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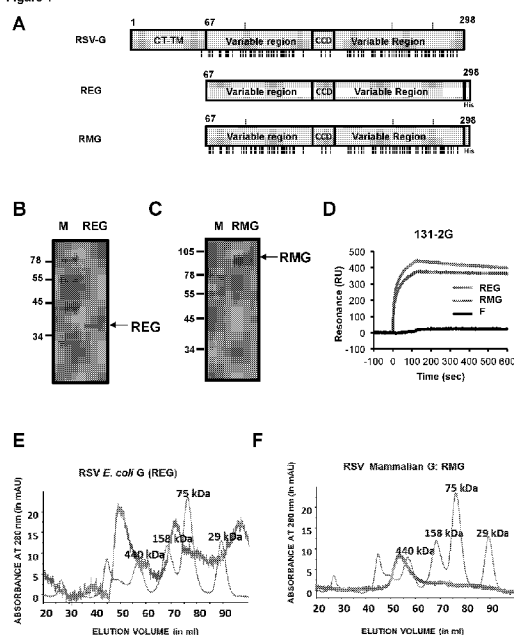
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[Continued on next page]

(54) Title: IMMUNOGENIC RSV POLYPEPTIDES

Figure 1



(57) **Abstract:** This invention provides immunogenic compositions comprising an immune stimulant and an respiratory syncytial virus (RSV) oligopeptide or an unglycosylated RSV polypeptide. The RSV oligopeptides are shown in SEQ ID NO: 3-33. The unglycosylated RSV polypeptide may consist essentially of the ectodomain of an RSV G protein, such as that shown in SEQ ID NO: 2 or the ectodomain of an RSV F protein such as the ectodomain of the F protein shown in SEQ ID NO: 39.



— with sequence listing part of description (Rule 5.2(a))

IMMUNOGENIC RSV POLYPEPTIDES

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CROSS-REFERENCES TO RELATED APPLICATIONS

[0001] This application claims priority to US Application No. 62/139,995, filed March 30, 2015, which is related to US Application No. 61/910,623, filed December 2, 2013, and
10 International Application No. PCT/US14/68144, filed December 2, 2014, all of which are incorporated herein by reference.

FIELD OF THE INVENTION

[0002] This invention relates to immunogenic polypeptides from human respiratory syncytial
15 virus (RSV) and their use to prevent, treat and diagnose RSV infection.

BACKGROUND OF THE INVENTION

[0003] Human respiratory syncytial virus (RSV) is the most important cause of virus mediated lower respiratory tract illness (LRI) in infants and children worldwide. In infants, ~ 2,345 per
20 100,000 hospitalizations are attributable to RSV infection in the United States annually and RSV is one of the leading causes of morbidity/mortality second only to influenza virus (1). Although traditionally regarded as a pediatric pathogen, RSV can cause life-threatening pulmonary disease in bone marrow transplant recipients and immunocompromised patients (2, 3). Most RSV mediated severe disease and mortality occurs in infants younger than 2 years (4). Among the
25 elderly, RSV is also a common cause of severe respiratory infections that requires hospitalization (4).

[0004] Although the importance of RSV as a respiratory pathogen has been recognized for over 50 years, a vaccine is not yet available because of several problems inherent in RSV vaccine development including the very young age of the target population, recurrent infections
30 in spite of prior exposure, and history of enhanced disease in young children that were

immunized with a formaldehyde inactivated RSV (FI-RSV) vaccine in the 1960s (3, 5). Subsequent studies with samples from these children showed poor functional antibody responses with low neutralization or fusion-inhibition titers (6, 7). There was also evidence for deposition of immune complexes in the small airways (8), however the mechanism of the FI-RSV vaccine induced enhanced disease is poorly understood. Animal models of the FI-RSV vaccine associated enhanced respiratory disease (VAERD) suggested a possible combination of poor functional antibody responses and Th2-biased hyper cytokine release leading to eosinophilic infiltration in the lungs (9, 10).

[0005] RSV live-attenuated vaccines (LAV) have a good safety record in infants. However, a recent RSV LAV candidate (rA2cp248/404/1030deltaSH) was found to be safe in infants but poorly immunogenic (11). New vaccine development efforts are now under way, using recombinant technologies, different cell substrates, and structure-based vaccine design (3). The RSV F glycoprotein (PFP-2) formulated in alum was evaluated in clinical trials and found to be well tolerated but only modestly immunogenic in adults, pregnant women, and in the elderly (12). A mixture of F, G, and M recombinant proteins was tested in subjects >65 years and was found to induce >4-fold increase in serum neutralizing activity in 58% of subjects with low pre-vaccine titers (13). Recently the structures of the F protective targets recognized by MAb palivizumab and 101F, as well as the pre-fusion form of the F protein trimer were resolved, leading to structure based design of new F-based vaccines (14-16). A subunit vaccine based on the central conserved region of the G attachment surface glycoprotein was also developed as a RSV polypeptide with the albumin-binding domain from streptococcal protein G, produced in prokaryotic cells and formulated with an alum-based adjuvant. Despite promising results in murine models, challenge studies in rhesus macaques showed no reduction of viral loads, and studies in adults showed a relatively low capacity for inducing neutralizing antibodies (17, 18). An optimally effective RSV vaccine must protect against antigenically divergent groups A and B RSV strains.

[0006] Despite advances in the development of subunit vaccines, a need still exists to produce RSV vaccines that will provide protective immunity without the potential for disease enhancement. The present invention addresses these and other needs.

BRIEF SUMMARY OF THE INVENTION

[0007] This invention provides immunogenic compositions comprising an immune stimulant and an RSV oligopeptide from the RSV F protein or G protein or a multimer thereof. In other embodiments, the composition may comprise an unglycosylated RSV polypeptide. Usually, the unglycosylated RSV polypeptide consists essentially of the ectodomain of an RSV G protein, such as one that is at least about 95% identical to that shown in SEQ ID NO: 2(RSV-A2) or SEQ ID NO: 42 (RSV-B1). Alternatively, the unglycosylated RSV polypeptide consists essentially of the ectodomain of an RSV F protein, such as one that is at least about 95% identical to the ectodomain of the F protein sequence shown in SEQ ID NO: 40. The unglycosylated RSV polypeptide can be made using a number of methods known in the art. In a typical embodiment, it is recombinantly expressed in a prokaryotic cell.

[0008] If the immunogenic composition comprises an RSV oligopeptide of the invention, the oligopeptide may be one or more of SEQ ID NO: 3-33. Thus, the composition of the invention may comprise SEQ ID NO: 3; alternatively, it may comprise SEQ ID NO: 4; alternatively, it may comprise SEQ ID NO: 5; alternatively, it may comprise SEQ ID NO: 6; alternatively, it may comprise SEQ ID NO: 7; alternatively, it may comprise SEQ ID NO: 8; alternatively, it may comprise SEQ ID NO: 9; alternatively, it may comprise SEQ ID NO: 10; alternatively, it may comprise SEQ ID NO: 11; alternatively, it may comprise SEQ ID NO: 12; alternatively, it may comprise SEQ ID NO: 13; alternatively, it may comprise SEQ ID NO: 14; alternatively, it may comprise SEQ ID NO: 15; alternatively, it may comprise SEQ ID NO: 16; alternatively, it may comprise SEQ ID NO: 17; alternatively, it may comprise SEQ ID NO: 18; alternatively, it may comprise SEQ ID NO: 19; alternatively, it may comprise SEQ ID NO: 20; alternatively, it may comprise SEQ ID NO: 21; alternatively, it may comprise SEQ ID NO: 22; alternatively, it may comprise SEQ ID NO: 23; alternatively, it may comprise SEQ ID NO: 24; alternatively, it may comprise SEQ ID NO: 25; alternatively, it may comprise SEQ ID NO: 26; alternatively, it may comprise SEQ ID NO: 27; alternatively, it may comprise SEQ ID NO: 28; alternatively, it may comprise SEQ ID NO: 29; alternatively, it may comprise SEQ ID NO: 30; alternatively, it may comprise SEQ ID NO: 31; alternatively, it may comprise SEQ ID NO: 32; alternatively, it may comprise or SEQ ID NO: 33.

[0009] The invention also provides methods of inducing a protective immune response against RSV infection in a subject. The methods comprise administering to the subject an immunologically effective amount of the immunogenic composition of the invention. Usually, the immunogenic composition is administered intramuscularly. The subject will typically be an infant, a pregnant mother, female of child bearing age, or an elderly person.

[0010] The invention further provides methods of detecting an immune response against RSV in a subject. The methods comprise contacting a biological sample from the subject with an RSV oligopeptide selected from SEQ ID NO: 3-33 and detecting an immune response. The immune response can be detected by detecting binding of antibodies in the biological sample to the RSV oligopeptide or by detecting a cellular immune response.

DEFINITIONS

[0011] The terms "adjuvant" and "immune stimulant" are used interchangeably herein, and are defined as one or more substances that cause stimulation of the immune system. The stimulation will augment or modify the immune response, *e.g.*, by intensifying or broadening the specificity of either or both the antibody and the cellular immune response to the RSV polypeptide. In this context, an adjuvant or immune stimulant is used in the immunogenic compositions of the invention to enhance an immune response to RSV proteins. In the case where the RSV polypeptide is expressed in a viral vector, the viral vector can act as the immune stimulant.

[0012] The term "expression vector" refers to a linear or circular DNA molecule that comprises a segment encoding a polypeptide of interest (*e.g.*, an RSV protein of the invention) operably linked to additional segments that provide for its transcription. Such additional segments include promoter and terminator sequences, and may also include one or more origins of replication, one or more selectable markers, an enhancer, a polyadenylation signal, and the like. Expression vectors are generally derived from bacterial or viral DNA, and may contain elements of both.

[0013] The terms "identical" or percent "identity," in the context of two or more nucleic acids or polypeptide sequences, (*e.g.*, RSV proteins/peptides of the invention and polynucleotides that encode them) refer to two or more sequences or subsequences that are the same or have a specified percentage of amino acid residues or nucleotides that are the same, when compared and

aligned for maximum correspondence, as measured using one of the following sequence comparison algorithms or by visual inspection.

[0014] The phrase "substantially identical," in the context of two nucleic acids or polypeptides of the invention, refers to two or more sequences or subsequences that have at least 50%, more preferably 65%, even more preferably 70%, still more preferably 75%, even more preferably 80%, and most preferably 90-95% nucleotide or amino acid residue identity, when compared and aligned for maximum correspondence, as measured using one of the following sequence comparison algorithms or by visual inspection. Preferably, the substantial identity exists over a region of the sequences that is at least about 10 or 50 residues in length, more preferably over a region of at least about 100 residues, and most preferably the sequences are substantially identical over at least about 150 residues. In a most preferred embodiment, the sequences are substantially identical over the entire length of the coding regions.

[0015] For sequence comparison, typically one sequence acts as a reference sequence (*e.g.*, SEQ ID NO: 2), to which test sequences are compared. When using a sequence comparison algorithm, test and reference sequences are input into a computer, subsequence coordinates are designated, if necessary, and sequence algorithm program parameters are designated. The sequence comparison algorithm then calculates the percent sequence identity for the test sequence(s) relative to the reference sequence, based on the designated program parameters.

[0016] Optimal alignment of sequences for comparison can be conducted, *e.g.*, by the local homology algorithm of Smith & Waterman, *Adv. Appl. Math.* 2:482 (1981), by the homology alignment algorithm of Needleman & Wunsch, *J. Mol. Biol.* 48:443 (1970), by the search for similarity method of Pearson & Lipman, *Proc. Nat'l. Acad. Sci. USA* 85:2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, WI), or by visual inspection (*see generally*, *Current Protocols in Molecular Biology*, F.M. Ausubel *et al.*, eds., Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc., (1995 Supplement) (Ausubel)).

[0017] Examples of algorithms that are suitable for determining percent sequence identity and sequence similarity are the BLAST and BLAST 2.0 algorithms, which are described in Altschul *et al.* (1990) *J. Mol. Biol.* 215: 403-410 and Altschul *et al.* (1977) *Nucleic Acids Res.* 25: 3389-

3402, respectively. Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information (<http://www.ncbi.nlm.nih.gov/>). This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold (Altschul *et al*, *supra*). These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are then extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always > 0) and N (penalty score for mismatching residues; always < 0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T, and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) of 10, M=5, N=-4, and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a wordlength (W) of 3, an expectation (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff & Henikoff, *Proc. Natl. Acad. Sci. USA* 89:10915 (1989)).

[0018] In addition to calculating percent sequence identity, the BLAST algorithm also performs a statistical analysis of the similarity between two sequences (see, e.g., Karlin & Altschul, *Proc. Nat'l. Acad. Sci. USA* 90:5873-5787 (1993)). One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a nucleic acid is considered similar to a reference sequence if the smallest sum probability in a comparison of the test nucleic acid to the reference nucleic acid is less than about 0.1, more preferably less than about 0.01, and most preferably less than about 0.001.

[0019] A further indication that two nucleic acid sequences or polypeptides of the invention are substantially identical is that the polypeptide encoded by the first nucleic acid is immunologically cross reactive with the polypeptide encoded by the second nucleic acid, as described below. Thus, a polypeptide is typically substantially identical to a second polypeptide, for example, where the two peptides differ only by conservative substitutions. Another indication that two nucleic acid sequences are substantially identical is that the two molecules hybridize to each other under stringent conditions, as described below.

[0020] "Operably linked" indicates that two or more DNA segments are joined together such that they function in concert for their intended purposes. For example, coding sequences are operably linked to promoter in the correct reading frame such that transcription initiates in the promoter and proceeds through the coding segment(s) to the terminator.

[0021] A "polynucleotide" is a single- or double-stranded polymer of deoxyribonucleotide or ribonucleotide bases typically read from the 5' to the 3' end. Polynucleotides include RNA and DNA, and may be isolated from natural sources, synthesized in vitro, or prepared from a combination of natural and synthetic molecules. When the term is applied to double-stranded molecules it is used to denote overall length and will be understood to be equivalent to the term "base pairs".

[0022] A "polypeptide" is a polymer of amino acid residues joined by peptide bonds, whether produced naturally or synthetically. Polypeptides of less than about 75 amino acid residues are also referred to here as peptides or oligopeptides.

[0023] The term "promoter" is used herein for its art-recognized meaning to denote a portion of a gene containing DNA sequences that provide for the binding of RNA polymerase and initiation of transcription of an operably linked coding sequence. Promoter sequences are typically found in the 5' non-coding regions of genes.

BRIEF DESCRIPTION OF THE DRAWINGS

[0024] Figure 1 shows purification of recombinant G protein from *E. coli* and 293 cells. (A) Schematic representation of the RSV G protein. RSV G protein purified from *E. coli* (REG) inclusion bodies is lacking the cytoplasmic and transmembrane domains (CT-TM) and is not

glycosylated, while the RSV G protein secreted from 293-Flp-In cells (RMG) is glycosylated. N-linked glycosylation sites, as predicted by NetNGlyc 1.0 software, are represented above the bar and predicted O-linked glycosylation sites, as predicted by NetOGlyc 4.0 software, are represented below the bar in the full length RSV G and RMG proteins. REG was purified from *E. coli* inclusion bodies as described in materials and methods. RMG was purified from the clarified supernatant of 293-Flp-In cells stably expressing the RSV G protein. Both REG (B) and RMG (C) were purified through a Ni-NTA column and the final products were detected as a single band in SDS PAGE under reducing conditions (D) SPR interaction profile of REG and RMG binding to the protective MAb 131-2G that targets the central conserved domain of RSV G. (E-F) Superdex S-200 gel filtration chromatography of RSV G protein produced in bacterial system (REG) (E) and mammalian system (RMG) (F). The panels present superimposed elution profiles of purified RSV G proteins (red lines) overlaid with calibration standards (grey lines).

