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(54) **METHOD AND SYSTEM FOR MULTIPLEX GENETIC ANALYSIS**

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(57) **ABSTRACT**

The present disclosure provides apparatus, systems and method for detecting separately and substantially simultaneously light emissions from a plurality of localized light-emitting analytes. A system according to exemplary embodiments of the present disclosure comprises a sample holder having structures formed thereon for spatially separating and constraining a plurality of light-emitting analytes each having a single nucleic acid molecule or a single nucleic acid polymerizing enzyme, a light source configured to illuminate the sample holder, an optical assembly configured to collect and detect separately and substantially simultaneously light emissions associated with the plurality of light emitting analytes. The system may further include a computer system configured to analyze the light emissions to determine the structures or properties of a target nucleic acid molecule associated with each analyte.

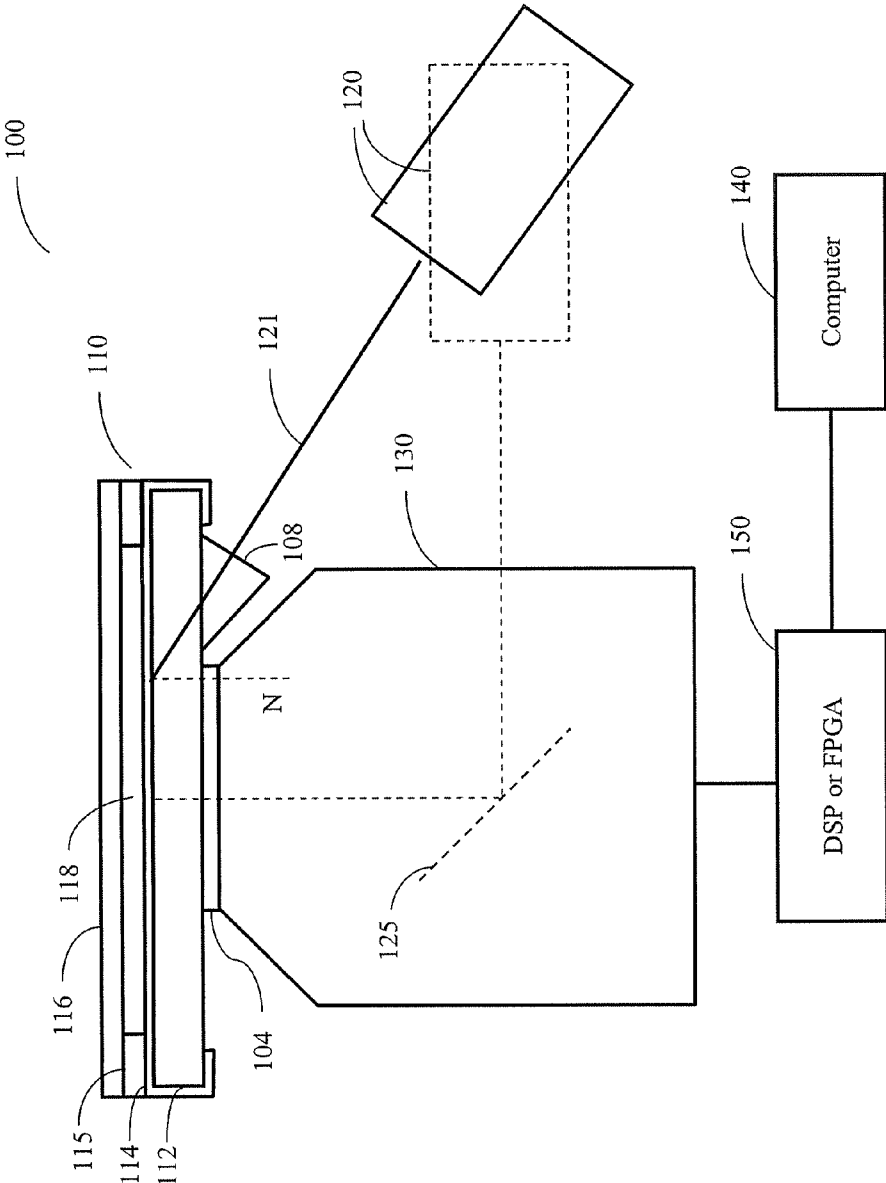


FIG. 1

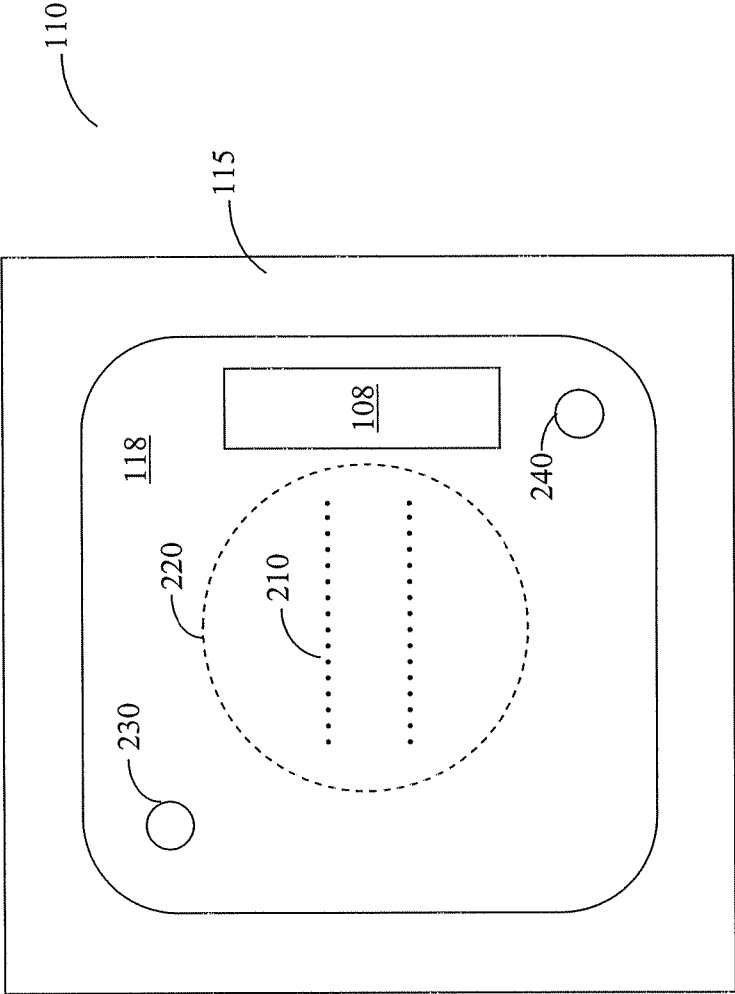


FIG. 2

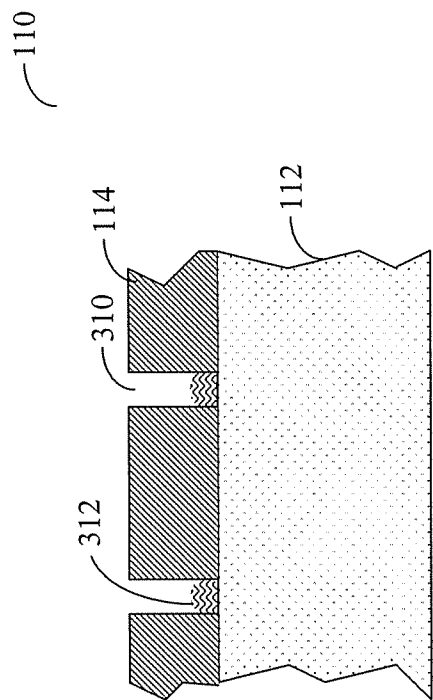


FIG. 3

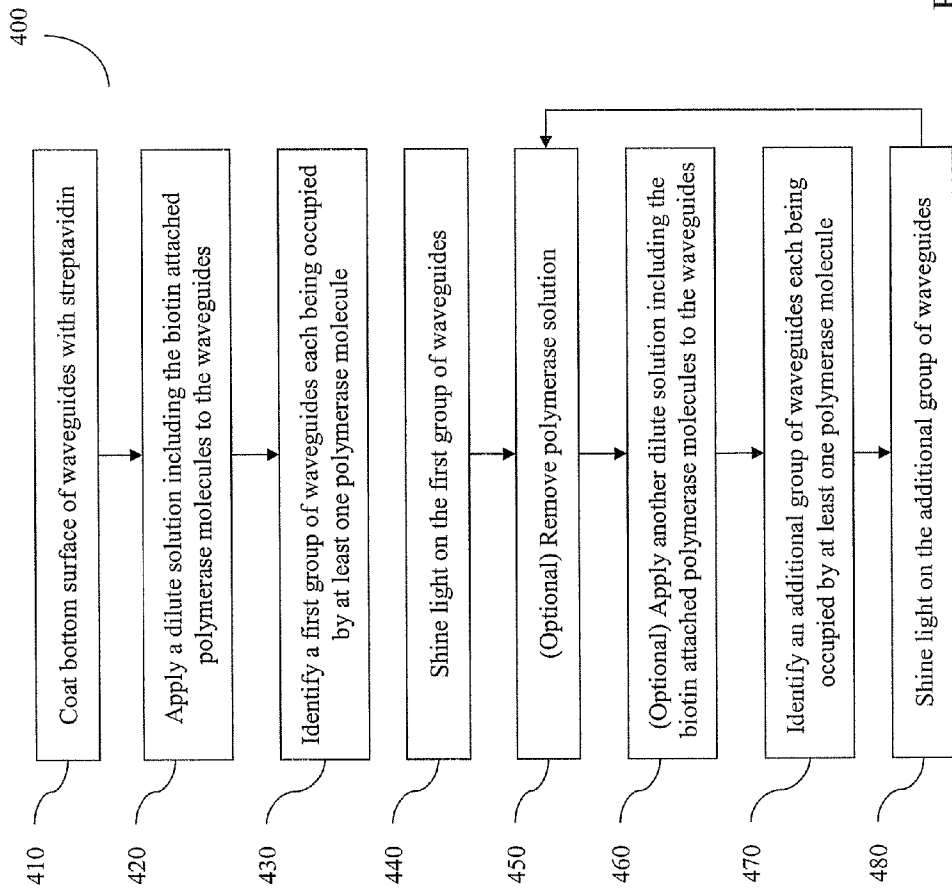


FIG. 4

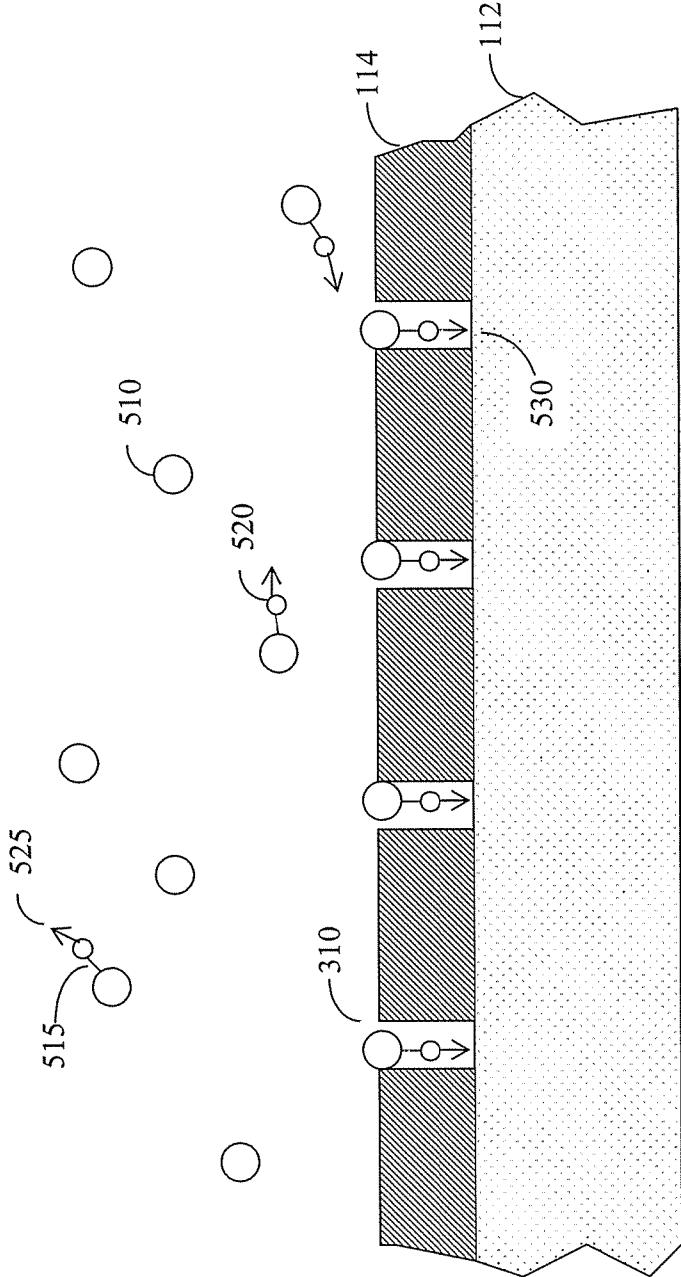


FIG. 5

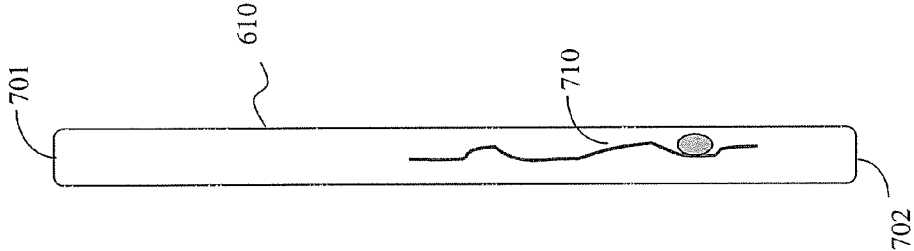


FIG. 7

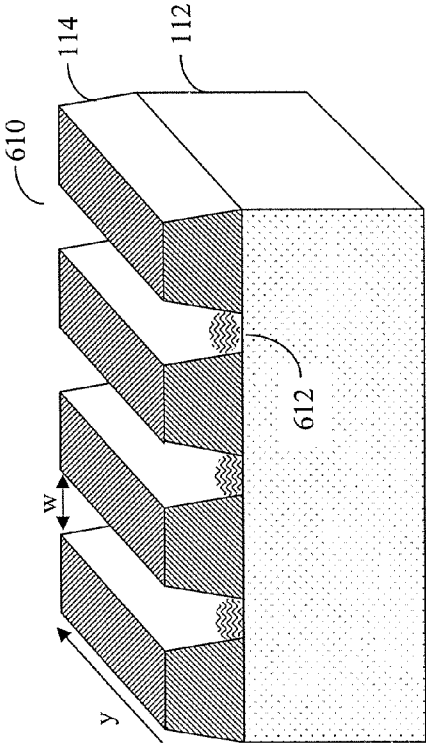


FIG. 6

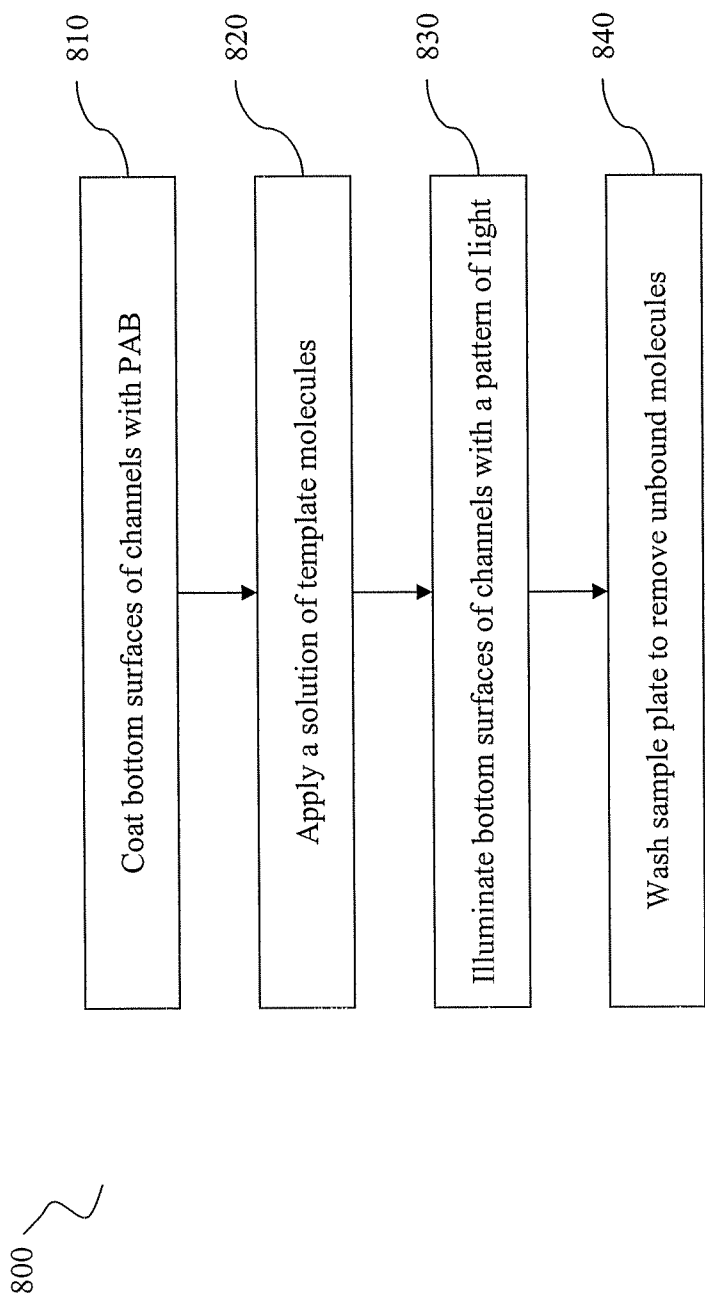


FIG. 8A

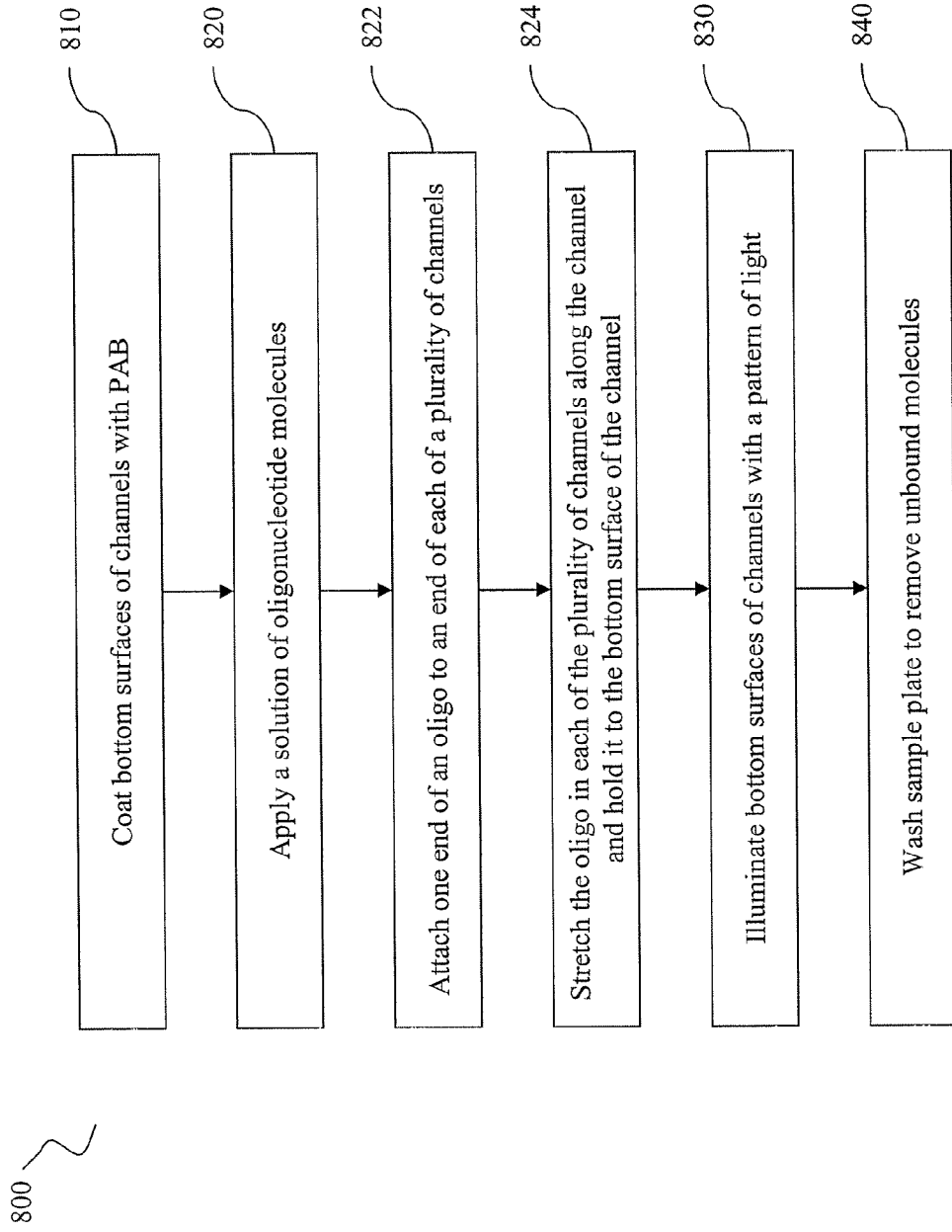


FIG. 8B

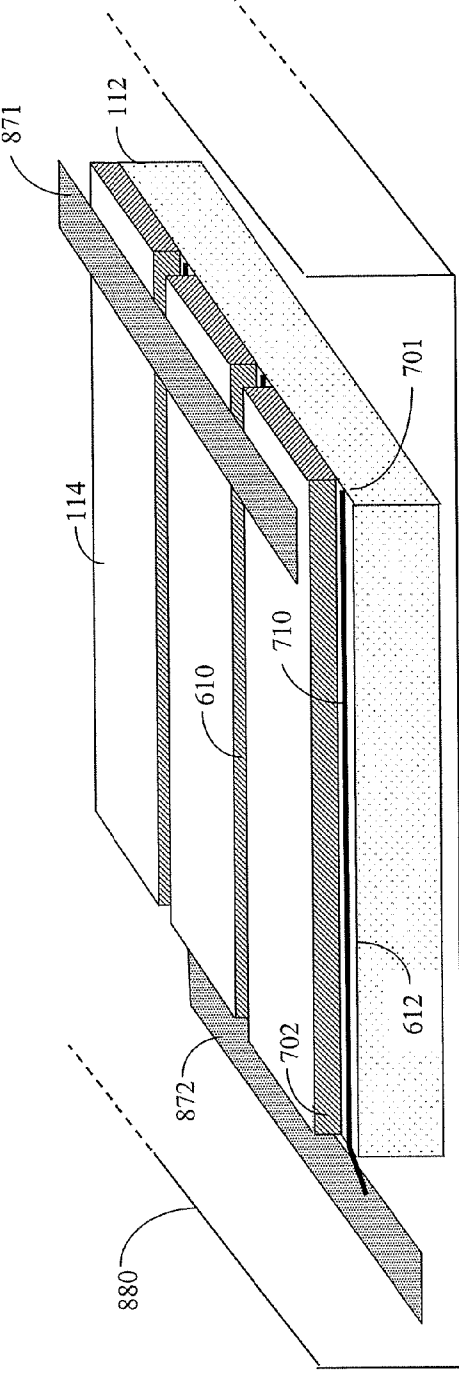


FIG. 8C

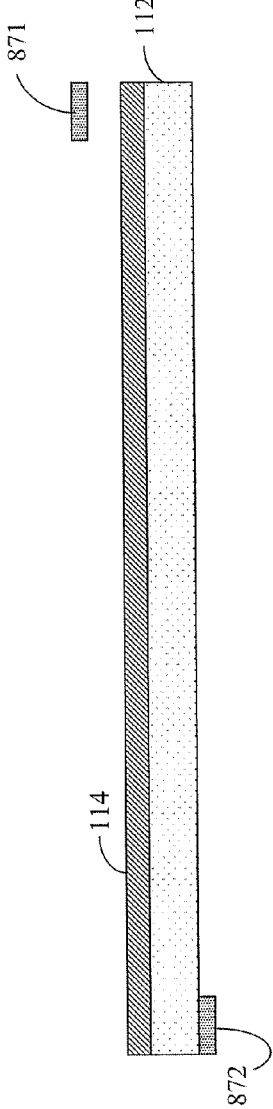


FIG. 8D

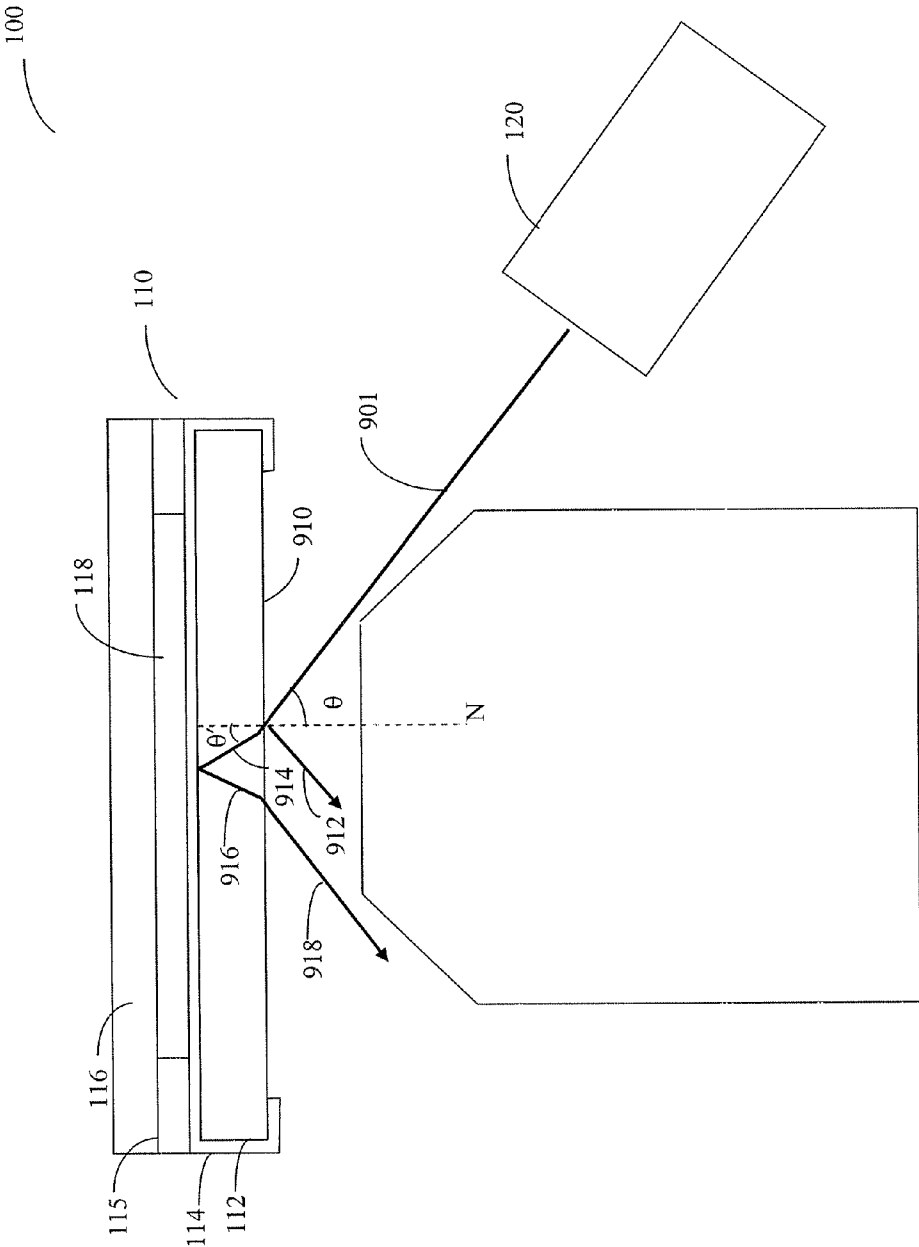


FIG. 9

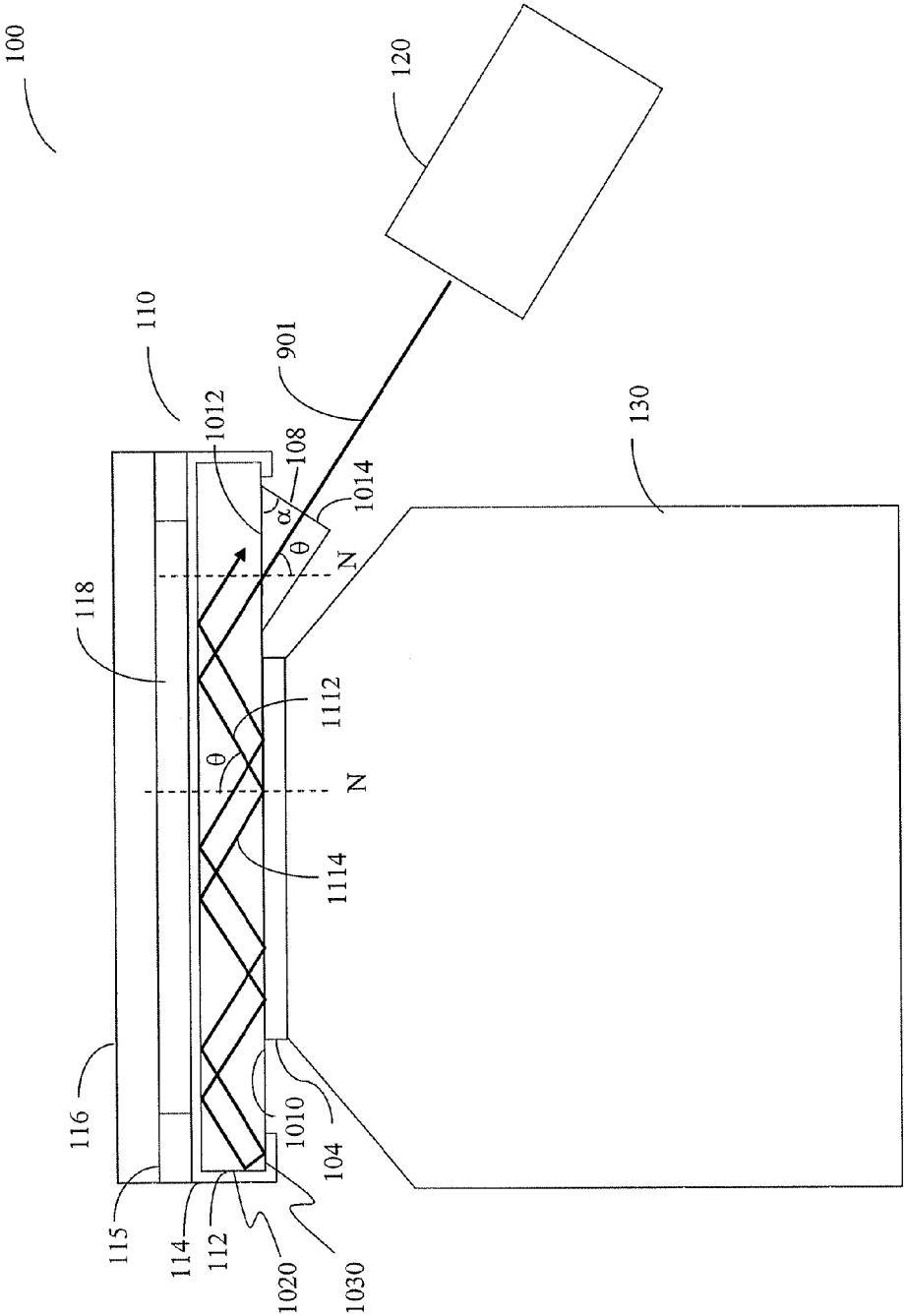


FIG. 10A

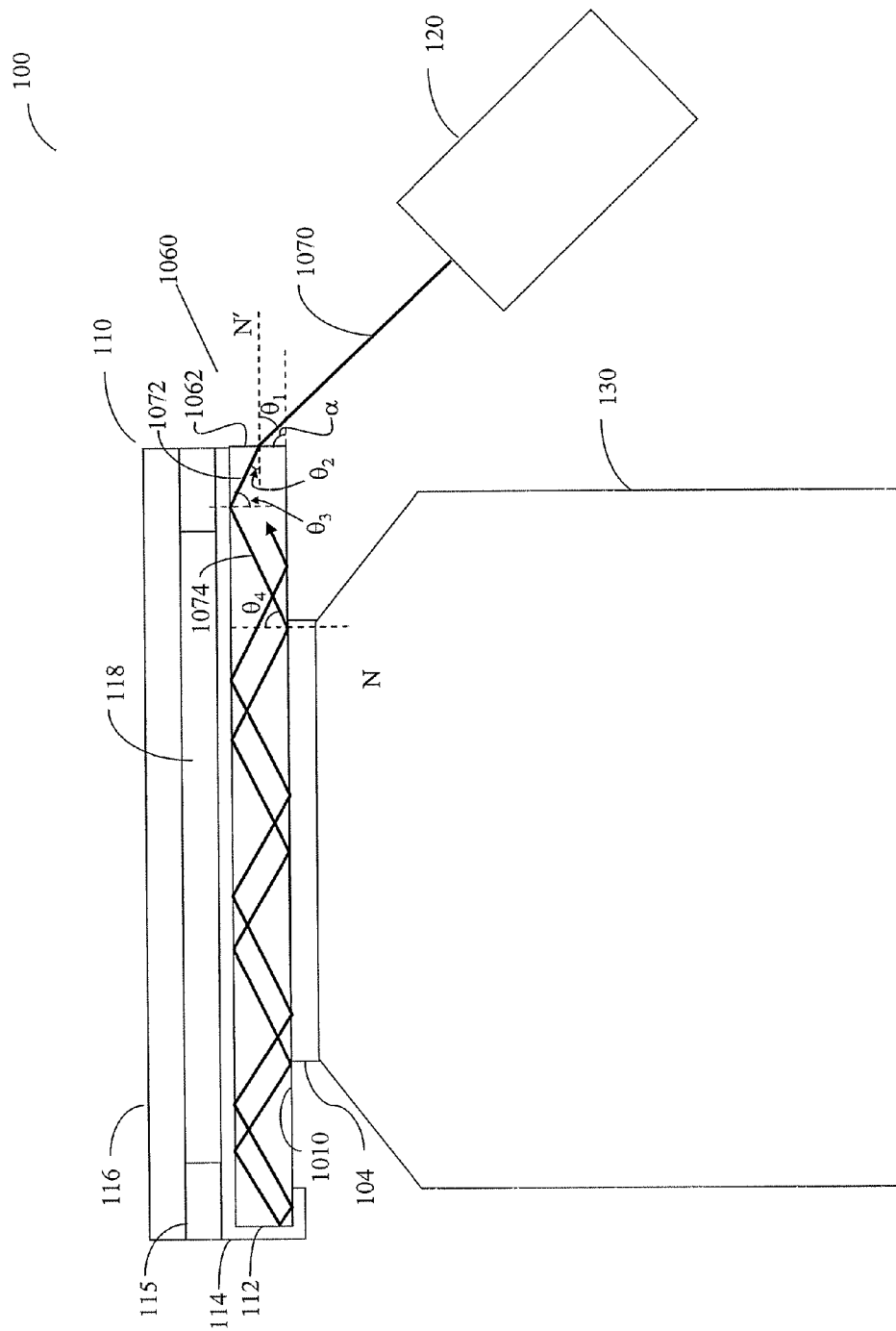


FIG. 10B

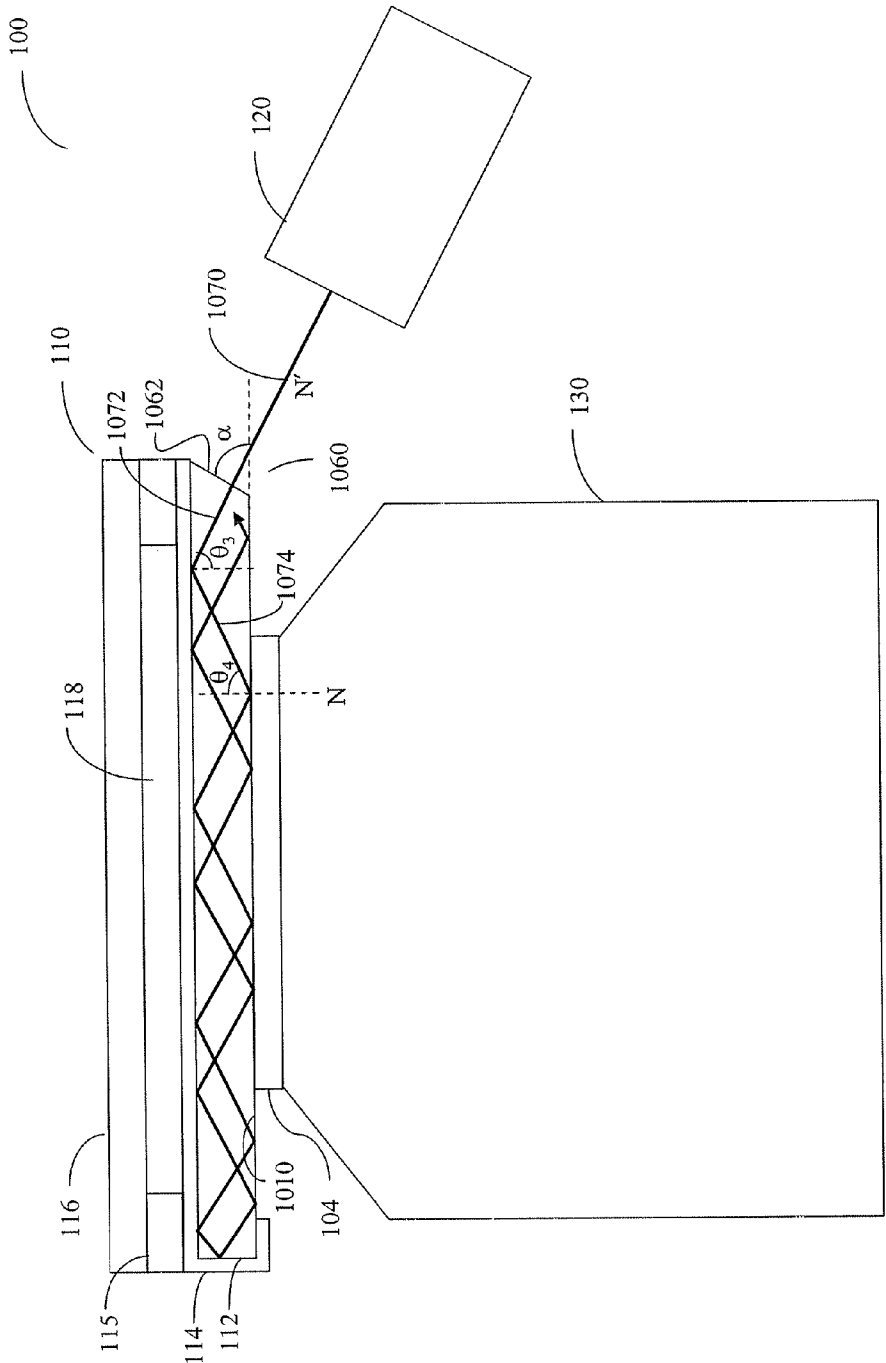


FIG. 10C

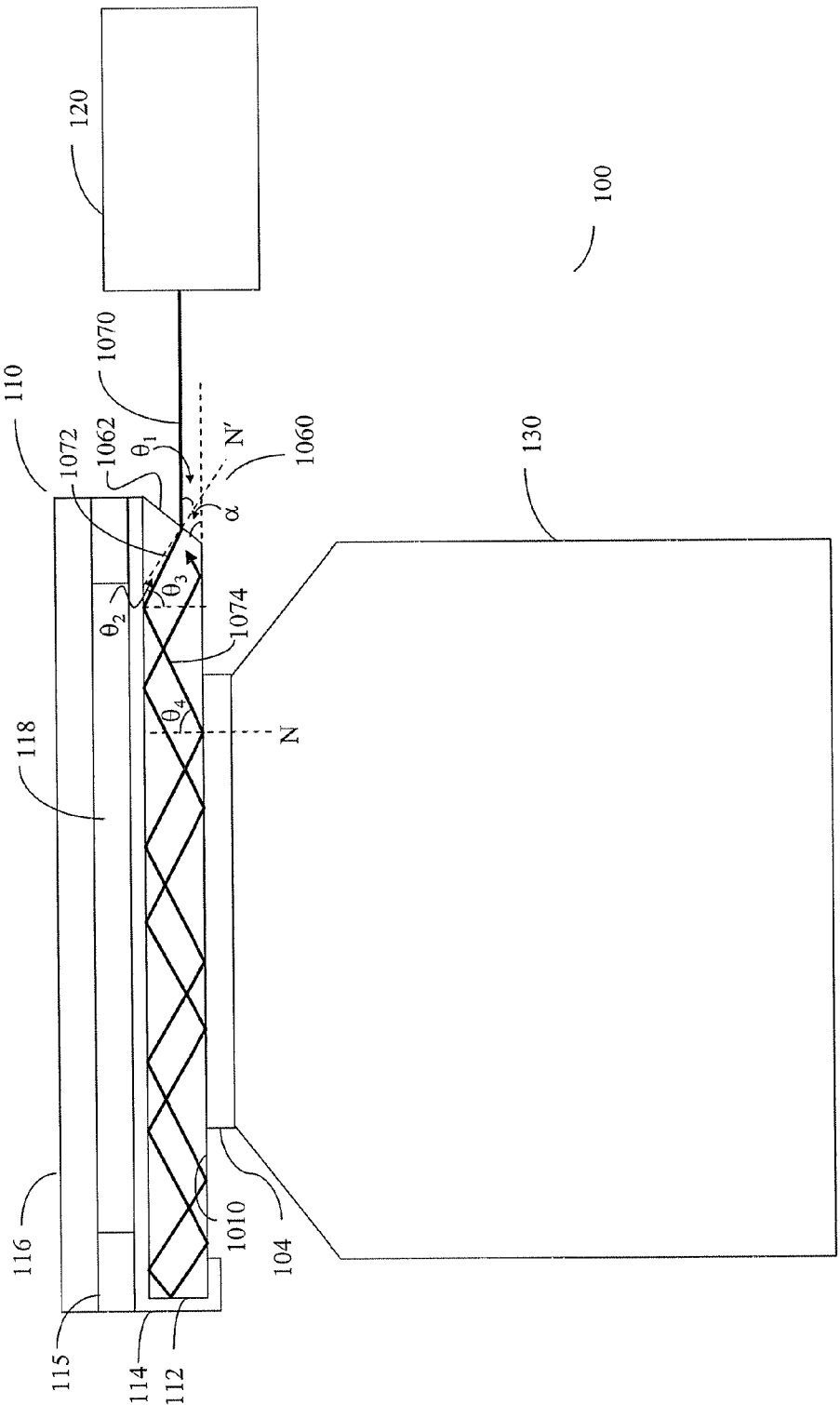


FIG. 10D

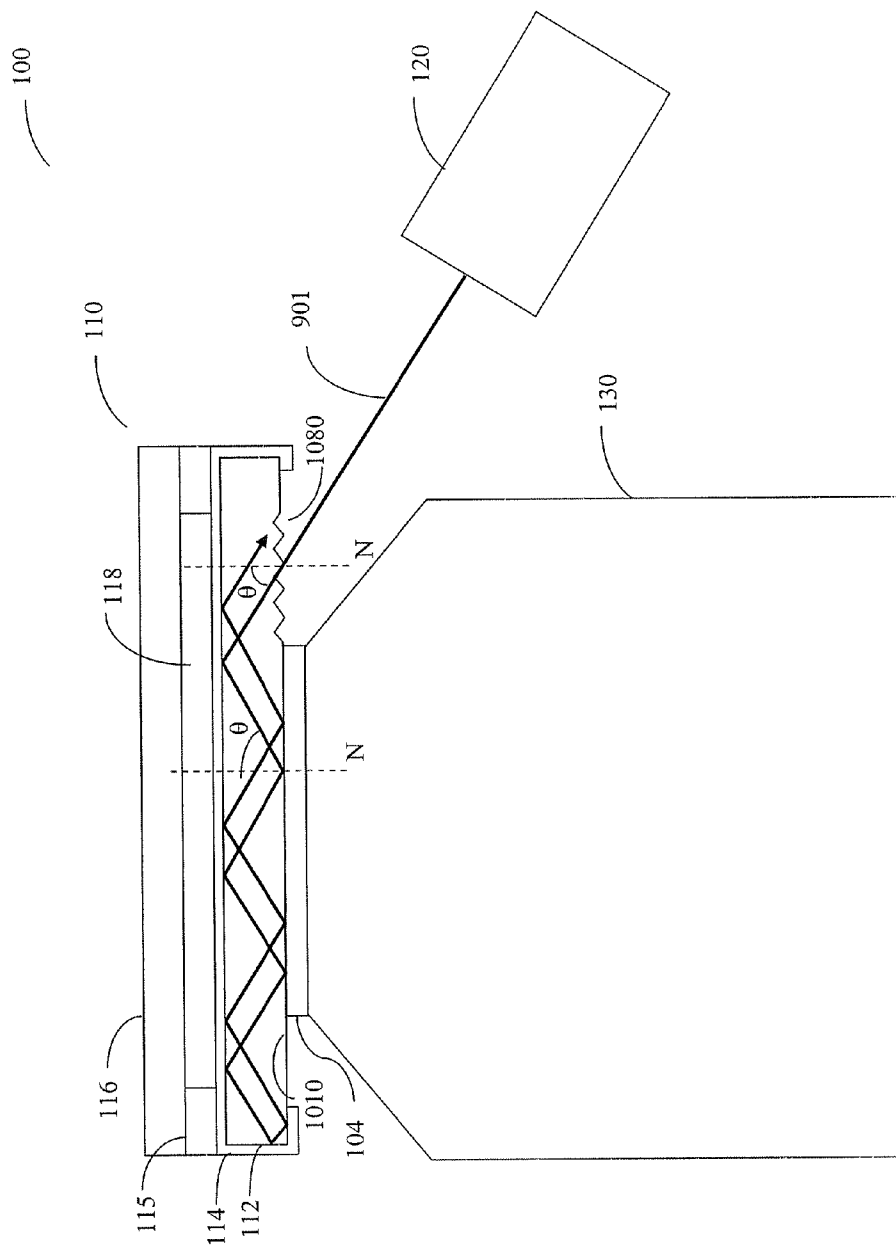


