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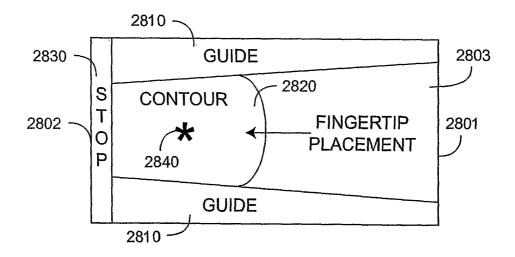
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(54) Title: MULTIPLE WAVELENGTH SENSOR ATTACHMENT



(57) Abstract: A physiological sensor is adapted to removably attach an emitter assembly and a detector assembly to a fingertip. The emitter assembly is adapted to transmit optical radiation having multiple wavelengths into fingertip tissue. The detector assembly is adapted to receive the optical radiation after attenuation by the fingertip tissue. The sensor has a first shell and a second shell hinged to the first shell. A spring is disposed between the shells and urges the shells together. An emitter pad is fixedly attached to the first shell and configured to retain the emitter assembly. A detector pad is fixedly attached to the second shell and configured to retain the detector assembly. A detector aperture is defined within the detector pad and adapted to pass optical radiation to the detector assembly. A contour is defined along the detector pad and generally shaped to conform to a fingertip positioned over the detector aperture.



## **MULTIPLE WAVELENGTH SENSOR ATTACHMENT**

### PRIORITY CLAIM TO RELATED PROVISIONAL APPLICATIONS

[0001] The present application claims priority benefit under 35 U.S.C. § 119(e) to U.S. Provisional Patent Application Serial No. 60/657,596, filed March 1, 2005, entitled "Multiple Wavelength Sensor," No. 60/657,281, filed March 1, 2005, entitled "Physiological Parameter Confidence Measure," No. 60/657,268, filed March 1, 2005, entitled "Configurable Physiological Measurement System," and No. 60/657,759, filed March 1, 2005, entitled "Noninvasive Multi-Parameter Patient Monitor." The present application incorporates the foregoing disclosures herein by reference.

# INCORPORATION BY REFERENCE OF COPENDING RELATED APPLICATIONS

[0002] The present application is related to the following copending U.S. utility applications:

App. Sr. No.	Filing Date	Title	Atty Dock.
11/###,###	March 1, 2006	Multiple Wavelength Sensor Emitters	MLR.002A
11/###,###	March 1, 2006	Multiple Wavelength Sensor Equalization	MLR.003A
11/###,###	March 1, 2006	Multiple Wavelength Sensor Substrate	MLR.004A
11/###,###	March 1, 2006	Multiple Wavelength Sensor Interconnect	MLR.005A
11/###,###	March 1, 2006	Multiple Wavelength Sensor Attachment	MLR.006A
11/###,###	March 1, 2006	Multiple Wavelength Sensor Drivers	MLR.009A
11/###,###	March 1, 2006	Physiological Parameter Confidence Measure	MLR.010A
11/###,###	March 1, 2006	Configurable Physiological Measurement System	MLR.011A
11/###,###	March 1, 2006	Noninvasive Multi- Parameter Patient Monitor	MLR.012A
11/###,###	March 1, 2006	Noninvasive Multi- Parameter Patient Monitor	MLR.013A
11/###,###	March 1, 2006	Noninvasive Multi- Parameter Patient Monitor	MLR.014A
	11/###,### 11/###,### 11/###,### 11/###,### 11/###,### 11/###,### 11/###,### 11/###,###	11/###,### March 1, 2006	11/###,### March 1, 2006 Multiple Wavelength Sensor Emitters  11/###,### March 1, 2006 Multiple Wavelength Sensor Equalization  11/###,### March 1, 2006 Multiple Wavelength Sensor Substrate  11/###,### March 1, 2006 Multiple Wavelength Sensor Interconnect  11/###,### March 1, 2006 Multiple Wavelength Sensor Attachment  11/###,### March 1, 2006 Multiple Wavelength Sensor Drivers  11/###,### March 1, 2006 Physiological Parameter Confidence Measure  11/###,### March 1, 2006 Configurable Physiological Measurement System  11/###,### March 1, 2006 Noninvasive Multi- Parameter Patient Monitor  11/###,### March 1, 2006 Noninvasive Multi- Parameter Patient Monitor  11/##################################

The present application incorporates the foregoing disclosures herein by reference.

#### **BACKGROUND OF THE INVENTION**

[0003] Spectroscopy is a common technique for measuring the concentration of organic and some inorganic constituents of a solution. The theoretical basis of this technique is the Beer-Lambert law, which states that the concentration  $c_i$  of an absorbent in solution can be determined by the intensity of light transmitted through the solution, knowing the pathlength  $d_\lambda$ , the intensity of the incident light  $I_{\varrho,\lambda}$ , and the extinction coefficient  $\varepsilon_{i,\lambda}$  at a particular wavelength  $\lambda$ . In generalized form, the Beer-Lambert law is expressed as:

$$I_{\lambda} = I_{0,\lambda} e^{-d_{\lambda} \cdot \mu_{a,\lambda}}$$
(1)
$$\mu_{a,\lambda} = \sum_{i=1}^{n} \varepsilon_{i,\lambda} \cdot c_{i}$$
(2)

where  $\mu_{a,\lambda}$  is the bulk absorption coefficient and represents the probability of absorption per unit length. The minimum number of discrete wavelengths that are required to solve EQS. **1-2** are the number of significant absorbers that are present in the solution.

[0004] A practical application of this technique is pulse oximetry, which utilizes a noninvasive sensor to measure oxygen saturation (SpO<sub>2</sub>) and pulse rate. In general, the sensor has light emitting diodes (LEDs) that transmit optical radiation of red and infrared wavelengths into a tissue site and a detector that responds to the intensity of the optical radiation after absorption (e.g., by transmission or transreflectance) by pulsatile arterial blood flowing within the tissue site. Based on this response, a processor determines measurements for SpO<sub>2</sub>, pulse rate, and can output representative plethysmographic waveforms. Thus, "pulse oximetry" as used herein encompasses its broad ordinary meaning known to one of skill in the art, which includes at least those noninvasive procedures for measuring parameters of circulating blood through spectroscopy. Moreover, "plethysmograph" as used herein (commonly referred to as "photoplethysmograph"), encompasses its broad ordinary meaning known to one of skill in the art, which includes at least data representative of a change in the absorption of particular wavelengths of light as a function of the changes in body

tissue resulting from pulsing blood. Pulse oximeters capable of reading through motion induced noise are available from Masimo Corporation ("Masimo") of Irvine, California. Moreover, portable and other oximeters capable of reading through motion induced noise are disclosed in at least U.S. Pat. Nos. 6,770,028, 6,658,276, 6,157,850, 6,002,952 5,769,785, and 5,758,644, which are owned by Masimo and are incorporated by reference herein. Such reading through motion oximeters have gained rapid acceptance in a wide variety of medical applications, including surgical wards, intensive care and neonatal units, general wards, home care, physical training, and virtually all types of monitoring scenarios.

## **SUMMARY OF THE INVENTION**

[0005] There is a need to noninvasively measure multiple physiological parameters, other than, or in addition to, oxygen saturation and pulse rate. For example, hemoglobin species that are also significant under certain circumstances are carboxyhemoglobin and methemoglobin. Other blood parameters that may be measured to provide important clinical information are fractional oxygen saturation, total hemaglobin (Hbt), bilirubin and blood glucose, to name a few.

[0006] One aspect of a physiological sensor that is adapted to removably attach an emitter assembly and a detector assembly to a fingertip, where the emitter assembly is adapted to transmit optical radiation having multiple wavelengths into fingertip tissue and the detector assembly is adapted to receive the optical radiation after attenuation by the fingertip tissue is a first shell and a second shell hinged to the first shell. A spring is disposed between the shells and urges the shells together. An emitter pad is fixedly attached to the first shell and configured to retain the emitter assembly. A detector pad is fixedly attached to the second shell and configured to retain the detector assembly. A detector aperture is defined within the detector pad and adapted to pass optical radiation to the detector assembly. A contour is defined along the detector pad and generally shaped to conform to a fingertip positioned over the detector aperture. [0007] Another aspect of a physiological sensor is a first shell, a second shell hinged to the first shell and a spring disposed between and urging together the

shells. The spring is configured so as to create a pivot point along a finger

gripped between the shells that is substantially behind a fingertip. A pad is configured to position the fingertip.

[0008] A further aspect of a physiological sensor provides housing elements comprising pads and shells. A sensor assembly is mounted within the housing elements. Pad features position a fingertip relative to the sensor assembly. The housing elements are urged against the fingertip so as to removably attach the sensor assembly to the fingertip.

#### BRIEF DESCRIPTION OF THE DRAWINGS

- [0009] FIG. 1 is a perspective view of a physiological measurement system utilizing a multiple wavelength sensor;
- [0010] FIGS. **2A-C** are perspective views of multiple wavelength sensor embodiments;
- [0011] FIG. 3 is a general block diagram of a multiple wavelength sensor and sensor controller;
- [0012] FIG. **4** is an exploded perspective view of a multiple wavelength sensor embodiment;
- [0013] FIG. 5 is a general block diagram of an emitter assembly;
- [0014] FIG. 6 is a perspective view of an emitter assembly embodiment:
- [0015] FIG. 7 is a general block diagram of an emitter array;
- [0016] FIG. 8 is a schematic diagram of an emitter array embodiment;
- [0017] FIG. 9 is a general block diagram of equalization;
- [0018] FIGS. 10A-D are block diagrams of various equalization embodiments;
- [0019] FIGS. 11A-C are perspective views of an emitter assembly incorporating various equalization embodiments;
- [0020] FIG. 12 is a general block diagram of an emitter substrate:
- [0021] FIGS. 13-14 are top and detailed side views of an emitter substrate embodiment;
- [0022] FIG. **15-16** are top and bottom component layout views of an emitter substrate embodiment;
- [0023] FIG. 17 is a schematic diagram of an emitter substrate embodiment;
- [0024] FIG. 18 is a plan view of an inner layer of an emitter substrate embodiment;

[0025] FIG. 19 is a general block diagram of an interconnect assembly in relationship to other sensor assemblies;

- [0026] FIG. 20 is a block diagram of an interconnect assembly embodiment;
- [0027] FIG. 21 is a partially-exploded perspective view of a flex circuit assembly embodiment of an interconnect assembly;
- [0028] FIG. 22 is a top plan view of a flex circuit;
- [0029] FIG. 23 is an exploded perspective view of an emitter portion of a flex circuit assembly;
- [0030] FIG. **24** is an exploded perspective view of a detector assembly embodiment;
- [0031] FIGS. **25-26** are block diagrams of adjacent detector and stacked detector embodiments;
- [0032] FIG. **27** is a block diagram of a finger clip embodiment of an attachment assembly;
- [0033] FIG. 28 is a general block diagram of a detector pad;
- [0034] FIGS. 29A-B are perspective views of detector pad embodiments;
- [0035] FIGS. **30A-H** are perspective bottom, perspective top, bottom, back, top, side cross sectional, side, and front cross sectional views of an emitter pad embodiment;
- [0036] FIGS. **31A-H** are perspective bottom, perspective top, top, back, bottom, side cross sectional, side, and front cross sectional views of a detector pad embodiment;
- [0037] FIGS. **32A-H** are perspective bottom, perspective top, top, back, bottom, side cross sectional, side, and front cross sectional views of a shoe box;
- [0038] FIGS. **33A-H** are perspective bottom, perspective top, top, back, bottom, side cross sectional, side, and front cross sectional views of a slim-finger emitter pad embodiment;
- [0039] FIGS. **34A-H** are perspective bottom, perspective top, top, back, bottom, side cross sectional, side, and front cross sectional views of a slim-finger detector pad embodiment;
- [0040] FIGS. **35A-B** are plan and cross sectional views, respectively, of a spring assembly embodiment;
- [0041] FIGS. 36A-C are top, perspective and side views of a finger clip spring;

[0042] FIGS. 37A-D are top, back, bottom, and side views of a spring plate;

[0043] FIGS. **38A-D** are front cross sectional, bottom, front and side cross sectional views of an emitter-pad shell;

[0044] FIGS. **39A-D** are back, top, front and side cross sectional views of a detector-pad shell;

[0045] FIG. 40 is a general block diagram of a monitor and a sensor;

[0046] FIGS. 41A-C are schematic diagrams of grid drive embodiments for a sensor having back-to-back diodes and an information element:

[0047] FIGS. **42** is a schematic diagrams of a grid drive embodiment for an information element;

[0048] FIGS. **43A-C** are schematic diagrams for grid drive readable information elements;

[0049] FIGS. 44A-B are cross sectional and side cut away views of a sensor cable;

[0050] FIG. 45 is a block diagram of a sensor controller embodiment; and

[0051] FIG. 46 is a detailed exploded perspective view of a multiple wavelength sensor embodiment.

# DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

#### **Overview**

[0052] In this application, reference is made to many blood parameters. Some references that have common shorthand designations are referenced through such shorthand designations. For example, as used herein, HbCO designates carboxyhemoglobin, HbMet designates methemoglobin, and Hbt designates total hemoglobin. Other shorthand designations such as COHb, MetHb, and tHb are also common in the art for these same constituents. These constituents are generally reported in terms of a percentage, often referred to as saturation, relative concentration or fractional saturation. Total hemoglobin is generally reported as a concentration in g/dL. The use of the particular shorthand designators presented in this application does not restrict the term to any particular manner in which the designated constituent is reported.

[0053] FIG. 1 illustrates a physiological measurement system 10 having a monitor 100 and a multiple wavelength sensor assembly 200 with enhanced

measurement capabilities as compared with conventional pulse oximetry. The physiological measurement system 10 allows the monitoring of a person, including a patient. In particular, the multiple wavelength sensor assembly 200 allows the measurement of blood constituent and related parameters in addition to oxygen saturation and pulse rate. Alternatively, the multiple wavelength sensor assembly 200 allows the measurement of oxygen saturation and pulse rate with increased accuracy or robustness as compared with conventional pulse oximetry.

[0054] In one embodiment, the sensor assembly 200 is configured to plug into a monitor sensor port 110. Monitor keys 160 provide control over operating modes and alarms, to name a few. A display 170 provides readouts of measured parameters, such as oxygen saturation, pulse rate, HbCO and HbMet to name a few.

