probed for rat glyceraldehyde-3-phosphate dehydrogenase (GAPDH) RNA (Clontech, Palo Alto, CA).

Hybridized membranes were visualized and quantitated using a PHOSPHORIMAGERJ and IMAGEQUANTJ Software V3.3 (Molecular Dynamics, Sunnyvale, CA). Data was normalized to GAPDH levels in untreated controls.

EXAMPLE 15: ANTISENSE INHIBITION OF HUMAN PTP1B
EXPRESSION BY CHIMERIC PHOSPHOROTHIOATE OLIGONUCLEOTIDES
HAVING 2'-MOE WINGS AND A DEOXY GAP

In accordance with the present invention, a series
5 of oligonucleotides were designed to target different
regions of the human PTP1B RNA, using published sequences
(GenBank accession number M31724, incorporated herein as
SEQ ID NO: 3). The oligonucleotides are shown in Table
1. ATarget siteA indicates the first (5'-most)
10 nucleotide number on the particular target sequence to
which the oligonucleotide binds. All compounds in Table
1 are chimeric oligonucleotides ("gapmers") 20
nucleotides in length, composed of a central "gap" region
consisting of ten 2'-deoxynucleotides, which is flanked
15 on both sides (5' and 3' directions) by five-nucleotide
"wings". The wings are composed of 2'-methoxyethyl (2'-
MOE)nucleotides. The internucleoside (backbone) linkages
are phosphorothioate (P=S) throughout the
oligonucleotide. All cytidine residues are 5-
20 methylcytidines. The compounds were analyzed for their
effect on human PTP1B mRNA levels by
quantitative real-time PCR as described in other examples
herein. Data are averages from two experiments. If
present, "N.D." indicates "no data".

25 TABLE 1: INHIBITION OF HUMAN PTP1B MRNA LEVELS BY
CHIMERIC PHOSPHOROTHIOATE OLIGONUCLEOTIDES HAVING 2'-MOE
WINGS AND A DEOXY GAP

ISIS #	REGION	TARGET SEQ ID NO	TARGET SITE	SEQUENCE	% INHI BIT.	SEQ ID NO
107769	5' UTR	3	1	cttagcccccaggcccgccc	0	17
107770	5' UTR	3	41	ctcggcccaactgcgcgtct	58	18
107771	Start Codon	3	74	catgacgggcccaggcggt	60	19
107772	Coding	3	113	cccggacttgtcgatctgct	95	20
107773	Coding	3	154	ctggcttcatgtcggatatc	88	21

107774	Coding	3	178	ttggccactctacatgggaa	77	22
107775	Coding	3	223	ggactgacgtctctgtacct	75	23
107776	Coding	3	252	gatgtagtttaatecgaacta	82	24
107777	Coding	3	280	ctagcgttgatatagtcatt	29	25
107778	Coding	3	324	gggtaagaatgtaactcctt	86	26
107779	Coding	3	352	tgaccgcatgtgttaggcaa	75	27
107780	Coding	3	381	ttttctgctcccacaccatc	30	28
107781	Coding	3	408	ctctgttgagcatgacgaca	78	29
107782	Coding	3	436	gcgcattttaacgaaccttt	83	30
107783	Coding	3	490	aaattttgtgtcttcaaagat	0	31
107784	Coding	3	519	tgatatcttcagagatcaat	57	32
107785	Coding	3	547	tctagctgtcgactgtata	74	33
107786	Coding	3	575	agtttcttgggttgtaaggt	33	34
107787	Coding	3	604	gtggtatagtggaaatgtaa	51	35
107788	Coding	3	632	tgattcagggactccaaagt	55	36
107789	Coding	3	661	ttgaaaagaaagttcaagaa	17	37
107790	Coding	3	688	gggctgagtgaccctgaetc	61	38
107791	Coding	3	716	gcagtgcaccacaacggggc	81	39
107792	Coding	3	744	aggttccagacctgccgatg	81	40
107793	Coding	3	772	agcaggaggcaggatcagc	2	41
107794	Coding	3	799	gaagaagggtcttctctctt	53	42
107795	Coding	3	826	tctaacagcactttcttgat	18	43
107796	Coding	3	853	atcaaccccatccgaaactt	0	44
107797	Coding	3	880	gagaagcgcagctggctcgc	82	45
107798	Coding	3	908	tttggcaccttcgatcacag	62	46
107799	Coding	3	952	agctccttcactgatcctg	70	47
107800	Coding	3	1024	tccaggattcgtttgggtgg	72	48
107801	Coding	3	1052	gaactccctgcatttcccat	68	49
107802	Coding	3	1079	ttccttcacccactgggtgat	40	50
107803	Coding	3	1148	gtagggtgcccatttaagg	0	51
107804	Coding	3	1176	cagtgtcttgactcatgctt	75	52
107805	Coding	3	1222	gcctggggcacctcgaagact	67	53
107806	Coding	3	1268	ctcgtccttctcgggcagtg	37	54
107807	Coding	3	1295	gggcttccagtaactcagtg	73	55
107808	Coding	3	1323	ccgtagccacgcacatgttg	80	56
107809	Coding	3	1351	tagcagaggtaagcgcgcgc	72	57
107810	Stop Codon	3	1379	ctatgtgttgctgttgaaca	85	58
107811	3' UTR	3	1404	ggaggtggagtggaggagg	51	59
107812	3' UTR	3	1433	ggctctgcgggcagaggcgg	81	60
107813	3' UTR	3	1460	ccgcggcatgcctgctagtc	84	61
107814	3' UTR	3	1489	tctctacgcggtccggcggc	84	62
107815	3' UTR	3	1533	aagatgggttttagtgcaga	65	63
107816	3' UTR	3	1634	gtactctctttcactctcct	69	64
107817	3' UTR	3	1662	ggccccttccctctgcgcgc	59	65
107818	3' UTR	3	1707	ctccaggaggagccctggg	57	66
107819	3' UTR	3	1735	gggctgttggcgtgcgcgc	54	67
107820	3' UTR	3	1783	tttaataaatatggagtgg	0	68
107821	3' UTR	3	1831	gttcaagaaaatgctagtgc	69	69

107822	3' UTR	3	1884	ttgataaaagccottgatgca	74	70
107823	3' UTR	3	1936	atggcaaaagccttccattcc	26	71
107824	3' UTR	3	1973	gtctctccttcccagttactgg	60	72
107825	3' UTR	3	2011	ttaccaccaaatatcactaaa	39	73
107826	3' UTR	3	2045	attatatattatagcattgt	24	74
107827	3' UTR	3	2080	tcacatcatgtttcttatta	48	75
107828	3' UTR	3	2115	ataacaggaggagagaataag	0	76
107829	3' UTR	3	2170	ttacatgcattctaatacac	21	77
107830	3' UTR	3	2223	gatcaaaagtcttctcatttca	81	78
107831	3' UTR	3	2274	ggatcatgcacaggcaggttg	82	79
107832	3' UTR	3	2309	caacaggccttaggaaccaca	65	80
107833	3' UTR	3	2344	aactgcaccctattgctgag	61	81
107834	3' UTR	3	2380	gtcatgccagggaattagcaa	0	82
107835	3' UTR	3	2413	acaggctgggcctcaccagg	58	83
107836	3' UTR	3	2443	tgagttacagcaagaccctg	44	84
107837	3' UTR	3	2473	gaatatggcttcccatacc	0	85
107838	3' UTR	3	2502	ccctaaatcatgtccagagc	87	86
107839	3' UTR	3	2558	gacttggaatggcggaggct	74	87
107840	3' UTR	3	2587	caaatacagggtctgctcaag	31	88
107841	3' UTR	3	2618	gaagtgtgggtttccagcagg	56	89
107842	3' UTR	3	2648	cctaaaggaccgtcaccacag	42	90
107843	3' UTR	3	2678	gtgaaccgggacagagacgg	25	91
107844	3' UTR	3	2724	gccccacagggtttgaggggt	53	92
107845	3' UTR	3	2755	cctttgcagggaagagtcgtg	75	93
107846	3' UTR	3	2785	aaagccacttaatgtggagg	79	94
107847	3' UTR	3	2844	gtgaaaatgctggcaagaga	86	95
107848	3' UTR	3	2970	tcagaatgcttacagcctgg	61	96

