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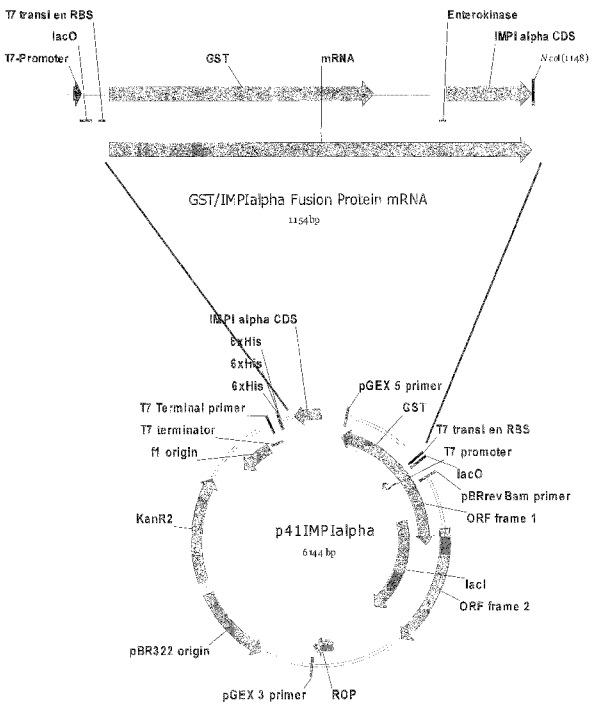
(54) Title: INSECT METALLOPROTEINASE INHIBITORS

Fig. 1



(57) Abstract: A polypeptide having at least 70% homology, in particular 80%, 90% or 95% homology to the polypeptide of SEQ ID No 2 representing the wild-type of the protein insect metalloproteinase inhibitor IMPIa and having at least one mutation at position 35, 36 and/or 39 of the amino acid sequence of IMPIa and the polypeptide having an IC_{50} value to thermolysine of less than the IC_{50} value of IMPIa wherein - the nonpolar amino acid isoleucine at position 35 of IMPIα is replaced either by a nonpolar amino acid selected from the group consisting of leucine, methionine and phenylalanine or by polar amino acid selected from the group consisting of cysteine, asparagine, glutamine, histidine, lysine and arginine; and/or - the nonpolar amino acid isoleucine at position 36 of IMPIa is replaced either by a nonpolar amino acid selected from the group consisting of valine, phenylalanine and tryptophan or by polar amino acid selected from the group consisting of tyrosine, serine, threonine, asparagine, glutamine, histidine, lysine and arginine; and/or - the polar amino acid position 39 of IMPIa is replaced either by the nonpolar amino acid valine or by the polar amino acids histidine or lysine.

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Insect Metalloproteinase Inhibitors

FIELD OF THE INVENTION

This invention relates to peptides and proteins inhibiting protease activity. More specifically, the invention relates to peptides exhibiting activity against a spectrum of proteases of microbial or fungal origin, in particular against the metzincin family including thermolysin, anthrax neutral protease 599 (npr599), pseudolysin, and aureolysin. The invention relates further to methods for producing the peptides, to pharmaceutical compositions comprising the peptides or proteins, and to methods of using the peptides or proteins to prevent and/or treat bacterial or fungal infections and their symptoms, in particular to reduce the toxic effects of secreted or membrane bound bacterial proteases such as aureolysin, bacillolysin, pseudolysin, vibriolysin, and anthrax npr599 by inhibiting their respective proteolytic activity.

BACKGROUND OF THE INVENTION

Antibiotics are the actual standard treatment for bacterial infections. The widespread and sometimes inadequate use of antibiotics, however, has led to pathogens developing resistance against one or even multiple antibiotics. Multidrug resistant bacteria pose an increasing and serious threat to infected patients. Therefore the search for new companion compounds dampening the bacterial virulence has become an urgent need during the last years. A a rapid effect of compounds against dangerous or life threatening symptoms of bacterial infection is desirable to protect the patient immediately against these symptoms even before antibiotics or alternative compounds can kill the microbes.

For decades it has been known that bacteria release proteases (Duthie et al. 1949). Later the pathogenic role of microbial proteases as virulence factors was recognized experimentally (Snell et al. 1978). Proteolytic enzymes produced by pathogens and parasites are essential for their successful infection of the host. Regardless of their substrate specificity the key function of individual pathogen-derived proteinases is the degradation of host proteins and peptides (Armstrong et al. 2006). Extracellular digestion of host proteins facilitates, for example, penetration across physical barriers of the host, for which the degradation of the extracellular matrix may serve as a prominent example. Furthermore, the degradation of immunity-related defense molecules of the host, including antimicrobial

peptides (AMP) (Otto 2008), and the acquisition of nutrients are important functions for the pathogen to survive. However, only those pathogen-associated proteolytic enzymes can serve as virulence factors that cannot or can only partially be inactivated by corresponding, endogenous host proteinase inhibitors. Virulence factors usually complement each other in degrading host peptides and proteins during infection. Consequently, pathogen-derived proteinases without corresponding endogenous inhibitors in the host will be evolutionarily selected because pathogens would otherwise waste their limited resources (Vilcinskas 2010).

Mammals and many invertebrates lack specific inhibitors for proteases of the M4 or Metzincin family of metalloproteinases, of which thermolysin from *Bacillus thermoproteolyticus* is a salient example. Thermolysin-like metalloproteinases encompass many prominent toxins produced by human pathogens, including aureolysin, bacillolysin, pseudolysin, vibriolysin, and anthrax npr599, which are presumably at the origin of many pathological symptoms associated with severe infections such as septicemia, hemorrhagic tissue bleeding, necrosis and enhancement of vascular permeability (Chung et al. 2006). Severe diseases like gastric and peptic ulcers and gastric carcinoma originate at least partly from the effect of metalloproteinases of the M4 family from pathogens (Schmidtchen et al. 2003, Smith et al. 1994).

For example, the major symptoms of an infection with bacillus anthracis are caused by the release of virulence factors from the bacteria. The well known virulence factor anthrax toxin leads to the destruction of the host macrophages, thereby enabling the bacteria to spread in the host tissue without encountering host defense. The anthrax toxin is a holoenzyme containing three different proteins, each of which is present in multiple copies. They are called protective antigen (PA), edema factor (EF), and lethal factor (LF). To be become active, it is required that at least PA and EF or PA and LF are combined. Just a single protein or just EF and LF without PA are not active in laboratory animals.

More recently it became evident that the combined activity of the PA, EF and LF alone does not cover all observed clinical anthrax symptoms. It was shown that anthrax harbors further genes coding for secreted proteins exhibiting proteolytic activity. Amongst those, gene BA0599 from bacillus anthracis (BA) codes for a protease called neutral protease 599 (npr599), which belongs to the M4 family of

proteases and is highly homologous to bacillolysins from other *bacillus* species (Popov et al. 2005, Chung et al. 2006, Chung et al. 2008).

Currently several treatment options for anthrax infections exist. A vaccine is available which protects with a safety of more than 90%, but usually the degree of vaccination against anthrax in a human population is low and so far it is not expected that this degree will increase. Anthrax can further be treated with antibiotics in case of an acute infection. However, the toxins shed by the bacteria continue to cause irreversible damage to tissue before the number of bacteria is sufficiently reduced by antibiotics. It is even likely that exposure to antibiotics stimulates bacteria to temporarily shed increased amounts of virulence factors, causing accelerated and irreversible damage of patient tissue. Furthermore, manmade synthetic or recombinant versions of the natural bacterial toxins may be inhaled by humans without bacteria being present, in which cases an antibiotic would have no curing effect at all.

Therefore the task of inhibiting or antagonizing bacterial toxins directly was recognized as a challenge, and inhibition strategies were published. In the case of Pseudomonas aeruginosa virulence factor LasB (Cathgart G.R. et al, 2011), N-mercaptoacetyl-Phe-Tyr-amide (K(i) = 41 nM) was suggested as inhibitor. Similarly, Anthrax Lethal Factor protease inhibitors were suggested in the past, for example (2R)-2-[(4-fluoro-3-methylphenyl)sulfonylamino]-*N*-hydroxy-2-(tetrahydro-2H-pyran-4-yl)acetamide (Xiong Y. et al, 2006, US 7,504,425 B2), as well as a compound named NSC12155 (Panchal R.G. et al. 2004), and substrate derived hydroxamates with Ki of up to 1 nM (In-2-LF) (Tonello F. et all 2002). A peptide derived from the substrate (MLARRKKVYPYPMEPTIAEG-amide) was found to inhibit LF with a Ki of 1 nM as well (Turk B.E. et al. 2002). No inhibitor of LF reached a clinical development stage so far. Side effects of some inhibitors are known, comprising, for example, the inhibition of endogenous furin in parallel to LF. In the laboratory, Nrp599 can be inhibited with EDTA and 1,10-phenanthroline (Chung et al. 2006), but further, therapeutically acceptable Nrp599 inhibitors are not published up to now.

Several inhibitors of other members of the thermolysin family were described earlier. Phosphoramidon is produced by the Bacterium *Streptomyces tanashiensis*, and Talopeptin is derived from it. Phosphonomadites termed ZG^{P}LL

and ZF^{P}LA display inhibitory constants in the low nanomolar range (Holden et al. 1987) with respect to thermolysin.

Khan et al described 1β-d-arabinofuranosyl-*N*^{4}-lauroylcytosine as potent inhibitor of thermolysin out of set of 12 inhibitors found in virtual screening run (Khan et al. 2009); and 3-Phenyl-2-(trifluoromethyl) quinazolin-4(3*H*)-one as most potent inhibitor out of a set of novel 38 quinazolin-4(3*H*)-ones (Khan et al. 2010).

Therapeutic inhibition of thermolysin activity by administering non-specific metalloproteinase inhibitors, however, is biased by negative side effects induced by collateral inhibition of essential host enzymes such as its matrix metalloproteinases. Phosphoramidon, for example, is inhibiting the endogenous protein *endothelin converting enzyme* (ECE). Furthermore, some of these compounds are toxic, while for others the toxicity is obviously unknown. Consequently, a strong demand for molecules capable of specifically inhibiting one microbial enzyme (Adekoya et al. 2009) or a corresponding enzyme family persists.

Streptomyces metalloproteinase inhibitor (SMPI) (Oda et al. 1979) is a known proteinaceous inhibitior of thermolysin. SMPI consists of 102 amino acids and exhibits two cystein bridges exhibiting an IC50 of 0.6 nM for Thermolysin (Hiraga et al. 1999). It was discovered, however, that SMPI is degraded by other bacterial proteases (Tsuru et al.1992), disqualifying SMPI for use as a compound for treating bacterial infections. Furthermore, its low molecular weight of ca. 12 kD means that it is rapidly cleared from the bloodstream by renal filtration.

The only other peptide inhibitor of thermolysin reported to date which specifically inhibits thermolysin-like enzymes is the insect metalloproteinase inhibitor IMPIα. It was originally discovered in and purified from the hemolymph of immunized *G. mellonella* larvae (Wedde et al. 1998). Its active moiety comprises 69 amino acids including intramolecular cystein bonds, and a molecular weight of 7667.7 Da. The molecule has a reported IC50 of 0.62, 0.86 and only 81.66 nM for thermolysin, bacillolysin and pseudolysin, respectively, all of which are enzymes belonging to the M4 protease family (Clermont et al.2004). IMPIα was tested against human metallo-matrix proteases MMP1,2,3,7,8, and -9, of which only MMP1 and MMP2 showed a negligible inhibition. IMPIα was recombinantly produced in Schneider cells (Clermont et al. 2004) and later in E.coli. Although the structure of IMPI was not known until recently (Gomes-Rueth. 2011), it was suggested by homology searches that IMPIα should belong to the I8 or Ascaris fami-

ly of serin protease inhibitors, despite the fact that IMPIα inhibits a metalloprotease and not a serin protease (Wedde et al., 2007). From this comparison it was further deduced that an active site loop would be present in IMPIα between aa 33 – aa40, including a cleavage site between aa37 (Asparagine) and aa38 (Isoleucin). Other known protein inhibitors of metalloproteinases do not inhibit proteinases of the M4 protease family.

In summary, most known inhibitors o of members of the M4 protease family are too toxic, unspecific, unstable or difficult to manufacture or express, or require a different specificity profile against metalloproteases..

SUMMARY OF THE INVENTION

It is therefore an object of the present invention to provide inhibitors of M4 protease family members with an increased potency for specifc M4 proteases, a selected if not engineered specificity profile including the potential requirement for a broad band metzincin inhibitior. Furthermore, higher stability, lower toxicity or more efficient manufacturing or expression profile is requested.

The object underlying the invention is accomplished by a polypeptide according to claim 1 and other independent claims. The claims depending on one or more independent claims are concerning specific embodiment of the invention.

Tfhe polypeptide is a polypeptide having at least 70% homology, in particular 80%, 90% or 95% homology to the polypeptide of SEQ ID No 2 representing the wild-type of the protein insect metalloproteinase inhibitor IMPIα and having at least one mutation at position 35, 36 and/or 39 of the amino acid sequence of IMPIα wherein

- the nonpolar amino acid isoleucine at position 35 of IMPIα is replaced either by a nonpolar amino acid selected from the group consisting of leucine, methionine and phenylalanine or by polar amino acid selected from the group consisting of cysteine, asparagine, glutamine, histidine, lysine and arginine; and/or
- the nonpolar amino acid isoleucine at position 36 of IMPIα is replaced either by a nonpolar amino acid selected from the group consisting of valine, phenylalanine and tryptophan or by polar amino acid selected from the group consisting of tyrosine, serine, threonine, asparagine, glutamine, histidine, lysine and arginine; and/or

- the polar amino acid position 39 of IMPIα is replaced either by the nonpolar amino acid valine or by the polar amino acids histidine or lysine.

The polypeptide of of the invention shows in particular an IC_{50} value to thermolysine of less than the IC_{50} value of IMPIα.

In an embodiment of the invention the polypeptide of the invention has the amino acid sequences of SEQ ID numbers 10, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, 78, 80, 82, 84.

In an another embodiment of the invention the polypeptide of the invention may comprise chemical modifications in the side chain or at the N and/or C terminal for improving biological or chemical properties such as bio availability, stability, effectivity. The modification may also provide for a detectable label, for example a chemiluminescent structural element, one or more radioactive isotopes in one or more side chains of an amino acid in the polypeptide, an enzyme which is able to generate a colour reaction and the like.

Subject matter of the present invention is also a fusion polypeptide comprising the polypeptide of the invention and at least one polypeptide having a physiological function in particular an antibody or antibody fragment, scaffolds such as lipocalin, ankyrin, fibronectin, transferrin, tetranectin, adnectin, albumin, uteroglobin, or protein A, functional peptides such as transferrin, peptides useful for diagnostic applications, such as green fluorescent protein (GFP), or peptide tags enabling immobilization on technical surfaces, such as hexahistidine, or glutathione-S-transferase (GST).

In an embodiment of the invention the polypeptide of the invention has the amino acid sequences of SEQ ID numbers ,6,8, 12, 86, 88, 90, 92.

There are three superfamilies (cytosolic, mitochondrial, and MAPEG) of GSTs: while classes from the cytosolic superfamily of GSTs possess more than 40% sequence homology, those from other classes may have less than 25%. Cytosolic GSTs are divided into 13 classes based upon their structure: alpha, beta, delta, epsilon, zeta, theta, mu, nu, pi, sigma, tau, phi, and omega. Mitochondrial GSTs are in class kappa. The MAPEG superfamily of microsomal GSTs consists of subgroups designated I-IV, between which amino acid sequences share less than 20% identity. Human cytosolic GSTs belong to the alpha, zeta, theta, mu, pi,

sigma, and omega classes, while six isozymes belonging to classes I, II, and IV of the MAPEG superfamily are known to exist:

GST Class	*Homo sapiens* GST Class Members
Alpha	GSTA1, GSTA2, GSTA3, GSTA4, GSTA5
Kappa	GSTK1
Mu	GSTM1, GSTM1L (RNAi), GSTM2, GSTM3, GSTM4, GSTM5
Omega	GSTO1, GSTO2
Pi	GSTP1
Theta	GSTT1, GSTT2, GSTT4
Zeta	GSTZ1 (aka GSTZ1 MAAI-Maleylacetoacetate isomerase)
Microsomal	MGST1, MGST2, MGST3

In a further embodiment of the invention the fusion polypeptide of the invention is linked by a peptide of 1 to 100 amino acids in length to the the polypeptide of the invention. Alternatively polypeptide of the invention can be conjugated by a chemical linking group with a polypeptide having a physiological function to yield the fusion polypeptide of the invention.

