Abstract:
Title: SYSTEMS AND METHODS FOR ANALYZING LIQUIDS

FIG. 1

(57) Abstract: In one embodiment, an optical spectroscopy probe includes an optical fiber having a distal tip and a microfluidic filtering chamber attached to the distal tip of the optical fiber, the chamber comprising a microfluidic membrane adapted to enable liquid to enter the chamber but prevent particles from entering the chamber.
SYSTEMS AND METHODS FOR ANALYZING LIQUIDS

Cross-Reference to Related Application

This application claims priority to co-pending U.S. Provisional Application serial number 62/000,130, filed May 19, 2014, which is hereby incorporated by reference herein in its entirety.

Background

There are various situations in which it is desirable to analyze a liquid to determine the makeup of its contents. One example is hemolysis detection. Hemolysis is the disruption of red blood cells and release of hemoglobin and other intercellular components into the blood plasma. Hundreds of millions of blood tests are performed annually in the United States and a significant number of these tests are compromised because of in vitro hemolysis. Unfortunately, there is currently no reliable way to detect hemolysis without plasma separation. Therefore, significant delays are introduced that negatively affect treatment and diagnosis of the patients. If early signs of in vivo
hemolysis could be detected, it would significantly improve the outcome for many patients, including pregnant women affected by HELLP syndrome, which is characterized by hemolysis, elevated liver enzymes, and low platelet count.

Another situation in which it may be desirable to analyze a liquid is drug identification. Drug administration errors account for approximately 32% of preventable medical errors that result in morbidity or mortality. While there are several specialized assay-based techniques have been used to monitor medication errors in specific therapeutic treatments, these techniques have several limitations that prevent them from being used for point-of-care sensing. The techniques are time consuming and often require large volumes of analyte to achieve the needed sensitivity. Furthermore, drug assays use complicated electrochemical measurements, suffer from background interference in complex solutions, and have poor thermal stability.

In view of the above discussion, it can be appreciated that there is a critical need for a system and method that enables real-time, point-of-care analysis of liquids.

**Brief Description of the Drawings**

The present disclosure may be better understood with reference to the following figures. Matching reference numerals designate corresponding parts throughout the figures, which are not necessarily drawn to scale.

Fig. 1 is a schematic diagram of an embodiment of a system for analyzing liquids.

Fig. 2 is a partial perspective view of an embodiment of an optical spectroscopy probe that can be used in the system of Fig. 1.

Fig. 3 is a perspective view of a microfluidic filtering chamber of the optical spectroscopy probe of Fig. 2.
Fig. 4 is a partial plan view of a wall of the microfluidic filtering chamber of Fig. 3.

Fig. 5 is a schematic side view of the optical spectroscopy probe of Fig. 2, illustrating reflection of light emitted from an optical fiber of the probe.

Figs. 6A-6D are renderings of microfluidic membranes having various pore sizes.

Fig. 7 includes graphs of the velocity and vector fields inside microfluidic chambers having various pore sizes.

Figs. 8A-8E illustrate sequential steps in an embodiment of a microfluidic filtering chamber fabrication process.

Fig. 9 is an image of a fabricated microfluidic membrane filtering particles that were larger that the pore size of the membrane.

Fig. 10 is an image of small particles having passed through a fabricated microfluidic membrane having pores larger than the particles.

Fig. 11 is an image of cells being blocked by a fabricated microfluidic membrane having pores smaller than the cells.

Fig. 12 is a graph that shows the theoretical absorption spectrum for cobalamin.

Fig. 13 is a graph that shows cobalamin measurements when an optical fiber was spaced from a reflective microfluidic membrane at distances of 75, 105, and 150 µm.

Fig. 14 is an image illustrating microfiltration of a liquid using a microfluidic membrane.