FIG. 10E

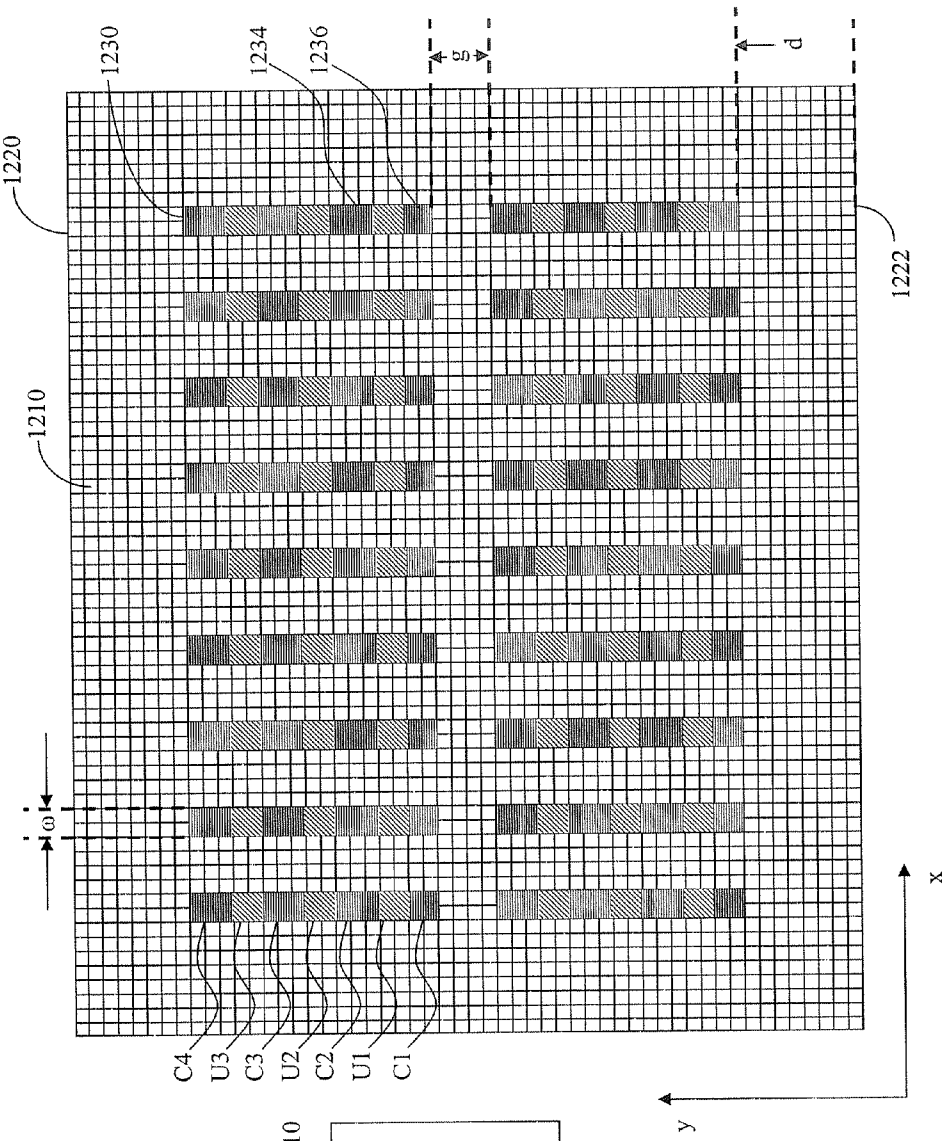


FIG. 11

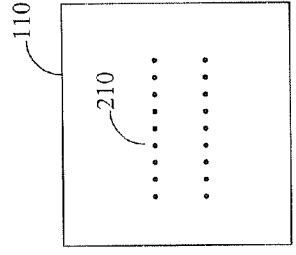


FIG. 12

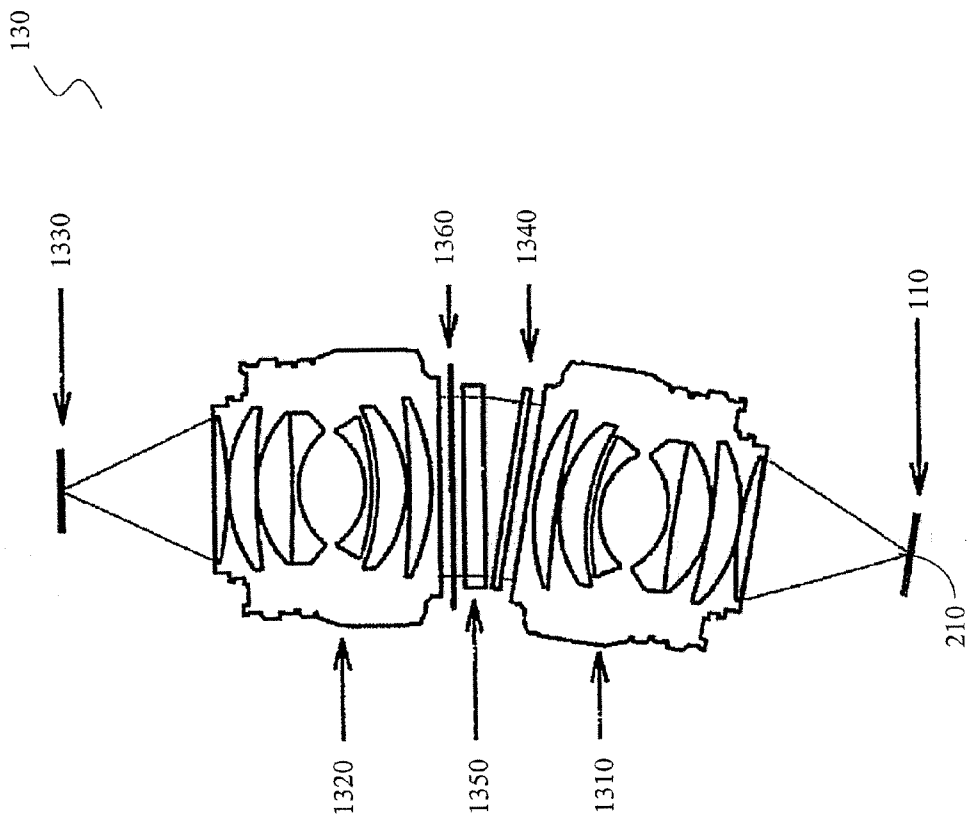


FIG. 13

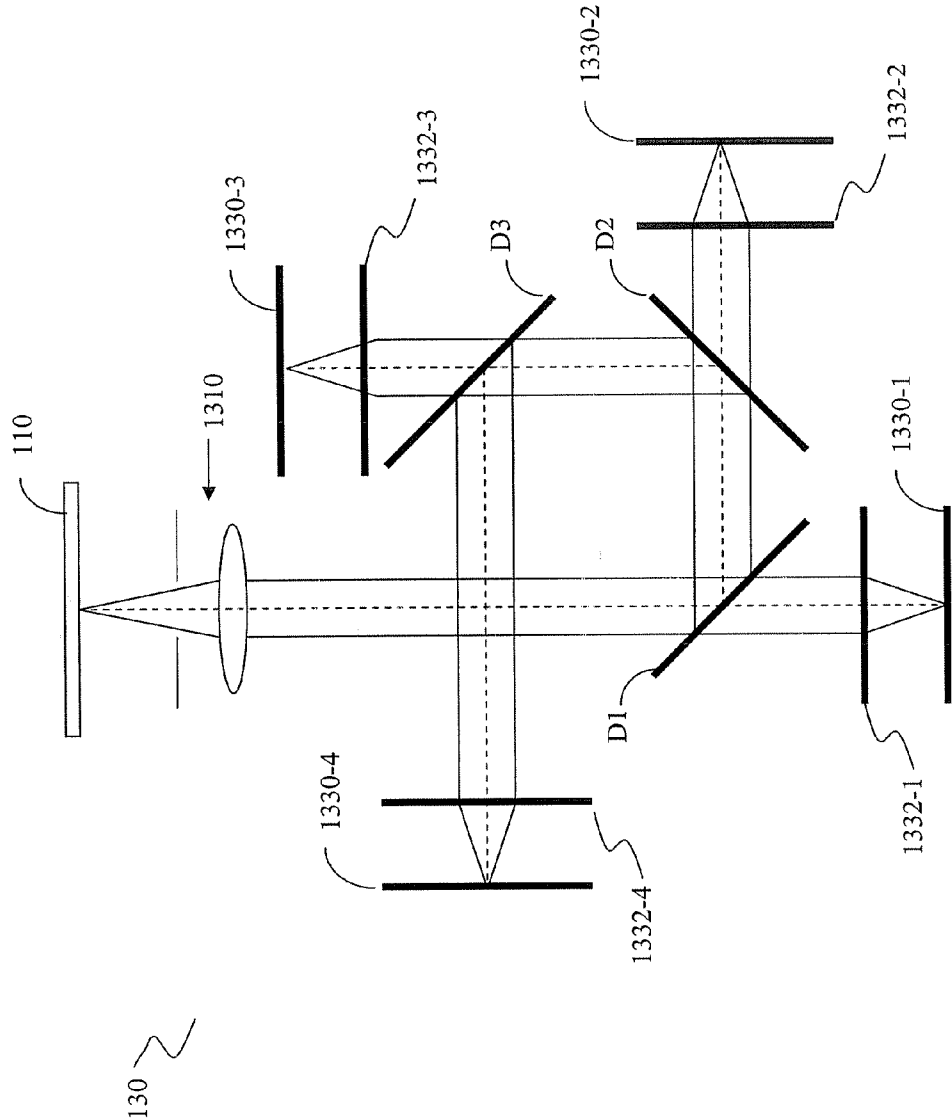


FIG. 14A

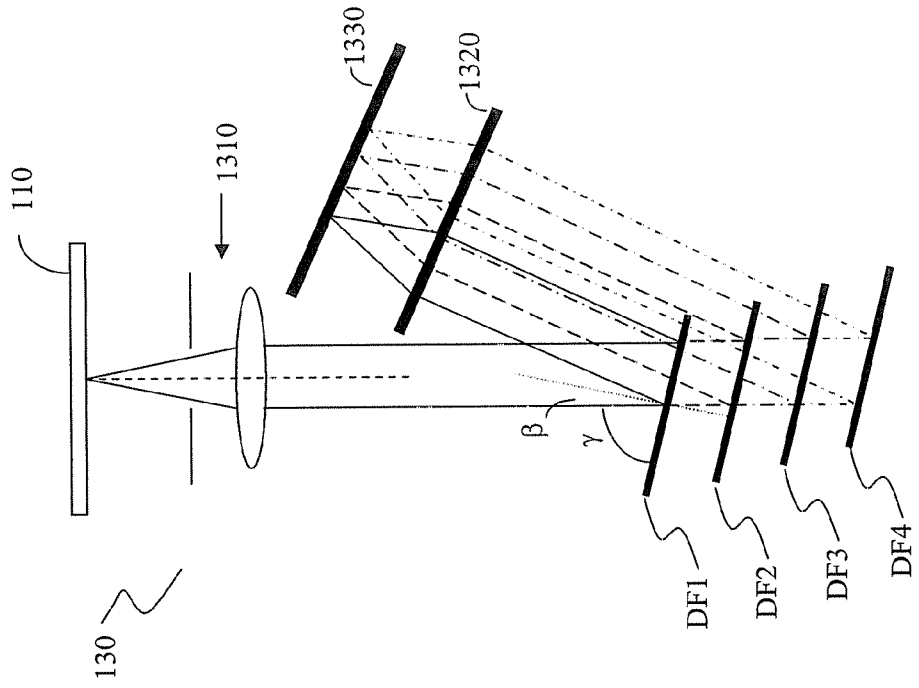


FIG. 14C

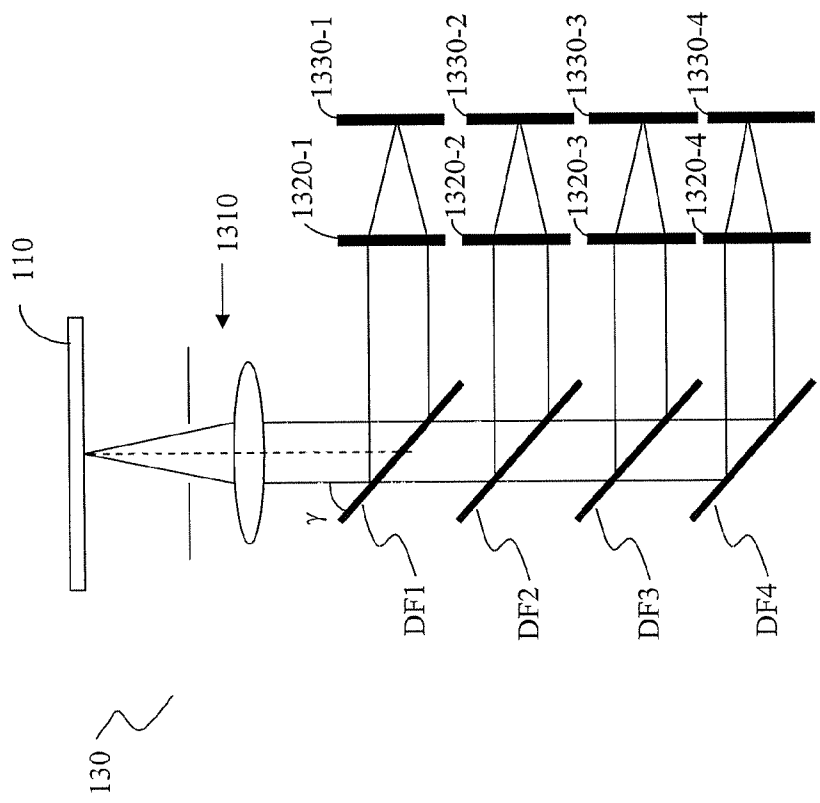


FIG. 14B

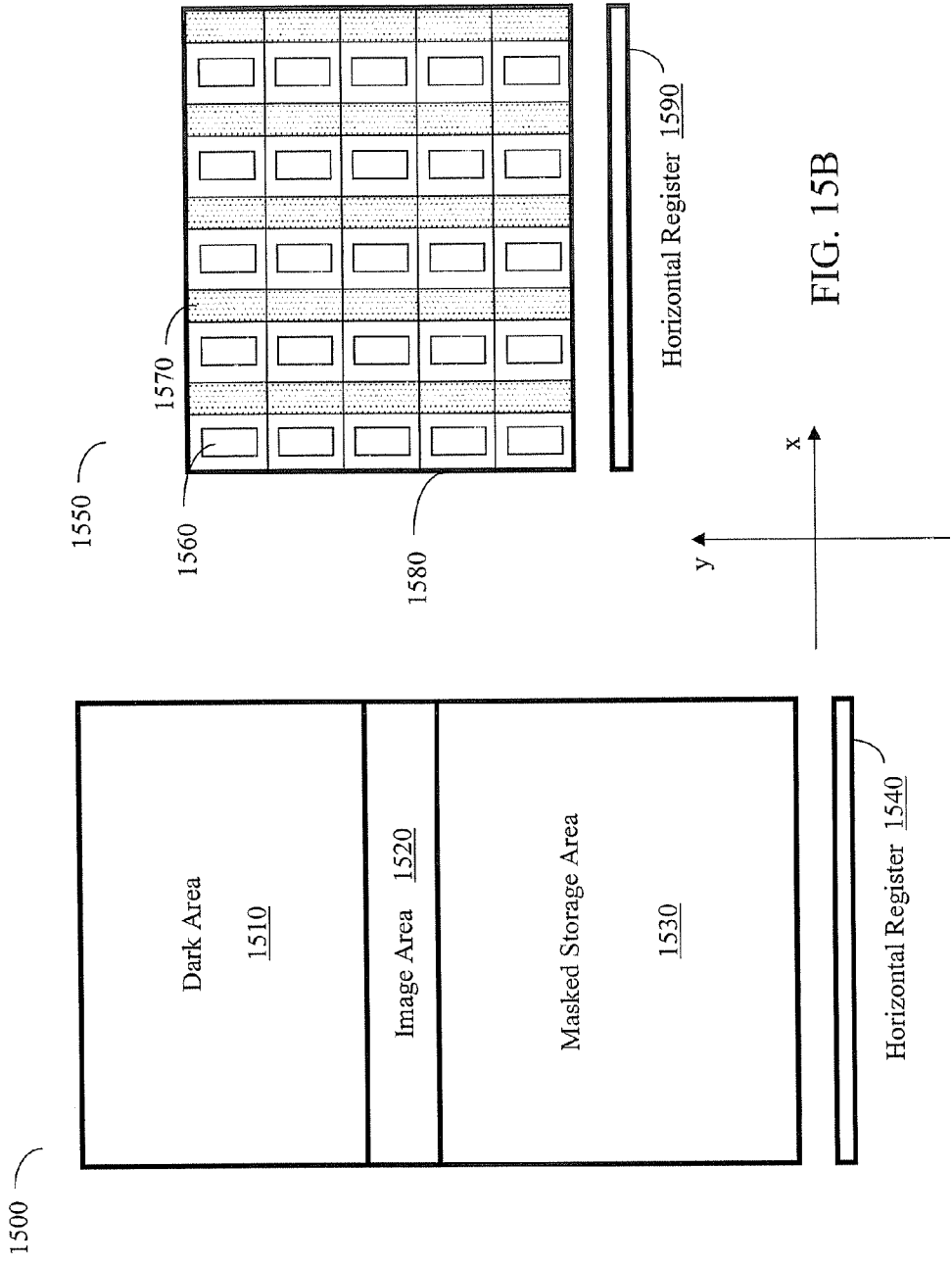


FIG. 15B

FIG. 15A

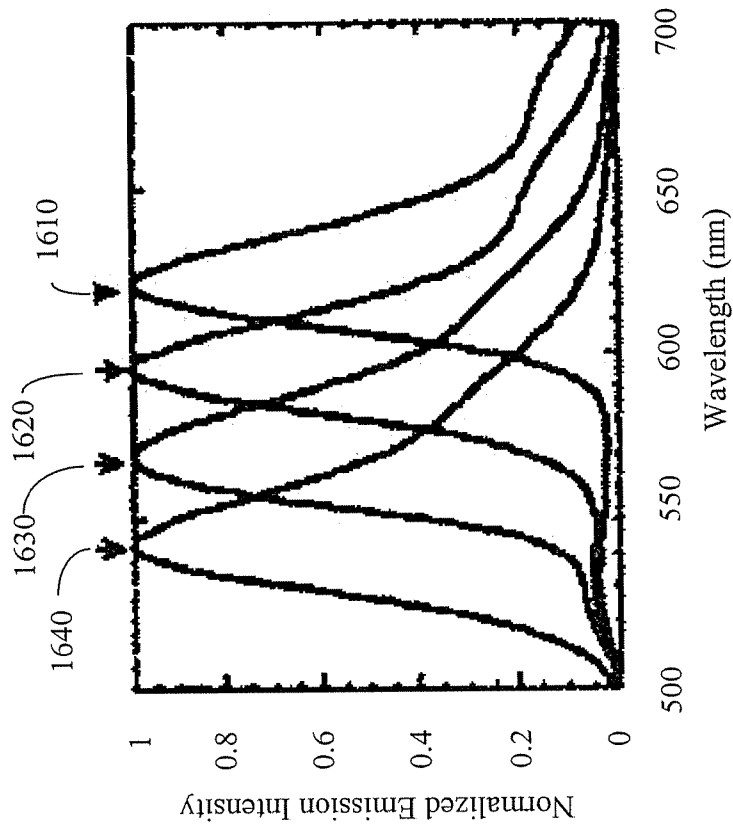


FIG. 16A

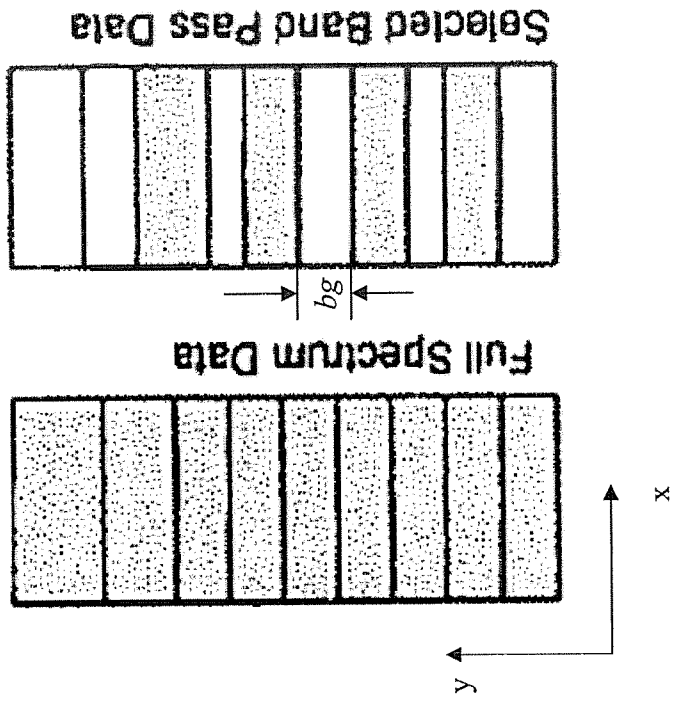


FIG. 16B

FIG. 16C

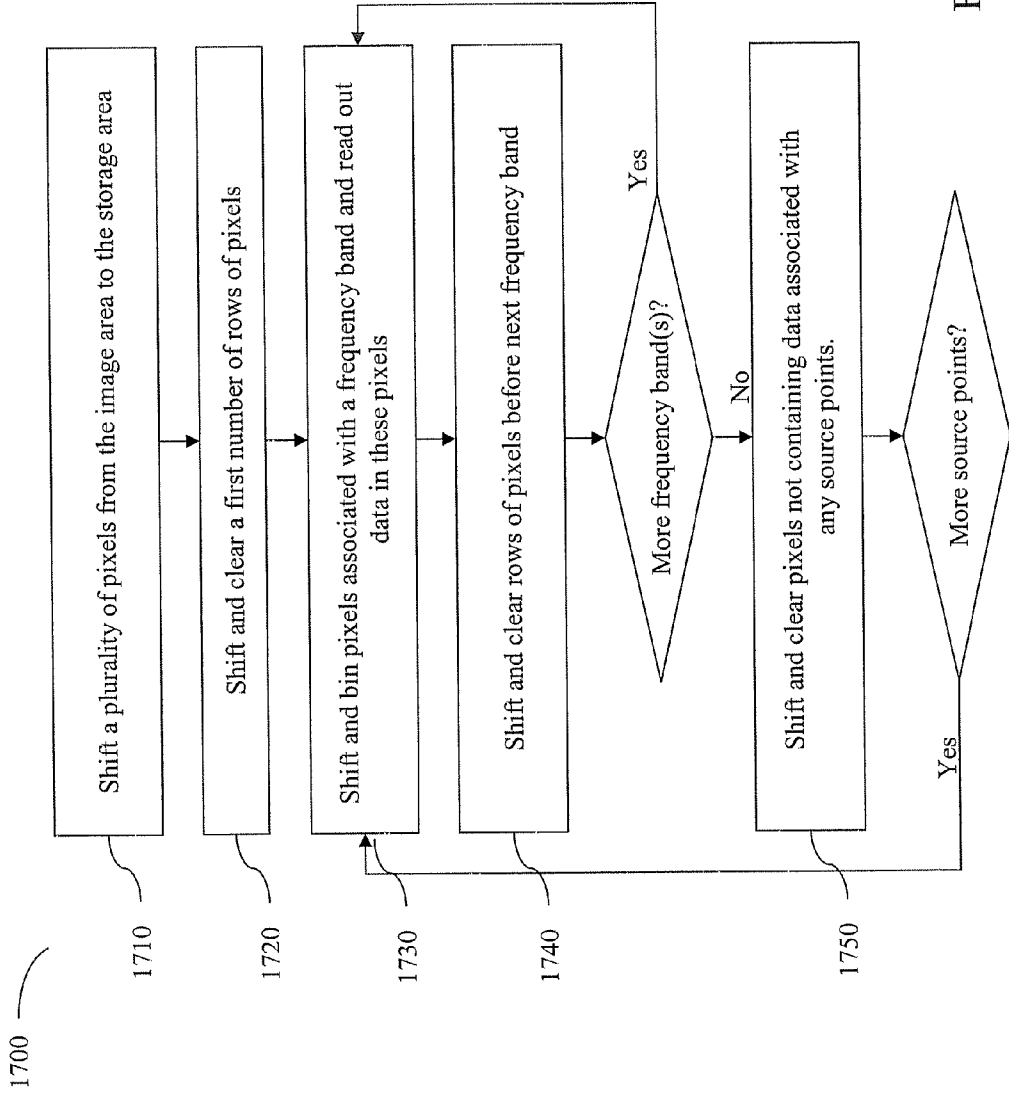


FIG. 17A

Source Point Thruput (Bold items are adjustable)	
CCD Specifications	
Horizontal Pixels =	1004
Vertical Pixels =	1002
Readout Time =	2.86E-08 sec
Vertical shift Time =	5.00E-07 sec
Horiz Reg Clear Time =	NA sec
Source Points Image	
Pitch between source points (p) =	4 pixels
Buffer rows (d) cleared =	8 rows
Bin color =	6
Colors Read =	4
Gap rows cleared =	4 rows
Bin gap or band gap (bg) =	4 rows
Source Point Gap (g) =	8 rows
Rows of source points =	4
Clear =	2.87E-05 sec
Readout Time per source point =	
Readout Time =	0.000282171 sec
# source points =	1004
% good =	25%
# good source points =	251
Polymerase Rate =	
Thruput =	100 bp/sec
	2168.64 Mbp/day
Data rate	
	886389 Bytes/sec
Number of rows/read =	
	224

FIG. 17C

Source Point Thruput (Bold items are adjustable)	
CCD Specifications	
Horizontal Pixels =	512
Vertical Pixels =	512
Readout Time =	1.00E-07 sec
Vertical shift Time =	4.00E-07 sec
Horiz Reg Clear Time =	5.00E-06 sec
Source Point Image	
Pitch between source points (p) =	4 pixels
Buffer rows (d) cleared =	8 rows
Bin color =	3
Colors Read =	4
Gap rows cleared =	2 rows
Bin gap or band gap (bg) =	2 rows
Source Point Gap (g) =	4 rows
Rows of source points =	4
Readout Time per source point =	
Readout Time =	0.000241 sec
# Source points =	512
% good =	25%
# good source points =	128
Polymerase Rate =	
Thruput =	100 bp/sec
	1105.92 Mbp/day
Data rate =	
	530240 Bytes/sec
Number of rows/read =	
	128

