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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶:
 C12N 15/12, C07K 14/705, C12N 5/10, A61K 38/17, G01N 33/50, C12Q 1/68, G01N 33/68

(11) International Publication Number:

WO 98/49293

(43) International Publication Date:

5 November 1998 (05.11.98)

(21) International Application Number:

PCT/GB98/01206

A1

(22) International Filing Date:

24 April 1998 (24.04.98)

(30) Priority Data:

9708479.2

25 April 1997 (25.04.97)

GB

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(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, GW, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).

Published

With international search report.

Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.

(54) Title: HUMAN THETA SUBUNIT OF THE GABA-A RECEPTOR

(57) Abstract

The present invention relates to the cloning of a novel cDNA sequence encoding the theta receptor subunit of the $GABA_A$ receptor; to stably co-transfected eukaryotic cell lines capable of expressing a $GABA_A$ receptor, which receptor comprises the novel theta receptor subunit; and to the use of such cell lines in screening for and designing medicaments which act upon the $GABA_A$ receptor.

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HUMAN THETA SUBUNIT OF THE GABA-A RECEPTOR

This invention concerns the cloning of a novel cDNA sequence encoding a particular subunit of the human GABA_A receptor. In addition, the invention relates to a stable cell line capable of expressing said cDNA and to the use of the cell line in a screening technique for the design and development of subtype-specific medicaments.

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Gamma-amino butyric acid (GABA) is a major inhibitory neurotransmitter in the central nervous system. It mediates fast synaptic inhibition by opening the chloride channel intrinsic to the GABA_A receptor. This receptor comprises a multimeric protein of molecular size 230-270 kDa with specific binding sites for a variety of drugs including benzodiazepines, barbiturates and β-carbolines, in addition to sites for the agonist ligand GABA (for reviews see MacDonald and Olsen, *Ann. Rev. Neurosci.*, 1994, 17, 569; and Whiting *et al*, *Int. Rev. Neurobiol.*, 1995, 38, 95).

Molecular biological studies demonstrate that the receptor is composed of several distinct types of subunit, which are divided into four classes (α , β , γ and δ) based on their sequence similarities. To date, in mammals, six types of α (Schofield *et al.*, *Nature* (*London*), 1987, **328**, 221; Levitan *et al.*, *Nature* (*London*), 1988, **335**, 76; Ymer *et al.*, *EMBO J.*, 1989, **8**, 1665; Pritchett & Seeberg, *J. Neurochem.*, 1990, **54**, 802; Luddens *et al.*, *Nature* (*London*), 1990, **346**, 648; and Khrestchatisky *et al.*, *Neuron*, 1989, **3**, 745), three types of β (Ymer *et al.*, *EMBO J.*, 1989, **8**, 1665), three types of γ (Ymer *et al.*, *EMBO J.*, 1990, **9**, 3261; Shivers *et al.*, *Neuron*, 1989, **3**, 327; and Knoflach *et al.*, *FEBS Lett.*, 1991, **293**, 191) and one δ subunit (Shivers *et al.*, *Neuron*, 1989, **3**, 327) have been identified. More recently, a further member of the GABA receptor gene family, ε , has been identified (Davies *et al.*, *Nature*, 1997, **385**, 820). The polypeptide is 506 amino acids in length and exhibits greatest amino acid sequence identity

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with the GABA_A receptor γ_3 subunit (47%), although this degree of homology is not sufficient for it to be classified as a fourth γ subunit.

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The differential distribution of many of the subunits has been characterised by *in situ* hybridisation (Shivers *et al.*, *Neuron*, 1989, 3, 327; Wisden *et al*, *J. Neurosci.*, 1992, 12, 1040; and Laurie *et al*, *J. Neurosci*, 1992, 12, 1063) and this has permitted it to be speculated which subunits, by their co-localisation, could theoretically exist in the same receptor complex.

Various combinations of subunits have been co-transfected into cells 10 to identify synthetic combinations of subunits whose pharmacology parallels that of bona fide GABAA receptors in vivo (Pritchett et al... Science, 1989, 245, 1389; Pritchett and Seeberg, J. Neurochem., 1990, 54, 1802; Luddens et al., Nature (London), 1990, 346, 648; Hadingham et al., Mol. Pharmacol., 1993, 43, 970; and Hadingham et al., Mol. Pharmacol., 15 1993, 44, 1211). This approach has revealed that, in addition to an α and β subunit, either γ₁ or γ₂ (Pritchett et al., Nature (London), 1989, **338**, 582; Ymer et al., EMBO J., 1990, 9, 3261; and Wafford et al., Mol. Pharmacol., 1993, 44, 437) or γ₃ (Herb et al., Proc. Natl. Acad. Sci. USA, 1992, 89, 1433; Knoflach et al., FEBS Lett., 1991, 293, 191; and Wilson-Shaw et al.. 20 FEBS Lett., 1991, 284, 211) is also generally required to confer benzodiazepine sensitivity, and that the benzodiazepine pharmacology of the expressed receptor is largely dependent on the identity of the α and γ subunits present. Receptors containing a δ subunit (i.e. $\alpha\beta\delta$) do not appear to bind benzodiazepines (Shivers et al., Neuron, 1989, 3, 327; and 25Quirk et al., J. Biol. Chem., 1994, 269, 16020). Combinations of subunits have been identified which exhibit the pharmacological profile of a BZ1 type receptor $(\alpha_1\beta_1\gamma_2)$ and a BZ₂ type receptor $(\alpha_2\beta_1\gamma_2)$ or $\alpha_3\beta_1\gamma_2$, Pritchett et al., Nature (London), 1989, 338, 582), as well as GABAA receptors with a novel pharmacology, α₅β₂γ₂ (Pritchett and Seeberg, J. Neurochem., 1990, 30 **54**, 1802), $\alpha_4\beta_2\gamma_2$ (Wisden *et al*, *FEBS Lett.*, 1991, **289**, 227) and $\alpha_6\beta_2\gamma_2$

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(Luddens et al., Nature (London), 1990, 346, 648). The pharmacology of these expressed receptors appears similar to that of those identified in brain tissue by radioligand binding, and biochemical experiments have begun to determine the subunit composition of native GABA receptors (McKernan & Whiting, Tr. Neurosci., 1996, 19, 139). The exact structure of receptors in vivo has yet to be definitively elucidated.

The present invention relates to a new class of GABA receptor subunit, hereinafter referred to as the theta subunit (θ subunit).

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The nucleotide sequence for the theta subunit, together with its deduced amino acid sequence corresponding thereto, is depicted in Figure 1 of the accompanying drawings.

The present invention accordingly provides, in a first aspect, a DNA molecule encoding the theta subunit of the human GABA receptor comprising all or a portion of the sequence depicted in Figure 1, or a modified human sequence.

In an alternative aspect, the present invention provides a DNA molecule encoding the theta subunit of the human GABA receptor comprising all or a portion of the sequence depicted in Figure 2, or a modified human sequence.

The term "modified human sequence" as used herein referes to a variant of the DNA sequences depicted in Figure 1 and Figure 2. Such variants may be naturally occurring allelic variants or non-naturally occurring or "engineered" variants. Allelic variation is well known in the art in which the nucleotide sequence may have a substitution, deletion or addition of one or more nucleotides without substantial alteration of the function of the encoded polypeptide. Particularly preferred allelic variants arise from nucleotide substitution based on the degeneracy of the genetic code.

The sequencing of the novel cDNA molecules in accordance with the invention can conveniently be carried out by the standard procedure described in accompanying Example 1; or may be accomplished by

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alternative molecular cloning techniques which are well known in the art, such as those described by Maniatis *et al.* in *Molecular Cloning*, *A Laboratory Manual*, Cold Spring Harbor Press, New York, 2nd edition, 1989.

In a further aspect, the present invention also relates to polynucleotides (for example, cDNA, genomic DNA or synthetic DNA) which hybridize under stringent conditions to the DNA molecules depicted in Figure 1 and Figure 2. As used herein, the term "stringent conditions" will be understood to require at least 95% and preferably at least 97% identity between the hybridized sequences. Polynucleotides which hybridize under stringent conditions to the DNA molecules depicted in Figure 1 and Figure 2 preferably encode polypeptides which exhibit substantially the same biological activity or function as the polypeptides depicted in Figure 1 and Figure 2, respectively.

The present invention further relates to a GABA theta subunit polypeptide which has the deduced amino acid sequence of Figure 1 or Figure 2, as well as fragments, analogs and derivatives thereof.

The terms "fragment", "derivative" and "analog" when referring to the polypeptide of Figure 1 or Figure 2, means a polypeptide which retains essentially the same biological activity or function as the polypeptide depicted in Figure 1 or Figure 2. Thus, an analog may be, for example, a proprotein which can be activated by cleavage of the proprotein portion to produce an active mature polypeptide.

The polypeptide of the present invention may be a recombinant polypeptide, a natural polypeptide or a synthetic polypeptide, preferably a recombinant polypeptide.

The fragment, derivative or analog of the polypeptide of Figure 1 or Figure 2 may be one in which one or more of the amino acid residues are substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue) and such substituted amino acid residues may or may not be one encoded by the genetic code; or one in

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which one or more of the amino acid residues includes a substituent group; or one in which the mature polypeptide is fused with another compound, such as a compound to increase the half-life of the polypeptide (for example, polyethylene glycol); or one in which the additional amino acids are fused to the mature polypeptide, such as a leader or secretory sequence or a sequence which is employed for purification of the mature polypeptide or a proprotein sequence. Such fragments, derivatives and analogs are deemed to be within the technical capabilities of those skilled in the art.

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The polypeptides and DNA molecules of the present invention are preferably provided in an isolated form, and preferably are purified to homogeneity.

The term "isolated" means that the material is removed from its original environment (e.g., the natural environment if it is naturally occurring). For example, a naturally-occurring DNA molecule or polypeptide present in a living animal is not isolated, but the same DNA molecule or polypeptide, separated from some or all of the coexisting materials in the natural system, is isolated. Such DNA molecules could be part of a vector and/or such DNA molecules or polypeptides could be part of a composition, and still be isolated in that such vector or composition is not part of its natural environment.

In another aspect, the invention provides a recombinant expression vector comprising the nucleotide sequence of the human GABA receptor theta subunit together with additional sequences capable of directing the synthesis of the said human GABA receptor theta subunit in cultures of stably co-transfected eukaryotic cells.

The term "expression vectors" as used herein refers to DNA sequences that are required for the transcription of cloned copies of recombinant DNA sequences or genes and the translation of their mRNAs in an appropriate host. Such vectors can be used to express eukaryotic genes in a variety of hosts such as bacteria, blue-green algae, yeast cells,

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insect cells, plant cells and animal cells. Specifically designed vectors allow the shuttling of DNA between bacteria-yeast, bacteria-plant or bacteria-animal cells. An appropriately constructed expression vector should contain: an origin of replication for autonomous replication in host cells, selective markers, a limited number of useful restriction enzyme sites, a high copy number, and strong promoters. A promoter is defined as a DNA sequence that directs RNA polymerase to bind to DNA and to initiate RNA synthesis. A strong promoter is one which causes mRNAs to be initiated at high frequency. Expression vectors may include, but are not limited to, cloning vectors, modified cloning vectors, specifically designed plasmids or viruses.

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The term "cloning vector" as used herein refers to a DNA molecule, usually a small plasmid or bacteriophage DNA capable of self-replication in a host organism, and used to introduce a fragment of foreign DNA into a host cell. The foreign DNA combined with the vector DNA constitutes a recombinant DNA molecule which is derived from recombinant technology. Cloning vectors may include plasmids, bacteriophages, viruses and cosmids.

