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(54) Titre : PROTEINE RV2386C TUBERCULOSIS, COMPOSITIONS ET UTILISATIONS CORRESPONDANTES
(54) Title: THE TUBERCULOSIS RV2386C PROTEIN, COMPOSITIONS AND USES THEREOF

(57) **Abrégé/Abstract:**

The present invention is directed to a polypeptide which comprises: (i) an Rv2386c protein sequence; (ii) a variant of an Rv2386c protein sequence; or (iii) an immunogenic fragment of an Rv2386c protein sequence. In other aspects the invention is directed to associated polynucleotides, fusion proteins and methods for the treatment or prevention of tuberculosis.

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(54) Title: THE TUBERCULOSIS RV2386C PROTEIN, COMPOSITIONS AND USES THEREOF

(57) Abstract: The present invention is directed to a polypeptide which comprises: (i) an Rv2386c protein sequence; (ii) a variant of an Rv2386c protein sequence; or (iii) an immunogenic fragment of an Rv2386c protein sequence. In other aspects the invention is directed to associated polynucleotides, fusion proteins and methods for the treatment or prevention of tuberculosis.

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THE TUBERCULOSIS RV2386C PROTEIN, COMPOSITIONS AND USES THEREOF

FIELD OF THE INVENTION

5 The present invention relates to polypeptides and polynucleotides for use in the treatment or prevention of tuberculosis, in particular for use in the treatment or prevention of latent tuberculosis and in the prevention or delay of reactivation of tuberculosis (and also to related methods). The present invention further relates to pharmaceutical and immunogenic compositions comprising said polypeptides and polynucleotides, and to methods for the
10 diagnosis of tuberculosis (in particular latent tuberculosis).

BACKGROUND OF THE INVENTION

Tuberculosis (TB) is a chronic infectious disease caused by infection with
15 *Mycobacterium tuberculosis* and other *Mycobacterium* species. It is a major disease in developing countries, as well as an increasing problem in developed areas of the world. More than 2 billion people are believed to be infected with TB bacilli, with about 9.2 million new cases of TB and 1.7 million deaths each year. 10% of those infected with TB bacilli will develop active TB, each person with active TB infecting an average of 10 to 15 others per
20 year. While annual incidence rates have peaked globally, the number of deaths and cases is still rising due to population growth (World Health Organisation *Tuberculosis Facts* 2008).

Mycobacterium tuberculosis infects individuals through the respiratory route. Alveolar macrophages engulf the bacterium, but it is able to survive and proliferate by inhibiting
25 phagosome fusion with acidic lysosomes. A complex immune response involving CD4+ and CD8+ T cells ensues, ultimately resulting in the formation of a granuloma. Central to the success of *Mycobacterium tuberculosis* as a pathogen is the fact that the isolated, but not eradicated, bacterium may persist for long periods, leaving an individual vulnerable to the later development of active TB.

30 Fewer than 5% of infected individuals develop active TB in the first years after infection. The granuloma can persist for decades and is believed to contain live *Mycobacterium tuberculosis* in a state of dormancy, deprived of oxygen and nutrients. However, recently it has been suggested that the majority of the bacteria in the dormancy state are located in non-
35 macrophage cell types spread throughout the body (Locht et al, *Expert Opin. Biol. Ther.* 2007 7(11):1665-1677). The development of active TB occurs when the balance between the host's natural immunity and the pathogen changes, for example as a result of an immunosuppressive

event (Anderson P *Trends in Microbiology* 2007 15(1):7-13; Ehlers S *Infection* 2009 37(2):87-95).

5 A dynamic hypothesis describing the balance between latent TB and active TB has also been proposed (Cardana P-J *Inflammation & Allergy – Drug Targets* 2006 6:27-39; Cardana P-J *Infection* 2009 37(2):80-86).

10 Although an infection may be asymptomatic for a considerable period of time, the active disease is most commonly manifested as an acute inflammation of the lungs, resulting in tiredness, weight loss, fever and a persistent cough. If untreated, serious complications and death typically result.

15 Tuberculosis can generally be controlled using extended antibiotic therapy, although such treatment is not sufficient to prevent the spread of the disease. Infected individuals may be asymptomatic, but contagious, for some time. In addition, although compliance with the treatment regimen is critical, patient behaviour is difficult to monitor. Some patients do not complete the course of treatment, which can lead to ineffective treatment and the development of drug resistance.

20 Multidrug-resistant TB (MDR-TB) is a form which fails to respond to first line medications. 5% of all TB cases are MDR-TB, with an estimated 490,000 new MDR-TB cases occurring each year. Extensively drug-resistant TB (XDR-TB) occurs when resistance to second line medications develops on top of MDR-TB. It is estimated that 40,000 new cases of the virtually untreatable XDR-TB arise annually (World Health Organisation *Tuberculosis Facts* 2008).

25 Even if a full course of antibiotic treatment is completed, infection with *M. tuberculosis* may not be eradicated from the infected individual and may remain as a latent infection that can be reactivated.

30 In order to control the spread of tuberculosis, effective vaccination and accurate early diagnosis of the disease are of utmost importance.

35 Diagnosis of latent TB infection is commonly achieved using the tuberculin skin test, which involves intradermal exposure to tuberculin protein-purified derivative (PPD). Antigen-specific T cell responses result in measurable induration at the injection site by 48-72 hours after injection, which indicates exposure to mycobacterial antigens. Sensitivity and specificity have, however, been a problem with this test, and individuals vaccinated with BCG cannot always be

- easily distinguished from infected individuals (this is particularly important in light of the fact that BCG does not protect against latent infection). In general, individuals who have received BCG but are not infected by *M. tuberculosis* show a PPD reaction below 10 mm in diameter whereas people who have a PPD reaction above 10 mm in diameter are considered to have
- 5 been infected by *M. tuberculosis*. However, this rule is not applicable to individuals with immunosuppression due to HIV infection, which may result in a PPD reaction below 10 mm in diameter); or in endemic countries, where people infected by non-tuberculosis mycobacteria can show a PPD reaction above 10 mm in diameter.
- 10 Progress over recent years has seen the development of *in vitro* T cell based assays, based on interferon-gamma release and using antigens which are more specific to *M. tuberculosis* than PPD, namely ESAT-6 and CFP-10. These high specificity tests appear to be at least as sensitive as the tuberculin skin test and also demonstrate less cross-reactivity due to BCG vaccination. See Pai M et al *Expert Rev. Mol. Diagn.* 2006 6(3):413-422 for a recent review of
- 15 latent TB diagnosis. However, since ESAT-6/CFP-10 are early stage antigens, assays based on ESAT-6/CFP-10 may only perform optimally in recently infected people. Consequently, the identification of new antigens specifically associated with latent tuberculosis may aid the development of more sensitive assays that could detect longer-term latent infections.
- 20 There remains a need for effective strategies for the treatment and prevention of tuberculosis, in particular the treatment and prevention of latent TB and the prevention of reactivation of TB.

BRIEF SUMMARY OF THE INVENTION

- 25 The present invention relates generally to the identification of Rv2386c as a TB antigen (in particular an antigen associated with latent TB) and to related methods and uses in the prevention and treatment of TB, especially the prevention and treatment of latent TB and the prevention or delay of TB reactivation.
- 30 The present invention provides an isolated polypeptide which comprises:
- (i) an Rv2386c protein sequence;
 - (ii) a variant of an Rv2386c protein sequence; or
 - (iii) an immunogenic fragment of an Rv2386c protein sequence.
- 35 The present invention also provides a polypeptide which comprises:
- (i) an Rv2386c protein sequence;
 - (ii) a variant of an Rv2386c protein sequence; or

(iii) an immunogenic fragment of an Rv2386c protein sequence;
for use as a medicament.

A further aspect of the invention relates to a method for the treatment or prevention of TB
5 comprising the administration of a safe and effective amount of a polypeptide comprising:
(i) an Rv2386c protein sequence;
(ii) a variant of an Rv2386c protein sequence; or
(iii) an immunogenic fragment of an Rv2386c protein sequence;
to a subject in need thereof, wherein said polypeptide induces an immune response, in
10 particular an immune response against *Mycobacterium tuberculosis*.

The use of a polypeptide comprising:
(i) an Rv2386c protein sequence;
(ii) a variant of an Rv2386c protein sequence; or
15 (iii) an immunogenic fragment of an Rv2386c protein sequence;
in the manufacture of a medicament for the treatment or prevention of TB, represents another
aspect of the invention.

The present invention provides an isolated polynucleotide comprising a nucleic acid sequence
20 encoding a polypeptide which comprises:
(i) an Rv2386c protein sequence;
(ii) a variant of an Rv2386c protein sequence; or
(iii) an immunogenic fragment of an Rv2386c protein sequence.

25 Also provided is a polynucleotide comprising a nucleic acid sequence encoding a polypeptide
which comprises:
(i) an Rv2386c protein sequence;
(ii) a variant of an Rv2386c protein sequence; or
(iii) an immunogenic fragment of an Rv2386c protein sequence;
30 for use as a medicament.

A further aspect of the invention relates to a method for the treatment or prevention of TB
comprising the administration of a safe and effective amount of a polynucleotide comprising a
nucleic acid sequence encoding a polypeptide comprising:
35 (i) an Rv2386c protein sequence;
(ii) a variant of an Rv2386c protein sequence; or
(iii) an immunogenic fragment of an Rv2386c protein sequence;

to a subject in need thereof, wherein said polynucleotide induces an immune response, in particular an immune response against *Mycobacterium tuberculosis*.

Use of a polynucleotide comprising a nucleic acid sequence encoding a polypeptide
5 comprising:

- (i) an Rv2386c protein sequence;
- (ii) a variant of an Rv2386c protein sequence; or
- (iii) an immunogenic fragment of an Rv2386c protein sequence;

10 in the manufacture of a medicament for the treatment or prevention of TB, represents another aspect of the invention.

Additionally, there is provided a pharmaceutical composition comprising:

(a) a polypeptide which comprises:

- (i) an Rv2386c protein sequence;
- 15 (ii) a variant of an Rv2386c protein sequence; or
- (iii) an immunogenic fragment of an Rv2386c protein sequence; or
- (b) a polynucleotide comprising a nucleic acid sequence encoding the polypeptide of (a);
- and
- 20 (c) a pharmaceutically acceptable carrier or excipient.

Further, there is provided an immunogenic composition comprising:

(a) a polypeptide which comprises:

- (i) an Rv2386c protein sequence;
- 25 (ii) a variant of an Rv2386c protein sequence; or
- (iii) an immunogenic fragment of an Rv2386c protein sequence; or
- (b) a polynucleotide comprising a nucleic acid sequence encoding the polypeptide of (a);
- and
- 30 (c) a non-specific immune response enhancer.

Suitably the pharmaceutical and immunogenic composition comprise a combination of an Rv2386c and a second heterologous tuberculosis antigen component.

35 Also provided is an expression vector comprising a nucleic acid sequence encoding a polypeptide which comprises:

- (i) an Rv2386c protein sequence;

- (ii) a variant of an Rv2386c protein sequence; or
- (iii) an immunogenic fragment of an Rv2386c protein sequence.

Host cells, transformed with said expression vector, form a further aspect of the invention.

- 5 Additionally provided is a host cell which recombinantly expresses a polypeptide which comprises:

- (i) an Rv2386c protein sequence;
- (ii) a variant of an Rv2386c protein sequence; or
- (iii) an immunogenic fragment of an Rv2386c protein sequence.

10

Further, there is provided a method for the production of a polypeptide comprising:

- (i) an Rv2386c protein sequence;
- (ii) a variant of an Rv2386c protein sequence; or
- (iii) an immunogenic fragment of an Rv2386c protein sequence;

- 15 said method comprising the step of recombinantly expressing said polypeptide within a host cell.

Additionally provided is an antibody or fragment thereof which specifically binds to a polypeptide comprising:

- 20
- (i) an Rv2386c protein sequence;
 - (ii) a variant of an Rv2386c protein sequence; or
 - (iii) an immunogenic fragment of an Rv2386c protein sequence.

- The use of said antibodies in diagnosis is also provided (such as methods for the diagnosis of tuberculosis comprising determining the presence of an antibody or fragment thereof which
- 25 specifically binds to the polypeptides of the invention in a biological sample from a test subject).

Also provided are diagnostic kits comprising:

- 30
- (a) a polypeptide of the present invention;
 - (b) apparatus sufficient to contact said polypeptide with a sample (e.g. whole blood or more suitably PBMC) from an individual; and
 - (c) means to quantify the T cell response of the sample.

Another aspect of the invention relates to a diagnostic kit comprising:

- 35
- (a) a polypeptide of the present invention; and
 - (b) apparatus sufficient to contact said polypeptide with the dermal cells of a patient.

In one embodiment the subject receiving a polypeptide, polynucleotide or composition of the invention may have active tuberculosis (e.g. active infection by *M. tuberculosis*). In a second embodiment the subject may have latent tuberculosis (e.g. dormant infection by *M.*

5 *tuberculosis*). In a third embodiment the subject may be free from tuberculosis (e.g. free from infection by *M. tuberculosis*).

A subject receiving a polypeptide, polynucleotide or composition of the invention may have previously been vaccinated for tuberculosis (e.g. vaccinated against infection by *M.*

10 *tuberculosis*), such as having been vaccinated with *Bacillus Calmette-Guerin* (BCG). Alternatively, a subject receiving a polypeptide, polynucleotide or composition of the invention may not have previously been vaccinated for tuberculosis (e.g. not vaccinated against infection by *M. tuberculosis*), such as not having been vaccinated with *Bacillus Calmette-Guerin* (BCG).

15 DESCRIPTION OF THE FIGURES

- Figure 1: Percentage of CD4 and CD8 cells from immunised CB6F1 mice expressing IFN-gamma and/or IL-2 and/or TNF-alpha cytokines at day 21 (i.e. 7 days post second immunisation).
- 20 Figure 2: Cytokine profile at day 21 (i.e. 7 days post second immunisation) of the antigen specific CD4 response in immunised CB6F1 mice.
- Figure 3: Cytokine profile at day 21 (i.e. 7 days post second immunisation) of the antigen specific CD8 response in immunised CB6F1 mice.
- Figure 4: Percentage of CD4 and CD8 cells from immunised CB6F1 mice expressing IFN-gamma and/or IL-2 and/or TNF-alpha cytokines at day
- 25 35 (i.e. 7 days post third immunisation).
- Figure 5: Cytokine profile at day 35 (i.e. 7 days post third immunisation) of the antigen specific CD4 response in immunised CB6F1 mice.
- Figure 6: Cytokine profile at day 35 (i.e. 7 days post third immunisation) of the
- 30 antigen specific CD8 response in immunised CB6F1 mice.
- Figure 7: Percentage of CD4 and CD8 cells from immunised C57BL/6 mice expressing IFN-gamma and/or IL-2 and/or TNF-alpha cytokines at day 21 (i.e. 7 days post second immunisation).
- Figure 8: Cytokine profile at day 21 (i.e. 7 days post second immunisation) of the
- 35 antigen specific CD4 response in immunised C57BL/6 mice.
- Figure 9: Cytokine profile at day 21 (i.e. 7 days post second immunisation) of the antigen specific CD8 response in immunised C57BL/6 mice.

DESCRIPTION OF THE LISTED SEQUENCES

- | | | |
|----|----------------|---|
| | SEQ ID No: 1: | polypeptide sequence of Rv2386c from <i>M. tuberculosis</i> H37Rv strain. |
| 5 | SEQ ID No: 2: | polynucleotide sequence of Rv2386c from <i>M. tuberculosis</i> H37Rv strain. |
| | SEQ ID No: 3: | polypeptide sequence of Rv2386c from <i>M. tuberculosis</i> CDC1551 strain. |
| | SEQ ID No: 4: | polypeptide sequence of Rv2386c from <i>M. tuberculosis</i> F11 strain. |
| | SEQ ID No: 5: | polypeptide sequence of Rv2386c from <i>M. tuberculosis</i> Haarlem A strain. |
| 10 | SEQ ID No: 6: | polypeptide sequence of Rv2386c from <i>M. tuberculosis</i> C strain. |
| | SEQ ID No: 7: | polypeptide sequence of Rv2386c from <i>BCG</i> . |
| | SEQ ID No: 8: | polypeptide sequence of Mtb8.4. |
| | SEQ ID No: 9: | polypeptide sequence of Mtb9.8. |
| | SEQ ID No: 10: | polypeptide sequence of Mtb9.9. |
| 15 | SEQ ID No: 11: | polypeptide sequence of Ra12. |
| | SEQ ID No: 12: | polypeptide sequence of Ra35. |
| | SEQ ID No: 13: | polypeptide sequence of TbH9. |
| | SEQ ID No: 14: | polypeptide sequence of Mtb40. |
| | SEQ ID No: 15: | polypeptide sequence of Mtb41. |
| 20 | SEQ ID No: 16: | polypeptide sequence of ESAT-6. |
| | SEQ ID No: 17: | polypeptide sequence of Ag85A. |
| | SEQ ID No: 18: | polypeptide sequence of Ag85B. |
| | SEQ ID No: 19: | polypeptide sequence of alpha-crystallin. |
| | SEQ ID No: 20: | polypeptide sequence of MPT64. |
| 25 | SEQ ID No: 21: | polypeptide sequence of Mtb32A. |
| | SEQ ID No: 22: | polypeptide sequence of Ser/Ala mutated mature Mtb32A. |
| | SEQ ID No: 23: | polypeptide sequence of TB10.4. |
| | SEQ ID No: 24: | polypeptide sequence of Mtb72f. |
| | SEQ ID No: 25: | polypeptide sequence of M72. |
| 30 | SEQ ID No: 26: | polypeptide sequence of Mtb71f. |
| | SEQ ID No: 27: | polypeptide sequence of M92 fusion. |
| | SEQ ID No: 28: | polypeptide sequence of M103 fusion. |
| | SEQ ID No: 29: | polypeptide sequence of M114 fusion. |
| | SEQ ID No: 30: | putative human CD4 cell epitope 1. |
| 35 | SEQ ID No: 31: | putative human CD4 cell epitope 2. |
| | SEQ ID No: 32: | putative human CD4 cell epitope 3. |
| | SEQ ID No: 33: | putative human CD4 cell epitope 4. |

	SEQ ID No: 34:	putative human CD4 cell epitope 5.
	SEQ ID No: 35:	putative human CD4 cell epitope 6.
	SEQ ID No: 36:	putative human CD4 cell epitope 7.
	SEQ ID No: 37:	putative human CD4 cell epitope 8.
5	SEQ ID No: 38:	putative human CD4 cell epitope 9.
	SEQ ID No: 39:	putative human CD4 cell epitope 10.
	SEQ ID No: 40:	putative human CD4 cell epitope 11.
	SEQ ID No: 41:	putative human CD4 cell epitope 12.
	SEQ ID No: 42:	putative human CD4 cell epitope 13.
10	SEQ ID No: 43:	putative human CD4 cell epitope 14.
	SEQ ID No: 44:	putative human CD4 cell epitope 15.
	SEQ ID No: 45:	putative human CD4 cell epitope 16.
	SEQ ID No: 46:	putative human CD4 cell epitope 17.
	SEQ ID No: 47:	putative human CD4 cell epitope 18.
15	SEQ ID No: 48:	putative human CD4 cell epitope 19.
	SEQ ID No: 49:	putative human CD4 cell epitope 20.
	SEQ ID No: 50:	putative human CD4 cell epitope 21.
	SEQ ID No: 51:	putative human CD4 cell epitope 22.
	SEQ ID No: 52:	putative human CD4 cell epitope 23.
20	SEQ ID No: 53:	putative human CD8 cell epitope 1.
	SEQ ID No: 54:	putative human CD8 cell epitope 2.
	SEQ ID No: 55:	putative human CD8 cell epitope 3.
	SEQ ID No: 56:	putative human CD8 cell epitope 4.
	SEQ ID No: 57:	putative human CD8 cell epitope 5.
25	SEQ ID No: 58:	putative human CD8 cell epitope 6.
	SEQ ID No: 59:	putative human CD8 cell epitope 7.
	SEQ ID No: 60:	putative human CD8 cell epitope 8.
	SEQ ID No: 61:	putative human CD8 cell epitope 9.
	SEQ ID No: 62:	putative human CD8 cell epitope 10.
30	SEQ ID No: 63:	putative human CD8 cell epitope 11.
	SEQ ID No: 64:	putative human CD8 cell epitope 12.
	SEQ ID No: 65:	putative human CD8 cell epitope 13.
	SEQ ID No: 66:	putative human CD8 cell epitope 14.
	SEQ ID No: 67:	putative human CD8 cell epitope 15.
35	SEQ ID No: 68:	putative human CD8 cell epitope 16.
	SEQ ID No: 69:	putative human CD8 cell epitope 17.
	SEQ ID No: 70:	putative human CD8 cell epitope 18.

	SEQ ID No: 71:	putative human CD8 cell epitope 19.
	SEQ ID No: 72:	putative human CD8 cell epitope 20.
	SEQ ID No: 73:	putative human CD8 cell epitope 21.
	SEQ ID No: 74:	putative human CD8 cell epitope 22.
5	SEQ ID No: 75:	putative human CD8 cell epitope 23.
	SEQ ID No: 76:	putative human CD8 cell epitope 24.
	SEQ ID No: 77:	putative human CD8 cell epitope 25.
	SEQ ID No: 78:	putative human CD8 cell epitope 26.
	SEQ ID No: 79:	putative human CD8 cell epitope 27.
10	SEQ ID No: 80:	putative human CD8 cell epitope 28.
	SEQ ID No: 81:	putative human CD8 cell epitope 29.
	SEQ ID No: 82:	putative human CD8 cell epitope 30.
	SEQ ID No: 83:	putative human CD8 cell epitope 31.
	SEQ ID No: 84:	putative human CD8 cell epitope 32.
15	SEQ ID No: 85:	putative human CD8 cell epitope 33.
	SEQ ID No: 86:	putative human CD8 cell epitope 34.
	SEQ ID No: 87:	putative human CD8 cell epitope 35.
	SEQ ID No: 88:	putative human CD8 cell epitope 36.
	SEQ ID No: 89:	putative human CD8 cell epitope 37.
20	SEQ ID No: 90:	putative human CD8 cell epitope 38.
	SEQ ID No: 91:	putative human CD8 cell epitope 39.
	SEQ ID No: 92:	putative human CD8 cell epitope 40.
	SEQ ID No: 93:	putative human CD8 cell epitope 41.
	SEQ ID No: 94:	putative human CD8 cell epitope 42.
25	SEQ ID No: 95:	putative human CD8 cell epitope 43.
	SEQ ID No: 96:	putative human CD8 cell epitope 44.
	SEQ ID No: 97:	putative human CD8 cell epitope 45.
	SEQ ID No: 98:	putative human CD8 cell epitope 46.
	SEQ ID No: 99:	putative human CD8 cell epitope 47.
30	SEQ ID No: 100:	putative human CD8 cell epitope 48.
	SEQ ID No: 101:	putative human CD8 cell epitope 49.
	SEQ ID No: 102:	putative human CD8 cell epitope 50.
	SEQ ID No: 103:	putative human CD8 cell epitope 51.
	SEQ ID No: 104:	putative human CD8 cell epitope 52.
35	SEQ ID No: 105:	putative human CD8 cell epitope 53.
	SEQ ID No: 106:	putative human CD8 cell epitope 54.
	SEQ ID No: 107:	putative human CD8 cell epitope 55.

	SEQ ID No: 108:	putative human CD8 cell epitope 56.
	SEQ ID No: 109:	putative human CD8 cell epitope 57.
	SEQ ID No: 110:	putative human CD8 cell epitope 58.
	SEQ ID No: 111:	putative human CD8 cell epitope 59.
5	SEQ ID No: 112:	putative human CD8 cell epitope 60.
	SEQ ID No: 113:	putative human CD8 cell epitope 61.
	SEQ ID No: 114:	putative human CD8 cell epitope 62.
	SEQ ID No: 115:	putative human CD8 cell epitope 63.
	SEQ ID No: 116:	putative human CD8 cell epitope 64.
10	SEQ ID No: 117:	putative human CD8 cell epitope 65.
	SEQ ID No: 118:	putative human CD8 cell epitope 66.
	SEQ ID No: 119:	putative human CD8 cell epitope 67.
	SEQ ID No: 120:	putative human CD8 cell epitope 68.
	SEQ ID No: 121:	putative human CD8 cell epitope 69.
15	SEQ ID No: 122:	putative human CD8 cell epitope 70.
	SEQ ID No: 123:	putative human CD8 cell epitope 71.
	SEQ ID No: 124:	putative human CD8 cell epitope 72.
	SEQ ID No: 125:	putative human CD8 cell epitope 73.
	SEQ ID No: 126:	putative human CD8 cell epitope 74.
20	SEQ ID No: 127:	putative human CD8 cell epitope 75.
	SEQ ID No: 128:	putative human CD8 cell epitope 76.
	SEQ ID No: 129:	putative human CD8 cell epitope 77.
	SEQ ID No: 130:	putative human CD8 cell epitope 78.
	SEQ ID No: 131:	putative human CD8 cell epitope 79.
25	SEQ ID No: 132:	putative human CD8 cell epitope 80.
	SEQ ID No: 133:	putative human CD8 cell epitope 81.
	SEQ ID No: 134:	putative human CD8 cell epitope 82.
	SEQ ID No: 135:	putative human CD8 cell epitope 83.
	SEQ ID No: 136:	putative human CD8 cell epitope 84.
30	SEQ ID No: 137:	putative human CD8 cell epitope 85.
	SEQ ID No: 138:	putative human CD8 cell epitope 86.
	SEQ ID No: 139:	putative human CD8 cell epitope 87.
	SEQ ID No: 140:	putative human CD8 cell epitope 88.
	SEQ ID No: 141:	putative human CD8 cell epitope 89.
35	SEQ ID No: 142:	putative human CD8 cell epitope 90.
	SEQ ID No: 143:	putative human CD8 cell epitope 91.
	SEQ ID No: 144:	putative human CD8 cell epitope 92.

- SEQ ID No: 145: putative human CD8 cell epitope 93.
 SEQ ID No: 146: putative human CD8 cell epitope 94.
 SEQ ID No: 147: putative human CD8 cell epitope 95.
 SEQ ID No: 148: putative human CD8 cell epitope 96.
 5 SEQ ID No: 149: putative human CD8 cell epitope 97.
 SEQ ID No: 150: putative human CD8 cell epitope 98.
 SEQ ID No: 151: putative human CD8 cell epitope 99.
 SEQ ID No: 152: putative human CD8 cell epitope 100.
 SEQ ID No: 153: putative human CD8 cell epitope 101.
 10 SEQ ID No: 154: putative human CD8 cell epitope 102.
 SEQ ID No: 155: polypeptide sequence of Rv1753c from *M. tuberculosis* H37Rv strain.
 SEQ ID No: 156: polypeptide sequence of Rv2707c from *M. tuberculosis* H37Rv strain.

DETAILED DESCRIPTION

15

Currently, vaccination with live bacteria is the most efficient method for inducing protective immunity. The most common *Mycobacterium* employed for this purpose is *Bacillus Calmette-Guerin* (BCG), an avirulent strain of *M. bovis* which was developed over 60 years ago.

However, the safety and efficacy of BCG is a source of controversy - while protecting against
 20 severe disease manifestation in children, BCG does not prevent the establishment of latent TB or reactivation of pulmonary disease in adult life. Additionally, some countries, such as the United States, do not vaccinate the general public with this agent.

Almost all new generation TB vaccines which are currently in clinical development have been
 25 designed as pre-exposure vaccines. These include subunit vaccines, which have been particularly effective in boosting immunity induced by prior BCG vaccination, and advanced live mycobacterial vaccines which aim to replace BCG with more efficient and/or safer strains. Although these vaccines aim to improve resistance to infection, they are likely to be less effective as post-exposure or therapeutic vaccines in latent TB cases (Lin MY et al *Endocrine*,
 30 *Metabolic & Immune Disorders – Drug Targets* 2008 8:15-29).

Several of the proteins which are strongly expressed during the early stages of *Mycobacterium* infection have been shown to provide strong protective efficacy in animal vaccination models. However, vaccination with antigens which are highly expressed during the early stages of
 35 infection may not provide an optimal immune response for dealing with later stages of infection. Adequate control during the later stages of infection may require T cells which are specific for the particular antigens which are expressed at that time.

Post-exposure vaccines which directly target the dormant persistent bacteria may aid in protecting against TB reactivation, thereby enhancing TB control, or even enabling clearance of the infection. A vaccine targeting latent TB could therefore significantly and economically
5 reduce global TB infection rates.

Subunit vaccines based on late stage antigens could also be utilised in combination with early stage antigens to provide a multiphase vaccine. Alternatively, late stage antigens could be used to complement and improve BCG vaccination (either by boosting the BCG response or
10 through the development of advanced recombinant BCG strains).

Rv2386c, also known as MbtI, catalyses the initial transformation in mycobactin biosynthesis by converting chorismate to salicylate (Zwahlen et al. *Biochemistry* 2007 46:954-964). Rv2386 has previously been identified as being associated with expression under stress conditions
15 although did not result in protection when administered as a DNA vaccine in a guinea pig model (Vipond et al. *Vaccine* 2006 24:6340-6350).

Recently, a range of *M. tuberculosis* vaccine candidates have been proposed based on a bioinformatics analysis of the whole genome *M. tuberculosis* genome (Zvi et al. *BMC Medical*
20 *Genetics* 2008 1:18) and on the testing of differentially expressed proteins in actively and latently infected individuals (Schuck SD et al. *PLoS ONE* 2009 4(5):e5590).

While macrophages have been shown to act as the principal effectors of *Mycobacterium* immunity, T cells are the predominant inducers of such immunity. The essential role of T cells
25 in protection against tuberculosis is illustrated by the increased rates of TB reactivation in human immunodeficiency virus infected individuals, due to the associated depletion of CD4+ T cells. Furthermore, adoptive transfer of CD4+ T cells taken at the height of the primary immune response to *M. tuberculosis* has been shown to confer protection against *M. tuberculosis* in T cell deficient mice (Orme et al *J. Exp. Med.* 1983 158:74-83).

30 *Mycobacterium*-reactive CD4+ T cells have been shown to be potent producers of γ -interferon (IFN- γ), which, in turn, has been shown to trigger the anti-mycobacterial effects of macrophages in mice (Flynn et al. *J. Exp. Med.* 1993 178:2249-2254). While the role of IFN- γ in humans is less clear, studies have shown that 1,25-dihydroxy-vitamin D3, either alone or in
35 combination with IFN- γ or tumor necrosis factor-alpha, activates human macrophages to inhibit *M. tuberculosis* infection. Furthermore, it is known that IFN- γ stimulates human macrophages to make 1,25-dihydroxy-vitamin D3. Similarly, interleukin-12 (IL-12) has been shown to play a

role in stimulating resistance to *M. tuberculosis* infection. For a review of the immunology of *M. tuberculosis* infection, see Chan & Kaufmann, *Tuberculosis: Pathogenesis, Protection and Control* (Bloom ed., 1994), *Tuberculosis* (2nd ed., Rom and Garay, eds., 2003), and *Harrison's Principles of Internal Medicine*, Chapter 150, pp. 953-966 (16th ed., Braunwald, et al., eds.,
5 2005).

