

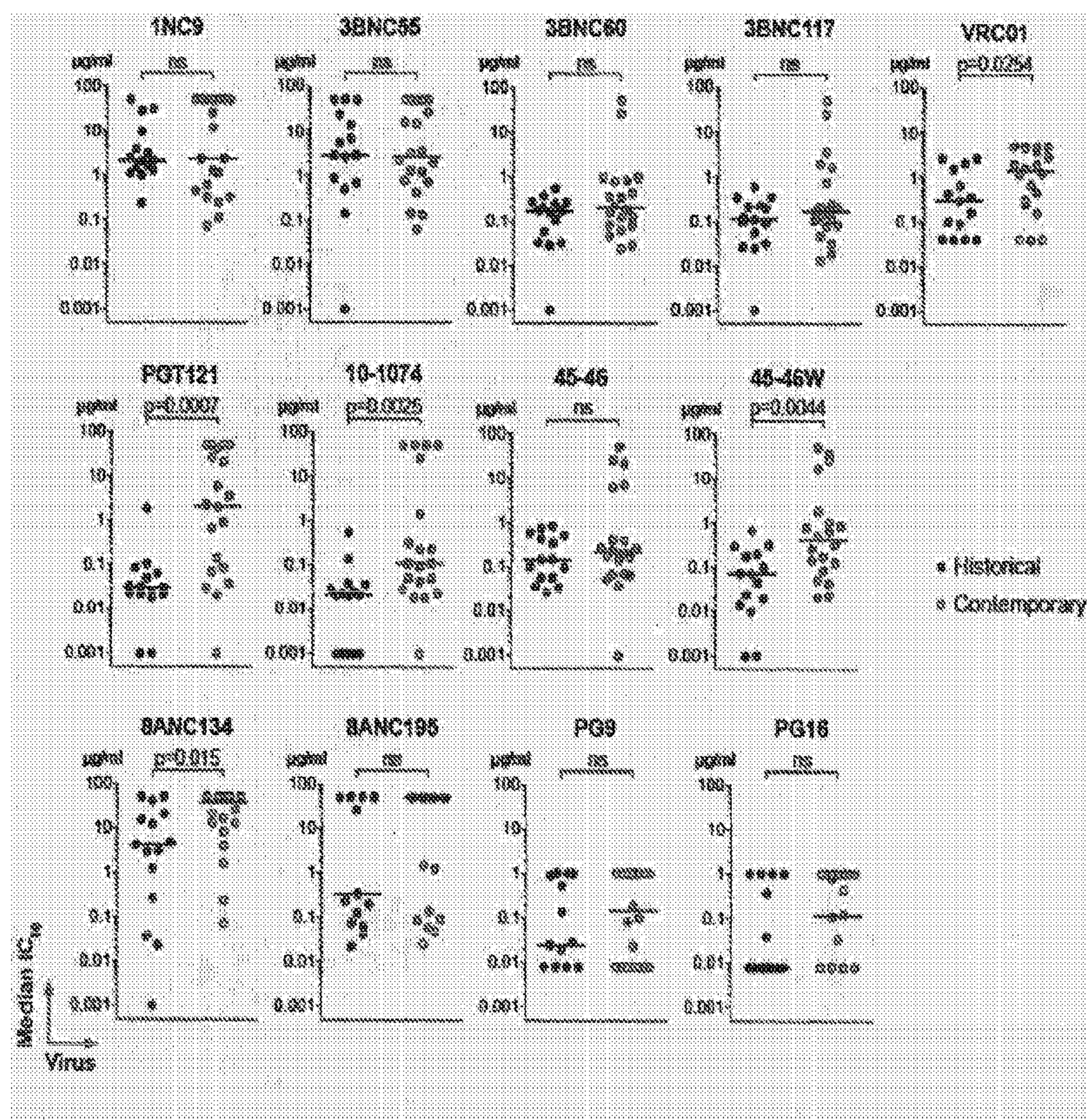


(86) Date de dépôt PCT/PCT Filing Date: 2013/10/18
(87) Date publication PCT/PCT Publication Date: 2014/04/24
(85) Entrée phase nationale/National Entry: 2015/04/16
(86) N° demande PCT/PCT Application No.: US 2013/065696
(87) N° publication PCT/PCT Publication No.: 2014/063059
(30) Priorité/Priority: 2012/10/18 (US61/715,642)

(51) Cl.Int./Int.Cl. *C07K 16/10* (2006.01),
A61K 39/42 (2006.01), *A61P 31/18* (2006.01),
C12N 15/13 (2006.01), *C12Q 1/68* (2006.01),
G01N 33/564 (2006.01)
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(54) Titre : ANTICORPS ANTI-VIH A NEUTRALISATION LARGE

(54) Title: BROADLY-NEUTRALIZING ANTI-HIV ANTIBODIES



(57) Abrégé/Abstract:

The present invention relates to anti-HIV antibodies. Also disclosed are related methods and compositions. HIV causes acquired immunodeficiency syndrome (AIDS), a condition in humans characterized by clinical features including wasting syndromes, central nervous system degeneration and profound immunosuppression that results in life-threatening opportunistic infections and malignancies. Since its discovery in 1981, HIV type 1 (HIV -1) has led to the death of at least 25 million people worldwide.



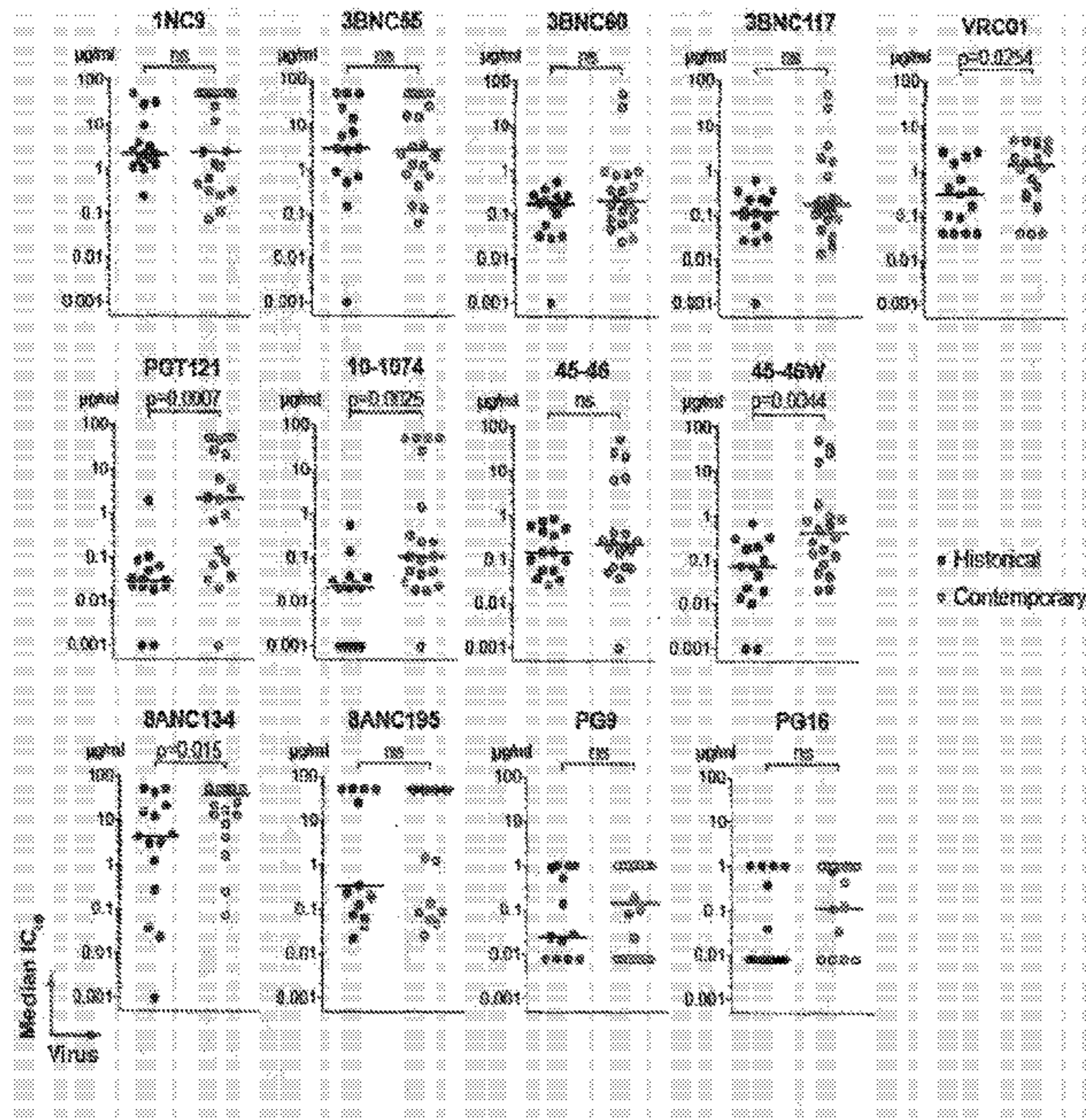
- (51) International Patent Classification:
A61P 31/18 (2006.01)
- (21) International Application Number:
PCT/US2013/065696
- (22) International Filing Date:
18 October 2013 (18.10.2013)
- (25) Filing Language: English
- (26) Publication Language: English
- (30) Priority Data:
61/715,642 18 October 2012 (18.10.2012) US
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- (81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BN, BR, BW, BY, BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IR, IS, JP, KE, KG, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LT, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PA, PE, PG, PH, PL, PT, QA, RO, RS, RU, RW, SA, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.
- (84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LR, LS, MW, MZ, NA, RW, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, RU, TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, KM, ML, MR, NE, SN, TD, TG).

Published:
— with international search report (Art. 21(3))

[Continued on next page]

(54) Title: BROADLY-NEUTRALIZING ANTI-HIV ANTIBODIES



(57) Abstract: The present invention relates to anti-HIV antibodies. Also disclosed are related methods and compositions. HIV causes acquired immunodeficiency syndrome (AIDS), a condition in humans characterized by clinical features including wasting syndromes, central nervous system degeneration and profound immunosuppression that results in life-threatening opportunistic infections and malignancies. Since its discovery in 1981, HIV type 1 (HIV -1) has led to the death of at least 25 million people worldwide.

WO 2014/063059 A1

WO 2014/063059 A1



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- *before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments (Rule 48.2(h))*

BROADLY-NEUTRALIZING ANTI-HIV ANTIBODIES

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims priority under 35 U.S.C. § 119(e) to U.S. Provisional Application No. 61/715,642 filed on October 18, 2012 and is hereby incorporated in its entirety.

5 GOVERNMENT INTERESTS

The invention disclosed herein was made, at least in part, with government support under Grant No. P01 AI081677 from the National Institutes of Health. Accordingly, the U.S. Government has certain rights in this invention.

FIELD OF THE INVENTION

10 This invention relates to broad and potent antibodies against Human Immunodeficiency Virus (“HIV”).

BACKGROUND OF THE INVENTION

HIV causes acquired immunodeficiency syndrome (AIDS), a condition in humans characterized by clinical features including wasting syndromes, central nervous system
15 degeneration and profound immunosuppression that results in life-threatening opportunistic infections and malignancies. Since its discovery in 1981, HIV type 1 (HIV-1) has led to the death of at least 25 million people worldwide. It is predicted that 20-60 million people will become infected over the next two decades even if there is a 2.5% annual decrease in HIV infections. There is a need for therapeutic agents and methods for treatment or inhibition of HIV infection.

20 Some HIV infected individuals show broadly neutralizing IgG antibodies in their serum. Yet, little is known regarding the specificity and activity of these antibodies, despite their potential importance in designing effective vaccines. In animal models, passive transfer of neutralizing antibodies can contribute to protection against virus challenge. Neutralizing antibody responses also can be developed in HIV-infected individuals but the detailed composition of the serologic
25 response is yet to be fully uncovered.

SUMMARY OF INVENTION

This invention relates to new categories of broadly-neutralizing anti-HIV antibodies. The consensus heavy and light chain amino acid sequences of the antibodies are listed below and shown in Figures 3a and 3b:

5 QVQLQESGPGGLVKPSETLSLTCSVSGX₁SX₂X₃DX₄YWSWIRQSPGKGLEWIGYVHDSGDTNYPNPSL
KSRVX₅X₆SLDTSKNQVSLKLX₇X₈VTAAADSAX₉YYCARAX₁₀HGX₁₁RIYGIVAFGEX₁₂FTYFYMDV
WGKGTTVTVSS (SEQ ID NO: 1)

10 SX₁VRPQPPSLSVAPGETARIX₂CGEX₃SLGSRAVQWYQQRPGQAPSLIIYNNQDRPSGIPERFSG
SPDX₄X₅FGTTATLTITX₆VEAGDEADYYCHIWDSRX₇PTX₈WVFGGGTTTLTVL (SEQ ID NO: 2)

In the sequence of SEQ ID NO: 1 or 2, each “X” can be any amino acid residue or no amino acid. Preferably, each of the Xs can be a residue at the corresponding location of clonal variants 10-259, 10-303, 10-410, 10-847, 10-996, 10-1074, 10-1121, 10-1130, 10-1146, 10-1341, and 10-1369 as
15 shown in Figures 3a and 3b, and an artificially modified version of 10-1074 antibody, 10-1074GM.

Accordingly, one aspect of this invention features an isolated anti-HIV antibody, or antigen binding portion thereof, having at least one complementarity determining region (CDR) having a sequence selected from the group consisting of SEQ ID NOs: 33-38, with a proviso that
20 the antibody is not antibody PGT-121, 122, or 123. SEQ ID NOs: 33-38 refer to the sequences of heavy chain CDRs (CDRH) 1-3 and the light chain CDRs (CDRL) 1-3 under the Kabat system as shown in Figures 3a and 3b. In one embodiment, the CDR can contain a sequence selected from the group consisting of SEQ ID NOs: 39-104, i.e., the CDR sequences under the KABAT system as shown in Table 1 below. Alternatively, the CDR can contain a sequence selected from those
25 corresponding antibodies’ CDR sequences under the IMGT system as shown in Table 1 below.

In one embodiment, the isolated anti-HIV antibody, or antigen binding portion thereof, contains a heavy chain variable region that includes CDRH 1, CDRH 2, and CDRH 3, wherein the CDRH 1, CDRH 2 and CDRH 3 include the respective sequences of SEQ ID NOs: 33-35. The CDRH 1, CDRH 2 and CDRH 3 can also include the respective sequences of a CDRH set selected
30 from the group consisting of SEQ ID NOs: 39-41, SEQ ID NOs: 45-47, SEQ ID NOs: 51-53, SEQ ID NOs: 57-59, SEQ ID NOs: 63-65, SEQ ID NOs: 69-71, SEQ ID NOs: 75-77, SEQ ID NOs: 81-83, SEQ ID NOs: 87-89, SEQ ID NOs: 93-95, SEQ ID NOs: 99-101, and SEQ ID NOs: 131-

133.. Alternatively, the CDRHs can contain the respective sequences selected from those corresponding antibodies' CDR sequences under the IMGT system as shown in Table 1 below.

In another embodiment, the isolated anti-HIV antibody, or antigen binding portion thereof, contains a light chain variable region that includes CDRL 1, CDRL 2 and CDRL 3, wherein the CDRL 1, CDRL 2 and CDRL 3 include the respective sequences of SEQ ID NOs: 36-38. For example, the CDRL 1, CDRL 2 and CDRL 3 can include the respective sequences of a CDRL set selected from the group consisting of SEQ ID NOs: 42-44, SEQ ID NOs: 48-50, SEQ ID NOs: 54-56, SEQ ID NOs: 60-62, SEQ ID NOs: 66-68, SEQ ID NOs: 72-74, SEQ ID NOs: 78-80, SEQ ID NOs: 84-86, SEQ ID NOs: 90-92, SEQ ID NOs: 96-98, SEQ ID NOs: 102-104, and SEQ ID NOs: 134-136. Alternatively, the CDRLs can contain the respective sequences selected from those corresponding antibodies' CDR sequences under the IMGT system as shown in Table 1 below.

In yet another embodiment, the above-mentioned isolated anti-HIV antibody, or antigen binding portion thereof, includes (i) a heavy chain variable region that include CDRH 1, CDRH 2, and CDRH 3, and (ii) a light chain variable region that include CDRL 1, CDRL 2 and CDRL 3. The CDRH 1, CDRH 2, CDRH 3, CDRL 1, CDRL 2 and CDRL 3 can include the respective sequences of a CDR set selected from the group consisting of SEQ ID NOs: 39-44, SEQ ID NOs: 45-50, SEQ ID NOs: 51-56, SEQ ID NOs: 57-62, SEQ ID NOs: 63-68, SEQ ID NOs: 69-74, SEQ ID NOs: 75-79, SEQ ID NOs: 81-86, SEQ ID NOs: 87-92, SEQ ID NOs: 93-98, SEQ ID NOs: 99-104, and SEQ ID NOs: 131-136. Alternatively, the CDRHs and CDRLs can contain the respective sequences selected from those corresponding antibodies' CDR sequences under the IMGT system as shown in Table 1 below.

In a further embodiment, the isolated anti-HIV antibody, or antigen binding portion thereof, contains one or both of (i) a heavy chain having the consensus amino acid sequence of SEQ ID NO: 1 and (ii) a light chain having the consensus amino acid sequence of SEQ ID NO: 2. The heavy chain can contain a sequence selected from the group consisting of SEQ ID NOs: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, and 129, and the light chain can contain a sequence selected from the group consisting of SEQ ID NOs: 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, and 130. For example, the heavy chain and the light chain can include the respective sequences of SEQ ID NOs: 3-4, SEQ ID NOs: 5-6, SEQ ID NOs: 7-8, SEQ ID NOs: 9-10, SEQ ID NOs: 11-12, SEQ ID NOs: 13-

14, SEQ ID NOs: 15-16, SEQ ID NOs: 17-18, SEQ ID NOs: 19-20, SEQ ID NOs: 21-22, SEQ ID NOs: 23-24, and 129-130.

In a preferred embodiment, the isolated anti-HIV antibody is one selected from the group consisting of 10-259, 10-303, 10-410, 10-847, 10-996, 10-1074, 10-1074GM, 10-1121, 10-1130, 5 10-1146, 10-1341, and 10-1369. Their corresponding heavy chain variable regions, light chain variable regions, CDRH 1-3 and CDRL 1-3 are shown in Figures 3a and 3b. In a more preferred embodiment, the isolated anti-HIV antibody is a 10-1074-like antibody, i.e., one reselected from the group consisting of 10-847, 10-996, 10-1074, 10-1074GM, 10-1146, and 10-1341. An antibody of this group is more potent in neutralizing contemporary viruses than PGT121. The 10 above-discussed antibody can be a human antibody, a humanized antibody, or a chimeric antibody.

In a second aspect, the invention provides an isolated nucleic acid having a sequence encoding a CDR, a heavy chain variable region, or a light chain variable region of the above-discussed anti-HIV antibody, or antigen binding portion thereof. Also featured are a vector 15 having the nucleic acid and a cultured cell having the vector.

The nucleic acid, vector, and cultured cell can be used in a method for making an anti-HIV antibody or a fragment thereof. The method includes, among others, the steps of: obtaining the cultured cell mentioned above; culturing the cell in a medium under conditions permitting expression of a polypeptide encoded by the vector and assembling of an antibody or fragment 20 thereof, and purifying the antibody or fragment from the cultured cell or the medium of the cell.

In a third aspect, the invention features a pharmaceutical composition containing (i) at least one anti-HIV antibody mentioned above, or antigen binding portion thereof, and (ii) a pharmaceutically acceptable carrier.

In a fourth aspect, the invention provides a method of preventing or treating an HIV 25 infection or an HIV-related disease. The method includes, among others, the steps of: identifying a patient in need of such prevention or treatment, and administering to said patient a first therapeutic agent containing a therapeutically effective amount of at least one anti-HIV antibody mentioned above, or antigen binding portion thereof. The method can further include administering a second therapeutic agent, such as an antiviral agent.

30 In a fifth aspect, the invention provides a kit having a pharmaceutically acceptable dose unit of a pharmaceutically effective amount of at least one isolated anti-HIV antibody mentioned a

above, or antigen binding portion thereof, and a pharmaceutically acceptable dose unit of a pharmaceutically effective amount of an anti-HIV agent. The two pharmaceutically acceptable dose units can optionally take the form of a single pharmaceutically acceptable dose unit. Exemplary anti-HIV agent can be one selected from the group consisting of a non-nucleoside reverse transcriptase inhibitor, a protease inhibitor, a entry or fusion inhibitor, and an integrase inhibitor.

In a sixth aspect, the invention provides a kit for the diagnosis, prognosis or monitoring the treatment of an HIV infection in a subject. The kit contains one or more detection reagents which specifically bind to anti-HIV neutralizing antibodies in a biological sample from a subject. The kit can further include reagents for performing PCR or mass spectrometry.

The details of one or more embodiments of the invention are set forth in the description below. Other features, objects, and advantages of the invention will be apparent from the description and from the claims.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 shows: Neutralization activity of PGT121-like and 10-1074-like variants. (A) Heat map comparing the neutralization potencies of PGT121-like and 10-1074-like antibodies in the TZM-bl assay. Darker colors = more potent neutralization; white = no neutralization. (B) Correlation between the mean IC_{80} against 9 viruses (y axis) and apparent K_D values for binding to gp120 and gp140 (x axis). (C) Graph comparing the neutralization breadth and potencies of PGT121, 10-996 and 10-1074 antibodies in the TZM-bl assay against an extended panel of 119 viruses. The y axis shows the cumulative frequency of IC_{50} values up to the concentration shown on the x axis. The spider graph (upper left corner) shows the frequency distribution of neutralized viruses according to HIV-1 clades. (D) Dot plot showing molar neutralization ratios (MNRs; ratio of the Fab and IgG IC_{50} concentrations). Horizontal bars represent the mean IC_{50} s for all viruses. (E) Bar graph comparing the neutralization potencies of PGT121 (dark gray) and 10-1074 (light gray) against viruses isolated from historical (Hist.) and contemporary (Cont.) seroconverters. ns, non significant; **, $p < 0.005$. Fold difference between median IC_{50} s for the neutralization of contemporary viruses by PGT121 and 10-1074 is indicated.

Fig. 2 shows: Binding and neutralization activities of PGT121_{GM} and 10-1074_{GM} mutant antibodies. (A) Bar graphs comparing apparent K_D values for the binding of 10-1074, PGT121, PGT121_{GM} and 10-1074_{GM} antibodies to gp120 and gp140. Error bars indicate the SEM

of K_D values obtained from three independent experiments. Fold differences between K_D values of “wildtype” vs “glycomutant” antibodies are indicated. (B) Bar graphs comparing binding of glycans (**Figure 7A**) by PGT121 and 10-1074 with mutant antibodies (PGT121_{GM} and 10-1074_{GM}). Numerical scores of binding are measured as fluorescence intensity (means at duplicate spots) for probes arrayed at 5 fmol per spot. (C) Coverage graph comparing the neutralization breadth and potencies of PGT121, PGT121_{GM}, 10-1074 and 10-1074_{GM} antibodies in the TZM-bl assay against a panel of 40 viruses.

Fig. 3 depicts: Sequence alignments of PGT121 and 10-1074 clonal variants. (A) Amino acid alignment of the heavy chains (IgH) of the PGT121-like and 10-1074-like antibodies, and the likely germline (GL) VH for all clonal variants. Amino acid numbering based on crystal structures, framework (FWR) and complementary determining regions (CDR) as defined by Kabat (. J Exp Med 132(2):211-250) and IMGT (Nucleic Acids Res 37(Database issue):D1006-1012) are indicated. Color shading shows acidic (red), basic (blue), and tyrosine (green) amino acids. (B) Same as A but for the light chains (IgL).

Fig. 4 shows: Binding affinity of PGT121 and 10-1074 clonal variants. (A) Binding affinity of the interaction of PGT121 IgG antibody variants with YU-2 gp140 and gp120 ligands as measured by surface plasmon resonance (SPR). M, mol/l; s, seconds; RU, response units; /, no binding detected. A χ^2 value (χ^2) < 10 indicates that the 1:1 binding model used to fit the curves adequately described the experimental data. Equilibrium and kinetic constants shown are considered as “apparent” constants to account for avidity effects resulting from bivalent binding of IgGs. (B) Dot plots showing the association (k_a) and dissociation (k_d) rate constants for PGT121-like (blue shading) and 10-1074-like (green shading). (C) Linear regression graphs comparing the k_a and k_d values of the IgG antibodies for their binding to gp120 and gp140 (x axis) vs their neutralization potencies (mean IC₈₀ values) against the 9 viruses shown in **Table 4** (y axis).

Fig. 5 depicts: Binding of PGT121 variants to gp120 “core” proteins, gp120^{GD324-5AA} mutant and linear gp120^{V3} peptides. (A) ELISA-based binding analyses of PGT121-like and 10-1074-like antibodies to HXB2 gp120^{core} and 2CC-core proteins compared to intact YU-2 gp120. The x axis shows the antibody concentration (M) required to obtain the ELISA values (OD_{405 nm}) indicated on the y axis. The anti-CD4bs antibody VRC01 (Science 329(5993):856-861), the anti-V3 loop antibody 10-188 (PLoS One 6(9):e24078), and the non HIV-reactive antibody mGO53

(Science 301(5638):1374-1377) were used as controls. (B) Same as (A) but for binding to gp120^{GD324-5AA} mutant protein (c) Bar graphs comparing the ELISA reactivities of the PGT121- and 10-1074-like antibodies and control antibodies (positive control, 10-188, 1-79, 2-59 and 2-1261 (Nature 458(7238):636-640)), and negative control, mGO53) against gp120^{V3-C3} overlapping peptides. The y axis indicates the ELISA values (OD_{405 nm}) obtained by testing the IgG antibodies at 2 µg/ml. The amino acid sequences of individual peptides are shown in the bottom right. All experiments were performed at least in duplicate. Representative data are shown.

Fig. 6 depicts: Binding of PGT121 to gp120 glycosylation mutants and deglycosylated gp120. (A) ELISA-based binding analyses of PGT121 and 10-1074 antibody variants to gp120, gp120^{NNT301-303AAA}, gp120^{N332A} and gp120^{N332A/NNT301-303AAA}. The x axis shows the antibody concentration (M) required to obtain the ELISA values (OD_{405 nm}) indicated on the y axis. The black dashed and continuous lines show the averaged reactivity against the four antigens of positive (10-188) and negative (mGO53) antibody controls. (B) Silver-stained SDS-PAGE gel comparing untreated gp120 (WT, wild type), PNGase F- and EndoH-digested gp120s. L, protein ladder. (C), Same as (A) but comparing untreated and PNGase F-treated gp120. (D) Same as (A) but comparing untreated and EndoH-treated gp120. All experiments were performed at least in duplicate.

Fig. 7 depicts: Binding of PGT121 and 10-1074 clonal variants to glycans. (A) Monosaccharide sequences of the set of 15 N-glycan probes used in the glycan microarray analyses to examine PGT121-like and 10-1074-like antibodies for direct binding to N-glycans. DH, designates the lipid tag 1,2-dihexadecyl-*sn*-glycero-3-phosphoethanolamine (DHPE) to which the N-glycans were conjugated by reductive amination. Key features of note are (i) PGT121-group antibodies bound the monoantennary N-glycan probe 10 (N2) with a galactose-terminating antenna joined by 1-3-linkage to the core mannose, but not the isomeric N-glycan probe 11 (designated N4) with the antenna 1-6-linked to the core mannose; (ii) the presence of this galactose-terminating 1-6-linked antenna, as in the biantennary probe 13 (NA2), was permissive to binding, as was the presence of α2-6-linked (but not α2-3-linked) sialic acid; (iii) the biantennary probe 12 (NGA2), lacking galactose and terminating in N-acetylglucosamine, was not bound. (B) Bar graphs comparing glycan binding by PGT121-like, 10-1074-like, and the germline version (GL) antibodies. 10-188, an anti-V3 loop antibody, was used as negative control.

Numerical scores of binding are measured as fluorescence intensity (means at duplicate spots) for probes arrayed at 2 fmol (white) and 5 fmol per spot (grey).

Fig. 8 depicts: Antibody binding and neutralization activity against high-mannose-only gp120 and viruses. (A) Silver-stained SDS-PAGE gel comparing YU-2 gp120 produced in cells treated with kifunensine (gp120_{kif}) and gp120 produced in untreated cells (WT, wild type). L, protein ladder. (B) ELISA comparison of the binding of PGT121-like (blue labels) and 10-1074-like (green labels) antibodies to YU-2 gp120 (gp120_{WT}) and gp120_{kif}. The *x* axis shows the antibody concentration (M) required to obtain the ELISA values (OD_{405 nm}) indicated on the *y* axis. (C) Neutralization curves for PGT121 evaluated against selected PGT121-sensitive/10-1074-resistant pseudoviruses produced in presence (Virus_{kif}) or absence (Virus_{WT}) of kifunensine. The dotted horizontal line indicates 50% neutralization, from which the IC₅₀ value can be derived from the antibody concentration on the *x* axis. Experiments were performed in triplicate. Error bars indicate the SD of triplicate measurements. (D) Bar graphs comparing the neutralization activity of selected antibodies against YU-2 and PVO.4 pseudoviruses produced in HEK 293S GnTI^{-/-} cells (Virus_{GnTI^{-/-}}) or in wild type cells (Virus_{WT}). The *y* axis shows the mean IC₅₀ values (μg/ml) for the neutralization of the viruses shown on the *x* axis. Error bars indicate the SEM of IC₅₀ values obtained from two independent experiments.

Fig. 9 shows: Neutralization activity of PGT121, 10-996 and 10-1074. (A) Graphs comparing the neutralization potencies of PGT121, 10-996 and 10-74 against viruses of the indicated HIV-1 clades (determined using the TZM-bl assay and a panel of 119 pseudoviruses). The *x* axis shows the antibody concentration (μg/ml) required to achieve 50% neutralization (IC₅₀). The *y* axis shows the cumulative frequency of IC₅₀ values up to the concentration shown on the *x* axis. (B) Graph comparing the neutralization breadth and potencies of PGT121, 10-996 and 10-1074 antibodies against the extended panel of 119 viruses as determined by the TZM-bl neutralization assay. The *y* axis shows the cumulative frequency of IC₈₀ values up to the concentration shown on the *x* axis. (C) Graphs show neutralization curves of the selected viruses by PGT121 and 10-1074. The dotted horizontal line indicates 50% neutralization, from which the IC₅₀ value can be derived from the antibody concentration on the *x*-axis. Experiments were performed in triplicate. Error bars indicate the SD of triplicate measurements.

Fig. 10 depicts: Neutralization activity against historical vs contemporary clade B viruses. Dot plots comparing neutralization potencies against clade B viruses isolated from

historical (Hist.) and contemporary (Cont.) seroconverters for the selected bNAbs. Horizontal bars represent the median IC₅₀ for all viruses per patient. Differences between groups were evaluated using Mann-Whitney test. ns, not significant.

Fig. 11 depicts: Neutralization of two R5 tropic SHIVs with a panel of 11 broadly acting anti-HIV-1 mAbs. The calculated IC₅₀ values for neutralizing SHIVAD8EO (A) and SHIVDH12-V3AD8 (B).

Fig. 12 depicts: The relationship of the plasma concentrations of passively administered neutralizing mAbs to virus acquisition following challenge of macaques with two different R5 SHIVs. Filled circles indicate protected (no acquisition) monkeys; open circles denote infected animals.

Fig. 13 depicts: Plasma concentration of bNAbs. The concentration of mAbs was determined by measuring neutralization activity in plasma samples. (A) ID₅₀-values measured in TZM.bl neutralization assay of 10-1074 and 3BNC117 against HIV-1 strains that are sensitive to one but not the other bNAb (i.e. HIV-1 strain X2088_9 (10-1074 sensitive); HIV-1 strain Q769_d22 (3BNC117 sensitive). (B) Neutralizing activity of plasma before antibody administration (preP), but spiked with 0.01, 0.1, 1, 10, and 100 µg/ml of antibodies 10-1074 (blue) or 3BNC117 (green). Neutralizing activity reported as plasma ID₅₀ titers (left columns) and converted to antibody concentrations (right columns) based on measured ID₅₀-values in (A). (C) ID₅₀ titers (left columns) and concentrations of bNAbs (right columns) measured in the indicated macaque plasma samples before (Prebleed) and following (Day) bNAb administration.

DETAILED DESCRIPTION OF THE INVENTION

This invention is based, at least in part, on an unexpected discovery of a new category of broadly neutralizing antibodies (bNAbs) against HIV that can recognize carbohydrate-dependent epitopes, including complex-type *N*-glycan, on gp120.

Antibodies are essential for the success of most vaccines, and antibodies against HIV appear to be the only correlate of protection in the recent RV144 anti-HIV vaccine trial. Some HIV-1 infected patients develop broadly neutralizing serologic activity against the gp160 viral spike 2-4 years after infection, but these antibodies do not generally protect infected humans because autologous viruses escape through mutation. Nevertheless, broadly neutralizing activity puts selective pressure on the virus and passive transfer of broadly neutralizing antibodies

(bNAbs) to macaques protects against SHIV infection. It has therefore been proposed that vaccines that elicit such antibodies may be protective against HIV infection in humans.

The development of single cell antibody cloning techniques revealed that bNAbs target several different epitopes on the HIV-1 gp160 spike. The most potent HIV-1 bNAbs recognize the CD4 binding site (CD4bs) (Science 333(6049):1633-1637; Nature 477(7365):466-470; Science 334(6060):1289-1293) and carbohydrate-dependent epitopes associated with the variable loops (Nature 477(7365):466-470; Science 326(5950):285-289; Science 334(6059):1097-1103; Nature 480(7377):336-343), including the V1/V2 (PG9/PG16) (Science 326(5950):285-289) and V3 loops (PGTs) (Nature 477(7365):466-470). Less is known about carbohydrate-dependent epitopes because the antibodies studied to date are either unique examples or members of small clonal families.

To better understand the neutralizing antibody response to HIV-1 and the epitope targeted by PGT antibodies, we isolated members of a large clonal family dominating the gp160-specific IgG memory response from the clade A-infected patient who produced PGT121. As disclosed herein, PGT121 antibodies segregate into two groups, a PGT121-like and a 10-1074-like group, according to sequence, binding affinity, neutralizing activity and recognition of carbohydrates and the V3 loop. 10-1074 and related family members exhibit unusual potent neutralization, including broad reactivity against newly-transmitted viruses. Unlike previously-characterized carbohydrate-dependent bNAbs, PGT121 binds to complex-type, rather than high-mannose, *N*-glycans in glycan microarray experiments. Crystal structures of PGT121 and 10-1074 compared with structures of their germline precursor and a structure of PGT121 bound to a complex-type *N*-glycan rationalize their distinct properties.

In one example, assays were carried out to isolate B-cell clones encoding PGT121, which is unique among glycan-dependent bNAbs in recognizing complex-type, rather than high-mannose, *N*-glycans. The PGT121 clones segregates into PGT121- and 10-1074-like groups distinguished by sequence, binding affinity, carbohydrate recognition and neutralizing activity. The 10-1074 group exhibit remarkable potency and breadth despite not binding detectably to protein-free glycans. Crystal structures of un-liganded PGT121, 10-1074, and their germline precursor reveal that differential carbohydrate recognition maps to a cleft between CDRH2 and CDRH3, which was occupied by a complex-type *N*-glycan in a separate PGT121 structure. Swapping glycan contact residues between PGT121 and 10-1074 confirmed the importance of

these residues in neutralizing activities. HIV envelopes exhibit varying proportions of high-mannose- and complex-type N-glycans, thus these results, including the first structural characterization of complex-type N-glycan recognition by anti-HIV bNAbs, are critical for understanding how antibodies and ultimately vaccines might achieve broad neutralizing activity.

5 The term "antibody" (Ab) as used herein includes monoclonal antibodies, polyclonal antibodies, multispecific antibodies (for example, bispecific antibodies and polyreactive antibodies), and antibody fragments. Thus, the term "antibody" as used in any context within this specification is meant to include, but not be limited to, any specific binding member, immunoglobulin class and/or isotype (e.g., IgG1, IgG2, IgG3, IgG4, IgM, IgA, IgD, IgE and
10 IgM); and biologically relevant fragment or specific binding member thereof, including but not limited to Fab, F(ab')₂, Fv, and scFv (single chain or related entity). It is understood in the art that an antibody is a glycoprotein having at least two heavy (H) chains and two light (L) chains interconnected by disulfide bonds, or an antigen binding portion thereof. A heavy chain is comprised of a heavy chain variable region (VH) and a heavy chain constant region (CH1, CH2 and CH3).
15 A light chain is comprised of a light chain variable region (VL) and a light chain constant region (CL). The variable regions of both the heavy and light chains comprise framework regions (FWR) and complementarity determining regions (CDR). The four FWR regions are relatively conserved while CDR regions (CDR1, CDR2 and CDR3) represent hypervariable regions and are arranged from NH₂ terminus to the COOH terminus as follows: FWR1, CDR1, FWR2, CDR2,
20 FWR3, CDR3, and FWR4. The variable regions of the heavy and light chains contain a binding domain that interacts with an antigen while, depending of the isotype, the constant region(s) may mediate the binding of the immunoglobulin to host tissues or factors.

Also included in the definition of "antibody" as used herein are chimeric antibodies, humanized antibodies, and recombinant antibodies, human antibodies generated from a transgenic
25 non-human animal, as well as antibodies selected from libraries using enrichment technologies available to the artisan.

The term "variable" refers to the fact that certain segments of the variable (V) domains differ extensively in sequence among antibodies. The V domain mediates antigen binding and defines specificity of a particular antibody for its particular antigen. However, the variability is
30 not evenly distributed across the 110-amino acid span of the variable regions. Instead, the V regions consist of relatively invariant stretches called framework regions (FRs) of 15-30 amino

acids separated by shorter regions of extreme variability called "hypervariable regions" that are each 9-12 amino acids long. The variable regions of native heavy and light chains each comprise four FRs, largely adopting a beta sheet configuration, connected by three hypervariable regions, which form loops connecting, and in some cases forming part of, the beta sheet structure. The hypervariable regions in each chain are held together in close proximity by the FRs and, with the hypervariable regions from the other chain, contribute to the formation of the antigen-binding site of antibodies (see, for example, Kabat *et al.*, Sequences of Proteins of Immunological Interest, 5th Ed. Public Health Service, National Institutes of Health, Bethesda, Md. (1991)).

The term "hypervariable region" as used herein refers to the amino acid residues of an antibody that are responsible for antigen binding. The hypervariable region generally comprises amino acid residues from a "complementarity determining region" ("CDR").

The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, i.e., the individual antibodies comprising the population are identical except for possible naturally occurring mutations that may be present in minor amounts. The term "polyclonal antibody" refers to preparations that include different antibodies directed against different determinants ("epitopes").

The monoclonal antibodies herein include "chimeric" antibodies in which a portion of the heavy and/or light chain is identical with, or homologous to, corresponding sequences in antibodies derived from a particular species or belonging to a particular antibody class or subclass, while the remainder of the chain(s) is identical with, or homologous to, corresponding sequences in antibodies derived from another species or belonging to another antibody class or subclass, as well as fragments of such antibodies, so long as they exhibit the desired biological activity (see, for example, U.S. Pat. No. 4,816,567; and Morrison *et al.*, Proc. Natl. Acad. Sci. USA, 81:6851-6855 (1984)). The described invention provides variable region antigen-binding sequences derived from human antibodies. Accordingly, chimeric antibodies of primary interest herein include antibodies having one or more human antigen binding sequences (for example, CDRs) and containing one or more sequences derived from a non-human antibody, for example, an FR or C region sequence. In addition, chimeric antibodies included herein are those comprising a human variable region antigen binding sequence of one antibody class or subclass and another sequence, for example, FR or C region sequence, derived from another antibody class or subclass.

