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 Vectors for Molecule Delivery to CD11b Expressing Cells

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ABSTRACT

5 The invention relates to a novel use of a *Bordetella* adenylylase toxin in the manufacturing of vectors for targeting *in vivo* a molecule of interest, specifically to CD11b expressing cells. The invention also relates to an immunogenic composition that primes immune responses, to pharmaceutical compositions and to a new vector for molecule delivery to CD11b expressing cells.

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**COMPLETE SPECIFICATION
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Invention Title: Vectors for Molecule Delivery to CD11b
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The following statement is a full description of the invention, including the best method of performing it, known to us:

BACKGROUND OF THE INVENTION

The invention relates to a novel use of a *Bordetella* adenylcyclase toxin in the manufacturing of vectors for targeting *in vivo* a molecule of interest, specifically to CD11b expressing cells. The invention also relates to an immunogenic composition that primes immune responses, to pharmaceutical compositions and a new vector for molecule delivery to CD11b expressing cells.

Bordetella pertussis, the causative agent of whooping cough, secretes several toxins including the well-known pertussis toxin (PT) and the adenylate cyclase toxin (CyaA) or also adenylcyclase. CyaA is a critical virulence factor of *B. pertussis* in the murine respiratory model that is required for the early steps of lung colonization. Indeed, genetic deletion of this toxin dramatically decreases the pathological effects of *B. pertussis* infection, reducing the number of bacteria recovered from the lung and almost abolishing the inflammatory cell recruitment and the lung lesions observed after infection [Weiss *et al.*, 1984; Weiss *et al.*, 1989; Gross *et al.*, 1992; Khelef *et al.*, 1992; Khelef *et al.*, 1994; Gueirard *et al.*, 1998]. Moreover, CyaA is an antigen protective against *B. pertussis* infection in the murine respiratory model [Guiso *et al.*, 1989; Guiso *et al.*, 1991].

Originally discovered by Hewlett *et al* in *B. pertussis* culture supernatants [Hewlett *et al.*, 1976], the adenylcyclase was later found to be activated by the eukaryotic calmodulin [Wolff *et al.*, 1980]. This striking feature quickly found a rationale when it was shown by Confer and Eaton that the adenylcyclase could enter into eukaryotic cells where, upon activation by calmodulin, it could trigger a large increase in cAMP within these target cells [Confer *et al.*, 1982].

Adenylcyclase is encoded by the *cyaA* gene, and its expression, like that of other virulence genes of *B. pertussis*, is coordinately regulated by environmental signals. The *cyaA* gene is part of an operon that also contains genes *cya B*, *D* and *E*, that are required for CyaA secretion [Ladant *et al.*, 1999].

The CyaA toxin is a bifunctional protein of 1706 residues that is made of a N-terminal catalytic domain of 400 amino acids and a C-terminal part of 1306 residues which is responsible for the binding of the toxin to target cell membrane and the subsequent delivery of the catalytic moiety into the cell cytosol [Sakamoto *et al.*, 1992] [Ladant *et al.*, 1999]. This part also exhibits a weak hemolytic activity due to its ability to form cation-selective channels in biological membranes [Benz *et al.*, 1994] [Gray *et al.*, 1998]. This region is homologous to *Escherichia coli* hemolysin and other members of the RTX (Repeat in ToXin) family of bacterial toxins. In particular, it contains a series of glycine and

aspartate-rich nonapeptide repeats that are involved in calcium binding [Rose *et al.*, 1995] [Coote *et al.*, 1992].

The CyaA polypeptide is synthesized as an inactive protoxin that is converted to an active toxin by posttranslational palmitoylation of two internal lysines (lysines 856 and 963). This modification requires the product of an accessory gene, *cyaC*, which is located nearby *cyaA* on *B. pertussis* chromosome.

CyaA has been shown to bind to and invade a variety of cell types including cells lacking membrane traffic like mammalian erythrocytes [Rogel *et al.*, 1992]. This suggested that the catalytic domain of CyaA is directly translocated across the plasma membrane of target cells. The internalization of the catalytic domain into the cell cytosol is calcium and temperature-dependent and depends upon the plasma membrane potential [Rogel *et al.*, 1992] [Karimova *et al.*, 1998] [Otero *et al.*, 1995]. However, the molecular mechanisms by which the toxin transports its N-terminus catalytic domain across the membrane remain largely unknown to date. Furthermore no specific receptor has been reported for CyaA binding.

The physiological consequences of cellular intoxication by CyaA were characterized *in vitro* in phagocytes. Confer and Eaton first showed that the Cya A extracted from *B. pertussis* increases the intracellular cAMP level in neutrophils or macrophages leading to an inhibition of chemotaxis and bactericidal functions such as superoxide generation and phagocytic abilities [Confer *et al.*, 1982]. These activities were later confirmed with purified toxins or with bacterial mutants genetically deleted of CyaA [Pearson *et al.*, 1987; Friedman *et al.*, 1987] [Njamkepo *et al.*, 2000]. On the contrary, and despite significant changes in their cAMP content, the viability of cell lines from non-hematopoietic origin appeared to be unaffected by the CyaA intoxication [Bassinat *et al.*, 2000]. Moreover, the present inventors previously demonstrated that *B. pertussis* CyaA can trigger macrophage apoptosis *in vitro* [Khelef *et al.*, 1993; Khelef *et al.*, 1995] and *in vivo* [Gueirard *et al.*, 1998]. In these models, genetic deletion of CyaA abolished macrophage apoptosis, but not neutrophil death, suggesting that CyaA i) is mainly responsible for macrophage apoptosis, ii) might be responsible for neutrophil apoptosis, but that another factor may also be responsible.

Besides that, *in vivo* studies performed in a murine model of *B. bronchiseptica* infection (the animal homologue of *B. pertussis* whose CyaA is closely related) demonstrated that the major target of *B. bronchiseptica* CyaA toxicity is a GM-CSF-dependent and cyclophosphamide-sensitive population that controls the early steps of infection [Harvill *et al.*, 1999]. These criterions identified neutrophils and possibly other

cells including macrophages or dendritic cells but no data is disclosed or suggested that the CD11b cell receptor is involved in the targeting by Cya A. These populations of target cells for CyaA is the same that limits the early phases of infection and favors the development of an adaptative immune response that controls the latter phases of infection [Harvill *et al.*, 1999].

Unlike other toxins, CyaA has been considered for a long time, as independent of any receptor binding. This is based on the observations that *i)* CyaA can intoxicate a wide variety of model cell lines from various origin [Ladant *et al.*, 1999] *ii)* CyaA binds to Jurkat cells and sheep erythrocytes in a non saturable fashion [Gray *et al.*, 1999]. However, some specificity has been found in respect of cells infected by CyaA. Indeed, *in vivo* studies showed that during murine respiratory infection with *Bordetella* species, CyaA destroyed specifically leukocytes (especially macrophages) without damaging dramatically epithelial cells [Gueirard *et al.*, 1998; Harvill *et al.*, 1999].

It has been proposed in patent application WO 93/21324 to use the recombinant *Bordetella* adenylylase to induce a CD4+ T cell or a CD8+ T cell response; however, since no specific receptor for *Bordetella* adenylylase was identified, it was not known if the antigen presentation was related to uptake by non professional antigen presenting cell followed by cross-priming and presentation by dendritic cells or if the antigen was targeted to professional Antigen Presenting Cells (pAPC).

In line with their surface phenotype, dendritic cells (high expression of MHC I and II, costimulatory and adhesion molecules) represent the most potent APC in many *in vitro* assay for the priming of naive T cells [Bell *et al.*, 1999; Viola *et al.*, 1999]. Other APC like resting naive B cells, for example, could even be tolerogenic since injection of resting, male B cells into female hosts leads to the specific tolerization of male-specific CD8+ T cells [Fuchs *et al.*, 1992]. *In vitro*, naive B cells could delete naive CD8+ T cells via a Fas dependent-mechanism [Bennett *et al.*, 1998].

Moreover, Ag presentation by dendritic cells correlates *in vivo* with the induction of T cell responses. This has been established for MHCII-restricted Ag presentation. Adjuvant-free Ag injection via the intravenous (iv) route usually does not induce T cell priming [Kyburz *et al.*, 1993; Aichele *et al.*, 1994; Aichele *et al.*, 1995] and leads to Ag presentation by non-specific B cells [Guery *et al.*, 1997] [Zhong *et al.*, 1997; Reis e Sousa *et al.*, 1999] and, eventually dendritic cells [Crowley *et al.*, 1990] [Zhong *et al.*, 1997; Reis e Sousa *et al.*, 1999]. In contrast, local immunization strategies like subcutaneous (sc) immunization in the presence of adjuvant usually, induces T cell priming and targeted Ag presentation by Langerhans cell migrating from the skin to the LN draining the

immunization site. In this case, B cells and macrophages are not involved [Guery *et al.*, 1996]. Similar results were obtained after sc or intradermic (id) DNA immunization for MHCII and MHCI -peptide complexes: dendritic cells can be directly transfected at the local site of injection and then migrate to the afferent LN via afferent lymphatics [Condon *et al.*, 1996; Casares *et al.*, 1997; Porgador *et al.*, 1998]. The migration is known as a key event of immunity since mechanical disruption of afferent lymphatics abrogates T cell response to skin sensitizers or skin grafts [Zinkernagel *et al.*, 1997].

Therefore, targeting to dendritic cells is essential for CD4⁺ and CD8⁺ T cell stimulation. Since most antibody responses are dependent upon CD4⁺ T cell help, targeting antigen to dendritic cells is a major goal in vaccination.

Applicants have been interested in studying the presentation of adenylylase of *Bordetella* species by T cell, and have identified a specific receptor molecule present on specific cells, that interacts with CyaA and opens new possibilities for the use of CyaA as a proteinaceous vector for molecules of interest.

Genetically detoxified bacterial toxins represent candidates as vaccine vectors, in particular for T epitope, due to their ability to invade eukaryotic cells (Ladant *et al.*, 1999). However, few proteinaceous vectors were shown to prime CTL responses *in vivo* (Ballard *et al.*, 1996, Cabonette *et al.*, 1999). Moreover, despite numerous *in vitro* promising studies, no vector was shown to be exclusively targeted to pAPC particularly to dendritic cells and more particularly to myeloid dendritic cells.

The inventors have further shown that other cells, especially neutrophils, could be targeted by the vectors of the present invention.

The invention provides means that may at least in part, fulfil these needs and proposes new vectors that would specifically target molecules to determined populations of pAPC for example, to enable stimulating immune response.

Moreover, molecule targeted to the pAPC and specific leukocytes would enable the manufacturing of new vectors useful to deliver biologically active molecule to the proximal environment of these cells. These molecules, for example, could modulate the functional properties of the targeted cells or those involved in the immune response or in the inflammatory response.

Indeed, the inventors have found that *Bordetella pertussis* adenylylase toxin binds specifically with a cellular receptor designated (CD11b/CD18) α_M/α_2 receptor and that this interaction is required for the intracellular delivery of the adenylylase domain to the cytosol of cells and subsequent for cell death. α_M/α_2 (CD11b/CD18) integrin is a dimer of the α_2 integrin family, the expression of these integrins being restricted to leukocytes.

CD11b/CD18 α_M/α_2 displays a pattern of expression in mouse and human, which is restricted to neutrophils/granulocytes, macrophages, dendritic cells, NK cells and subsets of B and T CD8+ lymphocytes (Jeyaseelan *et al.*, 2000, Arnaout *et al.*, 1990).

Therefore, this receptor would represent an ideal target for new vectors, designed in particular for T epitope immunization.

Applicants have shown in the present invention that the *Bordetella* adenylcyclase can be used to target a molecule *in vivo* specifically to CD11b expressing cells.

In particular, Applicants have shown in the present invention that a peptide antigen comprised in the *Bordetella pertussis* adenylcyclase toxin can efficiently be targeted specifically to the surface of dendritic cells, translocated in the cytosol of said dendritic cells and prime a CTL response.

In a specific embodiment, said response is obtained bypassing adjuvant requirement and CD4+ T-cell help.

It has also been shown that genetically modified adenylcyclase can be chemically coupled to a peptide of interest to target said peptide to CD11b expressing cells, especially the cytosol of dendritic cells.

This invention thus provides new efficient immunogenic composition as well as new drug delivery vector to CD11b expressing cells.

DETAILED DESCRIPTION OF THE INVENTION

This invention relates to the use of *Bordetella* adenylcyclase in the manufacturing of a proteinaceous vector for targeting a molecule of interest specifically to CD11b expressing cells.

The invention also relates to the use of a *Bordetella* adenylcyclase wherein said adenylcyclase is recombined with an antigen and especially modified by insertion of a peptide of interest or modified by insertion of a molecule of interest for the preparation of a composition for the targeting of said peptide or molecule specifically to CD11b expressing cells.

The term "specifically" means within the context of the present invention that the adenylcyclase when used as a vector for a molecule of interest, is preferentially directed to CD11b expressing cells, thereby offering means to target the molecule of interest at the surface of said cells or within said cells in a selective way with respect to other cells.

In one embodiment of the invention, the molecule of interest is essentially directed to CD11b expressing cells.

As used herein, the term "CD11b expressing cells" relates to the cells that express the CD11b/CD18 $\alpha_m\beta_2$ receptor on their surface. In particular, these cells are granulocytes/neutrophils, macrophages, NK cells, subsets of T CD8+ and B cells and myeloid dendritic cells.

5 Since CD11b expressing cells and more specifically the myeloid dendritic cells, the neutrophils and the macrophages are involved in essential functions of the immune and innate defence system, in particular in inflammatory and specific immune responses, the invention relates to the manufacturing of a proteinaceous vector or a composition capable of targeting a molecule or a peptide of interest to these CD11b expressing cells especially to myeloid dendritic cells, neutrophils or macrophages.

10 In particular, in one embodiment, the targeting of said molecule or peptide is effective *in vivo*.

The invention thereby provides means appropriate for the design of compositions suitable for administration to animal or human hosts requiring targeting of certain leukocytes and in particular myeloid dendritic cells, neutrophils or macrophages.

15 *Bordetella* adenylcyclase is the calmodulin-dependent adenylcyclase secreted in *Bordetella* species or fragment thereof, said fragment retaining the function properties of the adenylcyclase, a major virulence factor mandatory for the initial phases of bacterial colonization in the lung. The adenylcyclase is synthesized and secreted in the form of a polypeptide of 1706 amino acids: The calmodulin-dependent catalytic activity is localized in the first 400 amino acids. In order to be active, said adenylcyclase toxin is rendered invasive and hemolytic when post-traductionally modified by the coexpression of the *cyaC* gene product.

20 The following specific features of *Bordetella* adenylcyclase toxin indicate that this toxin can be used in the manufacturing of a proteinaceous vector for targeting *in vivo* a molecule of interest to CD11b expressing cells:

- 25 a) this adenylcyclase binds specifically to CD11b expressing cells
- b) the N-terminal catalytic domain is translocated into the cytosol of those CD11b expressing cells.
- 30 c) the C-terminal domain binds to the membrane of CD11b expressing cells and could internalize by endocytic pathway.
- d) epitope chemically coupled to genetically modified adenylcyclase can elicit *in vivo* specific CTL responses.

35 The expression "adenylcyclase" encompasses, within the present invention, natural or modified adenylcyclase, including genetically or chemically modified adenylcyclase,

providing the resulting product is able to target a molecule of interest specifically to CD11b expressing cells.

The invention thus relates to the use of *Bordetella* adenylcyclase as defined above and more particularly to the use of a modified or recombinant, adenylcyclase for targeting
5 a molecule of interest specifically to CD11b expressing cells.

More specifically, recombinant adenylclases include adenylcyclases which have been genetically modified to provide either adenylclases with peptide sequence or cysteine residues inserted within the catalytic domain, or truncated adenylcyclases lacking all or part of their catalytic domain.

10 Due to the specific interaction between *Bordetella* adenylcyclase toxin and CD11b/CD18 $\alpha_m\beta_2$ receptor, the molecule of interest is specifically targeted at least to the surface of CD11b expressing cells. In a particular embodiment of the invention, the *Bordetella* adenylcyclase toxin is used in the manufacturing of a proteinaceous vector to deliver the molecule of interest either in the cytosol of CD11b expressing cells, at the
15 surface of CD11b expressing cells or into the endocytic pathway of CD11b expressing cells.

Expression vectors for the preparation of recombinant *Bordetella* adenylcyclase are described in patent application WO 93/21324 (Institut Pasteur). Novel expression vectors for the preparation of genetically modified *Bordetella* adenylcyclase appropriate for
20 chemical coupling of a peptide of interest are also described in the experimental part hereafter. More specifically, expression vectors may be constructed directing the expression of both the *cyaA* gene and the *cyaC* gene (Sebo *et al.*, 1991). In parallel, a secondary plasmid carrying genes necessary for the secretion of the cytotoxic adenylcyclase in *E. coli*, such as *hlyB* and *hlyD* as described for example in Meckman *et al.*, 1985 can be constructed. In particular, the expression plasmid pCACT3 described in
25 WO 93/21324 can be used. Using this plasmid, the adenylcyclase may be expressed in *E. coli* and possibly secreted by this bacterium in large amounts. It is also readily purified for example using affinity chromatography on CaM Affi-Gel resin or other published procedures, as those using DEAE-sepharose and phenyl-sepharose (Guermonprez *et al.*,
30 2000).

In one embodiment of the invention, the adenylcyclase is a recombinant genetically modified adenylcyclase. In particular, mutations such as point mutations, deletions or insertions can be obtained using usual site-directed or random mutagenesis techniques, provided that the domains necessary for binding to CD11b expressing cells and,
35 optionnaly for translocation in the cytosol are still functional. Assays to evaluate specific

binding of recombinant toxins and fragments thereof, to CD11b expressing cells and optionally subsequent translocation of the catalytic domain are described in the following experimental part.

5 In another embodiment of the invention, the recombinant *Bordetella* species adenylylase is a fragment of the native, modified or recombinant *Bordetella* species adenylylase toxin, wherein said fragment is capable of binding the CD11b receptor. In particular, it has been found in the present invention that fragment encompassing residues 373 to 1706 (CyaA 373-1706) contains the structures essentially required for the interaction with the CD11b/CD18 receptor. Thus, a preferred fragment of the *Bordetella* species adenylylase toxin is the *Bordetella* adenylylase toxin lacking all or part of the N-terminal catalytic domain, and more specifically *Bordetella pertussis* adenylylase lacking all or part of residues 1-373.

Specific binding to CD11b can be assessed *in vitro* with anti-CD11b monoclonal antibodies as illustrated in the examples.

15 In order to be used in the manufacturing of proteinaceous vector or the preparation of a composition, the adenylylase is preferably non toxic. Non toxic mutants of the adenylylase toxin are well described in the Art. (Betsou *et al.*, 1993; Betsou *et al.*, 1995.

In a preferred embodiment of this invention, the adenylylase is isolated from *Bordetella pertussis*.

20 In specific embodiments, the molecule of interest is selected in the group comprising: peptides, glycopeptides, lipopeptides, polysaccharides, oligosaccharides, nucleic acids, lipids and chemicals.

In specific embodiments, a molecule of interest is a heterologous antigen. As used herein, the term "heterologous" refers to an antigen other than the adenylylase which is used in the vector itself.

25 In a preferred embodiment of the invention, the manufacturing of the proteinaceous vector comprises the step of inserting a heterologous molecule and especially a peptide in the catalytic domain of the adenylylase at a permissive site.

30 As used herein, the term "permissive site" relates to a site where the heterologous molecule and especially a peptide can be inserted without substantially affecting the desired functional properties of the adenylylase toxin, i.e. without affecting the domains necessary for the specific binding to CD11b/CD18 receptor and advantageously without affecting the process of translocation of the catalytic domain. In a preferred embodiment, the capacity of the CyaA toxin to promote the synthesis of cAMP in the targeted cells is further maintained.

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Methods to select for permissive sites are presented for example in WO93/21324 and in Ladant *et al.*, 1992. In particular, a methodology using a double selection (resistance to an antibiotic and calorimetric test on dishes by α -complementation) enables to identify readily oligonucleotides insertions (which preserve the reading frame) in the
5 portion of the gene coding for the N-terminal catalytic domain of the toxin. The functional consequences of these mutations on the catalytic activity of the toxin may be readily analysed, both genetically (functional complementation of an *E. coli cya*⁻ strain) and biochemically (characterization of the stability of the modified adenylyclases, of their enzymatic activity, of their interaction with caM, etc.). This methodology has enabled a
10 large number of mutations to be screened in order to identify the sites which are potentially advantageous for the insertion of antigenic determinants.

In specific embodiments of the invention, a permissive site is selected from the group consisting of residues 137-138, residues 224-225, residues 228-229, residues 235-236 residues 317-318 and residues 335-336 of the *Bordetella pertussis* adenylyclase.

