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(57) Abrégé/Abstract:

VEGF-B polypeptides from the PDGF family of growth factors having the property of promoting mitosis and proliferation of vascular endothelial cells, DNA sequences encoding these polypeptides, pharmaceutical compositions containing them and antibodies which react with them. The VEGF-B polypeptides are useful in stimulating angiogenesis as well as in diagnostic applications.

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<p>(21) International Application Number: PCT/US96/02957</p> <p>(22) International Filing Date: 1 March 1996 (01.03.96)</p> <p>(30) Priority Data:</p> <table border="0"> <tr> <td>08/397,651</td> <td>1 March 1995 (01.03.95)</td> <td>US</td> </tr> <tr> <td>08/469,427</td> <td>6 June 1995 (06.06.95)</td> <td>US</td> </tr> <tr> <td>08/569,063</td> <td>6 December 1995 (06.12.95)</td> <td>US</td> </tr> </table> <p>(71) Applicants: LUDWIG INSTITUTE FOR CANCER RESEARCH [US/US]; 1345 Avenue of the Americas, New York, NY 10105 (US). HELSINKI UNIVERSITY LICENSING LTD., OY [FI/FI]; P.O. Box 26, FIN-00014 Helsinki (FI).</p> <p>(72) Inventors: ERIKSSON, Ulf; Haegervaegen 27, S-746 34 Balsta (SE). OLOFSSON, Birgitta; Kling, Hornsgatan 106, 3 tr, S-117 26 Stockholm (SE). ALITALO, Kari; Nyyrikintie 4 A, FIN-02100 Espoo (FI). PAJUSOLA, Katri; Kasteholmantie 4 A 8, FIN-00900 Helsinki (FI).</p> <p>(74) Agent: EVANS, Joseph, D.; Evenson, McKeown, Edwards & Lenahan, Suite 700, 1200 G Street N.W., Washington, DC 20005 (US).</p>		08/397,651	1 March 1995 (01.03.95)	US	08/469,427	6 June 1995 (06.06.95)	US	08/569,063	6 December 1995 (06.12.95)	US	<p>(81) Designated States: AU, CA, CN, FI, JP, NO, NZ, European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).</p> <p>Published <i>With international search report.</i> <i>With amended claims.</i></p>
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<p>(57) Abstract</p> <p>VEGF-B polypeptides from the PDGF family of growth factors having the property of promoting mitosis and proliferation of vascular endothelial cells, DNA sequences encoding these polypeptides, pharmaceutical compositions containing them and antibodies which react with them. The VEGF-B polypeptides are useful in stimulating angiogenesis as well as in diagnostic applications.</p>											

VASCULAR ENDOTHELIAL GROWTH FACTOR-B

Background of the Invention

Angiogenesis, or the proliferation of new capillaries from pre-existing blood vessels, is a fundamental process necessary for normal growth and development of tissues. It is a prerequisite for the development and differentiation of the vascular tree, as well as for a wide variety of fundamental physiological processes including embryogenesis, somatic growth, tissue and organ repair and regeneration, cyclical growth of the corpus luteum and endometrium, and development and differentiation of the nervous system. In the female reproductive system, angiogenesis occurs in the follicle during its development, in the corpus luteum following ovulation and in the placenta to establish and maintain pregnancy. Angiogenesis additionally occurs as part of the body's repair processes, e.g. in the healing of wounds and fractures. Angiogenesis is also a factor in tumor growth, since a tumor must continuously stimulate growth of new capillary blood vessels in order to grow.

Capillary blood vessels consist of endothelial cells and pericytes. These two cell types carry all of the genetic information to form tubes, branches and entire capillary networks. Specific angiogenic molecules can initiate this process. In view of the physiological importance of angiogenesis, much effort has been devoted to the isolation, characterization and purification of factors that can stimulate angiogenesis. A number of polypeptides which stimulate angiogenesis have been purified and

characterized as to their molecular, biochemical and biological properties. For reviews of such angiogenesis regulators, see Klagsbrun et al., "Regulators of Angiogenesis", *Ann. Rev. Physiol.*, 53:217-39 (1991); and 5 Folkman et al., "Angiogenesis," *J. Biol. Chem.*, 267:10931-934 (1992). Recent results have implicated several endothelial receptor tyrosine kinases (RTKs) in the establishment and maintenance of the vascular system.

One such growth factor, which is highly specific as a 10 mitogen for vascular endothelial cells, is termed vascular endothelial growth factor (VEGF). See Ferrara et al., "The Vascular Endothelial Growth Factor Family of Polypeptides," *J. Cellular Biochem.*, 47:211-218 (1991); Connolly, "Vascular Permeability Factor: A Unique Regulator of Blood Vessel 15 Function," *J. Cellular Biochem.*, 47:219-223 (1991). VEGF is a potent vasoactive protein that has been detected in media conditioned by a number of cell lines including bovine pituitary follicular cells. VEGF is a glycosylated cationic 46-48 kD dimer made up of two 24 kD subunits. It is 20 inactivated by sulfhydryl reducing agents, resistant to acidic pH and to heating, and binds to immobilized heparin. VEGF is sometimes referred to as vascular permeability factor (VPF) because it increases fluid leakage from blood vessels following intradermal injection. It also has been 25 called by the name vasculotropin.

Four different molecular species of VEGF have been detected. The 165 amino acid species has a molecular weight of approximately 46 kD and is the predominant molecular form found in normal cells and tissues. A less abundant, shorter 30 form with a deletion of 44 amino acids between positions 116 and 159 (VEGF₁₂₁), a longer form with an insertion of 24 highly basic residues in position 116 (VEGF₁₈₉), and another longer form with an insertion of 41 amino acids (VEGF₂₀₆), which includes the 24 amino acid insertion found in VEGF₁₈₉, 35 are also known. VEGF₁₂₁ and VEGF₁₆₅ are soluble proteins. VEGF₁₈₉ and VEGF₂₀₆ appear to be mostly cell-associated. All

of the isoforms of VEGF are biologically active. For example, each of the species when applied intradermally is able to induce extravasation of Evans blue.

5 The various species of VEGF are encoded by the same gene and arise from alternative splicing of messenger RNA. This conclusion is supported by Southern blot analysis of human genomic DNA, which shows that the restriction pattern is identical using either a probe for VEGF₁₆₅ or one which contains the insertion in VEGF₂₀₆. Analysis of genomic clones
10 in the area of putative mRNA splicing also shows an intron/exon structure consistent with alternative splicing.

The different isoforms of VEGF have different chemical properties which may regulate cellular release, compartmentalization, bioavailability and possibly also
15 modulate the signalling properties of the growth factors.

Analysis of the nucleotide sequence of the VEGF gene indicates that VEGF is a member of the platelet-derived growth factor (PDGF) family. VEGF and PlGF are ligands for two endothelial RTKs, flt-1 (VEGF receptor 1, VEGFR1) and
20 flk-1/KDR (VEGF receptor 2, VEGFR2). The amino acid sequence of VEGF exhibits approximately 20% homology to the sequences of the A and B chains of PDGF, as well as complete conservation of the eight cysteine residues found in both mature PDGF chains. VEGF₁₆₅, VEGF₁₈₉ and VEGF₂₀₆ also contain
25 eight additional cysteine residues within the carboxy-terminal region. The amino-terminal sequence of VEGF is preceded by 26 amino acids corresponding to a typical signal sequence. The mature protein is generated directly following signal sequence cleavage without any intervening
30 prosequence. The existence of a potential glycosylation site at Asn⁷⁴ is consistent with other evidence that VEGF is a glycoprotein, but the polypeptide has been reported to exist in both glycosylated and deglycosylated species.

Like other cytokines, VEGF can have diverse effects
35 that depend on the specific biological context in which it is found. VEGF and its high affinity receptors flt-1 and

KDR/flk-1 are required for the formation and maintenance of the vascular system as well as for both physiological and pathological angiogenesis. VEGF is a potent endothelial cell mitogen and directly contributes to induction of angiogenesis *in vivo* by promoting endothelial cell growth during normal embryonic development, wound healing, and tissue regeneration and reorganization. VEGF is also involved in pathological processes such as growth and metastasis of solid tumors and ischemia-induced retinal disorders. A most striking property of VEGF is its specificity. It is mitogenic *in vitro* at 1 ng/ml for capillary and human umbilical vein endothelial cells, but not for adrenal cortex cells, corneal or lens epithelial cells, vascular smooth muscle cells, corneal endothelial cells, granulosa cells, keratinocytes, BHK-21 fibroblasts, 3T3 cells, rat embryo fibroblasts, human placental fibroblasts and human sarcoma cells. The target cell specificity of VEGF is thus restricted to vascular endothelial cells. VEGF can trigger the entire sequence of events leading to angiogenesis and stimulates angiogenesis *in vivo* in the cornea and in a healing bone graft model. It is able to stimulate the proliferation of endothelial cells isolated from both small and large vessels. Expression of VEGF mRNA is temporally and spatially related to the physiological proliferation of capillary blood vessels in the ovarian corpus luteum or in the developing brain. VEGF expression is triggered by hypoxia so that endothelial cell proliferation and angiogenesis appear to be especially stimulated in ischemic areas. VEGF is also a potent chemoattractant for monocytes. In addition, VEGF induces plasminogen activator and plasminogen activator inhibitor in endothelial cells.

Tumor cells release angiogenic molecules such as VEGF, and monoclonal antibodies to VEGF have been shown to inhibit the growth of certain types of tumor such as rhabdomyosarcoma. See Kim et al., "Inhibition of Vascular

Endothelial Growth Factor-Induced Angiogenesis Suppresses
Tumor Growth *in vivo*," *Nature*, 362:841-844 (1993). This
suggests that blocking VEGF action is of potential
therapeutic significance in treating tumors in general, and
5 highly-vascularized, aggressive tumors in particular.

Summary of the Invention

It is an object of the invention to provide a new
growth factor having the property of promoting proliferation
10 of endothelial cells.

Another object of the invention is to provide isolated
DNA sequences which encode a new growth factor which
promotes proliferation of endothelial cells.

It is also an object of the invention to provide new
15 products which may be useful in diagnostic and/or
therapeutic applications.

These and other objects are achieved in accordance with
the present invention by providing an isolated DNA which
codes for a protein exhibiting the following characteristic
20 amino acid sequence (SEQ ID NO:16):

Pro-Xaa-Cys-Val-Xaa-Xaa-Xaa-Arg-Cys-Xaa-Gly-Cys-Cys
and having the property of promoting proliferation of
endothelial cells or mesodermal cells, the DNA being
selected from the group consisting of the DNA of Figures 1
25 and 2 (SEQ ID NO:1), the DNA of Figure 3 (SEQ ID NO:4), the
DNA of Figure 5 (SEQ ID NO:6); the DNA of Figure 7 (SEQ ID
NO:8), the DNA of Figure 10 (SEQ ID NO:10), the DNA of
Figure 12 (SEQ ID NO:12), the DNA of Figure 14 (SEQ ID
NO:14), and DNA's which hybridize under stringent conditions
30 with at least one of the foregoing DNA sequences.

In accordance with further aspects of the invention,
the objects are also achieved by providing a protein
exhibiting the following characteristic amino acid sequence

Pro-Xaa-Cys-Val-Xaa-Xaa-Xaa-Arg-Cys-Xaa-Gly-Cys-Cys
35 (SEQ ID NO:16) and having the property of promoting
proliferation of endothelial cells or mesodermal cells,

which protein comprises a sequence of amino acids substantially corresponding to an amino acid sequence selected from the group consisting of the amino acid sequence of Figure 1 (SEQ ID NO:2), the amino acid sequence of Figure 2 (SEQ ID NO:3), the amino acid sequence of Figure 4 (SEQ ID NO:5), the amino acid sequence of Figure 6 (SEQ ID NO:7), the amino acid sequence of Figure 8 (SEQ ID NO:9), the amino acid sequence of Figure 11 (SEQ ID NO:11), the amino acid sequence of Figure 13 (SEQ ID NO:13), and the amino acid sequence of Figure 15 (SEQ ID NO:15).

In further aspects of the invention, the objects are achieved by providing pharmaceutical preparations which comprise such proteins; and by providing antibodies which react with or recognize such proteins.

The novel growth factor of the present invention, referred to hereinafter as vascular endothelial growth factor B or VEGF-B, has close structural similarities to VEGF and to placenta growth factor (PlGF). All of the VEGF-B forms contain the characteristic amino acid sequence

Pro-Xaa-Cys-Val-Xaa-Xaa-Xaa-Arg-Cys-Xaa-Gly-Cys-Cys (SEQ ID NO:16) (wherein Xaa represents a variable residue), which is an earmark of the PDGF/VEGF family of growth factors. This characteristic amino acid sequence can be found at amino acids 70 to 82 in Figures 4, 6, 8, 11, 13 and 15.

Clinical applications of the invention include diagnostic applications, acceleration of angiogenesis in wound healing, and inhibition of angiogenesis. Quantitation of VEGF-B in cancer biopsy specimens may be useful as an indicator of future metastatic risk. Topical application of VEGF-B preparations to chronic wounds may accelerate angiogenesis and wound healing. VEGF-B may be used in a manner analogous to VEGF.

According to yet further aspects of the invention, the objects are achieved by providing diagnostic/prognostic means typically in the form of test kits. For example, in

one embodiment of the invention there is provided a diagnostic/prognostic test kit comprising antibodies to the new growth factor of the invention and means for detecting, and more preferably evaluating, binding between the antibodies and the new growth factor of the invention. In one preferred embodiment of the diagnostic/prognostic means according to the invention, either the antibody or the new growth factor is labelled, and either the antibody or the growth factor is substrate-bound, such that the growth factor-antibody interaction can be established by determining the amount of label attached to the substrate following binding between the antibody and the growth factor. In a particularly preferred embodiment of the invention, the diagnostic/prognostic means may be provided as a conventional ELISA kit.

In another alternative embodiment, the diagnostic/prognostic means may comprise PCR means for establishing the genomic sequence structure of a VEGF-B gene of a test individual and comparing this sequence structure with that disclosed in this application in order to detect any abnormalities, with a view to establishing whether any aberrations in VEGF-B expression are related to a given disease condition.

A yet further aspect of the invention concerns an antibody which recognizes VEGF-B and which is suitably labelled.

Another aspect of the invention concerns the provision of a pharmaceutical composition comprising either VEGF-B protein or antibodies thereto. Compositions which comprise VEGF-B protein may optionally further comprise either VEGF or heparin or both.

According to an additional aspect of the invention the manufacture of a medicament is provided which comprises VEGF-B protein and heparin for treating conditions characterized by lack of, or reduction in, angiogenesis.

In another aspect, the invention relates to a protein dimer comprising VEGF-B protein, particularly a disulfide-linked dimer. The protein dimers of the invention include both homodimers of VEGF-B protein and heterodimers of VEGF-B and VEGF.

According to a yet further aspect of the invention there is provided a method for facilitating release of VEGF and/or VEGF-B from a cell comprising exposing a cell which expresses either or both of the aforementioned growth factors to heparin.

Another aspect of the invention involves providing a vector comprising an anti-sense nucleotide sequence which is complementary to at least a part of the DNA sequences disclosed herein which encode the new growth factor of the invention which promotes proliferation of endothelial cells. According to a yet further aspect of the invention such a vector comprising an anti-sense sequence may be used to inhibit, or at least mitigate, VEGF-B expression. The use of a vector of this type to inhibit VEGF-B expression is favored in instances where VEGF-B expression is associated with a disease such as in instances where tumors produce VEGF-B in order to provide for angiogenesis. Transformation of such tumor cells with a vector containing an anti-sense nucleotide sequence would suppress or retard angiogenesis and so would inhibit or retard growth of the tumor.

Brief Description of the Drawings

Figure 1 shows the nucleotide sequence of the (partial) cDNA clone of VEGF-B (SEQ ID NO:1) and the amino acid sequence of the protein segment (SEQ ID NO:2) coded by the first reading frame of the cDNA;

Figure 2 repeats the nucleotide sequence of the (partial) cDNA clone of VEGF-B (SEQ ID NO:1) and the amino acid sequence of the protein segment (SEQ ID NO:3) coded by the second reading frame of the cDNA;

Figure 3 shows the nucleotide sequence of the coding region of a full length cDNA clone of murine VEGF-B₁₆₇ (SEQ ID NO:4);

5 Figure 4 shows the amino acid sequence of murine VEGF-B₁₆₇ (SEQ ID NO:5);

Figure 5 shows the nucleotide sequence of the coding region of a cDNA clone of VEGF-B₁₇₄ (SEQ ID NO:6);

Figure 6 shows the amino acid sequence of VEGF-B₁₇₄ (SEQ ID NO:7);

10 Figure 7 shows the nucleotide sequence of a cDNA clone of VEGF-B₁₁₂ (SEQ ID NO:8);

Figure 8 shows the amino acid sequence of VEGF-B₁₁₂ (SEQ ID NO:9);

15 Figure 9 shows a comparison of the amino acid sequences of mVEGF-B₁₆₇, mVEGF₁₆₄, hPlGF, mPDGF A, and mPDGF B;

Figure 10 shows the nucleotide sequence of a clone of human VEGF-B₁₆₇ (SEQ ID NO:10);

Figure 11 shows the amino acid sequence of human VEGF-B₁₆₇ (SEQ ID NO:11); and

20 Figure 12 shows the nucleotide sequence of murine VEGF-B₁₈₆ (SEQ ID NO:12);

Figure 13 shows the amino acid sequence of murine VEGF-B₁₈₆ (SEQ ID NO:13);

25 Figure 14 shows the nucleotide sequence of human VEGF-B₁₈₆ (SEQ ID NO:14);

Figure 15 shows the amino acid sequence of human VEGF-B₁₈₆ (SEQ ID NO:15);

30 Figure 16 shows an amino acid sequence comparison of murine and human VEGF-B₁₆₇ and VEGF-B₁₈₆ isoforms (SEQ ID NOS: 5, 11, 13 & 15).

Figure 17 shows the schematic structure of mouse and human genes for VEGF-B;

Figure 18 shows a hydrophilicity analysis of murine VEGF-B₁₆₇ and VEGF-B₁₈₆ isoforms;

35 Figure 19 shows a phylogenetic analysis of the VEGF/PDGF family of growth factors;

Figure 20 is a graph showing the induction of [³H]thymidine incorporation by VEGF-B, VEGF and bFGF for human umbilical vein endothelial cells (HUVEC) and bovine capillary endothelial (BCE) cells;

5 Figure 21 is a Northern blot analysis of the expression of VEGF-B₁₈₆ transcripts in several mouse and human tissues;

Figure 22 shows the results of immunoprecipitation and SDS-PAGE analysis of cell culture media and detergent solubilized cell lysates from Cos-1 cells transiently
10 transfected with a murine VEGF-B cDNA;

Figure 23A shows the results of immunoprecipitation and SDS-PAGE analysis of cell culture media from transfected Cos-1 cells separately expressing murine VEGF-B₁₈₆ and human VEGF₁₆₅;

15 Figure 23B shows the results of immunoprecipitation and SDS-PAGE analysis of cell culture media (M) and detergent solubilized cell lysates (L) of Cos-1 cells which coexpress murine VEGF-B₁₈₆ and human VEGF₁₆₅;

Figure 23C shows the results of immunoprecipitation and
20 SDS-PAGE analysis of cell culture media from Cos-1 cells expressing murine VEGF-B₁₈₆ and human VEGF, either separately or in combination, and from mock transfected control cells;

Figure 24 is a schematic illustration of the derivation of VEGF-B promoter-reporter clones; and

25 Figure 25 shows the nucleotide sequence of a 1.55 kb human VEGF-B promoter fragment (SEQ ID NO:17).

Detailed Description of Preferred Embodiments

The present invention thus is directed to new vascular
30 endothelial growth factors, hereinafter referred to as VEGF-B growth factors, which share the angiogenic and other properties of VEGF, but which are distributed and expressed in tissues differently from VEGF.

35 VEGF-B growth factors are members of the family of platelet derived growth factors and are a growth factors which promote mitosis and proliferation of vascular

endothelial cells and/or mesodermal cells. They are produced by expression of DNA sequences which correspond to, or which are hybridizable under stringent conditions with, any one of the DNA sequences depicted in Figures 1 and 2 (SEQ ID NO:1), Figure 3 (SEQ ID NO:4), Figure 5 (SEQ ID NO:6), Figure 7 (SEQ ID NO:8), Figure 10 (SEQ ID NO:10), Figure 12 (SEQ ID NO:12) or Figure 14 (SEQ ID NO:14). It is intended to include within the scope of the invention all angiogenic proteins encoded by DNA sequences which hybridize under stringent conditions to any one of the foregoing DNA sequences. Suitable hybridization conditions include, for example, 50% formamide, 5 x SSPE buffer, 5 x Denhardt's solution, 0.5% SDS and 100 μ g/ml of salmon sperm DNA at 42°C overnight, followed by washing 2 x 30 minutes in 2 x SSC at 55°C.

The invention is also directed to an isolated and/or purified DNA which corresponds to, or which hybridizes under stringent conditions with, any one of the foregoing DNA sequences.

In a further aspect, the invention is directed to antibodies of VEGF-B growth factors, and particularly to monoclonal antibodies.

VEGF-B proteins are believed to interact with protein tyrosine kinase growth factor receptors. Details of such receptors are known in the art [See e.g. Wilks, A.F., "Protein Tyrosine Kinase Growth Factor Receptors and Their Ligands in Development, Differentiation, and Cancer," *Adv. Cancer Res.*, 60:43-73 (1993)].

Various adult mouse tissues were tested for expression of transcripts corresponding to VEGF-B by Northern blotting. The size of the mRNA was 1.3-1.4 kb. A mouse multiple tissue Northern blot (MTN, Clontech) was probed with the \approx 0.9 kb Sall/NotI fragment derived from the pPC67 yeast expression vectors described above. The probe was labelled with 32 P-dCTP using random priming (specific activity 10^8 - 10^9 cpm/ μ g of DNA). The blot was hybridized overnight at 42°C

using 50% formamide, 5 x SSPE buffer, 2% SDS, 10 x Denhardt's solution, 100 µg/ml salmon sperm DNA and 1x10⁶ cpm of the labelled probe/ml. The blot was washed at room temperature for 2 x 30 min in 2 x SSC containing 0.05% SDS and then for 2 x 20 min at 52°C in 0.1 x SSC containing 0.1% SDS. The blot was then exposed at -70°C for three days using intensifying screens. Kodak XAR film was used. The relative expression levels as determined by visual examinations of the film are listed in the following table:

10

Table 1

Distribution of VEGF-B Transcripts in the Adult Mouse

Tissue	Relative Expression Level
Heart	+++++
Brain	+++
Spleen	(+)
Lung	++
Liver	+
Skeletal Muscle	++++
Kidney	+++
Testis	(+)

15

20

+++++ = very strong expression; ++ = rather weak expression;
 ++++ = strong expression; + = weak expression;
 +++ = moderate expression; (+) = very weak expression.

25

A human multiple tissue Northern blot (MNT) from Clontech was probed using the murine partial cDNA to determine relative VEGF-B expression levels in various human tissues. The size of the transcript was 1.3-1.4 kb. The conditions were identical to those used for the mouse Northern blot described above. The relative VEGF-B transcript levels for the human Northern blot are listed in the following Table 2. For comparison purposes, Table 2

30

also lists relative expression level data from the literature for VEGF in various mammalian systems.

Table 2

5 Tissues	Relative Expression Levels			
	VEGF-B (Northern blot)	VEGF (from literature)		
	human	human	murine	guinea pig
heart	+++++	++	+++	+++
brain	+		+	+
placenta	+			
10 lung	+	++++		++
liver	(+)	++	(+)	+
skeletal muscle	++++		+++	+
kidney	+	++	+	++
15 pancreas	+++			
spleen	++		-	+
thymus	+		-	
prostate	+++			
testis	++			(+)
20 ovary	+++			-
small intestine	++			
colon	+++			
25 peripheral blood leucocytes	+			

From a comparison of Table 1 and Table 2 it can be seen that mouse and human tissue expression levels of VEGF-B transcripts are relatively similar with the highest expression levels being found in heart and skeletal muscle. Significant differences may be seen in brain and kidney tissue. It should also be noted that tissues containing a

large proportion of both muscular and epithelial cells, such as prostate, pancreas and colon from which some of the most common human tumors originate, express relatively high levels of VEGF-B.

5 A comparison of the relative expression levels of VEGF and VEGF-B in human tissues shows some striking differences. VEGF is expressed rather weakly by human heart tissue, but VEGF-B is very strongly expressed by the same tissue. On the other hand, VEGF is strongly expressed by human lung
10 tissue, but VEGF-B is only weakly expressed by human lung tissue. In a similar vein, human liver tissue expresses VEGF at a moderate level, but VEGF-B is expressed only very weakly. These data evidence that despite their general similarities, the actions of VEGF and VEGF-B are not
15 completely identical.

