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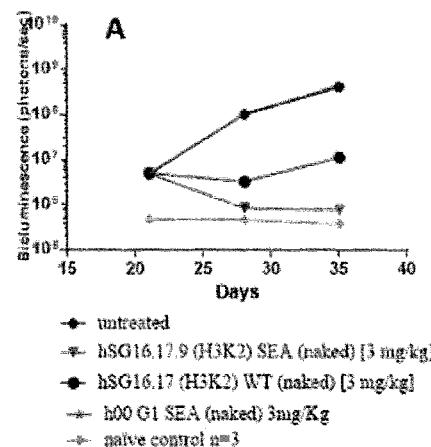
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(54) Title: BCMA ANTIBODIES AND USE OF SAME TO TREAT CANCER AND IMMUNOLOGICAL DISORDERS

Fig. 23. In Vivo Activity of Single Dosed hSG16.17-SEA in NCI-H929-luciferase Disseminated Tumor Model in NSG Mice.



(57) Abstract: The invention provides humanized antibodies that specifically bind to BCMA. The antibodies are useful for treatment and diagnosis of various cancers and immune disorders as well as detecting BCMA.

BCMA ANTIBODIES AND USE OF SAME TO TREAT CANCER AND IMMUNOLOGICAL DISORDERS**CROSS REFERENCE TO RELATED APPLICATION**

[0001] This application claims the benefit of U.S. provisional application no. US 62/296,594 filed February 17, 2016 and U.S. provisional application no. 62/396,084 filed September 16, 2016, both of which are incorporated herein by reference in their entirety for all purposes.

REFERENCE TO A SEQUENCE LISTING

[0002] Sequences disclosed in the application are contained in a sequence listing filed herewith.

BACKGROUND

[0003] B-cell maturation antigen (BCMA, CD269) is a member of the TNF receptor superfamily. Expression of BCMA is restricted to the B-cell lineage where it is predominantly expressed in the interfollicular region of germinal centers and on differentiated plasma cells and plasma blasts. BCMA binds to two distinct ligands, a proliferation inducing ligand (APRIL) and B-cell activating factor (BAFF, also known as BlyS, TALL-1, and THANK). The ligands for BCMA bind two additional TNF receptors, transmembrane activator and calcium modulator and cyclophilin ligand interactor (TACI) and BAFF receptor (BAFF-R also called BR3). TACI binds APRIL and BAFF, whereas BAFF-R shows restricted but high-affinity binding to BAFF. Together, BCMA, TACI, BAFF-R, and their corresponding ligands regulate different aspects of humoral immunity, B-cell development, and homeostasis.

[0004] BCMA is virtually absent on naïve and memory B cells (Novak et al., *Blood* 103, 689-94 (2004)) but it is selectively induced during plasma cell differentiation where it may support humoral immunity by promoting the survival of normal plasma cells and plasma blasts (O'Conner et al., *J. Exp Med.* 199, 91-98 (2004)). BCMA has been reported to be expressed in primary multiple myeloma (MM) samples. BCMA has also been detected on the Reed-

Sternberg cells (CD30⁺) from patients with Hodgkin's disease. It has been reported based on knockdown experiments that that BCMA contributed to both proliferation and survival of a Hodgkin's disease cell line (Chiu et al., Blood 109,729-39 (2007)).

SUMMARY OF THE CLAIMED INVENTION

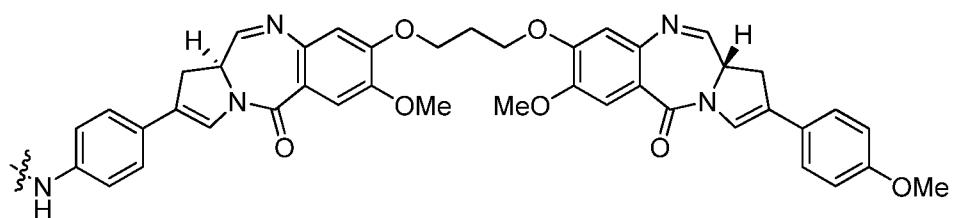
[0005] The invention provides a humanized, chimeric or veneered antibody, which is a humanized or chimeric form of an antibody deposited as ATCC PTC-6937. Optionally the antibody comprises a mature heavy chain variable region having at least 90% sequence identity to hSG16.17 VH3 (SEQ ID NO: 13) and a mature light chain variable region having at least 90% sequence identity to hSG16.17 VK2 (SEQ ID NO: 19). Optionally, the antibody comprises a mature heavy chain variable region having at least 95% sequence identity to hSG16.17 VH3 (SEQ ID NO: 13) and a mature light chain variable region having at least 95% sequence identity to hSG16.17 VK2 (SEQ ID NO: 19). Optionally, the antibody comprising the three Kabat CDRs (SEQ ID NOs: 60-62) of hSG16.17 VH3 (SEQ ID NO: 13) and three Kabat CDRs (SEQ ID NOs: 90-92) of hSG16.17 VK2 (SEQ ID NO: 19) provided that position H58 can be occupied by N or K, position H60 can be occupied by A or N, position H61 can be occupied by Q or E, position H62 can be occupied by K or N, position H64 can be occupied by Q or K, position H65 can be occupied by G or T, position L24 can be occupied by R or L and position L53 can be occupied by S or R. Optionally, the antibody comprises the three Kabat CDRs (SEQ ID NOs: 60-62) of hSG16.17 VH3 (SEQ ID NO: 13) and three Kabat CDRs (SEQ ID NOs: 90-92) of hSG16.17 VK2 (SEQ ID NO: 19). Optionally, positions H58, H60, H61, H62, H64 and H65 are occupied by N, A, Q, K, Q and G respectively and L24 and L53 are occupied by R and S respectively. Optionally, positions H20, H48, H69, H71, H73, H76, H80, H88, H91 and H93 are occupied by L, I, M, A, K, N, V, A, F, and T respectively, and positions L46, L48 and L87 are occupied by V, V and F respectively. Optionally, the mature heavy chain variable has the sequence of hSG16.17 VH3 (SEQ ID NO: 13) and the mature light chain variable region has the sequence of hSG16.17 VK2 (SEQ ID NO: 19).

[0006] The invention further provides a humanized, chimeric or veneered antibody, which is a humanized, chimeric or veneered form of the rat SG16.45 antibody having the VH (SEQ ID NO: 23) and VK (SEQ ID NO: 33) sequences. Optionally, the antibody comprises a heavy chain mature variable region having at least 90% sequence identity to hSG16.45 VH5 (SEQ ID NO: 31)

and a mature light chain variable region having at least 90% sequence identity to hSG16.45 VK2 (SEQ ID NO: 36). Optionally, the antibody comprises a mature heavy chain variable region having at least 95% sequence identity to hSG16.45 VH5 (SEQ ID NO: 31) and a mature light chain variable region having at least 95% sequence identity to hSG16.45 VK2 (SEQ ID NO: 36). Optionally, the antibody comprises the three Kabat CDRs (SEQ ID NOs: 152-154) of hSG16.45 VH5 (SEQ ID NO: 31) and three Kabat CDRs (SEQ ID NOs: 179-181) of hSG16.45 VK2 (SEQ ID NO: 36) provided that positions H50 can be occupied by A or S and position L24 can be occupied by R or L and position L26 can be occupied by S or T. Optionally, the antibody comprises the three Kabat CDRs (SEQ ID NOs: 152-154) of hSG16.45 VH5 (SEQ ID NO: 31) and three Kabat CDRs (SEQ ID NOs: 179-181) of hSG16.45 VK2 (SEQ ID NO: 36). Optionally positions H30, H93 and H94 are occupied by N, T and S respectively. Optionally, the mature heavy chain variable region has the sequence of hSG16.45 VH5 (SEQ ID NO: 31) and the mature light chain variable region has the sequence of hSG16.45 VK2 (SEQ ID NO: 36) or the mature heavy chain variable region has the sequence of hSG16.45 VH1 (SEQ ID NO: 27) and the mature light chain variable region has the sequence of hSG16.45 VK1 (SEQ ID NO: 35) or the mature heavy chain variable region has the sequence of hSG16.45 VH1 (SEQ ID NO: 27) and the mature light chain variable region has the sequence of hSG16.45 VK3 (SEQ ID NO: 37).

[0007] In any of the above antibodies, the mature heavy chain variable region can be fused to a heavy chain constant region and the mature light chain variable region can be fused to a light chain constant region. Optionally, the heavy chain constant region is a mutant form of natural human constant region which has reduced binding to an Fc_Y receptor relative to the natural human constant region. Optionally, the heavy chain constant region is of IgG1 isotype. Optionally, the heavy chain constant region has an amino acid sequence comprising SEQ ID NO: 5 and the light chain constant region has an amino acid sequence comprising SEQ ID NO: 3. Optionally, the heavy chain constant region has an amino acid sequence comprising SEQ ID NO: 7 (S239C) and the light chain constant region has an amino acid sequence comprising SEQ ID NO: 3. Optionally, the antibody is a naked antibody. Optionally, the antibody is conjugated to a cytotoxic or cytostatic agent. Optionally, the antibody is conjugated to a cytotoxic agent.

Optionally, the cytotoxic agent is conjugated to the via an enzyme cleavable linker. Optionally, the cytotoxic agent is a DNA minor groove binder, e.g., the cytotoxic agent having the formula



Optionally, the cytotoxic agent is MMAE or MMAF.

[0008] The invention further provides pharmaceutical compositions comprising any of the antibodies described above and a pharmaceutically acceptable carrier.

[0009] In one embodiment, the invention provides an antibody comprising the three Kabat CDRs (SEQ ID NOs: 60-62) of hSG16.17 VH3 (SEQ ID NO: 13) and three Kabat CDRs (SEQ ID NOs: 90-92) of hSG16.17 VK2 (SEQ ID NO: 19). In a further embodiment, the invention provides an antibody having a mature heavy chain variable with the sequence of hSG16.17 VH3 (SEQ ID NO: 13) and a mature light chain variable region with the sequence of hSG16.17 VK2 (SEQ ID NO: 19). In another embodiment, the mature heavy chain variable region is fused to a heavy chain constant region and the mature light chain variable region is fused to a light chain constant region. The antibody can be, e.g., an IgG1 antibody. In another embodiment, the antibody lacks core fucosylation by fucose or a fucose analogue. The antibodies can be formulated into a pharmaceutical composition, e.g., with addition of a pharmaceutically acceptable carrier.

[0010] In a further embodiment, the pharmaceutical composition has a plurality of antibodies having a mature heavy chain variable with the sequence of hSG16.17 VH3 (SEQ ID NO: 13) and a mature light chain variable region with the sequence of hSG16.17 VK2 (SEQ ID NO: 19). The variable regions of these antibodies are preferably fused to appropriate heavy and light chain constant regions. In another embodiment the antibodies are IgG1 antibodies. In a further embodiment, the plurality of antibodies has less than about 5% of the antibodies have core fucosylation by fucose or a fucose analogue. In a further embodiment, the plurality

of antibodies has less than about 10% of the antibodies have core fucosylation by fucose or a fucose analogue. In another embodiment, the plurality of antibodies includes about 2% antibodies with core fucosylation by fucose or a fucose analogue. In another embodiment, the plurality of antibodies includes 2% antibodies with core fucosylation by fucose or a fucose analogue.

[0011] The invention further provides a method of treating a patient having or at risk of having a cancer that expresses BCMA comprising administering to the patient an effective regime of an antibody as described above. Optionally the cancer is a hematological cancer. Optionally, the hematological cancer is a myeloma, leukemia or a lymphoma. Optionally, the hematological cancer is multiple myeloma. Optionally the hematological cancer is non-Hodgkin's lymphoma (NHL) or Hodgkin's lymphoma. Optionally, the hematological cancer is myelodysplastic syndromes (MDS), myeloproliferative syndromes (MPS), Waldenström's macroglobulinemia or Burkett's lymphoma.

[0012] The invention further provides a method of treating a patient having or at risk of having an immune disorder mediated by immune cells expressing BCMA comprising administering to the patient an effective regime of any of the above described antibodies. Optionally, the disorder is a B cell mediated disorder. Optionally, the immune disorder is rheumatoid arthritis, systemic lupus E (SLE), Type I diabetes, asthma, atopic dermatitis, allergic rhinitis, thrombocytopenic purpura, multiple sclerosis, psoriasis, Sjogren's syndrome, Hashimoto's thyroiditis, Grave's disease, primary biliary cirrhosis, Wegener's granulomatosis, tuberculosis, and graft versus host disease.

BRIEF DESCRIPTION OF THE FIGURES

[0013] Fig. 1A shows the structure of BCMA.

[0014] Fig. 1B shows the structural interaction of the extracellular domain of BCMA with BAFF.

[0015] Fig. 2 shows an antibody selection procedure.

[0016] Fig. 3 shows cell binding data and ligand blockade activity for uncloned hybridoma wells.

[0017] Fig. 4 shows blocking activity/percent inhibition of of anti-BCMA antibodies.

[0018] Fig. 5 shows inhibition of APRIL blocking titrated with anti-BCMA antibodies.

[0019] Fig. 6 shows a titration of BAFF blocking using anti-BCMA antibodies.

[0020] Fig. 7 shows alignment of hSG16.17 heavy chain variants with human VH acceptor sequence, HV1-2/HJ3. It shows rat SG16.17 vH (SEQ ID NO: 8) with Kabat CDRs (SEQ ID Nos: 39-41) and IMGT CDRs (SEQ ID NOs: 42 and 43); Hu HV1-2/HJ3 (SEQ ID NO: 9) with Kabat CDRs (SEQ ID NOs: 44 and 45) and IMGT CDRs (SEQ ID NO: 46 and “AR”); hSG16.17 vH1 (SEQ ID NO: 11) with Kabat CDRs (SEQ ID NOs: 50-52) and IMGT CDRs (SEQ ID NOs: 53 and 54); hSG16.17 vH2 (SEQ ID NO: 12) with Kabat CDRs (SEQ ID NOs: 55-57) and IMGT CDRs (SEQ ID NOs: 58 and 59); hSG16.17 vH3 (SEQ ID NO: 13) with Kabat CDRs (SEQ ID NOs: 60-62) and IMGT CDRs (SEQ ID NOs: 63 and 64); and hSG16.17 vH4 (SEQ ID NO: 14) with Kabat CDRs (SEQ ID NOs: 65-67) and IMGT CDRs (SEQ ID NOs: 68 and 69).

[0021] Fig. 8 shows alignment of hSG16.17 heavy chain variants with human VH acceptor sequence; HV1-46/HJ3. It shows the sequences of rat SG16.17 vH (SEQ ID NO: 8) with Kabat CDRs (SEQ ID NOs: 39-41) and IMGT CDRs (SEQ ID NOs: 42 and 43); Hu HV1-46/HJ3 (SEQ ID NO: 10) with Kabat CDRs (SEQ ID NOs: 47 and 48) and IMGT CDRs (SEQ ID NO: 49 and “AR”); hSG16.17 vH5 (SEQ ID NO: 15) with Kabat CDRs (SEQ ID NOs: 70-72) and IMGT CDRs (SEQ ID NOs: 73 and 74); and hSG16.17 vH6 (SEQ ID NO: 16) with Kabat CDRs (SEQ ID NOs: 75-77) and IMGT CDRs (SEQ ID NOs: 78 and 79).

[0022] Fig. 9 shows alignment of hSG16.17 heavy chain variants. It shows the sequences of hSG16.17 vH1-6 (SEQ ID NOs: 11-16).

[0023] Fig. 10 shows alignment of hSG16.17 light chain variants with human VK acceptor sequence; KV1-12/KJ5. It shows the sequences of rat SG16.17 vK (SEQ ID NO: 17) with Kabat CDRs (SEQ ID NOs: 80-82) and IMGT CDRs (SEQ ID NO: 83, “TTS”, and SEQ ID NO: 84, respectively); Hu KV1-12/KJ5 (SEQ ID NO: 18) with Kabat CDRs (SEQ ID NOs: 85-87) and IMGT CDRs (SEQ ID NO: 88, “AAS”, and SEQ ID NO: 89, respectively); hSG16.17 vK2 (SEQ ID NO: 19) with Kabat CDRs (SEQ ID NOs: 90-92) and IMGT CDRs (SEQ ID NO: 93, “TTS”, and SEQ ID NO: 94, respectively); hSG16.17 vK3 (SEQ ID NO: 20) with Kabat CDRs (SEQ ID NOs: 95-97) and IMGT CDRs (SEQ ID NO: 98, “TTS”, and SEQ ID NO: 99, respectively); hSG16.17 vK4 (SEQ ID NO: 21) with Kabat CDRs (SEQ ID NOs: 100-102) and IMGT CDRs (SEQ ID NO: 103, “TTS”, and SEQ

ID NO: 104, respectively); and hSG16.17 vK5 (SEQ ID NO: 22) with Kabat CDRs (SEQ ID NOs: 105-107) and IMGT CDRs (SEQ ID NO: 108, “TTS”, and SEQ ID NO: 109, respectively).

[0024] Fig. 11 shows alignment of hSG16.17 light chain variants. It shows the sequences of hSG16.17 vK2, vK3, vK4, vK5 (SEQ ID NOs: 19-22).

[0025] Fig. 12 shows competition binding assay showing binding of chimeric SG16.17 to human Fc γ RIIIa.

[0026] Fig. 13: shows chimeric SG16.17 induces signalling through Fc γ RIIIa.

[0027] Fig. 14 shows alignment of hSG16.45 heavy chain variants with human HV acceptor sequence HV3-23/HJ3. It shows the sequences of Rat SG16.45 vH (SEQ ID NO: 23) with Kabat CDRs (SEQ ID NOs: 110-112) and IMGT CDRs (SEQ ID NOs: 113-115); Hu HV3-23/HJ3 (SEQ ID NO: 24) with Kabat CDRs (SEQ ID NOs: 116 and 117) and IMGT CDRs (SEQ ID NOs: 118 and 119, and “AK”, respectively); hSG16.45 vH1 (SEQ ID NO: 27) with Kabat CDRs (SEQ ID NOs: 128-130) and IMGT CDRs (SEQ ID NOs: 131-133); hSG16.45 vH2 (SEQ ID NO: 28) with Kabat CDRs (SEQ ID NOs: 134-136) and IMGT CDRs (SEQ ID NOs: 137-139); hSG16.45 vH3 (SEQ ID NO: 29) with Kabat CDRs (SEQ ID NOs: 140-142) and IMGT CDRs (SEQ ID NOs: 143-145); and hSG16.45 vH4 (SEQ ID NO: 30) with Kabat CDRs (SEQ ID NOs: 146-148) and IMGT CDRs (SEQ ID NOs: 149-151).

[0028] Fig. 15 shows alignment of hSG16.45 heavy chain variants with human HV acceptor sequence HV3-74/HJ3. It shows the sequences of Rat SG16.45 vH (SEQ ID NO: 23) with Kabat CDRs (SEQ ID NOs: 110-112) and IMGT CDRs (SEQ ID NOs: 113-115); Hu HV3-74/HJ3 (SEQ ID NO: 25) with Kabat CDRs (SEQ ID NOs: 120 and 121) and IMGT CDRs (SEQ ID NOs: 122 and 123, and “AR”, respectively); hSG16.45 vH5 (SEQ ID NO: 31) with Kabat CDRs (SEQ ID NOs: 152-154) and IMGT CDRs (SEQ ID NOs: 155-157).

[0029] Fig. 16 shows alignment of hSG16.45 heavy chain variants with human HV acceptor sequence HV3-9/HJ3. It shows the sequences of Rat SG16.45 vH (SEQ ID NO: 23) with Kabat CDRs (SEQ ID NOs: 110-112) and IMGT CDRs (SEQ ID NOs: 113-115); Hu HV3-9/HJ3 (SEQ ID NO: 26) with Kabat CDRs (SEQ ID NOs: 124 and 125) and IMGT CDRs (SEQ ID NOs: 126 and 127, and “AR”, respectively); hSG16.45 vH6 (SEQ ID NO: 32) with Kabat CDRs (SEQ ID NOs: 158-160) and IMGT CDRs (SEQ ID NOs: 161-163).

[0030] Fig. 17 shows alignment of hSG16.45 heavy chain variants. It shows the sequences of hSG16.45 vH1-6 (SEQ ID NOs: 27-32).

[0031] Fig. 18 shows alignment of hSG16.45 light chain variants with human KV acceptor sequence KV3-20/KJ2. It shows the sequences of Rat SG16.45 vK (SEQ ID NO: 33) with Kabat CDRs (SEQ ID NOs: 164-166) and IMGT CDRs (SEQ ID NO: 167, "STS", and SEQ ID NO: 168, respectively); Hu KV3-20/KJ2 (SEQ ID NO: 34) with Kabat CDRs (SEQ ID NOs: 169-171) and IMGT CDRs (SEQ ID NO: 172, "STS", and SEQ ID NO: 173, respectively); hSG16.45 vK1 (SEQ ID NO: 35) with Kabat CDRs (SEQ ID NOs: 174-176) and IMGT CDRs (SEQ ID NO: 177, "STS", and SEQ ID NO: 178, respectively); hSG16.45 vK2 (SEQ ID NO: 36) with Kabat CDRs (SEQ ID NOs: 179-181) and IMGT CDRs (SEQ ID NO: 182, "STS", and SEQ ID NO: 183, respectively); hSG16.45 vK3 (SEQ ID NO: 37) with Kabat CDRs (SEQ ID NOs: 184-186) and IMGT CDRs (SEQ ID NO: 187, "STS", and SEQ ID NO: 188, respectively); and hSG16.45 vK5 (SEQ ID NO: 38) with Kabat CDRs (SEQ ID NOs: 189-191) and IMGT CDRs (SEQ ID NO: 192, "STS", and SEQ ID NO: 193, respectively).

[0032] Fig. 19. shows alignment of hSG16.45 light chain variants. It shows the sequences of hSG16.45 vK1, vK2, vK3, vK5 (SEQ ID NOs: 35-38).

[0033] Figs. 20A-C show in vivo activity of multi dosed hSG16.17-SEA in MM1S disseminated tumor model in SCID mice.

[0034] Figs. 21A-C show in vivo activity of single dosed hSG16.17-SEA in EJM disseminated tumor model in NSG mice.

[0035] Fig. 22 show in vivo activity of multi dosed hSG16.17-SEA in NCI-H929-luciferase disseminated tumor model in NSG mice.

[0036] Figs. 23A-B show in vivo activity of single dosed hSG16.17-SEA in NCI-H929-luciferase disseminated tumor model in NSG mice.

[0037] Figure 24 provides in vivo activity of single dosed hSG16.17-SEA in MOLP-8-luciferase disseminated tumor model in SCID mice.

[0038] Figure 25 provides ADCC activity of the SG16.17 SEA antibody on MM1R target cells.

DEFINITIONS

[0039] An “isolated” antibody refers to an antibody that has been identified and separated and/or recovered from components of its natural environment and/or an antibody that is recombinantly produced. A “purified antibody” is an antibody that is typically at least 50% w/w pure of interfering proteins and other contaminants arising from its production or purification but does not exclude the possibility that the monoclonal antibody is combined with an excess of pharmaceutical acceptable carrier(s) or other vehicle intended to facilitate its use. Interfering proteins and other contaminants can include, for example, cellular components of the cells from which an antibody is isolated or recombinantly produced. Sometimes monoclonal antibodies are at least 60%, 70%, 80%, 90%, 95 or 99% w/w pure of interfering proteins and contaminants from production or purification. The antibodies described herein, including rat, chimeric, veneered and humanized antibodies can be provided in isolated and/or purified form.

[0040] A “monoclonal antibody” refers to an antibody obtained from a population of substantially homogeneous antibodies, i.e., the individual antibodies comprising the population are identical except for possible naturally occurring mutations that may be present in minor amounts. The modifier “monoclonal” indicates the character of the antibody as being obtained from a substantially homogeneous population of antibodies, and is not to be construed as requiring production of the antibody by any particular method. For example, the monoclonal antibodies to be used in accordance with the present invention may be made by the hybridoma method first described by Kohler *et al.* (1975) *Nature* 256:495, or may be made by recombinant DNA methods (see, for example, U.S. Patent No. 4816567). The “monoclonal antibodies” may also be isolated from phage antibody libraries using the techniques described in Clackson *et al.* (1991) *Nature*, 352:624-628 and Marks *et al.* (1991) *J. Mol. Biol.*, 222:581-597, for example or may be made by other methods. The antibodies described herein are monoclonal antibodies.

[0041] Specific binding of a monoclonal antibody to its target antigen means an affinity of at least 10^6 , 10^7 , 10^8 , 10^9 , or 10^{10} M⁻¹. Specific binding is detectably higher in magnitude and distinguishable from non-specific binding occurring to at least one unrelated target. Specific binding can be the result of formation of bonds between particular functional groups or

particular spatial fit (e.g., lock and key type) whereas nonspecific binding is usually the result of van der Waals forces.

[0042] The basic antibody structural unit is a tetramer of subunits. Each tetramer includes two identical pairs of polypeptide chains, each pair having one "light" (about 25 kDa) and one "heavy" chain (about 50-70 kDa). The amino-terminal portion of each chain includes a variable region of about 100 to 110 or more amino acids primarily responsible for antigen recognition. This variable region is initially expressed linked to a cleavable signal peptide. The variable region without the signal peptide is sometimes referred to as a mature variable region. Thus, for example, a light chain mature variable region, means a light chain variable region without the light chain signal peptide. The carboxy-terminal portion of each chain defines a constant region primarily responsible for effector function.

[0043] Light chains are classified as either kappa or lambda. Heavy chains are classified as gamma, mu, alpha, delta, or epsilon, and define the antibody's isotype as IgG, IgM, IgA, IgD and IgE, respectively. Within light and heavy chains, the variable and constant regions are joined by a "J" region of about 12 or more amino acids, with the heavy chain also including a "D" region of about 10 or more amino acids. (*See generally, Fundamental Immunology* (Paul, W., ed., 2nd ed. Raven Press, N.Y., 1989, Ch. 7, incorporated by reference in its entirety for all purposes).