[0055] FIGS. 2A illustrates a multiple wavelength sensor assembly 200 having a sensor 400 adapted to attach to a tissue site, a sensor cable 4400 and a monitor connector 210. In one embodiment, the sensor 400 is incorporated into a reusable finger clip adapted to removably attach to, and transmit light through, a fingertip. The sensor cable 4400 and monitor connector 210 are integral to the sensor 400, as shown. In alternative embodiments, the sensor 400 may be configured separately from the cable 4400 and connector 210.

[0056] FIGS. 2B-C illustrate alternative sensor embodiments, including a sensor 401 (FIG. 2B) partially disposable and partially reusable (resposable) and utilizing an adhesive attachment mechanism. Also shown is a sensor 402 (FIG. 2C) being disposable and utilizing an adhesive attachment mechanism. In other embodiments, a sensor may be configured to attach to various tissue sites other than a finger, such as a foot or an ear. Also a sensor may be configured as a reflectance or transflectance device that attaches to a forehead or other tissue surface.

[0057] FIG. 3 illustrates a sensor assembly 400 having an emitter assembly 500, a detector assembly 2400, an interconnect assembly 1900 and an attachment assembly 2700. The emitter assembly 500 responds to drive signals received from a sensor controller 4500 in the monitor 100 via the cable 4400 so as to transmit optical radiation having a plurality of wavelengths into a tissue site.

The detector assembly 2400 provides a sensor signal to the monitor 100 via the cable 4400 in response to optical radiation received after attenuation by the tissue site. The interconnect assembly 1900 provides electrical communication between the cable 4400 and both the emitter assembly 500 and the detector assembly 2400. The attachment assembly 2700 attaches the emitter assembly 500 and detector assembly 2400 to a tissue site, as described above. The emitter assembly 500 is described in further detail with respect to FIG. 5, below. The interconnect assembly 1900 is described in further detail with respect to FIG. 19, below. The detector assembly 2400 is described in further detail with respect to FIG. 24, below. The attachment assembly 2700 is described in further detail with respect to FIG. 27, below.

[0058] FIG. 4 illustrates a sensor 400 embodiment that removably attaches to a fingertip. The sensor 400 houses a multiple wavelength emitter assembly 500 and corresponding detector assembly 2400. A flex circuit assembly 1900 mounts the emitter and detector assemblies 500, 2400 and interconnects them to a multiwire sensor cable 4400. Advantageously, the sensor 400 is configured in several respects for both wearer comfort and parameter measurement performance. The flex circuit assembly 1900 is configured to mechanically decouple the cable 4400 wires from the emitter and detector assemblies 500, 2400 to reduce pad stiffness and wearer discomfort. The pads 3000, 3100 are mechanically decoupled from shells 3800, 3900 to increase flexibility and wearer comfort. A spring 3600 is configured in hinged shells 3800, 3900 so that the pivot point of the finger clip is well behind the fingertip, improving finger attachment and more evenly distributing the clip pressure along the finger.

[0059] As shown in FIG. 4, the detector pad 3100 is structured to properly position a fingertip in relationship to the detector assembly 2400. The pads have flaps that block ambient light. The detector assembly 2400 is housed in an enclosure so as to reduce light piping from the emitter assembly to the detector assembly without passing through fingertip tissue. These and other features are described in detail below. Specifically, emitter assembly embodiments are described with respect to FIGS. 5-18. Interconnect assembly embodiments, including the flexible circuit assembly 1900, are described with respect to FIGS. 19-23. Detector assembly embodiments are described with respect to FIGS. 24-

26. Attachment assembly embodiments are described with respect to FIGS. 27-39.

#### **Emitter Assembly**

FIG. 5 illustrates an emitter assembly 500 having an emitter array 700, [0060] a substrate 1200 and equalization 900. The emitter array 700 has multiple light emitting sources, each activated by addressing at least one row and at least one column of an electrical grid. The light emitting sources are capable of transmitting optical radiation having multiple wavelengths. The equalization 900 accounts for differences in tissue attenuation of the optical radiation across the multiple wavelengths so as to at least reduce wavelength-dependent variations in detected intensity. The substrate 1200 provides a physical mount for the emitter array and emitter-related equalization and a connection between the emitter array and the interconnection assembly. Advantageously, the substrate 1200 also provides a bulk temperature measurement so as to calculate the operating wavelengths for the light emitting sources. The emitter array 700 is described in further detail with respect to FIG. 7, below. Equalization is described in further detail with respect to FIG. 9, below. The substrate 1200 is described in further detail with respect to FIG. 12, below.

[0061] FIG. 6 illustrates an emitter assembly 500 embodiment having an emitter array 700, an encapsulant 600, an optical filter 1100 and a substrate 1200. Various aspects of the emitter assembly 500 are described with respect to FIGS. 7-18, below. The emitter array 700 emits optical radiation having multiple wavelengths of predetermined nominal values, advantageously allowing multiple parameter measurements. In particular, the emitter array 700 has multiple light emitting diodes (LEDs) 710 that are physically arranged and electrically connected in an electrical grid to facilitate drive control, equalization, and minimization of optical pathlength differences at particular wavelengths. The optical filter 1100 is advantageously configured to provide intensity equalization across a specific LED subset. The substrate 1200 is configured to provide a bulk temperature of the emitter array 700 so as to better determine LED operating wavelengths.

#### **Emitter Array**

[0062] FIG. 7 illustrates an emitter array 700 having multiple light emitters (LE) 710 capable of emitting light 702 having multiple wavelengths into a tissue site 1. Row drivers 4530 and column drivers 4560 are electrically connected to the light emitters 710 and activate one or more light emitters 710 by addressing at least one row 720 and at least one column 740 of an electrical grid. In one embodiment, the light emitters 710 each include a first contact 712 and a second contact 714. The first contact 712 of a first subset 730 of light emitters is in communication with a first conductor 720 of the electrical grid. The second contact 714 of a second subset 750 of light emitters is in communication with a second conductor 740. Each subset comprises at least two light emitters, and at least one of the light emitters of the first and second subsets 730, 750 are not in common. A detector 2400 is capable of detecting the emitted light 702 and outputting a sensor signal 2500 responsive to the emitted light 702 after attenuation by the tissue site 1. As such, the sensor signal 2500 is indicative of at least one physiological parameter corresponding to the tissue site 1, as described above.

[0063] FIG. 8 illustrates an emitter array 700 having LEDs 801 connected within an electrical grid of n rows and m columns totaling n + m drive lines 4501, 4502, where n and m integers greater than one. The electrical grid advantageously minimizes the number of drive lines required to activate the LEDs 801 while preserving flexibility to selectively activate individual LEDs 801 in any sequence and multiple LEDs 801 simultaneously. The electrical grid also facilitates setting LED currents so as to control intensity at each wavelength, determining operating wavelengths and monitoring total grid current so as to limit power dissipation. The emitter array 700 is also physically configured in rows 810. This physical organization facilitates clustering LEDs 801 according to wavelength so as to minimize pathlength variations and facilitates equalization of LED intensities.

[0064] As shown in FIG. 8, one embodiment of an emitter array 700 comprises up to sixteen LEDs 801 configured in an electrical grid of four rows 810 and four columns 820. Each of the four row drive lines 4501 provide a common anode connection to four LEDs 801, and each of the four column drive lines 4502 provide a common cathode connection to four LEDs 801. Thus, the

sixteen LEDs **801** are advantageously driven with only eight wires, including four anode drive lines **812** and four cathode drive lines **822**. This compares favorably to conventional common anode or cathode LED configurations, which require more drive lines. In a particular embodiment, the emitter array **700** is partially populated with eight LEDs having nominal wavelengths as shown in TABLE **1**. Further, LEDs having wavelengths in the range of 610-630 nm are grouped together in the same row. The emitter array **700** is adapted to a physiological measurement system **10** (FIG. **1**) for measuring  $H_bCO$  and/or METHb in addition to  $S_pO_2$  and pulse rate.

LED	λ	Row	Col
D1	630	1	1
D2	620	1	2
D3	610	1	3
D4		1	4
D5	700	2	1
D6	730	2	2
D7	660	2	3
D8	805	2	4
D9		3	1
D10		3	2
D11		3	3
D12	905	3	4
D13		4	1
D14		4	2
D15		4	3
D16		4	4

TABLE 1: Nominal LED Wavelengths

[0065] Also shown in FIG. 8, row drivers 4530 and column drivers 4560 located in the monitor 100 selectively activate the LEDs 801. In particular, row and column drivers 4530, 4560 function together as switches to Vcc and current sinks, respectively, to activate LEDs and as switches to ground and Vcc, respectively, to deactivate LEDs. This push-pull drive configuration advantageously prevents parasitic current flow in deactivated LEDs. In a particular embodiment, only one row drive line 4501 is switched to Vcc at a time. One to four column drive lines 4502, however, can be simultaneously switched to a current sink so as to simultaneously activate multiple LEDs within a particular

row. Activation of two or more LEDs of the same wavelength facilitates intensity equalization, as described with respect to FIGS. 9-11, below. LED drivers are described in further detail with respect to FIG. 45, below.

[0066] Although an emitter assembly is described above with respect to an array of light emitters each configured to transmit optical radiation centered around a nominal wavelength, in another embodiment, an emitter assembly advantageously utilizes one or more tunable broadband light sources, including the use of filters to select the wavelength, so as to minimize wavelength-dependent pathlength differences from emitter to detector. In yet another emitter assembly embodiment, optical radiation from multiple emitters each configured to transmit optical radiation centered around a nominal wavelength is funneled to a tissue site point so as to minimize wavelength-dependent pathlength differences. This funneling may be accomplish with fiberoptics or mirrors, for example. In further embodiments, the LEDs 801 can be configured with alternative orientations with correspondingly different drivers among various other configurations of LEDs, drivers and interconnecting conductors.

#### Equalization

[0067] FIG. 9 illustrate a physiological parameter measurement system 10 having a controller 4500, an emitter assembly 500, a detector assembly 2400 and a front-end 4030. The emitter assembly 500 is configured to transmit optical radiation having multiple wavelengths into the tissue site 1. The detector assembly 2400 is configured to generate a sensor signal 2500 responsive to the optical radiation after tissue attenuation. The front-end 4030 conditions the sensor signal 2500 prior to analog-to-digital conversion (ADC).

[0068] FIG. 9 also generally illustrates equalization 900 in a physiological measurement system 10 operating on a tissue site 1. Equalization encompasses features incorporated into the system 10 in order to provide a sensor signal 2500 that falls well within the dynamic range of the ADC across the entire spectrum of emitter wavelengths. In particular, equalization compensates for the imbalance in tissue light absorption due to Hb and HbO<sub>2</sub> 910. Specifically, these blood constituents attenuate red wavelengths greater than IR wavelengths. Ideally, equalization 900 balances this unequal attenuation. Equalization 900 can be

introduced anywhere in the system **10** from the controller **4500** to front-end **4000** and can include compensatory attenuation versus wavelength, as shown, or compensatory amplification versus or both.

[0069] Equalization can be achieved to a limited extent by adjusting drive currents from the controller **4500** and front-end **4030** amplification accordingly to wavelength so as to compensate for tissue absorption characteristics. Signal demodulation constraints, however, limit the magnitude of these adjustments. Advantageously, equalization **900** is also provided along the optical path from emitters **500** to detector **2400**. Equalization embodiments are described in further detail with respect to FIGS. **10-11**, below.

[0070] FIGS. 10A-D illustrate various equalization embodiments having an emitter array 700 adapted to transmit optical radiation into a tissue site 1 and a detector assembly 2400 adapted to generate a sensor signal 2500 responsive to the optical radiation after tissue attenuation. FIG. 10A illustrates an optical filter 1100 that attenuates at least a portion of the optical radiation before it is transmitted into a tissue site 1. In particular, the optical filter 1100 attenuates at least a portion of the IR wavelength spectrum of the optical radiation so as to approximate an equalization curve 900 (FIG. 9). FIG. 10B illustrates an optical filter 1100 that attenuates at least a portion of the optical radiation after it is attenuated by a tissue site 1, where the optical filter 1100 approximates an equalization curve 900 (FIG. 9).

[0071] FIG. 10C illustrates an emitter array 700 where at least a portion of the emitter array generates one or more wavelengths from multiple light emitters 710 of the same wavelength. In particular, the same-wavelength light emitters 710 boost at least a portion of the red wavelength spectrum so as to approximately equalize the attenuation curves 910 (FIG. 9). FIG. 10D illustrates a detector assembly 2400 having multiple detectors 2610, 2620 selected so as to equalize the attenuation curves 910 (FIG. 9). To a limited extent, optical equalization can also be achieved by selection of particular emitter array 700 and detector 2400 components, e.g. LEDs having higher output intensities or detectors having higher sensitivities at red wavelengths. Although equalization embodiments are described above with respect to red and IR wavelengths, these equalization

embodiments can be applied to equalize tissue characteristics across any portion of the optical spectrum.

FIGS. 11A-C illustrates an optical filter 1100 for an emitter assembly [0072] **500** that advantageously provides optical equalization, as described above. LEDs within the emitter array 700 may be grouped according to output intensity or wavelength or both. Such a grouping facilitates equalization of LED intensity across the array. In particular, relatively low tissue absorption and/or relatively high output intensity LEDs can be grouped together under a relatively high attenuation optical filter. Likewise, relatively low tissue absorption and/or relatively low output intensity LEDs can be grouped together without an optical filter or under a relatively low or negligible attenuation optical filter. Further, high tissue absorption and/or low intensity LEDs can be grouped within the same row with one or more LEDs of the same wavelength being simultaneously activated, as described with respect to FIG. 10C, above. In general, there can be any number of LED groups and any number of LEDs within a group. There can also be any number of optical filters corresponding to the groups having a range of attenuation, including no optical filter and/or a "clear" filter having negligible attenuation.

As shown in FIGS. 11A-C, a filtering media may be advantageously [0073] added to an encapsulant that functions both as a cover to protect LEDs and bonding wires and as an optical filter 1100. In one embodiment, a filtering media 1100 encapsulates a select group of LEDs and a clear media 600 (FIG. 6) encapsulates the entire array 700 and the filtering media 1000 (FIG. 6). In a particular embodiment, corresponding to TABLE 1, above, five LEDs nominally emitting at 660-905 nm are encapsulated with both a filtering media 1100 and an overlying clear media 600 (FIG. 6), i.e. attenuated. In a particular embodiment, the filtering media 1100 is a 40:1 mixture of a clear encapsulant (EPO-TEK OG147-7) and an opaque encapsulate (EPO-TEK OG147) both available from Epoxy Technology, Inc., Billerica, MA. Three LEDs nominally emitting at 610-630 nm are only encapsulated with the clear media 600 (FIG. 6), i.e. unattenuated. In alternative embodiments, individual LEDs may be singly or multiply encapsulated according to tissue absorption and/or output intensity. In other alternative embodiments, filtering media may be separately attachable optical

filters or a combination of encapsulants and separately attachable optical filters. In a particular embodiment, the emitter assembly **500** has one or more notches along each side proximate the component end **1305** (FIG. **13**) for retaining one or more clip-on optical filters.

