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(71) Applicant: **EVELO BIOSCIENCES, INC.** [US/US]; 620 Memorial Drive, Suite 200 West, Cambridge, MA 02139 (US).

(72) Inventors: **SANDY, Peter**; 198 Prospect Avenue, Revere, MA 02151 (US). **PAPKOFF, Jacqueline**; 4373 21st Street, San Francisco, CA 94114 (US). **SIZOVA, Maria**; 152 Walter Street Unit 1, Roslindale, MA 02131 (US). **GOODMAN, Brian**; 16 McBride Street, #1, Jamaica Plain, MA 02130 (US). **GARDNER, Humphrey**; 23 Clifton Avenue, Marblehead, MA 01945 (US). **PONICHTERA, Holly**; 530 3rd Street Unit 504, Cambridge, MA 02142 (US).

(74) Agent: **JONES, Brendan T.** et al.; Foley Hoag LLP, 155 Seaport Boulevard, Boston, MA 02210-2600 (US).

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(54) Title: COMPOSITIONS AND METHODS OF TREATING CANCER USING *BIFIDOBACTERIUM ANIMALIS SSP. LACTIS*

(57) Abstract: Provided herein are methods and compositions related to *Bifidobacterium animalis ssp. lactis* useful as therapeutic agents.

Compositions and Methods of Treating Cancer Using *Bifidobacterium animalis ssp. lactis*

RELATED APPLICATION

[0001] This application claims the benefit of priority to U.S. Provisional Patent Application serial number 62/528,669, filed July 5, 2017, which is hereby incorporated herein by reference in its entirety.

SUMMARY

[0002] In certain aspects, provided herein are methods and compositions related to the treatment of a cancer in a subject (*e.g.*, a human subject) comprising administering a bacterial composition comprising *Bifidobacterium animalis ssp. lactis*. In some embodiments, the *Bifidobacterium animalis ssp. lactis* is *Bifidobacterium animalis ssp. lactis* Strain A (ATCC Deposit Number PTA-125097). In some embodiments, the *Bifidobacterium animalis ssp. lactis* is a strain comprising at least 99% sequence identity (*e.g.*, at least 99.5% sequence identity, at least 99.6% sequence identity, at least 99.7% sequence identity, at least 99.8% sequence identity, at least 99.9% sequence identity) to the nucleotide sequence of the *Bifidobacterium animalis ssp. lactis* Strain A (Table 1). In some embodiments, the administration of the bacterial composition induces an immune response against a tumor in the subject. In some embodiments, the administration of the bacterial composition induces CD3+ immune cell infiltration in the subject. In some embodiments, the administration of the bacterial composition induces MHC Class I upregulation at the tumor site. In some embodiments, the administration of the bacterial composition treats the cancer in the subject. In some embodiments, the administration augments a tumor microenvironment in the subject. In some embodiments, the cancer is a colorectal carcinoma.

[0003] In certain embodiments, provided herein are methods of treating a subject who has cancer, comprising administering to the subject a bacterial composition comprising *Bifidobacterium animalis ssp. lactis* (*e.g.*, a killed bacterium, a live bacterium and/or an attenuated bacterium). In some embodiments, the *Bifidobacterium animalis ssp. lactis* is *Bifidobacterium animalis ssp. lactis* Strain A (ATCC Deposit Number PTA-125097). In some embodiments, the *Bifidobacterium animalis ssp. lactis* is a strain comprising at least 99% sequence identity (*e.g.*, at least 99.5% sequence identity, at least 99.6% sequence identity, at

least 99.7% sequence identity, at least 99.8% sequence identity, at least 99.9% sequence identity) to the nucleotide sequence of the *Bifidobacterium animalis ssp. lactis* Strain A. In some embodiments, at least 50%, 60%, 70%, 80%, 85%, 90%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% of the bacteria in the bacterial composition are *Bifidobacterium animalis ssp. lactis.* In some embodiments, all or substantially all of the bacteria in the bacterial formulation are *Bifidobacterium animalis ssp. lactis.* In some embodiments, the bacterial formulation comprises at least 1×10^5, 5×10^5, 1×10^6, 2×10^6, 3×10^6, 4×10^6, 5×10^6, 6×10^6, 7×10^6, 8×10^6, 9×10^6, 1×10^7, 2×10^7, 3×10^7, 4×10^7, 5×10^7, 6×10^7, 7×10^7, 8×10^7, 9×10^7, 1×10^8, 2×10^8, 3×10^8, 4×10^8, 5×10^8, 6×10^8, 7×10^8, 8×10^8, 9×10^8 or 1×10^9 colony forming units of *Bifidobacterium animalis ssp. lactis.*

[0004] In certain embodiments, provided herein are bacterial compositions comprising *Bifidobacterium animalis ssp. lactis* (*e.g.,* a killed bacterium, a live bacterium and/or an attenuated bacterium). In some embodiments, at least 50%, 60%, 70%, 80%, 85%, 90%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% of the bacteria in the bacterial composition are *Bifidobacterium animalis ssp. lactis.* In some embodiments, the *Bifidobacterium animalis ssp. lactis* is *Bifidobacterium animalis ssp. lactis* Strain A (ATCC Deposit Number PTA-125097). In some embodiments, the *Bifidobacterium animalis ssp. lactis* is a strain comprising at least 99% sequence identity (*e.g.,* at least 99.5% sequence identity, at least 99.6% sequence identity, at least 99.7% sequence identity, at least 99.8% sequence identity, at least 99.9% sequence identity) to the nucleotide sequence of the *Bifidobacterium animalis ssp. lactis* Strain A. In some embodiments, all or substantially all of the bacteria in the bacterial formulation are *Bifidobacterium animalis ssp. lactis.* In some embodiments, the bacterial formulation comprises at least 1×10^5, 5×10^5, 1×10^6, 2×10^6, 3×10^6, 4×10^6, 5×10^6, 6×10^6, 7×10^6, 8×10^6, 9×10^6, 1×10^7, 2×10^7, 3×10^7, 4×10^7, 5×10^7, 6×10^7, 7×10^7, 8×10^7, 9×10^7, 1×10^8, 2×10^8, 3×10^8, 4×10^8, 5×10^8, 6×10^8, 7×10^8, 8×10^8, 9×10^8 or 1×10^9 colony forming units of *Bifidobacterium animalis ssp. lactis.* In some embodiments, the bacterial composition comprise at least one strain of *Bifidobacterium animalis ssp. lactis* having one gene comprising a nucleotide base change as identified in Table 6. In some embodiments, the bacterial composition comprise at least one strain of *Bifidobacterium animalis ssp. lactis* having 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, or 29, genes comprising a nucleotide base change as identified in Table 6. In some embodiments, the bacterial composition

comprise at least one strain of *Bifidobacterium animalis ssp. lactis* having genes comprising all the nucleotide base change as identified in Table 6.

[0005] In some embodiments, provided herein are pharmaceutically active biomasses (PhABs) derived from and/or comprising *Bifidobacterium animalis ssp. lactis* (e.g., *Bifidobacterium animalis ssp. lactis* Strain A (ATCC Deposit Number PTA-125097)). In some embodiments, the PhABs comprise whole cells, fractions of cells, supernatant from fermentation, fractions of supernatant and/or extracellular vesicles made from *Bifidobacterium animalis ssp. lactis* described herein. In some embodiments, the bacterial compositions provided herein comprise *Bifidobacterium animalis ssp. lactis* PhABs provided herein.

[0006] In some embodiments, the bacterial composition is administered orally, intravenously, intratumorally, or subcutaneously. In some embodiments, the bacterial composition is administered in 2 or more (*e.g.,* 3 or more, 4 or more or 5 or more doses). In some embodiments, the administration to the subject of the two or more doses are separated by at least 1 hour, 2 hours, 3 hours, 4 hours, 5 hours, 6 hours, 7 hours, 8 hours, 9 hours, 10 hours, 11 hours, 12 hours, 13 hours, 14 hours, 15 hours, 16 hours, 17 hours, 18 hours, 1 day, 2 days, 3 days, 4 days, 5 days, 6 days, 7 days, 8 days, 9 days, 10 days, 11 days, 12 days, 13 days, 14 days, 15 days, 16 days, 17 days, 18 days, 19 days, 20 days or 21 days. In some embodiments, a second bacterium is administered as part of an ecological consortium.

[0007] In some embodiments, the method further comprises administering to the subject an antibiotic. In some embodiments, the method further comprises administering to the subject one or more other cancer therapies. In some embodiments, the other cancer therapy is the surgical removal of a tumor, the administration of a chemotherapeutic agent, the administration of radiation therapy, the administration of an antibiotic, the administration of a cancer immunotherapy (*e.g.,* an immune checkpoint inhibitor, a cancer-specific antibody, a cancer vaccine, a primed antigen presenting cell, a cancer-specific T cell, a cancer-specific chimeric antigen receptor (CAR) T cell, an immune activating protein, an adjuvant), and/or the administration of another therapeutic bacterium.

[0008] In some embodiments, the subject is a mammal. In some embodiments, the subject is a human. In some embodiments, the subject is a non-human mammal (*e.g.,* a dog, a cat, a cow, a horse, a pig, a donkey, a goat, a camel, a mouse, a rat, a guinea pig, a sheep, a llama, a monkey, a gorilla or a chimpanzee).

[0009] In certain embodiments, provided herein is A bioreactor comprising *Bifidobacterium animalis ssp. Lactis.*

BRIEF DESCRIPTION OF THE FIGURES

[0010] **Figure 1** shows inhibition of tumor growth (by volume) by the oral administration of *Bifidobacterium animalis ssp. lactis* Strain A in a mouse colorectal carcinoma model.

[0011] **Figure 2** shows that in a mouse colorectal carcinoma model, the efficacy of orally administered *Bifidobacterium animalis ssp. lactis* Strain A is comparable to that of intraperitoneally (i.p.) administered anti-PD-1.

[0012] **Figure 3** shows that in a mouse colorectal carcinoma model, the efficacy of orally administered *Bifidobacterium animalis ssp. lactis* Strain A is comparable to that of intraperitoneally (i.p.) administered anti-PD-1.

[0013] **Figure 4** shows that in a mouse colorectal carcinoma model, the efficacy of intratumorally *Bifidobacterium animalis ssp. lactis* Strain A is comparable to that of intraperitoneally (i.p.) administered anti-PD-1.

[0014] **Figure 5** shows that in a mouse melanoma model, treating tumor-inoculated mice with a combination of *Bifidobacterium animalis ssp. lactis* Strain A and anti-PD-L1 inhibits tumor growth in a manner superior to either *B. animalis ssp. lactis* Strain A or anti-PD-L1 alone.

[0015] **Figure 6A** shows the tumor volume prior to CD3+ immune cell infiltrate and MHC Class I expression analysis. **Figure 6A** shows inhibition of tumor growth (by volume) by the oral administration of *Bifidobacterium animalis ssp. lactis* Strain A in a mouse colorectal carcinoma model.

[0016] **Figure 6B** shows the infiltration of CD3+ immune cells was significantly increased in the Bifidobacterium animalis ssp. lactis Strain A group relative to the vehicle group.

[0017] **Figure 6C** shows that *Bifidobacterium animalis ssp. lactis* Strain A induced a striking upregulation of MHC Class I expression relative to the vehicle group.

[0018] **Figure 7** shows an overview of a phase 1/2 open-label study of the safety, tolerability and efficacy of a *Bifidobacterium animalis ssp. lactis* Strain A as an oral therapeutic in patients with metastatic colorectal carcinoma.

[0019] **Figure 8** shows that in a mouse colorectal carcinoma model, the efficacy of a triple combination of *Bifidobacterium animalis ssp. lactis* Strain A, anti-PD-1, and anti-CTLA4 is greater than that of the combination of anti-PD-1 and anti-CTLA4.

[0020] **Figure 9A** and **Figure 9B** show a comparison of in vitro signatures of three strains of *Bifidobacterium animalis ssp. lactis* (Strain A, Strain B, and Strain C) from a mouse dendritic cell assay. Strain A exhibits an immune profile with a decreased induction of pro-tumorigenic cytokines (IL-10, IL-1b, IL-6, IL-8, TNFα) compared to Strain B and Strain C. The cytokine values reported represent geometric mean summary of groups (n=3) across the assay.

DETAILED DESCRIPTION

General

[0021] In certain aspects, provided herein are bacterial compositions comprising *Bifidobacterium animalis ssp. lactis* and methods of treating cancer in a subject comprising administering to the subject a bacterial composition comprising *Bifidobacterium animalis ssp. lactis.*

Definitions

[0022] "Adjuvant" or "Adjuvant therapy" broadly refers to an agent that affects an immunological or physiological response in a patient or subject. For example, an adjuvant might increase the presence of an antigen over time or to an area of interest like a tumor, help absorb an antigen presenting cell antigen, activate macrophages and lymphocytes and support the production of cytokines. By changing an immune response, an adjuvant might permit a smaller dose of an immune interacting agent to increase the effectiveness or safety of a particular dose of the immune interacting agent. For example, an adjuvant might prevent T cell exhaustion and thus increase the effectiveness or safety of a particular immune interacting agent.

[0023] "Administration" broadly refers to a route of administration of a composition to a subject. Examples of routes of administration include oral administration, rectal administration, topical administration, inhalation (nasal) or injection. Administration by injection includes intravenous (IV), intramuscular (IM), intratumoral (IT) and subcutaneous (SC) administration. The pharmaceutical compositions described herein can be administered in any form by any effective route, including but not limited to intratumoral, oral, parenteral, enteral, intravenous,

intraperitoneal, topical, transdermal (*e.g.*, using any standard patch), intradermal, ophthalmic, (intra)nasally, local, non-oral, such as aerosol, inhalation, subcutaneous, intramuscular, buccal, sublingual, (trans)rectal, vaginal, intra-arterial, and intrathecal, transmucosal (*e.g.*, sublingual, lingual, (trans)buccal, (trans)urethral, vaginal (*e.g.*, trans- and perivaginally), intravesical, intrapulmonary, intraduodenal, intragastrical, and intrabronchial. In preferred embodiments, the pharmaceutical compositions described herein are administered orally, rectally, intratumorally, topically, intravesically, by injection into or adjacent to a draining lymph node, intravenously, by inhalation or aerosol, or subcutaneously.

[0024] As used herein, the term "antibody" may refer to both an intact antibody and an antigen binding fragment thereof. Intact antibodies are glycoproteins that include at least two heavy (H) chains and two light (L) chains inter-connected by disulfide bonds. Each heavy chain includes a heavy chain variable region (abbreviated herein as V_H) and a heavy chain constant region. Each light chain includes a light chain variable region (abbreviated herein as V_L) and a light chain constant region. The V_H and V_L regions can be further subdivided into regions of hypervariability, termed complementarity determining regions (CDR), interspersed with regions that are more conserved, termed framework regions (FR). Each V_H and V_L is composed of three CDRs and four FRs, arranged from amino-terminus to carboxy-terminus in the following order: FR1, CDR1, FR2, CDR2, FR3, CDR3, FR4. The variable regions of the heavy and light chains contain a binding domain that interacts with an antigen. The term "antibody" includes, for example, monoclonal antibodies, polyclonal antibodies, chimeric antibodies, humanized antibodies, human antibodies, multispecific antibodies (*e.g.*, bispecific antibodies), single-chain antibodies and antigen-binding antibody fragments.

[0025] The terms "antigen binding fragment" and "antigen-binding portion" of an antibody, as used herein, refers to one or more fragments of an antibody that retain the ability to bind to an antigen. Examples of binding fragments encompassed within the term "antigen-binding fragment" of an antibody include Fab, Fab', $F(ab')_2$, Fv, scFv, disulfide linked Fv, Fd, diabodies, single-chain antibodies, NANOBODIES®, isolated CDRH3, and other antibody fragments that retain at least a portion of the variable region of an intact antibody. These antibody fragments can be obtained using conventional recombinant and/or enzymatic techniques and can be screened for antigen binding in the same manner as intact antibodies.

[0026] "Cancer" broadly refers to an uncontrolled, abnormal growth of a host's own cells leading to invasion of surrounding tissue and potentially tissue distal to the initial site of abnormal cell growth in the host. Major classes include carcinomas which are cancers of the epithelial tissue (*e.g.*, skin, squamous cells); sarcomas which are cancers of the connective tissue (*e.g.*, bone, cartilage, fat, muscle, blood vessels, etc.); leukemias which are cancers of blood forming tissue (*e.g.*, bone marrow tissue); lymphomas and myelomas which are cancers of immune cells; and central nervous system cancers which include cancers from brain and spinal tissue. "Cancer(s)," "neoplasm(s)," and "tumor(s)" are used herein interchangeably. As used herein, "cancer" refers to all types of cancer or neoplasm or malignant tumors including leukemias, carcinomas and sarcomas, whether new or recurring. Specific examples of cancers are: carcinomas, sarcomas, myelomas, leukemias, lymphomas and mixed type tumors. Non-limiting examples of cancers are new or recurring cancers of the brain, melanoma, bladder, breast, cervix, colon, head and neck, kidney, lung, non-small cell lung, mesothelioma, ovary, prostate, sarcoma, stomach, uterus and medulloblastoma. Pediatric and adult tumors include, but not limited to, those of bladder, brain, breast, bone, cervix, colon, connective tissue, fat, head and neck, kidney, liver, lung, mesothelium, melanocytes (melanoma), muscle, ovary, pancreas, prostate, stomach, small intestine, and uterus

[0027] The term "decrease" or "deplete" means a change, such that the difference is, depending on circumstances, at least 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 1/100, 1/1000, 1/10,000, 1/100,000, 1/1,000,000 or undetectable after treatment when compared to a pre-treatment state.

[0028] The term "ecological consortium" is a group of bacteria which trades metabolites and positively co-regulates one another, in contrast to two bacteria which induce host synergy through activating complementary host pathways for improved efficacy.

[0029] The term "epitope" means a protein determinant capable of specific binding to an antibody. Epitopes usually consist of chemically active surface groupings of molecules such as amino acids or sugar side chains. Certain epitopes can be defined by a particular sequence of amino acids to which an antibody is capable of binding.

[0030] The term "gene" is used broadly to refer to any nucleic acid associated with a biological function. The term "gene" applies to a specific genomic sequence, as well as to a cDNA or an mRNA encoded by that genomic sequence.

[0031] "Identity" as between nucleic acid sequences of two nucleic acid molecules can be determined as a percentage of identity using known computer algorithms such as the "FASTA" program, using for example, the default parameters as in Pearson *et al.* (1988) Proc. Natl. Acad. Sci. USA 85:2444 (other programs include the GCG program package (Devereux, J., *et al.*, Nucleic Acids Research 12(I):387 (1984)), BLASTP, BLASTN, FASTA Atschul, S. F., *et al.*, J Molec Biol 215:403 (1990); Guide to Huge Computers, Mrtin J. Bishop, ed., Academic Press, San Diego, 1994, and Carillo *et al.* (1988) SIAM J Applied Math 48:1073). For example, the BLAST function of the National Center for Biotechnology Information database can be used to determine identity. Other commercially or publicly available programs include, DNAStar "MegAlign" program (Madison, Wis.) and the University of Wisconsin Genetics Computer Group (UWG) "Gap" program (Madison Wis.)).

[0032] "Immunotherapy" is treatment that uses a subject's immune system to treat cancer and includes, for example, checkpoint inhibitors, cancer vaccines, cytokines, cell therapy, CAR-T cells, and dendritic cell therapy.

[0033] The term "increase" means a change, such that the difference is, depending on circumstances, at least 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 2-fold, 4-fold, 10-fold, 100-fold, 10^3 fold, 10^4 fold, 10^5 fold, 10^6 fold, and/or 10^7 fold greater after treatment when compared to a pre-treatment state. Properties that may be increased include immune cells, bacterial cells, stromal cells, myeloid derived suppressor cells, fibroblasts, metabolites, and cytokines.

[0034] "Innate immune agonists" or "immuno-adjuvants" are small molecules, proteins, or other agents that specifically target innate immune receptors including Toll-Like Receptors, NOD receptors, STING Pathway components. For example, LPS is a TLR-4 agonist that is bacterially derived or synthesized and aluminum can be used as an immune stimulating adjuvant. immuno-adjuvants are a specific class of broader adjuvant or adjuvant therapy.

[0035] The term "isolated" or "enriched" encompasses a microbe, bacteria or other entity or substance that has been (1) separated from at least some of the components with which it was associated when initially produced (whether in nature or in an experimental setting), and/or (2) produced, prepared, purified, and/or manufactured by the hand of man. Isolated microbes may be separated from at least about 10%, about 20%, about 30%, about 40%, about 50%, about 60%, about 70%, about 80%, about 90%, or more of the other components with which they were

initially associated. In some embodiments, isolated microbes are more than about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or more than about 99% pure. As used herein, a substance is "pure" if it is substantially free of other components. The terms "purify," "purifying" and "purified" refer to a microbe or other material that has been separated from at least some of the components with which it was associated either when initially produced or generated (*e.g.*, whether in nature or in an experimental setting), or during any time after its initial production. A microbe or a microbial population may be considered purified if it is isolated at or after production, such as from a material or environment containing the microbe or microbial population, and a purified microbe or microbial population may contain other materials up to about 10%, about 20%, about 30%, about 40%, about 50%, about 60%, about 70%, about 80%, about 90%, or above about 90% and still be considered "isolated." In some embodiments, purified microbes or microbial population are more than about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or more than about 99% pure. In the instance of microbial compositions provided herein, the one or more microbial types present in the composition can be independently purified from one or more other microbes produced and/or present in the material or environment containing the microbial type. Microbial compositions and the microbial components thereof are generally purified from residual habitat products.

[0036] As used herein, a gene is "overexpressed" in a bacteria if it is expressed at a higher level in an engineered bacteria under at least some conditions than it is expressed by a wild-type bacteria of the same species under the same conditions. Similarly, a gene is "underexpressed" in a bacteria if it is expressed at a lower level in an engineered bacteria under at least some conditions than it is expressed by a wild-type bacteria of the same species under the same conditions.

[0037] The terms "polynucleotide" and "nucleic acid" are used interchangeably. They refer to a polymeric form of nucleotides of any length, either deoxyribonucleotides or ribonucleotides, or analogs thereof. Polynucleotides may have any three-dimensional structure, and may perform any function. The following are non-limiting examples of polynucleotides: coding or non-coding regions of a gene or gene fragment, loci (locus) defined from linkage analysis, exons, introns, messenger RNA (mRNA), transfer RNA, ribosomal RNA, ribozymes,

cDNA, recombinant polynucleotides, branched polynucleotides, plasmids, vectors, isolated DNA of any sequence, isolated RNA of any sequence, nucleic acid probes, and primers. A polynucleotide may comprise modified nucleotides, such as methylated nucleotides and nucleotide analogs. If present, modifications to the nucleotide structure may be imparted before or after assembly of the polymer. A polynucleotide may be further modified, such as by conjugation with a labeling component. In all nucleic acid sequences provided herein, U nucleotides are interchangeable with T nucleotides.

[0038] "Operational taxonomic units" and "OTU(s)" refer to a terminal leaf in a phylogenetic tree and is defined by a nucleic acid sequence, e.g., the entire genome, or a specific genetic sequence, and all sequences that share sequence identity to this nucleic acid sequence at the level of species. In some embodiments the specific genetic sequence may be the 16S sequence or a portion of the 16S sequence. In other embodiments, the entire genomes of two entities are sequenced and compared. In another embodiment, select regions such as multilocus sequence tags (MLST), specific genes, or sets of genes may be genetically compared. For 16S, OTUs that share $\geq$ 97% average nucleotide identity across the entire 16S or some variable region of the 16S are considered the same OTU. See e.g. Claesson MJ, Wang Q, O'Sullivan O, Greene-Diniz R, Cole JR, Ross RP, and O'Toole PW. 2010. Comparison of two next-generation sequencing technologies for resolving highly complex microbiota composition using tandem variable 16S rRNA gene regions. Nucleic Acids Res 38: e200. Konstantinidis KT, Ramette A, and Tiedje JM. 2006. The bacterial species definition in the genomic era. Philos Trans R Soc Lond B Biol Sci 361: 1929–1940. For complete genomes, MLSTs, specific genes, other than 16S, or sets of genes OTUs that share $\geq$ 95% average nucleotide identity are considered the same OTU. See e.g., Achtman M, and Wagner M. 2008. Microbial diversity and the genetic nature of microbial species. Nat. Rev. Microbiol. 6: 431–440. Konstantinidis KT, Ramette A, and Tiedje JM. 2006. The bacterial species definition in the genomic era. Philos Trans R Soc Lond B Biol Sci 361: 1929–1940. OTUs are frequently defined by comparing sequences between organisms. Generally, sequences with less than 95% sequence identity are not considered to form part of the same OTU. OTUs may also be characterized by any combination of nucleotide markers or genes, in particular highly conserved genes (e.g., "house-keeping" genes), or a combination thereof. Operational Taxonomic Units (OTUs) with taxonomic assignments made to, e.g., genus, species, and phylogenetic clade are provided herein.

[0039] As used herein, "specific binding" refers to the ability of an antibody to bind to a predetermined antigen or the ability of a polypeptide to bind to its predetermined binding partner. Typically, an antibody or polypeptide specifically binds to its predetermined antigen or binding partner with an affinity corresponding to a K_D of about 10^{-7} M or less, and binds to the predetermined antigen/binding partner with an affinity (as expressed by K_D) that is at least 10 fold less, at least 100 fold less or at least 1000 fold less than its affinity for binding to a non-specific and unrelated antigen/binding partner (*e.g.*, BSA, casein). Alternatively, specific binding applies more broadly to a two component system where one component is a protein, lipid, or carbohydrate or combination thereof and engages with the second component which is a protein, lipid, carbohydrate or combination thereof in a specific way.

[0040] The terms "subject" or "patient" refers to any animal. A subject or a patient described as "in need thereof" refers to one in need of a treatment for a disease. Mammals (*i.e.*, mammalian animals) include humans, laboratory animals (*e.g.*, primates, rats, mice), livestock (*e.g.*, cows, sheep, goats, pigs), and household pets (*e.g.*, dogs, cats, rodents). For example, the subject may be a non-human mammal including but not limted to of a dog, a cat, a cow, a horse, a pig, a donkey, a goat, a camel, a mouse, a rat, a guinea pig, a sheep, a llama, a monkey, a gorilla or a chimpanzee. The subject or patient may be healthy, or may be suffering from a neoplasm at any developmental stage, wherein any of the stages are either caused by or opportunistically supported of a cancer associated or causative pathogen, or may be at risk of developing a neoplasm, or transmitting to others a cancer associated or cancer causative pathogen. In some embodiments patients have lung cancer, bladder cancer, prostate cancer, ovarian cancer, and/or melanoma. The patients may have tumors that show enhanced macropinocytosis with the underlying genomics of this process including Ras activation. In other embodiments patients suffer from other cancers. In some embodiments, the subject has undergone a cancer therapy.

[0041] "Strain" refers to a member of a bacterial species with a genetic signature such that it may be differentiated from closely-related members of the same bacterial species. The genetic signature may be the absence of all or part of at least one gene, the absence of all or part of at least on regulatory region (*e.g.*, a promoter, a terminator, a riboswitch, a ribosome binding site), the absence ("curing") of at least one native plasmid, the presence of at least one recombinant gene, the presence of at least one mutated gene, the presence of at least one foreign

gene (a gene derived from another species), the presence at least one mutated regulatory region (*e.g.*, a promoter, a terminator, a riboswitch, a ribosome binding site), the presence of at least one non-native plasmid, the presence of at least one antibiotic resistance cassette, or a combination thereof. Genetic signatures between different strains may be identified by PCR amplification optionally followed by DNA sequencing of the genomic region(s) of interest or of the whole genome. In the case in which one strain (compared with another of the same species) has gained or lost antibiotic resistance or gained or lost a biosynthetic capability (such as an auxotrophic strain), strains may be differentiated by selection or counter-selection using an antibiotic or nutrient/metabolite, respectively.

[0042] As used herein, the term "treating" a disease in a subject or "treating" a subject having or suspected of having a disease refers to subjecting the subject to a pharmaceutical treatment, *e.g.*, the administration of one or more agents, such that at least one symptom of the disease is decreased or prevented from worsening. Thus, in one embodiment, "treating" refers inter alia to delaying progression, expediting remission, inducing remission, augmenting remission, speeding recovery, increasing efficacy of or decreasing resistance to alternative therapeutics, or a combination thereof. In certain embodiments, a cancer is treated if the subject experiences a reduction in tumor size, a reduced number of tumors, a reduction in tumor growth, a reduction in cancer metastisis and/or a reduced number of total cancer cells following treatment than would be expected in the absence of treatment.

Bacteria

[0043] In certain aspects, provided herein are bacterial compositions comprising *Bifidobacterium animalis ssp. lactis* (e.g.,. an effective amount of *Bifidobacterium animalis ssp. lactis*) and methods of using a bacterial composition comprising *Bifidobacterium animalis ssp. lactis.* In some embodiments, the *Bifidobacterium animalis ssp. lactis* is *Bifidobacterium animalis ssp. lactis* Strain A (ATCC Deposit Number PTA-125097). In some embodiments, the bacterial composition comprise a PhAB made from or comprising a *Bifidobacterium animalis ssp. lactis* described herein. In some embodiments, the *Bifidobacterium animalis ssp. lactis* is a strain comprising at least 99% sequence identity (*e.g.,* at least 99.5% sequence identity, at least 99.6% sequence identity, at least 99.7% sequence identity, at least 99.8% sequence identity, at

least 99.9% sequence identity) to the nucleotide sequence of the *Bifidobacterium animalis ssp. lactis* Strain A.

[0044] Under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure, the *Bifidobacterium animalis ssp. Lactis* Strain A was deposited on April 27, 2018, with the American Type Culture Collection (ATCC) of 10801 University Boulevard, Manassas, Va. 20110-2209 USA and was assigned ATCC Accession Number PTA-125097.

[0045] Applicant represents that the ATCC is a depository affording permanence of the deposit and ready accessibility thereto by the public if a patent is granted. All restrictions on the availability to the public of the material so deposited will be irrevocably removed upon the granting of a patent. The material will be available during the pendency of the patent application to one determined by the Commissioner to be entitled thereto under 37 CFR 1.14 and 35 U.S.C. 122. The deposited material will be maintained with all the care necessary to keep it viable and uncontaminated for a period of at least five years after the most recent request for the furnishing of a sample of the deposited plasmid, and in any case, for a period of at least thirty (30) years after the date of deposit or for the enforceable life of the patent, whichever period is longer. Applicant acknowledges its duty to replace the deposit should the depository be unable to furnish a sample when requested due to the condition of the deposit.

[0046] In some embodiments, the bacteria described herein are modified to improve colonization and/or engraftment in the mammalian gastrointestinal tract (*e.g.,* modified metabolism, such as improved mucin degradation, enhanced competition profile, increased motility, increased adhesion to gut epithelial cells, modified chemotaxis). In some embodiments, the bacteria described herein are modified to enhance their immunomodulatory and/or therapeutic effect (*e.g.,* either alone or in combination with another therapeutic agent). In some embodiments, the bacteria described herein are modified to enhance immune activation (*e.g.,* through modified production of polysaccharides, pili, fimbriae, adhesins, outer membrane vesicles). In some embodiments, the bacteria described herein are modified to improve bacterial manufacturing (*e.g.,* higher oxygen tolerance, improved freeze-thaw tolerance, shorter generation times).

[0047] In certain embodiments, the *Bifidobacterium animalis ssp. lactis* is grown in Reinforced Clostridium medium (RCM medium) (ATCC 2107 or medium from other

manufacturer), Lactobacilli MRS medium (LMRS medium), and/or Bifido agar. Alternatively, *Bifidobactrium animalis ssp. lactis* is grown using RCM medium ATCC 1053 and/or Trypticase soy agar/broth with defibrimated sheep blood ATCC 260 per the manufacturer's instructions.

[0048] The nucleotide sequence of the *Bifidobacterium animalis ssp. lactis* Strain A is shown in Table 1.

Table 1: Nucleotide sequence of the Bifidobacterium animalis ssp. lactis Strain A:

Nucleotide Sequence
>1 length=1944302 depth=1.00x circular=true
CAACGATGCCTCATCCACAGCAGCCAACTCAAAGACATCCTTACACACTAACCTGAGAAGAGCCAACAAA
GGCATCCCATTGCAGCGCATCGCAGGCTTCCACGTCGACGGCGACGAGCAACGACTCGCCATGCTGCATC
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AGGGGGCCGTATCACCCGACCGCGTAGTCTCAATGCGGTGCCGCTCATTCCCGTACGTATACAGTCGGGG
TCGGATTTTGCGGATTGGCCTGCCGGTCAGTGCGATTACTGGGCGGAGCGCATAGGCAACTATGTGCTCA
CCCAATCCACCCGCAAATCGATGGAGGGACTCGACGACTTCCAGACGCGACGAGCCCGCATTCTGCGCAG
CCCAAGCTCACGCCAGTTCCCGTTGACGCGCGTGATCGGCGAACTTGAGAAGCTGACGCCGCAGTTCCTG
CAATTGCGGCAGCACCAGATCGTCGAGCTGATCGCACAGGCGTGGAACATCAATCCGGGGGAGCTGGGAT
CCTTGGCAGAGCCGCAGGGGCAGTCCGATCGCACAGGCACATCCGCGCATCGCAGTCGACGCACGTCGAA
GCGTGTGACGGTGGGCGAAGTGGTGCGCGCCGGTCTGCTCACGGTGGGGGACCGATTCGTATGGAACCGC
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Bacterial Compositions

[0049] In certain aspects, provided herein are bacterial compositions comprising *Bifidobacterium animalis ssp. Lactis*. In some embodiments, the *Bifidobacterium animalis ssp. lactis* is *Bifidobacterium animalis ssp. lactis* Strain A (ATCC Deposit Number PTA-125097). In some embodiments, the bacterial composition comprise a PhAB made from or comprising a *Bifidobacterium animalis ssp. lactis* described herein. In some embodiments, the *Bifidobacterium animalis ssp. lactis* is a strain comprising at least 99% sequence identity (*e.g.*, at least 99.5% sequence identity, at least 99.6% sequence identity, at least 99.7% sequence identity, at least 99.8% sequence identity, at least 99.9% sequence identity) to the nucleotide sequence of the *Bifidobacterium animalis ssp. lactis* Strain A. In some embodiments, the bacterial formulation comprises a bacterium and/or a combination of bacteria described herein and a pharmaceutically acceptable carrier.

[0050] In certain embodiments, at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% of the bacteria in the bacterial composition are *Bifidobacterium animalis ssp. lactis.* In certain embodiments, substantially all of the bacteria in the bacterial composition are *Bifidobacterium animalis ssp. lactis.* In certain embodiments, the bacterial composition comprises at least 1×10^3 colony forming units (CFUs), 1×10^4 colony forming units (CFUs), 1×10^5 colony forming units (CFUs), 5×10^5 colony forming units (CFUs), 1×10^6 colony forming units (CFUs), 2×10^6 colony forming units (CFUs), 3×10^6 colony forming units (CFUs), 4×10^6 colony forming units (CFUs), 5×10^6 colony forming units (CFUs), 6×10^6 colony forming units (CFUs), 7×10^6 colony forming units

(CFUs), 8 x 10^6 colony forming units (CFUs), 9 x 10^6 colony forming units (CFUs), 1 x 10^7 colony forming units (CFUs), 2 x 10^7 colony forming units (CFUs), 3 x 10^7 colony forming units (CFUs), 4 x 10^7 colony forming units (CFUs), 5 x 10^7 colony forming units (CFUs), 6 x 10^7 colony forming units (CFUs), 7 x 10^7 colony forming units (CFUs), 8 x 10^7 colony forming units (CFUs), 9 x 10^7 colony forming units (CFUs), 1 x 10^8 colony forming units (CFUs), 2 x 10^8 colony forming units (CFUs), 3 x 10^8 colony forming units (CFUs), 4 x 10^8 colony forming units (CFUs), 5 x 10^8 colony forming units (CFUs), 6 x 10^8 colony forming units (CFUs), 7 x 10^8 colony forming units (CFUs), 8 x 10^8 colony forming units (CFUs), 9 x 10^8 colony forming units (CFUs), 1 x 10^9 colony forming units (CFUs), 5 x 10^9 colony forming units (CFUs), 1 x 10^{10} colony forming units (CFUs) 5 x 10^{10} colony forming units (CFUs), 1 x 10^{11} colony forming units (CFUs) 5 x 10^{11} colony forming units (CFUs), 1 x 10^{12} colony forming units (CFUs) 5 x 10^{12} colony forming units (CFUs), 1 x 10^{13} colony forming units (CFUs) of *Bifidobacterium animalis ssp. lactis.*

[0051] In some embodiments, Probiotic formulations containing *Bifidobacterium animalis ssp. lactis* are provided as encapsulated, enteric coated, or powder forms, with doses ranging up to 10^{11} cfu (e.g., upto 10^{10} cfu). In some embodiments, the composition comprises 5 x 10^{11} cfu of *Bifidobacterium animalis ssp. lactis* and 10% (w/w) corn starch in a capsule. The capsule is enteric coated for duodenal release at pH5.5 In some embodiments, the capsule is enteric coated for duodenal release at pH 5.5. In some embodiments, the composition comprises a powder of freeze-dried *Bifidobacterium animalis ssp. lactis* which is deemed "Qualified Presumption of Safety" (QPS) status. In some embodiments, the composition is stable at frozen or refrigerated temperature.