The present invention relates generally to the identification of Rv2386c as a TB antigen (in particular an antigen associated with latent TB) and to related methods and uses in the prevention and treatment of TB, especially the prevention and treatment of latent TB and the
10 prevention or delay of TB reactivation.

The invention therefore provides an Rv2386c protein, variant thereof or immunogenic fragment thereof, or a polynucleotide encoding said protein, variant or fragment, for use in the treatment or prevention of TB. Suitably, the use may be specifically in the prevention and treatment of
15 latent TB (especially the treatment of latent TB). Alternatively, the use may be in the prevention or delay of TB reactivation (especially the delay of TB reactivation, for example by a period of months, years or even indefinitely).

The term "*Mycobacterium* species of the tuberculosis complex" includes those species
20 traditionally considered as causing the disease tuberculosis, as well as *Mycobacterium* environmental and opportunistic species that cause tuberculosis and lung disease in immune compromised patients, such as patients with AIDS, e.g., *M. tuberculosis*, *M. bovis*, or *M. africanum*, BCG, *M. avium*, *M. intracellulare*, *M. celatum*, *M. genavense*, *M. haemophilum*, *M. kansasii*, *M. simiae*, *M. vaccae*, *M. fortuitum*, and *M. scrofulaceum* (see, e.g., *Harrison's*
25 *Principles of Internal Medicine*, Chapter 150, pp. 953-966 (16th ed., Braunwald, et al., eds., 2005)). The present invention is particularly directed to infection with *M. tuberculosis*.

The term "active infection" refers to an infection (e.g. infection by *M. tuberculosis*) with manifested disease symptoms and/or lesions (suitably with manifested disease symptoms).
30

The terms "inactive infection", "dormant infection" or "latent infection" refer to an infection (e.g. infection by *M. tuberculosis*) without manifested disease symptoms and/or lesions (suitably without manifested disease symptoms).

35 The term "primary tuberculosis" refers to clinical illness (manifestation of disease symptoms) directly following infection (e.g. infection by *M. tuberculosis*). See, *Harrison's Principles of Internal Medicine*, Chapter 150, pp. 953-966 (16th ed., Braunwald, et al., eds., 2005).

The terms “secondary tuberculosis” or “postprimary tuberculosis” refer to the reactivation of a dormant, inactive or latent infection (e.g. infection by *M. tuberculosis*). See, *Harrison’s Principles of Internal Medicine*, Chapter 150, pp. 953-966 (16th ed., Braunwald, et al., eds., 2005).

The term “tuberculosis reactivation” refers to the later manifestation of disease symptoms in an individual that tests positive for infection (e.g. in a tuberculin skin test, suitably in an *in vitro* T cell based assay) test but does not have apparent disease symptoms. The positive diagnostic test indicates that the individual is infected, however, the individual may or may not have previously manifested active disease symptoms that had been treated sufficiently to bring the tuberculosis into an inactive or latent state. It will be recognised that methods for the prevention, delay or treatment of tuberculosis reactivation can be initiated in an individual manifesting active symptoms of disease.

The term “drug resistant” tuberculosis refers to an infection (e.g. infection by *M. tuberculosis*) wherein the infecting strain is not held static or killed (i.e. is resistant to) one or more of so-called “front-line” chemotherapeutic agents effective in treating tuberculosis (e.g., isoniazid, rifampin, ethambutol, streptomycin and pyrazinamide).

The term “multi-drug resistant” tuberculosis refers to an infection (e.g. infection by *M. tuberculosis*) wherein the infecting strain is resistant to two or more of “front-line” chemotherapeutic agents effective in treating tuberculosis.

A “chemotherapeutic agent” refers to a pharmacological agent known and used in the art to treat tuberculosis (e.g. infection by *M. tuberculosis*). Exemplified pharmacological agents used to treat tuberculosis include, but are not limited to amikacin, aminosalicylic acid, capreomycin, cycloserine, ethambutol, ethionamide, isoniazid, kanamycin, pyrazinamide, rifamycins (i.e., rifampin, rifapentine and rifabutin), streptomycin, ofloxacin, ciprofloxacin, clarithromycin, azithromycin and fluoroquinolones. “First-line” or “Front-line” chemotherapeutic agents used to treat tuberculosis that is not drug resistant include isoniazid, rifampin, ethambutol, streptomycin and pyrazinamide. “Second-line” chemotherapeutic agents used to treat tuberculosis that has demonstrated drug resistance to one or more “first-line” drugs include ofloxacin, ciprofloxacin, ethionamide, aminosalicylic acid, cycloserine, amikacin, kanamycin and capreomycin. Such pharmacological agents are reviewed in Chapter 48 of *Goodman and Gilman’s The Pharmacological Basis of Therapeutics*, Hardman and Limbird eds., 2001.

The terms "polypeptide", "peptide" and "protein" are used interchangeably herein to refer to a polymer of amino acid residues. The terms also apply to amino acid polymers in which one or more amino acid residue is an artificial chemical mimetic of a corresponding naturally occurring amino acid, as well as to naturally occurring amino acid polymers and non-naturally occurring amino acid polymer. Suitably a polypeptide according to the present invention will consist only of naturally occurring amino acid residues, especially those amino acids encoded by the genetic code.

The term "amino acid" refers to naturally occurring and synthetic amino acids, as well as amino acid analogs and amino acid mimetics that function in a manner similar to the naturally occurring amino acids. Naturally occurring amino acids are those encoded by the genetic code, as well as those amino acids that are later modified, e.g., hydroxyproline, γ -carboxyglutamate, and O-phosphoserine. Amino acid analogues refers to compounds that have the same basic chemical structure as a naturally occurring amino acid, i.e., an α carbon that is bound to a hydrogen, a carboxyl group, an amino group, and an R group, e.g., homoserine, norleucine, methionine sulfoxide, methionine methyl sulfonium. Such analogs have modified R groups (e.g., norleucine) or modified peptide backbones, but retain the same basic chemical structure as a naturally occurring amino acid. Amino acid mimetics refers to chemical compounds that have a structure that is different from the general chemical structure of an amino acid, but that functions in a manner similar to a naturally occurring amino acid. Suitably an amino acid is a naturally occurring amino acid or an amino acid analogue, especially a naturally occurring amino acid and in particular those amino acids encoded by the genetic code.

"Nucleic acid" refers to deoxyribonucleotides or ribonucleotides and polymers thereof in either single- or double-stranded form. The term encompasses nucleic acids containing known nucleotide analogs or modified backbone residues or linkages, which are synthetic, naturally occurring, and non-naturally occurring, which have similar binding properties as the reference nucleic acid, and which are metabolized in a manner similar to the reference nucleotides. Examples of such analogs include, without limitation, phosphorothioates, phosphoramidates, methyl phosphonates, chiral-methyl phosphonates, 2-O-methyl ribonucleotides, peptide-nucleic acids (PNAs). Suitably the term "nucleic acid" refers to naturally occurring deoxyribonucleotides or ribonucleotides and polymers thereof.

Unless otherwise indicated, a particular nucleic acid sequence also implicitly encompasses conservatively modified variants thereof (e.g., degenerate codon substitutions) and complementary sequences, as well as the sequence explicitly indicated (suitably it refers to the

sequence explicitly indicated). Specifically, degenerate codon substitutions may be achieved by generating sequences in which the third position of one or more selected (or all) codons is substituted with mixed-base and/or deoxyinosine residues (Batzner *et al.*, *Nucleic Acid Res.* 19:5081 (1991); Ohtsuka *et al.*, *J. Biol. Chem.* 260:2605-2608 (1985); Rossolini *et al.*, *Mol. Cell. Probes* 8:91-98 (1994)). The term nucleic acid is used interchangeably with gene, cDNA, mRNA, oligonucleotide, and polynucleotide.

Amino acids may be referred to herein by either their commonly known three letter symbols or by the one-letter symbols recommended by the IUPAC-IUB Biochemical Nomenclature Commission. Nucleotides, likewise, may be referred to by their commonly accepted single-letter codes.

By the term 'Rv2386c protein sequence' as used herein is meant the polypeptide sequence provided in SEQ ID No: 1 or a homologue thereof from a *Mycobacterium* species of the tuberculosis complex, e.g., a species such as *M. tuberculosis*, *M. bovis*, or *M. africanum*, or a *Mycobacterium* species that is environmental or opportunistic and that causes opportunistic infections such as lung infections in immune compromised hosts (e.g., patients with AIDS), e.g., *BCG*, *M. avium*, *M. intracellulare*, *M. celatum*, *M. genavense*, *M. haemophilum*, *M. kansasii*, *M. simiae*, *M. vaccae*, *M. fortuitum*, and *M. scrofulaceum* (see, e.g., *Harrison's Principles of Internal Medicine*, Chapter 150, pp. 953-966, 16th ed., Braunwald, *et al.*, eds., 2005).

To ensure a high efficacy rate among vaccinated hosts, the components of a vaccine should be well conserved among the strains of clinical significance. Suitably, the Rv2386c protein is derived from *M. tuberculosis* H37Rv (i.e. the polypeptide sequence provided in SEQ ID No: 1) or a homologue thereof from another *M. tuberculosis* strain (such as CDC1551, F11, Haarlem A and C strains). Strains of *M. tuberculosis* which are associated with drug resistance (e.g. MDR or especially XDR) are a particularly valuable basis for the Rv2386c protein sequence. Strains of interest include:

CDC1551 - transmissible and virulent strain

Haarlem family (such as Haarlem A) - Drug resistant strains found in crowded human populations. Members of the Haarlem family of *M. tuberculosis* strains have been found in many parts of the world. The first representative of the family was discovered in Haarlem, The Netherlands.

KZN4207 - Drug sensitive isolate from patients in KwaZulu-Natal, South Africa

KZN1435 - Multiple drug resistant (MDR) isolate from patients in KwaZulu-Natal, South Africa

5

KZN605 - Extensively drug resistant (XDR) isolate from patients in KwaZulu-Natal, South Africa

10 C - Highly transmitted in New York City. In one study this strain was found to be more common among injection drug users and resistant to reactive nitrogen intermediates (Friedman et al. *J. Infect. Dis.* 1997 176(2):478-84)

15 94_M4241A - Isolated in San Francisco in 1994 from a patient born in China. This strain was previously analysed by genomic deletion analysis (Gagneux et al., *PNAS* 2006 103(8):2869-2873).

20 02_1987 - Isolated in San Francisco in 2002 from a patient born in South Korea. This strain was previously analyzed by genomic deletion analysis (Gagneux et al., *PNAS* 2006 103(8):2869-2873).

T92 - Isolated in San Francisco in 1999 from a patient born in The Philippines. This strain was published in Hirsh et al. *PNAS* 2004 101:4871-4876).

25 T85 - Isolated in San Francisco in 1998 from a patient born in China. This strain was published in Hirsh et al. *PNAS* 2004 101:4871-4876).

30 EAS054 - Isolated in San Francisco in 1993 from a patient born in India. This strain was previously analyzed by genomic deletion analysis (Gagneux et al., *PNAS* 2006 103(8):2869-2873).

Gagneux et al., *PNAS* 2006 103(8):2869-2873 and Herbert et al. *Infect. Immun.* 2007 75(12):5798-5805 provide valuable background on the range of *M. tuberculosis* strains which are known to exist.

35 Most suitably, the Rv2386c protein is selected from the polypeptide sequences provided in SEQ ID Nos: 1 and 3-7, in particular SEQ ID Nos: 1 and 3-6, such as SEQ ID No: 1.

Polynucleotides of particular interest are those comprising (such as consisting of) a sequence encoding:

- (i) an Rv2386c protein sequence;
- (ii) a variant of an Rv2386c protein sequence; or
- 5 (iii) an immunogenic fragment of an Rv2386c protein sequence.

Polynucleotides will suitably comprise (such as consisting of) a variant of SEQ ID NO: 2 or fragment of SEQ ID NO: 2 which encodes an immunogenic fragment of an Rv2386c protein.

COMBINATIONS

10

The Rv2386c related polypeptides of the present invention can further comprise other components designed to enhance their immunogenicity or to improve these antigens in other respects. For example, improved isolation of the polypeptide antigens may be facilitated through the addition of a stretch of histidine residues (commonly known as a his-tag) towards

15 one end of the antigen.

20

The term "his-tag" refers to a string of histidine residues, typically six residues, that are inserted within the reference sequence. To minimise disruption of the activity associated with the reference sequence, a his-tag is typically inserted at the N-terminus, usually immediately after the initiating methionine residue, or else at the C-terminus. They are usually heterologous to the native sequence but are incorporated since they facilitate isolation by improving the protein binding to immobilised metal affinity chromatography resins (IMAC). Generally speaking the presence or absence of a his-tag is not of significance from the point of view of eliciting a desirable immune response against the reference protein. However, to avoid the

25 risk of an adverse reaction against the his-tag itself, it is considered best to minimise the length of the his-tag e.g. to four or fewer residues, in particular two residues (or to exclude the use of a his-tag entirely).

30

To improve the magnitude and/or breadth of the elicited immune response the compositions, polypeptides and nucleic acids of the invention can comprise multiple copies of the inventive antigen and/or additional heterologous polypeptides (or polynucleotides encoding them) from *Mycobacterium* species (in particular *M. tuberculosis*).

35

One skilled in the art will recognise that when a number of components are utilised in combination, the precise presentation can be varied. For example, a Rv2386c component and an additional copy of the inventive antigen or an additional heterologous antigen component could be presented:

- (1) as two individual polypeptide components;
- (2) as a fusion protein comprising both polypeptide components;
- (3) as one polypeptide and one polynucleotide component;
- 5 (4) as two individual polynucleotide components;
- (5) as a single polynucleotide encoding two individual polypeptide components; or
- (6) as a single polynucleotide encoding a fusion protein comprising both polypeptide components.

10 This flexibility applies equally to situations where three or more components are used in combination. However, for convenience, it is often desirable that when a number of components are present they are contained within a single fusion protein or a polynucleotide encoding a single fusion protein. In one embodiment of the invention all antigen components are provided as polypeptides (e.g. within a single fusion protein). In an alternative
15 embodiment of the invention all antigen components are provided as polynucleotides (e.g. a single polynucleotide, such as one encoding a single fusion protein).

The term "heterologous" when used with reference to portions of a nucleic acid indicates that the nucleic acid comprises two or more subsequences that are not found in the same
20 relationship to each other in nature. For instance, the nucleic acid is typically recombinantly produced, having two or more sequences from unrelated genes arranged to make a new functional nucleic acid, e.g., a promoter from one source and a coding region from another source. Similarly, a heterologous protein indicates that the protein comprises two or more subsequences that are not found in the same relationship to each other in nature (e.g., a
25 fusion protein).

"Fusion polypeptide" or "fusion protein" refers to a protein having at least two heterologous polypeptides (e.g. at least two *Mycobacterium* sp. polypeptides) covalently linked, either directly or via an amino acid linker. The polypeptides forming the fusion protein are typically
30 linked C-terminus to N-terminus, although they can also be linked C-terminus to C-terminus, N-terminus to N-terminus, or N-terminus to C-terminus. The polypeptides of the fusion protein can be in any order. This term also refers to conservatively modified variants, polymorphic variants, alleles, mutants, immunogenic fragments, and interspecies homologs of the antigens that make up the fusion protein. *Mycobacterium tuberculosis* antigens are described in Cole *et al.*, *Nature* 393:537 (1998), which discloses the entire *Mycobacterium tuberculosis* genome.
35 Antigens from other *Mycobacterium* species that correspond to *M. tuberculosis* antigens can be identified, e.g., using sequence comparison algorithms, as described herein, or other

methods known to those of skill in the art, e.g., hybridisation assays and antibody binding assays.

The term “fused” refers to the covalent linkage between two polypeptides in a fusion protein.

- 5 The polypeptides are typically joined via a peptide bond, either directly to each other or via an amino acid linker. Optionally, the peptides can be joined via non-peptide covalent linkages known to those of skill in the art.

Exemplary *M. tuberculosis* antigens which may be combined with Rv2386c include one or
10 more of (e.g. 1 to 5, such as 1 to 3, in particular 1) the following (such as one or more of (i) to (xii)):

(i) Mtb8.4 (also known as DPV and Rv1174c), the polypeptide sequence of which is described in SEQ ID No: 102 of WO97/09428 (cDNA in SEQ ID No: 101) and in Coler
15 et al *Journal of Immunology* 1998 161:2356-2364. Of particular interest is the mature Mtb8.4 sequence which is absent the leading signal peptide (i.e. amino acid residues 15-96 from SEQ ID No: 102 of WO97/09428). The full-length polypeptide sequence of Mtb8.4 is shown in SEQ ID No: 8;

20 (ii) Mtb9.8 (also known as MSL and Rv0287), the polypeptide sequence of which is described in SEQ ID No: 109 of WO98/53075 (fragments of MSL are disclosed in SEQ ID Nos: 110-124 of WO98/53075, SEQ ID Nos: 119 and 120 being of particular interest) and also in Coler et al *Vaccine* 2009 27:223-233 (in particular the reactive fragments shown in Figure 2 therein). The full-length polypeptide sequence for Mtb9.8
25 is shown in SEQ ID No: 9;

(iii) Mtb9.9 (also known as Mtb9.9A, MTI, MTI-A and Rv1793) the polypeptide sequence of which is described in SEQ ID No: 19 of WO98/53075 and in Alderson et al *Journal of Experimental Medicine* 2000 7:551-559 (fragments of MTI are disclosed in
30 SEQ ID Nos: 17 and 51-66 of WO98/53075, SEQ ID Nos: 17, 51, 52, 53, 56 and 62-65 being of particular interest). A number of polypeptide variants of MTI are described in SEQ ID Nos: 21, 23, 25, 27, 29 and 31 of WO98/53075 and in Alderson et al *Journal of Experimental Medicine* 2000 7:551-559. The full-length polypeptide sequence for Mtb9.9 is shown in SEQ ID No: 10;

35 (iv) Ra12 (also known as Mtb32A C-terminal antigen) the polypeptide sequence of which is described in SEQ ID No: 10 of WO01/98460 and in Skeiky et al *Journal of*

Immunology 2004 172:7618-7682. The full-length polypeptide sequence for Ra12 is shown in SEQ ID No: 11;

5 (v) Ra35 (also known as Mtb32A N-terminal antigen) the polypeptide sequence of which is described in SEQ ID No: 8 of WO01/98460 and in Skeiky et al *Journal of Immunology* 2004 172:7618-7682. The full-length polypeptide sequence for Ra35 is shown in SEQ ID No: 12;

10 (vi) TbH9 (also known as Mtb39, Mtb39A, TbH9FL and Rv1196) the polypeptide sequence of which is described in SEQ ID No: 107 of WO97/09428, and also in Dillon et al *Infection and Immunity* 1999 67(6):2941-2950 and Skeiky et al *Journal of Immunology* 2004 172:7618-7682. The full-length polypeptide sequence for TbH9 is shown in SEQ ID No: 13;

15 (vii) Mtb40 (also known as HTCC1 and Rv3616c) the polypeptide sequence of which is described in SEQ ID No: 138 of WO98/53075 (cDNA in SEQ ID No: 137). The full-length polypeptide sequence for Mtb40 is shown in SEQ ID No: 14;

20 (viii) Mtb41 (also known as MTCC2 and Rv0915c) the polypeptide sequence of which is described in SEQ ID No: 142 of WO98/53075 (cDNA in SEQ ID No: 140) and in Skeiky et al *Journal of Immunology* 2000 165:7140-7149. The full-length polypeptide sequence for Mtb41 is shown in SEQ ID No: 15;

25 (ix) ESAT-6 (also known as esxA and Rv3875) the polypeptide sequence of which is described in SEQ ID No: 103 of WO97/09428 (cDNA in SEQ ID No: 104) and in Sorensen et al *Infection and Immunity* 1995 63(5):1710-1717. The full-length polypeptide sequence for ESAT-6 is shown in SEQ ID No: 16;

30 (x) Ag85 complex antigens (e.g. Ag85A, also known as fbpA and Rv3804c; or Ag85B, also known as fbpB and Rv1886c) which are discussed, for example, in Content et al *Infection and Immunity* 1991 59:3205-3212 and in Huygen et al *Nature Medicine* 1996 2(8):893-898. The full-length polypeptide sequence for Ag85A is shown in SEQ ID No: 17 (the mature protein of residues 43-338, i.e. lacking the signal peptide, being of particular interest). The full-length polypeptide sequence for Ag85B is shown in SEQ ID
35 No: 18 (the mature protein of residues 41-325, i.e. lacking the signal peptide, being of particular interest);

- 5 (xi) Alpha-crystallin (also known as hspX and Rv2031c) which is described in Verbon et al *Journal of Bacteriology* 1992 174:1352-1359 and Friscia et al *Clinical and Experimental Immunology* 1995 102:53-57 (of particular interest are the fragments corresponding to residues 71-91, 21-40, 91-110 and 111-130). The full-length polypeptide sequence for alpha-crystallin is shown in SEQ ID No: 19;
- 10 (xii) Mpt64 (also known as Rv1980c) which is described in Roche et al *Scandinavian Journal of Immunology* 1996 43:662-670. The full-length polypeptide sequence for MPT64 is shown in SEQ ID No: 20 (the mature protein of residues 24-228, i.e. lacking the signal peptide, being of particular interest):
- 15 (xiii) Mtb32A, the polypeptide sequence of which is described in SEQ ID No: 2 (full-length) and residues 8-330 of SEQ ID No: 4 (mature) of WO01/98460, especially variants having at least one of the catalytic triad mutated (e.g. the catalytic serine residue, which may for example be mutated to alanine). The full-length polypeptide sequence for Mtb32A is shown in SEQ ID No: 21. The mature form of Mtb32A having a Ser/Ala mutation is shown in SEQ ID No: 22;
- 20 (xiv) TB10.4, the full-length polypeptide sequence for TB10.4 is shown in SEQ ID No: 23;
- (xv) Rv1753c, the full-length polypeptide sequence for Rv1753c from *Mycobacterium tuberculosis* H37Rv is shown in SEQ ID No: 155; and/or
- 25 (xvi) Rv2707c, the full-length polypeptide sequence for Rv2707c from *Mycobacterium tuberculosis* H37Rv is shown in SEQ ID No: 156.
- or combinations thereof, such as (for example combinations such as (a) to (g)):
- 30 (a) a combination of Ra12, TbH9 and Ra35 components, for example in the form of a fusion protein, such as Mtb72f. The polypeptide sequence of Mtb72f is described in SEQ ID No: 6 of WO2006/117240 (cDNA in SEQ ID No: 5) and in Skeiky et al *Journal of Immunology* 2004 172:7618-7682 (where it incorporates an optional His-tag to aid purification, when utilised in the present invention suitably Mtb72f is absent the optional
- 35 histidine residues). The polypeptide sequence for Mtb72f is shown in SEQ ID No: 24;

- 5 (b) a combination of Ra12, TbH9 and Ser/Ala mutated Ra35 (i.e. where the catalytic serine residue has been replaced with alanine) components, for example in the form of a fusion protein, such as M72. The polypeptide sequence of M72 is described in SEQ ID No: 4 of WO2006/117240 (cDNA in SEQ ID No: 3) where it incorporates an optional double histidine to aid manufacture, when utilised in the present invention M72 may also incorporate a double histidine though suitably M72 is absent the optional double histidine (i.e. residues 4-725 from SEQ ID No: 4 of WO2006/117240 are of particular interest). The polypeptide sequence for M72 is shown in SEQ ID No: 25;
- 10 (c) a combination of Mtb8.4, Mtb9.8, Mtb9.9 and Mtb41 components, for example in the form of a fusion protein, such as Mtb71f. The polypeptide sequence of Mtb71f is described in SEQ ID No: 16 of WO99/051748 (cDNA in SEQ ID No: 15), where it incorporates an optional His-tag to aid purification, when utilised in the present invention suitably Mtb71f corresponds to amino acid residues 9-710 of SEQ ID No: 16 from WO99/051748. The polypeptide sequence for Mtb71f is shown in SEQ ID No: 26;
- 15 (d) a combination of Mtb72f or M72 (suitably without optional histidine residues to aid expression) with Mtb9.8 and Mtb9.9, for example in a fusion protein. The polypeptide sequence for an M72-Mtb9.9-Mtb9.8 fusion is shown in SEQ ID No: 27 (M92 fusion), when used in the present invention, the M72-Mtb9.9-Mtb9.8 fusion may optionally incorporate a double histidine following the initiating methionine residue to aid manufacture;
- 20 (e) a combination of Mtb72f or M72 (suitably without optional histidine residues to aid expression) with Ag85B, for example in a fusion protein, such Mtb103f. The polypeptide sequence of Mtb103f is described in SEQ ID No: 18 of WO03/070187 (cDNA in SEQ ID No: 10), where it incorporates an optional His-tag to aid purification, when utilised in the present invention suitably Mtb103f corresponds to amino acid residues 8-1016 of SEQ ID No: 18 from WO03/070187. Also of particular interest is
- 25 M103, i.e. Mtb103f incorporating a Ser/Ala mutation in the Ra35 component, when utilised in the present invention suitably M103 corresponds to amino acid residues 8-1016 of SEQ ID No: 18 from WO03/070187 wherein the Ser residue at position 710 has been replaced with Ala. The polypeptide sequence for M103 is shown in SEQ ID No: 28, when used in the present invention, the M72-Mtb9.9-Mtb9.8 fusion may optionally
- 30 incorporate a double histidine following the initiating methionine residue to aid manufacture;
- 35

(f) a combination of Mtb72f or M72 (suitably without optional histidine residues to aid expression) with Mtb41, for example in a fusion protein, such as Mtb114f. The polypeptide sequence of Mtb114f is described in SEQ ID No: 16 of WO03/070187 (cDNA in SEQ ID No: 9), where it incorporates an optional His-tag to aid purification, when utilised in the present invention suitably Mtb114f corresponds to amino acid residues 8-1154 of SEQ ID No: 16 from WO03/070187. Also of particular interest is M114, i.e. Mtb114f incorporating a Ser/Ala mutation in the Ra35 component, when utilised in the present invention suitably M114 corresponds to amino acid residues 8-1154 of SEQ ID No: 16 from WO03/070187 wherein the Ser residue at position 710 has been replaced with Ala. The polypeptide sequence for M114 is shown in SEQ ID No: 29, when used in the present invention, the M72-Mtb9.9-Mtb9.8 fusion may optionally incorporate a double histidine following the initiating methionine residue to aid manufacture;

(g) a combination of Ag85B and ESAT-6 components, such as in a fusion described in Doherty et al *Journal of Infectious Diseases* 2004 190:2146–2153; and/or

(h) a combination of Ag85B and TB10.4 components, such as in a fusion described in Dietrich et al *Journal of Immunology* 2005 174(10):6332-6339 190:2146–2153.

Combinations of an Rv2386c component and an Mtb40 component are of particular interest. Obviously such combinations could optionally contain other additional antigen components (e.g. an M72 component).

Another combination of interest comprises an Rv2386c component and an M72 component.

A further combination of interest comprises an Rv2386c component and an Rv1753c component.

Other combinations of interest include those comprising an Rv2386c component and an Rv2707c component.

An additional combination of interest comprises an Rv2386c component and an alpha-crystallin component.

The skilled person will recognise that combinations need not rely upon the specific sequences described in above in (i)-(xvi) and (a)-(h), and that conservatively modified variants (e.g. having at least 70% identity, such as at least 80% identity, in particular at least 90% identity and

especially at least 95% identity) or immunogenic fragments (e.g. at least 20% of the full length antigen, such as at least 50% of the antigen, in particular at least 70% and especially at least 80%) of the described sequences can be used to achieve the same practical effect.

- 5 Each of the above individual antigen sequences is also disclosed in Cole et al *Nature* 1998 393:537-544 and Camus *Microbiology* 2002 148:2967-2973. The genome of *M. tuberculosis* H37Rv is publicly available, for example at the Wellcome Trust Sanger Institute website (www.sanger.ac.uk/Projects/M_tuberculosis/) and elsewhere.
- 10 Many of the above antigens are also disclosed in U.S. patent application numbers 08/523,435, 08/523,436, 08/658,800, 08/659,683, 08/818,111, 08/818,112, 08/942,341, 08/942,578, 08/858,998, 08/859,381, 09/056,556, 09/072,596, 09/072,967, 09/073,009, 09/073,010, 09/223,040, 09/287,849 and in PCT patent applications PCT/US98/10407, PCT/US98/10514, PCT/US99/03265, PCT/US99/03268, PCT/US99/07717, WO97/09428 and WO97/09429,
- 15 WO98/16645, WO98/16646.

The compositions, polypeptides, and nucleic acids of the invention can also comprise additional polypeptides from other sources. For example, the compositions and fusion proteins of the invention can include polypeptides or nucleic acids encoding polypeptides, wherein the

- 20 polypeptide enhances expression of the antigen, e.g., NS1, an influenza virus protein (see, e.g. WO99/40188 and WO93/04175). The nucleic acids of the invention can be engineered based on codon preference in a species of choice, e.g., humans (in the case of *in vivo* expression) or a particular bacterium (in the case of polypeptide production).

- 25 The Rv2386c component may also be administered with one or more chemotherapeutic agents effective against tuberculosis (e.g. *M. tuberculosis* infection). Examples of such chemotherapeutic agents include, but are not limited to, amikacin, aminosalicylic acid, capreomycin, cycloserine, ethambutol, ethionamide, isoniazid, kanamycin, pyrazinamide, rifamycins (*i.e.*, rifampin, rifapentine and rifabutin), streptomycin, ofloxacin, ciprofloxacin,
- 30 clarithromycin, azithromycin and fluoroquinolones. Such chemotherapy is determined by the judgment of the treating physician using preferred drug combinations. "First-line" chemotherapeutic agents used to treat tuberculosis (e.g. *M. tuberculosis* infection) that is not drug resistant include isoniazid, rifampin, ethambutol, streptomycin and pyrazinamide. "Second-line" chemotherapeutic agents used to treat tuberculosis (e.g. *M. tuberculosis*
- 35 infection) that has demonstrated drug resistance to one or more "first-line" drugs include ofloxacin, ciprofloxacin, ethionamide, aminosalicylic acid, cycloserine, amikacin, kanamycin and capreomycin.

