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(54) Title: ANTIBODIES THAT BLOCK RECEPTOR PROTEIN TYROSINE KINASE ACTIVATION, METHODS OF SCREENING FOR AND USES THEREOF

(57) Abstract: Molecules containing the antigen-binding portion of antibodies that block constitutive and/or ligand-dependent activation of a receptor protein tyrosine kinase, such as fibroblast growth factor receptor 3 (FGFR3), are found through screening methods, where a soluble dimeric form of a receptor protein tyrosine kinase is used as target for screening a library of antibody fragments displayed on the surface of bacteriophage. The molecules of the present invention which block constitutive activation can be administered to treat or inhibit skeletal dysplasia, craniosynostosis disorders, cell proliferative diseases or disorders, or tumor progression associated with the constitutive activation of a receptor protein tyrosine kinase.



WO 02/102973 A2

ANTIBODIES THAT BLOCK RECEPTOR PROTEIN TYROSINE KINASE ACTIVATION, METHODS OF SCREENING FOR AND USES THEREOF

Field of the Invention

- 5 The present invention relates to: immunoglobulins (and functional fragments thereof) useful for blocking activation of receptor protein tyrosine kinases, methods for screening for such immunoglobulins, compositions comprising said immunoglobulins and methods of using the same for treating or inhibiting disease, such as skeletal dysplasia, craniosynostosis disorders, cell proliferative diseases or disorders, or tumor progression.

10 Background of the Invention

- A wide variety of biological processes involves complex cellular communication mechanisms. One of the primary means of continual exchange of information between cells and their internal and external environments is via the secretion and specific binding of peptide growth factors. Growth factors exert pleiotropic effects and play important roles in
- 15 oncogenesis and the development of multicellular organisms regulating cell growth, differentiation and migration. Many of these factors mediate their effects by binding to specific cell surface receptors. The ligand-activated receptors start an enzymatic signal transduction cascade from the cell membrane to the cell nucleus, resulting in specific gene regulation and diverse cellular responses.

20 Protein Kinases

- One of the key biochemical mechanisms of signal transduction involves the reversible phosphorylation of proteins, which enables regulation of the activity of mature proteins by altering their structure and function.
- Protein kinases ("PKs") are enzymes that catalyze the phosphorylation of hydroxy groups on
- 25 tyrosine, serine and threonine residues of proteins. The consequences of this seemingly simple activity are staggering; cell growth, differentiation and proliferation; e.g., virtually all aspects of cell life in one way or another depend on PK activity. Furthermore, abnormal PK activity has been related to a host of disorders, ranging from relatively non-life threatening diseases such as psoriasis to extremely virulent diseases such as glioblastoma.

The kinases fall largely into two groups, those specific for phosphorylating serine and threonine, and those specific for phosphorylating tyrosine. Some kinases, referred to as “dual specificity” kinases, are able to phosphorylate tyrosine as well as serine/threonine residues.

Protein kinases can also be characterized by their location within the cell. Some kinases are transmembrane receptor proteins capable of binding ligands external to the cell membrane. Binding the ligands alters the receptor protein kinase’s catalytic activity. Others are non-receptor proteins lacking a transmembrane domain and yet others are ecto-kinases that have a catalytic domain on the extracellular (ecto) portion of a transmembrane protein or which are secreted as soluble extracellular proteins.

Many kinases are involved in regulatory cascades where their substrates may include other kinases whose activities are regulated by their phosphorylation state. Thus, activity of a downstream effector is modulated by phosphorylation resulting from activation of the pathway.

Receptor protein tyrosine kinases (RPTKs) are a subclass of transmembrane-spanning receptors endowed with intrinsic, ligand-stimulatable tyrosine kinase activity. RPTK activity is tightly controlled. When mutated or altered structurally, RPTKs can become potent oncoproteins, causing cellular transformation. In principle, for all RPTKs involved in cancer, oncogenic deregulation results from relief or perturbation of one or several of the auto-control mechanisms that ensure the normal repression of catalytic domains. More than half of the known RPTKs have been repeatedly found in either mutated or overexpressed forms associated with human malignancies (including sporadic cases; Blume-Jensen et al., 2001). RPTK over expression leads to constitutive kinase activation by increasing the concentration of dimers. Examples are Neu/ErbB2 and epidermal growth factor receptor (EGFR), which are often amplified in breast and lung carcinomas and the fibroblast growth factors (FGFR) associated with skeletal and proliferative disorders (Blume-Jensen et al., 2001).

Fibroblast Growth Factors

Normal growth, as well as tissue repair and remodeling, require specific and delicate control of activating growth factors and their receptors. Fibroblast Growth Factors (FGFs) constitute a family of over twenty structurally related polypeptides that are developmentally regulated and expressed in a wide variety of tissues. FGFs stimulate proliferation, cell migration and differentiation and play a major role in skeletal and limb development, wound healing, tissue repair, hematopoiesis, angiogenesis, and tumorigenesis (reviewed in Ornitz and Itoh, 2001).

The biological action of FGFs is mediated by specific cell surface receptors belonging to the RPTK family of protein kinases. These proteins consist of an extracellular ligand binding domain, a single transmembrane domain and an intracellular tyrosine kinase domain which undergoes phosphorylation upon binding of FGF. The FGF receptor (FGFR) extracellular
5 region contains three immunoglobulin-like (Ig-like) loops or domains (D1, D2 and D3), an acidic box, and a heparin binding domain. Five FGFR genes that encode for multiple receptor protein variants have been identified to date.

Another major class of cell surface binding sites includes binding sites for heparan sulfate proteoglycans (HSPG) that are required for high affinity interaction and activation of all
10 members of the FGF family. Tissue-specific expression of heparan sulfate structural variants confer ligand-receptor specificity and activity of FGFs.

FGFR-Related Disease

Recent discoveries show that a growing number of skeletal abnormalities, including achondroplasia, the most common form of human dwarfism, result from mutations in FGFRs.
15 Specific point mutations in different domains of FGFR3 are associated with autosomal dominant human skeletal disorders including hypochondroplasia, severe achondroplasia with developmental delay and acanthosis nigricans (SADDAN) and thanatophoric dysplasia (TD) (Cappellen et al., 1999; Webster et al., 1997; Tavormina et al., 1999). FGFR3 mutations have also been described in two craniosynostosis phenotypes: Muenke coronal craniosynostosis
20 (Bellus et al., 1996; Muenke et al., 1997) and Crouzon syndrome with acanthosis nigricans (Meyers et al., 1995). Crouzon syndrome is associated with specific point mutations in FGFR2 and both familial and sporadic forms of Pfeiffer syndrome are associated with mutations in FGFR1 and FGFR2 (Galvin et al., 1996; Schell et al., 1995). Mutations in FGFRs result in constitutive activation of the mutated receptors and increased receptor
25 protein tyrosine kinase activity, rendering cells and tissue unable to differentiate. Specifically, the achondroplasia mutation results in enhanced stability of the mutated receptor, dissociating receptor activation from down-regulation, leading to restrained chondrocyte maturation and bone growth inhibition (reviewed in Vajo et al., 2000).

There is accumulating evidence for mutations activating FGFR3 in various types of cancer.
30 Constitutively activated FGFR3 in a large proportion of two common epithelial cancers, bladder and cervix, as well as in multiple myeloma, is the first evidence of an oncogenic role for FGFR3 in carcinomas. FGFR3 currently appears to be the most frequently mutated

oncogene in bladder cancer where it is mutated in almost 50% of the cases and in about 70% of cases having recurrent superficial bladder tumors (Cappellen, et al, 1999; van Rhijn, et al, 2001; Billerey, et al, 2001). FGFR3 mutations are seen in 15-20% of multiple myeloma cases where point mutations that cause constitutive activation directly contribute to tumor
5 development and progression (Chesi, et al, 1997; Plowright, et al, 2000, Ronchetti, et al, 2001).

In this context, the consequences of FGFR3 signaling appear to be cell type-specific. In chondrocytes, FGFR3 hyperactivation results in growth inhibition (reviewed in Ornitz, 2001), whereas in the myeloma cell it contributes to tumor progression (Chesi et al., 2001).

10 In view of the link between RPTK-related cellular activities and a number of human disorders various strategies have been employed to target the receptors and/or their variants for therapy. Some of these have involved biomimetic approaches using large molecules patterned on those involved in the cellular processes, e.g., mutant ligands (US Patent 4,966,849); soluble receptors and antibodies (WO 94/10202, US 6,342,219); RNA ligands
15 (US Patent 5,459,015) and tyrosine kinase inhibitors (WO 94/14808; US Patent 5,330,992).

Antibody therapy

The search for new therapies to treat cancer and other diseases associated with growth factors and their corresponding cell surface receptors has resulted in the development of humanized antibodies capable of inhibiting receptor function. For example, US patents 5,942,602 and
20 6,365,157 disclose monoclonal antibodies specific for the HER2/neu and VEGF receptors, respectively. US patent 5,840,301 discloses a chimeric, humanized monoclonal antibody that binds to the extracellular domain of VEGF and neutralizes ligand-dependent activation.

There is an unmet need for highly selective molecules capable of blocking aberrant constitutive receptor protein tyrosine kinase activity, in particular FGFR activity, thereby
25 addressing the clinical manifestations associated with the above-mentioned mutations, and modulating various biological functions.

Citation of any document herein is not intended as an admission that such document is pertinent prior art, or considered material to the patentability of any claim of the present application. Any statement as to content or a date of any document is based on the
30 information available to applicant at the time of filing and does not constitute an admission as to the correctness of such a statement.

SUMMARY OF THE INVENTION

It is an object of the present invention to provide molecules which are able to block receptor protein tyrosine kinase (RPTK) activity.

5 It is an object of the present invention to provide molecules which are able to block fibroblast growth factor receptor (FGFR) activity, and more preferably fibroblast growth factor receptor 3 (FGFR3) activity.

It is another object of the present invention to provide a method to screen for molecules which are able to block said receptor activity.

10 It is yet another object of the present invention to provide a pharmaceutical composition comprising as an active ingredient a molecule of the invention useful in treating or preventing skeletal and proliferative diseases and disorders.

It is a further object of the present invention to provide a method for inhibiting growth of tumor cells associated with ligand-dependent or constitutive activation of a receptor protein tyrosine kinase, preferably a fibroblast growth factor receptor, and more preferably FGFR3.

15 It is yet a further object of the present invention to provide a method for treating skeletal disorders associated with ligand-dependent or constitutive activation of a receptor protein tyrosine kinase, preferably a fibroblast growth factor receptor, and more preferably FGFR3.

20 It is yet a further object of the present invention to provide a method for blocking receptor protein tyrosine kinase activation in the cells of patients in need thereof by treatment with molecules capable of inhibiting receptor protein tyrosine kinase function.

It is yet another object of the present invention to provide a method for inhibiting tumor growth, tumor progression or metastases.

It is still a further object to provide molecules useful for in vivo imaging of diseased states.

25 It is still a further object of the invention to provide a kit containing molecules of the invention.

These and other objects are met by the invention disclosed herein.

The present invention provides a molecule that contains the antigen-binding portion of an antibody which has a specific affinity for a receptor protein tyrosine kinase and which blocks constitutive activation of a receptor protein tyrosine kinase. The present invention further

provides a molecule that contains the antigen-binding portion of an antibody which has a specific affinity for a receptor protein tyrosine kinase and which blocks ligand-dependent activation of a fibroblast growth factor receptor (FGFR), including FGFR1 and FGFR3.

5 Certain molecules of the present invention were found to inhibit or block constitutive, or ligand independent, activation of the FGFR3. Generation of inhibitory molecules would be useful for developing medicaments for use in treating or preventing skeletal and proliferative diseases and disorders associated with constitutive activation of receptor protein tyrosine kinases.

10 Certain mutations in the genes of receptor protein tyrosine kinases result in activation of the receptor in a manner that is independent of the presence of a ligand. Such ligand-independent, or constitutive, receptor protein tyrosine kinase activation results in increased receptor activity. The clinical manifestations of certain mutations are skeletal and proliferative disorders and diseases, including achondroplasia and various cancers.

15 Furthermore, the present invention is directed to novel molecules comprising an antigen binding domain which binds to a receptor protein tyrosine kinase and blocks constitutive activation of said receptor protein tyrosine kinase. The molecules of the invention maybe antibodies or antigen binding fragments thereof.

A currently preferred embodiment of the present invention provides a molecule which binds to the extracellular domain of a receptor protein tyrosine kinase and blocks constitutive and 20 ligand-dependent activation of the receptor.

A currently more preferred embodiment of the present invention provides a molecule which binds to the extracellular domain of an FGF receptor and blocks constitutive and ligand-dependent activation of the receptor.

25 A currently most preferred embodiment of the present invention provides a molecule which binds FGFR3 and blocks constitutive and ligand-dependent activation of the receptor, comprising V_L -CDR3 and V_H -CDR3 regions having SEQ ID NO:25 and 24, respectively and the corresponding polynucleotide sequence SEQ ID NO:51 and 50.

30 A currently most preferred embodiment of the present invention provides a molecule which binds FGFR3 and blocks constitutive and ligand-dependent activation of the receptor, comprising V_L -CDR3 and V_H -CDR3 regions having SEQ ID NO:13 and 12 or SEQ ID NO:9

and 8, respectively and the corresponding polynucleotide sequence SEQ ID NO:35 and 34 or SEQ ID NO: 31 and 30.

Another currently preferred embodiment of the present invention provides a molecule herein denoted MSPRO12 comprising a light chain having SEQ ID NO:94 and a heavy chain having
5 SEQ ID NO:105 and the corresponding polynucleotide sequences having SEQ ID NO:75 and 89, respectively.

Another currently preferred embodiment of the present invention provides a molecule herein denoted MSPRO2 comprising a light chain having SEQ ID NO:92 and a heavy chain having
10 SEQ ID NO:103 and the corresponding polynucleotide sequences having SEQ ID NO:74 and 86,

Another currently most preferred embodiment of the present invention provides a molecule herein denoted MSPRO59 comprising a light chain having SEQ ID NO:102 and a heavy chain having SEQ ID NO:113 and the corresponding polynucleotide sequences having SEQ ID NO:76 and 91, respectively.

15 According to the principles of the present invention, molecules which bind FGFR and block ligand-dependent receptor activation are provided. These molecules are useful in treating disorders and diseases associated with an FGFR that is activated in a ligand-dependent manner including certain skeletal disorders, hyperproliferative diseases or disorders and non-neoplastic angiogenic pathologic conditions such as neovascular glaucoma, macular
20 degeneration, hemangiomas, angiofibromas, psoriasis and proliferative retinopathy including proliferative diabetic retinopathy.

A currently most preferred embodiment of the present invention provides a molecule which binds FGFR3 and blocks ligand-dependent activation of the receptor, comprising V_H-CDR3 and V_L-CDR3 regions having SEQ ID NO:20 and 21, respectively and the corresponding
25 polynucleotide sequence SEQ ID NO:44 and 45, respectively.

Other currently preferred embodiments of the present invention provides a molecule which binds FGFR3 and blocks ligand-dependent activation of the receptor, comprising V_H-CDR3 and V_L-CDR3 regions selected from the group consisting of SEQ ID NO:10 and SEQ ID NO:11; SEQ ID NO:14 and SEQ ID NO:15; SEQ ID NO:16 and SEQ ID NO:17; SEQ ID
30 NO:18 and SEQ ID NO:19; SEQ ID NO:20 and SEQ ID NO:21; SEQ ID NO:26 and SEQ ID NO:27 or SEQ ID NO:28 and SEQ ID NO:29 and the corresponding polynucleotide sequences according to table 1B.

Additional currently preferred embodiments of the present invention provide molecules having an antigen binding domain comprising a VL region and a VH region, respectively, selected from the group consisting of respectively, selected from the group consisting of SEQ ID NO: 92 and 103; SEQ ID NO: 93 and 104; SEQ ID NO: 94 and 105; SEQ ID NO:95 and 5 106; SEQ ID NO: 96 and 107 ; SEQ ID NO: 97 and 108; SEQ ID NO:98 and 109; SEQ ID NO: 99 and 110; SEQ ID NO: 101 and 112; and SEQ ID NO:102 and 113.

A currently preferred embodiment of the present invention provides a molecule comprising V_H-CDR3 and V_L-CDR3 domains having SEQ ID NO:22 and SEQ ID NO:23, which has specific affinity for FGFR1 and which blocks ligand-dependent activation of FGFR1, and the 10 corresponding polynucleotides having SEQ ID NO:46 and SEQ ID NO:47.

A currently preferred embodiment of the present invention provides a molecule comprising domains having SEQ ID NO: 100 and 111, which has specific affinity for FGFR1 and which blocks ligand-dependent activation of FGFR1, and the corresponding polynucleotides having SEQ ID NO:73 and SEQ ID NO:82.

15 In addition, the present invention also relates to methods for screening for the molecules according to the present invention by using a dimeric form of a receptor protein tyrosine kinase as a target for screening a library of antibody fragments.

According to one currently preferred embodiment the screening method comprises

20 screening a library of antibody fragments for binding to a dimeric form of a receptor protein tyrosine kinase;
identifying an antibody fragment which binds to the dimeric form of the receptor protein tyrosine kinase as a candidate molecule for blocking constitutive activation of the receptor protein tyrosine kinase; and
determining whether the candidate molecule blocks constitutive or ligand-dependent 25 activation of the receptor protein tyrosine kinase in a cell.

According to another currently preferred embodiment the dimeric form of the RPTK is a soluble extracellular domain of a receptor protein tyrosine kinase. Non-limiting examples of receptor protein tyrosine kinases disclosed in Blume-Jensen et al. (2001) include EGFR/ErbB1, ErbB2/HER2/Neu, ErbB/HER3, ErbB4/HER4, IGF-1R, PDGFR- α , PDGFR- β , 30 CSF-1R, kit/SCFR, Flk2/FH3, Flk1/VEGFR1, Flk1/VEGFR2, Flt4/VEGFR3, FGFR1,

FGFR2/K-SAM, FGFR3, FGFR4, TrkA, TrkC, HGFR, RON, EphA2, EphB2, EphB4, Axl, TIE/TIE1, Tek/TIE2, Ret, ROS, Alk, Ryk, DDR, LTK and MUSK.

By using a dimeric form of the RPTK as bait in the screen, a molecule which would bind to the dimeric form of the receptor has been identified. This presents a novel concept in
5 screening for antibodies or fragments thereof with the capacity to bind to a constitutively activated RPTK such as those associated with various disorders and diseases. It also presents an opportunity to screen for molecules which bind to a heterodimer RPTK. A further aspect of the present invention provides a pharmaceutical composition comprising as an active ingredient a molecule of the present invention useful for preventing or treating skeletal or
10 cartilage diseases or disorders and craniosynostosis disorders associated with constitutive or ligand-dependent activation of a receptor protein tyrosine kinase.

In a currently preferred embodiment the pharmaceutical compositions of the present invention may be used for treating or preventing skeletal disorders associated with aberrant FGFR signaling, including achondroplasia, thanatophoric dysplasia, Apert or Pfeiffer
15 syndrome and related craniosynostosis disorders.

A further aspect of the present invention provides a pharmaceutical composition comprising as an active ingredient a molecule of the present invention useful for preventing or treating cell proliferative diseases or disorders or tumor progression, associated with the constitutive or ligand-dependent activation of a receptor protein tyrosine kinase.

20 In a currently preferred embodiment the pharmaceutical compositions of the present invention may be used for treating or preventing proliferative diseases associated with aberrant FGFR signaling, including multiple myeloma, transitional cell carcinoma of the bladder, mammary and cervical carcinoma, chronic myeloid leukemia and osteo- or chondrosarcoma.

25 A further aspect of the invention provides molecules comprising an antigen binding domain which can be conjugated to cytotoxins useful for targeting cells expressing said antigen.

Another currently preferred aspect of the present invention provides molecules comprising an antigen binding domain which can be conjugated to appropriate detectable imaging moiety, useful for in vivo tumor imaging.

30 A still further aspect of the present invention provides methods for treating or inhibiting the aforementioned diseases and disorders by administering a therapeutically effective amount of

a pharmaceutical composition comprising a molecule of the present invention to a subject in need thereof.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows hFR3²³⁻³⁷⁴TDhis purification by Coomassie stained SDS-PAGE.

Figure 2 shows hFR3²³⁻³⁷⁴TDhis binding to heparin and FGF9.

Figure 3 shows the purification of FR3exFc and FR1exFc on SDS-PAGE.

- 5 Figure 4 shows the neutralization effect of the hFR3²³⁻³⁷⁴TDhis and FR3exFc soluble receptors in a ligand-dependent proliferation assay.

Figure 5 shows the effect of MSPRO Fabs on proliferation of FGFR1 and FGFR3-expressing cells.

Figure 6 shows the effect of MSPRO Fabs on proliferation of FGFR3-expressing cells.

- 10 Figures 7A and 7B show the neutralizing activity of several MSPRO Fabs in a proliferation assay using the FDCP-FR3 (C10; Fig. 7A) or the FDCP-FR1 cells (Fig. 7B).

Figure 8 shows the receptor specificity of MSPRO Fabs on RCJ-FR3 cells by Western blot using an anti-P-ERK antibody. Figure 8A shows different MSPRO Fabs while Figure 8B shows a dose response of MSPRO 12, 29 and 13 on RCJ-FR3 cells.

- 15 Figures 9A-9D demonstrates the specificity and potency of MS-PRO Fabs by Western blot with anti-P-ERK antibody.

Figure 10 shows a diagrammatic representation of FGFR3 and of FGFR3 truncations (D2-3, D2) and isoforms (IIIb, IIIc). The isoform IIIb differs from IIIc at the carboxy terminus of the IgIII domain as indicated with a dotted line.

- 20 Figure 11 shows that the FGFR3 neutralizing Fabs recognize IgII or IgII and III in the extracellular region of FGFR3.

Figure 12 shows that MSPRO29 specifically recognizes the IIIc isoform of FGFR3.

Figure 13 shows the results of a proliferation assay for FDCP-FR3IIIb or FDCP-FR3IIIc cells incubated with increasing dose of the indicated Fabs.

- 25 Figure 14 shows iodinated MSPRO29 binding to FGFR3.

Figure 15 shows results of a proliferation assay is a graph wherein iodinated MSPRO29 retained its activity against FGFR3.

Figures 16A-16F show the selective binding of radiolabelled MS-PRO29 to histological of growth plate.

Figure 17 shows a proliferation assay of FDCP-FR3 (C10) and FDCP-FR3ach cells incubated with FGF9 and with increasing doses of the indicated Fabs.

- 5 Figure 18B shows that MSPRO12 and MSPRO59 inhibit the ligand independent proliferation of FDCP-FR3ach cells. Fig. 18A shows analysis of the ligand-dependent FDCP-FR3wt cells.

Figure 19 shows the restoration of cell growth in RCS cells by MS-PRO54 and MSPRO59..

Figure 20 represents the growth rate of treated bone with MS-PRO 59.

- Figure 21 is a flow chart of the experimental protocol for assessing receptor activation and signaling.

Figure 22 shows ¹²⁵I labeled MSPRO59 localization to the FDCP-FR3ach derived tumor.

Figure 23 shows the effect of MSPRO59 on inhibiting ligand-independent tumor growth after 24 and 26 days.

Figure 24 shows the effect of MSPRO59 on inhibiting ligand-independent tumor growth.

- 15 Figure 25A shows the effect of MSPRO59 on inhibiting ligand-independent tumor growth.

Figure 25B shows scFv MSPRO59 blocking the proliferation of FDCP-FR3 (S375C) cells.

Figure 26 shows the effect of MSPRO59 single chain antibody on inhibiting ligand-independent tumor growth.

Figure 27 shows binding of Fab Miniantibodies to FGFR3-Fc and FGFR1-Fc (ELISA).

- 20 Figure 28A is an example of a Fab expression vector for use in accordance with the present invention.

Figure 28B is the DNA sequence of the vector according to Figure 28A

Figure 29A is an example of a phage display vector for use in accordance with the present invention.

- 25 Figure 29B is the DNA sequence of the vector according to Figure 29A.

Figure 30 depicts the polynucleotide sequences of the VL and VH of MSPRO antibodies of the present invention.

DETAILED DESCRIPTION OF THE INVENTION

The present invention is based on the discovery that neutralizing antibodies that block ligand-dependent and ligand-independent activation of fibroblast growth factor 3 (FGFR3), a
5 receptor protein tyrosine kinase (RPTK), can be obtained by screening an antibody library against a dimeric form of the extracellular portion of FGFR3. Until the present invention, the present inventors are unaware of any success in obtaining neutralizing antibodies that block constitutive activation of any RPTK including FGFR or ligand-dependent FGFR activation.

The term "receptor protein tyrosine kinase" or "RPTK" as used herein and in the claims
10 refers to a subclass of transmembrane-spanning receptors endowed with intrinsic, ligand-stimulatable tyrosine kinase activity. RPTKs comprise a large family of spatially and temporally regulated proteins that control many different aspects of growth and development. When mutated or altered structurally, RPTKs can undergo deregulation and become activated in a ligand-independent, or constitutive, manner. In certain cases they become potent
15 oncoproteins, causing cellular transformation.

As used herein and in the claims the term "fibroblast growth factor receptor" or "FGFR" denotes a receptor specific for FGF which is necessary for transducing the signal exerted by FGF to the cell interior, typically comprising an extracellular ligand-binding domain, a single transmembrane helix, and a cytoplasmic domain having tyrosine kinase activity. The FGFR
20 extracellular domain consists of three immunoglobulin-like (Ig-like) domains (D1, D2 and D3), a heparin binding domain and an acidic box. Alternative splicing of the FGF receptor mRNAs generates different variants of the receptors.

Molecules, including antibodies and fragments thereof, comprising an antigen binding domain to a receptor protein tyrosine kinase are highly necessary for the treatment of various
25 pathological conditions.

In the past, the laboratory of the present inventors encountered difficulties in raising neutralizing antibodies against FGFR3. When mice were immunized with the soluble monomeric FGFR3 receptor, by the time the antibody titers begins to increase, the mice died. The experiments performed in the laboratory of the present inventors that failed to obtain
30 anti-FGFR3 neutralizing antibodies in mice are presented in the Examples.

By using a soluble dimeric form of the extracellular domain of the FGFR3 receptor to screen for antibodies, e.g., Fabs, that bind from a phage display antibody library, the present inventors were able to overcome a problem in the prior art for which there was yet no solution and to obtain numerous high affinity ($K_D < 10$ nM) antibodies (Fabs) that bind
5 FGFR3 and interfere with ligand binding, thereby blocking ligand-dependent activation of FGFR3 signaling. Very surprisingly, from among the group of Fabs that block ligand-dependent activation, Fab antibodies which also block ligand-independent (constitutive) activation of FGFR3 by blocking signaling caused by constitutive dimerization of FGFR3 were identified. To the best of the present inventors' knowledge, the Fab antibodies obtained
10 which block constitutive activation of FGFR3 are the first antibodies against any receptor protein tyrosine kinase that blocks constitutive, ligand-independent activation/signaling.

Trastuzumab, an anti-human epidermal growth factor receptor 2 (HER2) antibody, was the first humanized monoclonal antibody approved for therapeutic use. Its mode of action appears to be manifold, including HER2 down regulation, prevention of heterodimer
15 formation, prevention of HER2 cleavage and others (Baselga and Albanell, 2001). US patents 5, 677171; 5772997; 6165464 and 6,399,063 disclose the anti-HER2 invention but neither teach nor suggest that the antibody blocks ligand-independent receptor activation.