The expression of VEGF-B transcripts was further analyzed in mouse and human tissues by Northern blotting and compared with the expression of VEGF transcripts. Mouse and human multiple tissue Northern (MTN) blots (Clontech) were
20 hybridized with a ³²P-labelled mouse VEGF-B probe (≈0.9 kb SalI/NotI insert of the clone pcif 2). VEGF expression was analyzed with ³²P-labelled VEGF₁₆₅ cDNA as the probe. The hybridizations were carried out at 42°C in 50% deionized formamide, 5 x SSC pH 7.0, 1% SDS, 5 x Denhardt's solution
25 and 100 µg/ml of denatured salmon sperm DNA. The filters were washed 2 x 30 min at 52°C in 2 x SSC containing 0.5% SDS and exposed to Kodak XAR film for 2-5 days at -70°C using intensifying screens. In situ hybridization analysis of adult mouse tissues from CBA mice and of embryos derived
30 from matings of CBA and NMRI mice were carried out essentially as previously described by Korhonen et al., *Blood*, 80, 2548-55 (1992). The RNA probes (a 383 bp antisense probe and a 169 bp sense probe) were generated from a linearized plasmid containing a 440 bp Sal I/Sac I
35 fragment derived from the pcif 2 cDNA clone. Radiolabelled RNA was synthesized using T7 and SP6 RNA polymerases and

[³⁵S]UTP (Amersham Inc.). Alkaline hydrolysis of the probes was omitted. Hematoxylin was used for counterstaining. Control hybridizations with sense strand and RNase A-treated sections did not give signals above background.

5 In mouse tissues the most abundant expression of the 1.4 kb VEGF-B transcript was detected in heart, brain, skeletal muscle, and kidney. The major 3.7 kb VEGF transcript was is expressed in heart, brain, lung, skeletal muscle and kidney. In human tissues, the most abundant
10 expression of the 1.4 kb VEGF-B transcript and the major 3.7 and 4.5 kb VEGF transcripts were detected in heart, skeletal muscle, pancreas and prostate. Thus, although clear quantitative differences exist, it appears that VEGF-B and VEGF are coexpressed in many human and mouse tissues.

15 The expression of VEGF-B transcripts was further examined by in situ hybridization in sections from adult mouse heart and skeletal muscle and from the early (E 10) mouse embryo. In the adult heart, VEGF-B transcripts are prominently expressed in the myocardium, while no specific
20 signal is detected in arterial smooth muscle. In adult striated muscle, VEGF-B transcripts are expressed by some of the myofibers whereas others seem to lack the transcript. In the E 10 mouse embryo, VEGF-B transcripts are detected mainly in the developing heart. The myocardium of the adult
25 mouse heart has a prominent signal. In striated muscle, VEGF-B expression is seen in subpopulations of myofibers. Strong signals were also obtained in the developing heart of the E10 mouse embryo. Other embryonic structures expressed lower or undetectable levels of transcripts for VEGF-B.
30 Taken together, these tests indicate that VEGF-B transcripts are expressed primarily in muscular tissues. VEGF-B is particularly abundant in heart and skeletal muscle and is co-expressed with VEGF in these and other tissues. In transfected cells, VEGF-B forms cell surface associated,
35 disulfide-linked homodimers and heterodimers with VEGF when coexpressed. A Northern blot analysis of the expression of

VEGF-B₁₈₆ transcripts in several mouse and human tissues using a VEGF-B₁₈₆ isoform specific probe is shown in Figure 21.

5 The chromosomal location of the VEGF-B gene was assessed by Southern blotting and polymerase chain reaction analysis of somatic cell hybrids and fluorescence *in situ* hybridization (FISH) of metaphase chromosomes. The VEGF-B gene was found on chromosome 11q13, proximal of the cyclin D1 gene. It is interesting that although the cyclin D1 gene is amplified in a number of human carcinomas, the VEGF-B
10 gene was not amplified in several mammary carcinoma cell lines which contained amplified cyclin D1. Nevertheless, mutations in the VEGF-B gene may be related to vascular malformations and/or cardiovascular diseases.

Unless otherwise indicated, the following Examples used
15 standard molecular biology techniques and procedures as disclosed in Ausubel et al. (eds.), *Current Protocols in Molecular Biology*, John Wiley & Sons, New York (1992).

Example 1: Partial cDNA clone with two reading frames.

20 A partial cDNA clone encoding murine VEGF-B was identified as follows. A cDNA library (E 14.5) derived from poly A+ mRNA isolated from 14.5 day old mouse embryos [Chevray P. and Nathans D., "Protein interaction cloning in yeast: Identification of mammalian proteins that react with
25 the leucine zipper of Jun," *Proc. Natl. Acad. Sci. USA*, 89:5789-93 (1992)] was screened for cellular proteins which potentially might interact with cellular retinoic acid-binding protein type 1 (CRABP-I) using a yeast two-hybrid interaction trap screening technique as described by Gyuris
30 J., Golemis E., Chertkov H. and Brent R., "Cdi1, a Human G1 and S Phase Protein Phosphatase That Associates with Cdk2," *Cell*, 75:791-803 (1993). This screening technique involves a fusion protein that contains a binding domain and that is known to be transcriptionally inert (the "bait"); reporter
35 genes that have no basal transcription and that are bound by the bait; and an expression library which encodes proteins

expressed as chimeras and whose amino termini contain an activation domain and other useful moieties (the "prey"). The screened library was a 14.5 day mouse embryo plasmid library in the yeast expression vector pPC67 obtained from
5 Dr. Pierre Chevray of the Johns Hopkins University, School of Medicine, 725 North Wolfe St., Baltimore, MD 21205. A positive cDNA clone (pcif-2) was recovered from the screening. The positive clone was sequenced using well known, conventional techniques and found to encode a protein
10 highly homologous to VEGF and the other members of the PDGF family of growth factors. The ≈0.9 kb SalI/NotI insert in the plasmid pPC67 was cloned into pBluescript and fully sequenced using T7 and T3 vector primers together with internal primers. The plasmid pBluescript is commercially
15 available from Stratagene Inc., LaJolla, California. The cDNA insert was found to be 886 base pairs long and to encode two polypeptides in different reading frames which were homologous to the N-terminal end and the C-terminal end, respectively, of VEGF. This novel growth factor is
20 referred to hereinafter as VEGF-B. The clone is partial and lacks several amino acids in the amino terminal region and the entire signal sequence.

Figure 1 shows the nucleotide sequence (SEQ ID NO:1) of this partial cDNA clone of VEGF-B and the amino acid
25 sequence (SEQ ID NO:2) encoded in the first reading frame thereof. The DNA sequence of Figure 1 was obtained by conventional sequencing of a clone (pcif-2) in the yeast expression vector pPC67. The clone comprised 886 base pairs and encoded a part of murine VEGF-B.

30 The isolated cDNA sequence will hybridize with the mammalian genomic DNA, e.g. either murine or human, which contains the VEGF-B gene. In addition to the coding sequence, the genomic DNA will contain one or more promoter sequence(s) which give and direct expression of VEGF-B in
35 one or more specific tissues. Thus the coding sequence of VEGF-B may be linked to muscle-specific promoter elements

which in turn are specific to certain types of muscle fibers.

The nucleotide sequence is translated in two different reading frames into two different amino acid sequences. There is a stop codon (TGA) within the coding sequence at base pairs 309-311. Thus, VEGF-B comes in several splicing variants. The 5' end of the cloned cDNA sequence encodes an 102 amino acid long protein with significant homology to the N-terminal domains of VEGF, PlGF and PDGF A and B. In particular, a number of cysteine residues are perfectly conserved within this group of proteins. In addition to the nucleotide sequence (SEQ ID NO:1), Figure 1 further depicts the deduced amino acid sequence (SEQ ID NO:2) of this first protein.

Translation of the C-terminal end of the cDNA (base pairs 308-475) in a different reading frame results in a protein which is highly homologous to the C-terminal part of VEGF₁₆₅, VEGF₁₈₉ and VEGF₂₀₆. Figure 2 again shows the nucleotide sequence (SEQ ID NO:1) of Figure 1, but this time includes the deduced amino acid sequence (SEQ ID NO:3) of the second protein, which is encoded in the second reading frame and is 54 amino acids long. It thus appears that the VEGF-B gene encodes different proteins using alternative splicing of the primary transcript. The last part of the clone, encoding the second peptide might be expressed as a functional protein in other spliced variants of VEGF-B.

The aforescribed proteins may exist in combined association with an additional N-terminal sequence of approximately five (5) to ten (10) amino acids, as well as a further leader sequence of approximately twenty-one (21) to twenty-eight (28) amino acids. Inasmuch such combined amino acid sequences exhibit the property of promoting the proliferation of endothelial cells and the DNA sequences which code for such combined peptide sequences will hybridize under stringent conditions with the DNA sequence of Figures 1 and 2, such amino acid sequences and the DNA

which codes for them are expressly contemplated to be within the scope of the present invention.

Example 2: Cloning of full length cDNA's for mouse VEGF-B.

5 Using the approximately 0.9 kb SalI/NotI cDNA insert of the previously identified cDNA clone of Example 1 as a probe, an adult mouse heart lambda ZAP-II cDNA library obtained from Stratagene Inc., of La Jolla, California was screened using standard techniques. The library was
10 titrated and plated as recommended and filters were prepared. Following prehybridization at 42 °C in 50 % formamide, 5 x SSPE, 5 x Denharts solution, 1 % SDS and 100 ug of salmon sperm DNA/ml, the filters were hybridized at the same temperature and in the same solution containing the
15 denatured radiolabelled probe using 10⁶ cpm/ml of hybridization solution. The probe was labelled using a random priming kit (Amersham). After 16 hours the filters were washed in 2 x SSC containing 0.5 % SDS for 2 x 30 mins at 52 °C. The filters were exposed overnight using
20 intensifying screens at -70 °C. Positive clones were rescreened two times until all plaques on a plate were positive. The inserts from several positive clones were subcloned into the plasmid pBluescript SK+ by *in vivo* excision as recommended by the supplier.

25 Several clones were mapped by restriction enzyme analysis and were found to fall into two distinct groups characterized by the length of a SpeI/BamHI restriction fragment. The first of these groups comprised three of the restriction mapped clones which each had a 240 bp SpeI/BamHI
30 restriction fragment. The other group comprised a clone which had a 340 bp SpeI/BamHI fragment. Analysis of this fragment is described in Example 5.

 The three clones which exhibited the 240 bp SpeI/BamHI restriction fragment were fully or partially sequenced
35 (Sequenase 2.0, U.S. Biochemicals), and the characteristics of the clones are summarized as follows:

Nucleotide sequence analyses revealed that two of the cDNA clones were substantially identical, although they differed in length, and one had a mutation. One of the clones was full length and contained an open reading frame encoding 188 amino acid residues in which the first 21 amino acids are a cleavable signal sequence. The other of the two substantially identical clones terminated at the G of the start initiation codon. It could be inferred by sequence analysis of additional clones that the sequence preceding the G reads ACCAT. Both of the clones were found to have the same coding region nucleotide sequence, which is depicted in Figure 3 (SEQ ID NO:4). The figure omits the initial thymine and adenine of the start codon (TAG) which were not present in the isolated clones. The deduced amino acid sequence of the open reading frame of the coding region of both of these two cDNA clones is shown in Figure 4 (SEQ ID NO:5). The resulting protein encoded by this sequence is referred to hereinafter as VEGF-B₁₆₇. In each of the protein names used herein, the subscript number refers to the number of amino acids in the mature protein without the signal sequence.

As would be expected, a comparison of the amino acid sequence encoded by these two clones with the partial amino acid sequence deduced from cDNA clone of Example 1 showed a striking similarity. However, the two open reading frames in the clone of Example 1, each of which encoded an amino acid sequence homologous to a different portion of VEGF, are both present in the same reading frame in each of these two clones according to Example 2. The frame shift in the clone of Example 1 is caused by an insertion of two extra adenine units which displace the C-terminal part of the clone of Example 1 out of frame. The reason for this is not presently understood, but may be due to a cloning artifact.

The coding part of the third clone had a nucleotide sequence identical to those of the preceding two clones except for a 21 bp insertion. Figure 5 shows the nucleotide

sequence of this third clone (SEQ ID NO:6). To facilitate identification, the 21 extra bases are underlined in the Figure. This insertion gives rise to 7 additional amino acid residues in the mature protein. Thus the resulting protein encoded by this longer cDNA is termed VEGF-B₁₇₄. The amino acid sequence of the protein encoded by the cDNA of Figure 5 is depicted in Figure 6 (SEQ ID NO:7). The seven additional amino acids also are underlined in the figure for ease of identification. The additional amino acids are inserted into the sequence in a splice site, and sequencing of mouse genomic DNA clones indicates that these additional amino acids are the result of true alternative splicing. Furthermore, based on what is known about the receptor binding site locations of PDGF, the insertion occurs in a position in the protein which is probably part of a receptor binding site. The insertion is thus likely to affect receptor binding and could be of functional importance in influencing antagonist and/or different receptor specificity.

20

Example 3: Hybrid cDNA clone.

As previously pointed out this original cDNA clone of Example 1 was not full length and may contain an artifact. However, if the extreme 5' nucleotide sequence of the clones which encode VEGF-B₁₆₇ and/or VEGF-B₁₇₄ is added, the open reading frame encodes a protein of 133 amino acids, yielding a mature protein which is 112 amino acids long and hence is named VEGF-B₁₁₂. The hybrid cDNA sequence encoding VEGF-B₁₁₂ (SEQ ID NO:8) is shown in Figure 7, and the amino acid sequence of the corresponding protein (SEQ ID NO:9) is illustrated in Figure 8.

Figure 9 shows a multiple amino acid sequence alignment for comparison purposes of the 167 amino acid variant of mouse Vascular Endothelial Growth Factor B (mVEGF-B₁₆₇), mouse Vascular Endothelial Growth Factor (mVEGF₁₆₄), human Placenta Growth Factor (hPlGF), mouse Platelet Derived Growth Factor

35

A (mPDGF A), and mouse Platelet Derived Growth Factor B (mPDGF B). Amino acid residues identical to mouse VEGF-B₁₆₇ are boxed. The homologous relationship of the sequences is apparent, and the figure clearly demonstrates the conserved structure of the growth factors belonging to the PDGF/VEGF family of growth factors, and that VEGF-B is a structural homolog of the other growth factors of this group. Pairwise comparisons of the amino acid sequences show that mouse VEGF-B is approximately 43% identical to mouse VEGF₁₆₄, approximately 30% identical to human PlGF, and approximately 20% identical to mouse PDGF A and B. The conserved cysteine residues are particularly noteworthy. It can be seen that the first eight cysteine residues in the N-terminal domains (i.e. the PDGF-like domains) of the five growth factors are shared by all members of this family, and it is thus evident that the eight cysteine residues, which are involved in intramolecular and intermolecular disulfide bonding, are invariant among these growth factors. Furthermore, the C-terminal domains of mouse VEGF-B₁₆₇ and VEGF₁₆₄ also display a significant similarity with eight additional conserved cysteine residues and several stretches of basic amino acids.

Example 4: Cloning of human VEGF-B cDNA.

10⁶ λ-clones of human fibrosarcoma cDNA library HT1080 in λgt11 (Clontech) were screened with the ≈0.9 kb insert of the mouse VEGF-B clone pcif 2 according to standard procedures. Among several positive clones, one, termed H.1 was analyzed more carefully and its nucleotide sequence was determined. The nucleotide sequence indicated that a 207 amino acid isoform of human VEGF-B was encoded. Analysis of this isoform is described subsequently in Example 6. Based on the H.1 sequence two oligonucleotides were designed that would amplify the whole coding region of putative human cDNA corresponding to mouse VEGF-B₁₆₇ isoform:

5'-CACCATGAGCCCTCTGCTCC-3' (forward) (SEQ ID NO:18)

5'-GCCATGTGTCACCTTCGCAG-3' (reverse) (SEQ ID NO:19)

These oligonucleotides were used to amplify by polymerase chain reaction (PCR) the whole coding region of human VEGF-B from oligo-dT primed human erythroleukemia cell (HEL) RNA. The amplified product was cloned into the pCR II-vector of TA cloning kit (Invitrogen) and sequenced using standard techniques. The nucleotide sequence of the human VEGF-B cDNA clone is shown in Fig. 10 (SEQ ID NO:10), and the deduced amino acid sequence of human VEGF-B₁₆₇ is shown in Fig. 11 (SEQ ID NO:11).

The full length mouse cDNA clone of Example 2 and the full length human cDNA clone of Example 4 each encode a polypeptide of 188 amino acids containing an N-terminal hydrophobic putative signal sequence. In analogy with VEGF, the signal peptidase cleavage site is believed to be located between Ala 21 and Pro 22. The putative cleavage site of the signal peptidase is indicated in Figure 16 by an arrow. Accordingly, the processed VEGF-B polypeptides of these two clones each contain 167 amino acids.

Example 5:

The clone which exhibited the 340 bp SpeI/BamHI fragment isolated in Example 2 was analyzed, and the major portion was found to be identical to the first two clones of Example 2 which exhibited the 240 bp SpeI/BamHI fragment. The difference is due to the presence of an insertion in the C-terminal part of the sequence.

This 340 bp SpeI/BamHI DNA fragment encodes a further isoform of mouse VEGF-B containing 207 amino acids. The coding portion of the DNA encoding this protein (SEQ ID NO:12) is illustrated in Figure 12, and the translated amino acid sequence (SEQ ID NO:13) is illustrated in Figure 13. After cleavage of the 21 amino acid leader sequence, the mature protein contains 186 amino acids and is referred to as mVEGF-B₁₈₆. This isoform is clearly a result of

alternative DNA splicing as described below with reference to Figure 17.

Example 6:

5 The H.1 clone isolated as described in Example 4 was found to encode a 207 amino acid isoform of human VEGF-B. The coding portion of the DNA (SEQ ID NO:14) encoding this protein is illustrated in Figure 14 and the translated amino acid sequence (SEQ ID NO:15) is illustrated in Figure 15.
10 Again, this isoform, which is designated hVEGF-B₁₈₆, appears to be a product of alternative splicing.

 Both the mVEGF-B₁₈₆ of Example 5 and the hVEGF-B₁₈₆ of Example 6 include a 101 base pair insertion between nucleotides 414 and 415 of the coding sequence of VEGF-B₁₆₇.
15 Following the insertion, the nucleotide sequences of these cDNA clones were identical to the corresponding VEGF-B₁₆₇ sequences. The position of the 101 base pair insertion corresponds to the exon 5-exon 6 junction in VEGF. The insertion results in a frameshift which causes the C-
20 terminal domains of the two VEGF-B isoforms to be entirely different.

 The divergence of the C-terminal amino acid sequences starting at amino acid 116 in SEQ ID NOS 11 and 15, which correspond to the two principal VEGF-B isoforms, VEGF-B₁₆₇ and
25 VEGF-B₁₈₆, is reflected by the different biochemical characteristics of the two isoforms. The C-terminal domain of VEGF-B₁₆₇ is strongly basic (net charge +13) and binds heparin. The C-terminal domain of VEGF-B₁₈₆ is weakly basic (net charge +5) and has a long stretch of hydrophobic amino
30 acid residues in its C-terminus. The hydrophobic tail in VEGF-B₁₈₆ is unlikely to behave as a transmembrane domain since this variant of VEGF-B is secreted from cells. Therefore, despite an identical N-terminal domain, these two principal isoforms of VEGF-B have very different biochemical
35 properties. The absence of the highly basic heparin-binding domain from VEGF-B₁₈₆ allows the protein to be freely secreted

from cells. However, the secretion of VEGF-B₁₈₆ is remarkably slow; in a pulse chase experiment using transfected cells, VEGF-B₁₈₆ homodimers were not found in the medium before 1 hour. In contrast, VEGF homodimers and VEGF-B₁₈₆·VEGF dimers
5 appear in the medium within 30 minutes.

Figure 16 shows the aligned amino acid sequences of mouse and human VEGF-B₁₆₇ and VEGF-B₁₈₆ (SEQ ID NOS:5, 11, 13 & 15) in one-letter code. Identical residues are enclosed in boxes, while amino acid residues which differ between
10 mouse and human VEGF-B₁₆₇ and VEGF-B₁₈₆ isoforms are outside the boxes. Mouse and human VEGF-B display approximately 88 % amino acid sequence identity and are highly basic, especially in their C-terminal regions. The C-terminal domains of murine and human VEGF-B₁₈₆ are approximately 85%
15 identical at the amino acid level. The C-terminal domains of murine and human VEGF-B₁₆₇ are approximately 84% identical at the amino acid level. Both polypeptides lack the consensus sequence for N-linked glycosylation (N-X-T/S). The arrow indicates the putative cleavage site for the
20 signal peptidase between Ala²¹ and Pro²². Excluding the signal sequences, the mouse and human VEGF-B₁₆₇ amino acid sequences are highly homologous with only 20 replacements out the the 167 residues. The replacements are clustered in the N-terminus, in two regions around amino acids 60 and
25 145. All cysteine residues in both VEGF-B₁₆₇ proteins are invariant, but the eight cysteine residues in the C-terminal end of VEGF-B₁₆₇ are not conserved in the VEGF-B₁₈₆ isoforms. It is notable that the mouse and human sequences in the region between residues 66 and 129 are identical apart from
30 one evolutionarily conserved replacement (Q105R). This is of importance since the receptor binding domains are found within this portion of the protein (compared to PDGF structure). From this it can be concluded that it is likely that mouse and human VEGF-B will exhibit cross-reactive
35 binding on the receptor level and thus display identical or similar biological activities.

Example 7: Exon-intron structures of mouse and human VEGF-B genes.

The structure of the human VEGF-B gene was determined by restriction mapping and nucleotide sequence analysis of 5 cloned PCR fragments obtained from PCR reactions employing human genomic DNA as the template, except in the case of the first exon and intron, which were identified from a genomic λ -clone. The structure of the mouse gene was determined by 10 restriction mapping and nucleotide sequence analysis of cloned PCR fragments amplified using different combinations of primers. As a template in these PCR amplifications an isolated genomic λ clone containing the entire mouse VEGF-B gene was used.

Procedure.

15 Several λ clones for the mouse VEGF-B gene were isolated from a 129/Sw λ FIX genomic library as recommended by the supplier (Stratagene, Inc.). The ≈ 0.9 kb SalI/NotI insert of the pcif2 cDNA for VEGF-B (SEQ ID NO:1) was used as the probe. λ DNA from several positive clones were 20 isolated from plate lysates. One of the positive λ -clones (clone 10) was subcloned as BamH1 fragments into pBluescript SK (Stratagene Inc.). Isolated DNA from this same clone was also used as the template in PCR reactions (100 ng of λ DNA/reaction) and the coding parts of the mouse VEGF-B gene 25 were amplified using different combinations of primers. The nucleotide sequences of these primers were derived from the cDNA clones encoding murine VEGF-B₁₆₇ and murine VEGF-B₁₈₆. Taq DNA polymerase (2.5 U/reaction) was used. The generated PCR fragments were directly cloned into the TA-cloning 30 vector pCR II (Invitrogen Inc.). The exon-intron structure of the mouse VEGF-B gene was established by nucleotide sequence analysis of the subcloned Bam H1 genomic fragments and of the cloned PCR products.

A human genomic λ -clone was isolated by screening 1 x 35 10^6 clones of a human genomic library in EMBL-3 SP6/T7 (Clontech) using high stringency conditions with a 90 bp

PCR-fragment spanning 5' sequences of human VEGF-B cDNA as the probe. The washing conditions were: one wash at 1 x SSC at room temperature for 30 minutes and two washes at 1 x SSC at 65°C for 30 minutes. Primers for the PCR were:

5' -CACCATGAGCCCTCTGCTCC-3' (forward) (SEQ ID NO:18) and
5' -GGGCATCAGGCTGGGAGACAG-3' (reverse) (SEQ ID NO:19).

The positive λ -clone was subcloned as SacI fragments into pGEM 3Z vector (Promega) and was found to carry the 5'-region of the gene. The remaining parts of the human VEGF-B gene were amplified by PCR using genomic DNA as the template. Different combinations of primers derived from the human cDNA sequence were used. Dynazyme DNA polymerase (2.5 U/reaction, Finnzymes) was used. The amplified PCR fragments were directly cloned into the TA-cloning vector pCR II (Invitrogen Inc.). The exon-intron boundaries and the length of the short introns of the mouse and human VEGF-B genes were determined by nucleotide sequence analysis using vector specific primers or suitable primers derived from the cDNA sequences. The length of the larger introns were calculated based on the length of the amplified PCR fragments when analyzed by agarose gel electrophoresis.

Results.

