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(54) **RECOMBINANT VWF FORMULATIONS**

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(57) **ABSTRACT**

The present invention provides long-term stable pharmaceu-
tical formulations of recombinant von-Willebrand Factor
(rVWF) and methods for making and administering said for-
mulations.

FIG. 1

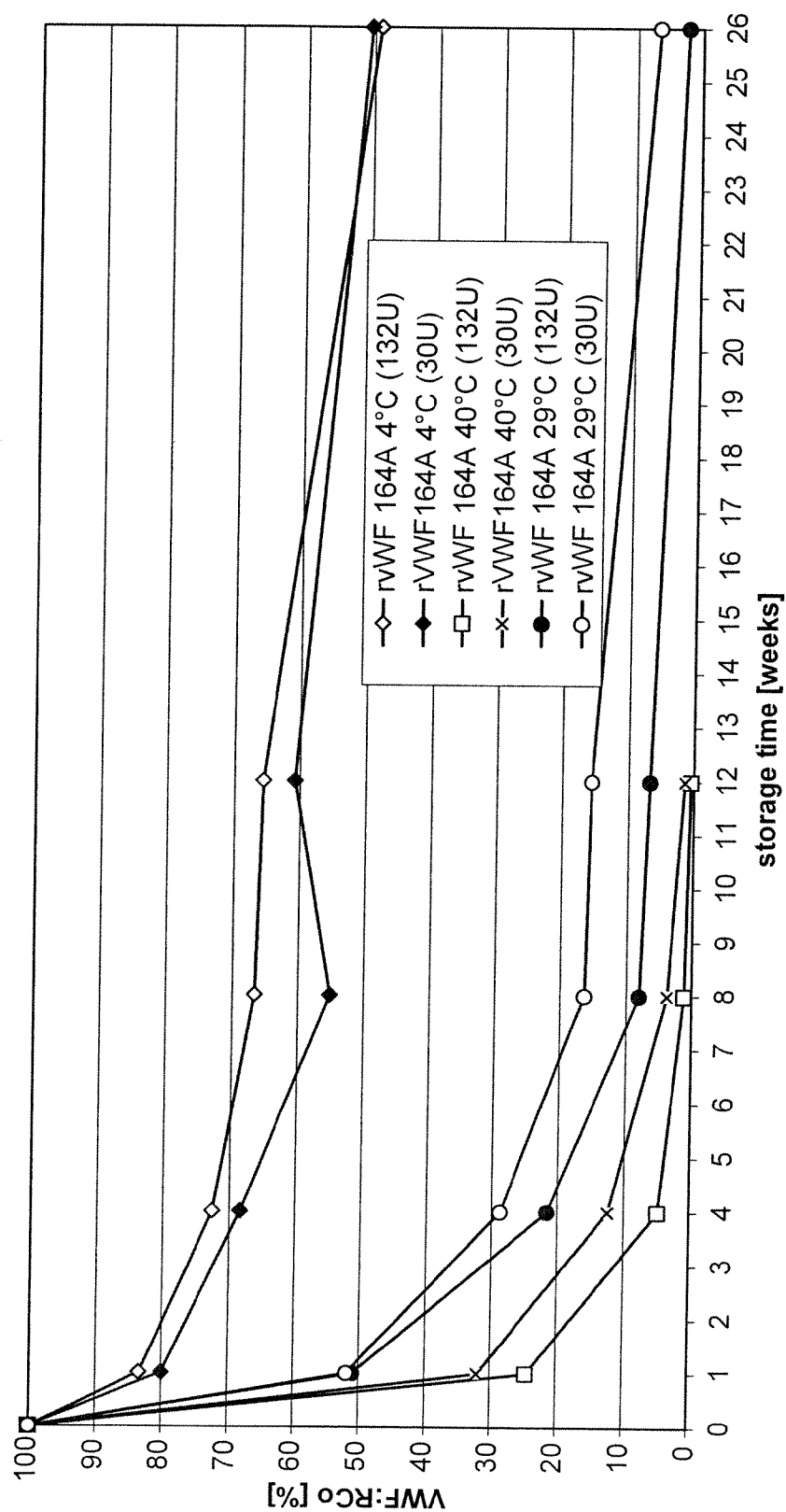


FIG. 2

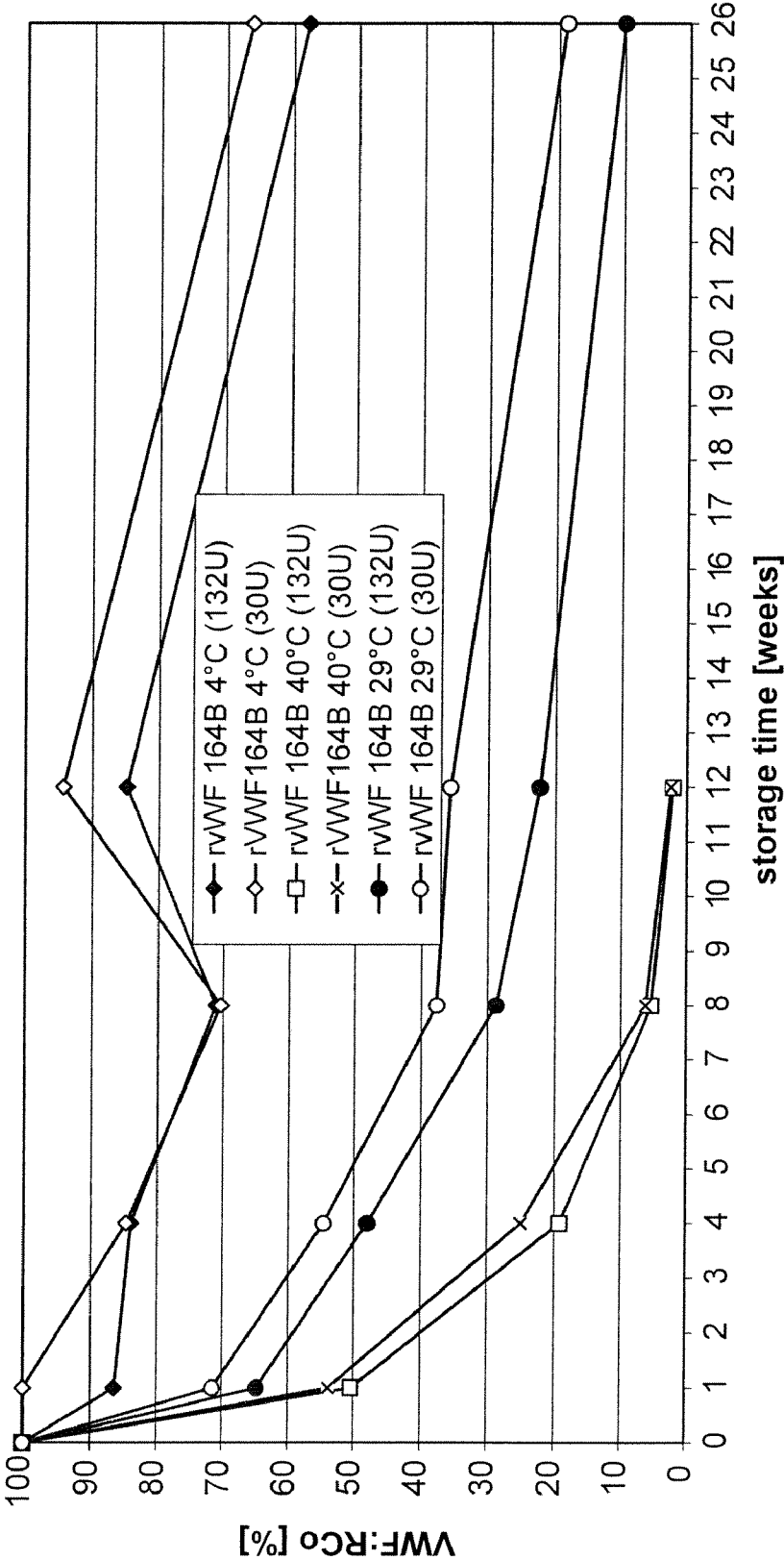


FIG. 4

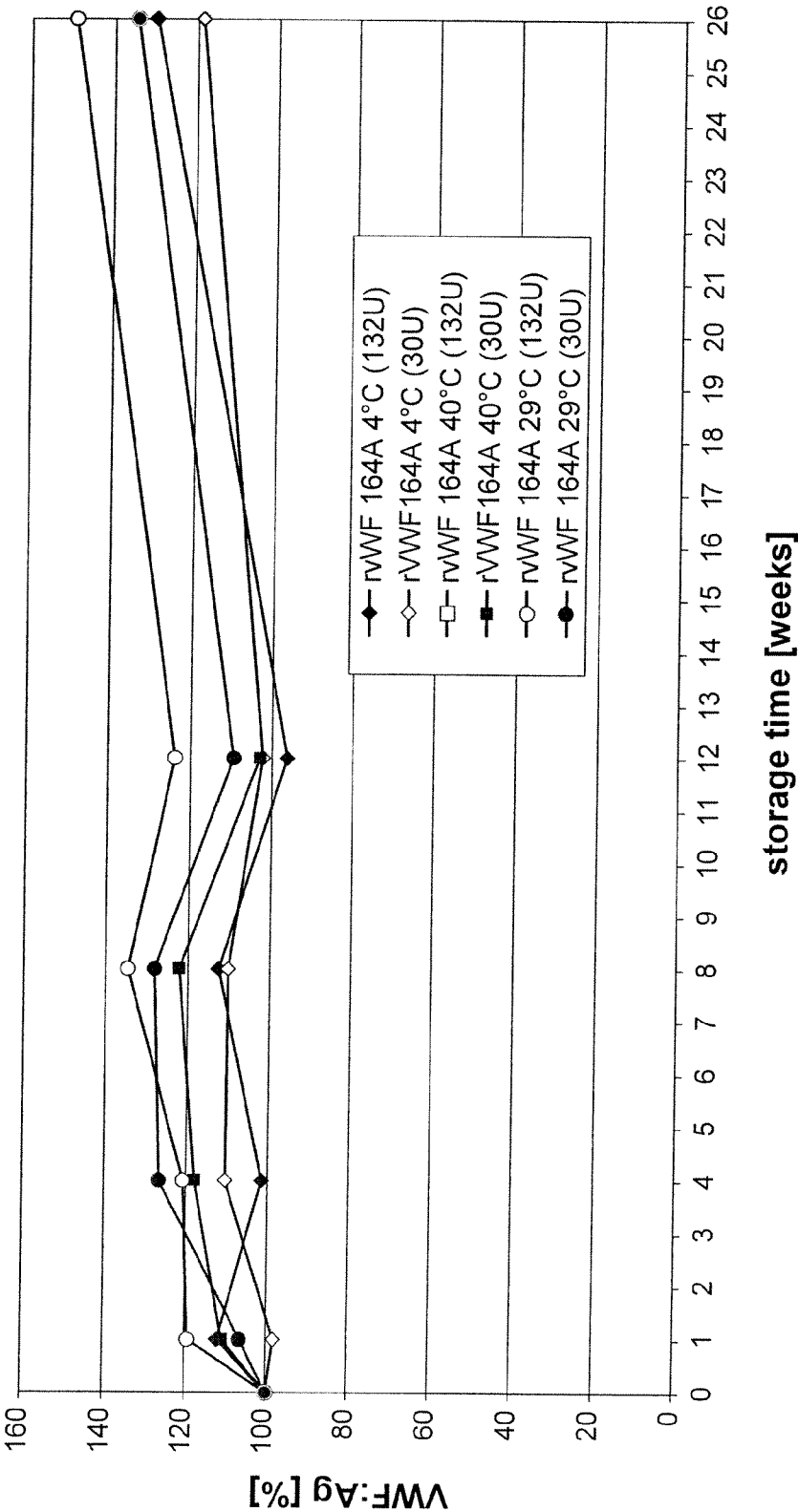


FIG. 5

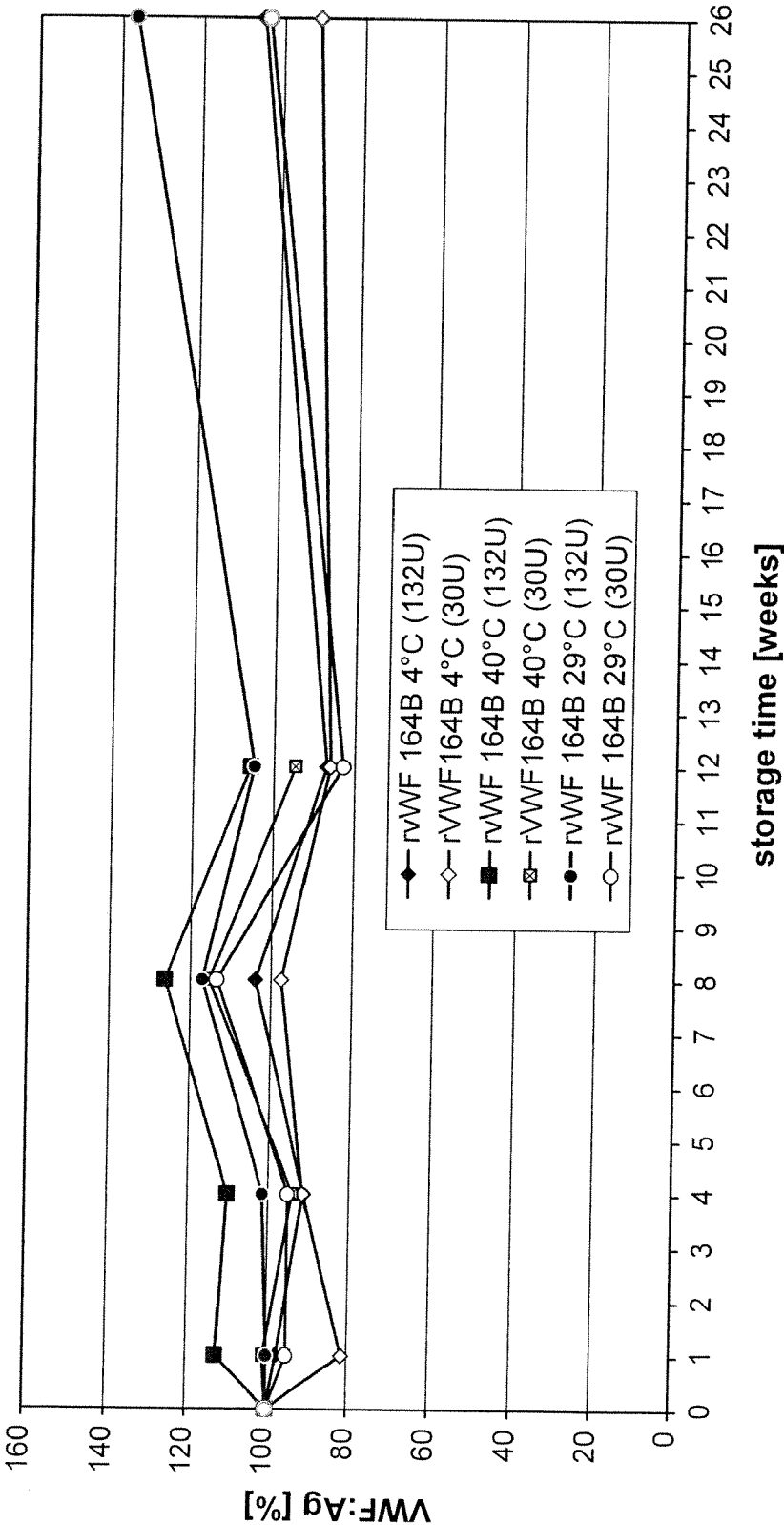


FIG. 6

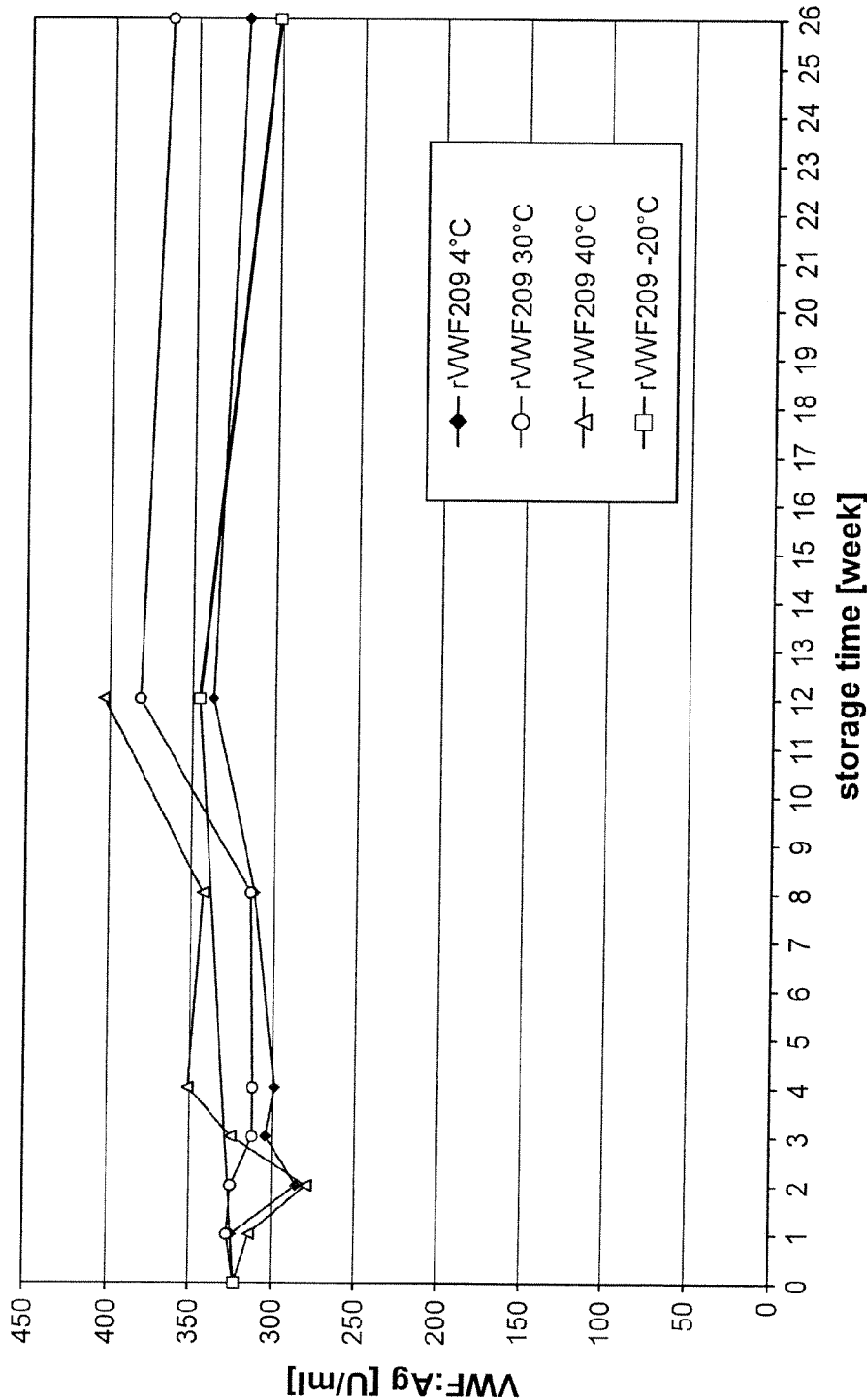


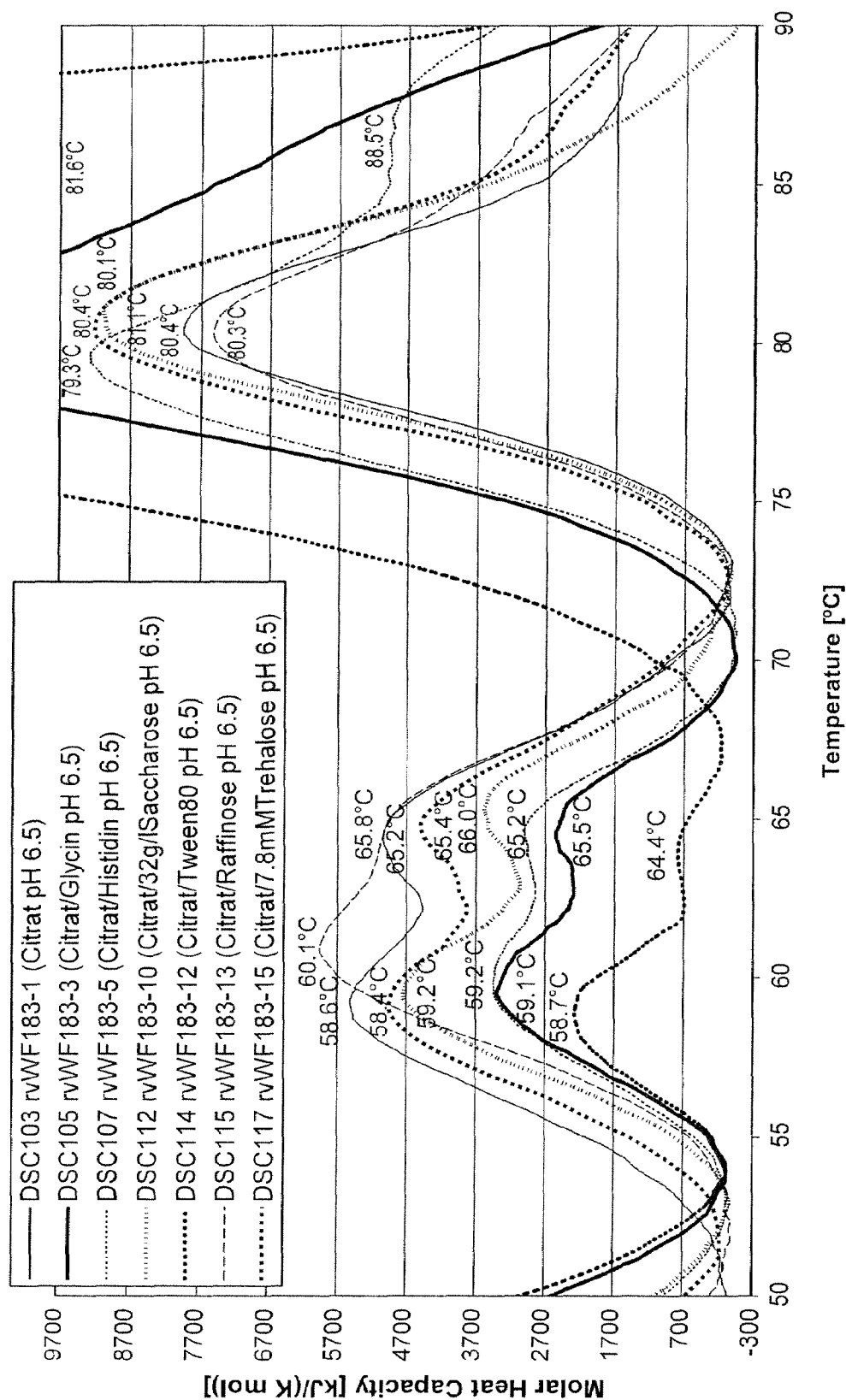
FIG. 7

FIG. 8

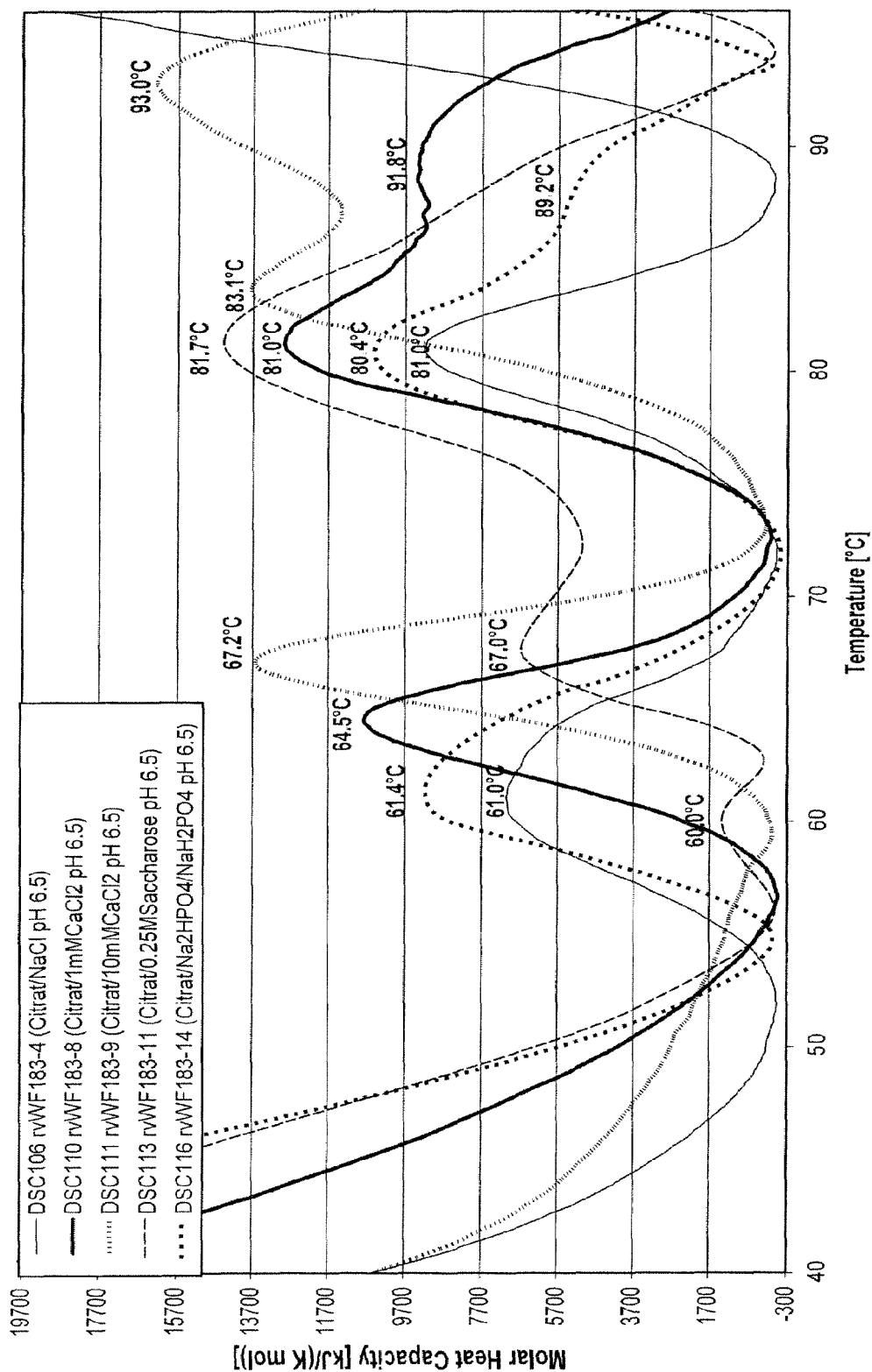
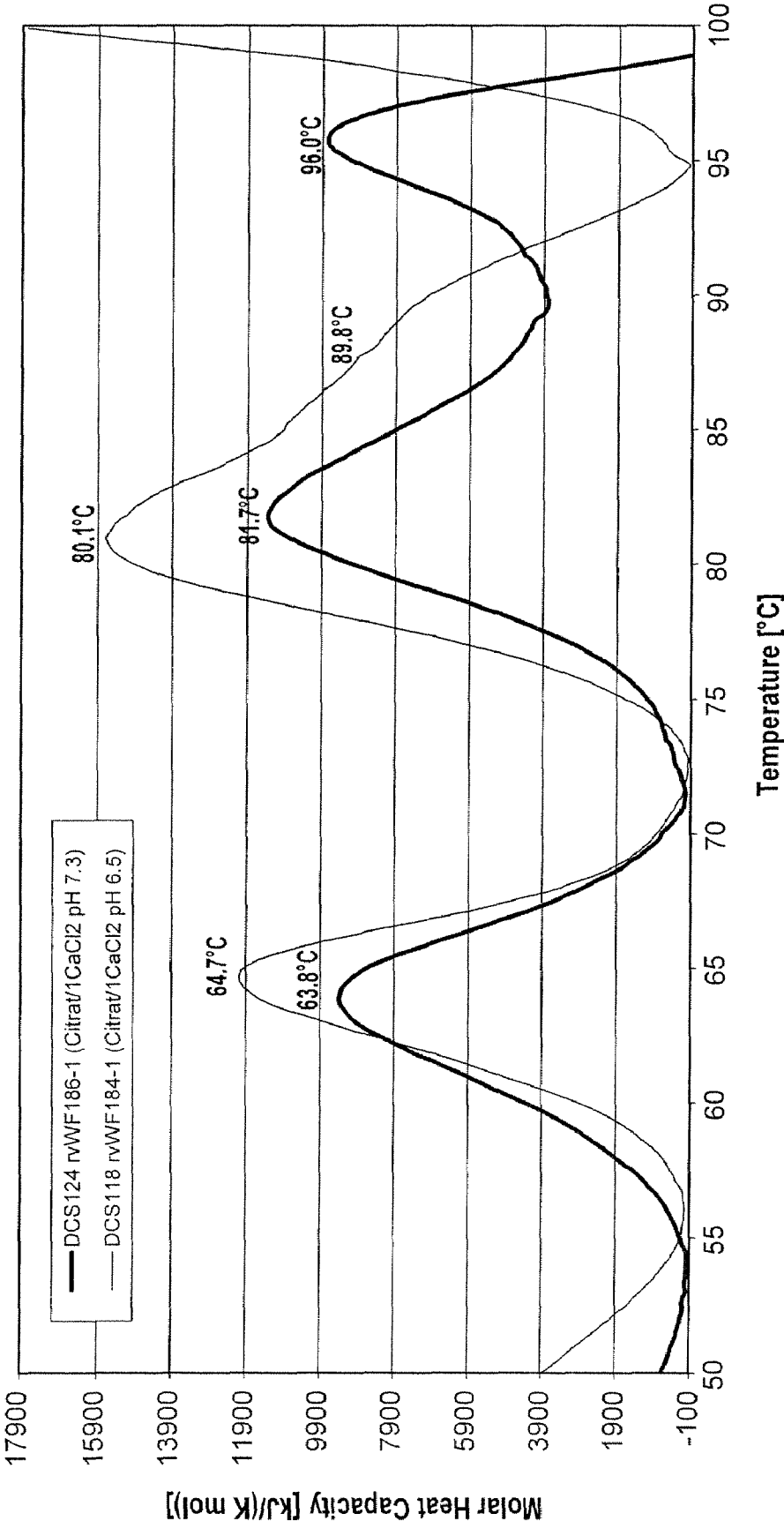


FIG. 9



RECOMBINANT VWF FORMULATIONS

[0001] This application claims priority of U.S. Provisional Application No. 61/017,418, filed Dec. 28, 2007, and U.S. Provisional Application No. 61/017,881 filed Dec. 31, 2007, each of which is incorporated by reference herein in its entirety.

FIELD OF THE INVENTION

[0002] Generally, the invention relates to formulations of recombinant VWF and methods for making a composition comprising recombinant VWF.

BACKGROUND OF THE INVENTION

[0003] Von Willebrand factor (VWF) is a glycoprotein circulating in plasma as a series of multimers ranging in size from about 500 to 20,000 kD. Multimeric forms of VWF are composed of 250 kD polypeptide subunits linked together by disulfide bonds. VWF mediates initial platelet adhesion to the sub-endothelium of the damaged vessel wall. Only the larger multimers exhibit hemostatic activity. It is assumed that endothelial cells secrete large polymeric forms of VWF and those forms of VWF which have a low molecular weight (low molecular weight VWF) arise from proteolytic cleavage. The multimers having large molecular masses are stored in the Weibel-Pallade bodies of endothelial cells and liberated upon stimulation.

[0004] VWF is synthesized by endothelial cells and megakaryocytes as prepro-VWF that consists to a large extent of repeated domains. Upon cleavage of the signal peptide, pro-VWF dimerizes through disulfide linkages at its C-terminal region. The dimers serve as protomers for multimerization, which is governed by disulfide linkages between the free end termini. The assembly to multimers is followed by the proteolytic removal of the propeptide sequence (Leyte et al., *Biochem. J.* 274 (1991), 257-261).

[0005] The primary translation product predicted from the cloned cDNA of VWF is a 2813-residue precursor polypeptide (prepro-VWF). The prepro-VWF consists of a 22 amino acid signal peptide and a 741 amino acid propeptide, with the mature VWF comprising 2050 amino acids (Ruggeri Z. A., and Ware, J., *FASEB J.*, 308-316 (1993)).

[0006] Defects in VWF are causal to Von Willebrand disease (VWD), which is characterized by a more or less pronounced bleeding phenotype. VWD type 3 is the most severe form in which VWF is completely missing, and VWD type 1 relates to a quantitative loss of VWF and its phenotype can be very mild. VWD type 2 relates to qualitative defects of VWF and can be as severe as VWD type 3. VWD type 2 has many sub forms, some being associated with the loss or the decrease of high molecular weight multimers. Von Willebrand syndrome type 2a (VWS-2A) is characterized by a loss of both intermediate and large multimers. VWS-2B is characterized by a loss of highest-molecular-weight multimers. Other diseases and disorders related to VWF are known in the art.

[0007] US. Pat. Nos. 6,531,577, 7,166,709, and European Patent Application No. 04380188.5, describe plasma-derived VWF formulations. However, in addition to quantity and purity issues with plasma-derived VWF, there is also a risk of blood-borne pathogens (e.g., viruses and Variant Creutzfeldt-Jakob disease (vCJD)).

[0008] Thus there exists a need in the art to develop a stable pharmaceutical formulation comprising recombinant VWF.