One aspect of the present invention is directed to neutralizing antibodies and more generally to a molecule that includes the antigen binding portion of an antibody which blocks ligand-
20 dependent activation and constitutive, ligand-independent activation of a receptor protein tyrosine kinase, preferably an FGFR and more preferably FGFR3.

Another aspect of the present invention is directed to molecules comprising an antigen binding domain which blocks ligand-dependent activation of an FGFR, more preferably FGFR3.

25 The molecule having the antigen-binding portion of an antibody according to the present invention can be used in a method for blocking the ligand-dependent activation and/or ligand independent (constitutive) activation of FGFR3. Preferred embodiments of such antibodies/molecules, obtained from an antibody library designated as HuCAL[®] (Human Combinatorial Antibody Library) clone, is presented in Table 1 with the unique VH-CDR3
30 and VL-CDR3 sequences given.

In addition to sequencing of the clones, a series of biochemical assays were performed to determine affinity and specificity of the molecules to the respective receptors.

TABLE 1A

HuCAL® Clone	VH-CDR3 Sequence	VL-CDR3 sequence	Framework
MSPRO2	DFLGYEFDY (SEQ ID NO: 8)	QSYDYSADY (SEQ ID NO: 9)	VH1B_L3
MSPRO11	YYGSSLYHYVFGGFIDY (SEQ ID NO: 10)	QSHHFYE (SEQ ID NO: 11)	VH1B_L2
MSPRO12	YHSWYEMGYYGSTVGYMFD (SEQ ID NO: 12)	QSYDFDFA (SEQ ID NO: 13)	VH2_L3
MSPRO21	DNWFKPFSDV (SEQ ID NO: 14)	QQYDSIPY (SEQ ID NO: 15)	VH1A_k4
MSPRO24	VNHWTYTFDY (SEQ ID NO: 16)	QQMSNYPD (SEQ ID NO: 17)	VH1A_k3
MSPRO26	GYWYAYFTYINYGYFDN (SEQ ID NO: 18)	QSYDNNSDV (SEQ ID NO: 19)	VH1B_L2
MSPRO28	GGGWVSHGYYYLFDL (SEQ ID NO: 26)	FQYGSIPP (SEQ ID NO: 27)	VH1A_k1
MSPRO29	TWQYSYFYLDGGYYFDI (SEQ ID NO: 20)	QQTNNAPV (SEQ ID NO: 21)	VH1B_k3
MSPRO54	NMAYTNYQYVNMPHFDY (SEQ ID NO: 22)	QSYDYFKL (SEQ ID NO: 23)	VH1B_L3
MSPRO55	SMNSTMYWYLRRVLFDH (SEQ ID NO: 28)	QSYDMYMYI (SEQ ID NO: 29)	VH1B_L2
MSPRO59	SYYPDFDY (SEQ ID NO: 24)	QSYDGPDW (SEQ ID NO: 25)	VH6_L3

VH refers to the variable heavy chain, VL refers to the variable light chain; L refers to the lambda light chain and k refers to the kappa light chain

Table 1B lists the corresponding polynucleotide sequences of the CDR domains.

TABLE 1B

HuCAL® Clone	VH-CDR3 polynucleotide sequence	VL-CDR3 polynucleotide Sequence
MSPRO2	GATTTTCTTGGTTATGAGTTTGATTAT (SEQ ID NO:30)	CAGAGCTATGAC TATTCTGCT GAT TAT (SEQ ID NO:31)
MSPRO11	TATTATGGTTCTTCTCTTTATCATTATGTTT TTGGTGGTTTTATTGATTAT (SEQ ID NO:32)	CAGTCTCATCAT TTTTATGAG (SEQ ID NO:33)
MSPRO12	TATCATTCTTGGTATGAGATGGGTTATTAT GGTTCTACTGTTGGTTATATGTTTGATTAT (SEQ ID NO:34)	CAGAGCTATGAC TTTGATTTT GCT (SEQ ID NO:35)
MSPRO21	GATAATTGGTTTAAGCCTTTTTCTGATGTT (SEQ ID NO:36)	CAGCAGTATGAT TCTATTCCT TAT (SEQ ID NO:37)
MSPRO24	GTTAATCATTGGACTTATACTTTTGATTAT (SEQ ID NO:38)	CAGCAGATGTCT AATTATCCTGAT (SEQ ID NO:39)
MSPRO26	GGTTATTGGTATGCTTATTTTACTTAT ATTAATTATGGTTATTTT GATAAT (SEQ ID NO:40)	CAGAGCTATGAC AATAATTCTGAT GTT (SEQ ID NO:41)
MSPRO28	GGTGGTGGTTGGGTTTCTCATGGTTATTAT TATCTTTTGTATCTT (SEQ ID NO:42)	TTTCAGTATGGT TCTATTCCT CCT (SEQ ID NO: 43)
MSPRO29	ACTTGGCAGTATTCTTATTTTATTAT CTTGATGGTGGTTATTATTTTGATATT (SEQ ID NO:44)	CAGCAGACTAAT AATGCTCCTGTT (SEQ ID NO:45)
MSPRO54	AATATGGCTTATACTAATTATCAGTATGTT AATATGCCTCATTTTGATTAT (SEQ ID NO:46)	CAGAGCTATGAC TATTTTAAGCTT (SEQ ID NO:47)
MSPRO55	TCTATGAATTCTACTATGTATTGGTATCTT CGTCGTGTTCTTTTGTAT CAT (SEQ ID NO:48)	CAGAGCTATGAC ATGTATAATTAT ATT (SEQ ID NO:49)
MSPRO59	TCTTATTAT CCTGATTTT GATTAT (SEQ ID NO:50)	CAGAGCTATGAC GGTCCTGATCTT TGG (SEQ ID NO:51)

Figure 30 provides the polynucleotide sequences of the preferred embodiments of the invention. The amino acid sequence of the VH and VL domains of the currently preferred embodiments of the present invention are presented below.

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5  MS-Pro-2-VL (SEQ ID NO:92)
      1   DIELTQPPSV SVAPGQTARI SCSGDALGDK YASWYQQKPG QAPVLVIYDD
      51   SDRPSGIPER FSGSNSGNTA TLTISGTQAE DEADYYCQSY DYSADYVFGG
10      101  GTKLTVLGQ

MS-Pro-11-VL (SEQ ID NO:93)
      1   DIALTQPPASV SGSPGQSITI SCTGTSSDVG GYNYVS WYQQ HPGKAPKLMI
15      51   YDVSNRPSGV SNRFSGSKSG NTASLTISGL QAEDADYYC QSHHFYEVFG
      101  GGTKLTVLGQ

20  MS-Pro-12-VL (SEQ ID NO:94)
      1   DIELTQPPSV SVAPGQTARI SCSGDALGDK YASWYQQKPG QAPVLVIYDD
      51   SDRPSGIPER FSGSNSGNTA TLTISGTQAE DEADYYCQSY DFDFAVFGG
25      101  TKLTVLGQ

MS-Pro-21-VL (SEQ ID NO:95)
      1   DIVMTQSPDS LAVSLGERAT INCRSQQSVL YSSNNKNYLA WYQQKPGQPP
30      51   KLLIYWASTR ESGVPDRFSG SSGSTDFTLT ISSLQAEDVA VYYCQQYDSI
      101  PYTFGQGTKV EIKRT

MS-Pro-24-VL (SEQ ID NO:96)
35      1   DIVLTQSPAT LSLSPGERAT LSCRASQSVS SSYLAWYQQK PGQAPRLLIY
      51   GASSRATGVP ARFSGSGSGT DFTLTISSE PEDFATYYCQ QMSNYPDTFG
      101  QGTKVEIKRT
40

MS-Pro-26-VL (SEQ ID NO:97)
      1   DIALTQPPASV SGSPGQSITI SCTGTSSDVG GYNYVS WYQQ HPGKAPKLMI
      51   YDVSNRPSGV SNRFSGSKSG NTASLTISGL QAEDADYYC QSYDNNSDVV
45      101  FGGGTKLTVL GQ

MS-Pro-28-VL (SEQ ID NO:98)
50      1   DIQMTQSPSS LSASVGDRVT ITCRASQGIS SYLAWYQQKPGKAPKLLIYA
      51   ASSLQSGVPS RFSGSGSGTD FTLTISSLQP EDFAVYYCFQ YGSIPPTFGQ
      101  GTKVEIKRT
55

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MS-Pro-29-VL (SEQ ID NO:99)
 1 DIVLTQSPAT LSLSPGERAT LSCRASQSVS SSYLAWYQQK PGQAPRLLIY
 5 51 GASSRATGVP ARFSGSGSGT DFTLT ISSLE PEDFATYYCQ QTNNAPVTFG
 101 QGTKVEIKRT

MS-Pro-54-VL (SEQ ID NO:100)
 10 1 DIELTQPPSV SVAPGQTARI SCSGDALGDK YASWYQQKPG QAPVLVIYDD
 51 SDRPSGIPER FSGSNSGNTA TLTISGTQAE DEADYYCQSY DYFKLVFGGG
 15 101 TKLTVLGQ

MS-Pro-55-VL (SEQ ID NO:101)
 1 DIALTQPASV SGSPGQSITI SCTGTSSDVG GYNYVS WYQQ HPGKAPKLMI
 20 51 YDVSNRPSGV SNRFSGSKSG NTASLTISGL QAEDDADYYC QSYDMYNYIV
 101 FGGGTKLTVL GQ

MS-Pro-59-VL (SEQ ID NO:102)
 25 1 DIELTQPPSV SVAPGQTARI SCSGDALGDK YASWYQQKPG QAPVLVIYDD
 51 SDRPSGIPER FSGSNSGNTA TLTISGTQAE DEADYYCQSY DGPDLWVFGG
 101 GTKLTVLGQ
 30

MS-Pro-2-VH (SEQ ID NO:103)
 1 QVQLVQSGAE VKKPGASVKV SCKASGYTFT SYMHWRQA PGQGLEWMGW
 51 INPNSSGGTNY AQKFQGRVTM TRDTSISTAY MELSSLRSED TAVYYCARD
 35 101 LGYEFDYWGQ GTLVTVSS

MS-Pro-11-VH (SEQ ID NO:104)
 40 1 QVQLVQSGAE VKKPGASVKV SCKASGYTFT SYMHWRQA PGQGLEWMGW
 51 INPNSSGGTNY AQKFQGRVTM TRDTSISTAY MELSSLRSED TAVYYCARYY
 101 GSSLYHYVFG GFIDYWGQGT LTVSS
 45

MS-Pro-12-VH (SEQ ID NO:105)
 1 QVQLKESGPA LVKPTQTLTL TCTFSGFSLT TSGVGVGWIR QPPGKALEWL
 51 ALIDWDDDKY YSTSLKTRLT ISKDTSKNQV VLTMTNMDPV DTATYYCARY
 50 101 HSWYEMGYYG STVG YMFYDW GQGLTVTVSS

MS-Pro-21-VH (SEQ ID NO:106)
 55 1 QVQLVQSGAE VKKPGSSVKV SCKASGGTFS SYAISWRQA PGQGLEWMGG
 51 IIPIFGTANY AQKFQGRVTI TADESTSTAY MELSSLRSED TAVYYCARDN
 101 WFKPFSDVWG QGTLTVTVSS
 60

MS-Pro-24-VH (SEQ ID NO:107)
 1 QVQLVQSGAE VKKP GSSVKV SCKAS GGTFS SYAISWVRQA PGQGLEWMGG
 5 51 IIPIFGTANY AQKFQGRVTI TADESTSTAY MELSSLRSED TAVYYCARVN
 101 HWTYTFDYWG QGTLVTVSS

MS-Pro-26-VH (SEQ ID NO:108)
 10 1 QVQLVQSGAE VKKP GASVKV SCKAS GYTFT SYMHWRQA PGQGLEWMGW
 51 INPNSGGTNY AQKFQGRVTM TRDTS ISTAY MELSSLRSED TAVYYCARGY
 101 WYAYFTYINY GYFDNWQGT LVTVSS

15 MS-Pro-28-VH (SEQ ID NO:109)
 1 QVQLVQSGAE VKKP GSSVKV SCKAS GGTFS SYAISWVRQA PGQGLEWMGG
 51 IIPIFGTANY AQKFQGRVTI TADESTSTAY MELSSLRSED TAVYYCARGG
 20 101 GWVSHGYYYL FDLWGQGTLV TVSS

MS-Pro-29-VH (SEQ ID NO:110)
 25 1 QVQLVQSGAE VKKP GASVKV SCKAS GYTFT SYMHWRQA PGQGLEWMGW
 51 INPNSGGTNY AQKFQGRVTM TRDTS ISTAY MELSSLRSED TAVYYCARTW
 101 QYSYFYLDG GYFDIWQGT TLVTVSS

30 MS-Pro-54-VH (SEQ ID NO:111)
 1 QVQLVQSGAE VKKP GASVKV SCKAS GYTFT SYMHWRQA PGQGLEWMGW
 51 INPNSGGTNY AQKFQGRVTM TRDTS ISTAY MELSSLRSED TAVYYCARNM
 35 101 AYTNYQYVNM PHFDYWGQGT LVTVSS

MS-Pro-55-VH (SEQ ID NO:112)
 1 QVQLVQSGAE VKKP GASVKV SCKAS GYTFT SYMHWRQA PGQGLEWMGW
 40 51 INPNSGGTNY AQKFQGRVTM TRDTS ISTAY MELSSLRSED TAVYYCARSM
 101 NSTMYWYLRRLVFDHWGQGT LVTVSS

45 MS-Pro-59-VH (SEQ ID NO:113)
 1 QVQLQQSGPG LVKPSQTLSTL TCAISGDSVS SNSAAWNWIR QSPGRGLEWL
 51 GRTYYRSKWY NDYAVSVKSR ITINPDTSKN QFSLQLNSVT PEDTAVYYCA
 50 101 RSYYPDFDYWGQGT LVTVSS

In addition to sequencing of the clones, a series of biochemical assays were performed to determine affinity and specificity of the molecules to the respective receptors. Table 1C lists the affinity of the respective molecules to FGFR3 and FGFR1 as measures by Biacore and/or
 55 FACS. In a binding assay to FGFR3-expressing cells the IC₅₀ of the molecules was

calculated (Example 6). Domain specificity was determined as described in Example 9. The ligand-independent inhibition of FGFR3 (neutralizing activity) was determined as described in Example 11. Finally, the molecules were synthesized in a number of different formats including Fab, miniantibody (Fab-dHLX), IgG1, IgG4, IgG3 and as single chain Fv (scFv).

5 **Table 1C**

Clone	Affinity to FGFR3 (BIAcore)	Affinity to FGFR3 (FACS)	Affinity to FGFR1	Koff (s ⁻¹)	IC50 FR3 (FGF9)	Domain Specificity	Ligand independent inhibition of FGFR3	Available formats
MSPRO59	1.5nM	<1nM	-	7.1x10e-4	19 nM	2	+	Fab, Fab-dHLX IgG1, IgG4, mIgG3, scFv
MSPRO2	37nM	43 nM	-	2x10e-2	360 nM	2	~	Fab(+/- tags), Fab-dHLX, IgG1, IgG4,
MSPRO12	14nM	6.5 nM	-	2.3x10e-3	58 nM	2	+	Fab (+/- tag), Fab-dHLX, IgG1, IgG4, scFv
MSPRO11	4	4 nM	108	4 x 10e-4	220 nM	3		Fab. Fab-dHLX
MSPRO21	9 nM		-	3.6x10e-3	50 nM	3c		Fab, IgG1, Fab-dHLX
MSPRO24	10 nM		-	5.4x10e-3	70 nM	3c		Fab, IgG1
MSPRO26	4 nM	1.4		5 x 10e-4	70 nM	3		Fab, Fab-dHLX
MSPRO28	9 nM	0.3 nM	160 nM	4 x 10e-3	50 nM	3		Fab
MSPRO29	6 nM	<1nM	29 nM	1.4x10e-3	20nM	3c	-	Fab (+/- tag), IgG1, IgG4, Fab-dHLX, scFv
MSPRO54	3.7nM		2.5nM	2x10e-3	45nM	3c		Fab, IgG1
MSPRO55	2.9nM		-	7.4x10e-4	34nM	3c		Fab

BiaCore results for certain molecules

In Table 1D the numbers are the IC₅₀s of the dimeric dHLX format of certain binders (molecule with antigen binding site) in the FDCP-FGFR3 proliferation assay performed with FGF9. The numbers in parentheses are the IC₅₀ of the monomeric Fabs in the same assay.

- 5 Table 1E presents the KD value for certain MSPRO molecules in miniantibody form, as determined in a Biacore.

Table 1D

binder	IC₅₀
MSPRO2	61 nM (360)
MSPRO12	26 nM (58)
MSPRO21	20 nM (50)
MSPRO26	8 nM (70)

Table 1E

- 10 **K_D determination for certain molecules**

Clone	BIAcore K_D [nM]	Number of measurements
MS-Pro-2-dHLX-MH	4.3 (37)	1
MS-Pro-11-dHLX-MH	0.7 (4)	1
MS-Pro-12-dHLX-MH	1.2 (14)	1
MS-Pro-21-dHLX-MH	2.2 (4.1)	1
MS-Pro-24-dHLX-MH	2 (10)	1
MS-Pro-26-dHLX-MH	2 (4)	1
MS-Pro-28-dHLX-MH	1.6 (9)	1

- The preferred, but non-limiting, embodiments of molecules according to the present invention that block constitutive (ligand-independent) activation of FGFR3 are referred to herein MSPRO2, MSPRO12 and MSPRO59 comprising VH-CDR3 and VL-CDR3 domains having SEQ ID Nos: 8 and 9; 12 and 13; and 24 and 25, respectively. The preferred, but non-
- 15 limiting, embodiments of molecules according to the present invention that block ligand-dependent activation of FGFR3 are referred to herein MSPRO11, MSPRO21, MSPRO24, MSPRO26, MSPRO29, and MSPRO54 comprising VH-CDR3 and VL-CDR3 domains

having SEQ ID Nos: 10 and 11; 14 and 15; 16 and 17, 18 and 19; 21 and 22; 23 and 24, respectively. Preferably, an antibody or a molecule of the present invention has an affinity (K_D) for binding a soluble dimeric form of FGFR3 of less than about 50 nM, preferably less than about 30 nM and more preferably less than about 10 nM, as determined by the BIAcore
5 chip assay for affinity, by a FACS-Scatchard analysis or other methods known in the art.

While the specific discovery of an antibody/molecule that blocks constitutive activation was made with respect to FGFR3 using a soluble dimeric form of FGFR3 to screen a phage display antibody library, it is believed that for all, or almost all receptor protein tyrosine kinases, antibodies/molecules that block constitutive activation can be similarly obtained
10 using a soluble dimeric form of a corresponding extracellular domain of a receptor protein tyrosine kinase. Non-limiting examples of receptor protein tyrosine kinases disclosed in Blume-Jensen et al. (2001) include EGFR/ErbB1, ErbB2/HER2/Neu, ErbB/HER3, ErbB4/HER4, IGF-1R, PDGFR- α , PDGFR- β , CSF-1R, kit/SCFR, Flk2/FH3, Flk1/VEGFR1, Flk1/VEGFR2, Flt4/VEGFR3, FGFR1, FGFR2/K-SAM, FGFR3, FGFR4, TrkA, TrkC,
15 HGFR, RON, EphA2, EphB2, EphB4, Axl, TIE/TIE1, Tek/TIE2, Ret, ROSA1k, Ryk, DDR, LTK and MUSK.

Furthermore, antibodies/molecules that block ligand-dependent or ligand independent activation of heterodimer receptor protein tyrosine kinases are intended to be included in the scope of the invention. Heterodimerization is well documented for members of the EGFR
20 subfamily of receptor protein tyrosine kinases and others. Non-limiting examples include EGFR/PDGFR β , Flt1/KDR and EGFR/ErbB2 heterodimers. EGFR and PDGFR β heterodimers have been identified as a mechanism for PDGF signal transduction in rat vascular smooth muscle cells (Saito et al., 2001) and Flt-1/KDR heterodimers are required for vinculin assembly in focal adhesions in response to VEGF signaling (Sato et al., 2000).
25 The present invention is also directed to a molecule having the antigen-binding portion of an antibody which binds to a dimeric form of an extracellular portion of a receptor protein tyrosine kinase (RPTK), such as a FGFR, and blocks the ligand-independent (constitutive) activation and/or ligand-dependent activation of the RPTK.

Antibodies

30 Antibodies, or immunoglobulins, comprise two heavy chains linked together by disulfide bonds and two light chains, each light chain being linked to a respective heavy chain by disulfide bonds in a "Y" shaped configuration. Proteolytic digestion of an antibody yields Fv

(Fragment variable and Fc (fragment crystalline) domains. The antigen binding domains, Fab', include regions where the polypeptide sequence varies. The term F(ab')₂ represents two Fab' arms linked together by disulfide bonds. The central axis of the antibody is termed the Fc fragment. Each heavy chain has at one end a variable domain (VH) followed by a number
5 of constant domains (CH). Each light chain has a variable domain (VL) at one end and a constant domain (CL) at its other end, the light chain variable domain being aligned with the variable domain of the heavy chain and the light chain constant domain being aligned with the first constant domain of the heavy chain (CH1).

The variable domains of each pair of light and heavy chains form the antigen binding site.

10 The domains on the light and heavy chains have the same general structure and each domain comprises four framework regions, whose sequences are relatively conserved, joined by three hypervariable domains known as complementarity determining regions (CDR1-3). These domains contribute specificity and affinity of the antigen binding site.

The isotype of the heavy chain (gamma, alpha, delta, epsilon or mu) determines
15 immunoglobulin class (IgG, IgA, IgD, IgE or IgM, respectively). The light chain is either of two isotypes (kappa,κ or lambda,λ) found in all antibody classes.

It should be understood that when the terms "antibody" or "antibodies" are used, this is intended to include intact antibodies, such as polyclonal antibodies or monoclonal antibodies (mAbs), as well as proteolytic fragments thereof such as the Fab or F(ab')₂ fragments. Further
20 included within the scope of the invention are chimeric antibodies; human and humanized antibodies; recombinant and engineered antibodies, and fragments thereof. Furthermore, the DNA encoding the variable region of the antibody can be inserted into the DNA encoding other antibodies to produce chimeric antibodies (see, for example, US patent 4,816,567).

Single chain antibodies fall within the scope of the present invention. Single chain antibodies
25 can be single chain composite polypeptides having antigen binding capabilities and comprising amino acid sequences homologous or analogous to the variable regions of an immunoglobulin light and heavy chain (linked VH-VL or single chain Fv (ScFv)). Both V_H and V_L may copy natural monoclonal antibody sequences or one or both of the chains may comprise a CDR-FR construct of the type described in US patent 5,091,513, the entire
30 contents of which are hereby incorporated herein by reference. The separate polypeptides analogous to the variable regions of the light and heavy chains are held together by a polypeptide linker. Methods of production of such single chain antibodies, particularly

where the DNA encoding the polypeptide structures of the V_H and V_L chains are known, may be accomplished in accordance with the methods described, for example, in US patents 4,946,778, 5,091,513 and 5,096,815, the entire contents of each of which are hereby incorporated herein by reference.

- 5 Additionally, CDR grafting may be performed to alter certain properties of the antibody molecule including affinity or specificity. A non-limiting example of CDR grafting is disclosed in US patent 5,225,539.

10 A "molecule having the antigen-binding portion of an antibody" as used herein is intended to include not only intact immunoglobulin molecules of any isotype and generated by any animal cell line or microorganism, but also the antigen-binding reactive fraction thereof, including, but not limited to, the Fab fragment, the Fab' fragment, the F(ab')₂ fragment, the variable portion of the heavy and/or light chains thereof, Fab miniantibodies (see WO 93/15210, US patent application 08/256,790, WO 96/13583, US patent application 08/817,788, WO 96/37621, US patent application 08/999,554, the entire contents of which are incorporated herein by reference) and chimeric or single-chain antibodies incorporating such reactive fraction, as well as any other type of molecule or cell in which such antibody reactive fraction has been physically inserted, such as a chimeric T-cell receptor or a T-cell having such a receptor, or molecules developed to deliver therapeutic moieties by means of a portion of the molecule containing such a reactive fraction. Such molecules may be provided by any known technique, including, but not limited to, enzymatic cleavage, peptide synthesis or recombinant techniques.

The term "Fc" as used herein is meant as that portion of an immunoglobulin molecule (Fragment crystallizable) that mediates phagocytosis, triggers inflammation and targets Ig to particular tissues; the Fc portion is also important in complement activation.

- 25 In one embodiment of the invention, a chimera comprising a fusion of the extracellular domain of the RPTK and an immunoglobulin constant domain can be constructed useful for assaying for ligands for the receptor and for screening for antibodies and fragments thereof

The "extracellular domain" when used herein refers the polypeptide sequence of the RPTKs disclosed herein which are normally positioned to the outside of the cell. The extracellular domain encompasses polypeptide sequences in which part of or all of the adjacent (C-terminal) hydrophobic transmembrane and intracellular sequences of the mature RPTK have

30

been deleted. Thus, the extracellular domain-containing polypeptide can comprise the extracellular domain and a part of the transmembrane domain. Alternatively, in the preferred embodiment, the polypeptide comprises only the extracellular domain of the RPTK. The truncated extracellular domain is generally soluble. The skilled practitioner can readily
5 determine the extracellular and transmembrane domains of a RPTK by aligning the RPTK of interest with known RPTK amino acid sequences for which these domains have been delineated. Alternatively, the hydrophobic transmembrane domain can be readily delineated based on a hydrophobicity plot of the polypeptide sequence. The extracellular domain is N-terminal to the transmembrane domain.

10 The term "epitope" is meant to refer to that portion of any molecule capable of being bound by an antibody or a fragment thereof which can also be recognized by that antibody. Epitopes or antigenic determinants usually consist of chemically active surface groupings of molecules such as amino acids or sugar side chains and have specific three-dimensional structural characteristics as well as specific charge characteristics.

15 An "antigen" is a molecule or a portion of a molecule capable of being bound by an antibody which is additionally capable of inducing an animal to produce antibody capable of binding to an epitope of that antigen. An antigen may have one or more than one epitope. The specific reaction referred to above is meant to indicate that the antigen will react, in a highly selective manner, with its corresponding antibody and not with the multitude of other
20 antibodies which may be evoked by other antigens.

A "neutralizing antibody" as used herein refers to a molecule having an antigen binding site to a specific receptor capable of reducing or inhibiting (blocking) activity or signaling through a receptor, as determined by *in vivo* or *in vitro* assays, as per the specification.

A monoclonal antibody (mAb) is a substantially homogeneous population of antibodies to a
25 specific antigen. MAbs may be obtained by methods known to those skilled in the art. See, for example Kohler et al (1975); US patent 4,376,110; Ausubel et al (1987-1999); Harlow et al (1988); and Colligan et al (1993), the contents of which references are incorporated entirely herein by reference. The mAbs of the present invention may be of any immunoglobulin class including IgG, IgM, IgE, IgA, and any subclass thereof. A hybridoma
30 producing an mAb may be cultivated *in vitro* or *in vivo*. High titers of mAbs can be obtained in *in vivo* production where cells from the individual hybridomas are injected intraperitoneally into pristine-primed Balb/c mice to produce ascites fluid containing high

concentrations of the desired mAbs. MAbs of isotype IgM or IgG may be purified from such ascites fluids, or from culture supernatants, using column chromatography methods well known to those of skill in the art.