[0025] Figure 2 is a schematic representation of mice immunization and challenge schedule. BALB/c mice were immunized i.m. with 5 µg of REG, RMG or F proteins of RSV-A2 virus strain with Emulsigen adjuvant, or with PBS control, on days 0 and 20. Ten days after the second immunization, blood was collected from the tail veins. 14 days after the second immunization, mice were challenged intranasally with 10⁶ pfu of either RSV-A2 or RSV-B1 viral strains (6-11 mice per group). Mice were sacrificed on days 2 or 4 post-challenge wherein lungs and blood were collected.

[0026] Figure 3 shows a neutralizing antibody response following RSV G (REG, RMG) and F protein immunization. Serum from individual mice collected on day 10 post second immunization was tested for neutralization in a plaque reduction neutralization test (PRNT) against the homologous RSV-A2 strain (A) or heterologous RSV-B1 strain (B). Neutralizing antibody titers represent 50% inhibition of plaque numbers. The average for each group is represented with a horizontal line. Pre-vaccination (day 0) serum samples from all the animals tested negative in PRNT (data not shown). 7/11 animals vaccinated with unglycosylated G ectodomain (REG) generated good neutralization titers against the RSV A2 strain. No animals vaccinated with the glycosylated G ectodomain (RMG) generated neutralizing antibodies against RSV A2. Statistical significance was tested with one-way ANOVA and Bonferroni multiple comparisons tests. *** p < 0.0001, ** p < 0.001, *p < 0.05.

[0027] Figure 4 shows surface plasmon resonance analysis of post-vaccination sera to REG, RMG and different antigenic regions within RSV G. (A-B) The same individual post-vaccination mice sera from Fig. 3 were tested for total antibody binding to the REG protein (A) or RMG protein (B) by Surface Plasmon Resonance (SPR). (C) Antigenic peptides representing amino acids 66-90, 90-110, 148-178, 169-207 and 236-263 of the RSV G protein were chemically synthesized and tested for binding to serum antibodies from REG or RMG immunized mice in a real time SPR kinetics experiment. Total antibody binding is represented as SPR resonance units. Sera from REG vaccinated mice showed >10 fold higher antibody binding to either REG protein (panel A) or to the glycosylated (RMG) G protein (panel B). peptide scanning showed more diverse repertoire for the REG vaccinated animals (panel C). Statistical significance was tested with one-way ANOVA and Bonferroni multiple comparisons tests. *** $p < 0.0001$, ** $p < 0.001$, * $p < 0.05$.

[0028] Figure 5 shows that immunization with REG protects against homologous and heterologous RSV challenge. Mice immunized twice with PBS, REG, RMG, or F protein were challenged intranasally (i.n.) with 1×10^6 pfu of RSV-A2 (A) or RSV-B1 (B) 14 days after the second immunization. Four days post virus challenge, mice lungs were collected and homogenized as described in materials and methods, and lung viral loads were determined by plaque assay. Statistical significance was tested with one-way ANOVA and Bonferroni multiple comparisons tests, * $p < 0.05$. (C) Relationship between the total anti-G antibody binding in post-vaccination sera (in Fig. 4A) and lung viral load at 4 days post-challenge with RSV-A2 (in Fig. 5A) was analyzed and Spearman's correlation was calculated. Total antibody binding to REG is represented in resonance units and lung viral loads are represented as pfu per gram lung weight. *p-values* less than 0.05 were considered significant. Most animals that were immunized with REG effectively eliminated virus from the lungs after challenge with homologous (panel A) or heterologous (panel B) viruses. Most of the RMG immunized animals did not clear virus.

[0029] Figure 6 shows histopathology analysis of lungs from REG and RMG immunized mice after virus challenge: REG immunization protected from lung disease, while RMG immunization resulted in perivascularitis and lung cellular infiltrates after challenge (enhanced disease/pathology). Lungs collected 2 days post-challenge with RSV-A2 or RSV-B1 were stained with hematoxylin and eosin (H-E) and scored for inflammation in bronchioles, near veins

(vascular), and alveoli. (A) Lung histology scores represent the average of 2 slides per mouse and 3 mice per group. Neutralizing antibody titers represent 50% inhibition of plaque numbers as measured by PRNT against the respective RSV strains for the 3 mice per group. 0, no lesions; 1, minimal; 2, mild; 3, modest; 4, severe. (B) 400X magnification images of H-E stained sectioned lungs

[0030] Figure 7 shows RMG (but not REG) immunization induces high levels of Th2 cytokines and chemokines following RSV i.n challenge. (A-H) Cytokines in lung homogenates from day 2 post challenge were measured in a Bio-Plex Pro™ Mouse Cytokine assay. Values represent the concentration in pg/mL of 3 mice per group. The box extends from the 25th to the 75th percentile and the error bars represent the lowest and highest values. Values below the assay's limit of detection were assigned a number according to the minimum detection limit of the cytokine. The mean for each group are represented. (I) Ratio of the observed concentration of Th2 vs Th1 cytokines as measured by IL4/IFN- γ . Statistical significance was analyzed using a one-way ANOVA and Bonferroni multiple comparisons tests. *** $p < 0.0001$, ** $p < 0.001$, * $p < 0.05$. High Th2 chemokine levels in the lung were associated with eosinophilic infiltration and severe lung pathology after challenge with RSV.

[0031] Figure 8 shows an alignment of the full-length G protein amino acid sequence from four different RSV strains: A2 (SEQ ID NO: 34), A-Long (SEQ ID NO: 35), B-18537 (SEQ ID NO: 36), and B1 (SEQ ID NO: 37).

DETAILED DESCRIPTION

[0032] The present invention provides unglycosylated RSV polypeptides that are useful as immunogens to induce protective immune response against RSV infection. In a typical embodiment, the RSV polypeptide comprises a RSV G protein ectodomain (corresponding to amino acids 67-298 of SEQ ID NO: 34) produced as an unglycosylated protein in a prokaryotic expression system. The present invention also provides shorter RSV polypeptides (*i.e.*, oligopeptides) that can be used in immunogenic compositions of the invention to induce protective immune responses.

RSV POLYPEPTIDES

[0033] The genome of RSV is a single, negative-sense strand of RNA of 15.2 kilobases that is transcribed by the viral polymerase into 10 mRNAs by a sequential stop-start mechanism that

initiates at a single viral promoter at the 3' end of the genome. Each mRNA encodes a single major protein, with the exception of the M2 mRNA, which has two overlapping open reading frames that encode two separate proteins. The 11 RSV proteins are: the RNA-binding nucleocapsid protein (N), the phosphoprotein (P), the large polymerase protein (L), the attachment glycoprotein (G), the fusion protein (F), the small hydrophobic (SH) surface glycoprotein, the internal matrix protein (M), the two nonstructural proteins NS1 and NS2, and the M2-1 and M2-2 proteins encoded by the M2 mRNA.

[0034] One of skill will recognize that any RSV protein could be used as an immunogen in the immunogenic compositions of the invention. In a typical embodiment, RSV polypeptides of the invention will be derived from either the F protein or the G protein.

[0035] The RSV G protein comprises a hydrophobic region near the N-terminus that serves as a membrane anchor, the transmembrane domain, and a C-terminal portion of the molecule exposed on the exterior side of the membrane, the ectodomain (*see* Figure 1). Those of skill can readily identify the domains of a G protein using standard sequence alignments to identify corresponding residues in G proteins isolated from different RSV strains. Such an alignment is provided in Figure 8. For example, the ectodomain of the RSV-A2 strain, which is exemplified below, extends from residues 67-298 of SEQ ID NO: 34). Using standard sequence alignment techniques, one of skill can readily determine the corresponding residues in other RSV strains. For example, to determine which amino acid of a first protein “corresponds” to the position of an amino acid in a second protein, the amino acid sequences of the two proteins are optimally aligned (e.g., using a BLAST algorithm).

[0036] The RSV polypeptides of the invention are typically fragments of a native RSV protein, such as an F protein or a G protein. In a typical embodiment, the RSV polypeptide is from about 10 amino acids to about 75 amino acids in length (also be referred to as RSV oligopeptides).

The RSV oligopeptides may also be less than about 50 amino acids in length. Exemplary RSV oligopeptides of the invention are provided in SEQ ID NOs: 3-33. Peptides of particular interest include the following peptides.

[0037] RSV- F Peptide Sequences:

101-157 PATNNRARRELPRFMNYTLNNAKKTNTLSKKRKRRFLGFLLGVG
SAIASGVAVSKV (SEQ ID NO: 5)

195-240 LKNYIDKQLLPVINKQSCSISNIETVIEFQQKNNRLLLEITREFSVN
(SEQ ID NO: 12)

425-450 SNKNRGIKTFSNGCDYVSNKGVDTV (SEQ ID NO: 19)

443-461 SNKGVDTVSVGNTLYYVVK (SEQ ID NO: 20)

[0038] RSV-G Peptide Sequences:

129-152 TTVKTKNTTTTQTQPSKPTTKQRQ (SEQ ID NO: 28)

169-207 NFVPCSICSNPTCWAICKRIPNKKPGKTTTKPTKKPT (SEQ ID NO: 30)

263-298 ETFHSTSS EGNPSPSQVS TTSEYPSQPS SPPNTPRQ (SEQ ID NO: 33)

[0039] The RSV polypeptides of the invention may be multimeric and comprise more than one of the RSV oligopeptides of the invention in a single polypeptide chain. Such multimeric polypeptides may, for example, comprise multiple copies of a single oligopeptide or may comprise different oligopeptide sequences. The multimeric RSV polypeptides may also comprise oligopeptide sequences from other pathogens so that immune protection against multiple pathogens is provided in a single composition. The multimeric RSV polypeptides will typically be from about 50 to about 300 amino acids in length.

[0040] In other embodiments, the RSV polypeptides of the invention are about 75 to about 250 amino acids in length and are derived from the ectodomain of an RSV G or F protein. More typically, the RSV polypeptide is from about 150 to about 230 amino acids in length. In these embodiments, the RSV polypeptide will consist essentially of the ectodomain of a G protein (*e.g.*, those residues corresponding to residues 67-298 of the RSV-A2 strain G protein, SEQ ID NO: 34 or 67-299 of the RSV-B1 strain G protein, SEQ ID NO: 37). Alternatively, the RSV polypeptide of the invention may consist essentially of the ectodomain of an F protein (*e.g.*, those residues corresponding to residues 23-524 of the RSV-A2 strain F protein, SEQ ID NO: 39). In such an embodiment, the RSV polypeptide consisting essentially of the ectodomain of a G or F protein may comprise, for example, an epitope tag or other short sequence to facilitate

purification of the polypeptide (*e.g.*, a polyhistidine tag). An RSV polypeptide consisting essentially of the ectodomain of a G or F protein may also comprise less than about 10 amino acids, usually less than about 5 amino acids, from the G or F protein transmembrane domain.

[0041] The RSV polypeptide is typically unglycosylated. An “unglycosylated RSV polypeptide” is one that lacks at least about 95% of the sugar moieties that occur on the naturally occurring RSV protein. Usually, the unglycosylated RSV polypeptide lacks at least about 99% of the sugar moieties. In the most embodiments, the RSV polypeptide is completely unglycosylated. In a typical embodiment, the unglycosylated RSV polypeptide is expressed in a prokaryotic organism (*e.g.* *E. coli*) that does not comprise the cellular machinery to glycosylate proteins. The unglycosylated RSV polypeptide may also be chemically synthesized or expressed in a glycosylation-deficient mammalian cell. Alternatively, appropriate enzymes may be used *in vitro* to remove all or most of the sugar moieties from the polypeptide.

[0042] The RSV polypeptides of the invention can be used in immunogenic compositions, as well a variety of other uses. For example, the polypeptides can be used in serological assays for diagnosis of RSV, in analysis of immune sera from pre-clinical and clinical trials of RSV vaccines, and in mapping of monoclonal and polyclonal antibodies against different RSV proteins. The oligopeptides are also useful in studies designed to understand viral pathogenesis and therefore useful in designing antiviral drugs or treatment. For example, the peptides can be used to study viral protein-protein, viral RNA-protein and viral-host protein interactions.

[0043] The RSV polypeptides of the invention can be made using standard methods well known to those of skill in the art. For example, shorter polypeptides (*i.e.*, oligopeptides) can be made synthetically. For longer polypeptides, recombinant expression can be conveniently used. Recombinant expression in a variety of host cells, including *E. coli*, or other prokaryotic hosts which lack the cellular machinery for glycosylation of proteins is well known in the art.

[0044] Polynucleotides encoding RSV polypeptides, recombinant expression vectors, and host cells containing the recombinant expression vectors, as well as methods of making such vectors and host cells by recombinant methods are useful to produce the polypeptides of the invention. These methods are well known to those of skill in the art.

[0045] The polynucleotides encoding RSV polypeptides may be synthesized or prepared by techniques well known in the art. Nucleotide sequences encoding the RSV polypeptides of the disclosure may be synthesized, and/or cloned, and expressed according to techniques well known to those of ordinary skill in the art. In some embodiments, the polynucleotide sequences will be codon optimized for a particular recipient using standard methodologies. For example, a DNA construct encoding a RSV polypeptide can be codon optimized for expression in prokaryotic hosts, e.g., bacteria.

[0046] Examples of useful bacteria include, but are not limited to, *Escherichia*, *Enterobacter*, *Azotobacter*, *Erwinia*, *Bacillus*, *Pseudomonas*, *Klebsiella*, *Proteus*, *Salmonella*, *Serratia*, *Shigella*, *Rhizobia*, *Vitreoscilla*, and *Paracoccus*. The nucleic acid encoding the RSV polypeptide is operably linked to appropriate expression control sequences for each host. For *E. coli* this includes a promoter such as the T7, trp, or lambda promoters, a ribosome binding site and preferably a transcription termination signal. For eukaryotic cells, the control sequences will include a promoter and preferably an enhancer derived from immunoglobulin genes, SV40, cytomegalovirus, *etc.*, and a polyadenylation sequence, and may include splice donor and acceptor sequences.

[0047] The RSV polypeptides may also be expressed in other cells that normally glycosylate proteins, such as mammalian, insect, plants or yeast cells. In some embodiments, the polypeptide is deglycosylated after expression. Removal of carbohydrate moieties may be accomplished chemically or enzymatically according to well-known techniques. Chemical deglycosylation typically involves contacting the polypeptide with trifluoromethanesulfonic acid, or an equivalent compound. This treatment results in the cleavage of most or all sugars except the linking sugar (N-acetylglucosamine or N-acetylgalactosamine), while leaving the polypeptide intact. Enzymatic cleavage of carbohydrate moieties on proteins can be achieved by the use of a variety of endo- and exo-glycosidases. Kits for enzymatic removal of carbohydrates are commercially available.

[0048] Once expressed, the recombinant RSV polypeptides can be purified according to standard procedures of the art, including ammonium sulfate precipitation, affinity columns, column chromatography, gel electrophoresis and the like. In a typical embodiment, the recombinantly produced RSV polypeptide is expressed as a fusion protein that has a "tag" at one

end which facilitates purification of the polypeptide. Suitable tags include epitope tags, which are amino acid sequences that are specifically recognized by an antibody. Epitope tags are generally incorporated into recombinantly expressed proteins to enable the use of a readily available antibody to detect or isolate the protein. Other suitable tags are known to those of skill in the art, and include, for example, an affinity tag such as a polyhistidine tag which will bind to metal ions such as nickel or cobalt ions.