[0044] The mature variable regions of each light/heavy chain pair form the antibody binding site. Thus, an intact antibody has two binding sites. Except in bifunctional or bispecific antibodies, the two binding sites are the same. The chains all exhibit the same general structure of relatively conserved framework regions (FR) joined by three hypervariable regions, also called complementarity determining regions or CDRs. The CDRs from the two chains of each pair are aligned by the framework regions, enabling binding to a specific epitope. From N-terminal to C-terminal, both light and heavy chains comprise the domains FR1, CDR1, FR2, CDR2, FR3, CDR3 and FR4. The assignment of amino acids to each domain is in accordance with the definitions of Kabat, *Sequences of Proteins of Immunological Interest* (National Institutes of Health, Bethesda, MD, 1987 and 1991), or Chothia & Lesk, *J. Mol. Biol.* 196:901-917 (1987); Chothia et al., *Nature* 342:878-883 (1989), or a composite of Kabat and Chothia, or IMGT, AbM or Contact or other conventional definition of CDRs. Kabat also provides a widely used

numbering convention (Kabat numbering) in which corresponding residues between different heavy chains or between different light chains are assigned the same number. Unless otherwise apparent from the context, Kabat numbering is used to designate the position of amino acids in the variable regions. Unless otherwise apparent from the context EU numbering is used to designated positions in constant regions.

[0045] The term “antibody” includes intact antibodies and binding fragments thereof. Typically, antibody fragments compete with the intact antibody from which they were derived for specific binding to the target including separate heavy chains, light chains Fab, Fab', F(ab')₂, F(ab)c, diabodies, Dabs, nanobodies, and Fv. Fragments can be produced by recombinant DNA techniques, or by enzymatic or chemical separation of intact immunoglobulins. The term “antibody” also includes a diabody (homodimeric Fv fragment) or a minibody (V_L-V_H-C_H3), a bispecific antibody or the like. A bispecific or bifunctional antibody is an artificial hybrid antibody having two different heavy/light chain pairs and two different binding sites (see, e.g., Songsivilai and Lachmann, Clin. Exp. Immunol., 79:315-321 (1990); Kostelny et al., J. Immunol., 148:1547-53 (1992)).

[0046] The term “antibody” includes an antibody by itself (naked antibody) or an antibody conjugated to a cytotoxic or cytostatic drug.

[0047] A chimeric antibody is an antibody in which the mature variable regions of light and heavy chains of a non-human antibody (e.g., a mouse) are combined with human light and heavy chain constant regions. Such antibodies substantially or entirely retain the binding specificity of the mouse antibody, and are about two-thirds human sequence.

[0048] A veneered antibody is a type of humanized antibody that retains some and usually all of the CDRs and some of the non-human variable region framework residues of a non-human antibody but replaces other variable region framework residues that may contribute to B- or T-cell epitopes, for example exposed residues (Padlan, Mol. Immunol. 28:489, 1991) with residues from the corresponding positions of a human antibody sequence. The result is an antibody in which the CDRs are entirely or substantially from a non-human antibody and the variable region frameworks of the non-human antibody are made more human-like by the substitutions.

[0049] The term “epitope” refers to a site on an antigen to which an antibody binds. An epitope can be formed from contiguous amino acids or noncontiguous amino acids juxtaposed by tertiary folding of one or more proteins. Epitopes formed from contiguous amino acids are typically retained on exposure to denaturing solvents whereas epitopes formed by tertiary folding are typically lost on treatment with denaturing solvents. An epitope typically includes at least 3, and more usually, at least 5 or 8-10 amino acids in a unique spatial conformation. Methods of determining spatial conformation of epitopes include, for example, x-ray crystallography and 2-dimensional nuclear magnetic resonance. See, e.g., *Epitope Mapping Protocols*, in *Methods in Molecular Biology*, Vol. 66, Glenn E. Morris, Ed. (1996).

[0050] Antibodies that recognize the same or overlapping epitopes can be identified in a simple immunoassay showing the ability of one antibody to compete with the binding of another antibody to a target antigen. The epitope of an antibody can also be defined by X-ray crystallography of the antibody bound to its antigen to identify contact residues. Alternatively, two antibodies have the same epitope if all amino acid mutations in the antigen that reduce or eliminate binding of one antibody reduce or eliminate binding of the other. Two antibodies have overlapping epitopes if some amino acid mutations that reduce or eliminate binding of one antibody reduce or eliminate binding of the other.

[0051] Competition between antibodies is determined by an assay in which an antibody under test inhibits specific binding of a reference antibody to a common antigen (see, e.g., Junghans et al., *Cancer Res.* 50:1495, 1990). A test antibody competes with a reference antibody if an excess of a test antibody (e.g., at least 2x, 5x, 10x, 20x or 100x) inhibits binding of the reference antibody by at least 50% but preferably 75%, 90% or 99% as measured in a competitive binding assay. Antibodies identified by competition assay (competing antibodies) include antibodies binding to the same epitope as the reference antibody and antibodies binding to an adjacent epitope sufficiently proximal to the epitope bound by the reference antibody for steric hindrance to occur. Antibodies that compete with the h2H12 antibody for binding to the human BCMA protein are included in this disclosure.

[0052] The term “patient” includes human and other mammalian subjects that receive either prophylactic or therapeutic treatment.

[0053] For purposes of classifying amino acids substitutions as conservative or nonconservative, amino acids are grouped as follows: Group I (hydrophobic side chains): met, ala, val, leu, ile; Group II (neutral hydrophilic side chains): cys, ser, thr; Group III (acidic side chains): asp, glu; Group IV (basic side chains): asn, gln, his, lys, arg; Group V (residues influencing chain orientation): gly, pro; and Group VI (aromatic side chains): trp, tyr, phe. Conservative substitutions involve substitutions between amino acids in the same class. Non-conservative substitutions constitute exchanging a member of one of these classes for a member of another.

[0054] Percentage sequence identities are determined with antibody sequences maximally aligned by the Kabat numbering convention. After alignment, if a subject antibody region (e.g., the entire mature variable region of a heavy or light chain) is being compared with the same region of a reference antibody, the percentage sequence identity between the subject and reference antibody regions is the number of positions occupied by the same amino acid in both the subject and reference antibody region divided by the total number of aligned positions of the two regions, with gaps not counted, multiplied by 100 to convert to percentage.

[0055] Compositions or methods “comprising” one or more recited elements may include other elements not specifically recited. For example, a composition that comprises antibody may contain the antibody alone or in combination with other ingredients.

[0056] Designation of a range of values includes all integers within or defining the range.

[0057] An antibody effector function refers to a function contributed by an Fc domain(s) of an Ig. Such functions can be, for example, antibody-dependent cellular cytotoxicity, antibody-dependent cellular phagocytosis or complement-dependent cytotoxicity. Such function can be effected by, for example, binding of an Fc effector domain(s) to an Fc receptor on an immune cell with phagocytic or lytic activity or by binding of an Fc effector domain(s) to components of the complement system. Typically, the effect(s) mediated by the Fc-binding cells or complement components result in inhibition and/or depletion of the BCMA targeted cell. Fc regions of antibodies can recruit Fc receptor (FcR)-expressing cells and juxtapose them with antibody-coated target cells. Cells expressing surface FcR for IgGs including Fc γ RIII (CD16), Fc γ RII (CD32) and Fc γ RIII (CD64) can act as effector cells for the destruction of IgG-coated cells.

Such effector cells include monocytes, macrophages, natural killer (NK) cells, neutrophils and eosinophils. Engagement of Fc γ R by IgG activates antibody-dependent cellular cytotoxicity (ADCC) or antibody-dependent cellular phagocytosis (ADCP). ADCC is mediated by CD16 $^{+}$ effector cells through the secretion of membrane pore-forming proteins and proteases, while phagocytosis is mediated by CD32 $^{+}$ and CD64 $^{+}$ effector cells (see *Fundamental Immunology*, 4th ed., Paul ed., Lippincott-Raven, N.Y., 1997, Chapters 3, 17 and 30; Uchida *et al.*, 2004, *J. Exp. Med.* 199:1659-69; Akewanlop *et al.*, 2001, *Cancer Res.* 61:4061-65; Watanabe *et al.*, 1999, *Breast Cancer Res. Treat.* 53:199-207). In addition to ADCC and ADCP, Fc regions of cell-bound antibodies can also activate the complement classical pathway to elicit complement-dependent cytotoxicity (CDC). C1q of the complement system binds to the Fc regions of antibodies when they are complexed with antigens. Binding of C1q to cell-bound antibodies can initiate a cascade of events involving the proteolytic activation of C4 and C2 to generate the C3 convertase. Cleavage of C3 to C3b by C3 convertase enables the activation of terminal complement components including C5b, C6, C7, C8 and C9. Collectively, these proteins form membrane-attack complex pores on the antibody-coated cells. These pores disrupt the cell membrane integrity, killing the target cell (see *Immunobiology*, 6th ed., Janeway *et al.*, Garland Science, N. Y., 2005, Chapter 2).

[0058] The term “antibody-dependent cellular cytotoxicity”, or ADCC, is a mechanism for inducing cell death that depends upon the interaction of antibody-coated target cells with immune cells possessing lytic activity (also referred to as effector cells). Such effector cells include natural killer cells, monocytes/macrophages and neutrophils. The effector cells attach to an Fc effector domain(s) of Ig bound to target cells via their antigen-combining sites. Death of the antibody-coated target cell occurs as a result of effector cell activity.

[0059] The term “antibody-dependent cellular phagocytosis”, or ADCP, refers to the process by which antibody-coated cells are internalized, either in whole or in part, by phagocytic immune cells (e.g., macrophages, neutrophils and dendritic cells) that bind to an Fc effector domain(s) of Ig.

[0060] The term “complement-dependent cytotoxicity”, or CDC, refers to a mechanism for inducing cell death in which an Fc effector domain(s) of a target-bound antibody activates a

series of enzymatic reactions culminating in the formation of holes in the target cell membrane. Typically, antigen-antibody complexes such as those on antibody-coated target cells bind and activate complement component C1q which in turn activates the complement cascade leading to target cell death. Activation of complement may also result in deposition of complement components on the target cell surface that facilitate ADCC by binding complement receptors (e.g., CR3) on leukocytes.

[0061] A “cytotoxic effect” refers to the depletion, elimination and/or the killing of a target cell. A “cytotoxic agent” refers to an agent that has a cytotoxic effect on a cell.

[0062] Cytotoxic agents can be conjugated to an antibody or administered in combination with an antibody.

[0063] A “cytostatic effect” refers to the inhibition of cell proliferation. A “cytostatic agent” refers to an agent that has a cytostatic effect on a cell, thereby inhibiting the growth and/or expansion of a specific subset of cells. Cytostatic agents can be conjugated to an antibody or administered in combination with an antibody.

[0064] The term “pharmaceutically acceptable” means approved or approvable by a regulatory agency of the Federal or a state government or listed in the U.S. Pharmacopeia or other generally recognized pharmacopeia for use in animals, and more particularly in humans. The term “pharmaceutically compatible ingredient” refers to a pharmaceutically acceptable diluent, adjuvant, excipient, or vehicle with which an anti-BCMA antibody is administered to a subject.

[0065] The phrase “pharmaceutically acceptable salt,” refers to pharmaceutically acceptable organic or inorganic salts of an anti-BCMA-1 antibody or conjugate thereof or agent administered with an anti-BCMA-1 antibody. Exemplary salts include sulfate, citrate, acetate, oxalate, chloride, bromide, iodide, nitrate, bisulfate, phosphate, acid phosphate, isonicotinate, lactate, salicylate, acid citrate, tartrate, oleate, tannate, pantothenate, bitartrate, ascorbate, succinate, maleate, gentisinate, fumarate, gluconate, glucuronate, saccharate, formate, benzoate, glutamate, methanesulfonate, ethanesulfonate, benzenesulfonate, p toluenesulfonate, and pamoate (i.e., 1,1' methylene bis -(2 hydroxy 3 naphthoate)) salts. A pharmaceutically acceptable salt may involve the inclusion of another molecule such as an

acetate ion, a succinate ion or other counterion. The counterion may be any organic or inorganic moiety that stabilizes the charge on the parent compound. Furthermore, a pharmaceutically acceptable salt may have more than one charged atom in its structure. Instances where multiple charged atoms are part of the pharmaceutically acceptable salt can have multiple counter ions. Hence, a pharmaceutically acceptable salt can have one or more charged atoms and/or one or more counterion.

[0066] Unless otherwise apparent from the context, the term “about” encompasses insubstantial variation having no significant effect on functional properties (e.g., within a margin of error or experimental measurement).

DETAILED DESCRIPTION

I. General

[0067] The invention provides monoclonal antibodies that specifically bind to BCMA. The antibodies are useful for treatment and diagnoses of various cancers and immunological disorders as well as detecting BCMA.

II. Target molecules

[0068] Unless otherwise indicated, BCMA means a human BCMA. Exemplary human nucleic acid and amino acid sequences are provided by SEQ ID NOS:1 and 2. Unless otherwise apparent from the context reference to BMCA means at least an extracellular domain of the protein (approximately residues 1-54 of SEQ ID NO: 2) and sometimes the complete protein. Likewise, unless otherwise apparent from the context reference to BAFF and APRIL and their receptors other than BCMA refers to wild type human sequences e.g., as provided in the Swiss Prot Database.

III. Antibodies of the invention

A. Binding specificity and functional properties

[0069] The SG16.17 antibody is a rat monoclonal antibody that specifically binds to human BCMA as described in the examples. An ATCC deposit was made on August 15, 2005 under the Budapest Treaty. The ATCC is located at 10801 University Boulevard, Manassas, Va. 20110-

2209, USA. The ATCC deposit was assigned accession number of PTA-6937. The SG16.17 antibody inhibits binding of BCMA to both of its ligands, APRIL and BAFF. The SG16.17 antibody when linked to a human IgG1 elicits ADCC, binds to and elicits signaling through Fc γ receptors. The SG16.17 antibody can also be incorporated into an antibody drug conjugate to deliver a linked drug into the interior of cells expressing BCMA. The SG16.45 antibody is another rat monoclonal antibody that specifically binds to human BCMA, inhibits its binding to its ligands and can deliver a linked drug to the interior of cells expressing BCMA.

[0070] The invention provides humanized, chimeric and veneered forms of the SG16.17 antibody (designated hSG16.17, cSG16.17 or vSG16.17) and SG16.45 (analogously designated). Such antibodies typically retain some or all of the properties for SG16.17 or SG16.45 noted above. For any given property, humanized, chimeric or veneered antibodies may exhibit the property to the same extent within experimental error or more or less than rat SG16.17 or SG16.45. The affinity of humanized, chimeric or veneered forms of the rat SG16.17 antibody (i.e., K_a) can be greater than that of the rat SG16.17 antibody, or within a factor of five or a factor of two (i.e., more than or less than) than that of the rat SG16.17 antibody for human BCMA. Preferred humanized, chimeric or veneered SG16.17 antibodies bind to the same epitope and/or compete with rat SG16.17 antibodies for binding to human BCMA. The affinity of humanized, chimeric or veneered forms of the rat SG16.45 antibody (i.e., K_a) can be greater than that of the rat SG16.45 antibody, or within a factor of five or a factor of two (i.e., more than or less than) than that of the rat SG16.45 antibody for human BCMA. Preferred humanized, chimeric or veneered SG16.45 antibodies bind to the same epitope and/or compete with rat SG16.45 antibodies for binding to human BCMA.

[0071] Preferred humanized, chimeric and veneered antibodies inhibit cancer (e.g., growth of cells, metastasis and/or lethality to the organisms) or B-cell mediated immune disorders as shown in vitro, in an animal model or clinical trial.

B. Antibodies

[0072] A humanized antibody is a genetically engineered antibody in which CDRs from a non-human “donor” antibody are grafted into human “acceptor” antibody sequences (see, e.g., Queen, US 5,530,101 and 5,585,089; Winter, US 5,225,539; Carter, US 6,407,213; Adair, US

5,859,205; and Foote, US 6,881,557). The acceptor antibody sequences can be, for example, a mature human antibody sequence, a composite of such sequences, a consensus sequence of human antibody sequences, or a germline region sequence. For humanization of SG16.17, a preferred acceptor sequence for the heavy chain is the germline V_H exon V_H1-2 and for the J exon (J_H), exon J_H-3 . For the light chain, a preferred acceptor sequence is exon V_L1-12 and J exon J_K5 . For humanization of SG16.45, a preferred heavy chain acceptor sequence is HV3-23/HJ3 (SEQ ID NO: 24) and a preferred light chain acceptor sequence is KV3-20/KJ2 (SEQ ID NO: 34).

[0073] Thus, a humanized antibody is an antibody having at least four CDRs entirely or substantially from a non-human donor antibody and variable region framework sequences and constant regions, if present, entirely or substantially from human antibody sequences. Similarly a humanized heavy chain has at least two and usually all three CDRs entirely or substantially from a donor antibody heavy chain, and a heavy chain variable region framework sequence and heavy chain constant region, if present, substantially from human heavy chain variable region framework and constant region sequences. Similarly a humanized light chain has at least two and usually all three CDRs entirely or substantially from a donor antibody light chain, and a light chain variable region framework sequence and light chain constant region, if present, substantially from human light chain variable region framework and constant region sequences. Other than nanobodies and dAbs, a humanized antibody comprises a humanized heavy chain and a humanized light chain. A CDR in a humanized or human antibody is substantially from or substantially identical to a corresponding CDR in a non-human antibody when at least 60%, 85%, 90%, 95% or 100% of corresponding residues (as defined by Kabat) are identical between the respective CDRs. The variable region framework sequences of an antibody chain or the constant region of an antibody chain are substantially from a human variable region framework sequence or human constant region respectively when at least 70%, 80%, 85%, 90%, 95% or 100% of corresponding residues defined by Kabat are identical.

[0074] Although humanized antibodies often incorporate all six CDRs (preferably as defined by Kabat, but alternatively as defined by IMGT, Chothia, composite Kabat-Chothia, AbM or Contact or other conventional definition) from a mouse antibody, they can also be made with

less than all CDRs (e.g., at least 4, or 5) CDRs from a mouse antibody (e.g., Pascalis *et al.*, *J. Immunol.* 169:3076, 2002; Vajdos *et al.*, *Journal of Molecular Biology*, 320: 415-428, 2002; Iwahashi *et al.*, *Mol. Immunol.* 36:1079-1091, 1999; Tamura *et al*, *Journal of Immunology*, 164:1432-1441, 2000).

[0075] Certain amino acids from the human variable region framework residues can be selected for substitution based on their possible influence on CDR conformation and/or binding to antigen. Investigation of such possible influences is by modeling, examination of the characteristics of the amino acids at particular locations, or empirical observation of the effects of substitution or mutagenesis of particular amino acids.

[0076] For example, when an amino acid differs between a murine variable region framework residue and a selected human variable region framework residue, the human framework amino acid can be substituted by the equivalent framework amino acid from the mouse antibody when it is reasonably expected that the amino acid:

- (1) noncovalently binds antigen directly,
- (2) is adjacent to a CDR region,
- (3) otherwise interacts with a CDR region (e.g. is within about 6 Å of a CDR region); or
- (4) mediates interaction between the heavy and light chains.

[0077] The invention provides humanized forms of the rat SG16.17 antibody including **six** exemplified humanized heavy chain mature variable regions (hSG16.17 VH1-6) (SEQ ID Nos: 11-16) and four exemplified humanized light chain mature variable regions (hSG16.17 VK2-5) (SEQ ID NOS: 19-22). The heavy and light chains can be combined in any permutations, with permutations including any of hSG16.17 VH1, VH3 or VH5 being preferred. The permutation having the best combination of binding affinity, percentage sequence identity to human germline, expression and percentage of monomeric content was hSG16.17 VH3 VK2. This antibody shows similar affinity within experimental error as the rat SG16.17, greater than 85% sequence identity with human germline in both heavy and light chain variable regions (thus, qualifying for “humanized” designation under the new INN guideliness), high expression in CHO

cells, and high proportion of monomers. Compared with most other humanized antibodies hSG16.17 VH3 VK2 is unusual in having a large number of variable region framework mutations in which human acceptor residues are changed to the corresponding rat residue (13) but also having a large number of “forward” CDR mutations, in which a rat residue in the Kabat CDRs is changed to the corresponding residue in the human acceptor sequence, such that overall the antibody has sufficient sequence identity to human germline sequences to be classified as humanized under INN guidelines. Most previous humanized antibodies have had Kabat CDR entirely from the donor antibody.

[0078] The invention provides antibodies in which the heavy chain variable region shows at least 90% identity to hSG16.17 VH3 (SEQ ID NO: 13) and a light chain variable region at least 90% identical to hSG16.17 VK2 (SEQ ID NO: 19). Some antibodies show at least 95%, 96%, 97%, 98% or 99% sequence identity to HV3 and at least 95%, 96%, 97%, 98% or 99% sequence identity to VK2. Some such antibodies include the the three Kabat CDRs (SEQ ID NOs: 60-62) of hSG16.17 VH3 (SEQ ID NO: 13) and three Kabat CDRs (SEQ ID NOs: 90-92) of hSG16.17 VK2 (SEQ ID NO: 19). Some such antibodies include the the three Kabat CDRs (SEQ ID NOs: 60-62) of hSG16.17 VH3 (SEQ ID NO: 13) and three Kabat CDRs (SEQ ID NOs: 90-92) of hSG16.17 VK2 (SEQ ID NO: 19) provided that position H58 can be occupied by N or K, position H60 can be occupied by A or N, position H61 can be occupied by Q or E, position H62 can be occupied by K or N, position H64 can be occupied by Q or K, position H65 can be occupied by G or T , position L24 can be occupied by R or L and position L53 can be occupied by S or R. Preferably positions H58, H60, H61, H62, H64 and H65 are occupied by N, A, Q, K, Q and G respectively and L24 and L53 are occupied by R and S respectively. These recited residues represent amino acids from a human acceptor sequence occupying positions within the Kabat CDRs. Some antibodies have at least 1, 2, 3, 4, 5, 6, 7 or 8 rat residues in the human Kabat CDRs replaced with corresponding residues from a human acceptor sequence. In some antibodies positions H58, H60, H61, H62, H64 and H65 are occupied by N, A, Q, K, Q and G respectively and L24 and L53 are occupied by R and S respectively. Some antibodies include at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13 or 14 backmutations representing replacement of variable region human acceptor sequence residues with corrsponding rat residues.

[0079] In some antibodies at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or 11 of positions H20, H48, H69, H71, H73, H76, H80, H88, H91 and H93 are occupied by L, I, M, A, K, N, V, A, F, and T respectively. In some antibodies at least 1, 2 or 3 of positions L46, L48 and L87 are occupied by V, V and F respectively. In some antibodies, each of positions H20, H48, H69, H71, H73, H76, H80, H88, H91 and H93 are occupied by L, I, M, A, K, N, V, A, F, and T respectively and each of L46, L48 and L87 are occupied by V, V and F respectively.

[0080] Insofar as humanized antibodies show any variation from the exemplified hSG16.17 VH3 VK2 humanized antibody, one possibility for such additional variation is additional backmutations in the variable region frameworks. Any or all of the positions backmutated in other exemplified humanized heavy or light chain mature variable regions can also be made (i.e., 1, 2, 3, 4, 5 or all 6) of H8 occupied by R, H67 occupied by A and H78 occupied by A, L40 occupied by S, L78 occupied by M and L85 occupied by D, or all 5 of H38 occupied by N, H40 occupied by R, H73 occupied by K, H82A occupied by S, and H83 occupied by T in the heavy chain and 1 or both of L3 occupied by K, and L20 occupied by I in the light chain. However, such additional backmutations are not preferred because they in general do not improve affinity and introducing more mouse residues may give increased risk of immunogenicity.

[0081] Another possible variation is to substitute more or fewer residues in the CDRs of the mouse antibody with corresponding residues from human CDRs sequences, typically from the CDRs of the human acceptor sequences used in designing the exemplified humanized antibodies. In some antibodies only part of the CDRs, namely the subset of CDR residues required for binding, termed the SDRs, are needed to retain binding in a humanized antibody. CDR residues not contacting antigen and not in the SDRs can be identified based on regions of Kabat CDRs lying outside CDRs according to other definitions, such as Chothia hypervariable loops (Chothia, *J. Mol. Biol.* 196:901, 1987), by molecular modeling and/or empirically, or as described in Gonzales et al., *Mol. Immunol.* 41: 863 (2004). In such humanized antibodies at positions in which one or more donor CDR residues is absent or in which an entire donor CDR is omitted, the amino acid occupying the position can be an amino acid occupying the corresponding position (by Kabat numbering) in the acceptor antibody sequence. The number of such substitutions of acceptor for donor amino acids in the CDRs to include reflects a balance

of competing considerations. Such substitutions are potentially advantageous in decreasing the number of mouse amino acids in a humanized antibody and consequently decreasing potential immunogenicity. However, substitutions can also cause changes of affinity, and significant reductions in affinity are preferably avoided. Positions for substitution within CDRs and amino acids to substitute can also be selected empirically.

[0082] Although not preferred other amino acid substitutions can be made, for example, in framework residues not in contact with the CDRs, or even some potential CDR-contact residues amino acids within the CDRs. Often the replacements made in the variant humanized sequences are conservative with respect to the replaced hSG16.17 VH3 VK2 amino. Preferably, replacements relative to hSG16.17 VH3 VK2 (whether or not conservative) have no substantial effect on the binding affinity or potency of the humanized mAb, that is, its ability to bind human BCMA and inhibit growth of cancer cells.