#### **Substrate**

[0074] FIG. 12 illustrates light emitters 710 configured to transmit optical radiation 1201 having multiple wavelengths in response to corresponding drive currents 1210. A thermal mass 1220 is disposed proximate the emitters 710 so as to stabilize a bulk temperature 1202 for the emitters. A temperature sensor 1230 is thermally coupled to the thermal mass 1220, wherein the temperature sensor 1230 provides a temperature sensor output 1232 responsive to the bulk temperature 1202 so that the wavelengths are determinable as a function of the drive currents 1210 and the bulk temperature 1202.

[0075] In one embodiment, an operating wavelength  $\lambda_a$  of each light emitter 710 is determined according to EQ. 3

$$\lambda_a = f(T_b, I_{drive}, \sum I_{drive})$$
3)

where  $T_b$  is the bulk temperature,  $I_{drive}$  is the drive current for a particular light emitter, as determined by the sensor controller **4500** (FIG. **45)**, described below, and  $\sum I_{drive}$  is the total drive current for all light emitters. In another embodiment, temperature sensors are configured to measure the temperature of each light emitter **710** and an operating wavelength  $\lambda_a$  of each light emitter **710** is determined according to EQ. **4** 

$$\lambda_a = f(T_a, I_{drive}, \sum I_{drive})$$
(4)

where  $T_a$  is the temperature of a particular light emitter,  $I_{drive}$  is the drive current for that light emitter and  $\sum I_{drive}$  is the total drive current for all light emitters.

[0076] In yet another embodiment, an operating wavelength for each light emitter is determined by measuring the junction voltage for each light emitter 710. In a further embodiment, the temperature of each light emitter 710 is

controlled, such as by one or more Peltier cells coupled to each light emitter **710**, and an operating wavelength for each light emitter **710** is determined as a function of the resulting controlled temperature or temperatures. In other embodiments, the operating wavelength for each light emitter **710** is determined directly, for example by attaching a charge coupled device (CCD) to each light emitter or by attaching a fiberoptic to each light emitter and coupling the fiberoptics to a wavelength measuring device, to name a few.

[0077] FIGS. 13-18 illustrate one embodiment of a substrate 1200 configured to provide thermal conductivity between an emitter array 700 (FIG. 8) and a thermistor 1540 (FIG. 16). In this manner, the resistance of the thermistor 1540 (FIG. 16) can be measured in order to determine the bulk temperature of LEDs 801 (FIG. 8) mounted on the substrate 1200. The substrate 1200 is also configured with a relatively significant thermal mass, which stabilizes and normalizes the bulk temperature so that the thermistor measurement of bulk temperature is meaningful.

[0078] FIGS. 13-14 illustrate a substrate 1200 having a component side 1301, a solder side 1302, a component end 1305 and a connector end 1306.

Alignment notches 1310 are disposed between the ends 1305, 1306. The substrate 1200 further has a component layer 1401, inner layers 1402-1405 and a solder layer 1406. The inner layers 1402-1405, e.g. inner layer 1402 (FIG. 18), have substantial metallized areas 1411 that provide a thermal mass 1220 (FIG. 12) to stabilize a bulk temperature for the emitter array 700 (FIG. 12). The metallized areas 1411 also function to interconnect component pads 1510 and wire bond pads 1520 (FIG. 15) to the connector 1530.

[0079] FIGS. 15-16 illustrate a substrate 1200 having component pads 1510 and wire bond pads 1520 at a component end 1305. The component pads 1510 mount and electrically connect a first side (anode or cathode) of the LEDs 801 (FIG. 8) to the substrate 1200. Wire bond pads 1520 electrically connect a second side (cathode or anode) of the LEDs 801 (FIG. 8) to the substrate 1200. The connector end 1306 has a connector 1530 with connector pads 1532, 1534 that mount and electrically connect the emitter assembly 500 (FIG. 23), including the substrate 1200, to the flex circuit 2200 (FIG. 22). Substrate layers 1401-1406 (FIG. 14) have traces that electrically connect the component pads 1510 and

wire bond pads **1520** to the connector **1532-1534**. A thermistor **1540** is mounted to thermistor pads **1550** at the component end **1305**, which are also electrically connected with traces to the connector **1530**. Plated thru holes electrically connect the connector pads **1532**, **1534** on the component and solder sides **1301**, **1302**, respectively.

[0080] FIG. 17 illustrates the electrical layout of a substrate 1200. A portion of the LEDs 801, including D1-D4 and D13-D16 have cathodes physically and electrically connected to component pads 1510 (FIG. 15) and corresponding anodes wire bonded to wire bond pads 1520. Another portion of the LEDs 801, including D5-D8 and D9-D12, have anodes physically and electrically connected to component pads 1510 (FIG. 15) and corresponding cathodes wire bonded to wire bond pads 1520. The connector 1530 has row pinouts J21-J24, column pinouts J31-J34 and thermistor pinouts J40-J41 for the LEDs 801 and thermistor 1540.

#### **Interconnect Assembly**

[0081] FIG. 19 illustrates an interconnect assembly 1900 that mounts the emitter assembly 500 and detector assembly 2400, connects to the sensor cable 4400 and provides electrical communications between the cable and each of the emitter assembly 500 and detector assembly 2400. In one embodiment, the interconnect assembly 1900 is incorporated with the attachment assembly 2700, which holds the emitter and detector assemblies to a tissue site. An interconnect assembly embodiment utilizing a flexible (flex) circuit is described with respect to FIGS. 20-24, below.

[0082] FIG. 20 illustrates an interconnect assembly 1900 embodiment having a circuit substrate 2200, an emitter mount 2210, a detector mount 2220 and a cable connector 2230. The emitter mount 2210, detector mount 2220 and cable connector 2230 are disposed on the circuit substrate 2200. The emitter mount 2210 is adapted to mount an emitter assembly 500 having multiple emitters. The detector mount 2220 is adapted to mount a detector assembly 2400 having a detector. The cable connector 2230 is adapted to attach a sensor cable 4400. A first plurality of conductors 2040 disposed on the circuit substrate 2200 electrically interconnects the emitter mount 2210 and the cable connector 2230.

A second plurality of conductors 2050 disposed on the circuit substrate 2200 electrically interconnects the detector mount 2220 and the cable connector 2230. A decoupling 2060 disposed proximate the cable connector 2230 substantially mechanically isolates the cable connector 2230 from both the emitter mount 2210 and the detector mount 2220 so that sensor cable stiffness is not translated to the emitter assembly 500 or the detector assembly 2400. A shield 2070 is adapted to fold over and shield one or more wires or pairs of wires of the sensor cable 4400.

[0083] FIG. 21 illustrates a flex circuit assembly 1900 having a flex circuit 2200, an emitter assembly 500 and a detector assembly 2400, which is configured to terminate the sensor end of a sensor cable 4400. The flex circuit assembly 1900 advantageously provides a structure that electrically connects yet mechanically isolates the sensor cable 4400, the emitter assembly 500 and the detector assembly 2400. As a result, the mechanical stiffness of the sensor cable 4400 is not translated to the sensor pads 3000, 3100 (FIGS. 30-31), allowing a comfortable finger attachment for the sensor 200 (FIG. 1). In particular, the emitter assembly 500 and detector assembly 2400 are mounted to opposite ends 2201, 2202 (FIG. 22) of an elongated flex circuit 2200. The sensor cable 4400 is mounted to a cable connector 2230 extending from a middle portion of the flex circuit 2200. Detector wires 4470 are shielded at the flex circuit junction by a fold-over conductive ink flap 2240, which is connected to a cable inner shield 4450. The flex circuit 2200 is described in further detail with respect to FIG. 22. The emitter portion of the flex circuit assembly 1900 is described in further detail with respect to FIG. 23. The detector assembly 2400 is described with respect to FIG. 24. The sensor cable 4400 is described with respect to FIGS. 44A-B, below.

[0084] FIG. 22 illustrates a sensor flex circuit 2200 having an emitter end 2201, a detector end 2202, an elongated interconnect 2204, 2206 between the ends 2201, 2202 and a cable connector 2230 extending from the interconnect 2204, 2206. The emitter end 2201 forms a "head" having emitter solder pads 2210 for attaching the emitter assembly 500 (FIG. 6) and mounting ears 2214 for attaching to the emitter pad 3000 (FIG. 30B), as described below. The detector end 2202 has detector solder pads for attaching the detector 2410 (FIG. 24).

The interconnect 2204 between the emitter end 2201 and the cable connector 2230 forms a "neck," and the interconnect 2206 between the detector end 2202 and the cable connector 2230 forms a "tail." The cable connector 2230 forms "wings" that extend from the interconnect 2204, 2206 between the neck 2204 and tail 2206. A conductive ink flap 2240 connects to the cable inner shield 4450 (FIGS. 44A-B) and folds over to shield the detector wires 4470 (FIGS. 44A-B) soldered to the detector wire pads 2236. The outer wire pads 2238 connect to the remaining cable wires 4430 (FIGS. 44A-B). The flex circuit 2200 has top coverlay, top ink, inner coverlay, trace, trace base, bottom ink and bottom coverlay layers.

The flex circuit 2200 advantageously provides a connection between a multiple wire sensor cable 4400 (FIGS. 44A-B), a multiple wavelength emitter assembly 500 (FIG. 6) and a detector assembly 2400 (FIG. 24) without rendering the emitter and detector assemblies unwieldy and stiff. In particular, the wings 2230 provide a relatively large solder pad area 2232 that is narrowed at the neck 2204 and tail 2206 to mechanically isolate the cable 4400 (FIGS. 44A-B) from the remainder of the flex circuit 2200. Further, the neck 2206 is folded (see FIG. 4) for installation in the emitter pad 3000 (FIGS. 30A-H) and acts as a flexible spring to further mechanically isolate the cable 4400 (FIGS. 44A-B) from the emitter assembly 500 (FIG. 4). The tail 2206 provides an integrated connectivity path between the detector assembly 2400 (FIG. 24) mounted in the detector pad 3100 (FIGS. 31A-H) and the cable connector 2230 mounted in the opposite emitter pad 3000 (FIGS. 30A-H).

[0086] FIG. 23 illustrates the emitter portion of the flex circuit assembly 1900 (FIG. 21) having the emitter assembly 500. The emitter assembly connector 1530 is attached to the emitter end 2210 of the flex circuit 2200 (FIG. 22). In particular, reflow solder 2330 connects thru hole pads 1532, 1534 of the emitter assembly 500 to corresponding emitter pads 2310 of the flex circuit 2200 (FIG. 22).

[0087] FIG. 24 illustrates a detector assembly 2400 including a detector 2410, solder pads 2420, copper mesh tape 2430, an EMI shield 2440 and foil 2450. The detector 2410 is soldered 2460 chip side down to detector solder pads 2420 of the flex circuit 2200. The detector solder joint and detector ground pads 2420

are wrapped with the Kapton tape 2470. EMI shield tabs 2442 are folded onto the detector pads 2420 and soldered. The EMI shield walls are folded around the detector 2410 and the remaining tabs 2442 are soldered to the back of the EMI shield 2440. The copper mesh tape 2430 is cut to size and the shielded detector and flex circuit solder joint are wrapped with the copper mesh tape 2430. The foil 2450 is cut to size with a predetermined aperture 2452. The foil 2450 is wrapped around shielded detector with the foil side in and the aperture 2452 is aligned with the EMI shield grid 2444.

#### **Detector Assembly**

FIG. 25 illustrates an alternative detector assembly 2400 embodiment [0088] having adjacent detectors. Optical radiation having multiple wavelengths generated by emitters 700 is transmitted into a tissue site 1. Optical radiation at a first set of wavelengths is detected by a first detector 2510, such as, for example, a Si detector. Optical radiation at a second set of wavelengths is detected by a second detector 2520, such as, for example, a GaAs detector. FIG. 26 illustrates another alternative detector assembly 2400 [0089] embodiment having stacked detectors coaxial along a light path. Optical radiation having multiple wavelengths generated by emitters 700 is transmitted into a tissue site 1. Optical radiation at a first set of wavelengths is detected by a first detector 2610. Optical radiation at a second set of wavelengths passes through the first detector 2610 and is detected by a second detector 2620. In a particular embodiment, a silicon (Si) detector and a gallium arsenide (GaAs) detector are used. The Si detector is placed on top of the GaAs detector so that light must pass through the Si detector before reaching the GaAs detector. The Si detector can be placed directly on top of the GaAs detector or the Si and GaAs detector can be separated by some other medium, such as a transparent medium or air. In another particular embodiment, a germanium detector is used instead of the GaAs detector. Advantageously, the stacked detector arrangement minimizes error caused by pathlength differences as compared with the adjacent detector embodiment.

#### Finger Clip

[0090] FIG. 27 illustrates a finger clip embodiment 2700 of a physiological sensor attachment assembly. The finger clip **2700** is configured to removably attach an emitter assembly 500 (FIG. 6) and detector assembly 2400 (FIG. 24), interconnected by a flex circuit assembly 1900, to a fingertip. The finger clip 2700 has an emitter shell 3800, an emitter pad 3000, a detector pad 2800 and a detector shell 3900. The emitter shell 3800 and the detector shell 3900 are rotatably connected and urged together by the spring assembly 3500. The emitter pad 3000 is fixedly retained by the emitter shell. The emitter assembly 500 (FIG. 6) is mounted proximate the emitter pad 3000 and adapted to transmit optical radiation having a plurality of wavelengths into fingertip tissue. The detector pad 2800 is fixedly retained by the detector shell 3900. The detector assembly 3500 is mounted proximate the detector pad 2800 and adapted to receive the optical radiation after attenuation by fingertip tissue. FIG. 28 illustrates a detector pad 2800 advantageously configured to [0091] position and comfortably maintain a fingertip relative to a detector assembly for accurate sensor measurements. In particular, the detector pad has fingertip positioning features including a guide 2810, a contour 2820 and a stop 2830. The guide **2810** is raised from the pad surface **2803** and narrows as the guide 2810 extends from a first end 2801 to a second end 2802 so as to increasingly conform to a fingertip as a fingertip is inserted along the pad surface 2803 from the first end 2801. The contour 2820 has an indentation defined along the pad surface 2803 generally shaped to conform to a fingertip positioned over a detector aperture 2840 located within the contour 2820. The stop 2830 is raised from the pad surface 2803 so as to block the end of a finger from inserting bevond the second end 2802. FIGS. 29A-B illustrate detector pad embodiments 3100, 3400 each having a guide 2810, a contour 2820 and a stop 2830, described in further detail with respect to FIGS. 31 and 34, respectively. [0092] FIGS. 30A-H illustrate an emitter pad 3000 having emitter pad flaps 3010, an emitter window 3020, mounting pins 3030, an emitter assembly cavity 3040, isolation notches 3050, a flex circuit notch 3070 and a cable notch 3080.