15 However, other permissive sites may be used in the present invention, that can be identified for example by use of the methodology indicated above, especially sites between residues 400 and 1700.

The manufacturing of the proteinaceous vector can also comprise a step of fusing a molecule of interest, for example a heterologous peptide, at the N-terminal extremity of a
20 *Bordetella* adenylyclase lacking all or part of its N-terminal catalytic domain, and more preferably, *Bordetella pertussis* adenylyclase lacking residues 1-373.

In a preferred embodiment of the invention, the adenylyclase according to one of the above definitions is used in the manufacturing of a proteinaceous vector or in the preparation of a composition specifically designed to prime CD8⁺ cytotoxic T-cell response
25 (CTL response) said response follows the targeting of the adenylyclase modified (especially recombined or conjugated) with a molecule of interest to CD11b expressing cells, followed by the translocation of the molecule of interest to the cytosol of said CD11b expressing cells, and in particular to myeloid dendritic cells. In this context, the molecule of interest is or comprises preferably an epitope or an antigen.

30 As used herein, the term "epitope" refers to a heterologous molecule and especially a heterologous peptide that can induce an immune response.

In specific embodiments, the antigen is selected from the group consisting of an intracellular bacterial cell antigen, a tumoral cell antigen, a viral antigen, a fungus antigen or a parasite cell antigen.

In a preferred embodiment of the invention, the adenylylase, i.e., a natural, modified or recombinant adenylylase according to one of the above definitions is used in the manufacturing of the proteinaceous vector or in the preparation of a composition specifically designed to prime CD4+ cells response said response follows the targeting of the adenylylase modified (especially recombined or conjugated) with a molecule of interest to CD11b expressing cells, in particular myeloid dendritic cells. In this context, the molecule of interest is or comprises preferably an epitope or an antigen.

A molecule of interest can be especially an antigen selected from the group consisting of: a poliovirus antigen, an HIV virus antigen, an influenza virus antigen, a choriomeningitis virus epitope, a tumor antigen.

The functional properties of the CD11b expressing cells define furthermore a novel use of the *Bordetella* adenylylase toxin in the manufacturing of a proteinaceous vector for drug targeting to these specific cells. In this context, in one specific embodiment of the invention, the so-called molecule of interest is a drug. Said drug may be chemically or genetically coupled to the adenylylase. Method for coupling a drug to a polypeptide are well known in the Art and comprise for example disulfide linkage by using N-pyridyl sulfonyl-activated sulfhydryl.

Advantageously, a molecule of interest is an anti-inflammatory drug which is, when coupled to the adenylylase toxin, specifically targeted to the surface of the cells involved of the inflammatory response, such as neutrophils.

In particular, it is shown for the first time in the Experimental part that it is possible to graft molecules to CyaA by a chemical linkage or by genetic insertion for *in vivo* targeting to CD11b+ antigen presenting cells and particularly to the cytosol of CD11b+ antigen presenting cells. Indeed, when coupling a molecule corresponding to a given CD8+ T-cell epitope to the catalytic domain of detoxified CyaA, either by means of a disulfide bond or by genetic insertion, it has been found that the engineered molecule can elicit *in vivo* specific CTL response, thereby showing that said CD8+ T-cell epitope is translocated into the cytosol of CD11b-expressing cells.

More specifically, antigen presentation for selective CD8+ cytotoxic cells priming is mainly performed by myeloid dendritic cells.

Accordingly, in a specific embodiment, the recombinant adenylylase used for the manufacturing of proteinaceous vector is a genetically modified adenylylase containing one or more molecule(s) chemically coupled by means of a disulfide bond to genetically inserted cysteine residue(s) located within the catalytic domain of said adenylylase.

Indeed, multiple molecules can be chemically coupled to the adenylcyclase by means of a disulfide bond to different cysteine residues located at different permissive sites within the catalytic domain.

5 Applicant has also shown that CTL specific for the vectorized antigen can be primed *in vivo* after a single intravenous injection of the recombinant toxin, especially with no need to provide an heterologous adjuvant. These results shown in the experimental part and in particular the specific targeting of the epitope to myeloid dendritic cells enable new immunization strategies that bypass the requirement for adjuvant and CD4+ T cell help.

10 Therefore, the invention also relates to the use of a *Bordetella* adenylcyclase toxin recombined with a molecule and especially a peptide of interest for the preparation of a composition formulated for intravenous administration and enabling a CD8+ T cell immune response *in vivo*, said composition being free of a heterologous adjuvant. The invention also concerns this composition as such.

15 The invention in particular also relates to a new immunogenic composition formulated for administration, especially intravenous administration, in an animal or human host, characterized in that it comprises a recombinant *Bordetella* adenylcyclase which comprises an antigen inserted in the catalytic domain.

20 The invention further relates to a pharmaceutical composition for administration in a human or an animal formulated for targeting a molecule of interest specifically to CD11b expressing cells characterized in that said molecule of interest is coupled to a *Bordetella* species adenylcyclase.

25 In one preferred embodiment, the molecule of interest is selected in the group comprising: peptides, glycopeptides, lipopeptides, polysaccharides, oligosaccharides, nucleic acids, lipids and chemicals.

In another preferred embodiment, the molecule of interest is an antigen.

30 In another specific embodiment, the pharmaceutical or immunogenic composition comprises a nucleic acid construction encoding the recombinant *Bordetella* species adenylcyclase comprising a recombinant *Bordetella* species adenylcyclase coupled to a molecule of interest.

In specific embodiments, the adenylcyclase is from *Bordetella pertussis* toxin.

In other specific embodiments, the adenylcyclase toxin is a genetically modified toxin. In one preferred embodiment, the adenylcyclase is a non toxic adenylcyclase, especially a detoxified adenylcyclase.

In one preferred embodiment, the genetically modified adenylcyclase is able to translocate the molecule of interest specifically in the cytosol of CD11b expressing cells.

In particular, said genetically modified adenylcyclase is a *Bordetella* adenylcyclase lacking all or part of its catalytic N-terminal domain, and more specifically *Bordetella pertussis* adenylcyclase lacking residues 1-373.

Advantageously, the genetically modified adenylcyclase comprises one or more cysteine residues inserted within the catalytic domain at permissive sites. Such genetically modified adenylcyclase can be coupled to one or more molecule(s) of interest by means of disulfide bond(s) at the inserted cysteine residue(s).

In a preferred embodiment, the molecule, and especially an antigen is inserted in the permissive sites selected from the group consisting of residues 137-138, residues 224-225, residues 228-229, residues 235-236 and residues 317-318, and residues 335-336 of the *Bordetella pertussis* adenylcyclase, or the molecule is fused to the N-terminal part of a *Bordetella* adenylcyclase lacking all or part of its N-terminal catalytic domain, and more specifically to the N-terminal part of a *Bordetella pertussis* adenylcyclase lacking residues 1-373.

In a specific embodiment, the molecule is an antigen which is an intracellular bacterial cell antigen, a tumoral cell antigen, a viral antigen, a fungus antigen or a parasite cell antigen.

In preferred embodiments, a molecule of interest is an antigen selected from the group consisting of: a poliovirus epitope, an HIV virus, an influenza virus antigen, a choriomeningitis virus antigen, a tumor antigen.

In another embodiment, the genetically modified toxin is able to deliver the molecule of interest specifically at the surface of CD11b expressing cells or into the endocytic pathway.

The applicants have shown that *in vivo* intravenous administration of the immunogenic composition in an animal or human host as defined in the present invention without adjuvant(s) is sufficient to promote efficiently an immune response in said animal or human host.

In particular, the immunogenic compositions of the invention are capable of inducing or stimulating, *in vivo* or *in vitro* an immune cell response involving specifically dendritic cells.

As a consequence, in a specific embodiment, the immunogenic or pharmaceutical composition is advantageously devoid of priming adjuvants commonly used in the Art, such as aluminium hydroxide.

In one specific embodiment, the molecule of interest is a drug, preferably an anti-inflammatory drug.

Furthermore, the invention also relates to the use of the immunogenic composition as defined above for the preparation of a vaccine or an immunotherapeutic composition, for administration to an animal or human host.

As used herein, the term "immunotherapeutic composition" relates to a composition which leads to an immunological response and which is associated to therapeutic treatments, such as treatment against cancers, viral infections, parasites infections or bacterial infections.

The invention further relates to a method to immunize an animal or human host, wherein said method comprises the steps of:

- a) providing an immunogenic composition as defined above;
- b) administering said immunogenic composition, preferably via intravenous route, to said host in order to promote an immune response.

The invention finally relates to a proteinaceous vector for delivery of a molecule of interest, specifically to CD11b expressing cells, characterized in that said vector comprises a *Bordetella* species adenylcyclase and more preferably a recombinant or modified *Bordetella* species adenylcyclase coupled to said molecule of interest.

The proteinaceous vector is able to target a molecule of interest to CD11b expressing cells via the specific binding of *Bordetella* adenylcyclase with CD11b/CD18 $\alpha_m\beta_2$ which is present on the surface of specific cells. In one specific embodiment, the vector is also able to deliver the molecule of interest specifically in the cytosol of CD11b expressing cells.

In specific embodiments, the proteinaceous vector is able to target a molecule of interest to dendritic cells, particularly myeloid dendritic cells, or to neutrophils.

The molecule of interest is chemically or genetically coupled to the adenylcyclase, more preferably the recombinant adenylcyclase toxin. Method for coupling a molecule to a polypeptide are well known in the Art. As an example, the inventors showed that a biotin derivative, biotin HDPD can be selectively coupled on an unique cysteine residue genetically inserted within the catalytic domain. Synthetic peptides have been coupled similarly to the cysteine containing adenylcyclase toxin, or to genetically modified adenylcyclase devoid of native cysteine residue but containing an unique genetically inserted cysteine residue.

In another specific embodiment, a molecule of interest coupled to the adenylcyclase is an epitope chemically coupled to a genetically modified adenylcyclase devoid of native

cysteine residue but containing genetically inserted cysteine residue(s) at a permissive site within its catalytic domain.

In a specific embodiment, the proteinaceous vector comprises a genetically modified adenylcyclase. In one preferred embodiment, the adenylcyclase is a recombinant non toxic adenylcyclase from *Bordetella pertussis*.

The invention more specifically relates to a proteinaceous vector which consists of a recombinant *Bordetella* adenylcyclase lacking all or part of the N-terminal catalytic domain, and more preferably a *Bordetella pertussis* lacking residues 1-373.

Another preferred proteinaceous vector of the invention is a genetically modified *Bordetella* adenylcyclase devoid of native cysteine residue, but containing genetically inserted cysteine residue(s) at a permissive site within the catalytic domain.

Advantageously, in one specific embodiment of the invention, a drug to be delivered is an anti-inflammatory drug which is, when coupled to the adenylcyclase toxin, targeted to the surface or to the cytosol of the cells involved of the inflammatory response, such as neutrophils.

Definitions of the specific embodiments of the invention as claimed herein follow.

According to a first embodiment of the invention, there is provided an immunogenic composition formulated for administration in an animal or a human host characterized in that it comprises a *Bordetella* adenylcyclase which comprises an antigen inserted in the catalytic domain.

According to a second embodiment of the invention, there is provided a pharmaceutical composition for administration in a human or an animal formulated for targeting a molecule of interest specifically to CD11b expressing cells characterized in that said molecule of interest is coupled to a *Bordetella* species adenylcyclase.

According to a third embodiment of the invention, there is provided use of the immunogenic or pharmaceutical composition according to the first or second embodiment for the preparation of a vaccine or an immunotherapeutic composition.

According to a fourth embodiment of the invention, there is provided a proteinaceous vector for delivery of a molecule of interest, specifically to CD11b expressing cells, characterized in that said vector comprises a recombinant *Bordetella* species adenylcyclase coupled to said molecule of interest.

The following examples will illustrate the present invention without limiting the scope of the claimed invention. More specifically, in part A, experimental data are shown that reveal the specific binding of adenylcyclase toxin to CD11b/CD18 receptor, and in particular the specific binding of adenylcyclase toxin to CD11b expressing cells *in vitro*. In

part B, the experimental results show the possibility to target a genetically coupled molecule, more particularly an antigen *in vitro* and *in vivo* specifically to the cytosol of CD11b expressing cells, and in particular to myeloid dendritic cells. Moreover, the results show that the targeting is mediated by the CD11b receptor and that CTL priming is observed after systemic immunization in the absence of adjuvant. In part C., it is shown that specific coupling of epitope peptides by means of a disulfide bond on CyaA fragments can be carried out to provide new proteinaceous vectors. In part D., it is shown that, only the C-terminal part of CyaA is necessary and sufficient for interaction with CD11b receptors.

Finally, the results indicate that, unlike many other CTL responses as those raised against cross-priming antigen, CD4+ T cell help was not mandatory for priming of CTL responses.

LEGENDS TO FIGURES

Figure 1: Saturable binding of CyaA correlates with CD11b expression

a: CyaA binding at the surface of macrophages (J774A.1), B cells (LB27.4), and T cells (EL4) was performed at 37°C for 20 minutes. Surface-bound CyaA was detected with a biotinylated anti-CyaA polyclonal antibody, revealed by streptavidin-PE and detected by flow cytometry on living cells, as described in the material and methods section. Binding is expressed as $\Delta\text{MFI} = (\text{mean fluorescence intensity value of cells incubated with CyaA}) - (\text{mean fluorescence intensity of cells without CyaA})$.

b, c, d, e: Surface expression of α_2 integrins on J774A.1, LB27.4 and EL4 cells. CD11a (b), CD11b (c), CD11c (d), and CD18(e) expression were determined by flow cytometry using specific mAbs coupled to PE. Integrin expression is expressed as $\Delta\text{MFI} = (\text{mean fluorescence intensity value of cells stained with specific mAb}) - (\text{mean fluorescence intensity of cells stained with an isotype control mAb})$.

Figure 2: CyaA binding to murine cell lines is blocked by anti-CD11b mAb.

Cells were preincubated at 4°C for 15 minutes with or without 20 $\mu\text{g/ml}$ of specific mAbs and then incubated at 4°C for 20 minutes with 5 $\mu\text{g/ml}$ CyaA and with 10 $\mu\text{g/ml}$ of specific mAbs if present during the preincubation. Surface-bound CyaA was detected with a biotinylated anti-CyaA polyclonal, revealed by streptavidin-PE and detected by flow cytometry on living cells, as described in the material and methods section.

a, b: Effect of the M1/70 anti-CD11b mAb on the binding of various doses of CyaA. FSDC dendritic cells (a) or J774A.1 macrophages (b) were preincubated with medium alone (m)

or with M1/70 anti-CD11b mAb (l) and then incubated with CyaA with or without M1/70 anti-CD11b mAb. Bmax was determined by fitting experimental points obtained from experiments performed without mAbs to $\Delta\text{MFI} = \text{Bmax} * [\text{CyaA}] / (K_d + [\text{CyaA}])$. Binding of CyaA is plot as a % of Bmax plot against CyaA concentration.

5 c, d: Effect of specific mAbs on a fixed dose of CyaA binding. FSDC (c) or J774A.1 (d) cells were preincubated with or without specific mAbs (anti-CD11a, 2D7, antiCD11b, M1/70 and 5C6, anti-CD11c, HL3, anti-CD18, C17/16, control A95-1) and incubated with CyaA at the fixed concentration of 5µg/ml. Values of ΔMFI obtained for CyaA binding on cells treated with specific mAbs were normalized as ΔMFI values obtained for CyaA binding without mAb.

Figure 3 : CyaA binding to the human neutrophils is blocked by anti-CD11b and anti-CD18 mAbs.

15 a, b, c: Fluorescence histograms of freshly purified neutrophils were preincubated with medium alone (a), the 44 anti-CD11b mAb (b) or an isotype-matched control mouse mAb (c) and then incubated with (gray) or without (blank) biotinylated CyaA and revealed by streptavidine-PE. Cell number is plotted against log of PE fluorescence.

20 d: Effect of specific mAbs on CyaA binding to neutrophils (anti-CD11b, 44, M1/70, anti-CD18, TS/18, control mouse IgG2a, control rat IgG2b, A95-1). Freshly purified neutrophils were preincubated with or without specific mAbs and incubated with CyaA. Values of ΔMFI obtained for CyaA binding on cells treated with specific mAbs were normalized as a % of the ΔMFI values obtained for CyaA binding without mAb.

Figure 4 : Intracellular cAMP increase and cell death mediated by CyaA are specifically blocked by an anti-CD11b mAb in J774A.1 cells.

25 a: Effect of specific mAbs on intracellular cAMP accumulation. J774A.1 cells were preincubated at 4°C for 1 h with or without 10 µg/ml of specific mAbs (anti-CD11b, M1/70, anti-CD18, C17/16) and then incubated at 37°C for 20 min with 5 µg/ml CyaA and with 10 µg/ml mAbs if present during the preincubation. Intracellular cAMP contents were determined as described in the materials and methods section.

30 b: Effect of specific mAbs on CyaA mediated cell death. J774A.1 cells were preincubated at 4°C for 1 h with medium alone or with 10 µg/ml of specific mAbs (anti-CD11a, 2D7 anti-CD11b, M1/70, anti-CD11c, HL3, anti-CD18, C71/16, control, 2.4G2). Then they were incubated at 37°C for 2 h with 0.5 µg/ml CyaA and with 10 µg/ml of specific mAbs when present during the preincubation. Cell lysis was determined by LDH release using the Cytotox 96™ assay.

Figure 5 : CHO cells bind CyaA and become sensitive to CyaA when transfected with CD11b, but not with CD11c

5 a, b: CyaA binding at the surface of CHO transfectants. CHO cells transfected with human CD11b/CD18 (l), human CD11c/CD18 (m) or mock-transfected (w) were incubated with various doses of CyaA for 20 min at 37°C (a) or 4°C (b). Surface-bound CyaA was detected with a biotinylated anti-CyaA polyclonal antibody, revealed by streptavidin-PE and detected by flow cytometry on living cells, as described in the material and methods section. Binding is expressed as \square MFI = (mean fluorescence intensity value of cells incubated with CyaA) - (mean fluorescence intensity of cells without CyaA).

10 c: Intracellular cAMP accumulation in CHO transfectants. CHO cells transfected with human CD11b/CD18 (l), human CD11c/CD18 (m) or mock-transfected (w) were incubated with or without CyaA for 20 min at 37°C. Intracellular cAMP contents were determined as described in the materials and methods section.

15 d: Cell lysis in CHO transfectants. CHO cells transfected with human CD11b/CD18, human CD11c/CD18 or mock-transfected were incubated with 5 μ g/ml CyaA for 4 h at 37 °C. Cell lysis was determined by LDH release using the Cytotox 96™ assay.

Figure 6 : Intravenous immunization with CyaAOVA primes anti-OVA CTL responses in a B cell, CD4 and CD40-independent way.

20 C57BL/6 WT +/+ (a), CD4-/- (b), CD40-/- (c) or IgM-/- (d) mice were intravenously immunized with 50 μ g of CyaAOVA, a genetically detoxified form of CyaA carrying the H-2K^b restricted SIINFEKL epitope from OVA (l, m) or with CyaAE5, a control detoxified toxin without the OVA epitope (s, Δ). Seven days after, animals were sacrificed and splenocytes were restimulated *in vitro* for 5 days with 10 μ g/ml of the pOVA synthetic peptide in the presence of irradiated C57BL/6 splenocytes. CTL activity was assessed in a 4 hours chromium⁵¹ release assay against H-2K^b EL4 cells previously pulsed (l, s) or not (m, \square) with pOVA at 10 μ g/ml.

Figure 7 : Identification of splenic antigen presenting cells involved in CyaAOVA presentation *in vitro* or *in situ*, after intravenous immunization.

30 The low density fraction of splenocytes presents CyaAOVA to a specific anti-OVA CD8+ T cell hybridoma (a, b):

In vitro assay (a) : Low (LDF, l) and high density (HDF, s) fractions or unfractionated total splenocytes from naive mice (TSC, n, r) were cultured with B3Z, a CD8⁺ T cell hybridoma specific for the pOVA peptide in the context of H-2K^b. After 18 hours of coculture in the presence of the recombinant detoxified CyaA carrying the OVA peptide (CyaAOVA, l, s, n) or a control peptide (CyaALCMV, r) at various concentrations, IL-2 released in

supernatants was measured in a CTLL proliferation assay. Results are expressed in \square cpm and plotted against CyaA concentration during the assay. \square cpm = [cpm + CyaA] - [cpm - CyaA]).

