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(71) Applicant (for all designated States except US): **BIOGEN IDEC MA INC.** [US/US]; 14 Cambridge Center, Cambridge, MA 02142 (US).

(72) Inventors; and

(75) Inventors/Applicants (for US only): **THORN, Karina** [DK/US]; 49 North Main St., Newmarket, NH 03857 (US). **TOBY, Garabet G.** [US/US]; 9 School Hill Lane, North Reading, MA 01864 (US). **MEZO, Adam R.** [CA/US]; 86 Arch Street, Needham, MA 02492 (US).(74) Agents: **COVERT, John M.** et al.; Sterne Kessler Goldstein & Fox PLLC, 1100 New York Avenue, N.W., Washington, DC 20005 (US).

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(54) Title: PRO-COAGULANT COMPOUNDS AND METHODS OF USE THEREOF

(57) Abstract: The present invention provides pro-coagulant compounds (e.g., pro-coagulant peptides or peptide derivatives) and methods of using and making those compounds. The present disclosure further provides conjugates between a pro-coagulant compound of the present disclosure (e.g., pro-coagulant peptide or peptide derivative) and a polypeptide selected from FIX, FVIIa, FVIII, and platelet targeting moieties (e.g., PDG-13), wherein the compound is linked to the polypeptide optionally via a linker. The compounds and conjugates of the present disclosure are useful for the treatment of coagulation disorders, such as hemophilia A and hemophilia B. The present disclosure further provides methods of using and making the conjugates.



PRO-COAGULANT COMPOUNDS AND METHODS OF USE THEREOF

BACKGROUND OF THE INVENTION

Field of the Invention

[0001] The present invention relates to pro-coagulant compounds useful for the treatment of coagulation disorders, such as hemophilia A and B.

Background of the Invention

[0002] The blood coagulation pathway, in part, involves the formation of an enzymatic complex of Factor VIIIa (FVIIIa) and Factor IXa (FIXa) (Xase complex) on the surface of platelets. FIXa is a serine protease with relatively weak catalytic activity without its cofactor FVIIIa. The Xase complex cleaves Factor X (FX) into Factor Xa (FXa), which in turn interacts with Factor Va (FVa) to cleave prothrombin and generate thrombin.

[0003] Hemophilia A is a bleeding disorder caused by mutations and/or deletions in the Factor VIII (FVIII) gene resulting in a deficiency of FVIII activity. In some cases, patients have reduced levels of FVIII due to the presence of FVIII inhibitors, such as anti-FVIII antibodies.

[0004] Hemophilia A is characterized by spontaneous hemorrhage and excessive bleeding after trauma. Over time, the repeated bleeding into muscles and joints, which often begins in early childhood, results in hemophilic arthropathy and irreversible joint damage. This damage is progressive and can lead to severely limited mobility of joints, muscle atrophy and chronic pain (Rodriguez-Merchan, E.C., *Semin. Thromb. Hemost.* 29:87-96 (2003), which is herein incorporated by reference in its entirety).

[0005] The disease can be treated by replacement therapy targeting restoration of FVIII activity to 1 to 5 % of normal levels to prevent spontaneous bleeding (see, e.g., Mannucci, P.M., et al., *N. Engl. J. Med.* 344:1773-9 (2001), herein incorporated by reference in its entirety). There are plasma-derived and recombinant FVIII products available to treat bleeding episodes on-demand or to prevent bleeding episodes from occurring by treating prophylactically. Based on the half-life of these products (10-12 hr) (White G.C., et al., *Thromb. Haemost.* 77:660-7 (1997); Morfini, M., *Haemophilia* 9 (suppl 1):94-99; discussion 100 (2003)), treatment regimens require frequent intravenous administration, commonly two to three times weekly for prophylaxis and one to three times daily for on-

demand treatment (Manco-Johnson, M.J., et al., *N. Engl. J. Med.* 357:535-544 (2007)), each of which is incorporated herein by reference in its entirety. Such frequent administration is painful and inconvenient.

[0006] Although on-demand treatment is frequently used, there is a trend toward prophylaxis and the prevention of joint damage (Blanchette P, *et al.*, *Haemophilia* 2004; 10:679-683, Manco-Johnson, MJ, *et al.*, *N. Engl. J. Med.* 2007; 357:535-544). Current FVIII products are administered every two to three days for prophylaxis due to the relatively short half-life of 10-12 hr in order to maintain a FVIII:C above 1 % in patients (Morfini, M, *Haemophilia* 2003; 9 (suppl 1):94-99;discussion 100, White GC, et al., *Thromb. Haemost.* 1997;77:660-7, Blanchette, P, et al., *J. Thromb. Haemost.* 2008 Aug;6(8):1319-26). Longer-acting FVIII therapies that provide prolonged protection from bleeding would represent an improvement in the quality of life for patients with hemophilia A.

[0007] Strategies to extend the half-life of clotting factors include pegylation (Rostin J, et al., *Bioconj. Chem.* 2000; 11:387-96), glycopegylation (Stennicke HR, et al., *Thromb. Haemost.* 2008; 100:920-8), formulation with pegylated liposomes (Spira J, *et al.*, *Blood* 2006;108:3668-3673, Pan J, et al., *Blood* 2009;114:2802-2811) and conjugation with albumin (Schulte S., *Thromb. Res.* 2008; 122 Suppl 4:S14-9).

[0008] D K Liles et al. (1997) *Blood* Vol 90 No 10 Supplement 1 (463a, poster abstract) discloses a peptide from FVIII which can promote FIXa mediated activation of FX on a phospholipid surface. However, in the presence of FVIIIa, the peptide inhibits FIXa mediated activation of FX. A peer-reviewed publication by these authors confirming the disclosed results was not available at time of this application.

[0009] Blostein *et al* (2000) *Biochemistry* 39:12000-12006 discloses that amphipathic alpha helices can interact with FIXa Gla domains and increases activation of FX in the absence of phospholipid.

[0010] Under normal conditions, activated platelets provide the lipid surface supporting coagulation. Since platelets are activated by thrombin, which is formed at sites of vascular injury, coagulation processes are restricted to the sites of injuries. However, it is undesirable to provide the body with peptides that are general substitutes for procoagulant lipids as this would cause systemic coagulation and ultimately lead to disseminated intravascular coagulation (DIC).

- [0011] U.S. Pat. Nos. 7,109,170 and 6,624,289 disclose regions of the FIXa protease domain that interact with FVIIIa and that comprise the FVIIIa binding site of FIXa. The peptides inhibit binding of FIXa to FVIIIa. The disclosed peptides may be useful as anticoagulants for preventing or treating thrombosis.
- [0012] US20010014456A1 discloses binding molecules for human FVIII and FVIII-like proteins. These polypeptides bind FVIII and/or FVIII-like polypeptides and are useful for the detection and purification of human FVIII and/or FVIII-like polypeptides from solutions such as blood or conditioned media.
- [0013] In U.S. Pat. No. 7,033,590 FIX/FIXa activating antibodies and antibody derivatives are used for increasing the amidolytic activity of FIXa, and for treating blood coagulation disorders such as hemophilia A and hemorrhagic diathesis.
- [0014] U.S. Pat. No. 7,084,109 discloses FVIIa antagonists that are peptides and inhibit FVIIa activity. The peptides may be useful for the prevention of arterial thrombosis in combination with thrombolytic therapy.
- [0015] Hemophilia B (also known as Christmas disease) is one of the most common inherited bleeding disorders in the world. It results in decreased *in vivo* and *in vitro* blood clotting activity and requires extensive medical monitoring throughout the life of the affected individual.
- [0016] In the absence of intervention, the afflicted individual may suffer from spontaneous bleeding in the joints, which produces severe pain and debilitating immobility. Bleeding into muscles results in the accumulation of blood in those tissues. Spontaneous bleeding in the throat and neck may cause asphyxiation if not immediately treated. Bleeding into the urine, and severe bleeding following surgery, minor accidental injuries, or dental extractions also are prevalent.
- [0017] Hemophilia B is caused by a deficiency in Factor IX that may result from either the decreased synthesis of the Factor IX protein or a defective molecule with reduced activity.
- [0018] Human FIX, one member of the group of vitamin K-dependent polypeptides, is a single-chain glycoprotein with a molecular weight of 57 kDa, which is secreted by liver cells into the blood stream as an inactive zymogen of 415 amino acids. It contains 12 γ -carboxy-glutamic acid residues localized in the N-terminal Gla-domain of the polypeptide. The Gla residues require vitamin K for their biosynthesis. Following the

Gla domain there are two epidermal growth factor domains, an activation peptide, and a trypsin-type serine protease domain. Further posttranslational modifications of FIX encompass hydroxylation (Asp 64), N-(Asn157 and Asn167) as well as O-type glycosylation (Ser53, Ser61, Thr159, Thr169, and Thr172), sulfation (Tyr155), and phosphorylation (Ser158). FIX is converted to its active form, Factor IXa, by proteolysis of the activation peptide at Arg145-Ala146 and Arg180-Val181 leading to the formation of two polypeptide chains, an N-terminal light chain (18 kDa) and a C-terminal heavy chain (28 kDa), which are held together by one disulfide bridge. Activation cleavage of Factor IX can be achieved *in vitro* e.g. by Factor XIa or Factor VIIa/TF. Factor IX is present in human plasma in a concentration of 5-10 µg/ml. Terminal plasma half-life of Factor IX in humans was found to be about 15 to 18 hours (White G C et al. 1997. Recombinant factor IX. *Thromb Haemost.* 78: 261-265; Ewenstein B M et al. 2002. Pharmacokinetic analysis of plasma-derived and recombinant F IX concentrates in previously treated patients with moderate or severe hemophilia B. *Transfusion* 42:190-197).

[0019] The treatment of hemophilia B occurs by replacement of the missing clotting factor by exogenous factor concentrates highly enriched in Factor IX. However, generating such a concentrate from blood is difficult. Purification of Factor IX from plasma (plasma derived Factor IX; pdFIX) almost exclusively yields active Factor IX. However, such purification of FIX from plasma is very difficult because FIX is only present in low concentration in plasma (Andersson, *Thrombosis Research* 7: 451 459 (1975). Further, purification from blood requires the removal or inactivation of infectious agents such as HIV and HCV. In addition, pdFIX has a short half-life and therefore requires frequent dosing. Recombinant FIX (rFIX) is also available, but suffers from the same short half-life and need for frequent dosing (e.g., 2-3 times per week for prophylaxis) as pdFIX.

[0020] A recombinant FVIIa product is marketed by Novo Nordisk (NovoSeven).

[0021] Reduced mortality, prevention of joint damage and improved quality of life have been important achievements due to the development of plasma-derived and recombinant clotting factors. Prolonged protection from bleeding would represent another key advancement in the treatment of hemophilia patients. However, to date, no products that allow for prolonged protection have been developed.

[0022] However, there remains a great need for improved pro-coagulant therapies for the treatment (e.g., prophylactic treatment) of hemophilia and other blood coagulation disorders that are more tolerable and more effective than current therapies. Small-molecule therapies, which can be administered by a non-intravenous route are particularly useful.

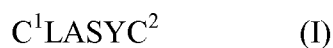
BRIEF SUMMARY OF THE INVENTION

[0023] The present invention provides low molecular weight compounds (e.g., peptides or peptide derivatives) with pro-coagulant activity useful for the treatment (e.g., intravenous or non-intravenous treatment) of bleeding diathesis (e.g., blood coagulation disorders/coagulopathies, such as hemophilia A and hemophilia B) or for the treatment of deficiencies in at least one of Factor V (FV), Factor FVII (FVII), Factor VIII (FVIII), Factor IX (FIX), Factor X (FX), Factor XI (FXI), Factor XII (FXII), Factor XIII (FXIII), and von Willebrand Factor (vWF). In one example, the current compounds exhibit greater *in vivo* stability than known treatments (e.g., FVIII, FIX, or FVIIa) and have the potential to significantly reduce the cost of treating coagulation disorders.

[0024] In various embodiments, the compounds of the present invention are capable of increasing the catalytic activity of a blood coagulation factor (e.g., FIXa or FVIIa). In other embodiments, the compounds of the invention exhibit biological activity in the presence of FVIII (i.e., possess additive activity with FVIII). In another example, the current compounds are useful for the treatment of impaired coagulation in FVIII inhibitor patients.

[0025] The present disclosure further provides conjugates containing a polypeptide selected from blood coagulation factors (e.g., FVIII, FIX, FVIIa), and platelet targeting moieties (e.g., PDG-13), wherein the polypeptide is linked to a compound of the present disclosure (e.g., a pro-coagulant peptide or a peptide derivative), optionally via a linker. The present disclosure further provides conjugates wherein a compound (e.g., a peptide or peptide derivative) has pro-coagulant activity.

[0026] The present invention provides a compound (e.g., a peptide or peptide derivative) comprising: (a) an amino acid sequence comprising Formula (I):



or (b) a retro-, an inverso- or a retro-inverso variant of the amino acid sequence of (a). A compound can be present as a pharmaceutically acceptable salt. In Formula (I), L is L-leucine; A is L-alanine; S is L-serine; Y is L-tyrosine, wherein one or two of L, A, S, and Y are optionally replaced with a replacement amino acid independently selected from D- and L-amino acids. Exemplary amino acids and replacement amino acids for L, A, S, and Y are described herein.

[0027] In some embodiments, a compound of the present invention includes at least 9 and not more than about 500 amino acid residues. In some embodiments, a compound of the present invention includes at least 12 and not more than about 100 amino acid residues. Other suitable ranges for the number of amino acids in the compounds of the present disclosure are described herein.

[0028] In Formula (I), C¹ and C² are independently selected from amino acids having a side chain, wherein the side chains of C¹ and C² are linked to form a loop. In one example, C¹ and C² are independently selected from amino acids having a side chain comprising a -S-H group, and wherein the side chains of C¹ and C² are reversibly linked via a disulfide bond. In another example, the side chains of C¹ and C² are covalently linked via an amide bond to form a lactam ring. In yet another example, the side chains of C¹ and C² are covalently linked via an optionally substituted triazole moiety.

[0029] The invention further provides a compound (e.g., a peptide or peptide derivative) containing an amino acid sequence having C¹ and C², wherein C¹ and C² are independently selected amino acids having a side chain, wherein the side chains of C¹ and C² are linked, and wherein C¹ and C² are separated by 4, 5 or 6 amino acids, wherein a compound includes at least 9 and not more than 500 amino acids. In one embodiment, C¹ and C² are separated by 4 amino acids.

[0030] In one example, compounds of the present disclosure have an EC₅₀ of about 5 μM or less in a Factor Xa (FXa) generation assay measuring conversion of Factor X (FX) to FXa (e.g., in the presence of FIXa). A suitable FXa generation assay is described in Example 2 of this application. In a particular example, a compound has an EC₅₀ of about 1 μM or less (e.g., 200 nM or less). Certain compounds of the present disclosure, at a concentration of 5 μM or less, increase the catalytic activity (k_{cat}) of at least one blood coagulation factor (e.g., FIXa or FVIIa). For example, compounds of the present disclosure, at a concentration of 5 μM or less, increase the catalytic activity (k_{cat}) of

Factor IXa (FIXa) or FVIIa for conversion of FX to FXa in a FXa generation assay when compared to a reference catalytic activity of the FIXa or the FVIIa measured in the absence of the compound. In one example, a compound increases the catalytic activity (k_{cat}) of FIXa by at least 50 fold or at least 100 fold.

[0031] In another example, a compound increases the catalytic activity (k_{cat}) of FVIIa by at least 200 fold, at least 400 fold or at least 1000 fold.

[0032] The invention further provides polypeptide conjugates comprising (a) a polypeptide selected from FVIII, FIX, FVIIa, and platelet targeting moieties, and (b) a compound of the present disclosure, wherein the compound is linked to the polypeptide, optionally via a linker.

[0033] The compounds and conjugates of the present disclosure, upon administration to a human or other animal, may produce an augmented prophylactic or therapeutic effect, lower dosing and/or dosing frequency of coagulation factors, or increased specific activity and catalytic activity of the coagulation factors.

[0034] The invention further provides a pharmaceutical composition containing at least one compound or conjugate of the present disclosure and a pharmaceutically acceptable carrier. Suitable pharmaceutically acceptable carriers are described herein.

[0035] The invention further provides a method of increasing the catalytic activity (k_{cat}) of a blood coagulation factor (e.g., FIXa or FVIIa). The method includes contacting the blood coagulation factor (*in vitro* or *in vivo*) with a compound or conjugate of the present disclosure. In one example, a compound or conjugate interacts with the blood coagulation factor at a region corresponding to amino acid sequence: MFCAG (SEQ ID NO: 1). In a particular example, the blood coagulation factor that is contacted with a compound or conjugate of the present disclosure is FIXa or FVIIa.

[0036] The invention further provides a method for treating bleeding diathesis in a mammalian subject (e.g., a human subject). An exemplary method includes administering to the subject a therapeutically effective amount of a compound or conjugate of the present disclosure or a pharmaceutical composition of the present disclosure. In one example, the bleeding diathesis is caused by a blood coagulation disorder, such as hemophilia (e.g., hemophilia A) or von Willebrand disease (vWD).

[0037] The invention further provides methods for making the compounds and conjugates of the present disclosure. An exemplary method includes forming a peptide incorporating

a desired amino sequence (or a retro-, inverso- or retro-inverso variant thereof) using solid-phase peptide synthesis. In one example, the method further includes covalently linking the peptide to a heterologous moiety that can extend the half-life of a compound, e.g., selected from a PEG moiety, Fc, IgG, FcRn binding ligand, albumin, albumin-binding ligand, transferrin, PAS, a half-life extension polypeptide (i.e., XTEN), or a hydroxyethyl starch

BRIEF DESCRIPTION OF THE FIGURES

- [0038] Figure 1 is a scheme illustrating an exemplary FXa generation assay. In one example, the FXa generation assay is performed using human FIXa (hFIXa), e.g., at 10 nM, and human FX (hFX), e.g., at 100 nM. In another example, the FXa generation assay is performed using human FVIIa (hFVIIa) and human FX (hFX).
- [0039] Figure 2 is a graph illustrating the additive effect between FVIII and compound 5 of the present invention in a thrombin generation assay (TGA) utilizing 0.1 pM of lipidated tissue factor (TF) as the clotting cascade activator. Results indicate that compound 5 does not compete with FVIII, but shows additive thrombin generation activity with FVIII. Compound 5 increases the thrombin peak in the presence of low amounts of rFVIII.
- [0040] Figure 3 is a graph illustrating that compound 5 enhances thrombin generation in human FVIII-deficient plasma. Thrombin generation was measured for compound 5 at 10 μ M and 5 μ M in a thrombin generation assay (TGA) using 0.1 pM lipTF as an activator. In this experiment, compound 5 has a shorter lag time than 0.1 or 0.25 U/mL of FVIII. The amount of thrombin generated by 5 or 10 μ M compound 5 is larger than the amount generated by 0.1 U/mL of FVIII. Compound 5 at 10 μ M generates similar amounts of thrombin than about 0.2 IU/mL of rFVIII.
- [0041] Figure 4 is a graph illustrating hydrogen-deuterium exchange (HDX) for amino acids 177-185 of hFIXa in the absence and presence of compound 4.
- [0042] Figure 5 is graph illustrating the activity of various compounds of the present disclosure in a thrombin generation assay using purified hemostatic components as described in Example 3.

- [0043] Figure 6 is a graph illustrating that compound 5 enhances thrombin generation in a dose-dependent manner when measured in a thrombin generation assay using purified hemostatic components as described in Example 3.
- [0044] Figure 7 contains three graphs illustrating that the thrombin generation activity of compound 5 is FIXa-dependent when measured in a thrombin generation assay using purified hemostatic components as described in Example 3.
- [0045] Figure 8 contains three graphs illustrating that the thrombin generation activity of compound 5 is FXIa-dependent when measured in a thrombin generation assay using purified hemostatic components as described in Example 3.
- [0046] Figure 9 is a scheme of a conjugate including FVIIa (AA), a heterologous moiety (represented by Fc) and a compound of the present disclosure. An exemplary conjugate according to Figure 9 is described in Example 14. In one example the FVIIa conjugate is formed from a precursor (BB) via further processing. Processing involves, e.g., cleavage of a cleavable linker, e.g., through intracellular activation/processing (e.g., by co-transfection of processing enzymes such as PC5 and PACE). In one example, in Figure 9, the compound is compound 21.
- [0047] Figure 10 is a scheme of a conjugate including FIX (CC), a heterologous moiety (represented by Fc) and a compound of the current disclosure. An exemplary conjugate according to Figure 10 is described in Example 15. In one example the FIX conjugate is formed from a precursor (DD) via further processing. Processing involves, e.g., cleavage of a cleavable linker, e.g., through intracellular activation/processing (e.g., by co-transfection of processing enzymes such as PC5 and PACE). In one example, in Figure 10, the compound is compound 21.
- [0048] Figure 11 is a scheme of a platelet targeting moiety conjugate (EE) including a platelet targeting moiety (represented by PDG-13), a heterologous moiety (represented by Fc) and a compound of the present disclosure. An exemplary conjugate according to Figure 11 is described in Example 16. In one example the conjugate (EE) is formed from a precursor (FF) via further processing. Processing involves, e.g., cleavage of a cleavable linker, e.g., through intracellular activation/processing (e.g., by co-transfection of processing enzymes such as PC5 and PACE). In one example, in Figure 11, the compound is compound 21.

- [0049] Figure 12 is a scheme of a conjugate (GG) including a heterologous moiety (represented by Fc) and a compound of the present disclosure. An exemplary conjugate according to Figure 12 is described in Example 17. In one example the conjugate (GG) is formed from a precursor (HH) via further processing. Processing involves, e.g., cleavage of a cleavable linker, e.g., through intracellular activation/processing (e.g., by co-transfection of processing enzymes such as PC5 and PACE). In one example, in Figure 12, the compound is compound 21.
- [0050] In Figures 9 to 12, the processing can involve intracellular activation/processing, which may be accomplished, e.g., by co-transfection of processing enzymes such as PC5 and PACE.
- [0051] Figure 13 is a gel showing the presence of various conjugates of the present disclosure (as prepared according to the procedures of Examples 14-17) after shFcRn-conjugated bead pulldown from conditioned media.
- [0052] Figure 14 is a graph illustrating the TGA activity of an Fc-compound 21 conjugate as described in Figure 12 and Example 17 at various concentrations in the assay mixture. The Fc-compound 21 conjugate enhances thrombin formation in a dose-dependent manner and maintains its TGA activity relative to compound 21 not conjugated to Fc.
- [0053] Figure 15 is a graph illustrating the TGA activity of a FVIIa-Fc-compound 21 (FVII-171) conjugate as described in Figure 33 and Example 14 at various concentrations. TGA activity is enhanced by the conjugate in a dose-dependent manner compared to the activity to FVIIa-Fc (FVII-002) not conjugated to compound 21.
- [0054] Figure 16 is a schematic illustration of conjugates containing a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative) covalently linked to a FIX-heterologous moiety construct, optionally via a linker. Fc is used as an example of a heterologous moiety and can be replaced by other heterologous moieties.
- [0055] Figure 17 is a schematic illustration of conjugates containing a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative) covalently linked to a FIX protein, optionally via a linker.
- [0056] Figure 18 is a schematic illustration of conjugates containing a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative) covalently linked to a FVIIa-heterologous moiety construct, optionally via a linker. Fc is used as an example of a heterologous moiety and can be replaced by other heterologous moieties.

- [0057] Figure 19 is a schematic illustration of the conjugates containing a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative) covalently linked to a FVIIa protein, optionally via a linker.
- [0058] Figure 20 is a schematic illustration of conjugates containing a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative) covalently linked to a FVIII-heterologous moiety construct, optionally via a linker. Fc is used as an example of a heterologous moiety and can be replaced by other heterologous moieties.
- [0059] Figure 21 is a schematic illustration of the conjugates containing a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative) covalently linked to a FVIII protein, optionally via a linker.
- [0060] Figure 22 is a schematic illustration of conjugates containing a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative) covalently linked to a platelet targeting moiety (construct H3) or a platelet targeting moiety-heterologous moiety construct (H1, H2, Ha, Hb) optionally via a linker. Fc is used as an example of a heterologous moiety and can be replaced by other heterologous moieties.
- [0061] Figure 23 is a schematic illustration of a general method for covalently linking a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative) to a FVIII-Fc, a FIX-Fc, or a FVIIa-Fc construct using a native ligation strategy. The cysteine residue used for the ligation can be replaced with other reactive amino acids and can be reacted with a complementary reactive group located on a compound.
- [0062] Figure 24 is a schematic illustration of a general method for covalently linking a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative) to a FVIII-Fc construct, a FIX-Fc construct, or a FVIIa-Fc construct using a site-directed ligation strategy. The cysteine residue used for the ligation can be replaced with other reactive amino acids and can be reacted with a complementary reactive group located on a compound.
- [0063] Figure 25 is a schematic illustration of a general method for covalently linking a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative) to a platelet targeting moiety-Fc construct using a native ligation strategy.
- [0064] Figure 26 is a schematic illustration of a general method for covalently linking a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative) to a platelet targeting moiety-Fc construct using a site-directed ligation strategy.

[0065] Linkers of Figures 1 to 26 can be any linker, e.g., those described herein.

DETAILED DESCRIPTION OF THE INVENTION

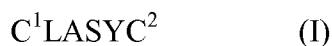
Compounds

[0066] In various aspects, the present invention provides compounds with pro-coagulant activities. In one example, a compound includes an amino acid sequence comprising C¹ and C², wherein C¹ and C² are independently selected from amino acids having a side chain, wherein the side chains of C¹ and C² are linked to form a loop structure. In one example, the amino acid side chains of C¹ and C² are reversibly linked via a disulfide bond. In another example, the side chains of C¹ and C² are covalently linked via an amide bond to form a lactam ring (e.g., formed between the amino group of a lysine residue and the carboxylic acid group of a glutamic acid or an aspartic acid residue). In one example, C¹ and C² are separated by 3, 4, 5 or 6 amino acids. In another example, C¹ and C² are separated by 3, 4 or 5 amino acids. In yet another example, C¹ and C² are separated by 4 or 5 amino acids. In yet another example, C¹ and C² are separated by 3 or 4 amino acids. In a further example, C¹ and C² are separated by 4 amino acids.

[0067] In one example according to any of the above embodiments, a compound that includes the amino acid sequence incorporating C¹ and C² contains at least 9 and not more than 500 amino acids. In another example, a compound comprises at least 12 and not more than 100 amino acids. In a further example, a compound comprises at least 20 and not more than 100 or 50 amino acids. Further suitable ranges for the number of amino acids in a compound of the present disclosure are described herein.

[0068] The present disclosure also provides a compound that includes:

(a) an amino acid sequence including Formula (I):



or (b) a retro-, an inverso- or a retro-inverso variant of the amino acid sequence of (a).

The present disclosure further provides pharmaceutically acceptable salts or solvates of the above compound.

[0069] In Formula (I), C¹ and C² are amino acids having a side chain, wherein the side chains of C¹ and C² are linked to form a loop. In one example, the side chains of C¹ and C² are covalently linked (e.g., via a disulfide bond or an amide bond).

- [0070] In one example, In Formula (I), one, two or three additional amino acids can be inserted anywhere between C^1 and C^2 . In one example according to any of the above embodiments, one or two additional amino acids are optionally inserted into Formula (I) anywhere between C^1 and C^2 . In another example, one amino acid is optionally inserted into Formula (I) anywhere between C^1 and C^2 . In another example, C^1 and C^2 are separated by exactly 4 amino acids.
- [0071] In Formula (I), L is L-leucine, A is L-alanine, S is L-serine, and Y is L-tyrosine. In Formula (I), one, two or three of L, A, S, and Y are optionally replaced with an independently selected replacement amino acid. In one example, one or two of L, A, S, and Y are optionally replaced with an independently selected replacement amino acid. In another example, exactly one of L, A, S, and Y is optionally replaced with an independently selected replacement amino acid. In one example, the amino acid sequence of a compound includes the sequence CLASYC.
- [0072] In one example, each replacement amino acid is independently selected from L- and D-amino acids. In another example, the replacement amino acid is a proteinogenic amino acid. In another example, the replacement amino acid is a non-proteinogenic amino acid. Exemplary non-proteinogenic amino acids are described herein and include, e.g., homo-analogs, such as homo-phenylalanine, and homo-cysteine. In yet another example, the replacement amino acid is a modified amino acid (including modified proteinogenic and modified non-proteinogenic amino acids). Examples of modified amino acids include, e.g., alpha-N-alkylated amino acids, tyrosine derivatives (e.g., those in which the hydroxyl group is converted to an ether or ester group), lysine derivatives (e.g., those in which the NH_2 group is converted to an amide group or sulfonamide group), and amino acids, in which a carboxylic acid group is derivatized, e.g., esterified, converted to an amide group, and the like. Other examples of suitable replacement amino acids are disclosed herein. In one example, L, A, S and Y are selected from amino acids other than those having a side chain comprising a -S-H group or a -Se-H group (e.g., other than cysteine).
- [0073] In one example in Formula (I), C^1 and C^2 are independently selected from amino acids having a side chain comprising a -S-H group or a -Se-H group and the side chains of C^1 and C^2 are linked (e.g., reversibly linked) via a disulfide bond (-S-S-), a diselenide bond (-Se-Se-), a -Se-S- or a -S-Se- bond. In another example in Formula (I), C^1 and C^2

are independently selected from cysteine, homo-cysteine (HCy), seleno-cysteine (U), homo-seleno cysteine, and D-amino acids thereof. In another example in Formula (I), C¹ is selected from cysteine, homo-cysteine (HCy), seleno-cysteine (U), homo-seleno cysteine, and D-amino acids thereof. In another example in Formula (I), C² is selected from L-cysteine, L-homo-cysteine (HCy), L-seleno-cysteine (U), and L-homo-seleno cysteine. In yet another example, C¹ in Formula (I) is selected from cysteine, homo-cysteine (HCy), seleno-cysteine (U), homo-seleno cysteine, and D-amino acids thereof, and C² is selected from L-cysteine, L-homo-cysteine (HCy), L-seleno-cysteine (U), and L-homo-seleno cysteine.

[0074] In one example in Formula (I), C¹ and C² are independently selected from amino acids having a side chain comprising a -S-H group, wherein the side chains of C¹ and C² are linked (e.g., reversibly linked) via a disulfide bond. In another example, C¹ and C² are independently selected from cysteine and homo-cysteine. In yet another example, C¹ and C² are both cysteine. In yet another example in Formula (I), C¹ is selected from L-cysteine and D-cysteine. In yet another example in Formula (I), C² is L-cysteine. In another example in Formula (I), C¹ is selected from L-cysteine and D-cysteine, and C² is L-cysteine.

[0075] The covalent linkage between the side chains of C¹ and C² can be reversible. In one example according to the above embodiments in which C¹ and C² are selected from amino acids having a side chain incorporating a -SH or Se-H group, a certain percentage (e.g., less than 50%, less than about 40%, less than about 30%, less than about 20%, less than about 10%, less than about 8%, less than about 6%, less than about 4%, or less than about 2%) of a compound can exist in an open form (i.e., in which the side chains of C¹ and C² are not linked to form a loop). Thus, those linkages are referred to as being reversible. For example, when side chains of C¹ and C² comprise an -SH group (e.g., cysteine), then C¹ and C² can be reversibly linked by a disulfide bond, wherein some molecules exist in an open form, but wherein the majority will be covalently linked by a disulfide bond. Whether or not the side chains of C¹ and C² are covalently linked can depend on the chemical environment in which a compound exists. For example, in a reducing environment, a disulfide bond may be broken or may not be formed.

[0076] In one example according to any of the above embodiments, the side chains of C¹ and C² are covalently linked via an amide bond to form a lactam ring. In one

embodiment according to this example, one of C¹ and C² is selected from amino acids having a side chain with a primary or secondary amino group (e.g., -NH₂ group), and the other of C¹ and C² is selected from an amino acid with a side chain having a carboxylic acid group (e.g., -COOH group), wherein the amino group and the carboxylic acid group form an amide bond. Methods for the formation of amide bonds between the side chains of C¹ and C² are described herein (see, e.g., Example 1). For example, the carboxylic acid group can first be activated prior to reaction with the amino group.

[0077] In another example according to any of the above embodiments, one of C¹ and C² is selected from amino acids having a straight or branched aminoalkyl, e.g., (C₁-C₁₀)aminoalkyl, side chain, and the other of C¹ and C² is selected from amino acids having a straight or branched carboxyalkyl, e.g., (C₁-C₁₀)carboxyalkyl, side chain, wherein an amino group of the aminoalkyl side chain and a carboxylic acid group of the carboxyalkyl side chain are linked to form an amide bond. In yet another example, one of C¹ and C² is selected from lysine (2,6-diamino-hexanoic acid), 2,5-diamino-pentanoic acid (ornithine; Orn), 2,4-diamino-butyric acid (Dab), 2,3-diamino-propionic acid (Dpr), 2,7-diamino-heptanoic acid, and 2,8-diamino-octanoic acid, and the other of C¹ and C² is selected from aspartic acid, glutamic acid (2-amino-pentanedioic acid), 2-amino-hexanedioic acid, 2-amino-heptanedioic acid, and 2-amino-octanedioic acid.

[0078] In yet another example according to any of the above embodiments, one of C¹ and C² is selected from lysine (K), 2,4-diaminobutyric acid (Dab), 2,3-diaminopropionic acid (Dpr), and ornithine (Orn), and the other of C¹ and C² is selected from aspartic acid and glutamic acid. In another example, one of C¹ and C² is lysine and the other of C¹ and C² is selected from aspartic acid and glutamic acid.

[0079] In a further example, the side chains of C¹ and C² are covalently linked via a triazole moiety. The triazole moiety can optionally be substituted, e.g., with alkyl, e.g., (C₁-C₄)alkyl, or OR⁹, wherein R⁹ is selected from H and (C₁-C₄)alkyl. Methods to form a triazole moiety are known to those of skill in the art (see e.g., Holland-Nell, K, and Meldal, M; *Angew. Chem Int. Ed.* 2011, 50: 5204-5206 and references cited therein, all of which are incorporated herein by reference in their entirety). In one example the triazole moiety is formed between an azide group of one of the side chains of C¹ and C², and an alkyne moiety of the side chain of the other of C¹ and C² (e.g., Huisgen cycloaddition). In one example, one of C¹ and C² is selected from amino acids having a straight or

branched azidoalkyl, e.g., (C₁-C₁₀)azidoalkyl, side chain and the other of C¹ and C² is selected from amino acids having a straight or branched alkynyl, e.g., (C₁-C₁₀)alkynyl, side chain, wherein an azide moiety of the azidoalkyl group and an alkyne moiety of the alkynyl group are linked to form a triazole moiety (e.g., a 1,4-triazole moiety or a 1,5-triazole moiety). In one example, the alkyno-functionalized amino acid is propargylglycine (Pra). In another example, the alkyno-functionalized amino acid is selected from 2-amino-4-azido-butyric acid (2Abu(γ N₃)) and 5-azido-norvaline (NVA(δ N₃)). Formation of the triazole ring can be accomplished using a suitable catalyst, such as a copper, e.g., Cu(I) catalyst (e.g., CuSO₄/tris(carboxyethyl)phosphine), or a suitable ruthenium-based catalyst.

[0080] In one example according to any of the above embodiments, S in Formula (I) is selected from serine and replacement amino acids having a side chain comprising a hydroxyl group. In another example, S in Formula (I) is serine. In another example, S in Formula (I) is L-serine.

[0081] In one example in Formula (I), each replacement amino acid for L, A, S, and Y, when present, is selected from L-amino acids. In a further example in Formula (I), S is L-serine or a replacement amino acid, L is L-leucine or a replacement amino acid, A is L-alanine or a replacement amino acid, and Y is L-tyrosine or a replacement amino acid, wherein each replacement amino acid for L, A, S and Y is independently selected from L-amino acids.

[0082] In another example in Formula (I), S is serine, L is L-leucine or a replacement amino acid, A is L-alanine or a replacement amino acid, and Y is L-tyrosine or a replacement amino acid, wherein each replacement amino acid for L, A and Y is independently selected from L-amino acids.

[0083] In another example in Formula (I), S is L-serine, L is L-leucine or a replacement amino acid, A is L-alanine or a replacement amino acid, and Y is L-tyrosine or a replacement amino acid, wherein each replacement amino acid for L, A, and Y is independently selected from L-amino acids.

[0084] In one example according to any of the above embodiments, replacement of one or two of L, A, S, and Y in Formula (I) with a replacement amino acid, or insertion of an additional amino acid, results in a neutral net-charge between C¹ and C². In another example according to any of the above embodiments, S is serine and replacement of one

or two of L, A, and Y in Formula (I) with a replacement amino acid, or insertion of an additional amino acid, results in a neutral net-charge between C¹ and C². A non-neutral net-charge between C¹ and C² results, e.g., when one of L, A, S and Y in Formula (I) is chosen from an amino acid having a side chain incorporating an acidic, e.g., carboxylic acid group (i.e., -COO⁻) or a basic, e.g., amino group (i.e., -NH₃⁺) while the remaining of L, A, S and Y are chosen from amino acids with a hydrophobic or polar uncharged side chain. Hence, in one example according to any of the above embodiments, each replacement amino acid for L, A, S and Y in Formula (I), when present, is independently selected from amino acids having a hydrophobic or a polar uncharged side chain; e.g., are independently selected from G, A, V, I, L, M, F, W, Y, S, T, N, Q, and P (e.g., G, A, V, I, L, M, F, W, Y, S, T, N, and Q).

[0085] In a further example, S is L-serine or a replacement amino acid, L is L-leucine or a replacement amino acid, A is L-alanine or a replacement amino acid, and Y is L-tyrosine or a replacement amino acid, wherein each replacement amino acid for L, A, S and Y is independently selected from amino acids having a hydrophobic or a polar uncharged side chain.

[0086] In a further example, S is L-serine or a replacement amino acid, L is L-leucine or a replacement amino acid, A is L-alanine or a replacement amino acid, and Y is L-tyrosine or a replacement amino acid, wherein each replacement amino acid for L, A, S and Y is independently selected from G, A, V, I, L, M, F, W, Y, S, T, N, Q and P (e.g., G, A, V, I, L, M, F, W, Y, S, T, N, and Q). In a further example, S is serine, L is L-leucine or a replacement amino acid, A is L-alanine or a replacement amino acid, and Y is L-tyrosine or a replacement amino acid, wherein each replacement amino acid for L, A, and Y, when present, is independently selected from G, A, V, I, L, M, F, W, Y, S, T, N, Q, and P (e.g., G, A, V, I, L, M, F, W, Y, S, T, N, and Q). In a further example, S is L-serine, L is L-leucine or a replacement amino acid, A is L-alanine or a replacement amino acid, and Y is L-tyrosine or a replacement amino acid, wherein each replacement amino acid for L, A, and Y is independently selected from G, A, V, I, L, M, F, W, Y, S, T, N, Q, and P (e.g., G, A, V, I, L, M, F, W, Y, S, T, N, and Q).

[0087] In another example according to any of the above embodiments, at least one of L, A, S and Y in Formula (I) is replaced with a replacement amino acid. In another example, exactly one of L, A, S and Y is replaced with a replacement amino acid. In

another example, exactly one of L, A, S and Y is replaced with a replacement amino acid, wherein the replacement amino acid is selected from L-amino acids. In another example, at least one of L, A, S and Y is replaced with a replacement amino acid, wherein the replacement amino acid is selected from G, A, V, I, L, M, F, W, Y, S, T, N, Q, and P (e.g., G, A, V, I, L, M, F, W, Y, S, T, N, and Q). In another example, exactly one of L, A, S and Y is replaced with a replacement amino acid, wherein the replacement amino acid is selected from G, A, V, I, L, M, F, W, Y, S, T, N, Q, and P (e.g., G, A, V, I, L, M, F, W, Y, S, T, N, and Q).

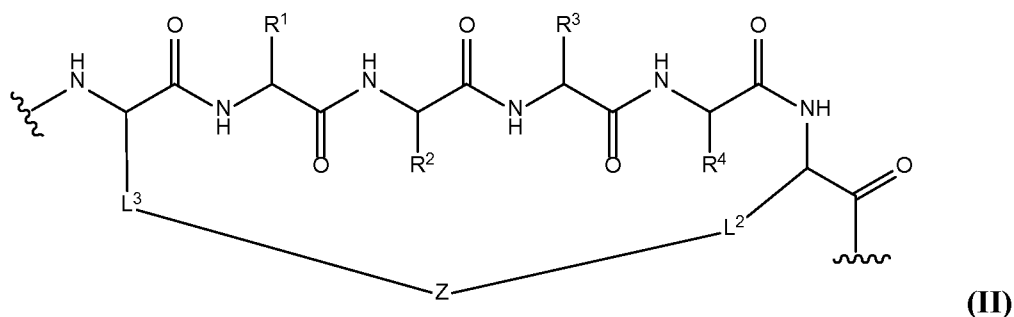
[0088] In another example, S is serine and at least one of L, A, and Y in Formula (I) is replaced with a replacement amino acid. In another example, S is serine and exactly one of L, A, and Y is replaced with a replacement amino acid. In another example, S is serine and at least one of L, A, and Y is replaced with a replacement amino acid, wherein the replacement amino acid is selected from L-amino acids. In another example, S is serine and exactly one of L, A, and Y is replaced with a replacement amino acid, wherein the replacement amino acid is selected from L-amino acids. In another example, S is serine and at least one of L, A, and Y is replaced with a replacement amino acid, wherein the replacement amino acid is selected from G, A, V, I, L, M, F, W, Y, S, T, N, Q, and P (e.g., G, A, V, I, L, M, F, W, Y, S, T, N, and Q). In another example, S is serine and exactly one of L, A, and Y is replaced with a replacement amino acid, wherein the replacement amino acid is selected from G, A, V, I, L, M, F, W, Y, S, T, N, Q and P (e.g., G, A, V, I, L, M, F, W, Y, S, T, N, and Q).

[0089] In one example, S is L-serine, and at least one of L, A, and Y in Formula (I) is replaced with a replacement amino acid. In another example, S is L-serine, and exactly one of L, A, and Y is replaced with a replacement amino acid. In another example, S is L-serine, and at least one of L, A, and Y is replaced with a replacement amino acid, wherein the replacement amino acid is selected from L-amino acids. In another example, S is L-serine, and exactly one of L, A, and Y is replaced with a replacement amino acid, wherein the replacement amino acid is selected from L-amino acids. In another example, S is L-serine, and at least one of L, A, and Y is replaced with a replacement amino acid, wherein the replacement amino acid is selected from G, A, V, I, L, M, F, W, Y, S, T, N, Q, and P (e.g., G, A, V, I, L, M, F, W, Y, S, T, N, and Q). In another example, S is L-serine, and exactly one of L, A, and Y is replaced with a replacement amino acid, wherein

the replacement amino acid is selected from G, A, V, I, L, M, F, W, Y, S, T, N, Q, and P (e.g., G, A, V, I, L, M, F, W, Y, S, T, N, and Q). In a further example in Formula (I), S is L-serine, L is L-leucine, A is L-alanine, and Y is L-tyrosine, and none of L, A, S and Y is replaced with a replacement amino acid.

[0090] In another example, exactly two of L, A, S and Y are replaced with a replacement amino acid. In another example, exactly two of L, A, S and Y are replaced with a replacement amino acid, wherein each replacement amino acid is independently selected from L-amino acids. In another example, S is serine and exactly two of L, A, and Y are replaced with a replacement amino acid. In another example, S is serine and exactly two of L, A, and Y are replaced with a replacement amino acid, wherein each replacement amino acid is independently selected from L-amino acids. In another example, S is serine and exactly two of L, A, and Y are replaced with a replacement amino acid, wherein each replacement amino acid for L, A and Y is independently selected from G, A, V, I, L, M, F, W, Y, S, T, N, Q, and P (e.g., G, A, V, I, L, M, F, W, Y, S, T, N, and Q). In another example, S is L-serine and exactly two of L, A, and Y are replaced with a replacement amino acid, wherein each replacement amino acid for L, A and Y is independently selected from G, A, V, I, L, M, F, W, Y, S, T, N, Q, and P (e.g., G, A, V, I, L, M, F, W, Y, S, T, N, and Q).

[0091] The present disclosure further provides a compound that contains a peptide of Formula (II):



or a retro-, an inverso- or a retro-inverso variant thereof.

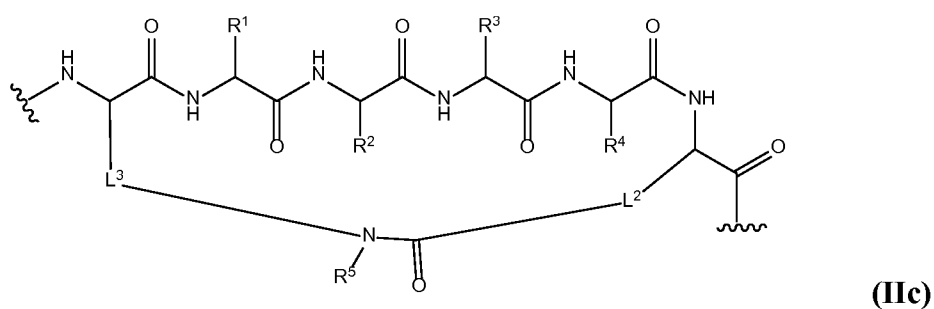
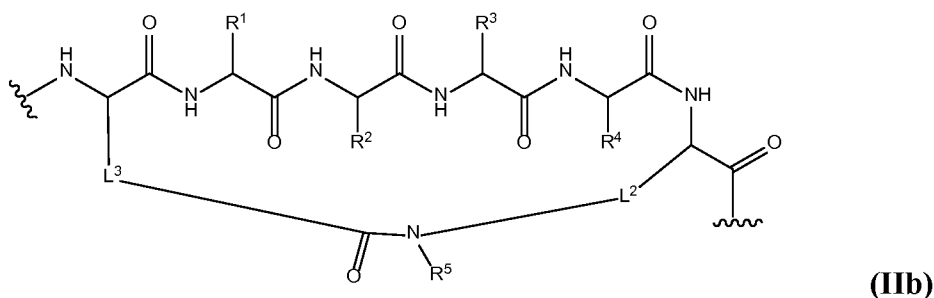
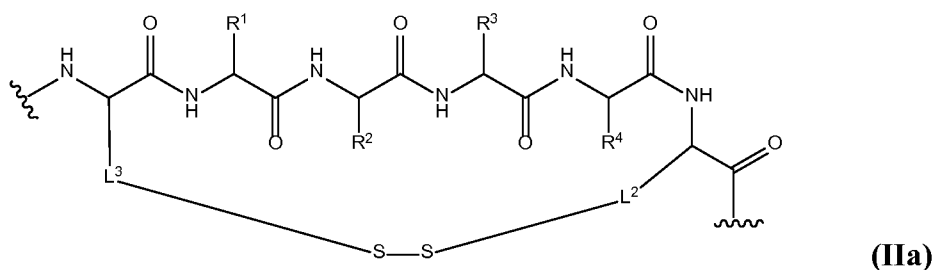
[0092] In Formula (II), R¹, R², R³ and R⁴ are members independently selected from amino acid side chains. In Formula (II), L² and L³ are linker groups independently selected from straight or branched alkylene, and straight or branched heteroalkylene. In one example, L² and L³ are independently selected from straight or branched (C₁-C₂₀)alkylene. In another example, L² and L³ are independently selected from straight or

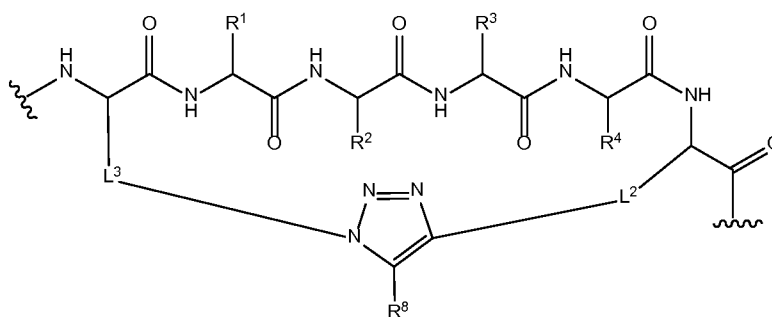
branched (C₁-C₁₀)alkylene. In yet another example, L² and L³ are independently selected from straight or branched (C₁-C₆)alkylene. In yet another example, L² and L³ are independently selected from straight or branched (C₁-C₄)alkylene. In a further example, L² and L³ are independently selected from heteroalkylene, e.g., (C₁-C₁₀)heteroalkylene. In one example, the heteroalkylene includes from 1 to 10 heteroatoms (e.g., from 1 to 7, from 1 to 5 heteroatoms, or from 1 to 3 heteroatoms) selected from O, S and N. In another example, at least one of L² and L³ incorporate a water-soluble polymeric moiety, such as a polyethylene glycol (PEG) or polypropylene glycol (PPG) moiety (e.g., with a molecular weight from about 1000 Da to about 60,000 Da. Other PEG or PPG moieties are described herein.

[0093] In Formula (II), Z is a linking moiety. In one example, Z is selected from an amino group, an amide group, a disulfide group, a diselenide group, a -S-Se- group, alkylene, e.g., (C₂-C₄)alkylene, alkenyl, e.g., (C₂-C₄)alkenyl, alkynyl, e.g., (C₂-C₄)alkynyl, cycloalkyl (e.g., (C₃-C₈)cycloalkyl containing from 1 to 4 double bonds), heterocycloalkyl (e.g., 3- to 8-membered heterocyclic ring comprising from 1 to 6 heteroatoms selected from O, S and N), aryl (e.g., (C₃-C₇)aryl), and heteroaryl (e.g., 3- to 8-membered heteroaryl comprising from 1 to 6 heteroatoms selected from O, S and N). In another example, Z in Formula (II) is selected from -NR⁵-, -NR⁵C(O)-, -S-S-, -S-Se-, -Se-Se-, -CR⁶=CR⁷-, -CR^{6a}R⁶-CR^{7a}R⁷-, and triazolenyl (e.g., 1,4-triazolenyl, or 1,5-triazolenyl), wherein R⁵, R⁶, R^{6a}, R⁷, and R^{7a} are independently selected from H, (C₁-C₄)alkyl, (C₁-C₄)heteroalkyl comprising from 1 to 3 heteroatoms selected from O, S and N. In one example, R⁶ and R⁷ are combined to form a 4- to 7-membered carbocyclic ring optionally comprising from 1 to 3 double bonds, a 3- to 7-membered heterocyclic ring comprising from 1 to 5 heteroatoms selected from O, S and N, a (C₅-C₇)aromatic ring, or a 5- to 7-membered heteroaromatic ring comprising from 1 to 5 heteroatoms selected from O, S and N. In another example, R⁶ and R⁷ are combined to form a 4- to 6-membered carbocyclic ring. In another example, R⁶ and R⁷ are combined to form a 4-membered carbocyclic ring. In another example, R⁶ and R⁷ are combined to form a 4- to 7-membered heterocyclic ring comprising from 1 to 3 heteroatoms selected from O, S and N, wherein the heterocyclic ring optionally comprises 1 or 2 double bonds. The carbocyclic or heterocyclic ring is optionally substituted with from 1 to 6 (e.g., 1 to 3) substituents selected from straight or branched (C₁-C₄)alkyl, straight or branched (C₁-

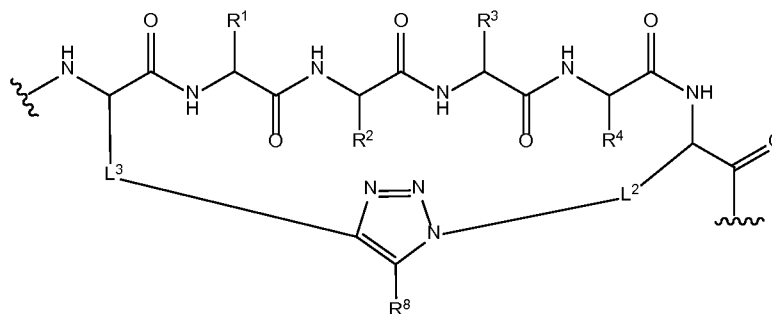
C₄)heteroalkyl comprising from 1 to 3 heteroatoms selected from O, S and N, halogen (e.g., F, Cl, Br), and OR¹⁰, wherein R¹⁰ is selected from H, straight or branched (C₁-C₄)alkyl, straight or branched (C₁-C₄)heteroalkyl comprising from 1 to 3 heteroatoms selected from O, S and N. In yet another example, Z in Formula (II) is selected from -NR⁵-, -NR⁵C(O)-, -S-S-, and a triazole moiety, wherein R⁵ is defined as above. In one example, the triazole moiety is a 1,4-triazole. In another example, the triazole moiety is a 1,5-triazole.

[0094] In another example, a compound of the present disclosure contains a peptide of Formula (IIa), Formula (IIb), Formula (IIc), Formula (IId), Formula (IIE), Formula (IIf), or Formula (IIg):

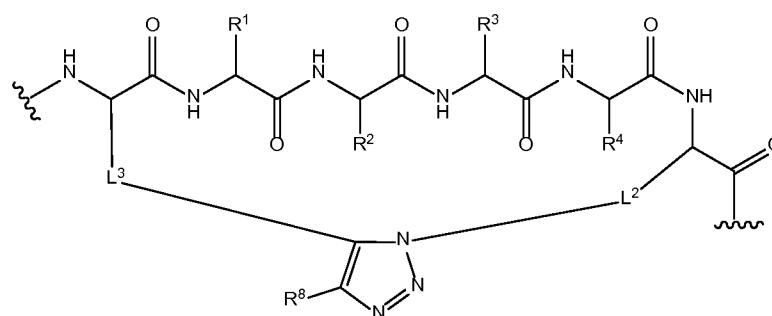




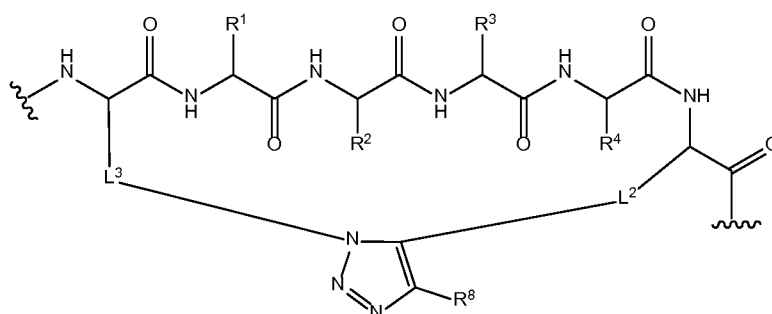
(IIId)



(IIe)



(IIIf)



(IIg)

or a retro-, an inverso- or a retro-inverso variant thereof, wherein R^1 , R^2 , R^3 , R^4 , R^5 , R^8 , L^2 , and L^3 are defined as for Formula (II) above. In one example in the above formulae, R^5 and R^8 are each selected from H and alkyl, e.g., (C₁-C₄)alkyl.

[0095] In another example, in Formula (II), (IIa), (IIb), (IIc), (IIId), (IIe), (IIIf), and (IIg), R^1 , R^2 , R^3 and R^4 are members independently selected from H, straight or branched alkyl, e.g., (C₁-C₆)alkyl, straight or branched heteroalkyl, e.g., (C₁-C₆)heteroalkyl comprising from 1 to 5 heteroatoms selected from O, S and N, and straight or branched aralkyl, e.g.,

(C₁-C₆)aralkyl. In one example, the aryl group in the aralkyl moiety is selected from aromatic and heteroaromatic rings disclosed herein. In one example, the aryl moiety of the aralkyl group is selected from phenyl, hydroxyphenyl, indolyl, and naphthyl. In another example, the aryl group in the aralkyl moiety is selected from phenyl, 4-hydroxyphenyl, and indolyl. In another example, R¹, R², R³ and R⁴ are independently selected from hydrophobic and polar uncharged side chains. In another example, R¹, R², R³ and R⁴ are independently selected from the side chains of G, A, V, I, L, M, F, W, Y, S, T, N, and Q. In another example, R¹ is 2-methyl-propyl, R² is methyl, R³ is hydroxymethyl, and R⁴ is (4-hydroxy-phenyl)methyl.

[0096] In another example according to any of the above embodiments, L² and L³ in Formula (II), (IIa), (IIb), (IIc), (IId), (IIe), (IIf), and (IIg) are independently selected from straight or branched (C₁-C₆)alkylene.

[0097] In various embodiments, a compound of the present disclosure comprises at least one of the following amino acid sequences:

KGASYE (SEQ ID NO: 560), KLGSYE (SEQ ID NO: 561), KLASGE (SEQ ID NO: 562), kGASYE (SEQ ID NO: 563), kLGSYE (SEQ ID NO: 564), kLASGE (SEQ ID NO: 565), KAASYE (SEQ ID NO: 566), KLASAE (SEQ ID NO: 567), kAASYE (SEQ ID NO: 568), kLASAE (SEQ ID NO: 569), KVASYE (SEQ ID NO: 570), KLVSYE (SEQ ID NO: 571), KLASVE (SEQ ID NO: 572), kVASYE (SEQ ID NO: 573), kLVSYE (SEQ ID NO: 574), kLASVE (SEQ ID NO: 575), KIASYE (SEQ ID NO: 576), KLISYE (SEQ ID NO: 577), KLASIE (SEQ ID NO: 578), kIASYE (SEQ ID NO: 579), kLISYE (SEQ ID NO: 580), kLASIE (SEQ ID NO: 581), KLASYE (SEQ ID NO: 582), KLLSYE (SEQ ID NO: 583), KLASLE (SEQ ID NO: 584), kLASYE (SEQ ID NO: 585), kLLSYE (SEQ ID NO: 586), kLASLE (SEQ ID NO: 587), KFASYE (SEQ ID NO: 588), KLFSYE (SEQ ID NO: 589), KLASFE (SEQ ID NO: 590), kFASYE (SEQ ID NO: 591), kLFSYE (SEQ ID NO: 592), kLASFE (SEQ ID NO: 593), KWASYE (SEQ ID NO: 594), KLWSYE (SEQ ID NO: 595), KLASWE (SEQ ID NO: 596), kWASYE (SEQ ID NO: 597), kLWSYE (SEQ ID NO: 598), kLASWE (SEQ ID NO: 599), KYASYE (SEQ ID NO: 600), KLYSYE (SEQ ID NO: 601), kYASYE (SEQ ID NO: 602), kLYSYE (SEQ ID NO: 603), KQASYE (SEQ ID NO: 604), KLQSYE (SEQ ID NO: 605), KLASQE (SEQ ID NO: 606), kQASYE (SEQ ID NO: 607), kLQSYE (SEQ ID NO: 608), kLASQE (SEQ ID NO: 609), EGASYK (SEQ ID NO: 610), ELGSYK (SEQ ID NO: 611),

ELASGK (SEQ ID NO: 612), eGASYK (SEQ ID NO: 613), eLGSYK (SEQ ID NO: 614), eLASGK (SEQ ID NO: 615), EAASYK (SEQ ID NO: 616), ELASAK (SEQ ID NO: 617), eAASYK (SEQ ID NO: 618), eLASAK (SEQ ID NO: 619), EVASYK (SEQ ID NO: 620), ELVSYK (SEQ ID NO: 621), ELASVK (SEQ ID NO: 622), eVASYK (SEQ ID NO: 623), eLVSYK (SEQ ID NO: 624), eLASVK (SEQ ID NO: 625), EIASYK (SEQ ID NO: 626), ELISYK (SEQ ID NO: 627), ELASIK (SEQ ID NO: 628), eIASYK (SEQ ID NO: 629), eLISYK (SEQ ID NO: 630), eLASIK (SEQ ID NO: 631), ELASYK (SEQ ID NO: 632), ELLSYK (SEQ ID NO: 633), ELASLK (SEQ ID NO: 634), eLASYK (SEQ ID NO: 635), eLLSYK (SEQ ID NO: 636), eLASLK (SEQ ID NO: 637), EFASYK (SEQ ID NO: 638), ELFSYK (SEQ ID NO: 639), ELASFK (SEQ ID NO: 640), eFASYK (SEQ ID NO: 641), eLFSYK (SEQ ID NO: 642), eLASFK (SEQ ID NO: 643), EWASYK (SEQ ID NO: 644), ELWSYK (SEQ ID NO: 645), ELASWK (SEQ ID NO: 646), eWASYK (SEQ ID NO: 647), eLWSYK (SEQ ID NO: 648), eLASWK (SEQ ID NO: 649), EYASYK (SEQ ID NO: 650), ELYSYK (SEQ ID NO: 651), eYASYK (SEQ ID NO: 652), eLYSYK (SEQ ID NO: 653), EQASYK (SEQ ID NO: 654), ELQSYK (SEQ ID NO: 655), ELASQK (SEQ ID NO: 656), eQASYK (SEQ ID NO: 657), eLQSYK (SEQ ID NO: 658), or eLASQK (SEQ ID NO: 659), or a retro-, an inverso- or a retro-inverso variant thereof.

[0098] In one example in the above sequences, each K (L-lysine) is optionally replaced with a replacement L-amino acid having a side chain comprising an amino group (e.g., -NH₂ group), each k (D-lysine) is optionally replaced with a replacement D-amino acid having a side chain comprising an amino group. Exemplary replacement amino acids for lysine (K) in the above sequences include ornithine (Orn), 2,4-diaminobutyric acid (Dab), and 2,3-diaminopropionic acid (Dap, also referred to as Dpr). In one example, a compound of the present disclosure comprises an amino acid sequence selected from Orn-LASYE (SEQ ID NO: 660), ELASY-Orn (SEQ ID NO: 661), Dab-LASYE (SEQ ID NO: 662), Dap-LASYE (SEQ ID NO: 663), and ELASY-Dap (SEQ ID NO: 664).

[0100] In another example in the above sequences, each E (L-glutamic acid) is optionally and independently replaced with L-aspartic acid (D) or another replacement L-amino acid having a side chain comprising a carboxylic acid (i.e., -COOH) group, and each e (D-glutamic acid) is optionally and independently replaced with D-aspartic acid (d) or another replacement D-amino acid having a side chain comprising a carboxylic acid

group. In certain embodiments, a compound of the present disclosure comprises a sequence selected from: DLASYK (SEQ ID NO: 665), DLASY-Orn (SEQ ID NO: 666), DLASY-Dpr (SEQ ID NO: 667), and Dab-LASYD (SEQ ID NO: 668).

[0101] In certain embodiments, a compound of the present disclosure comprises a sequence selected from: DLASYK (SEQ ID NO: 665) and DLASY-Orn (SEQ ID NO: 666).

[0102] In the above peptides, the amino acid side chains of K or k (or a replacement amino acid of K or k) and E or e (or a replacement amino acid of E or e) are covalently linked via a peptide bond formed between the amino group and the carboxylic acid group to form a lactam ring.

[0103] In other embodiments, a compound of the present disclosure comprises at least one of the following amino acid sequences:

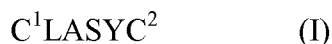
CGASYC (SEQ ID NO: 760), CLGSYC (SEQ ID NO: 761), CLASGC (SEQ ID NO: 762), cGASYC (SEQ ID NO: 763), cLGSYC (SEQ ID NO: 764), cLASGC (SEQ ID NO: 765), CAASYC (SEQ ID NO: 766), CLASAC (SEQ ID NO: 767), cAASYC (SEQ ID NO: 768), cLASAC (SEQ ID NO: 769), CVASYC (SEQ ID NO: 770), CLVSYC (SEQ ID NO: 771), CLASVC (SEQ ID NO: 772), cVASYC (SEQ ID NO: 773), cLVSYC (SEQ ID NO: 774), cLASVC (SEQ ID NO: 775), CIASYC (SEQ ID NO: 776), CLISYC (SEQ ID NO: 777), CLASIC (SEQ ID NO: 778), cIASYC (SEQ ID NO: 779), cLISYC (SEQ ID NO: 780), cLASIC (SEQ ID NO: 781), CLASYC (SEQ ID NO: 782), CLLSYC (SEQ ID NO: 783), CLASLC (SEQ ID NO: 784), cLASYC (SEQ ID NO: 785), cLLSYC (SEQ ID NO: 786), cLASLC (SEQ ID NO: 787), FASYC (SEQ ID NO: 788), CLFSYC (SEQ ID NO: 789), CLASFC (SEQ ID NO: 790), cFASYC (SEQ ID NO: 791), cLFSYC (SEQ ID NO: 792), cLASFC (SEQ ID NO: 793), CWASYC (SEQ ID NO: 794), CLWSYC (SEQ ID NO: 795), CLASWC (SEQ ID NO: 796), cWASYC (SEQ ID NO: 797), cLWSYC (SEQ ID NO: 798), cLASWC (SEQ ID NO: 799), CYASYC (SEQ ID NO: 800), CLYSYC (SEQ ID NO: 801), cYASYC (SEQ ID NO: 802), cLYSYC (SEQ ID NO: 803), CQASYC (SEQ ID NO: 804), CLQSYC (SEQ ID NO: 805), CLASQC (SEQ ID NO: 806), cQASYC (SEQ ID NO: 807), cLQSYC (SEQ ID NO: 808), and cLASQC (SEQ ID NO: 809), CLASSC (SEQ ID NO: 810), CLAsYC (SEQ ID NO: 811), CLASyC (SEQ ID NO: 812), or a retro-, an inverso- or a retro-inverso variant thereof.

[0104] In one example, a compound of the present disclosure comprises the sequence CLASYC.

[0105] In one example in the above sequences, each C (L-cysteine) is optionally and independently replaced with L-homo-cysteine (HCy), L-seleno-cysteine (U), or L-homo-seleno cysteine, and each c (D-cysteine) is optionally and independently replaced with D-homo-cysteine, D-seleno-cysteine (u), or D-homo-seleno cysteine.

[0106] In one example, the invention provides a compound comprising:

(a) an amino acid sequence comprising Formula (I):

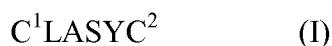


or (b) a retro-, an inverso- or a retro-inverso variant of the amino acid sequence of (a), wherein

[0107] S is L-serine, L is L-leucine, A is L-alanine, Y is L-tyrosine, wherein one or two of L, A and Y are optionally and individually replaced with a replacement L-amino acid. C^1 and C^2 are independently selected from amino acids having a side chain comprising a -S-H group, wherein the side chains of C^1 and C^2 are reversibly linked to form a disulfide bond. In one example, C^2 is L-cysteine and C^1 is selected from L-cysteine, D-cysteine, penicillamine, L-homocysteine, and D-homocysteine. In another example, C^1 and C^2 are both C. An additional L-amino acid is optionally inserted between Y (or a replacement amino acid thereof) and C^2 .

[0108] In another example, the invention provides a compound comprising:

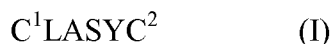
(a) an amino acid sequence comprising Formula (I):



or (b) a retro-, an inverso- or a retro-inverso variant of the amino acid sequence of (a), wherein S is L-serine, L is L-leucine, A is L-alanine, Y is L-tyrosine, wherein one of L, A and Y is optionally and individually replaced with a replacement L-amino acid. C^1 and C^2 are independently selected from cysteine and homo-cysteine. In one example, C^1 is selected from L-cysteine, D-cysteine, L-homocysteine, and D-homocysteine, and C^2 is selected from L-cysteine and L-homocysteine. In another example, C^1 and C^2 are both C.

[0109] In yet another example, the invention provides a compound comprising:

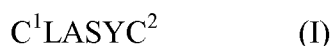
(a) an amino acid sequence comprising Formula (I):



or (b) a retro-, an inverso- or a retro-inverso variant of the amino acid sequence of (a), wherein S is L-serine; L is L-leucine; A is L-alanine; and Y is L-tyrosine. C^1 and C^2 are independently selected from amino acids having a side chain comprising a -S-H group. In one example, C^2 is selected from L-amino acids having a side chain comprising a -S-H group. In another example, C^1 and C^2 are independently selected from cysteine and homocysteine. In yet another example, C^1 is selected from L-cysteine, D-cysteine, L-homocysteine, and D-homocysteine, and C^2 is selected from L-cysteine and L-homocysteine. In another example, C^1 and C^2 are both C.

[0110] In yet another example, the invention provides a compound comprising:

(a) an amino acid sequence comprising Formula (I):



or (b) a retro-, an inverso- or a retro-inverso variant of the amino acid sequence of (a), wherein S is L-serine; L is L-leucine; A is L-alanine; and Y is L-tyrosine. C^1 and C^2 are selected from one of C^1 and C^2 is selected from amino acids having a side chain with a primary or secondary amino group (e.g., -NH₂ group), and the other of C^1 and C^2 is selected from an amino acid with a side chain having a carboxylic acid group (e.g., -COOH group), wherein the amino group and the carboxylic acid group form an amide bond. In one example, C^1 is selected from K, ornithine (Orn), 2,4-diaminobutyric acid (Dab), and 2,3-diaminopropionic acid (Dap, also referred to as Dpr), and C^2 is selected from glutamic acid (E) and aspartic acid (D). In another example, C^2 is selected from K, ornithine (Orn), 2,4-diaminobutyric acid (Dab), and 2,3-diaminopropionic acid (Dap, also referred to as Dpr), and C^1 is selected from glutamic acid (E) and aspartic acid (D).

[0111] In yet another example, the invention provides a compound comprising:

(a) an amino acid sequence comprising Formula (III):



or (b) a retro-, an inverso- or a retro-inverso variant of the amino acid sequence of (a).

[0112] In Formula (III), S, C^1 and C^2 are defined as for Formula (I) according to any of the above embodiments. In one example, S in Formula (III) is L-serine. In another example in Formula (III), C^1 and C^2 are independently selected from amino acids having a side chain comprising a -S-H group or a -Se-H group. In another example in Formula (III), C^1 is selected from amino acids having a side chain comprising a -S-H group or a -Se-H group, and C^2 is selected from L-amino acids having a side chain comprising a -S-H

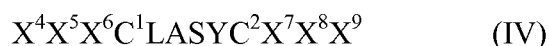
group or a -Se-H group. In another example, C¹ and C² are independently selected from cysteine and homo-cysteine. In another example, C¹ and C² are independently selected from cysteine and homo-cysteine.

[0113] In Formula (III), X¹, X² and X³ are independently selected from amino acids (e.g., L-amino acids) having a hydrophobic or a polar uncharged side chain. In one example, in Formula (III), X¹, X² and X³ are independently selected from G, A, V, I, L, M, F, W, Y, S, T, N, Q, and derivatives thereof.

[0114] In another example according to any of the above embodiments, X¹ is selected from A, L, M, V, and Q. In another example, X¹ is L. In another example according to any of the above embodiments, X² is selected from G, A, alpha-aminobutyric acid (Abu), V, L, I and S. In yet another example, X² is selected from G, A, alpha-aminobutyric acid (Abu), V, L, and I. In another example, X² is A. In another example according to any of the above embodiments, X³ is selected from A, V, Y, Q, F. In another example, X³ is Y.

[0115] In another example, a compound of the present disclosure includes:

(a) an amino acid sequence comprising Formula (IV):



or (b) a retro-, an inverso- or a retro-inverso variant of the amino acid sequence of (a).

[0116] In Formula (IV), L, A, S, Y, C¹ and C² are defined as for Formula (I) or any embodiment thereof.

[0117] In Formula (IV), X⁴, X⁵, X⁷, X⁸ and X⁹ are either absent or present, and when present are independently selected from amino acids. In one example, all of X⁴, X⁵, X⁷, X⁸ and X⁹ in Formula (IV) are present.

[0118] In one example in Formula (IV), X⁴ is either absent or present and, when present, is N, Q, an amino acid having a side chain comprising a basic moiety, or a modified (e.g., alpha-N-alkylated) amino acid thereof. In one example, X⁴ in Formula (IV) is an amino acid having a side chain comprising a basic moiety. In one example, X⁴ in Formula (IV) is selected from lysine (K), arginine (R), histidine (H), 2,4-diaminobutyric acid (Dab), 2,3-diaminopropionic acid (Dpr), ornithine (Orn), 2,7-diamino-heptanoic acid, 2,8-diamino-octanoic acid, and modified (e.g., alpha-N-alkylated) amino acids thereof. In another example X⁴ in Formula (IV) is lysine or N-methyl lysine. In another example, X⁴ in Formula (IV) is L-lysine (K).

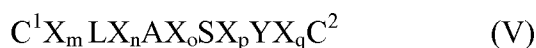
- [0119] In one example in Formula (IV), X^5 is either absent or present and, when present, is an amino acid having a hydrophobic or a polar uncharged side chain, or an alpha-N-alkylated amino acid thereof. In one example X^5 is selected from L, V, M, P, and modified (e.g., alpha-N-alkylated) amino acids thereof. In one example, X^5 in Formula (IV) is leucine or alpha-N-alkylated leucine. In another example, X^5 is L-leucine (L). In yet another example, X^5 is other than E and R.
- [0120] In Formula (IV), X^6 is an amino acid. In one example, X^6 is a modified (e.g., alpha-N-alkylated) amino acid. In another example, X^6 in Formula (IV) is selected from L-threonine, D-threonine, A, S, Q, R, and K. In one example, X^6 is selected from L-threonine and D-threonine, A, and K. In another example, X^6 is selected from L-threonine and D-threonine. In another example X^6 is T. In yet another example, X^6 is other than an amino acid with an acidic side chain (e.g., other than E).
- [0121] In another example in Formula (IV), X^7 , X^8 and X^9 are independently either absent or present and, when present, are independently selected from amino acids (e.g., L-amino acids). In another example in Formula (IV), X^7 , X^8 and X^9 , when present, are independently selected from amino acids having a hydrophobic side chain and amino acids having a polar uncharged side chain. In another example in Formula (IV), X^7 , X^8 and X^9 , when present, are independently selected from L-amino acids having a hydrophobic side chain and amino acids having a polar uncharged side chain. In yet another example in Formula (IV), X^7 , X^8 and X^9 are independently either absent or present and when present are independently selected from L, norleucine (Nle), I, V, M, F, W, Y, S, T, N, and Q.
- [0122] In one example, according to any of the above embodiments, X^7 is selected from G, L, Q and amino acids having a side chain containing an aromatic moiety. In another example, X^7 is selected from G, L, Q, W, F, Y, and 1-aryl-alanine (e.g., 1-naphthyl-alanine). In another example, X^7 is F. In another example, X^7 is L-tryptophan, D-tryptophan, F, L, or 1-naphthyl-alanine (e.g., L-1-naphthyl-alanine). In another example, X^7 is not an NMe amino acid. In another example, X^7 is 1-naphthyl-alanine. In yet another example, X^7 is L-tryptophan or D-tryptophan. In another example, X^7 is W. In another example, X^7 is Y. In a further example, X^7 is modified tyrosine as defined herein.
- [0123] In one example X^8 in Formula (IV) is selected from amino acids having a hydrophobic side chain and amino acids having a polar uncharged side chain. In one

example, X^8 is selected from L, I, nor-leucine (Nle), V, Y, Q, and M. In another example, X^8 is selected from hydrophobic L-amino acids. In one example, X^8 is selected from L, Y, I and L-norleucine. In another example, X^8 is L.

[0124] In one example, X^9 in Formula (IV) is selected from F, V, and L. In another example, X^9 in Formula (IV) is V. In another example, X^9 in Formula (IV) is selected from F and L. In another example, X^9 in Formula (IV) is F. I

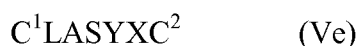
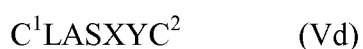
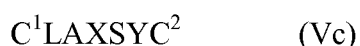
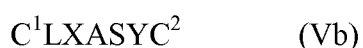
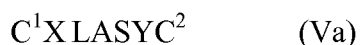
[0125] In one example, a compound of the present disclosure comprises:

(a) an amino acid sequence comprising Formula (V):



or (b) a retro-, an inverso- or a retro-inverso variant of the amino acid sequence of (a). In Formula (V), L is L-leucine; A is L-alanine; S is L-serine; Y is L-tyrosine, wherein one or two of L, A, S, and Y are optionally replaced with an independently selected replacement amino acid. In Formula (V), each X is independently selected from amino acids. In one example, each X is independently selected from amino acids having a hydrophobic or a polar uncharged side chain. In Formula (V), m, n, o, p, and q are integers independently selected from 1 and 0. In one example, m, n, o, p, and q are all zero. In another example, one of m, n, o, p, and q is 1 and the remaining of m, n, o, p, and q are zero. In Formula (V), C^1 and C^2 are independently selected from amino acids having a side chain comprising a -S-H group or a -Se-H group. In one example in Formula (V), both of C^1 and C^2 are C.

[0126] In another example according to the above embodiment, a compound of the present disclosure comprises an amino acid sequence comprising Formula (Va), Formula (Vb), Formula (Vc), Formula (Vd), or Formula (Ve):

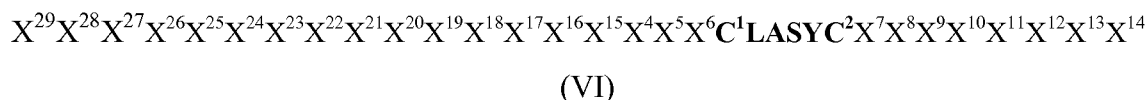


wherein X represents an amino acid. In one example in the above formulae, X is selected from amino acids having a hydrophobic or a polar uncharged side chain.

[0127] In one example according to any of the above embodiments, the amino acid sequence is (b) the retro-inverso variant of the amino acid sequence of (a).

[0128] In one example, a compound of the present disclosure comprises:

(a) an amino acid sequence comprising Formula (VI):



[0129] or (b) a retro-, an inverso- or a retro-inverso variant of the amino acid sequence of

(a). In Formula (VI), L, A, S, and Y are defined as for Formula (I). In one example, in Formula (VI), L is L-leucine; A is L-alanine; S is L-serine; Y is selected from A, F, L-tyrosine, D-tyrosine, L-tyrosine(OMe), and D-tyrosine(OMe). In one example, in Formula (VI), L is L-leucine; A is L-alanine; S is L-serine, and Y is L-tyrosine.

[0130] X^{29} (position 1) is either absent or present and when present is an amino acid. In one example X^{29} is S or P.

[0131] X^{28} (position 2) is either absent or present and when present is an amino acid. In one example X^{28} is selected from a basic amino acid (e.g., R) and S.

[0132] X^{27} (a position 3) is either absent or present and when present is an amino acid. In one example X^{27} is S or I.

[0133] X^{26} (at position 4) is either absent or present and when present is an amino acid. In one example X^{26} is selected from basic amino acids (e.g., L- or D-arginine).

[0134] X^{25} (position 5) is either absent or present and when present is an amino acid. In one example X^{25} is T or S.

[0135] X^{24} (position 6) is either absent or present and when present is an amino acid. In one example X^{24} is V or S.

[0136] X^{23} (position 7) is either absent or present and when present is an amino acid. In one example X^{23} is G or S.

[0137] X^{22} (position 8) is either absent or present and when present is an amino acid. In one example X^{22} is S or P.

[0138] X^{21} (position 9) is either absent or present and when present is an amino acid. In one example X^{21} is G or S.

[0139] X^{20} (position 10) is either absent or present and when present is an amino acid. In one example X^{20} is S.

[0140] X^{19} (position 11) is an amino acid. In one example X^{19} is a basic amino acid (e.g., L-arginine, D-arginine, or K).

- [0141] X^{18} (position 12) is an amino acid. In one example X^{18} is L-arginine, D-arginine or S.
- [0142] X^{17} (position 13) is an amino acid. In one example X^{17} is a hydrophobic amino acid. In another example, X^{17} is A or NMe-alanine.
- [0143] X^{16} (position 14) is an amino acid. In one example X^{16} is S, P or A. In another example X^{16} is not K or a D-amino acid (e.g., not D-proline).
- [0144] X^{15} (position 15) is an amino acid. In one example X^{15} is G, sarcosine, or A. In another example X^{15} is not K. In another example, X^{15} is selected from G and sarcosine.
- [0145] X^4 (position 16) is L-lysine or D-lysine, L-ornithine, D-ornithine, L-2,4-diaminobutyric acid, D-2,4-diaminobutyric acid, L-2,3-diaminopropionic acid, or D-2,3-diaminopropionic acid. In one example X^4 is selected from L-lysine and D-lysine.
- [0146] X^5 (position 17) is selected from hydrophobic amino acids. In one example, X^5 is selected from A, L-leucine, D-leucine, and S. In one example, X^5 is L.
- [0147] X^6 (position 18) is selected from L-threonine, D-threonine, A, Q, and K. In one example, X^6 is selected from T, A, Q, and K. In another example, X^6 is selected from L-threonine and D-threonine. In another example X^6 is T.
- [0148] C^1 (position 19) is selected from L-cysteine, D-cysteine, penicillamine, L-homocysteine, and D-homocysteine. In one example, C^1 is C.
- [0149] C^2 (position 24) is selected from L-cysteine, D-cysteine, penicillamine, L-homocysteine, and D-homocysteine. In one example, C^2 is C.
- [0150] In one example, in Formula (VI) both of C^1 and C^2 are C.
- [0151] X^7 (position 25) is selected from L-tryptophan, D-tryptophan, F, L or 1-naphthyl-alanine (e.g., L-1-naphthyl-alanine). In another example, X^7 is not an NMe amino acid. In another example, X^7 is W.
- [0152] X^8 (position 26) is selected from hydrophobic L-amino acids. In one example, X^8 is selected from L, Y, I and L-norleucine. In another example, X^8 is L.
- [0153] X^9 (position 27) is selected from F and L. In one example, X^9 is F.
- [0154] X^{10} (position 28) is either absent or present and when present is selected from W, A, S, F and L. In one example, X^{10} is W.
- [0155] X^{11} (position 29) is either absent or present, and when present is an amino acid. In one example, X^{11} is selected from L-threonine, D-threonine, and S.

- [0156] X^{12} (position 30) is either absent or present, and when present is an amino acid. In one example, X^{12} is selected from G, A, sarcosine, L, F and S. In one example, X^{12} is selected from G, A, and sarcosine. In one example, X^{12} is selected from G and sarcosine. In another example, X^{12} is G.
- [0157] X^{13} (position 31) is either absent or present, and when present is an amino acid. In one example, X^{13} is selected from I, L, F, L-norleucine, L-1-naphthyl-alanine, 3-cyclohexyl-L-alanine, and L-tert-leucine. In another example, X^{13} is selected from I, L, F, L-norleucine, and L-1-naphthyl-alanine. In one example, X^{13} is I.
- [0158] X^{14} (position 32) is either absent or present, and when present is an amino acid. In one example, X^{14} is A.
- [0159] In one example, all of X^{10} to X^{14} are present. In another example, all of X^{20} to X^{29} are present.
- [0160] In one example, the amino acid sequence of a compound of the present disclosure includes at least one of the following sequences:

KLTCCLASYCWLF
 k-MeLeu-TCLASYCWLF
 RRAPGKLQCLASYCWLFWTGIA
 RRAPGKLTCLASYCWLFWTGIA
 rRAPGKLTCLASYCWLFWTGIA
 rRAPGKSTCLASYCWLFWTGIA
 PRIRTVGPGSRSASGKLTCLASYCWLFWTGIA
 PRIrTVGPGSrSASGKLTCLASYCWLFWTGIA
 PRIRTVGPGSRSASGKSTCLASYCWLFWTGIA
 SRIRTVGPGSRSASGKSTCLASYCWLFWTGIA
 PRIRTVSPGSRASGKSTCLASYCWLFWTGIA
 SRIRTVSPGSRASGKSTCLASYCWLFWTGIA
 PRSRTVGPGRSASGKSTCLASYCWLFWTGIA
 SRSRTVSPGSRASGKSTCLASYCWLFWTGIA
 PRIrTVGPGSrSASGKSTCLASYCWLFWTGIA
 SKQGRPISPDRRAAGKLTCLASYCWLFWTGIA
 SKQGRPISPDrrAAGKLTCLASYCWLFWTGIA
 RRAPGKLTCLASYCWLFSGISLSRAPESAAP

RRFVGGSLSQRRAPGKLTCLASYCWLFWTGIA
PQTRDPSSRDRRAPGKLTCLASYCWLFWTGIA.

[0161] Additional amino acid residues may be added to the N- or C-terminus of the above sequences to form a compound of the present disclosure.

[0162] In various embodiments, a compound of the present disclosure includes an amino acid sequence incorporating a particular number of amino acid residues. The number of amino acids is defined by a lower and an upper limit. The length of the amino acid sequence can be defined by any combination of the lower and upper limits given below:

Lower limit

[0163] In one example according to any of the above embodiments, the amino acid sequence of a compound comprises at least 7, at least 8, at least 9, or at least 10 amino acids. In another example according to any of the above embodiments, the amino acid sequence of a compound comprises at least 11, at least 12, at least 13, at least 14, at least 15, at least 16, at least 17, at least 18, at least 19, or at least 20 amino acids. In another example according to any of the above embodiments, the amino acid sequence of a compound comprises at least 21, at least 22, at least 23, at least 24, at least 25, at least 26, at least 27, at least 28, at least 29, at least 30 amino acids, at least 31 amino acids, or at least 32 amino acids.

Upper limit

[0164] In one example according to any of the above embodiments, the amino acid sequence of a compound comprises not more than 500, not more than 400, not more than 300, not more than 200, or not more than 100 amino acids. In another example according to any of the above embodiments, the amino acid sequence of a compound comprises not more than 90, not more than 80, not more than 70, not more than 60, not more than 50, or not more than 40 amino acids.

Exemplary Ranges

[0165] In yet another example, the amino acid sequence of a compound comprises at least 9 and not more than 500 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 10 and not more than 500 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 11 and not more than 500 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 12 and not more than 500 amino acids. In yet another example, the

100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 12 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 13 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 14 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 15 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 16 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 17 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 18 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 19 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 20 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 21 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 22 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 23 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 24 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 25 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 26 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 27 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 28 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 29 and not more than 100 amino acids. In yet another example, the amino acid sequence of a compound comprises at least 30 and not more than 100 amino acids

[0168] In one example according to any of the above embodiments, a compound of the present disclosure contains not more than 500, not more than 400, not more than 300, not more than 200, or not more than 100 amino acids. In another example according to any of the above embodiments, a compound comprises not more than 90, not more than 80, not more than 70, not more than 60, or not more than 50 amino acids. In one example

according to any of the above embodiments, a compound comprises not more than 500, not more than 400, not more than 300, not more than 200, or not more than 100 consecutive amino acids.

[0169] In yet another example, a compound comprises at least 9 and not more than 500 amino acids. In yet another example, a compound comprises at least 10 and not more than 500 amino acids. In yet another example, a compound comprises at least 11 and not more than 500 amino acids. In yet another example, a compound comprises at least 12 and not more than 500 amino acids. In yet another example, a compound comprises at least 13 and not more than 500 amino acids. In yet another example, a compound comprises at least 14 and not more than 500 amino acids. In yet another example, a compound comprises at least 15 and not more than 500 amino acids. In yet another example, a compound comprises at least 16 and not more than 500 amino acids. In yet another example, a compound comprises at least 17 and not more than 500 amino acids. In yet another example, a compound comprises at least 18 and not more than 500 amino acids. In yet another example, a compound comprises at least 19 and not more than 500 amino acids. In yet another example, a compound comprises at least 20 and not more than 500 amino acids.

[0170] In yet another example, a compound comprises at least 9 and not more than 300 amino acids. In yet another example, a compound comprises at least 10 and not more than 300 amino acids. In yet another example, a compound comprises at least 11 and not more than 300 amino acids. In yet another example, a compound comprises at least 12 and not more than 300 amino acids. In yet another example, a compound comprises at least 13 and not more than 300 amino acids. In yet another example, a compound comprises at least 14 and not more than 300 amino acids. In yet another example, a compound comprises at least 15 and not more than 300 amino acids. In yet another example, a compound comprises at least 16 and not more than 300 amino acids. In yet another example, a compound comprises at least 17 and not more than 300 amino acids. In yet another example, a compound comprises at least 18 and not more than 300 amino acids. In yet another example, a compound comprises at least 19 and not more than 300 amino acids. In yet another example, a compound comprises at least 20 and not more than 300 amino acids.

[0171] In yet another example, a compound comprises at least 9 and not more than 100 amino acids. In yet another example, a compound comprises at least 10 and not more than 100 amino acids. In yet another example, a compound comprises at least 11 and not more than 100 amino acids. In yet another example, a compound comprises at least 12 and not more than 100 amino acids. In yet another example, a compound comprises at least 13 and not more than 100 amino acids. In yet another example, a compound comprises at least 14 and not more than 100 amino acids. In yet another example, a compound comprises at least 15 and not more than 100 amino acids. In yet another example, a compound comprises at least 16 and not more than 100 amino acids. In yet another example, a compound comprises at least 17 and not more than 100 amino acids. In yet another example, a compound comprises at least 18 and not more than 100 amino acids. In yet another example, a compound comprises at least 19 and not more than 100 amino acids. In yet another example, a compound comprises at least 20 and not more than 100 amino acids. In yet another example, a compound comprises at least 21 and not more than 100 amino acids. In yet another example, a compound comprises at least 22 and not more than 100 amino acids. In yet another example, a compound comprises at least 23 and not more than 100 amino acids. In yet another example, a compound comprises at least 24 and not more than 100 amino acids. In yet another example, a compound comprises at least 25 and not more than 100 amino acids. In yet another example, a compound comprises at least 26 and not more than 100 amino acids. In yet another example, a compound comprises at least 27 and not more than 100 amino acids. In yet another example, a compound comprises at least 28 and not more than 100 amino acids. In yet another example, a compound comprises at least 29 and not more than 100 amino acids. In yet another example, a compound comprises at least 30 and not more than 100 amino acids.