The recombinant expression vector in accordance with the invention may be prepared by inserting the nucleotide sequence of the GABA theta subunit into a suitable precursor expression vector (hereinafter referred to as the "precursor vector") using conventional recombinant DNA methodology known from the art. The precursor vector may be obtained commercially, or constructed by standard techniques from known expression vectors. The precursor vector suitably contains a selection marker, typically an antibiotic resistance gene, such as the neomycin or ampicillin resistance gene. The precursor vector preferably contains a neomycin resistance gene, adjacent the SV40 early splicing and polyadenylation region; an ampicillin resistance gene; and an origin of replication, e.g. pBR322 ori. The vector also preferably contains an inducible promoter, such as MMTV-LTR (inducible with dexamethasone)

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or metallothionin (inducible with zinc), so that transcription can be controlled in the cell line of this invention. This reduces or avoids any problem of toxicity in the cells because of the chloride channel intrinsic to the GABAA receptor.

One suitable precursor vector is pMAMneo, available from Clontech Laboratories Inc. (Lee *et al.*, *Nature*, 1981, **294**, 228; and Sardet *et al.*, *Cell*, 1989, **56**, 271). Alternatively the precursor vector pMSGneo can be constructed from the vectors pMSG and pSV2neo.

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The recombinant expression vector of the present invention is then produced by cloning the GABA receptor theta subunit cDNA into the above precursor vector. The receptor subunit cDNA is subcloned from the vector in which it is harboured, and ligated into a restriction enzyme site, e.g. the Hind III site, in the polylinker of the precursor vector, for example pMAMneo or pMSGneo, by standard cloning methodology known from the art, and in particular by techniques analogous to those described herein. Before this subcloning, it is often advantageous, in order to improve expression, to modify the end of the theta subunit cDNA with additional 5' untranslated sequences, for example by modifying the 5' end of the theta subunit DNA by addition of 5' untranslated region sequences from the α_1 subunit DNA. Alternatively, expression of the theta subunit cDNA may be modified by the insertion of an epitope tag sequence such as c-myc.

According to a further aspect of the present invention, there is provided a stably co-transfected eukaryotic cell line capable of expressing a GABA receptor, which receptor comprises the theta receptor subunit, at least one alpha receptor subunit and optionally one or more beta, gamma, delta, or epsilon receptor subunit.

This is achieved by co-transfecting cells with multiple expression vectors, each harbouring cDNAs encoding for an α , θ , and optionally one or more β , γ , δ , or ϵ GABA receptor subunits. In a further aspect, therefore, the present invention provides a process for the preparation of a eukaryotic cell line capable of expressing a GABA receptor, which

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comprises stably co-transfecting a eukaryotic host cell with at least two expression vectors, one such vector harbouring the cDNA sequence encoding the theta GABA receptor subunit, and another such vector harbouring the cDNA sequence encoding an alpha GABA receptor subunit. The stable cell-line which is established expresses an $\alpha\theta$ GABA receptor.

Each receptor thereby expressed, comprising a unique combination of α , θ and optionally one or more subunits selected from β , γ , δ or ϵ subunits, will be referred to hereinafter as a GABA receptor "subunit combination".

Expression of the GABA receptor may be accomplished by a variety of different promoter-expression systems in a variety of different host cells. The eukaryotic host cells suitably include yeast, insect and mammalian cells. Preferably the eukaryotic cells which can provide the host for the expression of the receptor are mammalian cells. Suitable host cells include rodent fibroblast lines, for example mouse Ltk-, Chinese hamster ovary (CHO) and baby hamster kidney (BHK); HeLa; and HEK293 cells. It is necessary to incorporate at least one α subunit, the θ subunit, and optionally one or more subunits selected from β , γ , δ or ϵ into the cell line in order to produce the required receptor. Within this limitation, the choice of receptor subunit combination is made according to the type of activity or selectivity which is being screened for.

In order to employ this invention most effectively for screening purposes, it is preferable to build up a library of cell lines, each with a different combination of subunits. Typically a library of 5 or 6 cell line types is convenient for this purpose. Preferred subunit combinations include: $\alpha\theta\beta$, $\alpha\theta\gamma$, $\alpha\theta\delta$, and $\alpha\theta\epsilon$, and most especially $\alpha_1\theta\gamma_2$. Further preferred subunit combinations include $\alpha\beta\theta\gamma$ and $\alpha\beta\theta\epsilon$, and most especially $\alpha_2\beta_1\theta\gamma_1$ and $\alpha_2\beta_3\theta\gamma_2$.

Cells are then co-transfected with the desired combination of the expression vectors. There are several commonly used techniques for transfection of eukaryotic cells *in vitro*. Calcium phosphate precipitation

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of DNA is most commonly used (Bachetti et al., Proc. Natl. Acad. Sci. USA, 1977, 74, 1590-1594; Maitland et al., Cell, 1977, 14, 133-141), and represents a favoured technique in the context of the present invention.

A small percentage of the host cells takes up the recombinant DNA.

In a small percentage of those, the DNA will integrate into the host cell chromosome. Because an antibitotic resistance marker gene, such as the neomycin or zeocin resistance gene, will have been incorporated into these host cells, they can be selected by isolating the individual clones which will grow in the presence of the chosen antibiotic, e.g. neomycin or zeocin.

Each such clone may then tested to identify those which will produce the receptor. This may be achieved by inducing the production, for example with dexamethasone, and then detecting the presence of receptor by means of radioligand binding.

Alternatively, expression of the GABA receptor may be effected in *Xenopus* oocytes (see, for instance, Hadingham *et al. Mol. Pharmacol.*, 1993, 44, 1211-1218). Briefly, isolated oocyte nuclei are injected directly with injection buffer or sterile water containing at least one alpha subunit, the theta subunit, and optionally one or more beta, gamma, delta or epsilon receptor subunits, engineered into a suitable expression vector. The oocytes are then incubated.

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The expression of subunit combinations in the transfected oocytes may be demonstrated using conventional patch clamp assay. This assay measures the charge flow into and out of an electrode sealed on the surface of the cell. The flow of chloride ions entering the cell via the GABA gated ion channel is measured as a function of the current that leaves the cell to maintain electrical equilibrium within the cell as the gate opens.

In a further aspect, the present invention provides protein preparations of GABA receptor subunit combinations, especially human GABA receptor subunit combinations, derived from cultures of stably transfected eukaryotic cells.

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The protein preparations of GABA receptor subunit combinations can be recovered and purified from recombinant cell cultures by methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography. Protein refolding steps can be used, as necessary, in completing configuration of the mature protein. Finally, high performance liquid chromatography (HPLC) can be employed for final purification steps.

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The polypeptides of the present invention may be a naturally purified product, or a product of chemical synthetic procedures, or produced by recombinant techniques from a prokaryotic or eukaryotic host (for example, by bacterial, yeast, higher plant, insect and mammalian cells in culture). Depending upon the host employed in a recombinant production procedure, the polypeptides of the present invention may be glycosylated or may be non-glycosylated. Polypeptides of the invention may also include an initial methionine amino acid residue.

The GABA theta subunit polypeptide of the present invention is also useful for identifying other subunits of the GABA receptor. An example of a procedure for identifying these subunits comprises raising high titre polyclonal antisera against unique, bacterially expressed GABA theta polypeptides. These polyclonal antisera are then used to immunoprecipitate detergent-solubilized GABA receptors from a mammalian brain, for example, a rat brain.

The invention also provides preparations of membranes containing subunit combinations of the GABA receptor, especially human GABA receptor subunit combinations, derived from cultures of stably transfected eukaryotic cells.

The cell line, and the membrane preparations therefrom, according to the present invention have utility in screening and design of drugs

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which act upon the GABA receptor, for example benzodiazepines, barbiturates, β -carbolines and neurosteroids.

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Receptor localisation studies using $in\ situ$ hybridization in monkey brains shows that the θ subunit has a restricted localisation; residing mainly in components of the limbic system (involved in emotions such as rage, fear, motivation sexual behaviours and feeding); medial septum, cingulate cortex, the amygdala and hippocampal fields, in various hypothalamic nuclei, and in regions that have been associated with pain perception; the cingulate cortex, insular cortex, and in mid brain and pons structures.

The present invention accordingly provides the use of stably cotransfected cell lines described above, and membrane preparations derived therefrom, in screening for and designing medicaments which act upon GABA receptors comprising the 0 subunit. Of particular interest in this context are molecules capable of interacting selectively with GABA receptors made up of varying subunit combinations. As will be readily apparent, the cell line in accordance with the present invention, and the membrane preparations derived therefrom, provide ideal systems for the study of structure, pharmacology and function of the various GABA receptor subtypes. In particular, preferred screens are functional assays utilizing the pharmacological properties of the GABA receptor subunit combinations of the present invention.

Thus, according to a further aspect of the present invention, there is provided a method for determining whether a ligand, not known to be capable of binding to a human GABAA receptor comprising the theta subunit, can bind to a human GABAA receptor comprising the theta subunit, which comprises contacting a mammalian cell comprising DNA molecules encoding at least one alpha receptor subunit, the theta receptor subunit, and optionally one or more beta, gamma, delta or epsilon receptor subunits with the ligand under conditions permitting binding of ligands known to bind to the GABAA receptor, detecting the presence of any of the

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ligand bound to the GABAA receptor comprising the theta subunit, and thereby determining whether the ligand binds to the GABAA receptor comprising the theta subunit. The theta subunit-encoding DNA in the cell may have a coding sequence substantially the same as the coding sequence shown in Figure 1 or Figure 2. Preferably, the mammalian cell is non-neuronal in origin. An example of a non-neuronal mammalian cell is a fibroblast cell such as an Ltk-cell. The preferred method for determining whether a ligand is capable of binding to a human GABAA receptor comprising the theta subunit comprises contacting a transfected non-neuronal mammalian cell (i.e. a cell that does not naturally express any type of GABAA receptor, and thus will only express such a receptor if it is transfected into the cell) expressing a GABAA receptor comprising the theta subunit on its surface, or contacting a membrane preparation from such a transfected cell, with the ligand under conditions which are known to prevail, and thus to be associated with, in vivo binding of the ligands to a GABA_A receptor comprising the theta subunit, detecting the presence of any of the ligand being tested bound to the GABAA receptor comprising the theta subunit on the surface of the cell, and thereby determining whether the ligand binds to a human GABAA receptor comprising the theta subunit. This response system may be based on ion flux changes measured, for example, by scintillation counting (where the ion is radiolabelled) or by interaction of the ion with a fluorescent marker. Particularly suitable ions are chloride ions. Such a host system is conveniently isolated from pre-existing cell lines. Such a transfection system provides a complete response system for investigation or assay of the activity of human GABAA receptors comprising the theta subunit with ligands as described above. Transfection systems are useful as living cell cultures for competitive binding assays between known or candidate drugs and ligands which bind to the receptor and which are labeled by radioactive, spectroscopic or other reagents. Membrane preparations containing the receptor isolated from transfected cells are also useful for

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these competitive binding assays. A transfection system constitutes a "drug discovery system" useful for the identification of natural or synthetic compounds with potential for drug development that can be further modified or used directly as therapeutic compounds to activate, inhibit or modulate the natural functions of human GABAA receptors comprising the theta subunit. The transfection system is also useful for determining the affinity and efficacy of known drugs at human GABAA receptor sites comprising the theta subunit.