5 *Ex vivo* assay (b): Spleen cells were obtained from mice previously immunized iv (6-12 hours) with 50 μ g of CyaAOVA (l, s, n) or CyaALCMV (m) and fractionated in LDF and HDF. Various numbers of cells recovered from TSC (n), LDF (l, m) or HDF (s) or unfractionated splenocytes (n) were directly put in culture with B3Z without addition of recombinant CyaA. IL-2 release was assessed after 18 hours of culture as described before. Results, expressed in cpm, are plotted against the number of APC present in each well.

Dendritic cells (CD11c⁺) are more efficient APC for CyaAOVA than the CD11b^{high}CD11c⁻ cells or B cells (CD45R⁺) (c,d):

10 *In vitro* assay (c): CD11c⁺ (l) sorted cells from LDF, CD11b^{high}CD11c⁻ (m) and CD45R⁺ (n) sorted cells from TSC, were put in culture with B3Z for 18 hours in the presence of various concentrations of CyaAOVA. IL-2 was assessed as above.

15 *Ex vivo* assay (d): Cells sorted by flow cytometry from C57BL/6 mice previously (6-12 hours) immunized with 50 μ g of CyaAOVA were used as APC. CD11c⁺ (l), CD11b^{high}CD11c⁻ (m), CD45R⁺ (n) sorted cells from low density splenocytes were directly put in culture for 18 hours with B3Z at various numbers of cells per well, without addition of CyaAOVA. IL-2 was assessed as above.

The CD8 α ⁻ myeloid dendritic cell subset is a more efficient APC for CyaA than the CD8 α ⁺ lymphoid dendritic cell subset (e, f):

20 CD11c⁺ low density cells from naive mice (e) or mice previously (6-12 hours) immunized iv with 50 μ g CyaAOVA (f) were fractionated in myeloid dendritic cells (CD11c⁺CD8 α ⁻) and lymphoid dendritic cells (CD11c⁺CD8 α ⁺,m) by flow cytometry and used as APC in *in vitro* (e) and *ex vivo* (f) assays for B3Z stimulation. IL-2 was assessed as above.

B cell genetic depletion does not impair CyaAOVA presentation by splenocytes (g, h):

30 TSC (n, r), LDF (l, m) or HDF (s, Δ) from C57BL/6 WT mice (n, l, s) or B cell deficient (r, m, \square) were used as APC in an *in vitro* assay (g, n, r) or an *ex vivo* assay (h, n, l, s, r, m, Δ) for B3Z stimulation as in a. Mice were either from naive (g) or previously (1.5 hours) immunized iv with 50 μ g CyaAOVA (h). IL-2 was assessed as above.

Figure 8 : The presentation of CyaAOVA by dendritic cells requires the TAP transporters *in vitro* and *in vivo* after intravenous immunization.

In vitro assay (a) : TSC (s, Δ) or CD11c⁺ (l, m) sorted cells from control C57BL6 TAP^{+/+} (s, l) or TAP^{-/-} mice (Δ, m) were cultured with B3Z in the presence or not of various doses of CyaAOVA. IL-2 was assessed as described in figure 6a. Results are expressed in cpm plotted against antigen concentration.

Ex vivo assay (b) : TSC (s, Δ) or CD11c⁺ (l, m) sorted cells from control C57BL6 TAP^{+/+} (s, l) or TAP^{-/-} mice (□, m) previously iv immunized with 50 μg of CyaAOVA were cultured with B3Z for 18 hours. IL-2 was assessed as described in figure 6a. Results are expressed in cpm plotted against the number of cocultured cells.

Figure 9 : Role of α_mβ₂ integrin (CD11b) in CyaAOVA binding to cells.

Binding of CyaAOVA-biotine to TSC is blocked by anti-CD11b (a) : TSC suspensions were incubated at 4°C with 10 μg/ml of the anti-CD11b M1/70 mAb or an isotype control mAb or nothing. Then, CyaAOVA-biotine at 2 μg/ml (left pannels) or various concentrations (right pannel) was added to the cells for 30 mn at 4°C. After a wash, CyaAOVA-biotine was revealed with streptavidine-PE for 30 mn (Strep-PE). Then, after washing, cells were resuspended in PBS containing propidium iodide. The size (FSC) of living cells gated by propidium iodide exclusion was plotted against the Strep-PE fluorescence. The percentage of leukocytes positive for CyaAOVA-biotine was plotted against CyaAOVA-biotine concentration during the staining.

Binding of CyaAOVA-biotine to low density cells correlates with the expression of CD11b (b) : LDF were triple stained for CD11c, CD8α and CyaOVBiotine (or medium) or, in separate experiments with CD11c, CD8α and CD11b (or a control mAb). After a wash, cells were stained for 30 mn with Strep-PE to reveal CyaAOVA-biotine, anti-CD11c-FITC and anti-CD8α-APC. Gates were done on lymphoid DC (CD11c⁺CD8α⁺), myeloid DC (CD11c⁺CD8α⁻), CD8⁺ T cells (CD11c⁻CD8α⁺) and other cells (CD11c⁻CD8α⁻). For each gate, CyaAOVA-biotine staining or CD11b staining is plotted against cell number in separate histograms. Left histograms : LDF suspensions were incubated with 0 (histograms filled in grey), 2.5 (narrow, open histograms) or 10 μg/ml (thick, open histograms) of CyaAOVA-biotine for 30 minutes at room temperature. Right histograms : isotype control-PE (histogram filled in grey), CD11b-PE (thin, open histograms).

Figure 10 : Role of α_mβ₂ integrin (CD11b) in CyaAOVA presentation by MHC I

In vitro antigen presenting assay with TSC (a, b): The same experiments than in a, b were performed with TSC from naive C57BL/6 mice as APC. The stimulation of B3Z was

assessed by IL-2 release in coculture supernatants measured in a CTLL proliferation assay. Results are plotted in cpm against CyaAOVA or pOVA concentration.

Ex vivo antigen presenting assay with TSC or CD11b⁺ and CD11b⁻ fractions (c, d): C57BL/6 mice were intravenously immunized with 50 µg of CyaAOVA (c) or 10 µg of pOVA (d). CD11b⁺ (l) and CD11b⁻ (m) cells were sorted by flow cytometry from TSC (Δ) and put in culture at various cell number per well with B3Z. After 18 hours of coculture, the stimulation of B3Z was assessed by IL-2 release. Results, expressed in cpm, are plotted against the numbers of APC from immunized animals present in each well.

Figure 11: Summary of the methodology for chemical coupling of epitopes to recombinant CyaA through disulfide bond.

Figure 12: A diagram of a vector for chemical coupling of CTL epitopes

Figure 13: A graph showing IL-2 release by B3Z measured in a CTLL proliferation assay.

3.10⁵ spleen cells from C57BL/6 mice were cocultured for 18 h with 105 B3Z cells in the presence of various concentrations of cyclases. The IL2 release by B3Z was measured in a CTLL proliferation assay.

Figure 14: A graph showing cytotoxic activity measured on ⁵¹Cr -labelled EL4 target cells pulsed (A) or not (B) with 50 µM of the OVA peptide.

C57BL/6 mice were iv injected with 50 µg of the various CyaA. Seven days later, spleen cells were *in vitro* stimulated with OVA peptide. The cytotoxic activity was measured on ⁵¹Cr-labelled target cells.

The figure 14 shows the *in vivo* capacity of the proteinaceous vectors of the invention to induce OVA-specific CTL responses.

Figure 15: Inhibition of CyaA binding to CD11b by CyaA-E5 and CyaA fragments

CHO cells transfected with CD11b/CD18 were preincubated on ice for 1 hour with different concentrations of CyaA-E5 (black triangle), CyaA 1-383 (black square) or CyaA-373-1706 (Black diamond) and then incubated on ice for 30 min with 5 µg/ml of biotinylated CyaA-E5. Surface bound cyclase was revealed using streptavidin-PE and analyzed by flow cytometry on living cells. Results are expressed as mean fluorescence intensity (A), percentage of positive cells (B) and percentage of inhibition (C).

Figure 16: Schematic representation of pTRAC-373-1706 expression vector.

The large arrows represent the open reading frames of β-lactamase (bla), the thermosensitive repressor cI⁸⁵⁷ of phage lambda (AcI⁸⁵⁷), the cyaC gene and truncated

'cyaA gene (the arrows are pointing to the direction of translation of the corresponding genes). The ColE1 origin (Ori), the Pr promoter (λ Pr), and some relevant restriction sites are also indicated. The intergenic region between the *cyaC* and truncated 'cyaA genes is detailed in the lower part. It shows the new C-terminus extension of *cyaC* (downstream to Arg182 of wild-type CyaC), the stop codon (underlined), the initiator Met and first codons of CyaA373-1706 (upstream to Ser373 of wild-type CyaA).

EXAMPLES

A. The *Bordetella* adenylate cyclase toxin interacts specifically with the $\alpha_M\beta_2$ integrin (CD11b/CD18)

A.1 Materials and Methods

A.1.1 Recombinant toxins and antibodies

A protocol for CyaA production has already been described elsewhere [Karimova, et al, 1998]. CyaA toxins were produced in *E. coli* BLR strain harboring an expression plasmid, pCACT3, which carries the *cyaA* structural gene CyaA under the lacUV5 promoter and the *cyaC* accessory gene required for activation of the protoxin. After solubilization in 8M urea, Hepes-Na 20mM, pH 7.5, CyaA was purified to more than 95% homogeneity (as judged by SDS-gel analysis, not shown) by sequential DEAE-Sepharose and Phenyl-Sepharose. A recombinant detoxified CyaA toxin, CACTE5-Cys-Ova, harboring a unique cysteine inserted within the genetically inactivated catalytic domain was constructed by inserting an appropriate double strand oligonucleotide between the BsiwI and KpnI sites of pCACT-Ova-E5 [Guermonprez *et al*, 2000]. In the resulting protein CACTE5-Cys-Ova, the amino acid sequence ASCGSIINFEKLGIT is inserted between residues 224 and 225 of CyaA. The recombinant toxin was expressed and purified as previously described. The purified protein was labeled on its unique Cys with the highly specific sulfhydryl reagent N-(6-(Biotinamido)hexyl)-3'-(2'-pyridyldithio) propionamide (Biotin-HPDP, PIERCE) according to the manufacturer's instructions. The biotinylated-CyaA was re-purified on DEAE-Sepharose to remove the unreacted Biotin-HPDP reagent. Toxin concentrations were determined spectrophotometrically from the adsorption at 280 nm using a molecular extinction coefficient of $142 \text{ M}^{-1} \times \text{cm}^{-1}$ (binding studies) or using the Biorad protein assay system (cAMP accumulation and cell death studies).

Purified mAbs specific for murine CD11a (2D7, Rat IgG2a, κ), murine and human CD11b (M1/70, Rat IgG2b, κ), murine CD11c (HL3, Hamster 1, λ), murine CD18 (C71/16, Rat IgG2a, κ), control (A95-1, or anti-CD16/32, 2.4G2, Rat IgG2b, κ) originated from Pharmingen (San Diego, USA). Supernatants from anti-human CD11b (44, Mouse IgG2a,

κ), and anti-human CD18 (TS/18, Mouse IgG1, κ) hybridoma were a kind gift and were used at 1/2 dilution in blocking experiments. Supernatants from an anti-murine CD11b (5C6, Rat IgG2b, κ) were a kind gift from G. Milon (Pasteur Institute, Paris) and were used at 1/2 final dilution in binding inhibition assays. Anti-CyaA polyclonal antibodies were obtained from a rabbit immunized subcutaneously with purified CyaA. Sera were precipitated from immune serum by ammonium sulfate (33%). After centrifugation the pelleted proteins were resuspended in 20 mM Hepes-Na, 150 mM NaCl, pH 7.5 (buffer C) and extensively dialysed against the same buffer. The antibodies were then biotinylated by incubation with Biotin-amidocaproate N-Hydroxysuccinimide ester (SIGMA, dissolved in dimethyl sulfoxide) for 130 min at room temperature. Then, 100 mM ethanolamine, pH 9.0 were added and after 30 min of additional incubation, the mixture was extensively dialysed at 4 °C against buffer C. Biotinylated antibodies were stored at - 20°C.

A.1.2 Cells and culture media

EL4, J774A.1, LB27.4, THP-1 were obtained from the American Type Culture Collection (ATCC) and were cultured in RPMI 1640 supplemented with 10% fetal calf serum, 100 U/ml penicillin, 100 µg/ml streptomycin, 2mM L-glutamine, with or without 5×10^{-5} M 2-mercaptoethanol (complete medium). FSDC [Girolomoni et al, 1995] were cultured in complete medium. CHO cells transfected for human CD11b/CD18 or CD11c/CD18 or transfected with the vector only were obtained from D. Golenbock (Boston, USA) and cultured in the presence of neomycin as previously described [Ingalls et al, 1998]. Human neutrophils were purified as previously described [Rieu et al, 1992].

A.1.3 CyaA binding assays

All binding assays were performed in DMEM 4.5 mg/ml glucose (Life Technologies) without serum in 96 well culture plates (Costar). 2×10^5 cells/well were incubated for 20 minutes (at 4°C or 37°C depending on the experiments) in a 200 µl final volume. In some experiments, cells were preincubated for 20 min at 4°C in the presence of blocking mAbs in 100 µl final volume. The toxin solution was added to the wells in the continuous presence of the mAbs in a total volume of 200 µl at 4°C. Then, plates were centrifuged at 1500 rpm for 5 min and supernatants were removed. Cells were incubated at 4°C for 25 minutes with biotinylated anti-CyaA rabbit polyclonal antibodies (1/400 in DMEM, 50 µl/well) in the presence of a control (non-immune or pre-immune) rabbit serum as a saturating agent (1/50).

After centrifugation and supernatant removal, cells were stained with streptavidin-phycoerythrin (PE) (Pharmingen) at 1/300 dilution (50 μ l/well). After washing, cells were analyzed by flow cytometry on a FACStar (Becton-Dickinson, Mountain View, USA) in the presence of 5 μ g/ml propidium iodide. Gatings were done to exclude cells aggregates and dead cells by propidium iodide exclusion. Experimental points were fitted to a hyperbolic model $\Delta\text{MFI} = \text{Bmax} * [\text{CyaA}] / (K_d + [\text{CyaA}])$, with Bmax = % of maximal binding, using the prism software.

A.1.4 cAMP assay

cAMP accumulation was measured by an antigen competition immunoassay [Karimova et al, 1998] in which the incubation medium was composed of DMEM without serum but containing 4.5 mg/ml glucose and 20 U/ml hexokinase. Hexokinase, which catalyzes the ATP-dependent phosphorylation of glucose, was added deplete the extracellular medium for any traces of ATP, thus preventing the extracellular synthesis of cAMP. Therefore, the amount of cAMP measured is representative of the accumulation of strictly intracellular cAMP. 5×10^5 cells were preincubated in 96 well plates in 100 μ l/well with or without 10 μ g/ml of specific mAbs at 4°C for 1 h and then incubated at 37°C for 20 min with 0.05, 0.5 or 5 g/ml CyaA and with 10 μ g/ml of specific mAbs when present during the preincubation. For the dose response effect of CyaA, cells were directly incubated with the toxin for 20 min at 37°C. After intoxication, cells were centrifuged at 2.500 rpm for 5 min. Samples were lysed with 100 μ l of HCl 0.1 N, boiled for 5 min at 120 °C, and neutralized with 100 μ l of Tris 0.125 M, NaCl 0.2 M. Microtiterplates were coated with cAMP-BSA conjugates diluted at 1/4.000 in Na_2CO_3 0.1 M pH 9.5. They were washed twice in PBS-Tween 0.1%, saturated for 1 h in PBS-BSA 2% and washed five times with PBS-Tween 0.1%. The samples and the cAMP standard (Sigma) were directly added to the plates coated with cAMP-BSA conjugates and serially diluted with a 1/1 mixture of HCl 0.1 N and Tris 0.125 M- NaCl 0.2 M. Anti-cAMP rabbit antibody was added at 1/2.500 in PBS-BSA 2% and incubated at 37°C for 3 h. Plates were washed five times with PBS-Tween 0.1%. Anti-rabbit antibodies coupled to horseradish peroxidase (Amersham) were added at 1/2.500 in PBS-BSA 2%, incubated at 37°C for 1 h and revealed using the classical peroxidase reaction. Experimental points of the standard curve were fitted to a sigmoid model using the Prism software.

A.1.5 Toxicity assay

Cell death was evaluated as previously described [Khelef et al, 1993 ; Khelef et al, 1995]. Briefly, 10^5 cells were incubated for 24 hours in a 96 well plate in complete medium

and washed once with serum free medium. All cell incubations were further performed in serum free medium. Dose response effects were evaluated by directly applying various concentrations of CyaA to CHO cells at 37 C for 4 h. For cytotoxicity inhibition, cells were preincubated at 4°C for 1 h with or without 10 µg/ml of specific mAbs and then incubated at 37°C with 0.5 µg/ml CyaA for 2 h for J774A.1A.1 cells or with 5 µg/ml for 4 h for CHO cells and with 10 µg/ml of specific mAbs when present during the preincubation. Cell lysis was evaluated using the Cytotox 96™ assay (Promega) which quantifies the amount of lactate dehydrogenase (LDH) released in the medium by dying cells.

A.2 Results

A.2.1 Saturable binding of CyaA correlates with the presence of CD11b at the surface of target cells

To characterize CyaA cellular specificity toward a population of leukocytes, we choose three representative murine cell lines expressing various combination of β₂ integrins: J774A.1, a tumoral macrophage; EL4, a T cell thymoma, and LB27.4, a B cell lymphoma. After a 20 minutes of incubation with CyaA at 37°C, binding of CyaA to the cell surface of these cells was monitored by flow cytometry using biotinylated anti-CyaA antibodies and streptavidine-PE. Under these conditions, we observed an efficient, dose-dependent and saturable binding of CyaA on J774A.1 cell line. The affinity of CyaA for J774A.1 cells was high since the apparent K_d were 9.2 ± 4.5 nM and 3.2 ± 1.9 nM, respectively. A low binding of CyaA to EL4 and LB27.4 cells was observed but it was not saturable at the concentrations tested.

In order to determine if the binding of CyaA to J774 cell lines was correlated to the expression of one of the members of the β₂ integrin family, we performed a phenotypic analysis of these cells by flow cytometry using monoclonal antibodies (mAbs) specific for the three α chains of the well characterized β₂ integrins (CD11a, Cd11b and CD11c) and for the common β chain (CD18) (Figure 1b, c, d, e). J774A.1 cells expressed mostly CD11b and CD18, but were also positive for CD11a. EL4 cells and LB27.4 cells expressed mostly CD11a and CD18. Taken together, these data show that the efficient and saturable binding of CyaA to J774A.1 was correlated with the presence of the integrin CD11b/CD18.

A.2.2 CyaA saturable binding is specifically blocked by anti-CD11b mAbs

We next examined if CD11b/CD18 could be directly involved in the binding of CyaA to the cells expressing this integrin. We performed a quantitative analysis of inhibition obtained with anti-CD11b M1/70 mAb by calculating percentage of mean fluorescence values in the absence of mAbs at a fixed or varying concentrations of CyaA (figure 2).

Inhibition of CyaA binding obtained with the M1/70 anti-CD11b mAb was almost total at most CyaA concentrations tested (figure 2a, b). This inhibition was specific since anti-CD11a, CD11c, CD18 or a control mAb did not inhibit CyaA binding. A second anti-CD11b mAb (clone 5c6) also inhibited binding of CyaA (figure 2 c,d). Similar results were obtained with FSDC, an immature dendritic cell line expressing CD11b (figure 2a, c), and the J774A.1 macrophage cell line (figure 2b, d).

To examine whether CyaA could similarly interact with human CD11b, CyaA binding studies were performed on human neutrophils, whose high expression of CD11b is well established. Since high background fluorescence was obtained following incubation of human myeloid cells with the anti-CyaA rabbit antibodies (data not shown), we set up an alternate binding assay. A detoxified form of CyaA was specifically biotinylated on unique cysteine residues, genetically introduced within its catalytic domain. Using this system, we were able to detect CyaA binding to neutrophils (Figure 3). Preincubation of neutrophils with the 44 or M1/70 anti-CD11b mAbs led to, a respectively complete or partial inhibition of the binding of CyaA (Figure 3a and b). Unlike the C71/16, anti-murine CD18 mAb that did not block CyaA binding to murine cells, preincubation with the human anti-CD18 TS/18 mAb led to a complete inhibition of CyaA binding to human neutrophils (Figure 3b). Similar results were obtained with the human monocytic THP-1 cell line (data not shown).

In conclusion, CyaA binding to the surface of three myeloid cell lines from both murine and human origin (J774A.1, FSDC, THP-1) as well as freshly purified human neutrophils appears to be mainly mediated through the CD11b/CD18 integrin.