IMMUNOGENIC COMPOSITIONS

[0049] The RSV polypeptides and oligopeptides of the invention may be formulated as a vaccine (also referred to as an “immunogenic composition”) according to methods well known in the art. Such compositions will typically include immune stimulants to enhance immune responses. The optimal ratios of each component in the formulation may be determined by techniques well known to those skilled in the art. The immunogenic compositions may be administered to a patient or subject according to standard techniques well-known to those of skill in the art. The subject will typically be a human, or non-human animal (*e.g.*, cattle, horses, pigs, dogs, and cats). The human patient or subject may be an adult and child. In a typical case the subject will be an infant (*e.g.*, a newborn), a young child, a pregnant mother, a woman of child bearing age, or the elderly.

[0050] The immunogenic compositions may also comprise one or more of the RSV oligopeptides shown in SEQ ID NOs: 3-33 or a multimeric RSV polypeptide comprising one or more the oligopeptide sequences. The oligopeptides or multimeric polypeptide may be unglycosylated or glycosylated. For example, the composition may comprise only one of the following oligopeptides: SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, SEQ ID NO: 30, SEQ ID NO: 31, SEQ ID NO: 32, or SEQ ID NO: 33. In some embodiments, the composition will further comprise additional oligopeptides selected from the same list of peptides. Alternatively, the oligopeptides may be included in a single, multimeric RSV polypeptide.

[0051] The preparation and use of immunogenic compositions are well known to those of skill in the art. Liquid pharmaceutical compositions generally include a liquid carrier such as water, petroleum, animal or vegetable oils, mineral oil or synthetic oil. Physiological saline solution, dextrose or other saccharide solution or glycols such as ethylene glycol, propylene glycol or polyethylene glycol may be included.

[0052] Polynucleotides encoding the RSV polypeptides of the invention can also be administered to the subject. Typically, an expression cassette suitable for driving expression in human cells is prepared. This approach is described, for instance, in Wolff (1990) Science 247:1465-1468; U.S. Patent Nos. 5,580,859 and 5,589,466.

[0053] The RSV polypeptides (*e.g.*, multimeric RSV polypeptides) may also be expressed in replication deficient or competent viral vectors that comprise nucleic acids encoding them. Such viral vectors include, for example, adenoviral vectors, vaccinia virus vectors, avipox vector such as fowlpox or canarypox, herpes virus vectors, a vesicular stomatitis virus vectors, or alphavirus vectors. One of skill will recognize that the immunogenic compositions of the invention may comprise multiple antigens and vectors. In such embodiments, the viral vector can act as an immune stimulant. Thus, the immune stimulant of the immunogenic compositions of the invention will be the vector itself and addition of further immune stimulants may not be necessary.

[0054] The RSV polypeptides (*e.g.*, multimeric RSV polypeptides and RSV oligopeptides) may also be administered in combination with nanoparticles (*i.e.*, particulate material with size 1–1000 nm). A great variety of materials can be used to prepare nanoparticles useful in the immunogenic compositions of the invention. For example, synthetic polymers such as poly(d,l-lactide-co-glycolide) (PLG), poly(d,l-lactic-coglycolic acid)(PLGA), poly(g-glutamic acid) (g-PGA), poly(ethylene glycol) (PEG) and polystyrene can be conveniently used. Polymeric nanoparticles entrap the RSV polypeptides for delivery to certain cells or sustain antigen release as a result of their slow biodegradation rate. Natural polymers based on polysaccharide may also be used. Examples include pullulan, alginate, inulin and chitosan. Inorganic nanoparticles useful in the invention include gold nanoparticles, which can be surface-modified with carbohydrates, carbon nanoparticles or silica-based nanoparticles.

[0055] The compositions are suitable for single administrations or a series of administrations. When given as a series, inoculations subsequent to the initial administration are given to boost the immune response and are typically referred to as booster inoculations. The compositions of the invention can be used as a boosting composition primed by antigen using any of a variety of different priming compositions, or as the priming composition. Thus, one aspect of the present invention provides a method of inducing and/or boosting an immune response to an antigen in an individual.

[0056] The timing of the administration of boosting compositions is well within the skill in the art. Boosting composition are generally administered weeks or months after administration of the priming composition, for example, about 2-3 weeks or 4 weeks, or 8 weeks, or 16 weeks, or 20 weeks, or 24 weeks, or 28 weeks, or 32 weeks.

[0057] The compositions of the invention may comprise other RSV immunogens or the priming or boosting inoculations may comprise other immunogens. The other immunogens used in combination with the RSV polypeptides of the invention are not critical to the invention.

Examples of such immunogens include those from Influenza, Hepatitis and other childhood infectious diseases.

[0058] The immunogens in the respective priming and boosting compositions (however many boosting compositions are employed) need not be identical, but should share epitopes. The immunogen may correspond to a complete antigen in a target pathogen or cell, or a fragment thereof. Peptide epitopes or artificial strings of epitopes may be employed, more efficiently cutting out unnecessary protein sequence in the antigen and encoding sequence in the vector or vectors. For example, one or more additional epitopes may be included, for instance epitopes which are recognized by T helper cells, especially epitopes recognized in individuals of different HLA types.

[0059] As noted above, the immunogenic compositions of the invention may comprise an immune stimulant (also referred to as an adjuvant). In those embodiments in which the RSV polypeptide is expressed in a viral vector, the immune stimulant is typically the viral vector, itself. Adjuvants suitable for co-administration in accordance with the present invention should be ones that are potentially safe, well tolerated and effective in people including QS-21, Detox-PC, MPL-

SE, MoGM-CSF, TiterMax-G, CRL- 1005, GERBU, TERamide, PSC97B, Adjumer, PG-026, GSK-I, GcMAF, B-aethine, MPC-026, Adjuvax, CpG ODN, Betafectin, Alum, and MF59.

[0060] Other immune stimulants that may be administered include lectins, growth factors, cytokines and lymphokines such as alpha-interferon, gamma interferon, platelet derived growth factor (PDGF), granulocyte-colony stimulating factor (GCSF), granulocyte macrophage colony stimulating factor (GMCSF), tumor necrosis factor (TNF), epidermal growth factor (EGF), IL-I, IL-2, IL-4, IL-6, IL-8, IL-10, and IL-12 or encoding nucleic acids therefore.

[0061] As noted above, the compositions of the invention may comprise a pharmaceutically acceptable excipient, carrier, buffer, stabilizer or other materials including particles (*e.g.*, nanoparticles) well known to those skilled in the art. Such materials should be non-toxic and should not interfere with the efficacy of the active ingredient. The precise nature of the carrier or other material may depend on the route of administration, *e.g.*, oral, intravenous, cutaneous or subcutaneous, intramucosal (*e.g.*, gut), intranasal, intramuscular, or intraperitoneal routes. Administration is typically intradermal, *e.g.*, subcutaneous or intramuscular.

[0062] Intramuscular administration of the immunogenic compositions may be achieved by using a needle to inject a suspension of the RSV polypeptide or nucleic acid encoding it. An alternative is the use of a needleless injection device to administer the composition (using, *e.g.*, Biojector(TM)) or a freeze-dried powder containing the vaccine.

[0063] For intravenous, cutaneous or subcutaneous injection, or injection at the site of affliction, the RSV polypeptide will be in the form of a parenterally acceptable aqueous solution which is pyrogen-free and has suitable pH, isotonicity and stability. Those of skill in the art are well able to prepare suitable solutions using, for example, isotonic vehicles such as Sodium Chloride Injection, Ringer's Injection, Lactated Ringer's Injection. Preservatives, stabilizers, buffers, antioxidants and/or other additives may be included, as required. A slow-release formulation may also be employed

[0064] Typically, administration will have a prophylactic aim to generate an immune response against a RSV antigen before infection or development of symptoms. Diseases and disorders that may be treated or prevented in accordance with the present invention include those in which

an immune response may play a protective or therapeutic role. In other embodiments, the RSV polypeptides can be administered for post-exposure prophylactics.

[0065] The immunogenic compositions containing the RSV polypeptide or polynucleotides encoding them are administered to a subject, giving rise to an anti-RSV immune response in the subject. An amount of a composition sufficient to induce a detectable immune response is defined to be an "immunologically effective dose." As shown below, the immunogenic compositions of the invention induce an antibody response, as well as a CD8+ T cell immune response. In a typical embodiment the immune response is a protective immune response.

[0066] The actual amount administered, and rate and time-course of administration, will depend on the nature and severity of what is being treated. Prescription of treatment, e.g., decisions on dosage etc., is within the responsibility of general practitioners and other medical doctors, or in a veterinary context a veterinarian, and typically takes account of the disorder to be treated, the condition of the individual patient, the site of delivery, the method of administration and other factors known to practitioners. Examples of the techniques and protocols mentioned above can be found in Remington's Pharmaceutical Sciences, 16th edition, Osol, A. ed., 1980.

[0067] In one exemplary regimen, the RSV polypeptide is administered (*e.g.*, intramuscularly) at a dose of 10 micrograms to 1 milligram/injection. A boost can be administered up to 4 weeks later. The composition may, if desired, be presented in a kit, pack or dispenser, which may contain one or more unit dosage forms containing the active ingredient. The kit, for example, may comprise metal or plastic foil, such as a blister pack. The kit, pack, or dispenser may be accompanied by instructions for administration.

[0068] The compositions of the invention may be administered alone or in combination with other treatments, either simultaneously or sequentially dependent upon the condition to be treated.

DETECTION OF IMMUNE RESPONSES

[0069] The RSV polypeptides (particularly oligopeptides) of the invention are useful in the detection of RSV infection. Methods for detecting humoral or cellular immune responses in biological samples are well known in the art. As used herein, a "biological sample" is any lymphocyte or antibody-containing sample obtained from a patient. For example, the sample can

be whole blood, sputum, serum, plasma, saliva cerebrospinal fluid or urine. The polypeptides of the invention are used in an assay, for example as described below, to determine the presence or absence of antibodies or immune cells (*e.g.*, lymphocytes, CD4⁺ T-cells, CD8⁺ T-cells, B-cells, NK cells, mast cells, antigen presenting cells) or cytokines induced by RSV antigens. The presence of such antibodies or immune cells indicates previous sensitization to RSV antigens which may be indicative of RSV infection.

[0070] There are a variety of assay formats known to those of ordinary skill in the art for using one or more oligopeptides to detect antibodies in a sample. For example, the assay may involve the use of an RSV polypeptide immobilized on a solid support to bind to and remove the antibodies from the sample. The assay may also involve the capture of the RSV polypeptide by an antibody or binding protein (*e.g.*, protein A, protein G, protein A/G, avidin, streptavidin, etc.) bound to a solid support. The assay may involve the labeling of the RSV polypeptide with small molecules such as biotin, enzymes, or fluorophores. Alternatively, antibodies from patients could be bound to a solid phase using binding proteins or type specific antibodies (*e.g.*, anti-human IgM or IgG antibodies), followed by treatment with the labeled or unlabeled RSV polypeptides, and finally detecting the bound RSV polypeptide with a conjugated antibody and/or a binding protein (*e.g.*, avidin-FITC, avidin-peroxidase, etc.). The bound antibody may then be detected using a detection reagent that contains a reporter group. Suitable detection reagents include antibodies that bind to the solid phase bound RSV polypeptide or antibody/polypeptide complex or to free polypeptide labeled with a reporter group (*e.g.*, in a semi-competitive assay). Alternatively, a competitive assay may be utilized, in which an antibody that binds to the polypeptide is labeled with a reporter group and allowed to bind to the immobilized antigen after incubation of the antigen with the sample. The extent to which components of the sample inhibit the binding of the labeled antibody to the polypeptide is indicative of the reactivity of the sample with the immobilized polypeptide. In certain embodiments, the assay is an enzyme linked immunosorbent assay (ELISA), chemiluminescent assay, or fluorescence assay.

[0071] Methods for detecting cellular immune responses are also well known. In such assays antigen presenting cells, immune cells (*e.g.*, CD8⁺ T-cells, CD4⁺ T-cells, B-cells, etc.) and RSV polypeptides of the invention are incubated under appropriate conditions to induce proliferation

or activation of immune effector cells (e.g., CD8⁺ T-cells, CD4⁺ T-cells, B-cells, NK and iNKT cells, etc) specific to RSV. The antigen-specific responses of induced immune effector cells (e.g., helper T-cells, CTLs, B-cells, NK, NKT, etc.) can be detected by a number of techniques (e.g., by a ⁵¹Cr release assay, ³H-thymidine incorporation, CFSE and other FACS-analysis proliferation assays, cytokine intracellular staining and FACS analysis, ELISPOT, detection of secreted cytokines such as INF-γ).

EXAMPLES

The following examples are offered to illustrate, but not to limit the claimed invention.

EXAMPLE 1

[0072] We have performed a large scale analysis of epitopes recognized by post-RSV vaccination sera that identified multiple protective vaccine targets against RSV. These peptide epitopes were chemically synthesized, tested for antibody binding. This was carried out using a “Whole Genome Phage Display Library” (GFPDL) expressing all the possible peptides from the RSV G and F protein and screening with polyclonal immune sera from immunized rabbits immunized with unglycosylated F and G proteins of RSV. Each phage in the library expresses on its surface a fragment of the RSV membrane proteins in fusion with the phage gIII protein. The antibody bound phages were amplified and the inserts are sequenced. Unlike the native RSV G and F proteins, which is heavily glycosylated, the phage library expresses only unglycosylated peptides. The peptides identified in the screen were then used to immunize rabbits using standard techniques. Following immunization these animal sera were tested in multiple RSV neutralization assays. The results, shown in Table 1, show that most of these peptides generated neutralizing antibodies against RSV. Those peptides that generated neutralizing antibodies are shown in SEQ ID NOs: 3-33. These peptides can be an important target for development of peptide based RSV diagnosis, treatment and vaccines against RSV infection and disease.