Polypeptides or proteins can be manufactured by chemical methods, such as solid phase syntheses or by means of genetic engineering by means of utilizing coding polynucletides in a suitable expression system. Subject matter of the present invention is also a polynucleotide coding for the polypeptide of the invention or the fusion polypeptide of the invention. The polynucleotide of the invention may be operably linked to a heterologous promoter.

Subject matter of the present invention is also a pharmaceutical composition comprising the polypeptide of the invention and/or the fusion polypeptide of the invention or their derivatives and/or a nucleic acid comprising a section coding for the polypeptide of invention.

The polypeptide of the invention or the fusion polypeptide of the invention can be used in the treatment of an animal or human infected by microorganisms secret-

ing bacterial toxins of the M4 or Metzincin family of metalloproteinases, in particular thermolysine, aureolysin, bacillolysin, pseudolysin, vibriolysin or anthrax npr599.

The polypeptide and/or the fusion polypeptide of the invention can be used for detection of the presence or activity of proteases belonging to the M4 family in a sample.

The invention is based on the result that mutations between amino acid 35 and 39 of IMPIα (SEQ ID NO: 10, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, 78, 80, 82, 84) could be used to alter systematically the specificity profile and potency related to specifically inhibiting members of the thermolysin M4 protease family. Accordingly, the polypeptide IMPIα of the invention inhibit further proteases of the thermolysinM4 protease family, particular thermolysine, aureolysin, bacillolysin, pseudolysin, vibriolysin or anthrax npr599, and that the mutein IMPIα exhibit different inhibitory constants with respect to the various proteases.

It was further found that all polypeptides of the invention and wtIMPIα consisting of SEQ ID NO:2 can all be produced very effectively employing the same recombinant process in bacteria, particularly in *E.coli*, in high quantities, high quality and without requiring refolding. In case of wtIMPIα, the DNA is including an oligonucleotide as SEQ ID NO:1. This discovery is surprising because wtIMPIα and all the polypeptide of the invention contain five cystein bridges each. Proteins or peptides containing many cystein bridges are usually considered as difficult or impossible to express in any bacteria, including E.coli., and published protocols reveal low production efficacy and requirement for cooling (see, for example, Comis Ruets, 2011) while according to the method of invention wtIMPIα and the polypeptide of the invention can be produced without requiring cooling of the fermenter during the protein expression phase, and without requiring protease inhibitors to be added to protect the protein product from degradation.

As used herein, the following terms have the following meanings unless expressly stated to the contrary.

The term "wtIMPIα" refers to a protein with an amino acid sequence as in SEQ ID NO:2 and consists of the N-terminal fragment of the full length IMPI molecule which is endogenously cleaved from the larger precursor molecule IMPI.

The term "wtIMPIβ" refers to a protein with an amino acid sequence as in SEQ ID NO:4 and consists of the C-terminal fragment of the full length IMPI molecule which is endogenously cleaved from the larger precursor molecule IMPI.

The term "loop mutein IMPIα" comprises all proteins comprising an amino acid sequence identical to SEQ ID NO:2 (wtIMPIα), with the exception of the amino acid stretch 35 to 39 , and in particular with exception of the amino acid stretch 35, 36, or 39, in which at least one amino acid is exchanged, added, or deleted. By this definition "Loop mutein IMPIα" includes, for example, amino acid sequence SEQ ID NO: 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, 78, 80, 82, 84, in which one amino acid at position 35,36, or 39 is exchanged. In SEQ ID NO 10 two additional amino acids at positions 37 and 38 are exchanged.

The term "mtIMPIα" or "Mutein IMPIα" is the polypeptide of the invention and includes all recombinant or synthetic proteins with an amino acid sequence which is at least 70%, especially 80% 90%, 95% homologous, but not identical to SEQ ID NO: 2 and which inhibits thermolysin or another protease of the M4 protease family with an IC_{50} < 1000 nM, especially IC_{50} < 100 nM, in particular those which can be expressed in E.coli and especially those including proteins of the group "loop mutein IMPIα" and "structure variants of IMPIα". mtIMPIα include inserted, substituted or deleted residues, including N-terminal and C-terminal and intrasequence deletions, provided the selected mutations preserve the biological activity of the Mutein IMPIα.

The term "M4 family of proteases" refers to the definition given in the MEROPS online database of peptidases (http://merops.sanger.ac.uk/cgi-bin/famsum?family=M4), and described also by Barret et al. (2004).

The term "biologically active" as used herein refers to wtIMPIα or the polypeptide of the invention demonstrating inhibition of thermolysin or another protease of the M4 protease family with an IC_{50} <1000 nM especially IC_{50} < 100 nM or better.

The term "IMPIα family" alone shall include biologically active mtIMPIα and wtIMPIα

BRIEF DESCRIPTION OF THE FIGURES

Fig.1: Vectormap of the hybridplasmid pET41IMPIα. Total mRNA was isolated from immunchallenged Galleria melonella und subsequently used as template for the amplification of DNA coding for IMPIα. The amplicon was then cloned into the PshA1 linearized pET-41A vector (Novagen) resulting in a GST (Glutathion-S-transferase) fusionprotein. Accurate in frame cloning of IMPIα into the vector was verified by sequencing.

Fig. 2: SDS-PAGE of the soluble cytoplasmatic fraction of Rosetta gami 2 three hours after induction of the T7-Expression system. The overexpressed fusionprotein is marked by an arrow. The corresponding controls (empty pET-41 and the uninduced T7-Expression system) show no corresponding bands.

Fig. 3 HPLC UV detector data from purification run of GST-tagged IMPIα. The purification has been carried out by affinity chromatography on a HPLC, using a resin with immobilized glutathion at room temperature. The purification has been carried out by affinity chromatography on a HPLC, using a resin with immobilized glutathion at room temperature.

Fig. 4a SDS-PAGE of the purified GST-tagged IMPIα For larger amounts of mutein GST Bind Fractogel Cartriges (Merck, Darmstadt) were used, while for amounts less than 40 mg glutathione spin columns (Pierce; Thermo Scientific, Rockford) have been deployed.

Fig. 4b: SDS-PAGE after the specific cleavage of the n-terminal GST-tagged IMPI alpha by recombinant enterokinase (Merck, Darmstadt). The cleavage was carried out for 16 hours at room temperature. Untagged IMPIα is marked by an arrow.

Fig. 5: Sucessfull downstream processing was verfied by mass spectroscopy on a micrOTOF-Q mass spectrometer (Bruker Daltonics). Analysis of samples was achieved by chromatography on a reverse-phase column (Acclaim 120, C8, 3 µm, 120 Å, 2.1 x 150 mm; Dionex) by applying a gradient of 1-80 % (1,63 %/min) acetonitril in water containing 0.1 percent formic acid. By-product (peak termed 1 and 3) and main product (peak 1) show the correct mass of 7,677 kDa.

Fig. 6: wild type protein assembly of full length IMPI, including IMPIα and IMPIβ in G. melonella.

Fig. 7: inhibitory plot of wtIMPIα and phenanthronin vs. the anthrax protein nrp599 and InhA

Fig. 8a: Multiple alignement of IMPIα homologues of *Galleria melonella*.

Fig. 8b: Multiple alignment of IMPI´s found in the transcriptome of other lepidoptera species (Sequences unbublished).

Fig. 8c: Pairwise alignment of the M4-metalloprotease thermolysin from *Bacillus thermoproteolyticus* with the metalloprotease npr599 from Bacillus anthracis (strain A0248)

Fig. 9: The binding of the peptidic inhibitor IMPIα to the M4-metalloprotease thermolysin (*Bacillus thermoproteolyticus*) is depicted. Red areas represent Van der Waals contact areas between inhibitor and protease. The isoleucin (P1´)(purple) fits perfectly into the S1´pocket which is responsible for specificity of the metalloptotease hence the inhibitor is binding in a lock and key kind of fashion

Fig. 10: Schematic drawing of a lateral flow device for detecting protease specific activity. The principle is based on thin layer chromatography. The eluent carries ichor collected by a cotton swap via tangential flow through the device. M4-metalloproteases produced by pathogens therefore also migrate through the device, until they get bound to IMPI muteins exhibiting certain specificity for different metalloproteases. The different muteins are bound at concentrated spots on the solid phase of the device, so that different metalloproteases can be clearly identified. Spots are made visible afterwards by detection antibodies present in the mobile phase.

Table 1: Overview of amino acid exchanges and respective functionality for Thermolysine and Pseudolysine, and respective ratio of IC_{50} values. The ratio changes mean that an inhibitor with designed ratio of IC_{50}s with respect to different proteases can be created.

Table 2: Detailled experimental parameters and results for inhibition of Thermolysine.

DETAILED DESCRIPTION OF THE INVENTION

In one aspect of the invention, biologically active loop polypeptide of the invention polypeptides are provided which exhibit a specificity profile different from the wtIMPIα profile for inhibiting members of the M4 protease family. The fact that the loop polypeptide of the invention are biologically active at all is surprising because the loop amino acids were particularly well conserved during evolution. It is even more surprising that altering a specific loop amino acid could increase the binding constant for thermolysin or pseudolysin, or even both.

New ways of improving the inhibitory constant by substituting I36 (Isoleucine) of wtIMPIα have been developed, which is surprising since it was assumed that at this position 36 isoleucine would already exhibit a perfect profile for a C-C interaction with amino acids H146 (histidine) and T157 (tyrosine) of the peptidase, and therefore no attempts were reported to design IMPI mutants or small molecules with the goal to strengthen this interaction further. The inventors found, however, that a long aliphatic side chain of an amino acid or modified amino acid at this position, preferably with an elevated pKa of 12 or higher, improves the inhibitory constant substantially. Examples of well suited amino acids comprise K, M, H, and in particular R.

Surprisingly it was found that further substitutions, for example those listed in the ensuing paragraph, could increase the potency of the inhibitor (Tables 1, 2):

Loop the polypeptide of the invention molecules exhibiting improved inhibition of Thermolysin comprise I35L, I36F, K, R, V, Y, Q, D, H, M, T, or W, R39K or V.

Loop the polypeptide of the invention molecules exhibiting improved inhibition of Pseudolysin comprise I35L or F, M, W or Y, I36R, Q, or M, R39K and A.

Loop polypeptide of the invention molecules exhibiting improved inhibition of both, Thermolysin and Pseudolysin in parallel comprise I35L, I36R, Q, or M, R39K.

Loop polypeptide of the invention exhibiting equal potency against both, Thermolysin and Pseudolysin, comprise I35L, or W.

This result is surprising because there no reasonable expectation of success if one amino acid is altered close to an area associated with the function of the respective protein. In particular this result is surprising considering the general assumption that a peptidic inhibitor of an enzyme should contain hydrophobic ami-

no acids binding to the enzyme close to its hydrophobic active center (Handbook of proteolytic Enzymes 2013, Biela et al. 2013) The results show, however, that hydrophobicity of the amino acids in the polypeptide of the invention is not the dominant factor for the inhibition.

It is understood that combinations of single site mutations could alter the properties of the polypeptide of the invention even further, and further increase the potency of the inhibitor.

The polypeptide of the invention may be obtained by known techniques for directed evolution as described by Arnold et al., 2003a and Arnold et al., 2003b, or may further be obtained by knowledge based engineering of the respective amino acid sequence, including application of rules for conservative or non-conservative amino acid replacement considering size, charge, polarity and other biochemical parameters of the amino acids as know by one of ordinary skill in the art by the teaching and principles disclosed in US Pat. Nos. 7,732,587 and 4,554,101, or by using suitable computer programs calculating or estimating the binding energy of the Mutant IMPIα and the selected target proteases, by assessing homologies to other proteins or the antigenicity of the protein sequence, and introducing the calculated, modified gene sequence into the bacterial genome by standard molecular biotechnology methods. Amino acid deletions within any IMPIα and its muteins may be made in regions outside the loop region (aa 35 to 39 of SEQ ID NO:2), where they are supposed to not directly interact with the target protease. Amino acid deletions may also be made within the loop region, where they are supposed to more likely modify the biological activity.

Comparison of wtIMPIα with similar proteins from other species, such as SEQ ID: NO 95 from Solenopsis, and 23 other insect related aa sequences depicted in Fig. 9 reveals that the loop region (aa 33 to 41 in SEQ ID: NO 2, and in particular in region 35 to 39) is highly conserved with the exception of aa39 (R). It is thus surprising that Loop Mutein IMPIα, for example, SEQ ID: NO 10, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, 78, 80, 82, 84 also exhibit a strong, undexpected thermolysin inhibiting activity.

In another aspect of the present invention it has been found that wtIMPIα and Mutein IMPIα, and in particular Loop polypeptide of the invention can be recombinantly produced in bacterial cells under conditions identified by the in-

ventors, especially at room temperature and up to 37°C, under conditions which do not require cooling of the fermenter or refolding of the protein produced. The method according to the invention, equipment and reagents used are described in detail in example 2 below. It should be understood, however, that modifications in concentrations of reagents used, and modifications in the genetic configuration of the producer organism (E.coli) could be made without affecting the output of correctly folded protein. For example, E.coli could be replaced by other bacteria as production host, additional plasmids or gene stretches with additional function could be added to the producer bacteria. Furthermore, medium components could be provided in different order or concentrations, and incubation times could be varied. It should be understood that those modifications which influence the production efficacy of biologically active Mutein or wtIMPIα negatively by 50% or less or even positively are considered within the scope of the invention.

The invention contemplates further modifications of elements of the expression system inasmuch as the expression of a functional Mutein or wtIMPIα without the need for refolding is maintained to at least 50% compared to the system disclosed in Example 3 and 4, and no cooling of the fermenter is required. For example, other expression vectors than pET-41a(+) may be used, which may not be linked to a fusion tag, or may be linked to a fusion tag other than GST, His-tag; and other promoters than T7, and may also include further regulators or other fusion elements. Depending on the operons employed, IPTG or other inducers may be employed. Other restriction sites than PshAI may be used in pET-41a(+) or in other vectors, for example XhoI, EcoR I, BamH 1. Other E.coli strains than Rosetta-gami 2 pLysS may be used, with or without additional t-RNA genes or ahp C mutation, and other bacteria than E.coli may also be employed.

Further modifications may relate to the downstream process steps which are known to the one skilled in the art, including but not limited, to incubation times, reagents and their concentrations and disposables used, for example capture resins and chromatography equipment and separation media for purification.

Recombinant production of Mutein or wtIMPIα in bacteria is very cost effective compared to all other production methods for proteins. Cloning and the subsequent development of a cell line producing the peptide are faster and less costly with bacteria as, for example, with mammalian or insect cells. During production,

the amount of protein produced per volume fermentation broth is much higher compared to eukaryotic cell cultures. Consequently there is a strong demand for methods of recombinant protein producing in bacteria, especially in E.coli since they are a proven host frequently used for protein production. It is known that wtIMPIα retains its biological activity after deglycosylation (Wedde et al. 1998); therefore bacterially produced Mutein or wtIMPIα lacking glycosylation are biologically active once folded correctly.

The skilled artisan would expect, however, that a protein like Mutein or wtIMPIα exhibiting 5 cystein bridges would not be correctly folded and thus be accumulated almost entirely in inclusion bodies of the E.coli host. While several processes were developed earlier to isolate and dissolve inclusion bodies, and to denature its protein content using chaotropic salts like urea or guanidylchloride, and to withdraw the salts again in an ensuing step, for example by dialysis, to enable refolding of the protein in a kinetically controlled manner. However, the success rate of refolding proteins with multiple cystein bridges, such as as IMPIα, is usually low, and even today time consuming and resource consuming trial and error methods are required to find adequate refolding conditions. It is generally not yet possible to predict or experimentally determine suitable refolding conditions.

It is thus surprising that, without any known exception, a family of cystein rich proteins such as Mutein or wtIMPIα with a complex tertiary structure can be recombinantly produced in bacteria by the method of invention and without the need for refolding steps, in particular that the production can take place without cooling of the fermenter, the latter being a common measure to reduce the protein production rate of the bacteria and thereby to reduce the ratio of incorrectly folded protein vs. correctly folded protein.

In another aspect of the invention, the use of Mutein or wtIMPIα, and in particular of wtIMPIα is provided for inhibition of the anthrax protease nrp599. Current approaches to treat Anthrax include vaccination such as BioThrax (Emergent Biosolutions, Rockville MD, U.S.A) for disease prevention, or antibiotics such as Ciprofloxacin or Doxycyclin (http://www.nationalterroralert.com/anthraxtreatment/). While the vaccine is only approved for preventive use, it takes several days until antibiotics exert effects on bacteria to an extent that the symptoms caused by their virulence factors are reduced. Compounds neutralizing virulence factors were developed re-

cently and are generally well known in the state of the art and for anthrax most compounds targeting virulence factors developed so far target LF. However, no specific inhibitor of nrp599 was reported so far, although recent evidence shows that nrp599 is one of the major anthrax virulence factors inducing hemorrhagic bleeding, for example (Chung et al., 2006) by targeting the fibrinolytic system (Chung et al, 2011), and that it also degrades the van Willebrand-Factor and thereby interferes with the coagulation cascade (Chung et al., 2008).