As shown in Table 1, SEQ ID NOS 18, 19, 20, 21, 22, 23, 24, 26, 27, 29, 30, 32, 33, 35, 36, 38, 39, 40, 42, 45, 46, 47, 48, 49, 50, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 69, 70, 72, 73, 75, 78, 79, 80, 81, 83, 84, 86, 87, 89, 90, 92, 93, 94, 95, and 96 demonstrated at least 35% inhibition of human PTP1B expression in this assay and are therefore preferred.

EXAMPLE 16: ANTISENSE INHIBITION OF RAT PTP1B EXPRESSION BY CHIMERIC PHOSPHOROTHIOATE OLIGONUCLEOTIDES HAVING 2'-MOE WINGS AND A DEOXY GAP.

In accordance with the present invention, a second series of oligonucleotides were designed to target different regions of the rat PTP1B RNA, using published

sequences (GenBank accession number M33962, incorporated herein as SEQ ID NO: 10). The oligonucleotides are shown in Table 2. ATarget siteA indicates the first (5'-most) nucleotide number on the particular target sequence to which the oligonucleotide binds. All compounds in Table 2 are chimeric oligonucleotides ("gapmers") 20 nucleotides in length, composed of a central "gap" region consisting of ten 2'-deoxynucleotides, which is flanked on both sides (5' and 3' directions) by five-nucleotide "wings". The wings are composed of 2'-methoxyethyl (2'-MOE)nucleotides. The internucleoside (backbone) linkages are phosphorothioate (P=S) throughout the oligonucleotide. All cytidine residues are 5-methylcytidines. The compounds were analyzed for their effect on rat PTP1B mRNA levels by quantitative real-time PCR as described in other examples herein. Data are averages from two experiments. If present, "N.D." indicates "no data".

TABLE 2: INHIBITION OF RAT PTP1B MRNA LEVELS BY CHIMERIC PHOSPHOROTHIOATE OLIGONUCLEOTIDES HAVING 2'-MOE WINGS AND A DEOXY GAP

ISIS #	REGION	TARGET SEQ ID NO	TARGET SITE	SEQUENCE	%INHIB	SEQ ID NO
111549	5' UTR	10	1	caacctccccagcagcgggt	32	97
111550	5' UTR	10	33	tcgaggcccgctcgcccgcca	27	98
111551	5' UTR	10	73	cctcgcccgctcgcccgct	34	99
111552	Coding	10	132	tcgatctgctcgaattcctt	49	100
113669	Coding	10	164	cctggtaaatagcgcgccag	36	101
113670	Coding	10	174	tgtcgaatatcctggtaaat	63	102
113671	Coding	10	184	actggcttcattgtcgaatat	58	103
113672	Coding	10	189	aagtcactggcttcattgtc	40	104
111553	Coding	10	190	gaagtcactggcttcattgtc	27	105
113673	Coding	10	191	ggaagtcactggcttcattgt	54	106
113674	Coding	10	192	gggaagtcactggcttcattg	41	107
113675	Coding	10	193	tgggaagtcactggcttcatt	56	108
113676	Coding	10	194	atgggaagtcactggcttca	31	109

113677	Coding	10	195	catgggaagtcactggcttc	59	110
113678	Coding	10	225	tttttgttcttaggaagttt	24	111
111554	Coding	10	228	cgggtttttgttcttaggaag	45	112
111555	Coding	10	269	tccgactgtggtcaaaaggg	39	113
113679	Coding	10	273	ttaatccgactgtggtcaaa	45	114
113680	Coding	10	298	atagtcattatcttctctgat	49	115
111556	Coding	10	303	ttgatatagtcattatcttc	29	116
113681	Coding	10	330	gcttctctccatttttatcaa	67	117
111557	Coding	10	359	ggccctgggtgaggatatag	20	118
113682	Coding	10	399	cacaccatctcccagaagt	29	119
111558	Coding	10	405	tgctcccacaccatctccca	48	120
113683	Coding	10	406	ctgctcccacaccatctccc	51	121
113684	Coding	10	407	tctgctcccacaccatctcc	37	122
113685	Coding	10	408	ttctgctcccacaccatctc	54	123
113686	Coding	10	417	ccctgctcttctgctccca	60	124
111559	Coding	10	438	atgcggttgagcatgaccac	15	125
113687	Coding	10	459	tttaacgagcctttctccat	33	126
113688	Coding	10	492	ttttctctcttctgtggcca	54	127
113689	Coding	10	502	gaccatctctttttctctct	58	128
111560	Coding	10	540	tcagagatcagtgctcagctt	21	129
113690	Coding	10	550	cttgacatcttcagagatca	64	130
113691	Coding	10	558	taatatgacttgacatcttc	46	131
111561	Coding	10	579	aactccaactgccgtactgt	14	132
111562	Coding	10	611	tctctcgagcctcctgggta	38	133
113692	Coding	10	648	ccaaagtccaggccaggtggt	63	134
111563	Coding	10	654	gggactccaaagtccaggcca	31	135
113693	Coding	10	655	agggactccaaagtccaggcc	50	136
113694	Coding	10	656	cagggactccaaagtccaggc	45	137
113695	Coding	10	657	tcagggactccaaagtccagg	49	138
113696	Coding	10	663	ggtgactcagggactccaaa	34	139
111564	Coding	10	705	cctgactctcggactttgaa	53	140
113697	Coding	10	715	gctgagtgagcctgactctc	57	141
113698	Coding	10	726	ccgtgctctgggctgagtga	48	142
111565	Coding	10	774	aagggtccctgacctgccaat	28	143
111566	Coding	10	819	tctttctctctgtccatcag	34	144
113699	Coding	10	820	gtctttctctctgtccatca	41	145
113700	Coding	10	821	ggctctttctctctgtccatc	66	146
113701	Coding	10	822	gggtctttctctctgtccat	71	147
113702	Coding	10	852	aacagcactttcttgatgtc	39	148
111567	Coding	10	869	ggaacctgcgcattctccaac	0	149
111568	Coding	10	897	tggtcggccgtctggatgag	29	150
113703	Coding	10	909	gagaagcgcagttggtcggc	48	151
113704	Coding	10	915	aggtaggagaagcgcagttg	31	152
113705	Coding	10	918	gccaggtaggagaagcgcag	41	153
111569	Coding	10	919	agccaggtaggagaagcgc	56	154
113706	Coding	10	920	cagccaggtaggagaagcgc	58	155
113707	Coding	10	921	acagccaggtaggagaagcg	43	156
113708	Coding	10	922	cacagccaggtaggagaagc	49	157
113709	Coding	10	923	tcacagccaggtaggagaag	47	158
111570	Coding	10	924	atcacagccaggtaggagaa	51	159