A "humanized antibody" generally is considered to be a human antibody that has one or more amino acid residues introduced into it from a source that is non-human. These non-human amino acid residues often are referred to as "import" residues, which typically are taken from an "import" variable region. Humanization may be performed following the method of Winter and co-workers (see, for example, Jones *et al.*, Nature, 321:522-525 (1986); Reichmann *et al.*, Nature, 332:323-327 (1988); Verhoeven *et al.*, Science, 239:1534-1536 (1988)), by substituting import hypervariable region sequences for the corresponding sequences of a human antibody. Accordingly, such "humanized" antibodies are chimeric antibodies (see, for example, U.S. Pat. No. 4,816,567), where substantially less than an intact human variable region has been substituted by the corresponding sequence from a non-human species.

An "antibody fragment" comprises a portion of an intact antibody, such as the antigen binding or variable region of the intact antibody. Examples of antibody fragments include, but are not limited to, Fab, Fab', F(ab')₂, and Fv fragments; diabodies; linear antibodies (see, for example, U.S. Pat. No. 5,641,870; Zapata *et al.*, Protein Eng. 8(10): 1057-1062 [1995]); single-chain antibody molecules; and multispecific antibodies formed from antibody fragments.

"Fv" is the minimum antibody fragment that contains a complete antigen-recognition and antigen-binding site. This fragment contains a dimer of one heavy- and one light-chain variable region domain in tight, non-covalent association. From the folding of these two domains emanate six hypervariable loops (three loops each from the H and L chain) that contribute the amino acid residues for antigen binding and confer antigen binding specificity to the antibody. However, even a single variable region (or half of an Fv comprising only three CDRs specific for an antigen) has the ability to recognize and bind antigen, although at a lower affinity than the entire binding site.

"Single-chain Fv" ("sFv" or "scFv") are antibody fragments that comprise the VH and VL antibody domains connected into a single polypeptide chain. The sFv polypeptide can further comprise a polypeptide linker between the VH and VL domains that enables the sFv to form the desired structure for antigen binding. For a review of sFv, see, for example, Pluckthun in The Pharmacology of Monoclonal Antibodies, vol. 113, Rosenberg and Moore eds., Springer-Verlag, New York, pp. 269-315 (1994); Borrebaeck 1995, *infra*.

The term "diabodies" refers to small antibody fragments prepared by constructing sFv fragments with short linkers (about 5-10 residues) between the VH and VL domains such that

inter-chain but not intra-chain pairing of the V domains is achieved, resulting in a bivalent fragment, i.e., fragment having two antigen-binding sites. Bispecific diabodies are heterodimers of two "crossover" sFv fragments in which the VH and VL domains of the two antibodies are present on different polypeptide chains. Diabodies are described more fully in, for example, EP 5 404,097; WO 93/11161; and Hollinger *et al.*, Proc. Natl. Acad. Sci. USA, 90:6444-6448 (1993).

Domain antibodies (dAbs), which can be produced in fully human form, are the smallest known antigen-binding fragments of antibodies, ranging from about 11 kDa to about 15 kDa. DAbs are the robust variable regions of the heavy and light chains of immunoglobulins (VH and VL, respectively). They are highly expressed in microbial cell culture, show favorable 10 biophysical properties including, for example, but not limited to, solubility and temperature stability, and are well suited to selection and affinity maturation by in vitro selection systems such as, for example, phage display. DAbs are bioactive as monomers and, owing to their small size and inherent stability, can be formatted into larger molecules to create drugs with prolonged serum half-lives or other pharmacological activities. Examples of this technology have been 15 described in, for example, WO9425591 for antibodies derived from Camelidae heavy chain Ig, as well in US20030130496 describing the isolation of single domain fully human antibodies from phage libraries.

Fv and sFv are the only species with intact combining sites that are devoid of constant regions. Thus, they are suitable for reduced nonspecific binding during in vivo use. sFv fusion 20 proteins can be constructed to yield fusion of an effector protein at either the amino or the carboxy terminus of an sFv. See, for example, Antibody Engineering, ed. Borrebaeck, *supra*. The antibody fragment also can be a "linear antibody", for example, as described in U.S. Pat. No. 5,641,870 for example. Such linear antibody fragments can be monospecific or bispecific.

In certain embodiments, antibodies of the described invention are bispecific or multi- 25 specific. Bispecific antibodies are antibodies that have binding specificities for at least two different epitopes. Exemplary bispecific antibodies can bind to two different epitopes of a single antigen. Other such antibodies can combine a first antigen binding site with a binding site for a second antigen. Alternatively, an anti-HIV arm can be combined with an arm that binds to a triggering molecule on a leukocyte, such as a T-cell receptor molecule (for example, CD3), or Fc 30 receptors for IgG (Fc gamma R), such as Fc gamma RI (CD64), Fc gamma RII (CD32) and Fc gamma RIII (CD16), so as to focus and localize cellular defense mechanisms to the infected cell.

Bispecific antibodies also can be used to localize cytotoxic agents to infected cells. Bispecific antibodies can be prepared as full length antibodies or antibody fragments (for example, F(ab')₂ bispecific antibodies). For example, WO 96/16673 describes a bispecific anti-ErbB2/anti-Fc gamma RIII antibody and U.S. Pat. No. 5,837,234 discloses a bispecific anti-ErbB2/anti-Fc gamma RI antibody. For example, a bispecific anti-ErbB2/Fc alpha antibody is reported in WO98/02463; U.S. Pat. No. 5,821,337 teaches a bispecific anti-ErbB2/anti-CD3 antibody. See also, for example, Mouquet *et al.*, Polyreactivity Increases The Apparent Affinity Of Anti-HIV Antibodies By Heteroligation. *Nature*. 467, 591-5 (2010), and Mouquet *et al.*, Enhanced HIV-1 neutralization by antibody heteroligation” *Proc Natl Acad Sci U S A*. 2012 Jan 17;109(3):875-80.

Methods for making bispecific antibodies are known in the art. Traditional production of full length bispecific antibodies is based on the co-expression of two immunoglobulin heavy chain-light chain pairs, where the two chains have different specificities (see, for example, Millstein *et al.*, *Nature*, 305:537-539 (1983)). Similar procedures are disclosed in, for example, WO 93/08829, Traunecker *et al.*, *EMBO J.*, 10:3655-3659 (1991) and see also Mouquet *et al.*, Enhanced HIV-1 neutralization by antibody heteroligation” *Proc Natl Acad Sci U S A*. 2012 Jan 17;109(3):875-80.

Alternatively, antibody variable regions with the desired binding specificities (antibody-antigen combining sites) are fused to immunoglobulin constant domain sequences. The fusion is with an Ig heavy chain constant domain, comprising at least part of the hinge, CH₂, and CH₃ regions. According to some embodiments, the first heavy-chain constant region (CH₁) containing the site necessary for light chain bonding, is present in at least one of the fusions. DNAs encoding the immunoglobulin heavy chain fusions and, if desired, the immunoglobulin light chain, are inserted into separate expression vectors, and are co-transfected into a suitable host cell. This provides for greater flexibility in adjusting the mutual proportions of the three polypeptide fragments in embodiments when unequal ratios of the three polypeptide chains used in the construction provide the optimum yield of the desired bispecific antibody. It is, however, possible to insert the coding sequences for two or all three polypeptide chains into a single expression vector when the expression of at least two polypeptide chains in equal ratios results in high yields or when the ratios have no significant affect on the yield of the desired chain combination.

Techniques for generating bispecific antibodies from antibody fragments also have been described in the literature. For example, bispecific antibodies can be prepared using chemical

linkage. For example, Brennan *et al.*, Science, 229: 81 (1985) describe a procedure wherein intact antibodies are proteolytically cleaved to generate F(ab')₂ fragments. These fragments are reduced in the presence of the dithiol complexing agent, sodium arsenite, to stabilize vicinal dithiols and prevent intermolecular disulfide formation. The Fab' fragments generated then are converted to thionitrobenzoate (TNB) derivatives. One of the Fab'-TNB derivatives then is reconverted to the Fab'-thiol by reduction with mercaptoethylamine and is mixed with an equimolar amount of the other Fab'-TNB derivative to form the bispecific antibody. The bispecific antibodies produced can be used as agents for the selective immobilization of enzymes.

Other modifications of the antibody are contemplated herein. For example, the antibody can be linked to one of a variety of nonproteinaceous polymers, for example, polyethylene glycol, polypropylene glycol, polyoxyalkylenes, or copolymers of polyethylene glycol and polypropylene glycol. The antibody also can be entrapped in microcapsules prepared, for example, by coacervation techniques or by interfacial polymerization (for example, hydroxymethylcellulose or gelatin-microcapsules and poly-(methylmethacrylate) microcapsules, respectively), in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nanoparticles and nanocapsules), or in macroemulsions. Such techniques are disclosed in, for example, Remington's Pharmaceutical Sciences, 16th edition, Oslo, A., Ed., (1980).

Typically, the antibodies of the described invention are produced recombinantly, using vectors and methods available in the art. Human antibodies also can be generated by in vitro activated B cells (see, for example, U.S. Pat. Nos. 5,567,610 and 5,229,275). General methods in molecular genetics and genetic engineering useful in the present invention are described in the current editions of Molecular Cloning: A Laboratory Manual (Sambrook, *et al.*, 1989, Cold Spring Harbor Laboratory Press), Gene Expression Technology (Methods in Enzymology, Vol. 185, edited by D. Goeddel, 1991. Academic Press, San Diego, CA), "Guide to Protein Purification" in Methods in Enzymology (M.P. Deutscher, ed., (1990) Academic Press, Inc.); PCR Protocols: A Guide to Methods and Applications (Innis, *et al.* 1990. Academic Press, San Diego, CA), Culture of Animal Cells: A Manual of Basic Technique, 2nd Ed. (R.I. Freshney. 1987. Liss, Inc. New York, NY), and Gene Transfer and Expression Protocols, pp. 109-128, ed. E.J. Murray, The Humana Press Inc., Clifton, N.J.). Reagents, cloning vectors, and kits for genetic manipulation are available from commercial vendors such as BioRad, Stratagene, Invitrogen, ClonTech and Sigma-Aldrich Co.

Human antibodies also can be produced in transgenic animals (for example, mice) that are capable of producing a full repertoire of human antibodies in the absence of endogenous immunoglobulin production. For example, it has been described that the homozygous deletion of the antibody heavy-chain joining region (JH) gene in chimeric and germ-line mutant mice results in complete inhibition of endogenous antibody production. Transfer of the human germ-line immunoglobulin gene array into such germ-line mutant mice results in the production of human antibodies upon antigen challenge. See, for example, Jakobovits *et al.*, Proc. Natl. Acad. Sci. USA, 90:2551 (1993); Jakobovits *et al.*, Nature, 362:255-258 (1993); Bruggemann *et al.*, Year in Immuno., 7:33 (1993); U.S. Pat. Nos. 5,545,806, 5,569,825, 5,591,669 (all of GenPharm); U.S. Pat. No. 5,545,807; and WO 97/17852. Such animals can be genetically engineered to produce human antibodies comprising a polypeptide of the described invention.

Various techniques have been developed for the production of antibody fragments. Traditionally, these fragments were derived via proteolytic digestion of intact antibodies (see, for example, Morimoto *et al.*, Journal of Biochemical and Biophysical Methods 24:107-117 (1992); and Brennan *et al.*, Science, 229:81 (1985)). However, these fragments can now be produced directly by recombinant host cells. Fab, Fv and ScFv antibody fragments can all be expressed in and secreted from E. coli, thus allowing the facile production of large amounts of these fragments. Fab'-SH fragments can be directly recovered from E. coli and chemically coupled to form F(ab')₂ fragments (see, for example, Carter *et al.*, Bio/Technology 10:163-167 (1992)). According to another approach, F(ab')₂ fragments can be isolated directly from recombinant host cell culture. Fab and F(ab')₂ fragment with increased in vivo half-life comprising a salvage receptor binding epitope residues are described in U.S. Pat. No. 5,869,046. Other techniques for the production of antibody fragments will be apparent to the skilled practitioner.

Other techniques that are known in the art for the selection of antibody fragments from libraries using enrichment technologies, including but not limited to phage display, ribosome display (Hanes and Pluckthun, 1997, Proc. Nat. Acad. Sci. 94: 4937-4942), bacterial display (Georgiou, *et al.*, 1997, Nature Biotechnology 15: 29-34) and/or yeast display (Kieckhefer, *et al.*, 1997, Protein Engineering 10: 1303-1310) may be utilized as alternatives to previously discussed technologies to select single chain antibodies. Single-chain antibodies are selected from a library of single chain antibodies produced directly utilizing filamentous phage technology. Phage display technology is known in the art (e.g., see technology from Cambridge Antibody

Technology (CAT)) as disclosed in U.S. Patent Nos. 5,565,332; 5,733,743; 5,871,907; 5,872,215; 5,885,793; 5,962,255; 6,140,471; 6,225,447; 6,291,650; 6,492,160; 6,521,404; 6,544,731; 6,555,313; 6,582,915; 6,593, 081, as well as other U.S. family members, or applications which rely on priority filing GB 9206318, filed 24 May 1992; see also Vaughn, *et al.* 1996, *Nature Biotechnology* 14: 309-314). Single chain antibodies may also be designed and constructed using available recombinant DNA technology, such as a DNA amplification method (e.g., PCR), or possibly by using a respective hybridoma cDNA as a template.

Variant antibodies also are included within the scope of the invention. Thus, variants of the sequences recited in the application also are included within the scope of the invention. Further variants of the antibody sequences having improved affinity can be obtained using methods known in the art and are included within the scope of the invention. For example, amino acid substitutions can be used to obtain antibodies with further improved affinity. Alternatively, codon optimization of the nucleotide sequence can be used to improve the efficiency of translation in expression systems for the production of the antibody.

Such variant antibody sequences will share 70% or more (i.e., 80%, 85%, 90%, 95%, 97%, 98%, 99% or greater) sequence identity with the sequences recited in the application. Such sequence identity is calculated with regard to the full length of the reference sequence (i.e., the sequence recited in the application). Percentage identity, as referred to herein, is as determined using BLAST version 2.1.3 using the default parameters specified by the NCBI (the National Center for Biotechnology Information; www.ncbi.nlm.nih.gov/) [Blosum 62 matrix; gap open penalty=11 and gap extension penalty=1]. For example, peptide sequences are provided by this invention that comprise at least about 5, 10, 15, 20, 30, 40, 50, 75, 100, 150, or more contiguous peptides of one or more of the sequences disclosed herein as well as all intermediate lengths there between. As used herein, the term "intermediate lengths" is meant to describe any length between the quoted values, such as 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, etc.; 21, 22, 23, etc.; 30, 31, 32, etc.; 50, 51, 52, 53, etc.; 100, 101, 102, 103, etc.; 150, 151, 152, 153, etc.

The present invention provides for antibodies, either alone or in combination with other antibodies, such as, but not limited to, VRC01, anti-V3 loop, CD4bs, and CD4i antibodies as well as PG9/PG16-like antibodies, that have broad neutralizing activity in serum.

According to another embodiment, the present invention provides methods for the preparation and administration of an HIV antibody composition that is suitable for administration

to a human or non-human primate patient having HIV infection, or at risk of HIV infection, in an amount and according to a schedule sufficient to induce a protective immune response against HIV, or reduction of the HIV virus, in a human.

According to another embodiment, the present invention provides a vaccine comprising at least one antibody of the invention and a pharmaceutically acceptable carrier. According to one embodiment, the vaccine is a vaccine comprising at least one antibody described herein and a pharmaceutically acceptable carrier. The vaccine can include a plurality of the antibodies having the characteristics described herein in any combination and can further include antibodies neutralizing to HIV as are known in the art.

It is to be understood that compositions can be a single or a combination of antibodies disclosed herein, which can be the same or different, in order to prophylactically or therapeutically treat the progression of various subtypes of HIV infection after vaccination. Such combinations can be selected according to the desired immunity. When an antibody is administered to an animal or a human, it can be combined with one or more pharmaceutically acceptable carriers, excipients or adjuvants as are known to one of ordinary skilled in the art. The composition can further include broadly neutralizing antibodies known in the art, including but not limited to, VRC01, b12, anti-V3 loop, CD4bs, and CD4i antibodies as well as PG9/PG16-like antibodies.

Further, with respect to determining the effective level in a patient for treatment of HIV, in particular, suitable animal models are available and have been widely implemented for evaluating the in vivo efficacy against HIV of various gene therapy protocols (Sarver *et al.* (1993b), *supra*). These models include mice, monkeys and cats. Even though these animals are not naturally susceptible to HIV disease, chimeric mice models (for example, SCID, bg/nu/xid, NOD/SCID, SCID-hu, immunocompetent SCID-hu, bone marrow-ablated BALB/c) reconstituted with human peripheral blood mononuclear cells (PBMCs), lymph nodes, fetal liver/thymus or other tissues can be infected with lentiviral vector or HIV, and employed as models for HIV pathogenesis. Similarly, the simian immune deficiency virus (SIV)/monkey model can be employed, as can the feline immune deficiency virus (FIV)/cat model. The pharmaceutical composition can contain other pharmaceuticals, in conjunction with a vector according to the invention, when used to therapeutically treat AIDS. These other pharmaceuticals can be used in their traditional fashion (i.e., as agents to treat HIV infection).

According to another embodiment, the present invention provides an antibody-based pharmaceutical composition comprising an effective amount of an isolated HIV antibody, or an affinity matured version, which provides a prophylactic or therapeutic treatment choice to reduce infection of the HIV virus. The antibody-based pharmaceutical composition of the present invention may be formulated by any number of strategies known in the art (e.g., see McGoff and Scher, 2000, *Solution Formulation of Proteins/Peptides*: In McNally, E.J., ed. *Protein Formulation and Delivery*. New York, NY: Marcel Dekker; pp. 139-158; Akers and Defilippis, 2000, *Peptides and Proteins as Parenteral Solutions*. In: *Pharmaceutical Formulation Development of Peptides and Proteins*. Philadelphia, PA: Talyor and Francis; pp. 145-177; Akers, *et al.*, 2002, *Pharm. Biotechnol.* 14:47-127). A pharmaceutically acceptable composition suitable for patient administration will contain an effective amount of the antibody in a formulation which both retains biological activity while also promoting maximal stability during storage within an acceptable temperature range. The pharmaceutical compositions can also include, depending on the formulation desired, pharmaceutically acceptable diluents, pharmaceutically acceptable carriers and/or pharmaceutically acceptable excipients, or any such vehicle commonly used to formulate pharmaceutical compositions for animal or human administration. The diluent is selected so as not to affect the biological activity of the combination. Examples of such diluents are distilled water, physiological phosphate-buffered saline, Ringer's solutions, dextrose solution, and Hank's solution. The amount of an excipient that is useful in the pharmaceutical composition or formulation of this invention is an amount that serves to uniformly distribute the antibody throughout the composition so that it can be uniformly dispersed when it is to be delivered to a subject in need thereof. It may serve to dilute the antibody to a concentration which provides the desired beneficial palliative or curative results while at the same time minimizing any adverse side effects that might occur from too high a concentration. It may also have a preservative effect. Thus, for the antibody having a high physiological activity, more of the excipient will be employed. On the other hand, for any active ingredient(s) that exhibit a lower physiological activity, a lesser quantity of the excipient will be employed.

The above described antibodies and antibody compositions or vaccine compositions, comprising at least one or a combination of the antibodies described herein, can be administered for the prophylactic and therapeutic treatment of HIV viral infection.

The present invention also relates to isolated polypeptides comprising the novel amino acid sequences of the light chains and heavy chains, as well as the consensus sequences for the heavy and light chains of SEQ ID NOs: 1 and 2, as listed in Figure 3.

5 In other related embodiments, the invention provides polypeptide variants that encode the amino acid sequences of the HIV antibodies listed in Figure 3; the consensus sequences for the heavy and light chains of SEQ ID NOs: 1 and 2. These polypeptide variants have at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99%, or greater, sequence identity compared to a polypeptide sequence of this invention, as determined using the methods described herein, (for example, BLAST analysis using standard parameters). One skilled in this art will recognize that
10 these values can be appropriately adjusted to determine corresponding identity of proteins encoded by taking into amino acid similarity and the like.

The term "polypeptide" is used in its conventional meaning, i.e., as a sequence of amino acids. The polypeptides are not limited to a specific length of the product. Peptides, oligopeptides, and proteins are included within the definition of polypeptide, and such terms can
15 be used interchangeably herein unless specifically indicated otherwise. This term also includes post-expression modifications of the polypeptide, for example, glycosylations, acetylations, phosphorylations and the like, as well as other modifications known in the art, both naturally occurring and non-naturally occurring. A polypeptide can be an entire protein, or a subsequence thereof. Particular polypeptides of interest in the context of this invention are amino acid
20 subsequences comprising CDRs, VH and VL, being capable of binding an antigen or HIV-infected cell.

A polypeptide "variant," as the term is used herein, is a polypeptide that typically differs from a polypeptide specifically disclosed herein in one or more substitutions, deletions, additions and/or insertions. Such variants can be naturally occurring or can be synthetically generated, for
25 example, by modifying one or more of the above polypeptide sequences of the invention and evaluating one or more biological activities of the polypeptide as described herein and/or using any of a number of techniques well known in the art.

For example, certain amino acids can be substituted for other amino acids in a protein structure without appreciable loss of its ability to bind other polypeptides (for example, antigens)
30 or cells. Since it is the binding capacity and nature of a protein that defines that protein's biological functional activity, certain amino acid sequence substitutions can be made in a protein

sequence, and, accordingly, its underlying DNA coding sequence, whereby a protein with like properties is obtained. It is thus contemplated that various changes can be made in the peptide sequences of the disclosed compositions, or corresponding DNA sequences that encode said peptides without appreciable loss of their biological utility or activity.

5 In many instances, a polypeptide variant will contain one or more conservative substitutions. A "conservative substitution" is one in which an amino acid is substituted for another amino acid that has similar properties, such that one skilled in the art of peptide chemistry would expect the secondary structure and hydrophobic nature of the polypeptide to be substantially unchanged.

10 Amino acid substitutions generally are based on the relative similarity of the amino acid side-chain substituents, for example, their hydrophobicity, hydrophilicity, charge, size, and the like. Exemplary substitutions that take various of the foregoing characteristics into consideration are well known to those of skill in the art and include: arginine and lysine; glutamate and aspartate; serine and threonine; glutamine and asparagine; and valine, leucine and isoleucine.

15 "Homology" or "sequence identity" refers to the percentage of residues in the polynucleotide or polypeptide sequence variant that are identical to the non-variant sequence after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent homology. In particular embodiments, polynucleotide and polypeptide variants have at least about 70%, at least about 75%, at least about 80%, at least about 90%, at least about 95%, at least
20 about 98%, or at least about 99% polynucleotide or polypeptide homology with a polynucleotide or polypeptide described herein.

Such variant polypeptide sequences will share 70% or more (i.e. 80%, 85%, 90%, 95%, 97%, 98%, 99% or more) sequence identity with the sequences recited in the application. In additional embodiments, the described invention provides polypeptide fragments comprising
25 various lengths of contiguous stretches of amino acid sequences disclosed herein. For example, peptide sequences are provided by this invention that comprise at least about 5, 10, 15, 20, 30, 40, 50, 75, 100, 150, or more contiguous peptides of one or more of the sequences disclosed herein as well as all intermediate lengths there between.

The invention also includes nucleic acid sequences encoding part or all of the light and
30 heavy chains of the described inventive antibodies, and fragments thereof. Due to redundancy of

the genetic code, variants of these sequences will exist that encode the same amino acid sequences.

The present invention also includes isolated nucleic acid sequences encoding the polypeptides for the heavy and light chains of the HIV antibodies listed in Figure 3 and the consensus sequences for the heavy and light chains of SEQ ID NOs: 1 and 2.

In other related embodiments, the described invention provides polynucleotide variants that encode the peptide sequences of the heavy and light chains of the HIV antibodies listed in Figure 3; the consensus sequences for the heavy and light chains of SEQ ID NOs: 1 and 2. These polynucleotide variants have at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99%, or greater, sequence identity compared to a polynucleotide sequence of this invention, as determined using the methods described herein, (for example, BLAST analysis using standard parameters). One skilled in this art will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning, and the like.

The terms "nucleic acid" and "polynucleotide" are used interchangeably herein to refer to single-stranded or double-stranded RNA, DNA, or mixed polymers. Polynucleotides can include genomic sequences, extra-genomic and plasmid sequences, and smaller engineered gene segments that express, or can be adapted to express polypeptides.

An "isolated nucleic acid" is a nucleic acid that is substantially separated from other genome DNA sequences as well as proteins or complexes such as ribosomes and polymerases, which naturally accompany a native sequence. The term encompasses a nucleic acid sequence that has been removed from its naturally occurring environment, and includes recombinant or cloned DNA isolates and chemically synthesized analogues or analogues biologically synthesized by heterologous systems. A substantially pure nucleic acid includes isolated forms of the nucleic acid. Accordingly, this refers to the nucleic acid as originally isolated and does not exclude genes or sequences later added to the isolated nucleic acid by the hand of man.

A polynucleotide "variant," as the term is used herein, is a polynucleotide that typically differs from a polynucleotide specifically disclosed herein in one or more substitutions, deletions, additions and/or insertions. Such variants can be naturally occurring or can be synthetically generated, for example, by modifying one or more of the polynucleotide sequences of the

invention and evaluating one or more biological activities of the encoded polypeptide as described herein and/or using any of a number of techniques well known in the art.

Modifications can be made in the structure of the polynucleotides of the described invention and still obtain a functional molecule that encodes a variant or derivative polypeptide with desirable characteristics. When it is desired to alter the amino acid sequence of a polypeptide to create an equivalent, or even an improved, variant or portion of a polypeptide of the invention, one skilled in the art typically will change one or more of the codons of the encoding DNA sequence.

Typically, polynucleotide variants contain one or more substitutions, additions, deletions and/or insertions, such that the immunogenic binding properties of the polypeptide encoded by the variant polynucleotide is not substantially diminished relative to a polypeptide encoded by a polynucleotide sequence specifically set forth herein.

In additional embodiments, the described invention provides polynucleotide fragments comprising various lengths of contiguous stretches of sequence identical to or complementary to one or more of the sequences disclosed herein. For example, polynucleotides are provided by this invention that comprise at least about 10, 15, 20, 30, 40, 50, 75, 100, 150, 200, 300, 400, 500 or 1000 or more contiguous nucleotides of one or more of the sequences disclosed herein as well as all intermediate lengths there between and encompass any length between the quoted values, such as 16, 17, 18, 19, etc.; 21, 22, 23, etc.; 30, 31, 32, etc.; 50, 51, 52, 53, etc.; 100, 101, 102, 103, etc.; 150, 151, 152, 153, etc.; and including all integers through 200-500; 500-1,000.

In another embodiment of the invention, polynucleotide compositions are provided that are capable of hybridizing under moderate to high stringency conditions to a polynucleotide sequence provided herein, or a fragment thereof, or a complementary sequence thereof. Hybridization techniques are well known in the art of molecular biology. For purposes of illustration, suitable moderate stringent conditions for testing the hybridization of a polynucleotide of this invention with other polynucleotides include prewashing in a solution of 5x SSC, 0.5% SDS, 1.0 mM EDTA (pH 8.0); hybridizing at 50-60° C., 5x SSC, overnight; followed by washing twice at 65° C for 20 minutes with each of 2x, 0.5x, and 0.2x SSC containing 0.1% SDS. One skilled in the art will understand that the stringency of hybridization can be readily manipulated, such as by altering the salt content of the hybridization solution and/or the temperature at which the hybridization is performed. For example, in another embodiment, suitable highly stringent

hybridization conditions include those described above, with the exception that the temperature of hybridization is increased, for example, to 60-65 °C or 65-70 °C.

5 In some embodiments, the polypeptide encoded by the polynucleotide variant or fragment has the same binding specificity (i.e., specifically or preferentially binds to the same epitope or HIV strain) as the polypeptide encoded by the native polynucleotide. In some embodiments, the described polynucleotides, polynucleotide variants, fragments and hybridizing sequences, encode polypeptides that have a level of binding activity of at least about 50%, at least about 70%, and at least about 90% of that for a polypeptide sequence specifically set forth herein.

10 The polynucleotides of the described invention, or fragments thereof, regardless of the length of the coding sequence itself, can be combined with other DNA sequences, such as promoters, polyadenylation signals, additional restriction enzyme sites, multiple cloning sites, other coding segments, and the like, such that their overall length can vary considerably. A nucleic acid fragment of almost any length is employed. For example, illustrative polynucleotide segments with total lengths of about 10000, about 5000, about 3000, about 2000, about 1000, 15 about 500, about 200, about 100, about 50 base pairs in length, and the like, (including all intermediate lengths) are included in many implementations of this invention.

Further included within the scope of the invention are vectors such as expression vectors, comprising a nucleic acid sequence according to the invention. Cells transformed with such vectors also are included within the scope of the invention.

20 The present invention also provides vectors and host cells comprising a nucleic acid of the invention, as well as recombinant techniques for the production of a polypeptide of the invention. Vectors of the invention include those capable of replication in any type of cell or organism, including, for example, plasmids, phage, cosmids, and mini chromosomes. In some embodiments, vectors comprising a polynucleotide of the described invention are vectors suitable for propagation or replication of the polynucleotide, or vectors suitable for expressing a polypeptide 25 of the described invention. Such vectors are known in the art and commercially available.

"Vector" includes shuttle and expression vectors. Typically, the plasmid construct also will include an origin of replication (for example, the ColE1 origin of replication) and a selectable marker (for example, ampicillin or tetracycline resistance), for replication and selection, 30 respectively, of the plasmids in bacteria. An "expression vector" refers to a vector that contains

the necessary control sequences or regulatory elements for expression of the antibodies including antibody fragment of the invention, in bacterial or eukaryotic cells.

As used herein, the term “cell” can be any cell, including, but not limited to, that of a eukaryotic, multicellular species (for example, as opposed to a unicellular yeast cell), such as, but
5 not limited to, a mammalian cell or a human cell. A cell can be present as a single entity, or can be part of a larger collection of cells. Such a “larger collection of cells” can comprise, for example, a cell culture (either mixed or pure), a tissue (for example, endothelial, epithelial, mucosa or other tissue), an organ (for example, lung, liver, muscle and other organs), an organ
10 system (for example, circulatory system, respiratory system, gastrointestinal system, urinary system, nervous system, integumentary system or other organ system), or an organism (e.g., a bird, mammal, or the like).

Polynucleotides of the invention may be synthesized, whole or in parts that then are combined, and inserted into a vector using routine molecular and cell biology techniques, including, for example, subcloning the polynucleotide into a linearized vector using appropriate
15 restriction sites and restriction enzymes. Polynucleotides of the described invention are amplified by polymerase chain reaction using oligonucleotide primers complementary to each strand of the polynucleotide. These primers also include restriction enzyme cleavage sites to facilitate subcloning into a vector. The replicable vector components generally include, but are not limited to, one or more of the following: a signal sequence, an origin of replication, and one or more
20 marker or selectable genes.

In order to express a polypeptide of the invention, the nucleotide sequences encoding the polypeptide, or functional equivalents, may be inserted into an appropriate expression vector, i.e., a vector that contains the necessary elements for the transcription and translation of the inserted coding sequence. Methods well known to those skilled in the art may be used to construct
25 expression vectors containing sequences encoding a polypeptide of interest and appropriate transcriptional and translational control elements. These methods include in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination. Such techniques are described, for example, in Sambrook, J., *et al.* (1989) *Molecular Cloning, A Laboratory Manual*, Cold Spring Harbor Press, Plainview, N.Y., and Ausubel, F. M. *et al.* (1989) *Current Protocols in*
30 *Molecular Biology*, John Wiley & Sons, New York, N.Y.

The present invention also provides kits useful in performing diagnostic and prognostic assays using the antibodies, polypeptides and nucleic acids of the present invention. Kits of the present invention include a suitable container comprising an HIV antibody, a polypeptide or a nucleic acid of the invention in either labeled or unlabeled form. In addition, when the antibody, polypeptide or nucleic acid is supplied in a labeled form suitable for an indirect binding assay, the kit further includes reagents for performing the appropriate indirect assay. For example, the kit may include one or more suitable containers including enzyme substrates or derivatizing agents, depending on the nature of the label. Control samples and/or instructions may also be included. The present invention also provides kits for detecting the presence of the HIV antibodies or the nucleotide sequence of the HIV antibody of the present invention in a biological sample by PCR or mass spectrometry.

"Label" as used herein refers to a detectable compound or composition that is conjugated directly or indirectly to the antibody so as to generate a "labeled" antibody. A label can also be conjugated to a polypeptide and/or a nucleic acid sequence disclosed herein. The label can be detectable by itself (for example, radioisotope labels or fluorescent labels) or, in the case of an enzymatic label, can catalyze chemical alteration of a substrate compound or composition that is detectable. Antibodies and polypeptides of the described invention also can be modified to include an epitope tag or label, for example, for use in purification or diagnostic applications. Suitable detection means include the use of labels such as, but not limited to, radionucleotides, enzymes, coenzymes, fluorescers, chemiluminescers, chromogens, enzyme substrates or co-factors, enzyme inhibitors, prosthetic group complexes, free radicals, particles, dyes, and the like.

According to another embodiment, the present invention provides diagnostic methods. Diagnostic methods generally involve contacting a biological sample obtained from a patient, such as, for example, blood, serum, saliva, urine, sputum, a cell swab sample, or a tissue biopsy, with an HIV antibody and determining whether the antibody preferentially binds to the sample as compared to a control sample or predetermined cut-off value, thereby indicating the presence of the HIV virus.

According to another embodiment, the present invention provides methods to detect the presence of the HIV antibodies of the present invention in a biological sample from a patient. Detection methods generally involve obtaining a biological sample from a patient, such as, for example, blood, serum, saliva, urine, sputum, a cell swab sample, or a tissue biopsy and isolating

HIV antibodies or fragments thereof, or the nucleic acids that encode an HIV antibody, and assaying for the presence of an HIV antibody in the biological sample. Also, the present invention provides methods to detect the nucleotide sequence of an HIV antibody in a cell. The nucleotide sequence of an HIV antibody may also be detected using the primers disclosed herein. The presence of the HIV antibody in a biological sample from a patient may be determined utilizing known recombinant techniques and/or the use of a mass spectrometer.

In another embodiment, the present invention provides a method for detecting an HIV antibody comprising a heavy chain comprising a highly conserved consensus sequence and a light chain comprising a highly conserved consensus sequence in a biological sample, comprising obtaining an immunoglobulin-containing biological sample from a mammalian subject, isolating an HIV antibody from said sample, and identifying the highly conserved consensus sequences of the heavy chain and the light chain. The biological sample may be blood, serum, saliva, urine, sputum, a cell swab sample, or a tissue biopsy. The amino acid sequences may be determined by methods known in the art including, for example, PCR and mass spectrometry.

The term "assessing" includes any form of measurement, and includes determining if an element is present or not. The terms "determining", "measuring", "evaluating", "assessing" and "assaying" are used interchangeably and include quantitative and qualitative determinations. Assessing may be relative or absolute. "Assessing the presence of" includes determining the amount of something present, and/or determining whether it is present or absent. As used herein, the terms "determining," "measuring," and "assessing," and "assaying" are used interchangeably and include both quantitative and qualitative determinations.

II. Method of Reducing Viral Replication

Methods for reducing an increase in HIV virus titer, virus replication, virus proliferation or an amount of an HIV viral protein in a subject are further provided. According to another aspect, a method includes administering to the subject an amount of an HIV antibody effective to reduce an increase in HIV titer, virus replication or an amount of an HIV protein of one or more HIV strains or isolates in the subject.

According to another embodiment, the present invention provides a method of reducing viral replication or spread of HIV infection to additional host cells or tissues comprising contacting a mammalian cell with the antibody, or a portion thereof, which binds to an antigenic epitope on gp120.

III. Method of Treatment

According to another embodiment, the present invention provides a method for treating a mammal infected with a virus infection, such as, for example, HIV, comprising administering to said mammal a pharmaceutical composition comprising the HIV antibodies disclosed herein.

5 According to one embodiment, the method for treating a mammal infected with HIV comprises administering to said mammal a pharmaceutical composition that comprises an antibody of the present invention, or a fragment thereof. The compositions of the invention can include more than one antibody having the characteristics disclosed (for example, a plurality or pool of antibodies). It also can include other HIV neutralizing antibodies as are known in the art, for example, but not
10 limited to, VRC01, PG9 and b12.

Passive immunization has proven to be an effective and safe strategy for the prevention and treatment of viral diseases. (See, for example, Keller *et al.*, Clin. Microbiol. Rev. 13:602-14 (2000); Casadevall, Nat. Biotechnol. 20:114 (2002); Shibata *et al.*, Nat. Med. 5:204-10 (1999); and Igarashi *et al.*, Nat. Med. 5:211-16 (1999). Passive immunization using human monoclonal
15 antibodies provides an immediate treatment strategy for emergency prophylaxis and treatment of HIV.

Subjects at risk for HIV-related diseases or disorders include patients who have come into contact with an infected person or who have been exposed to HIV in some other way. Administration of a prophylactic agent can occur prior to the manifestation of symptoms
20 characteristic of HIV-related disease or disorder, such that a disease or disorder is prevented or, alternatively, delayed in its progression.

For in vivo treatment of human and non-human patients, the patient is administered or provided a pharmaceutical formulation including an HIV antibody of the invention. When used for in vivo therapy, the antibodies of the invention are administered to the patient in
25 therapeutically effective amounts (i.e., amounts that eliminate or reduce the patient's viral burden). The antibodies are administered to a human patient, in accord with known methods, such as intravenous administration, for example, as a bolus or by continuous infusion over a period of time, by intramuscular, intraperitoneal, intracerebrospinal, subcutaneous, intra-articular, intrasynovial, intrathecal, oral, topical, or inhalation routes. The antibodies can be administered
30 parenterally, when possible, at the target cell site, or intravenously. In some embodiments, antibody is administered by intravenous or subcutaneous administration. Therapeutic

compositions of the invention may be administered to a patient or subject systemically, parenterally, or locally. The above parameters for assessing successful treatment and improvement in the disease are readily measurable by routine procedures familiar to a physician.

For parenteral administration, the antibodies may be formulated in a unit dosage injectable form (solution, suspension, emulsion) in association with a pharmaceutically acceptable, parenteral vehicle. Examples of such vehicles include, but are not limited, water, saline, Ringer's solution, dextrose solution, and 5% human serum albumin. Nonaqueous vehicles include, but are not limited to, fixed oils and ethyl oleate. Liposomes can be used as carriers. The vehicle may contain minor amounts of additives such as substances that enhance isotonicity and chemical stability, such as, for example, buffers and preservatives. The antibodies can be formulated in such vehicles at concentrations of about 1 mg/ml to 10 mg/ml.