The results showed that the coding parts of the mouse and human VEGF-B genes span approximately 4 kb of DNA and both genes are divided into seven coding exons ranging from 19 bp (E7) to 236 bp in length (E6). Figure 17 is a schematic representation of the structures of the mouse and human genes for VEGF-B. The exon sizes in base pairs are noted inside the boxes, and the sizes of the introns are noted between the boxes. The introns are not shown to scale. The structures of the untranslated flanking regions of mouse and human VEGF-B genes were not established and are represented by gray boxes. The exon-intron junctions in both genes are listed in the following Table 3:

Table 3

Exon Length	Donor Site	Intron Length (bp)	Acceptor Site
Mouse			
E1 60	T CGC ACC CAG/gtacgtgcgt a Arg Thr Gln	≈590	tttcccacag/GCC CCT GTG T Ala Pro Val S
E2 43	CAG AAG AAA G/gtaataatag Gln Lys Lys V	287	ctgcccacag/TG GTG CCA TG al Val Pro Tr
E3 197	C CGA ATG CAG/gtaccagggc l Arg Met Gln	161	ctgagcacag/ATC CTC ATG A Ile Leu Met I
E4 74	GT GAA TGC AG/gtgccagcca ys Glu Cys Ar	178	ctcctcctag/G GTT GCC ATA g Val Ala Ile
mVEGF-B ₁₈₆			
E5 36	AG CCA GAC AG/gtgagttttt ys Pro Asp Ar	≈200	ctcctcctag/G GTT GCC ATA g Val Ala Ile
E6A 211	Stop codon in exon 6 (TAG)	---	-----
mVEGF-B ₁₆₇			
E5 36	AG CCA GAC AG/gtgagttttt ys Pro Asp Se	≈300	cccactccag/C CCC AGG ATA r Pro Arg Ile
E6B 135	AC ACC TGT AG/gtaaggagtc sp Thr Cys Ar	≈2.6 kb	cactccccag/G TGC CGG AAG g Cys Arg Lys
E7 19	Stop codon in exon 7 (TGA)	---	-----
Human			
E1 60	C CCC GCC CAG/gtacgtgcgg a Pro Ala Gln	≈760	tctcccacag/GCC CCT GTC T Ala Pro Val S
E2 43	CAG AGG AAA G/gtaataactta Gln Arg Lys V	275	ctgctcccag/TG GTG TCA TG al Val Ser Tr
E3 197	C CGG ATG CAG/gtactgggca l Arg Met Gln	244	ctgagcacag/ATC CTC ATG A Ile Leu Met I
E4 74	GT GAA TGC AG/gtgccagcca ys Glu Cys Ar	≈710	tacttttcag/A CCT AAA AAA g Pro Lys Lys
hVEGF-B ₁₈₆			
E5 35	AG CCA GAC AG/gtgagtcttt ys Pro glu Ar	200	tctcctcctag/G GCT GCC ACT g Ala Ala Thr
E6A 211	Stop codon in exon 6 (TAG)	---	-----
hVEGF-B ₁₆₇			
E5 36	AG CCA GAC AG/gtgagtcttt ys Pro Glu Se	≈300	cccactccag/C CCC AGG CCC r Pro Arg Pro
E6B 135	AC ACC TGC AG/gtaggttttg sp Thr Cys Ar	736	ccctcctcag/G TGC CGG AAG g Cys Arg Lys
E7 19	Stop codon in exon 7 (TGA)	---	-----

As previously stated, exon 6 contains an alternative splice acceptor site which enables the gene to produce two different transcripts for VEGF-B isoforms. VEGF-B₁₆₇ uses exons 1-5, the last part of exon 6, and exon 7 (TGA). VEGF-B₁₈₆ uses exons 1 through 5, the first part of exon 6, and terminates in the last part of exon 6 (TAG). Exon 7 is not translated in VEGF-B₁₈₆ since the insertion of the first part of exon 6 introduces a frame shift and gives rise to a stop codon in the last part of exon 6. The position of the stop codon (TAG) for VEGF-B₁₈₆ is marked in exon 6B, and the stop codon (TGA) for VEGF-B₁₆₇ is marked in exon 7.

The introns in both genes vary from 161 bp to approximately 2.6 kb. The length of each exon and the locations of the splice junctions in the two genes were identical, and all splice donor and acceptor sites follow the canonical GT/AG rules, Padgett et al., *Annual Rev. of Biochemistry*, 55:1119-50 (1986). The only notable difference between the mouse and the human genes are the length of introns 1, 4 and 6 which are longer in the mouse gene. All exon-intron boundaries were found to be conserved between VEGF-B and VEGF, but the introns in the VEGF-B genes were generally smaller than in the VEGF gene.

The 300 bp-intron after the exon 5 in VEGF-B differs from the corresponding one in VEGF, which is 3 kb in length and contains an alternatively spliced exon found in the transcripts for VEGF₁₈₉ and VEGF₂₀₆, encoding many basic amino acid residues. When this intron in VEGF-B was analyzed more carefully, no exon corresponding to the 6th exon of VEGF could be found. Instead, the 3' end of this intron and the following exon were found to be identical with the corresponding sequences of the cDNA clones encoding VEGF-B₁₈₆. This is explainable by the fact that the mRNA for VEGF-B₁₈₆ is formed by use of an alternative splice acceptor site during mRNA splicing, resulting in an insertion of a 101 bp intron sequence into these mRNAs.

Figure 18 shows a comparative hydrophilicity analysis of murine VEGF-B₁₆₇ and VEGF-B₁₈₆. The profiles were generated according to Kyle and Dolittle using a window of nine (9) residues. As would be expected, the pattern of hydrophilicity/hydrophobicity is essentially identical from amino acid 1 through amino acid 115. After amino acid 115, the hydrophilicity/hydrophobicity patterns diverge because of the frame shift introduced by the first part of exon 6. Thus, VEGF-B₁₆₇ and VEGF-B₁₈₆ can be expected to exhibit both similar and dissimilar activities.

Figure 19 is a dendrogram showing the phylogenetic relationship of the amino acid sequences of five members of the VEGF/PDGF family of growth factors. The number of replacements or substitutions decreases from the left to the right of the chart. It can be seen that VEGF-B lies between VEGF and the platelet derived growth factor (PDGF) group.

The multiple amino acid sequence alignments of Figures 9 and 16 and the phylogenetic analysis of Figure 19 were carried out according to Hein, *Methods in Enzymology*, Vol. 183, pp. 626-45, Academic Press Inc., San Diego (1990) using the PAM 250 distance table.

Example 8: Antibody Production.

a. Antiserum to Mouse VEGF-B.

Antisera to mouse VEGF-B were raised by immunizing rabbits with a 18-mer oligopeptide comprising the N-terminal region of processed VEGF-B, coupled to keyhole limpet hemocyanin. Cysteine residues were introduced as the N-terminal and C-terminal amino acid residues to allow coupling of the peptide to the carrier protein using SPDP (Pharmacia). The sequence of the oligopeptide was

C-P-V-S-Q-F-D-G-P-S-H-Q-K-K-V-V-P-C (SEQ ID NO:21).

Each rabbit received a subcutaneous injection with 300 µg of the peptide conjugate emulsified in Complete Freund's Adjuvant. Subcutaneous booster injections were given every second week with the same amount of antigen emulsified in

Incomplete Freund's Adjuvant. Sera were obtained after the second booster injections.

b. Antiserum to Human VEGF-B.

Antipeptide antiserum to human VEGF-B was generated by immunizing rabbits with a branched 23-mer oligopeptide comprising the following N-terminal region amino acid residue sequence (SEQ ID NO:22):

S-Q-P-D-A-P-G-H-Q-R-K-V-V-S-W-I-D-V-Y-T-R-A-T.

The branched 23-mer oligo peptide was synthesized according to Tam, "Synthetic peptide vaccine design: synthesis and properties of a high-density multiple antigenic peptide system", *Proc. Natl. Acad. Sci. USA*, Vol. 85, pages 5409-413 (1988). In the first immunization, rabbits were subcutaneously injected with 500 μ g of the branched peptide emulsified in Complete Freund's Adjuvant. In the subsequent boosters, 200 μ g of the antigen emulsified in Incomplete Freund's Adjuvant was injected. Antisera were collected after the second and third boosters by conventional techniques.

20

Example 9: Biochemical properties of VEGF-B₁₆₇, homo-dimerization, and heterodimerization with VEGF.

The biochemical properties of human VEGF-B₁₆₇ were examined in transfected human embryonic kidney 293EBNA cells (Invitrogen, Inc.). cDNA inserts encoding human VEGF-B₁₆₇ and human VEGF₁₆₅ [see Keck et al., *Science*, Vol. 246, pages 1309-312 (1989)] were individually cloned into the pREP7 expression vector (Invitrogen, Inc.). Human embryo kidney 293EBNA cells (expressing Epstein-Barr virus nuclear antigen-1) were transfected by transient transfection with the respective expression plasmids using calcium phosphate precipitation, and the cells were incubated for 48 hours. As a control, cells also were transfected with an expression vector containing the VEGF-B₁₆₇ cDNA in reverse orientation. Monolayers of cells were incubated in methionine-free and cysteine-free medium for 30 minutes followed by labeling

35

with 100 $\mu\text{Ci/ml}$ [^{35}S]methionine and [^{35}S]cysteine (Promix, Amersham Inc.) in the same medium for 2 hours. The labeling medium was replaced with normal medium without serum, and labelled proteins were chased for 6 hours. Heparin was included during the chase when indicated (100 $\mu\text{g/ml}$). Media were collected after the chase period, and cells were solubilized in 10 mM Tris pH 7.5, 50 mM NaCl, 0.5% sodium deoxycholate, 0.5% Nonidet P-40, 0.1% SDS and 0.1 U/ml aprotinin.

10 VEGF-B₁₆₇ was expressed in the cells transfected with the plasmids containing the VEGF-B₁₆₇ DNA. Aliquots of the culture supernatants from cells treated or untreated with heparin and detergent solubilized cell lysates were subjected to immunoprecipitation with the specific antipeptide antiserum to VEGF-B obtained as described in Example 8 and analyzed by SDS-PAGE under reducing conditions unless otherwise indicated. The data show that VEGF-B₁₆₇ homodimers and VEGF-B₁₆₇ - VEGF₁₆₅ heterodimers are released from cells by heparin. By heparin treatment (1-100 $\mu\text{g/ml}$) or 1.2 M NaCl, VEGF-B₁₆₇ was released from cells and found in the supernatant. If cells were not treated with heparin, VEGF-B₁₆₇ remained cell-associated and was not released into the culture medium. Under the same conditions, VEGF₁₆₅ homodimers are secreted from the cells and found in the culture supernatants without heparin treatment.

Under reducing conditions, human VEGF-B₁₆₇ migrated with a Mr of 21 kDa. Analysis of culture supernatants under non-reducing conditions showed that VEGF-B₁₆₇ migrated as an Mr 42 kDa species indicating a dimeric structure. These results suggest that VEGF-B₁₆₇ forms disulfide-linked dimers associated with the cell surface, probably through ionic interactions with extracellular heparan sulfate proteoglycans. The association is likely to be mediated by the C-terminal basic domain, as observed for the longer splice variants of VEGF.

Since VEGF has been shown to form heterodimers with PlGF, it was decided to test whether VEGF₁₆₅ could also form heterodimers with VEGF-B₁₆₇. For this purpose 293EBNA cells were co-transfected with expression vectors encoding both
5 human VEGF₁₆₅ and human VEGF-B₁₆₇, and VEGF-B₁₆₇ was expressed in combination with VEGF₁₆₅. Metabolically labelled proteins were chased in the presence of heparin, and immunoprecipitations were carried out with antisera to either VEGF-B₁₆₇ or VEGF₁₆₅. The antiserum to human VEGF was
10 from R&D Systems. Under non-reducing conditions the VEGF-B₁₆₇·VEGF₁₆₅ heterodimers migrated as Mr 42-46 kDa species. The results show that VEGF-B can form disulfide linked heterodimers with VEGF, which, in the absence of heparin, remain cell-associated. Since homodimers of VEGF₁₆₅
15 are efficiently secreted into the media, VEGF-B appears to determine the secretion of the heterodimer.

VEGF-B is synthesized normally in the endoplasmic reticulum of the source cell for subsequent export. Recombinant VEGF-B may be produced by inserting a DNA
20 sequence encoding the VEGF-B protein together with suitable operatively linked promoter and control sequences into a suitable vector, such as the well known plasmid pBR322 or a derivative thereof, transforming or transfecting a suitable host cell, such as *E. coli* or a Cos cell, with the resulting
25 vector or other systems well known in the art, screening the resulting transformants or transfectants for VEGF-B expression, and then culturing cell lines or bacterial cell strains which are positive for the expression of VEGF-B. Either a eukaryotic vector or a prokaryotic vector may be
30 used, depending on the type of cell which is to be transfected or transformed therewith. A particularly preferred system for production of recombinant VEGF-B is the baculovirus - insect cell system, which has proved capable of producing excellent yields of recombinant protein.

Example 10: VEGF-B expression using the baculovirus system.

10.1 VEGF-B with its own signal peptide.

a) Cloning and Transfection.

The complete human VEGF-B₁₆₇ gene was inserted into a
5 commercially available plasmid pCRII (Invitrogen Corp.).
The HindIII-XbaI fragment from the resulting plasmid pCRII-
VEGF-B₁₆₇, which encodes the whole open reading frame of
VEGF-B₁₆₇ then was cloned into pFASTBAC1, and both the 3' - and
5'-junctions were sequenced. Bacmid-DNA was prepared
10 according to the manufacturers instructions for the "Bac-To-
Bac™ Baculovirus Expression System" (Life Technologies Inc.)
and lipofected to Sf900II-adapted Sf9 cells (obtained from
Dr. Christian Oker-Blom). Sf9 cells are from the American
Type Culture Collection Cell Repository Line Bank, Rockville
15 MD (ATCC CRL-1711). The transfected cells were then
cultured on standard TMN-FH medium in 25 cm² culture dishes.

b) Assay for protein expression.

About 72 hours after transfection, the cells were lysed
and 1 ml of culture supernatant and the cell lysate were
20 assayed for expressed VEGF-B by immunoprecipitation as
described in Example 9 and Western blotting. Lysates from
three out of four independently transfected cell cultures
were found positive for VEGF-B, although the intensity of
the signal in the Western blot varied. The expressed VEGF-B
25 polypeptide in each case was found to correspond in size to
the protein expressed in mammalian cells in Example 9.

The viral stock from the cells that gave the strongest
signal in Western blotting was amplified two rounds by
infecting cells and collecting new virus from the medium.
30 The resulting supernatant was analyzed. Uninfected cells
were also analyzed as a negative control. Time course
analysis showed that cells harvested between 48 and 72 hours
after infection contained the greatest amount of VEGF-B.
After 96 hours post infection, as a result of virus-induced
35 cell lysis, VEGF-B could also be detected in the culture
supernatant by immunoprecipitation and Western blotting.

Recombinant VEGF-B could be precipitated from the lysate between 20% and 40% $(\text{NH}_4)_2\text{SO}_4$.

10.2 VEGF-B with the Melittin signal peptide (pVTBac).

a) Cloning and transfection.

5 A polymerase chain reaction (PCR) fragment from nucleotide position 68 to 141 was used to introduce a BamHI restriction site immediately after the signal cleavage site in the plasmid pCRII-VEGF-B₁₆₇ from Example 10.1. The BamHI fragment from this modified pCRII-VEGF-B₁₆₇ construct was
10 cloned into BamHI opened pVTBac [Tessier et al., "Enhanced secretion from insect cells of a foreign protein fused to the honeybee mellitin signal peptide", *Gene*, Vol. 98, page 177 (1991)]. Both 3'- and 5'-junctions were sequenced. Sf9 cells were cotransfected with the aforescribed pVTBac
15 vector which contained the human VEGF-B₁₆₇ gene, and with linearized baculovirus DNA (Insectin™, Invitrogen Corp.). The transfected cells then were cultured in TMN-FH medium.

b) Assay for protein expression.

20 Forty-eight hours after transfection, the supernatant was collected and subjected to primary screening by immunoprecipitation. Four positive plaques were isolated.

10.3 A cDNA insert encoding murine VEGF-B₁₈₆ (EcoRI cut cDNA fragment from a murine VEGF-B₁₈₆ cDNA (SEQ ID NO:12) clone) was cloned into pFASTBAC 1. An EcoRI cut cDNA
25 fragment from murine VEGF-B₁₆₇ (SEQ ID NO:4) was also cloned into pFASTBAC 1. The resulting plasmids were transformed into bacteria as described in 10.1 above, and recombinant plasmids were isolated and lipofected into Sf9 and Sf21 cells. Supernatants containing recombinant baculovirus were
30 amplified by several rounds of reinfection of Sf21 cells. The final titers of the baculovirus stocks were determined by plaque titration and found to vary between 4×10^8 and 2×10^9 baculovirus particles per milliliter of stock supernatant.

Example 11: Large Scale Production of Recombinant VEGF-B.

Sf21 cells [see Vaughn et al., *In Vitro*, 13:213-17 (1977)] were infected with the baculovirus stocks of Example 10 at a multiplicity of infection of 10 virus particles per cell. The infected Sf21 cells were grown in roller flasks and seeded at a density of 2×10^6 cells per ml of serum free medium (Sf900II, Gibco-BRL) for 96 hours. Culture media and cells were then harvested. Aliquots of the cell lysates and of the media were analyzed by SDS-PAGE. Total protein patterns were visualized by staining the gels with Coomassie Brilliant Blue and the presence of expressed VEGF-B isoforms were visualized by immunoblotting using specific antipeptide antibodies to human and mouse VEGF-B as described above in Example 8. The analysis revealed that both human and mouse VEGF-B₁₆₇ polypeptides were of the expected sizes of 21.5 kDa. Both proteins were retained intracellularly in the infected cells and not released into the medium. In contrast, mouse VEGF-B₁₈₆ was readily secreted into the medium in a dimeric form. The VEGF-B₁₈₆ homodimers migrated as a 52-54 kDa species which suggested that insect cell produced protein did not undergo the same covalent modification as found for VEGF-B₁₈₆ secreted from transfected Cos-1 cells.

Example 12: Transfection and analysis of Cos-1 cells expressing VEGF-B₁₈₆.

cDNA inserts encoding mouse VEGF-B₁₈₆ and human VEGF₁₆₅ were cloned into the pSG5 expression vector [Green et al., *Nucleic Acid Res.*, 16:369 (1988)]. Cos-1 cells were maintained in minimal essential medium (MEM) containing 10% fetal calf serum, 2mM glutamine and appropriate antibiotics. For transfections, the cells were replated into 90 mm Petri dishes. The cells were transfected with the expression vectors, separately or in combination, using calcium phosphate precipitation and incubated for 36-48 hours. Monolayers of cells were incubated in medium free of methionine and cysteine for 30 min and then incubated in the

same medium containing 100 $\mu\text{Ci/ml}$ of [^{35}S]-methionine and [^{35}S]-cysteine for 2 hours (Promix Amersham Inc.).

For the pulse-chase experiments, the cells were labeled for 30 minutes, washed twice with normal medium and then
5 incubated for up to 6 hours in MEM without fetal calf serum. Media were collected after the chase period and the cells were solubilized in 10 mM Tris buffer pH 7.5 containing 50 mM NaCl, 0.5% deoxycholate, 0.5% nonidet P-40 and 0.1% SDS. Aliquots of the media and the cell lysates were subjected to
10 immunoprecipitation using the specific antiserum to mouse VEGF-B from Example 8a and a specific antiserum to human VEGF commercially available from R&D Systems. The precipitates were analyzed by SDS-PAGE.

15 Example 13: Biochemical properties of VEGF-B₁₈₆ expressed in transfected Cos-1 cells.

The biochemical properties of mouse VEGF-B₁₈₆ were examined in Cos-1 cells transiently transfected as described in Example 12 with an appropriate expression vector. The
20 cells were metabolically labelled, and proteins from the labelled cells were immunoprecipitated using an antipeptide antibody to VEGF-B. The precipitated material was subjected to SDS-PAGE analysis under reducing conditions. Both the cell culture medium (M) and a detergent solubilized cell
25 lysate (L) were analyzed. The results are shown in Figure 22. It can be seen that cell associated VEGF-B₁₈₆ migrated as an approximately M_r 24,000 polypeptide under reducing conditions. In contrast, VEGF-B₁₈₆ present in the medium of transfected cells migrated as a M_r 32,000 species, suggesting
30 that the protein was covalently modified during its intracellular transport and secretion. The corresponding molecules were not detected in cell lysates or media from mock transfected Cos-1 cells used as a control.

Immunoprecipitation of media and SDS-PAGE analysis
35 under non-reducing conditions, showed an approximately M_r 60,000 species suggesting that VEGF-B₁₈₆ formed disulfide-

linked homodimers. Including 100 ug/ml of heparin during the labelling did not affect the secretion or release of VEGF-B₁₈₆ homodimers from the transfected cells.

5 Example 14: Biosynthesis of VEGF-B₁₈₆ homodimers.

The biosynthesis of VEGF-B₁₈₆ homodimers was examined by pulse-chase experiments. Transfected Cos-1 cells were metabolically labelled for 30 minutes and then chased for up to 4 hours. Immunoprecipitation and SDS-PAGE analysis of
10 detergent solubilized cell lysates and media showed that the cell-associated approximately M_r 24,000 species was readily detected in the lysates throughout the chase period. The decrease in the intensity of this molecular species was associated with an increase in the M_r 32,000 protein present
15 in the media. The M_r 32,000 species appeared in the medium after 1 hour of chase. Highest levels of secreted VEGF-B₁₈₆ were obtained after the 4 hour chase period. No intermediates were detected in the cell lysates, but the secreted M_r 32,000 protein appeared slightly heterogenous.
20 The nature of the modification is presently unknown, but N-linked glycosylation can be excluded in the absence of consensus sites for this modification.

Example 15: Formation of heterodimers by VEGF-B₁₈₆.

25 As noted above, VEGF-B and VEGF are coexpressed in many tissues and VEGF-B₁₆₇·VEGF₁₆₅ heterodimers are readily formed when coexpressed in transfected cells. To examine whether VEGF-B₁₈₆ also could form heterodimers with VEGF₁₆₅, Cos-1 cells were transfected as described above with the
30 appropriate expression vectors, either alone or in combination. Metabolically labelled proteins present in the media from the transfected cells were subjected to immunoprecipitations using antisera to VEGF-B and VEGF. Figure 23A shows the results of SDS-PAGE analysis under
35 reducing conditions of the immunoprecipitates from the cell culture media of transiently transfected Cos-1 cells

separately expressing VEGF-B₁₈₆ and VEGF, respectively. It can be seen that the antisera were specific for VEGF-B and VEGF, respectively, with no detectable cross-reactivity.

5 Cos-1 cells were cotransfected with expression vectors for VEGF-B₁₈₆ and VEGF₁₆₅. Cell culture media (M) and detergent solubilized lysates (L) from the resulting cells which coexpressed VEGF-B₁₈₆ and VEGF₁₆₅ were subjected to immunoprecipitation and SDS-PAGE analysis under reducing conditions. The results are shown in Figure 23B. The test
10 showed that murine VEGF-B₁₈₆ and human VEGF₁₆₅ form intracellular and secreted heterodimers.

Culture media from cells expressing murine VEGF-B₁₈₆ and human VEGF₁₆₅, either separately or in combination, were subjected to immunoprecipitation using antibodies to VEGF-B
15 and VEGF and analyzed by SDS-PAGE under non-reducing conditions. As a control, cell culture medium from mock transfected cells was analyzed. The results are shown in Figure 23C. It was found that VEGF-B₁₈₆ forms secreted disulfide-linked homodimers and that VEGF-B₁₈₆ and VEGF₁₆₅
20 together form secreted disulfide-linked heterodimers.

To analyze whether heterodimer formation with VEGF affected the secretion and release of VEGF-B₁₈₆, pulse-chase experiments were carried out using Cos-1 cells transiently transfected with expression vectors for VEGF-B₁₈₆ and VEGF₁₆₅.
25 Cell associated disulfide-linked heterodimers could be recovered following the 30 minute labelling period, and secreted heterodimers were recovered from the medium already after a 30 minute chase period. The secreted heterodimers accumulated in the medium for up to 2 hours post labelling.
30 In the 4 hour chase time point there was a decrease in the amount of heterodimers in the medium, possibly due to the degradation of the complex. Some VEGF-B₁₈₆·VEGF heterodimers remained cell-associated throughout the chase. These results suggested that heterodimer formation with VEGF
35 promoted the secretion of VEGF-B₁₈₆ compared to the secretion of VEGF-B₁₈₆ homodimers. Furthermore, the presence of

heterodimers already following the 30 minute labelling period suggested that the slow release of VEGF-B₁₈₆ homodimers was not due an impaired ability of VEGF-B₁₈₆ to dimerize.

5 Example 16: Purification of Secreted VEGF-B₁₈₆ Homodimers.

Secreted VEGF-B₁₈₆ homodimers were isolated from serum free culture media of baculovirus infected Sf21 cells as follows:

a. Initial Separation.

10 The major contaminating protein in the culture media was the baculovirus protein gp64/67, an acidic protein secreted by baculovirus infected cells. To remove this protein, the culture media was concentrated twenty-fold by ultrafiltration and then passed over a Sephadex G-25 column
15 equilibrated in 20 mM phosphate buffer pH 6.5 containing 20 mM NaCl. Eluted proteins were then passed over a CM-Sepharose (Pharmacia) ion-exchange column equilibrated in the same buffer. The column was washed with the phosphate buffer to remove unbound proteins, and bound proteins were
20 eluted by stepwise increasing the NaCl concentration of the elution buffer. The major gp64/67 baculovirus encoded protein did not bind to the ion-exchange column under those conditions while VEGF-B₁₈₆ homodimers eluted at a NaCl concentration of 90 mM. As judged by SDS-PAGE analysis of
25 the eluted fraction, VEGF-B₁₈₆ homodimers were 5-15% pure by this procedure.

b. Purification to Homogeneity.

