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(71) Applicant: **MONSANTO TECHNOLOGY LLC**
[US/US]; 800 North Lindbergh Boulevard, St. Louis, Missouri 63167 (US).

(72) Inventors: **HUANG, Shihshieh**; 800 North Lindbergh Boulevard, St. Louis, Missouri 63167 (US). **IAN-DOLINO, Alberto B.**; 800 North Lindbergh Boulevard, St. Louis, Missouri 63167 (US). **PEEL, Gregory J.**; 800

North Lindbergh Boulevard, St. Louis, Missouri 63167 (US).

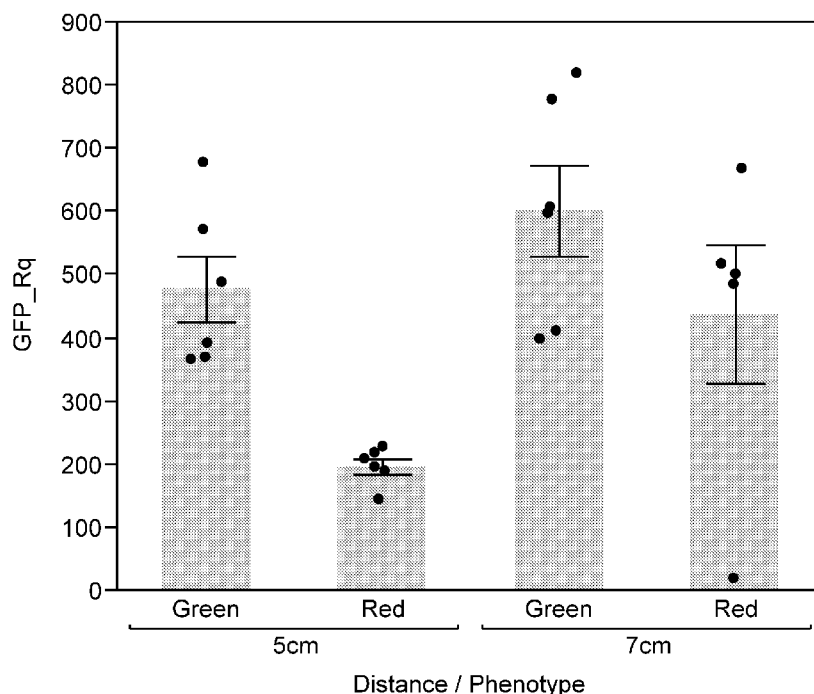
(74) Agents: **MARSH, David R.** et al.; 601 Massachusetts Ave., NW, Washington, District of Columbia 20001-3743 (US).

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[Continued on next page]

(54) Title: METHODS AND COMPOSITIONS FOR INTRODUCING NUCLEIC ACIDS INTO PLANTS



(57) Abstract: The present disclosure provides compositions and methods for introducing nucleic acids into plants. More specifically, disclosed herein are methods and compositions for introducing a nucleic acid, such as a double-stranded RNA, for silencing a target gene in a plant. Also disclosed herein are plants exhibiting modification of a target gene, obtained through the use of such methods and compositions.

FIG. 1



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LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE,
SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA,
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METHODS AND COMPOSITIONS FOR INTRODUCING NUCLEIC ACIDS INTO PLANTS

CROSS-REFERENCE TO RELATED APPLICATIONS AND INCORPORATION OF SEQUENCE

5 LISTINGS

[0001] This application claims priority to U. S. Provisional Patent Application No. 62/170,447 filed 3 June 2015, which is incorporated by reference in its entirety herein. The sequence listing contained in the file “40-77_61028_0000_US_ST25.txt” (40 kilobytes, created on 3 June 2015, and filed with U. S. Provisional Application number 62/170,447 on 3
10 June 2015) is incorporated by reference in its entirety herein. The sequence listing contained in the file “P34448WO00_Seq_Listing.txt” (40.8 kilobytes, created on June 1, 2016) is filed herewith and is incorporated by reference in its entirety herein.

FIELD

15 [0002] Disclosed herein are methods and compositions for introducing nucleic acids into plants. More specifically, disclosed herein are methods and compositions for introducing a nucleic acid, such as a double-stranded RNA, for silencing a target gene in a plant. Also disclosed herein are plants exhibiting modification of a target gene, obtained through the use of such methods and compositions.

20

BACKGROUND

[0003] There is a need for introducing nucleic acids, such as DNA or RNA for silencing a target gene, into plants, where the methods are scalable so as to be practical for use in multiple plants, such as plants in a greenhouse or growing in a field. Most methods of
25 introducing a nucleic acid for gene suppression are cumbersome and therefore generally of practical use only on individual plants in the laboratory or other small-scale environments. For example, “gene gun” or “biolistic” methods use gold or tungsten particles typically of 0.5 to 2 micrometers in size and coated with DNA or RNA that has been precipitated onto the particles; the particles are discharged using a “gene gun” powered by a gas at high pressure (typically
30 hundreds to thousands pounds per square inch) onto a plant held in an evacuated chamber. More recent biolistic methods using equipment such as the Helios® gene gun (Bio-Rad Laboratories, Inc.) use lower pressures (in the hundreds pounds per square inch) but still require the use of helium gas as a propellant and use of gold or tungsten particles typically of 0.5 to less than 2 micrometers in size that must be pre-coated with a nucleic acid; furthermore,

each plant must be treated individually and individual “cartridges” containing the particles must be prepared for each treatment. These limitations make the gene gun approaches inconvenient for true scalability for treating large numbers of plants such as in greenhouse and field use.

- 5 **[0004]** The present disclosure is related to various methods and compositions for introducing nucleic acids into a plant, where the methods are scalable and adaptable for use in multiple plants, and even for agricultural use in plants growing in a field.

SUMMARY

- 10 **[0005]** Several embodiments are related to a method for silencing a target gene in a plant, including, in any order, the steps of: (a) abrading a surface of a plant with a particulate of a size greater than about 2.5 micrometers; (b) applying an RNA to the surface of the plant, wherein the RNA comprises at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant; whereby the target gene is silenced. Several embodiments are also related to a single-step method, wherein a
15 surface of a plant is abraded with a composition including a particulate of a size greater than about 2.5 micrometers and an RNA that includes at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant; whereby the target gene is silenced.

- 20 **[0006]** Several embodiments are related to a spray apparatus for spraying multiple plants or multiple rows of plants, including a propellant source, at least one spray nozzle, and a reservoir containing a composition including a particulate and a nucleic acid that includes at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plants. In various embodiments, the nucleic acid is DNA or RNA, single- or double-stranded, or is provided as a synthetic nucleic acid or in a microbial
25 cell or as a microbial fermentation product. In some embodiments, the nucleic acid is an siRNA or a microRNA or a microRNA precursor.

- 30 **[0007]** Several embodiments are related to an apparatus for introducing a nucleic acid into a whole plant, including: (a) a matrix supporting an abrasive, and (b) a nucleic acid. In embodiments, the apparatus can be used in single-step methods, *e. g.*, when the nucleic acid is complexed with or otherwise attached or bonded to the abrasive. In some embodiments, the apparatus can be used in multi-step methods, *e. g.*, when the nucleic acid is applied before and/or after the plant or plant part is contacted by the abrasive.

- [0008]** Several embodiments are related to a method for introducing a nucleic acid into a plant, including: (a) applying a composition including a nucleic acid to a surface of a plant,

and (b) contacting a matrix supporting an abrasive with the surface; whereby the nucleic acid is introduced into the plant.

[0009] Several embodiments are related to a method for introducing a nucleic acid into a whole plant, including: (a) applying a composition including an abrasive and a nucleic acid to a surface of a whole plant, and (b) applying pressure to the surface; whereby the nucleic acid is introduced into the whole plant.

[0010] Several embodiments are related to a method for introducing a nucleic acid into a whole plant including, in any order, the steps of: (a) mechanical penetration of a surface of a whole plant with a non-particulate microstructure, and (b) contacting the surface of a whole plant with a nucleic acid. Embodiments include those where the non-particulate microstructure is a matrix on which is supported at least one selected from the group consisting of micro- or nano-needles, -whiskers, -fibers, and -wires. In embodiments, the non-particulate microstructure is removed after penetration of the plant surface, leaving the nucleic acid in the plant.

[0011] Several embodiments include a plant or a field of plants treated by a method, composition, or apparatus described herein, wherein the plant exhibits a desirable phenotype (such as improved yield, improved tolerance of biotic or abiotic stress, improved resistance to disease, improved herbicide susceptibility, improved herbicide resistance, and modified nutrient content) resulting from the treatment and when compared to an untreated plant.

Several embodiments include progeny seed or propagatable plant part of such plants, and commodity products produced from such plants. Several embodiments include a method for providing a plant exhibiting modification of a target gene in the plant, comprising directly regenerating a plant from a source plant treated by a method, composition, or apparatus described herein, wherein the modification of the target gene is non-heritable silencing of the target gene, or heritable or epigenetic silencing of the target gene, or a change in the nucleotide sequence of the target gene; embodiments include the directly regenerated plant exhibiting modification of the target gene and plants of subsequent generations grown from the directly regenerated plant and exhibiting modification of the target gene. Other aspects and specific embodiments are disclosed in the following detailed description and Examples.

BRIEF DESCRIPTION OF THE DRAWINGS

[0012] **Figure 1** depicts qPCR measurements of relative abundance of GFP mRNA, correlated to visual phenotype (see **Example 7**).

[0013] **Figure 2** depicts visual silencing efficacy for the different particulates tested, as described in **Example 10**. “AlO” = aluminum oxide (listed by mesh size), “DE” =

diatomaceous earth (listed as Celite grades), “SiC” = silicon carbide (listed by mesh size), “SLG” = soda lime glass (listed by bead diameter range in micrometers).

DETAILED DESCRIPTION

5 **[0014]** Unless defined otherwise, all technical and scientific terms used have the same meaning as commonly understood by one of ordinary skill in the art in the field. Generally, the nomenclature used and the manufacturing or laboratory procedures described below are well known and commonly employed in the art. Conventional methods are used for these procedures, such as those provided in the art and various general references. Where a term is provided in the singular, aspects described by the plural of that term are also contemplated. Where there are discrepancies in terms and definitions used in references that are incorporated by reference, the terms used in this application shall have the definitions given. Other technical terms used have their ordinary meaning in the art in which they are used, as exemplified by various art-specific dictionaries, for example, “The American Heritage® Science Dictionary” (Editors of the American Heritage Dictionaries, 2011, Houghton Mifflin Harcourt, Boston and New York), the “McGraw-Hill Dictionary of Scientific and Technical Terms” (6th edition, 2002, McGraw-Hill, New York), or the “Oxford Dictionary of Biology” (6th edition, 2008, Oxford University Press, Oxford and New York). No limitation to a mechanism or mode of action is intended by this disclosure. Reference thereto is provided for illustrative purposes only.

25 **[0015]** Unless otherwise stated, nucleic acid sequences in the text of this specification are given, when read from left to right, in the 5' to 3' direction. One of skill in the art would be aware that a given DNA sequence is understood to define a corresponding RNA sequence which is identical to the DNA sequence except for replacement of the thymine (T) nucleotides of the DNA with uracil (U) nucleotides. Thus, providing a specific DNA sequence is understood to define the exact RNA equivalent. A given first polynucleotide sequence, whether DNA or RNA, further defines the sequence of its exact complement (which can be DNA or RNA), *i. e.*, a second polynucleotide that hybridizes perfectly to the first polynucleotide by forming Watson-Crick base-pairs. By “essentially identical” or “essentially complementary” to a target gene or a fragment of a target gene is meant that a polynucleotide strand (or at least one strand of a double-stranded polynucleotide) is designed to hybridize (generally under physiological conditions such as those found in a living plant or animal cell) to a target gene or to a fragment of a target gene or to the transcript of the target gene or the fragment of a target gene; one of skill in the art would understand that such hybridization does

not necessarily require 100% sequence identity or complementarity. A first nucleic acid sequence is "operably" connected or "linked" with a second nucleic acid sequence when the first nucleic acid sequence is placed in a functional relationship with the second nucleic acid sequence. For example, a promoter sequence is "operably linked" to DNA if the promoter provides for transcription or expression of the DNA. Generally, operably linked DNA sequences are contiguous.

[0016] The term "polynucleotide" commonly refers to a DNA or RNA molecule containing multiple nucleotides and generally refers both to "oligonucleotides" (a polynucleotide molecule of 18-25 nucleotides in length) and longer polynucleotides of 26 or more nucleotides. Polynucleotides also include molecules containing multiple nucleotides including non-canonical nucleotides or chemically modified nucleotides as commonly practiced in the art; see, *e. g.*, chemical modifications disclosed in the technical manual "RNA Interference (RNAi) and DsiRNAs", 2011 (Integrated DNA Technologies Coralville, IA). Generally, polynucleotides or triggers described herein, whether DNA or RNA or both, and whether single- or double-stranded, include at least one segment of 18 or more contiguous nucleotides (or, in the case of double-stranded polynucleotides, at least 18 contiguous base-pairs) that are essentially identical or complementary to a fragment of equivalent size of the DNA of a target gene or the target gene's RNA transcript. Throughout this disclosure, "at least 18 contiguous" means "from about 18 to about 10,000, including every whole number point in between". Thus, embodiments include compositions including oligonucleotides having a length of 18-25 nucleotides (18-mers, 19-mers, 20-mers, 21-mers, 22-mers, 23-mers, 24-mers, or 25-mers), or medium-length polynucleotides having a length of 26 or more nucleotides (polynucleotides of 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, about 65, about 70, about 75, about 80, about 85, about 90, about 95, about 100, about 110, about 120, about 130, about 140, about 150, about 160, about 170, about 180, about 190, about 200, about 210, about 220, about 230, about 240, about 250, about 260, about 270, about 280, about 290, or about 300 nucleotides), or long polynucleotides having a length greater than about 300 nucleotides (*e. g.*, polynucleotides of between about 300 to about 400 nucleotides, between about 400 to about 500 nucleotides, between about 500 to about 600 nucleotides, between about 600 to about 700 nucleotides, between about 700 to about 800 nucleotides, between about 800 to about 900 nucleotides, between about 900 to about 1000 nucleotides, between about 300 to about 500 nucleotides, between about 300 to about 600 nucleotides, between about 300 to about 700 nucleotides, between about 300 to about 800 nucleotides, between about 300 to about 900

nucleotides, or about 1000 nucleotides in length, or even greater than about 1000 nucleotides in length, for example up to the entire length of a target gene including coding or non-coding or both coding and non-coding portions of the target gene). Where a polynucleotide is double-stranded, such as the dsRNA triggers described in the working Examples, its length can be similarly described in terms of base pairs. Double-stranded polynucleotides, such as the dsRNA triggers described in the working examples, can further be described in terms of one or more of the single-stranded components.

[0017] The polynucleotides or triggers described herein are generally designed to suppress or silence one or more genes (“target genes”). The term “gene” refers to any portion of a nucleic acid that provides for expression of a transcript or encodes a transcript. A “gene” can include, but is not limited to, a promoter region, 5’ untranslated regions, transcript encoding regions that can include intronic regions, 3’ untranslated regions, or combinations of these regions. In embodiments, the target genes can include coding or non-coding sequence or both. In other embodiments, the target gene has a sequence identical to or complementary to a messenger RNA, *e. g.*, in embodiments the target gene is a cDNA.

[0018] In embodiments, the target gene is an endogenous gene of the plant or a transgene expressed in the plant. In embodiments the target gene is not expressed by the plant itself but by a pathogen or parasite in or on the plant. In some embodiments, the target gene in the plant is a gene of a parasitic plant, fungus, or bacterium that is a pathogen or parasite of the plant; in other embodiments, the target gene in the plant is a gene of a virus that is a pathogen of the plant. In embodiments, the target gene is multiple target genes; the multiple target genes can be from a single species or can be from more than one species.

[0019] In embodiments, the methods, compositions, and apparatuses described herein are useful for obtaining a phenotype (*e. g.*, improved yield, improved resistance temperature, water, or nutrient stress, improved resistance to pathogens, improved herbicide susceptibility, improved herbicide resistance, and modified nutrient content or appearance) in a plant directly treated by a method as described herein. In other embodiments, the effect of treatment by a method of this disclosure is passed on to subsequent generations, for example in an epigenetic effect. In many embodiments the DNA or RNA employed in the methods is designed to silence a target gene. In related applications the methods can be used to deliver any nucleic acid of interest, including nucleic acids designed for gene editing, *e. g.*, using a CRISPR or Cas9 system.

[0020] In contrast to plant transformation techniques using a gene gun, the methods and compositions described herein use particulates made of materials less expensive than gold

or tungsten and of a size range greater than that of the particles used in gene gun transformation, typically use lower pressures, do not require treatment of the plant in a vacuum, and can be carried out in a whole plant or multiple plants. The methods and compositions are scalable so as to be useful in treating multiple plants at one time.

5 **[0021]** In general the methods described herein use mechanical disruption of a surface of the plant to assist in delivery of the nucleic acid to the plant, for example by contacting a surface of a plant with an abrasive such as a loose particulate or a particulate supported on a matrix, or by contacting a surface of a plant with a non-particulate microstructure. Generally the abrasion used in the methods superficially disrupts cells in the cuticle or epidermis or both
10 cuticle and epidermis of the plant, but does not damage cells in deeper tissues of the plant.

[0022] In embodiments, the target gene silencing resulting from treatment of a surface of the plant is systemic, *i. e.*, the silencing phenotype is observed in parts of the plant other than the treated parts. In embodiments, the target gene silencing resulting from treatment of a surface of the plant is local, *i. e.*, the silencing phenotype is observed in parts treated by the
15 methods and compositions described herein.

[0023] The methods described herein can additionally combine mechanical disruption of the plant surface with other treatments, for example with sequential or concomitant use of a surfactant, nuclease inhibitors, photoprotectants, rainfastness agents, and the like.

[0024] In embodiments, the methods and compositions described herein are useful for
20 obtaining a phenotype (*e. g.*, improved yield, improved resistance to temperature, water, or nutrient stress, improved resistance to pathogens, improved herbicide susceptibility, improved herbicide resistance, and modified nutrient content or appearance) in a plant directly treated by a method or composition described herein. In other embodiments, the effect of treatment by a method or composition as described herein is passed on to subsequent generations, for example
25 in an epigenetic effect. In many embodiments the DNA or RNA employed in the methods is designed to silence a target gene. In other applications the methods can be used to deliver any nucleic acid of interest, including nucleic acids designed for gene editing, *e. g.*, using a CRISPR or Cas9 system.

[0025] Also described herein are compositions and apparatuses useful in delivering a
30 nucleic acid into a plant, as well as plants treated by a method or composition as described herein. In embodiments, DNA- or RNA-coated aluminum oxide or silicon carbide particles are delivered into a plant using a pressurized gas. For example, RNA molecules (*e. g.*, synthetic dsRNA, or a dsRNA produced in a bacterial system) or DNA molecules (*e. g.*, a VIGS vector or a plasmid) are coated onto aluminum oxide (Al₂O₃) or silicon carbide (SiC, “carborundum”)

particles and allowed to dry; these nucleic-acid-coated particles are sprayed onto leaves of a plant using pressurized air or other gas and cause silencing of the gene targeted by the nucleic acid. An airbrush (*e. g.*, Master Airbrush Model G78 Single-Action Gravity Feed Air Abrasive Etching Airbrush Gun as used in the experiments described herein) using compressed air is one convenient means of applying the particulates to the plant. Pressurized gas can be provided by any convenient means, such as an air compressor or a compressed gas cylinder; when used with a dry powder composition, preferably a low-humidity pressurized gas is used.