Fig. 15 is a graph that shows the results of optical measurement of cobalamin concentrations.
Detailed Description

As described above, there is a critical need for a system and method that enables real-time, point-of-care analysis of liquids. Disclosed herein are examples of such systems and methods. In some embodiments, a system comprises an optical spectroscopy probe that includes a microfluidic filtering chamber that is used in conjunction with an optical fiber. The chamber of the probe can be immersed in a liquid and used to filter out components within the liquid that could otherwise skew the analysis results. The liquid can, for example, be a biological fluid, such as blood, or a fluid that is to be administered to a patient, such as intravenous fluid. Regardless, the chamber separates free-floating particles in the fluid from the remainder of the fluid to enable absorption spectroscopy to be performed on the fluid.

One goal for the disclosed systems and methods is to replace traditional hemolysis testing of blood samples, which requires large volume of blood to be collected, centrifugation, and bulk instrumentation, with a single setup that can work nearly instantaneously. A miniature, near-patient sensor for the detection of hemolysis will enhance patient diagnosis, treatments, costs, satisfaction, and experience. Another goal for the disclosed systems and methods is to overcome the limitations of drug identification testing by providing a new platform that can be used to detect the concentrations of drugs within a liquid by absorption spectroscopy, free of noise from particles and cells, without prior sample pre-processing.

These goals can be achieved through use of an optical spectroscopy system that comprises an optical spectroscopy probe that can be directly inserted into a liquid such as blood (in vivo or in vitro) or a solution that is to be administered to a patient. Fig. 1 illustrates an example embodiment of such a system. As shown in the figure, the system generally comprises an optical spectroscopy probe that is coupled to an
optical spectrometer 14 that can detect the intensity of absorbed light as a function of wavelength or frequency. The probe 12 includes an optical fiber 16 and a microfluidic filtering chamber (not visible in Fig. 1) that is provided at a distal end of the fiber. The optical spectrometer 14 is connected to a computing device 18 that executes software that can perform analysis on the light detected by the spectrometer and output conclusions based upon the analysis.

Fig. 2 illustrates an example configuration for the optical spectroscopy probe 12. As shown in this figure, the probe 12 comprises the optical fiber 16 identified in Fig. 1 and a microfluidic filtering chamber 20 that is attached to a cleaved distal tip 22 of the fiber. As shown in this figure, the microfluidic chamber 12 is configured as a generally rectangular box that is defined by a proximal (or top) wall 24, a distal (or bottom) wall 26, and multiple side walls 28. The proximal wall 24 and the distal wall 26 can be made of silicon nitride (Si$_3$N$_4$) and can be approximately 0.3 to 3 µm thick and have length and width dimensions of approximately 100 to 3,000 µm. The distal wall 26, and optionally the proximal wall 24, is perforated so as to comprise a plurality of pores that form a microfluidic filter or membrane through which fluid can flow, but particles larger than the pores cannot. Fig. 4 shows a partial detail view of one of the walls 24, 26. As indicated in this figure, the pores 30 can be generally circular and equidistantly spaced from each other across the plane of the wall 24, 26. In some embodiments, the pores 30 are approximately 1 to 10 µm in diameter or width.

Referring back to Fig. 2, the side walls 28 can be made of silicon and can be approximately 100 to 1,000 µm tall and approximately 10 to 500 µm thick. The optical fiber 16 can be approximately 100 to 400 µm in diameter and can have a core 32 that is approximately 8 to 50 µm in diameter. As shown in Fig. 3, which depicts the microfluidic filtering chamber 20 separate from the optical fiber 16, the proximal wall 24 of the
chamber 20 can have an opening 34 that is sized and configured to receive the distal tip 22 of the fiber. This opening 34 can also have a diameter of approximately 100 to 400 μm.

The distal wall 26 can be reflective to enable spectroscopic measurement of the fluid under evaluation. In some embodiments, the outer surface 36 of the distal wall 26 is coated with a reflective material, such as a reflective metal (see Fig. 5).

During use, the microfluidic filtering chamber 12 can be used to locally separate particles from fluid, which collects inside the chamber. For example, if the fluid is whole blood, the chamber 12 can separate red blood cells from the blood plasma. Because the distal wall 26 is reflective, light exiting the core 32 of the optical fiber 16 will propagate through the filtered liquid, reflect off of the distal wall, and couple back into the core, as depicted in Fig. 5. Hemolysis can be detected by measuring increased hemoglobin-related absorption in the locally filtered plasma. In some embodiments, the light has a wavelength that exhibits high hemoglobin absorption, such as approximately 532 nm. Alternatively, if the fluid is one that is to be administered to a patient, the absorption of the light by the fluid can provide an indication of a drug present in the fluid and its concentration.