Chimeric antibodies are molecules, the different portions of which are derived from different animal species, such as those having a variable region derived from a murine mAb and a human immunoglobulin constant region. Antibodies which have variable region framework residues substantially from human antibody (termed an acceptor antibody) and complementarity determining regions substantially from a mouse antibody (termed a donor antibody) are also referred to as humanized antibodies. Chimeric antibodies are primarily used to reduce immunogenicity in application and to increase yields in production, for example, where murine mAbs have higher yields from hybridomas but higher immunogenicity in humans, such that human/murine chimeric mAbs are used. Chimeric antibodies and methods for their production are known in the art (Better et al, 1988; Cabilly et al, 1984; Harlow et al, 1988; Liu et al, 1987; Morrison et al, 1984; Boulianne et al, 1984; Neuberger et al, 1985; Sahagan et al, 1986; Sun et al, 1987; Cabilly et al; European Patent Applications 125023, 171496, 173494, 184187, 173494, PCT patent applications WO 86/01533, WO 97/02671, WO 90/07861, WO 92/22653 and US patents 5,693,762, 5,693,761, 5,585,089, 5,530,101 and 5,225,539). These references are hereby incorporated by reference.

Besides the conventional method of raising antibodies *in vivo*, antibodies can be generated *in vitro* using phage display technology. Such a production of recombinant antibodies is much faster compared to conventional antibody production and they can be generated against an enormous number of antigens. In contrast, in the conventional method, many antigens prove to be non-immunogenic or extremely toxic, and therefore cannot be used to generate antibodies in animals. Moreover, affinity maturation (i.e., increasing the affinity and specificity) of recombinant antibodies is very simple and relatively fast. Finally, large numbers of different antibodies against a specific antigen can be generated in one selection procedure. To generate recombinant monoclonal antibodies one can use various methods all based on phage display libraries to generate a large pool of antibodies with different antigen recognition sites. Such a library can be made in several ways: One can generate a synthetic repertoire by cloning synthetic CDR3 regions in a pool of heavy chain germline genes and thus generating a large antibody repertoire, from which recombinant antibody fragments with

various specificities can be selected. One can use the lymphocyte pool of humans as starting material for the construction of an antibody library. It is possible to construct naive repertoires of human IgM antibodies and thus create a human library of large diversity. This method has been widely used successfully to select a large number of antibodies against
5 different antigens. Protocols for bacteriophage library construction and selection of recombinant antibodies are provided in the well-known reference text Current Protocols in Immunology, Colligan et al (Eds.), John Wiley & Sons, Inc. (1992-2000), Chapter 17, Section 17.1.

Another aspect of the present invention is directed to a method for screening for the antibody
10 or molecule of the present invention by screening a library of antibody fragments displayed on the surface of bacteriophage, such as disclosed in the Example herein and described in WO 97/08320, US Patent 6,300,064 , and Knappik et al. (2000), for binding to a soluble dimeric form of a receptor protein tyrosine kinase. An antibody fragment which binds to the soluble dimeric form of the RPTK used for screening is identified as a candidate molecule for
15 blocking ligand-dependent activation and/or constitutive activation of the RPTK in a cell. Preferably the RPTK of which a soluble dimeric form is used in the screening method is a fibroblast growth factor receptor (FGFR), and most preferably FGFR3.

As a first screening method, the soluble dimeric form of a receptor tyrosine kinase can be constructed and prepared in a number of different ways. For instance, the extracellular
20 domain of a RPTK joined to Fc and expressed as a fusion polypeptide that dimerizes naturally by means of the Fc portion of the RPTK-Fc fusion. Other suitable types of constructs of FGFR3, serving as guidance for other RPTKs, are disclosed in the Examples presented herein.

The assays for determining binding of antibody fragments to FGFR3, binding affinities,
25 inhibition of cell proliferation, etc., are also described in the Examples herein below.

The term "cell proliferation" refers to the rate at which a group of cells divides. The number of cells growing in a vessel can be quantified by a person skilled in the art when that person visually counts the number of cells in a defined volume using a common light microscope. Alternatively, cell proliferation rates can be quantified by laboratory apparatus that optically or
30 conductively measure the density of cells in an appropriate medium.

A second screen for antibody fragments as candidate molecules can be done using cells having very high over expression of the RPTK, such as for instance RCJ-M15 cells

overexpressing mutant (achondroplasia) FGFR3. In cells expressing very high levels of receptor some ligand-independent activation occurs even without the presence of a mutation, such as a constitutive activation mutation. It is believed that RPTK overexpression forces RPTKs to dimerize and signal even in the absence of ligand. These cells have monomeric
5 receptors as well as dimeric receptors present on their cell surface. Using this type of cell, one of skill in the art would be able to identify all different kinds of antibodies, i.e., blocking ligand-dependent activation, blocking constitutive activation, blocking activation and binding only to monomeric form, etc.

A third screen can be carried out on a cell line expressing a RPTK carrying a mutation, such
10 as the FDCP-FR3ach line expressing the FGFR3 achondroplasia mutation. The receptors of this line become constitutively active, e.g. are able to signal in the absence of a ligand as determined by ERK (MAPK) phosphorylation, a downstream effector.

A further aspect of the present invention relates to a method for treating or inhibiting a skeletal dysplasia or craniosynostosis disorder associated with constitutive activation of a
15 RPTK which involves administering the molecule of the present invention to a subject need thereof. Non-limiting examples of skeletal dysplasias include achondroplasia, thanatophoric dysplasia (TDI or TDII), hypochondroplasia, and severe achondroplasia with developmental delay and acanthosis nigricans (SADDAN) dysplasia. Non-limiting examples of craniosynostosis disorder are Muenke coronal craniosynostosis and Crouzon syndrome with
20 acanthosis nigricans. The symptoms and etiology of these diseases and disorders are reviewed in Vajo et al. (Vajo et al, 2000).

The present invention also provides for a method for treating or inhibiting a cell proliferative disease or disorder associated with the action of an abnormal constitutively activated RPTK, for example. tumor formation, primary tumors, tumor progression or tumor metastasis. A
25 molecule containing the antigen binding portion of an antibody that blocks constitutive activation of a RPTK is administered to a subject in need thereof to treat or inhibit such a cell proliferative disease or disorder.

The terms "treating or inhibiting a proliferative disease or disorder" or "treating or inhibiting a tumor" are used herein and in the claims to encompass tumor formation, primary tumors,
30 tumor progression or tumor metastasis.

Tumor formation or tumor growth are intended to encompass solid and non-solid tumors. Solid tumors include mammary, ovarian, prostate, colon, cervical, gastric, esophageal,

papillary thyroid, pancreatic, bladder, colorectal, melanoma, small-cell lung and non-small-cell lung cancers, granulose cell carcinoma, transitional cell carcinoma, vascular tumors, all types of sarcomas, e.g. osteosarcoma, chondrosarcoma, Kaposi's sarcoma, myosarcoma, hemangiosarcoma, and glioblastomas.

- 5 Non-solid tumors include for example hematopoietic malignancies such as all types of leukemia, e.g. chronic myelogenous leukemia (CML), acute myelogenous leukemia (AML), mast cell leukemia, chronic lymphocytic leukemia (CLL) and acute lymphocytic leukemia (ALL), lymphomas, and multiple myeloma (MM).

Tumor progression is the phenomenon whereby cancers become more aggressive with time.

- 10 Progression can occur in the course of continuous growth, or when a tumor recurs after treatment and includes progression of transitional cell carcinoma, osteo or chondrosarcoma, multiple myeloma, and mammary carcinoma (one of the known RPTKs involved in mammary carcinoma is ErbB3).

- 15 The role of the FGFR3 in tumor progression associated with transitional cell carcinoma and multiple myeloma has recently been elucidated (Cappellen, et al, 1999; Chesi, et al, 2001)

- In another aspect of the present invention, molecules which bind FGFR, more preferably FGFR3, and block ligand-dependent receptor activation are provided. These molecules are useful in treating hyperproliferative diseases or disorders and non-neoplastic angiogenic pathologic conditions such as neovascular glaucoma, proliferative retinopathy including
20 proliferative diabetic retinopathy, macular degeneration, hemangiomas, angiofibromas, and psoriasis. The role of FGFs and their receptors in neo- and hypervascularization has been well documented (Frank, 1997; Gerwins et al, 2000)

- In another aspect of the present invention, the pharmaceutical compositions according to the present invention is similar to those used for passive immunization of humans with other
25 antibodies. Typically, the molecules of the present invention comprising the antigen binding portion of an antibody will be suspended in a sterile saline solution for therapeutic uses. The pharmaceutical compositions may alternatively be formulated to control release of active ingredient (molecule comprising the antigen binding portion of an antibody) or to prolong its presence in a patient's system. Numerous suitable drug delivery systems are known and
30 include, e.g., implantable drug release systems, hydrogels, hydroxymethylcellulose, microcapsules, liposomes, microemulsions, microspheres, and the like. Controlled release preparations can be prepared through the use of polymers to complex or adsorb the molecule

according to the present invention. For example, biocompatible polymers include matrices of poly(ethylene-co-vinyl acetate) and matrices of a polyanhydride copolymer of a stearic acid dimer and sebacic acid (Sherwood et al, 1992). The rate of release molecule according to the present invention, i.e., of an antibody or antibody fragment, from such a matrix depends upon
5 the molecular weight of the molecule, the amount of the molecule within the matrix, and the size of dispersed particles (Saltzman et al., 1989 and Sherwood et al., 1992). Other solid dosage forms are described in (Ansel et al., 1990 and Gennaro, 1990).

The pharmaceutical composition of this invention may be administered by any suitable means, such as orally, intranasally, subcutaneously, intramuscularly, intravenously, intra-
10 arterially, intralesionally or parenterally. Ordinarily, intravenous (i.v.) or parenteral administration will be preferred.

It will be apparent to those of ordinary skill in the art that the therapeutically effective amount of the molecule according to the present invention will depend, *inter alia* upon the administration schedule, the unit dose of molecule administered, whether the molecule is
15 administered in combination with other therapeutic agents, the immune status and health of the patient, the therapeutic activity of the molecule administered and the judgment of the treating physician. As used herein, a "therapeutically effective amount" refers to the amount of a molecule required to alleviate one or more symptoms associated with a disorder being treated over a period of time.

20 Although an appropriate dosage of a molecule of the invention varies depending on the administration route, age, body weight, sex, or conditions of the patient, and should be determined by the physician in the end, in the case of oral administration, the daily dosage can generally be between about 0.01-200 mg, preferably about 0.01-10 mg, more preferably about 0.1-10 mg, per kg body weight. In the case of parenteral administration, the daily
25 dosage can generally be between about 0.001-100 mg, preferably about 0.001-1 mg, more preferably about 0.01-1 mg, per kg body weight. The daily dosage can be administered, for example in regimens typical of 1-4 individual administration daily. Various considerations in arriving at an effective amount are described, e.g., in Goodman and Gilman's: The Pharmacological Bases of Therapeutics, 8th ed., Pergamon Press, 1990; and Remington's
30 Pharmaceutical Sciences, 17th ed., Mack Publishing Co., Easton, Pa., 1990.

The molecule of the present invention as an active ingredient is dissolved, dispersed or admixed in an excipient that is pharmaceutically acceptable and compatible with the active

ingredient as is well known. Suitable excipients are, for example, water, saline, phosphate buffered saline (PBS), dextrose, glycerol, ethanol, or the like and combinations thereof. Other suitable carriers are well-known to those in the art. (See, for example, Ansel et al., 1990 and Gennaro, 1990). In addition, if desired, the composition can contain minor
5 amounts of auxiliary substances such as wetting or emulsifying agents, pH buffering agents. Combination therapy

The combined treatment of one or more of the molecules of the invention with an anti-neoplastic or anti-chemotherapeutic drug such as doxorubicin, cisplatin or taxol provides a more efficient treatment for inhibiting the growth of tumor cells than the use of the molecule
10 by itself. In one embodiment, the pharmaceutical composition comprises the antibody and carrier with an anti-chemotherapeutic drug.

The present invention also provides for a nucleic acid molecule, which contains a nucleotide sequence encoding the molecule having the antigen binding portion of an antibody that blocks ligand-dependent activation and/or constitutive activation of a receptor protein
15 tyrosine kinase such as FGFR3, and a host cell transformed with this nucleic acid molecule. Furthermore, also within the scope of the present invention is a nucleic acid molecule containing a nucleotide sequence having at least 90% sequence identity, preferably about 95%, and more preferably about 97% identity to the above encoding nucleotide sequence as would well understood by those of skill in the art.

20 The invention also provides nucleic acids that hybridize under high stringency conditions to polynucleotides having SEQ ID NOs: 8 through 29 and SEQ ID NOs: 62, 64-65, 67, 69-71, 73-76 78-80, 82-87, 89, 90-91 or the complement thereof. As used herein, highly stringent conditions are those which are tolerant of up to about 5-20% sequence divergence, preferably about 5-10%. Without limitation, examples of highly stringent (-10°C below the calculated
25 T_m of the hybrid) conditions use a wash solution of 0.1 X SSC (standard saline citrate) and 0.5% SDS at the appropriate T_i below the calculated T_m of the hybrid. The ultimate stringency of the conditions is primarily due to the washing conditions, particularly if the hybridization conditions used are those which allow less stable hybrids to form along with stable hybrids. The wash conditions at higher stringency then remove the less stable hybrids.
30 A common hybridization condition that can be used with the highly stringent to moderately stringent wash conditions described above is hybridization in a solution of 6 X SSC (or 6 X SSPE), 5 X Denhardt's reagent, 0.5% SDS, 100 µg/ml denatured, fragmented salmon sperm

DNA at an appropriate incubation temperature T_i . *See generally* Sambrook *et al.*, Molecular Cloning: A Laboratory Manual, 2d edition, Cold Spring Harbor Press (1989)) for suitable high stringency conditions.

Stringency conditions are a function of the temperature used in the hybridization experiment and washes, the molarity of the monovalent cations in the hybridization solution and in the wash solution(s) and the percentage of formamide in the hybridization solution. In general, sensitivity by hybridization with a probe is affected by the amount and specific activity of the probe, the amount of the target nucleic acid, the detectability of the label, the rate of hybridization, and the duration of the hybridization. The hybridization rate is maximized at a T_i (incubation temperature) of 20-25°C below T_m for DNA:DNA hybrids and 10-15°C below T_m for DNA:RNA hybrids. It is also maximized by an ionic strength of about 1.5M Na^+ . The rate is directly proportional to duplex length and inversely proportional to the degree of mismatching.

Specificity in hybridization, however, is a function of the difference in stability between the desired hybrid and "background" hybrids. Hybrid stability is a function of duplex length, base composition, ionic strength, mismatching, and destabilizing agents (if any).

The T_m of a perfect hybrid may be estimated for DNA:DNA hybrids using the equation of Meinkoth *et al* (1984), as

$$T_m = 81.5^\circ\text{C} + 16.6 (\log M) + 0.41 (\%GC) - 0.61 (\% \text{ form}) - 500/L$$

and for DNA:RNA hybrids, as

$$T_m = 79.8^\circ\text{C} + 18.5 (\log M) + 0.58 (\%GC) - 11.8 (\%GC)^2 - 0.56(\% \text{ form}) - 820/L$$

where M , molarity of monovalent cations, 0.01-0.4 M NaCl,
 $\%GC$, percentage of G and C nucleotides in DNA, 30%-75%,
 $\% \text{ form}$, percentage formamide in hybridization solution, and

L , length hybrid in base pairs.

T_m is reduced by 0.5-1.5°C (an average of 1°C can be used for ease of calculation) for each 1% mismatching.

The T_m may also be determined experimentally. As increasing length of the hybrid (L) in the above equations increases the T_m and enhances stability, the full-length rat gene sequence can be used as the probe.

Filter hybridization is typically carried out at 68°C, and at high ionic strength (e.g., 5 - 6 X SSC), which is non-stringent, and followed by one or more washes of increasing stringency, the last one being of the ultimately desired high stringency. The equations for T_m can be used to estimate the appropriate T_i for the final wash, or the T_m of the perfect duplex can be determined experimentally and T_i then adjusted accordingly.

The present invention also relates to a vector comprising the nucleic acid molecule of the present invention. The vector of the present invention may be, e.g., a plasmid, cosmid, virus, bacteriophage or another vector used e.g. conventionally in genetic engineering, and may comprise further genes such as marker genes which allow for the selection of said vector in a suitable host cell and under suitable conditions.

Furthermore, the vector of the present invention may, in addition to the nucleic acid sequences of the invention, comprise expression control elements, allowing proper expression of the coding regions in suitable hosts. Such control elements are known to the artisan and may include a promoter, a splice cassette, translation initiation codon, translation and insertion site for introducing an insert into the vector.

Preferably, the nucleic acid molecule of the invention is operatively linked to said expression control sequences allowing expression in eukaryotic or prokaryotic cells.

Control elements ensuring expression in eukaryotic or prokaryotic cells are well known to those skilled in the art. As mentioned herein above, they usually comprise regulatory sequences ensuring initiation of transcription and optionally poly-A signals ensuring termination of transcription and stabilization of the transcript.

Methods for construction of nucleic acid molecules according to the present invention, for construction of vectors comprising said nucleic acid molecules, for introduction of said vectors into appropriately chosen host cells, for causing or achieving the expression are well-known in the art (see, e.g., Sambrook et al., 1989; Ausubel et al., 1994).

The invention also provides for conservative amino acid variants of the molecules of the invention. Variants according to the invention also may be made that conserve the overall molecular structure of the encoded proteins. Given the properties of the individual amino acids comprising the disclosed protein products, some rational substitutions will be recognized by the skilled worker. Amino acid substitutions, *i.e.* "conservative substitutions," may be made, for instance, on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues involved.

For example: (a) nonpolar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan, and methionine; (b) polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine; (c) positively charged (basic) amino acids include arginine, lysine, and histidine; and (d) negatively charged (acidic) amino acids include aspartic acid and glutamic acid. Substitutions typically may be made within groups (a)-(d). In addition, glycine and proline may be substituted for one another based on their ability to disrupt α -helices. Similarly, certain amino acids, such as alanine, cysteine, leucine, methionine, glutamic acid, glutamine, histidine and lysine are more commonly found in α helices, while valine, isoleucine, phenylalanine, tyrosine, tryptophan and threonine are more commonly found in β -pleated sheets. Glycine, serine, aspartic acid, asparagine, and proline are commonly found in turns. Some preferred substitutions may be made among the following groups: (i) S and T; (ii) P and G; and (iii) A, V, L and I. Given the known genetic code, and recombinant and synthetic DNA techniques, the skilled scientist readily can construct DNAs encoding the conservative amino acid variants.

As used herein, "sequence identity" between two polypeptide sequences indicates the percentage of amino acids that are identical between the sequences. "Sequence similarity" indicates the percentage of amino acids that either are identical or that represent conservative amino acid substitutions.

Conjugates

One embodiment of the present invention provides molecules of the present invention conjugated to cytotoxins. The cytotoxic moiety of the antibody may be a cytotoxic drug or an enzymatically active toxin or bacterial or plant origin, or an enzymatically active fragment of such a toxin including, but not limited to, diphtheria A chain, nonbinding active fragments of diphtheria toxin, exotoxin A chain (from *Pseudomonas aeruginosa*), ricin A chain, abrin A chain, modeccin A chain, alpha-sarcin, Aleurites fordii proteins, dianthin proteins, curcin, crotonin, saponin, gelonin, mitogellin, restrictocin, phenomycin, and enomycin. In another embodiment, the molecules of the present invention are conjugate to small molecule anti-cancer drugs. Conjugates of the antibody and such cytotoxic moieties are made using a variety of bifunctional protein coupling agents. Examples of such reagents include SPDP, IT, bifunctional derivatives of imidoesters such as dimethyl adipimidate HCl, active esters such as disuccinimidyl suberate, aldehydes such as glutaraldehyde, bis-azido compounds such as bis-

(p-azidobenzoyl) hexanediamine, bis-diazonium derivatives, dissocyanates and bis-active fluorine compounds. The lysing portion of a toxin may be joined to the Fab fragment of the antibodies.

5 Additionally, the molecules of the present invention can also be detected *in vivo* by imaging, for example imaging of cells which have undergone tumor progression or have metastasized. Antibody labels or markers for *in vivo* imaging of RPTKs include those detectable by X-radiography, NMR, PET, or ESR. For X-radiography, suitable labels include radioisotopes such as barium or cesium, which emit detectable radiation but are not overtly harmful to the subject. Suitable markers for NMR and ESR include those with a detectable characteristic
10 spin, such as deuterium, which may be incorporated into the antibody.

A specific antibody or antibody portion which has been labeled with an appropriate detectable imaging moiety, such as a radioisotope (for example, ^{131}I , ^{111}In , ^{99}Tc), a radio-opaque substance, or a material detectable by nuclear magnetic resonance, is introduced (for example, parenterally, subcutaneously or intraperitoneally) into the mammal to be examined
15 for a disorder. It will be understood in the art that the size of the subject and the imaging system used will determine the quantity of imaging moieties needed to produce diagnostic images. In the case of a radioisotope moiety, for a human subject, the quantity of radioactivity injected will normally range from about 5 to 20 millicuries. The labeled antibody or antibody portion will then preferentially accumulate at the location of cells which
20 contain a specific RPTK. In vivo tumor imaging is described in Burchiel et al., (1982

The methods and compositions described herein may be performed, for example, by utilizing pre-packaged diagnostic test kits comprising in one or more containers (i) at least one immunoglobulin of the invention and (ii) a reagent suitable for detecting the presence of said immunoglobulin when bound to its target. A kit may be conveniently used, e.g., in clinical
25 settings or in home settings, to diagnose patients exhibiting a disease (e.g., skeletal dysplasia, craniosynostosis disorders, cell proliferative diseases or disorders, or tumor progression), and to screen and identify those individuals exhibiting a predisposition to such disorders. A composition of the invention also may be used in conjunction with a reagent suitable for detecting the presence of said immunoglobulin when bound to its target, as well as
30 instructions for use, to carry out one or more methods of the invention.

Having now generally described the invention, the same will be more readily understood through reference to the following examples, which are provided by way of illustration and are not intended to be limiting of the present invention.

EXAMPLES

- 5 An important approach to control FGFR3 activity is the generation of reagents that block receptor signaling. Without wishing to be bound by theory, molecules which bind the extracellular domain of the receptor may inhibit the receptor by competing with FGF or heparin binding or, alternatively, by preventing receptor dimerization. Additionally, binding to the extracellular domain may accelerate receptor internalization and turnover. Humanized
10 antibodies are expected to have inhibitory/neutralizing action and are of particular interest since they are considered to be valuable for therapeutic applications, avoiding the human anti-mouse antibody response frequently observed with rodent antibodies. The experiments in which the neutralizing antibodies are screened, identified and obtained using fully synthetic human antibody libraries (for discovering highly specific binders against a wide
15 variety of antigens) and FGFR3 extracellular domain are described below.

Example 1: Attempt to generate anti-FGFR3 antibodies

- One hundred micrograms of soluble FGFR3 in complete Freund's Adjuvant were injected into Balb/c 3T3 naive mice (9 animals). Two repeated injections of 20 micrograms were
20 performed at week intervals. 10 days after the second booster injection, blood was drawn from animals and serum was tested for the presence of polyclonal antibodies both by monitoring for binding to the receptor as well as for neutralizing activity at a dilution of 1:50. No significant neutralizing activity was observed in the tested serum (20% at most in some animals). A perfusion injection of 20 micrograms of soluble receptor was administered 1-2
25 days later but all the mice harboring some activity of neutralizing Ab died. The experiment was repeated twice with the same results.

Example 2: Generation of the FGFR3 Antigens

- Two dimeric forms of the extracellular domain of the human FGFR3 were prepared for use as antigen. One was a histidine-tagged domain with a Serine 371 to Cysteine (S371C)
30 substitution (thanatophoric dysplasia (TD) mutation) to facilitate dimerization and the second one an Fc fusion. The S371C variant was shown to bind heparin and FGF9 coated plates and

to inhibit FGF9-dependent FDCP-FR3 proliferation. The Fc fusion was similarly effective in binding assays demonstrating its potential as an inhibitor of FGFR function and as a target for selecting FGFR3 inhibitory molecules. Both soluble receptors were employed to select neutralizing human recombinant antibodies.

5 The two variants of the FGFR3 extracellular domain were prepared as follows:

1. A construct containing the extracellular portion of FGFR3 with a thanatophoric dysplasia (TD) mutation to facilitate dimer formation conjugated to a His-tag (histidine tag) was generated. A bluescript plasmid comprising the human FGFR3 gene (pBS-hFGFR3) was used as template for PCR with the following primers:

10 5'-ACGTGCTAGC TGAGTCCTTG GGGACGGAGC AG (SEQ ID NO:2).
5'-ACGTCTCGAG TTAATGGTGA TGGTGATGGT GTGCATACAC **ACAG**CCCCGCC TCGTC (SEQ ID NO:3),

wherein the Ser 371 Cys (S371C) substitution is bold and underlined.

The nucleotide sequence encoding the extracellular domain of FGFR3 with the TD

15 substitution is denoted herein SEQ ID NO:7:

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TGAGTCCTTG GGGACGGAGC AGCGCGTCGT GGGGC GAGCG GCAGAA GTCC CGGGCCC AGA 60
GCCCCGCCAG CAG GAGCAGT TGGTCTTCGG CAGC GGGGAT GCTGT GGAGC TGAGCT GTCC 120
CCCCCCCCGGG GGT GGTCCCA TGGG GCCCAC TGTCT GGGTC AAGGAT GGCA CAGGGCT GGT 180
GCCCTCGGAG CGT GTCCTGG TGGG GCCCCA GCGGC TGCAG GTGCTGAATG CCTCCCA CGA 240
20 GGACTCCGGG GCC TACAGCT GCCG GCAGCG GCTCA CGCAG CGCGTA CTGT GCCACTT CAG 300
TGTGCGGGTG ACA GACGCTC CATC CTCGGG AGATG ACGAA GACGGG GAGG ACCAGGC TGA 360
GGACACAGGT GTG GACACAG GGGC CCCTTA CTGGA CACGG CCGGAG CGGA TGGACAA GAA 420
GCTGCTGGCC GTG CCGGCCG CCAA CACCGT CCGCT TCCGC TGCCCA GCCG CTGGCAA CCC 480
CACTCCCTCC ATC TCCTGGC TGAA GAACGG CAGGG AGTTC CGCGGC GAGC ACCGCAT TGG 540
25 AGGCATCAAG CTG CGGCATC AGCA GTGGAG CCTGG TCATG GAAAGC GTGG TGCCCTC GGA 600
CCGCGGCAAC TAC ACCTGCG TCGT GGAGAA CAAGT TTGGC AGCATC CGGC AGACGTA CAC 660
GCTGGACGTG CTG GAGCGCT CCCC GCACCG GCCCA TCCTG CAGGCG GGGC TGCCGGC CAA 720
CCAGACGGCG GTG CTGGGCA GCGA CGTGGA GTTCC ACTGC AAGGTG TACA GTGACGC ACA 780
GCCCCACATC CAG TGGCTCA AGCA CGTGGA GGTGA ACGGC AGCAAG GTGG GCCCGGA CGG 840
30 CACACCCTAC GTT ACCGTGC TCAA GACGGC GGGCG CTAAC ACCACC GACA AGGAGCT AGA 900
GGTTCTCTCC TTG CACAACG TCAC CTTTGA GGACG CCGGG GAGTAC ACCT GCCTGG CGGG 960
CAATTCTATT GGG TTTTCTC ATCA CTCTGC GTGGC TGGTG GTGCTG CCAG CCGAGGA GGA 1020
GCTGGTGGAG GCT GACGAGG CGGG CTGTGT GTATG CACAC CATCAC CATC ACCATTAA 1078

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The PCR fragment was digested with XhoI and ligated into pBlueScript digested with

35 EcoRV and XhoI. The resulting plasmid, pBsFR3²³⁻³⁷⁴Tdhis, was cleaved with NdeI and

XhoI and the DNA fragment encoding the extracellular domain of FGFR3 was ligated into the same restriction sites in pCEP-Pu/Ac7 (Yamaguchi et al., 1999; Kohfeldt et al., 1997), generating the pCEP-hFR3²³⁻³⁷⁴TDhis plasmid construct.