The VEGF-B₁₈₆ homodimers are purified to homogeneity on a MonoS column coupled to a FLPC system (Pharmacia). Bound
30 protein is eluted with a linear gradient of NaCl in 20 mM phosphate buffer pH 6.5.

Example 17:

In order to find out whether the two VEGF-B splice
35 isoforms exhibited a differential tissue distribution and whether additional isoforms existed, an RT-PCR analysis was

carried out using total RNA extracted from mouse brain, heart, liver and kidney and from human embryonic heart and skeletal muscle. The transcripts were analyzed by PCR using four pairs of specific primers covering exons 4 to 7 and
5 exons 3 to 7 in the mouse and human VEGF-B genes, respectively.

Procedure.

Total RNA from mouse and human tissues were isolated using standard procedures as disclosed by Chirgwin et al.,
10 *Biochemistry*, 18:5294-99 (1979). Two to five μ g of total RNA per reaction were used for first strand cDNA synthesis using avian myelostosis virus reverse transcriptase (20 U/reaction). The reactions were primed with oligo-(dT)₁₈. Aliquots of these reactions were used as templates in PCR
15 reactions using *Taq* DNA polymerase (2.5 U/reaction). To amplify mouse cDNA, two pairs of primers were used. These pairs were obtained by combining a common 5'-primer
5'-CACAGCCAATGTGAATGCA (forward) (SEQ ID NO:23),
located in exon 4 with two different 3'-primers
20 5'-GCTCTAAGCCCCGCCCTTGGCAATGGAGGAA (reverse) (SEQ ID NO:24)
and 5'-ACGTAGATCTTCACTTTCGCGGCTTCCG (reverse) (SEQ ID NO:25)
(this last primer has a *Bgl* II site and 4 extra bases in the 5' end) located in exons 6B and 7, respectively. Following
analysis by agarose gel electrophoresis, the amplified bands
25 were transferred onto a nylon filter (Genescreen Plus) and sequentially hybridized with oligonucleotide probes specific for exons 6A and 6B. The oligonucleotide probes were 5'-CTCTGTTCCGGGCTGGGACTCTA (exon 6A) (SEQ ID NO:26) and 5'-TCAGGGCGTTGACGGCGCTGGGTGCAA (exon 6B) (SEQ ID NO:27).
30 The oligonucleotide probes were labeled with [³²P]dCTP using terminal transferase to high specific activity. Hybridizations, using 1 x 10⁶ cpm of labeled probe/ml of solution, were carried out at 37° C in 6 x SSC containing 5 X Denhardt's solution, 0.5% SDS and 100 μ g/ml of salmon sperm DNA. The filters were washed at the same temperature
35

in 6 x SSC containing 0.5% SDS for 2 x 15 min and exposed to film.

The two pairs of primers used for amplification of human cDNA were combined using two different 5'-primers,

5 5'-CCTGACGATGGCCTGGAGTGT (forward) (SEQ ID NO:28),
located in exon 3 and

5'-TGTCCCTGGAAGAACACAGCC (forward) (SEQ ID NO:29),
located in exon 4, with a common 3'-primer,

10 5'-GCCATGTGTCACCTTCGCAG (reverse) (SEQ ID NO:19)
located in exon 7. Aliquots of the amplified products were
analyzed by agarose gel electrophoresis. The aliquots were
directly cloned in the TA-cloning vector pCR II (Invitrogen,
Inc.), and generated plasmids were analyzed by nucleotide
sequencing. Amplification of GAPDH served as a control.

15 Results.

Analysis of amplified PCR products by agarose gel
electrophoresis showed two major bands of 215 and 316 bp,
respectively. These sizes are consistent with the two mRNAs
corresponding to VEGF-B₁₆₇ and VEGF-B₁₈₆. These two bands were
20 of the same intensity suggesting that the two isoforms were
expressed at approximately equal levels in all mouse and
human tissues examined.

To verify the identity of the amplified products from
mouse tissues, the PCR-amplified DNA was transferred to a
25 filter and probed with specific oligonucleotide probes for
exons 6A and 6B, respectively. The autoradiograms showed
that an exon 6-specific probe hybridized with the 316 bp
band while the exon 6B specific probe hybridized with both
the 215 bp and the 316 bp amplified bands. These results
30 are consistent with the alternative usage of acceptor site
in exon 6 to create the two isoforms of VEGF-B and thus all
the amplified products corresponded to those predicted from
the sequences of VEGF-B₁₆₇ and VEGF-B₁₈₆ isoforms.

Agarose gel electrophoresis of products of PCR analysis
35 of total RNA isolated from human embryonic heart and muscle
visualized two major amplified bands of 329 bp and 430 bp.

Taken together, these data demonstrate that VEGF-B₁₆₇ and VEGF-B₁₈₆ are the two major isoforms of VEGF-B in tissues. The pattern of the PCR products and the location of the primers indicate that if any still longer splice isoforms exist for VEGF-B, such transcripts use a splice acceptor site located a little more 5' than in the case of VEGF-B₁₈₆. Furthermore, PCR products corresponding to VEGF₁₂₁, which lacks heparin binding domains, i.e. sequences corresponding to exon 6 in VEGF-B, were not detected. However, splicing of exon 5 to exon 7 would give rise to a transcript encoding an isoform of VEGF-B corresponding to VEGF₁₂₁, and this putative isoform of VEGF-B might be expressed in tissues other than those analyzed in this example.

15 Example 18: Stimulation of Cell Proliferation.

The ability of VEGF-B₁₆₇ to stimulate endothelial cell proliferation was established through analysis of [³H]thymidine incorporation in human umbilical vein endothelial cells (HUVEC) and in bovine capillary endothelial (BCE) cells.

293EBNA cells were transfected as described above with expression vectors for VEGF-B₁₆₇, VEGF₁₆₅ or empty vector (mock) in the presence of 1 μg/ml heparin. Conditioned media from these cells were diluted in respective media, applied to human umbilical vein endothelial cells (HUVEC) and to bovine capillary endothelial (BCE) cells and incorporation of [³H]thymidine was measured. As a positive control recombinant bFGF was added to BCE cells.

To elaborate, conditioned media containing human VEGF-B and human VEGF₁₆₅ were collected from 293EBNA cells transfected with the appropriate expression vectors or with empty vector (mock) in the presence of heparin (1 μg/ml) 48 hours posttransfection. Second passage HUVEC were plated into 96-well plates (4 x 10³ cells/well) in M-199 medium supplemented with 10% fetal bovine serum and incubated for 24 hours. Conditioned media were diluted with the growth

medium, and cells were stimulated for 48 hours. Fresh conditioned media containing 10 μ Ci/ml of [3 H]thymidine (Amersham Inc.) were added to the cells, and stimulations were continued for another 48 hours. Cells were washed with
5 PBS and trypsinized, and incorporated radioactivity was determined by liquid scintillation counting. BCE cells were seeded into 24-well plates and grown until confluence in minimal essential medium (MEM) supplemented with 10% fetal calf serum. Cells were starved in MEM supplemented with 3%
10 fetal calf serum for 72 hours, after which conditioned media diluted into serum-free medium were added to the cells and the cells were stimulated for 24 hours. [3 H]Thymidine was included during the last 4 hours of the stimulation (1 μ Ci/ml). Stimulations with bFGF were carried out as above
15 using 6 ng/ml of recombinant bFGF (Synergen Inc.). Cells were washed with PBS, lysed with NaOH, and incorporated radioactivity was determined by liquid scintillation counting.

Figure 20 is a bar graph showing fold of induction of
20 [3 H]thymidine incorporation by VEGF-B₁₆₇ in human umbilical vein endothelial cells (HUVEC) and in bovine capillary endothelial (BCE) cells, as compared to basal activity induced by conditioned medium from the mock transfected cells. For comparison purposes, the induction by VEGF₁₆₅ and
25 by bFGF are also shown. The bars show the mean \pm standard deviation of parallel samples. Similar results were obtained in several other independent experiments. The test results clearly show that VEGF-B induced [3 H]thymidine incorporation in both HUVEC and BCE cells and stimulated
30 proliferation of endothelial cells in vitro, thereby demonstrating that VEGF-B is an endothelial growth factor.

Example 19: Identification of human VEGF-B promoter DNA clones and activity.

35 A human genomic DNA library in bacteriophage λ EMBL was screened using a 5' PCR fragment containing the sequences

from the VEGF-B first and second exons as a probe. Two positive clones were obtained, and one of these was subcloned in the Bluescript SKII plasmid as Sac I fragments. A 1.4 kb fragment was obtained, which contained about 0.4 kb of sequences upstream from an Nco I site present in the cDNA (located less than 100 bp upstream of the ATG translational initiation site).

In addition, an XhoI fragment of about 6 kb from the other λ clone was subcloned into the pGEMEX plasmid. This subclone contained about 1.5 kb of sequences upstream from the NcoI site. The SacI/NcoI fragment and an EcoRI (polylinker) - NcoI fragment were subcloned into pGL3 basic vector (Promega) in the respective transcriptional orientation. DNA of these subclones, and from the pGL3 control vector containing the SV40 promoter, was transfected into HeLa cells using calcium phosphate precipitation. Two days after transfection, the luciferase activities were measured from lysates of the transfected cells. The results indicated that the 400 bp SacI/NcoI fragment has promoter activity equal to about 30% of the activity of the pGL3 control vector, while the 1.5 kb fragment gave only background activity. Use of a stronger or more active promoter, for example the CMV promoter or the elongation factor 1-alpha promoter, would probably give higher activity in human cells and tissues. The structure of the cloned fragments is illustrated in Figure 24.

The 1.5 kb fragment upstream of the Nco I site was sequenced. The resulting sequence (SEQ ID NO:17) is illustrated in Figure 25. The sequence obtained revealed a putative silencer element [Weissman and Singer, *Molecular and Cellular Biology*, 11:4228-234 (1991)] composed of two eight-base pair stretches between nucleotides 166-187 (boxed in the drawing). This silencer may be responsible for the relative lack of activity of the 1.5 kb fragment.

Example 20: Analysis of VEGF-B mRNA in melanomas, normal skin and muscle by RT-PCR.

Normal skin and melanoma tissues were obtained from patients attending the Department of Radiotherapy and Oncology, Helsinki University Central Hospital. Four metastatic melanoma specimens were obtained freshly after surgical excision, immediately embedded in Tissue-tek (Miles) and frozen in liquid nitrogen. Samples of normal skin were obtained from volunteer patients undergoing surgery for mammary carcinoma and excision of a cutaneous naevus. All specimens were inspected by a pathologist to confirm the diagnosis.

Total RNA was isolated by the guanidium isothiocyanate procedure [Chomczynski et al., *Anal. Biochem.* 162:156-159 (1987)]. cDNA was synthesized using 0.2 μ g of random hexadeoxynucleotide primers, 5 units of murine reverse transcriptase, 5 μ g of total RNA as a template and a first-strand cDNA synthesis kit (Pharmacia). After incubation at 37°C for 1 hour, the reaction mixture was stored at -70°C. Negative control samples for PCR amplification were prepared similarly except that reverse transcriptase was not added. β -actin also was tested as an internal standard because it is expressed at a constitutive high level, and its expression does not show much variation in different cells.

For PCR amplification, the primer sequences were selected from the VEGF-B and β -actin genes as follows:

VEGF-B sense: 5'-GCCATGTGTCACCTTCGCAG-3' (SEQ ID NO:19)

VEGF-B antisense: 5'-TGTCCTGGAAGAACACAGCC-3' (SEQ ID NO:29)

β -actin sense: 5'-CGGGAAATCGTGCGTGACAT-3' (SEQ ID NO:30)

β -actin antisense: 5'-GGAGTTGAAGGTAGTTTCGTG-3' (SEQ ID NO:31)

[β -actin sequences comprise nucleotides 2105-2125 and 2411-2432 from Ng et al., *Mol. Cell Biol.* 5:2720-732 (1985)].

An aliquot of 4 μ l from the cDNA reaction product was heated to 94°C for 5 minutes and used as a template for PCR amplification with 20 pmol of primers, 10x PCR buffer, 1 μ l of 20 mM dNTPs and 2.5 U of Taq polymerase. Final volume

was adjusted to 100 μ l with DEPC treated water. Denaturation was at 95°C for 1 minute, annealing at 62°C for 45 seconds, and polymerization at 72°C for 50 seconds, for a total of 35 cycles for VEGF-B and 25 cycles for β -actin. After every 5 cycles, 15 μ l aliquots were taken for analysis.

Electrophoresis of 5 μ l of the PCR reaction mix was performed in a 2% agarose gel containing ethidium bromide. The size marker DNA fragments ranged in length from 24 to 726 base pairs (Φ X174 DNA/Hinf I marker from Promega, Madison, WI, USA). The tested samples thus included four metastatic melanomas, muscle, normal skin, a negative control (without reverse transcriptase), and the Φ X174 DNA/Hinf I size marker. The results of the RT-PCR analysis for VEGF-B (PCR product lengths 323 and 234 bp) and for β -actin show that VEGF-B is highly expressed in all melanomas studied, at levels approximately similar to the expression in muscle tissue. On the other hand, normal skin has very little of the VEGF-B RNA. Similar conclusions can be drawn from Northern blotting and hybridization analysis.

The foregoing results indicate that that VEGF-B is a novel growth factor for endothelial cells which plays a role in vascularization, in particular of muscle. Collateral artery growth in ischemic heart or limb may be promoted by arterial administration of a VEGF-B bolus using techniques described by Takeshita et al., *Am. J. Pathol.*, 147:1649-60 (1995). The cell-association of VEGF-B may have several implications for regulation of vascularization and endothelial cell growth. In developing embryos and in contractile tissues, cell-associated VEGF-B may provide spatial cues to outgrowing endothelial cells during establishment and maintenance of the vascular tree. It could also, through its cell-association, support the regeneration of damaged endothelium in adult vessels. Reendothelialization of arterial injury may be promoted by direct application of VEGF-B using techniques described by

Asahara et al., *Circulation*, 91(11):2793-802 (1995). The ability of VEGF-B to modulate the secretion of VEGF by heterodimer formation suggests an indirect role of VEGF-B in VEGF signalling, thereby regulating receptor binding and/or
5 activation as described by Potgens et al., *J. Biol. Chem.*, 269(52):32879-85 (1994). The formation of multiple heterodimeric complexes of these growth factors could provide a basis for a diverse array of regulatory signals for endothelial cells.

10 VEGF-B can be used as a growth factor for populations of endothelial cells *in vitro*. VEGF-B may be used to promote desirable angiogenesis, i.e. the formation of new blood vessels and capillaries; see Takeshita et al., *supra*. For example, it may be useful in promoting the development
15 of the corpus luteum and endometrium as an aid to initiating and/or maintaining pregnancy. It would also be useful in bone repair by virtue of its action on endothelial cells. Administration of VEGF-B may also be useful in supporting embryogenesis, as well as somatic growth and vascular
20 development and differentiation. Topical application of VEGF-B to wounds may be useful in promoting wound healing, and oral administration of VEGF-B may be useful to accelerate the healing of gastric and/or duodenal ulcers. The ability of VEGF-B to modulate the secretion of VEGF by
25 heterodimer formation could provide a therapeutic role for VEGF-B in diseases where VEGF agonists would be useful; see Potgens et al., *supra*.

30 VEGF-B may exert proliferative effects on mesodermal cells either directly or via improvements in the blood supply.

VEGF-B has been found to be overexpressed in tumors, such as melanomas. Consequently, assays for VEGF-B expression can be used as tools in tumor diagnosis, and suppression of VEGF-B expression, for example with
35 monoclonal antibodies, may be useful to retard tumor growth.

Tumor assays for VEGF-B may be useful as indicators of metastatic risk. For example, use of VEGF-B antibodies analogous to the procedures described by Takahashi et al., *Cancer Res.*, 55:3964-68 (1995) in order to quantitate
5 neovascularization and proliferation could be used as an indicator of metastatic risk from colon cancer. Assays of VEGF-B in body fluids or the tumor itself by histochemistry may be useful as a tumor prognostic factor. An ELISA analogous to the procedure described by Kondo et al.,
10 *Biochemica et Biophysica Acta*, 1221(2):211-14 (1994) may be useful to detect VEGF-B upregulation as a tumor screen. An enzyme linked immunoabsorbent assay of VEGF-B expression using techniques described by Boocock et al., *J. Natl. Cancer Inst.*, 87:506-16 (1995) may be useful as a diagnostic
15 index of ovarian cancer. An assay of VEGF-B expression similar to the VEGF assay described by Weindel et al., *Neurosurgery*, 35:439-48 (1994) may be useful as an indicator of malignancy in brain tumors.

Furthermore, because tumor growth requires
20 angiogenesis, administration of VEGF-B may also be useful in promoting tumor growth in laboratory animals in order to test anti-tumorigenic drugs. VEGF-B may also be useful to increase the microvasculature of hypoxic areas of tumors and make them more sensitive to radiation, radiation sensitizing
25 drugs, etc.

The angiogenic action of VEGF-B may be useful in treating ischemic conditions. Administration of an intra-arterial bolus of VEGF-B by the techniques described in Bouters et al., *American Journal of Physiology*, 267(4 Pt
30 2):H1263-71 (1994) may be useful to treat lower extremity ischemia and increase perfusion in the extremities. Using procedures described by Mesri et al., *Circulation Research*, 76:161-67 (1995) an angiogenic response may be produced in tissue injected with fibroblast cells transduced with a
35 virus which expresses VEGF-B in order to treat tissue ischemia (e.g. myocardial ischemia). VEGF-B or agonists

could be used to stimulate the development of collateral circulation in cases of arterial and/or venous obstruction, e.g. myocardial infarcts, ischaemic limbs, deep venous thrombosis, and/or postpartum vascular problems; see
5 Takeshita et al, *supra*.

A VEGF-B/VEGF-B receptor system may be used as an assay system to detect small molecules as agonists/antagonists for development as new drugs. Examples of small molecules which could be detected include, but are not limited to, organic
10 chemicals, peptides, and RNA molecules.

Pharmaceutical compositions may be produced by admixing a pharmaceutically effective amount of VEGF-B protein with one or more suitable carriers or adjuvants such as water, mineral oil, polyethylene glycol, starch, talcum, lactose,
15 thickeners, stabilizers, suspending agents, etc. Such compositions may be in the form of solutions, suspensions, tablets, capsules, creams, salves, ointments, or other conventional forms.

As demonstrated in Example 7, VEGF-B protein also can
20 be used to produce antibodies. In general, conventional antibody production techniques may be used to produce VEGF-B antibodies. For example, specific monoclonal antibodies may be produced via immunization of fusion proteins obtained by recombinant DNA expression.

25 Labelled monoclonal antibodies, in particular, should be useful in screening for conditions associated with abnormal levels of VEGF-B in the body. For example, an assay of VEGF-B in synovial fluids and/or joint tissue by immunofluorometric techniques analogous to the the procedure
30 described by Fava et al., *Journal of Experimental Medicine*, 180:341-46 (1994) may be useful as a diagnostic indicator of rheumatoid arthritis. A radioimmunoassay of VEGF-B in ocular fluid using techniques described by Aiello et al.,
in *New England Journal of Medicine*, 331(22):1480-87 (1994)
35 may be useful as a diagnostic indicator of diabetic retinopathy, neovascularization of the iris or retinal vein

occlusion. Immunoassays of VEGF-B levels in blood, urine or other bodily fluids may be useful also as a tumor marker; see Kondo et al., *supra*. These monoclonal antibodies to VEGF-B also may be useful in inhibiting angiogenesis associated with high levels of VEGF-B in the body, e.g. in rapidly proliferating, angiogenesis-dependent tumors in mammals, and thereby may retard the growth of such tumors. Treatment with a monoclonal antibody specific for VEGF-B using techniques analogous to those described by Kim et al., in *Nature*, 362(6243):841-44 (1993) may be useful to suppress or inhibit tumor growth *in vivo*. Intravenous and/or subcutaneous injection of monoclonal antibodies to VEGF-B using procedures like those described by Asano et al., in *Cancer Research*, 55:5296-5301 (1995) may be useful to inhibit neovascularization and primary and metastatic growth of solid tumors. For the therapy of humans, chiaserization or humanization of such monoclonal antibodies is to be preferred. Treatment may be effected, e.g., by twice weekly intraperitoneal injection of 10 to 500 μg , preferably 50-100 μg of monoclonal antibody.

VEGF-B antagonists such as antibodies also may be useful to inhibit new blood vessels in diabetic retinopathy, psoriasis, arthropathies and/or vascular tumors such as haemangiomas; see Aiello et al., *supra*.

The foregoing description and examples have been set forth merely to illustrate the invention and are not intended to be limiting. Since modifications of the disclosed embodiments incorporating the spirit and substance of the invention may occur to persons skilled in the art, the invention should be construed to include everything within the scope of the appended claims and equivalents thereof.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: ERIKSSON, Ulf
OLOFSSON, Birgitta
ALITALO, Kari
PAJUSOLA, Katri
- (ii) TITLE OF INVENTION: VASCULAR ENDOTHELIAL GROWTH FACTOR-B AND
DNA CODING THEREFOR
- (iii) NUMBER OF SEQUENCES: 31
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 - (D) STATE: DC
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- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: US
 - (B) FILING DATE: 01-MAR-1996
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 08/397,651
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- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 08/469,427
 - (B) FILING DATE: 06-JUN-1995
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 08/569,063
 - (B) FILING DATE: 06-DEC-1995
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(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 886 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO

WO 96/26736

PCT/US96/02957

(vi) ORIGINAL SOURCE:
(F) TISSUE TYPE: mouse embryo

(vii) IMMEDIATE SOURCE:
(B) CLONE: pcif2

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CGGGACGCC	AGTGGTGCCA	TGGATAGACG	TTTATGCACG	TGCCACATGC	CAGCCCAGGG	60
AGGTGGTGGT	GCCTCTGAGC	ATGGAACTCA	TGGGCAATGT	GGTCAAACAA	CTAGTGCCCA	120
GCTGTGTGAC	TGTGCAGCGC	TGTGGTGGCT	GCTGCCCTGA	CGATGGCCTG	GAATGTGTGC	180
CCACTGGGCA	ACACCAAGTC	CGAATGCAGA	TCCTCATGAT	CCAGTACCCG	AGCAGTCAGC	240
TGGGGGAGAT	GTCCCTGGAA	GAACACAGCC	AATGTGAATG	CAGACCAAAA	AAAAAAAGGA	300
GAGTGCTGTG	AAGCCAGACA	GCCCCAGGAT	CCTCTGCCCG	CCTTGCACCC	AGCGCCGTCA	360
ACGCCCTGAC	CCCCGGACCT	GCCGCTGCCG	CTGCAGACGC	CGCCGCTTCC	TCCATTGCCA	420
AGGGCGGGGC	TTAGAGCTCA	ACCCAGACAC	CTGTAGGTGC	CGGAAGCCGC	GAAAGTGACA	480
AGCTGCTTTC	CAGACTCCAC	GGGCCCGGCT	GCTTTTATGG	CCCTGCTTCA	CAGGGACGAA	540
GAGTGGAGCA	CAGGCAAACC	TCCTCAGTCT	GGGAGGTCAC	TGCCCCAGGA	CCTGGACCTT	600
TTAGAGAGCT	CTCTCGCCAT	CTTTTATCTC	CCAGAGCTGC	CATCTAACAA	TTGTCAAGGA	660
ACCTCATGTC	TCACCTCAGG	GGCCAGGGTA	CTCTCTCACT	TAACCACCCT	GGTCAAGTGA	720
GCATCTTCTG	GCTGGCTGTC	TCCCCTCACT	ATGAAAACCC	CAAACCTTCTA	CCAATAACGG	780
GATTTGGGTT	CTGTTATGAT	AACTGTGACA	CACACACACA	CTCACACTCT	GATAAAAGAG	840
AACTCTGATA	AAAGAGATGG	AAGACACTAA	AAAAAAAAAA	AAAAAA		886

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 102 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:
(F) TISSUE TYPE: mouse embryo

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Gly	Arg	Pro	Val	Val	Pro	Trp	Ile	Asp	Val	Tyr	Ala	Arg	Ala	Thr	Cys
1				5					10					15	
Gln	Pro	Arg	Glu	Val	Val	Val	Pro	Leu	Ser	Met	Glu	Leu	Met	Gly	Asn
			20					25					30		
Val	Val	Lys	Gln	Leu	Val	Pro	Ser	Cys	Val	Thr	Val	Gln	Arg	Cys	Gly
		35					40					45			

Gly Cys Cys Pro Asp Asp Gly Leu Glu Cys Val Pro Thr Gly Gln His
 50 55 60
 Gln Val Arg Met Gln Ile Leu Met Ile Gln Tyr Pro Ser Ser Gln Leu
 65 70 75 80
 Gly Glu Met Ser Leu Glu Glu His Ser Gln Cys Glu Cys Arg Pro Lys
 85 90 95
 Lys Lys Arg Arg Val Leu
 100

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 55 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:
 (F) TISSUE TYPE: mouse embryo