SolidWorks™ was used to evaluate different perforation configurations. Figs. 6A-6D are SolidWorks™ renderings of microfluidic membranes having pore sizes of 1, 2, 5, and 10 μm. The optimal pore size that will allow sufficient fluid flow inside the device was determined using only minimum external pressure as the driving force. The goal was to determine the smallest optimal pore size that would provide substantial fluid through the membrane while impeding access of the relatively large particles within the chamber in which the membrane could be provided. The fluid simulation was iterated until the bulk average velocity reached steady state. The study showed that, when the
membranes have a pore size less than 1 µm, the flow velocity through the membrane is negligible without driving the liquid through the membrane with elevated pressure. Reasonable flow can be achieved starting from 2 µm pores with the speed of flow significantly increasing for 5 and 10 µm (see Fig. 7). In view of this, for applications in which no driving force is provided, the optimal pore size may be around 2 µm. For smaller pore sizes, additional pressure may need to be applied. Of course, the optimal pore size may also be dependent upon the viscosity of the fluid.

After detailed design optimization was performed, prototype microfluidic filtering chambers were fabricated using standard micro-fabrication processes. Figs. 8A-8E schematically illustrate steps of one such fabrication process. Beginning with Fig. 8A, a double-sided polished silicon wafer from Nova Electronic Materials (Texas) was used as the membrane substrate 40. Referring next to Fig. 8B, a 1 µm thick layer 42, 44 of silicon nitride was grown on both sides of the substrate 40 using low stress, low pressure chemical vapor deposition (LPCVD). The coated substrate 40 was cleaned with acetone and isopropyl alcohol (IPA) and then dried with nitrogen. Additionally, the substrate 40 was baked on hotplate at 115°C for 5 minutes to dry it. The substrate 40 was cooled for a few minutes and then spin-coated with photoresist. Next, the Si₃N₄ layers were patterned using ultraviolet (UV) photolithography and reactive ion etching to form pores 46 on the layer 42 (i.e., the distal wall) and an optical fiber opening 48 on the opposite layer 44 (i.e., the proximal wall), as shown in Fig. 8C. The pores 46 were 10 µm in diameter and were patterned in a square array with the distance of 15 µm from center to center. This configuration enables filtration of particles larger than 10 µm in diameter.

After patterning the silicon nitride layers 42, 44, the exposed silicon was etched away in a solution of potassium hydroxide (KOH, 85°C) for 7 hours, to create side walls.
50 and a hollow inner chamber 52 between the two nitride layers 42, 44 and the side walls, as shown in Fig. 8D. Finally, as shown in Fig. 8E, a 200 nm layer 54 of gold was deposited on top of the silicon nitride layer 42 using a Denton Thermal Evaporator.

The filtering properties of fabricated membranes were tested using micro-particle filtration. Fluid was pumped through the membranes using a peristaltic pump with a minimum pressure of 2.14 μL/s working at this lowest setting. Fig. 9 shows particles larger than the pore size being stopped by a membrane, while Fig. 10 shows particles smaller than the pore size freely propagating through the pores (particles can be seen on both sides of the transparent membrane in Fig. 10).

After continuous testing using an initial setup with direct fluid drop on the membrane, the delay in filtering was observed due to the accumulation of large number of particles on surface of the membrane. Next, a two-head polydimethylsiloxane (PDMS) microfluidic setup was designed. It was made with a replaceable watertight seal for experiments with different flow parameters while little external pressure was provided using the peristaltic pump. Replaceable capillary tubes were used instead of molding columns for flow on the PDMS and glass slides were used to hold the setup in place.