[0026] Examples of blunt-ended dsRNA triggers designed to silence green fluorescent protein (GFP) are provided in **Table 1**.

Table 1

SEQ ID NO:*	size (base pairs)
1	50
2	78
3	124
4	125
5	249
6	258

*sequence of anti-sense strand, 5'→3'

[0027] Examples of particulate abrasives useful in the methods and compositions described herein are provided in **Table 2**.

Table 2

Abrasive	Composition	Median Size (micrometers)
280 mesh SiC	silicon carbide	33.0-36.0
320 mesh SiC	silicon carbide	26.3-29.2
360 mesh SiC	silicon carbide	20.1-23.1
400 mesh SiC	silicon carbide	15.5-17.5
500 mesh SiC	silicon carbide	11.3-13.3
600 mesh SiC	silicon carbide	8.0-10.0
CELITE 560	diatomaceous silica	95.7
CELITE 545	diatomaceous silica	46.5
CELITE 503	diatomaceous silica	33.9
CELITE 512	diatomaceous silica	23.9
CELITE 577	diatomaceous silica	20.8
CELITE S	diatomaceous silica	7.3
glass microsphere	soda lime glass	10-22
glass microsphere	soda lime glass	22-27
glass microsphere	soda lime glass	27-32

glass microsphere	soda lime glass	32-38
glass microsphere	soda lime glass	38-45
glass microsphere	soda lime glass	45-53
glass microsphere	soda lime glass	53-63
240 mesh AlO ₃	aluminum oxide	>37
280 mesh AlO ₃	aluminum oxide	33-36
320 mesh AlO ₃	aluminum oxide	<37
360 mesh AlO ₃	aluminum oxide	20.1-23.1
400 mesh AlO ₃	aluminum oxide	15.5-17.5
500 mesh AlO ₃	aluminum oxide	11.3-13.3
600 mesh AlO ₃	aluminum oxide	8.0-10.0
800 mesh AlO ₃	aluminum oxide	5.3-.73

METHODS FOR SILENCING A TARGET GENE IN A PLANT USING ABRASIVES

[0028] A first aspect provides a method for silencing a target gene in a plant, including, in any order, the steps of: (a) abrading a surface of a plant with a particulate of a size greater than about 2.5 micrometers; (b) applying an RNA to the surface of the plant, wherein the RNA includes at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant; whereby the target gene is silenced.

[0029] In embodiments, step (a) is followed by step (b). In other embodiments, step (b) is followed by step (a). In embodiments where the RNA is applied prior to the abrasion step, the RNA can be allowed to dry on the plant surface prior to abrasion. Additional treatments of the plant (for example, treatment with a surfactant, an RNase inhibitor, a photoprotectant, or a rainfastness agent) are optionally performed, subsequent to either abrasion or application of RNA, or concomitant with either or both steps.

[0030] The RNA applied to the plant surface can be single-stranded RNA (ssRNA), double-stranded RNA (dsRNA), or a combination of ssRNA and dsRNA. "Double-stranded" refers to the base-pairing that occurs between sufficiently complementary, anti-parallel nucleic acid strands to form a double-stranded or duplexed nucleic acid structure, generally under physiologically relevant conditions. In embodiments, the RNA is a dsRNA including two strands that are perfectly complementary and form a blunt-ended RNA duplex. In other embodiments, RNA is a dsRNA that is not perfectly complementary, for example, a dsRNA that has additional nucleotides at one or both ends so as to form an overhang, or a dsRNA that includes non-base-paired mismatches within the otherwise base-paired duplex. In embodiments, the dsRNA has an overhang at one or both ends (termini); the overhang can be a

single nucleotide or 2, 3, 4, 5, 6, or more nucleotides, and can be located on the 5' end or on the 3' end of a strand. The RNA can be provided as a single nucleotide sequence (as a single molecule) or multiple nucleotide sequences (as multiple molecules, or as multiple strands of RNA). The RNA can be chemically synthesized, or can be produced by expression in a microorganism, by expression in a plant cell, or by microbial fermentation. The RNA can be chemically modified, *e. g.*, to improve stability or efficacy.

[0031] The RNA includes at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant. The RNA can further include nucleotides having a sequence unrelated to the sequence of the target gene or genes to be suppressed; for example, the RNA can include flanking or intervening “filler” or “spacer” segments of consecutive nucleotides, wherein the segments have a sequence unrelated to the sequence of the target gene or genes to be suppressed. In embodiments, the RNA includes 19 – 500 nucleotides (or base pairs, where the RNA is dsRNA). In embodiments, the RNA includes 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, about 65, about 70, about 75, about 80, about 85, about 90, about 95, about 100, about 110, about 120, about 130, about 140, about 150, about 160, about 170, about 180, about 190, about 200, about 210, about 220, about 230, about 240, about 250, about 260, about 270, about 280, about 290, or about 300 nucleotides (or base pairs). For reasons of economy, the RNA can be relatively short, of about 300 nucleotides or base pairs in length. Nonetheless, in embodiments, the RNA has a length greater than about 300 nucleotides (or base pairs), *e. g.*, between about 300 to about 400, between about 400 to about 500, between about 500 to about 600, between about 600 to about 700, between about 700 to about 800, between about 800 to about 900, between about 900 to about 1000, between about 300 to about 500, between about 300 to about 600, between about 300 to about 700, between about 300 to about 800, between about 300 to about 900, or about 1000 nucleotides (or base pairs) in length, or even greater than about 1000 nucleotides (or base pairs) in length, for example up to the entire length of a target gene or genes including coding or non-coding or both coding and non-coding portions of the target gene or genes).

[0032] In embodiments, the RNA is ssRNA or dsRNA designed to silence a single target gene. In embodiments, the RNA is ssRNA or dsRNA including multiple repeating segments for targeting a single target gene. Multiple target genes (which can be target genes of the plant, or of a pathogen or parasite of the plant, or a combination of both), or multiple regions of a single target gene, can be targeted by a chimeric RNA. In an example, the RNA is a dsRNA including segments corresponding to different regions of the target gene, or multiple

copies of a segment. In an example, the RNA is a chimeric dsRNA including dsRNA segments targeting a gene expressed in the plant and dsRNA segments targeting a gene of a pathogen or parasite of the plant.

[0033] In embodiments, the RNA includes a miRNA precursor or a small RNA such as an siRNA or a mature miRNA known to have gene silencing functionality in a cell. In embodiments, the RNA is a 21-mer or 22-mer. The RNA is non-transcribable, and is not self-replicating. In embodiments, the RNA is not contained in a viral vector nor encoded by a plasmid.

[0034] In embodiments of the method, the RNA is in a solution, *e. g.*, a sprayable solution. Alternatively, the RNA can be provided in a powder, emulsion, or suspension. Solution, powders, emulsions, or suspensions can conveniently further include one or more components selected from the group consisting of a surfactant, a buffer, an osmoprotectant, a cryoprotectant, an RNase inhibitor (ideally compatible with RNAi activity), a nucleic acid condensing agent (*e. g.*, a polyamine), a transfection agent, a wound-response inhibitor (*e. g.*, callose, hydrogen peroxide inhibitors), a salt (*e. g.*, a Zn^{+2} or Mg^{+2} salt), a plant hormone, a sugar, and a sugar alcohol. Specific embodiments include an RNA solution (or powder, emulsion, or suspension) including one or more components selected from the group consisting of sucrose, mannitol, 2-(*N*-morpholino)ethanesulfonic acid, and Silwet L77.

[0035] In embodiments, the abrasion step includes application of the particulate in a spray. Embodiments include those wherein the spray is applied by an airbrush, by a compressed-gas sprayer, or by a canister sprayer, a track sprayer, or a boom sprayer.

[0036] The abrasion step can be accomplished by means other than by spraying. In embodiments, the particulate is supported by, attached to, or embedded in a matrix. The matrix can include a fibrous, porous, non-porous, or adhesive support. An example is a particulate abrasive that is permanently bonded to paper, such as sandpaper.

[0037] Particulates useful in the methods include a particulate abrasive selected from the group consisting of a mineral abrasive, a metal abrasive, a synthetic abrasive, and an organic abrasive. Embodiments include particulate abrasives selected from the group consisting of aluminum oxide, silicon carbide (“carborundum”, silicon dioxide, soda lime glass, diatomaceous silica (“diatomaceous earth”), flint, quartz, garnet, silicon dioxide, pumice, sand, feldspar, calcite, steel, tungsten, ceramic, boron carbide, tungsten carbide, an organic or biodegradable abrasive, or combinations of these. In embodiments, the particulate is composed of an organic or biodegradable material, such as, but not limited to wood particles, corn cob particles, grain or seed particles, or nut shell particles.

[0038] Particulate size is selected according to factors such as compatibility with a given formulation, suitability for use in a given apparatus (such as a spray nozzle), efficiency in delivering the RNA, or for minimizing damage to the treated plants. In embodiments, the particulate is of an average size range from about 2.5 micrometers to about 50 micrometers. In various embodiments, the particulate is of an average size range from 2.5 – 50, 2.5 – 40, 2.5 – 30, 2.5 – 20, 5 – 50, 5 – 40, 5 – 30, 5 – 20, 7 – 50, 7 – 40, 7 – 30, 7 – 20, 8 – 50, 8 – 40, 8 – 30, 8 – 20, 10 – 50, 10 – 40, 10 – 30, or 10 – 25 micrometers. The working Examples further illustrate embodiments of useful particulate size ranges.

[0039] In embodiments of the method, the particulate, the RNA, or both, are further applied with at least one selected from the group consisting of water, a biocide, a chelator, a buffer, a nonionic surfactant, a zwitterionic surfactant, a zinc or magnesium salt, a defoamer, a rainfastness agent, and a photoprotectant. In embodiments, the particulate, the RNA, or both, are further applied with at least one selected from the group consisting of a carrier agent, a surfactant, an osmolyte, an organosilicone, a polynucleotide herbicidal molecule, a non-polynucleotide herbicidal molecule, a non-polynucleotide pesticide, a safener, an insect attractant, and an insect growth regulator. In embodiments, the particulate, the RNA, or both, are further applied with at least one pesticidal agent selected from the group consisting of a patatin, a plant lectin, a phytoecdysteroid, a *Bacillus thuringiensis* insecticidal protein, a *Xenorhabdus* insecticidal protein, a *Photorhabdus* insecticidal protein, a *Bacillus laterosporous* insecticidal protein, and a *Bacillus sphaericus* insecticidal protein. In embodiments, the particulate, the RNA, or both, are further applied with at least one osmolyte such as an amino acid osmolyte, a methylamine osmolyte, or a polyol or sugar or sugar alcohol osmolyte; non-limiting examples suitable osmolytes include sorbitol, mannitol, xylitol, erythrol, glycerol, glucose, sucrose, proline, valine, isoleucine, ectoine, aspartic acid, trehalose, glycine betaine (betaine), carnitine, taurine, sarcosine, *myo*-inositol (inositol). Alternatively, any of these additional components or combinations thereof can be applied separately from the application of the particulate or of the RNA.

[0040] The abrasion used in these two-step methods preferably does minimal damage to the plant. In embodiments, the particulate disrupts cells only in the cuticle, or only in the cuticle and epidermis of the plant. In embodiments, cells deeper than the epidermis are essentially not damaged by the particulate abrasive. In embodiments, the silencing is systemic and the target gene is silenced in at least one location of the plant that is not the location of abrasion.

[0041] In embodiments, these methods are useful for obtaining a phenotype (*e. g.*, improved yield, improved resistance temperature, water, or nutrient stress, improved resistance to pathogens, improved herbicide susceptibility, improved herbicide resistance, and modified nutrient content or appearance) resulting from silencing of a target gene in a plant directly treated by these methods. In a related method, the effect of treatment is passed on to subsequent generations, for example where the RNA is designed to target non-coding sequence such as 5' untranslated sequence or a promoter of the target gene, and thereby causing an epigenetic effect. For example, use of a silencing RNA targeting the promoter region of an endogenous phytoene desaturase (PDS) gene in a plant results in bleaching not only of the treated plant but in progeny plants that have not themselves been treated directly with the silencing RNA.

[0042] In many embodiments the RNA employed in the methods is designed to silence a target gene. However, in related applications similar methods can be used to deliver any nucleic acid of interest, including nucleic acids designed for gene editing, *e. g.*, using a CRISPR or Cas9 system. In one aspect, a polynucleotide provided herein comprises a nucleic acid sequence encoding one or more elements of a NgAgo-gDNA system. In some embodiments, the polynucleotide encodes a prokaryotic Argonaute. In some embodiments, the polynucleotide encodes a guide sequence used by a prokaryotic Argonaute. In some embodiments, the prokaryotic Argonaute is from *Natronobacterium gregoryi* (NgAgo), *Thermus thermophilus* (TtAgo), or *Pyrococcus furiosus* (PfAgo). *See, e.g.*, Gao *et al.*, Nat. Biotechnol., May 2, 2016, published online; Swarts *et al.*, Nature, 2014, 507(7491):258-61; and Swarts *et al.*, Nucleic Acid Res., 2015, 43(10):5120-5129. Similar to Cas9, endonucleases from the Argonaute protein family also use oligonucleotides as guides to degrade invasive genomes. For example, the *Natronobacterium gregoryi* Argonaute (NgAgo) was found to be a DNA-guided endonuclease suitable for genome editing. NgAgo binds 5' phosphorylated single-stranded guide DNA (gDNA) of about 24 nucleotides, efficiently creates site-specific DNA double-strand breaks when loaded with the gDNA. The NgAgo-gDNA system does not require a protospacer-adjacent motif (PAM), as does Cas9, and it has been suggested that it has a low tolerance to guide-target mismatches and high efficiency in editing (G+C)-rich genomic targets. Gao *et al.*, Nat. Biotechnol., May 2, 2016.

SINGLE-STEP METHODS FOR SILENCING A TARGET GENE IN A PLANT USING ABRASIVES

[0043] Another aspect provides a single-step method for silencing a target gene in a plant, including: abrading a surface of a plant with a composition including a particulate of a

size greater than about 2.5 micrometers and an RNA that includes at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant; whereby the target gene is silenced.

[0044] In embodiments, the composition is a solid or a dry composition, such as a powder. In embodiments, the composition is a suspension of the particulate and the RNA in a liquid or a gel. In embodiments, the composition is an emulsion. In embodiments the RNA is physically closely associated with or in contact with the particulate, *e. g.*, simply by drying the RNA onto the particulate, or by bonding the RNA to the particulate by ionic or electrostatic attraction, direct covalent bonding, covalent bonding using a linker, or by means of an adhesive or other affixing agent.

[0045] The composition is applied to the plant using any convenient application method, which may involve the use of positive or negative pressure to achieve abrasion of the plant surface. Dry compositions, such as powders, can be applied by dusting or dry spraying. Liquids, including suspensions or emulsions, can be applied with a brush, roller, sprayer, or other apparatus. In embodiments, the abrasion is achieved by applying the composition by spraying, *e. g.*, by application using an airbrush, by a compressed-gas sprayer, or by a canister sprayer, a track sprayer, or a boom sprayer.

[0046] In embodiments, the composition includes “loose” (non-affixed) or discrete particulates, *e. g.*, a loose powder or dust. An embodiment includes a composition where a loose particulate abrasive is mixed with a solution of RNA and the mixture dried to form a dry powder which is then applied in a single step to a plant. In other embodiments, the composition includes a particulate supported by, attached to, or embedded in a matrix, such as a fibrous, porous, non-porous, or adhesive support. An embodiment includes a composition where a loose particulate abrasive is mixed with a solution of RNA, the resulting mixture is affixed to a matrix, and the matrix including the particulate and RNA is then applied to a plant. Another embodiment includes a composition where the RNA is added to a particulate already affixed to a matrix and the whole then applied to a plant; an example is sandpaper or other particulate-abrasive-bearing material to which RNA is added.

[0047] In embodiments, the composition includes a particulate selected from the group consisting of a mineral abrasive, a metal abrasive, a synthetic abrasive, and an organic abrasive. In embodiments, the composition includes particulate abrasives selected from the group consisting of aluminum oxide, silicon carbide (“carborundum”, silicon dioxide, soda lime glass, diatomaceous silica (“diatomaceous earth”), flint, quartz, garnet, silicon dioxide, pumice, sand, feldspar, calcite, steel, tungsten, ceramic, boron carbide, tungsten carbide, an

organic or biodegradable abrasive, or combinations of these. In embodiments, the particulate is composed of an organic or biodegradable material, such as, but not limited to wood particles, corn cob particles, grain or seed particles, or nut shell par In embodiments, the particulate used in the composition is of an average size range from about 2.5 micrometers to about 50

micrometers. In various embodiments, the particulate is of an average size range from 2.5 – 50, 2.5 – 40, 2.5 – 30, 2.5 – 20, 5 – 50, 5 – 40, 5 – 30, 5 – 20, 7 – 50, 7 – 40, 7 – 30, 7 – 20, 8 – 50, 8 – 40, 8 – 30, 8 – 20, 10 – 50, 10 – 40, 10 – 30, or 10 – 25 micrometers, as further illustrated in the working Examples.

[0048] In embodiments, the composition further includes one or more components selected from the group consisting of a surfactant, a buffer, an osmoprotectant, a cryoprotectant, an RNase inhibitor (ideally compatible with RNAi activity), a nucleic acid condensing agent (*e. g.*, a polyamine), a transfection agent, a wound-response inhibitor (*e. g.*, callose, hydrogen peroxide inhibitors), a salt (*e. g.*, a Zn^{+2} or Mg^{+2} salt), a plant hormone, a sugar, and a sugar alcohol. Specific embodiments of the composition include one or more components selected from the group consisting of sucrose, mannitol, 2-(*N*-morpholino)ethanesulfonic acid, and Silwet L77.

[0049] In embodiments, the composition further includes at least one selected from the group consisting of water, a biocide, a chelator, a buffer, a nonionic surfactant, a zwitterionic surfactant, a zinc or magnesium salt, a defoamer, a rainfastness agent, and a photoprotectant.