113710	Coding	10	925	gatcacagccaggtaggaga	51	160
113711	Coding	10	926	cgatcacagccaggtaggag	63	161
113712	Coding	10	927	tcgatcacagccaggtagga	71	162
113713	Coding	10	932	caccctcgatcacagccagg	75	163
113714	Coding	10	978	tccttccactgatcctgcac	97	164
111571	Coding	10	979	ctccttccactgatcctgca	89	165
113715	Coding	10	980	gctccttccactgatcctgc	99	166
107799	Coding	10	981	agtccttccactgatcctg	99	167
113716	Coding	10	982	aagctccttccactgatcct	97	168
113717	Coding	10	983	aaagctccttccactgatcc	95	169
113718	Coding	10	984	gaaagctccttccactgatc	95	170
113719	Coding	10	985	ggaaagctccttccactgat	95	171
111572	Coding	10	986	gggaaagctccttccactga	89	172
113720	Coding	10	987	tgggaaagctccttccactg	97	173
113721	Coding	10	1036	tggccggggagggtgggggca	20	174
111573	Coding	10	1040	tgggtggccggggagggtggg	20	175
113722	Coding	10	1046	tgcggttgggtggccggggga	18	176
111574	Coding	10	1073	tgcaactggcattgtgagggc	38	177
113723	Coding	10	1206	acttcagtgtcttgactcat	67	178
113724	Coding	10	1207	aacttcagtgtcttgactca	60	179
111575	Coding	10	1208	taacttcagtgtcttgactc	50	180
113725	Coding	10	1209	ctaacttcagtgtcttgact	53	181
111576	Coding	10	1255	gacagatgacctgagcacttt	32	182
106409	Coding	10	1333	gaccaggaagggttccagt	32	183
113726	Coding	10	1334	tgaccaggaagggttccag	39	184
111577	Coding	10	1335	ttgaccaggaagggttcca	32	185
113727	Coding	10	1336	gttgaccaggaagggttcc	41	186
113728	Coding	10	1342	gcacacgttgaccaggaagg	59	187
111578	Coding	10	1375	gaggtagcgccagtcgcca	45	188
111579	Coding	10	1387	taccggtaacagaggtacg	32	189
111580	Coding	10	1397	agtgaacacatacccggtaa	30	190
111581	3' UTR	10	1456	caaatcctaacctgggcagt	31	191
111582	3' UTR	10	1519	ttccagttccaccacaggct	24	192
111583	3' UTR	10	1552	ccagtgcacagatgcccttc	47	193
111584	3' UTR	10	1609	acaggttaaggccctgagat	29	194
111585	3' UTR	10	1783	gcctagcatcttttgttttc	43	195
111586	3' UTR	10	1890	aagccagcaggaactttaca	36	196
111587	3' UTR	10	2002	gggacacctgagggaaagcag	16	197
111588	3' UTR	10	2048	ggtcactctgcaagatggcgg	40	198
111589	3' UTR	10	2118	gccaacctctgatgacctg	25	199
111590	3' UTR	10	2143	tggaagccccagctctaagc	25	200
111591	3' UTR	10	2165	tagtaatgactttccaatca	44	201
111592	3' UTR	10	2208	tgagtcttgcctttacacctc	41	202
111593	3' UTR	10	2252	cctgcgcgcggagtgaacttc	22	203
111594	3' UTR	10	2299	aggacgtcactgcagcagga	43	204
111595	3' UTR	10	2346	tcaggacaagtcttggcagt	32	205
111596	3' UTR	10	2405	gaggctgcacagtaagcgct	34	206
111597	3' UTR	10	2422	tcagccaaccagcatcagag	20	207
111598	3' UTR	10	2449	acccacagtgtccacctccc	30	208
111599	3' UTR	10	2502	agtgcgggctgtgctgctgg	30	209

111600	3' UTR	10	2553	cagctcgcctctggcggcctc	8	210
111601	3' UTR	10	2608	aggaaggagctgcacgtcc	32	211
111602	3' UTR	10	2664	ccctcacgattgctcgtggg	24	212
111603	3' UTR	10	2756	cagtggagcggctcctctgg	18	213
111604	3' UTR	10	2830	caggctgacaccttacacgg	30	214
111605	3' UTR	10	2883	gtcctacctcaacctagga	37	215
111606	3' UTR	10	2917	ctgccccagcaccagccaca	12	216
111607	3' UTR	10	2946	attgcttctaagacctcag	33	217
111608	3' UTR	10	2978	ttacatgtcaccactgttgt	28	218
111609	3' UTR	10	3007	tacacatgtcatcagtagcc	37	219
111610	3' UTR	10	3080	ttttctaaactcacaggga	30	220
111611	3' UTR	10	3153	gtgcccgcagtgagcaggc	23	221
111612	3' UTR	10	3206	cggcctcggcactggacagc	27	222
111613	3' UTR	10	3277	gtggaatgtctgagatccag	31	223
111614	3' UTR	10	3322	agggcgggcctgcttgcaca	23	224
111615	3' UTR	10	3384	cggctcctggcctgctccaga	31	225
111616	3' UTR	10	3428	tacactgttcccaggagggt	42	226
111617	3' UTR	10	3471	tggtgccagcagcgtagca	10	227
111618	3' UTR	10	3516	cagtctcttcagcctcaaga	43	228
113729	3' UTR	10	3537	aagagtcattgagcaccatca	56	229
111619	3' UTR	10	3560	tgaaggtcaagtccccctca	40	230
111620	3' UTR	10	3622	ctggcaagagggcagactgga	30	231
111621	3' UTR	10	3666	ggctctgtgctggcttctct	52	232
111622	3' UTR	10	3711	gccatctcctcagcctgtgc	39	233
111623	3' UTR	10	3787	agcgctctgctctgaggcccc	16	234
111624	3' UTR	10	3854	tgctgagtaagtattgactt	35	235
111625	3' UTR	10	3927	ctatggccatttagagagag	36	236
113730	3' UTR	10	3936	tggtttattctatggccatt	59	237
111626	3' UTR	10	3994	cgctcctgcaaagggtgctat	11	238
111627	3' UTR	10	4053	gttggaacgggtgcagtcgg	39	239
111628	3' UTR	10	4095	atattattgttgcaactaatg	33	240

As shown in Table 2, SEQ ID NOs 97, 99, 100, 101, 102, 103, 104, 106, 107, 108, 109, 110, 112, 113, 114, 115, 117, 120, 121, 122, 123, 124, 126, 127, 128, 130, 131, 133, 134, 135, 136, 137, 138, 139, 140, 141, 142, 144, 145, 146, 147, 148, 151, 152, 153, 154, 155, 156, 157, 158, 159, 160, 161, 162, 163, 164, 165, 166, 167, 168, 169, 170, 171, 172, 173, 177, 178, 179, 180, 181, 182, 183, 184, 185, 186, 187, 188, 189, 191, 193, 195, 196, 198, 201, 202, 204, 205, 206, 211, 215, 217, 219, 223, 225, 226, 228, 229, 230, 232, 233, 235, 236, 237, 239 and 240 demonstrated at least 30% inhibition of rat

PTP1B expression in this experiment and are therefore preferred.