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This invention also provides a method of screening drugs to identify drugs which specifically interact with, and bind to, a human GABAA receptor comprising the theta subunit on the surface of a cell which comprises contacting a mammalian cell comprising DNA molecules encoding at least one alpha receptor subunit, the theta receptor subunit and optionally one or more beta, gamma, delta or epsilon receptor subunits on the surface of a cell with a plurality of drugs, determining those drugs which bind to the mammalian cell, and thereby identifying drugs which specifically interact with, and bind to, human GABAA receptors comprising the theta subunit. The theta subunit-encoding DNA in the cell may have a coding sequence substantially the same as the coding sequence shown in Figure 1 or Figure 2. Preferably, the mammalian cell is non-neuronal in origin. An example of a non-neuronal mammalian cell is a fibroblast cell such as an Ltk-cell. Drug candidates are identified by choosing chemical compounds which bind with high affinity to the expressed GABAA receptor protein in transfected cells, using radioligand binding methods well known in the art. Drug candidates are also screened for selectivity by identifying compounds which bind with high affinity to one particular GABAA receptor combination but do not bind with high affinity to any other GABAA receptor combination or to any other known receptor site. Because selective, high affinity compounds interact primarily with the target GABAA receptor site after administration to the patient, the chances of

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producing a drug with unwanted side effects are minimized by this approach.

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In the above screens, the mammalian cell may, for example, comprise DNA molecules encoding at least one alpha receptor subunit, the theta subunit, and optionally one or more gamma receptor subunits and optionally one or more beta receptor subunits.

More preferably, in the above screens, the mammalian cell comprises DNA molecules encoding at least one alpha receptor subunit, at least one gamma receptor subunit and the theta receptor subunit.

Ligands or drug candidates identified above may be agonists or antagonists at human GABAA receptors comprising the theta subunit, or may be agents which allosterically modulate a human GABAA receptor comprising the theta subunit. These ligands or drug candidates identified above may be employed as therapeutic agents, for example, for the modulation of emotions such as rage and fear, of sexual and appetite behaviours and of pain perception.

The ligands or drug candidates of the present invention thus identified as therapeutic agents may be employed in combination with a suitable pharmaceutical carrier. Such compositions comprise a therapeutically effective amount of the agonist or antagonist, and a pharmaceutically acceptable carrier or excipient.

Preferably the compositions containing the ligand or drug candidate identified according to the methods of the present invention are in unit dosage forms such as tablets, pills, capsules, wafers and the like.

Additionally, the therapeutic agent may be presented as granules or powders for extemporaneous formulation as volume defined solutions or suspensions. Alternatively, the therapeutic agent may be presented in ready-prepared volume defined solutions or suspensions. Preferred forms are tablets and capsules.

For preparing solid compositions such as tablets, the principal active ingredient is mixed with a pharmaceutical carrier, e.g. conventional

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tableting ingredients such as corn starch, lactose, sucrose, sorbitol, talc, stearic acid, magnesium stearate, dicalcium phosphate or gums, and other pharmaceutical diluents, e.g. water, to form a solid preformulation composition containing a homogeneous mixture of a compound of the present invention, or a non-toxic pharmaceutically acceptable salt thereof. When referring to these preformulation compositions as homogeneous, it is meant that the active ingredient is dispersed evenly throughout the composition so that the composition may be readily subdivided into equally effective unit dosage forms such as tablets, pills and capsules. This solid preformulation composition is then subdivided into unit dosage forms of the type described above containing from 0.1 to about 500 mg of the active ingredient of the present invention. The tablets or pills of the novel composition can be coated or otherwise compounded to provide a dosage form affording the advantage of prolonged action. For example, the tablet or pill can comprise an inner dosage and an outer dosage component, the latter being in the form of an envelope over the former. The two components can be separated by an enteric layer which serves to resist disintegration in the stomach and permits the inner component to pass intact into the duodenum or to be delayed in release. A variety of materials can be used for such enteric layers or coatings, such materials including a number of polymeric acids and mixtures of polymeric acids with such materials as shellac, cetyl alcohol and cellulose acetate.

The liquid forms in which the novel compositions of the present invention may be incorporated for administration orally include aqueous solutions, suitably flavoured syrups, aqueous or oil suspensions, and flavoured emulsions with edible oils such as cottonseed oil, sesame oil, coconut oil, peanut oil or soybean oil, as well as elixirs and similar pharmaceutical vehicles. Suitable dispersing or suspending agents for aqueous suspensions include synthetic and natural gums such as tragacanth, acacia, alginate, dextran, sodium carboxymethylcellulose, methylcellulose, polyvinyl-pyrrolidone or gelatin.

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Compositions of the present invention may also be administered via the buccal cavity using conventional technology, for example, absorption wafers.

Compositions in the form of tablets, pills, capsules or wafers for oral administration are particularly preferred.

A minimum dosage level for the ligand or drug candidate identified according to the methods of the present invention is about 0.05mg per day, preferably about 0.5mg per day and especially about 2.5mg per day. A maximum dosage level for the ligand or drug candidate is about 3000mg per day, preferably about 1500mg per day and especially about 500mg per day. The compounds are administered on a regimen of 1 to 4 times daily, preferably once or twice daily, and especially once a day.

It will be appreciated that the amount of the therapeutic agent required for use therapy will vary not only with the particular compounds or compositions selected but also with the route of administration, the nature of the condition being treated, and the age and condition of the patient, and will ultimately be at the discretion of the patient's physician or pharmacist.

20 <u>DESCRIPTION OF FIGURES</u>

Figure 1: Nucleotide sequence for the theta subunit, together with its deduced amino acid sequence corresponding thereto (SEQ.ID.NO.1 and SEQ.ID.NO.2, respectively)

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Figure 2: Preferred nucleotide sequence for the theta subunit, together with its deduced amino acid sequence corresponding thereto (SEQ.ID.NO.3 and SEQ.ID.NO.4, respectively).

Figure 3: GABA dose-response curves on HEK cells transiently transfected with and without θ subunit-containing GABA-A receptors ($\alpha_2\beta_1\theta\gamma_1$ and $\alpha_2\beta_1\gamma_1$).

5 The following non-limiting Examples illustrate the present invention.

EXAMPLE 1

10 ISOLATION AND SEQUENCING OF A cDNA ENCODING THE HUMAN GABAA RECEPTOR θ SUBUNIT.

The Genbank database was searched with GABAA receptor polypeptide amino acid sequences using the BLAST searching algorithm, 15 and a number of homologous sequences identified. One of these U47334 was investigated in more detail. U47334 contained sequences homologous to part of the amino-terminal extracellular domain and the TM4 spanning domain of other GABAA receptor subunits, but did not appear to contain any sequence homologous to the regions spanning these domains. 20 Polymerase chain reaction (PCR) was performed to determine if the size if the U47334 sequence was correct, or was for example, the result of an incorrect splicing event. For PCR, a sense (5' gcaaatgaagctgtggttc 3') (SEQ.ID.NO. 5) and antisense (5' caatgttgaacaacccaaag 3') (SEQ.ID.NO. 6) primer were generated from the U47334 sequence, and PCR performed 25 using standard conditions (Whiting et al, PNAS) using human whole brain cDNA (Clontech) as a template. A second PCR reation was then performed using nested sense (5' gcctgagaccgaattttgg 3') (SEQ.ID.NO. 7) and antisense (5' ggaaccgggaccacttgtc 3') (SEQ.ID.NO. 8) primers generated from the U47334 sequence, and using the products from the 30 first PCR as a template. A single PCR product of approximately 1600 bp was obtained suggesting that the U47334 sequence represents an

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incorrectly processed message. This product was sequenced directly using an Applied Biosystems 373 DNA sequencer and dye terminator chemistry.

cDNA sequences 5' and 3' of the U47334 sequence were obtained by 5'- and 3'-anchored PCR using human brain Marathon cDNA cloning kit (Clontech) according to the manufacturer's protocols. The nested antisense (5' tagtccagggtcaagttc 3' and 5' tagtatgctaagcgtgaatc 3') (SEQ.ID.NOS. 9 and 10) and sense (5' gagtttgaggatagttgc 3' and 5' tgctccttcactgaaggg 3') (SEQ.ID.NOS. 11 and 12) primers were derived from both the U47334 sequence and the sequence from the initial PCR amplifications. The PCR products were sequenced directly as previously described.

A full length cDNA was generated by PCR using primers derived from sequences downstream of the innitiating ATG (5' ccatgactcaagcttgccaccatgctgcgagccgcagtgatc 3', incorporating a HindIII site) (SEQ.ID.NO. 13) and in the 3' UT of the anchored PCR product (5' tgaaaggagcacagcacagtgctcccg 3') (SEQ.ID.NO. 14). The PCR product (1958 bp) was cloned into pMOS (Amersham), subcloned into pCDNAI Amp (Invitrogen), and sequenced completey on both strands by primer walking. Sequence analysis was performed using Inherit (Applied Biosystems), Sequencher (Genecodes), and Genetics Computer Group (Univ. Wisconsin) computer programs.

The coding region encodes 627 amino acids and has all the structural motifs expected of a ligand gated ion channel subunit. Comparison with other ligand gated ion channel subunits indicates that it is most similar to GABA_A receptor subunits, the highest homology being with the β_1 subunit (45 % identity). However, this sequence identity is sufficiently low as to indicate that the new subunit cannot be classified as a fourth human β subunit, but represents a novel class of subunit, classified as θ , within the GABA receptor gene family.

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EXAMPLE 2

LOCALISATION OF THE θ SUBUNIT IN MONKEY BRAIN BY *IN SITU* HYBRIDISATION.

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Antisense oligonucleotide probes to the human θ subunit sequence were generated on an Applied Biosystems Automated DNA synthesiser Probe 1

10 5' CTG-CTT-GCA-CAC-CCT-TCT-CGC-CAT-GGT-GAA-GCA-TGG-GCT-TCC 3' (SEQ.ID.NO. 15)

Probe 2

5'TGT-CGC-CTA-GGC-TGG-CGC-CGA-GGT-CCT-CGA-CTG-TAG-AAA-AGA-TAG 3' (SEQ.ID.NO. 16)

Each oligonucleotide was 3'-end labelled with [35S] deoxyadenosine 5'-(thiotriphosphate) in a 30:1 molar ratio of ³⁵S-isotope:oligonucleotide using terminal deoxynucleotidyl transferase for 15 min at 37°C in the reaction buffer supplied. Radiolabelled oligonucleotide was separated from unincorporated nucleotides using Sephadex G50 spin columns. The specific activities of the labelled probes in several labelling reactions varied from 1.2-2.3 x 109 cpm/mg. Monkey brains were removed and fresh frozen in 1 cm blocks. 12 µm sections were taken and fixed for in situ hybridisation. Hybridisation of the sections was carried out according to the method of Sirinathsingji and Dunnett (Imaging gene expression in neural graft; Molecular Imaging in Neuroscience: A Practical Approach. N.A. Sharif (ed), Oxford University Press, Oxford, pp43-70, 1993). Briefly, sections were removed from alcohol, air dried and 3 x105 cpm of each ³⁵S-labelled probe in 100μl of hybridisation buffer was applied to each slide. Labelled "antisense" probe was also used in the presence of an excess (100x) concentration of unlabelled antisense probe to define nonspecific hybridisation. Parafilm coverslips were placed over the sections

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which were incubated overnight (about 16 hr) at 37°C. Following hybridisation the sections were washed for 1 hr at 57°C in 1xSSC then rinsed briefly in 0.1 x SSC, dehydrated in a series of alcohols, air dried and exposed to Amersham Hyperfilm βmax X-ray film and the relative distribution of the mRNA assessed for a variety of brain regions.