[0083] Variants typically differ from the heavy and light chain mature variable region sequences of hSG16.17 VH3 VK2 by a small number (e.g., typically no more than 1, 2, 3, 5 or 10 in either the light chain or heavy chain mature variable region, or both) of replacements, deletions or insertions.

[0084] Other preferred combinations of humanized heavy and light chains include any of hSG16.17 VH1 VK2, VH1 VK3, VH1 VK4, VH1 VK4, VH3 VK2, VH3 VK3, VH3 VK4, and VH3 VK5, and VH5 VK2, VH5 VK3, VH5 VK4, VH5 VK5, as well as humanized antibodies in which the heavy and light chain variable regions show at least 90, 95, 96, 97, 98, or 99% identity with the heavy and light chain variable regions of any of these antibodies.

[0085] The invention provides humanized forms of the rat SG16.45 antibody including six exemplified humanized heavy chain mature variable regions (hSG16.45 VH1-6) (SEQ ID NOs: 27-32) and four exemplified humanized light chain mature variable regions (hSG16.45 VK1, 2, 3, and 5) (SEQ ID NOs: 35-38). The heavy and light chains can be combined in any permutations, with permutations hSG16.45 VH5 VK2, VH1 VK1 and VH1 VK5 being preferred. hSG16.45 HV5 VK2 shows greater than 85% sequence identity with human germline in both heavy and light chain variable regions (thus, qualifying for “humanized” designation under the new INN guideliness), high expression in CHO cells, a high proportion of monomers and adequate

binding albeit slightly less than that of rat or chimeric SG16.45. hSG16.45 VH5 VK2 has 3 variable region backmutations (all in the heavy chain) and 3 Kabat CDR forward mutations, in which a rat residue in the Kabat CDRs is changed to the corresponding residue in the human acceptor sequence, such that overall the antibody has sufficient sequence identity to human germline sequences to be classified as humanized under INN guidelines.

[0086] The invention provides antibodies in which the heavy chain variable region shows at least 90% identity to hSG16.45 VH5 (SEQ ID NO: 31) and a light chain variable region at least 90% identical to hSG16.45 VK2. Some antibodies show at least 95%, 96%, 97%, 98% or 99% sequence identity to hSG16.45 VH5 and at least 95%, 96%, 97%, 98% or 99% sequence identity to VK2. Some such antibodies include the three Kabat CDRs (SEQ ID NOs: 152-154) of hSG16.45 VH5 (SEQ ID NO: 31) and three Kabat CDRs (SEQ ID NOs: 179-181) of hSG16.45 VK2 (SEQ ID NO: 36). Some such antibodies include the three Kabat CDRs (SEQ ID NOs: 152-154) of hSG16.45 VH5 (SEQ ID NO: 31) and three Kabat CDRs (SEQ ID NOs: 179-181) of hSG16.45 VK2 (SEQ ID NO: 36) provided that position H50 can be occupied by A or S and position L24 can be occupied by R or L and position L26 can be occupied by S or T. Preferably positions H50 is occupied by A and positions L24 and L26 are occupied by R and S. These recited residues represent amino acids from a human acceptor sequence occupying positions within the Kabat CDRs. Some antibodies have at least 1, 2, or 3 rat residues in the human Kabat CDRs replaced with corresponding residues from a human acceptor sequence. In some antibodies positions H50, L24 and L26 are occupied by A, R and S respectively. Some antibodies include at least 1, 2, or 3 backmutations representing replacement of variable region human acceptor sequence residues with corresponding rat residues.

[0087] In some antibodies at least 1, 2, or 3, of positions H30, H93 and H94 are occupied by N, T and S respectively. In some antibodies, each of positions H30, H93 and H94 are occupied by N, T and S respectively

[0088] Insofar as humanized antibodies show any variation from the exemplified hSG16.45 VH5 VK2 humanized antibody, one possibility for such additional variation is additional backmutations in the variable region frameworks. Any or all of the positions backmutated in other exemplified humanized heavy or light chain mature variable regions can also be made

(i.e., 1, 2, 3, or 4) of H37, H48, H76, H107 occupied by I, I, N, and V respectively and/or 1, 2, 3, 4, 5, 6 or 7 of L14, L19, L21, L38, L58, L71 and L78 occupied by A, V, I, H, V, Y, and M respectively. However, such additional backmutations are not preferred because they in general do not improve affinity and introducing more mouse residues may give increased risk of immunogenicity.

[0089] Another possible variation is to substitute more or fewer residues in the CDRs of the mouse antibody with corresponding residues from human CDRs sequences, typically from the CDRs of the human acceptor sequences used in designing the exemplified humanized antibodies. In some antibodies only part of the CDRs, namely the subset of CDR residues required for binding, termed the SDRs, are needed to retain binding in a humanized antibody. CDR residues not contacting antigen and not in the SDRs can be identified based on regions of Kabat CDRs lying outside CDRs according to other definitions, such as Chothia hypervariable loops (Chothia, *J. Mol. Biol.* 196:901, 1987), by molecular modeling and/or empirically, or as described in Gonzales et al., *Mol. Immunol.* 41: 863 (2004). In such humanized antibodies at positions in which one or more donor CDR residues is absent or in which an entire donor CDR is omitted, the amino acid occupying the position can be an amino acid occupying the corresponding position (by Kabat numbering) in the acceptor antibody sequence. The number of such substitutions of acceptor for donor amino acids in the CDRs to include reflects a balance of competing considerations. Such substitutions are potentially advantageous in decreasing the number of mouse amino acids in a humanized antibody and consequently decreasing potential immunogenicity. However, substitutions can also cause changes of affinity, and significant reductions in affinity are preferably avoided. Positions for substitution within CDRs and amino acids to substitute can also be selected empirically.

[0090] Although not preferred other amino acid substitutions can be made, for example, in framework residues not in contact with the CDRs, or even some potential CDR-contact residues amino acids within the CDRs. Often the replacements made in the variant humanized sequences are conservative with respect to the replaced hSG16.45 VH3 VK2. Preferably, replacements relative to hSG16.45 VH5 VK2 (whether or not conservative) have no substantial

effect on the binding affinity or potency of the humanized mAb, that is, its ability to bind human BCMA and inhibit growth of cancer cells.

[0091] Variants typically differ from the heavy and light chain mature variable region sequences of SG16.45 VH5 VK2 by a small number (e.g., typically no more than 1, 2, 3, 5 or 10 in either the light chain or heavy chain mature variable region, or both) of replacements, deletions or insertions.

[0092] Other preferred combinations of humanized heavy and light chains include any of hSG16.45 VH1 VK1 and VH1 VK5, as well as humanized antibodies in which the heavy and light chain variable regions show at least 90, 95, 96, 97, 98, or 99% identity with the heavy and light chain variable regions of any of these antibodies.

C. Selection of Constant Region

[0093] Heavy and light chain variable regions of humanized antibodies can be linked to at least a portion of a human constant region. The choice of constant region depends, in part, whether antibody-dependent cell-mediated cytotoxicity, antibody dependent cellular phagocytosis and/or complement dependent cytotoxicity are desired. For example, human isotopes IgG1 and IgG3 have strong complement-dependent cytotoxicity, human isotype IgG2 weak complement-dependent cytotoxicity and human IgG4 lacks complement-dependent cytotoxicity. Human IgG1 and IgG3 also induce stronger cell mediated effector functions than human IgG2 and IgG4. Light chain constant regions can be lambda or kappa. Antibodies can be expressed as tetramers containing two light and two heavy chains, as separate heavy chains, light chains, as Fab, Fab', F(ab')2, and Fv, or as single chain antibodies in which heavy and light chain variable domains are linked through a spacer.

[0094] Human constant regions show allotypic variation and isoallotypic variation between different individuals, that is, the constant regions can differ in different individuals at one or more polymorphic positions. Isoallotypes differ from allotypes in that sera recognizing an isoallotype binds to a non-polymorphic region of a one or more other isotypes. Exemplary wild type human kappa and IgG1 constant region sequences (the latter with or without the C-terminal lysine) are provided in SEQ ID NOS: 3-5.

[0095] One or several amino acids at the amino or carboxy terminus of the light and/or heavy chain, such as the C-terminal lysine of the heavy chain, may be missing or derivatized in a proportion or all of the molecules. Substitutions can be made in the constant regions to reduce or increase effector function such as complement-mediated cytotoxicity or ADCC (see, e.g., Winter et al., US Patent No. 5,624,821; Tso et al., US Patent No. 5,834,597; and Lazar et al., Proc. Natl. Acad. Sci. USA 103:4005, 2006), or to prolong half-life in humans (see, e.g., Hinton et al., J. Biol. Chem. 279:6213, 2004).

[0096] Exemplary substitution include the amino acid substitution of the native amino acid to a cysteine residue is introduced at amino acid position 234, 235, 237, 239, 267, 298, 299, 326, 330, or 332, preferably an S239C mutation in a human IgG1 isotype (numbering is according to the EU index (Kabat, *Sequences of Proteins of Immunological Interest* (National Institutes of Health, Bethesda, MD, 1987 and 1991); see US 20100158909, which is herein incorporated reference). Sequences of a heavy chain constant regions with S239C with and without a C-terminal lysine are provided by SEQ ID NOS: 6 and 7. The presence of an additional cysteine residue allows interchain disulfide bond formation. Such interchain disulfide bond formation can cause steric hindrance, thereby reducing the affinity of the Fc region-Fc_YR binding interaction. The cysteine residue(s) introduced in or in proximity to the Fc region of an IgG constant region can also serve as sites for conjugation to therapeutic agents (i.e., coupling cytotoxic drugs using thiol specific reagents such as maleimide derivatives of drugs. The presence of a therapeutic agent causes steric hindrance, thereby further reducing the affinity of the Fc region-Fc_YR binding interaction. Other substitutions at any of positions 234, 235, 236 and/or 237 reduce affinity for Fc_Y receptors, particularly Fc_YRI receptor (see, e.g., US 6,624,821, US 5,624,821.) A preferred combination of mutations is S239D, A330L and I332E, which increases the affinity of the Fc domain for Fc_YRIIA and consequently increases ADCC.

[0097] The in vivo half-life of an antibody can also impact its effector functions. The half-life of an antibody can be increased or decreased to modify its therapeutic activities. FcRn is a receptor that is structurally similar to MHC Class I antigen that non-covalently associates with β 2-microglobulin. FcRn regulates the catabolism of IgGs and their transcytosis across tissues (Ghetie and Ward, 2000, *Annu. Rev. Immunol.* 18:739-766; Ghetie and Ward, 2002, *Immunol.*

Res. 25:97-113). The IgG-FcRn interaction takes place at pH 6.0 (pH of intracellular vesicles) but not at pH 7.4 (pH of blood); this interaction enables IgGs to be recycled back to the circulation (Ghetie and Ward, 2000, *Ann. Rev. Immunol.* 18:739-766; Ghetie and Ward, 2002, *Immunol. Res.* 25:97-113). The region on human IgG1 involved in FcRn binding has been mapped (Shields *et al.*, 2001, *J. Biol. Chem.* 276:6591-604). Alanine substitutions at positions Pro238, Thr256, Thr307, Gln311, Asp312, Glu380, Glu382, or Asn434 of human IgG1 enhance FcRn binding (Shields *et al.*, 2001, *J. Biol. Chem.* 276:6591-604). IgG1 molecules harboring these substitutions have longer serum half-lives. Consequently, these modified IgG1 molecules may be able to carry out their effector functions, and hence exert their therapeutic efficacies, over a longer period of time compared to unmodified IgG1. Other exemplary substitutions for increasing binding to FcRn include a Gln at position 250 and/or a Leu at position 428. EU numbering is used for all positions in the constant region.

[0098] Oligosaccharides covalently attached to the conserved Asn297 are involved in the ability of the Fc region of an IgG to bind Fc γ R (Lund *et al.*, 1996, *J. Immunol.* 157:4963-69; Wright and Morrison, 1997, *Trends Biotechnol.* 15:26-31). Engineering of this glycoform on IgG can significantly improve IgG-mediated ADCC. Addition of bisecting N-acetylglucosamine modifications (Umana *et al.*, 1999, *Nat. Biotechnol.* 17:176-180; Davies *et al.*, 2001, *Biotech. Bioeng.* 74:288-94) to this glycoform or removal of fucose (Shields *et al.*, 2002, *J. Biol. Chem.* 277:26733-40; Shinkawa *et al.*, 2003, *J. Biol. Chem.* 278:6591-604; Niwa *et al.*, 2004, *Cancer Res.* 64:2127-33) from this glycoform are two examples of IgG Fc engineering that improves the binding between IgG Fc and Fc γ R, thereby enhancing Ig-mediated ADCC activity.

[0099] A systematic substitution of solvent-exposed amino acids of human IgG1 Fc region has generated IgG variants with altered Fc γ R binding affinities (Shields *et al.*, 2001, *J. Biol. Chem.* 276:6591-604). When compared to parental IgG1, a subset of these variants involving substitutions at Thr256/Ser298, Ser298/Glu333, Ser298/Lys334, or Ser298/Glu333/Lys334 to Ala demonstrate increased in both binding affinity toward Fc γ R and ADCC activity (Shields *et al.*, 2001, *J. Biol. Chem.* 276:6591-604; Okazaki *et al.*, 2004, *J. Mol. Biol.* 336:1239-49).

[00100] Complement fixation activity of antibodies (both C1q binding and CDC activity) can be improved by substitutions at Lys326 and Glu333 (Idusogie *et al.*, 2001, *J. Immunol.* 166:2571-

2575). The same substitutions on a human IgG2 backbone can convert an antibody isotype that binds poorly to C1q and is severely deficient in complement activation activity to one that can both bind C1q and mediate CDC (Idusogie *et al.*, 2001, *J. Immunol.* 166:2571-75). Several other methods have also been applied to improve complement fixation activity of antibodies. For example, the grafting of an 18-amino acid carboxyl-terminal tail piece of IgM to the carboxyl-termini of IgG greatly enhances their CDC activity. This is observed even with IgG4, which normally has no detectable CDC activity (Smith *et al.*, 1995, *J. Immunol.* 154:2226-36). Also, substituting Ser444 located close to the carboxy-terminal of IgG1 heavy chain with Cys induced tail-to-tail dimerization of IgG1 with a 200-fold increase of CDC activity over monomeric IgG1 (Shopes *et al.*, 1992, *J. Immunol.* 148:2918-22). In addition, a bispecific diabody construct with specificity for C1q also confers CDC activity (Kontermann *et al.*, 1997, *Nat. Biotech.* 15:629-31).

[00101] Complement activity can be reduced by mutating at least one of the amino acid residues 318, 320, and 322 of the heavy chain to a residue having a different side chain, such as Ala. Other alkyl-substituted non-ionic residues, such as Gly, Ile, Leu, or Val, or such aromatic non-polar residues as Phe, Tyr, Trp and Pro in place of any one of the three residues also reduce or abolish C1q binding. Ser, Thr, Cys, and Met can be used at residues 320 and 322, but not 318, to reduce or abolish C1q binding activity. Replacement of the 318 (Glu) residue by a polar residue may modify but not abolish C1q binding activity. Replacing residue 297 (Asn) with Ala results in removal of lytic activity but only slightly reduces (about three fold weaker) affinity for C1q. This alteration destroys the glycosylation site and the presence of carbohydrate that is required for complement activation. Any other substitution at this site also destroys the glycosylation site. The following mutations and any combination thereof also reduce C1q binding: D270A, K322A, P329A, and P311S (see WO 06/036291).

[00102] Reference to a human constant region includes a constant region with any natural allotype or any permutation of residues occupying polymorphic positions in natural allotypes. Also, up to 1, 2, 5, or 10 mutations may be present relative to a natural human constant region, such as those indicated above to reduce Fcγ receptor binding or increase binding to FcRN.

D. Expression of Recombinant Antibodies

[00103] Humanized, chimeric or veneered antibodies are typically produced by recombinant expression. Recombinant polynucleotide constructs typically include an expression control sequence operably linked to the coding sequences of antibody chains, including naturally-associated or heterologous promoter regions. Preferably, the expression control sequences are eukaryotic promoter systems in vectors capable of transforming or transfecting eukaryotic host cells. Once the vector has been incorporated into the appropriate host, the host is maintained under conditions suitable for high level expression of the nucleotide sequences, and the collection and purification of the crossreacting antibodies.

[00104] Mammalian cells are a preferred host for expressing nucleotide segments encoding immunoglobulins or fragments thereof. See Winnacker, *From Genes to Clones*, (VCH Publishers, NY, 1987). A number of suitable host cell lines capable of secreting intact heterologous proteins have been developed in the art, and include CHO cell lines (e.g., DG44), various COS cell lines, HeLa cells, HEK293 cells, L cells, and non-antibody-producing myelomas including Sp2/0 and NS0. Preferably, the cells are nonhuman. Expression vectors for these cells can include expression control sequences, such as an origin of replication, a promoter, an enhancer (Queen et al., *Immunol. Rev.* 89:49 (1986)), and necessary processing information sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites, and transcriptional terminator sequences. Preferred expression control sequences are promoters derived from endogenous genes, cytomegalovirus, SV40, adenovirus, bovine papillomavirus, and the like. See Co et al., *J. Immunol.* 148:1149 (1992).

[00105] Once expressed, antibodies can be purified according to standard procedures of the art, including HPLC purification, column chromatography, gel electrophoresis and the like (see generally, Scopes, *Protein Purification* (Springer-Verlag, NY, 1982)).

E. Glycosylation Variants

[00106] Antibodies may be glycosylated at conserved positions in their constant regions (Jefferis and Lund, (1997) *Chem. Immunol.* 65:111-128; Wright and Morrison, (1997) *TibTECH* 15:26-32). The oligosaccharide side chains of the immunoglobulins affect the protein's function (Boyd et al., (1996) *Mol. Immunol.* 32:1311-1318; Wittwe and Howard, (1990) *Biochem.*

29:4175-4180), and the intramolecular interaction between portions of the glycoprotein which can affect the conformation and presented three-dimensional surface of the glycoprotein (Hefferis and Lund, *supra*; Wyss and Wagner, (1996) Current Opin. Biotech. 7:409-416). Oligosaccharides may also serve to target a given glycoprotein to certain molecules based upon specific recognition structures. For example, it has been reported that in agalactosylated IgG, the oligosaccharide moiety 'flips' out of the inter-CH2 space and terminal N-acetylglucosamine residues become available to bind mannose binding protein (Malhotra *et al.*, (1995) Nature Med. 1:237-243). Removal by glycopeptidase of the oligosaccharides from CAMPATH-1H (a recombinant humanized murine monoclonal IgG1 antibody which recognizes the CDw52 antigen of human lymphocytes) produced in Chinese Hamster Ovary (CHO) cells resulted in a complete reduction in complement mediated lysis (CMCL) (Boyd *et al.*, (1996) Mol. Immunol. 32:1311-1318), while selective removal of sialic acid residues using neuraminidase resulted in no loss of DMCL. Glycosylation of antibodies has also been reported to affect antibody-dependent cellular cytotoxicity (ADCC). In particular, CHO cells with tetracycline-regulated expression of β (1,4)-N-acetylglucosaminyltransferase III (GnTIII), a glycosyltransferase catalyzing formation of bisecting GlcNAc, was reported to have improved ADCC activity (Umana *et al.* (1999) Mature Biotech. 17:176-180).

[00107] Glycosylation of antibodies is typically either N-linked or O-linked. N-linked refers to the attachment of the carbohydrate moiety to the side chain of an asparagine residue. The tripeptide sequences asparagine-X-serine and asparagine-X-threonine, where X is any amino acid except proline, are the recognition sequences for enzymatic attachment of the carbohydrate moiety to the asparagine side chain. Thus, the presence of either of these tripeptide sequences in a polypeptide creates a potential glycosylation site. O-linked glycosylation refers to the attachment of one of the sugars N-acetylgalactosamine, galactose, or xylose to a hydroxyamino acid, most commonly serine or threonine, although 5-hydroxyproline or 5-hydroxylysine may also be used.

[00108] Glycosylation variants of antibodies are variants in which the glycosylation pattern of an antibody is altered. By altering is meant deleting one or more carbohydrate moieties found

in the antibody, adding one or more carbohydrate moieties to the antibody, changing the composition of glycosylation (glycosylation pattern), the extent of glycosylation, etc.

[00109] Addition of glycosylation sites to the antibody can be accomplished by altering the amino acid sequence such that it contains one or more of the above-described tripeptide sequences (for N-linked glycosylation sites). The alteration may also be made by the addition of, or substitution by, one or more serine or threonine residues to the sequence of the original antibody (for O-linked glycosylation sites). Similarly, removal of glycosylation sites can be accomplished by amino acid alteration within the native glycosylation sites of the antibody.

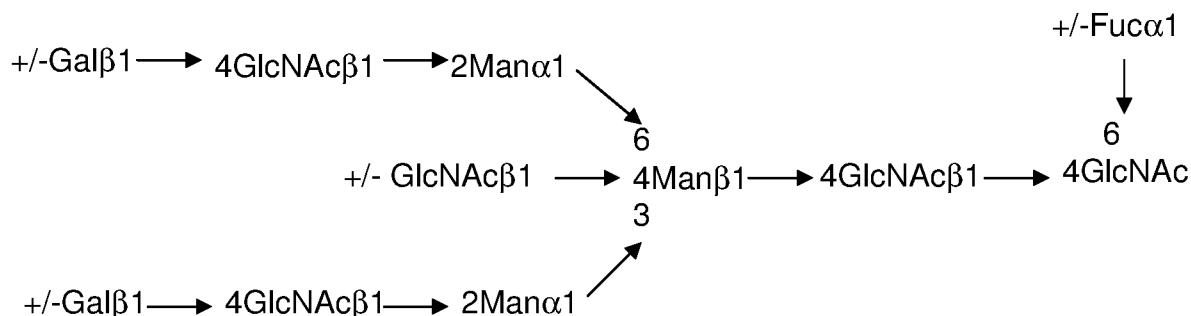
[00110] The amino acid sequence is usually altered by altering the underlying nucleic acid sequence. These methods include isolation from a natural source (in the case of naturally-occurring amino acid sequence variants) or preparation by oligonucleotide-mediated (or site-directed) mutagenesis, PCR mutagenesis, and cassette mutagenesis of an earlier prepared variant or a non-variant version of the antibody.

[00111] The glycosylation (including glycosylation pattern) of antibodies may also be altered without altering the amino acid sequence or the underlying nucleotide sequence. Glycosylation largely depends on the host cell used to express the antibody. Since the cell type used for expression of recombinant glycoproteins, *e.g.*, antibodies, as potential therapeutics is rarely the native cell, significant variations in the glycosylation pattern of the antibodies can be expected. See, *e.g.*, Hse *et al.*, (1997) *J. Biol. Chem.* 272:9062-9070. In addition to the choice of host cells, factors which affect glycosylation during recombinant production of antibodies include growth mode, media formulation, culture density, oxygenation, pH, purification schemes and the like. Various methods have been proposed to alter the glycosylation pattern achieved in a particular host organism including introducing or overexpressing certain enzymes involved in oligosaccharide production (U.S. Patent Nos. 5047335; 5510261; 5278299). Glycosylation, or certain types of glycosylation, can be enzymatically removed from the glycoprotein, for example using endoglycosidase H (Endo H). In addition, the recombinant host cell can be genetically engineered, *e.g.*, make defective in processing certain types of polysaccharides. These and similar techniques are well known in the art.

[00112] The glycosylation structure of antibodies can be readily analyzed by conventional techniques of carbohydrate analysis, including lectin chromatography, NMR, Mass spectrometry, HPLC, GPC, monosaccharide compositional analysis, sequential enzymatic digestion, and HPAEC-PAD, which uses high pH anion exchange chromatography to separate oligosaccharides based on charge. Methods for releasing oligosaccharides for analytical purposes are also known, and include, without limitation, enzymatic treatment (commonly performed using peptide-N-glycosidase F/endo- β -galactosidase), elimination using harsh alkaline environment to release mainly O-linked structures, and chemical methods using anhydrous hydrazine to release both N- and O-linked oligosaccharides

[00113] A preferred form of modification of glycosylation of antibodies is reduced core fucosylation. “Core fucosylation” refers to addition of fucose (“fucosylation”) to N-acetylglucosamine (“GlcNAc”) at the reducing terminal of an N-linked glycan.

[00114] A “complex N-glycoside-linked sugar chain” is typically bound to asparagine 297 (according to the number of Kabat). As used herein, the complex N-glycoside-linked sugar chain has a biantennary composite sugar chain, mainly having the following structure:



where \pm indicates the sugar molecule can be present or absent, and the numbers indicate the position of linkages between the sugar molecules. In the above structure, the sugar chain terminal which binds to asparagine is called a reducing terminal (at right), and the opposite side is called a non-reducing terminal. Fucose is usually bound to N-acetylglucosamine (“GlcNAc”) of the reducing terminal, typically by an α 1,6 bond (the 6-position of GlcNAc is linked to the 1-position of fucose). “Gal” refers to galactose, and “Man” refers to mannose.

[00115] A “complex N-glycoside-linked sugar chain” includes 1) a complex type, in which the non-reducing terminal side of the core structure has one or more branches of galactose-N-

acetylglucosamine (also referred to as “gal-GlcNAc”) and the non-reducing terminal side of Gal-GlcNAc optionally has a sialic acid, bisecting N-acetylglucosamine or the like; or 2) a hybrid type, in which the non-reducing terminal side of the core structure has both branches of a high mannose N-glycoside-linked sugar chain and complex N-glycoside-linked sugar chain.

[00116] In some embodiments, the “complex N-glycoside-linked sugar chain” includes a complex type in which the non-reducing terminal side of the core structure has zero, one or more branches of galactose-N-acetylglucosamine (also referred to as “gal-GlcNAc”) and the non-reducing terminal side of Gal-GlcNAc optionally further has a structure such as a sialic acid, bisecting N-acetylglucosamine or the like.