To express this FGFR3 variant, 293E cells (EBNA virus transfected 293 cells) were
 5 transfected with the aforementioned plasmid, pCEP-hFR3²³⁻³⁷⁴TDhis, clones were identified and grown. Cell supernatants analyzed by Western blot with anti-His antibody demonstrated high expression of the soluble receptor. Supernatants from large scale preparations were then subjected to batch affinity purification with Ni-NTA beads and the tagged soluble receptor was eluted by a step gradient ranging from 20 mM to 500 mM imidazol. A sample from each
 10 eluate was loaded onto a 7.5% SDS-PAGE and stained with GelCode (Pierce). In parallel Western blot analysis was performed and analyzed with anti-His antibodies. SDS-PAGE (Fig. 1) and immunoblot (not shown) analyses demonstrated peak amounts of purified extracellular FGFR3 in the 2nd (2) 50 mM imidazol fraction. About 0.5 mg of pure protein was obtained following this single step purification. In Figure 1, T=total protein, D= dialysed
 15 protein, U= unbound fraction.

To assess whether hFR3²³⁻³⁷⁴TDhis (hFR3-TDhis) retained the ability to associate with heparin and heparin-FGF complex, heparin coated wells were incubated with purified (2, 4 or 10 µg, labeled as FR3 2, FR3 4 or FR3 10, respectively in Fig. 2) or unpurified (FR3 sup) hFR3²³⁻³⁷⁴TDhis with (checkered bar) or without FGF9 (200ng/well, hatched bar). The
 20 binding of hFR3²³⁻³⁷⁴TDhis to each well was determined with anti-His antibody. Mock supernatant (M sup), PBS and unpurified mouse FR3AP (FGFR3-alkaline phosphatase, labeled as FRAP sup) were included as controls. This demonstrated that, like what was reported for the wild-type receptor, hFR3²³⁻³⁷⁴TDhis binds to heparin and that this interaction is augmented by the presence of FGF9 (Fig. 2). Finally, it was demonstrated that hFR3²³⁻³⁷⁴
 25 TDhis inhibits FDCP-FR3 FGF-dependent proliferation in a dose dependent manner. hFR3²³⁻³⁷⁴TDhis had no inhibitory effect on proliferation when FDCP-FR3 cells were grown in the presence of IL-3. Taken together, hFR3²³⁻³⁷⁴TDhis proved to be a good candidate as a target antigen for screening for FGFR3 neutralizing antibodies.

2. The extracellular domain of FGFR3 and FGFR1 were prepared as Fc fusions (FR3exFc
 30 and FR1exFc). The amino acid sequence of FGFR3 (NCBI access no: NP_000133) is denoted herein SEQ ID NO:1.

1 MGAPACALAL CVAVAIVAGA SSES LGTEQR VVGRAAEVPG PEPGQQEQLV FGSGDAVE LS

5

15

SEO ID NO:4:

20

25

30

35

AGGCGGG

1147

SEQ ID NO:5:

	GACGGATCGG GAGATCTCCC GATC CCCTAT GGTCGACTCT CAGTACAATC TGCTCTGATG	60
	CCGCATAGTT AAGCCAGTAT CTGCTCCCTG CTTGTGTGTT GGAGGT CGCT GAGTAGT GCG	120
5	CGAGCAAAAT TTAAGCTACA ACAAGGCAAG GCTTGACCGA CAATTGCATG AAGAATCTGC	180
	TTAGGGTTAG GCGTTTTGCG CTGCTTCGCG ATGTA CGGGC CAGATA TACG CGTTGAC ATT	240
	GATTATTGAC TAGTTATTAA TAGTAATCAA TTACG GGGTC ATTAGTTCAT AGCCCATATA	300
	TGGAGTTCCG CGTACATAA CTTA CGGTAA ATGGC CCGCC TGGCTGACCG CCCAACGACC	360
	CCCGCCCATTT GACGTCAATA ATGACGTATG TTCCC ATAGT AACGCCAATA GGGACTT TCC	420
10	ATTGACGTCA ATGGGTGGAC TATT TACGGT AAAC T GCCCA CTTGGCAGTA CATCAAGTGT	480
	ATCATATGCC AAGTACGCCC CCTATTGACG TCAATGACGG TAAATGCCCC GCCTGGCATT	540
	ATGCCCAGTA CATGACCTTA TGGGACTTTC CTACT TGGCA GTACATCTAC GTATTAGTCA	600
	TCGTATTAC CATGGTGATG CGGT TTTGGC AGTACATCAA TGGGCGTGGA TAGCGGTTTG	660
	ACTCACGGGG ATTTCCAAGT CTCCACCCCA TTGACGTCAA TGGGAGTTTG TTTTGGCACC	720
15	AAAATCAACG GGA CTTTCCA AAATGTCGTA ACAAC TCCGC CCCATTGACG CAAATGGGCG	780
	GTAGGCGTGT ACGGTGGGAG GTCTATATAA GCAGAGCTCT CTGGCTAACT AGAGAACCCA	840
	CTGCTTACTG GCTTATCGAA ATTAATACGA CTCAC TATAG GGAGACCCAA GCTGGCTAGC	900
	GTTTAAACTT AAGCTTGGTA CCGAGCTCGG ATCC CCGTCG TGCATCTATC GAAGGTCGTG	960
20	GA GAT CCC GAG GAG CCC AAA TCT TGT GAC AAA ACT CAC ACA TGC CCA	1007
	ASP PRO GLU GLU PRO LYS SER CYS ASP LYS THR HIS THR CYS PRO 15	
	CCG TGC CCA GCA CCT GAA CTC CTG GGG GGA CCG TCA GTC TTC CTC TTC	1055
25	PRO CYS PRO ALA PRO GLU LEU LEU GLY GLY PRO SER VAL PHE LEU PHE 31	
	CCC CCA AAA CCC AAG GAC ACC CTC ATG ATC TCC CGG ACC CCT GAG GTC	1103
	PRO PRO LYS PRO LYS ASP THR LEU MET ILE SER ARG THR PRO GLU VAL 47	
	ACA TGC GTG GTG GTG GAC GTG AGC CAC GAA GAC CCT GAG GTC AAG TTC	1151
30	THR CYS VAL VAL VAL ASP VAL SER HIS GLU ASP PRO GLU VAL LYS PHE 63	
	AAC TGG TAC GTG GAC GGC GTG GAG GTG CAT AAT GCC AAG ACA AAG CCG	1199
	ASN TRP TYR VAL ASP GLY VAL GLU VAL HIS ASN ALA LYS THR LYS PRO 79	
35	CGG GAG GAG CAG TAC AAC AGC ACG TAC CGG GTG GTC AGC GTC CTC ACC	1247
	ARG GLU GLU GLN TYR ASN SER THR TYR ARG VAL VAL SER VAL LEU THR 95	
	GTC CTG CAC CAG GAC TGG CTG AAT GGC AAG GAG TAC AAG TGC AAG GTC	1295
40	VAL LEU HIS GLN ASP TRP LEU ASN GLY LYS GLU TYR LYS CYS LYS VAL 111	
	TCC AAC AAA GCC CTC CCA GCC CCC ATC GAG AAA ACC ATC TCC AAA GCC	1343
	SER ASN LYS ALA LEU PRO ALA PRO ILE GLU LYS THR ILE SER LYS ALA 127	
	AAA GGG CAG CCC CGA GAA CCA CAG GTG TAC ACC CTG CCC CCA TCC CGG	1391
45	LYS GLY GLN PRO ARG GLU PRO GLN VAL TYR THR LEU PRO PRO SER ARG 143	

	GAT GAG CTG ACC AAG AAC CAG GTC AGC CTG ACC TGC CTG GTC AAA GGC	1439
	ASP GLU LEU THR LYS ASN GLN VAL SER LEU THR CYS LEU VAL LYS GLY	159
5	TTC TAT CCC AGC GAC ATC GCC GTG GAG TGG GAG AGC AAT GGG CAG CCG	1487
	PHE TYR PRO SER ASP ILE ALA VAL GLU TRP GLU SER ASN GLY GLN PRO	175
	GAG AAC AAC TAC AAG ACC ACG CCT CCC GTG CTG GAC TCC GAC GGC TCC	1535
	GLU ASN ASN TYR LYS THR THR PRO PRO VAL LEU ASP SER ASP GLY SER	191
10	TTC TTC CTC TAC AGC AAG CTC ACC GTG GAC AAG AGC AGG TGG CAG CAG	1583
	PHE PHE LEU TYR SER LYS LEU THR VAL ASP LYS SER ARG TRP GLN GLN	207
	GGG AAC GTC TTC TCA TGC TCC GTG ATG CAT GAG GCT CTG CAC AAC CAC	1631
15	GLY ASN VAL PHE SER CYS SER VAL MET HIS GLU ALA LEU HIS ASN HIS	223
	TAC ACG CAG AAG AGC CTC TCC CTG TCT CCG GGT AAA TGATCTAGAG	1677
	TYR THR GLN LYS SER LEU SER LEU SER PRO GLY LYS	235
	GGCCCCGTTTA AAC CCGCTGA TCAG CCTCGA CTGTG CCTTC TAGTTGCCAG CCATCTGTTG	1737
20	TTTGCCCCCTC CCC CGTGCCCT TCCTTGACCC TGGAA GGTGC CACTCC CACT GTCCTTT CCT	1797
	AATAAAATGA GGAAATTGCA TCGCATTTGTC TGAGTAGGTG TCATTC TATT CTGGGGG GTG	1857
	GGGTGGGGCA GGA CAGCAAG GGGGAGGATT GGGAA GACAA TAGCAGGCAT GCTGGGGATG	1917
	CGGTGGGCTC TATGGCTTCT GAGG CGGAAA GAACCACTG GGGCTC TAGG GGGTATC CCC	1977
	ACGCGCCCTG TAG CGGCGCA TTAA GCGCGG CGGGT GTGGT GGTAC GCGC AGCGTGA CCG	2037
25	CTACACTTGC CAG CGCCCTA GCGC CCGCTC CTTTC GCTTT CTTCC CTTCC TTTCTC GCCA	2097
	CGTTCGCCCG CTT TCCCCGT CAAG CTCTAA ATCGGGGCAT CCCTTT AGGG TTCCGAT TTA	2157
	GTGCTTTACG GCA CCTCGAC CCCA AAAAAC TTGAT TAGGG TGATGGTTCA CGTAGTGGC	2217
	CATCGCCCTG ATA GACGGT TTTC GCCCTT TGACGTTGGA GTCCAC GTTC TTTAATA GTG	2277
	GACTCTTGTT CCAACTGGA ACAACACTCA ACCCTATCTC GGTCTATTCT TTTGATTTAT	2337
30	AAGGGATTTT GGGGATTTTC GCCTATTGGT TAAAAAATGA GCTGATTTAA CAAAAAT TTA	2397
	ACGCGAATTA ATT CTGTGGA ATGT GTGTCA GTTAGGGTGT GGAAAGTCCC CAGGCTC CCC	2457
	AGGCAGGCAG AAG TATGCAA AGCA TGCATC TCAAT TAGTC AGCAA CCAGG TGTGGA AAGT	2517
	CCCCAGGCTC CCC AGCAGGC AGAAGTATGC AAAGCATGCA TCTCAA TTAG TCAGCAA CCA	2577
	TAGTCCCGCC CCT AACTCCG CCCA TCCCGC CCCTAACTCC GCCCAGTTCC GCCCATTCTC	2637
35	CGCCCCATGG CTGACTAATT TTTT TTATTT ATGCA GAGGC CGAGGC CGCC TCTGCCT CTG	2697
	AGCTATTCCA GAAGTAGTGA GGA GGCTTTT TTGGAGGCCT AGGCTTTTGC AAAAAGCTCC	2757
	CGGGAGCTTG TATATCCATT TTCG GATCTG ATCAG CACGT GTTGAC AATT AATCATC GGC	2817
	ATAGTATATC GGCATAGTAT AATA CGACAA GGTGA GGAAC TAAACCATGG CCAAGTTGAC	2877
	CAGTGCCGTT CCGGTGCTCA CCGC GCGCGA CGTCG CCGGA GCGGTC GAGT TCTGGAC CGA	2937
40	CCGGCTCGGG TTC TCCCGGG ACTT CGTGGA GGACGACTTC GCCGGTGTGG TCCGGGACGA	2997
	CGTGACCCTG TTCATCAGCG CCGT CCAGGA CCAGGTGGTG CCGGAC AACA CCCTGGC CTG	3057
	GGTGTGGGTG CGCGGCCTGG ACGA GCTGTA CGCCGAGTGG TCGGAGGTCG TGTCCAC GAA	3117
	CTTCCGGGAC GCC TCCGGGC CGGC CATGAC CGAGATCGGC GAGCAGCCGT GGGGGCGGGA	3177
	GTTCGCCCTG CGCGACCCGG CCGG CAACTG CGTGCACTTC GTGGCC GAGG AGCAGGACTG	3237

	ACACGTGCTA	CGA GATTTTCG	ATTC CACCGC	CGCCT TCTAT	GAAAGG TTGG	GCTTCGG AAT	3297
	CGTTTTCCGG	GAC GCCGGCT	GGAT GATCCT	CCAGC GCGGG	GATCT CATGC	TGGAGT TCTT	3357
	CGCCCACCCC	AAC TTGTTTA	TTGC AGCTTA	TAATG GTTAC	AAATAA AGCA	ATAGCAT CAC	3417
	AAATTTCA	AAT AAAGCAT	TTTT TTCACT	GCATT CTAGT	TGTGGT TTGT	CCAAACT CAT	3477
5	CAATGTATCT	TAT CATGTCT	GTAT ACCGTC	GACCT CTAGC	TAGAGC TTGG	CGTAATC ATG	3537
	GTCATAGCTG	TT TCCTGTGT	GAA ATTGTTA	TCCG CTCACA	ATTCC ACACA	ACATAC GAGC	3597
	CGGAAGCATA	AAG TGTAAG	CCTG GGGTGC	CTAAT GAGTG	AGCTAA CTCA	CATTAAT TGC	3657
	GTTCGCTCA	CTG CCCGCTT	TCCA GTCGGG	AAACC TGTCG	TGCCAG CTGC	ATTAATG AAT	3717
	CGGCCAACGC	GCG GGGAGAG	GCGG TTTGCG	TATTG GCGCG	TCTTCC GCTT	CCTCGC TCAC	3777
10	TGACTCGCTG	CGC TCGGTCG	TTGC GCTGCG	GCGAG CGGTA	TCAGCT CACT	CAAAGGC GGT	3837
	AATACGGTTA	TCC ACAGAAT	CAGG GGATAA	CGCAG GAAAG	AACATG TGAG	CAAAAGG CCA	3897
	GCAAAAAGGCC	AGG AACCGTA	AAAA GGCCGC	GTTCG TGGCG	TTTTTC CATA	GGCTCCG CCC	3957
	CCCTGACGAG	CAT CACAAAA	ATC GACGCTC	AAGT CAGAGG	TGGCG AAACC	CGACAG GACT	4017
	ATAAAGATAC	CAG GCGTTTC	CCCC TGGAG	CTCCC TCGTG	CGCTCT CCTG	TTCCGAC CCT	4077
15	GCCGCTTACC	GGA TACCTGT	CCGC CTTTCT	CCCTT CGGGA	AGCGTG GCGC	TTTCTCA ATG	4137
	CTCACGCTGT	AGG TATCTCA	GTTC GGTGTA	GGTCG TTCGC	TCCAAG CTGG	GCTGTGT GCA	4197
	CGAACCCCC	GTT CAGCCCC	ACCG CTGCGC	CTTAT CCGGT	AACATC CGTC	TTGAGTC CAA	4257
	CCCGGTAAGA	CAC GACTTAT	CGCC ACTGGC	AGCAG CCACT	GGTAAC AGGA	TTAGCAG AGC	4317
	GAGGTATGTA	GGC GGTGCTA	CAGA GTTCTT	GAAGT GGTGG	CCTAAC TACG	GCTACAC TAG	4377
20	AAGGACAGTA	TTT GGTATCT	GCGC TCTGCT	GAAGC CAGTT	ACCTTC GGAA	AAAGAGT TGG	4437
	TAGCTCTTGA	TCC GGCAAA	AAAC CACCGC	TGGTA GCGGT	GGTTTT TTTG	TTTGCAA GCA	4497
	GCAGATTACG	CGC AGAAAA	AAGG ATCTCA	AGAAG ATCCT	TTGATC TTTT	CTACGGG GTC	4557
	TGACGCTCAG	TGG AACGAAA	ACTC ACGTTA	AGGGA TTTTG	GTCATG AGAT	TATCAA AAAG	4617
	GATCTTCACC	TAG ATCCTTT	TAAA TTAAAA	ATGAA GTTTT	AAATCA ATCT	AAAGTAT ATA	4677
25	TGAGTAAACT	TGG TCTGACA	GTTA CCAATG	CTTAA TCAGT	GAGGCA CCTA	TCTCAGC GAT	4737
	CTGTCTATTT	CGT TCATCCA	TAGT TGCCTG	ACTCC CCGTC	GTGTAG ATAA	CTACGAT ACG	4797
	GGAGGGCTTA	CCA TCTGGCC	CCA GTGCTGC	AATG ATACCG	CGAGA CCCAC	GCTCAC CGGC	4857
	TCCAGATTTA	TCA GCAATAA	ACCA GCCAGC	CGGAA GGGCC	GAGCGC AGAA	GTGGTCC TGC	4917
	AACTTTATCC	GCC TCCATCC	AGTC TATTAA	TTGTT GCCGG	GAAGCT AGAG	TAAGTAG TTC	4977
30	GCCAGTTAAT	AGT TTGCGCA	ACGT TGTTCG	CATTC CTACA	GGCATC GTGG	TGTCAC GCTC	5037
	GTCGTTTGGT	ATG GCTTCAT	TCAG CTCCGG	TTCCC AACGA	TCAAGG CGAG	TTACATG ATC	5097
	CCCCATGTTG	TGC AAAAAAG	CGGT TAGCTC	CTTCG GTCCT	CCGATC GTTG	TCAGAAG TAA	5157
	GTTGGCCGCA	GTG TTATCAC	TCAT GGTAT	GGCAG CACTG	CATAAT TCTC	TTACTGT CAT	5217
	GCCATCCGTA	AGA TGCTTTT	CTG TGA CTGG	TGAG TACTCA	ACCAAGTCAT	TCTGAG AATA	5277
35	GTGTATGCGG	CGA CCGAGTT	GCTC TTGCCC	GGCGT CAATA	CGGGAT AATA	CCGCGCC ACA	5337
	TAGCAGAACT	TTA AAAGTGC	TCAT CATTGG	AAAAC GTTCT	TCGGGG CGAA	AACTCTC AAG	5397
	GATCTTACCG	CTG TTGAGAT	CCAG TTCGAT	GTAAC CCACT	CGTGCA CCCA	ACTGATC TTC	5457
	AGCATCTTTT	ACT TTCACCA	GCGT TTCTGG	GTGAG CAAAA	ACAGGA AGGC	AAAATGC CGC	5517
	AAAAAAGGGA	ATA AGGGCGA	CACG GAAATG	TTGAA TACTC	ATACTC TTCC	TTTTTCAATA	5577
40	TTATTGAAGC	ATT TATCAGG	GTTA TTGTCT	CATGA GCGGA	TACATA TTTG	AATGTAT TTA	5637

GAAAAATAAA CAAATAGGGG TTCCGCGCAC ATTTCCCCGA AAAGTGCCAC CTGACGT C 5695

SEQ ID NO:6

ASP PRO GLU GLU PRO LYS SER CYS ASP LYS THR HIS THR CYS PRO PRO 16
 CYS PRO ALA PRO GLU LEU LEU GLY GLY PRO SER VAL PHE LEU PHE PRO 32
 5 PRO LYS PRO LYS ASP THR LEU MET ILE SER ARG THR PRO GLU VAL THR 48
 CYS VAL VAL VAL ASP VAL SER HIS GLU ASP PRO GLU VAL LYS PHE ASN 64
 TRP TYR VAL ASP GLY VAL GLU VAL HIS ASN ALA LYS THR LYS PRO ARG 80
 GLU GLU GLN TYR ASN SER THR TYR ARG VAL VAL SER VAL LEU THR VAL 96
 LEU HIS GLN ASP TRP LEU ASN GLY LYS GLU TYR LYS CYS LYS VAL SER 112
 10 ASN LYS ALA LEU PRO ALA PRO ILE GLU LYS THR ILE SER LYS ALA LYS 128
 GLY GLN PRO ARG GLU PRO GLN VAL TYR THR LEU PRO PRO SER ARG ASP 144
 GLU LEU THR LYS ASN GLN VAL SER LEU THR CYS LEU VAL LYS GLY PHE 160
 TYR PRO SER ASP ILE ALA VAL GLU TRP GLU SER ASN GLY GLN PRO GLU 176
 ASN ASN TYR LYS THR THR PRO PRO VAL LEU ASP SER ASP GLY SER PHE 192
 15 PHE LEU TYR SER LYS LEU THR VAL ASP LYS SER ARG TRP GLN GLN GLY 208
 ASN VAL PHE SER CYS SER VAL MET HIS GLU ALA LEU HIS ASN HIS TYR 224
 THR GLN LYS SER LEU SER LEU SER PRO GLY LYS 235

Both FR3exFc and FR1exFc soluble receptors were demonstrated to be expressed to a high level in transiently transfected 293T cells (T-cell antigen infected human embryonic kidney
 20 293 cells). The observation that both soluble receptors remain bound to heparin-coated wells even following extensive washes led the laboratory of the present inventors to try to purify the proteins with the commercial heparin-Sepharose™ resin (Pharmacia). One hundred ml volume supernatants, harvested 48 hours post transfection with either FR3exFc or FR1exFc coding plasmids, were incubated overnight at 4°C with 1 ml heparin-Sepharose™ resin. The
 25 resin was washed and then subjected to PBS supplemented with increasing concentration of NaCl. Aliquots of each fraction were analyzed by 7.5% SDS-PAGE stained with GelCode (Pierce) demonstrating a purification profile of more than 90% homogeneity and a peak elution at 400 mM NaCl for FR3exFc (Fig. 3; T=total protein, U=unbound fraction, W=wash). In contrast, FR1exFc was hardly retained on the resin. This result was confirmed
 30 by Western analysis of the same fractions with anti-FGFR1ex antibodies demonstrating that most of FR1exFc is in the unbound fraction (not shown).

Functional analysis of FR3exFc and FR1exFc showed that both compete efficiently for FGF9 binding and stimulating FGFR3, thus, demonstrating their potential as inhibitors of FGFRs function and as a target (FR3exFc) for selecting FGFR3 inhibitory molecules.

Neutralizing effect of soluble receptors

The ability of hFR3-TDhis and FR3exFc to inhibit FGF-dependent FDCP-R3 cell proliferation was compared. Both soluble receptors inhibited FDCP-R3 cell proliferation, however, FR3exFc was about 60 times more potent than hFR3TDhis (Fig. 4; legend: ♦- FDCP-FR3²³⁻³⁷⁴TDhis on FDCP-FR3 cells + FGF9, ■-FR3exFc on FDCP-FR3 cells + FGF9, ▲ - FDCP-FR3²³⁻³⁷⁴TDhis on FDCP-FR3 cells + IL, X- FR3exFc on FDCP-FR3 cells + IL3). Neither had an effect on FDCP cells stimulated with IL3. The fact that FR3exFc is entirely in dimeric form whereas only a small proportion (1/10) of hFR3²³⁻³⁷⁴TDhis is in a dimeric form might explain, at least in part, this difference.

10 **Example 3: Screening for Antibodies**Panning and first screening of Ab Binding Characterization

The screening strategies to identify Fabs from the Human Combinatorial Antibody Library (HuCAL®), developed at MorphoSys, Munich, Germany and disclosed in WO 97/08320, US patent 6,300,064, and Knappik et al., (2000), the entire contents of which are incorporated
15 herein by reference, using soluble dimeric forms of the extracellular domain of the FGFR3 receptor are shown in Table 2.

TABLE 2**Panning Strategies**

	Panning Round 1	Panning Round 2	Panning Round 3
Screen 1	FR3-TDhis	HEK293	FR3-TDhis
Screen 2	FR3exFc captured with mouse anti-human IgG	RCJ-FR3ach	FR3exFc captured with mouse anti-human IgG
Screen 3	FR3-TDhis (Round 1 of panning 1)	RCJ-FR3ach & RCJ-FR3wt	FR3exFc Captured with mouse anti-human IgG

The screening was carried out, for example in Screen 1, by coating the wells of a 96 well
20 plate with hFR3²³⁻³⁷⁴TDhis (FR3-TDhis), panning with the bacteriophage library and

selecting the positive clones. The positive clones were then tested on HEK293 (293, human embryonic kidney) cells, expressing endogenous FGFR3. The positive clones were selected and rescreened on FR3-TDhis. Two additional similar screenings were carried out as shown in Table 2. In screen 2 the first and third pannings were carried out with the FR3exFc antigen and the second panning carried out with RCJ cells expressing a mutant (achondroplasia) form of FGFR3.

An overview of the number of initial hits and of the candidate clones is shown in Table 3.

Table 3

Overview of Screenings 1, 2 and 3 on FGFR3

	screened clones	primary hits	sequenced clones	consolidated candidate clones (ELISA & FACS)
Screen 1	1076	208	69	15 MSPRO 1-15
Screen 2	864	300	32	22 MSPRO 20-33 and 52-59
Screen 3	768	487	52	11 MSPRO 40-50

10 Sequence and Vector Data

A plasmid map and sequence (SEQ ID NO:52) of the dHLX-MH vector are presented in Fig. 28A and 28B.

Figure 29A shows the plasmid map of the phage display vector used in accordance with the present invention. Figure 29B is the corresponding sequence and restriction digest map (SEQ ID NO:53).