US 2004/018193 teaches the concurrent or offset administration of both, antibiotics and antigenic compounds, in particular to treat anthrax infected humans. The authors provide several examples of protease inhibitors including hypothetical antibodies, but they do not disclose nrp599 as a target.

Surprisingly it was found that polypeptides of the invention and in particular wt IMPIα is a potent and specific inhibitor of nrp599, despite the fact that bacillus anthracis is not known to infect insects. It was found that wtIMPIα inhibits npr599 at nanomolar concentrations.

To treat anthrax infection or symptoms caused by anthrax toxins, the polypeptide of the invention and in particular wt IMPIα may be combined with antibiotics, in particular Ciprofloxacin or Doxycyclin, with antimicrobial peptides, with other inhibitors of bacterial proteases, or with compounds improving the physiological state of the infected patient.

The polypeptide of the invention may also be produced effectively in recombinant cell lines and tissue, including mammalian and insect expression systems, plant and bacterial systems; or synthetically.

Mutein and wtIMPIα each may typically be isolated and purified to be free of other proteins and protein fragments. Preferably, Mutein and wtIMPIα is about 80% free of other proteins which may be present due to the production technique used in the manufacturing process. More preferably, the polypeptide of the invention or wtIMPIα is about 90% free of other proteins, particularly preferably about 95% free of other proteins, and most preferably about >98% free of other proteins. It will be appreciated, however, that the desired protein may be combined with other active ingredients, chemical compositions and/or suitable pharmaceutical formulation materials prior to administration as medication, as described in further detail below.

The biological activity of Mutein or wtIMPIα can be assessed in vitro by means of assays, such as measuring binding of the polypeptide of the invention or wtIMPIα to a protease immobilized on coated glass carrier used for Plasmon resonance spectroscopy. Alternatively, the polypeptide of the invention or wtIMPIα can be immobilized on the glass carrier and the protease could be added to start the assay. This assay is able to measure on- and off-rates for the respective interaction.

The biological activity can also be tested by means of an Parallel Line assay, for example in a microtiter plate format, and by detecting a coloration effect or by measuring changes in the fluorescence emitted upon excitation with a suitable light source.

Amino acid sequence additions may further include N- or C-terminal fusions ranging in length from one residue to 150 or more residues, as well as internal intrasequence insertions of single or multiple amino acid residues into Mutein or wtIMPIα ranging typically from about 1 to 10 amino acids, more typically from 1 to 5 amino acids, and most typical from 1 to 3 amino acids. N-terminal additions include the addition of a methionine or an additional amino acid residue or sequence. Another example of N-terminal addition includes the fusion of a signal sequence – provided the signal sequence SEQ ID NO: 94 is not bluntly added to wtIMPIα of SEQ ID NO: 2.

Further examples for additions to Mutein or wtIMPIα comprise fusions with antibodies (Abs) or antibody fragments. Ab fragments may comprise for example scFAb, scFv fragments, or single domain antibodies, or antibodies of camelide (hcIG) or shark (IgNAR) origin, or their respective VHH domains. The fusion may serve to provide a targeting moiety (the CDR domains) fused to to Mutein or wtIMPIα so that the fusion protein could be accumulated and enriched at surface displaying antigens specific for the antibody fragments. Another motivation for fusing Mutein or wtIMPIα to antibodies may be to benefit from the dimerization properties of the constant antibody regions to obtain a bivalent form of Mutein or wtIMPIα, in a way similar to Etanercept (Enbrel) in which 2 TNF receptor fragments are fused to the constant domains of an antibody. In addition, such bivalent antibody chimera is sufficiently large to escape from rapid renal clearance, for which a size exclusion barrier of roughly 65 kDa exists, so that the lifetime of the fusion protein in the patient body is strongly prolonged. Additions may also

comprise other scaffolds derived, for example, from lipocalin, ankyrin, fibronectin, transferrin, tetranectin, adnectin, albumin, uteroglobin, or protein A. Other additions contemplated comprise functional peptides, for example transferrin enabling the fusion protein to cross the blood brain barrier, a property potentially useful for inhibiting bacterial toxins accumulated in the brain or spinal cord, or additions useful for diagnostic applications, such as green fluorescent protein (GFP) enabling fluorescence detection, or peptide tags enabling immobilization on technical surfaces

Specific the polypeptide of the invention described herein may involve addition or deletion of or substitution with a non-native amino acid at the N- or C-terminus or at any site of the protein that is modified by the addition of an N-linked or O-linked carbohydrate. A cystein, for example, may be added for linking a water soluble polymer such as polyethylene glycol, or other amino acids like lysine, cysteine, histidine, arginine, asparaginic acid, glutamic acid, serine, threonine, or tyrosin could also be used for coupling polymers to the peptide.. Another example is the insertion of tripeptide sequences NXT or NXS or fragments thereof with X designating any amino acid except P, which may be recognized by a cellular enzyme adding glycosylation elements. Suitable, clinically acceptable, water soluble polymers include polyethylenglycol (PEG) and polysialic acid (PSA).

Another mutation or addition may include tags, in particular peptide tags, such as hexa-histidine which can be used to facilitate binding, for example, to a moiety of a purification column. These tags may comprise a peptide sequence serving as a substrate for a protease, so that the tag can be cleaved in purpose once it is not required anymore.

Further contemplated are Mutein or wtIMPIα to which chemical compounds or nanoparticles are added. This can be achieved for example by employing the enzyme O-6-Alkylguanin-alkyltransferase (AGT) or derivatives thereof to Mutein or wtIMPIα, or to a peptide tag. Other examples comprise conjugation to a cysteine being introduced by a mutation (Jagath et al. 2008), or to carbohydrate moieties (Fischer-Durand et al. 2010). Chemical compounds may include for example labeling moieties, in particular for in vivo imaging applications, targeting moieties such as receptor ligands to enrich the conjugated IMPIα family members in specific organs. Another compound suited to immobilize conjugated IMPIα family members on a surface or particle is biotin, for example for diagnostic purposes.

Nanoparticles such as quantum dots may be added for in vivo detection or imaging applications. Dendrimers or others particles increasing the size of the conjugate may be added to prevent the conjugate from cleared rapidly from the blood stream by the kidneys.

Modifications to the peptide backbone are also contemplated under the invention, such as beta peptides, where the amino group is attached to the beta carbon of the respective amino acid instead of the alpha carbon, rendering them invulnerable to proteases. Other modifications of the peptide backbone are possible, such as alkylation of the amid nitrogen and bioisosteric replacement of amide groups.

The present invention further provides polynucleotides which contain nucleotide sequences encoding Mutein or wtIMPIα, provided the polynucleotide is not identical to the complete wild type sequence SEQ ID NO:1, or SEQ ID NO:1 directly followed by SEQ ID NO:3. Further are considered nucleotide sequences in which oligonucleotide stretches are inserted which do not code for wtIMPIα. Based upon the present description and using the universal codon table, one of ordinary skill in the art can determine all of the nucleic acid sequences which encode Mutein or wtIMPIα. Presently preferred nucleic acid sequences include those encoding SEQ ID NO: 9, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, 79, 81, 83.

Chemical synthesis of Mutein or wtIMPIα is also contemplated, and is the obligatory route for synthesizing modified Mutein or wtIMPIα containing non-natural amino acids or backbone modifications. While chemical synthesis was traditionally regarded as being too expensive for larger peptides, recent advances in synthesizing technology like microwave assisted synthesis have recently shifted this limit towards larger proteins. Recent approaches also solve the issue of correct configuration of currently up to 3 cystein bridges by using several distinct protective groups in parallel.

While the invention contemplates the use of E.coli for producing Mutein or wtIMPIα with high quality and efficiency, other hosts may also be used for recombinantly producing Mutein or wtIMPIα, including other strains of E.coli, other bacteria like bacillus megaterium, fungal systems like yeast, pichia, or hansenula, eukaryotic cells like protozoa, insect cell lines like Schneider T7 or Sf9 mammalian animal cell lines like CHO, human cell lines like HEK293, or transgenic animals like goats, or plants like tobacco.

Mutein or wtIMPIα or modified versions thereof may be combined with ingredients to form a pharmaceutical composition. The pharmaceutical composition may include water and salts at physiological concentrations, solubilizing or dispersing agents, or anti-oxidant, or particles forming micelles, such as liposomes. Liposomes may also contain baIMPIα or modified versions thereof internally. This pharmaceutical composition may be filled in a glass or plastic vial, or in a syringe. The pharmaceutical composition may also contain additives supporting drying or freeze-drying of the pharmaceutical composition, for example cyclodextrins or saccharides, in particular disaccharides.

Mutein or wtIMPIα or modified versions thereof may be used as a medicine, in particular as a medicine for parenteral injection, to treat diseases in an animal, in particular in a mammal or bird, or in a human, and which are related to the activity of proteases from the M4 family. Such disease could be, for example, a bacterial infection, in particular infection by staphylococcus aureus, in particular multi-resistant staphylococcus aureus, bacillus anthracis, pseudomonas aeruginosa, helicobacter pylori, vibrio cholerae, or legionella pneumophilia. Such a disease could further be systemic inflammatory response syndrome (SIRS), or sepsis

Mutein or wtIMPIα or modified versions thereof may be administered parenterally, orally, or topically using suitable pharmaceutical compositions, or attached to a patch or wound debridement from where the medication elutes into a wound of the patient.

Mutein or wtIMPIα or modified versions thereof may be administered in combination with other treatments, in parallel or staggered, in particular with other inhibitors of virulence factors, antibiotics, or antimicrobial peptides.

Further contemplated are RNA molecules whose sequences comprise RNA sequences of Mutein or wtIMPIα, in particular sequences like SEQ ID 1, 5, 9, 11, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, 79, 81, 83, 85, 87, 89, 91, and which can be transcribed into Mutein or wtIMPIα protein in the target organism. Such RNA molecules may be administered in combination with transfection agent like liposomes or PEI.

The invention also contemplates the application of Mutein or wtIMPIα in devices for diagnostic purposes. In several cases it is beneficial to determine the prote-

ase activity in biological fluids, and in particular the protease activity profile if several of them are present.

For fluids contaminated with bacteria determining the protease activity would provide information not only about the presence of bacteria, but also of their activity and relative concentrations.

In wounds, and especially in slowly healing or chronic wounds, determining the protease activity profile also is of interest since the physiological stages of wound healing are characterized also by a specific evolution of protease activity during healing. It is even more important to distinguish between the activity of endogenous host proteases and the activity of bacterial proteases.

PCR or real time PCR assays do not provide sufficient information because these tests quantify the concentration of particular bacteria present, but they are not suited to test for the activity of the bacteria.

Currently laboratory tests for protease activity exist where the cleaved protease substrate becomes fluorigenic. Many of these tests determine overall protease presence; some of them are able to profile the protease concentrations (Chen et al. 2013). Currently the only Point-of-Care test for protease activity, Woundchek® manufactured by Systagenix, (Gatwick, Großbritannien) measures overall protease activity, but cannot quantify the concentration of each protease species separately (Strohal et al. 2012).

A diagnostic method according to the invention solves this issue. Mutein or wtIMPIα is used therein to bind M4 proteases specifically. Different concentrations of proteases can be determined when more than one type of Mutein or wtIMPIα is used. The difference in binding strength between the polypeptide of the invention or wtIMPIα is then used to calculate differences in concentrations of Mutein or wtIMPIα.

A preferred embodiment of the diagnostic method uses a device similar to Lateral Flow tests known in the state of the art; in which a membrane strip (suagfähig) is used to take up the fluid sample and to transport the sample across the membrane by capillary forces. In a preferred embodiment, the membrane consists of nitrocellulose or nitrocellulose endorsed by nylon fibres or tissue. At a particular location on that strip, Mutein or wtIMPIα are immobilized so that at least part of the sample fluid passes across this location during flow. Proteases present in the

sample are captured by the immobilized Mutein or wtIMPIα. In a second step one further solution containing detection capture molecules targeted against different binding epitopes of the proteases are added. These detection capture molecules are prepared to deliver a detection signal, which can be, for example, chromophoric, fluorescent, or electrochemical by means of a direct or enzymatic label. It is possible to increase signal strength by secondary capture molecules targeting the detection capture molecule. In a preferred embodiment, at least one kind of capture molecule is an antibody. A particular advantage of the diagnostic method according to the invention is that several proteases can be distinguished by means of specific binding to speiific locations where different Mutein or wtIMPIα are immobilized, while only a single sort of antibody is required. Furthermore, a control area is present at a different location on the membrane strip where detection capture molecules or secondary capture molecules bind to even if no proteases are present in the sample. In a preferred embodiment, the capture functionality in the control area is provided by immobilized proteases or immobilized detection capture molecules.

In a different embodiment, capture molecules other than Mutein or wtIMPIα, in particular antibodies are immobilized on the membrane strip and the proteases are added as detection capture molecule, in particular in combination with secondary capture molecules. This embodiment also requires a contral area containing immobilized proteases or Mutein or wtIMPIα.

The assessment of the protease content is made by visual inpection or a sensor device, including CMOS based imaging sensor or scanners. In case that sensors are used, a software can be employed to calculate the true concentration ratios of the proteases from potentially overlapping binding profiles of the polypeptide of the invention or wtIMPIα if the individual binding constants or other binding parameters are known. In particular such software may use a linear equation system to calculate the individual concentrations.

In another preferred embodiment, the binding kinetics are sampled, in particular by using an imaging sensor, to determine concentrations more accurately.

The sample may be added, for example, by a pipetting device, a paper strip, a cotton swab or a needle. The device may dispose of a seal preventing uncontrolled flow across the membrane strip.

To assist in understanding the present invention, the following examples are included which describe the results of a series of experiments. The following examples relating to this invention should not, of course, be construed in specifically limiting the invention and such variations of the invention, now known or later developed, which would be within the purview of one skilled in the art are considered to fall within the scope of the present invention as hereinafter claimed.

		Pos39													
	wt (avg)	RnG	RnK	RnL	RnV	RnW	RnF	RnI	RnM	RnA	RnQ	RnT	RnY		
IC50 mutein TLN		347,031	7,78	58,729	9,244	64,356	245,863	87,136	24,551	18,073	56,329				
IC50 wt TLN	12,53	12,728	12,524	12,41	12,538	12,362	12,489	12,548	12,571	12,542	12,605				
IC50 mutein PLN		1053,848	11,373	587,631	139,828	525,002	95,622	384,625	85,036	13,284	105,717	26,032	10000		
IC50 wt PLN	21,39	20,318	21,987	20,102	21,757	19,948	20,523	21,168	22,142	22,076	22,145	21,943	22,526		
ratio TLN:PLN	**0,59**	**0,33**	**0,68**	**0,10**	**0,07**	**0,12**	**2,57**	**0,23**	**0,29**	**1,36**	**0,53**				
		Pos38													
	wt (avg)	InF	InG	InQ	InL	InV	InH	InM	InW	InY					
IC50 mutein TLN		38,331	144,113	267,172	92,044	26,282	336,219	210,619	571,873	5,625					
IC50 wt TLN	12,52	12,571	12,481	12,614	12,563	12,563	12,491	12,478	12,375	12,553					
IC50 mutein PLN		86,095	484,676	291,099	273,497	35,181	777,328	280,957	128,451	10000					
IC50 wt PLN	21,80	21,085	22,796	22,201	21,734	21,523	23,139	21,397	21,148	21,148					
ratio TLN:PLN	**0,57**	**0,45**	**0,30**	**0,92**	**0,34**	**0,75**	**0,43**	**0,75**	**4,45**	**0,00**					
		Pos37													
	wt (avg)	NnC	NnD	NnE	NnG	NnH	NnQ	NnS	NnY	NnA	NnM	NnP	NnV	NnW	NnL
IC50 mutein TLN		307,492	1379,887	183,989	590,669	198,251	10000	488,91	179,264	5000	428,764	749,35	227,708	250,085	97,854
IC50 wt TLN	12,37	12,674	12,136	12,644	12,649	12,78	12,553	12,499	12,546	12,553	11,678	11,982	12,679	12,674	11,097
IC50 mutein PLN		332,781	45,714	5000	68,213	174,993	310,997	176,936	296,224	5000	136,086	137,03	234,271	125,939	137,03
IC50 wt PLN	22,17	20,731	22,607	21,945	21,827	21,18	22,224	21,472	21,501	21,945	30,792	20,761	21,194	21,464	20,761
ratio TLN:PLN	**0,56**	**0,92**	**30,19**	**0,04**	**8,66**	**1,13**	**32,15**	**2,76**	**0,61**	**1,00**	**3,15**	**5,47**	**0,97**	**1,99**	**0,71**
		Pos36													
	wt (avg)	InA	InF	InG	InK	InR	InV	InY	InQ	InD	InE	InH	InM	InT	InW
IC50 mutein TLN		13,887	11,627	10000	5,656	3,816	11,119	10,893	10,371	10,599	43,488	5,344	4,795	10,841	10,599
IC50 wt TLN	12,51	12,408	12,555	12,553	12,545	12,581	12,551	12,553	12,552	12,53	12,526	12,379	12,415	12,432	12,53
IC50 mutein PLN		74,4	105,107	28,916	45,25	16,069	22,275	34,934	15,404	47,007	5000	71,648	11,097	103,341	62,954
IC50 wt PLN	21,91	22,009	22,338	21,56	22,028	22,261	22,087	22,167	21,925	20,857	21,56	22,032	21,961	21,923	22,076
ratio TLN:PLN	**0,57**	**0,19**	**0,11**	**345,83**	**0,12**	**0,24**	**0,50**	**0,31**	**0,67**	**0,23**	**0,01**	**0,07**	**0,43**	**0,10**	**0,17**
		Pos35													
	wt (avg)	InA	InF	InL	InV	InM	InW	InY							
IC50 mutein TLN		34,34	13,26	9,06	14,93	12,55	14,10	16,25							
IC50 wt TLN	12,57	12,58	12,58	12,57	12,60	12,55	12,58	12,55							
IC50 mutein PLN		259,77	16,27	8,26	63,55	8,22	13,92	12,43							
IC50 wt PLN	21,98	21,81	21,95	21,91	22,19	21,94	22,12	21,95							
ratio TLN:PLN	**0,57**	**0,13**	**0,81**	**1,10**	**0,23**	**1,53**	**1,01**	**1,31**							