[0172] In one example, a compound of the present disclosure is a peptide or peptide derivative containing at least 21 amino acids, wherein amino acid 21 (counted from the N-terminus towards the C-terminus) is an amino acid with a hydrophobic side chain (e.g., G, A, L, I). In another example, a compound is a peptide or peptide derivative containing at least 21 amino acids, wherein the peptide has a positively charged N-terminal amino acid (i.e., basic amino acid, such as K or R). In another example a compound of the present disclosure is a peptide or peptide derivative having a neutral C-terminus. For

example, the final 6, final 5, final 4, final 3, final 2 or the final C-terminal amino acids are selected from amino acids having a hydrophobic or a polar uncharged side chain. In one example, the C-terminal amino acid is A. In another example, the C-terminal 3 amino acids are -GIA. In another example, the C-terminal 4 amino acids are -TGIA. In another example, the C-terminal 5 amino acids are -WTGIA. In another example, the C-terminal 6 amino acids are -FWTGIA. In another example, at least one of the N-terminal 3 amino acids is a D-amino acid. In one example, a compound of the present disclosure is a peptide or peptide derivative, wherein the N-terminal amino acid is D-arginine (r). In another example, the N-terminal two amino acids of the peptide are Rr- or rR-.

[0173] In one example according to any of the above embodiments, the amino acid sequence of a compound is acylated, e.g., acetylated at the N-terminus (i.e., -NHCOCH₃ or -NHAc). In another example according to any of the above embodiments, the amino acid sequence of a compound is amidated (i.e., -CONHCH₃ or -CONH₂) at the C-terminus. In yet another example according to any of the above embodiments, the amino acid sequence of a compound is acylated, e.g., acetylated at the N-terminus and amidated (i.e., -CONHCH₃ or -CONH₂) at the C-terminus. In a further example according to any of the above embodiments, the amino acid sequence of a compound has a free amino terminus (-NH₂ or a salt form thereof) and is amidated (i.e., -CONH₂) at the C-terminus.

[0174] In one example according to any of the above embodiments, a compound of the present disclosure is a peptide or a peptide derivative. In another example, a compound is a peptide derivative, in which the N-terminal amino acid is acylated, e.g., acetylated (i.e., -NHCOCH₃). In another example, a compound is a peptide derivative, in which the C-terminal amino acid is amidated (i.e., -CONHCH₃ or -CONH₂). In one example, a compound is a peptide derivative, which is acylated, e.g., acetylated at the N-terminus and amidated (i.e., -CONHCH₃ or -CONH₂) at the C-terminus. In one example, a compound is a peptide derivative, which has a free N-terminus (-NH₂ or a salt form thereof) and is amidated (i.e., -CONH₂) at the C-terminus.

[0175] In one example, a compound of the present disclosure contains an amino acid sequence selected from or is a peptide selected from those listed in Table 1, hereinbelow, or a retro-, an inverso- or a retro-inverso variant thereof. In one example in those peptides, the N-terminus is acetylated. In another example in those peptides, the N-terminus is free (-NH₂). In another example in those peptides, the C-terminus is amidated.

In another example in those peptides, the N-terminus is free and the C-terminus is amidated.

Conjugates

Compounds Linked to a Heterologous moiety

[0176] In certain embodiments, the compound of the present disclosure is linked to a heterologous moiety. For example, the pro-coagulant peptide of the present disclosure is covalently linked to a heterologous moiety, optionally via a linker (L_1) thereby forming a conjugate. Linker (L_1) is different than the linking moiety Z defined herein above.

[0177] The heterologous moiety is useful to increase the bioavailability or the *in vivo* stability/half-life of the compound. Exemplary heterologous moieties include, e.g., known half-life extending moieties, e.g., water-soluble polymers, such as polyethylene glycol (PEG) and polypropylene glycol (PPG), Fc, PAS, HES, XTEN, and albumin. In one example, the heterologous moiety is a polymer, e.g., a water-soluble polymer, such as polyethylene glycol (PEG). In another example, the heterologous moiety is a half-life prolonging protein, such as albumin. In another example, the heterologous moiety is an Fc moiety. In another example, the heterologous moiety is a XTEN moiety. In another example, the heterologous moiety is a HES moiety. In another example, the heterologous moiety is a PAS moiety. Other useful heterologous moieties are known in the art and others are further described herein.

[0178] In one embodiment the conjugate formed between the compound and the heterologous moiety has a structure according to Formula (A1) or Formula (A2):



wherein

Het is a heterologous moiety as described herein;

m is an integer selected from 0 and 1;

L_1 is either absent ($m=0$) or present ($m=1$), and when present is a linker as described herein; and

Pep is a compound (e.g., pro-coagulant peptide or peptide derivative) of the present disclosure.

Polypeptide Conjugates

- [0179] In other embodiments, the compound of the present disclosure is covalently linked to a polypeptide. In one example, the polypeptide is selected from a blood coagulation factor and platelet targeting moieties. In other embodiments, the blood coagulation factor is selected from FVIIa, FVIII, and FIX. In one example, the compound is linked to an internal amino acid residue of the polypeptide (e.g., FVIIa or FIX).
- [0180] The present disclosure further provides polypeptide conjugates comprising a polypeptide selected from FVIII, FIX, FVIIa, and platelet targeting moieties, and a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative), wherein the compound is linked, e.g., covalently linked, to the polypeptide, optionally via a linker.
- [0181] In another embodiment, the present disclosure provides conjugates comprising a polypeptide selected from FVIII, FIX, FVIIa, and platelet targeting moieties, a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative), and at least one linker, which links the compound to the polypeptide.
- [0182] In another example, the present disclosure provides polypeptide conjugates comprising a polypeptide, a heterologous moiety, a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative), and at least one linker, which covalently links the compound to the polypeptide and the heterologous moiety. In one example according to this embodiment, the compound (e.g., peptide or peptide derivative) is interposed between the polypeptide and the heterologous moiety. In another example according to this embodiment, the heterologous moiety is linked to the polypeptide, and the compound (e.g., peptide or peptide derivative) is linked to either the polypeptide or the heterologous moiety.
- [0183] In one example, the polypeptide is activatable, e.g., by an enzyme, which, e.g., cleaves a number of amino acids from the polypeptide sequence. In one example, the polypeptide is activatable by an enzyme of the blood coagulation cascade, e.g., thrombin. In one example, the polypeptide is a thrombin activatable FVII or FVIIa polypeptide. Exemplary thrombin-activatable FVII polypeptides are disclosed in WO2012/006635, incorporated herein by reference in its entirety.

FVIII Conjugates

- [0184] In various embodiments, the compound of the present disclosure (e.g., peptide or peptide derivative) is covalently linked to FVIII or a FVIII-heterologous moiety construct.
- [0185] In one example, the conjugate of the present disclosure includes a FVIII-heterologous moiety construct (e.g., FVIII-Fc, FVIII-albumin, FVIII-PEG) and the compound (e.g., peptide or peptide derivative) is covalently linked to the FVIII-portion of the construct. In another example, the conjugate of the present disclosure includes a FVIII-heterologous moiety construct (e.g., FVIII-Fc, FVIII-albumin, FVIII-PEG) and the compound (e.g., peptide or peptide derivative) is covalently linked to the heterologous moiety portion of the construct.
- [0186] In one example according to the above embodiments, the heterologous moiety is Fc. Accordingly, the present disclosure provides a conjugate comprising a FVIII-Fc construct (FVIII-Fc) and a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative), and a linker, which covalently links the compound (e.g., peptide or peptide derivative) to the FVIII-Fc. In one example according to this embodiment, the compound (e.g., peptide or peptide derivative) is covalently linked to the FVIII portion of the FVIII-Fc (e.g., via a linker). In another example, the compound (e.g., peptide or peptide derivative) is covalently linked to the Fc portion of the FVIII-Fc (e.g., via a linker).
- [0187] In one example according to any of the above embodiments, the FVIII is B-domain deleted FVIII.
- [0188] In one example, the compound of the present disclosure (e.g., the pro-coagulant peptide or peptide derivative) is covalently linked to the N-terminus of the FVIII heavy chain (HC). In another example, the compound is covalently linked to the C-terminus of the FVIII HC. In yet another example, the compound (e.g., the pro-coagulant peptide or peptide derivative) is covalently linked to the N-terminus of the FVIII light chain (LC). In yet another example, the compound (e.g., the pro-coagulant peptide or peptide derivative) is covalently linked to the C-terminus of the FVIII LC.
- [0189] In a further example according to any of the above embodiments, the compound (e.g., the pro-coagulant peptide or peptide derivative) is covalently linked to an internal amino acid residue of the FVIII molecule (internal conjugation), e.g., via a cysteine residue. In one example, the cysteine residue is engineered into the FVIII amino acid

sequence. In one example, the site of internal conjugation is selected from those described in Mei, B. *et al.* Rational design of a fully active, long-acting PEGylated FVIII for hemophilia A treatment. *Blood* (2010) 116:270-279; and U.S. Patent Application US2006/0115876 to Pan C. *et al.* (Site-directed modification of FVIII), each of which is incorporated herein by reference in its entirety.

[0190] Exemplary FVIII conjugates are illustrated in Figures 20 and 21.

[0191] Figure 20 illustrates various conjugates of the present disclosure, in which a compound (indicated as a peptide or peptide derivative) is covalently linked to a FVIII-heterologous moiety construct, wherein the heterologous moiety is represented in this figure as an Fc, optionally via a linker, wherein the peptide can be linked to the FVIII portion of the FVIII-fusion (e.g., constructs E3, E5-E7), to the heterologous moiety portion of the FVIII-fusion (constructs E1 and E2), or can be interposed between the FVIII and the heterologous moiety of the FVIII-fusion (E4). In constructs E1 and E2, in which the heterologous moiety is shown as an Fc, FVIII is linked to one chain of the Fc and the peptide or peptide derivative is linked to the other (free) Fc chain. In other constructs similar to E2, compound can be placed on either or both of the Fc chains. In constructs E3-E6, the compound is linked to the N- or C-terminal amino acid of the FVIII heavy chain (HC) or light chain (LC), respectively. In construct E7, the peptide or peptide derivative is covalently linked to an internal amino acid residue of the FVIII molecule (e.g., cysteine). It is understood that in other conjugates, the heterologous moiety (represented in this figure as Fc) can be e.g., PEG, PPG, albumin, XTEN, etc. In one example, constructs E1-E6 of Figure 20 can be made recombinantly. In another example, constructs E1 and E7 of Figure 20 can be made semi-recombinantly e.g., as illustrated in Figures 23 and 24.

[0192] Figure 21 illustrates various conjugates of the present disclosure, in which a compound (indicated as a peptide or peptide derivative) is covalently linked to a FVIII protein. In constructs F1-F4 the pro-coagulant peptide is linked to the N- or C-terminal amino acid of the FVIII heavy chain (HC) or light chain (LC), respectively. In construct F5, the peptide or peptide derivative is covalently linked to an internal amino acid residue of the FVIII. In one example, constructs F1-F4 of Figure 21 can be made recombinantly. In another example, construct F5 can be made semi-recombinantly as illustrated in Figures 23 and 24.

FVIIa Conjugates

- [0193] In various embodiments, the compound of the present disclosure (e.g., peptide or peptide derivative) is covalently linked to FVIIa or a FVIIa-heterologous moiety construct.
- [0194] In one example, the conjugate of the present disclosure includes a FVIIa-heterologous moiety construct (e.g., FVIIa-Fc, FVIIa-albumin, FVIIa-PEG, FVIIa-XTEN) and the compound (e.g., peptide or peptide derivative) is covalently linked to the FVIIa-portion of the construct. In another example, the conjugate of the present disclosure includes a FVIIa-heterologous moiety construct (e.g., FVIIa-Fc, FVIIa-albumin, FVIIa-PEG, FVIIa-XTEN) and the compound (e.g., peptide or peptide derivative) is covalently linked to the heterologous moiety portion of the construct.
- [0195] In one example according to the above embodiments, the heterologous moiety is Fc. Accordingly, the present disclosure provides a conjugate comprising a FVIIa-Fc construct (FVIIa-Fc) and a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative), and a linker, which covalently links the compound to the FVIIa-Fc. In one example according to this embodiment, the compound is covalently linked to the FVIIa portion of the FVIIa-Fc (e.g., via a linker). In another example according to this embodiment, the compound is covalently linked to the Fc portion of the FVIIa-Fc (e.g., via a linker). These optional linkers can be cleavable linkers as described herein elsewhere.
- [0196] In one example according to any of the above embodiments, the compound is covalently linked to the C-terminus of the FVIIa HC (e.g., the compound is interposed between the heterologous moiety and the FVIIa HC). In yet another example according to any of the above embodiments, the compound is covalently linked to the C-terminus of the FVIIa LC. In another example according to any of the above embodiments, the compound is covalently linked to the N-terminus of the FVIIa heavy chain (HC) with a cleavable linker to allow for conversion to an active protease. In a further example according to any of the above embodiments, the compound (e.g., the pro-coagulant peptide or peptide derivative) is covalently linked to an internal amino acid residue of the FVIIa molecule (internal conjugation), e.g., via a cysteine residue. In one example, the cysteine residue is engineered into the FVIIa amino acid sequence, e.g., according to the procedures described in Mei, B. *et. al.* Rational design of a fully active, long-acting

PEGylated FVIIa for hemophilia A treatment. *Blood* (2010) 116:270-279; and U.S. Patent Application US2006/0115876 to Pan C. *et al.* (Site-directed modification of FVIIa), each of which is incorporated herein by reference in its entirety.

[0197] Exemplary FVIIa conjugates are illustrated in Figures 18 and 19.

[0198] Figure 18 illustrates various conjugates of the present disclosure, in which a compound (shown as a peptide or peptide derivative) is covalently linked to a FVIIa heterologous moiety construct, wherein the heterologous moiety is represented in this figure as an Fc, optionally via a linker, wherein the peptide can be linked to the FVIIa portion of the FVIIa-fusion (e.g., constructs C4-C7), to the heterologous moiety portion of the FVIIa-fusion (constructs C1 and C2), or can be interposed between the FVIIa and the heterologous moiety of the FVIIa-fusion (C3). In constructs C1 and C2, in which the heterologous moiety is shown as an Fc, FVIIa is linked to one chain of the Fc and the peptide or peptide derivative is linked to the other (free) Fc chain. In other constructs similar to E2, compound can be placed on either or both of the Fc chains. In constructs C3 and C4, the peptide or peptide derivative is linked to the C-terminal amino acid of the FVIIa heavy chain (HC) or light chain (LC), respectively; for C3, the peptide or peptide derivative is also linked to the N-terminus of the heterologous moiety. In construct C5, the peptide or peptide derivative is linked to the N-terminus of the HC; in this case in particular, the linker could be cleavable by proteases activated during the clotting cascade (such as disclosed in International Patent Application No. PCT/US2011/043599 filed July 11, 2011), in order to generate the free N-terminus of the HC required for protease activity. In constructs C6 and C7, the peptide or peptide derivative is covalently linked to an internal amino acid residue (e.g., cysteine) of the FVIIa molecule HC or LC, respectively. It is understood that in other conjugates, the heterologous moiety (represented in this figure as Fc) can be e.g., PEG, PPG, albumin, XTEN, etc. In one example, constructs C1-C5 can be made recombinantly. In another example, constructs C1, C6 and C7 can be made semi-recombinantly e.g., as illustrated in Figures 23 and 24.

[0199] Figure 19 illustrates various conjugates of the present disclosure, in which a peptide or peptide derivative (e.g., a pro-coagulant peptide or peptide derivative, such as those disclosed herein) is covalently linked to a FVIIa protein. In constructs D1-D2, the peptide or peptide derivative is linked to the C-terminal amino acid of the FVIIa heavy chain (HC) or light chain (LC), respectively. In construct D3, the peptide or peptide

derivative is linked to the N-terminus of the HC; in this case in particular, the linker could be cleavable by proteases activated during the clotting cascade (such as disclosed in International Patent Application No. PCT/US2011/043599 filed July 11, 2011), in order to generate the free N-terminus of the HC required for protease activity. In constructs D4 and D5, the peptide or peptide derivative is covalently linked to an internal amino acid residue of the FVIIa HC or LC, respectively. In one example, constructs D1-D3 can be made recombinantly. In another example, constructs D4 and D5 can be made semi-recombinantly as illustrated in Figures 23 and 24.

FIX Conjugates

[0200] In various embodiments, the compound of the present disclosure (e.g., peptide or peptide derivative) is covalently linked to FIX or a FIXa-heterologous moiety construct.

[0201] In one example, the conjugate of the present disclosure includes a FIX-heterologous moiety construct (e.g., FIX-Fc, FIX-albumin, FIX-PEG) and the compound (e.g., peptide or peptide derivative) is covalently linked to the FIX-portion of the construct. In another example, the conjugate of the present disclosure includes a FIX-heterologous moiety construct (e.g., FIX-Fc, FIX-albumin, FIX-PEG) and the compound (e.g., peptide or peptide derivative) is covalently linked to the heterologous moiety portion of the construct, optionally via a linker.

[0202] In one example, the linker of the FIX conjugate is sufficiently long for the compound (e.g., the pro-coagulant peptide or peptide derivative) to bind to the FIX (e.g., once it is converted to FIXa) at an amino acid sequence around Tyr177 (FIXa numbering) as described herein. Such binding of the compound (e.g., peptide or peptide derivative) to the FIX can increase the catalytic activity of the FIXa or its inactivated form FIX. In another example, the linker is sufficiently long for the compound (e.g., peptide or peptide derivative) to be capable of binding to the amino acid sequence: MFCAG (SEQ ID NO: 1) of FIX or FIXa. In another example, the linker is sufficiently long to be capable of interacting with the amino acid sequence: YNNMFCAGFHE (SEQ ID NO: 2) of FIX or FIXa. In another example, the linker is sufficiently long to be capable of interacting with the amino acid sequence: RSTKFTIYNNMFCAGFHEGGRDSCQG (SEQ ID NO: 3) of FIX or FIXa.

[0203] In one example according to the above embodiments, the heterologous moiety is Fc. Accordingly, the present disclosure provides a conjugate comprising a FIX-Fc

construct (FIX-Fc) and a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative), and a linker, which covalently links the compound (e.g., peptide or peptide derivative) to the FIX-Fc. In one example according to this embodiment, the compound (e.g., peptide or peptide derivative) is covalently linked to the FIX portion of the FIX-Fc (e.g., via a linker). In another example according to this embodiment, the compound (e.g., peptide or peptide derivative) is covalently linked to the Fc portion of the FIX-Fc (e.g., via a linker).

[0204] In another example, the peptide or peptide derivative is interposed between the FIX and the Fc. In another example, the compound (e.g., peptide or peptide derivative) is inserted between the FIX heavy chain (HC) and the FIX light chain (LC), replacing all, part, or none of the FIX activation peptide, while maintaining a protease cleavage site at the N-terminus of the HC to enable the generation of an active protease.

[0205] In one example, the compound is covalently linked to the C-terminus of the FIX heavy chain (HC). In another example, the compound is covalently linked to the C-terminus of the FIX HC and is interposed between the FIX HC and a heterologous moiety (e.g., Fc). In yet another example according to any of the above embodiments, the compound (e.g., the pro-coagulant peptide or peptide derivative) is inserted between the FIX LC and the FIX HC, replacing all, part, or none of the FIX activation peptide, while maintaining a FXIa/FVIIa-TF cleavage site at the N-terminus of the HC to enable the generation of an active protease. Alternatively, the compound linked to the N-terminus of the FIX HC can be inserted with a linker that is cleavable by proteases activated during the clotting cascade in order to generate the free N-terminus of the HC required for protease activity (such as disclosed in International Patent Application No. PCT/US2011/043599 filed July 11, 2011). In a further example, the FIX LC and FIX HC could be separated with one or both chains covalently linked at the C-terminus to the N-terminus of a heterologous moiety (e.g. Fc), with the compound linked to the HC as described above to enable generation of an active protease after cleavage. In a further example according to any of the above embodiments, the compound (e.g., the pro-coagulant peptide or peptide derivative) is covalently linked to an internal amino acid residue of the FIX molecule (internal conjugation), e.g., via a cysteine residue. In one example, the cysteine residue is engineered into the FIX amino acid sequence, e.g., according to the procedures described in Mei, B. *et. al.* US2006/0115876 to Pan C. *et al.*

[0206] Exemplary FIX conjugates are illustrated in Figures 16 and 17.

[0207] Figure 16 illustrates various conjugates of the present disclosure, in which a compound (shown as a peptide or peptide derivative) is covalently linked to an FIX heterologous moiety construct, wherein the heterologous moiety is represented in this figure as an Fc and wherein the peptide can be linked to the FIX portion of the FIX-Fc (e.g., constructs A4 and A5), to the Fc portion of the FIX-Fc (constructs A1 and A2), or can be interposed between the FIX and the Fc of the FIX-Fc (A3). In constructs A1 and A2, FIX is linked to one chain of the Fc and the peptide or peptide derivative is linked to the other (free) Fc chain. In other constructs similar to E2, compound can be placed on either or both of the Fc chains. In construct A3, the peptide or peptide derivative is interposed between the FIX HC and the Fc. In construct A4, the peptide or peptide derivative is inserted between the FIX LC and the FIX HC, and replacing all, part, or none of the FIX activation peptide, while maintaining a FXIa/FVIIa-TF cleavage site at the N-terminus of the HC to enable the generation of an active protease. Alternatively, the compound linked to the N-terminus of the FIX HC can be inserted with a linker that is cleavable by proteases activated during the clotting cascade in order to generate the free N-terminus of the HC required for protease activity (such as disclosed in International Patent Application No. PCT/US2011/043599 filed July 11, 2011). In construct A5, the peptide or peptide derivative is covalently linked to an internal amino acid residue of the FIX molecule (e.g., cysteine). In one example, constructs A1-A4 can be made recombinantly. In another example, constructs A1 and A5 can be made semi-recombinantly e.g., as illustrated in Figures 23 and 24. It is understood that in other conjugates, the heterologous moiety (represented in this figure as Fc) can be e.g., PEG, PPG, albumin, XTEN, etc.

[0208] Figure 17 illustrates various conjugates of the present disclosure, in which a compound (shown as a peptide or peptide derivative) is covalently linked to a FIX protein. In construct B1, the peptide or peptide derivative is linked to the C-terminal amino acid of the FIX heavy chain (HC). In construct B2, the peptide or peptide derivative is inserted between the FIX LC and the FIX HC, and replacing all, part, or none of the FIX activation peptide, while maintaining a FXIa/FVIIa-TF cleavage site at the N-terminus of the HC to enable the generation of an active protease. Alternatively, the compound linked to the N-terminus of the FIX HC can be inserted with a linker that is

cleavable by proteases activated during the clotting cascade in order to generate the free N-terminus of the HC required for protease activity (such as disclosed in International Patent Application No. PCT/US2011/043599 filed July 11, 2011). In construct B3, the peptide or peptide derivative is covalently linked to an internal amino acid residue of the FIX. In one example, constructs B1 and B2 of Figure 17 can be made recombinantly. In another example, construct B3 can be made semi-recombinantly, e.g., as illustrated in Figures 23 and 24.

Platelet-Targeting Moiety (PTM) Conjugates

- [0209] In various embodiments, the compound of the present disclosure (e.g., peptide or peptide derivative) is covalently linked to a PTM (e.g., PDG-13) or a PTM-heterologous moiety construct. In various embodiments the compound of the present disclosure is covalently linked to a platelet targeting moiety
- [0210] Linking the compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative) to a platelet targeting moiety can be useful for targeting the compound (e.g., the pro-coagulant peptide or peptide derivative) to the surface of platelets (e.g., *in vivo*). In one example, the compound is targeted to platelets in order to enhance its efficacy by localizing the compound to the site of coagulation using a “targeting moiety” which binds to a molecule expressed on platelets. In addition to increasing the local concentration through the PTM, the concomitant interaction of the peptide or peptide derivative with FIXa may stabilize FIX association with the Xase complex on platelet surfaces, similar to the mechanism of action of FVIIIa. Preferably the targeted molecules are not expressed on cells or tissues other than platelets, *i.e.*, the targeting moieties specifically bind to platelets. Linking the peptide or peptide derivative to the targeting moiety may enhance its biological activity. This strategy may reduce the compound dose necessary to obtain a desired pharmaceutical effect, and thus can reduce potential side effects that the compound may have. Accordingly, in one example, the conjugates of the present disclosure bind (e.g., specifically) to platelets.
- [0211] In one example, the targeting moiety binds to receptors/conformations found on resting platelets. By doing so, sites for coagulation could be primed for enhanced efficacy. Targeting such molecule may also extend half life of the compound (e.g., peptide or peptide derivative) and/or prevent clearance. Exemplary platelet targeting

moieties according to this embodiment include GpIb (e.g., GpIb α) of the GpIb/V/IX complex, GpVI, and nonactive forms of GPIIb/IIIa.

[0212] In another example, the platelet targeting moiety binds to receptors/conformations only found on activated platelets in order to localize the compound (e.g., peptide or peptide derivative) to a site of active coagulation. Exemplary such molecules include the active form of GPIIb/IIIa as well as CD62P.

[0213] In another embodiment, the platelet targeting moiety selectively binds to a target selected from the group consisting of: P selectin, GMP-33, LAMP-1, LAMP-2, CD40L, and LOX-1.

[0214] The platelet targeting moiety can be, e.g. MB9, SCE5, scFv, AP3, or peptides (e.g., "RGD" peptides) targeting GPIIbIIIa, a fatty acid or small molecule capable of inserting into plasma membranes, OS1, OS2, and PS4 targeting GP1b. In one embodiment, the platelet targeting moiety is OS1, OS2 and PS4 targeting GP1b.

[0215] In another embodiment, the platelet targeting moiety is a moiety that binds to GPIIbIIIa (e.g., PDG-13).

[0216] In one example, the conjugate of the present disclosure includes a platelet targeting moiety-heterologous moiety construct (e.g., PTM-Fc, PTM-albumin, PTM-PEG) and the compound (e.g., peptide or peptide derivative) is covalently linked to the platelet targeting moiety-portion of the construct. In another example, the conjugate of the present disclosure includes a platelet targeting moiety-heterologous moiety construct (e.g., PTM-Fc, PTM-albumin, PTM-PEG) and the compound (e.g., peptide or peptide derivative) is covalently linked to the heterologous moiety portion of the construct.

[0217] In one example according to the above embodiments, the heterologous moiety is Fc. Accordingly, the present disclosure provides a conjugate comprising a PTM-Fc construct (PTM-Fc) and a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative), and a linker, which covalently links the compound (e.g., peptide or peptide derivative) to the PTM-Fc. In one example according to this embodiment, the compound (e.g., peptide or peptide derivative) is covalently linked to the PTM portion of the PTM-Fc (e.g., via a linker). In another example according to this embodiment, the compound (e.g., peptide or peptide derivative) is covalently linked to the Fc portion of the PTM-Fc (e.g., via a linker).

- [0218] In one example according to any of the above embodiments, the compound (e.g., the pro-coagulant peptide or peptide derivative) is covalently linked to the N-terminus of the platelet targeting moiety. In another example according to any of the above embodiments, the compound (e.g., the pro-coagulant peptide or peptide derivative) is covalently linked to the C-terminus of the platelet targeting moiety. In a further example according to any of the above embodiments, the compound (e.g., the pro-coagulant peptide or peptide derivative) is covalently linked to an internal amino acid residue of the platelet targeting moiety molecule (internal conjugation), e.g., via a cysteine residue. In one example, the cysteine residue is engineered into the platelet targeting moiety amino acid sequence, e.g., according to the procedures described in Mei, B. *et al.* and US2006/0115876 to Pan C. *et al.*
- [0219] The conjugates of the present disclosure can comprise one or more targeting moiety. Additionally, two or more targeting moieties may be linked to each other (e.g., via a polypeptide linker) in series. When two or more targeting moieties are present in a conjugate of the present disclosure, the moieties may be the same or different.
- [0220] Exemplary platelet targeting moiety conjugates are illustrated in Figure 22.
- [0221] Figure 22 illustrates various conjugates of the present disclosure, in which a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative) is covalently linked to a platelet targeting moiety (construct H3) or a platelet targeting moiety-heterologous moiety construct (H1, H2, Ha, Hb) optionally via a linker. The peptide or peptide derivative can be linked to the platelet targeting moiety portion of the construct (Ha), to the heterologous moiety (e.g., Fc) portion of the construct (H1, H2), or can be interposed between the platelet targeting moiety and the heterologous moiety (e.g., Fc) of the platelet targeting moiety-fusion construct (Hb). In Figure 22, the heterologous moiety is represented as an Fc, which can be replaced with other heterologous moieties described herein. In constructs H1 and H2, in which the heterologous moiety is shown as an Fc, the platelet targeting moiety is linked to one chain of the Fc and the peptide or peptide derivative is linked to the other (free) Fc chain. In constructs H3, the peptide or peptide derivative is linked to the N- or C-terminal amino acid of the platelet targeting moiety. In other constructs, the peptide or peptide derivative is covalently linked to an internal amino acid residue of the platelet targeting moiety (e.g., cysteine). It is

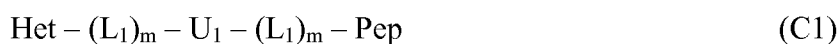
understood that in other conjugates, the heterologous moiety (represented in Figure 22 as Fc) can be any other heterologous moiety, e.g., PEG, PPG, albumin, XTEN, etc.

[0222] In one example, the conjugates of Figure 22 can be made recombinantly. In another example, the conjugates of Figure 22 (e.g., H1, H3) can be made semi-recombinantly e.g., as illustrated in Figures 25 and 26.

[0223] Heterologous moieties useful in any of the above embodiments are described herein. Exemplary heterologous moieties according to any of the above embodiments include, e.g., any half-life extending molecule known in the art, e.g., polyethylene glycol (PEG), polypropylene glycol (PPG), PAS, HES, XTEN, albumin, as well as antibodies and antibody fragments (e.g., Fc).

Exemplary Conjugates

[0224] In one example, the polypeptide conjugate of the present disclosure has a structure comprising a polypeptide (e.g., FVIII, FIX, FVIIa, or a platelet targeting moiety), a compound of the present disclosure (e.g., a pro-coagulant peptide or peptide derivative), optionally at least one linker, and optionally at least one heterologous moiety. Examples of such structures include, e.g., those according to one of Formula (B1) to Formula (G2) below:



[0225] In Formula (B1), (B2), (C1), (C2), (D1), (D2), (E1), and (E2), U_1 is a polypeptide selected from a blood coagulation factor and a platelet targeting moiety, wherein the blood coagulation factor is selected from FVIIa, FVIII, and FIX. In one example, FVIII is B-domain deleted FVIII. In Formulas (F1) and (F2), FIX(HC) is the heavy chain of FIX, and FIX(LC) is the light chain of FIX. In Formulas (FG) and (G2), FVIIa(HC) is the heavy chain of FVIIa, and FVIIa(LC) is the light chain of FVIIa.

- [0226] In Formulas (B1) through (E2), m is an integer selected from 0 and 1.
- [0227] Het is a heterologous moiety as defined herein. In one example, Het is a half-life extending moiety, selected from, e.g., PEG, PPG, HES, PAS, XTEN, albumin, and Fc.
- [0228] In Formulas (B1) through (E2), Pep is a compound of the present disclosure. In one example, the compound is a pro-coagulant peptide or peptide derivative of the present disclosure.
- [0229] In Formulas (B1) through (G2), each L_1 is either absent ($m=0$) or present ($m=1$), and when present is a linker as described herein. The linker covalently links the compound (e.g., the peptide or peptide derivative), directly or indirectly (e.g., via a heterologous moiety Het), to the polypeptide (e.g., FVIII, FIX, FVIIa, or a platelet targeting moiety). Exemplary linkers are described herein.
- [0230] In one embodiment, the present invention is directed to a conjugate comprising the compound disclosed herein and a heterologous moiety which are linked to each other via an optional linker. The conjugate can be represented by a structure comprising formula $\text{Het1}-(L_1)_m\text{---Pep}$ or $\text{Pep}-(L_1)_m\text{---Het1}$, wherein Het1 is a heterologous moiety; m is an integer selected from 0 and 1; L_1 is either absent ($m=0$) or present ($m=1$), and when present is a linker; Pep is a compound according to claim 1; and (—) is a covalent bond. In another embodiment, the heterologous moiety comprises a half-life extending molecule, e.g., an immunoglobulin constant region or a portion thereof, albumin, an XTEN sequence, transferrin, an albumin binding moiety, a PAS sequence, a HES sequence, the β subunit of the C-terminal peptide (CTP) of human chorionic gonadotropin, polyethylene glycol (PEG), hydroxyethyl starch (HES), albumin-binding small molecules, or any combinations thereof. In a specific embodiment, the immunoglobulin constant region or a portion thereof is an Fc moiety or an FcRn binding partner.
- [0231] In certain embodiments, the conjugate of the invention further comprises a second heterologous moiety (Het2), wherein the second heterologous moiety is linked to or associated with the heterologous moiety (Het or Het1). The conjugate can be represented as $\text{Het2:Het1}-(L_1)\text{---Pep}$ or $\text{Pep}-(L_1)\text{---Het1:Het2}$, wherein L_1 is an optional linker, Pep is the compound of the invention, Het1 is a first heterologous moiety, and Het2 is a second heterologous moiety. In still other embodiment, the conjugate comprises two polypeptide chains, a first chain comprising the compound of the invention linked to a first

heterologous moiety by a first linker and a second chain comprising a second heterologous moiety. In yet other embodiments, the second heterologous moiety is a half-life extending molecule. The second heterologous moiety can comprise an immunoglobulin constant region or a portion thereof, albumin, transferrin, an albumin binding moiety, a PAS sequence, a HES sequence, the β subunit of the C-terminal peptide (CTP) of human chorionic gonadotropin, polyethylene glycol (PEG), hydroxyethyl starch (HES), albumin-binding small molecules, or any combinations thereof. In a particular embodiment, the immunoglobulin constant region or a portion thereof is an Fc moiety or an FcRn binding partner. In yet other embodiments, the heterologous moiety and the second heterologous moiety are associated by a covalent bond, e.g., a peptide bond, a disulfide bond, a metal bond, a hydrogen bond, a disulfide bond, a sigma bond, a pi bond, a delta bond, a glycosidic bond, an agnostic bond, a bent bond, a dipolar bond, a Pi backbond, a double bond, a triple bond, a quadruple bond, a quintuple bond, a sextuple bond, conjugation, hyperconjugation, aromaticity, hapticity, or antibonding. In another embodiment, the heterologous moiety and the second heterologous moiety are associated by a non-covalent interaction, e.g., an ionic interaction, a hydrophobic interaction, a hydrophilic interaction, a Van der Waals interaction, or a hydrogen bond. In some embodiments, the association between the heterologous moiety and the second heterologous moiety is a covalent bond or a non-covalent bond. In a particular embodiment, the association is a disulfide bond.

[0232] In still other embodiments, the conjugate further comprises a scFc linker (X), which is linked to the second heterologous moiety and the compound. Therefore, the conjugate can be represented as Het2-X-Pep-L1-Het1 or Het1-L1-Pep-X-Het2, wherein the scFv linker comprises at least one intracellular processing site and a linker (Lx). In one embodiment, a first intracellular processing site (P1) interposed between the second heterologous moiety and the linker (Lx). In another embodiment, a second intracellular processing site (P2) interposed between the linker (Lx) and the compound. The intracellular processing site inserted therein can be processed (cleaved) by an intracellular processing enzyme upon expression in a host cell, thereby allowing formation of a zymogen-like heterodimer. Examples of the intracellular processing enzymes include furin, a yeast Kex2, PCSK1 (also known as PC1/Pc3), PCSK2 (also known as PC2), PCSK3 (also known as furin or PACE), PCSK4 (also known as PC4),

PCSK5 (also known as PC5 or PC6), PCSK6 (also known as PACE4), or PCSK7 (also known as PC7/LPC, PC8, or SPC7). Other processing sites are known in the art.

[0233] The conjugate comprising two polypeptide chains as shown above can further comprise FVIII, FIX, FVIIa, or a platelet targeting moiety, wherein the polypeptide is linked to the compound or to the second heterologous moiety via a second optional linker. In one embodiment, the polypeptide (either a heavy chain or a light chain) is linked to the second heterologous moiety via the second optional linker. Therefore, the conjugate can comprises two polypeptide chains, a first chain comprising the compound, the heterologous moiety, and the linker, and a second chain comprising the polypeptide, the second heterologous moiety, and the second optional linker, wherein the first polypeptide chain and the second polypeptide chain are associated with each other.

[0234] In other embodiments, the conjugate comprises two polypeptide chains selected from the group consisting of:

[0235] (a) the first polypeptide chain comprising the compound linked to the N-terminus of the first heterologous moiety by the first linker and the second polypeptide chain comprising the polypeptide linked to the N terminus of the second heterologous moiety by the second linker,

(i) wherein the polypeptide is FIX or a platelet targeting moiety, and

(ii) wherein the first polypeptide chain and the second polypeptide chain are chemically or physically associated with each other; (Figure 16 A1 and Figure 22 H1) and

(b) the first polypeptide chain comprising the compound linked to the C-terminus of the first heterologous moiety by the first linker and the second polypeptide chain comprising the polypeptide linked to the N terminus of the second heterologous moiety by the second linker,

(i) wherein the polypeptide is FIX or a platelet targeting moiety, and

(ii) wherein the first polypeptide chain and the second polypeptide chain are chemically or physically associated with each other; (Figure 16 A2 and Figure 22 H2).

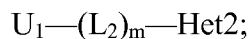
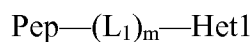
[0236] In yet other embodiments, the conjugate comprises three polypeptide chains selected from the group consisting of:

(a) the first polypeptide chain comprising the compound linked to the N-terminus of the first heterologous moiety by the first linker, the second polypeptide chain comprising the heavy chain of the polypeptide linked to the N-terminus of the second heterologous

moiety by the second linker, and the third polypeptide chain comprising the light chain of the polypeptide,

- (i) wherein the polypeptide is FVIII or FVIIa, and
- (ii) wherein the second polypeptide chain is chemically or physically associated with the first polypeptide chain and the third polypeptide chain; and (Figure 18 C1 and Figure 20 E1), and
- (b) the first polypeptide chain comprising the compound linked to the C-terminus of the first heterologous moiety by the first linker, the second polypeptide chain comprising the heavy chain of the polypeptide linked to the N-terminus of the second heterologous moiety by the second linker, and the third polypeptide chain comprising the light chain of the polypeptide
 - (i) wherein the polypeptide is FVIII or FVIIa, and
 - (ii) wherein the second polypeptide chain is chemically or physically associated with the first polypeptide chain and the third polypeptide chain. (Figure 18 C2 and Figure 20 E2).

[0237] In still yet other embodiments, the conjugate can be represented by formula:



wherein Het1 is the first heterologous moiety;

L1 is either absent ($m=0$) or present ($m=1$), and when present is a linker;

m is an integer selected from 0 and 1;

Pep is the compound of any one of claims 1 to 82,

Het2 is the second heterologous moiety;

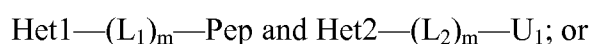
L2 is either absent ($m=0$) or present ($m=1$), and when present is a linker;

U1 is the polypeptide comprising FIX, FVIII, FVIIa, or the platelet targeting moiety;

(—) is a peptide bond; and

wherein Het1 and Het2 associate chemically or physically with each other. The chemical or physical association can be a covalent bond, e.g., a disulfide bond, or a non-covalent bond. Left represents N-terminus, and right represents C-terminus.

[0238] In another example the conjugate comprises:



Het1—(L₁)_m—Pep and U₁—(L₂)_m—Het2; or
 Pep—(L₁)_m—Het1 and Het2—(L₂)_m—U₁; or
 Pep—(L₁)_m—Het1 and U₁—(L₂)_m—Het2,
 U₁—(L₂)_m—Het2 and Het1—(L₁)_m—Pep, or
 U₁—(L₂)_m—Het2 and Pep—(L₁)_m—Het1, or
 wherein

Het1 is the first heterologous moiety;

L₁ is either absent (m=0) or present (m=1), and when present is a linker;

m is an integer selected from 0 and 1;

Pep is the compound of any one of claims 1 to 82,

Het2 is the second heterologous moiety;

L₂ is either absent (m=0) or present (m=1), and when present is a linker;

U₁ is the polypeptide comprising FIX, FVIII, FVIIa, or the platelet targeting moiety;

(—) is a covalent bond,

wherein Het1 and Het2 associate chemically or physically with each other.

[0239] In some embodiments, the conjugate comprising two polypeptide chains or three polypeptide chains as shown above can further comprise a scFc linker. In one embodiment, the scFc linker is linked to the second heterologous moiety and the compound. In another embodiment, the scFc linker comprises a linker (L_x) and a first intracellular processing site (P1) interposed between the second heterologous moiety and the linker (L_x). The scFc linker can further comprise a second intracellular processing site (P2) interposed between the linker (L_x) and the compound. In a further embodiment, the scFc linker comprises two intracellular processing sites which are recognized by the same or by different intracellular processing enzymes. The intracellular processing site can be recognized by a intracellular processing enzyme selected from the group consisting of a yeast Kex2, PCSK1, PCSK2, PCSK3, PCSK4, PCSK5, PCSK6, and PCSK7. In a specific embodiment, the at least one intracellular processing site is processed by PCSK5. In other embodiments, each of the two intracellular processing sites is processed by PCSK5. In one embodiment, the two intracellular processing sites are the same. In other embodiments, the two intracellular processing sites are different. Examples of the intracellular processing site processed by the intracellular processing enzyme comprises

the amino sequence R-X-[R/K]-R, wherein X can be any amino acid, and [R/K] indicated that the amino acid can be R or K. Each of the PCSK5 enzymatic cleavage sites independently comprises the sequence RRRR (SEQ ID NO: 900) or (RKR)_n (SEQ ID NO: 901), where n is 2. The PCSK5 enzymatic cleavage site at the C-terminal end of the scFc linker comprises the sequence RRRR (SEQ ID NO: 900) and the PCSK5 enzymatic cleavage site at the N-terminal end of the cscFc linker comprises the sequence (RKR)₂ (SEQ ID NO: 901). In one embodiment, the scFc linker has a length of about 10 to about 50 amino acids and about 20 to about 30 amino acids. In another embodiment, the scFc linker comprises a gly/ser peptide. In some embodiment, the gly/ser peptide comprises an amino acid sequence of formula (Gly₄Ser)_n or Ser(Gly₄Ser)_n, wherein n is a positive integer selected from the group consisting of 1, 2, 3, 4, 5, 6, 7, 8, 9 and 10. Examples of the linker include the (Gly₄ Ser)_n peptide, e.g., an amino acid sequence selected from the group consisting of (Gly₄Ser)₆, Ser(Gly₄Ser)₆, (Gly₄Ser)₄ and Ser(Gly₄Ser)₄.

[0240] In certain embodiments, FVIII and FVIIa can be heterodimers comprising a heavy chain and a light chain, wherein the heavy chain and the light chain are associated with each other by a metal bond.

[0241] In one aspect, a conjugate comprises (a) a polypeptide selected from a heavy chain of FVIII (FVIII HC), a light chain of FVIII (FVIII LC), a heavy chain of FIX (FIX HC), a light chain of FIX (FIX LC), a heavy chain of activatable or activated FVII (FVIIa HC), a light chain of activatable or activated FVII (FVIIa LC), and a platelet targeting moiety, and (b) a compound of any one of claims 1 to 82, wherein the compound is linked to the polypeptide, optionally via a linker. The N-terminus, C-terminus, or an internal amino acid residue of the compound of the invention is linked to the N-terminus, C-terminus, or an internal amino acid residue of the polypeptide via the linker. See Figure 17 B3, Figure 19 D4 and D5, and Figure 21 F5. One embodiment of the invention includes a conjugate comprising the compound of the invention linked to (1) the C-terminus of FIX HC via a linker, (2) the C-terminus of FIX LC via a linker, (3) the C-terminus of FIX LC via a first linker and the N-terminus of FIX HC via a second linker, or (4) an internal amino acid residue of FIX HC or FIX LC via a linker. See Figure 17 B1, B2, and B3. Another embodiment of the invention includes the compound of the invention linked to the N-terminus or the C-terminus of FVIIa HC (Figure 19 D1 and D3), the C-terminus of FVIIa LC (Figure 19 D2), or an internal amino acid residue of FVIIa HC or FVIIa LC (Figure

19 D4 and D5). In some embodiments, FVIIa HC or FVIIa LC forms a heterodimer with FVIIa LC or FVIIa HC, respectively. In other embodiments, the compound is linked to the N-terminal of activatable FVII HC via a linker, wherein said linker comprises a protease-cleavable substrate.

[0242] In one embodiment, the conjugate comprises the compound of the invention linked to the N terminus or the C terminus of FVIII HC (Figure 21 F1 and F3), the N-terminus or the C-terminus of FVIII LC (Figure 21 F2 and F4), or an internal amino acid residue of FVIII HC or FVIII LC. (Figure 21 F5). In this construct, FVIII HC or FVIII LC can form a heterodimer with FVIII LC or FVIII HC, respectively.

[0243] In another embodiment, the compound of the invention can be linked to the N-terminus or the C-terminus of the platelet targeting moiety. (See Figure 22, Ha, Hb, H3). In one embodiment, the compound or the Factor IX HC is further linked to a heterologous moiety (Het or Het1) by an optional linker. (See Figure 16, A3, A4, and A5). In another embodiment, the compound or FVIIa HC is further linked to a heterologous moiety (Het or Het1) by an optional linker. (See Figure 18, C3, C4, C5, and C6). In some embodiments, the compound or FVIII LC is further linked to a heterologous moiety (Het or Het1) by an optional linker. (Figure 20, E3, E4, E5, E6, and E7). In other embodiments, the compound or the targeting moiety is further linked to a heterologous moiety (Het or Het1) by an optional linker. (See Figure 22, Ha, Hb).

[0244] The heterologous moiety (Het or Het1) as used herein can comprise an immunoglobulin constant region or a portion thereof, albumin, transferrin, an albumin binding moiety, a PAS sequence, a HES sequence, an XTEN sequence, the β subunit of the C-terminal peptide (CTP) of human chorionic gonadotropin, polyethylene glycol (PEG), hydroxyethyl starch (HES), albumin-binding small molecules, or any combinations thereof. In one embodiment, the heterologous moiety is an Fc moiety or an FcRn binding partner. In another embodiment, the conjugate further comprises a second heterologous moiety (Het2). Examples of the second heterologous moiety (Het2) comprises an immunoglobulin constant region or a portion thereof, albumin, transferrin, an albumin binding moiety, an XTEN sequence, a PAS sequence, a HES sequence, the β subunit of the C-terminal peptide (CTP) of human chorionic gonadotropin, polyethylene glycol (PEG), hydroxyethyl starch (HES), albumin-binding small molecules, or any combinations thereof. In a specific embodiment, the immunoglobulin constant region or

a portion thereof of the second heterologous moiety is an Fc moiety or an FcRn binding partner.

[0245] In further embodiments, the conjugate is polysialylated, pegylated, glycosylated, hesylated, gamma-carboxylated, or any combinations thereof.

Linker (L₁)

[0246] The linker is used to covalently link the compound of the present disclosure (e.g., the pro-coagulant peptide) to the heterologous moiety or a polypeptide (e.g., a platelet targeting moiety, or a blood coagulation factor selected from FVIIa, FVIII, and FIX), or or polypeptide construct (e.g., FVIIa-Fc). In other embodiments, a linker is interposed between the polypeptide and a heterologous moiety. Each conjugate can contain multiple linkers.

[0247] When the compound (e.g., the peptide or peptide derivative) is interposed between the polypeptide (e.g., FVIII, FIX, FVIIa, or the platelet targeting moiety) and a heterologous moiety (e.g., Fc), then the compound (e.g., the peptide or peptide derivative) is divalent and the conjugate of the present disclosure can include more than one (e.g., 2 linkers), which are independently selected.

[0248] In some embodiments, the linker is a hydrophilic linker, e.g., a peptide linker, such as those comprised of Gly and Ser (e.g., (GGSGG)_n where n = 1-50) or Gly and Val. In other embodiments, the linker is a water-soluble polymer, such as polyethylene glycol (PEG) or polypropylene glycol (PPG). In one example, the linker is a PEG moiety. In other embodiments, the linker is a hydroxyethyl starch (HES) moiety.

[0249] The compound can be coupled with the heterologous moiety or the polypeptide chemically or using recombinant techniques, e.g., by recombinant expression of a fusion protein.

[0250] Chemical linkage can be achieved by linking together chemical moieties present on the compound and on the heterologous moiety or the polypeptide, e.g., using moieties such as amino, carboxyl, sulfhydryl, hydroxyl groups, and carbohydrate groups. A variety of homo- and hetero-bifunctional linker molecules can be used that have groups that are activated, or can be activated to link these moieties. Some useful reactive groups on linker molecules include maleimides, N-hydroxy-succinimide esters and hydrazides. Many different spacers of different chemical composition and length can be used for separating these reactive groups including, for example, polyethylene glycol (PEG),

aliphatic groups, alkylene groups, cycloalkylene groups, fused or linked aryl groups, peptides and/or peptidyl mimetics, e.g., peptides having from one to 20 amino acids or amino acid analogs.

[0251] In one embodiment, the linker is a non-cleavable linker. In one example according to this embodiment, the linker is a “gly-ser polypeptide linker”, e.g., includes the amino acid sequence (GGS)_n, (GGGS)_n, or (GGGGS)_n where n is 1-50 (e.g., n is 6).

[0252] In another example, the linker is cleavable (e.g., cleavable *in vivo*). In one example, the compound is linked to the heterologous moiety or the polypeptide in such a way that the compound is released from the heterologous moiety or the polypeptide *in vivo* (e.g., near the site of biological activity of the compound in the body). In one example, the cleavage of such linker liberates the compound from a potential activity-compromising steric hindrance that may be caused by the heterologous moiety (e.g., PEG moiety).

[0253] In other embodiments, the compound is linked to the polypeptide in such a way that *in vivo* a functional form of the polypeptide (e.g., the coagulation factor) is released (e.g., near the site of biological activity of the coagulation factor in the body). The cleavage of such linkers liberates the polypeptide from a potential activity-compromising steric hindrance that may be caused by the linked compound (e.g., peptide or peptide derivative), and thereby allows the generation of polypeptide conjugates which retain a high molar specific activity of the polypeptide.

[0254] In some embodiments, the linker is a peptide linker that is proteolytically cleavable. In other embodiments, the linker is cleavable by an enzyme from the coagulation cascade.

[0255] In some embodiments, the release of the polypeptide (e.g., a coagulation factor) from the conjugate form can be achieved by linking the compound (e.g., the peptide or peptide derivative) to a site on the polypeptide that is removed *in vivo*. For example, the release of a coagulation factor from the conjugate form can be achieved by linking the compound (e.g., the peptide or peptide derivative) to a site on the coagulation factor that is removed during the activation process.

[0256] “Proteolytic cleavage in a coagulation-related mode,” as used herein, means any proteolytic cleavage that occurs as a consequence of the activation of at least one coagulation factor or coagulation cofactor. The phrase “activated coagulation factor after

the linker is proteolytically cleaved in a coagulation-related mode,” as used herein means that the coagulation factor is either activated almost in parallel to the proteolytic cleavage of the linker, or that the coagulation factor was already activated before the proteolytic cleavage of the linker. Activation may occur, for example, by proteolytic cleavage of the coagulation factor or by binding to a cofactor.

[0257] The release of the compound from the conjugate form can be achieved using a linker that degrades in a controlled manner, e.g., by one or more proteolytic enzymes (e.g., in the blood). For example, sugar polymers or peptides can be used that are susceptible to general blood proteases or hydrolases. A variety of such technologies are known in the art and have been used to generate pro-drugs.

[0258] The linker could be further engineered to be cleaved specifically at sites where pro-coagulant compounds are most needed, such as sites of inflammation or blood coagulation sites triggered through trauma. For example, the linker may be susceptible to specific proteases produced only at the desired site of action, such as proteases released by the inflammation process or generated by the blood coagulation cascade. This selective release of the compound and/or the polypeptide (e.g., the coagulation factor) may lower the potential for side effects and increase the efficiency of the compound and/or the polypeptide at its site of action.

[0259] In one embodiment, the linker is cleavable by an enzyme from the coagulation cascade. In one example the linker includes a thrombin cleavable site (thrombin cleavable substrate moiety) or a FXIa cleavable site (FXIa cleavable substrate moiety). Exemplary FXIa cleavage sites include: TQSFNDFTR and SVSQTSKLTR. Exemplary thrombin cleavage sites include: DFLAEGGGVR, DFLAEEGGGVR, TTKIKPR, ALVPR, ALRPR and ALRPRVVGGA. In one embodiment, the thrombin cleavable site includes (D-Phe)-Pro-Arg. In one embodiment, the thrombin cleavable site includes (D-Phe)-Pip-Arg, wherein Pip is pipecolic acid.

[0260] The cleavable peptide linker may comprise a sequence derived from

- a) the coagulation factor itself if it contains proteolytic cleavage sites that are proteolytically cleaved during activation of the coagulation factor,
- b) a substrate polypeptide of this coagulation factor, or
- c) a substrate polypeptide cleaved by a protease which is activated or formed by the direct or indirect involvement of the coagulation factor.

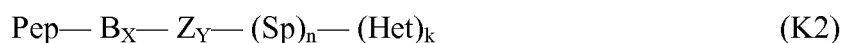
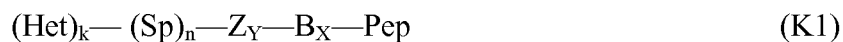
- [0261] Some embodiments of the invention are coagulation factors wherein the peptide linker is cleavable by the protease that normally activates the coagulation factor in vivo, thereby ensuring that the cleavage of the linker is linked to the activation of the coagulation factor at a site at which coagulation occurs.
- [0262] Other exemplary conjugates according to the invention are those wherein the linker is cleavable by the coagulation factor which is part of the conjugate once it is activated, thus also ensuring that cleavage of the conjugate is connected with a coagulatory event.
- [0263] Other exemplary therapeutic conjugates according to the invention are those wherein the linker is cleavable by a protease, which itself is activated directly or indirectly by the activity of the coagulation factor which is part of the conjugate, thus also ensuring that cleavage of the fusion protein is connected with a coagulatory event.
- [0264] In another example, the linker includes a thrombin-cleavable chimeric protein, such as those disclosed in U.S. Patent 7,589,178 to Le Bonniec, which is herein incorporated by reference in its entirety.
- [0265] In another embodiment, the compound of the present disclosure is covalently linked to the linker without further linkage of the linker to a heterologous moiety or protein. Such conjugate between a compound and a linker can be useful as a "pro-drug". For example, the linker is cleavable by a protease, which is activated directly or indirectly by the activity of the compound, thus ensuring that cleavage of the linker is connected with a coagulatory event.

Linkers containing a self-immolative moiety

- [0266] In one embodiment the linker includes a self-immolative moiety, e.g., interposed between the compound and a cleavable substrate moiety. For example, such self-immolating moiety has the advantage that the cleavability of the substrate moiety is not negatively impacted by the terminal amino acid residue of the compound (e.g., the pro-coagulant peptide).
- [0267] In one embodiment, the self-immolative moiety is derived from para-aminobenzylalcohol (PAB), e.g., connected to the compound via a carbamate, a carbonate or an ether bond. In one example, the self-immolative moiety is p-aminobenzyl carbamate (PABC). In another example, the self-immolative moiety is a heteroaromatic analog of PABC. Exemplary self-immolative moieties are disclosed, e.g., in U.S. Patents

7,375,078 and 7,754,681, each of which are incorporated herein by reference in its entirety.

[0268] In various embodiments, the conjugate includes a linker containing a self-immolative moiety (B_X). In one example according to this embodiment, the conjugate formed between the compound and the heterologous moiety, or between the compound and the polypeptide has a structure according to Formula (K1), (K2), (L1), or (L2):



wherein

k is an integer selected from 0 and 1;

Het is either absent ($k=0$) or present ($k=1$), and when present is a heterologous moiety (e.g., a half-life extending molecule);

U_1 is either absent ($k=0$) or present ($k=1$), and when present is a polypeptide selected from blood coagulation factor (e.g., FVIIa, FVIII, and FIXa) and a platelet targeting moiety, wherein U_1 is optionally further linked to a heterologous moiety;

n is an integer selected from 0 and 1;

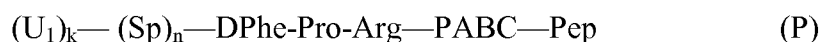
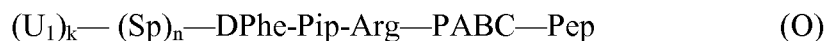
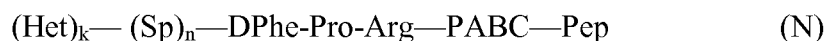
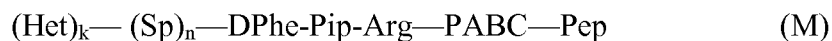
Sp is a spacer moiety, which is either absent ($n=0$) or present ($n=1$);

Z_Y is a cleavable substrate moiety (e.g., a protease-cleavable substrate moiety, e.g., a thrombin-cleavable substrate moiety, or a FXIa-cleavable substrate moiety);

B_X is a self-immolative moiety (e.g., a PABC moiety); and

Pep is a compound (e.g., pro-coagulant peptide or peptide derivative) of the present disclosure.

[0269] In one example, the conjugate has a structure according to Formula (M), (N), (O), or (P):



wherein k, Het, U₁, Sp, and Pep are defined as for Formulae K1-L2.

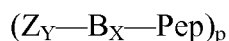
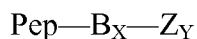
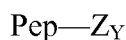
Compounds Linked to a Cleavable Substrate Moiety

[0270] In certain embodiments, the compound of the present disclosure is linked to a cleavable substrate moiety. The cleavable substrate moiety can be cleaved by a proteolytic enzyme as described herein (e.g., by an enzyme of the coagulation cascade).

[0271] In one example, the cleavable substrate moiety is linked to the compound via a self-immolative moiety (B_X). For example, the pro-coagulant compound of the present disclosure is covalently linked to a heterologous moiety, optionally via a linker (L₁) thereby forming a conjugate. The Linker (L₁) is different than the linking moiety Z previously defined.

[0272] In another example, two or more compounds are linked to each other via a cleavable linker (e.g., a cleavable linker having a self-immolative moiety).

[0273] In one example according to the various embodiments above, the conjugate can have a structure selected from the following formulae:



wherein Pep, Z_Y, and B_X are defined as above, and the integer p is selected from 1-50.

Heterologous Moiety

[0274] In some embodiments, the conjugate of the invention includes one heterologous moiety. For example, the compound of the present disclosure is linked to one heterologous moiety. In other embodiments, the conjugate includes two heterologous moieties, which may be the same or different. For example, a compound of the present disclosure is linked to two heterologous moieties (i.e., a first heterologous moiety, "Het 1", and a second heterologous moiety "Het2"). In yet other embodiments, the conjugate of the present disclosure includes more than two heterologous moieties, e.g., three, four, five, or more than five heterologous moieties. In some embodiments, all the heterologous moieties are identical. In some embodiments, at least one heterologous moiety is different from the other heterologous moieties. In some embodiments, the two, three or

more than three heterologous moieties are linked in tandem. In other embodiments, the conjugate of the invention can comprise two, three, or more than three heterologous moieties wherein at least an additional moiety (*e.g.*, a spacer moiety, a protease-cleavable substrate, a self-immolative moiety, or combinations thereof) is interposed between two heterologous moieties.

[0275] A heterologous moiety can comprise a heterologous polypeptide moiety, or a heterologous non-polypeptide moiety, or both. In some aspects, the heterologous moiety comprises a combination of a heterologous polypeptide and a heterologous non-polypeptide moiety.

[0276] In certain embodiments, the first heterologous moiety (*e.g.*, a first Fc region) and the second heterologous moiety (*e.g.*, a second Fc region) are associated with each other to form a dimer. In one embodiment, the second heterologous moiety is a second Fc region, wherein the second Fc region is linked to or associated with the first heterologous moiety, *e.g.*, the first Fc region. For example, the second heterologous moiety (*e.g.*, the second Fc region) can be linked to the first heterologous moiety (*e.g.*, the first Fc region) by a linker or associated with the first heterologous moiety by a covalent or non-covalent bond

[0277] In some embodiments, the heterologous moiety is a peptide or polypeptide with either unstructured or structured characteristics that are associated with the prolongation of *in vivo* half-life when incorporated in a procoagulant compound of the invention. Non-limiting examples include albumin, albumin fragments, Fc fragments of immunoglobulins, the β subunit of the C-terminal peptide (CTP) of human chorionic gonadotropin, a HAP sequence, XTEN, a transferrin or a fragment thereof, a PAS polypeptide, polyglycine linkers, polyserine linkers, albumin-binding moieties, or any fragments, derivatives, variants, or combinations of these polypeptides. In other related aspects a heterologous moiety can include an attachment site (*e.g.*, a cysteine amino acid) for a non-polypeptide moiety such as polyethylene glycol (PEG), hydroxyethyl starch (HES), polysialic acid, or any derivatives, variants, or combinations of these elements. In some aspects, a heterologous moiety consisting of a cysteine amino acid that function as an attachment site for a non-polypeptide moiety such as polyethylene glycol (PEG), hydroxyethyl starch (HES), polysialic acid, or any derivatives, variants, or combinations of these elements.

- [0278] In some embodiments, the heterologous moiety is a polypeptide comprising, consisting essentially of, or consisting of at least about 10, 100, 200, 300, 400, 500, 600, 700, 800, 900, 1000, 1100, 1200, 1300, 1400, 1500, 1600, 1700, 1800, 1900, 2000, 2500, 3000, or 4000 amino acids. In other embodiments, the heterologous moiety is a polypeptide comprising, consisting essentially of, or consisting of about 100 to about 200 amino acids, about 200 to about 300 amino acids, about 300 to about 400 amino acids, about 400 to about 500 amino acids, about 500 to about 600 amino acids, about 600 to about 700 amino acids, about 700 to about 800 amino acids, about 800 to about 900 amino acids, or about 900 to about 1000 amino acids.
- [0279] In certain embodiments, a heterologous moiety improves one or more pharmacokinetic properties of the compound or conjugate without significantly affecting its biological activity.
- [0280] In certain embodiments, a heterologous moiety increases the *in vivo* and/or *in vitro* half-life of the procoagulant compound or conjugate of the invention. In other embodiments, a heterologous moiety facilitates visualization or localization of the compound or conjugate of the invention. Visualization and/or location of the procoagulant compound of the invention or a fragment thereof can be *in vivo*, *in vitro*, *ex vivo*, or combinations thereof.
- [0281] In other embodiments, a heterologous moiety increases stability of the procoagulant compound or conjugate of the invention. As used herein, the term "stability" refers to an art-recognized measure of the maintenance of one or more physical properties of the procoagulant compound in response to an environmental condition (e.g., an elevated or lowered temperature). In certain aspects, the physical property can be the maintenance of the covalent structure of the procoagulant compound (e.g., the absence of proteolytic cleavage, unwanted oxidation or deamidation). In other aspects, the physical property can also be the presence of the procoagulant compound in a properly folded state (e.g., the absence of soluble or insoluble aggregates or precipitates). In one aspect, the stability of the procoagulant compound is measured by assaying a biophysical property of the procoagulant compound, for example thermal stability, pH unfolding profile, stable removal of glycosylation, solubility, biochemical function (e.g., ability to bind to a protein, receptor or ligand), etc., and/or combinations thereof. In another aspect, biochemical function is demonstrated by the binding affinity of the interaction. In one

aspect, a measure of protein stability is thermal stability, i.e., resistance to thermal challenge. Stability can be measured using methods known in the art, such as, HPLC (high performance liquid chromatography), SEC (size exclusion chromatography), DLS (dynamic light scattering), etc. Methods to measure thermal stability include, but are not limited to differential scanning calorimetry (DSC), differential scanning fluorimetry (DSF), circular dichroism (CD), and thermal challenge assay.

Half-Life Extender Heterologous Moieties

[0282] In certain aspects, the heterologous moiety is a half-like extender, i.e., a heterologous moiety which increases the *in vivo* half-life of the compound or conjugate with respect to the *in vivo* half-life of the corresponding procoagulant compound or conjugate lacking such heterologous moiety. The *in vivo* half-life can be determined by any method known to those of skill in the art, e.g., activity assays (chromogenic assay or one stage clotting aPTT assay), ELISA, etc.

[0283] In some embodiments, the presence of one or more half-life extenders results in the half-life of the procoagulant compound or conjugate to be increased compared to the half life of the corresponding compound or conjugate lacking such one or more half-life extenders. The half-life of the procoagulant conjugate comprising a half-life extender is at least about 1.5 times, at least about 2 times, at least about 2.5 times, at least about 3 times, at least about 4 times, at least about 5 times, at least about 6 times, at least about 7 times, at least about 8 times, at least about 9 times, at least about 10 times, at least about 11 times, or at least about 12 times longer than the *in vivo* half-life of the corresponding procoagulant compound lacking such half-life extender.

[0284] In one embodiment, the half-life of the compound or conjugate linked to a half-life extender is about 1.5-fold to about 20-fold, about 1.5 fold to about 15 fold, or about 1.5 fold to about 10 fold longer than the *in vivo* half-life of the corresponding compound or conjugate not linked to such half-life extender. In another embodiment, the half-life of the compound or conjugate linked to a half-life extender is extended about 2-fold to about 10-fold, about 2-fold to about 9-fold, about 2-fold to about 8-fold, about 2-fold to about 7-fold, about 2-fold to about 6-fold, about 2-fold to about 5-fold, about 2-fold to about 4-fold, about 2-fold to about 3-fold, about 2.5-fold to about 10-fold, about 2.5-fold to about 9-fold, about 2.5-fold to about 8-fold, about 2.5-fold to about 7-fold, about 2.5-fold to about 6-fold, about 2.5-fold to about 5-fold, about 2.5-fold to about 4-fold, about 2.5-fold

to about 3-fold, about 3-fold to about 10-fold, about 3-fold to about 9-fold, about 3-fold to about 8-fold, about 3-fold to about 7-fold, about 3-fold to about 6-fold, about 3-fold to about 5-fold, about 3-fold to about 4-fold, about 4-fold to about 6 fold, about 5-fold to about 7-fold, or about 6-fold to about 8 fold as compared to the *in vivo* half-life of the corresponding compound or conjugate not linked to such half-life extender.

[0285] In other embodiments, the half-life of the compound or conjugate linked to a half-life extender is at least about 10 hours, at least about 17 hours, at least about 18 hours, at least about 19 hours, at least about 20 hours, at least about 21 hours, at least about 22 hours, at least about 23 hours, at least about 24 hours, at least about 25 hours, at least about 26 hours, at least about 27 hours, at least about 28 hours, at least about 29 hours, at least about 30 hours, at least about 31 hours, at least about 32 hours, at least about 33 hours, at least about 34 hours, at least about 35 hours, at least about 36 hours, at least about 48 hours, at least about 60 hours, at least about 72 hours, at least about 84 hours, at least about 96 hours, or at least about 108 hours.

[0286] In still other embodiments, the half-life of the compound or conjugate linked to a half-life extender is about 15 hours to about two weeks, about 16 hours to about one week, about 17 hours to about one week, about 18 hours to about one week, about 19 hours to about one week, about 20 hours to about one week, about 21 hours to about one week, about 22 hours to about one week, about 23 hours to about one week, about 24 hours to about one week, about 36 hours to about one week, about 48 hours to about one week, about 60 hours to about one week, about 24 hours to about six days, about 24 hours to about five days, about 24 hours to about four days, about 24 hours to about three days, or about 24 hours to about two days.

[0287] In some embodiments, the average half-life per subject of the compound or conjugate linked to a half-life extender is about 15 hours, about 16 hours, about 17 hours, about 18 hours, about 19 hours, about 20 hours, about 21 hours, about 22 hours, about 23 hours, about 24 hours (1 day), about 25 hours, about 26 hours, about 27 hours, about 28 hours, about 29 hours, about 30 hours, about 31 hours, about 32 hours, about 33 hours, about 34 hours, about 35 hours, about 36 hours, about 40 hours, about 44 hours, about 48 hours (2 days), about 54 hours, about 60 hours, about 72 hours (3 days), about 84 hours, about 96 hours (4 days), about 108 hours, about 120 hours (5 days), about six days, about

seven days (one week), about eight days, about nine days, about 10 days, about 11 days, about 12 days, about 13 days, or about 14 days.

(a) *Low Complexity Polypeptides*

[0288] In certain aspects, the compound or conjugate of the disclosure is linked to a heterologous moiety comprising a polypeptide with low compositional and/or structural complexity (e.g., a disordered polypeptide with no secondary or tertiary structure in solution under physiologic conditions).

(b) *CTP*

[0289] In certain aspects, the compound or conjugate of the disclosure is linked to a heterologous moiety comprising one β subunit of the C-terminal peptide (CTP) of human chorionic gonadotropin or fragment, variant, or derivative thereof. One or more CTP peptides inserted into a recombinant protein is known to increase the *in vivo* half-life of that protein. See, e.g., U.S. Patent No. 5,712,122, incorporated by reference herein in its entirety. Exemplary CTP peptides include DPRFQDSSSSKAPPPSLPSPSRLPGPSDTPIL or SSSSKAPPPSLPSPSRLPGPSDTPILPQ. See, e.g., U.S. Patent Application Publication No. US 2009/0087411 A1, incorporated herein by reference in its entirety.

(c) *Immunoglobulin_Constant Region (Fc) or Portion Thereof*

[0290] In certain aspects, the compound or the conjugate of the invention is linked to at least one Fc region. The term "Fc" or "Fc region" as used herein, means a functional neonatal Fc receptor (FcRn) binding partner comprising an Fc domain, variant, or fragment thereof which maintains the desirable properties of an Fc region in a chimeric protein, e.g., an increase in *in vivo* half-life. Myriad mutants, fragments, variants, and derivatives are described, e.g., in PCT Publication Nos. WO 2011/069164 A2, WO 2012/006623 A2, WO 2012/006635 A2, or WO 2012/006633 A2, all of which are incorporated herein by reference in their entireties. An Fc region is comprised of domains denoted CH (constant heavy) domains (CH1, CH2, etc.). Depending on the isotype, (*i.e.* IgG, IgM, IgA IgD, or IgE), the Fc region can be comprised of three or four CH domains. Some isotypes (*e.g.* IgG) Fc regions also contain a hinge region. See Janeway *et al.* 2001, *Immunobiology*, Garland Publishing, N.Y., N.Y.

[0291] An Fc region or a portion thereof for producing the procoagulant conjugate of the present invention can be obtained from a number of different sources. In some embodiments, an Fc region or a portion thereof is derived from a human immunoglobulin. It is understood, however, that the Fc region or a portion thereof can be derived from an immunoglobulin of another mammalian species, including for example, a rodent (e.g. a mouse, rat, rabbit, guinea pig) or non-human primate (e.g. chimpanzee, macaque) species. Moreover, the Fc region or a portion thereof can be derived from any immunoglobulin class, including IgM, IgG, IgD, IgA and IgE, and any immunoglobulin isotype, including IgG1, IgG2, IgG3 and IgG4. In one embodiment, the human isotype IgG1 is used.

[0292] Conjugates comprising an Fc region of an immunoglobulin bestow several desirable properties on a chimeric protein including increased stability, increased serum half-life (see Capon *et al.*, 1989, *Nature* 337:525) as well as binding to Fc receptors such as the neonatal Fc receptor (FcRn) (U.S. Pat. Nos. 6,086,875, 6,485,726, 6,030,613; WO 03/077834; US2003-0235536A1), which are incorporated herein by reference in their entireties.

[0293] In certain embodiments, the compound or conjugate is linked to one or more truncated Fc regions that are nonetheless sufficient to confer Fc receptor (FcR) binding properties to the Fc region. For example, the portion of an Fc region that binds to FcRn (i.e., the FcRn binding portion) comprises from about amino acids 282-438 of IgG1, EU numbering (with the primary contact sites being amino acids 248, 250-257, 272, 285, 288, 290-291, 308-311, and 314 of the CH2 domain and amino acid residues 385-387, 428, and 433-436 of the CH3 domain. Thus, an Fc region in a procoagulant compound of the invention may comprise or consist of an FcRn binding portion. FcRn binding portions may be derived from heavy chains of any isotype, including IgG1, IgG2, IgG3 and IgG4. In one embodiment, an FcRn binding portion from an antibody of the human isotype IgG1 is used. In another embodiment, an FcRn binding portion from an antibody of the human isotype IgG4 is used.

[0294] In certain embodiments, an Fc region comprises at least one of: a hinge (e.g., upper, middle, and/or lower hinge region) domain (about amino acids 216-230 of an antibody Fc region according to EU numbering), a CH2 domain (about amino acids 231-340 of an antibody Fc region according to EU numbering), a CH3 domain (about amino acids 341-438 of an antibody Fc region according to EU numbering), a CH4 domain, or a

variant, portion, or fragment thereof. In other embodiments, an Fc region comprises a complete Fc domain (i.e., a hinge domain, a CH2 domain, and a CH3 domain). In some embodiments, an Fc region comprises, consists essentially of, or consists of a hinge domain (or a portion thereof) fused to a CH3 domain (or a portion thereof), a hinge domain (or a portion thereof) fused to a CH2 domain (or a portion thereof), a CH2 domain (or a portion thereof) fused to a CH3 domain (or a portion thereof), a CH2 domain (or a portion thereof) fused to both a hinge domain (or a portion thereof) and a CH3 domain (or a portion thereof). In still other embodiments, an Fc region lacks at least a portion of a CH2 domain (e.g., all or part of a CH2 domain). In a particular embodiment, an Fc region comprises or consists of amino acids corresponding to EU numbers 221 to 447.

[0295] An Fc in a procoagulant compound of the invention can include, for example, a change (e.g., a substitution) at one or more of the amino acid positions disclosed in Int'l. PCT Publs. WO88/07089A1, WO96/14339A1, WO98/05787A1, WO98/23289A1, WO99/51642A1, WO99/58572A1, WO00/09560A2, WO00/32767A1, WO00/42072A2, WO02/44215A2, WO02/060919A2, WO03/074569A2, WO04/016750A2, WO04/029207A2, WO04/035752A2, WO04/063351A2, WO04/074455A2, WO04/099249A2, WO05/040217A2, WO04/044859, WO05/070963A1, WO05/077981A2, WO05/092925A2, WO05/123780A2, WO06/019447A1, WO06/047350A2, and WO06/085967A2; U.S. Pat. Publ. Nos. US 2007/0231329, US2007/0231329, US2007/0237765, US2007/0237766, US2007/0237767, US2007/0243188, US2007/0248603, US2007/0286859, US2008/0057056; or U.S. Pat. Nos. 5,648,260; 5,739,277; 5,834,250; 5,869,046; 6,096,871; 6,121,022; 6,194,551; 6,242,195; 6,277,375; 6,528,624; 6,538,124; 6,737,056; 6,821,505; 6,998,253; 7,083,784; 7,404,956, and 7,317,091, each of which is incorporated by reference herein in its entirety. In one embodiment, the specific change (e.g., the specific substitution of one or more amino acids disclosed in the art) may be made at one or more of the disclosed amino acid positions. In another embodiment, a different change at one or more of the disclosed amino acid positions (e.g., the different substitution of one or more amino acid position disclosed in the art) may be made.

[0296] An Fc region used in the invention may also comprise an art recognized amino acid substitution which alters its glycosylation. For example, the Fc has a mutation

leading to reduced glycosylation (*e.g.*, N- or O-linked glycosylation) or may comprise an altered glycoform of the wild-type Fc moiety (*e.g.*, a low fucose or fucose-free glycan).

(d) Albumin or Fragment, or Variant Thereof

[0297] In certain embodiments, the compound or conjugate of the invention is linked to a heterologous moiety comprising albumin or a functional fragment thereof. Human serum albumin (HSA, or HA), a protein of 609 amino acids in its full-length form, is responsible for a significant proportion of the osmotic pressure of serum and also functions as a carrier of endogenous and exogenous ligands. The term “albumin” as used herein includes full-length albumin or a functional fragment, variant, derivative, or analog thereof. Examples of albumin or the fragments or variants thereof are disclosed in US Pat. Publ. Nos. 2008/0194481A1, 2008/0004206 A1, 2008/0161243 A1, 2008/0261877 A1, or 2008/0153751 A1 or PCT Appl. Publ. Nos. 2008/033413 A2, 2009/058322 A1, or 2007/021494 A2, which are incorporated herein by reference in their entireties.

[0298] In one embodiment, the heterologous moiety is albumin, a fragment, or a variant thereof which is further linked to a heterologous moiety selected from the group consisting of an immunoglobulin constant region or portion thereof (*e.g.*, an Fc region), a PAS sequence, HES, and PEG.

(e) Albumin Binding Moiety

[0299] In certain embodiments, the heterologous moiety is an albumin binding moiety, which comprises an albumin binding peptide, a bacterial albumin binding domain, an albumin-binding antibody fragment, or any combinations thereof.

[0300] For example, the albumin binding protein can be a bacterial albumin binding protein, an antibody or an antibody fragment including domain antibodies (see U.S. Pat. No. 6,696,245). An albumin binding protein, for example, can be a bacterial albumin binding domain, such as the one of streptococcal protein G (Konig, T. and Skerra, A. (1998) *J. Immunol. Methods* 218, 73-83). Other examples of albumin binding peptides that can be used as conjugation partner are, for instance, those having a Cys-Xaa₁-Xaa₂-Xaa₃-Xaa₄-Cys consensus sequence, wherein Xaa₁ is Asp, Asn, Ser, Thr, or Trp; Xaa₂ is Asn, Gln, His, Ile, Leu, or Lys; Xaa₃ is Ala, Asp, Phe, Trp, or Tyr; and Xaa₄ is Asp, Gly, Leu, Phe, Ser, or Thr as described in US patent application 2003/0069395 or Dennis et al. (Dennis et al. (2002) *J. Biol. Chem.* 277, 35035-35043).

[0301] Domain 3 from streptococcal protein G, as disclosed by Kraulis *et al.*, FEBS Lett. 378:190-194 (1996) and Linhult *et al.*, Protein Sci. 11:206-213 (2002) is an example of a bacterial albumin-binding domain. Examples of albumin-binding peptides include a series of peptides having the core sequence DICLPRWGCLW (SEQ ID NO:45). See, *e.g.*, Dennis *et al.*, J. Biol. Chem. 2002, 277: 35035-35043 (2002). Examples of albumin-binding antibody fragments are disclosed in Muller and Kontermann, Curr. Opin. Mol. Ther. 9:319-326 (2007); Rooverset *et al.*, Cancer Immunol. Immunother. 56:303-317 (2007), and Holt *et al.*, Prot. Eng. Design Sci., 21:283-288 (2008), which are incorporated herein by reference in their entireties. An example of such albumin binding moiety is 2-(3-maleimidopropanamido)-6-(4-(4-iodophenyl)butanamido) hexanoate ("Albu" tag) as disclosed by Trusset *et al.*, Bioconjugate Chem. 20:2286-2292 (2009).

(f) *PAS Sequence*

[0302] In other embodiments, the heterologous moiety is a PAS sequence. A PAS sequence, as used herein, means an amino acid sequence comprising mainly alanine and serine residues or comprising mainly alanine, serine, and proline residues, the amino acid sequence forming random coil conformation under physiological conditions. Accordingly, the PAS sequence is a building block, an amino acid polymer, or a sequence cassette comprising, consisting essentially of, or consisting of alanine, serine, and proline which can be used as a part of the heterologous moiety in the procoagulant compound. Yet, the skilled person is aware that an amino acid polymer also may form random coil conformation when residues other than alanine, serine, and proline are added as a minor constituent in the PAS sequence. The term "minor constituent" as used herein means that amino acids other than alanine, serine, and proline may be added in the PAS sequence to a certain degree, *e.g.*, up to about 12%, *i.e.*, about 12 of 100 amino acids of the PAS sequence, up to about 10%, *i.e.*, about 10 of 100 amino acids of the PAS sequence, up to about 9%, *i.e.*, about 9 of 100 amino acids, up to about 8%, *i.e.*, about 8 of 100 amino acids, about 6%, *i.e.*, about 6 of 100 amino acids, about 5%, *i.e.*, about 5 of 100 amino acids, about 4%, *i.e.*, about 4 of 100 amino acids, about 3%, *i.e.*, about 3 of 100 amino acids, about 2%, *i.e.*, about 2 of 100 amino acids, about 1%, *i.e.*, about 1 of 100 of the amino acids. The amino acids different from alanine, serine and proline may be selected from the group consisting of Arg, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Leu, Lys, Met, Phe, Thr, Trp, Tyr, and Val.

[0303] Under physiological conditions, the PAS sequence stretch forms a random coil conformation and thereby can mediate an increased *in vivo* and/or *in vitro* stability to procoagulant compound. Since the random coil domain does not adopt a stable structure or function by itself, the biological activity mediated by the Pep1 and/or Pep2 polypeptides in the procoagulant compound is essentially preserved. In other embodiments, the PAS sequences that form random coil domain are biologically inert, especially with respect to proteolysis in blood plasma, immunogenicity, isoelectric point/electrostatic behaviour, binding to cell surface receptors or internalisation, but are still biodegradable, which provides clear advantages over synthetic polymers such as PEG.

[0304] Non-limiting examples of the PAS sequences forming random coil conformation comprise an amino acid sequence selected from the group consisting of ASPAAPAPASPAAPAPSAPA, AAPASPAPAAPSAPAPAAPS, APSSPSPSAPSSPSPASPSS, APSSPSPSAPSSPSPASPS, SSPSAPSPSSPASPSPSSPA, AASPAAPSAPPAAASPAAPSAPPA, and ASAAAPAAASAAASAPSAAA, or any combinations thereof. Additional examples of PAS sequences are known from, e.g., US Pat. Publ. No. 2010/0292130 A1 and PCT Appl. Publ. No. WO 2008/155134 A1.

(g) *HAP Sequence*

[0305] In certain embodiments, the heterologous moiety is a glycine-rich homo-amino-acid polymer (HAP). The HAP sequence can comprise a repetitive sequence of glycine, which has at least 50 amino acids, at least 100 amino acids, 120 amino acids, 140 amino acids, 160 amino acids, 180 amino acids, 200 amino acids, 250 amino acids, 300 amino acids, 350 amino acids, 400 amino acids, 450 amino acids, or 500 amino acids in length. In one embodiment, the HAP sequence is capable of extending half-life of a moiety fused to or linked to the HAP sequence. Non-limiting examples of the HAP sequence includes, but are not limited to $(\text{Gly})_n$, $(\text{Gly}_4\text{Ser})_n$ or $\text{S}(\text{Gly}_4\text{Ser})_n$, wherein n is 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20. In one embodiment, n is 20, 21, 22, 23, 24, 25, 26, 26, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, or 40. In another embodiment, n is 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, or 200.

(h) *Transferrin or Fragment thereof*

[0306] In certain embodiments, the heterologous moiety is transferrin or a fragment thereof. Any transferrin may be used to make the conjugates of the invention. As an example, wild-type human Tf (Tf) is a 679 amino acid protein, of approximately 75 KDa (not accounting for glycosylation), with two main domains, N (about 330 amino acids) and C (about 340 amino acids), which appear to originate from a gene duplication. See GenBank accession numbers NM001063, XM002793, M12530, XM039845, XM 039847 and S95936 (www.ncbi.nlm.nih.gov/), all of which are herein incorporated by reference in their entirety. Transferrin comprises two domains, N domain and C domain. N domain comprises two subdomains, N1 domain and N2 domain, and C domain comprises two subdomains, C1 domain and C2 domain.

[0307] In one embodiment, the transferrin heterologous moiety includes a transferrin splice variant. In one example, a transferrin splice variant can be a splice variant of human transferrin, e.g., Genbank Accession AAA61140. In another embodiment, the transferrin portion of the chimeric protein includes one or more domains of the transferrin sequence, e.g., N domain, C domain, N1 domain, N2 domain, C1 domain, C2 domain or any combinations thereof.

(i) *Polymer, e.g., Polyethylene Glycol (PEG)*

[0308] In other embodiments, the heterologous moiety is a soluble polymer known in the art, including, but not limited to, polyethylene glycol, ethylene glycol/propylene glycol copolymers, carboxymethylcellulose, dextran, or polyvinyl alcohol. Also provided by the invention are procoagulant compounds of the invention comprising heterologous moieties which may provide additional advantages such as increased solubility, stability and circulating time of the polypeptide, or decreased immunogenicity (see U.S. Pat. No. 4,179,337). Such heterologous moieties for modification can be selected from the group consisting of water soluble polymers including, but not limited to, polyethylene glycol, ethylene glycol/propylene glycol copolymers, carboxymethylcellulose, dextran, and polyvinyl alcohol, poly(alkylene oxide), poly(vinyl pyrrolidone), polyoxazoline, or poly(acryloylmorpholine).

[0309] The polymer can be of any molecular weight, and can be branched or unbranched. For polyethylene glycol, in one embodiment, the molecular weight is between about 1

kDa and about 100 kDa for ease in handling and manufacturing. Other sizes may be used, depending on the desired profile (e.g., the duration of sustained release desired, the effects, if any on biological activity, the ease in handling, the degree or lack of antigenicity and other known effects of the polyethylene glycol to a protein or analog). For example, the polyethylene glycol may have an average molecular weight of about 200, 500, 1000, 1500, 2000, 2500, 3000, 3500, 4000, 4500, 5000, 5500, 6000, 6500, 7000, 7500, 8000, 8500, 9000, 9500, 10,000, 10,500, 11,000, 11,500, 12,000, 12,500, 13,000, 13,500, 14,000, 14,500, 15,000, 15,500, 16,000, 16,500, 17,000, 17,500, 18,000, 18,500, 19,000, 19,500, 20,000, 25,000, 30,000, 35,000, 40,000, 45,000, 50,000, 55,000, 60,000, 65,000, 70,000, 75,000, 80,000, 85,000, 90,000, 95,000, or 100,000 kDa.

[0310] In some embodiments, the polyethylene glycol may have a branched structure. Branched polyethylene glycols are described, for example, in U.S. Pat. No. 5,643,575; Morpurgo *et al.*, *Appl. Biochem. Biotechnol.* 56:59-72 (1996); Vorobjev *et al.*, *Nucleosides Nucleotides* 18:2745-2750 (1999); and Caliceti *et al.*, *Bioconjug. Chem.* 10:638-646 (1999), each of which is incorporated herein by reference in its entirety.

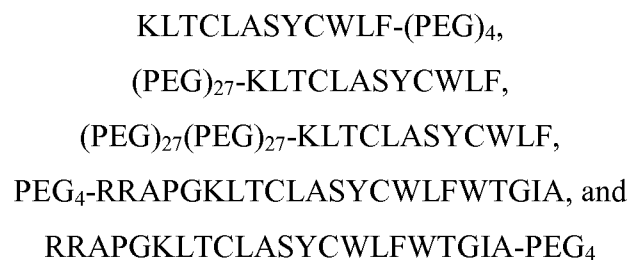
[0311] The number of polyethylene glycol moieties attached to each compound or conjugate of the invention (i.e., the degree of substitution) may also vary. For example, the PEGylated compound or conjugate may be linked, on average, to 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 12, 15, 17, 20, or more polyethylene glycol molecules. Similarly, the average degree of substitution within ranges such as 1-3, 2-4, 3-5, 4-6, 5-7, 6-8, 7-9, 8-10, 9-11, 10-12, 11-13, 12-14, 13-15, 14-16, 15-17, 16-18, 17-19, or 18-20 polyethylene glycol moieties per protein molecule. Methods for determining the degree of substitution are discussed, for example, in Delgado *et al.*, *Crit. Rev. Thera. Drug Carrier Sys.* 9:249-304 (1992).

[0312] In one example, the heterologous moiety is a water-soluble polymer, e.g., a straight or branched polyethylene glycol (PEG) moieties, straight or branched polypropylene glycol (PPG) moiety, a hydroxyethyl starch (HES) moiety, or a half-life extension polypeptide (e.g., XTEN; see, e.g., Schellenberger V. *et al.*, *Nat Biotechnol.* 2009 Dec;27(12):1186-90, US20100189682, PCT/US07/05952, US 12/228,859, US 12/646,566, PCT/US08/09787, US 12/699,761, and PCT/US2010/23106, each of which is incorporated herein by reference in its entirety). The polymer can be attached to the N-terminus, the C-terminus of the peptide sequence or an internal amino acid of the

compound or the polypeptide. In one example, the polymer is attached to the N-terminal amino acid, e.g. via an amide bond. In another example, the polymer is attached to the C-terminal amino acid via an amide bond. In yet another example, the polymer is attached to the peptide sequence through derivatization of an amino acid side chain located at an internal position of the amino acid sequence. For example, the polymer is attached to the peptide sequence via derivatization of a tyrosine side chain, a lysine side chain or a aspartic acid or glutamic acid side chain.

[0313] Suitable methods of PEGylation are disclosed, e.g., in U.S. Pat. Nos. 5,122,614 to Zalipsky *et al.*, and 5,539,063 to Hakimi *et al.*, all of which PEGylation methods are incorporated herein by reference. Various molecular weights of PEG may be used, suitably from 1000 Da to 80,000 Da (or from 5000 Da to 60,000 DA). In one example, the PEG is monodisperse, meaning that there is little variation in molecular weight between PEG molecules. PEGylation may improve the solubility and plasma half-life of a peptide.

[0314] In one example according to this embodiment, the compound of the present disclosure contains an amino acid sequence selected from or is a peptide selected from:



or a retro-, an inverso- or a retro-inverso variant thereof. In one example in the above peptides, the N-terminal amino acid is acetylated, and the C-terminal amino acid is amidated. In another example in the above peptides, the N-terminus is free (-NH₂ or a salt form thereof), and the C-terminal amino acid is amidated.