A.2.3 CyaA-mediated cAMP increase and toxicity are specifically blocked by an anti-CD11b mAb

To evaluate the physiological relevance of CD11b/CD18-dependent CyaA binding, we studied the effect of blocking mAbs on the cytotoxicity of CyaA. We first measured the amount of cAMP produced in J774A.1 cells exposed to CyaA in the presence of various mAbs. As shown in Figure 4a, the increase in the intracellular cAMP content induced by CyaA was totally abolished when cells were preincubated with the M1/70 anti-CD11b mAb. The C17/16 anti-murine CD18 mAb had that did not block CyaA cell binding (figure 2) had no effect on the intracellular cAMP content of CyaA treated cells. Thus, these data strongly suggest that the increase in intracellular cAMP induced by CyaA is dependent upon the interaction of the toxin with CD11b. To further analyze the requirement of this molecule for the toxicity of CyaA, we evaluated the effect of mAbs specific for the different chains of the β_2 integrin family on CyaA mediated cell death. Figure 4b shows that anti-CD11b mAb J774A.1 dramatically reduced the cell death induced by CyaA (88 % inhibition). The cell

death induced by CyaA was unaffected when J774A.1 were preincubated with mAbs that did not block toxin binding to cells (anti-CD11a, CD11c or CD18 or a control mAb).

Taken together, these data indicate that CyaA binding through CD11b is strictly required for CyaA mediated toxicity in J774A.1 cells.

5 **A.2.4 Transfection of CHO cells with CD11b/CD18 confers sensitivity to CyaA**

To confirm the role of CD11b in CyaA binding, we used CHO cells transfected with the human integrins CD11b/CD18 or CD11c/CD18 or mock transfected (vector alone). As shown in Figure 5a, CyaA could bind, at 37°C, to these cell lines. However, CyaA binding was efficient and saturable in CHO cells expressing CD11b/CD18 but not in CD11c/CD18 or mock-transfected cells. The affinity of CyaA for CD11b/CD18 transfected cells was in the nM range ($K_d = 0.7 \pm 0.09$ nM). At 4°C, the efficiency of CyaA binding was reduced as compared with the binding at 37°C (Figure 5b). At this temperature, the differences between CD11b/CD18 transfected cells and the two other cell lines were more pronounced.

15 Since we found that CD11b was required for CyaA mediated toxicity in J774A.1, we then determined if CD11b expression was sufficient to confer a CyaA-sensitive phenotype to CHO transfected cells. In line with previous reports [Gordon et al, 1988], CyaA induced a notable amount of intracellular cAMP in CHO cells transfected with CD11c/CD18 or in control mock-transfected cells, but only at high concentrations of toxin (5 µg/ml, figure 5c).
20 In contrast, CyaA increased intracellular level of cAMP in CD11b/CD18 transfected CHO cells even at the lowest concentration studied (0.05 µg/ml). Moreover, the cAMP production in response to 5 µg/ml CyaA was 4 to 5 times more elevated in CD11b/CD18 transfected cells as compared to CD11c/CD18- or mock-transfected cells.

We also evaluated the role of CD11b/CD18 in CyaA-mediated cell death. As shown 25 in figure 5d, more than 50% of CHO transfected with CD11b/CD18 were killed after 4 hours incubation with 5 µg/ml of CyaA, whereas CD11c/CD18- or mock-transfected cells were not affected by this treatment.

Altogether, these results thus clearly established that expression of human CD11b/CD18 integrin is sufficient to create a high affinity receptor for CyaA in CHO cells.

30 **A.3 Discussion: A receptor for CyaA**

Unlike other toxins, CyaA has been considered for a long time, as independent of any receptor binding. This is based on the observations that *i)* CyaA can intoxicate in vitro a wide variety of model cell lines from various origin [Ladant et al, 1999] *ii)* CyaA binds to Jurkat cells and sheep erythrocytes in a non saturable fashion [Gray, et al 1999]. In fact,

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these observations established that non-specific adsorption of CyaA to lipid membranes leads to some translocation of the catalytic domain into the cytosol. However, they did not rule out the existence of a specific receptor. We showed in this study on myeloid cell lines that the binding and the toxic properties of CyaA are dependent on its interaction with the integrin CD11b/CD18. Efficient and saturable binding correlates with the expression of CD11b and is fully and specifically blocked by anti-CD11b mAbs. Moreover, expression of CD11b/CD18 in CHO cells dramatically enhances the binding of CyaA, resulting in an increased sensitivity to intoxication by this toxin. Our results are the first evidence supporting the interaction of CyaA with a cell-surface molecule specifically expressed on leukocytes. The nearly complete blockade of CyaA binding by anti-CD11b mAbs suggests that CD11b is the main receptor for CyaA in the cell lines tested. The lack of efficient binding to CD11c/CD18 transfectants, or CD11a/CD18 expressing cells such as EL4 or LB27.4 suggest that CD11b/CD18 is the only integrin of the β_2 family involved in the binding of CyaA to target cells.

In line with previous studies, we observed a detectable binding of CyaA to all cell lines tested. Furthermore, CyaA at high concentrations triggered a small but detectable cAMP increase in mock-transfected CHO cells, that is not associated to cell death. Thus, at high concentration, CyaA can bind and translocate to a wide variety of cell lines but efficient and saturable binding, translocation and killing is a hallmark of CD11b expressing cells.

Binding of CyaA to a member of the β_2 integrin family is reminiscent of the behavior of other RTX toxins which were recently found to interact with these molecules [Lally et al, 1997; Li et al, 1999; Ambagala et al, 1999; Jeyaseelan et al, 2000]. The *E. coli* HlyA, that shares a strong homology with CyaA, forms cationic pores at the plasma membrane. HlyA exhibits a specificity for leukocytes but only at low concentration [Welch et al, 1991]. This relative specificity was shown to be mediated by its interaction with the integrin CD11a/CD18 [Lally et al, 1997]. Similarly, *A. actinomycetemcomitans* and *P. haemolytica* leukotoxin (LtxA and LktA, respectively), which are less promiscuous RTX toxins specific for human and bovine leukocytes, respectively, also interact with CD11a/CD18 [Lally et al, 1997; Li et al, 1999; Ambagala et al, 1999; Jeyaseelan et al, 2000]. Despite its strong homology with HlyA, CyaA recognizes another β_2 integrin (CD11b/CD18) whose cellular distribution is different. Indeed, CD11b is expressed mostly on macrophages, neutrophils and dendritic cells, but not on the majority of T and B cells, whereas CD11a is expressed on all leukocytes including T and B lymphocytes.

This specific targeting of CyaA to CD11b expressing cells is exploited in the present invention to specifically target this particular subset of cells. Detoxified mutants of CyaA remaining invasive could be used for the delivery of pharmacologically active molecules to CD11b positive cells, without noticeably affecting other cell types.

5 **B. Targeted antigen delivery to the cytosol of myeloid dendritic cells and selective CTL priming**

B.1 Materials and Methods

B.1.1 Recombinant Adenylate Cyclase toxins and peptide

10 The pOVA synthétique peptide (SIINFEKL) originated from NEOSYSTEM and was diluted in PBS at 1mg/ml.

B.1.2 Immunization and assay for the detection of cytotoxic T cells

Female C57BL/6 (H-2^b) from Iffa Credo (L'arbresle, France) were used between 6 and 8 weeks of age. TAP1 -/- (Van Kaer et al., 1992), CD4-/- (Killeen et al., 1993), CD40-/- (Kawabe et al, 1994) and B cell deficient μ MT (Kitamura et al, 1991) bred onto a C57BL/6
15 background originated from the CDTA facility (CNRS, Orleans, France) and were bred in the Institut Pasteur facilities. Animals were intravenously immunized with Ag in PBS. Seven days post-injection, animals were sacrificed and the spleen was removed. Single cell suspensions of splenocytes (2.5×10^7 cells) were restimulated in 10 ml CM (see below) with irradiated spleen cells (2.5×10^7 cells) for 5 days in the presence of 1 μ g/ml pOVA.
20 Cytotoxicity assay was performed exactly as previously described (Fayolle, et al., 1999).

B.1.3 Cell lines

B3Z (Karttunen et al., 1992), a CD8+ T cell hybridoma specific for the OVA 257-264 peptide (SIINFEKL) in the context of H-2K^b was a generous gift from Dr N. Shastri (University of California, Berkeley, USA).

25 **B.1.4 Antigen presentation assays**

All antigen presentation assays were performed by coculture of APC with B3Z in 96 well culture microplates (0.2 ml final volume) in RPMI 1640 supplemented with 10 % Fetal Calf Serum, 100 U/ml penicillin, 100 μ g/ml streptomycin, 2 mM L-glutamine, and 5×10^{-5} M 2-mercaptoethanol (complete medium, CM). The stimulation of B3Z cells (10^5 cells/well)
30 was monitored by IL-2 release in the supernatants of 18-24 hours cultures in the presence of APC. IL-2 was measured in CTLL assay as previously described (Guermontprez et al., 1999). In some experiments (see figure legends), B3Z stimulation was assessed using the

NF-AT lacZ reporter assay. LacZ activity in cell lysates was assessed with the CPRG substrate as previously described (Karttunen et al., 1992). Two antigen presentation assays were performed : i) *In vitro* assay: APC originated from naive mice were cocultured (10⁵/well) with B3Z in the presence of Ag at various concentrations. In some experiments, APC were preincubated or not with mAbs at 10 µg/ml for 40 minutes at 4°C, then Ag was added to the cells in a 100 µl final volume in the continuous presence of the mAbs. After a 4 hours pulse, APC were washed twice and put in coculture with B3Z. Purified mAbs used were against CD11b (M1/70 ratIgG2b, □) or isotype-matched control and originated from Pharmingen (San Diego, USA). ii) *Ex vivo* assay : APC originated from mice previously intravenously immunized with various Ags and were put in culture with B3Z in a 0.2 ml final volume at various numbers of APC per well.

B.1.5 Antigen presenting cells and sortings

Total spleen cells (TSC), low density (LDF) and high density (HDF) fractions were prepared according to the protocole of Steinman modified by Vremec et al. (Vremec et al., 1992). Briefly, spleens were digested with collagenase for 40 minutes at 37°C and then dilacerated and prepared in the continuous presence of EDTA 5mM. Cells were centrifugated on a dense BSA solution. Supernatant and pellet cells were collected appart and termed low density and high density fractions. CD11c staining was performed at 4°C in PBS supplemented with 5% of Fetal Calf Serum and 2mM EDTA (PBS-FACS) with the hamster HL3 mAb coupled to phycoerythrin (PE), fluoresceine isothiocyanate (FITC) or biotinylated and then revealed by Streptavidine-PE. CD8□ staining was performed with the 53-6.7 mAb coupled to PE. CD11b staining was performed with the M1/70 mAbs coupled to PE or FITC. CD54R staining was performed with the B220 mAb coupled to PE or biotinylated and revealed by streptavidine-PE. All mAbs originated from Pharmingen. After two washes, cells were sorted using a FACStar (Beckton Dickinson, Mountain View, USA). Cells were aseptically recovered in CM. Purity of the sorted cells was checked on an aliquot of the sorted cells analyzed on a FACScan apparatus (Beckton Dickinson, Mountain View, USA). Purity of the sorted cells was typically between 80 and 98%. In other experiments (as mentioned in figure legends), CD11c+ cells were sorted directly from spleen collagenase digests using the CD11c Micro Beads and the Magnetic Cell Sorting technology following the supplier protocole (MACS, MiltenyiBiotec, Bergish Gladbach, Germany). Purity of the sorted cells ranged around 80% with this technique.

B.2 Results

B.2.1 CD4- and CD40- independent CTL priming after systemic immunization with CyaAOVA in the absence of adjuvant

The chicken ovalbumin, H-2K^b restricted, SIINFEKL epitope was used as an experimental model epitope. It was genetically inserted in the catalytic domain of a detoxified, still invasive mutant CyaA. C57BL/6 (H-2^b) mice were immunized iv once with 50 µg of the recombinant toxin or control saline solution. Seven days after immunization, CTL activity specific for pOVA was detected within splenocytes of CyaAOVA-immunized C57BL/6 mice but not in mice injected with saline or a control CyaA (figure 6a). Similar results were obtained with CD4 or CD40 deficient mice indicating that, unlike many other CTL responses (as those raised against cross-priming Ag (Bennett et al, 1997; 1998; Schoenberger et al., 1998, Ridge et al., 1998) CD4⁺ T cell help was not mandatory for priming of CTL responses by CyaAOVA (figure 6b, c). Furthermore, B cells were not required since CTL responses were also obtained in B cell deficient mice (IgM^{-/-}, figure 6d). Contaminant LPS, possibly acting as adjuvants, are not involved in the stimulation of CTL responses CyaAOVA since C57BL10ScSn and the LPS-hyporesponsive mice C57BL10ScCr displayed similar OVA-specific CTL response after CyaAOVA injection (not shown).

B.2.2 In vitro and in vivo targeting of CyaAOVA presentation to CD11b-expressing cells

In order to better understand the immunogenicity of CyaAOVA, we intended to determine the APC involved in its presentation to CD8⁺ T cells. Using the IL-2 secretion as a readout for stimulation, we show that B3Z, an H-2K^b-restricted, anti-OVA CD8⁺ T cell hybridoma, is stimulated *in vitro* by bulk splenocytes in the presence of CyaAOVA but not CyaALCMV (figure 7a). We intended to analyze the APC ability of three well defined splenic APC: DC (CD11c⁺), B cells (CD45R⁺) and macrophages/granulocytes (CD11b^{high}+CD11c⁻). Due to the low percentage of CD11c⁺ in total splenocytes (<1%), we performed density fractionation and sorted the CD11c⁺ from the DC-enriched (4-10%) low density population by flow cytometry. The high density fraction contained only trace amounts of CD11c⁺ cells. In contrast, the distribution of CD45R⁺ and CD11b^{high}+CD11c⁻ (composed of granulocytes/macrophages) cells in both fractions allowed sortings on the total population for these markers. As shown in figure 7c, CyaAOVA was presented most efficiently by DC, less efficiently by CD11b^{high}+CD11c⁻ and B cells. This correlates with a quasi exclusive distribution of antigen presenting ability within the low density fraction of

splenocytes (figure 7a). In sharp contrast, both high and low density fractions were able to stimulate B3Z in response to pOVA (not shown).

To detect K^b-OVA complexes formed *in vivo* after immunization, the APC prepared from mice iv immunized 8-15 hrs before with 50 µg CyaAOVA were cocultured with B3Z *in vitro* without Ag addition (*ex vivo* assay). As for *in vitro* assays, APC responsible for CyaAOVA presentation were present exclusively in the DC-enriched low density fraction of splenocytes (figure 7b). The specificity of the assay was checked with APC from CyaALCMV-immunized mice that did not stimulated B3Z (figure 7b). In order to further characterize the low density APC involved in CyaAOVA presentation, we performed cell sortings: results shown in figure 7d show that DC (CD11c⁺) fraction was the more efficient APC. The macrophage/granulocyte (CD11b^{high}CD11c⁻) and the B cells (CD45R⁺) sorted from the same fraction were very inefficient for the stimulation of B3Z.

To further characterize APC involved in CyaAOVA, we performed subfractionation of the splenic low density CD11c⁺ DC in CD11c⁺CD8⁻ myeloid subset and CD11c⁺CD8⁺ lymphoid subset. *In vitro* and *ex vivo* assays (figure 7e and f, respectively) showed that antigen presenting ability for CyaAOVA was retained by the myeloid subset that also expressed CD11b⁺, unlike the lymphoid subset expressing low levels of this integrin (Pulendran *et al.*, 1997; Vremec *et al.*, 1997). The inability of the CD11c⁺CD8⁺ lymphoid subset to present CyaAOVA was specific for CyaAOVA since CD11c⁺CD8⁻ and CD11c⁺CD8⁺ presented equally well the pOVA synthetic peptide to B3Z.

In vitro and *ex vivo* assays performed with splenocytes from control (C57BL/6) or B cell-deficient mice (IgM^{-/-}) confirmed the poor contribution of B cells to CyaAOVA presentation *in vitro* and *in vivo* (figure 7g and h). In line with these results, CTL responses induced by CyaAOVA were not significantly affected in B cell-deficient mice as compared to control mice (figure 5d).

B.2.3 MHC-I-restricted presentation of CyaAOVA by dendritic cells depends on the cytosolic delivery of the recombinant toxin both in vitro and in vivo

To determine whether CyaAOVA presentation depends on the cytosolic delivery of the OVA epitope or extracellular loading, we performed Ag presentation assays with total splenocytes or CD11c⁺ DC from TAP^{-/-} or control TAP^{+/+} splenic purified from naive animals (*in vitro*, figure 8a) or from animals immunized iv with CyaAOVA (*ex vivo*, figure 8b). Results show that CyaAOVA presentation by dendritic cells is fully dependent on TAP either *in vitro* or *in vivo*. Since pOVA presentation is a TAP-independent phenomenon, we checked the functionality of TAP^{-/-} DC sorted from CyaAOVA-immunized mice by

stimulating B3Z with these cells loaded *in vitro* with the pOVA peptide (not shown). These results show that *in vivo* presentation of CyaAOVA depends on its cytosolic delivery.

B.2.4 Interaction of CyaA with CD11b is required for cell binding and delivery of the inserted antigen to the cytosolic pathway for antigen presentation by MHC I

5 We showed on part A that saturable and efficient binding of CyaA WT to CD11b⁺ cells can be blocked specifically by anti-CD11b mAbs. Moreover, CD11b transfection specifically conferred saturable binding and sensitivity to CyaA WT to otherwise CD11b⁻ cells resistant to CyaA WT. Cell binding blockade by anti-CD11b mAbs inhibited subsequent intracellular delivery of the catalytic adenylate cyclase domain, cAMP
10 elevation and cell death induced by CyaA WT. Since this results have been obtained on cell lines, it remained to determine if CyaA binds to splenocytes. We set up a flow cytometric assay to detect the fixation of a biotinylated, detoxified form of CyaA carrying the OVA peptide (CyaAOVA_{biotine}) to total splenocyte suspensions with streptavidin coupled to phycoerythrin. Using this assay, we observed that CyaAOVA binds to a subset
15 of leukocytes within the total splenocyte suspension (5-7%). Preincubation with the anti-CD11b M1/70 mAb but not a control mAb abrogated this binding (figure 9a). Furthermore, there is a correlation between the expression of CD11b and CyaAOVA binding to low density cells: CyaAOVA binds efficiently to CD11c⁺CD8α⁻ that expresses high levels of CD11b, less efficiently to CD11c⁺CD8α⁺ that expresses low levels of CD11b and very
20 weakly to CD11c⁻CD8α⁺ T cells that do not express CD11b (figure 9b). It is noticeable that CyaOVA binds efficiently to a low percentage of CD11c⁻CD8α⁻ cells in correlation with the presence of CD11b^{high+} into this CD11c⁻ population. Thus, CyaOVA biotine binding is mediated by CD11b (as for CyaA WT) and predicts the ability of a given cell type to present CyaOVA.

25 *In vitro*, we showed that the anti-CD11b mAb M1/70 blocks the CyaAOVA presentation by TSC cells to B3Z (figure 10a). This blockade is specific since i) a control mAb or mAbs specific for other $\alpha 2$ integrin family members (anti-CD11a, CD11c) had little or no effect (figure 10a and data not shown) ii) the presentation of pOVA was not affected by the anti-CD11b or none of these mAbs (figure 10b and data not shown). This confirms
30 that CD11b is the main receptor for CyaAOVA at least in the spleen, and that CyaAOVA-CD11b interaction is mandatory for the presentation of the inserted epitope.

Finally, to ascertain the role of CD11b expressing cells in CyaAOVA presentation, we performed sorting experiments on whole splenocyte cells from mice immunized with CyaAOVA or pOVA. Total splenocyte cells were sorted in CD11b⁺ and CD11b⁻ fractions.
35 Whereas the two subpopulations stimulated B3Z after pOVA iv immunization (figure 10d),

only the CD11b⁺ subpopulation stimulated B3Z after CyaAOVA iv immunization (figure 10c).

Taken together, these results clearly establish that the presentation of the OVA peptide from CyaOVA is dependent on cell binding and thus on interaction with CD11b.

5 **B.3 Discussion**

In the present study, using the detoxified adenylate cyclase of *Bordetella pertussis* as an epitope-delivery vector, we established a strategy for immunization that primes CTL responses after a single injection, bypassing the need for adjuvant requirement. We identified mechanisms that contribute to the high efficiency of detoxified CyaA as a vector :

10 **B.3.1 CyaA target myeloid dendritic cells through its interaction with the CD11b integrin**

Antigen presenting assay to a specific CD8⁺ T cell hybridoma using *in vitro*- or *in vivo*-loaded APC (*in vitro* and *ex vivo* assay, respectively) demonstrated that the most efficient APC for CyaAOVA are CD11c⁺ CD11b^{high+} DC. Indeed, all the Ag presenting ability for CyaAOVA belonged to the low density fraction of splenocytes that retains DC. Cell sorting of defined cell types revealed that CD11c⁺ CD11b^{high+} DC cells are much more efficient than CD11c⁺CD11b^{high+} cells to present CyAOVA. The minor contribution observed for B cells (CD45R⁺) was confirmed by efficient presentation of CyaAOVA (*in vitro* and *ex vivo*) and CTL responses in B cell-deficient mice.