In embodiments, the composition further includes at least one selected from the group consisting of a carrier agent, a surfactant, an osmolyte, an organosilicone, a polynucleotide herbicidal molecule, a non-polynucleotide herbicidal molecule, a non-polynucleotide pesticide, a safener, an insect attractant, and an insect growth regulator. In embodiments, the composition further includes at least one pesticidal agent selected from the group consisting of a patatin, a plant lectin, a phytoecdysteroid, a *Bacillus thuringiensis* insecticidal protein, a *Xenorhabdus* insecticidal protein, a *Photorhabdus* insecticidal protein, a *Bacillus laterosporous* insecticidal protein, and a *Bacillus sphaericus* insecticidal protein. In embodiments, the composition further includes at least one osmolyte such as an amino acid osmolyte, a methylamine osmolyte, or a polyol or sugar or sugar alcohol osmolyte; non-limiting examples suitable osmolytes include sorbitol, mannitol, xylitol, erythrol, glycerol, glucose, sucrose, proline, valine, isoleucine, ectoine, aspartic acid, trehalose, glycine betaine (betaine), carnitine, taurine, sarcosine, *myo*-inositol (inositol). Alternatively, any of these additional components or combinations thereof can be applied separately from the particulate-RNA composition. Thus, additional treatments of the plant (for example, treatment with a

surfactant, an RNase inhibitor, a photoprotectant, or a rainfastness agent) are optionally performed, subsequent to or following application of the particulate-RNA composition.

[0050] The abrasion used in these single-step methods preferably does minimal damage to the plant. In embodiments, the particulate disrupts cells only in the cuticle, or only in the cuticle and epidermis of the plant. In embodiments, cells deeper than the epidermis are essentially not damaged by the particulate abrasive. In embodiments, the silencing is systemic and the target gene is silenced in at least one location of the plant that is not the location of abrasion.

[0051] Other details of the single-step method are similar to those described above for the two-step method for silencing a target gene in a plant, as described above under the heading "METHODS FOR SILENCING A TARGET GENE IN A PLANT USING ABRASIVES".

[0052] In embodiments, these methods are useful for obtaining a phenotype (*e. g.*, improved yield, improved resistance temperature, water, or nutrient stress, improved resistance to pathogens, improved herbicide susceptibility, improved herbicide resistance, and modified nutrient content or appearance) resulting from silencing of a target gene in a plant directly treated by these methods. In a related method, the effect of treatment is passed on to subsequent generations, for example where the RNA is designed to target non-coding sequence such as 5' untranslated sequence or a promoter of the target gene, and thereby causing an epigenetic effect. For example, use of a silencing RNA targeting the promoter region of an endogenous phytoene desaturase (PDS) gene in a plant results in bleaching not only of the treated plant but in progeny plants that have not themselves been treated directly with the silencing RNA.

[0053] In many embodiments the RNA employed in the methods is designed to silence a target gene. However, in related applications similar methods can be used to deliver any nucleic acid of interest, including nucleic acids designed for gene editing, *e. g.*, using a CRISPR or Cas9 system.

A SPRAY APPARATUS FOR APPLYING A COMPOSITION FOR SILENCING A TARGET GENE IN A PLANT

[0054] Another aspect provides a spray apparatus for spraying multiple plants or multiple rows of plants, including a propellant source, at least one spray nozzle, and a reservoir containing a composition including a particulate and a nucleic acid that includes at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence

of a target gene in the plants. Generally, spraying by means of the spray apparatus results in abrading of the plants by the particulate and delivery of the nucleic acid to the plant.

[0055] In embodiments of the spray apparatus, the propellant includes a compressed gas, such as compressed air or nitrogen or other inert gas. In other embodiments, the propellant includes a compressed liquid. Embodiments of the spray apparatus include a canister sprayer, a track sprayer, or a boom sprayer. In embodiments, the spray apparatus is mounted on a vehicle or on other mechanical device to enable movement of the sprayer over the plants or rows of plants.

[0056] In embodiments of the spray apparatus, the nucleic acid is DNA or RNA or a mixture of both. In embodiments, the nucleic acid is synthetic. In embodiments, the nucleic acid is provided in a microbial cell or as a microbial fermentation product. Embodiments of the spray apparatus include those where the nucleic acid is single-stranded DNA or double-stranded DNA. Embodiments include those wherein the DNA includes coding sequence of the target gene, non-coding sequence of the target gene, or both. Embodiments include those wherein the DNA is contained in a plasmid, or in a viral vector. In embodiments, the DNA is non-transcribable, and is not self-replicating. Embodiments of the spray apparatus include those where the nucleic acid is RNA. Embodiments include those wherein the RNA includes single-stranded RNA (ssRNA), double-stranded RNA (dsRNA), or a combination of ssRNA and dsRNA. Other details relevant to nucleic acids of use in the spray apparatus are similar to those described above for the two-step method for silencing a target gene in a plant, as described above under the heading "METHODS FOR SILENCING A TARGET GENE IN A PLANT USING ABRASIVES".

[0057] Embodiments of the spray apparatus include those where the composition further includes one or more components selected from the group consisting of a surfactant, a buffer, an osmoprotectant, a cryoprotectant, an RNase inhibitor (ideally compatible with RNAi activity), a nucleic acid condensing agent (*e. g.*, a polyamine), a transfection agent, a wound-response inhibitor (*e. g.*, callose, hydrogen peroxide inhibitors), a salt (*e. g.*, a Zn^{+2} or Mg^{+2} salt), a plant hormone, a sugar, and a sugar alcohol. Specific embodiments of the composition include one or more components selected from the group consisting of sucrose, mannitol, 2-(*N*-morpholino)ethanesulfonic acid, and Silwet L77.

[0058] Embodiments of the spray apparatus include those where the composition further includes at least one selected from the group consisting of water, a biocide, a chelator, a buffer, a nonionic surfactant, a zwitterionic surfactant, a zinc or magnesium salt, a defoamer, a rainfastness agent, and a photoprotectant. In embodiments, the composition further includes at

least one selected from the group consisting of a carrier agent, a surfactant, an osmolyte, an organosilicone, a polynucleotide herbicidal molecule, a non-polynucleotide herbicidal molecule, a non-polynucleotide pesticide, a safener, an insect attractant, and an insect growth regulator. In embodiments, the composition further includes at least one pesticidal agent selected from the group consisting of a patatin, a plant lectin, a phytoecdysteroid, a *Bacillus thuringiensis* insecticidal protein, a *Xenorhabdus* insecticidal protein, a *Photorhabdus* insecticidal protein, a *Bacillus laterosporous* insecticidal protein, and a *Bacillus sphaericus* insecticidal protein. In embodiments, the composition further includes at least one osmolyte such as an amino acid osmolyte, a methylamine osmolyte, or a polyol or sugar or sugar alcohol osmolyte; non-limiting examples suitable osmolytes include sorbitol, mannitol, xylitol, erythrol, glycerol, glucose, sucrose, proline, valine, isoleucine, ectoine, aspartic acid, trehalose, glycine betaine (betaine), carnitine, taurine, sarcosine, *myo*-inositol (inositol). Alternatively, any of these additional components or combinations thereof can be applied separately from the particulate-RNA composition; in such instances the spray apparatus can include one or more additional reservoirs containing such components.

[0059] Embodiments of the spray apparatus include those where the composition includes a particulate selected from the group consisting of a mineral abrasive, a metal abrasive, a synthetic abrasive, and an organic abrasive. In embodiments, the composition includes particulate abrasives selected from the group consisting of aluminum oxide, silicon carbide (“carborundum”, silicon dioxide, soda lime glass, diatomaceous silica (“diatomaceous earth”), flint, quartz, garnet, silicon dioxide, pumice, sand, feldspar, calcite, steel, tungsten, ceramic, boron carbide, tungsten carbide, an organic or biodegradable abrasive, or combinations of these. In embodiments, the particulate is composed of an organic or biodegradable material, such as, but not limited to wood particles, corn cob particles, grain or seed particles, or nut shell par In embodiments, the particulate used in the composition is of an average size range from about 2.5 micrometers to about 50 micrometers. In various embodiments, the particulate is of an average size range from 2.5 – 50, 2.5 – 40, 2.5 – 30, 2.5 – 20, 5 – 50, 5 – 40, 5 – 30, 5 – 20, 7 – 50, 7 – 40, 7 – 30, 7 – 20, 8 – 50, 8 – 40, 8 – 30, 8 – 20, 10 – 50, 10 – 40, 10 – 30, or 10 – 25 micrometers, as further illustrated in the working Examples.

[0060] The abrasion achieved by use of the spray apparatus preferably does minimal damage to the plant. In embodiments, use of the spray apparatus results in disruption of cells only in the cuticle, or only in the cuticle and epidermis of the plant. In embodiments, cells deeper than the epidermis are essentially not damaged by the particulate abrasive. In

embodiments, use of the spray apparatus results in systemic silencing, wherein the target gene is silenced in at least one location of the plant that is not the location of abrasion.

[0061] Other details relevant to the spray apparatus or to its use are similar to those described above for the two-step method for silencing a target gene in a plant, as described above under the heading “METHODS FOR SILENCING A TARGET GENE IN A PLANT USING ABRASIVES”.

AN APPARATUS FOR INTRODUCING A NUCLEIC ACID INTO A WHOLE PLANT

[0062] Another aspect provides an apparatus for introducing a nucleic acid into a whole plant, including: (a) a matrix supporting an abrasive, and (b) a nucleic acid.

[0063] In embodiments of the apparatus, the matrix and abrasive are permanently bonded to each other. In other embodiments, the matrix and abrasive are not permanently bonded to each other. Embodiments include those wherein the matrix includes a fibrous, porous, non-porous, or adhesive support, and the abrasive includes discrete particles. In embodiments, the matrix includes cotton fiber or other natural fiber. In embodiments, the matrix supporting an abrasive is sandpaper or other solid material incorporating a particulate abrasive permanently bonded to the matrix. In embodiments, the nucleic acid is carried by or affixed to a particulate abrasive. An example of the apparatus includes a matrix including the particulate and RNA, prepared by mixing a loose particulate abrasive with a solution of RNA, drying the resulting mixture, and affixing the dried mixture to a matrix such as a fibrous, porous, non-porous, or adhesive matrix. Another embodiment of the apparatus includes a particulate already affixed to a matrix, to which the RNA is added and the whole then applied to a plant; an example is sandpaper or other particulate-abrasive-bearing material to which RNA is added.

[0064] In embodiments of the apparatus, the nucleic acid is DNA or RNA or a mixture of both. In embodiments, the nucleic acid is synthetic. In embodiments the nucleic acid includes at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant. In embodiments the nucleic acid includes at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene of a pathogen or parasite of the plant. In embodiments, the nucleic acid is provided in a microbial cell or as a microbial fermentation product. Embodiments of the apparatus include those where the nucleic acid is single-stranded DNA or double-stranded DNA. Embodiments include those wherein the DNA includes coding sequence of the target gene, non-coding sequence of the target gene, or both. Embodiments

include those wherein the DNA is contained in a plasmid, or in a viral vector. In embodiments, the DNA is non-transcribable, and is not self-replicating. Embodiments of the apparatus include those where the nucleic acid is RNA. Embodiments include those wherein the RNA includes single-stranded RNA (ssRNA), double-stranded RNA (dsRNA), or a combination of ssRNA and dsRNA. Other details relevant to RNAs of use in the apparatus are similar to those described above for RNAs used in the two-step method for silencing a target gene in a plant, as described above under the heading “METHODS FOR SILENCING A TARGET GENE IN A PLANT USING ABRASIVES”.

[0065] Embodiments of the apparatus include those where the apparatus further includes one or more components selected from the group consisting of a surfactant, a buffer, an osmoprotectant, a cryoprotectant, an RNase inhibitor (ideally compatible with RNAi activity), a nucleic acid condensing agent (*e. g.*, a polyamine), a transfection agent, a wound-response inhibitor (*e. g.*, callose, hydrogen peroxide inhibitors), a salt (*e. g.*, a Zn^{+2} or Mg^{+2} salt), a plant hormone, a sugar, and a sugar alcohol. Specific embodiments of the apparatus include one or more components selected from the group consisting of sucrose, mannitol, 2-(*N*-morpholino)ethanesulfonic acid, and Silwet L77.

[0066] Embodiments of those where the apparatus further includes at least one selected from the group consisting of water, a biocide, a chelator, a buffer, a nonionic surfactant, a zwitterionic surfactant, a zinc or magnesium salt, a defoamer, a rainfastness agent, and a photoprotectant. In embodiments, the apparatus further includes at least one selected from the group consisting of a carrier agent, a surfactant, an osmolyte, an organosilicone, a polynucleotide herbicidal molecule, a non-polynucleotide herbicidal molecule, a non-polynucleotide pesticide, a safener, an insect attractant, and an insect growth regulator. In embodiments, the apparatus further includes at least one pesticidal agent selected from the group consisting of a patatin, a plant lectin, a phytoecdysteroid, a *Bacillus thuringiensis* insecticidal protein, a *Xenorhabdus* insecticidal protein, a *Photorhabdus* insecticidal protein, a *Bacillus laterosporous* insecticidal protein, and a *Bacillus sphaericus* insecticidal protein. In embodiments, the apparatus further includes at least one osmolyte such as an amino acid osmolyte, a methylamine osmolyte, or a polyol or sugar or sugar alcohol osmolyte; non-limiting examples suitable osmolytes include sorbitol, mannitol, xylitol, erythrol, glycerol, glucose, sucrose, proline, valine, isoleucine, ectoine, aspartic acid, trehalose, glycine betaine (betaine), carnitine, taurine, sarcosine, *myo*-inositol (inositol). Alternatively, any of these additional components or combinations thereof can be applied in a separate step.

[0067] Embodiments include those where the abrasive includes a particulate selected from the group consisting of a mineral abrasive, a metal abrasive, a synthetic abrasive, and an organic abrasive. In embodiments, the apparatus includes particulate abrasives selected from the group consisting of aluminum oxide, silicon carbide (“carborundum”, silicon dioxide, soda lime glass, diatomaceous silica (“diatomaceous earth”), flint, quartz, garnet, silicon dioxide, pumice, sand, feldspar, calcite, steel, tungsten, ceramic, boron carbide, tungsten carbide, an organic or biodegradable abrasive, or combinations of these. In embodiments, the particulate is composed of an organic or biodegradable material, such as, but not limited to wood particles, corn cob particles, grain or seed particles, or nut shell par In embodiments, the particulate used in the apparatus is of an average size range from about 2.5 micrometers to about 50 micrometers. In various embodiments, the particulate is of an average size range from 2.5 – 50, 2.5 – 40, 2.5 – 30, 2.5 – 20, 5 – 50, 5 – 40, 5 – 30, 5 – 20, 7 – 50, 7 – 40, 7 – 30, 7 – 20, 8 – 50, 8 – 40, 8 – 30, 8 – 20, 10 – 50, 10 – 40, 10 – 30, or 10 – 25 micrometers, as further illustrated in the working Examples.

[0068] The abrasion achieved by use of the apparatus preferably does minimal damage to the plant. In embodiments, use of the apparatus results in disruption of cells only in the cuticle, or only in the cuticle and epidermis of the plant. In embodiments, cells deeper than the epidermis are essentially not damaged by the abrasive. In embodiments, use of the apparatus results in systemic silencing, wherein the target gene is silenced in at least one location of the plant that is not the location of abrasion.

[0069] Other details relevant to the apparatus or to its use are similar to those described above for the two-step method for silencing a target gene in a plant, as described above under the heading “METHODS FOR SILENCING A TARGET GENE IN A PLANT USING ABRASIVES”.

A METHOD FOR INTRODUCING A NUCLEIC ACID INTO A WHOLE PLANT

[0070] Another aspect provides a method for introducing a nucleic acid into a whole plant, including: (a) applying a composition including a nucleic acid to a surface of a plant, and (b) contacting a matrix supporting an abrasive with the surface; whereby the nucleic acid is introduced into the plant. In embodiments, the composition including a nucleic acid is applied to the plant’s surface and allowed to dry prior to contacting the plant’s surface with a matrix supporting an abrasive. The method is especially useful on intact or whole, growing plants.

[0071] In embodiments of the method, the nucleic acid is DNA or RNA or a mixture of both. In embodiments the nucleic acid includes at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant. In

embodiments the nucleic acid includes at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene of a pathogen or parasite of the plant. In embodiments, the nucleic acid is synthetic. In embodiments, the nucleic acid is provided in a microbial cell or as a microbial fermentation product. Embodiments of the method include those where the nucleic acid is single-stranded DNA or double-stranded DNA. Embodiments include those wherein the DNA includes coding sequence of the target gene, non-coding sequence of the target gene, or both. Embodiments include those wherein the DNA is contained in a plasmid, or in a viral vector. In embodiments, the DNA is non-transcribable, and is not self-replicating. Embodiments of the method include those where the nucleic acid is RNA. Embodiments include those wherein the RNA includes single-stranded RNA (ssRNA), double-stranded RNA (dsRNA), or a combination of ssRNA and dsRNA. Other details relevant to nucleic acids of use in the method are similar to those described above for RNAs used in the two-step method for silencing a target gene in a plant, as described above under the heading “METHODS FOR SILENCING A TARGET GENE IN A PLANT USING ABRASIVES”.

[0072] In embodiments of the method, the composition including a nucleic acid is a liquid, a solid, a powder, a solution, an emulsion, or a suspension. In embodiments, the composition including a nucleic acid further includes one or more components selected from the group consisting of a surfactant, a buffer, an osmoprotectant, a cryoprotectant, an RNase inhibitor (ideally compatible with RNAi activity), a nucleic acid condensing agent (*e. g.*, a polyamine), a transfection agent, a wound-response inhibitor (*e. g.*, callose, hydrogen peroxide inhibitors), a salt (*e. g.*, a Zn^{+2} or Mg^{+2} salt), a plant hormone, a sugar, and a sugar alcohol. In specific embodiments, the composition including a nucleic acid further include one or more components selected from the group consisting of sucrose, mannitol, 2-(*N*-morpholino)ethanesulfonic acid, and Silwet L77.

[0073] In embodiments of the method, the composition including a nucleic acid further includes at least one selected from the group consisting of water, a biocide, a chelator, a buffer, a nonionic surfactant, a zwitterionic surfactant, a zinc or magnesium salt, a defoamer, a rainfastness agent, and a photoprotectant. In embodiments, the composition further includes at least one selected from the group consisting of a carrier agent, a surfactant, an osmolyte, an organosilicone, a polynucleotide herbicidal molecule, a non-polynucleotide herbicidal molecule, a non-polynucleotide pesticide, a safener, an insect attractant, and an insect growth regulator. In embodiments, the composition further includes at least one pesticidal agent selected from the group consisting of a patatin, a plant lectin, a phytoecdysteroid, a *Bacillus thuringiensis* insecticidal protein, a *Xenorhabdus* insecticidal protein, a *Photorhabdus*

insecticidal protein, a *Bacillus laterosporous* insecticidal protein, and a *Bacillus sphaericus* insecticidal protein. In embodiments, the composition further includes at least one osmolyte such as an amino acid osmolyte, a methylamine osmolyte, or a polyol or sugar or sugar alcohol osmolyte; non-limiting examples suitable osmolytes include sorbitol, mannitol, xylitol, erythrol, glycerol, glucose, sucrose, proline, valine, isoleucine, ectoine, aspartic acid, trehalose, glycine betaine (betaine), carnitine, taurine, sarcosine, *myo*-inositol (inositol). Alternatively, any of these additional components or combinations thereof can be applied in a separate step.