[0052] Methods for producing microbial compositions may include three main processing steps. The steps are: organism banking, organism production, and preservation. In certain embodiments, a sample that contains an abundance of *Bifidobacterium animalis ssp. lactis* may be cultured by avoiding an isolation step.

[0053] For banking, the strains included in the microbial composition may be (1) isolated directly from a specimen or taken from a banked stock, (2) optionally cultured on a nutrient agar or broth that supports growth to generate viable biomass, and (3) the biomass optionally preserved in multiple aliquots in long-term storage.

[0054] In embodiments using a culturing step, the agar or broth may contain nutrients that provide essential elements and specific factors that enable growth. An example would be a medium composed of 20 g/L glucose, 10 g/L yeast extract, 10 g/L soy peptone, 2 g/L citric acid, 1.5 g/L sodium phosphate monobasic, 100 mg/L ferric ammonium citrate, 80 mg/L magnesium sulfate, 10 mg/L hemin chloride, 2 mg/L calcium chloride, 1 mg/L menadione. Another example would be a medium composed of 10 g/L beef extract, 10 g/L peptone, 5 g/L sodium chloride, 5 g/L dextrose, 3 g/L yeast extract, 3 g/L sodium acetate, 1 g/L soluble starch, and 0.5 g/L L-cysteine HCl, at pH 6.8. A variety of microbiological media and variations are well known in the art (*e.g.*, R.M. Atlas, *Handbook of Microbiological Media* (2010) CRC Press). Culture media can be added to the culture at the start, may be added during the culture, or may be intermittently/continuously flowed through the culture. The strains in the bacterial composition may be cultivated alone, as a subset of the microbial composition, or as an entire collection comprising the microbial composition. As an example, a first strain may be cultivated together with a second strain in a mixed continuous culture, at a dilution rate lower than the maximum growth rate of either cell to prevent the culture from washing out of the cultivation.

[0055] The inoculated culture is incubated under favorable conditions for a time sufficient to build biomass. For microbial compositions for human use this is often at 37°C temperature, pH, and other parameter with values similar to the normal human niche. The environment may be actively controlled, passively controlled (*e.g.*, via buffers), or allowed to drift. For example, for anaerobic bacterial compositions, an anoxic/reducing environment may be employed. This can be accomplished by addition of reducing agents such as cysteine to the broth, and/or stripping it of oxygen. As an example, a culture of a bacterial composition may be grown at 37°C, pH 7, in the medium above, pre-reduced with 1 g/L cysteine-HCl.

[0056] When the culture has generated sufficient biomass, it may be preserved for banking. The organisms may be placed into a chemical milieu that protects from freezing (adding 'cryoprotectants'), drying ('lyoprotectants'), and/or osmotic shock ('osmoprotectants'), dispensing into multiple (optionally identical) containers to create a uniform bank, and then treating the culture for preservation. Containers are generally impermeable and have closures that assure isolation from the environment. Cryopreservation treatment is accomplished by freezing a liquid at ultra-low temperatures (*e.g.*, at or below -80°C). Dried preservation removes water from the culture by evaporation (in the case of spray drying or 'cool drying') or by

sublimation (*e.g.*, for freeze drying, spray freeze drying). Removal of water improves long-term microbial composition storage stability at temperatures elevated above cryogenic conditions. If the microbial composition comprises, for example, spore forming species and results in the production of spores, the final composition may be purified by additional means such as density gradient centrifugation and preserved using the techniques [?]described above[?]. Microbial composition banking may be done by culturing and preserving the strains individually, or by mixing the strains together to create a combined bank. As an example of cryopreservation, a microbial composition culture may be harvested by centrifugation to pellet the cells from the culture medium, the supernatant decanted and replaced with fresh culture broth containing 15% glycerol. The culture can then be aliquoted into 1 mL cryotubes, sealed, and placed at -80°C for long-term viability retention. This procedure achieves acceptable viability upon recovery from frozen storage.

[0057] Microbial production may be conducted using similar culture steps to banking, including medium composition and culture conditions described above. It may be conducted at larger scales of operation, especially for clinical development or commercial production. At larger scales, there may be several subcultivations of the microbial composition prior to the final cultivation. At the end of cultivation, the culture is harvested to enable further formulation into a dosage form for administration. This can involve concentration, removal of undesirable medium components, and/or introduction into a chemical milieu that preserves the microbial composition and renders it acceptable for administration via the chosen route. For example, a microbial composition may be cultivated to a concentration of 10^{10} CFU/mL, then concentrated 20-fold by tangential flow microfiltration; the spent medium may be exchanged by diafiltering with a preservative medium consisting of 2% gelatin, 100 mM trehalose, and 10 mM sodium phosphate buffer. The suspension can then be freeze-dried to a powder and titrated.

[0058] After drying, the powder may be blended to an appropriate potency, and mixed with other cultures and/or a filler such as microcrystalline cellulose for consistency and ease of handling, and the bacterial composition formulated as provided herein.

[0059] In certain aspects, provided are bacterial compositions for administration subjects. In some embodiments, the bacterial compositions are combined with additional active and/or inactive materials in order to produce a final product, which may be in single dosage unit or in a multi-dose format.

[0060] In some embodiments, the composition comprises at least one carbohydrate. A "carbohydrate" refers to a sugar or polymer of sugars. The terms "saccharide," "polysaccharide," "carbohydrate," and "oligosaccharide" may be used interchangeably. Most carbohydrates are aldehydes or ketones with many hydroxyl groups, usually one on each carbon atom of the molecule. Carbohydrates generally have the molecular formula $C_nH_{2n}O_n$. A carbohydrate may be a monosaccharide, a disaccharide, trisaccharide, oligosaccharide, or polysaccharide. The most basic carbohydrate is a monosaccharide, such as glucose, sucrose, galactose, mannose, ribose, arabinose, xylose, and fructose. Disaccharides are two joined monosaccharides. Exemplary disaccharides include sucrose, maltose, cellobiose, and lactose. Typically, an oligosaccharide includes between three and six monosaccharide units (*e.g.*, raffinose, stachyose), and polysaccharides include six or more monosaccharide units. Exemplary polysaccharides include starch, glycogen, and cellulose. Carbohydrates may contain modified saccharide units such as 2'-deoxyribose wherein a hydroxyl group is removed, 2'-fluororibose wherein a hydroxyl group is replaced with a fluorine, or N-acetylglucosamine, a nitrogen-containing form of glucose (*e.g.*, 2'-fluororibose, deoxyribose, and hexose). Carbohydrates may exist in many different forms, for example, conformers, cyclic forms, acyclic forms, stereoisomers, tautomers, anomers, and isomers.

[0061] In some embodiments, the composition comprises at least one lipid. As used herein, a "lipid" includes fats, oils, triglycerides, cholesterol, phospholipids, fatty acids in any form including free fatty acids. Fats, oils and fatty acids can be saturated, unsaturated (cis or trans) or partially unsaturated (cis or trans). In some embodiments the lipid comprises at least one fatty acid selected from lauric acid (12:0), myristic acid (14:0), palmitic acid (16:0), palmitoleic acid (16:1), margaric acid (17:0), heptadecenoic acid (17:1), stearic acid (18:0), oleic acid (18:1), linoleic acid (18:2), linolenic acid (18:3), octadecatetraenoic acid (18:4), arachidic acid (20:0), eicosenoic acid (20:1), eicosadienoic acid (20:2), eicosatetraenoic acid (20:4), eicosapentaenoic acid (20:5) (EPA), docosanoic acid (22:0), docosenoic acid (22:1), docosapentaenoic acid (22:5), docosahexaenoic acid (22:6) (DHA), and tetracosanoic acid (24:0). In some embodiments the composition comprises at least one modified lipid, for example a lipid that has been modified by cooking.

[0062] In some embodiments, the composition comprises at least one supplemental mineral or mineral source. Examples of minerals include, without limitation: chloride, sodium,

calcium, iron, chromium, copper, iodine, zinc, magnesium, manganese, molybdenum, phosphorus, potassium, and selenium. Suitable forms of any of the foregoing minerals include soluble mineral salts, slightly soluble mineral salts, insoluble mineral salts, chelated minerals, mineral complexes, non-reactive minerals such as carbonyl minerals, and reduced minerals, and combinations thereof.

[0063] In some embodiments, the composition comprises at least one supplemental vitamin. The at least one vitamin can be fat-soluble or water soluble vitamins. Suitable vitamins include but are not limited to vitamin C, vitamin A, vitamin E, vitamin B12, vitamin K, riboflavin, niacin, vitamin D, vitamin B6, folic acid, pyridoxine, thiamine, pantothenic acid, and biotin. Suitable forms of any of the foregoing are salts of the vitamin, derivatives of the vitamin, compounds having the same or similar activity of the vitamin, and metabolites of the vitamin.

[0064] In some embodiments, the composition comprises an excipient. Non-limiting examples of suitable excipients include a buffering agent, a preservative, a stabilizer, a binder, a compaction agent, a lubricant, a dispersion enhancer, a disintegration agent, a flavoring agent, a sweetener, and a coloring agent.

[0065] In some embodiments, the excipient is a buffering agent. Non-limiting examples of suitable buffering agents include sodium citrate, magnesium carbonate, magnesium bicarbonate, calcium carbonate, and calcium bicarbonate.

[0066] In some embodiments, the excipient comprises a preservative. Non-limiting examples of suitable preservatives include antioxidants, such as alpha-tocopherol and ascorbate, and antimicrobials, such as parabens, chlorobutanol, and phenol.

[0067] In some embodiments, the composition comprises a binder as an excipient. Non-limiting examples of suitable binders include starches, pregelatinized starches, gelatin, polyvinylpyrolidone, cellulose, methylcellulose, sodium carboxymethylcellulose, ethylcellulose, polyacrylamides, polyvinyloxoazolidone, polyvinylalcohols, C_{12}-C_{18} fatty acid alcohol, polyethylene glycol, polyols, saccharides, oligosaccharides, and combinations thereof.

[0068] In some embodiments, the composition comprises a lubricant as an excipient. Non-limiting examples of suitable lubricants include magnesium stearate, calcium stearate, zinc stearate, hydrogenated vegetable oils, sterotex, polyoxyethylene monostearate, talc, polyethyleneglycol, sodium benzoate, sodium lauryl sulfate, magnesium lauryl sulfate, and light mineral oil.

[0069] In some embodiments, the composition comprises a dispersion enhancer as an excipient. Non-limiting examples of suitable dispersants include starch, alginic acid, polyvinylpyrrolidones, guar gum, kaolin, bentonite, purified wood cellulose, sodium starch glycolate, isoamorphous silicate, and microcrystalline cellulose as high HLB emulsifier surfactants.

[0070] In some embodiments, the composition comprises a disintegrant as an excipient. In some embodiments the disintegrant is a non-effervescent disintegrant. Non-limiting examples of suitable non-effervescent disintegrants include starches such as corn starch, potato starch, pregelatinized and modified starches thereof, sweeteners, clays, such as bentonite, micro-crystalline cellulose, alginates, sodium starch glycolate, gums such as agar, guar, locust bean, karaya, pectin, and tragacanth. In some embodiments the disintegrant is an effervescent disintegrant. Non-limiting examples of suitable effervescent disintegrants include sodium bicarbonate in combination with citric acid, and sodium bicarbonate in combination with tartaric acid.

[0071] In some embodiments, the bacterial formulation comprises an enteric coating or micro encapsulation. In certain embodiments, the enteric coating or micro encapsulation improves targeting to a desired region of the gastrointestinal tract. For example, in certain embodiments, the bacterial composition comprises an enteric coating and/or microcapsules that dissolves at a pH associated with a particular region of the gastrointestinal tract. In some embodiments, the enteric coating and/or microcapsules dissolve at a pH of about 5.5 – 6.2 to release in the duodenum, at a pH value of about 7.2 – 7.5 to release in the ileum, and/or at a pH value of about 5.6 – 6.2 to release in the colon. Exemplary enteric coatings and microcapsules are described, for example, in U.S. Pat. Pub. No. 2016/0022592, which is hereby incorporated by reference in its entirety.

[0072] In some embodiments, the composition is a food product (*e.g.*, a food or beverage) such as a health food or beverage, a food or beverage for infants, a food or beverage for pregnant women, athletes, senior citizens or other specified group, a functional food, a beverage, a food or beverage for specified health use, a dietary supplement, a food or beverage for patients, or an animal feed. Specific examples of the foods and beverages include various beverages such as juices, refreshing beverages, tea beverages, drink preparations, jelly beverages, and functional beverages; alcoholic beverages such as beers; carbohydrate-containing

foods such as rice food products, noodles, breads, and pastas; paste products such as fish hams, sausages, paste products of seafood; retort pouch products such as curries, food dressed with a thick starchy sauces, and Chinese soups; soups; dairy products such as milk, dairy beverages, ice creams, cheeses, and yogurts; fermented products such as fermented soybean pastes, yogurts, fermented beverages, and pickles; bean products; various confectionery products, including biscuits, cookies, and the like, candies, chewing gums, gummies, cold desserts including jellies, cream caramels, and frozen desserts; instant foods such as instant soups and instant soy-bean soups; microwavable foods; and the like. Further, the examples also include health foods and beverages prepared in the forms of powders, granules, tablets, capsules, liquids, pastes, and jellies.

[0073] In certain embodiments, the bacteria disclosed herein are administered in conjunction with a prebiotic to the subject. Prebiotics are carbohydrates which are generally indigestible by a host animal and are selectively fermented or metabolized by bacteria. Prebiotics may be short-chain carbohydrates (*e.g.,* oligosaccharides) and/or simple sugars (*e.g.,* mono- and di-saccharides) and/or mucins (heavily glycosylated proteins) that alter the composition or metabolism of a microbiome in the host. The short chain carbohydrates are also referred to as oligosaccharides, and usually contain from 2 or 3 and up to 8, 9, 10, 15 or more sugar moieties. When prebiotics are introduced to a host, the prebiotics affect the bacteria within the host and do not directly affect the host. In certain aspects, a prebiotic composition can selectively stimulate the growth and/or activity of one of a limited number of bacteria in a host. Prebiotics include oligosaccharides such as fructooligosaccharides (FOS) (including inulin), galactooligosaccharides (GOS), trans-galactooligosaccharides, xylooligosaccharides (XOS), chitooligosaccharides (COS), soy oligosaccharides (*e.g.,* stachyose and raffinose) gentiooligosaccharides, isomaltooligosaccharides, mannooligosaccharides, maltooligosaccharides and mannanoligosaccharides. Oligosaccharides are not necessarily single components, and can be mixtures containing oligosaccharides with different degrees of oligomerization, sometimes including the parent disaccharide and the monomeric sugars. Various types of oligosaccharides are found as natural components in many common foods, including fruits, vegetables, milk, and honey. Specific examples of oligosaccharides are lactulose, lactosucrose, palatinose, glycosyl sucrose, guar gum, gum Arabic, tagalose, amylose,

amylopectin, pectin, xylan, and cyclodextrins. Prebiotics may also be purified or chemically or enzymatically synthesized.

Production of PhABs

[0074] In certain aspects, the PhABs described herein can be prepared using any method known in the art.

[0075] In some embodiments, the PhABs described herein are prepared by fractionation. Bacterial cells and/or supernatants from cultured bacteria cells are fractionated into various pharmacologically active biomass (PhABs) and/or products derived therefrom. Bacterial cells and/or supernatants are fractionated using materials and methods known in the art (see e.g. Sandrini et al. Fractionation by ultracentrifugation of gram negative cytoplasmic and membrane proteins. 2014. Bio-Protocol. 4(21); Scholler et al. Protoplast and cytoplasmic membrane preparations from Streptococcus sanguis and Streptococcus mutans. 1983. J Gen Micro. 129: 3271-3279; Thein et al. Efficient subfractionation of gram-negative bacteria for proteomics studies. 2010. Am Chem Society. 9: 6135-6147; Hobb et al. Evaluation of procedures for outer membrane isolation from Campylobacter jejuni. 2009. 155(Pt. 3): 979-988).

[0076] Additionally, PhABs obtained by methods provided herein may be further purified by size based column chromatography, by affinity chromatography, and by gradient ultracentrifugation, using methods that may include, but are not limited to, use of a sucrose gradient or Optiprep gradient. Briefly, using a sucrose gradient method, if ammonium sulfate precipitation or ultracentrifugation were used to concentrate the filtered supernatants, pellets are resuspended in 60% sucrose, 30 mM Tris, pH 8.0. If filtration was used to concentrate the filtered supernatant, the concentrate is buffer exchanged into 60% sucrose, 30 mM Tris, pH 8.0, using an Amicon Ultra column. Samples are applied to a 35-60% discontinuous sucrose gradient and centrifuged at 200,000 x g for 3-24 hours at 4°C. Briefly, using an Optiprep gradient method, if ammonium sulfate precipitation or ultracentrifugation were used to concentrate the filtered supernatants, pellets are resuspended in 35% Optiprep in PBS. In some embodiments, if filtration was used to concentrate the filtered supernatant, the concentrate is diluted using 60% Optiprep to a final concentration of 35% Optiprep. Samples are applied to a 35-60% discontinuous sucrose gradient and centrifuged at 200,000 x g for 3-24 hours at 4°C.

[0077] In some embodiments, to confirm sterility and isolation of the PhAB preparations, PhABs are serially diluted onto agar medium used for routine culture of the bacteria being tested, and incubated using routine conditions. Non-sterile preparations are passed through a 0.22 um filter to exclude intact cells. To further increase purity, isolated PhABs may be DNase or proteinase K treated.

[0078] In some embodiments, for preparation of PhABs used for *in vivo* injections, purified PhABs are processed as described previously (G. Norheim, et al. *PLoS ONE.* 10(9): e0134353 (2015)). Briefly, after sucrose gradient centrifugation, bands containing PhABs are resuspended to a final concentration of 50 μg/mL in a solution containing 3% sucrose or other solution suitable for *in vivo* injection known to one skilled in the art. This solution may also contain adjuvant, for example aluminum hydroxide at a concentration of 0-0.5% (w/v).

[0079] In certain embodiments, to make samples compatible with further testing (e.g. to remove sucrose prior to TEM imaging or *in vitro* assays), samples are buffer exchanged into PBS or 30 mM Tris, pH 8.0 using filtration (e.g. Amicon Ultra columns), dialysis, or ultracentrifugation (200,000 x g, ≥ 3 hours, 4°C) and resuspension.

[0080] In some embodiments, the sterility of the PhAB preparations can be confirmed by plating a portion of the PhABs onto agar medium used for standard culture of the bacteria used in the generation of the PhABs and incubating using standard conditions.

[0081] In some embodiments select PhABs are isolated and enriched by chromatography and binding surface moieties on PhABs. In other embodiments, select PhABs are isolated and/or enriched by fluorescent cell sorting by methods using affinity reagents, chemical dyes, recombinant proteins or other methods known to one skilled in the art.

Administration

[0082] In certain aspects, provided herein is a method of delivering a bacterium and/or a bacterial composition described herein to a subject. In some embodiments of the methods provided herein, the bacteria are administered in conjunction with the administration of a cancer therapeutic. In some embodiments, the bacteria is co-formulated in a pharmaceutical composition with the cancer therapeutic. In some embodiments, the bacteria is co-administered with the cancer therapeutic. In some embodiments, the cancer therapeutic is administered to the subject before administration of the bacteria (*e.g.*, about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 30,

35, 40, 45, 50 or 55 minutes before, about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22 or 23 hours before, or about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13 or 14 days before). In some embodiments, the cancer therapeutic is administered to the subject after administration of the bacteria (*e.g.*, about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 30, 35, 40, 45, 50 or 55 minutes after, about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22 or 23 hours after, or about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13 or 14 days after). In some embodiments, the same mode of delivery is used to deliver both the bacteria and the cancer therapeutic. In some embodiments different modes of delivery are used to administer the bacteria and the cancer therapeutic. For example, in some embodiments, the bacteria is administered orally while the cancer therapeutic is administered via injection (*e.g.*, an intravenious, intramuscular and/or intratumoral injection).

[0083] In certain embodiments, the pharmaceutical compositions, dosage forms, and kits described herein can be administered in conjunction with any other conventional anti-cancer treatment, such as, for example, radiation therapy and surgical resection of the tumor. These treatments may be applied as necessary and/or as indicated and may occur before, concurrent with or after administration of the pharmaceutical compositions, dosage forms, and kits described herein.

[0084] The dosage regimen can be any of a variety of methods and amounts, and can be determined by one skilled in the art according to known clinical factors. As is known in the medical arts, dosages for any one patient can depend on many factors, including the subject's species, size, body surface area, age, sex, immunocompetence, and general health, the particular microorganism to be administered, duration and route of administration, the kind and stage of the disease, for example, tumor size, and other compounds such as drugs being administered concurrently. In addition to the above factors, such levels can be affected by the infectivity of the microorganism, and the nature of the microorganism, as can be determined by one skilled in the art. In the present methods, appropriate minimum dosage levels of microorganisms can be levels sufficient for the microorganism to survive, grow and replicate in a tumor or metastasis. The methods of treatment described herein may be suitable for the treatment of a primary tumor, a secondary tumor or metastasis, as well as for recurring tumors or cancers. The dose of the pharmaceutical compositions described herein may be appropriately set or adjusted in accordance with the dosage form, the route of administration, the degree or stage of a target

disease, and the like. For example, the general effective dose of the agents may range between 0.01 mg/kg body weight/day and 1000 mg/kg body weight/day, between 0.1 mg/kg body weight/day and 1000 mg/kg body weight/day, 0.5 mg/kg body weight/day and 500 mg/kg body weight/day, 1 mg/kg body weight/day and 100 mg/kg body weight/day, or between 5 mg/kg body weight/day and 50 mg/kg body weight/day. The effective dose may be 0.01, 0.05, 0.1, 0.5, 1, 2, 3, 5, 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 200, 500, or 1000 mg/kg body weight/day or more, but the dose is not limited thereto.

[0085] In some embodiments, the dose administered to a subject is sufficient to prevent cancer, delay its onset, or slow or stop its progression or prevent a relapse of a cancer. One skilled in the art will recognize that dosage will depend upon a variety of factors including the strength of the particular compound employed, as well as the age, species, condition, and body weight of the subject. The size of the dose will also be determined by the route, timing, and frequency of administration as well as the existence, nature, and extent of any adverse side-effects that might accompany the administration of a particular compound and the desired physiological effect.

[0086] Suitable doses and dosage regimens can be determined by conventional range-finding techniques known to those of ordinary skill in the art. Generally, treatment is initiated with smaller dosages, which are less than the optimum dose of the compound. Thereafter, the dosage is increased by small increments until the optimum effect under the circumstances is reached. An effective dosage and treatment protocol can be determined by routine and conventional means, starting *e.g.,* with a low dose in laboratory animals and then increasing the dosage while monitoring the effects, and systematically varying the dosage regimen as well. Animal studies are commonly used to determine the maximal tolerable dose ("MTD") of bioactive agent per kilogram weight. Those skilled in the art regularly extrapolate doses for efficacy, while avoiding toxicity, in other species, including humans.

[0087] In accordance with the above, in therapeutic applications, the dosages of the active agents used in accordance with the invention vary depending on the active agent, the age, weight, and clinical condition of the recipient patient, and the experience and judgment of the clinician or practitioner administering the therapy, among other factors affecting the selected dosage. Generally, the dose should be sufficient to result in slowing, and preferably regressing, the growth of the tumors and most preferably causing complete regression of the cancer.

[0088] Separate administrations can include any number of two or more administrations (e.g., doses), including two, three, four, five or six administrations. One skilled in the art can readily determine the number of administrations to perform, or the desirability of performing one or more additional administrations, according to methods known in the art for monitoring therapeutic methods and other monitoring methods provided herein. In some embodiments, the doses may be separated by at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29 or 30 days or 1, 2, 3, or 4 weeks. Accordingly, the methods provided herein include methods of providing to the subject one or more administrations of a bacterium, where the number of administrations can be determined by monitoring the subject, and, based on the results of the monitoring, determining whether or not to provide one or more additional administrations. Deciding on whether or not to provide one or more additional administrations can be based on a variety of monitoring results, including, but not limited to, indication of tumor growth or inhibition of tumor growth, appearance of new metastases or inhibition of metastasis, the subject's anti-bacterium antibody titer, the subject's anti-tumor antibody titer, the overall health of the subject and/or the weight of the subject.

[0089] The time period between administrations can be any of a variety of time periods. The time period between administrations can be a function of any of a variety of factors, including monitoring steps, as described in relation to the number of administrations, the time period for a subject to mount an immune response and/or the time period for a subject to clear the bacteria from normal tissue. In one example, the time period can be a function of the time period for a subject to mount an immune response; for example, the time period can be more than the time period for a subject to mount an immune response, such as more than about one week, more than about ten days, more than about two weeks, or more than about a month; in another example, the time period can be less than the time period for a subject to mount an immune response, such as less than about one week, less than about ten days, less than about two weeks, or less than about a month. In another example, the time period can be a function of the time period for a subject to clear the bacteria from normal tissue; for example, the time period can be more than the time period for a subject to clear the bacteria from normal tissue, such as more than about a day, more than about two days, more than about three days, more than about five days, or more than about a week.

[0090] In some embodiments, the delivery of a cancer therapeutic in combination with the bacteria described herein reduces the adverse effects and/or improves the efficacy of the cancer therapeutic.

[0091] The effective dose of a cancer therapeutic described herein is the amount of the therapeutic agent that is effective to achieve the desired therapeutic response for a particular patient, composition, and mode of administration, with the least toxicity to the patient. The effective dosage level can be identified using the methods described herein and will depend upon a variety of pharmacokinetic factors including the activity of the particular compositions administered, the route of administration, the time of administration, the rate of excretion of the particular compound being employed, the duration of the treatment, other drugs, compounds and/or materials used in combination with the particular compositions employed, the age, sex, weight, condition, general health and prior medical history of the patient being treated, and like factors well known in the medical arts. In general, an effective dose of a cancer therapy will be the amount of the therapeutic agent which is the lowest dose effective to produce a therapeutic effect. Such an effective dose will generally depend upon the factors described above.

[0092] The toxicity of a cancer therapy is the level of adverse effects experienced by the subject during and following treatment. Adverse events associated with cancer therapy toxicity include, but are not limited to, abdominal pain, acid indigestion, acid reflux, allergic reactions, alopecia, anaphylaxis, anemia, anxiety, lack of appetite, arthralgias, asthenia, ataxia, azotemia, loss of balance, bone pain, bleeding, blood clots, low blood pressure, elevated blood pressure, difficulty breathing, bronchitis, bruising, low white blood cell count, low red blood cell count, low platelet count, cardiotoxicity, cystitis, hemorrhagic cystitis, arrhythmias, heart valve disease, cardiomyopathy, coronary artery disease, cataracts, central neurotoxicity, cognitive impairment, confusion, conjunctivitis, constipation, coughing, cramping, cystitis, deep vein thrombosis, dehydration, depression, diarrhea, dizziness, dry mouth, dry skin, dyspepsia, dyspnea, edema, electrolyte imbalance, esophagitis, fatigue, loss of fertility, fever, flatulence, flushing, gastric reflux, gastroesophageal reflux disease, genital pain, granulocytopenia, gynecomastia, glaucoma, hair loss, hand-foot syndrome, headache, hearing loss, heart failure, heart palpitations, heartburn, hematoma, hemorrhagic cystitis, hepatotoxicity, hyperamylasemia, hypercalcemia, hyperchloremia, hyperglycemia, hyperkalemia, hyperlipasemia, hypermagnesemia, hypernatremia, hyperphosphatemia, hyperpigmentation, hypertriglyceridemia, hyperuricemia,

hypoalbuminemia, hypocalcemia, hypochloremia, hypoglycemia, hypokalemia, hypomagnesemia, hyponatremia, hypophosphatemia, impotence, infection, injection site reactions, insomnia, iron deficiency, itching, joint pain, kidney failure, leukopenia, liver dysfunction, memory loss, menopause, mouth sores, mucositis, muscle pain, myalgias, myelosuppression, myocarditis, neutropenic fever, nausea, nephrotoxicity, neutropenia, nosebleeds, numbness, ototoxicity, pain, palmar-plantar erythrodysesthesia, pancytopenia, pericarditis, peripheral neuropathy, pharyngitis, photophobia, photosensitivity, pneumonia, pneumonitis, proteinuria, pulmonary embolus, pulmonary fibrosis, pulmonary toxicity, rash, rapid heart beat, rectal bleeding, restlessness, rhinitis, seizures, shortness of breath, sinusitis, thrombocytopenia, tinnitus, urinary tract infection, vaginal bleeding, vaginal dryness, vertigo, water retention, weakness, weight loss, weight gain, and xerostomia. In general, toxicity is acceptable if the benefits to the subject achieved through the therapy outweigh the adverse events experienced by the subject due to the therapy.

[0093] In some embodiments, the administration of the bacterial composition treats the cancer. In some embodiments, the bacterial composition induces an anti-tumor immune response in the subject.

Therapeutic Agents

[0094] In certain aspects, the methods provided herein include the administration to a subject of a bacterium and/or a bacterial composition described herein (*e.g.*, a *Bifidobacterium animalis ssp. Lactis*-containing bacterial composition) either alone or in combination with another cancer therapeutic. The other cancer therapeutic may include *e.g.*, surgical resection, radiotherapy, or a cancer therapeutic agent. In some embodiments, the bacterial composition and the other cancer therapy can be administered to the subject in any order. In some embodiments, the bacterial composition and the other cancer therapy are administered conjointly.

[0095] In some embodiments the bacterium is administered to the subject before the cancer therapeutic is administered (*e.g.*, at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23 or 24 hours before or at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29 or 30 days before). In some embodiments the bacterium is administered to the subject after the cancer therapeutic is administered (*e.g.*, at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23 or 24 hours after

or at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29 or 30 days after). In some embodiments, the bacterium and the cancer therapeutic are administered to the subject simultaneously or nearly simultaneously (*e.g.*, administrations occur within an hour of each other). In some embodiments, the subject is administered an antibiotic before the bacterium is administered to the subject (*e.g.*, at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23 or 24 hours before or at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29 or 30 days before).In some embodiments, the subject is administered an antibiotic after the bacterium is administered to the subject (*e.g.*, at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23 or 24 hours before or at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29 or 30 days after). In some embodiments, the bacterium and the antibiotic are administered to the subject simultaneously or nearly simultaneously (*e.g.*, administrations occur within an hour of each other).

[0096] In certain embodiments, the subject may undergo surgery. Types of surgery include but are not limited to preventative, diagnostic or staging, curative and palliative surgery. Curative surgery is a cancer treatment that may be used in conjunction with other therapies, such as the treatment of the present invention, chemotherapy, radiotherapy, hormonal therapy, gene therapy, immunotherapy and/or alternative therapies.

[0097] Curative surgery includes resection in which all or part of cancerous tissue is physically removed, excised, and/or destroyed. Tumor resection refers to physical removal of at least part of a tumor. In addition to tumor resection, treatment by surgery includes laser surgery, cryosurgery, electrosurgery, and microscopically controlled surgery (Mohs' surgery). Upon excision of part of all of cancerous cells, tissue, or tumor, a cavity may be formed in the body.

[0098] In certain embodiments, the subject may undergo radiation therapy. Radiation therapy includes the administration or application of a radiotherapeutic agents and factors including but not limited to In addition to trays, UV-irradiation, microwaves, electronic emissions, and radioisotopes. The localized tumor site may be irradiated, including by one or more the above described forms of radiations. All of these factors may effect a broad range of damage DNA, on the precursors of DNA, the replication and repair of DNA, and the assembly and maintenance of chromosomes.

[0099] Dosage ranges for X-rays range from daily doses of 50 to 200 roentgens for prolonged periods of time (3 to 4 weeks), to single doses of 2000 to 6000 roentgens. Dosage ranges for radioisotopes vary widely, and depend on the half-life of the isotope, the strength and type of radiation emitted, and the uptake by the neoplastic cells.

[00100] In certain aspects, the methods provided herein further comprise administering another cancer therapeutic to the subject.

[00101] In some embodiments, the cancer therapeutic is a chemotherapeutic agent. Examples of such chemotherapeutic agents include, but are not limited to, alkylating agents such as thiotepa and cyclosphosphamide; alkyl sulfonates such as busulfan, improsulfan and piposulfan; aziridines such as benzodopa, carboquone, meturedopa, and uredopa; ethylenimines and methylamelamines including altretamine, triethylenemelamine, trietylenephosphoramide, triethiylenethiophosphoramide and trimethylolomelamine; acetogenins (especially bullatacin and bullatacinone); a camptothecin (including the synthetic analogue topotecan); bryostatin; callystatin; CC-1065 (including its adozelesin, carzelesin and bizelesin synthetic analogues); cryptophycins (particularly cryptophycin 1 and cryptophycin 8); dolastatin; duocarmycin (including the synthetic analogues, KW-2189 and CB1-TM1); eleutherobin; pancratistatin; a sarcodictyin; spongistatin; nitrogen mustards such as chlorambucil, chlornaphazine, cholophosphamide, estramustine, ifosfamide, mechlorethamine, mechlorethamine oxide hydrochloride, melphalan, novembichin, phenesterine, prednimustine, trofosfamide, uracil mustard; nitrosureas such as carmustine, chlorozotocin, fotemustine, lomustine, nimustine, and ranimnustine; antibiotics such as the enediyne antibiotics (*e.g.*, calicheamicin, especially calicheamicin gammaII and calicheamicin omegaI1; dynemicin, including dynemicin A; bisphosphonates, such as clodronate; an esperamicin; as well as neocarzinostatin chromophore and related chromoprotein enediyne antibiotic chromophores, aclacinomysins, actinomycin, authrarnycin, azaserine, bleomycins, cactinomycin, carabicin, caminomycin, carzinophilin, chromomycinis, dactinomycin, daunorubicin, detorubicin, 6-diazo-5-oxo-L-norleucine, doxorubicin (including morpholino-doxorubicin, cyanomorpholino-doxorubicin, 2-pyrrolino-doxorubicin and deoxydoxorubicin), epirubicin, esorubicin, idarubicin, marcellomycin, mitomycins such as mitomycin C, mycophenolic acid, nogalamycin, olivomycins, peplomycin, potfiromycin, puromycin, quelamycin, rodorubicin, streptonigrin, streptozocin, tubercidin, ubenimex, zinostatin, zorubicin; anti-metabolites such as methotrexate and 5-fluorouracil (5-

FU); folic acid analogues such as denopterin, methotrexate, pteropterin, trimetrexate; purine analogs such as fludarabine, 6-mercaptopurine, thiamiprine, thioguanine; pyrimidine analogs such as ancitabine, azacitidine, 6-azauridine, carmofur, cytarabine, dideoxyuridine, doxifluridine, enocitabine, floxuridine; androgens such as calusterone, dromostanolone propionate, epitiostanol, mepitiostane, testolactone; anti-adrenals such as aminoglutethimide, mitotane, trilostane; folic acid replenisher such as frolinic acid; aceglatone; aldophosphamide glycoside; aminolevulinic acid; eniluracil; amsacrine; bestrabucil; bisantrene; edatraxate; defofamine; demecolcine; diaziquone; elformithine; elliptinium acetate; an epothilone; etoglucid; gallium nitrate; hydroxyurea; lentinan; lonidainine; maytansinoids such as maytansine and ansamitocins; mitoguazone; mitoxantrone; mopidanmol; nitraerine; pentostatin; phenamet; pirarubicin; losoxantrone; podophyllinic acid; 2-ethylhydrazide; procarbazine; PSK polysaccharide complex); razoxane; rhizoxin; sizofuran; spirogermanium; tenuazonic acid; triaziquone; 2,2',2"-trichlorotriethylamine; trichothecenes (especially T-2 toxin, verracurin A, roridin A and anguidine); urethan; vindesine; dacarbazine; mannomustine; mitobronitol; mitolactol; pipobroman; gacytosine; arabinoside ("Ara-C"); cyclophosphamide; thiotepa; taxoids, *e.g.*, paclitaxel and doxetaxel; chlorambucil; gemcitabine; 6-thioguanine; mercaptopurine; methotrexate; platinum coordination complexes such as cisplatin, oxaliplatin and carboplatin; vinblastine; platinum; etoposide (VP-16); ifosfamide; mitoxantrone; vincristine; vinorelbine; novantrone; teniposide; edatrexate; daunomycin; aminopterin; xeloda; ibandronate; irinotecan (*e.g.*, CPT-11); topoisomerase inhibitor RFS 2000; difluoromethylomithine (DMFO); retinoids such as retinoic acid; capecitabine; and pharmaceutically acceptable salts, acids or derivatives of any of the above.