TABLE 1

F-peptides		Rabbit Sera
A.A.	Sequence	Neutralization titers
1-34	MELLILKANAITTILTAVTFCFASGQNITEEFYQ (SEQ ID NO: 3)	24
101-121	PATNNRRARRELPRFMNYTLNN (SEQ ID NO: 7)	13
174-203	TNKAVVSLSNGVSVLTSKVLDLKNYIDKQL (SEQ ID NO: 9)	13
195-240	LKNYIDKQLLPVIVNKQSCSISNIETVIEFQQKNNRLLLEITREFSVN (SEQ ID NO: 12)	17
234-287	TREFSVNAGVTTTPVSTYMLTNSELLSLINDMPITNDQKKLMSNNVQIVRQQSY S (SEQ ID NO: 14)	14
250-273	YMLTNSELLSLINDMPITNDQKKL (SEQ ID NO: 15)	12
425-450	SNKNRGIKTFSGCDYVSNKGVDTV (SEQ ID NO: 19)	59
443-461	SNKGVDTVSVGNTLYYVVK (SEQ ID NO: 20)	96
471-493	GEPIINFYDPLVFPSDEFDASIS (SEQ ID NO: 22)	18
552-572	ARSTPVTLSKQDLSGINNIAF (SEQ ID NO: 24)	13
G-peptides		
A.A.	Sequence	
33-61	LNLKSVAQITLSILAMIISTSLIIAAIIF (SEQ ID NO: 25)	
66-90	NHKVTFPTTAIIQDATSQIKNTTPTY (SEQ ID NO: 26)	24
90-110	YLTQNPQLGISPSNPSEITSQ (SEQ ID NO: 27)	16
129-152	TTVKTKNTTTTQTQPSKPTTKQRQ (SEQ ID NO: 28)	28
148-178	TTKQRQNKPPSKPNNDFFHFVFNFPVPCSICS (SEQ ID NO: 29)	47
169-207	NFVPCSICSNNPCTCWAICKRIPNKKPGKKTITTKPTKKPT 30)	683
236-263	INTTKTNIITTLTSTNTTGNPELTSQME (SEQ ID NO: 31)	19

5 EXAMPLE 2

[0073] New efforts are under way to develop vaccines against RSV that will provide protective immunity without the potential for disease enhancement. The G attachment protein represents an important candidate for inclusion in an effective RSV vaccine. In the current study, we evaluated the safety and protective efficacy of RSV A2 recombinant unglycosylated G protein ectodomain produced in *E. coli* (REG) vs. glycosylated G produced in mammalian cells (RMG) in a mouse RSV challenge model (A2 and B1 strains). The unglycosylated G generated high protective immunity and no lung pathology even in animals that lacked anti-RSV neutralizing antibodies prior to RSV challenge. Control of viral loads correlated with antibody binding to the

G protein. In contrast, the glycosylated G protein provided poor protection and enhanced lung pathology after RSV challenge. Therefore, bacterially produced unglycosylated G protein presents an economical approach for protective vaccine against RSV.

MATERIAL AND METHODS

5 Cell, Viruses and Plasmids

[0074] Vero cells (CCL-81) and A549 cells (CCL-185) were obtained from the ATCC 293-Flp-in cells (R750-07) were obtained from Invitrogen. Vero cells, A549 cells and 293-Flp-In cells were grown in EMEM medium, F12K medium and D-MEM (high glucose) respectively. All cell lines were supplemented with 10% heat inactivated fetal bovine serum (FBS) and 1X
10 Penicillin/Streptomycin (P/S) and L-glutamine and maintained in an incubator at 37°C and 5% CO₂.

[0075] RSV virus, strains A2 (NR-12149) and B1 (NR-4052) were obtained from BEI Resources, NIAID, NIH. All virus stocks were prepared by infecting a sub-confluent A549 cell monolayers with virus in F12K media with L-glutamine supplemented with 2 % FBS and 1X P/S
15 (infection media). Virus was collected 3-5 days post-infection (dpi) by freeze-thawing cells twice and combining with supernatant. Virus harvests were cleared of cell debris by centrifugation at 3,000 rpm for 15 min. Virus stocks to be used in challenged studies were pelleted by centrifugation at 7,000 rpm overnight. Pelleted virus was resuspended in F12K media supplemented with 50mM HEPES and 100mM MgSO₄, aliquoted and frozen at -80°C until
20 needed. Virus titers were determined by plaque assay in Vero cells.

[0076] Codon optimized RSV-G coding DNA for *E. coli* and mammalian cells were chemically synthesized. A *NotI* and *PacI* site was used for cloning the RSV A2 G ectodomain coding sequence (67-298) in the T7 based pSK expression vector (19) for bacterial expression and the pSecR vector for mammalian expression (20) to express G protein in *E. coli* and in
25 293Flp-In cells, respectively.

[0077] Purified RSV A2 F protein encoding amino acids 22-529 and fused to a polyhistidine tag produced in insect cells with endotoxin levels <1 EU/μg of protein was obtained from Sino biologicals.

Production of Recombinant G protein from *E. coli* (REG)

[0078] Recombinant RSV G 67-298 extracellular domain was expressed in *E. coli* BL21(DE3) cells (Novagen) and was purified as described previously (19, 21). Briefly, G protein expressed and localized in *E. coli* inclusion bodies (IB) was isolated by cell lysis and multiple washing steps with 1% Triton X-100. The pelleted IB containing G protein was resuspended in denaturation buffer and centrifuged to remove debris. The protein supernatant was renatured by slowly diluting the redox folding buffer. The renatured protein solution was dialyzed against 20 mM Tris-HCl pH 8.0 to remove the denaturing agents. The dialysate was filtered through a 0.45 µm filter and was purified through a HisTrap FF chromatography column (GE Healthcare). Protein concentration was analyzed by BCA (Pierce) and purity of the purified REG protein was determined by SDS-PAGE. Endotoxin levels of the purified protein were <1 EU/µg of protein

Production of Recombinant glycosylated G protein using 293 Flp-In cells (RMG)

[0079] 293-Flp-In cells and pOG44 (plasmid expressing Flp-In recombinase) were obtained from Invitrogen (Carlsbad, CA). The 293-Flp-In cell line stably expressing the RSV A2 G protein was developed as described previously (20). Briefly, 293-Flp-in cells were transfected with the plasmids expressing Flp-in recombinase and the RSV A2 G ectodomain in DMEM media (Invitrogen). Twenty four hours after transfection, culture medium was replaced with fresh DMEM containing 150 µg/mL of hygromycin for selection of stably transfected cells. For protein expression, cells were maintained in 293-Expression media (Invitrogen), and cell supernatant was collected every 3-4 days. Supernatant was cleared by centrifugation and filtered through a 0.45 µm filter before purification through a His-Trap FF column (GE healthcare).

Gel filtration chromatography

[0080] Proteins at a concentration of 5 mg/ml were analyzed on Superdex S200 XK 16/60 column (GE-Healthcare) pre-equilibrated with PBS, and the protein elution was monitored at 280 nm. Protein molecular weight marker standards (GE healthcare) were used for column calibration and generation of standard curves to identify the molecular weights of the test protein samples.

Plaque reduction neutralization test (PRNT)

[0081] For PRNT, heat-inactivated sera was diluted 4-fold and incubated with 20-60 pfu of RSV from strain A2 or B1 for 1hr at 37 °C and 5 % CO₂ and the assay was performed in presence of 5% Guinea Pig complement as described previously (22). Briefly, Vero cells were infected with sera:virus mix and incubated for 1hr before removing the inoculum and adding an overlay of 0.8 % methylcellulose in infection media. Plates were incubated for 5-7 days and plaques were detected by immunostaining. Neutralization titers were calculated by adding a trend-line to the neutralization curves and using the following formula to calculate 50% end-point titers: antilog of [(50 + y-intercept)/slope].

10 Mice immunization, RSV challenge and sample collection

[0082] All animal experiments were approved under animal protocol study number #2009-20 by the US FDA Institutional animal care and use committee. Four to six week old female BALB/c mice were obtained from NCI (Frederick, MD). Mice were immunized intramuscularly at day 0 and day 20 with 5 µg of purified RSV protein combined with Emulsigen adjuvant.

15 Blood was collected by tail vein on days 0, 14 and 30. On day 34, mice were anesthetized with ketamine/xylazine cocktail and infected intranasally (i.n.) with 10⁶ pfu of RSV from strain A2 or B1. Mice were sacrificed by CO₂ asphyxiation either two or four days post-RSV challenge, wherein blood and lungs were collected. For histopathology analysis of lungs, the right lobe of the lung was collected on days 2 and 4 post RSV challenge by inflation with 10% neutral
20 buffered formalin. For viral load determination and cytokine analysis the left lobe of the lung was collected.

Lung viral load determination

[0083] Lungs were weighed and homogenized in F12K/2% FBS/1X P/S (5 mL media/gram of lung) using Omni tissue homogenizer (Kennesaw, GA). The supernatant was cleared by
25 centrifugation at 3,000 rpm for 10 min and used immediately for viral titration with a plaque assay as described above in Vero cells.

Measurement of cytokine levels in the lung

[0084] All lungs were weighed and homogenized in 5mL media/gram of lung, as described above, to normalize amount of lung tissue used per sample. Homogenized lungs were further

diluted in infection culture media containing 2X concentration of Complete-EDTA Free protease inhibitor cocktail (Roche, Basel, Switzerland) and used in a Bio-Plex Pro™ Mouse Cytokine 23-plex assay following the manufacturers recommendations. Plates were read using a Bio-Plex 200 System (Biorad, Hercules, CA).

5 Lung Histopathology

[0085] Lungs were fixed in situ with 10% neutral buffered formalin and removed from chest cavity. Fixed lungs were embedded into a paraffin block, sectioned and stained with hematoxylin and eosin (H&E) by HistoServ. Three mice per immunization and two slides per mouse were analyzed for histopathology. The slides were reviewed with an Olympus BX41 microscope.

- 10 Photomicrographs were taken with the microscope, an Olympus DP71 digital camera and images captured using Olympus cellSens software. Lung lesions in each mouse were graded (scored) according to the severity of each category for following criteria: whole lung section, bronchiolar lesions, vascular lesion and alveolar lesions. Severity score was 0, no lesions; 1, minimal lesions; 2, mild lesions; 3, moderate lesions; 4, severe lesions.

15 Surface Plasmon Resonance (SPR)

[0086] Steady-state equilibrium binding of post-vaccinated mouse sera was monitored at 25°C using a ProteOn surface plasmon resonance biosensor (BioRad). The recombinant G proteins from *E. coli* (REG) or 293T cells (RMG) were coupled to a GLC sensor chip with amine coupling with 500 resonance units (RU) in the test flow channels. Samples of 100 µl freshly prepared sera at 10-fold dilution or Mabs (starting at 1 µg/ml) were injected at a flow rate of 50 µl/min (120 sec contact time) for association, and disassociation was performed over a 600 second interval. Responses from the protein surface were corrected for the response from a mock surface and for responses from a buffer only injection. Pre-vaccination mouse sera were used as a negative control. Total antibody binding and data analysis results were calculated with BioRad

20

25 ProteOn manager software (version 2.0.1).

Statistical Analyses

[0087] The statistical significances of group differences were determined using a one-way ANOVA and Bonferroni multiple comparisons test. Correlations were calculated with a

Spearman's two-tailed test. p-values less than 0.05 were considered significant with a 95% confidence interval.

RESULTS

5 Expression and purification of glycosylated and non-glycosylated RSV G protein

[0088] The RSV G protein from the A2 strain is a heavily glycosylated protein of 298 amino acids with several N-linked and O-linked glycosylation sites in its native form (Figure 1A) (23). To determine the impact of glycosylation on G protein immunogenicity, a non-glycosylated extracellular domain of G protein from RSV-A2 strain was recombinantly produced in *E. coli* and termed REG (Fig. 1A). Insoluble G protein was purified from *E. coli* inclusion bodies, 10 denatured, renatured under controlled redox refolding conditions and purified by Ni-NTA chromatography as described in materials and methods. Purified REG protein displayed a single ~36 kDa molecular weight protein under reducing conditions in SDS-PAGE (Figure 1B) that was recognized by rabbit anti-G antibodies when tested in western blot assay (data not shown). 15 To produce a native glycosylated extracellular G protein (Fig. 1A), 293-Flp-In cells were stably transfected with plasmid expressing the ectodomain of the RSV A2 G coding sequence and secreted glycosylated G-protein in the mammalian cell culture supernatant was purified by Ni-NTA chromatography and termed RMG. Purified glycosylated G protein (RMG) exists as a single band of ~90kDa under reducing conditions (Figure 1C) similar to native viral G protein 20 that was recognized by rabbit anti-G polyclonal sera in a western blot (data not shown). In addition, both G ectodomains were recognized equally by G-specific MAb 131-2G, which binds to the central conserved region of the RSV G protein (24, 25) (Fig. 1D)

[0089] To determine if these purified recombinant proteins contain higher order quaternary forms, the purified RSV G proteins were subjected to size exclusion gel filtration 25 chromatography (Fig. 1E and F). The *in-vitro* refolded bacterially produced recombinant RSV G (REG) extracellular domain (67-298) contained three molecular weight (MW) forms representing approximately equal amounts of monomers, tetramers and higher order oligomeric form (Fig. 1E). In comparison, the glycosylated RSV G (RMG) extracellular domain (67-298)

purified from proteins secreted in supernatant from 293-Flp-In mammalian cell culture contained only oligomeric form (Fig. 1F).

Immunization of mice with non-glycosylated REG protein generates higher binding and neutralizing antibody titers compared with glycosylated RMG protein

5 [0090] The RSV G protein is one of the most immunogenic proteins of RSV (26, 27). To test the antigenicity of glycosylated (RMG) versus non-glycosylated (REG) G protein, BALB/c mice were immunized twice, 20 days apart, with 5 µg of REG, RMG, or F protein from RSV A2 strain adjuvanted with Emulsigen, or were mock vaccinated with PBS (Fig. 2). To test if immunization with REG or RMG protects mice from RSV challenge, 14 days after the second immunization all
10 animals were challenged with either RSV A2 (vaccine strain) or with the heterologous RSV B1 strain. Sera collected after the second immunization were tested for RSV-neutralizing activity in plaque reduction neutralization test (PRNT). Pre-vaccination (day 0) serum samples from all the animals tested negative in PRNT (data not shown). As expected, mice immunized with the F protein had high levels of neutralizing antibodies against both RSV strains, while mice
15 immunized with PBS did not have detectable levels of anti-RSV neutralizing antibodies (Figure 3 A-B). In mice immunized with REG, 7 out 11 (67%) had neutralizing antibodies against homologous RSV-A2 virus (Fig. 3A), but all had very weak or no measurable neutralizing antibodies against heterologous RSV-B1 strain as measured by PRNT (Fig. 3B). Surprisingly, mice vaccinated with the glycosylated RMG did not develop RSV-neutralizing antibodies
20 against either strain (as measured in the PRNT) after two immunizations (Fig. 3 A and B).

[0091] To get more complete information about the antibody responses to the two recombinant G proteins in mice, total anti-G binding antibody titers were measured by surface plasmon resonance based real-time kinetics assay (SPR) using REG or RMG proteins captured on SPR chip surface (Fig. 4A and B, respectively). As can be seen, sera from the REG immunized mice
25 demonstrated high total binding antibodies to the REG (Fig. 4A, red dots). Importantly, strong binding to the glycosylated RMG was also observed for the REG immunized mice sera (Fig. 4B, red dots). However, sera from mice immunized with the adjuvanted glycosylated RMG protein gave >10 fold lower binding to both REG and RMG proteins in SPR (Fig. 4A and B, green dots).

REG vaccination generates higher diversity of the antibody immune response compared to RMG vaccination

[0092] To test which antigenic regions are recognized by antibodies generated following REG and RMG vaccination, a series of antigenic peptides derived from the N-terminus, CCD and C-terminus of the G protein were designed and tested in SPR. Serum samples from REG vaccinated mice showed strong binding to all G peptides, suggesting that REG induces a diverse antibody immune response that encompasses most of the RSV G protein (Figure 4C). In contrast, sera from mice vaccinated with RMG generated antibodies that recognized peptides from the CCD and C-terminus, but not the N-terminus of RSV-G protein. In addition, the RMG-induced antibody titers were lower than those generated following REG vaccination for all peptides tested. These data suggested that the non-glycosylated RSV G protein can induce strong binding antibodies against more diverse epitopes of the G protein compared with the glycosylated RMG immunogen.