Messenger RNA for the subunit was seen in components of the limbic system (involved in emotions such as rage, fear, motivation sexual behaviours and feeding); medial septum, cingulate cortex, the amygdala and hippocampal fields (dentate gyrus, CA3, CA2, CA1) and in various hypothalamic nuclei (often associated with the limbic system). Messenger RNA was also present in regions that have been associated with pain perception; the cingulate cortex, insular cortex, and in mid brain and pons structures (e.g. central grey and reticular formation).

EXAMPLE 3

LOCALISATION OF THE θ SUBUNIT IN MONKEY BRAIN BY WESTERN BLOT ANALYSIS AND IMMUNOCYTOCHEMISTRY

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Antibodies to the human GABAA Theta subunit were generated by sub-cutaneous injection of two New Zealand White rabbits with a glutathione-S-transferase (GST) fusion protein consisting of residues 353-595 of the large cytoplasmic loop region of the theta subunit. DNA encoding this region was cloned into the bacterial expression vector pGEX-2T (Pharmacia), transformed into $E.\ coli$ DH10B cells (Life Technologies), and expression of the fusion protein was carried out using the Pharmacia protocols. The bacterial cells were incubated on ice in STE solution (150 mM NaCl, 10 mM Tris-HCl pH 8, 1 mM EDTA) containing 100 µg/ml Lysozyme for 20 min before the addition of N-lauryl sarkosine to 1.5 % (w/v). The bacterial slurry was sonicated on ice, and any insoluble matter

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removed by centrifugation. Triton X-100 was added to 3 % (v/v) final and the GST-fusion protein purified by glutathione-agarose affinity chromatography. Columns were washed extensively with PBS and the bound protein eluted with 20 mM free glutathione in 150 mM NaCl, 100 mM Tris-HCl pH 9, 1 mM EDTA, 1 mM Dithiothreitol. Eluted protein was concentrated by precipitation with 5 volumes of cold acetone, resuspended in water, and stored at -70 °C until use.

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For western blot analysis tissue samples were removed and dissected out on a glass plate at 4°C. The tissue was homogenised in 50mM Tris, pH 7.5, containing 1mM PMSF, 1µM pepstatin A. The homogenate was centrifuged (2000 X g) for 10 minutes and the supernatant was centrifuged at 20,000 X g for 45 minutes. The pellet was resuspended in 50mM Tris and recentrifuged. The final pellet was resuspended in 50mM Tris pH 7.4 containing protease inhibitors and detergent (Na-deoxycholate:0.25%, 150mM NaCl, 1mM EDTA, 1mM PMSF, 1µM pepstatin and leupeptin. Membrane preparations were separated on a 10 % Tris tricine polyacrylamide gel and electrophoretically transferred to nitrocellulose. Nitrocellulose was blocked with 5% non-fat milk (marvelTM)/PBS/Tween (0.5%) for 1 hour at room temperature. The anti θ subunit antibody was used at a concentration of 1:500 made up in PBS/Tween/milk at 4°C overnight, washed and then incubated with anti-rabbit IgG HRP linked (Amersham) at 1:1000 in PBS/Tween/milk for one hour at room temperature. The filters were washed, incubated in ECL (Amersham) for 1min and opposed to film. A single band of approximately 60-66kDa was visualised in brainstem and striatal membranes, close to the predicted molecular weight for the θ subunit of 68-74 kDa.

For localisation of the θ subunit by immunocytochemistry a rhesus monkey was deeply anesthetised with ketamine and sodium

30 pentobarbitone and transcardially perfused with saline, followed by 10% formal saline. The brain was removed, post fixed for 24 hours, and sliced

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into coronal blocks, which were then dehydrated through graded alcohols, cleared and embedded in paraffin wax. Coronal sections (8µm) were cut on a base sledge microtome and mounted on glass microscope slides. Sections were deparaffinised, rehydrated and rinsed in 0.1M phosphate buffered saline (PBS). In order to enhance the immunoreactivity sections were subjected to antigen retrieval techniques. Briefly, sections were placed in 0.1M citrate buffer pH 6.0 and given two 5 minute bursts at full power in a conventional microwave oven (800W). Once rinsed in PBS, sections were incubated in 5% normal goat serum in PBS, for 1 hr to block background staining. Sections were then incubated overnight at +4°C in the anti θ subunit rabbit polyclonal antibody (1:1000 diluted in blocking buffer). Immunoreactivity was visualised using the Vector *elite*™ system (Vector Laboratories, Peterborough, U.K.), followed by development in diaminobenzidine (DAB) (Sigma, U.K.). Sections were counterstained in Gill's haematoxylin (Biomen, High Wycombe, U.K.), dehydrated and mounted for microscopical examination. For comparison, samples of 10% formalin immersion fixed post mortem human brainstem were processed in an identical manner. Comparable sections were used to detect θ subunit and tyrosine hydroxylase (Institut Jacques Boy, Reims, France) immunoreactivity by the application of 35S-labeled goat anti rabbit immunoglobulin 1:100 (Amersham Life Sciences, U.K.) for 1 hr. Slides were rinsed in distilled water, dehydrated to 95% ethanol, air dried and exposed to Amersham Hyperfilm βmax. Sections used for the immunofluorescent colocalisation of θ subunit and tyrosine hydroxylase were pretreated in the same manner, anti θ subunit immunoreactivity was detected using firstly a biotinylated anti rabbit ;1:200 (Vector Laboratories) followed by FITC conjugated streptavidin (Sigma, U.K.). The second rabbit polyclonal serum, anti tyrosine hydroxylase, was again visualised using biotinylated anti rabbit, reacted with Cy3 conjugated strepavidin (Sigma, U.K.). Sections were counterstained with Hoescht

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33258 (0.5μg/ml). To avoid any crossreactivity of the detection systems, sections were placed in boiling distilled water for 5 minutes prior to the application of the second primary antibody and its subsequent detection.. The distribution of the θ subunit immunoreactivity in monkey brain reflected the distribution of the θ mRNA observed by in situ hybridisation studies (Example 2). Labelled neurons were observed of hypothalamic and cortical pyramidal neurones. Significant labellingwas observed of cells in the brainstem, including the substantia nigra pars compacta, ventral and lateral tegmental areas, pigmented neurones of the locus coeruleus and restricted population within the dorsal raphe. Labelling of cell terminals within the caudate putamen was also observed. This distribution was found to closely resemble the distribution of tyrosine hydroxylase immunoreactivity, a marker of catocholaminergic neurones and their processes, visualised by immunoautoradiography. θ subunit colocalisation with tyrosine hydroxylase containing neurons was confirmed, using combination immunofluorescence. The expression of the θ subunit seen in both the catocholaminergic neurons of the substantia nigra pars compacta and locus coeruleus was further substantiated in sections of human post mortem brainstem.

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EXAMPLE 4

CONSTRUCTION OF AN LTK- CELL LINE EXPRESSING THE THETA RECEPTOR SUBUNIT

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A chimeric construct of the theta subunit was constructed in the mammalian expression vector pcDNA3.1Zeo (Invitrogen) that consisted of bases -224 to +99 of bovine GABAA α 1 gene, a sequence encoding the c-myc epitope tag (residues 410-419 of the human oncogene product c-myc), a cloning site encoding the amino acids aparagine - serine - glycine, and

DNA encoding residues 22-627 of the GABA_A θ gene product. This construct was linearised and the DNA transfected into a clonal population of mouse Ltk- cells that had previously been shown to be stably transfected with the GABA_A receptor subunits $\alpha_2\beta_1\gamma_1$ and separately an Ltk- line stably transfected with $\alpha_2\beta_3\gamma_2$. The resultant cells were clonally selected with Zeocin selection (100 µg/ml), and screened to verify stable intrgration and expression of $\alpha_2\beta_1\theta\gamma_1$ and $\alpha_2\beta_3\theta\gamma_2$ respectively.

EXAMPLE 5

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WHOLE CELL PATCH-CLAMP OF HEK 293 CELLS TRANSIENTLY TRANSFECTED WITH HUMAN GABA-A RECEPTORS

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Experiments were performed on HEK 293 cells transiently transfected with human cDNA combinations α2β1γ1, and α2β1θγ1 (4μgs of cDNA total per cover-slip) using calcium phosphate precipitation (Chen and Okayama, 1988) as previously described (Hadingham et al. 1993). Glass cover-slips containing the cells in a monolayer culture were transferred to a perspex chamber on the stage of Nikon Diaphot inverted microscope. Cells were continuously perfused with a solution containing 124mM NaCl, 2mM KCl, 2mM CaCl₂, 1mM MgCl₂, 1.25mM KH₂PO₄, 25mM NaHCO₃, 11mM D-glucose, at pH 7.2, and observed using phasecontrast optics. Patch-pipettes were pulled with an approximate tip diameter of $2\mu m$ and a resistance of $4M\Omega$ with borosilicate glass and filled with 130mM CsCl, 10mM HEPES, 10mM EGTA, 3mM Mg+-ATP, pH adjusted to 7.3 with CsOH. Cells were patch-clamped in whole-cell mode using an Axopatch 200B patch-clamp amplifier. Drug solutions were applied by a double-barrelled pipette assembly, controlled by a stepping motor attached to a Prior manipulator, enabling rapid equilibration

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around the cell. Increasing GABA concentrations were applied for 2sec pulses with a 30sec interval between applications. Non-cumulative concentration-response curves to GABA were constructed. Curves were fitted using a non-linear square-fitting program to the equation $f(x) = B_{\text{MAX}}/[1+(\text{EC}_{50}/x)^n]$ where x is the drug concentration, EC₅₀ is the concentration of drug eliciting a half-maximal response and n is the Hill coefficient. EC₅₀'s were analysed by unpaired students t-test.

The GABA EC₅₀ of HEK 293 cells transiently expressing the GABA_A receptor subunit combination $\alpha_2\beta_1\theta\gamma_1$ is significantly lower than that of HEK 293 cells transiently expressing the GABA_A receptor subunit combination $\alpha_2\beta_1\gamma_1$ (see Figure 3).