Table 1

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THERMOLYSINE					Pos 35		Pos 36		Pos 37		Pos 38		Pos 39	
					P3		P2		P1		P1"		P2"	
Amino acid	3-Letter	1- Letter	Side-chain polarity	Thresh.	IC_{50}	Std- Error	IC_{50}	Std- Error	IC_{50}	Std- Error	IC_{50}	Std- Error	IC_{50}	Std- Error
Alanine	Ala	A	nonploar	12,60	34,90	2,35	16,10	3,36	NM		320,00		18,00	1,37
Proline	Pro	P	nonpolar	12,60	NM		NM		NM		NM		NM	
Valine	Val	V	nonpolar	12,60	15,00	1,79	11,10	0,81	NM		26,10	2,70	9,20	0,34
Leucine	Leu	L	nonpolar	12,60	8,84	1,01			NM		96,20	10,00	67,50	5,49
Isoleucine	Ile	I	nonpolar	12,60	**12,60**	1,23	**12,60**	1,23	NM		**12,60**	1,23	82,80	15,50
Metheonin	Met	M	nonpolar	12,60	12,50	0,90	4,47	0,27	NM		285,00		24,80	0,96
Phenylalanin	Phe	F	nonpolar	12,60	12,30	1,34	11,90	1,12			34,10	2,47	248,00	53,10
Tryptophan	Trp	W	nonpolar	12,60	14,00	1,46	10,60	0,79	NM		NM		74,00	20,20
Tyrosine	Tyr	Y	polar	12,60	16,40	1,11	10,80	0,69	NM		NM		141,00	36,20
Glycine	Gly	G	nonpolar	12,60			1100,00		900,00		NM		314,00	110,00
Serine	Ser	S	polar	12,60	24,50	4,35	2,70	0,36	700,00		NM		14,30	1,35
Cysteine	Cys	C	nonpolar	12,60	11,00	0,81	400,00		NM		300,00		NM	
Threonine	Thr	T	polar	12,60	36,50	5,83	11,00	0,97			NM		26,80	1,41
Asparagine	Asn	N	polar	12,60	6,64	0,45	10,60	0,91	**12,60**	1,23	NM		65,90	8,68
Glutamine	Gln	Q	polar	12,60	9,24	0,81	10,50	0,98	NM		NM		57,00	7,22
Histidine	His	H	basic polar	12,60	10,70	1,90	4,97	0,33	181,00	75,10	NM		10,50	0,55
Lysine	Lys	K	basic polar	12,60	11,60	1,38	5,63	0,34			NM		7,77	0,22
Arginine	Arg	R	basic polar	12,60	8,76	0,81	3,30	0,23	NM		NM		**12,60**	1,23
Aspartic acid	Asp	D	acidic polar	12,60	40,00	6,24	600,00		NM		NM		54,70	6,49
Glutamic acid	Glu	E	acidic polar	12,60	30,40	2,45	41,50	3,75	NM		NM		460,00	

Table 2

NM = not measurable

Grey: Higher IC_{50} than wildtype (wildtype in bold and grey)

EXAMPLE 1

Total RNA isolation and cDNA synthesis

Last instar larvae were used for immunization using the following solutions: ($4.05*10^4$ cfu/ml) *Escherichia coli* strain BL21 (DE3) (Invitrogen, Carlsbad,CA) and ($0.7*10^4$ cfu/ml) *Micrococcus luteus* (DMSZ Reference number 495). Ten microliters of sample volume from each solution was injected dorsolaterally into the hemocoel using 1-ml disposable syringes and 0.4-mm needles mounted on a microapplicator. Larvae were homogenized at 8 h postinjection for total RNA isolation using the TRIzol reagent (Invitrogen, Germany) according to manufacturer´s instructions. cDNA was synthesized using the OneStep RT-PCR System (Qiagen, Germany) according to the manufacturer´s instructions.

EXAMPLE 2

preparation of IMPIs (Expression and purification)

1. Construction of a heterologous expression system

The T7-based expression system, pET-41a(+), which allows protein expression upon induction of the T7-polymerase in *Escherichia coli* cells by IPTG, was chosen to obtain strong and reproducible expression of the recombinant protein. The sequence of the mature IMPIα was amplified using the forward primer (5´- GATAGTCCTAATTTGTAACGGTGGACAC-3´) and the reverse primer (5´- CTACGAACGTATTTTAGGACAGTCTTTTATCG-3´). The fragment was cloned into the *Psh*AI site of vector pET-41a(+), which was then designated p41IMPIα and transformed into *E.coli* Rosetta-gami 2 pLysS (Novagen, Germany). The resulting clones were verified via sequencing (Eurofins MWG Operon, Germany)

2. Expression and production of recombinant IMPIα

   i. Preparatory Culture: LB-medium containing 1% (w/v) Glucose, Cm_{34}, Kan_{50} und $Tc_{12,5}$ was inoculated with cell material from a colony and cultivation started on an orbital shaker at 32°C up to a OD_{600} of 1. The preparatory culture was subsequently stored at 4°C in a refrigerator.

   ii. Main culture: LB-Medium containg 1% (w/v) Glucose, Cm_{34}, Kan_{50}, was inoculated with 3 %(v/v) of the preparatory culture and cultivation started on an orbital shaker at 32°C up to a OD_{600} of 1. Upon reaching

an OD_{600} of 1 the expression of the recombinant protein was induced with 1 mM IPTG. The main culture was subsequently incubated for further 3 hours at 32°C on an orbital shaker.

iii. Cells were harvested from liquid culture by centrifugation at 10,000 g for 10 min using a centrifuge tube of known weight. The pellet was decanted and allowed to drain, removing a maximum amount of liquid before the wet weight of the pellet was determined.

iv. The cell pellet was resuspended in BugBuster Reagent (Novagen, Germany) at room temperature by pipeting and gentle vortexing. 5 ml reagent per g wet cell paste was used. 1 µl (25 units) Benzonase® Nuclease (Novagen, Germany) per 1 ml of BugBuster Reagent was added.

v. The suspension obtained in step(iv) of this protocol was incubated at room temperature on a shaking platform at a slow setting for 20 min. Care was taken that the extract was not viscous at the end of the incubation.

vi. Insoluble cell debris was removed by centrifugation at 16,000 × *g* for 20 min at 4°C.

vii. The supernatant was transferred to a fresh tube. The soluble extract was applied directly to GST•Bind™ Resin (Novagen, Germany).

viii. Affinity chromatography was performed according to manufacturer´s instructions (GE Healthcare Life Sciences.

3. Main culture: LB-Medium containg 1% (w/v) Glucose, Cm_{34}, Kan_{50} was inoculated with 3 %(v/v) of the preparatory culture and cultivation started on an orbital shaker at 32°C until an OD_{600} of 1 was reached. At 1 OD the expression of the recombinant protein was induced with 1 mM IPTG. The main culture was subsequently incubated for further 3 hours at 32°C on an orbital shaker.

EXAMPLE 3

Bioinformatics Analysis

Protein motifs were identified using SMART (http://smart.embl-heidelberg.de/) and the Conserved Domain Database from NCBI (http://ncbi:nlm.nih.gov/Structure/cdd/wrpsb.cgi). The signal peptide was pre-

dicted using SignalP (http://www.cbs.dtu.dk/services /SignalP/), and the theoretical isolelectric point and molecular weight were predicted using Compute pI/MW (http://expasy.org/tools/protparam.html). Cysteine disulfide bonding state and connectivity prediction was done by using DISULFIND (http://disulfind.dsi.unifi.it/). Sequence similarity was analysed by BLAST from NCBI (http://www.ncbi.nlm.nih.gov/BLAST/). Multi-sequence alignment was generated using Vector NTI 9.0 (Invitrogen, Germany). Homology modeling was performed by the CPHmodels 3.0 server (http://www.cbs.dtu.dk/services/CPHmodels/) using the Swiss-Model server (http://swissmodel.expasy.org/) for structure assessment. All models were energy minimized by using GROMOS force field (http://www.gromacs.org/Documentation/Terminology/Force_Fields/GROMOS).

The subsequent table lists the sequences printed in the ensuing sequenc protocol. The leading number denotes the SEQ ID Nr. for the nucleotide sequence, the subsequent even number mssing number would denote the SEQ ID of the respective peptide sequence.

1 IMPIalpha (wild type or wtIMPIalpha)
3 IMPIbeta (wild type or wt IMPIbeta)
5 GST/IMPIalpha
7 GST/IMPIbeta
9 : IMPIalpha Pos37 NnG Pos38 InL Pos39 RnA
11 : GST/IMPIalpha Pos37 NnG Pos38 InL Pos39 RnA
13 : IMPIalpha Pos52 RnK
15 : GST/IMPIalpha Pos52 RnK
17 : IMPI Pos35 InL
19 : IMPI Pos35 InM
21 : IMPI Pos35 InF
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83 : IMPI Pos35 InH
85 : GST/IMPI Pos35 InN
87 : GST/IMPI Pos36 InS
89 : GST/IMPI Pos39 InK
91 : GST/IMPI Pos35 InC

93 : IMPI signal peptide,
95 : IMPI like (Solenopsis, Peptide only, no nucleotide sequence provided)

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Met Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser Met Leu Glu
85 90 95
gga gcg gtt ttg gat att aga tac ggt gtt tcg aga att gca tat agt 336
Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser
100 105 110
aaa gac ttt gaa act ctc aaa gtt gat ttt ctt agc aag cta cct gaa 384
Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu
115 120 125
atg ctg aaa atg ttc gaa gat cgt tta tgt cat aaa aca tat tta aat 432
Met Leu Lys Met Phe Glu Asp Arg Leu Cys His Lys Thr Tyr Leu Asn

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    130                 135                 140
ggt gat cat gta acc cat cct gac ttc atg ttg tat gac gct ctt gat      480
Gly Asp His Val Thr His Pro Asp Phe Met Leu Tyr Asp Ala Leu Asp
145                 150                 155                 160
gtt gtt tta tac atg gac cca atg tgc ctg gat gcg ttc cca aaa tta      528
Val Val Leu Tyr Met Asp Pro Met Cys Leu Asp Ala Phe Pro Lys Leu
                165                 170                 175
gtt tgt ttt aaa aaa cgt att gaa gct atc cca caa att gat aag tac      576
Val Cys Phe Lys Lys Arg Ile Glu Ala Ile Pro Gln Ile Asp Lys Tyr
            180                 185                 190
ttg aaa tcc agc aag tat ata gca tgg cct ttg cag ggc tgg caa gcc      624
Leu Lys Ser Ser Lys Tyr Ile Ala Trp Pro Leu Gln Gly Trp Gln Ala
        195                 200                 205
acg ttt ggt ggt ggc gac cat cct cca aaa tcg gat ggt tca act agt      672
Thr Phe Gly Gly Gly Asp His Pro Pro Lys Ser Asp Gly Ser Thr Ser
    210                 215                 220
ggt tct ggt cat cac cat cac cat cac tcc gcg ggt ctg gtg cca cgc      720
Gly Ser Gly His His His His His His Ser Ala Gly Leu Val Pro Arg
225                 230                 235                 240
ggt agt act gca att ggt atg aaa gaa acc gct gct gct aaa ttc gaa      768
Gly Ser Thr Ala Ile Gly Met Lys Glu Thr Ala Ala Ala Lys Phe Glu
                245                 250                 255
cgc cag cac atg gac agc cca gat ctg ggt acc ggt ggt ggc tcc ggt      816
Arg Gln His Met Asp Ser Pro Asp Leu Gly Thr Gly Gly Gly Ser Gly
            260                 265                 270
gat gac gac gac aag ata gtc cta att tgt aac ggt gga cac gaa tac      864
Asp Asp Asp Asp Lys Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr
        275                 280                 285
tac gag tgc ggt gga gcc tgc gat aat gta tgt gca gat tta cat ata      912
Tyr Glu Cys Gly Gly Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile
    290                 295                 300
cag aat aaa aca aac tgt ccc atc att ggt ctg gct tgt aat gac aag      960
Gln Asn Lys Thr Asn Cys Pro Ile Ile Gly Leu Ala Cys Asn Asp Lys
305                 310                 315                 320
tgc tac tgt gaa gat ggc tat gca agg gat gtc aat ggc aaa tgt ata     1008
Cys Tyr Cys Glu Asp Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile
                325                 330                 335
ccg ata aaa gac tgt cct aaa ata cgt tcg tag                        1041
Pro Ile Lys Asp Cys Pro Lys Ile Arg Ser
            340                 345
<210>  12
<211>  346
<212>  PRT
<213>  Galleria mellonella
<400>  12
Met Ser Pro Ile Leu Gly Tyr Trp Lys Ile Lys Gly Leu Val Gln Pro
1               5                   10                  15
Thr Arg Leu Leu Leu Glu Tyr Leu Glu Glu Lys Tyr Glu Glu His Leu
            20                  25                  30
Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu
        35                  40                  45
Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys
    50                  55                  60
Leu Thr Gln Ser Met Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn
65                  70                  75                  80
Met Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser Met Leu Glu
                85                  90                  95
Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser
            100                 105                 110
Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu
        115                 120                 125
Met Leu Lys Met Phe Glu Asp Arg Leu Cys His Lys Thr Tyr Leu Asn
    130                 135                 140
Gly Asp His Val Thr His Pro Asp Phe Met Leu Tyr Asp Ala Leu Asp
145                 150                 155                 160
Val Val Leu Tyr Met Asp Pro Met Cys Leu Asp Ala Phe Pro Lys Leu
                165                 170                 175
Val Cys Phe Lys Lys Arg Ile Glu Ala Ile Pro Gln Ile Asp Lys Tyr
            180                 185                 190
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Leu Lys Ser Ser Lys Tyr Ile Ala Trp Pro Leu Gln Gly Trp Gln Ala
195 200 205
Thr Phe Gly Gly Gly Asp His Pro Pro Lys Ser Asp Gly Ser Thr Ser
210 215 220
Gly Ser Gly His His His His His His Ser Ala Gly Leu Val Pro Arg
225 230 235 240
Gly Ser Thr Ala Ile Gly Met Lys Glu Thr Ala Ala Ala Lys Phe Glu
245 250 255
Arg Gln His Met Asp Ser Pro Asp Leu Gly Thr Gly Gly Gly Ser Gly
260 265 270
Asp Asp Asp Asp Lys Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr
275 280 285
Tyr Glu Cys Gly Gly Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile
290 295 300
Gln Asn Lys Thr Asn Cys Pro Ile Ile Gly Leu Ala Cys Asn Asp Lys
305 310 315 320
Cys Tyr Cys Glu Asp Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile
325 330 335
Pro Ile Lys Asp Cys Pro Lys Ile Arg Ser
340 345

<210> 13
<211> 207
<212> DNA
<213> Galleria mellonella
<220>
<221> CDS
<222> (1)..(207)
<400> 13

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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc atc att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Ile Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca aag gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Lys Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65