The dose and dosage regimen depends upon a variety of factors readily determined by a physician, such as the nature of the infection, for example, its therapeutic index, the patient, and the patient's history. Generally, a therapeutically effective amount of an antibody is administered to a patient. In some embodiments, the amount of antibody administered is in the range of about 0.1 mg/kg to about 50 mg/kg of patient body weight. Depending on the type and severity of the infection, about 0.1 mg/kg to about 50 mg/kg body weight (for example, about 0.1-15 mg/kg/dose) of antibody is an initial candidate dosage for administration to the patient, whether, for example, by one or more separate administrations, or by continuous infusion. The progress of this therapy is readily monitored by conventional methods and assays and based on criteria known to the physician or other persons of skill in the art. The above parameters for assessing successful treatment and improvement in the disease are readily measurable by routine procedures familiar to a physician.

Other therapeutic regimens may be combined with the administration of the HIV antibody of the present invention. The combined administration includes co-administration, using separate formulations or a single pharmaceutical formulation, and consecutive administration in either order, wherein preferably there is a time period while both (or all) active agents simultaneously exert their biological activities. Such combined therapy can result in a synergistic therapeutic effect. The above parameters for assessing successful treatment and improvement in the disease are readily measurable by routine procedures familiar to a physician.

The terms "treating" or "treatment" or "alleviation" are used interchangeably and refer to both therapeutic treatment and prophylactic or preventative measures; wherein the object is to prevent or slow down (lessen) the targeted pathologic condition or disorder. Those in need of treatment include those already with the disorder as well as those prone to have the disorder or those in whom the disorder is to be prevented. A subject or mammal is successfully "treated" for an infection if, after receiving a therapeutic amount of an antibody according to the methods of the present invention, the patient shows observable and/or measurable reduction in or absence of one or more of the following: reduction in the number of infected cells or absence of the infected cells; reduction in the percent of total cells that are infected; and/or relief to some extent, one or more of the symptoms associated with the specific infection; reduced morbidity and mortality, and improvement in quality of life issues. The above parameters for assessing successful treatment and improvement in the disease are readily measurable by routine procedures familiar to a physician.

The term "therapeutically effective amount" refers to an amount of an antibody or a drug effective to treat a disease or disorder in a subject or mammal.

Administration "in combination with" one or more further therapeutic agents includes simultaneous (concurrent) and consecutive administration in any order.

"Carriers" as used herein include pharmaceutically acceptable carriers, excipients, or stabilizers that are nontoxic to the cell or mammal being exposed thereto at the dosages and concentrations employed. Often the physiologically acceptable carrier is an aqueous pH buffered solution. Examples of physiologically acceptable carriers include, but not limited to, buffers such as phosphate, citrate, and other organic acids; antioxidants including, but not limited to, ascorbic acid; low molecular weight (less than about 10 residues) polypeptide; proteins, such as, but not limited to, serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as, but not limited to, polyvinylpyrrolidone; amino acids such as, but not limited to, glycine, glutamine, asparagine, arginine or lysine; monosaccharides, disaccharides, and other carbohydrates including, but not limited to, glucose, mannose, or dextrins; chelating agents such as, but not limited to, EDTA; sugar alcohols such as, but not limited to, mannitol or sorbitol; salt-forming counterions such as, but not limited to, sodium; and/or nonionic surfactants such as, but not limited to, TWEEN.; polyethylene glycol (PEG), and PLURONICS.

Where a value of ranges is provided, it is understood that each intervening value, to the tenth of the unit of the lower limit unless the context clearly dictates otherwise, between the upper and lower limit of that range and any other stated or intervening value in that stated range is encompassed within the invention. The upper and lower limits of these smaller ranges which may independently be included in the smaller ranges is also encompassed within the invention, subject to any specifically excluded limit in the stated range. Where the stated range includes one or both of the limits, ranges excluding either both of those included limits are also included in the invention.

EXAMPLE 1

This example describes materials and methods used in EXAMPLES 2-5 below.

HIV antibodies were cloned and produced following gp140-specific single B-cell capture as previously described (Mouquet, H. *et al.* PLoS One 6, e24078 (2011); Tiller, T. *et al.* J Immunol Methods 329, 112-24 (2008); and Scheid, J.F. *et al.* Nature 458, 636-40 (2009)). PGT121_{GM} and 10-1074_{GM} “glycomutant” antibodies were generated by substituting 10-1074 residues at HC positions 32, 53, 54, 58, 97, 100 into PGT121 and *vice versa*. Binding properties of anti-gp140 antibodies to HIV Env proteins were assayed by ELISA, SPR and glycan microarray assays as previously described (Scheid, J.F. *et al.* Science 333, 1633-7 (2011); Walker, L.M. *et al.* Nature 477, 466-70 (2011); and Mouquet, H. *et al.* PLoS One 6, e24078 (2011)). Neutralization was evaluated using (i) a luciferase-based assay in TZM.bl cells, and (ii) a PBMC-based assay using infection with primary HIV-1 variants as previously described (Li, M. *et al.* J Virol 79, 10108-25 (2005); Euler, Z. *et al.* Journal of virology 85, 7236-45 (2011); and Bunnik, E.M. *et al.* Nature medicine 16, 995-7 (2010)). Structures of PGT121 (“unliganded” and “liganded”), 10-1074 and GL Fab fragments were solved by molecular replacement to 2.8 Å, 2.3 Å, 1.8 Å and 2.4 Å resolution, respectively.

Single B cell RT-PCRs and Ig gene analyses

Single-cell sorting of gp140⁺CD19⁺IgG⁺ B cells from patient 10 (pt10; referred to as patient 17 in Nature 477(7365):466-470.) PBMCs, cDNA synthesis and nested PCR amplifications of Ig genes were performed in a previous study (PLoS One 6(9):e24078). Igλ genes expressed by PGT121 clonal variants were PCR amplified using a forward primer (L-Vλ3-21*02: 5' CTGGACCGTTCTCCTCCTCG 3') further upstream in the leader region to avoid the potentially mutated region (31). All PCR products were sequenced and analyzed for Ig gene

usage, CDR3 analyses and number of V_H/V_K somatic hypermutations (IgBLAST; <http://www.ncbi.nlm.nih.gov/igblast> and IMGT®; <http://www.imgt.org>). Multiple sequence alignments were performed using the MacVector program (v.12.5.0) with the ClustalW analysis function (default parameters), and were used to generate dendrograms by the Neighbor Joining method (with Best tree mode and outgroup rooting). Alternatively, dendrograms were generated using the UPGMA method (with Best tree mode).

The germline (GL) precursor gene segments of the PGT121-like and 10-1074-like antibodies were identified using IgBLAST (<http://www.ncbi.nlm.nih.gov/igblast>) and IMGT®/V-QUEST (http://www.imgt.org/IMGT_vquest/share/textes/) as V_H4-59*01, J_H6*03, V_L3-21*02 and J_L3*02. (These gene segments are among the most frequently used in the repertoire of human antibodies (PLoS One 6(8):e22365; Immunogenetics 64(5):337-350). To build a representative GL ancestor sequence, we aligned the IgH and IgL sequences of 10-996 (the antibody containing the fewest somatic hypermutations) to the GL sequences using IgBLAST (<http://www.ncbi.nlm.nih.gov/igblast>). The GL IgH sequence was constructed by replacing the mature V_H and J_H gene segments with their GL counterparts and using the 10-996 sequence for the CDRH3 region involving N-region nucleotides and the D_H gene segment. The GL IgL sequence was assembled from the V_L3-21*02 and J_L3*02 gene segment sequences.

Cloning and production of antibodies

Purified digested PCR products were cloned into human Igγ₁-, or Igλ-expressing vectors (J Immunol Methods 329(1-2):112-124). Vectors containing IgH and Igλ genes were then sequenced and compared to the original PCR product sequences. PGT121 and 10-303 shared the same Igλ gene and had one amino acid difference in position 2 of the IgH gene (**Fig. 4**); therefore to produce the PGT121 IgG, we used the 10-303 Igλ gene and a PGT121 IgH gene generated by introducing a single substitution (V2M) into the 10-303 IgH gene by site-directed mutagenesis (QuikChange Site-Directed Mutagenesis Kit; Stratagene). To generate His-tagged Fabs, the PGT121 and 10-1074 V_H genes were subcloned into a 6xHis-IgCγ1 expression vector generated by modifying our standard Igγ₁ vector (Science 301(5638):1374-1377) to encode the IgG1 C_H1 domain followed by a 6x-His tag. IgH DNA fragments encoding PGT121_{GM} (S32Y, K53D, S54R, N58T, H97R, T100IY) and 10-1074_{GM} (Y32S, D53K, R54S, T58N, R97H, Y100IT) mutant antibodies were obtained as a synthetic minigene (IDT) and subcloned into Igγ₁-expressing vectors.

Listed below is the heavy chain sequence for 10-1074_{GM} where the mutations are underlined. The light chain sequence of 10-1074_{GM} is the same as that of 10-1074.

QVQLQESGPGGLVKPSETLSVTCSVSGDSMNNSYWTWIRQSPGKGLEWIGYISKSESANYNPSLNS
RVVISRDTSKNQLSLKLNSVTPADTAVYYCATARHGQRIYGVVSFGEFFTYYSM DVWGKGTTVTV
SS

Antibodies and Fab fragments were produced by transient transfection of IgH and IgL expression plasmids into exponentially growing HEK 293T cells (ATCC, CRL-11268) using the polyethyleneimine (PEI)-precipitation method (PLoS One 6(9):e24078). IgG antibodies were affinity purified using Protein G sepharose beads (GE Healthcare) according to the manufacturer's instructions. Fab fragments were affinity purified using HisPur™ Cobalt Resin (Thermo scientific) as described below.

HIV-1 Env proteins

Alanine mutations were introduced into the pYU-2 gp120 vector (gift of J. Sodroski, Harvard Medical School) at positions 301 to 303 (Asn-Asn-Thr), 324 to 325 (Gly-Asp), and 332 (Asn) (HXBc2 amino acid numbering) using the QuikChange Site-Directed Mutagenesis kit (Stratagene) according to the manufacturer's instructions. The same procedure was used to generate "double glycan" mutants by introducing single alanine mutations in the pYU-2 gp120^{N332A} vector at each PNGS located between Asn262_{gp120} and Asn406_{gp120}. Site-directed mutations were verified by DNA sequencing.

Expression vectors encoding YU-2 gp140 (Journal of virology 74(12):5716-5725), YU-2 gp120, HXB2c gp120^{core} (Nature 393(6686):648-659), HXB2c 2CCcore (PLoS Pathog 5(5):e1000445) proteins, and YU-2 gp120 mutant proteins were used to transfect HEK 293T cells. To produce high-mannose-only YU-2 gp120 protein (gp120_{kif}), 25 μM kifunensine (Enzo Life Sciences) was added at the time of transfection. Culture supernatants were harvested and concentrated using centrifugation-based filtration devices (Vivacell 100, Sartorius Stedim Biotech GmbH) that allowed buffer exchange of the samples into 10 mM imidazole, 50 mM sodium phosphate, 300 mM sodium chloride; pH 7.4. Proteins were purified by affinity chromatography using HisPur™ Cobalt Resin (Thermo scientific) according to the manufacturer's instructions.

For deglycosylation reactions, 50 μg of HEK 293T cell-produced YU-2 gp120 in PBS was digested overnight at 37°C with 200 U of PNGase F (New England Biolabs) or 10,000 U of Endo H_f (New England Biolabs) in their respective reaction buffers without denaturing agents. After

buffer exchange into PBS using Centrifugal Filters (Amicon® Ultra, Millipore), glycosidase-treated gp120s (200 ng) were examined by SDS-PAGE using a 4-12% NuPAGE gel (Invitrogen) followed by silver staining (Pierce Silver Stain Kit, Thermo Scientific).

ELISAs

5 High-binding 96-well ELISA plates (Costar) were coated overnight with 100 ng/well of purified gp120 in PBS. After washing, the plates were blocked for 2 h with 2% BSA, 1 μM EDTA, 0.05% Tween-PBS (blocking buffer) and then incubated for 2 h with IgGs at concentrations of 26.7 nM (or 427.2 nM for ELISAs using the YU-2 gp120 double glycan mutants) and 7 consecutive 1:4 dilutions in PBS. After washing, the plates were developed by incubation with
10 goat HRP-conjugated anti-human IgG antibodies (Jackson ImmunoResearch) (at 0.8 μg/ml in blocking buffer) for 1 h, and by addition of HRP chromogenic substrate (ABTS solution, Invitrogen) (PLoS One 6(9):e24078). Antibody binding to the selected gp120^{V3} overlapping peptides was tested using a previously described peptide-ELISA method.

For competition ELISAs, gp120-coated plates were blocked for 2 h with blocking buffer
15 and then incubated for 2 h with biotinylated antibodies (at a concentration of 26.6 nM for PGT121, 0.21 nM for 10-1074, 0.43 nM for 10-996 and 1.67 nM for 10-1369) in 1:2 serially diluted solutions of antibody competitors in PBS (IgG concentration range from 5.2 to 667 nM). Plates were developed as described above using HRP-conjugated streptavidin (Jackson ImmunoResearch) (at 0.8 μg/ml in blocking buffer). All experiments were performed at least in
20 duplicate.

Glycan microarray analysis

Microarrays were generated by robotically printing glycan probes linked to lipid (neoglycolipids) onto nitrocellulose-coated glass slides (Methods Mol Biol 808:117-136) at two levels (2 and 5 fmol/spot) in duplicate. Binding assays were performed with microarrays
25 containing 15 neoglycolipids derived from *N*-glycans of high-mannose and complex-types. The sequences of the probes are shown in **Fig. 7A**. In brief, antibodies were tested at 50 μg/ml, and binding was detected with biotinylated anti-human IgG (Vector) followed by AlexaFluor 647-labeled streptavidin (Molecular Probes).

Surface plasmon resonance

30 Experiments were performed using a Biacore T100 (Biacore, Inc)(Nature 467(7315):591-595). Briefly, YU-2 gp140 and gp120 proteins were primary amine-coupled on CM5 chips

(Biacore, Inc.) at a coupling density of 300 RUs. Anti-gp120 IgGs and the germline precursor (GL) were injected over flow cells at 1 μ M and 10 μ M, respectively, at flow rates of 35 μ l/min with 3 min association and 5 min dissociation phases. The sensor surface was regenerated by a 30 sec injection of 10 mM glycine-HCl pH 2.5 at a flow rate of 50 μ l/min. Dissociation (k_d (s^{-1})), association (k_a ($M^{-1} s^{-1}$)) and binding constants (K_D (M) or K_A (M^{-1})) were calculated from kinetic analyses after subtraction of backgrounds using a 1:1 binding model without a bulk reflective index (RI) correction (Biacore T100 Evaluation software). Binding constants for bivalent IgGs calculated using a 1:1 binding model are referred to in the text as “apparent” affinities to emphasize that the K_D values include potential avidity effects

Neutralization assays

Virus neutralization was evaluated using a luciferase-based assay in TZM.bl cells (J Virol 79(16):10108-10125). The HIV-1 pseudoviruses tested contained mostly tier-2 and tier-3 viruses (Journal of virology 84(3):1439-1452)(**Tables 4 and 5**). High-mannose-only pseudoviruses were produced in wild-type cells treated with 25 μ M kifunensine (Enzo Life Sciences) (**Fig. 8C**) or in HEK 293S GnTI^{-/-} cells (**Fig. 8D**). Non-linear regression analysis was used to calculate concentrations at which half-maximal inhibition was observed (IC_{50} values). Neutralization activities were also evaluated with a previously characterized PBMC-based assay using infection with primary HIV-1 variants (n=95) isolated from clade B-infected donors with known seroconversion dates either between 1985 and 1989 (“historical seroconverters”, n=14) or between 2003 and 2006 (“contemporary seroconverters”, n=21)(Journal of virology 85(14):7236-7245; Nat Med 16(9):995-997). Neutralization activity for each antibody was calculated using GraphPad Prism software (v5.0b) as area under the best-fit curve, which fits the proportion of viruses neutralized over IC_{50} values ranging from 0.001 to 50 μ g/ml. Relative area under the curve (RAUC) values were derived by normalizing all AUC values by the highest value (obtained with 10-1074).

Statistical analyses

Statistical analyses were performed with the GraphPad Prism software (v5.0b). Neutralization potencies in the TZM-bl assay against the selected panel of 9 virus strains versus the apparent binding affinities of the antibodies for gp120 and gp140 were analyzed using a Spearman’s correlation test. The Mann Whitney test was used to compare: (i) affinities for

gp120/gp140 of antibodies belonging to the PGT121 or 10-1074 group, and (ii) neutralization activities against viruses isolated from historical and contemporary seroconverters.

Crystallization and structure determinations

6x-His tagged PGT121, 10-1074 and 10-996GL Fabs for crystallization were expressed. Fabs were purified from the supernatants of transiently-transfected HEK 293-6E cells by sequential Ni^{2+} -NTA affinity (Qiagen) and Superdex200 10/300 (GE Healthcare) size exclusion chromatography. For crystals of the unliganded PGT121 Fab, PGT121 IgG was isolated from the supernatants of transiently-transfected HEK 293-6E cells by Protein A affinity chromatography (Pierce), and Fab fragments were obtained by papain cleavage of the IgG and further purification using Superdex200 10/300 (GE Healthcare) size exclusion chromatography.

Purified Fabs were concentrated to 8-20 mg/mL (“unliganded” PGT121, 8 mg/mL; 10-1074 and GL, 20 mg/mL) in PBS buffer. The “liganded” PGT121 Fab crystals were prepared from a protein sample (final concentration: 15 mg/mL) that was mixed with a 3-fold molar excess of NA2 glycan and incubated at 20°C for 2 hours. Crystallization conditions were screened at 20°C using a Mosquito® crystallization robot (TTP labs) in 400 nL drops using a 1:1 protein to reservoir ratio. Crystals of “unliganded” PGT121 Fab ($\text{P2}_1\text{2}_1\text{2}_1$; $a = 56.8$, $b = 74.7$, $c = 114.9$ Å) were obtained in 24% PEG 4,000, 0.1 M Tris-HCl pH 8.5, 10 mM CuCl_2 and crystals of “liganded” PGT121 Fab ($\text{P2}_1\text{2}_1\text{2}_1$; $a = 67.8$, $b = 67.8$, $c = 94.1$ Å) grew in 17% PEG 10,000, 0.1M Bis-Tris pH 5.5, 0.1M $\text{CH}_3\text{COOHNH}_4$. Crystals of 10-1074 Fab (P2_1 ; $a = 61.4$, $b = 40.3$, $c = 84.5$ Å; $\beta = 95.39^\circ$) were obtained in 25% PEG 3,350, 0.1 M Bis-Tris pH 5.5, 0.2 M NaCl, and crystals of GL Fab (P2_1 ; $a = 54.9$, $b = 344.7$, $c = 55.2$ Å; $\beta = 91.95^\circ$) grew in 20% PEG 3,350, 0.24 M sodium malonate pH 7.0, 10 mM MnCl_2 . Crystals were cryoprotected by soaking in mother liquor containing 20% glycerol (“unliganded” and “liganded” PGT121 Fab) or 20% ethylene glycol (10-1074 Fab and GL Fab) and subsequently flash-cooled in liquid nitrogen.

Diffraction data were collected at beamline 12-2 (wavelength = 1.029 Å) at the Stanford Synchrotron Radiation Lightsource (SSRL) on a Pilatus 6M pixel detector (Dectris). Data were indexed, integrated and scaled using XDS. Using the data obtained from the “unliganded” PGT121 Fab crystals, we used Phenix to find a molecular replacement solution for one Fab per asymmetric unit (chains H and L for the heavy and light chain, respectively) using two search models, the C_H – C_L domains of PGT128 Fab (PDB code 3PV3) and the V_H – V_L domains of 2F5 (PDB code 3IDJ) after omitting residues in the CDRH3 and CDRL3 loops. Subsequently, we used

the “unliganded” PGT121 structure as a search model to find molecular replacement solutions for “liganded” PGT121 Fab (one Fab per asymmetric unit), 10-1074 Fab (one Fab per asymmetric unit) and GL (four Fabs per asymmetric unit).

Iterative refinement (including non-crystallographic symmetry restraints for GL) was performed using Phenix and manually fitting models into electron density maps using Coot. The atomic models were refined to 3.0 Å resolution for PGT121 Fab ($R_{\text{work}} = 21.6\%$; $R_{\text{free}} = 26.4\%$), 1.9 Å resolution for 10-1074 Fab ($R_{\text{work}} = 18.7\%$; $R_{\text{free}} = 22.3\%$), 2.4 Å resolution for four GL Fab molecules ($R_{\text{work}} = 19.4\%$; $R_{\text{free}} = 23.7\%$), and 2.4 Å resolution for “liganded” PGT121 Fab ($R_{\text{work}} = 20.1\%$; $R_{\text{free}} = 24.9\%$). The atomic model of PGT121 Fab contains 95.2%, 4.9% and 0.0% of the residues in the favored, allowed and disallowed regions of the Ramachandran plot, respectively (10-1074 Fab: 98.8%, 0.9%, 0.2%; GL Fab: 96.0%, 3.8%, 0.23%; “liganded PGT121 Fab: 96.7%, 3.1%, 0.2%). PyMOL was used for molecular visualization and to generate figures of the Fab structures. Buried surface area calculations were performed with Areaimol (CCP4 Suite) using a 1.4 Å probe.

Fab structures were aligned using the Super script in PyMOL. Pairwise Cα alignments were performed using PDBeFold.

EXAMPLE 2 *Predominance and diversity of PGT121 clonotype*

gp140-specific IgG memory B cells were isolated from a clade A-infected African donor using YU-2 gp140 trimers as “bait.” Eighty-seven matching immunoglobulin heavy (IgH) and light (IgL) chain genes corresponding to 23 unique clonal families were identified. The IgH anti-gp140 repertoire was dominated by one clonal family representing ~28% of all expanded B cell clones. This B cell family corresponds to the same clone as PGT121-123 (Nature 477(7365):466-470) and contained 38 members, 29 of which were unique variants at the nucleotide level (**Table 3**). Based on their IgH nucleotide sequence, the PGT121 family divides into two groups: a PGT121-like group containing PGT121-123 and 9 closely-related variants, and a second group, 10-1074-like, containing 20 members. Although our traditional primers (J Immunol Methods 329(1-2):112-124; Science 301(5638):1374-1377) did not amplify the IgL genes expressed by the PGT121 B cell clone due to the nucleotide deletions in the region encoding framework region 1, 24 of 38 Igλ genes were obtained using new Igλ-specific primers designed to amplify heavily somatically-mutated genes (**Table 3**). Consistent with the high levels of hypermutation in the IgH genes (18.2% of the VH gene on average), the amplified Igλ genes were highly mutated (18.2% of

the V λ gene on average) and carried nucleotide deletions in framework region 1 (FWR1) (12 to 21 nucleotides) and a 9-nucleotide insertion in framework region 3 (FWR3) (**Fig. 3B** and **Table 3**).

The sequence alignments of three PGT antibodies (PGT-121, -122, and -123), eleven PGT121 and 10-1074 clonal variants (10-259, 10-303, 10-410, 10-847, 10-996, 10-1074, 10-1121, 5 10-1130, 10-1146, 10-1341, 10-1369, and 10-1074GM), likely germline (GL), and consensus sequences are shown in Figures 3(a) and 3(b). The sequences for corresponding heavy chain variable regions, light chain variable regions, heavy chain CDRs, and light chain CDRs under both IMGT and KABT systems are listed in Table 1 below. Assigned sequence identification numbers for the sequences under the KABT systems are listed in Table 2 below:

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Table 1
IgH SEQUENCES

IMGT	FWR1	CDR1	FWR2	CDR2	FWR3	CDR3	FWR4
10-1369	QVQLQESGPGLVKPLETTLSTCN VS	GAFIADHY	WSWIRLPIGKG PEWIGY	VHDSGDI	NYNP SLKNRVHLSLDKSTNQVSLKLM AVTAGDSALYYC	ATTKHGRR IYG VVAFGE WFTYFYMDV	WGRGTTVTVSS
10-259	QVHLQESGPGLVKPPSETTLSTCN VS	GTLVRDNY	WSWMRQPLGKQ PEWIGY	VHDSGDT	NYNP SLKSRVHLSLDKSNNLVSLRLT AVTAADSATYYC	ATTKHGRR IYG I VAFNE WFTYFYMDV	WGKGT TTVTVSS
10-303	QVQLQESGPGLVKPPSETTLSTCS VS	GASISDSY	WSWIRRSPGKG LEWIGY	VHKSGDT	NYSPSLKSRVNLSDTSKNQVSLSLV AATAADSGKYYC	ARTLHGRR IYG I VAFNE WFTYFYMDV	WGNGTQVTVSS
10-410	QVQLQESGPGLVKPPETTLSTCS VS	GASVNDAY	WSWIRQSPGKR PEWVG Y	VHHS GDT	NYNP SLKRRTFSLDTAKNEVSLKLV ALTAADS AVYFC	ARALHGKR IYG I VALGE LFTYFYMDV	WGKGT TTVTVSS
10-1130	QVQLQESGPGLVKPPETTLSTCS VS	GASINDAY	WSWIRQSPGKR PEWVG Y	VHHS GDT	NYNP SLKRRTFSLDTAKNEVSLKLV DLTAADS AVYFC	ARALHGKR IYG I VALGE LFTYFYMDV	WGKGT TTVTVSS
10-1121	QVQLQESGPGLVKPPETTLSTCS VS	GASINDAY	WSWIRQSPGKR PEWVG Y	VHHS GDT	NYNP SLKRRTFSLDTAKNEVSLKLV DLTAADS AVYFC	ARALHGKR IYG I VALGE LFTYFYMDV	WGKGT TTVTVSS
10-1146	QVQLVESGPGLVTPSETTLSTCT VS	NGSVSGRF	WSWIRQSPGRG LEWIGY	FSDTDRS	EYSPSLRSRLTLSDASRNQLSLKLK SVTAADSATYYC	ARAQQGKR IYG I VSFGE FFYYYYMDA	WGKGTAVTVSS
10-996	QVQLQESGPGLVKPPSETTLSTCS VS	NGSVSGRF	WSWIRQSPGRG LEWIGY	FSDTEKS	NYNP SLRSRLTLSDASKNQLSLKLN SVTAADSATYYC	ARTQQGKR IYG VVSFGE FFHHYYMDA	WGKGTAVTVSS
GL	QVQLQESGPGLVKPPSETTLSTCT VS	GGSISSYY	WSWIRQPPGKG LEWIGY	IYYSGST	NYNP SLKSRVTISVDTSKNQFSLKLS SVTAADTAVYYC	ARTQQGKR IYG VVSFGD YYYYYYMDV	WGKGT TTVTVSS
10-1341	QVQLQESGPGLVKPPSETTLSVTCS VS	GDSMNNYY	WTWIRQSPGKG LEWIGY	ISDRESA	TYNP SLNSRVISRDTSTNQLSLKLN SVTPADTAVYYC	ATARRGQR IYG VVSFGE FFYYYSMDV	WGRGTTVTVSS
10-847	QVQLQESGPGLVKPPSETTLSVTCS VS	GDSMNNYY	WTWIRQSPGKG LEWIGY	ISDRASA	TYNP SLNSRVISRDTSKNQLSLKLN SVTPADTAVYYC	ATARRGQR IYG VVSFGE FFYYYSMDV	WGKGT TTVTVSS
10-1074	QVQLQESGPGLVKPPSETTLSVTCS VS	GDSMNNYY	WTWIRQSPGKG LEWIGY	ISDRESA	TYNP SLNSRVISRDTSKNQLSLKLN SVTPADTAVYYC	ATARRGQR IYG VVSFGE FFYYYSMDV	WGKGT TTVTVSS
10-1074GM	QVQLQESGPGLVKPPSETTLSVTCS VS	GDSMNNSY	WTWIRQSPGKG LEWIGY	ISKSESA	NYNP SLNSRVISRDTSKNQLSLKLN SVTPADTAVYYC	ATARHGQR IYG VVSFGE FFYYYSMDV	WGKGT TTVTVSS
KABAT	FWR1	CDR1	FWR2	CDR2	FWR3	CDR3	FWR4
10-1369	QVQLQESGPGLVKPLETTLSTCN VSGAFIA	DHYWS	WIRLP LIGKGE WIG	YVHDSGDIN Y NP SLKN	RVHLSLDKSTNQVSLKLM AVTAGDSA LYYCAT	TKHGRR IYG VVAFGEWF TYFYMDV	WGRGTTVTVSS
10-259	QVHLQESGPGLVKPPSETTLSTCN VSGTLVR	DNYWS	WMRQPLGKQPE WIG	YVHDSGDTNY NP SLKS	RVHLSLDKSNNLVSLRLTAVTAADSA TYYCAT	TKHGRR IYG I VAFNEWF TYFYMDV	WGKGT TTVTVSS
10-303	QVQLQESGPGLVKPPSETTLSTCS VSGASIS	DSYWS	WIRRSPGKGLE WIG	YVHKSGDTNY SPSLKS	RVNLSLDTSKNQVSLSLVAATAADSG KYYCAR	TLHGRR IYG I VAFNEWF TYFYMDV	WGNGTQVTVSS
10-410	QVQLQESGPGLVKPPETTLSTCS VSGASVN	DAYWS	WIRQSPGKRPE WVG	YVHHS GDTNY NP SLKR	RVTFSLDTAKNEVSLKLVALTAADSA VYFCAR	ALHGKR IYG I VALGELF TYFYMDV	WGKGT TTVTVSS
10-1130	QVQLQESGPGLVKPPETTLSTCS VSGASIN	DAYWS	WIRQSPGKRPE WVG	YVHHS GDTNY NP SLKR	RVTFSLDTAKNEVSLKLVDLTAADSA VYFCAR	ALHGKR IYG I VALGELF TYFYMDV	WGKGT TTVTVSS
10-1121	QVQLQESGPGLVKPPETTLSTCS VSGASIN	DAYWS	WIRQSPGKRPE WVG	YVHHS GDTNY NP SLKR	RVSFSLDTAKNEVSLKLVDLTAADSA IYFCAR	ALHGKR IYG I VALGELF TYFYMDV	WGKGT TTVTVSS
10-1146	QVQLVESGPGLVTPSETTLSTCT VSNGSVS	GRFWS	WIRQSPGRGLE WIG	YFSDTDRSEY SPSLRS	RLTLSDASRNQLSLKLKSVTAADSA TYYCAR	AQQGKR IYG I VSFGEFF YYYYMDA	WGKGTAVTVSS
10-996	QVQLQESGPGLVKPPSETTLSTCS VSNGSVS	GRFWS	WIRQSPGRGLE WIG	YFSDTEKSNY NP SLRS	RLTLSDASKNQLSLKLSVTAADSA TYYCAR	TQQGKR IYG VVSFGEFF HHYYMDA	WGKGTAVTVSS

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GL	QVQLQESGPGGLVKPSETLSLTCT VSGGSIS	SYAWS	WIRQPPGKGLE WIG	YIYSGSTNY NPSLKS	RVTISVDTSKNQFSLKLSSVTAADTA VYYCAR	TQQKRIYGVVSFGDIY YYYIMDV	WGKGTTVTVSS
10-1341	QVQLQESGPGGLVKPSETLSVTCS VSGDSMN	NIYWT	WIRQSPGKGLE WIG	YISDRESATY NPSLNS	RVVISRDTSTNQLSLKLSNSVTPADTA VYYCAT	ARRQRIYGVVSFGIEFF YYSMDV	WGRGTTVTVSS
10-847	QVQLQESGPGGLVKPSETLSVTCS VSGDSMN	NIYWT	WIRQSPGKGLE WIG	YISDRASATY NPSLNS	RVVISRDTSKNQLSLKLSNSVTPADTA VYYCAT	ARRQRIYGVVSFGIEFF YYSMDV	WGKGTTVTVSS
10-1074	QVQLQESGPGGLVKPSETLSVTCS VSGDSMN	NIYWT	WIRQSPGKGLE WIG	YISDRESATY NPSLNS	RVVISRDTSKNQLSLKLSNSVTPADTA VYYCAT	ARRQRIYGVVSFGIEFF YYSMDV	WGKGTTVTVSS
10-1074GM	QVQLQESGPGGLVKPSETLSVTCS VSGDSMN	NSYWT	WIRQSPGKGLE WIG	YISKSESANY NPSLNS	RVVISRDTSKNQLSLKLSNSVTPADTA VYYCAT	ARHGQRIYGVVSFGIEFF TYYSMDV	WGKGTTVTVSS

IgL SEQUENCES

IMGT	FWR1	CDR1	FWR2	CDR2	FWR3	CDR3	FWR4
GL	SYVLITQPPSVSVAPGQTARITCG GN	NIGSKS	VHWYQQKPGQA PVLVYV	DDS	DRPSGIPERFSGSNSGNTATLTISR V EAGDEADYYC	QVWDSSSDHPWV	FGGGTKLTLVL
10-1369	SSMSVSPGETAKITCGEK	SIGSRA	VQWYQKKPGQP PSLIY	NNQ	DRPSGVPERFSASPDIEFGTTATLT I TNVEAGDEADYYC	HIYDARRPTNWW	FDRGTTTLTVL
10-259	SSMSVSPGETAKISCGKE	SIGSRA	VQWYQQKSGQP PSLIY	NNQ	DRPSGVPERFSATPDFGAGTTATLT I TNVEADDEADYYC	HIYDARGGTNWW	FDRGATLTVL
10-303	SDISVAPGETARISCGEK	SLGSRA	VQWYQHRAGQA PSLIY	NNQ	DRPSGIPERFSGSPDSPFGTTATLT I TSVEAGDEADYYC	HIWDSRVPTKWV	FGGGTTLTVL
10-1121	SFVSVAPGQTARITCGEE	SLGSR	VIWYQQRPGQA PSLIMY	NNH	DRPSGIPERFSGSPGSTFGTTATLT I TSVEAGDEADYYC	HIWDSRRPTNWW	FGEGTTLTVL
10-410	SFVSVAPGQTARITCGEE	SLGSR	VIWYQQRPGQA PSLIY	NNN	DRPSGIPERFSGSPGSTFGTTATLT I TSVEAGDEADYYC	HIWDSRRPTNWW	FGEGTTLTVL
10-1130	SFVSVAPGQTARITCGEE	SLGSR	VIWYQQRPGQA PSLIY	NNN	DRPSGIPERFSGSPGSTFGTTATLT I TSVEAGDEADYYC	HIWDSRRPTNWW	FGEGTTLTVL
10-847	SYVRPLSVALGETASISCGRQ	ALGSRA	VQWYQHRPGQA PILLIY	NNQ	DRPSGIPERFSGTPDINFGTRATLT I SGVEAGDEADYYC	HMWDSRSGFSWS	FGGATRLTVL
10-1074	SYVRPLSVALGETARISCGRQ	ALGSRA	VQWYQHRPGQA PILLIY	NNQ	DRPSGIPERFSGTPDINFGTRATLT I SGVEAGDEADYYC	HMWDSRSGFSWS	FGGATRLTVL
10-1341	SYVRPLSVALGETARISCGRQ	ALGSRA	VQWYQHRPGQA PILLIY	NNQ	DRPSGIPERFSGTPDINFGTRATLT I SGVEAGDEADYYC	HMWDSRSGFSWS	FGGATRLTVL
10-996	SSLPLSVAPGATAKIAACGEK	SFASRA	VQWYQQKPGQA PVLIIY	NNQ	DRPAGVSERFSGTPDVGFGSTATLT I SRVEAGDEADYYC	HKWDSRSPLSWV	FGGGTQLTVL
10-1146	SSLPLSLAPGATAKIPCGEK	SRGSRA	VQWYQQKPGQA PTLIIY	NNQ	DRPAGVSERYSGNPDVAIGVTATLT I SRVEAGDEAEYYC	HYWDSRSPISWV	FGGWTQLTVL
KABAT	FWR1	CDR1	FWR2	CDR2	FWR3	CDR3	FWR4
GL	SYVLITQPPSVSVAPGQTARITC	GGNNIGSKSVH	WYQQKPGQAPV LVVY	DDSDRPS	GIPERFSGSNSGNTATLTISRVEAGD EADYYC	QVWDSSSDHPWV	FGGGTKLTVL
10-1369	SSMSVSPGETAKITC	GEKSIGRAVQ	WYQKKPGQPPS LIIY	NNQDRPS	GVPERFSASPDIEFGTTATLTITNVE AGDEADYYC	HIYDARRPTNWW	FDRGTTTLTVL
10-259	SSMSVSPGETAKISC	GKESIGRAVQ	WYQQKSGQPPS LIIY	NNQDRPS	GVPERFSATPDFGAGTTATLTITNVE ADDEADYYC	HIYDARGGTNWW	FDRGATLTVL
10-303	SDISVAPGETARISC	GEKSIGRAVQ	WYQHRAGQAPS LIIY	NNQDRPS	GIPERFSGSPDSPFGTTATLTITSVE AGDEADYYC	HIWDSRVPTKWV	FGGGTTLTVL

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10-1121	SFVSVAPGQTARITC	GEESLGSRSVI	WYQQRPGQAPS LIY	NNHDRPS	GIPERFSGSPGSTFGTTATLTITSVE AGDEADYYC	HIWDSRRPTNWV	FGEGTTLTLVL
10-410	SFVSVAPGQTARITC	GEESLGSRSVI	WYQQRPGQAPS LIY	NNNDRPS	GIPERFSGSPGSTFGTTATLTITSVE AGDEADYYC	HIWDSRRPTNWV	FGEGTTLTLVL
10-1130	SFVSVAPGQTARITC	GEESLGSRSVI	WYQQRPGQAPS LIY	NNNDRPS	GIPERFSGSPGSTFGTTATLTITSVE AGDEADYYC	HIWDSRRPTNWV	FGEGTTLTLVL
10-847	SYVRPLSVALGETASISC	GRQALGSRAVQ	WYQHRPGQAPI LIY	NNQDRPS	GIPERFSGTPDINFGTRATLTISGVE AGDEADYYC	HMWDSRSGFSWS	FGGATRLTLVL
10-1074	SYVRPLSVALGETARISC	GRQALGSRAVQ	WYQHRPGQAPI LIY	NNQDRPS	GIPERFSGTPDINFGTRATLTISGVE AGDEADYYC	HMWDSRSGFSWS	FGGATRLTLVL
10-1341	SYVRPLSVALGETARISC	GRQALGSRAVQ	WYQHRPGQAPI LIY	NNQDRPS	GIPERFSGTPDINFGTRATLTISGVE AGDEADYYC	HMWDSRSGFSWS	FGGATRLTLVL
10-996	SSLPLSVAPGATAKIIAC	GEKSFASTAVQ	WYQQKPGQAPV LIY	NNQDRPA	GVSERFSGTPDVGFGSTATLTISRVE AGDEADYYC	HKWDSRSPISWV	FGGGTQLTLVL
10-1146	SSLPLSLAPGATAKIPC	GEKSRGSRVAVQ	WYQQKPGQAPT LIY	NNQDRPA	GVSERYSGNPDVAIGVTATLTISRVE AGDEAEYYC	HYWDSRSPISWV	FGGWTQLTLVL

Table 2

Name	SEQ ID NOs			
	Variable Region		CDRs 1-3	
	Heavy chain (H)	Light chain (L)	H	L
consensus	SEQ ID NO: 1	SEQ ID NO: 2	SEQ ID NOs: 33-35	SEQ ID NOs: 36-38
10-259	SEQ ID NO: 3	SEQ ID NO: 4	SEQ ID NOs: 39-41	SEQ ID NOs: 42-44
10-303	SEQ ID NO: 5	SEQ ID NO: 6	SEQ ID NOs: 45-47	SEQ ID NOs: 48-50
10-410	SEQ ID NO: 7	SEQ ID NO: 8	SEQ ID NOs: 51-53	SEQ ID NOs: 54-56
10-847	SEQ ID NO: 9	SEQ ID NO: 10	SEQ ID NOs: 57-59	SEQ ID NOs: 60-62
10-996	SEQ ID NO: 11	SEQ ID NO: 12	SEQ ID NOs: 63-65	SEQ ID NOs: 66-68
10-1074	SEQ ID NO: 13	SEQ ID NO: 14	SEQ ID NOs: 69-71	SEQ ID NOs: 72-74
10-1121	SEQ ID NO: 15	SEQ ID NO: 16	SEQ ID NOs: 75-77	SEQ ID NOs: 78-80
10-1130	SEQ ID NO: 17	SEQ ID NO: 18	SEQ ID NOs: 81-83	SEQ ID NOs: 84-86
10-1146	SEQ ID NO: 19	SEQ ID NO: 20	SEQ ID NOs: 87-89	SEQ ID NOs: 90-92
10-1341	SEQ ID NO: 21	SEQ ID NO: 22	SEQ ID NOs: 93-95	SEQ ID NOs: 96-98
10-1369	SEQ ID NO: 23	SEQ ID NO: 24	SEQ ID NOs: 99-101	SEQ ID NOs: 102-104
PGT-121	SEQ ID NO: 25	SEQ ID NO: 26	SEQ ID NOs: 105-107	SEQ ID NOs: 108-110
PGT-122	SEQ ID NO: 27	SEQ ID NO: 28	SEQ ID NOs: 111-113	SEQ ID NOs: 114-116
PGT-123	SEQ ID NO: 29	SEQ ID NO: 30	SEQ ID NOs: 117-119	SEQ ID NOs: 120-122
GL	SEQ ID NO: 31	SEQ ID NO: 32	SEQ ID NOs: 123-125	SEQ ID NOs: 126-128
10-1074GM	SEQ ID NO: 129	SEQ ID NO: 130	SEQ ID NOs: 131-133	SEQ ID NOs: 134-136

Eleven new unique variants were expressed (**Table 3**) and demonstrated binding to YU-2
5 gp120 and gp140 by ELISA and surface plasmon resonance (SPR). Unless otherwise noted, the
gp120 and gp140 proteins for these and other experiments were expressed in mammalian cells that
can attach either a complex-type or a high-mannose *N*-glycan to a PNGS. The level of reactivity
with gp120 differed between antibodies belonging to the PGT121 and 10-1074 groups, the latter
exhibiting higher apparent affinities (**Fig. 3A**) mainly due to slower dissociation from
10 gp120/gp140 for the 10-1074-related antibodies (**Fig. 4B**).