(j) *XTEN Moieties*

[0315] In certain aspects, a compound of the invention is covalently linked to at least one heterologous moiety that is or comprises an XTEN polypeptide or fragment, variant, or derivative thereof. As used here "XTEN polypeptide" refers to extended length polypeptides with non-naturally occurring, substantially non-repetitive sequences that are composed mainly of small hydrophilic amino acids, with the sequence having a low degree or no secondary or tertiary structure under physiologic conditions. As a

heterologous moiety, XTENs can serve as a half-life extension moiety. In addition, XTEN can provide desirable properties including but are not limited to enhanced pharmacokinetic parameters and solubility characteristics.

[0316] The incorporation of a heterologous moiety comprising an XTEN sequence into a conjugate of the invention can confer one or more of the following advantageous properties to the resulting conjugate: conformational flexibility, enhanced aqueous solubility, high degree of protease resistance, low immunogenicity, low binding to mammalian receptors, or increased hydrodynamic (or Stokes) radii.

[0317] In certain aspects, an XTEN moiety can increase pharmacokinetic properties such as longer *in vivo* half-life or increased area under the curve (AUC), so that a compound or conjugate of the invention stays *in vivo* and has procoagulant activity for an increased period of time compared to a compound or conjugate with the same but without the XTEN heterologous moiety.

[0318] Examples of XTEN moieties that can be used as heterologous moieties in procoagulant conjugates of the invention are disclosed, *e.g.*, in U.S. Patent Publication Nos. 2010/0239554 A1, 2010/0323956 A1, 2011/0046060 A1, 2011/0046061 A1, 2011/0077199 A1, or 2011/0172146 A1, or International Patent Publication Nos. WO 2010091122 A1, WO 2010144502 A2, WO 2010144508 A1, WO 2011028228 A1, WO 2011028229 A1, or WO 2011028344 A2, each of which is incorporated by reference herein in its entirety.

(k) *Hydroxyethyl Starch (HES)*

[0319] In certain embodiments, the heterologous moiety is hydroxyethyl starch (HES) or a derivative thereof. Hydroxyethyl starch (HES) is a derivative of naturally occurring amylopectin and is degraded by alpha-amylase in the body. HES is a substituted derivative of the carbohydrate polymer amylopectin, which is present in corn starch at a concentration of up to 95% by weight. HES exhibits advantageous biological properties and is used as a blood volume replacement agent and in hemodilution therapy in the clinics (Sommermeyer *et al.*, *Krankenhauspharmazie*, 8(8), 271-278 (1987); and Weidler *et al.*, *Arzneim.-Forschung/Drug Res.*, 41, 494-498 (1991)).

[0320] Amylopectin contains glucose moieties, wherein in the main chain alpha-1,4-glycosidic bonds are present and at the branching sites alpha-1,6-glycosidic bonds are found. The physical-chemical properties of this molecule are mainly determined by the

type of glycosidic bonds. Due to the nicked alpha-1,4-glycosidic bond, helical structures with about six glucose-monomers per turn are produced. The physico-chemical as well as the biochemical properties of the polymer can be modified via substitution. The introduction of a hydroxyethyl group can be achieved via alkaline hydroxyethylation. By adapting the reaction conditions it is possible to exploit the different reactivity of the respective hydroxy group in the unsubstituted glucose monomer with respect to a hydroxyethylation. Owing to this fact, the skilled person is able to influence the substitution pattern to a limited extent.

[0321] HES is mainly characterized by the molecular weight distribution and the degree of substitution. The degree of substitution, denoted as DS, relates to the molar substitution, is known to the skilled people. See Sommermeyer *et al.*, *Krankenhauspharmazie*, 8(8), 271-278 (1987), as cited above, in particular p. 273.

[0322] In one embodiment, hydroxyethyl starch has a mean molecular weight (weight mean) of from 1 to 300 kD, from 2 to 200kD, from 3 to 100 kD, or from 4 to 70kD. hydroxyethyl starch can further exhibit a molar degree of substitution of from 0.1 to 3, preferably 0.1 to 2, more preferred, 0.1 to 0.9, preferably 0.1 to 0.8, and a ratio between C2:C6 substitution in the range of from 2 to 20 with respect to the hydroxyethyl groups. A non-limiting example of HES having a mean molecular weight of about 130 kD is a HES with a degree of substitution of 0.2 to 0.8 such as 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, or 0.8, preferably of 0.4 to 0.7 such as 0.4, 0.5, 0.6, or 0.7. In a specific embodiment, HES with a mean molecular weight of about 130 kD is VOLUVEN[®] from Fresenius. VOLUVEN[®] is an artificial colloid, employed, e.g., for volume replacement used in the therapeutic indication for therapy and prophylaxis of hypovolemia. The characteristics of VOLUVEN[®] are a mean molecular weight of 130,000+/-20,000 D, a molar substitution of 0.4 and a C2:C6 ratio of about 9:1. In other embodiments, ranges of the mean molecular weight of hydroxyethyl starch are, e.g., 4 to 70 kD or 10 to 70 kD or 12 to 70 kD or 18 to 70 kD or 50 to 70 kD or 4 to 50 kD or 10 to 50 kD or 12 to 50 kD or 18 to 50 kD or 4 to 18 kD or 10 to 18 kD or 12 to 18 kD or 4 to 12 kD or 10 to 12 kD or 4 to 10 kD. In still other embodiments, the mean molecular weight of hydroxyethyl starch employed is in the range of from more than 4 kD and below 70 kD, such as about 10 kD, or in the range of from 9 to 10 kD or from 10 to 11 kD or from 9 to 11 kD, or about 12 kD, or in the range of from 11 to 12 kD) or from 12 to 13 kD or from 11 to 13 kD, or

about 18 kD, or in the range of from 17 to 18 kD or from 18 to 19 kD or from 17 to 19 kD, or about 30 kD, or in the range of from 29 to 30, or from 30 to 31 kD, or about 50 kD, or in the range of from 49 to 50 kD or from 50 to 51 kD or from 49 to 51 kD.

[0323] In certain embodiments, the heterologous moiety can be a mixture of hydroxyethyl starches having different mean molecular weights and/or different degrees of substitution and/or different ratios of C2: C6 substitution. Therefore, mixtures of hydroxyethyl starches may be employed having different mean molecular weights and different degrees of substitution and different ratios of C2: C6 substitution, or having different mean molecular weights and different degrees of substitution and the same or about the same ratio of C2:C6 substitution, or having different mean molecular weights and the same or about the same degree of substitution and different ratios of C2:C6 substitution, or having the same or about the same mean molecular weight and different degrees of substitution and different ratios of C2:C6 substitution, or having different mean molecular weights and the same or about the same degree of substitution and the same or about the same ratio of C2:C6 substitution, or having the same or about the same mean molecular weights and different degrees of substitution and the same or about the same ratio of C2:C6 substitution, or having the same or about the same mean molecular weight and the same or about the same degree of substitution and different ratios of C2: C6 substitution, or having about the same mean molecular weight and about the same degree of substitution and about the same ratio of C2:C6 substitution.

(l) *Polysialic Acids (PSA)*

[0324] In certain embodiments, the heterologous moiety is a polysialic acids (PSAs) or a derivative thereof. Polysialic acids (PSAs) are naturally occurring unbranched polymers of sialic acid produced by certain bacterial strains and in mammals in certain cells Roth J., et al. (1993) in *Polysialic Acid: From Microbes to Man*, eds Roth J., Rutishauser U., Troy F. A. (Birkhäuser Verlag, Basel, Switzerland), pp 335–348.. They can be produced in various degrees of polymerisation from n=about 80 or more sialic acid residues down to n=2 by limited acid hydrolysis or by digestion with neuraminidases, or by fractionation of the natural, bacterially derived forms of the polymer. The composition of different polysialic acids also varies such that there are homopolymeric forms i.e. the alpha-2,8-linked polysialic acid comprising the capsular polysaccharide of *E. coli* strain K1 and the group-B meningococci, which is also found on

the embryonic form of the neuronal cell adhesion molecule (N-CAM). Heteropolymeric forms also exist—such as the alternating alpha-2,8 alpha-2,9 polysialic acid of *E. coli* strain K92 and group C polysaccharides of *N. meningitidis*. Sialic acid may also be found in alternating copolymers with monomers other than sialic acid such as group W135 or group Y of *N. meningitidis*. Polysialic acids have important biological functions including the evasion of the immune and complement systems by pathogenic bacteria and the regulation of glial adhesiveness of immature neurons during foetal development (wherein the polymer has an anti-adhesive function) Cho and Troy, *P.N.A.S., USA*, 91 (1994) 11427-11431, although there are no known receptors for polysialic acids in mammals. The alpha-2,8-linked polysialic acid of *E. coli* strain K1 is also known as ‘colominic acid’ and is used (in various lengths) to exemplify the present invention. Various methods of attaching or conjugating polysialic acids to a polypeptide have been described (for example, see U.S. Pat. No. 5,846,951; WO-A-0187922, and US 2007/0191597 A1, which are incorporated herein by reference in their entireties.

(m) *Clearance Receptors*

[0325] In some embodiments the heterologous moiety comprising a clearance receptor, fragment, variant, or derivative thereof. For example, soluble forms of clearance receptors, such as the low density lipoprotein-related protein receptor LRP1, or fragments thereof, can block binding of a polypeptide (e.g., FVIII or FIX) to clearance receptors and thereby extend its *in vivo* half-life.

[0326] LRP1 is a 600 kDa integral membrane protein that is implicated in the receptor-mediated clearance of a variety of proteins. See, e.g., Lenting *et al.*, *Haemophilia* 16:6-16 (2010). LRP1 also mediates clearance of Factor IX (see, e.g., Strickland & Medved. *J. Thromb. Haemostat.* 4:1484-1486 (2006).

[0327] Other suitable clearance receptors are, e.g., LDLR (low-density lipoprotein receptor), VLDLR (very low-density lipoprotein receptor), and megalin (LRP-2), or fragments thereof. See, e.g., Bovenschen *et al.*, *Blood* 106:906-912 (2005); Bovenschen, *Blood* 116:5439-5440 (2010); Martinelli *et al.*, *Blood* 116:5688-5697 (2010).

Visualization and Location

[0328] In certain embodiments, the heterologous moiety facilitates visualization or localization of the compounds or conjugate of the invention. Peptides and other moieties

for insertion or conjugation into a compound which facilitate visualization or localization are known in the art. Such moieties can be used to facilitate visualization or localization *in vitro*, *in vivo*, *ex vivo* or any combination thereof.

[0329] Since thrombin plays a central role in the coagulation cascade, detection of imaging of its activity *in vivo* is highly desired. Accordingly, various heterologous moieties facilitate visualization or localization of the compounds or conjugates of the invention (e.g., fluorescent dyes) and can be engineered into the conjugates of the invention. In some embodiments, fluorescent dyes can be engineered to be non-fluorescent until their amines are regenerated upon thrombin cleavage.

[0330] Non-limiting examples of peptides or polypeptides which enable visualization or localization include biotin acceptor peptides which can facilitate conjugation of avidin- and streptavidin-based reagents, lipoic acid acceptor peptides which can facilitate conjugation of thiol-reactive probes to bound lipoic acid or direct ligation of fluorescent lipoic acid analogs, fluorescent proteins, e.g., green fluorescent protein (GFP) and variants thereof (e.g., EGFP, YFP such as EYFP, mVenus, YPet or Citrine, or CFP such as Cerulean or ECFP) or red fluorescent protein (RFP), cysteine-containing peptides for ligation of biarsenical dyes such as 4',5'-bis(1,3,2-dithioarsolan-2-yl)fluorescein (FlAsH), or for conjugating metastable technetium, peptides for conjugating europium clathrates for fluorescence resonance energy transfer (FRET)-based proximity assays, any variants, thereof, and any combination thereof.

[0331] Procoagulant compounds of the present disclosure labeled by these techniques can be used, for example, for 3-D imaging of pathological thrombus formation and dissolution, tumor imaging in procoagulant malignancies, flow cytometric quantitation and characterization of procoagulant microparticles in blood and plasma, monitoring of thrombus formation by intravital microscopy.

Polypeptides

FVIII

[0332] The term "FVIII" or "Factor VIII", as used herein, means functional FVIII protein in its normal role in coagulation, unless otherwise specified (i.e., refers to any FVIII moiety which exhibits biological activity that is associated with native FVIII). Thus, the term FVIII includes FVIII variant proteins that are functional. Preferred FVIII proteins

are primate (e.g., chimpanzee), human, porcine, canine, and murine FVIII proteins. The full length polypeptide and polynucleotide sequences of FVIII are known, as are many functional fragments, mutants and modified versions. Exemplary FVIII sequences are disclosed, e.g., in WO2011/069164. FVIII polypeptides include, e.g., full-length FVIII, full-length FVIII minus Met at the N-terminus, mature FVIII (minus the signal sequence), mature FVIII with an additional Met at the N-terminus, and/or FVIII with a full or partial deletion of the B domain. In one example, the FVIII is a variant in which the B domain is deleted, either partially or fully. An exemplary sequence of FVIII can be found as NCBI Accession Number NP000123 or UniProtKB/Swiss-Prot entry P00451.

[0333] A number of functional FVIII molecules, including B-domain deletions, are disclosed in the following patents: US 6,316,226 and US 6,346,513, both assigned to Baxter; US 7,041,635 assigned to In2Gen; US 5,789,203, US 6,060,447, US 5,595,886, and US 6,228,620 assigned to Chiron; US 5,972,885 and US 6,048,720 assigned to Biovitrum, US 5,543,502 and US 5,610,278 assigned to Novo Nordisk; US 5,171,844 assigned to Immuno Ag; US 5,112,950 assigned to Transgene S.A.; US 4,868,112 assigned to Genetics Institute, each of which is incorporated herein by reference in its entirety.

[0334] As used herein, "plasma-derived FVIII" includes all forms of the protein found in blood obtained from a mammal having the property of activating the coagulation pathway.

[0335] "B domain" of FVIII, as used herein, is the same as the B domain known in the art that is defined by internal amino acid sequence identity and sites of proteolytic cleavage by thrombin, e.g., residues Ser741-Arg1648 of full length human FVIII. The other human FVIII domains are defined by the following amino acid residues: A1, residues Ala1-Arg372; A2, residues Ser373-Arg740; A3, residues Ser1690-Ile2032; C1, residues Arg2033-Asn2172; C2, residues Ser2173-Tyr2332. The A3-C1-C2 sequence includes residues Ser1690-Tyr2332. The remaining sequence, residues Glu1649-Arg1689, is usually referred to as the FVIII light chain activation peptide. The locations of the boundaries for all of the domains, including the B domains, for porcine, mouse and canine FVIII are also known in the art. Preferably, the B domain of FVIII is deleted ("B domain deleted FVIII" or "BDD FVIII"). An example of a BDD FVIII is REFACTO (recombinant BDD FVIII).

[0336] A "B domain deleted FVIII" may have the full or partial deletions disclosed in U.S. Patent Nos. 6,316,226, 6,346,513, 7,041,635, 5,789,203, 6,060,447, 5,595,886, 6,228,620, 5,972,885, 6,048,720, 5,543,502, 5,610,278, 5,171,844, 5,112,950, 4,868,112, and 6,458,563, each of which is incorporated herein by reference in its entirety. In some embodiments, a B domain deleted FVIII sequence of the present invention comprises any one of the deletions disclosed at col. 4, line 4 to col. 5, line 28 and examples 1-5 of U.S. Patent No. 6,316,226 (also in US 6,346,513). In some embodiments, a B domain deleted FVIII of the present invention has a deletion disclosed at col. 2, lines 26-51 and examples 5-8 of U.S. Patent No. 5,789,203 (also US 6,060,447, US 5,595,886, and US 6,228,620). In some embodiments, a B domain deleted FVIII has a deletion described in col. 1, lines 25 to col. 2, line 40 of US Patent No. 5,972,885; col. 6, lines 1-22 and example 1 of U.S. Patent no. 6,048,720; col. 2, lines 17-46 of U.S. Patent No. 5,543,502; col. 4, line 22 to col. 5, line 36 of U.S. Patent no. 5,171,844; col. 2, lines 55-68, figure 2, and example 1 of U.S. Patent No. 5,112,950; col. 2, line 2 to col. 19, line 21 and table 2 of U.S. Patent No. 4,868,112; col. 2, line 1 to col. 3, line 19, col. 3, line 40 to col. 4, line 67, col. 7, line 43 to col. 8, line 26, and col. 11, line 5 to col. 13, line 39 of U.S. Patent no. 7,041,635; or col. 4, lines 25-53, of U.S. Patent No. 6,458,563. In some embodiments, a B domain deleted FVIII has a deletion of most of the B domain, but still contains amino-terminal sequences of the B domain that are essential for *in vivo* proteolytic processing of the primary translation product into two polypeptide chain, as disclosed in WO 91/09122, which is incorporated herein by reference in its entirety. In some embodiments, a B domain deleted FVIII is constructed with a deletion of amino acids 747-1638, i.e., virtually a complete deletion of the B domain. Hoebe R.C., *et al. J. Biol. Chem.* 265 (13): 7318-7323 (1990), incorporated herein by reference in its entirety. A B domain deleted FVIII may also contain a deletion of amino acids 771-1666 or amino acids 868-1562 of FVIII. Meulien P., *et al. Protein Eng.* 2(4): 301-6 (1988), incorporated herein by reference in its entirety. Additional B domain deletions that are part of the invention include, e.g.,: deletion of amino acids 982 through 1562 or 760 through 1639 (Toole et al., Proc. Natl. Acad. Sci. U.S.A. (1986) 83, 5939-5942)), 797 through 1562 (Eaton, et al. Biochemistry (1986) 25:8343-8347)), 741 through 1646 (Kaufman (PCT published application No. WO 87/04187)), 747-1560 (Sarver, et al., DNA (1987) 6:553-564)), 741 through 1648 (Pasek (PCT application No.88/00831)), 816 through 1598 or 741 through 1689 (Lagner

(Behring Inst. Mitt. (1988) No 82:16-25, EP 295597)), each of which is incorporated herein by reference in its entirety. Each of the foregoing deletions may be made in any FVIII sequence.

[0337] In one example, the FVIII polypeptide is a single-chain FVIII.

[0338] In some embodiments, the FVIII has an increased half-life ($t_{1/2}$) due to linkage to a heterologous moiety that is a half-life extending moiety. Examples include, e.g., FVIII fused to Fc (including, e.g., FVIII constructs in the form of a hybrid such as a FVIII-Fc monomer dimer hybrid; see e.g., US Patent Nos. 7,404,956 and 7,348,004), FVIII fused to albumin, FVIII fused to XTEN, FVIII fused to PAS, FVIII fused to HES, and FVIII fused to a water-soluble polymer, such as PEG.

[0339] The half-life is increased compared to a "reference FVIII" not fused to the half-life extending moiety (e.g., the FVIII without the Fc portion, or without the albumin portion). Likewise, the reference FVIII in the case of a modified FVIII is the same FVIII without the modification, e.g., a FVIII without the pegylation.)

[0340] In one example, the FVIII is fused to one or more albumin polypeptides (FVIII-albumin construct), e.g., human albumin. FVIII can be fused to either the N-terminal end of the albumin or to the C-terminal end of the albumin, provided the FVIII component of the FVIII-albumin fusion protein can be processed by an enzymatically-active proprotein convertase to yield a processed FVIII-containing polypeptide. Examples of albumin, e.g., fragments thereof, that may be used in the present invention are known. e.g., U.S. Patent No. 7,592,010; U.S. Patent No. 6,686,179; and Schulte, Thrombosis Res. 124 Suppl. 2:S6-S8 (2009), each of which is incorporated herein by reference in its entirety.

[0341] Functional FVIII variants are known, as is discussed herein. In addition, hundreds of nonfunctional mutations in FVIII have been identified in hemophilia patients, and it has been determined that the effect of these mutations on FVIII function is due more to where they lie within the 3-dimensional structure of FVIII than on the nature of the substitution (Cutler et al., Hum. Mutat. 19:274-8 (2002)), incorporated herein by reference in its entirety. In addition, comparisons between FVIII from humans and other species have identified conserved residues that are likely to be required for function (Cameron et al., Thromb. Haemost. 79:317-22 (1998); US 6,251,632), incorporated herein by reference in its entirety.

- [0342] The human FVIII gene was isolated and expressed in mammalian cells (Toole, J. J., et al., Nature 312:342-347 (1984); Gitschier, J., et al., Nature 312:326-330 (1984); Wood, W. I., et al., Nature 312:330-337 (1984); Vehar, G. A., et al., Nature 312:337-342 (1984); WO 87/04187; WO 88/08035; WO 88/03558; U.S. Pat. No. 4,757,006), each of which is incorporated herein by reference in its entirety, and the amino acid sequence was deduced from cDNA. Capon et al., U.S. Pat. No. 4,965,199, incorporated herein by reference in its entirety, disclose a recombinant DNA method for producing FVIII in mammalian host cells and purification of human FVIII. Human FVIII expression in CHO (Chinese hamster ovary) cells and BHKC (baby hamster kidney cells) has been reported. Human FVIII has been modified to delete part or all of the B domain (U.S. Pat. Nos. 4,994,371 and 4,868,112, each of which is incorporated herein by reference in its entirety), and replacement of the human FVIII B domain with the human factor V B domain has been performed (U.S. Pat. No. 5,004,803, incorporated herein by reference in its entirety). The cDNA sequence encoding human FVIII and predicted amino acid sequence are shown in SEQ ID NOs:1 and 2, respectively, of US Application Publ. No. 2005/0100990, incorporated herein by reference in its entirety.
- [0343] U.S. Pat. No. 5,859,204, Lollar, J. S., incorporated herein by reference in its entirety, reports functional mutants of FVIII having reduced antigenicity and reduced immunoreactivity. U.S. Pat. No. 6,376,463, Lollar, J. S., incorporated herein by reference in its entirety, also reports mutants of FVIII having reduced immunoreactivity. US Application Publ. No. 2005/0100990, Saenko et al., incorporated herein by reference in its entirety, reports functional mutations in the A2 domain of FVIII.
- [0344] The porcine FVIII sequence is published, (Toole, J. J., et al., Proc. Natl. Acad. Sci. USA 83:5939-5942 (1986)), incorporated herein by reference in its entirety, and the complete porcine cDNA sequence obtained from PCR amplification of FVIII sequences from a pig spleen cDNA library has been reported (Healey, J. F., et al., Blood 88:4209-4214 (1996), incorporated herein by reference in its entirety). Hybrid human/porcine FVIII having substitutions of all domains, all subunits, and specific amino acid sequences were disclosed in U.S. Pat. No. 5,364,771 by Lollar and Runge, and in WO 93/20093, incorporated herein by reference in its entirety. More recently, the nucleotide and corresponding amino acid sequences of the A1 and A2 domains of porcine FVIII and a chimeric FVIII with porcine A1 and/or A2 domains substituted for the corresponding

human domains were reported in WO 94/11503, incorporated herein by reference in its entirety. U.S. Pat. No. 5,859,204, Lollar, J. S., also discloses the porcine cDNA and deduced amino acid sequences. 6,458,563, incorporated herein by reference in its entirety assigned to Emory discloses a B-domain deleted porcine FVIII.

- [0345] In one example, the FVIII is linked to Fc. Such constructs are known in the art. Exemplary FVIII and FVIII-Fc polypeptides include, e.g., SEQ ID NOs: 700-711 or a portion thereof.
- [0346] The FVIII may be at least 85%, 90%, 95%, 96%, 97%, 98% or 99% identical to a portion of the FVIII amino acid sequence of SEQ ID NOs:701, 705, 707, 709, or 711 without a signal sequence (e.g., amino acids 20 to 1457 of SEQ ID NO:701; amino acids 20 to 2351 of SEQ ID NO:705; amino acids 20 to 759 of SEQ ID NO:707; amino acids 20 to 764 of SEQ ID NO:709; or amino acids 20 to 703 of SEQ ID NO:711).
- [0347] The FVIII (or FVIII portion of a conjugate) may be identical to a portion of the FVIII amino acid sequence of SEQ ID NOs:701, 705, 707, 709, or 711 without a signal sequence (e.g., amino acids 20 to 1457 of SEQ ID NO:701; amino acids 20 to 2351 of SEQ ID NO:705; amino acids 20 to 759 of SEQ ID NO:707; amino acids 20 to 764 of SEQ ID NO:709; or amino acids 20 to 703 of SEQ ID NO:711).
- [0348] The FVIII (or FVIII portion of a conjugate) may be at least 85%, 90%, 95%, 96%, 97%, 98% or 99% identical to a portion of the FVIII amino acid sequence of SEQ ID NOs:701, 705, 707, 709, or 711, with a signal sequence (e.g., amino acids 1 to 1457 of SEQ ID NO:701; amino acids 1 to 2351 of SEQ ID NO:705; amino acids 1 to 759 of SEQ ID NO:707; amino acids 1 to 764 of SEQ ID NO:709; or amino acids 1 to 703 of SEQ ID NO:711).
- [0349] The FVIII (or FVIII portion of a chimeric polypeptide) may be identical to a FVIII amino acid sequence of SEQ ID NOs:701, 705, 707, 709, or 711, with a signal sequence (e.g., amino acids 1 to 1457 of SEQ ID NO:701; amino acids 1 to 2351 of SEQ ID NO:705; amino acids 1 to 759 of SEQ ID NO:707; amino acids 1 to 764 of SEQ ID NO:709; or amino acids 1 to 703 of SEQ ID NO:711).
- [0350] In one example, the FVIII is linked to Fc. Such constructs are known in the art. The FVIII-Fc may comprise a sequence at least 90% or 95% identical to the FVIII-Fc amino acid sequences of SEQ ID NOs:701, 705, 707, 709, or 711 without a signal sequence (e.g., amino acids 20 to 1684 of SEQ ID NO:701) or at least 90% or 95%

identical to the FVIII and Fc amino acid sequence of SEQ ID NOs:701, 705, 707, 709, or 711 with a signal sequence (e.g., amino acids 1 to 1684 of SEQ ID NO:701).

[0351] The FVIII-Fc may comprise a sequence identical to the FVIII and Fc amino acid sequence of SEQ ID NOs:701, 705, 707, 709, or 711 without a signal sequence (e.g., amino acids 20 to 1684 of SEQ ID NO:701) or identical to the FVIII and Fc amino acid sequence of SEQ ID NOs:701, 705, 707, 709, or 711 with a signal sequence (e.g., amino acids 1 to 1684 of SEQ ID NO:701).

[0352] The polynucleotide variants may contain alterations in the coding regions, non-coding regions, or both. Especially preferred are polynucleotide variants containing alterations which produce silent substitutions, additions, or deletions, but do not alter the properties or activities of the encoded polypeptide. Nucleotide variants produced by silent substitutions due to the degeneracy of the genetic code are preferred. Moreover, variants in which 5-10, 1-5, or 1-2 amino acids are substituted, deleted, or added in any combination are also preferred. Polynucleotide variants can be produced for a variety of reasons, e.g., to optimize codon expression for a particular host (change codons in the human mRNA to those preferred by a bacterial host such as *E. coli*).

[0353] In some embodiments, FVIII is modified, e.g., pegylated, at any convenient location. In some embodiments, FVIII is pegylated at a surface exposed amino acid of FVIII, preferably a surface exposed cysteine, which may be an engineered cysteine. Mei et al. (2010). In some embodiments, modified FVIII, e.g., pegylated FVIII, is a long-acting FVIII.

[0354] In one example, the FVIII of the present disclosure is a polypeptide having FVIII-like activity, but does not have the amino acid sequence of FVIII. For example, the polypeptide having FVIII-like activity increases the catalytic activity of FIXa. In one example, the polypeptide having FVIII-like activity is an antibody (e.g., FIX/FIXa activating antibodies and antibody derivatives). Exemplary polypeptides having FVIII-like activity are disclosed in U.S. Pat. No. 7,033,590, which is incorporated herein by reference in its entirety.

[0355] Methods for the preparation of recombinant FVIII or FVIII-Fc constructs are disclosed, e.g., in WO2011/069164, which is incorporated herein by reference in its entirety.

FIX

- [0356] "Factor IX" or "FIX," as used herein, means functional Factor IX polypeptide in its normal role in coagulation, unless otherwise specified. Thus, the term Factor IX includes FIX variant polypeptides that are functional and the polynucleotides that encode such functional variant polypeptides. Preferred Factor IX polypeptides are the human, bovine, porcine, canine, feline, and murine Factor IX polypeptides. The full length polypeptide and polynucleotide sequences of Factor IX are known, as are many functional variants, e.g., fragments, mutants and modified versions. Factor IX polypeptides include full-length Factor IX, full-length Factor IX minus Met at the N-terminus, mature Factor IX (minus the signal sequence), and mature Factor IX with an additional Met at the N-terminus.
- [0357] The FIX may be at least 85%, 90%, 95%, 96%, 97%, 98% or 99% identical to a FIX amino acid sequence without a signal sequence (e.g., amino acids 47 to 461 of SEQ ID NO:713). The FIX may be identical to a FIX amino acid sequence without a signal sequence (e.g., amino acids 47 to 461 of SEQ ID NO:713).
- [0358] The FIX may be at least 85%, 90%, 95%, 96%, 97%, 98% or 99% identical to a FIX amino acid sequence with a signal sequence (e.g., amino acids 1 to 461 of SEQ ID NO:713). The FIX may be identical to a FIX amino acid sequence with a signal sequence (e.g., amino acids 1 to 461 of SEQ ID NO:713).
- [0359] In one example, FIX is linked to Fc. Exemplary FIX-Fc amino acid and DNA sequences include SEQ ID NOs:712 and 713, with or without its signal sequence.
- [0360] The FIX-Fc polypeptide may comprise a sequence at least 85%, 90%, 95%, 96%, 97%, 98% or 99% identical to the FIX and Fc amino acid sequences without a signal sequence (amino acids 47 to 688 of SEQ ID NO:713) or at least 85%, 90%, 95%, 96%, 97%, 98% or 99% identical to the Factor IX and Fc amino acid sequence with a signal sequence (amino acids 1 to 688 of SEQ ID NO:713).
- [0361] The FIX-Fc polypeptide may comprise a sequence identical to the Factor IX and Fc amino acid sequence without a signal sequence (amino acids 47 to 688 of SEQ ID NO:713) or identical to the Factor IX and Fc amino acid sequence with a signal sequence (amino acids 1 to 688 of SEQ ID NO:713).
- [0362] A great many functional FIX variants are known. International publication number WO 02/040544 A3, which is herein incorporated by reference in its entirety,

discloses mutants that exhibit increased resistance to inhibition by heparin at page 4, lines 9-30 and page 15, lines 6-31. International publication number WO 03/020764 A2, which is herein incorporated by reference in its entirety, discloses Factor IX mutants with reduced T cell immunogenicity in Tables 2 and 3 (on pages 14-24), and at page 12, lines 1-27. International publication number WO 2007/149406 A2, which is herein incorporated by reference in its entirety, discloses functional mutant Factor IX molecules that exhibit increased protein stability, increased in vivo and in vitro half life, and increased resistance to proteases at page 4, line 1 to page 19, line 11. WO 2007/149406 A2 also discloses chimeric and other variant Factor IX molecules at page 19, line 12 to page 20, line 9. International publication number WO 08/118507 A2, which is herein incorporated by reference in its entirety, discloses Factor IX mutants that exhibit increased clotting activity at page 5, line 14 to page 6, line 5. International publication number WO 09/051717 A2, which is herein incorporated by reference in its entirety, discloses Factor IX mutants having an increased number of N-linked and/or O-linked glycosylation sites, which results in an increased half life and/or recovery at page 9, line 11 to page 20, line 2. International publication number WO 09/137254 A2, which is herein incorporated by reference in its entirety, also discloses Factor IX mutants with increased numbers of glycosylation sites at page 2, paragraph [006] to page 5, paragraph [011] and page 16, paragraph [044] to page 24, paragraph [057]. International publication number WO 09/130198 A2, which is herein incorporated by reference in its entirety, discloses functional mutant Factor IX molecules that have an increased number of glycosylation sites, which result in an increased half life, at page 4, line 26 to page 12, line 6. International publication number WO 09/140015 A2, which is herein incorporated by reference in its entirety, discloses functional Factor IX mutants that an increased number of Cys residues, which may be used for polymer (e.g., PEG) conjugation, at page 11, paragraph [0043] to page 13, paragraph [0053].

[0363] In addition, hundreds of non-functional mutations in Factor IX have been identified in hemophilia patients, many of which are disclosed in Table 1, at pages 11-14 of International publication number WO 09/137254 A2, which is herein incorporated by reference in its entirety. Such non-functional mutations are not included in the invention, but provide additional guidance for which mutations are more or less likely to result in a functional Factor IX polypeptide.

[0364] In various embodiments FIX is fused to one or more XTEN polypeptides. Schellenburger et al., Nat. Biotech. 27:1186-90 (2009), which is incorporated herein by reference in its entirety. FIX can be fused to either the N-terminal end of the XTEN polypeptide or to the C-terminal end of the XTEN polypeptide, provided the FIX component of the FIX-XTEN construct can be processed by a protease to yield a processed FIX containing polypeptide. A protease site may be included between the XTEN portion and the FIX portion to allow such processing. XTEN polypeptides include, e.g., those disclosed in WO 2009/023270, WO 2010/091122, WO 2007/103515, US 2010/0189682, and US 2009/0092582, each of which is incorporated herein by reference in its entirety.

[0365] Variant FIX polynucleotides may comprise, or alternatively consist of, a nucleotide sequence which is at least 85%, 90%, 95%, 96%, 97%, 98% or 99% identical to, for example, the nucleotide coding sequence in SEQ ID NO:712 (the Factor IX portion, the Fc portion, individually or together) or the complementary strand thereto, the nucleotide coding sequence of known mutant and recombinant Factor IX or Fc such as those disclosed in the publications and patents cited herein or the complementary strand thereto, a nucleotide sequence encoding the polypeptide of SEQ ID NO:713 or SEQ ID NO:703 (the Factor IX portion, the Fc portion, individually or together), and/or polynucleotide fragments of any of these nucleic acid molecules (e.g., those fragments described herein). Polynucleotides which hybridize to these nucleic acid molecules under stringent hybridization conditions or lower stringency conditions are also included as variants, as are polypeptides encoded by these polynucleotides as long as they are functional.

[0366] Variant FIX polypeptides may comprise, or alternatively consist of, an amino acid sequence which is at least 85%, 90%, 95%, 96%, 97%, 98%, 99% identical to, for example, the polypeptide sequence shown in SEQ ID NO:713 or 703 (the Factor IX portion, the Fc portion, individually or together), and/or polypeptide fragments of any of these polypeptides (e.g., those fragments described herein).

FVIIa

[0367] The term "Factor VII" or "FVII", includes "Factor VIIa", or "FVIIa", and herein, means functional FVII protein or functional FVIIa protein in its normal role in coagulation, unless otherwise specified (i.e., refers to any FVII moiety which exhibits

biological activity that is associated with native FVII). Thus, the term FVII includes variant proteins that are functional. Preferred FVII proteins are primate (e.g., chimpanzee), human, porcine, canine, and murine FVII proteins. The full length polypeptide and polynucleotide sequences of FVII are known, as are many functional fragments, mutants and modified versions. Exemplary chimeric and hybrid FVII sequences are disclosed, e.g., in US 2009/0291890 A1, US2009/0041744 A1, and US 2008/0318276 A1. Factor VII polypeptides include, e.g., full-length FVII, full-length FVII minus Met at the N-terminus, mature FVII (minus the signal sequence), and mature FVII with an additional Met at the N-terminus. An exemplary sequence of FVII can be found as NCBI Accession Number NP000122.

[0368] As used herein, "plasma-derived FVII" includes all forms of the protein found in blood obtained from a mammal having the property of activating the coagulation pathway.

[0369] The term Factor VII includes variant polypeptides that are functional. Preferred factor VII proteins are the human, porcine, canine, and murine factor VII proteins. As described in the Background Art section, the full length polypeptide and polynucleotide sequences are known, as are many functional fragments, mutants and modified versions. The one chain zymogen Factor VII is a polypeptide comprising 406 amino acid residues, 10 of which are γ -carboxylated glutamic acid residues, N-glycosylated asparagines residues (no 145 and no 322), and O-glycosylated serine residues in position 52 and 60. The variant forms of FVII encompasses i.a. molecules wherein one or more amino acid residues have been substituted, added or deleted, molecules with different number of GLA residues, molecules with a modified or uncomplete glycosylation pattern. Non-limiting examples of modifications of amino acid residues are amidation, alkylation, acylation and PEGylation.

[0370] Examples of human FVII amino acid and DNA sequences are shown as subsequences in SEQ ID NO:714 and SEQ ID NO:715. FVII polypeptides include, e.g., full-length FVII, full-length FVII minus Met at the N-terminus, mature FVII (minus the signal sequence), and/or mature FVII with an additional Met at the N-terminus.

[0371] The term "activated Factor VII" or "FVIIa" refers to the enzymatically active two-chain form of circulating FVII generated as well as variants thereof in case coagulation activity (e.g. thrombin generation) is needed. The two-chain Factor VIIa is a polypeptide

produced from FVII by hydrolysis of the Arg152-Ile153 peptide bond of FVII. FVIIa also comprises 406 amino acid residues, 10 of which are γ -carboxylated glutamic acid residues, N-glycosylated asparagines residues (no 145 and no 322), and O-glycosylated serine residues in position 52 and 60. The variant forms of FVIIa encompasses i.a. molecules wherein one or more amino acid residues have been substituted, added or deleted, molecules with different number of GLA residues, molecules with a modified or uncomplete glycosylation pattern. The terms "FVII" and "activated FVII" include naturally occurring FVII and activated FVII but also encompass function conservative variants and modified forms thereof.

[0372] The term "Factor VIIa" or "FVIIa" includes activatable FVII.

[0373] Exemplary FVII sequences are disclosed herein. For example, the FVII may be at least 90% or 95% identical to a FVII amino acid sequence without a signal sequence (e.g., amino acids 61 to 466 of SEQ ID NO:715). The FVII may be identical to a Factor VII amino acid sequence without a signal sequence (e.g., amino acids 61 to 466 of SEQ ID NO:715).

[0374] The Factor VII may be at least 90% or 95% identical to a Factor VII amino acid sequence with a signal sequence (e.g., amino acids 1 to 466 of SEQ ID NO:715). The Factor VII may be identical to a FVII amino acid sequence with a signal sequence (e.g., amino acids 1 to 466 of SEQ ID NO:715).

[0375] In one example FVII is linked to Fc. Exemplary FVII-Fc amino acid and DNA sequences are represented by SEQ ID NO:714 and 715. The FVII-Fc may comprise a sequence at least 90% or 95% identical to the FVII and Fc amino acid sequence without a signal sequence (e.g., amino acids 61 to 693 of SEQ ID NO:715) or at least 90% or 95% identical to the FVII and Fc amino acid sequence with a signal sequence (e.g., amino acids 1 to 693 of SEQ ID NO:715).

[0376] The FVII-Fc may comprise a sequence identical to the FVII and Fc amino acid sequence without a signal sequence (e.g., amino acids 61 to 693 of SEQ ID NO:715) or identical to the FVII and Fc amino acid sequence with a signal sequence (e.g., amino acids 1 to 693 of SEQ ID NO:715).

[0377] FVII polynucleotides include, e.g., those of SEQ ID NO:714 and fragments thereof, e.g., those that encode the FVII fragment.

- [0378] The term “Factor VIIa derivative” or “FVIIa derivative” as used herein, is intended to designate a FVIIa polypeptide exhibiting substantially the same or improved biological activity relative to wild-type Factor VIIa, in which one or more of the amino acids of the parent peptide have been genetically and/or chemically and/or enzymatically modified, e.g. by alkylation, glycosylation, deglycosylation, PEGylation, acylation, ester formation or amide formation or the like. This includes but is not limited to PEGylated Factor VIIa, cysteine-PEGylated human Factor VIIa and variants thereof.
- [0379] The term “PEGylated Factor VIIa” (and the like) means a Factor VIIa polypeptide conjugated with a PEG molecule. It is to be understood, that the PEG molecule may be attached to any part of the Factor VIIa polypeptide including any amino acid residue or carbohydrate moiety of the Factor VIIa polypeptide. The term “cysteine-PEGylated Factor VIIa” means Factor VIIa polypeptide having a PEG molecule conjugated to a sulfhydryl group of a non-native cysteine of the Factor VIIa polypeptide.
- [0380] Non-limiting examples of Factor VIIa derivatives includes GlycoPegylated FVIIa derivatives as disclosed in WO 03/31464 and US Patent applications US 20040043446, US 20040063911, US 20040142856, US 20040137557, and US 20040132640 (Neose Technologies, Inc.); FVIIa conjugates as disclosed in WO 01/04287, US patent application 20030165996, WO 01/58935, WO 03/93465 (Maxygen ApS) and WO 02/02764, US patent application 20030211094 (University of Minnesota).
- [0381] The term “improved biological activity” refers to FVIIa polypeptides with i) substantially the same or increased proteolytic activity compared to recombinant wild type human Factor VIIa or ii) to FVIIa polypeptides with substantially the same or increased TF binding activity compared to recombinant wild type human Factor VIIa or iii) to FVIIa polypeptides with substantially the same or increased half life in blood plasma compared to recombinant wild type human Factor VIIa.
- [0382] The term “PEGylated human Factor VIIa” means human Factor VIIa, having a PEG molecule conjugated to a human Factor VIIa polypeptide. It is to be understood, that the PEG molecule may be attached to any part of the Factor VIIa polypeptide including any amino acid residue or carbohydrate moiety of the Factor VIIa polypeptide.
- [0383] Non-limiting examples of FVIIa variants having increased biological activity compared to wild-type FVIIa include FVIIa variants as disclosed in WO 01/83725, WO 02/22776, WO 02/077218, PCT/DK02/00635 (corresponding to WO 03/027147), Danish

patent application PA 2002 01423 (corresponding to WO 04/029090), Danish patent application PA 2001 01627 (corresponding to WO 03/027147); WO 02/38162 (Scripps Research Institute); and FVIIa variants with enhanced activity as disclosed in IP 2001061479 (Chemo-Sero-Therapeutic Res Inst.).

[0384] Variant polynucleotides may comprise, or alternatively consist of, a nucleotide sequence which is at least 85%, 90%, 95%, 96%, 97%, 98% or 99% identical to, for example, the nucleotide coding sequence in SEQ ID NO:15 (the factor VII portion, the Fc portion, individually or together) or the complementary strand thereto, the nucleotide coding sequence of known mutant and recombinant factor VII or Fc such as those disclosed in the publications and patents cited herein or the complementary strand thereto, a nucleotide sequence encoding the polypeptide of SEQ ID NO:16 or 4 (the factor VII portion, the Fc portion, individually or together), and/or polynucleotide fragments of any of these nucleic acid molecules (e.g., those fragments described herein). Polynucleotides which hybridize to these nucleic acid molecules under stringent hybridization conditions or lower stringency conditions are also included as variants, as are polypeptides encoded by these polynucleotides as long as they are functional.

[0385] Variant polypeptides may comprise, or alternatively consist of, an amino acid sequence which is at least 85%, 90%, 95%, 96%, 97%, 98%, 99% identical to, for example, the polypeptide sequence shown in SEQ ID NO:715 or 703 (the FVII portion, the Fc portion, individually or together), and/or polypeptide fragments of any of these polypeptides (e.g., those fragments described herein).

[0386] Using known methods of protein engineering and recombinant DNA technology, variants may be generated to improve or alter the characteristics of the polypeptides. For instance, one or more amino acids can be deleted from the N-terminus or C-terminus of the secreted protein without substantial loss of biological function. The authors of Ron et al., J. Biol. Chem. 268: 2984-2988 (1993), incorporated herein by reference in its entirety, reported variant KGF proteins having heparin binding activity even after deleting 3, 8, or 27 amino-terminal amino acid residues. Similarly, Interferon gamma exhibited up to ten times higher activity after deleting 8-10 amino acid residues from the carboxy terminus of this protein. (Dobeli et al., J. Biotechnology 7:199-216 (1988), incorporated herein by reference in its entirety.)

[0387] As stated above, polypeptide variants include, e.g., modified polypeptides. Modifications include, e.g., acetylation, acylation, ADP-ribosylation, amidation, covalent attachment of flavin, covalent attachment of a heme moiety, covalent attachment of a nucleotide or nucleotide derivative, covalent attachment of a lipid or lipid derivative, covalent attachment of phosphatidylinositol, cross-linking, cyclization, disulfide bond formation, demethylation, formation of covalent cross-links, formation of cysteine, formation of pyroglutamate, formylation, gamma-carboxylation, glycosylation, GPI anchor formation, hydroxylation, iodination, methylation, myristoylation, oxidation, pegylation (Mei et al., Blood 116:270-79 (2010), which is incorporated herein by reference in its entirety), proteolytic processing, phosphorylation, prenylation, racemization, selenoylation, sulfation, transfer-RNA mediated addition of amino acids to proteins such as arginylation, and ubiquitination. In some embodiments, FVII is modified, e.g., pegylated, at any convenient location. In some embodiments, FVII is pegylated at a surface exposed amino acid of Factor VII, preferably a surface exposed.

[0388] In one embodiment, an enhanced clotting factor of the invention is manufactured in an activated form. For example, FVII, is generally produced recombinantly as a zymogen, and requires activation during manufacturing to produce the active form for administration. In one embodiment, an enhanced clotting factor of the invention is secreted from the cell in which it is expressed in active form to improve manufacturability. Such clotting factors can be produced by expressing the light chain of a clotting factor and the heavy chain of a clotting factor separately. In one embodiment, such a polypeptide comprises an intracellular processing site upstream of the heavy chain. Activation of such a construct is delayed until late in the secretory pathway, e.g., when the protein colocalizes with active processing enzymes in the trans-Golgi apparatus. In one embodiment, such a construct comprises an Fc scaffold moiety, for example an scFc scaffold in which the scFc linker comprises one or more intracellular processing sites.

[0389] In another embodiment, an enhanced clotting factor of the invention is made in activatable form. For use in bypass therapy exogenous clotting factors are only efficacious when given in the activated form. However, such activated clotting factors are rapidly inactivated by endogenous pathways (e.g. antithrombin III, TFPI), leading to clearance of the active form and a short effective half life. Giving higher doses does

not solve this problem as it can result in thrombogenic effects. Thus, in one embodiment, the invention pertains to an "activatable" enhanced clotting factor constructs which circulate as zymogens. These molecules have a longer half life, but can readily be activated at the site of clotting by cleavage by an enzyme. In one embodiment, such an enzyme is one produced during the clotting cascade. For example, in one embodiment, the cleavage site of an activatable construct comprises a Factor XIa, Xa, or thrombin cleavage site. Exemplary FXIa cleavage sites include: TQSFNDFTR and SVSQTSKLTR. Exemplary Thrombin cleavage sites include: DFLAEGGGVR, TTKIKPR, and ALRPRVVGGA.

[0390] As discussed above, exemplary polypeptides include FVII fused to one or more XTEN polypeptides. Schellenburger et al., Nat. Biotech. 27:1186-90 (2009), which is incorporated herein by reference in its entirety. FVII can be fused to either the N-terminal end of the XTEN polypeptide or to the C-terminal end of the XTEN polypeptide, provided the Factor VII component of the Factor VII-XTEN fusion protein can be processed by a protease to yield a processed Factor VII containing polypeptide. A protease site may be included between the XTEN portion and the Factor VII portion to allow such processing. XTEN polypeptides include, e.g., those disclosed in WO 2009/023270, WO 2010/091122, WO 2007/103515, US 2010/0189682, and US 2009/0092582, each of which is incorporated herein by reference in its entirety.

[0391] Exemplary polypeptides also include FVII fused to one or more albumin polypeptides. Preferably the albumin is human albumin. Factor VII can be fused to either the N-terminal end of the albumin or to the C-terminal end of the albumin, provided the Factor VII component of the Factor VII-albumin fusion protein can be processed by an enzymatically-active proprotein convertase to yield a processed Factor VII-containing polypeptide. Examples of albumin, e.g., fragments thereof, that may be used in the present invention are known. e.g., U.S. Patent No. 7,592,010; U.S. Patent No. 6,686,179; and Schulte, Thrombosis Res. 124 Suppl. 2:S6-S8 (2009), each of which is incorporated herein by reference in its entirety.

[0392] In some embodiments, a chimeric polypeptide comprising a Factor VII portion has an increased half-life ($t_{1/2}$) over a polypeptide consisting of the same Factor VII portion without the non Factor VII portion. A chimeric Factor VII polypeptide with an increased $t_{1/2}$ may be referred to herein as a long-acting Factor VII. Long-acting chimeric Factor

VII polypeptides include, e.g., Factor VII fused to Fc (including, e.g., chimeric Factor VII polypeptides in the form of a hybrid such as a FVIIFc monomer dimer hybrid; see Table 1), Factor VII fused to XTEN, and Factor VII fused to albumin.

Platelet Targeting Moiety

[0393] In one embodiment, the platelet targeting moiety comprises at least one of an antigen binding site (e.g., an antigen binding site of an antibody, antibody variant, or antibody fragment), a polypeptide, a receptor binding portion of ligand, or a ligand binding portion of a receptor which specifically binds to platelets, e.g., resting or activated platelets. Exemplary targeting moieties include scFv molecules or peptides which bind to molecules to be targeted. In one embodiment, the targeting moiety binds to resting platelets. In one embodiment, the targeting moiety selectively binds to activated platelets.

[0394] In one embodiment, the targeting moiety selectively binds to a target selected from the group consisting of: GPIIb, GPVI, and the nonactive form of GPIIb/IIIa. In another embodiment, the targeting moiety selectively binds to a target selected from the group consisting of: GPIIb/IIIa, P selectin, GMP-33, LAMP-1, LAMP-2, CD40L, and LOX-1. Examples of platelet targeting moieties are described below.

Antigen Binding Sites

[0395] In certain embodiments, the platelet targeting moiety is an antigen binding portion (e.g., binding site) of an antibody. In one embodiment, the antigen binding portion targets the composition to platelets

[0396] In other embodiments, a binding site of a polypeptide of the invention may comprise an antigen binding fragment. The term “antigen-binding portion” refers to a polypeptide fragment of an immunoglobulin, antibody, or antibody variant which binds antigen or competes with intact antibody (i.e., with the intact antibody from which they were derived) for antigen binding (i.e., specific binding). For example, said antigen binding fragments can be derived from any of the antibodies or antibody variants described *supra*. Antigen binding portions can be produced by recombinant or biochemical methods that are well known in the art. Exemplary antigen-binding fragments include Fv, Fab, Fab', and (Fab')₂ as well as scFv molecules.

[0397] In other embodiments, the targeting moiety is a binding site from a single chain binding molecule (*e.g.*, a single chain variable region or scFv). Techniques described for the production of single chain antibodies (U.S. Pat. No. 4,694,778; Bird, *Science* 242:423-442 (1988); Huston *et al.*, *Proc. Natl. Acad. Sci. USA* 85:5879-5883 (1988); and Ward *et al.*, *Nature* 334:544-554 (1989)) can be adapted to produce single chain binding molecules. Single chain antibodies are formed by linking the heavy and light chain fragments of the Fv region via an amino acid bridge, resulting in a single chain antibody. Techniques for the assembly of functional Fv fragments in *E coli* may also be used (Skerra *et al.*, *Science* 242:1038-1041 (1988)).

[0398] In certain embodiments, the platelet targeting moiety includes one or more binding sites or regions comprising or consisting of a single chain variable region sequence (scFv). Single chain variable region sequences comprise a single polypeptide having one or more antigen binding sites, *e.g.*, a V_L domain linked by a flexible linker to a V_H domain. The V_L and/or V_H domains may be derived from any of the antibodies or antibody variants described *supra*. ScFv molecules can be constructed in a V_H-linker-V_L orientation or V_L-linker-V_H orientation. The flexible linker that links the V_L and V_H domains that make up the antigen binding site preferably comprises from about 10 to about 50 amino acid residues. In one embodiment, the polypeptide linker is a gly-ser polypeptide linker. An exemplary gly/ser polypeptide linker is of the formula (Gly4Ser)_n, wherein *n* is a positive integer (*e.g.*, 1, 2, 3, 4, 5, or 6). Other polypeptide linkers are known in the art. Antibodies having single chain variable region sequences (*e.g.* single chain Fv antibodies) and methods of making said single chain antibodies are well-known in the art (see *e.g.*, Ho *et al.* 1989. *Gene* 77:51; Bird *et al.* 1988 *Science* 242:423; Pantoliano *et al.* 1991. *Biochemistry* 30:10117; Milenic *et al.* 1991. *Cancer Research* 51:6363; Takkinen *et al.* 1991. *Protein Engineering* 4:837).

[0399] In certain embodiments, a scFv molecule is a stabilized scFv molecule. In one embodiment, the stabilized cFv molecule may comprise a scFv linker interposed between a V_H domain and a V_L domain, wherein the V_H and V_L domains are linked by a disulfide bond between an amino acid in the V_H and an amino acid in the V_L domain. In other embodiments, the stabilized scFv molecule may comprise a scFv linker having an optimized length or composition. In yet other embodiments, the stabilized scFv molecule may comprise a V_H or V_L domain having at least one stabilizing amino acid

substitution(s). In yet another embodiment, a stabilized scFv molecule may have at least two of the above listed stabilizing features.

[0400] Stabilized scFv molecules have improved protein stability or impart improved protein stability to the polypeptide to which it is operably linked. Preferred scFv linkers of the invention improve the thermal stability of a polypeptide of the invention by at least about 2°C or 3°C as compared to a conventional polypeptide. Comparisons can be made, for example, between the scFv molecules of the invention. In certain preferred embodiments, the stabilized scFv molecule comprises a (Gly₄Ser)₄ scFv linker and a disulfide bond which links V_H amino acid 44 and V_L amino acid 100. Other exemplary stabilized scFv molecules which may be employed in the polypeptides of the invention are described in US Patent Application No. 11/725,970, filed on March 19, 2007, incorporated herein by reference in its entirety.

[0401] In one example, the platelet targeting moiety includes a variable region or portion thereof (*e.g.* a VL and/or VH domain) derived from an antibody using art recognized protocols. For example, the variable domain may be derived from antibody produced in a non-human mammal, *e.g.*, murine, guinea pig, primate, rabbit or rat, by immunizing the mammal with the antigen or a fragment thereof. See Harlow & Lane, *supra*, incorporated by reference for all purposes. The immunoglobulin may be generated by multiple subcutaneous or intraperitoneal injections of the relevant antigen (*e.g.*, purified tumor associated antigens or cells or cellular extracts comprising such antigens) and an adjuvant. This immunization typically elicits an immune response that comprises production of antigen-reactive antibodies from activated splenocytes or lymphocytes.

[0402] Optionally, antibodies may be screened for binding to platelets of a specific activation state or to a specific region or desired fragment of the antigen without binding to other nonoverlapping fragments of the antigen. The latter screening can be accomplished by determining binding of an antibody to a collection of deletion mutants of the antigen and determining which deletion mutants bind to the antibody. Binding can be assessed, for example, by Western blot or ELISA. The smallest fragment to show specific binding to the antibody defines the epitope of the antibody. Alternatively, epitope specificity can be determined by a competition assay in which a test and reference antibody compete for binding to the antigen. If the test and reference antibodies compete,

then they bind to the same epitope or epitopes sufficiently proximal such that binding of one antibody interferes with binding of the other.

[0403] In other embodiments, the binding site is derived from a fully human antibody. Human or substantially human antibodies may be generated in transgenic animals (e.g., mice) that are incapable of endogenous immunoglobulin production (see e.g., U.S. Pat. Nos. 6,075,181, 5,939,598, 5,591,669 and 5,589,369, each of which is incorporated herein by reference). Another means of generating human antibodies using SCID mice is disclosed in U.S. Pat. No. 5,811,524 which is incorporated herein by reference. It will be appreciated that the genetic material associated with these human antibodies may also be isolated and manipulated as described herein.

[0404] Yet another highly efficient means for generating recombinant antibodies is disclosed by Newman, *Biotechnology*, 10: 1455-1460 (1992). Specifically, this technique results in the generation of primatized antibodies that contain monkey variable domains and human constant sequences. This reference is incorporated by reference in its entirety herein. Moreover, this technique is also described in commonly assigned U.S. Pat. Nos. 5,658,570, 5,693,780 and 5,756,096 each of which is incorporated herein by reference.

[0405] In another embodiment, a variable region domain of an altered antibody of the invention consists of a V_H domain, e.g., derived from camelids, which is stable in the absence of a V_L chain (Hamers-Casterman *et al.* (1993). *Nature*, 363:446; Desmyter *et al.* (1996). *Nat. Struct. Biol.* 3: 803; Decanniere *et al.* (1999). *Structure*, 7:361; Davies *et al.* (1996). *Protein Eng.*, 9:531; Kortt *et al.* (1995). *J. Protein Chem.*, 14:167).

[0406] Further, the platelet targeting moiety may comprise a variable domain or CDR derived from a fully murine, fully human, chimeric, humanized, non-human primate or primatized antibody. Non-human antibodies, or fragments or domains thereof, can be altered to reduce their immunogenicity using art recognized techniques.

[0407] In one embodiment, the variable domains are altered by at least partial replacement of one or more CDRs. In another embodiment, variable domains can optionally be altered, e.g., by partial framework region replacement and sequence changing. In making a humanized variable region the CDRs may be derived from an antibody of the same class or even subclass as the antibody from which the framework regions are derived, however, it is envisaged that the CDRs will be derived from an antibody of different class and preferably from an antibody from a different species. It

may not be necessary to replace all of the CDRs with the complete CDRs from the donor variable region to transfer the antigen binding capacity of one variable domain to another. Rather, it may only be necessary to transfer those residues that are necessary to maintain the activity of the binding domain. Given the explanations set forth in U. S. Pat. Nos. 5,585,089, 5,693,761 and 5,693,762, it will be well within the competence of those skilled in the art, either by carrying out routine experimentation or by trial and error testing to obtain a functional antigen binding site with reduced immunogenicity.

[0408] In other aspects, the polypeptides of the invention may comprise antigen binding sites, or portions thereof, derived from modified forms of antibodies. Exemplary such forms include, e.g., minibodies, diabodies, triabodies, nanobodies, camelids, Dabs, tetravalent antibodies, intradiabodies (e.g., Jendreyko et al. 2003. J. Biol. Chem. 278:47813), fusion proteins (e.g., antibody cytokine fusion proteins, proteins fused to at least a portion of an Fc receptor), and bispecific antibodies. Other modified antibodies are described, for example in U.S. Pat. No. 4,745,055; EP 256,654; Faulkner et al., Nature 298:286 (1982); EP 120,694; EP 125,023; Morrison, J. Immun. 123:793 (1979); Kohler et al., Proc. Natl. Acad. Sci. USA 77:2197 (1980); Raso et al., Cancer Res. 41:2073 (1981); Morrison et al., Ann. Rev. Immunol. 2:239 (1984); Morrison, Science 229:1202 (1985); Morrison et al., Proc. Natl. Acad. Sci. USA 81:6851 (1984); EP 255,694; EP 266,663; and WO 88/03559. Reassorted immunoglobulin chains also are known. See, for example, U.S. Pat. No. 4,444,878; WO 88/03565; and EP 68,763 and references cited therein.

[0409] In another embodiment, the platelet targeting moiety includes an antigen binding site or region which is a diabody or an antigen binding site derived therefrom. Diabodies are dimeric, tetravalent molecules each having a polypeptide similar to scFv molecules, but usually having a short (e.g., less than 10 and preferably 1-5) amino acid residue linker connecting both variable domains, such that the V_L and V_H domains on the same polypeptide chain cannot interact. Instead, the V_L and V_H domain of one polypeptide chain interact with the V_H and V_L domain (respectively) on a second polypeptide chain (see, for example, WO 02/02781). In one embodiment, an immature polypeptide of the invention comprises a diabody which is operably linked to the N-terminus and/or C-terminus of at least one genetically-fused Fc region (i.e., scFc region).

[0410] Exemplary single-domain antibodies employed in the binding molecules of the invention include, for example, the Camelid heavy chain variable domain (about 118 to

136 amino acid residues) as described in Hamers-Casterman, et al., Nature 363:446-448 (1993), and Dumoulin, et al., Protein Science 11:500-515 (2002). Other exemplary single domain antibodies include single VH or VL domains, also known as Dabs® (Domantis Ltd., Cambridge, UK). Yet other single domain antibodies include shark antibodies (e.g., shark Ig-NARs). Shark Ig-NARs comprise a homodimer of one variable domain (V-NAR) and five C-like constant domains (C-NAR), wherein diversity is concentrated in an elongated CDR3 region varying from 5 to 23 residues in length. In camelid species (e.g., llamas), the heavy chain variable region, referred to as VHH, forms the entire antigen-binding domain. Methods for making single domain binding molecules are described in US Patent Nos 6,005,079 and 6,765,087, both of which are incorporated herein by reference. Exemplary single domain antibodies comprising VHH domains include Nanobodies® (Ablynx NV, Ghent, Belgium).

- [0411] Exemplary antibodies from which binding sites can be derived for use in the binding molecules of the invention are known in the art. Antibodies known to bind to platelets can be used to derive binding sites, for example, the MB9 antibody described in US 2007/0218067 or the variable region or an scFv molecule comprising the variable region can be used as a targeting moiety in a construct of the invention.

Non-Immunoglobulin Platelet Binding Molecules

- [0412] In certain other embodiments, the targeting moiety comprise one or more binding sites derived from a non-immunoglobulin binding molecule. As used herein, the term “non-immunoglobulin binding molecules” are binding molecules whose binding sites comprise a portion (e.g., a scaffold or framework) which is derived from a polypeptide other than an immunoglobulin, but which may be engineered (e.g., mutagenized) to confer a desired binding specificity.
- [0413] Other examples of binding molecules comprising binding sites not derived from antibody molecules include receptor binding sites and ligand binding sites which bind to platelets.
- [0414] Non-immunoglobulin binding molecules may be identified by selection or isolation of a target-binding variant from a library of binding molecules having artificially diversified binding sites. Diversified libraries can be generated using completely random approaches (e.g., error-prone PCR, exon shuffling, or directed evolution) or aided by art-recognized design strategies. For example, amino acid positions that are usually involved

when the binding site interacts with its cognate target molecule can be randomized by insertion of degenerate codons, trinucleotides, random peptides, or entire loops at corresponding positions within the nucleic acid which encodes the binding site (see *e.g.*, U.S. Pub. No. 20040132028). The location of the amino acid positions can be identified by investigation of the crystal structure of the binding site in complex with the target molecule. Candidate positions for randomization include loops, flat surfaces, helices, and binding cavities of the binding site. In certain embodiments, amino acids within the binding site that are likely candidates for diversification can be identified using techniques known in the art. Following randomization, the diversified library may then be subjected to a selection or screening procedure to obtain binding molecules with the desired binding characteristics, *e.g.* specific binding platelets using methods known in the art. Selection can be achieved by art-recognized methods such as phage display, yeast display, or ribosome display. In one embodiment, molecules known in the art to bind to platelets may be employed in the constructs of the invention. For example, peptides which bind to GPIIb as described in the art (*e.g.*, PS4, 0S1, or 0S2) may be used (Benard *et al.* 2008. *Biochemistry* 47:4674-4682).

[0415] In one example, the targeting moieties is linked to an Fc moiety.

Biological Activity

[0416] In various embodiments, the compounds and conjugates of the present disclosure have pro-coagulant activity. It will be appreciated that different assays are available to measure pro-coagulant activity. In one example, the conjugate has pro-coagulant activity when it shows activity in at least one of: a Fxa generation assay, a thrombin generation assay (TGA), and a rotational thromboelastometry (ROTEM) assay, which are described herein, *e.g.*, in Examples 2, 3, and 4, respectively.

[0417] A compound or conjugate of the present disclosure may promote coagulation in plasma depleted of FV, FVII, FVIII, FIX, FX, FXI, or vWF. In one example, the compound or conjugate of the present disclosure promotes thrombin generation and/or fibrin deposition in plasma in which FVIII is depleted or absent. This type of activity is referred to as coagulation FVIII activity. Where the plasma is from an individual lacking FVIII or having reduced levels of FVIII, the activity is typically referred to as FVIII equivalent activity. Where the plasma contains inhibitors against FVIII, the activity is

typically referred to as FVIII inhibitor bypassing equivalent activity. Other pro-coagulant activities include FV activity, FVII activity, FX activity and FXI activity.

[0418] Individual compounds and conjugates can vary in their relative efficacy between different types of assay. Therefore, even if a compound or conjugate appears to have a low efficacy in a particular assay, it may nevertheless possess a suitably high level of pro-coagulant activity in another assay.

[0419] Other suitable assays useful to determine pro-coagulant activity include those disclosed, e.g., in Patent Application Publication U.S. 2010/0022445 to Scheifflinger and Dockal, which is incorporated herein by reference in its entirety.

[0420] In one example according to any of the above embodiments, the compound of the present disclosure has an EC_{50} of about 20 μ M or less, e.g., of about 10 μ M or less, or about 5 μ M or less in a Factor Xa (FXa) generation assay measuring conversion of Factor X (FX) to FXa. In one example, the compound has an EC_{50} in the FXa generation assay of about 4 μ M or less, of about 3 μ M or less, about 2 μ M or less, or about 1 μ M or less. In another example, the compound has an EC_{50} in the FXa generation assay of about 900 nM or less, about 800 nM or less, about 700 nM or less, about 600 nM or less, about 500 nM or less, about 400 nM or less, about 300 nM or less, or of about 200 nM or less. An exemplary FXa generation assay useful to determine the EC_{50} of a compound of the present disclosure is described in Example 2 of this application.

[0421] In another example, the compound of the present disclosure increases the catalytic activity (e.g., increases the k_{cat}) of a blood coagulation factor, such as FIXa or FVIIa, e.g., when compared to a reference catalytic activity measured in the absence of the compound, e.g., by at least 2-fold. In one example according to any of the above embodiments, the compound of the present disclosure increases the catalytic activity (e.g., increases the k_{cat}) of a blood coagulation factor (e.g., FIXa or FIVIIa) *in vitro* (e.g., in a suitable *in vitro* assay system, such as a FXa generation assay), e.g., by at least 2-fold. In another example, the compound of the present disclosure increases the catalytic activity (e.g., increases the k_{cat}) of a blood coagulation factor (e.g., FIXa or FVIIa) *in vivo*, e.g., by at least 2-fold.

[0422] In various embodiments, compounds of the present disclosure lower the K_M and increase the k_{cat} of hFIXa or hFVIIa. In one example according to any of the above embodiments, the compound of the present disclosure increases the catalytic activity of

FIXa or FVIIa by at least 2-fold at a compound concentration from about 1 nM to about 1 mM, from about 10 nM to about 500 μ M, from about 50 nM to about 100 μ M, from about 100 nM to about 50 μ M, from about 200 nM to about 40 μ M, from about 300 nM to about 30 μ M, or from about 400 nM to about 20 μ M. In another example, the compound of the present disclosure increases the catalytic activity of the FIXa or FVIIa by at least 2-fold at a concentration from about 500 nM to about 20 μ M, or from about 1 μ M to about 20 μ M, or from about 2 μ M to about 20 μ M, or from about 4 μ M to about 20 μ M, or from about 5 μ M to about 10 μ M. In another example, the compound of the present disclosure increases the catalytic activity of the FIXa or FVIIa by at least 2-fold at a concentration of 100 μ M or less, e.g., 50 μ M or less, 40 μ M or less, 30 μ M or less, 20 μ M or less, 15 μ M or less, 10 μ M or less, or 5 μ M or less.

[0423] In one example according to any of the above embodiments, the compound is used *in vitro* and is present in the assay mixture at a concentration of from about 0.1 μ M to about 100 μ M, from about 1 μ M to about 100 μ M, from about 1 μ M to about 50 μ M, from about 1 μ M to about 20 μ M, from 1 μ M to about 10 μ M, from 5 μ M to about 20 μ M, or from about 5 μ M to about 10 μ M.

[0424] In one example according to any of the above embodiments, the compound of the present disclosure increases the catalytic activity (e.g., increases the k_{cat}) of FIXa (e.g., at a concentration of about 5 μ M or less, or at a concentration of about 1 μ M or less) measured for the conversion of FX to FXa (e.g., in a suitable FXa generation assay) when compared to a reference catalytic activity (e.g., reference k_{cat}) of the FIXa measured in the absence of the compound.

[0425] In another example according to any of the above embodiments, the compound of the present disclosure increases the catalytic activity (e.g., increases the k_{cat}) of FVIIa (e.g., at a concentration of about 5 μ M or less, or at a concentration of about 1 μ M or less) measured for the conversion of FX to FXa (e.g., in a suitable FXa generation assay) when compared to a reference catalytic activity (e.g., reference k_{cat}) of the FVIIa measured in the absence of the compound.

[0426] In other embodiments, the compound of the present disclosure increases the catalytic activity (e.g., increases the k_{cat}) of a blood coagulation factor selected from FXa (e.g., for the conversion of pro-thrombin to thrombin), and thrombin (e.g., for a

conversion selected from: FVIII to FVIIIa, fibrinogen to fibrin, FV to FVa, protein C to active protein C, FXI to FXIa, and FXIII to FXIIIa).

[0427] In another example, the compound of the present disclosure increases the catalytic activity of at least one blood coagulation factor selected from FIXa, FXa, FVIIa and thrombin. In another example, the compound of the present disclosure increases the catalytic activity of at least one blood coagulation factor selected from FIXa and FVIIa. In yet another example according to any of the above embodiments, the compound of the present disclosure increases the catalytic activity of at least two different blood coagulation factors selected from FIXa, FXa, FVIIa, and thrombin. In yet another example according to any of the above embodiments, the compound of the present disclosure increases the catalytic activity of FIXa as well as FVIIa.

[0428] In other embodiments, the compound of the present disclosure increases the catalytic activity of FIXa and/or FVIIa, e.g., as measured using a FXa generation assay, but does not substantially increase the catalytic activity of FXa (e.g., as measured using a thrombin generation assay). For example, the inventors have discovered that compound 5 does not significantly affect the activity of FXa (e.g., towards a chromogenic or macromolecular substrate when using a thrombin generation assay, e.g., a TGA with purified components, e.g., as described in Example 3). The inventors have further discovered that compound 5 does not significantly increase the activity of the prothrombinase complex (FXa/FVa) when using a thrombin generation assay, e.g., a TGA with purified components, e.g., as described in Example 3b).

[0429] In other embodiments, the compound of the present disclosure increases the catalytic activity of FIXa and/or FVIIa, e.g., as measured using a FXa generation assay, but does not substantially increase the catalytic activities of FXa (e.g., as measured using a thrombin generation assay), and does not significantly increase the catalytic activity of thrombin (e.g., as measured using a fibrinogen cleavage assay). For example, the inventors have discovered that compound 5 does not substantially affect the amidolytic activity of alpha-thrombin (*see, e.g.,* Example 12).

[0430] In other examples according to any of the above embodiments, the compound of the present disclosure increases the catalytic activity (k_{cat}) of a blood coagulation factor (e.g., FIXa or FVIIa) by at least about 5 fold, at least about 10 fold, at least about 20 fold, at least about 30 fold, at least about 40 fold, at least about 50 fold, at least about 60 fold,

at least about 70 fold, at least about 80 fold, at least about 90 fold, or at least about 100 fold. In another example, the compound increases the catalytic activity (k_{cat}) of the blood coagulation factor (e.g., FIXa or FVIIa) by at least about 120 fold, at least about 140 fold, at least about 160 fold, at least about 180 fold, or at least about 200 fold. In another example, compound increases the catalytic activity (k_{cat}) of the blood coagulation factor (e.g., FIXa or FVIIa) by at least about 250 fold, at least about 300 fold, at least about 350 fold, at least about 400 fold, at least about 450 fold, or at least about 500 fold.

[0431] In one example according to any of the above embodiments, the compound is present at a concentration sufficient to cause the specified increase in catalytic activity. In one example according to any of the above embodiments, the compound is present at a concentration of from about 0.1 μM to about 100 μM , from about 1 μM to about 100 μM , from about 1 μM to about 50 μM , from about 1 μM to about 20 μM , from 1 μM to about 10 μM , from 5 μM to about 20 μM , or from about 5 μM to about 10 μM to cause any of the above specified increase in catalytic activity. In another example according to any of the above embodiments, the compound is present at a concentration below about 100 nM to to cause any of the above specified increase in catalytic activity.

[0432] Exemplary FXa generation assays useful to measure the catalytic activities of either FIXa or FVIIa in the presence or absence of a compound of the present disclosure are described in Example 2 of this application.

[0433] In another example according to any of the above embodiments, the compound of the present disclosure is capable of reducing clotting time in a suitable coagulation assay, e.g., as measured in an activated partial thromboplastin time (aPTT) assay, a modified activated partial thromboplastin time (aPTT*) assay or a rotational thromboelastometry (ROTEM) assay when compared to a reference clotting time measured in the absence of the compound.

[0434] In one example according to any of the above embodiments, the compound is present at a concentration sufficient to reduce the clotting time (e.g., by at least 10% when compared to the reference clotting time). In one example according to any of the above embodiments, the compound reduces clotting time by at least 10% at a concentration of from about 0.1 μM to about 100 μM , from about 1 μM to about 100 μM , from about 1 μM to about 50 μM , from about 1 μM to about 20 μM , from 1 μM to about 10 μM , from 5 μM to about 20 μM , or from about 5 μM to about 10 μM . In another example

according to any of the above embodiments, the compound of the present disclosure reduces clotting time by at least 10% at a concentration from about 1 nM to about 1 mM, from about 10 nM to about 500 μ M, from about 50 nM to about 100 μ M, from about 100 nM to about 50 μ M, from about 200 nM to about 40 μ M, from about 300 nM to about 30 μ M, or from about 400 nM to about 20 μ M. In another example, the compound of the present disclosure reduces the clotting time by at least 10% at a concentration from about 500 nM to about 20 μ M, or from about 1 μ M to about 20 μ M, or from about 2 μ M to about 20 μ M, or from about 4 μ M to about 20 μ M, or from about 5 μ M to about 10 μ M. In another example, the compound of the present disclosure reduces clotting time by at least 10% at a concentration of 100 μ M or less, e.g., 50 μ M or less, 40 μ M or less, 30 μ M or less, 20 μ M or less, 15 μ M or less, 10 μ M or less, or 5 μ M or less. In one example according to any of the above embodiments, the compound is tested *in vitro* and is present in the assay mixture at a concentration of from about 0.1 μ M to about 100 μ M, from about 1 μ M to about 100 μ M, from about 1 μ M to about 50 μ M, from about 1 μ M to about 20 μ M, from 1 μ M to about 10 μ M, from 5 μ M to about 20 μ M, or from about 5 μ M to about 10 μ M.

[0435] For example, in a suitable assay, a 10% reduction of a 10-minute reference clotting time (measured in the absence of the compound), means that clotting occurs after 9 minutes in the presence of the compound. Exemplary assays useful to measure clotting time are known in the art. Suitable assays are described in Example 9 (aPPT assay) and Example 4 (ROTEM assay).

[0436] In one example according to any of the above embodiments, the compound reduces the clotting time by more than 10%, e.g., by at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, or at least about 90% compared to a reference clotting time measured in the absence of the compound.

[0437] In one embodiment the clotting time measured for a compound of the present disclosure (e.g., in a ROTEM assay) is calcium-dependent. This calcium-dependency of the clotting time is comparable to the calcium-dependency measured for FVIII. The procoagulant activities of compounds of the present disclosure were examined for calcium dependence using a Rotem assay, e.g., as described in Example 4. For example, compound 5 at 2.5 and 5 μ M and 0.1 IU/mL of FVIII were tested at 7 different calcium

concentrations ranging from 0 to about 16 mM calcium. Similar to the FVIII control, the procoagulant effect observed for compound 5 was sensitive to the calcium concentration (i.e., the clotting time measured for a particular compound concentration was higher at elevated calcium concentrations).

[0438] In another example according to any of the above embodiments, the compound of the present disclosure reduces clotting time or increases the α -angle (e.g., in a dose-dependent manner) in a ROTEM assay using blood coagulation factor-deficient plasma (e.g., human or canine coagulation factor-deficient plasma) when compared to a reference clotting time or a reference α -angle measured in the absence of the compound.

[0439] In one example, the compound of the present disclosure reduces clotting time (e.g., in a dose-dependent manner) in a ROTEM assay involving FVIII-deficient plasma (e.g., human or canine FVIII deficient plasma). An exemplary ROTEM assay useful to measure the clotting time and the α -angle is described in Example 4 of this application.

[0440] In another example, the compound of the present disclosure reduces the clotting time (e.g., in a dose-dependent manner) or increases the α -angle in a ROTEM assay involving FIX-deficient plasma (e.g., human or canine FIX-deficient plasma). The inventors have discovered that the compounds of the present disclosure reduce clotting time in FIX-deficient plasma even in the presence of anti-FIX antibodies (e.g., anti-FIX pAb). The addition of anti-FIX antibodies to FIX-deficient plasma removes residual FIX activity from the plasma sample. These results indicate that the compounds of the present disclosure can reduce clotting time or increase the α -angle employing a mechanism other than by increasing the catalytic activity of FIXa (e.g., via activation of the extrinsic pathway of the blood coagulation cascade).

[0441] In one example, the compounds of the present disclosure reduce clotting time or increase α -angle by increasing the catalytic activity of FVIIa. For example, the inventors have discovered that the clotting time measured in the presence of a current compound in a ROTEM assay employing FVIII-deficient plasma is significantly increased (i.e., clot formation is significantly reduced) in the presence of an anti-FVII antibody (e.g., anti-FVII pAb). Likewise the α -angle measured in the presence of a current compound is significantly reduced when anti-FVII antibody is added to the assay mixture. These results indicate that the current compounds can induce clotting by modulating FVIIa catalytic activity (e.g., in addition to increasing the catalytic activity of FIXa).

Additive Effect with FVIII

- [0442] In another example according to any of the above embodiments, the compound of the present disclosure does not compete with FVIII, but instead shows at least additive (or even synergistic) activity with FVIII in at least one suitable assay. Exemplary assays are selected from thrombin generation assays (e.g., those described herein) and ROTEM assays (e.g., those described herein). In one example, the compound shows additive activity with FVIII (i.e., the compound's activity is not inhibited or reduced by the presence of FVIII; or the activity of FVIII is not inhibited by the presence of the compound) when the FVIII is present at a low concentration (e.g., at physiological concentration). In one example, the FVIII is present at a concentration from about 0.005 U/mL to about 1.0 U/mL, or from about 0.01 U/mL to about 0.5 U/mL, or from about 0.01 U/mL to about 0.3 U/mL, or from about 0.01 U/mL to about 0.2 U/mL or from about 0.04 U/mL to about 0.2 U/mL).
- [0443] In one example, the compound shows additive activity with FVIII in a thrombin generation assay when FVIII is present at a concentration of about 0.01 U/mL to about 0.5 U/mL, or from about 0.05 U/mL to about 0.2 U/mL. In another example, the compound shows additive activity with FVIII in a ROTEM assay when FVIII is present at a concentration of about 0.01 U/mL to about 0.5 U/mL, or from about 0.05 U/mL to about 0.1 U/mL.
- [0444] In one example according to any of the above embodiments, the compound is present at a concentration sufficient to have additive activity with FVIII.
- [0445] In another example according to any of the above embodiments, the compound of the present disclosure is present (e.g., in the assay mixture used to measure the additive effect with FVIII) at a concentration from about 1 nM to about 1 mM, from about 10 nM to about 500 μ M, from about 50 nM to about 100 μ M, from about 100 nM to about 50 μ M, from about 200 nM to about 40 μ M, from about 300 nM to about 30 μ M, or from about 400 nM to about 20 μ M. In another example, the compound of the present disclosure is present at a concentration from about 500 nM to about 20 μ M, or from about 1 μ M to about 20 μ M, or from about 2 μ M to about 20 μ M, or from about 4 μ M to about 20 μ M, or from about 2 μ M to about 10 μ M. In another example, the compound of the present disclosure is present at a concentration of 100 μ M or less, e.g., 50 μ M or less, 40 μ M or less, 30 μ M or less, 20 μ M or less, 15 μ M or less, 10 μ M or less, or 5 μ M or less.

In one example according to any of the above embodiments, the compound is tested *in vitro* and is present in the assay mixture at a concentration of from about 0.1 μM to about 100 μM , from about 1 μM to about 100 μM , from about 1 μM to about 50 μM , from about 1 μM to about 20 μM , from 1 μM to about 10 μM , from 5 μM to about 20 μM , or from about 5 μM to about 10 μM .

- [0446] An exemplary thrombin generation assay or ROTEM assay useful to measure the additive (or synergistic) activity between the compound and FVIII, are described in Examples 3 and 4, respectively of this application.

Additive Effect with FIX

- [0447] In other embodiments, the compound of the present disclosure does not compete with FIX, but instead shows at least additive activity with FIX or FVIIa in at least one suitable assay. Exemplary assays are selected from thrombin generation assays and ROTEM assays (e.g., those described herein in Examples 3 and 4).

- [0448] In one example, the compound shows additive activity with FIX (*i.e.*, the compound's activity is not inhibited or reduced by the presence of FIX; or the activity of FIX is not inhibited by the presence of the compound) when the FIX is present at a low concentration (e.g., at physiological concentration). In one example, the FIX is present at a concentration from about 0.05 U/mL to about 2.0 U/mL, or from about 0.1 U/mL to about 1.0 U/mL, or from about 0.1 U/mL to about 0.3 U/mL, or about 0.25 U/mL. In one example, the FIX is FIX-Fc.

- [0449] In one example, the compound shows additive activity with FIX (e.g., FIX-Fc) in a thrombin generation assay, e.g., when the FIX is present at a concentration of about 0.1 U/mL to about 0.3 U/mL, or about 0.25 U/mL. In another example, the compound shows additive activity with FIX in a ROTEM assay (e.g., using FIX-deficient plasma), e.g. when FIX is present at a concentration of about 0.1 U/mL to about 0.3 U/mL, or about 0.25 U/mL (see, e.g., Example 12).

Additive Effect with FVIIa

- [0450] In other embodiments, the compound of the present disclosure does not compete with FVIIa, but instead shows at least additive activity with FVIIa in at least one suitable assay. Exemplary assays are selected from thrombin generation assays and ROTEM assays (e.g., those described herein in Examples 3 and 4).

- [0451] In one example, the compound shows additive activity with FVIIa (*i.e.*, the compound's activity is not inhibited or reduced by the presence of FIX; or the activity of FVIIa is not inhibited by the presence of the compound), *e.g.*, when the FVIIa is present at a low concentration (*e.g.*, at physiological concentration). In one example, the FVIIa is present in the assay mixture at a concentration from about 1 U/mL to about 50 U/mL, or from about 1 U/mL to about 30 U/mL, or from about 5 U/mL to about 25 U/mL, or from about 10 U/mL to about 22 U/mL. In one example, the FVIIa is linked to Fc.
- [0452] In one example, the compound shows additive activity with FVIIa (*e.g.*, FVIIa-Fc) in a thrombin generation assay, *e.g.*, when the FVIIa is present at a concentration of about 10 U/mL to about 20 U/mL. In another example, the compound shows additive activity with FVIIa in a ROTEM assay (*e.g.*, using FVIII-deficient plasma), *e.g.* when FVIIa is present at a concentration of about 10 U/mL to about 20 U/mL (*see, e.g.*, Example 12).
- [0453] In another example according to any of the above embodiments, the compound of the present disclosure is present (*e.g.*, in the assay mixture used to measure the additive effect with FIX or FVIIa) at a concentration from about 1 nM to about 1 mM, from about 10 nM to about 500 μ M, from about 50 nM to about 100 μ M, from about 100 nM to about 50 μ M, from about 200 nM to about 40 μ M, from about 300 nM to about 30 μ M, or from about 400 nM to about 20 μ M. In another example, the compound of the present disclosure is present at a concentration from about 500 nM to about 20 μ M, or from about 1 μ M to about 20 μ M, or from about 2 μ M to about 20 μ M, or from about 4 μ M to about 20 μ M, or from about 2 μ M to about 10 μ M. In another example, the compound of the present disclosure is present at a concentration of 100 μ M or less, *e.g.*, 50 μ M or less, 40 μ M or less, 30 μ M or less, 20 μ M or less, 15 μ M or less, 10 μ M or less, or 5 μ M or less. In one example according to any of the above embodiments, the compound is tested *in vitro* and is present in the assay mixture at a concentration of from about 0.1 μ M to about 100 μ M, from about 1 μ M to about 100 μ M, from about 1 μ M to about 50 μ M, from about 1 μ M to about 20 μ M, from 1 μ M to about 10 μ M, from about 1 μ M to about 5 μ M, from about 1 μ M to about 3 μ M, or about 2.5 μ M. In one example according to any of the above embodiments, the compound is present at a concentration sufficient to have additive activity with FIX. In one example according to any of the above embodiments, the compound is present at a concentration sufficient to have additive activity with FVIIa.

[0454] Because the compounds of the present disclosure do not compete with FVIII, FIX and FVII, e.g., with respect to their ability to reduce clotting time or enhance the formation of thrombin, they are suitable to be used in co-therapy with either of these clotting factors. In one example, the compound is used in a co-therapy with FVIII. In another example, the compound is used in a co-therapy with FIX (e.g., FIX-Fc). In yet another example, the compound is used in co-therapy with FVIIa (e.g., FVIIa-Fc).

Replacement of FVIII

[0455] In various embodiments, the compound of the present disclosure induces the formation of thrombin, e.g., in a suitable thrombin generation assay in the absence (or the presence of very low levels) of FVIII. In one example, the compound of the present disclosure induces the formation of thrombin in a thrombin generation assay utilizing FVIII-deficient plasma. In another example, the compound of the present disclosure induces the formation of thrombin in a thrombin generation assay (e.g., utilizing FVIII-deficient plasma) with a thrombin-generation activity comparable to a recombinant FVIII (rFVIII) standard. In one example, the compound of the present disclosure at an assay concentration of about 20 μ M exhibits at least as much thrombin-generation activity as about 0.1 U/mL of rFVIII. In another example, the compound of the present disclosure at an assay concentration of about 20 μ M exhibits at least as much thrombin-generation activity as about 0.25 U/mL of rFVIII. In yet another example, the compound of the present disclosure at an assay concentration of about 20 μ M exhibits at least as much thrombin-generation activity as about 0.5 U/mL of rFVIII. In a further example, the compound of the present disclosure at an assay concentration of about 20 μ M exhibits as much thrombin-generation activity as about 0.5 U/mL of rFVIII. In another example, the compound of the present disclosure at an assay concentration of about 20 μ M exhibits at least as much thrombin-generation activity as about 0.5 U/mL of rFVIII. A suitable thrombin-generation assay (TGA) to measure the above activities is described in Example 3.

[0456] Because the compounds of the present disclosure enhance thrombin formation in the absence of FVIII, they are suitable to be used instead of a FVIII therapy or in conjunction with FVIII therapy.

FIXa and FVIIa Binding

[0457] The compounds of the present disclosure can bind to soluble FIXa or FVIIa, e.g., with a dissociation constant (K_D) of about 300 nM or less, e.g., from about 80 nM to about 300 nM, or from about 100 nM to about 250 nM (*see, e.g.*, Example 8).

[0458] Unexpectedly, the inventors have found that certain compounds of the present disclosure activate FIXa by interacting with a region of the polypeptide sequence near Tyr₁₇₇ (which can also be referred to as the 170 loop) (Tyr₁₇₇: FIXa chymotrypsin numbering; corresponds to Tyr₃₄₅ when using FIX numbering). Since this region of FIXa is known to interact with FVIIIa, the compounds of the present disclosure may activate FIXa similarly to FVIII by moving the loop. In one example according to any of the above embodiments, the compound of the present disclosure is capable of interacting with (e.g., binding to) a peptide, which includes the amino acid sequence:

MFCAG (SEQ ID NO: 1).

[0459] In another example, the compound of the present disclosure is capable of interacting with (e.g., binding to) a peptide, which includes the amino acid sequence:

YNNMFCAGFHE (SEQ ID NO: 2).

[0460] In another example, the compound of the present disclosure is capable of interacting with (e.g., binding to) a peptide, which includes the amino acid sequence:

RSTKFTIYNNMFCAGFHEGGRDSCQG (SEQ ID NO: 3),

or an amino acid sequence having at least 20/26, at least 21/26, at least 22/26, at least 23/26, at least 24/26, or at least 25/26 homology with SEQ ID NO: 3 (e.g., at least 20, at least 21, at least 22, at least 23, at least 24, or at least 25 of the 26 amino acids are those as shown in SEQ ID NO: 3; or not more than 6, not more than 5, not more than 4, not more than 3, not more than 2, or not more than 1 of the 26 amino acids of SEQ ID NO: 3 are replaced by another amino acid).

[0461] In another example, the compound of the present disclosure is capable of interacting with a FIXa protein at a region corresponding to amino acid sequence:

MFCAG (SEQ ID NO: 1).

[0462] In another example, the compound of the present disclosure is capable of interacting with a FIXa protein at a region corresponding to amino acid sequence:

YNNMFCAGFHE (SEQ ID NO: 2).

[0463] In another example, the compound of the present disclosure is capable of interacting with a FIXa protein at a region corresponding to amino acid sequence:

RSTKFTIYNNMFCAGFHEGGRDSCQG (SEQ ID NO: 3),

or an amino acid sequence having at least 20/26, at least 21/26, at least 22/26, at least 23/26, at least 24/26, or at least 25/26 homology with SEQ ID NO: 3 (e.g., at least 20, at least 21, at least 22, at least 23, at least 24, or at least 25 of the 26 amino acids are those as shown in SEQ ID NO: 3; or not more than 6, not more than 5, not more than 4, not more than 3, not more than 2, or not more than 1 of the 26 amino acids of SEQ ID NO: 3 are replaced by another amino acid).

[0464] There is a strong homology between proteases in the coagulation cascade, e.g., FIX, FX, prothrombin, FVII, and protein C. One amino acid region that showed changes in HDX levels (amino acids FCAG) in FIXa is conserved across these peptides indicating that the compound of the invention can potentially interact with at least one of these blood coagulation factors other than FIXa. For example, the compounds of the present disclosure can potentially increase the catalytic activity of e.g. FVIIa, FXa or thrombin in addition to FIXa. For example, given the structural similarities between FIXa and FVIIa, the pro-coagulant compounds of the present disclosure may employ a similar mechanism for increasing the catalytic activities of FVIIa.