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Lys Pro Asp Ser Pro Arg Ile Leu Cys Pro Pro Cys Thr Gln Arg Arg
 1 5 10 15
 Gln Arg Pro Asp Pro Arg Thr Cys Arg Cys Arg Cys Arg Arg Arg
 20 25 30
 Phe Leu His Cys Gln Gly Arg Gly Leu Glu Leu Asn Pro Asp Thr Cys
 35 40 45
 Arg Cys Arg Lys Pro Arg Lys
 50 55

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 565 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:
 (F) TISSUE TYPE: adult mouse heart

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

GAGCCCCCTG CTCCGTCGCC TGCTGCTTGT TGCCTGCTG CAGCTGGCTC GCACCCAGGC 60
 CCCTGTGTCC CAGTTTGATG GCCCCAGCCA CCAGAAGAAA GTGGTGCCAT GGATAGACGT 120

TTATGCACGT GCCACATGCC AGCCCAGGGA GGTGGTGGTG CCTCTGAGCA TGGAACATCAT 180
 GGGCAATGTG GTCAAACAAC TAGTGCCCAG CTGTGTGACT GTGCAGCGCT GTGGTGGCTG 240
 CTGCCCTGAC GATGGCCTGG AATGTGTGCC CACTGGGCAA CACCAAGTCC GAATGCAGAT 300
 CCTCATGATC CAGTACCCGA GCAGTCAGCT GGGGGAGATG TCCCTGGAAG AACACAGCCA 360
 ATGTGAATGC AGACCAAAAA AAAAGGAGAG TGCTGTGAAG CCAGACAGCC CCAGGATCCT 420
 CTGCCCCGCT TGCACCCAGC GCCGTCAACG CCCTGACCCC CGGACCTGCC GCTGCCGCTG 480
 CAGACGCCGC CGCTTCCTCC ATTGCCAAGG GCGGGGCTTA GAGCTCAACC CAGACACCTG 540
 TAGGTGCCGG AAGCCGCGAA AGTGA 565

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 188 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

- (F) TISSUE TYPE: adult mouse heart

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Met	Ser	Pro	Leu	Leu	Arg	Arg	Leu	Leu	Leu	Val	Ala	Leu	Leu	Gln	Leu	1	5	10	15
Ala	Arg	Thr	Gln	Ala	Pro	Val	Ser	Gln	Phe	Asp	Gly	Pro	Ser	His	Gln	20	25	30	
Lys	Lys	Val	Val	Pro	Trp	Ile	Asp	Val	Tyr	Ala	Arg	Ala	Thr	Cys	Gln	35	40	45	
Pro	Arg	Glu	Val	Val	Val	Pro	Leu	Ser	Met	Glu	Leu	Met	Gly	Asn	Val	50	55	60	
Val	Lys	Gln	Leu	Val	Pro	Ser	Cys	Val	Thr	Val	Gln	Arg	Cys	Gly	Gly	65	70	75	80
Cys	Cys	Pro	Asp	Asp	Gly	Leu	Glu	Cys	Val	Pro	Thr	Gly	Gln	His	Gln	85	90	95	
Val	Arg	Met	Gln	Ile	Leu	Met	Ile	Gln	Tyr	Pro	Ser	Ser	Gln	Leu	Gly	100	105	110	
Glu	Met	Ser	Leu	Glu	Glu	His	Ser	Gln	Cys	Glu	Cys	Arg	Pro	Lys	Lys	115	120	125	
Lys	Glu	Ser	Ala	Val	Lys	Pro	Asp	Ser	Pro	Arg	Ile	Leu	Cys	Pro	Pro	130	135	140	
Cys	Thr	Gln	Arg	Arg	Gln	Arg	Pro	Asp	Pro	Arg	Thr	Cys	Arg	Cys	Arg	145	150	155	160

Cys Arg Arg Arg Arg Phe Leu His Cys Gln Gly Arg Gly Leu Glu Leu
 165 170 175
 Asn Pro Asp Thr Cys Arg Cys Arg Lys Pro Arg Lys
 180 185

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 591 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:
 - (F) TISSUE TYPE: adult mouse heart
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

```

ACCATGAGCC CCCTGCTCCG TCGCCTGCTG CTTGTTGCAC TGCTGCAGCT GGCTCGCACC      60
CAGGCCCTG TGTCCCAGTT TGATGGCCCC AGCCACCAGA AGAAAGTGGT GCCATGGATA      120
GACGTTTATG CACGTGCCAC ATGCCAGCCC AGGGAGGTGG TGGTGCCTCT GAGCATGGAA      180
CTCATGGGCA ATGTGGTCAA ACAACTAGTG CCCAGCTGTG TGACTGTGCA GCGCTGTGGT      240
GGCTGCTGCC CTGACGATGG CCTGGAATGT GTGCCCACTG GGCAACACCA AGTCCGAATG      300
CAGGTACCAG GGCCTATGGG TCAGATCCTC ATGATCCAGT ACCCGAGCAG TCAGCTGGGG      360
GAGATGTCCC TGGAAGAACA CAGCCAATGT GAATGCAGAC CAAAAAAAAA GGAGAGTGCT      420
GTGAAGCCAG ACAGCCCCAG GATCCTCTGC CCGCCTTGCA CCCAGCGCCG TCAACGCCCT      480
GACCCCCGGA CCTGCCGCTG CCGCTGCAGA CGCCGCCGCT TCCTCCATTG CCAAGGGCGG      540
GGCTTAGAGC TCAACCCAGA CACCTGTAGG TGCCGGAAGC CGCGAAAGTG A              591
    
```

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 195 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:
 - (F) TISSUE TYPE: adult mouse heart
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Met Ser Pro Leu Leu Arg Arg Leu Leu Leu Val Ala Leu Leu Gln Leu
 1 5 10 15

WO 96/26736

PCT/US96/02957

Ala Arg Thr Gln Ala Pro Val Ser Gln Phe Asp Gly Pro Ser His Gln
20 25 30

Lys Lys Val Val Pro Trp Ile Asp Val Tyr Ala Arg Ala Thr Cys Gln
35 40 45

Pro Arg Glu Val Val Val Pro Leu Ser Met Glu Leu Met Gly Asn Val
50 55 60

Val Lys Gln Leu Val Pro Ser Cys Val Thr Val Gln Arg Cys Gly Gly
65 70 75 80

Cys Cys Pro Asp Asp Gly Leu Glu Cys Val Pro Thr Gly Gln His Gln
85 90 95

Val Arg Met Gln Val Pro Gly Pro Met Gly Gln Ile Leu Met Ile Gln
100 105 110

Tyr Pro Ser Ser Gln Leu Gly Glu Met Ser Leu Glu Glu His Ser Gln
115 120 125

Cys Glu Cys Arg Pro Lys Lys Lys Glu Ser Ala Val Lys Pro Asp Ser
130 135 140

Pro Arg Ile Leu Cys Pro Pro Cys Thr Gln Arg Arg Gln Arg Pro Asp
145 150 155 160

Pro Arg Thr Cys Arg Cys Arg Cys Arg Arg Arg Phe Leu His Cys
165 170 175

Gln Gly Arg Gly Leu Glu Leu Asn Pro Asp Thr Cys Arg Cys Arg Lys
180 185 190

Pro Arg Lys
195

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 405 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

ACCATGAGCC CCCTGCTCCG TCGCCTGCTG CTTGTTGCAC TGCTGCAGCT GGCTCGCACC 60

CAGGCCCTG TGTCCAGTT TGATGGCCCC AGCCACCAGA AGAAAGTGGT GCCATGGATA 120

GACGTTTATG CACGTGCCAC ATGCCAGCCC AGGGAGGTGG TGGTGCCTCT GAGCATGGAA 180

CTCATGGGCA ATGTGGTCAA ACAACTAGTG CCCAGCTGTG TGA CTGTGCA GCGCTGTGGT 240

GGCTGCTGCC CTGACGATGG CCTGGAATGT GTGCCCACTG GGCAACACCA AGTCCGAATG 300

CAGATCCTCA TGATCCAGTA CCCGAGCAGT CAGCTGGGGG AGATGTCCCT GGAAGAACAC 360

AGCCAATGTG AATGCAGACC AAAAAAAAAA AGGAGAGTGC TGTGA 405

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 133 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

```

Met Ser Pro Leu Leu Arg Arg Leu Leu Leu Val Ala Leu Leu Gln Leu
1           5           10           15
Ala Arg Thr Gln Ala Pro Val Ser Gln Phe Asp Gly Pro Ser His Gln
20           25           30
Lys Lys Val Val Pro Trp Ile Asp Val Tyr Ala Arg Ala Thr Cys Gln
35           40           45
Pro Arg Glu Val Val Val Pro Leu Ser Met Glu Leu Met Gly Asn Val
50           55           60
Val Lys Gln Leu Val Pro Ser Cys Val Thr Val Gln Arg Cys Gly Gly
65           70           75           80
Cys Cys Pro Asp Asp Gly Leu Glu Cys Val Pro Thr Gly Gln His Gln
85           90           95
Val Arg Met Gln Ile Leu Met Ile Gln Tyr Pro Ser Ser Gln Leu Gly
100          105          110
Glu Met Ser Leu Glu Glu His Ser Gln Cys Glu Cys Arg Pro Lys Lys
115          120          125
Lys Arg Arg Val Leu
130

```

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 570 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

- (vi) ORIGINAL SOURCE:
 (F) TISSUE TYPE: human fibrosarcoma

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

```

ACCATGAGCC CTCTGCTCCG CCGCCTGCTG CTCGCCGCAC TCCTGCAGCT GGCCCCCGCC      60
CAGGCCCTG TCTCCAGCC TGATGCCCTT GGCCACCAGA GGAAAGTGGT GTCATGGATA      120
GATGTGTATA CTCGCGCTAC CTGCCAGCCC CGGGAGGTGG TGGTGCCCTT GACTGTGGAG      180
CTCATGGGCA CCGTGGCAA ACAGCTGGTG CCCAGCTGCG TGA CTGTGCA GCGCTGTGGT      240

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WO 96/26736

PCT/US96/02957

GGCTGCTGCC CTGACGATGG CCTGGAGTGT GTGCCCACTG GGCAGCACCA AGTCCGGATG 300
 CAGATCCTCA TGATCCGGTA CCCGAGCAGT CAGCTGGGGG AGATGTCCCT GGAAGAACAC 360
 AGCCAGTGTG AATGCAGACC TAAAAAAAAG GACAGTGCTG TGAAGCCAGA CAGCCCCAGG 420
 CCCCTCTGCC CACGCTGCAC CCAGCACCAC CAGCGCCCTG ACCCCCGGAC CTGCCGCTGC 480
 CGCTGCCGAC GCCGCAGCTT CCTCCGTTGC CAAGGGCGGG GCTTAGAGCT CAACCCAGAC 540
 ACCTGCAGGT GCCGGAAGCT GCGAAGGTGA 570

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 188 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(F) TISSUE TYPE: human fibrosarcoma

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Met Ser Pro Leu Leu Arg Arg Leu Leu Leu Ala Ala Leu Leu Gln Leu
 1 5 10 15
 Ala Pro Ala Gln Ala Pro Val Ser Gln Pro Asp Ala Pro Gly His Gln
 20 25 30
 Arg Lys Val Val Ser Trp Ile Asp Val Tyr Thr Arg Ala Thr Cys Gln
 35 40 45
 Pro Arg Glu Val Val Val Pro Leu Thr Val Glu Leu Met Gly Thr Val
 50 55 60
 Ala Lys Gln Leu Val Pro Ser Cys Val Thr Val Gln Arg Cys Gly Gly
 65 70 75 80
 Cys Cys Pro Asp Asp Gly Leu Glu Cys Val Pro Thr Gly Gln His Gln
 85 90 95
 Val Arg Met Gln Ile Leu Met Ile Arg Tyr Pro Ser Ser Gln Leu Gly
 100 105 110
 Glu Met Ser Leu Glu Glu His Ser Gln Cys Glu Cys Arg Pro Lys Lys
 115 120 125
 Lys Asp Ser Ala Val Lys Pro Asp Ser Pro Arg Pro Leu Cys Pro Arg
 130 135 140
 Cys Thr Gln His His Gln Arg Pro Asp Pro Arg Thr Cys Arg Cys Arg
 145 150 155 160
 Cys Arg Arg Arg Ser Phe Leu Arg Cys Gln Gly Arg Gly Leu Glu Leu
 165 170 175
 Asn Pro Asp Thr Cys Arg Cys Arg Lys Leu Arg Arg
 180 185

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 624 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

- (F) TISSUE TYPE: mouse

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

```

ATGAGCCCC TGCTCCGTCG CCTGCTGCTT GTTGCACTGC TGCAGCTGGC TCGCACCCAG      60
GCCCTGTGT CCCAGTTTGA TGGCCCCAGC CACCAGAAGA AAGTGGTGCC ATGGATAGAC      120
GTTTATGCAC GTGCCACATG CCAGCCCAGG GAGGTGGTGG TGCCTCTGAG CATGGAACTC      180
ATGGGCAATG TGGTCAAACA ACTAGTGCCC AGCTGTGTGA CTGTGCAGCG CTGTGGTGGC      240
TGCTGCCCTG ACGATGGCCT GGAATGTGTG CCCACTGGGC AACACCAAGT CCGAATGCAG      300
ATCCTCATGA TCCAGTACCC GAGCAGTCAG CTGGGGGAGA TGTCCCTGGA AGAACACAGC      360
CAATGTGAAT GCAGACCAA AAAAAAGGAG AGTGCTGTGA AGCCAGACAG GGTTGCCATA      420
CCCCACCACC GTCCCCAGCC CCGCTCTGTT CCGGGCTGGG ACTCTACCCC GGGAGCATCC      480
TCCCCAGCTG ACATCATCCA TCCCCTCCA GCCCCAGGAT CCTCTGCCCC CCTTGCACCC      540
AGCGCCGTCA ACGCCCTGAC CCCCAGACCT GCCGCTGCCG CTGCAGACGC CGCCGCTTCC      600
TCCATTGCCA AGGGCGGGGC TTAG                                             624
    
```

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 207 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:

- (F) TISSUE TYPE: mouse

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

```

Met Ser Pro Leu Leu Arg Arg Leu Leu Leu Val Ala Leu Leu Gln Leu
1           5           10           15
Ala Arg Thr Gln Ala Pro Val Ala Gln Phe Asp Gly Pro Ser His Gln
20           25           30
Lys Lys Val Val Pro Trp Ile Asp Val Tyr Ala Arg Ala Thr Cys Gln
35           40           45
    