EXAMPLE 3 *PGT121 and 10-1074 epitopes*

Asn332_{gp120} in the vicinity of the V3 loop stem was reported as critical for binding and
viral neutralization by PGT121 (Nature 477(7365):466-470), thus we examined the role of V3 in
antigen recognition by PGT121-like and 10-1074-like antibodies. ELISAs were performed using
15 HXB2 gp120 “core” proteins that lack V1-V3 loops (gp120^{core}) or retain a portion of V3 (2CC-
core), and using a YU-2 gp120 mutant protein carrying a double alanine substitution in the V3
stem (gp120^{GD324-5AA}). The tested antibodies showed decreased reactivity against variants lacking

the V3 loop and gp120^{GD324-5AA} when compared to intact YU-2 gp120, with the binding of 10-1074-group antibodies being the most affected (**Fig. 5A and B**). These results suggest that recognition by both antibody groups involves protein determinants in the vicinity of the V3 loop. None of the antibodies bound to overlapping peptides spanning V3, suggesting the targeted epitopes are discontinuous and/or require a particular conformation not achieved by isolated peptides (**Fig. 5C**).

Asn332_{gp120} (Asn337_{gp120} in earlier numbering (J Proteome Res 7(4):1660-1674)) is the N-terminal residue of a potential *N*-glycosylation site (PNGS) defined as the sequence Asn – X – Ser/Thr. To determine whether Asn332_{gp120} and/or its *N*-linked glycan are required for gp120 reactivity of the new PGT121- and 10-1074-group antibodies, we tested their binding to YU-2 gp120^{N332A} by ELISA. The N332A substitution diminished the binding of PGT121 and all the new antibody variants, whereas their reactivity against a mutant gp120 lacking a nearby glycosylation site (gp120^{NNT301-3AAA} mutant) was unchanged. To determine if a PNGS in addition to the Asn332_{gp120} PNGS affects recognition by the new antibodies, we constructed a series of 11 double glycan mutants in which the N332A mutation in YU-2 gp120 was combined with mutation of PNGSs located between Asn262_{gp120} and Asn406_{gp120}. All of the PGT121-like and 10-1074-like antibodies bound to each of the double glycan mutants with comparable affinity as to that for gp120^{N332A}.

To compare overall glycan recognition by the PGT121- and 10-1074-like antibodies, we examined their binding to YU-2 gp120 treated with PNGase F, which cleaves both complex-type and high-mannose *N*-glycans. Because gp120 cannot be fully deglycosylated enzymatically unless it is denatured, PNGase F treatment resulted in partial deglycosylation of natively-folded gp120 (**Fig. 6**). Nevertheless, the reactivities of the two groups of antibodies differed in that partial deglycosylation of gp120 by PNGase F decreased the binding activity of all PGT121-like antibodies but none of the 10-1074-like antibodies (**Fig. 6C**). Similar experiments conducted with YU-2 gp120 treated with Endo H, which cleaves high-mannose, but not complex-type, *N*-glycans, affected binding of 10-1074-like antibodies more than PGT121-like antibodies (**Fig. 6D**).

An *N*-glycan microarray revealed that six of seven tested PGT121-like antibodies showed detectable binding to complex-type mono- or bi-antennary *N*-glycans terminating with galactose or α 2-6-linked sialic acid but no detectable binding to high-mannose type glycans, corroborating and extending previous reports of no binding of PGT121-123 to high-mannose *N*-glycans and no

competition by Man₄ and Man₉ dendrons for gp120 binding (**Fig. 7**). In contrast, there was no detectable binding to protein-free glycans by 10-1074-like antibodies (**Fig. 7**). Although PGT121-like antibodies bound to protein-free complex-type, but not high-mannose, *N*-glycans, PGT121-like antibodies retained binding to YU-2 gp120 produced in cells treated with kifunensine (gp120_{kif}), a mannosidase inhibitor that results in exclusive attachment of high-mannose glycans to PNGSs (**Fig. 8B**). Most of the PGT121-like antibodies exhibited a small, but reproducible, decrease in binding to gp120_{kif}. By contrast, 10-1074-like antibodies retained full binding to gp120_{kif} (**Fig. 8B**). These results are consistent with the hypothesis that high-mannose, as well as complex-type, *N*-glycans can be involved in the epitope of PGT121-like antibodies.

Epitope mapping experiments were performed with two representative members of each group (PGT121 and 10-1369 for the PGT121-like group; 10-1074 and 10-996 for the 10-1074-like group) by competition ELISA. All four antibodies showed cross-competition, but PGT121 more modestly inhibited the binding of 10-996 and 10-1074 to gp120 than vice-versa. To further map the targeted epitopes, we used anti-gp120 antibodies that recognize the crown of the V3 loop (**Fig. 5**), the CD4bs, the co-receptor binding site (CD4-induced; CD4i), a constellation of high-mannose *N*-glycans (2G12) (Journal of virology 76(14):7293-7305; Proc Natl Acad Sci USA 102(38):13372-13377), or the V3 loop and *N*-linked glycans at positions 301 and 332 (PGT128). Anti-V3 crown antibodies inhibited the binding of PGT121 and 10-1369 but did not interfere with the binding of 10-996 and 10-1074. PGT128, and to a lesser extent 2G12, but not the CD4bs and CD4i antibodies, diminished the binding of all four antibodies to gp120.

Taken together, these data suggest that PGT121 clonal members recognize a site involving a protein determinant in the vicinity of the V3 loop and the Asn332_{gp120}-associated glycan. However, the clone segregates into two families, the PGT121-like and 10-1074-like groups, which differ in their affinities for gp120 and in the role of glycans in epitope formation.

EXAMPLE 4 *Broad and potent HIV neutralization*

To evaluate the neutralizing activity of the new PGT121 variants, we measured their ability to inhibit HIV infection of TZM-bl cells using 10 viral strains including R1166.c1, which lacks the PNGS at gp120 position 332. All PGT121 variants, including the 10-1074-like antibodies, neutralized 9 pseudoviruses and none neutralized the R1166.c1 control (**Fig. 1A** and **Table 4**). Neutralizing activity correlated with affinity for the HIV spike, with the 10-1074 group showing slightly greater potencies than the PGT121 group (**Fig. 1B** and **Fig. 4C**). A representative

germline version (GL) of the PGT121/10-1074 antibody clonotype failed to bind gp120/gp140 or neutralize any viruses in the panel, implying that somatic mutation is required for binding and neutralization. Pairing GL light chains with mutated 10-1074- or 10-996-group heavy chains failed to rescue binding or neutralization, suggesting that both mutated chains contribute to proper assembly of the antibody paratope.

Next assays were carried out to compared the neutralization activities of PGT121 and two 10-1074-like antibodies (10-996 and 10-1074) against an extended panel of 119 difficult-to-neutralize pseudoviruses (classified as tier-2 and tier-3) (**Tables 4 and 5**). 10-996 and 10-1074 showed neutralization potencies and breadth similar to PGT121 (**Fig. 1C, Fig. 9, and Tables 5 and 6**). As anticipated, most viruses bearing amino acid changes at gp120 positions 332 and/or 334 (spanning the Asn332–X–Ser334/Thr334 PNGS) were resistant to neutralization (83.8% were resistant to PGT121, 100% were resistant to 10-1074 and 10-996). Mutation at this PNGS accounted for the majority of viruses resistant to neutralization (68.5% for 10-996, 72.5% for 10-1074 and 60.8% for PGT121) (**Table 7**). Comparable neutralization activities were observed for the IgG and Fab forms of PGT121 and 10-1074, suggesting that bivalency is not critical for their activity (**Fig. 1D**).

To evaluate the potential role of complex-type *N*-glycans on the HIV envelope in neutralization by PGT121 and 10-1074, we produced high-mannose-only virions in two different ways: by assembling pseudoviruses in cells treated with kifunensine, which results in Man₉GlcNAc₂ *N*-linked glycans, or by assembly in HEK 293S GnTI^{−/−} cells, which results in Man₅GlcNAc₂ *N*-linked glycans. We found that PGT121 neutralized 2 of 3 kifunensine-derived PGT121-sensitive/10-1074-resistant strains equivalently to their counterparts produced in wildtype cells (**Fig. 8C**). Two PGT121-sensitive/10-1074-sensitive viral strains produced in GnTI^{−/−} cells were equally as sensitive to PGT121 and 10-1074 as their counterparts produced in wildtype cells. Consistent with previous reports that complex-type *N*-glycans partially protect the CD4 binding site from antibody binding, the viruses produced in GnTI^{−/−} cells were more sensitive to CD4-binding site antibodies (NIH45-46^{G54W} and 3BNC60) (**Fig. 8D**).

EXAMPLE 5 *Newly-transmitted HIV-1*

We next examined the activity of PGT121 and 10-1074 against transmitted founder viruses by evaluating neutralization in a peripheral blood mononuclear cell (PBMC)-based assay using 95 clade B viruses isolated from a cohort of individuals who seroconverted between 1985 and 1989

(historical seroconverters, n=14) or between 2003 and 2006 (contemporary seroconverters, n=25) (51, 52). We compared PGT121 and 10-1074 with anti-CD4bs bNAbs and other bNAbs including VRC01, PG9/PG16, b12, 2G12, 4E10 and 2F5. Clustering analyses of neutralization activity showed segregation into two groups; the PGT121/10-1074 group contained the most active HIV neutralizers including the anti-CD4bs and PG9 antibodies (**Table 8**). Remarkably, 10-1074 showed exceptional neutralization potency on this clade B virus panel, exhibiting the greatest breadth at 0.1 $\mu\text{g/ml}$ (67% of the 95 clade B viruses) of all bNAbs tested (**Table 8**). Although 10-1074 showed higher potency on contemporary clade B viruses than PGT121 (~20-fold difference), both antibodies were more effective against historical than contemporary viruses (**Fig. 1E** and **Fig. 10**).

EXAMPLE 6 *Crystal structures of PGT121, 10-1074 and GL*

To investigate the structural determinants of the differences between PGT121-like and 1074-like antibodies, we solved crystal structures of the Fab fragments of PGT121, 10-1074 and a representative germline precursor (GL) at 3.0 Å, 1.9 Å and 2.4 Å resolution, respectively (**Table 9**). Superimposition of the heavy and light chain variable domains (V_H and V_L) among the three Fabs showed conservation of the backbone structure, with differences limited to small displacements of the CDRH3 and CDRL3 loops of the affinity-matured Fabs relative to GL (**Table 10**).

An unusual feature shared by the antibodies is their long (25 residues) CDRH3 loop, which forms a two-stranded anti-parallel β -sheet extending the V_H domain F and G strands. In each Fab, the tip of the extended CDRH3 loop primarily contains non-polar residues. A similar structural feature was observed for the CDRH3 of PGT145, a carbohydrate-sensitive antibody whose epitope involves the gp120 V1V2 loop. However, the extended two-stranded β -sheet of PGT145's CDRH3 contains mostly negatively-charged residues, including two sulfated tyrosines at the tip. Aligning V_H - V_L of PGT121 and PGT145 (**Table 10**) shows that CDRH3_{PGT145} extends past CDRH3_{PGT121} and that its tip and V_H domain are aligned, whereas the CDRH3s of PGT121, 10-1074 and GL tilt towards V_L . The tilting of CDRH3_{PGT121}/CDRH3₁₀₋₁₀₇₄/CDRH3_{GL} towards V_L opens a cleft between CDRH2 and CDRH3, a feature not shared by related antibodies.

PGT121 and 10-1074 are highly divergent with respect to GL and each other (of 132 residues, PGT121_{VH} differs from 10-1074_{VH} and GL_{VH} by 36 and 45 residues, respectively, and 10-1074_{VH} and GL_{VH} differ by 29). The majority of the PGT121/10-1074 differences are located

in the CDR_{VH} loops and CDRL3. Interestingly, six substitutions in CDRH3 (residues 100d, 100f, 100h, 100j, 100l, 100n) alternate such that every second residue is substituted, causing resurfacing of the cleft between CDRH2 and CDRH3 that results from CDRH3 tilting towards V_L. This region likely contributes to the different fine specificities of PGT121 and 10-1074. Five other solvent-exposed substitutions in heavy chain framework region 3 (FWR3_{HC}) (residues 64, 78, 80-82; strands D and E) are potential antigen contact sites given that framework regions in HIV antibodies can contact gp120. Other differences that may contribute to fine specificity differences include a negative patch on PGT121 in the vicinity of Asp56_{HC} not present in 10-1074 or GL (Ser56_{HC} in 10-1074 and GL) and positive patches on the CDRL1 and CDRL3 surface not found on the analogous surface of GL .

Somatic mutations common to PGT121 and 10-1074 may be involved in shared features of their epitopes. The heavy chains of PGT121 and 10-1074 share only three common mutations (of 36 PGT121–GL and 29 10-1074–GL differences). In contrast, PGT121 and 10-1074 share 18 common light chain mutations (of 37 PGT121–GL and 36 10-1074–GL differences), including an insertion in light chain FWR3 that causes bulging of the loop connecting strands D and E, and the substitution of Asp50_{LC}-Asp51_{LC} in CDRL2_{GL} to Asn50_{LC}-Asn51_{LC} in both PGT121 and 10-1074, resulting in a less negatively-charged surface. The large number of common substitutions introduced into LC_{PGT121} and LC₁₀₋₁₀₇₄ (approximately 50% of LC substitutions) point to CDRL1, CDRL2 and FWR2_{LC} as potential contact regions for epitopes shared by PGT121 and 10-1074.

Next, comparisons were made with the structure of PGT128, which recognizes Asn332_{gp120}- and Asn301_{gp120}-linked glycans and V3 and was solved as a complex with an outer domain/mini-V3 loop gp120 expressed in cells that cannot produce complex-type *N*-glycan-modified proteins. Unlike the CDRH3 loops of PGT121 and 10-1074, PGT128_{CDRH3} is not tilted towards PGT128_{VL}, and CDRH3_{PGT128} does not include a two-stranded β -sheet. In addition, CDRH3_{PGT128} (18 residues) is shorter than the CDRH3s of PGT121 and 10-1074 (24 residues), whereas CDRH2_{PGT128} contains a six-residue insertion not found in PGT121 or 10-1074. Due to these differences, CDRH2 is the most prominent feature in PGT128, whereas CDRH3 is most prominent in PGT121 and 10-1074. CDRH2_{PGT128} and CDRL3_{PGT128} together recognize Man_{8/9} attached to Asn332_{gp120}, and CDRH3_{PGT128} contacts the V3 loop base. This mode of gp120 recognition is not possible for PGT121 and 10-1074 because the structural characteristics of their CDRH2 and CDRH3 loops differ significantly from those of PGT128, consistent with the ability

of PGT128, but not PGT121 and 10-1074 (**Fig. 7**), to recognize protein-free high-mannose glycans.

EXAMPLE 7 *Crystal structure of PGT121-glycan complex*

A 2.4 Å resolution structure of PGT121 associated with a complex-type sialylated bi-antennary glycan was solved (**Table 9**) using crystals obtained under conditions including NA2, a complex-type asialyl bi-antennary glycan (**Fig. 7**). Surprisingly, the glycan bound to PGT121 in our crystal structure was not NA2, but rather a complex-type *N*-glycan from a neighboring PGT121 Fab in the crystal lattice; specifically the *N*-glycan attached to Asn105_{HC}. The glycan identity is evident because there was electron density for the glycosidic linkage to Asn105_{HC} and for a terminal sialic acid on the Man α 1-3Man antenna (the galactose and sialic acid moieties of Man α 1-6Man antenna were unresolved). The composition of the bound glycan corresponds to a portion of the α 2-6-sialylated A2(2-6) glycan that was bound by PGT121 in microarray experiments (**Fig. 7**) and to the expected sialyl linkage on complex-type *N*-glycans attached to PNGS on proteins expressed in HEK293T cells. Although the V_H-V_L domains of this structure (“liganded” PGT121) superimpose with no significant differences onto the V_H-V_L domains of the PGT121 structure with no bound *N*-glycan (“unliganded” PGT121) (**Table 10**), the elbow bend angle (angle between the V_H-V_L and C_H1-C_L pseudo-dyads) differs between the structures. This difference likely reflects flexibility that allows the Fab to adopt variable elbow bend angles depending upon crystal lattice forces.

Given that we observed binding of complex-type *N*-glycan in one crystal structure (the “liganded” PGT121 structure) but not in another structure (the “unliganded” PGT121 structure), we estimate that the affinity of PGT121 for complex-type *N*-glycan not attached to gp120 is in the range of the concentration of PGT121 in crystals (~10 mM). If we assume that the *K_D* for binding isolated glycan is in the range of 1-10 mM, comparable to the 1.6 mM *K_D* derived for PG9 binding to Man₅GlcNAc₂-Asn, then the *K_D* for PGT121 binding of isolated glycan represents only a minor contribution to the affinity of PGT121 for gp120, which is in the nM range (**Fig. 4A**).

The glycan in the “liganded” PGT121 structure interacts exclusively with the V_H domain and makes extensive contacts with residues in all three CDRs (buried surface area on PGT121_{HC} = 600 Å²). Contacts include 10 direct and 18 water-mediated hydrogen bonds (**Table 11**) with 9 amino acids anchoring the glycan between the *N*-acetylglucosamine moiety linked to the branch-point mannose and the terminal sialic acid on the 1-3-antenna. Several contacts with PGT121 are

made by this sialic acid, including three direct hydrogen bonds with PGT121 residues Asp31_{HC} and His97_{HC} in addition to water-mediated hydrogen bonds with Asp31_{HC}. The sialic acid also contributes to a water-mediated intra-glycan hydrogen bond network. The direct contacts with sialic acid may explain the stronger binding of PGT121 to the sialylated A2(2-6) glycan than to the asialylated NA2 glycan in our glycan microarray analysis (**Fig. 7**). Extensive water-mediated protein contacts established by the *N*-acetylglucosamine and galactose moieties of the 1-3-antenna could explain the binding observed for asialylated mono- and bi-antennary glycans to PGT121 (**Fig. 7**).

Six of the residues contributing direct or likely amino acid side chain contacts to the glycan (Ser32_{HC-CDRH1}, Lys53_{HC-CDRH2}, Ser54_{HC-CDRH2}, Asn58_{HC-CDRH2}, His97_{HC-CDRH3}, Thr100_{HC-CDRH3}) differ from those on 10-1074 (Tyr32_{HC-CDRH1}, Asp53_{HC-CDRH2}, Arg54_{HC-CDRH2}, Thr58_{HC-CDRH2}, Arg97_{HC-CDRH3}, Tyr100_{HC-CDRH3}), and are highly conserved among PGT121-like, but not 10-1074-like, antibodies. The 10-1074 residues lack the corresponding functional groups to make the observed glycan contacts or have bulky side chains that would cause steric clashes. Four of these residues also differ from those on GL (Tyr32_{HC-CDRH1}, Tyr53_{HC-CDRH2}, Gln97_{HC-CDRH3}, Tyr100_{HC-CDRH3}), suggesting that the lack of binding of 10-1074-like antibodies and GL to protein-free complex-type glycans in our glycan microarrays results from missing hydrogen bonds and/or steric clashes (e.g., His97_{PGT121} versus Arg97₁₀₋₁₀₇₄; Thr100_{PGT121} versus Tyr100₁₀₋₁₀₇₄). As the majority of sequence differences between PGT121 and 10-1074 cluster in the CDRH loops, specifically to the surface of the cleft between CDRH2 and CDRH3 where we observe the bound complex-type *N*-glycan, differential recognition of complex-type glycans on gp120 may account for some or all of the differences in their fine specificity observed.

EXAMPLE 8 *Substitution of glycan-contacting antibody residues affects neutralization*

To evaluate the contributions of complex-type *N*-glycan contacting residues identified from the “liganded” PGT121 structure, we generated two mutant antibodies designed to exchange the complex-type glycan-contacting residues between PGT121 and 10-1074: a 10-1074 IgG with PGT121 residues (six substitutions in IgH Y32S, D53K, R54S, T58N, R97H, Y100IT) and a PGT121 IgG with reciprocal substitutions. The “glycomutant” antibodies (10-1074_{GM} and PGT121_{GM}) exhibited near-wildtype apparent affinity for YU-2 gp120/gp140 as measured by SPR (**Fig. 2A**), demonstrating that the substitutions did not destroy binding to an envelope spike derived from a viral strain neutralized by both PGT121 and 10-1074 (**Fig. 1A**). The fact that

PGT121 complex-type *N*-glycan contacting residues can be accommodated within the 10-1074 background without destroying binding to a gp120/gp140 bound by both wildtype antibodies implies overall similarity in antigen binding despite fine specificity differences.

Unlike wildtype PGT121, PGT121_{GM} showed no glycan binding in microarray
 5 experiments, confirming that 10-1074 residues at the substituted positions are not compatible with protein-free glycan binding (**Fig. 2B**) and supporting the suggestion that residues contacting the glycan in the “liganded” PGT121 structure are involved in recognition of complex-type glycans in the microarrays. 10-1074_{GM} also showed no binding to protein-free glycans (**Fig. 2B**), indicating the involvement of residues in addition to those substituted in creating the binding site for a
 10 protein-free complex-type *N*-glycan.

Next, a TZM-bl-based assay was used to compare neutralization of the wildtype and “glycomutant” antibodies. We tested 40 viral strains including strains differentially resistant to PGT121 or 10-1074 and strains sensitive to both wildtype antibodies (**Fig. 2C** and **Table 12**). Consistent with the binding of PGT121_{GM} and 10-1074_{GM} to purified YU-2 envelope proteins,
 15 both mutants neutralized the YU-2 virus; however, 64% of the PGT121-sensitive strains were resistant to PGT121_{GM} (**Fig. 2 C**, and **Table 12**) suggesting that the glycan-contacting residues identified in the “liganded” PGT121 structure are relevant to the neutralization activity of PGT121. Conversely, 10-1074_{GM} exhibited a higher average potency than wildtype 10-1074 against the 10-1074-sensitive strains (**Fig. 2C** and **Table 12**), including potency increases of >3-
 20 fold against four 10-1074-sensitive strains (WITO4160.33, ZM214M.PL15, Cel172_H1, and 3817.v2.c59). In general, the PGT121 substitutions into 10-1074 did not confer sensitivity to 10-1074_{GM} upon PGT121-sensitive/10-1074-resistant strains, however two of these strains (CNE19 and 62357_14_D3_4589) became sensitive to 10-1074_{GM} (IC₅₀s = 0.19 µg/ml and 40.8 µg/ml, respectively). Interestingly, these are the only PGT121-sensitive/10-1074-resistant strains that
 25 include an intact Asn332_{gp120}-linked PNGS. The other PGT121-sensitive/10-1074-resistant strains lack the Asn332_{gp120}-linked glycan and are resistant to PGT121_{GM} and 10-1074_{GM}, implying that their sensitivity to wildtype PGT121 involve a nearby *N*-glycan and/or compensation by protein portions of the epitope. Although a dramatic gain of function was observed only for 10-1074_{GM} against one strain (CNE19), this result, together with the general improvement observed for 10-
 30 1074_{GM} against 10-1074-sensitive strains (**Fig. 2C**), is consistent with the interpretation that the crystallographically-identified glycan-contacting residues can transfer PGT121-like recognition

properties to 10-1074 in some contexts and/or affect its potency in others. In addition, the loss of neutralization activity for PGT121_{GM} against PGT121-sensitive strains demonstrates that neutralization activity of PGT121 involves residues identified as contacting complex-type *N*-glycan in the “liganded” PGT121 structure.

5 *Results*

PGT121 is a glycan-dependent bNAb that was originally identified in the serum of a clade A-infected donor in a functional screen yielding only two clonally-related members. gp140 trimers were used as “bait” for single cell sorting to isolate 29 new clonal variants of. The PGT121 clonal family includes distinct groups of closely-related antibodies; the PGT121- and 10-1074-groups. The results suggest that the epitopes of both groups involve the PNGS at Asn332_{gp120} and the base of the V3 loop. The PGT121-like and 10-1074-like antibody groups differ in amino acid sequences, gp120/gp140 binding affinities, and neutralizing activities, with the 10-1074-like antibodies being completely dependent for neutralization upon an intact PNGS at Asn332_{gp120}, whereas PGT121-like antibodies were able to neutralize some viral strains lacking
15 the Asn332_{gp120} PNGS.

A notable difference between the two antibody groups is that the PGT121-like antibodies bound complex-type *N*-glycans in carbohydrate arrays, whereas the 10-1074-like antibodies showed no detectable binding to any of the protein-free *N*-glycans tested (**Fig. 7**). Protein-free glycan binding by anti-HIV antibodies is not always detectable; e.g., although PG9 recognizes a gp120-associated high-mannose glycan, no binding to protein-free glycans was detected in
20 microarrays. Thus although a positive result in a glycan microarray implies involvement of a particular glycan in an antibody epitope, a negative result does not rule out glycan recognition. For example, although not detectable in the glycan microarray experiments, high-mannose glycans may be involved in the PGT121 epitope, consistent with binding and neutralization of
25 high-mannose-only forms of gp120 protein and virions (**Fig. 8**).

The molecular basis for the differences between PGT121, 10-1074 and their GL progenitor was revealed in part by their crystal structures. The finding that the majority of light chain somatic mutations are shared between PGT121 and 10-1074, whereas mutations in the heavy chains differ, suggests that the light chain contacts shared portions of the gp120 epitope and the heavy chain
30 recognizes distinct features. All three antibodies exhibit an extended CDRH3 with a non-polar tip that may allow accessing of cryptic epitopes. Differences in the antigen-binding site of the two

mature Fabs were mainly localized to a cleft between CDRH2 and the extended CDRH3. Interestingly, the putative antigen-binding cleft between CDRH2 and CDRH3 was also found in a representative germline progenitor of PGT121 and 10-1074.

Structural information was obtained concerning glycan recognition by PGT121-like
 5 antibodies from a crystal structure in which a complex-type sialylated *N*-glycan attached to a V_H domain residue interacted with the combining site of a neighboring PGT121 Fab. Several features of the “liganded” PGT121 structure suggest it is relevant for understanding the recognition of complex-type *N*-glycans on gp120 by PGT121-like antibodies. First, the glycan in the structure corresponds to the α 2-6 sialylated glycan A2(2-6) PGT121 binds in microarrays (**Fig. 7**). Second,
 10 the glycan interacts with PGT121 using the cleft between CDRH3 and CDRH2 that was suggested by structural analyses to be involved in epitope recognition, potentially explaining the unusual tilting of CDRH3 towards V_L in the PGT121 and 10-1074 structures. Third, most of the V_H residues identified as interacting with the glycan differ between PGT121 and 10-1074, rationalizing different binding profiles in glycan microarrays and potentially explaining the
 15 different fine specificities revealed in protein binding experiments. Fourth, swapping crystallographically-identified glycan contact residues between PGT121 and 10-1074 in part transferred their properties: PGT121_{GM}, like 10-1074, did not bind to protein-free glycans, but both PGT121_{GM} and 10-1074_{GM} preserved near wildtype binding to purified YU-2 gp120/gp140. Although PGT121_{GM} retained the ability to neutralize some viral strains that were neutralized by
 20 wildtype PGT121 and 10-1074, it failed to neutralize strains that are PGT121-sensitive/10-1074-resistant, demonstrating that the glycan-binding motif is essential for the neutralizing activity of PGT121 against 10-1074-resistant strains. For the reciprocal swap, the neutralization potency of 10-1074_{GM} was increased or unaffected relative to 10-1074, and in one case, 10-1074_{GM} potently neutralized a PGT121-sensitive/10-1074-resistant strain, consistent with transfer of the
 25 crystallographically-identified glycan motif and the hypothesis that the epitopes of PGT121- and 10-1074-like antibodies are related. In analyses of gp120 sequences from strains for which PGT121 neutralization data are available, other than a correlation with the PNGS at Asn332gp120 for viruses sensitive to PGT121-like and 10-1074-like antibodies, no clear pattern of PNGS usage emerges for the different categories of viral strains (PGT121-sensitive/10-1074-sensitive,
 30 PGT121-sensitive/10-1074-resistant, PGT121-resistant/10-1074-sensitive) except that the 10-1074-resistant strains generally lack the Asn332gp120-associated PNGS.

EXAMPLE 9 *Passive Transfer of anti-HIV-1 neutralizing mAbs in-vivo*

Five isolated potent and broadly acting anti-HIV neutralizing monoclonal antibodies were administered to rhesus macaques and challenged them intrarectally 24h later with either of two different SHIVs. By combining the results obtained from 60 challenged animals, the protective neutralization titer in plasma preventing virus acquisition in 50% of the exposed monkeys was approximately 1:100.

Animal Experiments

The macaques used in this study were negative for the MHC class I *Mamu-A*01* allele.

Construction of the R5-tropic SHIVDH12-V3AD8

PCR mutagenesis, with primers corresponding to the 5' and 3' halves of the SHIVAD8EO (PNAS 109, 19769-19774 (2012)) gp120 V3 coding region (forward primer: AGAGCATTTTATACAACAGGAGACATAATAGGAGATATAAGACAAGCACATTGCAA CATTAGTAAAGTAAAATGGC and reverse primer: TCCTGGTCCTATATGTATACTTTTCCTTGTATTGTTGGGTCTTGTACAATTAATTT CTACAGTTTCATTC), was employed to introduce these V3 sequences into the genetic background of the pSHIVDH12.CL7 molecular clone (*J. of Virology* 78, 5513-5519 (2004)), using Platinum PFX DNA polymerase (Invitrogen). Following gel purification, the PCR product was treated with T4 polynucleotide kinase (GibcoBRL) and blunt-end ligated to create pSHIVDH12.V3AD8, which was used to transform competent cells.

Viruses

Virus stocks were prepared by first transfecting 293T cells with the SHIVAD8EO or SHIVDH12-V3AD8 molecular clones using Lipofectamine 2000 (Invitrogen, Carlsbad, CA). Culture supernatants were collected 48 h later and aliquots stored at -80°C until use. Concanavalin A-stimulated rhesus PBMCs (2×10^6 cells in 500 μl) were infected with transfected cell supernatants by spinoculation (*J. of Virology* 74, 10074-10080 (2000)) for 1 h, mixed with the same number/volume of activated PBMC, and cultures were maintained for at least 12 days with daily replacement of culture medium. Samples of supernatant medium were pooled around the times of peak RT production to prepare individual virus stocks.

Antibodies

Eleven monoclonal antibodies (VRC01, NIH45-46, 45-46G54W, 45-46m2, 3BNC117, 12A12, 1NC9, and 8ANC195, 10-1074, PGT121, and PGT126) were isolated and produced.

DEN3, a dengue virus NS1-specific human IgG1 monoclonal antibody (PNAS 109, 18921-18925 (2012)), or control human IgG (NIH Nonhuman Primate Reagent Resource <http://www.nhpreagents.org>) were used as the negative control antibodies in this study. The monoclonal antibodies selected for pre-exposure passive transfer were administered intravenously 24 h before virus challenge.

Quantitation of Plasma Viral RNA Levels.

Viral RNA levels in plasma were determined by real-time reverse transcription-PCR (ABI Prism 7900HT sequence detection system; Applied Biosystems).

Antibody concentrations in plasma.

The concentrations of administered monoclonal antibodies in monkey plasma were determined by enzyme-linked immunosorbent assay (ELISA) using recombinant HIV-1JRFL gp120 (Progenics Pharmaceuticals) or HIVIIIB (Advanced Biotechnology inc) (J. of Virology 75, 8340-8347 (2001)). Briefly, microtiter plates were coated with HIV-1 gp120 (2 µg/ml) and incubated overnight at 4 °C. The plates were washed with PBS/0.05% Tween-20 and blocked with 1% (vol/vol) BSA. After blocking, serial dilution of antibodies or plasma samples were added to the plate and incubated for 1 h at room temperature. Binding was detected with a goat anti-human IgG F(ab)2 fragments coupled to alkaline phosphatase (Pierce) and visualized with SIGMAFAST OPD (Sigma-Aldrich). The decay half-lives of neutralizing monoclonal antibodies were calculated by a single-exponential decay formula based on the plasma concentrations beginning on day 5 or day 7 post antibody administration (J. of Virology 84, 1302-1313 (2010)).

Neutralization Assays.

The in vitro potency of each mAb and the neutralization activity present in plasma samples collected from rhesus macaques were assessed by two types of neutralization assays; 1) TZM-bl entry assay with pseudotyped challenge virus (AIDS Res Hum Retroviruses 26, 89-98 (2010)) or 2) a 14 day PBMC replication assay with replication competent virus (J. of virology 76, 2123-2130 (2002)). For the TZM-bl assay, serially diluted mAb or plasma samples were incubated with pseudotyped viruses, expressing *env* gene derived from SHIVAD8EO or SHIVDH12.V3AD8 and prepared by cotransfecting 293T cells with pNLenv1 and pCMV vectors expressing the respective envelope proteins (J. of Virology 84, 4769-4781 (2010)). The 50% neutralization inhibitory dose (IC50) titer was calculated as the dilution causing a 50% reduction in relative luminescence units (RLU) compared with levels in virus control wells after subtraction of cell control RLU (J. of

Virology 84, 1439-1452 (2010)). The neutralization phenotype (tier levels) of the SHIVDH12-V3AD8 molecular clone was determined by TZM-bl cell assay using plasma samples from a cohort study, which exhibit a wide range of neutralizing activities against subtype B HIV-1 isolates (J. of General Virology 91, 2794-2803 (2010)).

5 *Determinations of animal protective titers and statistical analyses.*

Calculation of the neutralizing titer in plasma against each R5 SHIV, resulting in the prevention of virus acquisition of 50 or 80% of the virus-challenged animals, was performed using the method of Reed and Muench (Am J Hyg 27, 493-497 (1938)). One significant outlier animal (DEW7) was omitted from the calculation. Probit regression was used to model the relationship
10 between the titers in plasma required to confer sterilizing immunity *in vivo* using all 60 passively immunized monkeys (Cambridge University Press, Cambridge, England, ed. 3rd, 2007), with p-values from this model based on Likelihood ratio Tests. Plasma titers needed for different levels of *in vivo* protection (33%, 50%, 80%, 90%, and 95%) were determined from the probit model estimates and the method of bootstrapping was used to construct 90% confidence intervals.

15 *Results:*

SHIVDH12-V3AD8, like SHIVAD8EO, possesses Tier 2 anti-HIV-1 neutralization sensitivity properties (Table 13). Rhesus macaques inoculated intravenously or intrarectally with SHIVDH12-V3AD8 exhibited peak viremia ranging from 10^5 to 10^7 viral RNA copies/ml of plasma at weeks 2 to 3 post infection (PI). In most SHIVDH12-V3AD8 infected
20 animals, plasma viral loads decline to background levels between weeks 8 to 20 PI.

The neutralization sensitivity of SHIVAD8EO to 11 recently reported broadly reacting anti-HIV-1 mAbs was initially determined in the TZM-bl assay system (FIG. 11A and B). Eight of these antibodies, VRC01, NIH45-46 (23), 45-46G54W, 45-46m2, 3BNC117, 12A12, 1NC9, and 8ANC195 targeted the gp120 CD4 bs (Science 333, 1633-1637 (2011)) and three, 10-
25 1074, PGT121, and PGT126 (Nature 477, 466-470 (2011)), were dependent on the presence of the HIV-1 gp120 N332 glycan. When tested against SHIVAD8EO, all three glycan-dependent mAbs exhibited greater potency than the CD4 bs mAbs (FIG. 11 A). The IC₅₀ values for the three mAbs targeting the gp120 N332 glycan ranged from 0.09 to 0.15 μ g/ml. The CD4 bs mAbs exhibited a much broader range (0.14 to 6.34 μ g/ml) of IC₅₀ neutralizing activity with 3BNC117
30 being the most potent. A similar hierarchy (glycan-dependent > CD4 bs dependent) of neutralizing mAb potency was also observed with SHIVDH12-V3AD8, but the neutralizing activity was

distributed across a much wider (>100 fold) range compared to the IC₅₀ values observed for SHIVAD8EO (FIG. 11B). SHIVDH12-V3AD8 was somewhat more sensitive to the glycan targeting mAbs and more resistant to the CD4 bs neutralizing mAbs than SHIVAD8EO.

Based on the results shown in FIG. 11, five neutralizing mAbs were selected for a pre-exposure passive transfer study: VRC01, because it was the first CD4bs NAb of the newly isolated broadly acting NAbs to be characterized; the CD4 bs mAbs 45-46m2 and 3BNC117, both of which exhibited strong neutralizing activity against SHIVAD8EO and SHIVDH12-V3AD8; and the gp120 N332 glycan-dependent mAbs, PGT121 and 10-1074.

The protocol for passive transfer experiments was to administer decreasing amounts of neutralizing mAbs intravenously and challenge animals intrarectally 24h later. The goal was to block virus acquisition, coupled with the knowledge that repeated administrations of humanized anti-HIV mAbs to individual macaques could reduce their potency and/or possibly induce anaphylactic responses, a SHIV challenge dose of sufficient size to establish an *in vivo* infection following a single inoculation was chosen. In this regard, we had previously conducted intrarectal titrations of SHIVAD8 in rhesus monkeys and reported that the inoculation of 1×10^3 TCID₅₀, determined by endpoint dilution in rhesus macaque PBMC, was equivalent to administering approximately 3 animal infectious doses₅₀ (AID₅₀) (J. of virology 86, 8516-8526 (2012)). In fact, single intrarectal inoculations of 3 AID₅₀ have resulted in the successful establishment of infection in 10 of 10 rhesus macaques with SHIVAD8EO or SHIVDH12-V3AD8.