Conventional chemotherapeutic agents are generally administered over a relatively long period (ca. 9 months). Combination of conventional chemotherapeutic agents with the administration of a Rv2386c component according to the present invention may enable the chemotherapeutic treatment period to be reduced (e.g. to 8 months, 7 months, 6 months, 5 months, 4 months, 3 months or less) without a decrease in efficacy.

Of particular interest is the use of an Rv2386c component in conjunction with *Bacillus Calmette-Guerin* (BCG). For example, in the form of a modified BCG which recombinantly expresses Rv2386c (or a variant or fragment thereof as described herein). Alternatively, the Rv2386c component may be used to enhance the response of a subject to BCG vaccination, either by co-administration or by boosting a previous BCG vaccination. When used to enhance the response of a subject to BCG vaccination, the Rv2386c component may obviously be provided in the form of a polypeptide or a polynucleotide (optionally in conjunction with additional antigenic components as described above).

The skilled person will recognise that combinations of components need not be administered together and may be applied: separately or in combination; at the same time, sequentially or within a short period; though the same or through different routes. Nevertheless, for convenience it is generally desirable (where administration regimes are compatible) to administer a combination of components as a single composition.

The polypeptides, polynucleotides and compositions of the present invention will usually be administered to humans, but are effective in other mammals including domestic mammals (e.g., dogs, cats, rabbits, rats, mice, guinea pigs, hamsters, chinchillas) and agricultural mammals (e.g., cows, pigs, sheep, goats, horses).

IMMUNOGENIC FRAGMENTS

T cell epitopes are short contiguous stretches of amino acids which are recognised by T cells (e.g. CD4+ or CD8+ T cells). Identification of T cell epitopes may be achieved through epitope mapping experiments which are well known to the person skilled in the art (see, for example, Paul, *Fundamental Immunology*, 3rd ed., 243-247 (1993); Beißbarth et al *Bioinformatics* 2005 21(Suppl. 1):i29-i37).

Alternatively, epitopes may be predicted using the approaches discussed in the Examples.

As a result of the crucial involvement of the T cell response in tuberculosis, it is readily apparent that fragments of the full length Rv2386c polypeptide which contain at least one T cell epitope will be immunogenic and may contribute to immunoprotection. Such fragments are referred to herein as immunogenic fragments.

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Immunogenic fragments according to the present invention will typically comprise at least 9 contiguous amino acids from the full length polypeptide sequence (e.g. at least 10), such as at least 12 contiguous amino acids (e.g. at least 15 or at least 20 contiguous amino acids), in particular at least 50 contiguous amino acids, such as at least 100 contiguous amino acids (for example at least 200 contiguous amino acids). Suitably the immunogenic fragments will be at least 20%, such as at least 50%, at least 70% or at least 80% of the length of the full length polypeptide sequence.

It will be understood that in a diverse out-bred population, such as humans, different HLA types mean that specific epitopes may not be recognised by all members of the population. Consequently, to maximise the level of recognition and scale of immune response to a polypeptide, it is generally desirable that an immunogenic fragment contains a plurality of the epitopes from the full length sequence (suitably all epitopes).

Particular fragments of the Rv2386c protein which may be of use include those containing at least one CD4+ epitope, suitably at least two CD4+ epitopes and especially all CD4+ epitopes (such as those epitopes described in the Examples and in SEQ ID Nos: 30-52, particularly those associated with a plurality of HLA alleles, e.g. those associated with 2, 3, 4, 5 or more alleles).

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Other fragments of the Rv2386c protein which may be of use include those containing at least one CD8 epitope, suitably at least two CD8 epitopes and especially all CD8 epitopes (such as those epitopes described in the Examples and in SEQ ID Nos: 53-154, particularly those associated with a plurality of HLA alleles, e.g. those associated with 2, 3, 4, 5 or more alleles).

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Where an individual fragment of the full length polypeptide is used, such a fragment is considered to be immunogenic where it elicits a response which is at least 20%, suitably at least 50% and especially at least 75% (such as at least 90%) of the activity of the reference sequence in an *in vitro* restimulation assay of PBMC or whole blood with specific antigens (e.g. restimulation for a period of between several hours to up to two weeks, such as up to one day, 1 day to 1 week or 1 to 2 weeks) that measures the activation of the cells via lymphoproliferation, production of cytokines in the supernatant of culture (measured by ELISA,

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CBA etc) or characterisation of T and B cell responses by intra and extracellular staining (e.g. using antibodies specific to immune markers, such as CD3, CD4, CD8, IL2, TNFa, IFNg, CD40L, CD69 etc) followed by analysis with a flowcytometer. Suitably, a fragment is considered to be immunogenic where it elicits a response which is at least 20%, suitably at least 50% and especially at least 75% (such as at least 90%) of the activity of the reference sequence in a T cell proliferation and/or IFN-gamma production assay.

In some circumstances a plurality of fragments of the full length polypeptide (which may or may not be overlapping and may or may not cover the entirety of the full length sequence) may be used to obtain an equivalent biological response to the full length sequence itself. For example, at least two immunogenic fragments (such as three, four or five) as described above, which in combination provide at least 50%, suitably at least 75% and especially at least 90% activity of the reference sequence in an *in vitro* restimulation assay of PBMC or whole blood (e.g. a T cell proliferation and/or IFN-gamma production assay).

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VARIANTS

“Variants” or “conservatively modified variants” applies to both amino acid and nucleic acid sequences. With respect to particular nucleic acid sequences, conservatively modified variants refers to those nucleic acids which encode identical or essentially identical amino acid sequences, or where the nucleic acid does not encode an amino acid sequence, to essentially identical sequences.

Due to the degeneracy of the genetic code, a large number of functionally identical nucleic acids encode any given protein. For instance, the codons GCA, GCC, GCG and GCU all encode the amino acid alanine. Thus, at every position where an alanine is specified by a codon, the codon can be altered to any of the corresponding codons described without altering the encoded polypeptide. Such nucleic acid variations lead to “silent” or “degenerate” variants, which are one species of conservatively modified variations. Every nucleic acid sequence herein which encodes a polypeptide also describes every possible silent variation of the nucleic acid. One of skill will recognise that each codon in a nucleic acid (except AUG, which is ordinarily the only codon for methionine, and TGG, which is ordinarily the only codon for tryptophan) can be modified to yield a functionally identical molecule. Accordingly, each silent variation of a nucleic acid that encodes a polypeptide is implicit in each described sequence.

A polynucleotide of the invention may contain a number of silent variations (for example, 1-50, such as 1-25, in particular 1-5, and especially 1 codon(s) may be altered) when compared to

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the reference sequence. A polynucleotide of the invention may contain a number of non-silent conservative variations (for example, 1-50, such as 1-25, in particular 1-5, and especially 1 codon(s) may be altered) when compared to the reference sequence. Non-silent variations are those which result in a change in the encoded amino acid sequence (either through the substitution, deletion or addition of amino acid residues). Those skilled in the art will recognise that a particular polynucleotide sequence may contain both silent and non-silent conservative variations.

In respect of variants of a protein sequence, the skilled person will recognise that individual substitutions, deletions or additions to polypeptide, which alters, adds or deletes a single amino acid or a small percentage of amino acids is a "conservatively modified variant" where the alteration(s) results in the substitution of an amino acid with a functionally similar amino acid or the substitution/deletion/addition of residues which do not substantially impact the biological function of the variant.

Conservative substitution tables providing functionally similar amino acids are well known in the art. Such conservatively modified variants are in addition to and do not exclude polymorphic variants, interspecies homologs, and alleles of the invention.

A polypeptide of the invention may contain a number of conservative substitutions (for example, 1-50, such as 1-25, in particular 1-10, and especially 1 amino acid residue(s) may be altered) when compared to the reference sequence. In general, such conservative substitutions will fall within one of the amino-acid groupings specified below, though in some circumstances other substitutions may be possible without substantially affecting the immunogenic properties of the antigen. The following eight groups each contain amino acids that are typically conservative substitutions for one another:

- 1) Alanine (A), Glycine (G);
 - 2) Aspartic acid (D), Glutamic acid (E);
 - 3) Asparagine (N), Glutamine (Q);
 - 4) Arginine (R), Lysine (K);
 - 5) Isoleucine (I), Leucine (L), Methionine (M), Valine (V);
 - 6) Phenylalanine (F), Tyrosine (Y), Tryptophan (W);
 - 7) Serine (S), Threonine (T); and
 - 8) Cysteine (C), Methionine (M)
- (see, e.g., Creighton, *Proteins* 1984).

Suitably such substitutions do not occur in the region of an epitope, and do not therefore have a significant impact on the immunogenic properties of the antigen.

Protein variants may also include those wherein additional amino acids are inserted compared to the reference sequence, for example, such insertions may occur at 1-10 locations (such as 1-5 locations, suitably 1 or 2 locations, in particular 1 location) and may, for example, involve the addition of 50 or fewer amino acids at each location (such as 20 or fewer, in particular 10 or fewer, especially 5 or fewer). Suitably such insertions do not occur in the region of an epitope, and do not therefore have a significant impact on the immunogenic properties of the antigen. One example of insertions includes a short stretch of histidine residues (e.g. 2-6 residues) to aid expression and/or purification of the antigen in question.

Protein variants include those wherein amino acids have been deleted compared to the reference sequence, for example, such deletions may occur at 1-10 locations (such as 1-5 locations, suitably 1 or 2 locations, in particular 1 location) and may, for example, involve the deletion of 50 or fewer amino acids at each location (such as 20 or fewer, in particular 10 or fewer, especially 5 or fewer). Suitably such deletions do not occur in the region of an epitope, and do not therefore have a significant impact on the immunogenic properties of the antigen.

The skilled person will recognise that a particular protein variant may comprise substitutions, deletions and additions (or any combination thereof).

Methods of determining the epitope regions of an antigen are described and exemplified in the Examples.

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Variants preferably exhibit at least about 70% identity, more preferably at least about 80% identity and most preferably at least about 90% identity (such as at least about 95%, at least about 98% or at least about 99%) to the associated reference sequence.

The terms "identical" or percent "identity," in the context of two or more nucleic acids or polypeptide sequences, refer to two or more sequences or sub-sequences that are the same or have a specified percentage of amino acid residues or nucleotides that are the same (i.e., 70% identity, optionally 75%, 80%, 85%, 90%, 95%, 98% or 99% identity over a specified region), when compared and aligned for maximum correspondence over a comparison window, or designated region as measured using one of the following sequence comparison algorithms or by manual alignment and visual inspection. Such sequences are then said to be "substantially identical." This definition also refers to the complement of a test sequence.

Optionally, the identity exists over a region that is at least about 25 to about 50 amino acids or nucleotides in length, or optionally over a region that is 75-100 amino acids or nucleotides in length. Suitably, the comparison is performed over a window corresponding to the entire length of the reference sequence.

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For sequence comparison, typically one sequence acts as a reference sequence, to which test sequences are compared. When using a sequence comparison algorithm, test and reference sequences are entered into a computer, subsequence coordinates are designated, if necessary, and sequence algorithm program parameters are designated. Default program parameters can be used, or alternative parameters can be designated. The sequence comparison algorithm then calculates the percent sequence identities for the test sequences relative to the reference sequence, based on the program parameters.

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A "comparison window", as used herein, references to a segment in which a sequence may be compared to a reference sequence of the same number of contiguous positions after the two sequences are optimally aligned. Methods of alignment of sequences for comparison are well-known in the art. Optimal alignment of sequences for comparison can be conducted, e.g., by the local homology algorithm of Smith & Waterman, *Adv. Appl. Math.* 2:482 (1981), by the homology alignment algorithm of Needleman & Wunsch, *J. Mol. Biol.* 48:443 (1970), by the search for similarity method of Pearson & Lipman, *Proc. Nat'l. Acad. Sci. USA* 85:2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, WI), or by manual alignment and visual inspection (see, e.g., *Current Protocols in Molecular Biology* (Ausubel et al., eds. 1995 supplement)).

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One example of a useful algorithm is PILEUP. PILEUP creates a multiple sequence alignment from a group of related sequences using progressive, pairwise alignments to show relationship and percent sequence identity. It also plots a tree or dendrogram showing the clustering relationships used to create the alignment. PILEUP uses a simplification of the progressive alignment method of Feng & Doolittle, *J. Mol. Evol.* 35:351-360 (1987). The method used is similar to the method described by Higgins & Sharp, *CABIOS* 5:151-153 (1989). The program can align up to 300 sequences, each of a maximum length of 5,000 nucleotides or amino acids. The multiple alignment procedure begins with the pairwise alignment of the two most similar sequences, producing a cluster of two aligned sequences. This cluster is then aligned to the next most related sequence or cluster of aligned sequences. Two clusters of sequences are aligned by a simple extension of the pairwise alignment of two individual sequences. The final alignment is achieved by a series of progressive, pairwise alignments. The program is

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run by designating specific sequences and their amino acid or nucleotide coordinates for regions of sequence comparison and by designating the program parameters. Using PILEUP, a reference sequence is compared to other test sequences to determine the percent sequence identity relationship using the following parameters: default gap weight (3.00), default gap
5 length weight (0.10), and weighted end gaps. PILEUP can be obtained from the GCG sequence analysis software package, e.g., version 7.0 (Devereaux *et al.*, *Nuc. Acids Res.* 12:387-395 (1984).

Another example of algorithm that is suitable for determining percent sequence identity and
10 sequence similarity are the BLAST and BLAST 2.0 algorithms, which are described in Altschul *et al.*, *Nuc. Acids Res.* 25:3389-3402 (1977) and Altschul *et al.*, *J. Mol. Biol.* 215:403-410 (1990), respectively. Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information (website at www.ncbi.nlm.nih.gov/). This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short
15 words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighbourhood word score threshold (Altschul *et al.*, *supra*). These initial neighbourhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are extended in both directions along each sequence for as far as the
20 cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always > 0) and N (penalty score for mismatching residues; always < 0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its
25 maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T, and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) of 10, M=5, N=-4 and a comparison of both
30 strands. For amino acid sequences, the BLASTP program uses as defaults a wordlength of 3, and expectation (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff & Henikoff, *Proc. Natl. Acad. Sci. USA* 89:10915 (1989)) alignments (B) of 50, expectation (E) of 10, M=5, N=-4, and a comparison of both strands.

35 The BLAST algorithm also performs a statistical analysis of the similarity between two sequences (see, e.g., Karlin & Altschul, *Proc. Nat'l. Acad. Sci. USA* 90:5873-5787 (1993)). One measure of similarity provided by the BLAST algorithm is the smallest sum probability

(P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a nucleic acid is considered similar to a reference sequence if the smallest sum probability in a comparison of the test nucleic acid to the reference nucleic acid is less than about 0.2, more preferably less than about 0.01, and most preferably less than about 0.001.

The present invention also extends to polynucleotides comprising a first nucleotide sequence which selectively hybridises under moderately stringent conditions (such as under highly stringent conditions) to the complement of a second nucleotide sequence which encodes a polypeptide comprising:

- (i) an Rv2386c protein sequence;
- (ii) a variant of an Rv2386c protein sequence; or
- (iii) an immunogenic fragment of an Rv2386c protein sequence.

The phrase “highly stringent hybridisation conditions” refers to conditions under which a probe will hybridise to its target subsequence, typically in a complex mixture of nucleic acid, but to no other sequences. Highly stringent conditions are sequence-dependent and will be different in different circumstances. Longer sequences hybridise specifically at higher temperatures. An extensive guide to the hybridisation of nucleic acids is found in Tijssen, *Techniques in Biochemistry and Molecular Biology--Hybridization with Nucleic Probes*, “Overview of principles of hybridisation and the strategy of nucleic acid assays” (1993). Generally, highly stringent conditions are selected to be about 5-10°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength pH. The T_m is the temperature (under defined ionic strength, pH, and nucleic concentration) at which 50% of the probes complementary to the target hybridise to the target sequence at equilibrium (as the target sequences are present in excess, at T_m , 50% of the probes are occupied at equilibrium). Highly stringent conditions will be those in which the salt concentration is less than about 1.0 M sodium ion, typically about 0.01 to 1.0 M sodium ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes (e.g., 10 to 50 nucleotides) and at least about 60°C for long probes (e.g., greater than 50 nucleotides). Highly stringent conditions may also be achieved with the addition of destabilising agents such as formamide. For selective or specific hybridisation, a positive signal is at least two times background, optionally 10 times background hybridisation.

Exemplary highly stringent hybridisation conditions can be as following: 50% formamide, 5x SSC, and 1% SDS, incubating at 42°C, or, 5x SSC, 1% SDS, incubating at 65°C, with wash in 0.2x SSC, and 0.1% SDS at 65°C.

Nucleic acids that do not hybridise to each other under highly stringent conditions are still functionally equivalent if the polypeptides which they encode are substantially identical. This occurs, for example, when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code. In such cases, the nucleic acids typically hybridise under moderately stringent hybridisation conditions.

Exemplary “moderately stringent hybridisation conditions” include a hybridisation in a buffer of 40% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 1X SSC at 45°C. A positive hybridisation is at least twice background. Those of ordinary skill will readily recognise that alternative hybridisation and wash conditions can be utilised to provide conditions of similar stringency.

The phrase “selectively (or specifically) hybridises to” refers to the binding, duplexing, or hybridising of a molecule only to a particular nucleotide sequence under stringent hybridisation conditions when that sequence is present in a complex mixture (e.g., total cellular or library DNA or RNA).

In any event, variants of a polypeptide sequence will have essentially the same activity as the reference sequence (in the case of polynucleotides, variant polynucleotide sequences will encode a polypeptide which has essentially the same activity as the reference sequence). By essentially the same activity is meant at least 50%, suitably at least 75% and especially at least 90% activity of the reference sequence in an *in vitro* restimulation assay of PBMC or whole blood with specific antigens (e.g. restimulation for a period of between several hours to up to two weeks, such as up to one day, 1 day to 1 week or 1 to 2 weeks) that measures the activation of the cells via lymphoproliferation, production of cytokines in the supernatant of culture (measured by ELISA, CBA etc) or characterisation of T and B cell responses by intra and extracellular staining (e.g. using antibodies specific to immune markers, such as CD3, CD4, CD8, IL2, TNFa, IFNg, CD40L, CD69 etc) followed by analysis with a flowcytometer. Suitably, by essentially the same activity is meant at least 50%, suitably at least 75% and especially at least 90% activity of the reference sequence in a T cell proliferation and/or IFN-gamma production assay

POLYNUCLEOTIDE COMPOSITIONS

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As used herein, the term “polynucleotide” refers to a molecule that has been isolated free of total genomic DNA of a particular species. Therefore, a polynucleotide encoding a polypeptide

refers to a polynucleotide segment that contains one or more coding sequences yet is substantially isolated away from, or purified free from, total genomic DNA of the species from which the polynucleotide is obtained.

5 As will be understood by those skilled in the art, the polynucleotides of this invention can include genomic sequences, extra-genomic and plasmid-encoded sequences and smaller engineered gene segments that express, or may be adapted to express, proteins, polypeptides, peptides and the like. Such segments may be naturally isolated, or modified synthetically by the hand of man.

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“Isolated,” as used herein, means that a polynucleotide is substantially away from other coding sequences, and that the polynucleotide does not contain large portions of unrelated coding DNA, such as large chromosomal fragments or other functional genes or polypeptide coding regions. An isolated nucleic acid is separated from other open reading frames that flank the
15 gene and encode proteins other than the gene. Of course, this refers to the DNA segment as originally isolated, and does not exclude genes or coding regions later added to the segment by the hand of man.

As will be recognised by the skilled artisan, polynucleotides may be single-stranded (coding or antisense) or double-stranded, and may be DNA (genomic, cDNA or synthetic) or RNA
20 molecules. RNA molecules include HnRNA molecules, which contain introns and correspond to a DNA molecule in a one-to-one manner, and mRNA molecules, which do not contain introns. Additional coding or non-coding sequences may, but need not, be present within a polynucleotide of the present invention, and a polynucleotide may, but need not, be linked to other molecules and/or support materials.

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Polynucleotides may comprise a native sequence (i.e., an endogenous sequence that encodes a *Mycobacterium* antigen or a portion thereof) or may comprise a variant, or a biological or functional equivalent of such a sequence. Polynucleotide variants may contain one or more substitutions, additions, deletions and/or insertions, as further described below, preferably
30 such that the immunogenicity of the encoded polypeptide is not diminished, relative to the reference protein. The effect on the immunogenicity of the encoded polypeptide may generally be assessed as described herein.

In additional embodiments, the present invention provides isolated polynucleotides and
35 polypeptides comprising various lengths of contiguous stretches of sequence identical to or complementary to one or more of the sequences disclosed herein. For example,

polynucleotides are provided by this invention that comprise at least about 30, 40, 50, 75, 100, 150, 200, 300, 400, 500 or 1000 or more contiguous nucleotides of the reference sequence disclosed herein as well as all intermediate lengths there between. It will be readily understood that "intermediate lengths", in this context, means any length between the quoted values, such as 30, 31, 32, etc.; 50, 51, 52, 53, etc.; 100, 101, 102, 103, etc.; 150, 151, 152, 153, etc.; including all integers through 200-500; 500-1,000, and the like.

Moreover, it will be appreciated by those of ordinary skill in the art that, as a result of the degeneracy of the genetic code, there are many nucleotide sequences that encode a polypeptide as described herein. Some of these polynucleotides bear relatively low identity to the nucleotide sequence of any native gene. Nonetheless, polynucleotides that vary due to differences in codon usage are specifically contemplated by the present invention, for example polynucleotides that are optimized for human and/or primate codon selection. Further, alleles of the genes comprising the polynucleotide sequences provided herein are within the scope of the present invention. Alleles are endogenous genes that are altered as a result of one or more mutations, such as deletions, additions and/or substitutions of nucleotides. The resulting mRNA and protein may, but need not, have an altered structure or function. Alleles may be identified using standard techniques (such as hybridisation, amplification and/or database sequence comparison).

20

POLYNUCLEOTIDE IDENTIFICATION AND CHARACTERISATION

Polynucleotides may be identified, prepared and/or manipulated using any of a variety of well established techniques. For example, a polynucleotide may be identified, as described in more detail below, by screening a microarray of cDNAs. Such screens may be performed, for example, using a Synteni microarray (Palo Alto, CA) according to the manufacturer's instructions (and essentially as described by Schena *et al.*, *Proc. Natl. Acad. Sci. USA* 93:10614-10619 (1996) and Heller *et al.*, *Proc. Natl. Acad. Sci. USA* 94:2150-2155 (1997)). Alternatively, polynucleotides may be amplified from cDNA prepared from cells expressing the proteins described herein, such as *M. tuberculosis* cells. Such polynucleotides may be amplified via polymerase chain reaction (PCR). For this approach, sequence-specific primers may be designed based on the sequences provided herein, and may be purchased or synthesised.

An amplified portion of a polynucleotide may be used to isolate a full length gene from a suitable library (e.g., a *M. tuberculosis* cDNA library) using well known techniques. Within such techniques, a library (cDNA or genomic) is screened using one or more polynucleotide probes

or primers suitable for amplification. Preferably, a library is size-selected to include larger molecules. Random primed libraries may also be preferred for identifying 5' and upstream regions of genes. Genomic libraries are preferred for obtaining introns and extending 5' sequences.

5

For hybridisation techniques, a partial sequence may be labeled (e.g., by nick-translation or end-labeling with ^{32}P) using well known techniques. A bacterial or bacteriophage library is then generally screened by hybridising filters containing denatured bacterial colonies (or lawns containing phage plaques) with the labeled probe (see Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual* (2000)). Hybridising colonies or plaques are selected and expanded, and the DNA is isolated for further analysis. cDNA clones may be analyzed to determine the amount of additional sequence by, for example, PCR using a primer from the partial sequence and a primer from the vector. Restriction maps and partial sequences may be generated to identify one or more overlapping clones. The complete sequence may then be determined using standard techniques, which may involve generating a series of deletion clones. The resulting overlapping sequences can then be assembled into a single contiguous sequence. A full length cDNA molecule can be generated by ligating suitable fragments, using well known techniques.

Alternatively, there are numerous amplification techniques for obtaining a full length coding sequence from a partial cDNA sequence. Within such techniques, amplification is generally performed via PCR. Any of a variety of commercially available kits may be used to perform the amplification step. Primers may be designed using, for example, software well known in the art. Primers are preferably 22-30 nucleotides in length, have a GC content of at least 50% and anneal to the target sequence at temperatures of about 68°C to 72°C. The amplified region may be sequenced as described above, and overlapping sequences assembled into a contiguous sequence.

One such amplification technique is inverse PCR (see Triglia *et al.*, *Nucl. Acids Res.* 16:8186 (1988)), which uses restriction enzymes to generate a fragment in the known region of the gene. The fragment is then circularised by intramolecular ligation and used as a template for PCR with divergent primers derived from the known region. Within an alternative approach, sequences adjacent to a partial sequence may be retrieved by amplification with a primer to a linker sequence and a primer specific to a known region. The amplified sequences are typically subjected to a second round of amplification with the same linker primer and a second primer specific to the known region. A variation on this procedure, which employs two primers that initiate extension in opposite directions from the known sequence, is described in WO

96/38591. Another such technique is known as “rapid amplification of cDNA ends” or RACE. This technique involves the use of an internal primer and an external primer, which hybridises to a polyA region or vector sequence, to identify sequences that are 5’ and 3’ of a known sequence. Additional techniques include capture PCR (Lagerstrom *et al.*, *PCR Methods*
5 *Applic.* 1:111-19 (1991)) and walking PCR (Parker *et al.*, *Nucl. Acids. Res.* 19:3055-60 (1991)). Other methods employing amplification may also be employed to obtain a full length cDNA sequence.

10 In certain instances, it is possible to obtain a full length cDNA sequence by analysis of sequences provided in an expressed sequence tag (EST) database, such as that available from GenBank. Searches for overlapping ESTs may generally be performed using well known programs (e.g., NCBI BLAST searches), and such ESTs may be used to generate a contiguous full length sequence. Full length DNA sequences may also be obtained by analysis of genomic fragments.

15

POLYNUCLEOTIDE EXPRESSION IN HOST CELLS

Polynucleotide sequences or fragments thereof which encode polypeptides, or fusion proteins or functional equivalents thereof, may be used in recombinant DNA molecules to direct
20 expression of a polypeptide in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences that encode substantially the same or a functionally equivalent amino acid sequence may be produced and these sequences may be used to clone and express a given polypeptide.

25 As will be understood by those of skill in the art, it may be advantageous in some instances to produce polypeptide-encoding nucleotide sequences possessing non-naturally occurring codons. For example, codons preferred by a particular prokaryotic or eukaryotic host can be selected to increase the rate of protein expression or to produce a recombinant RNA transcript having desirable properties, such as a half-life which is longer than that of a transcript
30 generated from the naturally occurring sequence.

Moreover, the polynucleotide sequences can be engineered using methods generally known in the art in order to alter polypeptide encoding sequences for a variety of reasons, including but not limited to, alterations which modify the cloning, processing, and/or expression of the gene
35 product. For example, DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. In addition, site-directed mutagenesis may be used to insert new restriction sites, alter

glycosylation patterns, change codon preference, produce splice variants, or introduce mutations, and so forth.

5 Natural, modified, or recombinant nucleic acid sequences may be ligated to a heterologous sequence to encode a fusion protein. For example, to screen peptide libraries for inhibitors of polypeptide activity, it may be useful to encode a chimeric protein that can be recognised by a commercially available antibody. A fusion protein may also be engineered to contain a cleavage site located between the polypeptide-encoding sequence and the heterologous protein sequence, so that the polypeptide may be cleaved and purified away from the
10 heterologous moiety.

Sequences encoding a desired polypeptide may be synthesised, in whole or in part, using chemical methods well known in the art (see Caruthers, *M. H. et al., Nucl. Acids Res. Symp. Ser.* pp. 215-223 (1980), Horn *et al., Nucl. Acids Res. Symp. Ser.* pp. 225-232 (1980)).
15 Alternatively, the protein itself may be produced using chemical methods to synthesize the amino acid sequence of a polypeptide, or a portion thereof. For example, peptide synthesis can be performed using various solid-phase techniques (Roberge *et al., Science* 269:202-204 (1995)) and automated synthesis may be achieved, for example, using the ABI 431A Peptide Synthesizer (Perkin Elmer, Palo Alto, CA).

20 A newly synthesised peptide may be substantially purified by preparative high performance liquid chromatography (e.g., Creighton, *Proteins, Structures and Molecular Principles* (1983)) or other comparable techniques available in the art. The composition of the synthetic peptides may be confirmed by amino acid analysis or sequencing (e.g., the Edman degradation
25 procedure). Additionally, the amino acid sequence of a polypeptide, or any part thereof, may be altered during direct synthesis and/or combined using chemical methods with sequences from other proteins, or any part thereof, to produce a variant polypeptide.

In order to express a desired polypeptide, the nucleotide sequences encoding the polypeptide,
30 or functional equivalents, may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for the transcription and translation of the inserted coding sequence. Methods which are well known to those skilled in the art may be used to construct expression vectors containing sequences encoding a polypeptide of interest and appropriate transcriptional and translational control elements. These methods include *in vitro*
35 recombinant DNA techniques, synthetic techniques, and *in vivo* genetic recombination. Such techniques are described in Sambrook *et al., Molecular Cloning, A Laboratory Manual* (2000), and Ausubel *et al., Current Protocols in Molecular Biology* (updated annually).

- A variety of expression vector/host systems may be utilised to contain and express polynucleotide sequences. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression
5 vectors; yeast transformed with yeast expression vectors; insect cell systems infected with virus expression vectors (e.g., baculovirus); plant cell systems transformed with virus expression vectors (e.g., cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems.
- 10 The “control elements” or “regulatory sequences” present in an expression vector are those non-translated regions of the vector—enhancers, promoters, 5’ and 3’ untranslated regions—which interact with host cellular proteins to carry out transcription and translation. Such elements may vary in their strength and specificity. Depending on the vector system and host utilised, any number of suitable transcription and translation elements, including constitutive
15 and inducible promoters, may be used. For example, when cloning in bacterial systems, inducible promoters such as the hybrid lacZ promoter of the PBLUESCRIPT phagemid (Stratagene, La Jolla, Calif.) or PSFORT1 plasmid (Gibco BRL, Gaithersburg, MD) and the like may be used. In mammalian cell systems, promoters from mammalian genes or from mammalian viruses are generally preferred. If it is necessary to generate a cell line that
20 contains multiple copies of the sequence encoding a polypeptide, vectors based on SV40 or EBV may be advantageously used with an appropriate selectable marker.