20 **B.3.2 CyaA delivers Ag to the cytosolic pathway for MHC class I presentation in vivo**

Here we show the dependence of CyaAOVA presentation to total splenocytes *in vitro*. Strikingly, we also show that *in vivo* presentation of CyaAOVA takes also place according a TAP-dependent pathway. This lead to the conclusion that CyaAOVA presentation *in vivo* resulted effectively from cytosolic delivery and not from an eventual extracellular degradation.

B.3.3 CTL priming bypass CD4⁺ T cell help and is independent on CD40 signaling

Maturation from an immature stage toward a mature stage is characterized by i) a decrease in Ag capture ability, ii) an increase in T cell priming ability, iii) a migration from Ag sampling sites (marginal zone in the spleen) toward T cell area (peri arteriolar sheets in the spleen) were they maximize the probability of encounter with Ag-specific T cells (De smedt *et al.*, 1996). In addition to Ag presentation by DC, the maturation phase is now widely assumed as a prerequisite for T cell priming. *In vitro* studies have highlighted the role of CD4⁺ T cells in signalling DC maturation, notably via CD40L-CD40 interaction (Bell

et al., 1999). In the case of CD8⁺ T cells priming after the crosspriming of cellular Ag, CD4⁺ T cells dispensate their help to CD8⁺ T cells in a CD40-dependent mechanism (Schoenberger et al., 2000; Bennett et al., 1998; Schoenberger et al., 1998; Ridge et al., 1998). Since CyaAOVA primes CTL in a CD4 and CD40 independent way, it is tempting to speculate that detoxified CyaA could be endowed of intrinsic adjuvanticity.

B.3.4 Conclusion

The present study represents the first characterization, to our knowledge, of a proteinaceous vaccine vector that fits both targeting to professional APC, cytosolic delivery of the vectorized Ag and adjuvant-free CTL priming. Moreover, we elucidated the mechanism of cell targeting by demonstrating that Ag presentation is dependent on the interaction between CyaA and CD11b, its receptor. Hence, the cellular specificity of CyaA is serendipitously adapted to the Ag delivery purpose. Finally, the cellular specificity of CyaA or other bacterial toxins may serve the cytosolic delivery of a wide set of pharmaceutically-relevant molecules whose effects should be targeted on a restricted set of cells.

C. Use of adenylcyclase to deliver chemically coupled antigens to dendritic cells *in vivo*

C.1 Methods for coupling molecules of interest to CyaA-derived vectors

A general methodology is here described to create recombinant CyaA toxins by grafting molecules of interest to CyaA by means of disulfide bonds.

As an illustration, a synthetic 12 amino-acid long peptide corresponding to a CD8⁺ T-cell epitope from ovalbumin was chemically crosslinked through a disulfide bond to a cysteine residue genetically introduced into the CyaA catalytic domain at position 235 (wild type CyaA has no cysteine residues). The expected advantages of this novel architecture are:

- (i) versatility: a single CyaA carrier protein can be easily coupled to any desired synthetic peptides;
- (ii) immunogenicity: upon delivery into the APC cytosol, the epitopes chemically coupled to CyaA should be released from the vector (due to the reducing intracellular conditions) and introduced directly into the MHC class I presentation pathway, thus bypassing the potentially limiting step of proteolytic processing by proteasome.

The general procedure to couple synthetic peptides to CyaA by disulfide bonds is outlined in Figure 11.

In a first step, a recombinant CyaA toxin that contains a single cysteine residue genetically inserted within the catalytic domain of CyaA (wild type CyaA has no cysteine residue) is produced.

5 The recombinant CyaA toxin, ACTM235, has been previously characterized (Heveker and Ladant, 1997). In particular, ACTM235 harbors a Cys-Ser dipeptide inserted between amino-acid 235 and 236 and is fully cytotoxic. This toxin was expressed and purified to homogeneity as previously described in A.1.1.

10 In a second step a synthetic peptide corresponding to a CD8⁺ T cell epitope from ovalbumin was designed: in addition to the SIINFEKL (one letter code for amino acid) sequence that is the precise epitope sequence, a cysteine residue with an activated Nitro-pyridin-sulfonyl thiol group (Cys-NPys) was added at the N-terminus of the peptide during chemical synthesis. The activated Npys-cysteine was separated from the SIINFEKL sequence by a flexible GGA motif to facilitate further proteolytic processing of the peptide within APC. The peptide (Cys-Npys-OVA, molecular weight: 1405 da) was synthesized by
15 Neosystem (Strasbourg, France).

In a third step the Cys-Npys-OVA peptide was coupled to ACTM235 using the following procedure.

20 Ten mg of purified ACTM235 in 8 M urea, 20 mM Hepes-Na, pH 7.5, were reduced by incubation for 12 hrs in the presence of 10 mM Dithiothreitol (DTT). The efficiency of reduction was checked by an SDS-Page analysis under non reducing conditions: essentially all the ACTM235 protein migrated as a single band of 200 kDa corresponding to the monomeric species without evidence of any dimeric species (molecular weight of about 400 kDa). The reduced protein was then loaded on a DEAE-sepharose (Amersham Pharmacia Biotech) column (5 ml packed resin) equilibrated in 8 M urea, 20 mM Hepes-
25 Na, pH 7.5. The ACTM235 protein was fully retained on the DEAE-sepharose resin that was then extensively washed with 8 M urea, 20 mM Hepes-Na, pH 7.5, 0.1 M NaCl (> 100 ml) to remove any traces of DTT (the absence of residual DTT was checked using the classical Ellman's reaction with 5,5'-Dithio-bis-(2-nitrobenzoic acid), DTNB). The reduced ACTM235 protein was then eluted from the DEAE-sepharose resin in 7 ml of 8 M urea, 20
30 mM Hepes-Na, pH 7.5, 0.5 M NaCl. Three mg (about 2 μ mol) of Cys-Npys-OVA peptide were then added to the reduced ACTM235 protein (8 mg, about 45 nmol) and the mixture was incubated for 16 hrs at room temperature. Then, 25 ml of 20 mM Hepes-Na, pH 7.5 and 8 ml of 5 M NaCl were added and this diluted mixture was then loaded on a Phenyl-Sepharose (Amersham Pharmacia Biotech) column (10 ml packed resin) equilibrated in 20
35 mM Hepes-Na, pH 7.5, 1 M NaCl. The Phenyl-Sepharose resin was washed with 50 ml of

20 mM Hepes-Na, pH 7.5, 1 M NaCl and then with 50 ml of 20 mM Hepes-Na, pH 7.5. The derivatized ACTM235 protein was then eluted in 8 M urea, 20 mM Hepes-Na, pH 7.5. The toxin concentration was determined spectrophotometrically from the absorption at 280 nm using a molecular extinction coefficient of $142,000 \text{ M}^{-1} \cdot \text{cm}^{-1}$.

5 The Cys-Npys-OVA peptide was coupled using the same procedure to a second recombinant CyaA toxin, CyaAE5-LCMVgp, which is a detoxified variant (*i.e.* lacking the enzymatic activity as a result of the genetic insertion of 2 amino acid LQ between residues 188 and 189). This toxin also contains a 15 amino acid long polypeptide sequence (PASAKAVYNFATCGT) inserted between residues 224 and 225 of CyaA and that
10 contains a single Cys residues. The plasmid encoding this recombinant toxin is a derivative of pCACT-ova-E5 (Guermontprez *et al.* 2000) modified by the insertion between the *StuI* and *KpnI* restriction sites of an appropriate synthetic double stranded oligonucleotide encoding the PASAKAVYNFATCGT sequence. The CyaAE5-LCMVgp protein was expressed and purified as described previously in A.1.1.

15 The peptides shown in table 1 were also coupled similarly to another detoxified recombinant CyaA toxin, CyaAE5-CysOVA, which contains the same LQ dipeptide insertion in the catalytic site and a 14 amino acid sequence inserted between residues 224 and 225 of CyaA. This insert contains a Cys residue adjacent to the OVA epitope as shown in Figure 12. The plasmid encoding this recombinant toxin is a derivative of
20 pCACT-ova-E5 modified by the insertion between the *BsiWI* and *KpnI* restriction sites of an appropriate synthetic double stranded oligonucleotide encoding the ASCGSIIINFEKLG sequence. The CyaAE5-CysOVA protein was expressed and purified as described previously in A.1.1.

25 The CyaAE5-CysOVA can be considered as a general detoxified vector for chemical coupling of CTL epitopes by disulfide bridges. The presence of the OVA epitope within CyaAE5-CysOVA allows for an easy *in vitro* assay for functionality in epitope delivery by measuring the presentation of the OVA epitope to specific T-cell hybridoma as described previously in B.1.4.

TABLE 1	
NPys CTL peptides coupled to CyaAE5-CysOVA	
Name of epitopes	Amino-acid sequence of peptides
CEA 571	Cys(NPys)-GGYLSGANLNL
Gp100	Cys(NPys)-GGITDQVPFSV

MelanA	Cys(NPys)-GGEAAGIGILTV
Tyrosinase	Cys(NPys)-GGYMDGTMSQV

Alternatively, the thiol groups of recombinant CyaA toxins can be activated with 2,2'-dithiodipyridine (Sigma) and derivatised with peptides containing a reduced cysteine (the procedure to reduce the Cys in synthetic peptides is provided by the manufacturer).
 5 This would be especially appropriate if the desired peptide contains an internal cysteine residue.

C.2 Analysis of the *in vitro* and *in vivo* immunogenicity of the OVA epitope chemically coupled to CyaA

In vitro delivery of the OVA epitope to MHC class I molecules by CyaA after
 10 chemical (CyaA-gp-S-S-OVA E5) or genetic coupling (CyaA-OVA E5) was first analysed by studying the presentation of these molecules to the anti-OVA B3Z CD8⁺ T cell hybridoma by splenocytes. 3.10⁵ spleen cells from C57BL/6 mice were co-cultured for 18 h with 10⁵ B3Z cells in the presence of various concentrations of the recombinant CyaA. The IL-2 release by B3Z was measured in a CTLL proliferation assay.

15 As shown on the figure 13, a high IL-2 secretion was observed when B3Z was stimulated either with CyaA-gp-S-S-OVA E5 or CyaA-OVA E5, thus demonstrating that the OVA epitope chemically linked to CyaA was as efficiently delivered to the cytosolic pathway of the antigen-presenting cells than after genetic coupling. No IL-2 production was observed when B3Z was stimulated with CyaA molecules lacking the OVA epitope,
 20 thus showing the specificity of the IL-2 production.

In a second step, *in vivo* capacity of these molecules to induce OVA-specific CTL responses was then analysed. C57BL/6 mice (2 per group) were immunized by i.v. injection of 50 µg of the various CyaA. Seven days later, 25.10⁶ spleen cells of individual
 25 syngenic spleen cells for five days. The cytotoxic activity of the effector cells was measured on 51 Cr-labeled EL4 target cells pulsed or not with 50 µM of the OVA peptide.

As shown in figure 14, high CTL responses were induced in mice immunized by either CyaA-gp-S-S-OVA E5 or CyaA-OVA E5, thus demonstrating the comparable efficacy of chemical or genetic coupling of the OVA epitope. No CTL response was
 30 observed in mice immunized with the control CyaA molecules lacking the OVA epitope or when uncoated EL4 were used as target cells, thus showing the specificity of the CTL responses observed.

In conclusion, these results clearly demonstrate that a CD8⁺ T cell epitope chemically linked to CyaA-derived proteinaceous vectors is very efficiently delivered to the cytosolic pathway for MHC class I presentation and induces strong CTL response *in vivo*.

D. Identification of a CyaA-derived fragment that binds to CD11b/CD18 and can be used as efficient molecule delivery vectors.

As an attempt to map the region that is involved in the interaction between CyaA and the CD11b/CD18 receptor, various subfragments of CyaA were constructed and tested for their ability to compete with a biotinylated CyaA toxin for binding to CD11b/CD18 on the surface of transfected CHO cells, using methods previously described in paragraph A.

D.1 Expression of CyaA373-1706

CyaA373-1706 protein was produced in *E. coli* by using a novel expression vector pTRAC-373-1706 (fig.16). It is a derivative of plasmid pDL1312 (Ladant 1995), constructed by replacing the neurocalcin gene of plasmid pDL1312 by the *cyaC* gene (that encodes CyaC which is involved in the conversion of proCyaA into the active toxin by post-translational palmitoylation of Lys 860 and 983 of CyaA) and the 3' part of the *cyaA* gene that encodes the C-terminal domain - codons 373 to 1706- of CyaA (see fig. 16). Both *cyaC* and '*cyaA* genes are placed in the same transcriptional unit under the control of the λ phage Pr promoter. The 3' end of the *cyaC* gene was modified to introduce before its stop codon, a ribosome binding site to enhance the translation initiation of the downstream '*cyaA* gene (Fig. 16). The resulting modification of the CyaC polypeptide (the last 3 amino acid Gly-Thr-Ala at the C-terminus of CyaC were replaced by the Asn-Arg-Glu-Glu sequence) had no effect on its ability to acylate CyaA. The 5' end of the *cyaA* gene (coding for the catalytic domain of the toxin (upstream of the unique *Bst*BI site of *cyaA*) was deleted and replaced by an appropriate synthetic double stranded oligonucleotide encoding the MGCGN sequence (Fig. 16).

The pTRAC-373-1706 also encodes the thermosensitive λ repressor C1⁸⁵⁷ that strongly represses gene transcription at the λ Pr promoter at temperatures below 32° C, the origin of replication of colE1 and the beta-lactamase gene that confers ampicillin resistance.

Expression of the CyaA373-1706 protein was carried out in *E. coli* strain BLR. Cells transformed with pTRAC-373-1706 were grown at 30°C in LB medium containing 150 mg/L of ampicillin until mid-log phase and then synthesis of CyaC and of the truncated CyaA was induced by increasing the growth temperature to 42°C. Bacteria were harvested

after 3-4 hrs of further growth at 42°C. CyaA373-1706 protein was then purified as described for the wild type CyaA (Guermonez et al. 2000). CyaA373-1706 is devoid of cAMP synthesizing activity, it exhibits hemolytic activity of sheep erythrocytes and contains also a unique cysteine residues in its MGCGN N-terminal sequence.

5 **D.2 Inhibition of CyaA-E5 binding to CD11b by CyaA-E5 and CyaA fragments: 1-383 (catalytic domain) and 373-1706 (Hydrophobic and repeat domains)**

As shown on figure 15, a strong inhibition of biotinylated CyaA-E5 binding was achieved after incubation of transfected CHO cells with CyaA-E5. A similar inhibition was obtained after incubation of these cells with the fragment CyaA-373-1706 whereas CyaA 10 1-383 had no significant effect on the binding of the biotinylated CyaA-E5.

Thus, these results clearly demonstrate that the fragment of CyaA that encompasses residues 373 to 1706, CyaA373-1706 , contains the structures essentially required for the interaction of CyaA with the CD11b/CD18 receptor.

E. Conclusions

15 Since MHC class I molecules usually present peptides derived from endogenously synthesized proteins, epitopes must be delivered to the cytosol of antigen presenting cells to elicit CTL responses. It has been previously established that the CTL priming activity of recombinant ACT protein relies at least in part on its ability to deliver CD8⁺ T cell epitopes into the cytosol of antigen-presenting cells (APC). In one approach, epitopes were 20 genetically inserted within the catalytic domain of CyaA as it is known that, for the wild-type toxin, this part of the polypeptide reaches the cytosol of target cells where it exerts its toxic effect (*i.e.* cAMP synthesis). After entering the cells, the CyaA catalytic domain harboring the epitope insert must be proteolytically processed to release the matured CD8⁺ T cell epitope which then will be translocated to the endoplasmic reticulum to associate 25 with MHC class I molecules.

This proteolytic processing, carried out by the proteasome, might be a limiting step in numerous cases (Pamer and Cresswell, 1998; Cascio *et al.*, 2001), leading to a significant decrease of the overall yield of the mature epitope. The alternative approach, described here, that consist in the linkage of the epitopes to the catalytic domain of CyaA 30 by means of a disulfide bond, offers a significant advantage in that, with this design, matured epitopes (requiring only N-terminal trimming) will be quantitatively released from CyaA inside the cytosol of APC. In addition, this should be a very versatile system as a single CyaA carrier protein could be easily and rapidly coupled to any desired synthetic peptide. Furthermore, it will be easy to introduce by genetic engineering, several cysteine

residues within the catalytic domain (such as in previously mapped permissive sites, WO 93/21324 (INSTITUT PASTEUR)) of a recombinant detoxified CyaA toxin so that multiple peptides can be coupled on the same CyaA molecule. This should increase the overall epitope delivery and immunogenicity of the recombinant protein.

5 Deletion mapping allowed to identify the C-terminal part of CyaA (aa 373-1706) as the region that is involved in the interaction of the toxin with its CD11b/CD18 receptor. The truncated protein CyaA373-1706 can be therefore used as a protein module to target CD11b⁺ cells *in vivo*. In particular, polypeptides or proteins corresponding to antigens of interest can be genetically fused to CyaA373-1706 to be delivered to dendritic cells in
10 order to elicit specific immune responses. Similar coupling can be performed on the recombinant CyaA373-1706 protein that also contained a unique cysteine at its N terminal end.

The term "comprise" and variants of the term such as "comprises" or "comprising" are used herein to denote the inclusion of a stated integer or stated
15 integers but not to exclude any other integer or any other integers, unless in the context or usage an exclusive interpretation of the term is required.

Any reference to publications cited in this specification is not an admission that the disclosures constitute common general knowledge in Australia.

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The claims defining the invention are as follows:

1. An immunogenic composition formulated for administration in an animal or a human host characterized in that it comprises a *Bordetella* adenylcyclase which comprises an antigen inserted in the catalytic domain.
- 5 2. A pharmaceutical composition for administration in a human or an animal formulated for targeting a molecule of interest specifically to CD11b expressing cells characterized in that said molecule of interest is coupled to a *Bordetella* species adenylcyclase.
3. The immunogenic or pharmaceutical composition according to claim 1 or claim
10 2, wherein the adenylcyclase is a genetically modified adenylcyclase.
4. The immunogenic or pharmaceutical composition according to claim 3, wherein the genetically modified adenylcyclase is able to translocate the molecule of interest specifically in the cytosol of CD11b expressing cells.
5. The immunogenic or pharmaceutical composition according to claim 4, wherein
15 said genetically modified adenylcyclase is a *Bordetella* adenylcyclase lacking all or part of its catalytic N-terminal domain.
6. The immunogenic or pharmaceutical composition according to claim 4, wherein said genetically modified adenylcyclase is the *Bordetella pertussis* adenylcyclase lacking residues 1-373.
- 20 7. The immunogenic or pharmaceutical composition according to claim 3 or claim 4, wherein said genetically modified adenylcyclase comprises one or more cysteine residue(s) inserted within the catalytic domain at a permissive site.
8. The immunogenic or pharmaceutical composition according to any one of
25 claims 1 to 7, wherein said CD11b expressing cells are dendritic cells and more preferably myeloid dendritic cells.
9. The immunogenic or pharmaceutical composition according to any one of claims 1 to 7, wherein said CD11b expressing cells are neutrophils.

10. The pharmaceutical or immunogenic composition according to any one of claims 1 to 9, wherein the molecule of interest is selected from: peptides, glycopeptides, lipopeptides, polysaccharides, oligosaccharides, nucleic acids, lipids or chemicals.
- 5 11. The pharmaceutical or immunogenic composition according to any one of claims 1 to 9, wherein said composition comprises a nucleic acid construct encoding the *Bordetella* species adenylate cyclase coupled to a molecule of interest.
- 10 12. The pharmaceutical or immunogenic composition according to claim 10, wherein the molecule of interest is a drug.
13. The pharmaceutical composition according to claim 12, wherein the drug is an anti-inflammatory drug.
14. The pharmaceutical or immunogenic composition according to any one of claims 1 to 10, wherein the molecule of interest is an antigen.
- 15 15. The immunogenic or pharmaceutical composition according to claim 14, wherein the antigen is selected from the group consisting of: an intracellular bacterial cell antigen, a tumoral cell antigen, a viral antigen, a fungus antigen or a parasite cell antigen.
- 20 16. The immunogenic or pharmaceutical composition according to claim 15, wherein the antigen is selected from the group consisting of: a poliovirus antigen, an HIV virus, an influenza virus antigen, a choriomeningitis virus antigen, a tumor antigen.
- 25 17. The immunogenic or pharmaceutical composition according to any one of claims 14 to 16, wherein said antigen is fused to the N-terminal part of a genetically modified *Bordetella* adenylcyclase lacking its N-terminal catalytic domain.
18. The immunogenic or pharmaceutical composition according to any one of claims 14 to 17, wherein said antigen is fused to the genetically modified *Bordetella pertussis* adenylcyclase lacking residues 1-373.