FIG. 17B

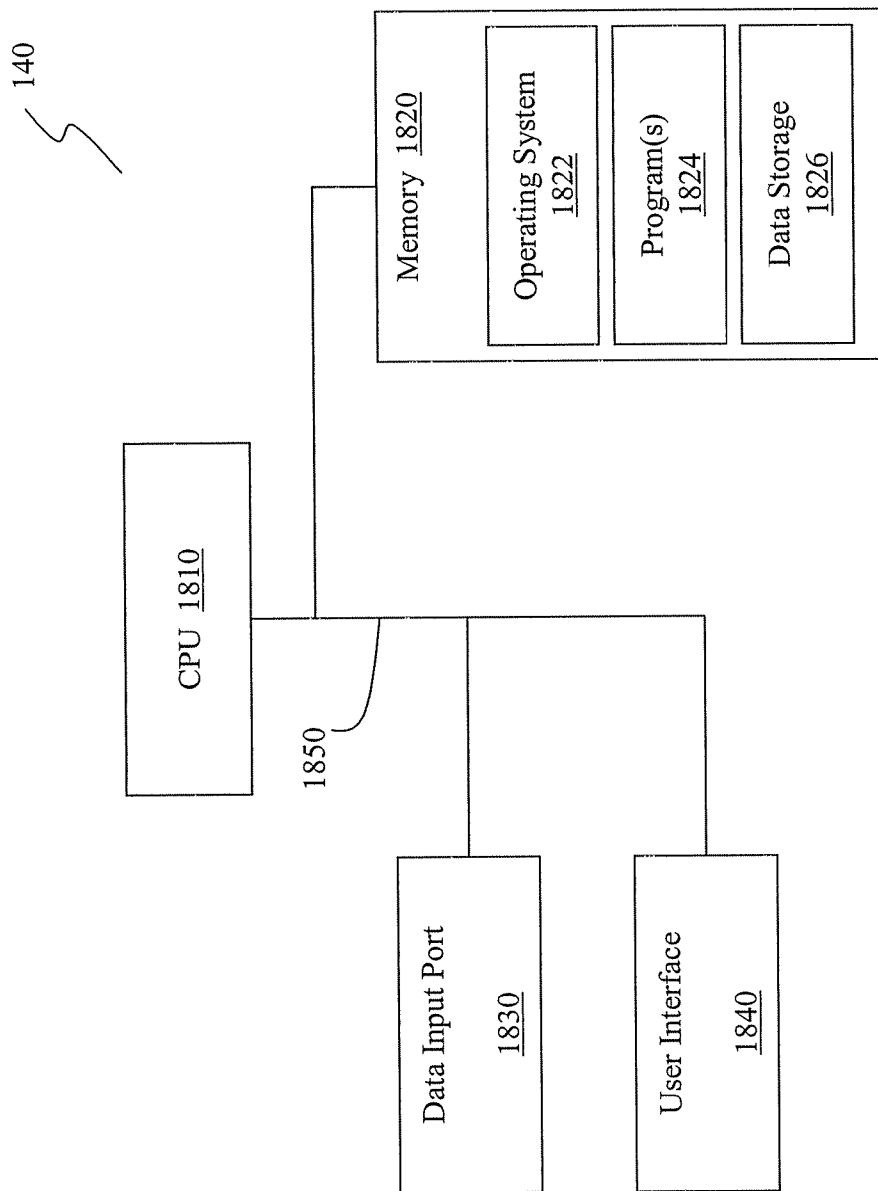


FIG. 18

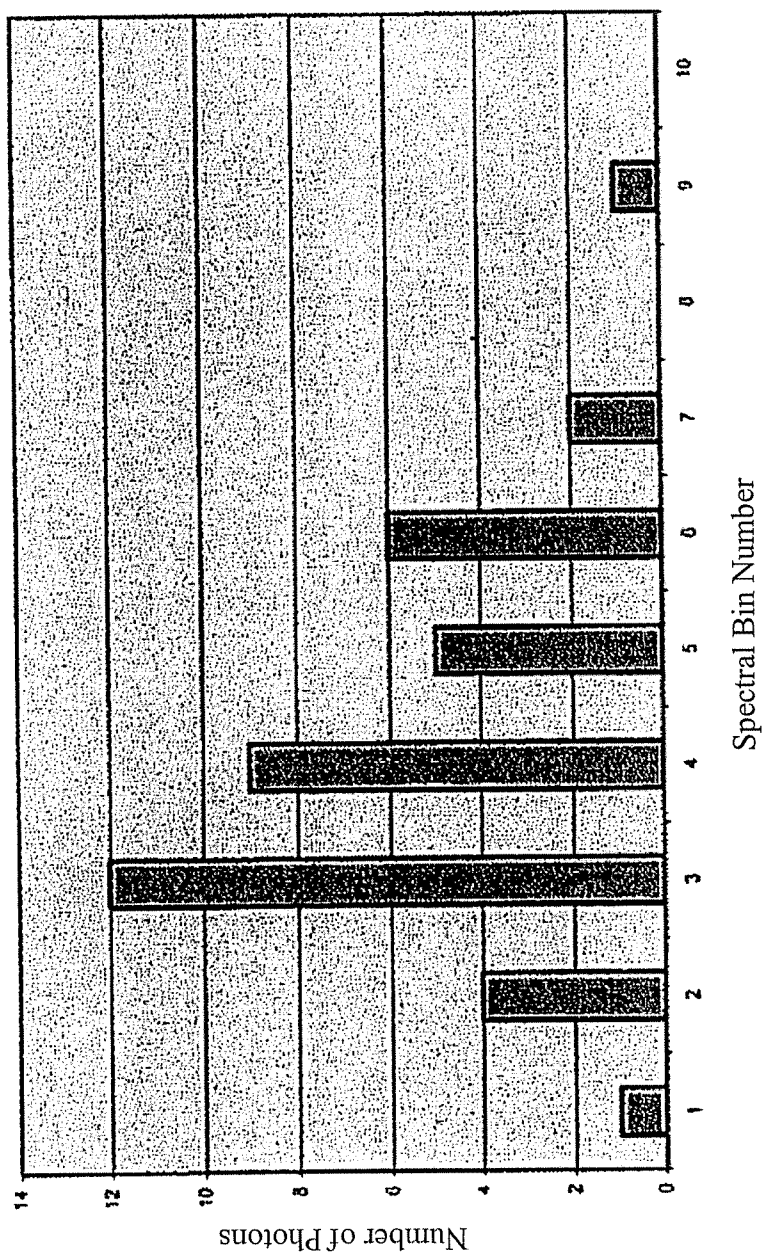


FIG. 19

2000

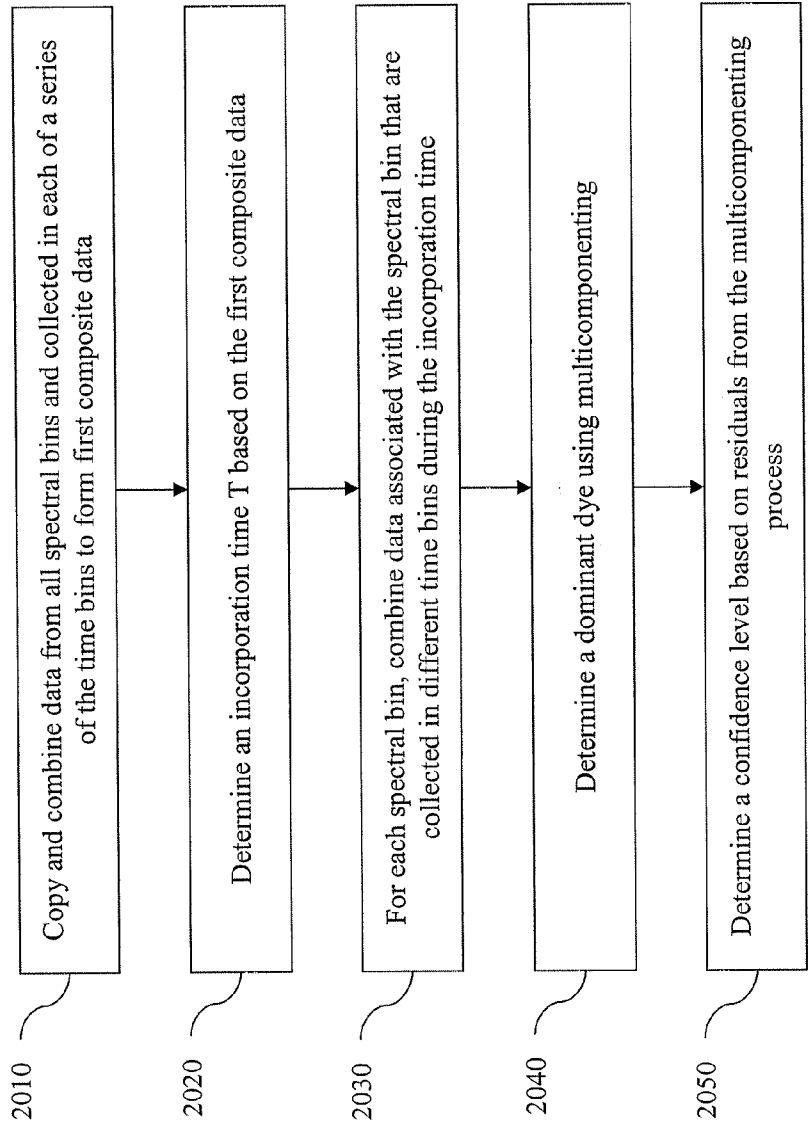


FIG. 20

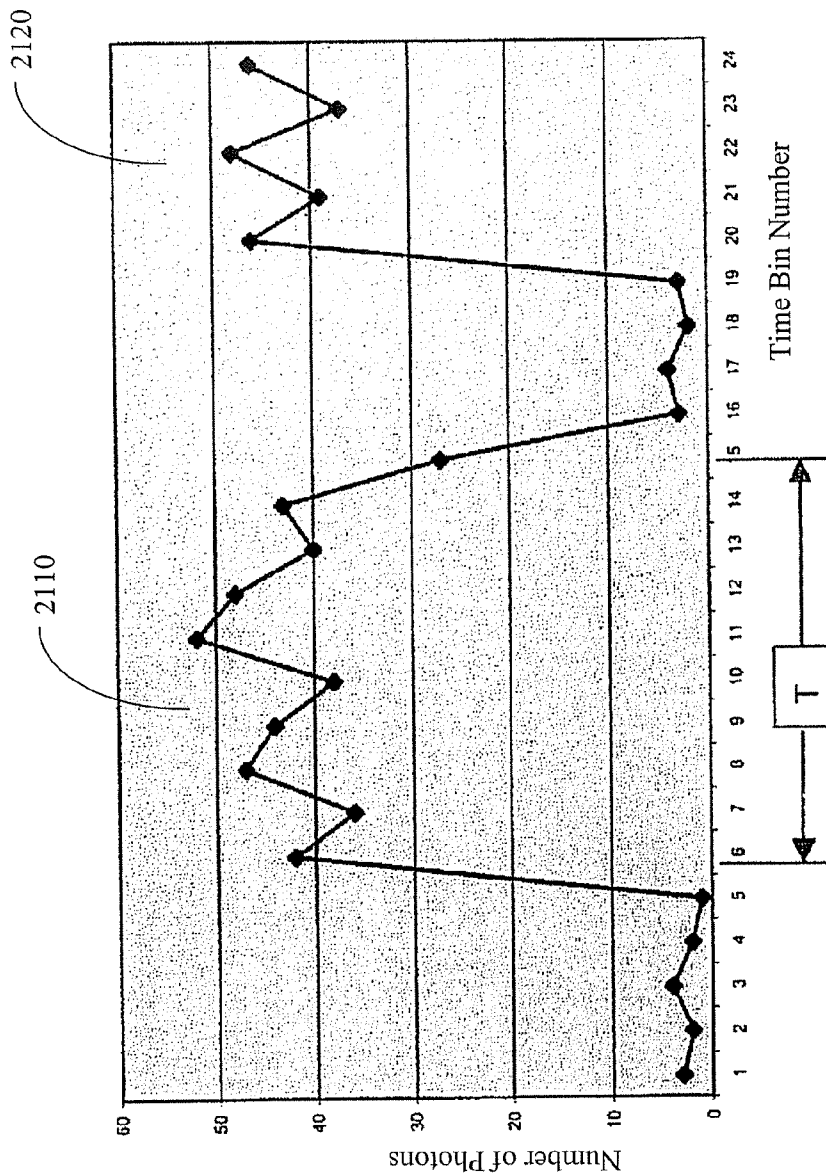


FIG. 21

METHOD AND SYSTEM FOR MULTIPLEX GENETIC ANALYSIS

CROSS REFERENCE TO RELATED APPLICATIONS

[0001] This application claims a priority benefit under 35 U.S.C. §119(e) from U.S. Provisional Application No. 60/689,692 filed Jun. 10, 2005, which is incorporated herein by reference.

FIELD

[0002] The present application relates to molecular analysis, and more particularly to methods and systems for multiplex genetic analysis of single molecule nucleic acid synthesis.

INTRODUCTION

[0003] The information stored in a DNA molecule depends on particular sequences of nucleotides, which are bases or building blocks of the DNA molecule. DNA sequencing allows the determination of the nucleotide sequence of a particular DNA segment. A conventional method of DNA sequencing starts with a defined fragment of a DNA molecule as a template. Based on this template, a population of molecules differing in size by one base of a known composition is generated. The population of molecules is then fractionated based on size using, for example, acrylamide or agarose gel electrophoresis of single-stranded DNA molecules. The base at the truncated end of each of the fractionated molecules is thereafter determined to establish the nucleotide sequence.

[0004] A sequencing method called dideoxy sequencing was developed by Fred Sanger. His method is based on DNA synthesis in the presence of dideoxy nucleotides (ddNTP), which differ from normal deoxynucleotides (dNTP) in that they lack a 3'-hydroxyl group so that once a dideoxy nucleotide is incorporated, it will terminate strand synthesis. The procedure for dideoxy sequencing starts with setting up four reactions each in a different tube containing the single strand DNA to be sequenced, labeled (tagged) primer, DNA polymerase, normal dNTPs, and a different ddNTP (i.e. for A, T, C, or G). A dideoxy nucleotide will be incorporated, randomly, at each point the corresponding nucleotide occurs in the template strand. Each time a dideoxy nucleotide is incorporated, it will stop further DNA replication. This will generate a set of fragments of various lengths, each fragment corresponding to the point at which there is a nucleotide complementary to the dideoxy nucleotide. The fragments are then separated based on their length by electrophoresis. With the smaller fragments migrating faster, the sequence can be determined by associating the base composition with each fragment.

[0005] The above technique for DNA sequencing suffer from the disadvantage that sample preparation is relatively complex in order to ensure that the tubes contain the same DNA molecules or fragments of the same DNA molecules to be sequenced. This leads to increased costs and the possibility of error. A simpler method results if molecule-based investigation techniques are used to observe the synthesis of a single DNA molecule. Because only one molecule is being observed, there is no need to ensure that all of the surrounding molecules are the same.

[0006] Specialized tools for imaging and spectroscopy have been developed to characterize nanomaterials and nanomaterials-related phenomenon. Techniques for constructing these tools comprise near-field scanning optical microscopy (NSOM) and single molecule spectroscopy (SMS). These techniques offer unique capabilities for investigating properties at the molecular level owing to their high spatial resolution, chemical sensitivity, and their ability to determine dynamical properties such as molecule binding/unbinding kinetics and the structural dynamics of polymers. For example, a sample-scanning confocal fluorescence microscope using SMS developed by McNeil et al. has demonstrated spatial resolution of ~400 nm, and single molecule sensitivity. It uses a detector system having a single-photon avalanche diode and a sensitive TE-cooled CCD spectrometer, permitting the ability to monitor fluorescence in the range of 400 to 1100 nm at a resolution of 20 nm and the ability to conduct time-lapse fluorescence spectroscopy with single molecule sensitivity.

[0007] The single-molecule techniques described above, however, often employ femtoliter-scale observation volumes and require the use of picomolar to nanomolar sample concentrations to ensure that on average only one molecule will be present in the sample volume. These concentrations are far lower than those that normally occur in nature. Thus, molecule dynamics that are affected by concentration cannot be suitably tested using the techniques. To overcome the deficiencies of NSOM and SMS techniques, other developments have been proposed. For example, Levene et al. describes a device for single molecule analysis employing a sample plate, which has 50 nm-diameter holes in a 100 nm thick aluminum film on a fused silica coverslip. When the holes are illuminated from under the fused silica coverslip, the holes act as zero-mode waveguides prohibiting the light from going through the aluminum film because the diameter of the holes are much smaller than the wavelength of the light, which is about 400-700 nm. The light, however, does generate an evanescent field that extends about 10 nm into the cavity of each illuminated hole producing a zeptoliter-scale effective observation volume near the opening of the hole. See Levene et al., US Patent Application Publication Number 2003/017492 A1, which is incorporated herein by reference.

[0008] The small observation volume provided by the zero-mode waveguides described by Levene, however, raises other challenges spanning from sample preparation, signal detection, noise or background suppression, data collection and data analysis algorithms. Accordingly, significant further developments are needed.

[0009] The present teaching in one aspect comprises an affordable high-sensitivity and high-throughput system and method for single-molecule analysis that performs at a lower cost relative to conventional systems used in sequencing, resequencing, and SNP detection. These and other features of the present teaching are set forth herein.

SUMMARY

[0010] The present disclosure provides apparatus, systems and method for analyzing a plurality of molecules by detecting separately and substantially simultaneously light emissions from a plurality of localized light-emitting analytes each including a single one of the plurality of molecules. The detected light emissions, after being properly analyzed, can be used to deduce the structure or properties

of each of the plurality of molecules. In some embodiments, the apparatus, systems and methods can be used for nucleic acid sequencing, nucleic acid resequencing, and/or detection and/or characterization of single nucleotide polymorphism (SNP analysis) including gene expression.

[0011] In various embodiments, the present invention can provide an apparatus for sequencing a plurality of target nucleic acid molecules including a sample holder configured to separate and confine a plurality of source points each including a single one of the target nucleic acid molecules, a fraction of a nucleic acid molecule, or a nucleic acid polymerizing enzyme molecule, a light source configured to direct excitation light toward the sample holder at an angle with respect to a normal of the sample holder, the excitation light illuminating the source points and causing the source points to fluoresce, at least one detector, and an optical assembly configured to collect fluorescent signals from illuminated source points to form images of the source points on the at least one detector.

[0012] In various embodiments, the present invention can provide a method for sequencing a plurality of target nucleic acid molecules, including subjecting a plurality of source points of a sample holder to nucleic acid polymerization reactions, wherein the source points each include fluorescence-labeled bases, primers, and at least one nucleic acid polymerizing enzyme molecule, and wherein the plurality of source points each has a single one of the target nucleic acid molecules, directing excitation light toward the sample holder at an angle with respect to a normal of the sample holder to illuminate the source points and to cause the source points to fluoresce, and collecting fluorescent signals from the illuminated source points and focusing the fluorescent signals onto at least one detector to form images of the source points on the at least one detector to determine time sequences of base incorporations in the polymerization reactions.

[0013] In various embodiments, the present invention can provide a method for sequencing a plurality of target nucleic acid molecules, including subjecting a plurality of source points of a sample holder to nucleic acid polymerization reactions, wherein the source points each include fluorescence-labeled bases, primers, and at least one of the target nucleic acid molecules, and wherein the plurality of source points each has a single nucleic acid polymerizing enzyme molecule, directing excitation light toward the sample holder at an angle with respect to a normal of the sample holder to illuminate the source points and to cause the source points to fluoresce, and collecting fluorescent signals from the illuminated source points and focusing the fluorescent signals onto at least one detector to form images of the source points on the at least one detector to determine time sequences of base additions in the polymerization reactions.

[0014] In various embodiments, the present invention can provide a method for sequencing a plurality of target nucleic acid molecules, including enriching a sample holder with a plurality of source points each having a single one of the target nucleic acid molecules and/or a single nucleic acid polymerizing enzyme molecule, subjecting the plurality of source points to nucleic acid polymerization reactions, (1) wherein when the source points have a single one of the target nucleic acid molecules, the source points each further include fluorescence-labeled bases, primers, and at least one nucleic acid polymerizing enzyme molecule, (2) wherein when the source points have a single nucleic acid polymer-

izing enzyme molecule, the source points each further include fluorescence-labeled bases, primers, and at least one of the target nucleic acid molecules, and (3) wherein when the source points have a single one of the target nucleic acid molecules and a single nucleic acid polymerizing enzyme molecule, the source points each further include fluorescence-labeled bases and primers, directing excitation light toward the sample holder at an angle with respect to a normal of the sample holder to illuminate the source points and to cause the source points to fluoresce, and collecting fluorescent signals from the illuminated source points and focusing the fluorescent signals onto at least one detector to form images of the source points on the at least one detector to determine time sequences of base incorporations in the polymerization reactions.

[0015] A system according to exemplary embodiments of the present disclosure comprises a sample holder having structures formed thereon for spatially separating and constraining a plurality of light-emitting analytes each having a single one of the plurality of molecules to be analyzed. In exemplary embodiments, each of the plurality of molecules is a single nucleic acid molecule, a fraction of the nucleic acid molecule, an oligonucleotide molecule, or a single nucleic acid polymerizing enzyme. The system further comprises a light source configured to illuminate the sample holder, an optical assembly configured to collect and detect separately and substantially simultaneously light emissions associated with the plurality of light emitting analytes. The system may further include a computer system configured to analyze the light emissions to determine the structures or properties of a target nucleic acid molecule associated with each analyte.

[0016] In one exemplary embodiment of the present invention, the light source is configured to produce excitation light that is directed toward the sample holder at an angle with respect to a normal of a plane associated with the sample holder. In further embodiments, excitation light is directed toward the sample holder such that total internal reflection occurs and the excitation light is recycled multiple times before exiting the sample holder.

[0017] In one exemplary embodiment of the present invention, the optical assembly comprises at least one pixelated sensor device such as a charge coupled device (CCD) detector or CMOS detector configured to detect substantially simultaneously light emissions from the multiple localized light-emitting analytes. In further embodiments, the optical assembly is configured to disperse spectrally the light emitted from the multiple localized light-emitting analytes onto the detector(s) so that different frequency bands of the emitted light are detected by different areas of the detector (s).

[0018] These and other features of the present teaching are set forth herein.

BRIEF DESCRIPTION OF THE DRAWINGS

[0019] The skilled artisan will understand that the drawings, described below, are for purposes of illustration only, and are not intended to limit the scope of the present teaching in any way.

[0020] FIG. 1 is a block diagram of an exemplary embodiment of a high throughput system for single molecule analysis.

[0021] FIG. 2 is a top view of a sample holder in the system.

[0022] FIG. 3 is a cross-sectional view of a portion of the sample holder.