[0465] In one example, the compound of the present disclosure has pro-coagulant activity. It will be appreciated that different assays are available to determine pro-coagulant activity. In one example, the compound of the present disclosure has pro-coagulant activity when it shows activity in at least one of: an activated partial thromboplastin time (aPTT) assay, a modified activated partial thromboplastin time (aPTT*) assay, a thrombin generation assay (TGA), and a rotational thromboelastometry (ROTEM) assay, which are described herein, e.g., in Examples 9, 3, and 4, respectively.

[0466] A compound of the present disclosure may promote coagulation in plasma depleted of FV, FVII, FVIII, FX, FXI, or vWF. In one example, a compound of the present disclosure promotes thrombin generation and/or fibrin deposition in plasma in which FVIII is depleted or absent. This type of activity is referred to as coagulation FVIII activity. Where the plasma is from an individual lacking FVIII or having reduced levels of FVIII, the activity is typically referred to as FVIII equivalent activity. Where the plasma contains inhibitors against FVIII, the activity is typically referred to as FVIII

inhibitor bypassing equivalent activity. Other procoagulant activities include FV activity, FVII activity, FX activity and FXI activity.

[0467] Individual compounds can vary in their relative efficacy between different types of assay. Therefore, even if a compound appears to have a low efficacy in a particular assay, it may nevertheless possess a suitably high level of procoagulant activity in another assay.

[0468] Other suitable assays useful to determine pro-coagulant activity include those disclosed, e.g., in Patent Application Publication U.S. 2010/0022445 to Scheifflinger and Dockal, which is incorporated herein by reference in its entirety.

[0469] In one example according to any of the above embodiments, certain compounds of the present disclosure (e.g., a compound listed in Table 1) inhibit heparin catalyzed (heparin accelerated) FIXa-AT complex formation when compared to FIXa-AT complex formation in the absence of the compound.

[0470] "FIXa-AT" is a covalent and equimolar complex formed between FIXa and antithrombin (AT). Antithrombin belongs to the serpin family of inhibitors and is a known physiological inhibitor of coagulation proteases, e.g. FIXa. On the surface of intact endothelium, AT interacts with heparin sulfate and its rate of protease inhibition accelerates. (see, e.g., Johnson, D. J. D. et al, *PNAS* 2010, 107, 645-650; Yang L. et al, *Journal of Biological Chemistry*, 2002, 277, 50756-50760).

[0471] FIXa-AT complex formation can be measured, e.g., as described herein in Example 11. In one embodiment FIXa-AT complex formation is measured in the presence of heparin at a concentration of from about 10 nM to about 200 nM. In another embodiment FIXa-AT complex formation is measured in the presence of heparin at a concentration of from about 50 nM to about 150 nM. In one embodiment FIXa-AT complex formation is measured in the presence of about 10 nM heparin. In another embodiment FIXa-AT complex formation is measured in the presence of about 50 nM heparin. In another embodiment FIXa-AT complex formation is measured in the presence of about 100 nM heparin. In yet another embodiment FIXa-AT complex formation is measured in the presence of about 150 nM heparin.

[0472] In another embodiment, FIXa-AT complex formation is measured at a compound concentration of from about 0.1 μ M to about 100 μ M. In another embodiment, FIXa-AT complex formation is measured at a compound concentration of from about 0.5 μ M to

about 50 μM . In another embodiment, FIXa-AT complex formation is measured at a compound concentration of from about 1 μM to about 20 μM . In another embodiment, FIXa-AT complex formation is measured at a compound concentration of from about 1 μM to about 10 μM . In another embodiment, FIXa-AT complex formation is measured at a compound concentration of about 0.1 μM , about 0.5 μM , about 1 μM , about 2 μM , about 3 μM , about 4 μM , about 5 μM , about 6 μM , about 7 μM , about 8 μM , about 9 μM , about 10 μM , about 12 μM , about 14 μM , about 16 μM , about 18 μM , or about 20 μM .

[0473] In another embodiment FIXa-AT complex formation is measured in the presence of heparin at a concentration of from about 10 nM to about 200 nM and at a compound concentration of from about 0.1 μM to about 100 μM . In another embodiment FIXa-AT complex formation is measured in the presence of heparin at a concentration of from about 50 nM to about 150 nM and at a compound concentration of from about 0.5 μM to about 50 μM . In another embodiment FIXa-AT complex formation is measured in the presence of heparin at a concentration of from about 50 nM to about 150 nM and at a compound concentration of from about 1 μM to about 20 μM .

[0474] In another embodiment FIXa-AT complex formation is measured in the presence of heparin at a concentration of about 100 nM and at a compound concentration of about 1 μM . In another embodiment FIXa-AT complex formation is measured in the presence of about 100 nM heparin and at a compound concentration of about 2 μM . In another embodiment FIXa-AT complex formation is measured in the presence of about 100 nM heparin and at a compound concentration of about 3 μM . In another embodiment FIXa-AT complex formation is measured in the presence of about 100 nM heparin and at a compound concentration of about 4 μM . In another embodiment FIXa-AT complex formation is measured in the presence of about 100 nM heparin and at a compound concentration of about 5 μM . In another embodiment FIXa-AT complex formation is measured in the presence of about 100 nM heparin and at a compound concentration of about 6 μM . In another embodiment FIXa-AT complex formation is measured in the presence of about 100 nM heparin and at a compound concentration of about 7 μM . In another embodiment FIXa-AT complex formation is measured in the presence of about 100 nM heparin and at a compound concentration of about 8 μM . In another embodiment FIXa-AT complex formation is measured in the presence of about 100 nM heparin and at a compound concentration of about 9 μM . In another embodiment FIXa-

AT complex formation is measured in the presence of about 100 nM heparin and at a compound concentration of about 10 μ M.

[0475] In one example according to any one of the above embodiments, the FIXa-AT complex formation measured for a compound of the present disclosure is inhibited by between about 5% and about 90% compared to the complex formation in the absence of the compound. In one example according to any one of the above embodiments, the FIXa-AT complex formation measured for a compound of the present disclosure is inhibited by between about 10% and about 80% compared to the complex formation in the absence of the compound. In one example according to any one of the above embodiments, the FIXa-AT complex formation measured for a compound of the present disclosure is inhibited by between about 20% and about 80% compared to the complex formation in the absence of the compound.

[0476] In another example according to any one of the above embodiments, the FIXa-AT complex formation measured for a compound of the present disclosure is inhibited by at least about 5% when compared to the complex formation in the absence of the compound. In another example according to any one of the above embodiments, the FIXa-AT complex formation is inhibited by at least about 10%. In another example according to any one of the above embodiments, the FIXa-AT complex formation is inhibited by at least about 20%. In yet another example according to any one of the above embodiments, the FIXa-AT complex formation is inhibited by at least about 30 %. In another example according to any one of the above embodiments, the FIXa-AT complex formation is inhibited by at least about 40%. In another example according to any one of the above embodiments, the FIXa-AT complex formation is inhibited by at least about 50%. In another example according to any one of the above embodiments, the FIXa-AT complex formation is inhibited by at least about 60%. In another example according to any one of the above embodiments, the FIXa-AT complex formation is inhibited by at least about 70%. In another example according to any one of the above embodiments, the FIXa-AT complex formation is inhibited by at least about 80%.

[0477] In one embodiment, the compounds of the present disclosure can compete with heparin for binding to FIXa, and can thus be viewed as heparin antagonists.

[0478] In one embodiment the compounds of the present disclosure antagonize the delay in plasma clotting caused by heparin (e.g., low molecular weight heparin). In another

example, the compounds of the present disclosure can be used as antidotes for hemorrhagic complications associated with heparin therapy. In another example, the compounds of the present disclosure can be used as antidotes for hemorrhagic complications associated with antithrombotic therapies (e.g., SH and LMWH antithrombotic therapies).

[0479] In one example, the invention provides a method of providing an antidote to heparin (e.g., low molecular weight heparin) overdose in a subject in need thereof, the method comprising administering to the subject an effective amount of a compound of the present disclosure, or an acceptable salt or solvate thereof, or a pharmaceutical composition comprising a compound of the present disclosure.

[0480] Assays for determining the heparin-neutralizing activity of a polymer or oligomer are either described herein or are well known to those of skill in the art. See, e.g., Kandrotas, R.J., *Clin. Pharmacokinet.* 22:359-374 (1992)), Diness, V.O., and Østergaard, P.B., *Thromb. Haemost.* 56:318-322 (1986)), and references cited therein, Wong, P.C., *et al.*, *J. Pharm. Exp. Therap.* 292:351-357 (2000), Ryn-McKenna, J.V., *et al.*, *Thromb. Haemost.* 63:271-274 (1990), and Wakefield, T.W., *et al.*, *J. Surg. Res.* 63:280-286 (1996).

Effect on platelets

[0481] In one embodiment, the compounds of the present disclosure do not impact platelet function and do not induce platelet aggregation as shown, e.g., in Example 13.

Animal model

[0482] In one example, a compound of the present disclosure can at least partially compensate for the absence of biologically active FVIII when administered in an animal model of severe human hemophilia A. For example, a compound can be active in controlling bleeding in FVIII deficient mice or dogs.

[0483] A exemplary assay to test the ability of a compound or conjugate to control bleeding is the tail clip assay (see, e.g., Pan J, *et al.*, *Blood* 2009;114:2802-2811). Compounds or conjugates are administered to mice in a suitable vehicle, typically i.v., i.p. or s.c. Different doses of each peptide or peptide derivative may be administered to different groups of mice to determine dose-dependency. In one example, mice administered the compound or conjugate have a blood loss in the tail clip assay at 62

minutes from tail clip of no more than 70% of the blood loss of mice administered the vehicle alone, more preferably no more than 60%, and most preferably no more than 50% of the blood loss of mice administered the vehicle alone.

[0484] In another example, survival of mice administered the compound or conjugate in the above assay is at least 40%, more preferably at least 60% and most preferably at least 80% at 2 hours after tail clip. Preferably, survival of mice administered the compound or conjugate in the tail clip assay is at least 20%, more preferably at least 30% and most preferably at least 40% at 24 hours after tail clip.

[0485] Exemplary compounds of the present disclosure and their *in vitro* biological activities measured using a FXa generation assay are described in Example 2.

[0486] Exemplary compounds of the present disclosure and their *in vitro* biological activities measured using a thrombin generation assay are described in Example 3.

[0487] In one example, the compound of the present disclosure (e.g., compound 5 or 6) has improved chemical stability in human plasma (e.g., in the presence of FIXa). For example, the compound of the present disclosure, after incubation of 30 minutes, 60 minutes, or 120 minutes in human plasma, is at least 50%, at least 70%, at least 80%, at least 90%, at least 95%, at least 98% or at least 99% stable. A stability of 100% indicates that no detectable degradation of the compound has occurred during the specified incubation time in human plasma. In one example, the compound is chemically stable (e.g., at least about 90% stable, or 100% stable) for at least about 60 min when incubated in human plasma. In another example, the compound is stable for at least about 80 min when incubated in human plasma. In yet another example, the compound is stable for at least about 100 min when incubated in human plasma. In a further example, the compound is stable for at least about 120 min. A suitable assay to determine stability in human plasma is described in Example 6.

[0488] In another example, the compound of the present disclosure has an aqueous solubility in phosphate buffered saline at pH 7.4 and 25°C of at least about 25 μ M, preferably at least about 60 μ M, and most preferably at least about 100 μ M.

Methods Of Making The Compounds

[0489] The compounds of the present disclosure (e.g., peptides or peptide derivatives) can be produced by chemical synthesis, recombinant DNA technology, biochemical or

enzymatic fragmentation of larger molecules, combinations of the foregoing or by any other method.

[0490] In one example, the method comprises forming the amino acid sequence of the compound, or a retro-, inverso- or retro-inverso variant thereof using solid-phase peptide synthesis. Exemplary methods of making the compounds of the invention are described herein in Example 1. Other methods to form peptides are known to those of skill in the art.

[0491] For example, the compounds of the present disclosure can be synthesised using solid-phase peptide synthesis as described in "*Fmoc Solid Phase Peptide Synthesis - A Practical Approach*", edited by W. C. Chan, P. D. White, Oxford University Press, New York 2000 and references therein. Temporary N-amino group protection is afforded, e.g., by a 9-fluorenylmethyloxycarbonyl (Fmoc) group. Repetitive cleavage of this highly base-labile protecting group is effected, e.g., using 20% piperidine in N,N-dimethylformamide. Side-chain functionalities may be protected as their butyl ethers (in the case of serine, threonine and tyrosine), butyl esters (in the case of glutamic acid and aspartic acid), butyloxycarbonyl derivative (in the case of lysine and histidine), trityl derivative (in the case of cysteine, asparagine and glutamine) and 4-methoxy-2,3,6-trimethylbenzenesulphonyl derivative (in the case of arginine). The solid-phase support can be based on a polydimethyl-acrylamide polymer constituted from the three monomers dimethylacrylamide (backbone-monomer), bisacryloylethylene diamine (cross linker) and acryloylsarcosine methyl ester (functionalising agent), or can be based on polyethylene glycol (PEG), such as Rink Amide resin (e.g., NovaPEG Rink Amide). The peptide-to-resin cleavable linked agent can be the acid-labile 4-hydroxymethyl-phenoxyacetic acid derivative, or in case of C-terminal amides, the Rink-amide linker. All amino acid derivatives can be added as their preformed symmetrical anhydride derivatives with the exception of asparagine and glutamine, which are added using a reversed N,N-dicyclohexyl-carbodiimide/1-hydroxybenzotriazole mediated coupling procedure. Alternatively, other peptide coupling reagents, such as O-benzotriazole-N,N,N',N'-tetramethyl-uronium-hexafluoro-phosphate (HBTU) or 2-(6-chloro-1-H-benzotriazole-1-yl)-1,1,3,3-tetramethylaminium hexafluorophosphate (HCTU) can be used (e.g., *in situ*). Coupling and deprotection reactions can be monitored using ninhydrin, trinitrobenzene sulphonic acid or isotin test procedures. Upon completion of synthesis, peptides are

cleaved from the resin support with concomitant removal of side-chain protecting groups, e.g., by treatment with 95% trifluoroacetic acid containing about 5-50% scavenger. Scavengers commonly used are TIPS (triisopropylsilane), ethanedithiol, phenol, anisole, water, and mixtures thereof. The exact choice depends on the constituent amino acids of the peptide being synthesised. For methionine containing peptides one can use, e.g., a mixture of TIPS (e.g., 2-5%) and ethanedithiol (e.g., 2-5%).

[0492] Trifluoroacetic acid can subsequently be removed by evaporation *in vacuo*, with subsequent trituration with diethyl ether affording the crude peptide. Any scavengers present can be removed by a simple extraction procedure which on lyophilisation of the aqueous phase affords the crude peptide free of scavengers.

[0493] Reagents for peptide synthesis are generally available, e.g., from Calbiochem-Novabiochem (UK), or EMD4Biosciences (U.S.).

[0494] Purification of the peptides may be effected by any one, or a combination of, techniques such as size exclusion chromatography, ion-exchange chromatography, affinity chromatography, differential solubility, and reverse-phase high performance liquid chromatography. Analysis of peptides may be carried out using thin layer chromatography, reverse-phase high performance liquid chromatography, mass spectroscopy (e.g., LC-MS), amino-acid analysis after acid hydrolysis and by fast atom bombardment (FAB) mass spectrometry.

[0495] SPOT-synthesis, which allows the positional addressable, chemical synthesis of peptides on continuous cellulose membranes may be also used (see, e.g., R. Frank, *Tetrahedron* (1992) 48, 9217).

[0496] The compounds of the present disclosure may also be produced by recombinant protein expression or *in vitro* translation systems (see, e.g., Sambrook *et al.*, "*Molecular cloning: A laboratory manual*", 2001, 3rd edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.). Recombinant methods are generally preferred when the peptide is particularly large, e.g., larger than 50 amino acids, or larger than 100 amino acids.

Methods of Making The Polypeptide Conjugates

[0497] The conjugates of the present disclosure can be made recombinantly, see, e.g., procedures for FVIII-Fc expression and purification described in WO2011/069164, which is incorporated herein by reference in its entirety.

[0498] For example, the suitable expression vector or vectors are transfected or co-transfected into a suitable target cell, which will express the polypeptides. Transfection techniques known in the art include, but are not limited to, calcium phosphate precipitation (Wigler *et al.* (1978) *Cell* 14:725), electroporation (Neumann *et al.* (1982) *EMBO J* 1:841), and liposome-based reagents. A variety of host-expression vector systems may be utilized to express the proteins described herein including both prokaryotic and eukaryotic cells. These include, but are not limited to, microorganisms such as bacteria (*e.g.*, *E. coli*) transformed with recombinant bacteriophage DNA or plasmid DNA expression vectors containing an appropriate coding sequence; yeast or filamentous fungi transformed with recombinant yeast or fungi expression vectors containing an appropriate coding sequence; insect cell systems infected with recombinant virus expression vectors (*e.g.*, baculovirus) containing an appropriate coding sequence; plant cell systems infected with recombinant virus expression vectors (*e.g.*, cauliflower mosaic virus or tobacco mosaic virus) or transformed with recombinant plasmid expression vectors (*e.g.*, Ti plasmid) containing an appropriate coding sequence; or animal cell systems, including mammalian cells (*e.g.*, HEK 293, CHO, Cos, HeLa, HKB11, and BHK cells).

[0499] In one embodiment, the host cell is a eukaryotic cell. As used herein, a eukaryotic cell refers to any animal or plant cell having a definitive nucleus. Eukaryotic cells of animals include cells of vertebrates, *e.g.*, mammals, and cells of invertebrates, *e.g.*, insects. Eukaryotic cells of plants specifically can include, without limitation, yeast cells. A eukaryotic cell is distinct from a prokaryotic cell, *e.g.*, bacteria.

[0500] In certain embodiments, the eukaryotic cell is a mammalian cell. A mammalian cell is any cell derived from a mammal. Mammalian cells specifically include, but are not limited to, mammalian cell lines. In one embodiment, the mammalian cell is a human cell. In another embodiment, the mammalian cell is a HEK 293 cell, which is a human embryonic kidney cell line. HEK 293 cells are available as CRL-1533 from American Type Culture Collection, Manassas, VA, and as 293-H cells, Catalog No. 11631-017 or 293-F cells, Catalog No. 11625-019 from Invitrogen (Carlsbad, Calif.). In some embodiments, the mammalian cell is a PER.C6[®] cell, which is a human cell line derived from retina. PER.C6[®] cells are available from Crucell (Leiden, The Netherlands). In other embodiments, the mammalian cell is a Chinese hamster ovary (CHO) cell. CHO

cells are available from American Type Culture Collection, Manassas, VA. (e.g., CHO-K1; CCL-61). In still other embodiments, the mammalian cell is a baby hamster kidney (BHK) cell. BHK cells are available from American Type Culture Collection, Manassas, Va. (e.g., CRL-1632). In some embodiments, the mammalian cell is a HKB11 cell, which is a hybrid cell line of a HEK293 cell and a human B cell line. Mei *et al.*, *Mol. Biotechnol.* 34(2): 165-78 (2006).

[0501] In another example, the conjugates of the present disclosure can be made semi-recombinantly e.g., as illustrated in Figures 23-26 (see, e.g., U.S. Patent 7,381,408 to Mezo, A.R. and Peters, R. P.; Dawson, P.E., Kent, S.B. *Ann. Rev. Biochem.* (2000) 69: 923-9600; Mei, B. *et. al.*, *Blood* (2010) 116:270-279; and U.S. Patent Application Publication US2006/0115876 to Pan, C. *et. al.*, each of which is incorporated herein in its entirety). In one example, the polypeptide or the polypeptide construct containing a suitable amino acid moiety is made recombinantly. The pro-coagulant compound is then attached to the polypeptide or polypeptide construct via chemical ligation as described herein.

[0502] Figure 23 illustrates a general method for covalently linking a peptide or peptide derivative to a FVIII-Fc, a FIX-Fc, or a FVIIa-Fc construct using a native ligation strategy (see, e.g., Mezo, A.R.; Peters, R. P., *Methods for Chemically Synthesizing Immunoglobulin Chimeric Proteins*. U.S. Patent 7,381,408; and Dawson, P.E., Kent, S.B. *Synthesis of native proteins by chemical ligation*. *Ann. Rev. Biochem.* (2000) 69: 923-9600). An exemplary method includes contacting a FVIII-Fc construct (e.g., a truncated FVIII-Fc construct) having a free sulfhydryl group located at the Fc portion of the construct (e.g., having an N-terminal cysteine) with a peptide or peptide derivative having a reactive group selected from a thioester moiety, a maleimide moiety and a iodoacetamide moiety, under reaction conditions sufficient to form a covalent bond between the peptide or peptide derivative and the FVIII-Fc construct. FVIII in the FVIII-Fc construct can be replaced by FIX or FVIIa, and the resulting construct can be ligated to a peptide or peptide derivative in the same way.

[0503] Figure 24 illustrates a general method for covalently linking a peptide or peptide derivative to a FVIII-Fc construct, a FIX-Fc construct, or a FVIIa-Fc construct using a site-directed ligation strategy (see, e.g., Mei, B. *et. al.* *Rational design of a fully active, long-acting PEGylated FVIII for hemophilia A treatment*. *Blood* (2010) 116:270-279;

and Pan, C. *et. al.* Site-directed modification of FVIII. U.S. Patent Application Publication US2006/0115876). This linking strategy is also useful for the preparation of FVIII conjugates, in which the peptide or peptide derivative is linked to FVIII instead of a FVIII-Fc construct. An exemplary method includes contacting FVIII having a free sulfhydryl group (e.g., an internal cysteine), or a FVIII-Fc construct having a free sulfhydryl group located at the FVIII portion of the construct (e.g., an internal cysteine) with a peptide or peptide derivative having a reactive group (e.g., a maleimide or iodoacetamide moiety), under reaction conditions sufficient to form a covalent bond between the peptide or peptide derivative and the FVIII or the FVIII-Fc construct. This linking strategy is also useful for the preparation of FIX or FVIIa conjugates by the same method described above for FVIII. Other reagents useful to form a covalent bond between a sulfhydryl group and another moiety are known to those of skill in the art.

[0504] Figure 25 illustrates a general method for covalently linking a peptide or peptide derivative to a platelet targeting moiety-Fc construct using a native ligation strategy (see, e.g., Mezo, A.R.; Peters, R. P., *Methods for Chemically Synthesizing Immunoglobulin Chimeric Proteins*. U.S. Patent 7,381,408; and Dawson, P.E., Kent, S.B. *Synthesis of native proteins by chemical ligation. Ann. Rev. Biochem.* (2000) 69: 923-9600). An exemplary method includes contacting a platelet targeting moiety-Fc construct (e.g., a truncated platelet targeting moiety-Fc construct) having a free sulfhydryl group located at the Fc portion of the construct (e.g., having an N-terminal cysteine) with a peptide or peptide derivative having a reactive group selected from a thioester moiety, a maleimide moiety and a iodoacetamide moiety, under reaction conditions sufficient to form a covalent bond between the peptide or peptide derivative and the platelet targeting moiety-Fc construct.

[0505] Figure 26 illustrates a general method for covalently linking a peptide or peptide derivative to a platelet targeting moiety-Fc construct using a site-directed ligation strategy (see, e.g., Mei, B. *et. al.* *Rational design of a fully active, long-acting PEGylated FVIII for hemophilia A treatment. Blood* (2010) 116:270-279; and Pan, C. *et. al.* *Site-directed modification of FVIII. U.S. Patent Application Publication US2006/0115876*). This linking strategy is also useful for the preparation of platelet targeting moiety conjugates, in which the peptide or peptide derivative is linked to the platelet targeting moiety instead of a platelet targeting moiety-Fc construct. An exemplary method includes contacting the

platelet targeting moiety having a free sulfhydryl group (e.g., an internal cysteine), or a platelet targeting moiety-Fc construct having a free sulfhydryl group located at the platelet targeting moiety portion of the construct (e.g., an internal cysteine) with a peptide or peptide derivative having a reactive group (e.g., a maleimide or iodoacetamide moiety), under reaction conditions sufficient to form a covalent bond between the peptide or peptide derivative and the platelet targeting moiety or the platelet targeting moiety-Fc construct. Other reagents useful to form a covalent bond between a sulfhydryl group and another moiety are known to those of skill in the art (see, e.g., U.S. Patent 7,381,408).

Pharmaceutical Formulations

- [0506] The invention also provides a pharmaceutical composition (also referred to as pharmaceutical formulation) containing at least one compound of the present disclosure according to any of the above embodiments and a pharmaceutically acceptable carrier.
- [0507] The invention further provides a pharmaceutical composition containing at least one conjugate of the present disclosure according to any of the above embodiments and a pharmaceutically acceptable carrier.
- [0508] The term "pharmaceutically acceptable carrier" means all pharmaceutically acceptable ingredients known to those of skill in the art, which are typically considered non-active ingredients. The term "pharmaceutically acceptable carrier" includes, e.g., solvents, solid or liquid diluents, additives, vehicles, adjuvants, excipients, glidants, binders, granulating agents, dispersing agents, suspending agents, wetting agents, lubricating agents, disintegrants, solubilizers, stabilizers, preservatives, emulsifiers, fillers, preservatives (e.g., anti-oxidants), flavoring agents, sweetening agents, thickening agents, buffering agents, coloring agents and the like, as well as any mixtures thereof. Exemplary carriers (i.e., excipients) are described in, e.g., Handbook of Pharmaceutical Manufacturing Formulations, Volumes 1-6, Niazi, Sarfaraz K., Taylor & Francis Group 2005, which is incorporated herein by reference in its entirety. Pharmaceutical compositions may additionally comprise, for example, one or more of water, buffers (e.g., neutral buffered saline or phosphate buffered saline), ethanol, mineral oil, vegetable oil, dimethylsulfoxide, carbohydrates (e.g., glucose, mannose, sucrose or dextrans), mannitol, proteins, adjuvants, polypeptides or amino acids such as glycine, antioxidants, chelating agents such as EDTA or glutathione and/or preservatives.

- [0509] Pharmaceutical compositions may be formulated for any appropriate manner of administration, including, for example, topical (e.g., transdermal or ocular), oral, buccal, nasal, vaginal, rectal or parenteral administration. In some embodiments, the compound or conjugate of the present disclosure is administered parenterally, e.g., intravenously or subcutaneously.
- [0510] The term parenteral as used herein includes subcutaneous, intradermal, intravascular (e.g., intravenous), intramuscular, spinal, intracranial, intrathecal, intraocular, periocular, intraorbital, intrasynovial and intraperitoneal injection, as well as any similar injection or infusion technique. It is preferred that subcutaneous, intraperitoneal, buccal, intravenous and other parenteral formulations are sterile and endotoxin free.
- [0511] Compounds or conjugates of the present disclosure may be administered parenterally in a sterile medium. The compound or conjugate, depending on the vehicle and concentration used, can either be suspended or dissolved in the vehicle. In one embodiment, adjuvants such as local anesthetics, preservatives and buffering agents can be dissolved in the vehicle.
- [0512] In one example, the compounds of the present disclosure are administered to the subject using a non-intravenous route, e.g., by subcutaneous, nasal, buccal, oral or pulmonary delivery.
- [0513] Forms suitable for oral use include, for example, tablets, troches, lozenges, aqueous or oily suspensions, dispersible powders or granules, emulsion, hard or soft capsules, or syrups or elixirs. Compositions provided herein may be formulated as a lyophilizate.
- [0514] Compositions intended for oral use may be prepared according to any method known for the manufacture of pharmaceutical compositions. Such compositions may contain one or more agents chosen from the group consisting of sweetening agents, flavoring agents, coloring agents and preservative agents in order to provide pharmaceutically elegant and palatable preparations. Tablets can contain the active ingredient in admixture with non-toxic pharmaceutically acceptable excipients that are suitable for the manufacture of tablets. These excipients may be for example, inert diluents, such as calcium carbonate, sodium carbonate, lactose, calcium phosphate or sodium phosphate; granulating and disintegrating agents, for example, corn starch, or

alginic acid; binding agents, for example starch, gelatin or acacia, and lubricating agents, for example magnesium stearate, stearic acid or talc. The tablets may be uncoated or they may be coated by known techniques. In some cases such coatings may be prepared by known techniques to delay disintegration and absorption in the gastrointestinal tract and thereby provide a sustained action over a longer period (i.e., tablets can be enterically coated). For example, a time delay material such as glyceryl monostearate or glyceryl distearate may be employed.

[0515] Formulations for oral use may also be presented as hard gelatin capsules, wherein the active ingredient is mixed with an inert solid diluent, for example, calcium carbonate, calcium phosphate or kaolin, or as soft gelatin capsules wherein the active ingredient is mixed with water or an oil medium, for example peanut oil, liquid paraffin or olive oil. In another example, the active ingredient is formulated in capsules containing optionally coated microtablets or micropellets. Formulations for oral use may also be presented as lozenges. The composition can be also for example a suspension, emulsion, sustained release formulation, cream, gel or powder. The composition can be formulated as a suppository, with traditional binders and carriers such as triglycerides.

[0516] Oily suspensions may be formulated by suspending the active ingredients in a vegetable oil, for example arachis oil, olive oil, sesame oil or coconut oil, or in a mineral oil such as liquid paraffin. The oily suspensions may contain a thickening agent, for example beeswax, hard paraffin or cetyl alcohol. Sweetening agents and flavoring agents may be added to provide palatable oral preparations. These compositions may be preserved by the addition of an anti-oxidant such as ascorbic acid.

[0517] In one example, the pharmaceutical formulation is a liquid formulation, e.g., a buffered, isotonic, aqueous solution. In one example, the pharmaceutical composition has a pH that is physiologic, or close to physiologic. In another example, the aqueous formulation has a physiologic or close to physiologic osmolarity and salinity. It can contain sodium chloride and/or sodium acetate.

[0518] Aqueous suspensions contain the active ingredient(s) in admixture with excipients suitable for the manufacture of aqueous suspensions. Such excipients include suspending agents (e.g., sodium carboxymethylcellulose, methylcellulose, hydropropylmethylcellulose, sodium alginate, polyvinyl-pyrrolidone, gum tragacanth and gum acacia); and dispersing or wetting agents (e.g., naturally-occurring phosphatides

such as lecithin, condensation products of an alkylene oxide with fatty acids such as polyoxyethylene stearate, condensation products of ethylene oxide with long chain aliphatic alcohols such as heptadecaethyleneoxycetanol, condensation products of ethylene oxide with partial esters derived from fatty acids and a hexitol such as polyoxyethylene sorbitol monooleate, or condensation products of ethylene oxide with partial esters derived from fatty acids and hexitol anhydrides such as polyethylene sorbitan monooleate). Aqueous suspensions may also comprise one or more preservatives, for example ethyl, or n-propyl p-hydroxybenzoate, one or more coloring agents, one or more flavoring agents, and one or more sweetening agents, such as sucrose or saccharin.

[0519] Dispersible powders and granules suitable for preparation of an aqueous suspension by the addition of water can provide the active ingredient in admixture with a dispersing or wetting agent, suspending agent and one or more preservatives. Suitable dispersing or wetting agents or suspending agents are exemplified by those already mentioned above. Additional excipients, for example sweetening, flavoring and coloring agents, may also be present.

[0520] Pharmaceutical compositions of the present disclosure may also be in the form of oil-in-water emulsions. The oily phase may be a vegetable oil or a mineral oil or mixtures of these. Suitable emulsifying agents may be naturally-occurring gums, for example gum acacia or gum tragacanth, naturally-occurring phosphatides, for example soy bean, lecithin, and esters or partial esters derived from fatty acids and hexitol, anhydrides, for example sorbitan monooleate, and condensation products of the said partial esters with ethylene oxide, for example polyoxyethylene sorbitan monooleate. The emulsions may also contain sweetening and flavoring agents.

[0521] Syrups and elixirs may be formulated with sweetening agents, for example glycerol, propylene glycol, sorbitol, glucose or sucrose. Such formulations may also contain a demulcent, a preservative, a flavoring agent or a coloring agent. The pharmaceutical compositions may be in the form of a sterile injectable aqueous or oleaginous suspension. This suspension may be formulated according to the known art using those suitable dispersing or wetting agents and suspending agents that have been mentioned above. The sterile injectable preparation may also be a sterile injectable solution or suspension in a non-toxic parentally acceptable diluent or solvent, for example

as a solution in 1,3-butanediol. Among the acceptable vehicles and solvents that may be employed are water, Ringer's solution and isotonic sodium chloride solution. In addition, sterile, fixed oils are conventionally employed as a solvent or suspending medium. For this purpose any bland fixed oil may be employed including synthetic mono- or diglycerides. In addition, fatty acids such as oleic acid find use in the preparation of injectables.

[0522] The compounds of the present disclosure may also be administered in the form of suppositories, e.g., for rectal administration of the drug. These compositions can be prepared by mixing the drug with a suitable non-irritating excipient that is solid at ordinary temperatures but liquid at the rectal temperature and will therefore melt in the rectum to release the drug. Such materials include cocoa butter and polyethylene glycols.

[0523] Compounds of the present disclosure can be formulated for local or topical administration, such as for topical application to the skin, wounds or mucous membranes, such as in the eye. Formulations for topical administration typically comprise a topical vehicle combined with active agent(s), with or without additional optional components. Suitable topical vehicles and additional components are well known in the art, and it will be apparent that the choice of a vehicle will depend on the particular physical form and mode of delivery. Topical vehicles include water; organic solvents such as alcohols (e.g., ethanol or isopropyl alcohol) or glycerin; glycols (e.g., butylene, isoprene or propylene glycol); aliphatic alcohols (e.g., lanolin); mixtures of water and organic solvents and mixtures of organic solvents such as alcohol and glycerin; lipid-based materials such as fatty acids, acylglycerols (including oils, such as mineral oil, and fats of natural or synthetic origin), phosphoglycerides, sphingolipids and waxes; protein-based materials such as collagen and gelatin; silicone-based materials (both non-volatile and volatile); and hydrocarbon-based materials such as microsponges and polymer matrices. A composition may further include one or more components adapted to improve the stability or effectiveness of the applied formulation, such as stabilizing agents, suspending agents, emulsifying agents, viscosity adjusters, gelling agents, preservatives, antioxidants, skin penetration enhancers, moisturizers and sustained release materials. Examples of such components are described in Martindale--The Extra Pharmacopoeia (Pharmaceutical Press, London 1993) and Martin (ed.), Remington's Pharmaceutical Sciences. Formulations may comprise microcapsules, such as hydroxymethylcellulose or gelatin-

microcapsules, liposomes, albumin microspheres, microemulsions, nanoparticles or nanocapsules.

[0524] For disorders of the eye or other external tissues, e.g., mouth and skin, the formulations are applied, for example, as a topical gel, spray, ointment or cream, or as a scleral suppository, containing the active ingredients in a total amount of, for example, 0.075 to 30% w/w, 0.2 to 20% w/w or such as 0.4 to 15% w/w. When formulated in an ointment, the active ingredients may be employed with either paraffinic or a water-miscible ointment base.

[0525] Formulations suitable for topical administration to the eye also include eye drops wherein the active ingredients are dissolved or suspended in suitable carrier, especially an aqueous solvent for the active ingredients. The anti-inflammatory active ingredients may, for example, be present in such formulations in a concentration of 0.5 to 20%, such as 0.5 to 10%, for example about 1.5% w/w. For therapeutic purposes, the active compounds of the present disclosure are ordinarily combined with one or more adjuvants appropriate to the indicated route of administration. The compounds may be admixed with lactose, sucrose, starch powder, cellulose esters of alkanolic acids, cellulose alkyl esters, talc, stearic acid, magnesium stearate, magnesium oxide, sodium and calcium salts of phosphoric and sulfuric acids, gelatin, acacia gum, sodium alginate, polyvinylpyrrolidone, and/or polyvinyl alcohol, and then tableted or encapsulated for convenient administration. Such capsules or tablets may contain a controlled-release formulation as may be provided in a dispersion of active compound in hydroxypropylmethyl cellulose. Formulations for parenteral administration may be in the form of aqueous or non-aqueous isotonic sterile injection solutions or suspensions. These solutions and suspensions may be prepared from sterile powders or granules having one or more of the carriers or diluents mentioned for use in the formulations for oral administration. The compounds may be dissolved in water, polyethylene glycol, propylene glycol, ethanol, corn oil, cottonseed oil, peanut oil, sesame oil, benzyl alcohol, sodium chloride, and/or various buffers. Other adjuvants and modes of administration are well and widely known in the pharmaceutical art.

[0526] Alternatively, the active ingredients may be formulated in a cream with an oil-in-water cream base. If desired, the aqueous phase of the cream base may include, for example at least 30% w/w of a polyhydric alcohol such as propylene glycol, butane-1,3-diol, mannitol, sorbitol, glycerol, polyethylene glycol and mixtures thereof. The topical

formulation may desirably include a compound, which enhances absorption or penetration of the active ingredient through the skin or other affected areas. Examples of such dermal penetration enhancers include dimethylsulfoxide and related analogs. The compounds of this present disclosure can also be administered by a transdermal device. In one embodiment, topical administration will be accomplished using a patch either of the reservoir and porous membrane type or of a solid matrix variety. In either case, the active agent is delivered continuously from the reservoir or microcapsules through a membrane into the active agent permeable adhesive, which is in contact with the skin or mucosa of the recipient. If the active agent is absorbed through the skin, a controlled and predetermined flow of the active agent is administered to the recipient. In the case of microcapsules, the encapsulating agent may also function as the membrane. The transdermal patch may include the compound in a suitable solvent system with an adhesive system, such as an acrylic emulsion, and a polyester patch. The oily phase of the emulsions of this present disclosure may be constituted from known ingredients in a known manner. While the phase may comprise merely an emulsifier, it may comprise a mixture of at least one emulsifier with a fat or oil or with both a fat and an oil. In one embodiment, a hydrophilic emulsifier is included together with a lipophilic emulsifier, which acts as a stabilizer. The phase may, for example, include both an oil and a fat. Together, the emulsifier(s) with or without stabilizer(s) make-up the so-called emulsifying wax, and the wax together with the oil and fat make up the so-called emulsifying ointment base, which forms the oily, dispersed phase of the cream formulations. Emulsifiers and emulsion stabilizers suitable for use in the formulation of the present disclosure include Tween 60, Span 80, cetostearyl alcohol, myristyl alcohol, glyceryl monostearate, and sodium lauryl sulfate, among others. The choice of suitable oils or fats for the formulation is based on achieving the desired cosmetic properties, since the solubility of the active compound in most oils likely to be used in pharmaceutical emulsion formulations is very low. Thus, the cream may, for example, be a non-greasy, non-staining and washable product with suitable consistency to avoid leakage from tubes or other containers. Straight or branched chain, mono- or dibasic alkyl esters such as di-isoadipate, isocetyl stearate, propylene glycol diester of coconut fatty acids, isopropyl myristate, decyl oleate, isopropyl palmitate, butyl stearate, 2-ethylhexyl palmitate or a blend of branched chain esters may be used. These may be used alone or in

combination depending on the properties required. Alternatively, high melting point lipids such as white soft paraffin and/or liquid paraffin or other mineral oils can be used.

[0527] A pharmaceutical composition may be formulated as inhaled formulations, including sprays, mists, or aerosols. For inhalation formulations, the compounds provided herein may be delivered via any inhalation methods known to those skilled in the art. Such inhalation methods and devices include, but are not limited to, metered dose inhalers with propellants such as CFC or HFA or propellants that are physiologically and environmentally acceptable. Other suitable devices are breath operated inhalers, multidose dry powder inhalers and aerosol nebulizers. Aerosol formulations for use in the subject method typically include propellants, surfactants and co-solvents and may be filled into conventional aerosol containers that are closed by a suitable metering valve.

[0528] Formulations suitable for inhalation or insufflation include solutions and suspensions in pharmaceutically acceptable aqueous or organic solvents, or mixtures thereof, and powders. The liquid or solid compositions may contain suitable pharmaceutically acceptable excipients as describe above. The compositions may be administered by oral or nasal respiratory route for local or systemic effect. Compositions may be nebulized by use of inert gases or vaporized, and breathed directly from the nebulizing/vaporizing device or the nebulizing device may be attached to a facemask tent or intermittent positive pressure-breathing machine.

[0529] Inhalant compositions may comprise liquid or powdered compositions containing the active ingredient that are suitable for nebulization and intrabronchial use, or aerosol compositions administered via an aerosol unit dispensing metered doses. Suitable liquid compositions comprise the active ingredient in an aqueous, pharmaceutically acceptable inhalant solvent, e.g., isotonic saline or bacteriostatic water. The solutions are administered by means of a pump or squeeze-actuated nebulized spray dispenser, or by any other conventional means for causing or enabling the requisite dosage amount of the liquid composition to be inhaled into the patient's lungs. Suitable formulations, wherein the carrier is a liquid, for administration, as for example, a nasal spray or as nasal drops, include aqueous or oily solutions of the active ingredient.

[0530] Formulations or compositions suitable for nasal administration, wherein the carrier is a solid, include a coarse powder having a particle size, for example, in the range of 20 to 500 microns which is administered in the manner in which snuff is administered

(i.e., by rapid inhalation through the nasal passage from a container of the powder held close up to the nose). Suitable powder compositions include, by way of illustration, powdered preparations of the active ingredient thoroughly intermixed with lactose or other inert powders acceptable for intrabronchial administration. The powder compositions can be administered via an aerosol dispenser or encased in a breakable capsule which may be inserted by the patient into a device that punctures the capsule and blows the powder out in a steady stream suitable for inhalation.

[0531] Pharmaceutical compositions may be formulated as sustained release formulations (i.e., a formulation such as a capsule that effects a slow release of modulator following administration). Such formulations may generally be prepared using well known technology and administered by, for example, oral, rectal or subcutaneous implantation, or by implantation at the desired target site. Carriers for use within such formulations are biocompatible, and may also be biodegradable; preferably the formulation provides a relatively constant level of modulator release. The amount of modulator contained within a sustained release formulation depends upon, for example, the site of implantation, the rate and expected duration of release and the nature of the condition to be treated or prevented.

[0532] In one example, the pharmaceutical formulations provided herein can include one or more additional active agent (i.e., other biologically active ingredient). In one example, the additional active agent is selected from known drugs approved for the treatment of a coagulation disorder, such as hemophilia A. For example, the pharmaceutical formulation can further include a blood coagulation factor.

[0533] In another example, the pharmaceutical formulation further contains a blood coagulation factor selected from FVIII and FIX. In another example, the pharmaceutical formulation further includes FVIII. In another example, the pharmaceutical formulation further includes FIX. In another example, the pharmaceutical formulation includes desmopressin (DDVAP).

[0534] Pharmaceutical compositions may be formulated with an agent to improve bioavailability, such as an organic solvent. For example, Cremophor EL.RTM. (Product No. 00647/1/63; BASF Aktiengesellschaft, Germany) is a polyethoxylated castor oil which is prepared by reacting 35 moles of ethylene oxide with each mole of castor oil. It may be used to stabilize emulsions of non-polar materials in aqueous systems.

Alternatively, peptide, peptide derivative or dual peptide may be incorporated within or bound to a proteinaceous micro or nano-particle for improved bioavailability. Suitable micro- and nano-particles are described in U.S. Pat. No. 5,439,686 (Desai et al; Vivorx Pharmaceuticals, Inc., CA) and U.S. Pat. No. 5,498,421 (Grinstaff et al; Vivorx Pharmaceuticals, Inc., CA). Suitably, the proteinaceous nano-particle comprises human serum albumin, particularly human serum albumin or a recombinant form thereof. WO 2007/077561 (Gabbai; Do-Coop Technologies Ltd., Israel) describe another suitable carrier comprising nanostructures and a liquid, referred to therein as Neowater.TM.

[0535] For veterinary use, a compound of the present disclosure is administered as a suitably acceptable formulation in accordance with normal veterinary practice and the veterinary surgeon will determine the dosing regimen and route of administration which will be most appropriate for a particular animal. For administration to non-human animals, the composition may be added to the animal feed or drinking water. It may be convenient to formulate the animal feed and drinking water compositions so that the animal takes in a therapeutically appropriate quantity of the composition along with its diet. It may also be convenient to present the composition as a premix for addition to the feed or drinking water.

Methods

[0536] The invention further provides a method (e.g., an *in vitro* or an *in vivo* method) of increasing/enhancing the catalytic activity (kcat) of a blood coagulation factor (i.e., activating the blood coagulation factor). An exemplary method includes contacting the blood coagulation factor (e.g., FIXa or FVIIa) with a compound of the present disclosure according to any of the above embodiments. In one example, the blood coagulation factor useful in this method includes the amino acid sequence: MFCAG (SEQ ID NO: 1) (e.g., FIXa, FXa, FVIIa and thrombin). In one example, the blood coagulation factor being activated is FXa. In another example, the blood coagulation factor being activated is thrombin.

[0537] The invention further provides a method of increasing/enhancing the catalytic activity (kcat) of FIXa (e.g., *in vitro* or *in vivo*), the method includes contacting the FIXa with a compound of the present disclosure according to any of the above embodiments.

[0538] The invention further provides a method of increasing/enhancing the catalytic activity (kcat) of FVIIa (e.g., *in vitro* or *in vivo*), the method includes contacting the

FVIIa with a compound of the present disclosure according to any of the above embodiments.

[0539] In one example according to the above method, the compound interacts with the blood coagulation factor (e.g., binds to the blood coagulation factor) at a region corresponding to amino acid sequence: MFCAG (SEQ ID NO: 1).

[0540] In a further example, the blood coagulation factor being activated is FIXa (e.g., human or canine FIXa). In one example according to this embodiment, the compound of the present disclosure interacts with the FIXa at a region corresponding to amino acid sequence: YNNMFCAGFHE (SEQ ID NO: 2). In yet another example, the compound of the present disclosure interacts with the FIXa at a region corresponding to amino acid sequence: RSTKFTIYNNMFCAGFHEGGRDSCQG (SEQ ID NO: 3).

[0541] In another example according to any of the above embodiments, the method is an *in vitro* method involving measuring conversion of FX to FXa. Such method is also generally referred to as a "FXa generation assay". An exemplary FXa generation assay is described in Example 2.

[0542] In one embodiment the compound is used in an *in vitro* assay system useful for the identification of other candidate compounds with pro-coagulant activity (e.g., a competition assay). In one example, the compound of the present disclosure is used as a reference compound in such assay system. In another example, the compound is used in a binding competition experiment as a probe. In one example, the compound of the present disclosure is used as a probe to measure binding of a candidate compound to a polypeptide (e.g., FIXa, FVIIa, or peptide including amino acid sequence of SEQ ID NO: 1, 2, or 3). For this purpose, the compound of the present disclosure can be linked to a detection molecule, such as an antibody (e.g., ELISA assay), biotin, a fluorescent molecule, a phage particle (e.g., phage display competition assay), and the like.

[0543] The invention further provides a method (e.g., an *in vitro* or an *in vivo* method) for identifying a candidate compound (e.g., a candidate compound with pro-coagulant activity) (e.g., within a screening procedure for the identification of compounds with pro-coagulant activity), the method comprising contacting a peptide or polypeptide comprising the amino acid sequence of SEQ ID NO: 1, 2, or 3 with a compound of the present disclosure according to any of the above embodiments.

Pharmaceutical Methods

- [0544] The present invention further provides methods for treating bleeding diathesis in a mammalian subject (e.g., a human patient) using the compounds or conjugates or the present disclosure.

Pharmaceutical Method 1

- [0545] In some embodiments, the present disclosure provides a method for treating bleeding diathesis in a mammalian subject (e.g., a human patient), comprising: administering to the subject in need thereof a therapeutically effective amount of a compound of the present disclosure or a pharmaceutical composition comprising a compound of the present disclosure.
- [0546] In some embodiments of pharmaceutical method 1, the compound or the pharmaceutical composition containing the compound is administered to the subject orally. In other embodiments, the compound or the pharmaceutical composition containing the compound is administered to the subject parenterally, e.g., intravenously or subcutaneously.

Pharmaceutical Method 2

- [0547] In some embodiments, the present disclosure provides a method for treating bleeding diathesis in a mammalian subject (e.g., a human patient), comprising: administering to the subject in need thereof a therapeutically effective amount of a conjugate of the present disclosure or a pharmaceutical composition comprising a conjugate of the present disclosure.
- [0548] In some embodiments of pharmaceutical method 2, the conjugate or the pharmaceutical composition comprising the conjugate is administered to the subject orally. In other embodiments, the conjugate or the pharmaceutical composition containing the conjugate is administered to the subject parenterally, e.g., intravenously or subcutaneously.
- [0549] In one example according to any of the above embodiments of pharmaceutical methods 1 and 2, the bleeding diathesis is caused by or associated with a blood coagulation disorder. A blood coagulation disorder can also be referred to as a coagulopathy. In a particular example, the blood coagulation disorder, which can be treated with a compound or conjugate of the current disclosure, is hemophilia or von

Willebrand disease (vWD). In a particular example, the blood coagulation disorder, which can be treated is hemophilia. In another example, the hemophilia is hemophilia A. In yet another example, the hemophilia is hemophilia B.

[0550] In another example, the type of bleeding associated with the bleeding diathesis is selected from hemarthrosis, muscle bleed, oral bleed, hemorrhage, hemorrhage into muscles, oral hemorrhage, trauma, trauma capitis, gastrointestinal bleeding, intracranial hemorrhage, intra-abdominal hemorrhage, intrathoracic hemorrhage, bone fracture, central nervous system bleeding, bleeding in the retropharyngeal space, bleeding in the retroperitoneal space, and bleeding in the iliopsoas sheath.

[0551] In another example, the subject suffering from bleeding diathesis is in need of treatment for surgery, including, e.g., surgical prophylaxis or peri-operative management. In one example, the surgery is selected from minor surgery and major surgery. Exemplary surgical procedures include tooth extraction, tonsillectomy, inguinal herniotomy, synovectomy, craniotomy, osteosynthesis, trauma surgery, intracranial surgery, intra-abdominal surgery, intrathoracic surgery, joint replacement surgery (e.g., total knee replacement, hip replacement, and the like), heart surgery, and caesarean section.

[0552] In some embodiments, the coagulation disorder is caused by a deficiency in at least one blood coagulation factor (e.g., FVIII). The current disclosure provides methods of treating a mammalian subject (e.g., a human subject) having a deficiency in at least one blood coagulation factor selected from von Willebrand Factor (vWF), FV, FVII, FVIII, FIX, FX, FXI, and activated forms thereof (e.g., for both the prophylaxis and for the treatment of acute bleeds).

Pharmaceutical Method 3

[0553] In some embodiments, the present disclosure provides a method for treating a coagulation disorder in a mammalian subject (e.g., a human patient) having a deficiency in at least one blood coagulation factor selected from von Willebrand Factor (vWF), FV, FVII, FVIII, FIX, FX, FXI, and activated forms thereof (e.g., for both the prophylaxis and for the treatment of acute bleeds), the method comprising: administering to the subject in need thereof a therapeutically effective amount of a compound of the present disclosure or a pharmaceutical composition comprising a compound of the present disclosure.

Pharmaceutical Method 4

[0554] In some embodiments, the present disclosure provides a method for treating a coagulation disorder in a mammalian subject (e.g., a human patient) having a deficiency in at least one blood coagulation factor selected from von Willebrand Factor (vWF), FV, FVII, FVIII, FIX, FX, FXI, and activated forms thereof (e.g., for both the prophylaxis and for the treatment of acute bleeds), the method comprising: administering to the subject in need thereof a therapeutically effective amount of a conjugate of the present disclosure or a pharmaceutical composition comprising a conjugate of the present disclosure.

[0555] In one example according to any of the above embodiments of pharmaceutical methods 1, 2, 3, and 4, the subject has a deficiency in FVIII. In another example, the subject responds to FVIII treatment.

Pharmaceutical Method 5

[0556] In some embodiments, the current disclosure provides a method of treating a mammalian subject (e.g., a human subject) having a deficiency in FVIII, the method includes administering to the subject a therapeutically effective amount of a compound of the current disclosure or a pharmaceutical composition containing a compound of the current disclosure.

Pharmaceutical Method 6

[0557] In some embodiments, the current disclosure provides a method of treating a mammalian subject (e.g., a human subject) having a deficiency in FVIII, the method includes administering to the subject a therapeutically effective amount of a conjugate of the current disclosure or a pharmaceutical composition containing a conjugate of the current disclosure.

[0558] Patients with a FVIII deficiency (i.e., hemophilia A) can develop inhibitor antibodies to FVIII. The biological activity of certain compounds or conjugates of the present disclosure is essentially not influenced by the presence of FVIII inhibitors, such as antibodies against FVIII. Hence, in one example according to any of the above embodiments of pharmaceutical methods 1-6, the mammalian subject (e.g., a human patient) suffering from a FVIII deficiency is a FVIII inhibitor patient (e.g., produces antibodies against FVIII). The magnitude of the antibody response to FVIII can be quantified using a functional inhibitor assay, such as that described in Kasper CK *et al.*

(1975) Proceedings: A more uniform measurement of factor VIII inhibitors. *Thromb. Diath. Haemorrh.* 34(2):612. FXI inhibitors could be quantified by an aPTT assay as described by Kasper CK *et al.*. Inhibitors of FV, FVII and FX could be quantified by a PT based assay following the procedure of Kasper CK *et al.*.

[0559] Inhibitor development (to FIX) is also known in FIX deficiency (i.e., hemophilia B). The biological activity (e.g., FXa generation assay activity) of certain compounds of the present disclosure is essentially not influenced by the presence of FIX inhibitors, such as antibodies against FIX. Since FV, FVII, FXI and FX deficiencies are very rare congenital disorders little is known about inhibitor development, although it is feasible that patients having such disorders might develop inhibitors. Treatment of inhibitor patients is contemplated by the current invention. Such inhibitor patients may have either a high titer response of greater than 5 BU or a low titer response of between 0.5 and 5 BU. Typically, the inhibitors are directed against FVIII and the patients have hemophilia A.

[0560] In one example according to any of the above embodiments, the mammalian subject is a human subject (i.e., a human patient). In another example according to any of the above embodiments, the mammalian subject (e.g, human patient) is concomitantly treated with at least one additional active agent, e.g., a drug approved for the treatment of coagulation disorders. In one example, the additional active agent is selected from a coagulation factor (e.g., FVIII or FIX) and desmopressin (DDVAP). In a particular example, the mammalian subject (e.g, human patient) is concomitantly treated with FVIII. In one example, the additional active agent is administered to the subject at the same time that the compound or conjugate of the present disclosure is administered to the subject. For example, the at least one additional active agent is contained in a pharmaceutical composition that also contains the compound or conjugate of the present disclosure. In another example, the additional active agent is administered to the subject at a different time but within the treatment period for the compound or conjugate of the present disclosure. For example, the additional active agent is administered alternately with the compound or conjugate of the present disclosure.

[0561] In another example, the mammalian subject (e.g, human patient) is concomitantly treated with FIX. Because the compounds or conjugates of the invention are capable of

activating FIXa, they could be used to pre-activate the FIXa polypeptide before administration of the FIXa to the subject.

[0562] In yet another example, the mammalian subject (e.g., human subject) is concomitantly treated with desmopressin (DDVAP).

[0563] The methods of the invention may be practiced on a subject in need of prophylactic treatment or on-demand treatment.

[0564] In various embodiments, the compounds of the present disclosure can be used as a replacement therapy (e.g., replacing FVIII treatment) in subjects in need of treatment for hemophilia A. In other embodiments, the compounds of the present disclosure can be used in conjunction with other therapies (e.g., FVIII or FIX therapy) in subjects in need of treatment for hemophilia A or B. In other embodiments, the compounds of the present disclosure are useful as a bypass therapy in subjects in need of treatment for hemophilia A (e.g., hemophilia A patients with inhibitors).

Administration of compounds

[0565] For oral and parenteral administration of compounds to patients, including human patients, the daily dosage level of the compound (e.g., peptide or peptide derivative) of the current disclosure will usually be from 2 to 2000 mg per adult (i.e. from about 0.03 to 30 mg/kg), administered in single or divided doses.

[0566] A unit dosage form (for example tablet or capsule) can contain from 2 mg to 2000 mg of active compound. The unit dosage form can be administered once, twice or more times per day as appropriate. The physician in any event will determine the actual dosage which will be most suitable for any individual patient and it will vary with the age, weight and response of the particular patient. The above dosages are exemplary of the average case. There can, of course, be individual instances where higher or lower dosage ranges are merited and such are within the scope of this invention.

Administration of polypeptide conjugates

[0567] The calculation of the required dosage for the polypeptide conjugates of the present disclosure is based upon the empirical finding that, on average, 1 IU of FVIII per kg body weight raises the plasma FVIII activity by approximately 2 IU/dL. The required dosage is determined using the following formula:

Required units = body weight (kg) x desired FVIII rise (IU/dL or % of normal) x 0.5 (IU/kg per IU/dL).

[0568] It will be understood, however, that the specific dose level for any particular patient will depend upon a variety of factors including the activity of the specific compound employed, the age, body weight, general health, sex, diet, time of administration, route of administration, and rate of excretion, drug combination and the severity of the particular disease undergoing therapy.

[0569] Dosage levels of the order of from about 0.005 mg to about 80 mg per kilogram of body weight per day are useful in the treatment of the diseases and conditions described herein (e.g., about 0.35 mg to about 5.6 g per human patient per day, based on an average adult person weight of 70 kg). The amount of active ingredient that may be combined with the carrier materials to produce a single dosage form will vary depending upon the host treated and the particular mode of administration. Dosage unit forms will generally contain between from about 1 mg to about 500 mg of an active ingredient. The daily dose can be administered in one to four doses per day. In the case of skin conditions, it may, for example, be applied as a topical preparation of compounds of this present disclosure on the affected area one to four times a day.

[0570] It will be understood, however, that the specific dose level for any particular patient will depend upon a variety of factors including the activity of the specific compound employed, the age, body weight, general health, sex, diet, time of administration, route of administration, and rate of excretion, drug combination and the severity of the particular disease undergoing therapy.

Definitions

[0571] The definitions and explanations below are for the terms as used throughout this entire document including both the specification and the claims. Throughout the specification and the appended claims, a given formula or name shall encompass all isomers thereof, such as stereoisomers, geometrical isomers, optical isomers, tautomers, and mixtures thereof where such isomers exist, as well as pharmaceutically acceptable salts and solvates thereof, such as hydrates.

[0572] It should be noted that, as used in this specification and the appended claims, the singular forms "a," "an," and "the" include plural referents unless the content clearly dictates otherwise. Thus, for example, reference to a composition containing "a

compound" includes a mixture of two or more compounds. It should also be noted that the term "or" is generally employed in its sense including "and/or" unless the content clearly dictates otherwise.

[0573] "Therapeutic dose," as used herein, means a dose that achieves a therapeutic goal, as described herein.

[0574] The phrase "therapeutically effective amount" as used herein means that amount of a compound, material, or composition of the present disclosure, which is effective for producing a desired therapeutic effect, at a reasonable benefit/risk ratio applicable to any medical treatment. For example, a "therapeutically effective amount" is an amount effective to reduce or lessen at least one symptom of the disease or condition being treated or to reduce or delay onset of one or more clinical markers or symptoms associated with the disease or condition, or to modify or reverse the disease process.

[0575] As used herein, the term "pharmaceutically acceptable" means approved by a regulatory agency of US or EU or other government or listed in the U.S. Pharmacopeia or other generally recognized pharmacopeia for use in humans. Hence, the term "pharmaceutically acceptable" refers to those properties and/or substances that are acceptable to a patient (e.g., human patient) from a toxicological and/or safety point of view.

[0576] The terms "treatment" or "treating" when referring to a disease or condition, means producing a desired therapeutic effect. Exemplary therapeutic effects include delaying onset or reducing at least one symptom associated with the disease, positively affecting (e.g., reducing or delaying onset) a clinical marker associated with the disease and slowing or reversing disease progression.

[0577] Where substituent groups are specified by their conventional chemical formulae, written from left to right, they equally encompass the chemically identical substituents, which would result from writing the structure from right to left. For example, "-CH₂O-" is intended to also recite "-OCH₂-".

[0578] The term "alkyl," by itself or as part of another substituent, means, unless otherwise stated, a straight or branched chain hydrocarbon radical having the number of carbon atoms designated (e.g., C₁-C₁₀ means one to ten carbon atoms). Typically, an alkyl group will have from 1 to 24 carbon atoms, for example having from 1 to 10 carbon atoms, from 1 to 8 carbon atoms or from 1 to 6 carbon atoms. A "lower alkyl" group is

an alkyl group having from 1 to 4 carbon atoms. The term “alkyl” includes di- and multivalent radicals. For example, the term “alkyl” includes “alkylene” wherever appropriate, e.g., when the formula indicates that the alkyl group is divalent or when substituents are joined to form a ring. Examples of alkyl radicals include, but are not limited to, methyl, ethyl, *n*-propyl, *iso*-propyl, *n*-butyl, *tert*-butyl, *iso*-butyl, *sec*-butyl, as well as homologs and isomers of, for example, *n*-pentyl, *n*-hexyl, *n*-heptyl and *n*-octyl.

[0579] The term “alkylene” by itself or as part of another substituent means a divalent (diradical) alkyl group, wherein alkyl is defined herein. “Alkylene” is exemplified, but not limited, by $-\text{CH}_2\text{CH}_2\text{CH}_2\text{CH}_2-$. Typically, an “alkylene” group will have from 1 to 24 carbon atoms, for example, having 10 or fewer carbon atoms (e.g., 1 to 8 or 1 to 6 carbon atoms). A “lower alkylene” group is an alkylene group having from 1 to 4 carbon atoms.

[0580] The term “alkenyl” by itself or as part of another substituent refers to a straight or branched chain hydrocarbon radical having from 2 to 24 carbon atoms and at least one double bond. A typical alkenyl group has from 2 to 10 carbon atoms and at least one double bond. In one embodiment, alkenyl groups have from 2 to 8 carbon atoms or from 2 to 6 carbon atoms and from 1 to 3 double bonds. Exemplary alkenyl groups include vinyl, 2-propenyl, 1-but-3-enyl, crotyl, 2-(butadienyl), 2,4-pentadienyl, 3-(1,4-pentadienyl), 2-isopentenyl, 1-pent-3-enyl, 1-hex-5-enyl and the like.

[0581] The term “alkynyl” by itself or as part of another substituent refers to a straight or branched chain, unsaturated or polyunsaturated hydrocarbon radical having from 2 to 24 carbon atoms and at least one triple bond. A typical “alkynyl” group has from 2 to 10 carbon atoms and at least one triple bond. In one aspect of the disclosure, alkynyl groups have from 2 to 6 carbon atoms and at least one triple bond. Exemplary alkynyl groups include prop-1-ynyl, prop-2-ynyl (i.e., propargyl), ethynyl and 3-butylnyl.

[0582] The terms “alkoxy,” “alkylamino” and “alkylthio” (or thioalkoxy) are used in their conventional sense, and refer to alkyl groups that are attached to the remainder of the molecule via an oxygen atom, an amino group, or a sulfur atom, respectively.

[0583] The term “heteroalkyl,” by itself or in combination with another term, means a stable, straight or branched chain hydrocarbon radical consisting of the stated number of carbon atoms (e.g., $\text{C}_2\text{-C}_{10}$, or $\text{C}_2\text{-C}_8$) and at least one heteroatom chosen, e.g., from N, O, S, Si, B and P (in one embodiment, N, O and S), wherein the nitrogen, sulfur and

phosphorus atoms are optionally oxidized, and the nitrogen atom(s) are optionally quaternized. The heteroatom(s) is/are placed at any interior position of the heteroalkyl group. Examples of heteroalkyl groups include, but are not limited to, $-\text{CH}_2-\text{CH}_2-\text{O}-\text{CH}_3$, $-\text{CH}_2-\text{CH}_2-\text{NH}-\text{CH}_3$, $-\text{CH}_2-\text{CH}_2-\text{N}(\text{CH}_3)-\text{CH}_3$, $-\text{CH}_2-\text{S}-\text{CH}_2-\text{CH}_3$, $-\text{CH}_2-\text{CH}_2-\text{S}(\text{O})-\text{CH}_3$, $-\text{CH}_2-\text{CH}_2-\text{S}(\text{O})_2-\text{CH}_3$, $-\text{CH}=\text{CH}-\text{O}-\text{CH}_3$, $-\text{CH}_2-\text{Si}(\text{CH}_3)_3$, $-\text{CH}_2-\text{CH}=\text{N}-\text{OCH}_3$, and $-\text{CH}=\text{CH}-\text{N}(\text{CH}_3)-\text{CH}_3$. Up to two heteroatoms can be consecutive, such as, for example, $-\text{CH}_2-\text{NH}-\text{OCH}_3$ and $-\text{CH}_2-\text{O}-\text{Si}(\text{CH}_3)_3$. Similarly, the term "heteroalkylene" by itself or as part of another substituent means a divalent radical derived from heteroalkyl, as exemplified, but not limited by, $-\text{CH}_2-\text{CH}_2-\text{S}-\text{CH}_2-\text{CH}_2-$ and $-\text{CH}_2-\text{S}-\text{CH}_2-\text{CH}_2-\text{NH}-\text{CH}_2-$. Typically, a heteroalkyl group will have from 3 to 24 atoms (carbon and heteroatoms, excluding hydrogen) (3- to 24-membered heteroalkyl). In another example, the heteroalkyl group has a total of 3 to 10 atoms (3- to 10-membered heteroalkyl) or from 3 to 8 atoms (3- to 8-membered heteroalkyl). The term "heteroalkyl" includes "heteroalkylene" wherever appropriate, e.g., when the formula indicates that the heteroalkyl group is divalent or when substituents are joined to form a ring.

[0584] The term "cycloalkyl" by itself or in combination with other terms, represents a saturated or unsaturated, non-aromatic carbocyclic radical having from 3 to 24 carbon atoms, for example, having from 3 to 12 carbon atoms (e.g., C_3 - C_8 cycloalkyl or C_3 - C_6 cycloalkyl). Examples of cycloalkyl include, but are not limited to, cyclopropyl, cyclobutyl, cyclopentyl, cyclohexyl, cycloheptyl, 1-cyclohexenyl, 3-cyclohexenyl, cycloheptyl and the like. The term "cycloalkyl" also includes bridged, polycyclic (e.g., bicyclic) structures, such as norbornyl, adamantyl and bicyclo[2.2.1]heptyl. The "cycloalkyl" group can be fused to at least one (e.g., 1 to 3) other ring selected from aryl (e.g., phenyl), heteroaryl (e.g., pyridyl) and non-aromatic (e.g., carbocyclic or heterocyclic) rings. When the "cycloalkyl" group includes a fused aryl, heteroaryl or heterocyclic ring, then the "cycloalkyl" group is attached to the remainder of the molecule via the carbocyclic ring.

[0585] The term "heterocycloalkyl," "heterocyclic," "heterocycle," or "heterocyclyl," by itself or in combination with other terms, represents a carbocyclic, non-aromatic ring (e.g., 3- to 8-membered ring and for example, 4-, 5-, 6- or 7-membered ring) containing at least one and up to 5 heteroatoms selected from, e.g., N, O, S, Si, B and P (for example, N, O and S), wherein the nitrogen, sulfur and phosphorus atoms are optionally oxidized,

and the nitrogen atom(s) are optionally quaternized (e.g., from 1 to 4 heteroatoms selected from nitrogen, oxygen and sulfur), or a fused ring system of 4- to 8-membered rings, containing at least one and up to 10 heteroatoms (e.g., from 1 to 5 heteroatoms selected from N, O and S) in stable combinations known to those of skill in the art. Exemplary heterocycloalkyl groups include a fused phenyl ring. When the "heterocyclic" group includes a fused aryl, heteroaryl or cycloalkyl ring, then the "heterocyclic" group is attached to the remainder of the molecule via a heterocycle. A heteroatom can occupy the position at which the heterocycle is attached to the remainder of the molecule. Exemplary heterocycloalkyl or heterocyclic groups of the present disclosure include morpholinyl, thiomorpholinyl, thiomorpholinyl S-oxide, thiomorpholinyl S,S-dioxide, piperazinyl, homopiperazinyl, pyrrolidinyl, pyrrolinyl, imidazolidinyl, tetrahydropyranyl, piperidinyl, tetrahydrofuranyl, tetrahydrothienyl, piperidinyl, homopiperidinyl, homomorpholinyl, homothiomorpholinyl, homothiomorpholinyl S,S-dioxide, oxazolidinonyl, dihydropyrazolyl, dihydropyrrolyl, dihydropyrazolyl, dihydropyridyl, dihydropyrimidinyl, dihydrofuryl, dihydropyranyl, tetrahydrothienyl S-oxide, tetrahydrothienyl S,S-dioxide, homothiomorpholinyl S-oxide, 1-(1,2,5,6-tetrahydropyridyl), 1-piperidinyl, 2-piperidinyl, 3-piperidinyl, 4-morpholinyl, 3-morpholinyl, tetrahydrofuran-2-yl, tetrahydrofuran-3-yl, tetrahydrothien-2-yl, tetrahydrothien-3-yl, 1-piperazinyl, 2-piperazinyl, and the like.

[0586] By "aryl" is meant a 5-, 6- or 7-membered, aromatic carbocyclic group having a single ring (e.g., phenyl) or being fused to other aromatic or non-aromatic rings (e.g., from 1 to 3 other rings). When the "aryl" group includes a non-aromatic ring (such as in 1,2,3,4-tetrahydronaphthyl) or heteroaryl group then the "aryl" group is bonded to the remainder of the molecule via an aryl ring (e.g., a phenyl ring). The aryl group is optionally substituted (e.g., with 1 to 5 substituents described herein). In one example, the aryl group has from 6 to 10 carbon atoms. Non-limiting examples of aryl groups include phenyl, 1-naphthyl, 2-naphthyl, quinoline, indanyl, indenyl, dihydronaphthyl, fluorenyl, tetralinyl, benzo[d][1,3]dioxolyl or 6,7,8,9-tetrahydro-5H-benzo[a]cycloheptenyl. In one embodiment, the aryl group is selected from phenyl, benzo[d][1,3]dioxolyl and naphthyl. The aryl group, in yet another embodiment, is phenyl.

- [0587] The term “arylalkyl” or “aralkyl” is meant to include those radicals in which an aryl group or heteroaryl group is attached to an alkyl group to create the radicals -alkyl-aryl and -alkyl-heteroaryl, wherein alkyl, aryl and heteroaryl are defined herein. Exemplary “arylalkyl” or “aralkyl” groups include benzyl, phenethyl, pyridylmethyl and the like.
- [0588] By “aryloxy” is meant the group -O-aryl, where aryl is as defined herein. In one example, the aryl portion of the aryloxy group is phenyl or naphthyl. The aryl portion of the aryloxy group, in one embodiment, is phenyl.
- [0589] The term “heteroaryl” or “heteroaromatic” refers to a polyunsaturated, 5-, 6- or 7-membered aromatic moiety containing at least one heteroatom (e.g., 1 to 5 heteroatoms, such as 1-3 heteroatoms) selected from N, O, S, Si and B (for example, N, O and S), wherein the nitrogen and sulfur atoms are optionally oxidized, and the nitrogen atom(s) are optionally quaternized. The “heteroaryl” group can be a single ring or be fused to other aryl, heteroaryl, cycloalkyl or heterocycloalkyl rings (e.g., from 1 to 3 other rings). When the “heteroaryl” group includes a fused aryl, cycloalkyl or heterocycloalkyl ring, then the “heteroaryl” group is attached to the remainder of the molecule via the heteroaryl ring. A heteroaryl group can be attached to the remainder of the molecule through a carbon- or heteroatom. In one example, the heteroaryl group has from 4 to 10 carbon atoms and from 1 to 5 heteroatoms selected from O, S and N. Non-limiting examples of heteroaryl groups include pyridyl, pyrimidinyl, quinolinyl, benzothienyl, indolyl, indolinyl, pyridazinyl, pyrazinyl, isoindolyl, isoquinolyl, quinazolinyl, quinoxalinyl, phthalazinyl, imidazolyl, isoxazolyl, pyrazolyl, oxazolyl, thiazolyl, indolizynyl, indazolyl, benzothiazolyl, benzimidazolyl, benzofuranyl, furanyl, thienyl, pyrrolyl, oxadiazolyl, thiadiazolyl, triazolyl, tetrazolyl, isothiazolyl, naphthyridinyl, isochromanlyl, chromanlyl, tetrahydroisoquinolinyl, isoindolinyl, isobenzotetrahydrofuranlyl, isobenzotetrahydrothienyl, isobenzothienyl, benzoxazolyl, pyridopyridyl, benzotetrahydrofuranlyl, benzotetrahydrothienyl, purinyl, benzodioxolyl, triazinyl, pteridinyl, benzothiazolyl, imidazopyridyl, imidazothiazolyl, dihydrobenzisoxazinyl, benzisoxazinyl, benzoxazinyl, dihydrobenzisothiazinyl, benzopyranlyl, benzothiopyranlyl, chromonyl, chromanonyl, pyridyl-N-oxide, tetrahydroquinolinyl, dihydroquinolinyl, dihydroquinolinonyl, dihydroisoquinolinonyl, dihydrocoumarinyl, dihydroisocoumarinyl, isoindolinonyl, benzodioxanyl, benzoxazolinonyl, pyrrolyl N-oxide, pyrimidinyl N-oxide,

pyridazinyl N-oxide, pyrazinyl N-oxide, quinolinyl N-oxide, indolyl N-oxide, indolinyl N-oxide, isoquinolyl N-oxide, quinazolinyl N-oxide, quinoxalinyl N-oxide, phthalazinyl N-oxide, imidazolyl N-oxide, isoxazolyl N-oxide, oxazolyl N-oxide, thiazolyl N-oxide, indolizinyl N-oxide, indazolyl N-oxide, benzothiazolyl N-oxide, benzimidazolyl N-oxide, pyrrolyl N-oxide, oxadiazolyl N-oxide, thiadiazolyl N-oxide, triazolyl N-oxide, tetrazolyl N-oxide, benzothiopyranyl S-oxide, benzothiopyranyl S,S-dioxide. Exemplary heteroaryl groups include imidazolyl, pyrazolyl, thiadiazolyl, triazolyl, isoxazolyl, isothiazolyl, imidazolyl, thiazolyl, oxadiazolyl, and pyridyl. Other exemplary heteroaryl groups include 1-pyrrolyl, 2-pyrrolyl, 3-pyrrolyl, 3-pyrazolyl, 2-imidazolyl, 4-imidazolyl, pyrazinyl, 2-oxazolyl, 4-oxazolyl, 2-phenyl-4-oxazolyl, 5-oxazolyl, 3-isoxazolyl, 4-isoxazolyl, 5-isoxazolyl, 2-thiazolyl, 4-thiazolyl, 5-thiazolyl, 2-furyl, 3-furyl, 2-thienyl, 3-thienyl, 2-pyridyl, 3-pyridyl, pyridin-4-yl, 2-pyrimidyl, 4-pyrimidyl, 5-benzothiazolyl, purinyl, 2-benzimidazolyl, 5-indolyl, 1-isoquinolyl, 5-isoquinolyl, 2-quinoxalinyl, 5-quinoxalinyl, 3-quinolyl, and 6-quinolyl. Substituents for each of the above noted aryl and heteroaryl ring systems are selected from the group of acceptable aryl group substituents described below.

[0590] For brevity, the term “aryl” when used in combination with other terms (*e.g.*, aryloxy, arylthioxy, arylalkyl) includes both aryl and heteroaryl rings as defined above.

[0591] Each of the above terms (*e.g.*, “alkyl,” “cycloalkyl,” “heteroalkyl,” “heterocycloalkyl,” “aryl” and “heteroaryl”) are meant to include both substituted and unsubstituted forms of the indicated radical. The term “substituted” for each type of radical is explained below. When a compound of the present disclosure includes more than one substituent, then each of the substituents is independently chosen.

[0592] The term “substituted” in connection with alkyl, alkenyl, alkynyl, cycloalkyl, heteroalkyl and heterocycloalkyl radicals (including those groups referred to as alkylene, heteroalkylene, heteroalkenyl, cycloalkenyl, heterocycloalkenyl, and the like) refers to one or more substituents, wherein each substituent is independently selected from, but not limited to, 3- to 10-membered heteroalkyl, C₃-C₁₀ cycloalkyl, 3- to 10-membered heterocycloalkyl, aryl, heteroaryl, -OR^a, -SR^a, =O, =NR^a, =N-OR^a, -NR^aR^b, -halogen, -SiR^aR^bR^c, -OC(O)R^a, -C(O)R^e, -C(O)OR^a, -C(O)NR^aR^b, -OC(O)NR^aR^b, -NR^cC(O)R^e, -NR^cC(O)NR^aR^b, -NR^cC(S)NR^aR^b, -NR^cC(O)OR^a, -NR^cC(NR^aR^b)=NR^d, -S(O)R^e, -S(O)₂R^e, -S(O)₂NR^aR^b, -NR^cS(O)₂R^a, -CN and -NO₂. R^a, R^b, R^c, R^d and R^e each

independently refer to hydrogen, C₁-C₂₄ alkyl (e.g., C₁-C₁₀ alkyl or C₁-C₆ alkyl), C₃-C₁₀ cycloalkyl, C₁-C₂₄ heteroalkyl (e.g., C₁-C₁₀ heteroalkyl or C₁-C₆ heteroalkyl), C₃-C₁₀ heterocycloalkyl, aryl, heteroaryl, arylalkyl and heteroarylalkyl, wherein, in one embodiment, R^e is not hydrogen. When two of the above R groups (e.g., R^a and R^b) are attached to the same nitrogen atom, they can be combined with the nitrogen atom to form a 5-, 6-, or 7-membered ring. For example, -NR^aR^b is meant to include pyrrolidinyl, N-alkyl-piperidinyl and morpholinyl.

[0593] The term “substituted” in connection with aryl and heteroaryl groups, refers to one or more substituents, wherein each substituent is independently selected from, but not limited to, alkyl (e.g., C₁-C₂₄ alkyl, C₁-C₁₀ alkyl or C₁-C₆ alkyl), cycloalkyl (e.g., C₃-C₁₀ cycloalkyl, or C₃-C₈ cycloalkyl), alkenyl (e.g., C₁-C₁₀ alkenyl or C₁-C₆ alkenyl), alkynyl (e.g., C₁-C₁₀ alkynyl or C₁-C₆ alkynyl), heteroalkyl (e.g., 3- to 10-membered heteroalkyl), heterocycloalkyl (e.g., C₃-C₈ heterocycloalkyl), aryl, heteroaryl, -R^a, -OR^a, -SR^a, =O, =NR^a, =N-OR^a, -NR^aR^b, -halogen, -SiR^aR^bR^c, -OC(O)R^a, -C(O)R^e, -C(O)OR^a, -C(O)NR^aR^b, -OC(O)NR^aR^b, -NR^cC(O)R^e, -NR^cC(O)NR^aR^b, -NR^cC(S)NR^aR^b, -NR^cC(O)OR^a, -NR^cC(NR^aR^b)=NR^d, -S(O)R^e, -S(O)₂R^e, -S(O)₂NR^aR^b, -NR^cS(O)₂R^a, -CN, -NO₂, -N₃, -CH(Ph)₂, fluoro(C₁-C₄)alkoxy, and fluoro(C₁-C₄)alkyl, in a number ranging from zero to the total number of open valences on the aromatic ring system, wherein R^a, R^b, R^c, R^d and R^e each independently refer to hydrogen, C₁-C₂₄ alkyl (e.g., C₁-C₁₀ alkyl or C₁-C₆ alkyl), C₃-C₁₀ cycloalkyl, C₁-C₂₄ heteroalkyl (e.g., C₁-C₁₀ heteroalkyl or C₁-C₆ heteroalkyl), C₃-C₁₀ heterocycloalkyl, aryl, heteroaryl, arylalkyl and heteroarylalkyl, wherein, in one embodiment, R^e is not hydrogen. When two R groups (e.g., R^a and R^b) are attached to the same nitrogen atom, they can be combined with the nitrogen atom to form a 5-, 6-, or 7-membered ring. For example, -NR^aR^b is meant to include pyrrolidinyl, N-alkyl-piperidinyl and morpholinyl.

[0594] The term “substituted” in connection with aryl and heteroaryl groups also refers to one or more fused ring(s), in which two hydrogen atoms on adjacent atoms of the aryl or heteroaryl ring are optionally replaced with a substituent of the formula -T-C(O)-(CRR')_q-U-, wherein T and U are independently -NR-, -O-, -CRR'- or a single bond, and q is an integer from 0 to 3. Alternatively, two of the hydrogen atoms on adjacent atoms of the aryl or heteroaryl ring can optionally be replaced with a substituent of the formula -A-(CH₂)_t-B-, wherein A and B are independently -CRR'-, -O-, -NR-, -S-, -S(O)-,

-S(O)₂-, -S(O)₂NR'- or a single bond, and r is an integer from 1 to 4. One of the single bonds of the ring so formed can optionally be replaced with a double bond. Alternatively, two of the hydrogen atoms on adjacent atoms of the aryl or heteroaryl ring can optionally be replaced with a substituent of the formula -(CRR')_s-X-(CR''R''')_d-, where s and d are independently integers from 0 to 3, and X is -O-, -NR'-, -S-, -S(O)-, -S(O)₂-, or -S(O)₂NR'-, wherein the substituents R, R', R'' and R''' in each of the formulas above are independently selected from hydrogen and (C₁-C₆)alkyl.

[0595] The terms "halo" or "halogen," by themselves or as part of another substituent, mean at least one of fluorine, chlorine, bromine and iodine.

[0596] By "haloalkyl" is meant an alkyl radical, wherein alkyl is as defined above and wherein at least one hydrogen atom is replaced by a halogen atom. The term "haloalkyl," is meant to include monohaloalkyl and polyhaloalkyl. For example, the term "halo(C₁-C₄)alkyl" or "(C₁-C₄)haloalkyl" is meant to include, but not limited to, chloromethyl, 1-bromoethyl, fluoromethyl, difluoromethyl, trifluoromethyl, 1,1,1-trifluoroethyl and 4-chlorobutyl, 3-bromopropyl.

[0597] As used herein, the term "acyl" describes the group -C(O)R^e, wherein R^e is selected from hydrogen, C₁-C₂₄ alkyl (e.g., C₁-C₁₀ alkyl or C₁-C₆ alkyl), C₁-C₂₄ alkenyl (e.g., C₁-C₁₀ alkenyl or C₁-C₆ alkenyl), C₁-C₂₄ alkynyl (e.g., C₁-C₁₀ alkynyl or C₁-C₆ alkynyl), C₃-C₁₀ cycloalkyl, C₁-C₂₄ heteroalkyl (e.g., C₁-C₁₀ heteroalkyl or C₁-C₆ heteroalkyl), C₃-C₁₀ heterocycloalkyl, aryl, heteroaryl, arylalkyl and heteroarylalkyl. In one embodiment, R^e is not hydrogen.

[0598] By "alkanoyl" is meant an acyl radical -C(O)-Alk-, wherein Alk is an alkyl radical as defined herein. Examples of alkanoyl include acetyl, propionyl, butyryl, isobutyryl, valeryl, isovaleryl, 2-methyl-butyryl, 2,2-dimethylpropionyl, hexanoyl, heptanoyl, octanoyl and the like.

[0599] As used herein, the term "heteroatom" includes oxygen (O), nitrogen (N), sulfur (S), silicon (Si), boron (B) and phosphorus (P). In one embodiment, heteroatoms are O, S and N.

[0600] By "oxo" is meant the group =O.

[0601] By "sulfonamide" is meant a group having the formula -S(O)₂NRR, where each of the R variables are independently selected from the variables listed below for R.

- [0602] The symbol "R" is a general abbreviation that represents a substituent group as described herein. Exemplary substituent groups include alkyl, alkenyl, alkynyl, cycloalkyl, heteroalkyl, aryl, heteroaryl and heterocycloalkyl groups, each as defined herein.
- [0603] As used herein, the terms "aromatic ring" and "non-aromatic ring" are consistent with the definitions commonly used in the art. For example, aromatic rings include phenyl and pyridyl. Non-aromatic rings include cyclohexanes.
- [0604] As used herein, the term "fused ring system" means at least two rings, wherein each ring has at least 2 atoms in common with another ring. "Fused ring systems can include aromatic as well as non-aromatic rings. Examples of "fused ring systems" are naphthalenes, indoles, quinolines, chromenes and the like. Likewise, the term "fused ring" refers to a ring that has at least two atoms in common with the ring to which it is fused.
- [0605] The term compound and molecule are used interchangeably. Other forms contemplated by the invention when the word "molecule" or "compound" is employed are salts, prodrugs, solvates, tautomers, stereoisomers and mixtures of stereoisomers. The compounds of this invention can be in the form of a pharmaceutically acceptable salt.
- [0606] The term "pharmaceutically acceptable salt" means a salt of the compounds of the present disclosure, which may be prepared with relatively nontoxic acids or bases, depending on the particular substituents found on the compounds described herein. When compounds of the present disclosure contain relatively acidic functionalities (e.g., -COOH group), base addition salts can be obtained by contacting the compound (e.g., neutral form of such compound) with a sufficient amount of the desired base, either neat or in a suitable inert solvent. Examples of pharmaceutically acceptable base addition salts include lithium, sodium, potassium, calcium, ammonium, organic amino, magnesium and aluminum salts and the like. When compounds of the present disclosure contain relatively basic functionalities (e.g., amines), acid addition salts can be obtained, e.g., by contacting the compound (e.g., neutral form of such compound) with a sufficient amount of the desired acid, either neat or in a suitable inert solvent. Examples of pharmaceutically acceptable acid addition salts include those derived from inorganic acids like hydrochloric, trifluoroacetic, hydrobromic, nitric, carbonic, monohydrogencarbonic, phosphoric, diphosphoric, monohydrogenphosphoric,

dihydrogenphosphoric, sulfuric, monohydrogensulfuric, hydriodic and the like, as well as the salts derived from relatively nontoxic organic acids like formic, acetic, propionic, isobutyric, malic, maleic, malonic, benzoic, succinic, suberic, fumaric, lactic, mandelic, phthalic, benzenesulfonic, p-tolylsulfonic, citric, tartaric, methanesulfonic, 2-hydroxyethylsulfonic, salicylic, stearic and the like. Also included are salts of amino acids such as arginate and the like, and salts of organic acids like glucuronic or galactunoric acids and the like (see, for example, Berge *et al.*, *Journal of Pharmaceutical Science*, 1977, 66: 1-19). Certain specific compounds of the present disclosure contain both, basic and acidic, functionalities that allow the compounds to be converted into either base or acid addition salts.

[0607] The neutral forms of the compounds can be regenerated, for example, by contacting the salt with a base or acid and isolating the parent compound in the conventional manner. The parent form of the compound can differ from the various salt forms in certain physical properties, such as solubility in polar solvents, but otherwise the salts are equivalent to the parent form of the compound for the purposes of the present disclosure.

[0608] When a substituent includes a negatively charged oxygen atom "O⁻", e.g., in "-COO⁻", then the formula is meant to optionally include a proton (i.e., H⁺) or an organic or inorganic cationic counterion (e.g., Na⁺). In one example, the resulting salt form of the compound is pharmaceutically acceptable. Further, when a compound of the present disclosure includes an acidic group, such as a carboxylic acid group, e.g., written as the substituent "-COOH", "-CO₂H" or "-C(O)₂H", then the formula is meant to optionally include the corresponding "de-protonated" form of that acidic group, e.g., "-COO⁻", "-CO₂⁻" or "-C(O)₂⁻", respectively.

[0609] In addition to salt forms, the present disclosure provides compounds, which are in a prodrug form. Prodrugs of the compounds described herein are those compounds that readily undergo chemical changes under physiological conditions to provide the compounds of the present disclosure. Non-limiting examples of "pharmaceutically acceptable derivative" or "prodrug" include pharmaceutically acceptable esters, phosphate esters or sulfonate esters thereof as well as other derivatives of a compound of this present disclosure which, upon administration to a recipient, is capable of providing, either directly or indirectly, a compound of this present disclosure. In one embodiment,

derivatives or prodrugs are those that increase the bioavailability of the compounds of this present disclosure when such compounds are administered to a mammal (e.g., by allowing an orally administered compound to be more readily absorbed into the blood stream) or which enhance delivery of the parent compound to a biological compartment (e.g., the brain or lymphatic system) relative to the parent species.

[0610] Prodrugs include a variety of esters (i.e., carboxylic acid ester). Ester groups, which are suitable as prodrug groups are generally known in the art and include benzyloxy, di(C₁-C₆)alkylaminoethoxy, acetoxymethyl, pivaloyloxymethyl, phthalidoyl, ethoxycarbonyloxyethyl, 5-methyl-2-oxo-1,3-dioxol-4-yl methyl, and (C₁-C₆)alkoxy esters, optionally substituted by N-morpholino and amide-forming groups such as di(C₁-C₆)alkylamino. For example, ester prodrug groups include C₁-C₆ alkoxy esters. Those skilled in the art will recognize various synthetic methodologies that may be employed to form pharmaceutically acceptable prodrugs of the compounds of the present disclosure (e.g., via esterification of a carboxylic acid group).

[0611] In an exemplary embodiment, the prodrug is suitable for treatment /prevention of those diseases and conditions that require the drug molecule to cross the blood brain barrier. In one embodiment, the prodrug enters the brain, where it is converted into the active form of the drug molecule. Additionally, prodrugs can be converted to the compounds of the present disclosure by chemical or biochemical methods in an *ex vivo* environment. For example, prodrugs can be slowly converted to the compounds of the present disclosure when placed in a transdermal patch reservoir together with a suitable enzyme or other chemical reagent.