19. The immunogenic or pharmaceutical composition according to any one of claims 1 to 18, wherein said composition is formulated for intravenous administration and especially is devoid of priming adjuvants.
20. The immunogenic composition according to any one of claims 1 to 19, wherein said immunogenic composition is capable of inducing or stimulating an immune response involving specifically dendritic cells.
21. Use of the immunogenic or pharmaceutical composition according to any one of claims 1 to 18 in the preparation of a vaccine or an immunotherapeutic composition.
22. A proteinaceous vector for delivery of a molecule of interest, specifically to CD11b expressing cells, characterized in that said vector comprises a recombinant *Bordetella* species adenylcyclase coupled to said molecule of interest.
23. The proteinaceous vector according to claim 22, wherein said vector is also able to deliver the molecule of interest specifically in the cytosol of CD11b expressing cells.
24. The proteinaceous vector according to claim 22 or claim 23, wherein said CD11b expressing cells are dendritic cells and more preferably myeloid dendritic cells.
25. The proteinaceous vector according to claim 22 or claim 23, wherein said CD11b expressing cells are neutrophils.
26. The proteinaceous vector according to any one of claims 22 to 25, wherein the adenylcyclase is a genetically modified adenylcyclase.
27. The proteinaceous vector according to claim 26, wherein said genetically modified adenylcyclase is devoid of native cysteine residue but contains genetically inserted cysteine residue(s) at permissive site(s) within the catalytic domain.

28. The proteinaceous vector according to claim 26 or claim 27, wherein the adenylyclase is a non toxic adenylyclase.
29. The proteinaceous vector according to any one of claims 26 to 28, wherein the genetically modified adenylyclase is a *Bordetella* adenylyclase lacking all or part of its N-terminal catalytic domain.
30. The proteinaceous vector according to any one of claims 22 to 29, wherein the adenylyclase is from *Bordetella pertussis*.
31. The proteinaceous vector according to claim 30, wherein said recombinant adenylyclase is the *Bordetella pertussis* adenylyclase lacking residues 1-373.
- 10 32. The proteinaceous vector according to any one of claims 22 to 31, wherein said molecule of interest is selected in the group comprising: peptides, glycopeptides, lipopeptides, polysaccharides, oligosaccharides, nucleic acids, lipids and chemicals.
- 15 33. The proteinaceous vector according to any one of claims 22 to 32, wherein said molecule of interest is a drug chemically or genetically coupled to said adenylyclase.
34. The proteinaceous vector according to claim 33, wherein the molecule of interest is an epitope.
- 20 35. The proteinaceous vector according to claim 33 or claim 34, wherein said molecule of interest is an epitope chemically coupled to a genetically modified adenylyclase devoid of native cysteine residue but containing genetically inserted cysteine residue(s) at permissive site(s) within its catalytic domain.

Date: 25 July 2007

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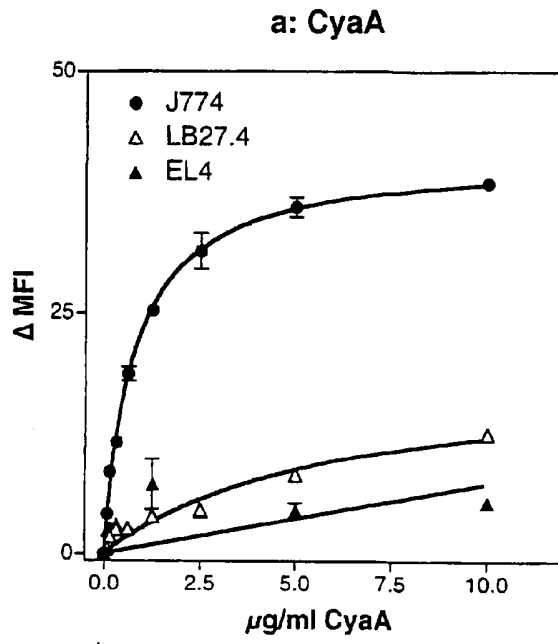


FIG. 1A

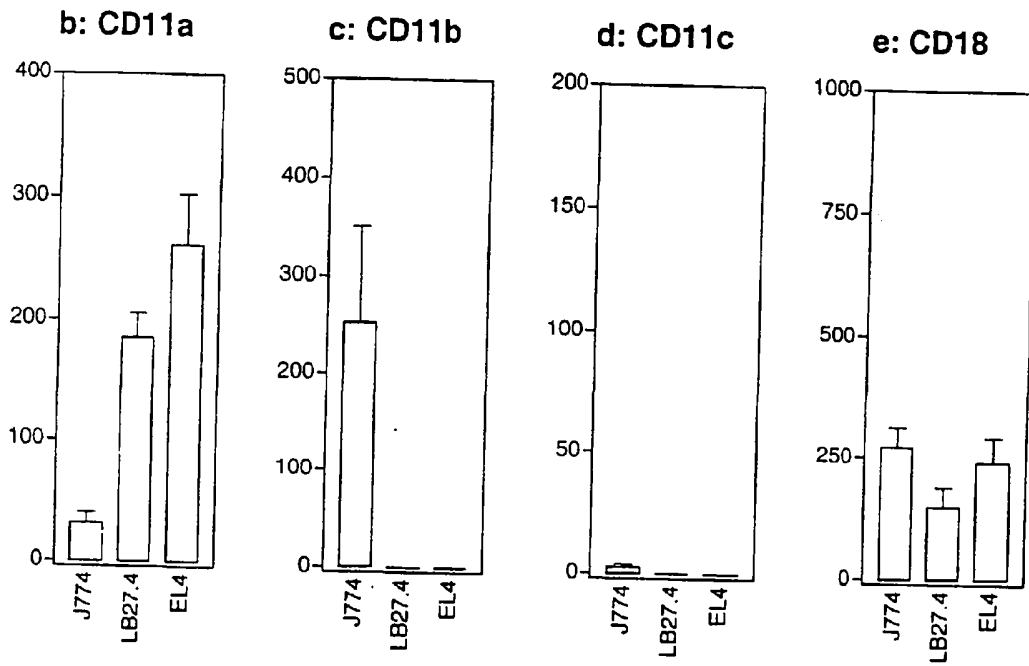
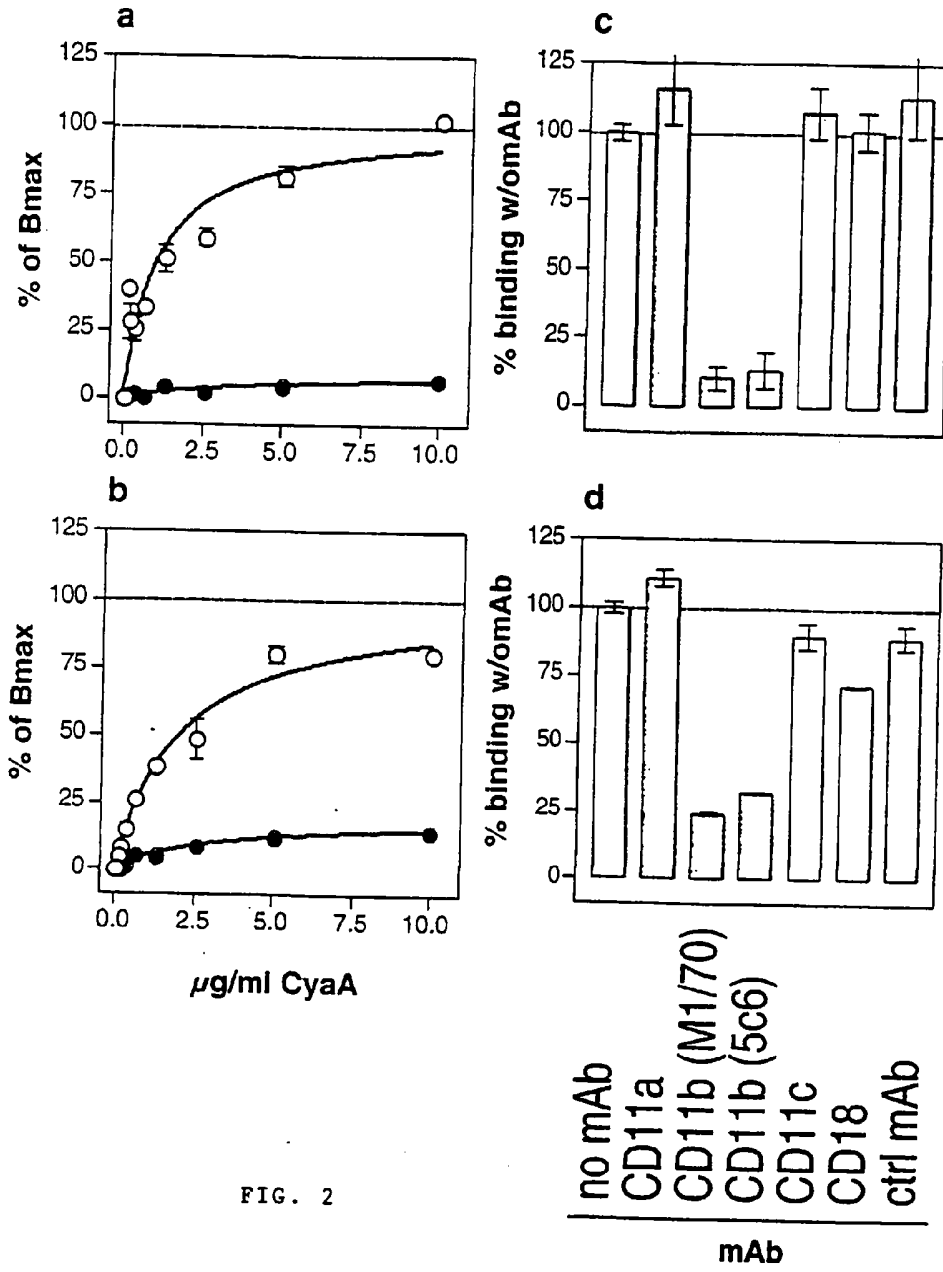