[0023] FIG. 4 is a flowchart illustrating an exemplary embodiment of a method useful for enriching the sample holder with a plurality of spatially restrained source points.

[0024] FIG. 5 is a diagram illustrating further embodiments of a method useful for enriching the sample holder with a plurality of spatially constrained source points.

[0025] FIG. 6 is a 3-dimensional view of a portion of the sample holder according to further embodiments of the present teaching.

[0026] FIG. 7 is a diagram illustrating a DNA sequencing process along a channel on the sample holder according to exemplary embodiments of the present teaching.

[0027] FIGS. 8A and 8B are flowcharts illustrating further embodiments of a method useful for placing a plurality of source points on the sample holder in FIG. 6.

[0028] FIGS. 8C and 8D are diagrams illustrating embodiments of a setup for stretching each of a plurality of oligonucleotide molecules along a bottom surface of a channel on the sample holder.

[0029] FIG. 9 is a block diagram illustrating exemplary embodiments of an optical arrangement useful for illuminating the source points placed on the sample holder.

[0030] FIGS. 10A-10E are block diagrams illustrating further embodiments of an optical arrangement useful for illuminating the source points placed on the sample holder.

[0031] FIG. 11 is top view of a sample holder showing a plurality of source points.

[0032] FIG. 12 is a top view of an image plane in a detector in the system in FIG. 1 according to exemplary embodiments.

[0033] FIG. 13 is a diagram illustrating an optical assembly for detecting light signals from the source points on the sample holder according to exemplary embodiments.

[0034] FIGS. 14A-14C are diagrams illustrating further embodiments of an optical assembly for detecting light signals from the source points on the sample holder.

[0035] FIG. 15A is a block diagram illustrating a frame transfer CCD array in a detector in the system in FIG. 1 according to exemplary embodiments of the present teaching.

[0036] FIG. 15B is a block diagram illustrating an inter-line CCD array in the detector in the system in FIG. 1 according to exemplary embodiments of the present teaching.

[0037] FIG. 16A are examples of normalized fluorescent spectra corresponding to four different fluorescent dyes.

[0038] FIG. 16B is a block diagram illustrating full spectrum data.

[0039] FIG. 16C is a block diagram of the full spectrum data with the most informative wavelengths distinguished from less informative wavelengths.

[0040] FIG. 17A is a flowchart illustrating a method for reading out CCD data associated with a plurality of source points according to exemplary embodiments.

[0041] FIGS. 17B-17C are each a spreadsheet for estimating the throughput of reading out data from a CCD array using the method in FIG. 17A.

[0042] FIG. 18 is a block diagram of a computer system used in the system in FIG. 1 according to exemplary embodiments of the present teaching;

[0043] FIG. 19 is a histogram illustrating the number of photons in different spectral bins detected from a single incorporation event;

[0044] FIG. 20 is a flowchart of an exemplary embodiment of a method useful for base determination according to the present teaching; and

[0045] FIG. 21 is a plot of composite data over a plurality of time bins according to exemplary embodiments of the present teaching.

DESCRIPTION OF VARIOUS EMBODIMENTS

[0046] It is to be understood that both the foregoing summary and the following description of various embodiments are exemplary and explanatory only and are not restrictive of the present teachings. In this application, the use of the singular comprises the plural unless specifically stated otherwise. Also, the use of "or" means "and/or" unless stated otherwise. Similarly, "comprise," "comprises," "comprising," "comprise," "comprises," and "including" are not intended to be limiting.

[0047] Additionally, while certain embodiments are described in detail herein, particularly embodiments suitable for analysis of single molecule nucleic acid synthesis, it is to be understood the apparatus, systems and methods of the present disclosure may be employed in other applications for analysis of single molecules, such as but not limited to directed resequencing, SNP detection, and gene expression.

[0048] Furthermore, the figures in this application are for illustration purposes and many of the figures are not to scale with corresponding hardware. Many parts of the features in the figures in this application are drawn out of scale purposefully for ease of illustration.

[0049] Systems according to some embodiments of the present disclosure generally comprise a sample holder configured to hold a plurality of localized light-emitting analytes each comprising a single one of a plurality of molecules to be analyzed, a light source configured to illuminate the sample holder, and an optical assembly configured to collect and detect light emitted from the source points.

[0050] FIG. 1 is a block diagram of an exemplary embodiment of a system 100 for detecting and analyzing light emitted from the plurality of light-emitting analytes. As shown in FIG. 1, system 100 comprises a sample holder 110, a light source 120, and an optical assembly 130. System 100 may further comprise a host computer system 140 (see also FIG. 18) configured to analyze optical data detected by optical assembly 130. System 100 may also comprise one or more digital signal processors (DSP) or field programmable gate arrays (FPGA) 150 coupled between optical assembly 130 and host computer system 140. DSPs or FPGAs 150 can be used to execute algorithms for base determination, as explained in more detail below.

[0051] System 100 may optionally comprise an index-matching prism 108. A space between the sample holder 110 and the optical assembly 130 may be filled with a fluid 104. The utility of the index-matching prism 108 and the fluid 104 is discussed below. Although for reasons discussed below, it may be advantageous to direct the excitation light from the light source 120 to the sample holder 110 at an angle, as shown by the solid line 121 in FIG. 1, system 100 is not limited to such use, and the excitation light can be directed via a dichroic filter 125 toward the sample holder along a normal N of the sample holder 110, as shown by the dashed lines in FIG. 1.

[0052] In exemplary embodiments of the present teaching, sample holder **110** is configured to support and confine the plurality of light-emitting analytes. For ease of discussion, each localized light-emitting analyte will hereafter be referred to as a dye or a “source point”. In various embodiments, a dye or a source point comprises a single nucleic acid molecule, a fraction of a nucleic acid molecule, an oligonucleotide molecule, or a single nucleic acid polymerizing enzyme. The dye or source point may also comprise one or more other molecules, constituents, or reactants. The emitted light from the complex can be used to deduce the structure or properties of a target nucleic acid molecule.

[0053] In one exemplary embodiment, in applications employing nucleic acid sequencing, each source point is a complex of a single nucleic acid polymerizing enzyme, a target nucleic acid molecule, and at least one incorporated or incorporating fluorescence-labeled nucleotide analog. The source point is localized or spatially constrained in at least one dimension that is less than the wavelength of the excitation light. The fluorescent label on the nucleotide analog emits fluorescent light upon illumination by light source **120**. In exemplary embodiments of the present teaching, four different nucleotide analogs are labeled with four different fluorescent dyes each having a unique emission spectrum. The four different fluorescent dyes can also be associated with four different frequency bands each corresponding to a peak in emission intensity according to the respective spectrum. The four different frequency bands are hereafter referred to as first, second, third, and fourth frequency bands.

[0054] Thus, the time sequence of base incorporation can be observed by detecting fluorescent signals from sequentially incorporated nucleotide analogs associated with a source point. The fluorescent light signals from different source points on the sample holder **110** are substantially simultaneously collected and detected by optical assembly **130** and are analyzed by computer system **140** to determine the identities of the incorporated nucleic acid molecule in each of the source points. To reduce or eliminate interference between fluorescent signals associated with consecutive incorporation events on a same source point, after detection of an incorporation event, fluorescent label on the newly incorporated nucleotide can be bleached, cleaved or otherwise removed with a known technique. Photo-cleavable linkers may be utilized to facilitate efficient and consistent removal of the fluorescent labels.

[0055] In some embodiments, the source points are localized or spatially constrained at different locations on sample holder **110** by immobilizing the single nucleic acid molecule or the single nucleic acid polymerizing enzyme in each source point at one of the locations. This allows separate and substantially simultaneous detection of fluorescent emission from the plurality of source points. A conventional method or one of the methods discussed below can be used to immobilize the enzymes or the template nucleic acid molecules.

[0056] FIG. 2 is a block diagram of a top-down view of sample holder **110** according to exemplary embodiments. As shown in FIGS. 1 and 2, in exemplary embodiments, sample holder **110** comprises a substrate **112** made of a material transparent to the excitation light from light source **120** and to the fluorescent emissions from the source points. A metallic film **114** is formed on a top surface of substrate **112**. Depending on specific applications, for reasons discussed

below, metallic film **114** may extend to the side surfaces and edge portions of a bottom surface of the substrate **112**, as shown in FIG. 1. Sample holder **110** may further comprise a sealer **115** and a cover **116** for evaporation control. A space **118** is formed between the cover **116** and the substrate **112**, which space serves as a sample chamber for holding a sample fluid that supplies at least one of the constituents or reactants in each source point. In various embodiments, in applications of nucleic acid sequencing, the sample fluid comprises a fluorophore solution of different types of fluorescent-labeled nucleotides. Sample holder **110** may further comprise a fill hole **230** for filling the sample chamber **118** with the sample fluid and a drain hole **240** for draining the sample fluid from the sample chamber. Fill hole **230** and drain hole **240** are preferably located near two opposite corners of sample holder **110**, as shown in FIG. 2, for more complete draining and washing away of sample fluid.

[0057] As shown in FIG. 2, sample holder **110** is configured to hold a plurality of spatially separated and constrained source points **210** in a field of view **220** of the optical assembly **130**. The spatial separation and confinement of the source points **210** help in one aspect to detect light signals from the source points **210** separately and substantially simultaneously. Although FIG. 2 shows that the source points **210** on the sample holder **110** are arranged in an array having two rows and a number of columns, such arrangement is not necessary as long as the source points are sufficiently spaced from each other so that the light signals from them can be effectively resolved by optical assembly **130**. When the source points are arranged in an array, the array may be perfect, meaning each array element site has an immobilized functional source point, or imperfect, meaning at least one array element site is missing a source point, has a source point that is not functional, or has multiple source points that are too close together to allow resolution by the optical assembly **130**.

[0058] In various embodiments, the metallic film **114** on the top surface of the substrate **112** has etched patterns forming cavities for housing the plurality of source points and separating the plurality of source points to allow resolution by the optical assembly **130**. In some embodiments, zero-mode waveguides, such as those described in Patent Application Number US 2003/0174992 by Levene et al, which is incorporated herein by reference, are formed in metallic film **114**, as shown in a cross-sectional view in FIG. 3. Zero-mode waveguides are known in the art and can be created using a variety of materials and methods. As a specific, non-limiting example, substrate **112** is a fused silica substrate, metallic film **114** is an aluminum film formed on the fused silica substrate, and an array of holes **310** are formed by masking and plasma etching the aluminum film to create holes **310** in the aluminum film. Each hole **310** has a diameter that is substantially smaller than a wavelength of the excitation light from light source **120** and a depth that is sufficient to block transmission of the excitation light through the hole. Thus, each hole **310** acts as a zero-mode waveguide for the excitation light from light source **120**, allowing the excitation light, which comes to the waveguides from the substrate side, to penetrate only a bottom portion **312** of the hole **310**. At the same time, the zero-mode waveguides also serves to block light emitted or scattered from the sample fluid on the sample holder **110** except emissions coming from any light emitting agents immobi-

lized in the bottom portions **312** of the waveguides or diffusing past the bottom portions **312** of the waveguides.

[0059] Thus, in some embodiments, to allow the detection and analysis of light emitted from the source points **210**, each source point **210** is immobilized in the bottom portion **312** of a zero-mode waveguide **310**, so that light emitted from the source point can escape the hole **310**, pass through substrate **112** and be collected by optical assembly **130**. Preferably, only one source point should be present in the bottom portion **310** of a hole **310** because it would be difficult for the optical assembly to distinguish the emitted light from more than one source point in a single hole **310** considering the size of the hole. Therefore, in the exemplary embodiment, holes **310** that either do not have any source point immobilized in the bottom portion **312** or have more than one source point immobilized in the bottom portion **312** do not contribute to the analysis and are considered as empty sites in an array of source points **210**.

[0060] For ease of discussion, the description hereafter will be illustrated in the context of nucleic acid sequencing, while the methods, systems and apparatus of the present teaching can be applied to other types of molecular analysis. Methods of immobilizing molecules involved in a genetic assay in waveguides **310** are described in detail in US Patent Application Number US 2003/0044781 by Korfach et al., which is incorporated herein by reference. Using the methods described by Korfach, some of the array of holes **310** can each contain a single DNA molecule or enzyme immobilized in the bottom portion **312**, while a large percentage of the holes may contain none or multiple molecules in each of them and are thus useless in the analysis.

[0061] FIG. 4 illustrates a flowchart of one embodiment of a method **400** for enriching the sample holder **110** with source points. Method **400** increases the efficiency and throughput of system **100** by maximizing the percentage of holes **310** that have in each of them a single source point in the bottom portion **312**. As shown in FIG. 4, according to exemplary embodiments of the present teaching, method **400** comprises the following steps: step **410** in which uncovered portions of the substrate **112** are coated with streptavidin, step **420** in which a dilute solution including a plurality of molecules each being a nucleic acid molecule or a nucleic acid polymerizing enzyme is applied to the waveguides **310**. Each of the plurality of molecules has a photoactivatable biotin attached to it. The concentration of the plurality of molecules in the solution is selected to be lower than the optimal Poisson distribution so that when the solution is applied to the waveguides **310** on the sample holder **110**, most of the waveguides **310** would be populated by zero number of the molecules and that statistically few of the waveguides **310** would be occupied by more than one of the molecules.

[0062] Still referring to FIG. 4, method **400** further comprises step **430** in which a first group of waveguides are identified as each being occupied by at least one nucleic acid molecule or a nucleic acid polymerizing enzyme. The first group of waveguides can be identified by using, for example, a simplified sequencing assay. Method **400** further comprises step **440** in which light is shown on each of the first group of waveguides. The light activates the biotin attachment in those waveguides and thus immobilizes the molecules using the biotin-streptavidin bound. Step **440** is followed by an optional step **450** in which the solution is removed from the sample holder by washing or inactivation,

leaving only those molecules bonded to the bottom of the waveguides. Step **450** is followed by an optional step **460** in which another dilute solution of the biotin attached molecules are applied to the waveguides **310** on sample holder **110**. Method **400** further comprises step **470** in which an additional group of waveguides are identified as each being occupied by at least one nucleic acid molecule or a nucleic acid polymerizing enzyme. In one embodiment, the additional group of waveguides do not overlap with previously identified group(s) of waveguides. Method **400** further comprises step **480** in which light is shown on each of the additional group of waveguides and thus immobilizes the nucleic acid molecules or enzymes in the additional group of waveguides. Method **400** then repeats steps **450-480** until most of the waveguides are populated by bound molecules. Note that steps **450** and **460** are optional because, instead of carrying out steps **450** and **460**, one can simply wait for a period of time to allow more of the plurality of molecules to diffuse into some of the waveguides.

[0063] Optionally, after populating the waveguides with polymerase molecules, a primer is attached to each polymerase molecule by a flexible linker. Attaching the primer to the polymerase molecule helps the analysis because the DNA template would be tethered and not float away, allowing subsequent synthesis to occur on the same template. In one aspect, this benefits the analysis by increasing read lengths and throughput. Longer read lengths help to simplify any fragment assembly problem.

[0064] In some embodiments, a method for enriching the sample holder involves the use of nanobeads. As shown in FIG. 5, the sample fluid comprise nanobeads **510**, and the enzyme or nucleic acid molecule **520** is attached to a nanobead **510** by a cleavable linker **515**, in a manner that most of the nanobeads in the sample fluid each have at most one enzyme or nucleic acid molecule attached. The nanobeads are sized such that only one nanobead is likely to fit in a waveguide **310**. The enzyme or nucleic acid molecule **520** has a photoactivatable linker **525** that allows attachment of the composite including the nanobead, the nucleic acid molecule or enzyme, and the cleavable linker **515** to an attachment site **530** at the bottom of a waveguide **310**. The attachment is activated by shining light from the bottom of substrate **112**. Since only those nanobeads each having an enzyme or nucleic acid molecule with the photoactivatable linker can bind to the substrate **312**, and the presence of a nanobead in a waveguide **310** excludes other enzyme or nucleic acid molecules from diffusing into the same waveguide, there is no need to determine which waveguides are occupied by the nanobeads before shining light on the waveguides to activate the linkers **525**. The shining of light can be repeated later when more nanobeads with enzyme or nucleic acid molecules attached thereon diffuse into other waveguides. Thus, the nanobeads can be used to increase the number of waveguides each having a single nucleic acid molecule or enzyme attached therein. After binding, the nanobeads are removed from the enzyme or nucleic acid molecules by, for example, chemically cleaving the linkers **515** or dissolving the nanobeads.

[0065] In alternative embodiments of the present teaching, sample holder **110** comprises slots or channels to facilitate confining the plurality of source points **210** on the sample holder **110**. FIG. 6 illustrates a 3-dimensional view of a plurality of channels **610** formed in metallic film **114** on substrate **112**. As a non-limiting example, channels **610** are

formed in an aluminum film over a fused silica substrate. Each channel **610** has a width w that is smaller than a wavelength associated with light source **120**. In exemplary embodiments of the present teaching, light from light source **120** is linearly polarized and the polarization direction is oriented with the electric field vector in the light wave along the width direction of the channels. Thus, only a bottom portion **612** in each channel **610** would be illuminated by the excitation light from light source **120**, as shown in FIG. 6. Channels **610** can be formed using conventional techniques, such as conventional semiconductor processing or integrated circuit (IC) fabrication techniques.

[0066] Sample holder **110** with channels **610** formed thereon has multiple advantages over a sample holder with zero-mode waveguide holes **310** formed thereon. Because the fluorescent emissions are largely unpolarized, they would not be attenuated when they try to exit the channels **610** as much as when they try to exit holes **310** of sub-wavelength dimension. So, more emitted light from sample holder **110** can be collected and detected by optical assembly **130**, resulting in increased signal to noise ratio. In addition, each channel **610** can house a larger DNA template molecule if the DNA molecule is oriented parallel to the channel, as shown in a top-down view of the channel in FIG. 7. This way, the polymerase can migrate down the template for a much longer distance without exiting the illuminated volume **612**. The DNA molecule can be tethered so that it can remain in one location while the polymerase, having a finite processivity, may fall off the template and be replaced by another polymerase. This can lead to longer read lengths and thus significantly simplified assembly processes, especially during denovo sequencing. Although FIG. 7 shows that channel **610** is closed at both ends **701** and **702**, the channels **610** on sample holder can be open on either or both ends by extending all the way to the edge(s) of the sample holder, as shown in FIG. 8C below.

[0067] The polymerase or template molecules can be attached to sample holder **110** using conventional photoactivatable linkers. In exemplary embodiments of the present teaching, channels **610** may house more than one polymerase or template molecules attached to sample holder **110** by flexible linkers that are placed in the channels **610**. The molecules should be attached to the channels **610** in a resolvable fashion, meaning that they are sufficiently spaced from each other to allow efficient resolution of the emissions therefrom by the optical assembly **130**.