Figure 30 displays the polynucleotide sequences of the specific VL and VH domains of MSPRO2 (SEQ ID NO:74 and 84); MSPRO11 (SEQ ID NO:70 and 85), MSPRO12 (SEQ ID NO:75 and 89); MSPRO21 (SEQ ID NO:67 and 78); MSPRO24 (SEQ ID NO:64 AND 79); MSPRO26 (SEQ ID NO:71 AND 86); MSPRO28 (SEQ ID NO:62 AND 80); MSPRO29 (SEQ ID NO:65 AND 87); MSPRO54 (SEQ ID NO:73 AND 82); MSPRO55 (SEQ ID NO:69 AND 83); MSPRO59 (SEQ ID NO:76 AND 91). The sequences include the framework domains 1-4 and the CDR domains 1-3. SEQ ID NO:61, 63, 66, 68, and 73

denote the polynucleotide sequences of the parent VL (kappa or lambda) strands. SEQ ID NO:77, 81, 88 and 90 denote the polynucleotide sequences of the VH parent strands.

Example 4: Analysis of Fabs identified by first screening.

Specificity of Antibody recognition

- 5 The first screening yielded 15 different Fabs that specifically recognize FGFR3 *in vitro* and on the cell surface. Fourteen of these were produced and sent to ProChon for further analysis. LY6.3, an anti-lysosyme antibody, was isolated from the same library and serves as a control. ELISA analysis, according to the following protocol was carried out to determine the specificity of the isolated Fabs for FGFR3 or FGFR1.

10 **Fab-FR3/Fc Binding Assay**

- MaxiSorp ELISA plates were coated with 100 µl anti-human Fc (10 µg/ml) in bicarbonate overnight at 4°C. Wells were washed five consecutive times with a PBS solution containing 0.1% Tween 20 (PBST). The well surface was blocked with 250 µl PBST+3%BSA (blocking solution) for 1 hour at 37°C. This was followed by capturing 1 µg of FGFR/Fc for 1 hour at
15 room temperature. To assess the antibody binding to the captured FGFR/Fc, 1 µg each of the tested Fabs was incubated in 100 µl blocking solution per well 1 hour at room temperature. Wells were washed 5 times with PBST. Reaction was initiated with the addition of 100 µl of 0.8µg/ml goat anti-human Fab-HRP diluted in blocking solution, subsequently washed and detected with TMB substrate (Pierce). The absorbance was measured at 450 nm. A
20 comparison of ELISA analyses done in the laboratory of the present inventors (Prochon) and at MorphoSys is presented in the following Table 4.

25

30

TABLE 4

<u>ProChon</u>	<u>MorphoSys</u>			
	FR1/Fc	FR3/Fc	FR1/Fc	FR3/Fc
MS-PRO1	++	++	+/-	+
MS-PRO2	-	++	-	++
MS-PRO3	+	++	-	++
MS-PRO4	-	+	-	++
MS-PRO5	-	++	+/-	+
MS-PRO6	-	++	-	+
MS-PRO7	-	++	-	+
MS-PRO8	+	++	-	+
MS-PRO9	-	+/-	+/-	+
MS-PRO10	+	++	-	++
MS-PRO11	-	+/-	+	++
MS-PRO12	-	+/-	-	++
MS-PRO13	-	+/-	+/-	+
MS-PRO14	-	-	-	+
LY6.3 (control)	-	-		

In most cases, the data generated at Morphosys and in the laboratory of the present inventors are in agreement. However, some Fabs behave differently. For example, MS-PRO3 and 10 were found to be completely FGFR3 specific under Morphosys conditions. In the laboratory of the present inventors, both show considerable cross-reaction with FGFR1. The FACS analysis, done at Morphosys, supports the Prochon results for MS-PRO3 but not for MS-PRO10. Taking into account the potency and specificity of the Fabs, MS-PRO2 has the highest score according to these preliminary data.

Example 5: Affinity of Fab to FGFR3

The affinity measurements were performed by BIAcore according to the standard procedure recommended by the supplier (Pharmacia). The anti-Fc antibody was coupled via the EDC/NHS chemistry to the chip and subsequently FGFR3 was captured. The Fabs of the invention were then bound to this surface.

Table 5 shows a comparison of affinities of Fabs candidates to FGFR3 as determined by BIAcore and by FACS-scatchard.

Table 5

Comparison of Antibody Affinities to FGFR3

5 **determined by BIAcore and FACS-Scatchard**

Fab clone	BIAcore [nM]	Indirect FACS-Scatchard [nM]
MSPRO2	37 ± 10	43
MSPRO11	4 ± 2	4
MSPRO12	14 ± 2	6.5
MSPRO21	9 ± 2	0.6
MSPRO24	10 ± 2	0.3
MSPRO26	4 ± 1	1.4
MSPRO28	9 ± 0.4	0.3
MSPRO29	6 ± 4	0.4

Table 1E shows the affinity as determined by BIAcore for the Fab candidates shown in Table 5 converted into the Fab mini-antibody format, Fab-dHLX-MH, where a dimer of the Fab monomer is produced after insertion into an expression vector as a fusion protein.

Table 6 shows the results of a competition assay wherein each MSPRO Fab was bound to the FGFR3 at a concentration of 500nM or 1, 000 nM and coinjected in pairs with the other MSPRO Fabs. The (-) indicates binding to the same or nearby epitope while (+) indicates binding to different epitope. The results show that MSPRO2 and 12 bind to the same or nearby epitope while MSPRO11, 21, 24, 26, 28 and 29 bind to an epitope different from that of MSPRO2 or 12.

Example 6: Specific Neutralizing Activity of the Antibodies

A: FDCP Cell Proliferation Assay

20 The FDCP cell line is a murine immortalized, interleukin 3 (IL3) dependent cell line of myelocytic bone marrow origin, which does not express endogenous FGF Receptors (FGFR).

Upon transfection with FGFR cDNA, the FDCP cell line exhibits an FGF dose dependent proliferative response that can replace the dependence on IL3. FDCP cell lines, transfected with FGFR cDNAs can therefore be used to screen for specific inhibitors or activators of FGFR, as well as for analyzing FGFR signaling. The FDCP cell response to various ligands was quantitated by a cell proliferation assay with XTT reagent (Cell Proliferation Kit, Biological Industries Co.). The method is based on the capability of mitochondrial enzymes to reduce tetrazolium salts into soluble colored formazan compounds which can be quantitated and is indicative of cell viability. Specifically, FDCP cells expressing FGFR3IIIb, FGFR3IIIc or FGFR1 were grown in "full medium" (Iscove's Medium containing 2ml glutamine, 10% FCS, 100ug/ml penicillin, 100ug/ml streptomycin) supplemented with 5ug/ml heparin and 10ng/ml FGF9. Cells were split every 3 days and kept in culture no more than one month. One day prior to the experiment, the cells were split. Before the experiment, the cells were washed 3 times (1000 rpm, 6 min) with full medium. Later, the cells were resuspended and counted with Trypan Blue. Twenty thousand (20,000) cells /well were added to wells in a 96-well plate in 50ul in full medium containing 5 ug/ml heparin. Conditioned medium was added in an additional volume of 50ul full medium containing FGF9 at varying concentrations to a final volume of 100ul. A primary stock solution (usually 2x the higher concentration) of the antibody (or Fabs) was prepared in Iscove's+++ containing 5µg/ml heparin and 2.5ng/ml FGF9 or IL-3 (final concentration 1.25 ng/ml). Dilutions were filtered in a 0.2 µm syringe nitrocellulose filter blocked first with 1mg/ml BSA and washed then with Iscove's+++ . Aliquots of requested serial dilutions were prepared. Dilutions were kept on ice until use. 50 µl of the corresponding 2x final concentration was added to each well and the plate was incubated at 37⁰C for either 40 hours or either 64 hours. After incubation, the reaction was developed as follows: 100 µl of activator solution was added to 5 ml XTT reagent and mixed gently. 50 µl of mixture was added to each well. Optical density (OD) at 490 nm at this point gave the zero time reading. Cells were then incubated at 37⁰C for 4 hours (in the case of a 40 hour incubation) or 2 hours (in the case of a 64 hour incubation) and proliferation was measured by O.D. at 490 nm (A490).

It is noted that the assay is successful when the O.D. of untreated control growing with saturated amounts of FGF (10 and 20 ng/ml) is at least 1.3 O.D. units. Furthermore, it is noted that the background of wells with no cells should be 0.2-0.35 O.D. units and that the

O.D. absorbance of 1.25 ng/ml FGF9 should not be less than 40% of the O.D. absorbance achieved with saturated FGF 9 concentration (10 and 20 ng/ml). Specific inhibition of FGF and FGF receptor mediated proliferation should always be accompanied with lack of any inhibition of the same antibody concentration on IL-3 dependent cell proliferation.

5 The following FDCP cell lines were used:

*FDCP-C10: FDCP cells transfected with the human wild-type FGF receptor 3IIIc.

*FDCP-R3: FDCP cells transfected with the human wild-type FGF receptor 3IIIb.

*FDCP-R1: FDCP cells transfected with the human wild-type FGFR1.

10 *FDCP-F3Ach: FDCP cells infected with human FGFR3 mutated at amino acid Glycine 380 to Arginine (G380R), analogous to the most common human achondroplasia mutation.

B: Neutralizing activity

The neutralizing activity of the antibodies was measured by the aforementioned cell proliferation analysis in FDCP-FR3 and FDCP-FR1 cell lines. Increasing amounts of the indicated Fabs (MSPRO 2, 3 and 4) were added to FDCP-FR3 (closed triangle ▲(2), star *
15 (3), and circle ● (4)) or FDCP-FR1 (diamond ◆ (2), square ■ (3) and open triangle Δ(4)) grown in the presence of FGF9 (Fig. 5). Two days later, an XTT proliferation assay was performed. While none of the Fabs inhibited FDCP-FR1 cell proliferation, MSPRO2 and 3 inhibited FDCP-FR3 proliferation with a similar IC50 of about 1.0 µg/ml (Fig. 5). In contrast, MS-PRO4 had no inhibitory effect on FDCP-FR3 proliferation. These data are in
20 agreement with those generated at Morphosys. The rest of the Fabs were similarly analyzed on FDCP-FR3 expressing cells. Increasing amounts of the indicated Fabs were added to FDCP-FR3 grown in the presence of FGF9 (Fig. 6). The results of the proliferation assay done at Morphosys and at Prochon are compared in Table 6. (NA- data not available)

25

30

Table 6

	<u>Prochon</u>		<u>Morphosys</u>	
	FDCP-FR1	FDCP-FR3	FDCP-FR1	FDCP-FR3
MSPRO1	-	++	NA	NA
MSPRO2	-	++	NA	++
MSPRO3	-	++	NA	++
MSPRO4	-	-	NA	-
MSPRO5	-	+	NA	+
MSPRO6	-	-	NA	+/-
MSPRO7	-	++	NA	+
MSPRO8	-	+/-	NA	+/-
MSPRO9	-	+	NA	+
MSPRO10	-	+	NA	NA
MSPRO11	-	+++	NA	++
MSPRO12	-	+++	NA	+++
MSPRO13	-	-	NA	NA
MSPRO14	-	-	NA	NA
LY6.3	-	-	NA	NA

As shown in Table 6, there is an excellent agreement between the Prochon and Morphosys data. About half of the Fabs show considerable neutralizing activity, MSPRO12 being the most potent. Most of the inhibitory Fabs performed well in the binding assay (Table 4), with MSPRO11 and MSPRO12 being the exception to the rule, however, clearly remain good candidates to pursue. None of the Fabs (including those that crossreact with FGFR1) inhibited FGF-dependent FDCP-FR1 proliferation. In addition, FDCP-FR3 grown in the presence of IL3 were not affected by any of the Fabs.

An additional 20 new Fabs were selected from the second panning done at Morphosys. Three of these new Fabs (MSPRO52, MSPRO54 and MSPRO55) were subjected to the FDCP cell proliferation test and all were found to neutralize the receptor (Fig. 7A). Interestingly (and in

accord with MorphoSys affinity data), one Fab (MSPRO54) showed strong neutralizing activity against FGFR1 (Fig. 7B).

Example 7: Receptor Expression and Activation in RCJ Cells

RCJ cell assay

5 RCJ cells (fetal rat calvaria-derived mesenchymal cells, RCJ 3.1C5.18; Grigoriadis, 1988) were generated to express various FGF Receptors an inducible manner, in the absence of tetracycline. The RCJ-M14 line (RCJ-FR3ach) expresses FGFR3-ach380 mutant upon induction by the removal of tetracycline. The cells were incubated in low serum after which FGF was added to stimulate receptor function and signaling. The cells were lysed and the
10 receptor level, receptor activation and signaling are assessed by Western with anti-FGFR3 (Santa Cruz), anti-phospho-tyrosine (Promega), and anti-active ERK (or JNK) (Promega) respectively.

RCJ-M14 cells were grown in α -MEM supplemented with 15% fetal calf serum, 1x penicillin/streptomycin/nystatin, 1x glutamine, 600 μ g/ml neomycin, 2 μ g/ml tetracycline, 50
15 μ g/ml hygromycin B to subconfluence. The medium was aspirated off and the cells washed with trypsin, 1 ml/10 cm dish, then trypsinized with 0.5 ml/10 cm dish. The cells were resuspended in 10 ml α -MEM supplemented with 15% fetal calf serum, 1x penicillin/streptomycin/nystatin, 1x glutamine, 600 μ g/ml neomycin, and 2 μ g/ml tetracycline.

20 Sixty thousand (6×10^5) cells/well were seeded in a 6-well dish. Alternatively, twice that number may be seeded. The cells were washed thrice 24 hours later (or 8 hours later if twice the amount of cells are seeded) with 1 ml α -MEM, and then incubated with α -MEM supplemented with 15% fetal calf serum, 1x penicillin/streptomycin/nystatin, and 1x glutamine (induction medium) for 16 hours. Cells were washed thrice with 1 ml α -MEM
25 and allowed to grow for 4 additional hours in 1 ml of 0.5% exhausted serum (prepared by diluting the induction medium X30 with α -MEM).

FGF9 (1 ng/ml) was added for 5 minutes and cells are then placed on ice. The cells were washed twice with ice-cold PBS and then lysed with 0.5 ml lysis buffer. The cells are scraped into an eppendorf tube, vortexed once and placed on ice for 10 minutes. The lysate
30 was microcentrifuged 10 minutes at 4⁰C and the cleared lysate transferred into a fresh Eppendorf tube.

- The protein content was determined by Bradford or DC protein assay (Bio-Rad, cat# 500-0116 - see manufacture instructions). Total protein aliquots, supplemented with 1/5 volume of 5x sample buffer, were boiled for 5 minutes and stored at -20°C until ready to load on gel. In parallel an immunoprecipitation (IP) assay was performed, 10 µl anti- FGFR3 antibodies
- 5 were added to the rest of the lysates and incubated for 4 hours at 4°C. 40 µl protein A-Sepharose was added and incubated for 1 hour at 4°C with continuous shaking. Afterwards, the mixture was microcentrifuged 15 seconds, and the fluid was aspirated, carefully leaving a volume of ~30 µl above the beads. The beads were washed 3 times with 1 ml lysis buffer. At this step, the protease inhibitor mix is omitted from the buffer.
- 10 After the final wash, 15 µl of 5x sample buffer was added, samples were boiled 5 minutes and stored at -20°C until ready to load onto gel. Samples were loaded on 7.5% SDS-PAGE, cast on a Mini-PROTEAN II electrophoresis cell, and run at 100 V through the upper gel and at 150 V through the lower gel. Proteins were transferred onto nitrocellulose sheet using the Mini trans-blot electrophoretic transfer cell at 100 V for 75 minutes or at 15 V overnight.
- 15 The lower part of the total lysate Western blots was probed with anti-active MAPK (ERK) and the upper part is probed with anti-phosphotyrosine, both diluted 5×10^3 . The IP lysate Western blots were probed with anti-anti-phosphotyrosine (R&D Systems). Hybridization was detected by ECL following the manufacturer's instructions.
- BIAcore and proliferation analyses done at MorphoSys showed that among the new Fabs,
- 20 MS-PRO54 is highly cross reactive with FGFR1. To further test the cross reactivity of the new Fabs, RCJ cells expressing either FGFR3ach (RCJ-M14; M14 on figure 9A) FGFR3 wild type (W11 on figure 9B), FGFR1 (R1-1 on figure 9C) or FGFR2 (R2-2 on figure 9D) were incubated with increasing amount of MS-PRO54 and MS-PRO59 for 1 hour. FGF9 was added for 5 minutes and cell lysates were analyzed by Western for pERK activation (Figs.
- 25 8A-B, 9A-9D). Figure 8A shows that MSPRO2 and MSPRO12 block FGFR3 receptor activation in W11 and RCJ-FR3ach expressing cells. Furthermore MSPRO13 was able to block FGFR1 activation while none of the Fabs blocked FGFR2 activation. Figures 8B and 9A-9D show the results of several Fabs on RCJ expressing wildtype FGFR3 (8B) or the different FGFR types. MSPRO29 appeared as the best FGFR3 blocker and was also effective
- 30 in blocking FGFR1 (Fig. 9c); however, MSPRO54 was the most effective Fab against FGFR1. None of the Fabs significantly inhibited FGFR2 activity. There are only a few amino acid residues, within the third Ig domain, that are shared by FGFR3 and FR1 but not by FR2.

Making mutants at these sites should clarify their role in Fab-receptor binding. Figure 8B depicts the dose effect of MS-PRO12, 29 and 13, stimulated with FGF9 and analyzed by Western blot using anti-ERK antibodies. . MSPRO29 strongly inhibits FGFR3 activation (5ug), MSPRO12 has an inhibitory effect but at a higher concentration (50 ug).

5 **Example 9: Epitope mapping of selected Fabs**

Constructs containing cDNAs that code for segments of the extracellular domain of FGFR3 were generated (Fig. 10). D2 comprises Ig domain 2, D2-3 comprises Ig domains 2 and 3 and D1-3 comprises Ig domains 1, 2 and 3. These include pChFR3^{D2}Fc that codes for Ig-like domain 2 of FGFR3 and pChFR3^{D2,3}Fc that codes for domain 2 and 3, both as human Fc
 10 fusions. The corresponding chimeric proteins, as well as the control hFR3exFc (containing domains 1, 2 and 3) were anchored to an ELISA plate coated with α human Fc antibody. A panel of 8 best Fabs, MSPRO2, 11, 12, 21, 24, 26, 28 and 29, were added, and bound Fab was determined with HRP- α human Fab (Fig. 11). The results in Fig. 11 demonstrate that MSPRO2 (speckled bar) and MSPRO12 (hatched bar) differ from the other tested Fabs. Both
 15 bind to the Ig like domain 2 while the others require domain 3 for binding. It was then tested whether or not Fabs that belong to the second group would distinguish the FGFR3IIIc isoform from the FGFR3IIIb from. FDCP-FR3IIIb or FDCP-FR3IIIc cells were incubated in the presence of 1.25 ng/ml FGF9 with increasing doses of either MSPRO12 or MSPRO29. Ly6.3 was included as control. After 2 days in culture, cell proliferation was measured with
 20 the XTT reagent. Clearly, MSPRO29 (open triangle) was completely ineffective against the IIIb isoform (Fig. 12). In contrast, MSPRO12 (square on hatched or solid lines) was equally effective against both isoforms. These data suggest that residues that differ between the two isoform are critical for MSPRO29 (and probably also for the other Fabs in the same group) FGFR3 binding.

25 **Domains in FGFR3 recognized by the new Fabs.**

In agreement with data generated at Morphosys, MSPROs can be divided into 2 groups, one that includes Fabs that bind the FGFR3 Ig II domain (MSPRO2 and 12) and a second with members that require the Ig III domain for binding (MSPRO11, 21, 24, 26, 28, and 29). To classify the new Fabs obtained from the last screen performed at Morphosys, as well as some
 30 previously obtained Fabs, a proliferation assay of FDCP cells expressing either FR3IIIb or FR3IIIc was performed. The cells were incubated in the presence of 10 (IIIb) or 5 (IIIc)

ng/ml FGF9 with increasing doses of the indicated Fabs. After 2 days in culture, cell proliferation was measured with the XTT reagent.

In agreement with Morphosys data, MSPRO59 efficiently inhibited both FDCP-FR3IIIb (Fig. 13A) and FDCP-FR3IIIc cells (Fig. 13B) while MSPRO21, 24, 26, 28, 29 and 54 inhibited
5 FDCP-FR3IIIc proliferation only.

Example 10: Bone culture

Radiolabeled MSPRO29 was used to determine whether or not MSPRO Fabs can enter the bone growth plate.

To determine the effect of iodination on Fab activity, 50 µg of MSPRO29 was first labeled
10 with cold iodine using Pierce IodoGen coated tubes. The process was carried out either without iodine, with 0.04 mM or with 1 mM NaI. MSPRO29 was then purified through a sephadex G-50 column. The ability of the modified Fab to bind FGFR3 was determined by ELISA. MaxiSorp wells were coated with anti-human Fc. FGFR3/Fc (checkered bars) was then anchored to the wells. In parallel, a similar set of wells was left in blocking buffer only
15 (no FR3/Fc, hatched bars). The unmodified (no I) or the modified MSPRO29 (low for that labeled with 0.04 mM NaI (low) and high for that labeled at 1 mM NaI (high); 2 G-50 fractions each) were added at approximately 5 µg/well and binding was measured with anti-human Fab. Fresh MSPRO29 and buffer alone were included as controls (Fig. 14)..

MSPRO29 labeled in the presence of 0.04 mM NaI showed equal binding to the receptor as
20 compared to the control unmodified Fab MSPRO29 labeled in the presence of 1 mM NaI (high I) also bound the receptor, however, the noise level of this sample was as high as the signal itself suggesting that at the high Iodide concentration the Fab was inactivated.

The neutralizing activity of the modified Fab was tested in a proliferation assay using FDCP-FR3 (C10) (Fig. 15). FDCP-FR3 (C10) cells were treated with the indicated amount of
25 labeled or unlabeled (without I) MSPRO29. The proliferation rate of the cells was determined by XTT analysis. The Fab was labeled at either 0.04 mM (Low) or 1 mM NaI (High). Two G-50 fraction (I and II) were analyzed. Fresh MSPRO29 and buffer alone (mock) were included as controls.

This experiment showed that MSPRO29, labeled at 0.04 mM NaI, kept its activity almost
30 entirely while that labeled at 1 mM NaI lost its activity completely. MS-PRO29 was labeled with 1 mCi ¹²⁵I. The specific activity of the Fab was 17 µCi/µg.

Ex vivo distribution of ^{125}I MSPRO29 in bone culture

Femora prepared from newborn mice were incubated with $2\text{ }\mu\text{g}$ ^{125}I -MSPRO29 ($17\text{ }\mu\text{Ci}/\mu\text{g}$) or ^{125}I -Ly6.3 ($20\text{ }\mu\text{Ci}/\mu\text{g}$) for 1, 3 or 5 days in culture. Then, sections were processed for radiomicroscopy. After 3 days in culture, MSPRO29 was predominantly visualized at the higher hypertrophic zone and to a lesser extent at the secondary ossification region (Figs. 16A-16F). Hematoxylin-eosin staining of growth plate treated with radiolabelled MS-PRO29 or Ly6.3 (Figs. 16A and 16D, respectively) x100 magnification. Radiomicroscopic sections of growth plate treated with radiolabelled MS-PRO29 or Ly6.3 (Figs. 16B and 16E) at X100 magnification. Figs. 16C and 16F are the same as Figs. 16B and 16E but at x400 magnification. The arrow in figure 16C indicates the location of the specific binding of the radiolabelled MS-PRO29 to the higher hypertrophic zone of the growth plate.

As compared to MSPRO29, the control Ly6.3 Fab was weakly and evenly distributed throughout the whole growth plate. At day 1 in culture, the signal was weaker but with similar distribution pattern. This distribution also holds at 5 days in culture with a less favorable signal to noise ratio (data not shown). This clearly demonstrates that MSPRO29 binds FGFR3 in our target organ.

Example 11: Neutralizing Activity on Constitutively Activating Receptors

The inhibitory activity of MSPRO antibodies on ligand-dependent and ligand-independent FDCP proliferation expressing FGFR3 Achondroplasia mutation was tested.

A proliferation assay was carried out using FDCP-FR3wt (C10) or FDCP-FR3ach cells incubated with 1.25 or 5 ng/ml FGF9 respectively and with increasing amounts of MSPRO54 or MSPRO59. As shown in Fig. 17, both MSPRO54 (diamond) and 59 (square) antibodies neutralize the mutant receptor. Few of the FDCP-FR3ach acquired ligand independent cell proliferation due to the high expression of the FGFR3ach mutation.

FDCP cells that express the achondroplasia FGFR3 (FDCP-FR3ach) and proliferate independently of ligand were incubated with the indicated amount of MSPRO12, 29, 59 or the control Ly6.3. Two days later, cell proliferation was determined by an XTT analysis. When inhibition of cell proliferation by the MS-PRO 12, 29, 54 and 59 were tested, only the antibodies 12 and 59 (the only Ab which recognized D2 domain) inhibited the ligand-independent cell proliferation (Figs. 18A and 18B). Previously, the activity of MSPRO Fabs generated in the first and second screens (MSPRO1-15 and MSPRO21-31, respectively) by XTT analysis of FDCP-FR3ach cells were tested. These cells, when generated, show ligand-

dependent proliferation. With time, however, they acquired a ligand-independent ability to proliferate. Accordingly, neutralizing Fabs were able to block the ligand-dependent, but not the ligand-independent, proliferation of these cells. To show whether this is also true for the new batch of Fabs, FDCP-FR3ach cells, which is the FDCP-derived cell line that expresses a constitutive FGFR3-G380R (Ach), were subjected to XTT analysis in the presence of MSPRO59 and MSPRO29. Surprisingly, and in contrast to the ineffective MSPRO29 (triangle), MSPRO59 (diamond) completely blocked cell proliferation (Fig. 18B). Whether other Fabs that, like MSPRO59, bind to the second Ig like domain would also inhibit FDCP-FR3ach cell proliferation was tested next. Indeed, it was found that MSPRO12 strongly inhibits the constitutive cell proliferation. However, the third member in this family, MSPRO2, had no effect on either the constitutive or the ligand-dependent cell growth, suggesting that the Fab may have lost its neutralizing activity (not shown).

Example 12: RCS Chondrocyte Culture

Effect of Fabs on growth arrest of RCS Chondrocytes

RCS is a rat chondrosarcoma derived cell line expressing preferentially high levels of FGFR2 and FGFR3 and low levels of FGFR1 (Sahni, 1999). In this cell line FGFR functions as an inhibitor of cell proliferation similar to its expected role in the achondroplasia phenotype. Analysis of RCS cell proliferation mediated by the addition of different molecules of the invention, showed that MSPRO54 and MSPRO59 were able to restore cell proliferation. The screening was performed on RCS parental cells in 96 wells plates. Cells were seeded at a concentration of 2,000 cells/well. The following day 10ng/ml FGF-9 and 5µg/ml heparin were added to the cells. 50ug/ml of the antibodies were added. Positive and negative controls for cell proliferation are included in this assay at the same concentrations as the tested molecules. On the fourth day of incubation, plates were observed under the microscope. If all cells were viable, no quantitative assay to measure the effect of the variants was performed. If cell death was observed, the Cy-Quant assay kit is used to measure the amount of the cells. The results are measured in a fluoro ELISA reader. Figure 19 shows the ELISA results in bar graph form. Untreated cells are shown speckled, ligand treated cells are shown in gray, control antibody (LY6.3)treated cells are in black while MSPRO54 and MSPRO59 treated cells are shown in hatched or checkered bars, respectively.

