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(54) Titre : METHODE DE TRAITEMENT DU CANCER A L'AIDE D'UN INHIBITEUR DE BCL-2 EN ASSOCIATION AVEC UNE RADIOIMMUNOTHERAPIE A EMISSION ALPHA

(54) Title: METHOD FOR TREATING CANCER USING A BCL-2 INHIBITOR IN CONJUNCTION WITH AN ALPHA-EMITTING RADIOIMMUNOTHERAPEUTIC

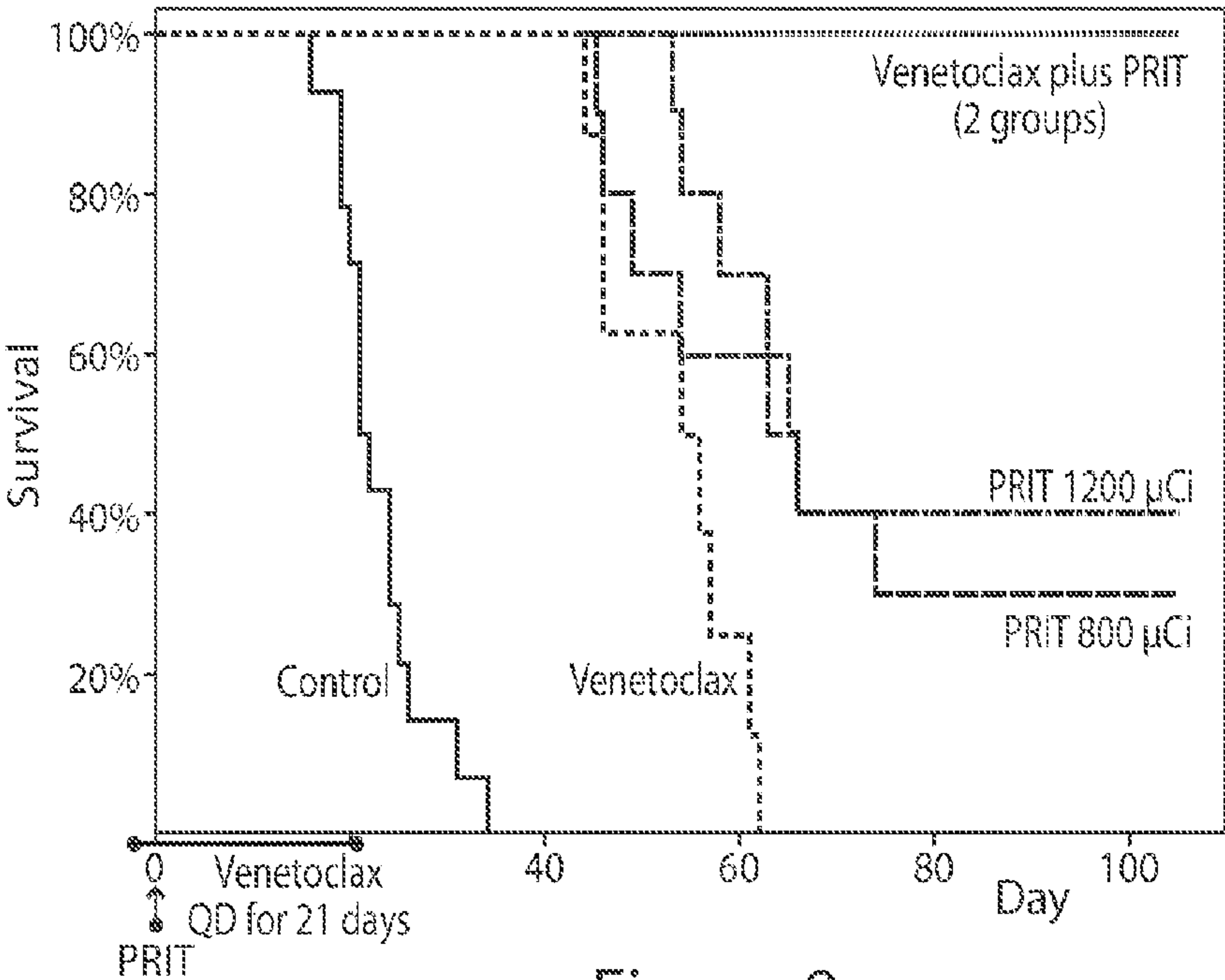


Figure 8

(57) **Abrégé/Abstract:**
This invention provides a method for treating a subject afflicted with cancer, comprising administering to the subject (i) a BCL-2 inhibitor in conjunction with (ii) an alpha-emitting isotope-labeled agent that targets cancer cells in the subject, wherein the amounts of the BCL-2 inhibitor and labeled agent, when administered in conjunction with one another, are therapeutically effective. This invention also provides a method for inducing the death of a cancer cell, comprising contacting the cell with (i) a BCL-2 inhibitor in conjunction with (ii) an alpha-emitting isotope-labeled agent that targets the cancer cell, wherein the amounts of BCL-2 inhibitor and labeled agent, when concurrently contacted with the cell, are effective to induce the cell's death.

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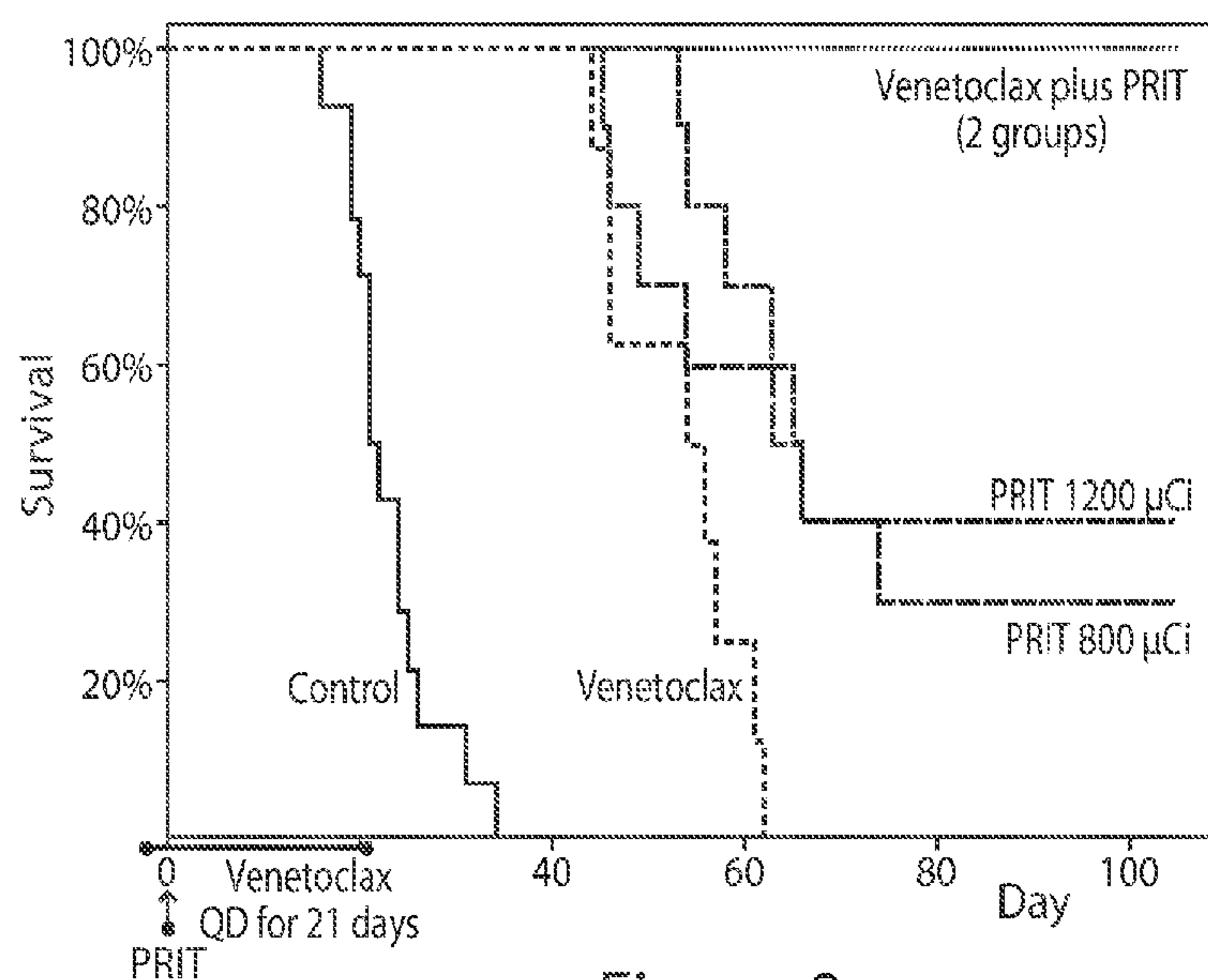


Figure 8

(57) Abstract: This invention provides a method for treating a subject afflicted with cancer, comprising administering to the subject (i) a BCL-2 inhibitor in conjunction with (ii) an alpha-emitting isotope-labeled agent that targets cancer cells in the subject, wherein the amounts of the BCL-2 inhibitor and labeled agent, when administered in conjunction with one another, are therapeutically effective. This invention also provides a method for inducing the death of a cancer cell, comprising contacting the cell with (i) a BCL-2 inhibitor in conjunction with (ii) an alpha-emitting isotope-labeled agent that targets the cancer cell, wherein the amounts of BCL-2 inhibitor and labeled agent, when concurrently contacted with the cell, are effective to induce the cell's death.

[Continued on next page]



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METHOD FOR TREATING CANCER USING A BCL-2 INHIBITOR IN
CONJUNCTION WITH AN ALPHA-EMITTING
RADIOIMMUNOTHERAPEUTIC

This application claims the benefit of U.S. Provisional Application No.
62/491,803, filed April 28, 2017, the contents of which are incorporated herein
by reference.

Throughout this application, various publications are cited. The disclosure of
these publications is hereby incorporated by reference into this application to
describe more fully the state of the art to which this invention pertains.

Field of the Invention

The present invention relates to treating a subject afflicted with cancer using a
therapeutically effective regimen of a BCL-2 inhibitor in conjunction with an
alpha-emitting isotope-labeled agent that targets cancer cells in the subject.

Background of the Invention

BCL-2 Inhibitors

BCL-2 inhibitors have potential for treating malignancies. One such BCL-2
inhibitor is venetoclax, a drug that has been approved for treating chronic
lymphocytic leukemia ("CLL") (1). Venetoclax binds to the BH3-binding
groove of BCL-2, displacing pro-apoptotic proteins like BIM to initiate
mitochondrial outer membrane permeabilization ("MOMP"), the release of
cytochrome c, and caspase activation, ultimately resulting in programmed
cancer cell death (i.e., apoptosis) (2).

Apoptosis is a mechanism of cell death in cancer cells, in addition to necrosis and autophagy (3). Ideally, by changing the balance between pro-apoptotic and anti-apoptotic stimuli, venetoclax would facilitate programmed cell death of cancer cells and thus improve cancer patient outcomes.

5

However, apoptosis is a complex pathway. Cancer cells can develop various mechanisms to circumvent and/or abrogate a given treatment strategy intended to cause apoptotic death (4) (as presented in Figure 1 of that reference). For example, X-linked XIAP can abrogate the blocking of BCL-2.

10 XIAP is a well-characterized inhibitor of apoptosis proteins (IAPs) (5). Indeed, the majority of human cancers harbor high levels of IAPs such as XIAP (6).

Other possible mechanisms of circumventing the effect of BCL-2 inhibitors can be seen in Figure 7. These include blocking activation of caspase 8 to
15 prevent the downstream activity of venetoclax on the BAX/BCL-2 axis. Also, stimulating or un-blocking one part of the apoptotic pathway may not be sufficient to cause apoptosis, as pro-apoptotic stimuli are still needed to trigger an apoptotic pathway (7), (8).

20 Consequently, not all cancer cells respond to BCL-2 inhibitors. In one venetoclax trial, for example, the complete response rate (including complete responses with incomplete marrow recovery) was 7.5%, even though a majority of patients (79.4%) had some level of response to venetoclax (2). In addition, venetoclax has a significant myelosuppressive effect on neutrophils,
25 with 40% of patients experiencing grade 3 and/or 4 neutropenia (2).