EXAMPLE 17: WESTERN BLOT ANALYSIS OF PTP1B PROTEIN LEVELS

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Western blot analysis (immunoblot analysis) is carried out using standard methods. Cells are harvested 16-20 h after oligonucleotide treatment, washed once with PBS, suspended in Laemmli buffer (100 ul/well),
10 boiled for 5 minutes and loaded on a 16% SDS-PAGE gel. Gels are run for 1.5 hours at 150 V, and transferred to membrane for western blotting. Appropriate primary antibody directed to PTP1B is used, with a radiolabelled or fluorescently labeled secondary antibody directed
15 against the primary antibody species. Bands are visualized using a PHOSPHORIMAGERJ (Molecular Dynamics, Sunnyvale CA).

EXAMPLE 18: EFFECTS OF ANTISENSE INHIBITION OF PTP1B (ISIS 113715) ON BLOOD GLUCOSE LEVELS

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db/db mice are used as a model of Type 2 diabetes. These mice are hyperglycemic, obese, hyperlipidemic, and insulin resistant. The db/db phenotype is due to a mutation in the leptin receptor on a C57BLKS background.
25 However, a mutation in the leptin gene on a different mouse background can produce obesity without diabetes (ob/ob mice). Leptin is a hormone produced by fat that regulates appetite and animals or humans with leptin deficiencies become obese. Heterozygous db/wt mice
30 (known as lean littermates) do not display the hyperglycemia/hyperlipidemia or obesity phenotype and are used as controls.

In accordance with the present invention, ISIS 113715 (GCTCCTTCCACTGATCCTGC, SEQ ID No: 166) was investigated in experiments designed to address the role of PTP1B in glucose metabolism and homeostasis. ISIS 113715 is completely complementary to sequences in the coding region of the human, rat, and mouse PTP1B nucleotide sequences incorporated herein as SEQ ID No: 3 (starting at nucleotide 951 of human PTP1B; Genbank Accession No. M31724), SEQ ID No: 10 (starting at nucleotide 980 of rat PTP1B; Genbank Accession No. M33962) and SEQ ID No: 241 (starting at nucleotide 1570 of mouse PTP1B; Genbank Accession No. U24700). The control used is ISIS 29848 (NNNNNNNNNNNNNNNNNNNN, SEQ ID No: 242) where N is a mixture of A, G, T and C.

Male db/db mice and lean (heterozygous, i.e., db/wt) littermates (age 9 weeks at time 0) were divided into matched groups (n=6) with the same average blood glucose levels and treated by intraperitoneal injection once a week with saline, ISIS 29848 (the control oligonucleotide) or ISIS 113715. db/db mice were treated at a dose of 10, 25 or 50 mg/kg of ISIS 113715 or 50 mg/kg of ISIS 29848 while lean littermates were treated at a dose of 50 or 100 mg/kg of ISIS 113715 or 100 mg/kg of ISIS 29848. Treatment was continued for 4 weeks with blood glucose levels being measured on day 0, 7, 14, 21 and 28.

By day 28 in db/db mice, blood glucose levels were reduced at all doses from a starting level of 300 mg/dL to 225 mg/dL for the 10 mg/kg dose, 175 mg/dL for the 25 mg/kg dose and 125 mg/dL for the 50 mg/kg dose. These final levels are within normal range for wild-type mice (170 mg/dL). The mismatch control and saline treated

levels levels were 320 mg/dL and 370 mg/dL at day 28, respectively.

In lean littermates, blood glucose levels remained constant throughout the study for all treatment groups (average 120 mg/dL). These results indicate that treatment with ISIS 113715 reduces blood glucose in db/db mice and that there is no hypoglycemia induced in the db/db or the lean littermate mice as a result of the oligonucleotide treatment.

In a similar experiment, ob/ob mice and their lean littermates (heterozygous, i.e., ob/wt) were dosed twice a week at 50 mg/kg with ISIS 113715, ISIS 29848 or saline control and blood glucose levels were measured at the end of day 7, 14 and 21. Treatment of ob/ob mice with ISIS 113715 resulted in the largest decrease in blood glucose over time going from 225 mg/dL at day 7 to 95 mg/dL at day 21. Ob/ob mice displayed an increase in plasma glucose over time from 300 mg/dL to 325 mg/dL while treatment with the control oligonucleotide reduced plasma glucose from an average of 280 mg/dL to 130 mg/dL. In the lean littermates plasma glucose levels remained unchanged in all treatment groups (average level 100 mg/dL).

EXAMPLE 19: EFFECTS OF ANTISENSE INHIBITION OF PTP1B (ISIS 113715) ON MRNA EXPRESSION IN LIVER

Male db/db mice and lean littermates (age 9 weeks at time 0) were divided into matched groups (n=6) with the same average blood glucose levels and treated by intraperitoneal injection once a week with saline, ISIS 29848 (the control oligonucleotide) or ISIS 113715. db/db mice were treated at a dose of 10, 25 or 50 mg/kg of ISIS 113715 or 50 mg/kg of ISIS 29848 while lean

littermates were treated at a dose of 50 or 100 mg/kg of
ISIS 113715 or 100 mg/kg of ISIS 29848. Treatment was
continued for 4 weeks after which the mice were
sacrificed and tissues collected for mRNA analysis. RNA
5 values were normalized and are expressed as a percentage
of saline treated control.

ISIS 113715 successfully reduced PTP1B mRNA levels
in the livers of db/db mice at all doses examined (60%
reduction of PTP1B mRNA), whereas the control
10 oligonucleotide treated animals showed no reduction in
PTP1B mRNA, remaining at the level of the saline treated
control. Treatment of lean littermates with ISIS 113715
also reduced mRNA levels to 45% of control at the 50
mg/kg dose and 25% of control at the 100 mg/kg dose. The
15 control oligonucleotide (ISIS 29848) failed to show any
reduction in mRNA levels.

EXAMPLE 20: EFFECTS OF ANTISENSE INHIBITION OF PTP1B
(ISIS 113715) ON BODY WEIGHT

20 Male db/db mice and lean littermates (age 9 weeks at
time 0) were divided into matched groups (n=6) with the
same average blood glucose levels and treated by
intraperitoneal injection once a week with saline, ISIS
29848 (the control oligonucleotide) or ISIS 113715.
25 db/db mice were treated at a dose of 10, 25 or 50 mg/kg
of ISIS 113715 or 50 mg/kg of ISIS 29848 while lean
littermates were treated at a dose of 50 or 100 mg/kg of
ISIS 113715 or 100 mg/kg of ISIS 29848. Treatment was
continued for 4 weeks. At day 28 mice were sacrificed
30 and final body weights were measured.

Treatment of ob/ob mice with ISIS 113715 resulted in
an increase in body weight which was constant over the
dose range with animals gaining an average of 11.0 grams

while saline treated controls gained 5.5 grams. Animals treated with the control oligonucleotide gained an average of 7.8 grams of body weight.

5 Lean littermate animals treated with 50 or 100 mg/kg of ISIS 113715 gained 3.8 grams of body weight compared to a gain of 3.0 grams for the saline controls.