	$\alpha_2\beta_1\gamma_1$	$\alpha_2\beta_1\theta\gamma_1$
EC_{50}	$16.7\pm3.7~\mathrm{nM}$	62.7±6.7 nM*
Slope	1.6 ± 0.2	1.5 ± 0.1

* p<0.001

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT:
 - (A) NAME: Merck Sharp & Dohme Limited
 - (B) STREET: Terlings Park, Eastwick Road
 - (C) CITY: Harlow
 - (D) STATE: Essex
 - (E) COUNTRY: England
 - (F) POSTAL CODE (ZIP): CM20 2QR
 - (G) TELEPHONE: +44 1279 440175
 - (H) TELEFAX: +44 1279 440717
- (ii) TITLE OF INVENTION: Human theta subunit of the GABA-A receptor
- (iii) NUMBER OF SEQUENCES: 16
- (iv) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)
- (2) INFORMATION FOR SEQ ID NO: 1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1884 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:
- ATGCTGCGAG CCGCAGTGAT CCTGCTGCTC ATCAGGACCT GGCTCGCGGA GGGCAACTAC 60

 CCCAGTCCCA TCCCGAAATT CCACTTCGAG TTCTCCTCTG CTGTGCCCGA AGTCGTCCTG 120

 AACCTCTTCA ACTGCAAAAA TTGTGCAAAT GAAGCTGTGG TTCAAAAGAT TTTGGACAGG 180

 GTGCTGTCAA GATACGATGT CCGCCTGAGA CCGAATTTTG GAGGTGCCCC TGTGCCTGTG 240

 AGAATATCTA TTTATGTCAC GAGCATTGAA CAGATCTCAG AAATGAATAT GGACTACACG 300

 ATCACGATGT TTTTTCATCA GACTTGGAAA GATTCACGCT TAGCATACTA TGAGACCACC 360

CTGAACTTGA CCCTGGACTA TCGGATGCAT GAGAAGTTGT GGGTCCCTGA CTGCTACTTT 420 CTGAACAGCA AGGATGCTTT CGTGCATGAT GTGACTGTGG AGAATCGCGT GTTTCAGCTT 480 CACCCAGATG GAACGGTGCG GTACGGCATC CGACTCACCA CTACAGCAGT TTGTTCCCTG 540 GATCTGCATA AATTCCCTAT GGACAAGCAG GCCTGCAACC TGGTGGTAGA GAGCTATGGT 600 TACACGGTTG AAGACATCAT ATTATTCTGG GATGACAATG GGAACGCCAT CCACATGACT 660 GAGGAGCTGC ATATCCCTCA GTTCACTTTC CTGGGAAGGA CGATTACTAG CAAGGAGGTG 720 TATTTCTACA CAGGTTCCTA CATACGCCTG ATACTGAAGT TCCAGGTTCA GAGGGAAGTT 780 AACAGCTACC TTGTGCAAGT CTACTGGCCT ACTGTCCTCA CCACTATTAC CTCTTGGATA 840 TCGTTTTGGA TGAACTATGA TTCCTCTGCA GCCAGGGTGA CAATTGGCTT AACTTCAATG 900 CTCATCCTGA CCACCATCGA CTCACATCTG CGGGATAAGC TCCCCAACAT TTCCTGTATC 960 AAGGCCATTG ATATCTATAT CCTCGTGTGC TTGTTCTTTG TGTTCCTGTC CTTGCTGGAG 1020 TATGTCTACA TCAACTATCT TTTCTACAGT CGAGGACCTC GGCGCCAGCC TAGGCGACGC 1080 AGGAGACCCC GAAGAGTCAT TGCCCGCTAC CGCTACCAGC AAGTGGTGGT AGGAAACGTG 1140 CAGGATGGCC TGATTAACGT GGAAGACGGA GTCAGCTCTC TCCCCATCAC CCCAGCGCAG 1200 GCCCCCTGG CAAGCCCGGA AAGCCTCGGT TCTTTGACGT CCACCTCCGA GCAGGCCCAG 1260 ACTGGAGAAA GCCTGAGCGA TCTCCCCTCC ACCTCAGAGC AGGCCCGGCA CAGCTATGGT 1380 GTTCGCTTTA ATGGTTTCCA GGCTGATGAC AGTATTATTC CTACCGAAAT CCGCAACCGT 1440 GTCGAAGCCC ATGGCCATGG TGTTACCCAT GACCATGAAG ATTCCAATGA GAGCTTGAGC 1500 TCGGATGAGC GCCATGGCCA TGGCCCCAGT GGGAAGCCCA TGCTTCACCA TGGCGAGAAG 1560 GGTGTGCAAG AAGCAGGCTG GGACCTTGAT GACAACAATG ACAAGAGCGA CTGCCTTGCC 1620 ATTAAGGAGC AATTCAAGTG TGATACTAAC AGTACCTGGG GCCTTAATGA TGATGAGCTC 1680 GTGGCCCATG GCCAAGAGAA GGACAGTAGC TCAGAGTCTG AGGATAGTTG CCCCCCAAGC 1740 CCTGGGTGCT CCTTCACTGA AGGGTTCTCC TTCGATCTCT TTAATCCTGA CTACGTCCCA 1800 AAGGTCGACA AGTGGTCCCG GTTCCTCTTC CCTCTGGCCT TTGGGTTGTT CAACATTGTT 1860 TACTGGGTAT ACCATATGTA TTAG 1884

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 627 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Met Leu Arg Ala Ala Val Ile Leu Leu Leu Ile Arg Thr Trp Leu Ala Glu Gly Asn Tyr Pro Ser Pro Ile Pro Lys Phe His Phe Glu Phe Ser 25 Ser Ala Val Pro Glu Val Val Leu Asn Leu Phe Asn Cys Lys Asn Cys 40 Ala Asn Glu Ala Val Val Gln Lys Ile Leu Asp Arg Val Leu Ser Arg 50 55 60 Tyr Asp Val Arg Leu Arg Pro Asn Phe Gly Gly Ala Pro Val Pro Val Arg Ile Ser Ile Tyr Val Thr Ser Ile Glu Gln Ile Ser Glu Met Asn 90 Met Asp Tyr Thr Ile Thr Met Phe His Gln Thr Trp Lys Asp Ser 100 Arg Leu Ala Tyr Tyr Glu Thr Thr Leu Asn Leu Thr Leu Asp Tyr Arg Met His Glu Lys Leu Trp Val Pro Asp Cys Tyr Phe Leu Asn Ser Lys 130 135 Asp Ala Phe Val His Asp Val Thr Val Glu Asn Arg Val Phe Gln Leu 145 150 His Pro Asp Gly Thr Val Arg Tyr Gly Ile Arg Leu Thr Thr Ala 165 170 Val Cys Ser Leu Asp Leu His Lys Phe Pro Met Asp Lys Gln Ala Cys 180 Asn Leu Val Val Glu Ser Tyr Gly Tyr Thr Val Glu Asp Ile Ile Leu 200 Phe Trp Asp Asp Asn Gly Asn Ala Ile His Met Thr Glu Glu Leu His 215 Ile Pro Gln Phe Thr Phe Leu Gly Arg Thr Ile Thr Ser Lys Glu Val 230 235 240

Tyr	Phe	Tyr	Thr	Gly 245	Ser	Tyr	Ile	Arg	Leu 250	Ile	Leu	Lys	Phe	Gln 255	Val
Gln	Arg	Glu	Val 260	Asn	Ser	Tyr	Leu	Val 265	Gln	Val	Tyr	Trp	Pro 270	Thr	Val
Leu	Thr	Thr 275	Ile	Thr	Ser	Trp	Ile 280	Ser	Phe	Trp	Met	Asn 285	Tyr	Asp	Ser
Ser	Ala 290	Ala	Arg	Val	Thr	Ile 295	Gly	Leu	Thr	Ser	Met 300	Leu	Ile	Leu	Thr
Thr 305	Ile	Asp	Ser	His	Leu 310	Arg	Asp	Lys	Leu	Pro 315	Asn	Ile	Ser	Cys	Ile 320
Lys	Ala	Ile	Asp	Ile 325	Tyr	Ile	Leu	Val	Cys 330	Leu	Phe	Phe	Val	Phe 335	Leu
Ser	Leu	Leu	Glu 340	Tyr	Val	Tyr	Ile	Asn 345	Tyr	Leu	Phe	Tyr	Ser 350	Arg	Gly
Pro	Arg	Arg 355	Gln	Pro	Arg	Arg	Arg 360	Arg	Arg	Pro	Arg	Arg 365	Val	Ile	Ala
Arg	Tyr 370	Arg	Tyr	Gln	Gln	Val 375	Val	Val	Gly	Asn	Val 380	Gln	Asp	Gly	Leu
Ile 385	Asn	Val	Glu	Asp	Gly 390	Val	Ser	Ser	Leu	Pro 395	Ile	Thr	Pro	Ala	Gln 400
Ala	Pro	Leu	Ala	Ser 405	Pro	Glu	Ser	Leu	Gly 410	Ser	Leu	Thr	Ser	Thr 415	Ser
Glu	Gln	Ala	Gln 420	Leu	Ala	Thr	Ser	Glu 425	Ser	Leu	Ser	Pro	Leu 430	Thr	Ser
Leu	Ser	Gly 435	Gln	Ala	Pro	Leu	Ala 440	Thr	Gly	Glu	Ser	Leu 445	Ser	Asp	Leu
Pro	Ser 450	Thr	Ser	Glu	Gln	Ala 455	Arg	His	Ser	Tyr	Gly 460	Val	Arg	Phe	Asn
Gly 465	Phe	Gln	Ala	Asp	Asp 470	Ser	Ile	Ile	Pro	Thr 475	Glu	Ile	Arg	Asn	Arg 480
Val	Glu	Ala	His	Gly 485	His	Gly	Val	Thr	His 490	Asp	His	Glu	Asp	Ser 495	Asn
Glu	Ser	Leu	Ser 500	Ser	Asp	Glu	Arg	His 505	Gly	His	Gly	Pro	Ser 510	Gly	Lys
Pro	Met	Leu 515	His	His	Gly	Glu	Lys 520	Gly	Val	Gln	Glu	Ala 525	Gly	Trp	Asp
Leu	Asp 530	Asp	Asn	Asn	Asp	Lys 535	Ser	Asp	Cys	Leu	Ala 540	Ile	Lys	Glu	Gln

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Phe Lys Cys Asp Thr Asn Ser Thr Trp Gly Leu Asn Asp Asp Glu Leu 545 550 555 555

Val Ala His Gly Gln Glu Lys Asp Ser Ser Ser Glu Ser Glu Asp Ser 565 570 575

Cys Pro Pro Ser Pro Gly Cys Ser Phe Thr Glu Gly Phe Ser Phe Asp 580 585 590

Leu Phe Asn Pro Asp Tyr Val Pro Lys Val Asp Lys Trp Ser Arg Phe 595 600 605

Leu Phe Pro Leu Ala Phe Gly Leu Phe Asn Ile Val Tyr Trp Val Tyr 610 620

His Met Tyr 625

(2) INFORMATION FOR SEQ ID NO: 3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1884 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