[0612] Certain compounds of the present disclosure can exist in unsolvated forms as well as solvated forms, including hydrated forms. In general, the solvated forms are equivalent to unsolvated forms and are encompassed within the scope of the present disclosure. Certain compounds of the present disclosure can exist in multiple crystalline or amorphous forms ("polymorphs"). In general, all physical forms are of use in the methods contemplated by the present disclosure and are intended to be within the scope of the present disclosure. "Compound or a pharmaceutically acceptable salt, hydrate, polymorph or solvate of a compound" intends the inclusive meaning of "and/or", in that materials meeting more than one of the stated criteria are included, e.g., a material that is both a salt and a solvate is encompassed.

- [0613] The term "solvate" is intended to refer to a complex formed by combination of solute molecules or ions with solvent molecules. The solvent can be an organic compound, an inorganic compound, or a mixture of both. Exemplary solvents for the formation of solvates include, but are not limited to, methanol, N,N-dimethylformamide (DMF), tetrahydrofuran, dimethylsulfoxide, toluene, and water. In one embodiment, solvents having a higher boiling point, such as for example, DMF, DMA, and the like.
- [0614] The compounds of the present disclosure can contain unnatural proportions of atomic isotopes at one or more of the atoms that constitute such compounds. For example, the compounds can be radiolabeled with radioactive isotopes, such as for example tritium (^3H), iodine-125 (^{125}I) or carbon-14 (^{14}C). All isotopic variations of the compounds of the present disclosure, whether radioactive or not, are intended to be encompassed within the scope of the present disclosure. Compounds described herein, in which one or more of the hydrogen atoms are replaced with another stable isotope of hydrogen (i.e., deuterium) or a radioactive isotope (i.e., tritium), are part of this disclosure.
- [0615] The term "tautomer" is intended to refer to alternate forms of a compound that differ in the position of a proton, such as enol keto and imine enamine tautomers, or the tautomeric forms of heteroaryl groups containing a ring atom attached to both a ring NH moiety and a ring =N moiety such as pyrazoles, imidazoles, benzimidazoles, triazoles, and tetrazoles.
- [0616] The term "amino acid" within the scope of the present invention means alpha-amino acid unless otherwise specified. The term "amino acid" includes naturally occurring, and non-naturally occurring (i.e., non-proteinogenic) amino acids. The one and three letter abbreviations for naturally occurring amino acids are used herein (see, e.g., Lehninger, Biochemistry, 2nd ed., Worth Publishers, New York, 1995: 71-92). The term "amino acid" also includes stereoisomers (for example D-amino acids) and modifications of naturally occurring amino acids (e.g., alpha-N-alkylated amino acids; modified phenylalanine or tyrosine derivatives, and the like).
- [0617] Conventionally, L-amino acids are designated using upper case letters (e.g., the upper case letter "C" refers to L-cysteine), and D-amino acids are designated in lower case (e.g., the lower case letter "c" refers to D-cysteine). Where no indication of the isomer is given, both isomers are intended. For example, when the amino acid is referred

to with its name, both L- and D-amino acids are included (e.g., the term "cysteine" includes both, L-cysteine and D-cysteine).

[0618] The term "modified amino acid" refers to amino acids, which are altered with respect to their original structure, e.g., by substitution. Examples of modified amino acids include, e.g., alpha-N-alkylated amino acids, tyrosine derivatives (e.g., those in which the hydroxyl group is converted to an ether or ester group, or those in which the phenyl ring is substituted, e.g., with a halogen atom), phenylalanine derivatives (e.g., those in which the phenyl ring is substituted, e.g., with a halogen atom), lysine derivatives (e.g., those in which the NH₂ group is converted to an amide group or sulfonamide group), and amino acids, in which a carboxylic acid group is derivatized, e.g., esterified, converted to an amide group, and the like. In one example, the modified amino acid is a tyrosine residue modified at the hydroxyl group and having the formula Y-OR^b, wherein R^b is selected from straight or branched alkyl, e.g., (C₁-C₁₀)alkyl, straight or branched heteroalkyl, e.g., (C₁-C₁₀)heteroalkyl comprising from 1 to 5 heteroatoms selected from O, S and N, and a water-soluble polymer (e.g., a PEG moiety). In another example, the modified tyrosine is methoxy-tyrosine (Y-OMe).

[0619] Introduction of at least one non-natural amino acid, or modification of at least one amino acid can improve the stability or solubility of a peptide. It can further increase the resistance to protease degradation or alter the biological (e.g., *in vitro* biological) activity of the peptide.

[0620] Other modified and non-proteinogenic amino acids useful in the present invention are described, e.g., in Grant, *Synthetic Peptides: A Users Guide*, Oxford University Press, 1992, which is incorporated herein by reference in its entirety.

[0621] Non-proteinogenic amino acids may include but are not limited to norvaline (Nva), norleucine (Nle), 4-aminobutyric acid (gamma-Abu), 2-aminoisobutyric acid (Aib), 6-aminohexanoic acid (gamma-Ahx), ornithine (Orn), hydroxyproline (Hyp), sarcosine, citrulline, cysteic acid (Coh), cyclohexylalanine, methioninesulfoxide (Meo), methioninesulfone (Moo), homoserinemethylester (Hsm), propargylglycine (Eag), 5-fluorotryptophan (5Fw), 6-fluorotryptophan (6Fw), 3',4'-dimethoxyphenyl-alanine (Ear), 3',4'-difluorophenylalanine (Dff), 4'-fluorophenyl-alanine (Pff), 1-naphthyl-alanine (1-Nal), 1-naphthyl-alanine, 1-methyltryptophan (1Mw), penicillamine (Pen), homoserine (HSe). Further, such amino acids may include but are not limited to, alpha-amino

isobutyric acid, t-butylglycine, t-butylalanine, phenylglycine (Phg), benzothienylalanine (Bta), L-homo-cysteine (L-Hcys), N-methyl-phenylalanine (NMF), 2-thienylalanine (Thi), 3,3-diphenylalanine (Ebw), homophenylalanine (Hfe), s-benzyl-L-cysteine (Ece) or cyclohexylalanine (Cha). These and other non-proteinogenic amino acids may exist as D- or L-isomers.

[0622] The term "hydrophobic amino acids" or "amino acids having a hydrophobic side chain" means any amino acid with an alkyl side chains (e.g., A, L, I, V, and P), any amino acid with an aromatic side chains (e.g., F, W, and Y), and other amino acids having a non-polar side chain, e.g., those containing an ether or thioether group (i.e., M), or having a side chain, which is not ionized under physiological conditions. The term "hydrophobic amino acid" includes glycine and amino acids having a cycloalkyl side chain, such as P.

[0623] The term "polar uncharged amino acid," or "amino acid having a polar uncharged side chain," means any amino acid having a side chain incorporating a functional group (e.g., -OH or -SH group, amide -C(O)NH₂ group, etc.), wherein the functional group is not ionized under physiological conditions.

[0624] The term "hydrophobic or polar uncharged amino acid", or "amino acid having a hydrophobic or polar uncharged side chain", means any amino acid, which is not ionized under physiological conditions (i.e., other than basic and acidic amino acids). Exemplary "amino acids having a hydrophobic or polar uncharged side chain" include G, A, V, I, L, M, F, W, Y, S, T, N, Q and P (e.g., G, A, V, I, L, M, F, W, Y, S, T, N, and Q).

[0625] The term "acidic amino acid," or "amino acid having an acidic side chain," or any variation thereof, means any amino acid having a side chain with an ionizable (negatively charged) group, such as a carboxylic acid group. Examples of "acidic amino acids" include D and E.

[0626] The term "basic amino acid," or "amino acid having a basic side chain," or any variation thereof, means any amino acid having a side chain with an ionizable (positively charged) group, such as a primary or secondary amino group. Examples of "basic amino acids" include K, R, and H.

[0627] The term "peptide" or "peptide sequence" includes molecules in which amino acids are joined by peptide (-CO-NH-) linkages (also referred to as amide bonds).

[0628] "Polypeptide" and "protein" are used interchangeably herein and refer to a polymeric compound comprised of covalently linked amino acid residues. In one

example, the protein or polypeptide incorporates at least 500 amino acids. The term "polypeptide" as used herein further refers to a blood coagulation factor or a platelet targeting moiety as described herein.

[0629] The term "inverso-variant" of a peptide as used in this application means an enantiomer or mirror-image of a peptide. An "inverso-variant" is a peptide having the same amino acid sequence as its corresponding native peptide, but includes the corresponding D-amino acid for each L-amino acid in the native peptide, and the corresponding L-amino acid for each D-amino acid present in the native peptide (*i.e.*, the chirality for each amino acid is inverted). For example, the inverso variant of the native peptide kCLASYC is Kclasyc.

[0630] The term "retro-variant" of a peptide, means a peptide having the same amino acid sequence but in which the amino acids are assembled in opposite direction (reverse order) to the native peptide. For example, the retro-variant of the native peptide kCLASYC is CYSALCk.

[0631] The term "retro-inverso variant" as used in this application means a retro-peptide as described herein, which is also reversed in its amino acid sequence as in a retro-variant described herein. Thus, a "retro-inverso variant" of a peptide refers to a peptide is made up of amino acid residues which are assembled in the opposite direction and which have inverted chirality with respect to the native peptide to which it is retro-inverso modified. For example, the retro-inverso variant of the peptide kCLASYC is cysalcK.

[0632] A retro-inverso variant can maintain the topology of the side chains as in the native peptide sequence. For example, Guichard *et al.* (1994) *Proc. Natl. Acad. Sci. USA* 91:9765-9769 described that a retro-inverso peptide mimicked the structure and antigenic activity of the natural L-peptide IRGERA, but not of the D- and retro peptides. Such retro-inverso peptidomimetics may be made using methods known in the art, for example such as those described in Meziere *et al.* (1997) *J. Immunol.* 159, 3230-3237, incorporated herein by reference in its entirety. Partial retro-inverso peptide analogues are polypeptides in which only part of the sequence is reversed and replaced with enantiomeric amino acid residues. Processes for making such analogues are described, e.g., in European Patent EP0097994 to Pessi *et al.*, which is incorporated by reference herein in its entirety.

- [0633] Conventionally, where the amino acids are joined by peptide bonds, a peptide is represented such that the amino group at the N-terminus appears to the left and the carboxyl group at the C-terminus to the right. Peptides and peptide derivatives according to the present invention are represented in this manner.
- [0634] A "peptide derivative" contains a modification of one or more amino acid residues or a linker group or other covalently linked group. Examples of amino acid derivatives include N-acyl derivatives of the amino terminal or of another free amino group, esters of the carboxyl terminal or of another free carboxyl or hydroxy group, amides of the carboxyl terminal or of another free carboxyl group produced by reaction with ammonia or with a suitable amine, glycosylated derivatives, hydroxylated derivatives, nucleotidylated derivatives, ADP-ribosylated derivatives, pegylated derivatives, phosphorylated derivatives, biotinylated derivatives, derivatives conjugated to an antibody or antibody fragment, albumin, transferrin, HES, and the like. Also included among the chemical derivatives are those obtained by modification of the peptide bond-CO-NH-, for example by reduction to -CH₂-NH- or alkylation to -CO-N(alkyl)-. Other derivatives include amide bond bioisosteres, ketomethylene and hydroxyethylene derivatives, as well as thioesters, thioamides and the like.
- [0635] Other modifications include, e.g., acetylation, acylation, ADP-ribosylation, amidation, covalent attachment of flavin, covalent attachment of a heme moiety, covalent attachment of a nucleotide or nucleotide derivative, covalent attachment of phosphatidylinositol, cross-linking, cyclization, disulfide bond formation, demethylation, formation of covalent cross-links, formation of cysteine, formation of pyroglutamate, formylation, gamma-carboxylation, glycosylation, GPI anchor formation, hydroxylation, halogenation (e.g., iodination), methylation, myristoylation, oxidation, pegylation (Mei *et al.*, Blood 116:270-79 (2010), which is incorporated herein by reference in its entirety), proteolytic processing, phosphorylation, prenylation, racemization, selenoylation, sulfation, transfer-RNA mediated addition of amino acids to proteins such as arginylation, and ubiquitination.
- [0636] In one example, derivatisation is C-terminal amidation. C-terminal amidation of a peptide removes the negative charge of the C-terminal carboxyl group. Peptide derivatives having a C-terminal amide can be represented with "NH₂" at the C-terminus, for example KLTCLASYCWLF-NH₂. Another derivatisation is N-terminal acetylation.

This removes the positive charge at the N-terminus. Blocking of the C- or N-terminus, such as by C-terminal amidation or N-terminal acetylation, may improve proteolytic stability due to reduced susceptibility to exoproteolytic digestion.

[0637] "Administering," as used herein, means to give a pro-coagulant compound of the present disclosure, or pharmaceutical composition containing a pro-coagulant compound of the present disclosure, to a subject (e.g., human subject) in need thereof via a pharmaceutically acceptable route of administration. In some embodiments, the route of administration is parenteral. In one embodiment, the pro-coagulant compound of the present disclosure is administered subcutaneously. In some embodiments, the route of administration is intravenous, e.g., intravenous injection or intravenous infusion. In other embodiments, the route of administration is selected from subcutaneous, intramuscular, oral, nasal, and pulmonary administration. In other embodiments, the route of administration is selected from subcutaneous and intravenous. The pro-coagulant compounds of the invention can be administered as part of a pharmaceutical composition comprising at least one pharmaceutically acceptable carrier.

[0638] "Area under the plasma concentration versus time curve (AUC)," as used herein, is the same as the term of art in pharmacology, and is based upon the rate and extent of absorption of the compound following administration. AUC is determined over a specified time period, such as 12, 18, 24, 36, 48, or 72 hours, or for infinity using extrapolation based on the slope of the curve. Unless otherwise specified herein, AUC is determined for infinity. The determination of AUC may be carried out in a single subject, or in a population of subjects for which the average is calculated.

[0639] "Equivalent amount," as used herein, means the same amount of FVIII activity as expressed in International Units, which is independent of molecular weight of the polypeptide in question. One International Unit (IU) of FVIII activity corresponds approximately to the quantity of FVIII in one milliliter of normal human plasma. Several assays are available for measuring FVIII activity, including the European Pharmacopoeia chromogenic substrate assay and a one stage clotting assay.

[0640] "Dosing interval," as used herein, means the amount of time that elapses between multiple doses being administered to a subject. The comparison of dosing interval may be carried out in a single subject or in a population of subjects and then the average obtained in the population may be calculated.

[0641] "On-demand treatment," as used herein, means treatment that is intended to take place over a short course of time and is in response to an existing condition, such as a bleeding episode, or a perceived need such as planned surgery. Conditions that may require on-demand treatment include, e.g., a bleeding episode, hemarthrosis, muscle bleed, oral bleed, hemorrhage, hemorrhage into muscles, oral hemorrhage, trauma, trauma capitis, gastrointestinal bleeding, intracranial hemorrhage, intra-abdominal hemorrhage, intrathoracic hemorrhage, bone fracture, central nervous system bleeding, bleeding in the retropharyngeal space, bleeding in the retroperitoneal space, or bleeding in the iliopsoas sheath. The subject may be in need of surgical prophylaxis, peri-operative management, or treatment for surgery. Such surgeries include, e.g., minor surgery, major surgery, tooth extraction, tonsillectomy, inguinal herniotomy, synovectomy, total knee replacement, craniotomy, osteosynthesis, trauma surgery, intracranial surgery, intra-abdominal surgery, intrathoracic surgery, or joint replacement surgery.

[0642] Preferably, on-demand treatment resolves greater than 80% (greater than 80%, greater than 81%, greater than 82%, greater than 83%, greater than 84%, greater than 85%, greater than 86%, greater than 87%, greater than 88%, greater than 89%, greater than 90%, greater than 91%, greater than 92%, greater than 93%, greater than 94%, greater than 95%, greater than 96%, greater than 97%, greater than 98%, greater than 99%, or 100%) or 80-100%, 80-90%, 85-90%, 90-100%, 90-95%, or 95-100% of bleeds (e.g., spontaneous bleeds) in a single dose. Preferably, greater than 80% (greater than 81%, greater than 82%, greater than 83%, greater than 84%, greater than 85%, greater than 86%, greater than 87%, greater than 88%, greater than 89%, greater than 90%, greater than 91%, greater than 92%, greater than 93%, greater than 94%, greater than 95%, greater than 96%, greater than 97%, greater than 98%, or 100%) or 80-100%, 80-90%, 85-90%, 90-100%, 90-95%, or 95-100% of bleeding episodes are rated excellent or good by physicians after on-demand treatment. Preferably, greater than 5%, (greater than 6%, greater than 7%, greater than 8%, greater than 9%, greater than 10%, greater than 11%, greater than 12%, greater than 13%, greater than 14%, greater than 15%, greater than 16%, greater than 17%, greater than 18%, greater than 19%, greater than 20%), or 5-20%, 5-15%, 5-10%, 10-20%, or 10-15% of bleeding episodes are rated as fair by physicians after on-demand treatment.

[0643] "Prophylactic treatment," as used herein, means administering a pro-coagulant compound of the present disclosure to a subject over a course of time to increase the level of activity in a subject's plasma. Preferably, the increased level is sufficient to decrease the incidence of spontaneous bleeding or to prevent bleeding, e.g., in the event of an unforeseen injury. Preferably, during prophylactic treatment, the plasma protein level in the subject does not fall below the baseline level for that subject, or below the level that characterizes severe hemophilia.

[0644] Preferably, the prophylaxis regimen is "tailored" to the individual patient, preferably by determining PK data for each patient and administering the pro-coagulant compound of the present disclosure at a dosing interval that maintains a trough level equivalent to 1-3% FVIII activity. Adjustments may be made when a subject experiences unacceptable bleeding episodes defined as ≥ 2 spontaneous bleeding episodes over a rolling two-month period. In this case, adjustment will target trough levels of 3-5%. Preferably, prophylactic treatment results in prevention and control of bleeding, sustained control of bleeding, sustained protection from bleeding, and/or sustained benefit. Preferably, prophylaxis results in no spontaneous bleeding episodes within about 24, 36, 48, 72, or 96 hours (e.g., 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, or 96 hours, preferably within 72 hours), after treatment (e.g., the last injection). Preferably, prophylaxis results in greater than 30% (e.g., greater than 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, or 90%, preferably greater than 50%), mean reduction in annualized bleeding episodes (e.g., with once weekly dosing).

[0645] "Subject," as used herein means a human or a non-human mammal. Non-human mammals include, e.g., mice, dogs, primates, monkeys, cats, horses, cows, pigs, and other domestic animals and small animals. In one embodiment, the "subject" is a human patient.

[0646] "Therapeutic dose," as used herein, means a dose that achieves a therapeutic goal, as described herein. The therapeutic doses that may be used in the methods of the invention are about 10-100 mg/kg, more specifically, 10-20, 20-30, 30-40, 40-50, 50-60,

60-70, 70-80, 80-90, or 90-100 mg/kg, and more specifically, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, or 100 mg/kg.

[0647] Additional therapeutic doses that may be used in the methods of the invention are about 10 to about 150 mg/kg, more specifically, about 100-110, 110-120, 120-130, 130-140, 140-150 mg/kg, and more specifically, about 110, 115, 120, 125, 130, 135, 140, 145, or 150 mg/kg.

[0648] "About," as used herein for a range, modifies both ends of the range. Thus, "about 10-20" means "about 10 to about 20."

[0649] By "procoagulant activity" is meant the ability to promote thrombin generation and/or fibrin deposition in a suitable test system. Exemplary assays useful to measure the pro-coagulant activity of a compound or conjugate of the present are described herein and include, e.g., FXa generation assays (see, e.g., Example 2), thrombin generation assays (see, e.g., Example 3), and ROTEM assays (see, e.g., Example 4).

[0650] The term "antibody variant" or "modified antibody" includes an antibody which does not occur in nature and which has an amino acid sequence or amino acid side chain chemistry which differs from that of a naturally-derived antibody by at least one amino acid or amino acid modification as described herein. As used herein, the term "antibody variant" includes synthetic forms of antibodies which are altered such that they are not naturally occurring, e.g., antibodies that comprise at least two heavy chain portions but not two complete heavy chains (such as, domain deleted antibodies or minibodies); multispecific forms of antibodies (e.g., bispecific, trispecific, etc.) altered to bind to two or more different antigens or to different epitopes on a single antigen); heavy chain molecules joined to scFv molecules; single-chain antibodies; diabodies; triabodies; and antibodies with altered effector function and the like.

[0651] As used herein the term "scFv molecule" includes binding molecules which consist of one light chain variable domain (VL) or portion thereof, and one heavy chain variable domain (VH) or portion thereof, wherein each variable domain (or portion thereof) is derived from the same or different antibodies. scFv molecules preferably comprise an scFv linker interposed between the VH domain and the VL domain. ScFv molecules are known in the art and are described, e.g., in US patent 5,892,019, Ho et al. 1989. Gene 77:51; Bird et al. 1988 Science 242:423; Pantoliano et al. 1991.

Biochemistry 30:10117; Milenic et al. 1991. Cancer Research 51:6363; Takkinen et al. 1991. Protein Engineering 4:837.

[0652] A “scFv linker” as used herein refers to a moiety interposed between the VL and VH domains of the scFv. scFv linkers preferably maintain the scFv molecule in a antigen binding conformation. In one embodiment, a scFv linker comprises or consists of an scFv linker peptide. In certain embodiments, a scFv linker peptide comprises or consists of a gly-ser polypeptide linker. In other embodiments, a scFv linker comprises a disulfide bond.

[0653] As used herein, the term “gly-ser polypeptide linker” refers to a peptide that consists of glycine and serine residues. An exemplary gly/ser polypeptide linker comprises the amino acid sequence (Gly⁴ Ser)ⁿ. In one embodiment, n=1. In one embodiment, n=2. In another embodiment, n=3, i.e., (Gly⁴ Ser)³. In another embodiment, n=4, i.e., (Gly⁴ Ser)⁴. In another embodiment, n=5. In yet another embodiment, n=6. In another embodiment, n=7. In yet another embodiment, n=8. In another embodiment, n=9. In yet another embodiment, n=10. Another exemplary gly/ser polypeptide linker comprises the amino acid sequence Ser(Gly⁴Ser)ⁿ. In one embodiment, n=1. In one embodiment, n=2. In a preferred embodiment, n=3. In another embodiment, n=4. In another embodiment, n=5. In yet another embodiment, n=6.

[0654] The term “glycosylation” refers to the covalent linking of one or more carbohydrates to a polypeptide. Typically, glycosylation is a posttranslational event which can occur within the intracellular milieu of a cell or extract therefrom. The term glycosylation includes, for example, N-linked glycosylation (where one or more sugars are linked to an asparagine residue) and/or O-linked glycosylation (where one or more sugars are linked to an amino acid residue having a hydroxyl group (e.g., serine or threonine). In one embodiment, a molecule of the invention is glycosylated. In another embodiment, a molecule of the invention is aglycosylated. In yet another embodiment, a molecule of the invention has reduced glycosylation as compared to that in a wild type Fc region.

[0655] As used herein the term “disulfide bond” includes the covalent bond formed between two sulfur atoms. The amino acid cysteine comprises a thiol group that can form a disulfide bond or bridge with a second thiol group. In most naturally occurring IgG molecules, the CH1 and CL regions are linked by native disulfide bonds and the two

heavy chains are linked by two native disulfide bonds at positions corresponding to 239 and 242 using the Kabat numbering system (position 226 or 229, EU numbering system).

[0656] The term “vector” or “expression vector” is used herein to mean vectors used in accordance with the present invention as a vehicle for introducing into and expressing a desired polynucleotide in a cell. As known to those skilled in the art, such vectors may easily be selected from the group consisting of plasmids, phages, viruses and retroviruses. In general, vectors compatible with the instant invention will comprise a selection marker, appropriate restriction sites to facilitate cloning of the desired gene and the ability to enter and/or replicate in eukaryotic or prokaryotic cells.

[0657] Numerous expression vector systems may be employed to produce the conjugates of the invention. For example, one class of vector utilizes DNA elements which are derived from animal viruses such as bovine papilloma virus, polyoma virus, adenovirus, vaccinia virus, baculovirus, retroviruses (RSV, MMTV or MOMLV) or SV40 virus. Additionally, cells which have integrated the DNA into their chromosomes may be selected by introducing one or more markers which allow selection of transfected host cells. The marker may provide for prototrophy to an auxotrophic host, biocide resistance (e.g., antibiotics) or resistance to heavy metals such as copper. The selectable marker gene can either be directly linked to the DNA sequences to be expressed, or introduced into the same cell by cotransformation. In one embodiment, an inducible expression system can be employed. Additional elements may also be needed for optimal synthesis of mRNA. These elements may include signal sequences, splice signals, as well as transcriptional promoters, enhancers, and termination signals. In one embodiment, a secretion signal, e.g., any one of several well characterized bacterial leader peptides (e.g., pelB, phoA, or ompA), can be fused in-frame to the N terminus of a polypeptide of the invention to obtain optimal secretion of the polypeptide. (Lei et al. (1988), *Nature*, 331:543; Better et al. (1988) *Science*, 240:1041; Mullinax et al., (1990). *PNAS*, 87:8095).

[0658] The term “host cell” refers to a cell that has been transformed with a vector constructed using recombinant DNA techniques and encoding at least one heterologous gene. In descriptions of processes for isolation of proteins from recombinant hosts, the terms “cell” and “cell culture” are used interchangeably to denote the source of protein unless it is clearly specified otherwise. In other words, recovery of protein from the “cells” may mean either from spun down whole cells, or from the cell culture containing

both the medium and the suspended cells. The host cell line used for protein expression is most preferably of mammalian origin; those skilled in the art are credited with ability to preferentially determine particular host cell lines which are best suited for the desired gene product to be expressed therein. Exemplary host cell lines include, but are not limited to, DG44 and DUXB11 (Chinese Hamster Ovary lines, DHFR minus), HELA (human cervical carcinoma), CVI (monkey kidney line), COS (a derivative of CVI with SV40 T antigen), R1610 (Chinese hamster fibroblast) BALBC/3T3 (mouse fibroblast), HAK (hamster kidney line), SP2/O (mouse myeloma), P3x63-Ag3.653 (mouse myeloma), BFA-1c1BPT (bovine endothelial cells), RAJI (human lymphocyte) and 293 (human kidney). CHO cells are particularly preferred. Host cell lines are typically available from commercial services, the American Tissue Culture Collection or from published literature. The conjugates of the invention can also be expressed in non-mammalian cells such as bacteria or yeast or plant cells. In this regard it will be appreciated that various unicellular non-mammalian microorganisms such as bacteria can also be transformed; i.e. those capable of being grown in cultures or fermentation. Bacteria, which are susceptible to transformation, include members of the enterobacteriaceae, such as strains of *Escherichia coli* or *Salmonella*; *Bacillaceae*, such as *Bacillus subtilis*; *Pneumococcus*; *Streptococcus*, and *Haemophilus influenzae*. It will further be appreciated that, when expressed in bacteria, the polypeptides typically become part of inclusion bodies. The polypeptide conjugates must be isolated, purified and then assembled into functional molecules.

[0659] In addition to prokaryotes, eukaryotic microbes may also be used. *Saccharomyces cerevisiae*, or common baker's yeast, is the most commonly used among eukaryotic microorganisms although a number of other strains are commonly available including *Pichia pastoris*. For expression in *Saccharomyces*, the plasmid YRp7, for example, (Stinchcomb et al., (1979), *Nature*, 282:39; Kingsman et al., (1979), *Gene*, 7:141; Tschemper et al., (1980), *Gene*, 10:157) is commonly used. This plasmid already contains the TRP1 gene which provides a selection marker for a mutant strain of yeast lacking the ability to grow in tryptophan, for example ATCC No. 44076 or PEP4-1 (Jones, (1977), *Genetics*, 85:12). The presence of the *trp1* lesion as a characteristic of the yeast host cell genome then provides an effective environment for detecting transformation by growth in the absence of tryptophan.

- [0660] As used herein, the term "cleavage site" refers to a site recognized by an enzyme. In one embodiment, such an enzyme is one that is activated during the clotting cascade, such that cleavage of such sites occurs at the site of clot formation. Exemplary such sites include those recognized by thrombin, Factor XIa or Factor Xa
- [0661] In constructs that include more than one processing or cleavage site, it will be understood that such sites may be the same or different.
- [0662] "Blood coagulation factor" or "coagulation factor" as used herein means FVIIa, FVIII, or FIX.
- [0663] "Culture," "to culture" and "culturing," as used herein, means to incubate cells under in vitro conditions that allow for cell growth or division or to maintain cells in a
- [0664] "Polynucleotide" and "nucleic acid" are used interchangeably and refer to a polymeric compound comprised of covalently linked nucleotide residues. Polynucleotides may be DNA, cDNA, RNA, single stranded, or double stranded, vectors, plasmids, phage, or viruses.
- [0665] "Variant," as used herein, refers to a polynucleotide or polypeptide differing from the original polynucleotide or polypeptide, but retaining essential properties thereof, e.g., FVIII coagulant activity or Fc (FcRn binding) activity. Generally, variants are overall closely similar, and, in many regions, identical to the original polynucleotide or polypeptide. Variants include, e.g., polypeptide and polynucleotide fragments, deletions, insertions, and modified versions of original polypeptides.
- [0666] Naturally occurring variants are called "allelic variants," and refer to one of several alternate forms of a gene occupying a given locus on a chromosome of an organism (Genes II, Lewin, B., ed., John Wiley & Sons, New York (1985)). These allelic variants can vary at either the polynucleotide and/or polypeptide level and are included in the present invention. Alternatively, non-naturally occurring variants may be produced by mutagenesis techniques or by direct synthesis.
- [0667] Using known methods of protein engineering and recombinant DNA technology, variants may be generated to improve or alter the characteristics of the polypeptides. For instance, one or more amino acids can be deleted from the N-terminus or C-terminus of the secreted protein without substantial loss of biological function. The authors of Ron et al., *J. Biol. Chem.* 268: 2984-2988 (1993), incorporated herein by reference in its entirety, reported variant KGF proteins having heparin binding activity even after deleting 3, 8, or

27 amino-terminal amino acid residues. Similarly, Interferon gamma exhibited up to ten times higher activity after deleting 8-10 amino acid residues from the carboxy terminus of this protein. (Dobeli et al., J. Biotechnology 7:199-216 (1988), incorporated herein by reference in its entirety.)

[0668] Moreover, ample evidence demonstrates that variants often retain a biological activity similar to that of the naturally occurring protein. For example, Gayle and coworkers (J. Biol. Chem 268:22105-22111 (1993), incorporated herein by reference in its entirety) conducted extensive mutational analysis of human cytokine IL-1a. They used random mutagenesis to generate over 3,500 individual IL-1a mutants that averaged 2.5 amino acid changes per variant over the entire length of the molecule. Multiple mutations were examined at every possible amino acid position. The investigators found that "most of the molecule could be altered with little effect on either binding or biological activity." In fact, only 23 unique amino acid sequences, out of more than 3,500 nucleotide sequences examined, produced a protein that significantly differed in activity from wild-type.

[0669] As stated above, polypeptide variants include, e.g., modified polypeptides. Modifications include, e.g., acetylation, acylation, ADP-ribosylation, amidation, covalent attachment of flavin, covalent attachment of a heme moiety, covalent attachment of a nucleotide or nucleotide derivative, covalent attachment of a lipid or lipid derivative, covalent attachment of phosphatidylinositol, cross-linking, cyclization, disulfide bond formation, demethylation, formation of covalent cross-links, formation of cysteine, formation of pyroglutamate, formylation, gamma-carboxylation, glycosylation, GPI anchor formation, hydroxylation, iodination, methylation, myristoylation, oxidation, pegylation (Mei et al., Blood 116:270-79 (2010), which is incorporated herein by reference in its entirety), proteolytic processing, phosphorylation, prenylation, racemization, selenoylation, sulfation, transfer-RNA mediated addition of amino acids to proteins such as arginylation, and ubiquitination.

[0670] By a nucleic acid having a nucleotide sequence at least, for example, 95% "identical" to a reference nucleotide sequence, it is intended that the nucleotide sequence of the nucleic acid is identical to the reference sequence except that the nucleotide sequence may include up to five point mutations per each 100 nucleotides of the reference nucleotide sequence. In other words, to obtain a nucleic acid having a nucleotide

sequence at least 95% identical to a reference nucleotide sequence, up to 5% of the nucleotides in the reference sequence may be deleted or substituted with another nucleotide, or a number of nucleotides up to 5% of the total nucleotides in the reference sequence may be inserted into the reference sequence.

[0671] By a polypeptide having an amino acid sequence at least, for example, 95% "identical" to a query amino acid sequence of the present invention, it is intended that the amino acid sequence of the subject polypeptide is identical to the query sequence except that the subject polypeptide sequence may include up to five amino acid alterations per each 100 amino acids of the query amino acid sequence. In other words, to obtain a polypeptide having an amino acid sequence at least 95% identical to a query amino acid sequence, up to 5% of the amino acid residues in the subject sequence may be inserted, deleted, (indels) or substituted with another amino acid. These alterations of the reference sequence may occur at the amino or carboxy terminal positions of the reference amino acid sequence or anywhere between those terminal positions, interspersed either individually among residues in the reference sequence or in one or more contiguous groups within the reference sequence.

[0672] As a practical matter, whether any particular nucleic acid molecule or polypeptide is at least 85%, 90%, 95%, 96%, 97%, 98% or 99% identical to a nucleotide sequence or polypeptide of the present invention can be determined conventionally using known computer programs. A preferred method for determining the best overall match between a query sequence (reference or original sequence) and a subject sequence, also referred to as a global sequence alignment, can be determined using the FASTDB computer program based on the algorithm of Brutlag et al. (Comp. App. Biosci. (1990) 6:237-245), which is herein incorporated by reference in its entirety. In a sequence alignment the query and subject sequences are both DNA sequences. An RNA sequence can be compared by converting U's to T's. The result of said global sequence alignment is in percent identity. Preferred parameters used in a FASTDB alignment of DNA sequences to calculate percent identity are: Matrix=Unitary, k-tuple=4, Mismatch Penalty=1, Joining Penalty=30, Randomization Group Length=0, Cutoff Score=1, Gap Penalty=5, Gap Size Penalty 0.05, Window Size=500 or the length of the subject nucleotide sequence, whichever is shorter.

[0673] If the subject sequence is shorter than the query sequence because of 5' or 3' deletions, not because of internal deletions, a manual correction must be made to the results. This is because the FASTDB program does not account for 5' and 3' truncations of the subject sequence when calculating percent identity. For subject sequences truncated at the 5' or 3' ends, relative to the query sequence, the percent identity is corrected by calculating the number of bases of the query sequence that are 5' and 3' of the subject sequence, which are not matched/aligned, as a percent of the total bases of the query sequence. Whether a nucleotide is matched/aligned is determined by results of the FASTDB sequence alignment. This percentage is then subtracted from the percent identity, calculated by the above FASTDB program using the specified parameters, to arrive at a final percent identity score. This corrected score is what is used for the purposes of the present invention. Only bases outside the 5' and 3' bases of the subject sequence, as displayed by the FASTDB alignment, which are not matched/aligned with the query sequence, are calculated for the purposes of manually adjusting the percent identity score.

[0674] For example, a 90 base subject sequence is aligned to a 100 base query sequence to determine percent identity. The deletions occur at the 5' end of the subject sequence and therefore, the FASTDB alignment does not show a matched/alignment of the first 10 bases at 5' end. The 10 unpaired bases represent 10% of the sequence (number of bases at the 5' and 3' ends not matched/total number of bases in the query sequence) so 10% is subtracted from the percent identity score calculated by the FASTDB program. If the remaining 90 bases were perfectly matched the final percent identity would be 90%. In another example, a 90 base subject sequence is compared with a 100 base query sequence. This time the deletions are internal deletions so that there are no bases on the 5' or 3' of the subject sequence which are not matched/aligned with the query. In this case the percent identity calculated by FASTDB is not manually corrected. Once again, only bases 5' and 3' of the subject sequence which are not matched/aligned with the query sequence are manually corrected for. No other manual corrections are made for the purposes of the present invention.

[0675] If the subject sequence is shorter than the query sequence due to N- or C-terminal deletions, not because of internal deletions, a manual correction must be made to the results. This is because the FASTDB program does not account for N- and C-terminal

truncations of the subject sequence when calculating global percent identity. For subject sequences truncated at the N- and C-termini, relative to the query sequence, the percent identity is corrected by calculating the number of residues of the query sequence that are N- and C-terminal of the subject sequence, which are not matched/aligned with a corresponding subject residue, as a percent of the total bases of the query sequence. Whether a residue is matched/aligned is determined by results of the FASTDB sequence alignment. This percentage is then subtracted from the percent identity, calculated by the above FASTDB program using the specified parameters, to arrive at a final percent identity score. This final percent identity score is what is used for the purposes of the present invention. Only residues to the N- and C-termini of the subject sequence, which are not matched/aligned with the query sequence, are considered for the purposes of manually adjusting the percent identity score. That is, only query residue positions outside the farthest N- and C-terminal residues of the subject sequence.

[0676] For example, a 90 amino acid residue subject sequence is aligned with a 100 residue query sequence to determine percent identity. The deletion occurs at the N-terminus of the subject sequence and therefore, the FASTDB alignment does not show a matching/alignment of the first 10 residues at the N-terminus. The 10 unpaired residues represent 10% of the sequence (number of residues at the N- and C- termini not matched/total number of residues in the query sequence) so 10% is subtracted from the percent identity score calculated by the FASTDB program. If the remaining 90 residues were perfectly matched the final percent identity would be 90%. In another example, a 90 residue subject sequence is compared with a 100 residue query sequence. This time the deletions are internal deletions so there are no residues at the N- or C-termini of the subject sequence which are not matched/aligned with the query. In this case the percent identity calculated by FASTDB is not manually corrected. Once again, only residue positions outside the N- and C-terminal ends of the subject sequence, as displayed in the FASTDB alignment, which are not matched/aligned with the query sequence are manually corrected for. No other manual corrections are to made for the purposes of the present invention.

[0677] Having now described the present invention in detail, the same will be more clearly understood by reference to the following examples, which are included herewith

for purposes of illustration only and are not intended to be limiting of the invention. All patents and publications referred to herein are expressly incorporated by reference.

Example 1

Preparation of Compounds

[0678] Compounds of the present disclosure (i.e., peptides) were synthesized by solid phase peptide synthesis using 9-fluorenylmethoxycarbonyl/tertiary-butyl (Fmoc/tBu) chemistry. Heating was accomplished using a microwave oven or other means. In most cases, the peptides were synthesized in 0.1 mmol scale using NovaPEG Rink Amide resin (35 mL reaction vessel). Standard methods for resin load, amino acid coupling, Fmoc deprotection and washing steps were performed on a CEM Liberty peptide synthesizer, whereas the trifluoroacetic acid (TFA) cleavage of the peptide was performed manually. Briefly, 5 eq. Fmoc protected amino acids dissolved in *N,N*-dimethylformamide (DMF) were linked subsequently to the resin in the presence of 5 eq. 2(6-chloro-1H-benzotriazole-1-yl)-1,1,3,3-tetramethylaminium hexafluorophosphate (HCTU) and 10 eq. (diisopropylethylamine) DIPEA. The microwave method for the coupling step was single coupling at 75 °C (20W for 300 sec), except for cysteine and histidine at 50 °C (0W for 120 sec, 20W for 240 sec), and arginine was double coupled at 75 °C (0W for 1500 sec, 20W for 300 sec). The Fmoc deprotection was performed with 5% piperazine, 0.1M 1-hydroxybenzotriazole (HOBt) in DMF at 75 °C (45W for 30 sec, 45W for 180 sec). Most amino acids and coupling reagents were purchased from Novabiochem EMD.

[0679] Following the automated peptide synthesis, the peptides were cleaved from the resin with 95% TFA and 5% triisopropylsilane (TIPS) for 4 hrs. Peptides containing methionine were cleaved from the resin with a mixture of 95% TFA, 2.5% TIPS and 2.5% ethanedithiol (EDT). Next, the peptides were filtered into round bottom reaction flasks, and in case of the methionine containing peptides 2.5% bromotrimethylsilane (TMSBr) was added. The solvents were removed *in vacuo*, and the concentrates containing the peptides were precipitated and further triturated with ice cold diethyl ether (Et₂O). The synthesized peptides were confirmed by mass spectral analysis.

[0680] Some of the peptides required modifications prior to cleavage from the resin, e.g. the peptides containing lactam loops. Here the orthogonal protection groups allyloxycarbonyl (Alloc) and Allyl or methyltrityl (Mtt) and 2-phenylisopropyl (2-PhiPr)

were removed by Pd[P(Ph)₃]₄ or 1% TFA treatment, respectively. The subsequent lactam formation between the carboxylic acid and amine side chains occurred in the presence of 10 eq. benzotriazol-1-yl-oxytripyrrolidinophosphonium hexafluorophosphate (PyBOP) and 10 eq. DIPEA in DMF.

Peptide purification

[0681] The synthesized peptides were purified by preparative reverse phase high performance liquid chromatography (RP-HPLC) using Waters 600 controller and pump system with 2489 UV detector and fraction collector III. The purifications were typically performed on a Phenomenex Jupiter C18 10 micron 250 x 21.20 mm column with a flow rate of 20 mL/min. The acetonitrile/water (0.1% TFA) gradient was modified for each specific peptide based on hydrophobicity. The peptides were detected at two wavelengths 228 and 280 nm, and the fractions were further analyzed by liquid chromatography mass spectrometry (LC-MS). Fractions containing peptide of adequate purity were pooled, flash frozen and lyophilized.

Peptide characterization

[0682] The peptides were characterized by LC-MS (Agilent LC-MS TOF 6220 with 1200 series pump, auto handler and UV detection system). The LC separation was performed on a Phenomenex Jupiter C18 5 micron 250 x 2.00 mm column using a mobile phase of A (water + 0.08% formic acid + 0.02% trifluoroacetic acid) and B (acetonitrile + 0.08% formic acid + 0.02% trifluoroacetic acid). The general LC method had a gradient from 0-70% B over 12 min. Mass determination was achieved by electrospray ionization in positive mode. The purity of the peptides was determined by measuring the absorbance of UV light at 228 nm over the chromatogram.

Example 2

FXa Generation Assay

Primary Screening Assay

[0683] FXa generation was utilized to assess the capability of the compounds of the present disclosure to enhance the catalytic activities of FIXa and/or FVIIa.

- [0684] To measure the catalytic activity of FIXa, FIXa was mixed with FX and phospholipids in the presence of calcium chloride. Under these conditions FIXa cleaves the zymogen FX into the active form FXa.
- [0685] The generation of FXa is monitored by the change in absorbance of the reaction at 405 nm due to the presence of a cleavable FXa chromogenic substrate (S-2765; Z-D-Arg-Gly-Arg-pNA; cleavage of p-nitroaniline). The assay components were obtained from the Coatest SP FVIII FXa generation kit (Chromogenix) utilized in the industry to assess the activity of FVIII. The kit contains all the FXa generation components, however, FIXa and FX obtained from this kit are from a bovine origin and were replaced with purified human FIXa or human FVIIa and human FX (Haematologic Technologies).
- [0686] The assay was also modified to enhance the screening throughput. Instead of the sequential addition, all the assay components were mixed and added simultaneously to the purified compound to be assayed. A 125 μ l reaction mix of the assay components was prepared in the buffer supplied by the manufacturer (50 mM Tris pH 7.3, and 1% BSA). The reaction mix contained hFIXa (12 nM), hFX (120 nM), S-2765 substrate (720 μ M), calcium chloride (5 mM) and phospholipids (8.3 μ l of the mixture of highly purified phospholipid stock supplied) based on the kit recommendations. The reaction mix (125 μ l) was then added to 25 μ l of compound diluted in water in a Costar-3651 flat bottom 96-well plate. This resulted in a final reaction concentration of 10 nM hFIXa, 100 nM hFX, 600 μ M FXa substrate S-2765, and 4.17 mM calcium chloride. The absorbance was monitored over a period of 1 hour at 405 nm using a plate reader (Synergy 2, Biotek).
- [0687] The 405 nm absorbance data were then analyzed to obtain the slope of the first derivative which reflects the rate of change in absorbance. The first derivative slope data for each compound concentration were plotted against the compound concentration and fitted to a four-parameter equation to obtain the EC50 and Vmax values for each compound tested.
- [0688] Similarly, in order to measure the catalytic activity of FVIIa, a FXa generation assay measuring the ability of the compounds of the present disclosure to enhance the catalytic activity of FVIIa was established. The difference between the two assays was that the final concentration of 10 nM hFIXa was replaced with 10 nM hFVIIa. The remaining reagents and assay procedures described above were unchanged.

K_m/K_{cat}

[0689] To determine the K_m and K_{cat} of FIXa or FVIIa for the substrate FX, the FXa generation assay described above was used with the following modifications. A single peptide concentration was tested (typically one that gave the maximal rate of FXa generation) and the concentration of all assay components was similar to that described above with the exception of FX. Reactions were set up with varying concentrations of FX ranging between 400 and 0.8 nM and the rates of FXa generation were determined for each peptide at the different FX concentrations tested as described above. For each peptide assayed, the data were fitted to the following equation that gives the Michaelis-Menten constant (K_m) and the V_{max} for FIXa or FVIIa: $v = V_{max} [FX] / K_m + [FX]$, wherein v is the rate of FXa generation determined as described above. The catalytic constant (K_{cat}) was calculated by normalizing the V_{max} to the enzyme concentration.

[0690] Exemplary compounds of the present disclosure and their *in vitro* biological activities measured using the FXa generation assay as described above (using human FIXa) are summarized in Table 1, below. In Table 1, peptides are amidated (-CONH₂) at the C-terminus and have a free N-terminus, unless otherwise indicated.

Table 1

Exemplary Compounds of the Present Disclosure and their Activities in the FXa Generation Assay

Sequence	EC ₅₀ [μM]	V _{max}	
RRAPGKLQCLASYCWLFWTGIA (compound 25)	(+++)	(+++)	SEQ ID NO: 4
PLKWTASGCRWLGCQLARFAY (compound B)	(+)	(++)	SEQ ID NO: 5
LYTAWIKCQFARLPGCALSGRW (compound C)	>25	>0.6	SEQ ID NO: 6
Biotin-PEG ₂ -LYTAWIKCQFARLPGCALSGRW	>10	>1.5	SEQ ID NO: 7
RRAPG-k-NMeLeu-TCLASYCWLFWTGIA	(++)	(++++)	SEQ ID NO: 9
k-NMeLeu-TCLASYCWLFWTGIA (compound 10)	(+++)	(++)	SEQ ID NO: 10
RRAPGKLQCLASYCWLFWTGAA	(+)	(++++)	SEQ ID NO: 11
RRAPGKLQCLASYCWLFWTAIA	(+++)	(+++)	SEQ ID NO: 12
RRAPGKLQCLASYCWLFWAGIA	(++)	(++++)	SEQ ID NO: 13
RRAPGKLQCLASYCWLFATAIA	(+++)	(++++)	SEQ ID NO: 14

Sequence	EC ₅₀ [μM]	Vmax	
RRAPAKLQCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 15
RRAAGKLQCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 16
RAAPGKLQCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 17
ARAPGKLQCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 18
rRAAGKLTCLASYCWLFATGIA	(+++)	(++)	SEQ ID NO: 414
rRAAGKATCLASYCWLFATGIA	(++)	(++)	SEQ ID NO: 415
rRAAGKLTCLASACWLFATGIA	(+)	(++)	SEQ ID NO: 416
rRAAGKLTCLASYCWLAATGIA	>10	>0.8	SEQ ID NO: 417
rRAAGKATCLASACWLAATGIA	>10	>3	SEQ ID NO: 418
rRASGKLTCLASYCWLFWTGIA	(+++)	(++)	SEQ ID NO: 419
rRAPGKLTCLASYCWLFSTGIA	(+++)	(++)	SEQ ID NO: 420
rRAPGKSTCLASYCWLFWTGIA	(+++)	(++)	SEQ ID NO: 421
rRAPGKLTCLASSCWLFWTGIA	(++)	(++)	SEQ ID NO: 422
rRAPGKLTCLASYCWLSWTGIA	>5	>1.8	SEQ ID NO: 423
RRAPGKLQCLASYCWLFWTGI	(++)	(+++)	SEQ ID NO: 19
RRAPGKLQCLASYCWLFWTG	(+)	(+++)	SEQ ID NO: 20
RRAPGKLQCLASYCWLFWT	(+)	(+++)	SEQ ID NO: 21
RRAPGKLQCLASYCWLFW	(+)	(+++)	SEQ ID NO: 22
RRAPGKLQCLASYCWLF	>5	>1.1	SEQ ID NO: 23
RAPGKLQCLASYCWLFWTGIA	(+++)	(++)	SEQ ID NO: 24
APGKLQCLASYCWLFWTGIA	(+++)	(++)	SEQ ID NO: 25
PGKLQCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 26
GKLQCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 27
KLQCLASYCWLFWTGIA	(++)	(+++)	SEQ ID NO: 28
RRAPGKLTCLASYCWLFWTGIA (compound 4)	(+++)	(+++)	SEQ ID NO: 29
RRAPGKLQCLASYCWLFWTGLA	(+++)	(+++)	SEQ ID NO: 30
RRAPGKLQCLASYCWLFWTG-Nle-A	(+++)	(+++)	SEQ ID NO: 31
RRAPGKLQCLASYCWLFWTG-Tle-A	(+++)	(+++)	SEQ ID NO: 32
RRAPGKLQCLASYCWLFWTGFA	(+++)	(+++)	SEQ ID NO: 33
RRAPGKLQCLASYCWLFWTG-Cha-A	(++)	(++++)	SEQ ID NO: 34
RRAPGKLQCLASYCWLFWTG-(1-Nal)-A	(+++)	(+++)	SEQ ID NO: 35
RRAPGKLQCLASYCWLFWT-Aib-IA	(+)	(++)	SEQ ID NO: 36
RRAPGKLQCLASYCWLFWTGIAAAAAGAP	(++)	(++)	SEQ ID NO: 37
RRAPGKLQCLASYCWLFWTGIK	(+)	(++)	SEQ ID NO: 38
RRAPGKLQCLASYCWLFWTGKA	>5	>1	SEQ ID NO: 39
RRAPGKLQCLASYCWLFWTKIA	(+)	(++)	SEQ ID NO: 40
RRAPGKLQCLASYCWLFWKGIA	>5	>1	SEQ ID NO: 41

Sequence	EC ₅₀ [μM]	Vmax	
RRAPGKLQCLASYCWLFKTGIA	>10	>0.2	SEQ ID NO: 42
RRAPKKLQCLASYCWLFWTGIA	(+)	(++)	SEQ ID NO: 43
RRAKGKLQCLASYCWLFWTGIA	(+)	(++)	SEQ ID NO: 44
RRKPGKLQCLASYCWLFWTGIA	(+)	(+++)	SEQ ID NO: 45
RKAPGKLQCLASYCWLFWTGIA	(+)	(+++)	SEQ ID NO: 46
KRAPGKLQCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 47
MeArg-RAPGKLTCCLASYCWLFWTGIA	(+)	(+)	SEQ ID NO: 48
R-MeArg-APGKLTCCLASYCWLFWTGIA	(+++)	(++)	SEQ ID NO: 49
RR-MeAla-PGKLTCCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 50
RRAP-Sar-KLTCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 51
RRAPG-MeLys-LTCLASYCWLFWTGIA	(++)	(++)	SEQ ID NO: 52
RRAPGK-MeLeu-TCLASYCWLFWTGIA	(+)	(++)	SEQ ID NO: 53
RRAPGKL-MeThr-CLASYCWLFWTGIA	(+)	(+)	SEQ ID NO: 54
RRAPGKLTC-MeLeu-ASYCWLFWTGIA	(+)	(++)	SEQ ID NO: 55
RRAPGKLTC-MeAla-SYCWLFWWTGIA	(++)	(++)	SEQ ID NO: 56
RRAPGKLTC-MeSer-YCWLFWTGIA	(+++)	(++)	SEQ ID NO: 57
RRAPGKLTC-MeTyr-CWLFWWTGIA	(+)	(++)	SEQ ID NO: 58
RRAPGKLTCCLASYC-MeTrp-LFWWTGIA	>10	>1.1	SEQ ID NO: 59
RRAPGKLTCCLASYCW-MeLeu-FWTGIA	>10		SEQ ID NO: 60
RRAPGKLTCCLASYCWL-MePhe-WTGTGIA	(+)	(+++)	SEQ ID NO: 61
RRAPGKLTCCLASYCWLF-MeTrp-TGTGIA	(++)	(+)	SEQ ID NO: 62
RRAPGKLTCCLASYCWLF-MeThr-GTGTGIA	(++)	(+)	SEQ ID NO: 63
RRAPGKLTCCLASYCWLFWT-Sar-IA	(+++)	(++++)	SEQ ID NO: 64
RRAPGKLTCCLASYCWLFWTG-Melle-A	>4	>2.3	SEQ ID NO: 65
RRAPGKLTCCLASYCWLFWTGI-MeAla	(+)	(++)	SEQ ID NO: 66
rRAPGKLTCCLASYCWLFWTGIA (compound 5)	(+++)	(+++)	SEQ ID NO: 67
RrAPGKLTCCLASYCWLFWTGIA (compound 6)	(+++)	(+++)	SEQ ID NO: 68
RRaPGKLTCCLASYCWLFWTGIA	(+)	(++)	SEQ ID NO: 69
RRaPGKLTCCLASYCWLFWTGIA	(+++)	(++)	SEQ ID NO: 70
RRAPGkLTCCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 71
RRAPGKITCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 72
RRAPGKLtCLASYCWLFWTGIA	(+)	(++)	SEQ ID NO: 73
RRAPGKLtCLASYCWLFWTGIA	(++)	(++)	SEQ ID NO: 74
RRAPGKLTCIASYCWLFWWTGIA	(+)	(++)	SEQ ID NO: 75
RRAPGKLTClaSYCWLFWTGIA	(+)	(+)	SEQ ID NO: 76
RRAPGKLTClaSYCWLFWTGIA	(+++)	(++)	SEQ ID NO: 77

Sequence	EC ₅₀ [μM]	Vmax	
RRAPGKLTCLASyCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 78
RRAPGKLTCLASyCwLFWTGIA	(+)	(++)	SEQ ID NO: 79
RRAPGKLTCLASyCwLFWTGIA	(++)	(+++)	SEQ ID NO: 80
RRAPGKLTCLASyCWlFWWTGIA	(+)	(++)	SEQ ID NO: 81
RRAPGKLTCLASyCWLfWTGIA	(+++)	(++)	SEQ ID NO: 82
RRAPGKLTCLASyCWLFWtGIA	(++)	(++)	SEQ ID NO: 83
RRAPGKLTCLASyCWLFWtGIA	(+++)	(++)	SEQ ID NO: 84
RRAPGKLTCLASyCWLFWTGiA	(++)	(++)	SEQ ID NO: 85
RRAPGKLTCLASyCWLFWTGIa	(+)	(++)	SEQ ID NO: 86
CRRAPGKLQCLASyCWLFWTGIAC	(+)	(+++)	SEQ ID NO: 87
CGGSGGRRAPGKLQCLASyCWLFWTGIAC	(+)	(++)	SEQ ID NO: 88
CRRAPGKLQCLASyCWLFWTGIAGGSGGC	(+)	(+++)	SEQ ID NO: 89
CGGSGGRRAPGKLQCLASyCWLFWTGIAGGSGGC	(+)	(++)	SEQ ID NO: 90
PEG4-RRAPGKLTCLASyCWLFWWTGIA	(+++)	(++++)	SEQ ID NO: 91
Glu(Biotinyl-PEG)-RAPGKLTCLASyCWLFWWTGIA	>4	>0.8	SEQ ID NO: 92
RK(PEG2-Biotin)APGKLTCLASyCWLFWWTGIA	(+++)	(++)	SEQ ID NO: 93
RRK(PEG2-Biotin)PGKLTCLASyCWLFWWTGIA	(+++)	(++)	SEQ ID NO: 94
RRAK(PEG2-Biotin)GKLTCLASyCWLFWWTGIA	(+)	(++)	SEQ ID NO: 95
RRAPK(PEG2-Biotin)KLTCLASyCWLFWWTGIA (compound 20)	(+++)	(+++)	SEQ ID NO: 96
RRAPGK(PEG2-Biotin)LTCLASyCWLFWWTGIA	(+++)	(++)	SEQ ID NO: 97
RRAPGKK(PEG2-Biotin)TCLASyCWLFWWTGIA	>5	>2	SEQ ID NO: 98
RRAPGKLK(PEG2-Biotin)CLASyCWLFWWTGIA	(++)	(++)	SEQ ID NO: 99
RRAPGKLTCK(PEG2-Biotin)ASyCWLFWWTGIA	(+++)	(++)	SEQ ID NO: 100
RRAPGKLTCLK(PEG2-Biotin)SYCWLFWWTGIA	(+++)	(++)	SEQ ID NO: 101
RRAPGKLTCLAK(PEG2-Biotin)YCWLFWWTGIA (compound 18)	(+++)	(+++)	SEQ ID NO: 102
RRAPGKLTCLASK(PEG2-Biotin)CWLFWTGIA	(+++)	(++)	SEQ ID NO: 103
RRAPGKLTCLASYCK(PEG2-Biotin)LFWWTGIA	(+++)	(++)	SEQ ID NO: 104
RRAPGKLTCLASYCWK(PEG2-Biotin)FWWTGIA	(+)	(++++)	SEQ ID NO: 105
RRAPGKLTCLASYCWLK(PEG2-Biotin)WTGIA	(+)	(+++)	SEQ ID NO: 106
RRAPGKLTCLASYCWLFK(PEG2-Biotin)TGIA (compound 19)	(+++)	(+++)	SEQ ID NO: 107
RRAPGKLTCLASYCWLFWK(PEG2-Biotin)GIA	(+++)	(++)	SEQ ID NO: 108
RRAPGKLTCLASYCWLFWTK(PEG2-Biotin)IA	(++)	(+++)	SEQ ID NO: 109
RRAPGKLTCLASYCWLFWTGK(PEG2-Biotin)A	(+++)	(++)	SEQ ID NO: 110
RRAPGKLTCLASYCWLFWTGIK(PEG2-Biotin)	(+++)	(++)	SEQ ID NO: 111
RRAPGKLTCLASYCWLFWTGIA-PEG4	(+++)	(+++)	SEQ ID NO: 112

Sequence	EC ₅₀ [μM]	Vmax	
QWQIAGQVLK RRAPA KLTCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 113
LQLSYGEQRQ SRAPG KLTCLASYCWLFWTGIA	(+++)	(++)	SEQ ID NO: 114
WMSAEGIVGV RRATG KLTCLASYCWLFWTGIA			SEQ ID NO: 115
TSGPFGFGGS SRAQG KLTCLASYCWLFWTGIA	(+)	(++)	SEQ ID NO: 116
HLFGADWLGA RTAPG KLTCLASYCWLFWTGIA	(+)	(++)	SEQ ID NO: 117
QRAGRVARLH RRAPN KLTCLASYCWLFWTGIA	(+)	(+++)	SEQ ID NO: 118
WRAGLDESQR DRAPG KLTCLASYCWLFWTGIA	(+++)	(++)	SEQ ID NO: 119
PSGWAGWAPG RREPG KLTCLASYCWLFWTGIA	(+)	(+++)	SEQ ID NO: 120
AVDSLPLYGA RSAPS KLTCLASYCWLFWTGIA	(+++)	(++)	SEQ ID NO: 121
ASVWGALALV RRASG KLTCLASYCWLFWTGIA			SEQ ID NO: 122
GYRVPVGGLV RRAHG KLTCLASYCWLFWTGIA	(+)	(+++)	SEQ ID NO: 123
TQWAQVGPRG RRAQG KLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 124
VGSGDERALP SRASG KLTCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 125
TLWPWGGQGG RNAPG KLTCLASYCWLFWTGIA	(+)	(+++)	SEQ ID NO: 126
TGLLQGRRDE RARPP KLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 127
RGGFFVWFLS RIAPG KLTCLASYCWLFWTGIA			SEQ ID NO: 128
RRAPGKLTCLASYCWLF STGVA THANTTATAQ	(+)	(++++)	SEQ ID NO: 129
RRAPGKLTCLASYCWLF WAGFA ASTLAPAHHQ	(++)	(+++)	SEQ ID NO: 130
RRAPGKLTCLASYCWLF WSGFA SLGGLLWPVA			SEQ ID NO: 131
RRAPGKLTCLASYCWLF WTGYA SGKPSRVYVI	(+)	(++++)	SEQ ID NO: 132
RRAPGKLTCLASYCWLF WTGLS RYQWQAQEDV	(+)	(+++)	SEQ ID NO: 133
RRAPGKLTCLASYCWLF GSGIS LSRAPESAAP	(++)	(++++)	SEQ ID NO: 134
RRAPGKLTCLASYCWLF WTGWA VLARVPVGGT	(+++)	(+++)	SEQ ID NO: 135
RRAPGKLTCLASYCWLF WTGLA PGRGQGGVAG	(++)	(++++)	SEQ ID NO: 136
RRAPGKLTCLASYCWLF WTGIA DRLVWGVIST			SEQ ID NO: 137
RRAPGKLTCLASYCWLF WTGFA FRVGLASSLY	(+++)	(++++)	SEQ ID NO: 138
RRAPGKLTCLASYCWLF WTGLA STLYKTYTRE	(+)	(++)	SEQ ID NO: 139
RRAPGKLTCLASYCWLF RTQIA TPESEYRQQA	(++)	(++++)	SEQ ID NO: 140
RRAPGKLTCLASYCWLF WAGYP SLRGSLLVGV	(++)	(++++)	SEQ ID NO: 141
RRAPGKLTCLASYCWLF QTGWA YWGYRQHPGS	(+++)	(+++)	SEQ ID NO: 142
RRAPGKLTCLASYCWLF WTGWC RDTASHACDS	(+)	(++)	SEQ ID NO: 143
RRAPGKLTCLASYCWLF WTGWS RDTASHASDS	(++)	(+++)	SEQ ID NO: 144
RRAPGKLTCLASYCWLF WRGFA ERASEDTNQG	(+)	(+++)	SEQ ID NO: 145
RRAPGKLTCLASYCWLF EPGIA QPYAKSPTRN	(+)	(+++)	SEQ ID NO: 146
RRAPGKLTCLASYCWLF STPVA RKSLRRHQPT	>10	(++++)	SEQ ID NO: 147
PRIRTVGPGS RSASG KLTCLASYCWLFWTGIA (compound 21)	(+++)	(++++)	SEQ ID NO: 148
TVGPGSRSASGKLTCLASYCWLFWTGIA			SEQ ID NO: 424

Sequence	EC ₅₀ [μM]	Vmax	
PRIrTVGPGSRsASGKLTCLASYCWLFWTGIA			SEQ ID NO: 425
PRIrTVGPGSrSASGKLTCLASYCWLFWTGIA (compound 23)	(+++)	(++++)	SEQ ID NO: 426
PRIRTVGPGSRsASGKLTCLASYCWLFWTGIA-PEG4			SEQ ID NO: 427
PEG4-PRIRTVGPGSRsASGKLTCLASYCWLFWTGIA			SEQ ID NO: 428
PRIRTVGPGSRsASGKLTCLASYCWLFWTGIA-PEG4- Pra			SEQ ID NO: 429
Pra-PEG4- PRIRTVGPGSRsASGKLTCLASYCWLFWTGIA			SEQ ID NO: 430
Ac-PRIRTVGPGSRsASGKLTCLASYCWLFWTGIA			SEQ ID NO: 431
SRIRTVGPGSRsASGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 432
PSIRTVGPGSRsASGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 433
PRSRTVGPGSRsASGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 434
PRISTVGPGSRsASGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 435
PRIRSVGPGSRsASGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 436
PRIRTSVGPGSRsASGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 437
PRIRTVSPGSRsASGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 438
PRIRTVSGSGSRsASGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 439
PRIRTVGPSSRSASGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 440
PRIRTVGPGSRsASGKSTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 441
PRIrTVGPGSrSASGKSTCLASYCWLFWTGIA			SEQ ID NO: 442
PRIrTVGPGSrSASGKSTCLASYCWLFWTGIA-PEG4-Pra	(+++)	(++++)	SEQ ID NO: 443
Pra-PEG4-PRIrTVGPGSrSASGKSTCLASYCWLFWTGIA	(+)	(++)	SEQ ID NO: 444
SRIRTVGPGSRsASGKSTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 445
PRIRTVSPGSRsASGKSTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 446
SRIRTVSPGSRsASGKSTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 447
PRSRTVGPGSRsASGKSTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 448
SRSRTVSPGSRsASGKSTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 449
K(PEG2-biotin)-PEG4- PRIRTVGPGSRsASGKLTCLASYCWLFWTGIA			SEQ ID NO: 450
PRIRTVGPGSRsASGKLTCLASYCWLAWTGIA	(+)	(++++)	SEQ ID NO: 451
PRIRTVGPGSRsASGKLTCLASYCWLLWTGIA	(+++)	(++++)	SEQ ID NO: 452
PRIRTVGPGSRsASGKLTCLASYCWLFATGIA	(+++)	(++++)	SEQ ID NO: 453
PRIRTVGPGSRsASGKLTCLASYCWLFFTGIA	(+++)	(++++)	SEQ ID NO: 454
PRIRTVGPGSRsASGKLTCLASYCWLFITGIA	(+++)	(++++)	SEQ ID NO: 455
PRIRTVGPGSRsASGKLTCLASYCWLFWSGIA	(+++)	(++++)	SEQ ID NO: 456
PRIRTVGPGSRsASGKLTCLASYCWLFWTLIA	(+++)	(++++)	SEQ ID NO: 457
PRIRTVGPGSRsASGKLTCLASYCWLFWTFIA	(+++)	(+++)	SEQ ID NO: 458
PRIRTVGPGSRsASGKLTCLASYCWLFWTSIA	(+++)	(++++)	SEQ ID NO: 459

Sequence	EC ₅₀ [μM]	Vmax	
PRIRTVGPGRSASGKLTCLASYCWLFWTGI	(+++)	(++++)	SEQ ID NO: 460
SRIRTVSPGSRsSASGKSTCLASYCWLFWTGIA	(+)	(++++)	SEQ ID NO: 461
PRIRTVGPGRSRRASGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 462
PRIRTVGPGRSASGKLSCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 463
PRIRTVGPGRSASGKLTCAASYCWLFWTGIA	(+)	(+++)	SEQ ID NO: 464
PRIRTVGPGRSASGKLTCSASYCWLFWTGIA	(+)	(++)	SEQ ID NO: 465
PRIRTVGPGRSASGKLTCIASYCWLFWTGIA	(++)	(+++)	SEQ ID NO: 466
PRIRTVGPGRSASGKLTCVASYCWLFWTGIA	(+)	(+++)	SEQ ID NO: 467
PRIRTVGPGRSASGKLTCFASYCWLFWTGIA	(+)	(+++)	SEQ ID NO: 468
PRIRTVGPGRSASGKLTCLASFCWLFWTGIA			SEQ ID NO: 469
PRIRTVGPGRSASGKLTCLASACWLFWTGIA	(+)	(++)	SEQ ID NO: 470
PTDTGPVISG LRAPG KLTCLASYCWLFWTGIA	(+++)	(++)	SEQ ID NO: 149
GSVRRALFVA ARAPA KLTCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 150
RRFVGGSLSQ RRAPG KLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 151
SKQGRPISPD RRAAG KLTCLASYCWLFWTGIA (compound 22)	(++)	(++++)	SEQ ID NO: 152
SKQGRPISPD _r RAAGKLTCLASYCWLFWTGIA (compound 24)	(+++)	(++++)	SEQ ID NO: 471
SKQGRPISPDRRAAGKLTCLASYCWLFWTGIA-PEG2- K(PEG2-Biotin)	(+)	(++)	SEQ ID NO: 472
SKQGRPISPD _r RAAGKLTCLASYCWLFWTGI	(+++)	(++++)	SEQ ID NO: 473
SKQGRPISPD _r RAAGKLTCLASYCWLFWTG	(+++)	(+++)	SEQ ID NO: 474
SKQGRPISPD _r RAAGKLTCLASYCWLFWT	(+++)	(+++)	SEQ ID NO: 475
SKQGRPISPD _r RAAGKLTCLASYCWLFW	(+++)	(++++)	SEQ ID NO: 476
SKQGRPISPD _r RAAGKLTCLASYCWLF	(++)	(++++)	SEQ ID NO: 477
AKQGRPISPD _r RAAGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 478
SAQGRPISPD _r RAAGKLTCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 479
SKAGRPISPD _r RAAGKLTCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 480
SKQARPISPD _r RAAGKLTCLASYCWLFWTGIA	(+)	(++++)	SEQ ID NO: 481
SKQGAPISPD _r RAAGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 482
SKQGRAISPD _r RAAGKLTCLASYCWLFWTGIA	(+)	(++++)	SEQ ID NO: 483
SKQGRPASPDrRAAGKLTCLASYCWLFWTGIA	(++)	(+++)	SEQ ID NO: 484
SKQGRPIAPDrRAAGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 485
SKQGRPISADrRAAGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 486
SKQGRPISPA _r RAAGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 487
SKQGRPISPD _r RAAGKLTCLaSYCWLFWTGIA	>10	>0.5	SEQ ID NO: 488
SKQGRPISPD _r RAAGKLTCLASYCWIFWTGIA	>10	>2.4	SEQ ID NO: 489
SKQGRPISPD _r RAAGKLTCLAS-NMeTyr-CWLFWTGIA	>10	>0.3	SEQ ID NO: 490

Sequence	EC ₅₀ [μM]	Vmax	
SKQGRPISPDrRAAGKLTCLASYCW-NMeLeu-FWTGIA	>10	>1.9	SEQ ID NO: 491
SKQGRPISSDrRAAGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 492
SKQGRPISSDrRAAGKSTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 493
SKQGRPISSDrRASGKLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 494
SKQGRPISSDrRASGKSTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 495
RPRSSAHRP RRAAG KLTCLASYCWLFWTGIA	(+)	(++++)	SEQ ID NO: 153
TALSRLVTM RTAPD KLTCLASYCWLFWTGIA	(+++)	(++)	SEQ ID NO: 154
PARGKERELM RRAPG KLTCLASYCWLFWTGIA	(+)	(++++)	SEQ ID NO: 155
GRAMAAEPWP RQAPG KLTCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 156
LYPRLYTPGS RRAYG KLTCLASYCWLFWTGIA	(++)	(+++)	SEQ ID NO: 157
AQWVGRGQWA IRAPG KLTCLASYCWLFWTGIA	(+)	(+++)	SEQ ID NO: 158
MQIRQAHQPR RSAPQ KLTCLASYCWLFWTGIA	(++)	(+++)	SEQ ID NO: 159
PRTTANRRSS RRAPA KLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 160
PNLLRVRTSE VRNPG KLTCLASYCWLFWTGIA	(+++)	(++)	SEQ ID NO: 161
SLISMTNPSG RRVPG KLTCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 162
NGALGFRSVV PRAAG KLTCLASYCWLFWTGIA	(+++)	(+++)	SEQ ID NO: 163
RSHSLDRMAG RRAPG KLTCLASYCWLFWTGIA	(++)	(++++)	SEQ ID NO: 164
AVVRGQEPHT RRTPG KLTCLASYCWLFWTGIA	(++)	(++++)	SEQ ID NO: 165
PQTRDPSSRD RRAPG KLTCLASYCWLFWTGIA	(+++)	(++++)	SEQ ID NO: 166
RRAPGKLTCLASYCWLF LGGIA PEGASTRTAN	(++)	(++++)	SEQ ID NO: 167
RRAPGKLTCLASYCWLF VTGTA HAPRVAPAPA	(+)	(++++)	SEQ ID NO: 168
RRAPGKLTCLASYCWLF WTAVG GQPYMLLAWR	(+++)	(++)	SEQ ID NO: 169
RRAPGKLTCLASYCWLF WAGIA PHRPLKERVER	(+)	(+++)	SEQ ID NO: 170
RRAPGKLTCLASYCWLF WAVIA PPKVKGTRQS	(+)	(+++)	SEQ ID NO: 171
RRAPGKLTCLASYCWLF ATGVT FPQIWAIPSP	(+)	(++++)	SEQ ID NO: 172
RRAPGKLTCLASYCWLF YTGIA HGHPMEHRKS	(+)	(++++)	SEQ ID NO: 173
RRAPGKLTCLASYCWLF LTGWA RVPLPPRPHP	(+)	(++)	SEQ ID NO: 174
RRAPGKLTCLASYCWLF WTGIA RWPSHRSGPS	(+)	(++)	SEQ ID NO: 175
RRAPGKLTCLASYCWLF WTTYA SYATKPADTT	(+++)	(+++)	SEQ ID NO: 176
RRAPGKLTCLASYCWLF FTGVA RSTTATTNTQ	(++)	(+++)	SEQ ID NO: 177
RRAPGKLTCLASYCWLF WSGIA PQPPNMRPSV	(+++)	(++++)	SEQ ID NO: 178
RRAPGKLTCLASYCWLF WTTFA WVFVVAVYGS			SEQ ID NO: 179
RRAPGKLTCLASYCWLF WTALA TLSVSMRSPF			SEQ ID NO: 180
RRAPGKLTCLASYCWLF WTTWT TAPTPPLTT	(+++)	(+++)	SEQ ID NO: 181
RRAPGKLTCLASYCWLF WTGLL DYPTPQSHEP	(+++)	(+++)	SEQ ID NO: 182
RRAPGKLTCLASYCWLF WTGIA LGHPPRPAPK			SEQ ID NO: 183
SKQGRPISPDRRAAGKLTCLASYCWLF GSGISLSRAPE-SAAP	>10	(+)	SEQ ID NO: 496

Sequence	EC ₅₀ [μM]	Vmax	
RRAPGKLCALASYCWLFWTGIA-COOH	(+)	(+++)	SEQ ID NO: 184
RRAPGKCTALASYCWLFWTGIA-COOH	(+)	(++)	SEQ ID NO: 185
RRAPGCLTALASYCWLFWTGIA-COOH	(+)	(+++)	SEQ ID NO: 186
RRAPGKLTCLASYACLFWTGIA-COOH	(++)	(+++)	SEQ ID NO: 187
RRAPGKLTCLASYAWCFWTGIA-COOH	(+)	(+++)	SEQ ID NO: 188
RRAPGKLTCLASYAWLCWTGIA-COOH	>10	>0.5	SEQ ID NO: 189
RRAPGKLCALASYACLFWTGIA-COOH	>10	>0.3	SEQ ID NO: 190
RRAPGKCTALASYAWCFWTGIA-COOH	(+)	(+++)	SEQ ID NO: 191
RRAPGCLTALASYAWLCWTGIA-COOH	>20	>0.1	SEQ ID NO: 192
RRAPGKLCALASYCWLFWTGIA-CONH ₂	(+)	(+++)	SEQ ID NO: 497
RRAPGKCTALASYCWLFWTGIA-CONH ₂	(+)	(+++)	SEQ ID NO: 498
RRAPGCLTALASYCWLFWTGIA-CONH ₂	(+)	(++)	SEQ ID NO: 499
RRAPGKLTCLASYACLFWTGIA-CONH ₂	(+)	(++++)	SEQ ID NO: 500
RRAPGKLTCLASYAWCFWTGIA-CONH ₂	>20	(+)	SEQ ID NO: 501
RRAPGKLTCLASYAWLCWTGIA-CONH ₂	>10	(+)	SEQ ID NO: 502
RRAPGKLCALASYACLFWTGIA-CONH ₂	>10	(+)	SEQ ID NO: 503
RRAPGKCTALASYAWCFWTGIA-CONH ₂	(+)	(++)	SEQ ID NO: 504
RRAPGCLTALASYAWLCWTGIA-CONH ₂	(+)	(++)	SEQ ID NO: 505
RRAPGKLT-Lys-LASY-Asp-WLFWTGIA-COOH	>10	>0.5	SEQ ID NO: 193
RRAPGKLT-Asp-LASY-Lys-WLFWTGIA-COOH	(+++)	(++)	SEQ ID NO: 194
RRAPGKLT-Orn-LASY-Asp-WLFWTGIA-COOH	(+)	(+++)	SEQ ID NO: 195
RRAPGKLT-Asp-LASY-Orn-WLFWTGIA-COOH	(+++)	(++)	SEQ ID NO: 196
RRAPGKLT-Dab-LASY-Asp-WLFWTGIA-COOH	(++)	(++)	SEQ ID NO: 506
RRAPGKLT-Asp-LASY-Dab-WLFWTGIA-COOH	>10	>1.0	SEQ ID NO: 197
RRAPGKLT-Dap-LASY-Asp-WLFWTGIA-COOH	>10	>0.8	SEQ ID NO: 198
RRAPGKLT-Asp-LASY-Dap-WLFWTGIA-COOH	(+)	(++)	SEQ ID NO: 199
RRAPGKLT-Lys-LASY-Glu-WLFWTGIA-COOH	(+++)	(++)	SEQ ID NO: 507
RRAPGKLT-Lys-LASY-Asp-WLFWTGIA-CONH ₂	(+)	(++)	SEQ ID NO: 508
RRAPGKLT-Asp-LASY-Lys-WLFWTGIA-CONH ₂	(+++)	(+++)	SEQ ID NO: 509
RRAPGKLT-Orn-LASY-Asp-WLFWTGIA-CONH ₂	>10	(+)	SEQ ID NO: 510
RRAPGKLT-Asp-LASY-Orn-WLFWTGIA-CONH ₂	(+++)	(+++)	SEQ ID NO: 511
RRAPGKLT-Dab-LASY-Asp-WLFWTGIA-CONH ₂	(+)	(+++)	SEQ ID NO: 512
RRAPGKLT-Dap-LASY-Asp-WLFWTGIA-CONH ₂	>10	(+)	SEQ ID NO: 513
RRAPGKLT-Asp-LASY-Dap-WLFWTGIA-CONH ₂	(+)	(+++)	SEQ ID NO: 514
RRAPGKLT-Lys-LASY-Glu-WLFWTGIA-CONH ₂	(+)	(++)	SEQ ID NO: 515
RRAPGKLT-Asp-LASY-Dab-WLFWTGIA-CONH ₂	>10	(++)	SEQ ID NO: 516
SKQGRPISPDRRAAGKLT-Asp-LASY-Orn-WLFWTGIA	(+++)	(++++)	SEQ ID NO: 517
RRAPGKLT-Asp-LASY-Orn-WLFGSGISLSRAPESAAP			SEQ ID NO: 518

Sequence	EC ₅₀ [μM]	Vmax	
PRIRTVGPGSRSASGKLT-Asp-LASY-Orn-WLFWTGIA			SEQ ID NO: 519
RRFVGGSLSQRRAPGKLT-Asp-LASY-Orn-WLFWTGIA			SEQ ID NO: 520
PQTRDPSSRRRAPGKLT-Asp-LASY-Orn-WLFWTGIA			SEQ ID NO: 521
C(PEG5k)GGG-RRAPGKLT-Asp-LASY-Lys-WLFWTGIA	(+)	(++)	SEQ ID NO: 522
CGGG-RRAPGKLT-Asp-LASY-Lys-WLFWTGIA	(+++)	(+++)	SEQ ID NO: 523
CGGGLVPRGGG-RRAPGKLT-Asp-LASY-Lys-WLFWTGIA	(+++)	(+++)	SEQ ID NO: 524
C(PEG5K)GGGLVPRGGG-RRAPGKLT-Asp-LASY-Lys-WLFWTGIA	(++)	(++)	SEQ ID NO: 525
C(N-ethylmaleimide)-PEG4-LVPR-PEG4-rRAPGKLTCLASYCWLFWTGIA	(+)	(++)	SEQ ID NO: 526
rRAPGKLTCLASYCWLFWTGIA-PEG4-C(N-PEG5k maleimide)	(+)	(+++)	SEQ ID NO: 527
C(N-PEG5k maleimide)-PEG4-LVPR-PEG4-rRAPGKLTCLASYCWLFWTGIA	(+)	(++)	SEQ ID NO: 528
C(N-PEG5k maleimide)-PEG4-SKQGRPISPDrRAAGKLTCLASYCWLFWTGIA	(+)	(+)	SEQ ID NO: 529
SKQGRPISPDrRAAGKLTCLASYCWLFWTGIA-PEG4-C(N-PEG5k maleimide)	>5	>0.5	SEQ ID NO: 530
C(Acm)GGGGfpipR-PEG5-rRAPGKLTCLASYCWLFWTGIA	(+)	(++++)	SEQ ID NO: 531
CKTYFWKpGNIMVTFC-PEG12-Lys(PEG2-Biotin)-PEG12-rRAPGKLTCLASYCWLFWTGIA	>20		SEQ ID NO: 532
rrapgkltclasycwlfwtgia (inverso)	(+++)	(+++)	SEQ ID NO: 533
AIGTWFLWCYSALCTLKGPARR (retro)	(+)	(++)	SEQ ID NO: 534
aigtwflwcysalctlkgparr (retro-inverso)	(+)	(+++)	SEQ ID NO: 535
RRAPGKLTCLASYCWLFWTGIA-COOH	(++)	(+++)	SEQ ID NO: 536
rRAPGKLTCLASYCWLFWTGIA-COOH	(++)	(+++)	SEQ ID NO: 537
KLTCLASYCWLF (compound 1)	(+++)	(+++)	SEQ ID NO: 200
RRRKLTCLASYCWLFRRR	(+)	(++)	SEQ ID NO: 201
RRRRRKLTCLASYCWLFRRRRR	(++)	(+++)	SEQ ID NO: 202
KKKKLTCLASYCWLFKKK	>5		SEQ ID NO: 203
KLTCLASYCWLFK	(++)	(++++)	SEQ ID NO: 204
KKLTCLASYCWLF	(+++)	(++++)	SEQ ID NO: 205
ALTCLASYCWLF	(+)	(++)	SEQ ID NO: 206
KATCLASYCWLF	(+++)	(+++)	SEQ ID NO: 207
KLACLASYCWLF	(+++)	(+++)	SEQ ID NO: 208
KLTALASYCWLF	(+)	(++)	SEQ ID NO: 209
KLTCAASYCWLF	(+++)	(++)	SEQ ID NO: 210
KLTCLAAYCWLF	(++)	(++)	SEQ ID NO: 211

Sequence	EC ₅₀ [μM]	Vmax	
KLTCLASACWLF	(+++)	(+++)	SEQ ID NO: 212
KLTCLASYAWLF	(+)	(++)	SEQ ID NO: 213
KLTCLASYCalf	(+)	(++++)	SEQ ID NO: 214
KLTCLASYCWAF	(++)	(+++)	SEQ ID NO: 215
KLTCLASYCWLA	(+++)	(++++)	SEQ ID NO: 216
ALTCLAAYCalf	(+)	(++)	SEQ ID NO: 217
KAACLASACWLA	>20		SEQ ID NO: 218
KLTCLASYCWLF	(+++)	(+++)	SEQ ID NO: 219
KITCLASYCWLF	(+++)	(+++)	SEQ ID NO: 220
KLtCLASYCWLF	(++)	(+++)	SEQ ID NO: 221
KLTcLASYCWLF	(++)	(+++)	SEQ ID NO: 222
KLTClASYCWLF	>5		SEQ ID NO: 223
KLTCLaSYCWLF	>5		SEQ ID NO: 224
KLTCLAsYCWLF	(+)	(+++)	SEQ ID NO: 225
KLTCLASyCWLF	>5		SEQ ID NO: 226
KLTCLASYcWLF	(+)	(++++)	SEQ ID NO: 227
KLTCLASYCwLF	(++)	(++++)	SEQ ID NO: 228
KLTCLASYCWIF	(+)	(++++)	SEQ ID NO: 229
KLTCLASYCWlf	(+)	(++++)	SEQ ID NO: 230
MeLys-LTCLASYCWLF	(+++)	(++)	SEQ ID NO: 231
K-MeLeu-TCLASYCWLF	(+++)	(+++)	SEQ ID NO: 232
KL-MeThr-CLASYCWLF	(+++)	(++)	SEQ ID NO: 233
KLTC-MeLeu-ASYCWLF	>20	(+)	SEQ ID NO: 234
KLTCCL-MeAla-SYCWLF	>20	(+)	SEQ ID NO: 235
KLTCCL-MeSer-YCWLF	>20	(+)	SEQ ID NO: 236
KLTCCL-MeTyr-CWLF	>20	(+)	SEQ ID NO: 237
KLTCCLASYC-MeTrp-LF	>20	(+)	SEQ ID NO: 238
KLTCCLASYCW-MeLeu-F	(+)	(+++)	SEQ ID NO: 239
KLTCCLASYCWL-MePhe	(+)	(++)	SEQ ID NO: 240
k-MeLeu-TCLASYCWLF (compound 2)	(+++)	(++++)	SEQ ID NO: 241
KLTCLASYCWL	(++)	(++++)	SEQ ID NO: 242
KLTCLASYCW	>10	(+)	SEQ ID NO: 243
KLTCLASYC	(+)	(++++)	SEQ ID NO: 244
LTCLASYCWLF	(++)	(++)	SEQ ID NO: 245
TCLASYCWLF	(+++)	(+)	SEQ ID NO: 246
CLASYCWLF	>20	(+)	SEQ ID NO: 247
WSLCFKLTCAYL	>100	(+)	SEQ ID NO: 248

Sequence	EC ₅₀ [μM]	Vmax	
(compound A)			
KLTALASYAWLF	>5.8	(+++)	SEQ ID NO: 249
KLTSLASYSWLF	>15	(+)	SEQ ID NO: 250
KLT-Pen-LASYCWLF	>20	(+)	SEQ ID NO: 251
KLTCLASY-Pen-WLF	>20	(+)	SEQ ID NO: 252
KLT-HCy-LASYCWLF	(++)	(+++)	SEQ ID NO: 253
KLTCLASY-HCy-WLF	(++)	(++++)	SEQ ID NO: 254
KLT-Lys-LASY-Glu-WLF	(+++)	(++)	SEQ ID NO: 255
KLT-Glu-LASY-Lys-WLF	(++)	(++)	SEQ ID NO: 256
KLT-Glu-LASY-Orn-WLF	(+++)	(++)	SEQ ID NO: 257
KLT-Orn-LASY-Glu-WLF-COOH	(+++)	(+)	SEQ ID NO: 258
KLT-Orn-LASY-Glu-WLF-CONH ₂	(+++)	(++)	SEQ ID NO: 538
KLT-Glu-LASY-Dab-WLF	(+)	(++)	SEQ ID NO: 539
KLT-Dab-LASY-Glu-WLF	(++)	(++)	SEQ ID NO: 540
KLT-Glu-LASY-Dap-WLF	(+++)	(++)	SEQ ID NO: 541
KLT-Dap-LASY-Glu-WLF	(+++)	(++)	SEQ ID NO: 542
KKTCLASYCWLF	>50	(++)	SEQ ID NO: 259
KLKCLASYCWLF	(++)	(++++)	SEQ ID NO: 260
KLTCKASYCWLF	(+)	(++++)	SEQ ID NO: 261
KLTCLKSYCWLF	(+)	(++)	SEQ ID NO: 262
KLTCLASKCWLF	>100	(+)	SEQ ID NO: 263
KLTCLASYCWLK	>100	(+)	SEQ ID NO: 264
RLTCLASYCWLF	(++)	(+++)	SEQ ID NO: 265
Dpr-LTCLASYCWLF	(+)	(+++)	SEQ ID NO: 266
Dab-LTCLASYCWLF	(++)	(++)	SEQ ID NO: 267
QLTCLASYCWLF	>20	(+)	SEQ ID NO: 268
Orn-LTCLASYCWLF	(+++)	(++)	SEQ ID NO: 269
Lys(Me)-LTCLASYCWLF	(++)	(++)	SEQ ID NO: 270
Lys(Me) ₂ -LTCLASYCWLF	(+)	(+)	SEQ ID NO: 271
Lys(Me ₃ Cl)-LTCLASYCWLF	(+)	(+)	SEQ ID NO: 272
KLTCLSSYCWLF	(+)	(+++)	SEQ ID NO: 273
KLTCLVSYCWLF	(++)	(+++)	SEQ ID NO: 274
KLTCL-Dpr-SYCWLF	(+)	(++)	SEQ ID NO: 275
KLTCL-Abu-SYCWLF	(+)	(+++)	SEQ ID NO: 276
KLTCLGSYCWLF	(+)	(+++)	SEQ ID NO: 277
KLTCL-Aib-SYCWLF	>20	(+)	SEQ ID NO: 278
KLTCLA-HSe-YCWLF	(+)	(++)	SEQ ID NO: 279
KLTCLA-Dpr-YCWLF	>20	(+)	SEQ ID NO: 280