- In bacterial systems, a number of expression vectors may be selected depending upon the use intended for the expressed polypeptide. For example, when large quantities are needed, for
25 example for the induction of antibodies, vectors which direct high level expression of fusion proteins that are readily purified may be used. Such vectors include, but are not limited to, the multifunctional *E. coli* cloning and expression vectors such as BLUESCRIPT (Stratagene), in which the sequence encoding the polypeptide of interest may be ligated into the vector in frame with sequences for the amino-terminal Met and the subsequent 7 residues of β -galactosidase so that a hybrid protein is produced; pIN vectors (Van Heeke & Schuster, *J. Biol. Chem.* 264:5503-5509 (1989)); and the like. pGEX Vectors (Promega, Madison, Wis.) may
30 also be used to express foreign polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified from lysed cells by adsorption to glutathione-agarose beads followed by elution in the presence of free
35 glutathione. Proteins made in such systems may be designed to include heparin, thrombin, or factor XA protease cleavage sites so that the cloned polypeptide of interest can be released from the GST moiety at will.

In the yeast, *Saccharomyces cerevisiae*, a number of vectors containing constitutive or inducible promoters such as alpha factor, alcohol oxidase, and PGH may be used. Other vectors containing constitutive or inducible promoters include GAP, PGK, GAL and ADH. For reviews, see Ausubel *et al.* (*supra*) and Grant *et al.*, *Methods Enzymol.* 153:516-544 (1987) and Romas *et al.* *Yeast* 8 423-88 (1992).

In cases where plant expression vectors are used, the expression of sequences encoding polypeptides may be driven by any of a number of promoters. For example, viral promoters such as the 35S and 19S promoters of CaMV may be used alone or in combination with the omega leader sequence from TMV (Takamatsu, *EMBO J.* 6:307-311 (1987)). Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used (Coruzzi *et al.*, *EMBO J.* 3:1671-1680 (1984); Broglie *et al.*, *Science* 224:838-843 (1984); and Winter *et al.*, *Results Probl. Cell Differ.* 17:85-105 (1991)). These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. Such techniques are described in a number of generally available reviews (see, e.g., Hobbs *in McGraw Hill Yearbook of Science and Technology* pp. 191-196 (1992)).

An insect system may also be used to express a polypeptide of interest. For example, in one such system, *Autographa californica* nuclear polyhedrosis virus (AcNPV) is used as a vector to express foreign genes in *Spodoptera frugiperda* cells or in *Trichoplusia larvae*. The sequences encoding the polypeptide may be cloned into a non-essential region of the virus, such as the polyhedrin gene, and placed under control of the polyhedrin promoter. Successful insertion of the polypeptide-encoding sequence will render the polyhedrin gene inactive and produce recombinant virus lacking coat protein. The recombinant viruses may then be used to infect, for example, *S. frugiperda* cells or *Trichoplusia larvae* in which the polypeptide of interest may be expressed (Engelhard *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 91:3224-3227 (1994)).

In mammalian host cells, a number of viral-based expression systems are generally available. For example, in cases where an adenovirus is used as an expression vector, sequences encoding a polypeptide of interest may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain a viable virus which is capable of expressing the polypeptide in infected host cells (Logan & Shenk, *Proc. Natl. Acad. Sci. U.S.A.* 81:3655-3659 (1984)). In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. Methods and protocols for working with adenovirus vectors are reviewed in Wold, *Adenovirus*

Methods and Protocols, 1998. Additional references regarding use of adenovirus vectors can be found in *Adenovirus: A Medical Dictionary, Bibliography, and Annotated Research Guide to Internet References*, 2004.

- 5 Specific initiation signals may also be used to achieve more efficient translation of sequences encoding a polypeptide of interest. Such signals include the ATG initiation codon and adjacent sequences. In cases where sequences encoding the polypeptide, its initiation codon, and upstream sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only
10 coding sequence, or a portion thereof, is inserted, exogenous translational control signals including the ATG initiation codon should be provided. Furthermore, the initiation codon should be in the correct reading frame to ensure translation of the entire insert. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers which
15 are appropriate for the particular cell system which is used, such as those described in the literature (Scharf. *et al.*, *Results Probl. Cell Differ.* 20:125-162 (1994)).

In addition, a host cell strain may be chosen for its ability to modulate the expression of the inserted sequences or to process the expressed protein in the desired fashion. Such
20 modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" form of the protein may also be used to facilitate correct insertion, folding and/or function. Different host cells such as CHO, HeLa, MDCK, HEK293, and WI38, which have specific cellular machinery and characteristic mechanisms for such post-translational
25 activities, may be chosen to ensure the correct modification and processing of the foreign protein.

For long-term, high-yield production of recombinant proteins, stable expression is generally preferred. For example, cell lines which stably express a polynucleotide of interest may be
30 transformed using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for 1-2 days in an enriched media before they are switched to selective media. The purpose of the selectable marker is to confer resistance to selection, and its presence allows growth and recovery of
35 cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be proliferated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase (Wigler *et al.*, *Cell* 11:223-32 (1977)) and adenine phosphoribosyltransferase (Lowy *et al.*, *Cell* 22:817-23 (1990))
5 genes which can be employed in tk.sup.- or aprt.sup.- cells, respectively. Also, antimetabolite, antibiotic or herbicide resistance can be used as the basis for selection; for example, dhfr which confers resistance to methotrexate (Wigler *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 77:3567-70 (1980)); npt, which confers resistance to the aminoglycosides, neomycin and G-418 (Colbere-Garapin *et al.*, *J. Mol. Biol.* 150:1-14 (1981)); and als or pat, which confer resistance
10 to chlorsulfuron and phosphinotricin acetyltransferase, respectively (Murry, *supra*). Additional selectable genes have been described, for example, trpB, which allows cells to utilise indole in place of tryptophan, or hisD, which allows cells to utilise histinol in place of histidine (Hartman & Mulligan, *Proc. Natl. Acad. Sci. U.S.A.* 85:8047-51 (1988)). Recently, the use of visible
15 markers has gained popularity with such markers as anthocyanins, β -glucuronidase and its substrate GUS, and luciferase and its substrate luciferin, being widely used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system (Rhodes *et al.*, *Methods Mol. Biol.* 55:121-131 (1995)).

Although the presence/absence of marker gene expression suggests that the gene of interest
20 is also present, its presence and expression may need to be confirmed. For example, if the sequence encoding a polypeptide is inserted within a marker gene sequence, recombinant cells containing sequences can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a polypeptide-encoding sequence under the control of a single promoter. Expression of the marker gene in response to induction
25 or selection usually indicates expression of the tandem gene as well.

Alternatively, host cells which contain and express a desired polynucleotide sequence may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridisations and protein bioassay or
30 immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein.

A variety of protocols for detecting and measuring the expression of polynucleotide-encoded products, using either polyclonal or monoclonal antibodies specific for the product are known in
35 the art. Examples include enzyme-linked immunosorbent assay (ELISA), radioimmunoassay (RIA), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilising monoclonal antibodies reactive to two non-interfering epitopes on a

given polypeptide may be preferred for some applications, but a competitive binding assay may also be employed. These and other assays are described, among other places, in Hampton *et al.*, *Serological Methods, a Laboratory Manual* (1990) and Maddox *et al.*, *J. Exp. Med.* 158:1211-1216 (1983).

5

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labelled hybridisation or PCR probes for detecting sequences related to polynucleotides include oligolabeling, nick translation, end-labelling or PCR amplification using a labelled nucleotide.

10

Alternatively, the sequences, or any portions thereof may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes *in vitro* by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits. Suitable reporter molecules or labels, which may be used include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

15

Host cells transformed with a polynucleotide sequence of interest may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a recombinant cell may be secreted or contained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides may be designed to contain signal sequences which direct secretion of the encoded polypeptide through a prokaryotic or eukaryotic cell membrane.

20

Other recombinant constructions may be used to join sequences encoding a polypeptide of interest to nucleotide sequence encoding a polypeptide domain which will facilitate purification of soluble proteins. Such purification facilitating domains include, but are not limited to, metal chelating peptides such as histidine-tryptophan modules that allow purification on immobilized metals, protein A domains that allow purification on immobilised immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system (Immunex Corp., Seattle, Wash.). The inclusion of cleavable linker sequences such as those specific for Factor XA or enterokinase (Invitrogen, San Diego, Calif.) between the purification domain and the encoded polypeptide may be used to facilitate purification. One such expression vector provides for expression of a fusion protein containing a polypeptide of interest and a nucleic acid encoding 6 histidine residues preceding a thioredoxin or an enterokinase cleavage site. The histidine residues facilitate purification on IMIAC (immobilised metal ion affinity chromatography) as described in Porath *et al.*, *Prot. Exp. Purif.* 3:263-281 (1992) while the enterokinase cleavage

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site provides a means for purifying the desired polypeptide from the fusion protein. A discussion of vectors which contain fusion proteins is provided in Kroll *et al.*, *DNA Cell Biol.* 12:441-453 (1993)).

5 IN VIVO POLYNUCLEOTIDE DELIVERY TECHNIQUES

In additional embodiments, genetic constructs comprising one or more of the polynucleotides of the invention are introduced into cells *in vivo*. This may be achieved using any of a variety or well known approaches, several of which are outlined below for the purpose of illustration.

10

1. ADENOVIRUS

One of the preferred methods for *in vivo* delivery of one or more nucleic acid sequences involves the use of an adenovirus expression vector. "Adenovirus expression vector" is meant to include those constructs containing adenovirus sequences sufficient to (a) support packaging of the construct and (b) to express a polynucleotide that has been cloned therein in a sense or antisense orientation. Of course, in the context of an antisense construct, expression does not require that the gene product be synthesised.

15

The expression vector comprises a genetically engineered form of an adenovirus. Knowledge of the genetic organisation of adenovirus, a 36 kb, linear, double-stranded DNA virus, allows substitution of large pieces of adenoviral DNA with foreign sequences up to 7 kb (Grunhaus & Horwitz, 1992). In contrast to retrovirus, the adenoviral infection of host cells does not result in chromosomal integration because adenoviral DNA can replicate in an episomal manner without potential genotoxicity. Also, adenoviruses are structurally stable, and no genome rearrangement has been detected after extensive amplification. Adenovirus can infect virtually all epithelial cells regardless of their cell cycle stage. So far, adenoviral infection appears to be linked only to mild disease such as acute respiratory disease in humans.

20

Adenovirus is particularly suitable for use as a gene transfer vector because of its mid-sized genome, ease of manipulation, high titre, wide target-cell range and high infectivity. Both ends of the viral genome contain 100-200 base pair inverted repeats (ITRs), which are *cis* elements necessary for viral DNA replication and packaging. The early (E) and late (L) regions of the genome contain different transcription units that are divided by the onset of viral DNA replication. The E1 region (E1A and E1B) encodes proteins responsible for the regulation of transcription of the viral genome and a few cellular genes. The expression of the E2 region (E2A and E2B) results in the synthesis of the proteins for viral DNA replication. These proteins

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are involved in DNA replication, late gene expression and host cell shut-off (Renan, 1990). The products of the late genes, including the majority of the viral capsid proteins, are expressed only after significant processing of a single primary transcript issued by the major late promoter (MLP). The MLP, (located at 16.8 m.u.) is particularly efficient during the late
5 phase of infection, and all the mRNA's issued from this promoter possess a 5'-tripartite leader (TPL) sequence which makes them preferred mRNA's for translation.

In a current system, recombinant adenovirus is generated from homologous recombination between shuttle vector and provirus vector. Due to the possible recombination between two
10 proviral vectors, wild-type adenovirus may be generated from this process. Therefore, it is critical to isolate a single clone of virus from an individual plaque and examine its genomic structure.

Generation and propagation of the current adenovirus vectors, which are replication deficient,
15 depend on a unique helper cell line, designated 293, which was transformed from human embryonic kidney cells by Ad5 DNA fragments and constitutively expresses E1 proteins (Graham *et al.*, 1977). Since the E3 region is dispensable from the adenovirus genome (Jones & Shenk, 1978), the current adenovirus vectors, with the help of 293 cells, carry foreign DNA in either the E1, the D3 or both regions (Graham & Prevec, 1991). In nature, adenovirus can
20 package approximately 105% of the wild-type genome (Ghosh-Choudhury *et al.*, 1987), providing capacity for about 2 extra kB of DNA. Combined with the approximately 5.5 kB of DNA that is replaceable in the E1 and E3 regions, the maximum capacity of the current adenovirus vector is under 7.5 kB, or about 15% of the total length of the vector. More than
25 vector-borne cytotoxicity. Also, the replication deficiency of the E1-deleted virus is incomplete. For example, leakage of viral gene expression has been observed with the currently available vectors at high multiplicities of infection (MOI) (Mulligan, 1993).

Helper cell lines may be derived from human cells such as human embryonic kidney cells,
30 muscle cells, hematopoietic cells or other human embryonic mesenchymal or epithelial cells. Alternatively, the helper cells may be derived from the cells of other mammalian species that are permissive for human adenovirus. Such cells include, e.g., Vero cells or other monkey embryonic mesenchymal or epithelial cells. As stated above, the currently preferred helper cell line is 293.

35
Racher *et al.* (1995) have disclosed improved methods for culturing 293 cells and propagating adenovirus. In one format, natural cell aggregates are grown by inoculating individual cells

into 1 litre siliconised spinner flasks (Techne, Cambridge, UK) containing 100-200 ml of medium. Following stirring at 40 rpm, the cell viability is estimated with trypan blue. In another format, Fibra-Cel microcarriers (Bibby Sterlin, Stone, UK) (5 g/l) is employed as follows. A cell inoculum, resuspended in 5 ml of medium, is added to the carrier (50 ml) in a 250 ml

5 Erlenmeyer flask and left stationary, with occasional agitation, for 1 to 4 h. The medium is then replaced with 50 ml of fresh medium and shaking initiated. For virus production, cells are allowed to grow to about 80% confluence, after which time the medium is replaced (to 25% of the final volume) and adenovirus added at an MOI of 0.05. Cultures are left stationary overnight, following which the volume is increased to 100% and shaking commenced for

10 another 72 h.

Other than the requirement that the adenovirus vector be replication defective, or at least conditionally defective, the nature of the adenovirus vector is not believed to be crucial to the successful practice of the invention. The adenovirus may be of any of the 42 different known

15 serotypes or subgroups A-F. Adenovirus type 5 of subgroup C is the preferred starting material in order to obtain a conditional replication-defective adenovirus vector for use in the present invention, since Adenovirus type 5 is a human adenovirus about which a great deal of biochemical and genetic information is known, and it has historically been used for most constructions employing adenovirus as a vector.

20 As stated above, the typical vector according to the present invention is replication defective and will not have an adenovirus E1 region. Thus, it will be most convenient to introduce the polynucleotide encoding the gene of interest at the position from which the E1-coding sequences have been removed. However, the position of insertion of the construct within the

25 adenovirus sequences is not critical to the invention. The polynucleotide encoding the gene of interest may also be inserted in lieu of the deleted E3 region in E3 replacement vectors as described by Karlsson *et al.* (1986) or in the E4 region where a helper cell line or helper virus complements the E4 defect.

30 Adenovirus is easy to grow and manipulate and exhibits broad host range *in vitro* and *in vivo*. This group of viruses can be obtained in high titres, e.g., 10^9 - 10^{11} plaque-forming units per ml, and they are highly infective. The life cycle of adenovirus does not require integration into the host cell genome. The foreign genes delivered by adenovirus vectors are episomal and, therefore, have low genotoxicity to host cells. No side effects have been reported in studies of

35 vaccination with wild-type adenovirus (Couch *et al.*, 1963; Top *et al.*, 1971), demonstrating their safety and therapeutic potential as *in vivo* gene transfer vectors.

Adenovirus vectors have been used in eukaryotic gene expression (Levrero *et al.*, 1991; Gomez-Foix *et al.*, 1992) and vaccine development (Grunhaus & Horwitz, 1992; Graham & Prevec, 1992). Recently, animal studies suggested that recombinant adenovirus could be used for gene therapy (Stratford-Perricaudet & Perricaudet, 1991; Stratford-Perricaudet *et al.*, 1990; Rich *et al.*, 1993). Studies in administering recombinant adenovirus to different tissues include trachea instillation (Rosenfeld *et al.*, 1991; Rosenfeld *et al.*, 1992), muscle injection (Ragot *et al.*, 1993), peripheral intravenous injections (Herz & Gerard, 1993) and stereotactic inoculation into the brain (Le Gal La Salle *et al.*, 1993).

Adenovirus vectors may originate from human adenovirus. Alternatively they may originate from adenovirus of other species e.g. chimpanzee which may have the advantage that the viral vectors are not neutralised by antibodies against human adenovirus circulating in many human subjects (see e.g.: Tatsis N *et al* *Gene Therapy* 2006 13:421-429).

Adenovirus type 35, which is relatively uncommon and therefore there are low levels of pre-existing immunity to the vector itself, has been used as a delivery system in certain tuberculosis vaccines which are being developed (see for example, Radosevich *et al* *Infection and Immunity* 2007 75(8):4105-4115). Adenovirus type 35 may also be of particular value in the present invention as a delivery vector.

20

2. RETROVIRUSES

The retroviruses are a group of single-stranded RNA viruses characterised by an ability to convert their RNA to double-stranded DNA in infected cells by a process of reverse-transcription (Coffin, 1990). The resulting DNA then stably integrates into cellular chromosomes as a provirus and directs synthesis of viral proteins. The integration results in the retention of the viral gene sequences in the recipient cell and its descendants. The retroviral genome contains three genes, gag, pol, and env that code for capsid proteins, polymerase enzyme, and envelope components, respectively. A sequence found upstream from the gag gene contains a signal for packaging of the genome into virions. Two long terminal repeat (LTR) sequences are present at the 5' and 3' ends of the viral genome. These contain strong promoter and enhancer sequences and are also required for integration in the host cell genome (Coffin, 1990).

In order to construct a retroviral vector, a nucleic acid encoding one or more oligonucleotide or polynucleotide sequences of interest is inserted into the viral genome in the place of certain viral sequences to produce a virus that is replication-defective. In order to produce virions, a

packaging cell line containing the gag, pol, and env genes but without the LTR and packaging components is constructed (Mann *et al.*, 1983). When a recombinant plasmid containing a cDNA, together with the retroviral LTR and packaging sequences is introduced into this cell line (by calcium phosphate precipitation for example), the packaging sequence allows the RNA transcript of the recombinant plasmid to be packaged into viral particles, which are then secreted into the culture media (Nicolas & Rubenstein, 1988; Temin, 1986; Mann *et al.*, 1983). The media containing the recombinant retroviruses is then collected, optionally concentrated, and used for gene transfer. Retroviral vectors are able to infect a broad variety of cell types. However, integration and stable expression require the division of host cells (Paskind *et al.*, 1975).

A novel approach designed to allow specific targeting of retrovirus vectors was recently developed based on the chemical modification of a retrovirus by the chemical addition of lactose residues to the viral envelope. This modification could permit the specific infection of hepatocytes *via* sialoglycoprotein receptors.

A different approach to targeting of recombinant retroviruses was designed in which biotinylated antibodies against a retroviral envelope protein and against a specific cell receptor were used. The antibodies were coupled *via* the biotin components by using streptavidin (Roux *et al.*, 1989). Using antibodies against major histocompatibility complex class I and class II antigens, they demonstrated the infection of a variety of human cells that bore those surface antigens with an ecotropic virus *in vitro* (Roux *et al.*, 1989).

3. ADENO-ASSOCIATED VIRUSES

AAV (Ridgeway, 1988; Hermonat & Muzyczka, 1984) is a parovirus, discovered as a contamination of adenoviral stocks. It is a ubiquitous virus (antibodies are present in 85% of the US human population) that has not been linked to any disease. It is also classified as a dependovirus, because its replication is dependent on the presence of a helper virus, such as adenovirus. Five serotypes have been isolated, of which AAV-2 is the best characterised. AAV has a single-stranded linear DNA that is encapsidated into capsid proteins VP1, VP2 and VP3 to form an icosahedral virion of 20 to 24 nm in diameter (Muzyczka & McLaughlin, 1988).

The AAV DNA is approximately 4.7 kilobases long. It contains two open reading frames and is flanked by two ITRs. There are two major genes in the AAV genome: *rep* and *cap*. The *rep* gene codes for proteins responsible for viral replications, whereas *cap* codes for capsid protein VP1-3. Each ITR forms a T-shaped hairpin structure. These terminal repeats are the only

essential *cis* components of the AAV for chromosomal integration. Therefore, the AAV can be used as a vector with all viral coding sequences removed and replaced by the cassette of genes for delivery. Three viral promoters have been identified and named p5, p19, and p40, according to their map position. Transcription from p5 and p19 results in production of rep proteins, and transcription from p40 produces the capsid proteins (Hermonat & Muzyczka, 1984).

There are several factors that prompted researchers to study the possibility of using rAAV as an expression vector. One is that the requirements for delivering a gene to integrate into the host chromosome are surprisingly few. It is necessary to have the 145-bp ITRs, which are only 6% of the AAV genome. This leaves room in the vector to assemble a 4.5-kb DNA insertion. While this carrying capacity may prevent the AAV from delivering large genes, it is amply suited for delivering antisense constructs.

AAV is also a good choice of delivery vehicles due to its safety. There is a relatively complicated rescue mechanism: not only wild type adenovirus but also AAV genes are required to mobilise rAAV. Likewise, AAV is not pathogenic and not associated with any disease. The removal of viral coding sequences minimises immune reactions to viral gene expression, and therefore, rAAV does not evoke an inflammatory response.

4. OTHER VIRAL VECTORS AS EXPRESSION CONSTRUCTS

Other viral vectors may be employed as expression constructs in the present invention for the delivery of oligonucleotide or polynucleotide sequences to a host cell. Vectors derived from viruses such as vaccinia virus (Ridgeway, 1988; Coupar *et al.*, 1988), lentiviruses, polio viruses and herpes viruses may be employed. Other poxvirus derived vectors, such as fowl-pox derived vectors, may also be expected to be of use. They offer several attractive features for various mammalian cells (Friedmann, 1989; Ridgeway, 1988; Coupar *et al.*, 1988; Horwich *et al.*, 1990).

With the recent recognition of defective hepatitis B viruses, new insight was gained into the structure-function relationship of different viral sequences. *In vitro* studies showed that the virus could retain the ability for helper-dependent packaging and reverse transcription despite the deletion of up to 80% of its genome (Horwich *et al.*, 1990). This suggested that large portions of the genome could be replaced with foreign genetic material. The hepatotropism and persistence (integration) were particularly attractive properties for liver-directed gene transfer. Chang *et al.* (1991) introduced the chloramphenicol acetyltransferase (CAT) gene

into duck hepatitis B virus genome in the place of the polymerase, surface, and pre-surface coding sequences. It was cotransfected with wild-type virus into an avian hepatoma cell line. Culture media containing high titres of the recombinant virus were used to infect primary duckling hepatocytes. Stable CAT gene expression was detected for at least 24 days after
5 transfection (Chang *et al.*, 1991).

Additional 'viral' vectors include virus like particles (VLPs) and phages.

5. NON-VIRAL VECTORS

10

In order to effect expression of the oligonucleotide or polynucleotide sequences of the present invention, the expression construct must be delivered into a cell. This delivery may be accomplished *in vitro*, as in laboratory procedures for transforming cells lines, or *in vivo* or *ex vivo*, as in the treatment of certain disease states. As described above, one preferred
15 mechanism for delivery is *via* viral infection where the expression construct is encapsulated in an infectious viral particle.

20

Once the expression construct has been delivered into the cell the nucleic acid encoding the desired oligonucleotide or polynucleotide sequences may be positioned and expressed at different sites. In certain embodiments, the nucleic acid encoding the construct may be stably integrated into the genome of the cell. This integration may be in the specific location and orientation *via* homologous recombination (gene replacement) or it may be integrated in a random, non-specific location (gene augmentation). In yet further embodiments, the nucleic acid may be stably maintained in the cell as a separate, episomal segment of DNA. Such
25 nucleic acid segments or "episomes" encode sequences sufficient to permit maintenance and replication independent of or in synchronisation with the host cell cycle. How the expression construct is delivered to a cell and where in the cell the nucleic acid remains is dependent on the type of expression construct employed.

30

In certain embodiments of the invention, the expression construct comprising one or more oligonucleotide or polynucleotide sequences may simply consist of naked recombinant DNA or plasmids. Transfer of the construct may be performed, for example, by any method which physically or chemically permeabilises the cell membrane. This is particularly applicable for transfer *in vitro* but it may be applied to *in vivo* use as well. Dubensky *et al.* (1984)
35 successfully injected polyomavirus DNA in the form of calcium phosphate precipitates into liver and spleen of adult and newborn mice demonstrating active viral replication and acute infection. Benvenisty & Reshef (1986) also demonstrated that direct intraperitoneal injection of

calcium phosphate-precipitated plasmids results in expression of the transfected genes. It is envisioned that DNA encoding a gene of interest may also be transferred in a similar manner *in vivo* and express the gene product.

- 5 Another embodiment of the invention for transferring a naked DNA expression construct into cells may involve particle bombardment. This method depends on the ability to accelerate DNA-coated microprojectiles to a high velocity allowing them to pierce cell membranes and enter cells without killing them (Klein *et al.*, 1987). Several devices for accelerating small particles have been developed. One such device relies on a high voltage discharge to
10 generate an electrical current, which in turn provides the motive force (Yang *et al.*, 1990). The microprojectiles used have consisted of biologically inert substances such as tungsten or gold beads.

- Selected organs including the liver, skin, and muscle tissue of rats and mice have been
15 bombarded *in vivo* (Yang *et al.*, 1990; Zelenin *et al.*, 1991). This may require surgical exposure of the tissue or cells, to eliminate any intervening tissue between the gun and the target organ, i.e., *ex vivo* treatment. Again, DNA encoding a particular gene may be delivered *via* this method and still be incorporated.

- 20 Bacteria may also be utilised as a delivery method (e.g. listeria, see WO2004/11048) and in particular BCG.

POLYPEPTIDE COMPOSITIONS

- 25 The present invention, in other aspects, provides polypeptide compositions.

Generally, a polypeptide of the invention will be an isolated polypeptide (i.e. separated from those components with which it may usually be found in nature).

- 30 For example, a naturally-occurring protein is isolated if it is separated from some or all of the coexisting materials in the natural system. Preferably, such polypeptides are at least about 90% pure, more preferably at least about 95% pure and most preferably at least about 99% pure. A polynucleotide is considered to be isolated if, for example, it is cloned into a vector that is not a part of the natural environment.

35

Polypeptides may be prepared using any of a variety of well known techniques. Recombinant polypeptides encoded by DNA sequences as described above may be readily prepared from

the DNA sequences using any of a variety of expression vectors known to those of ordinary skill in the art. Expression may be achieved in any appropriate host cell that has been transformed or transfected with an expression vector containing a DNA molecule that encodes a recombinant polypeptide. Suitable host cells include prokaryotes, yeast, and higher
5 eukaryotic cells, such as mammalian cells and plant cells. Preferably, the host cells employed are *E. coli*, yeast or a mammalian cell line such as COS or CHO. Supernatants from suitable host/vector systems which secrete recombinant protein or polypeptide into culture media may be first concentrated using a commercially available filter. Following concentration, the concentrate may be applied to a suitable purification matrix such as an affinity matrix or an ion
10 exchange resin. Finally, one or more reverse phase HPLC steps can be employed to further purify a recombinant polypeptide.

Polypeptides of the invention, immunogenic fragments thereof, and other variants having less than about 100 amino acids, and generally less than about 50 amino acids, may also be
15 generated by synthetic means, using techniques well known to those of ordinary skill in the art. For example, such polypeptides may be synthesised using any of the commercially available solid-phase techniques, such as the Merrifield solid-phase synthesis method, where amino acids are sequentially added to a growing amino acid chain. See Merrifield, *J. Am. Chem. Soc.* 85:2149-2146 (1963). Equipment for automated synthesis of polypeptides is
20 commercially available from suppliers such as Perkin Elmer/Applied BioSystems Division (Foster City, CA), and may be operated according to the manufacturer's instructions.

Within certain specific embodiments, a polypeptide may be a fusion protein that comprises multiple polypeptides as described herein, or that comprises at least one polypeptide as
25 described herein and an unrelated sequence, examples of such proteins include tetanus, tuberculosis and hepatitis proteins (see, e.g., Stoute *et al.*, *New Engl. J. Med.* 336:86-91 (1997)). A fusion partner may, for example, assist in providing T helper epitopes (an immunological fusion partner), preferably T helper epitopes recognised by humans, or may assist in expressing the protein (an expression enhancer) at higher yields than the native
30 recombinant protein. Certain preferred fusion partners are both immunological and expression enhancing fusion partners. Other fusion partners may be selected so as to increase the solubility of the protein or to enable the protein to be targeted to desired intracellular compartments. Still further fusion partners include affinity tags, which facilitate purification of the protein.

35 Fusion proteins may generally be prepared using standard techniques, including chemical conjugation. Preferably, a fusion protein is expressed as a recombinant protein, allowing the

production of increased levels, relative to a non-fused protein, in an expression system. Briefly, DNA sequences encoding the polypeptide components may be assembled separately, and ligated into an appropriate expression vector. The 3' end of the DNA sequence encoding one polypeptide component is ligated, with or without a peptide linker, to the 5' end of a DNA
5 sequence encoding the second polypeptide component so that the reading frames of the sequences are in phase. This permits translation into a single fusion protein that retains the biological activity of both component polypeptides.

A peptide linker sequence may be employed to separate the first and second polypeptide
10 components by a distance sufficient to ensure that each polypeptide folds into its secondary and tertiary structures. Such a peptide linker sequence is incorporated into the fusion protein using standard techniques well known in the art. Suitable peptide linker sequences may be chosen based on the following factors: (1) their ability to adopt a flexible extended
15 conformation; (2) their inability to adopt a secondary structure that could interact with functional epitopes on the first and second polypeptides; and (3) the lack of hydrophobic or charged residues that might react with the polypeptide functional epitopes. Preferred peptide linker sequences contain Gly, Asn and Ser residues. Other near neutral amino acids, such as Thr and Ala may also be used in the linker sequence. Amino acid sequences which may be
20 usefully employed as linkers include those disclosed in Maratea *et al.*, *Gene* 40:39-46 (1985); Murphy *et al.*, *Proc. Natl. Acad. Sci. USA* 83:8258-8262 (1986); U.S. Patent No. 4,935,233 and U.S. Patent No. 4,751,180. The linker sequence may generally be from 1 to about 50 amino acids in length. Linker sequences are not required when the first and second polypeptides have non-essential N-terminal amino acid regions that can be used to separate the functional domains and prevent steric interference.