[0074] Embodiments of the method include those wherein the matrix supports an abrasive that is permanently bonded to the matrix. In other embodiments, the matrix supports an abrasive that is not permanently bonded to the matrix. Embodiments include those wherein the matrix includes a fibrous, porous, non-porous, or adhesive support, and the abrasive includes discrete particles. In embodiments, the matrix includes cotton fiber or other natural fiber. In embodiments, the matrix supporting an abrasive is sandpaper or other solid material incorporating a particulate abrasive permanently bonded to the matrix. An example of the method includes applying the composition including a nucleic acid to a surface of a whole plant, optionally allowing the applied composition to dry, and then applying a matrix supporting an abrasive (such as a fibrous support carrying a particulate abrasive, or sandpaper, or other particulate-abrasive-bearing material) to the same surface to abrade the surface and allow the nucleic acid to be delivered into the plant.

[0075] Embodiments of the method include those where the abrasive includes a particulate selected from the group consisting of a mineral abrasive, a metal abrasive, a synthetic abrasive, and an organic abrasive. In embodiments, the abrasive includes particulate abrasives selected from the group consisting of aluminum oxide, silicon carbide (“carborundum”, silicon dioxide, soda lime glass, diatomaceous silica (“diatomaceous earth”), flint, quartz, garnet, silicon dioxide, pumice, sand, feldspar, calcite, steel, tungsten, ceramic, boron carbide, tungsten carbide, an organic or biodegradable abrasive, or combinations of these. In embodiments, the particulate is composed of an organic or biodegradable material, such as, but not limited to wood particles, corn cob particles, grain or seed particles, or nut shell par In embodiments, the abrasive used in the method is a particulate abrasive of an average size range from about 2.5 micrometers to about 50 micrometers. In various embodiments, the abrasive used in the method is a particulate is of an average size range from 2.5 – 50, 2.5 – 40, 2.5 – 30, 2.5 – 20, 5 – 50, 5 – 40, 5 – 30, 5 – 20, 7 – 50, 7 – 40, 7 – 30, 7 – 20, 8 – 50, 8 – 40, 8 – 30, 8 – 20, 10 – 50, 10 – 40, 10 – 30, or 10 – 25 micrometers, as further illustrated in the working Examples.

[0076] The method preferably does minimal damage to the plant. In embodiments, the method results in disruption of cells only in the cuticle, or only in the cuticle and epidermis of the plant. In embodiments, cells deeper than the epidermis are essentially not damaged by the method. In embodiments, the method results in systemic silencing, wherein the target gene is
5 silenced in at least one location of the plant that is not the location where the nucleic acid is applied.

[0077] Other details relevant to the method are similar to those described above for the two-step method for silencing a target gene in a plant, as described above under the heading “METHODS FOR SILENCING A TARGET GENE IN A PLANT USING ABRASIVES”, and under the
10 heading “AN APPARATUS FOR INTRODUCING A NUCLEIC ACID INTO A WHOLE PLANT”.

[0078] In embodiments, these methods are useful for obtaining a phenotype (*e. g.*, improved yield, improved resistance temperature, water, or nutrient stress, improved resistance to pathogens, improved herbicide susceptibility, improved herbicide resistance, and modified nutrient content or appearance) resulting from silencing of a target gene in a plant directly
15 treated by these methods. In a related method, the effect of treatment is passed on to subsequent generations, for example where the nucleic acid is designed to target non-coding sequence such as 5' untranslated sequence or a promoter of the target gene, and thereby causing an epigenetic effect. For example, use of a silencing RNA targeting the promoter region of an endogenous phytoene desaturase (PDS) gene in a plant results in bleaching not
20 only of the treated plant but in progeny plants that have not themselves been treated directly with the silencing RNA.

[0079] In many embodiments the nucleic acid employed in the methods is designed to silence a target gene. However, in related applications similar methods can be used to deliver any nucleic acid of interest, including nucleic acids designed for gene editing, *e. g.*, using a
25 CRISPR or Cas9 system.

A METHOD FOR INTRODUCING A NUCLEIC ACID INTO A WHOLE PLANT

[0080] Another aspect provides a method for introducing a nucleic acid into a whole plant, including: (a) applying a composition including an abrasive and a nucleic acid to a
30 surface of a whole plant, and (b) applying pressure to the surface; whereby the nucleic acid is introduced into the plant. The method is especially useful on intact or whole, growing plants. In embodiments, the composition including an abrasive and a nucleic acid is applied to the plant's surface and allowed to dry prior to applying pressure to the plant's surface. In embodiments, the pressure is positive pressure and is applied by mechanical or pneumatic or

hydraulic force, *e. g.*, by means of a pressurized gas or liquid or by means of a solid (such as a roller surface or a flat planar surface) applied to the plant's surface. In other embodiments, the pressure includes negative pressure applied by means of a vacuum, similar to vacuum-assisted *Agrobacterium* infiltration.

5 **[0081]** In embodiments of the method, the nucleic acid in the composition is DNA or RNA or a mixture of both. In embodiments the nucleic acid includes at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant. In embodiments the nucleic acid includes at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene of a
10 pathogen or parasite of the plant. In embodiments, the nucleic acid is synthetic. In embodiments, the nucleic acid is provided in a microbial cell or as a microbial fermentation product. Embodiments of the method include those where the nucleic acid is single-stranded DNA or double-stranded DNA. Embodiments include those wherein the DNA includes coding sequence of the target gene, non-coding sequence of the target gene, or both. Embodiments
15 include those wherein the DNA is contained in a plasmid, or in a viral vector. In embodiments, the DNA is non-transcribable, and is not self-replicating. Embodiments of the method include those where the nucleic acid is RNA. Embodiments include those wherein the RNA includes single-stranded RNA (ssRNA), double-stranded RNA (dsRNA), or a combination of ssRNA and dsRNA. Other details relevant to nucleic acids of use in the method are similar to those
20 described above for RNAs used in the two-step method for silencing a target gene in a plant, as described above under the heading "METHODS FOR SILENCING A TARGET GENE IN A PLANT USING ABRASIVES".

[0082] In embodiments of the method, the composition including an abrasive and a nucleic acid is a liquid, a solid, a powder, a solution, an emulsion, or a suspension. In
25 embodiments, the composition is a solid or a dry composition, such as a powder. In embodiments, the composition is a suspension of the particulate and the nucleic acid in a liquid or a gel. In embodiments, the composition is an emulsion. In embodiments the nucleic acid is physically closely associated with or in contact with the particulate, *e. g.*, simply by drying the nucleic acid onto the particulate, or by bonding the nucleic acid to the particulate by ionic or
30 electrostatic attraction, direct covalent bonding, covalent bonding using a linker, or by means of an adhesive or other affixing agent.

[0083] In embodiments, the composition further includes one or more components selected from the group consisting of a surfactant, a buffer, an osmoprotectant, a cryoprotectant, an RNase inhibitor (ideally compatible with RNAi activity), a nucleic acid

condensing agent (*e. g.*, a polyamine), a transfection agent, a wound-response inhibitor (*e. g.*, callose, hydrogen peroxide inhibitors), a salt (*e. g.*, a Zn^{+2} or Mg^{+2} salt), a plant hormone, a sugar, and a sugar alcohol. In specific embodiments, the composition further includes one or more components selected from the group consisting of sucrose, mannitol, 2-(*N*-

morpholino)ethanesulfonic acid, and Silwet L77.

[0084] In embodiments of the method, the composition including an abrasive and a nucleic acid further includes at least one selected from the group consisting of water, a biocide, a chelator, a buffer, a nonionic surfactant, a zwitterionic surfactant, a zinc or magnesium salt, a defoamer, a rainfastness agent, and a photoprotectant. In embodiments, the composition further includes at least one selected from the group consisting of a carrier agent, a surfactant, an osmolyte, an organosilicone, a polynucleotide herbicidal molecule, a non-polynucleotide herbicidal molecule, a non-polynucleotide pesticide, a safener, an insect attractant, and an insect growth regulator. In embodiments, the composition further includes at least one pesticidal agent selected from the group consisting of a patatin, a plant lectin, a phytoecdysteroid, a *Bacillus thuringiensis* insecticidal protein, a *Xenorhabdus* insecticidal protein, a *Photorhabdus* insecticidal protein, a *Bacillus laterosporous* insecticidal protein, and a *Bacillus sphaericus* insecticidal protein. In embodiments, the composition further includes at least one osmolyte such as an amino acid osmolyte, a methylamine osmolyte, or a polyol or sugar or sugar alcohol osmolyte; non-limiting examples suitable osmolytes include sorbitol, mannitol, xylitol, erythrol, glycerol, glucose, sucrose, proline, valine, isoleucine, ectoine, aspartic acid, trehalose, glycine betaine (betaine), carnitine, taurine, sarcosine, *myo*-inositol (inositol). Alternatively, any of these additional components or combinations thereof can be applied in a separate step.

[0085] Embodiments of the method include those where the abrasive includes a particulate selected from the group consisting of a mineral abrasive, a metal abrasive, a synthetic abrasive, and an organic abrasive. In embodiments, the abrasive includes particulate abrasives selected from the group consisting of aluminum oxide, silicon carbide (“carborundum”, silicon dioxide, soda lime glass, diatomaceous silica (“diatomaceous earth”), flint, quartz, garnet, silicon dioxide, pumice, sand, feldspar, calcite, steel, tungsten, ceramic, boron carbide, tungsten carbide, an organic or biodegradable abrasive, or combinations of these. In embodiments, the particulate is composed of an organic or biodegradable material, such as, but not limited to wood particles, corn cob particles, grain or seed particles, or nut shell par In embodiments, the abrasive used in the method is a particulate abrasive of an average size range from about 2.5 micrometers to about 50 micrometers. In various

embodiments, the abrasive used in the method is a particulate is of an average size range from 2.5 – 50, 2.5 – 40, 2.5 – 30, 2.5 – 20, 5 – 50, 5 – 40, 5 – 30, 5 – 20, 7 – 50, 7 – 40, 7 – 30, 7 – 20, 8 – 50, 8 – 40, 8 – 30, 8 – 20, 10 – 50, 10 – 40, 10 – 30, or 10 – 25 micrometers, as further illustrated in the working Examples. In embodiments, the abrasive includes discrete particles.

5 **[0086]** The method preferably does minimal damage to the plant. In embodiments, the method results in disruption of cells only in the cuticle, or only in the cuticle and epidermis of the plant. In embodiments, cells deeper than the epidermis are essentially not damaged by the method. In embodiments, the method results in systemic silencing, wherein the target gene is silenced in at least one location of the plant that is not the location where the nucleic acid is
10 applied.

[0087] Other details relevant to the method are similar to those described above for the two-step method for silencing a target gene in a plant, as described above under the heading “METHODS FOR SILENCING A TARGET GENE IN A PLANT USING ABRASIVES” and for the one-step
15 method for silencing a target gene in a plant, as described above under the heading “SINGLE-STEP METHODS FOR SILENCING A TARGET GENE IN A PLANT USING ABRASIVES”.

[0088] In embodiments, these methods are useful for obtaining a phenotype (*e. g.*, improved yield, improved resistance temperature, water, or nutrient stress, improved resistance to pathogens, improved herbicide susceptibility, improved herbicide resistance, and modified nutrient content or appearance) resulting from silencing of a target gene in a plant directly
20 treated by these methods. In a related method, the effect of treatment is passed on to subsequent generations, for example where the nucleic acid is designed to target non-coding sequence such as 5' untranslated sequence or a promoter of the target gene, and thereby causing an epigenetic effect. For example, use of a silencing RNA targeting the promoter region of an endogenous phytoene desaturase (PDS) gene in a plant results in bleaching not
25 only of the treated plant but in progeny plants that have not themselves been treated directly with the silencing RNA.

[0089] In many embodiments the nucleic acid employed in the methods is designed to silence a target gene. However, in related applications similar methods can be used to deliver any nucleic acid of interest, including nucleic acids designed for gene editing, *e. g.*, using a
30 CRISPR or Cas9 system.

A METHOD FOR INTRODUCING A NUCLEIC ACID INTO A WHOLE PLANT

[0090] Another aspect provides a method for introducing a nucleic acid into a whole plant including, in any order, the steps of: (a) mechanical penetration of a surface of a whole

plant with a non-particulate microstructure, and (b) contacting the surface of a whole plant with a nucleic acid. In embodiments, the non-particulate microstructure is removed after it has penetrated the plant, so that no part of the non-particulate microstructure remains within the plant. The method is especially useful on intact or whole, growing plants.

5 **[0091]** In embodiments, step (a) is followed by step (b). In other embodiments, step (b) is followed by step (a). In embodiments where the nucleic acid is applied prior to the abrasion step, the nucleic acid can be allowed to dry on the plant surface prior to abrasion. Additional treatments of the plant (for example, treatment with a surfactant, an nuclease inhibitor, a photoprotectant, or a rainfastness agent) are optionally performed, subsequent to
10 either abrasion or application of nucleic acid, or concomitant with either or both steps. A related, one-step method includes the step of mechanically penetrating the a surface of a whole plant with a non-particulate microstructure bearing a nucleic acid, whereby the nucleic acid is delivered into the plant.

15 **[0092]** In embodiments of the method, the nucleic acid in the composition is DNA or RNA or a mixture of both. In embodiments the nucleic acid includes at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant. In embodiments the nucleic acid includes at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene of a pathogen or parasite of the plant. In embodiments, the nucleic acid is synthetic. In
20 embodiments, the nucleic acid is provided in a microbial cell or as a microbial fermentation product. Embodiments of the method include those where the nucleic acid is single-stranded DNA or double-stranded DNA. Embodiments include those wherein the DNA includes coding sequence of the target gene, non-coding sequence of the target gene, or both. Embodiments include those wherein the DNA is contained in a plasmid, or in a viral vector. In embodiments,
25 the DNA is non-transcribable, and is not self-replicating. Embodiments of the method include those where the nucleic acid is RNA. Embodiments include those wherein the RNA includes single-stranded RNA (ssRNA), double-stranded RNA (dsRNA), or a combination of ssRNA and dsRNA. Other details relevant to nucleic acids of use in the method are similar to those described above for RNAs used in the two-step method for silencing a target gene in a plant, as
30 described above under the heading “METHODS FOR SILENCING A TARGET GENE IN A PLANT USING ABRASIVES”.

[0093] In embodiments, the nucleic acid is ssRNA or dsRNA designed to silence a single target gene. In embodiments, the nucleic acid is ssRNA or dsRNA including multiple repeating segments for targeting a single target gene. Multiple target genes (which can be

target genes of the plant, or of a pathogen or parasite of the plant, or a combination of both), or multiple regions of a single target gene, can be targeted by a chimeric nucleic acid, such as by a chimeric RNA. In an example, the nucleic acid is a chimeric dsRNA including dsRNA segments targeting a gene expressed in the plant and dsRNA segments targeting a gene of a pathogen or parasite of the plant.

[0094] In embodiments of the method, the nucleic acid is in the form of a liquid, a solid, a powder, a solution, an emulsion, or a suspension. In embodiments, the nucleic acid is in the form of a solid or a dry composition, such as a powder. In embodiments, the nucleic acid is in the form of a suspension of the nucleic acid in a liquid or a gel. In embodiments, the nucleic acid is in the form of an emulsion. In embodiments the nucleic acid is physically closely associated with or in contact with the non-particulate microstructure, *e. g.*, simply by drying the nucleic acid onto the non-particulate microstructure, or by bonding the nucleic acid to the particulate by ionic or electrostatic attraction, or by means of an adhesive or other affixing agent. In embodiments the nucleic acid is bonded to the non-particulate microstructure by direct covalent bonding or by covalent bonding using a linker, wherein the covalent bond can be broken (*e. g.*, by enzymatic action) in order to deliver the nucleic acid to the plant.

[0095] In embodiments, the nucleic acid is in the form of a composition wherein the composition further includes one or more components selected from the group consisting of a surfactant, a buffer, an osmoprotectant, a cryoprotectant, an RNase inhibitor (ideally compatible with RNAi activity), a nucleic acid condensing agent (*e. g.*, a polyamine), a transfection agent, a wound-response inhibitor (*e. g.*, callose, hydrogen peroxide inhibitors), a salt (*e. g.*, a Zn^{+2} or Mg^{+2} salt), a plant hormone, a sugar, and a sugar alcohol. In specific embodiments, the nucleic acid is in the form of a composition wherein the composition further includes one or more components selected from the group consisting of sucrose, mannitol, 2-(*N*-morpholino)ethanesulfonic acid, and Silwet L77.

[0096] In embodiments of the method, the nucleic acid is in the form of a composition wherein the composition further includes at least one selected from the group consisting of water, a biocide, a chelator, a buffer, a nonionic surfactant, a zwitterionic surfactant, a zinc or magnesium salt, a defoamer, a rainfastness agent, and a photoprotectant. In embodiments, the composition further includes at least one selected from the group consisting of a carrier agent, a surfactant, an osmolyte, an organosilicone, a polynucleotide herbicidal molecule, a non-polynucleotide herbicidal molecule, a non-polynucleotide pesticide, a safener, an insect attractant, and an insect growth regulator. In embodiments, the composition further includes at

least one pesticidal agent selected from the group consisting of a patatin, a plant lectin, a phytoecdysteroid, a *Bacillus thuringiensis* insecticidal protein, a *Xenorhabdus* insecticidal protein, a *Photorhabdus* insecticidal protein, a *Bacillus laterosporous* insecticidal protein, and a *Bacillus sphaericus* insecticidal protein. In embodiments, the composition further includes at

5 least one osmolyte such as an amino acid osmolyte, a methylamine osmolyte, or a polyol or sugar or sugar alcohol osmolyte; non-limiting examples suitable osmolytes include sorbitol, mannitol, xylitol, erythrol, glycerol, glucose, sucrose, proline, valine, isoleucine, ectoine, aspartic acid, trehalose, glycine betaine (betaine), carnitine, taurine, sarcosine, *myo*-inositol (inositol). Alternatively, any of these additional components or combinations thereof can be
10 applied in a separate step.

[0097] Embodiments of the method include those where the non-particulate microstructure includes a matrix or apparatus on which is supported at least one selected from the group consisting of micro- or nano- needles, -whiskers, -fibers, and -wires.

[0098] The method preferably does minimal damage to the plant. In embodiments, the
15 method results in disruption of cells only in the cuticle, or only in the cuticle and epidermis of the plant. In embodiments, cells deeper than the epidermis are essentially not damaged by the method. In embodiments, the method results in systemic silencing, wherein the target gene is silenced in at least one location of the plant that is not the location where the nucleic acid is applied.

[0099] Other details relevant to the method are similar to those described above for the
20 two-step method for silencing a target gene in a plant, as described above under the heading “METHODS FOR SILENCING A TARGET GENE IN A PLANT USING ABRASIVES” and for the one-step method for silencing a target gene in a plant, as described above under the heading “SINGLE-STEP METHODS FOR SILENCING A TARGET GENE IN A PLANT USING ABRASIVES”.

[0100] In embodiments, these methods are useful for obtaining a phenotype (*e. g.*,
25 improved yield, improved resistance temperature, water, or nutrient stress, improved resistance to pathogens, improved herbicide susceptibility, improved herbicide resistance, and modified nutrient content or appearance) resulting from silencing of a target gene in a plant directly treated by these methods. In a related method, the effect of treatment is passed on to
30 subsequent generations, for example where the nucleic acid is designed to target non-coding sequence such as 5' untranslated sequence or a promoter of the target gene, and thereby causing an epigenetic effect. For example, use of a silencing RNA targeting the promoter region of an endogenous phytoene desaturase (PDS) gene in a plant results in bleaching not

only of the treated plant but in progeny plants that have not themselves been treated directly with the silencing RNA.