10 In a similar experiment, ob/ob mice and their lean littermates were dosed twice a week at 50 mg/kg with ISIS 113715, ISIS 29848 or saline control and body weights were measured at the end of day 7, 14 and 21.

15 Treatment of the ob/ob mice with ISIS 113715, ISIS 29848 or saline control all resulted in a similar increase in body weight across the 21-day timecourse. At the end of day 7 all ob/ob treatment groups had an average weight of 42 grams. By day 21, animals treated with ISIS 113715 had an average body weight of 48 grams, while those in the ISIS 29848 (control oligonucleotide) and saline control group each had an average body weight of 52 grams. All of the lean littermates had an average
20 body weight of 25 grams at the beginning of the timecourse and all lean littermate treatment groups showed an increase in body weight, to 28 grams, by day 21.

25 EXAMPLE 21: EFFECTS OF ANTISENSE INHIBITION OF PTP1B (ISIS 113715) ON PLASMA INSULIN LEVELS

Male db/db mice (age 9 weeks at time 0) were divided into matched groups (n=6) with the same average blood glucose levels and treated by intraperitoneal injection
30 twice a week with saline, ISIS 29848 (the control oligonucleotide) or ISIS 113715 at a dose of 50 mg/kg. Treatment was continued for 3 weeks with plasma insulin levels being measured on day 7, 14, and 21.

Mice treated with ISIS 113715 showed a decrease in plasma insulin levels from 15 ng/mL at day 7 to 7.5 ng/mL on day 21. Saline treated animals has plasma insulin levels of 37 ng/mL at day 7 which dropped to 25 ng/mL on day 14 but rose again to 33 ng/mL by day 21. Mice treated with the control oligonucleotide also showed a decrease in plasma insulin levels across the timecourse of the study from 25 ng/mL at day 7 to 10 ng/mL on day 21. However, ISIS 113715 was the most effective at reducing plasma insulin over time.

EXAMPLE 22: ANTISENSE INHIBITION OF HUMAN PTP1B
EXPRESSION BY ADDITIONAL CHIMERIC PHOSPHOROTHIOATE
OLIGONUCLEOTIDES HAVING 2'-MOE WINGS AND A DEOXY GAP

In accordance with the present invention, an additional series of oligonucleotides were designed to target different genomic regions of the human PTP1B RNA, using published sequences (GenBank accession number M31724, incorporated herein as SEQ ID NO: 3), and concatenated genomic sequence derived from nucleotide residues 1-31000 of Genbank accession number AL034429 followed by nucleotide residues 1-45000 of Genbank accession number AL133230, incorporated herein as SEQ ID NO: 243). The oligonucleotides are shown in Table 3. "Target site" indicates the first (5'-most) nucleotide number on the particular target sequence to which the oligonucleotide binds. All compounds in Table 3 are chimeric oligonucleotides ("gapmers") 20 nucleotides in length, composed of a central "gap" region consisting of ten 2'-deoxynucleotides, which is flanked on both sides (5' and 3' directions) by five-nucleotide "wings". The wings are composed of 2'-methoxyethyl (2'- MOE) nucleotides. The internucleoside (backbone) linkages are

phosphorothioate (P=S) throughout the oligonucleotide.
All cytidine residues are 5-methylcytidines. The
compounds were analyzed for their effect on human PTP1B
mRNA levels by quantitative real-time PCR as described in
other examples herein. Data are averages from two
experiments. If present, "N.D." indicates "no data".

TABLE 3: INHIBITION OF HUMAN PTP1B MRNA LEVELS BY
CHIMERIC PHOSPHOROTHIOATE OLIGONUCLEOTIDES HAVING 2'-MOE
WINGS AND A DEOXY GAP

Isis #	REGION	TARGET	TARGET	SEQUENCE	%	SEQ ID
142020	5' UTR	3	6	GCGCTCTTAGCCCCGAGGCC	61	244
142021	5' UTR	3	65	CCAGGGCGGCTGCTGCGCCT	56	245
142022	Start Codon	3	80	CATCTCCATGACGGGCCAGG	4	246
142023	Start Codon	3	85	TTTCCATCTCCATGACGGG	67	247
142024	Start Codon	3	90	ACTCCTTTTCCATCTCCATG	71	248
142025	Exon 1	3	106	TTGTCGATCTGCTCGAACTC	61	249
142026	Exon 1	3	109	GACTTGTGATCTGCTCGAA	66	250
142027	Exon 1	3	116	GCTCCCGGACTTGTGATCT	95	251
142028	Exon 1	3	119	CCAGCTCCCGGACTTGTGGA	92	252
142029	Exon:Exo n Junction	3	945	TCCACTGATCCTGCACGGAA	44	253
142030	Exon:Exo n Junction	3	948	CCTTCCACTGATCCTGCACG	55	254
142031	3' UTR	3	1453	ATGCCTGCTAGTCGGGCGTG	67	255
142032	3' UTR	3	1670	CGGGTGTAGGCCCTTCCCT	74	256
142033	3' UTR	3	1772	ATGGAGTGGAGAGTTGCTCC	63	257
142034	3' UTR	3	1893	TTGTAATTTTGATAAAGCC	61	258
142035	3' UTR	3	1962	CAGTACTGGTCTGACGCAGC	68	259
142036	3' UTR	3	2018	TCTCACGTTACCCACAATAT	74	260
142037	3' UTR	3	2070	TTTCTTATTAAATACCCACG	61	261
142038	3' UTR	3	2088	AAGTAATCTCACATCATGTT	79	262
142039	3' UTR	3	2314	TTCAGCAACAGGCTTAGGAA	51	263
142040	3' UTR	3	2323	GACAATGACTTCAGCAACAG	43	264
142041	3' UTR	3	2359	TGCCTATTCTGGAAAACCTG	43	265
142042	3' UTR	3	2395	GGAAGTCACTAGAGTGTCTAT	14	266
142043	3' UTR	3	2418	CCAGGACAGGCTGGGCTCA	67	267
142044	3' UTR	3	2426	CTGCTGTACCAGGACAGGCT	73	268
142045	3' UTR	3	2452	TGGAATGTCTGAGTTACAGC	74	269
142046	3' UTR	3	2566	AGAGTGTGACTTGAATGG	43	270