[00102] In some embodiments, the cancer therapeutic is a cancer immunotherapy agent. Immunotherapy refers to a treatment that uses a subject's immune system to treat cancer, *e.g.*, checkpoint inhibitors, cancer vaccines, cytokines, cell therapy, CAR-T cells, and dendritic cell therapy. Non-limiting examples of immunotherapies are checkpoint inhibitors include Nivolumab (BMS, anti-PD-1), Pembrolizumab (Merck, anti-PD-1), Ipilimumab (BMS, anti-CTLA-4), MEDI4736 (AstraZeneca, anti-PD-L1), and MPDL3280A (Roche, anti-PD-L1). Other immunotherapies may be tumor vaccines, such as Gardail, Cervarix, BCG, sipulencel-T, Gp100:209-217, AGS-003, DCVax-L, Algenpantucel-L, Tergenpantucel-L, TG4010, ProstAtak, Prostvac-V/R-TRICOM, Rindopepimul, E75 peptide acetate, IMA901, POL-103A,

Belagenpumatucel-L, GSK1572932A, MDX-1279, GV1001, and Tecemotide. Immunotherapy may be administered via injection (*e.g.*, intravenously, intratumorally, subcutaneously, or into lymph nodes), but may also be administered orally, topically, or via aerosol. Immunotherapies may comprise adjuvants such as cytokines. .

[00103] In some embodiments, the immunotherapy agent is an immune checkpoint inhibitor. Immune checkpoint inhibition broadly refers to inhibiting the checkpoints that cancer cells can produce to prevent or downregulate an immune response. Examples of immune checkpoint proteins include, but are not limited to, CTLA4, PD-1, PD-L1, PD-L2, A2AR, B7-H3, B7-H4, BTLA, KIR, LAG3, TIM-3 or VISTA. Immune checkpoint inhibitors can be antibodies or antigen binding fragments thereof that bind to and inhibit an immune checkpoint protein. Examples of immune checkpoint inhibitors include, but are not limited to, nivolumab, pembrolizumab, pidilizumab, AMP-224, AMP-514, STI-A1110, TSR-042, RG-7446, BMS-936559, MEDI-4736, MSB-0020718C, AUR-012 and STI-A1010. In some embodiments, the cancer immunotherapy includes administering an additional immune checkpoint inhibitor (e.g., administering 2, 3, 4, or 5, immune checkpoint inhibitors) to the subject

[00104] In certain embodiments, immune checkpoint inhibitors can be an inhibitory nucleic acid molecule (e.g., an siRNA molecule, an shRNA molecule or an antisense RNA molecule) that inhibits expression fo an immune checkpoint protein that inhibits expression of an immune checkpoint protein.

[00105] In some embodiments, the immune checkpoint inhibitor is a siRNA molecule. Such siRNA molecules should include a region of sufficient homology to the target region, and be of sufficient length in terms of nucleotides, such that the siRNA molecule down-regulate target RNA (e.g., RNA of an immune checkpoint protein). The term "ribonucleotide" or "nucleotide" can, in the case of a modified RNA or nucleotide surrogate, also refer to a modified nucleotide, or surrogate replacement moiety at one or more positions. It is not necessary that there be perfect complementarity between the siRNA molecule and the target, but the correspondence must be sufficient to enable the siRNA molecule to direct sequence-specific silencing, such as by RNAi cleavage of the target RNA. In some embodiments, the sense strand need only be sufficiently complementary with the antisense strand to maintain the overall double-strand character of the molecule.

[00106] In addition, an siRNA molecule may be modified or include nucleoside surrogates. Single stranded regions of an siRNA molecule may be modified or include nucleoside surrogates, *e.g.*, the unpaired region or regions of a hairpin structure, *e.g.*, a region which links two complementary regions, can have modifications or nucleoside surrogates. Modification to stabilize one or more 3′- or 5′-terminus of an siRNA molecule, *e.g.*, against exonucleases, or to favor the antisense siRNA agent to enter into RISC are also useful. Modifications can include C3 (or C6, C7, C12) amino linkers, thiol linkers, carboxyl linkers, non-nucleotidic spacers (C3, C6, C9, C12, abasic, triethylene glycol, hexaethylene glycol), special biotin or fluorescein reagents that come as phosphoramidites and that have another DMT-protected hydroxyl group, allowing multiple couplings during RNA synthesis.

[00107] Each strand of an siRNA molecule can be equal to or less than 35, 30, 25, 24, 23, 22, 21, or 20 nucleotides in length. In some embodiments, the strand is at least 19 nucleotides in length. For example, each strand can be between 21 and 25 nucleotides in length. In some embodiments, siRNA agents have a duplex region of 17, 18, 19, 29, 21, 22, 23, 24, or 25 nucleotide pairs, and one or more overhangs, such as one or two 3′ overhangs, of 2-3 nucleotides.

[00108] In some embodiments, the immune checkpoint inhibitor is a shRNA molecule. A "small hairpin RNA" or "short hairpin RNA" or "shRNA" includes a short RNA sequence that makes a tight hairpin turn that can be used to silence gene expression via RNA interference. The shRNAs provided herein may be chemically synthesized or transcribed from a transcriptional cassette in a DNA plasmid. The shRNA hairpin structure is cleaved by the cellular machinery into siRNA, which is then bound to the RNA-induced silencing complex (RISC).

[00109] In some embodiments, shRNAs are about 15-60, 15-50, or 15-40 (duplex) nucleotides in length, about 15-30, 15-25, or 19-25 (duplex) nucleotides in length, or are about 20-24, 21-22, or 21-23 (duplex) nucleotides in length (*e.g.*, each complementary sequence of the double-stranded shRNA is 15-60, 15-50, 15-40, 15-30, 15-25, or 19-25 nucleotides in length, or about 20-24, 21-22, or 21-23 nucleotides in length, and the double-stranded shRNA is about 15-60, 15-50, 15-40, 15-30, 15-25, or 19-25 base pairs in length, or about 18-22, 19-20, or 19-21 base pairs in length). shRNA duplexes may comprise 3' overhangs of about 1 to about 4 nucleotides or about 2 to about 3 nucleotides on the antisense strand and/or 5'-phosphate termini on the sense strand. In some embodiments, the shRNA comprises a sense strand and/or antisense strand sequence of from about 15 to about 60 nucleotides in length (*e.g.*, about 15-60, 15-55, 15-

50, 15-45, 15-40, 15-35, 15-30, or 15-25 nucleotides in length),or from about 19 to about 40 nucleotides in length (*e.g.*, about 19-40, 19-35, 19-30, or 19-25 nucleotides in length), or from about 19 to about 23 nucleotides in length (*e.g.*, 19, 20, 21, 22, or 23 nucleotides in length).

[00110] Non-limiting examples of shRNA include a double-stranded polynucleotide molecule assembled from a single-stranded molecule, where the sense and antisense regions are linked by a nucleic acid-based or non-nucleic acid-based linker; and a double-stranded polynucleotide molecule with a hairpin secondary structure having self-complementary sense and antisense regions. In some embodiments, the sense and antisense strands of the shRNA are linked by a loop structure comprising from about 1 to about 25 nucleotides, from about 2 to about 20 nucleotides, from about 4 to about 15 nucleotides, from about 5 to about 12 nucleotides, or 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, or more nucleotides.

[00111] Additional embodiments related to the shRNAs, as well as methods of designing and synthesizing such shRNAs, are described in U.S. patent application publication number 2011/0071208, the disclosure of which is herein incorporated by reference in its entirety for all purposes.

[00112] In some embodiments, the immune checkpoint inhibitor is an antisense oligonucleotide compounds that inhbits expression of an immune checkpoint protein. In certain embodiments, the degree of complementarity between the target sequence and antisense targeting sequence is sufficient to form a stable duplex. The region of complementarity of the antisense oligonucleotides with the target RNA sequence may be as short as 8-11 bases, but can be 12-15 bases or more, e.g., 10-40 bases, 12-30 bases, 12-25 bases, 15-25 bases, 12-20 bases, or 15-20 bases, including all integers in between these ranges. An antisense oligonucleotide of about 14-15 bases is generally long enough to have a unique complementary sequence.

[00113] In certain embodiments, antisense oligonucleotides may be 100% complementary to the target sequence, or may include mismatches, *e.g.*, to improve selective targeting of allele containing the disease-associated mutation, as long as a heteroduplex formed between the oligonucleotide and target sequence is sufficiently stable to withstand the action of cellular nucleases and other modes of degradation which may occur *in vivo*. Hence, certain oligonucleotides may have about or at least about 70% sequence complementarity, *e.g.*, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%,

87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or 100% sequence complementarity, between the oligonucleotide and the target sequence. Oligonucleotide backbones that are less susceptible to cleavage by nucleases are discussed herein. Mismatches, if present, are typically less destabilizing toward the end regions of the hybrid duplex than in the middle. The number of mismatches allowed will depend on the length of the oligonucleotide, the percentage of G:C base pairs in the duplex, and the position of the mismatch(es) in the duplex, according to well understood principles of duplex stability.

[00114] The inhibitory nucleic acid molecule can be prepared, for example, by chemical synthesis, in vitro transcription, or digestion of long dsRNA by Rnase III or Dicer. These can be introduced into cells by transfection, electroporation, or other methods known in the art. See Hannon, GJ, 2002, RNA Interference, Nature 418: 244-251; Bernstein E et al., 2002, The rest is silence. RNA 7: 1509-1521; Hutvagner G et al., RNAi: Nature abhors a double-strand. Curr. Opin. Genetics & Development 12: 225-232; Brummelkamp, 2002, A system for stable expression of short interfering RNAs in mammalian cells. Science 296: 550-553; Lee NS, Dohjima T, Bauer G, Li H, Li M-J, Ehsani A, Salvaterra P, and Rossi J. (2002). Expression of small interfering RNAs targeted against HIV-1 rev transcripts in human cells. Nature Biotechnol. 20:500-505; Miyagishi M, and Taira K. (2002). U6-promoter-driven siRNAs with four uridine 3' overhangs efficiently suppress targeted gene expression in mammalian cells. Nature Biotechnol. 20:497-500; Paddison PJ, Caudy AA, Bernstein E, Hannon GJ, and Conklin DS. (2002). Short hairpin RNAs (shRNAs) induce sequence-specific silencing in mammalian cells. Genes & Dev. 16:948-958; Paul CP, Good PD, Winer I, and Engelke DR. (2002). Effective expression of small interfering RNA in human cells. Nature Biotechnol. 20:505-508; Sui G, Soohoo C, Affar E-B, Gay F, Shi Y, Forrester WC, and Shi Y. (2002). A DNA vector-based RNAi technology to suppress gene expression in mammalian cells. Proc. Natl. Acad. Sci. USA 99(6):5515-5520; Yu J-Y, DeRuiter SL, and Turner DL. (2002). RNA interference by expression of short-interfering RNAs and hairpin RNAs in mammalian cells. Proc. Natl. Acad. Sci. USA 99(9):6047-6052.

[00115] In the present methods, the inhibitory nucleic acid molecule can be administered to the subject, for example, as naked nucleic acid, in combination with a delivery reagent, and/or as a nucleic acid comprising sequences that express an interfering nucleic acid molecule. In some embodiments the nucleic acid comprising sequences that express the interfering nucleic acid molecules are delivered within vectors, *e.g.* plasmid, viral and bacterial vectors. Any nucleic acid

delivery method known in the art can be used in the methods described herein. Suitable delivery reagents include, but are not limited to, *e.g.*, the Mirus Transit TKO lipophilic reagent; lipofectin; lipofectamine; cellfectin; polycations (*e.g.*, polylysine), atelocollagen, nanoplexes and liposomes. The use of atelocollagen as a delivery vehicle for nucleic acid molecules is described in Minakuchi *et al.* Nucleic Acids Res., 32(13):e109 (2004); Hanai *et al.* Ann NY Acad Sci., 1082:9-17 (2006); and Kawata *et al.* Mol Cancer Ther., 7(9):2904-12 (2008); each of which is incorporated herein in their entirety. Exemplary interfering nucleic acid delivery systems are provided in U.S. Patent Nos. 8,283,461, 8,313,772, 8,501,930. 8,426,554, 8,268,798 and 8,324,366, each of which is hereby incorporated by reference in its entirety.

[00116] In some embodiments, the immunotherapy agent is an antibody or antigen binding fragment thereof that, for example, binds to a cancer-associated antigen. Examples of cancer-associated antigens include, but are not limited to, adipophilin, AIM-2, ALDH1A1, alpha-actinin-4, alpha-fetoprotein ("AFP"), ARTC1, B-RAF, BAGE-1, BCLX (L), BCR-ABL fusion protein b3a2, beta-catenin, BING-4, CA-125, CALCA, carcinoembryonic antigen ("CEA"), CASP-5, CASP-8, CD274, CD45, Cdc27, CDK12, CDK4, CDKN2A, CEA, CLPP, COA-1, CPSF, CSNK1A1, CTAG1, CTAG2, cyclin D1, Cyclin-A1, dek-can fusion protein, DKK1, EFTUD2, Elongation factor 2, ENAH (hMena), Ep-CAM, EpCAM, EphA3, epithelial tumor antigen ("ETA"), ETV6-AML1 fusion protein, EZH2, FGF5, FLT3-ITD, FN1, G250/MN/CAIX, GAGE-1,2,8, GAGE-3,4,5,6,7, GAS7, glypican-3, GnTV, gp100/Pmel17, GPNMB, HAUS3, Hepsin, HER-2/neu, HERV-K-MEL, HLA-A11, HLA-A2, HLA-DOB, hsp70-2, IDO1, IGF2B3, IL13Ralpha2, Intestinal carboxyl esterase, K-ras, Kallikrein 4, KIF20A, KK-LC-1, KKLC1, KM-HN-1, KMHN1 also known as CCDC110, LAGE-1, LDLR-fucosyltransferaseAS fusion protein, Lengsin, M-CSF, MAGE-A1, MAGE-A10, MAGE-A12, MAGE-A2, MAGE-A3, MAGE-A4, MAGE-A6, MAGE-A9, MAGE-C1, MAGE-C2, malic enzyme, mammaglobin-A, MART2, MATN, MC1R, MCSP, mdm-2, ME1, Melan-A/MART-1, Meloe, Midkine, MMP-2, MMP-7, MUC1, MUC5AC, mucin, MUM-1, MUM-2, MUM-3, Myosin, Myosin class I, N-raw, NA88-A, neo-PAP, NFYC, NY-BR-1, NY-ESO-1/LAGE-2, OA1, OGT, OS-9, P polypeptide, p53, PAP, PAX5, PBF, pml-RARalpha fusion protein, polymorphic epithelial mucin ("PEM"), PPP1R3B, PRAME, PRDX5, PSA, PSMA, PTPRK, RAB38/NY-MEL-1, RAGE-1, RBAF600, RGS5, RhoC, RNF43, RU2AS, SAGE, secernin 1, SIRT2, SNRPD1, SOX10, Sp17, SPA17, SSX-2, SSX-4, STEAP1, survivin, SYT-SSX1 or -SSX2 fusion protein, TAG-1, TAG-2,

Telomerase, TGF-betaRII, TPBG, TRAG-3, Triosephosphate isomerase, TRP-1/gp75, TRP-2, TRP2-INT2, tyrosinase, tyrosinase ("TYR"), VEGF, WT1, XAGE-1b/GAGED2a. In some embodiments, the antigen is a neo-antigen.

[00117] In some embodiments, the immunotherapy agent is a cancer vaccine and/or a component of a cancer vaccine (*e.g.,* an antigenic peptide and/or protein). The cancer vaccine can be a protein vaccine, a nucleic acid vaccine or a combination thereof. For example, in some embodiments, the cancer vaccine comprises a polypeptide comprising an epitope of a cancer-associated antigen. In some embodiments, the cancer vaccine comprises a nucleic acid (*e.g.,* DNA or RNA, such as mRNA) that encodes an epitope of a cancer-associated antigen. In some embodiemnts, the nucleic acid is a vector (e.g., a bacterial vector, viral vector). Examples of bacterial vectors include, but are not limited to, Mycobacterium bovis (BCG), Salmonella Typhimurium ssp., Salmonella Typhi ssp., Clostridium sp. spores, Escherichia coli Nissle 1917, Escherichia coli K-12/LLO, Listeria monocytogenes, and Shigella flexneri. Examples of viral vectors include, but are not limited to, vaccinia, adenovirus, RNA viruses, and replication-defective avipox, replication-defective fowlpox, replication-defective canarypox, replication-defective MVA and replication-defective adenovirus.

[00118] In some embodiments, the cancer immunotherapy comprises administration of an antigen presenting cell (APC) primed with a cancer-specific antigen. In some embodiments, the APC is a dendritic cell, a macrophage or a B cell.

[00119] Examples of cancer-associated antigens include, but are not limited to, adipophilin, AIM-2, ALDH1A1, alpha-actinin-4, alpha-fetoprotein ("AFP"), ARTC1, B-RAF, BAGE-1, BCLX (L), BCR-ABL fusion protein b3a2, beta-catenin, BING-4, CA-125, CALCA, carcinoembryonic antigen ("CEA"), CASP-5, CASP-8, CD274, CD45, Cdc27, CDK12, CDK4, CDKN2A, CEA, CLPP, COA-1, CPSF, CSNK1A1, CTAG1, CTAG2, cyclin D1, Cyclin-A1, dek-can fusion protein, DKK1, EFTUD2, Elongation factor 2, ENAH (hMena), Ep-CAM, EpCAM, EphA3, epithelial tumor antigen ("ETA"), ETV6-AML1 fusion protein, EZH2, FGF5, FLT3-ITD, FN1, G250/MN/CAIX, GAGE-1,2,8, GAGE-3,4,5,6,7, GAS7, glypican-3, GnTV, gp100/Pmel17, GPNMB, HAUS3, Hepsin, HER-2/neu, HERV-K-MEL, HLA-A11, HLA-A2, HLA-DOB, hsp70-2, IDO1, IGF2B3, IL13Ralpha2, Intestinal carboxyl esterase, K-ras, Kallikrein 4, KIF20A, KK-LC-1, KKLC1, KM-HN-1, KMHN1 also known as CCDC110, LAGE-1, LDLR-fucosyltransferaseAS fusion protein, Lengsin, M-CSF, MAGE-A1, MAGE-

A10, MAGE-A12, MAGE-A2, MAGE-A3, MAGE-A4, MAGE-A6, MAGE-A9, MAGE-C1, MAGE-C2, malic enzyme, mammaglobin-A, MART2, MATN, MC1R, MCSP, mdm-2, ME1, Melan-A/MART-1, Meloe, Midkine, MMP-2, MMP-7, MUC1, MUC5AC, mucin, MUM-1, MUM-2, MUM-3, Myosin, Myosin class I, N-raw, NA88-A, neo-PAP, NFYC, NY-BR-1, NY-ESO-1/LAGE-2, OA1, OGT, OS-9, P polypeptide, p53, PAP, PAX5, PBF, pml-RARalpha fusion protein, polymorphic epithelial mucin ("PEM"), PPP1R3B, PRAME, PRDX5, PSA, PSMA, PTPRK, RAB38/NY-MEL-1, RAGE-1, RBAF600, RGS5, RhoC, RNF43, RU2AS, SAGE, secernin 1, SIRT2, SNRPD1, SOX10, Sp17, SPA17, SSX-2, SSX-4, STEAP1, survivin, SYT-SSX1 or -SSX2 fusion protein, TAG-1, TAG-2, Telomerase, TGF-betaRII, TPBG, TRAG-3, Triosephosphate isomerase, TRP-1/gp75, TRP-2, TRP2-INT2, tyrosinase, tyrosinase ("TYR"), VEGF, WT1, XAGE-1b/GAGED2a. In some embodiments, the antigen is a neo-antigen.

[00120] In some embodiments, the cancer immunotherapy comprises administration of a cancer-specific chimeric antigen receptor (CAR). In some embodiments, the CAR is administered on the surface of a T cell. In some embodiments, the CAR binds specifically to a cancer-associated antigen.

[00121] In some embodiments, the cancer immunotherapy comprises administration of a cancer-specific T cell to the subject. In some embodiments, the T cell is a $CD4^+$ T cell. In some embodiments, the $CD4^+$ T cell is a T_H1 T cell, a T_H2 T cell or a T_H17 T cell. In some embodiments, the T cell expresses a T cell receptor specific for a cancer-associated antigen.

[00122] In some embodiments, the cancer vaccine is administered with an adjuvant. Examples of adjuvants include, but are not limited to, an immune modulatory protein, Adjuvant 65, α-GalCer, aluminum phosphate, aluminum hydroxide, calcium phosphate, β-Glucan Peptide, CpG DNA, GPI-0100, lipid A, lipopolysaccharide, Lipovant, Montanide, N-acetyl-muramyl-L-alanyl-D-isoglutamine, Pam3CSK4, quil A and trehalose dimycolate.

[00123] In some embodiments, the immunotherapy agent is an immune modulating protein to the subject. In some embodiments, the immune modulatory protein is a cytokine. Examples of immune modulating proteins include, but are not limited to, B lymphocyte chemoattractant ("BLC"), C-C motif chemokine 11 ("Eotaxin-1"), Eosinophil chemotactic protein 2 ("Eotaxin-2"), Granulocyte colony-stimulating factor ("G-CSF"), Granulocyte macrophage colony-stimulating factor ("GM-CSF"), 1-309, Intercellular Adhesion Molecule 1 ("ICAM-1"), Interferon gamma ("IFN-gamma"), Interlukin-1 alpha ("IL-1 alpha"), Interlukin-1

beta ("IL-1 beta"), Interleukin 1 receptor antagonist ("IL-1 ra"), Interleukin-2 ("IL-2"), Interleukin-4 ("IL-4"), Interleukin-5 ("IL-5"), Interleukin-6 ("IL-6"), Interleukin-6 soluble receptor ("IL-6 sR"), Interleukin-7 ("IL-7"), Interleukin-8 ("IL-8"), Interleukin- 10 ("IL-10"), Interleukin- 11 ("IL-11"), Subunit beta of Interleukin- 12 ("IL-12 p40" or "IL-12 p70"), Interleukin-13 ("IL-13"), Interleukin-15 ("IL-15"), Interleukin-16 ("IL-16"), Interleukin-17 ("IL-17"), Chemokine (C-C motif) Ligand 2 ("MCP-1"), Macrophage colony-stimulating factor ("M-CSF"), Monokine induced by gamma interferon ("MIG"), Chemokine (C-C motif) ligand 2 ("MIP-1 alpha"), Chemokine (C-C motif) ligand 4 ("MIP-1 beta"), Macrophage inflammatory protein- 1 -delta ("MIP-1 delta"), Platelet-derived growth factor subunit B ("PDGF-BB"), Chemokine (C-C motif) ligand 5, Regulated on Activation, Normal T cell Expressed and Secreted ("RANTES"), TIMP metallopeptidase inhibitor 1 ("TIMP-1"), TIMP metallopeptidase inhibitor 2 ("TIMP-2"), Tumor necrosis factor, lymphotoxin-alpha ("TNF alpha"), Tumor necrosis factor, lymphotoxin-beta ("TNF beta"), Soluble TNF receptor type 1 ("sTNFRI"), sTNFRIIAR, Brain-derived neurotrophic factor ("BDNF"), Basic fibroblast growth factor ("bFGF"), Bone morphogenetic protein 4 ("BMP-4"), Bone morphogenetic protein 5 ("BMP-5"), Bone morphogenetic protein 7 ("BMP-7"), Nerve growth factor ("b-NGF"), Epidermal growth factor ("EGF"), Epidermal growth factor receptor ("EGFR"), Endocrine-gland-derived vascular endothelial growth factor ("EG-VEGF"), Fibroblast growth factor 4 ("FGF-4"), Keratinocyte growth factor ("FGF-7"), Growth differentiation factor 15 ("GDF-15"), Glial cell-derived neurotrophic factor ("GDNF"), Growth Hormone, Heparin-binding EGF-like growth factor ("HB-EGF"), Hepatocyte growth factor ("HGF"), Insulin-like growth factor binding protein 1 ("IGFBP-1"), Insulin-like growth factor binding protein 2 ("IGFBP-2"), Insulin-like growth factor binding protein 3 (" IGFBP-3"), Insulin-like growth factor binding protein 4 ("IGFBP-4"), Insulin-like growth factor binding protein 6 ("IGFBP-6"), Insulin-like growth factor 1 ("IGF-1"), Insulin, Macrophage colony-stimulating factor ("M-CSF R"), Nerve growth factor receptor ("NGF R"), Neurotrophin-3 ("NT-3"), Neurotrophin-4 ("NT-4"), Osteoclastogenesis inhibitory factor ("Osteoprotegerin"), Platelet-derived growth factor receptors ("PDGF-AA"), Phosphatidylinositol-glycan biosynthesis ("PIGF"), Skp, Cullin, F-box containing comples ("SCF"), Stem cell factor receptor ("SCF R"), Transforming growth factor alpha ("TGFalpha"), Transforming growth factor beta-1 ("TGF beta 1"), Transforming growth factor beta-3 ("TGF beta 3"), Vascular endothelial growth factor ("VEGF"), Vascular endothelial growth factor

receptor 2 ("VEGFR2"), Vascular endothelial growth factor receptor 3 ("VEGFR3"), VEGF-D 6Ckine, Tyrosine-protein kinase receptor UFO ("Axl"), Betacellulin ("BTC"), Mucosae-associated epithelial chemokine ("CCL28"), Chemokine (C-C motif) ligand 27 ("CTACK"), Chemokine (C-X-C motif) ligand 16 ("CXCL16"), C-X-C motif chemokine 5 ("ENA-78"), Chemokine (C-C motif) ligand 26 ("Eotaxin-3"), Granulocyte chemotactic protein 2 ("GCP-2"), GRO, Chemokine (C-C motif) ligand 14 ("HCC-l"), Chemokine (C-C motif) ligand 16 ("HCC-4"), Interleukin-9 ("IL-9"), Interleukin-17 F ("IL-17F"), Interleukin- 18-binding protein ("IL-18 BPa"), Interleukin-28 A ("IL-28A"), Interleukin 29 ("IL-29"), Interleukin 31 ("IL-31"), C-X-C motif chemokine 10 ("IP-10"), Chemokine receptor CXCR3 ("I-TAC"), Leukemia inhibitory factor ("LIF"), Light, Chemokine (C motif) ligand ("Lymphotactin"), Monocyte chemoattractant protein 2 ("MCP-2"), Monocyte chemoattractant protein 3 ("MCP-3"), Monocyte chemoattractant protein 4 ("MCP-4"), Macrophage-derived chemokine ("MDC"), Macrophage migration inhibitory factor ("MIF"), Chemokine (C-C motif) ligand 20 ("MIP-3 alpha"), C-C motif chemokine 19 ("MIP-3 beta"), Chemokine (C-C motif) ligand 23 ("MPIF-1"), Macrophage stimulating protein alpha chain ("MSPalpha"), Nucleosome assembly protein 1-like 4 ("NAP-2"), Secreted phosphoprotein 1 ("Osteopontin"), Pulmonary and activation-regulated cytokine ("PARC"), Platelet factor 4 ("PF4"), Stroma cell-derived factor- 1 alpha ("SDF-1 alpha"), Chemokine (C-C motif) ligand 17 ("TARC"), Thymus-expressed chemokine ("TECK"), Thymic stromal lymphopoietin ("TSLP 4- IBB"), CD 166 antigen ("ALCAM"), Cluster of Differentiation 80 ("B7-1"), Tumor necrosis factor receptor superfamily member 17 ("BCMA"), Cluster of Differentiation 14 ("CD14"), Cluster of Differentiation 30 ("CD30"), Cluster of Differentiation 40 ("CD40 Ligand"), Carcinoembryonic antigen-related cell adhesion molecule 1 (biliary glycoprotein) ("CEACAM-1"), Death Receptor 6 ("DR6"), Deoxythymidine kinase ("Dtk"), Type 1 membrane glycoprotein ("Endoglin"), Receptor tyrosine-protein kinase erbB-3 ("ErbB3"), Endothelial-leukocyte adhesion molecule 1 ("E-Selectin"), Apoptosis antigen 1 ("Fas"), Fms-like tyrosine kinase 3 ("Flt-3L"), Tumor necrosis factor receptor superfamily member 1 ("GITR"), Tumor necrosis factor receptor superfamily member 14 ("HVEM"), Intercellular adhesion molecule 3 ("ICAM-3"), IL-1 R4, IL-1 RI, IL-10 Rbeta, IL-17R, IL-2Rgamma, IL-21R, Lysosome membrane protein 2 ("LIMPII"), Neutrophil gelatinase-associated lipocalin ("Lipocalin-2"), CD62L ("L-Selectin"), Lymphatic endothelium ("LYVE-1"), MHC class I polypeptide-related sequence A ("MICA"), MHC class I polypeptide-related sequence B

("MICB"), NRGl-betal, Beta-type platelet-derived growth factor receptor ("PDGF Rbeta"), Platelet endothelial cell adhesion molecule ("PECAM-1"), RAGE, Hepatitis A virus cellular receptor 1 ("TIM-1"), Tumor necrosis factor receptor superfamily member IOC ("TRAIL R3"), Trappin protein transglutaminase binding domain ("Trappin-2"), Urokinase receptor ("uPAR"), Vascular cell adhesion protein 1 ("VCAM-1"), XEDARActivin A, Agouti-related protein ("AgRP"), Ribonuclease 5 ("Angiogenin"), Angiopoietin 1, Angiostatin, Catheprin S, CD40, Cryptic family protein IB ("Cripto-1"), DAN, Dickkopf-related protein 1 ("DKK-1"), E-Cadherin, Epithelial cell adhesion molecule ("EpCAM"), Fas Ligand (FasL or CD95L), Fcg RIIB/C, FoUistatin, Galectin-7, Intercellular adhesion molecule 2 ("ICAM-2"), IL-13 Rl, IL-13R2, IL-17B, IL-2 Ra, IL-2 Rb, IL-23, LAP, Neuronal cell adhesion molecule ("NrCAM"), Plasminogen activator inhibitor- 1 ("PAI-1"), Platelet derived growth factor receptors ("PDGF-AB"), Resistin, stromal cell-derived factor 1 ("SDF-1 beta"), sgpl30, Secreted frizzled-related protein 2 ("ShhN"), Sialic acid-binding immunoglobulin-type lectins ("Siglec-5"), ST2, Transforming growth factor-beta 2 ("TGF beta 2"), Tie-2, Thrombopoietin ("TPO"), Tumor necrosis factor receptor superfamily member 10D ("TRAIL R4"), Triggering receptor expressed on myeloid cells 1 ("TREM-1"), Vascular endothelial growth factor C ("VEGF-C"), VEGFRlAdiponectin, Adipsin ("AND"), Alpha-fetoprotein ("AFP"), Angiopoietin-like 4 ("ANGPTL4"), Beta-2-microglobulin ("B2M"), Basal cell adhesion molecule ("BCAM"), Carbohydrate antigen 125 ("CA125"), Cancer Antigen 15-3 ("CA15-3"), Carcinoembryonic antigen ("CEA"), cAMP receptor protein ("CRP"), Human Epidermal Growth Factor Receptor 2 ("ErbB2"), Follistatin, Follicle-stimulating hormone ("FSH"), Chemokine (C-X-C motif) ligand 1 ("GRO alpha"), human chorionic gonadotropin ("beta HCG"), Insulin-like growth factor 1 receptor ("IGF-1 sR"), IL-1 sRII, IL-3, IL-18 Rb, IL-21, Leptin, Matrix metalloproteinase-1 ("MMP-1"), Matrix metalloproteinase-2 ("MMP-2"), Matrix metalloproteinase-3 ("MMP-3"), Matrix metalloproteinase-8 ("MMP-8"), Matrix metalloproteinase-9 ("MMP-9"), Matrix metalloproteinase-10 ("MMP-10"), Matrix metalloproteinase-13 ("MMP-13"), Neural Cell Adhesion Molecule ("NCAM-1"), Entactin ("Nidogen-1"), Neuron specific enolase ("NSE"), Oncostatin M ("OSM"), Procalcitonin, Prolactin, Prostate specific antigen ("PSA"), Sialic acid-binding Ig-like lectin 9 ("Siglec-9"), ADAM 17 endopeptidase ("TACE"), Thyroglobulin, Metalloproteinase inhibitor 4 ("TIMP-4"), TSH2B4, Disintegrin and metalloproteinase domain-containing protein 9 ("ADAM-9"), Angiopoietin 2, Tumor necrosis factor ligand superfamily

member 13/ Acidic leucine-rich nuclear phosphoprotein 32 family member B ("APRIL"), Bone morphogenetic protein 2 ("BMP-2"), Bone morphogenetic protein 9 ("BMP-9"), Complement component 5a ("C5a"), Cathepsin L, CD200, CD97, Chemerin, Tumor necrosis factor receptor superfamily member 6B ("DcR3"), Fatty acid-binding protein 2 ("FABP2"), Fibroblast activation protein, alpha ("FAP"), Fibroblast growth factor 19 ("FGF-19"), Galectin-3, Hepatocyte growth factor receptor ("HGF R"), IFN-gammalpha/beta R2, Insulin-like growth factor 2 ("IGF-2"), Insulin-like growth factor 2 receptor ("IGF-2 R"), Interleukin-1 receptor 6 ("IL-1R6"), Interleukin 24 ("IL-24"), Interleukin 33 ("IL-33", Kallikrein 14, Asparaginyl endopeptidase ("Legumain"), Oxidized low-density lipoprotein receptor 1 ("LOX-1"), Mannose-binding lectin ("MBL"), Neprilysin ("NEP"), Notch homolog 1, translocation-associated (Drosophila) ("Notch-1"), Nephroblastoma overexpressed ("NOV"), Osteoactivin, Programmed cell death protein 1 ("PD-1"), N-acetylmuramoyl-L-alanine amidase ("PGRP-5"), Serpin A4, Secreted frizzled related protein 3 ("sFRP-3"), Thrombomodulin, Tolllike receptor 2 ("TLR2"), Tumor necrosis factor receptor superfamily member 10A ("TRAIL Rl"), Transferrin ("TRF"), WIF-lACE-2, Albumin, AMICA, Angiopoietin 4, B-cell activating factor ("BAFF"), Carbohydrate antigen 19-9 ("CA19-9"), CD 163 , Clusterin, CRT AM, Chemokine (C-X-C motif) ligand 14 ("CXCL14"), Cystatin C, Decorin ("DCN"), Dickkopf-related protein 3 ("Dkk-3"), Delta-like protein 1 ("DLL1"), Fetuin A, Heparin-binding growth factor 1 ("aFGF"), Folate receptor alpha ("FOLR1"), Furin, GPCR-associated sorting protein 1 ("GASP-1"), GPCR-associated sorting protein 2 ("GASP-2"), Granulocyte colony-stimulating factor receptor ("GCSF R"), Serine protease hepsin ("HAI-2"), Interleukin-17B Receptor ("IL-17B R"), Interleukin 27 ("IL-27"), Lymphocyte-activation gene 3 ("LAG-3"), Apolipoprotein A-V ("LDL R"), Pepsinogen I, Retinol binding protein 4 ("RBP4"), SOST, Heparan sulfate proteoglycan ("Syndecan-1"), Tumor necrosis factor receptor superfamily member 13B ("TACI"), Tissue factor pathway inhibitor ("TFPI"), TSP-1, Tumor necrosis factor receptor superfamily, member 10b ("TRAIL R2"), TRANCE, Troponin I, Urokinase Plasminogen Activator ("uPA"), Cadherin 5, type 2 or VE-cadherin (vascular endothelial) also known as CD144 ("VE-Cadherin"), WNTl-inducible-signaling pathway protein 1 ("WISP-1"), and Receptor Activator of Nuclear Factor κ B ("RANK").

[00124] In some embodiments, the cancer therapeutic is a radioactive moiety that comprises a radionuclide. Exemplary radionuclides include, but are not limited to Cr-51, Cs-131,

Ce-134, Se-75, Ru-97, I-125, Eu-149, Os-189m, Sb-119, I-123, Ho-161, Sb-117, Ce-139, In-111, Rh-103m, Ga-67, Tl-201, Pd-103, Au-195, Hg-197, Sr-87m, Pt-191, P-33, Er-169, Ru-103, Yb-169, Au-199, Sn-121, Tm-167, Yb-175, In-113m, Sn-113, Lu-177, Rh-105, Sn-117m, Cu-67, Sc-47, Pt-195m, Ce-141, I-131, Tb-161, As-77, Pt-197, Sm-153, Gd-159, Tm-173, Pr-143, Au-198, Tm-170, Re-186, Ag-111, Pd-109, Ga-73, Dy-165, Pm-149, Sn-123, Sr-89, Ho-166, P-32, Re-188, Pr-142, Ir-194, In-114m/In-114, and Y-90.

[00125] In some embodiments, the cancer therapeutic is an angiogenesis inhibitor to the subject. Examples of such angiogenesis inhibitors include, but are not limited to Bevacizumab (Avastin®), Ziv-aflibercept (Zaltrap®), Sorafenib (Nexavar®), Sunitinib (Sutent®), Pazopanib (Votrient®), Regorafenib (Stivarga®), and Cabozantinib (Cometriq™).

[00126] In some embodiments, the cancer therapeutic is an antibiotic. For example, if the presence of a cancer-associated bacteria and/or a cancer-associated microbiome profile is detected according to the methods provided herein, antibiotics can be administered to eliminate the cancer-associated bacteria from the subject. "Antibiotics" broadly refers to compounds capable of inhibiting or preventing a bacterial infection. Antibiotics can be classified in a number of ways, including their use for specific infections, their mechanism of action, their bioavailability, or their spectrum of target microbe (*e.g.*, Gram-negative vs. Gram-positive bacteria, aerobic vs. anaerobic bacteria, etc.) and these may be used to kill specific bacteria in specific areas of the host ("niches") (Leekha, et al 2011. General Principles of Antimicrobial Therapy. Mayo Clin Proc. 86(2): 156-167). In certain embodiments, antibiotics can be used to selectively target bacteria of a specific niche. In some embodiments, antibiotics known to treat a particular infection that includes a cancer niche may be used to target cancer-associated microbes, including cancer-associated bacteria in that niche. In other embodiments, antibiotics are administered after the bacterial treatment. In some embodiments, antibiotics are administered after the bacterial treatment to remove the engraftment.