FIG. 1B



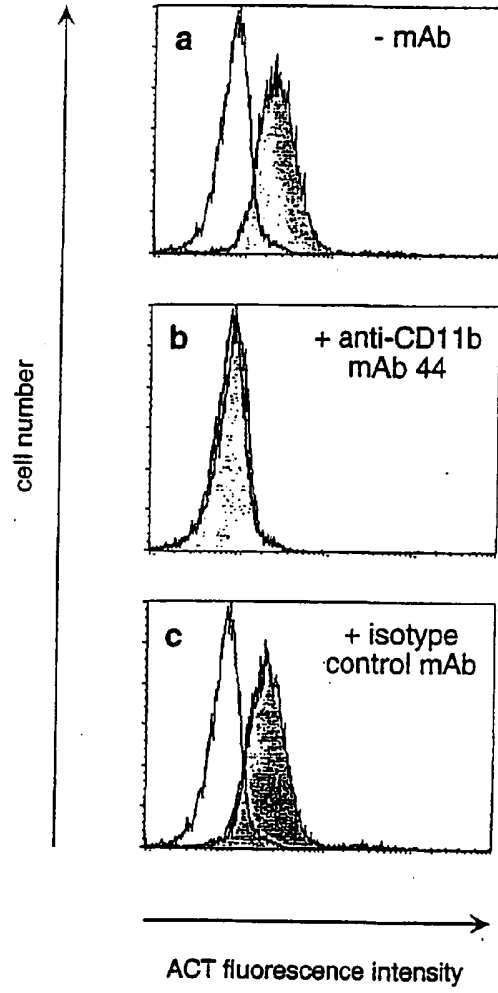


FIG. 3A

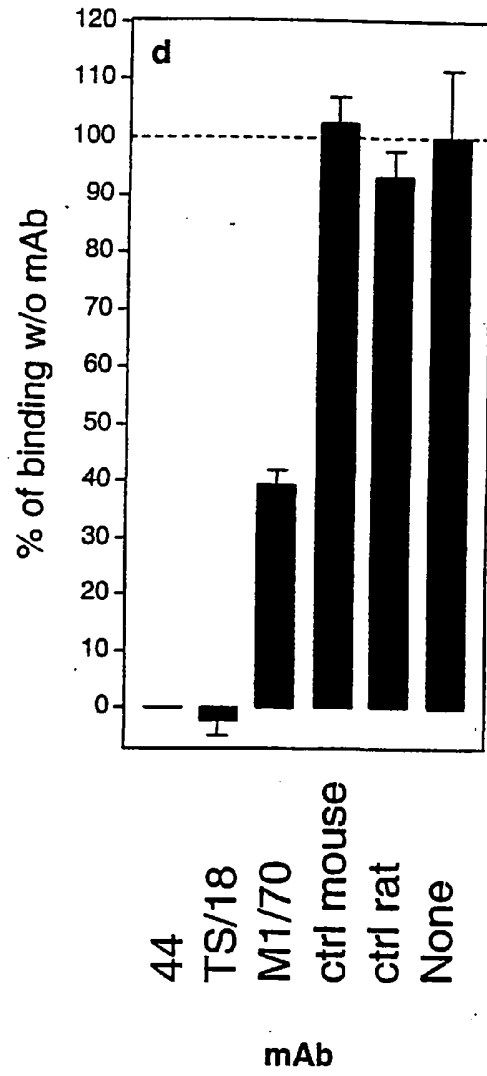


FIG. 3B

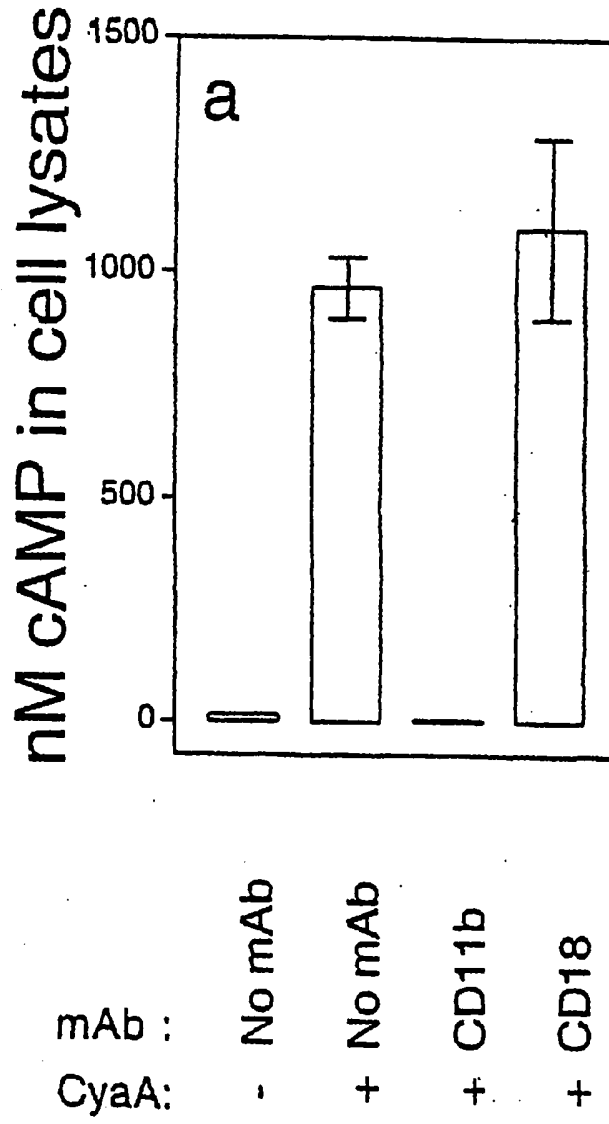


FIG. 4A

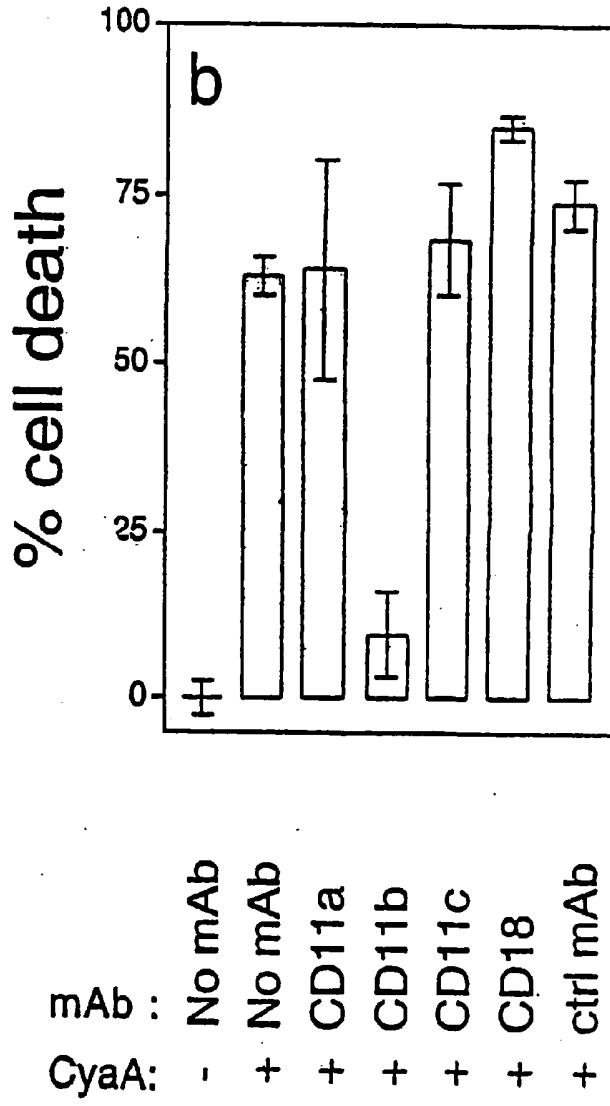


FIG. 4B

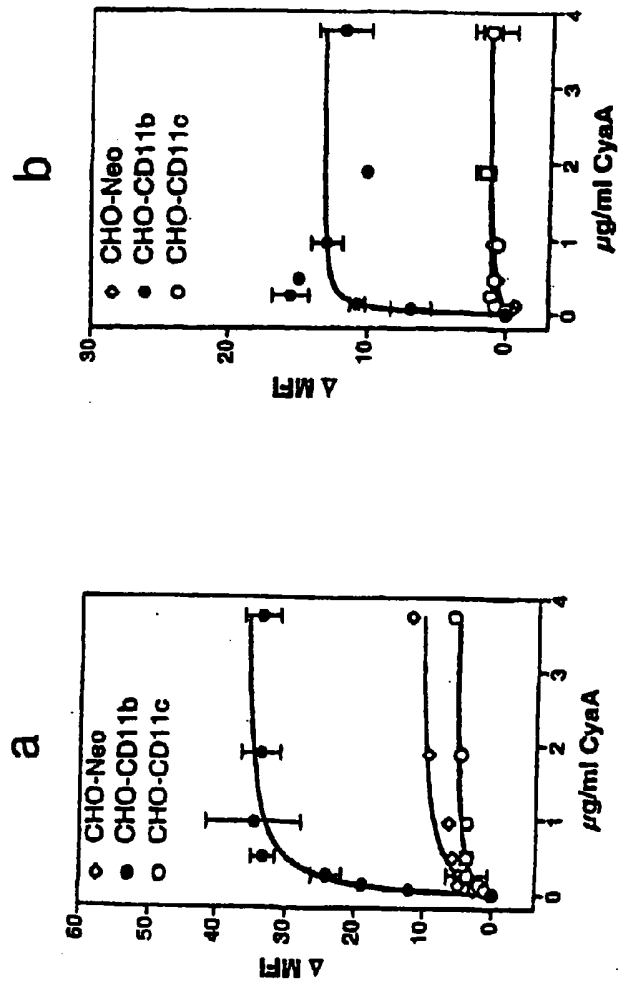


FIG. 5A

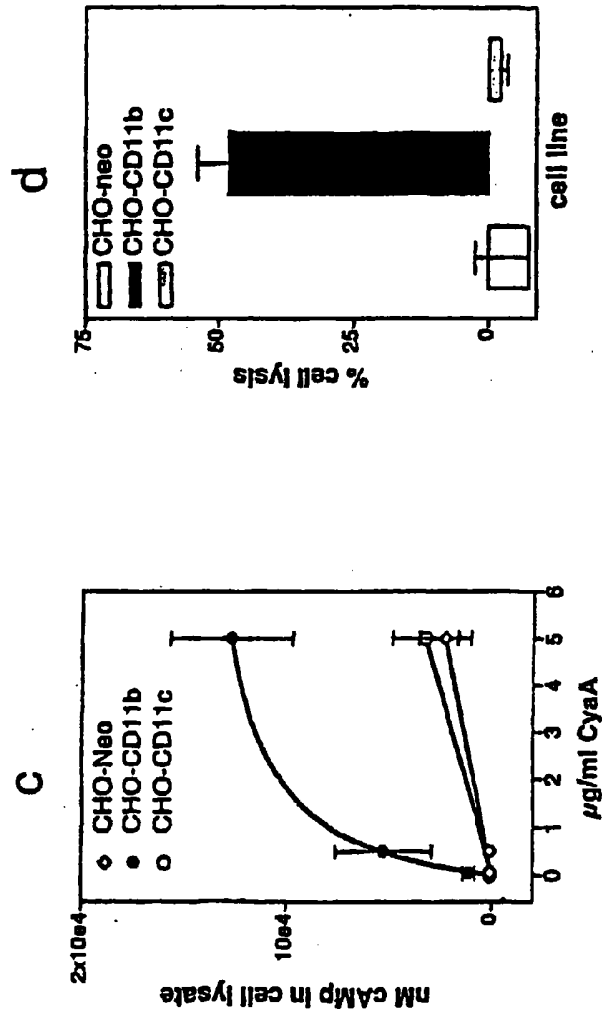


FIG. 5B

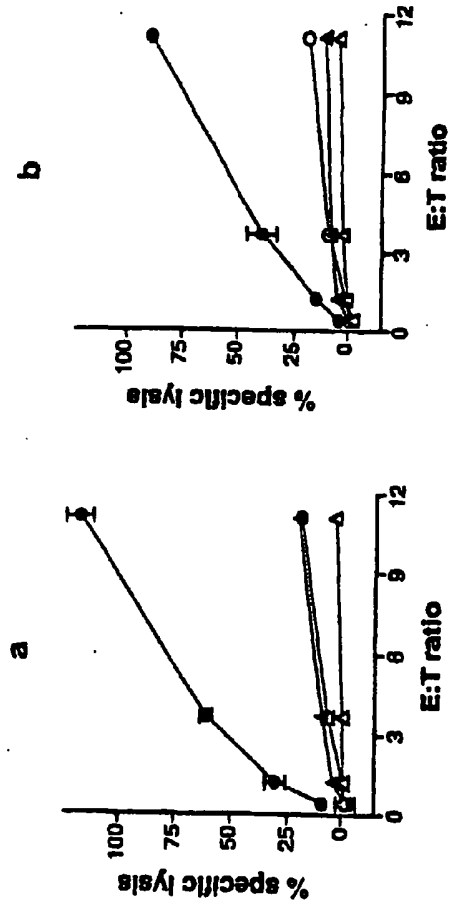


FIG. 6A

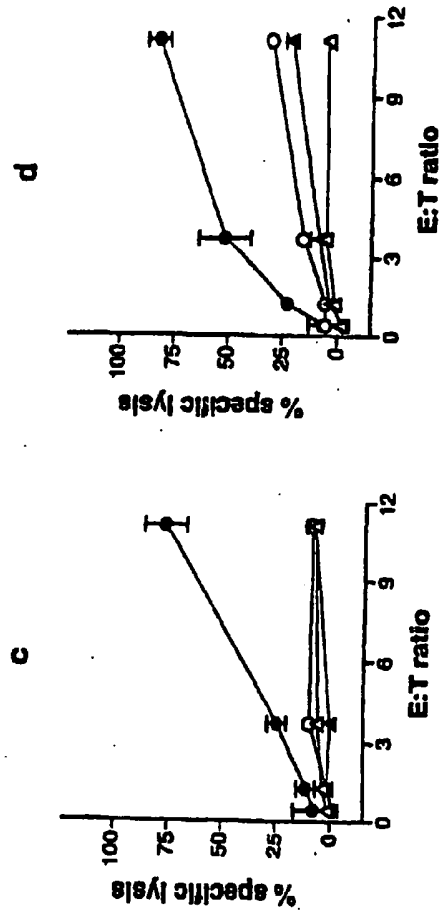


FIG. 6B

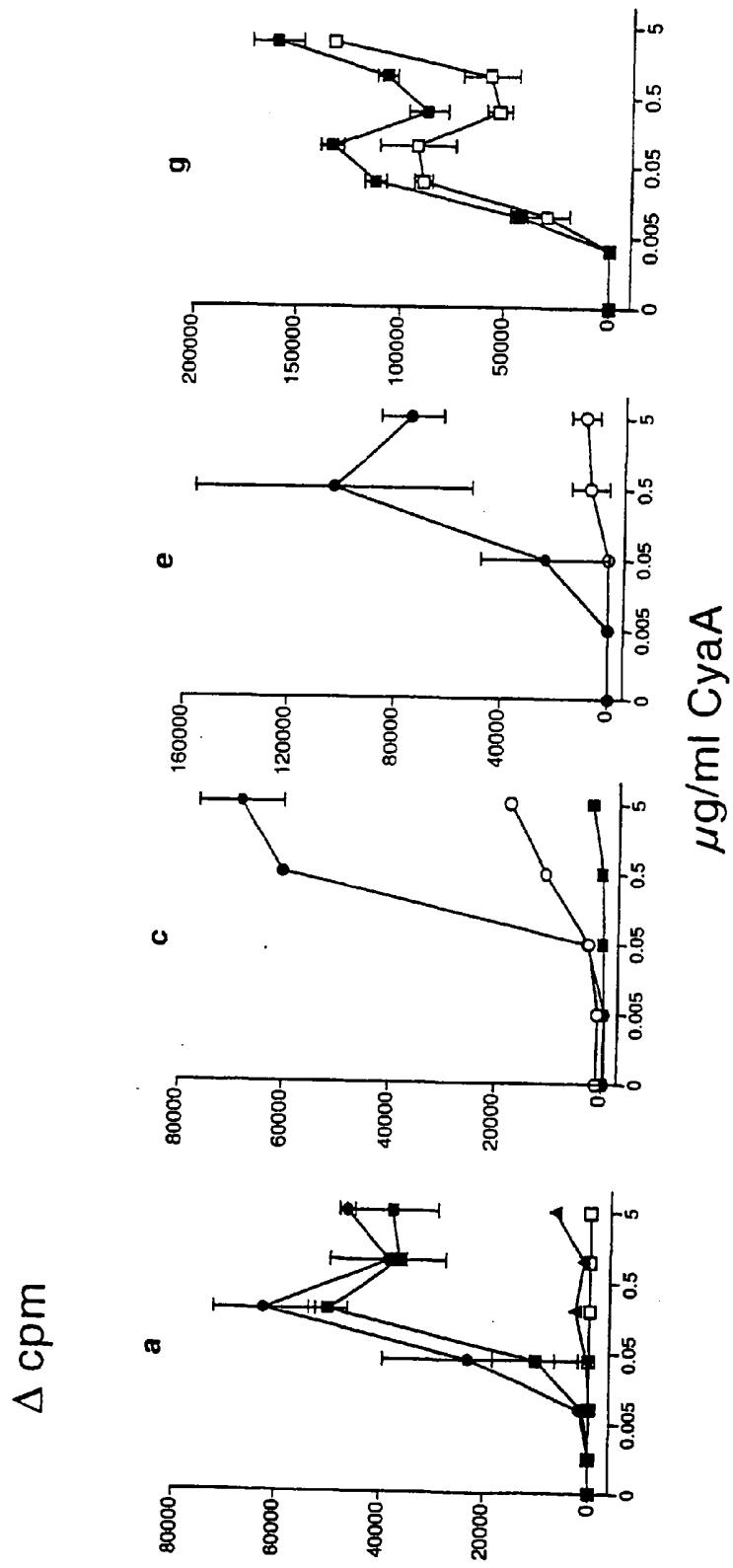


FIG. 7A

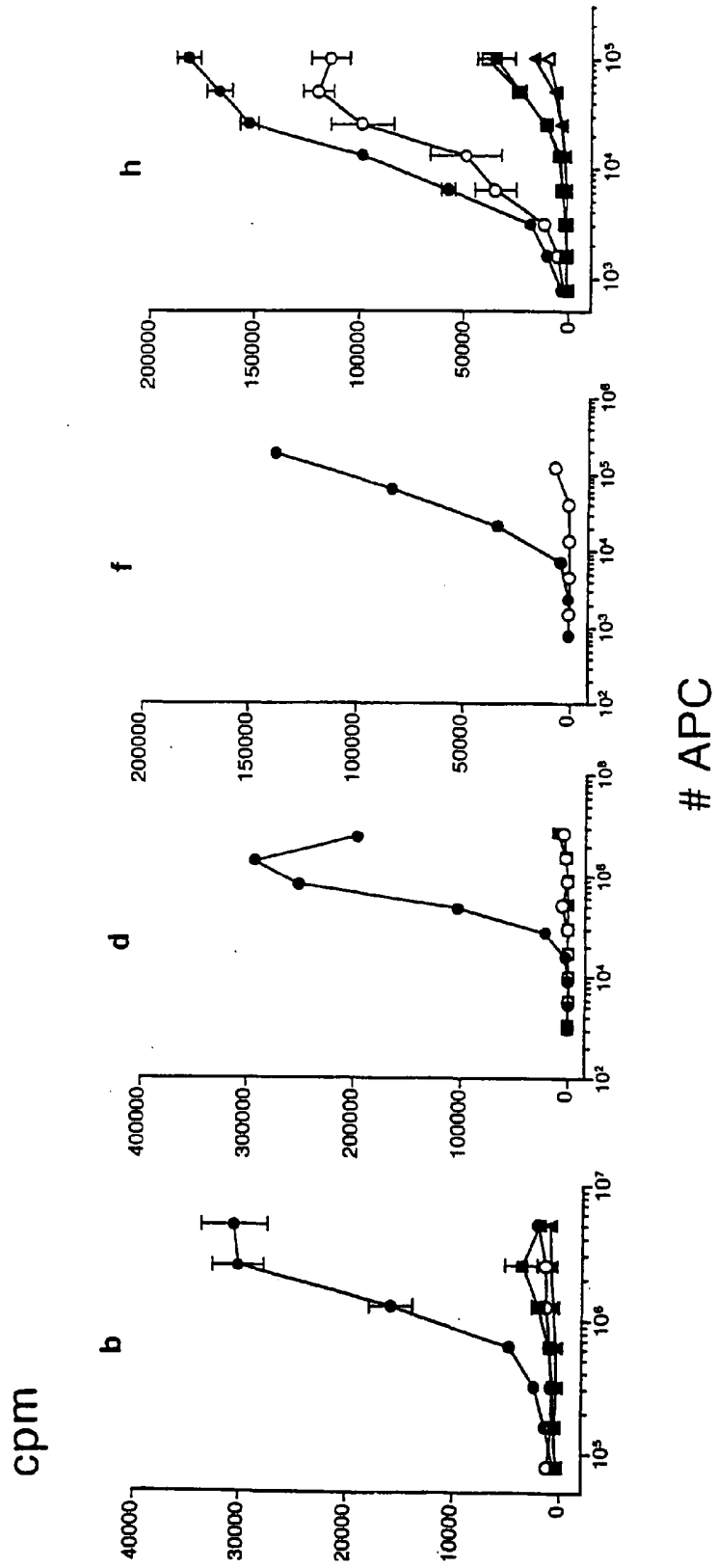


FIG. 7B

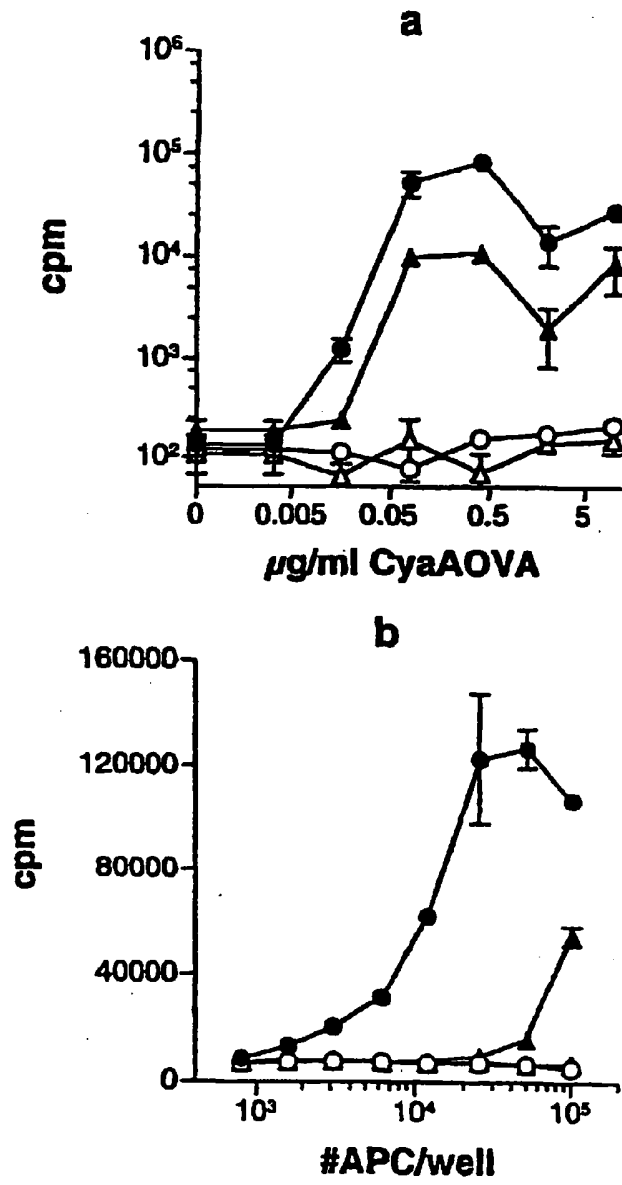


FIG. 8

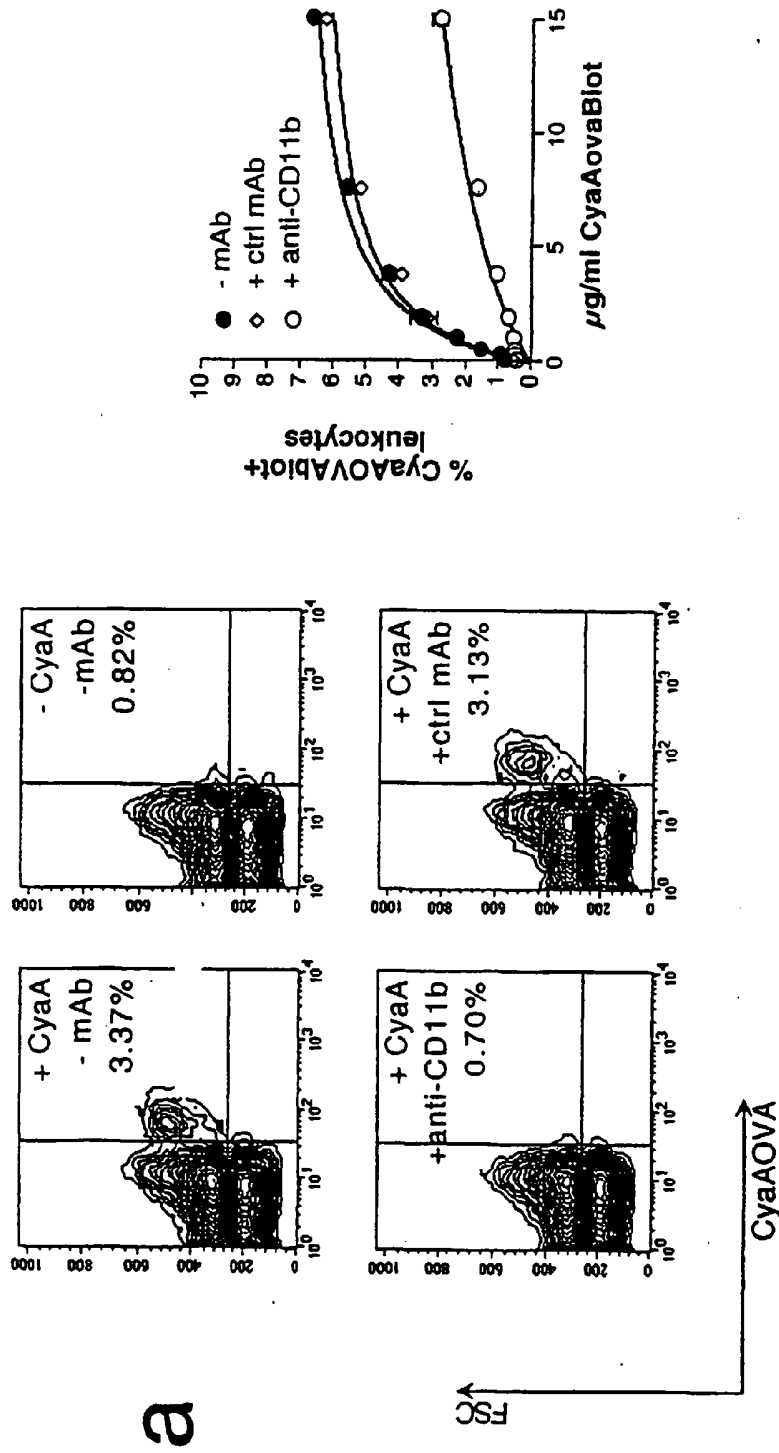


FIG. 9a

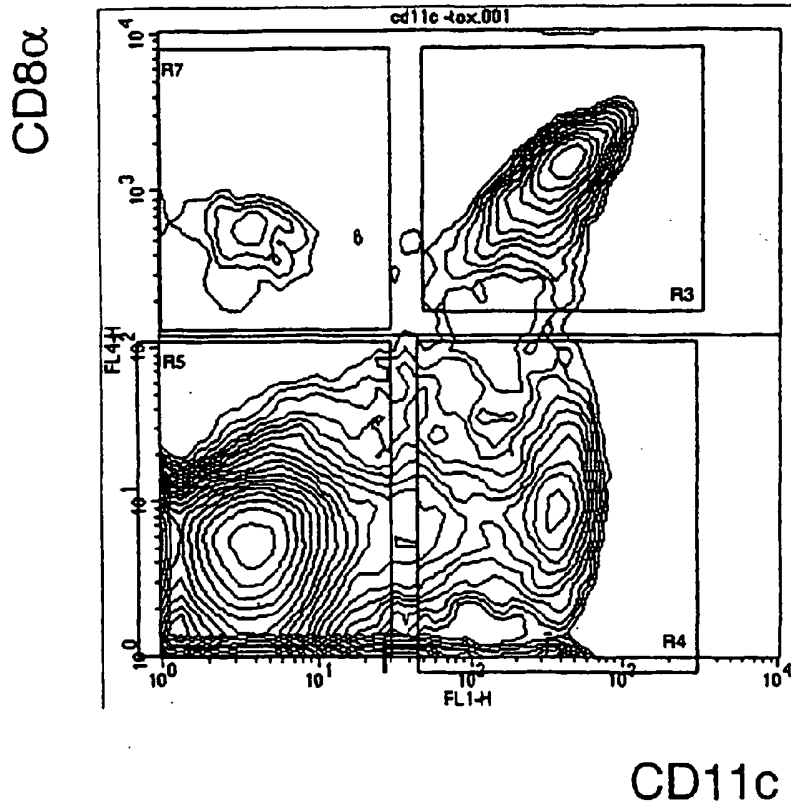
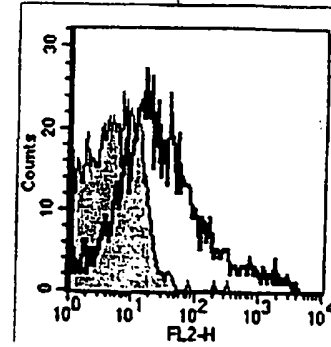
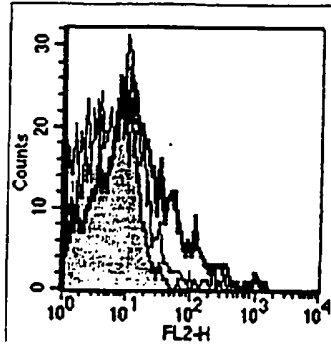
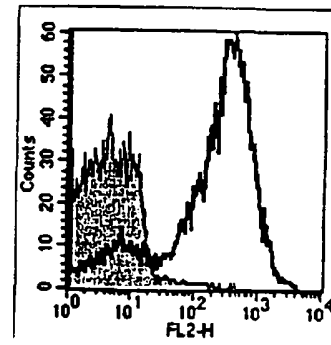
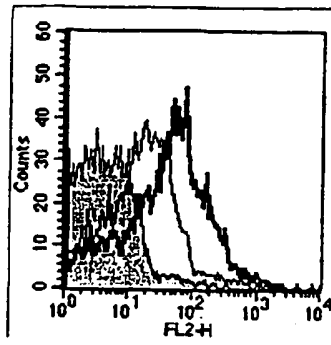