142047	3' UTR	3	2574	GCTCAAGAAGAGTGTGACT	76	271
142048	3' UTR	3	2598	TGCCTCTCTTCCAAATCAG	43	272
142049	3' UTR	3	2800	TGTTTTTCATGTTAAAAAGC	44	273
142050	3' UTR	3	2895	TCCCACCACAGAAATTCTCT	21	274
142051	3' UTR	3	2921	GCTCTGCAGGGTGACACCTC	74	275
142052	3' UTR	3	3066	AGGAGGTTAAACCAGTACGT	78	276
142053	3' UTR	3	3094	GGTGGAGAGCCAGCTGCTCT	59	277
142054	3' UTR	3	3153	TATTGGCTTAAGGCATATAG	72	278
142055	3' UTR	3	3168	GACCTGATGAGTAAATATTG	58	279
142084	5' UTR	243	859	TTCTTCATGTCAACCGGCAG	11	280
142085	5' UTR	243	919	GCCCCGAGGCCCGCTGCAT	83	281
142056	Intron 1	243	4206	TAGTGAACATTGTTACAAC	70	282
142057	Intron 1	243	27032	TGCTAAGCCACTTCTAATCA	72	283
142058	Intron 1	243	27203	CAGGATTCCTAAGTTATTAAA	32	284
142059	Intron 1	243	33720	TGGGCAGGATGGCTCTGGTA	21	285
142060	Intron 1	243	48065	TACAATACTATCTGTGACTA	34	286
142061	Exon: Intron function	243	51931	GATACTTACAGGACTGACG	39	287
142086	Intron 2	243	52005	AACCCTGAGGCGAAAGGAGT	64	288
142062	Intron 2	243	54384	CCCCAGGTCACTAAAATTAA	48	289
142063	Intron 2	243	55362	AAAGCAAAGGTGAGTTGGTG	56	290
142064	Intron 3	243	56093	GCTCAATTATTAAACCACTT	64	291
142065	Intron 3	243	56717	AGTCCTCAAGAACTCACTTT	70	292
142066	Intron 4	243	61780	GAAAGCAGGAGCTGTGGCA	39	293
142067	Intron 4	243	64554	AAAACCTGGAGAGACAGCAG	71	294
142068	Intron 4	243	64869	ACATGGAAGCCATGGTCAGC	24	295
142069	Intron 5	243	67516	ATTGCTAGACTCACACTAGG	68	296
142070	Intron 5	243	68052	GGCTGTGATCAAAAGGCAGC	51	297
142087	Intron 5	243	68481	CACTGGCTCTGGGCACTTT	70	298
142088	Intron 5	243	68563	GCTGGGCAGCCACCATAAAA	71	299
142071	Intron 5	243	68648	AGTCCCCTCACCTCTTTTCT	59	300
142072	Exon: Intron function	243	69107	CCTCCTTACCAGCAAGAGGC	26	301
142089	Intron 6	243	69198	TGTATTTTGGAAAGAGGAGCG	53	302
142090	Intron 6	243	69220	ACAGACTAACACAGTGAGTC	53	303
142073	Intron 6	243	69264	ACAAATTACCGAGTCTCAGG	47	304
142074	Intron 6	243	69472	TCATGAAAGGCTTGGTGCCC	41	305
142075	Intron 7	243	70042	TTGGAAGATGAAATCTTTTG	30	306
142076	Intron 7	243	70052	AGCCATGTACTTGGGAAGATG	69	307
142077	Intron 8	243	70661	CGAGCCCCCTCATTCCAACAA	42	308
142078	Intron 8	243	71005	CACCTCAGCGGACACCTCTA	6	309
142079	Exon: Intron	243	71938	GAAACATACCCTGTAGCAGA	52	310
142091	Intron 9	243	72131	CAGAGGGCTCCTTAAAACCC	61	311
142092	Intron 9	243	72430	ATTCGTAAAAGTTTGGGATT	34	312
142080	Intron 9	243	72453	CCCTCTTCTCCAAGGGAGTT	73	313
142081	Intron 9	243	73158	GGAATGAAACCAACAGTTC	42	314
142082	Exon 10	243	75012	AAATGGTTTATTCCATGGCC	66	315

142083	Exon 10	243	75215	AAAAATTTTATTGTTGCAGC	48	316
142093	3' UTR	243	75095	CCGGTCATGCAGCCACGTAT	85	317
142094	3' UTR	243	75165	GTTGGAAACTGTACAGTCT	77	318
142095	3' UTR	243	75211	ATTTTATTGTTGCAGCTAAA	46	319

As shown in Table 3, SEQ ID NOS: 244, 245, 247, 248, 249, 250, 251, 252, 254, 255, 256, 257, 258, 259, 260, 261, 262, 263, 267, 268, 269, 271, 275, 276, 277, 278, 279, 281, 282, 283, 288, 290, 291, 292, 294, 296, 297, 298, 299, 300, 302, 303, 307, 310, 311, 313, 315, 317, and 318, demonstrated at least 50% inhibition of human PTP1B expression in this assay and are therefore preferred.

EXAMPLE 23: ANTISENSE INHIBITION OF HUMAN PTP1B
EXPRESSION BY ADDITIONAL CHIMERIC PHOSPHOROTHIOATE
OLIGONUCLEOTIDES HAVING 2'-MOE WINGS AND A DEOXY GAP

In accordance with the present invention, an additional series of oligonucleotides were designed to target either the 3'UTR or the 5'UTR of the human PTP1B RNA, using published sequences (GenBank accession number M31724, incorporated herein as SEQ ID NO: 3) and concatenated genomic sequence derived from nucleotide residues 1-31000 of Genbank accession number AL034429 followed by nucleotide residues 1-45000 of Genbank accession number AL133230, incorporated herein as SEQ ID NO: 243. The oligonucleotides are shown in Table4. "Target site" indicates the first (5'-most) nucleotide number on the particular target sequence to which the oligonucleotide binds. All compounds in Table 3 are chimeric oligonucleotides ("gapmers") 20 nucleotides in length, composed of a central "gap" region consisting of ten 2'-deoxynucleotides, which is flanked on both sides (5' and 3' directions) by five-nucleotide "wings". The

wings are composed of 2'-methoxyethyl (2'-MOE) nucleotides. The internucleoside (backbone) linkages are phosphorothioate (P=S) throughout the oligonucleotide. All cytidine residues are 5-methylcytidines. The compounds were analyzed for their effect on human PTP1B mRNA levels by quantitative real-time PCR as described in other examples herein. Data are averages from two experiments. If present, "N.D." indicates "no data".

TABLE 4: INHIBITION OF HUMAN PTP1B MRNA LEVELS BY CHIMERIC PHOSPHOROTHIOATE OLIGONUCLEOTIDES HAVING 2'-MOE WINGS AND A DEOXY GAP