Example 13: Ex vivo Bone Culture

The femoral bone cultures were performed by excising the hind limbs of 369-mice, heterozygous or homozygous mice for the achondroplasia G369C mutation (age P0). The limbs were carefully cleaned up from the surrounding tissue (skin and muscles) and the femora exposed. The femora were removed and further cleared from tissue remains and ligaments. The femora were measured for their initial length, using a binocular with an eyepiece micrometer ruler. The bones were grown in 1 ml of medium in a 24 well tissue culture dish. The growing medium is α -MEM supplemented with penicillin (100 units/ml), streptomycin (0.1 mg/ml) and nystatin (12.5 units/ml). In addition, the medium contains BSA (0.2%), α -glycerophosphate (1 mM) and freshly prepared ascorbic acid (50 μ g/ml). The bones were cultured for 15 days. Measurements of bone length and medium replacement were performed every three days.

At the end of the experiment, the growth rate of the bones was determined. The growth rate of bones is calculated from the slope of a linear regression fit on the length measurements obtained from day 3 to 9.

The results shown in Fig. 20 demonstrate a dose dependent increase in the growth rate of bones treated with MS-PRO 59 in comparison to non-relevant control LY6.3 Fab. The LY6.3-treated control femurs, marked with a circle, grew at the slowest rate. The MSPRO59 treated femurs exhibited a higher growth rate, with the optimal rate achieved at MSPRO59 concentration of 100ug/ml (square) while the higher cocentration (400ug/ml, triangle) showed inhibition. Moreover, the growth rates achieved by 400 microgram/ml of MSPRO59 doubled in comparison to the control Ab (3.55 U/day as compared to 1.88 U/day, respectively). This experiment shows the neutralizing effect of the MSPRO59 antibody on constitutively active FGFR3, in an *ex vivo* model.

Example 14: In-vivo trials

FDCP-FR3ach cells, but not FDCP (control) cells, were found to be tumorigenic when injected into nude mice. Each of 9 mice received two sub-cutaneous injections with different amount of transfected cells. Fourteen days after injection, progressively growing tumors started to appear at the site of FDCP-FR3ach injection but not at the FDCP site of injection. External examination of the tumors showed a high vascular capsule. ¹²⁵I-labeled MSPRO59 and LY6.3 were injected I.P. into nude mice carrying the FDCP-FR3ach derived tumor. The

tumors were dissected 4 and 24 hrs later and radioactivity was measured. Concentration of MSPRO59 Abs in FDCP-FR3ach derived tumors is shown in Fig. 22.

Example 15: Animal Model for Bladder Carcinoma

- 5 Recent studies have shown that the IIIb isoform of FGFR3 is the only form expressed in bladder carcinoma, in particular an FGFR3 with an amino acid substitution wherein Serine 249 is replaced by Cysteine (S249C). The progression of the cancer is believed to be a result of the constitutive activation resulting from this amino acid substitution. In order to create the FGFR3 IIIb form, we isolated the IIIb region of FGFR3 from HeLa cells and generated a full length FGFR3IIIb isoform in
10 pLXSN. Retroviruses, expressing either normal FGFR3 (FR3wt) or mutant FGFR3 (FR3-S249C) were produced and used to infect FDCP cells. Stable pools were generated and further used for *in-vitro* and *in-vivo* experiments.

A. MSPRO59 reduces tumor size in mice

- Twelve nude mice were injected with 2×10^6 FDCP-S249C cells subcutaneous at 2 locations, one on
15 each flank. A week later MSPRO59 was administered i.p. at 400ug per mouse (3 mice in total), followed by 3 injections of 275 ug each, in 2 to 3 days intervals. Following 24 and 26 days the tumor size was measures. Figure 23 shows the inhibitory effect of MSPRO59 on tumor size.

B. Treating FDCP-S249C-derived tumors with MSPRO59

- Nude mice (3 in each group), were injected subcutaneous at 2 locations, one on each flank,
20 with 2×10^6 FDCP-S249C cells each. A week later, 400 or 80 μ g MSPRO59 were injected IP. Three days later, mice were injected with 400 μ g followed by 5 additional injections with 275 μ g MSPRO59, each, every 3 or 4 days. Mice initially treated with 80 μ g MSPRO59 were similarly given an additional 80 μ g MSPRO59 followed by 5 injections with 50 μ g MSPRO59 at the same schedule. Mice injected with PBS were used as control. Tumors
25 typically appeared three weeks post injection of the cells. Tumor volume was estimated from measurements in 3 dimensions at 16,20, 23 or 32 days post cell injection.

As shown in Figure 24 there is both a delay in tumor appearance and an inhibitory effect on tumor progression in the treated mice. This indicates that these FGFR3 inhibitors are potent *in-vivo*.

- 30 These data may also help us understand the mechanism by which the S249C-derived tumors were developed. Since we are using pools of cells, treatment with MSPRO59 inhibited the

susceptible cells, leading to delay in tumor appearance. However, over time, the resistant cells survived and proliferated, giving rise to a solid tumor.

C. MSPRO59 inhibits FDCP-FR3ach380 derived tumor growth.

Nude mice were injected subcutaneously in the flank with 2×10^6 FDCP-FR3ach380 cells, each. Treatment with MSPRO59 began at the day of tumor appearance. Three mice were treated with a known tyrosine kinase inhibitor (TKI -50 mg/Kg/injection) and three with 400 μ g followed by 3 additional injections with 300 μ g MSPRO59, every 3 or 4 days. Three mice were treated with PBS alone as control. The tumor size was estimated as before at the indicated days after cell injection. The dose schedule is shown in Table 7 below.

10

Table 7

	Days After FDCP-FR3 ^{ach380} Cell Injection			
	21	25	28	31
MSPRO59 (μ g)	400 μ g	300 μ g	300 μ g	300 μ g
PBS (μ l)	50	50	50	50

Results are shown in bar graph format in Figure 25A.

D. MSPRO59 inhibits FDCP-S249C induced tumor growth

To overcome the instability of the FDCP-derived pools, clones from each pool FDCP-S249C clone #2) were isolated and characterized. These clones were tested in an XTT proliferation assay and were shown to be inhibited by MSPRO59. 2×10^6 cells from each clone were injected into nude mice. Tumors appeared 18-30 after injection.

FDCP-S249C clone #2 was injected subcutaneously on the flank. A week later mice were injected with 280 μ g MSPRO59 single chain (SC) I.P. every day. Mice injected with PBS were used as control. Tumor volume was estimated from measurements in 3 dimensions at 18 or 24 days post cell injection. An apparent inhibition of tumor growth by MSPRO59(SC) was observed in tumors derived from clone 2 (figure 26). Figure 25B shows the inhibition effected by MSPRO59scFv and MSPRO59 Fab compared to the control. Both inhibit growth of the tumor resulting from constitutively activated cells.

Having now fully described this invention, it will be appreciated by those skilled in the art that the same can be performed within a wide range of equivalent parameters, concentrations, and conditions without departing from the spirit and scope of the invention and without undue experimentation.

While this invention has been described in connection with specific embodiments thereof, it will be understood that it is capable of further modifications. This application is intended to cover any variations, uses, or adaptations of the inventions following, in general, the principles of the invention and including such departures from the present disclosure as come
5 within known or customary practice within the art to which the invention pertains and as may be applied to the essential features hereinbefore set forth as follows in the scope of the appended claims.

All references cited herein, including journal articles or abstracts, published or corresponding U.S. or foreign patent applications, issued U.S. or foreign patents, or any other references, are
10 entirely incorporated by reference herein, including all data, tables, figures, and text presented in the cited references. Additionally, the entire contents of the references cited within the references cited herein are also entirely incorporated by references.

Reference to known method steps, conventional methods steps, known methods or conventional methods is not in any way an admission that any aspect, description or
15 embodiment of the present invention is disclosed, taught or suggested in the relevant art.

The foregoing description of the specific embodiments will so fully reveal the general nature of the invention that others can, by applying knowledge within the skill of the art (including the contents of the references cited herein), readily modify and/or adapt for various applications such specific embodiments, without undue experimentation, without departing
20 from the general concept of the present invention. Therefore, such adaptations and modifications are intended to be within the meaning and range of equivalents of the disclosed embodiments, based on the teaching and guidance presented herein. It is to be understood that the phraseology or terminology herein is for the purpose of description and not of limitation, such that the terminology or phraseology of the present specification is to be
25 interpreted by the skilled artisan in light of the teachings and guidance presented herein, in combination with the knowledge of one of ordinary skill in the art.

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15

CLAIMS

1. A molecule comprising the antigen binding portion of an isolated antibody which has specific binding affinity for a receptor protein tyrosine kinase and which blocks constitutive activation of said receptor protein tyrosine kinase.
- 5 2. The molecule according to claim 1, wherein said molecule binds to the extracellular domain of the receptor protein tyrosine kinase.
3. The molecule according to claim 1 wherein the antibody binds the dimeric form of the receptor.
4. The molecule according to claim 1, wherein the receptor protein tyrosine kinase is
10 selected from the group consisting of EGFR/ErbB1, ErbB2/HER2/Neu, ErbB/HER3, ErbB4/HER4, IGF-1R, PDGFR- α , PDGFR- β , CSF-1R, kit/SCFR, Flk2/FH3, Flk1/VEGFR1, Flk1/VEGFR2, Flt4/VEGFR3, FGFR1, FGFR2/K-SAM, FGFR3, FGFR4, TrkA, TrkC, HGFR, RON, EphA2, EphB2, EphB4, Axl, TIE/TIE1, Tek/TIE2, Ret, ROS, Alk, Ryk, DDR, LTK and MUSK, and
15 heterodimeric combinations thereof.
5. The molecule according to claim 4, wherein said receptor protein tyrosine kinase is a fibroblast growth factor receptor (FGFR).
6. The molecule according to claim 5, wherein said FGFR is FGFR3.
7. A pharmaceutical composition, comprising, as an active ingredient, the molecule
20 according to any one of claims 1 through 6 and a pharmaceutically acceptable carrier, excipient, or auxiliary agent.
8. A molecule comprising the antigen-binding portion of an antibody which has specific binding affinity for a fibroblast growth factor receptor (FGFR) and which blocks ligand-dependent activation of said FGFR.
9. The molecule according to claim 8, wherein said molecule binds to the extracellular
25 domain of the FGFR.
10. The molecule according to claim 9, wherein the FGFR is FGFR3.
11. A pharmaceutical composition, comprising the molecule according to any one of claims 8-10 and a pharmaceutically acceptable carrier, excipient, or auxiliary agent.

12. A kit comprising a molecule of any one of claims 1-6 and 8-10 and at least one reagent suitable for detecting the presence of said molecule when bound to said receptor protein tyrosine kinase and instructions for use.
- 5 13. A method for treatment of bone and cartilage related disorders, comprising administering a therapeutically effective amount of the pharmaceutical composition according to claim 7 or 11 to a subject in need thereof.
14. The method according to claim 13 wherein the skeletal disorder is a skeletal dysplasia or a craniosynostosis disorder.
- 10 15. The method according to claim 14 wherein said craniosynostosis disorder is Muenke coronal craniosynostosis or Crouzon syndrome with acanthosis nigricans.
16. The method according to claim 13 wherein the skeletal dysplasia is selected from achondroplasia, thanatophoric dysplasia (TD), hypochondroplasia, severe achondroplasia with developmental delay and acanthosis nigricans (SADDAN) dysplasia.
- 15 17. The method according to claim 16, wherein the skeletal dysplasia is achondroplasia.
18. The method according to claim 13 for treating or inhibiting a malignant cell proliferative disease or disorder associated with abnormal RPTK activity.
19. The method according to claim 18 wherein the malignant cell proliferative disease or disorder is a hematopoietic malignancy.
- 20 20. The method according to claim 19 wherein the hematopoietic malignancy is multiple myeloma.
21. The method according to claim 18 for the treatment or inhibition of solid tumors.
22. The method according to claim 21 wherein the solid tumors are selected from
25 mammary, colon, cervical, bladder, colorectal, chondrosarcoma or osteosarcoma.
23. The method according to claim 18 for treating or inhibiting tumor formation, primary tumors, tumor progression or tumor metastasis.
24. The method according to claim 23 wherein tumor progression is the progression of transitional cell carcinoma.

25. The method according to claim 19 wherein the disorder is associated with the action of a constitutively activated receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 7.
- 5 26. The method according to claim 19, wherein the disorder is associated with ligand-dependent activation of a receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 11.
- 10 27. The method according to claim 18 for treatment of hyperproliferative diseases and disorders associated with ligand dependent fibroblast growth factor receptor signaling.
28. The method according to claim 27 wherein the hyperproliferative diseases and disorders are vision disorders such as neovascular glaucoma, macular degeneration and proliferative retinopathy including diabetic retinopathy.
- 15 29. The method according to claim 27 wherein the hyperproliferative diseases are non-neoplastic angiogenic pathologic conditions such as hemangiomas, angiofibromas and psoriasis
- 20 30. The method according to claim 18, wherein the disorder is associated with constitutive activation of a receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 7.
- 25 31. The method according to claim 18, wherein the disorder is associated with ligand-dependent activation of a receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 11.
32. A method for treating or inhibiting a cell proliferative disease or disorder, comprising administering a therapeutically effective amount of the pharmaceutical composition according to claim 7 or 11 to a subject in need thereof.
- 30 33. The method according to claim 19, wherein the cell proliferative disease or disorder is tumor progression.

34. The method according to claim 20, wherein the tumor progression is the progression of transitional cell carcinoma.
35. The method according to claim 20, wherein the tumor progression is the progression of osteo or chondrosarcoma.
- 5 36. The method according to claim 20, wherein the tumor progression is the progression of multiple myeloma.
37. The method according to claim 19 wherein the receptor protein tyrosine kinase is FGFR3 and the tumor progression is the progression of mammary carcinoma.
- 10 38. A method for screening a molecule comprising the antigen-binding portion of an antibody which blocks ligand-dependent activation of a receptor protein tyrosine kinase, comprising:
screening a library of antibody fragments for binding to a dimeric form of a receptor protein tyrosine kinase;
identifying an antibody fragment which binds to the dimeric form of the receptor
15 protein tyrosine kinase as a candidate molecule for blocking ligand-dependent activation of the receptor protein tyrosine kinase;
and determining whether or not the candidate molecule can block ligand-dependent activation of the receptor protein tyrosine kinase in a cell.
- 20 39. The method according to claim 38, wherein the receptor protein tyrosine kinase is a fibroblast growth factor receptor
40. The method according to claim 39, wherein the fibroblast growth factor receptor is FGFR3.
- 25 41. A molecule according to claim 1 comprising V_H-CDR3 and V_L-CDR3 regions, selected from the group consisting of SEQ ID NO: 8 and 9; SEQ ID NO: 12 and 13; and SEQ ID NO: 24 and 25.
42. The molecule according to claim 41, comprising V_L region and V_H regions, selected from the group consisting of SEQ ID NO: 92 and 103; SEQ ID NO: 94 and 105 and SEQ ID NO: 102 and 113.
- 30 43. A pharmaceutical composition, comprising, as an active ingredient, the molecule according to any one of claims 41 or 42 and a pharmaceutically acceptable carrier, excipient, or auxiliary agent.

44. An isolated nucleic acid molecule, comprising a sequence selected from SEQ ID NO: 30, 31, 34, 35, 50 or 51 or a nucleotide sequence hybridizing under high stringency conditions thereto.
- 5 45. An isolated nucleic acid molecule, comprising a sequence selected from SEQ ID NO: 74, 75, 76, 84, 89 or 91 and 87 or a nucleotide sequence hybridizing under high stringency conditions thereto.
- 10 46. The isolated nucleic acid molecule of claim 44, comprising nucleotides encoding a V_L -CDR3 DNA region and a V_H -CDR3 DNA region, respectively, selected from the group consisting of SEQ ID NO: 30 and 31; SEQ ID NO:34 and 35; SEQ ID NO: 50 and 51.
47. The isolated nucleic acid molecule of claim 45 comprising nucleotides encoding a V_L region and a V_H region, respectively, selected from the group consisting of SEQ ID NO: 74 and 84 ; SEQ ID NO:75 and 89; and SEQ ID NO: 76 and 91.
48. A vector comprising a nucleic acid molecule according to claim 46 or 47.
- 15 49. The vector according to claim 48 wherein the vector is an expression vector.
50. The host cell transformed with the vector according to claim 48 or 49.
51. A molecule according to claim 8 comprising the combination of V_H -CDR3 and V_L -CDR3 amino acid sequences selected from the group consisting of SEQ ID NO:10 and SEQ ID NO:11; SEQ ID NO:14 and SEQ ID NO:15; SEQ ID NO:16 and SEQ ID NO:17; SEQ ID NO:18 and SEQ ID NO:19; SEQ ID NO:20 and SEQ ID NO:21; SEQ ID NO:22 and SEQ ID NO:23, SEQ ID NO:26 and SEQ ID NO:27 or SEQ ID NO:28 and SEQ ID NO:29.
- 20 52. The molecule according to claim 51, comprising a V_L region and a V_H region, respectively, selected from the group consisting of respectively, selected from the group consisting of SEQ ID NO: 92 and 103; SEQ ID NO: 93 and 104; SEQ ID NO: 94 and 105; SEQ ID NO:95 and 106; SEQ ID NO: 96 and 107 ; SEQ ID NO: 97 and 108; SEQ ID NO:98 and 109; SEQ ID NO: 99 and 110; SEQ ID NO: 100 and 111; SEQ ID NO: 101 and 112; and SEQ ID NO:102 and 113.
- 25 53. A pharmaceutical composition, comprising the molecule according to any one of claims 51-52 and a pharmaceutically acceptable carrier, excipient, or auxiliary agent.
- 30

54. An isolated nucleic acid molecule, comprising SEQ ID NO: NO:32, 33, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 48, and 49 or a nucleotide sequence hybridizing under high stringency conditions thereto.
55. An isolated nucleic acid molecule, comprising SEQ ID NO: 62, 64, 65, 67, 69, 70, 76, 78, 79, 80, 83, 85, 86, and 87 or a nucleotide sequence hybridizing under high stringency conditions thereto.
56. An isolated nucleic acid molecule, comprising nucleotides encoding a V_L-CDR3 DNA region and a V_H-CDR3 DNA region, respectively, selected from the group consisting of SEQ ID NO: 32 and 33; SEQ ID NO:36 and 37; SEQ ID NO: 38 and 39, SEQ ID NO:40 and 41, SEQ ID NO: 42 and 43, SEQ ID NO: 44 and 45, SEQ ID NO: 48 and 49.
57. An isolated nucleic acid molecule, comprising nucleotides encoding a V_L region and a V_H region, respectively, selected from the group consisting of SEQ ID NO: 70 and 85; SEQ ID NO:67 and 78; SEQ ID NO:64 and 79; SEQ ID NO:71 and 86; SEQ ID NO:62 and 80; SEQ ID NO:65 and 87; SEQ ID NO:69 and 83.
58. A vector comprising a nucleic acid molecule according to claim 56 or 57.
59. The vector according to claim 58 which is an expression vector.
60. The host cell transformed with the vector according claim 58 or 59.
61. A method for treatment of bone and cartilage related disorders, comprising administering a therapeutically effective amount of the pharmaceutical composition according to claim 43 or 53 to a subject in need thereof.
62. The method according to claim 61 wherein the skeletal disorder is a skeletal dysplasia or a craniosynostosis disorder.
63. The method according to claim 62 wherein said craniosynostosis disorder is Muenke coronal craniosynostosis or Crouzon syndrome with acanthosis nigricans.
64. The method according to claim 63 wherein the skeletal dysplasia is selected from achondroplasia, thanatophoric dysplasia (TD), hypochondroplasia, severe achondroplasia with developmental delay and acanthosis nigricans (SADDAN) dysplasia.
65. The method according to claim 64, wherein the skeletal dysplasia is achondroplasia.

66. The method according to claim 61 for treating or inhibiting a malignant cell proliferative disease or disorder associated with abnormal RPTK activity.
67. The method according to claim 66 wherein the malignant cell proliferative disease or disorder is a hematopoietic malignancy.
- 5 68. The method according to claim 67 wherein the hematopoietic malignancy is multiple myeloma.
69. The method according to claim 61 for the treatment or inhibition of solid tumors.
70. The method according to claim 69 wherein the solid tumors are selected from mammary, colon, cervical, bladder, colorectal, chondrosarcoma or osteosarcoma.
- 10 71. The method according to claim 61 for treating or inhibiting tumor formation, primary tumors, tumor progression or tumor metastasis.
72. The method according to claim 71 wherein tumor progression is the progression of transitional cell carcinoma.
73. The method according to claim 61 wherein the disorder is associated with the
15 action of a constitutively activated receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 43.
74. The method according to claim 61, wherein the disorder is associated with ligand-
dependent activation of a receptor protein tyrosine kinase, and wherein the
20 administered pharmaceutical composition is the pharmaceutical composition according to claim 53.
75. The method according to claim 74 for treatment of hyperproliferative diseases and disorders associated with ligand dependent fibroblast growth factor receptor signaling.
- 25 76. The method according to claim 75 wherein the hyperproliferative diseases and disorders are vision disorders such as neovascular glaucoma, macular degeneration and proliferative retinopathy including diabetic retinopathy.
77. The method according to claim 75 wherein the hyperproliferative diseases
are non-neoplastic angiogenic pathologic conditions such as hemangiomas,
30 angiofibromas and psoriasis

78. The method according to claim 73, wherein the disorder is associated with constitutive activation of a receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 43.
- 5 79. The method according to claim 74, wherein the disorder is associated with ligand-dependent activation of a receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 53.
- 10 80. A method for treating or inhibiting a cell proliferative disease or disorder, comprising administering a therapeutically effective amount of the pharmaceutical composition according to claim 43 or 53 to a subject in need thereof.
81. The method according to claim 80, wherein the cell proliferative disease or disorder is tumor progression.
- 15 82. The method according to claim 81, wherein the tumor progression is the progression of transitional cell carcinoma.
83. The method according to claim 81, wherein the tumor progression is the progression of osteo or chondrosarcoma.
84. The method according to claim 81, wherein the tumor progression is the progression of multiple myeloma.
- 20 85. The method according to claim 81 wherein the receptor protein tyrosine kinase is FGFR3 and the tumor progression is the progression of mammary carcinoma.