Next, breast cancer cells in a cell medium were used to demonstrate cell filtering and capture. Using a confocal microscope, images of human breast cancer cell line were captured as droplets of cells in the medium were introduced to the microfluidic membrane. The Hs578t epithelial breast cancer cells, as shown in Fig. 11, are on the larger scale than normal cells and had an average size of 11 μm. The cells clustered on top of the membrane.

In further testing, fabricated membranes were positioned underneath an upright optical microscope for observation, where the membrane surfaces were oriented
perpendicular to the microscope objective. The membranes were slightly elevated, allowing for space to exist beneath them. In order to observe filtration with these membranes, a droplet of deionized water containing naturally occurring contaminants (dust) was placed on top of the membrane surface. By relying on gravitational forces alone, the droplet of water was allowed to pass through the membrane, while contaminants were effectively filtered out. These effects were recorded using a microscope camera. After the filtration experiment was completed, the membranes were easily cleaned by rinsing with acetone.

After testing the filtering properties of the membranes, fabricated microfluidic filtering chambers comprising the membranes were attached to optical fiber tips. For this, an 8 \( \mu \text{m} \) optical fiber was cleaved and inserted into an adjustable fiber holder to provide mechanical support to the otherwise flexible fiber. Following this, the fiber holder was inserted into a high precision XYZ-stage and the tip was positioned approximately 105 \( \mu \text{m} \) from a reflective metal membrane by adjusting the dial of the optical stage. Finally, the entire device was fixed in place by epoxying the metal surrounding the membranes to a small PDMS tube. This tube was created to fit tightly to the optical fiber holder.

In order to optimize coupling, the optical fiber was set perpendicular to the membrane. Angular alignment of the system was performed by adjusting the XYZ-stage and observing the reflected power in air. An approximation of fiber angle was made qualitatively through visual observation. However, for added precision, reflected power was recorded while the fiber angle was finely tuned. The fiber angle was set once the reflected power reach a maximum value. Following angular alignment, the fiber needed to be placed at a set spacing from the membrane surface. The XYZ-stage allows for vertical adjustment of the fiber. However, micron-level precision was needed to
effectively determine the spacing, and thus Fabry-Perot resonance was used for high precision measurements. The resonance was formed between the gold-coated membrane and the cleaved fiber interface. The spectrum was recorded in air (refractive index: 1.0) and conducted using infrared light between 1,400 and 1,500 nm. Using the collected infrared (IR) spectrum, the distance could be calculated using the following expression:

$$d = \frac{X_1}{2n(\lambda_M - \lambda_r)}$$  \hspace{2cm} (Equation 1)

where $X_1$ and $\lambda_{i-1}$ are consecutive resonance wavelengths (nm) and $n$ is the refractive index. By recording resonance patterns for different vertical settings of the XYZ-stage, a correlation between stage setting and the actual distance, determined by Equation (1), was obtained. Once the appropriate vertical setting was found, the fiber was fixed in position and the distance was verified again using Fabry-Perot resonance. The Fabry-Perot resonance was recorded for the vertical setting once the fiber was set 105 $\mu$m from the membrane.

In order to demonstrate that the microfluidic filtering chamber is capable of identifying drugs and their concentrations, absorption spectroscopy was conducted for cobalamin (vitamin $B_{12}$). Cobalamin is an essential water-soluble vitamin, of which a deficiency can lead to abnormal neurologic and psychiatric symptoms. There are a variety of doses that are used for injections, from 0.2 $\mu$g/kg for neonates and infants to 1,000 $\mu$g/kg total for adults with severe vitamin deficiency.

The absorption spectrum of cobalamin dissolved in water with a concentration of 60 mg/dL is shown in Fig. 12. When drug is known, its concentration can be measured
at one specific wavelength, since it is much faster than measurements of the whole spectrum. The goal was to construct a sensor that would be able to conduct measurements for the broad range of concentrations from 0.1 mg/dL to 500 mg/dL. Theoretical modeling of this sensors transfer function was conducted using Beer-Lambert law:

\[ \frac{P}{P_0} = \exp(-2adC) \]  