[00127] In some aspects, antibiotics can be selected based on their bactericidal or bacteriostatic properties. Bactericidal antibiotics include mechanisms of action that disrupt the cell wall (*e.g.*, β-lactams), the cell membrane (*e.g.*, daptomycin), or bacterial DNA (*e.g.*, fluoroquinolones). Bacteriostatic agents inhibit bacterial replication and include sulfonamides, tetracyclines, and macrolides, and act by inhibiting protein synthesis. Furthermore, while some drugs can be bactericidal in certain organisms and bacteriostatic in others, knowing the target

organism allows one skilled in the art to select an antibiotic with the appropriate properties. In certain treatment conditions, bacteriostatic antibiotics inhibit the activity of bactericidal antibiotics. Thus, in certain embodiments, bactericidal and bacteriostatic antibiotics are not combined.

[00128] Antibiotics include, but are not limited to aminoglycosides, ansamycins, carbacephems, carbapenems, cephalosporins, glycopeptides, lincosamides, lipopeptides, macrolides, monobactams, nitrofurans, oxazolidonones, penicillins, polypeptide antibiotics, quinolones, fluoroquinolone, sulfonamides, tetracyclines, and anti-mycobacterial compounds, and combinations thereof.

[00129] Aminoglycosides include, but are not limited to Amikacin, Gentamicin, Kanamycin, Neomycin, Netilmicin, Tobramycin, Paromomycin, and Spectinomycin. Aminoglycosides are effective, *e.g.*, against Gram-negative bacteria, such as Escherichia coli, Klebsiella, Pseudomonas aeruginosa, and Francisella tularensis, and against certain aerobic bacteria but less effective against obligate/facultative anaerobes. Aminoglycosides are believed to bind to the bacterial 30S or 50S ribosomal subunit thereby inhibiting bacterial protein synthesis.

[00130] Ansamycins include, but are not limited to, Geldanamycin, Herbimycin, Rifamycin, and Streptovaricin. Geldanamycin and Herbimycin are believed to inhibit or alter the function of Heat Shock Protein 90.

[00131] Carbacephems include, but are not limited to, Loracarbef. Carbacephems are believed to inhibit bacterial cell wall synthesis.

[00132] Carbapenems include, but are not limited to, Ertapenem, Doripenem, Imipenem/Cilastatin, and Meropenem. Carbapenems are bactericidal for both Gram-positive and Gram-negative bacteria as broad-spectrum antibiotics. Carbapenems are believed to inhibit bacterial cell wall synthesis.

[00133] Cephalosporins include, but are not limited to, Cefadroxil, Cefazolin, Cefalotin, Cefalothin, Cefalexin, Cefaclor, Cefamandole, Cefoxitin, Cefprozil, Cefuroxime, Cefixime, Cefdinir, Cefditoren, Cefoperazone, Cefotaxime, Cefpodoxime, Ceftazidime, Ceftibuten, Ceftizoxime, Ceftriaxone, Cefepime, Ceftaroline fosamil,and Ceftobiprole. Selected Cephalosporins are effective, *e.g.*, against Gram-negative bacteria and against Gram-positive bacteria, including *Pseudomonas*, certain Cephalosporins are effective against methicillin-

resistant *Staphylococcus aureus* (MRSA). Cephalosporins are believed to inhibit bacterial cell wall synthesis by disrupting synthesis of the peptidoglycan layer of bacterial cell walls.

[00134] Glycopeptides include, but are not limited to, Teicoplanin, Vancomycin, and Telavancin. Glycopeptides are effective, *e.g.*, against aerobic and anaerobic Gram-positive bacteria including MRSA and *Clostridium difficile*. Glycopeptides are believed to inhibit bacterial cell wall synthesis by disrupting synthesis of the peptidoglycan layer of bacterial cell walls.

[00135] Lincosamides include, but are not limited to, Clindamycin and Lincomycin. Lincosamides are effective, *e.g.,* against anaerobic bacteria, as well as Staphylococcus, and Streptococcus. Lincosamides are believed to bind to the bacterial 50S ribosomal subunit thereby inhibiting bacterial protein synthesis.

[00136] Lipopeptides include, but are not limited to, Daptomycin. Lipopeptides are effective, *e.g.*, against Gram-positive bacteria. Lipopeptides are believed to bind to the bacterial membrane and cause rapid depolarization.

[00137] Macrolides include, but are not limited to, Azithromycin, Clarithromycin, Dirithromycin, Erythromycin, Roxithromycin, Troleandomycin, Telithromycin, and Spiramycin. Macrolides are effective, *e.g.*, against Streptococcus and Mycoplasma. Macrolides are believed to bind to the bacterial or 50S ribosomal subunit, thereby inhibiting bacterial protein synthesis.

[00138] Monobactams include, but are not limited to, Aztreonam. Monobactams are effective, *e.g.*, against Gram-negative bacteria. Monobactams are believed to inhibit bacterial cell wall synthesis by disrupting synthesis of the peptidoglycan layer of bacterial cell walls.

[00139] Nitrofurans include, but are not limited to, Furazolidone and Nitrofurantoin.

[00140] Oxazolidonones include, but are not limited to, Linezolid, Posizolid, Radezolid, and Torezolid. Oxazolidonones are believed to be protein synthesis inhibitors.

[00141] Penicillins include, but are not limited to, Amoxicillin, Ampicillin, Azlocillin, Carbenicillin, Cloxacillin, Dicloxacillin, Flucloxacillin, Mezlocillin, Methicillin, Nafcillin, Oxacillin, Penicillin G, Penicillin V, Piperacillin, Temocillin and Ticarcillin. Penicillins are effective, *e.g.*, against Gram-positive bacteria, facultative anaerobes, *e.g.*, Streptococcus, Borrelia, and Treponema. Penicillins are believed to inhibit bacterial cell wall synthesis by disrupting synthesis of the peptidoglycan layer of bacterial cell walls.

[00142] Penicillin combinations include, but are not limited to, Amoxicillin/clavulanate, Ampicillin/sulbactam, Piperacillin/tazobactam, and Ticarcillin/clavulanate.

[00143] Polypeptide antibiotics include, but are not limited to, Bacitracin, Colistin, and Polymyxin B and E. Polypeptide Antibiotics are effective, *e.g.*, against Gram-negative bacteria. Certain polypeptide antibiotics are believed to inhibit isoprenyl pyrophosphate involved in synthesis of the peptidoglycan layer of bacterial cell walls, while others destabilize the bacterial outer membrane by displacing bacterial counter-ions.

[00144] Quinolones and Fluoroquinolone include, but are not limited to, Ciprofloxacin, Enoxacin, Gatifloxacin, Gemifloxacin, Levofloxacin, Lomefloxacin, Moxifloxacin, Nalidixic acid, Norfloxacin, Ofloxacin, Trovafloxacin, Grepafloxacin, Sparfloxacin, and Temafloxacin. Quinolones/Fluoroquinolone are effective, *e.g.*, against *Streptococcus* and *Neisseria*. Quinolones/Fluoroquinolone are believed to inhibit the bacterial DNA gyrase or topoisomerase IV, thereby inhibiting DNA replication and transcription.

[00145] Sulfonamides include, but are not limited to, Mafenide, Sulfacetamide, Sulfadiazine, Silver sulfadiazine, Sulfadimethoxine, Sulfamethizole, Sulfamethoxazole, Sulfanilimide, Sulfasalazine, Sulfisoxazole, Trimethoprim-Sulfamethoxazole (Co-trimoxazole), and Sulfonamidochrysoidine. Sulfonamides are believed to inhibit folate synthesis by competitive inhibition of dihydropteroate synthetase, thereby inhibiting nucleic acid synthesis.

[00146] Tetracyclines include, but are not limited to, Demeclocycline, Doxycycline, Minocycline, Oxytetracycline, and Tetracycline. Tetracyclines are effective, *e.g.*, against Gram-negative bacteria. Tetracyclines are believed to bind to the bacterial 30S ribosomal subunit thereby inhibiting bacterial protein synthesis.

[00147] Anti-mycobacterial compounds include, but are not limited to, Clofazimine, Dapsone, Capreomycin, Cycloserine, Ethambutol, Ethionamide, Isoniazid, Pyrazinamide, Rifampicin, Rifabutin, Rifapentine, and Streptomycin.

[00148] Suitable antibiotics also include arsphenamine, chloramphenicol, fosfomycin, fusidic acid, metronidazole, mupirocin, platensimycin, quinupristin/dalfopristin, tigecycline, tinidazole, trimethoprim amoxicillin/clavulanate, ampicillin/sulbactam, amphomycin ristocetin, azithromycin, bacitracin, buforin II, carbomycin, cecropin Pl, clarithromycin, erythromycins, furazolidone, fusidic acid, Na fusidate, gramicidin, imipenem, indolicidin, josamycin, magainan II, metronidazole, nitroimidazoles, mikamycin, mutacin B-Ny266, mutacin B-JHl 140, mutacin

J-T8, nisin, nisin A, novobiocin, oleandomycin, ostreogrycin, piperacillin/tazobactam, pristinamycin, ramoplanin, ranalexin, reuterin, rifaximin, rosamicin, rosaramicin, spectinomycin, spiramycin, staphylomycin, streptogramin, streptogramin A, synergistin, taurolidine, teicoplanin, telithromycin, ticarcillin/clavulanic acid, triacetyloleandomycin, tylosin, tyrocidin, tyrothricin, vancomycin, vemamycin, and virginiamycin.

[00149] In some embodiments, the the cancer therapy comprises administering a therapeutic bacteria and/or a therapeutic combination of bacteria to the subject so a healthy microbiome can be reconstituted in the subject. In some embodiments, the therapeutic bacteria is a non-cancer-associated bacteria. In some embodiments the therapeutic bacteria is a probiotic bacteria.

Cancer

[00150] In some embodiments, the methods and compositions described herein relate to the treatment of cancer. Examples of cancers that may treated by methods described herein include, but are not limited to, hematological malignancy, acute nonlymphocytic leukemia, chronic lymphocytic leukemia, acute granulocytic leukemia, chronic granulocytic leukemia, acute promyelocytic leukemia, adult T-cell leukemia, aleukemic leukemia, a leukocythemic leukemia, basophilic leukemia, blast cell leukemia, bovine leukemia, chronic myelocytic leukemia, leukemia cutis, embryonal leukemia, eosinophilic leukemia, Gross' leukemia, Rieder cell leukemia, Schilling's leukemia, stem cell leukemia, subleukemic leukemia, undifferentiated cell leukemia, hairy-cell leukemia, hemoblastic leukemia, hemocytoblastic leukemia, histiocytic leukemia, stem cell leukemia, acute monocytic leukemia, leukopenic leukemia, lymphatic leukemia, lymphoblastic leukemia, lymphocytic leukemia, lymphogenous leukemia, lymphoid leukemia, lymphosarcoma cell leukemia, mast cell leukemia, megakaryocytic leukemia, micromyeloblastic leukemia, monocytic leukemia, myeloblastic leukemia, myelocytic leukemia, myeloid granulocytic leukemia, myelomonocytic leukemia, Naegeli leukemia, plasma cell leukemia, plasmacytic leukemia, promyelocytic leukemia, acinar carcinoma, acinous carcinoma, adenocystic carcinoma, adenoid cystic carcinoma, carcinoma adenomatosum, carcinoma of adrenal cortex, alveolar carcinoma, alveolar cell carcinoma, basal cell carcinoma, carcinoma basocellulare, basaloid carcinoma, basosquamous cell carcinoma, bronchioalveolar carcinoma, bronchiolar carcinoma, bronchogenic carcinoma, cerebriform carcinoma, cholangiocellular

carcinoma, chorionic carcinoma, colloid carcinoma, comedo carcinoma, corpus carcinoma, cribriform carcinoma, carcinoma en cuirasse, carcinoma cutaneum, cylindrical carcinoma, cylindrical cell carcinoma, duct carcinoma, carcinoma durum, embryonal carcinoma, encephaloid carcinoma, epiennoid carcinoma, carcinoma epitheliale adenoides, exophytic carcinoma, carcinoma ex ulcere, carcinoma fibrosum, gelatiniform carcinoma, gelatinous carcinoma, giant cell carcinoma, signet-ring cell carcinoma, carcinoma simplex, small-cell carcinoma, solanoid carcinoma, spheroidal cell carcinoma, spindle cell carcinoma, carcinoma spongiosum, squamous carcinoma, squamous cell carcinoma, string carcinoma, carcinoma telangiectaticum, carcinoma telangiectodes, transitional cell carcinoma, carcinoma tuberosum, tuberous carcinoma, verrucous carcinoma, carcinoma villosum, carcinoma gigantocellulare, glandular carcinoma, granulosa cell carcinoma, hair-matrix carcinoma, hematoid carcinoma, hepatocellular carcinoma, Hurthle cell carcinoma, hyaline carcinoma, hypernephroid carcinoma, infantile embryonal carcinoma, carcinoma in situ, intraepidermal carcinoma, intraepithelial carcinoma, Krompecher's carcinoma, Kulchitzky-cell carcinoma, large-cell carcinoma, lenticular carcinoma, carcinoma lenticulare, lipomatous carcinoma, lymphoepithelial carcinoma, carcinoma medullare, medullary carcinoma, melanotic carcinoma, carcinoma molle, mucinous carcinoma, carcinoma muciparum, carcinoma mucocellulare, mucoepidermoid carcinoma, carcinoma mucosum, mucous carcinoma, carcinoma myxomatodes, naspharyngeal carcinoma, oat cell carcinoma, carcinoma ossificans, osteoid carcinoma, papillary carcinoma, periportal carcinoma, preinvasive carcinoma, prickle cell carcinoma, pultaceous carcinoma, renal cell carcinoma of kidney, reserve cell carcinoma, carcinoma sarcomatodes, schneiderian carcinoma, scirrhous carcinoma, carcinoma scroti, chondrosarcoma, fibrosarcoma, lymphosarcoma, melanosarcoma, myxosarcoma, osteosarcoma, endometrial sarcoma, stromal sarcoma, Ewing' s sarcoma, fascial sarcoma, fibroblastic sarcoma, giant cell sarcoma, Abemethy's sarcoma, adipose sarcoma, liposarcoma, alveolar soft part sarcoma, ameloblastic sarcoma, botryoid sarcoma, chloroma sarcoma, chorio carcinoma, embryonal sarcoma, Wilms' tumor sarcoma, granulocytic sarcoma, Hodgkin's sarcoma, idiopathic multiple pigmented hemorrhagic sarcoma, immunoblastic sarcoma of B cells, lymphoma, immunoblastic sarcoma of T-cells, Jensen's sarcoma, Kaposi's sarcoma, Kupffer cell sarcoma, angiosarcoma, leukosarcoma, malignant mesenchymoma sarcoma, parosteal sarcoma, reticulocytic sarcoma, rhabdosarcoma, serocystic sarcoma, synovial sarcoma, telangiectaltic sarcoma, Hodgkin's Disease, Non-Hodgkin's

Lymphoma, multiple myeloma, neuroblastoma, bladder cancer, breast cancer, ovarian cancer, lung cancer, colorectal cancer, rhabdomyosarcoma, primary thrombocytosis, primary macroglobulinemia, small-cell lung tumors, primary brain tumors, stomach cancer, colon cancer, malignant pancreatic insulanoma, malignant carcinoid, premalignant skin lesions, testicular cancer, lymphomas, thyroid cancer, neuroblastoma, esophageal cancer, genitourinary tract cancer, malignant hypercalcemia, cervical cancer, endometrial cancer, adrenal cortical cancer, Harding-Passey melanoma, juvenile melanoma, lentigo maligna melanoma, malignant melanoma, acral-lentiginous melanoma, amelanotic melanoma, benign juvenile melanoma, Cloudman's melanoma, S91 melanoma, nodular melanoma subungal melanoma, and superficial spreading melanoma.

[00151] In some embodiments, the methods and compositions provided herein relate to the treatment of a leukemia. The term "leukemia" is meant broadly progressive, malignant diseases of the hematopoietic organs/systems and is generally characterized by a distorted proliferation and development of leukocytes and their precursors in the blood and bone marrow. Non-limiting examples of leukemia diseases include, acute nonlymphocytic leukemia, chronic lymphocytic leukemia, acute granulocytic leukemia, chronic granulocytic leukemia, acute promyelocytic leukemia, adult T-cell leukemia, aleukemic leukemia, a leukocythemic leukemia, basophilic leukemia, blast cell leukemia, bovine leukemia, chronic myelocytic leukemia, leukemia cutis, embryonal leukemia, eosinophilic leukemia, Gross' leukemia, Rieder cell leukemia, Schilling's leukemia, stem cell leukemia, subleukemic leukemia, undifferentiated cell leukemia, hairy-cell leukemia, hemoblastic leukemia, hemocytoblastic leukemia, histiocytic leukemia, stem cell leukemia, acute monocytic leukemia, leukopenic leukemia, lymphatic leukemia, lymphoblastic leukemia, lymphocytic leukemia, lymphogenous leukemia, lymphoid leukemia, lymphosarcoma cell leukemia, mast cell leukemia, megakaryocytic leukemia, micromyeloblastic leukemia, monocytic leukemia, myeloblastic leukemia, myelocytic leukemia, myeloid granulocytic leukemia, myelomonocytic leukemia, Naegeli leukemia, plasma cell leukemia, plasmacytic leukemia, and promyelocytic leukemia.

[00152] In some embodiments, the methods and compositions provided herein relate to the treatment of a carcinoma. The term "carcinoma" refers to a malignant growth made up of epithelial cells tending to infiltrate the surrounding tissues, and/or resist physiological and non-physiological cell death signals and gives rise to metastases. Non-limiting exemplary types of

carcinomas include, acinar carcinoma, acinous carcinoma, adenocystic carcinoma, adenoid cystic carcinoma, carcinoma adenomatosum, carcinoma of adrenal cortex, alveolar carcinoma, alveolar cell carcinoma, basal cell carcinoma, carcinoma basocellulare, basaloid carcinoma, basosquamous cell carcinoma, bronchioalveolar carcinoma, bronchiolar carcinoma, bronchogenic carcinoma, cerebriform carcinoma, cholangiocellular carcinoma, chorionic carcinoma, colloid carcinoma, comedo carcinoma, corpus carcinoma, cribriform carcinoma, carcinoma en cuirasse, carcinoma cutaneum, cylindrical carcinoma, cylindrical cell carcinoma, duct carcinoma, carcinoma durum, embryonal carcinoma, encephaloid carcinoma, epiennoid carcinoma, carcinoma epitheliale adenoides, exophytic carcinoma, carcinoma ex ulcere, carcinoma fibrosum, gelatiniform carcinoma, gelatinous carcinoma, giant cell carcinoma, signet-ring cell carcinoma, carcinoma simplex, small-cell carcinoma, solanoid carcinoma, spheroidal cell carcinoma, spindle cell carcinoma, carcinoma spongiosum, squamous carcinoma, squamous cell carcinoma, string carcinoma, carcinoma telangiectaticum, carcinoma telangiectodes, transitional cell carcinoma, carcinoma tuberosum, tuberous carcinoma, verrucous carcinoma, carcinoma villosum, carcinoma gigantocellulare, glandular carcinoma, granulosa cell carcinoma, hair-matrix carcinoma, hematoid carcinoma, hepatocellular carcinoma, Hurthle cell carcinoma, hyaline carcinoma, hypernephroid carcinoma, infantile embryonal carcinoma, carcinoma in situ, intraepidermal carcinoma, intraepithelial carcinoma, Krompecher's carcinoma, Kulchitzky-cell carcinoma, large-cell carcinoma, lenticular carcinoma, carcinoma lenticulare, lipomatous carcinoma, lymphoepithelial carcinoma, carcinoma medullare, medullary carcinoma, melanotic carcinoma, carcinoma molle, mucinous carcinoma, carcinoma muciparum, carcinoma mucocellulare, mucoepidermoid carcinoma, carcinoma mucosum, mucous carcinoma, carcinoma myxomatodes, naspharyngeal carcinoma, oat cell carcinoma, carcinoma ossificans, osteoid carcinoma, papillary carcinoma, periportal carcinoma, preinvasive carcinoma, prickle cell carcinoma, pultaceous carcinoma, renal cell carcinoma of kidney, reserve cell carcinoma, carcinoma sarcomatodes, schneiderian carcinoma, scirrhous carcinoma, and carcinoma scroti.

[00153] In some embodiments, the methods and compositions provided herein relate to the treatment of a sarcoma. The term "sarcoma" generally refers to a tumor which is made up of a substance like the embryonic connective tissue and is generally composed of closely packed cells embedded in a fibrillar, heterogeneous, or homogeneous substance. Sarcomas include, but are not limited to, chondrosarcoma, fibrosarcoma, lymphosarcoma, melanosarcoma, myxosarcoma,

osteosarcoma, endometrial sarcoma, stromal sarcoma, Ewing' s sarcoma, fascial sarcoma, fibroblastic sarcoma, giant cell sarcoma, Abemethy's sarcoma, adipose sarcoma, liposarcoma, alveolar soft part sarcoma, ameloblastic sarcoma, botryoid sarcoma, chloroma sarcoma, chorio carcinoma, embryonal sarcoma, Wilms' tumor sarcoma, granulocytic sarcoma, Hodgkin's sarcoma, idiopathic multiple pigmented hemorrhagic sarcoma, immunoblastic sarcoma of B cells, lymphoma, immunoblastic sarcoma of T-cells, Jensen's sarcoma, Kaposi's sarcoma, Kupffer cell sarcoma, angiosarcoma, leukosarcoma, malignant mesenchymoma sarcoma, parosteal sarcoma, reticulocytic sarcoma, Rous sarcoma, serocystic sarcoma, synovial sarcoma, and telangiectaltic sarcoma.

[00154] Additional exemplary neoplasias that can be treated using the methods and compositions described herein include Hodgkin's Disease, Non-Hodgkin's Lymphoma, multiple myeloma, neuroblastoma, breast cancer, ovarian cancer, lung cancer, rhabdomyosarcoma, primary thrombocytosis, primary macroglobulinemia, small-cell lung tumors, primary brain tumors, stomach cancer, colon cancer, malignant pancreatic insulanoma, malignant carcinoid, premalignant skin lesions, testicular cancer, lymphomas, thyroid cancer, neuroblastoma, esophageal cancer, genitourinary tract cancer, malignant hypercalcemia, cervical cancer, endometrial cancer, and adrenal cortical cancer.

[00155] In some embodiments, the cancer treated is a melanoma. The term "melanoma" is taken to mean a tumor arising from the melanocytic system of the skin and other organs. Non-limiting examples of melanomas are Harding-Passey melanoma, juvenile melanoma, lentigo maligna melanoma, malignant melanoma, acral-lentiginous melanoma, amelanotic melanoma, benign juvenile melanoma, Cloudman's melanoma, S91 melanoma, nodular melanoma subungal melanoma, and superficial spreading melanoma.

[00156] Particular categories of tumors that can be treated using methods and compositions described herein include lymphoproliferative disorders, breast cancer, ovarian cancer, prostate cancer, cervical cancer, endometrial cancer, bone cancer, liver cancer, stomach cancer, colon cancer, colorectal cancer, pancreatic cancer, cancer of the thyroid, head and neck cancer, cancer of the central nervous system, cancer of the peripheral nervous system, skin cancer, kidney cancer, as well as metastases of all the above. Particular types of tumors include hepatocellular carcinoma, hepatoma, hepatoblastoma, rhabdomyosarcoma, esophageal carcinoma, thyroid carcinoma, ganglioblastoma, fibrosarcoma, myxosarcoma, liposarcoma,

chondrosarcoma, osteogenic sarcoma, chordoma, angiosarcoma, endotheliosarcoma, Ewing's tumor, leimyosarcoma, rhabdotheliosarcoma, invasive ductal carcinoma, papillary adenocarcinoma, melanoma, pulmonary squamous cell carcinoma, basal cell carcinoma, adenocarcinoma (well differentiated, moderately differentiated, poorly differentiated or undifferentiated), bronchioloalveolar carcinoma, renal cell carcinoma, hypernephroma, hypernephroid adenocarcinoma, bile duct carcinoma, choriocarcinoma, seminoma, embryonal carcinoma, Wilms' tumor, testicular tumor, lung carcinoma including small cell, non-small and large cell lung carcinoma, bladder carcinoma, glioma, astrocyoma, medulloblastoma, craniopharyngioma, ependymoma, pinealoma, retinoblastoma, neuroblastoma, colon carcinoma, rectal carcinoma, hematopoietic malignancies including all types of leukemia and lymphoma including: acute myelogenous leukemia, acute myelocytic leukemia, acute lymphocytic leukemia, chronic myelogenous leukemia, chronic lymphocytic leukemia, mast cell leukemia, multiple myeloma, myeloid lymphoma, Hodgkin' s lymphoma, non-Hodgkin' s lymphoma.

[00157] Cancers treated in certain embodiments also include precancerous lesions, *e.g.*, actinic keratosis (solar keratosis), moles (dysplastic nevi), acitinic chelitis (farmer's lip), cutaneous horns, Barrett's esophagus, atrophic gastritis, dyskeratosis congenita, sideropenic dysphagia, lichen planus, oral submucous fibrosis, actinic (solar) elastosis and cervical dysplasia.

[00158] Cancers treated in some embodiments include non-cancerous or benign tumors, *e.g.*, of endodermal, ectodermal or mesenchymal origin, including, but not limited to cholangioma, colonic polyp, adenoma, papilloma, cystadenoma, liver cell adenoma, hydatidiform mole, renal tubular adenoma, squamous cell papilloma, gastric polyp, hemangioma, osteoma, chondroma, lipoma, fibroma, lymphangioma, leiomyoma, rhabdomyoma, astrocytoma, nevus, meningioma, and ganglioneuroma.

EXAMPLES

Example 1: *Bifidobacterium animalis ssp. lactis* production

[00159] Enriched media is used to grow and prepare *Bifidobacterium animalis ssp. lactis* for in vitro and in vivo use. For example, media may contain sugar, yeast extracts, peptones, trace elements, and vitamins. Alternatively, media may be prepared and *Bifidobacterium animalis ssp. lactis* grown as shown by Saarela *et al., J. Applied Microbiology.* 2005. 99: 1330-1339, which is hereby incorporated by reference. Influence of fermentation time, cryoprotectant and neutralization of cell concentrate on freeze-drying survival, storage stability, and acid and bile exposure of *Bifidobacterium animalis ssp. lactis* cells produced without milk-based ingredients,

[00160] At large scale, the media is sterilized. Sterilization may be by Ultra High Temperature (UHT) processing. The UHT processing is performed at very high temperature for short periods of time. The UHT range may be from 135-180°C. For example, the medium may be sterilized from between 10 to 30 seconds at 135°C.

[00161] Inoculum can be prepared in flasks or in smaller bioreactors and growth is monitored. For example, the inoculum size may be between approximately 0.5 and 3% of the total bioreactor volume. Depending on the application and need for material, bioreactor volume can be at least 2L, 10L, 80L, 100L, 250L, 1000L, 2500L, 5000L, 10,000L.

[00162] Before the inoculation, the bioreactor is prepared and ready with medium at desired pH, temperature, and oxygen concentration. For example, pH may be set between 4.5 and 8.0 During the fermentation the pH can be controlled through the use of sodium hydroxide, potassium hydroxide, or ammonium hydroxide. The temperature may be controlled from 25^0C to 45^0C, for example at 37^0C. Anaerobic conditions are created by reducing the level of oxygen in the culture broth from around 8mg/L to 0mg/L. For example, nitrogen or gas mixtures (N2, CO2, and H2) may be used in order to establish anaerobic conditions. Alternatively, no gases are used and anaerobic conditions are established by cells consuming remaining oxygen from the medium. Depending on strain and inoculum size, the bioreactor fermentation time can vary. For example, fermentation time can vary from approximately 5 hours to 48 hours.

[00163] Harvesting can be performed by continuous centrifugation. Product will be resuspended with various excipients to a desired final concentration. Excipients can be added for

cryo protection or for protection during lyophilization. Excipients can include, but are not limited to, sucrose, trehalose, or lactose, and these may be alternatively mixed with buffer and anti-oxidants. Prior to lyophilization, droplets of cell pellets mixed with excipients are submerged in liquid nitrogen.

[00164] Lyophilization of material, including live bacteria, begins with primary drying. During the primary drying phase, the ice is removed. Here, a vacuum is generated and an appropriate amount of heat is supplied to the material for the ice to sublime. During the secondary drying phase, product bound water molecules are removed. Here, the temperature is raised higher than in the primary drying phase to break any physico-chemical interactions that have formed between the water molecules and the product material. The pressure may also be lowered further to enhance desorption during this stage. After the freeze-drying process is complete, the chamber may be filled with an inert gas, such as nitrogen. The product may be sealed within the freeze dryer under dry conditions, preventing exposure to atmospheric water and contaminants.

Example 2: Orally Administered *Bifidobacterium animalis ssp. lactis* inhibits colorectal carcinoma tumor growth

[00165] Female 6-8 week old Balb/c mice were obtained from Taconic (Germantown, NY). 100,000 CT-26 colorectal tumor cells (ATCC CRL-2638) were resuspended in sterile PBS and inoculated in the presence of 50% Matrigel. CT-26 tumor cells were subcutaneously injected into one hind flank of each mouse. When tumor volumes reached an average of $100mm^3$ (approximately 10-12 days following tumor cell inoculation), animals were distributed into the following groups: 1) Vehicle + isotype control antibody IgG2a; and 3) *Bifidobacterium animalis ssp. lactis* Strain A (ATCC accession number PTA-125097) + IgG2a. *Bifidobacterium animalis ssp. lactis* bacteria ($1x10^9$) were administered by oral gavage (p.o.) daily, starting on day 1 until the conclusion of the study. The *Bifidobacterium animalis ssp. lactis* + IgG2a group showed significant tumor growth inhibition compared to the vehicle + IgG2a group (See **Figure 1**).

Example 3: Orally Administered *Bifidobacterium animalis ssp. lactis* inhibits colorectal carcinoma tumor growth comparable to inhibition seen with anti-PD-1 treatment

[00166] CT-26 tumor cells were subcutaneously injected into one hind flank of each Balb/c mouse as above. When tumor volumes reached an average of $100mm^3$ (approximately 10-12 days following tumor cell inoculation), animals were distributed into the following groups: 1) Vehicle + PBS; 2) Vehicle + isotype control antibody IgG2a; and 3) Vehicle + anti-PD-1; 4) *Bifidobacterium animalis ssp. lactis* Strain A (ATCC accession number PTA-125097) + IgG2a; and 5) *Bifidobacterium animalis ssp. lactis* Strain A + anti-PD-1. Antibodies were administered intraperitoneally (i.p.) at 100ug/mouse (100ul final volume) every four days, starting on day 1, and *Bifidobacterium animalis ssp. lactis* bacteria ($1x10^9$) were administered by oral gavage (p.o.) daily, starting on day 1 until the conclusion of the study. The *Bifidobacterium animalis ssp. lactis* + IgG2a group showed tumor growth inhibition comparable to that seen in the vehicle + anti-PD-1 group (See **Figure 2**).

Example 4: Treatment of colorectal carcinoma using different *Bifidobacterium animalis ssp. lactis* strains

As described in Examples 2 and 3, CT-26 tumor cells were subcutaneously injected into one hind flank of mice and animals were assigned into groups receiving the following treatments: 1) Vehicle; 2) *Bifidobacterium animalis ssp. lactis* Strain A (ATCC accession number PTA-125097); 3) *B. animalis lactis* Strain B; 4) *B. animalis ssp. lactis* Strain C; and 5) anti-PD-1. $1x10^9$ bacterial cells were administered daily (p.o.), beginning on day 1 of animal group assignment, and tumors measured as described above. Anti-PD-1 antibodies were administered intraperitoneally (i.p.) at 100ug/mouse (100ul final volume) every four days, starting on day 1. As seen in Table 2, Strain A showed significant tumor growth inhibition compared to the other *B. animalis lactis* treatment groups (**Table 2** and **Figure 3**).

Table 2. Colorectal carcinoma tumor volume (mm^3) on Day 22 (12th day of dosing)

	Anti PD-1	Vehicle	Strain C	Strain B	Strain A
Median tumor vol. (mm^3)	1096.97	1775.89	1294.94	1819.68	1788.83
Mean tumor vol. (mm^3)	1029.97	1757.75	1231.34	1753.12	1704.33
SEM (mm^3)	193.37	154.75	190.00	89.48	119.03

Example 5: Intratumorally administered *Bifidobacterium animalis ssp. lactis* inhibits colorectal carcinoma tumor growth

[00167] As described in Examples 2 and 3, CT-26 tumor cells were subcutaneously injected into one hind flank of mice and animals were assigned into groups receiving the following treatments: 1) Vehicle; 2) *Bifidobacterium animalis ssp. lactis* Strain A (ATCC accession number PTA-125097); and 3) anti-PD-1. $2x10^9$ bacterial cells were administered intratumorally (IT) on day 1 of animal group assignment, and tumors measured as described above. Mice received a second dose on day 4. Anti-PD-1 antibodies were administered intraperitoneally (i.p.) at 100ug/mouse (100ul final volume) every four days, starting on day 1. The *Bifidobacterium animalis ssp. lactis* Strain A group showed significant tumor growth inhibition compared to the control group (See **Figure 4**).

Example 6: *Bifidobacterium animalis ssp. lactis* in a mouse melanoma model

[00168] *B. animalis ssp. lactis* is tested for efficacy in a mouse melanoma model. For example, Female 6-8 week old C57Bl/6 mice are obtained from Taconic (Germantown, NY). Between 1x10^4 and 1x10^6 B16-F10 tumor cells (ATCC CRL-6475) may be used to inoculate various groups of mice. B16-F10 tumor cells may be resuspended in sterile PBS containing 50% Matrigel and inoculated in a 100ul final volume into one hind flank (the first flank) of each mouse. Treatment with *Bifidobacterium animalis ssp. lactis* is initiated at some point following tumor cell inoculation at varied doses and at defined intervals. For example, some mice receive between 1-5x10^9 CFU (100µl final volume) per dose. Possible routes of administration include oral gavage (p.o.), intravenous injection, intratumoral injection (IT) or peritumoral or subtumoral or subcutaneous injection. In order to assess the systemic anti-tumoral effects of *B. animalis lactis* treatment, additional mice may be inoculated with tumor cells in the contralateral (untreated, second) flank prior to IT, peritumoral, or subtumoral treatment with *B. animalis lactis* in the first flank.

[00169] Some mice may receive *Bifidobacterium animalis ssp. lactis* (p.o.) on day 1 (the day following tumor cell injection). Other mice may receive seven (7) consecutive doses of *Bifidobacterium animalis ssp. lactis* (one dose per day on days 14-21). Other mice receive daily dosing or, alternatively, some mice receive dosing every other day. Alternatively, mice are randomized into various treatment groups at a defined timepoint (*e.g.* on day 13) or when the tumors reach a certain size (*e.g.* 100 mm^3) and treatment is then initiated accordingly. For example, when tumor volumes reach an average of $100mm^3$ (approximately 10-12 days

following tumor cell inoculation), animals are distributed into groups and treated with either vehicle or *Bifidobacterium animalis ssp. lactis* (p.o. or IT). Some additional groups of mice may be treated with an additional cancer therapeutic or appropriate control antibody. One example of a cancer therapeutic that may be administered is an inhibitor of an immune checkpoint, for example anti-PD-1, anti-PD-L1, or other treatment that blocks the binding of an immune checkpoint to its ligand(s). Checkpoint inhibitors anti-PD-1 and anti-PD-L1 may be formulated in PBS and administered intraperitoneally (i.p.) in effective doses. For example, mice are given 100ug of anti-PD-1 (i.p.) every four days starting on day 1, and continuing for the duration of the study.

[00170] In addition, some mice are treated with antibiotics prior to treatment. For example, vancomycin (0.5g/L), ampicillin (1.0g/L), gentamicin (1.0g/L) and amphotericin B (0.2g/L) are added to the drinking water, and antibiotic treatment is halted at the time of treatment or a few days prior to treatment. Some mice are inoculated with tumor cells without receiving prior treatment with antibiotics. Also, in some groups the B16-f10 cells may be cultured with 100 U/ml penicillin and 10mg/ml streptomycin prior to inoculation.

[00171] For example, B16-F10 cells were expanded using standard tissue culture technique, maintained in DMEM medium supplemented with 10% fetal bovine serum (FBS), 100 U/ml penicillin, and 10mg/ml streptomycin in a 37^0C incubator with 5% CO_2. On the day of tumor cell implantation, B16-F10 cells were resuspended in cold HBSS at 1x10^6 tumor cells/ml and each mouse was injected with 100ul of resuspended tumor cells. Tumor growth was measured three times per week using digital calipers according to methods known in the art. When the average tumor size reached approximately 100mm^3, mice were randomly separated into treatment groups and treatment initiated according to Table 3 below.