[0101] In many embodiments the nucleic acid employed in the methods is designed to silence a target gene. However, in related applications similar methods can be used to deliver any nucleic acid of interest, including nucleic acids designed for gene editing, *e. g.*, using a CRISPR or Cas9 system.

REGENERATED PLANTS AND PROGENY THEREOF

[0102] Another aspect provides for providing a plant exhibiting modification of a target gene in the plant, comprising directly regenerating a plant from a source plant, wherein the source plant has been contacted with a composition comprising an RNA that comprises at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant, wherein the directly regenerated plant exhibits modification of the target gene. Related aspects provide a plant exhibiting modification of a target gene in the plant, comprising directly regenerating a plant from a source plant, wherein the source plant has been contacted with a composition comprising at least one nucleic acid (an RNA or a DNA or a combination of RNA and DNA) that comprises at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant, wherein the directly regenerated plant exhibits modification of the target gene.

[0103] In embodiments, the source plant is a growing seedling or a growing plant in a post-seedling growth stage. In embodiments, the source plant is an intact plant. Embodiments include use of the various methods, compositions, and apparatuses described herein. In embodiments, the source plant is topically treated with the RNA.

[0104] In embodiments, the modification of the target gene is (1) non-heritable silencing of the target gene, (2) heritable or epigenetic silencing of the target gene, or (3) a change in the nucleotide sequence of the target gene. In embodiments, the directly regenerated plant exhibits partial silencing of the target gene, or the directly regenerated plant exhibits complete systemic silencing of the target gene, or the directly regenerated plant exhibits a homogeneous morphological or physiological phenotype resulting from modification of the target gene. In embodiments, the modification of the target gene is heritable to plants of subsequent generations grown from the directly regenerated plant. Thus, aspects include the directly regenerated plant exhibiting modification of the target gene as well as plants of subsequent generations exhibiting modification of the target gene.

RELATED INFORMATION AND TECHNIQUES

Plants

[0105] The methods, apparatuses, and compositions described herein are useful across a broad range of plants. Suitable plants in which the methods apparatuses, and compositions disclosed herein can be used include, but are not limited to, cereals and forage grasses (rice, maize, wheat, barley, oat, sorghum, pearl millet, finger millet, cool-season forage grasses, and bahiagrass), oilseed crops (soybean, oilseed brassicas including canola and oilseed rape, sunflower, peanut, flax, sesame, and safflower), legume grains and forages (common bean, cowpea, pea, faba bean, lentil, tepary bean, Asiatic beans, pigeonpea, vetch, chickpea, lupine, alfalfa, and clovers), temperate fruits and nuts (apple, pear, peach, plums, berry crops, cherries, grapes, olive, almond, and Persian walnut), tropical and subtropical fruits and nuts (citrus including limes, oranges, and grapefruit; banana and plantain, pineapple, papaya, mango, avocado, kiwifruit, passionfruit, and persimmon), vegetable crops (solanaceous plants including tomato, eggplant, and peppers; vegetable brassicas; radish, carrot, cucurbits, alliums, asparagus, and leafy vegetables), sugar, tuber, and fiber crops (sugarcane, sugar beet, stevia, potato, sweet potato, cassava, and cotton), plantation crops, ornamentals, and turf grasses (tobacco, coffee, cocoa, tea, rubber tree, medicinal plants, ornamentals, and turf grasses), and forest tree species.

[0106] In certain embodiments, the plant is a weedy plant. Weedy plants are plants that compete with cultivated plants, those of particular importance include, but are not limited to important invasive and noxious weeds and herbicide resistant biotypes in crop production, such as, *Amaranthus* species -*A. albus*, *A. blitoides*, *A. hybridus*, *A. palmeri*, *A. powellii*, *A. retroflexus*, *A. spinosus*, *A. tuberculatus*, and *A. viridis*; *Ambrosia* species - *A. trifida*, *A. artemisifolia*; *Lolium* species -*L. multiflorum*, *L. rigidum*, *L. perenne*; *Digitaria* species -*D. insularis*; *Euphorbia* species -*E. heterophylla*; *Kochia* species - *K. scoparia*; *Sorghum* species -*S. halepense*; *Conyza* species -*C. bonariensis*, *C. canadensis*, *C. sumatrensis*; *Chloris* species -*C. truncate*; *Echinochola* species - *E. colona*, *E. crus-galli*; *Eleusine* species -*E. indica*; *Poa* species -*P. annua*; *Plantago* species -*P. lanceolata*; *Avena* species - *A. fatua*; *Chenopodium* species - *C. album*; *Setaria* species - *S. viridis*, *Abutilon theophrasti*, *Ipomoea* species, *Sesbania*, species, *Cassia* species, *Sida* species, *Brachiaria*, species and *Solanum* species.

Additional weedy plant species found in cultivated areas include *Alopecurus myosuroides*, *Avena sterilis*, *Avena sterilis ludoviciana*, *Brachiaria plantaginea*, *Bromus diandrus*, *Bromus rigidus*, *Cynosurus echinatus*, *Digitaria ciliaris*, *Digitaria ischaemum*,

- Digitaria sanguinalis*, *Echinochloa oryzicola*, *Echinochloa phyllopogon*, *Eriochloa punctata*, *Hordeum glaucum*, *Hordeum leporinum*, *Ischaemum rugosum*, *Leptochloa chinensis*, *Lolium persicum*, , *Phalaris minor*, *Phalaris paradoxa*, *Rottboellia exalta*, *Setaria faberi*, *Setaria viridis* var, *robusta-alba schreiber*, *Setaria viridis* var, *robusta -purpurea*, *Snowdenia polystachea*, *Sorghum sudanese*, *Alisma plantago-aquatica*, *Amaranthus lividus*, *Amaranthus quitensis*, *Ammania auriculata*, *Ammania coccinea*, *Anthemis cotula*, *Apera spica-venti*, *Bacopa rotundifolia*, *Bidens pilosa*, *Bidens subalternans*, *Brassica tournefortii*, *Bromus tectorum*, *Camelina microcarpa*, *Chrysanthemum coronarium*, *Cuscuta campestris*, *Cyperus difformis*, *Damasonium minus*, *Descurainia sophia*, *Diplotaxis tenuifolia*, *Echium plantagineum*, *Elatine triandra* var, *pedicellata*, *Euphorbia heterophylla*, *Fallopia convolvulus*, *Fimbristylis miliacea*, *Galeopsis tetrahit*, *Galium spurium*, *Helianthus annuus*, *Iva xanthifolia*, *Ixophorus unisetus*, *Ipomoea indica*, *Ipomoea purpurea*, *Ipomoea sepiaria*, *Ipomoea aquatic*, *Ipomoea triloba*, *Lactuca serriola*, *Limncharis flava*, *Limnophila erecta*, *Limnophila sessiliflora*, *Lindernia dubia*, *Lindernia dubia* var, *major*, *Lindernia micrantha*, *Lindernia procumbens*, *Mesembryanthemum crystallinum*, *Monochoria korsakowii*, *Monochoria vaginalis*, *Neslia paniculata*, *Papaver rhoeas*, *Parthenium hysterophorus*, *Pentzia suffruticosa*, *Phalaris minor*, *Raphanus raphanistrum*, *Raphanus sativus*, *Rapistrum rugosum*, *Rotala indica* var, *uliginosa*, *Sagittaria guyanensis*, *Sagittaria montevidensis*, *Sagittaria pygmaea*, *Salsola iberica*, *Scirpus juncoides* var, *ohwianus*, *Scirpus mucronatus*, *Setaria lutescens*, *Sida spinosa*, *Sinapis arvensis*, *Sisymbrium orientale*, *Sisymbrium thellungii*, *Solanum ptycanthum*, *Sonchus asper*, *Sonchus oleraceus*, *Sorghum bicolor*, *Stellaria media*, *Thlaspi arvense*, *Xanthium strumarium*, *Arctotheca calendula*, *Conyza sumatrensis*, *Crassocephalum crepidiodes*, *Cuphea carthagenensis*, *Epilobium adenocaulon*, *Erigeron philadelphicus*, *Landoltia punctata*, *Lepidium virginicum*, *Monochoria korsakowii*, *Solanum americanum*, *Solanum nigrum*, *Vulpia bromoides*, *Youngia japonica*, *Hydrilla verticillata*, *Carduus nutans*, *Carduus pycnocephalus*, *Centaurea solstitialis*, *Cirsium arvense*, *Commelina diffusa*, *Convolvulus arvensis*, *Daucus carota*, *Digitaria ischaemum*, *Echinochloa crus-pavonis*, *Fimbristylis miliacea*, *Galeopsis tetrahit*, *Galium spurium*, *Limnophila erecta*, *Matricaria perforate*, *Papaver rhoeas*, *Ranunculus acris*, *Soliva sessilis*, *Sphenoclea zeylanica*, *Stellaria media*, *Nassella trichotoma*, *Stipa neesiana*, *Agrostis stolonifera*, *Polygonum aviculare*, *Alopecurus japonicus*, *Beckmannia syzigachne*, *Bromus tectorum*, *Chloris inflata*, *Echinochloa erecta*, *Portulaca oleracea*, and *Senecio vulgaris*.

Additional Nucleic Acid Elements

[0107] Embodiments of the nucleic acids useful in the compositions and methods described herein can include additional elements, such as promoters, small RNA recognition sites, aptamers or ribozymes, additional and additional expression cassettes for expressing coding sequences (*e. g.*, to express a transgene such as an insecticidal protein or selectable marker) or non-coding sequences (*e. g.*, to express additional suppression elements). In an embodiment, a nucleic acid useful in methods or compositions as described herein includes a recombinant DNA construct including a heterologous promoter operably linked to DNA encoding an RNA transcript including at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant or to the sequence of a target gene of a pathogen or parasite of the plant. In an embodiment, a nucleic acid useful in methods and compositions described herein is a DNA or RNA including an aptamer that serves to guide the nucleic acid to a desired location in the plant. In an embodiment, a nucleic acid useful in methods and compositions described herein is an RNA molecule including one or more recognition sites for binding and cleavage by a small RNA (*e. g.*, by a miRNA or an siRNA that is expressed only in a particular cell or tissue), which allows for more precise control of the suppression of the target gene in the plant.

EXAMPLES

Example 1

[0108] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes silencing a target gene by way of particulate-assisted delivery of a nucleic acid such as an RNA “trigger” or silencing element into a plant. The methods generally involve treatment of the surface of a plant (or of plant cells or tissues) with an abrasive or particulate, and with a nucleic acid.

[0109] Four dsRNA “triggers” (silencing elements) of 50, 78, 124, and 249 base-pairs (bp) (SEQ ID NOs:1, 2, 3, and 5, respectively), and targeting green fluorescent protein (GFP) were used to silence the GFP gene in a transgenic *Nicotiana benthamiana* line (16c) expressing GFP. For each trigger, 420 micrograms of total RNA were dissolved in 210 microliters; 10 microliters were removed for later analysis and the remaining 200 microliters was added to 200 milligrams of aluminum oxide (~220 mesh) particles in a 15 milliliter culture tube. The preparation was incubated overnight at 37 degrees Celsius, then centrifuged at 250 rpm with the cap off. One milliliter of 100% ethanol was added to transfer the RNA-coated aluminum oxide particles into a weighing tray; excess liquid was removed by pipette and the particles

allowed to air-dry. Each preparation of the dry particles was loaded into the chamber of an airbrush and sprayed at 45-65 pounds per square inch (psi) onto a single leaf of each of six plants. Local silencing in the treated leaf was observed in 3 of the 6 plants sprayed with the 124 bp dsRNA trigger, but not in the plants treated with the 50 or 78 bp dsRNA triggers. No silencing was observed in plants treated with the 249 bp dsRNA trigger but these results were not considered based on subsequent analysis of trigger quality. Systemic GFP silencing (outside of the treated leaves) was not observed in this experiment.

Example 2

[0110] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes silencing a target gene by way of particulate-assisted delivery of a nucleic acid such as an RNA “trigger” or silencing element into a plant.

[0111] In another example, 1500 micrograms of dsRNA trigger in 1 milliliter water was added to 200 milligrams of aluminum oxide (320 mesh (20.1 – 23.1 micrometers) or 400 mesh (15.5 – 17.5 micrometers)) in a 6-well plate and incubated overnight at room temperature on a shaker (150 rpm). One milliliter of 100% ethanol was added to transfer the RNA-coated aluminum oxide particles into a weighing tray; excess liquid was removed by pipette and the particles allowed to air-dry. Each preparation of the dry particles was loaded into the chamber of an airbrush and sprayed at 55 pounds per square inch (psi) onto leaves of nine transgenic *Nicotiana benthamiana* 16c plants. Results are provided in **Table 3**. Local silencing in the treated leaf was observed in nearly all plants treated with the GFP dsRNA trigger (**SEQ ID NO:3**), with less efficient GFP silencing observed in the plants treated with the GFP/PDS fusion dsRNA trigger (**SEQ ID NO:7**, which contains the intact sequence of **SEQ ID NO:3** at its 3’ end). The larger particle size (320 mesh) provided better silencing efficiency than the smaller particles (400 mesh). Systemic GFP silencing (outside of the treated leaves) was not observed in this experiment.

Table 3

SEQ ID NO:*	dsRNA trigger size (base pairs)	Target gene	Aluminum oxide mesh size	Number of plants where GFP silencing was observed
3	124	GFP	320	9/9
3	124	GFP	400	7/9
7	300	GFP/PDS fusion	320	5/9
7	300	GFP/PDS fusion	400	2/9

*sequence of anti-sense strand, 5'→3'

Example 3

5 **[0112]** This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene by way of particulate-assisted delivery of a nucleic acid such as an RNA “trigger” or silencing element into a plant.

10 **[0113]** In another example, 1.5 milligrams of total RNA (124 bp dsRNA, **SEQ ID NOs:3**) were coated onto aluminum oxide or silicon carbide particles and applied using an airbrush spray (65 psi) onto 9 two- to three-week-old transgenic *Nicotiana benthamiana* 16c plants. Phenotype was recorded 17 days after the treatment. Plants showing GFP silencing (red spots/sectors under ultraviolet light) on sprayed leaves only were scored as displaying

15 local silencing. Plants additionally showing GFP silencing (red spots/sectors under ultraviolet light) in parts of the plants other than the sprayed leaves were scored as displaying systemic silencing; in this experiment the systemic silencing was observed as a vasculature-associated GFP silencing pattern in newly grown leaves. Results are provided in **Table 4**.

Table 4

SEQ ID NO:*	Particulate type	Particulate mesh size	Number of plants displaying local silencing	Number of plants displaying systemic silencing	Number of plants displaying no silencing
3	Al ₂ O ₃	320	8/9	1/9	0/9
3	Al ₂ O ₃	360	7/9	1/9	1/9
3	SiC	320	8/9	1/9	0/9
3	SiC	360	6/9	3/9	0/9

*sequence of anti-sense strand, 5'→3'

Example 4

[0114] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene by way of particulate-assisted delivery of a nucleic acid such as an RNA “trigger” or silencing element into a plant.

[0115] In another example, different RNA triggers designed to silence GFP were compared. Several triggers were blunt-ended dsRNAs; one (**SEQ ID NO:8**) was a single-stranded miRNA precursor transcript designed to self-hybridize and be processed to produce a mature miRNA targeting GFP. For each RNA trigger, 1.5 milligrams of total RNA were coated onto SiC particles. Each individual RNA trigger was dissolved in water to make up 1 milliliter, added to 200 milligrams SiC (320 mesh) in a well of a 6-well plate. The plate was placed in a fume hood to air-dry with gentle shaking. After the plate was completely dry, 100% ethanol was added to transfer the RNA-coated SiC particles into a weighing tray; excess liquid was removed by pipette and the particles allowed to air-dry overnight. The dried RNA-coated particles were transferred to 2-milliliter microcentrifuge tubes, ground briefly in the tubes, and applied using an airbrush spray (60 psi) onto 9 three-week-old transgenic *Nicotiana benthamiana* 16c plants. Local silencing was observed beginning at 4 – 5 days after treatment. Phenotype was recorded at 9 days (for local silencing) and at 19 days (for systemic silencing) after treatment. In this experiment, systemic silencing was again observed as a vasculature-associated GFP silencing pattern in newly grown leaves. Results are provided in **Table 5**.

Table 5

SEQ ID NO:	dsRNA trigger size (base pairs)	Target gene	Number of plants displaying local silencing	Number of plants displaying systemic silencing	Number of plants displaying no silencing
1	50	GFP	4/9	0/9	5/9
2	78	GFP	8/9	2/9	1/9
3	124	GFP	9/9	5/9	0/9
4	125	GFP	4/9	0/9	5/9
5	249	GFP	3/9	0/9	6/9
8**	355	GFP	1/9	0/9	8/9
6	258	PDS	0/9	0/9	9/9
--	--	(none)	0/9	0/9	9/9
--	--	(none)	0/9	0/9	9/9

*sequence of anti-sense strand, 5'→3'

****SEQ ID NO:8** is a synthetic miRNA precursor; the sequence is of the complete transcript, 5'→3'

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Example 5

[0116] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene by way of particulate-assisted delivery of a nucleic acid such as DNA viral vector into a plant.

[0117] A viral vector was used to silence either a green fluorescent protein (GFP) transgene or an endogenous phytoene desaturase (PDS) target gene in treated plants. Plasmid A1 (**SEQ ID NO:9**) targeting PDS or plasmid A2 (**SEQ ID NO:10**) targeting GFP was combined with plasmid B (ToGMoV DNA-B in the pUC19 vector, **SEQ ID NO:11**) to produce a VIGS system. 250 micrograms of either plasmid A1 (**SEQ ID NO:9**) or plasmid A2 (**SEQ ID NO:10**) was added to 250 micrograms plasmid B (**SEQ ID NO:11**) in 600 microliters water. The DNA mixtures were each added to 150 milligrams of aluminum oxide particles (400 mesh or 600 mesh) in wells of a 6-well plate and incubated overnight at room temperature on a shaker (150 rpm) in a fume hood to air dry. After the plate was completely dry, 1 milliliter of 70% ethanol was added to transfer the RNA-coated aluminum oxide particles into a weighing tray; excess liquid was removed by pipette and the particles allowed to air-dry. Each preparation of the dried DNA-coated particles was applied using an airbrush spray (55 psi) onto six transgenic *Nicotiana benthamiana* 16c plants. Results are shown in **Table 6**. The results demonstrate that particle-assisted delivery of a viral vector results in systemic silencing of transgenes or endogenous genes expressed in a whole plant. This

technique is useful in other applications, such as in virus resistance assays, as the method does not involve *Agrobacterium*-mediated infection.