[00117] According to the present methods, typically only a minor amount of fucose is incorporated into the complex N-glycoside-linked sugar chain(s) of humanized, chimeric or veneered SG16.17 or SG16.45 antibodies. For example, in various embodiments, less than about 60%, less than about 50%, less than about 40%, less than about 30%, less than about 20%, less than about 15%, less than about 10%, less than about 5%, or less than about 3% of the molecules of an antibody have core fucosylation by fucose. In some embodiments, about 2% of the molecules of the antibody has core fucosylation by fucose.

[00118] In certain embodiments, only a minor amount of a fucose analog (or a metabolite or product of the fucose analog) is incorporated into the complex N-glycoside-linked sugar chain(s). For example, in various embodiments, less than about 60%, less than about 50%, less than about 40%, less than about 30%, less than about 20%, less than about 15%, less than about 10%, less than about 5%, or less than about 3% of humanized, chimeric or veneered SG16.17 or SG16.45 antibodies have core fucosylation by a fucose analog or a metabolite or product of the fucose analog. In some embodiments, about 2% of humanized, chimeric or veneered SG16.17 antibodies have core fucosylation by a fucose analog or a metabolite or product of the fucose analog.

[00119] Methods of making non-fucosylated antibodies by incubating antibody-producing cells with a fucose analogue are described, e.g., in WO2009/135181. Briefly, cells that have been engineered to express humanized, chimeric or veneered SG16.17 antibodies antibody are incubated in the presence of a fucose analogue or an intracellular metabolite or product of the

fucose analog. An intracellular metabolite can be, for example, a GDP-modified analog or a fully or partially de-esterified analog. A product can be, for example, a fully or partially de-esterified analog. In some embodiments, a fucose analogue can inhibit an enzyme(s) in the fucose salvage pathway. For example, a fucose analog (or an intracellular metabolite or product of the fucose analog) can inhibit the activity of fucokinase, or GDP-fucose-pyrophosphorylase. In some embodiments, a fucose analog (or an intracellular metabolite or product of the fucose analog) inhibits fucosyltransferase (preferably a 1,6-fucosyltransferase, *e.g.*, the FUT8 protein). In some embodiments, a fucose analog (or an intracellular metabolite or product of the fucose analog) can inhibit the activity of an enzyme in the *de novo* synthetic pathway for fucose. For example, a fucose analog (or an intracellular metabolite or product of the fucose analog) can inhibit the activity of GDP-mannose 4,6-dehydratase or/or GDP-fucose synthetase. In some embodiments, the fucose analog (or an intracellular metabolite or product of the fucose analog) can inhibit a fucose transporter (*e.g.*, GDP-fucose transporter).

[00120] In one embodiment, the fucose analogue is 2-fluorofucose. Methods of using fucose analogues in growth medium and other fucose analogues are disclosed, *e.g.*, in WO/2009/135181, which is herein incorporated by reference.

[00121] Other methods for engineering cell lines to reduce core fucosylation included gene knock-outs, gene knock-ins and RNA interference (RNAi). In gene knock-outs, the gene encoding FUT8 (alpha 1,6- fucosyltransferase enzyme) is inactivated. FUT8 catalyzes the transfer of a fucosyl residue from GDP-fucose to position 6 of Asn-linked (N-linked) GlcNac of an N-glycan. FUT8 is reported to be the only enzyme responsible for adding fucose to the N-linked biantennary carbohydrate at Asn297. Gene knock-ins add genes encoding enzymes such as GNTIII or a golgi alpha mannosidase II. An increase in the levels of such enzymes in cells diverts monoclonal antibodies from the fucosylation pathway (leading to decreased core fucosylation), and having increased amount of bisecting N-acetylglucosamines. RNAi typically also targets FUT8 gene expression, leading to decreased mRNA transcript levels or knocking out gene expression entirely. Any of these methods can be used to generate a cell line that would be able to produce a non-fucosylated antibody, *e.g.*, a humanized, chimeric or veneered SG16.17 antibody.

[00122] Many methods are available to determine the amount of fucosylation on an antibody. Methods include, e.g., LC-MS via PLRP-S chromatography and electrospray ionization quadrupole TOF MS.

IV. Nucleic Acids

[00123] The invention further provides nucleic acids encoding any of the humanized heavy and light chains described above. Typically, the nucleic acids also encode a signal peptide fused to the mature heavy and light chains. Coding sequences on nucleic acids can be in operable linkage with regulatory sequences to ensure expression of the coding sequences, such as a promoter, enhancer, ribosome binding site, transcription termination signal and the like. The nucleic acids encoding heavy and light chains can occur in isolated form or can be cloned into one or more vectors. The nucleic acids can be synthesized by for example, solid state synthesis or PCR of overlapping oligonucleotides. Nucleic acids encoding heavy and light chains can be joined as one contiguous nucleic acid, e.g., within an expression vector, or can be separate, e.g., each cloned into its own expression vector.

V. Antibody Drug Conjugates

[00124] Anti-MCMA antibodies can be conjugated to cytotoxic moieties to form antibody-drug conjugates (ADCs). Particularly suitable moieties for conjugation to antibodies are cytotoxic agents (e.g., chemotherapeutic agents), prodrug converting enzymes, radioactive isotopes or compounds, or toxins (these moieties being collectively referred to as therapeutic agents or drugs). For example, an anti-BCMA antibody can be conjugated to a cytotoxic agent such as a chemotherapeutic agent, or a toxin (e.g., a cytostatic or cytocidal agent such as, e.g., abrin, ricin A, *pseudomonas* exotoxin, or diphtheria toxin). Examples of useful classes of cytotoxic agents include, for example, DNA minor groove binders, DNA alkylating agents, and tubulin inhibitors. Exemplary cytotoxic agents include, for example, auristatins, camptothecins, duocarmycins, etoposides, maytansines and maytansinoids (e.g., DM1 and DM4), taxanes, benzodiazepines (e.g., pyrrolo[1,4]benzodiazepines (PBDs), indolinobenzodiazepines, and oxazolidinobenzodiazepines) and vinca alkaloids. Techniques for conjugating therapeutic

agents to proteins, and in particular to antibodies, are well-known. (See, e.g., Alley et al., *Current Opinion in Chemical Biology* 2010 14:1-9; Senter, *Cancer J.*, 2008, 14(3):154-169.)

[00125] The therapeutic agent (e.g., cytotoxic agent) can be conjugated to the antibody in a manner that reduces its activity unless it is detached from the antibody (e.g., by hydrolysis, by antibody degradation, or by a cleaving agent). Such therapeutic agent can be attached to the antibody via a linker. A therapeutic agent conjugated to a linker is also referred to herein as a drug linker. The nature of the linker can vary widely. The components that make up the linker are chosen on the basis of their characteristics, which may be dictated in part, by the conditions at the site to which the conjugate is delivered.

[00126] The therapeutic agent can be attached to the antibody with a cleavable linker that is sensitive to cleavage in the intracellular environment of the anti-BCMA-expressing cancer cell but is not substantially sensitive to the extracellular environment, such that the conjugate is cleaved from the antibody when it is internalized by the anti-BCMA-expressing cancer cell (e.g., in the endosomal or, for example by virtue of pH sensitivity or protease sensitivity, in the lysosomal environment or in the caveolar environment). The therapeutic agent can also be attached to the antibody with a non-cleavable linker.

[00127] As indicated, the linker may comprise a cleavable unit. In some such embodiments, the structure and/or sequence of the cleavable unit is selected such that it is cleaved by the action of enzymes present at the target site (e.g., the target cell). In other embodiments, cleavable units that are cleavable by changes in pH (e.g. acid or base labile), temperature or upon irradiation (e.g. photolabile) may also be used.

[00128] In some embodiments, the cleavable unit may comprise one amino acid or a contiguous sequence of amino acids. The amino acid sequence may be the target substrate for an enzyme.

[00129] In some aspects, the cleavable unit is a peptidyl unit and is at least two amino acids long. Cleaving agents can include cathepsins B and D and plasmin (see, e.g., Dubowchik and Walker, 1999, *Pharm. Therapeutics* 83:67-123). Most typical are cleavable unit that are cleavable by enzymes that are present in anti-BCMA expressing cells, i.e., an enzyme cleavable linker. Accordingly, the linker can be cleaved by an intracellular peptidase or protease enzyme,

including a lysosomal or endosomal protease. For example, a linker that is cleavable by the thiol-dependent protease cathepsin-B, which is highly expressed in cancerous tissue, can be used (e.g., a linker comprising a Phe-Leu or a Val-Cit peptide or a Val-Ala peptide).

[00130] In some embodiments, the linker will comprise a cleavable unit (e.g., a peptidyl unit) and the cleavable unit will be directly conjugated to the therapeutic agent. In other embodiments, the cleavable unit will be conjugated to the therapeutic agent via an additional functional unit, e.g., a self-immolative spacer unit or a non-self-immolative spacer unit. A non self-immolative spacer unit is one in which part or all of the spacer unit remains bound to the drug unit after cleavage of a cleavable unit (e.g., amino acid) from the antibody drug conjugate. To liberate the drug, an independent hydrolysis reaction takes place within the target cell to cleave the spacer unit from the drug.

[00131] With a self-immolative spacer unit, the drug is released without the need for drug for a separate hydrolysis step. In one embodiment, wherein the linker comprises a cleavable unit and a self immolative group, the cleavable unit is cleavable by the action of an enzyme and after cleavage of the cleavable unit, the self-immolative group(s) release the therapeutic agent. In some embodiments, the cleavable unit of the linker will be directly or indirectly conjugated to the therapeutic agent on one end and on the other end will be directly or indirectly conjugated to the antibody. In some such embodiments, the cleavable unit will be directly or indirectly (e.g., via a self-immolative or non-self-immolative spacer unit) conjugated to the therapeutic agent on one end and on the other end will be conjugated to the antibody via a stretcher unit. A stretcher unit links the antibody to the rest of the drug and/or drug linker. In one embodiment, the connection between the antibody and the rest of the drug or drug linker is via a maleimide group, e.g., via a maleimidocaproyl linker. In some embodiments, the antibody will be linked to the drug via a disulfide, for example the disulfide linked maytansinoid conjugates SPDB-DM4 and SPP-DM1.

[00132] The connection between the antibody and the linker can be via a number of different routes, e.g., through a thioether bond, through a disulfide bond, through an amide bond, or through an ester bond. In one embodiment, the connection between the anti-BCMA antibody and the linker is formed between a thiol group of a cysteine residue of the antibody and a

maleimide group of the linker. In some embodiments, the interchain bonds of the antibody are converted to free thiol groups prior to reaction with the functional group of the linker. In some embodiments, a cysteine residue is introduced into the heavy or light chain of an antibody and reacted with the linker. Positions for cysteine insertion by substitution in antibody heavy or light chains include those described in Published U.S. Application No. 2007-0092940 and International Patent Publication WO2008070593, each of which are incorporated by reference herein in its entirety and for all purposes.

[00133] In some embodiments, the antibody- drug conjugates have the following formula I:



wherein L is an anti-BCMA antibody, LU is a Linker unit and D is a Drug unit (i.e., the therapeutic agent). The subscript p ranges from 1 to 20. Such conjugates comprise an anti-BCMA antibody covalently linked to at least one drug via a linker. The Linker Unit is connected at one end to the antibody and at the other end to the drug.

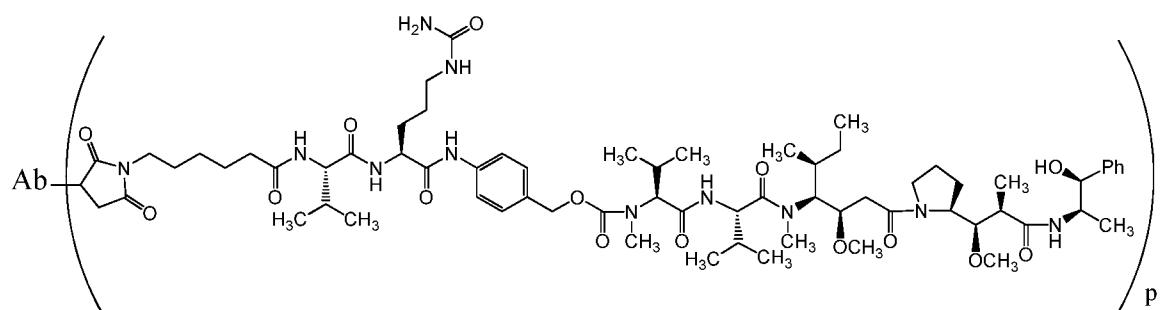
[00134] The drug loading is represented by p, the number of drug molecules per antibody. Drug loading may range from 1 to 20 Drug units (D) per antibody. In some aspects, the subscript p will range from 1 to 20 (i.e., both integer and non-integer values from 1 to 20). In some aspects, the subscript p will be an integer from 1 to 20, and will represent the number of drug-linkers on a singular antibody. In other aspects, p represents the average number of drug-linker molecules per antibody, e.g., the average number of drug-linkers per antibody in a reaction mixture or composition (e.g., pharmaceutical composition), and can be an integer or non-integer value. Accordingly, in some aspects, for compositions (e.g., pharmaceutical compositions), p represents the average drug loading of the antibody-drug conjugates in the composition, and p ranges from 1 to 20.

[00135] In some embodiments, p is from about 1 to about 8 drugs per antibody. In some embodiments, p is 1. In some embodiments, p is 2. In some embodiments, p is from about 2 to about 8 drugs per antibody. In some embodiments, p is from about 2 to about 6, 2 to about 5, or 2 to about 4 drugs per antibody. In some embodiments, p is about 2, about 4, about 6 or about 8 drugs per antibody.

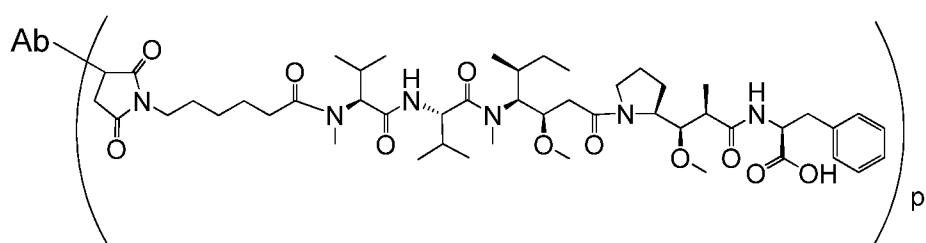
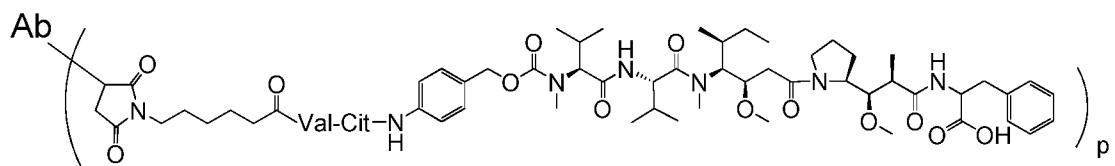
[00136] The average number of drugs per antibody unit in a preparation from a conjugation reaction may be characterized by conventional means such as mass spectroscopy, ELISA assay, HIC, and HPLC. The quantitative distribution of conjugates in terms of *p* may also be determined.

[00137] Exemplary antibody-drug conjugates include auristatin based antibody-drug conjugates, *i.e.*, conjugates wherein the drug component is an auristatin drug. Auristatins bind tubulin, have been shown to interfere with microtubule dynamics and nuclear and cellular division, and have anticancer activity. Typically the auristatin based antibody-drug conjugate comprises a linker between the auristatin drug and the anti-BCMA antibody. The auristatins can be linked to the anti-BCMA antibody at any position suitable for conjugation to a linker. The linker can be, for example, a cleavable linker (e.g., a peptidyl linker) or a non-cleavable linker (e.g., linker released by degradation of the antibody). The auristatin can be auristatin E or a derivative thereof. The auristatin can be, for example, an ester formed between auristatin E and a keto acid. For example, auristatin E can be reacted with paraacetyl benzoic acid or benzoylvaleric acid to produce AEB and AEVB, respectively. Other typical auristatins include MMAF (monomethyl auristatin F), and MMAE (monomethyl auristatin E). The synthesis and structure of exemplary auristatins are described in U.S. Publication Nos. 7,659,241, 7,498,298, 2009-0111756, 2009-0018086, and 7,968, 687 each of which is incorporated herein by reference in its entirety and for all purposes.

[00138] Exemplary auristatin based antibody-drug conjugates include vcMMAE, vcMMAF and mcMMAF antibody-drug conjugates as shown below wherein Ab is an antibody as described herein and val-cit represents the valine-citrulline dipeptide:



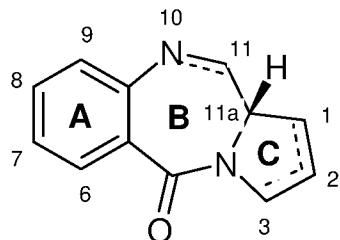
Ab-vcMMAE



or a pharmaceutically acceptable salt thereof. The drug loading is represented by p , the number of drug-linker molecules per antibody. Depending on the context, p can represent the average number of drug-linker molecules per antibody, also referred to the average drug loading. The variable p ranges from 1 to 20 and is preferably from 1 to 8. In some preferred embodiments, when p represents the average drug loading, p ranges from about 2 to about 5. In some embodiments, p is about 2, about 3, about 4, or about 5. In some aspects, the antibody is conjugated to the linker via a sulfur atom of a cysteine residue. In some aspects, the cysteine residue is one that is engineered into the antibody. In other aspects, the cysteine residue is an interchain disulfide cysteine residue.

[00139] Exemplary antibody-drug conjugates include PBD based antibody-drug conjugates; *i.e.*, antibody-drug conjugates wherein the drug component is a PBD drug.

[00140] PBDs are of the general structure:



[00141] They differ in the number, type and position of substituents, in both their aromatic A rings and pyrrolo C rings, and in the degree of saturation of the C ring. In the B-ring there is

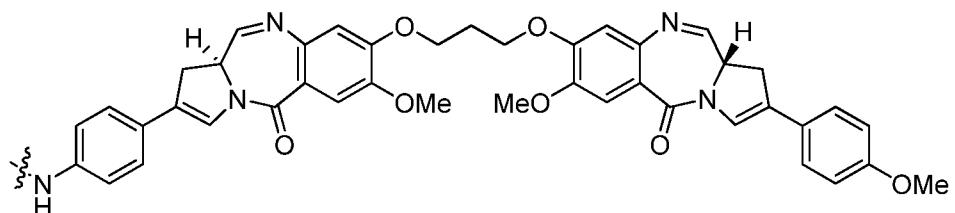
either an imine (N=C), a carbinolamine (NH-CH(OH)), or a carbinolamine methyl ether (NH-CH(OMe)) at the N10-C11 position, which is the electrophilic center responsible for alkylating DNA. All of the known natural products have an (S)-configuration at the chiral C11a position which provides them with a right-handed twist when viewed from the C ring towards the A ring. This gives them the appropriate three-dimensional shape for isohelicity with the minor groove of B-form DNA, leading to a snug fit at the binding site. The ability of PBDs to form an adduct in the minor groove enables them to interfere with DNA processing, hence their use as antitumor agents.

[00142] The biological activity of these molecules can be potentiated by joining two PBD units together through their C8/C'-hydroxyl functionalities via a flexible alkylene linker. The PBD dimers are thought to form sequence-selective DNA lesions such as the palindromic 5'-Pu-GATC-Py-3' interstrand cross-link, which is thought to be mainly responsible for their biological activity.

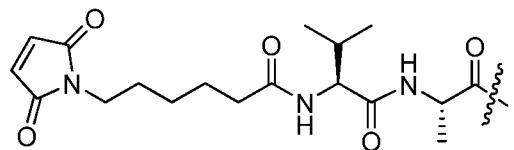
[00143] In some embodiments, PBD based antibody-drug conjugates comprise a PBD dimer linked to an anti-BCMA antibody. The monomers that form the PBD dimer can be the same or different, *i.e.*, symmetrical or unsymmetrical. The PBD dimer can be linked to the anti-BCMA antibody at any position suitable for conjugation to a linker. For example, in some embodiments, the PBD dimer will have a substituent at the C2 position that provides an anchor for linking the compound to the anti-BCMA antibody. In alternative embodiments, the N10 position of the PBD dimer will provide the anchor for linking the compound to the anti-BCMA antibody.

[00144] Typically the PBD based antibody-drug conjugate comprises a linker between the PBD drug and the anti-BCMA antibody. The linker may comprise a cleavable unit (*e.g.*, an amino acid or a contiguous sequence of amino acids that is a target substrate for an enzyme) or a non-cleavable linker (*e.g.*, linker released by degradation of the antibody). The linker may further comprise a maleimide group for linkage to the antibody, *e.g.*, maleimidocaproyl. The linker may, in some embodiments, further comprise a self-immolative group, such as, for example, a p-aminobenzyl alcohol (PAB) unit.

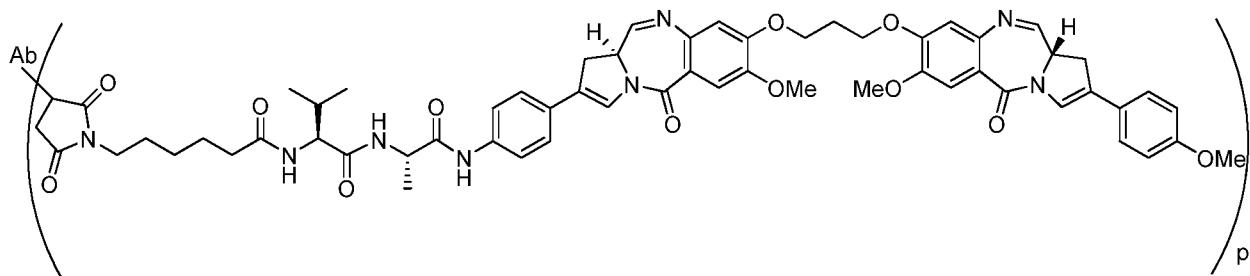
[00145] An exemplary PBD for use as a conjugate is described in International Application No. WO 2011/130613 and is as follows wherein the wavy line indicates the site of attachment to the linker:



or a pharmaceutically acceptable salt thereof. An exemplary linker is as follows wherein the wavy line indicates the site of attachment to the drug and the antibody is linked via the maleimide group.



[00146] Exemplary PBDs based antibody-drug conjugates include antibody-drug conjugates as shown below wherein Ab is an antibody as described herein:



or a pharmaceutically acceptable salt thereof. The drug loading is represented by p, the number of drug-linker molecules per antibody. Depending on the context, p can represent the average number of drug-linker molecules per antibody, also referred to as the average drug loading. The variable p ranges from 1 to 20 and is preferably from 1 to 8. In some preferred

embodiments, when p represents the average drug loading, p ranges from about 2 to about 5. In some embodiments, p is about 2, about 3, about 4, or about 5. In some aspects, the antibody is conjugated to the drug linker via a sulfur atom of a cysteine residue that is engineered into the antibody. In some aspects, the cysteine residue is engineered into the antibody at position 239 (IgG1) as determined by the EU index (Kabat, *Sequences of Proteins of Immunological Interest* (National Institutes of Health, Bethesda, MD, 1987 and 1991)).

VI. Animal Models of Immunological Disorders or BCMA-Expressing Cancers

[00147] The anti-BCMA antibodies or derivatives can be tested or validated in animal models of immunological disorders or BCMA-expressing cancers. Examples for animal models of systemic and organ-specific autoimmune diseases including diabetes, lupus, systemic sclerosis, Sjögren's Syndrome, experimental autoimmune encephalomyelitis (multiple sclerosis), thyroiditis, myasthenia gravis, arthritis, uveitis, inflammatory bowel disease have been described by Bigazzi, "Animal Models of Autoimmunity: Spontaneous and Induced," in The Autoimmune Diseases (Rose and Mackay eds., Academic Press, 1998) and in "Animal Models for Autoimmune and Inflammatory Disease," in Current Protocols in Immunology (Coligan et al. eds., Wiley and Sons, 1997).

[00148] Allergic conditions, e.g., asthma and dermatitis, can also be modeled in rodents. Airway hypersensitivity can be induced in mice by ovalbumin (Tomkinson et al., 2001, J. Immunol. 166:5792-800) or *Schistosoma mansoni* egg antigen (Tesciuba et al., 2001, J. Immunol. 167:1996-2003). The Nc/Nga strain of mice show marked increase in serum IgE and spontaneously develop atopic dermatitis-like lesions (Vestergaard et al., 2000, Mol. Med. Today 6:209-10; Watanabe et al., 1997, Int. Immunol. 9:461-66; Saskawa et al., 2001, Int. Arch. Allergy Immunol. 126:239-47).

[00149] Injection of immuno-competent donor lymphocytes into a lethally irradiated histocompatible host is a classical approach to induce GVHD in mice. Alternatively, the parent B6D2F1 murine model provides a system to induce both acute and chronic GVHD. In this model the B6D2F1 mice are F1 progeny from a cross between the parental strains of C57BL/6 and DBA/2 mice. Transfer of DBA/2 lymphoid cells into non-irradiated B6D2F1 mice causes chronic GVHD, whereas transfer of C57BL/6, C57BL/10 or B10.D2 lymphoid cells causes acute GVHD

(Slabback et al., 2000, *Bone Marrow Transpl.* 26:931-938; Kataoka et al., 2001, *Immunology* 103:310-318).

[00150] Additionally, both human hematopoietic stem cells and mature peripheral blood lymphoid cells can be engrafted into SCID mice, and these human lympho-hematopoietic cells remain functional in the SCID mice (McCune et al., 1988, *Science* 241:1632-1639; Kamel-Reid and Dick, 1988, *Science* 242:1706-1709; Mosier et al., 1988, *Nature* 335:256-259). This has provided a small animal model system for the direct testing of potential therapeutic agents on human lymphoid cells. (See, e.g., Tournay et al., 2001, *J. Immunol.* 166:6982-6991).