<210> 14
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 14

Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Ile Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Lys Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65

<210> 15
<211> 1041
<212> DNA
<213> Galleria mellonella
<220>
<221> CDS
<222> (1)..(1041)
<400> 15

atg tcc cct ata cta ggt tat tgg aaa att aag ggc ctt gtg caa ccc 48
Met Ser Pro Ile Leu Gly Tyr Trp Lys Ile Lys Gly Leu Val Gln Pro
1 5 10 15

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act cga ctt ctt ttg gaa tat ctt gaa gaa aaa tat gaa gag cat ttg       96
Thr Arg Leu Leu Leu Glu Tyr Leu Glu Glu Lys Tyr Glu Glu His Leu
            20                  25                  30
tat gag cgc gat gaa ggt gat aaa tgg cga aac aaa aag ttt gaa ttg      144
Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu
        35                  40                  45
ggt ttg gag ttt ccc aat ctt cct tat tat att gat ggt gat gtt aaa      192
Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys
    50                  55                  60
tta aca cag tct atg gcc atc ata cgt tat ata gct gac aag cac aac      240
Leu Thr Gln Ser Met Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn
65                  70                  75                  80
atg ttg ggt ggt tgt cca aaa gag cgt gca gag att tca atg ctt gaa      288
Met Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser Met Leu Glu
                85                  90                  95
gga gcg gtt ttg gat att aga tac ggt gtt tcg aga att gca tat agt      336
Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser
            100                 105                 110
aaa gac ttt gaa act ctc aaa gtt gat ttt ctt agc aag cta cct gaa      384
Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu
        115                 120                 125
atg ctg aaa atg ttc gaa gat cgt tta tgt cat aaa aca tat tta aat      432
Met Leu Lys Met Phe Glu Asp Arg Leu Cys His Lys Thr Tyr Leu Asn
    130                 135                 140
ggt gat cat gta acc cat cct gac ttc atg ttg tat gac gct ctt gat      480
Gly Asp His Val Thr His Pro Asp Phe Met Leu Tyr Asp Ala Leu Asp
145                 150                 155                 160
gtt gtt tta tac atg gac cca atg tgc ctg gat gcg ttc cca aaa tta      528
Val Val Leu Tyr Met Asp Pro Met Cys Leu Asp Ala Phe Pro Lys Leu
                165                 170                 175
gtt tgt ttt aaa aaa cgt att gaa gct atc cca caa att gat aag tac      576
Val Cys Phe Lys Lys Arg Ile Glu Ala Ile Pro Gln Ile Asp Lys Tyr
            180                 185                 190
ttg aaa tcc agc aag tat ata gca tgg cct ttg cag ggc tgg caa gcc      624
Leu Lys Ser Ser Lys Tyr Ile Ala Trp Pro Leu Gln Gly Trp Gln Ala
        195                 200                 205
acg ttt ggt ggt ggc gac cat cct cca aaa tcg gat ggt tca act agt      672
Thr Phe Gly Gly Gly Asp His Pro Pro Lys Ser Asp Gly Ser Thr Ser
    210                 215                 220
ggt tct ggt cat cac cat cac cat cac tcc gcg ggt ctg gtg cca cgc      720
Gly Ser Gly His His His His His His Ser Ala Gly Leu Val Pro Arg
225                 230                 235                 240
ggt agt act gca att ggt atg aaa gaa acc gct gct gct aaa ttc gaa      768
Gly Ser Thr Ala Ile Gly Met Lys Glu Thr Ala Ala Ala Lys Phe Glu
                245                 250                 255
cgc cag cac atg gac agc cca gat ctg ggt acc ggt ggt ggc tcc ggt      816
Arg Gln His Met Asp Ser Pro Asp Leu Gly Thr Gly Gly Gly Ser Gly
            260                 265                 270
gat gac gac gac aag ata gtc cta att tgt aac ggt gga cac gaa tac      864
Asp Asp Asp Asp Lys Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr
        275                 280                 285
tac gag tgc ggt gga gcc tgc gat aat gta tgt gca gat tta cat ata      912
Tyr Glu Cys Gly Gly Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile
    290                 295                 300
cag aat aaa aca aac tgt ccc atc att aat ata aga tgt aat gac aag      960
Gln Asn Lys Thr Asn Cys Pro Ile Ile Asn Ile Arg Cys Asn Asp Lys
305                 310                 315                 320
tgc tac tgt gaa gat ggc tat gca aag gat gtc aat ggc aaa tgt ata     1008
Cys Tyr Cys Glu Asp Gly Tyr Ala Lys Asp Val Asn Gly Lys Cys Ile
                325                 330                 335
ccg ata aaa gac tgt cct aaa ata cgt tcg tag                        1041
Pro Ile Lys Asp Cys Pro Lys Ile Arg Ser
            340                 345
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<210> 16
<211> 346
<212> PRT
<213> Galleria mellonella
<400> 16

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Met Ser Pro Ile Leu Gly Tyr Trp Lys Ile Lys Gly Leu Val Gln Pro
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1 5 10 15
Thr Arg Leu Leu Leu Glu Tyr Leu Glu Glu Lys Tyr Glu Glu His Leu
20 25 30
Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu
35 40 45
Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys
50 55 60
Leu Thr Gln Ser Met Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn
65 70 75 80
Met Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser Met Leu Glu
85 90 95
Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser
100 105 110
Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu
115 120 125
Met Leu Lys Met Phe Glu Asp Arg Leu Cys His Lys Thr Tyr Leu Asn
130 135 140
Gly Asp His Val Thr His Pro Asp Phe Met Leu Tyr Asp Ala Leu Asp
145 150 155 160
Val Val Leu Tyr Met Asp Pro Met Cys Leu Asp Ala Phe Pro Lys Leu
165 170 175
Val Cys Phe Lys Lys Arg Ile Glu Ala Ile Pro Gln Ile Asp Lys Tyr
180 185 190
Leu Lys Ser Ser Lys Tyr Ile Ala Trp Pro Leu Gln Gly Trp Gln Ala
195 200 205
Thr Phe Gly Gly Gly Asp His Pro Pro Lys Ser Asp Gly Ser Thr Ser
210 215 220
Gly Ser Gly His His His His His His Ser Ala Gly Leu Val Pro Arg
225 230 235 240
Gly Ser Thr Ala Ile Gly Met Lys Glu Thr Ala Ala Ala Lys Phe Glu
245 250 255
Arg Gln His Met Asp Ser Pro Asp Leu Gly Thr Gly Gly Gly Ser Gly
260 265 270
Asp Asp Asp Asp Lys Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr
275 280 285
Tyr Glu Cys Gly Gly Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile
290 295 300
Gln Asn Lys Thr Asn Cys Pro Ile Ile Asn Ile Arg Cys Asn Asp Lys
305 310 315 320
Cys Tyr Cys Glu Asp Gly Tyr Ala Lys Asp Val Asn Gly Lys Cys Ile
325 330 335
Pro Ile Lys Asp Cys Pro Lys Ile Arg Ser
340 345

<210> 17
<211> 207
<212> DNA
<213> Galleria mellonella
<220>
<221> CDS
<222> (1)..(207)
<220>
<221> misc_feature
<222> (185)..(185)
<223> n is a, c, g, or t

<400> 17
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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc ttg att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Leu Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata ana gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Xaa Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser

65
<210> 18
<211> 69
<212> PRT
<213> Galleria mellonella
<220>
<221> misc_feature
<222> (62)..(62)
<223> The 'Xaa' at location 62 stands for Lys, Arg, Thr, or Ile.
<400> 18
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1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Leu Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Xaa Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65
<210> 19
<211> 207
<212> DNA
<213> Galleria mellonella
<220>
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<222> (1)..(207)
<400> 19
ata gtc cta att tgt aac ggt gga cac gaa tac tac gag tgc ggt gga 48
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc atg att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Met Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65
<210> 20
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 20
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Met Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
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<210> 21
<211> 207
<212> DNA
<213> Galleria mellonella
<220>
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<222> (1)..(207)
<400> 21
ata gtc cta att tgt aac ggt gga cac gaa tac tac gag tgc ggt gga 48
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15

gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc ttc att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Phe Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65

<210> 22
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 22
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Phe Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65

<210> 23
<211> 207
<212> DNA
<213> Galleria mellonella
<220>
<221> CDS
<222> (1)..(207)
<400> 23
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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc tgt att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Cys Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65

<210> 24
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 24
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Cys Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65

<210> 25
<211> 207
<212> DNA
<213> Galleria mellonella

<220>
<221> CDS
<222> (1)..(207)
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1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc aat att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Asn Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65
<210> 26
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 26
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Asn Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65
<210> 27
<211> 207
<212> DNA
<213> Galleria mellonella
<220>
<221> CDS
<222> (1)..(207)
<400> 27
ata gtc cta att tgt aac ggt gga cac gaa tac tac gag tgc ggt gga 48
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc cag att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Gln Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65
<210> 28
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 28
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Gln Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys

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    50                  55                  60
Pro Lys Ile Arg Ser
65
<210>  29
<211>  207
<212>  DNA
<213>  Galleria mellonella
<220>
<221>  CDS
<222>  (1)..(207)
<400>  29
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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1               5                   10                  15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac       96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
            20                  25                  30
tgt ccc cac att aat ata aga tgt aat gac aag tgc tac tgt gaa gat      144
Cys Pro His Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
        35                  40                  45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt      192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
    50                  55                  60
cct aaa ata cgt tcg                                                  207
Pro Lys Ile Arg Ser
65
<210>  30
<211>  69
<212>  PRT
<213>  Galleria mellonella
<400>  30
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1               5                   10                  15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
            20                  25                  30
Cys Pro His Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
        35                  40                  45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
    50                  55                  60
Pro Lys Ile Arg Ser
65
<210>  31
<211>  207
<212>  DNA
<213>  Galleria mellonella
<220>
<221>  CDS
<222>  (1)..(207)
<400>  31
ata gtc cta att tgt aac ggt gga cac gaa tac tac gag tgc ggt gga       48
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1               5                   10                  15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac       96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
            20                  25                  30
tgt ccc aag att aat ata aga tgt aat gac aag tgc tac tgt gaa gat      144
Cys Pro Lys Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
        35                  40                  45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt      192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
    50                  55                  60
cct aaa ata cgt tcg                                                  207
Pro Lys Ile Arg Ser
65
<210>  32
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<213>  Galleria mellonella
<400>  32
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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Lys Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65

<210> 33
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<400> 33

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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc cgt att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Arg Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65

<210> 34
<211> 69
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<213> Galleria mellonella
<400> 34

Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Arg Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
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<400> 35

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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc atc gtt aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Ile Val Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207

Pro Lys Ile Arg Ser
65

<210> 36
<211> 69
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<213> Galleria mellonella
<400> 36

Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Ile Val Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65

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<222> (1)..(207)
<400> 37

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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc atc atg aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Ile Met Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65

<210> 38
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 38

Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Ile Met Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
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<210> 39
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<222> (1)..(207)
<400> 39

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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30

tgt ccc atc ttc aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Ile Phe Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65

<210> 40
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 40
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Ile Phe Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65

<210> 41
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<222> (7)..(7)
<223> n is a, c, g, or t
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<222> (17)..(17)
<223> n is a, c, g, or t
<220>
<221> misc_feature
<222> (33)..(33)
<223> n is a, c, g, or t
<220>
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<222> (35)..(36)
<223> n is a, c, g, or t
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<223> n is a, c, g, or t
<220>
<221> misc_feature
<222> (186)..(186)
<223> n is a, c, g, or t
<220>
<221> misc_feature
<222> (188)..(189)
<223> n is a, c, g, or t
<400> 41
ata gtc nta att tgt anc ggt gga cac gaa tan tnn gng tgc ggt gga 48
Ile Val Xaa Ile Cys Xaa Gly Gly His Glu Xaa Xaa Xaa Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aan naa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Xaa Xaa Thr Asn

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            20                  25                  30
tgt ccc atc tgg aat ata aga tgt aat gac aag tgc tac tgt gaa gat      144
Cys Pro Ile Trp Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
        35                  40                  45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg att aan gnn tgt      192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Xaa Xaa Cys
    50                  55                  60
cct aaa ata cgt tcg                                                  207
Pro Lys Ile Arg Ser
65
<210>  42
<211>  69
<212>  PRT
<213>  Galleria mellonella
<220>
<221>  misc_feature
<222>  (3)..(3)
<223>  The 'Xaa' at location 3 stands for Ile, Val, or Leu.
<220>
<221>  misc_feature
<222>  (6)..(6)
<223>  The 'Xaa' at location 6 stands for Asn, Ser, Thr, or Ile.
<220>
<221>  misc_feature
<222>  (11)..(11)
<223>  The 'Xaa' at location 11 stands for Tyr.
<220>
<221>  misc_feature
<222>  (12)..(12)
<223>  The 'Xaa' at location 12 stands for Tyr, Trp, Cys, Ser, Leu, or
       Phe.
<220>
<221>  misc_feature
<222>  (13)..(13)
<223>  The 'Xaa' at location 13 stands for Glu, Gly, Ala, or Val.
<220>
<221>  misc_feature
<222>  (29)..(29)
<223>  The 'Xaa' at location 29 stands for Lys, or Asn.
<220>
<221>  misc_feature
<222>  (30)..(30)
<223>  The 'Xaa' at location 30 stands for Lys, Glu, or Gln.
<220>
<221>  misc_feature
<222>  (62)..(62)
<223>  The 'Xaa' at location 62 stands for Lys, or Asn.
<220>
<221>  misc_feature
<222>  (63)..(63)
<223>  The 'Xaa' at location 63 stands for Glu, Asp, Gly, Ala, or Val.
<400>  42
Ile Val Xaa Ile Cys Xaa Gly Gly His Glu Xaa Xaa Xaa Cys Gly Gly
1               5                  10                  15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Xaa Xaa Thr Asn
            20                  25                  30
Cys Pro Ile Trp Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
        35                  40                  45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Xaa Xaa Cys
    50                  55                  60
Pro Lys Ile Arg Ser
65
<210>  43
<211>  207
<212>  DNA
<213>  Galleria mellonella
<220>
<221>  CDS
<222>  (1)..(207)
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<400> 43
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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc atc tac aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Ile Tyr Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65

<210> 44
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 44
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Ile Tyr Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65

<210> 45
<211> 207
<212> DNA
<213> Galleria mellonella
<220>
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<222> (1)..(207)
<400> 45
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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc atc agt aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Ile Ser Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65

<210> 46
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 46
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Ile Ser Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65

<210> 47
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<212> DNA
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<220>
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<222> (1)..(207)
<400> 47
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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc atc act aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Ile Thr Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65

<210> 48
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 48
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Ile Thr Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65

<210> 49
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<212> DNA
<213> Galleria mellonella
<220>
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<222> (1)..(207)
<400> 49
ata gtc cta att tgt aac ggt gga cac gaa tac tac gag tgc ggt gga 48
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc atc aat aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Ile Asn Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65

<210> 50
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 50
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn

20 25 30
Cys Pro Ile Asn Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65

<210> 51
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<212> DNA
<213> Galleria mellonella
<220>
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<222> (1)..(207)
<400> 51

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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1               5                  10                  15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac       96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
            20                  25                  30
tgt ccc atc cag aat ata aga tgt aat gac aag tgc tac tgt gaa gat      144
Cys Pro Ile Gln Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
        35                  40                  45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt      192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
    50                  55                  60
cct aaa ata cgt tcg                                              207
Pro Lys Ile Arg Ser
65
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<210> 52
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 52

Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Ile Gln Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65

<210> 53
<211> 207
<212> DNA
<213> Galleria mellonella
<220>
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<222> (1)..(207)
<400> 53

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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1               5                  10                  15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac       96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
            20                  25                  30
tgt ccc atc cat aat ata aga tgt aat gac aag tgc tac tgt gaa gat      144
Cys Pro Ile His Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
        35                  40                  45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt      192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
    50                  55                  60
cct aaa ata cgt tcg                                              207
Pro Lys Ile Arg Ser
65
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<210> 54

<211> 69
<212> PRT
<213> Galleria mellonella
<400> 54
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Ile His Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65
<210> 55
<211> 207
<212> DNA
<213> Galleria mellonella
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<222> (1)..(207)
<400> 55
ata gtc cta att tgt aac ggt gga cac gaa tac tac gag tgc ggt gga 48
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc atc cgt aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Ile Arg Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65
<210> 56
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 56
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Ile Arg Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65
<210> 57
<211> 207
<212> DNA
<213> Galleria mellonella
<220>
<221> CDS
<222> (1)..(207)
<400> 57
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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc atc aag aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Ile Lys Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45

ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65

<210> 58
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 58
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Ile Lys Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65

<210> 59
<211> 207
<212> DNA
<213> Galleria mellonella
<220>
<221> CDS
<222> (1)..(207)
<400> 59
ata gtc cta att tgt aac ggt gga cac gaa tac tac gag tgc ggt gga 48
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc atc att aat ata gta tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Ile Ile Asn Ile Val Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65