ATGCTGCGAG CCGCAGTGAT CCTGCTGCTC ATCAGGACCT GGCTCGCGGA GGGCAACTAC 60

CCCAGTCCCA TCCCGAAATT CCACTTCGAG TTCTCCTCTG CTGTGCCCGA AGTCGTCCTG 120

AACCTCTTCA ACTGCAAAAA TTGTGCAAAT GAAGCTGTGG TTCAAAAGAT TTTGGACAGG 180

GTGCTGTCAA GATACGATGT CCGCCTGAGA CCGAATTTTG GAGGTGCCCC TGTGCCTGTG 240

AGAATATCTA TTTATGTCAC GAGCATTGAA CAGATCTCAG AAATGAATAT GGACTACACG 300

ATCACGATGT TTTTCATCA GACTTGGAAA GATTCACGCT TAGCATACTA TGAGACCACC 360

CTGAACTTGA CCCTGGACTA TCGGATGCAT GAGAAGTTGT GGGTCCCTGA CTGCTACTTT 420

TTGAACAGCA AGGATGCTTT CGTGCATGAT GTGACTGTGG AGAATCGCGT GTTTCAGCTT 480

CACCCAGATG GAACGGTGCG GTACGGCATC CGACTCACCA CTACAGCAGC TTGTTCCCTG 540

GATCTGCATA AATTCCCTAT GGACAAGCAG GCCTGCAACC TGGTGGTAGA GAGCTATGGT 600

TACACGGTTG AAGACATCAT ATTATTCTGG GATGACAATG GGAACGCCAT CCACATGACT 660

GAGGAGCTGC ATATCCCTCA GTTCACTTTC CTGGGAAGGA CGATTACTAG CAAGGAGGTG 720 TATTTCTACA CAGGTTCCTA CATACGCCTG ATACTGAAGT TCCAGGTTCA GAGGGAAGTT 780 AACAGCTACC TTGTGCAAGT CTACTGGCCT ACTGTCCTCA CCACTATTAC CTCTTGGATA 840 TCGTTTTGGA TGAACTATGA TTCCTCTGCA GCCAGGGTGA CAATTGGCTT AACTTCAATG 900 CTCATCCTGA CCACCATCGA CTCACATCTG CGGGATAAGC TCCCCAACAT TTCCTGTATC 960 AAGGCCATTG ATATCTATAT CCTCGTGTGC TTGTTCTTTG TGTTCCTGTC CTTGCTGGAG 1020 TATGTCTACA TCAACTATCT TTTCTACAGT CGAGGACCTC GGCGCCAGCC TAGGCGACAC 1080 AGGAGACCCC GAAGAGTCAT TGCCCGCTAC CGCTACCAGC AAGTGGTGGT AGGAAACGTG 1140 CAGGATGGCC TGATTAACGT GGAAGACGGA GTCAGCTCTC TCCCCATCAC CCCAGCGCAG 1200 GCCCCCTGG CAAGCCCGGA AAGCCTCGGT TCTTTGACGT CCACCTCCGA GCAGGCCCAG 1260 ACTGGAGAAA GCCTGAGCGA TCTCCCCTCC ACCTCAGAGC AGGCCCGGCA CAGCTATGGT 1380 GTTCGCTTTA ATGGTTTCCA GGCTGATGAC AGTATTTTTC CTACCGAAAT CCGCAACCGT 1440 GTCGAAGCCC ATGGCCATGG TGTTACCCAT GACCATGAAG ATTCCAATGA GAGCTTGAGC 1500 TCGGATGAGC GCCATGGCCA TGGCCCCAGT GGGAAGCCCA TGCTTCACCA TGGCGAGAAG 1560 GGTGTGCAAG AAGCAGGCTG GGACCTTGAT GACAACAATG ACAAGAGCGA CTGCCTTGCC 1620 ATTAAGGAGC AATTCAAGTG TGATACTAAC AGTACCTGGG GCCTTAATGA TGATGAGCTC 1680 ATGGCCCATG GCCAAGAGAA GGACAGTAGC TCAGAGTCTG AGGATAGTTG CCCCCCAAGC 1740 CCTGGGTGCT CCTTCACTGA AGGGTTCTCC TTCGATCTCT TTAATCCTGA CTACGTCCCA 1800 AAGGTCGACA AGTGGTCCCG GTTCCTCTC CCTCTGGCCT TTGGGTTGTT CAACATTGTT 1860 TACTGGGTAT ACCATATGTA TTAG 1884

(2) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 627 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

Met 1	Leu	Arg	Ala	Ala 5	Val	Ile	Leu	Leu	Leu 10	Ile	Arg	Thr	Trp	Leu 15	Ala
Glu	Gly	Asn	Tyr 20	Pro	Ser	Pro	Ile	Pro 25	Lys	Phe	His	Phe	Glu 30	Phe	Ser
Ser	Ala	Val 35	Pro	Glu	Val	Val	Leu 40	Asn	Leu	Phe	Asn	Cys 45	Lys	Asn	Cys
Ala	Asn 50	Glu	Ala	Val	Val	Gln 55	Lys	Ile	Leu	Asp	Arg 60	Val	Leu	Ser	Arg
Tyr 65	Asp	Val	Arg	Leu	Arg 70	Pro	Asn	Phe	Gly	Gly 75	Ala	Pro	Val	Pro	Val 80
Arg	Ile	Ser	Ile	Tyr 85	Val	Thr	Ser	Ile	Glu 90	Gln	Ile	Ser	Glu	Met 95	Asn
Met	Asp	Tyr	Thr 100	Ile	Thr	Met	Phe	Phe 105	His	Gln	Thr	Trp	Lys 110	Asp	Ser
Arg	Leu	Ala 115	Tyr	Tyr	Glu	Thr	Thr 120	Leu	Asn	Leu	Thr	Leu 125	Asp	Tyr	Arg
Met	His 130	Glu	Lys	Leu	Trp	Val 135	Pro	Asp	Cys	Tyr	Phe 140	Leu	Asn	Ser	Lys
Asp 145	Ala	Phe	Val	His	Asp 150	Val	Thr	Val	Glu	Asn 155	Arg	Val	Phe	Gln	Leu 160
His	Pro	Asp	Gly	Thr 165	Val	Arg	Tyr	Gly	Ile 170	Arg	Leu	Thr	Thr	Thr 175	Ala
Ala	Cys	Ser	Leu 180	Asp	Leu	His	Lys	Phe 185	Pro	Met	Asp	Lys	Gln 190	Ala	Cys
Asn	Leu	Val 195	Val	Glu	Ser	Tyr	Gly 200	Tyr	Thr	Val	Glu	Asp 205	Ile	Ile	Leu
Phe	Trp 210	Asp	Asp	Asn	Gly	Asn 215	Ala	Ile	His	Met	Thr 220	Glu	Glu	Leu	His
Ile 225	Pro	Gln	Phe	Thr	Phe 230	Leu	Gly	Arg	Thr	Ile 235	Thr	Ser	Lys	Glu	Val 240
Tyr	Phe	Tyr	Thr	Gly 245	Ser	Tyr	Ile	Arg	Leu 250	Ile	Leu	Lys	Phe	Gln 255	Val
Gln	Arg	Glu	Val 260	Asn	Ser	Tyr	Leu	Val 265	Gln	Val	Tyr	Trp	Pro 270	Thr	Val
Leu	Thr	Thr 275	Ile	Thr	Ser	Trp	Ile 280	Ser	Phe	Trp	Met	Asn 285	Tyr	Asp	Ser

Ser		Ala	Arg	Val	Thr		Gly	Leu	Thr	Ser		Leu	Ile	Leu	Thr
	290					295					300				
Thr 305	Ile	Asp	Ser	His	Leu 310	Arg	Asp	Lys	Leu	Pro 315	Asn	Ile	Ser	Cys	Ile 320
Lys	Ala	Ile	Asp	Ile 325	Tyr	Ile	Leu	Val	Cys 330	Leu	Phe	Phe	Val	Phe 335	Leu
Ser	Leu	Leu	Glu 340	Tyr	Val	Tyr	Ile	Asn 345	Tyr	Leu	Phe	Tyr	Ser 350	Arg	Gly
Pro	Arg	Arg 355	Gln	Pro	Arg	Arg	His 360	Arg	Arg	Pro	Arg	Arg 365	Val	Ile	Ala
Arg	Tyr 370	Arg	Tyr	Gln	Gln	Val 375	Val	Val	Gly	Asn	Val 380	Gln	Asp	Gly	Leu
Ile 385	Asn	Val	Glu	Asp	Gly 390	Val	Ser	Ser	Leu	Pro 395	Ile	Thr	Pro	Ala	Gln 400
Ala	Pro	Leu	Ala	Ser 405	Pro	Glu	Ser	Leu	Gly 410	Ser	Leu	Thr	Ser	Thr 415	Ser
Glu	Gln	Ala	Gln 420	Leu	Ala	Thr	Ser	Glu 425	Ser	Leu	Ser	Pro	Leu 430	Thr	Ser
Leu	Ser	Gly 435	Gln	Ala	Pro	Leu	Ala 440	Thr	Gly	Glu	Ser	Leu 445	Ser	Asp	Leu
Pro	Ser 450	Thr	Ser	Glu	Gln	Ala 455	Arg	His	Ser	Tyr	Gly 460	Val	Arg	Phe	Asn
Gly 465	Phe	Gln	Ala	Asp	Asp 470	Ser	Ile	Phe	Pro	Thr 475	Glu	Ile	Arg	Asn	Arg 480
Val	Glu	Ala	His	Gly 485	His	Gly	Val	Thr	His 490	Asp	His	Glu	Asp	Ser 495	Asn
Glu	Ser	Leu	Ser 500	Ser	Asp	Glu	Arg	His 505	Gly	His	Gly	Pro	Ser 510	Gly	Lys
Pro	Met	Leu 515	His	His	Gly	Glu	Lys 520	Gly	Val	Gln	Glu	Ala 525	Gly	Trp	Asp
Leu	Asp 530	Asp	Asn	Asn	Asp	Lys 535	Ser	Asp	Cys	Leu	Ala 540	Ile	Lys	Glu	Gln
Phe 545	Lys	Cys	Asp	Thr	Asn 550	Ser	Thr	Trp	Gly	Leu 555	Asn	Asp	Asp	Glu	Leu 560
Met	Ala	His	Gly	Gln 565	Glu	Lys	Asp	Ser	Ser 570	Ser	Glu	Ser	Glu	Asp 575	Ser

595 600 605

Leu Phe Pro Leu Ala Phe Gly Leu Phe Asn Ile Val Tyr Trp Val Tyr 610 615 620

His Met Tyr 625

(2) INFORMATION FOR SEQ ID NO: 5:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 19 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

GCAAATGAAG CTGTGGTTC

19

- (2) INFORMATION FOR SEQ ID NO: 6:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

CAATGTTGAA CAACCCAAAG

20

- (2) INFORMATION FOR SEQ ID NO: 7:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 19 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

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(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:	
GCCTGAGACC GAATTTTGG	19
(2) INFORMATION FOR SEQ ID NO: 8:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 19 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:	
GGAACCGGGA CCACTTGTC	19
(2) INFORMATION FOR SEQ ID NO: 9:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:	
TAGTCCAGGG TCAAGTTC	18
(2) INFORMATION FOR SEQ ID NO: 10:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	·
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:	
TAGTATGCTA AGCGTGAATC	20

(2) INFORMATION FOR SEQ ID NO: 11:

 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:	
GAGTTTGAGG ATAGTTGC	18
(2) INFORMATION FOR SEQ ID NO: 12:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:	
TGCTCCTTCA CTGAAGGG	18
(2) INFORMATION FOR SEQ ID NO: 13:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 42 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:	
CCATGACTCA AGCTTGCCAC CATGCTGCGA GCCGCAG	TGA TC 42
(2) INFORMATION FOR SEQ ID NO: 14:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 27 base pairs	

- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

TGAAAGGAGC ACAGCACAGT GCTCCCG

27

- (2) INFORMATION FOR SEQ ID NO: 15:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 45 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

CTGCTTCTTG CACACCCTTC TCGCCATGGT GAAGCATGGG CTTCC 45

- (2) INFORMATION FOR SEQ ID NO: 16:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 45 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

TGTCGCCTAG GCTGGCGCCG AGGTCCTCGA CTGTAGAAAA GATAG 4

CLAIMS:

- 1. A stably co-transfected eukaryotic cell line capable of expressing a GABA receptor, which receptor comprises the theta receptor subunit, at least one alpha receptor subunit and optionally one or more receptor subunits selected from the beta, gamma, delta and epsilon subunits.
- 2. A cell line according to claim 1 which is a rodent fibroblast 10 cell line.
 - 3. A process for the preparation of an eukaryotic cell line capable of expressing a GABA receptor, which comprises stably cotransfecting a eukaryotic host cell with at least two expression vectors, one such vector harbouring the cDNA sequence encoding the theta GABA receptor subunit, another such vector harbouring the cDNA sequence encoding an alpha GABA receptor subunit, and optionally one or more additional vectors harbouring the cDNA sequence encoding a beta, gamma, delta or epsilon GABA receptor subunit.