[00151] Moreover, small animal models to examine the *in vivo* efficacies of the anti-BCMA antibodies or derivatives can be created by implanting BCMA-expressing human tumor cell lines into appropriate immunodeficient rodent strains, e.g., athymic nude mice or SCID mice. Examples of BCMA-expressing human lymphoma cell lines include, for example, Daudi (Ghetie et al., 1994, *Blood* 83:1329-36; Ghetie et al., 1990, *Int. J. Cancer* 15:481-85; de Mont et al., 2001, *Cancer Res.* 61:7654-59), Ramos (Ma et al., 2002, *Leukemia* 16:60-6; Press et al., 2001, *Blood* 98:2535-43), HS-Sultan (Cattan and Maung, 1996, *Cancer Chemother. Pharmacol.* 38:548-52; Cattan and Douglas, 1994, *Leuk. Res.* 18:513-22), Raji (Ochakovskaya et al., 2001, *Clin. Cancer Res.* 7:1505-10; Breisto et al., 1999, *Cancer Res.* 59:2944-49), and CA46 (Kreitman et al., 1999, *Int. J. Cancer* 81:148-55). Non-limiting example of a BCMA-expressing Hodgkin's lymphoma line is L540cy (Barth et al., 2000, *Blood* 95:3909-14; Wahl et al., 2002, *Cancer Res.* 62:3736-42). Non-limiting examples of BCMA expressing human renal cell carcinoma cell lines include 786-O (Ananth et al., 1999, *Cancer Res.* 59:2210-16; Datta et al., 2001, *Cancer Res.* 61:1768-75), ACHN (Hara et al., 2001, *J. Urol.* 166:2491-94; Miyake et al., 2002, *J. Urol.* 167:2203-08), Caki-1 (Prewett et al., 1998, *Clin. Cancer Res.* 4:2957-66; Shi and Siemann, 2002, *Br. J. Cancer* 87:119-26), and Caki-2 (Zellweger et al., 2001, *Neoplasia* 3:360-67). Non-limiting examples of BCMA-expressing nasopharyngeal carcinoma cell lines include C15 and C17 (Busson et al., 1988, *Int. J. Cancer* 42:599-606; Bernheim et al., 1993, *Cancer Genet. Cytogenet.* 66:11-5). Non-limiting examples of BCMA-expressing human glioma cell lines include U373 (Palma et al., 2000, *Br. J. Cancer* 82:480-7) and U87MG (Johns et al., 2002, *Int. J. Cancer* 98:398-408). These tumor cell lines can be established in immunodeficient rodent hosts either as solid

tumor by subcutaneous injections or as disseminated tumors by intravenous injections. Once established within a host, these tumor models can be applied to evaluate the therapeutic efficacies of the anti-BCMA antibody or derivatives as described herein on modulating in vivo tumor growth.

VII. Therapeutic Applications

[00152] The anti-BCMA antibodies of the invention can be used to treat cancer. Some such cancers show detectable levels of BCMA measured at either the protein (e.g., by immunoassay using one of the exemplified antibodies) or mRNA level. Some such cancers show elevated levels of BCMA relative to noncancerous tissue of the same type, preferably from the same patient. An exemplary level of BCMA on cancer cells amenable to treatment is 5000-150000 BCMA molecules per cell, although higher or lower levels can be treated. Optionally, a level of BCMA in a cancer is measured before performing treatment.

[00153] Cancers treatable with antibodies of the invention include solid tumors and hematological cancers, such as leukemias and lymphomas. The antibodies are particularly suitable for cancers of B-cells. Examples of cancers treatable with the antibodies include: adult and pediatric acute myeloid leukemia (AML), chronic myeloid leukemia (CML), acute lymphocytic leukemia (ALL), chronic lymphocytic leukemia (CLL) and secondary leukemia; non-Hodgkin's lymphoma (NHL) and Hodgkin's disease; myelodysplastic syndromes (MDS), myeloproliferative syndromes (MPS) multiple myeloma, Waldenström's macroglobulinemia or Burkett's lymphoma., malignant plasma cell neoplasms, BCMA+high-grade lymphoma, Kahler's disease and myelomatosis; plasma cell leukemia; plasmacytoma; B-cell prolymphocytic leukemia; hairy cell leukemia; follicular lymphoma (including follicular non-Hodgkin's lymphoma types); Burkitt's lymphoma (Endemic Burkitt's lymphoma; sporadic Burkitt's lymphoma); marginal zone lymphoma (Mucosa-Associated Lymphoid Tissue: MALT 1 MALToma; Monocytoid B cell lymphoma; splenic lymphoma with villous lymphocytes); mantle cell lymphoma; large cell lymphoma (diffuse large cell; diffuse mixed cell; immunoblastic lymphoma; primary mediastinal B cell lymphoma; angiocentric lymphoma pulmonary B cell); small lymphocytic lymphoma (SLL); recursor B-lymphoblastic lymphoma; myeloid leukemia (granulocytic; myelogenous; acute myeloid leukemia; chronic myeloid leukemia; subacute

myeloid leukemia; myeloid sarcoma; chloroma; granulocytic sarcoma; acute promyelocytic leukemia; acute myelomonocytic leukemia); Waldenstrom's macroglobulinemia, or other B-cell leukemia or lymphoma.

[00154] The antibodies of the invention are also useful for immune disorders mediated by immune cells expressing BCMA, particularly B-cell mediated disorders. Examples of such diseases include rheumatoid arthritis, systemic lupus E (SLE), Type I diabetes, asthma, atopic dermatitis, allergic rhinitis, thrombocytopenic purpura, multiple sclerosis, psoriasis, Sjorgren's syndrome, Hashimoto's thyroiditis, Grave's disease, primary biliary cirrhosis, Wegener's granulomatosis, tuberculosis, and graft versus host disease

immune-mediated thrombocytopenia, haemolytic anaemia, bullous pemphigoid, myasthenia gravis, Graves' disease, Addison's disease, pemphigus foliaceus, psoriasis, psoriatic arthritis, and ankylosing spondylitis.

[00155] Anti-BCMA antibodies alone or as drug-conjugates thereof, are administered in an effective regime meaning a dosage, route of administration and frequency of administration that delays the onset, reduces the severity, inhibits further deterioration, and/or ameliorates at least one sign or symptom of cancer. If a patient is already suffering from cancer, the regime can be referred to as a therapeutically effective regime. If the patient is at elevated risk of the cancer relative to the general population but is not yet experiencing symptoms, the regime can be referred to as a prophylactically effective regime. In some instances, therapeutic or prophylactic efficacy can be observed in an individual patient relative to historical controls or past experience in the same patient. In other instances, therapeutic or prophylactic efficacy can be demonstrated in a preclinical or clinical trial in a population of treated patients relative to a control population of untreated patients.

[00156] Exemplary dosages for a monoclonal antibody are 0.1 mg/kg to 50 mg/kg of the patient's body weight, more typically 1 mg/kg to 30 mg/kg, 1 mg/kg to 20 mg/kg, 1 mg/kg to 15 mg/kg, 1 mg/kg to 12 mg/kg, or 1 mg/kg to 10 mg/kg, or 2 mg/kg to 30 mg/kg, 2 mg/kg to 20 mg/kg, 2 mg/kg to 15 mg/kg, 2 mg/kg to 12 mg/kg, or 2 mg/kg to 10 mg/kg, or 3 mg/kg to 30 mg/kg, 3 mg/kg to 20 mg/kg, 3 mg/kg to 15 mg/kg, 3 mg/kg to 12 mg/kg, or 3 mg/kg to 10 mg/kg. Exemplary dosages for active monoclonal antibody drug conjugates thereof, e.g.,

auristatins, are 1 mg/kg to 7.5 mg/kg, or 2 mg/kg to 7.5 mg/kg or 3 mg/kg to 7.5 mg/kg of the subject's body weight, or 0.1-20, or 0.5-5 mg/kg body weight (e.g., 0.5, 1, 2, 3, 4, 5, 6, 7, 8, 9 or 10 mg/kg) or 10-1500 or 200-1500 mg as a fixed dosage. Exemplary dosages for highly active monoclonal antibody drug conjugates thereof, e.g., PBDs, are 1.0 μ g/kg to 1.0 mg/kg, or 1.0 μ g/kg to 500.0 μ g/kg of the subject's body weight. In some methods, the patient is administered then antibody or ADC every two, three or four weeks. The dosage depends on the frequency of administration, condition of the patient and response to prior treatment, if any, whether the treatment is prophylactic or therapeutic and whether the disorder is acute or chronic, among other factors.

[00157] Administration can be parenteral, intravenous, oral, subcutaneous, intra-arterial, intracranial, intrathecal, intraperitoneal, topical, intranasal or intramuscular. Administration can also be localized directly into a tumor. Administration into the systemic circulation by intravenous or subcutaneous administration is preferred. Intravenous administration can be, for example, by infusion over a period such as 30-90 min or by a single bolus injection.

[00158] The frequency of administration depends on the half-life of the antibody or antibody-drug conjugate in the circulation, the condition of the patient and the route of administration among other factors. The frequency can be daily, weekly, monthly, quarterly, or at irregular intervals in response to changes in the patient's condition or progression of the cancer being treated. An exemplary frequency for intravenous administration is between twice a week and quarterly over a continuous course of treatment, although more or less frequent dosing is also possible. Other exemplary frequencies for intravenous administration are between once weekly or once monthly over a continuous course of treatment, although more or less frequent dosing is also possible. For subcutaneous administration, an exemplary dosing frequency is daily to monthly, although more or less frequent dosing is also possible.

[00159] The number of dosages administered depends on the nature of the cancer or autoimmune disease (e.g., whether presenting acute or chronic symptoms) and the response of the disorder to the treatment. For acute disorders or acute exacerbations of a chronic disorder between 1 and 10 doses are often sufficient. Sometimes a single bolus dose, optionally in divided form, is sufficient for an acute disorder or acute exacerbation of a chronic disorder.

Treatment can be repeated for recurrence of an acute disorder or acute exacerbation. For chronic disorders, an antibody can be administered at regular intervals, e.g., weekly, fortnightly, monthly, quarterly, every six months for at least 1, 5 or 10 years, or the life of the patient.

[00160] Pharmaceutical compositions for parenteral administration are preferably sterile and substantially isotonic and manufactured under GMP conditions. Pharmaceutical compositions can be provided in unit dosage form (i.e., the dosage for a single administration).

Pharmaceutical compositions can be formulated using one or more physiologically acceptable carriers, diluents, excipients or auxiliaries. The formulation depends on the route of administration chosen. For injection, antibodies can be formulated in aqueous solutions, preferably in physiologically compatible buffers such as Hank's solution, Ringer's solution, or physiological saline or acetate buffer (to reduce discomfort at the site of injection). The solution can contain formulatory agents such as suspending, stabilizing and/or dispersing agents. Alternatively antibodies can be in lyophilized form for constitution with a suitable vehicle, e.g., sterile pyrogen-free water, before use. The concentration of antibody in a liquid formulation can be e.g., .01-10 mg/ml, such as 1.0 mg/ml.

[00161] Treatment with antibodies of the invention can be combined with chemotherapy, radiation, stem cell treatment, surgery other treatments effective against the disorder being treated. Useful classes of other agents that can be administered with antibodies to BCMA include, for example, antibodies to other receptors expressed on cancerous cells, antitubulin agents (e.g., auristatins), DNA minor groove binders (e.g., PBDs), DNA replication inhibitors, alkylating agents (e.g., platinum complexes such as *cis*-platin, mono(platinum), bis(platinum) and tri-nuclear platinum complexes and carboplatin), anthracyclines, antibiotics, antifolates, antimetabolites, chemotherapy sensitizers, duocarmycins, etoposides, fluorinated pyrimidines, ionophores, lexitropsins, nitrosoureas, platinols, pre-forming compounds, purine antimetabolites, puromycins, radiation sensitizers, steroids, taxanes, topoisomerase inhibitors, vinca alkaloids, and the like. The same additional treatments just mentioned for cancer can also be used for immune mediated disorders. Additional agents for immune mediated disorders include immune suppressors such as mast cell degranulation inhibitors, anti-histamines,

corticosteroids, NSAIDs, azathioprine, cyclophosphamide, leukeran, and cyclosporine and biologic anti-inflammatory agents, such as Tysabri® or Humira®.

[00162] Treatment with anti-BCMA antibodies, optionally in combination with any of the other agents or regimes described above alone or as an antibody drug conjugate, can increase the median progression-free survival or overall survival time of patients with cancer, especially when relapsed or refractory, by at least 30% or 40% but preferably 50%, 60% to 70% or even 100% or longer, compared to the same treatment (e.g., chemotherapy) but without the anti-BCMA antibody. In addition or alternatively, treatment (e.g., standard chemotherapy) including the anti-BCMA antibody, alone or as an antibody-drug conjugate, can increase the complete response rate, partial response rate, or objective response rate (complete + partial) of patients with tumors by at least 30% or 40% but preferably 50%, 60% to 70% or even 100% compared to the same treatment (e.g., chemotherapy) but without the anti-BCMA antibody.

[00163] Typically, in a clinical trial (e.g., a phase II, phase II/III or phase III trial), the aforementioned increases in median progression-free survival and/or response rate of the patients treated with standard therapy plus the anti-BCMA antibody, relative to the control group of patients receiving standard therapy alone (or plus placebo), are statistically significant, for example at the p = 0.05 or 0.01 or even 0.001 level. The complete and partial response rates are determined by objective criteria commonly used in clinical trials for cancer, e.g., as listed or accepted by the National Cancer Institute and/or Food and Drug Administration.

VIII. Other Applications

[00164] The anti-BCMA antibodies disclosed herein can be used for detecting BCMA in the context of clinical diagnosis or treatment or in research. Expression of BCMA on a cancer provides an indication that the cancer is amenable to treatment with the antibodies of the present invention. The antibodies can also be sold as research reagents for laboratory research in detecting cells bearing BCMA and their response to various stimuli. In such uses, monoclonal antibodies can be labeled with fluorescent molecules, spin-labeled molecules, enzymes or radioisotypes, and can be provided in the form of kit with all the necessary reagents to perform the assay for BCMA. The antibodies can also be used to purify BCMA protein, e.g., by affinity chromatography.

[00165] Any feature, step, element, embodiment, or aspect of the invention can be used in combination with any other unless specifically indicated otherwise. Although the present invention has been described in some detail by way of illustration and example for purposes of clarity and understanding, it will be apparent that certain changes and modifications may be practiced within the scope of the appended claims.

EXAMPLES

Example 1: Antibody development

Preparation of Recombinant BCMA Extracellular Domain (BCMA ECD)

[00166] The extracellular domain (ECD) of human (amino acids 1-51) and mouse BCMA (amino acids 1-46) were cloned and expressed as a GST fusion protein (pGEX4T1; Amersham Biosciences). Purified BCMA ECD was obtained by capturing the BCMA fusion protein with glutathione-sepharose and releasing the BCMA ECD by protease digestion with thrombin. Thrombin was subsequently removed by benzamidine sepharose.

Identification of BCMA Expression on Malignant B-cell Lines

[00167] Quantitative flow cytometry was performed on multiple myeloma cell lines using Vicky-1, a commercial antibody for BCMA (Alexis Biotechnology). Results showed that BCMA is prevalent among myeloma lines tested. NCI H929 showed positive cell surface staining for BCMA but lacked expression of either BR3 or TACI. Because NCI H929 expressed BCMA but not BR3 or TACI, it was used for cell-based screening of the BCMA hybridomas.

Development of a Transfected BCMA Cell Line.

[00168] Stable cell lines were developed by transfecting HEK 293 cells with either a full-length BCMA clone or an empty vector. Flow cytometry confirmed positive expression of BCMA on the surface of the BCMA transfected (293: BCMA) but not the vector empty control plasmid (293: vector). These cell lines were subsequently used as a tool to confirm the specificity of cloned BCMA antibodies.

Example 2: Immunization and Screening of Uncloned Hybridoma Wells

Immunization and Screening of Antiserum

[00169] Our immunization strategy used amino acids 1-50 of the BCMA ECD so that epitopes internal and external to ligand binding domain could be targeted by antibodies (Figs. 1A and 1B) KLH-conjugated BCMA ECD was generated from a commercial source (Alexis Biochemicals). Rats were immunized KLH-conjugated BCMA using Titermax adjuvant until a maximum immune response was detected by ELISA. Immunized rats serum was also screened for ability to block APRIL binding in a plate-based assay. Rat 2-3 was selected for fusion because the antiserum had a significant titer of human BCMA antibodies and it displayed robust blocking activity.

[00170] Spleen cells from rat 2-3 were harvested, fused to X-63.Ag8.653.3.12.11 mouse myeloma cells and selected as described (Goding, 1989). Culture supernatants from the resulting hybridomas were screened by ELISA using purified hBCMA-GST (see flow chart in Fig. 2). Eighty positive wells were identified and selected for expansion. Sixty of the eighty positives wells continued to showed an OD > 0.5 by ELISA following expansion. These sixty uncloned hybridoma wells were then screened in secondary assays for cell-based binding, ligand blockade activity, and cross-reactivity to mouse BCMA. This led to the identification of twelve lead BCMA hybridoma wells. Cell binding data and ligand blockade activity from these twelve lead wells is summarized in Fig. 3. Hybridoma well 17 showed cell binding and ligand blockade activity that superseded the commercial monoclonal Vicky-1 (Alexis Biochemicals). Eight wells (indicated with a red asterisk in Fig. 3) were taken forward for cloning based on their ability to bind BCMA-positive cells or block ligand binding.

Example 3: Characterization of Clonal Hybridomas

Cell binding and Ligand Blockade Activity.

[00171] Hybridoma wells 11, 17, 20, 29, 40, 45 and 70 were taken through 2 rounds of limited dilution cloning. From this point forward, the antibodies will be designated with the formal clone ID shown in Table 1. The specific binding of the antibodies to 293: BCMA cells but not to the 293: vector control cells confirms that the antibodies are binding to BCMA.

Table 1: Formal Clone IDs.

Uncloned Designation	Cloned ID
11	SG16.11

17	SG16.17
20	SG16.20
29	SG16.29
40	SG16.40
45	SG16.45
70	SG16.70

[00172] Ligand blockade activity of the new BCMA antibodies was compared using supernatant from the uncloned master well, supernatant from the cloned well and purified antibody from a cloned well (Fig. 4). A commercial antibody was used as a positive control. SG-16.17 gave significant blocking of APRIL binding using culture supernatant from the cloned hybridoma well. A titration of the SG16.17 blockade of APRIL binding was performed in a separate experiment using purified SG16.17 and the commercial antibody (Fig. 5). Purified SG16.17 displayed improved blocking activity across similar concentrations when compared to the commercial antibody. SG-16.45 showed dose-dependent inhibition of April binding although not as strongly as SG-16.17. Ligand blockade activity for the remaining BCMA antibodies (SG-16.11, SG16.20, SG16.29, SG16.40, and SG16.70) was more modest. Certain blocking BCMA antibodies show >75% inhibition of APRIL binding as was observed with SG-16.17. More “modest” blocking antibodies including SG-16.11, SG-16.20, SG-16.29, SG-16.40, and SG-16.70 showed about 30% inhibition for APRIL binding (Fig. 4).

[00173] The ability of BAFF to bind immobilized BCMA was also analyzed in the presence and absence of purified BCMA antibodies. Pretreatment with BCMA antibodies SG16.17, SG16.40, SG16.20 and SG17.70 all resulted in a titratable inhibition of BAFF binding to BCMA (Fig. 6). The relative inhibition was determined by binding BAFF to immobilized BCMA in the absence of antibody treatment (Fig. 6, asterisk). Taken together, the data in Figs. 5 and 6 shows that BCMA antibodies can block ligand binding of APRIL and BAFF to BCMA and thereby interfere with B cell survival signals.

Example 4: Testing SG16.17 and SG16.45 antibodies for ADCC and cytotoxicity as an ADC

[00174] The SG16.17 antibody was converted into a rat-human chimeric IgG by fusing the rat V_H and V_L domains to wild-type human IgG1 heavy chain and κ light chain constant domains, respectively. The chimerized antibody, designated cSG16.17 wild-type, showed similar antigen binding properties when compared with the parental antibody SG16.17. Next, we installed Fc mutations, S239D:A330L:I332E, known to enhance ADCC, to generate cSG16.17 mutant. Similar to cSG16.17 wild-type, generation of the Fc triple mutant did not alter the antigen-binding properties of cSG16.17 mutant. Evaluation of cSG16.17 wild-type and cSG16.17 mutant in an ADCC assay with purified natural killer cells resulted in dose-dependent lysis of JJN3 and U266 cells whereas no significant lysis was observed with a nonbinding human IgG control. The cSG16.17 wild-type antibody displayed limited ADCC activity on JJN3 cells, which was increased ~100-fold in potency and >2-fold in efficacy (maximal lysis) by cSG16.17 mutant. Similarly, for U266 cells, the ADCC activity of cSG16.17 mutant was enhanced ~100-fold in potency and 2-fold in efficacy compared with the parent chimeric antibody. The concentration of cSG16.17 mutant required for maximal lysis of both JJN3 and U266 cells was ~100 pmol/L. In contrast, the dissociation constant (K_D) of cSG16.17 on JJN3 and U266 cells was estimated as 15 and 10 nmol/L, respectively. Thus, maximal lysis by cSG16.17 mutant was achieved at concentrations well below those required to reach saturation binding.

[00175] We assessed the ability of SG16.17 and SG16.45 to induce cytotoxicity as ADCs using vcMMAF with a stoichiometry of eight drugs per antibody. SG16.17 or SG16.45-vcMMAF8 was potently cytotoxic against H929 cells. No decline in cell viability was observed using a nonbinding control ADC or unconjugated antibodies. We also examined the potency of SG16.17 ADC across other MM cell lines, including JJN3 and U266 cell lines. SG16.17-vcMMAF8 showed consistent and high potency (IC_{50} values \leq 130 pmol/L) across all three MM cell lines whereas SG16.45-vcMMAF8 showed more variability and less overall potency.

Example 5: Testing SG16.17 antibody for binding to Fc γ RIIIa, and signaling through Fc γ RIIIa

[00176] For the binding assay, CHO cells were transfected with Fc γ RIIIa (hCD16) and binding of labelled h00 antibody measured in competition with chimeric SG16.17 with wild type IgG1 and IgG1 S239D, A330L, I332E genotype, and various IgG1 control antibodies. Fig. 12 shows that chimeric SG16.17 competed more strongly than two control antibodies, rituximab and cOKT9.

The mutant form of SG16.17 competed more strongly than the wild type IgG1 form. The signaling assay uses U266 target cells expressing BCMA, Jurkat effector cells expressing Fc γ RIIIa and engineered to express a luciferase reporter from a NFAT response element and Bio-Glo indicator. cSG16.17 G1 WT & S239D, A330L, I332E both elicited Fc γ RIIIa signaling with that from the S239D, A330L, I332E form being stronger (Fig. 13).

Example 6: Humanization of SG16.17

Table 2: Humanizing Mutations in hSG16.17 Heavy Chain Variants

vH Variant	HV Exon Acceptor Sequence	Donor Framework Residues	Acceptor CDR Residues
hvH1	HV1-2/HJ3	H8, H20, H48, H67, H69, H71, H73, H76, H80, H88, H91, H93	none
hvH2	HV1-2/HJ3	H20, H48, H69, H71, H73, H76, H80, H88, H91, H93	H34, H50, H58, H60, H61, H62, H64, H65
hvH3	HV1-2/HJ3	H20, H48, H67, H69, H71, H73, H76, H80, H88, H91, H93	H58, H60, H61, H62, H64, H65
hvH4	HV1-2/HJ3	H48, H67, H69, H71, H73, H76, H80, H88, H91, H93	H34, H50, H58, H60, H61, H62, H64, H65
hvH5	HV1-46/HJ3	H48, H67, H71, H73, H76, H78, H80, H91, H93	none
hvH6	HV1-46/HJ3	H8, H20, H48, H71, H73, H76, H78, H80, H91, H93	none

Table 3: Humanizing Mutations in hSG16.17 Kappa Light Chain Variants

vK Variant	KV Exon Acceptor Sequence	Donor Framework Residues	Acceptor CDR Residues
hVK2	KV1-12/KJ5	L46, L48, L87	L53
hVK3	KV1-12/KJ5	L46, L48, L87	L24, L53
hVK4	KV1-12/KJ5	L46, L48, L78, L85, L87	none
hVK5	KV1-12/KJ5	L40, L46, L48, L87	L24, L53

Table 4: Specific Framework Mutations in hSG16.17 Heavy Chain Variants

Variant	H8	H20	H48	H67	H69	H71	H73	H76	H78	H80	H88	H91	H93	% Human
hvH1	R*	L*	I*	A*	M*	A*	K*	N*	A	V*	A*	F*	T*	79.6
hvH2	G	L*	I*	V	M*	A*	K*	N*	A	V*	A*	F*	T*	88.8
hvH3	G	L*	I*	A*	M*	A*	K*	N*	A	V*	A*	F*	T*	86.7
hvH4	G	V	I*	A*	M*	A*	K*	N*	A	V*	A*	F*	T*	88.8
hvH5	G	V	I*	A*	M	A*	K*	N*	A*	V*	A	F*	T*	78.6
hvH6	R*	L*	I*	V	M	A*	K*	N*	A*	V*	A	F*	T*	85.7

*Rat residues

Table 5: Specific Framework Mutations in hSG16.17 Kappa Light Chain Variants

Variant	L40	L46	L48	L78	L85	L87	% Human
hvK2	P	V*	V*	L	T	F*	86.3
hvK3	P	V*	V*	L	T	F*	87.4
hvK4	P	V*	V*	M*	D*	F*	83.2
hvK5	S*	V*	V*	L	T	F*	86.3

*Rat residues

[00177] The rat heavy and light chain variable regions of the rat hybridoma expressing SG16.17 were sequenced. HV1-2/HJ3 (SEQ ID NO: 9) or HV1-46/HJ3 (SEQ ID NO: 10) was used as the human acceptor sequence for the heavy chain and KV1-12/KJ5 (SEQ ID NO: 18) was used as the human acceptor sequence for the light chain.