REG immunization provides better protection than RMG against both homologous and heterologous RSV challenge

[0093] To examine whether REG or RMG immunization could protect from homologous and heterologous RSV viral challenge, mice were intra-nasally (i.n.) infected with 1×10^6 pfu of either RSV A2 or RSV B1 virus strains 14 days after the second immunization with REG, RMG, F (positive control) and PBS (negative control) (Figure 5A-B). Since both viruses are not lethal for mice, we measured viral loads in the lungs on day 4 post-challenge. Two log reduction in lung viral loads compared to mock (PBS) vaccinated control animals is considered good control of virus replication. The majority of F-vaccinated animals reduced viral loads completely after challenge with both RSV A2 and B1 strains (Fig. 5A-B, purple symbols). Among the REG vaccinated animals, seven out of eight mice completely protected from viral replication in the lungs and the eighth animal in each group showed >2 fold reduction in viral loads following challenge with either RSV A2 or B1 strains (Fig. 5A-B, red symbols). In contrast, RMG immunization conferred more variable protection and reduction of viral loads after homologous RSV A2 (2/8 animals protected) or heterologous RSV B1 (4/8 protected) virus challenge (Fig. 5A-B, green symbols).

[0094] The lack of detectable neutralizing antibodies in mice that were completely or partially protected from challenge suggested that protection was mediated by immunological functions

not captured in the traditional RSV-PRNT, unlike the case in RSV F-immunized animals. A relationship plot between the individual mice anti-G binding serum antibody titers after the second immunization and lung viral loads after RSV challenge, (Figure 5C) showed that anti-G binding antibody titers have statistically significant inverse correlation with lung viral loads against the homologous RSV A2 strain (Fig. 5C) ($r = -0.6248$; $p = 0.0014$). In the case of RSV B1, a weaker inverse correlation was found between anti-G binding antibodies and lung viral loads ($r = -0.3961$; $p = 0.0613$; data not shown).

[0095] Therefore, it was apparent that REG immunogen conferred better protective immunity than RMG against both homologous and heterologous RSV challenge, but the mechanisms of protection may include immune mechanisms not measured using a classical RSV PRNT.

Glycosylated RMG induces enhanced pathology in the lungs following virus infection, while nonglycosylated REG protein provides protection from lung pathology

[0096] Previous studies have suggested that vaccines based on RSV G protein can prime mice for enhanced lung pathology following RSV challenge that was associated with increased cytokine production and cellular infiltration due to eosinophils, neutrophils and NK cells (28-31) (32). Therefore, we investigated whether REG or RMG mice immunization leads to enhanced lung pathology following RSV challenge. The two RSV strains used in current study, RSV A2 and B1, are known to cause mild lung pathology in mice. Lungs were collected from RSV-infected animals and were analyzed by histopathology as described in materials and methods. At 2 days following virus challenge, lungs from PBS (control) vaccinated mice showed mild lung pathology after RSV infection (Fig. 6). Lungs from RMG-immunized mice demonstrated more cellular infiltrates and higher scores of perivascularitis and interstitial pneumonia than placebo (PBS) immunized mice following either homologous or heterologous RSV viral challenge. In contrast, REG immunized mice showed lung pathology scores that were lower compared to PBS immunized mice after infection with RSV A2. Surprisingly, scores were similarly low in REG immunized mice following challenge with RSV B1 virus even though these mice lacked serum neutralizing antibodies against RSV as measured in PRNT (Figure 6A-B). In addition, cellular infiltrates in the lungs of PBS and REG immunized mice were composed mostly of lymphocytes with few eosinophils, while lungs in RMG immunized mice contained mostly eosinophils. Similar lung pathology was observed in animals on day 4 following RSV challenge (data not

shown). These results suggest that nonglycosylated REG immunogen provides good protection from RSV mediated disease by control of lung viral loads and reduces lung disease, while the glycosylated RMG counterpart induces enhanced pathology in the lungs following RSV infection.

5 Cytokine profile in lungs of REG and RMG immunized mice following virus challenge

[0097] A hallmark of RSV vaccine-induced enhanced pathology in mice is the increase in cytokine expression and the development of a skewed Th2 response. Therefore, levels of Th1 or Th2 cytokines and chemokines were measured in lung homogenates of immunized mice following RSV infection (Fig. 7). IL-4, known to be important for the development of a Th2 response, was >100-fold higher in the lungs of RMG immunized mice compared to placebo (PBS), while IL-4 levels in REG immunized mice were comparable to the PBS control (Figure 7A). IFN- γ , a Th1 cytokine, was only ~2 fold higher in RMG immunized mice than placebo or REG immunized mice (Figure 7E). As a consequence, the ratio of Th2/Th1 (IL-4/IFN γ) cytokines was significantly higher in the lungs of mice immunized with RMG immunogen compared with placebo or REG immunized mice following RSV infection (Figure 7I). IL-5, and IL-13, involved in eosinophil recruitment, airway hyper responsiveness and mucus production (29, 33), were found at significantly elevated levels in mice immunized with RMG compared to placebo, while in lungs from REG immunized mice these cytokines were at similar ranges as in the placebo control lungs (Figure 7B-D). We also identified significant production of IL-6, a proinflammatory cytokine often associated with increased C-reactive protein (CRP) and elevation in body temperature. The chemokines Eotaxin, MCP-1 and MIP-1 α , previously found to be involved in RSV induced enhanced lung pathology (34), were also observed at significantly higher levels in the lungs of RSV-challenged RMG-immunized mice compared with the REG-immunized or PBS control mice (Figure 7F-H). The elevated levels of cytokines and chemokines in the RMG immunized mice following viral challenge seems to be immune mediated and were independent of lung viral loads following RSV viral challenge since similar or higher viral loads were observed in the PBS control mice (Fig. 5).

[0098] Together, this study demonstrated that RMG vaccination induced enhanced lung cellular infiltration, as measured by both histopathology and cytokine levels after RSV challenge and did not elicit strong protective immunity, while the REG vaccine did not induce enhanced

pathology and conferred protection against both homologous and heterologous RSV strains even in the absence of a neutralizing antibody response.

DISCUSSION

5 [0099] New efforts are under way to develop subunit vaccines against RSV that will provide protective immunity without the potential for disease enhancement as was observed in infants following vaccination with the formalin-inactivated RSV (FI-RSV) vaccine in the 1960s (5, 35-37). Most recent RSV vaccine development efforts are focused on the F protein, which is more conserved among strains (3) and can be expressed in mammalian and insect cells (15, 16, 38, 10 39). The G attachment protein expressed on the virus surface also represents an important candidate for inclusion in RSV vaccines. However, glycosylated G protein expressed in mammalian cells or recombinant vaccinia virus expressing G protein was shown in mouse models to induce pulmonary eosinophilia upon RSV infection, mimicking the response seen with FI-RSV vaccine (33, 40, 41). Therefore, we decided to evaluate side by side the immunogenicity and safety of recombinant unglycosylated G protein ectodomain (amino acids 67-298) expressed 15 in *E. coli* (REG) and a fully glycosylated G ectodomain produced in 293-Flp-In mammalian cells (RMG) in a mouse RSV-challenge model. The FI-RSV model was not included in the current study since it has been documented in previous studies, wherein the effect in immune responses, lung cytokines and pathology can be influenced by multiple viral proteins. Mice were vaccinated 20 twice with low dose of REG or RMG (5 µg) in combination with the veterinary adjuvant Emulsigen (Figs. 1-2). Two vaccinations with REG generated neutralizing antibodies against RSV A2 in 7/11 animals in PRNT. In contrast, the glycosylated RMG did not elicit neutralizing antibodies as measured in the PRNT (Fig. 3). Previous studies wherein glycosylated G protein was shown to be protective in mice typically used three vaccinations with ≥ 10 µg protein per 25 dose. In this study, we gave only two vaccinations of a lower dose of the G proteins (5 µg) to differentiate the impact of glycosylation on virus neutralization and disease enhancement phenomenon after RSV infection of G-vaccinated animals. Total binding antibodies against the recombinant proteins (both REG and RMG) was measured by SPR and was found to be >10-fold higher in the REG immunized mice sera compared with sera from RMG vaccinated animals. 30 Furthermore, REG immune sera contained antibodies against peptides spanning the entire G

ectodomain, while RMG sera recognized more restricted targets in central conserved domain (CCD) and the C-terminus (Fig. 4C).

[0100] Homologous and heterologous protection was evaluated by measuring lung viral loads on day 4 after challenge with either RSV A2 (homologous) or RSV B1 (heterologous) strains.

5 Complete control of viral loads was observed in the majority of animals vaccinated with adjuvanted REG and challenged with either RSV strain. In contrast, the majority of RMG-vaccinated animals did not control virus replication after homologous or heterologous virus challenge (Fig. 5). The correlate of protection needs to be further investigated as classical PRNT
10 did not measure neutralizing antibodies in all animals that were subsequently protected against challenge. There were animals with low or no serum neutralization titers that controlled lung viral loads very well. This was most evident in animals challenged with the heterologous RSV B1 strain. It has been shown before that anti-G antibodies may neutralize virus *in vivo* but not in the *in vitro* PRNT (25). Cell mediated immunity may also play a role in virus clearance after RSV challenge. However, previous studies showed that BALB/c mice immunized with either FI-
15 RSV or recombinant vaccinia virus that expresses the RSV G attachment protein generated RSV specific CD4⁺ T cells but did not prime CD8⁺ T cells (42, 43). In our study, a strong inverse correlation was observed between the total anti-G binding serum antibody titers (measured by SPR) and lung viral loads after RSV challenge. Therefore, it is likely that some anti-G binding antibodies that are protective *in vivo* do not neutralize RSV very well in the PRNT as was found
20 for anti-G MAb 131-2G (25).

[0101] An important safety aspect of subunit and non-replicating candidate vaccines against RSV is its potential to enhance the occurrence of severe disease after infection. Elevated levels of Th2 cytokines were implicated in the enhanced cellular infiltration predominated by eosinophils. The RSV G glycoprotein in particular has been implicated as an RSV antigen that
25 promotes Th2 CD4⁺ T lymphocytes and induces eosinophilic infiltrates in the lungs after RSV challenge (28, 29, 33, 44-47). Therefore, lungs were isolated two days or four days after RSV challenge and evaluated by histopathology and quantification of cytokines and chemokines. It was observed that the glycosylated RMG vaccinated animals demonstrated significant lung histopathology after either RSV-A2 or RSV-B1 challenge (Fig. 6). Furthermore, a Th2-dominant
30 cytokine response was measured in the lungs of all RMG vaccinated RSV-challenged animals,

and chemokine levels (Eotaxin, MCP-1 and MIP-2 α) were elevated as well (Fig. 7). In addition, elevated IL-6, a known proinflammatory cytokine that is associated with upregulation of C-reactive protein and fever was found in the lungs of RMG-vaccinated animals. Importantly, the *E. coli* derived REG-vaccinated animals had no evidence of an increase in lung cellular infiltrates compared with PBS control animals and no elevation of Th2 cytokines and chemokines. The IL4/IFN γ ratios were significantly higher in the RMG vaccinated group compared with the PBS and REG vaccinated animals following virus challenge. The elevated levels of cytokines and chemokines and enhanced lung pathology in the RMG immunized mice following viral challenge seems to be immune mediated and was independent of lung viral loads following RSV viral challenge. Similar or higher viral loads were observed in the PBS control mice that showed lower cytokine levels and less lung pathology. The combination of low virus neutralizing activity and enhanced lung pathology observed in the RMG vaccinated animals is reminiscent of the findings in children and animals vaccinated with FI-RSV. The lack of enhanced lung pathology after REG vaccination is a very important finding that is an absolute requirement for further development of any RSV G based vaccine.

[0102] Both purified recombinant G proteins were identical in terms of their primary amino acid sequence and were administered in combination with the same adjuvant, Emulsigen. Therefore, the observed increase in Th2 cytokines (IL-3, IL-5, IL-13) and multiple chemokines in the lungs of RMG-vaccinated animals (but not REG-vaccinated animals) after RSV challenge could not be simply attributed to binding of the CX3C motif within CCD to the CX3CR1 (fractalkine receptor) on several cell types, as previously suggested (48). Instead, the enhanced Th2 and chemokine levels could possibly be induced by the high level of O-linked sugars characteristic of mammalian cell expressed glycosylated RMG and also of the native virus-associated G attachment protein. A role for processing by different glycosylation specific antigen presenting cell subsets that could influence the subsequent Th2 vs. Th1 cytokines has been reported and should be further investigated (49, 50).

[0103] Our findings are in agreement with previous studies using *E. coli* produced fusion protein, BBG2Na that contained the central conserved region of A2 G gene (amino acids 130-230) (G2Na) fused to the albumin-binding domain of streptococcal protein G protein (BB) formulated with aluminum adjuvant (51-54). In comparison with BBG2Na, our REG encompass

the entire G-ectodomain with no “foreign” sequences or fusion protein that may influence the immune response to the irrelevant target (BB rather than G2Na) during vaccinations. Most of the studies with BBG2Na used 20 µg of the vaccine in a series of three vaccinations. In contrast, in our study, only two vaccinations with 5 µg REG protein/dose elicited strong protection. Also, REG-immunization induced antibodies with diverse epitope repertoire that recognized multiple targets in the G protein spanning the N-terminus, central conserved domain, and C-terminus of the protein (Fig. 4C and data not shown). We will continue to investigate the contribution of antibodies against different parts of the G protein in homologous and heterologous protection against RSV infection.

[0104] In summary, our study for the first time compared side by side fully glycosylated and unglycosylated G proteins for immunogenicity and safety in a murine RSV challenge model. It also provided data supporting the feasibility of developing simple recombinant RSV G-based vaccine produced in *E. coli* expression system. This immunogen is unlikely to induce the Th2/Th1 imbalance observed after vaccination with fully glycosylated G protein that resulted in significant cellular infiltration and lung pathology. Moreover, when combined with the appropriate adjuvant it elicits protective antibodies against both homologous and heterologous RSV challenge. The bacterial expression system for G protein based vaccine provides economical alternative to cell-based subunit vaccines as was previously demonstrated for influenza HA1 proteins from multiple virus strains (21, 55-57), that should be evaluated alone or in combination with F- subunit vaccines for effective protection against RSV disease.

[0105] It is understood that the examples and embodiments described herein are for illustrative purposes only and that various modifications or changes in light thereof will be suggested to persons skilled in the art and are to be included within the spirit and purview of this application and scope of the appended claims. All publications, patents, and patent applications cited herein are hereby incorporated by reference in their entirety for all purposes.