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- 4. A process according to claim 3 wherein the cell line is a rodent fibroblast cell line.
- 5. A DNA molecule encoding the theta subunit of the human GABA receptor comprising all or a portion of the sequence depicted in Figure 1 or Figure 2, or a modified human sequence.
 - 6. A recombinant expression vector comprising the nucleotide sequence of the human GABA receptor theta subunit together with additional sequences capable of directing the synthesis of the said human

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GABA receptor theta subunit in cultures of stably co-transfected eukaryotic cells.

- 7. A protein preparation of GABA receptor subunit combinations derived from a cell line according to claim 1 or 2.
 - 8. A membrane preparation containing subunit combinations of the GABA receptor derived from a cell line according to claim 1 or 2.
- 9. A preparation according to claim 7 or 8 wherein the subunit combination derived is the $\alpha_1\theta\gamma_2$, $\alpha_2\beta_1\theta\gamma_1$ or $\alpha_2\beta_3\theta\gamma_2$ subunit combination of the GABA receptor.
 - 10. The use of a cell according to claim 1 or 2 or a membrane preparation derived therefrom in screening for and designing medicaments which act upon a GABA receptor comprising the theta subunit.

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- 11. A method for determining whether a ligand, not known to be capable of binding to a human GABAA receptor comprising the theta subunit, can bind to a human GABAA receptor comprising the theta subunit, which comprises contacting a mammalian cell comprising DNA molecules encoding at least one alpha receptor subunit, the theta receptor subunit and optionally one or more beta, gamma, delta or epsilon receptor subunits, with the ligand under conditions permitting binding of ligands known to bind to the GABAA receptor, detecting the presence of any of the ligand bound to the GABAA receptor comprising the theta subunit and thereby determining whether the ligand binds to the GABAA receptor comprising the theta subunit.
- 30 12. A method of screening drugs to identify drugs which specifically interact with, and bind to, a human GABAA receptor

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comprising the theta subunit on the surface of a cell which comprises contacting a mammalian cell comprising a DNA molecule encoding at least one alpha receptor subunit, the theta receptor subunit and optionally one or more beta, gamma, delta or epsilon receptor subunits, on the surface of a cell with a plurality of drugs, determining those drugs which bind to the mammalian cell, and thereby identifying drugs which specifically interact with, and bind to, human GABAA receptors comprising the theta subunit.

- 13. A polynucleotide which hybridizes under stringent conditions10 to the DNA molecule depicted in Figure 1 or Figure 2.
 - 14. A GABA_A receptor theta subunit polypeptide which has the deduced amino acid sequence of Figure 1 or Figure 2, or a fragment, analog or derivative thereof.

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FIGURE 1

<u>Human θ Subunit</u>

1	AT	GCT	GCG	AGC	CGC	'AGT	'GA'T	CCT	GCT	GCT	'CAT	'CAG	GAC	CTG	GCT	CGC	GGA	GGG	CAA	CTAC	60
	M	L		Α				L					T	W	L		E	G	N	Y	
61	CC	CAG	TCC	CAT	CCC	GAA	ATT	'CCA	CTT	CGA	GTT	CTC	CTC	TGC	TGT	GCC	CGA	AGT	CGT	CCTG	120
	P	s	P	I	P		F	Н		E	F	S	S	A		P	E	V	V	L	120
121	AA	CCT	CTT	CAA	.CTG	CAA	AAA	TTG	TGC	AAA	TGA	AGC	TGT	GGT	TCA	AAA	GAT	TTT	GGA	CAGG	180
	N	L	F	N		K		С					V			K	I	L	D	R	200
181	GT	GCT	GTC	AAG	ATA	.CGA	TGT	CCG	CCT	GAG	ACC	GAA	TTT	TGG	AGG	TGC	CCC	TGT	GCC'	TGTG	240
	V	L		R	Y			R			P		F			A	P	V	P	V	
241	AG.	AAT	ATC	TAT	TTA	TGT	'CAC	GAG	CAT	TGA	ACA	GAT	CTC	AGA	AAT	GAA	TAT	GGA	CTA	CACG	300
	R	Ι	S	Ι	Y	V	Т	S	I	E	Q	I	S	E	M	N	M	D	Y	T	
301	AT	CAC	GAT	GTT	TTT	TCA	TCA	.GAC	TTG	GAA	AGA	TTC	ACG	CTT.	AGC.	ATA	CTA	TGA	GAC	CACC	360
	Ι	Т	M	F	F		Q		W		D	S	R		A	Y	Y	E	Т	T	
361	CT	GAA	CTT	GAC	CCT	GGA	.CTA	TCG	GAT	GCA	TGA	GAA	GTT	GTG	GGT	CCC'	TGA	CTG	CTA	CTTT	420
	L	N	L	Т	L	D	Y	R	M	Н	E	K	L	W	V	P	D	С	Y	F	
421	CT	GAA	CAG	CAA	.GGA	TGC	TTT	CGT	GCA	TGA	TGT	GAC	TGT	GGA	GAA	TCG	CGT	GTT'	TCA	GCTT	480
	L	N	S	K	D	Α	F	٧	H	D	V	Т	V	E	N	R	V	F	Q	L	
481	CA	CCC	AGA	TGG	AAC	GGT	GCG	GTA	CGG	CAT	CCG	ACT	CAC	CAC	TAC.	AGC.	AGT	TTG'	TTC	CCTG	540
	Η	P	D	G	Т	V	R	Y	G	Ι	R	L	Т	Т	Т	A	V	С	S	L	
541	GA'	TCT	GCA	TAA	ATT	CCC	TAT	GGA	CAA	GÇA	GGC	CTG	CAA	CCT	GGT	GGT.	AGA	GAG	CTA!	rggt	600
	D	L	H	K	F	P	M	D	K	Q	A	С	N	L	V	V	E	S	Y	G	
601	TA				AGA	CAT	CAT	ATT.	ATT	CTG	GGA	TGA	CAA	TGG	GAA	CGC	CAT	CCA	CAT	GACT	660
	Y	_	V		D	Ι	Ι	L	F	M	D	D	N	G	N	A	Ι	H	М	T	
661				GCA	TAT	CCC	TCA	GTT	CAC	TTT	CCT	GGG	AAG	GAC	GAT	TAC	TAG	CAA	GGA(GGTG	720
	Ε	Е	L	H	Ι	P	Q	F	Т	F	L	_	R	T	Ι	T	S	K	E	V	
721																				AGTT	780
			Y												Q					V	
781																				GATA	840
															Т					I	
841																				AATG	900
	S	F	W	M	N	Y	D	S	s	Α	A	R	V	T	Ι	G	L	Т	s	М	
901																					960
	L	I	L	Т	T	I	D	S	Н	L	R	D	K	L	P	N	I	s	С	I	
961																					1020
	K	Α	I	$^{\mathrm{D}}$	I	Y	I	L	V	C	L	F	F	V	F	L	S	L	L	E	

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1021	TA	TGT	CTA	CAT	CAA	.CTA	TCT	ттт	СТА	CAG	TCG	AGG	ACC	TCG	GCG	CCA	GCC	TAG	GCG	ACGC	1080
	Y	V	Y	Ι	N							G						R	R	R	
1081	AG	GAG	ACC	CCG	AAG	AGT	CAT	TGC	CCG	CTA	.CCG	CTA	CCA	GCA	AGT	GGT	'GGT	'AGG	AAA	.CGTG	1140
	R	R	P	R	R	V	Ι	A	R	Y	R	Y	Q	Q	V	V	V	G	N	V	
1141	CA	GGA	TGG	CCT	GAT	TAA	CGT	GGA	AGA	CGG	AGT	CAG	CTC	TCT	CCC	CAT	'CAC	ccc	AGC	GCAG	1200
	Q	D	G	L	I	N	V	E	D	G	V	S	S	L	P	I	Т	P	A	Q	
1201	GC	CCC	CCT	'GGC	AAG	CCC	GGA	AAG	CCT	'CGG	TTC	TTT	GAC	GTC	CAC	CTC	CGA	.GCA	.GGC	CCAG	1260
				A								L		S			E	Q	A		
1261	СТ	GGC	CAC	CTC	GGA	AAG	CCT	CAG	CCC	ACT	'CAC	TTC	TCT	CTC	AGG	CCA	.GGC	CCC	CCT	GGCC	1320
				S								S					A		L	A	
1321	AC	TGG	AGA	AAG	CCT	GAG	CGA	TCT	ccc	CTC	CAC	CTC	AGA	GCA	GGC	CCG	GCA	CAG	СТА	TGGT	1380
	T	G	E	s								s						s	Y	G	1300
1381	GT	TCG	CTT	'TAA	TGG	TTT	CCA	.GGC	TGA	TGA	.CAG	TAT	TAT	TCC	TAC	CGA	AAT	CCG	CAA	CCGT	1440
	V	R	F	N	G	F	Q	A	D	D	S	I	I	P	T	E	I	R	N	R	
1441	GT	CGA	AGC	CCA	TGG	CCA	TGG	TGT	TAC	CCA	TGA	.CCA	TGA	AGA	TTC	CAA	TGA	.GAG	CTT	GAGC	1500
	V											Н		D	s	N	E	S	L	S	
1501	TC	GGA	TGA	.GCG	CCA	TGG	CCA	TGG	CCC	CAG	TGG	GAA	GCC	CAT	GCT	TCA	.CCA	TGG	CGA	GAAG	1560
	S		E									K					Н		E	K	
1561	GG	TGT	GCA	AGA	AGC	AGG	CTG	GGA	CCT	'TGA	TGA	.CAA	CAA	TGA	CAA	GAG	CGA	.CTG	CCT	TGCC	1620
	G	V	Q	E	A	G	W	D	L	D	D	N	N	D	K	S	D	C	L	A	
1621	AT	TAA	GGA	.GCA	ATT	CAA	GTG	TGA	TAC	TAA	.CAG	TAC	CTG	GGG	CCT	TAA	TGA	TGA	TGA	GCTC	1680
				Q							S			G		N	D	D	E	L	2000
1681	GТ	GGC	CCA	TGG	CCA	AGA	GAA	GGA	.CAG	TAG	CTC	AGA	GTC	TGA	GGA	TAG	TTG	ccc	CCC	AAGC	1740
											S			E	D	s	C	P	P		2,10
1741	CC															TCC	TGA	.CTA	.CGT	CCCA	1800
	P	G	С	S	F	T	E	G	F	S	F	D	L	F	N	P	D	Y	V	P	
1801	AΑ	GGT	CGA	CAA	.GTG	GTC	CCG	GTT	CCT	'CTT	'CCC	TCT	GGC	CTT	TGG	GTT	'GTT	CAA	CAT	TGTT	1860
												L									
1861	TA	CTG	GGT	'ATA	.CCA	TAT	GTA	TTA	G.		188	4									
	Y	W	V	Y	Н	M	Y	*													