[0068] FIG. 8A illustrates a method **800** for enriching the sample holder **110** with source points by attaching the polymerase or template molecules in channels **610** in a resolvable fashion according to exemplary embodiments of the present teaching. As shown in FIG. 8A, method **800** comprises step **810** in which exposed portions of the top surface of the substrate **112** are coated with a photoactivatable linking substance such as PHOTOACTIVATABLE BIOTIN™ (PAB), step **820** in which a solution of enzyme or template molecules is applied to sample holder **110**, and step **830** in which the PAB is exposed to a pattern of light shone from the bottom surface of the substrate **112**. The pattern of light may be created by interference or refraction using grating or other conventional techniques and has interleaving lighted and dark areas in each channel **610**. The distance between two neighboring lighted areas is selected based on the resolution of the optical assembly **130** so that emissions from the two lighted areas can be separately and

substantially simultaneously detected by the optical assembly **130**. The PAB in the lighted area will be activated causing the template molecules to be attached to the sample holder in those areas, while the PAB in the dark areas will not be activated so no template molecules will be attached to those areas. Method **800** further comprises step **840** in which sample holder **110** is washed to remove unbound template molecules, leaving the bound enzyme or nucleic acid molecules in each channel **610** and allowing the formation of optically resolvable source points **210** on sample holder **110**.

[0069] Referring to FIG. 8B, instead of enzyme or template molecules, a solution of oligonucleotide (oligo) molecules can be applied to the substrate in step **820**, and additional steps can be used to stretch an oligo along the bottom surface of each of a plurality of channels **610** on the sample holder **110**. As shown in FIG. 8B, to attach an oligo to the bottom surface of a channel **610**, method **800** further comprises step **822** in which an end of the oligo is attached at one end **701** of the channel using a chemical linker, such as a biotin-streptavidin or PNA-PNA hybridization binding, where one part of the linker is attached to the end of the oligo and the other part of the linker is bound to the substrate. Method **800** further comprises step **824** in which the oligo is stretched along the channel and held to the bottom surface of the channel. Many conventional techniques of stretching DNA molecules can be used in step **824**, including but not limited to hydrodynamic, electrostatic, and magnetic manipulations. For example, the oligo molecules can be stretched using dielectrophoresis, in which the oligo is stretched by a direct current (DC) electric field or a high-frequency (e.g., 1 MHz) and high-density (e.g., 1 MV/m) alternating-current (AC) field applied between two electrodes.

[0070] In one embodiment, as shown in FIG. 8C, the sample holder **110** can be placed in a container **880** for holding the fluid containing the oligonucleotides, an electrode **871** made of, for example, Indium Ti, is placed above the end **701** of the channel, and another electrode **872** made of the same or different conductive material as electrode **871** is placed in the container **880** below the bottom surface **612** of the channel **610** and near the other end **702** of the channel. A field is provided between the two electrodes **871** and **872** such that the oligo **710** is stretched along the channel **610** and held by the field along the bottom of the channel. Depending on the relative lengths of the oligo **710** and the channel **610**, the oligo **710** may extend beyond the channel toward the electrode **872**, if the channel is open at the end **702**. Electrode **872** may also be placed under the substrate **112**, as shown in FIG. 8C.

[0071] With the electric field still on, step **830** is performed to further attach the oligo **710** so that the field can be removed later, preventing the field from interfering with sequencing operation afterwards. The oligo in each of the plurality of channels may be stretched and attached simultaneously using the same or different electrodes.

[0072] After binding the enzyme, oligonucleotide, or target nucleic acid molecules to the sample holder **110**, the sample holder **110** is placed in system **100**. A fluorophore solution comprising fluorescence labeled nucleotide analogs is applied to the sample holder **110**. In exemplary embodiments of the present teaching, the speed of chemistry of incorporation can be altered by changing the temperature, viscosity, and concentration of the fluorophore solution,

and/or by modifying the base chemistry. For example, adding molecules such as dye molecules to the fluorophore solution has been found to slow the rate of base incorporation. In addition, the sample holder 110 in system 100 should ideally be under temperature control to insure consistency. The temperature could be changed during detection. For example, the temperature of the sample holder 110 can be reduced to slow down or stop incorporation activities until the rest of system 100 is ready to collect signals from the sample holder 110, as discussed below.

[0073] To observe light emitted from the source points, excitation light from light source 120 is directed towards the substrate side of the sample holder 110, and signals from fluorescing nucleotides are collected by optical assembly 130. The confinement of the source points on sample holder 110 helps to distinguish the fluorescent signals emitted by incorporated nucleotides in the source points 210 from those emitted by freely diffusing fluorescent ligands.

[0074] As explained in more detail below, multiple methods can be used in exemplary embodiments for base determination. For example, color, signal strength, bleaching life, fluorescent lifetime, and incorporation time can be combined to gain better base discrimination. The consistency of these measurements can be used to predict a confidence value for the base determination. Confidence values can be used to sort or weight the data and to discard data of low quality, thus allowing automated consensus generation from large amount of data. This can improve the quality of the consensus as well as providing a measure of confidence.

[0075] Prior art systems, such as the one described by Levene et al., 2003 in "Zero-Mode Waveguides for Single-Molecule Analysis at High Concentrations," SCIENCE, Vol. 299:682, which article is incorporated herein by reference, uses a confocal fluorescent set up. The confocal fluorescent set up has multiple shortcomings. First, the aluminum film reflects the excitation light directly back into the collection optics. The reflected excitation light is very intense compared to the fluorescent signals from incorporated nucleotides. To attenuate the reflected light, multiple filters are used, and each filter attenuates a significant percentage of the already weak fluorescent signals. Furthermore, the excitation light in the set up of Korlach and Levene, supra, can also excite fluorescence in the optics. This unwanted fluorescence could pass through the filters, increasing the background noise.

[0076] In exemplary embodiments of the present teachings, excitation light from light source 120 is directed to the source points in sample holder 110 in an off-axis manner such that reflected excitation light, or a significant amount of it, could not enter the optical assembly 130. In some embodiments, where prism or wedge 108 is not provided, a light ray 901 from light source 120 is directed to sample holder 110 at an angle θ with respect to a normal direction N of substrate 112, as shown in FIG. 9. As the substrate 112 is made of a transparent material, such as fused silica, a relatively small first portion of the incident light 901 is reflected by the bottom surface 910 of substrate 112 and comes toward the optical assembly 103 as a first reflected light ray 912, while a second portion of the incident light enters the substrate at a different angle θ' with respect to the normal N as a refracted light ray 914. Angle θ' depends on angle θ and the refractive index n of the substrate 112. The refracted light ray 914 impinges on the metallic film at the angle θ' and a relatively large portion of the refracted light

ray 914 is likely to be reflected by the metallic film 114 and comes toward the bottom surface 910 of substrate 112 as light ray 916. Light ray 916 when crossing the bottom surface 910 is refracted again and comes off the bottom surface 910 at the angle θ as light ray 918. With the off-axis arrangement and a proper selection of the angle θ , little of the light ray 918 should enter the optical assembly 130 placed under sample holder 110, as shown in FIG. 9.

[0077] To eliminate or reduce reflection at the bottom surface 910 of substrate 112, θ can be chosen to be within 10° of the Brewster's angle θ_B . Furthermore, to achieve zero or near zero reflection at the bottom surface 910 of substrate 112, the light from the light source 120 is linearly polarized with the E vector in the light parallel to the plane of incidence, which is the plane containing the incident ray 901 and the normal N of substrate 112. According to Brewster's Law, when the angle of incidence θ is equal to or near the Brewster's angle θ_B , the transmittance, i.e., the ratio of transmitted power in ray 914 to the incident power in ray 901 across bottom surface 910 of substrate 112 should be one or near to one and the reflected power in ray 912 from surface 910 should be zero or near zero. Brewster's angle θ_B is given by:

$$\theta_B = \tan^{-1}\left(\frac{n_2}{n_1}\right) = \tan^{-1}\sqrt{\frac{\epsilon_2}{\epsilon_1}}$$

where n_1 and n_2 are the refractive indices of the respective media, i.e., air and substrate 112, and ϵ_1 and ϵ_2 are their respective electric permittivity values.

[0078] In some embodiments, system 100 is configured to achieve total internal reflection so that a significant amount of the excitation light from light source 120 is recycled within substrate 112, as shown in FIGS. 10A-10C. Total internal reflection is a phenomenon that light incident upon a boundary from a denser medium to a less dense medium is completely reflected off the boundary. Since the light ray 916 reflected from metallic film 114 has to travel through the substrate 112 toward the boundary 1010 between the substrate 112 and air, it is possible to achieve total reflection such that the light ray 916 is recycled in the substrate 112.

[0079] FIG. 10A illustrates an optical arrangement for achieving total internal reflection according to exemplary embodiments of the present teaching. As shown in FIG. 10A, prism 108 is ideally made of the same material as substrate 112 and is in direct or fluidic contact with the substrate. In exemplary embodiments, prism 108 is fused with substrate 112 at a first surface 1012 of the prism. Prism 108 has a second surface 1014 disposed at an angle α with respect to the first surface 1012. In some embodiments, angle α is selected to be equal to the incident angle θ of light ray 901 from light source 120. Thus, light ray 901 from light source 120 is directed toward the second surface 1012 of prism 108 along a normal direction of the second surface 1014 of the prism 108. While a small portion of light ray 901 may be reflected by surface 1014, the rest of light ray 901 enters substrate 112 without any change in direction because prism 108 and substrate 112 are ideally made of a same material and are fused together or optically coupled with each other with a fluid. A large portion of light ray 901 is reflected by metallic film 114 and comes off from the metallic film 114 as a light ray 1112. Light ray 1112 impinges

on the boundary **1010** between substrate **112** at angle equal to the angle θ with respect to the normal N of the substrate **112** from the inside of substrate **112**.

[0080] In exemplary embodiments of the present teaching, θ is selected to be equal or larger than a critical angle θ_c such that light ray **1112** is totally reflected from boundary **1010** and comes back towards metallic film **114** as light ray **1114**. The above reflection from the metallic film **114** and the total reflection at the boundary **1010** are repeated for light ray **1114** and its derivatives, which are the reflected portion of light ray **1114** and reflected portion thereof and so on, as shown in FIG. 10A. In exemplary embodiments, metallic film **114** is formed to extend to the side surfaces **1020** and in some embodiments to the edge portions **1030** of the bottom surface **910** of the substrate **112** so that light rays reflected from the metallic film **114** have little chance of escaping substrate **112** at the side surfaces **1020** but are recycled and used repeatedly as excitation light for the source points **210**, as shown in FIG. 10A. According to Snell's Law, the critical angle θ_c is determined by:

$$\theta_c = \sin^{-1}\left(\frac{n_1}{n_2}\right),$$

where n_1 and n_2 are the refractive indices of the respective media, i.e., air and substrate **112**, respectively.

[0081] In further embodiments, collection efficiency of optical assembly can be increased by using a fluid **104** having a refractive index between that of the air and that of the transparent material used to construct the substrate **112**. For example, when substrate **112** is made of fused silica having a refractive index of about 1.46, water can be used as the fluid **104** because it has a refractive index of 1.33, which is between the refractive index of air (~1) and that of fused silica (~1.46). The fluid **104** is placed between the substrate **112** and the optical assembly **130**. In the embodiments employing the fluid **104**, the critical angle is determined by:

$$\theta_c = \sin^{-1}\left(\frac{n_f}{n_2}\right),$$

where n_f is the refractive index of the fluid. The critical angle θ_c is therefore increased by employing the fluid. With the increase in the critical angle θ_c , the collection efficiency is increased because more emitted light from the source points is able to escape through the bottom surface **910** of the substrate **112** without going through total internal reflection, and can therefore be collected by the optical assembly **130**. The angle α of the prism **108** and the incident angle θ of the excitation light may be adjusted accordingly to allow total internal reflection of the excitation light to still occur in the presence of the fluid **104**.

[0082] In another exemplary embodiment, as shown in FIG. 10B, total internal reflection is facilitated by directing the excitation light toward a side **1060** of the substrate **112**. Excitation light **1070** from the light source **120** is directed to a side surface **1062** at an angle θ_1 with respect to a normal N' of the side surface **1062**. A refracted portion **1072** of the excitation light **1070** leaves the side surface **1062** at an angle θ_2 that is dependent on the angle θ_1 according to the Snell's Law, and impinges on the aluminum film **114** on top of the

substrate **112** at an angle $\theta_3 = \alpha - \theta_2$, where α is the angle between the side surface **1062** and the bottom surface **1010** of the substrate **112**. After reflection from the aluminum film, a reflected portion **1074** of the excitation light impinges on the surface **1010** at an angle θ_4 that is equal to the angle θ_3 . θ_2 and α can be selected such that θ_3 or θ_4 is equal to or larger than the critical angle θ_c for total reflection at the bottom surface **1010** of the substrate **112**. For example, α in this case can be selected to be at or near 90° in order to create a large θ_3 or θ_4 angle. Thus, most of the refracted portion **1072** of the excitation light **1070** can be repeated used to illuminate the source points on the sample holder **110** before exiting the substrate **112**.

[0083] In another exemplary embodiment, as shown in FIG. 10C, the side **1060** has a beveled surface **1062** forming an angle α with the bottom surface **910** of the substrate **112**, where α is less than 90° and larger or equal to the critical angle θ_c . Excitation light **1070** from the light source **120** is directed to the beveled surface **1062** along a normal N' of the beveled surface **1062**. A refracted portion **1072** of the excitation light **1070** leaves the beveled surface **1062** at an angle θ_2 that is dependent on the angle θ_1 according to the Snell's Law, and impinges on the aluminum film **114** on top of the substrate **112** at an angle $\theta_3 = \alpha$. After reflection from the aluminum film, a reflected portion **1074** of the excitation light impinges on the surface **1010** at an angle θ_4 that is equal to the angle θ_3 . With α being equal or larger than the critical angle θ_c , θ_4 is also equal or larger than θ_c and total reflection occurs at the bottom surface **1010** of the substrate **112**.

[0084] In another exemplary embodiment, as shown in FIG. 10D, the side **1060** has a beveled surface **1062** forming an angle α with the bottom surface **910** of the substrate **112**, and excitation light **1070** from the light source **120** is directed to the beveled surface **1062** at an angle θ_1 with respect to a normal N' of the beveled surface **1062**. A refracted portion **1072** of the excitation light **1070** leaves the beveled surface **1062** at an angle θ_2 that is dependent on the angle θ_1 according to the Snell's Law, and impinges on the aluminum film **114** on top of the substrate **112** at an angle $\theta_3 = \theta_2 + \alpha$. After reflection from the aluminum film, a reflected portion **1074** of the excitation light impinges on the surface **1010** at an angle θ_4 that is equal to the angle θ_3 . θ_2 and α can be selected such that θ_3 or θ_4 is equal to or larger than the critical angle θ_c for total reflection at the bottom surface **1010** of the substrate **112**. Thus, most of the refracted portion **1072** of the excitation light **1070** can be repeated used to illuminate the source points on the sample holder **110** before exiting the substrate **112**.

[0085] In another exemplary embodiment, the excitation light **901** from the light source **120** is coupled into the substrate **112** through a grism, which is a prism and grating combination, or grating **1080** formed on or attached to a portion of the bottom surface **1010** of the substrate **112**, as shown in FIG. 10E. With the use of the grism or grating **1080**, the excitation light **901** can enter the substrate at an angle θ with respect to the normal N of the substrate that is equal or larger than the critical angle θ_c , and after being reflected from the aluminum film **114**, would impinge on the bottom surface **1010** of the substrate **112** at the angle θ and be totally reflected from the bottom surface **1010** back into the substrate. Thus, the excitation light is recycled within the substrate, as shown in FIG. 10C.

[0086] The arrangements in FIGS. 9 through 10E are advantageous over conventional systems in part because, by placing the detector(s) and the optical assembly 130 under sample holder 110, fluorescent signals can be collected through the bottom surface 910 of the sample holder 110 without the interference of reflected light from metallic film 114. Furthermore, since the excitation light and the fluorescent signals entering the optical assembly 130 do not have a common light path, there is no need of heavy filtering to separate the excitation light entering the substrate 112 from the fluorescent signals exiting the substrate 112 through the bottom surface 910.

[0087] In various embodiments, optical assembly 130 comprises at least one pixilated or multi-element detector configured to sense light signals landed thereon and a set of optical components configured to direct light emissions from the source points toward the multi-element detector(s). FIG. 11 illustrates a top view of a portion of the sample holder 110 showing a plurality of source points 210. In exemplary embodiments, as shown in FIG. 12, the pixilated or multi-element detector comprises a plurality of addressable light-sensing elements 1210 organized in an imaging plane 1220, such as the x-y plane. The set of optical components is configured to direct light emissions from different source points toward different areas 1230 of the imaging plane 1220 so that light emissions from different source points 210 can be separately and substantially simultaneously detected.

[0088] Thus, as shown in FIG. 12, light emissions from each source point 210 form an image of the source point in an area 1230 on the imaging plane 1220. In exemplary embodiments, the set of optical components further comprises a light-dispersing setup configured to separate light emissions from the multiple source points 210 into multiple spectral components so that the detected light from each source point is spread out spectrally along the y axis and images 1230 represent spectrally resolved images of the source points 210, as shown in FIG. 12. When the light-dispersing setup is provided, enough separation between neighboring source points 210 on the sample holder 110 is provided to insure that the spectrally resolved images 1230 of the source points do not overlap with each other. In addition, sufficient gap g along the y-direction between an image 1230 of a source point and an image 1230 of a neighbor source point are provided to prevent overlap of data associated with the two source points due to cutoff filter tolerances.

[0089] The position of the images 1230 can be determined by a spatial calibration to associate each source point on the sample holder with an area 1230 on the image plane 1220. The calibration can be done by using a dye solution or a light source that is not blocked by system filters. Such calibration, however, may not be required if there is no need to correspond the images 1230 with the source points 210. In addition, tolerance should be allowed to insure that there is sufficient separation d between the areas 1230 and the edges of the image plane 1220, and the separation should be controlled to allow detection of all of the source points 210 on the image plane 1220. As a non-limiting example, the buffer zone d between a side 1232 of an areas 1230 facing an edge 1222 of the image plane 1220 is no more than 8 pixels wide.

[0090] Although FIG. 12 shows that the source point images 1230 each having a width w of 2 pixels, a pitch between two neighboring images in the x-direction being 4

pixels, a gap g between two adjacent rows of images being 4 pixels, and a buffer distance d of the images 1230 from each edge of the image plane 1220 being 8 pixels, these numbers are shown as examples and can be different in different applications or are adjustable to suit different applications. Moreover, although FIG. 12 shows only two rows of source points 210 and two rows of images 1230 corresponding to the source points, in practice, there may be more or less rows of source points or images. Also, the source points 210 do not have to be arranged in rows and can be spread out on the sample holder in any order or even randomly as long as they are sufficiently separated so that their images 1230 do not overlap on the image plane 1220.

[0091] In exemplary embodiments, the optical assembly 130 is similar to the one in the optical system disclosed in U.S. Pat. No. 6,690,467 B1 by Reel, which is incorporated herein by reference. As shown in FIG. 13, as a non-limiting example, the optical assembly comprises a collection lens assembly 1310, a reimaging lens assembly 1320, and at least one CCD detector 1330 as the pixilated detector. The collection lens assembly 1310 comprises at least one collection lens configured to collect light emissions from the source points 210. The reimaging lens assembly 1320 comprises at least one reimaging lens configured to focus the collected light emissions from different source points into different areas 1230 of the imaging plane 1220 of the detector 1330.