25 Within preferred embodiments, an immunological fusion partner is derived from protein D, a surface protein of the gram-negative bacterium *Haemophilus influenza* B (WO 91/18926). Preferably, a protein D derivative comprises approximately the first third of the protein (e.g., the first N-terminal 100-110 amino acids), and a protein D derivative may be lipidated. Within
30 certain preferred embodiments, the first 109 residues of a lipoprotein D fusion partner is included on the N-terminus to provide the polypeptide with additional exogenous T-cell epitopes and to increase the expression level in *E. coli* (thus functioning as an expression enhancer). The lipid tail ensures optimal presentation of the antigen to antigen presenting cells. Other fusion partners include the non-structural protein from influenzae virus, NS1
35 (hemagglutinin). Typically, the N-terminal 81 amino acids are used, although different fragments that include T-helper epitopes may be used.

In another embodiment, the immunological fusion partner is the protein known as LYTA, or a portion thereof (preferably a C-terminal portion). LYTA is derived from *Streptococcus pneumoniae*, which synthesizes an N-acetyl-L-alanine amidase known as amidase LYTA (encoded by the *LytA* gene; *Gene* 43:265-292 (1986)). LYTA is an autolysin that specifically
5 degrades certain bonds in the peptidoglycan backbone. The C-terminal domain of the LYTA protein is responsible for the affinity to the choline or to some choline analogues such as DEAE. This property has been exploited for the development of *E. coli* C-LYTA expressing plasmids useful for expression of fusion proteins. Purification of hybrid proteins containing the C-LYTA fragment at the amino terminus has been described (see *Biotechnology* 10:795-798
10 (1992)). Within a preferred embodiment, a repeat portion of LYTA may be incorporated into a fusion protein. A repeat portion is found in the C-terminal region starting at residue 178. A particularly preferred repeat portion incorporates residues 188-305.

T CELLS

15 Immunotherapeutic compositions may also, or alternatively, comprise T cells specific for a *Mycobacterium* antigen. Such cells may generally be prepared *in vitro* or *ex vivo*, using standard procedures. For example, T cells may be isolated from bone marrow, peripheral blood, or a fraction of bone marrow or peripheral blood of a patient, using a commercially
20 available cell separation system, such as the Isolex™ System, available from Nexell Therapeutics, Inc. (Irvine, CA; see also U.S. Patent No. 5,240,856; U.S. Patent No. 5,215,926; WO 89/06280; WO 91/16116 and WO 92/07243). Alternatively, T cells may be derived from related or unrelated humans, non-human mammals, cell lines or cultures.

25 T cells may be stimulated with a polypeptide of the invention, polynucleotide encoding such a polypeptide, and/or an antigen presenting cell (APC) that expresses such a polypeptide. Such stimulation is performed under conditions and for a time sufficient to permit the generation of T cells that are specific for the polypeptide. Preferably, the polypeptide or polynucleotide is present within a delivery vehicle, such as a microsphere, to facilitate the generation of specific
30 T cells.

T cells are considered to be specific for a polypeptide of the invention if the T cells specifically proliferate, secrete cytokines or kill target cells coated with the polypeptide or expressing a gene encoding the polypeptide. T cell specificity may be evaluated using any of a variety of
35 standard techniques. For example, within a chromium release assay or proliferation assay, a stimulation index of more than two fold increase in lysis and/or proliferation, compared to negative controls, indicates T cell specificity. Such assays may be performed, for example, as

described in Chen *et al.*, *Cancer Res.* 54:1065-1070 (1994)). Alternatively, detection of the proliferation of T cells may be accomplished by a variety of known techniques. For example, T cell proliferation can be detected by measuring an increased rate of DNA synthesis (e.g., by pulse-labelling cultures of T cells with tritiated thymidine and measuring the amount of tritiated
5 thymidine incorporated into DNA). Contact with a polypeptide of the invention (100 ng/ml - 100 µg/ml, preferably 200 ng/ml - 25 µg/ml) for 3 - 7 days should result in at least a two fold increase in proliferation of the T cells. Contact as described above for 2-3 hours should result in activation of the T cells, as measured using standard cytokine assays in which a two fold increase in the level of cytokine release (e.g., TNF or IFN-γ) is indicative of T cell activation
10 (see Coligan *et al.*, *Current Protocols in Immunology*, vol. 1 (1998)). T cells that have been activated in response to a polypeptide, polynucleotide or polypeptide-expressing APC may be CD4⁺ and/or CD8⁺. Protein-specific T cells may be expanded using standard techniques. Within preferred embodiments, the T cells are derived from a patient, a related donor or an unrelated donor, and are administered to the patient following stimulation and expansion.

15

For therapeutic purposes, CD4⁺ or CD8⁺ T cells that proliferate in response to a polypeptide, polynucleotide or APC can be expanded in number either *in vitro* or *in vivo*. Proliferation of such T cells *in vitro* may be accomplished in a variety of ways. For example, the T cells can be re-exposed to a polypeptide, or a short peptide corresponding to an immunogenic portion of
20 such a polypeptide, with or without the addition of T cell growth factors, such as interleukin-2, and/or stimulator cells that synthesise a polypeptide. Alternatively, one or more T cells that proliferate in the presence of the protein can be expanded in number by cloning. Methods for cloning cells are well known in the art, and include limiting dilution.

25 PHARMACEUTICAL COMPOSITIONS

In additional embodiments, the polynucleotide, polypeptide, T-cell and/or antibody compositions disclosed herein will be formulated in pharmaceutically-acceptable or physiologically-acceptable solutions for administration to a cell or an animal, either alone, or in
30 combination with one or more other modalities of therapy.

It will also be understood that, if desired, the nucleic acid segment (e.g., RNA or DNA) that expresses a polypeptide as disclosed herein may be administered in combination with other agents as well, such as, e.g., other proteins or polypeptides or various pharmaceutically-active
35 agents, including chemotherapeutic agents effective against a *M. tuberculosis* infection. In fact, there is virtually no limit to other components that may also be included, given that the

additional agents do not cause a significant adverse effect upon contact with the target cells or host tissues. The compositions may thus be delivered along with various other agents as required in the particular instance. Such compositions may be purified from host cells or other biological sources, or alternatively may be chemically synthesised as described herein.

- 5 Likewise, such compositions may further comprise substituted or derivatised RNA or DNA compositions.

Formulation of pharmaceutically-acceptable excipients and carrier solutions is well-known to those of skill in the art, as is the development of suitable dosing and treatment regimens for
10 using the particular compositions described herein in a variety of treatment regimens, including e.g., oral, parenteral, intravenous, intranasal, and intramuscular administration and formulation. Other routes of administration include via the mucosal surfaces.

Typically, formulations comprising a therapeutically effective amount deliver about 0.1 ug to about 1000 ug of polypeptide per administration, more typically about 2.5 ug to about 100 ug of
15 polypeptide per administration. In respect of polynucleotide compositions, these typically deliver about 10 ug to about 20 mg of the inventive polynucleotide per administration, more typically about 0.1 mg to about 10 mg of the inventive polynucleotide per administration.

Naturally, the amount of active compound(s) in each therapeutically useful composition may be
20 prepared in such a way that a suitable dosage will be obtained in any given unit dose of the compound. Factors such as solubility, bioavailability, biological half-life, route of administration, product shelf life, as well as other pharmacological considerations will be contemplated by one skilled in the art of preparing such pharmaceutical formulations, and as such, a variety of dosages and treatment regimens may be desirable.

25

1. ORAL DELIVERY

In certain applications, the pharmaceutical compositions disclosed herein may be delivered via oral administration to an animal. As such, these compositions may be formulated with an inert
30 diluent or with an assimilable edible carrier, or they may be enclosed in hard- or soft-shell gelatin capsule, or they may be compressed into tablets, or they may be incorporated directly with the food of the diet.

The active compounds may even be incorporated with excipients and used in the form of
35 ingestible tablets, buccal tables, troches, capsules, elixirs, suspensions, syrups, wafers, and the like (Mathiowitz *et al.*, 1997; Hwang *et al.*, 1998; U. S. Patent 5,641,515; U. S. Patent

5,580,579 and U.S. Patent 5,792,451). The tablets, troches, pills, capsules and the like may also contain the following: a binder, as gum tragacanth, acacia, cornstarch, or gelatin; excipients, such as dicalcium phosphate; a disintegrating agent, such as corn starch, potato starch, alginic acid and the like; a lubricant, such as magnesium stearate; and a sweetening agent, such as sucrose, lactose or saccharin may be added or a flavouring agent, such as peppermint, oil of wintergreen, or cherry flavouring. When the dosage unit form is a capsule, it may contain, in addition to materials of the above type, a liquid carrier. Various other materials may be present as coatings or to otherwise modify the physical form of the dosage unit. For instance, tablets, pills, or capsules may be coated with shellac, sugar, or both. A syrup or elixir may contain the active component, sucrose as a sweetening agent methyl and propylparabens as preservatives, a dye and flavouring, such as cherry or orange flavour. Of course, any material used in preparing any dosage unit form should be pharmaceutically pure and substantially non-toxic in the amounts employed. In addition, the active components may be incorporated into sustained-release preparation and formulations.

For oral administration the compositions of the present invention may alternatively be incorporated with one or more excipients in the form of a mouthwash, dentifrice, buccal tablet, oral spray, or sublingual orally-administered formulation. For example, a mouthwash may be prepared incorporating the active ingredient in the required amount in an appropriate solvent, such as a sodium borate solution (Dobell's Solution). Alternatively, the active ingredient may be incorporated into an oral solution such as one containing sodium borate, glycerin and potassium bicarbonate, or dispersed in a dentifrice, or added in a therapeutically-effective amount to a composition that may include water, binders, abrasives, flavoring agents, foaming agents, and humectants. Alternatively the compositions may be fashioned into a tablet or solution form that may be placed under the tongue or otherwise dissolved in the mouth.

2. INJECTABLE DELIVERY

In certain circumstances it will be desirable to deliver the pharmaceutical compositions disclosed herein parenterally, intravenously, intramuscularly, intradermally, or even intraperitoneally as described in U.S. Patent 5,543,158; U.S. Patent 5,641,515 and U.S. Patent 5,399,363. Solutions of the active compounds as free base or pharmacologically acceptable salts may be prepared in water suitably mixed with a surfactant, such as hydroxypropylcellulose. Dispersions may also be prepared in glycerol, liquid polyethylene glycols, and mixtures thereof and in oils.

Under ordinary conditions of storage and use, these preparations contain a preservative to prevent the growth of microorganisms.

5 The pharmaceutical forms suitable for injectable use include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions (U.S. Patent 5,466,468). In all cases the form must be sterile and must be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms, such as bacteria and fungi. The carrier can be a solvent or
10 dispersion medium containing, for example, water, ethanol, polyol (e.g., glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and/or vegetable oils. Proper fluidity may be maintained, for example, by the use of a coating, such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. The prevention of the action of microorganisms can be facilitated by
15 various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

20

For parenteral administration in an aqueous solution, for example, the solution should be suitably buffered if necessary and the liquid diluent first rendered isotonic with sufficient saline or glucose. These particular aqueous solutions are especially suitable for intravenous, intramuscular, subcutaneous and intraperitoneal administration. In this connection, a sterile
25 aqueous medium that can be employed will be known to those of skill in the art in light of the present disclosure. For example, one dosage may be dissolved in 1 ml of isotonic NaCl solution and either added to 1000 ml of hypodermoclysis fluid or injected at the proposed site of infusion (see, e.g., *Remington's Pharmaceutical Sciences*, 15th Edition, pp. 1035-1038 and 1570-1580). Some variation in dosage will necessarily occur depending on the condition of the
30 subject being treated. The person responsible for administration will, in any event, determine the appropriate dose for the individual subject. Moreover, for human administration, preparations should meet sterility, pyrogenicity, and the general safety and purity standards as required by FDA Office of Biologics standards.

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Sterile injectable solutions are prepared by incorporating the active compounds in the required amount in the appropriate solvent with various of the other ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating

the various sterilized active ingredients into a sterile vehicle which contains the basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum-drying and freeze-drying techniques which yield a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

The compositions disclosed herein may be formulated in a neutral or salt form. Pharmaceutically-acceptable salts, include the acid addition salts (formed with the free amino groups of the protein) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups can also be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, histidine, procaine and the like. Upon formulation, solutions will be administered in a manner compatible with the dosage formulation and in such amount as is therapeutically effective. The formulations are easily administered in a variety of dosage forms such as injectable solutions, drug-release capsules, and the like.

As used herein, "carrier" includes any and all solvents, dispersion media, vehicles, coatings, diluents, antibacterial and antifungal agents, isotonic and absorption delaying agents, buffers, carrier solutions, suspensions, colloids, and the like. The use of such media and agents for pharmaceutical active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active ingredient, its use in the therapeutic compositions is contemplated. Supplementary active ingredients can also be incorporated into the compositions.

The phrase "pharmaceutically-acceptable" refers to molecular entities and compositions that do not produce an allergic or similar untoward reaction when administered to a human. The preparation of an aqueous composition that contains a protein as an active ingredient is well understood in the art. Typically, such compositions are prepared as injectables, either as liquid solutions or suspensions; solid forms suitable for solution in, or suspension in, liquid prior to injection can also be prepared. The preparation can also be emulsified.

3. NASAL AND BUCCAL DELIVERY

In certain embodiments, the pharmaceutical compositions may be delivered by intranasal sprays, buccal sprays, inhalation, and/or other aerosol delivery vehicles. Methods for

delivering genes, nucleic acids, and peptide compositions directly to the lungs e.g. *via* nasal and buccal aerosol sprays has been described e.g., in U.S. Patent 5,756,353 and U.S. Patent 5,804,212. Likewise, the delivery of drugs using intranasal microparticle resins (Takenaga *et al.*, 1998) and lysophosphatidyl-glycerol compounds (U.S. Patent 5,725,871) are also well-known in the pharmaceutical arts. Likewise, transmucosal drug delivery in the form of a polytetrafluoroethylene support matrix is described in U.S. Patent 5,780,045.

10

4. LIPOSOME-, NANOCAPSULE-, AND MICROPARTICLE-MEDIATED DELIVERY

In certain embodiments, the inventors contemplate the use of liposomes, nanocapsules, microparticles, microspheres, lipid particles, vesicles, and the like, for the introduction of the compositions of the present invention into suitable host cells. In particular, the compositions of the present invention may be formulated for delivery either encapsulated in a lipid particle, a liposome, a vesicle, a nanosphere, or a nanoparticle or the like.

Such formulations may be preferred for the introduction of pharmaceutically-acceptable formulations of the nucleic acids or constructs disclosed herein. The formation and use of liposomes is generally known to those of skill in the art (see for example, Couvreur *et al.*, 1977; Couvreur, 1988; Lasic, 1998; which describes the use of liposomes and nanocapsules in the targeted antibiotic therapy for intracellular bacterial infections and diseases). Recently, liposomes were developed with improved serum stability and circulation half-times (Gabizon & Papahadjopoulos, 1988; Allen and Choun, 1987; U.S. Patent 5,741,516). Further, various methods of liposome and liposome like preparations as potential drug carriers have been reviewed (Takakura, 1998; Chandran *et al.*, 1997; Margalit, 1995; U.S. Patent 5,567,434; U.S. Patent 5,552,157; U.S. Patent 5,565,213; U.S. Patent 5,738,868 and U.S. Patent 5,795,587).

30

Liposomes have been used successfully with a number of cell types that are normally resistant to transfection by other procedures including T cell suspensions, primary hepatocyte cultures and PC 12 cells (Renneisen *et al.*, 1990; Muller *et al.*, 1990). In addition, liposomes are free of the DNA length constraints that are typical of viral-based delivery systems. Liposomes have been used effectively to introduce genes, drugs (Heath & Martin, 1986; Heath *et al.*, 1986; Balazsovits *et al.*, 1989; Fresta & Puglisi, 1996), radiotherapeutic agents (Pikul *et al.*, 1987),

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enzymes (Imaizumi *et al.*, 1990a; Imaizumi *et al.*, 1990b), viruses (Faller & Baltimore, 1984), transcription factors and allosteric effectors (Nicolau & Gersonde, 1979) into a variety of cultured cell lines and animals. In addition, several successful clinical trials examining the effectiveness of liposome-mediated drug delivery have been completed (Lopez-Berestein *et al.*, 1985a; 1985b; Coune, 1988; Sculier *et al.*, 1988). Furthermore, several studies suggest that the use of liposomes is not associated with autoimmune responses, toxicity or gonadal localization after systemic delivery (Mori & Fukatsu, 1992).

Liposomes are formed from phospholipids that are dispersed in an aqueous medium and spontaneously form multilamellar concentric bilayer vesicles (also termed multilamellar vesicles (MLVs). MLVs generally have diameters of from 25 nm to 4 μ m. Sonication of MLVs results in the formation of small unilamellar vesicles (SUVs) with diameters in the range of 200 to 500 Å, containing an aqueous solution in the core.

Liposomes bear resemblance to cellular membranes and are contemplated for use in connection with the present invention as carriers for the peptide compositions. They are widely suitable as both water- and lipid-soluble substances can be entrapped, i.e. in the aqueous spaces and within the bilayer itself, respectively. It is possible that the drug-bearing liposomes may even be employed for site-specific delivery of active agents by selectively modifying the liposomal formulation.

In addition to the teachings of Couvreur *et al.* (1977; 1988), the following information may be utilized in generating liposomal formulations. Phospholipids can form a variety of structures other than liposomes when dispersed in water, depending on the molar ratio of lipid to water. At low ratios the liposome is the preferred structure. The physical characteristics of liposomes depend on pH, ionic strength and the presence of divalent cations. Liposomes can show low permeability to ionic and polar substances, but at elevated temperatures undergo a phase transition which markedly alters their permeability. The phase transition involves a change from a closely packed, ordered structure, known as the gel state, to a loosely packed, less-ordered structure, known as the fluid state. This occurs at a characteristic phase-transition temperature and results in an increase in permeability to ions, sugars and drugs.

In addition to temperature, exposure to proteins can alter the permeability of liposomes. Certain soluble proteins, such as cytochrome c, bind, deform and penetrate the bilayer, thereby causing changes in permeability. Cholesterol inhibits this penetration of proteins, apparently by packing the phospholipids more tightly. It is contemplated that the most useful liposome formations for antibiotic and inhibitor delivery will contain cholesterol.

The ability to trap solutes varies between different types of liposomes. For example, MLVs are moderately efficient at trapping solutes, but SUVs are extremely inefficient. SUVs offer the advantage of homogeneity and reproducibility in size distribution, however, and a compromise
5 between size and trapping efficiency is offered by large unilamellar vesicles (LUVs). These are prepared by ether evaporation and are three to four times more efficient at solute entrapment than MLVs.

In addition to liposome characteristics, an important determinant in entrapping compounds is
10 the physicochemical properties of the compound itself. Polar compounds are trapped in the aqueous spaces and nonpolar compounds bind to the lipid bilayer of the vesicle. Polar compounds are released through permeation or when the bilayer is broken, but nonpolar compounds remain affiliated with the bilayer unless it is disrupted by temperature or exposure to lipoproteins. Both types show maximum efflux rates at the phase transition temperature.

15 Liposomes interact with cells *via* four different mechanisms: endocytosis by phagocytic cells of the reticuloendothelial system such as macrophages and neutrophils; adsorption to the cell surface, either by nonspecific weak hydrophobic or electrostatic forces, or by specific interactions with cell-surface components; fusion with the plasma cell membrane by insertion
20 of the lipid bilayer of the liposome into the plasma membrane, with simultaneous release of liposomal contents into the cytoplasm; and by transfer of liposomal lipids to cellular or subcellular membranes, or vice versa, without any association of the liposome contents. It often is difficult to determine which mechanism is operative and more than one may operate at the same time.

25 The fate and disposition of intravenously injected liposomes depend on their physical properties, such as size, fluidity, and surface charge. They may persist in tissues for h or days, depending on their composition, and half lives in the blood range from min to several h. Larger liposomes, such as MLVs and LUVs, are taken up rapidly by phagocytic cells of the
30 reticuloendothelial system, but physiology of the circulatory system restrains the exit of such large species at most sites. They can exit only in places where large openings or pores exist in the capillary endothelium, such as the sinusoids of the liver or spleen. Thus, these organs are the predominate site of uptake. On the other hand, SUVs show a broader tissue distribution but still are sequestered highly in the liver and spleen. In general, this *in vivo*
35 behaviour limits the potential targeting of liposomes to only those organs and tissues accessible to their large size. These include the blood, liver, spleen, bone marrow, and lymphoid organs.

Targeting is generally not a limitation in terms of the present invention. However, should specific targeting be desired, methods are available for this to be accomplished. Antibodies may be used to bind to the liposome surface and to direct the antibody and its drug contents to specific antigenic receptors located on a particular cell-type surface. Carbohydrate determinants (glycoprotein or glycolipid cell-surface components that play a role in cell-cell recognition, interaction and adhesion) may also be used as recognition sites as they have potential in directing liposomes to particular cell types. Mostly, it is contemplated that intravenous injection of liposomal preparations would be used, but other routes of administration are also conceivable.

Alternatively, the invention provides for pharmaceutically-acceptable nanocapsule formulations of the compositions of the present invention. Nanocapsules can generally entrap compounds in a stable and reproducible way (Henry-Michelland *et al.*, 1987; Quintanar-Guerrero *et al.*, 1998; Douglas *et al.*, 1987). To avoid side effects due to intracellular polymeric overloading, such ultrafine particles (sized around 0.1 μm) should be designed using polymers able to be degraded *in vivo*. Biodegradable polyalkyl-cyanoacrylate nanoparticles that meet these requirements are contemplated for use in the present invention. Such particles may be easily made, as described (Couvreur *et al.*, 1980; 1988; zur Muhlen *et al.*, 1998; Zambaux *et al.* 1998; Pinto-Alphandry *et al.*, 1995 and U. S. Patent 5,145,684).

Skin patches may also be utilised for transcutaneous delivery.

25 IMMUNOGENIC COMPOSITIONS

In certain preferred embodiments of the present invention, immunogenic compositions are provided. The immunogenic compositions will generally comprise one or more polypeptides or polynucleotides, such as those discussed above, in combination with an immunostimulant. An immunostimulant may be any substance that enhances or potentiates an immune response (antibody and/or cell-mediated) to an exogenous antigen. Examples of immunostimulants include adjuvants, biodegradable microspheres (e.g., polylactic galactide) and liposomes (into which the compound is incorporated; see, e.g., Fullerton, U.S. Patent No. 4,235,877).

Preparation of immunogenic compositions is generally described in, for example, Powell & Newman, eds., *Vaccine Design* (the subunit and adjuvant approach) (1995). Pharmaceutical compositions and immunogenic compositions within the scope of the present invention may

also contain other compounds, which may be biologically active or inactive. For example, one or more immunogenic portions of other *M. tuberculosis* antigens may be present, either incorporated into a fusion polypeptide or as a separate compound, within the pharmaceutical or immunogenic composition.

5 Illustrative immunogenic compositions may contain a polynucleotide (e.g. DNA) encoding one or more of the polypeptides as described above, such that the polypeptide is generated *in situ* (thereby eliciting an immune response). As noted above, the DNA may be present within any of a variety of delivery systems known to those of ordinary skill in the art, including nucleic acid
10 expression systems, bacteria and viral expression systems. Numerous gene delivery techniques are well known in the art, such as those described by Rolland, *Crit. Rev. Therap. Drug Carrier Systems* 15:143-198 (1998), and references cited therein. Appropriate nucleic acid expression systems contain the necessary DNA sequences for expression in the patient (such as a suitable promoter and terminating signal). Bacterial delivery systems involve the
15 administration of a bacterium host cell (*for example*, a *Mycobacterium*, *Bacillus* or *Lactobacillus* strain, including *Bacillus-Calmette-Guerrin* or *Lactococcus lactis*) that expresses the polypeptide (e.g. on its cell surface or secretes the polypeptide) (*see, for example*, Ferreira, *et al.*, *An Acad Bras Cienc* (2005) 77:113-124; and Raha, *et al.*, *Appl Microbiol Biotechnol* (2005) PubMedID 15635459). In a preferred embodiment, the DNA may be introduced using a viral
20 expression system (e.g., vaccinia or other pox virus, retrovirus, or adenovirus), which may involve the use of a non-pathogenic (defective), replication competent virus. Suitable systems are disclosed, for example, in Fisher-Hoch *et al.*, *Proc. Natl. Acad. Sci. USA* 86:317-321 (1989); Flexner *et al.*, *Ann. N.Y. Acad. Sci.* 569:86-103 (1989); Flexner *et al.*, *Vaccine* 8:17-21 (1990); U.S. Patent Nos. 4,603,112, 4,769,330, and 5,017,487; WO 89/01973; U.S. Patent No.
25 4,777,127; GB 2,200,651; EP 0,345,242; WO 91/02805; Berkner, *Biotechniques* 6:616-627 (1988); Rosenfeld *et al.*, *Science* 252:431-434 (1991); Kolls *et al.*, *Proc. Natl. Acad. Sci. USA* 91:215-219 (1994); Kass-Eisler *et al.*, *Proc. Natl. Acad. Sci. USA* 90:11498-11502 (1993); Guzman *et al.*, *Circulation* 88:2838-2848 (1993); and Guzman *et al.*, *Cir. Res.* 73:1202-1207 (1993). Techniques for incorporating DNA into such expression systems are well known to
30 those of ordinary skill in the art. The DNA may also be "naked," as described, for example, in Ulmer *et al.*, *Science* 259:1745-1749 (1993) and reviewed by Cohen, *Science* 259:1691-1692 (1993). The uptake of naked DNA may be increased by coating the DNA onto biodegradable beads, which are efficiently transported into the cells. It will be apparent that a immunogenic composition may comprise both a polynucleotide and a polypeptide component. Such
35 immunogenic composition may provide for an enhanced immune response.

It will be apparent that an immunogenic composition may contain pharmaceutically acceptable salts of the polynucleotides and polypeptides provided herein. Such salts may be prepared from pharmaceutically acceptable non-toxic bases, including organic bases (e.g., salts of primary, secondary and tertiary amines and basic amino acids) and inorganic bases (e.g.,
5 sodium, potassium, lithium, ammonium, calcium and magnesium salts).

While any suitable carrier known to those of ordinary skill in the art may be employed in the immunogenic compositions of this invention, the type of carrier will vary depending on the mode of administration. Compositions of the present invention may be formulated for any
10 appropriate manner of administration, including for example, topical, oral, nasal, intravenous, intracranial, intraperitoneal, subcutaneous or intramuscular administration. For parenteral administration, such as subcutaneous injection, the carrier preferably comprises water, saline, alcohol, a fat, a wax or a buffer. For oral administration, any of the above carriers or a solid carrier, such as mannitol, lactose, starch, magnesium stearate, sodium saccharine, talcum,
15 cellulose, glucose, sucrose, and magnesium carbonate, may be employed. Biodegradable microspheres (e.g., polylactate polyglycolate) may also be employed as carriers for the pharmaceutical compositions of this invention. Suitable biodegradable microspheres are disclosed, for example, in U.S. Patent Nos. 4,897,268; 5,075,109; 5,928,647; 5,811,128; 5,820,883; 5,853,763; 5,814,344 and 5,942,252. One may also employ a carrier comprising
20 the particulate-protein complexes described in U.S. Patent No. 5,928,647, which are capable of inducing a class I-restricted cytotoxic T lymphocyte responses in a host.

Such compositions may also comprise buffers (e.g., neutral buffered saline or phosphate buffered saline), carbohydrates (e.g., glucose, mannose, sucrose or dextrans), mannitol,
25 proteins, polypeptides or amino acids such as glycine, antioxidants, bacteriostats, chelating agents such as EDTA or glutathione, adjuvants (e.g., aluminum hydroxide), solutes that render the formulation isotonic, hypotonic or weakly hypertonic with the blood of a recipient, suspending agents, thickening agents and/or preservatives. Alternatively, compositions of the present invention may be formulated as a lyophilizate. Compounds may also be encapsulated
30 within liposomes using well known technology.

Any of a variety of immunostimulants may be employed in the immunogenic compositions of this invention. For example, an adjuvant may be included. Most adjuvants contain a substance designed to protect the antigen from rapid catabolism, such as aluminium hydroxide
35 or mineral oil, and a stimulator of immune responses, such as lipid A, *Bordetella pertussis* or *Mycobacterium* species or *Mycobacterium* derived proteins. For example, delipidated, deglycolipidated *M. vaccae* ("pVac") can be used. Suitable adjuvants are commercially

available as, for example, Freund's Incomplete Adjuvant and Complete Adjuvant (Difco Laboratories, Detroit, MI); Merck Adjuvant 65 (Merck and Company, Inc., Rahway, NJ); AS01B, AS02A, AS15, AS-2 and derivatives thereof (GlaxoSmithKline, Philadelphia, PA); CWS (cell wall skeleton from a tubercule bacillus), TDM (trehalose dicorynomycolate), Leif
 5 (Leishmania elongation initiation factor), aluminium salts such as aluminium hydroxide gel (alum) or aluminum phosphate; salts of calcium, iron or zinc; an insoluble suspension of acylated tyrosine; acylated sugars; cationically or anionically derivatized polysaccharides; polyphosphazenes; biodegradable microspheres; monophosphoryl lipid A (MPL®); and quil A (e.g. QS-21). Cytokines, such as GM-CSF or interleukin-2, -7, or -12, may also be used as
 10 adjuvants.

An adjuvant refers to the components in a vaccine or therapeutic composition that increase the specific immune response to the antigen (see, e.g., Edelman, *AIDS Res. Hum Retroviruses* 8:1409-1411 (1992)). Adjuvants induce immune responses of the Th1-type and Th-2 type
 15 response. Th1-type cytokines (e.g., IFN- γ , IL-2, and IL-12) tend to favour the induction of cell-mediated immune response to an administered antigen, while Th-2 type cytokines (e.g., IL-4, IL-5, IL-6, IL-10) tend to favour the induction of humoral immune responses. Adjuvants capable of preferential stimulation of a Th-1 cell-mediated immune response are described in WO 94/00153 and WO 95/17209.

20 Within the immunogenic compositions provided herein, the adjuvant composition is preferably designed to induce an immune response predominantly of the Th1 type. Following application of a immunogenic composition as provided herein, a patient will typically support an immune response that includes Th1- and Th2-type responses. Within a preferred embodiment, in
 25 which a response is predominantly Th1-type, the level of Th1-type cytokines will increase to a greater extent than the level of Th2-type cytokines. The levels of these cytokines may be readily assessed using standard assays. For a review of the families of cytokines, see Janeway, *et al.*, *Immunobiology*, 5th Edition, 2001.