[00178] Positions differing between rat donor and human acceptor sequences included H8, H20, H48, H67, H69, H71, H76, H78, H80, H88, H91, H93, L40, L46, L48, L78, L85 and L87. Different permutations of these residues were included as back mutations in different humanized heavy chain and light chain sequences. Several rat residues in the Kabat CDRs were also tested for replacement with corresponding residues of the human acceptor sequences. The positions of these residues were H34, H50, H58, H60, H61, H62, H64 and H65, and L24 and L53. Six humanized heavy chain variants and four humanized light chain variants were designed and expressed. Tables 2 and 3 indicate the human acceptor sequence, back mutations (donor framework residues), and CDR substitutions (Acceptor CDR residues) in each humanized variant chain. Tables 4 and 5 indicate the amino acids occupying each of the positions considered for back mutation in each of the humanized variant chain. These tables also indicate the percent of residue identical to the closest human germline sequence. According to recent INN Guidelines

only antibodies with at least 85% identity to a human germline sequence in both heavy and light chains can be referred to as humanized. Figs. 7-9 show alignments of humanized heavy chain variable regions with the rat variable region and human acceptor sequences. Figs. 10 and 11 show alignment of the humanized light chain variable regions with the rat variable region and human acceptor sequences. The C-terminal arginine (R) of the variable light chains can alternatively be regarded as the N-terminal arginine of the light chain constant region.

[00179] The six humanized heavy chains and four humanized light chains were tested in all 24 possible permutations for binding to BCMA expressed on NCI-H929 cells, which express about 50,000 molecules of BCMA per cell. The results are shown in Table 6 below. In brief, all of the humanized light chains showed good binding. Of the humanized heavy chains, variants VH1, VH3 and VH5 all showed improved binding compared with either chimeric or rat SG16.17 antibody.

Table 6: Humanized Antibodies hSG16.17 Binding to BCMA Expressed on NCI-H929 Cells

hSG16.17	vH	vK	NCI-H929 3-pt Assay
1	vH1	vK2	++++
2	vH1	vK3	++++
3	vH1	vK4	++++
4	vH1	vK5	++++
5	vH2	vK2	-
6	vH2	vK3	-
7	vH2	vK4	-
8	vH2	vK5	-
9	vH3	vK2	++++
10	vH3	vK3	++++
11	vH3	vK4	++++
12	vH3	vK5	++++
13	vH4	vK2	-
14	vH4	vK3	-
15	vH4	vK4	-

hSG16.17	vH	vK	NCI-H929 3-pt Assay
16	vH4	vK5	-
17	vH5	vK2	++++
18	vH5	vK3	++++
19	vH5	vK4	++++
20	vH5	vK5	++++
21	vH6	vK2	++
22	vH6	vK3	++
23	vH6	vK4	++
24	vH6	vK5	++
cSG16.17			+++
rSG16.17			+++

[00180] The humanized antibodies performing best on the NCI-H929 assay (i.e., those containing VH1, VH3 or VH5 heavy chains, were further tested for binding to U266 cells at a full range of concentration points. In this assay, humanized antibodies containing VH1 heavy chains (regardless of which humanized light chain variant was included) showed enhanced binding relative to rat or chimeric SG16.17. Humanized antibodies containing VH3 or VH5 heavy chains (regardless of which humanized light chain variant was included) showed the same binding within experimental error as rat or chimeric SG16.17 binding. Humanized antibodies containing VH2 or VH6 variable regions showed reduced binding relative to rat or chimeric SG16.17 regardless of which humanized light chain variant was included.

[00181] The humanized antibodies performing best on the NCI-H929 assay were also compared for protein expression level, monomer level and percentage sequence identity to human germ line as shown in Table 7 below.

Table 7:

hSG16.17	vH	vK	hBCMA Binding	Transient Titer (mg/L)	aSEC (% Monomer)	≥85% human (vH, vK) & INN Designation			Lead Selection
						79.6	86.3	Mix	
1	vH1	vK2	++++	139	90.4	79.6	86.3	Mix	Y
2	vH1	vK3	++++	126	89.6	79.6	87.4	Mix	Y
3	vH1	vK4	++++	80	94.6	79.6	83.2	Chimeric	N
4	vH1	vK5	++++	119	89.5	79.6	86.3	Mix	N

9	vH3	vK2	++++	129	94.1	86.7	86.3	Humanized	Y
10	vH3	vK3	++++	116	94.1	86.7	87.4	Humanized	Y
11	vH3	vK4	++++	82	95.2	86.7	83.2	Mix	Y
12	vH3	vK5	++++	117	93.5	86.7	86.3	Humanized	Y
17	vH5	vK2	++++	97	96.2	78.6	86.3	Mix	Y
18	vH5	vK3	++++	86	96.1	78.6	87.4	Mix	Y
19	vH5	vK4	++++	65	96.5	78.6	83.2	Chimeric	N
20	vH5	vK5	++++	73	95.0	78.6	86.3	Mix	Y

[00182] The VH3 VK2 humanized antibody was selected as the lead humanized antibody based on it having the same binding affinity for human BCMA as rat and mouse SG16.17 antibodies (within experimental error); greater than 85% identity to human germline sequence in both heavy and light chain variable regions, good expression and high percentage of monomers.

Example 7: Humanization of SG16.45

Table 8: Humanizing Mutations in hSG16.45 Heavy Chain Variants

vH Variant	HV Exon Acceptor Sequence	Donor Framework Residues	Acceptor CDR Residues
hvH1	HV3-23/HJ3	H30, H37, H48, H93, H94, H107	none
hvH2	HV3-23/HJ3	H30, H37, H48, H93, H94, H107	H50, H60
hvH3	HV3-23/HJ3	H30, H37, H48, H76, H93, H94, H107	H50, H60
hvH4	HV3-23/HJ3	H30, H48, H76, H93, H94	H50
hvH5	HV3-74/HJ3	H30, H93, H94	H50
hvH6	HV3-9/HJ3	H30, H93, H94	H50, H60

Table 9: Humanizing Mutations in hSG16.45 Kappa Light Chain Variants

vK Variant	KV Exon Acceptor Sequence	Donor Framework Residues	Acceptor CDR Residues
hvK1	KV3-20/KJ2	L14, L19, L21, L38, L58, L71, L78	L24, L26
hvK2	KV3-20/KJ2	none	L24, L26
hvK3	KV3-20/KJ2	L21, L38, L58, L71	L24, L26
hvK5	KV3-20/KJ2	L38, L71	none

Table 10: Specific Framework Mutations in hSG16.45 Heavy Chain Variants

Variant	H30	H37	H48	H76	H93	H94	H107	% Human
hvH1	N*	I*	I*	N	T*	S*	V*	86.5

hvH2	N*	I*	I*	N	T*	S*	V*	88.5
hvH3	N*	I*	I*	S*	T*	S*	V*	87.5
hvH4	N*	V	I*	S*	T*	S*	T	87.5
hvH5	N*	V	V	N	T*	S*	T	88.5
hvH6	N*	V	V	N	T*	S*	T	88.5

*Rat residues

Table 11: Specific Framework Mutations in hSG16.45 Kappa Light Chain Variants

Variant	L14	L19	L21	L38	L58	L71	L78	% Human
hvK1	A*	V*	I*	H*	V*	Y*	M*	79.2
hvK2	L	A	L	Q	I	F	L	86.5
hvK3	L	A	I*	H*	V*	Y*	L	82.3
hvK5	L	A	L	H*	I	Y*	L	82.3

*Rat residues

[00183] The rat heavy and light chain variable regions of the rat hybridoma expressing SG16.45 were sequenced. HV3-23/HJ3 (SEQ ID NO: 24) was used as the human acceptor sequence for the heavy chain and KV3-20/KJ2 (SEQ ID NO: 34) was used as the human acceptor sequence for the light chain.

[00184] Variable region framework positions differing between rat donor and human acceptor sequences included H30, H37, H48, H67, H93, H94 and H107 and positions L14, L19, L21, L38, L58, L71 and L78. Different permutations of these residues were included as back mutations in different humanized heavy chain and light chain sequences. Several rat residues in the Kabat CDRs were also tested for replacement with corresponding residues of the human acceptor sequences. The positions of these residues were H50, H60, L24 and L26. Six humanized heavy chain variants and four humanized light chain variants were designed and expressed. Tables 8 and 9 indicate the human acceptor sequence, back mutations (donor framework residues), and CDR substitutions (Acceptor CDR residues) in each humanized variant chain. Tables 10 and 11 indicate the amino acids occupying each of the positions considered for back mutation in each of the humanized variant chain. These tables also indicate the percent of residue identical to the closest human germline sequence. According to recent INN Guidelines only antibodies with at least 85% identity to a human germline sequence in both heavy and light chains can be

referred to as humanized. Figs. 14-17 show an alignment of humanized heavy chain variable regions with the rat variable region and human acceptor sequences. Figs. 18 and 19 show alignments of the light chain variable regions. The C-terminal arginine (R) of the variable light chains can alternatively be regarded as the N-terminal arginine of the light chain constant region.

[00185] The six humanized heavy chains and four humanized light chains were tested in all 24 possible permutations for binding to BCMA expressed on NCI-H929 cells, which express about 50,000 molecules of BCMA per cell. The results are shown in Table 12 below.

Table 12: Humanized Antibodies hSG16.45 Binding to BCMA Expressed on NCI-H929 Cells

hSG16.45	vH	vK	NCI-H929 3-pt Assay
1	vH1	vK1	+++
2	vH1	vK2	+++
3	vH1	vK3	+++
4	vH1	vK5	+++
5	vH2	vK1	-
6	vH2	vK2	-
7	vH2	vK3	-
8	vH2	vK5	-
9	vH3	vK1	-
10	vH3	vK2	-
11	vH3	vK3	-
12	vH3	vK5	++
13	vH4	vK1	+
14	vH4	vK2	+
15	vH4	vK3	+

hSG16.45	vH	vK	NCI-H929 3-pt Assay
16	vH4	vK5	++
17	vH5	vK1	++
18	vH5	vK2	++
19	vH5	vK3	++
20	vH5	vK5	++
21	vH6	vK1	+
22	vH6	vK2	+
23	vH6	vK3	+
24	vH6	vK5	++
cSG16.45			+++
rSG16.45			+++

[00186] The humanized antibodies performing best on the NCI-H929 assay, were further tested for binding to U266 cells at a full range of concentration points, as well as for expression and monomer content, as well as sequence identity to human germline (Table 13).

Table 13:

hSG16.45	VH	VK	hBCMA	IgG mg	aSEC%	VH%	VK%	INN
1	VH1	VK1	+++	0.67	94.5	86.5	79.2	Mix
3	VH1	VK3	+++	0.54	94.6	86.5	82.3	Mix
4	VH1	VK5	+++	0.16	76.0	86.5	82.3	Mix
17	VH5	VK1	++	0.64	94.4	88.5	79.2	Mix
18	VH5	VK2	++	0.65	93.7	88.5	86.5	Hu
19	VH5	VK3	++	0.64	94.1	88.5	82.3	Mix

[00187] The VH5 VK2, VH1 VK1 and VH1 VK3 were the best antibodies overall based on binding affinity for human, sequence identity to human germline sequence in both heavy and light chain variable regions, good expression and high percentage of monomers VH1 VK1 and VH1 VK3 have somewhat higher binding (the same as rat or chimeric within experimental error) but lower sequence identity to human germline.

Example 8: Synthesis of a reduced-fucosylated hSG16.17 or hSG16.45 antibody

[00188] The hSG16.17 VH3 VK2 or hSG16.45 VH5 VK2 antibody was expressed in CHO cells. A fucosylation inhibitor, 2-fluorofucose, was included in the cell culture media during the production of antibodies resulted in non- fucosylated antibody. *See, e.g.,* Okeley et al., *Proc. Nat'l Acad. Sci.* 110:5404-55409 (2013). The base media for cell growth was fucose free and 2-fluorofucose was added to the media to inhibit protein fucosylation. *Ibid.* Incorporation of fucose into antibodies was measured by LC-MS via PLRP-S chromatography and electrospray ionization quadropole TOF MS. *Ibid.*

Example 9: In vivo activity of hSG16.17-SEA in SCID or NSG mice

[00189] Figs. 20A-C showed in vivo activity of multi dosed hSG16.17-SEA in MM1S disseminated tumor model in SCID mice. Animals were implanted with MM1S cells IV, and antibody dosing was initiated 9 days post implant. Animal survival was followed over time. N=8 animals per group. BCMA copy# =7,000, CD38 copy# = 14,000. A) 1mg/kg weekly ip for 5 weeks B) 3mg/kg weekly ip for 5 weeks and C) 10 mg/kg weekly ip for 5 weeks. SCID animals contain effector cells to mediate ADCC and ADCP. Data in this figure show that hSG16.17 SEA improves survival comparable to daratumumab (CD38 targeted Ab. Non-binding h00 control showed no activity.

[00190] Figs. 21A-C showed In vivo activity of single dosed hSG16.17-SEA in EJM disseminated tumor model in NSG mice. NSG animals contain no NK cells and minimally active macrophages. Animals were implanted with EJM cells IV, and a single dose of antibody was given ip 5 days post implant. Animal survival was followed over time. N=8 animals per group. BCMA copy#=45,000. CD38 copy# =47,000. CS1 copy# = 14,000. A) 1mg/kg dose B) 3mg/kg dose C) 10 mg/kg dose. Data in this figure show that hSG16.17 SEA increases survival to an equal or

greater extent than daratumumab (CD38 targeted Ab) and elotuzumab (CS1 targeted Ab). WT SG16.17 can also induce increased survival. Non-binding h00 control showed no activity at the highest dose. Since there are minimal effector cells in these animals, activity of WT and SEA hSG16.17 antibodies is likely due to blocking of the APRIL and BAFF proliferation signals.

[00191] Fig. 22 showed in vivo activity of multi dosed hSG16.17-SEA in NCI-H929-luciferase disseminated tumor model in NSG mice. NSG animals were implanted with NCI-H929 luciferase cells. Antibody dosing was initiated 21 days post implant when bioluminescence was observed in the bone marrow. Dosed ip weekly for 5 doses total. N=5 animals per group. BCMA copy#=25,000. CD38 copy# =45,000. CS1 copy# = 3,000. Average luminescence is plotted over time in comparison to untreated and naïve animals. hSG16.17 SEA displayed much better activity compared to daratumumab (CD38 targeted Ab) and elotuzumab (CS1 targeted Ab). The increased luminescence observed in the hSG16.17-SEA 10mg/kg group is driven by a single animal.

[00192] Figs. 23A and 23B showed in vivo activity of single dosed hSG16.17-SEA in NCI-H929-luciferase disseminated tumor model in NSG mice. NSG animals were implanted with NCI-H929 luciferase cells. Antibody dosing was initiated 21 days post injection when bioluminescence was observed in the bone marrow. Dosed once IP. N=5 animals per group. . A) 3mg/kg WT vs SEA antibodies. B) Dose range of hSG16.17 SEA. Data in this figure show that hSG16.17 SEA can be active at 0.3mg/kg single dose and hSG16.17SEA can be more active than its WT (fucosylated) counterpart.

[00193] Figs. 23A and 23B showed in vivo activity of single dosed hSG16.17-SEA in NCI-H929-luciferase disseminated tumor model in NSG mice. NSG animals were implanted with NCI-H929 luciferase cells. Antibody dosing was initiated 21 days post injection when bioluminescence was observed in the bone marrow. Dosed once IP. N=5 animals per group. . A) 3mg/kg WT vs SEA antibodies. B) Dose range of hSG16.17 SEA. Data in this figure show that hSG16.17 SEA can be active at 0.3mg/kg single dose and hSG16.17SEA can be more active than its WT (fucosylated) counterpart. Effects on luminescence translates to prolonged animal survival (data not shown).

[00194] Fig 24 In vivo activity of single dosed hSG16.17-SEA in MOLP-8-luciferase disseminated tumor model in SCID mice. SCID animals were implanted with MOLP-8 luciferase

cells by IV. Antibody dosing was initiated 13 days post injection when bioluminescence was observed in the bone marrow. Dosed once IP. N=5 animals per group. BCMA copy# =2,000. Luminescence is plotted over time. These data show that even with only 2000 BCMA copies the hSG16.17-SEA displays significant antitumor activity. Deglycosylated SEA BCMA antibody, which does not bind Fc γ RII or Fc γ RIII, showed no activity similar to h00 SEA non-binding control. This reveals the importance of Fc mediated activity in this model.

[00195] Figure 25 The SG16.17 SEA antibody displays improved ADCC activity on MM1R target cells in comparison to WT antibody in vitro. NK cells were isolated from PBMCs via negative selection using an EasySep Human NK cell enrichment kit, and resulting CD16+ cells were quantitated. Multiple myeloma MM1R ADCC target cells were labeled with chromium-51 for 1 hr. A dilution series of antibodies was added to the assay plate, followed by target cells (T) and NK effector cells (E) at a 13:1 E:T ratio. Lysis was calculated based on total and spontaneous release controls after 4hrs at 37°C. These data show a significant improvement in ADCC activity of the afucosylated SEA SG16.17 antibody over WT antibody as well as clinical antibodies, daratumumab and elotuzumab.

Although the invention has been described in detail for purposes of clarity of understanding, certain modifications may be practiced within the scope of the appended claims. All publications including accession numbers, websites and the like, and patent documents cited in this application are hereby incorporated by reference in their entirety for all purposes to the same extent as if each were so individually denoted. To the extent difference version of a sequence, website or other reference may be present at different times, the version associated with the reference at the effective filing date is meant. The effective filing date means the earliest priority date at which the accession number at issue is disclosed. Unless otherwise apparent from the context any element, embodiment, step, feature or aspect of the invention can be performed in combination with any other.

WHAT IS CLAIMED IS:

1. A humanized, chimeric or veneered antibody, which is a humanized, chimeric or veneered form of an antibody deposited as ATCC PTC-6937.
2. The antibody of claim 1, comprising a mature heavy chain variable region having at least 90% sequence identity to hSG16.17 VH3 (SEQ ID NO: 13) and a mature light chain variable region having at least 90% sequence identity to hSG16.17 VK2 (SEQ ID NO: 19).
3. The antibody of claim 2, comprising a mature heavy chain variable region having at least 95% sequence identity to hSG16.17 VH3 (SEQ ID NO: 13) and a mature light chain variable region having at least 95% sequence identity to hSG16.17 VK2 (SEQ ID NO: 19).
4. The antibody of any preceding claim, comprising the three Kabat CDRs (SEQ ID NOs: 60-62) of hSG16.17 VH3 (SEQ ID NO: 13) and three Kabat CDRs (SEQ ID NOs: 90-92) of hSG16.17 VK2 (SEQ ID NO: 19) provided that position H58 can be occupied by N or K, position H60 can be occupied by A or N, position H61 can be occupied by Q or E, position H62 can be occupied by K or N, position H64 can be occupied by Q or K, position H65 can be occupied by G or T, position L24 can be occupied by R or L, and position L53 can be occupied by S or R .
5. The antibody of any preceding claim comprising the three Kabat CDRs (SEQ ID NOs: 60-62) of hSG16.17 VH3 (SEQ ID NO: 13) and three Kabat CDRs (SEQ ID NOs: 90-92) of hSG16.17 VK2 (SEQ ID NO: 19).
6. The antibody of any preceding claim, wherein positions H20, H48, H69, H71, H73, H76, H80, H88, H91 and H93 are occupied by L, I, M, A, K, N, V, A, F, and T respectively, and positions L46, L48 and L87 are occupied by V, V and F respectively.
7. The antibody of claim 1, wherein the mature heavy chain variable has the sequence of hSG16.17 VH3 (SEQ ID NO: 13) and the mature light chain variable region has the sequence of hSG16.17 VK2 (SEQ ID NO: 19).
8. The antibody of any preceding claim, wherein the mature heavy chain variable region is fused to a heavy chain constant region and the mature light chain variable region is fused to a light chain constant region.

9. The antibody of claim 6, wherein the heavy chain constant region is a mutant form of natural human constant region which has reduced binding to an Fc_Y receptor relative to the natural human constant region.

10. The antibody of claim 8 or 9, wherein the heavy chain constant region is of IgG1 isotype.

11. The antibody of claims 8, wherein the heavy chain constant region has an amino acid sequence comprising SEQ ID NO: 5 and the light chain constant region has an amino acid sequence comprising SEQ ID NO: 3.

12. The antibody of claim 8, wherein the heavy chain constant region has an amino acid sequence comprising SEQ ID NO:7 (S239C) and the light chain constant region has an amino acid sequence comprising SEQ ID NO:3.

13. The antibody of any preceding claim, which is a naked antibody.

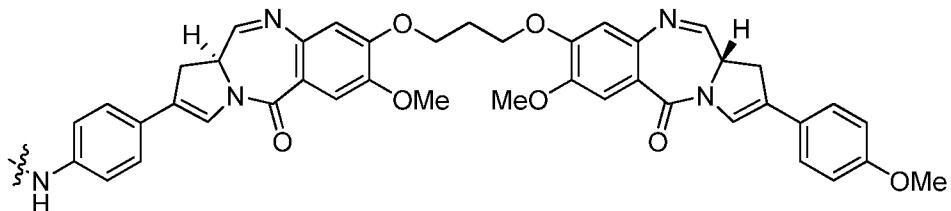
14. The antibody of any of claims 1-12, wherein the antibody is conjugated to a cytotoxic or cytostatic agent.

15. The antibody of claim 14, wherein the antibody is conjugated to a cytotoxic agent.

16. The antibody of claim 15, wherein the cytotoxic agent is conjugated to the antibody via an enzyme cleavable linker.

17. The antibody of claim 15 or 16, wherein the cytotoxic agent is a DNA minor groove binder.

18. The antibody of claim 17, wherein the cytotoxic agent has the formula



19. The antibody of claim 15 or 16, wherein the cytotoxic agent is MMAE or MMAF.

20. A pharmaceutical composition comprising an antibody of any preceding claim and a pharmaceutically acceptable carrier.

21. A method of treating a patient having or at risk of having a cancer that expresses BCMA comprising administering to the patient an effective regime of an antibody of any of the preceding claims.
22. The method of claim 20, wherein the cancer is a hematological cancer.
23. The method of claim 22, wherein the hematological cancer is a myeloma, leukemia or a lymphoma.
24. The method of claim 22, wherein the hematological cancer is multiple myeloma.
25. The method of claim 22, wherein the hematological cancer is non-Hodgkin's lymphoma (NHL) or Hodgkin's lymphoma.
26. The method of claim 22, wherein the hematological cancer is myelodysplastic syndromes (MDS), myeloproliferative syndromes (MPS), Waldenström's macroglobulinemia or Burkett's lymphoma.
27. A method of treating a patient having or at risk of having an immune disorder mediated by immune cells expressing BCMA comprising administering to the patient an effective regime of a humanized antibody of any of the preceding claims.
28. The method of claim 27, which is a B cell mediated disorder.
29. The method of claim 27, wherein the immune disorder is rheumatoid arthritis, systemic lupus E (SLE), Type I diabetes, asthma, atopic dermatitis, allergic rhinitis, thrombocytopenic purpura, multiple sclerosis, psoriasis, Sjögren's syndrome, Hashimoto's thyroiditis, Grave's disease, primary biliary cirrhosis, Wegener's granulomatosis, tuberculosis, and graft versus host disease.
30. A humanized, chimeric or veneered antibody, which is a humanized, chimeric or veneered form of the rat SG16.45 antibody having a mature heavy chain variable region of SEQ ID NO: 23 and a mature light chain variable region of SEQ ID NO: 33.
31. The antibody of claim 30, comprising a mature heavy chain variable region having at least 90% sequence identity to hSG16.45 VH5 (SEQ ID NO: 31) and a mature light chain variable region having at least 90% sequence identity to hSG16.45 VK2 (SEQ ID NO: 36).

32. The antibody of claim 31, comprising a mature heavy chain variable region having at least 95% sequence identity to hSG16.45 VH5 (SEQ ID NO: 31) and a mature light chain variable region having at least 95% sequence identity to hSG16.45 VK2 (SEQ ID NO: 36).

33. The antibody of any of claims 30-32, comprising the three Kabat CDRs (SEQ ID NOs: 152-154) of hSG16.45 VH5 (SEQ ID NO: 31) and three Kabat CDRs (SEQ ID NOs: 179-181) of hSG16.45 VK2 (SEQ ID NO: 36) provided that position H50 can be occupied by A or S, position L24 can be occupied by R or L and position L26 can be occupied by S or T.

34. The antibody of any of claims 30-33 comprising the three Kabat CDRs (SEQ ID NOs: 152-154) of hSG16.45 VH5 (SEQ ID NO: 31) and three Kabat CDRs (SEQ ID NOs: 179-181) of hSG16.45 VK2 (SEQ ID NO: 36). , .

35. The antibody of any one of claims 30-34, wherein positions H30, H93 and H94 are occupied by N, T and S respectively.

36. The antibody of claim 30, wherein the mature heavy chain variable region has the sequence of hSG16.45 VH5 (SEQ ID NO: 31) and the mature light chain variable region has the sequence of hSG16.45 VK2 (SEQ ID NO: 36) or the mature heavy chain variable region has the sequence of hSG16.45 VH1 (SEQ ID NO: 27) and the mature light chain variable region has the sequence of hSG16.45 VK1 (SEQ ID NO: 35) or the mature heavy chain variable region has the sequence of hSG16.45 VH1 (SEQ ID NO: 27) and the mature light chain variable region has the sequence of hSG16.45 VK3 (SEQ ID NO: 37).