SUMMARY OF THE INVENTION

[0009] The present invention provides formulations useful for compositions comprising recombinant VWF, resulting in a highly stable pharmaceutical composition. The stable pharmaceutical composition is useful as a therapeutic agent in the treatment of individuals suffering from disorders or conditions that can benefit from the administration of recombinant VWF.

[0010] In one embodiment, the invention provides a stable liquid pharmaceutical formulation of a recombinant von Willebrand Factor (rVWF) comprising: (a) a rVWF; (b) a buffering agent; (c) one or more salts; (d) optionally a stabilizing agent; and (e) optionally a surfactant; wherein the rVWF comprises a polypeptide selected from the group consisting of: a) the amino acid sequence set out in SEQ ID NO: 3; b) a biologically active analog, fragment or variant of a); c) a polypeptide encoded by the polynucleotide set out in SEQ ID NO: 1; d) a biologically active analog, fragment or variant of c); and e) a polypeptide encoded by a polynucleotide that hybridizes to the polynucleotide set out in SEQ ID NO: 1 under moderately stringent hybridization conditions; wherein the buffer is comprised of a pH buffering agent in a range of about 0.1 mM to about 500 mM and wherein the pH is in a range of about 2.0 to about 12.0; wherein the salt is at a concentration of about 1 to 500 mM; wherein the stabilizing agent is at a concentration of about 0.1 to 1000 mM; and wherein the surfactant is at a concentration of about 0.01 g/L to 0.5 g/L.

[0011] In another embodiment, the aforementioned formulation is provided wherein the rVWF comprises the amino acid sequence set out in SEQ ID NO: 3. In another embodiment, an aforementioned formulation is provided wherein the buffering agent is selected from the group consisting of sodium citrate, glycine, histidine, Tris and combinations of these agents. In yet another embodiment, an aforementioned formulation is provided wherein the buffering agent is citrate. In still another embodiment of the invention, the aforementioned formulation is provided wherein pH is in the range of 6.0-8.0, or 6.5-7.3. In a related embodiment, the aforementioned formulation is provided wherein the pH is 7.0. In another embodiment, an aforementioned formulation is provided wherein the buffering agent is citrate and the pH is 7.0.

[0012] In still another embodiment, an aforementioned formulation is provided wherein the salt is selected from the group consisting of calcium chloride, sodium chloride and magnesium chloride. In another embodiment, the aforementioned formulation is provided wherein the salt is at a concentration range of 0.5 to 300 mM. In another embodiment, the aforementioned formulation is provided wherein the salt is calcium chloride at a concentration of 10 mM.

[0013] In another embodiment, an aforementioned formulation is provided wherein the rVWF comprises the amino acid sequence set out in SEQ ID NO: 3; wherein the buffering agent is citrate and the pH is 7.0; and wherein the salt is calcium chloride at a concentration of 10 mM. In still another embodiment, an aforementioned formulation is provided wherein the rVWF comprises the amino acid sequence set out in SEQ ID NO: 3; wherein the buffering agent is sodium citrate and the pH is 7.0; and wherein the salt is calcium chloride at a concentration of 10 mM and NaCl at a concentration of 100 mM.

[0014] Other formulations are also contemplated by the instant invention. For example, in one embodiment, an aforementioned formulation is provided wherein the one or more buffering agents is histidine and Tris at a concentration of 3.3 mM each. In another embodiment, the aforementioned formulation is provided wherein the pH is 7.0. In yet another embodiment, an aforementioned formulation is provided wherein the first salt is sodium chloride at a concentration of 30 mM and the second salt is calcium chloride at a concentration of 0.56 mM.