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SEQUENCE LISTING

SEQ ID NO: 1 RSV-A2 ectodomain (67-298) codon optimized sequence for E. coli:

5 CATAAAGTCACCCCGACCCACGGCGATTATCCAGGATGCCACCTCTCAAATCAAAAA
CACCACGCCGACGTACCTGACCCAGAATCCGCAACTGGGCATTTACCGTCGAACCC
GTCAGAAATCACCTCGCAGATTACCACGATCCTGGCAAGCACCACGCCGGGTGTCA
AAAGCACGCTGCAATCTACCACGGTGAAAACCAAAAATACCACGACCACGCAGACC
CAACCGAGCAAACCGACCACGAAACAGCGTCAAAAATAAACCGCCGTCTAAACCGAA
CAATGATTTTCACTTCGAAGTGTTTAACTTCGTTCCGTCAGTATTTGTTCCAACAAT
10 CCGACCTGCTGGGCCATTTGTAAACGCATCCCGAACAAAAACCGGGCAAGAAAAAC
CACGACCAAACCGACGAAAAAACGACCCTGAAAAACGACCAAAAAAGACCCGAAA
CCGACGACGACCAAAAGCAAAGAGTGCCGACGACCAAAACCGACGGAAGAACCGA
CCATTAACACGACCAAAACCAATATTATCAGACCCCTGCTGACCTCCAACACGACCG
GCAATCCGGAACCTGACCTCACAGATGGAACGTTCCATTGACACGCTCTGAAGGT
AATCCGAGCCCGTCTCAGGTCAGCACGACCTCCGAATACCCGAGCCAGCCGTCTTCT
15 CCGCCGAATACCCCGCGTCAG

SEQ ID NO: 2 RSV-A2 ectodomain (67-298) protein sequence:

20 HKVTPTTAIIQDATSQIKNTTPTYLTONPQLGISPSNPSEITSQITTIILASTTPGVKSTLQSTTVKTKNTTTTQTQP
SKPTTKQRQNKPPSKPNNDFHFVFNFPVPCSI CSNNPTCWAICKRIP
NKKPGKKT'TTKPTKKPTLKT'TKDPKPKQTTKSKEVPTTKPTTEPTINTTKNIIITLLTSN
TTGNPELTSQMETFHSSTSEGNPSPSQVSTTSEYPSQPSSFPNTPRQ

25

RSV-F PEPTIDE SEQUENCES

SEQ ID NO:	Amino Acid No	Sequence	
3	1-34	MELLILKANAITTILTAVTFCFASGQNITEEFYQ	
4	23-74	ASGQNITEEFYQSTCSAVSKGYLSALRTGWYTSVITIELSNIKKKNCNGTDA	
30	5	101-157	PATNNRARRELPRFMNYTLNNAKNTNVTLSKKRKRFLGFLGVSIAIASGVAVSKV
6	82-115	ELDKYKNAVTELQLLMQSTPATNNRARRELPRFM	
7	101-121	PATNNRARRELPRFMNYTLNN	
8	137-155	FLGFLGVSIAIASGVAVS	
9	174-203	TNKAVVSLSNGVSVLTSKVLDLKNYIDKQL	
35	10	176-240	KAVVSLSNGVSVLTSKVLDLKNYIDKQLLPVNVKQSCSIS NIETVIEFQQKNNRLEITREFSVN
11	190-233	SKVLDLKNYIDKQLLPVNVKQSCSISNIETVIEFQQKNNRLEI	
12	195-240	LKNYIDKQLLPVNVKQSCSISNIETVIEFQQKNNRLEITREFSVN	
13	216-244	NIETVIEFQQKNNRLEITREFSVNAGVT	
40	14	234-287	TREFSVNAGVTTPVSTYMLTNSSELLSLINDMPITNDQKKLMSNNVQIVRQQSYS
15	250-273	YMLTNSSELLSLINDMPITNDQKKL	
16	310-368	DTPCWKLHTSPLCTTNTKEGSNICLTRTRDRGWYCDNAGSVSFFPQAETCKVQSNRVFCD	
17	341-358	WYCDNAGSVSFFPQAETC	
18	371-400	NSLTLPSEVNLCNVDIFNPKYDCKIMTSKT	
45	19	425-450	SNKNRGLIKTFSGCDYVSNKGVDTV
20	443-461	SNKGVDTVSVGNTLYYVVK	
21	475-521	INFYDPLVFPSPDEFDASISQVNEKINQSLAFIRKSDELLHNVNAVKS	
22	471-493	GEPIINFYDPLVFPSPDEFDASIS	
23	497-521	EKINQSLAFIRKSDELLHNVNAVKS	
50	24	552-572	ARSTPVTLSKDQLSGINNIAF

RSV-G PEPTIDE SEQUENCES

SEQ ID NO: Amino Acid No Sequence

	25	33-61	LNLKSVAQITLSILAMIISTSLIIAAIIF
	26	66-90	NHKVTPTTAIIQDATSQIKNTTPTY
5	27	90-110	YLTQNPQLGISPSNPSEITSQ
	28	129-152	TTVKTKNTTTTQTQPSKPTTKQRQ
	29	148-178	TTKQRQNKPPSKPNNDFFHFVFNFPVCSICS
	30	169-207	NFVPCSICSNNPWCWAICKRIPNKKPGKKTITTKPTKKPT
	31	236-263	INTTKTNIITLLTSNTTGNPELTSQME
10	32	245-274	TTLLTSNTTGNPELTSQMETFHSSTSEGNP
	33	263-298	ETFHSTSEGNPSPSQVSTTSEYPSQPSPPNTP

SEQ ID NO: 38 RSV-A2 F codon optimized sequence for E. coli:

15 ATGGAAGCTGCTGATTCTGAAAGCCAAACGCAATTACCACGATTCTGACCGCTGTTACCTTCTGTTTCGCCTCGGGC
 CAAAATATCACGGAAGAATTTTATCAGAGTACCTGCTCCGCGAGTTTCAAAGGCTATCTGAGCGCTCTGCGTACC
 GGTTGGTACACGTCGGTGATCACCATTGAAGTGAAGCAACATTAAGAAAGCAAGTGAATGGCAGGATGCCAAA
 ATCAAGCTGATTAAACAGGAACTGGACAAGTACAAGAACGCGAGTTACCGAAGTGCAGCTGCTGATGCAAGCACC
 CCGGCAACGAACAATCGTGCTCGTCCGGAAGTGCAGCGCTTCATGAAGTACACCTGAACAATGCTAAAAAGACC
 20 AATGTTACGCTGTCCAAAAGCGCAACGTCGCTTTCTGGGCTTCTGCTGGGTGTGGGTAGCGCCATTGCCAGC
 GCGCTTGACAGTCTCTAAAGTGTGCTGATCTGGAAGGTGAAGTTAACAAAATCAAGTCTGCGCTGCTGAGTACGAAC
 AAAGCCGTGGTTTCCCTGTCAAATGGTGTGTCGGTTCTGACCTCAAAGTGTGGATCTGAAGAATTATATCGAC
 AAACAGCTGCTGCCGATTGTGAACAAGCAATCGTGCGAGCATCTCTAACATCGAAACGGTTATCGAATTTTCAGCAG
 AAAAACAATCGTCTGCTGGAAATCACCCGTGAATTTTCGGTTAACGCGGTGTCAACACGCTGTGAGCACGTAT
 25 ATGCTGACCAATTCGGAAGTGTGAGCCTGATCAACGATATGCCGATTACCAATGACCAGAAAAAGCTGATGTCG
 AACAAATGTTCAAATTGTCCGTCAGCAAGTTACTCCATCATGAGCATATCAAAGAAGAAGTTCTGGCGTATGTC
 GTGCAGCTGCCGCTGTACGGCGTCATTGTATACGCGGTGCTGGAAGCTGCACACCTCTCCGCTGTGTACCACGAAC
 ACCAAAGAAGGTAGTAATATTTGCCCTGACCCGTACGGATCGCGGCTGGTATTGTGACAACGCGGGTTCAAGTTTCG
 TTTTCCCGCAGGCCGAAACCTGCAAAGTGCAAAGCAACCGTGTGTTTTCGATACGATGAACAGCCTGACCCGTG
 30 CCGAGTGAAGTCAACCTGTGCAATGTGGATATTTTCAATCCGAAATACGACTGTAAAGATCATGACCAGTAAACG
 GATGTTAGCTCTAGTGTATACCTCCCTGGGCGGATGTGTATGCTATGGTAAACCAAGTGTACGGCCAGC
 AACAAAGAATCGCGGCATTATCAAAACCTTTTCTAACGGTTGCGATTACGTTAGTAATAAAGGCGTCGACACGCTC
 TCTGTGGGTAACACCTGTATTACGTCAATAAACAGGAAGGCAAGAGTCTGTATGTGAAAGGTGAACCGATTATC
 AACTTTTACGATCCGCTGGTCTTTCCGTCGATGAATTTGACGCAAGCATCTCTCAGGTGAACGAAAGATTAT
 35 CAATCCCTGGCTTTCATCCGTAAATCAGACGAACTGCTGCATAACGTGAATGCGGTTAAAGCACCACGAATATC
 ATGATTACCACGATTATCATTGTATCATTTGTGATCCTGCTGTCCCTGATTGCAGTGGGCCTGCTGCTGTATTGT
 AAAGCTCGCTCAACGCCGTGACGCTGAGCAAAGACCAACTGTCGGGTATCAATAACATCGCATTAGCAAC

SEQ ID NO: 39 RSV-A2 F protein sequence:

40 MELLILKANAITTILTAFTFCFASGQNITEEFYQSTCSAVSKGYLSALRTGWYTSVITIELSNIKKKNCNGTDAKIK
 LIKQELDKYKNAVTELQLLMQSTPATNNRARRRELPRFMNYTLNNAKKTNVTLSKKRKRRLGLFLLGVGSAIASGVAV
 SKVLHLEGEVKNKISALLSTNKAVVSLSNVSVLTSKVLDLKNYIDKQLLPINVKQSCSISNIETVIEFQQKNNRLL
 EITREFSVNAGVTFPVSTYMLTNSSELLSLINDMPI TNDQKKLMSNNVQIVRQQSYSIMSIIKEEVLAYVVQLPLYGV
 IDTPCWKLHTSPLCTTNTKEGSNICLTRTRDGRWYCDNAGSVSFFPQAETCKVQSNRVFCDTMNSLTLPSEVNLNVD
 45 IFNPKYDCKIMTSKTDVSSSVITSLGAIVSCYGKTKCTASNKNRGIKTFNSNGCDYVSNKGVDTVSVGNTLYVYVVKQ
 EGKSLYVKGEPI INFYDPLVFPSPDEFDASISQVNEKINQSLAFIRKSDELLHNVNAVKSTTNIMITTIIVIVILL
 SLIAVGLLLYCKARSTPVTLSKDQLSGINNI AFNS

SEQ ID NO: 40 RSV-A2 F Ectodomain (23-524) protein sequence:

ASGQNIITEEFYQSTCSAVSKGYLSALRTGWYTSVITIELSNIKKKNCNGTDAKIKLIKQELDKYKNAVTELQLLMQS
 5 TPATNNRRARRELPRFMNYTLNNAKKTNVTLSSKKRKRRLGFLLGVSIAISGVAVSKVLHLEGEVNIKSALLSTNK
 AVVSLSNGVSVLTSTKVLDLKNYIDKQLLPVINKQSCSISNIETVIEFQQKNNRLLLEITREFSVNAGVTTTPVSTYMLT
 NSELLSLINDMPITNDQKKLMSNNVQIVRQQSYSIMSIIKEEVLAYVVQLPLYGVIDTPCWKLHTSPLCTTNTKEGS
 NICLTRTRDRGWYCDNAGSVSFFPQAETCKVQSNRVFCDTMNSLTLPSEVNLCNVDIFNPKYDCKIMTSKTDVSSSVI
 TSLGAIVSCYGKTKCTASNKNRGI IKTFSNGCDYVSNKGVDTVSVGNTLYYVVKQEGKSLEYVKGEPIINFYDPLVFP
 10 SDEFDASISQVNEKINQSLAFIRKSDELLHNVNAVKSTTN

SEQ ID NO: 41 RSV-B1 ectodomain (67-299) codon optimized sequence for E. coli:

CATAAAGTTACCCTGACCACGGTGACGGTTCAGACCATTAAAAACCACACCGAGAAAAACATCACCACGTACCTGAC
 CCAAGTTCGCCCGGAACGCGTCAGCTCTAGTAAACAGCCGACCACGACCAGCCCGATTATACCAACTCAGCGACGA
 15 CCTCGCCGAATACGAAATCTGAAACCCATCACACGACCGCCCCAAACCAAAGGCCGTACGACCACGAGTACGCAGACC
 AACAAACCGTCCACCAAACCGCGCTGAAAAATCCGCCGAAAAAACCGAAAGATGACTATCACTTCGAAGTCTTTAA
 CTTTCGTGCCGTGCAGCATTGTGTGTAACAATCAGCTGTGCAAATCAATTTGTAAAACCATCCCGTCGAATAAACCGA
 AGAAAAAACCGACGATCAAACCGACCAACAAACCGACACGAAAAACCGAATAAACGTGATCCGAAAAACCCCGCA
 AAAACCACCAAGAAAGAAACCAAGACCAACCCGACGAAAAAACCGACCCTGACGACCACGGAACGCGATACGTGAC
 20 CAGCCAATCTACCGTCCTGGACACCAAGACCCCTGGAACATACCATTAGCAACAGAGTCTGCACTCCACGACCCCGG
 AAAACACGCCGAATAGCACCAGACCCCGACCGCAAGCGAACCGAGCACCAGCAACTCCACCCAAAAACCCCAATCC
 CACGCA

SEQ ID NO: 42 RSV-B1 ectodomain (67-299) protein sequence:

HKVTLTFTVTVQTIKNHTEKNITTYLTQVPPERVSSSKQPTTTSPIHNTSATTSPNTKSETHHTTAQTKGRTTTSTQT
 25 NKPSTKPRLLKNPPKKPKDDYHFEVFNFPVPSICGNNQLCKSI CKTI PSNPKPKKPTIKPTNKPTTKTTNKRDPKTPA
 KTTKKETTTNPTKKPTLTTERDTSTSQSTVLDTTLEHTIQQSLHSTTPENTPNSTQTPTASEPSTSNSTQNTQS
 HA

30

1 WHAT IS CLAIMED IS:

2 1. An immunogenic composition comprising an immune stimulant and an
3 RSV oligopeptide selected from the group consisting of SEQ ID NO: 3 to 33 or a multimeric
4 RSV polypeptide comprising at least one sequence selected from the group consisting of SEQ ID
5 NO: 3 to 33.

1 2. The immunogenic composition of claim 1, wherein the composition
2 comprises an RSV oligopeptide selected from the the group consisting of SEQ ID NO: 3 to 33.

1 3. The immunogenic composition of claim 2, further comprising a second
2 immunogen.

1 4. The immunogenic composition of claim 3, wherein the second
2 immunogen is from RSV.

1 5. The immunogenic composition of claim 4, wherein the second
2 immunogen from RSV is selected from the group consisting of SEQ ID NO: 3 to 33.

1 6. The immunogenic composition of claim 3, wherein the second
2 immunogen is from a virus other than RSV.

1 7. The immunogenic composition of claim 1, wherein the composition
2 comprises a multimeric RSV polypeptide comprising at least one sequence selected from the
3 group consisting of SEQ ID NO: 3 to 33.

1 8. The immunogenic composition of claim 7, wherein the multimeric RSV
2 polypeptide comprises two different sequences selected from the group consisting of SEQ ID
3 NO: 3 to 33.

1 9. An immunogenic composition comprising an immune stimulant and an
2 unglycosylated respiratory syncytial virus (RSV) polypeptide.

1 10. The immunogenic composition of claim 9, wherein the unglycosylated
2 RSV polypeptide consists essentially of the ectodomain of an RSV G protein.

1 11. The immunogenic composition of claim 9 or 10, wherein the
2 unglycosylated RSV polypeptide consists of the amino acid sequence shown in SEQ ID NO: 2.

1 12. The immunogenic composition of claim 9, wherein the unglycosylated
2 RSV polypeptide consists essentially of the ectodomain of an RSV F protein.

1 13. The immunogenic composition of any of the proceeding claims, wherein
2 the unglycosylated RSV polypeptide is recombinantly expressed in a prokaryotic cell.

1 14. The immunogenic composition of any of the proceeding claims, further
2 comprising a second immunogen.

1 15. The immunogenic composition of claim 14, wherein the second
2 immunogen is from RSV.

1 16. The immunogenic composition of claim 14, wherein the second
2 immunogen is from a virus other than RSV.

1 17. A method of inducing a protective immune response against RSV
2 infection in a subject, the method comprising administering to the subject an immunologically
3 effective amount of the immunogenic composition of any one of claims 1 to 16.