Radiation

Radiation is a recognized way to treat cancer. It is known that cellular effects
30 of radiation include cell cycle arrest, mutation, apoptosis, necrosis and autophagy (9). Radiation-related mediators of cellular damage include: (i) direct LED (linear energy deposition); (ii) ROS (reactive oxygen species); and (iii) RNS (reactive nitrogen species) (9).

These mediators lead to cell damage/kill/arrest via the following mechanisms:

- (i) DNA damage (9) (e.g., double-strand DNA breaks (most efficient), single-strand DNA breaks (less efficient, repairable), DNA base damage (least efficient, repairable), and DNA crosslinks); (ii) direct effects on the apoptotic cascade (e.g., direct activation of caspases, and damage to IAPs) (10); and (iii) bystander effects (i.e., damage or killing of cells not directly damaged by radiation, which damage or killing occurs through mediation via gap junction communication and/or cytokines from target cells) (11).

10 *The Unpredictability of Combination Therapies*

In a mouse xenograft model of venetoclax and radiation synergy (12), mice treated with a combination of venetoclax and ^{90}Y -based radioimmunotherapy had better survival rates compared to mice treated with either venetoclax or the radioimmunotherapy alone. Survival outcomes in xenografted mouse cohorts are shown in Figure 8.

Importantly, however, these mouse results may not be applicable to humans. Indeed, there are various factors that could render infeasible the treatment of cancer in humans using radiation in conjunction with venetoclax.

One such factor is oxygenation. Xenografted mice had small tumor masses in Fred Hutchinson Cancer Research Center experiments. Diffuse large B-cell lymphoma ("DLBCL") tumor xenografts were treated at a volume of 50 mm^3 , implying a tumor diameter of under 0.5 cm. For DLBCL patients in an MD Anderson study, ~25% of patients had tumor diameters greater than 7 cm (13). In large xenografted tumors in rats, it was found that in tumors larger than 3.5 cm^3 , baseline hypoxia was greater than 80%, while tumors smaller than 2.5 cm^3 had baseline hypoxia of ~20% (14). Hypoxia confers resistance to irradiation by lowering the creation of ROS (15), (16), (17). High tumor burden with hypoxic areas in human disease would significantly abrogate beta radiation-induced ROS and RNS.

Another such factor is the range of feasible dose levels. In mouse experiments on candidate therapeutics, the mice typically receive doses of drug weight per body weight that cannot be applied to humans. For example, in the ^{90}Y / venetoclax combination experiment described above, mice were
5 treated with doses of 800 μCi and 1,200 μCi per mouse (whereby 800 μCi was used in combination with venetoclax). Eight hundred μCi in a mouse would correspond to 3,000 mCi in an average human. By comparison, Zevalin[®] (^{90}Y -RIT, ibritumomab tiuxetan) can be administered to patients at a dose not exceeding 32 mCi (18).

10

There remains a need for a cancer therapy that solves the problems seen with BCL-2 inhibitors such as venetoclax and radiation therapies such as ^{90}Y -based therapies.

15 **Summary of the Invention**

This invention provides a method for treating a subject afflicted with cancer, comprising administering to the subject (i) a BCL-2 inhibitor in conjunction with (ii) an alpha-emitting isotope-labeled agent that targets cancer cells in the
20 subject, wherein the amounts of the BCL-2 inhibitor and labeled agent, when administered in conjunction with one another, are therapeutically effective.

This invention also provides a method for treating a human subject afflicted with acute myeloid leukemia, comprising administering to the subject (i)
25 venetoclax in conjunction with (ii) ^{225}Ac -labeled HuM195, wherein the amounts of venetoclax and ^{225}Ac -labeled HuM195, when administered in conjunction with one another, are therapeutically effective.

This invention further provides a method for inducing the death of a cancer
30 cell, comprising contacting the cell with (i) a BCL-2 inhibitor in conjunction with (ii) an alpha-emitting isotope-labeled agent that targets the cancer cell, wherein the amounts of BCL-2 inhibitor and labeled agent, when concurrently contacted with the cell, are effective to induce the cell's death.

Finally, this invention also provides a method for inducing the death of an acute myeloid leukemic cell, comprising contacting the cell with (i) venetoclax in conjunction with (ii) ^{225}Ac -labeled HuM195, wherein the amounts of venetoclax and ^{225}Ac -labeled HuM195, when concurrently contacted with the cell, are effective to induce the cell's death.

Brief Description of the Figures

Figure 1

10

This figure shows a schematic diagram of the expression plasmids for HuM195. The humanized VL and VH exons of HuM195 are flanked by XbaI sites. The VL exon was inserted into mammalian expression vector pVk, and the VH exon into pVg1 (Co, et al., J. Immunol. 148:1149-1154, 1992).

15

Figure 2

This figure shows the complete sequence of the HuM195 light chain gene cloned in pVk between the XbaI and BamHI sites. The nucleotide number indicates its position in the plasmid pVk-HuM195. The VL and CK exons are translated in single letter code; the dot indicates the translation termination codon. The mature light chain begins at the double-underlined aspartic acid (D). The intron sequence is in italics. The polyA signal is underlined.

25 Figure 3

This figure shows the complete sequence of the HuM195 heavy chain gene cloned in pVg1 between the XbaI and BamHI sites. The nucleotide number indicates its position in the plasmid pVg1-HuM195. The VH, CH1, H, CH2 and CH3 exons are translated in single letter code; the dot indicates the translation termination codon. The mature heavy chain begins at the double-underlined glutamine (Q). The intron sequences are in italics. The polyA signal is underlined.

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Figure 4

This figure shows the structure of ^{225}Ac -Lintuzumab (^{225}Ac -HuM195).

5 Figure 5

This figure shows a flowchart for the production of ^{225}Ac -HuM195.

10 Figure 6

This figure shows a dosing protocol for ^{225}Ac -Lintuzumab (^{225}Ac -HuM195) treatment of AML.

15 Figure 7

This figure shows a schematic of apoptotic cell death and mechanisms of cancer cells resistance to apoptosis (modified from (4)).

20 Figure 8

This figure shows a diagram of survival of xenografted mice treated with venetoclax alone, targeted beta radioimmunotherapy alone, and a combination of venetoclax and targeted beta radioimmunotherapy. In Rec-1-bearing mice, venetoclax had no effect alone ($p = .12$), 800 μCi PRIT lengthened survival time 111% beyond controls ($p = .0001$), while the combination extended survival 483% beyond controls and cured 40% ($p = .001$, combination group > PRIT alone). In the U2932 xenograft model, venetoclax alone doubled survival time compared to controls ($p < .0001$) and 800 μCi PRIT alone doubled survival and cured 30%. Combination treatments cured 100% (12).

Figure 9

This figure shows a comparison between beta and alpha radiation mechanisms of apoptotic cell killing. As the figure shows, alpha radiation is significantly more potent than beta radiation (~700 times); causes more dsDNA breaks than beta radiation; does not depend on tissue oxygenation and the cell division phase; and can overcome cellular resistance to beta and gamma radiation and cytotoxic chemotherapy. These findings are collectively supported by (9)-(11) and (22)-(24).

Detailed Description of the Invention

This invention provides methods for treating a subject afflicted with cancer. These methods comprise administering to the subject two types of agents in conjunction with one another. The first type of agent is a BCL-2 inhibitor such as venetoclax. The second type is an alpha-emitting isotope-labeled agent, such as ^{225}Ac -labeled HuM195, that targets cancer cells in the subject.

Definitions

In this application, certain terms are used which shall have the meanings set forth as follows.

5

As used herein, “administer”, with respect to an agent, means to deliver the agent to a subject’s body via any known method. Specific modes of administration include, without limitation, intravenous, oral, sublingual, transdermal, subcutaneous, intraperitoneal, intrathecal and intra-tumoral
10 administration.

In addition, in this invention, the various antibodies and other antigen-targeting agents used can be formulated using one or more routinely used pharmaceutically acceptable carriers. Such carriers are well known to those
15 skilled in the art. For example, injectable drug delivery systems include solutions, suspensions, gels, microspheres and polymeric injectables, and can comprise excipients such as solubility-altering agents (e.g., ethanol, propylene glycol and sucrose) and polymers (e.g., polycaprylactones and PLGA's).

20

As used herein, the term “agent”, whether in reference to a BCL-2 inhibitor or an alpha-emitting isotope-labeled agent, can be any type of compound or composition useful for such purpose. Types of agents include, without
25 limitation, antibodies, other protein-based drugs, peptides, nucleic acids, carbohydrates and small molecules drugs.

As used herein, the term “alpha-emitting isotope” includes, without limitation, ^{225}Ac , ^{213}Bi and ^{213}Po . Methods for affixing an alpha-emitting isotope to an antibody (i.e., “labeling” an antibody with an alpha-emitting isotope) are well
30 known.

As used herein, the term “antibody” includes, without limitation, (a) an immunoglobulin molecule comprising two heavy chains and two light chains and which recognizes an antigen; (b) polyclonal and monoclonal

immunoglobulin molecules; (c) monovalent and divalent fragments thereof, and (d) bi-specific forms thereof. Immunoglobulin molecules may derive from any of the commonly known classes, including but not limited to IgA, secretory IgA, IgG and IgM. IgG subclasses are also well known to those in the art and include, but are not limited to, human IgG1, IgG2, IgG3 and IgG4. Antibodies can be both naturally occurring and non-naturally occurring. Furthermore, antibodies include chimeric antibodies, wholly synthetic antibodies, single chain antibodies, and fragments thereof. Antibodies may be human, humanized or nonhuman.

10

As used herein, an “anti-CD33 antibody” is an antibody that binds to any available epitope of CD33. In one embodiment, the anti-CD33 antibody binds to the epitope recognized by the antibody HuM195.

15

As used herein, the term “burden”, when used in connection with a cancerous cell, means quantity. So, a cancerous cell “burden” means the quantity of cancerous cells. Cancerous cells have a burden with respect to their tissue of origin (i.e., the primary site of disease), such as the “bone marrow blast burden” in the case of AML. Cancerous cells also have a burden with respect to one or more tissues other than those of origin, such as the blast burden in blood, liver and spleen in the case of AML. The term “peripheral burden” relates to such cells. The peripheral burden of cancerous cells, such as blasts in the case of AML, can be measured in different ways with different outcomes. For example, in the case of AML, the “peripheral blast burden” can be measured as the total blast population outside of the bone marrow, or the total blast population of the blood, spleen and liver combined, or simply the blast population of the blood as measured in cells per unit volume. As used herein in connection with AML and other cancers originating in the bone marrow, and unless stated otherwise, the term “peripheral cancerous cell burden” (e.g., peripheral blast burden) refers to the cancerous cell population of the blood as measured in cells per unit volume (e.g., cells/ μ l). This blood-based measurement is a useful proxy for the more cumbersome measurements of spleen and liver burdens, for example.

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Herein, a peripheral cancerous cell burden in a subject is “high” if, when the subject is administered an agent (e.g., an antibody) targeting a hematologic malignancy-associated antigen at the maximum safe dose, the agent does not reach the primary site of disease in a sufficient amount to bind to more than
5 90% of its target antigens at that site. Conversely, a peripheral cancerous cell burden in a subject is “low” if, when the subject is administered an agent (e.g., an antibody) targeting a hematologic malignancy-associated antigen at the maximum safe dose, the agent reaches the primary site of disease in a sufficient amount to bind to more than 90% of its target antigens at that site.

10 In the case of AML, examples of low peripheral blast burden are those yielding blood blast burdens at or below 1,000 blast cells/ μ l, at or below 500 blast cells/ μ l, at or below 400 blast cells/ μ l, at or below 300 blast cells/ μ l, at or below 200 blast cells/ μ l, at or below 100 blast cells/ μ l, and at or below 50 blast cells/ μ l.

15

A “hematologic malignancy”, also known as a blood cancer, is a cancer that originates in blood-forming tissue, such as the bone marrow or other cells of the immune system. Hematologic malignancies include, without limitation, leukemias (such as AML, acute promyelocytic leukemia, acute lymphoblastic
20 leukemia, acute mixed lineage leukemia, chronic myeloid leukemia, chronic lymphocytic leukemia, hairy cell leukemia and large granular lymphocytic leukemia), myelodysplastic syndrome (MDS), myeloproliferative disorders (polycythemia vera, essential thrombocytosis, primary myelofibrosis and chronic myeloid leukemia), lymphomas, multiple myeloma, and MGUS and
25 similar disorders.