[0092] The use of the collection lens assembly 1310 may also provide a substantially collimated region between the collection lens assembly 1310 and the reimaging lens assembly 1320, which is suitable for insertion of a variety of optical devices such as an aperture 1340, a light-dispersing assembly 1350, and/or a laser line filter 1360. In exemplary embodiments, the light-dispersing assembly 1350 comprises at least one grating, prism, or grism configured to spread spectrally rays of light that pass through it. For example, a transmission grating deflects rays of light that strike thereon at an angle roughly proportional to the wavelength of the light. Thus, the collimated light emissions from the source points 210, after going through the transmission gratings, are dispersed spectrally. With the spectral dispersion, a first light ray of a first wavelength and a second light ray of a second wavelength coming from a same source point 210 should arrive at the reimaging lens assembly 1320 at different angles with respect to an optical axis of the reimaging lens assembly 1320 and thus be focused onto different locations 1234 and 1236 of the area 1230 corresponding to the source point, as shown in FIG. 12. Locations 1234 and 1236 are spaced apart from each other along the y-axis because of the spectral dispersion.

[0093] Instead of prism, grating, or grism in the light dispersing assembly 1350, dichroic or bandpass filters can be used to separate the spectral components in the fluorescent signals from each source point. FIG. 14A illustrates other embodiments of optical assembly 130. As shown in FIG. 14A, optical assembly 130 comprises a collection assembly 1310, an imaging assembly comprising imaging lenses 1332-1, 1332-2, 1332-3, and 1332-4 disposed at 90° angles with respect to each other, a plurality of CCD detectors 1330, and a light dispersing assembly comprising dichroic or bandpass filters D1, D2, and D3 placed at 90° angles with respect to each other. Each dichroic or bandpass filter is configured to allow passage of one of the first, second, and third bands of fluorescent signals, respectively, and to reflect all other frequencies. The plurality of CCD detectors 1330

comprises CCD detectors **1330-1**, **1330-2**, **1330-3**, and **1330-4**. CCD detector **1330-1** is placed behind dichroic filter **D1** to collect the first band of fluorescent signals, CCD detector **1330-2** is placed behind dichroic filter **D2** to collect the second band of fluorescent signals, CCD detector **1330-3** is placed behind dichroic filter **D3** to collect the third band of fluorescent signals, and CCD detector **1330-4** is placed in front of dichroic filter **D3** to collect the signals reflected therefrom, which should comprise the fourth band of fluorescent signals. Other filters (not shown) can be placed before CCD detectors **1330**, respectively, for improved frequency selection.

[0094] Alternatively, a dichroic or bandpass filter can be configured to reflect the first, second, third, or fourth band of fluorescent signals, and to allow passage of all other frequencies. It is also possible to combine bandpass, notch, lowpass and highpass filters in any combination that permits appropriate separation of the emission wavelengths. FIG. **14B** illustrates still other embodiments of optical assembly **130**. As shown in FIG. **14B**, optical assembly **130** comprises a collection lens assembly **1310**, a reimaging assembly comprising reimaging lenses **1320-1**, **1320-2**, **1320-3** and **1320-4**, and CCD detectors **1330-1**, **1330-2**, **1330-3**, and **1330-4** each behind a respective one of the reimaging lenses **1320-1**, **1320-2**, **1320-3** and **1320-4**. Optical assembly **130** further comprises a light dispersing assembly comprising dichroic or bandpass filters **DF1**, **DF2**, **DF3** and **DF4** placed in a row under collection assembly **1310** and each at an angle γ to an optical axis (shown by the dashed line) of the collection assembly **1310**. Dichroic or bandpass filters **DF1**, **DF2**, **DF3**, and **DF4** are each placed in front of a respective one of the reimaging lenses **1320-1**, **1320-2**, **1320-3** and **1320-4** and are configured to reflect the first, second, third and fourth bands of fluorescent signals, respectively, while allowing passage of signals of other frequencies. Other filters (not shown) can be placed before CCD detectors **1330**, respectively, for improved frequency selection.

[0095] Imaging lenses **1320-1**, **1320-2**, **1320-3** and **1320-4** can be separate lenses or sections of a single lens, CCD detectors **1330-1**, **1330-2**, **1330-3**, and **1330-4** in FIG. **14B** can be separate CCD detectors or sections of a single CCD detector. Although FIG. **14B** shows that the dichroic or bandpass filters **DF1**, **DF2**, **DF3** and **DF4** are at a roughly 45° angle with respect to the optical axis of the collection assembly **1310**, such placement is not necessary and the angle γ can be larger or smaller than 45° . FIG. **14C** illustrates an exemplary configuration of the optical assembly **130** when the angle γ is close to 90° so that the collimated light emissions from a source point would impinge on the dichroic or bandpass filters **DF1**, **DF2**, **DF3** and **DF4** at a small incident angle β and be imaged by the reimaging lens assembly **1320** onto the CCD detector(s) **1330**.

[0096] The CCD assembly **1330** comprises at least one charge-coupled device (CCD) array, such as a regular CCD array, a complimentary metal-oxide-semiconductor (CMOS) array, an electron-multiplying CCD (EMCCD) array, an intensified CCD (ICCD) array, or an electron-bombarded CCD (EBCCD) array. A CCD array is advantageous over other multi-element detectors, such as an array of avalanche photodiode (APD) based detectors or photomultiplier tube (PMT) based detectors, because the number of elements in a CCD array is much higher, as the size of a CCD pixel in the CCD array can be as small as $3\ \mu\text{m}$ or even smaller. Therefore, signals from different source points can

be differentiated by detecting them using different groups of elements in the CCD array, as discussed above. A CCD array can be much less costly than an APD or PMT array.

[0097] To amplify the low light signals from fluorescing labels on the incorporated bases above background noises in CCD arrays, a high-sensitivity CCD-based device such as EMCCD, ICCD, or EB-CCD, is used in exemplary embodiments. Due to fast base incorporation rates of DNA molecules, in addition to sensitivity, the speed of reading data out from a CCD detector is also important because it is associated with the ability to capture event data and to readout the data out over a short period of time to allow the next event to be observed. Through careful design of a readout scheme, a CCD array can be made to be fast enough to resolve fluorescent emissions from two consecutive incorporation events associated with a same source point. Moreover, a CCD with multiple outputs or taps can be used to increase the CCD readout speed. For example, a CCD with **4** taps can allow a 4-times increase in readout speed, which allows images of more source points to be read for increased throughput.

[0098] To further improve the readout speed, a frame transfer CCD (FTCCD) array **1500**, as illustrated in FIG. **15A**, is used in detector **1330** according to exemplary embodiments of the present teaching. The FTCCD array **1500** allows data readout operations to be performed concurrently with data collection operations. As shown in FIG. **15A**, CCD array **1500** comprises a dark area **1510**, an image area **1520**, a masked storage area **1530**, and a horizontal register **1540**. The image area **1520** of CCD array **1500** is where light signals are detected and is constructed as a two-dimensional array of light-sensitive elements or pixels. Storage area **1530** comprises an array of storage elements covered with an opaque mask to provide temporary storage for an image frame transferred from the image area. The signal that is accumulated in each pixel in image area **1520** is read out by a process of parallel transfer in the negative y-direction shown in the figure, whereby charges in each horizontal row within image area **1520** and storage area **1530** are transferred to the next row and so forth until ultimately they reach the horizontal register **1540**, which is a serial readout register that allows charges to be transferred in the x-direction to an output node (not shown), from which they are read out. While data from storage array **1530** is read, image area **1520** is available to collect a next round of light signals.

[0099] Dark area **1510** is a region of excess pixels. Because these pixels are not illuminated, they do not have to be cleared during each readout. Usually, the combination of dark area **1510** and image areas **1520** maps directly onto the storage area **1530**. In one embodiment, image area **1520** occupies a small fraction (e.g. $\frac{1}{10}$) of the combination so that source point data can be read out at, for example, 10 times the normal frame rate. In exemplary embodiments, CCD array **1500** is kept cool at about 80°C . below zero so that minimal dark current charges are generated. In certain embodiments, the dark area is eliminated when CCD **1500** is custom built to have just the right amount of rows in the image area **1520**.

[0100] In some embodiments, an interline CCD or a combined interline and frame transfer CCD may be employed. FIG. **15B** illustrates an interline CCD array **1550** in CCD detector **1330** according to exemplary embodiments of the present teaching. As shown in FIG. **15B**, interline

CCD array 1550 comprises separate image regions 1560 and CCD storage regions 1570. CCD storage regions 1570 are protected with a mask structure and positioned alongside respective ones of image regions 1560 such that CCD storage regions 1570 and image regions 1560 together form an alternating parallel array. Image regions 1560 may comprise circuit elements such as photodiodes for capturing the images of the source points while CCD storage regions 1570 shift previously acquired images in a parallel fashion towards a horizontal register 1590. The horizontal register 1590 then sequentially shifts image information from each CCD storage region to an output amplifier or other processing circuits (not shown) as a serial data stream. The readout process is repeated until data from all of the CCD storage regions 1570 are transferred to the output amplifier.

[0101] Where an actual two-dimensional image is desired from the CCD, the image data in a digital format is reconstructed to yield the final image. Where the data is to be used for non-pictorial or non-imaging applications, the relevant pixel data may be identified and processed according to its intended purpose. One advantage of the interline CCDs is their ability to operate without a shutter or synchronized strobe, allowing for an increase in device speed and faster frame rates. An interline CCD array can be used to eliminate blurring or image smear, which is a common problem with frame-transfer CCDs, by effectively doing horizontal shifts directly from the image regions to the respective ones of the storage regions.

[0102] In exemplary embodiments, readout speed is further improved by limiting the number of source points to be imaged on the CCD so that the number of data rows to be read are minimized. The number of data rows to be read may also be minimized by binning vertically (in the y-direction), especially when the source points 210 and thus the images of the source points 1230 are in an array so that the positions of the images can be fairly accurately predicted, as shown in FIG. 12. With the fluorescent signals dispersed spectrally over a range of pixels along the y-direction, the pixels can be binned in the y-direction to capture informative wavelength groupings for base determination. For example, as shown in FIG. 12, each source point image 1230 may comprise four interested wavelength groups C1, C2, C3, and C4 interleaved with uninterested wavelength groups U1, U2, and U3. Data in each of the interested wavelength groups can be binned, while data in the uninterested wavelength groups can be cleared without being read, as discussed below. Binning can also be done in the x-direction for further increase in readout speed because shifting is often faster than reading. With a CCD array, binning can be done on-chip in a conventional manner that does not introduce noise.

[0103] The CCD array in detector 1330 may also be made to allow clearing of the horizontal register 1540 or 1590. This can speed up readouts if desired data is separated by rows of unneeded pixels. FIG. 16A illustrates examples of four normalized fluorescent spectra associated with the four different fluorescent labels used to label respective ones of four different nucleotides. The four spectra have peak regions 1610, 1620, 1630, and 1640 corresponding to respective ones of the first, second, third and fourth frequency bands. Only four fluorescent frequency bands need to be collected by detector 1330. Data can be over-determined, however, to comprise more than the spectrum of data associated with an incorporation event. With the light dis-

persing setup shown in FIG. 13, a continuous portion of a spectrum of data is spread along the y-direction, as shown in FIG. 16B. FIG. 16C illustrates the over-determined data with the most informative wavelengths corresponding to the four frequency bands in gray and the less informative wavelength ranges shown in white. Data rows in the less informative wavelength range between two gray bands is referred to as a band gap bg, which may not be needed and can be cleared without being read out for increased readout speed. The width and position of each informative frequency band in FIG. 16C can be optimized for best signal to noise (S/N) ratio after multicomponenting, as explained in more detail below.

[0104] While the frequency bands in FIGS. 12, 16B, and 16C are shown to spread along the y-direction, this is not essential and orienting the frequency bands in a different direction on the CCD may be advantageous in some cases.

[0105] FIG. 17A is a flow chart of a method 1700 for reading out image data associated with a plurality of source points according to exemplary embodiments of the present teaching. As a non-limiting example, method 1700 is described in the context of a frame transfer EMCCD array of 512x512 pixels in the image area that allows clearing of the horizontal register. Nevertheless, the method can be readily adapted to other types of CCD array. As shown in FIG. 17A, method 1700 comprises step 1710 in which the pixels in the image area are shifted down by a frame having, for example, 128 rows. So, 128 rows of pixels are shifted from the image area into the masked storage area. These pixels are not read out for a number of (e.g., 4) frames (512/128=4). Pixels from the dark area will be shifted down to collect the next images, but the provision of a dark area is not necessary because the CCD is kept cool (-80° C.) so minimal dark current charge is generated. Method 1700 further comprises step 1720 in which a first number of (e.g., 8) buffer rows in a frame is shifted down to the register and cleared. The number 8 is arbitrarily chosen. This number should correspond to the distance d shown in FIG. 12. Method 1700 further comprises step 1730 in which the pixels associated with the first frequency band is vertically binned and shifted. As a non-limiting example, 4 rows of pixels are associated with a frequency band, and these rows are binned and read out as one row. Method 1700 further comprises step 1740 in which at least some of the rows in the band gap bg before the next frequency band are cleared. Steps 1730 and step 1740 are thereafter repeated for each of the other frequency bands. Method 1700 further comprises step 1750 in which at least some of the buffer rows in the gap g between two rows of images 1230 are shifted and cleared. Steps 1730 through 1750 are repeated for each row of source points when the source points are arranged in an array.

[0106] FIG. 17B is a spreadsheet for estimating the throughput for system 100 using a frame transfer EMCCD as detector 1330 according to exemplary embodiments. As shown in FIG. 17B, for a frame transfer CCD having 512x512 pixels in the image area, a normal read out time of 1×10^{-7} second, a vertical shift time of 4×10^{-7} second, and a horizontal register clear time of 5×10^{-6} second, using method 1700, the time to read out emission data from 4 rows of source points is about 0.00096 seconds, resulting in a readout speed higher than 1000 Hz, which is much faster than the normal readout speed of 30 Hz for the CCD. FIG. 17C is a spreadsheet for estimating the throughput for system 100 using a 1004x1002 EMCCD as detector 1330

according to exemplary embodiments. As shown in FIG. 17C, the time for reading out emission data from 4 rows of source points in this case is about 0.00113 seconds, resulting in a readout speed close to 1000 Hz.

[0107] In exemplary embodiments of the present teaching, optical data collected by detector 1330 is sent to computer system 140 and optionally or additionally DSP or FPGA 150 for base determination. FIG. 18 is a block diagram of computer system 140 according to exemplary embodiments of the present teaching. As shown in FIG. 18, computer system 140 comprises a central processing unit (CPU) 1810, a memory unit 1820, a data input port 1830, and a user interface 1840, interconnected by one or more buses 1850. Memory unit 1820 preferably stores operating system software 1822 and other software programs including a program 1824 for base determination. Memory unit 1820 further comprises a data storage unit 1826 for storing optical data collected by detector 1330 and sent to computer system 140 through data input port 1830. Program 1824 comprises coded instructions, which, when executed by CPU 1810, cause computer system 140 to carry out methods for detecting light emissions from multiple source points as described above and/or methods for base determination based on the detected data according to exemplary embodiments of the present teaching, as explained in more detail below.

[0108] With the illumination of excitation light, a labeled and incorporated nucleotide should fluoresce by emitting photons from an associated source point. The spectrum of the photons collected at detector 130 from this single incorporation event should be a collection of photons with different energies or frequencies. When the number of collected photons is large, the spectrum should resemble the normal dye spectrum corresponding to the fluorescent dye used to label the incorporated nucleotide. The spectrum will vary, however, due to the small number of photons that are typically collected by detector 1330 from the single incorporation event in each collection time period.

[0109] For example, a fluorescing dye may emit 10,000 photons over a 10 micro second period, and about 4% of the 10,000 photons may be detected by detector 1330 in ten time bins each corresponding to, for example, a one micro second period. Thus, roughly 40 photons may be collected in each time bin. Plotted spectrally over 10 spectral bins, the 40 expected photons might spread out like the histogram shown in FIG. 19. Because of the small number of photons, the distribution in FIG. 19 may not match the normal dye spectrum corresponding to the incorporated nucleotide, and the mismatch may lead to a chance of incorrect base determination.

[0110] In exemplary embodiments, to avoid a base determination problem caused by a small number of photons from a single incorporation event, the present teaching comprises a method 2000 for base determination illustrated by the flowchart in FIG. 20. As shown in FIG. 20, method 2000 comprises step 2010 in which data from all spectral bins collected in each of a plurality of consecutive time bins are copied and combined to form composite light data. Method 2000 further comprises step 2020 in which the composite data from the plurality of consecutive time bins are used to determine an incorporation time interval T.

[0111] FIG. 21 is a plot of the composite data over a plurality of 24 time bins showing the number of photons detected in each time bin. As shown in FIG. 21, during time bins 1, 2, 3, 4, and 5, and time bins 16, 17, 18, and 19, the

composite data indicate small numbers of photons coming from a background of diffusion events, trial events, and substrate fluorescence. In other time bins, the numbers of photons are significantly larger, resulting in large rises 2110 and 2120 above the background noises. Rises 2110 and 2120 indicate incorporation events. The incorporation time T for the incorporation event corresponding to rise 2110 can be determined by measuring the width of rise 2110, as shown in FIG. 21.

[0112] Since most of the photons detected during the incorporation time interval T are from a single incorporation event, for each color bin, data associated with the same spectral bin but collected in different time bins in the incorporation time interval can be combined, resulting in increased data points for the spectral bin. The increase in the number of data points leads to an improved multicomponenting process, which is used to convert color data to dye composition. Thus, method 2000 further comprises step 2030 in which data associated with each spectral bin or frequency band of interest but collected in different time bins in the incorporation time interval T are combined, and step 2040 in which the combined data is used in a conventional multicomponenting process to determine a dominant dye, which is used to determine the base being incorporated. Method 2000 further comprises step 2050 in which the residuals of the multicomponenting process is used to determine a confidence level.

[0113] Method 2000 for improving the signal to noise ratio by combining data from multiple time bins may be coded as a computer program and executed by computer system 140. Alternatively, since the same algorithm in method 2000 is executed a large number of times, hardware solutions such as field program gate arrays (FPGA) or digital signal processors (DSP) 150 and the like can be used to reduce the computation load and data stream size. The FPGAs or DSPs 150 could be integrated in detector(s) 1330, between detector 1330 and computer system 140, as shown in FIG. 1, or installed into computer system 140.

[0114] The foregoing descriptions of specific embodiments of the present teaching have been presented for purposes of illustration and description. They are not intended to be exhaustive or to limit the teaching to the precise forms disclosed, and obviously many modifications and variations are possible in light of the above teaching. The embodiments were chosen and described in order to best explain the principles of the teaching and its practical application, to thereby enable others skilled in the art to best use the teaching and various embodiments with various modifications as are suited to the particular use contemplated. It is intended that the scope of the teaching be defined by the claims appended hereto and their equivalents.

1.-60. (canceled)

61. A method of interrogating target molecules within optical confinement regions disposed upon a substrate, comprising:

- providing photo-activated coupling groups on the substrate, including within the optical confinement regions;
- directing activating radiation at the substrate to selectively activate the photo-activated coupling groups within the optical confinement regions;
- depositing target molecules over the surface of the substrate such that at least a portion of the target molecules

are retained within the optical confinement regions by coupling with the activated photo-activated coupling groups;
removing uncoupled target molecules from the substrate that are not retained within the optical confinement regions; and
directing light toward the substrate at an angle off-axis with respect to normal of the substrate such that at least a portion of the light is propagated by total internal reflection through at least a portion of the substrate illuminating coupled target molecules within a multiplicity of optical confinement regions.

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