Sequence	EC ₅₀ [μM]	Vmax	
KLTCLATYCWLF	(+)	(++)	SEQ ID NO: 281
KLTCLASYC-Nal-LF	(+++)	(++++)	SEQ ID NO: 282
KLTCLASYCFLF	(++)	(++++)	SEQ ID NO: 283
KLTCLASYCCLF	(++)	(+++)	SEQ ID NO: 284
KLTCLASYCW-Nle-F	(+++)	(+++)	SEQ ID NO: 285
KLTCLASYCWYF	(+++)	(+++)	SEQ ID NO: 286
KLTCLASYCWIF	(++)	(+++)	SEQ ID NO: 287
TGS-KLTCLASYCWLF-APG	(++)	(+++)	SEQ ID NO: 288
KLTCLASYCWLF-COOH	(+)	(++)	SEQ ID NO: 289
Ac-KLTCLASYCWLF	(+)	(++)	SEQ ID NO: 290
KLTCLASYCWLF-(PEG)4-CONH ₂	(+)	(++)	SEQ ID NO: 291
(PEG) ₂₇ -KLTCLASYCWLF	(+)	(+++)	SEQ ID NO: 292
(PEG) ₂₇ (PEG) ₂₇ -KLTCLASYCWLF	(+)	(+)	SEQ ID NO: 293
KLTCQASYCWLF	(+)	(++)	SEQ ID NO: 294
KLTCLASQCWLF	(+++)	(+++)	SEQ ID NO: 295
KLTCLASYCQLF	(++)	(++++)	SEQ ID NO: 296
QQTCQASQCQLF	>20	(+)	SEQ ID NO: 297
NPTCQASYCQLF	>20	(+)	SEQ ID NO: 298
QLTCLASECGLS	>20	(+)	SEQ ID NO: 299
AQTRVARCCQLF	>20	(+)	SEQ ID NO: 300
KKTCVASFCQMI	>5	>4	SEQ ID NO: 301
NLTGRASYGWLP	>20	(+)	SEQ ID NO: 302
KGRCLTSHCWLF	>20	(+)	SEQ ID NO: 303
TLTCRASYCQLF	(+)	(+)	SEQ ID NO: 304
KLTCRASYCQLF	(+)	(+++)	SEQ ID NO: 305
KLSCQAGQCWVF	(+)	(+++)	SEQ ID NO: 306
KLTCLASYCQLV	(+++)	(+++)	SEQ ID NO: 307
KLKCLSSECQLL	>20	(+)	SEQ ID NO: 308
QLTCLASYCGGV	>20	(+)	SEQ ID NO: 309
EQTCLASYCWLF	>20	(+)	SEQ ID NO: 310
QLPCLASYCGLF	>20	(+)	SEQ ID NO: 311
MLTCIASYCQLG	>20	(+)	SEQ ID NO: 312
SLADTQLTWLARQYWLSVSEGS	>20	(+)	SEQ ID NO: 313
RRCPGKLQALASYCWLFWTGIA	(+)	(+++)	SEQ ID NO: 314
RRAPGKLQCLASYCWLFWTGIA (compound 3)	(+++)	(++++)	SEQ ID NO: 315
DSRSAKRKCLASYCWLFGIGQA	>20	(+)	SEQ ID NO: 316
GASSDKLTCRTRHCSMFQPLSV	>20	(+)	SEQ ID NO: 317

Sequence	EC ₅₀ [μM]	Vmax	
GCSSDKLTARTRHCSMFQPLSV	>20	(+)	SEQ ID NO: 318
GSCRDQLTCLSSDRWQFFRRVS	>20	(+)	SEQ ID NO: 319
KEGFAQLPCLVCQGGLFSPRAI	>20	(+)	SEQ ID NO: 320
LRTQPKVTGLASCSGLVNCSD	>20	(+)	SEQ ID NO: 321
RCAQSRLPWLVSYCWLFSPYGM	>5	>1.5	SEQ ID NO: 322
LQELTKLTCLARSGWLVCNPGY	>20	(+)	SEQ ID NO: 323
WVPQWKVTCLASCSRLFHGFDA	>5	>1.5	SEQ ID NO: 324
SCVKHELKCLSSDSRLFSAVQR	>20	(+)	SEQ ID NO: 325
VAHYGKVTCLASYCQPLPSVGA	>20	(+)	SEQ ID NO: 326
MEGTRQPTCLASYCSPFQYVAP	>20	(+)	SEQ ID NO: 327
RPGGGKVTQCASYCWPFARAG	>20	(+)	SEQ ID NO: 328
ERYRLDMTCMASQCWQFPPAAG	(+++)	(++)	SEQ ID NO: 329
VGEHRKISCVASNCQLLRSGLA	>20	(+)	SEQ ID NO: 330
SISGQQLTCRASHCWLNLPWHS	>20	(+)	SEQ ID NO: 331
TLDSKNLQCLGSSCWLFSSGLS	>20	(+)	SEQ ID NO: 332
VQRSTQLTCLYGGCRLFGWNYH	>20	(+)	SEQ ID NO: 333
VSGTGRLTCVASYCWMFQLGSF	(+++)	(++)	SEQ ID NO: 334
MAGMLKLTCFASYCGLFPLVSS	(+++)	(++)	SEQ ID NO: 335
GAQLDKETCLASYCQLFSTVRR	>20	(+)	SEQ ID NO: 336
HMQWGKLPCLASYCWLFWYGIG	(+)	(+++)	SEQ ID NO: 337
LRQRLAKTCVASYCWLFSLVAS	>10	>1.5	SEQ ID NO: 338
WHERQQLTCLASYCGLFVGQVA	(+++)	(+)	SEQ ID NO: 339
RYQRARLTCLASYCGLLFMSA	>15	>0.5	SEQ ID NO: 340
AVAINKVPVASYCQLFESKIH	>20	(+)	SEQ ID NO: 341
AWPYHKPTCLASQCWQFLAQGS	>20	(+)	SEQ ID NO: 342
SYGRTKLTCLASSCWLFQVHG	>10	(+)	SEQ ID NO: 343
GVEDRQLTCLASSCWVFSRHSV	(+)	(++)	SEQ ID NO: 344
RSFTSELTCLASSCRRFHHVPP	>20	(+)	SEQ ID NO: 345
AQLRRKLTCLASYCWLFQFFSP	(+)	(++)	SEQ ID NO: 346
HMQWGKLPCLASYCWLFWYGIG	(+)	(+++)	SEQ ID NO: 347
QQRQIKMSCLASWCWLFSGIPW	(++)	(++)	SEQ ID NO: 348
GGALQQLTCPASYCWLFPMES	>10	(+)	SEQ ID NO: 349
HYARVQLRCLDGYCWLLTKSRM	>10	>2.5	SEQ ID NO: 350
YARDSTLTCQARTCQLVDYLG	>50	(+)	SEQ ID NO: 351
QQQARKLACLASYCWLFPSAG	>50	(+)	SEQ ID NO: 352
APPGGKRMCLVSGCQLFPWSAS	>50	(+)	SEQ ID NO: 353
QDGDGKLTCRASYCRRFLVGVH	>50	(+)	SEQ ID NO: 354
GIQGSEVACRASFCRLFEGGHV	>50	(+)	SEQ ID NO: 355

Sequence	EC ₅₀ [μM]	Vmax	
RFQTTQLTCLGSASCLFNLSVR	(+)	(+)	SEQ ID NO: 356
AWPYHKPTCLASQCWQFLAQGS	>50	(+)	SEQ ID NO: 357
GFGSRKLTSLASYGWLIQDRLP	>50	(+)	SEQ ID NO: 358
SGRGGKLTQCASFQFLGNGLS	(+++)	(++)	SEQ ID NO: 359
RSQGRKLTCLASYCWFLVWHR	(++)	(++)	SEQ ID NO: 360
EGRRDKLTCLASYCWLVGHGQH	>25	>0.3	SEQ ID NO: 361
RGRSAKLRLCLASYCWLFFGVIL	(+++)	(+++)	SEQ ID NO: 362
LLQIPNLTCLGSYCWLDNGVYA	>50	(+)	SEQ ID NO: 363
FGQPSRLTCLASYCWFLGNLVT	(++)	(++)	SEQ ID NO: 364
GEGGGKLSCVAIQCGLFKGLGR	>50	(+)	SEQ ID NO: 365
VDKGHQLRCQAGYCWLLGYNRE	>50	(+)	SEQ ID NO: 366
SGFGMKLTCLASYCGLFQGEIG	(+++)	(+)	SEQ ID NO: 367
LLHAQKLSCLASYCWVFDAEWD	(+++)	(+)	SEQ ID NO: 368
SGGSGKLTCLASYCWPFQGSQVR	>50	(+)	SEQ ID NO: 369
QDGVEKLTCLASYCWRFGDHGA	>50	(+)	SEQ ID NO: 370
DAGPNKLRCLASYCQLFGGGHA	>50	(+)	SEQ ID NO: 371
TLLYQNRTCLASYCWLFDKRSV	>50	(+)	SEQ ID NO: 372
LTWREKLACLASYCWFLWGAP	(++)	(++)	SEQ ID NO: 373
RQLWNKLTCLASYCALIGLSGT	(+)	(++)	SEQ ID NO: 374
KGAYQKLTCLASYCLLFLTAQ	(+++)	(++)	SEQ ID NO: 375
QEQPAKLTCRGSYCWLFKRGDQ	>50	(+)	SEQ ID NO: 376
HDSL DQLTCLASVCQLASMGAR	(+)	(+)	SEQ ID NO: 377
SRQSDKPTCLAISCSLLTSNVR	>50	(+)	SEQ ID NO: 378
HGLADRLTCLSSDCWLQPFQTS	>50	(+)	SEQ ID NO: 379
VARASKVECLASYCQLFVGGEV	(+)	(+)	SEQ ID NO: 380
GASGRRRTCVAASYCLLFQSGLP	>25	>0.1	SEQ ID NO: 381
FPIQHKLTCLSSDCWLFP SHSY	>50	(+)	SEQ ID NO: 382
QAKMLKLTCLASYCWLFVWTRS	(+++)	(+++)	SEQ ID NO: 383
RGRSAKLRLCLASYCWLFFGVIL	(+++)	(+++)	SEQ ID NO: 384
RGRSAKLTCLASYCWLFFGVIL	(+++)	(+++)	SEQ ID NO: 385
RGRSAKLRLCLASYCWLFFTVIL	(+++)	(+++)	SEQ ID NO: 386
VSGTGRLTCVASYCWMFQLGSF	(+++)	(++)	SEQ ID NO: 387
VSGTGRLTCVASYCWMFQLGIF	(+++)	(++)	SEQ ID NO: 388
VSGTGKLTCLASYCWLFQLGSF	(+++)	(++)	SEQ ID NO: 389
HQQRRLTCLAGYCWFLVGLPS	(+)	(+++)	SEQ ID NO: 390
GDSGRKLSCLGSYCWLSVQFMA	(+++)	(++)	SEQ ID NO: 391
RSTVSQMRCLASYCWLF PALVS	(+++)	(++)	SEQ ID NO: 392
NGGMQKPACLASQCWLFANPLP	(+)	(++)	SEQ ID NO: 393

Sequence	EC ₅₀ [μM]	Vmax	
RHSNHNLTQCASYCWLLPAGLQ	(+)	(+)	SEQ ID NO: 394
HLGSPKLTGASQCWLLNHEVS	>50	(+)	SEQ ID NO: 395
DAKVAKLRLGSGCWLLQYAPG	(+++)	(++)	SEQ ID NO: 396
SKWEHQRGCLANNCWLFTLAPG	>50	(+)	SEQ ID NO: 397
RGSVHQPTCLGGYCGR LHSSWV	>50	(+)	SEQ ID NO: 398
KRYVYRQMCLVSACWLLQLGYA	(+++)	(++)	SEQ ID NO: 399
k-MeLeu-TCLASYCWLF	(+++)	(++++)	SEQ ID NO: 400
K-k-MeLeu-TCLASYCWLF	(+)	(++++)	SEQ ID NO: 401
Ac-K-k-MeLeu-TCLASYCWLF	(+++)	(+++)	SEQ ID NO: 402
KK-k-MeLeu-TCLASYCWLF	>10		SEQ ID NO: 403
KKK-k-MeLeu-TCLASYCWLF	>20		SEQ ID NO: 404
k-k-MeLeu-TCLASYCWLF	>2.5		SEQ ID NO: 405
R-k-MeLeu-TCLASYCWLF	(+)	(++++)	SEQ ID NO: 406
k-MeLeu-TCLASFCWLF	(++)	(+++)	SEQ ID NO: 407
k-MeLeu-TCLAS-(Y-OMe)-CWLF	(+++)	(++++)	SEQ ID NO: 408
k-MeLeu-TCLASYCQLF	(+)	(++++)	SEQ ID NO: 409
k-MeLeu-TCLASYCWLA	(+)	(+++)	SEQ ID NO: 410
(KLTCLASYCWLF) ₂ =N-O-(CH ₂) ₃ -O-N=	(++)	(++++)	SEQ ID NO: 411
(KLTCLASYCWLF) ₂ =N-O-(CH ₂) ₃ -O-N=	(+++)	(++++)	SEQ ID NO: 412
"=N-O-(CH ₂) ₃ -O-N=(COCH ₂ CH ₂ CO-KLTCLASYCWLF) ₂ "	(++)	(++++)	SEQ ID NO: 413
KLLKLLKLLKLLK-k-MeLeu-TCLASYCWLF	(+++)	(+++)	SEQ ID NO: 543
FAM-GGSGG-k-MeLeu-TCLASYCWLF			SEQ ID NO: 544
Palmitoyl-(PEG)27-KLTCLASYCWLF	(+)	(+)	SEQ ID NO: 545
Palmitoyl-(PEG)27(PEG)27-KLTCLASYCWLF	>20	>0.3	SEQ ID NO: 546
H3N-KLTCLASYCWLF=N-O-CH ₂ -CO-PEG27-CNPRGD-(Y-OEt)-RC	>50		SEQ ID NO: 547
(CNPRGD-(Y-OEt)-RC)-PEG27-CO-CH ₂ -O-N=CH ₂ NHCOCH ₂ CH ₂ CO-KLTCLASYCWLF			SEQ ID NO: 548
5-FAM-CNPRGD(Y-OEt)RC			SEQ ID NO: 549
rRAPGKLTCLASYCWLFWTGIA-PEG16-Lys(PEG2-Biotin)-PEG16-Lys(palmitoyl)	(+)	(++++)	SEQ ID NO: 550
Lys(palmitoyl)-PEG16-Lys(PEG2-Biotin)-PEG16-rRAPGKLTCLASYCWLFWTGIA	(++)	(++++)	SEQ ID NO: 551
rRAPGKLTCLASYCWLFWTGIA-PEG16-Lys(PEG2-Biotin)-PEG16	(+++)	(++++)	SEQ ID NO: 552
rRAPGKLTCLASYCWLFWTGIA-PEG16-Lys(PEG2-Biotin)-PEG16-CNPRGD-Tyr(OEt)-RC	(++)	(++++)	SEQ ID NO: 553
PEG16-Lys(PEG2-Biotin)-PEG16-rRAPGKLTCLASYCWLFWTGIA	(+)	(++++)	SEQ ID NO: 554
CNPRGD-Tyr(OEt)-RC-PEG16-Lys(PEG2-Biotin)-PEG16-	(++)	(++++)	SEQ ID NO: 555

Sequence	EC ₅₀ [μM]	Vmax	
rRAPGKLTCLASYCWLFWTGIA			

[0691] In Table 1, above, the following indicators were used:

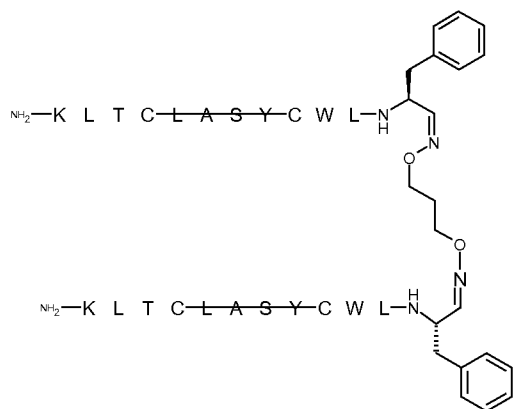
EC₅₀:

(+) > 1
 (++) 0.5 – 1.0
 (+++) < 0.5

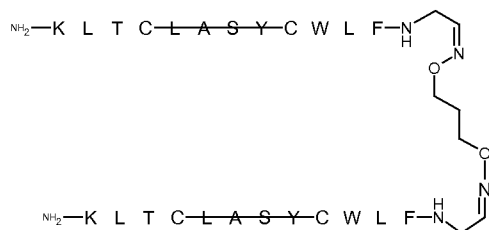
Vmax:

(+) < 1
 (++) 1 -2
 (+++) > 2
 (++++) >3

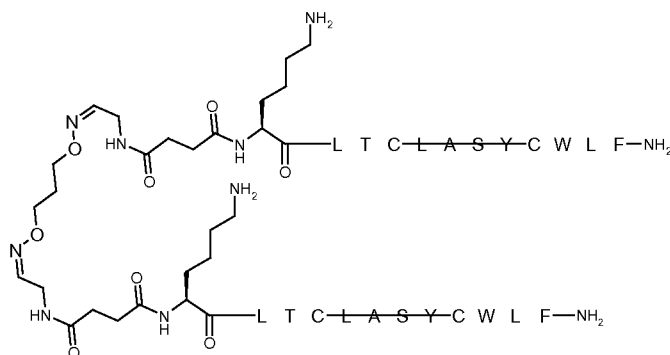
[0692] In Table 1, "(H₃N-KLTCLASYCWLF)₂=N-O-(CH₂)₃-O-N=" has the structure:



[0693] In Table 1, "(H₃N-KLTCLASYCWLF₂)₂=N-O-(CH₂)₃-O-N=" has the structure:



[0694] In Table 1, "=N-O-(CH₂)₃-O-N=(COCH₂CH₂CO-KLTCLASYCWLF-CONH₂)₂" has the structure:



Activity of compound 1 and its derivatives

[0695] Alanine mutants, D-amino acid mutants, and alpha-N-methyl derivatives of compound 1, and D-amino acid mutants, and alpha-N-methyl derivatives of compound 4 were prepared and tested according to the procedure of Example 2. Results indicate that the critical amino acid residues of the compounds of the present disclosure (e.g., compound 1 and compound 4) are located within the cysteine loop (between C¹ and C²) and towards the C-terminus of the amino acid sequence of compound 1.

Catalytic activities of FIXa in the absence and presence of various compounds

[0696] The catalytic constants k_{cat} and K_{M} for human FIXa (10 nM) measured in the absence and presence of various compounds of the present disclosure in the above described FXa generation assay are summarized below. Compound C (scrambled peptide; see Table 1) was used as a control at 10.0 μM . Compound 1 was used at 3 μM , compounds 3, 4, and 5 were used at 1 μM , and compound 10 was used at 0.3 μM . FIXa was used at 10 nM, and FX was used at 10-1000 nM. The assay was performed in the presence of phospholipids (PL) and calcium chloride. Certain compounds of the present disclosure lower the K_{M} and increase the k_{cat} of hFIXa. In this experiment, compounds of the present disclosure increased the catalytic activity of FIXa (*i.e.*, increased the k_{cat} of hFIXa) by at least 160-fold. As a comparison, known FIXa mutants increase k_{cat} up to 20-fold (see, e.g., *J Thromb Haemost*, 2009, 7, 1656-1662), and known FIXa enhancing antibodies increase the k_{cat} about 10-fold (see, e.g., *J Thromb Haemost*, 2008, 6, 315-322).

	k_{cat} (min^{-1})	Δk_{cat}	K_{M} (nM)	$\frac{\Delta K_{\text{M}}}{K_{\text{M,compound}}/K_{\text{M,buffer}}}$
Buffer	0.00009	1.0	78 ± 30	1.0
Compound 1	0.0144	160	24 ± 4.8	0.31
Compound 3	0.0177	197	16 ± 3.5	0.21
Compound 4	0.0171	190	11 ± 2.7	0.14
Compound 5	0.0179	199	11 ± 2.3	0.14
Compound C	0.00044	6	99 ± 76	~ 1

[0697] Comparison of the catalytic activities of FIXa and FVIIa in the absence and presence of compound 5

[0698] The the catalytic activities (k_{cat}) and K_{M} values for FIXa and FVIIIa were measured in the absence and presence of compound 5 using the above described modified FXa generation assay. The results are summarized below:

Δk_{cat}		Δk_{M}	
FVIIa	FIXa	FVIIa	FIXa
2419	199	0.3	0.14

$k_{\text{cat}}/K_{\text{M}}$

[0699] The $k_{\text{cat}}/K_{\text{M}}$ for FIXa was measured using various concentrations of FIXa (4, 2, 1, 0.5, and 0.25 μM) and a FIXa substrate in the absence and presence of compound 1 at 10 μM (~ 25 fold EC_{50}) and the positive control ethylene glycol (EG) 33%. The results for the chromogenic FIXa substrate, $\text{CH}_3\text{SO}_2\text{-(D)-CHG-Gly-Arg-pNA}$ (AcOH, BIOPHEN CS-51; $[\text{S}]_0 = 50$ nM constant; $[\text{S}]_0 \ll K_{\text{M}}$) are summarized below.

Sample	$(k_{\text{cat}}/K_{\text{M}})_{\text{app}} (\text{M}^{-1}\text{s}^{-1})$	Rel $k_{\text{cat}}/K_{\text{M}}$
FIXa only	1,591	-
FIXa + compound 1	1,358	0.85
FIXa + EG	29,730	18.7

[0700] Similar results were obtained for a fluorogenic FIXa substrate (Pefluor FIXa 3688). The above results indicate that the compounds of the present disclosure do not increase the amidolytic activity of FIXa directly (similarly to FVIIIa).

Mouse versus human FIXa

The EC₅₀ values for compound 1 in a FXa generation assay using either mouse FIXa (mFIXa) or human FIXa (hFIXa) were measured. The EC₅₀ values of compound 1 measured for mouse and human FIXa are similar (0.44 μ M and 0.30 μ M, respectively), however, the V_{max} is greatly reduced with mFIXa compared to hFIXa. The data confirms that the peptide was selected for binding to hFIXa.

Compounds of the present disclosure are not a substrate for FIXa

[0701] Compound 3 (100 μ M) was incubated in FXa generation assay buffer containing various concentrations of FIXa (0, 10 nM, 1 μ M) in the absence or presence of bovine serum albumin (BSA) for 3 hours. The mixtures were then analyzed by LC-MS. No significant differences were seen in the chromatograms or the mass spectra for compound 3 in the above samples. The results indicate that compound 3 is not a substrate for FIXa.

Example 3

(a) Thrombin Generation Assay (TGA)

[0702] A thrombogram was generated for selected compounds of the present disclosure in order to assess the thrombin potential in a physiological plasma environment. The TGA assay was carried out using a Calibrated Automated Thrombogram (CAT) test. Briefly, lyophilized and citrated FVIII deficient human plasma (Siemens) was reconstituted with 1 ml of distilled water, allowed to stand for 20 min at RT and mixed well before use. Alternatively, frozen FVIII-deficient plasma from Precision Biologics or HRF was used. FVIII deficient plasma was in some cases spiked with lyophilized platelets (Helena Laboratories, final concentration 0.66×10^8 cells/mL). Dilutions of peptide or controls such as FVIII were prepared in PBS (- Ca, -Mg) or in deficient plasma. In a 96-well round bottom plate, 20 μ l of activator solution (PRP reagent [final 0.1 pM rTF], Thrombinoscope bv) was added to each sample well and 20 μ l of Thrombin calibrator (Thrombinoscope bv) was added to calibrator well. Whenever plasma samples were used without platelets, PPP Low reagent [final 1 pM rTF, 4 μ M phospholipids], Thrombinoscope bv was used as activator instead of the PRP reagent. Subsequently, 80 μ l

of blank plasma was added to the calibrator well and 80 μ L of plasma containing compounds of the present disclosure, controls such as FVIII or blank was added to sample wells. All samples were prepared in duplicates and the plate was incubated at 37 °C for 10 minutes. After the incubation, 20 μ L of Fluca solution (Ca^{2+} + fluorogenic substrate, Diagnostica Stago) was added to each well and fluorescence was measured in a microtiter plate fluorometer (Fluoroskan Ascent, Thermo Scientific, Thrombinoscope software) for 1 hr. Similarly, TGA experiments were performed in FIX-deficient plasma.

[0703] Exemplary compounds of the present disclosure and their *in vitro* biological activities measured using the thrombin generation assay are summarized in Table 2, below. In Table 2, peptides are amidated ($-\text{CONH}_2$) at the C-terminus and have a free N-terminus, unless otherwise indicated. In Table 2, compound activities are based on thrombin peak height (nM thrombin). TGA experiments were performed in FVIII-deficient plasma from two different sources exhibiting different baseline values: HRF FVIII-deficient plasma (1) and Precision Biologics FVIII-deficient plasma (2).

Table 2

Exemplary Compounds of the Present Disclosure and their Activities in the TGA

Compound	Concentration (μ M)	Thrombin Above Baseline (nM) ^{a)}
Baseline (1)	0	-
FVIII	0.1 IU/mL	+
FVIII	0.25 IU/mL	++
FVIII	0.5 IU/mL	+++
FVIII	0.75 IU/mL	++++
FVIII	1 IU/mL	++++
KLTCCLASYCWLF	2.5 μ M	-
	5	-
	10	-
	20	-
k-MeLeu-TCLASYCWLF	2.5	+
	5	-
	10	-
	20	+
rRAPGKLTCCLASYCWLFWTGIA	2.5	+
	5	+
	10	++
	20	++
PRIRTVGPGSRASGKLTCCLASYCWLFWTGIA	2.5	+
	5	++
	10	+++
	20	+++

Compound	Concentration (μM)	Thrombin Above Baseline (nM) ^{a)}
PRIrTVGPGSrSASGKLTCLASYCWLFWTGIA	2.5	++
	5	+++
	10	+++
Baseline (2)		
SRIRTVGPGSRsSASGKLTCLASYCWLFWTGIA	2.5	++
	5	+++
	10	++++
PSIRTVGPGSRsSASGKLTCLASYCWLFWTGIA	2.5	+
	5	+
	10	+
PRSRTVGPGSRsSASGKLTCLASYCWLFWTGIA	2.5	++
	5	+++
	10	+++
PRISTVGPGRsSASGKLTCLASYCWLFWTGIA	2.5	+
	5	+
	10	+
PRIRSVGPGRsSASGKLTCLASYCWLFWTGIA	2.5	++
	5	+++
	10	+++
PRIRTSVGPGRsSASGKLTCLASYCWLFWTGIA	2.5	+
	5	++
	10	++
PRIRTVSPGRsSASGKLTCLASYCWLFWTGIA	2.5	++
	5	+++
	10	++++
PRIRTVGSGSRsSASGKLTCLASYCWLFWTGIA	2.5	+
	5	+
	10	+
PRIRTVGPSSRsSASGKLTCLASYCWLFWTGIA	2.5	+
	5	++
	10	++
PRIRTVGPGSRsSASGKSTCLASYCWLFWTGIA	2.5	+
	5	++
	10	+++
PRIRTVGPGSRsSASGKLTCLASYCWLFWTGIA	2.5	++
	5	+++
	10	+++
Baseline (2)		
PRIRTVGPGSRsSASGKSTCLASYCWLFWTGIA	2.5	+
	5	+++
	10	++++
	20	++++
SRIRTVGPGSRsSASGKSTCLASYCWLFWTGIA	2.5	+++
	5	++++
	10	++++
PRIRTVSPGRsSASGKSTCLASYCWLFWTGIA	2.5	++
	5	+++
	10	++++
	20	++++
SRIRTVSPGRsSASGKSTCLASYCWLFWTGIA	2.5	+++
	5	+++

Compound	Concentration (μM)	Thrombin Above Baseline (nM) ^{a)}
	10	++++
	20	++++
PRSRTVGPGSRSASGKSTCLASYCWLFWTGIA	2.5	++
	5	+++
	10	++++
	20	++++
SRSRTVSPGSRASGKSTCLASYCWLFWTGIA	2.5	+++
	5	++++
	10	++++

[0704] In Table 2, above, the following indicators were used:

Thrombin above baseline

1-10 nM (1) or 1-29 nM (2)	(+)
11-20 nM (1) or 30-59 nM (2)	(++)
21-30 nM (1) or 60-90 nM (2)	(+++)
>30 nM (1) or >90 nM (2)	(++++)
At or about baseline	(-)

[0705] The TGA activities of compound 7 (GWKPFLWDPRVLLSSGWYGRG), compound 8 (PWRRFWAWNPRSLALSTWFGRGCD), and compound 9 (GWKPFLWDPRVLLSSGWYGRGGGGGWKPFLWDPRVLLSSGWYGRG), which are less potent and structurally unrelated to the compounds of the present disclosure, were measured in the presence of FVIII. Compound 7 was used at at 100, 50, 25, 12.5 and 6.3 μM in the presence of 0.1 U/mL of FVIII; compound 8 was used at at 50, 25, 12.5, 6.3 and 3.1 μM in the presence of 0.1 U/mL of FVIII; and compound 9 was used at 50, 25, 12.5, 6.3 and 3.1 μM in the presence of 0.1 U/mL of FVIII. Under these conditions, each compound competes with FVIII in the TGA. The opposite phenomena was observed for the compounds of the present disclosure.

(b) Thrombin Generation Assay Using Purified Hemostatic Components

[0706] A purified thrombin generation assay was used to measure the selectivity of the compounds of the present disclosure. The assay was modified from the procedures described in Aljamali MN *et al.*, Epub 2009 Jul 29, Thrombin generation and platelet activation induced by rFVIIa (NovoSeven) and NN1731 in a reconstituted cell-based model mimicking haemophilia conditions, *Haemophilia* 2009, 15(6):1318-26; and

Christiansen ML *et al.*, Functional characteristics of N8, a new recombinant FVIII, *Haemophilia* 2010, 16(6):878-87.

[0707] The thrombin generation assay mixture was composed of FXI (3.1 ug/mL), FIX (3.1 ug/mL), FVII (6.3 nM), FVIIa (6.3 pM), tissue factor (10 fM), FV (4.4 ug/mL), FX (5 ug/mL), FII (54 ug/mL), antithrombin III (ATIII) (75 ug/mL), tissue factor pathway inhibitor (TFPI) (0.06 ug/mL) and lyophilized platelets (6E+07/mL). Purified recombinant human coagulation factors V, XI, II, X, IX, VII, VIIa, recombinant tissue factor and antithrombin III (ATIII) were obtained from Haematologic Technologies (Essex Junction, VT, USA). Lyophilized platelets were from Helena Laboratoires (Beaumont, TX); TFPI was from American Diagnostica. Compound 5 (1 uM), compound 4 (1 µM), compound 10 (1 µM), FVIII at 10% (0.1 IU/mL) or 100% (1 IU/mL) or buffer was added to the assay mixture together with the thrombin substrate (Fluca-kit, Thrombinoscope BV, The Netherlands) and the thrombin generation was measured with a Thrombinoscope Instrument (Thrombinoscope BV, The Netherlands). Both, FVIII and the pro-coagulant compounds of the present disclosure induce thrombin generation in this assay (i.e., in the presence of purified hemostatic components). The thrombin generation activities of compound 5, 4, and 10 are illustrated in Figure 5.

[0708] In a second experiment, compound 5 was tested at various concentrations (1 nM, 10 nM, 100 nM, 1 µM, and 2 µM). The results of this experiment are illustrated in Figure 6, which shows that compound 5 enhances thrombin generation in a dose-dependent manner.

[0709] In a third experiment it was investigated whether thrombin formation induced by FVIII or compound 5 is dependent on FIXa. The components of the thrombin generation assay were combined to generate the following final concentrations: FV (4.4 ug/mL), FX (5 ug/mL), FII (54 ug/mL), ATIII (75 ug/mL), and lyophilized platelets (6E+07/ml) in the presence of either 1%, 10% or 100% (3.1 µg/mL) of physiological FIXa. FVIII (0.01 U/mL) or compound 5 (1 µM) were added together with the thrombin substrate (Fluca-kit) and the plate was read using a thrombinoscope. Results show that the thrombin generation induced by compound 5 is dependent on the FIXa concentration present in the assay mixture. The results of this experiment are illustrated in Figure 7.

[0710] In a fourth experiment it was investigated whether thrombin formation induced by FVIII or compound 5 is dependent on FXIa. The components of the thrombin generation

assay were combined to generate the following final concentrations: FIX (3.1 ug/mL), FV (4.4 ug/mL), FX (5 ug/mL), FII (54 ug/mL), ATIII (75 ug/mL), and lyophilized platelets ($6E+07$ /ml) in the absence or presence of either 50%, or 100% FXIa. (100% FXIa corresponds to the physiological concentration of 3.1 μ g/mL). FVIII (0.01 U/mL) or compound 5 (1 μ M) were added together with the thrombin substrate (Fluca-kit) and the plate was read using a thrombinoscope. Results show that the thrombin generation induced by compound 5 is dependent on the FXIa concentration present in the assay mixture. The results of this experiment are illustrated in Figure 8.

[0711] In another experiment compound 5 in the above assay system enhanced thrombin generation in the absence of FIX/FIXa indicating that compounds of the present disclosure, in addition to increasing the intrinsic pathway, can enhance thrombin formation through the extrinsic pathway (*e.g.*, via increasing the catalytic activity of FVIIa). In this experiment, the thrombin generation assay mixture contained the following components: FVII (6.3 nM), FVIIa (6.3 pM), tissue factor (10 fM), FV (4.4 ug/mL), FX (5 ug/mL), FII (54 ug/mL), ATIII (75 ug/mL), TFPI (0.06 ug/mL) and lyophilized platelets ($6E+07$ /mL). Compound 5 (1 μ M) or buffer was added to the assay mixture together with the thrombin substrate (Fluca-kit) and the kinetics were read using a thrombinoscope.

Compounds of the present disclosure do not directly increase FXa activity

[0712] In another experiment it was determined whether or not the compounds of the present disclosure directly enhance the activity of FXa or the prothrombinase complex. In this experiment, compound 5 (1 μ M), compound 4 (1 μ M) or buffer was added to a thrombin generation assay mixture containing the following components at the indicated final concentrations: FII (54 ug/mL), FXa (0.05 ug/mL), ATIII (75 ug/mL), TFPI (0.06 ug/mL) and lyophilized platelets ($6E+07$ /mL) with or without FV (4.4 ug/mL).

[0713] A thrombin substrate (Fluca-kit) was added and the thrombin generation was measured using a thrombinoscope. Under these conditions, compound 5 and compound 4 did not substantially increase the catalytic activity of FXa or the prothrombinase complex.

Compounds of the present disclosure do not directly increase thrombin activity

[0714] In another experiment it was investigated whether or not the compounds of the present disclosure increase the thrombin activity directly. The thrombin activity was measured in the absence or presence of compound 5 using a fibrin generation assay with

the following assay components having the indicated final concentrations in the assay mixture: fibrinogen (0.45 g/L), calcium (16.5 mM), platelets (6×10^7 /mL), α -thrombin (0 or 0.1 IU/mL). The generation of fibrin was determined by measuring the absorbance at OD405 with the Biotek Synergy 2 multi-detection microplate reader.

[0715] In this experiment, compound 5 (1 μ M) did not substantially increase the catalytic activity of thrombin directly (*i.e.*, did not directly increase the amidolytic activity of α -thrombin).

Example 4

Rotational Thromboelastometry (ROTEM®) Assay

[0716] Compounds of the present disclosure were tested by rotational thromboelastometry to evaluate coagulation parameters such as clotting time (CT), α -angle, clot formation time (CFT), maximum clot firmness (MCF). Briefly, lyophilized and citrated FVIII deficient human plasma (Siemens) was reconstituted with 1 mL of distilled water, allowed to stand for 20 min at RT and mixed well before use. Alternatively, an aliquot of citrated (non-lyophilized) FVIII deficient human plasma (HRF, George King, Precision Biologics, or real-time donors) was thawed for 10 min in a 37 °C water bath and when needed centrifuged at 2800g for 5 min at 25 °C. FVIII deficient plasma was spiked with lyophilized platelets (Helena Laboratories, final concentration 0.5×10^8 cells/ml) and compounds, FVIII, or both compound and FVIII. Dilutions of compounds of the present disclosure and controls, such as FVIII, were prepared in PBS (-Ca, -Mg). Next, 300 μ l of the spiked plasma was transferred to a ROTEM cup already containing 20 μ l of StarTEM (concentrated calcium chloride solution) and 20 μ l of lipidated TF (American Diagnostica, final concentration 10 fM) or Kaolin (Sigma-Aldrich, final concentration 0.4 μ g/ml) and recording was initiated. The cups were maintained at 37 °C during the testing. All tests were run for 1.5 to 2 hrs.

[0717] Clotting times and α -angles for FVIII and compound 3 measured in a ROTEM assay using FVIII-deficient human plasma containing lyophilized platelets and 10 fM lipTF were compared. Clotting times and α -angles measured in the presence of FVIII and compound 3 at various concentrations are summarized below. In this experiment, compound 3 (at 5 μ M) has a faster clotting time than about 100% FVIII (100 % FVIII corresponding to 1 IU/mL). In this experiment, compound 3 (at 10 μ M) has an α -angle corresponding to about 20% FVIII. Results are summarized in Tables 3 and 4, below.

Table 3

	Compound 3 in human HemA plasma					
Conc., μ M	0	0.625	1.25	2.5	5	10
Clotting time, sec (avg.)	989	792	588	452	329	205
α -angle, degrees (avg.)	18	29	29	33	38	51

Table 4

	rFVIII in human HemA plasma					
Conc., IU/mL	0.01	0.05	0.10	0.25	0.50	1
Clotting time, sec (avg.)	852	761	610	521	458	363
α -angle, degrees (avg.)	26	35	48	60	63	71

[0718] The clotting times and α -angles for compound 3 in FVIII-deficient plasma in the absence and presence of neutralizing anti-FVIII antibodies (polyclonal sheep IgG against human FVIII, 18.4 mg/mL at 1:100 dilution) were also measured. In this experiment, the clotting time for compound 3 at 10 μ M is independent of residual FVIII, and the presence of a neutralizing anti-FVIII antibody. The α -angle measured for compound 3 at 10 μ M is independent of residual FVIII and the presence of a neutralizing anti-FVIII antibody. Results are summarized in Table 5, below.

Table 5

	Compound 5 in human HemA plasma					
Conc., μ M	0	0	5	5	10	10
Anti-FVIII pab	-	+	-	+	-	+
Clotting time, sec	777; 775	1,063; 1,202	154; 164	197; 194	101; 105	108; 110
α -angle, degrees	42; 41	10; 11	51; 58	48; 53	70; 68	67; 69

[0719] Similarly, clotting times and α -angles for FVIII and compound 23 at various concentrations were measured in a ROTEM assay using FVIII-deficient human plasma (HRF) were compared. Results are summarized in Tables 6 and 7, below.

Table 6

	Compound 23 in human HemA plasma (HRF)				
Conc., μM	0	2.5	5	10	20
Clotting time, sec (avg.)	2,208	983	673	486	472
α -angle, degrees (avg.)	9	13	22	32	39

Table 7

	rFVIII in human HemA plasma (HRF)					
Conc., IU/mL	0	0.10	0.25	0.50	0.75	1
Clotting time, sec (avg.)	1,823.5	1,062	821	783	654.5	608
α -angle, degrees (avg.)	18	21.5	25	32	38.5	35

[0720] Clotting times of compound 5 at 5 μM and 10 μM were measured using FIX-deficient human plasma containing lyophilized platelets and 10 fM TF. In this assay, compound 5 significantly reduces clotting time in FIX-deficient plasma. Results are summarized in Table 8, below.

Table 8

	Compound 5 in human HemB plasma			rFIX in HemB plasma
Conc. (μM)	0	5	10	1 IU/mL
Clotting time (sec)	1,188; 1,288	836; 810	804; 773	468; 468
α -angle (degrees)	19; 18	24; 20	22; 21	54; 51

[0721] The clotting time (CT) in a ROTEM assay using either FVIII-deficient human plasma (containing platelets) or FIX-deficient plasma (containing platelets) was measured in the absence and presence of compound 5. The results, summarized in Table 9, below, indicate that compound 5 significantly reduced clotting time in both FVIII- and FIX-deficient plasma. The effect is more pronounced in FVIII-deficient human plasma.

Table 9

	FVIII-deficient plasma		FIX-deficient plasma	
Compound 5, μM	0	10	0	10
Clotting time, sec	1,563; 1,666	277; 298	1,617; 1,628	1,311; 1,328
α -angle, degrees	14; 17	34; 27	20; 15	15; 19

[0722] The clotting time α -angle for compound 5 in FIX-deficient plasma in the absence and presence of neutralizing anti-FIX antibodies (polyclonal sheep IgG against human FIX, 3.96 mg/mL at 1:50 dilution) was measured. In these experiments, compound 5 reduces clotting time and increases α -angle in FIX-inhibited FIX-deficient plasma indicating that compound 5 is capable of enhancing the catalytic activity of other proteins, e.g., other blood clotting factors, in addition to FIXa. Results are summarized in Table 10, below.

Table 10

	Compound 5 in human HemB plasma					
Conc., μ M	0	0	5	5	10	10
Anti-FIX pab	-	+	-	+	-	+
Clotting time, sec	3,450; 3,267	5,138; 5,131	1,420; 1,556	1,660; 1,661	1,182; 1,194	1,125; 989
α -angle, degrees	8; 6	-	11; 12	9	12; 12	13; 10

[0723] The clotting time in the absence and presence of compound 5 at various concentrations in a ROTEM assay using canine FVIII-deficient plasma containing lyophilized human platelets and 10 fM lipTF was measured. Results are summarized in Table 11, below.

Table 11

	Compound 5 in canine HemA plasma						Compound C in canine HemA plasma	Normal canine plasma
Conc. (μ M)	0	0.625	1.25	2.5	5	10	10	0
Clotting time (sec)	1,062; 1,029	687; 539	404	346; 356	295; 369	286; 294	1,136; 1,491	364; 390
α -angle (degrees)	9; 14	12	14	15; 15	29; 28	58; 57	7	55; 44

[0724] The clotting time α -angle for compound 21 in whole blood was measured. The only difference from the above described plasma Rotem is that the experiment is done in human HemA (severe) whole blood. No platelets were added as they are already present in whole blood. Rotem activity was measured after 30 min pre-incubation of compound 21 in whole blood. Results are summarized in Table 12, below:

Table 12

	Compound 21 in human HemA (severe) whole blood						rFVIII in HemA WB
Conc. (μ M)	0	1.25	2.5	5	10	20	1 IU/mL
Clotting time (sec)	2,360; 2,910	1,140; 1,137	1,057; 1,090	930	748, 703	952; 625	702; 618
α -angle (degrees)	15	22; 24	32	32	44; 52	42; 43	42; 46

Example 5**Hydrogen Deuterium Exchange (HDX)**

[0725] Hydrogen/deuterium exchange mass spectrometry (H/DX-MS) was utilized to study the higher-order structural analysis of recombinant human factor IXa (hFIXa) in combination with and without compound 4. The general method can be found in the following references: Engen and Smith (2001) *Anal. Chem.* 73, 256A-265A.; Wales and Engen (2006) *Mass Spectrom. Rev.* 25, 158-170.

Experimental Conditions:***Samples and buffers:***

[0726] Samples contained human FIXa (1 μ M, Hematologic Technologies) in 50 mM Tris, pH 7.4; 100 mM NaCl; 2 mM CaCl₂ (98.8 % D₂O) with or without the presence of compound 4 (12 μ M).

H/DX-MS Analysis:

[0727] FIXa samples were equilibrated at ambient temperature (20 $\pm$ 1 °C) for 1 hour before labeling with deuterated buffer (reaction buffers described above). The samples were then diluted 1:15 with deuterated buffer and incubated, allowing hydrogen exchange to occur for various amounts of time (10 seconds, 1, 10, 60, and 240 minutes), before the reaction was quenched with a 1:1 dilution of 200 mM citrate, 8 M guanidinium HCl, and 0.5 M TCEP, pH 2.33. Quenched samples were then incubated for 20 seconds before being further diluted 1:1 with 0.1% formic acid and immediately injected into the LC-MS system for analysis.

[0728] Approximately 7 μ g of exchanged/quenched FIXa was injected onto an immobilized pepsin column where the digestion and peptide trapping were performed for 3 minutes with a flow rate of 0.1 mL/min in 0.1% formic acid at 10 °C (Houde D. et al., 2010 Post-Translational Modifications Differentially Affect IgG1 conformation and receptor binding. *Mol. Cell. Proteomics*, 9 (8): p1716). The peptic peptides were trapped on an ACQUITY BEH C18 1.7 μ m peptide pre-column trap (Waters Corp. Milford, MA) maintained at 0 °C. Flow was diverted by a switching valve and the trapped peptides flushed from the trap onto an ACQUITY BEH C18 1.7 μ m, 1mm x 100mm column (Waters Corp. Milford, MA) to separate the peptides at 0 °C using a 9 minute linear acetonitrile gradient (2-55%) with 0.1 % formic acid at a flow rate of 40 μ L/min (Wales TE, Fadgen KE, Gerhardt GC, Engen JR. 2008. High-speed and high-resolution UPLC separation at zero degrees Celsius. *Anal. Chem.* 80(17): p6815). Eluate from the C18 column was directed into a Waters Synapt HD mass spectrometer with electrospray ionization and lock-mass correction (using Glu-fibrinogen peptide). Mass spectra were acquired and peptic peptides were identified using a combination of exact mass and MS, aided by Waters Identity software (Silva JC et al., 2006. Absolute quantification of proteins by LCMSE: a virtue of parallel MS acquisition. *Mol Cell Proteomics*, 5(1): p144). All data was averaged from duplicate injections. The amount of deuterium in each peptide was determined by subtracting the mass of the undeuterated peptide from the mass of the deuterated peptide, incubated at various HDX time points (which were uncorrected for any back exchange). The mass data was then plotted as a function of deuterium exposure time.

Results:

[0729] 69 peptides were identified, representing approximately 75% of the hFIXa amino acid sequence, which corresponded to 37% coverage of the light chain and 99% coverage of the heavy chain. The resulting data produced 69 deuterium incorporation (HDX) graphs for both the FIXa control and FIXa with compound 4.

[0730] The difference in deuterium exchange between FIXa and FIXa with compound 4 was determined by subtracting the mass of each FIXa peptide from the mass of the corresponding FIXa with compound 4. This subtraction was done for each H/D exchange time point (0.17, 1, 10, 60, and 240 minutes). The sum of these differences, across all time points, was calculated. The values for the mass differences and the sum can be

either positive or negative. Positive values indicate that FIXa exchanges more rapidly than FIXa with compound 4, which may indicate that FIXa has a more open, flexible, or weakly H-bonded structure. Negative values mean the reverse. For a sample to be considered not comparable, the following criteria must be achieved: (1) At least one time point must fall outside of the ± 0.5 Da threshold; and (2) the corresponding difference sum value, for a peptide containing a difference value that exceeded the ± 0.5 Da threshold (criteria 1), must also exceed the ± 1.1 Da threshold. These criteria represent a rigorous measure for assessing comparability and are based on pure statistical experimental uncertainties associated with the H/DX-MS method. The rigor of how large a difference needs to be before establishing non-comparability between two nearly identical biopharmaceuticals is a variable that depends on the nature of the biopharmaceutical and where in the biopharmaceutical a difference is observed.

[0731] In order to further assess the presence of non-comparability in H/DX-MS experiments two quantitative difference indices, DI(1) and DI(2) were developed and reported as whole numbers. The DI(1) value is determined by summing up all the absolute values for the difference sum that exceed the threshold (1.1) and meet the criteria above. If a difference sum value is negative, the number is assigned a value of zero. In expressing DI values, a final value > 0 indicates that the samples are not comparable. The DI(2) value is determined similarly, but for each individual time point. The value for DI(1) and DI(2) for these experiments are 2 and 0, respectively. There were two peptides that showed statistically significant differences in H/D exchange when comparing FIXa to FIXa with compound 4: light chain peptide 85-97 (peptide number 4) and most importantly heavy chain peptide 177-185 (peptide number 57). The region of difference within the heavy chain peptide 177-185 could be further resolved from overlapping peptides. Heavy chain peptide 169-180 was found and the H/D exchange was shown to be similar between FIXa and FIXa with compound 4. Results are illustrated in Figure 4.

[0732] As a result, the difference observed in heavy chain peptide 177-185 could be localized to residues 180-185. The difference detected within the FIXa light chain is very small but appears significant at the later H/D exchange time points (60 and 240 minutes). The difference seen within the FIXa heavy chain is significant and is visible across nearly all time points. These residues are within close proximity to those regions on FIXa which are reported to interact with factor VIIIa (Bajaj SP et al. 2001. Factor

IXa:factorVIIIa interaction: helix 330-338 of factor IXa interacts with residues 558-565 and spacially adjacent regions of the A2 subunit of factor VIIIa. *J. Biol. Chem.* 276(19): p16302).

- [0733] Amino acid sequence alignments of human, canine and mouse FIXa suggest that the measured selectivity of the compounds of the present invention for human and canine FIXa as compared to mouse FIXa can be due to F184Y and H185R.

Example 6

In Vitro Stability of The Compounds

Plasma Stability

- [0734] Certain compounds of the present disclosure have limited chemical stability in plasma. For example, compound 1 shows significant degradation after 0.5 hours of incubation in mouse plasma at 37°C. However, compound 2 is stable in mouse plasma for at least 3 h.
- [0735] The stability, e.g., *in vitro* plasma stability of the compounds of the present disclosure can be increased by including D-amino acids or N-methylated amino acid residues into the peptide sequence. For example, the stability of a compound can be increased by replacing arginine (R) with D-arginine (r).
- [0736] The *in vitro* plasma stabilities for the compounds of the present disclosure were determined using a LC-MS method. Selected compounds were spiked into 120 µL of human FVIII deficient plasma (Siemens) to give a final concentration of 50 µg/mL. An aliquot of 20 µL was taken from the sample at time 0 as control. The remaining sample was incubated at 37°C for 2hrs. Additional aliquots of 20 µL were taken at time points 30 min, 1hr, 2hrs. All samples (20 µL aliquots) were treated immediately with 100 µL of cold 100% acetonitrile, vortexed for 10 min and centrifuged at 13000 rpm for 8 min. The 100 µL of supernant was transferred to a new vial and dried by speedvac and reconstituted with 100 µL of 20% acetonitrile and 0.1% formic acid. 90 µL of reconstituted sample was injected for LC-UV-MS/MS analysis. The MS was set at triple play with full scan, zoom scan and MS/MS scans with top five ions using dynamic exclusions. 5 µL of 10 µg/mL neat peptide in water was injected as a standard control. HPLC conditions: A. 0.1% FA in water; B. 0.1% formic acid in acetonitrile, column

temperature 50°C; flow rate 0.4 mL/min; 8 min run time with fast gradient. Samples were analyzed by LC-MS, and the data was reviewed to identify any breakdown products.

[0737] The chemical stability of various compounds of the present disclosure in human plasma was measured according to the above procedure. After two hours of incubation at 37°C, compounds 5 and 6 showed no detectable degradation, while compound 3 showed about 85% degradation after 2 hours. The results indicate that compounds of the present disclosure, which incorporate a D-amino acid at the N-terminus or close to the N-terminus (i.e., D-arginine), such as compounds 5 and 6, are more stable in plasma (e.g., human plasma) than a corresponding compound, which does not incorporate such D-amino acid (e.g., compound 3). The results are summarized in Table 13, below:

Table 13

Compound	Time (min)	Degradation (%)
3	0	0.00
3	30	64.72
3	60	75.71
3	120	85.26
5	0	0.00
5	30	0.00
5	60	0.00
5	120	0.00
6	0	0.00
6	30	0.00
6	60	0.00
6	120	0.00

[0738] In a similar fashion, the plasma stability of compound 23 was improved when compared to compound 21, and the chemical stability of compound 24 was improved compared to the stability of compound 22, while maintaining biological activity in each case.

Degradation products in mouse plasma

[0739] The stabilities of compound 1 and 2 were tested in mouse plasma. Compound 1 showed significant degradation after 0.5 h of incubation. Two degradation products were found for compound 1, which were identified using mass spectroscopy:

1. TCLASYCWLF (m/z theoretical: 1204.50; found 1204.44)
2. LTCLASYCWLF (m/z theoretical: 1317.50; found 1317.60).

[0740] Contrarily, no degradation products were detectable for compound 2, even after incubation at 37 °C for 3 hours in mouse plasma.

Whole Blood Stability

[0741] Compound 5 was also stable in whole blood for at least 120 min.

Example 7

Screening of Phage Libraries for FIXa binders

[0742] Selected Peptides capable of binding to FIXa were identified by screening filamentous phage display libraries licensed from Dyax Corp. (Cambridge, MA). More specifically, the following seven libraries were used in combination; TN6.VII, TN7.IV, TN8.IX, TN-9-IV, TN10-X, TN-11-I and TN-12-I were used in the screen. The total number of individual viable phage contained in each library was reflected by the number of transformants established for each library when the libraries were expressed in *E. coli* and plated at a clonal dilution as described by the Dyax protocol. The number of transformants for TN6.VII, TN7.IV, TN8.IX, TN-9-IV, TN10-X, TN-11-I and TN-12-I was 1.2×10^9 , 2.3×10^9 , 5.0×10^9 , 3.2×10^9 , 2×10^9 , 2.7×10^9 and 1.4×10^9 , respectively. Another way to refer to the absolute number of viable phage in a given volume is by stating the plaque forming units (pfu) per unit volume.

Reagents

[0743] The following reagents were used for the screening of FIXa-binding peptides:

1. Ampicillin: 100 g ampicillin in 1L MQ water; filter sterilized (0.22 μ m).
2. NZCYM medium: 10 g Casein Hydrolysate Enzymatic, 5 g NaCl (sodium chloride), 5 g Bacto Yeast Extract, 1 g Bacto Casamino Acids (Casein Digest), 1 g MgSO₄ anhydrous powder (magnesium sulfate). Ingredients were dissolved in 800 mL MQ water and pH adjusted to 7.5 with 1 N NaOH (sodium hydroxide), then brought up to a total volume of 1L with MQ water and autoclaved at 120 °C for 20 min.
3. NZCYM-A50 plates: NZCYM medium containing 15 g Bacto Agar / 1L and 100 μ g ampicillin/mL.

4. NZCYM-T12.5 medium: NZCYM medium containing tetracycline at 12.5 µg/mL
NZCYM-T12.5 plates: NZCYM medium containing 15 g Bacto Agar / L and tetracycline at 12.5 µg/mL.
5. PBS: 150 mM NaCl (sodium chloride), 8 mM Na₂HPO₄ anhydrous powder (sodium phosphate, dibasic), 1.5 mM KH₂PO₄ anhydrous powder (potassium phosphate, monobasic). Adjust pH to 7.4-7.6.
6. PEG/NaCl: 20% polyethylene glycol 6000 (PEG), 2.5 M NaCl (sodium chloride). The buffer was filter sterilized (0.22 µm). To make this solution, a 40% PEG solution stock was made in MQ water. An equal volume of 5 M NaCl was added while stirring to make the 20% PEG / 2.5 M NaCl stock solution.
7. TBS: 10 mM Tris-HCl (pH 7.5), 150 mM NaCl (sodium chloride)
8. TEA, 100 mM: 100 mM triethylamine (TEA). Buffer was freshly prepared (pH).
9. Tetracycline: 12.5 g tetracycline in 1L ethanol. Buffer was stored in dark at -20°C.
10. Tris-HCl, pH 7.4: 1 M Tris Base in MQ water. Adjust pH with HCl to pH = 7.4. The buffer was filter sterilized (0.22 µm).

Screening Protocol: Round 1

- [0744] Nunc plates were coated with 5, 50 and 500 µg/mL human Factor IXa (hFIXa, Hematologic Technologies) in TBS/5mM CaCl₂/pH 7.4 overnight at 4 °C. The solution was removed and the plate was blocked with 2% milk in TBS/5mM CaCl₂/pH 7.4 for 1-2 hours at room temperature.
- [0745] Aliquots (10 µL for each condition) of the 7 Tn libraries (Tn6~Tn12) were pooled and mixed with an equal volume of PBS/2%Milk for 1-2 hours at room temperature. From this solution, 100 µL was added to each target well. The phages were allowed to bind to hFIXa for 1 hour at room temperature. Subsequently, the solution was removed and the wells were washed 13 times with 2% milk in TBS/5mM CaCl₂/pH 7.4. Next, phage were eluted with 100 µL of the TEA solution per well, the solution was removed after 2-5 minutes and neutralized with 50 µL per well of 1 M Tris-HCl pH 7.4. The eluted phages were used to transfect competent *E. coli* XL1-Blue MRF' (Stratagene) and amplified overnight at 37 °C as described below.

Phage Infection

- [0746] A single colony of *E. Coli* XL1-Blue MRF' from a NZCYM-T12.5 plate was inoculated into 25 mL of NZCYM-T12.5 broth (tetracycline at 12.5 µg/mL). The culture was grown overnight at 37°C at 250 rpm. The following day, the XL1 blue MRF' *E. coli* cells were diluted 1:100 into 25 mL NZCYM-T12.5 and grown for about 2 hours until the culture reached an optical density of 0.5 at 600 nm. At this stage, 10 mL of the XL1 blue MRF' culture was infected with phage at 37°C for 15 mins.

Phage Titer

- [0747] A 20 µL aliquot of the above phage-infected XL1 blue MRF' culture was diluted with NZCYM broth in a serial manner (10^4 , 10^5 and 10^6). Each dilution was spread onto NZCYM-A50 plates which prior had been dried in a 37°C incubator for 1 hour. The plates were incubated in an inverted position overnight at 37°C. The titer was calculated the following day from a plate containing 30 to 300 plaques. The phage titer was derived from the equation: Phage titer = number of plaque x 1/dilution x 1/fraction plated.

Phage Amplification

- [0748] The infected cells were concentrated by centrifuging them at 3000 rpm, followed by resuspension in 10 mL of NZCYM overnight at 37°C without shaking. Next, the suspension was centrifuged at 10K at 4°C for 10 min and the supernatant was precipitated with 0.5 mL 30% PEG on ice for 1 hour. The precipitated phage was isolated by centrifugation and the supernatant was discarded. The phage pellet was resuspended in 100 µL per well of PBS/2%Milk.

Round 2 and 3

- [0749] The amplified phage library was used for a similar panning as described above. Screening Protocol: Round 1. At the completion of Round 3, the phages in the eluent were titered and assayed for FIXa binding using phage enzyme linked immunosorbent assay (ELISA).

Phage ELISA

- [0750] The following steps were carried out to identify phages encoding peptides that were able to bind hFIXa. Individual agar plugs containing phage plaques were picked

with autoclaved Pasteur pipets. Phage Titer. Plugs were deposited in 96-well sterile deep well plates (Nunc), to which 400 μ L per well of NZCYM media was added per well. Phage were grown overnight at 37°C. In addition, 100 μ L of 1 μ g/mL of hFIXa in TBS/Ca was coated on Nunc plates overnight at 4°C. The hFIXa solution was removed the following day and the plate was washed one time with TBS/Ca/tween. Subsequently, the wells were blocked with TBS/Ca/2% milk for 1-2 hours at room temperature. The wells were again washed with TBS/Ca/tween before 100 μ L of phage were added. Phage were allowed to bind to hFIXa for 1-2 hours at room temperature. Next, the wells were washed three times with TBS/Ca/tween before adding 100 μ L anti-M13 HRP antibody (diluted 1:5000 in TBS/Ca/2% milk) for 1 hour. Again, the wells were washed three times with TBS/Ca/tween, and the ELISA signal was detected by adding 100 μ L TMB, and scanning the plate at 650 nm. About 5% of the peptides identified as binding to FIXa from the phage ELISA possessed activity in the FXa generation assay.

Sequencing

- [0751] An aliquot (50 μ L) of the supernatant from positive phage clones (ELISA target signal/milk background signal >2.0) was collected and sequenced using Dyax's designed primer (3seq-80: 5'gataaacgatacaattaaaggctcc 3'). Among them the sequence of compound 1 was discovered.

Isolation of compound 3

- [0752] A secondary phage library was built based on the compound 1 primary sequence. The library contained the compound 1 sequence engineered with a 30% chance of an amino acid change within the compound 1 sequence, as well as an additional five randomized amino acid residues flanking both ends of the core sequence.
- [0753] DNA fragments coding for the peptides within the 22 amino acid secondary library were generated in the following manner: A 105-base oligonucleotide was synthesized to contain the sequence (NNB)₅ compound 1 sequence (NNB)₅, where N = A, C, T or G and B = C, G or T. This oligonucleotide was used as the template (0.5 nM, 1 μ L) in PCR amplification along with two shorter oligonucleotide primers (10 μ M, 1 μ L), both of which complement the 5' and the 3' end of the oligo (oligos A and B, respectively). The resulting PCR product was purified and concentrated with QIAquick spin columns (Qiagen), then digested with NcoI and NotI. The pSYNPHE phagemid

(Syntonix) was also digested with NcoI and NotI, followed by phosphatase treatment. The digested DNAs were resolved using a 1% agarose gel, excised and purified by QIAEX II treatment (Qiagen). The vector and insert were ligated overnight at 15°C. The ligation product was purified using QIAquick spin columns and electroporation was performed in an electroporation cuvette (0.1mm gap; 0.5ml volume) containing 12.5 µg of DNA and 500 µl of TG1 electrocompetent cells (Stratagene). Immediately after the pulse, 12.5 mL of pre-warmed (37°C) 2YT medium containing 2% glucose was added and the transformants were grown at 37°C for 1 hour. Cell transformants were pooled, the volume measured and an aliquot was plated onto 2YT containing 100 µg/ml amp plates to determine the total number of transformants.

[0754] Cells were grown to 0.5 (A_{600}) in 2YT-amp/2% glucose at 30°C at 250rpm (shaker). M13K07 helper phage (Biolab) was then added (moi =10), and the cells were incubated for 1 hr at the conditions described above. Cells were pelleted at 2500 x g for 10 min and the supernatant discarded. The cell pellet was re-suspended in the initial culture volume of media, containing 100 mg/mL amp and 50 mg/mL kanamycin and grown overnight at 30°C at 250rpm. The cells were then pelleted at 2500 x g for 10 min and the supernatant was transferred to another container and precipitated by adjusting the solution to 4% PEG, 500 mM NaCl, and chilled at 4°C for 1 hr before centrifugation at 10,000 g for 10 min. The pellet was re-suspended in TBS (1:100 of the initial culture volume). The phage was titered by infecting TG1 cells.

5'ATGGGCCCAGCCGGCCATGGCA(NNB)₅AAGCTGACGTGTCTGGCCAGTTATTGTTG
GCTGTTC(NNB)₅GCGGCCGCAGGTAGCTA3'

oligo A: 5'ATGGGCCCAGCCGGCCATG 3'

oligo B: 5'TAGCTACCTGCGGCCGC 3'

PCR conditions: 95°C 5min

95°C 30sec

65°C 30sec 7- 11x

72°C 30sec

72°C 5min

4°C ∞

Panning and the subsequent ELISA were performed as described above.

Example 8

hFIXa or hFVIIa binding across captured biotinylated compounds of the disclosure via immobilized streptavidin probes

[0755] The affinity of soluble human FIXa or soluble FVIIa to captured biotinylated peptides of the disclosure were measured using Streptavidin (SA) probes. Bio-Layer Interferometry (BLI) based measurements were obtained at 25°C with a ForteBio Octet 384 instrument using HBS-P buffer (10 mM HEPES, pH 7.4, 150 mM NaCl, 5 mM CaCl and 0.05% surfactant P20). Briefly, a layer of molecules attached to the tip of an optical fiber creates an interference pattern upon incident white light. The combination of the optical fiber and biocompatible surface creates the physical conditions needed for BLI, allowing the detection of changes in mass accumulation on the tip of the probe. This is measured by alterations in the spectral shift (nm). These changes are recorded as a binding profile, a continuous, real-time monitoring of the association and dissociation of interacting molecules.

[0756] All biotinylated peptides (20 µg/mL) were diluted in 4M guanidinium chloride and loaded across streptavidin (SA) biosensors for 120 sec, yielding approximately 0.5 – 1.0 nm binding on the reaction probes. Control SA probes were loaded with 4M guanidinium chloride in the absence of biotinylated peptide for reference subtraction. After loading, probes were incubated in HBS-P for 300 sec to establish a new baseline. Subsequently, biosensor probes were incubated in solutions of human FIXa or FVIIa (at 0, 2, 6, 20, 60, 200, 600, 2000 nM) for 1 hour at room temperature to establish an equilibrium state known as the association phase. The probes were then incubated in HBS-P buffer, permitting human FIXa or FVIIa to dissociate from the probe. This is described as the dissociation phase. The equilibrium KD was derived from the non-linear regression analysis of the subtracted data (Reaction probe minus Reference probe) using a 1:1 binding model with ForteBio software (Version 7.0).

[0757] hFIXa binding to immobilized compound 19 (compound 4 biotinylated at K18), RRAPGKLTCLASYCWLFK(PEG₂-Biotin)TGIA), was measured (steady state analysis). Association and dissociation time curves using various hFIXa concentrations were also recorded. In this experiment, the R_{max} for the binding of compound 19 to human FIXa

was found to be 1.65 ± 0.116 , and the dissociation constant (K_D) was found to be $120 \text{ nM} \pm 33 \text{ nM}$.

[0758] In a similar fashion, binding to human FVIIa (hFVIIa) was assessed. For example, hFVIIa binding to compound 25 was measured, and the K_D was found to be $250 \text{ nM} \pm 36 \text{ nM}$.

Example 9

Activated Partial Thromboplastin Time (aPTT) Assay

[0759] For the activated Partial Thromboplastin Time (aPTT) assay peptides to be tested were first diluted in 10% FVIII-deficient human plasma. For the aPTT* the peptides were diluted in 10% FVIII-deficient human plasma that contained traces of hFIXa (anywhere between 0 and 8.8 nM hFIXa depending on the assay) in Tris/NaCl/BSA buffer. The peptides were incubated for 10 minutes at room temperature and then assayed for clotting on a Sysmex instrument that recorded the clotting time observed for each sample. The times were compared to those of a standard curve derived from standards with known FVIII activity (U/mL) in order to determine the clotting outcome.

[0760] Clotting times for compound 1, compound 2, and compound A (scrambled control) were measured using the above described modified activated partial thromboplastin time (aPTT*) assay. Compounds were tested at 5, 10, and 20 μM . Results summarized in the Table 14, below, indicate that the decrease of clotting time observed for compounds of the present disclosure is FIXa dependent.

Table 14

	Compound 1			Compound 2			Compound A		Blank + FIXa
Conc. (μM)	20	10	5	20	10	5	20	10	-
Clotting time, (sec)	43.5;43.8	44.2;43.3	47.3;47.9	42.4;42.3	42.9;42.2	51.1;50.3	122.0;123.2	120.6;123.2	120.2;120.2

[0761] Clotting times for compound 1 and compound 2 were further measured in the presence of various FIXa concentrations, and activities were compared to a blank sample (containing no peptide). The results are summarized in Table 15, below:

Table 15

	Compound 1	Compound 2	Blank
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	Compound 1				Compound 2				Blank			
FIXa (nM)	1	2	4	8	1	2	4	8	1	2	4	8
Clotting time (sec)	105.8; 107.4	84.4; 82.8	50.9; 50.6	40.1; 40.6	86.8; 91.4	51.3; 49.6	46.0; 42.7	34.4; 37.1	119.8; 120.6	119; 120.4	114.6; 115.4	107.8; 108.4

[0762] Clotting times were measured in a modified activated partial thromboplastin time (aPTT*) assay for D-amino acid mutants of compound 1 and were compared to EC₅₀ values obtained using a FXa generation assay (compare Table 1). Results summarized in Table 16 below indicate a strong correlation between the FXa generation activity and the clotting time values measured for the compound 1 family of D-amino acid mutants. These results further confirm that D-amino acid replacement of the loop amino acids (positions 5 to 9), and certain C-terminal amino acids within compound 1 results in reduced biological activity compared to the native peptide.

Table 16

	D-amino acid Mutations of Compound 1, per position													
D-amino acid scan	1	2	3	4	5	6	7	8	9	10	11	12	WT	Blank
EC50, μ M	(+++)	(+++)	(++)	(++)	>5	>5	(+)	>5	(+)	(++)	(+)	(+)	(+++)	-
Clotting time, sec	60.6; 61.8	69.2; 69.8	79.4; 80.6	60.8; 61.8	115.6; 115.4	117.2; 117.8	117; 118	113.8; 115	115; 113.8	63.1; 68.8	96; 98.4	87.6; 87.8	47.9; 47.8	116; 117

[0763] Clotting times were measured for compound 3 in a modified activated partial thromboplastin time (aPTT*) assay in the absence and presence of various concentrations of exogenous hFIXa as indicated. In this assay, compound 3 reduces clotting time in a FIXa-dependent manner. The results are summarized in the table 17, below:

Table 17

	Compound 3			Blank		
FIXa, nM	0.5	2.2	8.8	0.5	2.2	8.8
Clotting time, sec	115.6; 115.2	90.8; 89	45.4; 41.9	122.6; 124	119.8; 119.8	106.8; 106.8

Example 10

Phospholipid Dependency

[0764] (a) EC_{50} and V_{max} values were measured for various compounds of the present disclosure in a FXa generation assay (Example 2) in the absence and presence of phospholipids (PL). The results indicate limited phospholipid dependence for the EC_{50} and V_{max} values. Results are shown in Table 18, below:

Table 18

Compound	PL	EC_{50} (μ M)	V_{max} (mOD/min min)
1	Yes	0.49	3.98
1	No	0.40	2.60
2	Yes	0.23	4.22
2	No	0.20	3.05
3	Yes	0.125	3.90
3	no	0.115	2.77

[0765] (b) In another experiment, the impact of phospholipids on hFIXa in the presence of FVIIIa in the FXa generation assay was tested. Assay components of the control were FVIIIa, FIXa, PL, FX, and FXa substrate. FIXa (100 nM) was pre-incubated with anti-FIX antibodies (1000 nM) specific for the Gla domain of FIX or IgG control for 20 minutes on ice. Following this incubation the activity of FIXa was assayed using a FXa generation assay in the presence of FVIIIa.

[0766] (b.1.) FVIII (4 nM) was activated with thrombin (0.5 nM) for 5 minutes at RT. Hirudin (5 nM) was then added to the reaction to inhibit thrombin activity. FVIIIa (2 nM) and FIXa or FIXa pre-treated with antibodies (4 nM) were then incubated at RT for 10 minutes in the absence or presence of 200 μ M phospholipid (PL) vesicles to form the Xase complex. Following this 50 μ l of the reaction was mixed with FX (100 nM) and a FXa-specific chromogenic substrate (0.5 mM) and the absorbance was monitored at 405 nm. The FXa generation rates (nM/min) were measured. The results relative to the control (100 %) are summarized in Table 19, below:

Table 19

Experiment	Percentage of control
Control	100
No FIXa	0.0
No PL	0.01
Anti-FIX IgG (Gla)	12.5
control IgG	98.6

In this experiment, FXa generation depends on the presence of phospholipids. Removal of the phospholipid vesicles reduces the FXa generation rate by 10,000 fold. FXa generation further depends on FIXa. In the presence of anti-FIXa antibodies, FXa generation rate is reduced by 8-fold.

[0767] (b.2.) In another experiment the impact of phospholipids on hFIXa in the presence of compound 3 in the FXa generation assay was measured.

[0768] For the assay involving a compound of the invention, FIXa (100 nM) was pre-incubated with anti-FIX antibodies (1000 nM) specific for the Gla domain of FIX. Following this incubation FIXa (10 nM) was mixed with compound 3 (1 μ M) in the presence of FX (100 nM), calcium and phospholipids from the Coatest SP FVIII chromogenic assay kit. Formation of FXa was monitored using a chromogenic substrate as described in the FXa generation assay directly above (see b.1.). The results relative to the control (presence of PL, 100 %) are summarized in Table 20, below:

Table 20

	Experiment	Percentage of control
compound 3	+ PL	100
compound 3	- PL	90
compound 3	+ PL + anti-FIX IgG (Gla)	58.0
water	+ PL	0.62
water	- PL	1.85
water	+ PL + anti-FIX IgG (Gla)	1.48

[0769] In this experiment, FXa generation does not depend on the presence of phospholipids and is significantly less dependent on a functional Gla domain of FIXa than in the FVIIIa experiment above (The Gla domain of FIXa is responsible for

phospholipid binding). Treatment with anti-FIX Gla antibody reduces the FIXa generation rate by less than 2-fold. In this experiment, water was used as the control, which did essentially not generate any FIXa.

[0770] The above results indicate that compound 3 increases the catalytic activity of FIXa with limited phospholipid dependency (*i.e.*, the absence of phospholipids in the above assay system does not substantially decrease the enhancement of FIXa activity by compounds of the present disclosure (*e.g.*, compound 3).

In similar experiments the impact of phospholipids on hFVIIa in the FIXa generation assay was tested in the absence and presence of compound 5. Results indicate that while the increase of FIXa catalytic activity by compound 5 is marginally phospholipid dependent (about 80 % of control in the absence of PL), the increase of FVIIa activity by compound 5 is significantly phospholipid dependent; *i.e.*, significantly reduced in the absence of PL (about 10 % of control). These results indicate that compounds of the present disclosure (*e.g.*, compound 5) enhance FVIIa activity in a phospholipid dependent manner.

Example 11

A shared FIXa binding site for the pro-coagulant compounds and heparin

[0771] Procoagulant peptides enhance the activity of hFIXa by interacting near the FVIIIa binding site on the protease domain.

[0772] The interaction between certain pro-coagulant peptides and FIXa was studied by hydrogen/deuterium exchange mass spectrometry (H/DX-MS). The data revealed that the pro-coagulant peptides interact near the 330 loop (FIX numbering) on FIXa which is close to the postulated FVIIIa binding site, as well as the heparin binding site.

[0773] Heparin is believed to have two functions in the FIXa inactivation: Heparin catalyzed inhibition of FIXa by antithrombin (heparin causes a conformational change of antithrombin), and heparin mediated bridging of FIXa and antithrombin (PNAS 2010, 107, 645-650; J. Biol. Chem. 2002, 277, 50756-50760; J. Biol. Chem. 2003, 278, 35767-35774; J. Biol. Chem. 1998, 273, 120).

Heparin accelerated FIXa-AT complex formation assay

[0774] Competition studies between the pro-coagulant peptides and heparin: compounds of the present disclosure were tested in a heparin competition assay in an effort to further map the peptide's binding site on FIXa. FIXa inactivation by antithrombin (AT) was assessed by gel electrophoresis in the presence of heparin and varying procoagulant peptide concentrations. The rate of FIXa-AT complex formation was quantified over a 15 minute time course. Test samples contained 2 μ M human FIXa (Haemotologic Technologies Inc.); 20 μ M antithrombin (Haemotologic Technologies Inc.); 0, 1, 10, or 100 nM heparin (Heparin Sodium Injection USP APPP Pharmaceuticals LLC) and 0, 0.1, 1 or 10 μ M of compound 5 in 50 mM Tris pH 7.4, 0.1 M NaCl, 10 mM CaCl_2 buffer. The samples were incubated at 37 $^{\circ}\text{C}$ in a water bath and 12.5 μ L aliquots were removed at each time point (0.5, 2, 5, 10, 15 min) and immediately mixed with 12.5 μ L SDS 2x non-reducing sample buffer. The samples were heated at 90 $^{\circ}\text{C}$ for 3 min and loaded onto a 4-20% BioRad gel (20 μ L/lane). The gel was run at 300V for 25 min, and the bands were quantified using Quantity One software from BioRad.

[0775] The rate of heparin-catalyzed FIXa-antithrombin (AT) complex formation in the absence or presence of compound 5 was measured. In the absence of heparin, the inactivation of FIXa by AT was unaffected by compound 5. At heparin concentrations that significantly accelerated the AT inactivation of FIXa (e.g., about 100 nM), certain compounds of the present disclosure (e.g., compound 5) inhibited the heparin-catalyzed FIXa-AT complex formation (e.g., in a concentration-dependent manner). The results show that in the absence of heparin, the inactivation of FIXa by AT was unaffected by the presence of compound 5. However, at heparin concentrations (100 nM) that significantly accelerated the AT inactivation of FIXa, compound 5 inhibited the FIXa-AT complex formation in a concentration-dependent manner. Results suggest a shared FIXa binding site for compounds of the present disclosure (e.g., compound 5) and heparin. Results are summarized in Table 21, below:

Table 21

	% FIXa-AT complex formation in the presence of Compound 5			
Compound 5	0 μ M	0.1 μ M	1 μ M	10 μ M
0 min	0%	0%	0%	0%
0.5 min	52%	57%	30%	18%
2 min	71%	72%	45%	28%
5 min	79%	82%	55%	41%
10 min	81%	82%	70%	54%
15 min	83%	83%	74%	62%

Example 12

**Additive effect between compounds of the present disclosure
and FIX-Fc or FVIIa-Fc**

[0776] The additive effect between the compounds of the present disclosure and FVIIa-Fc or FIX-Fc was assessed by a rotational thromboelastometry (ROTEM®) assay as described in Example 4 using either FIX-deficient plasma or FVIII-deficient plasma.

Additive effect between compound 5 and FVIIa-Fc

[0777] The effect of 5, 10 and 20 IU/mL of FVIIa-Fc was tested in the absence or presence of 2.5 or 5 μ M of compound 5 in FVIII-deficient plasma.

[0778] In the absence of compound 5, FVIIa-Fc (10 or 20 IU/mL) reduced the clotting time to 1058 and 581 seconds and improved the α -angle to 12 and 21 degrees, respectively. The baseline clotting time was 2439 seconds and α -angle 9 degrees with trigger alone. In the presence of compound 5 (2.5 μ M), the clotting time was further reduced to 303.5 and 115 seconds and the α -angle was further increased to 38.5 and 63 degrees, respectively. Similar trends were observed for 5 IU/mL of FVIIa-Fc and 5 μ M of compound 5.

Additive effect between compound 5 and FIX-Fc

[0779] The additive effect between FIX-Fc and compound 5 was also evaluated in FIX-deficient plasma. FIX-Fc at 0.25 IU/mL reduced the clotting time to 1173 seconds and the α -angle to 26 degrees compared to the baseline clotting time of 3204 seconds and α -angle of 10 degrees with trigger alone. In the presence of 2.5 μ M compound 5, the

clotting time was further reduced to 876.5 seconds and the α -angle was further enhanced to 39.5 degrees. All values listed in this example are averages of duplicates.

The above results show that compounds of the present disclosure enhance the procoagulant effect of FVIIa-Fc and FIX-Fc in FVIII- and FIX-deficient plasma, respectively.

[0780] Because a combination of FVIIa-Fc and a compound of the present disclosure reduces clotting time and increases α -angle further than either component alone, FVIIa-Fc and compounds of the present disclosure are suitable to be used in conjunction (*i.e.*, in a combination therapy, e.g. to treat hemophilia, such as hemophilia A).

[0781] Likewise, because a combination of FIX-Fc and a compound of the present disclosure reduces clotting time and increases α -angle further than either component alone, FIX-Fc and compounds of the present disclosure are suitable to be used in conjunction (*i.e.*, in a combination therapy, e.g. to treat hemophilia, such as hemophilia A).

[0782] The above results also indicate that conjugates between FVIIa, FVIIa-Fc, FIX, or FIX-Fc with compounds of the present disclosure (e.g., cleavable conjugates) are useful in the therapy of hemophilia (e.g., hemophilia A). Exemplary conjugates are disclosed herein.

Example 13

Compounds of the present disclosure have no effect on platelet aggregation

[0783] In order to determine whether the compounds of the present disclosure cause platelet aggregation directly, compound 5 and compound 22 were tested using an aggregometer (Platelet Aggregation Profiler PAP8 v.2.0 from BIO/DATA Corporation). ADP was used as a positive control for platelet aggregation.

[0784] For example, compound 5 did not exhibit a significant effect on adenosine 5'-diphosphate-activated (ADP-activated) platelets when used at a concentration of up to 10 μ M, and did exhibit only a moderate effect on ADP-activated platelets when used at a very high concentration of 30 μ M. Furthermore, compound 5 did not induce platelet aggregation even at a high concentration of about 30 μ M. Compound 22 did also not induce platelet aggregation at 10 μ M and 30 μ M.

Example 14

Preparation of FVIIa Conjugates

- [0785] Cloning was performed using Rapid DNA Ligation Kit from Roche Diagnostics (cat # 11635379001) following manufacturer's guidelines. Briefly, 10-50 ng of vector or fragment DNA were added to a final volume of 10 ul ligation mixture containing 1 x Rapid DNA Ligation buffer and 1 ul Rapid DNA Ligase. Reaction was allowed to proceed for 5-30 min at room temperature and placed on ice. 2 ul of ligation reaction were transformed in 50 ul Mach 1 E. coli competent cells from Invitrogen (cat # C869601) and plated on an appropriate antibiotic for selection.
- [0786] The 3.1 kb DNA fragment comprising the region from HindIII to EcoRI of pSYN-FVII-171 was synthesized and subcloned into the HindIII/EcoRI sites of pcDNA4 vector (Invitrogen) to generate pSYN-FVII-171.
- [0787] For expression of FVII-171, HEK-293-F cells were grown in suspension in Freestyle media (Invitrogen) supplemented with vitamin K3 (Sigma Aldrich, St. Louis, MO) to 2 µg/liter (growth media) as suspension cells at 37° C/10% CO₂. Cells were subcultured every three to four days by seeding at cell density of 5x10⁵ cells/ml.
- [0788] Twenty-four hours prior to transfection cells were seeded at a density of 7x10⁵ cells/ml in growth media. On the day of transfection, a transfection solution was made with a volume equal to 5% of the total volume of the cell culture to be transfected. In the transfection solution DNA was added (final concentration 20 mg/L) to a freshly made solution of PEI (60 mg/L) in growth media. The solution was swirled for 30 seconds and incubated for five minutes at room temperature before adding directly to the cell culture. Four hours later a volume equal to the cell culture volume of OptiCHO (Invitrogen) supplemented with vitamin K3 and 200 mM L-glutamine was added to the cells. The cell culture was allowed to grow as shown above and daily media samples were taken to assess protein expression. On the day of harvest, the cells were spun down and the media filtered in preparation for protein purification or protein analysis by protein A pulldown. For expression of FVII-171, a plasmid encoding FVII-171 was cotransfected with a plasmid encoding the propeptide endopeptidase PC5 or PACE to ensure cleavage of the propeptide endopeptidase sites in the linker connecting the Fc to compound and between the HC and LC of FVII (Figure 9).

FVII-171 DNA sequence (SEQ ID NO: 830):

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1   ATGGTCTCCC AGGCCCTCAG GCTCCTCTGC CTTCTGCTTG GGCTTCAGGG CTGCCTGGCT
61  GCAGTCTTCG TAACCCAGGA GGAAGCCCAC GGCGTCCTGC ACCGGCGCCG GCGCGCCAAC
121 GCGTTCTCTG AGGAGCTGCG GCCGGGCTCC CTGGAGAGGG AGTGCAAGGA GGAGCAGTGC
181 TCCTTCGAGG AGGCCCGGGA GATCTTCAAG GACGCGGAGA GGACGAAGCT GTTCTGGATT
241 TCTTACAGTG ATGGGGACCA GTGTGCCTCA AGTCCATGCC AGAATGGGGG CTCCTGCAAG
301 GACCAGCTCC AGTCCTATAT CTGCTTCTGC CTCCCTGCCT TCGAGGGCCG GAACTGTGAG
361 ACGCACAAAG ATGACCAGCT GATCTGTGTG AACGAGAACG GCGGCTGTGA GCAGTACTGC
421 AGTGACCACA CGGGCACCAA GCGCTCCTGT CGGTGCCACG AGGGGTACTC TCTGCTGGCA
481 GACGGGGTGT CCTGCACACC CACAGTTGAA TATCCATGTG GAAAAATACC TATTCTAGAA
541 AAAAGAAATG CCAGCAAACC CCAAGGCCGA AGGAAGAGGA GGAAGAGGAT TGTGGGGGGC
601 AAGGTGTGCC CCAAAGGGGA GTGTCCATGG CAGGTCTGTG TGTGTGGTAA TGGAGCTCAG
661 TTGTGTGGGG GGACCCTGAT CAACACCATC TGGGTGGTCT CCGCGGCCCA CTGTTTCGAC
721 AAAATCAAGA ACTGGAGGAA CCTGATCGCG GTGCTGGGCG AGCACGACCT CAGCGAGCAC
781 GACGGGGATG AGCAGAGCCG GCGGGTGGCG CAGGTCATCA TCCCCAGCAC GTACGTCCCG
841 GGCACCACCA ACCACGACAT CGCGCTGCTC CGCCTGCACC AGCCCGTGGT CCTCACTGAC
901 CATGTGGTGC CCCTCTGCCT GCCCGAACGG ACGTTCTCTG AGAGGACGCT GGCCTTCGTG
961 CGCTTCTCAT TGGTCAGCGG CTGGGGCCAG CTGCTGGACC GTGGCGCCAC GGCCCTGGAG
1021 CTCATGGTCC TCAACGTGCC CCGCTGATG ACCCAGGACT GCCTGCAGCA GTCACGGAAG
1081 GTGGGAGACT CCCCAAATAT CACGGAGTAC ATGTTCTGTG CCGGCTACTC GGATGGCAGC
1141 AAGGACTCCT GCAAGGGGGA CAGTGGAGGC CCACATGCCA CCCACTACCG GGGCACGTGG
1201 TACCTGACGG GCATCGTCAG CTGGGGCCAG GGCTGCGCAA CCGTGGGGCA CTTTGGGGTG
1261 TACACCAGGG TCTCCCAGTA CATCGAGTGG CTGCAAAAGC TCATGCGCTC AGAGCCACGC
1321 CCAGGAGTCC TCCTGCGAGC CCCATTTCCC GGTGGCGGTG GCTCCGGCGG AGGTGGGTCC
1381 GGTGGCGGCG GATCAGGTGG GGGTGGATCA GGCGGTGGAG GTTCCGGTGG CGGGGGCTCC
1441 GACAAAACTC ACACATGCCC ACCGTGCCCC GCTCCGGAAC TCCTGGGAGG ACCGTCAGTC
1501 TTCTCTTTCC CCCCAAAACC CAAGGACACC CTCATGATCT CCCGGACCCC TGAGGTCACA
1561 TGCGTGGTGG TGGACGTGAG CCACGAAGAC CCTGAGGTCA AGTTCAACTG GTACGTGGAC
1621 GGCGTGGAGG TGCATAATGC CAAGACAAAG CCGCGGGAGG AGCAGTACAA CAGCACGTAC
1681 CGTGTGGTCA GCGTCCTCAC CGTCCTGCAC CAGGACTGGC TGAATGGCAA GGAGTACAAG
1741 TGCAAGGTCT CCAACAAAGC CCTCCCAGCC CCCATCGAGA AAACCATCTC CAAAGCCAAA
1801 GGGCAGCCCC GAGAACCACA GGTGTACACC CTGCCCCCAT CCCGGGATGA GCTGACCAAG
1861 AACCAGGTCA GCCTGACCTG CCTGGTCAAA GGCTTCTATC CCAGCGACAT CGCCGTGGAG
1921 TGGGAGAGCA ATGGGCAGCC GGAGAACAAC TACAAGACCA CGCTCCCGT GTTGGACTCC
1981 GACGGCTCCT TCTTCTCTA CAGCAAGCTC ACCGTCGACA AGAGCAGGTG GCAGCAGGGG
2041 AACGTCTTCT CATGCTCCGT GATGCATGAG GCTCTGCACA ACCACTACAC GCAGAAGAGC
2101 CTCTCCCTGT CTCCGGGTAA ACGGCGCCGC CGGAGCGGTG GCGGCGGATC AGGTGGGGGT
2161 GGATCAGGCG GTGGAGGTTC CGGTGGCGGG GGATCTGGCG GTGGAGGTTC CGGTGGGGGT
2221 GGATCCAGGA AGAGGAGGAA GAGGGGCCCC CGGATCCGGA CAGTGGGGCC CGGCAGCCGG

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2281  AGCGCCAGCG GCAAGCTGAC CTGCCTGGCC AGCTACTGCT GGCTGTTCTG GACCGGCATC
2341  GCCGGTGGCG GTGGATCCGG CGGAGGTGGG TCCGGTGGCG GCGGATCAGG TGGGGGTGGA
2401  TCAGGCGGTG GAGGTTCGGG TGGCGGGGGA TCAGACAAAA CTCACACATG CCCACCGTGC
2461  CCAGCACCGG AACTCCTGGG CGGACCGTCA GTCTTCCTCT TCCCCCAAAA ACCCAAGGAC
2521  ACCCTCATGA TCTCCCGGAC CCCTGAGGTC ACATGCGTGG TGGTGGACGT GAGCCACGAA
2581  GACCCTGAGG TCAAGTTCAA CTGGTACGTG GACGGCGTGG AGGTGCATAA TGCCAAGACA
2641  AAGCCGCGGG AGGAGCAGTA CAACAGCACG TACCGTGTGG TCAGCGTCCT CACCGTCTCTG
2701  CACCAGGACT GGCTGAATGG CAAGGAGTAC AAGTGCAAGG TCTCCAACAA AGCCCTCCCA
2761  GCCCCATCG AGAAAACCAT CTCAAAGCC AAAGGGCAGC CCCGAGAACC ACAGGTGTAC
2821  ACCCTGCCCC CATCCCGGGA TGAGCTGACC AAGAACCAGG TCAGCCTGAC CTGCCTGGTC
2881  AAAGGCTTCT ATCCCAGCGA CATCGCCGTG GAGTGGGAGA GCAATGGGCA GCCGGAGAAC
2941  AACTACAAGA CCACGCCTCC CGTGTTGGAC TCCGACGGCT CCTTCTTCCT CTACAGCAAG
3001  CTCACCGTGG ACAAGAGCAG GTGGCAGCAG GGGAACGTCT TCTCATGCTC CGTGATGCAT
3061  GAGGCTCTGC ACAACCACTA CACGCAGAAG AGCCTCTCCC TGTCTCCGGG TAAATGA

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FVII-171 amino acid sequence (SEQ ID NO: 831)

[0789] Signal sequence is underlined, propeptide is double underlined, furin cleavage site separating light chain and heavy chain is in dotted underline, linker region connecting heavy chain to Fc region is in dashed underline, furin cleavage site separating Fc and linker is in thick underline, furin cleavage site separating linker and compound 21 is in wave underline and linker region separating compound 21 and Fc is in dot-dot-dash underline.