1 18. The method of claim 17, wherein the immunogenic composition is
2 administered intramuscularly.

1 19. The method of claim 17, wherein the subject is an infant.

1 20. A method of detecting an immune response against RSV in a subject, the
2 method comprising contacting a biological sample from the subject with an RSV oligopeptide
3 selected from SEQ ID NO: 3-33 and detecting an immune response.

1 21. The method of claim 20, wherein the step of detecting an immune
2 response includes the step of detecting binding of antibodies in the biological sample to the RSV
3 oligopeptide.

1 22. The method of claim 21, wherein the step of detecting binding of
2 antibodies is carried by ELISA, a chemiluminescence assay, or a fluorescence assay.

1 23. The method of claim 20, wherein the step of detecting an immune
2 response includes the step of detecting a cellular immune response.

Figure 1

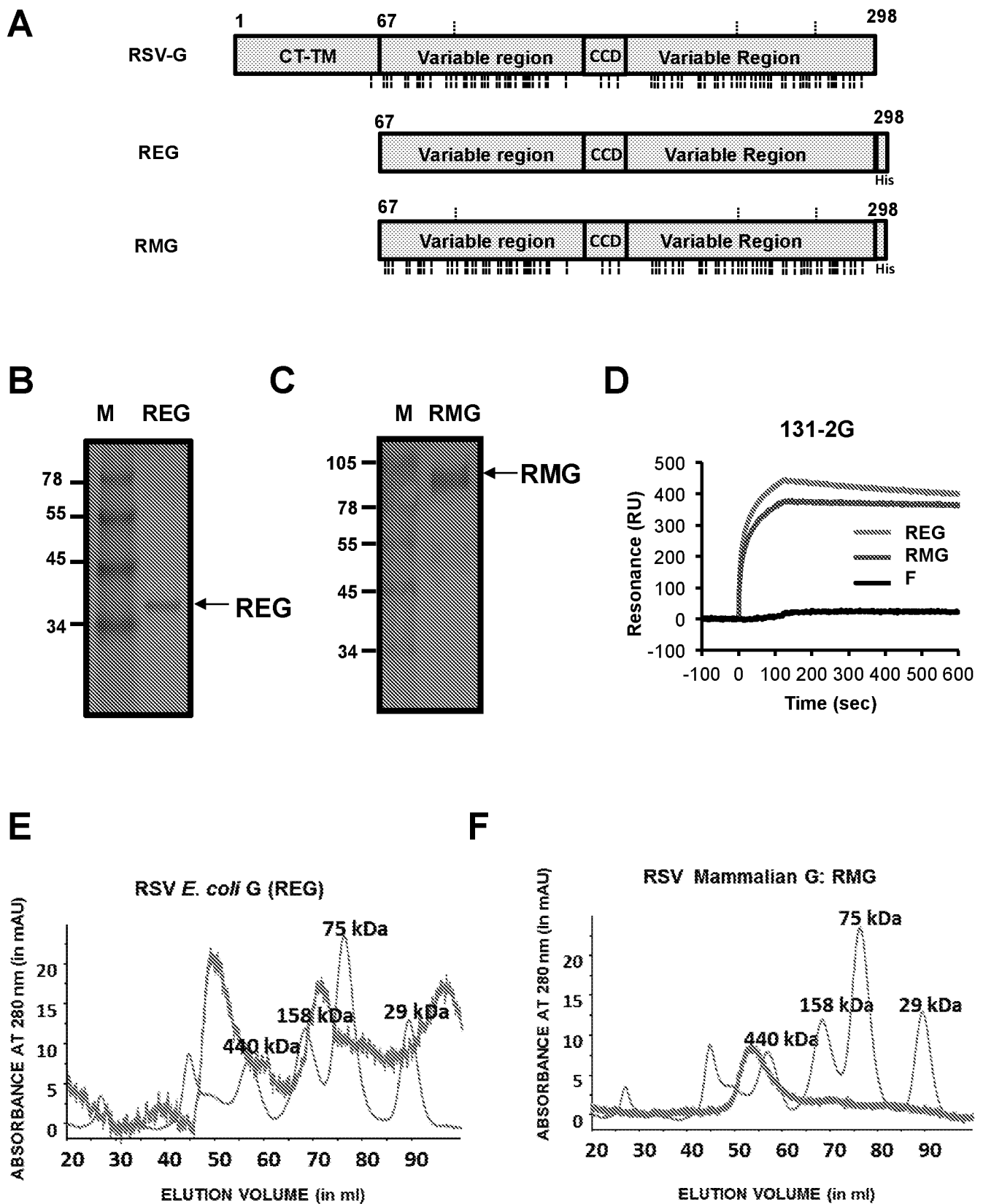
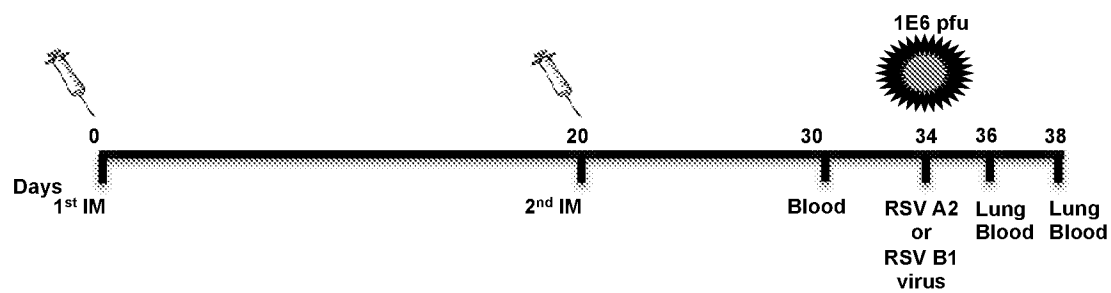


Figure 2



Group	Dose	Virus Challenge	N
PBS	-	RSV A2	9
		RSV B1	9
RSV A2 G <i>E. Coli</i> (REG)	5 ug	RSV A2	11
		RSV B1	11
RSV A2 G Mammalian (RMG)	5 ug	RSV A2	11
		RSV B1	11
RSV A2 F	5 ug	RSV A2	6
		RSV B1	6

Figure 3

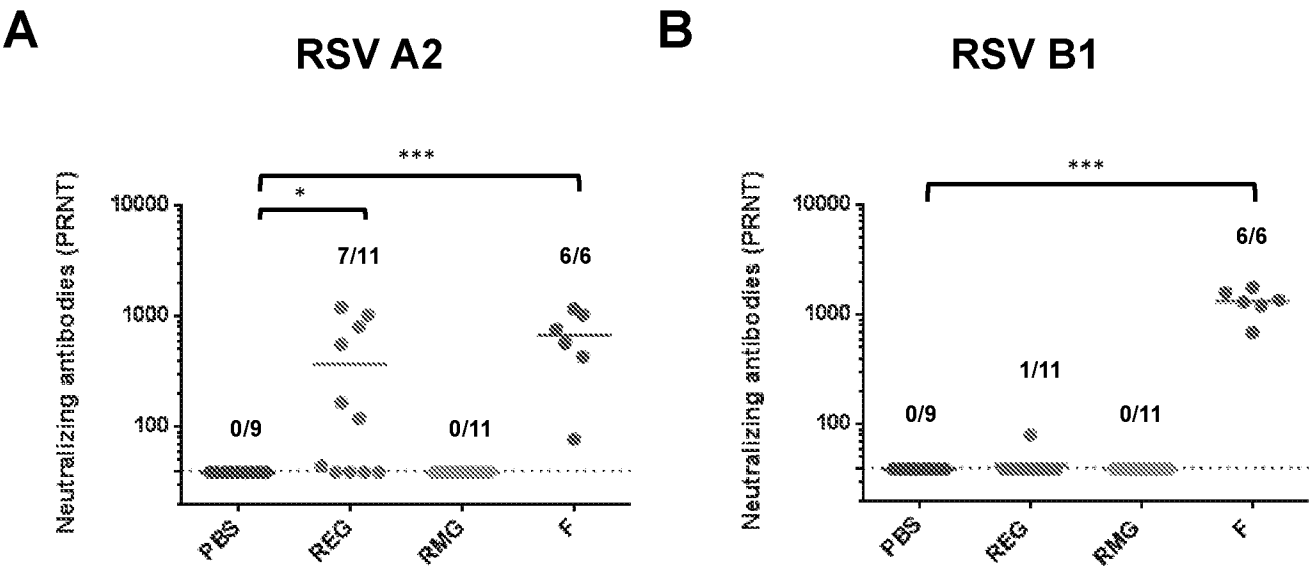


Figure 4

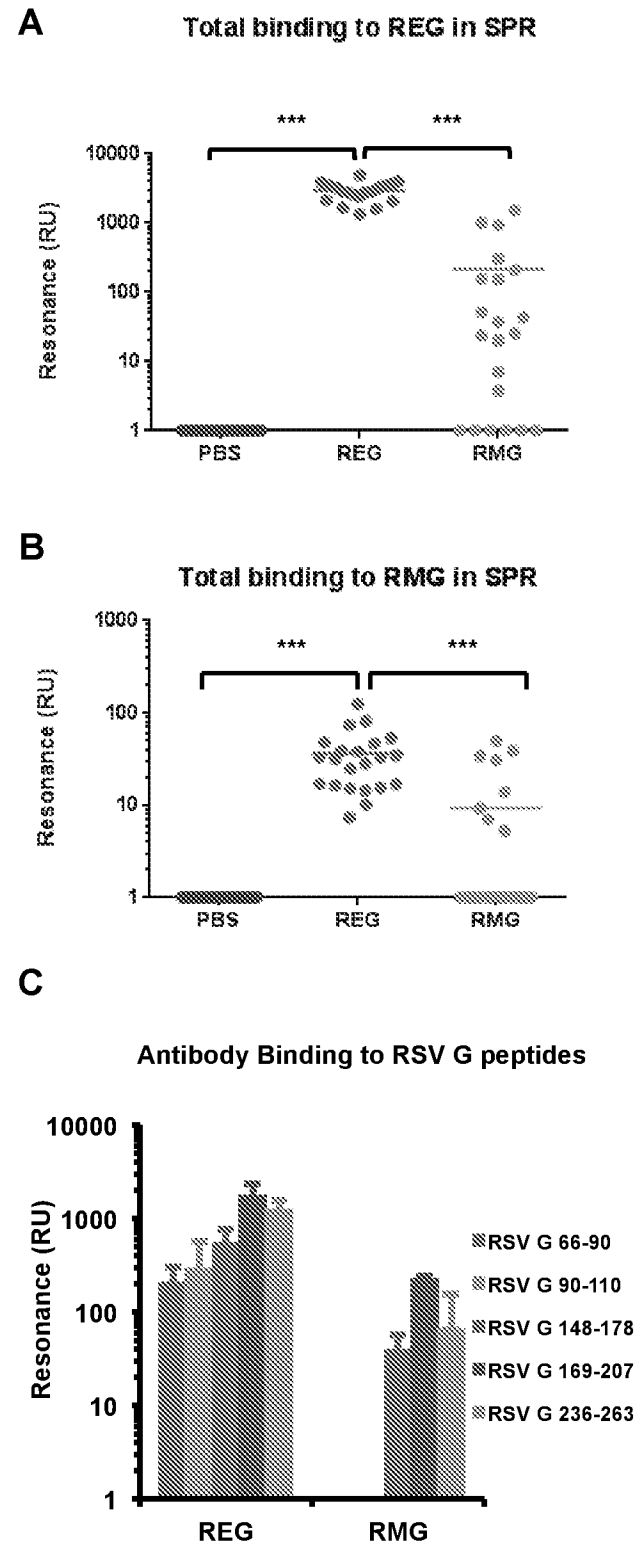


Figure 5

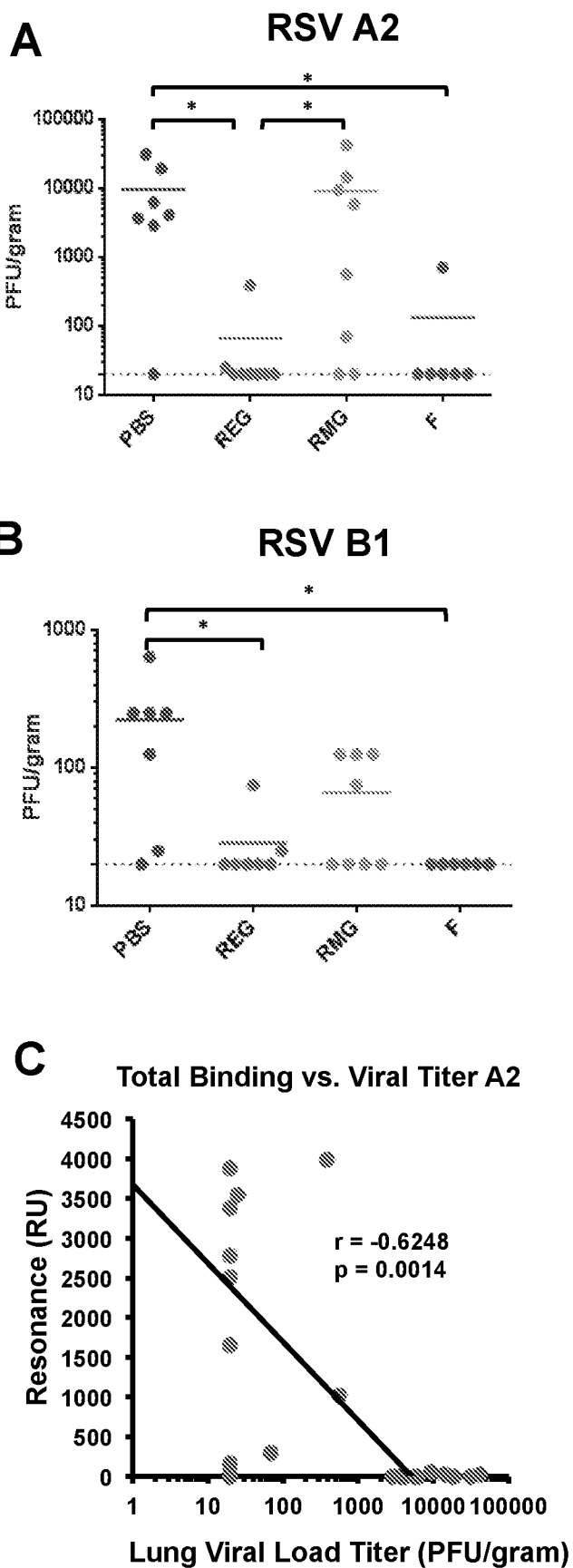


Figure 6

A

Immunization PRNT titers			Lung Pathology Scores			
			Peribronchiolitis	Perivascularitis	Interstitial Pneumonia	Total 12 max
RSV A2	PBS	<40	1	0.8	1	2.8
	REG	503 - 1198	0.5	0.8	0.5	1.8
	RMG	<40	1.3	1.8	1.8	4.9
RSV B1	PBS	<40	0	1.3	1	2.3
	REG	<40	0	0.7	0	0.7
	RMG	<40	1	1.7	1.2	3.9