Table 6

Plasmid ID	SEQ ID NO:	Target gene	Aluminum oxide mesh size	Number of plants displaying systemic PDS silencing	Number of plants displaying systemic GFP silencing
A1	9	PDS	400	5/6	--
A2	10	GFP	400	--	4/6
A1	9	PDS	600	1/6	--
A2	10	GFP	600	--	6/6

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Example 6

[0118] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene by way of abrasion of a plant surface with particulates to disrupt the cuticle or epidermis, thereby delivering a nucleic acid such as an RNA “trigger” or silencing element into a plant.

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[0119] Double-stranded RNA was labelled with Cy3 as a fluorescent marker and coated onto SiC particles (320 mesh) which were then sprayed onto a leaf. The leaf was imaged with confocal fluorescence microscopy 1 day after treatment. The images obtained showed that the fluorescently labelled particles were located at the bottom of “craters” formed by the particle impact some layers deep in the leaf epidermis and suggested that, while most of the fluorescence was still associated with the particles, some of the fluorescence diffused into adjacent undamaged cells. The images suggest that the nucleic acid on the particles is not delivered directly into cells in the manner seen with gene gun delivery using much smaller particles, but by diffusion into cells adjacent to the larger particles used here with relatively low-pressure delivery.

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Example 7

[0120] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene by way of particulate-assisted delivery of a nucleic acid such as an RNA “trigger” or silencing element into a plant.

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[0121] This experiment compared varying distances between airbrush nozzle and plant surface. 1.5 milligrams of blunt-ended dsRNA targeting GFP (SEQ ID NO:3) was coated onto 100 milligrams of silicon carbide (360 mesh) and air dried overnight. After drying, the mixture was ground to singulate the particles, and loaded into a G78 airbrush mounted to a ring stand. Transgenic *Nicotiana benthamiana* 16c plants were each sprayed with three 1-second bursts at 3, 5, and 7 centimeters nozzle-to-leaf distance). Phenotype (GFP silencing) was visually assessed using blue light excitation 7 days after treatment. In addition, GFP expression was quantified in the red (silenced) and green (non-silenced) sectors using qPCR. Results: the 3-centimeter spray distance damaged the plants and resulted in little silencing; approximately equivalent silencing was observed with the 5- and 7-centimeter spray distances. The qPCR measurements demonstrated that GFP expression was correlated to visual phenotype (Figure 1).

Example 8

[0122] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene by way of particulate-assisted delivery of a nucleic acid such as an RNA “trigger” or silencing element into a plant.

[0123] An experiment to test sequential application of RNA and particulate abrasive was performed. Blunt-ended dsRNA targeting GFP (SEQ ID NO:3) was dissolved in water at 1, 5, and 10 milligrams/milliliter, with a silicone surfactant (0.1% Silwet L77) added to aid spreading on the leaf surface. 20 microliters of the RNA solution was applied to three leaves of transgenic *Nicotiana benthamiana* 16c plants and allowed to dry briefly. Dry uncoated silicon carbide (360 mesh) particles were sprayed onto the RNA-coated leaves at 60 psi using a G78 airbrush mounted to a ring stand at 5 centimeters nozzle-to-leaf distance from the plants. GFP silencing was assessed visually using blue light excitation at 7 days after treatment. Leaf damage prevented full interpretation of the dsRNA rate data, but GFP silencing was observed using this sequential method, where applying an RNA to the surface of the plant is followed by abrading the surface of a plant with a particulate of a size greater than about 2.5 micrometers.

Example 9

[0124] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant.

More specifically, this example describes systemic silencing of a target gene by way of particulate-assisted delivery of a nucleic acid such as an RNA “trigger” or silencing element into a plant.

[0125] This experiment compared the silencing efficiency of a single-step application of RNA-coated particulates and a two-step sequential application. The effects of mannitol and a surfactant were also examined.

[0126] For single-step application of dry, RNA-coated particulates, 1.5 milligrams of a blunt-ended dsRNA trigger targeting GUS (SEQ ID NO:12) or of a blunt-ended dsRNA trigger targeting GFP (SEQ ID NO:3) were dissolved in either water or 200 millimolar mannitol. 100 milligrams SiC particles (360 mesh) were added to the RNA solutions, and the mixture was air dried overnight. The dry RNA-coated particles were sprayed at 60 psi in three 1-second bursts onto the leaves of transgenic *Nicotiana benthamiana* 16c plants using a G78 airbrush mounted to a ring stand at 5 centimeters nozzle-to-leaf distance from the plants. For two-step sequential application, the dsRNA triggers were dissolved in water, with or without 0.2% Silwet L77, and with or without 200 millimolar mannitol. Twenty microliters of RNA solution was applied to each of three leaves of transgenic *Nicotiana benthamiana* 16c plants and allowed to dry briefly. Dry uncoated silicon carbide (360 mesh) particles were sprayed onto the RNA-coated leaves at 60 psi using a G78 airbrush mounted to a ring stand at 5 centimeters nozzle-to-leaf distance from the plants. GFP silencing was assessed visually using blue light excitation at 7 days after treatment. The silencing frequency of dry coated particles and sequential application was found to be approximately the same. The addition of mannitol had no effect in the single-step application of dry, RNA-coated but had a positive effect on the two-step sequential application, by apparent reduction in leaf damage.

Example 10

[0127] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene by way of particulate-assisted delivery of a nucleic acid such as an RNA “trigger” or silencing element into a plant.

[0128] This experiment compared the silencing efficiency of different particulate abrasives in a two-step sequential application method, where applying an RNA to the surface of the plant is followed by abrading the surface of a plant with a particulate of a size greater than about 2.5 micrometers.

[0129] Particulate abrasives tested included silicon carbide (SiC, angular), aluminum oxide (Al₂O₃, angular), soda lime glass (“SLG”, round) and diatomaceous silica (“diatomaceous earth”, “DE”, angular) particles with the size ranges provided in **Table 2** (see **Example 1**). Blunt-ended dsRNA triggers targeting GFP (**SEQ ID NO:3**) or magnesium chelatase (**SEQ ID NO:13**) were diluted to 5 milligrams/milliliter in 200 millimolar mannitol containing 0.05% Silwet L77. Fifteen microliters of RNA solution was hand applied using a pipette onto two leaves of transgenic *Nicotiana benthamiana* 16c plants and allowed to dry briefly. Dry uncoated particles were sprayed in three 1-second bursts onto the RNA-coated leaves at 60 psi using a G78 airbrush mounted to a ring stand at 7 centimeters nozzle-to-leaf distance from the plants. GFP silencing was assessed visually using blue light excitation at 7 days after treatment. The results comparing visual silencing efficacy for the different particulates is depicted in **Figure 2**. Under the conditions in this experiment, the greatest silencing with lowest leaf damage generally resulted from use of particles of about 10 to about 25 micrometers in size. The use of larger particles also resulted in GFP silencing but also caused heavier leaf damage. The use of smaller particles resulted in less silencing and less leaf damage. Particulate shape (angular or round) had little effect on silencing efficiency. Density appeared to be an important factor as little silencing was observed with diatomaceous silica, the least dense particle tested.

Example 11

[0130] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene by way of particulate-assisted delivery of a nucleic acid such as an RNA “trigger” or silencing element into a plant.

[0131] This experiment compared the silencing efficiency of a single-step application of RNA-coated particulates and a two-step sequential application. The GFP silencing efficacy of a 124 bp dsRNA trigger (**SEQ ID NO:3**) and of a 22 bp dsRNA trigger (**SEQ ID NO:14**) was also compared.

[0132] In the single-step method, the dsRNA trigger was diluted in water and added to 200 milligrams SiC (400 mesh), air dried overnight with gentle agitation, ground gently and sieved through 270 mesh. Thirty microliters of an aqueous solution of 0.05% Silwet L77, 200 millimolar mannitol was applied by hand to the top two expanded leaves and terminal leaf of transgenic *Nicotiana benthamiana* 16c plants; the dry dsRNA-coated particles were sprayed in

three 1-second bursts onto the Silwet L77/mannitol-coated leaves at 60 psi using a G78 airbrush mounted to a ring stand at 7 centimeters nozzle-to-leaf distance from the plants. To estimate the amount of dsRNA delivered, three 1-second bursts of the dry dsRNA-coated particles were sprayed into 100 microliters of water in a centrifuge tube which was then
5 vortexed, and the dsRNA concentration estimated by UV spectrometry.

[0133] In the two-step method, the dsRNA trigger was diluted in water. Silwet L77 and mannitol was added to the dsRNA solution to final concentrations of 0.05% and 200 millimolar, respectively. Thirty microliters of the dsRNA solution was applied by hand to the top two expanded leaves and terminal leaf of transgenic *Nicotiana benthamiana* 16c plants; the
10 treated leaves were allowed to air dry 10 minutes. SiC (400 mesh) particles were sprayed in three 1-second bursts onto the Silwet L77/mannitol-coated leaves at 60 psi using a G78 airbrush mounted to a ring stand at 7 centimeters nozzle-to-leaf distance from the plants.

[0134] GFP silencing was assessed visually using blue light excitation at 7 days after treatment. In this experiment, GFP silencing efficiency for the single-step and two-step
15 application methods appeared to be similar, and, while on a mass basis the 22 bp dsRNA trigger was more efficient than the 124 bp dsRNA trigger, the efficiency was similar when compared on a mole basis.

Example 12

[0135] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene by way of particulate-assisted delivery of a nucleic acid such as an RNA “trigger” or silencing element
20 into a plant.

[0136] This experiment compared the silencing efficiency of a single-step application of RNA-coated particulates, a two-step sequential application, a single-step high-pressure spray application of RNA without particulates, and a single-step high-pressure spray application of an RNA/particulate suspension. A commercial spray tip fitted in a track sprayer was used.
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[0137] Blunt-ended dsRNA triggers targeting GFP (SEQ ID NO:3) or magnesium chelatase (SEQ ID NO:13) were diluted to 5 milligrams/milliliter in 200 millimolar mannitol containing 0.05% Silwet L77. For the RNA/particulate suspensions, diatomaceous silica (Celite 512) or SiC (360 mesh) was added to the above RNA solutions at 20
30 milligrams/milliliter. The RNA preparations were sprayed onto transgenic *Nicotiana*

benthamiana 16c plants at either 60 or 85 psi using a canister sprayer fitted with a TeeJet 40005E flat fan nozzle positioned 7 centimeters from the plants. The plants sprayed at 60 psi were sprayed a second time with dry uncoated particles applied at 80 psi with a canister sprayer fitted with a TeeJet DG110015 nozzle 10 centimeters from the plants. GFP silencing was assessed visually using blue light excitation at 7 days after treatment. Silencing efficiency was very low in the plants sprayed only with RNA solutions (no particulates). Silencing using either the RNA/Celite or RNA/SiC suspensions was observed for both GFP and magnesium chelatase; for GFP the silencing efficacy was less than that resulting from a two-step sequential application, but for magnesium chelatase the silencing efficacy was comparable. These results indicate that a single-step application of an RNA/particulate suspension is efficacious and can be advantageously used with commercial spraying equipment.

Example 13

[0138] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene by way of particulate-assisted delivery of an RNA “trigger” into maize plants.

[0139] This experiment demonstrates silencing of a GFP transgene in maize (*Zea mays*). A 121 bp dsRNA targeting GFP (**SEQ ID NO:15**) was diluted to 5 milligrams/milliliter in water containing 0.05% Silwet L77. Thirty microliters of the RNA solution was applied to a single corn (*Zea mays*) leaf and allowed to dry briefly. Dry uncoated silicon carbide particles (280, 320, 360, and 400 mesh) were sprayed at 60 psi on the dsRNA-coated leaves using a G78 airbrush mounted to a ring stand 5 centimeters from the plants. GFP silencing was assessed visually using blue light excitation at 7 days after treatment. GFP silencing was observed in plants sprayed with 280, 320, and 360 mesh SiC. The silenced sectors manifested as a long stripe (in one plant treated with 360 mesh SiC) or multiple small spots (in two plants treated respectively with 280 and 320 mesh SiC). Silenced and non-silenced sectors were sampled in the leaves and GFP expression was measured. GFP expression was reduced by about 30 to about 50 percent in silenced sectors compared to non-silenced sectors was observed in both silenced sector types (stripe and spots).

Example 14

[0140] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene by various

methods including contacting a plant surface with a matrix supporting an abrasive. In these experiments, cotton swabs supporting a particulate abrasive, uncoated or coated with dsRNA trigger, are used to abrade a plant surface and deliver a dsRNA trigger to the plant.

[0141] In a first experiment, dry dsRNA-coated particles were prepared by mixing 100 milligrams of silicon carbide (360 mesh) particles per 1 milliliter of formulations containing 1.5 milligrams/milliliter 78 bp dsRNA against GFP (**SEQ ID NO:2**) in: a) water, b) 4 millimolar MES buffer, c) 200 millimolar mannitol, or, d) 4 millimolar MES buffer and 200 millimolar mannitol. The dsRNA-SiC mixtures were air dried overnight on a rotational shaker. A cotton swab was loaded with the dry, dsRNA-coated SiC particles by pressing the swab into the prepared SiC particles, and then used to gently abrade the upper leaf surface of approximately 4-week old transgenic *Nicotiana benthamiana* 16c plants by gently rolling the swab along the leaf surface with the leaf supported from below by the worker's finger. GFP silencing was assessed visually using blue light excitation at 7 days after treatment. In this experiment, addition of 200 millimolar mannitol to the dsRNA formulation prevented leaf dehydration after abrasion using cotton-swab rolling with dsRNA coated SiC particles. Addition of 4 millimolar MES and 200 millimolar mannitol to the dsRNA formulation enhanced frequency of GFP silencing foci in the treated leaves.

[0142] In a second experiment, dry dsRNA-coated SiC particles were manufactured by prepared by mixing 100 milligrams of silicon carbide (360 mesh) particles per 1 milliliter of an aqueous dsRNA solution at the following trigger concentrations: a) 1.5 milligrams/milliliter of a 78 bp dsRNA trigger against GFP (**SEQ ID NO:2**), b) 1.5 milligrams/milliliter of a 76 bp dsRNA trigger against the *N. benthamiana* 16C magnesium chelatase (**SEQ ID NO:16**), and c) a mix of both triggers at 0.75 milligrams/milliliter each. The dsRNA-SiC mixtures were air dried overnight on a rotational shaker. A cotton swab was loaded with the dry, dsRNA-coated SiC particles by pressing the swab into the prepared SiC particles, and then used to gently abrade the upper leaf surface of approximately 4-week old transgenic *Nicotiana benthamiana* 16c plants by gently rolling the swab along the leaf surface with the leaf supported from below by the worker's finger. The same dry, dsRNA-coated SiC particle preparations were delivered to a second set of plants using an airbrush. Silencing was assessed visually using ambient light or blue light excitation at 7 days after treatment. In this experiment, GFP and magnesium chelatase silencing foci were observed in treated leaves with all particle coating protocols and delivery methods. The expected gene-target-specific phenotypes were observed in plants treated with a single dsRNA trigger, and phenotype co-localization was observed in plants treated with both dsRNA triggers.

[0143] In a third experiment, efficacy of three different two-step sequential delivery methods using the cotton-swab rolling technique was tested in *N. benthamiana* 16C seedlings. In these methods, the dsRNA trigger is applied to the plant surface prior to abrasion of the plant surface with uncoated particulates supported on a cotton swab.

5 [0144] The two-step sequential delivery methods tested were:

(a) Method 1: the dsRNA formulation was pipetted onto the leaf surface and spread with a pipette tip to ensure uniform coverage, followed by abrasion by rolling a cotton-swab carrying uncoated SiC particles;

(b) Method 2: leaves were abraded by rolling a cotton-swab carrying uncoated SiC particles, followed by pipette delivery and spreading of the dsRNA formulation; and
10

(c) Method 3: the cotton swab was first submerged in the dsRNA formulation, and then rolled over uncoated SiC particles, and finally gently rolled on the leaf surface.

[0145] Three liquid formulations of a 78 bp dsRNA trigger against GFP (**SEQ ID NO:2**) were tested: 2 milligrams/milliliter dsRNA in water; 2 milligrams/milliliter dsRNA in 200 millimolar mannitol and 20 millimolar MES; and 0.0125 milligrams/milliliter dsRNA in a Lipofectamine® formulation. For each treatment, a total of 20 microliters dsRNA formulation was applied per treated leaf of approximately 4-week old transgenic *Nicotiana benthamiana* 16c plants (three plants per treatment). Silencing was assessed visually using blue light excitation at 4 and 7 days after treatment. In this experiment, all three delivery methods and all
20 dsRNA formulations produced GFP silencing foci in treated leaves. Plants treated by Method 1 maintained normal leaf growth and displayed a higher frequency of GFP silencing foci per treated leaf. The frequency of GFP silencing foci was significantly greater in plants treated with a dsRNA concentration of 2 milligrams/milliliter, compared to plants treated with dsRNA of 0.0125 milligrams/milliliter in the Lipofectamine® formulation. Addition of 200 millimolar
25 mannitol and 20 millimolar MES increased frequency of GFP silencing foci across delivery treatment types.

Example 15

[0146] This example illustrates non-limiting embodiments of methods, apparatuses,
30 and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene by various methods including contacting a plant surface with a matrix supporting an abrasive. In these examples, sandpaper serves as a matrix supporting a particulate abrasive, and is used to abrade a plant surface and deliver a dsRNA trigger to the plant.

[0147] Sandpapers for wet sanding were used to deliver a 78 bp dsRNA trigger against GFP (SEQ ID NO:2) into approximately 3-week old transgenic *Nicotiana benthamiana* 16c plants. Three different grit sizes were used: P180, P600, and P2500, which have an average particle size diameter of 82, 25.8, and 8.4 micrometers, respectively. The sandpaper was supported by a 3/4-inch diameter PVC tube to facilitate gentle rolling on the surface of the treated leaves. Formulations of the dsRNA at a final concentration of 2 milligrams/milliliter were prepared in water or in aqueous 0.05% Silwet L77. Ten or 20 microliters of dsRNA formulation were pipetted onto the surface of two leaves per plant, and spread with a pipette tip to ensure uniform coverage, followed by abrasion by gently rolling the sandpaper over the treated leaf surface. For comparison, additional plants were treated only with the dsRNA formulation (no abrasion), or with the dsRNA formulation followed by abrasion with a cotton swab supporting SiC particles (360 mesh). Silencing was assessed visually using blue light excitation at 4 and 7 days after treatment.

[0148] The results are summarized as follows. No signs of leaf damage or turgor loss was observed in treated *Nicotiana benthamiana* leaves. Treated plants showed no signs of wilting or severe leaf damage immediately after treatment or 1 day after treatment. The observed frequency of GFP silencing foci depended on sandpaper grit size; plants abraded with the 600 sandpaper roller had greater frequency of GFP silencing foci than plants abraded with other sandpaper grit sizes with the cotton swab supporting uncoated SiC particles. In a two-step sequential application (dsRNA applied first, followed by abrasion), abrasion with sandpaper was found to be more efficient in inducing GFP silencing foci than abrasion with a cotton swab supporting uncoated SiC particles, independently of the dsRNA formulation or timing of abrasive treatment.