Isis #	REGION	TARGET	TARGET	SEQUENCE	%	SEQ ID
146879	5' UTR	3	50	CGCCTCCTTCTCGGCCCACT	29	320
146880	5' UTR	3	62	GGGCGGCTGCTGCGCCTCCT	34	321
146881	3' UTR	3	1601	GTGGATTGGTACTCAAAGT	72	322
146882	3' UTR	3	1610	AAATGGCTTGTGGATTGGT	72	323
146883	3' UTR	3	1637	ATGGTACTCTCTTCACTCT	61	324
146884	3' UTR	3	1643	GCCAGCATGGTACTCTCTTT	63	325
146885	3' UTR	3	1764	GAGAGTTGCTCCCTGCAGAT	62	326
146886	3' UTR	3	1770	GGAGTGGAGAGTTGCTCCCT	57	327
146887	3' UTR	3	1874	CCTTGATGCAAGGCTGACAT	65	328
146888	3' UTR	3	1879	AAAGCCCTTGATGCAAGGCT	59	329
146889	3' UTR	3	1915	AGTACTACCTGAGGATTTAT	46	330
146890	3' UTR	3	1925	TTCCATTCCCAGTACTACCT	41	331
146891	3' UTR	3	1938	CCATGGCAAAGCCTTCCATT	65	332
146892	3' UTR	3	1943	CAGGCCCATGGCAAAGCCTT	52	333
146893	3' UTR	3	1988	CAACTGCTTACAACCGTCCT	60	334
146894	3' UTR	3	2055	CCACGTGTTTATTATATATT	42	335
146895	3' UTR	3	2063	TTAAATACCCAGTGTTCAT	27	336
146896	3' UTR	3	2099	TAAGCGGACAAAGTATCT	47	337
146897	3' UTR	3	2118	CAGATAACAGGGAGGAGAAT	31	338
146898	3' UTR	3	2133	GAGAACTAGATCTAGCAGAT	0	339
146899	3' UTR	3	2140	AGTGATTGAGAACTAGATCT	62	340
146900	3' UTR	3	2184	GACACAAGAAGACCTTACAT	49	341
146901	3' UTR	3	2212	CTCATTTCAGACATATTT	60	342
146902	3' UTR	3	2263	GGCAGGTTGGACTTGGACAT	49	343
146903	3' UTR	3	2296	AACCACAGCCATGTAATGAT	43	344
146904	3' UTR	3	2332	TTGCTGAGCGACAATGACTT	42	345
146905	3' UTR	3	2350	CTGGAAAACGACCCCTATT	31	346
146906	3' UTR	3	2409	GCTGGGCCTCACCAGGAAGT	77	347
146907	3' UTR	3	2439	TTACAGCAAGACCCTGCTGT	28	348
146908	3' UTR	3	2457	ACCCTTGAATGTCTGAGTT	65	349
146909	3' UTR	3	2464	TTCCCATACCTTGGAATGT	62	350
146910	3' UTR	3	2471	ATATGGCTTCCCATACCTT	47	351
146911	3' UTR	3	2477	GTGTGAATATGGCTTCCCAT	54	352
146912	3' UTR	3	2509	CCTGCTTCCCTAAATCATGT	65	353

146913	3' UTR	3	2514	GTGTCCCTGCTTCCCTAAAT	55	354
146914	3' UTR	3	2546	CGGAGGCTGATCCCAAAGGT	55	355
146915	3' UTR	3	2602	CAGGTGCCCTCTCTTCAAAT	60	356
146916	3' UTR	3	2613	GTGGTTTCCAGCAGGTGCCT	63	357
146917	3' UTR	3	2628	GCTGTTTCAAGAAGTGTGGT	43	358
146918	3' UTR	3	2642	GGACCGTCACCCAGGCTGTT	32	359
146919	3' UTR	3	2655	CAGGCTGCCATAAGGACCGT	60	360
146920	3' UTR	3	2732	ACCATCAGGCCCCACAGGGT	58	361
146921	3' UTR	3	2759	GTTCCTTTGCAAGGAAGAGT	69	362
146922	3' UTR	3	2772	GTGGAGGTCTTCAGTTCCT	64	363
146923	3' UTR	3	2781	CCACTTAATGTGGAGGTCTT	54	364
146924	3' UTR	3	2814	AGCTACAGCTGCCGTGTTTT	51	365
146925	3' UTR	3	2862	CCACGAGAAAGCAAAATGT	50	366
146926	3' UTR	3	2885	GAATTTCTCTGACTGGCTT	23	367
146927	3' UTR	3	2890	CCACAGAAATTTCTCTGTACT	61	368
146928	3' UTR	3	2901	GAATGTTCCCAACACAGAAT	61	369
146929	3' UTR	3	2956	GCCTGGCACCTAAGCCTTAT	0	370
146930	3' UTR	3	2965	ATGCTTACAGCCTGGCACCT	55	371
146931	3' UTR	3	3008	CTACATACATATACAGGACT	65	372
146932	3' UTR	3	3042	TTTGAAATGCTACTATATAT	44	373
146933	3' UTR	3	3070	GGATAGGAGGTTAAACCAGT	67	374
146934	3' UTR	3	3086	GCCAGCTGCTCTCCAAGGAT	42	375
146935	3' UTR	3	3121	CTACCTCTCTAACATAATGT	39	376
146936	3' UTR	3	3126	GCTCGCTACCTCTCTAACAT	68	377
146937	3' UTR	3	3143	AGGCATATAGCAGAGCAGCT	61	378
146938	5' UTR	243	851	GTCAACCGGCAGCCGGAAT	14	379
146942	5' UTR	243	891	CCTGCAGCTACCGCCGCCCT	69	380
146943	5' UTR	243	908	CGCTGCAATCCCGACCCCT	87	381
146944	3' UTR	243	75050	ACCAAAACACCTTGCTTTTT	27	382
146945	3' UTR	243	75057	GTATTATACCAAACACCTT	39	383
146946	3' UTR	243	75072	CACACACCTGAAAAGGTATT	42	384
146947	3' UTR	243	75097	ACCCGGTCATGCAGCCACGT	49	385
146948	3' UTR	243	75136	GTGAGGTCACAGAAGACCCT	49	386
146949	3' UTR	243	75154	GTACAGTCTGACAGTTCTGT	40	387
146950	3' UTR	243	75172	ATGGCAAGTTGGAAAACCTGT	65	388
146951	3' UTR	243	75192	AATGCAAACCCATCATGAAT	43	389

As shown in Table 4, SEQ ID NOs, 322, 323, 324, 325, 326, 327, 328, 329, 330, 331, 332, 333, 334, 335, 337, 340, 341, 342, 343, 344, 345, 347, 349, 350, 351, 352, 353, 354, 355, 356, 357, 358, 360, 361, 362, 363, 364, 365, 366, 368, 369, 371, 372, 373, 374, 375, 377, 378, 380, 381, 384, 385, 386, 387, 388, and 389 demonstrated at least 40% inhibition of human PTP1B expression in this assay and are therefore preferred.

EXAMPLE 24: ANTISENSE INHIBITION OF PTP1B EXPRESSION
(ISIS 113715) IN LIVER, MUSCLE AND ADIPOSE TISSUE OF THE
CYNOMOLGUS MONKEY

5 In a further embodiment, male cynomolgus monkeys
were treated with ISIS 113715 (SEQ ID NO: 166) and levels
of PTP1B mRNA and protein were measured in muscle,
adipose and liver tissue. Serum samples were also
measured for insulin levels.

10 Male cynomolgus monkeys were divided into two
treatment groups, control animals (n=4; saline treatment
only) and treated animals (n=8; treated with ISIS
113715). All animals had two pre-dosing glucose
tolerance tests (GTTs) performed to establish insulin and
glucose baseline values. Animals in the treatment group
15 were dosed subcutaneously on days 1, 8, and 15 with
3mg/kg, 6 mg/kg and 12 mg/kg of ISIS 113715,
respectively. Animals in the control group were
untreated. All animals had GTTs performed on days 5, 13
and 19, four days post-dosing. Ten days after the last
20 dose of 12 mg/kg, all animals in the treatment group
(ISIS 113715) received a one-time dose of 6 mg/kg of ISIS
113715. Three days later, all animals were sacrificed
and tissues were taken for analysis of PTP1B mRNA and
protein levels. Levels of mRNA and protein were
25 normalized to those of the saline treated animals. Of
the tissue examined, PTP1B mRNA levels were reduced to
the greatest extent in the fat and liver, being reduced
by 41% and 40%, respectively. mRNA levels in muscle were
reduced by 10%. Protein levels were reduced by 60% in
30 the liver and by 45% in the muscle but were shown to
increase by 10% in the fat.