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FIGURE 2

Human 0 Subunit

-				,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,		JAG.	. GAI		. GC	i GC.	CA	CAC	CAC	CTU	الكافاة	CGC	.GG.	re:	CAP	CTAC	60
	M	L	R	A	A	V	Ι	L	L	L	I	R	T	W	L	A	Е	G	N	Y	
61	CC	CCAC	TCC	CAI	ccc	GAA	ATT	CCF	CTI	CGA	GTT	CTC	стс	TGC	<u>'</u> ጥርብ	יפרנ	rcaz	ממו	ירפיו	CCTG	120
	P	s	P		P										V		E	v	v	L	120
121	AA	CCI	CTI	CAA	CTG	CAF	AAA	TTG	TGC	'AAA'	TGA	AGC	TGT	GGT	TCA	LAA.	GAT	TTT	'GGA	CAGG	180
	N	L	F	N	С	K	N	С	Α	N	E	A	V	V	Q	K	I	L	D	R	
181	GI	GCI	GTC	'AAC	ATA	CGA	TGT	'CCG	CCI	GAG	ACC	:GAA	TTT	TGG	AGG	TGC	ccc	TGT	GCC	TGTG	240
	V	L	S	R	Y	D	V	R	L	R	P	N	F	G	G	A	P	v	P	v	
241	AG	raa:	TATC	TAT:	'TTA	TGI	'CAC	'GAG	CAT	'TGA	ACA	GAI	'CTC	AGA	TAA	'GAA	rat	'GGA	CTA	CACG	300
	R	Ι	S	Ι	Y	V	Т	S	Ι	Е	Q	I	s	E	M	N	M	D	Y	T	
301	ΑT	'CAC	GAT	GTI	TTT	'TCA	TCA	.GAC	'TTG	GAA	AGA	TTC	ACG	CTT	'AGC	'ATA	CTA	TGA	.GAC	CACC	360
	I	T	М	F	F	н	Q	Т	W	ĸ	D	s	R	L	A	Y	Y	E	т	T	
361	CT	'GAA	CTT	GAC	CCT	'GGA	СТА	TCG	GAT	'GCA	TGA	GAA	GTT.	GTG	GGT	ccc	TGA	CTG	СТА	CTTT	420
	L	N	L	Т	L	D	Y	R	М	Н	Ε	K	L	W	V	P	D	С	Y	F	
421	TT	'GAA	CAG	CAA	GGA	TGC	TTT	CGT	GCA	TGA	TGT	'GAC	TGT	GGA	.GAA	TCG	CGT	GTT	TCA	GCTT	480
	L	N	S	K	D	A	F	V	Н	D	V	T	v	E	N	R	V	F	Q	L	
481	CA	.CCC	AGA	TGG	AAC	GGT	GCG	GTA	CGG	CAT	CCG	ACT	CAC	CAC	TAC	AGC	AGC	TTG	TTC	CCTG	540
	Н	P	D	G	Т	V	R	Y	G	I	R	L	T	Т	T	A	A	С	s	L	
541	GA	TCT	GCA	TAA	ATT	ccc	TAT	GGA	CAA	GCA	GGC	CTG	CAA	CCT	GGT	GGT	AGA	GAG	CTA	TGGT	600
	D	L	H	K	F	P	М	D	K	Q	A	С	N	L	V	V	E	S	Y	G	
601	TA	CAC	GGT	TGA	AGA	CAT	CAT	ATT.	ATT	CTG	GGA	TGA	CAA	TGG	GAA	CGC	CAT	CCA	CAT	GACT	660
	Y	Т	v	E	D	I	I	L	F	W	D	D	N	G	N	A	I	Н	М	т	
661	GA	GGA	GCT	GCA	TAT	CCC	TCA	GTT	CAC	TTT	CCT	GGG	AAG	GAC	GAT	TAC	TAG	CAA	GGA	GGTG	720
	E	E	L	Н	Ι	P	Q	F	Т	F	L	G	R	Т	I	Т	s	K	E	v	
721	TA	TTT	CTA	CAC	AGG'	TTC	СТА	CAT	ACG	CCT	GAT.	ACT	GAA	GTT	CCA	GGT	TCA	GAG	gga.	AGTT	780
	Y	F	Y	Т	G	S	Y	I	R	L	I	L	K	F	Q	v	Q	R	E	v	
781																TAT	TAC	CTC'	TTG	GATA	840
	N	S	Y	L	V	Q	V	Y	W	P	T	V	L	T	T	I	Т	S	W	I	

41	TC	GTT	TTG	GAT	'GAA	CTA	TGA	TTC	CTC	TGC	AGC	CAG	GGT	GAC	AAT	TGG	CTT	AAC	TTC	AATG	900
	S	F	W	М	N	Y	D	s	S	A	Α	R	V	Т	I	G	L	T	s	М	
01	CT	CAT	CCI	'GAC	CAC	CAT	'CGA	.CTC	ACA	TCT	'GCG	GGA	TAA	.GCT	ccc	CAA	CAT	TTC	CTG	TATC	960
	L	I	L	Т	Т	I	D	s	Н	L	R	D	K	L	P	N	I	s	С	I	
51.	AA	GGC	CAT	TGA	TAT	CTA	TAT.	CCT	CGT	GTG	CTT	GTT	CTT	TGT	GTT	CCT	GTC	CTT	GCT	GGAG	1020
	K	A	I	D	I	Y	I	L	v	С	L	F	F	V	F	L	s	L	L	E	
21																			GCG	ACAC	1080
	Y	V	Y	Ι	N	Y	L	F	Y	S	R	G	P	R	R	Q	P	R	R	Н	
81	AG	GAG	ACC	:CCG	AAG	AGT	CAT	TGC	CCC	CTA	.CCG	CTA	.CCA	.GCA	AGT	GGT	GGT.	AGG	AAA	.CGTG	1140
	R	R	P	R	R	V	I	A	R	Y	R	Y	Q	Q	V	V	V	G	N	V	
41	CA	GGA	TGG	CCT	GAT	TAA	.CGT	GGA	AGA	.CGG	AGT	CAG	CTC	TCT	ccc	CAT	CAC	CCC	AGC	GCAG	1200
	Q	D	G	L	Ι	N	V	Ε	D	G	V	S	S	L	P	I	T	P	A	Q	
01															CAC	CTC	CGA	GCA	GGC	CCAG	1260
	Α	P	L	A	S	P	Ε	S	L	G	S	L	Т	S	Т	S	E	Q	A	Q	
61	CT	GGC	CAC	CTC	:GGA	AAG	CCT	CAG	ccc	ACT	CAC	TTC	TCT	CTC	AGG	CCA	GGC	ccc	CCT	GGCC	1320
	L	Α	Т	s	Е	S	L	S	P	L	T	s	Ъ	S	G	Q	A	P	L	A	
21	AC	TGG	AGA	AAG	CCT	'GAG	CGA	TCT	'CCC	CTC	CAC	CTC	'AGA	.GCA	GGC	CCG	GCA	CAG	CTA	TGGT	1380
	Т	G	E	S	L	S	D	L	P	S	Т	S	Е	Q	A	R	H	S	Y	G	
81	GT	TCG	CTI	'TAA	TGG	TTT	CCA	.GGC	TGA	TGA	CAG	TAT	'TTT	TCC	TAC	CGA	AAT	CCG	CAA	CCGT	1440
	V	R	F	N	G	F	Q	A	D	D	S	I	F	P	Т	E	Ι	R	N	R	
41	GT	CGA	AGC	CCA	TGG	CCA	TGG	TGT	TAC	CCA	TGA	.CCA	TGA	AGA	TTC	CAA	TGA	GAG	CTT	'GAGC	1500
	V	E	A	Н	G	Н	G	v	T	Н	D	Н	Е	D	s	N	Е	s	L	S	
01	TC	GGA	TGA	.GCG	CCA	TGG	CCA	TGG	ccc	'CAG	TGG	GAA	GCC	CAT	GCT	TCA	CCA	TGG	CGA	GAAG	1560
	S	D	E	R	Н	G	Н	G	P	S	G	K	P	M	L	Н	H	G	E	K	
61	GG	TGT	GCA	AGA	AGC	'AGG	CTG	GGA	CCI	TGA	TGA	.CAA	CAA	TGA	.CAA	.GAG	CGA	CTG	CCI	TGCC	1620
	G	V	Q	Ε	A	G	W	D	L	D	D	N	N	D	K	S	D	С	L	A	
21	ΑT	'TAA	.GGA	.GCA	TTA	'CAA	.GTG	TGA	TAC	TAA	CAG	TAC	CTG	GGG	CCT	'TAA	TGA	TGA	TGA	GCTC	1680
	I	K	Е	Q	F	K	С	D	Т	N	s	Т	W	G	ь	N	D	D	Ε	L	
81	ΑT	'GGC	CCA	TGG	CCA	AGA	GAA	.GGA	CAC	TAC	CTC	AGA	GTC	TGA	.GGA	TAG	TTG	CCC	ccc	AAGC	1740
	М	Α	Н	G	Q	E	K	D	S	S	S	E	S	Е	D	s	С	Р	Þ	S	

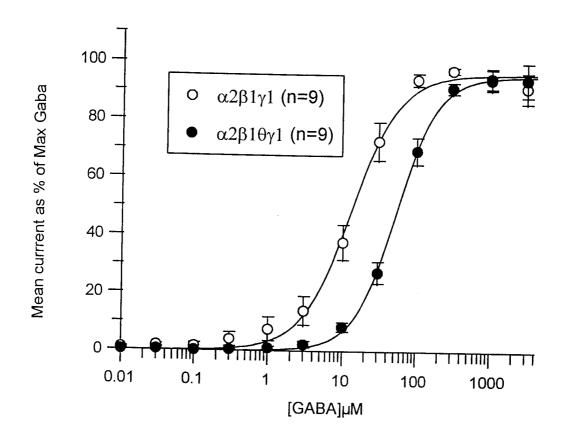
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1741	CC	TGG	GTG	CTC	CTT	CAC	TGA	AGG	GTT	CTC	CTT	CGA	TCT	CTT	TAA	TCC	TGA	CTA	CGT	CCCA	1800
	P	G	С	s	F	Т	Е	G	F	S	F	D	L	F	N	P	D	Y	v	P	

- 1801 AAGGTCGACAAGTGGTCCCGGTTCCTCTCCCTCTGGCCTTTGGGTTGTTCAACATTGTT 1860

 K V D K W S R F L F P L A F G L F N I V
 - 1861 TACTGGGTATACCATATGTATTAG 1884
 Y W V Y H M Y *

FIGURE 3



INTERNATIONAL SEARCH REPORT

lı ational Application No PCT/GB 98/01206

A CLASS	FICATION OF SUBJECT MATTER		·
IPC 6	C12N15/12 C07K14/705 C12N5/1 C12Q1/68 G01N33/68	0 A61K38/17 G01N	33/50
According to	o International Patent Classification(IPC) or to both national classific	cation and IPC	
	SEARCHED	-	
Minimum do	commentation searched (classification system followed by classificat ${\tt C07K}$	ion symbols)	44-44
Documenta	tion searched other than minimumdocumentation to the extent that	such documents are included in the fields se	arched
Electronic d	ata base consulted during the international search (name of data ba	ase and, where practical, search terms used	
C. DOCUM	ENTS CONSIDERED TO BE RELEVANT		· · · · · · · · · · · · · · · · · · ·
Category °	Citation of document, with indication, where appropriate, of the re	levant passages	Relevant to claim No.
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"A" docume consid "E" earlier of filing docume which citation "O" docume other n "P" docume	ont defining the general state of the art which is not ered to be of particular relevance locument but published on or after the international ate nt which may throw doubts on priority claim(s) or is cited to establish the publication date of another or or other special reason (as specified) ent referring to an oral disclosure, use, exhibition or	"T" later document published after the inte or priority date and not in conflict with cited to understand the principle or the invention "X" document of particular relevance; the cannot be considered novel or cannot involve an inventive step when the document of particular relevance; the cannot be considered to involve an in document is combined with one or moments, such combination being obvious in the art. "&" document member of the same patent	the application but early underlying the laimed invention be considered to cument is taken alone laimed invention ventive step when the tre other such docu- us to a person skilled
Date of the a	actual completion of theinternational search	Date of mailing of the international sea	***************************************
2!	5 August 1998	04/09/1998	
Name and m	nailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo ni, Fax: (+31-70) 340-3016	Authorized officer Le Cornec N	And the second s

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