As a control for the first passive transfer experiment, an anti-dengue virus NS1 IgG1 mAb was administered intravenously to animals, which were challenged with SHIVAD8EO 24h later. Both monkeys (ML1 and MAA) rapidly became infected, generating peak levels of plasma viremia at week 2 PI. VRC01 was the first anti-HIV-1 neutralizing mAb tested for protection against virus acquisition and was administered to two macaques at a dose of 50 mg/kg. One (DEGF) of the two inoculated macaques was completely protected from the SHIVAD8EO challenge, with no evidence of plasma viremia or cell-associated viral DNA over a 45 week observation period. The other recipient of 50 mg/kg VRC01 (DEH3) became infected, but peak plasma viremia was delayed until week 5 PI. Two additional macaques administered lower amounts (20 mg/kg) of VRC01 were not protected from the SHIVAD8EO challenge. These results are summarized in Table 13.

Examined next, the protective properties of PGT121 against a SHIVAD8EO challenge. PGT121 was one of the most potent glycan targeting neutralizing mAbs measured in the TZM-bl assay (**FIG. 11**). Based on the results obtained with VRC01, *in vivo* PGT121 mAb titration at 20 mg/kg was chosen to begin with. The two challenged monkeys (KNX and MK4) resisted the SHIVAD8EO challenge. When lower amounts (*viz.* 5 mg/kg, 1 mg/kg, or 0.2 mg/kg) of PGT121 were administered, 1 of 2, 2 of 2, and 0 of 2 animals, respectively, were protected (Table 13).

The capacity of VRC01 and PGT121 mAbs to block SHIVDH12-V3AD8 acquisition was similarly evaluated (Table 13). The results obtained with VRC01 were comparable to those observed with the SHIVAD8EO challenge: 1 of 2 recipients of 30 mg/kg was protected from the establishment of a SHIVDH12-V3AD8 infection. The PGT121 mAb was considerably more potent than VRC01 in preventing SHIVDH12-V3AD8 acquisition: 2 of 2 recipients of 0.2 mg/kg PGT121 resisted infection. PGT121 also appeared to be somewhat more effective in preventing SHIVDH12-V3AD8 versus SHIVAD8EO *in vivo* infections (Table 13). This result is consistent with the 8-fold difference in IC₅₀ values for PGT121 for neutralizing the two SHIVs in *in vitro* assays (**FIG. 11**).

The results of passively transferring 10-1074, 3BNC117, or 45-46m2 neutralizing mAbs to rhesus monkeys, followed by a challenge with either SHIVAD8EO or SHIVDH12-V3AD8, are summarized in Table 13. The 10-1074 mAb potently blocked the *in vivo* acquisition of both SHIVs. The CD4bs 3BNC117 and 45-46m2 mAbs were selected for passive transfer to macaques based on their IC₅₀ values against both SHIVs in the *in vitro* neutralization experiments shown in **FIG. 11**. 3BNC117 successfully blocked SHIVAD8EO infection in 2 of 2 monkeys at 5 mg/kg but not in 2 other animals given a dose of 1 mg/kg (Table 13). This was similar to the results observed when the same amounts of 3BNC117 were administered to macaques challenged with SHIVDH12-V3AD8: 1 of 2 became infected at 5 mg/kg; 1 of 2 became infected at 1 mg/kg.

Plasma samples collected at various times from passively transferred macaques were analyzed by HIV-1 gp120 ELISA to determine neutralizing mAb concentrations. In general, the plasma concentrations of each mAb at the time of challenge (24h following antibody administration) correlated with the dose of antibody administered (Table 13).

The relationships of plasma mAb concentrations to *in vivo* protection are shown in **FIG. 12**. Of the 5 neutralizing mAbs evaluated, PGT121 was clearly the most effective against both viruses, with SHIVDH12-V3AD8 exhibiting somewhat greater sensitivity to this mAb (2 of 2

monkeys protected at a plasma concentration of 0.2 µg/ml). In contrast, a plasma concentration of nearly 400 µg/ml of VRC01 was required to protect 1 of 2 animals against the same SHIVDH12-V3AD8 challenge virus (Table 13). The most potent CD4 bs mAb administered to macaques in this study, 3BNC117, was approximately 6 to 10-fold more effective than VRC01 in preventing the acquisition of either SHIV (FIG. 12, Table 13).

The calculated half lives of PGT121, 10-1074, 3BNC117, and VRC01 mAbs were quite similar: 3.5 days, 3.5 days, 3.3 days, and 3.1 days, respectively. In contrast, the half-life of 45-46m2 was extremely short and could not be determined. Based on the plasma mAb concentrations in several macaques 24h following the administration of 20 mg/kg of humanized neutralizing mAbs (*viz.* approximately 250 µg/ml [Table 13]), the two monkeys receiving 20 mg/kg of 45-46m2 had plasma mAb concentrations of only 15.0 and 17.6 µg/ml, a decay of more than 95% relative to other neutralizing mAbs in 24 h.

Neutralization titers were measured on plasma samples collected 24h following mAb administration when the macaques were challenged with SHIVAD8EO or SHIVDH12-V3AD8. As shown in Table 13, good correlation was observed between anti-viral plasma neutralization titers and protection from SHIV infection. The administration of the two glycan-dependent mAbs (PGT121 and 10-1074) clearly resulted in the highest titers of anti-HIV-1 neutralizing activity at the time of virus challenge. The titers measured in recipients of the 45-46m2 mAb were at the limits of detection or undetectable due to its extremely short half-life *in vivo*.

The method described by Reed and Muench (Am J Hyg 27, 493-497 (1938)) was used to calculate the neutralization titers, measured in plasma, needed to prevent virus acquisition in 50% of challenged monkeys. These protective titers for the 28 monkeys, challenged with SHIVAD8EO, or the 32 monkeys, challenged with SHIVDH12-V3AD8, were separately deduced (Tables 15 and 16). The plasma neutralization titers required for protecting 50% of the SHIVAD8EO or SHIVDH12-V3AD8 challenged animals were calculated to be 1:115 and 1:96, respectively. Because these similar titers were obtained following: 1) SHIV challenges by identical routes and inoculum size and 2) the administration of the same ensemble of neutralizing mAbs, the neutralization data from all 60 animals were combined and subjected to probit regression to examine the relationship between plasma neutralization titers and *in vivo* protection. As a further check, when a term for the SHIV virus was included in the probit regression model on all 60 macaques, there was no evidence of a difference between the two SHIV viruses

($p=0.16$). When applied to the entire group of 60 macaques, probit regression estimated that plasma neutralization titers of 1:104 would prevent virus acquisition in 50% of animals. Probit analysis of the data also estimates that 50% plasma neutralization titers of 1:57 or 1:329 would protect 33% or 80%, respectively, of exposed animals.

5 EXAMPLE 10 *Administration of neutralizing mAbs to chronically infected HIV in-vivo models*

Methods Summary: The neutralization activities of the broadly acting 3BNC11724 and 10-107423 neutralizing mAbs against SHIVAD8EO were initially determined in the TZM-bl cell system against SHIVAD8EO. Their capacities to block virus acquisition or to control plasma viremia in chronically infected animals challenged with the R5-tropic SHIVAD8EO were
10 assessed by monitoring plasma viral loads and cell-associated viral nucleic acids; levels of CD4+ T cell subsets were measured by flow cytometry. SGA analyses of circulating viral variants and the determination of antibody levels in plasma. Plasma concentration of NAbs was determined by measuring neutralizing activity against HIV-1 pseudovirus preparations only susceptible to either 10-1074 or 3BNC117.

15 *Results:*

Two groups of chronically infected macaques were assessed. The first group consisted of two clinically asymptomatic animals (DBZ3 and DC99A) that had been infected for 159 weeks and had sustained similar and significant declines of circulating CD4+ T cells (Table 17). The regimen for treating ongoing SHIV infections was to co-administer 101074 and 3BNC117, at a
20 dose of 10mg/kg. At the time of mAb administration, the plasma viral loads in macaques DBZ3 and DC99A were 1.08×10^4 and 7.6×10^3 RNA copies/ml, respectively. Both monkeys responded to combination anti-HIV-1 mAb treatment with immediate and rapid reductions of plasma viremia to undetectable levels within 7 to 10 days. Suppression of measurable SHIVAD8EO in the plasma of macaques DBZ3 and DC99A, following a single administration of
25 the two mAbs, lasted 27 and 41 days, respectively. In each case, plasma viremia rebounded to pretreatment levels.

A second group of three animals (DBX3, DCF1, and DCM8), each of which were also infected with SHIVAD8EO for more than 3 years and were clinically symptomatic with intermittent diarrhea and or anorexia, were treated with the two neutralizing antibodies (Table 17).
30 At the time of mAb administration, the level of circulating CD4+ T cells in macaque DCM8 was only 43 cells/ μ l and somewhat higher in animals DCF1 (105 cells/ μ l) and DBXE (158 cells/ μ l).

Plasma viral loads exceeded 10^5 RNA copies/ml in animals DBXE and DCF1 and were significantly lower (1.59×10^3 RNA copies/ml) in monkey DCM8. The administration of the two mAbs to monkey DBXE resulted in a biphasic reduction of viremia from 2.0×10^5 RNA copies at day 0 to undetectable levels in plasma at day 20. This was followed, within a few days, by a resurgence of high levels of circulating virus in DBXE. Macaque DCM8, with more modest plasma virus loads and very low numbers of circulating CD4⁺ T cells, experienced a rapid decline of viremia to undetectable levels between days 6 and 20 following the initiation of mAb treatment. Finally, animal DCF1, previously reported to have generated broadly reacting anti-HIV-1 NAb, exhibited a transient and a comparatively modest 27-fold reduction of plasma viremia by day 6 in response to combination mAb therapy, before the viral loads returned to high pretreatment levels.

PBMC associated viral RNA and DNA levels were also determined prior to and following antibody administration (Table 18). For each animal, mAb treatment resulted in reduced levels of cell associated viral RNA, correlating well with the plasma viral load measurements. No consistent pattern was observed for cell associated viral DNA levels as a result of antibody treatment. Administration of neutralizing mAbs to chronically SHIVAD8EO infected monkeys also had beneficial effects on circulating CD4⁺ T cell levels, particularly in animals with very high virus loads. The CD4⁺ T cell numbers in macaques DBXE and DCF1 increased 2 to 3 fold during the period of mAb mediated virus suppression, but gradually declined to pretreatment levels as viremia again became detectable.

Plasma concentrations of each mAb were determined by measuring the plasma neutralizing activity against selected HIV-1 pseudovirus strains sensitive to one or the other, but not to both antibodies (FIG. 13A). In every treated animal, suppression of SHIVAD8EO viremia was maintained until a threshold plasma mAb concentration of approximately 1 to 3 μ g/ml was reached (FIGS. 13B and 13C). This was even the case for macaque DCF1, for which a modest and transient reduction of plasma viral RNA levels was observed. Interestingly, the mAbs administered to clinically symptomatic macaques DCM8 and DCF1 had shortened half-lives or were undetectable. As noted earlier, macaque DCM8 had extremely low CD4⁺ T cell levels (43 cells/ μ l plasma) and macaque DCF1 had to be euthanized on day 56 post treatment initiation due to its deteriorating clinical condition. A necropsy of DCF1 revealed severe enteropathy, characterized by disseminated gastrointestinal cryptosporidiosis, pancreatitis, and cholangitis.

SGA analysis was used to determine whether amino acid substitutions had arisen in gp120 regions previously shown to affect the sensitivity to 10-1074 or 3BNC117 mAbs. In each case the rebound virus present in plasma following immunotherapy was unchanged. To further test the sensitivity of the re-emerging viruses, 10-1074 plus 3BNC117 combination therapy (10 mg/kg of each) was re-administered to the two clinically asymptomatic monkeys (DBZ3 and DC99A). The viral loads in each animal again rapidly fell, becoming undetectable at day 7 of the second immunotherapy cycle. Viremia was suppressed for 7 days in macaque DBZ3 and more than 21 days in monkey DC99A. Taken together, these results suggest that the re-emergence of virus following the first treatment cycle in these two animals represented insufficient mAb levels in vivo rather than antibody selected virus resistance.

Table 3

Repertoire of PGT121 and 10-1074 clonal variants

pt10-mAbid	YH	DH	CH	CDR3 ¹	VHmut	Length ²	(-) (+)	Y	LC	VL	DL	CDR3 ³	VHmut	FRW1 _{del}	FRW2 _{ins}	Length ²	(-) (+)	Y		
10-180	4-59	3-3-15	5	ARRDQRIVGVVFQGFFFYYISKDV	25	24	2	3	4	?	?	?	?	?	?	?	?	?		
10-185	4-59	3-3-15	5	ARRDQRIVGVVFQGFFFYYISKDV	52	24	2	3	4	?	3-21	3	RRNDGRSGPFSSNS	47	12	3	12	?	2	5
10-248	4-59	3-3-16	6	ARRDQRIVGVVFQGFFFYYISKDV	46	34	2	3	4	?	?	?	?	?	?	?	?	?		
10-259	4-59	?	5	THDGRIVGVVAFQGFFFYYISKDV	53	24	2	4	3	?	3-21	3	RRNDGRSGTFSSNS	58	21	3	12	?	2	?
10-266	4-59	3-3-16	6	ARRDQRIVGVVFQGFFFYYISKDV	46	34	2	3	4	?	3-21	3	RRNDGRSGPFSSNS	46	12	3	12	?	2	5
10-267	4-59	?	6	ARRDQRIVGVVFQGFFFYYISKDV	58	24	2	2	5	?	?	?	?	?	?	?	?	?		
10-302	4-59	3-3-15	5	TLHGRIVGVVAFQGFFFYYISKDV	54	24	2	3	3	?	3-21	3	RRNDGRSGTFSSNS	52	23	3	12	?	3	5
10-324	4-59	3-3-16	6	ARRDQRIVGVVFQGFFFYYISKDV	48	24	2	3	4	?	?	?	?	?	?	?	?	?		
10-417	4-59	3-10-3	5	ALHGRIVGVVAFQGFFFYYISKDV	63	34	2	3	3	?	3-21	3	RRNDGRSGTFSSNS	44	21	3	12	?	3	5
10-418	4-59	3-3-15	5	ARRDQRIVGVVFQGFFFYYISKDV	47	24	2	3	4	?	3-21	3	RRNDGRSGPFSSNS	45	12	3	12	?	2	5
10-498	4-59	3-3-16	6	ARRDQRIVGVVFQGFFFYYISKDV	47	34	2	3	4	?	3-21	3	RRNDGRSGPFSSNS	46	12	3	12	?	2	5
10-543	4-59	3-10-3	5	ALHGRIVGVVAFQGFFFYYISKDV	68	24	2	3	3	?	?	?	?	?	?	?	?	?		
10-570	4-59	3-3-16	6	ARRDQRIVGVVFQGFFFYYISKDV	42	34	2	3	4	?	?	?	?	?	?	?	?	?		
10-621	4-59	3-3-15	5	TLHGRIVGVVAFQGFFFYYISKDV	54	24	2	3	3	?	?	?	?	?	?	?	?	?		
10-654	4-59	3-3-16	6	ARRDQRIVGVVFQGFFFYYISKDV	47	34	2	3	4	?	?	?	?	?	?	?	?	?		
10-730	4-59	3-3-16	6	ARRDQRIVGVVFQGFFFYYISKDV	45	24	2	3	4	?	?	?	?	?	?	?	?	?		
10-730	4-59	3-3-16	6	ARRDQRIVGVVFQGFFFYYISKDV	46	34	2	3	4	?	?	?	?	?	?	?	?	?		
10-814	4-59	3-3-15	5	TLHGRIVGVVAFQGFFFYYISKDV	43	24	2	3	4	?	3-21	3	RRNDGRSGTFSSNS	52	16	3	12	?	3	5
10-847	4-59	3-3-16	6	ARRDQRIVGVVFQGFFFYYISKDV	47	34	2	3	4	?	3-21	3	RRNDGRSGPFSSNS	46	12	3	12	?	2	5
10-943	4-59	3-3-15	5	TLHGRIVGVVAFQGFFFYYISKDV	58	24	2	3	3	?	3-21	3	RRNDGRSGTFSSNS	46	21	3	12	?	3	5
10-996	4-59	3-3-10	6	TLHGRIVGVVAFQGFFFYYISKDV	49	34	2	3	4	?	3-21	3	RRNDGRSGTFSSNS	50	16	3	12	?	3	5
10-1022	4-59	3-3	5	ARRDQRIVGVVFQGFFFYYISKDV	51	24	2	3	4	?	?	?	?	?	?	?	?	?		
10-1053	4-59	3-3-16	6	THHGRIVGVVAFQGFFFYYISKDV	59	34	2	4	3	?	3-21	3	RRNDGRSGTFSSNS	46	21	3	12	?	3	5
10-1074	4-59	3-3-15	5	ARRDQRIVGVVFQGFFFYYISKDV	45	24	2	3	4	?	3-21	3	RRNDGRSGPFSSNS	45	12	3	12	?	2	5
10-1121	4-59	3-10-3-3	5	ALHGRIVGVVAFQGFFFYYISKDV	63	24	2	3	3	?	3-21	3	RRNDGRSGTFSSNS	42	21	3	12	?	3	5
10-1130	4-59	3-10-3-2	5	ALHGRIVGVVAFQGFFFYYISKDV	65	34	2	3	3	?	3-21	3	RRNDGRSGTFSSNS	42	21	3	12	?	3	5
10-1141	4-59	3-10-3-3	5	ALHGRIVGVVAFQGFFFYYISKDV	53	24	2	3	3	?	3-21	3	RRNDGRSGTFSSNS	46	21	3	12	?	3	5
10-1142	4-59	3-3-15	5	ARRDQRIVGVVFQGFFFYYISKDV	38	24	2	3	5	?	3-21	3	RRNDGRSGTFSSNS	61	16	3	12	?	2	?
10-1188	4-59	3-3	5	ARRDQRIVGVVFQGFFFYYISKDV	46	24	2	3	4	?	3-21	3	RRNDGRSGPFSSNS	46	12	3	12	?	2	5
10-1197	4-59	3-3-15	5	ARRDQRIVGVVFQGFFFYYISKDV	44	24	2	3	4	?	?	?	?	?	?	?	?	?		
10-1223	4-59	3-3-16	6	ARRDQRIVGVVFQGFFFYYISKDV	47	34	2	3	4	?	?	?	?	?	?	?	?	?		
10-1233	4-59	3-10-3-3	5	ALHGRIVGVVAFQGFFFYYISKDV	58	24	2	3	3	?	3-21	?	?	?	?	?	?	?		
10-1248	4-59	3-3-15	5	ARRDQRIVGVVFQGFFFYYISKDV	45	24	2	3	4	?	3-21	3	RRNDGRSGPFSSNS	45	12	3	12	?	2	5
10-1294	4-59	3-3-16	6	ARRDQRIVGVVFQGFFFYYISKDV	46	34	2	3	4	?	3-21	3	RRNDGRSGPFSSNS	46	12	3	12	?	2	5
10-1341	4-59	3-3-15	5	ARRDQRIVGVVFQGFFFYYISKDV	45	24	2	3	4	?	3-21	3	RRNDGRSGPFSSNS	45	12	3	12	?	2	5
10-1342	4-59	3-3-16	6	ARRDQRIVGVVFQGFFFYYISKDV	46	34	2	3	4	?	3-21	3	RRNDGRSGPFSSNS	46	12	3	12	?	2	5
10-1357	4-59	3-3-16	6	THHGRIVGVVAFQGFFFYYISKDV	57	24	2	4	3	?	3-21	3	RRNDGRSGTFSSNS	43	21	3	12	?	3	?
10-1375	4-59	3-3-16	6	ARRDQRIVGVVFQGFFFYYISKDV	47	34	2	3	4	?	3-21	3	RRNDGRSGPFSSNS	46	12	3	12	?	2	5

VHmut and VHmut indicate the total number of mutations in the VH and VL, Ig genes. (-) and (+) indicate the number of negatively and positively charged amino acids in the Ig complementary determin region (CDR3), respectively. Y indicates the number of Tyrosine residues in the IgHL CDR3s (in green). Based on Kabat nomenclature (JBL457, <http://www.ncbi.nlm.nih.gov/jbl457/>). Acidic and basic are in red and blue, respectively. FRW1_{del}, number of deleted nucleotides in framework region 1 (FRW1) of the IgL; FRW2_{ins}, number of inserted nucleotides in framework region 2 (FRW2) of the IgL. Color shading shows clonal members with identical IgH sequences and among them, IgL sequence identity that defines clones is indicated in bold. ? indicates the representative antibody variants that produced and analyzed, 10-265 IgL was not cloned, and 10-1141 IgG was not produced.

Table 4

In vitro TZM-bl neutralization assay on the basic panel

IC ₅₀	10-1369	10-259	PGT121	10-302	10-410	10-1120	10-1121	10-1146	10-996	10-1341	10-947	10-1074
BaL.26	0.009	0.001	0.02	0.045	0.018	0.013	0.043	0.064	0.046	0.032	0.003	0.003
S51196.1	0.003	0.012	0.003	0.015	0.006	0.006	0.009	0.007	0.007	0.011	0.009	0.016
6535.3	0.003	0.005	0.007	0.014	0.003	0.003	0.009	0.003	0.016	0.009	0.011	0.007
QH0692.42	0.503	0.155	1.085	3.122	2.630	4.971	4.137	0.500	0.395	0.335	0.259	0.259
TRJ04551.58	0.589	0.189	3.898	14.401	12.511	38.888	15.380	0.548	0.516	0.333	0.210	0.170
SC422861.8	0.195	0.096	0.283	0.333	0.132	0.070	0.173	0.195	0.255	0.189	0.137	0.145
PVO.4	0.225	0.175	0.147	0.670	0.494	0.385	0.570	0.310	0.211	0.238	0.172	0.178
CAAN5342.A2	0.070	0.003	0.013	0.020	0.012	0.009	0.003	0.003	0.007	0.009	0.006	0.007
YU-2	0.210	0.135	0.098	0.190	0.089	0.078	0.152	0.278	0.258	0.234	0.161	0.143
R1168.c1	> 40	> 40	> 40	> 40	> 40	> 40	> 40	> 40	> 40	> 40	> 40	> 40
MuLV	> 40	> 40	> 40	> 40	> 40	> 40	> 40	> 40	> 40	> 40	> 40	> 40

IC ₈₀	10-1369	10-259	PGT121	10-302	10-410	10-1120	10-1121	10-1146	10-996	10-1341	10-947	10-1074
BaL.26	0.288	0.101	0.08	0.155	0.068	0.082	0.154	0.203	0.228	0.189	0.113	0.124
S51196.1	0.003	0.007	0.006	0.005	0.003	0.007	0.005	0.013	0.003	0.040	0.006	0.027
6535.3	0.003	0.002	0.001	0.003	0.001	0.010	0.003	0.008	0.005	0.008	0.004	0.004
QH0692.42	1.714	0.551	14.976	18.122	12.571	> 40	21.943	1.883	1.404	1.100	0.808	0.881
TRJ04551.58	3.818	0.905	38.930	> 40	> 40	> 40	> 23	3.804	4.265	1.236	0.768	0.893
SC422861.8	0.940	0.333	0.714	1.156	0.449	0.264	0.741	0.883	0.845	0.501	0.358	0.362
PVO.4	0.787	0.716	1.097	2.189	1.572	1.793	2.485	1.319	1.715	0.784	0.774	0.788
CAAN5342.A2	0.168	0.003	0.006	0.002	0.003	0.005	0.005	0.008	0.003	0.004	0.006	0.004
YU-2	0.738	0.382	0.356	0.582	0.243	0.213	0.340	0.758	0.881	0.766	0.537	0.398
R1168.c1	> 40	> 40	> 40	> 40	> 40	> 40	> 23	> 40	> 40	> 40	> 40	> 40
MuLV	> 40	> 40	> 40	> 40	> 40	> 40	> 23	> 40	> 40	> 40	> 40	> 40

Numbers indicate antibody (IgG) concentrations in $\mu\text{g/ml}$ to reach the IC₅₀ (top) and IC₈₀ (bottom) in the TZM-bl neutralization assay. IC_{50/80} values are color coded and indicate from dark green to dark red, an increasing neutralization sensitivity. > indicates that the IC₅₀ for a given virus was not reached at the concentration tested. Murine leukemia virus (MuLV) and R1168.c1 (clade AE) are negative controls.

Table 5

In vitro TZM-bl neutralization assay on the extended panel - IC₅₀ values

Virus ID	Clade	10-288	10-1074	PQY121	Virus ID	Clade	10-288	10-1074	PQY121
8535.3	B	0.007	0.014	0.008	CNE58	BQ	0.573	0.007	>50
Q40893.43	B	0.326	0.191	1.041	MS208.A1	A	>50	>50	>50
3042265.8	B	0.173	0.091	0.101	Q23.17	A	0.008	0.008	0.008
8V01.2	B	0.186	0.074	0.131	Q481.50	A	>50	>50	>50
TRO.11	B	0.005	0.008	0.005	Q789.022	A	>50	>50	>50
AC10.0.29	B	0.087	0.023	0.037	Q259.02.17	A	>50	>50	0.008
RHP44059.7	B	0.034	0.01	0.014	Q343.012	A	>50	>50	0.003
THR04158.12	B	>50	>50	>50	3419.41.1	A	0.008	>50	>50
REJ04541.57	B	>50	>50	0.007	3385.42.2	A	0.386	0.131	0.001
TRJ04551.58	B	1.147	0.172	0.718	3260.45.135	A	0.160	0.035	0.004
X0704160.33	B	0.539	0.185	0.459	191555.A.1	A (T/F)	>50	>50	>50
044N5343.A2	B	0.015	0.007	0.011	191084.B7.15	A (T/F)	0.087	0.003	0.043
YLA.2	B	0.155	0.143	0.039	300402.A2.4	A (T/F)	0.012	0.01	0.003
WEAL.015.410.757	B (T/F)	0.147	0.104	0.083	7257.51	CRF01_AG	>50	>50	>50
1506.11.03.1601	B (T/F)	0.003	0.003	0.003	909.38	CRF02_AG	1.033	0.047	>50
1054.07.704.1495	B (T/F)	0.363	0.139	0.115	263.8	CRF01_AG	10.919	0.656	1.047
1506.10.741.1805	B (T/F)	0.117	0.038	0.068	7251.4	CRF02_AG	>50	>50	>50
1012.11.703.1257	B (T/F)	0.015	0.008	0.008	7251.18	CRF01_AG	0.035	1.091	>50
6140.06.745.4630	B (T/F)	0.065	0.065	0.108	7275.50	CRF02_AG	14.010	2.145	>50
8044.13.55.4575	B (T/F)	0.353	0.203	0.249	7255.34	CRF01_AG	39.869	>50	0.708
82357.14.03.4598	B (T/F)	29.000	>50	1.036	211.9	CRF02_AG	0.750	0.112	1.459
8035.3011.2344	B (T/F)	0.064	0.043	0.053	325.47	CRF01_AG	0.109	0.000	0.333
D0158.10	C	0.015	0.015	0.007	620145.001	CRF01_AE	>50	>50	>50
D0171.17	C	0.173	0.121	0.115	CNE8	CRF01_AE	>50	>50	>50
D0423.1	C	0.055	0.044	0.009	C1080.003	CRF01_AE	>50	>50	>50
ZM197M.P57	C	>50	>50	>50	R1364.004	CRF01_AE	>50	>50	>50
ZM121M.P115	C	0.413	0.174	0.335	R1156.001	CRF01_AE	>50	>50	>50
ZM133M.P55	C	0.050	0.063	1.451	Q2101.001	CRF01_AE	>50	>50	>50
ZM124M.P11	C	>50	>50	>50	Q247.011	CRF01_AE	>50	>50	>50
ZM13M.P513	C	>50	>50	>50	Q4119.008	CRF01_AE	>50	>50	>50
ZM105M.P54	C	>50	>50	7.024	CNE5	CRF01_AE	>50	>50	>50
ZM135M.P1106	C	0.059	0.069	0.076	B1000900002.4	CRF01_AE	>50	>50	3.628
C4945.2.02.03	C	>50	>50	0.006	B10015000.11.5	CRF01_AE (T/F)	>50	>50	>50
C4910.2.02.05	C	24.783	>50	0.082	B1001000006.2	CRF01_AE (T/F)	>50	>50	>50
HW-101429-0.42	C	0.040	0.044	0.006	B10010000001.1	CRF01_AE (T/F)	>50	>50	>50
HW-1013035-0.11	C	0.180	>50	>50	B100100000.10.3	CRF01_AE (T/F)	>50	>50	>50
HW-10055-0.2	C	>50	>50	0.444	A1152.11	G	0.144	0.003	0.040
HW-15645-0.22	C	1.325	1.159	12.955	P0402.02.11	G	0.000	0.01	0.000
Ce1086_B0	C (T/F)	>50	>50	>50	X1254.13	G	0.121	0.085	0.055
Ce2393_C2	C (T/F)	>50	>50	>50	X0362.19	G	0.003	0.009	0.011
Ce1176_A3	C (T/F)	0.043	0.019	0.017	Q2121.01.055	G	0.010	0.015	0.010
Ce2010_F5	C (T/F)	>50	>50	>50	F1561.05.3	G	0.007	0.005	0.004
Ce2652_E4	C (T/F)	>50	>50	>50	X1632.02.010	G	>50	>50	>50
Ce1172_H1	C (T/F)	0.055	0.047	0.003	Q215.15.045	G	>50	>50	>50
Ce2050_09	C (T/F)	>50	>50	>50	A07412M1.1.1.12	G	0.005	0.00	0.00
Ce702010024.242	C (T/F)	>50	>50	>50	211565.001	G	>50	>50	>50
BP1255.431.2	C (T/F)	>50	>50	>50	211565.002	G	>50	>50	>50
349F.013	C (T/F)	0.050	0.023	0.003	191921.00.1	G (T/F)	>50	>50	>50
249M.B10	C (T/F)	>50	>50	>50	3517.42.059	GD	0.147	1.148	>50
ZM1247y1(Rev-)	C (T/F)	0.050	0.043	0.007	5450.44.125	GD	0.010	0.009	0.007
703010102155(Rev-)	C (T/F)	0.015	0.015	0.000	6552.41.120	GD	0.044	0.037	0.055
1394053(Rev-)	C (T/F)	0.050	0.063	0.456	5811.47.115	GD	0.001	0.000	0.004
Ce704802021.183	C (T/F)	0.243	0.135	0.035	65-F1_0_25	GD	>50	>50	>50
CNE19	BQ	1.453	0.106	0.018	3301.41.074	AD	0.016	0.019	0.014
CNE20	BQ	>50	>50	0.003	6041.42.123	AD	>50	>50	>50
CNE21	BQ	0.085	0.087	0.003	6543.44.01	AD	>50	>50	>50
CNE17	BQ	4.040	0.686	0.019	6545.44.01	AD	>50	>50	>50
CNE35	BQ	0.514	0.353	0.101	6515.43.03	ACD	0.061	0.030	0.003
CNE32	BQ	4.628	1.228	0.741	3103.43.110	ACD	0.003	0.007	0.043
CNE53	BQ	0.057	0.029	0.055					

Numbers indicate antibody (µg) concentrations or µg/ml to reach the IC₅₀ in the TZM-bl neutralization assay. IC₅₀ values are color-coded and indicate from dark green to dark red, an increasing neutralization sensitivity. > indicates that the IC₅₀ for a given virus was not reached at the concentration tested.

Table 6

In vitro TZM-bl neutralization assay on the extended panel - IC80 values

Virus ID	Clade	10-388	10-1874	PRT151	Virus ID	Clade	10-388	10-1874	PRT151
6E25.3	B	0.396	0.396	0.396	CNE53	BC	1.230	0.963	>50
QW0592.42	B	1.834	0.923	1.945	MS208.A1	A	>50	>50	>50
00470651.3	B	0.307	0.418	0.460	Q23.17	A	0.800	0.380	0.390
PR0.4	B	0.350	0.360	0.345	Q451.50	A	>50	>50	>50
TR0.11	B	0.391	0.397	0.390	Q732.022	A	>50	>50	>50
AD10.1.03	B	0.350	0.110	1.159	Q059.00.17	A	>50	>50	>50
RHFAA259.7	B	0.183	0.118	0.304	Q042.092	A	>50	>50	0.304
THRO4156.18	B	>50	>50	>50	3419.01.01	A	>50	>50	>50
REJQ4541.57	B	>50	>50	>50	3365.02.02	A	1.380	0.450	7.353
TRJQ4551.58	B	7.069	0.634	20.300	Q050.05.036	A	0.436	0.160	0.162
WITQ4150.33	B	6.484	2.110	6.077	191955.A11	A (T/F)	>50	>50	>50
CAAN5340.A2	B	0.379	0.330	0.361	191034.07-19	A (T/F)	0.144	0.108	0.138
YU-8	B	0.331	0.339	0.329	500430.03_4	A (T/F)	0.350	0.320	0.304
WEAU_01E_410_787	B (T/F)	0.402	0.175	0.295	T257-31	CRF01_AG	>50	>50	>50
1008_11_02_1801	B (T/F)	0.330	0.330	0.330	505-18	CRF01_AG	7.181	4.696	>50
1054_07_00_1499	B (T/F)	0.391	0.343	0.356	263-6	CRF01_AG	>50	6.327	24.376
1059_10_01_1826	B (T/F)	0.663	0.372	1.323	T255-4	CRF01_AG	0.354	0.355	>50
1012_11_00_1157	B (T/F)	0.111	0.029	0.338	T251-15	CRF01_AG	7.029	7.392	>50
0040_08_04_4610	B (T/F)	0.348	0.304	1.624	T278-10	CRF01_AG	>50	16.376	>50
0244_11_05_4575	B (T/F)	1.286	0.922	1.878	T255-34	CRF01_AG	>50	>50	>50
00357_04_03_4689	B (T/F)	>50	>50	40.009	211-9	CRF01_AG	0.343	0.425	6.843
0005_0511_2344	B (T/F)	0.114	0.113	0.175	T35-47	CRF01_AG	0.381	0.163	1.575
D0136.12	C	0.181	0.076	0.333	020345.031	CRF01_AE	>50	>50	>50
D0170.17	C	0.607	0.420	0.820	CNE3	CRF01_AE	>50	>50	>50
D0420.1	C	0.315	0.166	0.191	Q1080.003	CRF01_AE	>50	>50	>50
ZK197M.F87	C	>50	>50	>50	R0124.004	CRF01_AE	>50	>50	>50
ZK0144M.F115	C	3.251	3.357	3.150	R1165.001	CRF01_AE	>50	>50	>50
ZK0233M.F84	C	4.624	0.349	8.377	Q0101.001	CRF01_AE	>50	>50	>50
ZK0249M.F01	C	>50	>50	>50	Q0147.011	CRF01_AE	>50	>50	>50
ZK023M.F012	C	>50	>50	0.000	Q4118.002	CRF01_AE	>50	>50	>50
ZK029F.F84	C	>50	>50	>50	CNE5	CRF01_AE	>50	>50	>50
ZK125M.F108	C	0.653	0.367	1.825	B00002000.00.4	CRF01_AE	>50	>50	0.000
CAP45.2.00.03	C	>50	>50	6.544	B000015000.01.5	CRF01_AE (T/F)	>50	>50	>50
CAP010.1.00.08	C	>50	>50	>50	B000010000.02.3	CRF01_AE (T/F)	>50	>50	>50
H01-001438-2.42	C	0.104	0.161	0.153	B000005000.01.1	CRF01_AE (T/F)	>50	>50	>50
H01-0010095-0.11	C	>50	>50	>50	B000002000.02.3	CRF01_AE (T/F)	>50	>50	>50
H01-16055-2.3	C	>50	>50	4.050	X1153_01	G	0.461	0.475	0.301
H01-16045-2.12	C	9.923	5.636	>50	PE001_02_11	G	0.063	0.038	0.028
C01086_01	C (T/F)	>50	>50	0.000	X1054_03	G	0.400	0.397	0.169
C01083_03	C (T/F)	>50	>50	>50	X0065_09	G	0.014	0.018	0.009
C01176_03	C (T/F)	0.136	0.073	0.038	X0111_01_05	G	0.085	0.054	0.063
C02010_05	C (T/F)	>50	>50	>50	P1001_05_3	G	0.018	0.011	0.014
C02001_04	C (T/F)	>50	>50	>50	X1610_02_010	G	>50	>50	>50
C01172_01	C (T/F)	0.178	0.155	0.029	0016.04.045	D	>50	>50	>50
C02000_09	C (T/F)	>50	>50	>50	A074 (2M) 0010	D	0.070	0.048	0.406
C0200000254_0A2	C (T/F)	>50	>50	>50	001955.001	D	>50	>50	>50
BF1365.4103	C (T/F)	>50	>50	>50	221955.002	D	>50	>50	>50
245F C1G	C (T/F)	0.070	0.111	0.287	191621_01_1	D (T/F)	>50	>50	>50
249M B10	C (T/F)	>50	>50	>50	1917_01_059	DD	0.619	0.660	>50
ZK0247M (Rev-)	C (T/F)	0.050	0.181	0.156	8490.04.025	DD	0.049	0.041	0.073
T03010300103 (Rev-)	C (T/F)	0.044	0.031	0.049	8952.01.020	DD	0.185	0.128	0.505
13540560 (Rev-)	C (T/F)	0.028	0.181	0.070	8511.07.019	DD	0.011	0.016	0.017
C0204905221_1B3	C (T/F)	1.208	0.596	0.480	89-F1_02_25	DD	>50	>50	>50
CNE19	BC	>50	>50	0.163	1301.01.024	AC	0.024	0.043	0.043
CNE20	BC	0.000	0.000	0.000	0041.03.023	AC	>50	>50	>50
CNE21	BC	0.335	0.181	0.261	0540.04.01	AC	>50	>50	>50
CNE17	BC	29.701	13.287	>50	0545.04.01	AC	>50	>50	>50
CNE30	BC	1.298	1.220	0.555	0915.03.03	ACD	0.251	0.138	0.105
CNE51	BC	12.024	13.147	22.006	0125.03.015	ACD	0.150	0.101	0.113
CNE53	BC	0.323	0.141	0.200					

Numbers indicate antibody (µg) concentrations in µg/ml to reach the IC₈₀ in the TZM-bl neutralization assay. IC₈₀ values are color coded and indicate from dark green to dark red, an increasing neutralization sensitivity. > indicates that the IC₈₀ for a given virus was not reached at the concentration tested.