The emitter pad flaps 3010 overlap with detector pad flaps 3110 (FIGS. 31A-H)

to block ambient light. The emitter window 3020 provides an optical path from the emitter array 700 (FIG. 8) to a tissue site. The mounting pins 3030 accommodate apertures in the flex circuit mounting ears 2214 (FIG. 22), and the cavity 3040 accommodates the emitter assembly 500 (FIG. 21). Isolation notches 3050 mechanically decouple the shell attachment 3060 from the remainder of the emitter pad 3000. The flex circuit notch 3070 accommodates the flex circuit tail 2206 (FIG. 22) routed to the detector pad 3100 (FIGS. 31A-H). The cable notch 3080 accommodates the sensor cable 4400 (FIGS. 44A-B). FIGS. 33A-H illustrate an alternative slim finger emitter pad 3300 embodiment. FIGS. 31A-H illustrate a detector pad 3100 having detector pad flaps [0093] 3110, a shoe box cavity 3120 and isolation notches 3150. The detector pad flaps 3110 overlap with emitter pad flaps 3010 (FIGS. 30A-H), interleaving to block ambient light. The shoe box cavity 3120 accommodates a shoe box 3200 (FIG. 32A-H) described below. Isolation notches 3150 mechanically decouple the attachment points 3160 from the remainder of the detector pad 3100. FIGS. **34A-H** illustrate an alternative slim finger detector pad **3400** embodiment. FIGS. 32A-H illustrate a shoe box 3200 that accommodates the detector assembly 2400 (FIG. 24). A detector window 3210 provides an optical path from a tissue site to the detector 2410 (FIG. 24). A flex circuit notch 3220 accommodates the flex circuit tail 2206 (FIG. 22) routed from the emitter pad 3000 (FIGS. 30A-H). In one embodiment, the shoe box 3200 is colored black or other substantially light absorbing color and the emitter pad 3000 and detector pad 3100 are each colored white or other substantially light reflecting color. [0095] FIGS. 35-37 illustrate a spring assembly 3500 having a spring 3600 configured to urge together an emitter shell 3800 (FIG. 46) and a detector shell **3900**. The detector shell is rotatably connected to the emitter shell. The spring is disposed between the shells 3800, 3900 and adapted to create a pivot point along a finger gripped between the shells that is substantially behind the fingertip. This advantageously allows the shell hinge 3810, 3910 (FIGS. 38-39) to expand so as to distribute finger clip force along the inserted finger, comfortably keeping the fingertip in position over the detector without excessive force.

[0096] As shown in FIGS 36A-C, the spring 3600 has coils 3610, an emitter shell leg 3620 and a detector shell leg 3630. The emitter shell leg 3620 presses against the emitter shell 3800 (FIGS. 38A-D) proximate a grip 3820 (FIGS. 38A-D). The detector shell legs 3630 extend along the detector shell 3900 (FIGS. 39A-D) to a spring plate 3700 (FIGS. 37A-D) attachment point. The coil 3610 is secured by hinge pins 410 (FIG. 46) and is configured to wind as the finger clip is opened, reducing its diameter and stress accordingly.

[0097] As shown in FIGS. 37A-D the spring plate 3700 has attachment apertures 3710, spring leg slots 3720, and a shelf 3730. The attachment apertures 3710 accept corresponding shell posts 3930 (FIGS. 39A-D) so as to secure the spring plate 3700 to the detector shell 3900 (FIG. 39A-D). Spring legs 3630 (FIG. 36A-C) are slidably anchored to the detector shell 3900 (FIG. 39A-D) by the shelf 3730, advantageously allowing the combination of spring 3600, shells 3800, 3900 and hinges 3810, 3910 to adjust to various finger sizes and shapes.

[0098] FIGS. 38-39 illustrate the emitter and detector shells 3800, 3900, respectively, having hinges 3810, 3910 and grips 3820, 3920. Hinge apertures 3812, 3912 accept hinge pins 410 (FIG. 46) so as to create a finger clip. The detector shell hinge aperture 3912 is elongated, allowing the hinge to expand to accommodate a finger.

#### Monitor And Sensor

[0099] FIG. 40 illustrates a monitor 100 and a corresponding sensor assembly 200, as described generally with respect to FIGS. 1-3, above. The sensor assembly 200 has a sensor 400 and a sensor cable 4400. The sensor 400 houses an emitter assembly 500 having emitters responsive to drivers within a sensor controller 4500 so as to transmit optical radiation into a tissue site. The sensor 400 also houses a detector assembly 2400 that provides a sensor signal 2500 responsive to the optical radiation after tissue attenuation. The sensor signal 2500 is filtered, amplified, sampled and digitized by the front-end 4030 and input to a DSP (digital signal processor) 4040, which also commands the sensor controller 4500. The sensor cable 4400 electrically communicates drive signals from the sensor controller 4500 to the emitter assembly 500 and a sensor signal

2500 from the detector assembly 2400 to the front-end 4030. The sensor cable 4400 has a monitor connector 210 that plugs into a monitor sensor port 110. [0100] In one embodiment, the monitor 100 also has a reader 4020 capable of obtaining information from an information element (IE) in the sensor assembly 200 and transferring that information to the DSP 4040, to another processor or component within the monitor 100, or to an external component or device that is at least temporarily in communication with the monitor 100. In an alternative embodiment, the reader function is incorporated within the DSP 4040, utilizing one or more of DSP I/O, ADC, DAC features and corresponding processing routines, as examples.

In one embodiment, the monitor connector 210 houses the information [0101] element 4000, which may be a memory device or other active or passive electrical component. In a particular embodiment, the information element 4000 is an EPROM, or other programmable memory, or an EEPROM, or other reprogrammable memory, or both. In an alternative embodiment, the information element 4000 is housed within the sensor 400, or an information element 4000 is housed within both the monitor connector 4000 and the sensor 400. In yet another embodiment, the emitter assembly 500 has an information element 4000, which is read in response to one or more drive signals from the sensor controller 4500, as described with respect to FIGS. 41-43, below. In a further embodiment, a memory information element is incorporated into the emitter array 700 (FIG. 8) and has characterization information relating to the LEDs 801 (FIG. 8). In one advantageous embodiment, trend data relating to slowly varying parameters, such as perfusion index, HbCO or METHb, to name a few, are stored in an IE memory device, such as EEPROM.

#### Back-to-Back LEDs

[0102] FIGS. 41-43 illustrate alternative sensor embodiments. A sensor controller 4500 configured to activate an emitter array 700 (FIG. 7) arranged in an electrical grid, is described with respect to FIG. 7, above. Advantageously, a sensor controller 4500 so configured is also capable of driving a conventional

two-wavelength (red and IR) sensor **4100** having back-to-back LEDs **4110**, **4120** or an information element **4300** or both.

[0103] FIG. 41A illustrates a sensor 4100 having an electrical grid 4130 configured to activate light emitting sources by addressing at least one row conductor and at least one column conductor. A first LED 4110 and a second LED 4120 are configured in a back-to-back arrangement so that a first contact 4152 is connected to a first LED 4110 cathode and a second LED 4120 anode and a second contact 4154 is connected to a first LED 4110 anode and a second LED 4120 cathode. The first contact 4152 is in communications with a first row conductor 4132 and a first column conductor 4134. The second contact is in communications with a second row conductor 4136 and a second column conductor 4138. The first LED 4110 is activated by addressing the first row conductor 4132 and the second column conductor 4138. The second LED 4120 is activated by addressing the second LED 4120 is activated by addressing the second column conductor 4136 and the first column conductor 4134.

[0104] FIG. 41B illustrates a sensor cable 4400 embodiment capable of communicating signals between a monitor 100 and a sensor 4100. The cable 4400 has a first row input 4132, a first column input 4134, a second row input 4136 and a second column input 4138. A first output 4152 combines the first row input 4132 and the first column input 4134. A second output 4154 combines a second row input 4136 and second column input 4138.

[0105] FIG. 41C illustrates a monitor 100 capable of communicating drive signals to a sensor 4100. The monitor 4400 has a first row signal 4132, a first column signal 4134, a second row signal 4136 and a second column signal 4138. A first output signal 4152 combines the first row signal 4132 and the first column signal 4134. A second output signal 4154 combines a second row signal 4136 and second column signal 4138.

#### Information Elements

[0106] FIGS. 42-43 illustrate information element 4200-4300 embodiments in communications with emitter array drivers configured to activate light emitters connected in an electrical grid. The information elements are configured to provide information as DC values, AC values or a combination of DC and AC

values in response corresponding DC, AC or combination DC and AC electrical grid drive signals. FIG. **42** illustrates information element embodiment **4200** advantageously driven directly by an electrical grid having rows **710** and columns **720**. In particular, the information element **4200** has a series connected resistor R<sub>2</sub> **4210** and diode **4220** connected between a row line **710** and a column line **720** of an electrical grid. In this manner, the resistor R<sub>2</sub> value can be read in a similar manner that LEDs **810** (FIG. **8**) are activated. The diode **4220** is oriented, e.g. anode to row and cathode to column as the LEDs so as to prevent parasitic currents from unwanted activation of LEDs **810** (FIG. **8**).

FIGS. 43A-C illustrate other embodiments where the value of R<sub>1</sub> is [0107] read with a DC grid drive current and a corresponding grid output voltage level. In other particular embodiments, the combined values of R<sub>1</sub>, R<sub>2</sub> and C or, alternatively, R<sub>1</sub>, R<sub>2</sub> and L are read with a varying (AC) grid drive currents and a corresponding grid output voltage waveform. As one example, a step in grid drive current is used to determine component values from the time constant of a corresponding rise in grid voltage. As another example, a sinusoidal grid drive current is used to determine component values from the magnitude or phase or both of a corresponding sinusoidal grid voltage. The component values determined by DC or AC electrical grid drive currents can represent sensor types, authorized suppliers or manufacturers, emitter wavelengths among others. Further, a diode D (FIG. 43C) can be used to provide one information element reading R<sub>1</sub> at one drive level or polarity and another information element reading, combining R<sub>1</sub> and R<sub>2</sub>, at a second drive level or polarity, i.e. when the diode is forward biased.

[0108] Passive information element 4300 embodiments may include any of various combinations of resistors, capacitors or inductors connected in series and parallel, for example. Other information element 4300 embodiments connected to an electrical grid and read utilizing emitter array drivers incorporate other passive components, active components or memory components, alone or in combination, including transistor networks, PROMs, ROMs, EPROMs, EEPROMs, gate arrays and PLAs to name a few.

#### Sensor Cable

[0109] FIGS. 44A-B illustrate a sensor cable 4400 having an outer jacket 4410, an outer shield 4420, multiple outer wires 4430, an inner jacket 4440, an inner shield 4450, a conductive polymer 4460 and an inner twisted wire pair 4470. The outer wires 4430 are advantageously configured to compactly carry multiple drive signals to the emitter array 700 (FIG. 7). In one embodiment, there are twelve outer wires 4430 corresponding to four anode drive signals 4501 (FIG. 45), four cathode drive signals 4502 (FIG. 45), two thermistor pinouts 1450 (FIG. 15) and two spares. The inner twisted wire pair 4470 corresponds to the sensor signal 2500 (FIG. 25) and is extruded within the conductive polymer 4460 so as to reduce triboelectric noise. The shields 4420, 4450 and the twisted pair 4470 boost EMI and crosstalk immunity for the sensor signal 2500 (FIG. 25).

#### Controller

[0110] FIG. 45 illustrates a sensor controller 4500 located in the monitor 100 (FIG. 1) and configured to provide anode drive signals 4501 and cathode drive signals 4502 to the emitter array 700 (FIG. 7). The DSP (digital signal processor) 4040, which performs signal processing functions for the monitor, also provides commands 4042 to the sensor controller 4500. These commands determine drive signal 4501, 4502 levels and timing. The sensor controller 4500 has a command register 4510, an anode selector 4520, anode drivers 4530, current DACs (digital-to-analog converters) 4540, a current multiplexer 4550, cathode drivers 4560, a current meter 4570 and a current limiter 4580. The command register 4510 provides control signals responsive to the DSP commands 4042. In one embodiment, the command register 4510 is a shift register that loads serial command data 4042 from the DSP 4040 and synchronously sets output bits that select or enable various functions within the sensor controller 4500, as described below.

[0111] As shown in FIG. 45, the anode selector 4520 is responsive to anode select 4516 inputs from the command register 4510 that determine which emitter array row 810 (FIG. 8) is active. Accordingly, the anode selector 4520 sets one of the anode on 4522 outputs to the anode drivers 4530, which pulls up to Vcc one of the anode outputs 4501 to the emitter array 700 (FIG. 8).

Also shown in FIG. 45, the current DACs 4540 are responsive to [0112] command register data 4519 that determines the currents through each emitter array column 820 (FIG. 8). In one embodiment, there are four, 12-bit DACs associated with each emitter array column 820 (FIG. 8), sixteen DACs in total. That is, there are four DAC outputs 4542 associated with each emitter array column 820 (FIG. 8) corresponding to the currents associated with each row 810 (FIG. 8) along that column 820 (FIG. 8). In a particular embodiment, all sixteen DACs 4540 are organized as a single shift register, and the command register 4510 serially clocks DAC data 4519 into the DACs 4540. A current multiplexer 4550 is responsive to cathode on 4518 inputs from the command register 4510 and anode on 4522 inputs from the anode selector 4520 so as to convert the appropriate DAC outputs 4542 to current set 4552 inputs to the cathode drivers 4560. The cathode drivers 4560 are responsive to the current set 4552 inputs to pull down to ground one to four of the cathode outputs 4502 to the emitter array 700 (FIG. 8).