30 The Rv2386c compositions usually comprise one or more adjuvants, e.g., AS01B (3-de-O-acylated monophosphoryl lipid A (3D-MPL®) and QS21 in a liposome formulation; see, U.S. Patent Publication No. 2003/0143240); AS02A (3D-MPL® and QS21 and an oil in water emulsion; see, Bojang, *et al.*, *Lancet* (2001) 358:1927); , ENHANZYN® (Detox); 3D-MPL®; saponins including Quil A and its components e.g. QS21 and saponin mimetics; CWS (cell wall
 35 skeleton from a tubercule bacillus); TDM (trehalose dicorynomycolate); aminoalkyl glucosaminide 4-phosphates (AGPs); immunostimulatory oligonucleotides e.g. CPG; Leif

(Leishmania elongation initiation factor); and derivatives thereof. In a preferred embodiment, an Rv2386c polypeptide is administered with one or more adjuvants selected from the group consisting of 3D-MPL® and QS21 in a liposome formulation eg AS01B and 3D-MPL® and QS21 and an oil in water emulsion (e.g. AS02A). Adjuvant systems AS01B and AS02A are
5 further described in Pichyangkul, *et al.*, *Vaccine* (2004) 22:3831-40.

When delivering the Rv2386c antigen as a nucleic acid, it can be delivered, for example, in a viral vector (*i.e.*, an adenovirus vector), or in a mutant bacterium host cell (*i.e.*, a mutant, avirulent *Mycobacterium*, *Lactobacillus* or *Bacillus* host cell including *Bacillus* Calmette-Guerin
10 (BCG) and *Lactococcus lactis*).

Preferred adjuvants for use in eliciting a predominantly Th1-type response include, for example, a combination of monophosphoryl lipid A (MPL ®), preferably 3-O-deacylated monophosphoryl lipid A (3D-MPL®), optionally with an aluminium salt (*see, for example*, Ribi, *et al.*, 1986, *Immunology and Immunopharmacology of Bacterial Endotoxins*, Plenum Publ. Corp., NY, pp. 407-419; GB 2122204B; GB 2220211; and US 4,912,094). A preferred form of
15 3D-MPL® is in the form of an emulsion having a small particle size less than 0.2mm in diameter, and its method of manufacture is disclosed in WO 94/21292. Aqueous formulations comprising monophosphoryl lipid A and a surfactant have been described in WO 98/43670.

20 Exemplified preferred adjuvants include AS01B (MPL® and QS21 in a liposome formulation), 3D-MPL® and QS21 in a liposome formulation, AS02A (MPL® and QS21 and an oil in water emulsion), 3D-MPL® and QS21 and an oil in water emulsion, and AS15, available from GlaxoSmithKline. MPL® adjuvants are available from GlaxoSmithKline (*see* US Patent Nos. 4,436,727; 4,877,611; 4,866,034 and 4,912,094).

25 CpG-containing oligonucleotides (in which the CpG dinucleotide is unmethylated) also induce a predominantly Th1 response. CpG is an abbreviation for cytosine-guanosine dinucleotide motifs present in DNA. Such oligonucleotides are well known and are described, for example, in WO 96/02555, WO 99/33488 and U.S. Patent Nos. 6,008,200 and 5,856,462.

30 Immunostimulatory DNA sequences are also described, for example, by Sato *et al.*, *Science* 273:352 (1996). CpG when formulated into immunogenic compositions, is generally administered in free solution together with free antigen (WO 96/02555; McCluskie and Davis, *supra*) or covalently conjugated to an antigen (WO 98/16247), or formulated with a carrier such as aluminium hydroxide ((Hepatitis surface antigen) Davis *et al.* *supra*; Brazolot-Millan *et al.*,
35 *Proc.Natl.Acad.Sci.*, USA, 1998, 95(26), 15553-8). CpG is known in the art as being an adjuvant that can be administered by both systemic and mucosal routes (WO 96/02555, EP

468520, Davis *et al.*, *J.Immunol*, 1998, 160(2):870-876; McCluskie and Davis, *J.Immunol.*, 1998, 161(9):4463-6).

Another preferred adjuvant is a saponin or saponin mimetics or derivatives, such as Quil A,
5 preferably QS21 (Aquila Biopharmaceuticals Inc., Framingham, MA), which may be used alone
or in combination with other adjuvants. For example, an enhanced system involves the
combination of a monophosphoryl lipid A (MPL®) and saponin derivative, such as the
combination of QS21 and 3D-MPL® as described in WO 94/00153, or a less reactogenic
composition where the QS21 is quenched with cholesterol, as described in WO 96/33739.
10 Other preferred formulations comprise an oil-in-water emulsion and tocopherol. A particularly
potent adjuvant formulation involving QS21, 3D-MPL® and tocopherol in an oil-in-water
emulsion is described in WO 95/17210. Additional saponin adjuvants of use in the present
invention include QS7 (described in WO 96/33739 and WO 96/11711) and QS17 (described in
U.S. Patent No. 5,057,540 and EP 0 362 279 B1).

15

Alternatively the saponin formulations may be combined with vaccine vehicles composed of
chitosan or other polycationic polymers, polylactide and polylactide-co-glycolide particles, poly-
N-acetyl glucosamine-based polymer matrix, particles composed of polysaccharides or
chemically modified polysaccharides, liposomes and lipid-based particles, particles composed
20 of glycerol monoesters, etc. The saponins may also be formulated in the presence of
cholesterol to form particulate structures such as liposomes or ISCOM®s. Furthermore, the
saponins may be formulated together with a polyoxyethylene ether or ester, in either a non-
particulate solution or suspension, or in a particulate structure such as a paucilamellar
liposome or ISCOM®. The saponins may also be formulated with excipients such as
25 CARBOPOL® to increase viscosity, or may be formulated in a dry powder form with a powder
excipient such as lactose.

In one embodiment, the adjuvant system includes the combination of a monophosphoryl lipid A
and a saponin derivative, such as the combination of QS21 and 3D-MPL® adjuvant, as
30 described in WO 94/00153, or a less reactogenic composition where the QS21 is quenched
with cholesterol containing liposomes, as described in WO 96/33739. Other suitable
formulations comprise an oil-in-water emulsion and tocopherol. Another suitable adjuvant
formulation employing QS21, 3D-MPL® adjuvant and tocopherol in an oil-in-water emulsion is
described in WO 95/17210.

35

Another enhanced adjuvant system involves the combination of a CpG-containing oligonucleotide and a saponin derivative particularly the combination of CpG and QS21 as disclosed in WO 00/09159. Suitably the formulation additionally comprises an oil in water emulsion and tocopherol.

5

Other suitable adjuvants include MONTANIDE® ISA 720 (Seppic, France), SAF (Chiron, California, United States), ISCOMS® (CSL), MF-59 (Chiron), the SBAS series of adjuvants (SmithKline Beecham, Rixensart, Belgium), Detox (Corixa), RC-529 (Corixa) and other aminoalkyl glucosaminide 4-phosphates (AGPs), such as those described in pending U.S. Patent Application Serial Nos. 08/853,826 and 09/074,720, and polyoxyethylene ether adjuvants such as those described in WO 99/52549A1. SmithKline Beecham and Corixa Corporation are now part of GlaxoSmithKline.

10

15 Other suitable adjuvants include adjuvant molecules of the general formula (I):



wherein, n is 1-50, A is a bond or $-\text{C}(\text{O})-$, R is C_{1-50} alkyl or Phenyl C_{1-50} alkyl.

20

A further adjuvant of interest is shiga toxin b chain, used for example as described in WO2005/112991.

One embodiment of the present invention consists of an immunogenic composition comprising a polyoxyethylene ether of general formula (I), wherein n is between 1 and 50, preferably 4-24, most preferably 9; the R component is C_{1-50} , preferably $\text{C}_4\text{-C}_{20}$ alkyl and most preferably C_{12} alkyl, and A is a bond. The concentration of the polyoxyethylene ethers should be in the range 0.1-20%, preferably from 0.1-10%, and most preferably in the range 0.1-1%. Preferred polyoxyethylene ethers are selected from the following group: polyoxyethylene-9-lauryl ether, polyoxyethylene-9-stearyl ether, polyoxyethylene-8-stearyl ether, polyoxyethylene-4-lauryl ether, polyoxyethylene-35-lauryl ether, and polyoxyethylene-23-lauryl ether. Polyoxyethylene ethers such as polyoxyethylene lauryl ether are described in the Merck index (12th edition: entry 7717). These adjuvant molecules are described in WO 99/52549.

25

30

Any immunogenic composition provided herein may be prepared using well known methods that result in a combination of antigen, immune response enhancer and a suitable carrier or excipient. The compositions described herein may be administered as part of a sustained release formulation (i.e., a formulation such as a capsule, sponge or gel (composed of

35

polysaccharides, for example) that effects a slow release of compound following administration). Such formulations may generally be prepared using well known technology (see, e.g., Coombes *et al.*, *Vaccine* 14:1429-1438 (1996)) and administered by, for example, oral, rectal or subcutaneous implantation, or by implantation at the desired target site.

- 5 Sustained-release formulations may contain a polypeptide, polynucleotide or antibody dispersed in a carrier matrix and/or contained within a reservoir surrounded by a rate controlling membrane.

- Carriers for use within such formulations are biocompatible, and may also be biodegradable; preferably the formulation provides a relatively constant level of active component release. Such carriers include microparticles of poly(lactide-co-glycolide), polyacrylate, latex, starch, cellulose, dextran and the like. Other delayed-release carriers include supramolecular biovectors, which comprise a non-liquid hydrophilic core (e.g., a cross-linked polysaccharide or oligosaccharide) and, optionally, an external layer comprising an amphiphilic compound, such as a phospholipid (see, e.g., U.S. Patent No. 5,151,254 and PCT applications WO 94/20078, WO/94/23701 and WO 96/06638). The amount of active compound contained within a sustained release formulation depends upon the site of implantation, the rate and expected duration of release.

- 20 Any of a variety of delivery vehicles may be employed within pharmaceutical compositions and immunogenic compositions to facilitate production of an antigen-specific immune response. Delivery vehicles include antigen presenting cells (APCs), such as dendritic cells, macrophages, B cells, monocytes and other cells that may be engineered to be efficient APCs. Such cells may, but need not, be genetically modified to increase the capacity for presenting the antigen, to improve activation and/or maintenance of the T cell response and/or to be immunologically compatible with the receiver (i.e., matched HLA haplotype). APCs may generally be isolated from any of a variety of biological fluids and organs, and may be autologous, allogeneic, syngeneic or xenogeneic cells.

- 30 Certain preferred embodiments of the present invention use dendritic cells or progenitors thereof as antigen-presenting cells. Dendritic cells are highly potent APCs (Banchereau & Steinman, *Nature* 392:245-251 (1998)) and have been shown to be effective as a physiological adjuvant for eliciting prophylactic or therapeutic immunity (see Timmerman & Levy, *Ann. Rev. Med.* 50:507-529 (1999)). In general, dendritic cells may be identified based on their typical shape (stellate *in situ*, with marked cytoplasmic processes (dendrites) visible *in vitro*), their ability to take up, process and present antigens with high efficiency and their ability to activate naïve T cell responses. Dendritic cells may, of course, be engineered to express specific cell-

surface receptors or ligands that are not commonly found on dendritic cells *in vivo* or *ex vivo*, and such modified dendritic cells are contemplated by the present invention. As an alternative to dendritic cells, secreted vesicles antigen-loaded dendritic cells (called exosomes) may be used within an immunogenic composition (see Zitvogel *et al.*, *Nature Med.* 4:594-600 (1998)).

5

Dendritic cells and progenitors may be obtained from peripheral blood, bone marrow, lymph nodes, spleen, skin, umbilical cord blood or any other suitable tissue or fluid. For example, dendritic cells may be differentiated *ex vivo* by adding a combination of cytokines such as GM-CSF, IL-4, IL-13 and/or TNF α to cultures of monocytes harvested from peripheral blood.

10

Alternatively, CD34 positive cells harvested from peripheral blood, umbilical cord blood or bone marrow may be differentiated into dendritic cells by adding to the culture medium combinations of GM-CSF, IL-3, TNF α , CD40 ligand, LPS, flt3 ligand and/or other compound(s) that induce differentiation, maturation and proliferation of dendritic cells.

15

Dendritic cells are conveniently categorised as “immature” and “mature” cells, which allows a simple way to discriminate between two well characterised phenotypes. However, this nomenclature should not be construed to exclude all possible intermediate stages of differentiation. Immature dendritic cells are characterised as APC with a high capacity for antigen uptake and processing, which correlates with the high expression of Fc γ receptor and mannose receptor. The mature phenotype is typically characterized by a lower expression of these markers, but a high expression of cell surface molecules responsible for T cell activation such as class I and class II MHC, adhesion molecules (e.g., CD54 and CD11) and costimulatory molecules (e.g., CD40, CD80, CD86 and 4-1BB).

20

25

APCs may generally be transfected with a polynucleotide encoding a protein (or portion or other variant thereof) such that the polypeptide, is expressed on the cell surface. Such transfection may take place *ex vivo*, and a pharmaceutical composition or immunogenic composition comprising such transfected cells may then be used, as described herein.

30

Alternatively, a gene delivery vehicle that targets a dendritic or other antigen presenting cell may be administered to a patient, resulting in transfection that occurs *in vivo*. *In vivo* and *ex vivo* transfection of dendritic cells, for example, may generally be performed using any methods known in the art, such as those described in WO 97/24447, or the gene gun approach described by Mahvi *et al.*, *Immunology and Cell Biology* 75:456-460 (1997). Antigen loading of dendritic cells may be achieved by incubating dendritic cells or progenitor cells with the polypeptide, DNA (naked or within a plasmid vector) or RNA; or with antigen-expressing recombinant bacterium or viruses (e.g., vaccinia, fowlpox, adenovirus or lentivirus vectors).

35

Prior to loading, the polypeptide may be covalently conjugated to an immunological partner that provides T cell help (e.g., a carrier molecule). Alternatively, a dendritic cell may be pulsed with a non-conjugated immunological partner, separately or in the presence of the polypeptide.

- 5 Immunogenic compositions and pharmaceutical compositions may be presented in unit-dose or multi-dose containers, such as sealed ampoules or vials. Such containers are preferably hermetically sealed to preserve sterility of the formulation until use. In general, formulations may be stored as suspensions, solutions or emulsions in oily or aqueous vehicles. Alternatively, an immunogenic composition or pharmaceutical composition may be stored in a
10 freeze-dried condition requiring only the addition of a sterile liquid carrier immediately prior to use.

- In some embodiments, a “priming” or first administration of an Rv2386c polypeptide (including variants, immunogenic fragments or fusion proteins), or polynucleotide encoding said
15 polypeptide, is followed by one or more “boosting” or subsequent administrations of an Rv2386c polypeptide (including variants, immunogenic fragments or fusion proteins) or polynucleotide encoding said polypeptide (“prime and boost” method). For instance, a first administration with an Rv2386c polypeptide (including variants, immunogenic fragments or fusion proteins) or polynucleotide encoding said polypeptide is followed by one or more
20 subsequent administrations of an Rv2386c polypeptide (including variants, immunogenic fragments or fusion proteins) or polynucleotide encoding said polypeptide.

- In one embodiment, a first administration with an Rv2386c polypeptide or polynucleotide is followed by one or more subsequent administrations of an Rv2386c polypeptide. In one
25 embodiment, a first administration with an Rv2386c polypeptide or polynucleotide is followed by one or more subsequent administrations of an Rv2386c polynucleotide. Usually the first or “priming” administration and the second or “boosting” administration are given about 2-12 weeks apart, or up to 4-6 months apart. Subsequent “booster” administrations are given about 6 months apart, or as long as 1, 2, 3, 4 or 5 years apart. Conventional booster treatment (e.g.,
30 a protein priming administration followed by a protein boosting administration) may also be useful in preventing or treating tuberculosis (e.g. preventing or treating latent tuberculosis, in particular preventing or delay tuberculosis reactivation).

ANTIBODIES

35

“Antibody” refers to a polypeptide comprising a framework region from an immunoglobulin gene or fragments thereof that specifically binds and recognises an antigen. The recognised

immunoglobulin genes include the kappa, lambda, alpha, gamma, delta, epsilon, and mu constant region genes, as well as the myriad immunoglobulin variable region genes. Light chains are classified as either kappa or lambda. Heavy chains are classified as gamma, mu, alpha, delta, or epsilon, which in turn define the immunoglobulin classes, IgG, IgM, IgA, IgD and IgE, respectively.

An exemplary immunoglobulin (antibody) structural unit comprises a tetramer. Each tetramer is composed of two identical pairs of polypeptide chains, each pair having one "light" (about 25 kDa) and one "heavy" chain (about 50-70 kDa). The N-terminus of each chain defines a variable region of about 100 to 110 or more amino acids primarily responsible for antigen recognition. The terms variable light chain (V_L) and variable heavy chain (V_H) refer to these light and heavy chains respectively.

Antibodies exist, e.g., as intact immunoglobulins or as a number of well-characterised fragments produced by digestion with various peptidases. Thus, for example, pepsin digests an antibody below the disulfide linkages in the hinge region to produce $F(ab)'_2$, a dimer of Fab which itself is a light chain joined to V_H-C_H1 by a disulfide bond. The $F(ab)'_2$ may be reduced under mild conditions to break the disulfide linkage in the hinge region, thereby converting the $F(ab)'_2$ dimer into an Fab' monomer. The Fab' monomer is essentially Fab with part of the hinge region (see *Fundamental Immunology* (Paul ed., 3d ed. 1993)). While various antibody fragments are defined in terms of the digestion of an intact antibody, one of skill will appreciate that such fragments may be synthesized *de novo* either chemically or by using recombinant DNA methodology. Thus, the term antibody, as used herein, also includes antibody fragments either produced by the modification of whole antibodies, or those synthesised *de novo* using recombinant DNA methodologies (e.g., single chain Fv) or those identified using phage display libraries (see, e.g., McCafferty *et al.*, *Nature* 348:552-554 (1990)).

For preparation of monoclonal or polyclonal antibodies, any technique known in the art can be used (see, e.g., Kohler & Milstein, *Nature* 256:495-497 (1975); Kozbor *et al.*, *Immunology Today* 4: 72 (1983); Cole *et al.*, pp. 77-96 in *Monoclonal Antibodies and Cancer Therapy* (1985)). Techniques for the production of single chain antibodies (U.S. Patent 4,946,778) can be adapted to produce antibodies to polypeptides of this invention. Also, transgenic mice, or other organisms such as other mammals, may be used to express humanised antibodies. Alternatively, phage display technology can be used to identify antibodies and heteromeric Fab fragments that specifically bind to selected antigens (see, e.g., McCafferty *et al.*, *Nature* 348:552-554 (1990); Marks *et al.*, *Biotechnology* 10:779-783 (1992)).

The phrase “specifically (or selectively) binds” to an antibody or “specifically (or selectively) immunoreactive with,” when referring to a protein or peptide, refers to a binding reaction that is determinative of the presence of the protein in a heterogeneous population of proteins and other biologics. Thus, under designated immunoassay conditions, the specified antibodies
5 bind to a particular protein at least two times the background and do not substantially bind in a significant amount to other proteins present in the sample. Specific binding to an antibody under such conditions may require an antibody that is selected for its specificity for a particular protein. For example, polyclonal antibodies raised to fusion proteins can be selected to obtain only those polyclonal antibodies that are specifically immunoreactive with fusion protein and
10 not with individual components of the fusion proteins. This selection may be achieved by subtracting out antibodies that cross-react with the individual antigens. A variety of immunoassay formats may be used to select antibodies specifically immunoreactive with a particular protein. For example, solid-phase ELISA immunoassays are routinely used to select antibodies specifically immunoreactive with a protein (see, e.g., Harlow & Lane, *Antibodies, A Laboratory Manual* (1988) and *Using Antibodies: A Laboratory Manual* (1998), for a description
15 of immunoassay formats and conditions that can be used to determine specific immunoreactivity). Typically a specific or selective reaction will be at least twice background signal or noise and more typically more than 10, 20 or 100 times background (e.g. binding to other *Mycobacterium* proteins, such as other *Mycobacterium tuberculosis* proteins).

20

DIAGNOSTICS

In another aspect, this invention provides methods for using one or more of the polypeptides described above to diagnose tuberculosis (for example using T cell response based assays or
25 antibody based assays of conventional format).

For example, there is provided a method for determining prior *M. tuberculosis* infection in an individual comprising:

- (a) obtaining a sample from the individual;
- 30 (b) contacting said sample with an isolated polypeptide which comprises:
 - (i) an Rv2386c protein sequence;
 - (ii) a variant of an Rv2386c protein sequence; or
 - (iii) an immunogenic fragment of an Rv2386c protein sequence;
- (c) quantifying the sample response.

35

The sample may for example be whole blood or purified cells. Suitably the sample will contain peripheral blood mononucleated cells (PBMC). In one embodiment of the invention the

individual will be seropositive. In a second embodiment of the invention the individual will be seronegative.

Suitably the individual will not previously have been vaccinated against *M. tuberculosis*
5 infection (e.g. suitably the individual will not previously have been vaccinated with BCG).

The sample response may be quantified by a range of means known to those skilled in the art, including the monitoring of lymphocyte proliferation or the production of specific cytokines or antibodies. For example, T-cell ELISPOT may be used to monitor cytokines such as interferon
10 gamma (IFN γ), interleukin 2 (IL2) and interleukin 5 (IL5). B-cell ELLISPOT may be used to monitor the stimulation of *M. tuberculosis* specific antigens. The cellular response may also be characterised by the use of by intra- and extra-cellular staining and analysis by a flow cytometer.

15 Methods of quantifying a sample proliferation response include:

- (i) pulsing cultured cells with a radiolabel (e.g. tritiated thymidine) and monitoring tritium uptake (e.g. gas scintillation);
- (ii) carboxyfluorosecein diacetate succinimidyl ester (CFSE) labelling and
20 fluorescence monitoring of cell division using flow cytometry.

Quantifying a sample cytokine response includes in particular the monitoring of interferon gamma production.

When using such quantification methods, a positive response to an antigen may be defined by
25 a signal to noise ratio (S/N ratio) of at least 2:1 (for example, at least 3:1 or at least 5:1).

In a further aspect of the present invention methods are provided to diagnose *M. tuberculosis* infection using a skin test. As used herein, a "skin test" is any assay performed directly on a patient in which a delayed-type hypersensitivity (DTH) reaction (such as swelling, reddening or
30 dermatitis) is measured following intradermal injection of an Rv153c polypeptide as described above (or variant, immunogenic fragments thereof or nucleotides encoding them). Such injection may be achieved using any suitable device sufficient to contact the antigen combinations with dermal cells of the patient, such as a tuberculin syringe or 1 mL syringe. The reaction is measured after a period of time, for example at least 48 hours after injection,
35 especially 48-72 hours.

The DTH reaction is a cell-mediated immune response, which is greater in patients that have been exposed previously to the test antigen. The response may be measured visually, using a ruler. In general, a response that is greater than about 0.5 cm in diameter, especially greater than about 1.0 cm in diameter, is a positive response, indicative of prior *M. tuberculosis* infection, which may or may not be manifested as an active disease.

For use in a skin test, the Rv2386c component is suitably formulated as a pharmaceutical composition containing a physiologically acceptable carrier. Suitably, the carrier employed in such pharmaceutical compositions is a saline solution with appropriate preservatives, such as phenol and/or Tween 80™.

The present invention further provides kits for use within any of the above diagnostic methods. Such kits typically comprise two or more components necessary for performing a diagnostic assay. Components may be compounds, reagents, containers and/or equipment.

For example, one container within a kit may contain a monoclonal antibody or fragment thereof that specifically binds to a protein. Such antibodies or fragments may be provided attached to a support material, as described above. One or more additional containers may enclose elements, such as reagents or buffers, to be used in the assay. Such kits may also, or alternatively, contain a detection reagent as described above that contains a reporter group suitable for direct or indirect detection of antibody binding.

Alternatively, a kit may be designed to detect the level of mRNA encoding a protein in a biological sample. Such kits generally comprise at least one oligonucleotide probe or primer, as described above, that hybridises to a polynucleotide encoding a protein. Such an oligonucleotide may be used, for example, within a PCR or hybridization assay. Additional components that may be present within such kits include a second oligonucleotide and/or a diagnostic reagent or container to facilitate the detection of a polynucleotide encoding a protein of the invention.

Other diagnostics kits include those designed for the detection of cell mediated responses (which may, for example, be of use in the diagnostic methods of the present invention). Such kits will typically comprise:

- (i) apparatus for obtaining an appropriate cell sample from a subject;
- (ii) means for stimulating said cell sample with an Rv2386c polypeptide (or variant thereof, immunogenic fragments thereof, or DNA encoding such polypeptides);
- (iii) means for detecting or quantifying the cellular response to stimulation.

Suitable means for quantifying the cellular response include a B-cell ELISPOT kit or alternatively a T-cell ELISPOT kit, which are known to those skilled in the art.

5 One possible kit comprises:

- (a) a polypeptide of the invention; and
- (b) a detection reagent suitable for direct or indirect detection of antibody binding.

Of particular interest are diagnostic kits tailored for quantifying T cell responses:

10

A diagnostic kit comprising:

- (a) a polypeptide of the invention; and
- (b) apparatus sufficient to contact said polypeptide with the dermal cells of an individual.

15

A diagnostic kit comprising:

- (a) a polypeptide of the invention;
- (b) apparatus sufficient to contact said polypeptide with a sample (e.g. whole blood or more suitably PBMC) from an individual; and
- 20 (c) means to quantify the T cell response (e.g. proliferation or IFN-gamma production).

EXAMPLES

25 The following examples are provided by way of illustration only and not by way of limitation. Those of skill in the art will readily recognize a variety of noncritical parameters that could be changed or modified to yield essentially similar results.

EXAMPLE 1 – IDENTIFICATION OF Rv2386c AS A LATENT TB VACCINE TARGET

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The gene Rv2386c also known as MbtI, encodes for a protein which is involved in the biogenesis of hydroxyphenyloxazoline-containing siderophore mycobactins where it converts chorismate to salicylate, the starter unit for mycobacterin siderophore synthesis.

35 Rv2386c was selected based on a genome-wide analysis of *Mycobacterium tuberculosis* genes associated with dormancy phase maintenance and infectivity as in Murphy and Brown *BMC.Infect.Dis.* 2007 7:84-99. Potential dormancy phase gene targets in *Mycobacterium*

tuberculosis were prioritised through a bioinformatics meta-analysis of published genome-wide DNA microarray datasets of bacterial gene expression under simulated dormancy conditions. Subcellular localisation of *M. tuberculosis* proteins encoded by genes, was subsequently carried out on the entire genome to identify vaccine targets.

5

Briefly, experimental conditions in the dormancy models were quite varied so a zero to five scoring system was developed to normalise these data based upon two criteria: 1) the relevance of the experimental conditions to the dormant state and 2) the rank order of expression. The maximum score for a particular experimental dataset was adjusted based on potential relevance to the clinical occurrence of dormancy phase *M. tuberculosis* infections. Table 1 shows the data sets collected for Step 1 together with the adjusted maximum scores for each dataset. Additional datasets on gene essentiality for growth were obtained from published studies using transposon-based knockout experiments (TraSH). Genes which had no effect on growth received a score of zero.

10
15

Table 1 – Sources, experimental models, and scoring criteria for *M. tuberculosis* DNA microarray gene expression and genome-wide gene knock-out (growth phase essentiality).

Reference	Experimental model	Timepoint: Maximum score ^a
Betts JC et al. <i>Mol. Microbiol.</i> 2002 43:717-731	Starvation under controlled O ₂	96h: 3 24h: 2 4h: 1
Hampshire T et al. <i>Tuberculosis.(Edinb.)</i> 2004 84:228-238	Nutrient depletion under controlled O ₂	62 and 75d: 5 49d: 4 18d: 2
Muttucumaru DG et al. <i>Tuberculosis.(Edinb.)</i> 2004 84:239-246	Wayne model of hypoxia [#]	14d (NRP-2): 4 7d (NRP-1): 2
Voskuil MI et al. <i>Tuberculosis.(Edinb.)</i> 2004 84:218-227	Wayne model of hypoxia [#]	30 and 80d: 5 14 and 20d: 4 10 and 12d: 3 6 and 8d: 2
Schnappinger D et al. <i>J. Exp. Med.</i> 2003 198:693-704	Infection of mouse macrophages, +/- γ-INF	24 and 48h: 5

Reference	Experimental model	Timepoint: Maximum score ^a
Karakousis PC et al. <i>J. Exp. Med.</i> 2004 200:647-657	Hollow fiber subcutaneous implant in mice	10d: 3
Talaat AM et al. <i>Proc. Natl. Acad. Sci. U.S.A</i> 2004, 101:4602-4607	Infection of mice. MTB harvested from lung ^b	28d: 3
Sasseti CM et al. <i>Mol. Microbiol.</i> 2003 48:77-84	TraSH mutated libraries grown on solid media	14d:5
Rengarajan J et al. <i>Proc.Natl.Acad.Sci.U.S.A</i> 2005, 102:8327-8332	Infection of mouse macrophages, +/- γ -INF with TraSH mutated libraries of <i>M. tuberculosis</i>	7d:5
Sasseti CM et al. <i>Proc.Natl.Acad.Sci.U.S.A</i> 2003 100:12989-12994	C57BL/6J mice infected with TraSH mutated libraries of <i>M.</i> <i>tuberculosis</i>	7, 14, 28 and 56d:5

^aMaximum score based on relevance as a dormancy model; h = hour ; d = day.

^bRatio of *M. tuberculosis* from Balb/c lung to MTB in aerated culture for 28d.

Wayne LG and Hayes LG *Infect. Immun.* 1996 64:2062-2069

- 5 Step 2 - In applying the second criterion, the rank order of gene expression, gene scores from each dataset were ordered from highest to lowest based on expression ratio (fold expression in the experimental condition versus cells in log-phase liquid culture). The highest scoring gene received the maximum score for that particular dataset (listed in column 3 of Table 1. (e.g. 5, 4 ..., 1 point)). The score was decreased by 0.005 points for each gene in order until zero, or the
- 10 end of the data set was reached. Thus when the maximum score was 4 points, the 100th ranked gene would receive a score of 3.500. For a maximum score of 5 points, 1000 genes or 25% of the *M. tuberculosis* genome received a score. For experiments where data from multiple time points were collected, the maximum score across all time points was used as the final score.

15

In Step 3 scores for each gene in each of the experimental conditions were collected into a Microsoft Access database. Reference fields were added to facilitate prioritisation, such as the Refseq ID, Genbank function, Genbank note, Tuberculist classification, and KEGG and Sanger Center links. By combining the data from different studies and sources, a consensus view was

20 reached about the particular genes and pathways most critical for survival in the dormant state.

In Step 4, a prioritised list of therapeutic targets was derived utilising the top 400 scoring genes (~10% of the genome) supplemented by expert computational and manual analysis of biochemical pathways, enzymology, drug tractability, homology to human genes and other prior knowledge. The great majority of the high scoring genes come from the subset where two or three of the groups intersect.