[0149] Results from these and similar experiments provided further inferences. Silencing activity was observed to be retained in plants where the dsRNA-treated leaf was left for a day prior to abrasion; a stronger phenotype and more frequent GFP silencing foci were observed when the dsRNA formulation was left to dry on the surface of the leaf for at least 20 minutes prior to abrasion. Experiments with a “flat” roller, which gave reduced silencing efficacy, suggested that leaf surface abrasion and not pressure alone is the mechanism for dsRNA delivery. Sequential abrasive methods have shown consistently high efficacy levels and success rate. Systemic GFP silencing was observed in sandpaper-abraded *N. benthamiana* 16C plants grown under different conditions and in different locations, approximately 10-13 days after treatment, independent of the dsRNA trigger size used. Efficacy of mechanical

abrasion methods was also demonstrated against endogenous gene targets including magnesium chelatase, PAT1, and PDS.

[0150] Similar experiments demonstrating localized target gene silencing induced by particle-assisted nucleic acid delivery were carried out in *Arabidopsis thaliana*. The sandpaper abrasion method was modified for developing *Arabidopsis thaliana* leaves from small plants grown in 24-well blocks. Round-tip tweezers were modified by wrapping one end with a paper pad and laboratory film (Parafilm M® Bemis NA, Neenah, WI) (to support the leaf and prevent leaf damage), and attaching sandpaper to the other end with double-sided sticky tape. Similarly, methods using a cotton-swab rolling technique for abrasion can also be used on *Arabidopsis thaliana* seedlings.

[0151] Similar experiments were also carried out in a transgenic tomato line expressing GFP. GFP and magnesium chelatase silencing foci were observed in tomato seedlings treated with a two-step sequential method including dsRNA application followed by sandpaper abrasion. The frequency of putative GFP silencing foci was low (1-2 foci per treated leaves) but was present in 6 to 7 of 10 treated tomato seedlings. Magnesium chelatase silencing foci was observed with low frequency in treated tomato seedlings, tomato seedlings treated with mixed dsRNA triggers displayed the expected co-localized GFP and magnesium chelatase silencing foci.

Example 16

[0152] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene by applying a relatively small (22 base-pair) dsRNA trigger to a plant surface, followed by abrasion with a matrix supporting particulate abrasives.

[0153] This example describes systemic silencing of GFP in transgenic *Nicotiana benthamiana* 16c plants by a 22 bp dsRNA trigger (SEQ ID NO:14) in combination with sandpaper abrasion. Non-specific dsRNA was used as a control in the experiment. The dsRNA was dissolved in water to 1 milligram/milliliter final concentration and a total of 20 microliters dsRNA was applied to 2 young leaves on individual transgenic *Nicotiana benthamiana* 16c plants. The treated leaves were abraded with a 600 sandpaper roller. Samples for Northern blot analysis of GFP mRNA levels were collected at 24 and 48 hours after treatment. Silencing was assessed visually using blue light excitation at 2, 5, 8, and 13 days after treatment. A reduction of GFP mRNA expression in T52623 dsRNA-treated plants was

observed at 1 day after treatment, and strong GFP expression reduction observed at 2 days after treatment. Localized GFP silencing was observed on treated leaves at 2 days after treatment, and the localized silencing phenotype became much clearer and stronger from 5 days after treatment onward. Systemic GFP silencing was observed on untreated young leaves at 10 to 13 days after treatment.

[0154] In a similar experiment, 22 bp dsRNA trigger (SEQ ID NO:17) targeting an endogenous gene, magnesium chelatase. The dsRNA was dissolved in water to 1 milligram/milliliter final concentration and a total of 20 microliters dsRNA was applied to 2 young leaves on individual transgenic *Nicotiana benthamiana* 16c plants. The treated leaves were abraded with a 600 sandpaper roller. Silencing was assessed visually under visible light at 2, 5, 8, and 13 days after treatment. Localized silencing was observed as the expected chlorophyll-deficient phenotype in leaves treated with SEQ ID NO:17.

Example 17

[0155] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene by treatment with a dsRNA targeting the promoter region of the target gene, followed by abrasion with a matrix supporting particulate abrasives.

[0156] The region upstream of the transgenic GFP insert from *Nicotiana benthamiana* 16c was cloned and sequenced. The size of the sequenced region is 2278 bp and contains an 835 bp region encoding the cauliflower mosaic virus (CaMV) 35S promoter. An upstream expression cassette containing a *nos* terminator is located 698 bp from the 5' end of the CaMV 35S promoter. Three dsRNA triggers, ranging in size from 122-127 bp, were designed to match the DNA sequence from 3' end of the CaMV 35S promoter region: CaMV.35S-1 (SEQ ID NO:18), CaMV.35S-2 (SEQ ID NO:19), CaMV.35S-3 (SEQ ID NO:20), and (as a control) a 124 bp dsRNA targeting the coding region of GFP (T41817, SEQ ID NO:3). The dsRNA was dissolved in water to 4 milligram/milliliter final concentration and a total of 10 – 20 microliters dsRNA was applied to leaves 3 and 4 from 2 week-old plants transgenic *Nicotiana benthamiana* 16c plants. After the RNA was aliquoted on the leaves, a pipette tip was used to evenly spread the RNA over the surface of the adaxial side of each leaf. The RNA solution was allowed to dry for 30 minutes and then the top of the leaf was abraded once with P600 sandpaper glued to a dowel that was rolled over the leaf. The plants were then placed in a growth chamber set for 263 micromoles of light set for 14 hour/10 hour (light/dark cycle)

with a temperature setting of 23 degrees Celsius /18 degrees Celsius (day/night). Silencing was assessed visually using blue light excitation at 7 days after treatment. The first 2 triggers closest to the end of the promoter, CaMV.35S-1 (SEQ ID NO:18) and CaMV.35S-2 (SEQ ID NO:19), produced a strong silencing phenotype with many small silencing foci on the treated leaves. CaMV.35S-3 (SEQ ID NO:20) produced the weakest phenotype with only slight levels of silencing in only a few areas. The control dsRNA targeting the coding region of GFP (SEQ ID NO:3) gave the strongest phenotype with many large silencing spots that merge to cover most of the treated leaves.

Example 18

[0157] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene by treatment with a nucleic acid, followed by abrasion with a particulate that disrupts cells in the cuticle or epidermis or both cuticle and epidermis of the plant.

[0158] Double-stranded RNA was fluorescently labelled with Cy3 and coated onto SiC particles (320 mesh) or soda lime glass beads of three size ranges (10 – 22, 22 – 27, and 35 – 45 micrometer). Control particles were made in the same way but without Cy3 labelling. The dry dsRNA-coated SiC or glass beads were sprayed onto leaves and central axis of 3-week old *Nicotiana benthamiana* 16c plants at 65 psi using a G78 airbrush mounted to a ring stand at 5 – 7 centimeters nozzle-to-leaf distance from the plants. Equipment was cleaned with ethanol between treatments to minimize cross-contamination.

[0159] For live imaging studies regions of interest (silenced spots identified as red areas under UV light) were removed with 4 – 5 millimeter biopsy punches and the live tissues were imaged with confocal fluorescence microscopy. In addition, tissue samples were fixed with paraformaldehyde, cryoprotected with sucrose, mounted in OCT medium, and cryosectioned for epifluorescent and bright-field imaging. These microscopic studies demonstrated that the sprayed particles primarily impacted epidermal cells.

[0160] Similar microscopic studies were performed on tomato leaves treated with a two-step sequential method including dsRNA application followed by abrasion with sandpapers of different grit sizes. The results demonstrated that silencing efficiency increased in the grit size order P200 < P400 < P2000 (*i. e.*, from coarser to finer grits), indicating that the most efficacious sandpapers have grit sizes that can disrupt the leaf cuticle and only compromise or partially compromise the epidermal cell layer but do not cause deeper damage.

Example 19

[0161] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant.

5 More specifically, this example describes systemic silencing of a target gene by treatment with a dsRNA, followed by abrasion with a matrix supporting particulate abrasives.

[0162] This experiment compared the silencing efficiency of sandpapers of different grit sizes in a two-step sequential application. The effects of nuclease inhibitors were also examined.

10 [0163] Three dsRNA formulations were prepared. The base formulation contained 124 bp dsRNA trigger (**SEQ ID NO:3**) at 2 milligram/milliliter, 200 millimolar mannose, 4 millimolar MES buffer final concentration in water. A second formulation was identical to the base formulation but included 4.8 millimolar Zn₂SO₄ as an RNase inhibitor. A third formulation was identical to the base formulation but included 3.7% RNasin® Ribonuclease
15 Inhibitor (Promega Corporation, Madison, WI) as an RNase inhibitor. A total of 10 or 20 microliters dsRNA was applied to two leaves of 3-week old plants transgenic *Nicotiana benthamiana* 16c plants. After the RNA was aliquoted on the leaves, a pipette tip was used to evenly spread the RNA over the surface of the adaxial side of each leaf. The RNA solution was allowed to dry for 30 minutes and then the top of the leaf was abraded once with
20 sandpaper of two different grit sizes (P180 or P600) attached to a ¾-inch PVC tube that was rolled over the leaf. Silencing was assessed visually using blue light excitation at 7 days after treatment. Results are provided in **Table 7**.

Table 7

Sandpaper grit	RNase inhibitor	Average number of GFP silencing loci per leaf	Standard error
P600	None	50	15
P600	Zn ₂ SO ₄	78	13
P600	RNasin ®	66	12
P180	None	4	2
P180	Zn ₂ SO ₄	9	3
P180	RNasin ®	6	2

25 [0164] These results show that across all formulations, P600 abraded leaves had ~10X more GFP silencing foci per leaf than those abraded with a coarser P180 sandpaper. Independently of the sandpaper grit used, formulations including an RNase inhibitor had more GFP silencing foci per leaf. The effect of nuclease inhibitor on increasing number of GFP

silencing foci per leaf was relatively stronger for the coarser P180 sandpaper abraded leaves than for the P600 abraded leaves. At the concentrations used, Zn_2SO_4 had the strongest effect on increasing the number of GFP silencing foci per leaf.

5 Example 20

[0165] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene by way of particulate-assisted delivery of a nucleic acid such as DNA viral vector into a plant. This
10 example demonstrates the effect of directly applied dsRNA triggers in preventing systemic infection of TSWV.

[0166] An experiment was conducted to assess the capacity of dsRNA triggers applied without bacterial lysate to prevent infection with tomato spotted wilt virus (TSWV) in *Nicotiana benthamiana*. GFP silencing served as a tracer for trigger delivery and processing.
15 Two 298 bp chimeric dsRNA triggers were produced; the first trigger TSWV-GFP-TSWV (SEQ ID NO:21) included two dsRNA regions targeting GFP flanking a dsRNA region targeting the TSWV N-gene, and the second trigger GFP-TSWV-GFP (SEQ ID NO:22) included two dsRNA regions targeting the TSWV N-gene flanking a dsRNA region targeting GFP. The blunt-ended 141 bp dsRNA trigger (SEQ ID NO:23) targeting GFP was used as a
20 control.

[0167] The chimeric and control dsRNA triggers were applied directly to *N. benthamiana* 16c plants showing 3 true leaves (approximately 26 days after germination), followed by abrasion with 600 grit sandpaper. Local silencing of GFP was observed on the treated leaves in all treatments 4 days after treatment; at this time, TSWV was rub-inoculated
25 onto the leaves showing local GFP silencing. Fourteen days after TSWV challenge, plants were assessed for development of TSWV symptoms. All plants treated with the GFP trigger alone were strongly symptomatic for TSWV. Less than 20% of plants treated with the chimeric GFP/TSWV dsRNA triggers were obviously infected with TSWV. Similar results occurred in a similar experiment where plants were inoculated with TSWV 7 days after
30 treatment, demonstrating that direct application of the chimeric dsRNA triggers protected plants from TSWV infection for at least 7 days after treatment.

Example 21

[0168] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant. More specifically, this example describes systemic silencing of a target gene using a dsRNA trigger targeting a non-coding regulatory region of the gene to be silenced, and heritability of the phenotype in a progeny plant.

[0169] The sequence of the promoter region of the chalcone synthase chs (A) gene in *Petunia hybrida* was published by Van der Meer *et al.* (1990) *Plant Mol. Biol.*, 15:95 – 190. A 258 bp blunt-ended dsRNA trigger (anti-sense strand having **SEQ ID NO:24**) targeting the upstream promoter region was produced. The dsRNA trigger is applied to *Petunia hybrida* leaves with abrasion, using any of the single-step or two-step methods described in the preceding working Examples. The treated leaves are regenerated into R0 plants. R0 plants displaying the expected phenotype of white flowers are selected. The white flower phenotype is heritable by an epigenetic effect in plants of the subsequent generation.

[0170] Additional experiments were carried out to test for silencing in plants regenerated directly from treated tissue. *Nicotiana benthamiana* plants (line 16c) transgenically expressing green fluorescent protein (GFP) driven by the 35S promoter were topically treated using any of the single-step or two-step methods described in the preceding working Examples; a 127 bp blunt-ended dsRNA trigger with the anti-sense (*i. e.*, anti-sense relative to the direction of the GFP gene's coding sequence) strand having **SEQ ID NO:29** targeting the 35S promoter was applied with abrasion to either the second or the third true leaves on 3-week-old plants. Both local silencing and systemic silencing of the target gene GFP were observed in the treated 16c plants. Samples of leaf tissue displaying local silencing were taken from the topically treated leaves and cultured on solid media (using a protocol similar to that described in Horsch *et al.* (1984) *Science*, 223:496-498). Sixteen R0 16c plants were regenerated; fifteen of the sixteen R0 plants exhibited a homogenous phenotype with complete silencing of the target gene GFP throughout the entire R0 life cycle.

[0171] In additional experiments, *Nicotiana benthamiana* plants (line 16c) transgenically expressing green fluorescent protein (GFP) driven by the 35S promoter are topically treated using any of the single-step or two-step methods described in the preceding working Examples; a 311 bp blunt-ended dsRNA trigger with the anti-sense (*i. e.*, anti-sense relative to the direction of the GFP gene's coding sequence) strand having **SEQ ID NO:30** targeting the 35S promoter is applied with abrasion to two leaves of each plant. Both local silencing and systemic silencing of the target gene GFP are observed in the treated 16c plants.

Samples of leaf tissue from the topically treated leaves displaying local silencing are taken for culture on solid media and R0 plants regenerated. The R0 plants include 16c plants that exhibited a homogenous phenotype with complete silencing of the target gene GFP. R0 plants displaying the expected phenotype of GFP silencing are selected. The silenced GFP phenotype is heritable by an epigenetic effect in plants of the subsequent generation.

Example 22

[0172] This example illustrates non-limiting embodiments of methods, apparatuses, and compositions useful for delivering a nucleic acid into a plant or cells or tissues of a plant.

More specifically, this example describes use of a method including application of nucleic acids to the surface of a plant, followed by abrasion with a particulate, whereby the nucleic acids are delivered to the plant and result in *in vivo* editing or sequence replacement of a gene in the plant.

[0173] Methods for *in vivo* editing or sequence replacement of a gene are known in the art, for example through the use of zinc-finger nucleases, CRISPR, and Cas9. See, for example, Townsend *et al.* (2009) *Nature*, 459:442 – 446; Qi *et al.* (2012) *Nature Biotechnol.*, 30:1002 – 1007; Cong *et al.* (2013) *Science*, 339:819 – 823; and Hsu *et al.* (2013) *Nature Biotechnol.*, 31:827 – 834. In this example, nucleic acids for *in vivo* editing are used with methods similar to those described herein in the preceding Examples to modify the sequences of an endogenous gene in a plant.

[0174] Specific amino acid point mutations of the endogenous acetolactate synthase genes (ALS *SuRA* and *SuRB*) in tobacco (*Nicotiana tabacum*), which share highly conserved coding regions, result in resistance to certain herbicides. Three such amino acid point mutations are P191A (conferring resistance to chlorsulphuron), W568L (conferring resistance to both chlorsulphuron and imazaquin), and S647T (conferring resistance to imazaquin), for which the corresponding nucleotide mutations have been reported (depicted in Figure 1b of Townsend *et al.* (2009) *Nature*, 459:442 – 446).

[0175] Three nucleic acids are prepared: (1) a CAS9 expression DNA plasmid; (2) a synthetic ssRNA containing a fused target sequence/guide RNA, wherein the target RNA includes about 20 nucleotides of the selected region to be edited *in vivo*, fused to a guide RNA having the sequence **SEQ ID NO:25**; and (3) a donor DNA (provided as either a plasmid or as a dsDNA fragment) including a replacement sequence selected from P191A (**SEQ ID NO:26**), W568L (**SEQ ID NO:27**), and S647T (**SEQ ID NO:28**), plus additional 5' and 3' flanking sequence as needed. The three nucleic acids are applied to *Nicotiana tabacum* leaves with

abrasion, using any of the single-step or two-step methods described in the preceding working Examples. Herbicide-resistant R0 tobacco plants are regenerated from treated leaves on selective media containing the appropriate herbicide.

[0176] A separate series of experiments employed a commercially available CRISPR-Cas9 genome editing system (“Alt-R™ CRISPR-Cas9 System”, Integrated DNA Technologies (IDT, 1710 Commercial Park, Coralville, IA 52241, USA). Nucleic acids were obtained from Integrated DNA Technologies (IDT, 1710 Commercial Park, Coralville, IA 52241, USA): (1) a crRNA having the sequence of **SEQ ID NO:31**, designed to target a region near the 5’ end of the GFP mRNA; and (2) a “universal tracrRNA oligonucleotide” with the primary nucleotide sequence of **SEQ ID NO:32**, modified to improve RNase resistance using an IDT proprietary modification. The two nucleic acids are applied with abrasion, together with a source of Cas9 or a modified Cas9, to leaves of hemizygous *Nicotiana benthamiana* plants (line 16c) transgenically expressing green fluorescent protein (GFP) driven by the 35S promoter, using any of the single-step or two-step methods described in the preceding working Examples. The Cas9 or modified Cas9 is provided in a convenient form, *e. g.*, as a CAS9 expression DNA plasmid or as a commercially available Cas9 nuclease (*e. g.*, purified, codon-optimized, recombinant *S. pyogenes* Cas9 nuclease catalogue number 1074181, Integrated DNA Technologies (IDT, 1710 Commercial Park, Coralville, IA 52241, USA). The nucleic acids and Cas9 source are applied with abrasion to either the second or the third true leaves on 3-week-old plants. Both local silencing and systemic silencing of the target gene GFP are observed in the treated 16c plants. Samples of leaf tissue from the topically treated leaves displaying local silencing are taken for culture on solid media and R0 plants regenerated. The R0 plants include 16c plants that exhibited a homogenous phenotype with complete silencing of the target gene GFP.

[0177] All of the materials and methods disclosed and claimed herein can be made and used without undue experimentation as instructed by the above disclosure. Although the methods and compositions as described herein have been described in terms of preferred embodiments and illustrative examples, it will be apparent to those of skill in the art that variations can be applied to the materials and methods described herein without departing from the concept, spirit and scope of this disclosure. All such similar substitutes and modifications apparent to those skilled in the art are deemed to be within the spirit, scope and concept of this disclosure as defined by the appended claims.