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1    MVSQALRLLC LLLGLQGCLA AVFVTQEEAH GVLHRRRRAN AFLEELRPGS LERECKEEQC
61   SFEEAREIFK DAERTKLFWI SYSDGDQCAS SPCQNGGSK DQLQSYICFC LPAFEGRNCE
121  THKDDQLICV NENGGCEQYC SDHTGTRKSC RCHEGYSLA DGVSCTPTVE YPCGKIPILE
181  KRNASKPQGR RKRRKRIVGG KVC PKGEC PW QVLLLVNGAQ LCGGTINTI WVVSAAHCFD
241  KIKNWRNLIA VLGEHDLSEH DGDEQSRRVA QVIIPSTYVP GTTNHDIAL RLHQPVVLT D
301  HVVPLCLPER TFSERTLAFV RFSLVSGWGQ LLDRGATALE LMVLNVPRLM TQDCLQQSRK
361  VGDSPNITEY MFCAGYSDGS KDSCKGDSGG PHATHYRG TW YLTGIVSWGQ GCATVGHFGV
421  YTRVSQYIEW LQKLMRSEPR PGVLLRAPFP GGGSGGGGS GGGSGGGGS GGGSGGGGS
481  DKTHTCPPCP APELLGGPSV FLFPPKPKDT LMISRTPEVT CVVVDVSHED PEVKFNWYVD
541  GVEVHNAKTK PREEQYNSTY RVVSVLTVLH QDWLNGKEYK CKVSNKALPA PIEKTISKAK
601  GQPREPQVYT LPPSRDELTK NQVSLTCLVK GFYPSDIAVE WESNGQPENN YKTTTPVLD S
661  DGSFFLYSKL TVDKSRWQQG NVFSCSV MHE ALHNHYTQKS LSLSPGKRRR RSGGGSGGGG
721  GSGGGSGGGG GSGGGSGGGG GSRKRRKRG RIRTVGPGSR SASGKLTCLA SYCWLFWTGI
781  AGGGGSGGGG SGGGGSGGGG SGGGSGGGG SDKTHTCPPC PAPELLGGPS VFLFPPKPKD
841  TLMISRTPEV TCVVVDVSHE DPEVKFNWYV DGVEVHNAKT KPREEQYNST YRVVSVLTVL

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901 HQDWLNGKEY KCKVSNKALP APIEKTISKA KGQPREPQVY TLPPSRDELT KNQVSLTCLV
 961 KGFYPSDIAV EWESNGQPEN NYKTTTPVLD SDGSFFLYSK LTVDKSRWQQ GNVFSCSVMH
 1021 EALHNHYTQK SLSLSPGK

Example 15

Preparation of FIX Conjugates

FIX-124 construction

[0790] Cloning and expression was performed as described in Example 14.

[0791] The 3.4 kb DNA fragment comprising the region from HindIII to EcoRI of pSYN-FIX-124 was synthesized and subcloned into the HindIII/EcoRI sites of pcDNA4 vector (Invitrogen) to generate pSYN-FIX-124 (FIX-Fc-compound 21 conjugate).

FIX-124 DNA sequence (SEQ ID NO: 832)

1 ATGCAGCGCG TGAACATGAT CATGGCAGAA TCACCAGGCC TCATCACCAT CTGCCTTTTA
 61 GGATATCTAC TCAGTGCTGA ATGTACAGTT TTTCTTGATC ATGAAAACGC CAACAAAATT
 121 CTGAATCGGC CAAAGAGGTA TAATTCAGGT AAATTGGAAG AGTTTGTTCA AGGGAATCTA
 181 GAGAGAGAAT GTATGGAAGA AAAGTGTAGT TTTGAAGAAG CACGAGAAGT TTTTGAAAAC
 241 ACTGAAAGAA CAACTGAATT TTGGAAGCAG TATGTTGATG GAGATCAGTG TGAGTCCAAT
 301 CCATGTTTTAA ATGGCGGCAG TTGCAAGGAT GACATTAATT CCTATGAATG TTGGTGTCCC
 361 TTTGGATTTG AAGGAAAGAA CTGTGAATTA GATGTAACAT GTAACATTAA GAATGGCAGA
 421 TGCGAGCAGT TTTGTAAAAA TAGTGCTGAT AACAAAGGTGG TTTGCTCCTG TACTGAGGGA
 481 TATCGACTTG CAGAAAACCA GAAGTCCTGT GAACCAGCAG TGCCATTTCC ATGTGGAAGA
 541 GTTTCTGTTT CACAACTTC TAAGCTCACC CGTGCTGAGA CTGTTTTTCC TGATGTGGAC
 601 TATGTAAATT CTAAGAAGC TGAAACCATT TTGGATAACA TCACTCAAAG CACCCAATCA
 661 TTTAATGACT TCACTCGGGT TGTGTTGGTGA GAAGATGCCA AACCAGGTCA ATTCCCTTGG
 721 CAGGTTGTTT TGAATGGTAA AGTTGATGCA TTCTGTGGAG GCTCTATCGT TAATGAAAAA
 781 TGGATTGTAA CTGCTGCCCC CTGTGTTGAA ACTGGTGTTA AAATTACAGT TGTCGCAGGT
 841 GAACATAATA TTGAGGAGAC AGAACATACA GAGCAAAAAGC GAAATGTGAT TCGAATTATT
 901 CCTCACCACA ACTACAATGC AGCTATTAAT AAGTACAACC ATGACATTGC CCTTCTGGAA
 961 CTGGACGAAC CCTTAGTGCT AAACAGCTAC GTTACACCTA TTTGCATTGC TGACAAGGAA
 1021 TACACGAACA TCTTCCTCAA ATTTGGATCT GGCTATGTAA GTGGCTGGGG AAGAGTCTTC
 1081 CACAAAGGGA GATCAGCTTT AGTTCTTCAG TACCTTAGAG TTCCACTTGT TGACCGAGCC
 1141 ACATGTCTTC GATCTACAAA GTTCACCATC TATAACAACA TGTTCTGTGC TGGCTTCCAT
 1201 GAAGGAGGTA GAGATTCATG TCAAGGAGAT AGTGGGGGAC CCCATGTTAC TGAAGTGGAA
 1261 GGGACCAGTT TCTTAACTGG AATTATTAGC TGGGGTGAAG AGTGTGCAAT GAAAGGCAAA
 1321 TATGGAATAT ATACCAAGGT GTCCCGGTAT GTCAACTGGA TTAAGGAAAA AACAAAGCTC
 1381 ACTGACAAAA CTCACACATG CCCACCGTGC CCAGCTCCGG AACTCCTGGG AGGACCGTCA
 1441 GTCTTCCTCT TCCCCCAAA ACCCAAGGAC ACCCTCATGA TCTCCCGGAC CCCTGAGGTC

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1501  ACATGCGTGG TGGTGGACGT GAGCCACGAA GACCCTGAGG TCAAGTTCAA CTGGTACGTG
1561  GACGGCGTGG AGGTGCATAA TGCCAAGACA AAGCCGCGGG AGGAGCAGTA CAACAGCACG
1621  TACCGTGTGG TCAGCGTCCT CACCGTCTCG CACCAGGACT GGCTGAATGG CAAGGAGTAC
1681  AAGTGCAAGG TCTCCAACAA AGCCCTCCCA GCCCCATCG AGAAAACCAT CTCCAAAGCC
1741  AAAGGGCAGC CCCGAGAACC ACAGGTGTAC ACCCTGCCCC CATCCCGGGA TGAGCTGACC
1801  AAGAACCAGG TCAGCCTGAC CTGCCTGGTC AAAGGCTTCT ATCCCAGCGA CATCGCCGTG
1861  GAGTGGGAGA GCAATGGGCA GCCGGAGAAC AACTACAAGA CCACGCCTCC CGTGTTGGAC
1921  TCCGACGGCT CCTTCTTCCT CTACAGCAAG CTCACCGTCG ACAAGAGCAG GTGGCAGCAG
1981  GGAACGTCT TCTCATGCTC CGTGATGCAT GAGGCTCTGC ACAACCACTA CACGCAGAAG
2041  AGCCTCTCCC TGTCTCCGGG TAAACGGCGC CGCCGGAGCG GTGGCGGCGG ATCAGGTGGG
2101  GGTGGATCAG GCGGTGGAGG TTCCGGTGGC GGGGGATCTG GCGGTGGAGG TTCCGGTGGG
2161  GGTGGATCCA GGAAGAGGAG GAAGAGGGGC CCCCGGATCC GGACAGTGGG CCCCGGCAGC
2221  CGGAGCGCCA GCGGCAAGCT GACCTGCCTG GCCAGCTACT GCTGGCTGTT CTGGACCGGC
2281  ATCGCCGGTG GCGGTGGATC CGGCGGAGGT GGGTCCGGTG GCGGCGGATC AGGTGGGGGT
2341  GGATCAGGCG GTGGAGGTTT CGGTGGCGGG GGATCAGACA AAATCACAC ATGCCACCG
2401  TGCCCAGCAC CGGAATCCTT GGGCGGACCG TCAGTCTTCC TCTTCCCCC AAAACCCAAG
2461  GACACCCTCA TGATCTCCCG GACCCCTGAG GTCACATGCG TGGTGGTGA CGTGAGCCAC
2521  GAAGACCCTG AGGTCAAGTT CAACTGGTAC GTGGACGGCG TGGAGGTGCA TAATGCCAAG
2581  ACAAAGCCGC GGGAGGAGCA GTACAACAGC ACGTACCGTG TGGTCAGCGT CCTCACCCTC
2641  CTGACACAGG ACTGGCTGAA TGGCAAGGAG TACAAGTGCA AGGTCTCCAA CAAAGCCCTC
2701  CCAGCCCCCA TCGAGAAAAC CATCTCCAAA GCCAAAGGGC AGCCCCGAGA ACCACAGGTG
2761  TACACCCTGC CCCCATCCCG GGATGAGCTG ACCAAGAACC AGGTCAGCCT GACCTGCCTG
2821  GTCAAAGGCT TCTATCCCAG CGACATCGCC GTGGAGTGGG AGAGCAATGG GCAGCCGGAG
2881  AACAATAACA AGACCACGCC TCCCGTGTTG GACTCCGACG GTCCTTCTT CCTCTACAGC
2941  AAGCTACCG TGGACAAGAG CAGGTGGCAG CAGGGGAACG TCTTCTCATG CTCCGTGATG
3001  CATGAGGCTC TGCACAACCA CTACACGCAG AAGAGCCTCT CCCTGTCTCC GGGTAAATGA

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FIX-124 amino acid sequence (SEQ ID NO: 833)

[0792] Signal sequence is underlined, propeptide is double underlined, Fc separating Factor IX and furin cleavage site is in dotted underline, linker separating 2 furin cleavage sites is in wave underline, compound 21 separating furin cleavage site and linker is in dased underline, Fc is in dot-dot-dash underline.

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1    MQRVNMIMAE SPGLITICLL GYLLSAECTV FLDHENANKI LNRPKRYNSG KLEEFVQGNL
61   ERECMEEKCS FEEAREVFEN TERTTEFWKQ YVDGDQCESN PCLNGGSKCD DINSYECWCP
121  FGFEGKNCEL DVTCNIKNGR CEQFCKNSAD NKVVCSTEG YRLAENQKSC EPAVPFPCGR
181  VSVSQTSKLT RAETVFPDVD YVNSTEAETI LDNITQSTQS FNDFTRVVGG EDAKPGQFPW
241  QVVLNGKVDA FCGGSIVNEK WIVTAAHCVE TGVKITVVAG EHNIEETEHT EQKRNVIIRI
301  PHHNYNAAIN KYNHDIALLE LDEPLVLNSY VTPICIADKE YTNIFLKFGS GYVSGWGRVF

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361  HKGRSALVLQ YLRVPLVDRA TCLRSTKFTI YNNMFCAGFH EGGRDSCQGD SGGPHVTEVE
421  GTSFLTGIIS WGEECAMKGK YGIYTKVSRV VNWIKETKL TDKTHTCPPC PAPELLGGPS
481  VFLFPPKPKD TLMISRTPEV TCVVVDVSHE DPEVKFNWYV DGVEVHNAKT KPREEQYNST
541  YRVVSVLTVL HQDWLNGKEY KCKVSNKALP APIEKTISKA KGQPREPQVY TLPPSRDELT
601  KNQVSLTCLV KGFYPSDIAV EWESNGQOPEN NYKTTTPVLD SDGSFFLYSK LTVDKSRWQQ
661  GNVFSCSVMH EALHNHYTQK SLSLSPGKRR RRSGGGGSGG GSGGGGGSGG GSGGGGGSGG
721  GGSRRKRRKG PRIRTVGPGS RSASGKLTCL ASYCWLFWTG IAGGGGSGGG GSGGGGGSGG
781  GSGGGGGSGG GSDKTHTCPP CPAPELLGGP SVFLFPPKPK DTLMISRTPE VTCVVVDVSH
841  EDPEVKFNWY VDGVEVHNAK TKPREEQYNS TYRVVSVLTV LHDWLNGKE YKCKVSNKAL
901  PAPIEKTISK AKGQPREPQV YTLPPSRDEL TKNQVSLTCL VKGFYPSDIA VEWESNGQPE
961  NNYKTTTPVL DSDGSFFLYS KLTVDKSRWQ QGNVFSCSVM HEALHNHYTQ KSLSLSPGK

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[0793] The TGA activity of FIX-124 was compared to FIX-Fc (without peptide).

[0794] TGA activity of FIX-124 was measured in pooled FIX-deficient HRF plasma with the results summarized in Table 22, below:

Table 22

	TGA activity of FIX-124 and FIXFc in FIX-deficient plasma				
Conc. (nM)	0	6.7	33.5	67	134
FIX-124, nM thrombin peak height	32.48; 29.88	97.98; 96.48	111.59; 102.01	106.71; 106.15	103.19; 101.87
FIXFc, nM thrombin peak height		35.11; 35.42	40.42; 40.93	46.45; 48.00	75.08; 80.78

[0795] TGA activity of FIX-124 was measured in FVIII-deficient Precision Biologics plasma with the results summarized in Table 23, below:

Table 23

	TGA activity of FIX-124 and FIXFc in FVIII-deficient plasma			
Conc. (nM)	0	6.7	33.5	134
FIX-124, nM thrombin peak height	18.24; 14.64	32.09; 29.17	39.72; 38.02	40.79; 36.89
FIXFc, nM thrombin peak height		21.06; 20.56	18.54; 18.4	22.23; 21.83

Example 16**Preparation of Platelet-Targeting Moiety Conjugates****pSYN-Fc-046 construction**

[0796] Cloning and expression was performed as described in Example 14.

[0797] The 2.6 kb DNA fragment comprising the region from HindIII to EcoRI of pSYN-Fc-046 was synthesized and subcloned into the HindIII/EcoRI sites of pcDNA4 vector (Invitrogen) to generate pSYN-Fc-046 (PDG13-Fc-compound 21 conjugate).