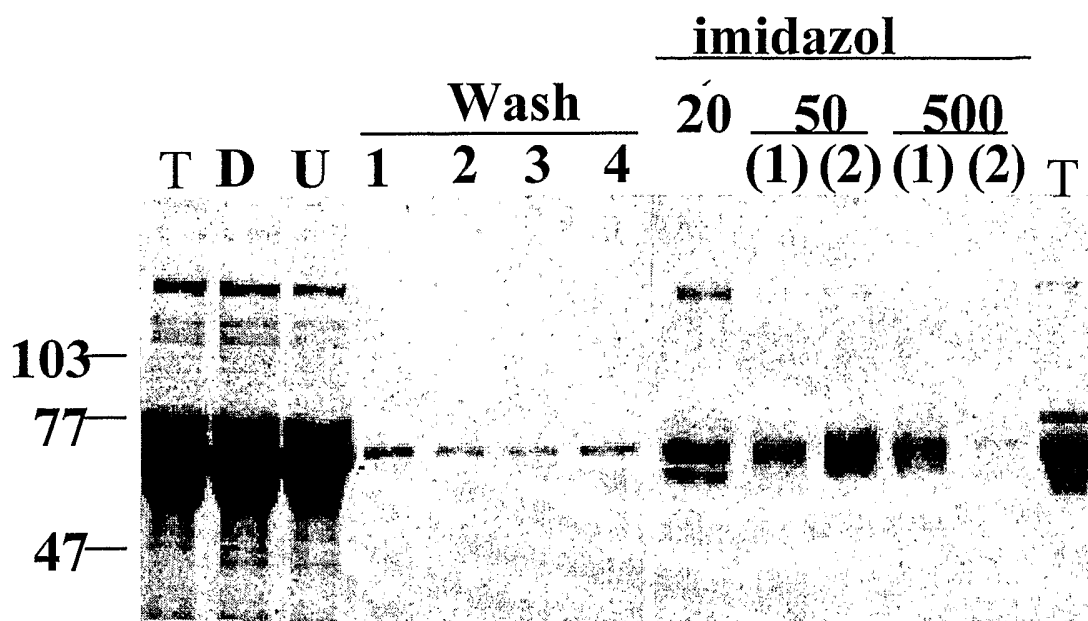


Figure 1

Figure 2

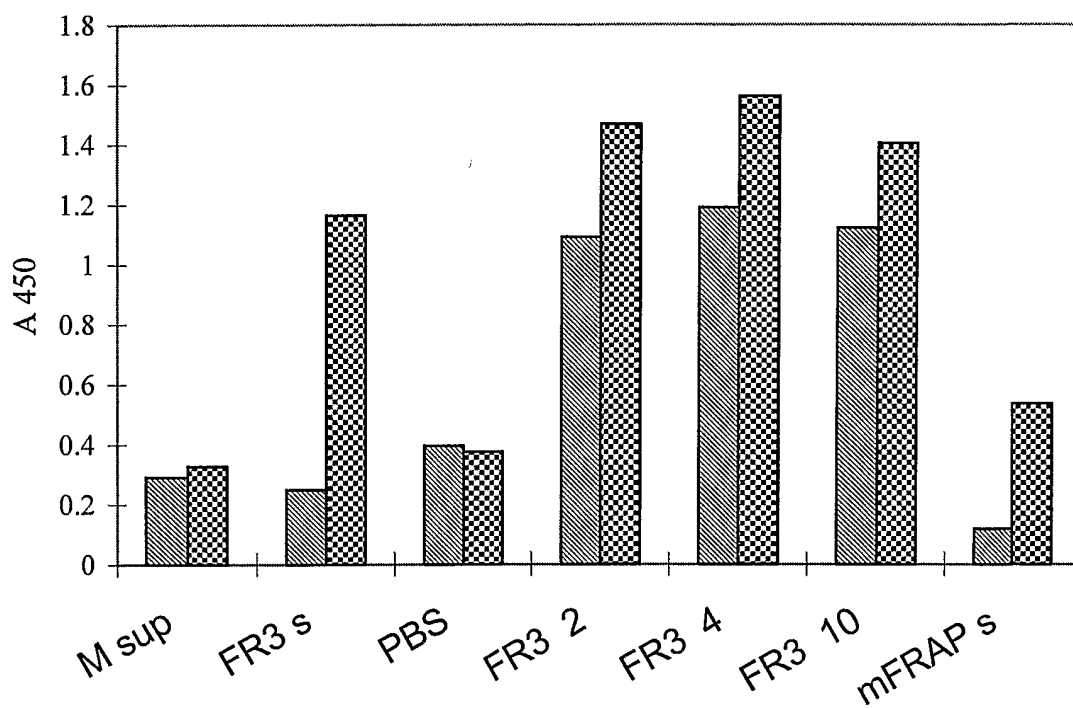


Figure 3

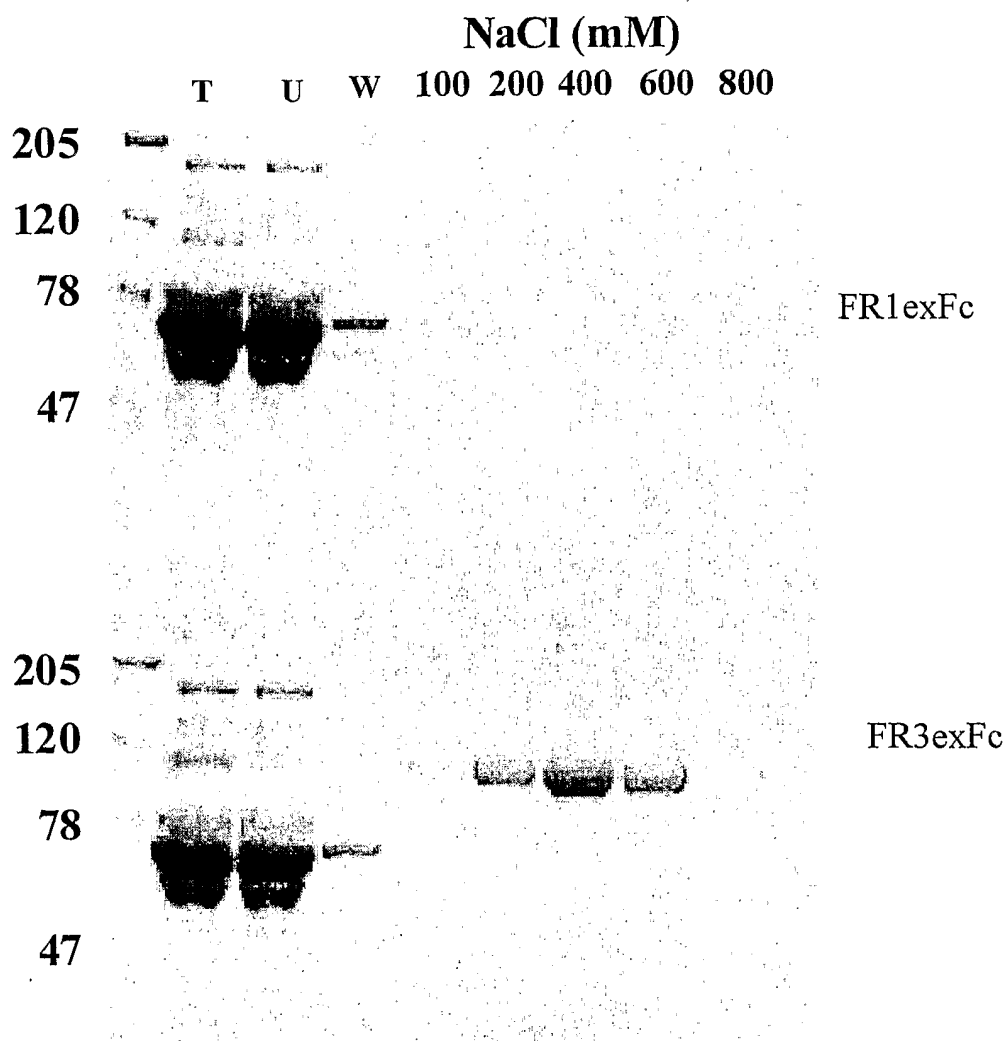


Figure 4

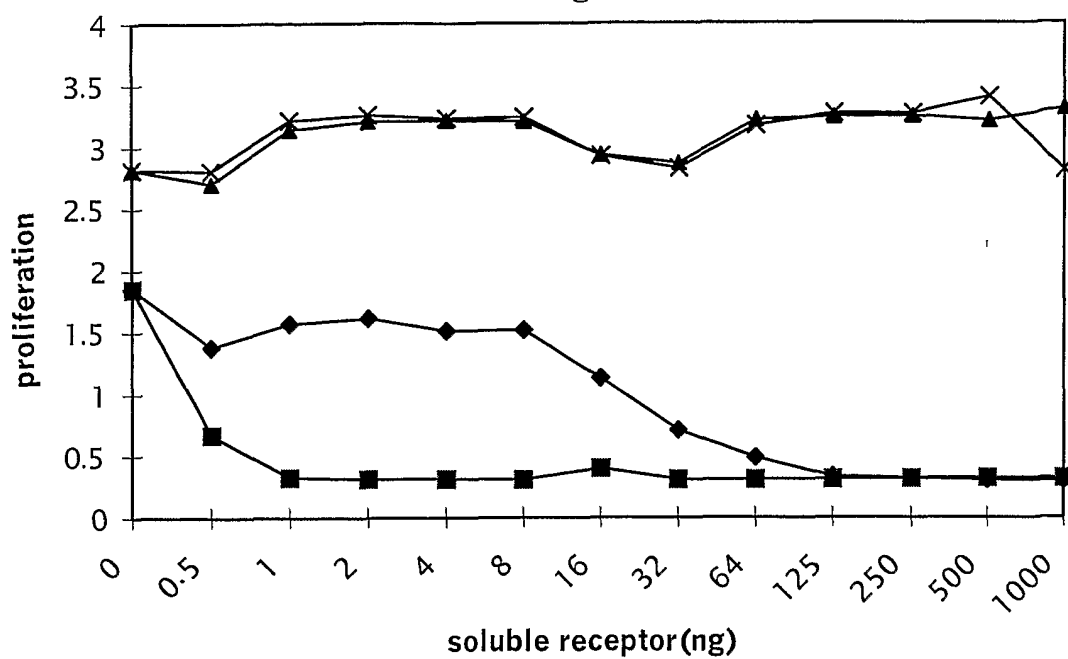


Figure 5

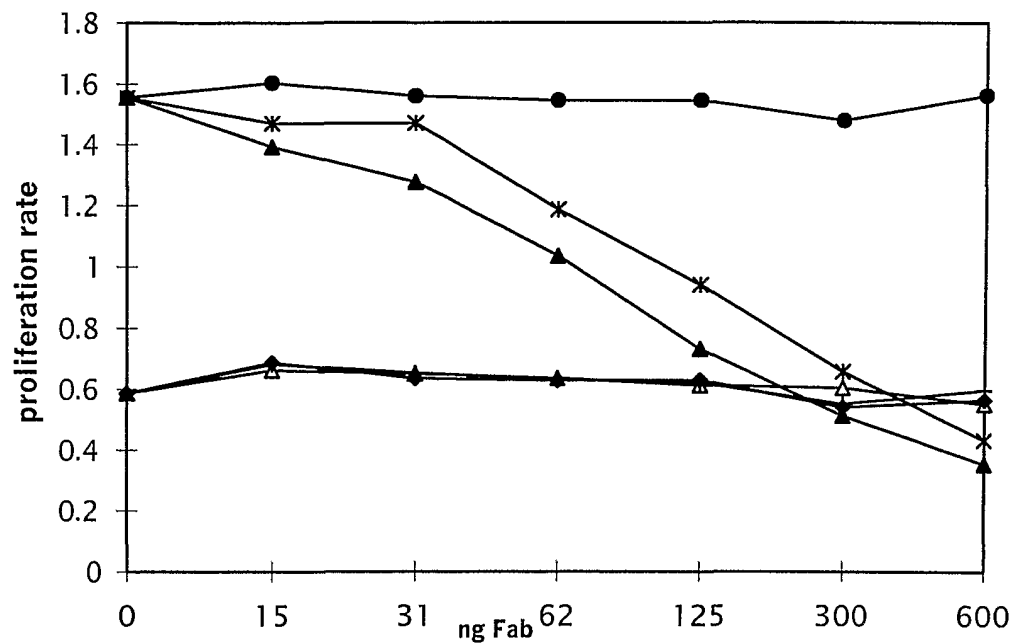


Figure 6

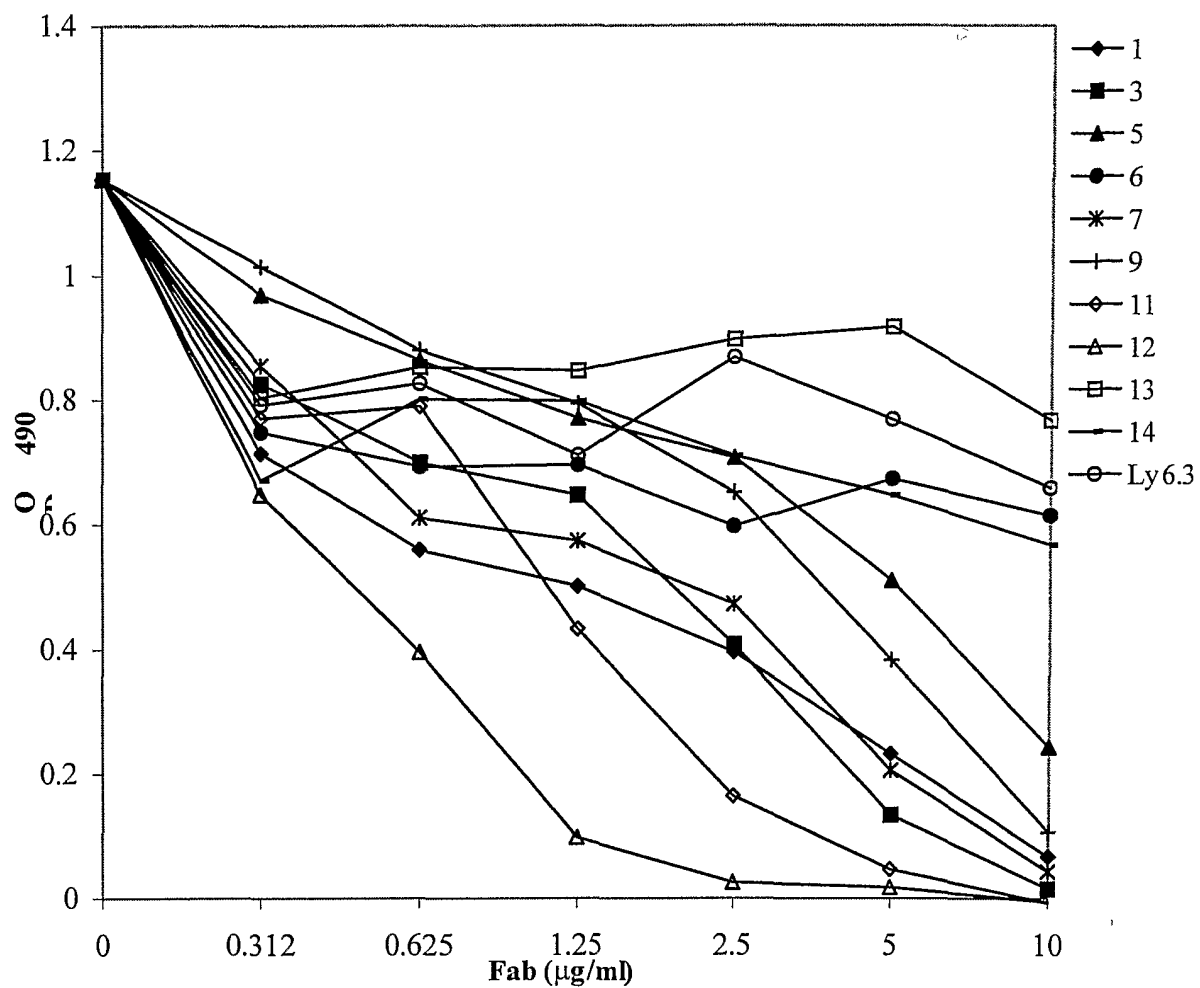


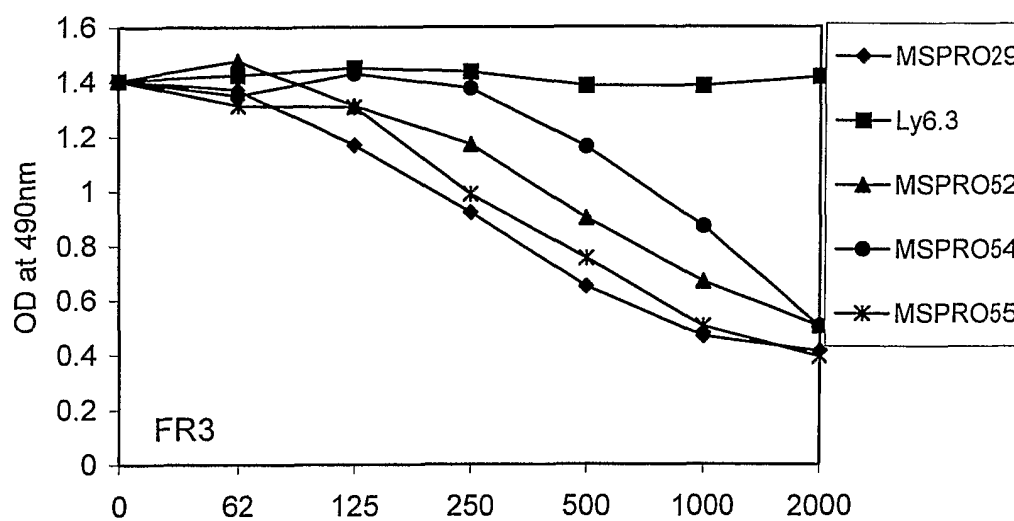
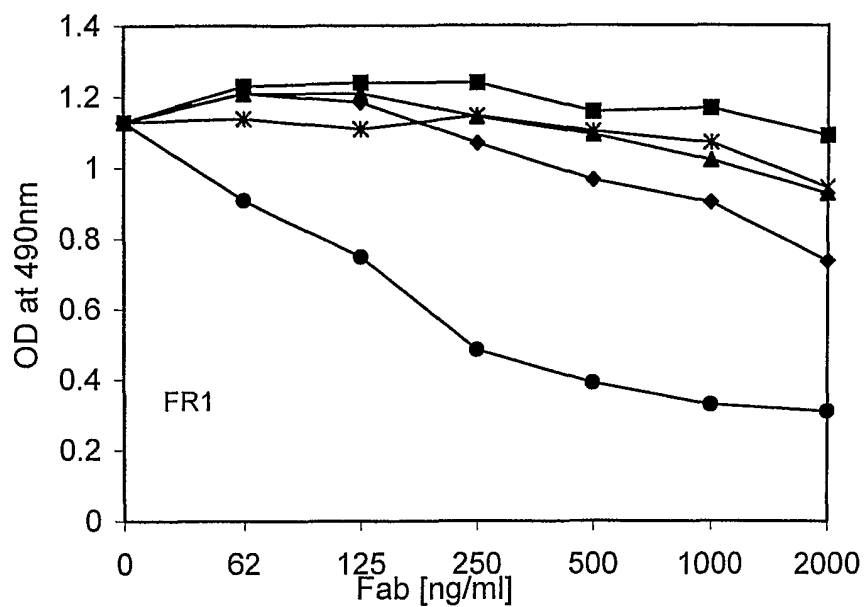
Figure 7A**Figure 7B**

Figure 8A

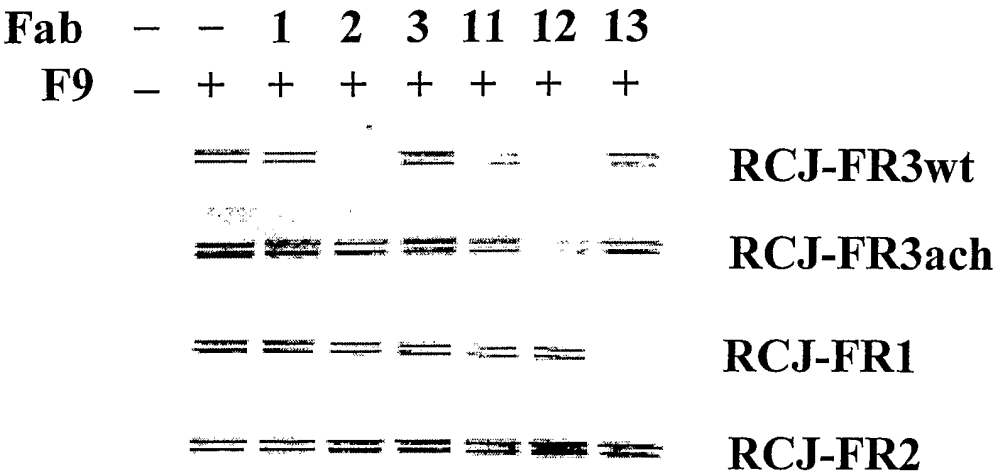


Figure 8B

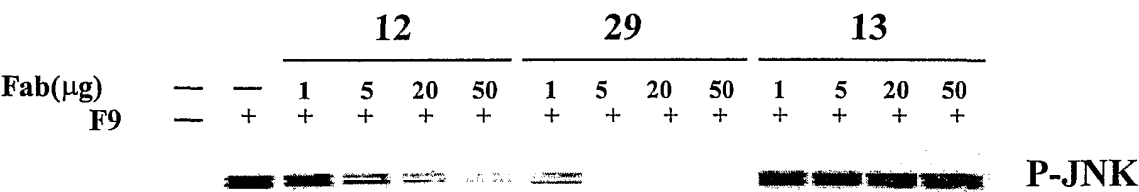


FIGURE 9A
M14

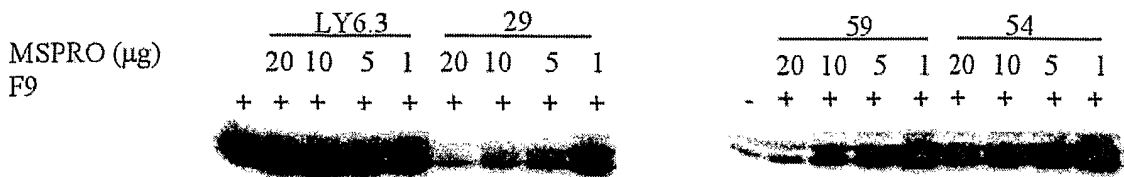


FIGURE 9B
W11

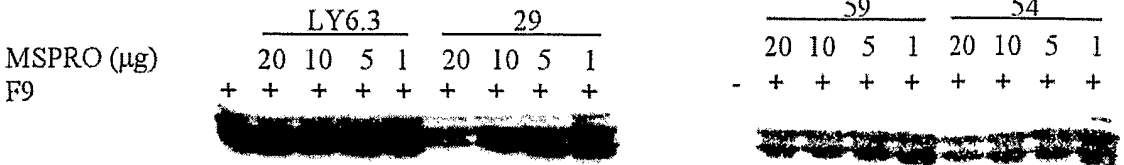


FIGURE 9C
R1-1

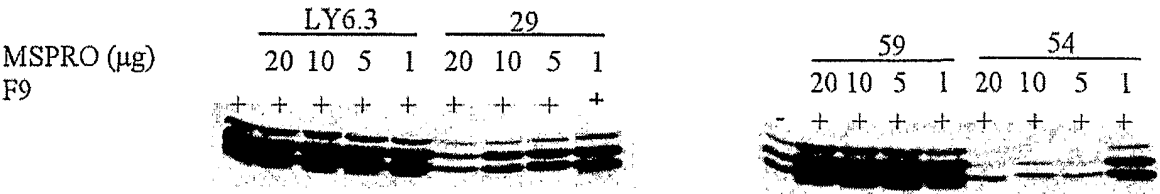


FIGURE 9D
R2-2

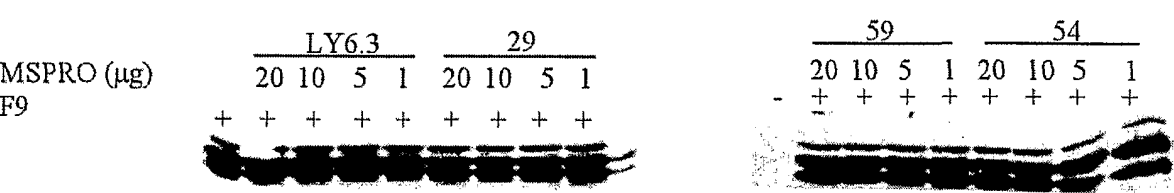


Figure 10

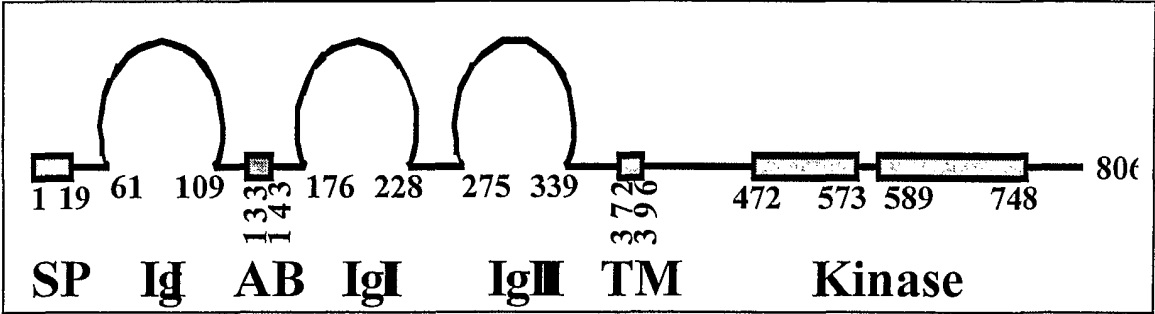
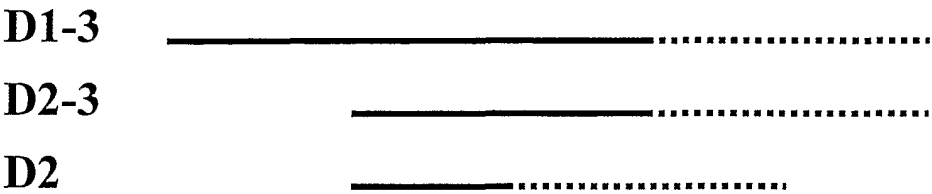
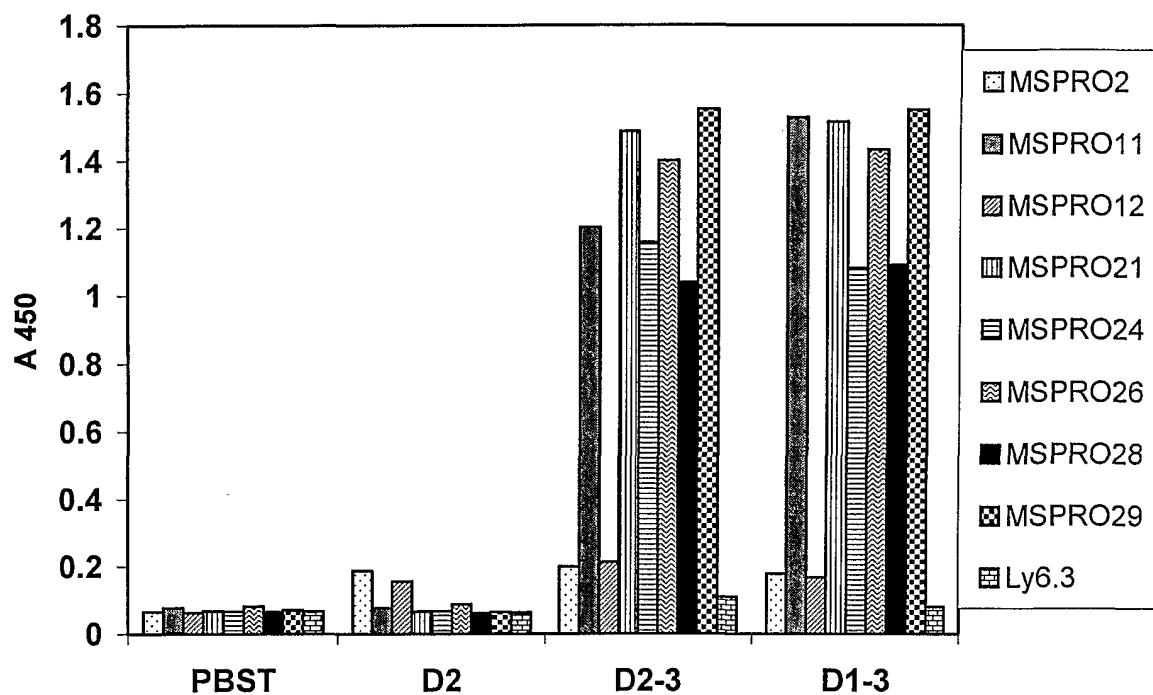