Table 7

Neutralization sensitivity according to N332 PNGS

Virus ID	Class	N332	S334	10-296	10-3074	P21121	Virus ID	Class	N332	S334	10-296	10-3074	P21121
8535.3	B	.	.				QNEE8	BC	.	(T)			
QNEE82.42	B	.	.				QNEE8.A1	A	.	.			
Q0412691.3	B	.	.				Q23.17	A	.	(T)			
PV01.4	B	.	.				Q441.42	A	V	N			
TRO.11	B	.	.				Q755.622	A	.	N			
AD180.22	B	.	.				Q259.62.17	A	T	N			
RHFA4153.7	B	.	.				Q542.612	A	.	N			
TAR04155.18	B	T	N				Q415.61.61	A	.	.			
RE304541.67	B	T	N				Q555.62.22	A	.	.			
TRJ04551.36	B	.	.				Q160.65.636	A	.	.			
INT04160.32	B	.	.				181952_A11	A (TF)	N	N			
CAAN5342.42	B	.	.				181050_57-15	A (TF)	.	.			
YU-2	B	.	.				901483_A3_4	A (TF)	.	.			
WEAU_015_A10_787	B (TF)	.	.				T257-31	CRF01_AG	N	N			
Y05_11_03_1501	B (TF)	.	.				921-05	CRF01_AG	.	.			
Y054_07_T04_1459	B (TF)	.	.				261-8	CRF01_AG	.	.			
Y056_10_TA11_1626	B (TF)	.	.				T252-4	CRF01_AG	.	.			
Y012_11_T021_1227	B (TF)	.	.				T251-18	CRF01_AG	.	.			
6245_05_TA5_4523	B (TF)	.	.				T078-52	CRF01_AG	.	.			
6244_12_65_4575	B (TF)	.	.				T255-34	CRF01_AG	.	.			
62357_14_D1_4565	B (TF)	.	.				211-9	CRF01_AG	.	.			
6005_0011_2344	B (TF)	.	.				235-47	CRF01_AG	.	.			
D0126.12	C	.	.				920345.031	CRF01_AE	N	N			
D0172.17	C	.	.				QNE9	CRF01_AE	N	N			
D0422.1	C	.	.				Q1080.105	CRF01_AE	N	N			
DN197M.P87	C	D	.				R2154.104	CRF01_AE	N	N			
DN214M.PL18	C	.	.				R1155.021	CRF01_AE	N	N			
DN213M.P86	C	.	.				Q2101.021	CRF01_AE	N	N			
DN249M.PL1	C	.	.				Q3347.511	CRF01_AE	N	N			
DN453M.P512	C	.	N				Q4118.102	CRF01_AE	N	D			
DN103F.P84	C	N	N				QNE5	CRF01_AE	N	N			
DN115M.PL103	C	.	.				9100028002.02.4	CRF01_AE	N	N			
QAP45.1001.03	C	.	N				91000215002.11.5	CRF01_AE (TF)	N	N			
QAPT10.1001.05	C	.	.				91000210002.06.2	CRF01_AE (TF)	N	N			
H01-201429-2.42	C	.	.				91000225002.01.1	CRF01_AE (TF)	N	N			
H01-2013055-2.13	C	.	.				9100022002.10.3	CRF01_AE (TF)	.	.			
H01-16355-2.3	C	.	N				X1193_01	G	.	.			
H01-16345-2.22	C	.	.				F0420_02_11	G	.	.			
C01036_02	C (TF)	.	N				X1254_03	G	.	.			
C02391_03	C (TF)	.	D				N0088_05	G	.	(T)			
C01175_A3	C (TF)	.	.				X2131_01_85	G	.	.			
C02010_F5	C (TF)	T	N				F1261_05_3	G	.	.			
C03692_54	C (TF)	.	.				X1432_02_543	G	.	N			
C01173_H1	C (TF)	.	.				3215.65.645	D	T	D			
C02060_58	C (TF)	.	(T)				AE7412M1.0212	D	.	.			
C0203010054_0A2	C (TF)	D	N				221565.031	D	.	.			
BF1065.4312	C (TF)	T	N				231955.032	D	.	.			
245F.013	C (TF)	.	.				191821_E6_7	D (TF)	.	.			
245M.513	C (TF)	.	.				3517.02.058	DD	.	.			
DN247.115F01	C (TF)	.	(T)				6453.04.025	DD	.	.			
T03010220155(F01)	C (TF)	.	.				6552.01.022	DD	.	.			
13540301(F01)	C (TF)	.	.				6511.07.018	DD	.	.			
C0704029211_187	C (TF)	.	.				89-F1_2_25	DD	T	.			
QNE19	BC	.	.				3201.01.014	AC	.	(T)			
QNE25	BC	.	.				5541.01.012	AC	D	N			
QNE21	BC	.	.				6543.04.01	AC	T	N			
QNE17	BC	.	.				5545.04.01	AC	T	N			
QNE10	BC	.	.				Q215.03.02	AGD	.	.			
QNE52	BC	.	.				3103.03.010	AGD	.	.			
QNE13	BC	.	.										

Cells indicate the presence of an asparagine and of a serine residue in position 332 and 334, respectively. Mutations at positions 332 and 334 (H352 sequence numbering) are indicated by the substituting amino acid. IC50 values are color coded and indicate from dark green to dark red, an increasing neutralization sensitivity in the TZM-bl assay.

Table 8

In vitro PBMC-based neutralization assay

Further, please indicate whether you intend to use the χ^2 or the GFI-based goodness-of-fit measures to determine and indicate that the model is not well representing the data. Indicate that the χ^2 is a poor measure of fit for the measurement model GFI not determined.

Table 9

Data collection and refinement statistics (molecular replacement)

	PGT121 Fab "unliganded"	10-1074 Fab	GL Fab	PGT121 Fab "liganded"
Data collection				
Space group	P2 ₁ 2 ₁ 2 ₁	P2 ₁	P2 ₁	P2 ₁ 2 ₁ 2 ₁
Cell dimensions (Å)	58.75, 74.87, 114.917	61.38, 40.36, 84.46	54.93, 344.74, 55.23	67.79, 67.79, 94.11
α, β, γ (°)	90.00, 90.00, 90.00	90.00, 95.39, 90.00	90.00, 91.95, 90.00	90.00, 90.00, 90.00
Resolution (Å)	2.78-35.5 (2.78-2.93)	1.80- 36.31 (1.80-1.91)	2.42-38.60 (2.42-2.55)	2.33-38.66 (2.33-2.47)
R _{merge}	0.099 (0.293)	0.075 (0.558)	0.072 (0.482)	0.161 (0.603)
I / σ _i	8.8 (3.1)	8.7 (1.8)	11.0 (1.9)	6.7 (2.9)
Completeness (%)	96.7 (84.8)	93.49 (98.0)	95.5 (80.1)	92.2 (98.9)
Redundancy	3.2 (2.7)	2.7 (2.8)	3.1 (2.6)	5.3 (5.6)
Refinement				
Resolution (Å)	3.0	1.9	2.42	2.4
No. reflections	10,076	31,363	74,237	16,831
R _{work} / R _{free}	0.216/0.264	0.167/0.223	0.134/0.237	0.201/0.249
No. atoms				
Protein	3,276	3,346	12,881	3,127
Ligand/ion	0	0	0	129
Water	0	300	527	203
B-factors				
Protein	32.78	29.17	44.67	31.48
Ligand/ion	-	-	-	45.1
Water	-	37.37	40.27	36.78
R.m.s. deviations				
Bond lengths (Å)	0.005	0.007	0.005	0.008
Bond angles (°)	0.971	1.234	0.951	0.949

*Data for each structure were acquired from a single crystal.

*Values in parentheses are for the highest-resolution shell.

Table 10

RMSD values for Cα alignments of Fabs

Fab1/Fab2	RMSD _{vh} (Å)	# residues	RMSD _{vl} (Å)	# residues	RMSD _{vh+vl} (Å)	# residues
PGT121/PGT128	1.159	116/130	1.63	95/100	1.462	207/235
PGT121/PGT145	2.93	124/130	1.91	94/105	1.75	208/235
PGT121/10-1074	0.74	128/130	1.2	102/105	1.26	226/235
PGT121/GL	1.33	129/130	1.37	94/105	1.6	225/235
10-1074/GL	1.38	130/130	1.35	92/105	1.39	229/235
PGT121/PGT121 _{glycosylated}	0.79	125/126	0.5	100/100	0.78	225/226

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Table 11

Contacts between PGT121 Fab and bound glycan

Glycan atom	Protein atom	Water	Distance (Å)	Glycan atom	Protein atom	Water	Distance (Å)
GlcNAc ⁵ -O3	Asn ⁵⁶ -N52		2.91	Sia ¹⁰ -O8	Asp ³¹ -O		2.72
GlcNAc ⁵ -O7	Asn ⁵⁶ -O51		2.94	Sia ¹⁰ -O10	His ²⁷ -N		3.18
GlcNAc ⁵ -O6		H ₂ O ⁴⁷¹	3.15	Sia ¹⁰ -O10	His ²⁷ -O		3.19
GlcNAc ⁵ -O4		H ₂ O ⁴⁷⁷	3.05	Sia ¹⁰ -O9		H ₂ O ⁴⁸⁰	3.19
GlcNAc ⁵ -O3		H ₂ O ⁴⁸¹	2.94	Sia ¹⁰ -O8	Ser ³³ -Oγ		3.70*
Man ¹ -O4		H ₂ O ⁴¹⁰	3.02	Man ³ -O3	Asn ⁵⁸ -O51		2.58
Man ¹ -O4		H ₂ O ⁴²⁰	2.88	Man ³ -O6		H ₂ O ⁴⁷⁷	3.35
Man ¹ -O3		H ₂ O ⁴¹⁰	3.35	GlcNAc ⁴ -O5	Thr ²⁷ -O		3.33
Man ¹ -O2		H ₂ O ⁴⁷⁷	3.14	GlcNAc ⁴ -N2		H ₂ O ⁴⁷⁹	3.2
Man ¹ -O5		H ₂ O ⁴⁷⁷	2.82	Fuc ⁶ -O2		H ₂ O ⁴⁷¹	2.57
Man ² -O6	Thr ¹²⁰ -Oγ1		3.34		Asp ³¹ -O	H ₂ O ⁴³⁵	3.09
Man ² -O2		H ₂ O ⁴¹⁰	3.41		Asp ³¹ -O51	H ₂ O ⁴⁸⁵	3.32
Man ² -O5		H ₂ O ⁴⁴⁸	2.95		Tyr ⁵² -OH	H ₂ O ⁴⁸¹	2.8
Man ² -O6		H ₂ O ⁴⁴⁵	3.28		His ⁵² -Nε2	H ₂ O ⁴³⁵	3.18
GlcNAc ⁷ -N2	Tyr ²³ -OH		2.72		Ser ⁵⁴ -Oγ	H ₂ O ⁴¹²	3.2
GlcNAc ⁷ -O5		H ₂ O ⁴¹⁰	3.38		Ser ⁵⁴ -O	H ₂ O ⁴²⁰	3.16
GlcNAc ⁷ -O7		H ₂ O ⁴¹¹	3.03		Gly ⁵³ -O	H ₂ O ⁴⁷⁹	2.85
GlcNAc ⁷ -O3	His ²⁷ -N52		3.60*		Asp ⁵⁶ -O51	H ₂ O ⁴⁸¹	3.49
GlcNAc ⁷ -O7	His ²⁷ -Nε2		3.70*		Asp ⁵⁶ -O52	H ₂ O ⁴⁴⁵	3.07
Gal ⁶ -O3	Lys ⁵³ -Nε		2.97		Asn ⁵⁸ -N52	H ₂ O ⁴⁸¹	3.15
Gal ⁶ -O4		H ₂ O ⁴⁸⁰	3.47		Arg ²² -Nε	H ₂ O ⁴⁷¹	2.58
Gal ⁶ -O4		H ₂ O ⁴³⁵	2.76		Thr ¹²⁰ -Oγ1	H ₂ O ⁴¹¹	2.94
Gal ⁶ -O5		H ₂ O ⁴³⁵	3.17				

Hydrogen bond criteria: bond distance < 3.5 Å, O-H...O/N-H...O angle > 90°

*Contacts are close to hydrogen bond distance cutoff and are included as possible interactions

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Table 12

In vitro neutralization activity of PGT121GM and 10-1074GM

Virus ID	Clade	PGT121	PGT121 _{GM}	10-1074	10-1074 _{GM}
Q842.d12	A	0.074	>50	>50	>50
3365.v2.c2	A	7.353	>50	0.453	0.457
0260.v5.c36	A	0.153	>50	0.180	0.518
YU.2	B	0.356	1.355	0.308	0.262
TRO.11	B	0.051	0.358	0.057	0.049
TRJO4551.58	B	35.294	>50	0.634	0.721
GH0682.42	B	8.545	>50	0.929	0.378
PVO.4	B	0.045	47.884	0.960	0.139
BHPA4259.7	B	0.054	20.801	0.138	0.087
WITO4160.33	B	6.007	>50	2.112	0.406
1054_07_TC4_1498	B (T/F)	0.606	>50	0.583	0.193
6244_13_B5_4578	B (T/F)	1.879	48.880	0.922	0.304
62357_14_D3_4589	B (T/F)	45.650	>50	>50	40.792
CNE19	BC	0.189	48.092	50	0.379
CNE17	BC	>50	>50	13.297	4.818
CNE58	BC	>50	>50	0.968	1.158
CNE30	BC	0.559	8.401	1.200	1.045
CNE52	BC	32.936	>50	13.147	6.884
ZM233M.PB6	C	8.977	>50	0.348	0.232
ZM53M.PB12	C	0.002	>50	>50	>50
CAP45.2.00.03	C	6.544	>50	>50	>50
HIV-16855-2.3	C	4.290	>50	>50	>50
HIV-16845-2.22	C	>50	>50	5.835	2.878
ZM214M.PL15	C	3.150	>50	2.367	0.200
ZM135M.PL10a	C	5.885	>50	0.367	0.184
Ce1086_B2	C (T/F)	0.008	>50	>50	>50
Ce1172_H1	C (T/F)	0.066	0.180	0.165	0.064
1394C8G1(Rev-)	C (T/F)	0.372	2.120	0.191	0.075
3817.v2.c59	CD	>50	>50	14.330	3.423
6852.v1.c20	CD	0.608	>50	0.138	0.124
BJOX009000.02.4	CRF01_AE	37.239	>50	>50	>50
211-8	CRF02_AG	8.640	>50	0.425	0.978
928-28	CRF02_AG	>50	>50	4.698	3.121
T251-18	CRF02_AG	>50	>50	7.296	3.459
T278-50	CRF02_AG	>50	>50	18.278	12.017
263-8	CRF02_AG	24.678	>50	8.627	7.779
235-47	CRF02_AG	1.076	>50	0.166	0.089
A07412M1.vrc12	D	0.408	18.947	0.048	0.048
X1193_c1	G	0.202	11.859	0.475	0.195
X1254_c3	G	0.188	0.222	0.297	0.112

Numbers indicate antibody IgG concentrations in µg/ml to reach the IC₅₀ in the TZM-bl neutralization assay. IC₅₀ values are color coded and indicate from dark green to dark red, an increasing neutralization sensitivity. > indicates that the IC₅₀ for a given virus was not reached at the concentration tested.

Table 13

SHIV _{AD262}						SHIV _{GENE-03458}					
Animal ID	Age	Doseage	PROTECTED	Abs conc ($\mu\text{g/ml}$)	Titer (TCID ₅₀)	Animal ID	Age	Doseage	PROTECTED	Abs conc ($\mu\text{g/ml}$)	Titer (TCID ₅₀)
				at Day 0	at Day 0					at Day 0	at Day 0
R000037	18/03	20mg/kg	No	200.0	1.000	R000038	18/03	20mg/kg	No	200.0	1.00
R000040			No	211.0	<1.000	R000039			No	200.0	1.00
R000041		20mg/kg	No	200.0	1.00	R000040	18/03	20mg/kg	Yes	210.1	1.00000
R000042			No	198.1	1.00	R000041			Yes	200.7	1.00000
R000043	18/03	20mg/kg	Yes	207.0	1.0000	R000042			Yes	202.7	1.00000
R000044			No	203.0	1.0000	R000043			Yes	199.1	1.00000
R000045		20mg/kg	No	20.7	1.000	R000044		10mg/kg	Yes	19.1	1.400
R000046			No	47.2	1.000	R000045			No	20.3	1.000
R000047		10mg/kg	Yes	24.0	1.000	R000046		0.0mg/kg	Yes	3.1	1.000
R000048			No	19.7	1.00	R000047			Yes	1.0	1.000
R000049		0.0mg/kg	No	1.0	<1.00	R000048		0.0mg/kg	No	1.0	<1.00
R000050			No	1.0	1.00	R000049			No	1.0	<1.00
R000051	10-10/14	20mg/kg	No	200.0	1.0000	R000050	10-10/14	20mg/kg	Yes	200.0	1.0000
R000052			No	207.7	1.0000	R000051			Yes	173.0	1.0000
R000053		20mg/kg	Yes	112.0	1.000	R000052		20mg/kg	Yes	90.0	1.400
R000054			No	117.0	1.000	R000053		20mg/kg	Yes	90.0	1.000
R000055		10mg/kg	No	100.0	1.00	R000054			No	100.0	1.000
R000056			No	24.0	1.00	R000055		0.0mg/kg	No	100.0	1.000
R000057	00/00/17	20mg/kg	No	100.0	1.000	R000056	00/00/17	20mg/kg	Yes	200.0	1.000
R000058			No	90.1	1.000	R000057			Yes	200.0	1.000
R000059		10mg/kg	No	90.0	1.00	R000058		20mg/kg	Yes	114.0	1.00
R000060			No	10.1	1.00	R000059			No	100.1	1.00
R000061	40-40m2	20mg/kg	No	10.0	1.00	R000060		20mg/kg	Yes	100.0	1.00
R000062			No	17.0	<1.00	R000061			No	100.0	1.00
R000063		10mg/kg	No	0.0	<1.00	R000062		10mg/kg	No	100.0	1.00
R000064			No	0.0	<1.00	R000063		0.0mg/kg	No	0.0	<1.00
R000065			No	0.0	<1.00	R000064			No	0.0	<1.00
R000066	10/00	20mg/kg	No	0.0	<1.00	R000065	40-40m2	20mg/kg	No	0.1	0.0
R000067			No	0.0	<1.00	R000066			No	4.0	0.0
R000068			No	0.0	<1.00	R000067	10/00	20mg/kg	No	0.0	0.0
R000069			No	0.0	<1.00	R000068			No	0.0	0.0

Table 14

Sample ID	IC ₅₀ in TZM-bl cells							HIVIG ($\mu\text{g/ml}$)	Tier Phenotype
	S321	C500	B320	G435	T5306	M263	M600c		
R5 SHIV _{GENE-03458}	321	289	77	172	160	429	134	132	2
R5 SHIV _{AD262}	48	36	39	31	41	44	40	1760	2
X4 SHIV _{GENE-03458}	110	94	50	65	109	115	65	630	2
HIV-1 _{CAADPR01A1}	84	<20	27	<20	<20	77	185	630	2
HIV-1 _{CAADPR}	13944	9152	812	8432	3960	43712	1709	1.81	1

*Values are the serum dilution at which relative luminescence units (RLUs) were reduced 50% compared to virus control wells (no test sample).

Table 15

SHVAD06EC

Endpoint neutralization Animal number		Accumulated value		Projected	
titers in plasma	Protected	Infected	Protected ^a	Infected ^b	Ratio
2773	1	0	12	0	12/12
2496	1	0	11	0	11/11
2076	1	0	10	0	10/10
2004	1	0	9	0	9/9
633	1	0	8	0	8/8
638	0	1	7	1	7/8
583	1	0	7	1	7/8
384	1	0	6	1	6/7
372	1	0	5	1	5/6
372	1	0	4	1	4/5
178	0	1	3	2	3/5
182	1	0	3	2	3/5
115	1	0	2	2	2/4
78	0	1	1	3	1/4
88	0	1	1	4	1/5
65	0	1	1	5	1/6
56	0	1	1	6	1/7
95	1	0	1	6	1/7
55	0	1	0	7	0/7
59	0	1	0	8	0/8
27	0	1	0	8	0/8
20	0	1	0	10	0/10
20	0	1	0	11	0/11
20	0	1	0	12	0/12
20	0	1	0	13	0/13
17	0	1	0	14	0/14

^a Sum from the bottom.
^b Sum from the top.
^c Endpoint protection titer (50% protective titer) was calculated to be 1:115

Table 16

SHVEDH12-V3AD6

Endpoint neutralization		Animal number		Accumulated value		Protected	
Uter in plasma		Protected	Infected	Protected	Infected	Ratio	%
13805		1	0	16	0	16/16	100%
13120		1	0	15	0	15/15	100%
12659		1	0	14	0	14/14	100%
13056		1	0	13	0	13/13	100%
2282		1	0	12	0	12/12	100%
1372		1	0	11	0	11/11	100%
539		0	1	10	1	10/11	91%
422		1	0	10	1	10/11	91%
420		1	0	9	1	9/10	90%
378		1	0	8	1	8/9	89%
159		1	0	7	1	7/8	88%
143		1	0	6	1	6/7	86%
142		1	0	5	1	5/6	83%
136		0	1	4	2	4/6	67%
106		1	0	4	2	4/6	67%
101		1	0	3	2	3/5	60%
90		0	2	2	3	2/5	40%
85		1	0	2	2	2/5	40%
79		0	1	1	4	1/5	20%
62		1	0	1	4	1/5	20%
39		0	1	0	6	0/6	0%
35		0	1	0	6	0/6	0%
20		0	1	0	7	0/7	0%
20		0	1	0	8	0/8	0%
20		0	1	0	9	0/9	0%
20		0	1	0	10	0/10	0%
20		0	1	0	11	0/11	0%
20		0	1	0	12	0/12	0%
20		0	1	0	13	0/13	0%

^a Sum from the bottom.

^b Sum from the top.

^c Endpoint protection titer (50% protective titer) was calculated to be 1:95.5

Table 17

Animal	Weeks Post infection	Pre-Infection	Pre mAb Treatment		Clinical Status
		CD4+ T Cells cells/ μ l	CD4+ T cells cells/ μ l	Viral Load RNA Copies/ml	
DBZ3	159	650	118	1.08E+04	Asymptomatic
DC99A	159	623	165	7.50E+03	Asymptomatic
DBXE	163	1585	158	1.96E+05	Intermittent
DCF1	157	1203	105	1.44E+05	Intermittent
DCM8	163	508	43	1.59E+03	Intermittent

Table 18

Animal	Treatment Time (Days)	SIV Gag RNA Copies per 10^8 Cell Eq	SIV Gag DNA Copies per 10^8 Cell Eq
DBZ3	0	9000	6700
DBZ3	10	360	7500
DBZ3	20	2400	14000
DC99A	0	31000	1400
DC99A	14	18000	5600
DC99A	20	8100	2700
DBXE	0	470000	71000
DBXE	14	17000	33000
DBXE	17	11000	22000
DCM8	0	110000	8600
DCM8	14	1700	1600
DCM8	20	22000	6600
DCF1	0	240000	15000
DCF1	14	190000	11000
DCF1	20	1100000	14000

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The foregoing examples and description of the preferred embodiments should be taken as illustrating, rather than as limiting the present invention as defined by the claims. As will be readily appreciated, numerous variations and combinations of the features set forth above can be utilized without departing from the present invention as set forth in the claims. Such variations are not regarded as a departure from the scope of the invention, and all such variations are intended to be included within the scope of the following claims. All references cited herein are incorporated herein in their entireties.

WHAT IS CLAIMED IS:

1. An isolated anti-HIV antibody, or antigen binding portion thereof, comprising at least one complementarity determining region (CDR) having a sequence selected from the group consisting of SEQ ID NOs: 33-38.
2. The isolated anti-HIV antibody of claim 1, or antigen binding portion thereof, wherein the CDR comprises a sequence selected from the group consisting of SEQ ID NOs: 39-104.
3. The isolated anti-HIV antibody of claim 1, or antigen binding portion thereof, comprising a heavy chain variable region that comprises CDRH 1, CDRH 2, and CDRH 3, wherein the CDRH 1, CDRH 2 and CDRH 3 comprise the respective sequences of SEQ ID NOs: 33-35.
4. The isolated anti-HIV antibody of claim 3, or antigen binding portion thereof, wherein the CDRH 1, CDRH 2 and CDRH 3 comprise the respective sequences of a CDRH set selected from the group consisting of SEQ ID NOs: 39-41, SEQ ID NOs: 45-47, SEQ ID NOs: 51-53, SEQ ID NOs: 57-59, SEQ ID NOs: 63-65, SEQ ID NOs: 69-71, SEQ ID NOs: 75-77, SEQ ID NOs: 81-83, SEQ ID NOs: 87-89, SEQ ID NOs: 93-95, SEQ ID NOs: 99-101, and SEQ ID NOs: 131-133.
5. The isolated anti-HIV antibody of claim 1, or antigen binding portion thereof, comprising a light chain variable region that comprises CDRL 1, CDRL 2 and CDRL 3, wherein the CDRL 1, CDRL 2 and CDRL 3 comprise the respective sequences of SEQ ID NOs: 36-38.
6. The isolated anti-HIV antibody of claim 5, or antigen binding portion thereof, wherein the CDRL 1, CDRL 2 and CDRL 3 comprise the respective sequences of a CDRL set selected from the group consisting of SEQ ID NOs: 42-44, SEQ ID NOs: 48-50, SEQ ID NOs: 54-56, SEQ ID NOs: 60-62, SEQ ID NOs: 66-68, SEQ ID NOs: 72-74, SEQ ID NOs: 78-80, SEQ ID NOs: 84-86, SEQ ID NOs: 90-92, SEQ ID NOs: 96-98, SEQ ID NOs: 102-104, and SEQ ID NOs: 134-136.

7. The isolated anti-HIV antibody of claim 1, or antigen binding portion thereof, comprising

a heavy chain variable region that comprises CDRH 1, CDRH 2, and CDRH 3, and
a light chain variable region that comprises CDRL 1, CDRL 2 and CDRL 3.

8. The isolated anti-HIV antibody of claim 7, or antigen binding portion thereof, wherein the CDRH 1, CDRH 2, CDRH 3, CDRL 1, CDRL 2 and CDRL 3 comprise the respective sequences of a CDR set selected from the group consisting of SEQ ID NOs: 39-44, SEQ ID NOs: 45-50, SEQ ID NOs: 51-56, SEQ ID NOs: 57-62, SEQ ID NOs: 63-68, SEQ ID NOs: 69-74, SEQ ID NOs: 75-79, SEQ ID NOs: 81-86, SEQ ID NOs: 87-92, SEQ ID NOs: 93-98, SEQ ID NOs: 99-104, and SEQ ID NOs: 131-136.

9. The isolated anti-HIV antibody of claim 1, or antigen binding portion thereof, comprising one or both of (i) a heavy chain comprising the consensus amino acid sequence of SEQ ID NO: 1 and (ii) a light chain comprising the consensus amino acid sequence of SEQ ID NO: 2.

10. The isolated anti-HIV antibody of claim 9, or antigen binding portion thereof, wherein the heavy chain comprises a sequence selected from the group consisting of SEQ ID NOs: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, and 129 and the light chain comprises a sequence selected from the group consisting of SEQ ID NOs: 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, and 130.

11. The isolated anti-HIV antibody of claim 9, or antigen binding portion thereof, wherein the heavy chain and the light chain comprise the respective sequences of SEQ ID NOs: 3-4, SEQ ID NOs: 5-6, SEQ ID NOs: 7-8, SEQ ID NOs: 9-10, SEQ ID NOs: 11-12, SEQ ID NOs: 13-14, SEQ ID NOs: 15-16, SEQ ID NOs: 17-18, SEQ ID NOs: 19-20, SEQ ID NOs: 21-22, SEQ ID NOs: 23-24, and 129-130.

12. The isolated anti-HIV antibody of claim 1, or antigen binding portion thereof, wherein the antibody is one selected from the group consisting of 10-259, 10-303, 10-410, 10-847, 10-996, 10-1074, 10-1121, 10-1130, 10-1146, 10-1341, 10-1369, and 10-1074GM..

13. The isolated anti-HIV antibody of anyone of claims 1-12, or antigen binding portion thereof, wherein the antibody is a human antibody, a humanized antibody, or a chimeric antibody.

14. An isolated nucleic acid comprising a sequence encoding a CDR, a heavy chain variable region, or a light chain variable region of the anti-HIV antibody of anyone of claims 1-13, or antigen binding portion thereof.

15. A vector comprising the nucleic acid of claim 14.

16. A cultured cell comprising the vector of claim 15.

17. A pharmaceutical composition comprising (i) at least one anti-HIV antibody of anyone of claims 1-13, or antigen binding portion thereof, and (ii) a pharmaceutically acceptable carrier.

18. A method of preventing or treating an HIV infection or an HIV-related disease comprising the steps of:

identifying a patient in need of such prevention or treatment, and

administering to said patient a first therapeutic agent comprising a therapeutically effective amount of at least one anti-HIV antibody of anyone of claims 1-13, or antigen binding portion thereof.

19. The method of claim 18, further comprising administering a second therapeutic agent.

20. The method of claim 19, wherein the second therapeutic agent is an antiviral agent.

21. A method for making an anti-HIV antibody or a fragment thereof, comprising obtaining the cultured cell of claim 16;
culturing the cell in a medium under conditions permitting expression of a polypeptide encoded by the vector and assembling of an antibody or fragment thereof, and purifying the antibody or fragment from the cultured cell or the medium of the cell.

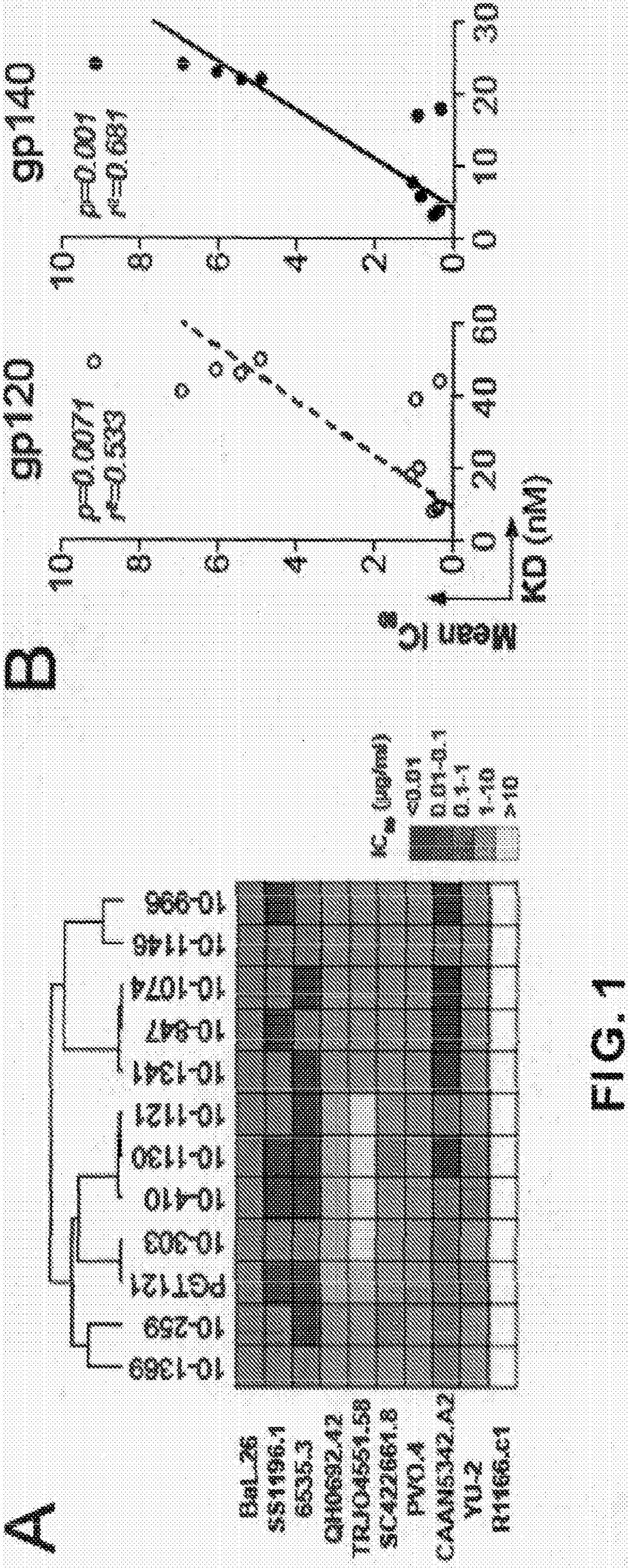
22. A kit comprising
a pharmaceutically acceptable dose unit of a pharmaceutically effective amount of at least one isolated anti-HIV antibody according to any of claims 1-13, and
a pharmaceutically acceptable dose unit of a pharmaceutically effective amount of an anti-HIV agent,
wherein the two pharmaceutically acceptable dose units can optionally take the form of a single pharmaceutically acceptable dose unit.

23. The kit of claim 22, wherein the anti-HIV agent is one selected from the group consisting of a non-nucleoside reverse transcriptase inhibitor, a protease inhibitor, a entry or fusion inhibitor, and an integrase inhibitor.

24. A kit for the diagnosis, prognosis or monitoring the treatment of an HIV infection in a subject comprising one or more detection reagents which specifically bind to anti-HIV neutralizing antibodies in a biological sample from a subject.

25. The kit of claim 24, further comprising reagents for performing PCR.

26. The kit of claim 25, further comprising reagents for performing mass spectrometry.



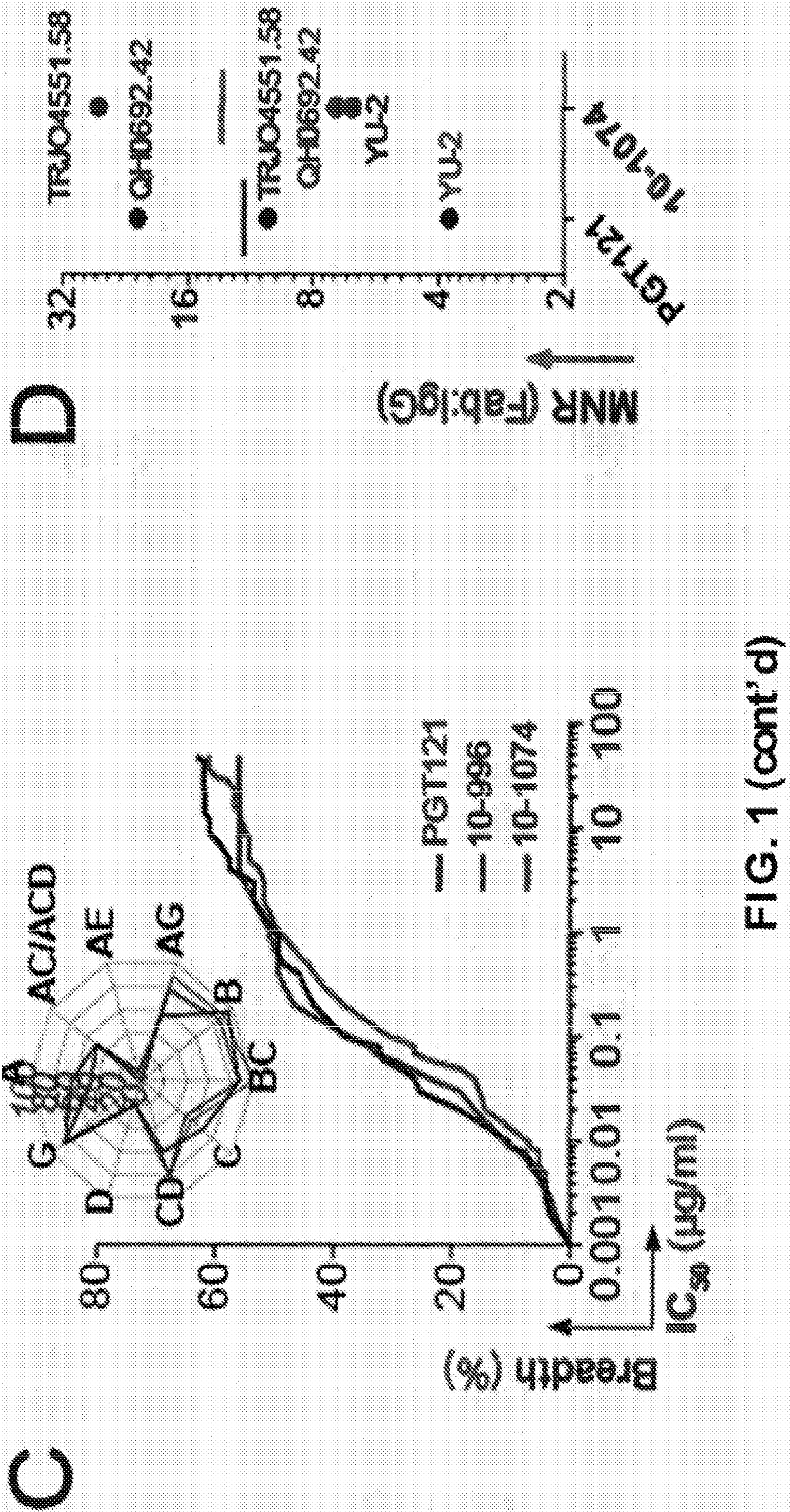


FIG. 1 (cont'd)

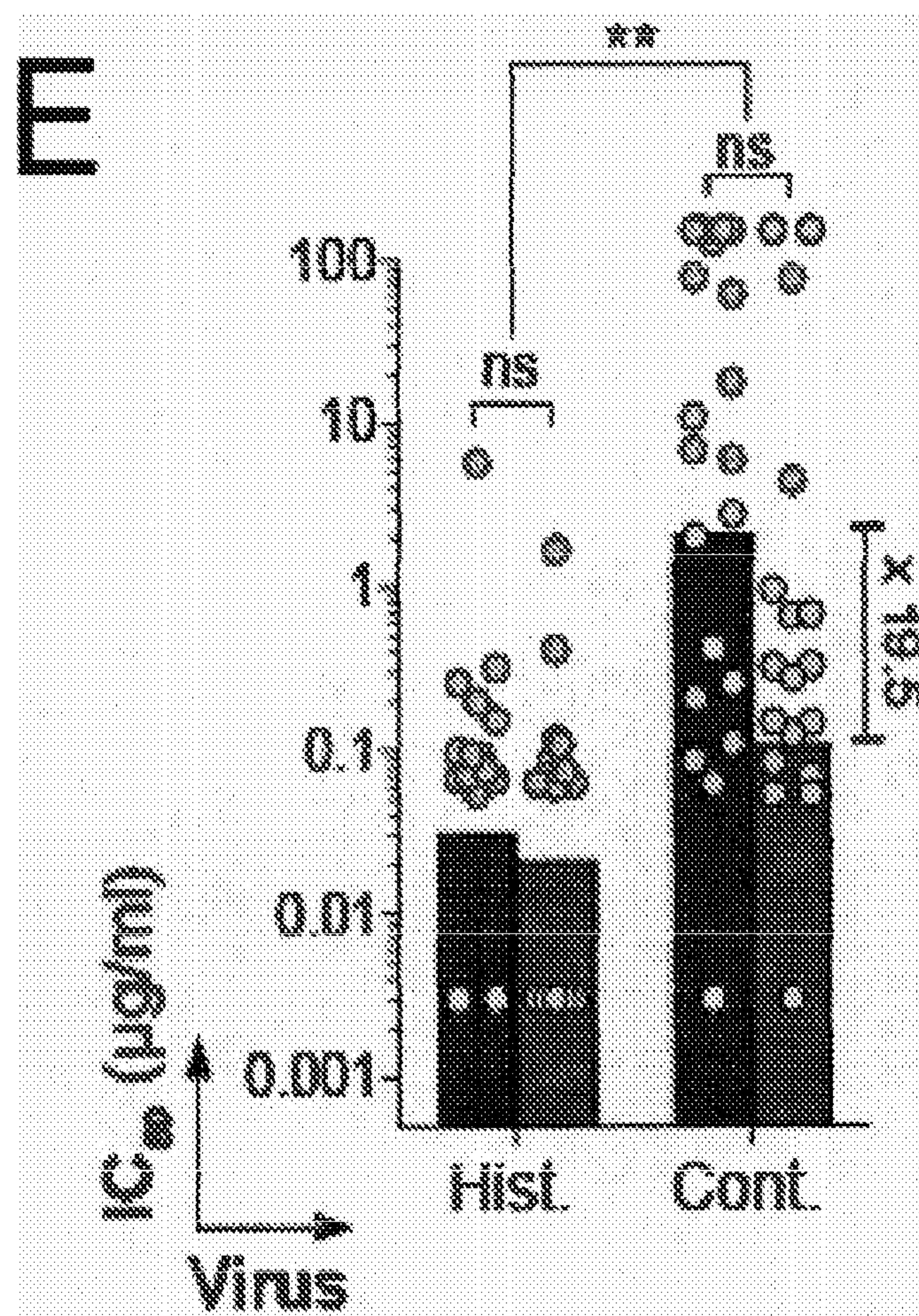


FIG. 1 (cont'd)