Fc-046 DNA sequence (SEQ ID NO: 834)

```
1   ATGGAGACAG ACACACTCCT GCTATGGGTA CTGCTGCTCT GGGTTCCAGG TTCCACTGGT
61  GGCCCCCGGA TTCGGACAGT GGGCCCCGGC AGCCGGAGCG CCAGCGGCAA GCTGACCTGC
121 CTGGCCAGCT ACTGCTGGCT GTTCTGGACC GGCATCGCCG GTGGCGGTGG ATCCGGCGGA
181 GGTGGGTCCG GTGGCGGCGG ATCAGGTGGG GGTGGATCAG GCGGTGGAGG TTCCGGTGGC
241 GGGGGATCAG ACAAACCTCA CACATGCCCA CCGTGCCAG CTCCGGAACCT CCTGGGAGGA
301 CCGTCAGTCT TCCTCTTCCC CCCAAAACCC AAGGACACCC TCATGATCTC CCGGACCCCT
361 GAGGTCACAT GCGTGGTGGT GGACGTGAGC CACGAAGACC CTGAGGTCAA GTTCAACTGG
421 TACGTGGACG GCGTGGAGGT GCATAATGCC AAGACAAAGC CGCGGGAGGA GCAGTACAAC
481 AGCACGTACC GTGTGGTCAG CGTCCTCACC GTCTGCACC AGGACTGGCT GAATGGCAAG
541 GAGTACAAGT GCAAGGTCTC CAACAAAGCC CTCCCAGCCC CCATCGAGAA AACCATCTCC
601 AAAGCCAAAG GGCAGCCCCG AGAACCACAG GTGTACACCC TGCCCCCATC CCGGGATGAG
661 CTGACCAAGA ACCAGGTCAG CCTGACCTGC CTGGTCAAAG GCTTCTATCC CAGCGACATC
721 GCCGTGGAGT GGGAGAGCAA TGGGCAGCCG GAGAACAACCT ACAAGACCAC GCCTCCCGTG
781 TTGGAATCCG ACGGCTCCTT CTTCTCTTAC AGCAAGCTCA CCGTCGACAA GAGCAGGTGG
841 CAGCAGGGGA ACGTCTTCTC ATGCTCCGTG ATGCATGAGG CTCTGCACAA CCACTACACG
901 CAGAAGAGCC TCTCCCTGTC TCCGGGTAAA CGGCGCCGCC GGAGCGGTGG CGGCGGATCA
961 GGTGGGGGTG GATCAGGCGG TGGAGGTTCC GGTGGCGGGG GATCCGGCGG TGGAGGTTCC
1021 GGTGGGGGTG GATCAAGGAA GAGGAGGAAAG AGGCAGGTGA AACTGCTCGA GTCTGGGGGA
1081 GGCGTGGTCC AGCCTGGGAG GTCCCTGAGA CTCTCCTGTG CAGCCTCTGG ATTCACCTTC
1141 AGTAGCTATG CTATGCACTG GGTCCGCCAG GCTCCAGGCA AGGGGCTGGA GTGGGTGGCA
1201 GTTATATCAT ATGATGGAAG CAATAAATAC TACGCAGACT CCGTGAAGGG CCGATTCGCC
1261 ATCTCCAGAG ACAATTCCAA GAACACGCTG TATCTGCAAA TGAACAGCCT GAGAGCTGAG
1321 GACACGGCTG TGTATTACTG TGCAGAGAGC CTGGGGAGCT GGGGGGGTTG GGACCACCTAC
1381 ATGGACGTCT GGGGCAAAGG GACCACGGTC ACCGTCTCCT CAGGTGGCGG CGGATCAGGT
1441 GGGGGTGGAT CAGGTGGCGG TGGCTCCGGT GGCGGGGGAT CAGTGGTGAC TCAGCCACCC
1501 TCAGCGTCTG GGACCCCCGG GCAGAGGGTC ACCATCTCTT GTTCTGGAAG CAGCTCCAAC
1561 ATCGGAAGTA AACTGTAAA CTGGTACCAG CAGCTCCAG GAACGGCCCC CAACTCCTC
1621 ATCTATAGTA ATAATCAGCG GCCCTCAGGG GTCCCTGACC GATTCTCTGG CTCCAAGTCT
1681 GGCACCTCAG CCTCCCTGGC CATCAGTGGG CTCCAGTCTG AGGATGAGGC TGATTATTAC
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1741   TGTGCAGCAT GGGATGACAG CCTGAATGGT TGGGTGTTCG GCGGAGGGAC CAAGCTGACC
1801   GTCCTAGGTC AGCCCGGTGG CGGTGGCTCC GGCGGAGGTG GGTCCGGTGG CGGCGGATCA
1861   GGTGGGGGTG GATCAGGCGG TGGAGGTTCG GGTGGCGGGG GATCAGACAA AACTCACACA
1921   TGCCACCGT GCCCAGCACC GGAAGTACTG GGCGGACCGT CAGTCTTCCT CTTCCCCCCA
1981   AAACCCAAGG ACACCCTCAT GATCTCCCGG ACCCCTGAGG TCACATGCGT GGTGGTGGAC
2041   GTGAGCCACG AAGACCCTGA GGTCAAGTTC AACTGGTACG TGGACGGCGT GGAGGTGCAT
2101   AATGCCAAGA CAAAGCCGCG GGAGGAGCAG TACAACAGCA CGTACCGTGT GGTGAGCGTC
2161   CTCACCGTCC TGCACCAGGA CTGGCTGAAT GGCAAGGAGT ACAAGTGCAA GGTCTCCAAC
2221   AAAGCCCTCC CAGCCCCCAT CGAGAAAACC ATCTCCAAAG CCAAAGGGCA GCCCCGAGAA
2281   CCACAGGTGT ACACCCTGCC CCCATCCCGG GATGAGCTGA CCAAGAACCA GGTGAGCCTG
2341   ACCTGCCTGG TCAAAGGCTT CTATCCCAGC GACATCGCCG TGGAGTGGGA GAGCAATGGG
2401   CAGCCGGAGA ACAACTACAA GACCACGCCT CCCGTGTTGG ACTCCGACGG CTCCTTCTTC
2461   CTCTACAGCA AGCTCACCGT GGACAAGAGC AGGTGGCAGC AGGGGAACGT CTTCTCATGC
2521   TCCGTGATGC ATGAGGCTCT GCACAACCAC TACACGCAGA AGAGCCTCTC CCTGTCTCCG
2581   GGTAAATGA

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Fc-046 amino acid sequence (SEQ ID NO: 835):

[0798] Signal sequence is underlined, linker connecting compound 21 to Fc is in dotted underline, furin cleavage site separating Fc and the linker region is in thick underline, furin cleavage site separating the linker region and PDG13 scFv is in wave underline, and the linker connecting PDG13 and Fc is in dashed underline.

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1     METDTLLLLWV LLLWVPGSTG GPRIRTVGPG SRSASGKLTC LASYCWLFWT GIAGGGGSGG
61    GGSGGGGSGG GGSGGGGSGG GGSDKTHTCP PCPAPELLGG PSVFLFPPKP KDTLMISRTP
121   EVTCVVVDVS HEDPEVKFNW YVDGVEVHNA KTKPREEQYN STYRVVSVLT VLHQDWLNGK
181   EYKCKVSNKA LPAPIEKTIS KAKGQPREPQ VYTLPPSRDE LTKNQVSLT LVKGFIYPSDI
241   AVEWESNGQP ENNYKTTTPV LDSDGSFFLY SKLTVDKSRW QQGNVFSCSV MHEALHNHYT
301   QKSLSLSLSPGK RRRRSGGGGS GGGGSGGGGS GGGGSGGGGS GGGGSRKRRK RQVKLLESGG
361   GVVQPGRSLR LSCAASGFTF SSYAMHWVRQ APGKGLEWVA VISYDGSNKY YADSVKGRFA
421   ISRDNSKNTL YLQMNSLRAE DTAVYYCARA LGSWGGWDHY MDVWGKGTTV TVSSGGGGSG
481   GGGSGGGGSG GGGSVVTQPP SASGTPGQRV TISCSGSSSN IGSNTVNWYQ QLPGTAPKLL
541   IYSNNQRPSG VPDRFSGSKS GTSASLAISG LQSEDEADYY CAAWDDSLNG WVFGGGTKLT
601   VLGQPGGGGS GGGGSGGGGS GGGGSGGGGS GGGGSDKTHT CPPCPAPELL GGPSVFLFPP
661   KPKDTLMISR TPEVTCVVVD VSHEDPEVKF NWYVDGVEVH NAKTKPREEQ YNSTYRVVSV
721   LTVLHQDWLN GKEYKCKVSN KALPAPIEKT ISKAKGQPRE PQVYTLPPSR DELTKNQVSL
781   TCLVKGFYPS DIAVEWESNG QPENNYKTP PVLDSDGSFF LYSKLTVDKS RWQQGNVFSC
841   SVMHEALHNH YTQKSLSLSP GK

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Example 17**Preparation of Fc Conjugates****pSYN-Fc-045 construction**

[0799] Cloning and expression was performed as described in Example 14.

[0800] The 1.8 kb DNA fragment comprising the region from HindIII to EcoRI of pSYN-Fc-045 was synthesized and subcloned into the HindIII/EcoRI sites of pcDNA4 vector (Invitrogen) to generate pSYN-Fc-045 (Fc-compound 21 conjugate).

Fc-045 DNA sequence (SEQ ID NO: 836)

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1      ATGGAGACAG ACACACTCCT GCTATGGGTA CTGCTGCTCT GGGTTCCAGG TTCCACTGGT
61     GACAAAACCTC ACACATGCCC ACCGTGCCCCA GCTCCGGAAC TCCTGGGAGG ACCGTCAGTC
121    TTCTCTTTCC CCCCCAAAACC CAAGGACACC CTCATGATCT CCCGACCCC TGAGGTCACA
181    TGGCTGGTGG TGGACGTGAG CCACGAAGAC CCTGAGGTCA AGTTCAACTG GTACGTGGAC
241    GGGCTGGAGG TGCATAATGC CAAGACAAAG CCGCGGGAGG AGCAGTACAA CAGCACGTAC
301    CGTGTGGTCA GCGTCCTCAC CGTCCTGCAC CAGGACTGGC TGAATGGCAA GGAGTACAAG
361    TGCAAGGTCT CCAACAAAGC CCTCCCAGCC CCCATCGAGA AAACCATCTC CAAAGCCAAA
421    GGGCAGCCCC GAGAACCACA GGTGTACACC CTGCCCCCAT CCCGGGATGA GCTGACCAAG
481    AACCAGGTCA GCCTGACCTG CCTGGTCAAA GGCTTCTATC CCAGCGACAT CGCCGTGGAG
541    TGGGAGAGCA ATGGGCAGCC GGAGAACAAC TACAAGACCA CGCCTCCCGT GTTGGACTCC
601    GACGGCTCCT TCTTCCTCTA CAGCAAGCTC ACCGTCGACA AGAGCAGGTG GCAGCAGGGG
661    AACGTCTTCT CATGCTCCGT GATGCATGAG GCTCTGCACA ACCACTACAC GCAGAAGAGC
721    CTCTCCCTGT CTCCGGGTAA ACGGCGCCGC CGGAGCGGTG GCGGCGGATC AGGTGGGGGT
781    GGATCAGGCG GTGGAGGTTT CGGTGGCGGG GGATCTGGCG GTGGAGGTTT CGGTGGGGGT
841    GGATCCAGGA AGAGGAGGAA GAGGGGCCCC CGGATCCGGA CAGTGGGCCC CGGCAGCCGG
901    AGCGCCAGCG GCAAGCTGAC CTGCCTGGCC AGCTACTGCT GGCTGTTCTG GACCGGCATC
961    GCCGGTGGCG GTGGATCCGG CGGAGGTGGG TCCGGTGGCG GCGGATCAGG TGGGGGTGGA
1021   TCAGGCGGTG GAGGTTCCGG TGGCGGGGGA TCAGACAAAA CTCACACATG CCCACCGTGC
1081   CCAGCACCGG AACTCCTGGG CGGACCGTCA GTCTTCTCTT TCCCCC AAA ACCCAAGGAC
1141   ACCCTCATGA TCTCCCGGAC CCCTGAGGTC ACATGCGTGG TGGTGGACGT GAGCCACGAA
1201   GACCCTGAGG TCAAGTTCAA CTGGTACGTG GACGGCGTGG AGGTGCATAA TGCCAAGACA
1261   AAGCCGCGGG AGGAGCAGTA CAACAGCACG TACCGTGTGG TCAGCGTCCT CACCGTCCTG
1321   CACCAGGACT GGCTGAATGG CAAGGAGTAC AAGTGCAAGG TCTCCAACAA AGCCCTCCCA
1381   GCCCCATCG AGAAAACCAT CTCCAAAGCC AAAGGGCAGC CCCGAGAACC ACAGGTGTAC
1441   ACCCTGCCCC CATCCCGGGA TGAGCTGACC AAGAACCAGG TCAGCCTGAC CTGCCTGGTC
1501   AAAGGCTTCT ATCCCAGCGA CATCGCCGTG GAGTGGGAGA GCAATGGGCA GCCGAGAAC
1561   AACTACAAGA CCACGCCTCC CGTGTTGGAC TCCGACGGCT CCTTCTTCTT CTACAGCAAG
1621   CTCACCGTGG ACAAGAGCAG GTGGCAGCAG GGAACGTCT TCTCATGCTC CGTGATGCAT
1681   GAGGCTCTGC ACAACCACTA CACGCAGAAG AGCCTCTCCC TGCTCCGGG TAAATGA

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Fc-045 amino acid sequence (SEQ ID NO: 837)

[0801] Signal sequence is underlined, furin cleavage site separating Fc and linker is in dotted...underline, furin cleavage site separating linker and compound 21 is in thick underline, linker region connecting compound 21 to the Fc region is in wave underline.

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1   METDTLLLWV LLLWVPGSTG DKTHTCPPCP APELLGGPSV FLFPPKPKDT LMISRTPEVT
61  CVVVVDVSHED PEVKFNWYVD GVEVHNAKTK PREEQYNSTY RVVSVLTVLH QDWLNGKEYK
121 CKVSNKALPA PIEKTISKAK GQPREPQVYT LPPSRDELTK NQVSLTCLVK GFYPSDIAVE
181 WESNGQPENN YKTTTPVLDS DGSFFLYSKL TVDKSRWQQG NVFSCSVMHE ALHNHYTQKS
241 LSLSPGKRRR RSGGGGSGGG GSGGGGSGGG GSGGGGSGGG GSRKRRKRGP RIRTVGPGR
301 SASGKLTCLA SYCWLFWTGI AGGGGSGGGG SGGGGSGGGG SGGGGSGGGG SDKTHTCPPC
361 PAPELLGGPS VFLFPPKPKD TLMISRTPEV TCVVVDVSH EPEVKFNWYV DGVEVHNAKT
421 KPREEQYNST YRVVSVLTVL HQDWLNGKEY KCKVSNKALP APIEKTISKA KGQPREPQVY
481 TLPPSRDELT KNQVSLTCLV KGFYPSDIAV EWESNGQPEN NYKTTTPVLD SDGSFFLYSK
541 LTVDKSRWQQ GNVFSCSVMH EALHNHYTQK SLSLSPGK

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Example 18**Preparation of Conjugates Useful for Subsequent Chemical Ligation To Compounds**pSYN-CysFc-044 construction

[0802] Cloning was performed as described in Example 14.

[0803] The 3.2 kb DNA fragment comprising the region from HindIII to EcoRI of pSYN-CysFc-044 was synthesized and subcloned into the HindIII/EcoRI sites of pcDNA4 vector (Invitrogen) to generate pSYN-CysFc-044 (PDG13-CysFc).

CysFc-044 DNA sequence (SEQ ID NO: 838)

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1   ATGGAGACAG ACACACTCCT GCTATGGGTA CTGCTGCTCT GGGTTCCAGG TTCCACTGGT
61  TGCCCGCCGT GCCCGGCTCC GGAACCTCTG GGAGGACCGT CAGTCTTCCT CTTCCCCCA
121 AAACCCAAGG ACACCCTCAT GATCTCCCGG ACCCCTGAGG TCACATGCGT GGTGGTGGAC
181 GTGAGCCACG AAGACCCTGA GGTCAAGTTC AACTGGTACG TGGACGGCGT GGAGGTGCAT
241 AATGCCAAGA CAAAGCCGCG GGAGGAGCAG TACAACAGCA CGTACCGTGT GGTGAGCGTC
301 CTCACCGTCC TGCACCAGGA CTGGCTGAAT GGCAAGGAGT ACAAGTGCAA GGTCTCCAAC
361 AAAGCCCTCC CAGCCCCCAT CGAGAAAACC ATCTCCAAAG CCAAAGGGCA GCCCCGAGAA
421 CCACAGGTGT ACACCCTGCC CCCATCCCGG GATGAGCTGA CCAAGAACCA GGTGAGCCTG
481 ACCTGCCTGG TCAAAGGCTT CTATCCCAGC GACATCGCCG TGGAGTGGGA GAGCAATGGG

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541   CAGCCGGAGA ACAACTACAA GACCACGCCT CCCGTGTTGG ACTCCGACGG CTCCTTCTTC
601   CTCTACAGCA AGCTCACCGT CGACAAGAGC AGGTGGCAGC AGGGGAACGT CTTCTCATGC
661   TCCGTGATGC ATGAGGCTCT GCACAACCAC TACACGCAGA AGAGCCTCTC CCTGTCTCCG
721   GGTAACCGGC GCCGCCGGAG CGGTGGCGGC GGATCAGGTG GGGGTGGATC AGGCGGTGGA
781   GGTTCGGTG GCGGGGGATC CGGCGGTGGA GGTTCGGTG GGGGTGGATC AAGGAAGAGG
841   AGGAAGAGGC AGGTGAAACT GCTCGAGTCT GGGGGAGGCG TGGTCCAGCC TGGGAGGTCC
901   CTGAGACTCT CCTGTGCAGC CTCTGGATTG ACCTTCAGTA GCTATGCTAT GCACTGGGTC
961   CGCCAGGCTC CAGGCAAGGG GCTGGAGTGG GTGGCAGTTA TATCATATGA TGGAAGCAAT
1021  AAATACTACG CAGACTCCGT GAAGGGCCGA TTCGCCATCT CCAGAGACAA TTCCAAGAAC
1081  ACGCTGTATC TGCAAATGAA CAGCCTGAGA GCTGAGGACA CGGCTGTGTA TTA CTGTGCG
1141  AGAGCGCTGG GGAGCTGGGG GGGTTGGGAC CACTACATGG ACGTCTGGGG CAAAGGGACC
1201  ACGGTCACCG TCTCCTCAGG TGGCGGCGGA TCAGGTGGGG GTGGATCAGG TGGCGGTGGC
1261  TCCGGTGGCG GGGGATCAGT GGTGACTCAG CCACCCTCAG CGTCTGGGAC CCCC GGCGAG
1321  AGGGTCACCA TCTCTTGTTT TGGAAGCAGC TCCAACATCG GAAGTAATAC TGTA AACTGG
1381  TACCAGCAGC TCCCAGGAAC GGCCCCAAA CTCCTCATCT ATAGTAATAA TCAGCGGCCC
1441  TCAGGGGTCC CTGACCGATT CTCTGGCTCC AAGTCTGGCA CCTCAGCCTC CCTG GCCATC
1501  AGTGGGCTCC AGTCTGAGGA TGAGGCTGAT TATTACTGTG CAGCATGGGA TGACAGCCTG
1561  AATGGTTGGG TGTTCCGCGG AGGGACCAAG CTGACCGTCC TAGGTCAGCC CGGTGGCGGT
1621  GGCTCCGGCG GAGGTGGGTC CGGTGGCGGC GGATCAGGTG GGGGTGGATC AGGCGGTGGA
1681  GGTTCGGTG GCGGGGGATC AGACAAA AACT CACACATGCC CACCGTGCCC AGCACCGGAA
1741  CTACTGGGCG GACCGTCAGT CTTCTCTTTC CCCCCAAAAC CCAAGGACAC CCTCATGATC
1801  TCCCGGACCC CTGAGGTCAC ATGCGTGGTG GTGGACGTGA GCCACGAAGA CCCTGAGGTC
1861  AAGTTCAACT GGTACGTGGA CGGCGTGAG GTGCATAATG CCAAGACAAA GCCGCGGGAG
1921  GAGCAGTACA ACAGCACGTA CCGTGTGGTC AGCGTCCTCA CCGTCTGCA CCAGGACTGG
1981  CTGAATGGCA AGGAGTACAA GTGCAAGGTC TCCAACAAAAG CCCTCCCAGC CCCCATCGAG
2041  AAAACCATCT CCAAAGCCAA AGGGCAGCCC CGAGAACCAC AGGTGTACAC CCTG CCCCCA
2101  TCCCGGATG AGCTGACCAA GAACCAGGTC AGCCTGACCT GCCTGGTCAA AGGCTTCTAT
2161  CCCAGCGACA TCGCCGTGGA GTGGGAGAGC AATGGGCAGC CGGAGAACAA CTACAAGACC
2221  ACGCCTCCCG TGTTGGACTC CGACGGCTCC TTCTTCTCT ACAGCAAGCT CACCGTGGAC
2281  AAGAGCAGGT GGCAGCAGGG GAACGTCTTC TCATGCTCCG TGATGCATGA GGCTCTGCAC
2341  AACCACTACA CGCAGAAGAG CCTCTCCCTG TCTCCGGGTA AATGA

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CysFc-044 amino acid sequence (SEQ ID NO: 839):

[0804] Signal sequence is underlined, furin cleavage site separating truncated Fc and linker is in thick underline, furin cleavage site separating linker and PDG13 scFv is dotted underline and linker region connecting PDG-13 scFv to Fc region is in dashed underline.

1 METDTLLLWV LLLWVPGSTG CPPCPAPELL GGPSVFLFPP KPKDTLMISR TPEVTCVVVD

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61    VSHEDPEVKF NQYVDGVEVH NAKTKPREEQ YNSTYRVVSV LTVLHQDWLN GKEYKCKVSN
121   KALPAPIEKT ISKAKGQPRE PQVYTLPPSR DELTKNQVSL TCLVKGFYPS DIAVEWESNG
181   QPENNYKTTP PVLDSGDSFF LYSKLTVDKS RWQQGNVFSC SVMHEALHNH YTQKSLSLSP
241   GKRRRRSGGG GSGGGGSGGG GSGGGGSGGG GSGGGGSRKR_RKRQVKLLES GGGVVQPGRS
301   LRLSCAASGF TFSSYAMHWV RQAPGKGLEW VAVISYDGSN KYYADSVKGR FAISRDNSKN
361   TLYLQMNSLR AEDTAVYYCA RALGSWGGWD HYMDVWGKGT TVTVSSGGGG SGGGSGGGG
421   SGGGGSVVVQ PPSASGTPGQ RVTISCSGSS SNIGSNTVNW YQQLPGTAPK LLIYSNNQRP
481   SGVPDRFSGS KSGTSASLAI SGLQSEDEAD YYCAAWDDSL NGWVFGGGTK LTVLGQPGGG
541   _SGGGGSGGGG_GSGGGGSGGGG_GSGGGGSDKT HTCPCPAPE LLGGPSVFLF PPKPKDTLMI
601   SRTPEVTCVV VDVSHEDPEV KFNWYVDGVE VHNAKTKPRE EQYNSTYRVV SVLTVLHQDW
661   LNGKEYKCKV SNKALPAPIE KTISKAKGQP REPQVYTLPP SRDELTKNQV SLTCLVKGFY
721   PSDIAVEWES NGQPENNYKT TPPVLDSGDS FFLYSKLTVD KSRWQQGNVF SCSVMHEALH
781   NHYTQKSLSL SPGK

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Example 19

Preparation of Compounds With a Cleavable Linker

(a) Synthesis of Compounds Incorporating a Self-Immolative Linker

[0805] Outlined below is the preparation for a conjugate (conjugate A) in which compound 5 is covalently connected to a linker comprising the self-immolative moiety *p*-aniline benzyl carbamate (PABC) and a thrombin substrate moiety. In a similar fashion conjugates incorporating other thrombin substrate moieties were prepared and tested for cleavability in the presence of thrombin.

Fmoc-Cys(Acm)-Gly-Gly-Gly-Gly-Dphe-Pip-Arg(Pbf)

[0806] The fully protected peptide was cleaved from the NovaPEG TGT resin by 30% HFIP/DCM and filtered into a round bottom reaction flask. The solvents were removed *in vacuo*, and the concentrate containing the peptide was precipitated and further triturated with ice cold diethyl ether (Et₂O). This material was directly used without further purification. ESI-MS *m/z* : 1309.51 (MH)⁺.

Fmoc-Cys(Acm)-Gly-Gly-Gly-Gly-Dphe-Pip-Arg(Pbf)-PABOH (p-aniline benzyl alcohol)

[0807] A stirred solution of Fmoc-Cys(Acm)-Gly-Gly-Gly-Gly-Dphe-Pip-Arg(Pbf) (268 mg, 0.2 mmol) and *p*-aniline benzyl alcohol (28 mg, 1.1 equiv) in THF (2 mL) at room temperature was treated with EEDQ (55.6 mg, 1.1 equiv). After 16 h, the mixture was evaporated to dryness, and the residue was triturated with ether. The resulting white solid

product was collected by centrifugation and dried in vacuo (200 mg, 70 %). ESI-MS m/z : 1414.61 (MH)⁺.

Fmoc-Cys(Acm)-Gly-Gly-Gly-Gly-Dphe-Pip-Arg(Pbf)-PABC-PNP

- [0808] A stirred solution of Fmoc-Cys(Acm)-Gly-Gly-Gly-Gly-Dphe-Pip-Arg(Pbf)-PABOH (180 mg, 0.127 mmol) in dry THF (4 mL) and DCM (4mL) at room temperature was treated with PNP chloroformate (38.5 mg, 1.5 equiv) and dry pyridine (15 mg, 1.5 equiv). After 16 h, the mixture was concentrated to 1 mL, and the product was precipitated and triturated with cold ether. The resulting white solid product was collected by centrifugation and dried in vacuo (150 mg, 75 %). ESI-MS m/z : 1579.61 (MH)⁺.

rRAPGK(Alloc)LTCLASYCWLFWTGIA-NH₂ (disulfide)

- [0809] Linear alloc-protected compound 5 was synthesized on NovaPEG Rink Amide resin (0.2 mmol) as described in the general method. The Cys-Cys disulfidic bond was formed by stirring the crude peptide in 50% DMSO/H₂O overnight at 37°C. 35 mg of peptide was obtained after purification by preparative HPLC. ESI-MS m/z : 1298.17 (MH₂)²⁺, 865.78 (MH₃)³⁺.

Fmoc-Cys(Acm)-Gly-Gly-Gly-Gly-Dphe-Pip-Arg(Pbf)-PABC- rRAPGK(Alloc)LTCLA SYCWLFWTGIA-NH₂ (disulfide)

- [0810] Fmoc-Cys(Acm)-Gly-Gly-Gly-Gly-Dphe-Pip-Arg(Pbf)-PABC-PNP (12.5 mg, 0.008 mmol) and rRAPGK(Alloc)LTCLASYCWLFWTGIA (30 mg, 0.011 mmol) in DMF (1 mL) at room temperature were treated with DIPEA (6.5 μL, 5 equiv). The mixture was allowed to stand in the dark overnight. The crude product was precipitated, and triturated with cold ether. The resulting crude product was collect by centrifugation, dried in vacuo, and used for the next step without further purification. ESI-MS m/z : 2018.32 (MH₂)²⁺, 1345.86 (MH₃)³⁺.

Fmoc-Cys(Acm)-Gly-Gly-Gly-Gly-Dphe-Pip-Arg-PABC- rRAPGK(Alloc)LTCLA SYCWLFWTGIA-NH₂ (disulfide)

- [0811] Pbf deprotection of Fmoc-Cys(Acm)-Gly-Gly-Gly-Gly-Dphe-Pip-Arg(Pbf)-PABC-rRAPGK(Alloc)LTCLA SYCWLFWTGIA from the previous step was carried out in 1 mL of solvent mixture (72% TFA, 5% DMF, 5% H₂O, 18% DCM) for 75 min. Since the PABC linker was unstable under this condition, aliquots were taken at various time points to monitor progress of the reaction. At 75 min, cold ether (50 mL) was added

to stop the reaction. The resulting solid was purified by preparative HPLC, to give a white powder (8 mg, 25% for 2 steps). ESI-MS m/z : 1261.83 (MH_3)³⁺.

Fmoc-Cys(Acm)-Gly-Gly-Gly-Gly-Dphe-Pip-Arg-PABC- rRAPGKLTCLA
SYCWLFWTGIA-NH₂ (disulfide)

- [0812] Fmoc-Cys(Acm)-Gly-Gly-Gly-Gly-Dphe-Pip-Arg-PABC-rRAPGK(Alloc)LTCLASYCWLFWTGIA (8 mg, 0.002 mmol) in MeOH/Dioxane (1: 1, 180 μ L) under N₂ at room temperature was treated with Pd(PPh₃)₄ (0.0002 mmol, 0.1 equiv, 20 μ L of a THF solution of Pd(PPh₃)₄ (23 mg/mL), followed by PhSiH₃ (0.01 mmol, 5 equiv). After 20 min, the crude mixture was precipitated and triturated with cold ether. The resulting crude product was used for the next step without purification. ESI-MS m/z : 1233.82 (MH_3)³⁺.

Cys(Acm)-Gly-Gly-Gly-Gly-Dphe-Pip-Arg-PABC- rRAPGKLTCLASYCWLFWTGIA-NH₂
(disulfide)

- [0813] Fmoc deprotection of Fmoc-Cys(Acm)-Gly-Gly-Gly-Gly-Dphe-Pip-Arg-PABC-rRAPGKLTCLA SYCWLFWTGIA from the previous step was carried out in DMSO (200 μ L) with Et₂NH (50 μ L, excess). After 20 min, the reaction was complete and the mixture was purified by preparative HPLC, to give conjugate A a white powder (0.83 mg, 12% over 2 steps). ESI-MS m/z : 1159.80 (MH_3)³⁺, 1159.80 (MH_4)⁴⁺.

- [0814] The above procedures can be used to synthesize conjugates comprising other compounds of the present disclosure.

(b) Thrombin Cleavage of Conjugate A

- [0815] Conjugate A (21 μ L, 0.24 mM) in water was added to 476.5 μ L PBS. The mixture was incubated at 37°C for 30min, followed by 2.5 μ L of thrombin (278 nM, 10 μ g/mL), giving the following approximate initial concentrations: thrombin (1.4 nM, physiological concentration), conjugate A (10 μ M). The mixture was incubated at 37°C. Aliquots (60 μ L) at various time points were quenched with 1 μ L of hirudin (10 μ M) and injected into the HPLC (C-18 column, CH₃CN/H₂O, 0 to 70% over 12 minutes, 60°C 0.5 mL/min, λ = 280 nm). Under these conditions, conjugate A was cleaved rapidly by 1.4 nM thrombin to release the active procoagulant compound 5 (i.e., the conjugate was fully cleaved after about 60 min of incubation).

Example 20

Preparation of Fc-compound Conjugates by Copper-catalyzed Azide-Alkyne Cycloaddition

[0816] A semi-synthetic method to prepare conjugates in which a compound of the present disclosure is linked to Fc is outlined below. This method allows linkage of Fc with a compound containing an unnatural amino acid. Incorporation of an unnatural amino acids may increase biological activity and/or stability. This method is an alternative approach to the native chemical ligation with a N-terminal Cys on Fc and a peptide-thioester directly.

NH₂-PEG₂₇-COSBn

[0817] A stirred solution of Boc-NH-PEG₂₇-COOH (500mg, 0.35 mmol) and benzyl mercaptan (174 mg, 4 equiv) in DMF (2 mL) at room temperature was treated with DIC (53 mg, 1.2 equiv) and DMAP (4.3 mg, 0.1 equiv). After 16 h, the crude product was precipitated, triturated with cold ether, and collected by centrifugation. The Boc group was cleaved by addition of 10 mL of 95 % TFA/TIPS into the resulting white pellet. After 30 min, the mixture was concentrated to 1 mL, and the product was precipitated and triturated with cold ether. The resulting off-white oil product was collected by centrifugation, dried in vacuo, and used for next step without further purification (550 mg). ESI-MS *m/z* : 1428.83 (MH)⁺.

N₃-PEG₂₇-COSBn:

[0818] A stirred solution of NH₂-PEG₂₇-COSBn (300 mg, 0.21 mmol) and 5-Azidopentanoic acid (60 mg, 2 equiv) in DMF (1 mL) at room temperature was treated with PyBOP (164 mg, 1.5 equiv) and DIEA (136 mg, 5 equiv). After 16 h, the crude product was precipitated, triturated with cold ether, and collected by centrifugation. The resulting solid was purified by preparative HPLC, to give a white powder (120 mg, 37 %). ESI-MS *m/z* : 1553.77 (MH)⁺.

Fc-PEG₂₇-N₃ dimer conjugate

[0819] Cys-Fc (2580 μL, 8mg/mL in PBS, pH 7.4) was treated with 2-mercaptoethanesulfonic acid, sodium salt (MESNA) (300 μL, 100 mM in PBS) and N₃ - PEG₂₇-COSBn (120 μL, 50 mM in water) such that the final concentration of Cys-Fc, MESNA, and N₃ -PEG₂₇-COSBn were 6.9 mg/mL, 10 mM, and 2 mM respectively. The reaction mixture was allowed to stand at room temperature for 16 h. The crude reaction

mixture was dialyzed exhaustively against PBS (pH 7.4) (7 changes over 24 hours). SDS-PAGE gel and LC/MS showed greater than 90% conversion. MW observed (reduced): 27707.02.

Fc-PEG₂₇-procoagulant peptide conjugate (copper-catalyzed azide-alkyne cycloaddition

- [0820] Fc-PEG₂₇-N₃ dimer conjugate (20 μ L, 13 mg/mL in PBS pH 7.4) was treated with premixed solution of CuSO₄ and Tris(3-hydroxypropyltriazolylmethyl)amine (THPTA) (10 μ L, 1:1 ratio, 10 mM in water), and Pra-PEG4-PRIRTVGPGSRASGKLTCLASYCWLFWTGIA-NH₂ (30 μ L, 2.15 mM) (Pra = L-propargylglycine). The pH was adjusted to 5.5 by addition of MES buffer (40 μ L, 1M pH 5.5) and water (90 μ L). Reducing agent sodium ascorbate (10 μ L, 100 mM) was added to initiate the reaction. Final concentrations of Fc-PEG₂₇-N₃ dimer conjugate, CuSO₄, THPTA, and SYN4002 were 49 μ M, 500 μ M, 500 μ M, and 322.5 μ M, respectively. After 2 h, LC/MS showed greater than 90% conversion. MW observed (reduced): 31501.96.

Example A

Hemophilia A/B In Vivo Studies

- [0821] The pro-coagulant compounds and conjugates of the present disclosure can be tested using hemophilia A and/or B animals. In one example, the animal is a hemophilia A mouse. In another example, the test animal is a hemophilia A dog (e.g., in-bred colony maintained at the Francis Owen Blood Research Laboratory at the University of North Carolina, Chapel Hill). These dogs have a severe hemophilic phenotype comparable to the severe form of the human disease (Graham, JB, et al., *J. Exp. Med.* 1949;90:97-111; Lozier, JN, et al., *Proc. Natl. Acad. Sci.* 2002;99:12991-12996, each of which is incorporated by reference herein in its entirety).
- [0822] In one example, pro-coagulant compounds and/or conjugates of the present disclosure are injected, e.g. IV or SC, into hemophilia A mice. Blood is collected by vena cava puncture at different time points, e.g. 2, 15, 30, 60, 120, 240 and 480 min (3-5 mice per time point) and citrated immediately. In another example blood is collected at 2 min, 1h, 6h, 12h, 24h, 48h and 96h. Activity is measured by ex vivo Rotem, and remaining blood is centrifuged at 5000 rpm for 2x 10 min to generate plasma for PK analysis.
- [0823] In one example, pro-coagulant compounds and/or conjugates of the present disclosure are injected, e.g. IV or SC, into hemophilia A dogs. Blood samples are drawn

at different time points, e.g. 2, 15, 30, 60, 120, 240 and 480 min. In another example blood is drawn at 2 min, 1h, 6h, 12h, 24h, 48h and 96h. Activity is measured by ex vivo WBCT, and remaining blood is centrifuged at 5000 rpm for 2x 10 min to generate plasma for PK analysis (Dumont, JA, et al. *Blood* 2012, 119, 3024-3030 which is incorporated by reference herein in its entirety).

[0824] In one example, pro-coagulant compounds and/or conjugates of the present disclosure are injected, e.g. IV or SC, into hemophilia A mice and acute efficacy is tested by a tail clip model. Shortly after injection, e.g. 5 min, mice are injured by tail clip and blood loss is measured for 30 min (10-15 mice per dose) and compared to vehicle and FVIII treated mice (Dumont, JA, et al. *Blood* 2012, 119, 3024-3030 which is incorporated by reference herein in its entirety).

[0825] In one example, pro-coagulant compounds and/or conjugates of the present disclosure are injected, e.g. IV or SC, into hemophilia A mice and prophylactic efficacy is tested by a tail vein transection (TVT) model. Following the injection, e.g. after 24 hours, mice are injured by TVT and the survival rate is measured at different time points, e.g. hourly up to 24 hours (10-15 mice per dose) and compared to vehicle and FVIII treated mice. (Dumont, JA, et al. *Blood* 2012, 119, 3024-3030 which is incorporated by reference herein in its entirety).

[0826] Additional bleeding and thrombogenicity models can be used such as described in Tranholm, M, et al, *Blood* 2003, 102, 3615-3620; Tranholm, M, et al, *Thrombosis Research* 2003, 109, 217-223; Lauritzen, B, et al, *Journal of Thrombosis and Hemostasis* 2009, 7, 651-657.

[0827] Having now fully described this invention, it will be understood by those of ordinary skill in the art that the same can be performed within a wide and equivalent range of conditions, formulations and other parameters without affecting the scope of the invention or any embodiment thereof.

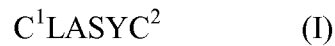
[0828] Other embodiments of the invention will be apparent to those skilled in the art from consideration of the specification and practice of the invention disclosed herein. It is intended that the specification and examples be considered as exemplary only, with a true scope and spirit of the invention being indicated by the following claims.

All patents and publications cited herein are incorporated by reference herein in their entirety.

WHAT IS CLAIMED IS:

1. A compound comprising:

- (a) an amino acid sequence comprising Formula (I):



wherein

L is L-leucine;

A is L-alanine;

S is L-serine;

Y is L-tyrosine;

one or two of L, A, S, and Y are optionally replaced with a replacement amino acid independently selected from D- and L-amino acids;

one additional amino acid is optionally inserted into Formula (I), anywhere between C^1 and C^2 ; and

C^1 and C^2 are independently selected from amino acids having a side chain, wherein the side chains of C^1 and C^2 are linked to form a loop; or

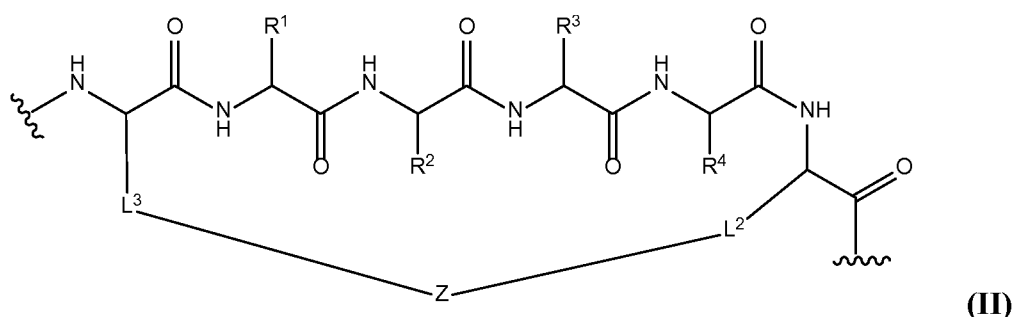
- (b) a retro-, an inverso- or a retro-inverso variant of the amino acid sequence of (a);

or a pharmaceutically acceptable salt or solvate thereof.

2. The compound of claim 1, wherein C^1 and C^2 are independently selected from amino acids having a side chain comprising a -S-H group or a -Se-H group, wherein the side chains of C^1 and C^2 are reversibly linked via a disulfide bond (-S-S-), a diselenide bond (-Se-Se-), or a -S-Se- bond.
3. The compound according to claim 2, wherein C^1 and C^2 are independently selected from cysteine, homo-cysteine (HCy), seleno-cysteine (U), homo-seleno cysteine, and D-amino acids thereof.
4. The compound according to claim 3, wherein
 - (a) C^1 is selected from cysteine, homo-cysteine (HCy), seleno-cysteine (U), homo-seleno cysteine, and D-amino acids thereof; and

- (b) C² is selected from L-cysteine, L-homo-cysteine (HCy), L-seleno-cysteine (U), and L-homo-seleno cysteine.
5. The compound according to claim 1, wherein C¹ and C² are independently selected from amino acids having a side chain comprising a -S-H group, and wherein the side chains of C¹ and C² are reversibly linked via a disulfide bond (-S-S-).
 6. The compound of claim 5 wherein C¹ is selected from L-cysteine and D-cysteine; and C² is L-cysteine.
 7. The compound of claim 1, wherein the side chains of C¹ and C² are covalently linked via an amide bond to form a lactam ring.
 8. The compound of claim 7, wherein one of C¹ and C² is selected from amino acids having a straight or branched (C₁-C₁₀)aminoalkyl side chain, and the other of C¹ and C² is selected from amino acids having a straight or branched (C₁-C₁₀)carboxyalkyl side chain.
 9. The compound of claim 8, wherein one of C¹ and C² is selected from lysine (2,6-diamino-hexanoic acid), 2,5-diamino-pentanoic acid, 2,4-diamino-butyric acid, 2,3-diamino-propionic acid, 2,7-diamino-heptanoic acid, 2,8-diamino-octanoic acid, and the other of C¹ and C² is selected from aspartic acid, glutamic acid (2-amino-pentanedioic acid), 2-amino-hexanedioic acid, 2-amino-heptanedioic acid, and 2-amino-octanedioic acid.
 10. The compound of claim 1, wherein the side chains of C¹ and C² are covalently linked via an optionally substituted triazole moiety.
 11. The compound according to any one of claims 1 to 10, wherein replacement of one or two of L, A, S, and Y with a replacement amino acid, or insertion of an additional amino acid results in a neutral net-charge between C¹ and C².
 12. The compound according to claim 11, wherein each replacement amino acid for L, A, S and Y, when present, is independently selected from amino acids having a hydrophobic or a polar uncharged side chain.
 13. The compound according to any one of claims 1 to 12, wherein each replacement amino acid for L, A, S and Y, when present, is independently selected from L-amino acids.

14. The compound according to any one of claims 1 to 13 wherein each replacement amino acid for L, A, S and Y, when present, is independently selected from G, A, V, I, L, M, F, W, Y, S, T, N and Q.
15. The compound according to any one of claims 1 to 14, wherein the S in Formula (I) is serine.
16. The compound according to claim 15, wherein the S in Formula (I) is L-serine.
17. The compound according to any one of claims 1 to 16, wherein S in Formula (I) is serine, and each replacement amino acid for L, A, and Y, when present, is independently selected from G, A, V, I, L, M, F, W, Y, S, T, N and Q.
18. The compound according to any one of claims 1 to 17, wherein at least one of L, A, S and Y is replaced with a replacement amino acid.
19. The compound according to claim 18, wherein exactly one of L, A, S and Y is replaced with a replacement amino acid.
20. The compound according to any one of claims 1 to 19, comprising a peptide of Formula (II):



wherein

R¹, R², R³ and R⁴ are members independently selected from amino acid side chains;

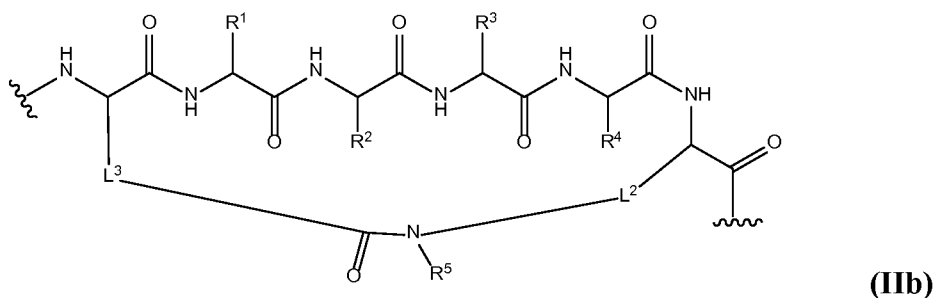
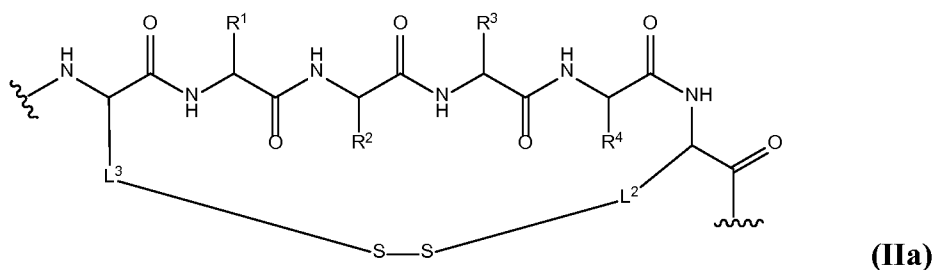
L² and L³ are linker groups independently selected from straight or branched (C₁-C₁₀) alkylene and straight or branched (C₁-C₁₀) heteroalkylene comprising from 1 to 7 heteroatoms selected from O, S and N; and

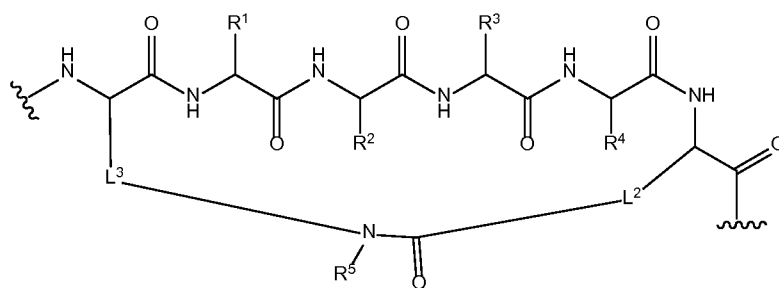
Z is a linking moiety selected from an amino group, an amide group, a disulfide group, a diselenide group, a -S-Se- group, (C₂-C₄)alkylene, (C₂-C₄)alkenyl, (C₂-C₄)alkynyl, a (C₃-

C₈)carbocyclic ring optionally containing from 1 to 4 double bonds, a 3- to 8-membered heterocyclic ring comprising from 1 to 6 heteroatoms selected from O, S and N, a (C₃-C₇)aromatic ring, and a 3- to 8-membered hetero-aromatic ring comprising from 1 to 6 heteroatoms selected from O, S and N,

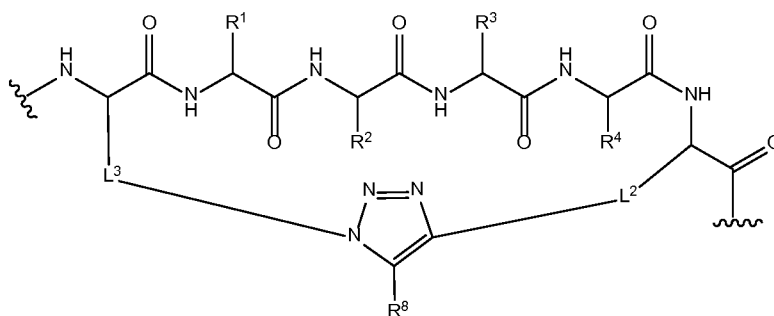
or a retro-, an inverso- or a retro-inverso variant thereof.

21. The compound of claim 20, wherein Z is selected from: -NR⁵-, -NR⁵C(O)-, -S-S-, -S-Se-, -Se-Se-, -CR⁶=CR⁷-, -CR^{6a}R⁶-CR^{7a}R⁷-, and triazolenyl, wherein R⁵, R⁶, R^{6a}, R⁷, and R^{7a} are independently selected from H, (C₁-C₄)alkyl, (C₁-C₄)heteroalkyl comprising from 1 to 3 heteroatoms selected from O, S and N, wherein R⁶ and R⁷ are optionally combined to form a 4- to 7-membered carbocyclic ring optionally comprising from 1 to 3 double bonds, a 3- to 7-membered heterocyclic ring comprising from 1 to 5 heteroatoms selected from O, S and N, a (C₅-C₇)aromatic ring, or a 5- to 7-membered heteroaromatic ring comprising from 1 to 5 heteroatoms selected from O, S and N.
22. The compound of claim 21, wherein Z is selected from -NR⁵-, -NR⁵C(O)-, -S-S-, 1,4-triazolenyl, and 1,5-triazolenyl.
23. The compound according to claim 22, wherein the peptide of Formula (II) has a structure according to Formula (IIa), Formula (IIb), Formula (IIc), Formula (IId), Formula (IIe), Formula (IIf), or Formula (IIg):

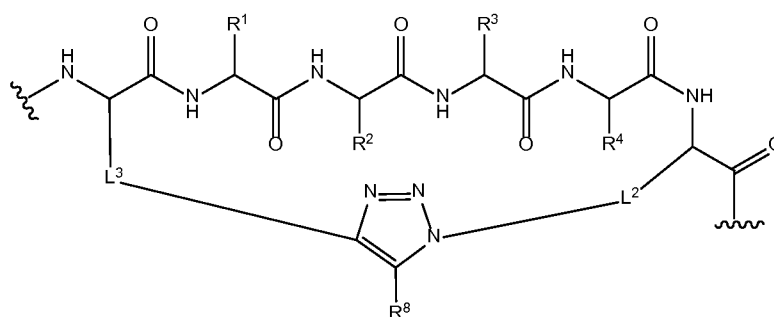




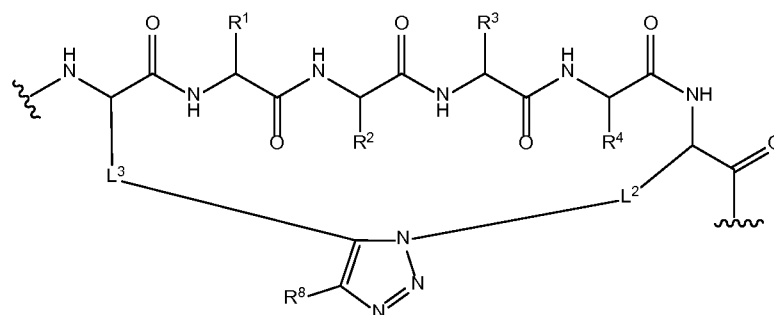
(IIc)



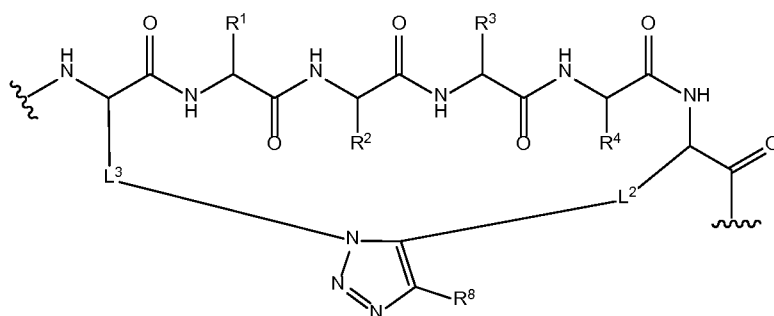
(IIId)



(IIe)



(IIIf)



(IIg)

wherein R^5 and R^8 are independently selected from H and (C_1-C_4) alkyl;

or a retro-, an inverso- or a retro-inverso variant thereof.

24. The compound according to any one of claims 20 to 23, wherein R^1 , R^2 , R^3 and R^4 are members independently selected from H, straight or branched (C_1 - C_6)alkyl, straight or branched (C_1 - C_6)heteroalkyl comprising from 1 to 5 heteroatoms selected from O, S and N, and (C_1 - C_6)aralkyl.
25. The compound according to any one of claims 20 to 24, wherein L^1 and L^2 are independently selected from straight or branched (C_1 - C_6)alkylene.
26. The compound according to claim 1, wherein the amino acid sequence comprises:

CGASYC (SEQ ID NO: 760), CLGSYC (SEQ ID NO: 761), CLASGC (SEQ ID NO: 762), cGASYC (SEQ ID NO: 763), cLGSYC (SEQ ID NO: 764), cLASGC (SEQ ID NO: 765), CAASYC (SEQ ID NO: 766), CLASAC (SEQ ID NO: 767), cAASYC (SEQ ID NO: 768), cLASAC (SEQ ID NO: 769), CVASYC (SEQ ID NO: 770), CLVSYC (SEQ ID NO: 771), CLASVC (SEQ ID NO: 772), cVASYC (SEQ ID NO: 773), cLVSYC (SEQ ID NO: 774), cLASVC (SEQ ID NO: 775), CIASYC (SEQ ID NO: 776), CLISYC (SEQ ID NO: 777), CLASIC (SEQ ID NO: 778), cIASYC (SEQ ID NO: 779), cLISYC (SEQ ID NO: 780), cLASIC (SEQ ID NO: 781), CLASYC (SEQ ID NO: 782), CLLSYC (SEQ ID NO: 783), CLASLC (SEQ ID NO: 784), cLASYC (SEQ ID NO: 785), cLLSYC (SEQ ID NO: 786), cLASLC (SEQ ID NO: 787), FASYC (SEQ ID NO: 788), CLFSYC (SEQ ID NO: 789), CLASFC (SEQ ID NO: 790), cFASYC (SEQ ID NO: 791), cLFSYC (SEQ ID NO: 792), cLASFC (SEQ ID NO: 793), CWASYC (SEQ ID NO: 794), CLWSYC (SEQ ID NO: 795), CLASWC (SEQ ID NO: 796), cWASYC (SEQ ID NO: 797), cLWSYC (SEQ ID NO: 798), cLASWC (SEQ ID NO: 799), CYASYC (SEQ ID NO: 800), CLYSYC (SEQ ID NO: 801), cYASYC (SEQ ID NO: 802), cLYSYC (SEQ ID NO: 803), CQASYC (SEQ ID NO: 804), CLQSYC (SEQ ID NO: 805), CLASQC (SEQ ID NO: 806), cQASYC (SEQ ID NO: 807), cLQSYC (SEQ ID NO: 808), and cLASQC (SEQ ID NO: 809), CLASSC (SEQ ID NO: 810), CLAsYC (SEQ ID NO: 811), or CLASyC (SEQ ID NO: 812),

wherein

each C (L-cysteine) is optionally and independently replaced with L-homo-cysteine (HCy), L-seleno-cysteine (U), or L-homo-seleno cysteine, and

each c (D-cysteine) is optionally and independently replaced with D-homo-cysteine, D-seleno-cysteine (u), or D-homo-seleno cysteine, or a retro-, an inverso- or a retro-inverso variant thereof.

27. The compound according to claim 1, wherein the amino acid sequence comprises:

KGASYE (SEQ ID NO: 560), KLGSYE (SEQ ID NO: 561), KLASGE (SEQ ID NO: 562), kGASYE (SEQ ID NO: 563), kLGSYE (SEQ ID NO: 564), kLASGE (SEQ ID NO: 565), KAASYE (SEQ ID NO: 566), KLASAE (SEQ ID NO: 567), kAASYE (SEQ ID NO: 568), kLASAE (SEQ ID NO: 569), KVASYE (SEQ ID NO: 570), KLVSYE (SEQ ID NO: 571), KLASVE (SEQ ID NO: 572), kVASYE (SEQ ID NO: 573), kLVSYE (SEQ ID NO: 574), KLASVE (SEQ ID NO: 575), KIASYE (SEQ ID NO: 576), KLISYE (SEQ ID NO: 577), KLASIE (SEQ ID NO: 578), kIASYE (SEQ ID NO: 579), kLISYE (SEQ ID NO: 580), KLASIE (SEQ ID NO: 581), KLASYE (SEQ ID NO: 582), KLLSYE (SEQ ID NO: 583), KLASLE (SEQ ID NO: 584), kLASYE (SEQ ID NO: 585), kLLSYE (SEQ ID NO: 586), KLASLE (SEQ ID NO: 587), KFASYE (SEQ ID NO: 588), KLFSYE (SEQ ID NO: 589), KLASFE (SEQ ID NO: 590), kFASYE (SEQ ID NO: 591), kLFSYE (SEQ ID NO: 592), kLASFE (SEQ ID NO: 593), KWASYE (SEQ ID NO: 594), KLWSYE (SEQ ID NO: 595), KLASWE (SEQ ID NO: 596), kWASYE (SEQ ID NO: 597), kLWSYE (SEQ ID NO: 598), kLASWE (SEQ ID NO: 599), KYASYE (SEQ ID NO: 600), KLYSYE (SEQ ID NO: 601), kYASYE (SEQ ID NO: 602), kLYSYE (SEQ ID NO: 603), KQASYE (SEQ ID NO: 604), KLQSYE (SEQ ID NO: 605), KLASQE (SEQ ID NO: 606), kQASYE (SEQ ID NO: 607), kLQSYE (SEQ ID NO: 608), kLASQE (SEQ ID NO: 609), EGASYK (SEQ ID NO: 610), ELGSYK (SEQ ID NO: 611), ELASGK (SEQ ID NO: 612), eGASYK (SEQ ID NO: 613), eLGSYK (SEQ ID NO: 614), eLASGK (SEQ ID NO: 615), EAASYK (SEQ ID NO: 616), ELASAK (SEQ ID NO: 617), eAASYK (SEQ ID NO: 618), eLASAK (SEQ ID NO: 619), EVASYK (SEQ ID NO: 620), ELVSYK (SEQ ID NO: 621), ELASVK (SEQ ID NO: 622), eVASYK (SEQ ID NO: 623), eLVSYK (SEQ ID NO: 624), eLASVK (SEQ ID NO: 625), EIASYK (SEQ ID NO: 626), ELISYK (SEQ ID NO: 627), ELASIK (SEQ ID NO: 628), eIASYK (SEQ ID NO: 629), eLISYK (SEQ ID NO: 630), eLASIK (SEQ ID NO: 631), ELASYK (SEQ ID NO: 632), ELLSYK (SEQ ID NO: 633), ELASLK (SEQ ID NO: 634), eLASYK (SEQ ID NO: 635), eLLSYK (SEQ ID NO: 636), eLASLK (SEQ ID NO: 637),

EFASYK (SEQ ID NO: 638), ELFSYK (SEQ ID NO: 639), ELASFK (SEQ ID NO: 640), eFASYK (SEQ ID NO: 641), eLFSYK (SEQ ID NO: 642), eLASFK (SEQ ID NO: 643), EWASYK (SEQ ID NO: 644), ELWSYK (SEQ ID NO: 645), ELASWK (SEQ ID NO: 646), eWASYK (SEQ ID NO: 647), eLWSYK (SEQ ID NO: 648), eLASWK (SEQ ID NO: 649), EYASYK (SEQ ID NO: 650), ELYSYK (SEQ ID NO: 651), eYASYK (SEQ ID NO: 652), eLYSYK (SEQ ID NO: 653), EQASYK (SEQ ID NO: 654), ELQSYK (SEQ ID NO: 655), ELASQK (SEQ ID NO: 656), eQASYK (SEQ ID NO: 657), eLQSYK (SEQ ID NO: 658), or eLASQK (SEQ ID NO: 659),

wherein

each K (L-lysine) is optionally replaced with a replacement L-amino acid having a side chain comprising an amino group,

each k (D-lysine) is optionally replaced with a replacement D-amino acid having a side chain comprising an amino group,

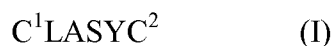
each E (L-glutamic acid) is optionally and independently replaced with L-aspartic acid (D) or another replacement L-amino acid having a side chain comprising a carboxylic acid group,

each e (D-glutamic acid) is optionally and independently replaced with D-aspartic acid (d) or another replacement D-amino acid having a side chain comprising a carboxylic acid group,

wherein the amino acid side chains of K or k (or a replacement amino acid of K or k) and E or e (or a replacement amino acid of E or e) are covalently linked via a peptide bond formed between the amino group and the carboxylic acid group, or a retro-, an inverso- or a retro-inverso variant thereof.

28. The compound according to claim 1, comprising:

(a) (a) an amino acid sequence comprising Formula (I):



wherein

S is L-serine;

L is L-leucine;

A is L-alanine;

Y is L-tyrosine,

wherein one of L, A and Y are optionally and individually replaced with a replacement L-amino acid; and

C¹ and C² are independently selected from amino acids having a side chain comprising a -S-H group, and wherein the side chains of C¹ and C² are linked via a disulfide bond, or (b) a retro-, an inverso- or a retro-inverso variant of the amino acid sequence of (a).

29. The compound of claim 28, wherein:

S is L-serine;

L is L-leucine;

A is L-alanine;

Y is L-tyrosine;

C¹ is selected from L-cysteine, D-cysteine, L-homo-cysteine, and D-homo-cysteine; and

C² is selected from L-cysteine and L-homo-cysteine.

30. The compound according to claim 1, comprising:

(a) (a) an amino acid sequence comprising Formula (III):



wherein

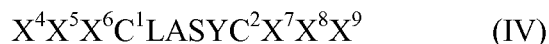
S is L-serine, and

X¹, X² and X³ are independently selected from L-amino acids having a hydrophobic or a polar, uncharged side chain; or

(b) a retro-, an inverso- or a retro-inverso variant of the amino acid sequence of (a).

31. The compound according to claim 30, wherein X¹ is selected from A, L, M, V, and Q.
32. The compound according to claim 30 or 31, wherein X² is selected from G, A, alpha-aminobutyric acid (Abu), V, L, I and S.
33. The compound according to any one of claims 30 to 32, wherein X³ is selected from V, Q, F, Y, and derivatives thereof.
34. The compound according to any one of claims 1 to 33, comprising

(a) an amino acid sequence comprising Formula (IV):



wherein

X⁴ is either absent or present and, when present, is N, Q, an amino acid having a side chain comprising a basic moiety, or an alpha-N-alkylated amino acid thereof;

X⁵ is either absent or present and, when present, is an amino acid having a hydrophobic or a polar, uncharged side chain, or an alpha-N-alkylated amino acid thereof;

X⁶ is an amino acid;

X⁷ and X⁹ are independently either absent or present and when present are independently selected from amino acids;

X⁸ is either absent or present and when present is selected from L-amino acids; or

(b) a retro-, an inverso- or a retro-inverso variant of the amino acid sequence of (a), or a pharmaceutically acceptable salt or solvate thereof.

35. The compound according to any one of claims 1 to 34, wherein the amino acid sequence comprises at least 9 amino acids and not more than 500 amino acids.
36. The compound according to claim 35, wherein the amino acid sequence comprises at least 12 amino acids.
37. The compound of claim 36 wherein the amino acid sequence comprises at least 16 amino acids.
38. The compound of claim 37 wherein the amino acid sequence comprises at least 20 amino acids.
39. The compound of any one of claims 34 to 38, wherein X⁷, X⁸ and X⁹ are independently either absent or present and, when present, are independently selected from amino acids having a hydrophobic side chain and amino acids having a polar uncharged side chain.
40. The compound according to claim 39, wherein X⁷, X⁸ and X⁹ are independently either absent or present and, when present, are independently selected from L, norleucine (Nle), I, V, M, F, W, Y, S, T, N, and Q.
41. The compound according to any one of claims 34 to 40 wherein X⁴ is selected from lysine (K), arginine (R), histidine (H), 2,4-diaminobutyric acid (Dab), 2,3-diaminopropionic

acid (Dpr), ornithine (Orn), 2,7-diamino-heptanoic acid, 2,8-diamino-octanoic acid and alpha-N-alkylated amino acids thereof.

42. The compound according to claim 41, wherein X^4 is lysine.
43. The compound according to any one of claims 34 to 42, wherein X^5 is selected from L, V, M, P, and alpha-N-methylated amino acids thereof.
44. The compound according to claim 43, wherein X^5 is L or alpha-N-alkylated L.
45. The compound according to any one of claims 34 to 44, wherein X^6 is selected from A, T, S, Q, R, and alpha-N-methylated amino acids thereof.
46. The compound according to any one of claims 34 to 45, wherein X^7 is selected from G, L, Q and amino acids having a side chain comprising an aromatic moiety.
47. The compound according to claim 46, wherein X^7 is selected from G, L, Q, W, F, Y, and 1-aryl-alanine.
48. The compound according to claim 47, wherein X^7 is W.
49. The compound according to any one of claims 34 to 48, wherein X^8 is selected from L, I, nor-leucine (Nle), V, Y, Q, and M.
50. The compound according to any one of claims 34 to 49, wherein X^9 is selected from F, V, and L.
51. The compound according to any one of claims 1 to 50, wherein the amino acid sequence is a retro-inverso variant of the amino acid sequence of (a).
52. The compound according to any one of claims 1 to 50, wherein the compound comprises not more than 300 amino acids.
53. The compound according to claim 52 comprising not more than 100 amino acids.
54. The compound according to any one of the preceding claims, wherein the amino acid sequence of the compound comprises at least one of:

KLTCCLASYCWLF

k-MeLeu-TCLASYCWLF

RRAPGKLQCLASYCWLFWTGIA

RRAPGKLTCLASYCWLFWTGIA

rRAPGKLTCLASYCWLFWTGIA

rRAPGKSTCLASYCWLFWTGIA

PRIRTVGPGSRSASGKLTCLASYCWLFWTGIA

PRIrTVGPGSrSASGKLTCLASYCWLFWTGIA

PRIRTVGPGSRSASGKSTCLASYCWLFWTGIA

SRIRTVGPGSRSASGKSTCLASYCWLFWTGIA

PRIRTVSPGSRASGKSTCLASYCWLFWTGIA

SRIRTVSPGSRASGKSTCLASYCWLFWTGIA

PRSRTVGPGSRSASGKSTCLASYCWLFWTGIA

SRSRTVSPGSRASGKSTCLASYCWLFWTGIA

PRIrTVGPGSrSASGKSTCLASYCWLFWTGIA

SKQGRPISPDRRAAGKLTCLASYCWLFWTGIA

SKQGRPISPD_rRAAGKLTCLASYCWLFWTGIA

RRAPGKLTCLASYCWLF_{FGSGISLSRAPESAAP}

RRFVGGSLSQRRAPGKLTCLASYCWLFWTGIA

PQTRDPSSRDRRAPGKLTCLASYCWLFWTGIA.

55. The compound according to any one of claims 1 to 54, wherein the compound is covalently linked to a half-life extending moiety.
56. The compound according to claim 55, wherein the half-life extending moiety is selected from Fc, FcRn binding ligand, albumin, albumin-binding ligand, transferrin, a PEG moiety, a PPG moiety, a PAS moiety, and a HES moiety.
57. The compound according to any one of claims 1 to 56, wherein the compound is a peptide or a peptide derivative.
58. The compound according to any of claims 1 to 57, wherein the peptide sequence has a free N-terminus (-NH₂).

59. The compound according to any one of claims 1 to 58, wherein the peptide sequence is amidated at the C-terminus.
60. The compound according to claim 1 wherein the compound is
- (a) a peptide or peptide derivative selected from those listed in Table 1, or
 - (b) a retro-, an inverso- or a retro-inverso variant of the peptide or peptide derivative, or a salt or solvate thereof.
61. A compound comprising an amino acid sequence comprising C¹ and C², wherein C¹ and C² are independently selected amino acids having a side chain, wherein the side chains of C¹ and C² are linked, and wherein C¹ and C² are separated by 4 amino acids, wherein the compound comprises at least 9 and not more than 100 amino acids.
62. The compound of claim 61 comprising:
- (a) an amino acid sequence comprising Formula (I):
$$C^1LASYC^2 \quad (I)$$
wherein
 - L is L-leucine;
 - A is L-alanine;
 - S is L-serine;
 - Y is L-tyrosine;one of L, A, S, and Y is optionally replaced with an independently selected replacement amino acid; and
 - C¹ and C² are independently selected from amino acids having a side chain, wherein the side chains of C¹ and C² are covalently linked; or
 - (b) a retro-, an inverso- or a retro-inverso variant of the amino acid sequence of (a).
63. The compound according to any one of the preceding claims, wherein the compound is covalently linked to a half-life extension moiety.

64. The compound according to any one of claims 1 to 63 wherein the compound has an EC_{50} of about 5 μ M or less in a Factor Xa (FXa) generation assay measuring conversion of FX to FXa in the presence of FIXa.
65. The compound according to claim 64, wherein the EC_{50} is about 1 μ M or less.
66. The compound according to claim 65, wherein the EC_{50} is about 200 nM or less.
67. The compound according to any one of claims 1 to 66, wherein the compound, at a concentration of 5 μ M or less, increases the catalytic activity (k_{cat}) of Factor IXa (FIXa) for conversion of FX to FXa in a FXa generation assay when compared to a reference catalytic activity of the FIXa measured in the absence of the compound.
68. The compound according to claim 67, wherein the compound increases the catalytic activity (k_{cat}) of FIXa by at least 50 fold.
69. The compound according to claim 68, wherein the compound increases the catalytic activity (k_{cat}) of FIXa by at least 100 fold.
70. The compound according to any one of claims 1 to 69, wherein the compound, at a concentration of 5 μ M or less, increases the catalytic activity (k_{cat}) of FVIIa for conversion of FX to FXa in a FXa generation assay when compared to a reference catalytic activity of the FVIIa measured in the absence of the compound.
71. The compound according to any one of claims 1 to 70, wherein the compound, at a concentration of 5 μ M or less, reduces clotting time in at least one coagulation assay selected from: an activated partial thromboplastin time (aPTT) assay, a modified activated partial thromboplastin time (aPTT*) assay, and a rotational thromboelastometry (ROTEM) assay when compared to a reference clotting time measured in the absence of the compound.
72. The compound according to any one of claims 1 to 71, wherein the compound, at a concentration of 10 μ M or less, reduces clotting time in a rotational thromboelastometry (ROTEM) assay using FVIII-deficient or FIX-deficient human plasma.

73. The compound according to claim 72 wherein the compound reduces the clotting time in a dose-dependent manner.
74. The compound according to any one of claims 1 to 73, wherein the compound, at a concentration of 5 μ M or less, has additive activity with Factor VIII (FVIII) in at least one assay selected from a thrombin generation assay and a ROTEM assay.
75. The compound according to claim 74, wherein FVIII is present at a concentration of from about 0.05 U/mL to about 0.2U/mL in the thrombin generation assay, and from about 0.05 U/mL to about 0.1U/mL in the ROTEM assay.
76. The compound according to any one of claims 1 to 75, wherein the compound has pro-coagulant activity.
77. The compound according to claim 76, wherein the pro-coagulant activity is measured using a method selected from an activated partial thromboplastin time (aPTT) assay, a thrombin generation assay (TGA), and a ROTEM assay.
78. The compound according to any of the preceding claims, wherein the compound is capable of interacting with a peptide comprising the amino acid sequence:

MFCAG (SEQ ID NO: 1).

79. The compound according to claim 78, wherein the compound is capable of interacting with a peptide comprising the amino acid sequence:

YNNMFCAGFHE (SEQ ID NO: 2).

80. The compound according to claim 78, wherein the compound is capable of interacting with a peptide comprising the amino acid sequence:

RSTKFTIYNNMFCAGFHEGGRDSCQG (SEQ ID NO: 3),

or an amino acid sequence that has at least 23/26 homology with SEQ ID NO: 3.

81. The compound according to any of the preceding claims, wherein the compound is capable of interacting with a FIXa protein at a region corresponding to amino acid sequence:

YNNMFCAGFHE (SEQ ID NO: 2).

82. The compound according to claim 81, wherein the compound is capable of interacting with a FIXa protein at a region corresponding to amino acid sequence:

RSTKFTIYNNMFCAGFHEGGRDSCQG (SEQ ID NO: 3),

or an amino acid sequence that has at least 23/26 homology with SEQ ID NO: 3.

83. A pharmaceutical composition comprising a compound according to any one of claims 1 to 82, and a pharmaceutically acceptable carrier.
84. A method of increasing the catalytic activity (k_{cat}) of a blood coagulation factor, comprising contacting the blood coagulation factor with a compound according to any one of claims 1 to 82.
85. The method of claim 84, wherein the compound interacts with the blood coagulation factor at a region corresponding to amino acid sequence: MFCAG (SEQ ID NO: 1).
86. The method according to claim 84 or 85, wherein the blood coagulation factor is selected from FIXa and FVIIa.
87. The method according to claim 86, wherein the blood coagulation factor is FIXa.
88. The method according to claim 87, wherein the compound interacts with the FIXa at a region corresponding to amino acid sequence: YNNMFCAGFHE (SEQ ID NO: 2).
89. The method according to claim 87, wherein the FIXa is human FIXa or canine FIXa.
90. The method according to any one of claims 84 to 89, wherein the method is an *in vitro* method measuring conversion of FX to FXa (FXa generation assay).
91. The method according to claim 86, wherein the blood coagulation factor is FVIIa.
92. A method for treating bleeding diathesis in a mammalian subject, the method comprising administering to the subject a therapeutically effective amount of a compound according to any one of claims 1 to 82 or the pharmaceutical composition of claim 83.

93. The method according to claim 92, wherein the bleeding diathesis is caused by a blood coagulation disorder.
94. The method according to claim 93, wherein the blood coagulation disorder is selected from hemophilia and von Willebrand disease (vWD).
95. The method according to claim 94, wherein the blood coagulation disorder is hemophilia A.
96. The method of any one of claims 92 to 95, wherein the bleeding is selected from hemarthrosis, muscle bleed, oral bleed, hemorrhage, hemorrhage into muscles, oral hemorrhage, trauma, trauma capitis, gastrointestinal bleeding, intracranial hemorrhage, intra-abdominal hemorrhage, intrathoracic hemorrhage, bone fracture, central nervous system bleeding, bleeding in the retropharyngeal space, bleeding in the retroperitoneal space, and bleeding in the iliopsoas sheath.
97. The method of claim 96, wherein the subject is in need of surgical prophylaxis, peri-operative management, or treatment for surgery.
98. The method of claim 97, wherein the surgery is minor surgery, major surgery, tooth extraction, tonsillectomy, inguinal herniotomy, synovectomy, total knee replacement, craniotomy, osteosynthesis, trauma surgery, intracranial surgery, intra-abdominal surgery, intrathoracic surgery, or joint replacement surgery.
99. A method of treating a mammalian subject having a deficiency in at least one blood coagulation factor selected from FV, FVII, FVIIa, FVIII, FIX, FX, FXI, and vWF the method comprising administering to the subject a therapeutically effective amount of the compound according to any one of claims 1 to 82 or a pharmaceutical composition according to claim 83.
100. The method according to claim 99, wherein the subject has a deficiency in FVIII, responds to FVIII treatment, or produces antibodies against FVIII.
101. The method of any one of claims 92 to 100, wherein the mammalian subject is a human subject.

102. The method of any one of claims 92 to 101, wherein the mammalian subject is concomitantly treated with at least one of FVIII, FIX and desmopressin (DDVAP).
103. The compound according to any one of claims 1 to 82 or the pharmaceutical formulation of claim 83 for treating a mammalian subject having a blood coagulation disorder.
104. The compound according to claim 103, wherein the mammalian subject is a human subject.
105. Use of the compound according to any one of claims 1 to 82 or the pharmaceutical formulation of claim 83 for the manufacture of a medicament for the treatment of a blood coagulation disorder.
106. A method for making the compound according to any one of claims 1 to 82, the method comprising forming a peptide having the amino acid sequence, or a retro-, inverso- or retro-inverso variant thereof using solid-phase peptide synthesis.
107. The method of claim 106 further comprising covalently linking the peptide to a heterologous moiety.
108. The method of claim 107, wherein the heterologous moiety is a half-life extending moiety.
109. The method according to 108, wherein the half-life extending moiety is selected from Fc, FcRn binding ligand, albumin, albumin-binding ligand, transferrin, a PEG moiety, a PPG moiety, a PAS moiety, and a HES moiety.
110. The compound according to any one of claims 1 to 82, wherein the compound inhibits heparin catalyzed (heparin accelerated) FIXa-AT complex formation when compared to FIXa-AT complex formation in the absence of the compound.
111. The compound of claim 110, wherein the FIXa-AT complex formation is measured in the presence of 100 nM heparin and the compound is present at a concentration of from about 1 μ M to about 10 μ M.

112. The compound according to any one of claims 1 to 82, wherein the compound is linked to a heterologous moiety (Het), optionally via a linker.
113. A conjugate comprising the compound of any one of claims 1 to 82 and a first heterologous moiety which are linked to each other via a first optional linker.
114. The conjugate of claim 113, which comprises a structure according to Formula (A1 or (A2):



wherein

Het1 is the first heterologous moiety;

m is an integer selected from 0 and 1;

L₁ is either absent (m=0) or present (m=1), and when present is a linker;

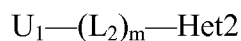
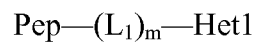
Pep is a compound according to claim 1; and

(—) is a covalent bond.

115. The conjugate of claim 113 or 114, wherein the first heterologous moiety is a half-life extending molecule.
116. The conjugate of claim 115, wherein the first heterologous moiety is an immunoglobulin constant region or a portion thereof, albumin, transferrin, an albumin binding moiety, a PAS sequence, a HES sequence, the β subunit of the C-terminal peptide (CTP) of human chorionic gonadotropin, polyethylene glycol (PEG), hydroxyethyl starch (HES), albumin-binding small molecules, or any combination thereof.
117. The conjugate of claim 116, wherein the immunoglobulin constant region or a portion thereof is an Fc moiety or an FcRn binding partner.
118. The conjugate of claim 113 to 117, further comprising a second heterologous moiety (Het2), wherein the second heterologous moiety is linked to or associated with the first heterologous moiety (Het1).
119. The conjugate of claim 118, wherein the second heterologous moiety is a half-life extending moiety.

120. The conjugate of claim 119, wherein the second heterologous moiety is an immunoglobulin constant region or a portion thereof, albumin, transferrin, an albumin binding moiety, a PAS sequence, a HES sequence, the β subunit of the C-terminal peptide (CTP) of human chorionic gonadotropin, polyethylene glycol (PEG), hydroxyethyl starch (HES), albumin-binding small molecules, or any combinations thereof.
121. The conjugate of claim 120, wherein the immunoglobulin constant region or a portion thereof is an Fc moiety, a single chain Fc, or an FcRn binding partner.
122. The conjugate of any one of claims 118 to 121, wherein the first heterologous moiety and the second heterologous moiety are associated by a disulfide bond.
123. The conjugate of claim 118 to 122, further comprising a scFc linker, which is linked to the second heterologous moiety and the compound.
124. The conjugate of claim 123, wherein the scFc linker comprises a linker (Lx) and a first intracellular processing site (P1) interposed between the second heterologous moiety and the linker (Lx).
125. The conjugate of claim 124, wherein the scFc linker further comprises a second intracellular processing site (P2) interposed between the linker (Lx) and the compound.
126. The conjugate of claim 114 to 125, further comprising a polypeptide comprising FVIII, FIX, FVIIa, or a platelet targeting moiety, wherein the polypeptide is linked to the compound or to the second heterologous moiety via a second optional linker.
127. The conjugate of claim 126, wherein the polypeptide is linked to the second heterologous moiety via the second optional linker.
128. The conjugate of claim 127, wherein the compound, the first heterologous moiety, and the first optional linker are in a first polypeptide chain, and the polypeptide, the second heterologous moiety, and the second optional linker are in a second polypeptide chain, wherein the first polypeptide chain and the second polypeptide chain associate chemically or physically with each other.

129. The conjugate of claim 128, wherein the first heterologous moiety (Het1) of the first polypeptide chain and the second heterologous moiety (Het2) of the second polypeptide chain associate chemically or physically with each other.
130. The conjugate of claim 129, wherein each of the first heterologous moiety and the second heterologous moiety is a Fc single chain.
131. The conjugate of claim 127 to 130, wherein the conjugate comprises:



wherein

Het1 is the first heterologous moiety;

L₁ is either absent (m=0) or present (m=1), and when present is a linker;

m is an integer selected from 0 and 1;

Pep is the compound of any one of claims 1 to 82,

Het2 is the second heterologous moiety;

L₂ is either absent (m=0) or present (m=1), and when present is a linker;

U₁ is the polypeptide comprising FIX, FVIII, FVIIa, or the platelet targeting moiety;

(—) is a covalent bond,

wherein Het1 and Het2 associate chemically or physically with each other.

132. The conjugate of claim 126 to 131, further comprising a scFc linker.
133. The conjugate of claim 132, wherein the scFc linker is linked to the second heterologous moiety and the compound.
134. The conjugate of claim 132 or 133, wherein the scFc linker comprises a linker (Lx) and a first intracellular processing site (P1) interposed between the second heterologous moiety and the linker (Lx).
135. The conjugate of claim 134, wherein the scFc linker further comprises a second intracellular processing site (P2) interposed between the linker (Lx) and the compound.

136. The conjugate of claim 124, 125, 134, or 135, wherein the scFc linker comprises two intracellular processing sites which are recognized by the same or by different intracellular processing enzymes.
137. The conjugate according to claim 136, wherein the intracellular processing site is recognized by a intracellular processing enzyme selected from the group consisting of a yeast Kex2, PCSK1, PCSK2, PCSK3, PCSK4, PCSK5, PCSK6, or PCSK7.
138. The conjugate according to claim 136 or 137, wherein the at least one intracellular processing site is processed by PCSK5.
139. The conjugate according to claim 136 or 137, wherein each of the two intracellular processing sites is processed by PCSK5.
140. The conjugate according to claim 139, wherein the two intracellular processing sites are the same.
141. The conjugate according to claim 139, wherein the two intracellular processing sites are different.
142. The conjugate according to any one of claims 124, 125, and 134 to 141, wherein the intracellular processing site processed by the intracellular processing enzyme comprises the amino sequence R-X-[R/K]-R, wherein X can be any amino acid, and [R/K] indicated that the amino acid can be R or K.
143. The conjugate according to claim 142, wherein each of the PCSK5 enzymatic cleavage sites independently comprises the sequence RRRR (SEQ ID NO: 900) or (RKR)_n (SEQ ID NO: 901), where n is 2.
144. The conjugate according to claim 142, wherein the PCSK5 enzymatic cleavage site at the C-terminal end of the scFc linker comprises the sequence RRRR (SEQ ID NO: 900) and the PCSK5 enzymatic cleavage site at the N-terminal end of the scFc linker comprises the sequence (RKR)₂ (SEQ ID NO: 901).
145. The conjugate according to any one of claims 124, 125, and 132 to 144, wherein the scFc linker has a length of about 10 to about 50 amino acids.

146. The conjugate according to any one of claims 124, 125, and 134 to 144, wherein the scFc linker has a length of about 20 to about 30 amino acids.
147. The conjugate according to any one of claims 124, 125, and 134 to 146, wherein the scFc linker comprises a gly/ser peptide.
148. The conjugate according to claim 147, wherein the gly/ser peptide comprises an amino acid sequence of formula $(\text{Gly}_4\text{Ser})_n$ or $\text{Ser}(\text{Gly}_4\text{Ser})_n$, wherein n is a positive integer selected from the group consisting of 1, 2, 3, 4, 5, 6, 7, 8, 9 and 10.
149. The conjugate according to claim 147, wherein the $(\text{Gly}_4\text{Ser})_n$ peptide comprises an amino acid sequence selected from the group consisting of $(\text{Gly}_4\text{Ser})_6$, $\text{Ser}(\text{Gly}_4\text{Ser})_6$, $(\text{Gly}_4\text{Ser})_4$ and $\text{Ser}(\text{Gly}_4\text{Ser})_4$.
150. The conjugate according to claim 149, wherein the $(\text{Gly}_4\text{Ser})_n$ peptide is a $(\text{Gly}_4\text{Ser})_6$ peptide.
151. The conjugate of claim 126 to 150, wherein FIX is full-length FIX or mature FIX.
152. The conjugate of claim 126 to 150, wherein FVIII is full-length FVIII, mature FVIII, or B domain deleted FVIII (BDD FVIII).
153. The conjugate of claim 126 to 150 and 152, wherein FVIII is a heterodimer comprising a heavy chain and a light chain, wherein the heavy chain and the light chain are associated with each other by a metal bond.
154. The conjugate of claim 126 to 150, wherein FVIIa is a heterodimer comprising a heavy chain and a light chain, wherein the heavy chain and the light chain are associated with each other by a covalent bond.
155. A conjugate comprising,
 - (a) a polypeptide selected from a heavy chain of FVIII (FVIII HC), a light chain of FVIII (FVIII LC), a heavy chain of FIX (FIX HC), a light chain of FIX (FIX LC), a heavy chain of activatable or activated FVII (FVIIa HC), a light chain of activatable or activated FVII (FVIIa LC), and a platelet targeting moiety, and

- (b) a compound of any one of claims 1 to 82, wherein the compound is linked to the polypeptide, optionally via a linker.
156. The conjugate of claim 155, wherein the N-terminus, C-terminus, or an internal amino acid residue of the compound of any one of claims 1 to 82 is linked to the N-terminus, C-terminus, or an internal amino acid residue of the polypeptide via the linker.
157. The conjugate of claim 155, wherein the compound of any one of claims 1 to 82 is linked to (1) the C-terminus of FIX HC via a linker, (2) the C-terminus of FIX LC via a linker, (3) the C-terminus of FIX LC via a first linker and the N-terminus of FIX HC via a second linker, or (4) an internal amino acid residue of FIX HC or FIX LC via a linker.
158. The conjugate of claim 155 or 156, wherein the compound of any one of claims 1 to 82 is linked to the N-terminus or the C-terminus of FVIIa HC, the C-terminus of FVIIa LC, or an internal amino acid residue of FVIIa HC or FVIIa LC.
159. The conjugate of claim 158, wherein FVIIa HC or FVIIa LC forms a heterodimer with FVIIa LC or FVIIa HC, respectively.
160. The conjugate of claim 155 or 156, wherein the compound is linked to the N-terminal of activatable FVII HC via a linker, wherein said linker comprises a protease-cleavable substrate.
161. The conjugate of claim 155 or 156, wherein the compound of any one of claims 1 to 82 is linked to the N terminus or the C terminus of FVIII HC, the N-terminus or the C-terminus of FVIII LC, or an internal amino acid residue of FVIII HC or FVIII LC.
162. The conjugate of claim 161, wherein FVIII HC or FVIII LC forms a heterodimer with FVIII LC or FVIII HC, respectively.
163. The conjugate of claim 155 or 156, wherein the compound of any one of claims 1 to 82 is linked to the N-terminus or the C-terminus of the platelet targeting moiety.
164. The conjugate of claim 157, wherein the compound or the Factor IX HC is further linked to a heterologous moiety (Het or Het1) by an optional linker.

165. The conjugate of claim 158 to 160, wherein the compound or FVIIa HC is further linked to a heterologous moiety (Het or Het1) by an optional linker.
166. The conjugate of claim 161 or 162, wherein the compound or FVIII LC is further linked to a heterologous moiety (Het or Het1) by an optional linker.
167. The conjugate of claim 155 or 156, wherein the compound or the targeting moiety is further linked to a heterologous moiety (Het or Het1) by an optional linker.
168. The conjugate of claim 164 to 167, wherein the heterologous moiety (Het or Het1) comprises an immunoglobulin constant region or a portion thereof, albumin, transferrin, an albumin binding moiety, a PAS sequence, a HES sequence, the β subunit of the C-terminal peptide (CTP) of human chorionic gonadotropin, polyethylene glycol (PEG), hydroxyethyl starch (HES), albumin-binding small molecules, or any combinations thereof.
169. The conjugate according to claim 168, wherein the heterologous moiety is an Fc moiety or an FcRn binding partner.
170. The conjugate according to claim 164 to 169, further comprising a second heterologous moiety (Het2).
171. The conjugate of claim 170, wherein the second heterologous moiety (Het2) comprises an immunoglobulin constant region or a portion thereof, albumin, transferrin, an albumin binding moiety, a PAS sequence, a HES sequence, the β subunit of the C-terminal peptide (CTP) of human chorionic gonadotropin, polyethylene glycol (PEG), hydroxyethyl starch (HES), albumin-binding small molecules, or any combinations thereof.
172. The conjugate of claim 171, wherein the immunoglobulin constant region or a portion thereof is an Fc moiety or an FcRn binding partner.
173. The conjugate according to claim 155, 156, and 167, wherein the platelet targeting moiety is capable of binding to a molecule selected from GpIb (of the GpIb/V/IX complex), GpVI, a nonactive form of GPIIb/IIIa, the active form of GPIIb/IIIa, CD62P/P selectin, GMP-33, LAMP-1, LAMP-2, CD40L, and LOX-1.

174. The conjugate according to claim 173, wherein the platelet targeting moiety is selected from MB9, SCE5, scFv, AP3, peptides targeting GPIIb/IIIa, OS1, OS2 and PS4 targeting GP1b.
175. The conjugate according to claim 174, wherein the platelet targeting moiety is PDG-13.
176. The conjugate according to any one of claims 113 to 175, which is polysialylated, pegylated, glycosylated, hesylated, gamma-carboxylated, or any combinations thereof.
177. A nucleic acid molecule or a set of nucleic acid molecules encoding a conjugate according to any one of claims 113 to 176 or a complement thereof.
178. A vector or a set of vectors comprising the nucleic acid molecule or the set of the nucleic acid molecules of claim 177 or a complement thereof.
179. The vector or a set of vectors according to claim 178, further comprising a nucleotide sequence encoding an intracellular processing enzyme which processes at least one of the intracellular processing sites in the chimeric protein or a complement thereof.
180. A host cell comprising the vector according to any one of claims 178 to 179.
181. The host cell of claim 180, which is a HEK293 cell, a BHK cell, or a CHO cell.
182. A method for producing a chimeric protein comprising culturing the host cell according to claim 180 or 182 and recovering the chimeric protein from the culture medium.
183. A pharmaceutical composition comprising the conjugate of any one of claims 112 to 176, the nucleic acid molecule or the set of nucleic acid molecules according to claim 177, the vector or the set of vectors of any one of claims 178 or 179 and a pharmaceutically acceptable carrier.
184. A method for treating, ameliorating, or preventing a bleeding disease or disorder in a subject in need thereof, comprising administering to the subject an effective amount of the conjugate of any one of claims 112 to 176, the nucleic acid molecule or the set of nucleic acid molecules according to claim 177, the vector or the set of vectors of any one of claims 178 or 179, or a pharmaceutical composition according to claim 183.

185. The method according to claim 184, wherein the bleeding disease or disorder is caused by a blood coagulation disorder.
186. The method according to claim 185, wherein the blood coagulation disorder is hemophilia A or hemophilia B.
187. The method according to any one of claims 184 to 186, wherein the bleeding disease or disorder is selected from hemarthrosis, muscle bleed, oral bleed, hemorrhage, hemorrhage into muscles, oral hemorrhage, trauma, trauma capitis, gastrointestinal bleeding, intracranial hemorrhage, intra-abdominal hemorrhage, intrathoracic hemorrhage, bone fracture, central nervous system bleeding, bleeding in the retropharyngeal space, bleeding in the retroperitoneal space, and bleeding in the iliopsoas sheath.
188. A method of treating, ameliorating, or preventing a coagulation factor deficiency in a mammalian subject comprising administering to the subject an effective amount of the conjugate of any one of claims 112 to 176, the nucleic acid molecule or the set of nucleic acid molecules according to claim 177, the vector or the set of vectors of any one of claims 178 or 179, or the pharmaceutical composition of claim 183, wherein the coagulation factor is selected from the group consisting of FVII, FVIIa, FVIII, FIX, and FXI.
189. The method according to any one of claims 184 to 188, wherein the subject is a human subject.
190. The conjugate of any one of claims 112 to 176, the nucleic acid molecule or the set of nucleic acid molecules according to claim 177, the vector or the set of vectors of any one of claims 178 or 179, or the pharmaceutical composition of claim 183 for treating, ameliorating, or preventing a subject having a blood coagulation disorder.
191. Use of the conjugate of any one of claims 112 to 176, the nucleic acid molecule or the set of nucleic acid molecules according to claim 177, the vector or the set of vectors of any one of claims 178 or 179, or the pharmaceutical composition of claim 183 for the manufacture of a medicament for the treatment, prevention, or amelioration of a blood coagulation disorder.

192. A method for making the conjugate of any one of claims 112 to 176, the method comprising using solid-phase peptide synthesis.
193. The method according to claim 192, comprising using orthogonal solid-phase peptide synthesis.

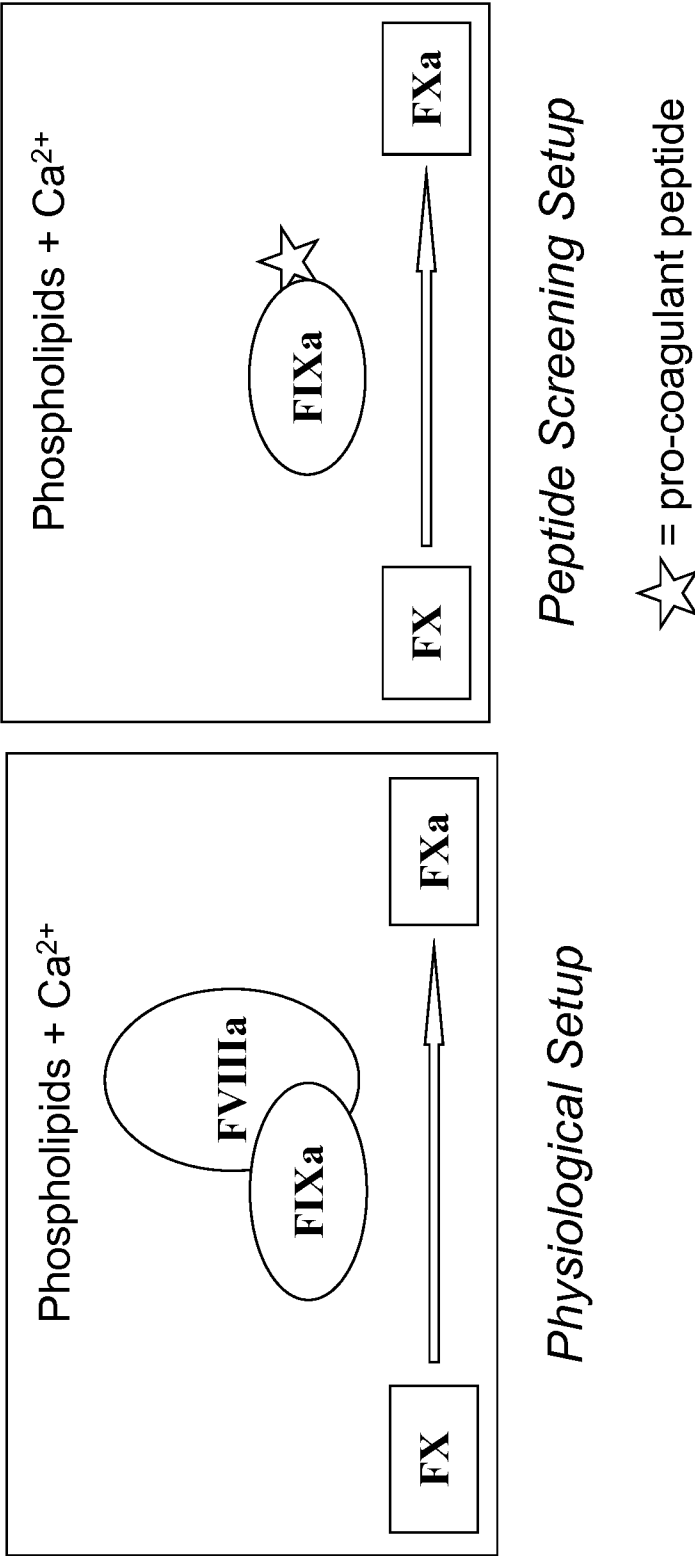
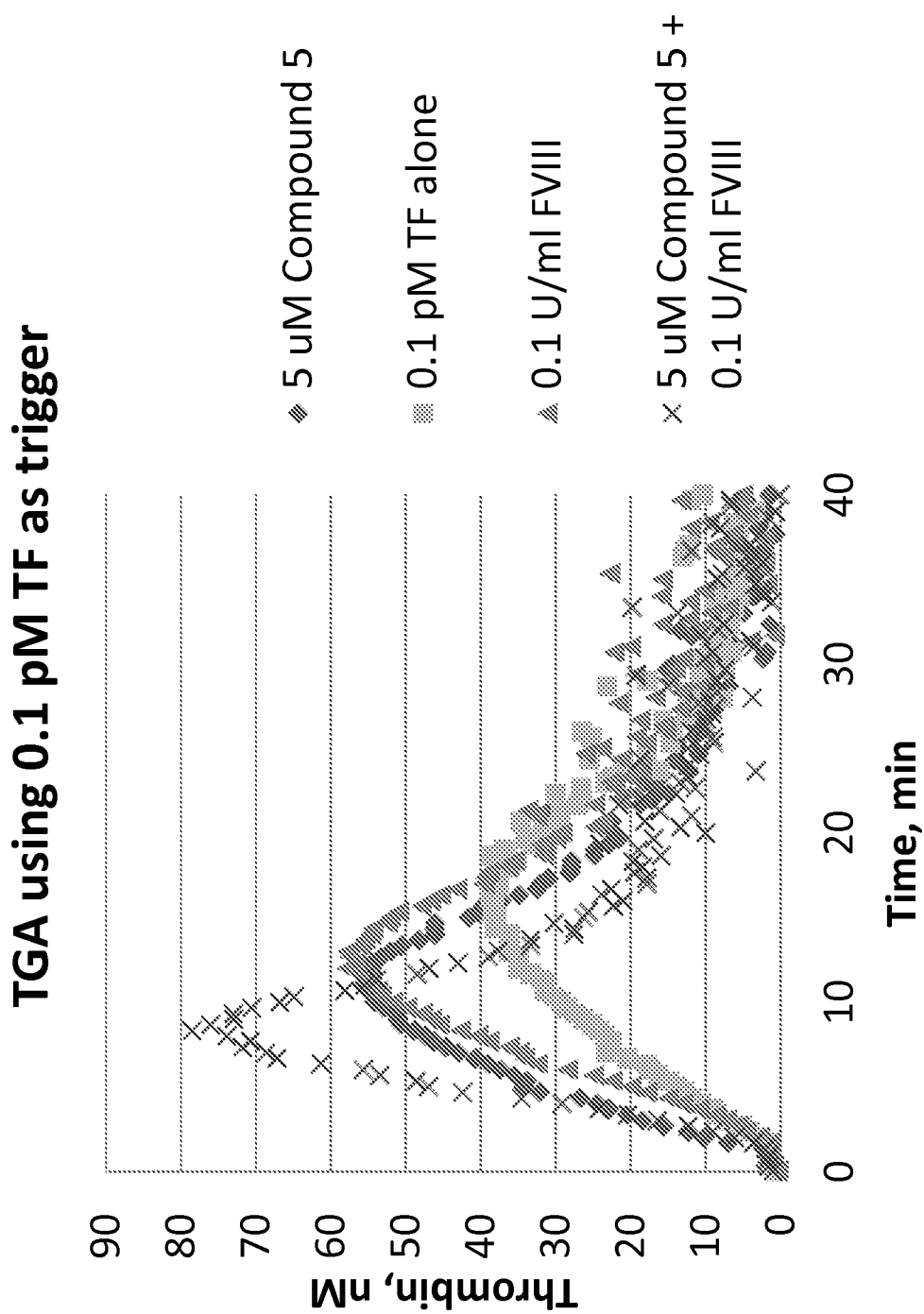
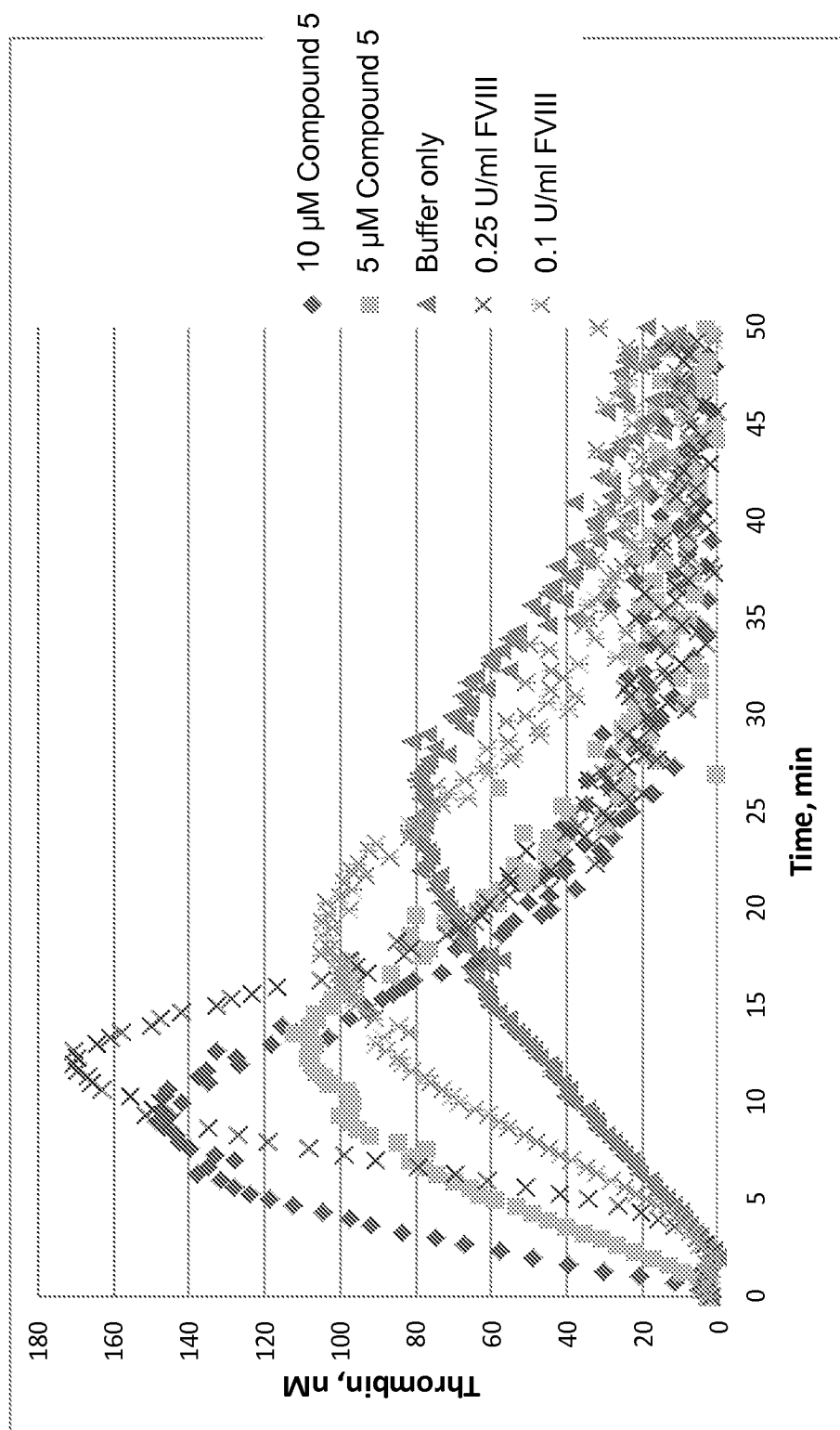


FIG. 1

**FIG. 2**

**FIG. 3**

Deuterium Level, HC 177-185

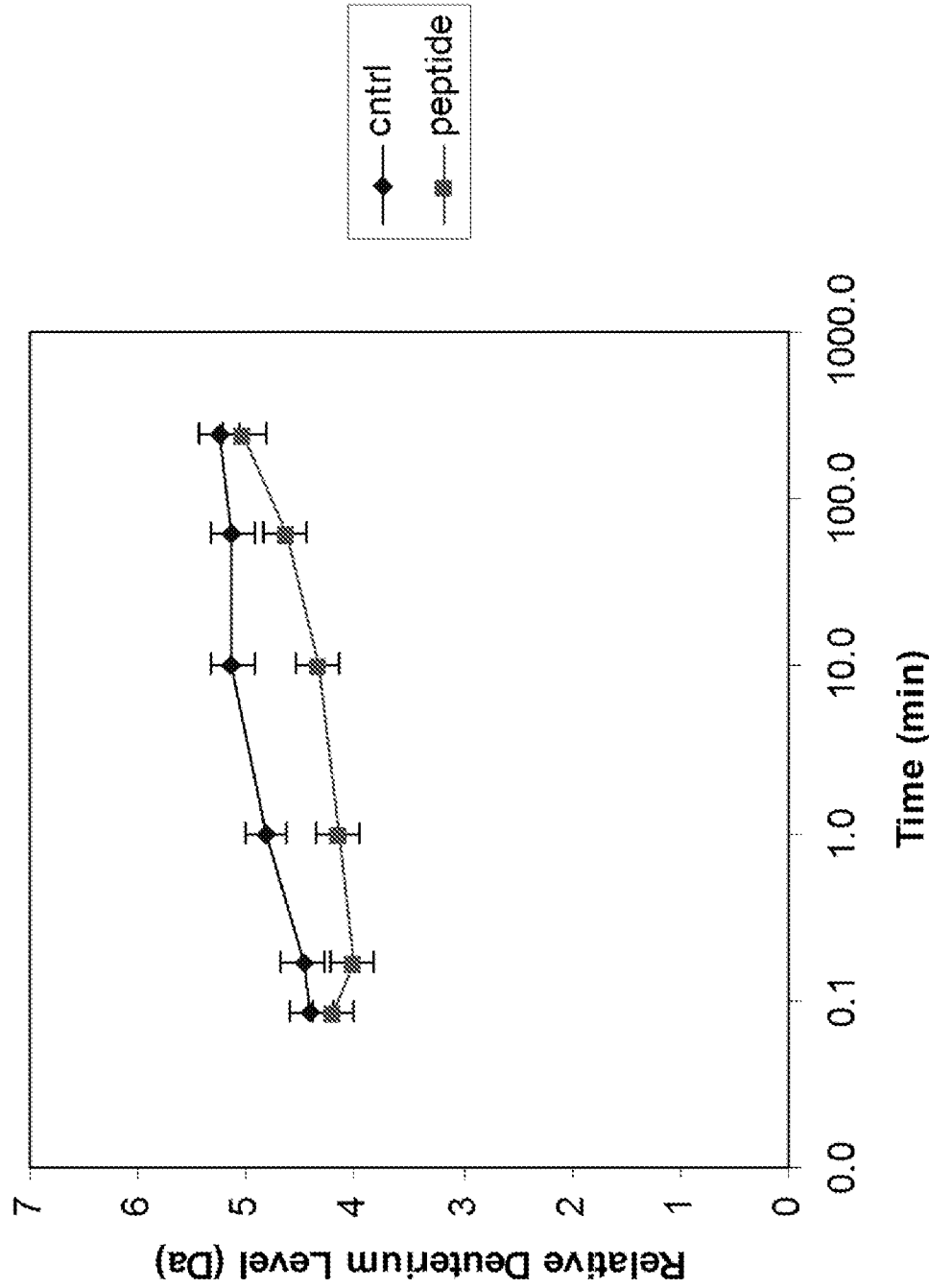
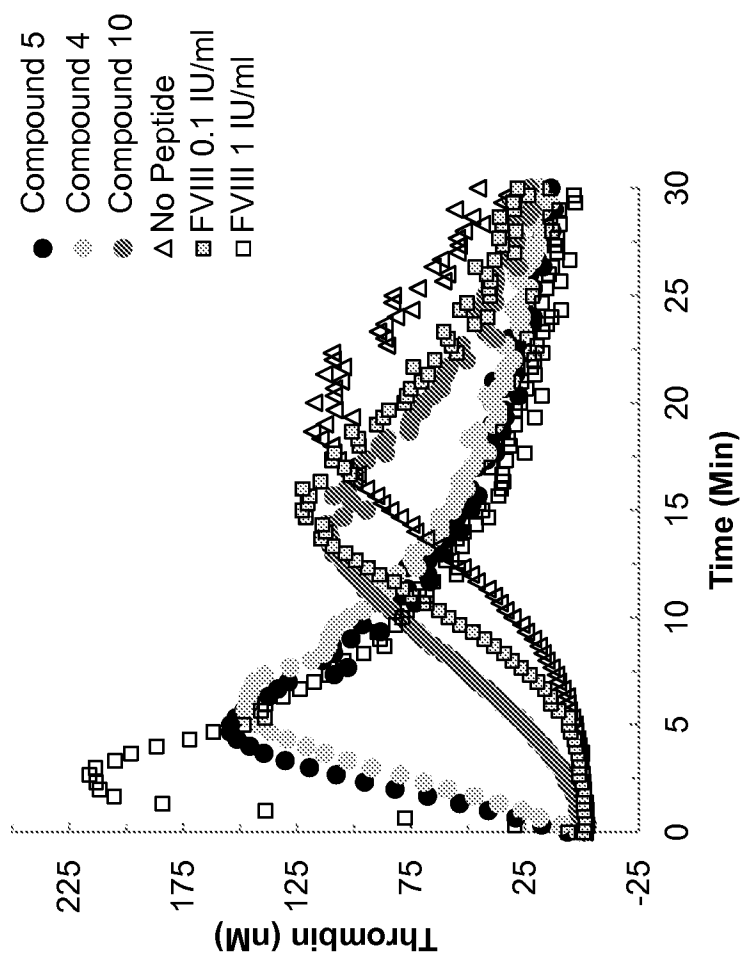


FIG. 4

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**FIG. 5**

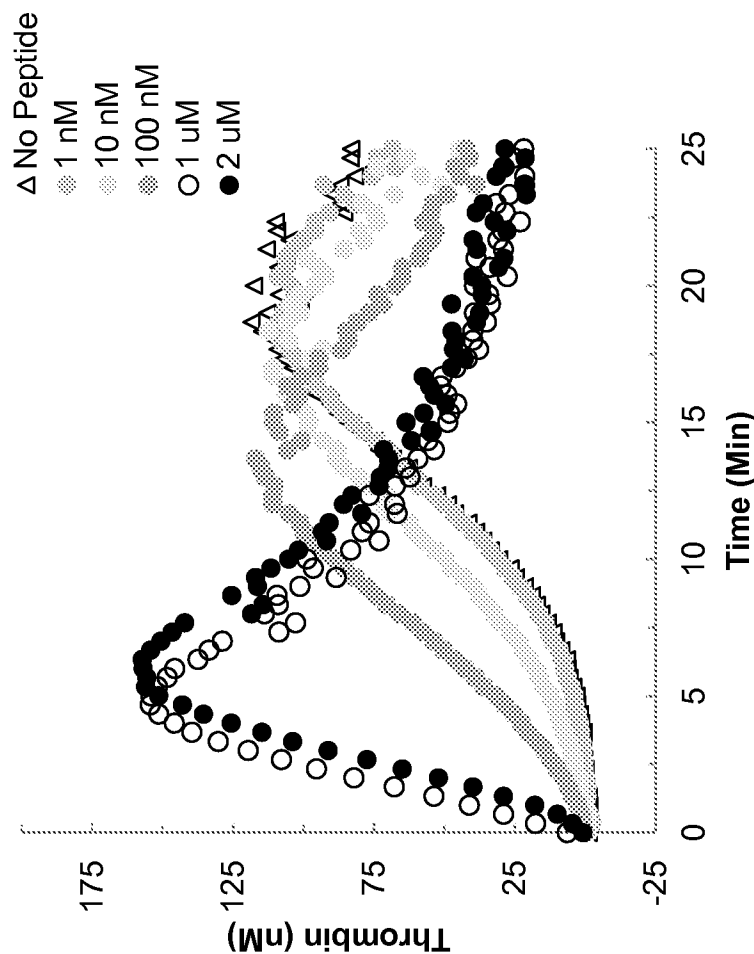


FIG. 6

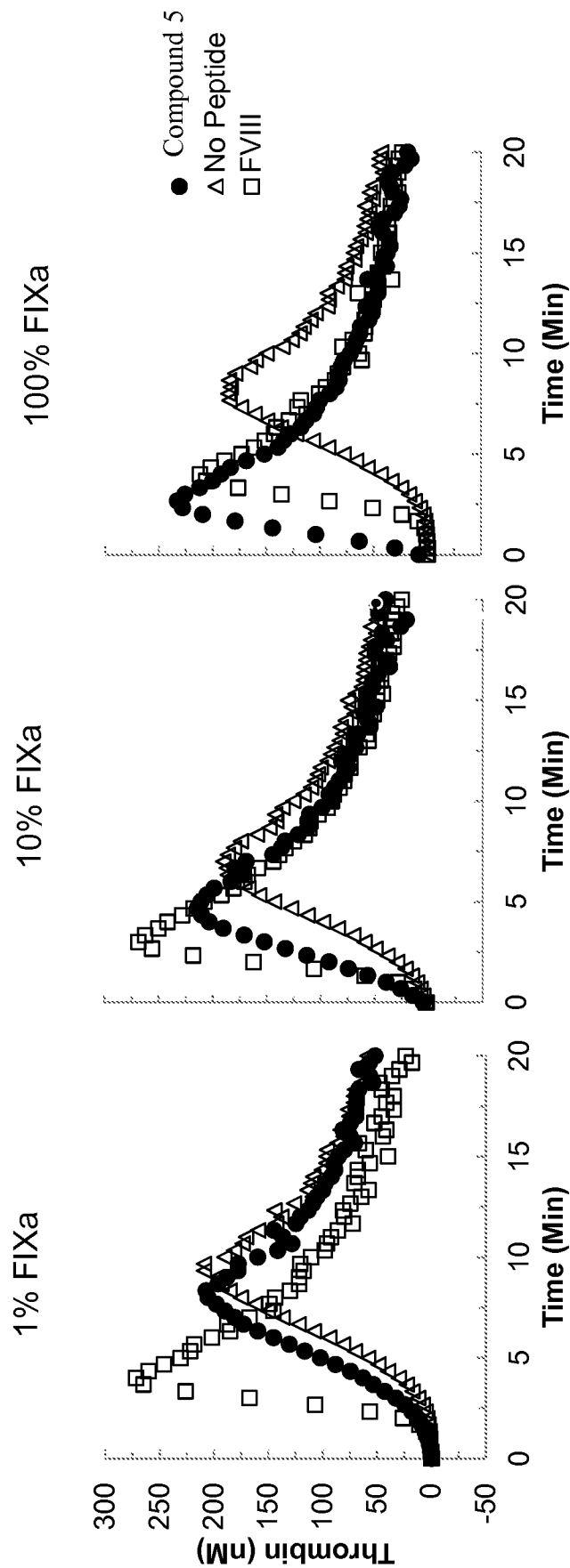


FIG. 7

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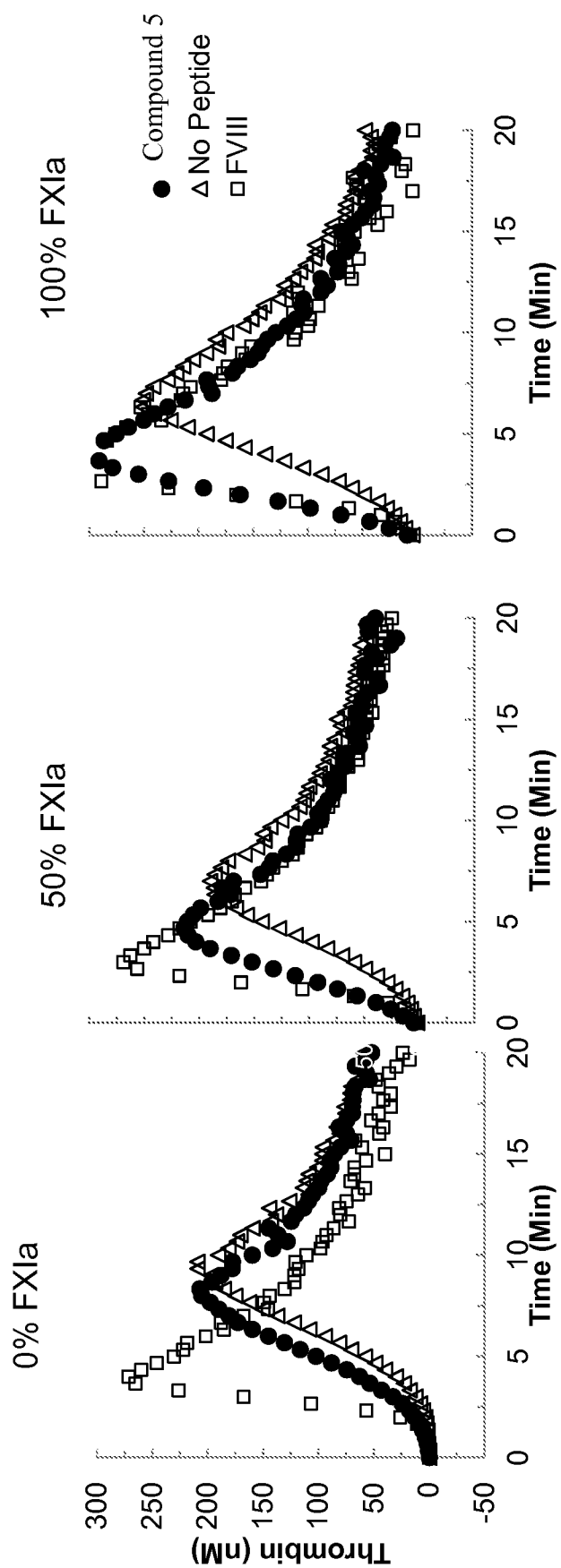


FIG. 8

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FVIIa-Fc-Compound Conjugate

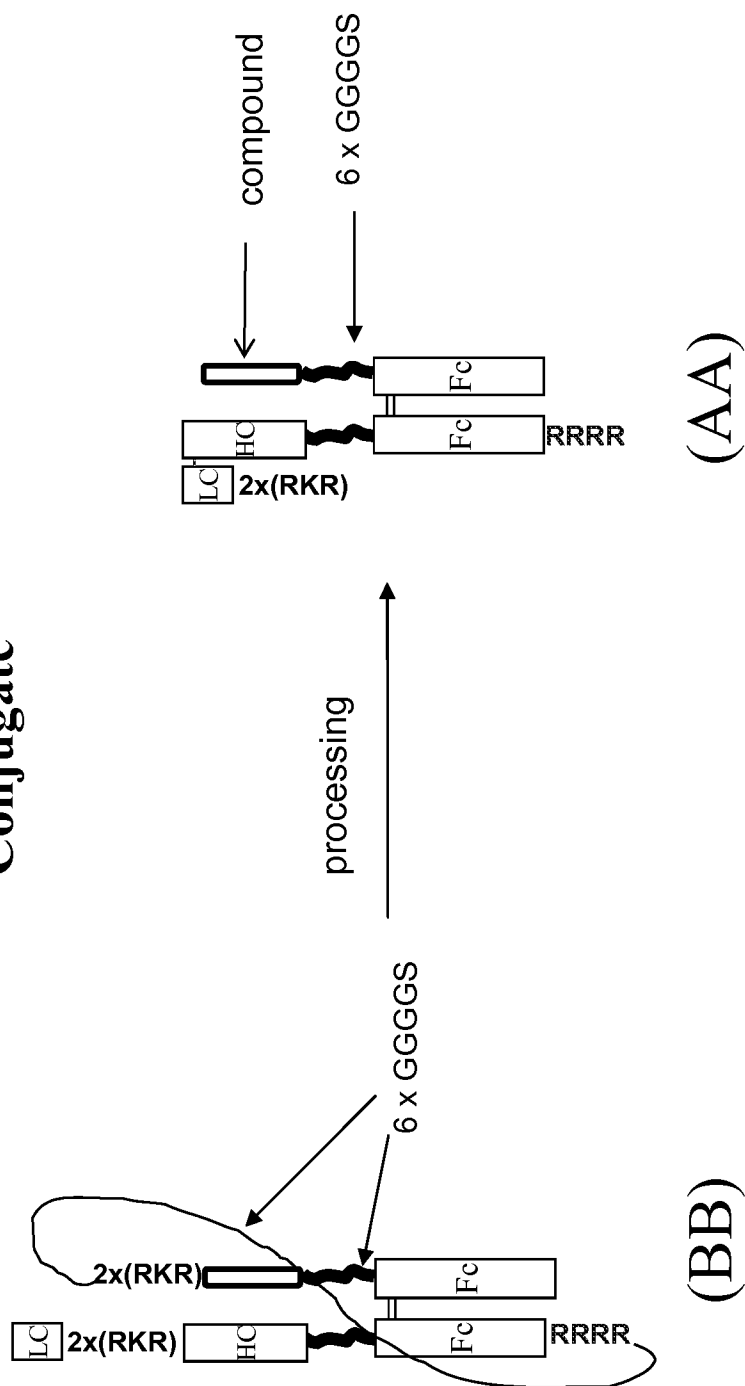


FIG. 9

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FIX-Fc-Compound Conjugate

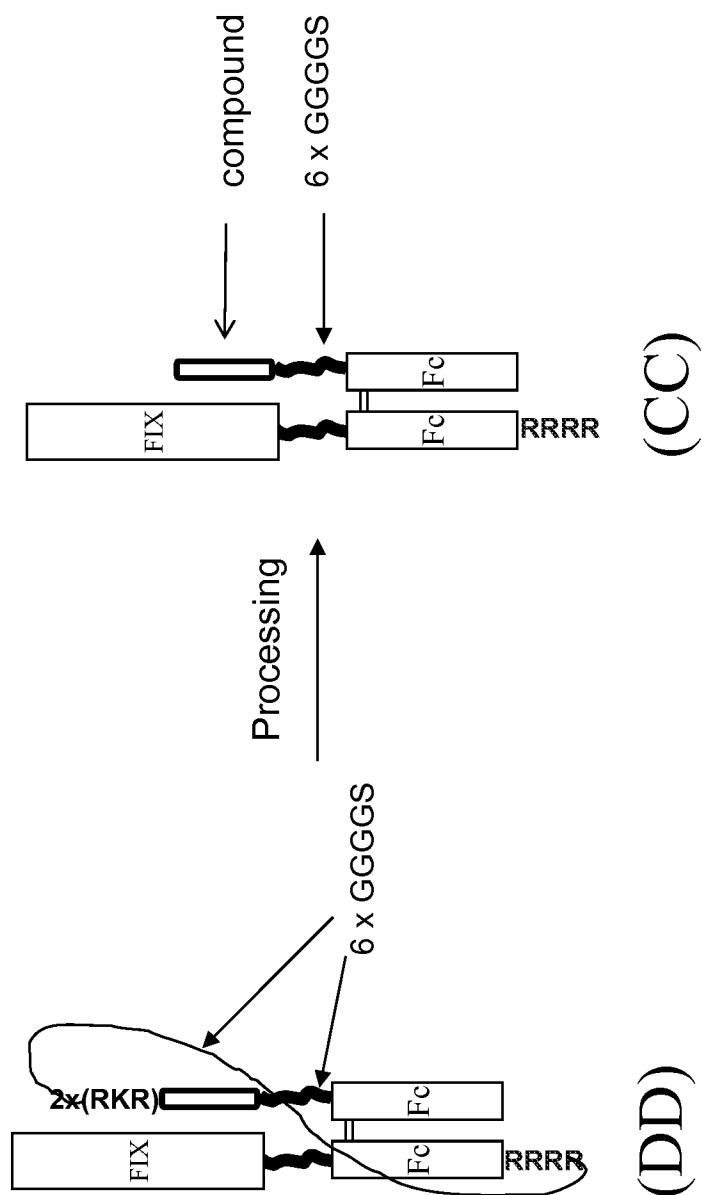


FIG. 10

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PDG-13-Fc-Compound Conjugate

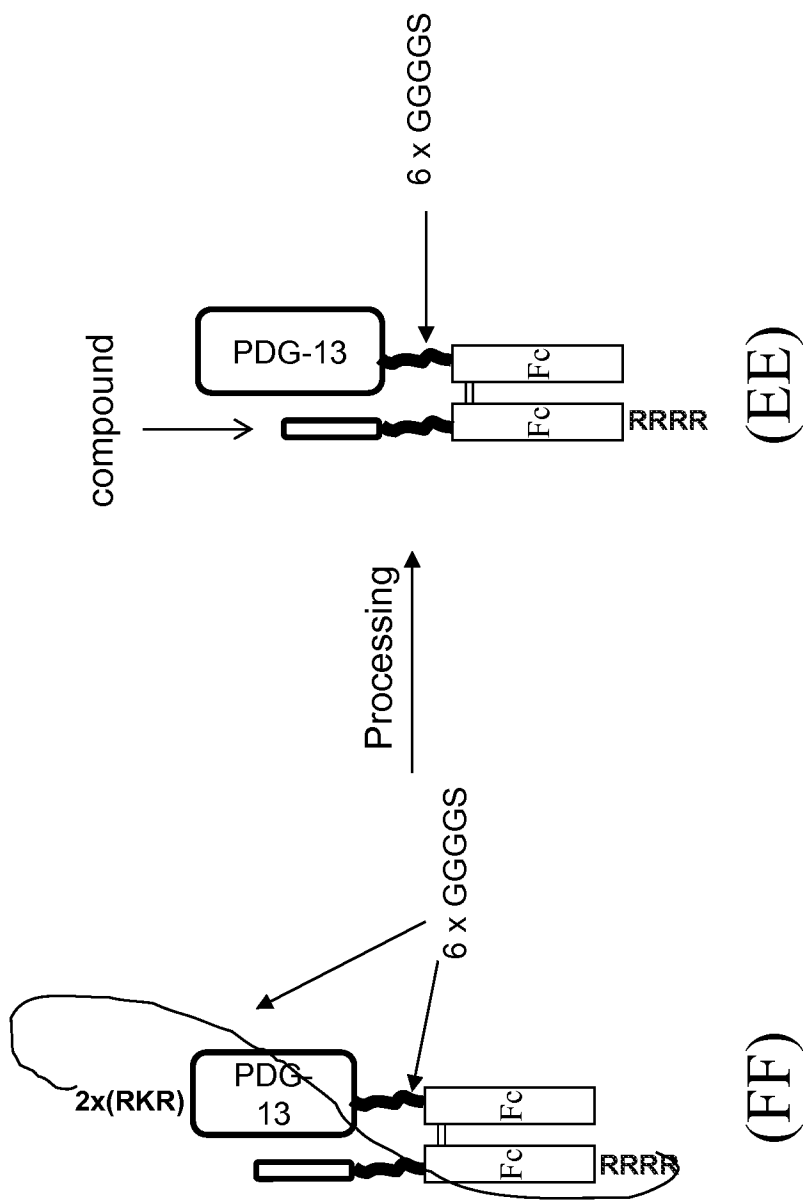


FIG. 11

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Fc-Compound Conjugate

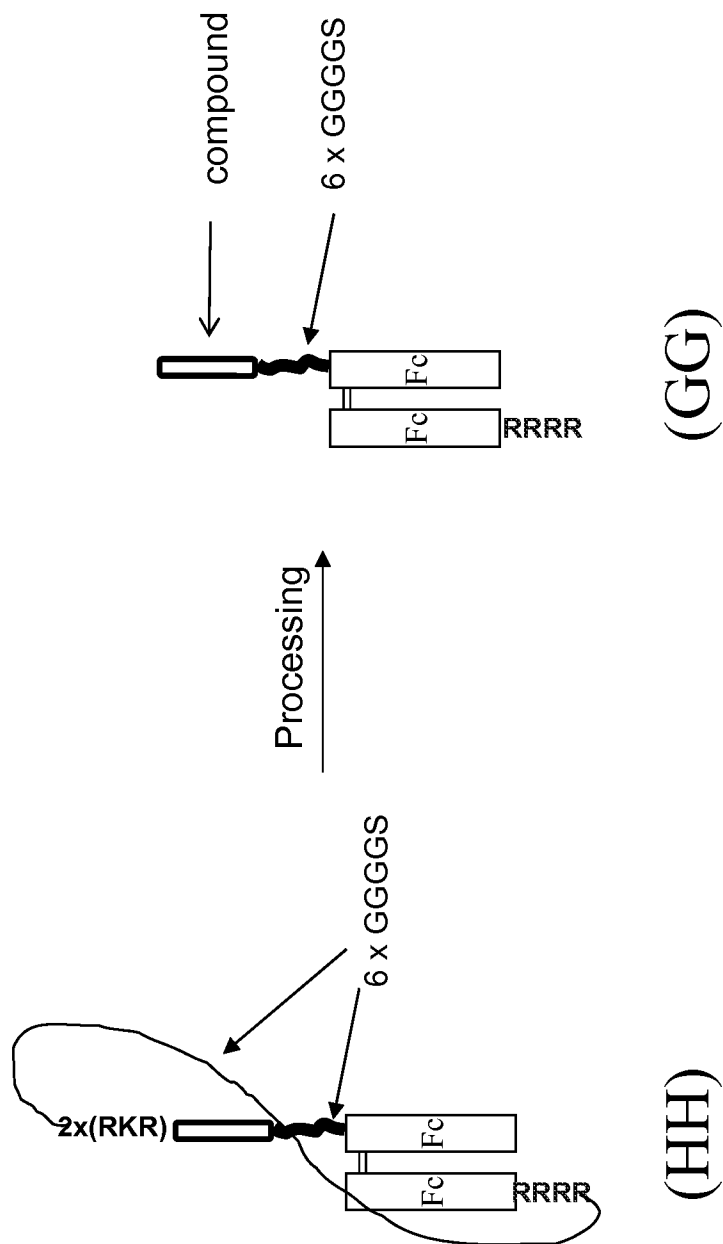


FIG. 12

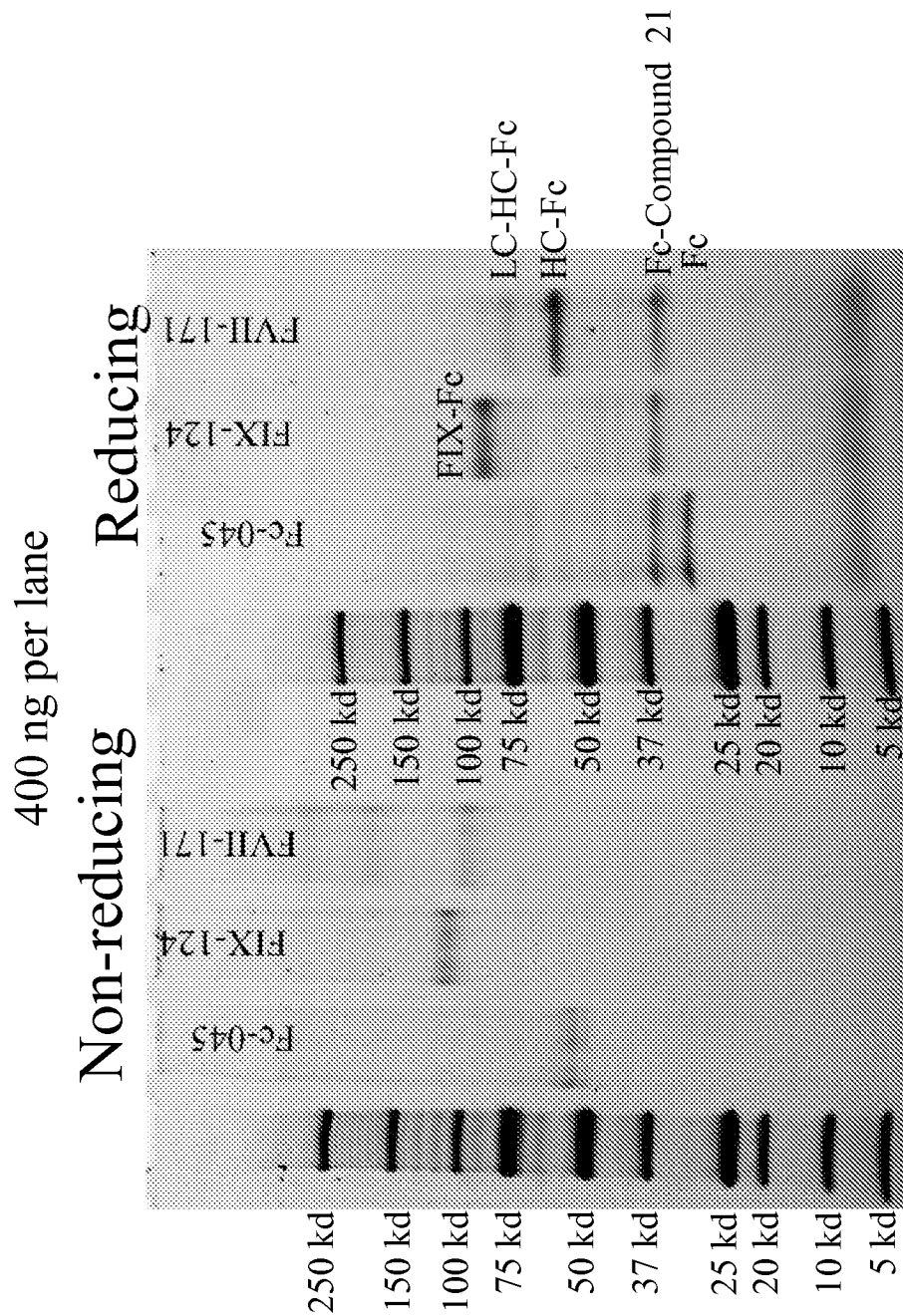


FIG. 13

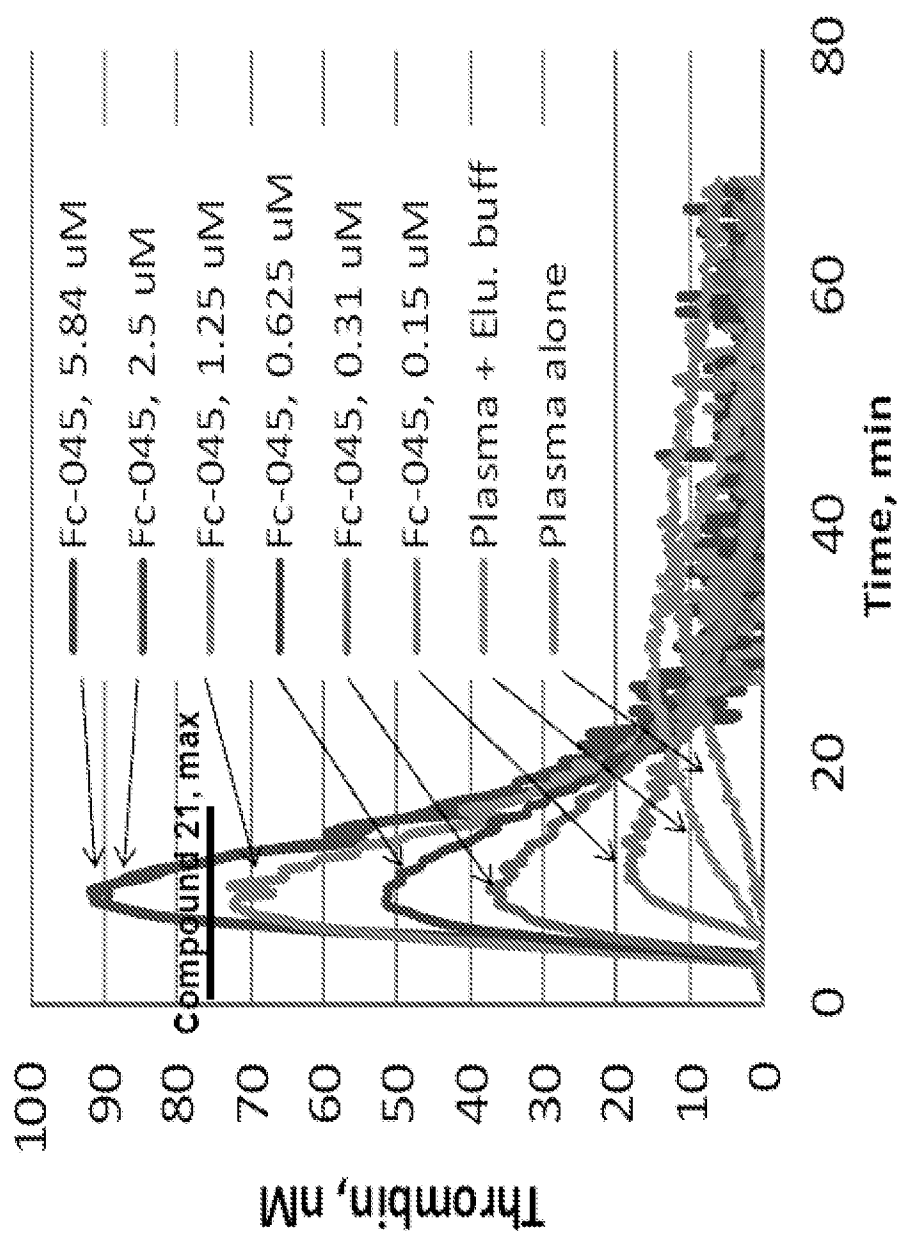
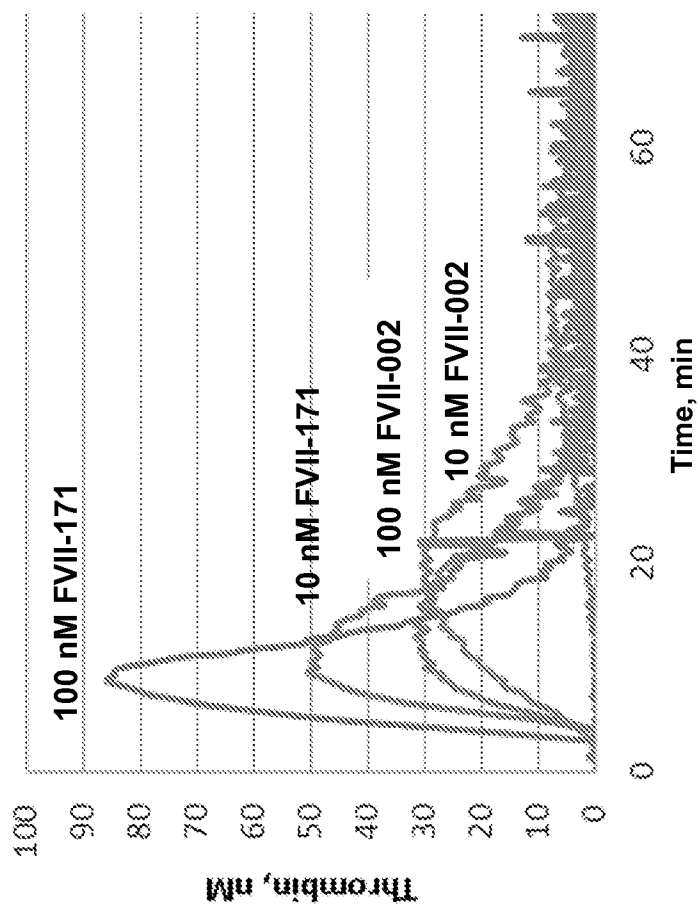


FIG. 14

**FIG. 15**

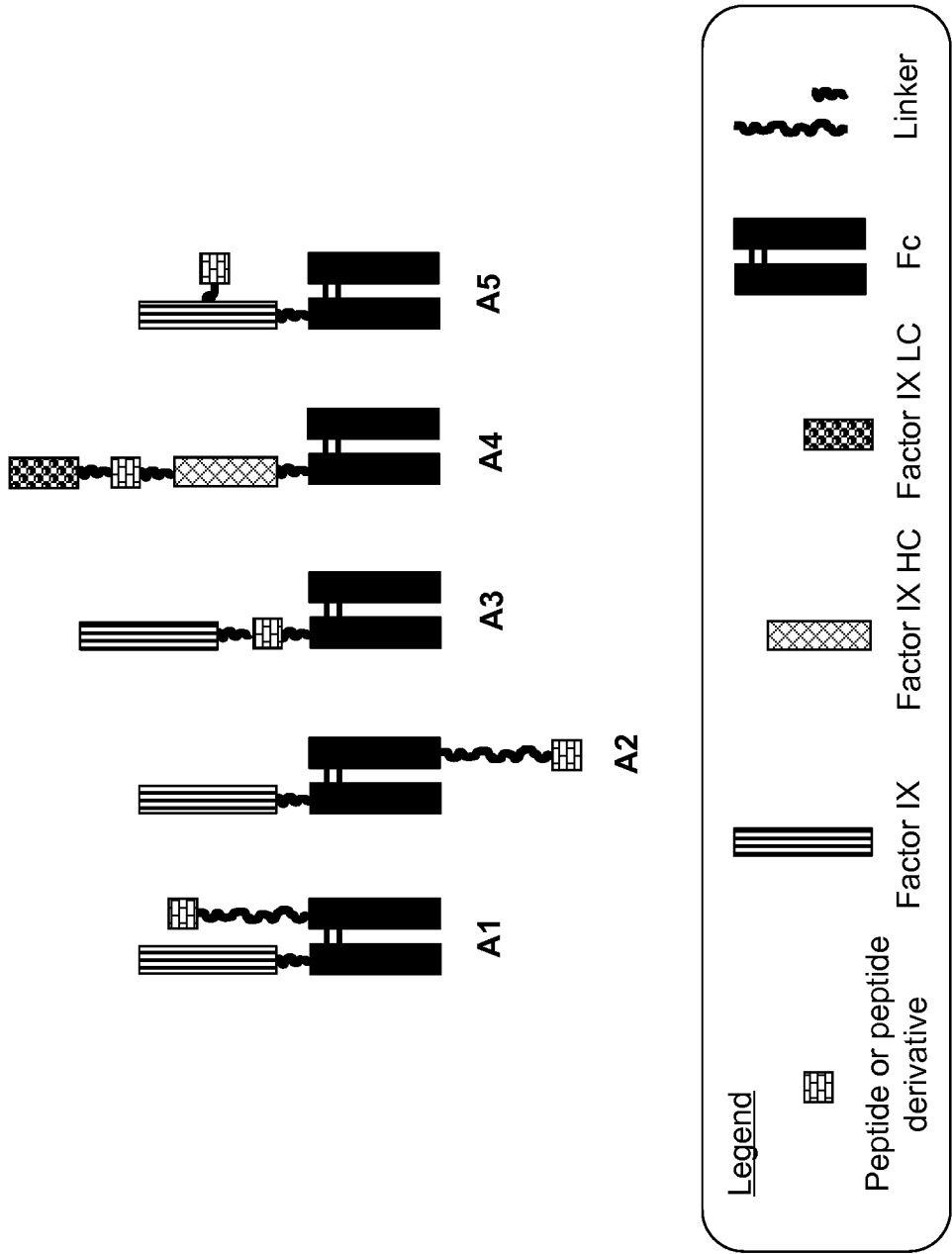


FIG. 16

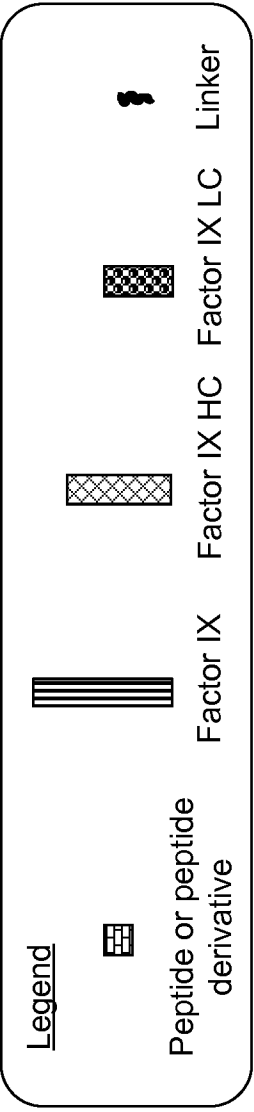
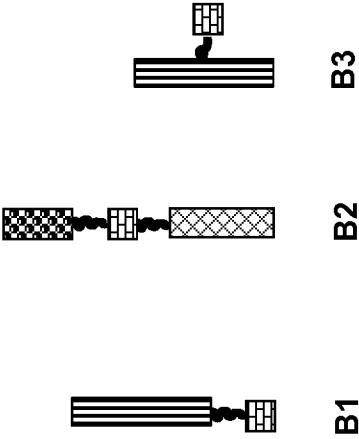


FIG. 17

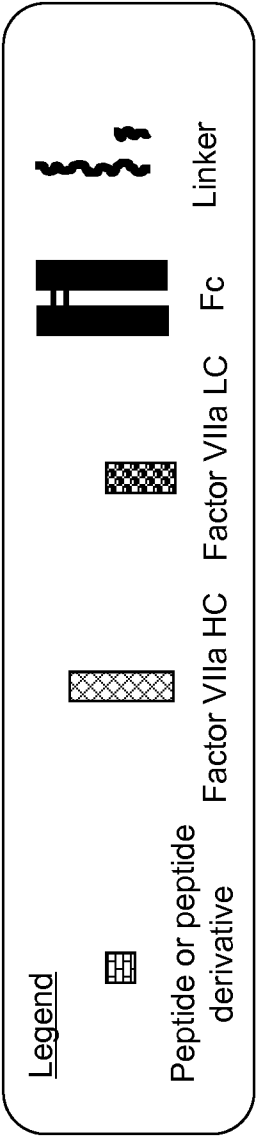
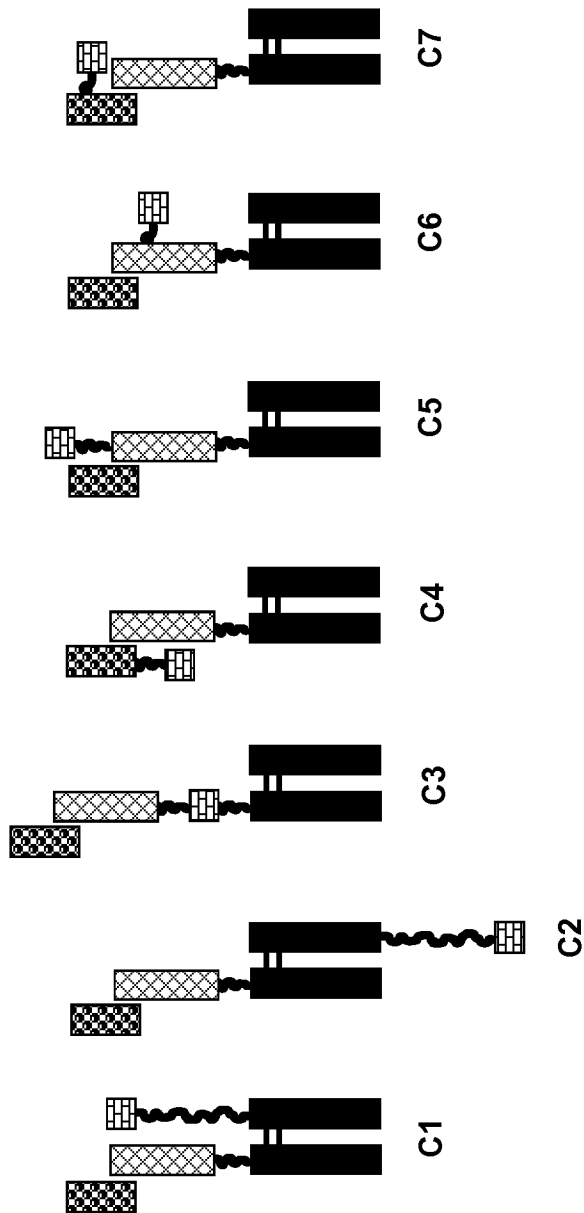


FIG. 18

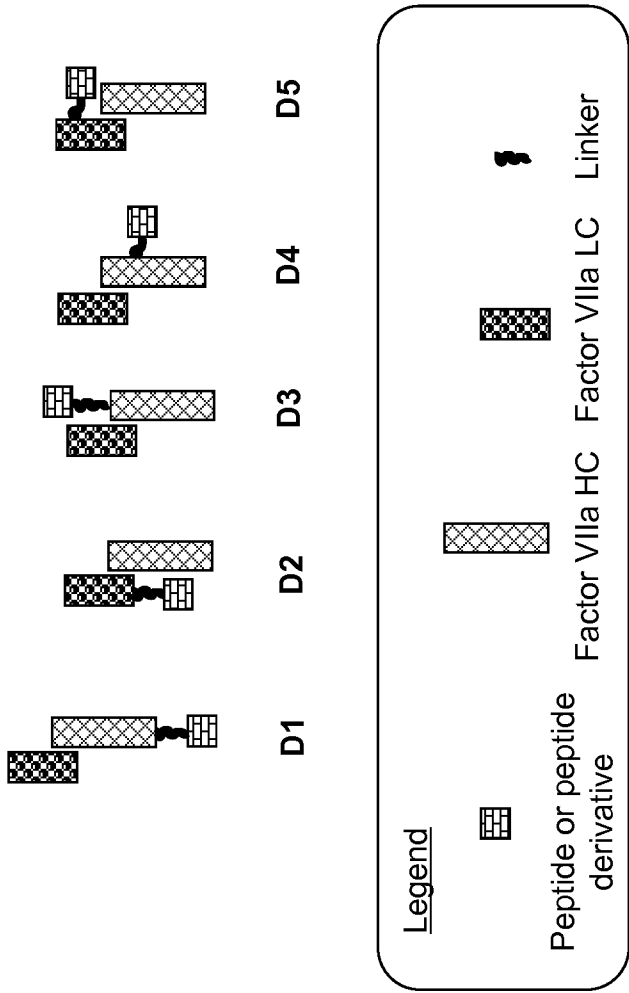


FIG. 19

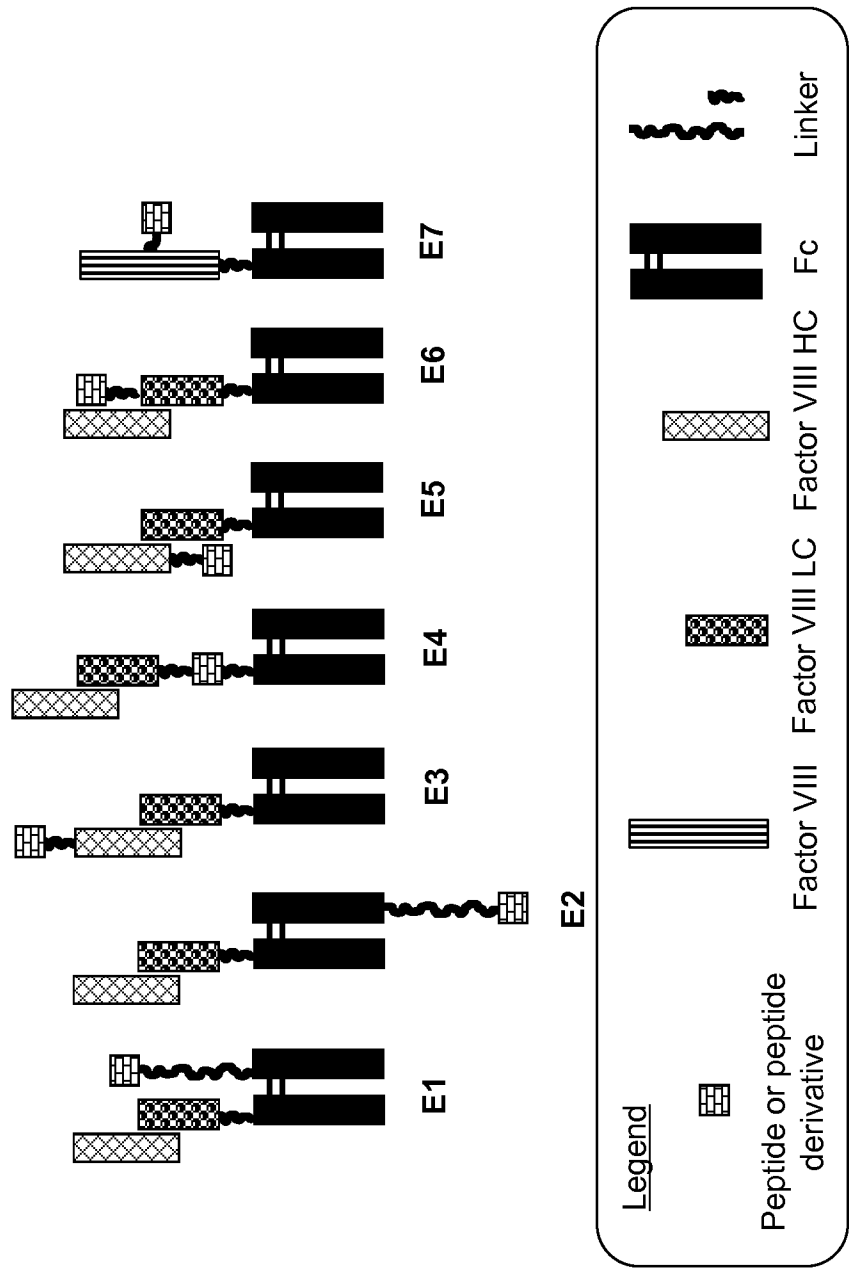


FIG. 20

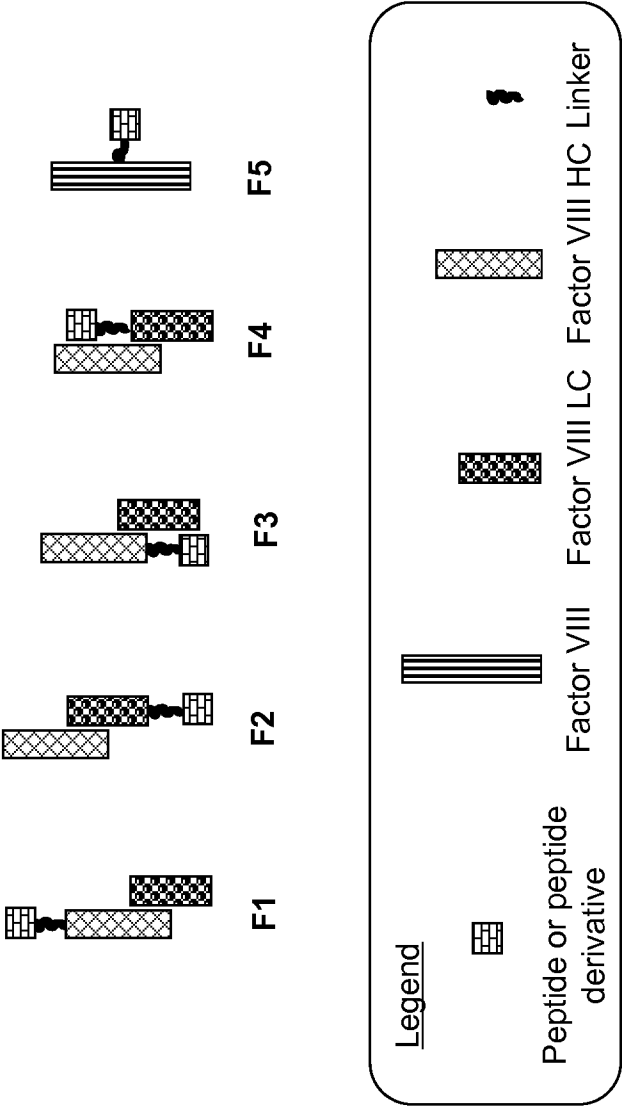


FIG. 21

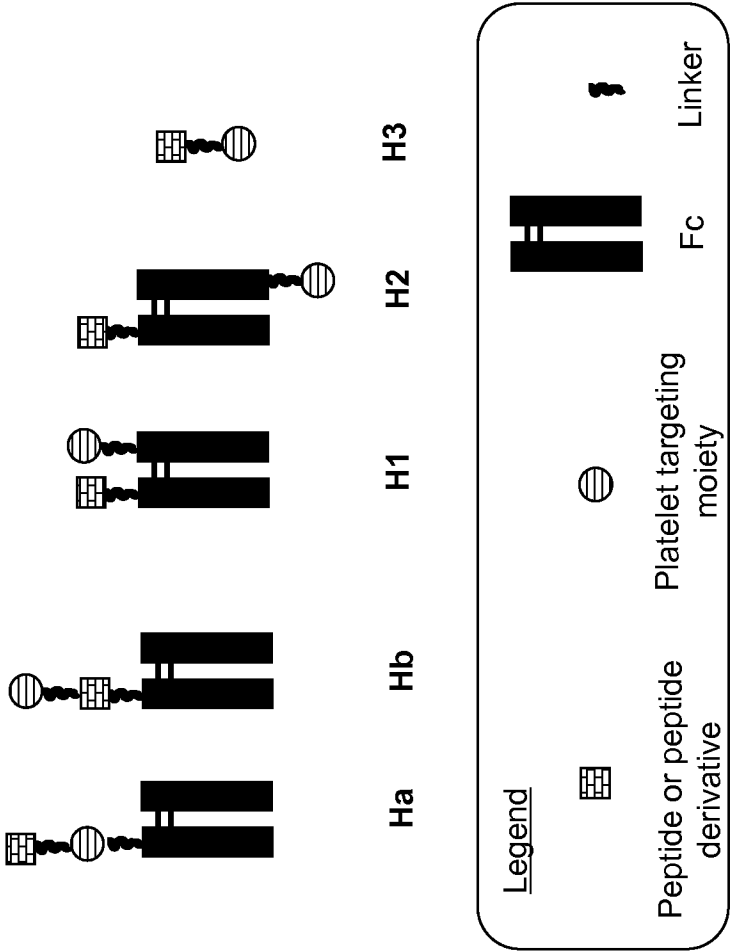


FIG. 22

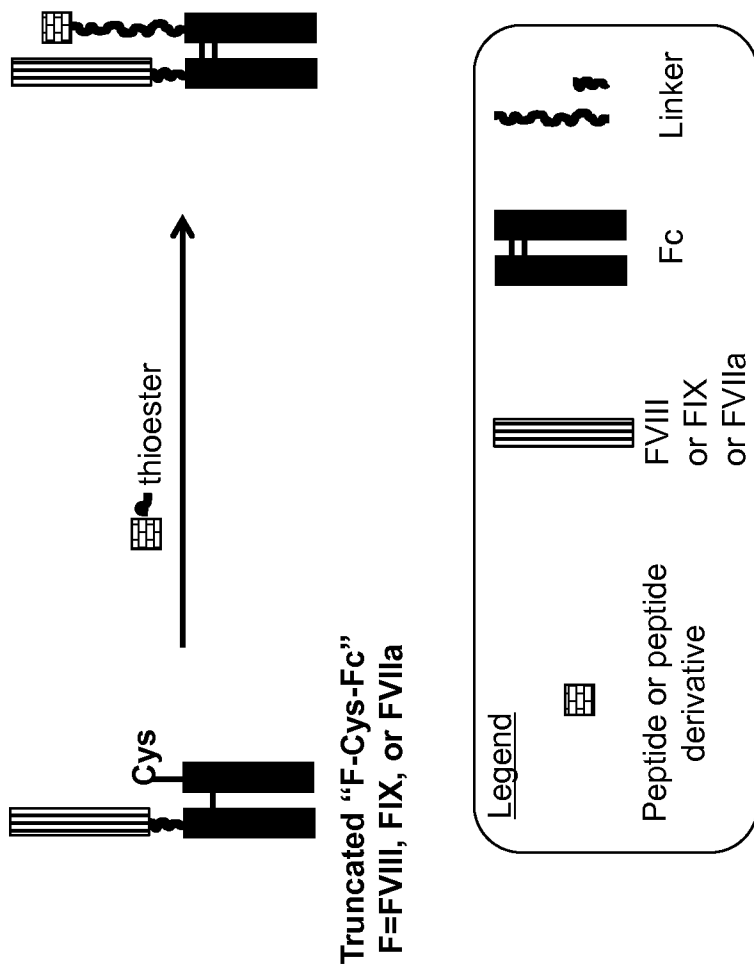


FIG. 23

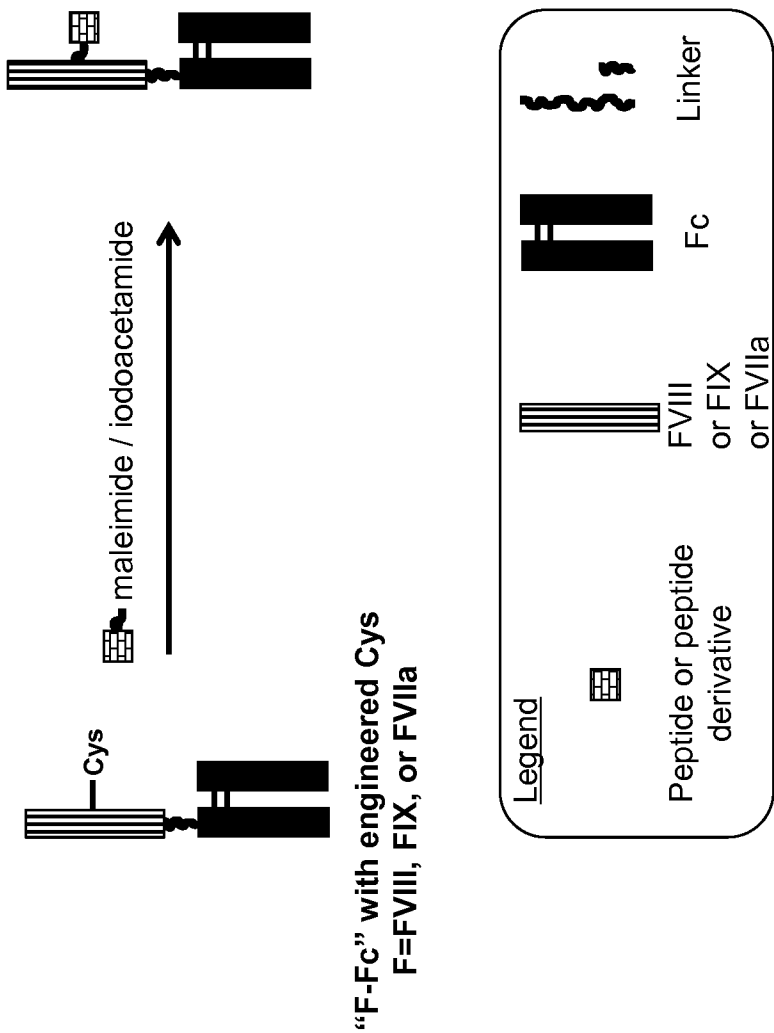
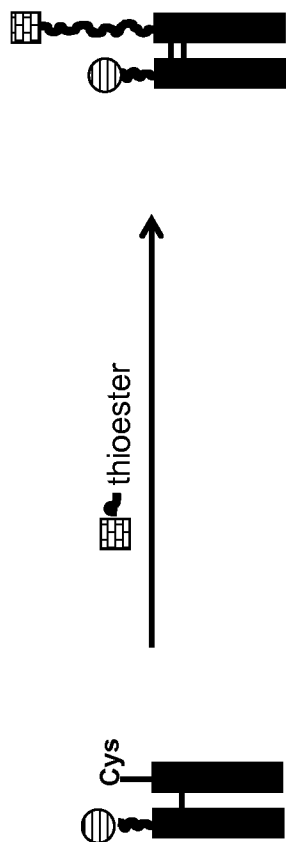


FIG. 24

Native ligation:



Truncated "Platelet-Targeting-Fc"

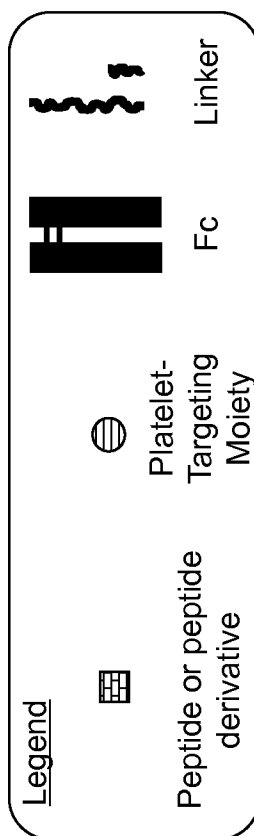
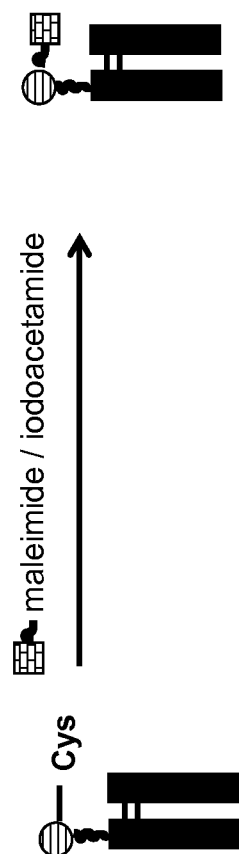


FIG. 25

Site-directed modification:



“Platelet Targeting Moiety-Fc”
with engineered Cys

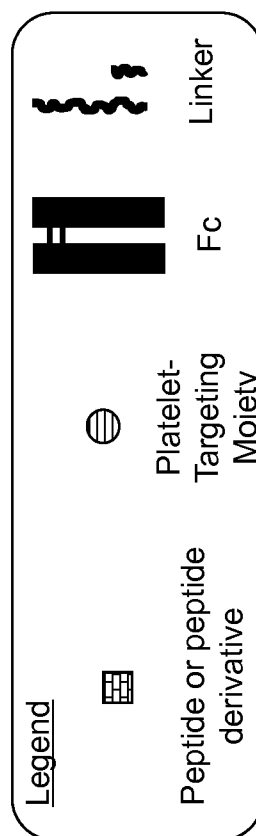


FIG. 26