FIG. 9b(1)

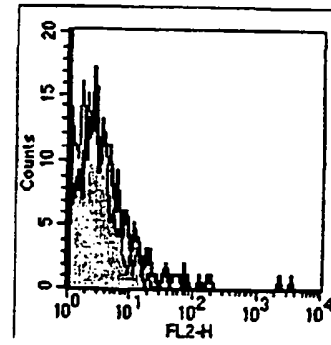
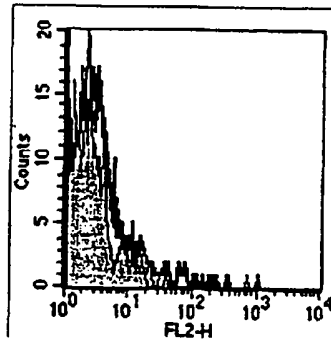
11c+CD8 α +



11c+CD8 α -



11c-CD8 α +



11c-CD8 α -

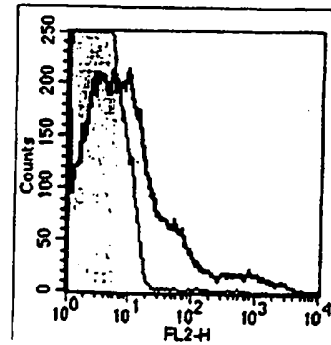
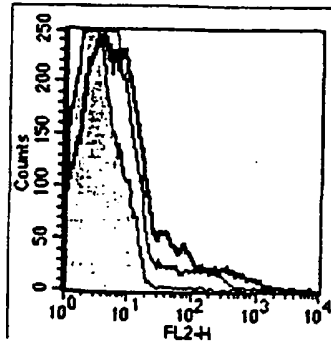
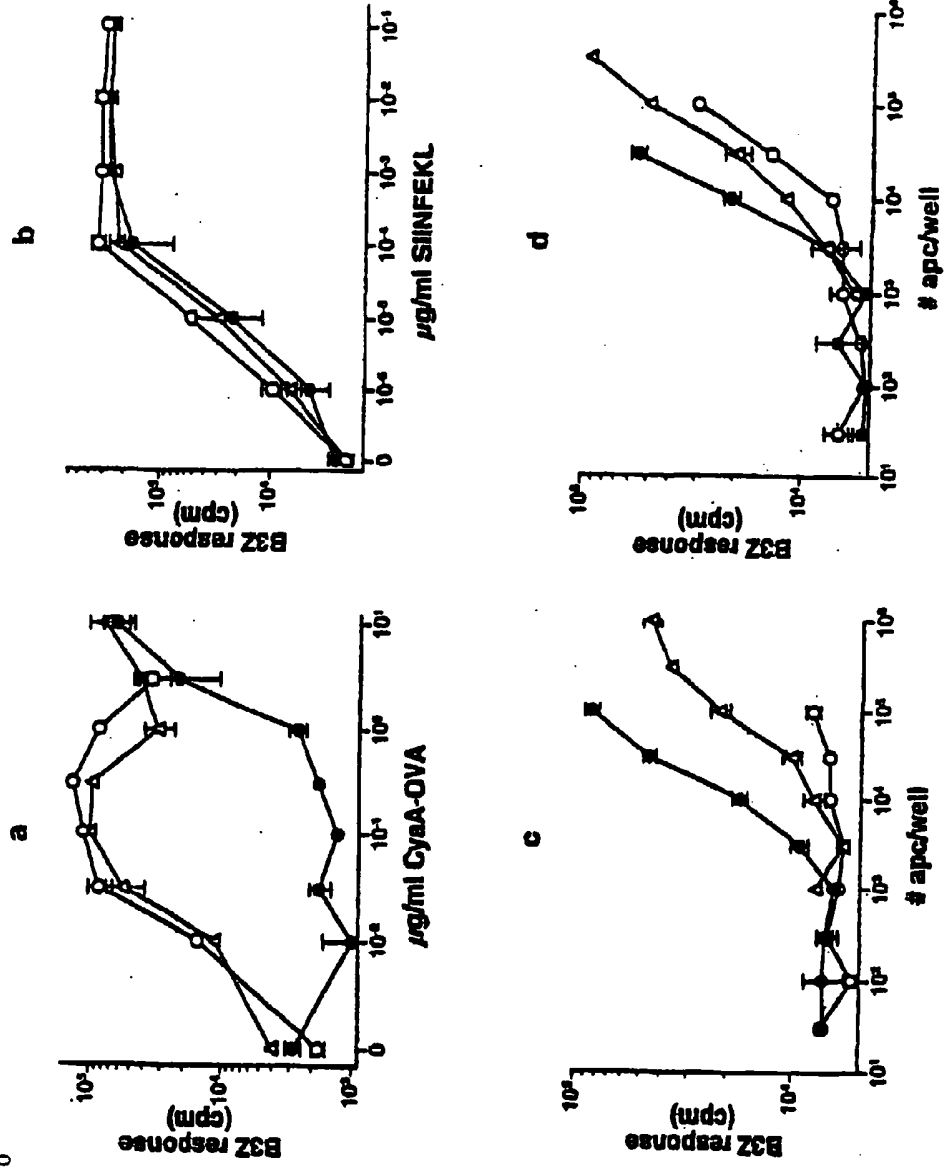


FIG. 9b(2)

CyaAOVA

CD11b

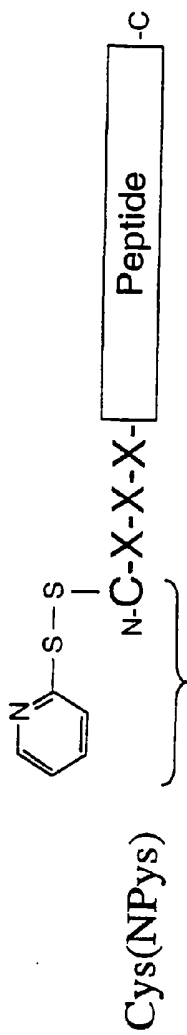
FIG. 10



1 - Insertion of Cys in CyaA (devoid of Cys residue)



2 - Chemical Synthesis of Cys(NPys)-peptide



3 - Crosslinking Cys(NPys)-peptide to CyaA-Cys

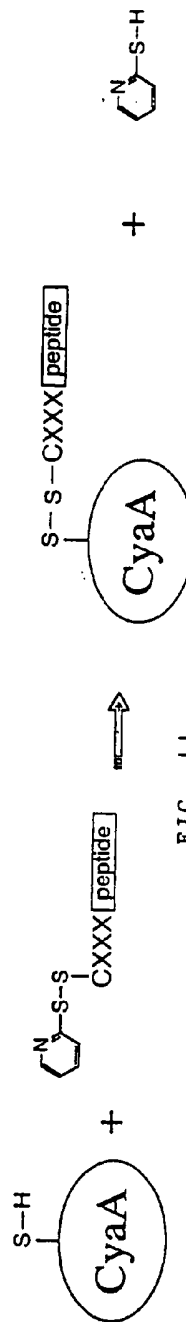
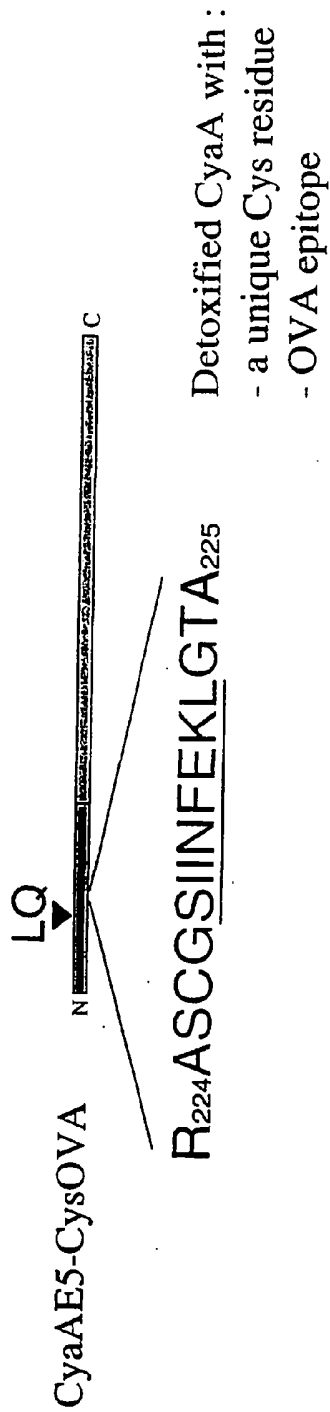


FIG. 11



 **Efficient presentation of OVA to B3Z *in vitro***
&
Selective binding to CD11b cells *in vitro*

 **Chemical coupling of NPys-peptides
HPDP-biotin
& proteins**

FIG. 12

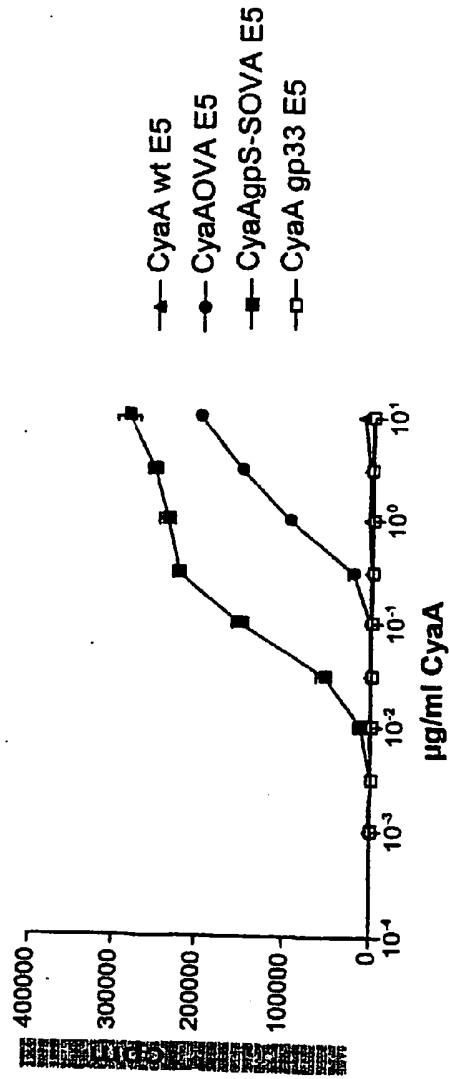


FIG. 13

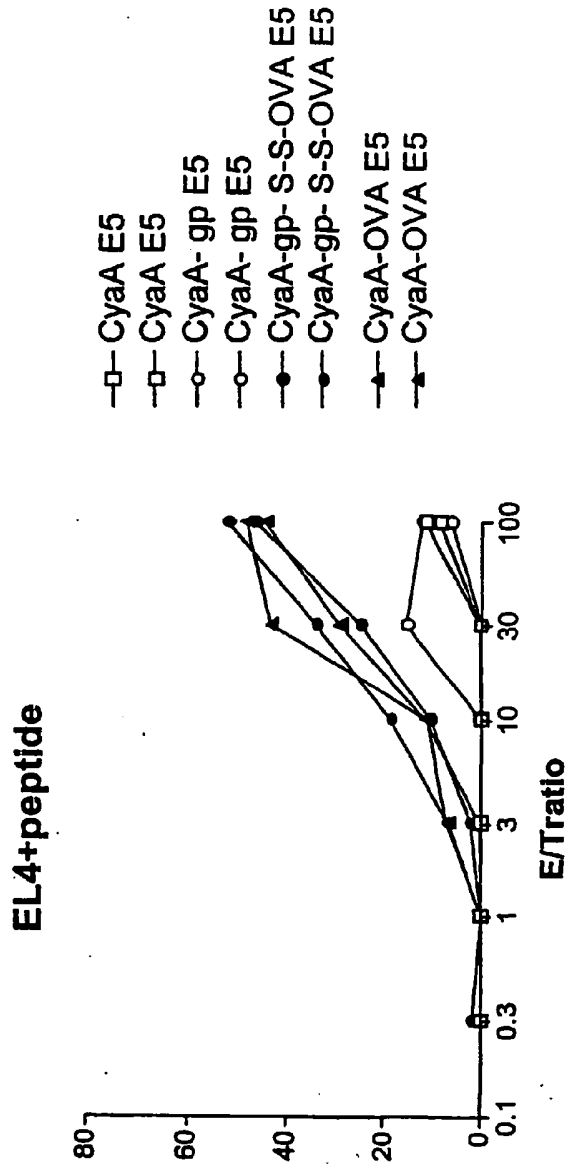


FIG. 14A

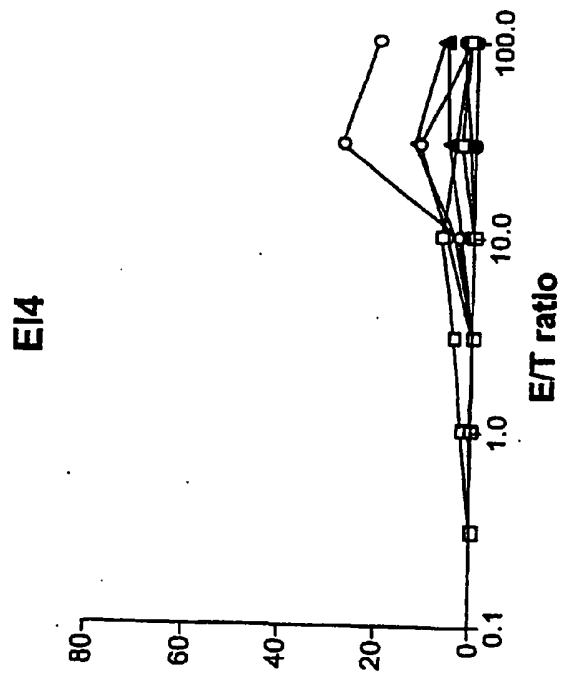


FIG. 14B

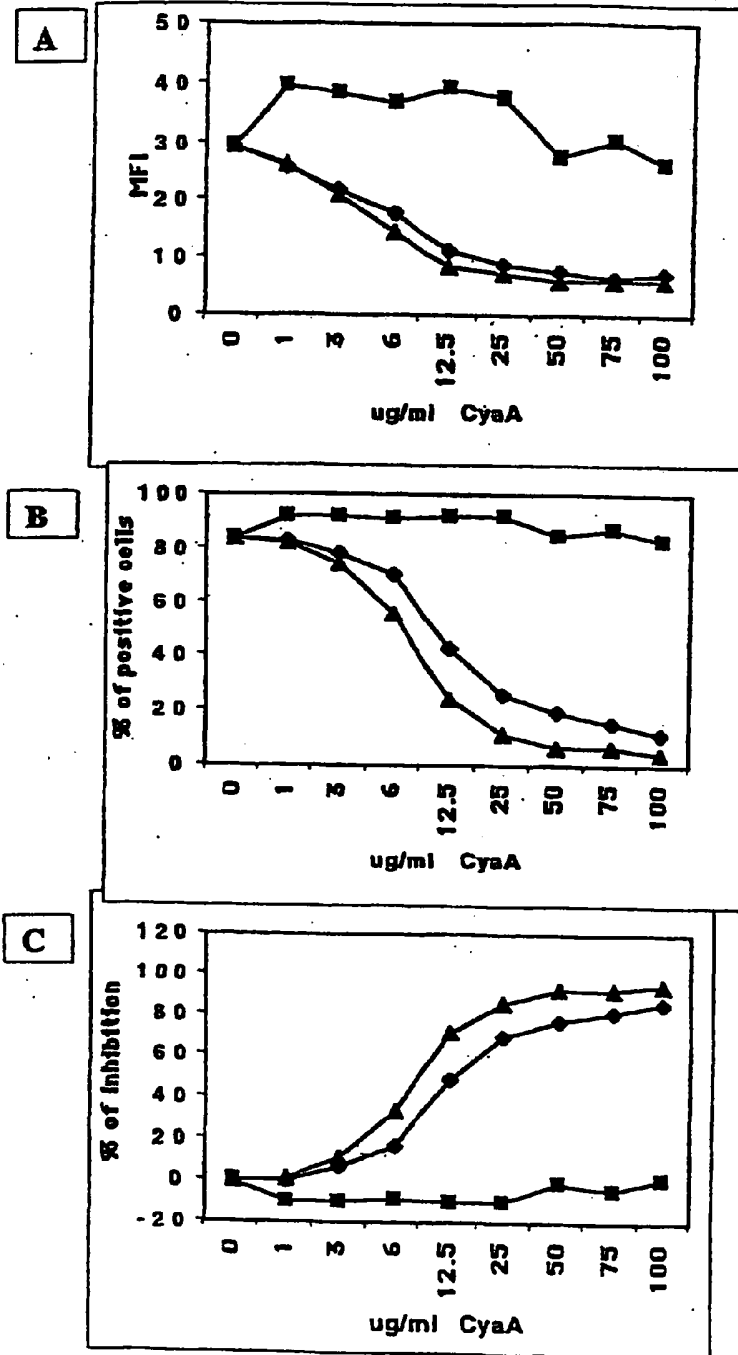


FIG. 15

pTRAC-373-1706

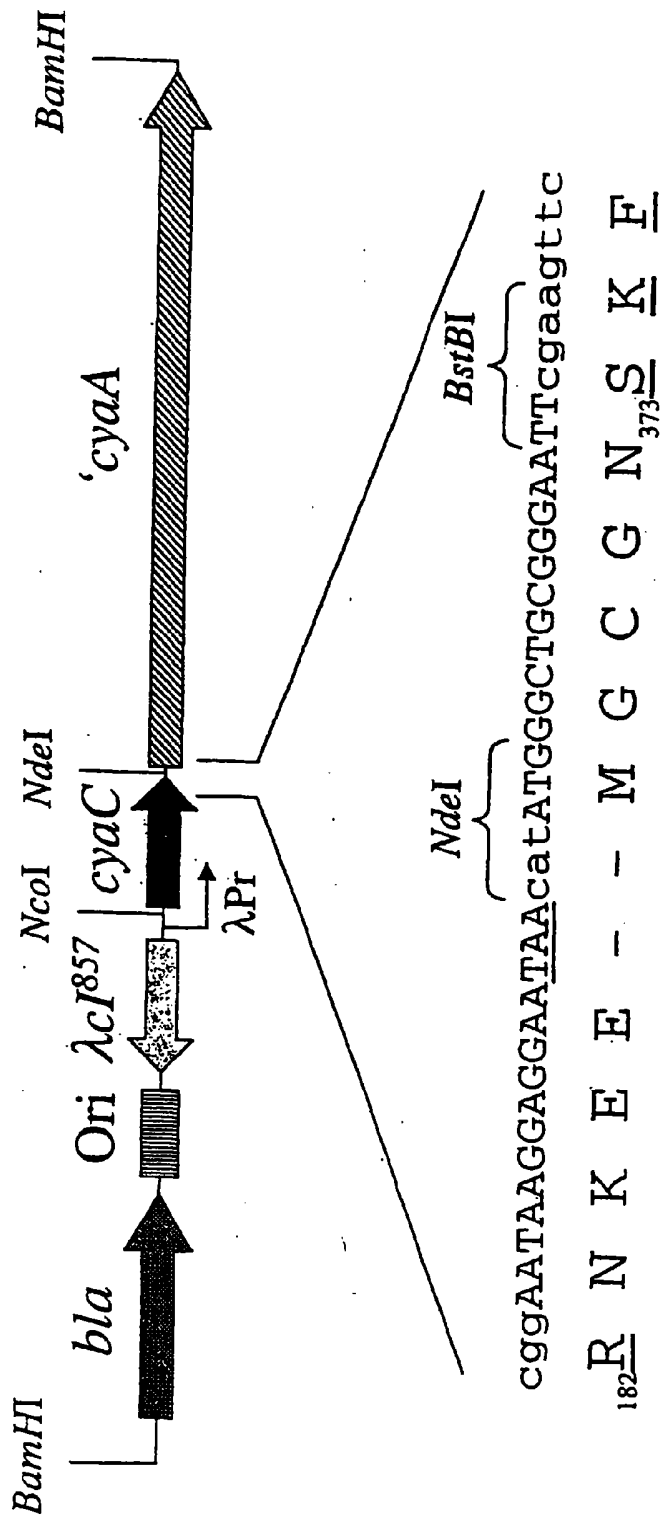


Figure 16