(Equation 2)

where \( P \) is the power of transmitted light (W) for the fluid under study, \( P_0 \) is the transmitted power (W) for a pure sample, \( a \) is the molar absorptivity with units of L/mol cm, \( 2d \) is the total optical path where \( d \) was the spacing between the fiber and reflective surface (cm), and \( C \) is the concentration of the cobalamin expressed in mol/L. Fig. 13 shows the theoretical transfer functions plotted for the needed range of concentrations with 75, 105, and 150 \( \mu \text{m} \) spacing between the fiber and the membrane. While the 150 \( \mu \text{m} \) gap is better for the measurements of lower concentrations and the 75 \( \mu \text{m} \) works better for higher concentrations, the 105 \( \mu \text{m} \) gap is suitable for both ranges and thus was chosen for the experimental testing.

During the experiments, the concentration of cobalamin was varied from approximately 0.1 to 515 mg/dL while the reflected power was recorded for each concentration. All measurements were conducted over 5 minute timeframes, during which the power was averaged. A high stability green laser (532 nm) was used at a fixed power at 30 mW. After a cobalamin measurement was made, the sample was removed, and the sensor was thoroughly rinsed with water. This cleaning was conducted to prevent build-up of cobalamin on surfaces. After cleaning, the setup was
allowed to air dry for approximately 1 minute, ensuring that all water was removed from the system. Samples were tested sequentially with increasing concentration. In order to ensure reproducibility of results, every test was calibrated with respect to the water control measurements.

As previously mentioned, the filtering properties of the microfabricated porous membrane were demonstrated with a drop of deionized water. The water passed through the pores in the membrane and forms a drop on the other side, while all particles were filtered by the pores and remained on the membrane surface (see Figs. 14B-14F). Fig. 14A demonstrates the outline of the water drop that is not fully seen since it is already under the membrane and the dark spots are particles that are present in all real world samples and were successfully filtered out by the membrane. For cleaning purposes, the membrane was flushed with acetone. It was observed that this also improved its wetting properties. While without applying additional pressure, water takes several minutes to completely pass through the membrane. However, prior prewashing decreases this time to seconds. The membrane was also tested continuously with the lowest setting of a peristaltic pump pumping fluid with the flow rate approximately 2 µL/s, and it could withstand the external pressure still demonstrating successful particle filtration.

The aforementioned 0.2 µg/kg to 1,000 µ/kg dosage range translates to a range of concentrations from 1 µg/ml to 1,000 µg/ml. Fig. 15 shows measurements of concentrations between 1 µg /ml to 5 mg/mL to monitor the physiological range and potential overdose. Because cobalamin was the only compound sensed, all the measurements were conducted with a single wavelength, 532 nm, where cobalamin has high absorption and the sensor would have the highest sensitivity. The experimental points are well fitted by the theoretical curve obtained using Beer-Lambert Law ($R^2 =$
0.994).

Various specific embodiments have been described in the preceding disclosure. It is to be understood that those embodiments are example implementations of the disclosed inventions and that alternative embodiments are possible. All such embodiments are intended to fall within the scope of this disclosure.
CLAIMS

Claimed are:

1. An optical spectroscopy probe comprising:
   an optical fiber having a distal tip; and
   a microfluidic filtering chamber attached to the distal tip of the optical fiber, the chamber comprising a microfluidic membrane adapted to enable liquid to enter the chamber but prevent particles from entering the chamber.

2. The probe of claim 1, wherein the membrane is formed by a distal wall of the microfluidic filtering chamber.

3. The probe of claim 2, wherein the distal wall comprises a plurality of pores through which the fluid can flow.

4. The probe of claim 3, wherein the pores are approximately 1 to 10 \( \mu \text{m} \) in diameter or width.

5. The probe of claim 3, wherein the distal wall is reflective so as to be adapted to reflect light emitted from the end of the optical fiber back to the optical fiber.

6. The probe of claim 5, wherein the distal wall is coated with a reflective metal.

7. The probe of claim 2, wherein the microfluidic filtering chamber further includes a proximal wall that connects the chamber to the optical fiber.
8. The probe of claim 7, wherein the proximal wall comprises a plurality of pores through which the fluid can flow.