As used herein, a “hematologic malignancy-associated antigen” can be, for example, a protein and/or carbohydrate marker found exclusively or predominantly on the surface of a cancer cell associated with that particular
30 malignancy. Examples of hematologic malignancy-associated antigens include, without limitation, CD20, CD33, CD38, CD45, CD52, CD123 and CD319.

The antibody “HuM195” (also known as lintuzumab) is known, as are methods of making it. Likewise, methods of labeling HuM195 with ²²⁵Ac are known. These methods are exemplified, for example, in Scheinberg, et al., U.S. Patent No. 6,683,162. This information is also exemplified in the examples and figures below.

As used herein, administering to a subject a BCL-2 inhibitor “in conjunction with” an alpha-emitting isotope-labeled agent that targets cancer cells in the subject means administering the BCL-2 inhibitor before, during or after administration of the labeled agent. This administration includes, without limitation, the following scenarios: (i) the BCL-2 inhibitor is administered first (e.g., orally once per day for 21 days, 28 days, 35 days, 42 days, 49 days, or a longer period during which the cancer being treated does not progress and during which the BCL-2 inhibitor does not cause unacceptable toxicity), and the labeled agent is administered second (e.g., intravenously in a single dose or a plurality of doses over a period of weeks); (ii) the BCL-2 inhibitor is administered concurrently with the labeled agent (e.g., the BCL-2 inhibitor is administered orally once per day for n days, and the labeled agent is administered intravenously in a single dose on one of days 2 through n-1 of the BCL-2 inhibitor regimen); (iii) the BCL-2 inhibitor is administered concurrently with the labeled agent (e.g., the BCL-2 inhibitor is administered orally for a duration of greater than one month (e.g., orally once per day for 35 days, 42 days, 49 days, or a longer period during which the cancer being treated does not progress and during which the BCL-2 inhibitor does not cause unacceptable toxicity), and the labeled agent is administered intravenously in a single dose on a day within the first month of the BCL-2 inhibitor regimen); and (iv) the labeled agent is administered first (e.g., intravenously in a single dose or a plurality of doses over a period of weeks), and the BCL-2 inhibitor is administered second (e.g., orally once per day for 21 days, 28 days, 35 days, 42 days, 49 days, or a longer period during which the cancer being treated does not progress and during which the BCL-2 inhibitor does not cause unacceptable toxicity). Additional permutations are provided below in the Examples section.

As used herein, the term “subject” includes, without limitation, a mammal such as a human, a non-human primate, a dog, a cat, a horse, a sheep, a goat, a cow, a rabbit, a pig, a rat and a mouse. Where the subject is human, the subject can be of any age. For example, the subject can be 60 years or older, 65 or older, 70 or older, 75 or older, 80 or older, 85 or older, or 90 or older. Alternatively, the subject can be 50 years or younger, 45 or younger, 40 or younger, 35 or younger, 30 or younger, 25 or younger, or 20 or younger. For a human subject afflicted with AML, the subject can be newly diagnosed, or relapsed and/or refractory, or in remission.

10

As used herein, a “sub-saturating dose” of an agent targeting an antigen (e.g., CD33) or marker (e.g., BCL-2) is one that introduces into the subject’s body fewer target antigen-binding sites (e.g., Fab’s) than there are target antigens, or fewer target marker-binding sites (e.g., venetoclax molecules) than there are target markers, as applicable. By way of example, for an anti-CD33 antibody, a sub-saturating dose is one that introduces into the subject’s body fewer CD33-binding sites than there are CD33 molecules. In one embodiment, a sub-saturating dose of an agent targeting a hematologic malignancy-associated antigen is one where the ratio of target antigen-binding sites to target antigens is less than or equal to 9:10. In another embodiment, the ratio of target antigen-binding sites to target antigens is less than or equal to 1:2, less than or equal to 1:5, less than or equal to 1:10, less than or equal to 1:20, or less than or equal to 1:100. By way of additional example, for a BCL-2 inhibitor, a sub-saturating dose is one that introduces into the subject’s body fewer BCL-2-binding sites than there are BCL-2 proteins. In one embodiment, a sub-saturating dose of a BCL-2 inhibitor is one where the ratio of inhibitor to BCL-2 protein is less than or equal to 9:10. In another embodiment, the ratio of target antigen-binding sites to target antigens is less than or equal to 1:2, less than or equal to 1:5, less than or equal to 1:10, less than or equal to 1:20, or less than or equal to 1:100. In a further embodiment, a “sub-saturating dose” of a BCL-2 inhibitor (e.g., venetoclax) is a dose lower than the inhibitor’s maximum approved dose in humans (e.g., below 400 mg per day, below 300 mg per day, below 200 mg

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per day, below 100 mg per day, below 50 mg per day, or below 10 mg per day).

For an agent such as an antibody labeled with an alpha-emitting isotope, the majority of the drug administered to a subject typically consists of non-labeled antibody, with the minority being the labeled antibody. Thus, in one embodiment, a sub-saturating dose of an agent targeting a hematologic malignancy-associated antigen is one where the ratio of total (i.e., labeled and unlabeled) target antigen-binding sites to target antigens is less than or equal to 9:10 (and can be less than or equal to 1:2, less than or equal to 1:5, less than or equal to 1:10, less than or equal to 1:20, or less than or equal to 1:100). In another embodiment, a sub-saturating dose of an agent targeting a hematologic malignancy-associated antigen is one where the ratio of labeled target antigen-binding sites to target antigens is less than or equal to 9:10 (and can be less than or equal to 1:2, less than or equal to 1:5, less than or equal to 1:10, less than or equal to 1:20, or less than or equal to 1:100).

Sub-saturating doses of labeled agent used in connection with this invention include, for example, a single administration, and two or more administrations (i.e., fractions). The amount administered in each dose can be measured, for example, by labeled radiation activity (e.g., $\mu\text{Ci/kg}$) or antibody weight (e.g., $\mu\text{g/kg}$ or $\mu\text{g/m}^2$). In the case of $^{225}\text{Ac-HuM195}$ for treating AML, human dosing regimens include the following, without limitation: (i) $2 \times < 0.5 \mu\text{Ci/kg}$, $2 \times 0.5 \mu\text{Ci/kg}$, $2 \times 1.0 \mu\text{Ci/kg}$, $2 \times 1.5 \mu\text{Ci/kg}$, or $2 \times 2.0 \mu\text{Ci/kg}$, where the fractions are administered one week apart; (ii) $< 0.5 \mu\text{Ci/kg}$, or from $0.5 \mu\text{Ci/kg}$ to $10 \mu\text{Ci/kg}$; (iii) $2 \times < 7.5 \mu\text{g/kg}$, $2 \times 7.5 \mu\text{g/kg}$, $2 \times 10 \mu\text{g/kg}$, or $2 \times 12.5 \mu\text{g/kg}$, where the fractions are administered one week apart; or (iv) $< 15 \mu\text{g/kg}$, or from $15 \mu\text{g/kg}$ to $50 \mu\text{g/kg}$.

As used herein, an amount of BCL-2 inhibitor and an amount of alpha-emitting isotope-labeled agent that targets cancer cells in the subject, when administered in conjunction with each other, are “therapeutically effective” if the subject is treated.

As used herein, “treating” a subject afflicted with a disorder shall include, without limitation, (i) slowing, stopping or reversing the disorder's progression, (ii) slowing, stopping or reversing the progression of the disorder's symptoms, (iii) reducing the likelihood of the disorder's recurrence, and/or (iv) reducing the likelihood that the disorder's symptoms will recur. In the preferred embodiment, treating a subject afflicted with a disorder means (i) reversing the disorder's progression, ideally to the point of eliminating the disorder, and/or (ii) reversing the progression of the disorder's symptoms, ideally to the point of eliminating the symptoms, and/or (iii) reducing or eliminating the likelihood of relapse (i.e., consolidation, which is a common goal of post-remission therapy for AML and, ideally, results in the destruction of any remaining leukemia cells).

The treatment of hematologic malignancy, such as the treatment of AML, can be measured according to a number of clinical endpoints. These include, without limitation, survival time (such as weeks, months or years of improved survival time, e.g., one, two or more months' of additional survival time), and response status (such as complete remission (CR), complete remission with incomplete platelet recovery (CRp), complete remission with incomplete peripheral blood recovery (CRi), morphologic leukemia-free state (MLFS) and partial remission (PR)).

In one embodiment, treatment of hematologic malignancy, such as the treatment of AML, can be measured in terms of remission. Included here are the following non-limiting examples. (1) Morphologic complete remission (“CR”): ANC \geq 1,000/mcl, platelet count \geq 100,000/mcl, $<$ 5% bone marrow blasts, no Auer rods, no evidence of extramedullary disease. (No requirements for marrow cellularity, hemoglobin concentration). (2) Morphologic complete remission with incomplete blood count recovery (“CRi”): Same as CR but ANC may be $<$ 1,000/mcl and/or platelet count $<$ 100,000/mcl. (3) Partial remission (PR): ANC \geq 1,000/mcl, platelet count $>$ 100,000/mcl, and at least a 50% decrease in the percentage of marrow aspirate blasts to 5-25%, or marrow blasts $<$ 5% with persistent Auer rods. These criteria and others are known, and are described, for example, in

SWOG Oncology Research Professional (ORP) Manual Volume I, Chapter 11A, Leukemia (2014).

Embodiments of the Invention

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This invention employs the use of alpha particles. These particles induce apoptosis in target cells, e.g., leukemic cells (10), (19). Alpha-emitters and beta-emitters induce apoptosis with different efficiencies at comparable activities in leukemic cells (10). Alpha particles can overcome doxorubicin-
10 resistance, CD95-resistance, and radio-resistance to beta-irradiation and gamma-irradiation in leukemic cells (10). The particles induce apoptosis via: (i) double-strand DNA breaks (20), (21); (ii) activation of caspases; (iii) the fact that [^{213}Bi]anti-CD45 activates caspases 2, 3, 8 and 9 through the mitochondrial pathway independent of the CD95 ligand/receptor system (10),
15 (19); and (iv) inactivation of XIAP and Bcl-XL (19).

Specifically, this invention provides a first therapeutic method. This first method is for treating a subject afflicted with cancer, comprising administering to the subject (i) a BCL-2 inhibitor in conjunction with (ii) an alpha-emitting
20 isotope-labeled agent that targets cancer cells in the subject, wherein the amounts of the BCL-2 inhibitor and labeled agent, when administered in conjunction with one another, are therapeutically effective.

This invention also provides a second therapeutic method. This second
25 method is for treating a human subject afflicted with acute myeloid leukemia, comprising administering to the subject (i) venetoclax in conjunction with (ii) ^{225}Ac -labeled HuM195, wherein the amounts of venetoclax and ^{225}Ac -labeled HuM195, when administered in conjunction with one another, are therapeutically effective.

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Preferably in the first and second therapeutic methods, the subject is human. In one embodiment of the first and second therapeutic methods, the cancer is a hematologic malignancy, and preferably is a leukemia such as acute myeloid leukemia.

In the preferred embodiment of the first and second therapeutic methods, the BCL-2 inhibitor is venetoclax. Also in the preferred embodiment of the first and second therapeutic methods, the alpha-emitting isotope-labeled agent is an anti-CD33 antibody labeled with an alpha-emitting isotope, ideally ^{225}Ac -labeled HuM195. In these methods, the BCL-2 inhibitor, the labeled agent, or both, are preferably administered (i) in sub-saturating doses, and/or (ii) in doses that are less than (and/or shorter duration than) those presently prescribed on their respective labels. Also in these methods, the subject's peripheral blast burden is preferably low, and the methods preferably do not cause unacceptable levels of neutropenia.

This invention provides a third method. This third method is for inducing the death of a cancer cell, comprising contacting the cell with (i) a BCL-2 inhibitor in conjunction with (ii) an alpha-emitting isotope-labeled agent that targets the cancer cell, wherein the amounts of BCL-2 inhibitor and labeled agent, when concurrently contacted with the cell, are effective to induce the cell's death.