[0015] In still another embodiment of the invention, an aforementioned formulation is provided wherein the stabilizing agent is selected from the group consisting of mannitol, lactose, sorbitol, xylitol, sucrose, trehalose, mannose, maltose, lactose, glucose, raffinose, cellobiose, gentiobiose, isomaltose, arabinose, glucosamine, fructose and combinations of these stabilizing agents. In another embodiment, the aforementioned formulation is provided wherein the stabilizing agents are trehalose at a concentration of 7.8 mM and mannitol at a concentration of 58.6 mM.

[0016] In another embodiment, an aforementioned formulation is provided wherein the surfactant is selected from the group consisting of digitonin, Triton X-100, Triton X-114, TWEEN-20, TWEEN-80 and combinations of these surfactants. In another embodiment, the aforementioned formulation is provided wherein the surfactant is TWEEN-80 at 0.03 g/L.

[0017] In one embodiment of the invention, an aforementioned formulation is provided wherein the rVWF comprises amino acid sequence set out in SEQ ID NO: 3; wherein the buffering agents are histidine at a concentration of 3.3 mM and Tris at a concentration of 3.3 mM at pH 7.0; wherein the first salt is sodium chloride at a concentration of 30 mM and the second salt is calcium chloride at a concentration of 0.56 mM; wherein the stabilizing agents are trehalose at a concentration of 7.8 mM mannitol at a concentration of 58.6 mM; and wherein the surfactant is TWEEN-80 at 0.03 g/L.

BRIEF DESCRIPTION OF THE FIGURES

[0018] FIG. 1 shows that rVWF is not stable in Advate buffer after 26 weeks, due to the presence of glutathione.

[0019] FIG. 2 shows that rVWF is stable in Advate 1:3 buffer for up to 12 weeks at 4° C.

[0020] FIG. 3 shows that the stability of a citrate-based formulation is better than Advate 1:3 buffer formulation containing 0.1M glutathione.

[0021] FIG. 4 shows that rVWF concentration is stable over 26 weeks in Advate buffer.

[0022] FIG. 5 shows that rVWF concentration is stable over time in Advate 1:3 buffer.

[0023] FIG. 6 shows that rVWF concentration is stable over time in citrate-based buffer.

[0024] FIG. 7 shows that most excipients increase the unfolding temperature of rVWF by about 1 or 2° C.

[0025] FIG. 8 shows that 10 mM CaCl₂ increases unfolding temperature of rVWF by about 8° C. to about 67° C.

[0026] FIG. 9 shows that the effect of CaCl₂ is similar at pH 7.3 and pH 6.5.

DETAILED DESCRIPTION OF THE INVENTION

Definition of Terms

[0027] Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly

understood by one of ordinary skill in the art to which this invention belongs. The following references provide one of skill with a general definition of many of the terms used in this invention: Singleton, et al., *DICTIONARY OF MICROBIOLOGY AND MOLECULAR BIOLOGY* (2d ed. 1994); *THE CAMBRIDGE DICTIONARY OF SCIENCE AND TECHNOLOGY* (Walker ed., 1988); *THE GLOSSARY OF GENETICS*, 5TH ED., R. Rieger, et al. (eds.), Springer Verlag (1991); and Hale and Marham, *THE HARPER COLLINS DICTIONARY OF BIOLOGY* (1991).

[0028] Each publication, patent application, patent, and other reference cited herein is incorporated by reference in its entirety to the extent that it is not inconsistent with the present disclosure.

[0029] It is noted here that, as used in this specification and the appended claims, the singular forms “a,” “an,” and “the” include plural reference unless the context clearly dictates otherwise.

[0030] As used herein, the following terms have the meanings ascribed to them unless specified otherwise.

[0031] The term “comprising,” with respect to a peptide compound, means that a compound may include additional amino acids at either or both amino and carboxy termini of the given sequence. Of course, these additional amino acids should not significantly interfere with the activity of the compound. With respect to a composition of the instant invention, the term “comprising” means that a composition may include additional components. These additional components should not significantly interfere with the activity of the composition.

[0032] The term “pharmacologically active” means that a substance so described is determined to have activity that affects a medical parameter (e.g., but not limited to blood pressure, blood cell-count, cholesterol level) or disease state (e.g., but not limited to cancer, autoimmune disorders).

[0033] As used herein the terms “express,” “expressing” and “expression” mean allowing or causing the information in a gene or DNA sequence to become manifest, for example, producing a protein by activating the cellular functions involved in transcription and translation of a corresponding gene or DNA sequence. A DNA sequence is expressed in or by a cell to form an “expression product” such as a protein. The expression product itself, e.g. the resulting protein, may also be said to be “expressed.” An expression product can be characterized as intracellular, extracellular or secreted. The term “intracellular” means inside a cell. The term “extracellular” means outside a cell, such as a transmembrane protein. A substance is “secreted” by a cell if it appears in significant measure outside the cell, from somewhere on or inside the cell.

[0034] As used herein a “polypeptide” refers to a polymer composed of amino acid residues, structural variants, related naturally-occurring structural variants, and synthetic non-naturally occurring analogs thereof linked via peptide bonds. Synthetic polypeptides can be prepared, for example, using an automated polypeptide synthesizer. The term “protein” typically refers to large polypeptides. The term “peptide” typically refers to short polypeptides.

[0035] As used herein a “fragment” of a polypeptide is meant to refer to any portion of a polypeptide or protein smaller than the full-length polypeptide or protein expression product.

[0036] As used herein an “analog” refers to any of two or more polypeptides substantially similar in structure and hav-

ing the same biological activity, but can have varying degrees of activity, to either the entire molecule, or to a fragment thereof. Analogs differ in the composition of their amino acid sequences based on one or more mutations involving substitution of one or more amino acids for other amino acids. Substitutions can be conservative or non-conservative based on the physico-chemical or functional relatedness of the amino acid that is being replaced and the amino acid replacing it.

[0037] As used herein a "variant" refers to a polypeptide, protein or analog thereof that is modified to comprise additional chemical moieties not normally a part of the molecule. Such moieties may modulate the molecule's solubility, absorption, biological half-life, etc. The moieties may alternatively decrease the toxicity of the molecule and eliminate or attenuate any undesirable side effect of the molecule, etc. Moieties capable of mediating such effects are disclosed in Remington's Pharmaceutical Sciences (1980). Procedure for coupling such moieties to a molecule are well known in the art. For example, the variant may be a blood clotting factor having a chemical modification which confers a longer half-life in vivo to the protein. In various aspects, polypeptides are modified by glycosylation, pegylation, and/or polysialylation.

[0038] Recombinant VWF

[0039] The polynucleotide and amino acid sequences of prepro-VWF are set out in SEQ ID NO:1 and SEQ ID NO:2, respectively, and are available at GenBank Accession Nos. NM_000552 and NP_000543, respectively. The amino acid sequence corresponding to the mature VWF protein is set out in SEQ ID NO: 3 (corresponding to amino acids 764-2813 of the full length prepro-VWF amino acid sequence).

[0040] One form of useful rVWF has at least the property of in vivo-stabilizing, e.g. binding, of at least one Factor VIII (FVIII) molecule and having optionally a glycosylation pattern which is pharmacologically acceptable. Specific examples thereof include VWF without A2 domain thus resistant to proteolysis (Lankhof et al., *Thromb. Haemost.* 77: 1008-1013, 1997), and the VWF fragment from Val 449 to Asn 730 including the glycoprotein Ib-binding domain and binding sites for collagen and heparin (Pietu et al., *Biochem. Biophys. Res. Commun.* 164: 1339-1347, 1989). The determination of the ability of a VWF to stabilize at least one FVIII molecule can be carried out in VWF-deficient mammals according to methods known in the state in the art.

[0041] The rVWF of the present invention may be produced by any method known in the art. One specific example is disclosed in WO86/06096 published on Oct. 23, 1986 and U.S. patent application Ser. No. 07/559,509, filed on Jul. 23, 1990, which is incorporated herein by reference with respect to the methods of producing recombinant VWF. Thus, methods are known in the art for (i) the production of recombinant DNA by genetic engineering, e.g. via reverse transcription of RNA and/or amplification of DNA, (ii) introducing recombinant DNA into procaryotic or eucaryotic cells by transfection, e.g. via electroporation or microinjection, (iii) cultivating said transformed cells, e.g. in a continuous or batchwise manner, (iv) expressing VWF, e.g. constitutively or upon induction, and (v) isolating said VWF, e.g. from the culture medium or by harvesting the transformed cells, in order to (vi) obtain purified rVWF, e.g. via anion exchange chromatography or affinity chromatography. A recombinant VWF may be made in transformed host cells using recombinant DNA techniques well known in the art. For instance,

sequences coding for the polypeptide could be excised from DNA using suitable restriction enzymes.

[0042] Alternatively, the DNA molecule could be synthesized using chemical synthesis techniques, such as the phosphoramidate method. Also, a combination of these techniques could be used.

[0043] The invention also provides vectors encoding polypeptides of the invention in an appropriate host. The vector comprises the polynucleotide that encodes the polypeptide operatively linked to appropriate expression control sequences. Methods of effecting this operative linking, either before or after the polynucleotide is inserted into the vector, are well known. Expression control sequences include promoters, activators, enhancers, operators, ribosomal binding sites, start signals, stop signals, cap signals, polyadenylation signals, and other signals involved with the control of transcription or translation. The resulting vector having the polynucleotide therein is used to transform an appropriate host. This transformation may be performed using methods well known in the art.

[0044] Any of a large number of available and well-known host cells may be used in the practice of this invention. The selection of a particular host is dependent upon a number of factors recognized by the art, including, for example, compatibility with the chosen expression vector, toxicity of the peptides encoded by the DNA molecule, rate of transformation, ease of recovery of the peptides, expression characteristics, bio-safety and costs. A balance of these factors must be struck with the understanding that not all host cells are equally effective for the expression of a particular DNA sequence. Within these general guidelines, useful microbial host cells include bacteria, yeast and other fungi, insects, plants, mammalian (including human) cells in culture, or other hosts known in the art.

[0045] Next, the transformed host is cultured and purified. Host cells may be cultured under conventional fermentation conditions so that the desired compounds are expressed. Such fermentation conditions are well known in the art. Finally, the polypeptides are purified from culture by methods well known in the art.

[0046] Depending on the host cell utilized to express a compound of the invention, carbohydrate (oligosaccharide) groups may conveniently be attached to sites that are known to be glycosylation sites in proteins. Generally, O-linked oligosaccharides are attached to serine (Ser) or threonine (Thr) residues while N-linked oligosaccharides are attached to asparagine (Asn) residues when they are part of the sequence Asn-X-Ser/Thr, where X can be any amino acid except proline. X is preferably one of the 19 naturally occurring amino acids not counting proline. The structures of N-linked and O-linked oligosaccharides and the sugar residues found in each type are different. One type of sugar that is commonly found on both is N-acetylneuraminic acid (referred to as sialic acid). Sialic acid is usually the terminal residue of both N-linked and O-linked oligosaccharides and, by virtue of its negative charge, may confer acidic properties to the glycosylated compound. Such site(s) may be incorporated in the linker of the compounds of this invention and are preferably glycosylated by a cell during recombinant production of the polypeptide compounds (e.g., in mammalian cells such as CHO, BHK, COS). However, such sites may further be glycosylated by synthetic or semi-synthetic procedures known in the art.

[0047] Alternatively, the compounds may be made by synthetic methods. For example, solid phase synthesis techniques may be used. Suitable techniques are well known in the art, and include those described in Merrifield (1973), *Chem. Polypeptides*, pp. 335-61 (Katsoyannis and Panayotis eds.); Merrifield (1963), *J. Am. Chem. Soc.* 85: 2149; Davis et al. (1985), *Biochem. Intl.* 10: 394-414; Stewart and Young (1969), *Solid Phase Peptide Synthesis*; U.S. Pat. No. 3,941,763; Finn et al. (1976), *The Proteins* (3rd ed.) 2: 105-253; and Erickson et al. (1976), *The Proteins* (3rd ed.) 2: 257-527. Solid phase synthesis is the preferred technique of making individual peptides since it is the most cost-effective method of making small peptides.

[0048] Fragments, Variants and Analogs of VWF

[0049] Methods for preparing polypeptide fragments, variants or analogs are well-known in the art.

[0050] Fragments of a polypeptide are prepared using, without limitation, enzymatic cleavage (e.g., trypsin, chymotrypsin) and also using recombinant means to generate a polypeptide fragments having a specific amino acid sequence. Polypeptide fragments may be generated comprising a region of the protein having a particular activity, such as a multimerization domain or any other identifiable VWF domain known in the art.

[0051] Methods of making polypeptide analogs are also well-known. Amino acid sequence analogs of a polypeptide can be substitutional, insertional, addition or deletion analogs. Deletion analogs, including fragments of a polypeptide, lack one or more residues of the native protein which are not essential for function or immunogenic activity. Insertional analogs involve the addition of, e.g., amino acid(s) at a non-terminal point in the polypeptide. This analog may include insertion of an immunoreactive epitope or simply a single residue. Addition analogs, including fragments of a polypeptide, include the addition of one or more amino acids at either of both termini of a protein and include, for example, fusion proteins.

[0052] Substitutional analogs typically exchange one amino acid of the wild-type for another at one or more sites within the protein, and may be designed to modulate one or more properties of the polypeptide without the loss of other functions or properties. In one aspect, substitutions are conservative substitutions. By "conservative amino acid substitution" is meant substitution of an amino acid with an amino acid having a side chain of a similar chemical character. Similar amino acids for making conservative substitutions include those having an acidic side chain (glutamic acid, aspartic acid); a basic side chain (arginine, lysine, histidine); a polar amide side chain (glutamine, asparagine); a hydrophobic, aliphatic side chain (leucine, isoleucine, valine, alanine, glycine); an aromatic side chain (phenylalanine, tryptophan, tyrosine); a small side chain (glycine, alanine, serine, threonine, methionine); or an aliphatic hydroxyl side chain (serine, threonine).

[0053] Analogs may be substantially homologous or substantially identical to the recombinant VWF from which they are derived. Preferred analogs are those which retain at least some of the biological activity of the wild-type polypeptide, e.g. blood clotting activity.

[0054] Polypeptide variants contemplated include polypeptides chemically modified by such techniques as ubiquitination, glycosylation, including polysialation, conjugation to therapeutic or diagnostic agents, labeling, covalent polymer attachment such as pegylation (derivatization with

polyethylene glycol), introduction of non-hydrolyzable bonds, and insertion or substitution by chemical synthesis of amino acids such as ornithine, which do not normally occur in human proteins. Variants retain the same or essentially the same binding properties of non-modified molecules of the invention. Such chemical modification may include direct or indirect (e.g., via a linker) attachment of an agent to the VWF polypeptide. In the case of indirect attachment, it is contemplated that the linker may be hydrolyzable or non-hydrolyzable.

[0055] Preparing pegylated polypeptide analogs will generally comprise the steps of (a) reacting the polypeptide with polyethylene glycol (such as a reactive ester or aldehyde derivative of PEG) under conditions whereby the binding construct polypeptide becomes attached to one or more PEG groups, and (b) obtaining the reaction product(s). In general, the optimal reaction conditions for the acylation reactions will be determined based on known parameters and the desired result. For example, the larger the ratio of PEG: protein, the greater the percentage of poly-pegylated product. In some embodiments, the binding construct will have a single PEG moiety at the N-terminus. Polyethylene glycol (PEG) may be attached to the blood clotting factor via acylation or reductive alkylation through a natural or engineered reactive group on the PEG moiety (e.g., an aldehyde, amino, thiol, or ester group) to a reactive group on the blood clotting factor (e.g., an aldehyde, amino, or ester group) or by any other technique known in the art.

[0056] Methods for preparing polysialylated polypeptide are described in United States Patent Publication 20060160948, Fernandes et Gregoriadis; *Biochim. Biophys. Acta* 1341: 26-34, 1997, and Saenko et al., *Haemophilia* 12:42-51, 2006. Briefly, a solution of colominic acid containing 0.1 M NaIO₄ is stirred in the dark at room temperature to oxidize the CA. The activated CA solution is dialyzed against, e.g., 0.05 M sodium phosphate buffer, pH 7.2 in the dark and this solution was added to a rVWF solution and incubated for 18 h at room temperature in the dark under gentle shaking. Free reagents can then be separated from the rVWF-polysialic acid conjugate by ultrafiltration/diafiltration. Conjugation of rVWF with polysialic acid may also be achieved using glutaraldehyde as cross-linking reagent (Migneault et al., *Biotechniques* 37: 790-796, 2004).

[0057] It is further contemplated that a polypeptide of the invention may be a fusion protein with a second agent which is a polypeptide. In one embodiment, the second agent which is a polypeptide, without limitation, is an enzyme, a growth factor, an antibody, a cytokine, a chemokine, a cell-surface receptor, the extracellular domain of a cell surface receptor, a cell adhesion molecule, or fragment or active domain of a protein described above. In a related embodiment, the second agent is a blood clotting factor such as Factor VIII, Factor VII, Factor IX. The fusion protein contemplated is made by chemical or recombinant techniques well-known in the art.

[0058] It is also contemplated that prepro-VWF and pro-VWF polypeptides may provide a therapeutic benefit in the formulations of the present invention. For example, U.S. Pat. No. 7,005,502 describes a pharmaceutical preparation com-

prising substantial amounts of pro-VWF that induces thrombin generation in vitro. In addition to recombinant, biologically active fragments, variants, or analogs of the naturally-occurring mature VWF, the present invention contemplates the use of recombinant biologically active fragments, variants, or analogs of the prepro-VWF (set out in SEQ ID NO:2) or pro-VWF polypeptides (amino acid residues 23 to 764 of SEQ ID NO: 2) in the formulations described herein.

[0059] Polynucleotides encoding fragments, variants and analogs may be readily generated by a worker of skill to encode biologically active fragments, variants, or analogs of the naturally-occurring molecule that possess the same or similar biological activity to the naturally-occurring molecule. These polynucleotides can be prepared using PCR techniques, digestion/ligation of DNA encoding molecule, and the like. Thus, one of skill in the art will be able to generate single base changes in the DNA strand to result in an altered codon and a missense mutation, using any method known in the art, including, but not limited to site-specific mutagenesis. As used herein, the phrase “moderately stringent hybridization conditions” means, for example, hybridization at 42° C. in 50% formamide and washing at 60° C. in 0.1×SSC, 0.1% SDS. It is understood by those of skill in the art that variation in these conditions occurs based on the length and GC nucleotide base content of the sequences to be hybridized. Formulas standard in the art are appropriate for determining exact hybridization conditions. See Sambrook et al., 9.47-9.51 in *Molecular Cloning*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1989).