[0113] The current meter **4570** outputs a current measure **4572** that indicates the total LED current driving the emitter array **700** (FIG. **8**). The current limiter **4580** is responsive to the current measure **4572** and limits specified by the command register **4510** so as to prevent excessive power dissipation by the emitter array **700** (FIG. **8**). The current limiter **4580** provides an enable **4582** output to the anode selector **4520**. A Hi Limit **4512** input specifies the higher of two preset current limits. The current limiter **4580** latches the enable **4582** output in an off condition when the current limit is exceeded, disabling the anode selector **4520**. A trip reset **4514** input resets the enable **4582** output to re-enable the anode selector **4520**.

#### Sensor Assembly

[0114] As shown in FIG. 46, the sensor 400 has an emitter shell 3800, an emitter pad 3000, a flex circuit assembly 2200, a detector pad 3100 and a detector shell 3900. A sensor cable 4400 attaches to the flex circuit assembly 2200, which includes a flex circuit 2100, an emitter assembly 500 and a detector assembly 2400. The portion of the flex circuit assembly 2200 having the sensor cable 4400 attachment and emitter assembly 500 is housed by the emitter shell

3800 and emitter pad 3000. The portion of the flex circuit assembly 2200 having the detector assembly 2400 is housed by the detector shell 3900 and detector pad 3100. In particular, the detector assembly 2400 inserts into a shoe 3200, and the shoe 3200 inserts into the detector pad 3100. The emitter shell 3800 and detector shell 3900 are fastened by and rotate about hinge pins 410, which insert through coils of a spring 3600. The spring 3600 is held to the detector shell 3900 with a spring plate 3700. A finger stop 450 attaches to the detector shell. In one embodiment, a silicon adhesive 420 is used to attach the pads 3000, 3100 to the shells 3800, 3900, a silicon potting compound 430 is used to secure the emitter and detector assemblies 500, 2400 within the pads 3000, 3100, and a cyanoacrylic adhesive 440 secures the sensor cable 4400 to the emitter shell 3800.

[0115] A multiple wavelength sensor has been disclosed in detail in connection with various embodiments. These embodiments are disclosed by way of examples only and are not to limit the scope of the claims that follow. One of ordinary skill in art will appreciate many variations and modifications.

#### WHAT IS CLAIMED IS:

1. A physiological sensor adapted to removably attach an emitter assembly and a detector assembly to a fingertip, the emitter assembly adapted to transmit optical radiation having a plurality of wavelengths into fingertip tissue, the detector assembly adapted to receive the optical radiation after attenuation by the fingertip tissue, the sensor comprising:

- a first shell;
- a second shell hinged to the first shell;
- a spring disposed between and urging together the shells;
- an emitter pad fixedly attached to the first shell and configured to retain the emitter assembly;
- a detector pad fixedly attached to the second shell configured to retain the detector assembly;
- a detector aperture defined within the detector pad and adapted to pass optical radiation to the detector assembly; and
- a contour defined along the detector pad and generally shaped to conform to a fingertip positioned over the detector aperture.
- 2. The physiological sensor according to claim 1 further comprising a guide defined along, and raised from, the detector pad,

wherein the guide narrows from a first pad end to an opposite second pad end so as to increasingly conform to a fingertip inserted from the first pad end to the second pad end.

- 3. The physiological sensor according to claim 2 further comprising a stop raised from the detector pad and adapted to block the fingertip from extending beyond the second pad end.
- The physiological sensor according to claim 3 further comprising: a plurality of emitter pad flaps disposed along both sides of the emitter pad;
- a plurality of detector pad flaps disposed along both sides of the detector pad,

wherein the emitter pad flaps and the detector pad flaps are configured to interleave so as to block ambient light from the detector assembly.

- 5. The physiological sensor according to claim 4 further comprising a plurality of isolating notches defined along the emitter pad and along the detector pad and adapted to mechanically decouple shell attachment areas from the remainder of the emitter pad and the detector pad.
- 6. The physiological sensor according to claim 5 further comprising a cavity defined within the detector pad so as to accommodate a shoe that contains the detector assembly.
- 7. The physiological sensor according to claim 6 wherein the spring is configured so as to create a pivot point along a finger gripped between the shells that is substantially behind a fingertip.
  - 8. A physiological sensor comprising:
  - a first shell;
  - a second shell hinged to the first shell;
  - a spring disposed between and urging together the shells,

the spring configured so as to create a pivot point along a finger gripped between the shells that is substantially behind a fingertip; and

- a pad configured to position the fingertip.
- 9. The physiological sensor according to claim 8 wherein the spring comprises:
  - a first leg pressing against a grip portion of the first shell; and a second leg extending along a clip portion of the second shell.
- 10. The physiological sensor according to claim 9 further comprising a plate that slidably anchors the second leg to the clip portion of the second shell distal a grip portion of the second shell.

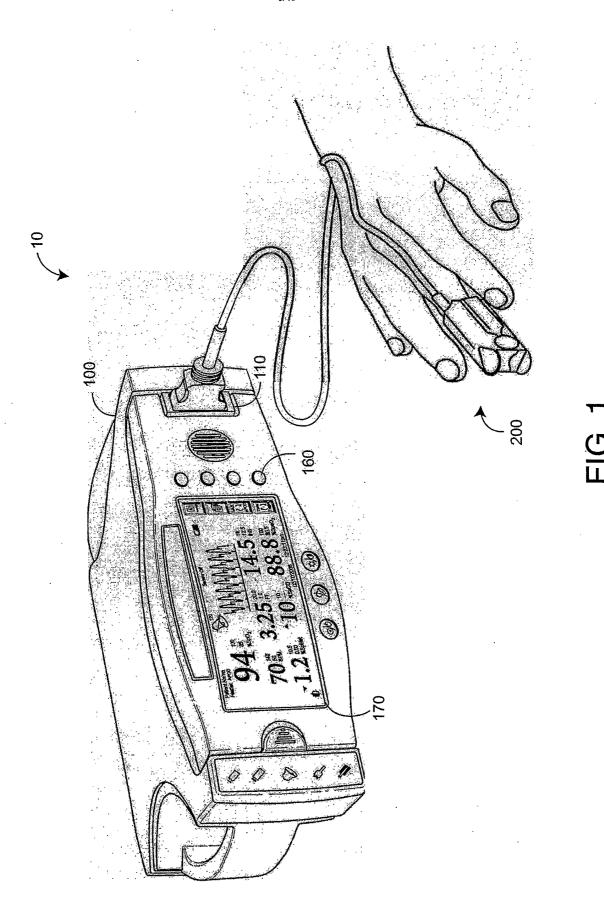
11. The physiological sensor according to claim 10 further comprising a contour defined along the pad and generally shaped to conform to the fingertip.

- 12. The physiological sensor according to claim 11 further comprising a guide raised from, and defined along, the pad and adapted to increasingly conform to a fingertip inserted along the pad.
- 13. The physiological sensor according to claim 12 further comprising a stop raised from the pad and adapted to block the fingertip from extending beyond the contour.
- 14. A physiological sensor method comprising: providing a plurality of housing elements comprising pads and shells; mounting a sensor assembly within the housing elements; positioning a fingertip relative to the sensor assembly with a plurality of pad features; and

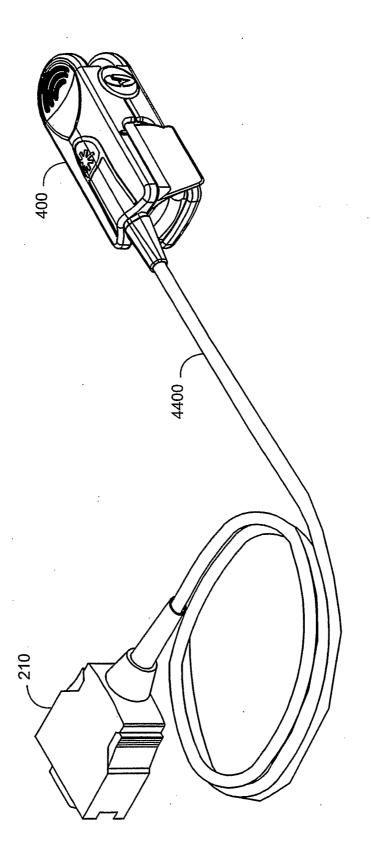
urging the housing elements against the fingertip so as to removably attach the sensor assembly to the fingertip.

- 15. The physiological sensor method according to claim 14 wherein the positioning comprises guiding a fingertip to a detector of the sensor assembly.
- 16. The physiological sensor method according to claim 15 wherein the positioning further comprises stopping the fingertip at the detector.
- 17. The physiological sensor method according to claim 16 wherein the positioning further comprises conforming to the fingertip at the detector.
- 18. The physiological sensor method according to claim 17 wherein the urging comprises pivotably gripping behind the fingertip with the housing elements.

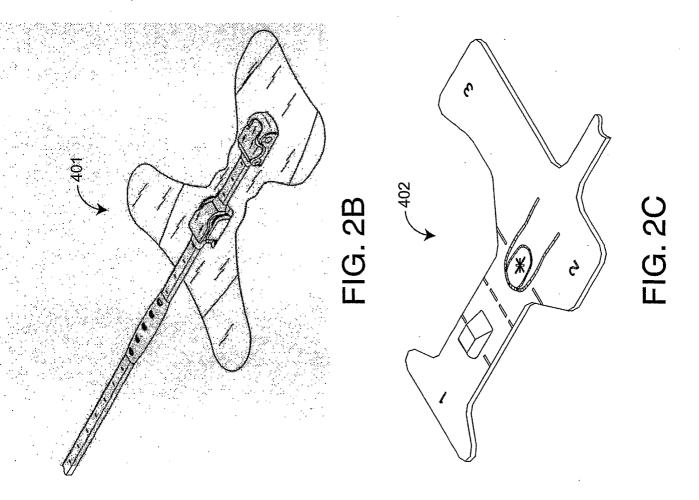
19. The physiological sensor method according to claim 18 wherein the gripping comprises anchoring an extended spring leg to one of the shells distal the detector.







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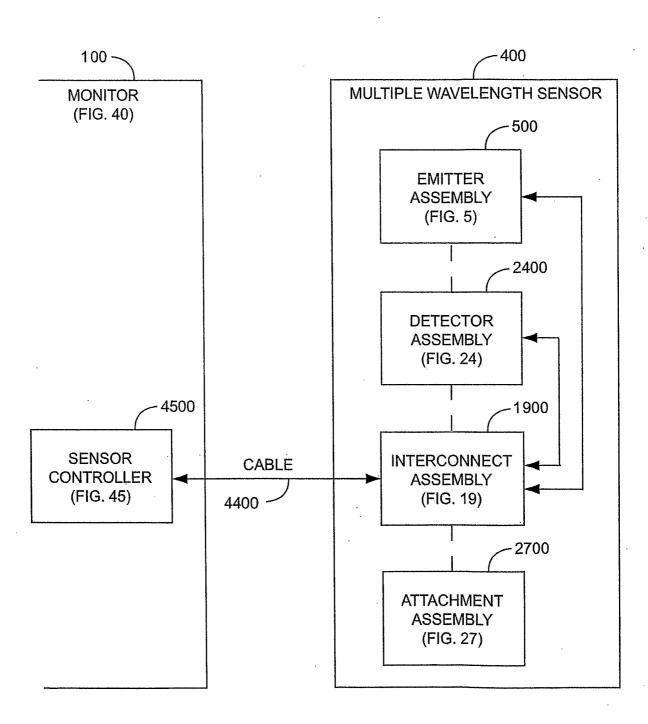


FIG. 3

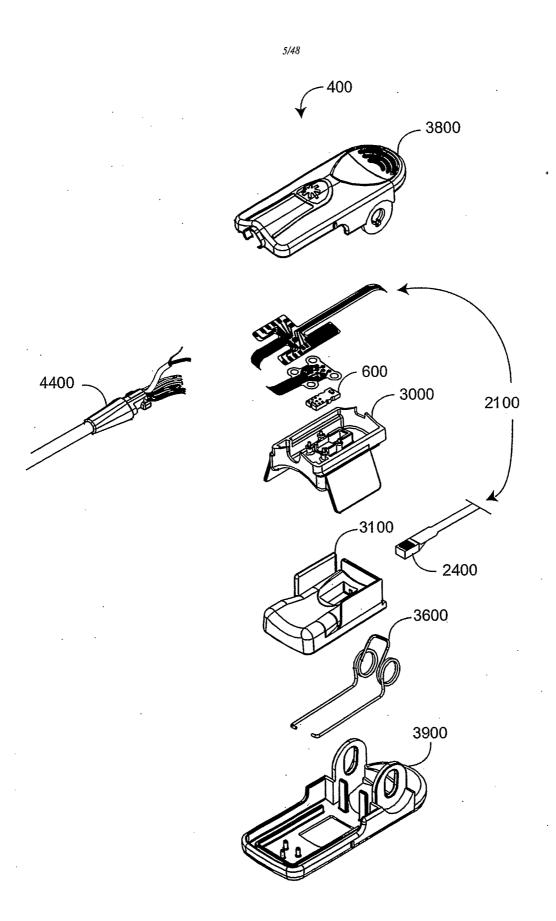


FIG. 4

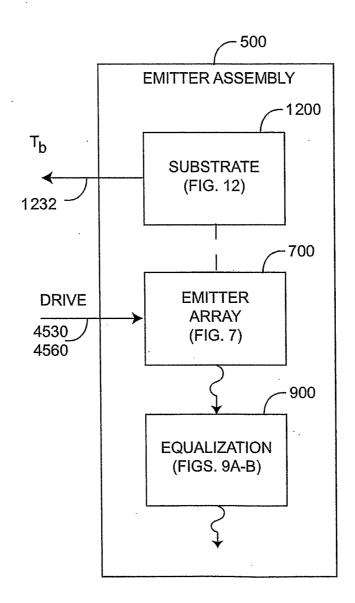
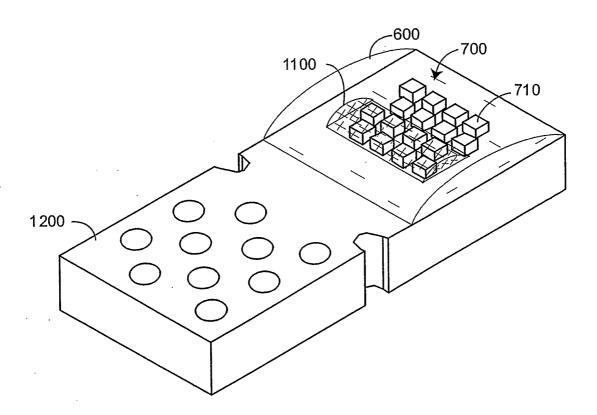