Preferably, the cancer cell is a human cancer cell. In one embodiment, the cancer cell is a hematologic cell, and preferably is a leukemic cell such as an acute myeloid leukemic cell.

In the preferred embodiment, the BCL-2 inhibitor is venetoclax. Also in the preferred embodiment, the alpha-emitting isotope-labeled agent is an anti-CD33 antibody labeled with an alpha-emitting isotope, ideally ^{225}Ac -labeled HuM195.

This invention also provides a fourth method. This fourth method is for inducing the death of an acute myeloid leukemic cell, comprising contacting the cell with (i) venetoclax in conjunction with (ii) ^{225}Ac -labeled HuM195, wherein the amounts of venetoclax and ^{225}Ac -labeled HuM195, when concurrently contacted with the cell, are effective to induce the cell's death.

Finally, this invention provides two articles of manufacture. The first article comprises (i) a BCL-2 inhibitor (e.g., venetoclax) and (ii) a label instructing the user (e.g., the patient or healthcare provider) to treat a subject afflicted with cancer (e.g., acute myeloid leukemia) by administering the BCL-2 inhibitor to the subject in conjunction with an alpha-emitting isotope-labeled agent that targets cancer cells in the subject (e.g., ^{225}Ac -labeled HuM195), wherein the amounts of the BCL-2 inhibitor and labeled agent, when administered in conjunction with one another, are therapeutically effective. The second article comprises (i) an alpha-emitting isotope-labeled agent that targets cancer cells (e.g., ^{225}Ac -labeled HuM195) and (ii) a label instructing the user to treat a subject afflicted with cancer (e.g., acute myeloid leukemia) by administering the labeled agent to the subject in conjunction with a BCL-2 inhibitor (e.g., venetoclax), wherein the amounts of the BCL-2 inhibitor and labeled agent, when administered in conjunction with one another, are therapeutically effective.

Wherever applicable, the methods of the subject invention may also be performed using pre-targeted radioimmunotherapy (PRIT). A PRIT-based method comprises the steps of (i) administering a monoclonal antibody labeled with a marker (e.g., streptavidin), (ii) then administering a suitable clearing agent (e.g., a biotin galactose clearing agent), and (iii) administering an alpha-emitting isotope-labeled agent that specifically binds to the marker (e.g., ^{225}Ac -labeled biotin). Therefore, the various embodiments of the invention relating to non-PRIT-based methods for administering an alpha-emitting isotope-labeled agent apply, *mutatis mutandis*, to these PRIT-based methods.

This invention will be better understood by reference to the examples which follow, but those skilled in the art will readily appreciate that the specific examples detailed are only illustrative of the invention as described more fully in the claims which follow thereafter.

Examples

Example 1 – Structure of ^{225}Ac -Lintuzumab (^{225}Ac -HuM195)

5 ^{225}Ac -Lintuzumab includes three key components; humanized monoclonal antibody HuM195 (generic name, lintuzumab), the alpha-emitting radioisotope ^{225}Ac , and the bi-functional chelate 2-(p-isothiocyanatobenzyl)-1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid (p-SCN-Bn-DOTA). As depicted in Figure 4, HuM195 is radiolabeled using the bi-functional chelate p-SCN-Bn-DOTA that binds to ^{225}Ac and that is covalently attached to the IgG
10 via a lysine residue on the antibody.

Example 2 – p-SCN-Bn-DOTA

15 DOTA, 2-(4-Isothiocyanatobenzyl)-1,4,7,10-tetraazacyclododecane tetraacetic acid (Macrocylics item code B205-GMP) is synthesized by a multi-step organic synthesis that is fully described in U.S. Patent No. 4,923,985.

Example 3 – Preparation of ^{225}Ac -Lintuzumab (^{225}Ac -HuM195)

20 The procedure for preparing ^{225}Ac -Lintuzumab is based on the method described by Michael R. McDevitt, "Design and synthesis of ^{225}Ac radioimmuno-pharmaceuticals, Applied Radiation and Isotope", 57 (2002), 841-847. The procedure involves radiolabeling the bi-functional chelate, p-SCN-Bn-DOTA, with the radioisotope ^{225}Ac , followed by binding of the radiolabeled p-SCN-Bn-DOTA to the antibody (HuM195). The construct, ^{225}Ac -p-SCN-Bn-DOTA-HuM195, is purified using 10 DG size exclusion chromatography and eluted with 1% human serum albumin (HSA). The resulting drug product, Ac^{225} -Lintuzumab, is then passed through a 0.2 μm
25
30 sterilizing filter.

Example 4 – Process Flow for Preparation of ^{225}Ac -Lintuzumab (^{225}Ac -HuM195)

The procedure, shown in Figure 5, begins with confirming the identity of all components and the subsequent QC release of the components to production. The ^{225}Ac is assayed to confirm the level of activity and is reconstituted to the desired activity concentration with hydrochloric acid. A vial of lyophilized p-SCN-Bn-DOTA is reconstituted with metal-free water to a concentration of 10 mg/mL. To the actinium reaction vial, 0.02 ml of ascorbic acid solution (150 mg/mL) and 0.05 ml of reconstituted p-SCN-Bn-DOTA are added and the pH adjusted to between 5 and 5.5 with 2M tetramethylammonium acetate (TMAA). The mixture is then heated at $55 \pm 4^\circ\text{C}$ for 30 minutes.

To determine the labeling efficiency of the ^{225}Ac -p-SCN-Bn-DOTA, an aliquot of the reaction mixture is removed and applied to a 1 ml column of Sephadex C25 cation exchange resin. The product is eluted in 2-4 ml fractions with a 0.9% saline solution. The fraction of ^{225}Ac activity that elutes is ^{225}Ac -p-SCN-Bn-DOTA and the fraction that is retained on the column is un-chelated, unreactive ^{225}Ac . Typically, the labeling efficiency is greater than 95%.

To the reaction mixture, 0.22 ml of previously prepared HuM195 in DTPA (1 mg HuM195) and 0.02 ml of ascorbic acid are added. The DTPA is added to bind any trace amounts of metals that may compete with the labeling of the antibody. The ascorbic acid is added as a radio-protectant. The pH is adjusted with carbonate buffer to pH 8.5-9. The mixture is heated at $37 \pm 3^\circ\text{C}$ for 30 minutes.

The final product is purified by size exclusion chromatography using 10DG resin and eluted with 2 ml of 1% HSA. Typical reaction yields are 10%.

Example 5 – Venetoclax And Its Normal Dosing Regimen

Venetoclax is sold by Genentech (San Francisco, CA) under the brand name Venclexta™. According to the FDA's Venclexta™ label, this drug "is a BCL-2 inhibitor indicated for the treatment of patients with chronic lymphocytic leukemia (CLL) with 17p deletion ... who have received at least one prior

therapy.” Venclexta™ is sold in tablet form at 10 mg, 50 mg and 100 mg. Therapy is to be initiated “at 20 mg once daily for 7 days, followed by a weekly ramp-up dosing schedule to the recommended daily dose of 400 mg.” The ramp-up dosing schedule is as follows: week 1, 20mg/day; week 2, 50mg/day; week 3, 100mg/day; week 4, 200mg/day; and week 5 and beyond, 400mg/day. This dosing regimen is referred to herein as the “normal” human dosing regimen for venetoclax, regardless of the disorder treated. Any dosing regimen having a shorter duration (e.g., 21 days) or involving the administration of less venetoclax (e.g., 20mg/day for a total of 21 days) is referred to herein as a “reduced” human dosing regimen. The terms “normal” human dosing regimen and “reduced” human dosing regimen also apply, *mutatis mutandis*, to any other BCL-2 inhibitor with respect to its approved or otherwise customary dosing regimen.

Also envisioned is a “normal” murine dosing regimen and a “reduced” murine dosing regimen, each being commensurate with mouse body weight and tumor xenograft size. Moreover, the “normal” murine dosing regimen has a duration of at least 21 days.

Example 6 – ²²⁵Ac-HuM195 And Its Normal Dosing Regimen

In the case of ²²⁵Ac-HuM195, the “normal” human dosing regimen (regardless of the disorder treated), as this term is used herein, includes either of the following: (i) 2 x 2.0 μCi/kg, where the fractions are administered one week apart; and (ii) 4.0 μCi/kg when delivered in a single administration. Any dosing regimen involving the administration of less ²²⁵Ac-HuM195 (e.g., 2.0 μCi/kg when delivered in a single administration) is referred to herein as a “reduced” human dosing regimen (which may also be considered a sub-saturating dose). The terms “normal” human dosing regimen and “reduced” human dosing regimen also apply, *mutatis mutandis*, to any other alpha-emitting isotope-labeled agent with respect to its approved or otherwise customary dosing regimen.

Also envisioned is a “normal” murine dosing regimen and a “reduced” murine dosing regimen, each being commensurate with mouse body weight and tumor xenograft size.

5 Example 7 – Dosing Scenario I for ²²⁵Ac-HuM195 and Venetoclax

A human AML patient is treated according to the following regimen.

Venetoclax is orally administered according to its normal dosing regimen (i.e., for at least five weeks), followed by intravenous administration of ²²⁵Ac-
10 HuM195 according to its normal dosing regimen (either single or fractional administration). In one embodiment, the first (and only, if applicable) dose of ²²⁵Ac-HuM195 is administered on the same day as, or one day following, the last dose of venetoclax.

15 Also envisioned is the treatment of an experimental mouse model according to the treatment regimen in this scenario, whereby the appropriate dosing regimens are commensurate with mouse body weight and tumor xenograft size.

20 Example 8 – Dosing Scenario II for ²²⁵Ac-HuM195 and Venetoclax

A human AML patient is treated according to the following regimen.

Venetoclax is orally administered according to its normal dosing regimen (i.e., for at least five weeks), followed by intravenous administration of a reduced
25 dosing regimen of ²²⁵Ac-HuM195 (either single or fractional administration). In one embodiment, the first (and only, if applicable) dose of ²²⁵Ac-HuM195 is administered on the same day as, or one day following, the last dose of venetoclax. In another embodiment, the reduced dosing regimen of ²²⁵Ac-HuM195 is (i) 2 x 0.5 µCi/kg, 2 x 1.0 µCi/kg, or 2 x 1.5 µCi/kg, where the
30 fractions are administered one week apart; or (ii) 1 x 0.5 µCi/kg, 1 x 1.0 µCi/kg, 1 x 2.0 µCi/kg, or 1 x 3.0 µCi/kg, for a single administration.

Also envisioned is the treatment of an experimental mouse model according to the treatment regimen in this scenario, whereby the appropriate dosing

regimens are commensurate with mouse body weight and tumor xenograft size.

Example 9 – Dosing Scenario III for ^{225}Ac -HuM195 and Venetoclax

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A human AML patient is treated according to the following regimen.

Venetoclax is orally administered according to a reduced dosing regimen, followed by intravenous administration of the normal dosing regimen of ^{225}Ac -HuM195 (either single or fractional administration). In one embodiment, the first (and only, if applicable) dose of ^{225}Ac -HuM195 is administered on the same day as, or one day following, the last dose of venetoclax. In another embodiment, the reduced dosing regimen of venetoclax is one of the following: (i) 20 mg once daily for 7 days; (ii) 20 mg once daily for 14 days; (iii) 20 mg once daily for 21 days; (iv) 50 mg once daily for 7 days; (v) 50 mg once daily for 14 days; (vi) 50 mg once daily for 21 days; (vii) 100 mg once daily for 7 days; (viii) 100 mg once daily for 14 days; (ix) 100 mg once daily for 21 days; (x) 200 mg once daily for 7 days; (xi) 200 mg once daily for 14 days; (xii) 200 mg once daily for 21 days; (xiii) 400 mg once daily for 7 days; and (xiv) week 1 at 20mg/day, week 2 at 50mg/day and week 3 at 100mg/day.

20

Also envisioned is the treatment of an experimental mouse model according to the treatment regimen in this scenario, whereby the appropriate dosing regimens are commensurate with mouse body weight and tumor xenograft size.

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Example 10 – Dosing Scenario IV for ^{225}Ac -HuM195 and Venetoclax

A human AML patient is treated according to the following regimen.