What is claimed is:

1. A method for silencing a target gene in a plant, comprising, in any order, the steps of:
 - (a) abrading a surface of a plant with a particulate of a size greater than about 2.5 micrometers;
 - 5 (b) applying an RNA to the surface of the plant, wherein the RNA comprises at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant;
 whereby the target gene is silenced.
2. The method of claim 1, wherein (a) the target gene is an endogenous gene of the plant or a transgene expressed in the plant, (b) the target gene in the plant is a gene of a parasitic plant, fungus, or bacterium that is a pathogen or parasite of the plant, (c) the target gene is multiple target genes, or (d) the target gene is multiple target genes from more than one species.
3. The method of claim 1, wherein (a) the RNA is single-stranded RNA, (b) the RNA is double-stranded RNA, (c) the RNA is a miRNA precursor or a small RNA, (d) the RNA is non-transcribable, (e) the RNA is not self-replicating, (f) the RNA comprises 19 – 500 nucleotides or base pairs, or (g) the RNA is not contained in a viral vector nor encoded by a plasmid.
4. The method of claim 1, wherein (a) the particulate is selected from the group consisting of a mineral abrasive, a metal abrasive, a synthetic abrasive, and an organic abrasive, (b) the particulate is selected from the group consisting of aluminum oxide, silicon carbide, silicon dioxide, soda lime glass, diatomaceous silica (diatomaceous earth), flint, quartz, garnet, silicon dioxide, pumice, sand, feldspar, calcite, steel, tungsten, ceramic, boron carbide, tungsten carbide, (c) the particulate is of an average size range from about 2.5 micrometers to about 50 micrometers, or (d) the particulate is supported by, attached to, or embedded in a matrix.
5. The method of claim 1, wherein the particulate, the RNA, or both, are further applied with
 - (a) at least one selected from the group consisting of water, a biocide, a chelator, a buffer, a nonionic surfactant, a zwitterionic surfactant, a defoamer, a rainfastness agent, and a photoprotectant, (b) at least one selected from the group consisting of a carrier agent, a surfactant, an osmolyte, an organosilicone, a polynucleotide herbicidal molecule, a non-polynucleotide herbicidal molecule, a non-polynucleotide pesticide, a safener, an insect attractant, and an insect growth regulator, or (c) at least one pesticidal agent selected from the group consisting of a patatin, a plant lectin, a phytoecdysteroid, a *Bacillus thuringiensis*

insecticidal protein, a *Xenorhabdus* insecticidal protein, a *Photorhabdus* insecticidal protein, a *Bacillus laterosporous* insecticidal protein, and a *Bacillus sphaericus* insecticidal protein.

6. A method for introducing a nucleic acid into a plant, comprising:

5 (a) in any order, the steps of (i) applying a composition comprising a nucleic acid to a surface of a plant, and (ii) contacting a matrix supporting an abrasive with the surface; or,

(b) in any order, the steps of (i) abrading a surface of a plant with a particulate of a size greater than about 2.5 micrometers, and (ii) applying a composition comprising a
10 nucleic acid to the surface of the plant, wherein the nucleic acid comprises at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant;

whereby the nucleic acid is introduced into the plant.

7. The method of claim 6, wherein (a) the nucleic acid is DNA, (b) the nucleic acid is DNA in
15 the form of a plasmid or viral vector, (c) the nucleic acid is single-stranded RNA, (d) the nucleic acid is double -stranded RNA, (e) the nucleic acid is a combination of DNA and RNA, (f) the nucleic acid is a miRNA precursor or a small RNA, (g) the nucleic acid is a non-transcribable RNA, (h) the nucleic acid is a non-self-replicating DNA or RNA, (i) the nucleic acid is a DNA that comprises 19 – 500 nucleotides or base pairs, (j) the nucleic acid
20 is an RNA that comprises 19 – 500 nucleotides or base pairs, (k) the nucleic acid is an RNA that is not contained in a viral vector nor encoded by a plasmid, (l) the nucleic acid is provided in a microbial cell or as a microbial fermentation product, or (m) the nucleic acid comprises at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant.

25 8. The method of claim 6, wherein (a) the target gene is an endogenous gene of the plant or a transgene expressed in the plant, (b) the target gene in the plant is a gene of a parasitic plant, fungus, or bacterium that is a pathogen or parasite of the plant, (c) the target gene is multiple target genes, or (d) the target gene is multiple target genes from more than one species.

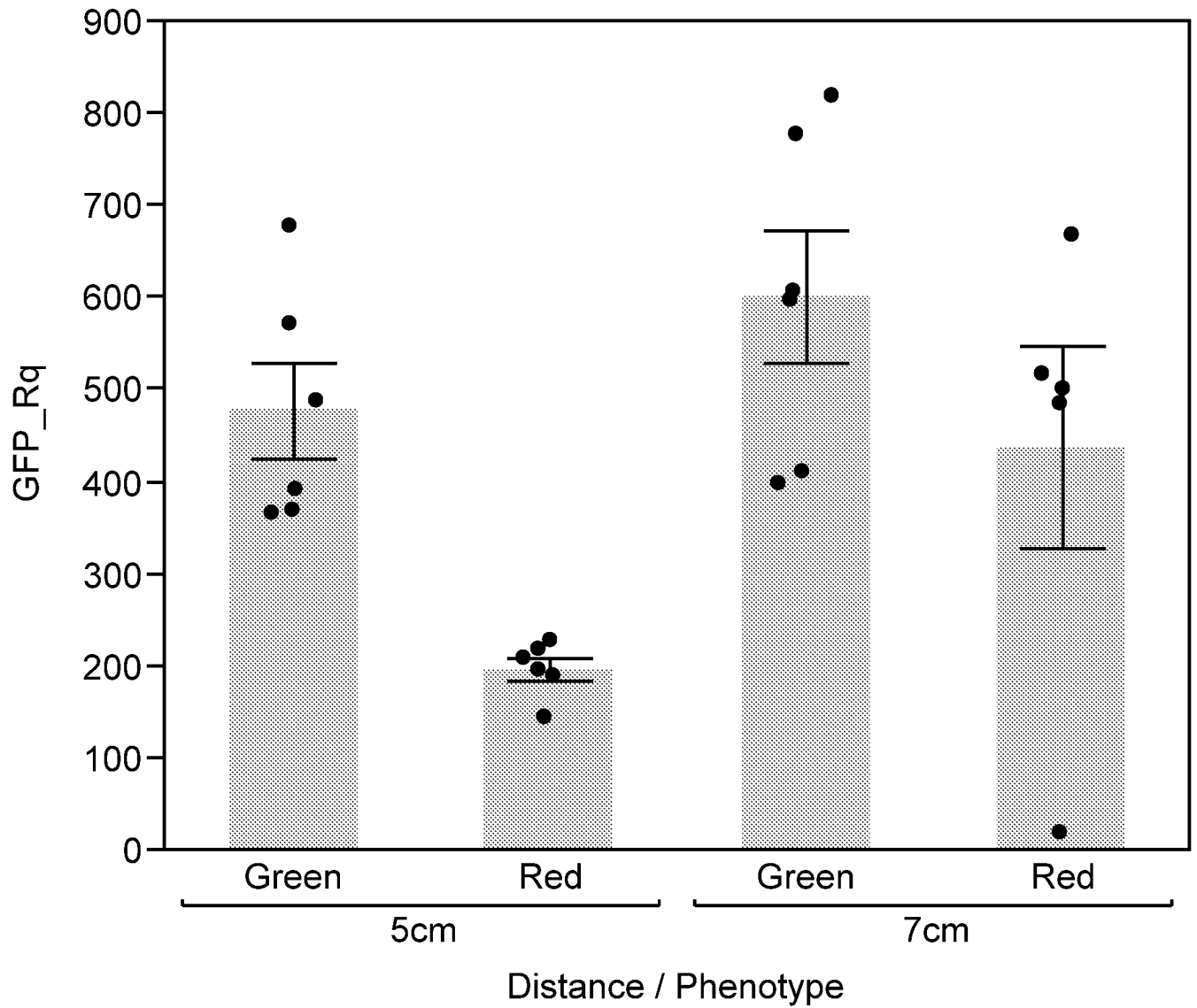
30 9. The method of claim 6, wherein (a) the abrasive or the particulate is selected from the group consisting of a mineral abrasive, a metal abrasive, a synthetic abrasive, and an organic abrasive, (b) the abrasive or the particulate is selected from the group consisting of aluminum oxide, silicon carbide, silicon dioxide, soda lime glass, diatomaceous silica (diatomaceous earth), flint, quartz, garnet, silicon dioxide, pumice, sand, feldspar, calcite,

steel, tungsten, ceramic, boron carbide, tungsten carbide, (c) the abrasive or the particulate is of an average size range from about 2.5 micrometers to about 50 micrometers, or (d) the abrasive or the particulate is supported by, attached to, or embedded in a matrix.

10. The method of claim 6, wherein the abrasive or the particulate, or the composition comprising a nucleic acid, or both, are further applied with (a) at least one selected from the group consisting of water, a biocide, a chelator, a buffer, a nonionic surfactant, a zwitterionic surfactant, a defoamer, a rainfastness agent, and a photoprotectant, (b) at least one selected from the group consisting of a carrier agent, a surfactant, an osmolyte, an organosilicone, a polynucleotide herbicidal molecule, a non-polynucleotide herbicidal molecule, a non-polynucleotide pesticide, a safener, an insect attractant, and an insect growth regulator, or (c) at least one pesticidal agent selected from the group consisting of a patatin, a plant lectin, a phytoecdysteroid, a *Bacillus thuringiensis* insecticidal protein, a *Xenorhabdus* insecticidal protein, a *Photorhabdus* insecticidal protein, a *Bacillus laterosporous* insecticidal protein, and a *Bacillus sphaericus* insecticidal protein.
11. A method for providing a plant exhibiting modification of a target gene in the plant, comprising directly regenerating a plant from a source plant, wherein the source plant has been contacted with a composition comprising an RNA that comprises at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant, wherein the directly regenerated plant exhibits modification of the target gene.
12. The method of claim 11, wherein the source plant has been topically contacted with the RNA.
13. The method of claim 11, wherein the source plant is an intact, growing plant.
14. The method of claim 11, wherein the modification of the target gene is (1) non-heritable silencing of the target gene, (2) heritable or epigenetic silencing of the target gene, or (3) a change in the nucleotide sequence of the target gene.
15. The method of claim 11, wherein (a) the directly regenerated plant exhibits partial silencing of the target gene, or (b) the directly regenerated plant exhibits complete silencing of the target gene, or (c) the directly regenerated plant exhibits a homogeneous morphological or physiological phenotype resulting from modification of the target gene.
16. The method of claim 11, wherein the modification of the target gene is heritable to plants of subsequent generations grown from the directly regenerated plant.
17. The directly regenerated plant exhibiting modification of the target gene, provided by the method of claim 11.

18. The plants of subsequent generations exhibiting modification of the target gene, provided by the method of claim 16.

1/2

**FIG. 1**

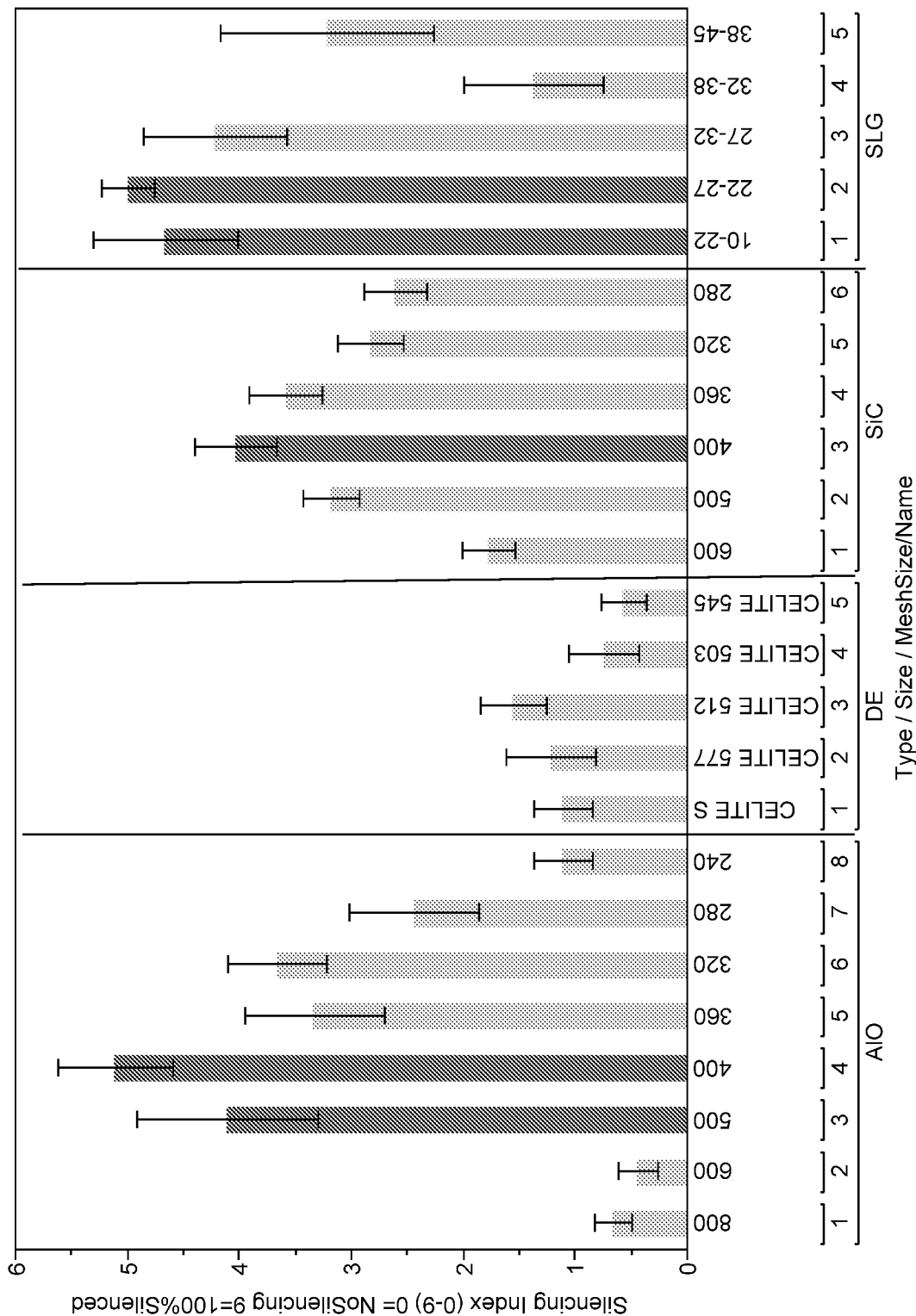


FIG. 2

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US16/35500

A. CLASSIFICATION OF SUBJECT MATTER

IPC(8) - A01H 1/00, 1/06, 5/00; C12N 5/04 (2016.01)

CPC - A01H 1/00, 1/06, 5/00; C12N 5/04

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC(8) Classifications: A01H 1/00, 1/06, 5/00; C12N 5/04, 15/00, 15/82, 15/09 (2016.01)

CPC Classifications: A01H 1/00, 1/06, 5/00; C12N 5/04, 15/00, 15/82, 15/09

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

PatSeer (US, EP, WO, JP, DE, GB, CN, FR, KR, ES, AU, IN, CA, INPADOC Data); Google; Google Scholar; PubMed; EBSCO; Search Terms Used: silenc*, inhibit*, suppress*, introduc*, gene*, plant*, DNA, RNA, 'nucleic acid', abrad*, abrad*, scrap*, wear*, surfac*, particul*, nucleotid*, endogen*, transgen*, grit*, 'Monsanto Technology', 'Shihshieh Huang', 'Alberto landolino', 'Gregory Peel'

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 2011/0296556 A1 (SAMMONS, RD et al.) 1 December 2011; paragraphs [0004], [0006], [0050]-[0052], [0054], [0064], [0066], [0070], [0086]-[0087], [0108]; claim 63.	1-10
A	(ZIPPERIAN, D) Silicon Carbide Abrasive Grinding. Quality Matters Newsletter. PACE Technologies. November 2002, Vol. 1, Issue 2; pages 1-3.	1-10

☐ Further documents are listed in the continuation of Box C.☐ See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

23 September 2016 (23.09.2016)

Date of mailing of the international search report

13 OCT 2016

Name and mailing address of the ISA/

Mail Stop PCT, Attn: ISA/US, Commissioner for Patents
P.O. Box 1450, Alexandria, Virginia 22313-1450

Facsimile No. 571-273-8300

Authorized officer

Shane Thomas

PCT Helpdesk: 571-272-4300
PCT OSP: 571-272-7774

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US16/35500

Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box No. III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

****-Please See Supplemental Page-****

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying additional fees, this Authority did not invite payment of additional fees.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Group I, 1-10

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee.
- ☐ The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation.
- ☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT
Information on patent family members

International application No.

PCT/US16/35500

-***-Continued from Box No. III: Observations Where Unity Of Invention Is Lacking:

This application contains the following inventions or groups of inventions which are not so linked as to form a single general inventive concept under PCT Rule 13.1. In order for all inventions to be examined, the appropriate additional examination fees must be paid.

Group I, Claims 1-10 are directed toward methods for silencing a target gene in a plant by introducing a nucleic acid into the plant, comprising, in any order, the steps of: (a) abrading a surface of a plant with a particulate of a size greater than about 2.5 micrometers; (b) applying an RNA to the surface of the plant, wherein the RNA comprises at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant; whereby the target gene is silenced.

Group II, Claims 11-18 are directed toward a method for providing a plant exhibiting modification of a target gene in the plant, comprising directly regenerating a plant from a source plant, wherein the source plant has been contacted with a composition comprising an RNA that comprises at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant, wherein the directly regenerated plant exhibits modification of the target gene.

The inventions listed as Groups I and II do not relate to a single general inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: the special technical features of Group I include abrading a surface of a plant, not present in Group II; the special technical features of Group II include regenerating a plant from a source plant, not present in Group I.

Groups I and II share the technical features including: a method for providing a plant exhibiting modification of a target gene in the plant, comprising contacting a plant with a composition comprising an RNA that comprises at least 19 consecutive nucleotides having a sequence that is complementary or identical to the sequence of a target gene in the plant.

However, these shared technical features are previously disclosed by US 2011/0296556 A1 to Sammons et al. (hereinafter 'Sammons').

Sammons discloses a method for providing a plant exhibiting modification of a target gene in the plant (a method for effecting regulation of a target gene in a plant (a method for providing a plant exhibiting modification of a target gene in the plant); paragraph [0004]), comprising contacting a plant with a composition (comprising topically contacting a plant with a composition that can permeate or be adsorbed into plant tissue (comprising contacting a plant with a composition); paragraphs [0004], [0006]) comprising an RNA that comprises at least 19 consecutive nucleotides (comprising an RNA that comprises at least 19 consecutive nucleotides; paragraph [0050]) having a sequence that is complementary or identical to the sequence of a target gene in the plant (having a sequence that is complementary or identical to the sequence of a target gene in the plant; paragraphs [0004], [0054]).

Since none of the special technical features of the Groups I and II inventions is found in more than one of the inventions, and since all of the shared technical features are previously disclosed by the Sammons reference, unity of invention is lacking.