[00172] **Table 3. Treatments**

Group, N=20	Treatment #1	Treatment #2
1	Vehicle (PO, 100 µL vol. QD)	PBS (IP, 100 µ L, Q4D)
2	Vehicle (PO, 100 µ L vol. QD)	anti-PD-L1 (IP, 200 µ g/100 µ L vol, Q4D)
3	B. animalis ssp. lactis Strain A (PO, 6.5x10^9 CFU in 100 µ L vol. QD)	PBS (IP, 100 µ L, Q4D)

4	B. animalis ssp. lactis Strain A (PO, 6.5x10^9 CFU in 100 µ L vol. QD)	anti-PD-L1 (IP, 200 µ g/100 µ L vol, Q4D)

[00173] Animals received Treatment #1 (Vehicle or B. animalis ssp. lactis Strain A) via oral gavage every day. Mice received Treatment #2 (PBS or anti-PD-L1 intraperitoneally every 4 days until the termination of the study. Treating tumor-inoculated mice with a combination of *Bifidobacterium animalis ssp. lactis* Strain A and anti-PD-L1 inhibits tumor growth in a manner superior to either *B. animalis ssp. lactis* Strain A or anti-PD-L1 alone **(Figure 5).**

Example 7: Infiltration of CD3+ cells is significantly increased in CT26 tumors in mice orally gavaged with B. animalis lactis Strain A.

[00174] In some studies, at various timepoints, mice are sacrificed and tumors, lymph nodes, or other tissues may be removed for *ex vivo* flow cytometric analysis using methods known in the art. For example, tumors are dissociated using a Miltenyi tumor dissociation enzyme cocktail according to the manufacturer's instructions. Tumor weights are recorded and tumors are chopped then placed in 15ml tubes containing the enzyme cocktail and placed on ice. Samples are then placed on a gentle shaker at 37°C for 45 minutes and quenched with up to 15ml complete RPMI. Each cell suspension is strained through a 70µm filter into a 50ml falcon tube and centrifuged at 1000 rpm for 10 minutes. Cells are resuspended in FACS buffer and washed to remove remaining debris. If necessary, samples are strained again through a second 70µm filter into a new tube. Cells are stained for analysis by flow cytometry using techniques known in the art. Staining antibodies can include anti-CD11c (dendritic cells), anti-CD80, anti-CD86, anti-CD40, anti-MHCII, anti-CD8a, anti-CD4, and anti-CD103. Other markers that may be analyzed include pan-immune cell marker CD45, T cell markers (CD3, CD4, CD8, CD25, Foxp3, T-bet, Gata3, Rorγt, Granzyme B, CD69, PD-1, CTLA-4), and macrophage/myeloid markers (CD11b, MHCII, CD206, CD40, CSF1R, PD-L1, Gr-1). In addition to immunophenotyping, cytokines are analyzed including, but not limited to, IP-10, TNFa, IL-17, IL-13, IL-12p70, IL12p40, IL-10, IL-6, IL-5, IL-4, IL-2, IL-1b, IFNy, GM-CSF, G-CSF, M-CSF, MIG, IP10, MIP1b, RANTES, and MCP-1, using techniques known in the art. Cytokine analysis may be analyzed on immune cells obtained from lymph nodes or other tissue, tumor sections, dissociated tumor cells, and/or on purified CD45+ tumor-infiltrated immune cells obtained ex vivo. Cytokine levels may be

ascertained using various staining techniques, qPCR, or other techniques known in the arts. Immunohistochemistry may also be performed on tumor sections to measure T cells, macrophages, dendritic cells, and checkpoint molecule or other protein expression using techniques known in the art.

[00175] Mice were inoculated with CT26 tumor cells, divided into groups as described above, and orally gavaged with *B. animalis ssp. lactis* Strain A 8.3x10^9 CFU/mouse daily. Other mice were given vehicle as a negative control. Mice from this study were sacrificed on day 10 and tumors harvested for analysis. Using known techniques described above, tumor sections were stained with anti-CD3 antibodies (T cell marker) to assess the average CD3+ cells per tumor section, and tumor gene expression was analyzed by qPCR using a TaqMan assay as follows: Tumor RNA was isolated with Qiagen's RNeasy Mini kit (Catalog No. 74104), using the manufacturer's protocol. cDNA was isolated using BioRad's iScript™ cDNA Synthesis Kit according to the manufacturer's protocol (Catalog No. 1708891). The qPCR reaction was run using SsoAdvanced™ Universal Probes Supermix 1725281 using manufacturer's protocol on the BioRad CFX384 instrument. The TaqMan mouse specific probes were from Invitrogen, MHC I (b2m) Mm00437762_m1 and β-actin Mm01205647_g1 for normalizing gene expression. The delta Ct value for each tumor was calculated for MHC I using β-actin as the reference gene. The expression for MHC I was calculated using the formula 2^-delta Ct for each mouse tumor and this value was divided by its respective final tumor volume at the end of the study.

[00176] Tumor volumes were taken prior to CD3+ immune cell infiltrate and MHC Class I expression analysis. As described in **Figure 1**, tumor volume was notably decreased on day 9 in the *B. animalis ssp. lactis* group (**Figure 6A**). Infiltration of CD3+ immune cells was significantly increased in the *Bifidobacterium animalis ssp. lactis* Strain A group relative to vehicle **(Figure 6B)**, and *B. animalis ssp. lactis* Strain A induced a striking upregulation of MHC Class I expression **(Figure 6C).**

[00177] Rather than being sacrificed, some mice may be rechallenged with tumor cell injection into the contralateral flank (or other area) to determine the impact of the immune system's memory response on tumor growth.

Example 8: *Bifidobacterium animalis ssp. lactis* in a mouse lung cancer model

[00178] *Bifidobacterium animalis ssp. lactis* is tested for its efficacy in the mouse lung cancer model, either alone or in combination with other cancer therapies, including checkpoint inhibitor(s). Mice are divided into groups receiving *Bifidobacterium animalis ssp. lactis*, with or without checkpoint inhibitor treatment. As described in Example 6, *Bifidobacterium animalis ssp. lactis* is administered at varied doses at defined intervals. For example, some mice receive *Bifidobacterium animalis ssp. lactis* (p.o.) on the day following tumor cell injection (day 1). Some mice receive seven (7) consecutive doses of *Bifidobacterium animalis ssp. lactis* (one dose per day on days 14-21). Other mice receive daily dosing or, alternatively, some mice receive dosing every other day. Alternatively, mice are randomized into various treatment groups at a defined timepoint (*e.g.* on day 13) or when the tumors reach a certain size (*e.g.* 100 mm^3) and treatment is then initiated accordingly.

[00179] $1x10^6$ LLC1 cells, or an appropriate number of lung cancer cells from another lung cancer cell line, are injected into the hind flank of syngeneic mice. Tumors from the various treatment groups are measured with calipers at regular intervals. As described in Example 6, some mice are sacrificed for ex vivo tumor analysis using flow cytometry. Other mice may be rechallenged with tumor cell injection into the contralateral flank to determine the impact of the immune system's memory response on tumor growth.

Example 9: *Bifidobacterium animalis ssp. lactis* in a mouse breast cancer model

[00180] *Bifidobacterium animalis ssp. lactis* is tested for its efficacy in the mouse breast cancer model, either alone or in combination with other cancer therapies, including checkpoint inhibitor(s). Mice are divided into groups receiving *Bifidobacterium animalis ssp. lactis*, with or without checkpoint inhibitor treatment. As described in Example 6, *Bifidobacterium animalis ssp. lactis* is administered at varied doses at defined intervals. For example, some mice receive *Bifidobacterium animalis ssp. lactis* (p.o.) on the day following tumor cell injection (day 1). Some mice receive seven (7) consecutive doses of *Bifidobacterium animalis ssp. lactis* (one dose per day on days 14-21). Other mice receive daily dosing or, alternatively, some mice receive dosing every other day. Alternatively, mice are randomized into various treatment groups at a defined timepoint (*e.g.* on day 13) or when the tumors reach a certain size (*e.g.* 100 mm^3) and treatment is then initiated accordingly.

[00181] 4T1 mouse mammary carcinoma cells are obtained from ATCC and $1x10^6$ cells in 50ul PBS are injected subcutaneously into one or both hind limbs of Balb/c female mice (as described by Wang *et al.* 2003, Systemic dissemination of viral vectors during intratumoral injection. Molecular Cancer Therapeutics; 2(11)). Alternatively, EMT6 mouse mammary carcinoma cells are obtained from ATCC and $1x10^6$ cells in 50µl PBS are injected subcutaneously into one or both of the hind limbs of Balb/c female mice 6-8 weeks old (as described by Guo *et al.* 2014, Combinatorial Photothermal and Immuno Cancer Therapy Using Chitosan-Coated Hollow Copper Sulfide Nanoparticles. ASC Nano.; 8(6): 5670-5681). In addition, other available mouse mammary cell lines may be used.

[00182] Tumors from the various treatment groups are measured with calipers at regular intervals. As described in Example 6, *Bifidobacterium animalis ssp. lactis* is administered at varied doses at defined intervals. For example, some mice are sacrificed for ex vivo tumor analysis using flow cytometry. Other mice may be rechallenged with tumor cell injection into the contralateral flank to determine the impact of the immune system's memory response on tumor growth.

[00183] Alternatively, 4T1 cells can be used in an orthotopic murine model of breast cancer as described by Tao *et al.* (Tao *et al.* 2008. Imagable 4T1 model for the study of late stage breast cancer. 8: 288). Mice are sacrificed for ex vivo tumor analysis. Tumors are analyzed by flow cytometry and immunohistochemistry.

Example 10: *Bifidobacterium animalis ssp. lactis* in a mouse pancreatic cancer model

[00184] *Bifidobacterium animalis ssp. lactis* is tested for its efficacy in the mouse model of pancreatic cancer, either alone or in combination with other cancer therapies, including checkpoint inhibitor(s). Mice are divided into groups receiving *Bifidobacterium animalis ssp. lactis*, with or without checkpoint inhibitor treatment. As described in Example 6, some mice receive *Bifidobacterium animalis ssp. lactis* (p.o.) on the day following tumor cell injection (day 1). Some mice receive seven (7) consecutive doses of *Bifidobacterium animalis ssp. lactis* (one dose per day on days 14-21). Other mice receive daily dosing or, alternatively, some mice receive dosing every other day. Alternatively, mice are randomized into various treatment groups at a defined timepoint (*e.g.* on day 13) or when the tumors reach a certain size (*e.g.* 100 mm^3) and treatment is then initiated accordingly.

[00185] Panc02 cells are maintained in DMEM, supplemented with 10% fetal calf serum and 1% penicillin/streptomycin, and incubated at 37°C at 5% CO2. Female 8–10 week-old C57Bl/6 mice are obtained from Charles River, Inc. or other certified vendor. Female C57Bl/6 mice are injected subcutaneously into the right hind flank with $1x10^6$ Panc02 cells. This protocol is based on standard Panc02 tumor models (Maletzki *et al.* 2008. Pancreatic cancer regression by intratumoral injection of live streptococcus pyogenes in a syngeneic mouse model. Gut. 57:483-491). Tumors from the various treatment groups are measured with calipers at regular intervals. As described in Example 6, some mice are sacrificed for ex vivo tumor analysis using flow cytometry, while other mice are rechallenged to determine the impact of the memory response on tumor growth.

[00186] Alternatively, Panc02, 6606PDA, or Capan-1 cells lines can be used in an orthotopic murine model of pancreatic cancer as described by Partecke *et al.* (Partecke *et al.* 2011. A syngeneic orthotopic murine model of pancreatic adenocarcinoma in the C57/Bl6 mouse using the Panc02 and 6606PDA cell lines. Eur. Surg. Res. 47(2):98-107) or Chai *et al.* (Chai *et al.* 2013. Bioluminescent orthotopic model of pancreatic cancer progression. J. Vis. Exp. 76: 50395). Mice are sacrificed for ex vivo tumor analysis. Tumors are analyzed by flow cytometry and immunohistochemistry.

Example 11: *Bifidobacterium animalis ssp. lactis* in a mouse model of hepatocellular carcinoma

[00187] *Bifidobacterium animalis ssp. lactis* is tested for its efficacy in the mouse model of hepatocellular carcinoma, either alone or in combination with other cancer therapies, including checkpoint inhibitor(s). Mice are divided into groups receiving *Bifidobacterium animalis ssp. lactis*, with or without checkpoint inhibitor treatment. As described in Example 6, *Bifidobacterium animalis ssp. lactis* is administered at varied doses at defined intervals. For example, some mice receive *Bifidobacterium animalis ssp. lactis* (p.o.) on the day following tumor cell injection (day 1). Some mice receive seven (7) consecutive doses of *Bifidobacterium animalis ssp. lactis* (one dose per day on days 14-21). Other mice receive daily dosing or, alternatively, some mice receive dosing every other day. Alternatively, mice are randomized into various treatment groups at a defined timepoint (*e.g.* on day 13) or when the tumors reach a certain size (*e.g.* 100 mm^3) and treatment is then initiated accordingly.

[00188] Hepatocellular carcinoma is induced in mice by subcutaneous inoculation of $1x10^6$ Hepa129 cells (obtained from NCI or other source), or an appropriate number of cells from other hepatocellular carcinoma cell line (as described by Gonzalez-Carmona *et al.* 2008. CD40 ligand-expressing dendritic cells induce regression of hepatocellular carcinoma by activating innate and acquired immunity in vivo. Hepatology. 48(1):157-168). Tumor cells are inoculated into one or both flanks. Tumors from the various treatment groups are measured with calipers at regular intervals. As described in Example 6, some mice are sacrificed for ex vivo tumor analysis using flow cytometry, while other mice are rechallenged to determine the impact of the memory response on tumor growth.

Example 12: *Bifidobacterium animalis ssp. lactis* in a mouse lymphoma model

[00189] *Bifidobacterium animalis ssp. lactis* is tested for its efficacy in the mouse model of lymphoma, either alone or in combination with other cancer therapies, including checkpoint inhibitor(s). Mice are divided into groups receiving *Bifidobacterium animalis ssp. lactis*, with or without checkpoint inhibitor treatment. As described in Example 6, *Bifidobacterium animalis ssp. lactis* is administered at varied doses at defined intervals. For example, some mice receive *Bifidobacterium animalis ssp. lactis* (p.o.) on the day following tumor cell injection (day 1). Some mice receive seven (7) consecutive doses of *Bifidobacterium animalis ssp. lactis* (one dose per day on days 14-21). Other mice receive daily dosing or, alternatively, some mice receive dosing every other day. Alternatively, mice are randomized into various treatment groups at a defined timepoint (*e.g.* on day 13) or when the tumors reach a certain size (*e.g.* 100 mm^3) and treatment is then initiated accordingly.

[00190] One lymphoma cell line is the A20 lymphoma, although other lymphoma cell lines may be used with syngeneic mice. A20 lymphoma cells are obtained from ATCC and $5x10^6$ cells in 50ul PBS are injected subcutaneously into one or both of the hind limbs of Balb/c female mice (as described by Houot *et al.* 2009. T-cell modulation combined with intratumoral CpG cures lymphoma in a mouse model without the need for chemotherapy. Blood. 113(15): 3546-3552). Tumors from the various treatment groups are measured with calipers at regular intervals. As described in Example 6, some mice are sacrificed for ex vivo tumor analysis using flow cytometry, while other mice are rechallenged to determine the impact of the memory response on tumor growth.

Example 13: *Bifidobacterium animalis ssp. lactis* in a mouse prostate cancer model

[00191] *Bifidobacterium animalis ssp. lactis* is tested for its efficacy in the mouse model of prostate cancer, either alone or in combination with other cancer therapies, including checkpoint inhibitor(s). Mice are divided into groups receiving *Bifidobacterium animalis ssp. lactis*, with or without checkpoint inhibitor treatment. As described in Example 6, *Bifidobacterium animalis ssp. lactis* is administered at varied doses at defined intervals. For example, some mice receive *Bifidobacterium animalis ssp. lactis* (p.o.) on the day following tumor cell injection (day 1). Some mice receive seven (7) consecutive doses of *Bifidobacterium animalis ssp. lactis* (one dose per day on days 14-21). Other mice receive daily dosing or, alternatively, some mice receive dosing every other day. Alternatively, mice are randomized into various treatment groups at a defined timepoint (*e.g.* on day 13) or when the tumors reach a certain size (*e.g.* 100 mm^3) and treatment is then initiated accordingly.

[00192] Mouse prostate cancer cells ($1x10^5$ RM-1 cells or an appropriate number of cells from another prostate cancer cell line) are injected into syngeneic mice. Tumors from the various treatment groups are measured with calipers at regular intervals. As described in Example 6, some mice are sacrificed for ex vivo tumor analysis using flow cytometry, while other mice are rechallenged to determine the impact of the memory response on tumor growth.

Example 14: *Bifidobacterium animalis ssp. lactis* in a mouse plasmacytoma model

[00193] *Bifidobacterium animalis ssp. lactis* is tested for its efficacy in the mouse model of plasmacytoma, either alone or in combination with other cancer therapies, including checkpoint inhibitor(s). Mice are divided into groups receiving *Bifidobacterium animalis ssp. lactis*, with or without checkpoint inhibitor treatment. As described in Example 6, *Bifidobacterium animalis ssp. lactis* is administered at varied doses at defined intervals. For example, some mice receive *Bifidobacterium animalis ssp. lactis* (p.o.) on the day following tumor cell injection (day 1). Some mice receive seven (7) consecutive doses of *Bifidobacterium animalis ssp. lactis* (one dose per day on days 14-21). Other mice receive daily dosing or, alternatively, some mice receive dosing every other day. Alternatively, mice are randomized into various treatment groups at a defined timepoint (*e.g.* on day 13) or when the tumors reach a certain size (*e.g.* 100 mm^3) and treatment is then initiated accordingly.

Mineral Oil Induced model of plasmacytoma

[00194] To examine the efficacy of *Bifidobacterium animalis ssp. lactis* in a plasmacytoma or multiple myeloma model, mice are injected intraperitoneally three times with 500ul of 2,6,10,12-tetramethylpentadecane ("pristane oil") at various time points between 0 and 60 days, as described by Potter *et al.* 1983. Peritoneal plasmacytomagenesis in mice: comparison of different pristane dose regimens. J. Natl. Cancer Inst. 71(2):391-5 (see also Lattanzio *et al.* 1997. Defective Development of Pristane-Oil Induced Plasmacytomas in Interleukin-6-Deficient BALB/C Mice. Am. J. Pathology: 151(3):689696). Progression of disease is measured by the degree of abdominal swelling and immune cells and particles in the ascites. Ascites fluid is analyzed for immune cell phenotype by flow cytometry as described in Example 2.

Cell-line induced model of plasmacytoma

[00195] To examine the efficacy of *Bifidobacterium animalis ssp. lactis* in a plasmacytoma or multiple myeloma model, either MOPC-104E cells or J558 plasmacytoma cells (TIB-6 ATCC) are injected subcutaneously into one or more hind flanks of Balb/c mice ($5x10^6$ cells), based on model described by Bhoopalam *et al.* 1980. Effect of dextran-S (alpha, 1-3 dextran) on the growth of plasmacytomas MOPC-104E and J558. J. Immunol. 125(4):1454-8 (see also Wang *et al.* 2015. IL-10 enhances CTL-mediated tumor rejection by inhibiting highly suppressive CD4+ T cells and promoting CTL persistence in a murine model of plasmacytoma. OncoImmunology. 4(7): e1014232-1-9). Mice are divided into groups receiving *Bifidobacterium animalis ssp. lactis* by oral gavage, and with or without checkpoint inhibitor treatment. Tumors from the various treatment groups are measured with calipers at regular intervals. As described in Example 6, some mice are sacrificed for ex vivo tumor analysis using flow cytometry, while other mice are rechallenged to determine the impact of the memory response on tumor growth.

Example 15: *Bifidobacterium animalis ssp. lactis* in a SCID mouse model of mouse myeloma

[00196] *Bifidobacterium animalis ssp. lactis* is tested for its efficacy in the SCID mouse model of myeloma, either alone or in combination with other cancer therapies, including checkpoint inhibitor(s). Mice are divided into groups receiving *Bifidobacterium animalis ssp. lactis*, with or without checkpoint inhibitor treatment. As described in Example 6, *Bifidobacterium animalis ssp. lactis* is administered at varied doses at defined intervals. For example, some mice receive *Bifidobacterium animalis ssp. lactis* (p.o.) on the day following

tumor cell injection (day 1). Some mice receive seven (7) consecutive doses of *Bifidobacterium animalis ssp. lactis* (one dose per day on days 14-21). Other mice receive daily dosing or, alternatively, some mice receive dosing every other day. Alternatively, mice are randomized into various treatment groups at a defined timepoint (*e.g.* on day 13) or when the tumors reach a certain size (*e.g.* 100 mm^3) and treatment is then initiated accordingly.

[00197] To examine the efficacy of *Bifidobacterium animalis ssp. lactis* using a human plasma cell leukemia, $1x10^7$ human myeloma cell lines, ARH77 cells (ARH77-ATCC CRL-1621, or an appropriate number of cells from another myeloma cell line such as KPMM2) are used. Myeloma cells are injected subcutaneously into one or both hind flanks of SCID mice (See Caers *et al.* 2004. Of mice and men: disease models of multiple myeloma. Drug Discovery Today: Disease Models. 1(4):373-380. Tumors from the various treatment groups are measured with calipers at regular intervals. As described in Example 6, some mice are sacrificed for ex vivo tumor analysis using flow cytometry, while other mice are rechallenged to determine the impact of the memory response on tumor growth.

Example 16: *Bifidobacterium animalis ssp. lactis* in a mouse renal cell carcinoma model

[00198] *Bifidobacterium animalis ssp. lactis* is tested for its efficacy in the mouse model of renal cell carcinoma, either alone or in combination with other cancer therapies, including checkpoint inhibitor(s). Mice are divided into groups receiving *Bifidobacterium animalis ssp. lactis*, with or without checkpoint inhibitor treatment. As described in Example 6, *Bifidobacterium animalis ssp. lactis* is administered at varied doses at defined intervals. For example, some mice receive *Bifidobacterium animalis ssp. lactis* (p.o.) on the day following tumor cell injection (day 1). Some mice receive seven (7) consecutive doses of *Bifidobacterium animalis ssp. lactis* (one dose per day on days 14-21). Other mice receive daily dosing or, alternatively, some mice receive dosing every other day. Alternatively, mice are randomized into various treatment groups at a defined timepoint (*e.g.* on day 13) or when the tumors reach a certain size (*e.g.* 100 mm^3) and treatment is then initiated accordingly.

[00199] To examine the efficacy of *Bifidobacterium animalis ssp. lactis* in a mouse model of renal cell carcinoma, Renca cells (ATCC CRL-2947) or other renal cell carcinoma cells are injected subcutaneously into one or both flanks of 7-8 week old syngeneic Balb/c mice ($5x10^6$ in

0.1 ml PBS). Tumors from the various treatment groups are measured with calipers at regular intervals. As described in Example 6, some mice are sacrificed for ex vivo tumor analysis using flow cytometry, while other mice are rechallenged to determine the impact of the memory response on tumor growth.

Example 17: *Bifidobacterium animalis ssp. lactis* in a mouse bladder cancer model

[00200] *Bifidobacterium animalis ssp. lactis* is tested for its efficacy in the mouse model of bladder cancer, either alone or in combination with other cancer therapies, including checkpoint inhibitor(s). Mice are divided into groups receiving *Bifidobacterium animalis ssp. lactis*, with or without checkpoint inhibitor treatment. As described in Example 6, *Bifidobacterium animalis ssp. lactis* is administered at varied doses at defined intervals. For example, some mice receive *Bifidobacterium animalis ssp. lactis* (p.o.) on the day following tumor cell injection (day 1). Some mice receive seven (7) consecutive doses of *Bifidobacterium animalis ssp. lactis* (one dose per day on days 14-21). Other mice receive daily dosing or, alternatively, some mice receive dosing every other day. Alternatively, mice are randomized into various treatment groups at a defined timepoint (*e.g.* on day 13) or when the tumors reach a certain size (*e.g.* 100 mm^3) and treatment is then initiated accordingly.

[00201] On the day of inoculation, MBT-2 cells (or other bladder cancer cell line) are harvested and resuspended in 1:1 PBS/Matrigel mixture. $2x10^5$ MBT-2 cells are suspended in 100 ul of mixture and injected subcutaneously into one or both hind flanks of syngeneic mice. Tumors are measured with calipers at regular intervals.

[00202] As described in Example 6, some mice are sacrificed for ex vivo tumor analysis using flow cytometry, while other mice are rechallenged to determine the impact of the memory response on tumor growth.

Example 18: The efficacy of a *Bifidobacterium animalis ssp. lactis* strain is compared to other *Bifidobacterium* strains for efficacy in cancer models

[00203] Using methods described above, various *Bifidobacterium animalis ssp. lactis* and other *Bifidobacterium* strains are tested and compared for efficacy in various cancer models. Such models may include, but are not limited to, melanoma, lung cancer, breast cancer, colon

cancer, colorectal cancer, pancreatic cancer, hepatocellular cancer, lymphoma, prostate cancer, plasmacytoma, a SCID model of myeloma, renal cell carcinoma, and/or bladder cancer.

Example 19: An open-label study of the safety, tolerability and efficacy of a *Bifidobacterium animalis ssp. lactis* Strain A oral therapeutic in patients with metastatic colorectal carcinoma

[00204] A multi-center, open-label clinical study with dose escalations and dose expansions to assess preliminary safety, tolerability, and efficacy of the *Bifidobacterium animalis ssp. lactis* Strain A is performed. The study proceeds in three parts: Part A: Dose escalation in microsatellite stable (MSS) metastatic colorectal carcinoma (CRC) and alternative indications, Part B: MSS CRC and alternative indications, and Part C: microsatellite instable (MSI) CRC. Dose escalation occurs in a standard 3+3 design. In part A, from 9 to 18 patients are enrolled. Parts B and C are enrolled concurrently. In Part B, up to 15 and no fewer than 10 pateints are enrolled. In Part C no fewer than 5 and up to 10 patients are enrolled. Based on in vivo studies of allograft bearing mice, an exemplary estimated human effective dose is $2x10^{11}$ organisms per day, delivered per enteric capsule. A schematic overview of the study is depicted in Figure 7.

[00205] The primary objectives of the study include the determination of the maximum tolerated dose within the tested dose range for *Bifidobacterium animalis ssp. lactis* Strain A in patients wiuth advanced malignancies; the determination of the recommended phase 2 dose for *Bifidobacterium animalis ssp. lactis* Strain A in CRC; the evaluation of the safety and tolerability, including dose limiting toxicities of *Bifidobacterium animalis ssp. lactis* Strain; and the demonstration of anti-tumor activity of *Bifidobacterium animalis ssp. lactis* Strain A in patients with CRC (*MSI and MSS*). The anti-tumor activity will be assessed by monitoring changes in circulating tumor cells, immune cell subsets in blood and tumor, and tumor viability; objective response rate and duration of response; progression-free survival; overall survival; clinical benefit rate and duration of clinical benefit rate; and disease control rate and duration of disease control rate. The secondary objective is to determine the human distribution and elimination of *Bifidobacterium animalis ssp. lactis* Strain A. The exploratory objectives is to evaluate correlations of *Bifidobacterium animalis ssp. lactis* Strain A response with molecular

markers relevant to each tumor type and to identify possible mechanisms of resistance to *Bifidobacterium animalis ssp. lactis* Strain A.

Inclusion and Exclusion Criteria:

[00206] The inclusion criteria for all parts of the study include the following:

1. Written informed consent obtained prior to any screening procedures and in accordance with federal, local, and institutional guidelines.
2. Age ≥ 18 years.
3. Adequate hepatic function:
    a. total bilirubin ≤ 2 times the upper limit of normal (ULN) (except patients with Gilbert's syndrome [hereditary indirect hyperbilirubinemia] who must have a total bilirubin of ≤ 3 times ULN),
    b. aspartate aminotransferase (AST) and alanine aminotransferase (ALT) ≤ 2.5 times ULN (except patients with known liver involvement of their tumor who must have their AST and ALT ≤ 5.0 times ULN).
4. Adequate renal function: estimated creatinine clearance of ≥ 30 mL/min, calculated using the formula of Cockcroft and Gault (140-Age) • Mass (kg)/(72 • creatinine mg/dL); multiply by 0.85 if female.
5. All patients in part B + C must be willing to have fresh biopsies at start of therapy and after 4 weeks of therapy.
6. Contraception: Female patients of child-bearing potential must agree to use dual methods of contraception (including one highly effective and one effective method of contraception) and have a negative serum pregnancy test at Screening, and male patients must use an effective barrier method of contraception if sexually active with a female of child-bearing potential. For both male and female patients, effective methods of contraception must be used throughout the study and for 3 months following the last dose.

[00207] For the portions of the study testing relapsed/refactory colorectal cancer, the inclusion criteria also include the following:

7. Histological or cytological documentation of adenocarcinoma of the colon or rectum.
8. Known MSI/MSS status.

9. Measurable disease by RECIST v1.1.
10. Metastatic disease not suitable for upfront curative-intent surgery.
11. Documented evidence of progressive disease according to RECIST v1.1.
12. Prior treatment (with completion of a course of therapy, or to disease progression or intolerability) with each of the following:
    a. Fluoropyrimidine-, oxaliplatin-, irinotecan-based chemotherapies (e.g., FOLFOX and FOLFIRI)
    b. if KRAS wild-type, an anti-EGFR therapy,
    c. Regorafenib or TAS 102 (Past A required, parts B+C optional)
    d. Radiation and surgery are not considered as prior anticancer regimens
13. Patients should not be transfusion dependent.
14. Adequate hematopoietic function: ANC $\geq$ 1000/mm3, hemoglobin (Hb) $\geq$ 9.0 g/dL, and platelet count $\geq$ 100,000/mm3.
15. Eastern Cooperative Oncology Group (ECOG) performance status of $\leq$ 1.
16. Life expectancy of $\geq$ 3 months.

[00208] The following categories of patient are excluded from the study:

1. Female patients who are pregnant or lactating.
2. Major surgery within 4 weeks before C1D1.
3. Impaired cardiac function or clinically significant cardiac diseases, including any of the following:
    a. Unstable angina or acute myocardial infarction $\leq$ 3 months prior to C1D1;
    b. Clinically significant heart disease (e.g., symptomatic congestive heart failure [e.g., >NYHA Class 2]; uncontrolled arrhythmia, or hypertension; history of labile hypertension or poor compliance with an antihypertensive regimen).
4. Uncontrolled active severe systemic infection requiring parenteral antibiotics, antivirals, or antifungals within one week prior to C1D1.
5. Any ongoing antibiotic treatment which has not been discontinued at least 3 daus prior to initiation of therapy
6. Patients with known symptomatic brain metastasis are not suitable for enrollment. Patients with asymptomatic, stable, treated brain metastases are eligible for study entry.

7. Patients with a known history of human immunodeficiency virus (HIV); HIV testing is not required as part of this study.
8. Known, active hepatitis A, B, or C infection; or known to be positive for HCV RNA or HBsAg (HBV surface antigen).
9. Prior malignancies:
    a. Patients with adequately resected basal or squamous cell carcinoma of the skin, or adequately resected carcinoma in situ (i.e. cervix) may enroll irrespective of the time of diagnosis.
    b. Prior malignancies which may interfere with the interpretation of the study. Cancer treated with curative intent < 5 years previously will not be allowed unless approved by the Sponsor. Cancer treated with curative intent > 5 years previously and without evidence of recurrence will be allowed.
10. Patients with active central nervous system (CNS) malignancy. Patients who have only had prophylactic intrathecal or intravenous chemotherapy against CNS disease are eligible.
11. Patients with gastrointestinal tract disease (or uncontrolled vomiting or diarrhea) that could interfere with the absorption of EVP001.
12. Serious psychiatric or medical conditions that, in the opinion of the Investigator, could interfere with treatment, compliance, or the ability to give consent.
13. Patients unwilling to comply with the protocol including required biopsies and sample collections required to measure disease.
14. Radiotherapy within two weeks prior to screening. Patients must have recovered from clinically significant toxicities.

[00209] The use of any concomitant medication/therapy, including over-the-counter (OTC) medications deemed necessary for the care of the patient is permitted during the study. Medications required to treat AEs, manage cancer symptoms, concurrent stable diseases and supportive care agents (e.g. blood product transfusions), pain medications, anti-emetics, and anti-diarrheals are allowed. Concurrent therapy with growth factors is allowed. The use of any immunosuppressive agents must be discussed between the Investigator and the Medical Monitor on a case-by-case basis. Any diagnostic, therapeutic, or surgical procedure performed during the

study period should be recorded, including the dates, description of the procedure(s), and any clinical findings, if applicable. All antibiotics are contraindicated.

[00210] Hormonal contraceptives are permitted in women of child-bearing potential. Hormonal contraceptives include any marketed contraceptive agent that includes an estrogen and/or a progestational agent.

[00211] Investigational or commercial anticancer agents other than *Bifidobacterium animalis ssp. lactis* Strain A is not allowed during the study. The initiation of any non-protocol specific anti-tumor treatment is considered an indication of disease relapse/progression and should be recorded appropriately in the electronic case report forms.

[00212] Palliative radiation therapy to non-target lesions is permitted but study treatment is held for ≥1 day before the start of palliative radiation therapy and ≥ 1 day following each dose of palliative radiation therapy. Treatment with *Bifidobacterium animalis ssp. lactis* Strain A is not discontinued solely due to palliative radiation.

[00213] Supportive measures for optimal medical care should be provided to patients during participation in this study. These should be based on institutional and/or National Comprehensive Cancer Network (NCCN) guidelines.

Dose escalation study

[00214] Patients receive all *Bifidobacterium animalis ssp. lactis* Strain A doses during the 7 day treatment period, or have had a dose-limiting toxicity (DLT) within the treatment period to be considered evaluable for dose escalation decisions. *Bifidobacterium animalis ssp. lactis* Strain A is orally administered as tablets or enteric coated capsules. Dose escalation decisions occur when the cohort of patients has met these criteria.

[00215] A DLT is defined as an adverse event (AE) or abnormal laboratory value that occurs within the first 7 days of treatment with *Bifidobacterium animalis ssp. lactis* Strain A, except for those that are clearly and incontrovertibly due to underlying disease, disease progression, or extraneous causes, and meets any of the criteria included in Table 4. National Cancer Institute Common Terminology Criteria for Adverse Events (NCI CTCAE) (Version 4.03) is used for all grading. In addition, > 4 missed doses of *Bifidobacterium animalis ssp. lactis* Strain A will be considered a DLT. Dose escalation decisions occur when the cohort of patients has met these criteria.

Table 4: Criteria for defining DLT

Toxicity	Any of the following criteria (based on CTCAE [Version 4.03]):
Non-Hematologic	Grade ≥ 3 nausea/vomiting, while taking optimal supportive medications.
	Any other Grade ≥ 3 non-hematological toxicity except alopecia or electrolyte abnormalities correctable with supportive therapy.
Hematologic	Grade 4 neutropenia lasting more than 5 days.
	Febrile neutropenia (absolute neutrophil count [ANC] < 1 x 10^9/L, fever > 38.5°C.).
	Grade 4 thrombocytopenia, or Grade 3 thrombocytopenia with bleeding, or any requirement for platelet transfusion.
CTCAE Version 4.03 will be used for grading all AEs and laboratory abnormalities. Patients may receive supportive care as per local institutional guidelines.	

[00216] To implement dose escalation decisions, the available toxicity information (i.e., all AEs and all laboratory abnormalities regardless of DLT assessment) is evaluated by the enrolling Investigators and Sponsor medical monitor at a dose decision meeting or teleconference. Decisions are based on an evaluation of all relevant data available from all dose cohorts evaluated in the ongoing study including safety information, DLTs, all NCI CTCAE, Version 4.03 toxicity data during Cycle 1 from evaluable patients. Drug administration at the next higher dose cohort may not proceed until the Investigator receives written confirmation from Sponsor indicating that the results of the previous dose cohort were evaluated and that it is permissible to proceed to the next higher dose cohort.

[00217] Table 5 below describes the starting dose and the dose levels that may be evaluated during the study for all parts of the study.

Table 5: *Bifidobacterium animalis ssp. lactis* Strain A Dose Escalation Levels

Cohort	Dose Levels *QD dosing; 3 week cycles*	Number of Patients
1	2x10^8 organisms	3+3
2	1 x10^9 organisms	3+3
3	2x10^9 organisms	3+3

4 (optional)	$4x10^9$ organisms	3

[00218] A standard 3+3 dose escalation is conducted as follows:

- If 0 of 3 patients experiences a DLT, escalate to next higher dose cohort.
- If 1 of 3 patients experiences a DLT, that cohort will be expanded to 6 patients. If 1 of 6 patients experiences a DLT, escalate to the next higher dose cohort;
- If $\geq$ 2 of 3 or $\geq$ 2 of 6 patients experience a DLT, the maximum tolerated dose (MTD) is exceeded.
- If the MTD is exceeded, enrollment of additional patients will be at a lower dose level. If a starting dose does not clear DLT assessment, dose de-escalation will proceed using standard 3+3 rules

[00219] Intra-patient dose escalations are permitted for all cohorts after the intended dose level has been shown to be safe (i.e., all patients treated at the intended dose level completed DLT assessments and $\leq$ 1 patient experienced a DLT).