In Step 5, the identification of subcellular localisation of *M. tuberculosis* proteins encoded by genes, was carried out on the entire genome. The heuristic used for membrane protein prediction is described in Chalker et al. *J. Bacteriol.* 2001 183:1259-1268. Average hydropathy profiles (H) (von Heijne G *J. Mol. Biol.* 1992 225:487-494) were generated using GES hydropathy values (Engelman DM et al. *Annu. Rev. Biophys. Biophys. Chem.* 1986 15:321-353) weighted using a trapezoid window. Using a process similar to the initial steps of the TopPred II algorithm (Claros MG et al. *Comput. Appl. Biosci.* 1994 10:685-686), helical transmembrane segments (TMS) were predicted for each peptide sequence by selecting 19 amino acids centered on the highest H value (MaxH), masking these from further consideration, and repeating the process until no peaks with a H of >0.5 remained. Subcellular locations were assigned based on the peak MaxH value, number of segments with a H of >1.0, and distribution and peak H values of the putative TMS. A MaxH cutoff of 1.15 was chosen to maximize the discrimination between two SwissProtein release 34 test datasets containing transmembrane and cytoplasmic proteins, respectively (Boyd D et al. *Protein Sci.* 1998 7:201-205). Proteins with a MaxH of <1.15 were classified as cytoplasmic, while those with a MaxH of >1.15 and at least three possible TMS were classified as membrane proteins. Anchored proteins were defined as having exactly two TMS, one starting before amino acid (aa) 35 and one having a H of >1.15 with the other having a H not lower than 0.5. SignalP with Gram positive settings was specifically used for *M. bacterium* to identify secreted proteins amongst those classified as either cytoplasmic or "unknown" in the heuristic analysis (Nielsen H et al. *Protein Eng.* 1997 10:1-6).

Rv2386c ranked very high as a vaccine antigen according to several criteria:

(i) Rv2386c is consistently up-regulated across all models of dormancy. Among the entire suite of 3999 genes scored in the meta-analysis, Rv2386c was ranked 120th as one of the top 10 % of over-expressed genes across all dormancy models. The up-regulated score for Rv2386c was 13.165 which favourably compared with the top gene score of 22.28. Rv2386c was minimally down-regulated, scoring only slightly less than 0 (-2.316 compared to -18.13 for the most widely down regulated gene).

5 (ii) The scavenging of iron from macrophages is crucial for *M. tuberculosis* survival and infectivity. Small molecules called siderophores function in bacteria to bind iron for intra-cellular uptake, MbtI catalyses the first step in the formation of the only *M. tuberculosis* siderophore, mycobactin T, the biosynthesis of which depends on the production of salicylate from chorismate (Harrison et al. *J. Bacteriol.* 2006 188:6081-6091). Expression of MbtI is induced during *M. tuberculosis* infection of human THP-1 macrophages (Gold et al. *Mol. Microbiol.* 2001 42:851-865). The 3D protein structure is available and the mechanism of enzymatic action MbtI has been well-studied (Zwahlen et al. *Biochemistry* 2007 46:954-964). Rv2386c ranked high in essentiality scoring and is required by *M. tuberculosis* for survival in in vitro growth models (scoring 2.6 out of a possible scoring of 5).

15 (iii) Subcellular localisation predicted that the Rv2386c protein is secreted and thus has significant extracellular exposure, indicating suitability as a vaccine target.

EXAMPLE 2 – Rv2386c EPITOPE IDENTIFICATION

Method

20 T cell epitope prediction was based on the following approaches:

Prediction	Name	URL/References
CD4 and CD8	Multipred	Zhang,G.L., Khan,A.M., Srinivasan,K.N., August,J.T. and Brusic,V. (2005) "MULTIPRED: a computational system for prediction of promiscuous HLA binding peptides" Nucleic Acids Res. 33, W172 - W179.
	SVMHC	"Prediction of MHC class I binding peptides, using SVMHC." Pierre Dönnès and Arne Elofsson in: BMC Bioinformatics 2002 3: 25

Prediction	Name	URL/References
CD4	ProPred	Singh,H. and Raghava,G.P.S.(2001) "ProPred: Prediction of HLA-DR binding sites." <i>Bioinformatics</i> ,17(12), 1236-37.
	Tepitope2	In house program based on : H. Bian, J. Hammer (2004) "Discovery of promiscuous HLA-II-restricted T cell epitopes with TEPITOPE." <i>Methods</i> 34 : 468-75
CD8	nHLA	Bhasin M. and Raghava G P S (2006) "A hybrid approach for predicting promiscuous MHC class I restricted T cell epitopes"; <i>J. Biosci.</i> 32:31-42
	NetCTL	"An integrative approach to CTL epitope prediction. A combined algorithm integrating MHC-I binding, TAP transport efficiency, and proteasomal cleavage predictions." Larsen M.V., Lundegaard C., Kasper Lamberth, Buus S., Brunak S., Lund O., and Nielsen M. <i>European Journal of Immunology</i> . 35(8): 2295-303. 2005
	EpiJen	Doytchinova, I. A., P. Guan, D. R. Flower. "EpiJen: a server for multi-step T cell epitope prediction." <i>BMC Bioinformatics</i> , 2006, 7, 131.
	Syfpeithi	Hans-Georg Rammensee, Jutta Bachmann, Niels Nikolaus Emmerich, Oskar Alexander Bachor, Stefan Stevanovic: "SYFPEITHI: database for MHC ligands and peptide motifs." <i>Immunogenetics</i> (1999) 50: 213-219

Prediction	Name	URL/References
	PredTAP	Zhang,G.L., Petrovsky,N., Kwoh,C.K., August,J.T. and Brusic,V. (2006) "PRED ^{TAP} : a system for prediction of peptide binding to the human transporter associated with antigen processing." Immunome Res. 2(1), 3.
	PAPROC	C. Kuttler, A.K. Nussbaum, T.P. Dick, H.-G. Rammensee, H. Schild, K.P. Haderler, "An algorithm for the prediction of proteasomal cleavages", J. Mol. Biol. 298 (2000), 417-429 A.K. Nussbaum, C. Kuttler, K.P. Haderler, H.-G. Rammensee, H. Schild, "PAProC: A Prediction Algorithm for Proteasomal Cleavages available on the WWW", Immunogenetics 53 (2001), 87-94

Results

5

Table 2 – Putative Rv2386c human CD4⁺ T cell epitopes

Putative CD4 epitope number	Amino acid position	Epitope sequence	SEQ ID No:	HLA allele
1	54	WVLAAGVQA	SEQ ID No: 30	DRB1_0401
2	55	VLAAGVQAM	SEQ ID No: 31	DRB1_0301, DRB1_1301
3	73	VIRDGVTRR	SEQ ID No: 32	DRB1_0301, DRB1_0401
4	125	LAPHTPLAR	SEQ ID No: 33	DRB1_1301
5	148	IRLFDAGIR	SEQ ID No: 34	DRB1_1101, DRB1_1301, DRB1_1501
6	155	IRHREAIDR	SEQ ID No: 35	DRB1_0801, DRB1_1301
7	164	LLATGVREV	SEQ ID No: 36	DRB1_1301
8	187	FRRRVAVAV	SEQ ID No: 37	DRB1_0101, DRB1_0801
9	216	FAIDFPLTY	SEQ ID No: 38	DRB1_0301, DRB1_0401
10	220	FPLTYRLGR	SEQ ID No: 39	DRB1_1101
11	224	YRLGRRHNT	SEQ ID No: 40	DRB1_0801, DRB1_1101, DRB1_1501
12	234	VRSFLLQLG	SEQ ID No: 41	DRB1_1301
13	238	LLQLGGIRA	SEQ ID No: 42	DRB1_1501
14	253	LVTAVRADG	SEQ ID No: 43	DRB1_1301
15	257	VRADGVVIT	SEQ ID No: 44	DRB1_0301, DRB1_0401

Putative CD4 epitope number	Amino acid position	Epitope sequence	SEQ ID No:	HLA allele
16	262	VVITEPLAG	SEQ ID No: 45	DRB1_0301, DRB1_0401, DRB1_1301, DRB1_1501
17	295	IVEHAISVR	SEQ ID No: 46	DRB1_1301
18	321	IDFMTVRER	SEQ ID No: 47	DRB1_1301
19	375	FRLDECPRG	SEQ ID No: 48	DRB1_0301, DRB1_0401
20	384	LYSGAVVML	SEQ ID No: 49	DRB1_0301
21	385	YSGAVVMLS	SEQ ID No: 50	DRB1_0401, DRB1_1101
22	389	VVMLSADGG	SEQ ID No: 51	DRB1_0401, DRB1_1301
23	415	WLRAGAGII	SEQ ID No: 52	DRB1_0101

Table 3 – Putative Rv2386c human CD8+ T cell epitopes

Putative CD8 epitope number	Amino acid position	Epitope sequence	SEQ ID No:	HLA allele
1	3	ELSVATGAV	SEQ ID No: 53	A_0201
2	5	SVATGAVST	SEQ ID No: 54	A_0201
3	18	IPMPAGVNP	SEQ ID No: 55	B_0702, B51, Cw_0401
4	19	PMPAGVNPA	SEQ ID No: 56	A_0201
5	21	PAGVNPADL	SEQ ID No: 57	A_2402, B_3501
6	23	GVNPADLAA	SEQ ID No: 58	A3
7	25	NPADLAAEL	SEQ ID No: 59	A24, B_0702, B44, Cw_0401, Cw_0602, B7, B_3501, B51
8	28	DLAAELAAV	SEQ ID No: 60	A_0201
9	33	LAADVTESV	SEQ ID No: 61	A2, A_0201, B51
10	37	VTESVDEDY	SEQ ID No: 62	A1, A_0101, A_0301
11	38	TESVDEDYL	SEQ ID No: 63	B44
12	40	SVDEDYLLY	SEQ ID No: 64	A1, A_0101, A3
13	48	YECDGQWVL	SEQ ID No: 65	A24, B_4403, B51, Cw_0401, B44
14	52	GQWVLAAGV	SEQ ID No: 66	A2, A_0201
15	55	VLAAGVQAM	SEQ ID No: 67	A2, B8
16	65	ELDSDELRV	SEQ ID No: 68	A_0201
17	67	DSDELRVIR	SEQ ID No: 69	A_0101, A_0301
18	82	QQWSGRPGA	SEQ ID No: 70	A_0201, A_0301
19	86	GRPGAALGE	SEQ ID No: 71	A3, B8
20	87	RPGAALGEA	SEQ ID No: 72	B7, B_0702, B_3501, B51
21	91	ALGEAVDRL	SEQ ID No: 73	A2, A_0201
22	93	GEAVDRLLL	SEQ ID No: 74	B44
23	106	AFGWVAFEF	SEQ ID No: 75	A24
24	124	RLAPHTPLA	SEQ ID No: 76	A2, A_0201, A3
25	125	LAPHTPLAR	SEQ ID No: 77	A_0101, A_0301
26	126	APHTPLARV	SEQ ID No: 78	B7, B_3501, B51
27	127	PHTPLARVF	SEQ ID No: 79	A24, B44
28	130	PLARVFSR	SEQ ID No: 80	A_0101, A3, A_0301
29	133	RVFSRPRTRI	SEQ ID No: 81	B7
30	134	VFSRPRTRIM	SEQ ID No: 82	A24, B8

Putative CD8 epitope number	Amino acid position	Epitope sequence	SEQ ID No:	HLA allele
31	143	VSEKEIRLF	SEQ ID No: 83	A1, A24
32	153	AGIRHREAI	SEQ ID No: 84	B8, B51
33	164	LLATGVREV	SEQ ID No: 85	A2, A_0201
34	170	REVPQSRSV	SEQ ID No: 86	B44
35	172	VPQSRSVDV	SEQ ID No: 87	B7, B8
36	180	VSDDPSGFR	SEQ ID No: 88	A_0101, A_0301
37	183	DPSGFRRRV	SEQ ID No: 89	B_3501, B51
38	185	SGFRRRVAV	SEQ ID No: 90	B8, B51
39	186	GFRRRVAVA	SEQ ID No: 91	A_0301, B8
40	187	FRRRVAVAV	SEQ ID No: 92	A3, B8, B51
41	190	RVAVAVDEI	SEQ ID No: 93	A24, B7
42	198	IAAGRYHKV	SEQ ID No: 94	B8, B51
43	200	AGRYHKVIL	SEQ ID No: 95	B7, B8
44	202	RYHKVILSR	SEQ ID No: 96	A3, A24
45	206	VILSRCVEV	SEQ ID No: 97	A2, A_0201
46	214	VPFAIDFPL	SEQ ID No: 98	B7, B_3501, B51
47	216	FAIDFPLTY	SEQ ID No: 99	A1, A_0101, B_3501, B51
48	218	IDFPLTYRL	SEQ ID No: 100	B44
49	226	LGRRHNTPV	SEQ ID No: 101	B8, B51
50	228	RRHNTPVRS	SEQ ID No: 102	A3, B8
51	231	NTPVRSFLL	SEQ ID No: 103	A_0101, A24
52	233	PVRSFLLQL	SEQ ID No: 104	A_0201, B7
53	245	RALGYSPLE	SEQ ID No: 105	A2, A_0201, B_3501, B51
54	246	ALGYSPELV	SEQ ID No: 106	A_0101, A2, A_0201
55	249	YSPELVTAV	SEQ ID No: 107	A2
56	250	SPELVTA VR	SEQ ID No: 108	B_0702, B_3501, B51
57	256	AVRADGVVI	SEQ ID No: 109	A3, A_0301, B7
58	264	ITEPLAGTR	SEQ ID No: 110	A_0101, A_0301
59	266	EPLAGTRAL	SEQ ID No: 111	B7, B8, B_3501, B51
60	270	GTRALGRGP	SEQ ID No: 112	A3, B8
61	272	RALGRGPAI	SEQ ID No: 113	A24, B7, B51
62	288	LESNSKEIV	SEQ ID No: 114	B44
63	294	EIVEHAISV	SEQ ID No: 115	A_0201
64	295	IVEHAISVR	SEQ ID No: 116	A_0101
65	298	HAISVRSSL	SEQ ID No: 117	B7, B8, B_3501
66	301	SVRSSLEEI	SEQ ID No: 118	B7
67	305	SLEEITDIA	SEQ ID No: 119	A2
68	311	DIAEPGSAA	SEQ ID No: 120	A_0101, A_0301
69	313	AEPGSAAVI	SEQ ID No: 121	B44
70	327	RERGSVQHL	SEQ ID No: 122	B7, B44
71	331	SVQHLGSTI	SEQ ID No: 123	A24, B7
72	342	RLDPSSDRM	SEQ ID No: 124	A2, A_0201
73	344	DPSSDRMAA	SEQ ID No: 125	B7, B_3501
74	346	SSDRMAALE	SEQ ID No: 126	A1, B8
75	348	DRMAALEAL	SEQ ID No: 127	B7, Cw_0401, Cw_0602
76	349	RMAALEALF	SEQ ID No: 128	A24
77	351	AALEALFPA	SEQ ID No: 129	A2, A_0201, B_3501
78	352	ALEALFPAV	SEQ ID No: 130	A_0101, A2, A_0201
79	353	LEALFPAVT	SEQ ID No: 131	B8, B_4403

Putative CD8 epitope number	Amino acid position	Epitope sequence	SEQ ID No:	HLA allele
80	357	FPAVTASGI	SEQ ID No: 132	B7, B8, B_3501, B51
81	359	AVTASGIPK	SEQ ID No: 133	A3, A_0301, B7
82	360	VTASGIPKA	SEQ ID No: 134	A_0201
83	364	GIPKAAGVE	SEQ ID No: 135	A_0301, B8
84	365	IPKAAGVEA	SEQ ID No: 136	B7, B_3501
85	367	KAAGVEAIF	SEQ ID No: 137	A24, B_3501
87	369	AGVEAIFRL	SEQ ID No: 138	A_0201, B51
87	376	RLDECPRGL	SEQ ID No: 139	A2, A_0201
88	380	CPRGLYSGA	SEQ ID No: 140	B7, B_3501
89	383	GLYSGAVVM	SEQ ID No: 141	A2, A_0201, A_0301, A3
90	384	LYSGAVVML	SEQ ID No: 142	A24, B44
91	390	VMLSADGGL	SEQ ID No: 143	A2, A_0201, A24
92	397	GLDAALTLR	SEQ ID No: 144	A_0101, A3, A_0301
93	398	LDAALTLRA	SEQ ID No: 145	A_0201, B_4403
94	400	AALTLRAAY	SEQ ID No: 146	A_0101, A_0301, B_3501
95	402	LTLRAAYQV	SEQ ID No: 147	A_0201
96	407	AYQVGGRTW	SEQ ID No: 148	A24
97	414	TWLRAGAGI	SEQ ID No: 149	A24
98	424	EESEPEREF	SEQ ID No: 150	B44
99	430	REFEETCEK	SEQ ID No: 151	B44
100	438	KLSTLTPYL	SEQ ID No: 152	A2, A_0201
101	440	STLTPYLVA	SEQ ID No: 153	A_0101, A_0201
102	441	TLTPYLVAR	SEQ ID No: 154	A3, A_0301

As can be seen from Tables 2 and 3, Rv2386c contains a number of predicted CD4+ and CD8 T cell epitopes. Furthermore, this information suggests that the protein carries epitopes that can be recognised by HLAs which occur worldwide (that is HLAs from Caucasian, African,
5 Asian or Latin-American individuals – see website at www.allele frequencies.net).

EXAMPLE 3 – H37Rv HOMOLOGUES

Rv2386c sequences from a number of *M. tuberculosis* strains and BCG were identified using
10 BLASTP searches of GenBank (H37Rv reference sequence accession number YP_177877.1

Strain	Accession Number	% identity
CDC1551	NP_336935.1	100
F11	YP_001288342.1	100
Haarlem	ZP_02247771.1	100
C	ZP_00878160.1	99
BCG	CAL72388.1	100

Alignment of the homologue sequences indicates a high level of identity.

BIOLOGICAL ASSAYS

Quantification of T cell responses to Rv2386c

5

Polypeptides may be screened for their ability to activate T-cells (induction of proliferation and/or production of cytokines) in peripheral blood mononuclear cell (PBMC) or in whole blood preparations from infected (such as latently infected) individuals.

10

Latently infected individuals are usually identified by a skin test that has a diameter above 10mm and without symptoms, with no Mtb positive culture, with a negative sputum negative and with no lesion (as detected by chest X-Ray).

15

A range of *in vitro* assays can be used based on PBMC samples or whole blood: after restimulation in presence of the antigen (or variant/immunogenic fragment thereof as appropriate) the proliferation of the cells may be determined (as measured by CFSE/flow cytometry) or the production of cytokines quantified (present in the supernatant of cultured cells and measured by ELISA, or, after intracellular staining of CD4 and CD8 T cells and analysis by flow cytometry).

20

For example, PBMC samples may be obtained from heparinised whole blood by Ficoll-Hypaque density gradient centrifugation following standard procedures. The cells may then be washed and cryopreserved in liquid nitrogen until testing (for further details see Lalvani A et al. *J. Infect. Dis.* 1999 180:1656-1664).

T cell proliferation

25

The specific immune response may be characterised by performing lymphocyte proliferation analysis using the tritiated thymidine. This technique assesses the cellular expansion upon *in vitro* stimulation to an antigen. In practice, cell proliferation is determined by estimating incorporation of tritiated-thymidine into DNA, a process closely related to underlying changes in cell number.

30

More suitably, lymphocyte proliferation may be performed using the succinimidyl ester of carboxyfluorescein diacetate (CFSE). CFSE spontaneously and irreversibly couples to both intracellular and cell surface proteins by reaction with lysine side chains and other available amine groups. When lymphocyte cells divide, CFSE labelling is distributed equally between

the daughter cells, which are therefore half as fluorescent as the parents. As a result, halving of cellular fluorescence intensity marks each successive generation in a population of proliferating cells and is readily followed by flow cytometry (for further details see Hodgkins, PD et al *J. Exp. Med.* 1996 184:277-281).

5

Practically, after thawing, PMBC may be washed and stained with CFSE before being cultivated (2×10^5 cells) for 72 hrs with 10 ug/ml of antigen in culture media (RPMI-1640 supplemented with glutamine, non essential amino acid, pyruvate and heat inactivated human AB serum). Cells may then then harvested and their phenotype characterised using surface staining to identify memory CD8 and CD4+ T-Cells. Subsequently, flow cytometry analysis can be used to indicate the extent of lymphocyte proliferation in response to each antigen (proportion of cells with decreased CFSE intensity upon *in vitro* stimulation).

10

Cytokine production

15

IFN- γ production (or the production of other cytokines such as e.g. IL2, TNF- α , IL5, IL12 etc) may be measured using an enzyme-linked immunosorbent assay (ELISA). ELISA plates may be coated with a mouse monoclonal antibody directed to human IFN- γ (PharMingen, San Diego, CA) in PBS for four hours at room temperature. Wells are then blocked with PBS containing 5% (W/V) non-fat dried milk for 1 hour at room temperature. The plates are then washed, for example, six times in PBS/0.2% TWEEN-20 and samples diluted 1:2 in culture medium in the ELISA plates are incubated overnight at room temperature. The plates are again washed and a polyclonal rabbit anti-human IFN- γ serum, for example, diluted 1:3000 in PBS/10% normal goat serum may be added to each well. The plates are then incubated for two hours at room temperature, washed and horseradish peroxidase-coupled anti-rabbit IgG (Sigma Chemical So., St. Louis, MO) may be added, for example, at a 1:2000 dilution in PBS/5% non-fat dried milk. After a further two hour incubation at room temperature, the plates are washed and TMB substrate added. The reaction may be stopped after 20 min with 1 N sulfuric acid. Optical density can then be determined at 450 nm using 570 nm as a reference wavelength. Typically, fractions that result in both replicates giving an OD two fold greater than the mean OD from cells cultured in medium alone may be considered positive.

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EXAMPLE 4 – IMMUNOGENICITY IN CB6F1 MICE

The immunogenicity of the antigen was evaluated in CB6F1 mice (first generation cross of BALB/c and C57BL/6 mice).

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CB6F1 mice were immunised intramuscularly three times (on day 0, day 14 and day 28) with 0.5 ug or 2 ug of protein antigen in combination with the Adjuvant System AS01E (a liposomal adjuvant formulation comprising 3D-MPL and QS21).

- 5 The experimental design was as follows:

Group	Day 0	Day 14	Day 28
1	2 ug Rv2386c/AS01E	2 ug Rv2386c/AS01E	2 ug Rv2386c/AS01E
2	0.5 ug Rv2386c/AS01E	0.5 ug Rv2386c/AS01E	0.5 ug Rv2386c/AS01E

A total of 24 mice were used in each protocol group.

- 10 Peripheral blood lymphocytes (PBL) were collected and pooled on day 21 (i.e. 7 days post second immunisation) and day 35 (i.e. 7 days post third immunisation) and the antigen-specific CD4 & CD8 T cell responses (as determined by CD4 or CD8 T cells producing IL-2 and/or IFN-gamma and/or TNF-alpha) were measured by flow cytometry after overnight *in vitro* restimulation with pools of 15mer peptides covering the sequences of interest.
- 15 The detection of mouse T cells that express IL-2 and/or IFN-gamma and/or TNF-alpha was done by using short-term antigen-driven *in vitro* amplification of cytokine expression.

- Briefly, PharmLyse solution (BD-Pharmingen) was added to heparinised mouse peripheral blood in order to lyse the red blood cells. The PBLs (Peripheral Blood Lymphocytes) obtained
- 20 were washed and then incubated in the presence of a pool of 15-mer peptides - overlapping by 11 amino acids - covering the sequence of the antigen of interest and of 1 ug/ml of antibodies to CD28 and CD49d (BD-Pharmingen). Each 15-mer peptide was used at a final concentration of 1 ug/ml. Medium controls were also stimulated with antibodies to CD28 and CD49d.

25

The cytokine secretion blocking compound brefeldin-A (BD-Pharmingen) was added 2 h after the onset of the cultures at 37°C, 5% CO₂ and the cells maintained at 37°C, 5% CO₂ for 4 additional hours followed by overnight incubation at +4°C.

- 30 Cells were then harvested and stained with Pacific Blue-coupled anti-CD4 (BD – clone RM4-5, BD-Pharmingen) and peridinin chlorophyll A protein (PerCp) cyanin5.5 (Cy5.5)-coupled anti-CD8 alpha (clone 53-6.7, BD-Pharmingen) antibodies.

Cells were then washed, fixed, permeabilised (Cytofix-cytoperm kit, BD-Pharmingen) and stained with allophycocyanin-coupled anti-IFN-g antibodies (clone XMG1.2, BDPharmingen), fluorescein isothiocyanate (FITC)-coupled anti-IL-2 antibodies (clone JES 6-5H4, Beckman Coulter) and phycoerythrin (PE)-coupled anti-TNF alpha antibodies (clone MP6-XT22, BDPharmingen). After final washes, stained cells were analysed on a LSR II flow cytometer (Beckton-Dickinson). A minimum number of 10,000 cells were acquired in the CD8+ subset.

For further background see Walzer T et al *Cell Immunol.* 2000 206(1):16-25 and Maecker HT et al *J. Immunol. Methods* 2001 255(1-2):27-40.

10

As negative controls, some cells were also cultured overnight *in vitro* in culture medium (unstimulated). The antigen-specific responses were calculated by subtracting the average cytokine response produced by unstimulated cells from the average cytokine response produced by the peptide-stimulated cells.

15

At each timepoint and for each group, the data was collected from 4 pools of 6 mice each. The data below is presented as the % of CD4 or CD8 T cells producing IL-2 and/or IFN-gamma and/or TNF-alpha. Each individual pool of mice is plotted (triangles) as well as the average value of the group (bar).

20

Figure 1 shows that on day 21 (i.e. 7 days post second immunisation), Rv2386c-specific CD4 and CD8 T cell responses are detected in mice immunised with either dose of Rv2386c/AS01E. The levels of Rv2386c-specific CD4 T cell responses are similar regardless of the immunising dose of Rv2386. In contrast, mice immunised with 2 ug Rv2386c/AS01E displayed higher Rv2386c-specific CD8 T cell responses than mice immunised with 0.5 ug Rv2386c/AS01E.

25

Figure 2 shows the cytokine profile of CD4 T cell response from the Rv2386c peptide pool-stimulated PBL (not medium removed) on day 21 (i.e. 7 days post second immunisation).

30

Figure 3 shows the cytokine profile of CD8 T cell response from the Rv2386c peptide pool-stimulated PBL (not medium removed) on day 21 (i.e. 7 days post second immunisation).

Figure 4 shows that on day 35 (i.e. 7 days post third immunisation), Rv2386c-specific CD4 and CD8 T cell responses are detected in mice immunised with either dose of Rv2386c/AS01E. The levels of Rv2386c-specific CD4 T cell responses are similar regardless of the immunising

35

dose of Rv2386. In contrast, mice immunised with 2 ug Rv2386c/AS01E displayed higher Rv2386c-specific CD8 T cell responses than mice immunised with 0.5 ug Rv2386c/AS01E. The third immunisation dose increases the CD4 T cell response but not the CD8 T cell response.

5

Figure 5 shows the cytokine profile of CD4 T cell response from the Rv2386c peptide pool-stimulated PBL (not medium removed) on day 35 (i.e. 7 days post third immunisation).

Figure 6 shows the cytokine profile of CD8 T cell response from the Rv2386c peptide pool-stimulated PBL (not medium removed) on day 35 (i.e. 7 days post third immunisation).

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EXAMPLE 5 – IMMUNOGENICITY IN C57BL/6 MICE

The immunogenicity of the antigen was also evaluated in C57BL/6 mice.

15

C57BL/6 mice were immunised intramuscularly three times (on day 0, day 14 and day 28) with 1 ug or 4 ug of protein antigen in combination with a the Adjuvant System AS01E (a liposomal adjuvant formulation comprising 3D-MPL and QS21)..

20 The experimental design was the following:

Group	Day 0	Day 14	Day 28
1	4 ug Rv2386c/AS01E	4 ug Rv2386c/AS01E	4 ug Rv2386c/AS01E
2	1 ug Rv2386c/AS01E	1 ug Rv2386c/AS01E	1 ug Rv2386c/AS01E

Peripheral blood lymphocytes (PBL) were collected and pooled on day 21 (i.e. 7 days post second immunisation) and day 35 (i.e. 7 days post third immunisation) and the antigen-specific CD4 & CD8 T cell responses (as determined by CD4 or CD8 T cells producing IL-2 and/or IFN-gamma and/or TNF-alpha) were measured by flow cytometry after overnight *in vitro* restimulation with pools of 15mer peptides covering the sequences of interest. The procedure followed was as described previously.

25

30 As negative controls, some cells were also cultured overnight *in vitro* in culture medium (unstimulated). The antigen-specific responses were calculated by subtracting the average cytokine response produced by unstimulated cells from the average cytokine response produced by the peptide-stimulated cells.

At each timepoint and for each group, the data was collected from 4 pools of 6 mice each. The data below is presented as the % of CD4 or CD8 T cells producing IL-2 and/or IFN-gamma and/or TNF-alpha. Each individual pool of mice is plotted (triangles) as well as the average
5 value of the group (bar).

Figure 7 shows that on day 21 (i.e. 7 days post second immunisation), Rv2386c-specific CD4 and CD8 T cell responses are detected in mice immunised with either dose of Rv2386c/AS01E. The levels of Rv2386c-specific CD4 T cell responses are similar regardless
10 of the immunising dose of Rv2386c. In contrast, mice immunised with 4 ug Rv2386c/AS01E displayed higher Rv2386c-specific CD8 T cell responses than mice immunized with 1 ug Rv2386c/AS01E. Due to technical difficulties, data for the first pool of cells at the 4 ug dose are not available.

15 Figure 8 shows the cytokine profile of CD4 T cell response from the Rv2386c peptide pool-stimulated PBL (not medium removed) on day 21 (i.e. 7 days post second immunisation). Due to technical difficulties, data for the first pool of cells at the 4 ug dose are not available.

Figure 9 shows the cytokine profile of CD8 T cell response from the Rv2386c peptide pool-stimulated PBL (not medium removed) on day 21 (i.e. 7 days post second immunisation).
20 Due to technical difficulties, data for the first pool of cells at the 4 ug dose are not available.

Immunological data for day 35 were not yet available at the time this application was prepared.

25 In conclusion it may be noted that the Rv2386c antigen is capable of eliciting an immune response in both CB6F1 and C57BL/6 mice. Furthermore, the profile of cytokine production indicates that a large proportion of antigen-specific T-cells express a plurality of Th1 associated cytokines (i.e. a polyfunctional T-cell response is elicited). Importantly both CD4
30 and CD8 antigen-specific T-cells are present after immunisation, CD8 cells may be particularly important in a latent TB scenario. Rv2386c may therefore be expected to be of substantial value in the prevention, treatment and diagnosis of latent tuberculosis infection.