Venetoclax is orally administered according to a reduced dosing regimen, followed by intravenous administration of a reduced dosing regimen of ^{225}Ac -HuM195 (either single or fractional administration). In one embodiment, the first (and only, if applicable) dose of ^{225}Ac -HuM195 is administered on the same day as, or one day following, the last dose of venetoclax. In another embodiment, (a) the reduced dosing regimen of venetoclax is one of (i) 20 mg

once daily for 7 days; (ii) 20 mg once daily for 14 days; (iii) 20 mg once daily for 21 days; (iv) 50 mg once daily for 7 days; (v) 50 mg once daily for 14 days; (vi) 50 mg once daily for 21 days; (vii) 100 mg once daily for 7 days; (viii) 100 mg once daily for 14 days; (ix) 100 mg once daily for 21 days; (x) 200 mg once daily for 7 days; (xi) 200 mg once daily for 14 days; (xii) 200 mg once daily for 21 days; (xiii) 400 mg once daily for 7 days; and (xiv) week 1 at 20mg/day, week 2 at 50mg/day and week 3 at 100mg/day; and (b) the reduced dosing regimen of ^{225}Ac -HuM195 is one of (i) 2 x 0.5 $\mu\text{Ci/kg}$, 2 x 1.0 $\mu\text{Ci/kg}$, or 2 x 1.5 $\mu\text{Ci/kg}$, where the fractions are administered one week apart; or (ii) 1 x 0.5 $\mu\text{Ci/kg}$, 1 x 1.0 $\mu\text{Ci/kg}$, 1 x 2.0 $\mu\text{Ci/kg}$, or 1 x 3.0 $\mu\text{Ci/kg}$, for a single administration.

Also envisioned is the treatment of an experimental mouse model according to the treatment regimen in this scenario, whereby the appropriate dosing regimens are commensurate with mouse body weight and tumor xenograft size.

Example 11 – Dosing Scenario V for ^{225}Ac -HuM195 and Venetoclax

A human AML patient is treated according to the following regimen. Venetoclax is orally administered according to its normal dosing regimen (i.e., for at least five weeks), and ^{225}Ac -HuM195 is intravenously administered according to its normal single dose regimen during the course of the venetoclax dosing regimen. In one embodiment, the single dose of ^{225}Ac -HuM195 is administered (a) on day 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 or 21 of the venetoclax dosing regimen, or (b) on the last day of, the penultimate day of, or 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 or 21 days prior to the last day of, the venetoclax dosing regimen.

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Also envisioned is the treatment of an experimental mouse model according to the treatment regimen in this scenario, whereby the appropriate dosing regimens are commensurate with mouse body weight and tumor xenograft size.

Example 12 – Dosing Scenario VI for ^{225}Ac -HuM195 and Venetoclax

A human AML patient is treated according to the following regimen.

Venetoclax is orally administered according to its normal dosing regimen (i.e.,

5 for at least five weeks), and ^{225}Ac -HuM195 is intravenously administered according to a reduced single dose regimen during the course of the venetoclax dosing regimen. In one embodiment, the single dose of ^{225}Ac -HuM195 is administered (a) on day 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 or 21 of the venetoclax dosing regimen, or (b) on the
10 last day of, the penultimate day of, or 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 or 21 days prior to the last day of, the venetoclax dosing regimen. In another embodiment, the reduced dosing regimen of ^{225}Ac -HuM195 is (i) 2 x 0.5 $\mu\text{Ci/kg}$, 2 x 1.0 $\mu\text{Ci/kg}$, or 2 x 1.5 $\mu\text{Ci/kg}$, where the fractions are administered one week apart; or (ii) 1 x 0.5 $\mu\text{Ci/kg}$, 1 x 1.0
15 $\mu\text{Ci/kg}$, 1 x 2.0 $\mu\text{Ci/kg}$, or 1 x 3.0 $\mu\text{Ci/kg}$, for a single administration.

Also envisioned is the treatment of an experimental mouse model according to the treatment regimen in this scenario, whereby the appropriate dosing regimens are commensurate with mouse body weight and tumor xenograft
20 size.

Example 13 – Dosing Scenario VII for ^{225}Ac -HuM195 and Venetoclax

A human AML patient is treated according to the following regimen.

25 Venetoclax is orally administered according to a reduced dosing regimen, and ^{225}Ac -HuM195 is intravenously administered according to its normal single dose regimen during the course of the venetoclax dosing regimen. In one embodiment, the single dose of ^{225}Ac -HuM195 is administered (a) on day 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 or 21 of the
30 venetoclax dosing regimen, or (b) on the last day of, the penultimate day of, or 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 or 21 days prior to the last day of, the venetoclax dosing regimen. In another embodiment, the reduced dosing regimen of venetoclax is one of the following: (i) 20 mg once daily for 7 days; (ii) 20 mg once daily for 14 days; (iii) 20 mg once daily for 21

days; (iv) 50 mg once daily for 7 days; (v) 50 mg once daily for 14 days; (vi) 50 mg once daily for 21 days; (vii) 100 mg once daily for 7 days; (viii) 100 mg once daily for 14 days; (ix) 100 mg once daily for 21 days; (x) 200 mg once daily for 7 days; (xi) 200 mg once daily for 14 days; (xii) 200 mg once daily for 21 days; (xiii) 400 mg once daily for 7 days; and (xiv) week 1 at 20mg/day, week 2 at 50mg/day and week 3 at 100mg/day.

Also envisioned is the treatment of an experimental mouse model according to the treatment regimen in this scenario, whereby the appropriate dosing regimens are commensurate with mouse body weight and tumor xenograft size.

Example 14 – Dosing Scenario VIII for ^{225}Ac -HuM195 and Venetoclax

A human AML patient is treated according to the following regimen. Venetoclax is orally administered according to a reduced dosing regimen, and ^{225}Ac -HuM195 is intravenously administered according to a reduced single dose regimen during the course of the venetoclax dosing regimen. In one embodiment, the single dose of ^{225}Ac -HuM195 is administered (a) on day 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 or 21 of the venetoclax dosing regimen, or (b) on the last day of, the penultimate day of, or 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 or 21 days prior to the last day of, the venetoclax dosing regimen. In another embodiment, (a) the reduced dosing regimen of venetoclax is one of (i) 20 mg once daily for 7 days; (ii) 20 mg once daily for 14 days; (iii) 20 mg once daily for 21 days; (iv) 50 mg once daily for 7 days; (v) 50 mg once daily for 14 days; (vi) 50 mg once daily for 21 days; (vii) 100 mg once daily for 7 days; (viii) 100 mg once daily for 14 days; (ix) 100 mg once daily for 21 days; (x) 200 mg once daily for 7 days; (xi) 200 mg once daily for 14 days; (xii) 200 mg once daily for 21 days; (xiii) 400 mg once daily for 7 days; and (xiv) week 1 at 20mg/day, week 2 at 50mg/day and week 3 at 100mg/day; and (b) the reduced dosing regimen of ^{225}Ac -HuM195 is one of (i) 2 x 0.5 $\mu\text{Ci/kg}$, 2 x 1.0 $\mu\text{Ci/kg}$, or 2 x 1.5 $\mu\text{Ci/kg}$, where the fractions are administered one week apart; or (ii) 1 x 0.5 $\mu\text{Ci/kg}$, 1 x 1.0 $\mu\text{Ci/kg}$, 1 x 2.0 $\mu\text{Ci/kg}$, or 1 x 3.0 $\mu\text{Ci/kg}$, for a single administration.

Also envisioned is the treatment of an experimental mouse model according to the treatment regimen in this scenario, whereby the appropriate dosing regimens are commensurate with mouse body weight and tumor xenograft size.

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What is claimed is:

1. A method for treating a subject afflicted with cancer, comprising administering to the subject (i) a BCL-2 inhibitor in conjunction with (ii) an alpha-emitting isotope-labeled agent that targets cancer cells in the subject, wherein the amounts of the BCL-2 inhibitor and labeled agent, when administered in conjunction with one another, are therapeutically effective.
2. The method of claim 1, wherein the subject is human.
3. The method of claim 1, wherein the cancer is a hematologic malignancy.
4. The method of claim 3, wherein the hematologic malignancy is a leukemia.
5. The method of claim 4, wherein the leukemia is acute myeloid leukemia.
6. The method of claim 1, wherein the BCL-2 inhibitor is venetoclax.
7. The method of claim 1, wherein the alpha-emitting isotope-labeled agent is an anti-CD33 antibody labeled with an alpha-emitting isotope.
8. The method of claim 7, wherein the anti-CD33 antibody labeled with an alpha-emitting isotope is ^{225}Ac -labeled HuM195.
9. A method for treating a human subject afflicted with acute myeloid leukemia, comprising administering to the subject (i) venetoclax in conjunction with (ii) ^{225}Ac -labeled HuM195, wherein the amounts of venetoclax and ^{225}Ac -labeled HuM195, when administered in conjunction with one another, are therapeutically effective.

10. A method for inducing the death of a cancer cell, comprising contacting the cell with (i) a BCL-2 inhibitor in conjunction with (ii) an alpha-emitting isotope-labeled agent that targets the cancer cell, wherein the amounts of BCL-2 inhibitor and labeled agent, when concurrently contacted with the cell, are effective to induce the cell's death.
11. The method of claim 10, wherein the cancer cell is a human cancer cell.
12. The method of claim 10, wherein the cancer cell is a hematologic cell.
13. The method of claim 12, wherein the cancer cell is a leukemic cell.
14. The method of claim 13, wherein the leukemic cell is an acute myeloid leukemic cell.
15. The method of claim 10, wherein the BCL-2 inhibitor is venetoclax.
16. The method of claim 10, wherein the alpha-emitting isotope-labeled agent is an anti-CD33 antibody labeled with an alpha-emitting isotope.
17. The method of claim 16, wherein the anti-CD33 antibody labeled with an alpha-emitting isotope is ^{225}Ac -labeled HuM195.
18. A method for inducing the death of an acute myeloid leukemic cell, comprising contacting the cell with (i) venetoclax in conjunction with (ii) ^{225}Ac -labeled HuM195, wherein the amounts of venetoclax and ^{225}Ac -labeled HuM195, when concurrently contacted with the cell, are effective to induce the cell's death.

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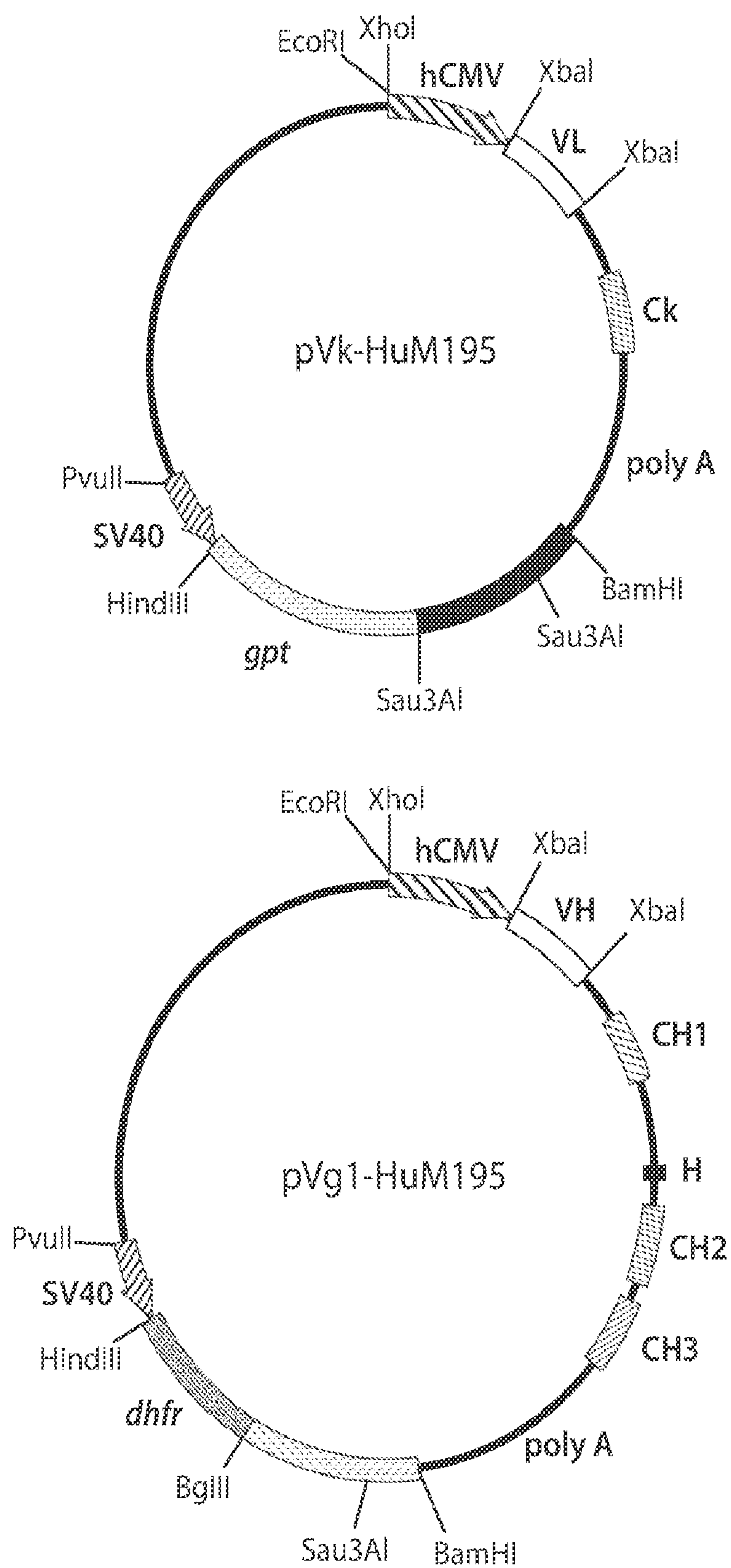