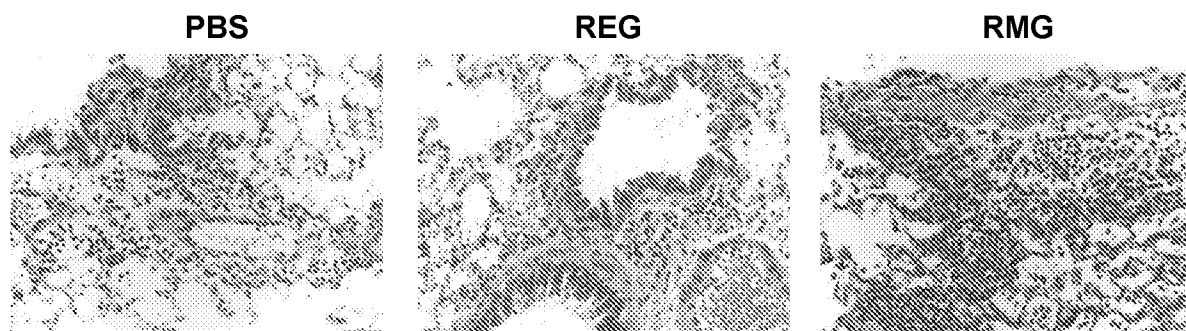
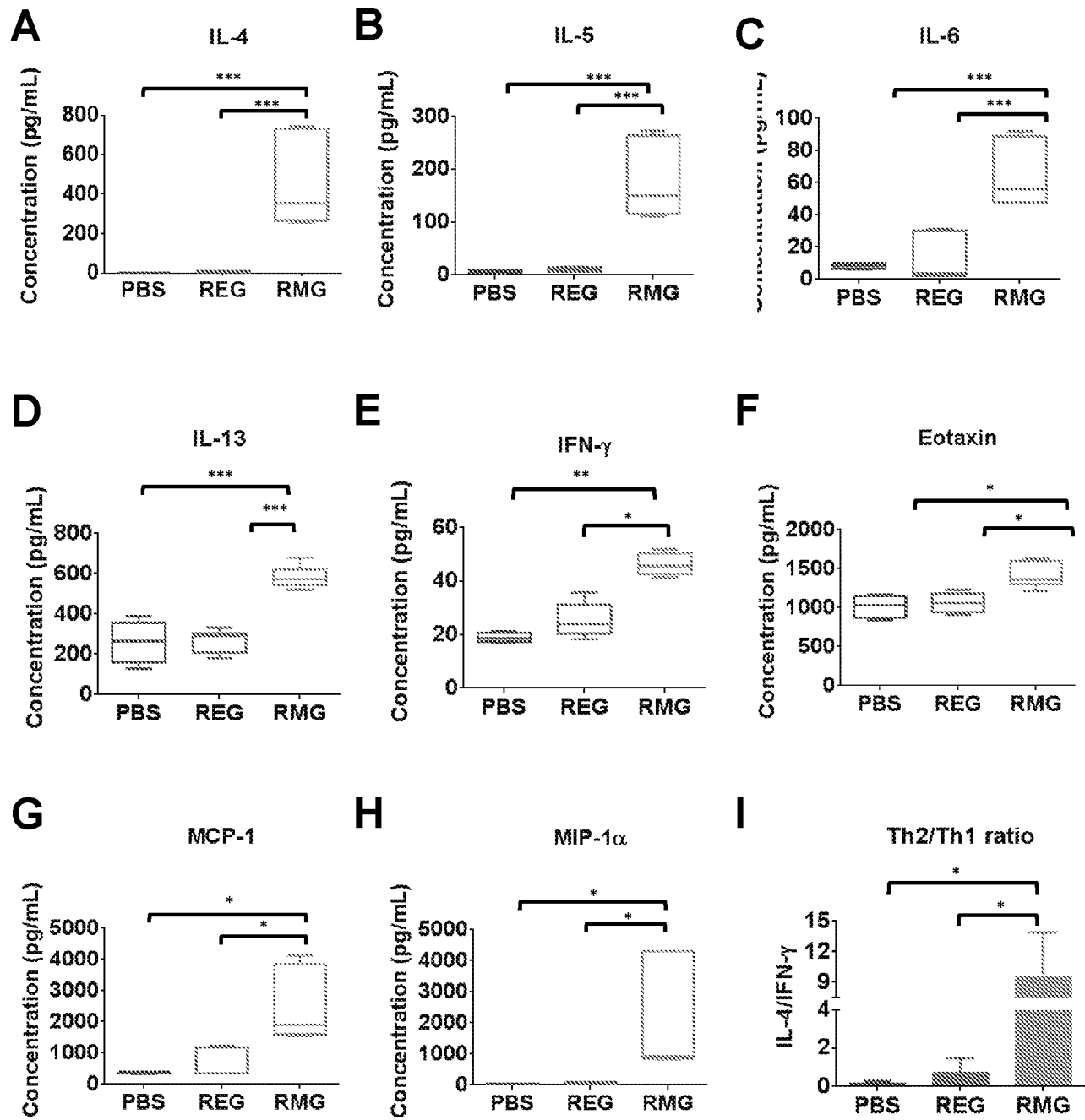
B

Figure 7



		1	50
RSV_A_ST_A2	(1)	MSKMKDQRTARTLEKTWDTLNHLEFISSCLYKLNKLSVAQITLSILAMII	
RSV_A_ST_LONG	(1)	MSKMKDQRTARTLEKTWDTLNHLEFISSGLYKLNKLSIAQITLSILAMII	
RSV_B_ST_18537	(1)	MSKHKNQRTARTLEKTWDTLNHLEVISSCLYRLNLKSIAQIALSVLAMII	
RSV_B_ST_B1	(1)	MSKHKNQRTARTLEKTWDTLNHLEVISSCLYRLNLKSIAQIALSVLAMII	
		51	100
RSV_A_ST_A2	(51)	STSLIIAAIIFIASANHKVTPPTAIQDATSQIKNTTPTYLTQNPQLGIS	
RSV_A_ST_LONG	(51)	STSLIITAIIFIASANHKVLTETTAIQDATSQIKNTTPTYLTQDPOLGIS	
RSV_B_ST_18537	(51)	STSLIIAAIIFIISANHKVLTETTVQTIKNHTEKNISTYLTQVPPERVN	
RSV_B_ST_B1	(51)	STSLIIAAIIFIISANHKVLTETTVQTIKNHTEKNITTYLTQVPPERVS	
		101	150
RSV_A_ST_A2	(101)	PSNPSEITSQITTLASTTPGVKSTLQSTTVKTKNTTTTQTOPSKPPTKQ	
RSV_A_ST_LONG	(101)	FSNLSEITSQITTLASTTPGVKSNLQPTTVKTKNTTTTQTOPSKPPTKQ	
RSV_B_ST_18537	(101)	SSKQPTTSPIHNTSATTSPNTKSETHHTTAQTKGRITTSQTENKPSKTS	
RSV_B_ST_B1	(101)	SSKQPTTSPIHNTSATTSPNTKSETHHTTAQTKGRITTSQTENKPSKTP	
		151	200
RSV_A_ST_A2	(151)	RQNKPPSKPNRDFHFEVFNFVPCISCSNNPTCWAICKRIPNKKPGKKTTT	
RSV_A_ST_LONG	(151)	RQNKPPNKPNDFFHFEVFNFVPCISCSNNPTCWAICKRIPNKKPGKKTTT	
RSV_B_ST_18537	(151)	RSKNPPKPKDDYHFEVFNFVPCISCGNNQLCKSICKTIPENKPKKKPTI	
RSV_B_ST_B1	(151)	RLKNPPKPKDDYHFEVFNFVPCISCGNNQLCKSICKTIPENKPKKKPTI	
		201	250
RSV_A_ST_A2	(201)	KPTKKPTLKTT-KKDPKPQTTKSKEVPPTTKPTEEPTINTTKTNIITLLT	
RSV_A_ST_LONG	(201)	KPTKKPTFKTT-KKDHPQTTKPKKEVPPTTKPTEEPTINTTKTNIITLLT	
RSV_B_ST_18537	(201)	KPTNKPTTKTTNKRDPKTPAKMPKKEIITNPAKKPTLKTTERDTSISQST	
RSV_B_ST_B1	(201)	KPTNKPTTKTTNKRDPKTPAKTTKETTTNPTKKPTLTTTERDTSISQST	
		251	299
RSV_A_ST_A2	(250)	SNITGNFELTSQMEFFHSTSEEGNPSPEQVETTSEYPSQPSPPNTTPO	
RSV_A_ST_LONG	(250)	NNITGNPKLTSQMEFFHSTSEEGNLSPEQVETTSEHPSQPSPPNTTPO	
RSV_B_ST_18537	(251)	VLDTIIPKYTIQQQSLHSTTSENTPSSTQIPTASEPSTLNPN-----	
RSV_B_ST_B1	(251)	VLDTTTLEHTIQQQSLHSTTPENTPNSTQTPASEPSTSNSTQNTQSHA	

Figure 8

INTERNATIONAL SEARCH REPORT

International application No
PCT/US2016/018530

A. CLASSIFICATION OF SUBJECT MATTER
INV. A61K39/155 C07K14/135
ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
C12N A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

EP0-Internal, BIOSIS, Sequence Search, WPI Data, EMBASE, CHEM ABS Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	WO 2014/140190 A1 (EMERGENT PRODUCT DEV GERMANY GMBH [DE]) 18 September 2014 (2014-09-18) the whole document	1-8, 17-23
Y	-& DATABASE Geneseq [Online] 6 November 2014 (2014-11-06), "Respiratory syncytial virus conserved G protein, SEQ ID 6.", XP002756649, retrieved from EBI accession no. GSP:BBN97565 Database accession no. BBN97565 sequence 6 ----- -/--	1-8, 17-23

☒ Further documents are listed in the continuation of Box C.

☒ See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

19 April 2016

Date of mailing of the international search report

22/07/2016

Name and mailing address of the ISA/

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Authorized officer

Renggli-Zulliger, N

INTERNATIONAL SEARCH REPORT

International application No
PCT/US2016/018530

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 2014/079842 A1 (NOVARTIS AG [CH]; CARFI ANDREA [US]; SWANSON KURT [US]) 30 May 2014 (2014-05-30)	1-5,7,8, 17-19
Y	claims 28,29; figure 1c; table 1; sequences 23,26	1-8, 17-23
X	----- WO 2014/144756 A1 (VLP BIOTECH INC [US]; MEDIMMUNE LLC [US]) 18 September 2014 (2014-09-18)	1-3,6, 17-19
Y	figure 9	1-8, 17-23
Y	----- WO 2011/091279 A2 (LIGOCYTE PHARMACEUTICALS INC [US]; STEADMAN BRYAN [US]; TAYLOR ROSS [U]) 28 July 2011 (2011-07-28)	1-8, 17-23
Y	----- DE GRAAFF P M A ET AL: "HLA-DP4 presents an immunodominant peptide from the RSV G protein to CD4 T cells", VIROLOGY, ELSEVIER, AMSTERDAM, NL, vol. 326, no. 2, 1 September 2004 (2004-09-01), pages 220-230, XP004525545, ISSN: 0042-6822, DOI: 10.1016/J.VIROL.2004.06.008 table 1	1-8, 17-23
X	----- WO 2014/174018 A1 (CRUCCELL HOLLAND BV [NL]) 30 October 2014 (2014-10-30)	1-5,7, 17-19
Y	example 12; sequence 21	1-8, 17-23
X	-& DATABASE Geneseq [Online] 18 December 2014 (2014-12-18), "RSV A2 strain fusion protein A2_F24 N67I+S215P, SEQ: 21.", XP002756650, retrieved from EBI accession no. GSP:BBP75385 Database accession no. BBP75385	1-5,7, 17-19
Y	sequence 21	1-8, 17-23

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2016/018530

Box No. I Nucleotide and/or amino acid sequence(s) (Continuation of item 1.c of the first sheet)

1. With regard to any nucleotide and/or amino acid sequence disclosed in the international application, the international search was carried out on the basis of a sequence listing:
 - a. ☒ forming part of the international application as filed:
 - ☒ in the form of an Annex C/ST.25 text file.
 - ☐ on paper or in the form of an image file.
 - b. ☐ furnished together with the international application under PCT Rule 13~~ter~~.1(a) for the purposes of international search only in the form of an Annex C/ST.25 text file.
 - c. ☐ furnished subsequent to the international filing date for the purposes of international search only:
 - ☐ in the form of an Annex C/ST.25 text file (Rule 13~~ter~~.1(a)).
 - ☐ on paper or in the form of an image file (Rule 13~~ter~~.1(b) and Administrative Instructions, Section 713).
2. ☐ In addition, in the case that more than one version or copy of a sequence listing has been filed or furnished, the required statements that the information in the subsequent or additional copies is identical to that forming part of the application as filed or does not go beyond the application as filed, as appropriate, were furnished.
3. Additional comments:

INTERNATIONAL SEARCH REPORT

International application No.
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Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box No. III Observations where unity of invention is lacking (Continuation of Item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.

2. ☐ As all searchable claims could be searched without effort justifying an additional fees, this Authority did not invite payment of additional fees.

3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

1-8, 17-23(all partially)

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee.
- ☐ The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation.
- ☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is SEQ.3, its use as a vaccine or in a method of detecting an immune response against RSV and a multimeric RSV polypeptide comprising at least SEQ. ID 3 and its use as a vaccine

2. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is SEQ.4, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least SEQ. ID 4 and its use as a vaccine

3. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is SEQ.5, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least SEQ. ID 5 and its use as a vaccine

4. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is selected from SEQ.6 or 7, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least one selected from SEQ. ID 6 or 7 and its use as a vaccine

5. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is SEQ.8, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least SEQ. ID 8 and its use as a vaccine

6. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is SEQ.9, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least SEQ. ID

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

9 and its use as a vaccine

7. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is selected from SEQ.ID 10, 11, 12 or 13, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least one sequence selected from SEQ. ID 10,11,12 or 13 and its use as a vaccine

8. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is selected from SEQ.ID 14 or 15, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least one sequence selected from SEQ. ID 14 or 15 and its use as a vaccine

9. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is selected from SEQ.ID 16 or 17 its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least one sequence selected from SEQ. ID 16 or 17 and its use as a vaccine

10. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is SEQ.ID 18, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least one sequence selected from SEQ. ID 18 and its use as a vaccine

11. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is selected from SEQ.ID 19 or 20, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least a sequence selected from SEQ. ID 19 or 20 and its use as a vaccine

12. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

and a RSV oligopeptide which is selected from SEQ.ID 21, 22 or 23, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least a sequence selected from SEQ. ID 21, 22 or 23 and its use as a vaccine

13. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is SEQ.ID 24, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least SEQ. ID 24 and its use as a vaccine

14. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is SEQ.ID 25, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least SEQ. ID 25 and its use as a vaccine

15. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is SEQ.ID 26, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least SEQ. ID 26 and its use as a vaccine

16. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is SEQ.ID 27, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least SEQ. ID 27 and its use as a vaccine

17. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is SEQ.ID 30, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least SEQ. ID 30 and its use as a vaccine

18. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

and a RSV oligopeptide which is selected from SEQ.ID 28, 29, 31 or 32, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least a sequence selected from SEQ. ID 28, 29, 31 or 32 and its use as a vaccine

19. claims: 1-8, 17-23(all partially)

An immunogenic composition comprising an immune stimulant and a RSV oligopeptide which is SEQ.ID 33, its use as a vaccine or in a method of detecting an immune response against RSV, and a multimeric RSV polypeptide comprising at least SEQ. ID 33 and its use as a vaccine

20. claims: 9-16(completely); 17-19(partially)

An immunogenic composition comprising an immune stimulant and an unglycosylated respiratory virus (RSV) polypeptide and its use as a vaccine

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No

PCT/US2016/018530

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