[0060] Formulations and Excipients in General

[0061] Excipients are additives that are included in a formulation because they either impart or enhance the stability and delivery of a drug product. Regardless of the reason for their inclusion, excipients are an integral component of a drug product and therefore need to be safe and well tolerated by patients. For protein drugs, the choice of excipients is particularly important because they can affect both efficacy and immunogenicity of the drug. Hence, protein formulations need to be developed with appropriate selection of excipients that afford suitable stability, safety, and marketability.

[0062] The principal challenge in developing formulations for therapeutic proteins is stabilizing the product against the stresses of manufacturing, shipping and storage. The role of formulation excipients is to provide stabilization against these stresses. Excipients may also be employed to reduce viscosity of high concentration protein formulations in order to enable their delivery and enhance patient convenience. In general, excipients can be classified on the basis of the mechanisms by which they stabilize proteins against various chemical and physical stresses. Some excipients are used to alleviate the effects of a specific stress or to regulate a particular susceptibility of a specific protein. Other excipients have more general effects on the physical and covalent stabilities of proteins. The excipients described herein are organized either by their chemical type or their functional role in formulations. Brief descriptions of the modes of stabilization are provided when discussing each excipient type.

[0063] Given the teachings and guidance provided herein, those skilled in the art will know what amount or range of excipient can be included in any particular formulation to achieve a biopharmaceutical formulation of the invention that promotes retention in stability of the biopharmaceutical (e.g., a polypeptide). For example, the amount and type of a salt to be included in a biopharmaceutical formulation of the inven-

tion can be selected based on the desired osmolality (i.e., isotonic, hypotonic or hypertonic) of the final solution as well as the amounts and osmolality of other components to be included in the formulation. Similarly, by exemplification with reference to the type of polyol or sugar included in a formulation, the amount of such an excipient will depend on its osmolality.

[0064] By way of example, inclusion of about 5% sorbitol can achieve isotonicity while about 9% of a sucrose excipient is needed to achieve isotonicity. Selection of the amount or range of concentrations of one or more excipients that can be included within a biopharmaceutical formulation of the invention has been exemplified above by reference to salts, polyols and sugars. However, those skilled in the art will understand that the considerations described herein and further exemplified by reference to specific excipients are equally applicable to all types and combinations of excipients including, for example, salts, amino acids, other tonicity agents, surfactants, stabilizers, bulking agents, cryoprotectants, lyoprotectants, anti-oxidants, metal ions, chelating agents and/or preservatives.

[0065] Further, where a particular excipient is reported in molar concentration, those skilled in the art will recognize that the equivalent percent (%) w/v (e.g., (grams of substance in a solution sample/mL of solution)×100%) of solution is also contemplated.

[0066] Of course, a person having ordinary skill in the art would recognize that the concentrations of the excipients described herein share an interdependency within a particular formulation. By way of example, the concentration of a bulking agent may be lowered where, e.g., there is a high polypeptide concentration or where, e.g., there is a high stabilizing agent concentration. In addition, a person having ordinary skill in the art would recognize that, in order to maintain the isotonicity of a particular formulation in which there is no bulking agent, the concentration of a stabilizing agent would be adjusted accordingly (i.e., a “tonicifying” amount of stabilizer would be used). Common excipients are known in the art and can be found in Powell et al., *Compendium of Excipients for Parenteral Formulations* (1998), *PDA J. Pharm. Sci. Technology*, 52:238-311.

[0067] Buffers and Buffering Agents

[0068] The stability of a pharmacologically active polypeptide formulation is usually observed to be maximal in a narrow pH range. This pH range of optimal stability needs to be identified early during pre-formulation studies. Several approaches, such as accelerated stability studies and calorimetric screening studies, have been demonstrated to be useful in this endeavor (Remmele R. L. Jr., et al., *Biochemistry*, 38(16): 5241-7 (1999)). Once a formulation is finalized, the drug product must be manufactured and maintained throughout its shelf-life. Hence, buffering agents are almost always employed to control pH in the formulation.

[0069] Organic acids, phosphates and Tris have been employed routinely as buffers in protein formulations. The buffer capacity of the buffering species is maximal at a pH equal to the pKa and decreases as pH increases or decreases away from this value. Ninety percent of the buffering capacity exists within one pH unit of its pKa. Buffer capacity also increases proportionally with increasing buffer concentration.

[0070] Several factors need to be considered when choosing a buffer. First and foremost, the buffer species and its concentration need to be defined based on its pKa and the

desired formulation pH. Equally important is to ensure that the buffer is compatible with the polypeptide and other formulation excipients, and does not catalyze any degradation reactions. A third important aspect to be considered is the sensation of stinging and irritation the buffer may induce upon administration. For example, citrate is known to cause stinging upon injection (Laursen T, et al., *Basic Clin Pharmacol Toxicol.*, 98(2): 218-21 (2006)). The potential for stinging and irritation is greater for drugs that are administered via the subcutaneous (SC) or intramuscular (IM) routes, where the drug solution remains at the site for a relatively longer period of time than when administered by the IV route where the formulation gets diluted rapidly into the blood upon administration. For formulations that are administered by direct IV infusion, the total amount of buffer (and any other formulation component) needs to be monitored. One has to be particularly careful about potassium ions administered in the form of the potassium phosphate buffer, which can induce cardiovascular effects in a patient (Hollander-Rodriguez J C, et al., *Am. Fam. Physician.*, 73(2): 283-90 (2006)).

[0071] The buffer system present in the compositions is selected to be physiologically compatible and to maintain a desired pH of the pharmaceutical formulation. In one embodiment, the pH of the solution is between pH 2.0 and pH 12.0. For example, the pH of the solution may be 2.0, 2.3, 2.5, 2.7, 3.0, 3.3, 3.5, 3.7, 4.0, 4.3, 4.5, 4.7, 5.0, 5.3, 5.5, 5.7, 6.0, 6.3, 6.5, 6.7, 7.0, 7.3, 7.5, 7.7, 8.0, 8.3, 8.5, 8.7, 9.0, 9.3, 9.5, 9.7, 10.0, 10.3, 10.5, 10.7, 11.0, 11.3, 11.5, 11.7, or 12.0.

[0072] The pH buffering compound may be present in any amount suitable to maintain the pH of the formulation at a predetermined level. In one embodiment, the pH buffering concentration is between 0.1 mM and 500 mM (1 M). For example, it is contemplated that the pH buffering agent is at least 0.1, 0.5, 0.7, 0.8 0.9, 1.0, 1.2, 1.5, 1.7, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 30, 40, 50, 60, 70, 80, 90, 100, 200, 500 mM.

[0073] Exemplary pH buffering agents used to buffer the formulation as set out herein include, but are not limited to glycine, histidine, glutamate, succinate, phosphate, acetate, citrate, Tris and amino acids or mixtures of amino acids, including, but not limited to aspartate, histidine, and glycine.

[0074] Salts

[0075] Salts are often added to increase the ionic strength of the formulation, which can be important for protein solubility, physical stability, and isotonicity. Salts can affect the physical stability of proteins in a variety of ways. Ions can stabilize the native state of proteins by binding to charged residues on the protein's surface. Alternatively, salts can stabilize the denatured state by binding to peptide groups along the protein backbone ($-\text{CONH}-$). Salts can also stabilize the protein native conformation by shielding repulsive electrostatic interactions between residues within a protein molecule. Salts in protein formulations can also shield attractive electrostatic interactions between protein molecules that can lead to protein aggregation and insolubility. In formulations provided, the salt concentration is between 0.1, 1, 10, 20, 30, 40, 50, 80, 100, 120, 150, 200, 300, and 500 mM.

[0076] Stabilizers and Bulking Agents

[0077] In the present pharmaceutical formulations, a stabilizer (or a combination of stabilizers) may be added to prevent or reduce storage-induced aggregation and chemical degradation. A hazy or turbid solution upon reconstitution indicates that the protein has precipitated or at least aggregated. The term "stabilizer" means an excipient capable of preventing

aggregation or other physical degradation, as well as chemical degradation (for example, autolysis, deamidation, oxidation, etc.) in an aqueous state. Stabilizers that are conventionally employed in pharmaceutical compositions include, but are not limited to, sucrose, trehalose, mannose, maltose, lactose, glucose, raffinose, cellobiose, gentiobiose, isomaltose, arabinose, glucosamine, fructose, mannitol, sorbitol, glycine, arginine HCL, poly-hydroxy compounds, including polysaccharides such as dextran, starch, hydroxyethyl starch, cyclodextrins, N-methylpyrrolidene, cellulose and hyaluronic acid, sodium chloride, [Carpenter et al., *Develop. Biol. Standard* 74:225, (1991)]. In the present formulations, the stabilizer is incorporated in a concentration of about 0.1, 0.5, 0.7, 0.8 0.9, 1.0, 1.2, 1.5, 1.7, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 30, 40, 50, 60, 70, 80, 90, 100, 200, 500, 700, 900, or 1000 mM.

[0078] If desired, the formulations also include appropriate amounts of bulking and osmolarity regulating agents. Bulking agents include, for example, mannitol, glycine, sucrose, polymers such as dextran, polyvinylpyrrolidone, carboxymethylcellulose, lactose, sorbitol, trehalose, or xylitol. In one embodiment, the bulking agent is mannitol. The bulking agent is incorporated in a concentration of about 0.1, 0.5, 0.7, 0.8 0.9, 1.0, 1.2, 1.5, 1.7, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 30, 40, 50, 60, 70, 80, 90, 100, 200, 500, 700, 900, or 1000 mM.

[0079] Surfactants

[0080] Protein molecules have a high propensity to interact with surfaces making them susceptible to adsorption and denaturation at air-liquid, vial-liquid, and liquid-liquid (silicone oil) interfaces. This degradation pathway has been observed to be inversely dependent on protein concentration and results in either the formation of soluble and insoluble protein aggregates or the loss of protein from solution via adsorption to surfaces. In addition to container surface adsorption, surface-induced degradation is exacerbated with physical agitation, as would be experienced during shipping and handling of the product.

[0081] Surfactants are commonly used in protein formulations to prevent surface-induced degradation. Surfactants are amphipathic molecules with the capability of out-competing proteins for interfacial positions. Hydrophobic portions of the surfactant molecules occupy interfacial positions (e.g., air/liquid), while hydrophilic portions of the molecules remain oriented towards the bulk solvent. At sufficient concentrations (typically around the detergent's critical micellar concentration), a surface layer of surfactant molecules serve to prevent protein molecules from adsorbing at the interface. Thereby, surface-induced degradation is minimized. The most commonly used surfactants are fatty acid esters of sorbitan polyethoxylates, i.e. polysorbate 20 and polysorbate 80. The two differ only in the length of the aliphatic chain that imparts hydrophobic character to the molecules, C-12 and C-18, respectively. Accordingly, polysorbate-80 is more surface-active and has a lower critical micellar concentration than polysorbate-20.

[0082] Detergents can also affect the thermodynamic conformational stability of proteins. Here again, the effects of a given detergent excipient will be protein specific. For example, polysorbates have been shown to reduce the stability of some proteins and increase the stability of others. Detergent destabilization of proteins can be rationalized in terms of the hydrophobic tails of the detergent molecules that can engage in specific binding with partially or wholly unfolded

protein states. These types of interactions could cause a shift in the conformational equilibrium towards the more expanded protein states (i.e. increasing the exposure of hydrophobic portions of the protein molecule in complement to binding polysorbate). Alternatively, if the protein native state exhibits some hydrophobic surfaces, detergent binding to the native state may stabilize that conformation.

[0083] Another aspect of polysorbates is that they are inherently susceptible to oxidative degradation. Often, as raw materials, they contain sufficient quantities of peroxides to cause oxidation of protein residue side-chains, especially methionine. The potential for oxidative damage arising from the addition of stabilizer emphasizes the point that the lowest effective concentrations of excipients should be used in formulations. For surfactants, the effective concentration for a given protein will depend on the mechanism of stabilization. It has been postulated that if the mechanism of surfactant stabilization is related to preventing surface-denaturation the effective concentration will be around the detergent's critical micellar concentration. Conversely, if the mechanism of stabilization is associated with specific protein-detergent interactions, the effective surfactant concentration will be related to the protein concentration and the stoichiometry of the interaction (Randolph T. W., et al., *Pharm Biotechnol.*, 13:159-75 (2002)).

[0084] Surfactants may also be added in appropriate amounts to prevent surface related aggregation phenomenon during freezing and drying [Chang, B, J. Pharm. Sci. 85:1325, (1996)]. Exemplary surfactants include anionic, cationic, nonionic, zwitterionic, and amphoteric surfactants including surfactants derived from naturally-occurring amino acids. Anionic surfactants include, but are not limited to, sodium lauryl sulfate, dioctyl sodium sulfosuccinate and dioctyl sodium sulfonate, chenodeoxycholic acid, N-lauroylsarcosine sodium salt, lithium dodecyl sulfate, 1-octanesulfonic acid sodium salt, sodium cholate hydrate, sodium deoxycholate, and glycodeoxycholic acid sodium salt. Cationic surfactants include, but are not limited to, benzalkonium chloride or benzethonium chloride, cetylpyridinium chloride monohydrate, and hexadecyltrimethylammonium bromide. Zwitterionic surfactants include, but are not limited to, CHAPS, CHAPSO, SB3-10, and SB3-12. Non-ionic surfactants include, but are not limited to, digitonin, Triton X-100, Triton X-114, TWEEN-20, and TWEEN-80. Surfactants also include, but are not limited to laurumacrogol 400, polyoxyl 40 stearate, polyoxyethylene hydrogenated castor oil 10, 40, 50 and 60, glycerol monostearate, polysorbate 40, 60, 65 and 80, soy lecithin and other phospholipids such as dioleoyl phosphatidyl choline (DOPC), dimyristoylphosphatidyl glycerol (DMPG), dimyristoylphosphatidyl choline (DMPC), and (dioleoyl phosphatidyl glycerol) DOPG; sucrose fatty acid ester, methyl cellulose and carboxymethyl cellulose. Compositions comprising these surfactants, either individually or as a mixture in different ratios, are therefore further provided. In the present formulations, the surfactant is incorporated in a concentration of about 0.01 to about 0.5 g/L.

[0085] Other Common Excipient Components

[0086] Amino Acids

[0087] Amino acids have found versatile use in protein formulations as buffers, bulking agents, stabilizers and antioxidants. Histidine and glutamic acid are employed to buffer protein formulations in the pH range of 5.5-6.5 and 4.0-5.5 respectively. The imidazole group of histidine has a pKa=6.0 and the carboxyl group of glutamic acid side chain has a pKa

of 4.3 which makes these amino acids suitable for buffering in their respective pH ranges. Glutamic acid is particularly useful in such cases (e.g., Stemgen®). Histidine is commonly found in marketed protein formulations (e.g., Xolair®, Herceptin®, Recombinate®), and this amino acid provides an alternative to citrate, a buffer known to sting upon injection. Interestingly, histidine has also been reported to have a stabilizing effect, as observed in formulations with ABX-IL8 (an IgG2 antibody), with respect to aggregation when used at high concentrations in both liquid and lyophilized presentations (Chen B, et al., *Pharm Res.*, 20(12): 1952-60 (2003)). Histidine (up to 60 mM) was also observed to reduce the viscosity of a high concentration formulation of this antibody. However, in the same study, the authors observed increased aggregation and discoloration in histidine containing formulations during freeze-thaw studies of the antibody in stainless steel containers. The authors attributed this to an effect of iron ions leached from corrosion of steel containers. Another note of caution with histidine is that it undergoes photo-oxidation in the presence of metal ions (Tomita M, et al., *Biochemistry*, 8(12): 5149-60 (1969)). The use of methionine as an antioxidant in formulations appears promising; it has been observed to be effective against a number of oxidative stresses (Lam X M, et al., *J Pharm Sci.*, 86(11): 1250-5 (1997)).

[0088] The amino acids glycine, proline, serine and alanine have been shown to stabilize proteins by the mechanism of preferential exclusion. Glycine is also a commonly used bulking agent in lyophilized formulations (e.g., Neumega®, Genotropin®, Humatrope®). Arginine has been shown to be an effective agent in inhibiting aggregation and has been used in both liquid and lyophilized formulations (e.g., Activase®, Avonex®, Enbrel® liquid). Furthermore, the enhanced efficiency of refolding of certain proteins in the presence of arginine has been attributed to its suppression of the competing aggregation reaction during refolding.

[0089] Antioxidants

[0090] Oxidation of protein residues arises from a number of different sources. Beyond the addition of specific antioxidants, the prevention of oxidative protein damage involves the careful control of a number of factors throughout the manufacturing process and storage of the product such as atmospheric oxygen, temperature, light exposure, and chemical contamination. The most commonly used pharmaceutical antioxidants are reducing agents, oxygen/free-radical scavengers, or chelating agents. Antioxidants in therapeutic protein formulations are water-soluble and remain active throughout the product shelf-life. Reducing agents and oxygen/free-radical scavengers work by ablating active oxygen species in solution. Chelating agents such as EDTA are effective by binding trace metal contaminants that promote free-radical formation. For example, EDTA was utilized in the liquid formulation of acidic fibroblast growth factor to inhibit the metal ion catalyzed oxidation of cysteine residues. EDTA has been used in marketed products like Kineret® and Ontak®.

[0091] In addition to the effectiveness of various excipients to prevent protein oxidation, the potential for the antioxidants themselves to induce other covalent or physical changes to the protein is of concern. For example, reducing agents can cause disruption of intramolecular disulfide linkages, which can lead to disulfide shuffling. In the presence of transition metal ions, ascorbic acid and EDTA have been shown to promote methionine oxidation in a number of proteins and peptides (Akers M J, and Defelippis M R. Peptides and Pro-

teins as Parenteral Solutions. In: Pharmaceutical Formulation Development of Peptides and Proteins. Sven Frokjaer, Lars Hovgaard, editors. Pharmaceutical Science. Taylor and Francis, UK (1999); Fransson J. R., *J. Pharm. Sci.* 86(9): 4046-1050 (1997); Yin J, et al., *Pharm. Res.*, 21(12): 2377-83 (2004)). Sodium thiosulfate has been reported to reduce the levels of light and temperature induced methionine-oxidation in rhuMab HER2; however, the formation of a thiosulfate-protein adduct was also reported in this study (Lam X M, Yang JY, et al., *J Pharm Sci.* 86(11): 1250-5 (1997)). Selection of an appropriate antioxidant is made according to the specific stresses and sensitivities of the protein.