FIG. 5





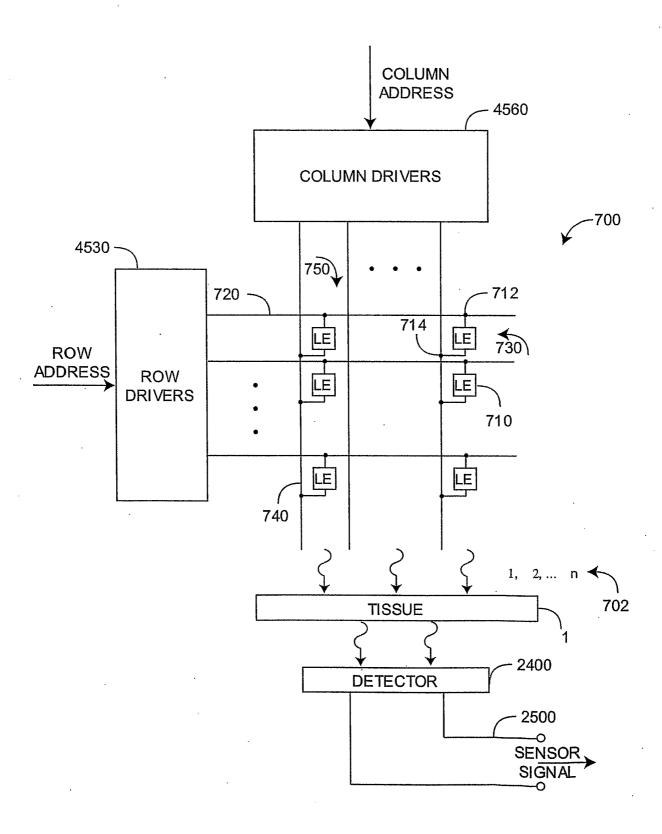


FIG. 7

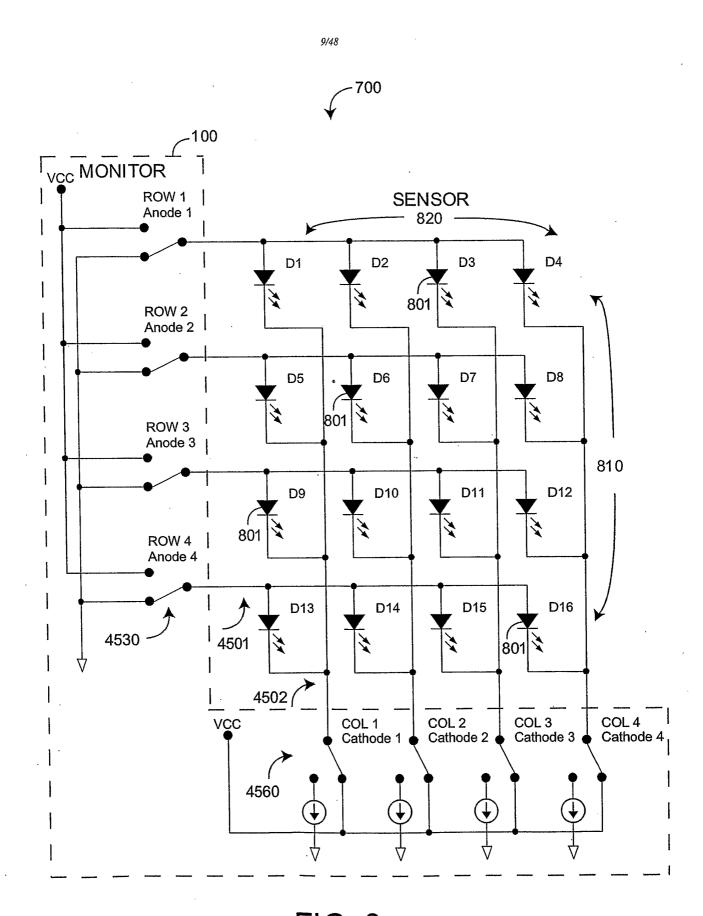
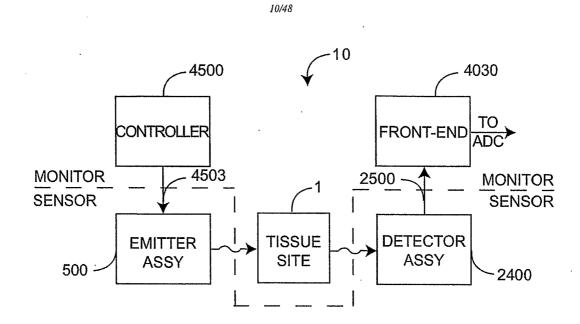


FIG. 8



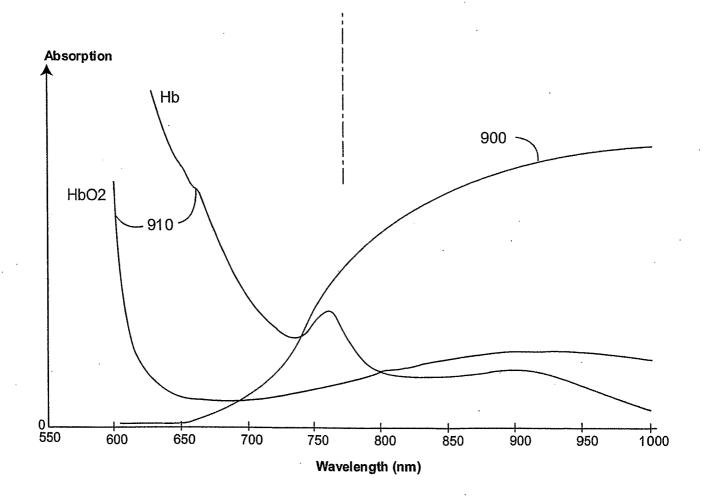
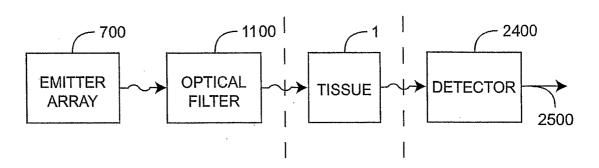


FIG. 9



**FIG. 10A** 

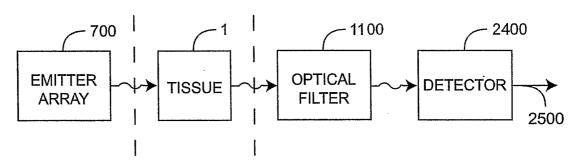


FIG. 10B

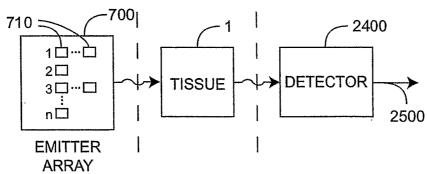
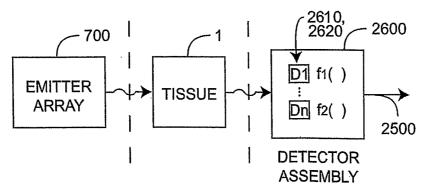
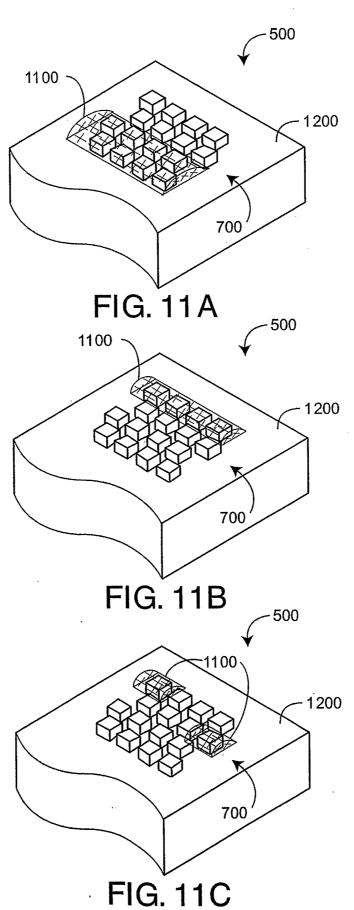


FIG. 10C



**FIG. 10D** 



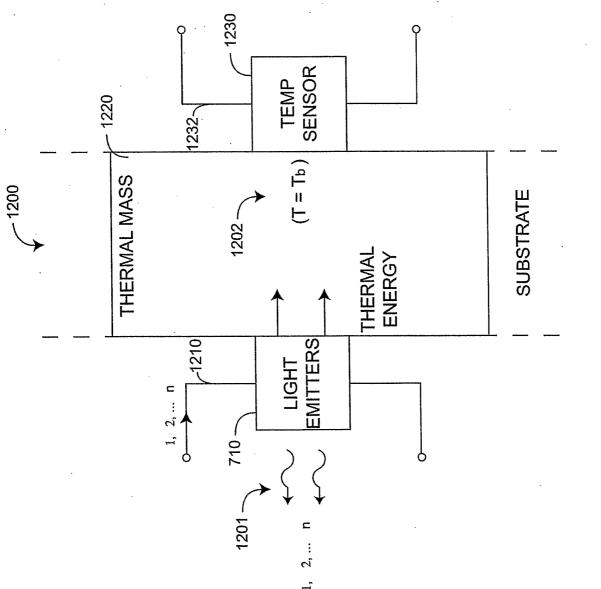


FIG. 12

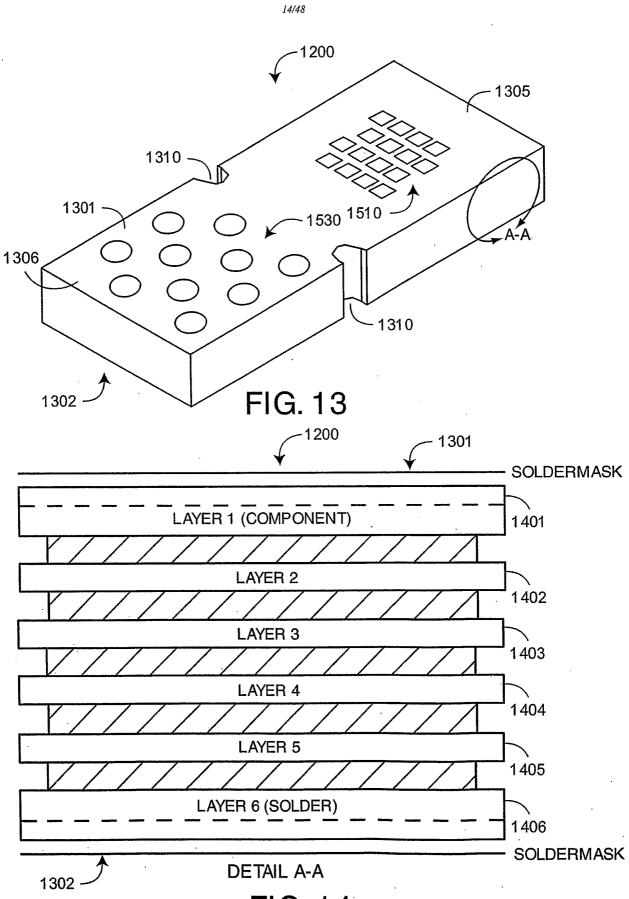


FIG. 14

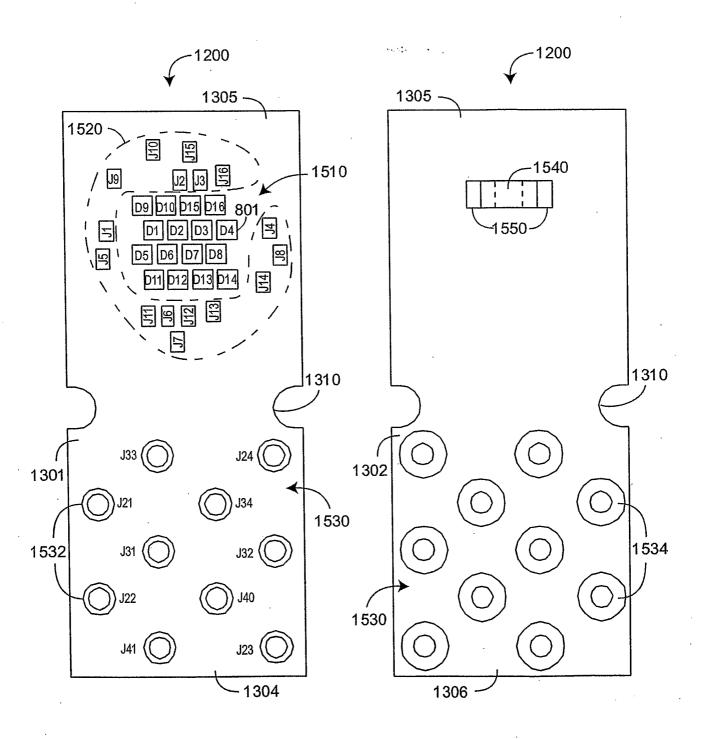
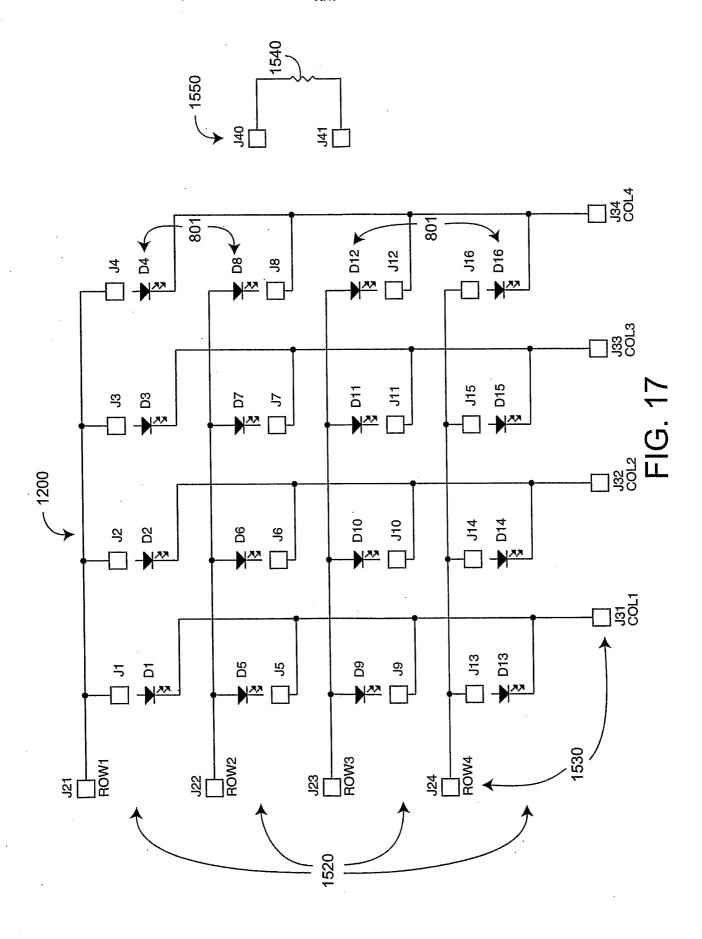