37. The antibody of any one of claims 30-36, wherein the mature heavy chain variable region is fused to a heavy chain constant region and the mature light chain variable region is fused to a light chain constant region.

38. The antibody of claim 37, wherein the heavy chain constant region is a mutant form of natural human constant region which has reduced binding to an Fc_Y receptor relative to the natural human constant region.

39. The antibody of claim 37 or 38, wherein the heavy chain constant region is of IgG1 isotype.

40. The antibody of claims 37, wherein the heavy chain constant region has an amino acid sequence comprising SEQ ID NO: 5 and the light chain constant region has an amino acid sequence comprising SEQ ID NO: 3.

41. The antibody of claim 37, wherein the heavy chain constant region has an amino acid sequence comprising SEQ ID NO:7 (S239C) and the light chain constant region has an amino acid sequence comprising SEQ ID NO:3.

42. The antibody of any one of claims 30-41, which is a naked antibody.

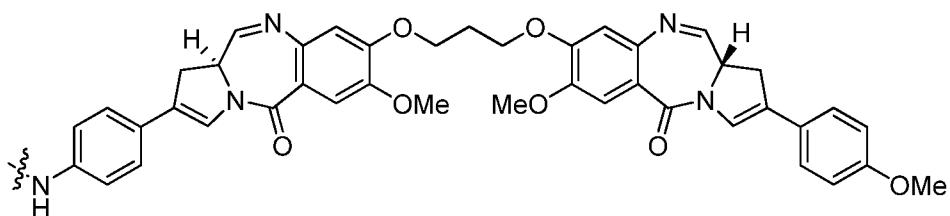
43. The antibody of any one of claims 30-41, wherein the antibody is conjugated to a cytotoxic or cytostatic agent.

44. The antibody of claim 43, wherein the antibody is conjugated to a cytotoxic agent.

45. The antibody of claim 44, wherein the cytotoxic agent is conjugated to the antibody via an enzyme cleavable linker.

46. The antibody of claim 43 or 44, wherein the cytotoxic agent is a DNA minor groove binder.

47. The antibody of claim 46, wherein the cytotoxic agent has the formula



48. The antibody of claim 44 or 45, wherein the cytotoxic agent is MMAE or MMAF.

49. The antibody of any preceding claim, wherein less than 5% of N-glycoside linked sugar chains at an asn residue at EU position 297 of heavy chain constant region include fucose or an analog thereof in which cells expressing the antibody were cultured to reduce fucosylation of the antibody.

50. A pharmaceutical composition comprising an antibody of any one of claims 30-48 and a pharmaceutically acceptable carrier.

51. A method of treating a patient having or at risk of having a cancer that expresses BCMA comprising administering to the patient an effective regime of a humanized antibody of any one of claims 30-49.

52. The method of claim 51, wherein the cancer is a hematological cancer.
53. The method of claim 52, wherein the hematological cancer is a myeloma, leukemia or a lymphoma.
54. The method of claim 52, wherein the hematological cancer is multiple myeloma.
55. The method of claim 52, wherein the hematological cancer is non- Hodgkin's lymphoma (NHL) or Hodgkin's lymphoma.
56. The method of claim 52, wherein the hematological cancer is myelodysplastic syndromes (MDS), myeloproliferative syndromes (MPS), Waldenström's macroglobulinemia or Burkett's lymphoma.
57. A method of treating a patient having or at risk of having an immune disorder mediated by immune cells expressing BCMA comprising administering to the patient an effective regime of an antibody of any of the preceding claims.
58. The method of claim 56, which is a B cell mediated disorder.
59. The method of claim 56, wherein the immune disorder is rheumatoid arthritis, systemic lupus E (SLE), Type I diabetes, asthma, atopic dermitis, allergic rhinitis, thrombocytopenic purpura, multiple sclerosis, psoriasis, Sjorgren's syndrome, Hashimoto's thyroiditis, Grave's disease, primary biliary cirrhosis, Wegener's granulomatosis, tuberculosis, and graft versus host disease.
60. A humanized antibody the specifically binds to the human BCMA protein, the antibody comprising a mature heavy chain variable region having at least 90% sequence identity to hSG16.17 VH3 (SEQ ID NO: 13) and a mature light chain variable region having at least 90% sequence identity to hSG16.17 VK2 (SEQ ID NO: 19).
61. The antibody of claim 60, comprising a mature heavy chain variable region having at least 95% sequence identity to hSG16.17 VH3 (SEQ ID NO: 13) and a mature light chain variable region having at least 95% sequence identity to hSG16.17 VK2 (SEQ ID NO: 19).
62. The antibody of claim 60, comprising the three Kabat CDRs (SEQ ID NOs: 60-62) of hSG16.17 VH3 (SEQ ID NO: 13) and three Kabat CDRs (SEQ ID NOs: 90-92) of hSG16.17 VK2 (SEQ ID NO: 19) provided that position H58 can be occupied by N or K, position H60 can be occupied by A or N, position H61 can be occupied by Q or E, position H62 can be occupied by K or N,

position H64 can be occupied by Q or K, position H65 can be occupied by G or T, position L24 can be occupied by R or L, and position L53 can be occupied by S or R .

63. The antibody of claim 60 comprising the three Kabat CDRs (SEQ ID NOs: 60-62) of hSG16.17 VH3 (SEQ ID NO: 13) and three Kabat CDRs (SEQ ID NOs: 90-92) of hSG16.17 VK2 (SEQ ID NO: 19).

64. The antibody of claim 60, wherein positions H20, H48, H69, H71, H73, H76, H80, H88, H91 and H93 are occupied by L, I, M, A, K, N, V, A, F, and T respectively, and positions L46, L48 and L87 are occupied by V, V and F respectively.

65. The antibody of claim 60, wherein the mature heavy chain variable has the sequence of hSG16.17 VH3 (SEQ ID NO: 13) and the mature light chain variable region has the sequence of hSG16.17 VK2 (SEQ ID NO: 19).

66. The antibody of claim 60, wherein the mature heavy chain variable region is fused to a heavy chain constant region and the mature light chain variable region is fused to a light chain constant region.

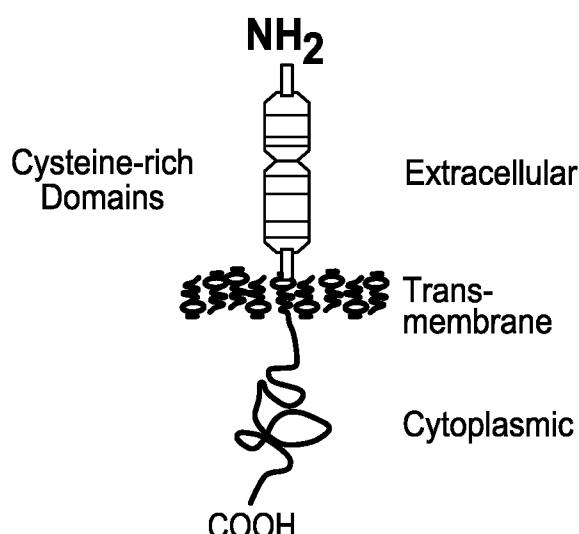
67. The antibody of claim 65, wherein the mature heavy chain variable region is fused to a heavy chain constant region and the mature light chain variable region is fused to a light chain constant region.

68. A pharmaceutical composition comprising an antibody of claim 67 and a pharmaceutically acceptable carrier.

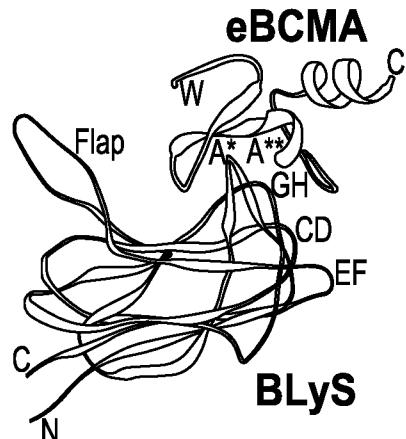
69. The pharmaceutical composition of claim 68, wherein less than about 10% of the antibodies have core fucosylation by fucose or a fucose analogue.

70. The pharmaceutical composition of claim 68, wherein less than about 5% of the antibodies have core fucosylation by fucose or a fucose analogue.

71. The pharmaceutical composition of claim 69, wherein about 2% of the antibodies have core fucosylation by fucose or a fucose analogue.



Jelinek and Darce (2005) *Curr Dir Autoimmun.* Basel, Karger 8: 266-288



Wallweber (2004) *J Mol Biol* 343: 283-290

Fig. 1A

Fig. 1B

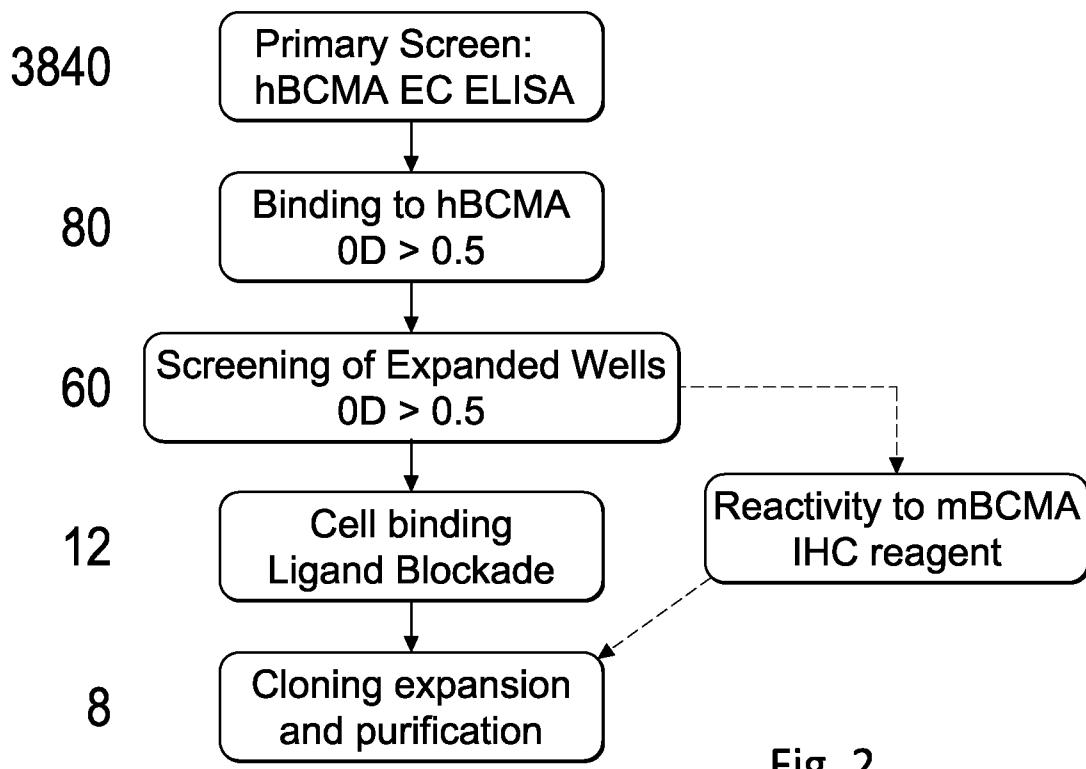


Fig. 2

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Fig. 3

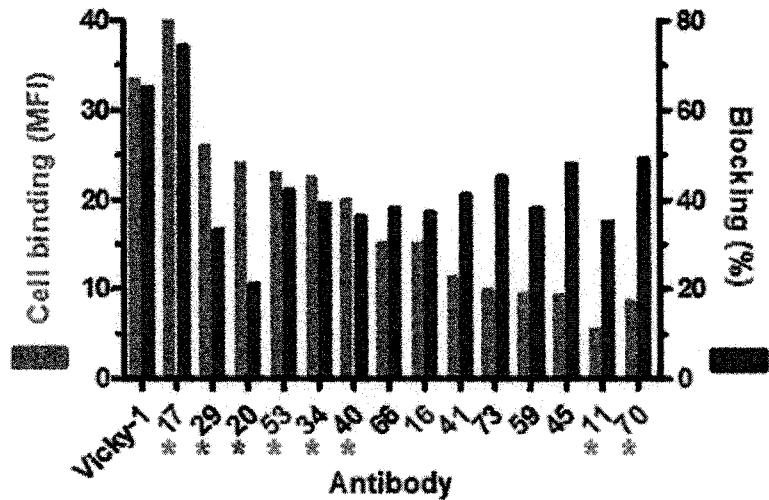
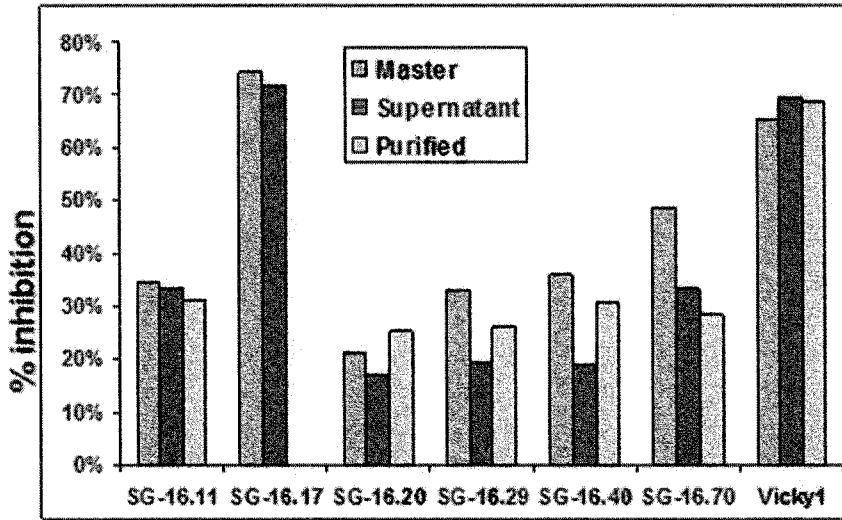


Fig. 4



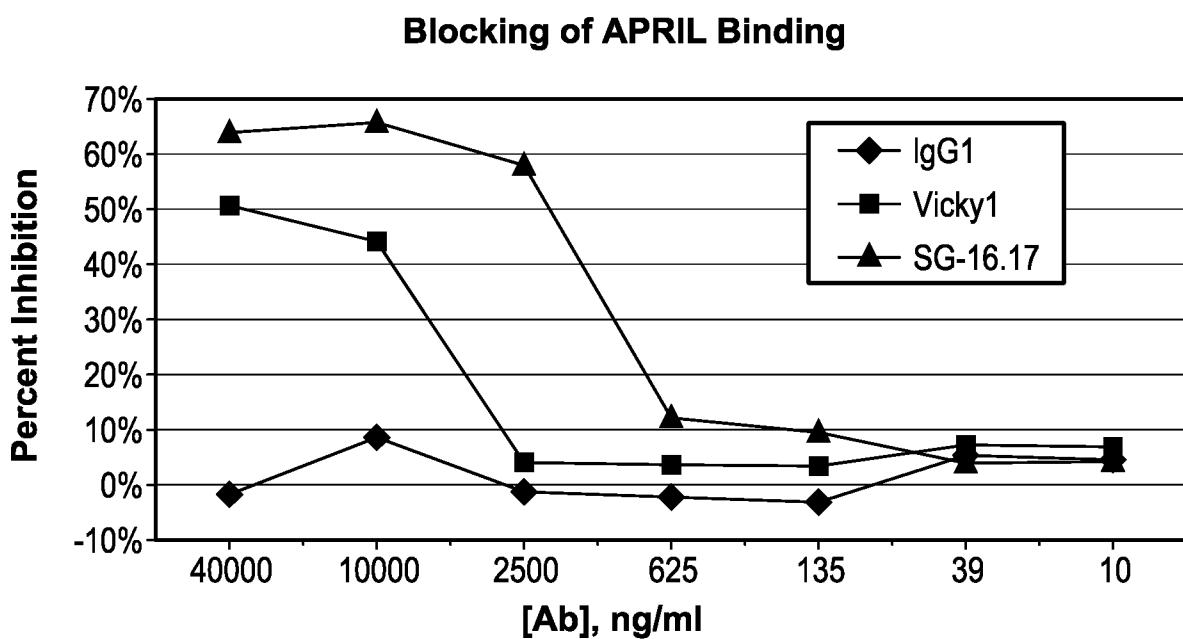


Fig. 5

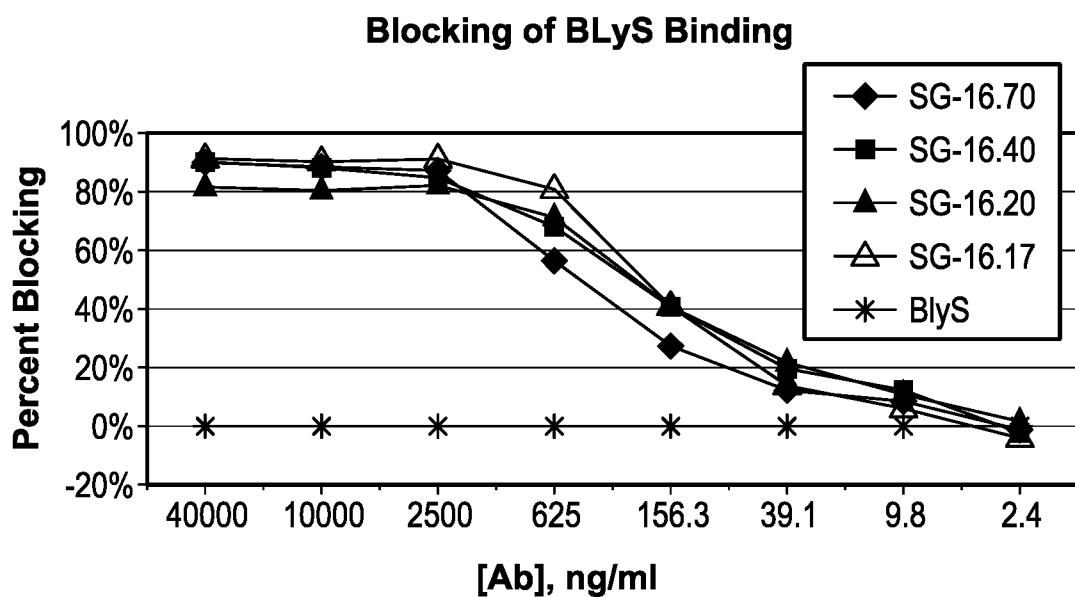


Fig. 6

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Fig. 7: Alignment of hSG16.17 Heavy Chain Variants with Human VH Acceptor Sequence, HV1-2/HJ3.

Fig. 8: Alignment of hSG16.17 Heavy Chain Variants with Human VH Acceptor Sequence; HV1-46/HJ3.

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Fig. 9: Alignment of hSG16.17 Heavy Chain Variants

	10	20	30	40	50	
hSG16.17 vh1					
hSG16.17 vh2	QVQLVQSRAEVKKPGASVQLSCKASGYFTDYYIHWVRQAPGQGLEWIGYINPNSGYTKYG.....M.....R.....N.....	
hSG16.17 vh3G.....G.....N.....	
hSG16.17 vh4G.....	V.....	M.....N.....	
hSG16.17 vh5G.....	V.....	
hSG16.17 vh6M.....I.....S.....		
<u>Kabat</u> CDRs		*****		*****		
IMGT CDRs		++++++		++++++		
	60	70	80	90	100	110
hSG16.17 vh1					
hSG16.17 vh2	NENFKTRATMTADKSINTAYVELSRLRSDDTAVYFCTRYMWERVTGFFDFWGQGMVTVSS	AQK.QG.V.....
hSG16.17 vh3	AQK.QG.....
hSG16.17 vh4	AQK.QG.....
hSG16.17 vh5	T.....S...E.....
hSG16.17 vh6	AQK.QG.V.....T.....S...E.....
<u>Kabat</u> CDRs	*****		*****	*****	*****	
IMGT CDRs			++++++	++++++	++++++	

Fig. 10: Alignment of hSG16.17 Light Chain Variants with Human VK Acceptor Sequence; KV1-12/KJ5.

	10	20	30	40	50	60
Rat SG16.17 VK
Hu KV1-12/KJ5A.L...L.E.T.S.E.L...E.D...S...S.QV.V.TT.R...D...
hSG16.17 VK2L...E.D...V.V.TT...
hSG16.17 VK3E.D...V.V.TT...
hSG16.17 VK4L...E.D...V.V.TT.R...
hSG16.17 VK5E.D...S...V.V.TT...
<u>Kabat</u> CDRs	*****					*****
IMGT CDRs	+++++					+++
	70	80	90	100		
Rat SG16.17 VK
Hu KV1-12/KJ5R.S.K.IVM...E.D.F.QQTYYKFPPT...A.RLDL...			
hSG16.17 VK2F.QQTYYKFPPT...				
hSG16.17 VK3F.QQTYYKFPPT...				
hSG16.17 VK4M...D.F.QQTYYKFPPT...				
hSG16.17 VK5F.QQTYYKFPPT...				
<u>Kabat</u> CDRs	*****					
IMGT CDRs	++++++					

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Fig. 11: Alignment of hSG16.17 Light Chain Variants

Fig. 12

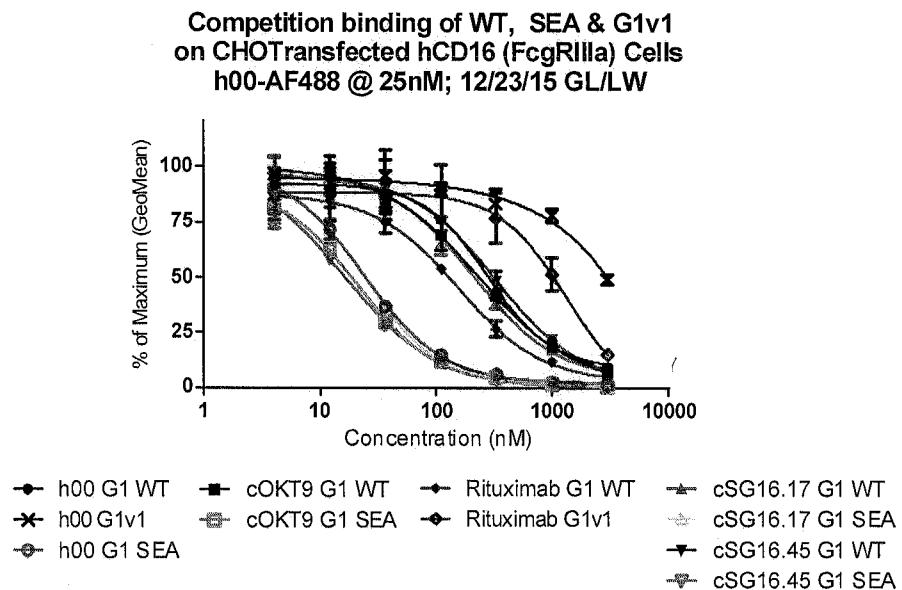


Fig. 13

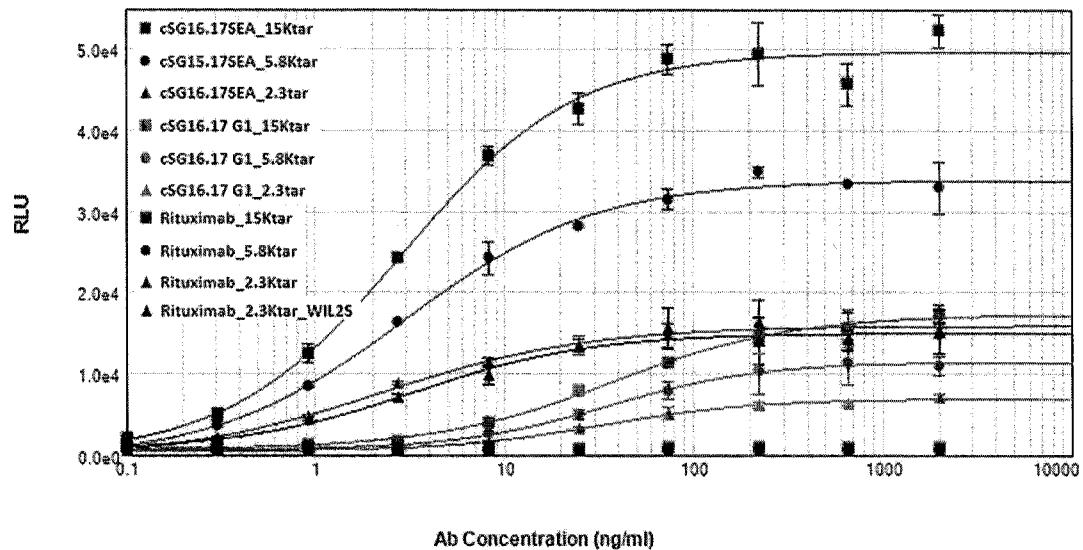


Fig. 14: Alignment of hSG16.45 Heavy Chain Variants with Human HV Acceptor Sequence, HV3-23/HJ3.