[0092] Metal Ions

[0093] In general, transition metal ions are undesired in protein formulations because they can catalyze physical and chemical degradation reactions in proteins. However, specific metal ions are included in formulations when they are co-factors to proteins and in suspension formulations of proteins where they form coordination complexes (e.g., zinc suspension of insulin). Recently, the use of magnesium ions (10-120 mM) has been proposed to inhibit the isomerization of aspartic acid to isoaspartic acid (WO 2004039337).

[0094] Two examples where metal ions confer stability or increased activity in proteins are human deoxyribonuclease (rhDNase, Pulmozyme®), and Factor VIII. In the case of rhDNase, Ca^{+2} ions (up to 100 mM) increased the stability of the enzyme through a specific binding site (Chen B, et al., *J Pharm Sci.*, 88(4): 477-82 (1999)). In fact, removal of calcium ions from the solution with EGTA caused an increase in deamidation and aggregation. However, this effect was observed only with Ca^{+2} ions; other divalent cations Mg^{+2} , Mn^{+2} and Zn^{+2} were observed to destabilize rhDNase. Similar effects were observed in Factor VIII. Ca^{+2} and Sr^{+2} ions stabilized the protein while others like Mg^{+2} , Mn^{+2} and Zn^{+2} , Cu^{+2} and Fe^{+2} destabilized the enzyme (Fatouros, A., et al., *Int. J. Pharm.*, 155, 121-131 (1997)). In a separate study with Factor VIII, a significant increase in aggregation rate was observed in the presence of Al^{+3} ions (Derrick T S, et al., *J. Pharm. Sci.*, 93(10): 2549-57 (2004)). The authors note that other excipients like buffer salts are often contaminated with Al^{+3} ions and illustrate the need to use excipients of appropriate quality in formulated products.

[0095] Preservatives

[0096] Preservatives are necessary when developing multi-use parenteral formulations that involve more than one extraction from the same container. Their primary function is to inhibit microbial growth and ensure product sterility throughout the shelf-life or term of use of the drug product. Commonly used preservatives include benzyl alcohol, phenol and m-cresol. Although preservatives have a long history of use, the development of protein formulations that includes preservatives can be challenging. Preservatives almost always have a destabilizing effect (aggregation) on proteins, and this has become a major factor in limiting their use in multi-dose protein formulations (Roy S, et al., *J Pharm Sci.*, 94(2): 382-96 (2005)).

[0097] To date, most protein drugs have been formulated for single-use only. However, when multi-dose formulations are possible, they have the added advantage of enabling patient convenience, and increased marketability. A good example is that of human growth hormone (hGH) where the development of preserved formulations has led to commercialization of more convenient, multi-use injection pen presentations. At least four such pen devices containing pre-

served formulations of hGH are currently available on the market. Norditropin® (liquid, Novo Nordisk), Nutropin AQ® (liquid, Genentech) & Genotropin (lyophilized—dual chamber cartridge, Pharmacia & Upjohn) contain phenol while Somatrop® (Eli Lilly) is formulated with m-cresol.

[0098] Several aspects need to be considered during the formulation development of preserved dosage forms. The effective preservative concentration in the drug product must be optimized. This requires testing a given preservative in the dosage form with concentration ranges that confer anti-microbial effectiveness without compromising protein stability. For example, three preservatives were successfully screened in the development of a liquid formulation for interleukin-1 receptor (Type I), using differential scanning calorimetry (DSC). The preservatives were rank ordered based on their impact on stability at concentrations commonly used in marketed products (Remmele R L Jr., et al., *Pharm. Res.*, 15(2): 200-8 (1998)).

[0099] Some preservatives can cause injection site reactions, which is another factor that needs consideration when choosing a preservative. In clinical trials that focused on the evaluation of preservatives and buffers in Norditropin, pain perception was observed to be lower in formulations containing phenol and benzyl alcohol as compared to a formulation containing m-cresol (Kappelgaard A. M., *Horm. Res.* 62 Suppl 3:98-103 (2004)). Interestingly, among the commonly used preservative, benzyl alcohol possesses anesthetic properties (Minogue S C, and Sun D A., *Anesth. Analg.*, 100(3): 683-6 (2005)).

[0100] Lyophilization

[0101] It is also contemplated that the formulations comprising a VWF polypeptide of the invention may be lyophilized prior to administration. Lyophilization is carried out using techniques common in the art and should be optimized for the composition being developed [Tang et al., *Pharm. Res.* 21:191-200, (2004) and Chang et al., *Pharm. Res.* 13:243-9 (1996)].

[0102] A lyophilization cycle is, in one aspect, composed of three steps: freezing, primary drying, and secondary drying [A. P. Mackenzie, *Phil. Trans. R. Soc. London, Ser. B, Biol. Sci.* 278:167 (1977)]. In the freezing step, the solution is cooled to initiate ice formation. Furthermore, this step induces the crystallization of the bulking agent. The ice sublimates in the primary drying stage, which is conducted by reducing chamber pressure below the vapor pressure of the ice, using a vacuum and introducing heat to promote sublimation. Finally, adsorbed or bound water is removed at the secondary drying stage under reduced chamber pressure and at an elevated shelf temperature. The process produces a material known as a lyophilized cake. Thereafter the cake can be reconstituted with either sterile water or suitable diluent for injection.

[0103] The lyophilization cycle not only determines the final physical state of the excipients but also affects other parameters such as reconstitution time, appearance, stability and final moisture content. The composition structure in the frozen state proceeds through several transitions (e.g., glass transitions, wettings, and crystallizations) that occur at specific temperatures and can be used to understand and optimize the lyophilization process. The glass transition temperature (T_g and/or T_g') can provide information about the physical state of a solute and can be determined by differential scanning calorimetry (DSC). T_g and T_g' are an important parameter that must be taken into account when designing the lyophilization cycle. For example, T_g' is important for pri-

mary drying. Furthermore, in the dried state, the glass transition temperature provides information on the storage temperature of the final product.

[0104] Methods of Preparation

[0105] The present invention further contemplates methods for the preparation of pharmaceutical formulations. A variety of aqueous carriers, e.g., sterile water for injection, water with preservatives for multi dose use, or water with appropriate amounts of surfactants (for example, polysorbate-20), 0.4% saline, 0.3% glycine, or aqueous suspensions may contain the active compound in admixture with excipients suitable for the manufacture of aqueous suspensions. In various aspects, such excipients are suspending agents, for example sodium carboxymethylcellulose, methylcellulose, hydroxypropylmethylcellulose, sodium alginate, polyvinylpyrrolidone, gum tragacanth and gum acacia; dispersing or wetting agents may be a naturally-occurring phosphatide, for example lecithin, or condensation products of an alkylene oxide with fatty acids, for example polyoxyethylene stearate, or condensation products of ethylene oxide with long chain aliphatic alcohols, for example heptadecaethyl-eneoxycetanol, or condensation products of ethylene oxide with partial esters derived from fatty acids and a hexitol such as polyoxyethylene sorbitol monooleate, or condensation products of ethylene oxide with partial esters derived from fatty acids and hexitol anhydrides, for example polyethylene sorbitan monooleate. The aqueous suspensions may also contain one or more preservatives, for example ethyl, or n-propyl, p-hydroxybenzoate.

[0106] Administration

[0107] To administer compositions to human or test animals, in one aspect, the compositions comprises one or more pharmaceutically acceptable carriers. The phrases "pharmaceutically" or "pharmacologically" acceptable refer to molecular entities and compositions that are stable, inhibit protein degradation such as aggregation and cleavage products, and in addition do not produce allergic, or other adverse reactions when administered using routes well-known in the art, as described below. "Pharmaceutically acceptable carriers" include any and all clinically useful solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents and the like, including those agents disclosed above.

[0108] The pharmaceutical formulations may be administered orally, topically, transdermally, parenterally, by inhalation spray, vaginally, rectally, or by intracranial injection. The term parenteral as used herein includes subcutaneous injections, intravenous, intramuscular, intracisternal injection, or infusion techniques. Administration by intravenous, intradermal, intramuscular, intramammary, intraperitoneal, intrathecal, retrobulbar, intrapulmonary injection and or surgical implantation at a particular site is contemplated as well. Generally, compositions are essentially free of pyrogens, as well as other impurities that could be harmful to the recipient.

[0109] Single or multiple administrations of the compositions can be carried out with the dose levels and pattern being selected by the treating physician. For the prevention or treatment of disease, the appropriate dosage will depend on the type of disease to be treated, as defined above, the severity and course of the disease, whether drug is administered for preventive or therapeutic purposes, previous therapy, the patient's clinical history and response to the drug, and the discretion of the attending physician.

[0110] Kits

[0111] As an additional aspect, the invention includes kits which comprise one or more pharmaceutical formulations packaged in a manner which facilitates their use for administration to subjects. In one embodiment, such a kit includes pharmaceutical formulation described herein (e.g., a composition comprising a therapeutic protein or peptide), packaged in a container such as a sealed bottle or vessel, with a label affixed to the container or included in the package that describes use of the compound or composition in practicing the method. In one embodiment, the pharmaceutical formulation is packaged in the container such that the amount of headspace in the container (e.g., the amount of air between the liquid formulation and the top of the container) is very small. Preferably, the amount of headspace is negligible (i.e., almost none). In one embodiment, the kit contains a first container having a therapeutic protein or peptide composition and a second container having a physiologically acceptable reconstitution solution for the composition. In one aspect, the pharmaceutical formulation is packaged in a unit dosage form. The kit may further include a device suitable for administering the pharmaceutical formulation according to a specific route of administration. Preferably, the kit contains a label that describes use of the pharmaceutical formulations.

[0112] Dosages

[0113] The dosage regimen involved in a method for treating a condition described herein will be determined by the attending physician, considering various factors which modify the action of drugs, e.g. the age, condition, body weight, sex and diet of the patient, the severity of any infection, time of administration and other clinical factors. By way of example, a typical dose of a recombinant VWF of the present invention is approximately 50 U/kg, equal to 500 µg/kg.

[0114] Formulations of the invention may be administered by an initial bolus followed by a continuous infusion to maintain therapeutic circulating levels of drug product. As another example, the inventive compound may be administered as a one-time dose. Those of ordinary skill in the art will readily optimize effective dosages and administration regimens as determined by good medical practice and the clinical condition of the individual patient. The frequency of dosing will depend on the pharmacokinetic parameters of the agents and the route of administration. The optimal pharmaceutical formulation will be determined by one skilled in the art depending upon the route of administration and desired dosage. See for example, Remington's Pharmaceutical Sciences, 18th Ed. (1990, Mack Publishing Co., Easton, Pa. 18042) pages 1435-1712, the disclosure of which is hereby incorporated by reference. Such formulations may influence the physical state, stability, rate of in vivo release, and rate of in vivo clearance of the administered agents. Depending on the route of administration, a suitable dose may be calculated according to body weight, body surface area or organ size. Appropriate dosages may be ascertained through use of established assays for determining blood level dosages in conjunction with appropriate dose-response data. The final dosage regimen will be determined by the attending physician, considering various factors which modify the action of drugs, e.g. the drug's specific activity, the severity of the damage and the responsiveness of the patient, the age, condition, body weight, sex and diet of the patient, the severity of any infection, time of administration and other clinical factors. As studies are con-

ducted, further information will emerge regarding the appropriate dosage levels and duration of treatment for various diseases and conditions.

[0115] The following examples are not intended to be limiting but only exemplary of specific embodiments of the invention.

Example 1

Shaking Experiments

[0116] In order to determine the amount of precipitation of rVWF in various formulations, the percent recovery of rVWF following turbulent shaking was tested under a variety of conditions.

[0117] rVWF in Advate buffer (90 mM NaCl, 1.68 mM CaCl_2 , 10 mM L-histidine, 1 mM tris, 0.26 mM glutathione,

23.4 mM trehalose, 175.7 mM mannitol, and 0.1 g/L TWEEN-80, pH 7.0) or Advate 1:3 buffer (Advate buffer diluted 3-fold in water) was subjected to turbulent shaking on a shaker at room temperature (RT) for 0 min, 1 min, 2.5 hrs, or 4 days, and percent recovery of the rVWF was measured relative to the starting material prior to shaking. As shown in Table 1, losses of about 40-80% were observed in the Advate buffer while losses of about 20-30% were observed in the Advate 1:3 buffer. VWF antigen VWF:Ag corresponds to the amount of VWF which can be detected in an VWF-specific ELISA using polyclonal anti-VWF antibody, while VWF:RCo corresponds to the amount of VWF which causes agglutination of stabilized platelets in the presence of ristocetin. In both cases human reference plasma calibrated against the actual WHO standard was used as standard (1 ml of reference plasma usually contains 1 U VWF).

TABLE 1

Influence of turbulent shaking time on rVWF recovery						
rVWF	Turbulent shaking at RT	VWF:Ag [U/ml]	Recovery [%]	VWF:RCo [U/ml]	Recovery [%]	RCo/VWF:Ag [U/U]
Advate	0 min	213	100%	104	100%	0.49
	1 min	120	56%			
	2.5 hr	139	65%			
	4 d	37	17%	7	7%	0.19
Advate 1:3	0 min	206	100%	134	100%	0.65
	1 min	152	74%			
	2.5 hr	170	82%			
	4 d	138	67%	131	98%	0.95

[0118] The effect of freeze/thawing and lyophilization was also tested in the shaking experiments. Freezing was performed at -20°C . in an -20°C . cold room or on dry ice, thawing in both cases at RT and both started from the liquid formulations. As for lyophilization, the formulated VWF samples described herein were frozen within a pilot scale lyophilizer at $\leq -40^\circ\text{C}$. and were lyophilized using a standard lyo program. Shaking was performed directly with the liquid formulations (2 ml in 5 ml vials). As shown in Table 2, percent recovery of rVWF was higher in Advate 1:3 buffer compared to Advate buffer.

TABLE 2

RVWF		VWF:Ag [U/ml]	VWF:Ag recovery [%]	VWF:RCo [U/ml]	VWF:RCo recovery [%]	RCo:Ag [U/U]
Advate	Frozen	213	100%	104	100%	0.49
	Frozen - 3x at -20°C .	229	107%	84	81%	0.37
	Frozen - 3x with dry ice	231	108%	72	69%	0.31
	Lyo	242	113%	61	59%	0.25
	Starting material	213	100%	104	100%	0.49
	Heavily shaken for 4 days at RT	37.0	17%	7.2	6.9%	0.19
	Frozen	206	100%	134	100%	0.65
	Frozen - 3x at -20°C .	184	89%	132	99%	0.72
Advate 1:3	Frozen - 3x with dry ice	195	94%	128	96%	0.66

TABLE 2-continued

RVWF	VWF:Ag [U/ml]	VWF:Ag recovery [%]	VWF:RCo [U/ml]	VWF:RCo recovery [%]	RCo:Ag [U/U]
Lyo	195	94%	107	80%	0.55
Starting	206	100%	134	100%	0.65
material					
Heavily	138	67%	131	98%	0.95
shaken for					
4 days at					
RT					

[0119] Percent recovery was also measured in the shaking experiments with rVWF being stored in syringes with headspace and without headspace. Interestingly, when rVWF is stored in syringes without headspace and shaken as described above, no rVWF precipitation was observed. In contrast, when rVWF is stored in syringes with headspace, some precipitation was observed.

[0120] In summary, turbulent shaking resulted in at least 30% loss of rVWF in Advate buffer or Advate 1:3 buffer, with Advate buffer showing higher loss of recovery compared to Advate 1:3 buffer. Interestingly, the same precipitates observed in the turbulent shaking experiments were not observed when rVWF was stored and transported ~5000 km in an automobile (representing the expected shaking during transport). Precipitation of rVWF could be eliminated by storage in syringes without headspace.

Example 2

Stability of Recombinant VWF

[0121] The stability of rVWF was tested by assessing the activity level of rVWF present in a various formulations.

[0122] As shown in FIG. 1, rVWF is not stable in Advate buffer after 26 weeks due to the presence of 0.3 mM glutathione. As shown in FIG. 2, however, rVWF is more stable in Advate 1:3 buffer (e.g., for up 12 weeks at 4° C.)

[0123] As shown in FIG. 3, the stability of a citrate-based formulation (15 mM sodium citrate, 10 mM CaCl₂, 100 mM NaCl, pH 7.0) is better than Advate 1:3 buffer formulation containing 0.1M glutathione.

[0124] Likewise, the concentration of rVWF was measured over time in various buffers. As shown in FIG. 4, FIG. 5 and FIG. 6, rVWF concentration is stable over time in Advate buffer, Advate 1:3 buffer, and citrate-based buffer, respectively.

Example 4

Characterization of the Liquid Formulations

[0125] Differential scanning calorimetry (DSC) was used to assess the extent of protein (rVWF) unfolding in various buffers. As shown in Table 3, Advate buffer pH 7.0 is the optimum for stabilization.

[0126] DSC is a thermoanalytical technique in which the difference in the amount of heat required to increase the temperature of a sample and references are measured as a function of temperature. The result of a DSC experiment is a curve of heat flux versus temperature or versus time.

[0127] The Differential Scanning Calorimeter can scan through a range of temperatures while heating and cooling

and it determines a phase transition, i.e. melting, crystallization, or glass transition, by measuring the amount of heat needed to reach a set temperature. The calorimeter was calibrated with a set of pure metals (zinc, indium, and tin) that have a known heat capacity, Cp and melting point, Tm. The respective reference buffer was placed into the reference capillary and the rVWF sample was placed into the sample capillary of the instrument.

TABLE 3

Unfolding temperature in various buffers			
Lot	Buffer	pH	T unfold [° C]
rVWF161A	Advate	7.0	66.0
rVWF161B	Immunate	6.8	64.5
rVWF161C	Citrate	6.8	61.2
rVWF161D	NovoSeven	6.8	64.9
rVWF158	Hepes	7.4	61.3

Buffer components and concentrations:

A) Advate:	5.26 g/l NaCl 0.248 g/l CaCl ₂ 32 g/l D-Mannitol 8 g/l Trehalose 1.56 g/l L-Histidine 1.2 g/l Tris 0.08 g/l Glutadione red.	pH = 7.0
B) Immunate:	5.25 g/l Glycin 2.2 g/l NaCl 5.25 g/l NaCit3 5.25 g/l Lysin-HCl 0.62 g/l CaCl ₂	pH = 6.8
C) Citrat:	3 g/l Glycin 2.92 g/l NaCl 2.5 g/l NaCit3 30 g/l D-Mannitol 10 g/l Trehalose	pH = 6.8
D) NovoSeven:	0.75 g/l Glycin 2.92 g/l NaCl 1.47 g/l CaCl ₂ 30 g/l D-Mannitol	pH = 6.8

[0128] rVWF158: 20 mM Hepes, 150 mM NaCl, 5 g/L sucrose, pH 7.4

[0129] Further, as shown in FIG. 7, most formulation excipients increase the unfolding temperature by about 1-2° C. FIG. 8 shows that 10 mM CaCl₂ increases the unfolding temperature by ~8° C. to ~67° C., an unfolding temperature which can also be reached by Advate buffer. This effect of CaCl₂ is similar at pH 7.3 and 6.5, as shown in FIG. 9. Finally, the effect of trehalose and sucrose were analyzed on the

unfolding temperature. Compared to citrate alone, neither trehalose nor sucrose increased the unfolding temperature of rVWF. A summary of the unfolding temperature (T_{max}) data for rVWF in the presence of various excipients is set out in Table 4.