```

WO 96/26736

PCT/US96/02957

Pro	Arg	Glu	Val	Val	Val	Pro	Leu	Ser	Met	Glu	Leu	Met	Gly	Asn	Val
	50					55					60				
Val	Lys	Gln	Leu	Val	Pro	Ser	Cys	Val	Thr	Val	Gln	Arg	Cys	Gly	Gly
65					70					75					80
Cys	Cys	Pro	Asp	Asp	Gly	Leu	Glu	Cys	Val	Pro	Thr	Gly	Gln	His	Gln
			85						90					95	
Val	Arg	Met	Gln	Ile	Leu	Met	Ile	Gln	Tyr	Pro	Ser	Ser	Gln	Leu	Gly
			100					105					110		
Glu	Met	Ser	Leu	Glu	Glu	His	Ser	Gln	Cys	Glu	Cys	Arg	Pro	Lys	Lys
		115					120					125			
Lys	Glu	Ser	Ala	Val	Lys	Pro	Asp	Arg	Val	Ala	Ile	Pro	His	His	Arg
	130					135					140				
Pro	Gln	Pro	Arg	Ser	Val	Pro	Gly	Trp	Asp	Ser	Thr	Pro	Gly	Ala	Ser
145					150					155					160
Ser	Pro	Ala	Asp	Ile	Ile	His	Pro	Thr	Pro	Ala	Pro	Gly	Ser	Ser	Ala
				165					170					175	
Arg	Leu	Ala	Pro	Ser	Ala	Val	Asn	Ala	Leu	Thr	Pro	Gly	Pro	Ala	Ala
			180					185					190		
Ala	Ala	Ala	Asp	Ala	Ala	Ala	Ser	Ser	Ile	Ala	Lys	Gly	Gly	Ala	
		195					200					205			

(2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 624 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:
 (F) TISSUE TYPE: human

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

ATGAGCCCTC	TGCTCCGCCG	CCTGCTGCTC	GCCGCACTCC	TGCAGCTGGC	CCCCGCCAG	60
GCCCCTGTCT	CCCAGCCTGA	TGCCCCTGGC	CACCAGAGGA	AAGTGGTGTC	ATGGATAGAT	120
GTGTATACTC	GCGCTACCTG	CCAGCCCCGG	GAGGTGGTGG	TGCCCTTGAC	TGTGGAGCTC	180
ATGGGCACCG	TGGCAAACA	GCTGGTGCCC	AGCTGCGTGA	CTGTGCAGCG	CTGTGGTGGC	240
TGCTGCCCTG	ACGATGGCCT	GGAGTGTGTG	CCCACTGGGC	AGCACCAAGT	CCGGATGCAG	300
ATCCTCATGA	TCCGGTACCC	GAGCAGTCAG	CTGGGGGAGA	TGTCCCTGGA	AGAACACAGC	360
CAGTGTGAAT	GCAGACCTAA	AAAAAAGGAC	AGTGCTGTGA	AGCCAGACAG	GGCTGCCACT	420
CCCCACCACC	GTCCCCAGCC	CCGTTCTGTT	CCGGGCTGGG	ACTCTGCCCC	CGGAGCACCC	480
TCCCCAGCTG	ACATCACCCA	TCCCACTCCA	GCCCCAGGCC	CCTCTGCCCA	CGCTGCACCC	540

AGCACCACCA GCGCCCTGAC CCCC GGACCT GCCGCCGCCG CTGCCGACGC CGCAGCTTCC 600
 TCCGTTGCCA AGGGCGGGGC TTAG 624

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 207 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:

- (F) TISSUE TYPE: human

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Met	Ser	Pro	Leu	Leu	Arg	Arg	Leu	Leu	Leu	Ala	Ala	Leu	Leu	Gln	Leu	1	5	10	15
Ala	Pro	Ala	Gln	Ala	Pro	Val	Ser	Gln	Pro	Asp	Ala	Pro	Gly	His	Gln	20	25	30	
Arg	Lys	Val	Val	Ser	Trp	Ile	Asp	Val	Tyr	Thr	Arg	Ala	Thr	Cys	Gln	35	40	45	
Pro	Arg	Glu	Val	Val	Val	Pro	Leu	Thr	Val	Glu	Leu	Met	Gly	Thr	Val	50	55	60	
Ala	Lys	Gln	Leu	Val	Pro	Ser	Cys	Val	Thr	Val	Gln	Arg	Cys	Gly	Gly	65	70	75	80
Cys	Cys	Pro	Asp	Asp	Gly	Leu	Glu	Cys	Val	Pro	Thr	Gly	Gln	His	Gln	85	90	95	
Val	Arg	Met	Gln	Ile	Leu	Met	Ile	Arg	Tyr	Pro	Ser	Ser	Gln	Leu	Gly	100	105	110	
Glu	Met	Ser	Leu	Glu	Glu	His	Ser	Gln	Cys	Glu	Cys	Arg	Pro	Lys	Lys	115	120	125	
Lys	Asp	Ser	Ala	Val	Lys	Pro	Asp	Arg	Ala	Ala	Thr	Pro	His	His	Arg	130	135	140	
Pro	Gln	Pro	Arg	Ser	Val	Pro	Gly	Trp	Asp	Ser	Ala	Pro	Gly	Ala	Pro	145	150	155	160
Ser	Pro	Ala	Asp	Ile	Thr	His	Pro	Thr	Pro	Ala	Pro	Gly	Pro	Ser	Ala	165	170	175	
His	Ala	Ala	Pro	Ser	Thr	Thr	Ser	Ala	Leu	Thr	Pro	Gly	Pro	Ala	Ala	180	185	190	
Ala	Ala	Ala	Asp	Ala	Ala	Ala	Ser	Ser	Val	Ala	Lys	Gly	Gly	Ala	195	200	205		

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 13 amino acids

WO 96/26736

PCT/US96/02957

(B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Pro Xaa Cys Val Xaa Xaa Xaa Arg Cys Xaa Gly Cys Cys
 1 5 10

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 1550 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

CTCGAGATCT	GTTTGTGTC	TTGGAACAAT	ACGGTTTAGA	GGTGACTGGC	GGGTGACGAG	60
AACATATGCG	AGTTCACCTA	AGAGAAAAGC	TGAATGAGGC	AATGCCTCTT	CCTGACCATA	120
TCTCTTACTC	AGATAACTAT	AGAATTTATT	GTCCAGTAAA	GGGTATATTA	AAAAATCATA	180
TTAAAAGTCA	TACAGTGAAG	TTGTCCAGGG	AAATCAAGAC	TTAACAGTCT	CACTCTGACA	240
ATAATGAACA	GGGGGATTCC	CTCAAGATAG	ACTAGGACAT	GACCCACAC	TGGCAGGTAG	300
TAGTACCAGA	AAAGAACGCA	TGGAAAATCT	TTACCTTATG	CTTGAGGTAG	GGACCAGGCT	360
AAAGTGAAGG	CCAGACCTAA	AATTCTATCT	AAAATAAATC	CACAATCGAA	GAAAATATGT	420
GGTGTACAGG	TATAGAATGT	CTTTACTGGA	TCATTGAAAT	AGTAAGATAA	ATTCAACTTT	480
TTACATTGTT	TTCTTTTCCT	CCAGTTAGGG	CTTGAGACCT	TCGTCTCTGG	AGAGTGACTG	540
TCAATTGGAG	CCCTGCTTTC	TGGGTTTCTG	GCCAGGGGGG	TTGTGGATGC	TTAACATGTG	600
CCTTTCACAG	GACACTTCCT	TACCCAGCA	GTGGCCANGT	GTGCATCCCA	CGACCAGGCC	660
TCCCTCTCAC	GGAACATCTG	TTGAGACTAG	GAGATGCCTG	GTGACTGTTG	CCTGACCTGT	720
GTCCTGTGTA	TTTCTGACAA	GAGCCACTCT	CAAAGACCCT	GGCCAGGAGG	AGAGTTAGGT	780
TCCAGTGTAG	GTCAGCTCAG	ACAGATGGAG	GCCACAGAA	CAAACATGGG	AAATCACAGA	840
AGTAGGTTTA	TTACTCACAG	ATCCCTATCC	CAACCACCCA	GGTGCCCTCT	CCTCCAGGGC	900
CAACAGAGGC	ATCCTTCAGC	AGGAGCGACA	ACGGCTAGGG	CAGCGGCAAG	CCGCCACCAT	960
CCGAGCCAAC	CCAGGCCCCG	AGATCGTGCC	CCGGGGCGCC	GGCCCCTGAG	GGGCTCACCT	1020
GGATGGGGCC	TGCATGCGTT	CCCGCTTTGC	TTCCTTCCCT	GGACGGCCCG	CTCCCCGAA	1080
ACGCGCCGCC	AATAAAGTGA	TTCGCAGAGC	TCGTGTGCGG	CTCCCTCCTT	AAGGCCCGAC	1140
GCCCCCGGCC	CCGGCCTCGC	CAAGGGCAGC	GCCCCGGCCT	CCGGGTAGTG	GCGGCCGGCG	1200

ACTGGGGAGC CCAGCCTCCT GGGCGGTGCG TCCCCTTCCC CCTGCCGCGG CGGGAGGCGG 1260
 GAGGGGGTGT GTGGAGGAGG CGGGCCCCGC CGACGGCCTC GCCCCCCCAC CCCGCCGCC 1320
 CGCCCCCGCC CCACGGGCCC GGTGGGGAGC GCGTGTCTGG GTCACATGAG CCGCCTGCCC 1380
 GCCAGCCCGG GCCCAGCCCC CCGCCGCCCC CGCCGTCCCC GCCGCCGCTG CCCGCCGCCA 1440
 CCGGCCGCC GCGCCCGG CTCCTCCGGC CGCCTTCGCT GCGCTGCNTG CGCTGCCTGC 1500
 ACCCAGGGCT CGGGAGGGGG CCGCGGAGGA GCCGCCCCC GCGCCCGGCC 1550

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

CACCATGAGC CCTCTGCTCC 20

(2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

GCCATGTGTC ACCTTCGCAG 20

(2) INFORMATION FOR SEQ ID NO:20:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 21 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

GGGCATCAGG CTGGGAGACA G 21

(2) INFORMATION FOR SEQ ID NO:21:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 18 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single

WO 96/26736

PCT/US96/02957

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Cys Pro Val Ser Gln Phe Asp Gly Pro Ser His Gln Lys Lys Val Val
 1 5 10 15

Pro Cys

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 23 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Ser Gln Pro Asp Ala Pro Gly His Gln Arg Lys Val Val Ser Trp Ile
 1 5 10 15

Asp Val Tyr Thr Arg Ala Thr
 20

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 19 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

CACAGCCAAT GTGAATGCA

19

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 31 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

GCTCTAAGCC CCGCCCTTGG CAATGGAGGA A

31

(2) INFORMATION FOR SEQ ID NO:25:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 28 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:
 ACGTAGATCT TCACTTTCGC GGCTTCCG 28
- (2) INFORMATION FOR SEQ ID NO:26:
- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 23 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:
 CTCTGTTCCG GGCTGGGACT CTA 23
- (2) INFORMATION FOR SEQ ID NO:27:
- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 27 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:
 TCAGGGCGTT GACGGCGCTG GGTGCAA 27
- (2) INFORMATION FOR SEQ ID NO:28:
- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 21 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:
 CCTGACGATG GCCTGGAGTG T 21
- (2) INFORMATION FOR SEQ ID NO:29:
- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 21 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

TGTCCCTGGA AGAACACAGC C

21

(2) INFORMATION FOR SEQ ID NO:30:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

CGGAAATCG TCGTGACAT

20

(2) INFORMATION FOR SEQ ID NO:31:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

GGAGTTGAAG GTAGTTTCGT G

21

CLAIMS:

1. An isolated nucleic acid which codes for a protein which comprises the amino acid sequence:

Pro-Xaa-Cys-Val-Xaa-Xaa-Xaa-Arg-Cys-Xaa-Gly-Cys-Cys

(SEQ ID NO:16) and has the property of promoting proliferation of endothelial cells or mesodermal cells, said nucleic acid being the DNA of SEQ ID NO:1, the DNA of SEQ ID NO:4, the DNA of SEQ ID NO:6, the DNA of SEQ ID NO:8, the DNA of SEQ ID NO:10, the DNA of SEQ ID NO:12, the DNA of SEQ ID NO:14, or a nucleic acid which hybridizes at 42°C in 50% formamide, 5 x 0.75M sodium chloride and 0.075M sodium citrate pH 7.0 or 5 x buffer composed of 0.9M sodium chloride, 0.05M sodium phosphate, pH 7.7 and 0.005M ethylene diamine tetraacetic acid, 1% to 2% sodium dodecyl sulfate, 5 to 10 x Denhardt's solution and 100 µg/ml of salmon sperm DNA followed by washing 2 times for 30 minutes in 2 x SSC at 55°C with the complement of at least one of the foregoing DNA sequences.

2. An isolated nucleic acid according to claim 1, wherein said nucleic acid is a cDNA.

3. An isolated nucleic acid according to claim 1, comprising the sequence set forth as SEQ ID NO:1.

4. An isolated nucleic acid according to claim 1, wherein said nucleic acid is a mammalian DNA.

5. An isolated nucleic acid according to claim 4, wherein said nucleic acid is a murine DNA.

6. An isolated nucleic acid according to claim 4, wherein said nucleic acid is a human DNA.

7. An isolated nucleic acid according to claim 1, wherein said nucleic acid codes for a protein which promotes proliferation of vascular endothelial cells.

8. An isolated nucleic acid according to claim 1, comprising the sequence set forth as SEQ ID NO:4.

9. An isolated nucleic acid according to claim 1, comprising the sequence set forth as SEQ ID NO:6.

10. An isolated nucleic acid according to claim 1, comprising the sequence set forth as SEQ ID NO:8.

11. An isolated nucleic acid according to claim 1, comprising the sequence set forth as SEQ ID NO:10.

12. An isolated nucleic acid according to claim 1, comprising the sequence set forth as SEQ ID NO:12.

13. An isolated nucleic acid according to claim 1, comprising the sequence set forth as SEQ ID NO:14.

14. A vector comprising the isolated nucleic acid according to claim 1, which nucleic acid is operably linked with a promoter sequence.

15. A vector according to claim 14, wherein said vector is a eukaryotic vector.

16. A vector according to claim 14, wherein said vector is a prokaryotic vector.

17. A vector according to claim 14, wherein said vector is a plasmid.

18. An isolated protein which comprises a characteristic sequence

Pro-Xaa-Cys-Val-Xaa-Xaa-Xaa-Arg-Cys-Xaa-Gly-Cys-Cys

(SEQ ID NO:16) and has the property of promoting proliferation of endothelial cells or mesodermal cells, said protein comprising an amino acid sequence which is the amino acid sequence of FIG. 1 (SEQ ID NO:2), the amino acid sequence of FIG. 2 (SEQ ID NO:3), the amino acid sequence of FIG. 4 (SEQ ID NO:5), the amino acid sequence of FIG. 6 (SEQ ID NO:7), the amino acid sequence of FIG. 8 (SEQ ID NO:9), the amino acid sequence of FIG. 11 (SEQ ID NO:11), the amino acid sequence of FIG. 13 (SEQ ID NO:13), or the amino acid sequence of FIG. 15 (SEQ ID NO:15).

19. An isolated protein according to claim 18, wherein said protein comprises the amino acid sequence of FIG. 1 (SEQ ID NO:2).

20. An isolated protein according to claim 18, wherein said isolated protein comprises the amino acid sequence of FIG. 2 (SEQ ID NO:3).

21. An isolated protein according to claim 18, wherein said isolated protein comprises the amino acid sequence of FIG. 4 (SEQ ID NO:5).

22. An isolated protein according to claim 18, wherein said isolated protein comprises the amino acid sequence of FIG. 6 (SEQ ID NO:7).

23. An isolated protein according to claim 18, wherein said isolated protein comprises the amino acid sequence of FIG. 8 (SEQ ID NO:9).

24. An isolated protein according to claim 18, wherein said isolated protein comprises the amino acid sequence of FIG. 11 (SEQ ID NO:11).

25. An isolated protein according to claim 18, wherein said protein comprises the amino acid sequence of FIG. 13 (SEQ ID NO:13).

26. An isolated protein according to claim 18, wherein said protein comprises the amino acid sequence of FIG. 15 (SEQ ID NO:15).

27. An isolated protein according to claim 18, wherein said isolated protein is a mammalian protein.

28. An isolated protein according to claim 27, wherein said mammalian protein is a murine protein.

29. An isolated protein according to claim 27, wherein said mammalian protein is a human protein.

30. An isolated protein according to claim 18, wherein said isolated protein promotes proliferation of vascular endothelial cells.

31. An isolated protein having VEGF-B activity produced by expression of a DNA which is the DNA of FIGS. 1 and 2 (SEQ ID NO:1), the DNA of FIG. 3 (SEQ ID NO:4), the DNA of FIG. 5 (SEQ ID NO:6), the DNA of FIG. 7 (SEQ ID NO:8), the DNA of FIG. 10 (SEQ ID NO:10), or DNA which hybridizes at 42°C in 50% formamide, 5 x 0.75M sodium chloride and 0.075M sodium

citrate pH 7.0 or 5 x buffer composed of 0.9M sodium chloride, 0.05M sodium phosphate, pH 7.7 and 0.005M ethylene diamine tetraacetic acid, 1% to 2% sodium dodecyl sulfate, 5 to 10 x Denhardt's solution and 100 µg/ml of salmon sperm DNA followed by washing 2 times for 30 minutes in 2 x SSC at 55°C with the compliment of at least one of the foregoing DNA sequences.

32. A pharmaceutical composition comprising an effective endothelial or mesodermal cell proliferation promoting amount of the isolated protein according to claim 18, and at least one pharmaceutical carrier or diluent.

33. An antibody which reacts specifically with the isolated protein according to claim 18.

34. A host cell transformed or transfected with the vector according to claim 14.

35. A host cell according to claim 34, wherein said host cell expresses a protein having the property of promoting proliferation of endothelial or mesodermal cells.

36. A transfected host cell according to claim 34, wherein said host cell is a eukaryotic cell.

37. A transfected host cell according to claim 34, wherein said host cell is a COS cell.

38. A transformed host cell according to claim 34, wherein said host cell is a prokaryotic cell.

39. A transformed host cell according to claim 34, wherein said host cell is a 293EBNA cell.

40. A transformed host cell according to claim 34, wherein said host cell is an insect cell.

41. A method for quantitatively detecting VEGF-B in a test sample, the method comprising reacting the antibody according to claim 33 with a sample suspected of containing VEGF-B.

42. The method of Claim 41, wherein the antibody is a labeled antibody.

43. A method for detecting VEGF-B in a test sample, the method comprising providing at least one pair of primers complementary to the isolated nucleic acid according to claim 1; amplifying the VEGF-B polynucleotide by polymerase chain reaction and comparing the VEGF-B polynucleotide with the nucleic acid according to claim 1.

44. A pharmaceutical composition comprising an effective VEGF-B-binding amount of the antibody according to Claim 33, and at least one pharmaceutically acceptable carrier or adjuvant.

45. A pharmaceutical composition comprising an effective endothelial or mesodermal cell proliferation promoting amount of the isolated protein according to Claim 18, and VEGF.

46. A pharmaceutical composition of Claim 45, further comprising heparin.

47. A pharmaceutical composition of Claim 44, further comprising heparin.

48. A pharmaceutical composition comprising an effective angiogenesis-stimulating amount of the isolated protein according to claim 18, and heparin.

49. An isolated protein dimer comprising the isolated protein according to claim 18.

50. An isolated protein dimer according to claim 49, wherein said protein dimer is a homodimer of said protein.

51. An isolated protein dimer according to claim 49, wherein said protein dimer is a heterodimer of said protein and VEGF.

52. An isolated protein dimer according to claim 49, wherein said protein dimer is a disulfide-linked dimer.

53. A method of promoting release of at least one protein according to claim 18, from a cell in a cell culture, which cell expresses said at least one protein, said method comprising exposing the cell to heparin.

54. The method according to claim 53, further comprising promoting release of VEGF.

55. A vector comprising (a) the nucleic acid according to claim 1 or (b) a fragment of the nucleic acid according to claim 1, the nucleic acid or fragment in antisense orientation, wherein a nucleic acid complementary to said nucleic acid or fragment retards expression of a VEGF-B protein.

56. A method of retarding expression of VEGF-B from a cell *in vitro* which expresses VEGF-B, said method comprising transfecting said cell with the vector according to claim 55.

57. A method according to claim 56, wherein said cell is a tumor cell.

58. An isolated DNA comprising a nucleotide sequence comprising the sequence of Figure 25 (SEQ ID NO:17) or a DNA which hybridizes to the complement of said nucleotide sequence at 42° in 50% formamide, 5 x 0.75M sodium chloride and 0.075M sodium citrate pH 7.0 or 5 x buffer composed of 0.9M sodium chloride, 0.05M sodium phosphate, pH 7.7 and 0.005M ethylene diamine tetraacetic acid, 1% to 2% sodium dodecyl sulphate, 5 to 10 x Denhardt's solution and 100 µg/ml of salmon sperm DNA followed by washing 2 times for 30 minutes in 2 x SSC at 55°C, wherein the DNA which hybridizes to the complement of SEQ. ID NO:17 has promoter activity.

59. A host cell transformed or transfected with a vector comprising the isolated nucleic acid sequence according to claim 1, operatively linked to a promoter, such that said host cell expresses a VEGF-B protein.

60. A host cell according to claim 59, wherein said VEGF-B protein comprises a sequence of amino acids which is the amino acid sequence of FIG. 1 (SEQ ID NO:2), the amino acid sequence of FIG. 2 (SEQ ID NO: 3), the amino acid sequence of FIG. 4 (SEQ ID NO:5), the amino acid sequence of FIG. 6 (SEQ ID NO:7), the amino acid sequence of FIG. 8 (SEQ ID NO:9), the amino acid sequence of FIG.

11 (SEQ ID NO:11), the amino acid sequence of FIG. 13 (SEQ ID NO:13), or the amino acid sequence of FIG. 15 (SEQ ID NO: 15), wherein said VEGF-B protein has VEGF-B activity.

61. A kit comprising at least one oligonucleotide primer complementary to a nucleic acid sequence which is the DNA of SEQ ID NO:1, the DNA of SEQ ID NO:4, the DNA of SEQ ID NO:6, the DNA of SEQ ID NO:8, the DNA of SEQ ID NO:10, the DNA of SEQ ID NO:12, the DNA of SEQ ID NO:14, or the DNA substantially identical to at least one of the foregoing sequences wherein the DNA codes for a protein which comprises the amino acid sequence Pro-Xaa-Cys-Val-Xaa-Xaa-Xaa-Arg-Cys-Xaa-Gly-Cys-Cys, and a suitable container.

62. A method for obtaining the isolated protein according to claim 18, from a cell which expresses said protein, said method comprising:

- exposing the cell to heparin to induce release of said protein, and
- collecting the released protein.

63. The method according to claim 62, further comprising obtaining VEGF.

64. A method of making a vector which expresses VEGF-B protein, said method comprising incorporating the isolated nucleic acid according to claim 1, into a vector in operatively linked relation with a promoter.

65. An isolated nucleic acid molecule which encodes a human VEGF-B molecule, wherein said isolated nucleic acid molecule hybridizes to the complement of at least one

nucleic acid molecule which is SEQ ID NO:1, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8 or SEQ ID NO:12, at 42° in 50% formamide, 5 x 0.75M sodium chloride and 0.075M sodium citrate pH 7.0 or 5 x buffer composed of 0.9M sodium chloride, 0.05M sodium phosphate, pH 7.7 and 0.005M ethylene diamine tetraacetic acid, 1% to 2% sodium dodecyl sulphate, 5 to 10 x Denhardt's solution and 100 µg/ml of salmon sperm DNA followed by washing 2 times for 30 minutes in 2 x SSC at 55°C.

66. A kit for amplifying a vascular endothelial growth factor-B (VEGF-B) polynucleotide in a test sample, said kit comprising a polymerase and at least one pair of primers complementary to the isolated nucleic acid according to claim 1, for amplifying the VEGF-B polynucleotide by polymerase chain reaction in order to facilitate a sequence comparison of the VEGF-B polynucleotide with the nucleic acid according to claim 1.

67. An isolated protein according to Claim 18, comprising the amino acid sequence Lys-Pro-Asp-Ser-Pro-Arg (amino acids 1-6 of SEQ ID NO:3).

68. An isolated protein according to Claim 18, comprising the amino acid sequence Gln-Arg-Pro-Asp-Pro-Arg (amino acids 17-22 of SEQ ID NO:3).

69. An isolated protein according to Claim 18, comprising the amino acid sequence Met-Ser-Pro-Leu-Leu-Arg-Arg-Leu-Leu-Leu-Ala-Ala-Leu-Leu-Gln-Leu-Ala (amino acids 1-17 of SEQ ID NO:11).

70. An isolated protein according to Claim 18, comprising the amino acid sequence Met-Ser-Pro-Leu-Leu-Arg-Arg-Leu-Leu-Leu (amino acids 1-10 of SEQ ID NO:5).

71. An isolated protein according to Claim 18, comprising the amino acid sequence Ala-Leu-Leu-Gln-Leu-Ala (amino acids 12-17 of SEQ ID NO:5).

72. An isolated protein according to Claim 18, comprising the amino acid sequence His-Pro-Thr-Pro-Ala-Pro-Gly (amino acids 167-173 of SEQ ID NO:13).

73. An isolated protein having VEGF-B activity produced by expression of a DNA which is the DNA of FIGS. 1 and 2 (SEQ ID NO:1), the DNA of FIG. 3 (SEQ ID NO:4), the DNA of FIG. 5 (SEQ ID

NO:6), the DNA of FIG. 7 (SEQ ID NO:8), the DNA of FIG. 10, (SEQ ID NO:10), the DNA of FIG. 12 (SEQ ID NO:12), the DNA of FIG. 14 (SEQ ID NO:14), or DNA which hybridizes with the complement of least one of the foregoing DNA sequences at 42°C. in 50% formamide, 5 x 0.75M sodium chloride and 0.075M sodium citrate pH 7.0 or 5 x buffer composed of 0.9M sodium chloride, 0.05M sodium phosphate. pH 7.7 and 0.005M ethylene diamine tetraacetic acid, 1% to 2% sodium dodecyl sulfate, 5 to 10 x Denhardt's solution and 100 µg/ml of salmon sperm DNA followed by washing 2 times for 30 minutes in 2 x SSC at 55°C.