Figure 1

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624 TCTAGACCACCATGGAGAAAGACACACTCCTGCTATGGGTCTTCTCTGGGTCCAGGTCCACAGGTGACATTCAG
 M E K D T L L L W V L L L W V P G S T G D I Q
 704 ATGACCCAGTCTCCGAGCTCTCTGTCCGCATCAGTAGGAGACAGGGTCACCATCACATGCAGAGCCAGCGAAAGTGTGGA
 M T Q S P S S L S A S V G D R V T I T C R A S E S V D
 784 CAATTATGGCATTAGCTTTATGAAGTGGTTCCAACAGAAACCCGGGAAGGCTCCTAAGCTTCTGATTTACGCTGCATCCA
 N Y G I S F M N W F Q Q K P G K A P K L L I Y A A S
 864 ACCAAGGCTCCGGGGTACCCTCTCGCTTCTCAGGCAGTGGATCTGGGACAGACTTCACTCTCACCATTTCATCTCTGCAG
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 P D D F A T Y Y C Q Q S K E V P W T F G Q G T K V E I
 1024 CAAACGTAAGTAGAATCCAAAGTCTAGAAATTCTAAACTCTGAGGGGGTCCGATGACGTGGCCATTCTTTGCCTAAAGCA
 K R
 1104 TTGAGTTTACTGCAAGGTCAGAAAAGCATGCAAAGCCCTCAGAATGGCTGCAAAGAGCTCCAACAAAACAATTTAGAACT

 1184 TTATTAAGGAATAGGGGGAAGCTAGGAAGAACTCAAAACATCAAGATTTTAAATACGCTTCTTGGTCTCCTTGCTATAA

 1264 TTATCTGGGATAAGCATGCTGTTTTCTGTCTGTCCCTAACATGCTCTGTGATTATCCGCAAACAACACACCCCAAGGGCAG

 1344 AACTTTGTTACTTAAACACCATCCTGTTTGCTTCTTTCTCAGGAAGTGTGGCTGCACCATCTGTCTTCATCTTCCCGCC
 T V A A P S V F I F P P
 1424 ATCTGATGAGCAGTTCAAATCTGGAAGTGCCTCTGTTGTGTGCCTGCTGAATAACTTCTATCCCAGAGAGGCCAAAGTAC
 S D E Q L K S G T A S V V C L L N N F Y P R E A K V
 1504 AGTGAAGGTGGATAACGCCCTCCAATCGGGTAACTCCCAGGAGAGTGTACAGAGCAGGACAGCAAGGACAGCACCTAC
 Q W K V D N A L Q S G N S Q E S V T E Q D S K D S T Y
 1584 AGCCTCAGCAGCACCTGACGCTGAGCAAAGCAGACTACGAGAAACACAAAGTCTACGCCGTGCGAAGTCACCCATCAGGG
 S L S S T L T L S K A D Y E K H K V Y A C E V T H Q G
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 1744 GCCTGACCCCTCCCATCCTTTGGCTCTGACCCTTTTTCCACAGGGGACCTACCCCTATTGCGGTCTCCAGCTCATCT

 1824 TTCACCTCACCCCCCTCCTCCTCTTGGCTTTAATTATGCTAATGTTGGAGGAGAATGAATAAATAAAGTGAATCTTTGC

 1904 ACCTGFGGTTTCTCTCTTTCTCCTCATTTAATAATTATTATCTGTTGTTTTACCAACTACTCAATTTCTCTTATAAGGGACT

 1984 AAATATGTAGTCATCCTAAGGCGCATAACCATTATATAAAATCATCCTTCATTCTATTTTACCCTATCATCCTCTGCAAG

 2064 ACAGTCCTCCCTCAAACCCACAAGCCTTCTGTCTCAGAGTCCCTGGGCCATGGTAGGAGAGACTTGCTTCCTTGTTTT

 2144 CCCCTCCTCAGCAAGCCCTCATAGTCCTTTTTAAGGGTGACAGGTCTTACAGTCATATATCCTTTGATTCAATTCCTGA

 2224 GAATCAACCAAAGCAAATTTTTCAAAGAAGAAACCTGCTATAAAGAGAATCATTCAATTGCAACATGATATAAAATAACA

 2304 ACACAATAAAGCAATTAAATAAACAAACAATAGGGAAATGTTTAAGTTCATCATGGTACTTAGACTTAATGGAATGTCA

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 2464 TTAATCCACACTATACTGTGAGATTAAAAACATTCATTAAATGTTGCAAAGGTTCTATAAAGCTGAGAGACAAATATAT

 2544 TCTATAACTCAGCAATCCCACTTCTAGGATCC

Figure 2

SUBSTITUTE SHEET (RULE 26)

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624 TCTAGACCACCATGGGATGGAGCTGGATCTTTCTCTTCCTCCTGTCAGGAACTGCTGGCGTCCACTCTCAGGTCAGCTG
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 704 GTGCAGTCTGGAGCTGAGGTGAAGAAGCCTGGGAGCTCAGTGAAGGTTTCCTGCAAAGCTTCTGGCTACACCTTCACTGA
 V Q S G A E V K K P G S S V K V S C K A S G Y T F T D
 784 CTACAACATGCACTGGGTGAGGCAGGCTCCTGGCCAAGGCCTGGAATGGATTGGATATATTTATCCTTACAATGGTGGTA
 Y N M H W V R Q A P G Q G L E W I G Y I Y P Y N G G
 864 CCGGCTACAACCAGAAGTTCAAGAGCAAGGCCACAATTACAGCAGACGAGAGTACTAACACAGCCTACATGGAAGTCTCC
 T G Y N Q K F K S K A T I T A D E S T N T A Y M E L S
 944 AGCCTGAGGTCTGAGGACACTGCAGTCTATTACTGCGCAAGAGGGCGCCCGCTATGGACTACTGGGGCCAAGGACTCT
 S L R S E D T A V Y Y C A R G R P A M D Y W G Q G T L
 1024 GGTCACTGTCTCTTCAGGTAAGAATGGCCTCTAGACCACCATGGGATGGAGCTTTCTGGGGCAGGCCAGGCCTGACCTTG
 V T V S S
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 1184 GGACGCTGAACCTCGCGGACAGTTAAGAACCCAGGGGCCCTCTGCGCCCTGGGCCCAGCTCTGTCCCACACCGCGGTCACA

 1264 TGGCACCACCTCTCTTGCAGCCTCCACCAAGGGCCCATCGGTCTTCCCCCTGGCACCCTCCTCCAAGAGCACCTCTGGGG
 A S T K G P S V F P L A P S S K S T S G
 1344 GCACAGCGGGCCCTGGGCTGCCTGGTCAAGCACTACTTCCCCGAACCGGTGACGGTGTGCTGGAAGTCAAGCGCCCTGACC
 G T A A L G C L V K D Y F P E P V T V S W N S G A L T
 1424 AGCGCGGTGCACACCTTCCCGGCTGTCTACAGTCCTCAGGACTCTACTCCCTCAGCAGCGTGGTGACCGTCCCCTCCAG
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 S L G T Q T Y I C N V N H K P S N T K V D K K V
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 1664 AGTCCAGGGCAGCAAGGCAGGCCCCGTCTGCCTCTTCACCCGAGGCCTCTGCCCCCCCCACTCATGCTCAGGGAGAGGG

 1744 TCTTCTGGCTTTTTTCCCCAGGCTCTGGGCAGGCACAGGCTAGGTGCCCCCTAAGCCAGGCCCTGCACACAAAGGGGCAGGT

 1824 GCTGGGCTCAGACCTGCCAAGAGCCATATCCGGGAGGACCTGCCCCCTGACCTAAGCCCACCCCAAAGGCCAAACTCTCC

 1904 ACTCCCTCAGCTCGGACACCTTCTCTCCTCCCAGATTCCAGTAACTCCCAATCTTCTCTCTGCAGAGCCCCAAATCTTGTG
 E P K S C
 1984 ACAAACTCACACATGCCACCGTGCCAGGTAAGCCAGCCCAGGCCTCGCCCTCCAGCTCAAGGCGGGACAGGTGCCCT
 D K T H T C P P C P
 2064 AGAGTAGCCTGCATCCAGGGACAGGCCCCAGCCGGGTGCTGACACGTCCACCTCCATCTCTTCCCTCAGCACCTGAACTCC
 A P E L
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 L G G P S V F L F P P K P K D T L M I S R T P E V T C
 2224 GTGGTGGTGGACGTGAGCCACGAAGACCTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATAATGCCAA
 V V V D V S H E D P E V K F N W Y V D G V E V H N A K
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 2384 ATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCCAGCCCCCATCGAGAAAACCATCTCCAAGCCAAAGGT
 N G K E Y K C K V S N K A L P A P I E K T I S K A K
 2464 GGGACCCGTGGGGTGCGAGGGCCACATGGACAGAGGCGGGCTCGGCCACCCCTCTGCCCTGAGAGTGACCGCTGTACCAA

Figure 3

SUBSTITUTE SHEET (RULE 26)

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2544 CCTCTGTCCCTACAGGGCAGCCCCGAGAACCACAGGTGTACACCCTGCCCCCATCCCGGGATGAGCTGACCAAGAACCAG
G Q P R E P Q V Y T L P P S R D E L T K N Q
2624 GTCAGCCTGACCTGCCTGGTCAAAGGCTTCTATCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAGAA
V S L T C L V K G F Y P S D I A V E W E S N G Q P E N
2704 CAACTACAAGACCACGCCTCCCGTGTGACTCCGACGGCTCCTTCTTCTCTACAGCAAGCTCACCGTGGACAAGAGCA
N Y K T T P P V L D S D G S F F L Y S K L T V D K S
2784 GGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCACTACACGCAGAAGAGCCTCTCC
R W Q Q G N V F S C S V M H E A L H N H Y T Q K S L S
2864 CTGTCTCCGGTAAATGAGTGGCAGCGCCCGCAAGCCCCGCTCCCGGGCTCTCGCGGTGGCAGAGGATGCTTGGCAC
L S P G K *
2944 GTACCCCTGTACATACTTCCCGGGCGCCCAGCATGGAAATAAAGCACCCAGCGCTGCCCTGGGCCCCCTGGGAGACTGTG
3024 ATGGTTCCTTCCACGGGTCAGGCCGAGTCTGAGGCCTGAGTGGCATGAGGGAGGCAGAGCGGGTCCCACTGTCCCCACAC
3104 TGGCCCAGGCTGTGCAGGTGTGCCTGGCCCCCTAGGGTGGGGCTCAGCCAGGGGTGCCCTCGGCAGGGTGGGGGATTT
3184 GCCAGCGTGGCCCTCCCTCCAGCAGCACCTGCCCTGGGCTGGGCCACGGGAAGCCCTAGGAGCCCTGGGGACAGACACA
3264 CAGCCCCCTGCCTCTGTAGGAGACTGTCTGTCTGTGAGCGCCCTGTCTCTCCGACCTCCATGCCCACTCGGGGGCATGCC
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3664 GACACAGGCCCCCACCAGCCCCACGGGCACTCAAGGCCACAGGCTCTCGGCAGCTTCTCCACATGCTGACCTGCTC
3744 AGACAAACCCAGCCCTCCTCTCACAAGGGTGGCCCTGCAGCCGCCACACACACACAGGGGATCACACACCACGTACGTC
3824 CCTGGCCCTGGCCCACTTCCAGTGCCGCCCTTCCCTGCAGGATCC