TABLE 4

15 mM Sodium Citrate buffer	—	15 mM Tris	15 mM Glycine	50 mM NaCl
ΔH [kJ/mol]	128494.3	656259.7	157352.2	124985.8
Unfolding T [° C.] - Peak 1	58.6		59.1	61
Peak 2	65.2	68.5	65.5	
Peak 3	80.4		80.1	81
Peak 4				
15 mM Sodium Citrate buffer	15 mM Histidine	20.52 g/L Mannitol	10.26 g/L Trehalose	
ΔH [kJ/mol]	134044.5	1588590.1	612235.9	
Unfolding T [° C.] - Peak 1	59.2	58.5	58.5	
Peak 2	65.2	65.5	71.3	
Peak 3	79.3	78.2	81.5	
Peak 4	88.5		92.7	
15 mM Sodium Citrate buffer	1 mM CaCl ₂	10 mM CaCl ₂	32 g/L Saccharose	0.25 mM Saccharose
ΔH [kJ/mol]	266008.2	308171.3	115082.4	246904.6
Unfolding T [° C.] - Peak 1	64.5	67.2	59.2	60
Peak 2			66	67
Peak 3	81	83.1	81.1	81.7
Peak 4	91.8	93		
15 mM Sodium Citrate buffer	0.1 g/L TWEEN-80	32 g/L Raf-fucose	Na ₂ HPO ₄ /NaHPO ₄	7.8 mM Trehalose
ΔH [kJ/mol]	338792.7	127329.2	197967.5	135573.3
Unfolding T [° C.] - Peak 1	58.7	60.1	61.4	58.4
Peak 2	64.4	65.8		65.4
Peak 3	81.6	80.3	80.4	80.4
Peak 4			89.2	

[0130] In addition to the various buffers, DSC was used to assess unfolding temperature of rVWF at various pH values in Advate buffer. The results are shown in Table 5, below.

Advate buffer pH 7.0 is the optimum for stabilization (i.e., highest unfolding temperature; Peak 1) of rVWF.

TABLE 5

pH	Peak 1	Peak 2
5.0	59.5	62.0
6.0	65.2	75.4
7.0	67.2	82.8
8.0	66.6	85.6
9.0	65.0	84.9

[0131] The fluorescence spectrum of rVWF in Advate buffer and Advate 1:3 buffer was assessed after storage at various temperatures for various lengths of time. No (or only slight) change in fluorescence spectrum was observed after storage at 40° C. from 0 to 28 days in either Advate or Advate 1:3 buffers. No difference was observed at other temperatures.

[0132] Likewise, degradation of rVWF was assessed using gelfiltration (Superose 6). While some degradation was observed after 26 weeks at 4° C. in Advate buffer, almost no degradation of rVWF in Advate 1:3 buffer was observed after 26 weeks at 4° C. At 40° C., glutathione increased the amount of degradation over time (albeit to a slower extent in Advate 1:3 buffer).

[0133] Based on the above Examples, Advate 1:3 buffer offers an advantage with respect to freeze/thawing and recovery after lyophilization as compared to the undiluted Advate buffer. Moreover, Advate 1:3 buffer can stabilize (e.g., maintain biological activity) rVWF activity during incubation at 40° C. better than Advate buffer. rVWF in Advate 1:3 buffer is stable for 4 weeks of incubation at 4° C. Finally, DSC has demonstrated that pH 7.0 is optimum for preventing degradation of rVWF (i.e., showed the highest unfolding temperature).

[0134] Thus, in view of the data presented herein, a formulation was proposed for rVWF including 15 mM citrate (or glycine or histidine), 10 mM CaCl₂, pH 6.5-7.3, adjusted to the desired osmolarity by NaCl. For example, in one embodiment, the citrate-based formula is 15 mM sodium citrate, 10 mM CaCl₂, 100 mM NaCl, pH 7.0.

[0135] Alternatively, an Advate or Advate 1:3 buffer, without glutathione, is also contemplated: Advate: 90 mM NaCl, 1.68 mM CaCl₂, 10 mM L-histidine, 10 mM Tris, 0.26 mM glutathione, 23.4 mM trehalose, 175.7 mM mannitol, and 0.1 g/L TWEEN-80, pH 7.0; Advate 1:3: 30 mM NaCl, 0.56 mM CaCl₂, 3.3 mM L-histidine, 3.3 mM tris, 7.8 mM trehalose, 58.6 mM mannitol, and 0.03 g/L TWEEN-80, pH 7.0.

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tgctgtgag gtggtgactg gctcaccgcy gggggactcc cagtcttctt ggaagagtgt 7800
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cagctgcatg ggtgcctgct gctgcctgcc ttggcctgat ggccaggcca gagtgtgcc 8760
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<210> SEQ ID NO 2

<211> LENGTH: 2783

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic Polypeptide
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: prepro-vWF

<400> SEQUENCE: 2

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Thr  Leu  Cys  Ala  Glu  Gly  Thr  Arg  Gly  Arg  Ser  Ser  Thr  Ala  Arg  Cys
20     25     30

Ser  Leu  Phe  Gly  Ser  Asp  Phe  Val  Asn  Thr  Phe  Asp  Gly  Ser  Met  Tyr
35     40     45

Ser  Phe  Ala  Gly  Tyr  Cys  Ser  Tyr  Leu  Leu  Ala  Gly  Gly  Cys  Gln  Lys
50     55     60

Arg  Ser  Phe  Ser  Ile  Ile  Gly  Asp  Phe  Gln  Asn  Gly  Lys  Arg  Val  Ser
65     70     75     80

Leu  Ser  Val  Tyr  Leu  Gly  Glu  Phe  Phe  Asp  Ile  His  Leu  Phe  Val  Asn
85     90     95

Gly  Thr  Val  Thr  Gln  Gly  Asp  Gln  Arg  Val  Ser  Met  Pro  Tyr  Ala  Ser
100    105    110

Lys  Leu  Glu  Thr  Glu  Ala  Gly  Tyr  Tyr  Lys  Leu  Ser  Gly  Glu  Ala  Tyr
115    120    125

Gly  Phe  Val  Ala  Arg  Ile  Asp  Gly  Ser  Gly  Asn  Phe  Gln  Val  Leu  Leu
130    135    140

Ser  Asp  Arg  Tyr  Phe  Asn  Lys  Thr  Cys  Gly  Leu  Cys  Gly  Asn  Phe  Asn
145    150    155    160

Ile  Phe  Ala  Glu  Asp  Asp  Phe  Met  Thr  Gln  Glu  Gly  Thr  Leu  Thr  Ser
165    170    175

Asp  Pro  Tyr  Asp  Phe  Ala  Asn  Ser  Trp  Ala  Leu  Ser  Ser  Gly  Glu  Gln
180    185    190

Trp  Cys  Glu  Arg  Pro  Ser  Ser  Ser  Cys  Asn  Ile  Ser  Ser  Gly  Glu  Met
195    200    205

Gln  Lys  Gly  Leu  Trp  Glu  Gln  Cys  Gln  Leu  Leu  Lys  Ser  Thr  Ser  Val
210    215    220

Phe  Ala  Arg  Cys  His  Pro  Leu  Val  Asp  Pro  Glu  Pro  Phe  Cys  Glu  Lys
225    230    235    240

Thr  Leu  Cys  Glu  Cys  Ala  Gly  Gly  Leu  Glu  Cys  Ala  Cys  Pro  Ala  Leu
245    250    255

Leu  Glu  Tyr  Ala  Arg  Thr  Cys  Ala  Gln  Glu  Gly  Met  Val  Leu  Tyr  Gly
260    265    270

Trp  Thr  Asp  His  Ser  Ala  Cys  Ser  Pro  Val  Cys  Pro  Ala  Gly  Met  Glu
275    280    285

Tyr  Arg  Gln  Cys  Val  Ser  Pro  Cys  Ala  Arg  Thr  Cys  Gln  Ser  Leu  His
290    295    300

Ile  Asn  Glu  Met  Cys  Gln  Glu  Arg  Cys  Val  Asp  Gly  Cys  Ser  Cys  Pro
305    310    315    320

Glu  Gly  Gln  Leu  Leu  Asp  Glu  Gly  Leu  Cys  Val  Glu  Ser  Thr  Glu  Cys
325    330    335

Pro  Cys  Val  His  Ser  Gly  Lys  Arg  Tyr  Pro  Pro  Gly  Thr  Ser  Leu  Ser
340    345    350

Arg  Asp  Cys  Asn  Thr  Cys  Ile  Cys  Arg  Asn  Ser  Gln  Trp  Ile  Cys  Ser
355    360    365

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Asn Glu Glu Cys Pro	Gly Glu Cys Leu Val	Thr Gly Gln Ser His Phe
370	375	380
Lys Ser Phe Asp Asn Arg Tyr Phe Thr Phe	Ser Gly Ile Cys Gln Tyr	
385	390	395 400
Leu Leu Ala Arg Asp Cys Gln Asp His Ser	Phe Ser Ile Val Ile Glu	
405	410	415
Thr Val Gln Cys Ala Asp Asp Arg Asp Ala	Val Cys Thr Arg Ser Val	
420	425	430
Thr Val Arg Leu Pro Gly Leu His Asn Ser	Leu Val Lys Leu Lys His	
435	440	445
Gly Ala Gly Val Ala Met Asp Gly Gln Asp	Val Gln Leu Pro Leu Leu	
450	455	460
Lys Gly Asp Leu Arg Ile Gln His Thr Val	Thr Ala Ser Val Arg Leu	
465	470	475 480
Ser Tyr Gly Glu Asp Leu Gln Met Asp Trp	Asp Gly Arg Gly Arg Leu	
485	490	495
Leu Val Lys Leu Ser Pro Val Tyr Ala Gly	Lys Thr Cys Gly Leu Cys	
500	505	510
Gly Asn Tyr Asn Gly Asn Gln Gly Asp Asp	Phe Leu Thr Pro Ser Gly	
515	520	525
Leu Ala Glu Pro Arg Val Glu Asp Phe Gly	Asn Ala Trp Lys Leu His	
530	535	540
Gly Asp Cys Gln Asp Leu Gln Lys Gln His	Ser Asp Pro Cys Ala Leu	
545	550	555 560
Asn Pro Arg Met Thr Arg Phe Ser Glu Glu	Ala Cys Ala Val Leu Thr	
565	570	575
Ser Pro Thr Phe Glu Ala Cys His Arg Ala	Val Ser Pro Leu Pro Tyr	
580	585	590
Leu Arg Asn Cys Arg Tyr Asp Val Cys Ser	Cys Ser Asp Gly Arg Glu	
595	600	605
Cys Leu Cys Gly Ser Tyr Ala Ala Ala Cys	Ala Gly Arg Gly Val Arg	
610	615	620
Val Ala Trp Arg Glu Pro Gly Arg Cys Glu	Leu Asn Cys Pro Lys Gly	
625	630	635 640
Gln Val Tyr Leu Gln Cys Gly Thr Pro Cys	Asn Leu Thr Cys Arg Ser	
645	650	655
Leu Ser Tyr Pro Asp Glu Glu Cys Asn Glu	Ala Cys Leu Glu Gly Cys	
660	665	670
Phe Cys Pro Pro Met Asp Glu Arg Gly Asp	Cys Val Pro Lys Ala Gln	
675	680	685
Cys Pro Cys Tyr Tyr Asp Gly Glu Ile Phe	Gln Pro Glu Asp Ile Phe	
690	695	700
Ser Asp His His Thr Met Cys Tyr Cys Glu	Asp Gly Phe Met His Cys	
705	710	715 720
Thr Met Ser Gly Val Pro Gly Ser Leu Leu	Pro Asp Ala Val Leu Ser	
725	730	735
Ser Pro Leu Ser His Arg Ser Lys Arg Ser	Leu Ser Cys Arg Pro Pro	
740	745	750
Met Val Lys Leu Val Cys Pro Ala Asp Asn	Leu Arg Ala Glu Gly Leu	
755	760	765

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Glu Cys Thr Lys Thr	Cys Gln Asn Tyr Asp	Leu Glu Cys Met Ser Met
770	775	780
Gly Cys Val Ser Gly	Cys Leu Cys Pro Pro	Gly Met Val Arg His Glu
785	790	795 800
Asn Arg Cys Glu Arg	Cys Pro Cys Phe His	Gln Gly Lys Glu Tyr Ala
805	810	815
Pro Gly Glu Thr Val	Lys Ile Gly Cys Asn Thr	Cys Val Cys Arg Asp
820	825	830
Arg Lys Trp Asn Cys	Thr Asp His Val Cys	Asp Ala Thr Cys Ser Thr
835	840	845
Ile Gly Met Ala His	Tyr Leu Thr Phe Asp	Gly Leu Lys Tyr Leu Phe
850	855	860
Pro Gly Glu Cys Gln	Tyr Val Leu Val Gln	Asp Tyr Cys Gly Ser Asn
865	870	875 880
Pro Gly Thr Phe Arg	Ile Leu Val Gly Asn	Lys Gly Cys Ser His Pro
885	890	895
Ser Val Lys Cys Lys	Lys Arg Val Thr Ile	Leu Val Glu Gly Gly Glu
900	905	910
Ile Glu Leu Phe Asp	Gly Glu Val Asn Val	Lys Arg Pro Met Lys Asp
915	920	925
Glu Thr His Phe Glu	Val Val Glu Ser Gly	Arg Tyr Ile Ile Leu Leu
930	935	940
Leu Gly Lys Ala Leu	Ser Val Val Trp Asp	Arg His Leu Ser Ile Ser
945	950	955 960
Val Val Leu Lys Gln	Thr Tyr Gln Glu Lys	Val Cys Gly Leu Cys Gly
965	970	975
Asn Phe Asp Gly Ile	Gln Asn Asn Asp Leu	Thr Ser Ser Asn Leu Gln
980	985	990
Val Glu Glu Asp Pro	Val Asp Phe Gly Asn	Ser Trp Lys Val Ser Ser
995	1000	1005
Gln Cys Ala Asp Thr	Arg Lys Val Pro	Leu Asp Ser Ser Pro Ala
1010	1015	1020
Thr Cys His Asn Asn	Ile Met Lys Gln	Thr Met Val Asp Ser Ser
1025	1030	1035
Cys Arg Ile Leu Thr	Ser Asp Val Phe	Gln Asp Cys Asn Lys Leu
1040	1045	1050
Val Asp Pro Glu Pro	Tyr Leu Asp Val	Cys Ile Tyr Asp Thr Cys
1055	1060	1065
Ser Cys Glu Ser Ile	Gly Asp Cys Ala	Cys Phe Cys Asp Thr Ile
1070	1075	1080
Ala Ala Tyr Ala His	Val Cys Ala Gln	His Gly Lys Val Val Thr
1085	1090	1095
Trp Arg Thr Ala Thr	Leu Cys Pro Gln	Ser Cys Glu Glu Arg Asn
1100	1105	1110
Leu Arg Glu Asn Gly	Tyr Glu Cys Glu	Trp Arg Tyr Asn Ser Cys
1115	1120	1125
Ala Pro Ala Cys Gln	Val Thr Cys Gln	His Pro Glu Pro Leu Ala
1130	1135	1140
Cys Pro Val Gln Cys	Val Glu Gly Cys	His Ala His Cys Pro Pro
1145	1150	1155
Gly Lys Ile Leu Asp	Glu Leu Leu Gln	Thr Cys Val Asp Pro Glu

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1160		1165		1170	
Asp Cys	Pro Val Cys	Glu Val	Ala Gly Arg	Arg Phe	Ala Ser Gly
1175		1180		1185	
Lys Lys	Val Thr Leu	Asn Pro	Ser Asp Pro	Glu His	Cys Gln Ile
1190		1195		1200	
Cys His	Cys Asp Val	Val Asn	Leu Thr Cys	Glu Ala	Cys Gln Glu
1205		1210		1215	
Pro Gly	Gly Leu Val	Val Pro	Pro Thr Asp	Ala Pro	Val Ser Pro
1220		1225		1230	
Thr Thr	Leu Tyr Val	Glu Asp	Ile Ser Glu	Pro Pro	Leu His Asp
1235		1240		1245	
Phe Tyr	Cys Ser Arg	Leu Leu	Asp Leu Val	Phe Leu	Leu Asp Gly
1250		1255		1260	
Ser Ser	Arg Leu Ser	Glu Ala	Glu Phe Glu	Val Leu	Lys Ala Phe
1265		1270		1275	
Val Val	Asp Met Met	Glu Arg	Leu Arg Ile	Ser Gln	Lys Trp Val
1280		1285		1290	
Arg Val	Ala Val Val	Glu Tyr	His Asp Gly	Ser His	Ala Tyr Ile
1295		1300		1305	
Gly Leu	Lys Asp Arg	Lys Arg	Pro Ser Glu	Leu Arg	Arg Ile Ala
1310		1315		1320	
Ser Gln	Val Lys Tyr	Ala Gly	Ser Gln Val	Ala Ser	Thr Ser Glu
1325		1330		1335	
Val Leu	Lys Tyr Thr	Leu Phe	Gln Ile Phe	Ser Lys	Ile Asp Arg
1340		1345		1350	
Pro Glu	Ala Ser Arg	Ile Thr	Leu Leu Leu	Met Ala	Ser Gln Glu
1355		1360		1365	
Pro Gln	Arg Met Ser	Arg Asn	Phe Val Arg	Tyr Val	Gln Gly Leu
1370		1375		1380	
Lys Lys	Lys Lys Val	Ile Val	Ile Pro Val	Gly Ile	Gly Pro His
1385		1390		1395	
Ala Asn	Leu Lys Gln	Ile Arg	Leu Ile Glu	Lys Gln	Ala Pro Glu
1400		1405		1410	
Asn Lys	Ala Phe Val	Leu Ser	Ser Val Asp	Glu Leu	Glu Gln Gln
1415		1420		1425	
Arg Asp	Glu Ile Val	Ser Tyr	Leu Cys Asp	Leu Ala	Pro Glu Ala
1430		1435		1440	
Pro Pro	Pro Thr Leu	Pro Pro	Asp Met Ala	Gln Val	Thr Val Gly
1445		1450		1455	
Pro Gly	Leu Leu Gly	Val Ser	Thr Leu Gly	Pro Lys	Arg Asn Ser
1460		1465		1470	
Met Val	Leu Asp Val	Ala Phe	Val Leu Glu	Gly Ser	Asp Lys Ile
1475		1480		1485	
Gly Glu	Ala Asp Phe	Asn Arg	Ser Lys Glu	Phe Met	Glu Glu Val
1490		1495		1500	
Ile Gln	Arg Met Asp	Val Gly	Gln Asp Ser	Ile His	Val Thr Val
1505		1510		1515	
Leu Gln	Tyr Ser Tyr	Met Val	Thr Val Glu	Tyr Pro	Phe Ser Glu
1520		1525		1530	
Ala Gln	Ser Lys Gly	Asp Ile	Leu Gln Arg	Val Arg	Glu Ile Arg
1535		1540		1545	