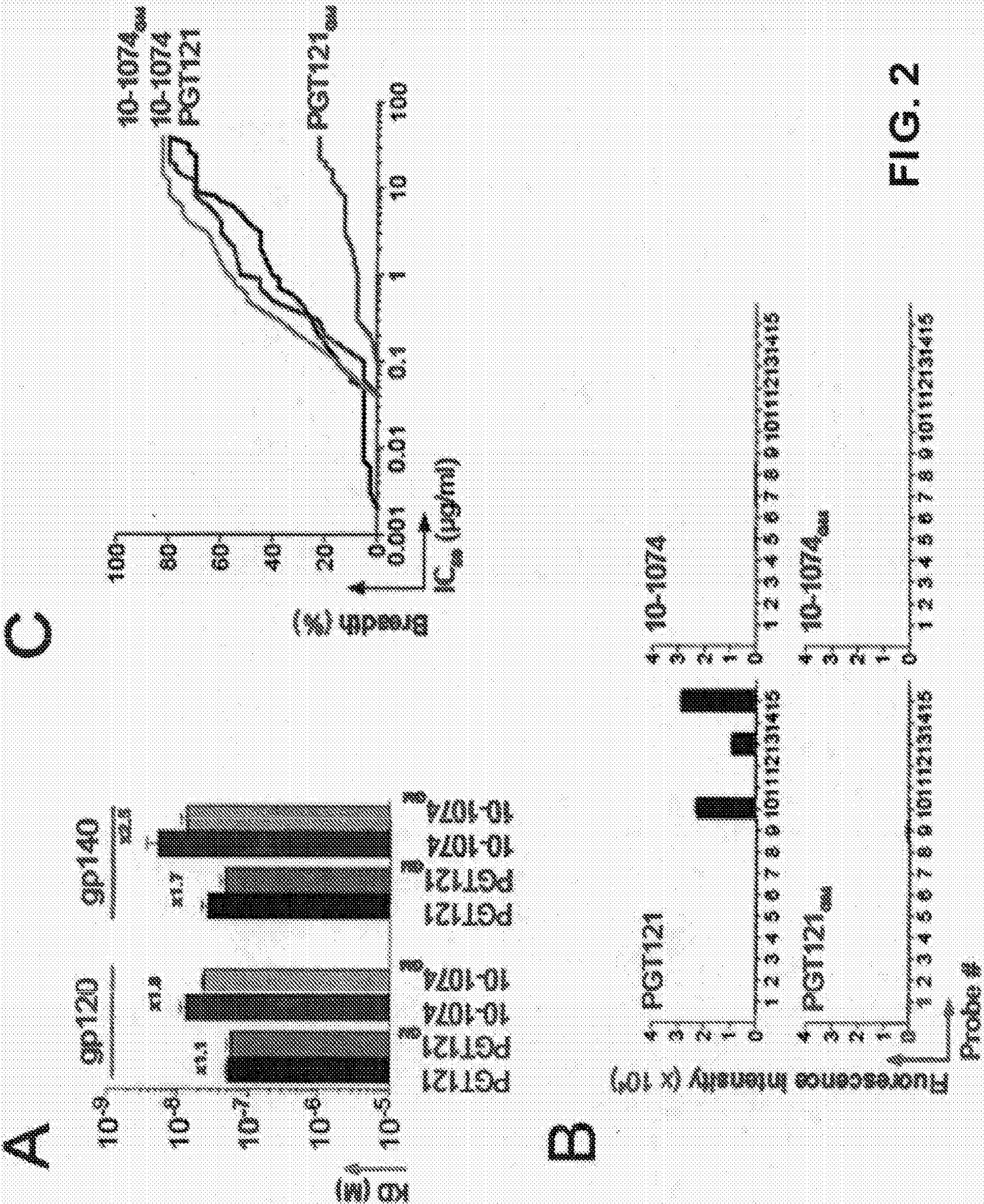
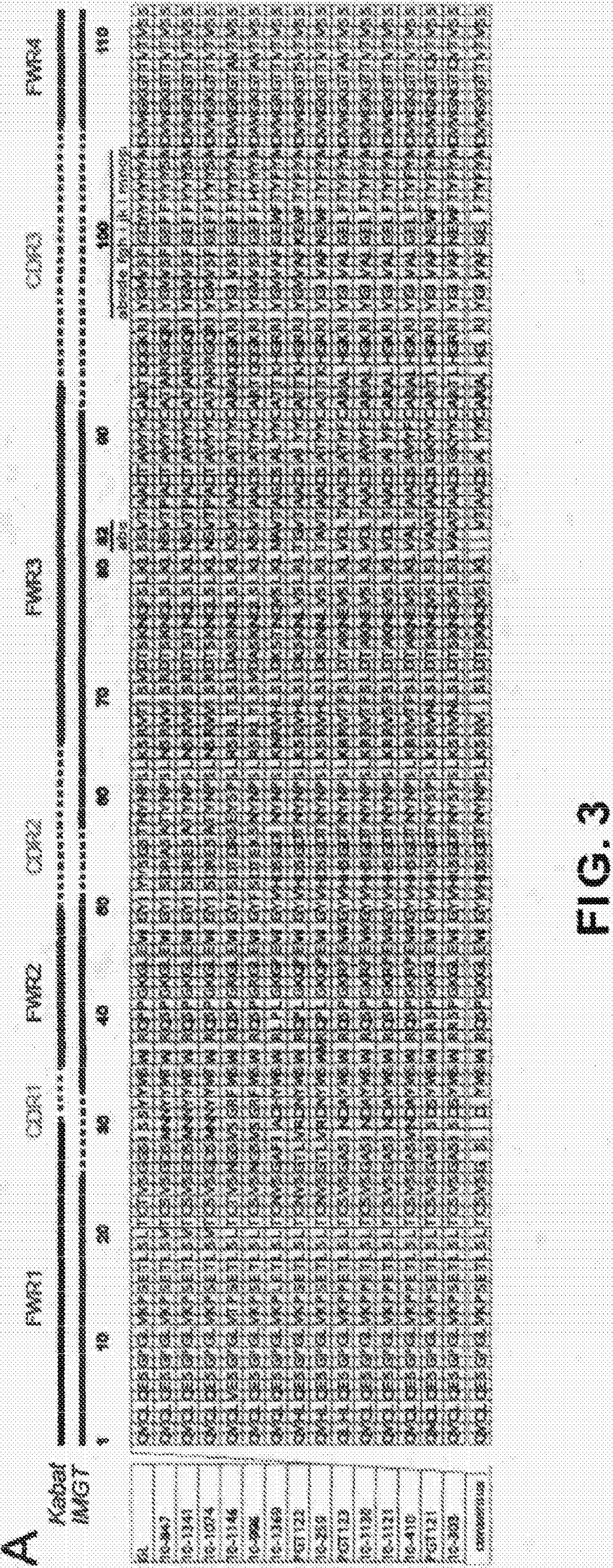


FIG. 2



B

Kabat
IMGT

FWR1 CDR1 FWR2 CDR2 FWR3 CDR3 FWR4

	12	20	30	40	50	60	70	80	90	100
GL	57	57	57	57	57	57	57	57	57	57
PGT121	57	57	57	57	57	57	57	57	57	57
10-303	57	57	57	57	57	57	57	57	57	57
10-1368	57	57	57	57	57	57	57	57	57	57
PGT123	57	57	57	57	57	57	57	57	57	57
10-258	57	57	57	57	57	57	57	57	57	57
10-847	57	57	57	57	57	57	57	57	57	57
10-1074	57	57	57	57	57	57	57	57	57	57
10-1341	57	57	57	57	57	57	57	57	57	57
10-996	57	57	57	57	57	57	57	57	57	57
10-1146	57	57	57	57	57	57	57	57	57	57
10-410	57	57	57	57	57	57	57	57	57	57
PGT122	57	57	57	57	57	57	57	57	57	57
10-1130	57	57	57	57	57	57	57	57	57	57
10-1121	57	57	57	57	57	57	57	57	57	57
consensus	57	57	57	57	57	57	57	57	57	57

FIG. 3 cont'd)

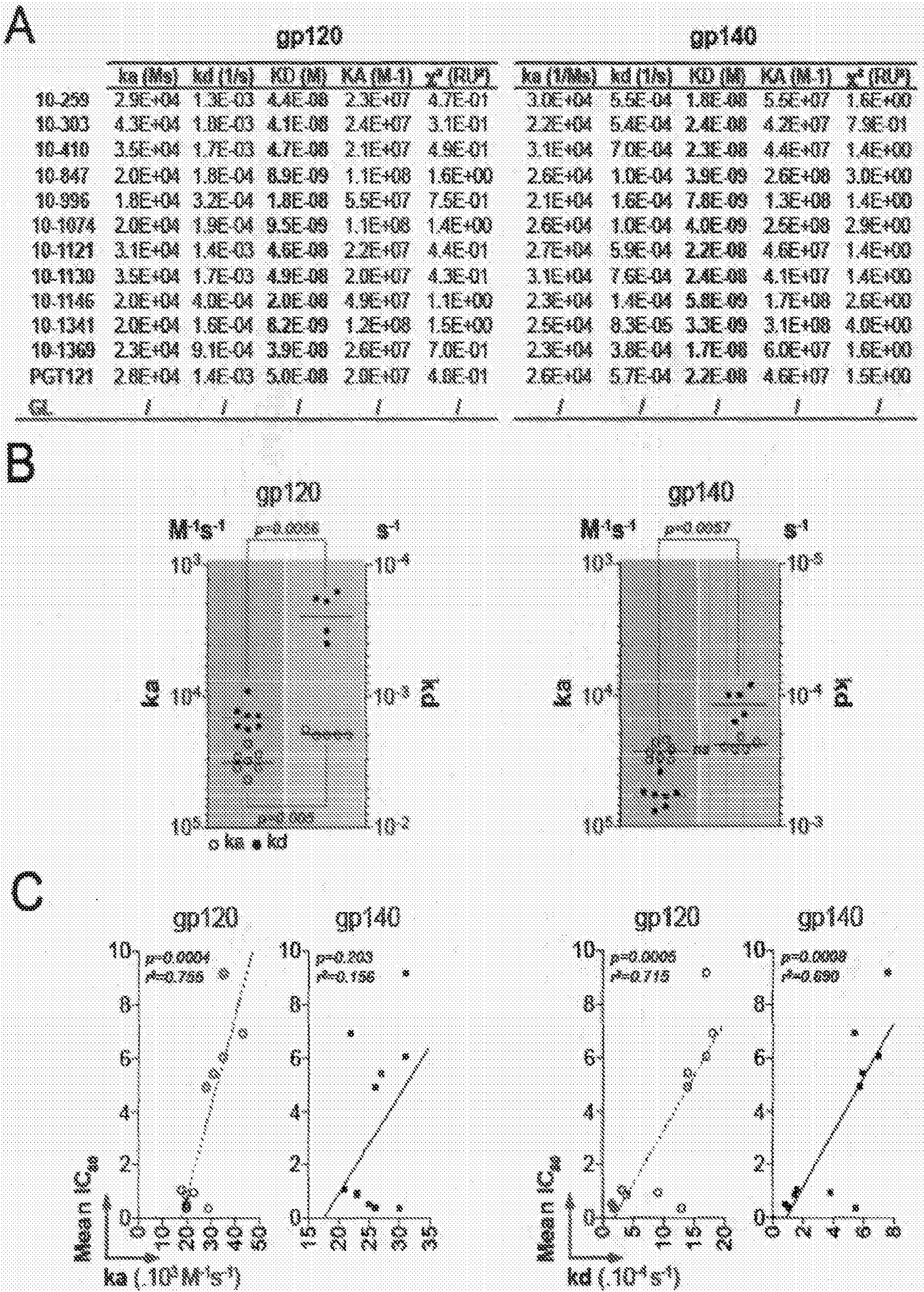


FIG. 4

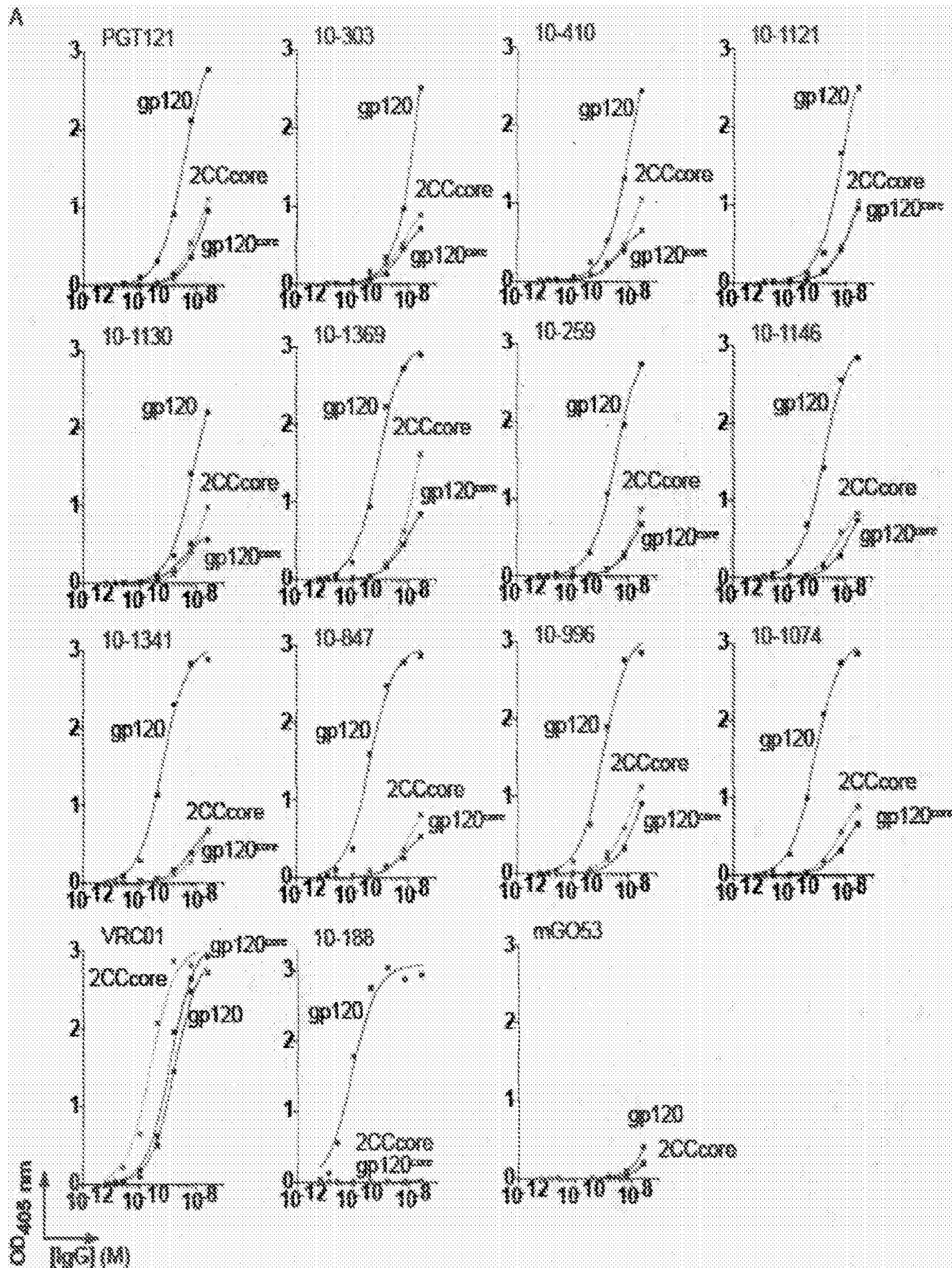


FIG. 5

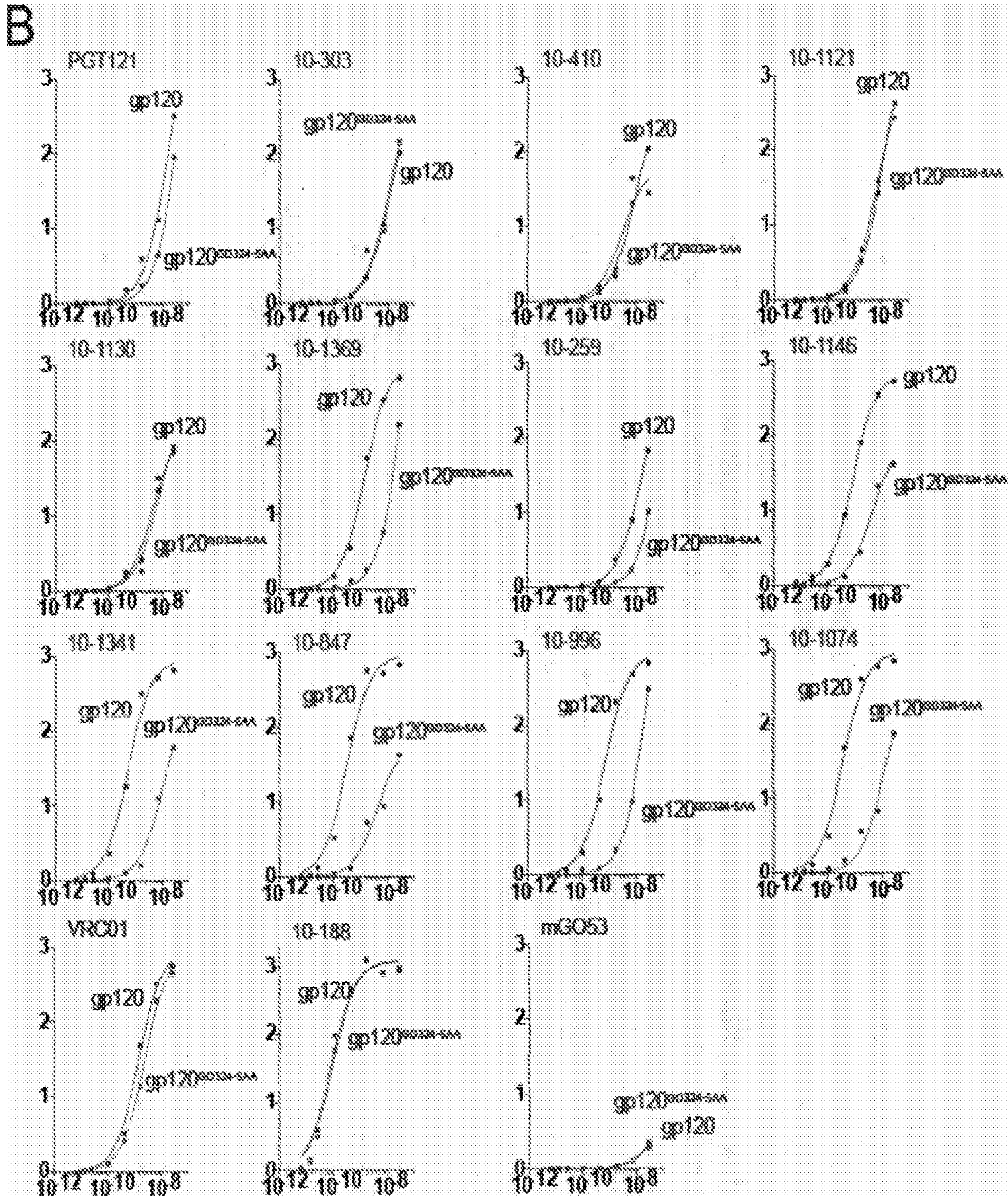
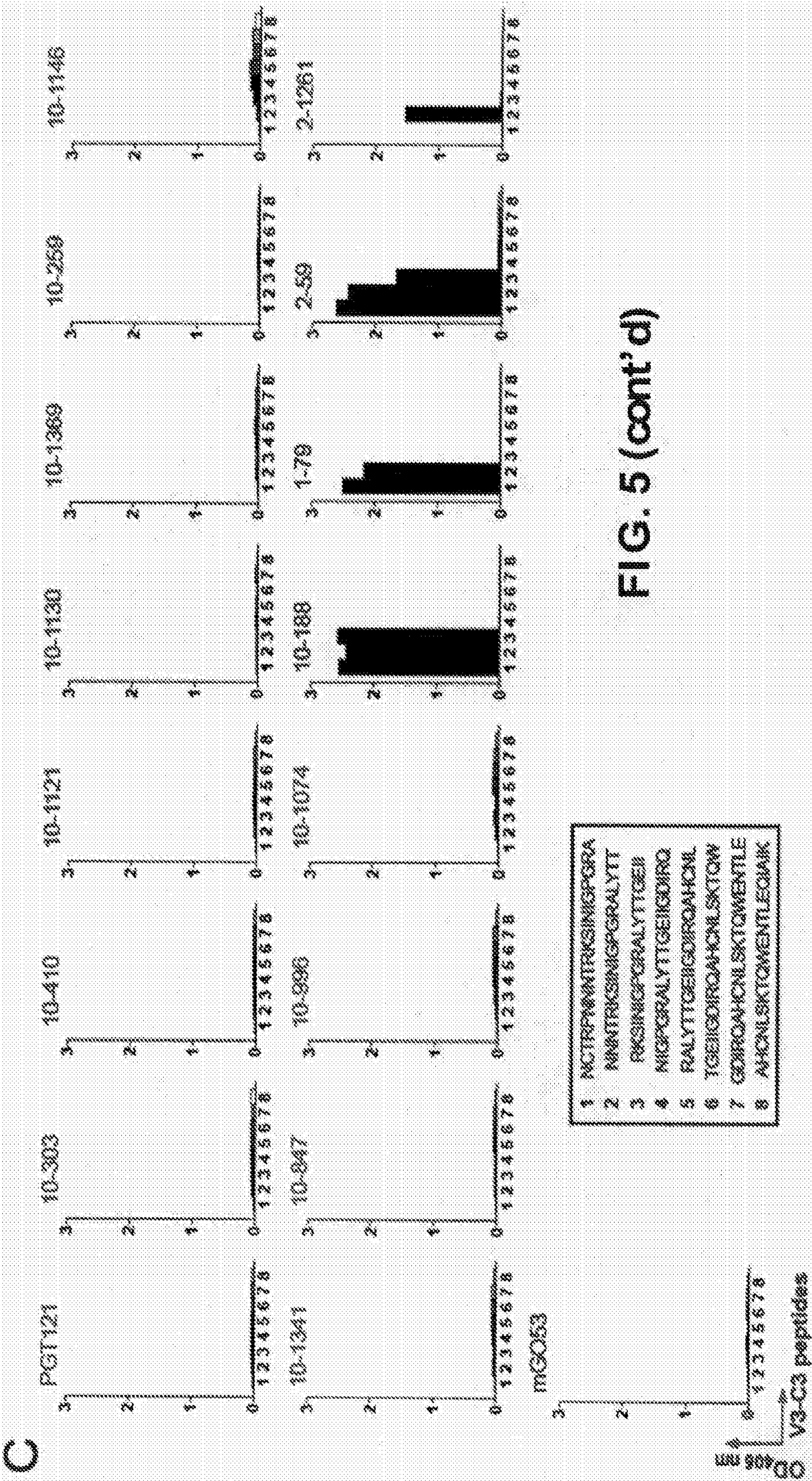
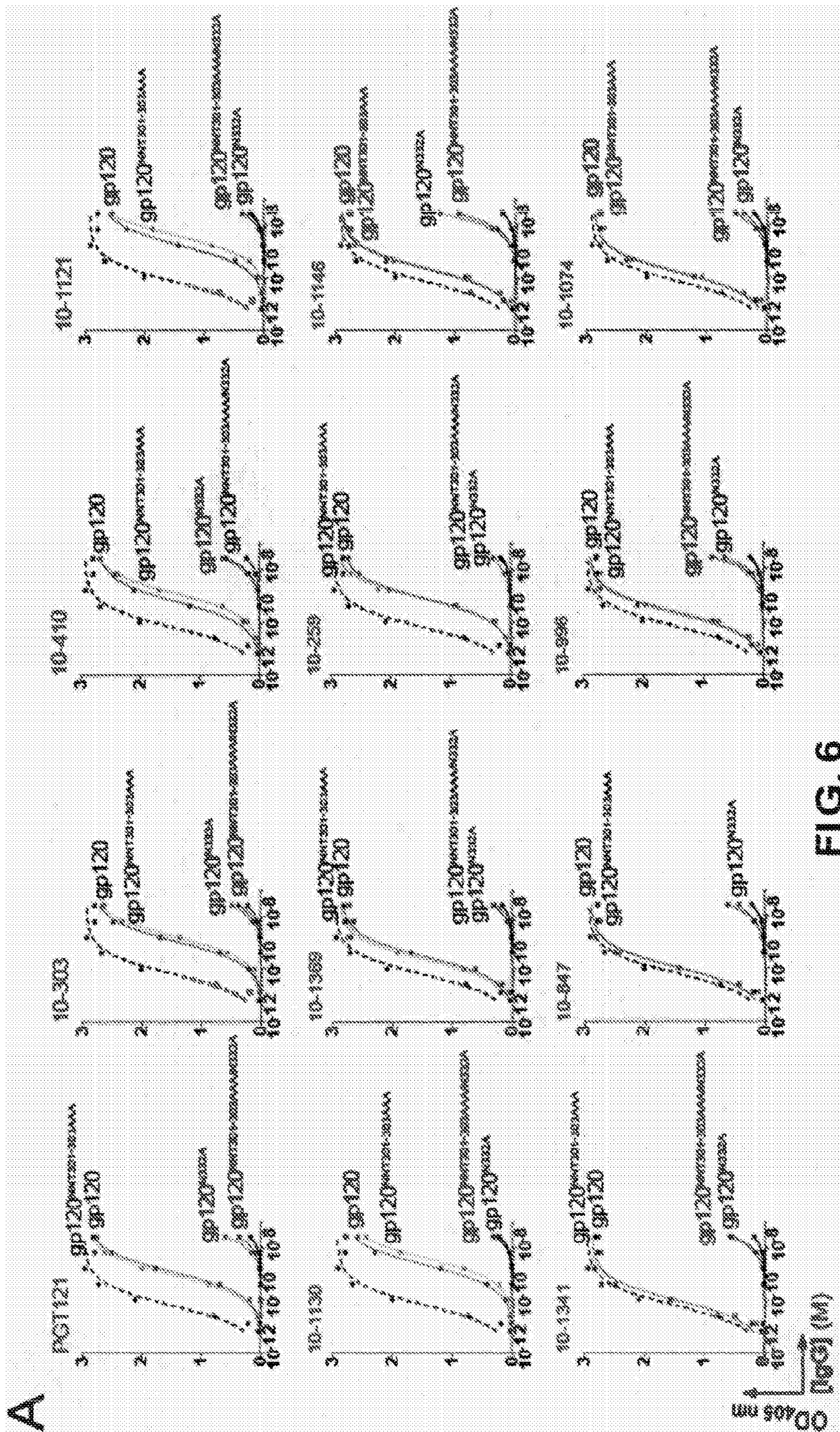
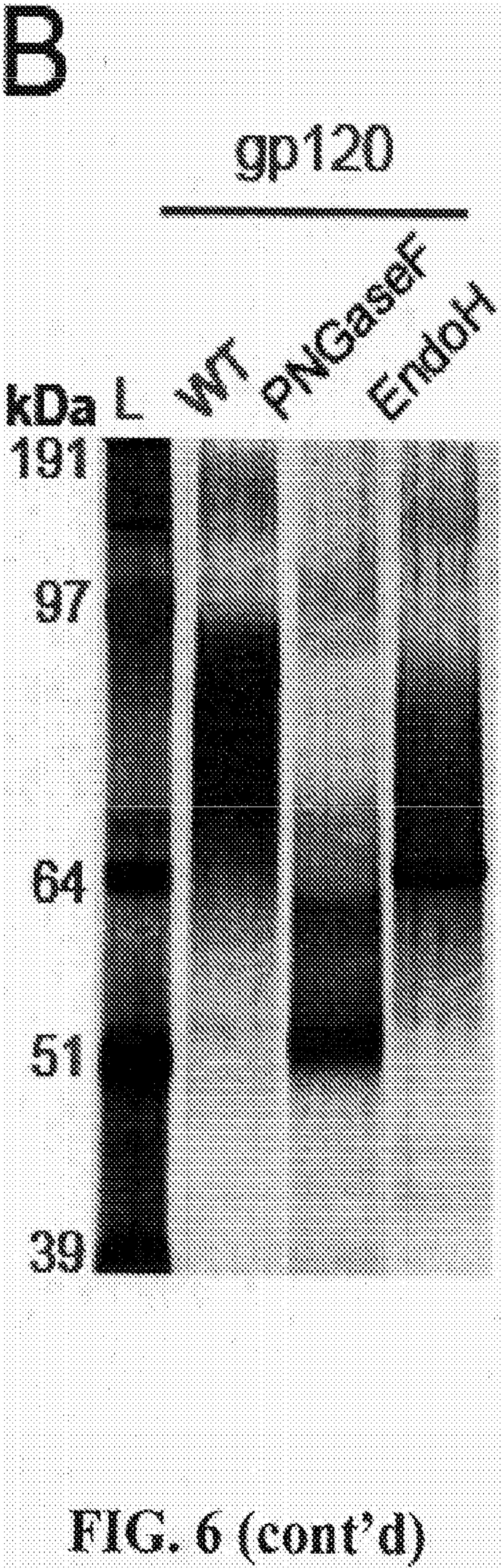
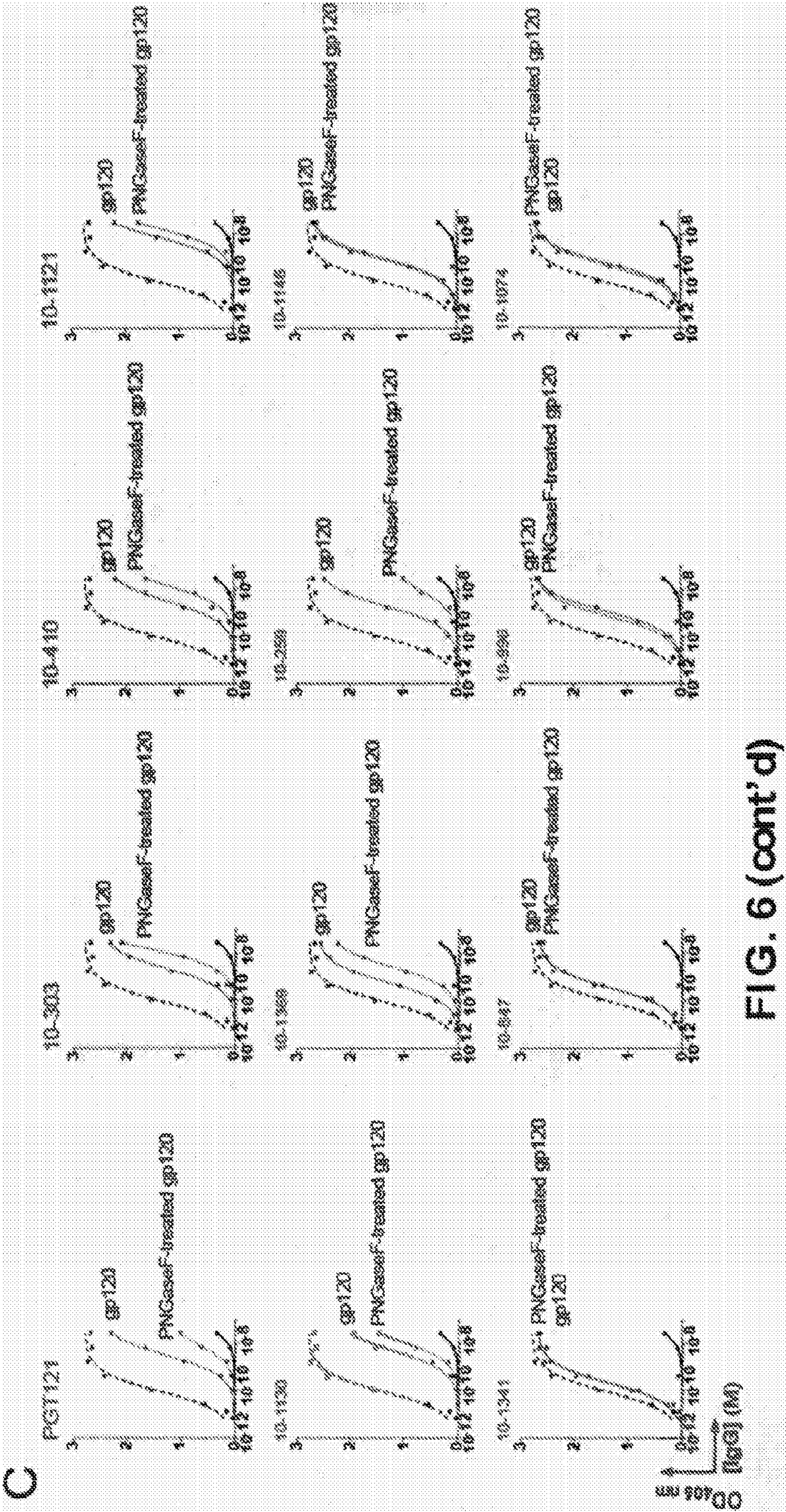


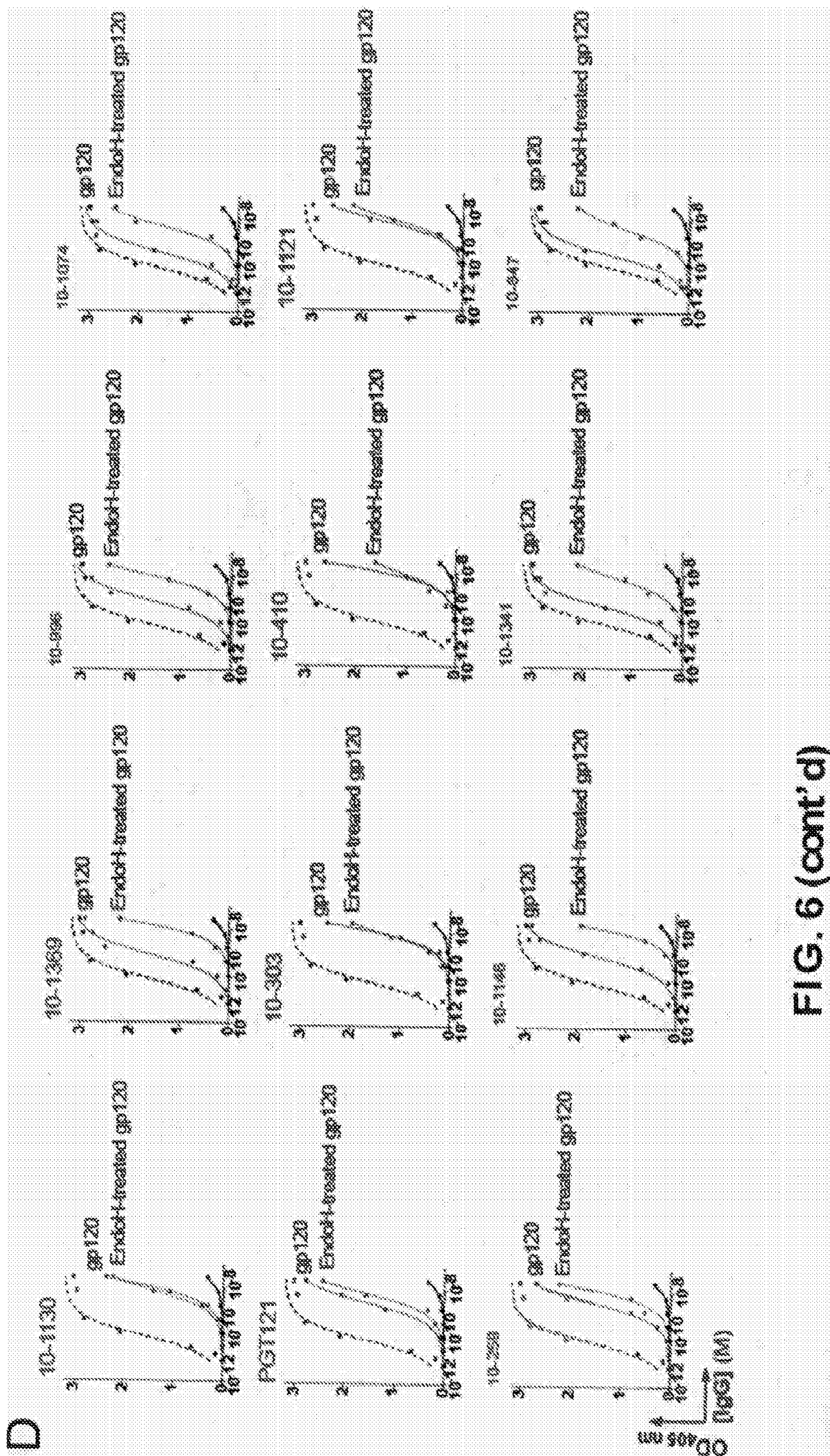
FIG. 5 (cont'd)