35 Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be readily apparent to one of ordinary

skill in the art in light of the teachings of this invention that certain changes and modifications may be made thereto without departing from the spirit or scope of the appended claims.

5 Throughout the specification and the claims which follow, unless the context requires otherwise, the word 'comprise', and variations such as 'comprises' and 'comprising', will be understood to imply the inclusion of a stated integer, step, group of integers or group of steps but not to the exclusion of any other integer, step, group of integers or group of steps.

WHAT IS CLAIMED IS:

1. An isolated polypeptide which comprises:
 - (i) an Rv2386c protein sequence of any one of SEQ ID NO: 1 and 3-7; or
 - (ii) an immunogenic fragment of at least 10 amino acid residues from the Rv2386c protein sequence of SEQ ID NO: 1
 for use as a medicament for the treatment or prevention of tuberculosis.
2. The polypeptide for use according to claim 1, which comprises an Rv2386c protein sequence of any one of SEQ ID NO: 1 and 3-7.
3. The polypeptide for use according to claim 1, which comprises the Rv2386c protein sequence of SEQ ID NO: 1.
4. The polypeptide for use according to claim 1, which comprises an immunogenic fragment of at least 10 amino acid residues from the Rv2386c protein sequence of SEQ ID NO: 1.
5. The polypeptide for use according to any one of claims 1 to 4, for use in the treatment of latent tuberculosis.
6. The polypeptide for use according to any one of claims 1 to 4, for use in the prevention of latent tuberculosis.
7. The polypeptide for use according to any one of claims 1 to 4, for use in the prevention of reactivation of tuberculosis.
8. The polypeptide for use according to any one of claims 1 to 4, for use in the delay of reactivation of tuberculosis.
9. An isolated polynucleotide comprising a nucleic acid sequence encoding a polypeptide which comprises:
 - (i) an Rv2386c protein sequence of any one of SEQ ID NO: 1 and 3-7; or
 - (ii) an immunogenic fragment of at least 10 amino acid residues from the Rv2386c protein sequence of SEQ ID NO: 1
 for use as a medicament for the treatment of tuberculosis.

10. The polynucleotide for use according to claim 9, comprising a nucleic acid sequence encoding a polypeptide which comprises an Rv2386c protein sequence of any one of SEQ ID NO: 1 and 3-7.
11. The polynucleotide for use according to claim 9, comprising a nucleic acid sequence encoding a polypeptide which comprises an immunogenic fragment of at least 10 amino acid residues from the Rv2386c protein sequence of SEQ ID NO: 1.
12. The polynucleotide for use according to any one of claims 9 to 11, for use in the treatment of latent tuberculosis.
13. An isolated polynucleotide comprising a nucleic acid sequence encoding a polypeptide which comprises:
 - (i) an Rv2386c protein sequence of any one of SEQ ID NO: 1 and 3-7; or
 - (ii) an immunogenic fragment of at least 10 amino acid residues from the Rv2386c protein sequence of SEQ ID NO: 1,for use in the prevention of reactivation of tuberculosis.
14. An isolated polynucleotide comprising a nucleic acid sequence encoding a polypeptide which comprises:
 - (i) an Rv2386c protein sequence of any one of SEQ ID NO: 1 and 3-7; or
 - (ii) an immunogenic fragment of at least 10 amino acid residues from the Rv2386c protein sequence of SEQ ID NO: 1,for use in the delay of reactivation of tuberculosis.
15. The polynucleotide for use according to claim 13 or 14, comprising a nucleic acid sequence encoding a polypeptide which comprises an Rv2386c protein sequence of any one of SEQ ID NO: 1 and 3-7.
16. The polynucleotide for use according to claim 13 or 14, comprising a nucleic acid sequence encoding a polypeptide which comprises an immunogenic fragment of at least 10 amino acid residues from the Rv2386c protein sequence of SEQ ID NO: 1.
17. A pharmaceutical composition comprising:
 - (a) a polypeptide which comprises:
 - (i) an Rv2386c protein sequence of any one of SEQ ID NO: 1 and 3-7; or

- (ii) an immunogenic fragment of at least 10 amino acid residues from the Rv2386c protein sequence of SEQ ID NO: 1; and
 - (b) a pharmaceutically acceptable carrier or excipient.
- 18. An immunogenic composition comprising:
 - (a) a polypeptide which comprises:
 - (i) an Rv2386c protein sequence of any one of SEQ ID NO: 1 and 3-7; or
 - (ii) an immunogenic fragment of at least 10 amino acid residues from the Rv2386c protein sequence of SEQ ID NO: 1; and
 - (b) a non-specific immune response enhancer.
- 19. The polypeptide for use, polynucleotide for use or composition according to any one of claims 1 to 18, for treatment in a subject that has active tuberculosis.
- 20. The polypeptide for use, polynucleotide for use or composition according to any one of claims 1 to 18, for treatment in a subject that has latent tuberculosis.
- 21. The polypeptide for use according to any one of claims 1 to 4, for the prevention of tuberculosis in a subject that does not have tuberculosis.
- 22. Use of a polypeptide comprising:
 - (i) an Rv2386c protein sequence of any one of SEQ ID NO: 1 and 3-7; or
 - (ii) an immunogenic fragment of at least 10 amino acid residues from the Rv2386c protein sequence of SEQ ID NO: 1;in the manufacture of a medicament for the treatment or prevention of tuberculosis.
- 23. Use of a polynucleotide comprising a nucleic acid sequence encoding a polypeptide comprising:
 - (i) an Rv2386c protein sequence of any one of SEQ ID NO: 1 and 3-7; or
 - (ii) an immunogenic fragment of at least 10 amino acid residues from the Rv2386c protein sequence SEQ ID NO: 1;in the manufacture of a medicament for the treatment of tuberculosis.
- 24. An isolated polypeptide which comprises:
 - (i) an Rv2386c protein sequence of any one of SEQ ID NO: 1 and 3-7; or
 - (ii) an immunogenic fragment of at least 10 amino acid residues from the Rv2386c protein sequence SEQ ID NO: 1;

for administration with one or more chemotherapeutic agents effective against a *M. tuberculosis* infection.

25. An isolated polynucleotide comprising a nucleic acid sequence encoding a polypeptide which comprises:
- (i) an Rv2386c protein sequence of any one of SEQ ID NO: 1 and 3-7; or
 - (ii) an immunogenic fragment of at least 10 amino acid residues from the Rv2386c protein sequence SEQ ID NO: 1;

for administration with one or more chemotherapeutic agents effective against a *M. tuberculosis* infection.

26. A composition comprising an Rv2386c component and an M72 component, wherein said Rv2386c component comprises:

- (i) an Rv2386c protein sequence of any one of SEQ ID NO: 1 and 3-7; or
- (ii) an immunogenic fragment of at least 10 amino acid residues from the Rv2386c protein sequence of SEQ ID NO: 1;

and said M72 component is:

- (i) a polypeptide which comprises an amino acid sequence having at least 90% identity to SEQ ID NO: 25; or
- (ii) a polynucleotide comprising a nucleic acid sequence encoding a polypeptide comprising an amino acid sequence having at least 90% identity to SEQ ID NO: 25;

and wherein when said M72 component is provided in the form of a polypeptide, the Rv2386c component and an M72 component may be provided as either two individual polypeptide components or as a fusion protein comprising both polypeptide components.

27. A composition comprising an Rv2386c component and an M72 component, wherein said Rv2386c component is a polynucleotide comprising a nucleic acid sequence encoding a polypeptide which comprises:

- (i) an Rv2386c protein sequence of any one of SEQ ID NO: 1 and 3-7; or
- (ii) an immunogenic fragment of at least 10 amino acid residues from the Rv2386c protein sequence of SEQ ID NO: 1;

and said M72 component is:

- (i) a polypeptide which comprises an amino acid sequence having at least 90% identity to SEQ ID NO: 25; or

(ii) a polynucleotide comprising a nucleic acid sequence encoding a polypeptide comprising an amino acid sequence having at least 90% identity to SEQ ID NO: 25:

and wherein when said M72 component is provided in the form of a polynucleotide, the Rv2386c component and an M72 component may be provided as two individual polynucleotide components, as a single polynucleotide encoding two individual polypeptide components or as a single polynucleotide encoding a fusion protein comprising both polypeptide components.

Figure 1

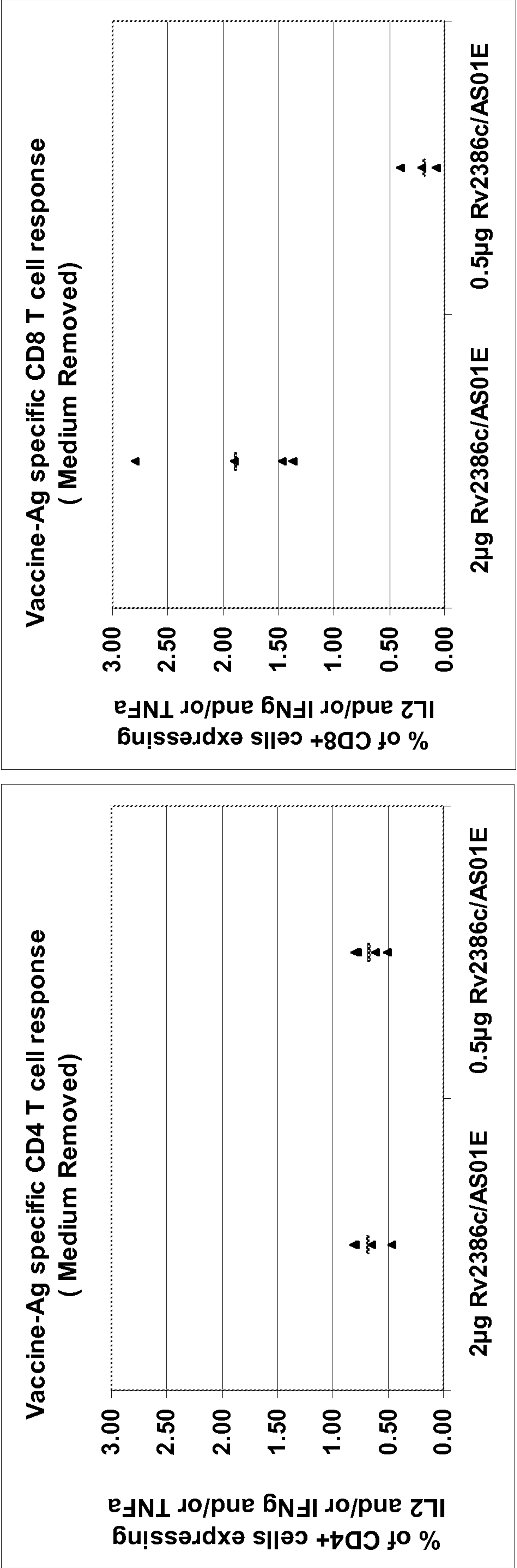


Figure 2

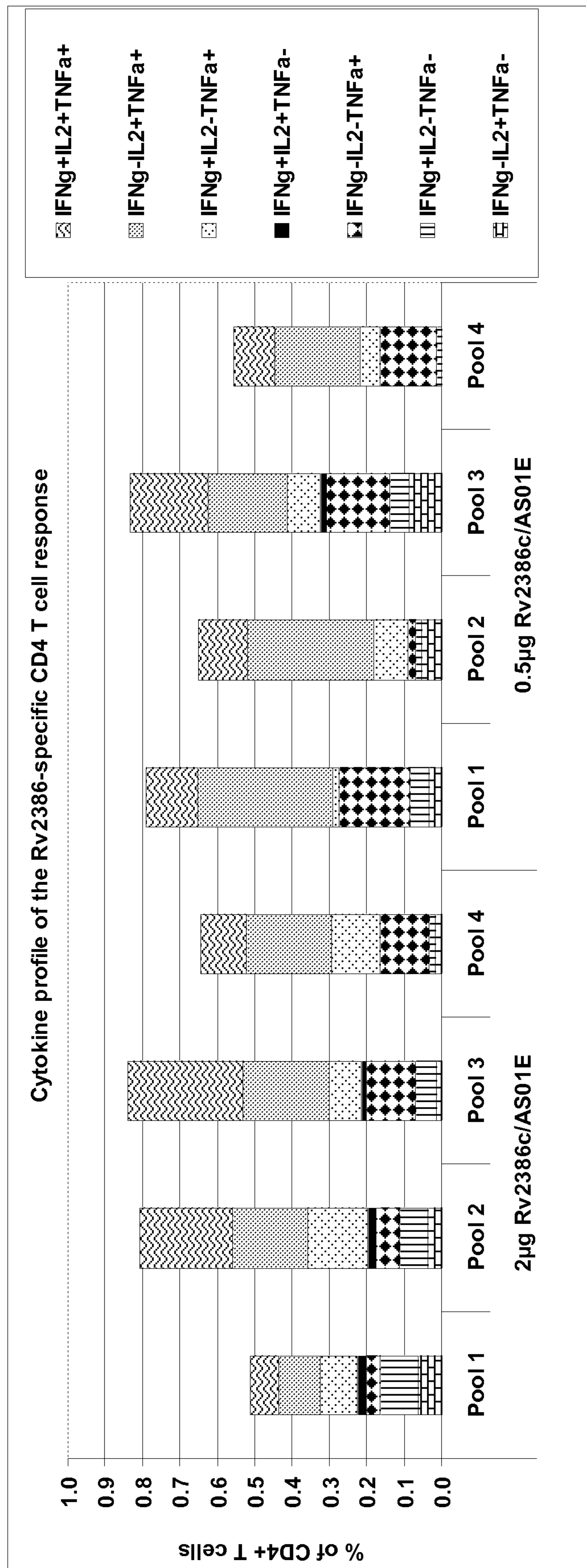


Figure 3

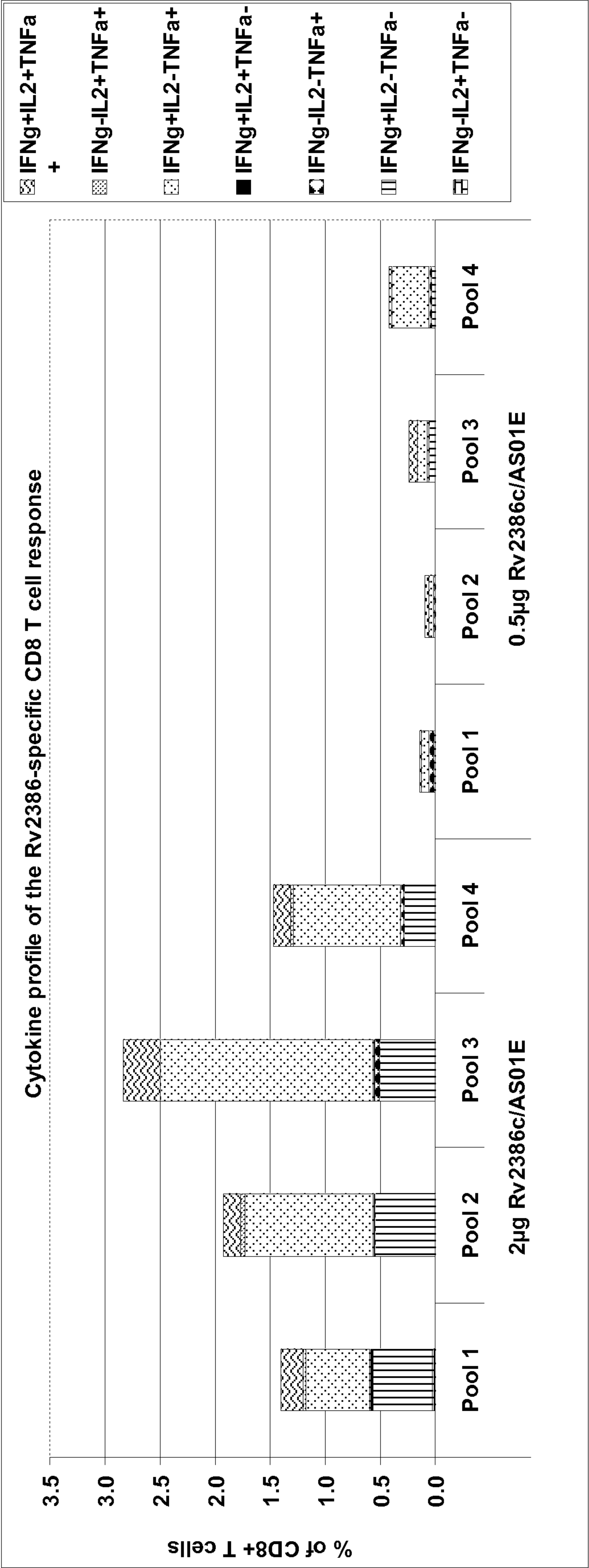


Figure 4

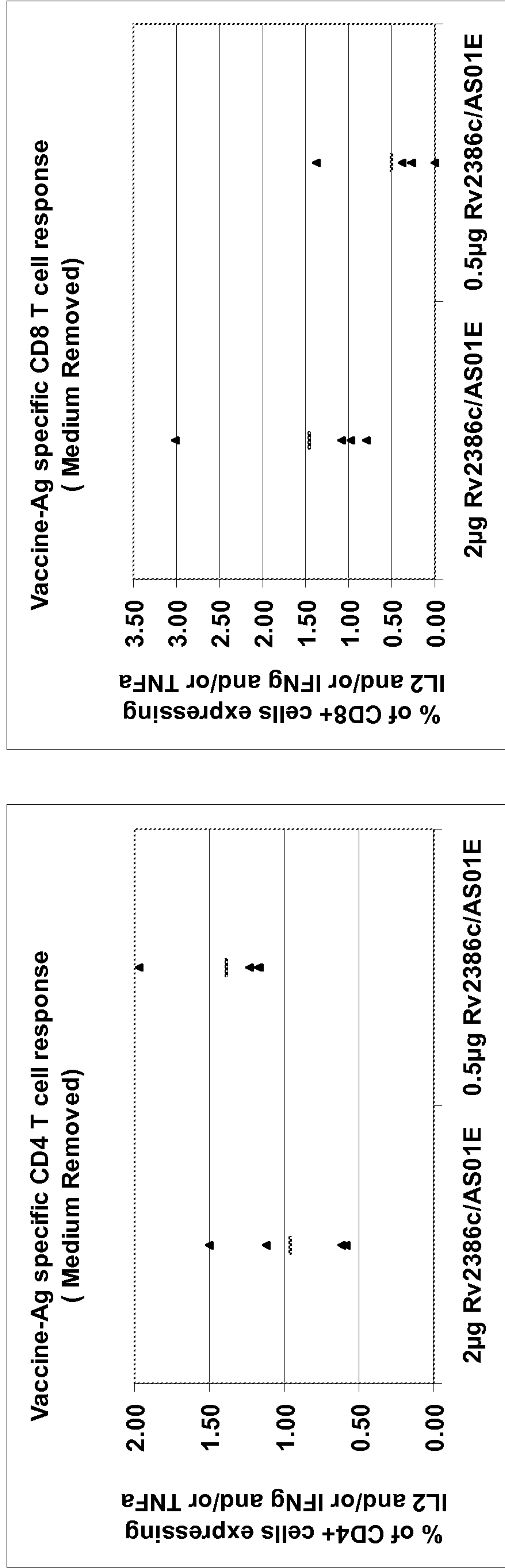


Figure 5

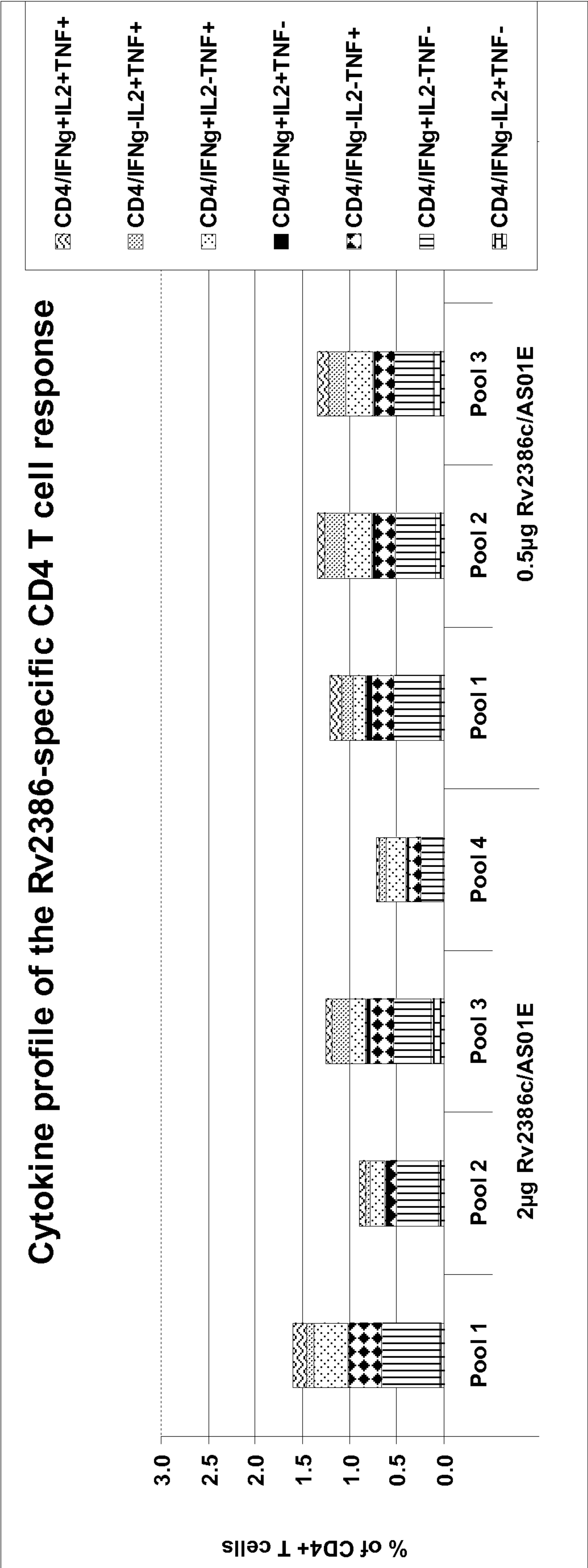


Figure 6

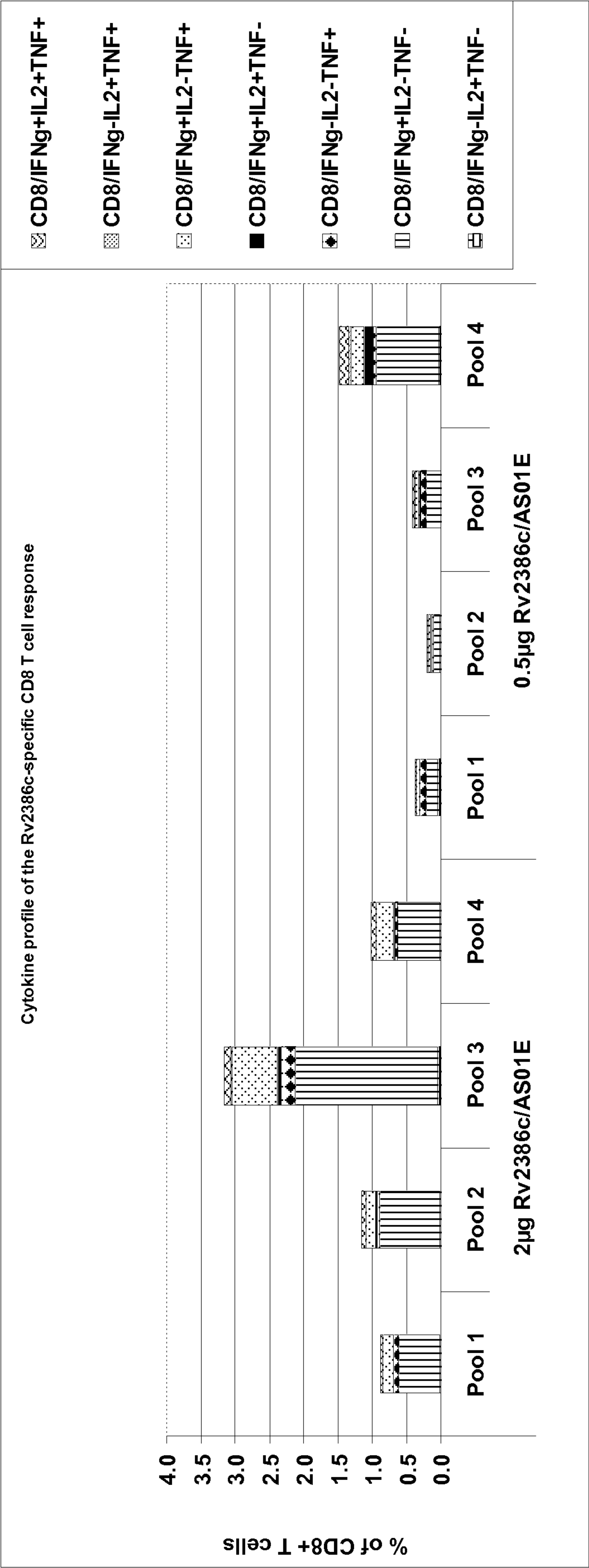


Figure 7

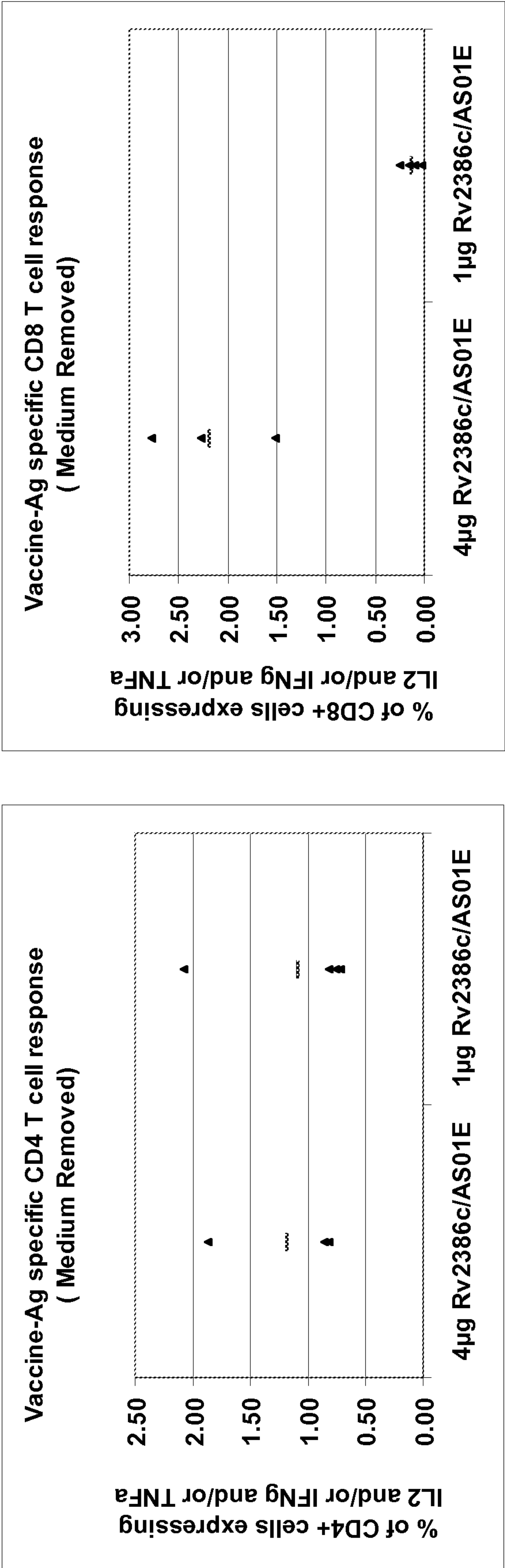


Figure 8

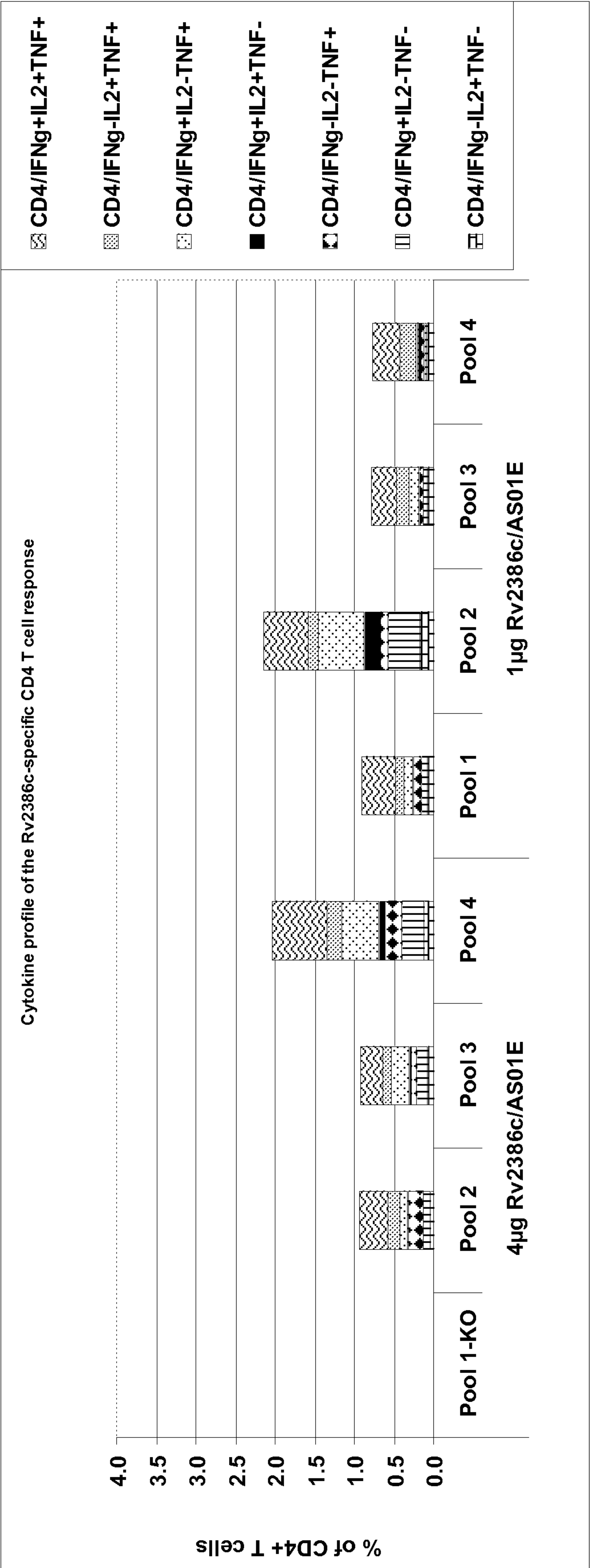


Figure 9