Figure 3 Cont'd

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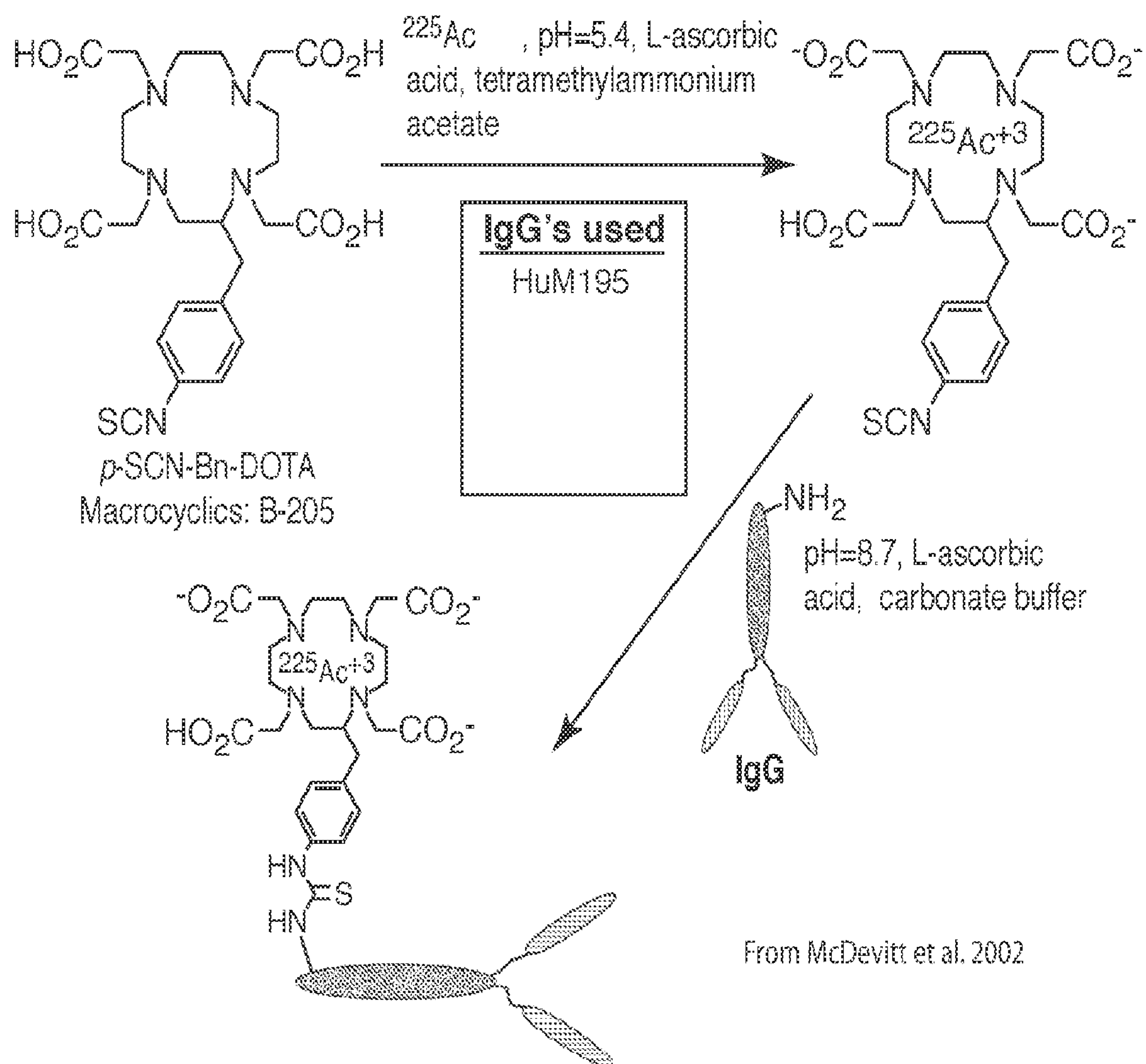


Figure 4

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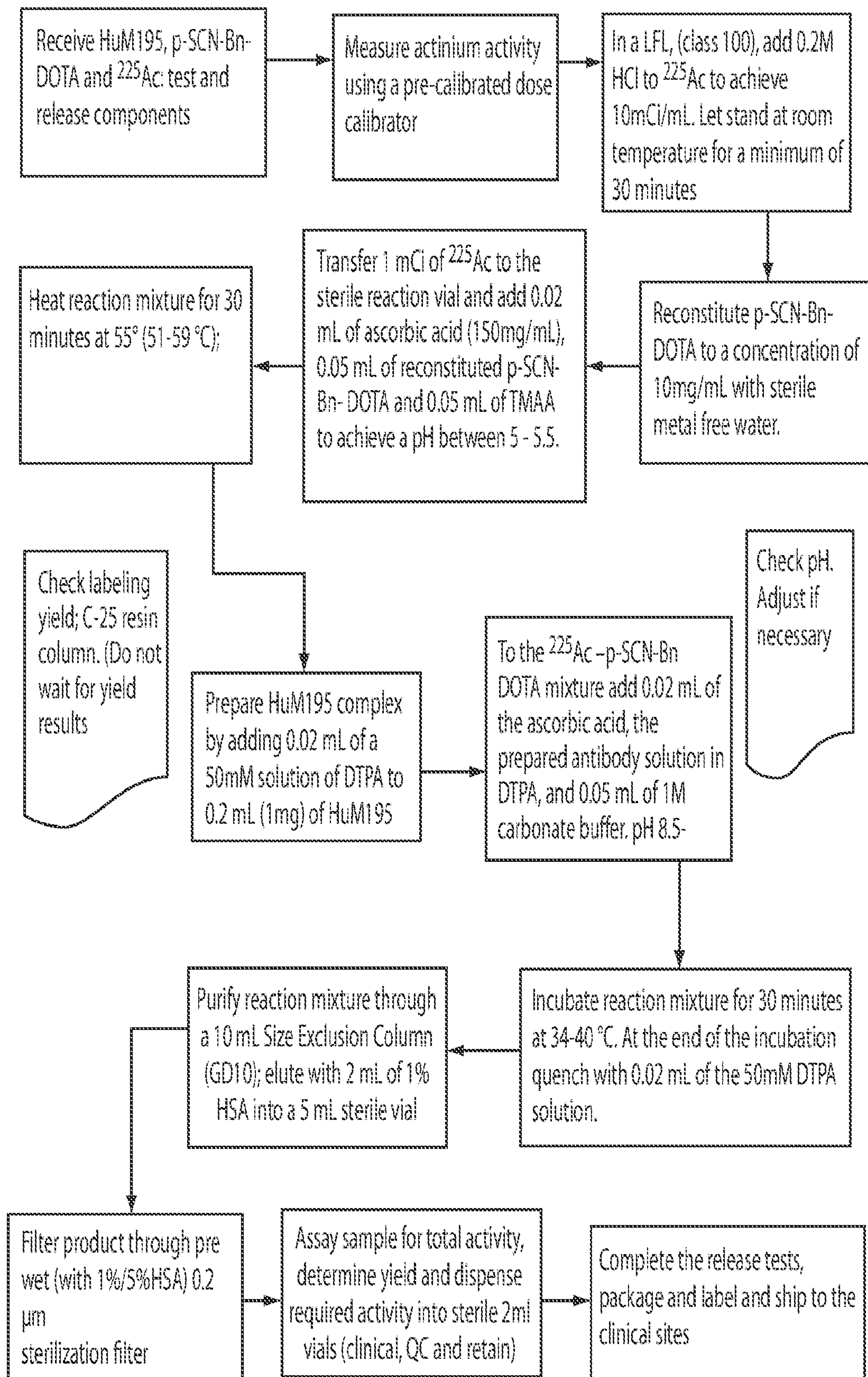