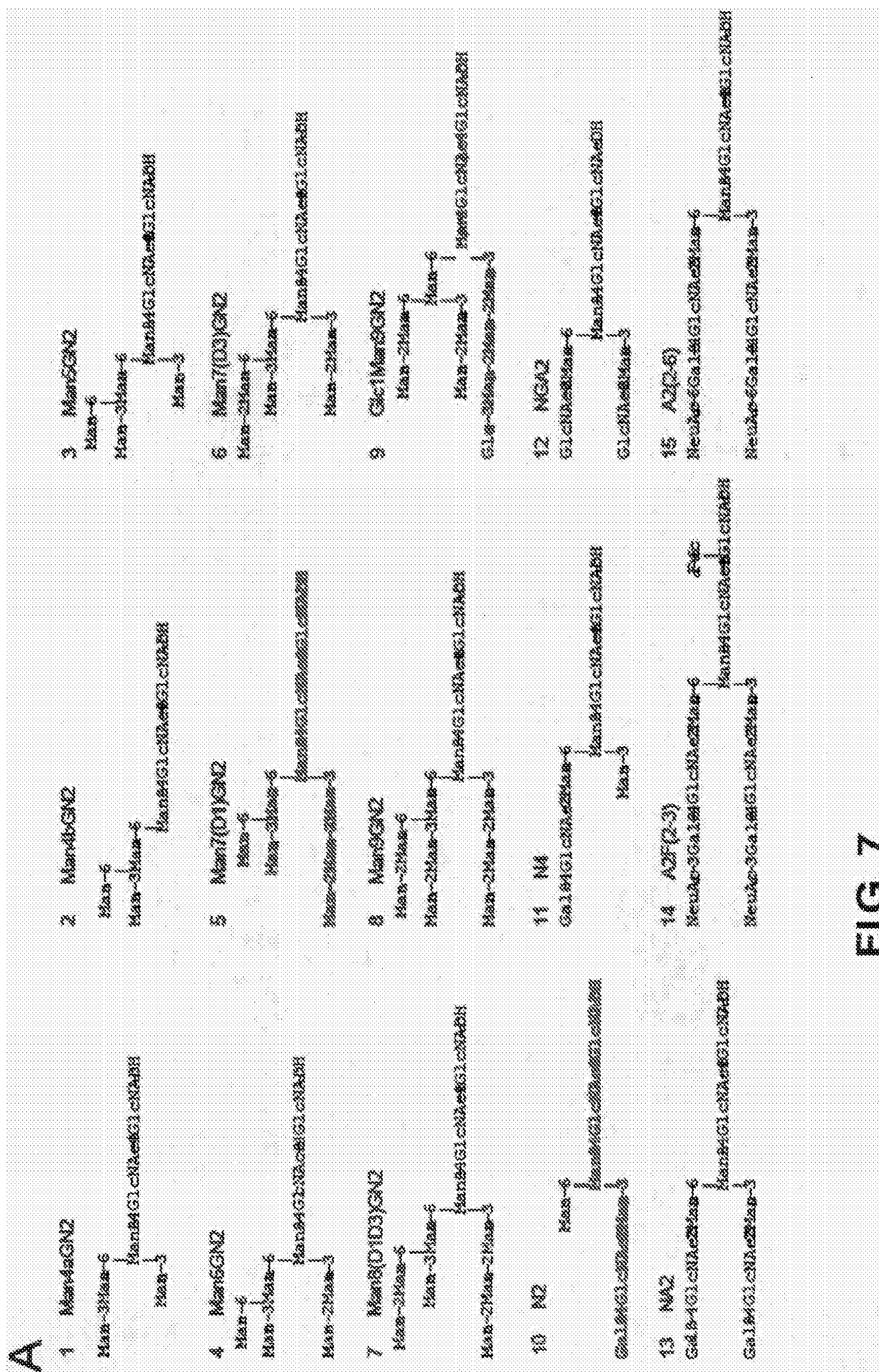












75
G
45
L

B

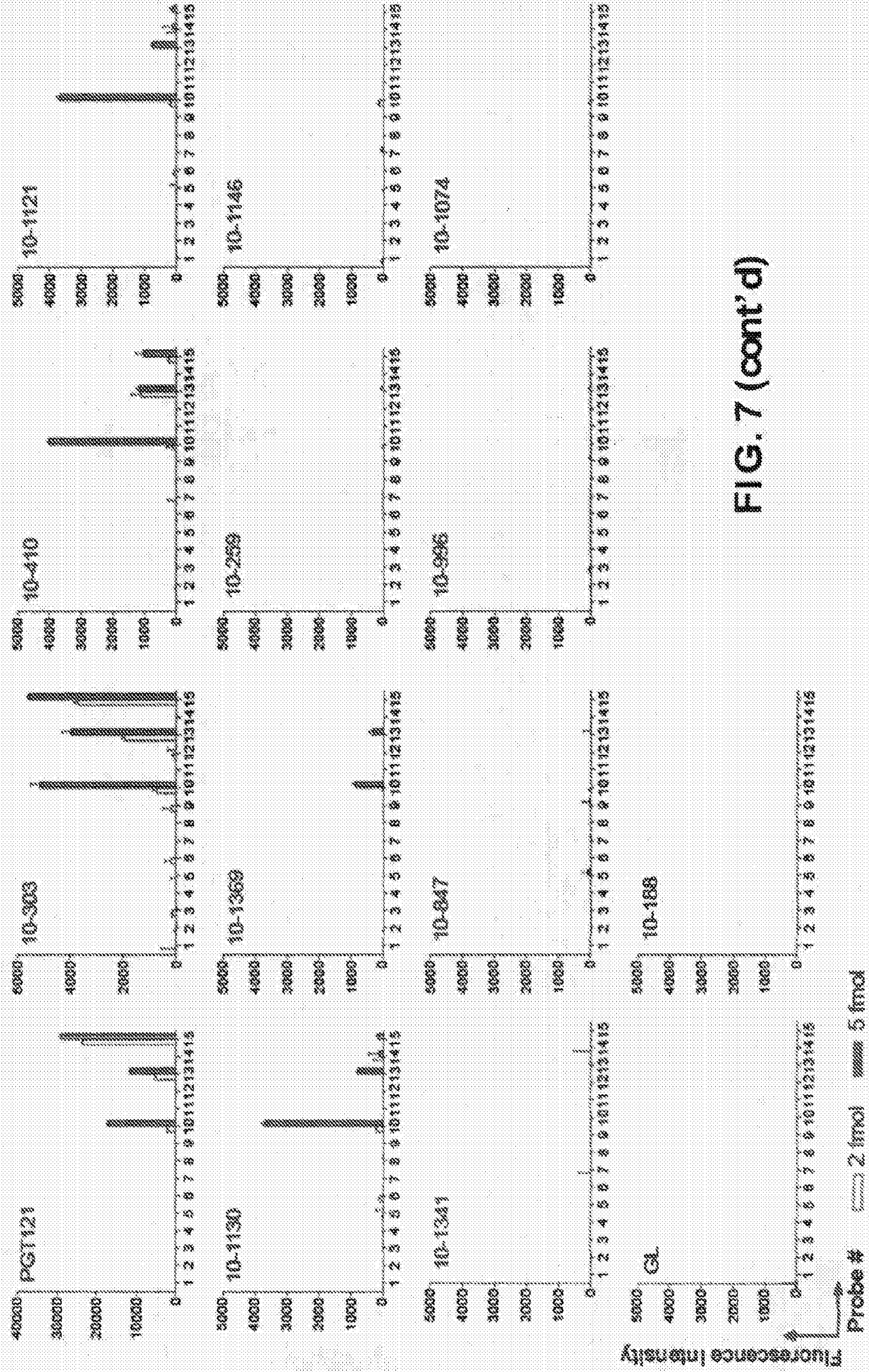
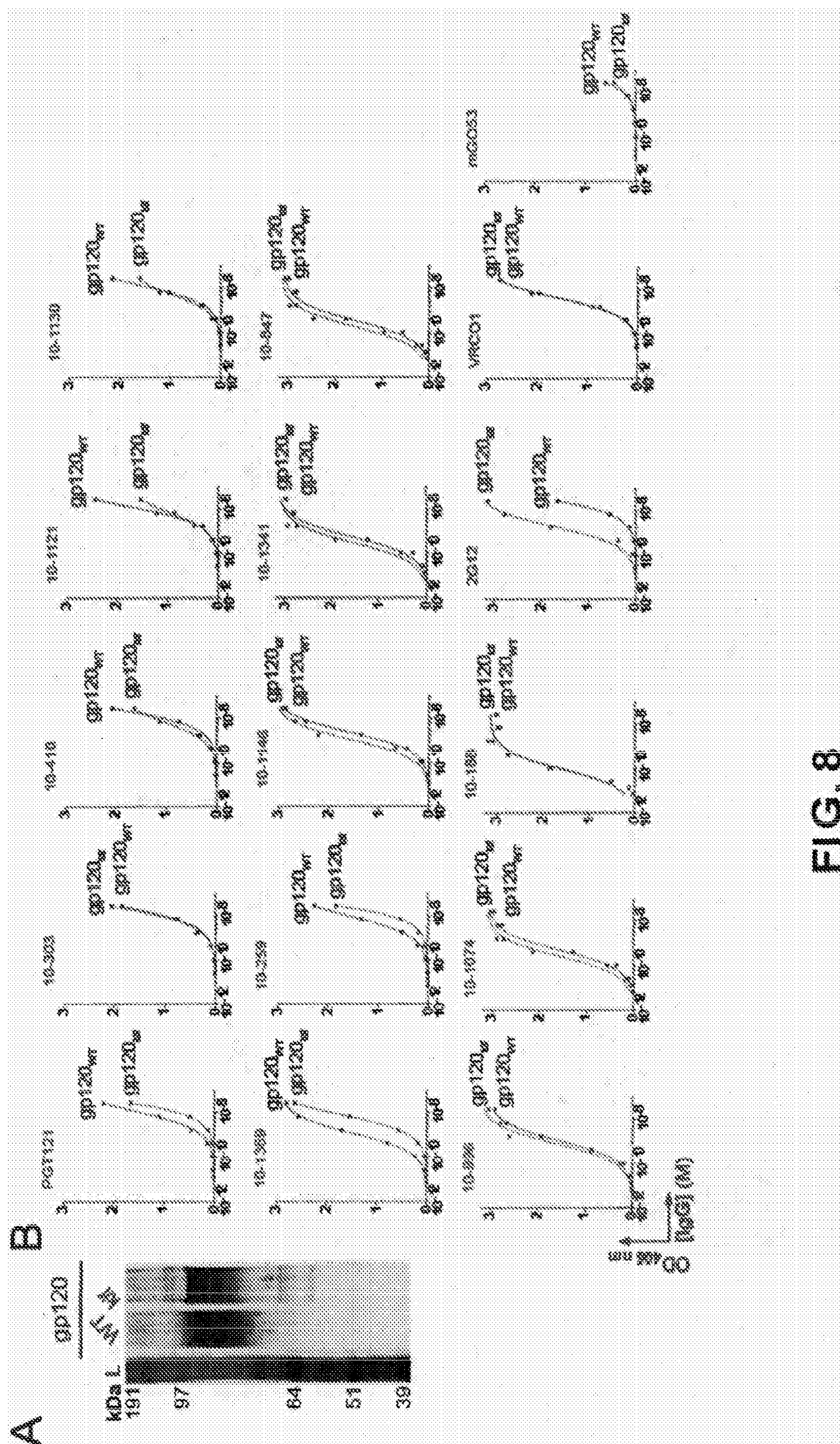
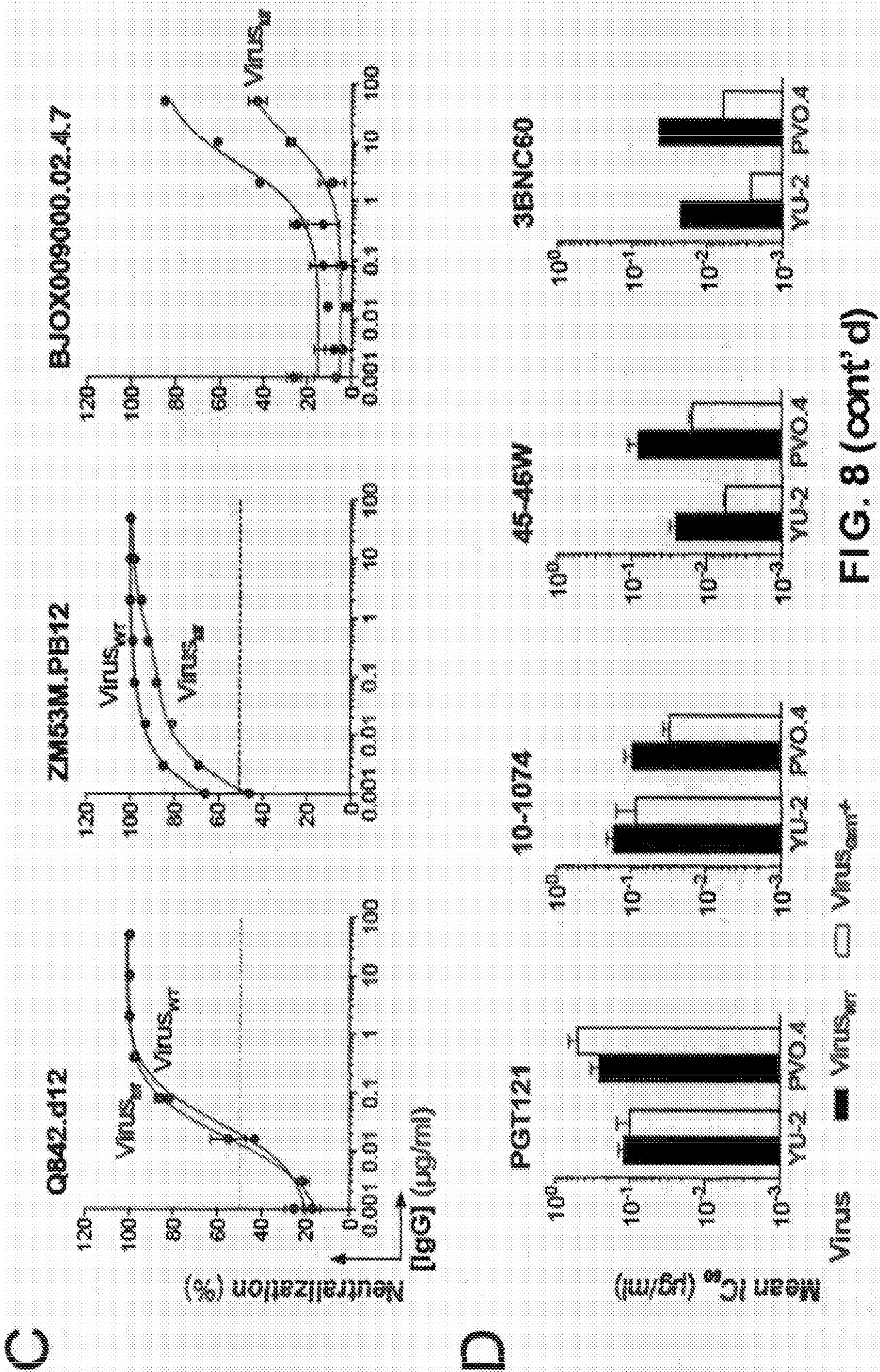


FIG. 7 (cont'd)





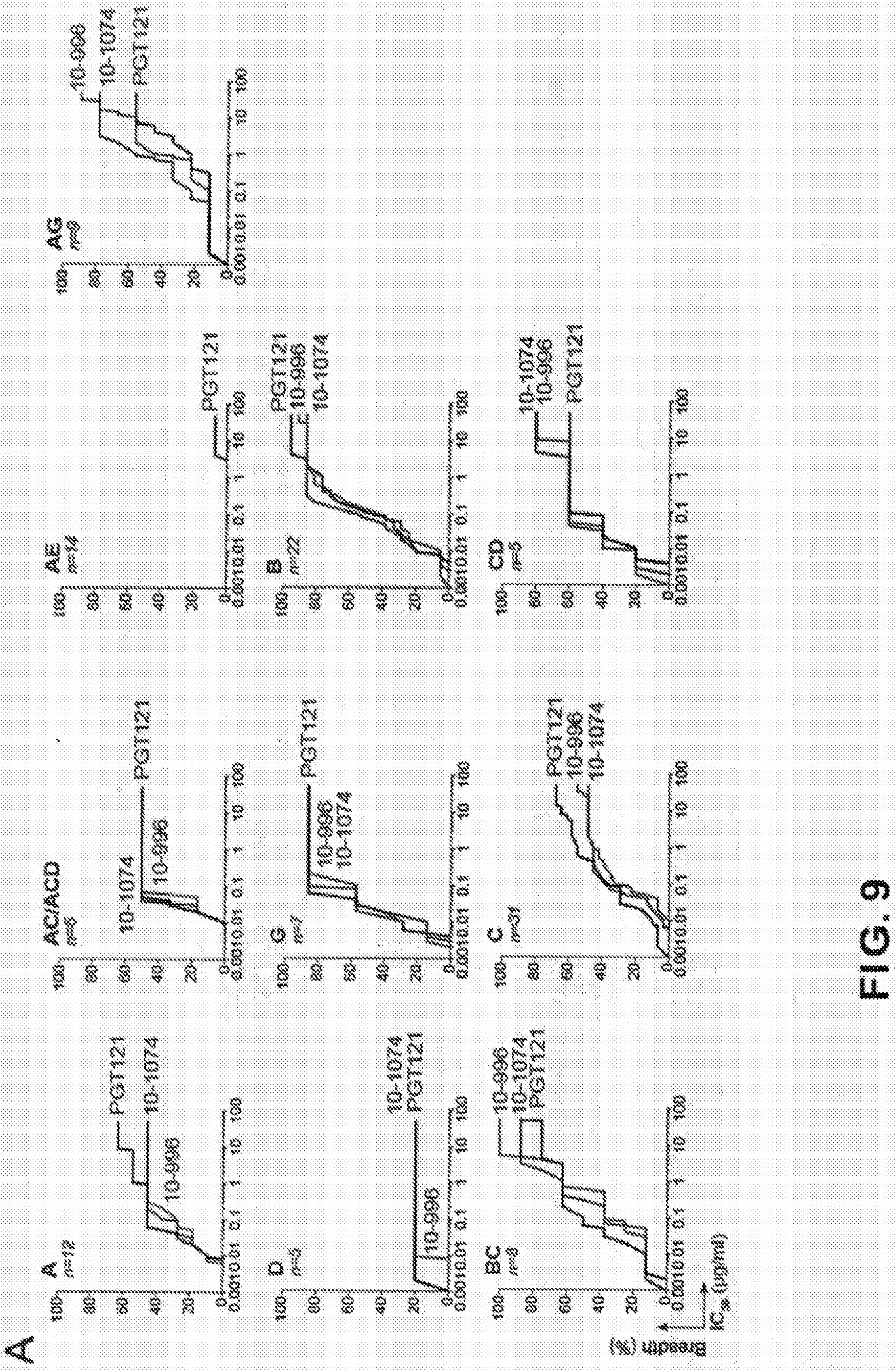
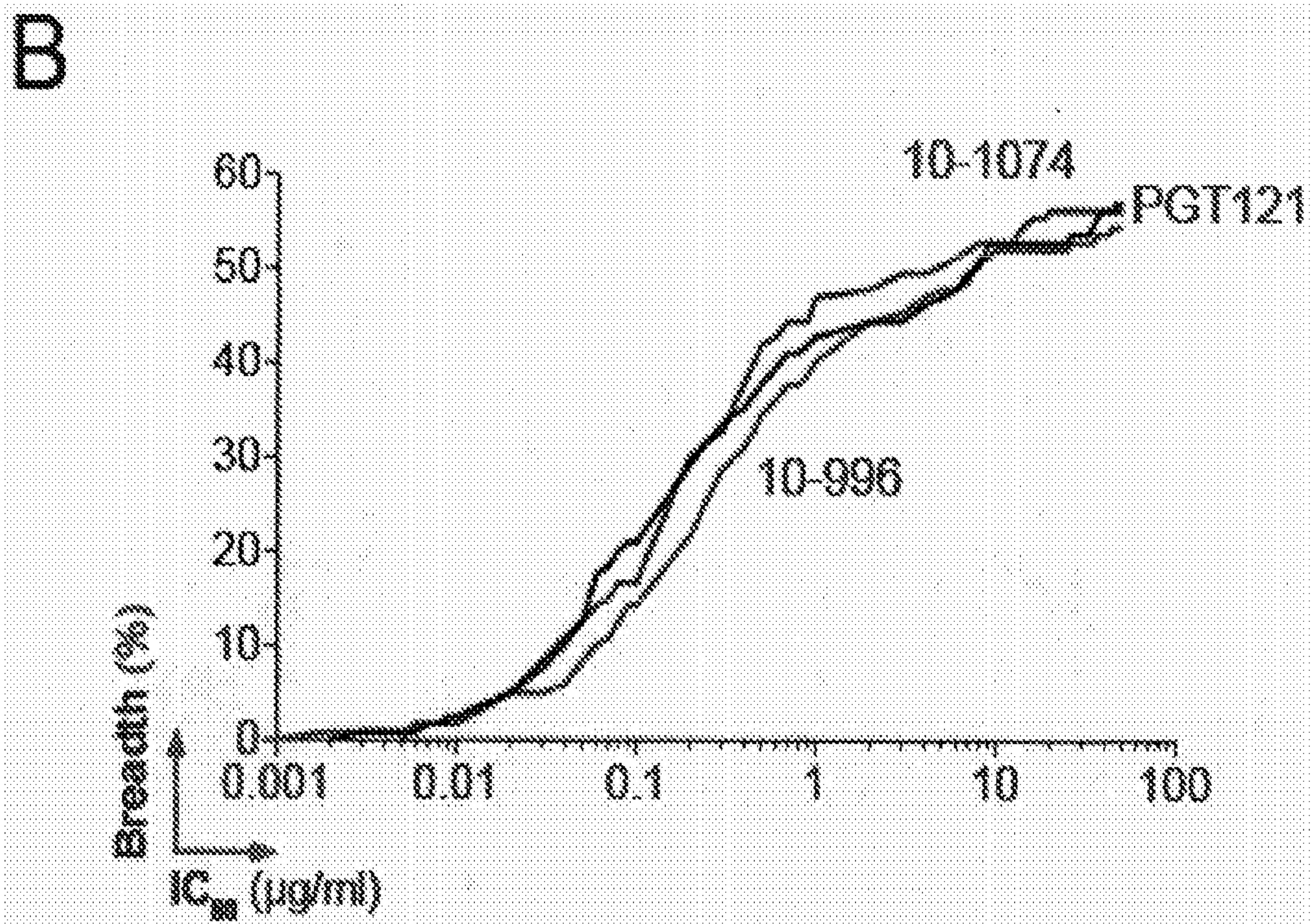


FIG. 9

**FIG. 9 (cont'd)**

C

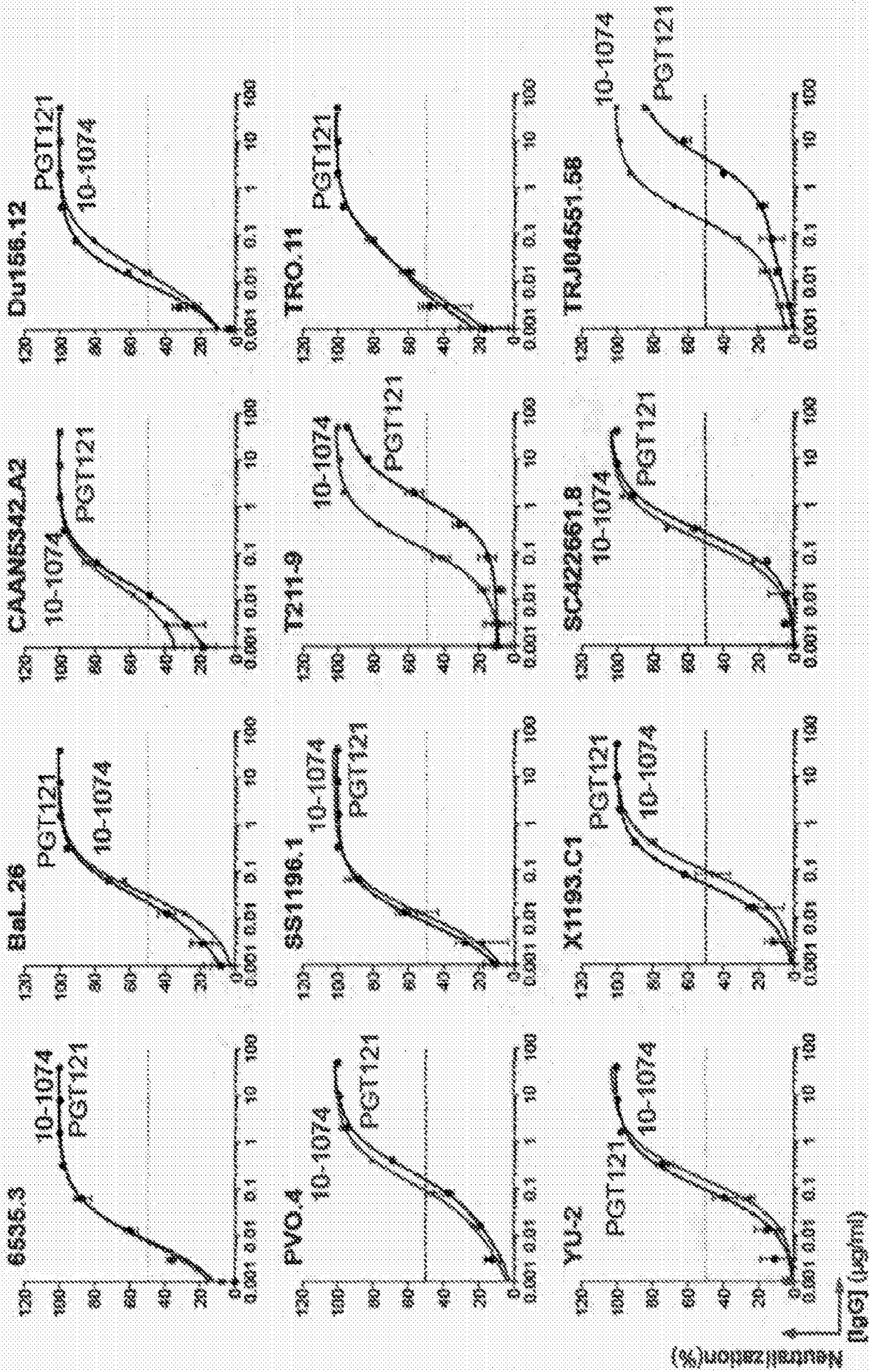


FIG. 9 (cont'd)

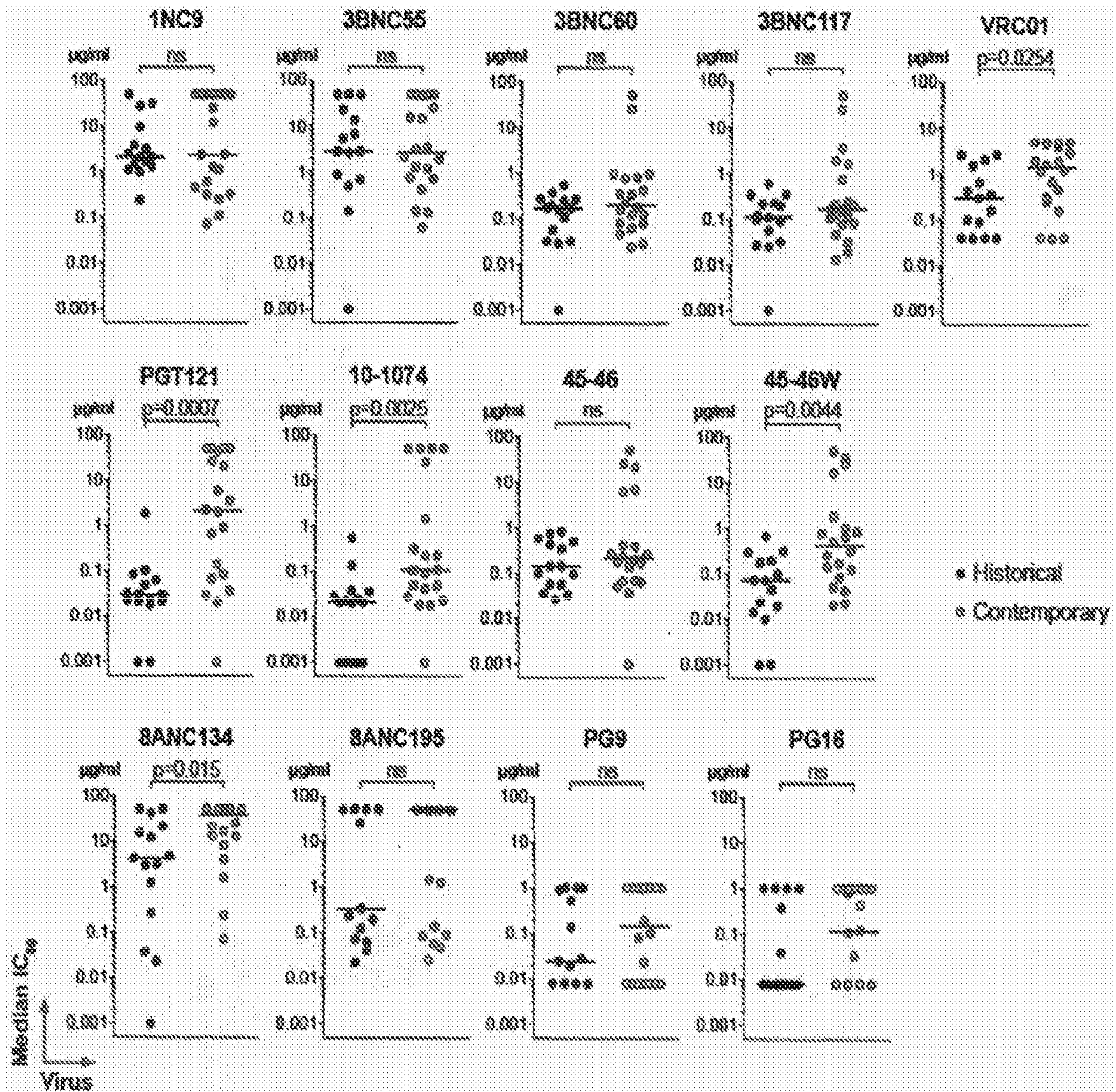


FIG. 10

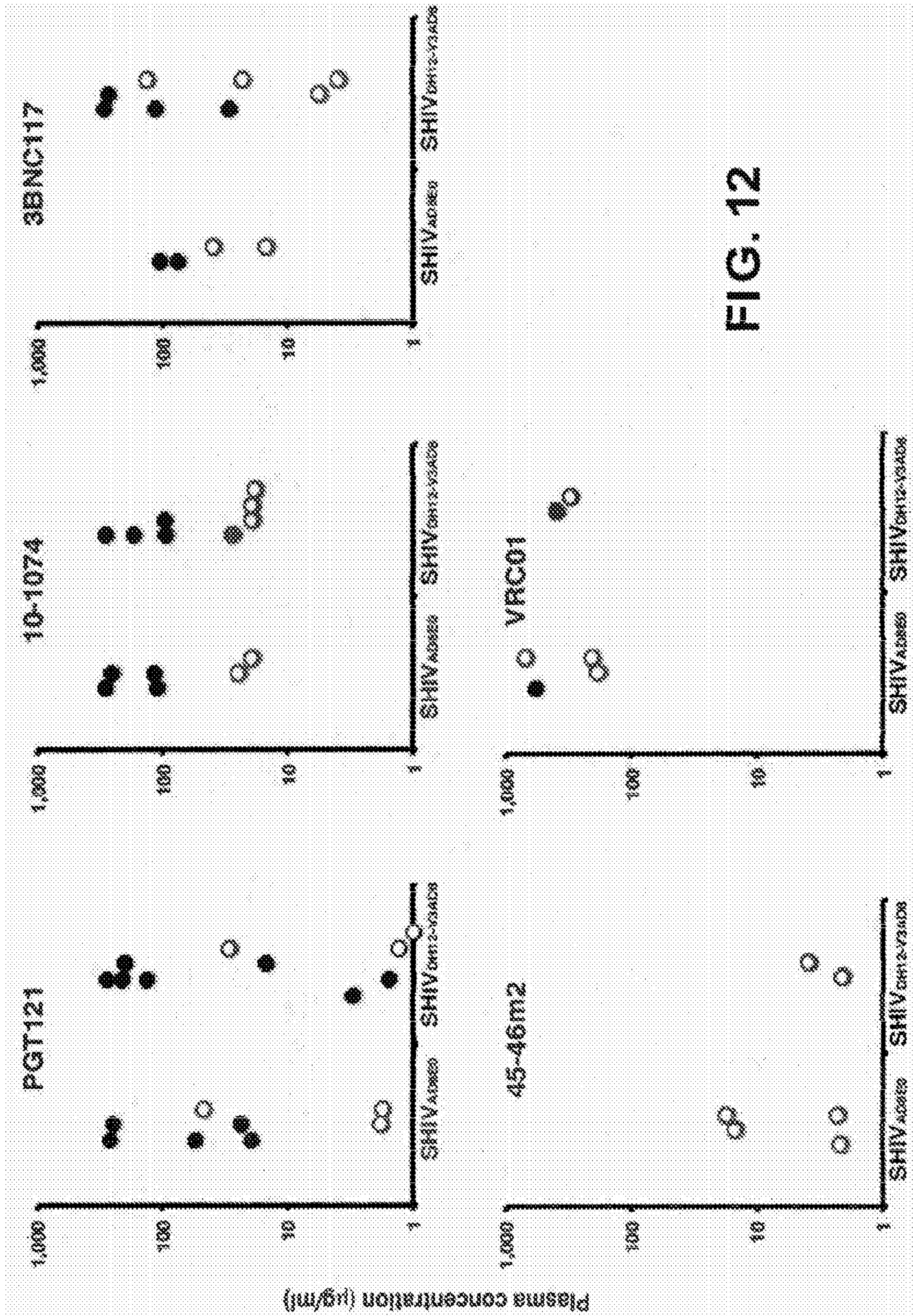
A

	IC50 TZM-bl ($\mu\text{g/ml}$)
VRC01	0.94
NIH 45-46	0.92
45-46m2	0.29
45-46 G54W	0.28
3BNC117	0.14
12A12	6.36
1NC9	2.04
8ANC195	3.38
10-1074	0.15
PGT-121	0.10
PGT-126	0.09

B

	IC50 TZM-bl ($\mu\text{g/ml}$)
VRC01	2.35
NIH 45-46	1.48
45-46m2	0.52
45-46 G54W	0.44
3BNC117	0.39
12A12	86.27
1NC9	30.26
8ANC195	3.08
10-1074	0.06
PGT-121	0.01
PGT-126	0.16

FIG. 11



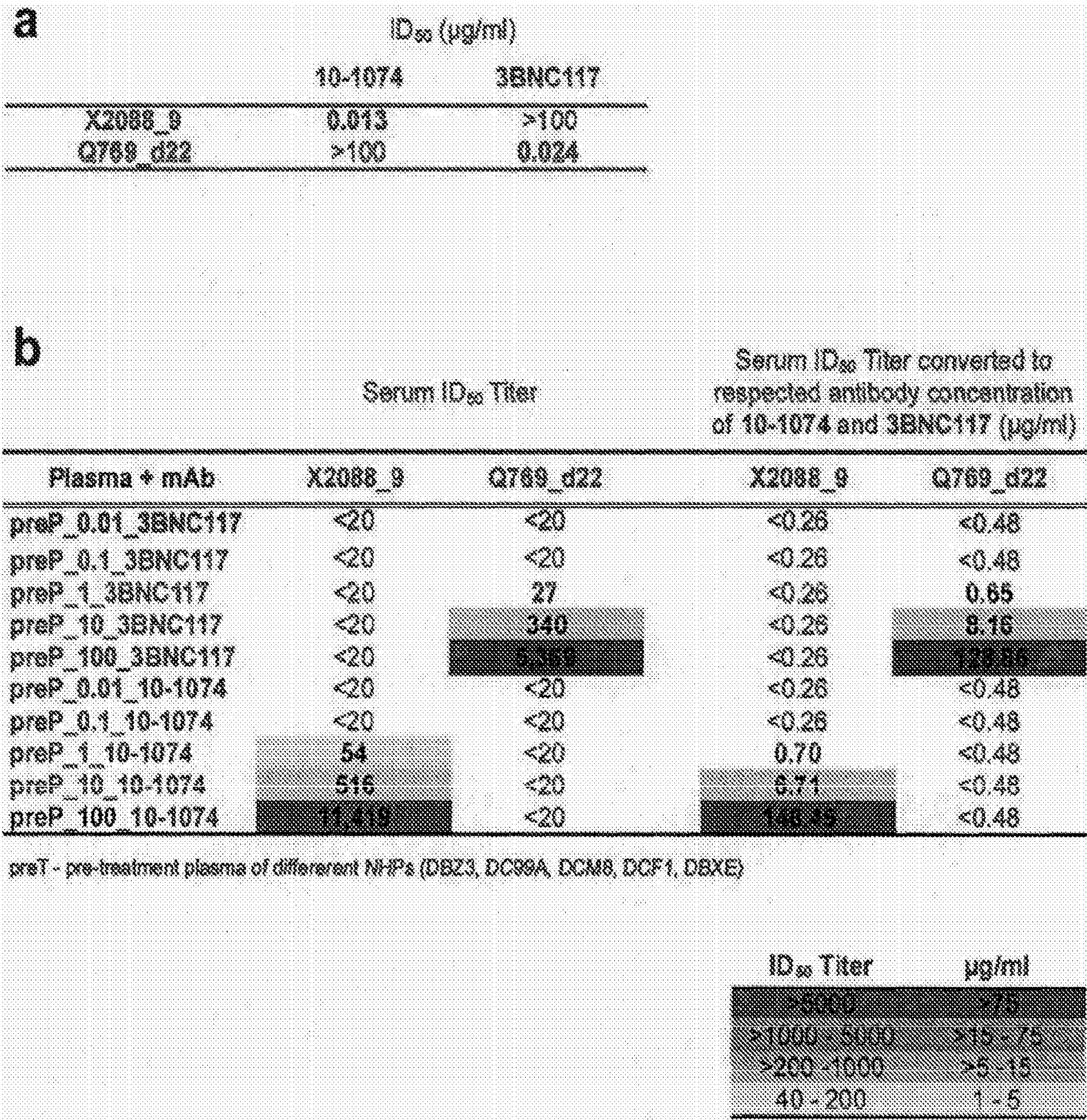


FIG. 13

mAbs levels in NHPs

C

		Plasma ID ₅₀ Titer		Plasma ID ₅₀ Titer converted to respected antibody concentration of 18-1074 and 3BNC117 (µg/ml)	
NHP	days post injection	X2888_9	Q759_622	18-1074	3BNC117
DBZ3	Prebleed	<20	<20	<0.26	<0.48
	3	1500	3179	20.21	25.22
	6	1734	1459	22.12	24.04
	10	3150	848	40.36	20.35
	14	3114	567	37.48	13.61
	17	1331	317	17.38	7.61
	20	1300	188	16.90	4.51
	24	485	<20	6.31	<0.48
	27	333	<20	4.33	<0.48
	34	85	<20	1.11	<0.48
	41	<20	<20	<0.26	<0.48
DCMB	49	<20	<20	<0.26	<0.48
	Prebleed	<20	<20	<0.26	<0.48
	3	1555	1977	20.24	27.45
	6	1794	1388	22.12	23.34
	10	2432	877	31.62	21.08
	14	1222	477	16.34	11.45
	17	787	249	10.36	5.88
	20	298	37	3.85	0.88
DBXE	23	148	<20	1.82	<0.48
	Prebleed	<20	24	<0.26	0.58
	3	1554	1495	20.21	26.93
	6	1827	927	22.12	22.26
	10	1445	504	18.80	12.10
	14	957	242	12.44	6.81
	17	686	208	9.85	4.99
	20	418	180	6.40	4.56
DC99A	23	378	131	4.91	3.14
	Prebleed	<20	<20	<0.26	<0.48
	3	2522	3506	28.12	35.54
	6	1874	1685	19.88	24.28
	10	1756	1852	18.84	36.73
	14	1429	717	14.56	17.21
	17	2655	647	34.92	15.83
	20	2247	540	28.15	12.86
	24	1639	384	21.28	9.22
	27	2436	676	31.71	13.82
	34	1007	185	13.01	4.66
DCF1	41	968	<20	12.68	<0.48
	49	461	35	5.99	0.84
	Prebleed	<20	<20	<0.26	<0.48
	3	<20	2134	<0.26	63.34
	6	29	1172	0.38	28.33
	10	<20	133	<0.26	3.19
	14	21	35	0.27	0.84
	17	<20	30	<0.26	0.72
	20	23	28	0.30	0.67
	23	<20	23	<0.26	0.58

ID₅₀ Titer

µg/ml

<20	<0.26
20-100	0.27-0.38
>200-1000	>0.39-1.19
40-200	1-5

FIG. 13 (cont'd)