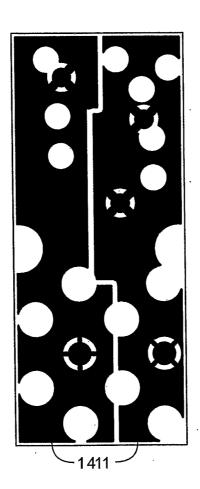
FIG. 15

FIG. 16



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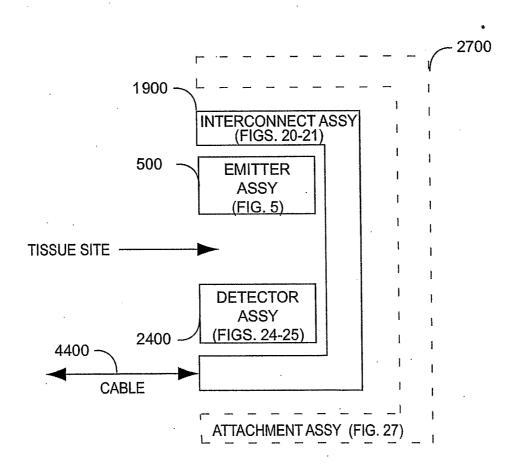
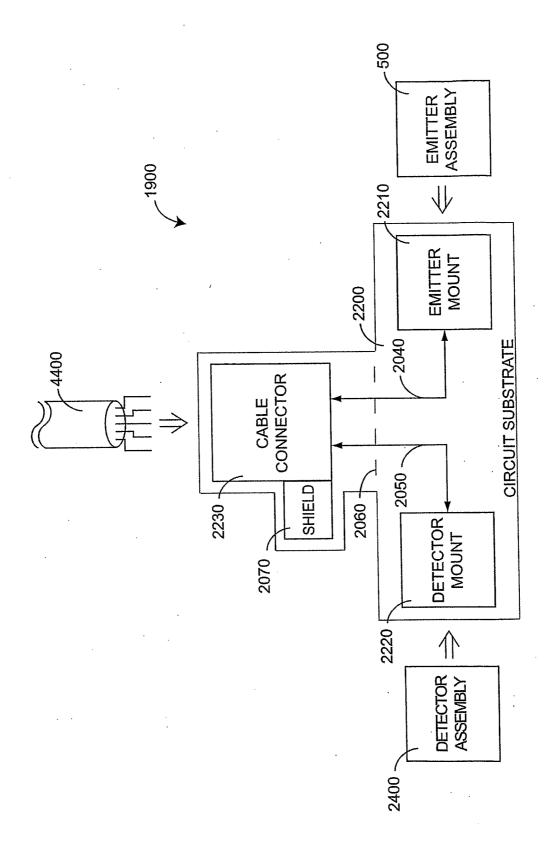
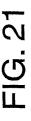
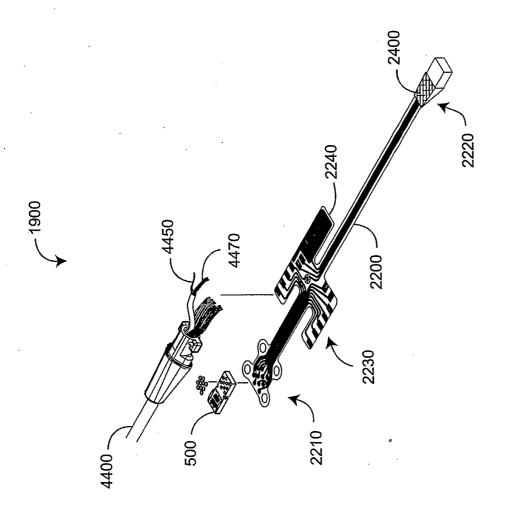


FIG. 19









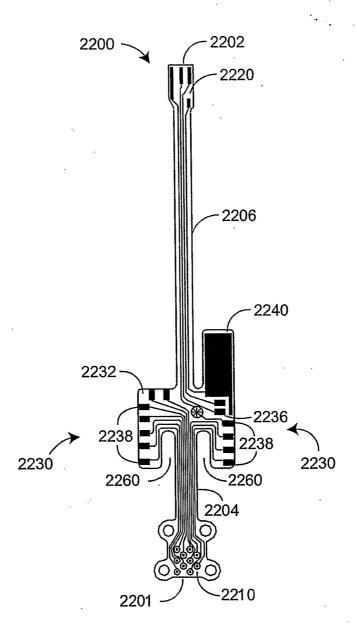


FIG. 22

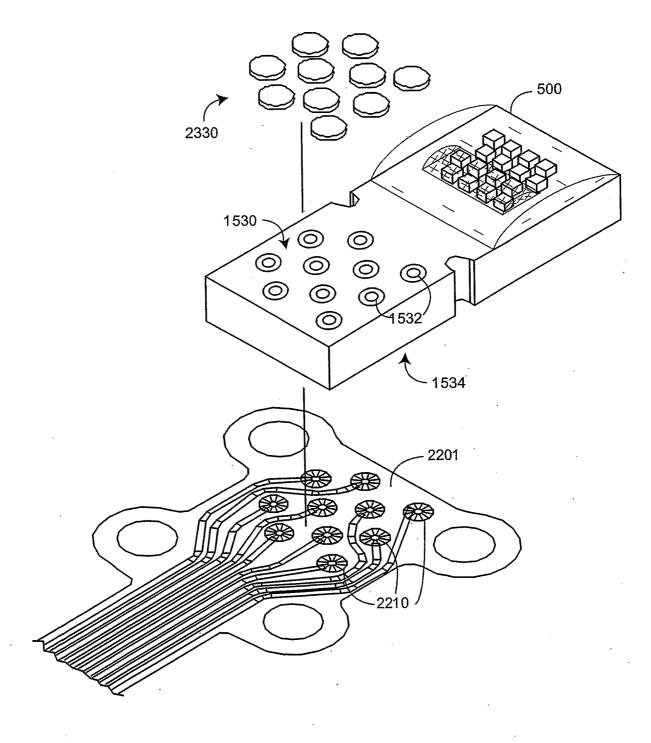
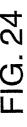
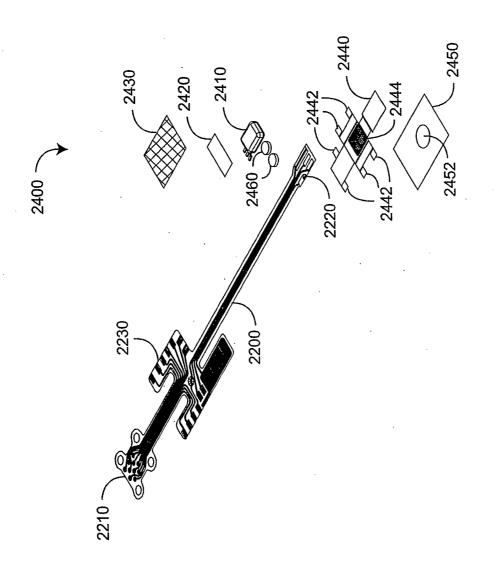


FIG. 23





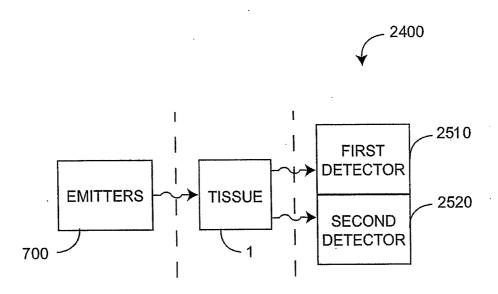


FIG. 25

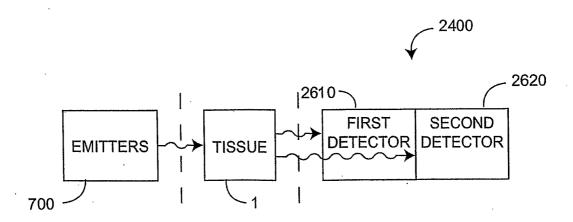


FIG. 26

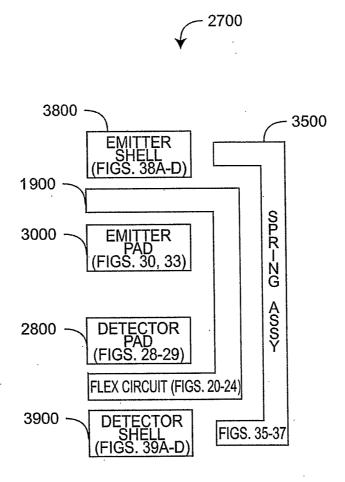
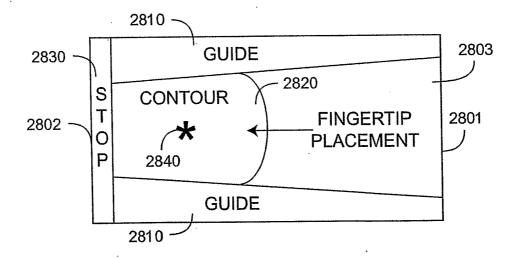


FIG. 27





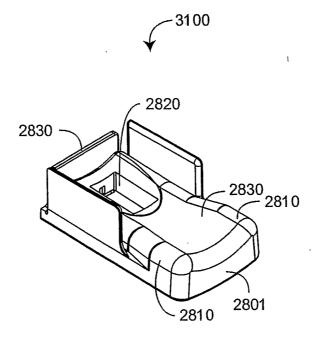


FIG. 29A

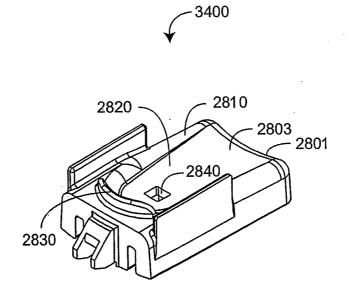


FIG. 29B

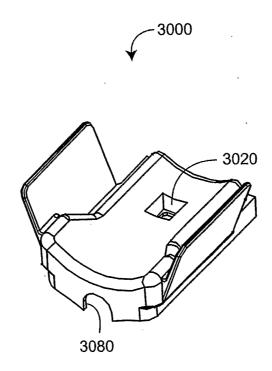


FIG. 30A

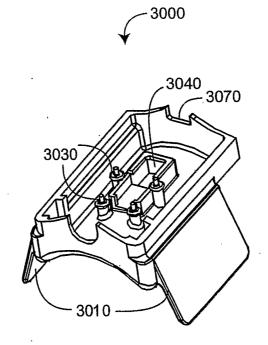


FIG. 30B

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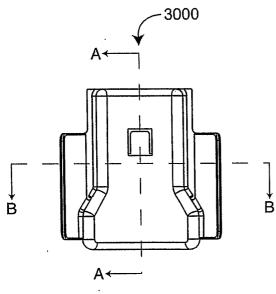


FIG. 30C

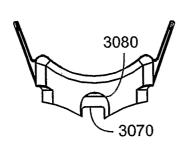


FIG. 30D

3000

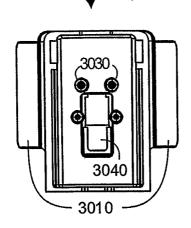


FIG. 30E

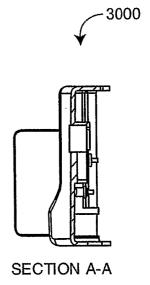


FIG. 30F

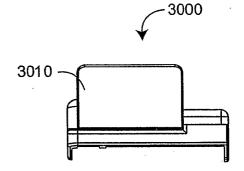


FIG. 30G

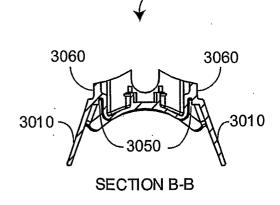


FIG. 30H



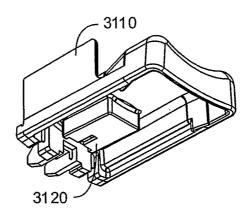


FIG. 31 A

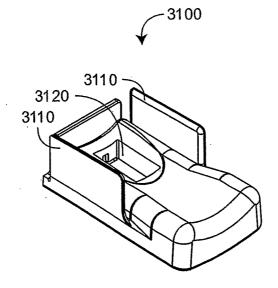


FIG. 31B

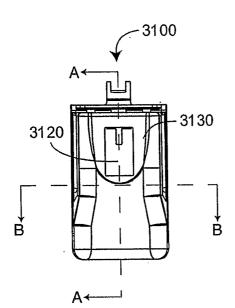


FIG. 31C

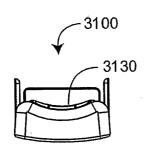


FIG. 31D

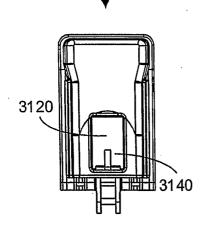
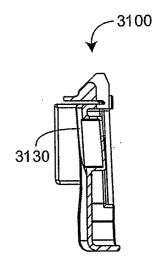


FIG. 31E



**SECTION A-A** 

FIG. 31F

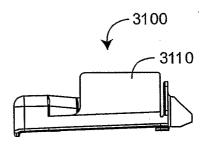
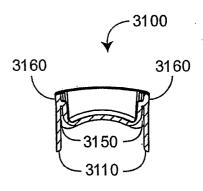
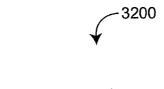