```

1      Gly Arg Pro Val Val Val Pro Trp Ile Asp Val Tyr Ala Arg Ala Thr Cys Gln
      17  GGA CGC CCA GTG GTG GTG CCA CCA TGG ATA GAC GTT TAT TAT GCA CGT CGC GCC ACA TGC CAG
      34  Pro Arg Glu Val Val Val Val Pro Val Val Val Val Val Val Val Val Val Val Val Val
      55  CCC AGG GAG GTG GTG GTG CCA CCA TGG CCT CCT CTG AGC Thr Val Thr Arg Cys Cys
      51  Lys Gln Leu Val Val Pro Ser Ser AGC TGT TGT GTG ACT ACT Pro Val Pro Thr Gly Gln Val Arg Met
      68  AAA CAA CTA GTG CCC AGC Glu Glu Gly Leu Cys Cys TGT TGT GGT GGC Gly Cys Cys TGC
      106 Pro Asp Asp Gly Leu Ile Ile Gln Tyr Cys Cys Val Pro Thr Gly Gln His Gln Val Arg Met
      157 CCT GAC GAT GGC CTG GAA TGT TGT GTG CCC ACT ACT Thr Ser Ser Ser Ser Ser Ser Ser Ser
      208 Gln Ile Leu Met Ile Ile Gln Tyr Cys Cys Val Pro Ser Ser Ser Ser Ser Ser Ser Ser
      259 CAG ATC CTC ATG ATC CAG TAC CCG AGC AGT AGT CAG CTG CAG CTG Lys Lys Lys Arg Arg Val Leu
      102 Glu Glu His Ser Gln Cys Glu Cys Glu Cys Arg Pro Pro Lys Lys Lys Arg Arg Val Leu
      GAA GAA CAC AGC CAA TGT GAA TGC AGA CCA AAA AAA AAA AGG AGA GTG CTG
      Stop
      310 TGA AGCCAGACAGCCCCAGGATCCTCTGCCCGCCTTGCACCCAGCGCCGTC AACGCCCTGACCCCC
      376 GGACCTGCCCGCTGCCGCTGCAGACGCCCGCCTTCCCTCCATTGCCAAGGGCGGGCTTAGAGCTCAA
      443 CCCAGACACCTGTAGGTGCCGGAAGCCGGAAGTGACAAGCTGCTTCCAGACTCCACGGGCCCGG
      510 CTGCTTTTATGGCCCTGCTTCACAGGGACGAGAGTGGAGCACAGGCA AACCTCCTCAGTCTGGGAG
      577 GTCACTGCCCCAGGACCTGGACCCTTTAGAGAGCTCTCTCGCCATCTTTTATCTCCAGAGCTGCCA
      644 TCTAACAAATTGTCAAGGAACCTCATGTCTCACCTCAGGGCCAGGGTACTCTCTCACTTAACCACCC
      711 TGGTCAAGTGAGCATCTTCTGGCTGGCTGTCTCCCTCACTATGA AAAACCCCAACTTCTACCAATA
      778 ACGGGATTGGGTCTGTATGATAACTGTGACACACACACTCACACTCTGATAAAAGAGAAC
      845 TCTGATAAAAGAGATGGAGACACTAA AAAAAA AAAA AAAAAA

```

(SEQ ID NOS: 1 & 2)

FIG. 1

1 CAGTGGTGCCATGGATAGACGTTTATGCACGTGCCACATGCCAGCCAGGGAGGTGGTGGTGCCTCT
 10 CAGTGGTGCCATGGATAGACGTTTATGCACGTGCCACATGCCAGCCAGGGAGGTGGTGGTGCCTCT
 77 GAGCATGGAACTCATGGCAATGTGGTCAAACTAGTGCCAGCTGTGACTGTGCAGCGCTGT
 144 GGTGGCTGCCCTGACGATGGCCCTGGAAATGTGCCCAGTGGCAACCAAGTCCGAAATGCAGA
 211 TCCTCATGATCCAGTACCCGAGCAGTGGGGAGATGTCCCTGGAAGAACAACAGCCAATGTGA
 278 ATG CAG ACC AAA AAA AAG GAG AGT GCT GTG AAG CCA GAC AGC CCC AGG
 Ile Leu Cys Pro Cys Thr Gln Arg Arg Gln Arg Pro Asp Pro Arg Thr
 330 ATC CTC TGC CCG CCT TGC ACC CAG CGC CGT CAA CGC CCT GAC CCC CGG ACC
 Cys Arg Cys Arg Cys Arg Arg Phe Leu His Cys Gln Gly Arg Gly
 381 TGC CGC TGC CGC TGC AGA CGC CGC TTC CTC CAT TGC CAA GGG CGG GGC
 Leu Glu Leu Asn Pro Asp Thr Cys Arg Cys Arg Lys Pro Arg Lys Stop
 432 TTA GAG CTC AAC CCA GAC ACC TGT AGG TGC CGG AAG CCG CGA AAG TGA CAA
 483 GCTGCTTCCAGACTCCACGGGCCGGCTGCTTTATGGCCCTGCTTACAGGGACGGAAGAGTGGAG
 550 CACAGGCAAACTCCTCAGTCTGGGAGGTCACCTGCCCAGGACCTGGACCTTTAGAGAGCTCTC
 617 GCCATCTTTATCTCCAGAGCTGCCATCTAACAAATGTCAAGGAACCTCATGTCTCACCTCAGGGG
 684 CCAGGTACTCTCTCACTTAACCCCTGGTCAAGTGAGCATCTTCTGGCTGGCTGTCCCTCAC
 751 TATGAAAACCCCAACTTCTACCAATAACGGGATTTGGGTTCTGTATGATAACTGTGACACACACA
 816 CACACTCACACTCTGATAAAAGAGAACTCTGATAAAAGAGATGGAAAGACACTAAATAAAAAA
 885 AAA

7
 24
 41
 56

(SEQ ID NOS: 1 & 3)

FIG. 2

GAGCCCCCTG CTCCTGTCGCC TGCTGCTTGT TGCAC TGCTG CAGCTGGCTC
 GCACCCAGGC CCTGTGTCC CAGTTGATG GCCCCAGCCA CCAGAGAAA
 GTGGTGCCAT GGATAGACGT TTATGCACGT GCCACATGCC AGCCAGGGA
 GGTGGTGGTG CCTCTGAGCA TGGAACTCAT GGGCAATGTG GTCAAACAAC
 TAGTGCCCAG CTGTGTGACT GTGCAGCGCT GTGGTGGCTG CTGCCCTGAC
 GATGGCCTGG AATGTGTGCC CACTGGCAA CACCAAGTCC GAATGCAGAT
 CCTCATGATC CAGTACCCGA GCAGTCAGCT GGGGAGATG TCCCTGGAAG
 AACACAGCCA ATGTGAATGC AGACCAAAA AAAAGGAGAG TGCTGTGAAG
 CCAGACAGCC CCAGGATCCT CTGCCCGCCT TGCACCCAGC GCCGTCAACG
 CCTGACCCC CGGACCTGCC GCTGCCGCTG CAGACGCCGC CGCTTCCTCC
 ATTGCCAAGG GCGGGCTTA GAGCTCAACC CAGACACCTG TAGGTGCCGG
 AAGCCGCGAA AGTGA
 (SEQ ID NO: 4)

FIG. 3

MSPLLRLLLL VALLQLARTQ APVSQFDGPS HQKKVVPWID VYARATCQPR
 EVVPLSMEL MGNVVKQLVP SCVTVQRCGG CCPDDGLECV PTGQHQVRMQ
 ILMIQYSSQ LGMSLEEHS QCECRPKKE SAVKPSPRI LCPPCTQRRQ
 RPDPRTCRCR CRRRRFLHCQ GRGLELNPDY CRCRKRK
 (SEQ ID NO: 5)

FIG. 4

ACCATGAGCC CCCTGCTCCG TCGCCTGCTG CTTGTTGCAC TGCTGCAGCT
 GGCTCGCACC CAGGCCCTG TGTCCAGTT TGATGGCCCC AGCCACCAGA
 AGAAAGTGGT GCCATGGATA GACGTTATG CACGTGCCAC ATGCCAGCCC
 AGGAGGTGG TGGTGCCCTCT GAGCATGGAA CTCATGGGCA ATGTGGTCAA
 ACAACTAGTG CCCAGCTGTG TGACTGTGCA GCGTGTGGT GGCTGCTGCC
 CTGACGATGG CCTGGAATGT GTGCCCACTG GGCAACACCA AGTCCGAATG
 CAGGTACCAG GGCCTATGG TCAGATCCTC ATGATCCAGT ACCCGAGCAG
 TCAGCTGGGG GAGATGTCCC TGGAGAACA CAGCCAATGT GAATGCAGAC
 CAAAAAAA GGAGAGTGCT GTGAAGCCAG ACAGCCCCAG GATCCTCTGC
 CCGCCTTGCA CCCAGCGCCG TCAACGCCCT GACCCCCGA CCTGCCGCTG
 CCGCTGCAGA CGCCGCCGCT TCCTCCATTG CCAAGGGCGG GGCTTAGAGC
 TCAACCCAGA CACCTGTAGG TGCCGGAAGC CGCGAAAGTG A
 (SEQ ID NO: 6)

4/22

FIG. 5

MSPLLRLLLL VALLQLARTQ APVSQFDGPS HQKKVVPWID VYARATCQPR
 EVVPLSMEL MGNVVKQLVP SCVTVQRCGG CCPDDGLECV PTGQHQVRMQ
VGPMGQILM IQYPSSQLGE MSL EHSQCE CRPKKESAV KPDSPRILCP
 PCTQRRQRPD PRTCRCRR RRFLHCQGRG LELNPDTCRC RKPRK
 (SEQ ID NO: 7)

FIG. 6

ACCATGAGCC CCCTGCTCCG TCGCCTGCTG CTTGTTCAC TGCAGCCT
 GGCTCGCACC CAGGCCCTG TGTCCAGTT TGATGGCCCC AGCCACAGA
 AGAAAGTGGT GCCATGGATA GACGTTATG CACGTGCCAC ATGCCAGCCC
 AGGGAGGTGG TGGTGCCTCT GAGCATGGAA CTCATGGGCA ATGTGGTCAA
 ACAACTAGTG CCAGCTGTG TGA CTGTGGT GCGCTGTGGT GGTGTGCC
 CTGACGATGG CCTGGAATGT GTGCCACTG GGCACACCA AGTCCGAATG
 CAGATCCTCA TGATCCAGTA CCCGAGCAGT CAGCTGGGG AGATGTCCCT
 GGAAGAACAC AGCCAATGTG AATGCAGACC AAAAAAAAAA AGGAGAGTGC
 TGTGA

(SEQ ID NO: 8)

FIG. 7

MSPLLRRLLL VALLQLARTQ APVSQFDGPS HQKKVVPWID VYARATCQPR
 EVVPLSMEL MGNVVKQLVP SCVTVQRCGG CCPDDGLECV PTGQHQRMQ
 ILMIQYPSQ LGEMSLEHS QCECRPKKR RVL
 (SEQ ID NO: 9)

FIG. 8

mVEGF-B 167
mVEGF 164
hPIGF
mPDGF A
mPDGF B

M N R T W A C L L L G C Y L A H A L A E A E A E
M N R C W A L F L P L C C Y L R L V S A E D P I P E E L
- -

mVEGF-B 167
mVEGF 164
hPIGF
mPDGF A
mPDGF B

M S P L L R R L L
L S W V H W T L A L L
L M R L F P C F L L L
L V M R L S V G A E L L
L P V E I D S V D E G A L L
L H R D S V D E G A L L
M N F L L M R I D S V D E G A L L
M - R L L Q R L L H R D S V D E G A L L
I E R L A R S Q I R S F I R D L Q R L L H R D S V D E G A L L
E M L S D H S I R S F I R D L Q R L L H R D S V D E G A L L

mVEGF-B 167
mVEGF 164
hPIGF
mPDGF A
mPDGF B

L V A L L Q L A R T Q A P V S Q F D G P S H Q K K V V P W I
L L Y L H H A K W S Q A P - T T E G E Q K S H E V I K F M
L A G L L A L P A Q W A L S - A G N G S S E R K E V I K F M
E T S L R A H G S P H A I N V P E K R P V R S L G S L A A E
D L N M T R A H S G V E L E S S R G R R S L G S L A A E

mVEGF-B 167
mVEGF 164
hPIGF
mPDGF A
mPDGF B

D V Y A R A T
D V Y Q R S Y
F Q E V W G R S
E A I P A V
P A V I A E

C Q P R E V V V P L S M E L M G N V V K Q
C R P I E T L V D I F Q E Y P D E I E Y I
C R A L E R L V D V S E Y P S E V E H M
C K T R T V I Y E I P R S Q V D P T S A N
C K T R T E V F Q I S R N L I D R T N A N

FIG. 9A

mVEGF-B 167
 mVEGF 164
 hPIGF
 mPDGF A
 mPDGF B

L - - V P S C V T V Q R C G G C C C C C C C C C C C C C C
 E - - K P S C V V P L M R R C A G G C C C C C C C C C C C C
 F - - S P S C V V S L L R R C A T G G C C C C C C C C C C C C
 F I W P P P C V V S L L R R C V K T S G G C C C C C C C C C C
 F L V W P P C V V S E V V Q R C T S G G C C C C C C C C C C C C

mVEGF-B 167
 mVEGF 164
 hPIGF
 mPDGF A
 mPDGF B

Q V R M Q I L M I Q Y P S S Q
 N I T M Q I M R I K P H Q S Q
 N V T M Q L L K I R S G D R K P
 H R S V K V A K V E Y V R K K P
 M R P V Q V R K I E I V R K K P

mVEGF-B 167
 mVEGF 164
 hPIGF
 mPDGF A
 mPDGF B

C E C C C A C C R P K K K E S A V K P D S P R I L C P P C T Q R R Q R
 C E C C C R R P K K D R T - - K P E N H - - C E P C S E R R K H
 C E C C R R P L R E K M - - K P E - - R C G D A V P R R
 C A C C A T S N L N P D H R E E T D V R
 C K C E T I V T P R P V T R S R E Q R A K T P Q A R

mVEGF-B 167
 mVEGF 164
 hPIGF
 mPDGF A