[00220] For dose escalation patients after 7 days and dose expansion patients, if an event meeting the definition of a DLT, but without necessarily occurring within the first 21 days, is observed in >33% of patients at any time, or if >33% of treated patients have withdrawn consent due to toxicity, enrollment will be held and a meeting with all Investigators and Sponsor will take place to review the events and discuss their clinical significance. Based on this review, the Sponsor may elect to reduce the dose for enrolled patients and to resume enrollment of the expansion cohort at this lower dose, or the enrollment into the expansion cohort may be stopped.

[00221] Safety and tolerability are evaluated by means of DLTs (dose escalation cohorts only), AE reports, physical examinations, electrocardiograms and laboratory safety evaluations.

[00222] Anti-tumor activity are assessed by the Investigator according to disease specific response criteria and described in terms of objective response rate, duration of response, progression-free survival, clinical benefit rate, overall survival, and disease control rate.

[00223] In Part A: Peripheral blood WBC subsets and cytokines are analyzed at baseline and 7 days. Cytokines are additionally analyzed at 24 and 48 hours.

[00224] In Parts B and C: Peripheral blood WBC subsets and cytokines are analyzed at baseline and 7 and 21 days. Baseline and day 28 tumor biopsies are analyzed for immune subset infiltration, as well as makers of angiogenesis, proliferation, and death.

[00225] Patients continue to receive *Bifidobacterium animalis ssp. lactis* Strain A until the patient has confirmed pharmacodynamics, withdraws consent, is lost to follow-up, experiences intolerable toxicity which precludes further treatment with *Bifidobacterium animalis ssp. lactis* Strain A, or treatment is discontinued at the discretion of the patient, Investigator, or Sponsor. Patients who have objective disease progression but have evidence of overall clinical benefit may, at the request of the treating physician, continue treatment with *Bifidobacterium animalis ssp. lactis* Strain A after discussion with the Medical Monitor.

Example 20: Intratumorally administered *Bifidobacterium animalis ssp. lactis* inhibits colorectal carcinoma tumor growth

[00226] As described in Examples 2 and 3, CT-26 tumor cells were subcutaneously injected into one hind flank of mice and animals were assigned into groups receiving the following treatments: 1) Sucrose Vehicle; 2) anti-PD-1 and anti-CTLA4; and 3) *Bifidobacterium animalis ssp. lactis* Strain A, anti-PD-1, and anti-CTLA4. $2x10^9$ bacterial cells were administered by oral gavage (p.o.) every day for 21 days. Anti-PD-1 antibodies were administered intraperitoneally (i.p.) at 200ug/mouse (100ul final volume) every four days, starting on day 1, six times in total. Anti-CTLA4 antibodies were administered intraperitoneally (i.p.) at 100ug/mouse (100ul final volume) every four days, starting on day 1, six times in total. The tumor volumes were measured as described above. The triple combination group of *Bifidobacterium animalis ssp. lactis* Strain A, anti-PD-1, and anti-CTLA4 showed significant tumor growth inhibition compared to the control group and anti-PD-1 and anti-CTLA4 group (See **Figure 8**).

Example 21: Genomic analysis of *Bifidobacterium animalis ssp. Lactis* Strain A

[00227] Over the duration of a CT26 model experiments as described above, tumor volume varied when some stains of Bifidobacterium animalis ssp. Lactis were administered. Negative performing microbes were determined through a three-way comparison of tumor volumes at termination of groups treated with microbes, anti-PD1, and vehicle. The distributions of tumor volumes at termination of the anti-PD1 group vs. the vehicle group were used as a benchmark to classify negative performance of microbes. If the distributions were more closely

overlaid with vehicle than the group treated with anti-PD1, the strain was considered a negative performing microbe.

[00228] For example, tumor volume was found to be higher for groups treated with two species of Bifidobacterium animalis lactis, Strain B and Strain C, compared to the group treated with Strain A.

[00229] Consistent with these findings, a comparison of in vitro signatures of the three strains from a mouse dendritic cell assay show that Strain A exhibits an immune profile with a decreased induction of pro-tumorigenic cytokines compared to Strain B and Strain C (see **Figures 9A** and **9B**). Several of these cytokines have been found to be associated with poorer prognosis in the serum of CRC patients (Mager et al., (2016) Cytokine-Induced Modulation of Colorectal Cancer. Frontiers in Oncology. 6. 96. 10.3389).

[00230] To elucidate the differences between the strains, Stains A, B, and C were sequenced to ~500x using Illumina MiSeq. The complete and circular Strain A genome (Table 1) was used as the reference to call single nucleotide polymorphisms (SNPs) between the strains. Coding regions were predicted, and SNPs that occurred in coding regions were recorded, along with the nature of the coding change (synonymous, missense, frameshift, ect.) Using groupings established through results in the CT26 tumor model, SNPs that were present in negative strains (Strains B and C) when compared to the reference Strain A were identified, and structures unique to positive strains (e.g., Strain A) were identified.

[00231] When compared to highly homologous non-efficacious strains, Strain A contains single nucleotide polymorphisms (SNPs) and or insertion/deletion events in the following genes listed in Tables 6 and 7.

Table 6: SNPs and/or insertion/deletions in Strain A compared to non-efficacious strains

gene	product	Type of Variant	Position in Gene	Strain A Base change	Strain A Amino Acid change
glcU	putative glucose uptake protein GlcU	Missense	301	C	Arg
glcU	putative glucose uptake protein GlcU	Missense	814	C	Pro

relA	Bifunctional (p)ppGpp synthase/hydrolase RelA	Missense	251	G	Gly
relA	Bifunctional (p)ppGpp synthase/hydrolase RelA	Missense	2053	G	Val
metF	5,10-methylenetetrahydrofolate reductase	Missense	527	T	Phe
nnr	Bifunctional NAD(P)H-hydrate repair enzyme Nnr	Missense	574	C	Pro
ykoE_1	Putative HMP/thiamine permease protein YkoE	Missense	533	C	Ala
tcrY	putative sensor histidine kinase TcrY	Missense	300	G	Gln
tcrY	putative sensor histidine kinase TcrY	Missense	49	C	Pro
dac	D-alanyl-D-alanine carboxypeptidase	Frameshift	199	DELETION	His
hypothetical protein	hypothetical protein	Missense	739	A	Ile
hypothetical protein	hypothetical protein	Frameshift	990	DELETION	His
rdgB	dITP/XTP pyrophosphatase	Missense	547	A	Ser
hypothetical protein	hypothetical protein	Missense	290	C	Ala
glnB	Nitrogen regulatory protein P-II	Missense	161	A	Asp
accC	Biotin carboxylase	Frameshift	826	DELETION	-
trxB_2	Thioredoxin reductase	Missense	926	T	Val

afsK	Serine/threonine-protein kinase AfsK	Missense	686	G	Gly
nhaK	Sodium, potassium, lithium and rubidium/H(+) antiporter	Missense	989	T	Val
hypothetical protein	hypothetical protein	Inframe_Insertion	45-77	INSERTION (GGCGCTCGGCACCCTGGCGATTGGCGCGGCAAC)	-
oxc_1	Oxalyl-CoA decarboxylase	Frameshift	183-184	DELETION	-
wbgU	UDP-N-acetylglucosamine 4-epimerase	Missense	7	G	Val
wcaJ	UDP-glucose:undecaprenyl-phosphate glucose-1-phosphate transferase	Missense	292	G	Gly
hypothetical protein	hypothetical protein	Inframe_Deletion	126-128	DELETION	-
yciV	5'-3' exoribonuclease	Missense	100	T	Ser
aspA	Aspartate ammonia-lyase	Missense	1355	C	Ala
tcrY	putative sensor histidine kinase TcrY	Missense	368	T	Phe
tcrY	putative sensor histidine kinase TcrY	Missense	278	C	Pro
nagC_2	N-acetylglucosamine repressor	Missense	457	G	Ala

Table 7: Genes with synonymous variants

gene	product	Position in Gene	Strain A Base change	Strain A Amino Acid change
hypothetical protein	hypothetical protein	753	T	Leu
hemN	Oxygen-independent coproporphyrinogen-III oxidase-like protein	912	A	Gly
hypothetical protein	hypothetical protein	951	G	Pro
hypothetical protein	hypothetical protein	1722	T	Pro
cytR_2	HTH-type transcriptional repressor CytR	372	C	Ile
egtC	Gamma-glutamyl-hercynylcysteine sulfoxide hydrolase	135	C	Ile
hypothetical protein	hypothetical protein	30	G	Arg

Incorporation by Reference

[00232] All publications patent applications mentioned herein are hereby incorporated by reference in their entirety as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference. In case of conflict, the present application, including any definitions herein, will control.

Equivalents

[00233] Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following claims.

What is claimed is:

1. A method of treating cancer in a subject comprising administering to the subject a bacterial composition comprising *Bifidobacterium animalis ssp. lactis*.

2. The method of claim 1, wherein the *Bifidobacterium animalis ssp. lactis* is a strain comprising at least 99% sequence identity to the nucleotide sequence of the *Bifidobacterium animalis ssp. lactis* Strain A (ATCC Deposit Number PTA-125097).

3. The method of claim 1, wherein the *Bifidobacterium animalis ssp. lactis* is a strain comprising at least 99.9% sequence identity to the nucleotide sequence of the *Bifidobacterium animalis ssp. lactis* Strain A (ATCC Deposit Number PTA-125097).

4. The method of claim 1, wherein the *Bifidobacterium animalis ssp. lactis* is the *Bifidobacterium animalis ssp. lactis* Strain A (ATCC Deposit Number PTA-125097).

5. The method of any one of claims 1 to 4, wherein the cancer is selected from the group consisting of hematological malignancy, acute nonlymphocytic leukemia, chronic lymphocytic leukemia, acute granulocytic leukemia, chronic granulocytic leukemia, acute promyelocytic leukemia, adult T-cell leukemia, aleukemic leukemia, a leukocythemic leukemia, basophilic leukemia, blast cell leukemia, bovine leukemia, chronic myelocytic leukemia, leukemia cutis, embryonal leukemia, eosinophilic leukemia, Gross' leukemia, Rieder cell leukemia, Schilling's leukemia, stem cell leukemia, subleukemic leukemia, undifferentiated cell leukemia, hairy-cell leukemia, hemoblastic leukemia, hemocytoblastic leukemia, histiocytic leukemia, stem cell leukemia, acute monocytic leukemia, leukopenic leukemia, lymphatic leukemia, lymphoblastic leukemia, lymphocytic leukemia, lymphogenous leukemia, lymphoid leukemia, lymphosarcoma cell leukemia, mast cell leukemia, megakaryocytic leukemia, micromyeloblastic leukemia, monocytic leukemia, myeloblastic leukemia, myelocytic leukemia, myeloid granulocytic leukemia, myelomonocytic leukemia, Naegeli leukemia, plasma cell leukemia, plasmacytic leukemia, promyelocytic leukemia, acinar carcinoma, acinous carcinoma, adenocystic carcinoma, adenoid cystic carcinoma, carcinoma adenomatosum, carcinoma of adrenal cortex, alveolar carcinoma, alveolar cell carcinoma, basal cell carcinoma, carcinoma basocellulare, basaloid carcinoma, basosquamous cell carcinoma, bronchioalveolar carcinoma, bronchiolar carcinoma, bronchogenic carcinoma, cerebriform carcinoma, cholangiocellular carcinoma,

chorionic carcinoma, colloid carcinoma, comedo carcinoma, corpus carcinoma, cribriform carcinoma, carcinoma en cuirasse, carcinoma cutaneum, cylindrical carcinoma, cylindrical cell carcinoma, duct carcinoma, carcinoma durum, embryonal carcinoma, encephaloid carcinoma, epiennoid carcinoma, carcinoma epitheliale adenoides, exophytic carcinoma, carcinoma ex ulcere, carcinoma fibrosum, gelatiniform carcinoma, gelatinous carcinoma, giant cell carcinoma, signet-ring cell carcinoma, carcinoma simplex, small-cell carcinoma, solanoid carcinoma, spheroidal cell carcinoma, spindle cell carcinoma, carcinoma spongiosum, squamous carcinoma, squamous cell carcinoma, string carcinoma, carcinoma telangiectaticum, carcinoma telangiectodes, transitional cell carcinoma, carcinoma tuberosum, tuberous carcinoma, verrucous carcinoma, carcinoma villosum, carcinoma gigantocellulare, glandular carcinoma, granulosa cell carcinoma, hair-matrix carcinoma, hematoid carcinoma, hepatocellular carcinoma, Hurthle cell carcinoma, hyaline carcinoma, hypernephroid carcinoma, infantile embryonal carcinoma, carcinoma in situ, intraepidermal carcinoma, intraepithelial carcinoma, Krompecher's carcinoma, Kulchitzky-cell carcinoma, large-cell carcinoma, lenticular carcinoma, carcinoma lenticulare, lipomatous carcinoma, lymphoepithelial carcinoma, carcinoma medullare, medullary carcinoma, melanotic carcinoma, carcinoma molle, mucinous carcinoma, carcinoma muciparum, carcinoma mucocellulare, mucoepidermoid carcinoma, carcinoma mucosum, mucous carcinoma, carcinoma myxomatodes, naspharyngeal carcinoma, oat cell carcinoma, carcinoma ossificans, osteoid carcinoma, papillary carcinoma, periportal carcinoma, preinvasive carcinoma, prickle cell carcinoma, pultaceous carcinoma, renal cell carcinoma of kidney, reserve cell carcinoma, carcinoma sarcomatodes, schneiderian carcinoma, scirrhous carcinoma, carcinoma scroti, chondrosarcoma, fibrosarcoma, lymphosarcoma, melanosarcoma, myxosarcoma, osteosarcoma, endometrial sarcoma, stromal sarcoma, Ewing' s sarcoma, fascial sarcoma, fibroblastic sarcoma, giant cell sarcoma, Abemethy's sarcoma, adipose sarcoma, liposarcoma, alveolar soft part sarcoma, ameloblastic sarcoma, botryoid sarcoma, chloroma sarcoma, chorio carcinoma, embryonal sarcoma, Wilms' tumor sarcoma, granulocytic sarcoma, Hodgkin's sarcoma, idiopathic multiple pigmented hemorrhagic sarcoma, immunoblastic sarcoma of B cells, lymphoma, immunoblastic sarcoma of T-cells, Jensen's sarcoma, Kaposi's sarcoma, Kupffer cell sarcoma, angiosarcoma, leukosarcoma, malignant mesenchymoma sarcoma, parosteal sarcoma, reticulocytic sarcoma, Rous sarcoma, serocystic sarcoma, synovial sarcoma, telangiectaltic sarcoma, Hodgkin's Disease, Non-Hodgkin's Lymphoma, multiple myeloma, neuroblastoma,

bladder cancer, breast cancer, ovarian cancer, lung cancer, rhabdomyosarcoma, primary thrombocytosis, primary macroglobulinemia, small-cell lung tumors, primary brain tumors, stomach cancer, colon cancer, malignant pancreatic insulanoma, malignant carcinoid, premalignant skin lesions, testicular cancer, lymphomas, thyroid cancer, neuroblastoma, esophageal cancer, genitourinary tract cancer, malignant hypercalcemia, cervical cancer, endometrial cancer, adrenal cortical cancer, Harding-Passey melanoma, juvenile melanoma, lentigo maligna melanoma, malignant melanoma, acral-lentiginous melanoma, amelanotic melanoma, benign juvenile melanoma, Cloudman's melanoma, S91 melanoma, nodular melanoma subungal melanoma, and superficial spreading melanoma.

6. The method of any one of claims 1 to 4, wherein the cancer is prostate cancer, lung cancer, colon cancer, colorectal cancer, melanoma, breast cancer, pancreatic cancer, hepatocellular carcinoma, or lymphoma.

7. The method of any preceding claim, wherein the bacterial composition is administered orally, rectally, intravenously, intratumorally, or subcutaneously.

8. The method of any one of claims 1 to 7, wherein at least 50% of the bacteria in the bacterial composition are *Bifidobacterium animalis ssp. lactis*.

9. The method of any one of claims 1 to 7, wherein at least 90% of the bacteria in the bacterial composition are *Bifidobacterium animalis ssp. lactis*.

10. The method of any one of claims 1 to 7, wherein substantially all of the bacteria in the bacterial composition are *Bifidobacterium animalis ssp. lactis*.

11. The method of any one of claims 1 to 10, wherein the bacterial composition comprises at least 1×10^6 colony forming units (CFUs) of *Bifidobacterium animalis ssp. lactis*.

12. The method of claim 11, wherein the bacterial composition comprises at least 1×10^7 CFUs of *Bifidobacterium animalis ssp. lactis*.

13. The method of claim 1, wherein the bacterial composition comprises at least 1×10^8 CFUs of *Bifidobacterium animalis ssp. lactis*.

14. The method of any one of claims 1 to 13, wherein the bacterial composition is administered in two or more doses.

15. The method of claim 14, wherein the administration to the subject of the two or more doses are separated by at least 1 day.

16. The method of claim 15, wherein the administration of the two or more doses are separated by at least 1 week.

17. The method of any one of claims 1 to 16, wherein the bacterial composition comprises live bacteria.

18. The method of any one of claims 1 to 16, wherein the bacterial composition comprises attenuated bacteria.

19. The method of any one of claims 1 to 19, wherein the bacterial composition comprises killed bacteria.

20. The method of any one of claims 1 to 19, wherein administration of the bacterial composition treats the cancer.

21. The method of any one of claims 1 to 20, wherein administration of the bacterial composition induces an anti-cancer immune response.

22. The method of any one of claims 1 to 21, wherein the method further comprises administering to the subject a second cancer therapy.

23. The method of claim 22, wherein the second cancer therapy comprises the administration of a chemotherapy agent to the subject.

24. The method of claim 23, wherein the chemotherapy agent is selected from the group consisting of thiotepa, cyclosphosphamide, busulfan, improsulfan, piposulfan, benzodopa, carboquone, meturedopa, uredopa, altretamine, triethylenemelamine, trietylenephosphoramide, triethiylenethiophosphoramide, trimethylolomelamine, bullatacin, bullatacinone, camptothecin, topotecan, bryostatin, callystatin, CC-1065, cryptophycin 1, cryptophycin 8, dolastatin, duocarmycin, eleutherobin, pancratistatin, sarcodictyin, spongistatin, chlorambucil, chlornaphazine, cholophosphamide, estramustine, ifosfamide, mechlorethamine, mechlorethamine oxide hydrochloride, melphalan, novembichin, phenesterine, prednimustine, trofosfamide, uracil mustard, carmustine, chlorozotocin, fotemustine, lomustine, nimustine, ranimnustine, calicheamicin, dynemicin, clodronate, esperamicin; neocarzinostatin chromophore,

aclacinomysins, actinomycin, authrarnycin, azaserine, bleomycins, cactinomycin, carabicin, caminomycin, carzinophilin, chromomycinis, dactinomycin, daunorubicin, detorubicin, 6-diazo-5-oxo-L-norleucine, doxorubicin, epirubicin, esorubicin, idarubicin, marcellomycin, mitomycin, mitomycin C, mycophenolic acid, nogalamycin, olivomycin, peplomycin, potfiromycin, puromycin, quelamycin, rodorubicin, streptonigrin, streptozocin, tubercidin, ubenimex, zinostatin, zorubicin, methotrexate, 5-fluorouracil (5-FU), denopterin, methotrexate, pteropterin, trimetrexate, fludarabine, 6-mercaptopurine, thiamiprine, thioguanine, ancitabine, azacitidine, 6-azauridine, carmofur, cytarabine, dideoxyuridine, doxifluridine, enocitabine, floxuridine, calusterone, dromostanolone propionate, epitiostanol, mepitiostane, testolactone, aminoglutethimide, mitotane, trilostane, frolinic acid, aceglatone, aldophosphamide glycoside, aminolevulinic acid, eniluracil, amsacrine, bestrabucil, bisantrene, edatraxate, defofamine, demecolcine, diaziquone, elformithine, elliptinium acetate, epothilone, etoglucid, gallium nitrate, hydroxyurea, lentinan, lonidainine, maytansine, ansamitocins, mitoguazone, mitoxantrone, mopidanmol, nitraerine, pentostatin, phenamet, pirarubicin, losoxantrone, podophyllinic acid, 2-ethylhydrazide, procarbazine, PSK polysaccharide complex, razoxane, rhizoxin, sizofuran, spirogermanium, tenuazonic acid, triaziquone; 2,2',2''-trichlorotriethylamine, trichothecene, T-2 toxin, verracurin A, roridin A, anguidine, urethane, vindesine, dacarbazine, mannomustine, mitobronitol, mitolactol, pipobroman, gacytosine, arabinoside, cyclophosphamide, thiotepa, paclitaxel, doxetaxel, chlorambucil, gemcitabine, 6-thioguanine, mercaptopurine, methotrexate, cisplatin, oxaliplatin, carboplatin, vinblastine, platinum, etoposide, ifosfamide, mitoxantrone, vincristine, vinorelbine, novantrone, teniposide, edatrexate, daunomycin, aminopterin, xeloda, ibandronate, irinotecan, RFS 2000, difluoromethylomithine, retinoic acid and capecitabine.

25. The method of any one of claims 22 to 24, wherein the second cancer therapy comprises cancer immunotherapy.

26. The method of claim 25, wherein the cancer immunotherapy comprises administering an immune checkpoint inhibitor to the subject.

27. The method of claim 26, wherein the cancer immunotherapy comprises administering an additional immune checkpoint inhibitor to the subject.

28. The method of claim 26 or claim 27, wherein the immune checkpoint inhibitor is an antibody or antigen-binding fragment thereof that specifically binds to an immune checkpoint protein.

29. The method of claim 28, wherein the immune checkpoint inhibitor is an siRNA molecule, an shRNA molecule or an antisense RNA molecule that inhibits expression of an immune checkpoint protein.

30. The method of claim 28 or 29, wherein the immune checkpoint protein is CTLA4, PD-1, PD-L1, PD-L2, A2AR, B7-H3, B7-H4, BTLA, KIR, LAG3, TIM-3 or VISTA.

31. The method of claim 26 or claim 27, wherein the immune checkpoint inhibitor is atezolizumab, avelumab, durvalumab, ipilimumab, nivolumab, pembrolizumab, pidilizumab, AMP-224, AMP-514, BGB-A317, STI-A1110, TSR-042, RG-7446, BMS-936559, MEDI-4736, MSB-0020718C, AUR-012 or STI-A1010.

32. The method of any one of claims 25 to 31, wherein the cancer immunotherapy comprises administration of a cancer-specific antibody or antigen-binding fragment thereof to the subject.

33. The method of claim 32, wherein the cancer-specific antibody or antigen-binding fragment thereof binds specifically to a cancer-associated antigen.

34. The method of claim 33, wherein the cancer-associated antigen is selected from the group consisting of adipophilin, AIM-2, ALDH1A1, alpha-actinin-4, alpha-fetoprotein ("AFP"), ARTC1, B-RAF, BAGE-1, BCLX (L), BCR-ABL fusion protein b3a2, beta-catenin, BING-4, CA-125, CALCA, carcinoembryonic antigen ("CEA"), CASP-5, CASP-8, CD274, CD45, Cdc27, CDK12, CDK4, CDKN2A, CEA, CLPP, COA-1, CPSF, CSNK1A1, CTAG1, CTAG2, cyclin D1, Cyclin-A1, dek-can fusion protein, DKK1, EFTUD2, Elongation factor 2, ENAH (hMena), Ep-CAM, EpCAM, EphA3, epithelial tumor antigen ("ETA"), ETV6-AML1 fusion protein, EZH2, FGF5, FLT3-ITD, FN1, G250/MN/CAIX, GAGE-1,2,8, GAGE-3,4,5,6,7, GAS7, glypican-3, GnTV, gp100/Pmel17, GPNMB, HAUS3, Hepsin, HER-2/neu, HERV-K-MEL, HLA-A11, HLA-A2, HLA-DOB, hsp70-2, IDO1, IGF2B3, IL13Ralpha2, Intestinal carboxyl esterase, K-ras, Kallikrein 4, KIF20A, KK-LC-1, KKLC1, KM-HN-1, KMHN1 also known as CCDC110, LAGE-1, LDLR-fucosyltransferaseAS fusion protein, Lengsin, M-CSF, MAGE-A1, MAGE-A10, MAGE-A12, MAGE-A2, MAGE-A3, MAGE-A4, MAGE-A6, MAGE-A9,

MAGE-C1, MAGE-C2, malic enzyme, mammaglobin-A, MART2, MATN, MC1R, MCSP, mdm-2, ME1, Melan-A/MART-1, Meloe, Midkine, MMP-2, MMP-7, MUC1, MUC5AC, mucin, MUM-1, MUM-2, MUM-3, Myosin, Myosin class I, N-raw, NA88-A, neo-PAP, NFYC, NY-BR-1, NY-ESO-1/LAGE-2, OA1, OGT, OS-9, P polypeptide, p53, PAP, PAX5, PBF, pml-RARalpha fusion protein, polymorphic epithelial mucin ("PEM"), PPP1R3B, PRAME, PRDX5, PSA, PSMA, PTPRK, RAB38/NY-MEL-1, RAGE-1, RBAF600, RGS5, RhoC, RNF43, RU2AS, SAGE, secernin 1, SIRT2, SNRPD1, SOX10, Sp17, SPA17, SSX-2, SSX-4, STEAP1, survivin, SYT-SSX1 or -SSX2 fusion protein, TAG-1, TAG-2, Telomerase, TGF-betaRII, TPBG, TRAG-3, Triosephosphate isomerase, TRP-1/gp75, TRP-2, TRP2-INT2, tyrosinase, tyrosinase ("TYR"), VEGF, WT1 and XAGE-1b/GAGED2a.

35. The method of claim 33, wherein the cancer associated antigen is a neo-antigen.

36. The method any one of claims 25 to 35, wherein the cancer immunotherapy comprises administration of a cancer vaccine to the subject.

37. The method of claim 36, wherein the cancer vaccine comprises a polypeptide comprising an epitope of a cancer-associated antigen.

38. The method of claim 37, wherein the cancer-associated antigen is selected from the group consisting of adipophilin, AIM-2, ALDH1A1, alpha-actinin-4, alpha-fetoprotein ("AFP"), ARTC1, B-RAF, BAGE-1, BCLX (L), BCR-ABL fusion protein b3a2, beta-catenin, BING-4, CA-125, CALCA, carcinoembryonic antigen ("CEA"), CASP-5, CASP-8, CD274, CD45, Cdc27, CDK12, CDK4, CDKN2A, CEA, CLPP, COA-1, CPSF, CSNK1A1, CTAG1, CTAG2, cyclin D1, Cyclin-A1, dek-can fusion protein, DKK1, EFTUD2, Elongation factor 2, ENAH (hMena), Ep-CAM, EpCAM, EphA3, epithelial tumor antigen ("ETA"), ETV6-AML1 fusion protein, EZH2, FGF5, FLT3-ITD, FN1, G250/MN/CAIX, GAGE-1,2,8, GAGE-3,4,5,6,7, GAS7, glypican-3, GnTV, gp100/Pmel17, GPNMB, HAUS3, Hepsin, HER-2/neu, HERV-K-MEL, HLA-A11, HLA-A2, HLA-DOB, hsp70-2, IDO1, IGF2B3, IL13Ralpha2, Intestinal carboxyl esterase, K-ras, Kallikrein 4, KIF20A, KK-LC-1, KKLC1, KM-HN-1, KMHN1 also known as CCDC110, LAGE-1, LDLR-fucosyltransferaseAS fusion protein, Lengsin, M-CSF, MAGE-A1, MAGE-A10, MAGE-A12, MAGE-A2, MAGE-A3, MAGE-A4, MAGE-A6, MAGE-A9, MAGE-C1, MAGE-C2, malic enzyme, mammaglobin-A, MART2, MATN, MC1R, MCSP,

mdm-2, ME1, Melan-A/MART-1, Meloe, Midkine, MMP-2, MMP-7, MUC1, MUC5AC, mucin, MUM-1, MUM-2, MUM-3, Myosin, Myosin class I, N-raw, NA88-A, neo-PAP, NFYC, NY-BR-1, NY-ESO-1/LAGE-2, OA1, OGT, OS-9, P polypeptide, p53, PAP, PAX5, PBF, pml-RARalpha fusion protein, polymorphic epithelial mucin ("PEM"), PPP1R3B, PRAME, PRDX5, PSA, PSMA, PTPRK, RAB38/NY-MEL-1, RAGE-1, RBAF600, RGS5, RhoC, RNF43, RU2AS, SAGE, secernin 1, SIRT2, SNRPD1, SOX10, Sp17, SPA17, SSX-2, SSX-4, STEAP1, survivin, SYT-SSX1 or -SSX2 fusion protein, TAG-1, TAG-2, Telomerase, TGF-betaRII, TPBG, TRAG-3, Triosephosphate isomerase, TRP-1/gp75, TRP-2, TRP2-INT2, tyrosinase, tyrosinase ("TYR"), VEGF, WT1 and XAGE-1b/GAGED2a.

39. The method of claim 37, wherein the cancer-associated antigen is a neo-antigen.

40. The method of any one of claim 37 to 39 wherein the polypeptide is a fusion protein.

41. The method of claim 36, wherein the cancer vaccine comprises a nucleic acid encoding an epitope of a cancer-associated antigen.

42. The method of claim 41, wherein the cancer-associated antigen is selected from the group consisting of adipophilin, AIM-2, ALDH1A1, alpha-actinin-4, alpha-fetoprotein ("AFP"), ARTC1, B-RAF, BAGE-1, BCLX (L), BCR-ABL fusion protein b3a2, beta-catenin, BING-4, CA-125, CALCA, carcinoembryonic antigen ("CEA"), CASP-5, CASP-8, CD274, CD45, Cdc27, CDK12, CDK4, CDKN2A, CEA, CLPP, COA-1, CPSF, CSNK1A1, CTAG1, CTAG2, cyclin D1, Cyclin-A1, dek-can fusion protein, DKK1, EFTUD2, Elongation factor 2, ENAH (hMena), Ep-CAM, EpCAM, EphA3, epithelial tumor antigen ("ETA"), ETV6-AML1 fusion protein, EZH2, FGF5, FLT3-ITD, FN1, G250/MN/CAIX, GAGE-1,2,8, GAGE-3,4,5,6,7, GAS7, glypican-3, GnTV, gp100/Pmel17, GPNMB, HAUS3, Hepsin, HER-2/neu, HERV-K-MEL, HLA-A11, HLA-A2, HLA-DOB, hsp70-2, IDO1, IGF2B3, IL13Ralpha2, Intestinal carboxyl esterase, K-ras, Kallikrein 4, KIF20A, KK-LC-1, KKLC1, KM-HN-1, KMHN1 also known as CCDC110, LAGE-1, LDLR-fucosyltransferaseAS fusion protein, Lengsin, M-CSF, MAGE-A1, MAGE-A10, MAGE-A12, MAGE-A2, MAGE-A3, MAGE-A4, MAGE-A6, MAGE-A9, MAGE-C1, MAGE-C2, malic enzyme, mammaglobin-A, MART2, MATN, MC1R, MCSP, mdm-2, ME1, Melan-A/MART-1, Meloe, Midkine, MMP-2, MMP-7, MUC1, MUC5AC, mucin, MUM-1, MUM-2, MUM-3, Myosin, Myosin class I, N-raw, NA88-A, neo-PAP, NFYC, NY-

BR-1, NY-ESO-1/LAGE-2, OA1, OGT, OS-9, P polypeptide, p53, PAP, PAX5, PBF, pml-RARalpha fusion protein, polymorphic epithelial mucin ("PEM"), PPP1R3B, PRAME, PRDX5, PSA, PSMA, PTPRK, RAB38/NY-MEL-1, RAGE-1, RBAF600, RGS5, RhoC, RNF43, RU2AS, SAGE, secernin 1, SIRT2, SNRPD1, SOX10, Sp17, SPA17, SSX-2, SSX-4, STEAP1, survivin, SYT-SSX1 or -SSX2 fusion protein, TAG-1, TAG-2, Telomerase, TGF-betaRII, TPBG, TRAG-3, Triosephosphate isomerase, TRP-1/gp75, TRP-2, TRP2-INT2, tyrosinase, tyrosinase ("TYR"), VEGF, WT1 and XAGE-1b/GAGED2a.

43. The method of claim 41, wherein the cancer-associated antigen is a neo-antigen.

44. The method of any one of claims 41 to 43, wherein the nucleic acid is DNA.

45. The method of any one of claims 41 to 43, wherein the nucleic acid is RNA.

46. The method of claim 45, wherein the RNA is mRNA.

47. The method of any one of claims 41 to 46, wherein the nucleic acid is in a vector.

48. The method of claim 47, wherein the vector is a bacterial vector.

49. The method of claim 48, wherein the bacterial vector is selected from the group consisting of Mycobacterium bovis (BCG), Salmonella Typhimurium ssp., Salmonella Typhi ssp., Clostridium sp. spores, Escherichia coli Nissle 1917, Escherichia coli K-12/LLO, Listeria monocytogenes, and Shigella flexneri.

50. The method of claim 47, wherein the vector is a viral vector.

51. The method of claim 50, wherein the viral vector is selected from the group consisting of vaccinia, adenovirus, RNA viruses, and replication-defective avipox, replication-defective fowlpox, replication-defective canarypox, replication-defective MVA and replication-defective adenovirus.

52. The method any one of claims 25 to 51, wherein the cancer immunotherapy comprises administration of an antigen presenting cell (APC) primed with a cancer-specific antigen.

53. The method of claim 51, wherein the APC is a dendritic cell, a macrophage or a B cell.

54. The method of claim 52 or claim 53, wherein the cancer-associated antigen is selected from the group consisting of adipophilin, AIM-2, ALDH1A1, alpha-actinin-4, alpha-fetoprotein

("AFP"), ARTC1, B-RAF, BAGE-1, BCLX (L), BCR-ABL fusion protein b3a2, beta-catenin, BING-4, CA-125, CALCA, carcinoembryonic antigen ("CEA"), CASP-5, CASP-8, CD274, CD45, Cdc27, CDK12, CDK4, CDKN2A, CEA, CLPP, COA-1, CPSF, CSNK1A1, CTAG1, CTAG2, cyclin D1, Cyclin-A1, dek-can fusion protein, DKK1, EFTUD2, Elongation factor 2, ENAH (hMena), Ep-CAM, EpCAM, EphA3, epithelial tumor antigen ("ETA"), ETV6-AML1 fusion protein, EZH2, FGF5, FLT3-ITD, FN1, G250/MN/CAIX, GAGE-1,2,8, GAGE-3,4,5,6,7, GAS7, glypican-3, GnTV, gp100/Pmel17, GPNMB, HAUS3, Hepsin, HER-2/neu, HERV-K-MEL, HLA-A11, HLA-A2, HLA-DOB, hsp70-2, IDO1, IGF2B3, IL13Ralpha2, Intestinal carboxyl esterase, K-ras, Kallikrein 4, KIF20A, KK-LC-1, KKLC1, KM-HN-1, KMHN1 also known as CCDC110, LAGE-1, LDLR-fucosyltransferaseAS fusion protein, Lengsin, M-CSF, MAGE-A1, MAGE-A10, MAGE-A12, MAGE-A2, MAGE-A3, MAGE-A4, MAGE-A6, MAGE-A9, MAGE-C1, MAGE-C2, malic enzyme, mammaglobin-A, MART2, MATN, MC1R, MCSP, mdm-2, ME1, Melan-A/MART-1, Meloe, Midkine, MMP-2, MMP-7, MUC1, MUC5AC, mucin, MUM-1, MUM-2, MUM-3, Myosin, Myosin class I, N-raw, NA88-A, neo-PAP, NFYC, NY-BR-1, NY-ESO-1/LAGE-2, OA1, OGT, OS-9, P polypeptide, p53, PAP, PAX5, PBF, pml-RARalpha fusion protein, polymorphic epithelial mucin ("PEM"), PPP1R3B, PRAME, PRDX5, PSA, PSMA, PTPRK, RAB38/NY-MEL-1, RAGE-1, RBAF600, RGS5, RhoC, RNF43, RU2AS, SAGE, secernin 1, SIRT2, SNRPD1, SOX10, Sp17, SPA17, SSX-2, SSX-4, STEAP1, survivin, SYT-SSX1 or -SSX2 fusion protein, TAG-1, TAG-2, Telomerase, TGF-betaRII, TPBG, TRAG-3, Triosephosphate isomerase, TRP-1/gp75, TRP-2, TRP2-INT2, tyrosinase, tyrosinase ("TYR"), VEGF, WT1 and XAGE-1b/GAGED2a.

55. The method of claim 52 or 53, wherein the cancer-associated antigen is a neo-antigen.

56. The method any one of claims 25 to 55, wherein the cancer immunotherapy comprises administration of a cancer-specific chimeric antigen receptor (CAR).