	10	20	30	40	50	
Rat SG16.45 vHV.....R.....K.....V.....NDHW.T.I.....R.....I.S.TNT..A...	EVQLLESGGGLVQPGGSLRLSCAASGFTFSSYAMSWVRQAPGKGLEWVSAISGSGGSTYY			
Hu HV3-23/HJ3						
hSG16.45 vH1						NDHW.T.I.....I.S.TNT..A...
hSG16.45 vH2						NDHW.T.I.....I...TNT..A...
hSG16.45 vH3						NDHW.T.I.....I...TNT..A...
hSG16.45 vH4						NDHW.T.....I.S.TNT..A...
Kabat CDRs					*****	*****
IMGT CDRs				++++++		++++++
	60	70	80	90	101	110
Rat SG16.45 vH	L.....A.S.....S.....T.....TSPGLYFDY.....V.....	ADSVKGRFTISRDNSKNTLYLQMNSLRAEDTAVYYCAK-----WGQGTMVTVSS			
Hu HV3-23/HJ3						
hSG16.45 vH1						TSPGLYFDY.....V.....
hSG16.45 vH2						TSPGLYFDY.....V.....
hSG16.45 vH3						TSPGLYFDY.....V.....
hSG16.45 vH4						TSPGLYFDY.....V.....
Kabat CDRs	*****				*****	
IMGT CDRs					++++++	++++++

Figure 15: Alignment of hSG16.45 Heavy Chain Variants with Human HV Acceptor Sequence; HV3-74/HJ3.

Figure 16: Alignment of hSG16.45 Heavy Chain Variants with Human HV Acceptor Sequence; HV3-9/HJ3.

	10	20	30	40	50	
Rat SG16.45 VH	EVQLVESGGGLVQPGRSLKLSCVASGFTFNDHWMWTIRQAPGRGLEWISSITNTGGATYY				
Hu HV3-9/HJ3	R...A.....D.YA.H.V....K....V.G.SWNS.SIG.				
hSG16.45 VH6	R...A.....V.....K....V.G.....				
Kabat CDRs			*****		*****	
IMGT CDRs			++++++		++++++	
	60	70	80	90	101	110
Rat SG16.45 VH	LDSVKGRFTISRDNAKSTLYLQMNSLRSEDTATYYCTSPGLYFDYWQGVMTVSS				
Hu HV3-9/HJ3	A.....	NS.....A....L...AK-----T.....				
hSG16.45 VH6	A.....	NS.....A....L.....T.....				
Kabat CDRs	*****			*****		
IMGT CDRs			++++++		++++++	

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Figure 17: Alignment of hSG16.45 Heavy Chain Variants.

	10	20	30	40	50	
hSG16.45 vh1					
hSG16.45 vh2	EVQLLESGGGLVQPGGSLRLSCAASGFTFNDHWMTWIRQAPGKGLEWISSITNTGGATYY					
hSG16.45 vh3					
hSG16.45 vh4					
hSG16.45 vh5					
hSG16.45 vh6					
Kabat CDRs				V		
IMGT CDRs				V	V.V	
				V	V.G	
				*****	*****	
				++++++	++++++	
	60	70	80	90	101	110
hSG16.45 vh1					
hSG16.45 vh2	LDSVKGRFTISRDN SKNTLYLQMNSLRAEDTAVYYCTSPGLYFDYWGQGVMTVSS					
hSG16.45 vh3	A.....					
hSG16.45 vh4	A.....					
hSG16.45 vh5	A.....					
hSG16.45 vh6	A.....					
Kabat CDRs	A.....					
IMGT CDRs	A.....					
	*****				*****	
					++++++	

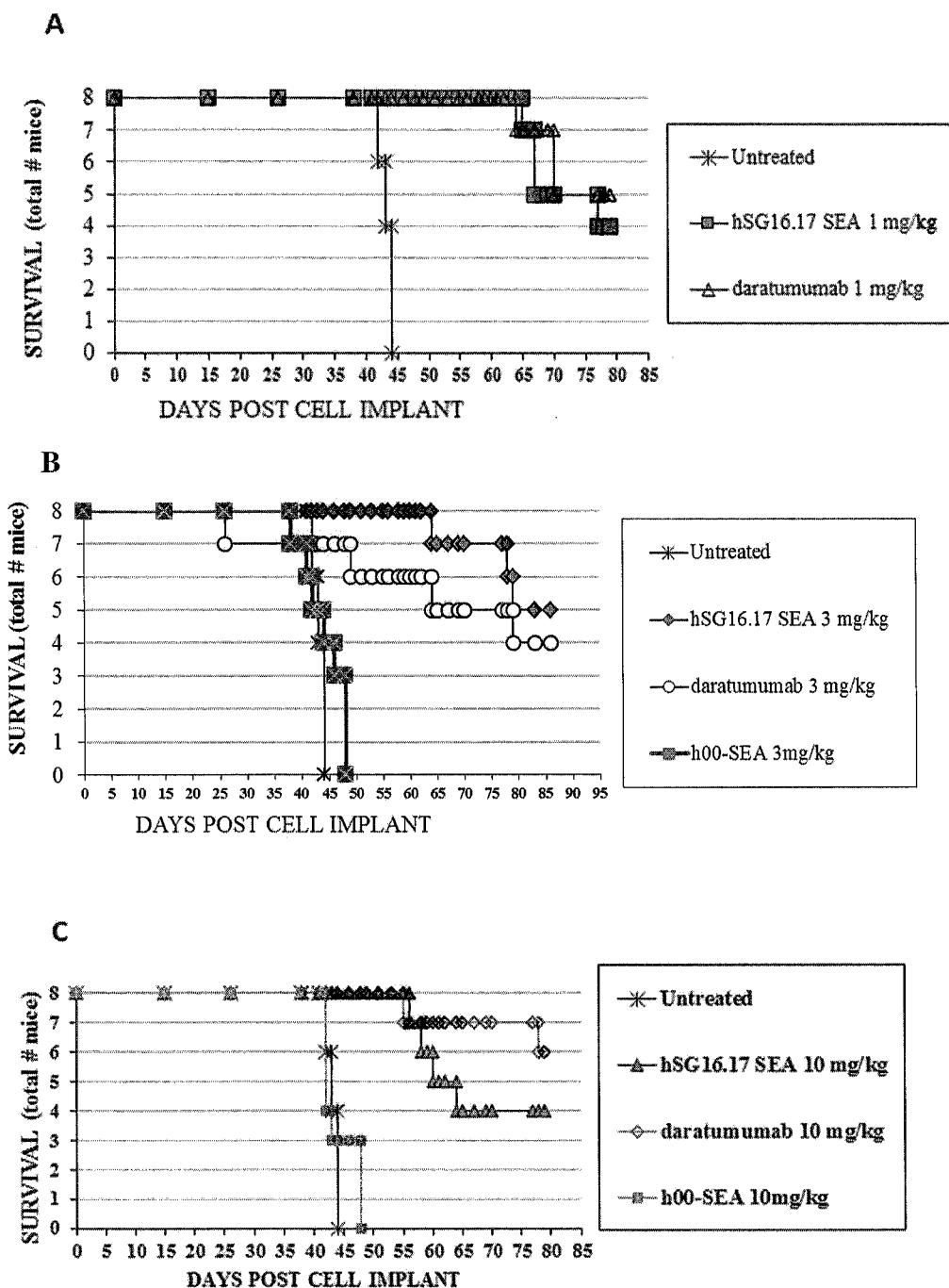
Fig. 18: Alignment of hSG16.45 Light Chain Variants with Human KV Acceptor Sequence; KV3-20/KJ2.

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Fig. 19: Alignment of hSG16.45 Light Chain Variants.

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Fig. 20: In Vivo Activity of Multi Dosed hSG16.17-SEA in MM1S Disseminated Tumor Model in SCID Mice.



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Fig. 21. In Vivo Activity of Single Dosed hSG16.17-SEA in EJM Disseminated Tumor Model in NSG Mice.

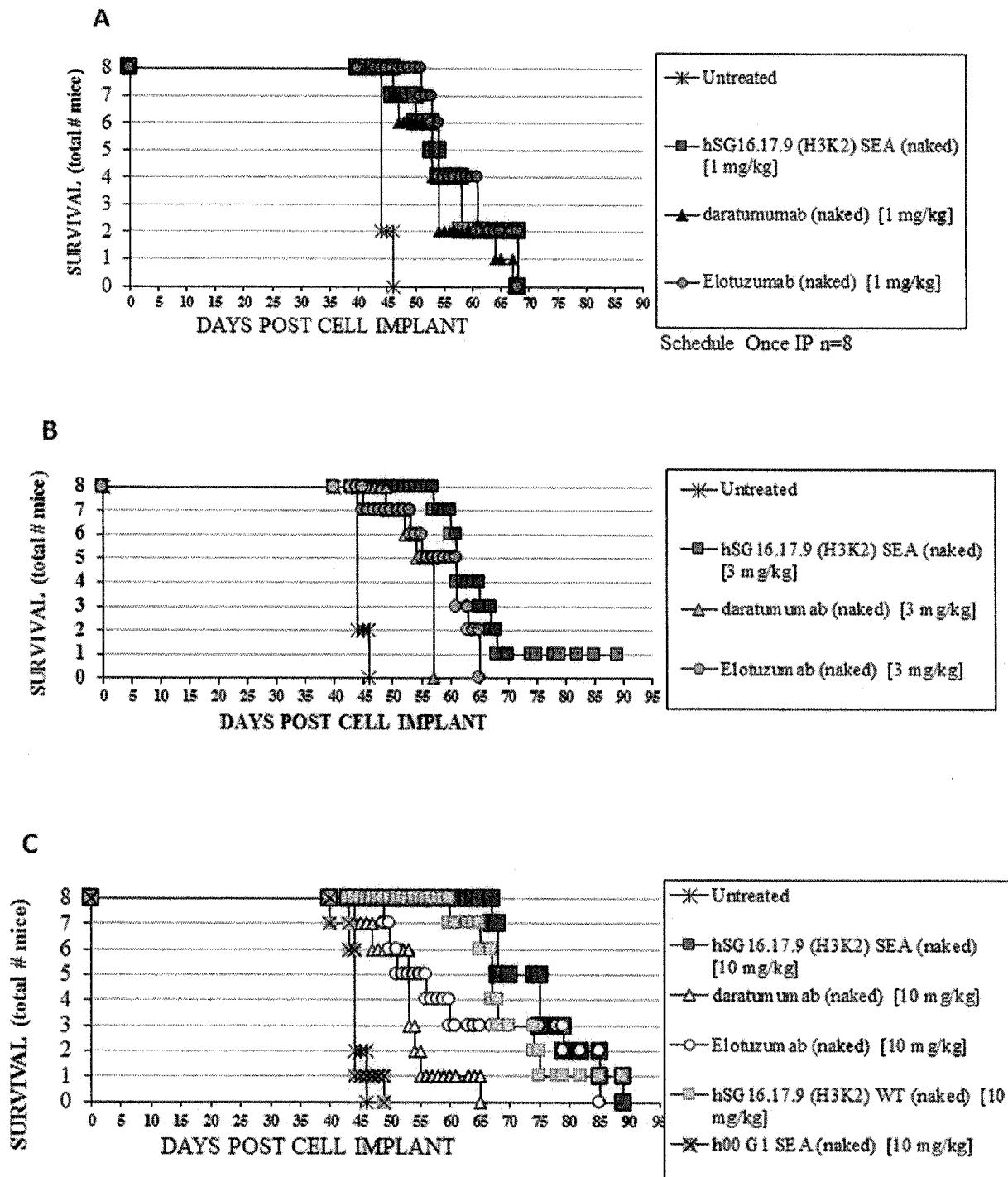


Fig. 22. In Vivo Activity of Multi Dosed hSG16.17-SEA in NCI-H929-luciferase Disseminated Tumor Model in NSG Mice.

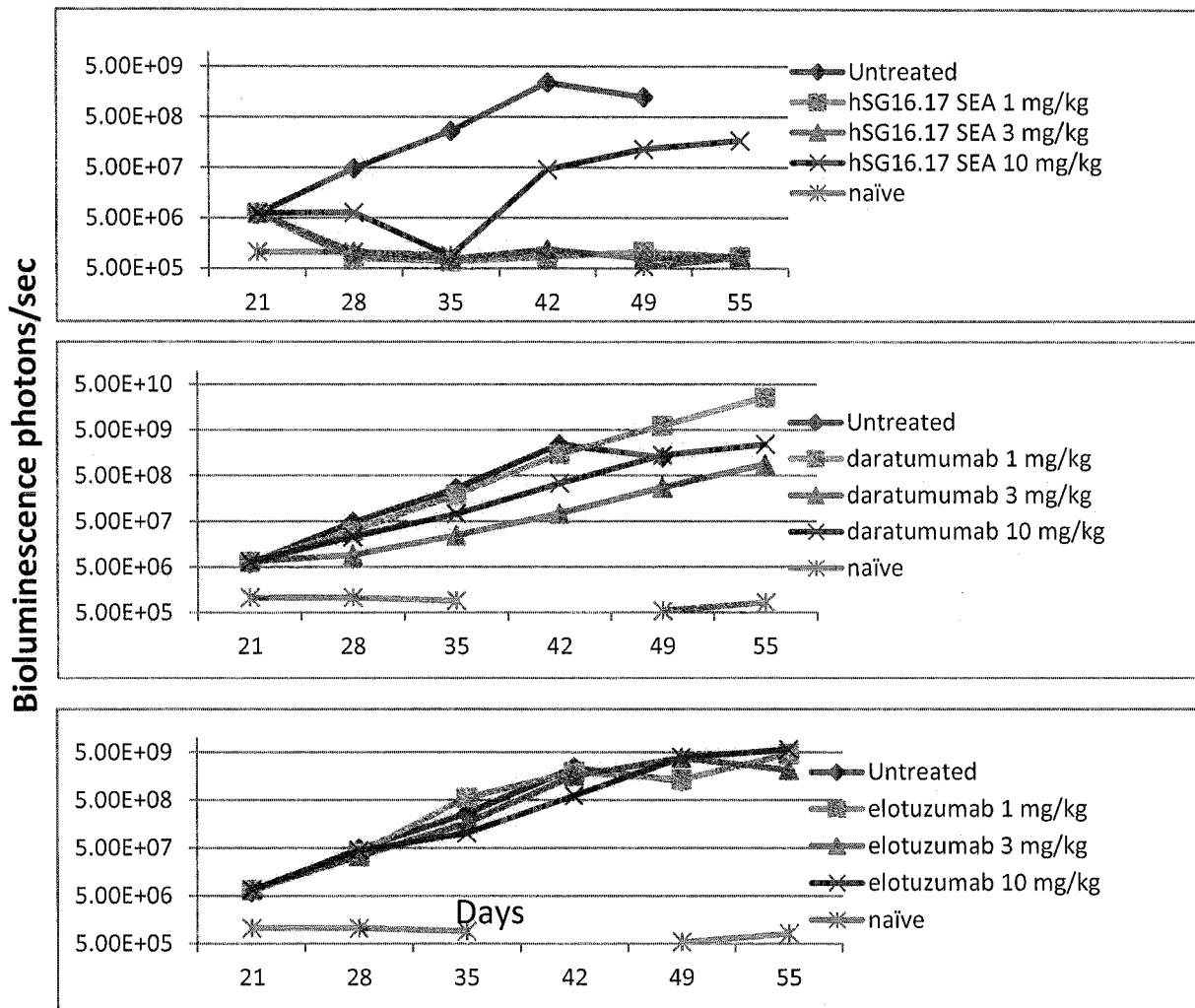


Fig. 23. In Vivo Activity of Single Dosed hSG16.17-SEA in NCI-H929-luciferase Disseminated Tumor Model in NSG Mice.

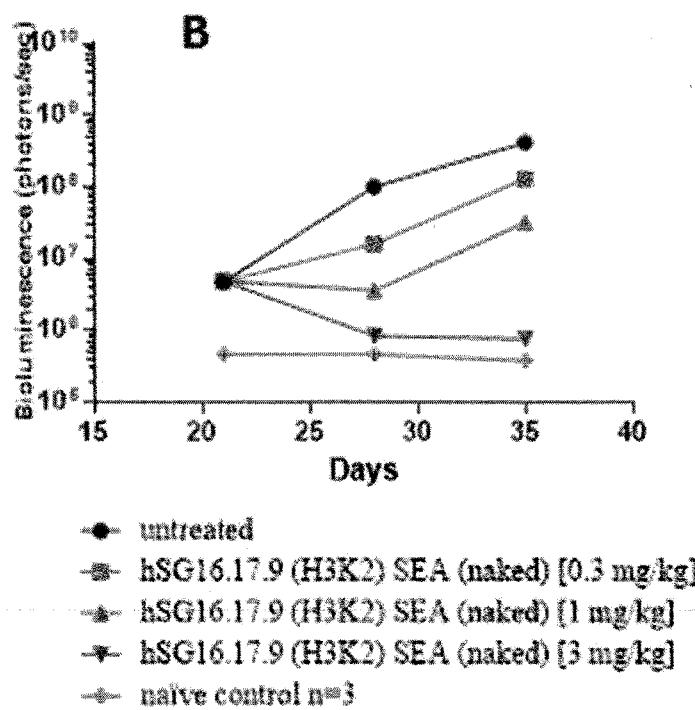
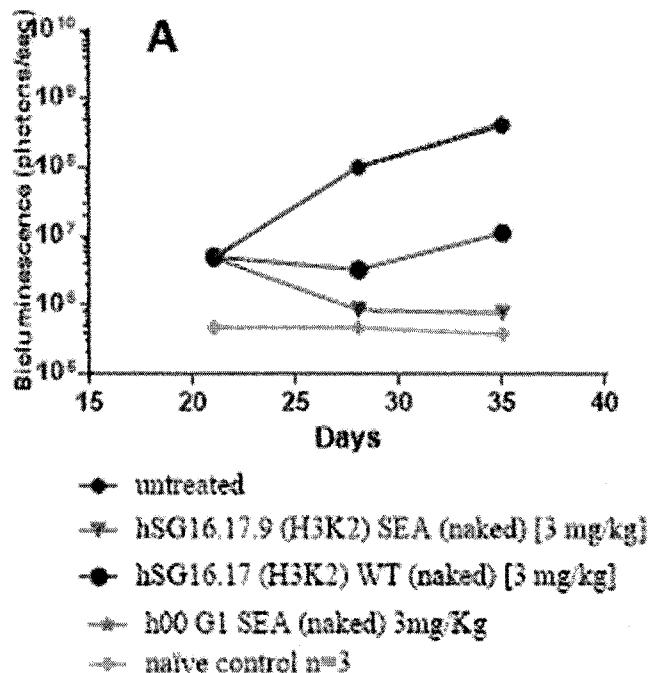
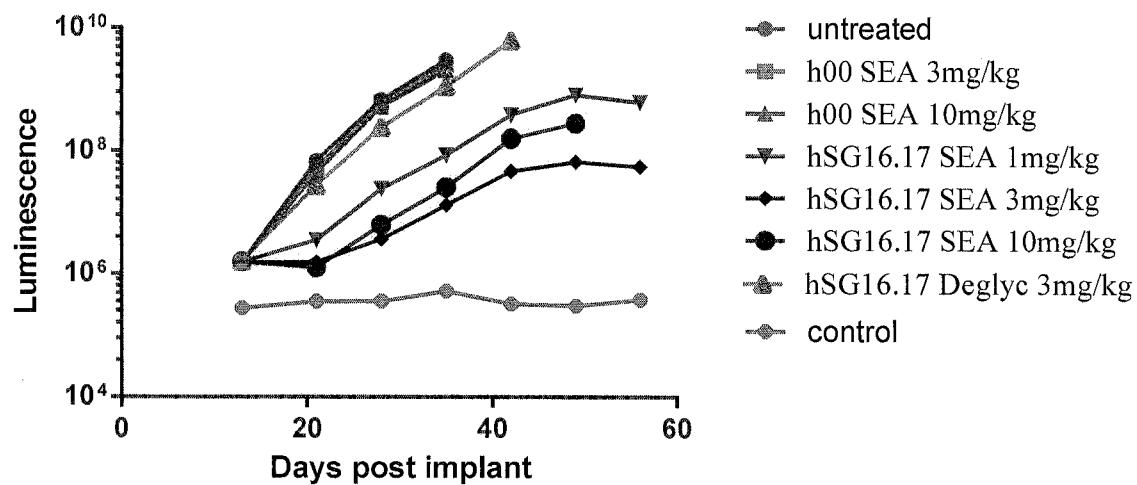
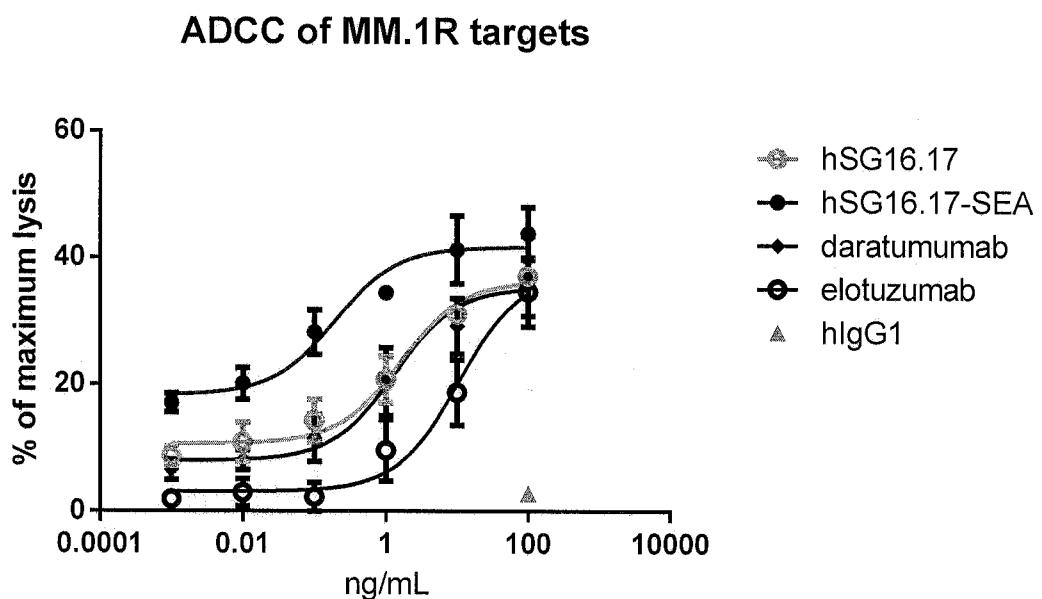


Fig. 24. In Vivo Activity of Single Dosed hSG16.17-SEA in MOLP-8-luciferase Disseminated Tumor Model in SCID Mice.



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Fig. 25. The SG16.17 SEA Antibody Displays Improved ADCC Activity on MM1R Target Cells.



INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2017/018177

A. CLASSIFICATION OF SUBJECT MATTER

IPC(8) - A61K 39/395; A61K 45/06; C07K 16/28; C07K 16/30 (2017.01)

CPC - A61K 39/39558; A61K 45/06; A61K 2039/505; C07K 16/2809; C07K 16/2878; C07K 16/3061; C07K 2317/21; C07K 2317/31; C07K 2317/33; C07K 2317/34; C07K 2317/515; C07K 2317/565; C07K 2317/76; C07K 2317/92 (2017.02)

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

See Search History document

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

USPC - 424/130.1; 424/131.1; 424/133.1; 424/139.1; 435/328; 435/331; 530/387.1; 530/387.3; 536/23.53 (keyword delimited)

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

See Search History document

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 2010/104949 A2 (BIOGEN IDEC MA INC. et al) 16 September 2010 (16.09.2010) entire document	1-4, 7, 30-33, 36, 60-71
A	WO 2015/158671 A1 (CELLLECTIS) 22 October 2015 (22.10.2015) entire document	1-4, 7, 30-33, 36, 60-71
A	RYAN et al. "Antibody targeting of B-cell maturation antigen on malignant plasma cells," Molecular Cancer Therapeutics, 19 November 2007 (19.11.2007), Vol. 6, Iss 11, Pgs. 3009-3018. entire document	1-4, 7, 30-33, 36, 60-71
A	WO 2012/163805 A1 (GLAXO GROUP LIMITED et al) 06 December 2012 (06.12.2012) entire document	1-4, 7, 30-33, 36, 60-71
A	TAI et al. "Targeting B-cell maturation antigen in multiple myeloma," Immunotherapy, 15 September 2015 (15.09.2015), Vol. 7, No. 11, Pgs. 1187-1199. entire document	1-4, 7, 30-33, 36, 60-71

Further documents are listed in the continuation of Box C.

See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"E" earlier application or patent but published on or after the international filing date

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"O" document referring to an oral disclosure, use, exhibition or other means

"&" document member of the same patent family

"P" document published prior to the international filing date but later than the priority date claimed

Date of the actual completion of the international search

17 April 2017

Date of mailing of the international search report

08 MAY 2017

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PCT OSP: 571-272-7774

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2017/018177

Box No. I Nucleotide and/or amino acid sequence(s) (Continuation of item 1.c of the first sheet)

1. With regard to any nucleotide and/or amino acid sequence disclosed in the international application, the international search was carried out on the basis of a sequence listing:
 - a. forming part of the international application as filed:
 in the form of an Annex C/ST.25 text file.
 on paper or in the form of an image file.
 - b. furnished together with the international application under PCT Rule 13*ter*.1(a) for the purposes of international search only in the form of an Annex C/ST.25 text file.
 - c. furnished subsequent to the international filing date for the purposes of international search only:
 in the form of an Annex C/ST.25 text file (Rule 13*ter*.1(a)).
 on paper or in the form of an image file (Rule 13*ter*.1(b) and Administrative Instructions, Section 713).
2. In addition, in the case that more than one version or copy of a sequence listing has been filed or furnished, the required statements that the information in the subsequent or additional copies is identical to that forming part of the application as filed or does not go beyond the application as filed, as appropriate, were furnished.

3. Additional comments:

SEQ ID NOs:3, 5, 7, 13, 19, 23, 27, 31, 33, 35, 36, and 37 were searched.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2017/018177

Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:

2. Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. Claims Nos.: 5, 6, 8-29, 34, 35, 37-59 because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box No. III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying additional fees, this Authority did not invite payment of additional fees.
3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee.
- The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation.
- No protest accompanied the payment of additional search fees.