Figure 5

Protocol Schema

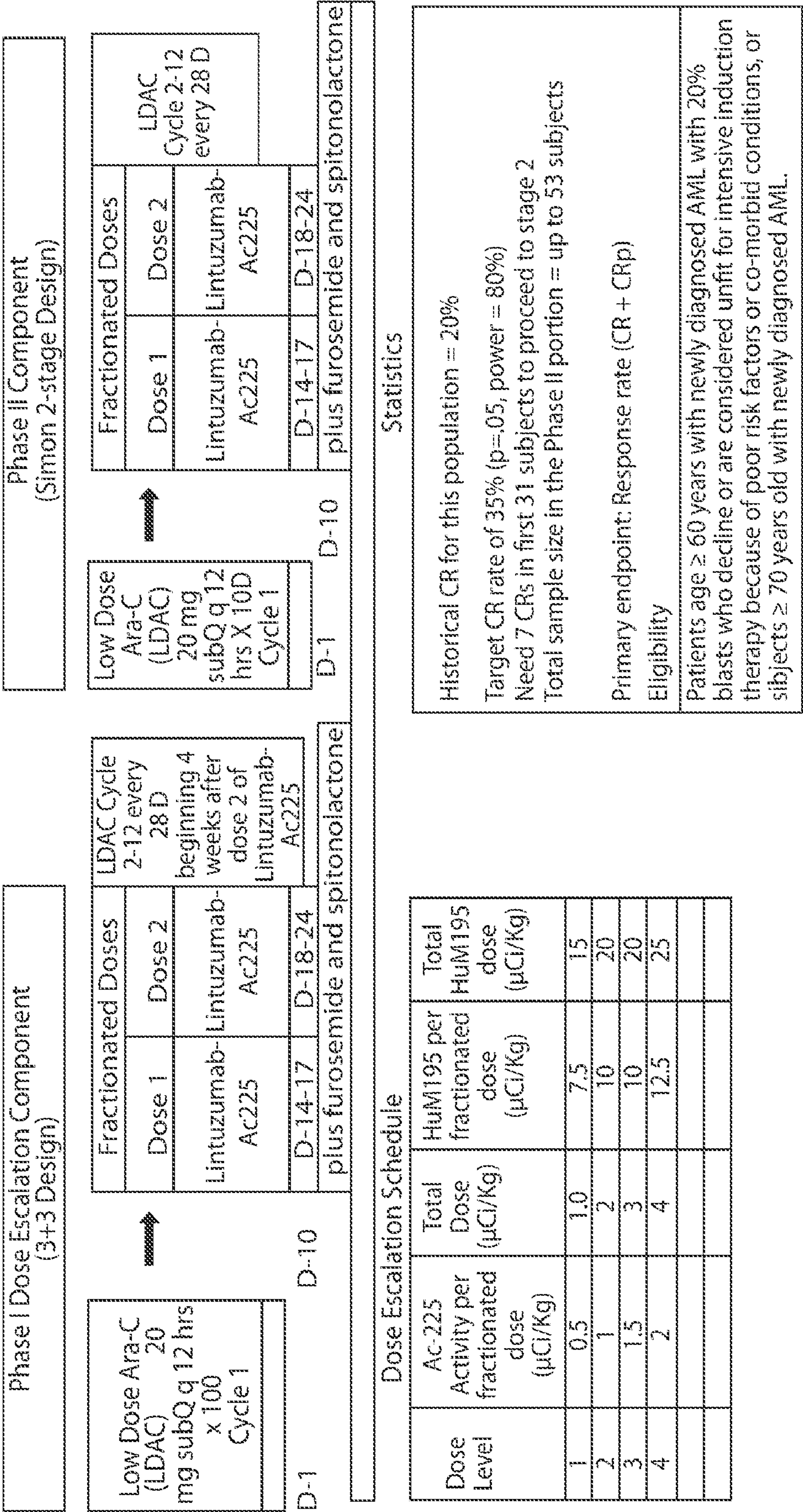


Figure 6

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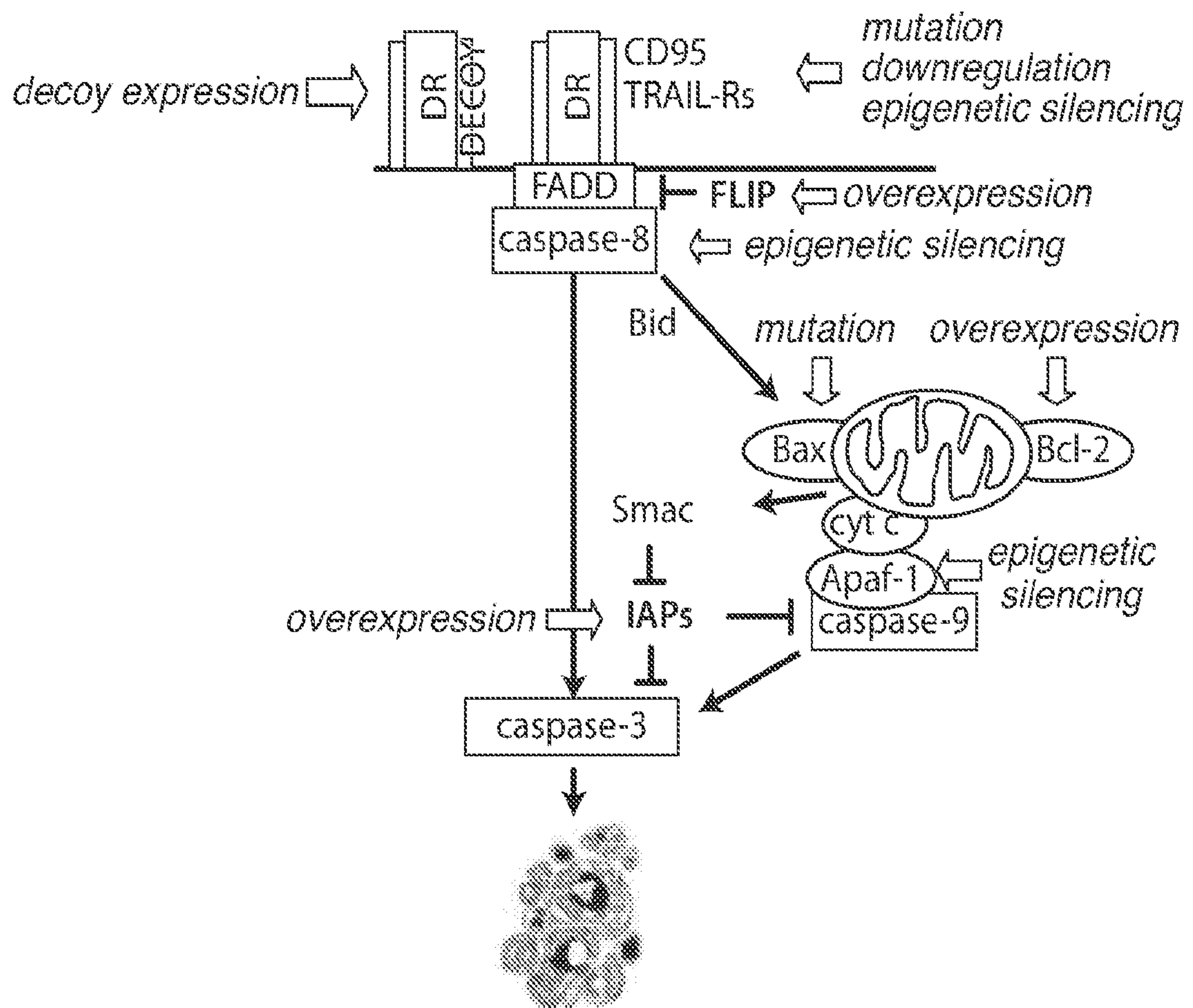


Figure 7

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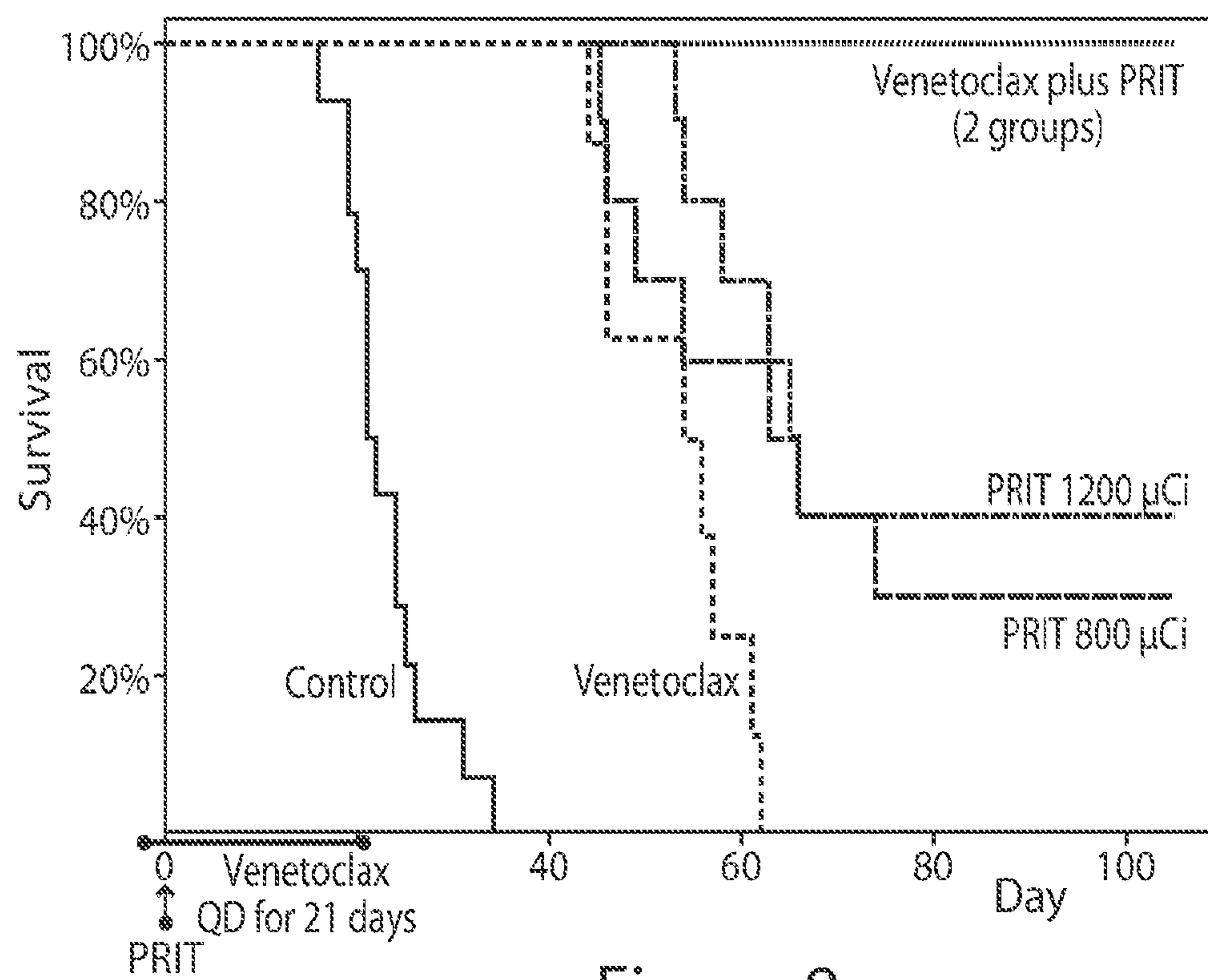


Figure 8

MoA	Beta (low LED)	Alpha (high LED)
dsDNA breaks (1), (2), (3), (4)	+/-	++
ssDNA breaks (1), (4)	+	+
Direct effect on apoptotic cascade (5), (6)	+/-	++
Bystander effect (5)	+	+
Dependence on tissue oxygenation (5)	+	-
Dependence on cell cycle phase (5)	+	-

Figure 9

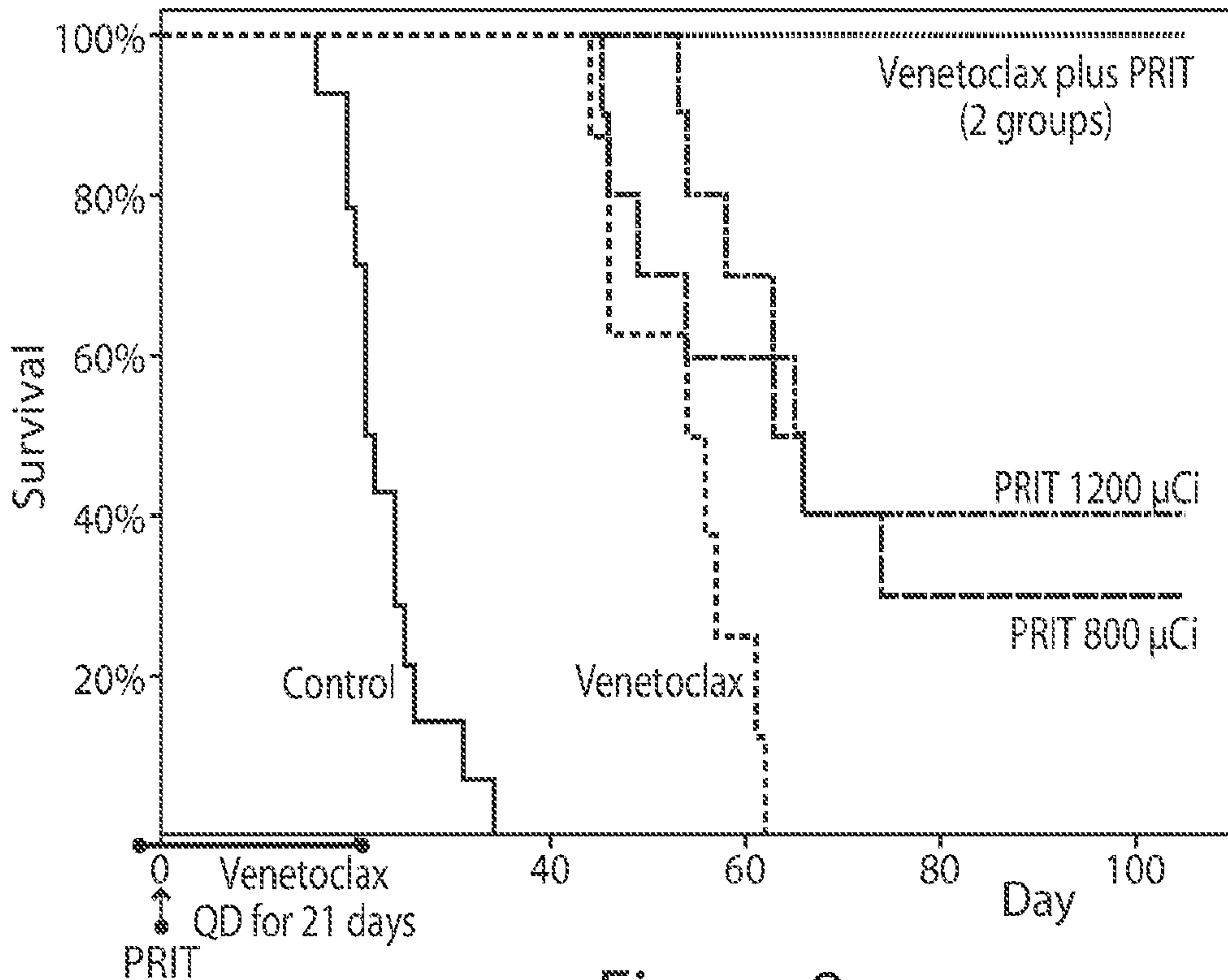


Figure 8