57. The method of claim 56, wherein the CAR is administered on the surface of a T cell.

58. The method of claim 56 or 57, wherein the CAR binds specifically to a cancer-associated antigen.

59. The method of claim 58, wherein the cancer-associated antigen is selected from the group consisting of adipophilin, AIM-2, ALDH1A1, alpha-actinin-4, alpha-fetoprotein ("AFP"),

ARTC1, B-RAF, BAGE-1, BCLX (L), BCR-ABL fusion protein b3a2, beta-catenin, BING-4, CA-125, CALCA, carcinoembryonic antigen ("CEA"), CASP-5, CASP-8, CD274, CD45, Cdc27, CDK12, CDK4, CDKN2A, CEA, CLPP, COA-1, CPSF, CSNK1A1, CTAG1, CTAG2, cyclin D1, Cyclin-A1, dek-can fusion protein, DKK1, EFTUD2, Elongation factor 2, ENAH (hMena), Ep-CAM, EpCAM, EphA3, epithelial tumor antigen ("ETA"), ETV6-AML1 fusion protein, EZH2, FGF5, FLT3-ITD, FN1, G250/MN/CAIX, GAGE-1,2,8, GAGE-3,4,5,6,7, GAS7, glypican-3, GnTV, gp100/Pmel17, GPNMB, HAUS3, Hepsin, HER-2/neu, HERV-K-MEL, HLA-A11, HLA-A2, HLA-DOB, hsp70-2, IDO1, IGF2B3, IL13Ralpha2, Intestinal carboxyl esterase, K-ras, Kallikrein 4, KIF20A, KK-LC-1, KKLC1, KM-HN-1, KMHN1 also known as CCDC110, LAGE-1, LDLR-fucosyltransferaseAS fusion protein, Lengsin, M-CSF, MAGE-A1, MAGE-A10, MAGE-A12, MAGE-A2, MAGE-A3, MAGE-A4, MAGE-A6, MAGE-A9, MAGE-C1, MAGE-C2, malic enzyme, mammaglobin-A, MART2, MATN, MC1R, MCSP, mdm-2, ME1, Melan-A/MART-1, Meloe, Midkine, MMP-2, MMP-7, MUC1, MUC5AC, mucin, MUM-1, MUM-2, MUM-3, Myosin, Myosin class I, N-raw, NA88-A, neo-PAP, NFYC, NY-BR-1, NY-ESO-1/LAGE-2, OA1, OGT, OS-9, P polypeptide, p53, PAP, PAX5, PBF, pml-RARalpha fusion protein, polymorphic epithelial mucin ("PEM"), PPP1R3B, PRAME, PRDX5, PSA, PSMA, PTPRK, RAB38/NY-MEL-1, RAGE-1, RBAF600, RGS5, RhoC, RNF43, RU2AS, SAGE, secernin 1, SIRT2, SNRPD1, SOX10, Sp17, SPA17, SSX-2, SSX-4, STEAP1, survivin, SYT-SSX1 or -SSX2 fusion protein, TAG-1, TAG-2, Telomerase, TGF-betaRII, TPBG, TRAG-3, Triosephosphate isomerase, TRP-1/gp75, TRP-2, TRP2-INT2, tyrosinase, tyrosinase ("TYR"), VEGF, WT1 and XAGE-1b/GAGED2a.

60. The method of claim 58, wherein the cancer associated antigen is a neo-antigen.

61. The method any one of claims 25 to 60, wherein the cancer immunotherapy comprises administration of a cancer-specific T cell to the subject.

62. The method of claim 61, wherein the T cell is a $CD4^+$ T cell.

63. The method of claim 62, wherein the $CD4^+$ T cell is a T_H1 T cell, a T_H2 T cell or a T_H17 T cell.

64. The method of any one of claims 61 to 63, wherein the T cell expresses a T cell receptor specific for a cancer-associated antigen.

65. The method of claim 64, wherein the cancer-associated antigen is selected from the group consisting of adipophilin, AIM-2, ALDH1A1, alpha-actinin-4, alpha-fetoprotein ("AFP"), ARTC1, B-RAF, BAGE-1, BCLX (L), BCR-ABL fusion protein b3a2, beta-catenin, BING-4, CA-125, CALCA, carcinoembryonic antigen ("CEA"), CASP-5, CASP-8, CD274, CD45, Cdc27, CDK12, CDK4, CDKN2A, CEA, CLPP, COA-1, CPSF, CSNK1A1, CTAG1, CTAG2, cyclin D1, Cyclin-A1, dek-can fusion protein, DKK1, EFTUD2, Elongation factor 2, ENAH (hMena), Ep-CAM, EpCAM, EphA3, epithelial tumor antigen ("ETA"), ETV6-AML1 fusion protein, EZH2, FGF5, FLT3-ITD, FN1, G250/MN/CAIX, GAGE-1,2,8, GAGE-3,4,5,6,7, GAS7, glypican-3, GnTV, gp100/Pmel17, GPNMB, HAUS3, Hepsin, HER-2/neu, HERV-K-MEL, HLA-A11, HLA-A2, HLA-DOB, hsp70-2, IDO1, IGF2B3, IL13Ralpha2, Intestinal carboxyl esterase, K-ras, Kallikrein 4, KIF20A, KK-LC-1, KKLC1, KM-HN-1, KMHN1 also known as CCDC110, LAGE-1, LDLR-fucosyltransferaseAS fusion protein, Lengsin, M-CSF, MAGE-A1, MAGE-A10, MAGE-A12, MAGE-A2, MAGE-A3, MAGE-A4, MAGE-A6, MAGE-A9, MAGE-C1, MAGE-C2, malic enzyme, mammaglobin-A, MART2, MATN, MC1R, MCSP, mdm-2, ME1, Melan-A/MART-1, Meloe, Midkine, MMP-2, MMP-7, MUC1, MUC5AC, mucin, MUM-1, MUM-2, MUM-3, Myosin, Myosin class I, N-raw, NA88-A, neo-PAP, NFYC, NY-BR-1, NY-ESO-1/LAGE-2, OA1, OGT, OS-9, P polypeptide, p53, PAP, PAX5, PBF, pml-RARalpha fusion protein, polymorphic epithelial mucin ("PEM"), PPP1R3B, PRAME, PRDX5, PSA, PSMA, PTPRK, RAB38/NY-MEL-1, RAGE-1, RBAF600, RGS5, RhoC, RNF43, RU2AS, SAGE, secernin 1, SIRT2, SNRPD1, SOX10, Sp17, SPA17, SSX-2, SSX-4, STEAP1, survivin, SYT-SSX1 or -SSX2 fusion protein, TAG-1, TAG-2, Telomerase, TGF-betaRII, TPBG, TRAG-3, Triosephosphate isomerase, TRP-1/gp75, TRP-2, TRP2-INT2, tyrosinase, tyrosinase ("TYR"), VEGF, WT1 and XAGE-1b/GAGED2a.

66. The method of any one of claims 25 to 65, wherein the cancer immunotherapy comprises administration of an immune modulatory protein to the subject.

67. The method of claim 66, wherein the immune modulatory protein is a cytokine.

68. The method of claim 67, wherein the immune modulatory protein is selected from the group consisting of B lymphocyte chemoattractant ("BLC"), C-C motif chemokine 11 ("Eotaxin-1"), Eosinophil chemotactic protein 2 ("Eotaxin-2"), Granulocyte colony-stimulating factor ("G-CSF"), Granulocyte macrophage colony-stimulating factor ("GM-CSF"), 1-309, Intercellular

Adhesion Molecule 1 ("ICAM-1"), Interferon gamma ("IFN-gamma"), Interlukin-1 alpha ("IL-1 alpha"), Interlukin-1 beta ("IL-1 beta"), Interleukin 1 receptor antagonist ("IL-1 ra"), Interleukin-2 ("IL-2"), Interleukin-4 ("IL-4"), Interleukin-5 ("IL-5"), Interleukin-6 ("IL-6"), Interleukin-6 soluble receptor ("IL-6 sR"), Interleukin-7 ("IL-7"), Interleukin-8 ("IL-8"), Interleukin- 10 ("IL-10"), Interleukin- 11 ("IL-11"), Subunit beta of Interleukin- 12 ("IL-12 p40" or "IL-12 p70"), Interleukin-13 ("IL-13"), Interleukin-15 ("IL-15"), Interleukin-16 ("IL-16"), Interleukin-17 ("IL-17"), Chemokine (C-C motif) Lignad 2 ("MCP-1"), Macrophage colony-stimulating factor ("M-CSF"), Monokine induced by gamma interferon ("MIG"), Chemokine (C-C motif) ligand 2 ("MIP-1 alpha"), Chemokine (C-C motif) ligand 4 ("MIP-1 beta"), Macrophage inflammatory protein- 1 -delta ("MIP-1 delta"), Platelet-derived growth factor subunit B ("PDGF-BB"), Chemokine (C-C motif) ligand 5, Regulated on Activation, Normal T cell Expressed and Secreted ("RANTES"), TIMP metallopeptidase inhibitor 1 ("TIMP-1"), TIMP metallopeptidase inhibitor 2 ("TIMP-2"), Tumor necrosis factor, lymphotoxin-alpha ("TNF alpha"), Tumor necrosis factor, lymphotoxin-beta ("TNF beta"), Soluble TNF receptor type 1 ("sTNFRI"), sTNFRIIAR, Brain-derived neurotrophic factor ("BDNF"), Basic fibroblast growth factor ("bFGF"), Bone morphogenetic protein 4 ("BMP-4"), Bone morphogenetic protein 5 ("BMP-5"), Bone morphogenetic protein 7 ("BMP-7"), Nerve growth factor ("b-NGF"), Epidermal growth factor ("EGF"), Epidermal growth factor receptor ("EGFR"), Endocrine-gland-derived vascular endothelial growth factor ("EG-VEGF"), Fibroblast growth factor 4 ("FGF-4"), Keratinocyte growth factor ("FGF-7"), Growth differentiation factor 15 ("GDF-15"), Glial cell-derived neurotrophic factor ("GDNF"), Growth Hormone, Heparin-binding EGF-like growth factor ("HB-EGF"), Hepatocyte growth factor ("HGF"), Insulin-like growth factor binding protein 1 ("IGFBP-1"), Insulin-like growth factor binding protein 2 ("IGFBP-2"), Insulin-like growth factor binding protein 3 (" IGFBP-3"), Insulin-like growth factor binding protein 4 ("IGFBP-4"), Insulin-like growth factor binding protein 6 ("IGFBP-6"), Insulin-like growth factor 1 ("IGF-1"), Insulin, Macrophage colony-stimulating factor ("M-CSF R"), Nerve growth factor receptor ("NGF R"), Neurotrophin-3 ("NT-3"), Neurotrophin-4 ("NT-4"), Osteoclastogenesis inhibitory factor ("Osteoprotegerin"), Platelet-derived growth factor receptors ("PDGF-AA"), Phosphatidylinositol-glycan biosynthesis ("PIGF"), Skp, Cullin, F-box containing complex ("SCF"), Stem cell factor receptor ("SCF R"), Transforming growth factor alpha ("TGFalpha"), Transforming growth factor beta-1 ("TGF beta 1"), Transforming growth factor beta-3 ("TGF

beta 3"), Vascular endothelial growth factor ("VEGF"), Vascular endothelial growth factor receptor 2 ("VEGFR2"), Vascular endothelial growth factor receptor 3 ("VEGFR3"), VEGF-D 6Ckine, Tyrosine-protein kinase receptor UFO ("Axl"), Betacellulin ("BTC"), Mucosae-associated epithelial chemokine ("CCL28"), Chemokine (C-C motif) ligand 27 ("CTACK"), Chemokine (C-X-C motif) ligand 16 ("CXCL16"), C-X-C motif chemokine 5 ("ENA-78"), Chemokine (C-C motif) ligand 26 ("Eotaxin-3"), Granulocyte chemotactic protein 2 ("GCP-2"), GRO, Chemokine (C-C motif) ligand 14 ("HCC-l"), Chemokine (C-C motif) ligand 16 ("HCC-4"), Interleukin-9 ("IL-9"), Interleukin-17 F ("IL-17F"), Interleukin- 18-binding protein ("IL-18 BPa"), Interleukin-28 A ("IL-28A"), Interleukin 29 ("IL-29"), Interleukin 31 ("IL-31"), C-X-C motif chemokine 10 ("IP-10"), Chemokine receptor CXCR3 ("I-TAC"), Leukemia inhibitory factor ("LIF"), Light, Chemokine (C motif) ligand ("Lymphotactin"), Monocyte chemoattractant protein 2 ("MCP-2"), Monocyte chemoattractant protein 3 ("MCP-3"), Monocyte chemoattractant protein 4 ("MCP-4"), Macrophage-derived chemokine ("MDC"), Macrophage migration inhibitory factor ("MIF"), Chemokine (C-C motif) ligand 20 ("MIP-3 alpha"), C-C motif chemokine 19 ("MIP-3 beta"), Chemokine (C-C motif) ligand 23 ("MPIF-1"), Macrophage stimulating protein alpha chain ("MSPalpha"), Nucleosome assembly protein 1-like 4 ("NAP-2"), Secreted phosphoprotein 1 ("Osteopontin"), Pulmonary and activation-regulated cytokine ("PARC"), Platelet factor 4 ("PF4"), Stroma cell-derived factor- 1 alpha ("SDF-1 alpha"), Chemokine (C-C motif) ligand 17 ("TARC"), Thymus-expressed chemokine ("TECK"), Thymic stromal lymphopoietin ("TSLP 4- IBB"), CD 166 antigen ("ALCAM"), Cluster of Differentiation 80 ("B7-1"), Tumor necrosis factor receptor superfamily member 17 ("BCMA"), Cluster of Differentiation 14 ("CD14"), Cluster of Differentiation 30 ("CD30"), Cluster of Differentiation 40 ("CD40 Ligand"), Carcinoembryonic antigen-related cell adhesion molecule 1 (biliary glycoprotein) ("CEACAM-1"), Death Receptor 6 ("DR6"), Deoxythymidine kinase ("Dtk"), Type 1 membrane glycoprotein ("Endoglin"), Receptor tyrosine-protein kinase erbB-3 ("ErbB3"), Endothelial-leukocyte adhesion molecule 1 ("E-Selectin"), Apoptosis antigen 1 ("Fas"), Fms-like tyrosine kinase 3 ("Flt-3L"), Tumor necrosis factor receptor superfamily member 1 ("GITR"), Tumor necrosis factor receptor superfamily member 14 ("HVEM"), Intercellular adhesion molecule 3 ("ICAM-3"), IL-1 R4, IL-1 RI, IL-10 Rbeta, IL-17R, IL-2Rgamma, IL-21R, Lysosome membrane protein 2 ("LIMPII"), Neutrophil gelatinase-associated lipocalin ("Lipocalin-2"), CD62L ("L-Selectin"), Lymphatic endothelium ("LYVE-1"), MHC

class I polypeptide-related sequence A ("MICA"), MHC class I polypeptide-related sequence B ("MICB"), NRGl-betal, Beta-type platelet-derived growth factor receptor ("PDGF Rbeta"), Platelet endothelial cell adhesion molecule ("PECAM-1"), RAGE, Hepatitis A virus cellular receptor 1 ("TIM-1"), Tumor necrosis factor receptor superfamily member IOC ("TRAIL R3"), Trappin protein transglutaminase binding domain ("Trappin-2"), Urokinase receptor ("uPAR"), Vascular cell adhesion protein 1 ("VCAM-1"), XEDARActivin A, Agouti-related protein ("AgRP"), Ribonuclease 5 ("Angiogenin"), Angiopoietin 1, Angiostatin, Catheprin S, CD40, Cryptic family protein IB ("Cripto-1"), DAN, Dickkopf-related protein 1 ("DKK-1"), E-Cadherin, Epithelial cell adhesion molecule ("EpCAM"), Fas Ligand (FasL or CD95L), Fcg RIIB/C, FoUistatin, Galectin-7, Intercellular adhesion molecule 2 ("ICAM-2"), IL-13 Rl, IL-13R2, IL-17B, IL-2 Ra, IL-2 Rb, IL-23, LAP, Neuronal cell adhesion molecule ("NrCAM"), Plasminogen activator inhibitor- 1 ("PAI-1"), Platelet derived growth factor receptors ("PDGF-AB"), Resistin, stromal cell-derived factor 1 ("SDF-1 beta"), sgpl30, Secreted frizzled-related protein 2 ("ShhN"), Sialic acid-binding immunoglobulin-type lectins ("Siglec-5"), ST2, Transforming growth factor-beta 2 ("TGF beta 2"), Tie-2, Thrombopoietin ("TPO"), Tumor necrosis factor receptor superfamily member 10D ("TRAIL R4"), Triggering receptor expressed on myeloid cells 1 ("TREM-1"), Vascular endothelial growth factor C ("VEGF-C"), VEGFRlAdiponectin, Adipsin ("AND"), Alpha-fetoprotein ("AFP"), Angiopoietin-like 4 ("ANGPTL4"), Beta-2-microglobulin ("B2M"), Basal cell adhesion molecule ("BCAM"), Carbohydrate antigen 125 ("CA125"), Cancer Antigen 15-3 ("CA15-3"), Carcinoembryonic antigen ("CEA"), cAMP receptor protein ("CRP"), Human Epidermal Growth Factor Receptor 2 ("ErbB2"), Follistatin, Follicle-stimulating hormone ("FSH"), Chemokine (C-X-C motif) ligand 1 ("GRO alpha"), human chorionic gonadotropin ("beta HCG"), Insulin-like growth factor 1 receptor ("IGF-1 sR"), IL-1 sRII, IL-3, IL-18 Rb, IL-21, Leptin, Matrix metalloproteinase-1 ("MMP-1"), Matrix metalloproteinase-2 ("MMP-2"), Matrix metalloproteinase-3 ("MMP-3"), Matrix metalloproteinase-8 ("MMP-8"), Matrix metalloproteinase-9 ("MMP-9"), Matrix metalloproteinase-10 ("MMP-10"), Matrix metalloproteinase-13 ("MMP-13"), Neural Cell Adhesion Molecule ("NCAM-1"), Entactin ("Nidogen-1"), Neuron specific enolase ("NSE"), Oncostatin M ("OSM"), Procalcitonin, Prolactin, Prostate specific antigen ("PSA"), Sialic acid-binding Ig-like lectin 9 ("Siglec-9"), ADAM 17 endopeptidase ("TACE"), Thyroglobulin, Metalloproteinase inhibitor 4 ("TIMP-4"), TSH2B4, Disintegrin and metalloproteinase domain-

containing protein 9 ("ADAM-9"), Angiopoietin 2, Tumor necrosis factor ligand superfamily member 13/ Acidic leucine-rich nuclear phosphoprotein 32 family member B ("APRIL"), Bone morphogenetic protein 2 ("BMP-2"), Bone morphogenetic protein 9 ("BMP-9"), Complement component 5a ("C5a"), Cathepsin L, CD200, CD97, Chemerin, Tumor necrosis factor receptor superfamily member 6B ("DcR3"), Fatty acid-binding protein 2 ("FABP2"), Fibroblast activation protein, alpha ("FAP"), Fibroblast growth factor 19 ("FGF-19"), Galectin-3, Hepatocyte growth factor receptor ("HGF R"), IFN-gammalpha/beta R2, Insulin-like growth factor 2 ("IGF-2"), Insulin-like growth factor 2 receptor ("IGF-2 R"), Interleukin-1 receptor 6 ("IL-1R6"), Interleukin 24 ("IL-24"), Interleukin 33 ("IL-33", Kallikrein 14, Asparaginyl endopeptidase ("Legumain"), Oxidized low-density lipoprotein receptor 1 ("LOX-1"), Mannose-binding lectin ("MBL"), Neprilysin ("NEP"), Notch homolog 1, translocation-associated (Drosophila) ("Notch-1"), Nephroblastoma overexpressed ("NOV"), Osteoactivin, Programmed cell death protein 1 ("PD-1"), N-acetylmuramoyl-L-alanine amidase ("PGRP-5"), Serpin A4, Secreted frizzled related protein 3 ("sFRP-3"), Thrombomodulin, Tolllike receptor 2 ("TLR2"), Tumor necrosis factor receptor superfamily member 10A ("TRAIL Rl"), Transferrin ("TRF"), WIF-lACE-2, Albumin, AMICA, Angiopoietin 4, B-cell activating factor ("BAFF"), Carbohydrate antigen 19-9 ("CA19-9"), CD 163 , Clusterin, CRT AM, Chemokine (C-X-C motif) ligand 14 ("CXCL14"), Cystatin C, Decorin ("DCN"), Dickkopf-related protein 3 ("Dkk-3"), Delta-like protein 1 ("DLL1"), Fetuin A, Heparin-binding growth factor 1 ("aFGF"), Folate receptor alpha ("FOLR1"), Furin, GPCR-associated sorting protein 1 ("GASP-1"), GPCR-associated sorting protein 2 ("GASP-2"), Granulocyte colony-stimulating factor receptor ("GCSF R"), Serine protease hepsin ("HAI-2"), Interleukin-17B Receptor ("IL-17B R"), Interleukin 27 ("IL-27"), Lymphocyte-activation gene 3 ("LAG-3"), Apolipoprotein A-V ("LDL R"), Pepsinogen I, Retinol binding protein 4 ("RBP4"), SOST, Heparan sulfate proteoglycan ("Syndecan-1"), Tumor necrosis factor receptor superfamily member 13B ("TACI"), Tissue factor pathway inhibitor ("TFPI"), TSP-1, Tumor necrosis factor receptor superfamily, member 10b ("TRAIL R2"), TRANCE, Troponin I, Urokinase Plasminogen Activator ("uPA"), Cadherin 5, type 2 or VE-cadherin (vascular endothelial) also known as CD144 ("VE-Cadherin"), WNTl-inducible-signaling pathway protein 1 ("WISP-1"), and Receptor Activator of Nuclear Factor κ B ("RANK").

69. The method of any one of claims 25 to 68, wherein the cancer immunotherapy comprises administering an adjuvant to the subject.

70. The method of claim 69, wherein the adjuvant is selected from the group consisting of an immune modulatory protein, Adjuvant 65, α-GalCer, aluminum phosphate, aluminum hydroxide, calcium phosphate, β-Glucan Peptide, CpG DNA, GPI-0100, lipid A, lipopolysaccharide, Lipovant, Montanide, N-acetyl-muramyl-L-alanyl-D-isoglutamine, Pam3CSK4, quil A and trehalose dimycolate.

71. The method of any one of claims 22 to 70, wherein second the cancer therapy comprises administering an angiogenesis inhibitor to the subject.

72. The method of claim 71, wherein the angiogenesis inhibitor is selected from the group consisting of Bevacizumab (Avastin®), Ziv-aflibercept (Zaltrap®), Sorafenib (Nexavar®), Sunitinib (Sutent®), Pazopanib (Votrient®), Regorafenib (Stivarga®), and Cabozantinib (Cometriq™).

73. The method of any one of claims 22 to 72, wherein the second cancer therapy comprises radiation therapy.

74. The method of any one of claims 22 to 73, wherein the cancer therapy comprises administering an antibiotic to the subject.

75. The method of claim 74, wherein the antibiotic is selected from the group consisting of aminoglycosides, ansamycins, carbacephems, carbapenems, cephalosporins, glycopeptides, lincosamides, lipopeptides, macrolides, monobactams, nitrofurans, oxazolidonones, penicillins, polypeptide antibiotics, quinolones, fluoroquinolone, sulfonamides, tetracyclines, anti-mycobacterial compounds and combinations thereof.

76. The method of any one of claims 22 to 75, wherein the cancer therapy comprises administering to the subject a second therapeutic bacteria.

77. The method of any one of claims 1 to 76, wherein the method further comprises administering a prebiotic to the subject.

78. The method of claim 77, wherein the prebiotic is a fructooligosaccharide, a galactooligosaccharide, a trans-galactooligosaccharide, a xylooligosaccharide, a

chitooligosaccharide, a soy oligosaccharides, a gentiooligosaccharide, an isomaltooligosaccharide, a mannooligosaccharide, a maltooligosaccharide, a mannanoligosaccharide, lactulose, lactosucrose, palatinose, glycosyl sucrose, guar gum, gum Arabic, tagalose, amylose, amylopectin, pectin, xylan, or a cyclodextrin.

79. A method of augmenting a tumor environment containing an immune cell in a subject comprising administering a bacterial composition comprising *Bifidobacterium animalis ssp. lactis* to the subject.

80. The method of claim 79, wherein the immune cell is an antigen presenting cell, a dendritic cell, a T cell, a B cell, an NK cell, an NKT cell, a macrophage, or a neutrophil.

81. The method of claim 79 or claim 80, wherein the composition is administered orally.

82. A method of augmenting a tumor environment that comprises a biomarker of patient selection in a subject comprising administering a bacterial composition comprising *Bifidobacterium animalis ssp. lactis* to the subject.

83. The method of claim 82, wherein the biomarker is PD-L1 and the therapy is a checkpoint inhibitor.

84. A method of augmenting a tumor environment comprising a biomarker associated with immune cell activity in a subject comprising administering a bacterial composition comprising *Bifidobacterium animalis ssp. lactis* to the subject.

85. The method of claim 84, wherein the biomarker is CD86, CD80 or CD206.

86. The method of claim 84 or claim 85, wherein the composition is administered orally.

87. The method of any one of claims 1 to 86, wherein the subject is a human.

88. The method of any one of claims 1 to 86, wherein the subject is a non-human mammal.

89. The method of claim 88, wherein the mammal is selected from the group consisting of a dog, a cat, a cow, a horse, a pig, a donkey, a goat, a camel, a mouse, a rat, a guinea pig, a sheep, a llama, a monkey, a gorilla or a chimpanzee.

90. The method of any one of claims 1 to 89, wherein a second bacterium is administered as part of an ecological consortium.

91. A bacterial composition comprising *Bifidobacterium animalis ssp. Lactis.*

92. The bacterial composition of claim 91, further comprising a pharmaceutically acceptable carrier.

93. A bacterial composition comprising a pharmaceutically active biomass (PhABs) derived from *Bifidobacterium animalis ssp. lactis*.

94. A bacterial composition comprising a pharmaceutically active biomass (PhABs) isolated from *Bifidobacterium animalis ssp. lactis*.

95. The bacterial composition of any one of claims 91 to 94, wherein the *Bifidobacterium animalis ssp. lactis* is a strain comprising at least 99% sequence identity to the nucleotide sequence of the *Bifidobacterium animalis ssp. lactis* Strain A (ATCC Deposit Number PTA-125097).

96. The bacterial composition of any one of claims 91 to 94, wherein the *Bifidobacterium animalis ssp. lactis* is a strain comprising at least 99.9% sequence identity to the nucleotide sequence of the *Bifidobacterium animalis ssp. lactis* Strain A (ATCC Deposit Number PTA-125097).

97. The bacterial composition of any one of claims 91 to 94, wherein the *Bifidobacterium animalis ssp. lactis* is the *Bifidobacterium animalis ssp. lactis* Strain A (ATCC Deposit Number PTA-125097).

98. The bacterial composition of any one of claims 91 to 94, wherein the composition comprises an effective amount of *Bifidobacterium animalis ssp. lactis* Strain A (ATCC Deposit Number PTA-125097).

99. The bacterial composition of any one of claims 91 to 98, wherein the bacterial composition formulated for oral, rectal, intravenous, intratumoral, or subcutaneous administration.

100. The bacterial composition of any one of claims 91 to 99, wherein at least 50% of the bacteria in the bacterial composition are *Bifidobacterium animalis ssp. lactis*.

101. The bacterial composition of any one of claims 91 to 99, wherein at least 90% of the bacteria in the bacterial composition are *Bifidobacterium animalis ssp. lactis*.

102. The bacterial composition of any one of claims 91 to 101, wherein substantially all of the bacteria in the bacterial composition are *Bifidobacterium animalis ssp. lactis*.

103. The bacterial composition of any one of claims 91 to 103, wherein the bacterial composition comprises at least 1 x 10^6 colony forming units (CFUs) of *Bifidobacterium animalis ssp. lactis*.

104. The bacterial composition of claim 103, wherein the bacterial composition comprises at least 1 x 10^7 CFUs of *Bifidobacterium animalis ssp. lactis*.

105. The bacterial composition of claim 103, wherein the bacterial composition comprises at least 1 x 10^8 CFUs of *Bifidobacterium animalis ssp. lactis*.

106. The bacterial composition of any one of claims 91 to 105, wherein the bacterial composition comprises live bacteria.

107. The bacterial composition of any one of claims 91 to 105, wherein the bacterial composition comprises attenuated bacteria.

108. The bacterial composition of any one of claims 91 to 105, wherein the bacterial composition comprises killed bacteria.

109. The bacterial composition of any one of claims 91 to 108, wherein administration of the bacterial composition treats the cancer.

110. The bacterial composition of any one of claims 91 to 109, wherein administration of the bacterial composition induces an anti-cancer immune response.

111. The bacterial composition of any one of claims 91 to 109, wherein administration of the bacterial composition induces CD3+ immune cell infiltration.

112. The bacterial composition of any one of claims 91 to 109, wherein administration of the bacterial composition induces MHC Class I upregulation at the tumor site.

113. The bacterial composition of any one of clams 91 to 112, wherein the *Bifidobacterium animalis ssp. lactis* is formulated with an enteric coating or micro encapsulation.

114. A bioreactor comprising *Bifidobacterium animalis ssp. lactis*.

115. The bioreactor of claim 114, wherein the *Bifidobacterium animalis ssp. lactis* is a strain comprising at least 99% sequence identity to the nucleotide sequence of the *Bifidobacterium animalis ssp. lactis* Strain A (ATCC Deposit Number PTA-125097).

116. The bioreactor of claim 114, wherein the *Bifidobacterium animalis ssp. lactis* is a strain comprising at least 99.9% sequence identity to the nucleotide sequence of the *Bifidobacterium animalis ssp. lactis* Strain A (ATCC Deposit Number PTA-125097).

117. The bioreactor of claim 114,wherein the *Bifidobacterium animalis ssp. lactis* is the *Bifidobacterium animalis ssp. lactis* Strain A (ATCC Deposit Number PTA-125097).

118. A bacterial composition comprising at least one strain of *Bifidobacterium animalis ssp. lactis* having a gene comprising a nucleotide base change as identified in Table 6 .

119. The bacterial composition of claim 118, comprising at least one strain of *Bifidobacterium animalis ssp. lactis* having two genes comprising a nucleotide base change as identified in Table 6.

120. The bacterial composition of claim 118, comprising at least one strain of *Bifidobacterium animalis ssp. lactis* having three genes comprising a nucleotide base change as identified in Table 6.

121. The bacterial composition of claim 118, comprising at least one strain of *Bifidobacterium animalis ssp. lactis* having four genes comprising a nucleotide base change as identified in Table 6.

122. The bacterial composition of claim 118, comprising at least one strain of *Bifidobacterium animalis ssp. lactis* having five genes comprising a nucleotide base change as identified in Table 6.

123. The bacterial composition of claim 118, comprising at least one strain of *Bifidobacterium animalis ssp. lactis* having six genes comprising a nucleotide base change as identified in Table 6.

124. The bacterial composition of claim 118, comprising at least one strain of *Bifidobacterium animalis ssp. lactis* having seven genes comprising a nucleotide base change as identified in Table 6.

125. The bacterial composition of claim 118, comprising at least one strain of *Bifidobacterium animalis ssp. lactis* having eight genes comprising a nucleotide base change as identified in Table 6.

126. The bacterial composition of claim 118, comprising at least one strain of *Bifidobacterium animalis ssp. lactis* having nine genes comprising a nucleotide base change as identified in Table 6.

127. The bacterial composition of claim 118, comprising at least one strain of *Bifidobacterium animalis ssp. lactis* having ten genes comprising a nucleotide base change as identified in Table 6.

128. The bacterial composition of claim 118, comprising at least one strain of *Bifidobacterium animalis ssp. lactis* having eleven genes comprising a nucleotide base change as identified in Table 6.

129. The bacterial composition of claim 118, comprising at least one strain of *Bifidobacterium animalis ssp. lactis* having twelve genes comprising a nucleotide base change as identified in Table 6.

130. The bacterial composition of claim 118, comprising at least one strain of *Bifidobacterium animalis ssp. lactis* having thirteen genes comprising a nucleotide base change as identified in Table 6.

131. The bacterial composition of claim 118, comprising at least one strain of *Bifidobacterium animalis ssp. lactis* having fourteen genes comprising a nucleotide base change as identified in Table 6.

132. The bacterial composition of claim 118, comprising at least one strain of *Bifidobacterium animalis ssp. lactis* having fifteen genes comprising a nucleotide base change as identified in Table 6.

133. The bacterial composition of claim 118, comprising at least one strain of *Bifidobacterium animalis ssp. lactis* having genes comprising all the nucleotide base changes as identified in Table 6.