 mPDGF B

- - - P D P R T C R C R C R R R R F L H C Q G R G L E L N P
 L F V Q D P Q T C K C S C K N T D - S R C K A R Q L E L N E

V T I R T V R I R R P P K G K H R K F K H T H D K A A L K E

mVEGF-B 167
 mVEGF 164
 hPIGF
 mPDGF A
 mPDGF B

D T C R C R K P R K
 R T C R C D K P R R

T L G A

FIG. 9B

ACCATGAGCC CTCGTCTCCG CCGCCTGCTG CTCGCCGCAC TCCTGCAGCT
 GGCCCCCGCC CAGGCCCTG TCTCCAGCC TGATGCCCTT GGCCACCAGA
 GGAAAGTGGT GTCATGGATA GATGTGATA CTCGCGCTAC CTGCCAGCCC
 CGGGAGGTGG TGGTGCCCTT GACTGTGGAG CTCATGGGCA CCGTGGCCAA
 ACAGCTGGTG CCCAGCTGCG TGA CTGTGCA GCGCTGTGGT GGCTGCTGCC
 CTGACGATGG CCTGGAGTGT GTGCCACTG GGCAGCACCA AGTCCGGATG
 CAGATCCTCA TGATCCGGTA CCCGAGCAGT CAGCTGGGG AGATGTCCCT
 GGAAGAACAC AGCCAGTGT AATGCAGACC TAAATAAAG GACAGTGTG
 TGAAGCCAGA CAGCCCAGG CCCCTCTGCC CACGCTGCAC CCAGCACCCAC
 CAGCGCCCTG ACCCCCAGC CTGCCGCTGC CGCTGCCGAC GCCGCAGCTT
 CCTCCGTTGC CAAGGGCGGG GCTTAGAGCT CAACCCAGAC ACCTGCAGGT
 GCCGGAAGCT GCGAAGGTGA
 (SEQ ID NO: 10)

8/22

FIG. 10

MSPLLRLLLL AALLQLAPAQ APVSQPDAPG HQRKVVSWID VYTRATCQPR
 EVVVPLTVEL MGTVAKQLVP SCVTVQRCGG CCPDDGLECV PTGQHQVRMQ
 ILMIRYSSQ LGEMSLEHS QCECRPKKD SAVKPDSPRP LCPRCTQHHQ
 RPDPRTCRCR CRRRSFLRCQ GRGLELNPDT CRCRKLRR
 (SEQ ID NO: 11)

FIG. 11

ATGAGCCCCC TGCTCCGTCG CCTGCTGCTT GTTGCACTGC TGCAGCTGGC
 TCGCACCCAG GCCCCTGTGT CCCAGTTTGA TGGCCCCAGC CACCAGAAGA
 AAGTGGTGCC ATGGATAGAC GTTTATGCAC GTGCCACATG CCAGCCCAGG
 GAGGTGGTGG TGCCCTCTGAG CATGGAATC ATGGGCAATG TGGTCAAACA
 ACTAGTGCCC AGCTGTGTGA CTGTGCAGCG CTGTGGTGGC TGTGCCCTG
 ACGATGGCCT GGAATGTGTG CCCACTGGGC AACACCAAGT CCGAATGCAG
 ATCCTCATGA TCCAGTACCC GAGCAGTCAG CTGGGGAGA TGTCCCTGGA
 AGAACACAGC CAATGTGAAT GCAGACCAAA AAAAAGGAG AGTGCTGTGA
 AGCCAGACAG GGTGCCATA CCCACACC GTCCCAGCC CCGCTCTGTT
 CCGGGCTGGG ACTCTACCC GGGAGCATCC TCCCAGCTG ACATCATCCA
 TCCCACCTCCA GCCCCAGGAT CCTCTGCCCG CCTTGCACCC AGCGCCGTCA
 ACGCCCTGAC CCCCGGACCT GCCGCTGCCG CTGCAGACGC CGCCGCTTCC
 TCCATTGCCA AGGGCGGGC TTAG
 (SEQ ID NO:12)

FIG. 12

9/22

MSPLLRLLLL VALLQLARTQ APVSQFDGPS HQKKVVPWID VYARATCQPR
 EVVVP LSMEL MGNVVKQLVP SCVTVQRCGG CCPDDGLECV PTGQHQVRMQ
 ILMIQYSSQ LGEMSLEEHS QCECRPKKE SAVKPDRAI PHHRPQPRSV
 PGWDSTPGAS SPADIIHPTP APGSSARLAP SAVNALTPGP AAAAADAAAS
 SIAKGA
 (SEQ ID NO:13)

FIG. 13

ATGAGCCCTC TGCTCCGCCG CCTGCTGCTC GCCGCACTCC TGCAGCTGGC
 CCCCGCCAG GCCCCTGTCT CCCAGCCTGA TGCCCTGGC CACCAGAGGA
 AAGTGGTGC ATGGATAGAT GTGTACTC GCGTACCTG CCAGCCCCTGG
 GAGTGGTGG TGCCCTTGAC TGTGAGCTC ATGGCACC GGGCCAAACA
 GCTGGTGCC AGCTGCGTGA CTGTGAGCG CTGTGGTGG TGCTGCCCTG
 ACGATGGCCT GGAGTGTGT CCCCCTGGC AGCACCAGT CCGATGCAG
 ATCCTCATGA TCCGGTACCC GAGCAGTCAG CTGGGGGAGA TGTCCCTGGA
 AGAACACAGC CAGTGTGAAT GCAGACCTAA AAAAAAGGAC AGTGTGTGA
 AGCCAGACAG GGCTGCCACT CCCCACACC GTCCCAGCC CCGTCTGT
 CCGGGCTGG ACTCTGCCCC CGGAGCACCC TCCCAGCTG ACATCACCCA
 TCCCACCTCA GCCCAGGCC CCTCTGCCCA CGCTGCACC AGCACCACCA
 GCGCCCTGAC CCCCAGACCT GCCGCCGCC CTGCCGACGC CGCAGCTTCC
 TCCGTTGCCA AGGGCGGGC TTAG
 (SEQ ID NO:14)

10/22

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FIG. 14

MSPLLRLLL AALLQLAPAQ APVSQPDAPG HQRKVSWID VYTRATCQPR
 EVVPLTVEL MGTVAKQLVP SCVTVQRCGG CCPDDGLECV PTGQHQVRMQ
 ILMIRYPSSQ LGEMSLEEHS QCECRPKKD SAVKPDRAAT PHHRPQPRSV
 PGWDSAPGAP SPADITHPTP APGPSAHAAP STTSALTGP AAAAAAAS
 SVAKGGA
 (SEQ ID NO:15)

FIG. 15



M	S	P	L	L	R	R	L	L	V	A	L	L	Q	L	A	R	T	Q	A	P	V	S	Q	F	D	G	P	S
M	S	P	L	L	R	R	L	L	A	A	L	L	Q	L	A	P	A	Q	A	P	V	S	Q	P	D	A	P	G
M	S	P	L	L	R	R	L	L	V	A	L	L	Q	L	A	R	T	Q	A	P	V	S	Q	F	D	G	P	S
M	S	P	L	L	R	R	L	L	A	A	L	L	Q	L	A	P	A	Q	A	P	V	S	Q	P	D	A	P	G

mVEGF-B₁₈₆
hVEGF-B₁₈₆
mVEGF-B₁₆₇
hVEGF-B₁₆₇

H	Q	K	K	V	V	P	W	I	D	V	Y	A	R	A	T	C	Q	P	R	E	V	V	V	P	L	S	M	E	L
H	Q	R	K	V	V	S	W	I	D	V	Y	T	R	A	T	C	Q	P	R	E	V	V	V	P	L	T	V	E	L
H	Q	K	K	V	V	P	W	I	D	V	Y	A	R	A	T	C	Q	P	R	E	V	V	V	P	L	S	M	E	L
H	Q	R	K	V	V	S	W	I	D	V	Y	T	R	A	T	C	Q	P	R	E	V	V	V	P	L	T	V	E	L

mVEGF-B₁₈₆
hVEGF-B₁₈₆
mVEGF-B₁₆₇
hVEGF-B₁₆₇

M	G	N	V	V	K	Q	L	V	P	S	C	V	T	V	Q	R	C	G	G	C	C	P	D	D	G	L	E	C	V
M	G	T	V	A	K	Q	L	V	P	S	C	V	T	V	Q	R	C	G	G	C	C	P	D	D	G	L	E	C	V
M	G	N	V	V	K	Q	L	V	P	S	C	V	T	V	Q	R	C	G	G	C	C	P	D	D	G	L	E	C	V
M	G	T	V	A	K	Q	L	V	P	S	C	V	T	V	Q	R	C	G	G	C	C	P	D	D	G	L	E	C	V

mVEGF-B₁₈₆
hVEGF-B₁₈₆
mVEGF-B₁₆₇
hVEGF-B₁₆₇

P	T	G	Q	H	Q	V	R	M	Q	I	L	M	I	Q	Y	P	S	S	Q	L	G	E	M	S	L	E	E	H	S
P	T	G	Q	H	Q	V	R	M	Q	I	L	M	I	R	Y	P	S	S	Q	L	G	E	M	S	L	E	E	H	S
P	T	G	Q	H	Q	V	R	M	Q	I	L	M	I	Q	Y	P	S	S	Q	L	G	E	M	S	L	E	E	H	S
P	T	G	Q	H	Q	V	R	M	Q	I	L	M	I	R	Y	P	S	S	Q	L	G	E	M	S	L	E	E	H	S

mVEGF-B₁₈₆
hVEGF-B₁₈₆
mVEGF-B₁₆₇
hVEGF-B₁₆₇

FIG. 16A

mVEGF-B₁₈₆ Q C E C R P K K K E S A V K P D R V A I P H H R P Q P R S V
hVEGF-B₁₈₆ Q C E C R P K K K D S A V K P D R A A T P H H R P Q P R S V
mVEGF-B₁₆₇ Q C E C R P K K K E S A V K P D S P R I L C P P C T Q R R Q
hVEGF-B₁₆₇ Q C E C R P K K K D S A V K P D S P R P L C P R C T Q H H Q

mVEGF-B₁₈₆ P G W D S T P G A S S P A D I I H P T P A P G S S A R L A P
hVEGF-B₁₈₆ P G W D S A P G A P S P A D I T H P T P A P G P S A H A A P
mVEGF-B₁₆₇ R P D P R T C R C R C R R R F L H C Q G R G L E L N P D T
hVEGF-B₁₆₇ R P D P R T C R C R C R R R S F L R C Q G R G L E L N P D T

mVEGF-B₁₈₆ S A V N A L T P G P A A A A D A A S S I A K G G A
hVEGF-B₁₈₆ S T T S A L T P G P A A A A D A A S S V A K G G A
mVEGF-B₁₆₇ C R C R K P R K
hVEGF-B₁₆₇ C R C R K L R R

FIG. 16B

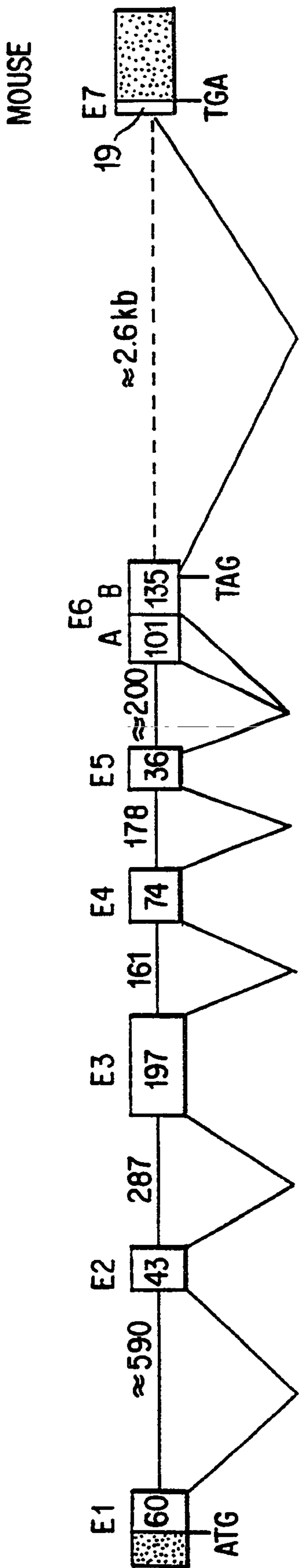


FIG.17

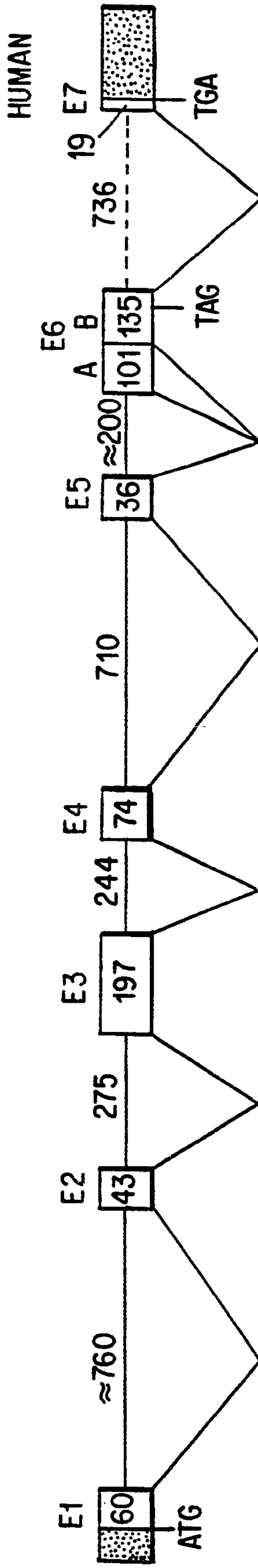


FIG. 17A

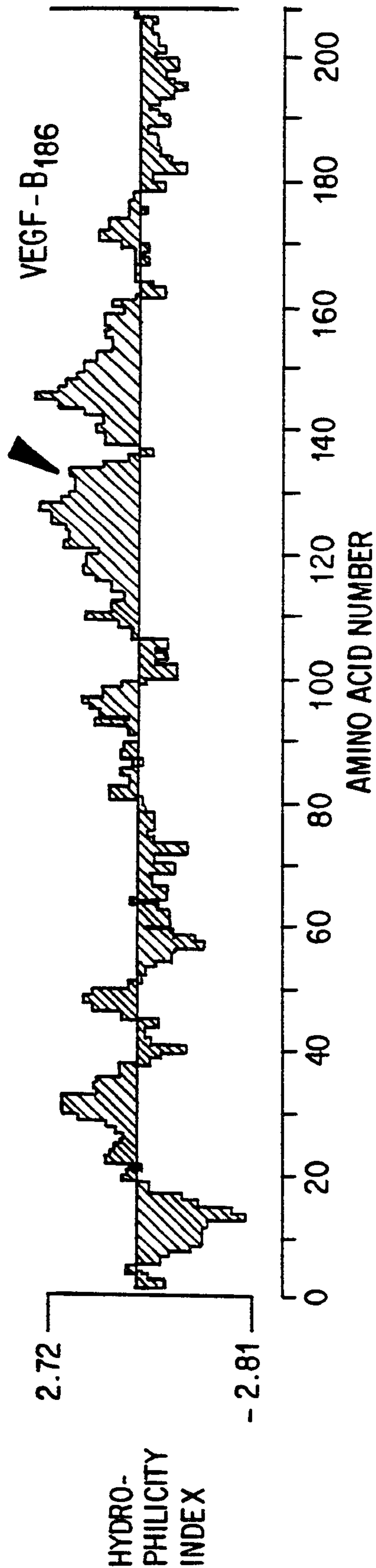


FIG. 18

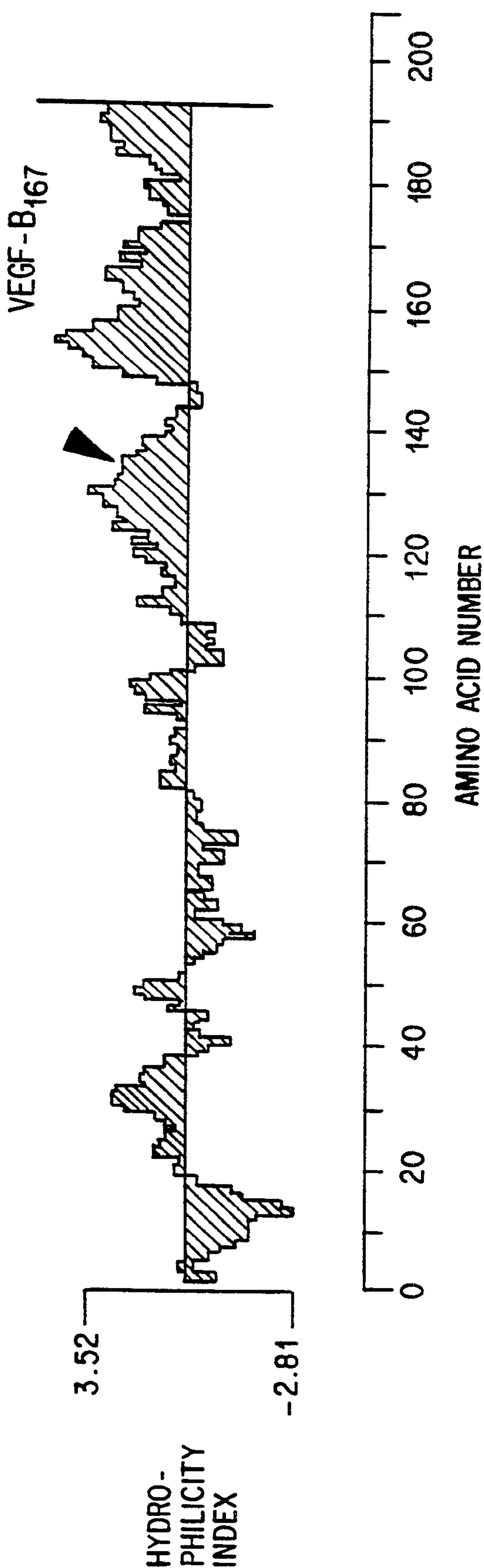


FIG. 18A

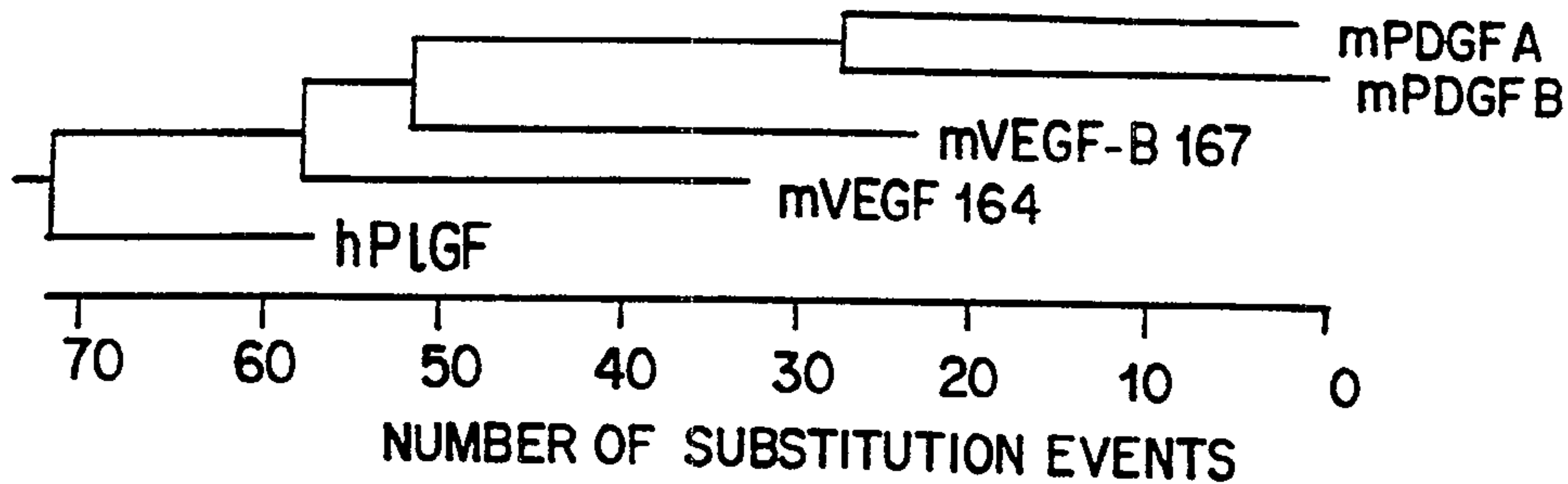


FIG. 19

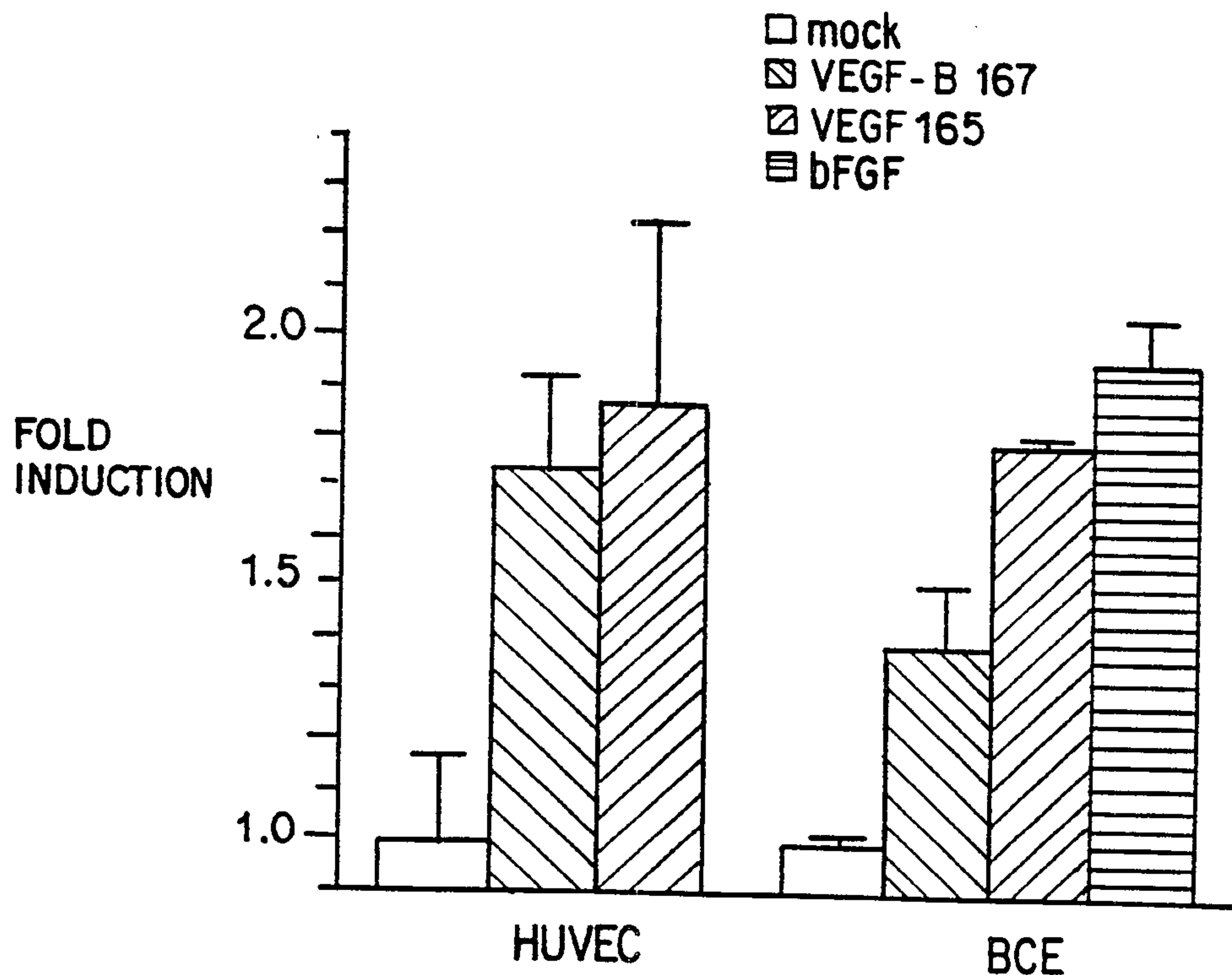


FIG. 20

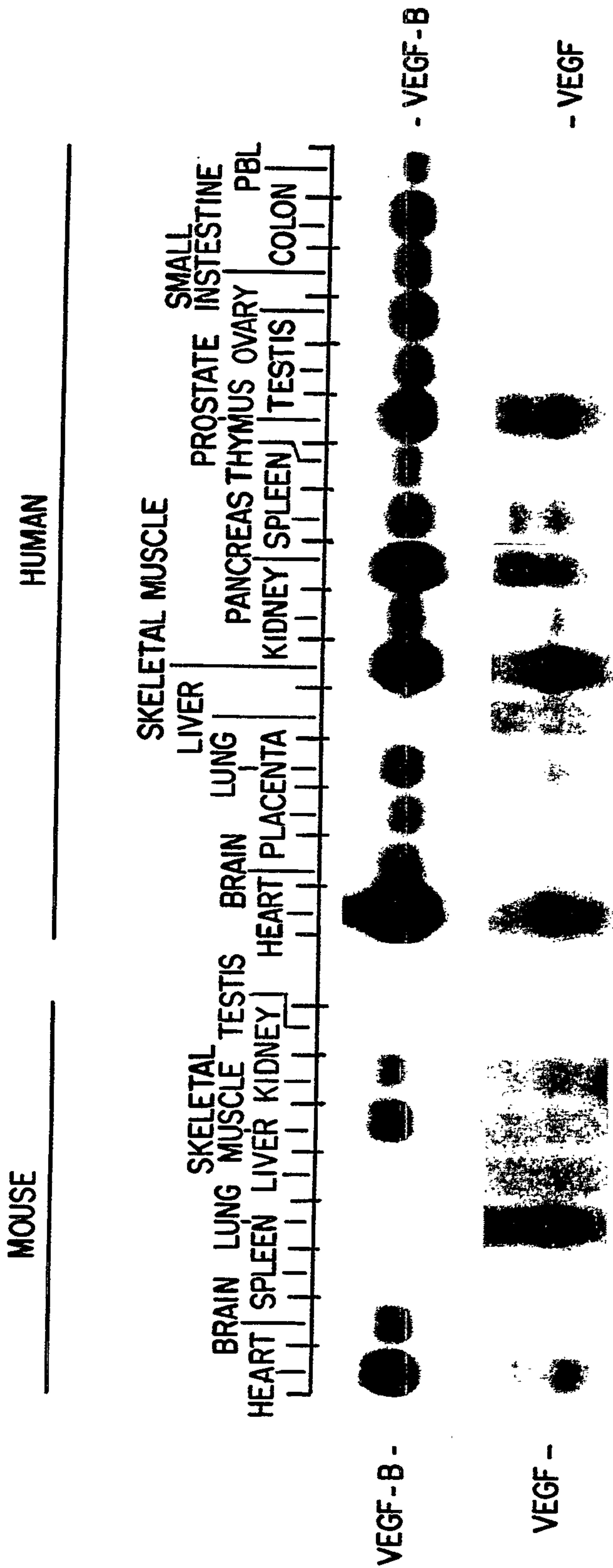


FIG. 21

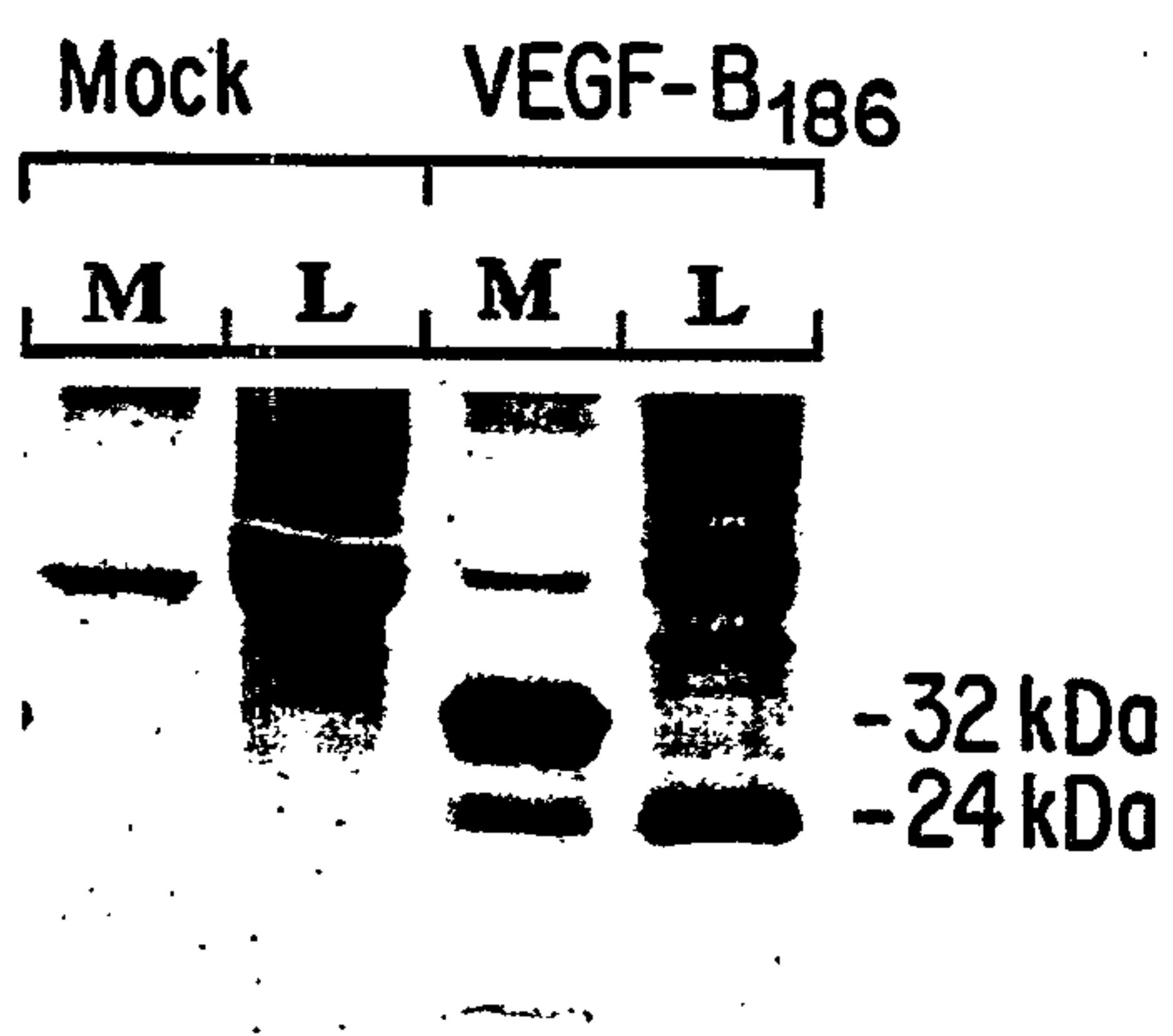


FIG. 22

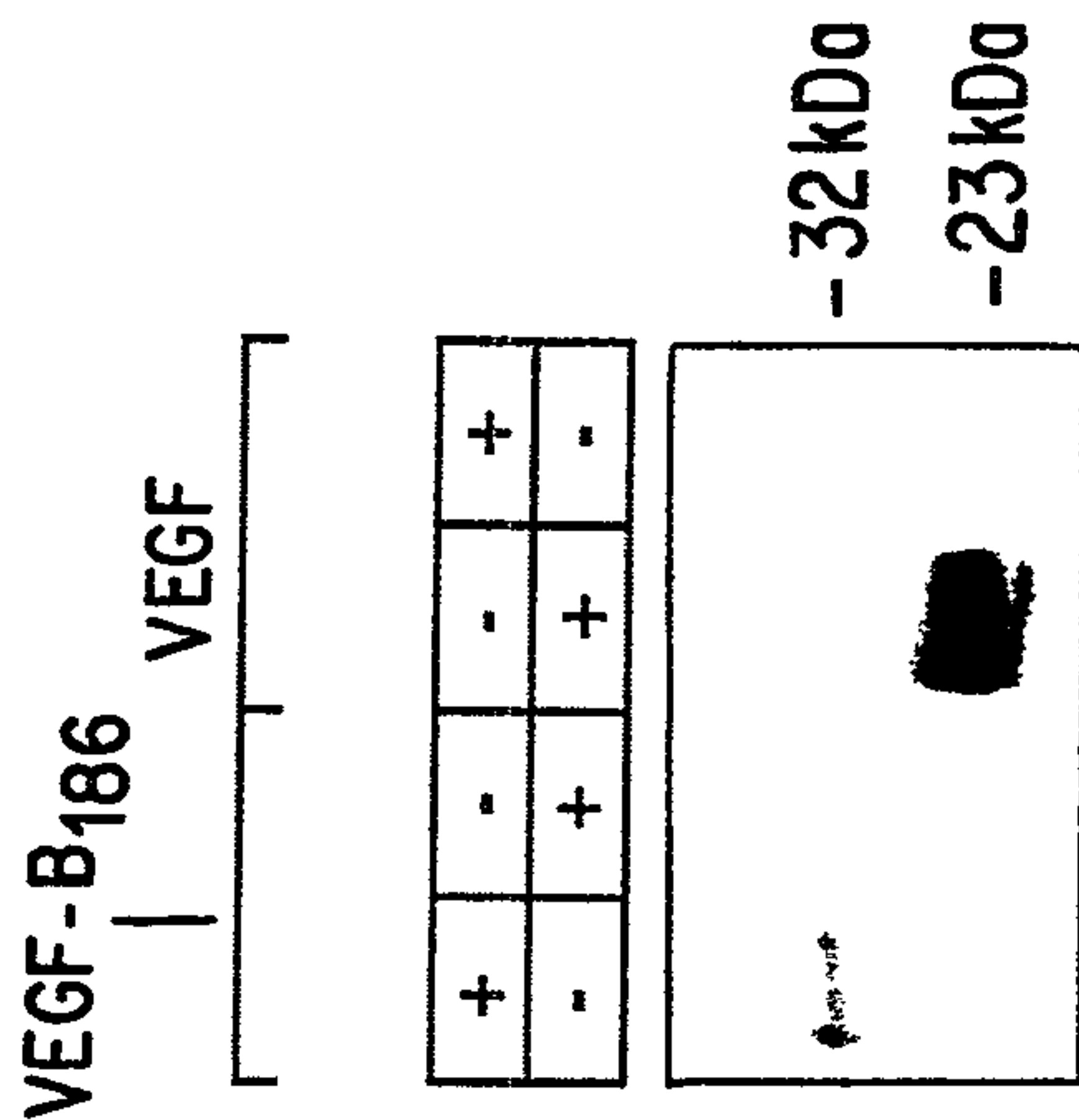


FIG. 23A



FIG. 23B

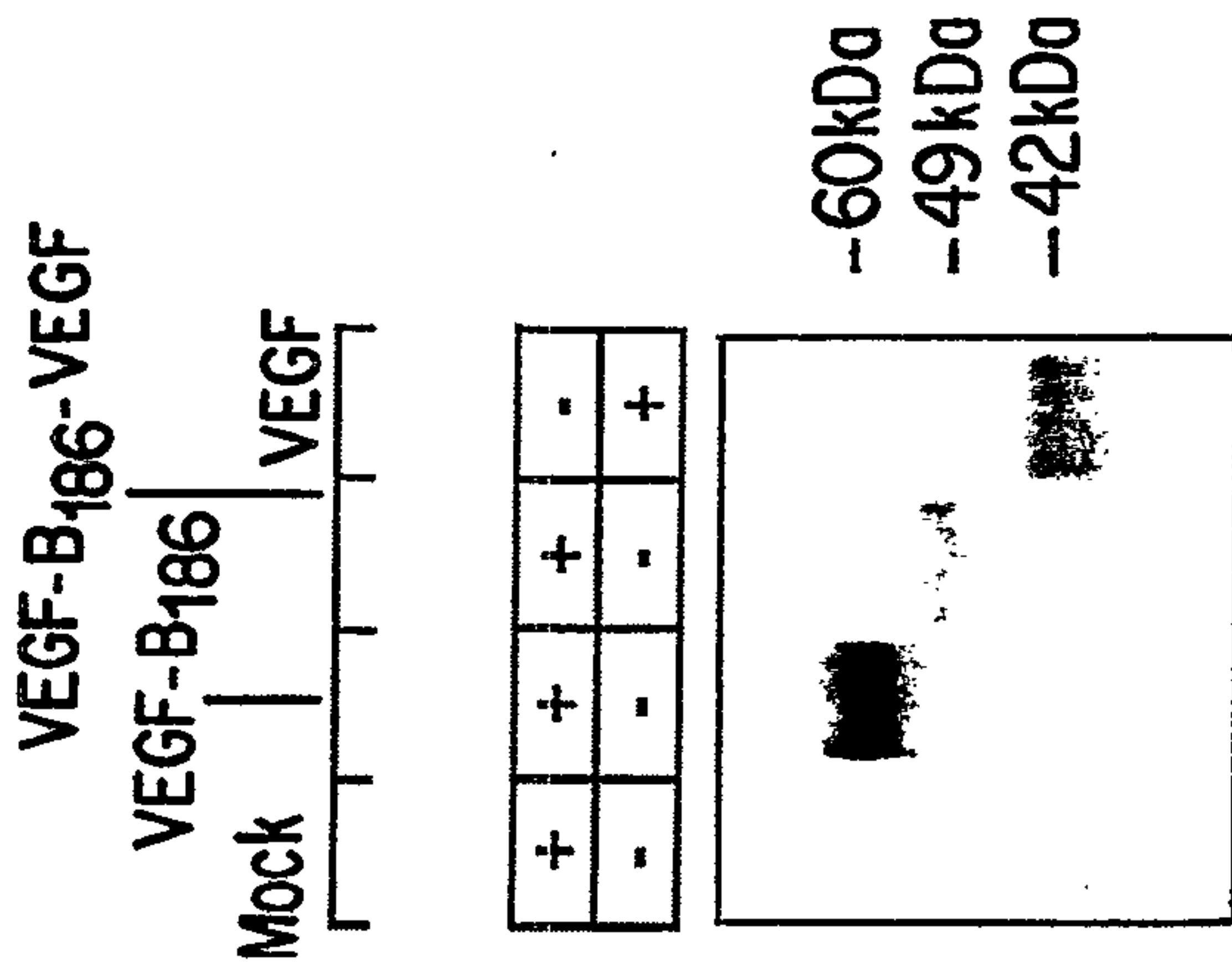


FIG. 23C

Antibody to

VEGF-B
VEGF

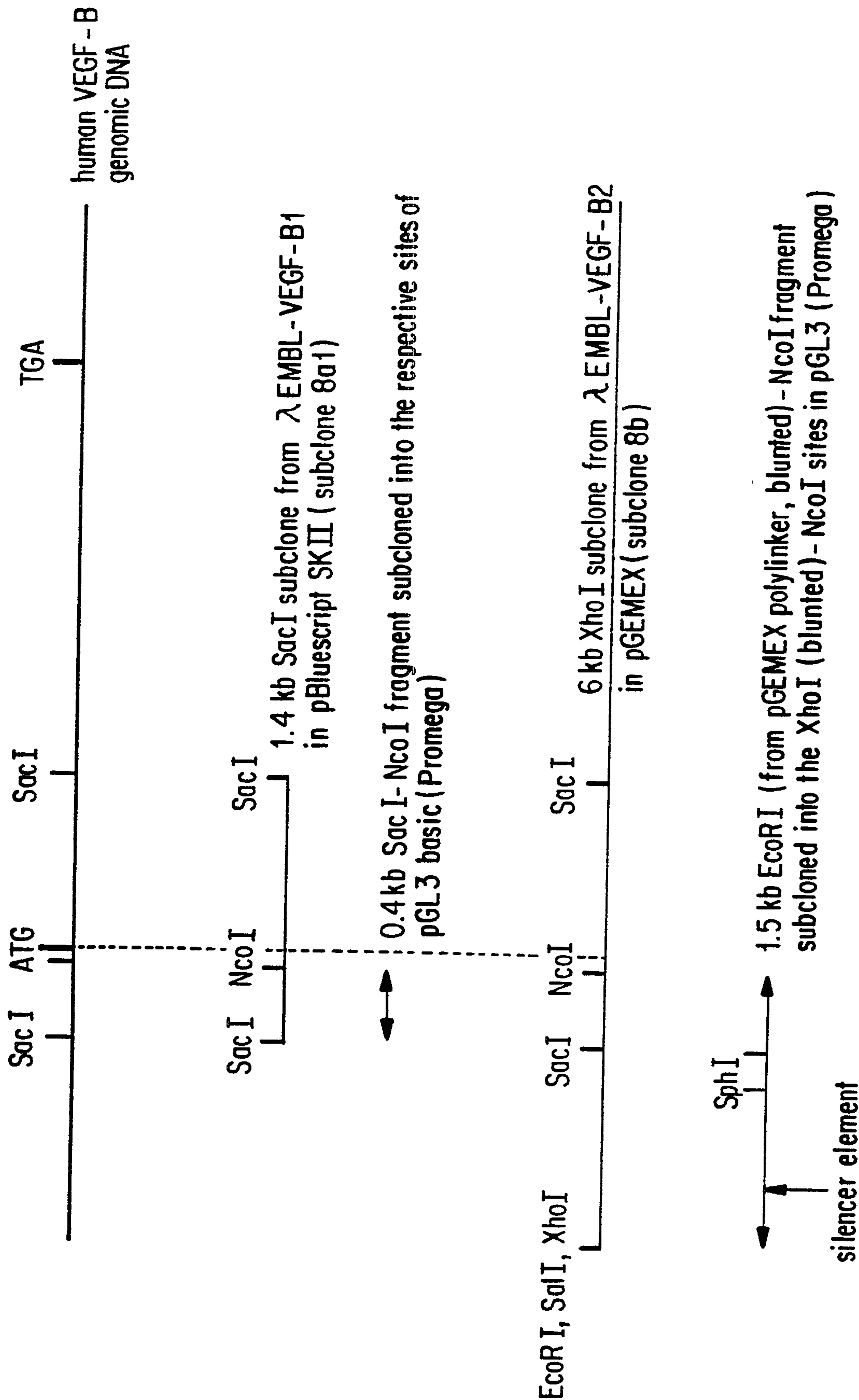


FIG. 24

1 ctcgagatct gtttgttgtc ttggaacaat acggtttaga ggtgactggc
Xho I
51 gggtagcgag aacatatgcg agttcaccta agagaaaagc tgaatgaggc
101 aatgcctctt cctgaccata tctcttactc agataactat agaatttatt
151 gtccagtaaa gggta^{tatta} ^{aaa}aatc^{ata} ^{ta} t^{taaa}agtca tacagtgaag
201 ttgtccaggg aatcaagac ^{silencer binding site 2} t^{aacag}t^{ct} cactctgaca ataatgaaca
251 ggggattcc ctcaagatag actaggacat gaccacacac tggcaggtag
301 tagtaccaga aaagaacgca tggaaaatct ttaccttatg cttgaggtag
351 ggaccaggct aaagtgaagg ccagacctaa aattctatct aaaataaadc
401 cacaatcгаа gaaaatatgt ggtgtacagg tatagaatgt ctttactgga
451 tcattgaaat agtaagataa attcaacttt ttacattggt ttcttttctt
501 ccagttaggg cttgagacct tcgtctctgg agagtgactg tcaattggag
551 ccctgcttcc tgggtttctg gccagggggg ttgtggatgc ttaacatgtg
601 cctttcacag gacacttctt taccacagca gtggccangt gtgcatccca

651 cgaccaggcc tccctctcac ggaacatctg ttgagactag gagatgcctg
701 gtgactgttg cctgacctgt gtctgtgta ttctgacaa gagccactct
751 caagaccct ggccaggagg agagttaggt tccagtgtag gtcagctcag
801 acagatggag gccacagaan caaacatggg aatcacaga agtaggttta
851 ttactcacag atccctatcc caaccacca ggtgccctct cctccagggc
901 caacagaggc atccttcagc aggagcaca acggctaggg cagcggcaag
951 ccgccaccat ccgagccaac ccaggccccg agatcgtgcc ccggggcgcc
1001 ggccccctgag gggctcacct ggatggggcc tgcatgcgtt cccgctttgc
1051 ttccttccct ggacggccccg ctccccgaa ^{Sph I} acgcgccgcc aataaagtga
1101 ttcgcagagc tcgtgtgcgg ctccctcctt aaggcccgac gccccggcc
1151 ccggcctcgc ^{SacI} caagggcagc gccccggcct ccgggtagtg gcggccggcg
1201 actggggagc ccagcctcct gggcgggtgcg tccccctccc cctgccgcgg
1251 cgggaggcgg gagggggtgt gtggaggagg cgggccccgc cgacggcctc

FIG. 25B

```
1301 gccccccac cccgccgccc cgcccccgcc ccacgggccc ggtggggagc
1351 gcgtgtctgg gtcacatgag ccgcctgccc gccagcccgg gccagcccc
1401 ccgccgccc cgccgtccc gccgccgctg ccgcggcca ccggccgccc
1451 gccgcccggt ctctccggc cgcctcgct gcgctgcntg cgctgcctgc
1501 acccagggt cggaggagg ccgcggagg gccgcccccc gcgccccggc
... Nco I site is about 30 bp downstream) (SEQ ID NO:17)
```

FIG. 25C