Levels of the liver enzymes ALT and AST were measured weekly and showed no change, indicating no ongoing toxic effects of the oligonucleotide treatment.

5 The results of this study demonstrate a significant reduction in liver PTP1B mRNA and protein upon treatment with ISIS 113715. Furthermore, there was no change seen in the fasting insulin levels either between groups or between pre-treatment and post-treatment of the same group. There was, however, a significant lowering of
10 insulin levels with no decrease in fasting glucose levels in all groups suggesting that insulin efficiency (sensitivity) was increased upon treatment with ISIS 113715.

15 Throughout this specification and the claims which follow, unless the context requires otherwise, the word "comprise", or variations such as "comprises" or "comprising", will be understood to imply the inclusion of a stated integer or step or group of integers or steps
20 but not the exclusion of any other integer or step or group of integers or steps.

The reference to any prior art in this specification is not, and should not be taken as an acknowledgment or
25 any form of suggestion that that prior art forms part of the common general knowledge in Australia.

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The claims defining the invention are as follows:

1. A compound up to 30 nucleobases in length comprising one of SEQ ID NOs: 164-173.
2. The compound according to claim 1, wherein the compound is an antisense oligonucleotide.
3. The antisense oligonucleotide of claim 2, wherein the oligonucleotide has a sequence comprising SEQ ID NO: 164, 165, 166, 168, 169, 170, 171, 172 or 173.
4. The compound of claim 1 which is 12 to 30 nucleobases in length.
5. The compound of claim 1 which is 20 nucleobases in length.
6. The antisense oligonucleotide of claim 2, wherein said oligonucleotide includes at least one modified internucleoside linkage, sugar moiety, or nucleobase.
7. The antisense oligonucleotide of claim 2, wherein said oligonucleotide includes at least one 2'-O-methoxyethyl sugar moiety.
8. The antisense oligonucleotide of claim 2, wherein said oligonucleotide includes at least one phosphorothioate internucleoside linkage.
9. The antisense oligonucleotide of claim 2, wherein said oligonucleotide includes at least one 5-methylcytidine.
10. The antisense oligonucleotide of claim 2, wherein the oligonucleotide is a chimeric oligonucleotide.
11. The compound of claim 1 comprising SEQ ID NO: 166.
12. The compound of claim 11, wherein nucleotides 1-5 and 16-20 of SEQ ID NO: 166 are 2'-O-methoxyethyl nucleotides; nucleotides 6-15 of SEQ ID NO: 166 are 2'-deoxynucleotides; all cytidine residues of SEQ ID NO: 166 are 5-methylcytidines and all internucleoside linkages of SEQ ID NO: 166 are phosphorothioate linkages.
13. The compound of claim 12, wherein the compound is a pharmaceutically acceptable salt.
14. The compound of claim 12, wherein the compound is a sodium salt.

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15. A composition comprising a compound according to any one of claims 1 to 12 and a pharmaceutically acceptable carrier or diluent.
16. A composition according to claim 15, further comprising a colloidal dispersion system.
17. A compound according to any one of claims 1-12 for use in medicine.
18. Use of a compound according to any one of claims 1-12 in the preparation of a medicament for treating an animal having or suspected of having a disease or condition associated with PTP1B.
19. Use of a compound according to any one of claims 1-12 in the preparation of a medicament for treating diabetes, obesity, cancer or a hyperproliferative disorder.
20. A method of inhibiting the expression of PTP1B in cells or tissues, comprising contacting said cells or tissues with a compound according to any one of claims 1-12 so that expression of PTP1B is inhibited.
21. The method of claim 20, wherein the cells or tissues are human cells or tissues.
22. The method of claim 21, wherein the cells or tissues are liver, kidney or adipose cells or tissues.
23. A method of treating an animal having or suspected of having a disease or condition associated with PTP1B comprising administering to said animal a therapeutically or prophylactically effective amount of a compound according to any one of claims 1-12 so that expression of PTP1B is inhibited.
24. A method of decreasing blood glucose levels in an animal comprising administering to said animal a compound according to any one of claims 1-12.
25. A method of preventing or delaying the onset of a disease or condition associated with PTP1B in an animal comprising administering to said animal a therapeutically or prophylactically effective amount of a compound according to any one of claims 1-12.
26. A method of preventing or delaying the onset of an increase in blood glucose levels in an animal comprising administering to said animal a compound according to any one of claims 1-12.
27. The method of any one of claims 23-26, wherein the animal is human.

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Printed on acid-free paper. ISBN 0-896-03772-6. 217726350 claims doc. 5-2002097

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28. The method of claim 23 or 25, wherein the disease or condition is a metabolic disease or condition.
29. The method of claim 28, wherein the disease or condition is diabetes.
30. The method according to claim 29, wherein the disease or condition is Type 2 diabetes.
31. The method according to claim 28, wherein the disease or condition is obesity.
32. The method according to claim 23 or 25, wherein the disease or condition is a hyperproliferative condition.
33. The method according to claim 32, wherein the hyperproliferative condition is cancer.
34. The method of claim 24 or 26, wherein the blood glucose levels are plasma glucose levels or serum glucose levels.
35. The method of claim 24 or 26, wherein the animal is a diabetic human.

SEQUENCE LISTING

<110> ISIS Pharmaceuticals, Inc.
Lex M. Cowser
Jacqueline Wyatt
Susan M. Freier
Brett P. Monia
Madeline M. Butler
Robert McKay

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Met Glu Met

1

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tac cag gac att cga cat gaa gcc agc gac ttc cca tgc aaa gtc gcg 814
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 Lys Leu Pro Lys Asn Lys Asn Arg Asn Arg Tyr Arg Asp Val Ser Pro
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ttt gac cac agt cgg att aaa ttg cac cag gaa gat aat gac tat atc 910
 Phe Asp His Ser Arg Ile Lys Leu His Gln Glu Asp Asn Asp Tyr Ile
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aat gcc agc ttg ata aaa atg gaa gaa gcc cag agg agc tat att ctc 958
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acc cag ggc cct tta cca aac aca tgt ggg cac ttc tgg gag atg gtg 1006
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tgg gag cag aag agc agg ggc gtg gtc atg ctc aac cgc atc atg gag 1054
 Trp Glu Gln Lys Ser Arg Gly Val Val Met Leu Asn Arg Ile Met Glu
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64

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Asp Val Lys Ser Tyr Tyr Thr Val Arg Gln Leu Glu Leu Glu Asn Leu
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act acc aag gag act cga gag atc ctg cat ttc cac tac acc aca tgg 1246
Thr Thr Lys Glu Thr Arg Glu Ile Leu His Phe His Tyr Thr Thr Trp
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cct gac ttt gga gtc ccc gag tca ccg gct tct ttc ctc aat ttc ctt 1294
Pro Asp Phe Gly Val Pro Glu Ser Pro Ala Ser Phe Leu Asn Phe Leu
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Pro Pro Pro Arg Pro Pro Lys Arg Thr Leu Glu Pro His Asn Gly Lys
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Cys Lys Glu Leu Phe Ser Ser His Gln Trp Val Ser Glu Glu Thr Cys
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Arg Met Val Gly Gly Gly Leu Gln Ser Ala Gln Ala Ser Val Pro Thr
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Glu Glu Glu Leu Ser Ser Thr Glu Glu Glu His Lys Ala His Trp Pro
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