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Tyr Gln 1550	Gly Gly Asn 1555	Arg Thr 1555	Asn Thr Gly 1560	Leu Ala 1560	Leu Arg Tyr
Leu Ser 1565	Asp His Ser 1570	Phe Leu 1570	Val Ser Gln 1575	Gly Asp 1575	Arg Glu Gln
Ala Pro 1580	Asn Leu Val 1585	Tyr Met 1585	Val Thr Gly 1590	Asn Pro 1590	Ala Ser Asp
Glu Ile 1595	Lys Arg Leu 1600	Pro Gly 1600	Asp Ile Gln 1605	Val Val 1605	Pro Ile Gly
Val Gly 1610	Pro Asn Ala 1615	Asn Val 1615	Gln Glu Leu 1620	Glu Arg 1620	Ile Gly Trp
Pro Asn 1625	Ala Pro Ile 1630	Leu Ile 1630	Gln Asp Phe 1635	Glu Thr 1635	Leu Pro Arg
Glu Ala 1640	Pro Asp Leu 1645	Val Leu 1645	Gln Arg Cys 1650	Cys Ser 1650	Gly Glu Gly
Leu Gln 1655	Ile Pro Thr 1660	Leu Ser 1660	Pro Ala Pro 1665	Asp Cys 1665	Ser Gln Pro
Leu Asp 1670	Val Ile Leu 1675	Leu Leu 1675	Asp Gly Ser 1680	Ser Ser 1680	Phe Pro Ala
Ser Tyr 1685	Phe Asp Glu 1690	Met Lys 1690	Ser Phe Ala 1695	Lys Ala 1695	Phe Ile Ser
Lys Ala 1700	Asn Ile Gly 1705	Pro Arg 1705	Leu Thr Gln 1710	Val Ser 1710	Val Leu Gln
Tyr Gly 1715	Ser Ile Thr 1720	Thr Ile 1720	Asp Val Pro 1725	Trp Asn 1725	Val Val Pro
Glu Lys 1730	Ala His Leu 1735	Leu Ser 1735	Leu Val Asp 1740	Val Met 1740	Gln Arg Glu
Gly Gly 1745	Pro Ser Gln 1750	Ile Gly 1750	Asp Ala Leu 1755	Gly Phe 1755	Ala Val Arg
Tyr Leu 1760	Thr Ser Glu 1765	Met His 1765	Gly Ala Arg 1770	Pro Gly 1770	Ala Ser Lys
Ala Val 1775	Val Ile Leu 1780	Val Thr 1780	Asp Val Ser 1785	Val Asp 1785	Ser Val Asp
Ala Ala 1790	Ala Asp Ala 1795	Ala Arg 1795	Ser Asn Arg 1800	Val Thr 1800	Val Phe Pro
Ile Gly 1805	Ile Gly Asp 1810	Arg Tyr 1810	Asp Ala Ala 1815	Gln Leu 1815	Arg Ile Leu
Ala Gly 1820	Pro Ala Gly 1825	Asp Ser 1825	Asn Val Val 1830	Lys Leu 1830	Gln Arg Ile
Glu Asp 1835	Leu Pro Thr 1840	Met Val 1840	Thr Leu Gly 1845	Asn Ser 1845	Phe Leu His
Lys Leu 1850	Cys Ser Gly 1855	Phe Val 1855	Arg Ile Cys 1860	Met Asp 1860	Glu Asp Gly
Asn Glu 1865	Lys Arg Pro 1870	Gly Asp 1870	Val Trp Thr 1875	Leu Pro 1875	Asp Gln Cys
His Thr 1880	Val Thr Cys 1885	Gln Pro 1885	Asp Gly Gln 1890	Thr Leu 1890	Leu Lys Ser
His Arg 1895	Val Asn Cys 1900	Asp Arg 1900	Gly Leu Arg 1905	Pro Ser 1905	Cys Pro Asn
Ser Gln 1910	Ser Pro Val 1915	Lys Val 1915	Glu Glu Thr 1920	Cys Gly 1920	Cys Arg Trp

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Thr Cys 1925	Pro Cys Val	Cys Thr 1930	Gly Ser Ser	Thr Arg 1935	His Ile Val
Thr Phe 1940	Asp Gly Gln	Asn Phe 1945	Lys Leu Thr	Gly Ser 1950	Cys Ser Tyr
Val Leu 1955	Phe Gln Asn	Lys Glu 1960	Gln Asp Leu	Glu Val 1965	Ile Leu His
Asn Gly 1970	Ala Cys Ser	Pro Gly 1975	Ala Arg Gln	Gly Cys 1980	Met Lys Ser
Ile Glu 1985	Val Lys His	Ser Ala 1990	Leu Ser Val	Glu Leu 1995	His Ser Asp
Met Glu 2000	Val Thr Val	Asn Gly 2005	Arg Leu Val	Ser Val 2010	Pro Tyr Val
Gly Gly 2015	Asn Met Glu	Val Asn 2020	Val Tyr Gly	Ala Ile 2025	Met His Glu
Val Arg 2030	Phe Asn His	Leu Gly 2035	His Ile Phe	Thr Phe 2040	Thr Pro Gln
Asn Asn 2045	Glu Phe Gln	Leu Gln 2050	Leu Ser Pro	Lys Thr 2055	Phe Ala Ser
Lys Thr 2060	Tyr Gly Leu	Cys Gly 2065	Ile Cys Asp	Glu Asn 2070	Gly Ala Asn
Asp Phe 2075	Met Leu Arg	Asp Gly 2080	Thr Val Thr	Thr Asp 2085	Trp Lys Thr
Leu Val 2090	Gln Glu Trp	Thr Val 2095	Gln Arg Pro	Gly Gln 2100	Thr Cys Gln
Pro Glu 2105	Gln Cys Leu	Val Pro 2110	Asp Ser Ser	His Cys 2115	Gln Val Leu
Leu Leu 2120	Pro Leu Phe	Ala Glu 2125	Cys His Lys	Val Leu 2130	Ala Pro Ala
Thr Phe 2135	Tyr Ala Ile	Cys Gln 2140	Gln Asp Ser	Cys His 2145	Gln Glu Gln
Val Cys 2150	Glu Val Ile	Ala Ser 2155	Tyr Ala His	Leu Cys 2160	Arg Thr Asn
Gly Val 2165	Cys Val Asp	Trp Arg 2170	Thr Pro Asp	Phe Cys 2175	Ala Met Ser
Cys Pro 2180	Pro Ser Leu	Val Tyr 2185	Asn His Cys	Glu His 2190	Gly Cys Pro
Arg His 2195	Cys Asp Gly	Asn Val 2200	Ser Ser Cys	Gly Asp 2205	His Pro Ser
Glu Gly 2210	Cys Phe Cys	Pro Pro 2215	Asp Lys Val	Met Leu 2220	Glu Gly Ser
Cys Val 2225	Pro Glu Glu	Ala Cys 2230	Thr Gln Cys	Ile Gly 2235	Glu Asp Gly
Val Gln 2240	His Gln Phe	Leu Glu 2245	Ala Trp Val	Pro Asp 2250	His Gln Pro
Cys Gln 2255	Ile Cys Thr	Cys Leu 2260	Ser Gly Arg	Lys Val 2265	Asn Cys Thr
Thr Gln 2270	Pro Cys Pro	Thr Ala 2275	Lys Ala Pro	Thr Cys 2280	Gly Leu Cys
Glu Val 2285	Ala Arg Leu	Arg Gln 2290	Asn Ala Asp	Gln Cys 2295	Cys Pro Glu
Tyr Glu	Cys Val Cys	Asp Pro	Val Ser Cys	Asp Leu	Pro Pro Val

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2300		2305		2310	
Pro His	Cys Glu Arg	Gly Leu	Gln Pro Thr	Leu Thr	Asn Pro Gly
2315		2320		2325	
Glu Cys	Arg Pro Asn	Phe Thr	Cys Ala Cys	Arg Lys	Glu Glu Cys
2330		2335		2340	
Lys Arg	Val Ser Pro	Pro Ser	Cys Pro Pro	His Arg	Leu Pro Thr
2345		2350		2355	
Leu Arg	Lys Thr Gln	Cys Cys	Asp Glu Tyr	Glu Cys	Ala Cys Asn
2360		2365		2370	
Cys Val	Asn Ser Thr	Val Ser	Cys Pro Leu	Gly Tyr	Leu Ala Ser
2375		2380		2385	
Thr Ala	Thr Asn Asp	Cys Gly	Cys Thr Thr	Thr Thr	Cys Leu Pro
2390		2395		2400	
Asp Lys	Val Cys Val	His Arg	Ser Thr Ile	Tyr Pro	Val Gly Gln
2405		2410		2415	
Phe Trp	Glu Glu Gly	Cys Asp	Val Cys Thr	Cys Thr	Asp Met Glu
2420		2425		2430	
Asp Ala	Val Met Gly	Leu Arg	Val Ala Gln	Cys Ser	Gln Lys Pro
2435		2440		2445	
Cys Glu	Asp Ser Cys	Arg Ser	Gly Phe Thr	Tyr Val	Leu His Glu
2450		2455		2460	
Gly Glu	Cys Cys Gly	Arg Cys	Leu Pro Ser	Ala Cys	Glu Val Val
2465		2470		2475	
Thr Gly	Ser Pro Arg	Gly Asp	Ser Gln Ser	Ser Trp	Lys Ser Val
2480		2485		2490	
Gly Ser	Gln Trp Glu	Asn Pro	Cys Leu Ile	Asn Glu	Cys Val Arg
2495		2500		2505	
Val Lys	Glu Glu Val	Phe Ile	Gln Gln Arg	Asn Val	Ser Cys Pro
2510		2515		2520	
Gln Leu	Glu Val Pro	Val Cys	Pro Ser Gly	Phe Gln	Leu Ser Cys
2525		2530		2535	
Lys Thr	Ser Ala Cys	Cys Pro	Ser Cys Arg	Cys Glu	Arg Met Glu
2540		2545		2550	
Ala Cys	Met Leu Asn	Gly Thr	Val Ile Gly	Pro Gly	Lys Thr Val
2555		2560		2565	
Met Ile	Asp Val Cys	Thr Thr	Cys Arg Cys	Met Val	Gln Val Gly
2570		2575		2580	
Val Ile	Ser Gly Phe	Lys Leu	Glu Cys Arg	Lys Thr	Thr Cys Asn
2585		2590		2595	
Pro Cys	Pro Leu Gly	Tyr Lys	Glu Glu Asn	Asn Thr	Gly Glu Cys
2600		2605		2610	
Cys Gly	Arg Cys Leu	Pro Thr	Ala Cys Thr	Ile Gln	Leu Arg Gly
2615		2620		2625	
Gly Gln	Ile Met Thr	Leu Lys	Arg Asp Glu	Thr Leu	Gln Asp Gly
2630		2635		2640	
Cys Asp	Thr His Phe	Cys Lys	Val Asn Glu	Arg Gly	Glu Tyr Phe
2645		2650		2655	
Trp Glu	Lys Arg Val	Thr Gly	Cys Pro Pro	Phe Asp	Glu His Lys
2660		2665		2670	
Cys Leu	Ala Glu Gly	Gly Lys	Ile Met Lys	Ile Pro	Gly Thr Cys
2675		2680		2685	

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Cys Asp Thr Cys Glu Glu Pro Glu Cys Asn Asp Ile Thr Ala Arg
 2690 2695 2700
 Leu Gln Tyr Val Lys Val Gly Ser Cys Lys Ser Glu Val Glu Val
 2705 2710 2715
 Asp Ile His Tyr Cys Gln Gly Lys Cys Ala Ser Lys Ala Met Tyr
 2720 2725 2730
 Ser Ile Asp Ile Asn Asp Val Gln Asp Gln Cys Ser Cys Cys Ser
 2735 2740 2745
 Pro Thr Arg Thr Glu Pro Met Gln His Cys Thr Asn Gly Ser Val
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 Arg Lys Cys Ser Lys
 2780

<210> SEQ ID NO 3
 <211> LENGTH: 2050
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic Polypeptide
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: mature vWF

<400> SEQUENCE: 3

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 35 40 45
 Pro Gly Met Val Arg His Glu Asn Arg Cys Val Ala Leu Glu Arg Cys
 50 55 60
 Pro Cys Phe His Gln Gly Lys Glu Tyr Ala Pro Gly Glu Thr Val Lys
 65 70 75 80
 Ile Gly Cys Asn Thr Cys Val Cys Arg Asp Arg Lys Trp Asn Cys Thr
 85 90 95
 Asp His Val Cys Asp Ala Thr Cys Ser Thr Ile Gly Met Ala His Tyr
 100 105 110
 Leu Thr Phe Asp Gly Leu Lys Tyr Leu Phe Pro Gly Glu Cys Gln Tyr
 115 120 125
 Val Leu Val Gln Asp Tyr Cys Gly Ser Asn Pro Gly Thr Phe Arg Ile
 130 135 140
 Leu Val Gly Asn Lys Gly Cys Ser His Pro Ser Val Lys Cys Lys Lys
 145 150 155 160
 Arg Val Thr Ile Leu Val Glu Gly Gly Glu Ile Glu Leu Phe Asp Gly
 165 170 175
 Glu Val Asn Val Lys Arg Pro Met Lys Asp Glu Thr His Phe Glu Val
 180 185 190
 Val Glu Ser Gly Arg Tyr Ile Ile Leu Leu Leu Gly Lys Ala Leu Ser
 195 200 205
 Val Val Trp Asp Arg His Leu Ser Ile Ser Val Val Leu Lys Gln Thr
 210 215 220

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Tyr Gln Glu Lys Val	Cys Gly Leu Cys Gly	Asn Phe Asp Gly Ile Gln
225	230	235 240
Asn Asn Asp Leu Thr	Ser Ser Asn Leu Gln	Val Glu Glu Asp Pro Val
245	250	255
Asp Phe Gly Asn Ser	Trp Lys Val Ser Ser	Gln Cys Ala Asp Thr Arg
260	265	270
Lys Val Pro Leu Asp	Ser Ser Pro Ala Thr	Cys His Asn Asn Ile Met
275	280	285
Lys Gln Thr Met Val	Asp Ser Ser Cys Arg	Ile Leu Thr Ser Asp Val
290	295	300
Phe Gln Asp Cys Asn	Lys Leu Val Asp Pro	Glu Pro Tyr Leu Asp Val
305	310	315 320
Cys Ile Tyr Asp Thr	Cys Ser Cys Glu Ser	Ile Gly Asp Cys Ala Cys
325	330	335
Phe Cys Asp Thr Ile	Ala Ala Tyr Ala His	Val Cys Ala Gln His Gly
340	345	350
Lys Val Val Thr Trp	Arg Thr Ala Thr Leu	Cys Pro Gln Ser Cys Glu
355	360	365
Glu Arg Asn Leu Arg	Glu Asn Gly Tyr Glu	Cys Glu Trp Arg Tyr Asn
370	375	380
Ser Cys Ala Pro Ala	Cys Gln Val Thr Cys	Gln His Pro Glu Pro Leu
385	390	395 400
Ala Cys Pro Val Gln	Cys Val Glu Gly Cys	His Ala His Cys Pro Pro
405	410	415
Gly Lys Ile Leu Asp	Glu Leu Leu Gln Thr	Cys Val Asp Pro Glu Asp
420	425	430
Cys Pro Val Cys Glu	Val Ala Gly Arg Arg	Phe Ala Ser Gly Lys Lys
435	440	445
Val Thr Leu Asn Pro	Ser Asp Pro Glu His	Cys Gln Ile Cys His Cys
450	455	460
Asp Val Val Asn Leu	Thr Cys Glu Ala Cys	Gln Glu Pro Gly Gly Leu
465	470	475 480
Val Val Pro Pro Thr	Asp Ala Pro Val Ser	Pro Thr Thr Leu Tyr Val
485	490	495
Glu Asp Ile Ser Glu	Pro Pro Leu His Asp	Phe Tyr Cys Ser Arg Leu
500	505	510
Leu Asp Leu Val Phe	Leu Leu Asp Gly Ser	Ser Arg Leu Ser Glu Ala
515	520	525
Glu Phe Glu Val Leu	Lys Ala Phe Val Val	Asp Met Met Glu Arg Leu
530	535	540
Arg Ile Ser Gln Lys	Trp Val Arg Val Ala	Val Val Glu Tyr His Asp
545	550	555 560
Gly Ser His Ala Tyr	Ile Gly Leu Lys Asp	Arg Lys Arg Pro Ser Glu
565	570	575
Leu Arg Arg Ile Ala	Ser Gln Val Lys Tyr	Ala Gly Ser Gln Val Ala
580	585	590
Ser Thr Ser Glu Val	Leu Lys Tyr Thr Leu	Phe Gln Ile Phe Ser Lys
595	600	605
Ile Asp Arg Pro Glu	Ala Ser Arg Ile Thr	Leu Leu Leu Met Ala Ser
610	615	620