<210> 60
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 60
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Ile Ile Asn Ile Val Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65

<210> 61
<211> 207
<212> DNA
<213> Galleria mellonella
<220>
<221> CDS
<222> (1)..(207)
<400> 61
ata gtc cta att tgt aac ggt gga cac gaa tac tac gag tgc ggt gga 48
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly

1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc atc att aat ata aaa tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Ile Ile Asn Ile Lys Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65

<210> 62
<211> 69
<212> PRT
<213> Galleria mellonella
<400> 62
Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Ile Ile Asn Ile Lys Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
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1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc tgg att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Trp Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
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20 25 30
Cys Pro Trp Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
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1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc tac att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Tyr Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
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20 25 30
Cys Pro Tyr Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
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1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc atc att aat ata gca tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Ile Ile Asn Ile Ala Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
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20 25 30
Cys Pro Ile Ile Asn Ile Ala Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45

Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
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1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc tgt att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Cys Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
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20 25 30
Cys Pro Cys Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc aag att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Lys Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65

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<212> PRT
<213> Galleria mellonella

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Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Lys Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc cgt att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Arg Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
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<210> 74
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<213> Galleria mellonella
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Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Arg Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
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1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc ttg att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Leu Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60

cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65
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<211> 69
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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
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Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Leu Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc atg att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Met Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
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20 25 30
Cys Pro Met Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn

20 25 30
tgt ccc ttc att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Phe Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65

<210> 80
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<213> Galleria mellonella
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20 25 30
Cys Pro Phe Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65

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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc cag att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro Gln Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65

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<213> Galleria mellonella
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Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro Gln Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
65

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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
1 5 10 15
gcc tgc gat aat gta tgt gca gat tta cat ata cag aat aaa aca aac 96
Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
tgt ccc cac att aat ata aga tgt aat gac aag tgc tac tgt gaa gat 144
Cys Pro His Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
ggc tat gca agg gat gtc aat ggc aaa tgt ata ccg ata aaa gac tgt 192
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
cct aaa ata cgt tcg 207
Pro Lys Ile Arg Ser
65
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<211> 69
<212> PRT
<213> Galleria mellonella
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Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr Tyr Glu Cys Gly Gly
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Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile Gln Asn Lys Thr Asn
20 25 30
Cys Pro His Ile Asn Ile Arg Cys Asn Asp Lys Cys Tyr Cys Glu Asp
35 40 45
Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile Pro Ile Lys Asp Cys
50 55 60
Pro Lys Ile Arg Ser
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1 5 10 15
act cga ctt ctt ttg gaa tat ctt gaa gaa aaa tat gaa gag cat ttg 96
Thr Arg Leu Leu Leu Glu Tyr Leu Glu Glu Lys Tyr Glu Glu His Leu
20 25 30
tat gag cgc gat gaa ggt gat aaa tgg cga aac aaa aag ttt gaa ttg 144
Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu
35 40 45
ggt ttg gag ttt ccc aat ctt cct tat tat att gat ggt gat gtt aaa 192
Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys
50 55 60
tta aca cag tct atg gcc atc ata cgt tat ata gct gac aag cac aac 240
Leu Thr Gln Ser Met Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn
65 70 75 80
atg ttg ggt ggt tgt cca aaa gag cgt gca gag att tca atg ctt gaa 288
Met Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser Met Leu Glu
85 90 95
gga gcg gtt ttg gat att aga tac ggt gtt tcg aga att gca tat agt 336
Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser
100 105 110
aaa gac ttt gaa act ctc aaa gtt gat ttt ctt agc aag cta cct gaa 384
Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu
115 120 125
atg ctg aaa atg ttc gaa gat cgt tta tgt cat aaa aca tat tta aat 432
Met Leu Lys Met Phe Glu Asp Arg Leu Cys His Lys Thr Tyr Leu Asn
130 135 140
ggt gat cat gta acc cat cct gac ttc atg ttg tat gac gct ctt gat 480
Gly Asp His Val Thr His Pro Asp Phe Met Leu Tyr Asp Ala Leu Asp

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145                 150                 155                 160
gtt gtt tta tac atg gac cca atg tgc ctg gat gcg ttc cca aaa tta      528
Val Val Leu Tyr Met Asp Pro Met Cys Leu Asp Ala Phe Pro Lys Leu
                165                 170                 175
gtt tgt ttt aaa aaa cgt att gaa gct atc cca caa att gat aag tac      576
Val Cys Phe Lys Lys Arg Ile Glu Ala Ile Pro Gln Ile Asp Lys Tyr
            180                 185                 190
ttg aaa tcc agc aag tat ata gca tgg cct ttg cag ggc tgg caa gcc      624
Leu Lys Ser Ser Lys Tyr Ile Ala Trp Pro Leu Gln Gly Trp Gln Ala
        195                 200                 205
acg ttt ggt ggt ggc gac cat cct cca aaa tcg gat ggt tca act agt      672
Thr Phe Gly Gly Gly Asp His Pro Pro Lys Ser Asp Gly Ser Thr Ser
    210                 215                 220
ggt tct ggt cat cac cat cac cat cac tcc gcg ggt ctg gtg cca cgc      720
Gly Ser Gly His His His His His His Ser Ala Gly Leu Val Pro Arg
225                 230                 235                 240
ggt agt act gca att ggt atg aaa gaa acc gct gct gct aaa ttc gaa      768
Gly Ser Thr Ala Ile Gly Met Lys Glu Thr Ala Ala Ala Lys Phe Glu
                245                 250                 255
cgc cag cac atg gac agc cca gat ctg ggt acc ggt ggt ggc tcc ggt      816
Arg Gln His Met Asp Ser Pro Asp Leu Gly Thr Gly Gly Gly Ser Gly
            260                 265                 270
gat gac gac gac aag ata gtc cta att tgt aac ggt gga cac gaa tac      864
Asp Asp Asp Asp Lys Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr
        275                 280                 285
tac gag tgc ggt gga gcc tgc gat aat gta tgt gca gat tta cat ata      912
Tyr Glu Cys Gly Gly Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile
    290                 295                 300
cag aat aaa aca aac tgt ccc aat att aat ata aga tgt aat gac aag      960
Gln Asn Lys Thr Asn Cys Pro Asn Ile Asn Ile Arg Cys Asn Asp Lys
305                 310                 315                 320
tgc tac tgt gaa gat ggc tat gca agg gat gtc aat ggc aaa tgt ata     1008
Cys Tyr Cys Glu Asp Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile
                325                 330                 335
ccg ata aaa gac tgt cct aaa ata cgt tcg tag                        1041
Pro Ile Lys Asp Cys Pro Lys Ile Arg Ser
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Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu
        35                  40                  45
Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys
    50                  55                  60
Leu Thr Gln Ser Met Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn
65                  70                  75                  80
Met Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser Met Leu Glu
                85                  90                  95
Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser
            100                 105                 110
Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu
        115                 120                 125
Met Leu Lys Met Phe Glu Asp Arg Leu Cys His Lys Thr Tyr Leu Asn
    130                 135                 140
Gly Asp His Val Thr His Pro Asp Phe Met Leu Tyr Asp Ala Leu Asp
145                 150                 155                 160
Val Val Leu Tyr Met Asp Pro Met Cys Leu Asp Ala Phe Pro Lys Leu
                165                 170                 175
Val Cys Phe Lys Lys Arg Ile Glu Ala Ile Pro Gln Ile Asp Lys Tyr
            180                 185                 190
Leu Lys Ser Ser Lys Tyr Ile Ala Trp Pro Leu Gln Gly Trp Gln Ala
        195                 200                 205
Thr Phe Gly Gly Gly Asp His Pro Pro Lys Ser Asp Gly Ser Thr Ser
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210 215 220
Gly Ser Gly His His His His His His Ser Ala Gly Leu Val Pro Arg
225 230 235 240
Gly Ser Thr Ala Ile Gly Met Lys Glu Thr Ala Ala Ala Lys Phe Glu
245 250 255
Arg Gln His Met Asp Ser Pro Asp Leu Gly Thr Gly Gly Gly Ser Gly
260 265 270
Asp Asp Asp Asp Lys Ile Val Leu Ile Cys Asn Gly Gly His Glu Tyr
275 280 285
Tyr Glu Cys Gly Gly Ala Cys Asp Asn Val Cys Ala Asp Leu His Ile
290 295 300
Gln Asn Lys Thr Asn Cys Pro Asn Ile Asn Ile Arg Cys Asn Asp Lys
305 310 315 320
Cys Tyr Cys Glu Asp Gly Tyr Ala Arg Asp Val Asn Gly Lys Cys Ile
325 330 335
Pro Ile Lys Asp Cys Pro Lys Ile Arg Ser
340 345

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<213> Galleria mellonella
<220>
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<400> 87

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1               5                  10                  15
act cga ctt ctt ttg gaa tat ctt gaa gaa aaa tat gaa gag cat ttg       96
Thr Arg Leu Leu Leu Glu Tyr Leu Glu Glu Lys Tyr Glu Glu His Leu
            20                  25                  30
tat gag cgc gat gaa ggt gat aaa tgg cga aac aaa aag ttt gaa ttg      144
Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu
        35                  40                  45
ggt ttg gag ttt ccc aat ctt cct tat tat att gat ggt gat gtt aaa      192
Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys
    50                  55                  60
tta aca cag tct atg gcc atc ata cgt tat ata gct gac aag cac aac      240
Leu Thr Gln Ser Met Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn
65                  70                  75                  80
atg ttg ggt ggt tgt cca aaa gag cgt gca gag att tca atg ctt gaa      288
Met Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser Met Leu Glu
                85                  90                  95
gga gcg gtt ttg gat att aga tac ggt gtt tcg aga att gca tat agt      336
Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser
            100                 105                 110
aaa gac ttt gaa act ctc aaa gtt gat ttt ctt agc aag cta cct gaa      384
Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu
        115                 120                 125
atg ctg aaa atg ttc gaa gat cgt tta tgt cat aaa aca tat tta aat      432
Met Leu Lys Met Phe Glu Asp Arg Leu Cys His Lys Thr Tyr Leu Asn
    130                 135                 140
ggt gat cat gta acc cat cct gac ttc atg ttg tat gac gct ctt gat      480
Gly Asp His Val Thr His Pro Asp Phe Met Leu Tyr Asp Ala Leu Asp
145                 150                 155                 160
gtt gtt tta tac atg gac cca atg tgc ctg gat gcg ttc cca aaa tta      528
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Claims

1. A polypeptide having at least 70% homology, in particular 80%, 90% or 95% homology to the polypeptide of SEQ ID No 2 representing the wild-type of the protein insect metalloproteinase inhibitor IMPIα and having at least one mutation at position 35, 36 and/or 39 of the amino acid sequence of IMPIα and the polypeptide having an IC_{50} value to thermolysine of less than the IC_{50} value of IMPIα wherein
   - the nonpolar amino acid isoleucine at position 35 of IMPIα is replaced either by a nonpolar amino acid selected from the group consisting of leucine, methionine and phenylalanine or by polar amino acid selected from the group consisting of cysteine, asparagine, glutamine, histidine, lysine and arginine; and/or
   - the nonpolar amino acid isoleucine at position 36 of IMPIα is replaced either by a nonpolar amino acid selected from the group consisting of valine, phenylalanine and tryptophan or by polar amino acid selected from the group consisting of tyrosine, serine, threonine, asparagine, glutamine, histidine, lysine and arginine; and/or
   - the polar amino acid position 39 of IMPIα is replaced either by the nonpolar amino acid valine or by the polar amino acids histidine or lysine.
2. The polypeptide of claim 1 having the amino acid sequences of SEQ ID numbers 10, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, 78, 80, 82, 84.
3. The polypeptide of claim 1 or 2having chemical modifications in the side chain or at the N and/or C terminal for improving biological or chemical properties such as bio availability, stability, effectivity or comprising a detectable label.
4. A fusion polypeptide comprising a wtIMPIα, wtIMPIβ and/or a polypeptide of at least one of the claims 1 to 3 fused with at least one polypeptide having a physiological function selected from the group consisting of an antibody or antibody fragment, scaffolds such as lipocalin, ankyrin, fibronectin, transferrin, tetranectin, adnectin, albumin, uteroglobin, or protein A; functional peptides such as transferrin; peptides useful for diagnostic applications, such as green fluorescent protein (GFP); or peptide tags

enabling immobilization on technical surfaces, such as hexahistidine; or a transferase such as glutathione-S-transferase (GST).

5. The fusion polypeptide of claim 4 being linked by a peptide of 1 to 100 amino acids in length.

6. The fusion polypeptide of claim 4 wherein a chemical linking group conjugates the polypeptide of at least one of the claims 1 to 3 and the at least one polypeptide having a physiological function.

7. A polynucleotide coding for the polypeptide of at least one of the claims 1 or 2 or the fusion polypeptide of claim 5 or 6.

8. The polynucleotide of claim 7 operably linked to the heterologous promoter.

9. A host cell comprising the recombinant nucleic acid of claim 7 or 8.

10. A method of producing a polypeptide comprising culturing the host cell of claim 9, expression of said nucleic acid in the host cell and isolation of the polypeptide.

11. A kit for diagnosis comprising a peptide of claim 3 and/or a fusion polypeptide of anyone of the claims 4 to 6 the polypeptide.

12. A pharmaceutical composition comprising the
polypeptide having at least 70% homology, in particular 80%, 90% or 95% homology to the polypeptide of SEQ ID No 2 representing the wild-type of the protein insect metalloproteinase inhibitor IMPIα, in particular having at least one mutation at position 35, 36 and/or 39 of the amino acid sequence of IMPIα wherein

- the nonpolar amino acid isoleucine at position 35 of IMPIα is replaced either by a nonpolar amino acid selected from the group consisting of leucine, methionine and phenylalanine or by polar amino acid selected from the group consisting of cysteine, asparagine, glutamine, histidine, lysine and arginine; and/or
- the nonpolar amino acid isoleucine at position 36 of IMPIα is replaced either by a nonpolar amino acid selected from the group consisting of valine, phenylalanine and tryptophan or by polar amino acid selected from the group consisting of tyrosine, serine, threonine, asparagine, glutamine, histidine, lysine and arginine; and/or the polar amino acid position 39 of IMPIα is replaced either by the nonpolar amino acid valine or by the polar amino acids histidine or lysine. and/or the fusion polypeptide of at least

one of the claims 4 to 6 or a polynucleotide comprising a section coding for at least one of the polypeptides of claim 1 to 6.

13. The polypeptide having at least 70% homology, in particular 80%, 90% or 95% homology to the polypeptide of SEQ ID No 2 representing the wild-type of the protein insect metalloproteinase inhibitor IMPIα and having at least one mutation at position 35, 36 and/or 39 of the amino acid sequence of IMPIα wherein
   - the nonpolar amino acid isoleucine at position 35 of IMPIα is replaced either by a nonpolar amino acid selected from the group consisting of leucine, methionine and phenylalanine or by polar amino acid selected from the group consisting of cysteine, asparagine, glutamine, histidine, lysine and arginine; and/or
   - the nonpolar amino acid isoleucine at position 36 of IMPIα is replaced either by a nonpolar amino acid selected from the group consisting of valine, phenylalanine and tryptophan or by polar amino acid selected from the group consisting of tyrosine, serine, threonine, asparagine, glutamine, histidine, lysine and arginine; and/or
   - the polar amino acid position 39 of IMPIα is replaced either by the nonpolar amino acid valine or by the polar amino acids histidine or lysine. or the fusion polypeptide of anyone of the claims 4 to 6 or the polynucleotide of claim 7 for use in the treatment of an animal or human infected by microorganisms secreting bacterial toxins of the M4 or Metzincin family of metalloproteinases, in particular thermolysine, aureolysin, bacillolysin, pseudolysin, vibriolysin or anthrax npr599.

14. Use of a polypeptide of claim 3 and/or a fusion polypeptide of anyone of the claims 4 to 6 for detection of the presence or activity of proteases belonging to the M4 family in a sample.