Figure 1

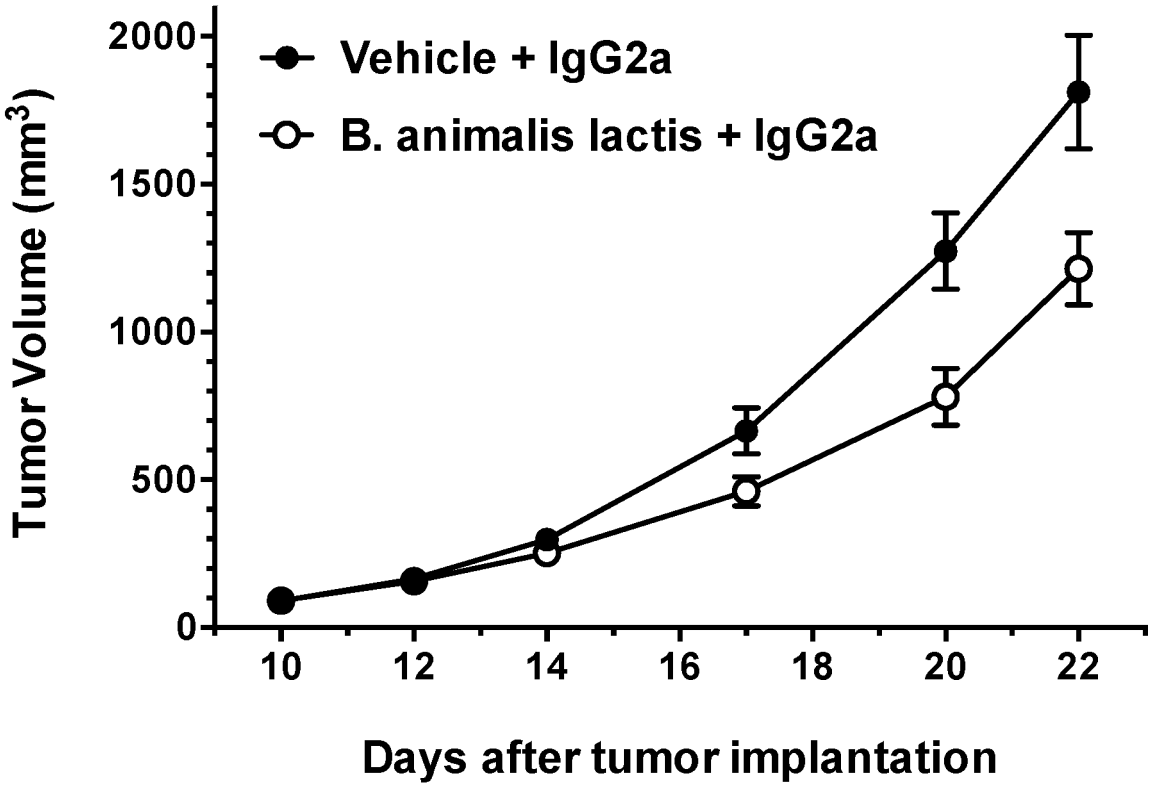

Vehicle + IgG2a
B. animalis lactis + IgG2a
Tumor Volume (mm3)
2000
1500
1000
500
0
10
12
14
16
18
20
22
Days after tumor implantation

Figure 2

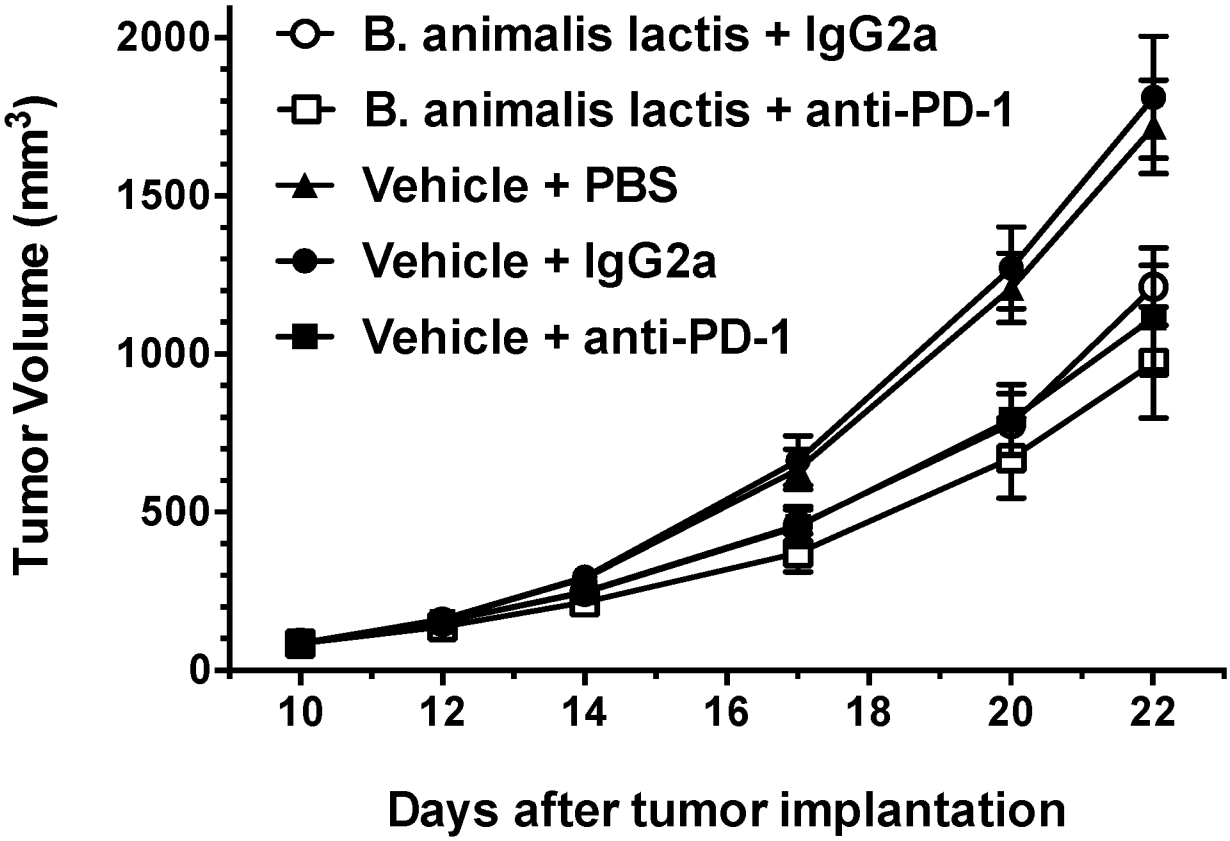

B. animalis lactis + IgG2a
B. animalis lactis + anti-PD-1
Vehicle + PBS
Vehicle + IgG2a
Vehicle + anti-PD-1
Tumor Volume (mm3)
2000
1500
1000
500
0
10
12
14
16
18
20
22
Days after tumor implantation

Figure 3

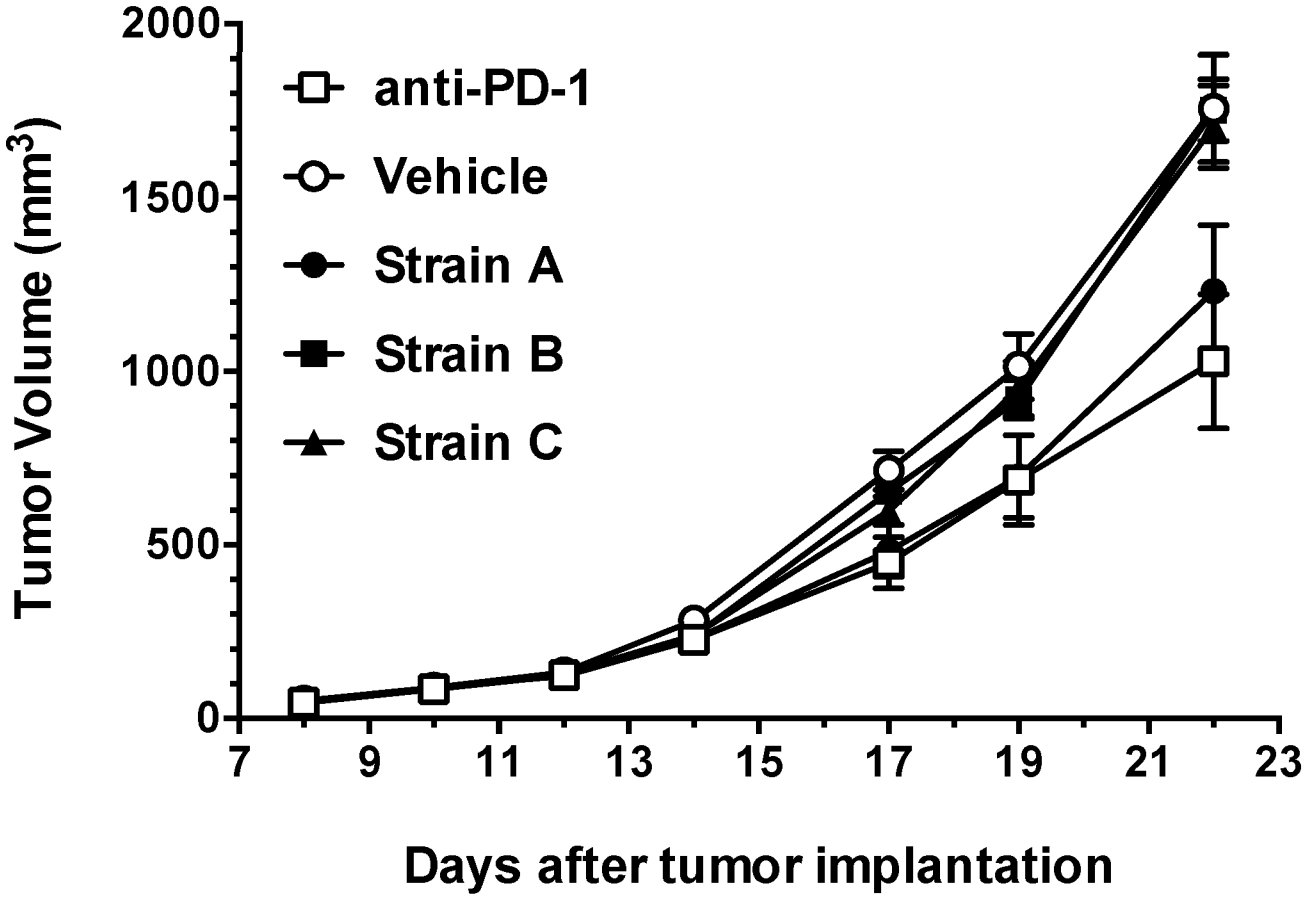

2000
1500
1000
500
0
Tumor Volume (mm3)
anti-PD-1
Vehicle
Strain A
Strain B
Strain C
7
9
11
13
15
17
19
21
23
Days after tumor implantation

Figure 4

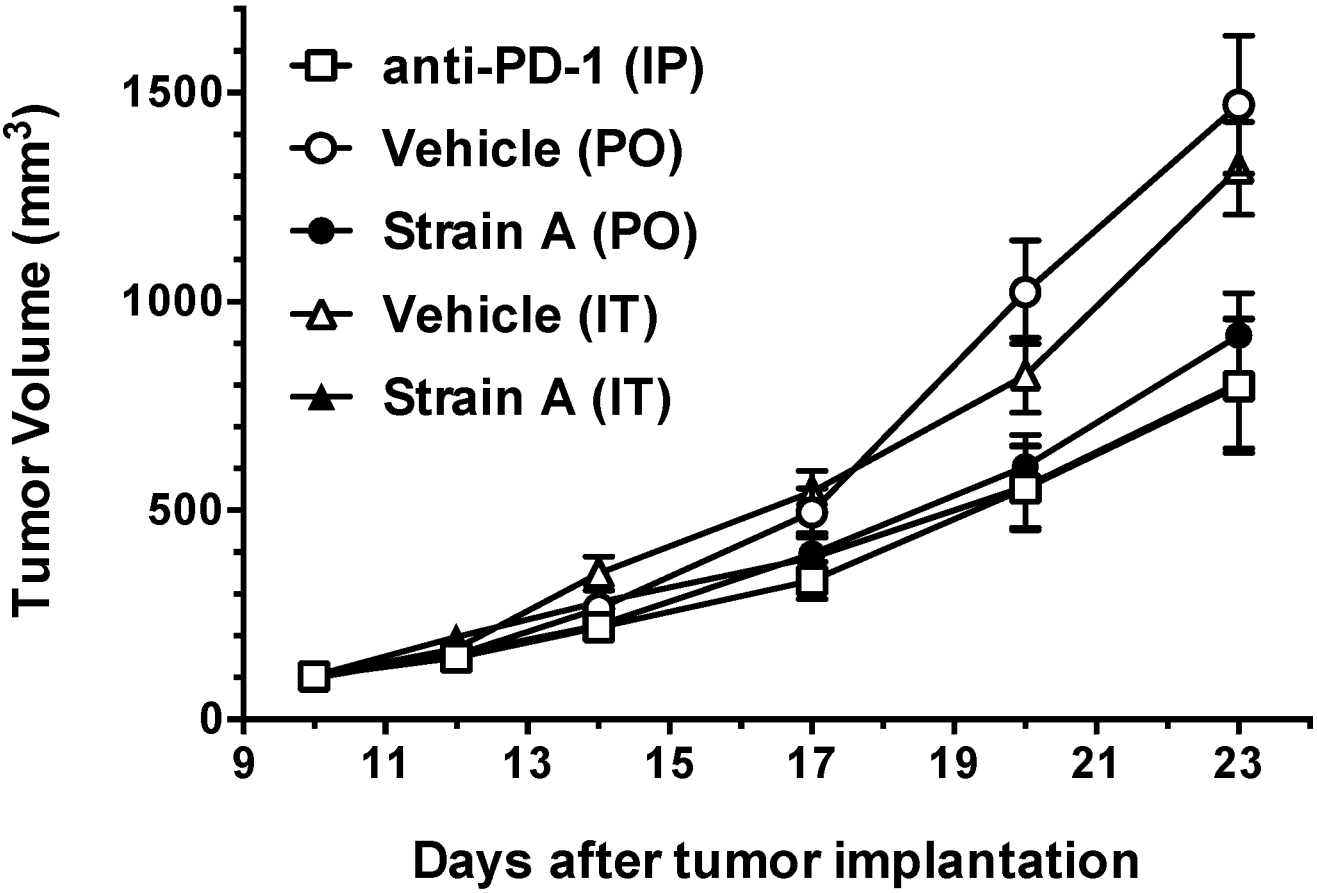

anti-PD-1 (IP)
Vehicle (PO)
Strain A (PO)
Vehicle (IT)
Strain A (IT)
Tumor Volume (mm3)
1500
1000
500
0
9
11
13
15
17
19
21
23
Days after tumor implantation

Figure 5

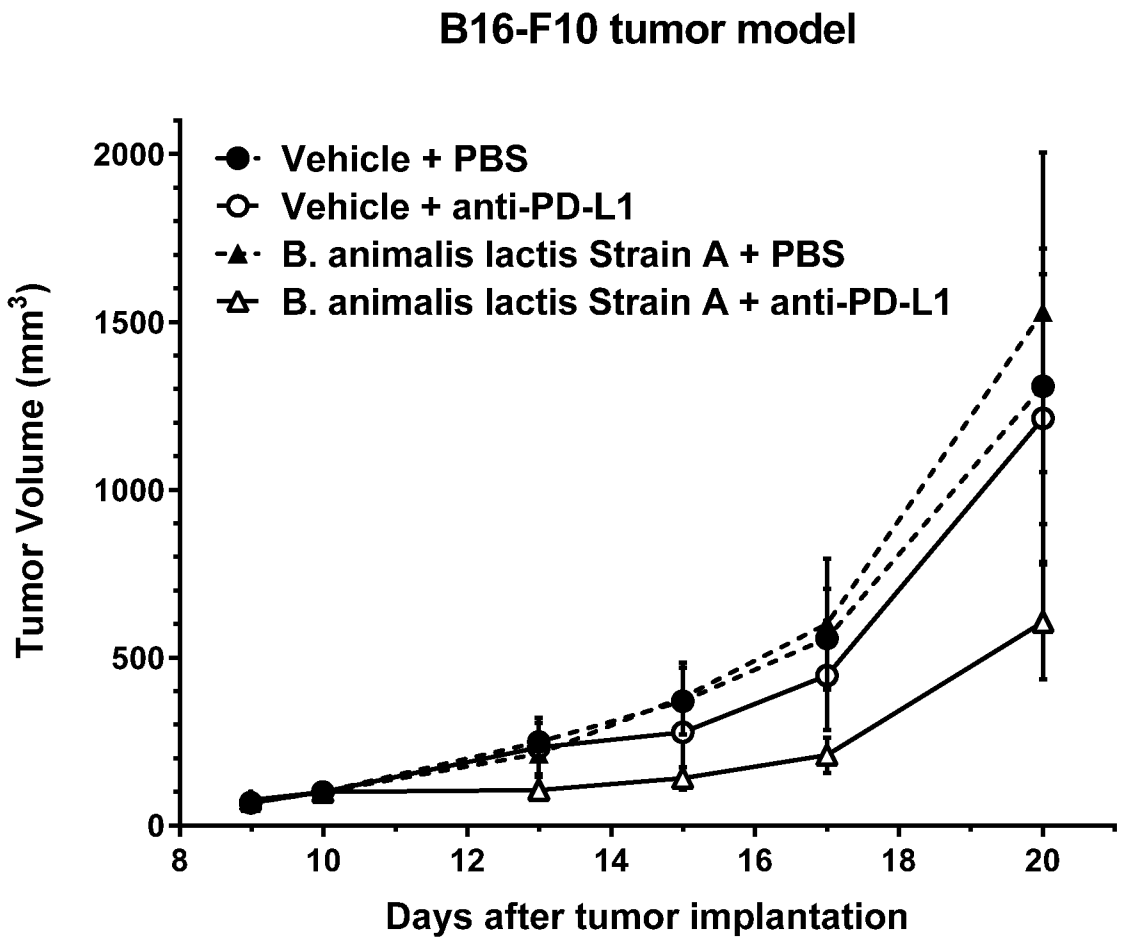

B16-F10 tumor model
Vehicle + PBS
Vehicle + anti-PD-L1
B. animalis lactis Strain A + PBS
B. animalis lactis Strain A + anti-PD-L1
Tumor Volume (mm3)
2000
1500
1000
500
0
8
10
12
14
16
18
20
Days after tumor implantation

Figure 6A

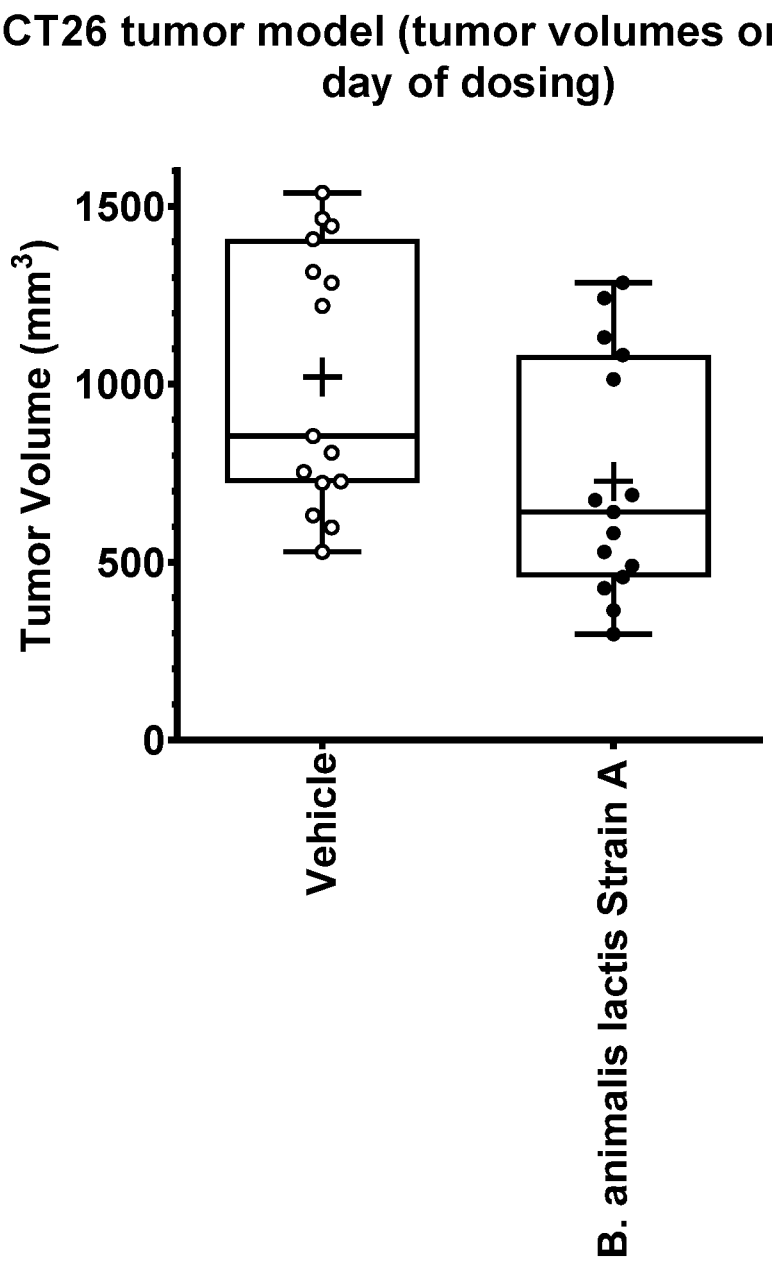

CT26 tumor model (tumor volumes on the 9th day of dosing)
Tumor Volume (mm3)
1500
1000
500
0
Vehicle
B. animalis lactis Strain A

Figure 6B

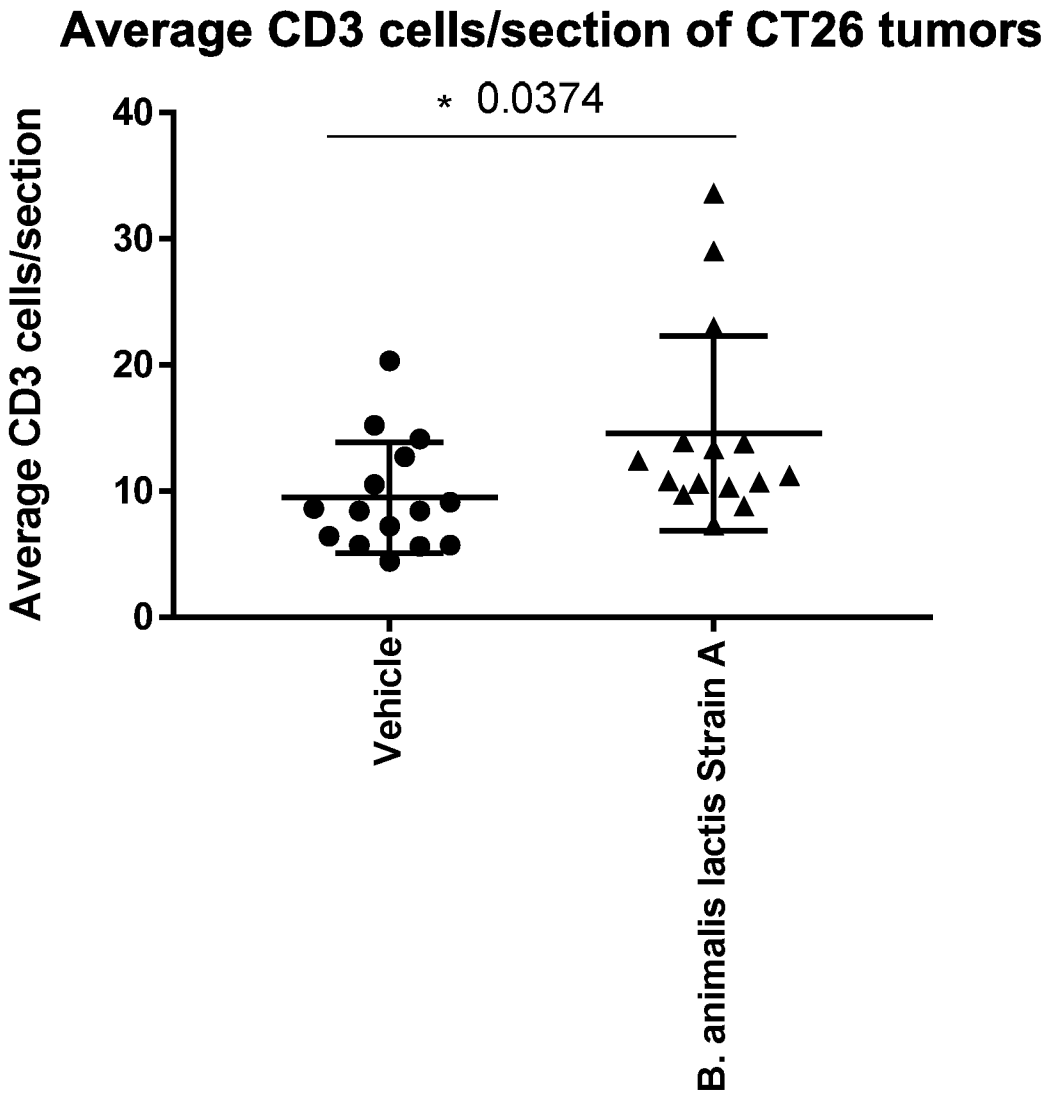

Average CD3 cells/section of CT26 tumors
* 0.0374
Average CD3 cells/section
40
30
20
10
0
Vehicle
B. animalis lactis Strain A

Figure 6C

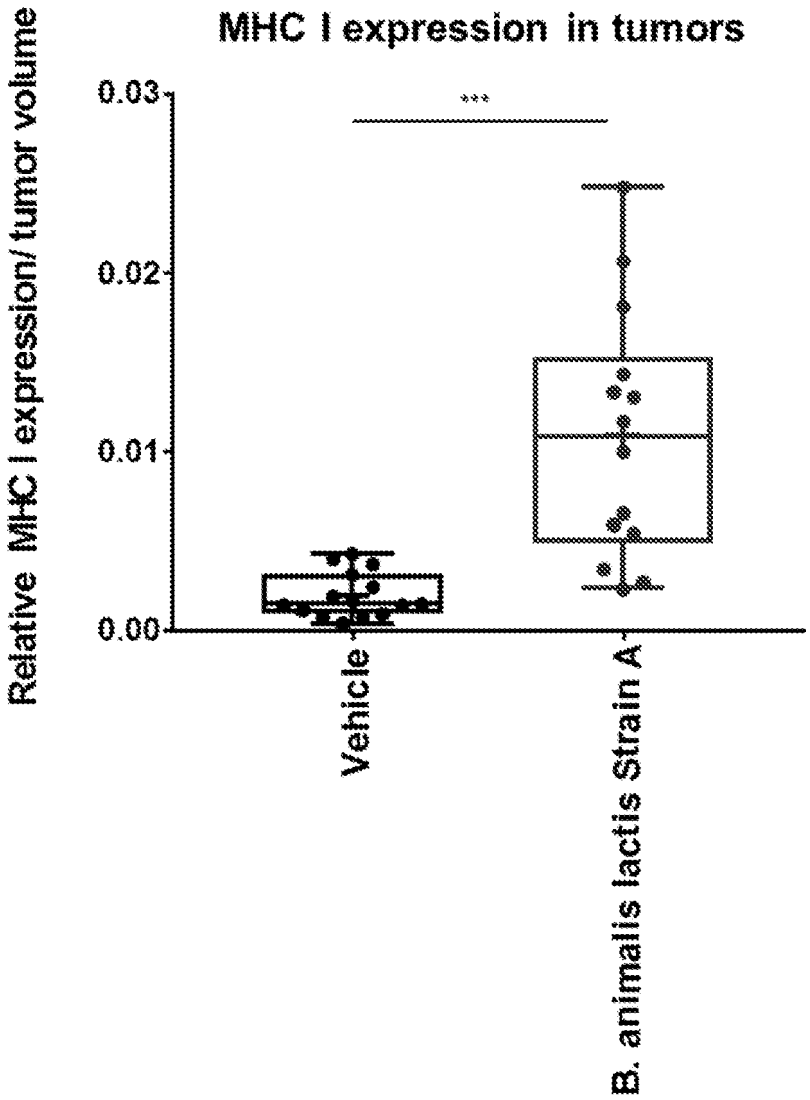

MHC I expression in tumors
Relative MHC I expression/ tumor volume
0.03
0.02
0.01
0.00
***
Vehicle
B. animalis lactis Strain A

Figure 7

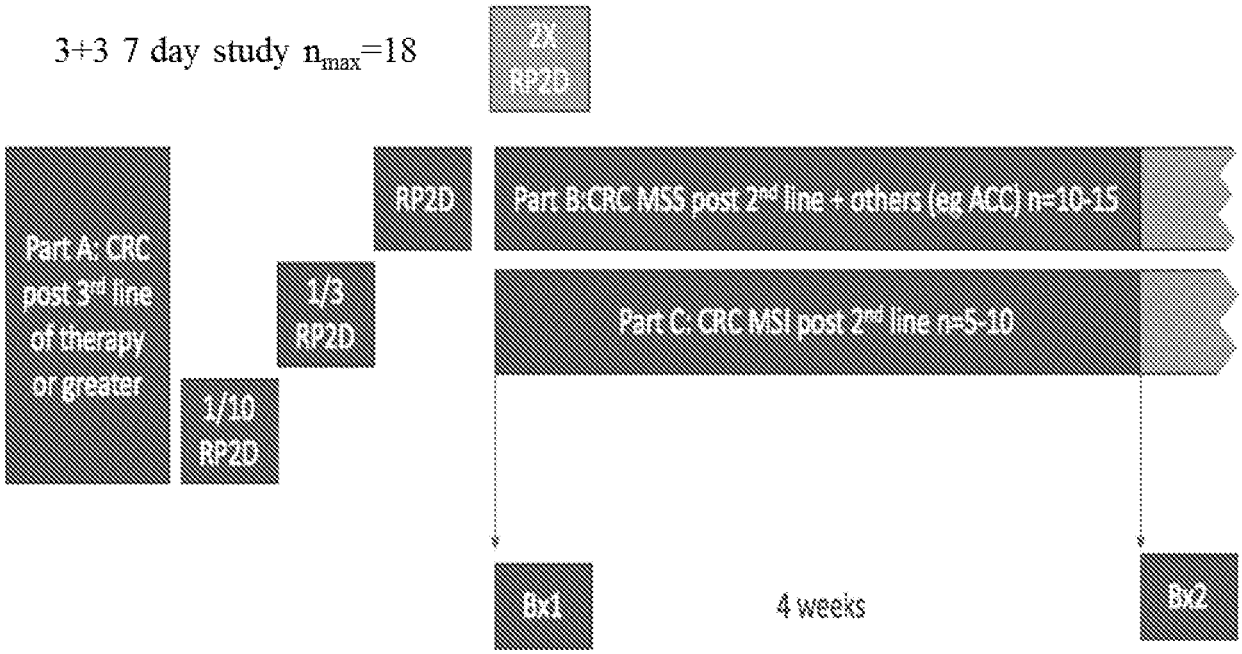

3+3 7 day study n_{max}=18
2X RP2D
Part A: CRC post 3rd line of therapy or greater
1/10 RP2D
1/3 RP2D
RP2D
Part B: CRC MSS post 2nd line + others (eg ACC) n=10-15
Part C: CRC MSI post 2nd line n=5-10
Bx1
4 weeks
Bx2

Figure 8

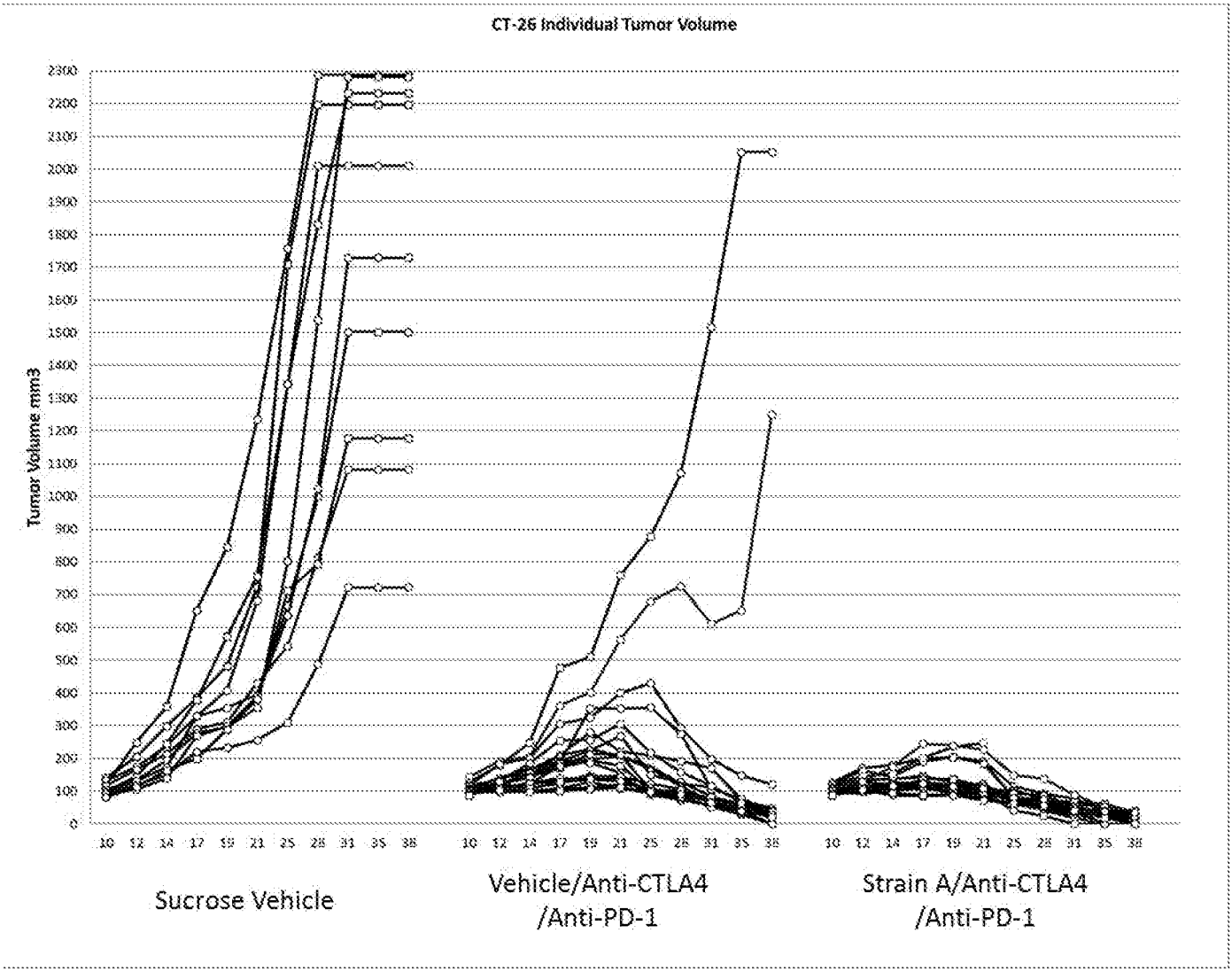

CT-26 Individual Tumor Volume
Tumor Volume mm3
0
100
200
300
400
500
600
700
800
900
1000
1100
1200
1300
1400
1500
1600
1700
1800
1900
2000
2100
2200
2300
10 12 14 17 19 23 25 28 31 35 38
Sucrose Vehicle
Vehicle/Anti-CTLA4 /Anti-PD-1
Strain A/Anti-CTLA4 /Anti-PD-1

Figure 9A

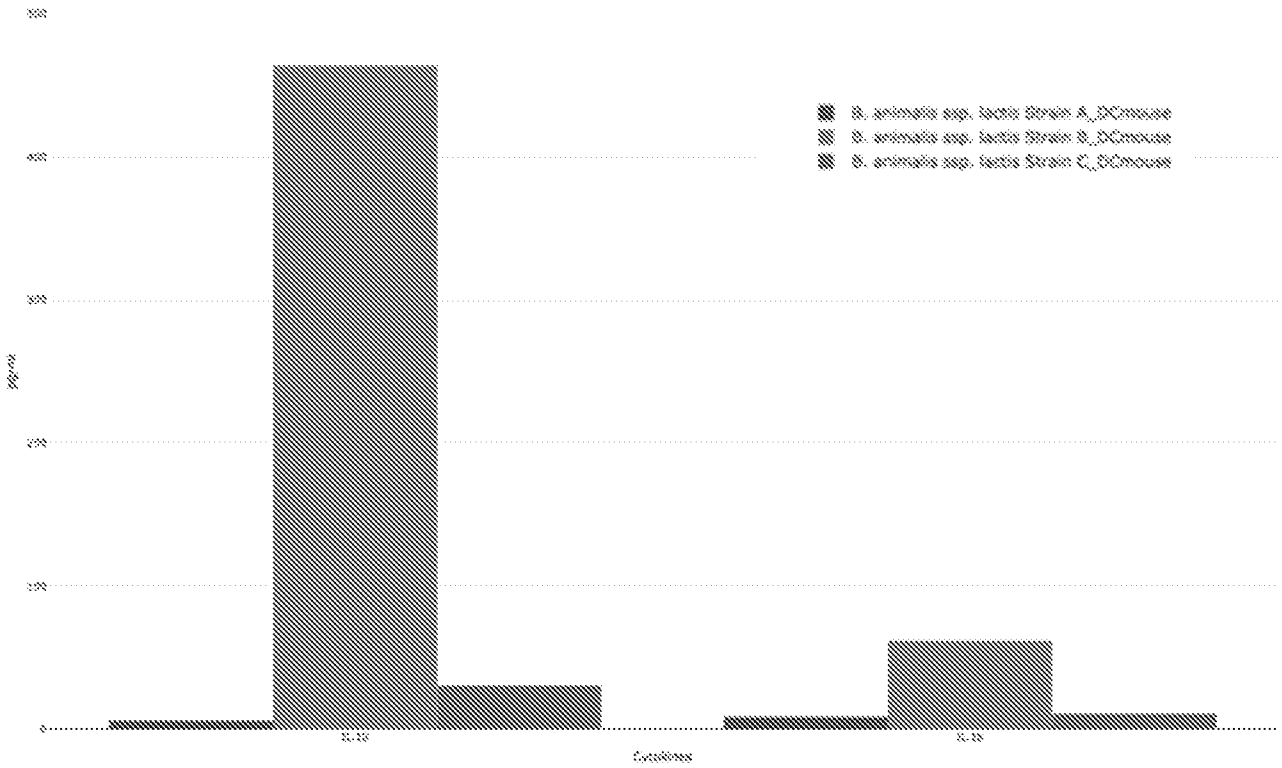
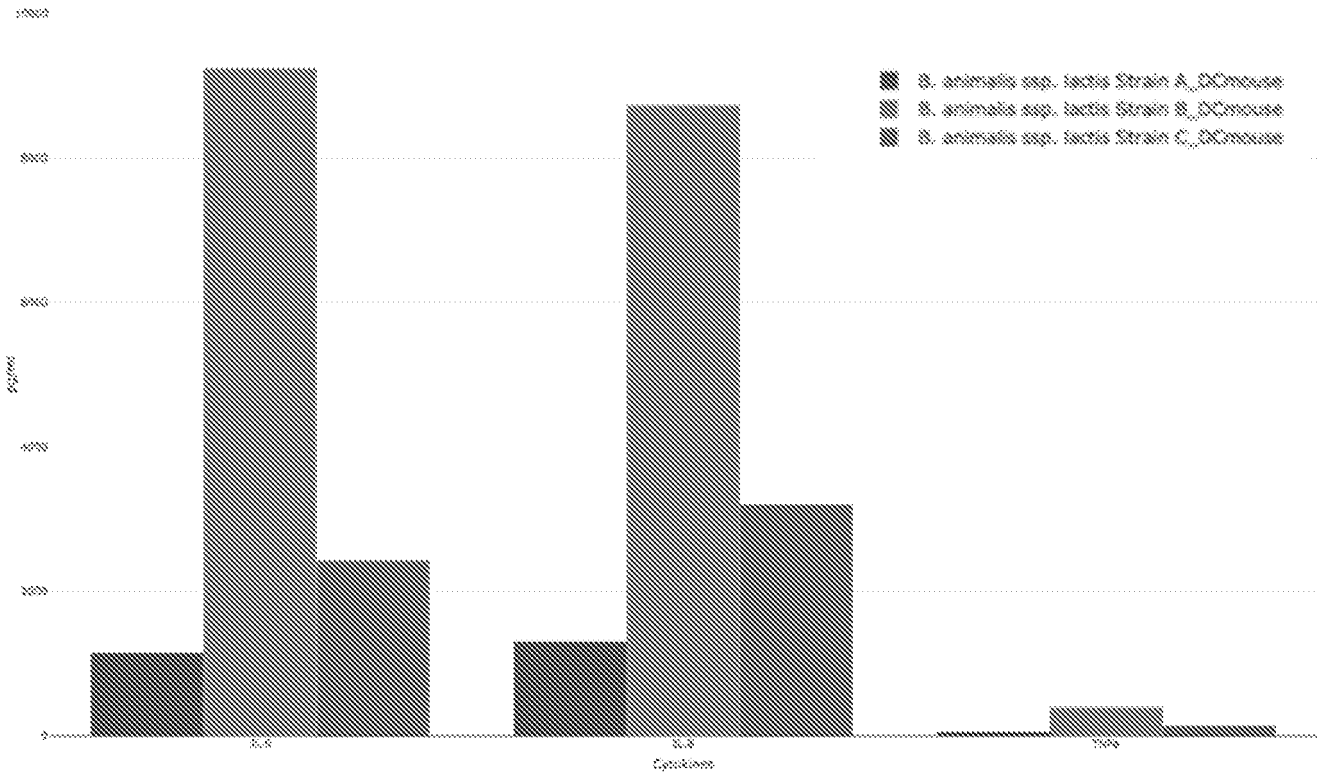


Figure 9B


B. animalis ssp. lactis Strain A_2Omouse
B. animalis ssp. lactis Strain B_2Omouse
B. animalis ssp. lactis Strain C_2Omouse