9. The probe of claim 7, wherein the proximal wall comprises an opening adapted to receive the distal tip of the optical fiber.

10. The probe of claim 7, wherein the microfluidic filtering chamber is box-shaped and further comprises multiple side walls that extend between the distal and proximal walls.

11. A system for analyzing liquid samples, the system comprising:
   an optical spectroscopy probe adapted for immersion into a liquid under evaluation, the probe including an optical fiber having a distal tip and a microfluidic chamber attached to the distal tip of the optical fiber, the chamber comprising a microfluidic membrane adapted to enable liquid to enter the chamber but prevent particles from entering the chamber; and
   an optical spectrometer to which the optical fiber is connected that receives reflected light from within the microfluidic chamber.

12. The system of claim 11, wherein the membrane is formed by a distal wall of the microfluidic chamber that comprises a plurality of pores through which the fluid can flow.

13. The system of claim 12, wherein the pores are approximately 1 to 10 µm in diameter or width.
14. The system of claim 12, wherein the distal wall is reflective so as to be adapted to reflect light emitted from the end of the optical fiber back to the optical fiber.

15. The system of claim 11, wherein the distal wall is coated with a reflective metal.

16. The sensor of claim 10, further comprising a computing device that executes software configured to analyze the light reflected back from the microfluidic chamber.

17. A method for analyzing a liquid, the method comprising:

immersing an optical spectroscopy probe in the liquid, the probe including an optical fiber having a distal tip and a microfluidic chamber attached to the distal tip of the optical fiber, the chamber comprising a microfluidic membrane adapted to enable liquid to enter the chamber but prevent particles from entering the chamber; and

analyzing the liquid within the microfluidic chamber to determine what it contains.

18. The method of claim 17, wherein analyzing comprises performing optical spectroscopy on the liquid.

19. The method of claim 17, wherein the liquid is whole blood and analyzing comprises determining if there is hemoglobin in the blood's plasma.

20. The method of claim 17, wherein the liquid contains a drug and analyzing
comprises determining the concentration of the drug.
FIG. 5

FIG. 6A

FIG. 6B

FIG. 6C

FIG. 6D
FIG. 13

FIG. 14
FIG. 15

Normalized Power ($P_{avg}/P_{water}$)

Cobalamin concentration (mg/mL)

$R^2 = 0.9944$
## A. CLASSIFICATION OF SUBJECT MATTER

<table>
<thead>
<tr>
<th>IPC(8)</th>
<th>CPC</th>
</tr>
</thead>
</table>

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC(8) - C12M 1/34; A61 B 5/145, 5/1455, 5/1459, 5/1495, 5/157 (2015.01)  
USPC - 435/278.1; 287.3; 288.7; 500/183, 342, 476, 478

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched


Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

Orbit, Google Patents, Google Scholar  
Search terms used: Spectroscopy, probe, fiber, microfluidic, membrane, filter, particle, reflect, metal, liquid, fluid, blood, immersion, hemoglobin, software

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

<table>
<thead>
<tr>
<th>Category*</th>
<th>Citation of document, with indication, where appropriate, of the relevant passages</th>
<th>Relevant to claim No.</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>6, 9, 15</td>
</tr>
</tbody>
</table>

Further documents are listed in the continuation of Box C.

See patent family annex.

- **A** special categories of cited documents:
  - **A** document defining the general state of the art which is not considered to be of particular relevance
  - **E** earlier application or patent but published on or after the international filing date
  - **L** document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
  - **D** document referring to an oral disclosure, use, exhibition or other means
  - **P** document published prior to the international filing date but later than the priority date claimed

**T** later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

**Y** document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

**Y** document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

- **S** document member of the same patent family

Date of the actual completion of the international search: 21 July 2015

Date of mailing of the international search report: 14 AUG 2015

Name and mailing address of the ISA:

Mail Stop PCT, Attn: ISA/US, Commissioner for Patents  
P.O. Box 1450, Alexandria, Virginia 22313-1450  
Facsimile No. 571-273-8300  
Authorized officer: Blaine Copenheaver