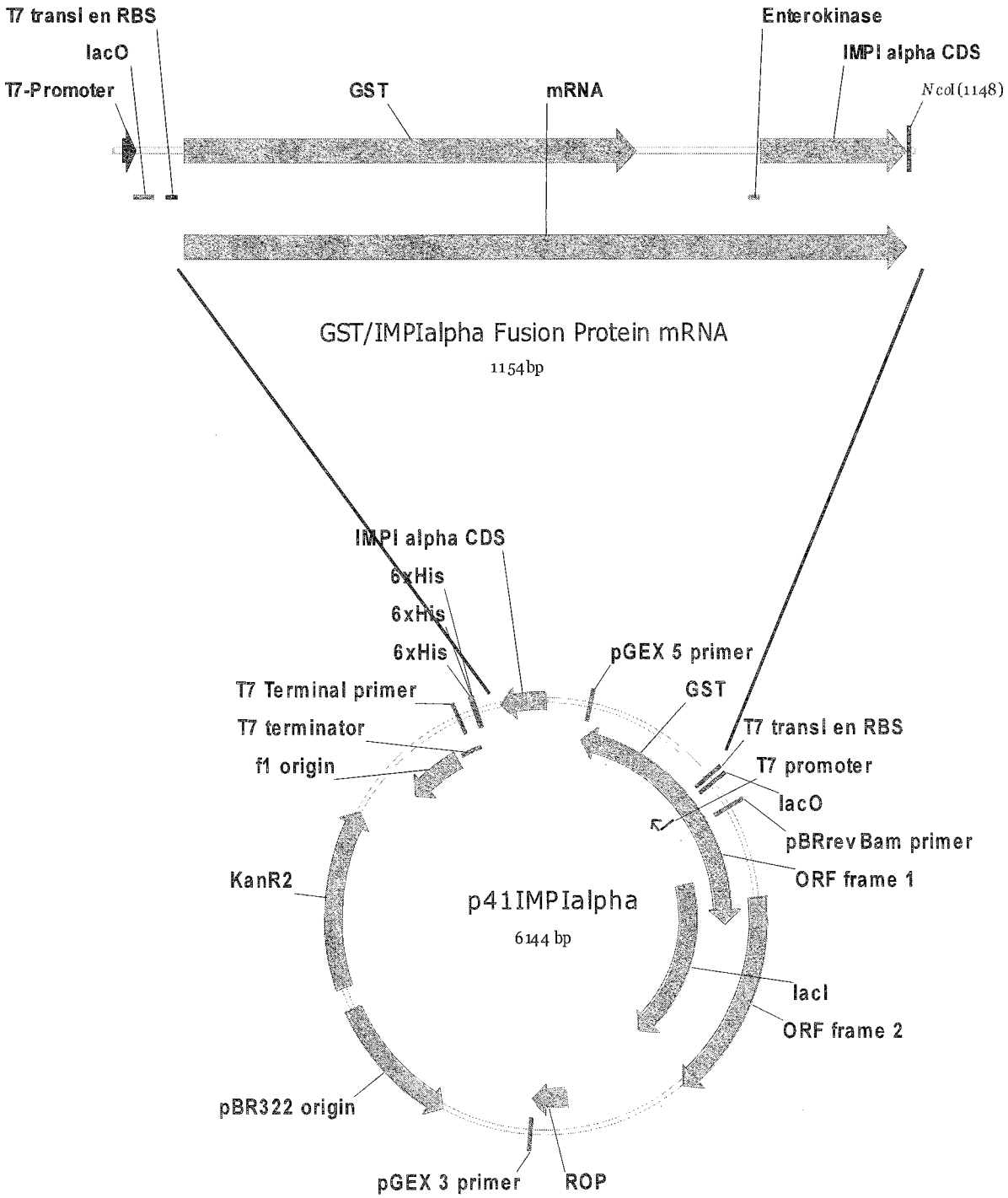
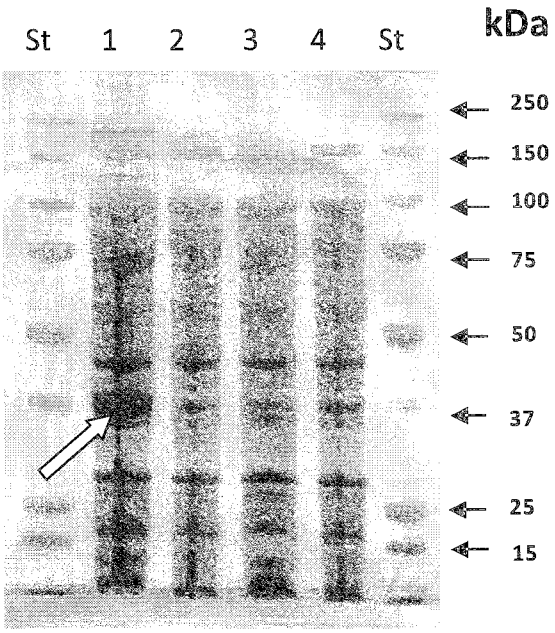
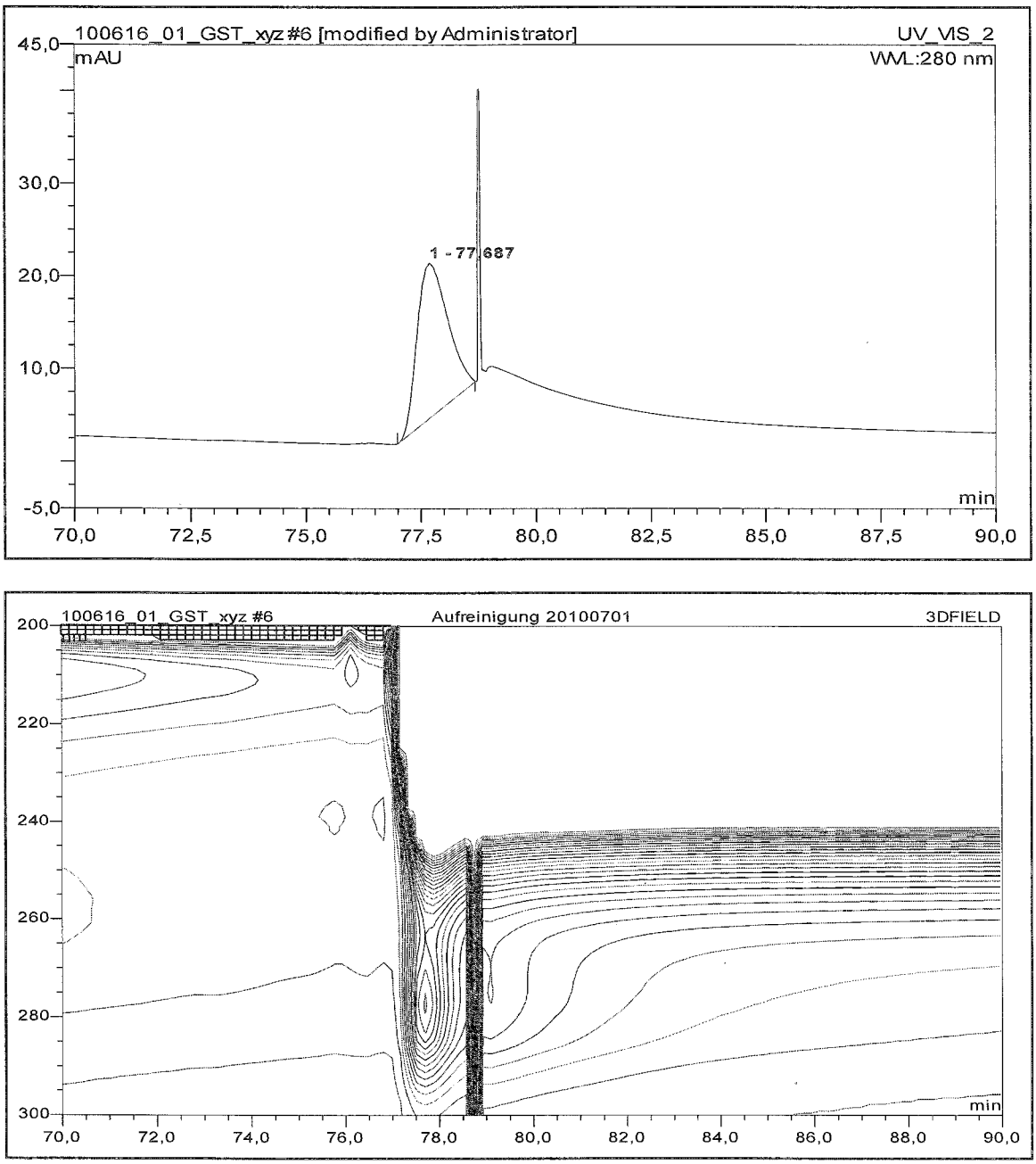

T7 transl en RBS
lacO
T7-Promoter
GST
mRNA
Enterokinase
IMPI alpha CDS
NcoI(1148)
GST/IMPIalpha Fusion Protein mRNA
1154bp
IMPI alpha CDS
6xHis
6xHis
6xHis
T7 Terminal primer
T7 terminator
f1 origin
pGEX 5 primer
GST
T7 transl en RBS
T7 promoter
lacO
pBRrev Bam primer
ORF frame 1
KanR2
p41IMPIalpha
6144 bp
lacI
ORF frame 2
pBR322 origin
pGEX 3 primer
ROP

Fig. 1



St: Precision Plus All Blue (Fa. Biorad)
1: E. coli Rosetta-gami 2 p41IMPialpha Soluble Cytoplasmatic Fraction (induced)
2: E. coli Rosetta-gami 2 p41IMPialpha Soluble Cytoplasmatic Fraction (uninduced)
3: E. coli Rosetta-gami 2 pET-41a(+) Soluble Cytoplasmatic Fraction (induced)
4: E. coli Rosetta-gami 2 pET-41a(+) Soluble Cytoplamatic Fraction (uninduced)

Fig. 2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WVL:280 nm
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20,0
10,0
-5,0
1 - 77,687
min
70,0
72,5
75,0
77,5
80,0
82,5
85,0
87,5
90,0
100616_01_GST_xyz #6
Aufreinigung 20100701
3DFIELD
nm
200
220
240
260
280
300
min
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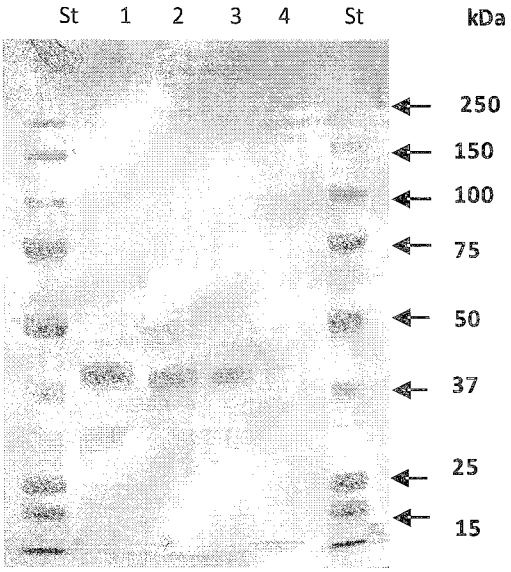
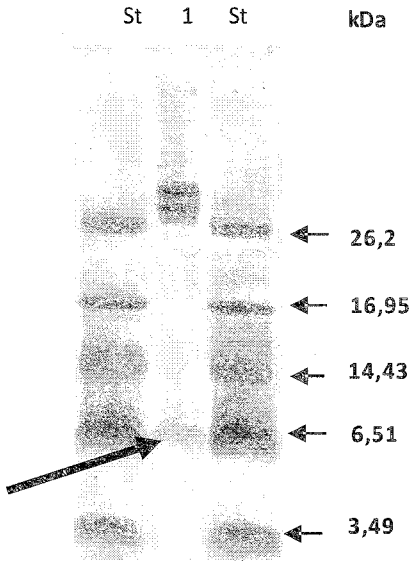
Fig. 3



St: Precision Plus All Blue (Fa. Biorad)
Lane 1-4: Pooled Fractions

Fig. 4a



St: Polypeptide SDS-PAGEStandard (Fa. Biorad)
Lane 1- Enterokinase digested fusion-protein

Fig. 4b

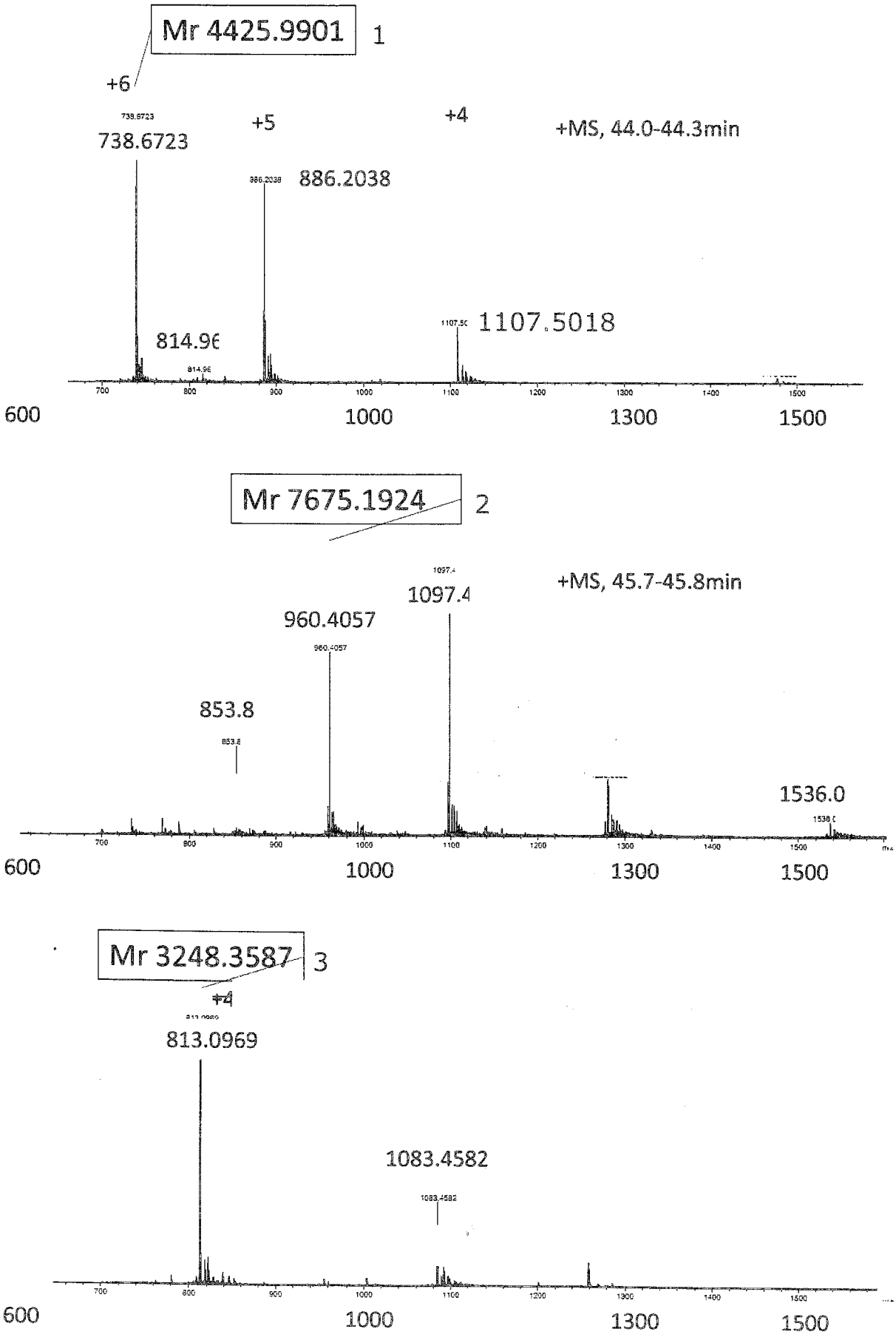

Mr 4425.9901
1
+6
+5
+4
+MS, 44.0-44.3min
738.6723
886.2038
814.9
1107.5018
600
1000
1300
1500
Mr 7675.1924
2
+MS, 45.7-45.8min
1097.4
960.4057
853.8
1536.0
600
1000
1300
1500
Mr 3248.3587
3
+4
813.0969
1083.4582
600
1000
1300
1500