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Gln	Glu	Pro	Gln	Arg	Met	Ser	Arg	Asn	Phe	Val	Arg	Tyr	Val	Gln	Gly	625	630	635	640
Leu	Lys	Lys	Lys	Lys	Val	Ile	Val	Ile	Pro	Val	Gly	Ile	Gly	Pro	His	645	650	655	
Ala	Asn	Leu	Lys	Gln	Ile	Arg	Leu	Ile	Glu	Lys	Gln	Ala	Pro	Glu	Asn	660	665	670	
Lys	Ala	Phe	Val	Leu	Ser	Ser	Val	Asp	Glu	Leu	Glu	Gln	Gln	Arg	Asp	675	680	685	
Glu	Ile	Val	Ser	Tyr	Leu	Cys	Asp	Leu	Ala	Pro	Glu	Ala	Pro	Pro	Pro	690	695	700	
Thr	Leu	Pro	Pro	Asp	Met	Ala	Gln	Val	Thr	Val	Gly	Pro	Gly	Leu	Leu	705	710	715	720
Gly	Val	Ser	Thr	Leu	Gly	Pro	Lys	Arg	Asn	Ser	Met	Val	Leu	Asp	Val	725	730	735	
Ala	Phe	Val	Leu	Glu	Gly	Ser	Asp	Lys	Ile	Gly	Glu	Ala	Asp	Phe	Asn	740	745	750	
Arg	Ser	Lys	Glu	Phe	Met	Glu	Glu	Val	Ile	Gln	Arg	Met	Asp	Val	Gly	755	760	765	
Gln	Asp	Ser	Ile	His	Val	Thr	Val	Leu	Gln	Tyr	Ser	Tyr	Met	Val	Thr	770	775	780	
Val	Glu	Tyr	Pro	Phe	Ser	Glu	Ala	Gln	Ser	Lys	Gly	Asp	Ile	Leu	Gln	785	790	795	800
Arg	Val	Arg	Glu	Ile	Arg	Tyr	Gln	Gly	Gly	Asn	Arg	Thr	Asn	Thr	Gly	805	810	815	
Leu	Ala	Leu	Arg	Tyr	Leu	Ser	Asp	His	Ser	Phe	Leu	Val	Ser	Gln	Gly	820	825	830	
Asp	Arg	Glu	Gln	Ala	Pro	Asn	Leu	Val	Tyr	Met	Val	Thr	Gly	Asn	Pro	835	840	845	
Ala	Ser	Asp	Glu	Ile	Lys	Arg	Leu	Pro	Gly	Asp	Ile	Gln	Val	Val	Pro	850	855	860	
Ile	Gly	Val	Gly	Pro	Asn	Ala	Asn	Val	Gln	Glu	Leu	Glu	Arg	Ile	Gly	865	870	875	880
Trp	Pro	Asn	Ala	Pro	Ile	Leu	Ile	Gln	Asp	Phe	Glu	Thr	Leu	Pro	Arg	885	890	895	
Glu	Ala	Pro	Asp	Leu	Val	Leu	Gln	Arg	Cys	Cys	Ser	Gly	Glu	Gly	Leu	900	905	910	
Gln	Ile	Pro	Thr	Leu	Ser	Pro	Ala	Pro	Asp	Cys	Ser	Gln	Pro	Leu	Asp	915	920	925	
Val	Ile	Leu	Leu	Leu	Asp	Gly	Ser	Ser	Ser	Phe	Pro	Ala	Ser	Tyr	Phe	930	935	940	
Asp	Glu	Met	Lys	Ser	Phe	Ala	Lys	Ala	Phe	Ile	Ser	Lys	Ala	Asn	Ile	945	950	955	960
Gly	Pro	Arg	Leu	Thr	Gln	Val	Ser	Val	Leu	Gln	Tyr	Gly	Ser	Ile	Thr	965	970	975	
Thr	Ile	Asp	Val	Pro	Trp	Asn	Val	Val	Pro	Glu	Lys	Ala	His	Leu	Leu	980	985	990	
Ser	Leu	Val	Asp	Val	Met	Gln	Arg	Glu	Gly	Gly	Pro	Ser	Gln	Ile	Gly	995	1000	1005	
Asp	Ala	Leu	Gly	Phe	Ala	Val	Arg	Tyr	Leu	Thr	Ser	Glu	Met	His		1010	1015	1020	
Gly	Ala	Arg	Pro	Gly	Ala	Ser	Lys	Ala	Val	Val	Ile	Leu	Val	Thr					

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1025				1030				1035			
Asp Val	Ser Val	Asp	Ser Val	Asp Ala	Ala	Ala	Ala Asp	Ala Ala	Arg		
1040			1045				1050				
Ser Asn	Arg Val	Thr	Val Phe	Pro Ile	Gly	Ile Gly	Asp Arg	Tyr			
1055			1060			1065					
Asp Ala	Ala Gln	Leu	Arg Ile	Leu Ala	Gly	Pro Ala	Gly Asp	Ser			
1070			1075			1080					
Asn Val	Val Lys	Leu	Gln Arg	Ile Glu	Asp	Leu Pro	Thr Met	Val			
1085			1090			1095					
Thr Leu	Gly Asn	Ser	Phe Leu	His Lys	Leu	Cys Ser	Gly Phe	Val			
1100			1105			1110					
Arg Ile	Cys Met	Asp	Glu Asp	Gly Asn	Glu	Lys Arg	Pro Gly	Asp			
1115			1120			1125					
Val Trp	Thr Leu	Pro	Asp Gln	Cys His	Thr	Val Thr	Cys Gln	Pro			
1130			1135			1140					
Asp Gly	Gln Thr	Leu	Leu Lys	Ser His	Arg	Val Asn	Cys Asp	Arg			
1145			1150			1155					
Gly Leu	Arg Pro	Ser	Cys Pro	Asn Ser	Gln	Ser Pro	Val Lys	Val			
1160			1165			1170					
Glu Glu	Thr Cys	Gly	Cys Arg	Trp Thr	Cys	Pro Cys	Val Cys	Thr			
1175			1180			1185					
Gly Ser	Ser Thr	Arg	His Ile	Val Thr	Phe	Asp Gly	Gln Asn	Phe			
1190			1195			1200					
Lys Leu	Thr Gly	Ser	Cys Ser	Tyr Val	Leu	Phe Gln	Asn Lys	Glu			
1205			1210			1215					
Gln Asp	Leu Glu	Val	Ile Leu	His Asn	Gly	Ala Cys	Ser Pro	Gly			
1220			1225			1230					
Ala Arg	Gln Gly	Cys	Met Lys	Ser Ile	Glu	Val Lys	His Ser	Ala			
1235			1240			1245					
Leu Ser	Val Glu	Leu	His Ser	Asp Met	Glu	Val Thr	Val Asn	Gly			
1250			1255			1260					
Arg Leu	Val Ser	Val	Pro Tyr	Val Gly	Gly	Asn Met	Glu Val	Asn			
1265			1270			1275					
Val Tyr	Gly Ala	Ile	Met His	Glu Val	Arg	Phe Asn	His Leu	Gly			
1280			1285			1290					
His Ile	Phe Thr	Phe	Thr Pro	Gln Asn	Asn	Glu Phe	Gln Leu	Gln			
1295			1300			1305					
Leu Ser	Pro Lys	Thr	Phe Ala	Ser Lys	Thr	Tyr Gly	Leu Cys	Gly			
1310			1315			1320					
Ile Cys	Asp Glu	Asn	Gly Ala	Asn Asp	Phe	Met Leu	Arg Asp	Gly			
1325			1330			1335					
Thr Val	Thr Thr	Asp	Trp Lys	Thr Leu	Val	Gln Glu	Trp Thr	Val			
1340			1345			1350					
Gln Arg	Pro Gly	Gln	Thr Cys	Gln Pro	Ile	Leu Glu	Glu Gln	Cys			
1355			1360			1365					
Leu Val	Pro Asp	Ser	Ser His	Cys Gln	Val	Leu Leu	Leu Pro	Leu			
1370			1375			1380					
Phe Ala	Glu Cys	His	Lys Val	Leu Ala	Pro	Ala Thr	Phe Tyr	Ala			
1385			1390			1395					
Ile Cys	Gln Gln	Asp	Ser Cys	His Gln	Glu	Gln Val	Cys Glu	Val			
1400			1405			1410					

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Ile Ala 1415	Ser Tyr Ala 1420	His Leu 1420	Cys Arg Thr 1425	Asn Gly 1425	Val Cys Val
Asp Trp 1430	Arg Thr Pro 1435	Asp Phe 1435	Cys Ala Met 1440	Ser Cys 1440	Pro Pro Ser
Leu Val 1445	Tyr Asn His 1450	Cys Glu 1450	His Gly Cys 1455	Pro Arg 1455	His Cys Asp
Gly Asn 1460	Val Ser Ser 1465	Cys Gly 1465	Asp His Pro 1470	Ser Glu 1470	Gly Cys Phe
Cys Pro 1475	Pro Asp Lys 1480	Val Met 1480	Leu Glu Gly 1485	Ser Cys 1485	Val Pro Glu
Glu Ala 1490	Cys Thr Gln 1495	Cys Ile 1495	Gly Glu Asp 1500	Gly Val 1500	Gln His Gln
Phe Leu 1505	Glu Ala Trp 1510	Val Pro 1510	Asp His Gln 1515	Pro Cys 1515	Gln Ile Cys
Thr Cys 1520	Leu Ser Gly 1525	Arg Lys 1525	Val Asn Cys 1530	Thr Thr 1530	Gln Pro Cys
Pro Thr 1535	Ala Lys Ala 1540	Pro Thr 1540	Cys Gly Leu 1545	Cys Glu 1545	Val Ala Arg
Leu Arg 1550	Gln Asn Ala 1555	Asp Gln 1555	Cys Cys Pro 1560	Glu Tyr 1560	Glu Cys Val
Cys Asp 1565	Pro Val Ser 1570	Cys Asp 1570	Leu Pro Pro 1575	Val Pro 1575	His Cys Glu
Arg Gly 1580	Leu Gln Pro 1585	Thr Leu 1585	Thr Asn Pro 1590	Gly Glu 1590	Cys Arg Pro
Asn Phe 1595	Thr Cys Ala 1600	Cys Arg 1600	Lys Glu Glu 1605	Cys Lys 1605	Arg Val Ser
Pro Pro 1610	Ser Cys Pro 1615	Pro His 1615	Arg Leu Pro 1620	Thr Leu 1620	Arg Lys Thr
Gln Cys 1625	Cys Asp Glu 1630	Tyr Glu 1630	Cys Ala Cys 1635	Asn Cys 1635	Val Asn Ser
Thr Val 1640	Ser Cys Pro 1645	Leu Gly 1645	Tyr Leu Ala 1650	Ser Thr 1650	Ala Thr Asn
Asp Cys 1655	Gly Cys Thr 1660	Thr Thr 1660	Thr Cys Leu 1665	Pro Asp 1665	Lys Val Cys
Val His 1670	Arg Ser Thr 1675	Ile Tyr 1675	Pro Val Gly 1680	Gln Phe 1680	Trp Glu Glu
Gly Cys 1685	Asp Val Cys 1690	Thr Cys 1690	Thr Asp Met 1695	Glu Asp 1695	Ala Val Met
Gly Leu 1700	Arg Val Ala 1705	Gln Cys 1705	Ser Gln Lys 1710	Pro Cys 1710	Glu Asp Ser
Cys Arg 1715	Ser Gly Phe 1720	Thr Tyr 1720	Val Leu His 1725	Glu Gly 1725	Glu Cys Cys
Gly Arg 1730	Cys Leu Pro 1735	Ser Ala 1735	Cys Glu Val 1740	Val Thr 1740	Gly Ser Pro
Arg Gly 1745	Asp Ser Gln 1750	Ser Ser 1750	Trp Lys Ser 1755	Val Gly 1755	Ser Gln Trp
Ala Ser 1760	Pro Glu Asn 1765	Pro Cys 1765	Leu Ile Asn 1770	Glu Cys 1770	Val Arg Val
Lys Glu 1775	Glu Val Phe 1780	Ile Gln 1780	Gln Arg Asn 1785	Val Ser 1785	Cys Pro Gln

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Leu Glu 1790	Val Pro Val 1795	Cys Pro 1795	Ser Gly Phe 1800	Gln Leu 1800	Ser Cys Lys
Thr Ser 1805	Ala Cys Cys 1810	Pro Ser 1810	Cys Arg Cys 1815	Glu Arg 1815	Met Glu Ala
Cys Met 1820	Leu Asn Gly 1825	Thr Val 1825	Ile Gly Pro 1830	Gly Lys 1830	Thr Val Met
Ile Asp 1835	Val Cys Thr 1840	Thr Cys 1840	Arg Cys Met 1845	Val Gln 1845	Val Gly Val
Ile Ser 1850	Gly Phe Lys 1855	Leu Glu 1855	Cys Arg Lys 1860	Thr Thr 1860	Cys Asn Pro
Cys Pro 1865	Leu Gly Tyr 1870	Lys Glu 1870	Glu Asn Asn 1875	Thr Gly 1875	Glu Cys Cys
Gly Arg 1880	Cys Leu Pro 1885	Thr Ala 1885	Cys Thr Ile 1890	Gln Leu 1890	Arg Gly Gly
Gln Ile 1895	Met Thr Leu 1900	Lys Arg 1900	Asp Glu Thr 1905	Leu Gln 1905	Asp Gly Cys
Asp Thr 1910	His Phe Cys 1915	Lys Val 1915	Asn Glu Arg 1920	Gly Glu 1920	Tyr Phe Trp
Glu Lys 1925	Arg Val Thr 1930	Gly Cys 1930	Pro Pro Phe 1935	Asp Glu 1935	His Lys Cys
Leu Ala 1940	Glu Gly Gly 1945	Lys Ile 1945	Met Lys Ile 1950	Pro Gly 1950	Thr Cys Cys
Asp Thr 1955	Cys Glu Glu 1960	Pro Glu 1960	Cys Asn Asp 1965	Ile Thr 1965	Ala Arg Leu
Gln Tyr 1970	Val Lys Val 1975	Gly Ser 1975	Cys Lys Ser 1980	Glu Val 1980	Glu Val Asp
Ile His 1985	Tyr Cys Gln 1990	Gly Lys 1990	Cys Ala Ser 1995	Lys Ala 1995	Met Tyr Ser
Ile Asp 2000	Ile Asn Asp 2005	Val Gln 2005	Asp Gln Cys 2010	Ser Cys 2010	Cys Ser Pro
Thr Arg 2015	Thr Glu Pro 2020	Met Gln 2020	Val Ala Leu 2025	His Cys 2025	Thr Asn Gly
Ser Val 2030	Val Tyr His 2035	Glu Val 2035	Leu Asn Ala 2040	Met Glu 2040	Cys Lys Cys
Ser Pro 2045	Arg Lys Cys 2050	Ser Lys 2050			

What is claimed is:

1. A stable liquid pharmaceutical formulation of a recombinant von Willebrand Factor (rVWF) comprising: (a) a rVWF; (b) a buffering agent; (c) one or more salts; (d) optionally a stabilizing agent; and (e) optionally a surfactant;

wherein said rVWF comprises a polypeptide selected from the group consisting of:

- the amino acid sequence set out in SEQ ID NO: 3;
- a biologically active analog, fragment or variant of a);
- a polypeptide encoded by the polynucleotide set out in SEQ ID NO: 1;
- a biologically active analog, fragment or variant of c); and
- a polypeptide encoded by a polynucleotide that hybridizes to the polynucleotide set out in SEQ ID NO: 1 under moderately stringent hybridization conditions;

wherein said buffer is comprised of a pH buffering agent in a range of about 0.1 mM to about 500 mM and wherein the pH is in a range of about 2.0 to about 12.0; wherein said salt is at a concentration of about 1 to 500 mM;

wherein said stabilizing agent is at a concentration of about 0.1 to 1000 mM; and

wherein said surfactant is at a concentration of about 0.01 g/L to 0.5 g/L.

2. The formulation of claim 1 wherein the rVWF comprises the amino acid sequence set out in SEQ ID NO: 3.

3. The formulation of claim 1 wherein the buffering agent is selected from the group consisting of sodium citrate, glycine, histidine, Tris and combinations of these agents.

4. The formulation of claim 3 wherein the buffering agent is sodium citrate at a concentration of 15 mM.

5. The formulation of claim 1 wherein pH is in the range of 6.0-8.0.

6. The formulation of claim 5 wherein pH is in the range of 6.5-7.3.

7. The formulation of claim 4 wherein the pH is 7.0.

8. The formulation of claim 1 wherein the buffering agent is citrate and the pH is 7.0.

9. The formulation of claim 1 wherein the salt is selected from the group consisting of calcium chloride, sodium chloride and magnesium chloride.

10. The formulation of claim 9 wherein the salt is at a concentration range of 0.5 to 300 mM.

11. The formulation of claim 10 wherein the salt is calcium chloride at a concentration of 10 mM.

12. The formulation of claim 1 wherein the rVWF comprises the amino acid sequence set out in SEQ ID NO: 3; wherein the buffering agent is citrate and the pH is 7.0; and wherein the salt is calcium chloride at a concentration of 10 mM.

13. The formulation of claim 1 wherein the rVWF comprises the amino acid sequence set out in SEQ ID NO: 3; wherein the buffering agent is sodium citrate at a concentration of 15 mM and the pH is 7.0; and wherein the salt is calcium chloride at a concentration of 10 mM and NaCl at a concentration of 100 mM.

14. The formulation of claim 3 wherein the one or more buffering agents is histidine and Tris at a concentration of 3.3 mM each.

15. The formulation of claim 3 wherein the pH is 7.0.

16. The formulation of claim 9 wherein the one or more salts is sodium chloride at a concentration of 30 mM and calcium chloride at a concentration of 0.56 mM.

17. The formulation of claim 1 wherein the stabilizing agent is selected from the group consisting of mannitol, lactose, sorbitol, xylitol, sucrose, trehalose, mannose, maltose, lactose, glucose, raffinose, cellobiose, gentiobiose, isomaltose, arabinose, glucosamine, fructose and combinations of these stabilizing agents.

18. The formulation of claim 17 wherein the stabilizing agents are trehalose at a concentration of 7.8 mM and mannitol at a concentration of 58.6 mM.

19. The formulation of claim 1 wherein the surfactant is selected from the group consisting of digitonin, Triton X-100, Triton X-114, TWEEN-20, TWEEN-80 and combinations of these surfactants.

20. The formulation of claim 1 wherein the surfactant is TWEEN-80 at 0.03 g/L.

21. The formulation of claim 1 wherein the rVWF comprises amino acid sequence set out in SEQ ID NO: 3; wherein the buffering agents are histidine at a concentration of 3.3 mM and Tris at a concentration of 3.3 mM at pH 7.0; wherein the salts are sodium chloride at a concentration of 30 mM and calcium chloride at a concentration of 0.56 mM; wherein the stabilizing agents are trehalose at a concentration of 7.8 mM and mannitol at a concentration of 58.6 mM.; and wherein the surfactant is TWEEN-80 at 0.03 g/L.

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