FIG. 31G



**SECTION B-B** 

FIG. 31H



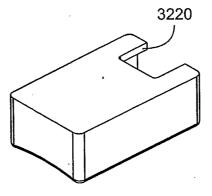


FIG. 32A

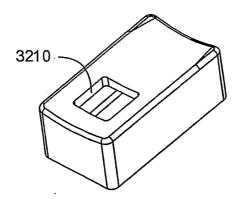


FIG. 32B

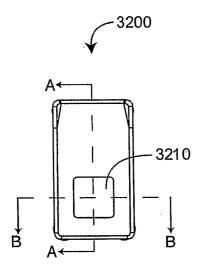


FIG. 32C

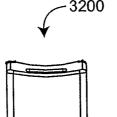


FIG. 32D

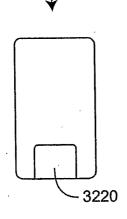
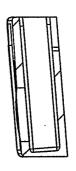


FIG. 32E





SECTION A-A

FIG. 32F

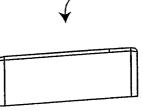


FIG. 32G





**SECTION B-B** 

FIG. 32H



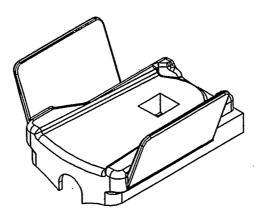


FIG. 33A

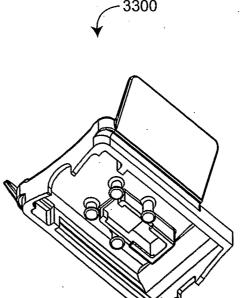
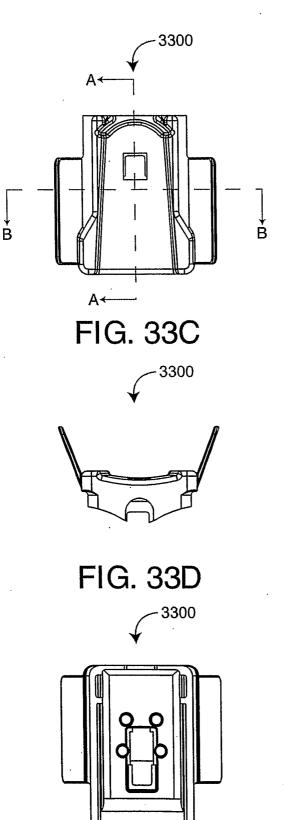
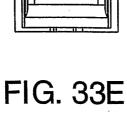


FIG. 33B





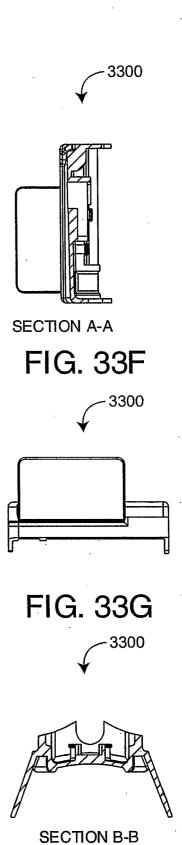


FIG. 33H



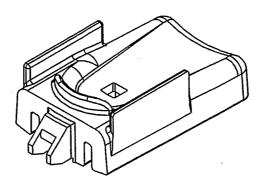


FIG. 34A

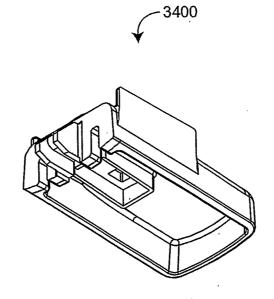


FIG. 34B

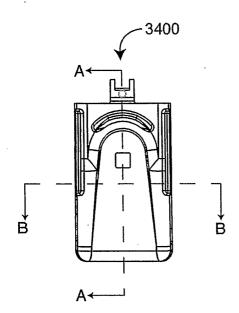


FIG. 34C





FIG. 34D



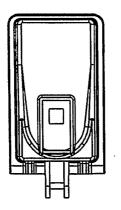
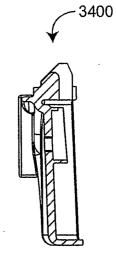


FIG. 34E



**SECTION A-A** 

FIG. 34F

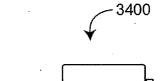
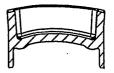




FIG. 34G





SECTION B-B

FIG. 34H

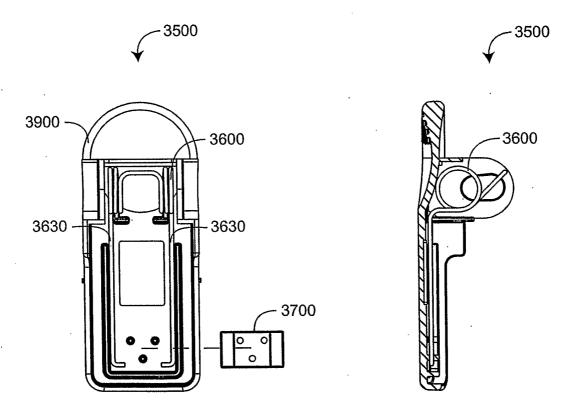


FIG. 35A FIG. 35B

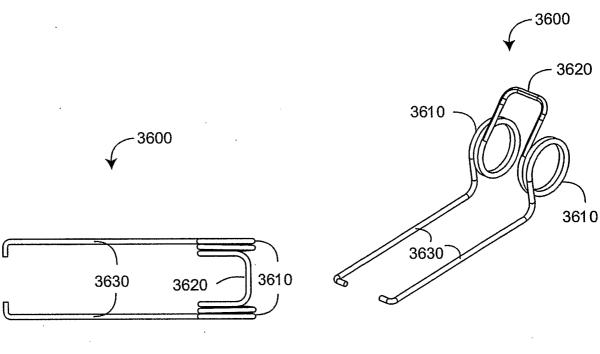


FIG. 36A

FIG. 36B

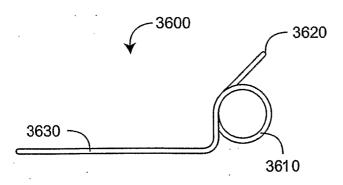


FIG. 36C

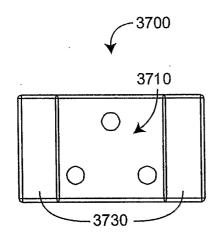


FIG. 37A

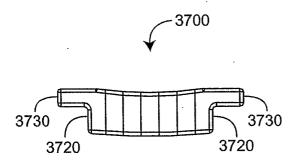


FIG. 37B

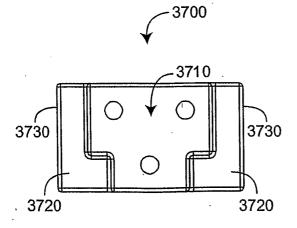


FIG. 37C

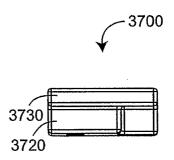


FIG. 37D

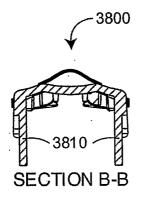


FIG. 38A

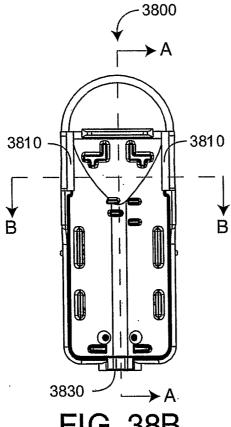


FIG. 38B

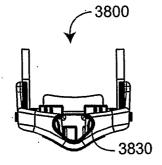


FIG. 38C

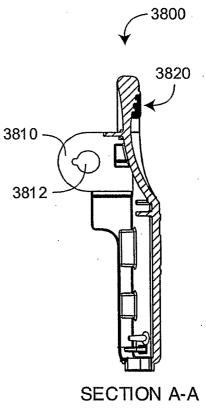


FIG. 38D

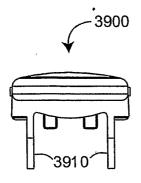


FIG. 39A

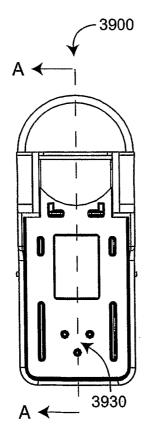


FIG.39B

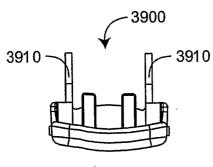


FIG. 39C

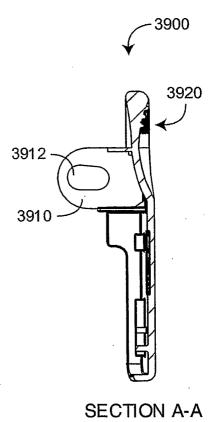


FIG. 39D

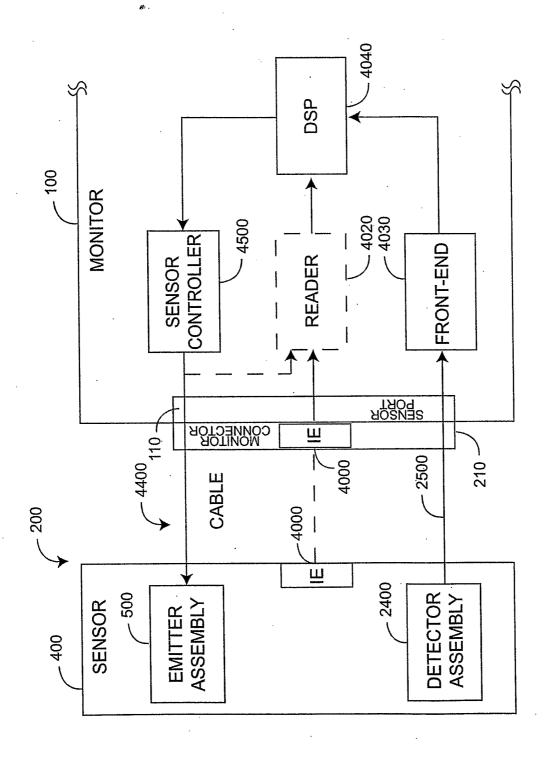


FIG. 40

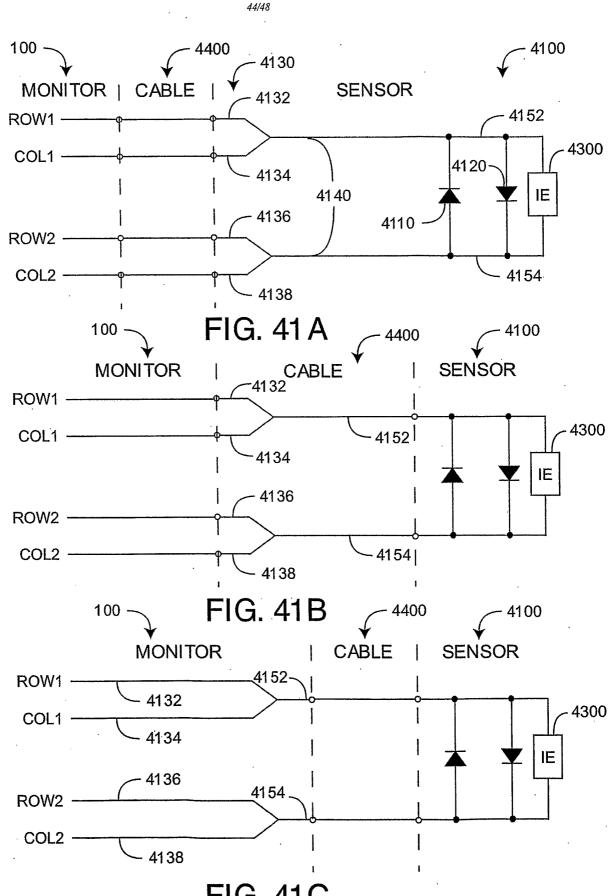
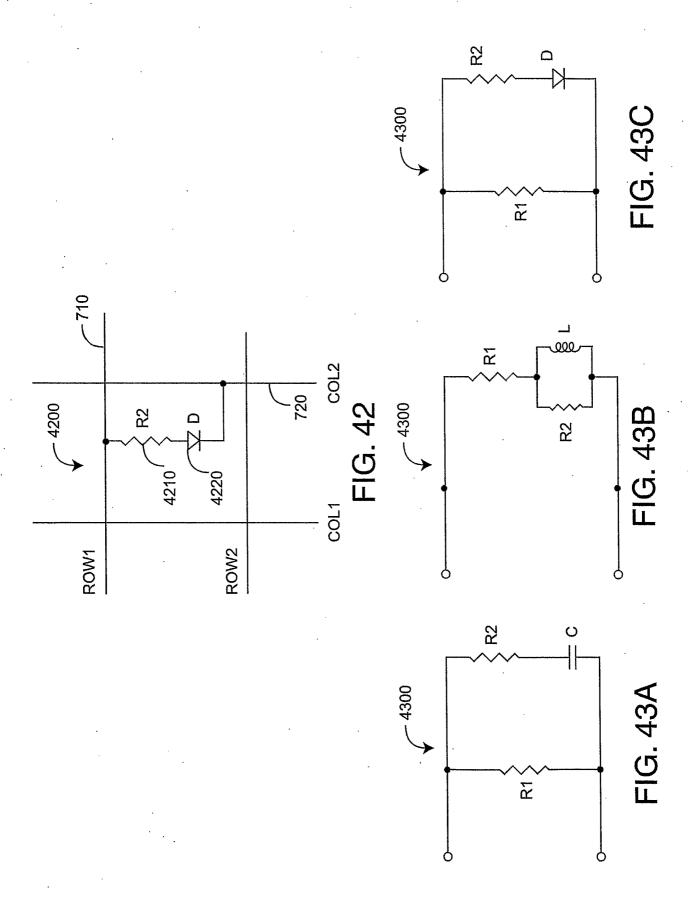


FIG. 41C



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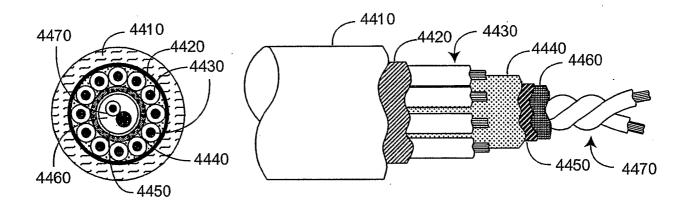


FIG. 44A

FIG. 44B