Fig. 5

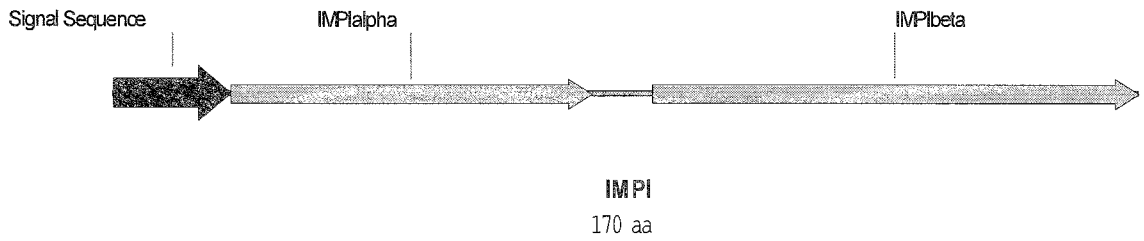

Signal Sequence
IMPIalpha
IMPIbeta
IMPI
170 aa

Fig. 6

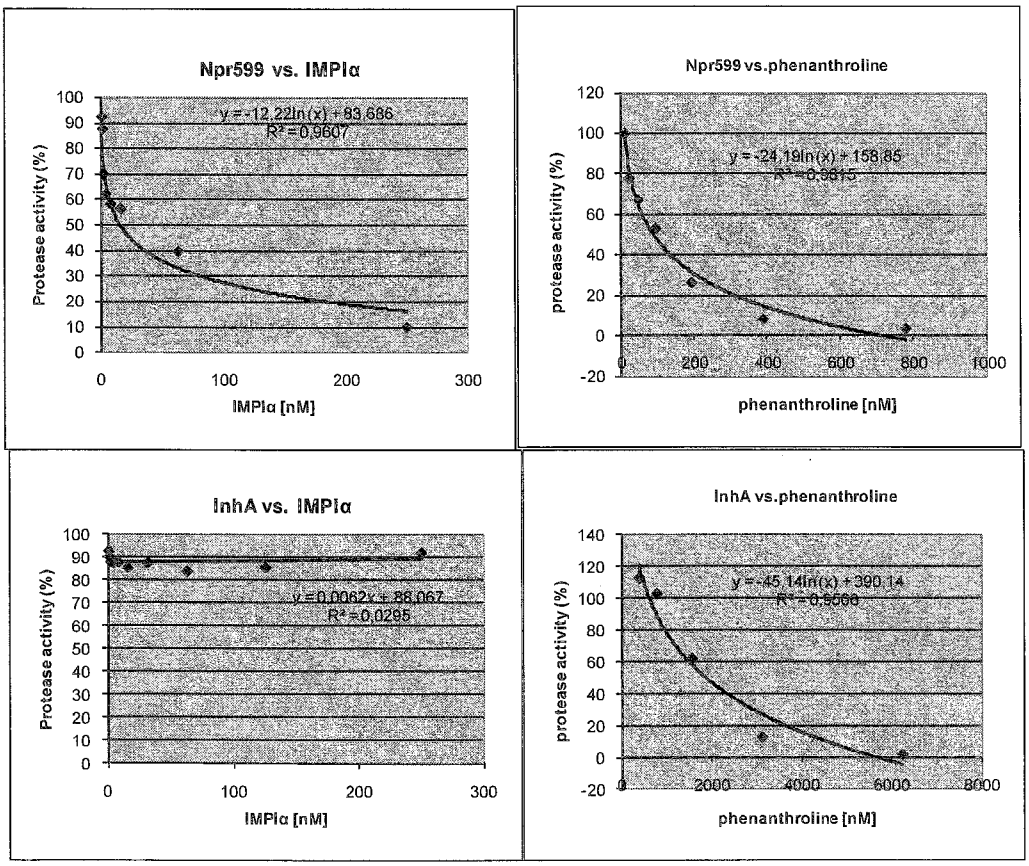

Npr599 vs. IMPIα
y = -12,22ln(x) + 83,686
R² = 0,9607
Protease activity (%)
IMPIα [nM]
Npr599 vs.phenanthroline
y = -24,19ln(x) + 158,85
R² = 0,9815
protease activity (%)
phenanthroline [nM]
InhA vs. IMPIα
y = 0,0062x + 88,067
R² = 0,0295
Protease activity (%)
IMPIα [nM]
InhA vs.phenanthroline
y = -45,14ln(x) + 390,14
R² = 0,9666
protease activity (%)
phenanthroline [nM]

Fig. 7

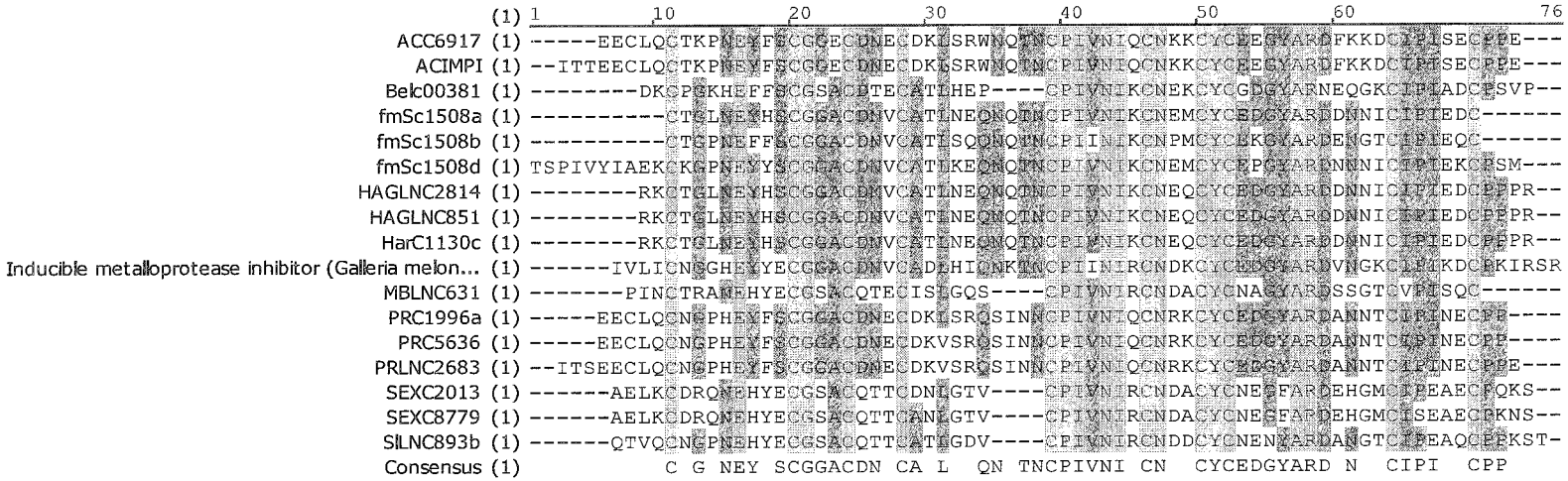


Fig 8a

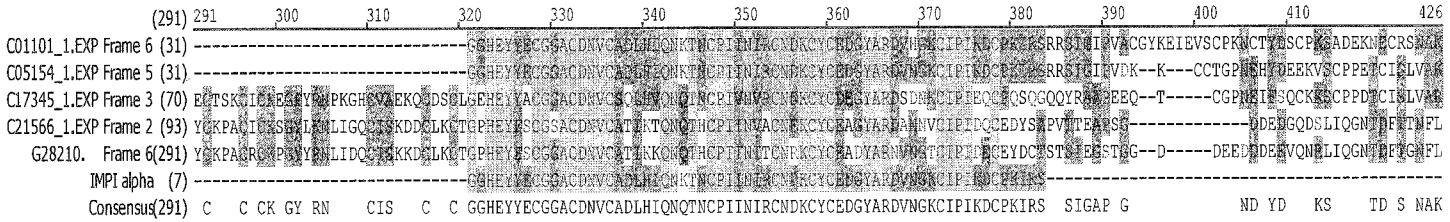


Fig. 8b

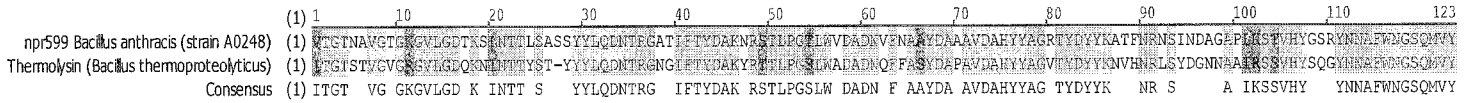


Fig. 8c

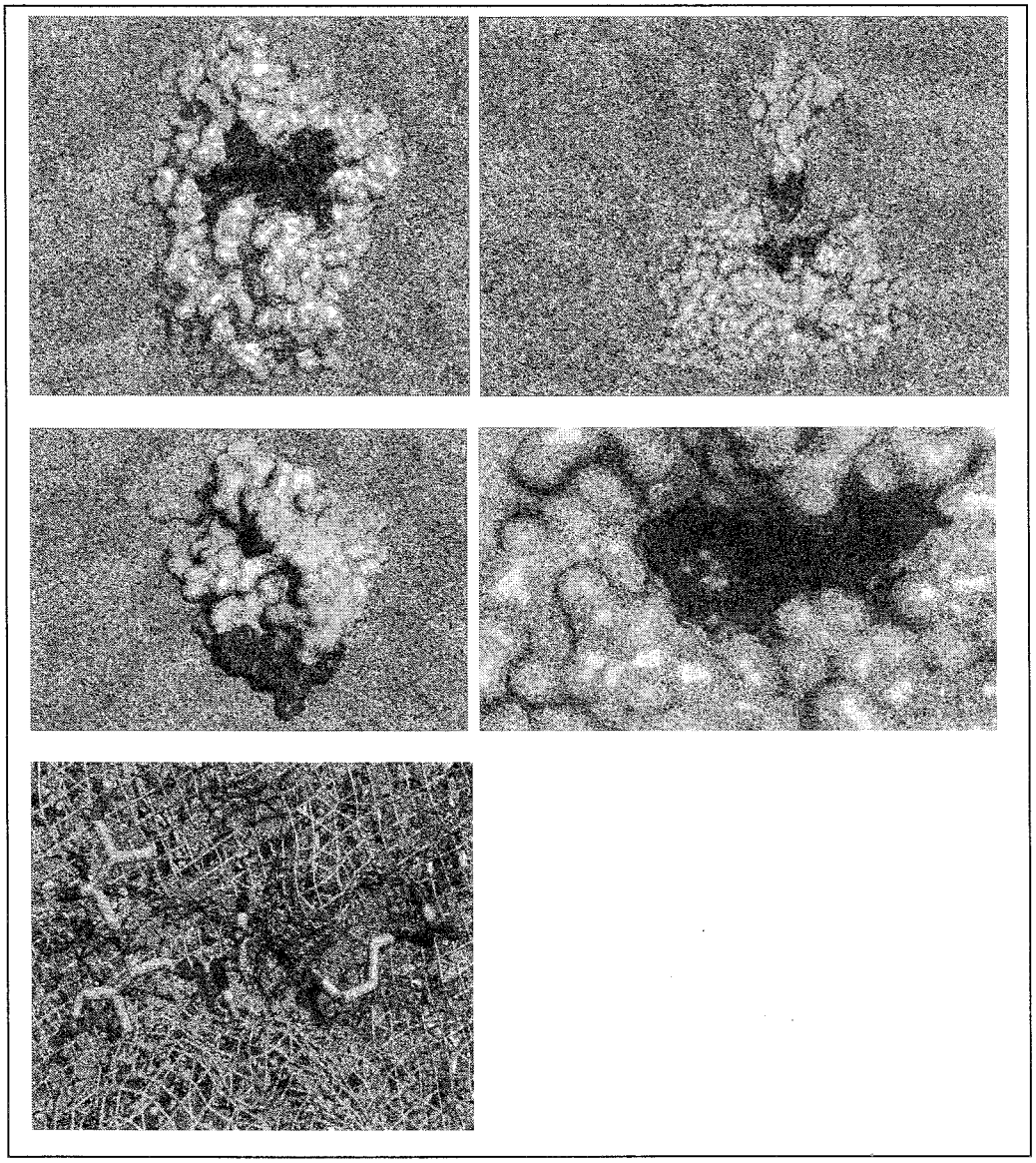


Fig. 9

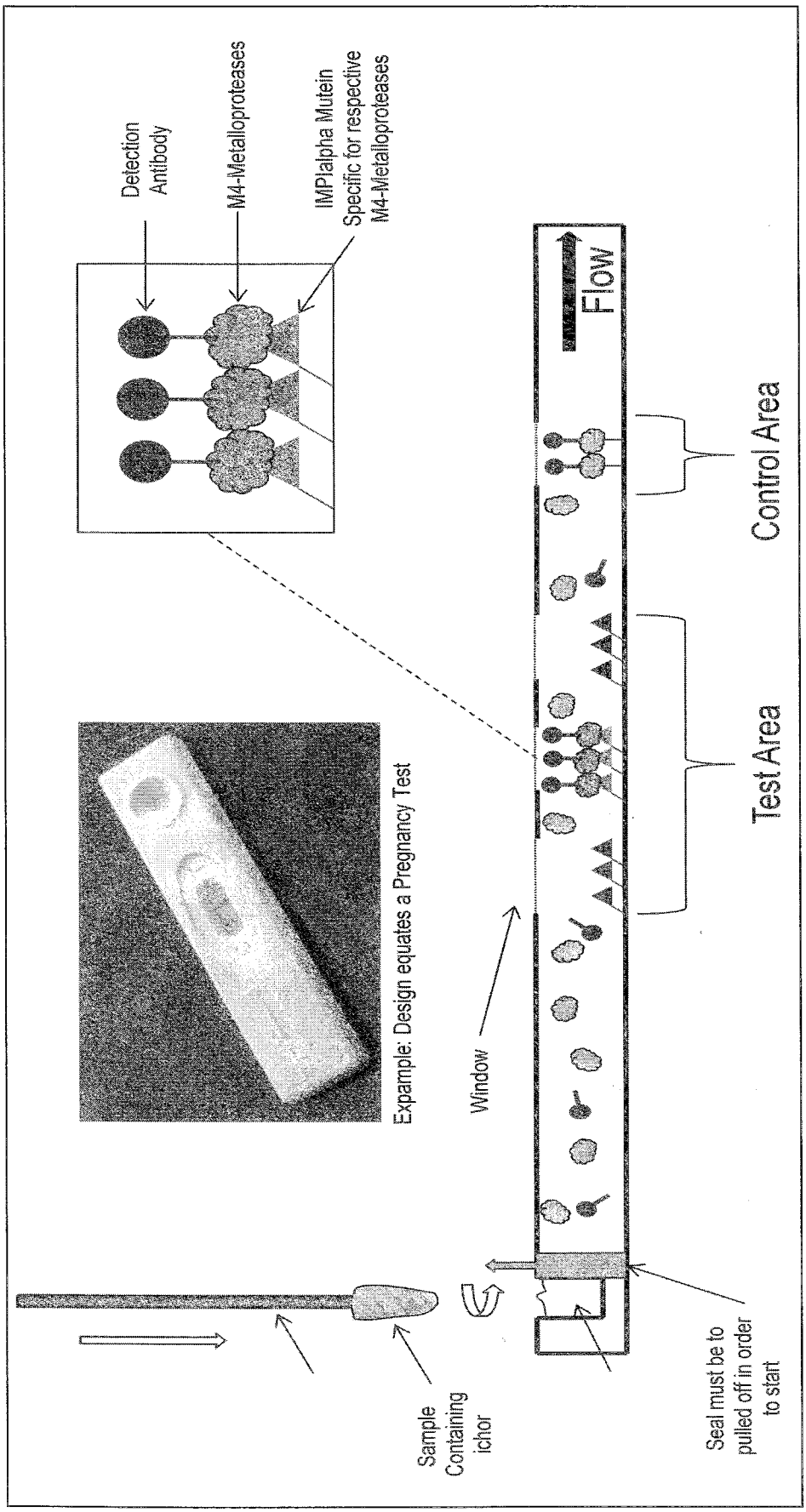

Detection Antibody
M4-Metalloproteases
IMPlalpha Mutein Specific for respective M4-Metalloproteases
Flow
Control Area
Test Area
Expample: Design equates a Pregnancy Test
Window
Sample Containing ichor
Seal must be to pulled off in order to start

Fig. 10

INTERNATIONAL SEARCH REPORT

International application No
PCT/EP2014/066165

A. CLASSIFICATION OF SUBJECT MATTER
INV. A61K38/57 C07K14/81
ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
A61K C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
EPO-Internal, BIOSIS, Sequence Search, WPI Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WEDDE MARIANNE ET AL: "Purification and characterization of an inducible metalloprotease inhibitor from the hemolymph of greater wax moth larvae, Galleria mellonella", EUR. J. BIOCHEM., vol. 255, no. 3, 1 August 1998 (1998-08-01), pages 535-543, XP002214760, cited in the application the whole document ----- -/--	4-7,9,10

[X] Further documents are listed in the continuation of Box C. [ ] See patent family annex.

* Special categories of cited documents :

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"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
20 October 2014	03/11/2014

Name and mailing address of the ISA/	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Fax: (+31-70) 340-3016	Scheffzyk, Irmgard

INTERNATIONAL SEARCH REPORT

International application No
PCT/EP2014/066165

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	ANJA CLERMONT ET AL: "Cloning and expression of an inhibitor of microbial metalloproteinases from insects contributing to innate immunity", THE BIOCHEMICAL JOURNAL, vol. 382, 15 August 2004 (2004-08-15), pages 315-322, XP055101475, England DOI: 10.1042/BJ20031923 cited in the application the whole document -----	4-7,10, 11