Figure 11



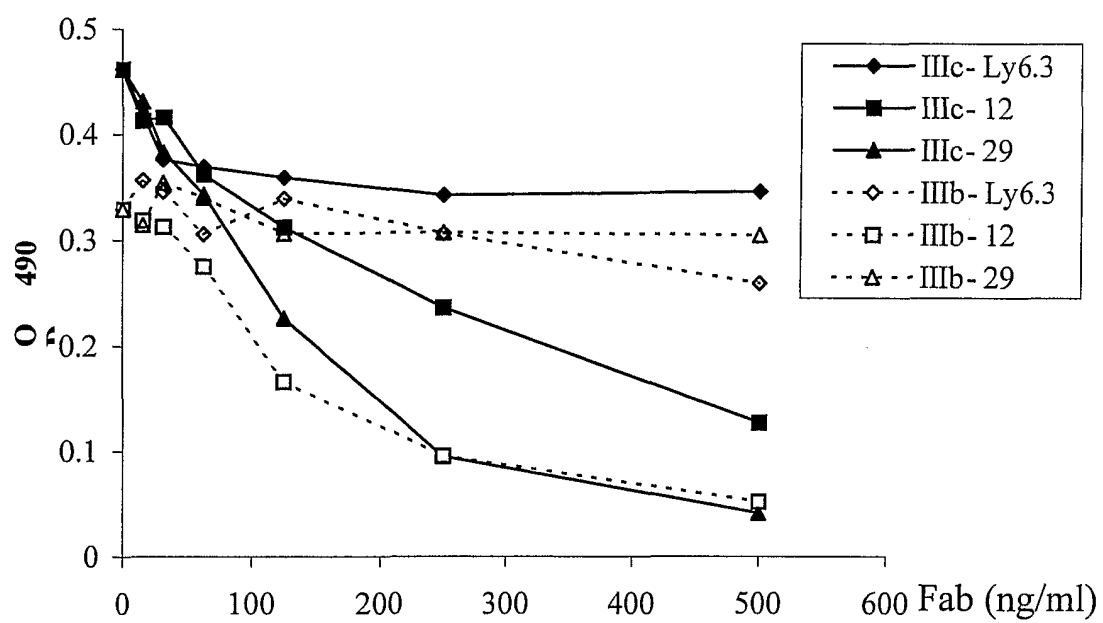


Figure 12

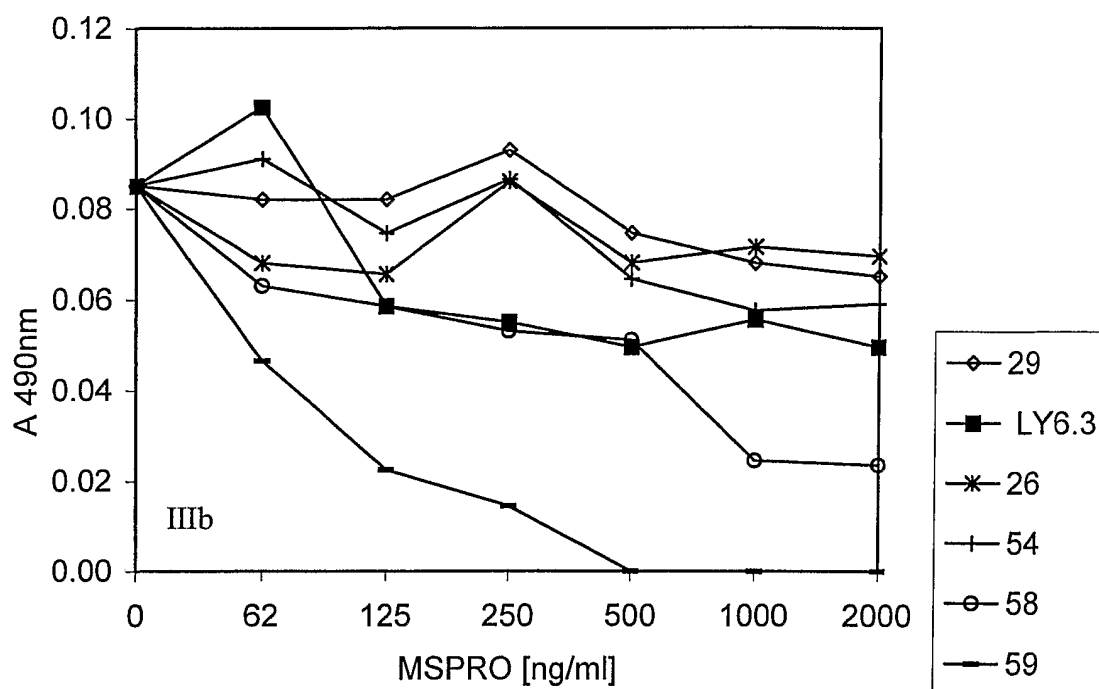
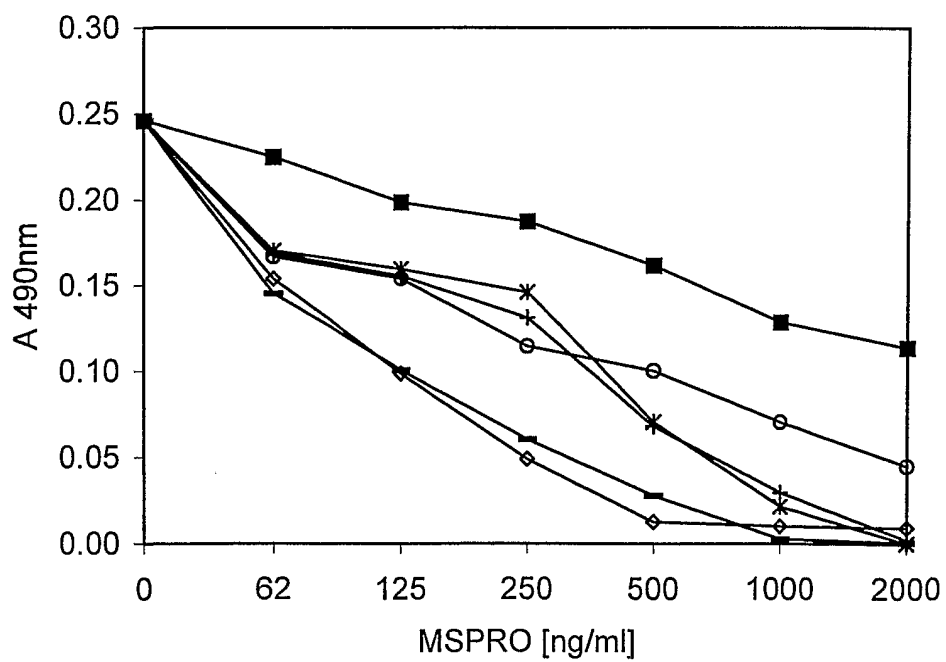
Figure 13A**Figure 13B**

Figure 14

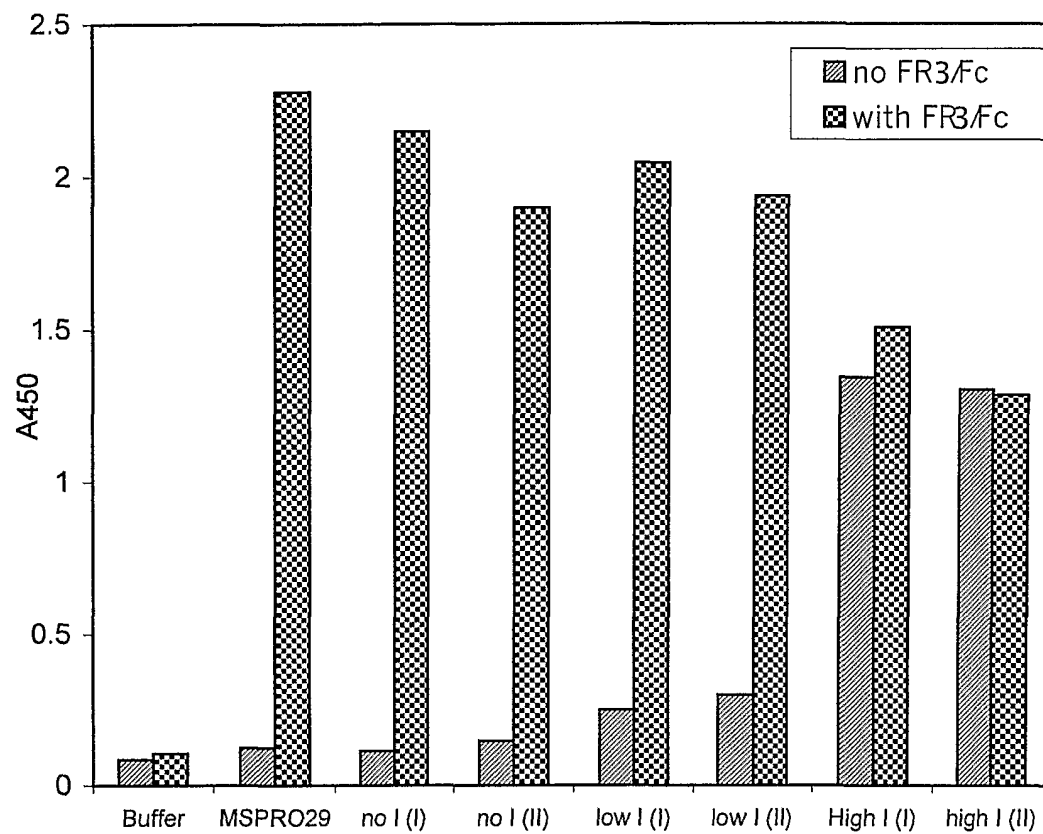


Figure 15

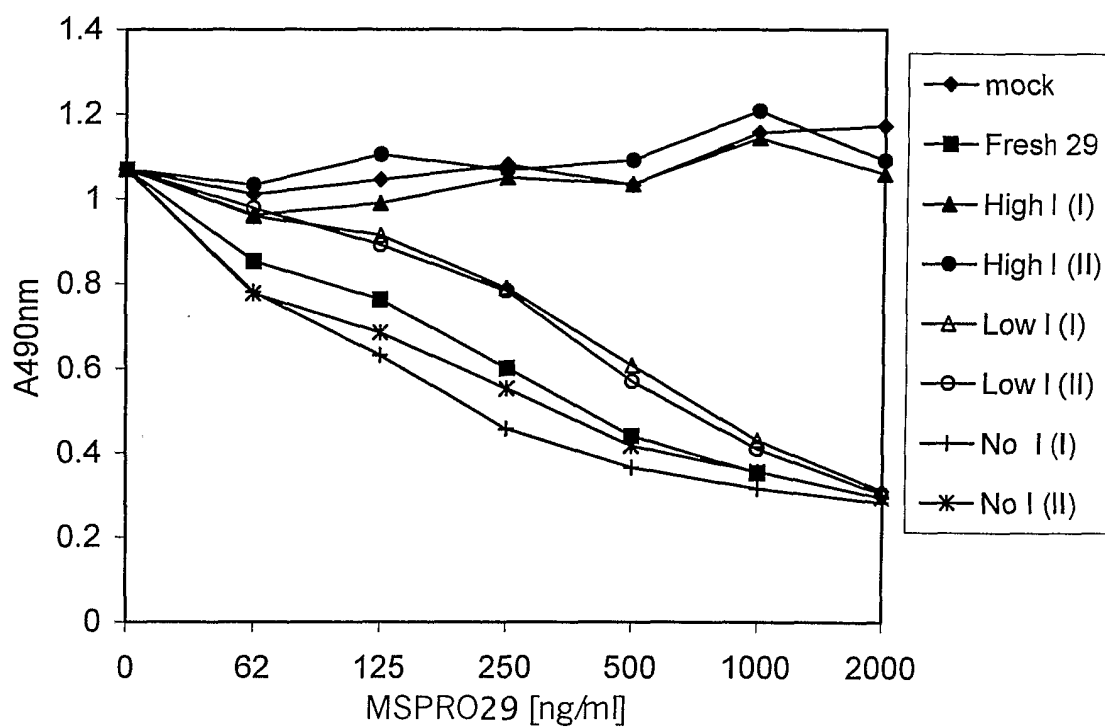


Figure 16

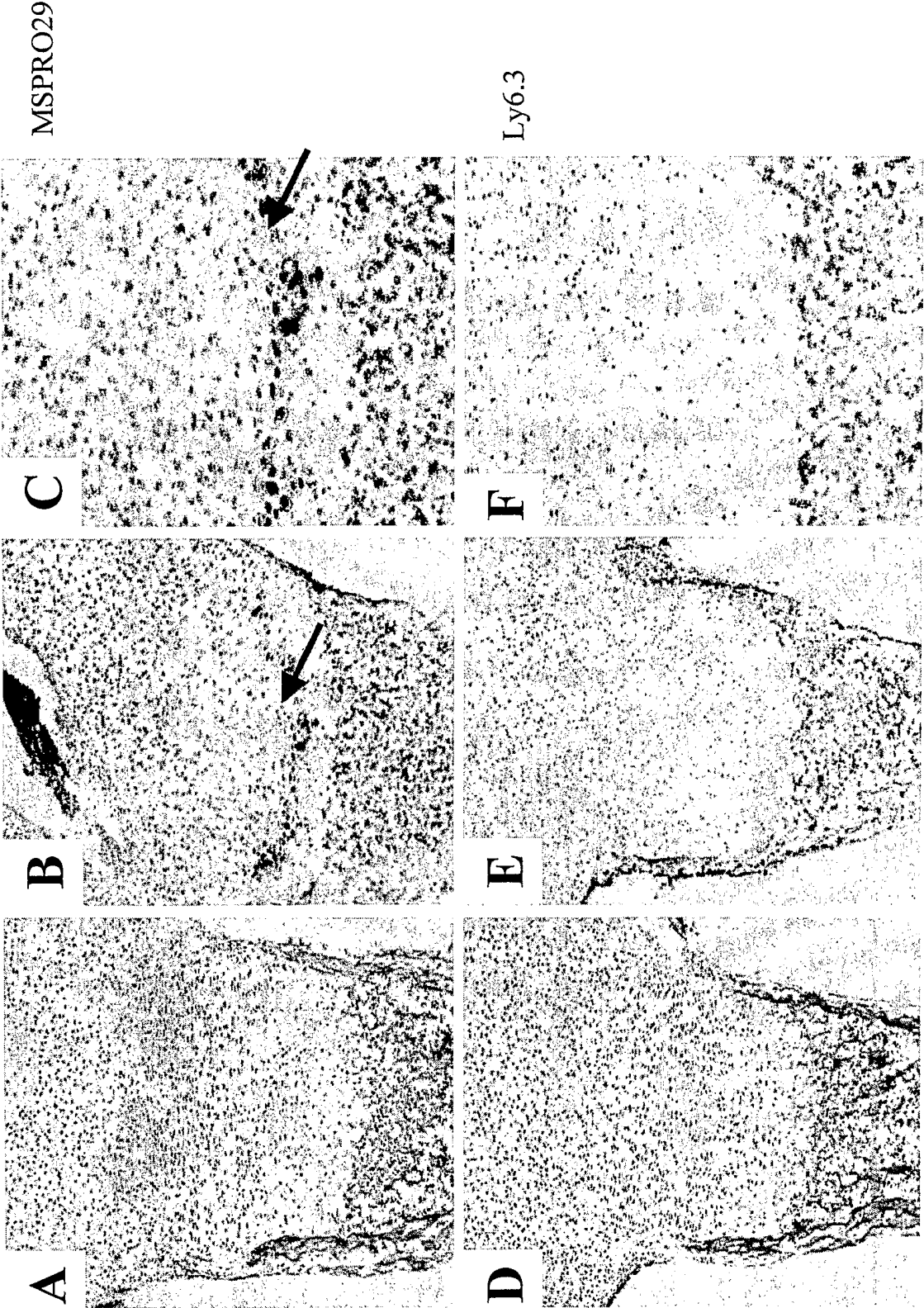


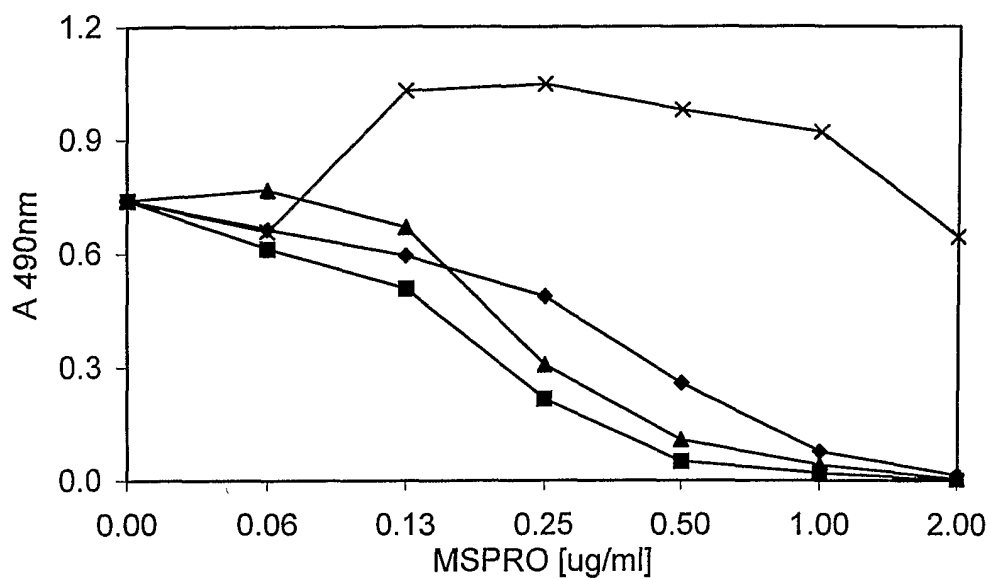
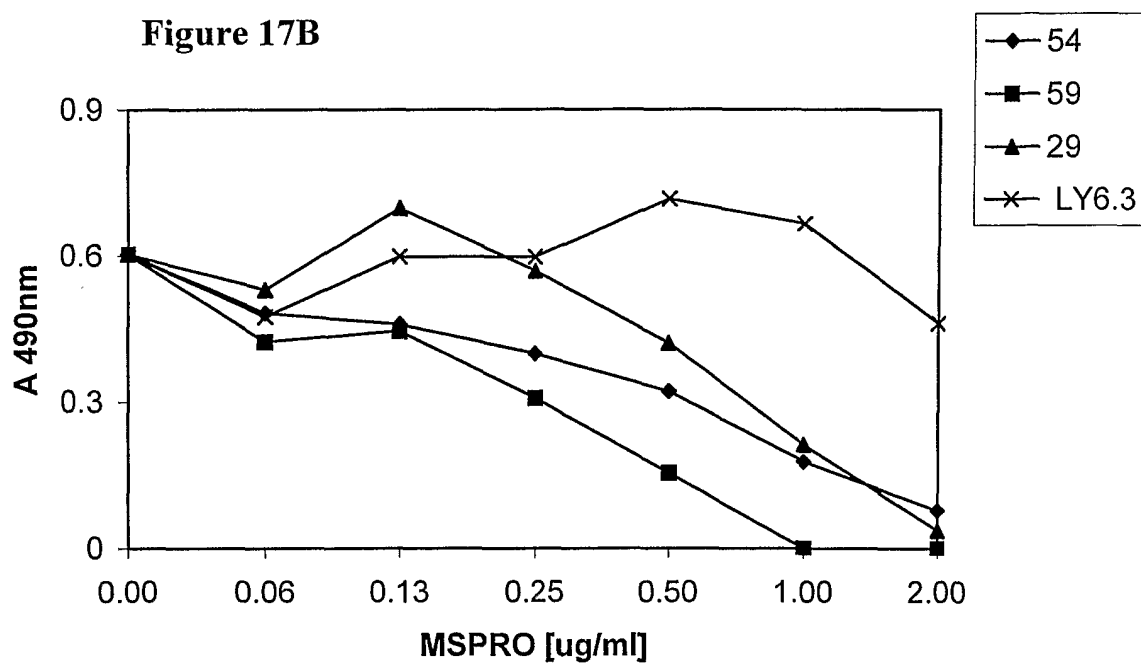
Figure 17A**Figure 17B**

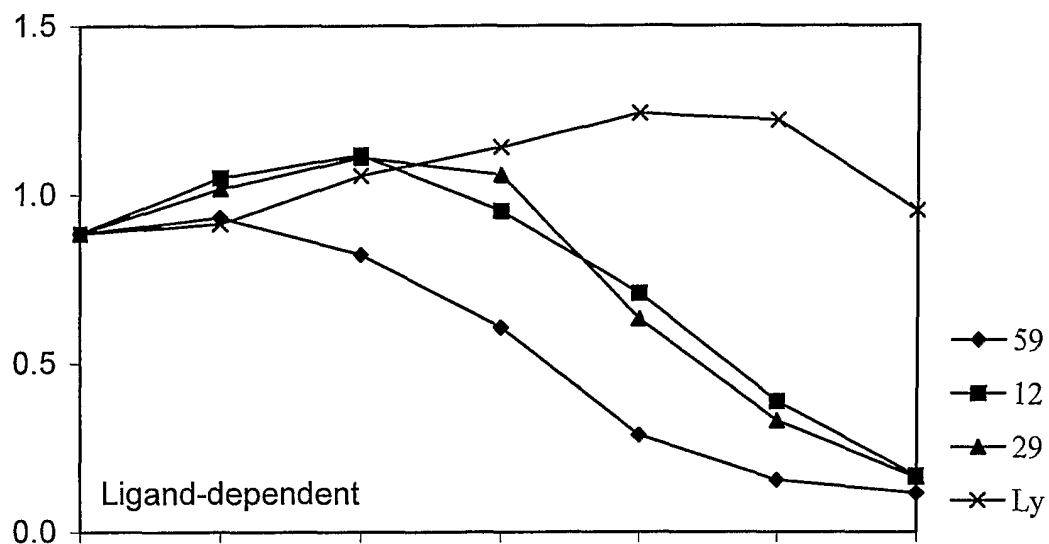
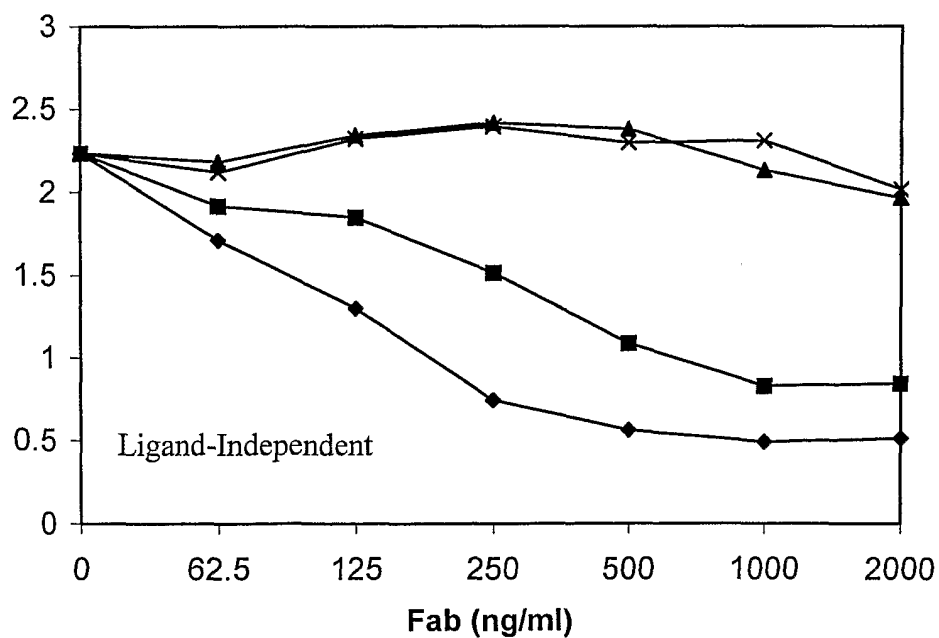
Figure 18A**Figure 18B**

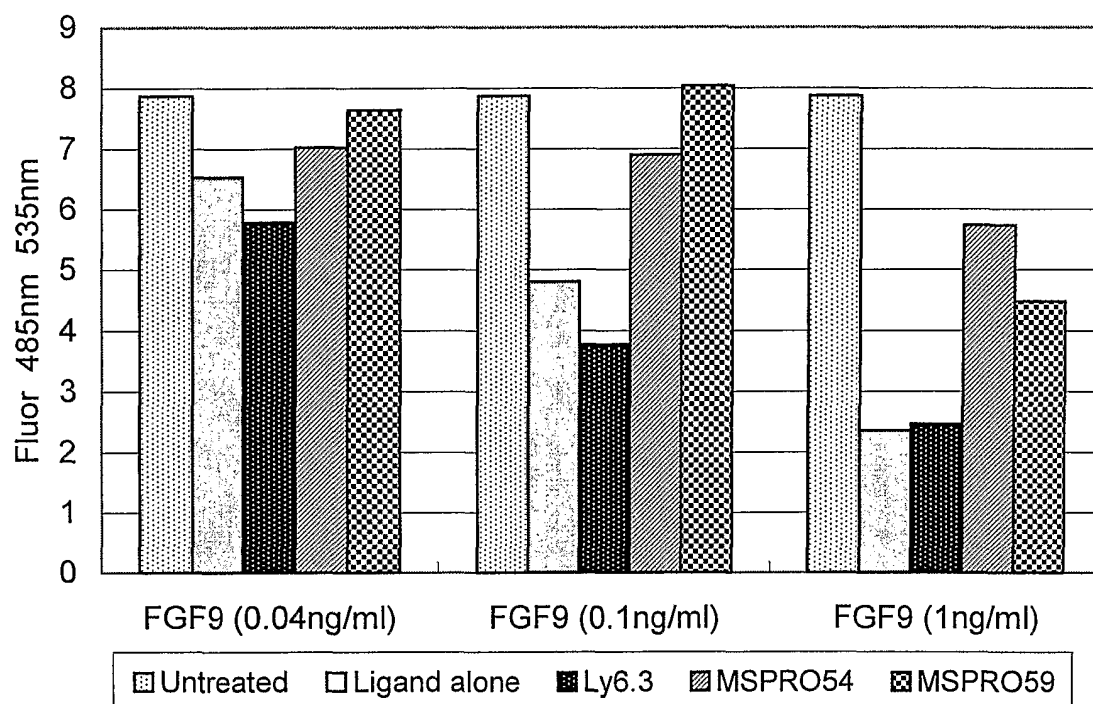
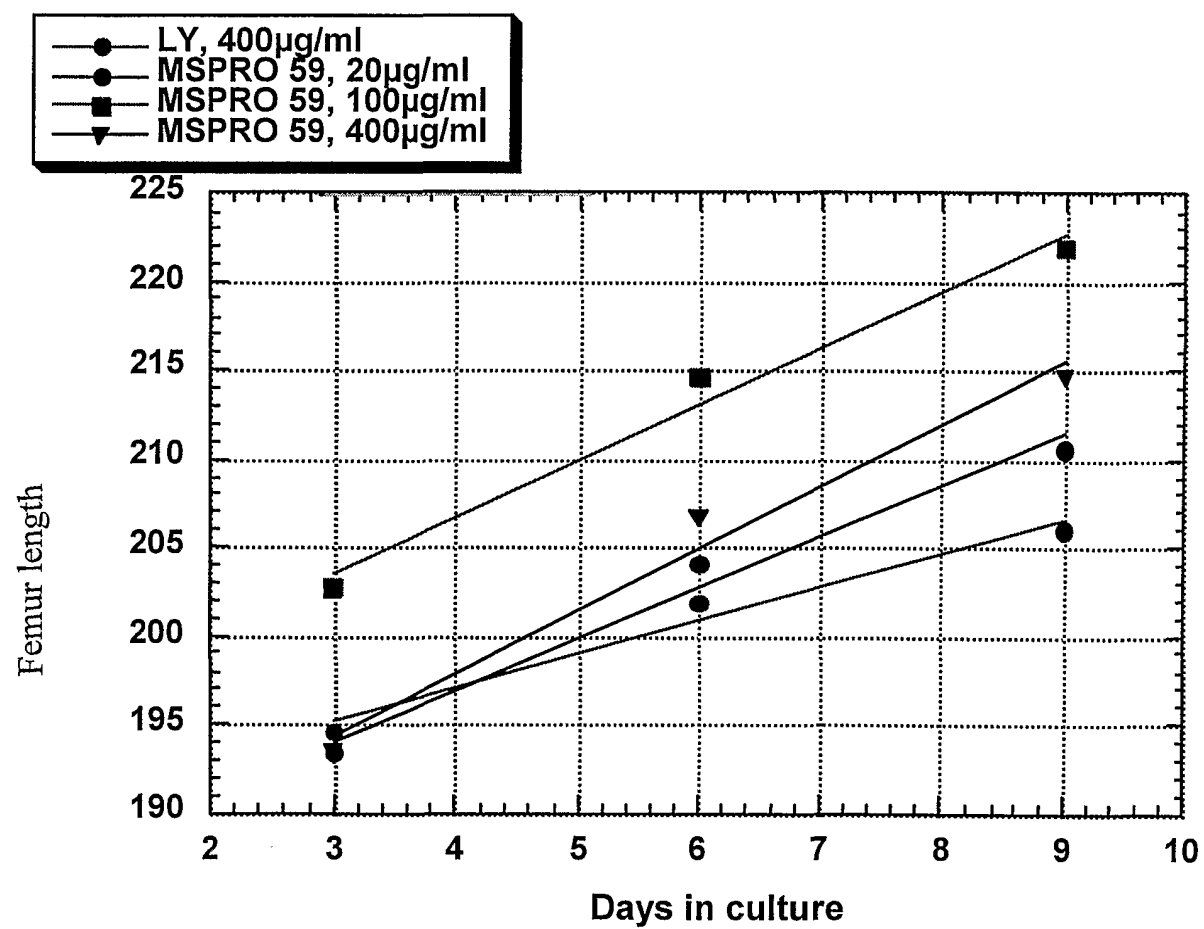
Figure 19

Figure 20



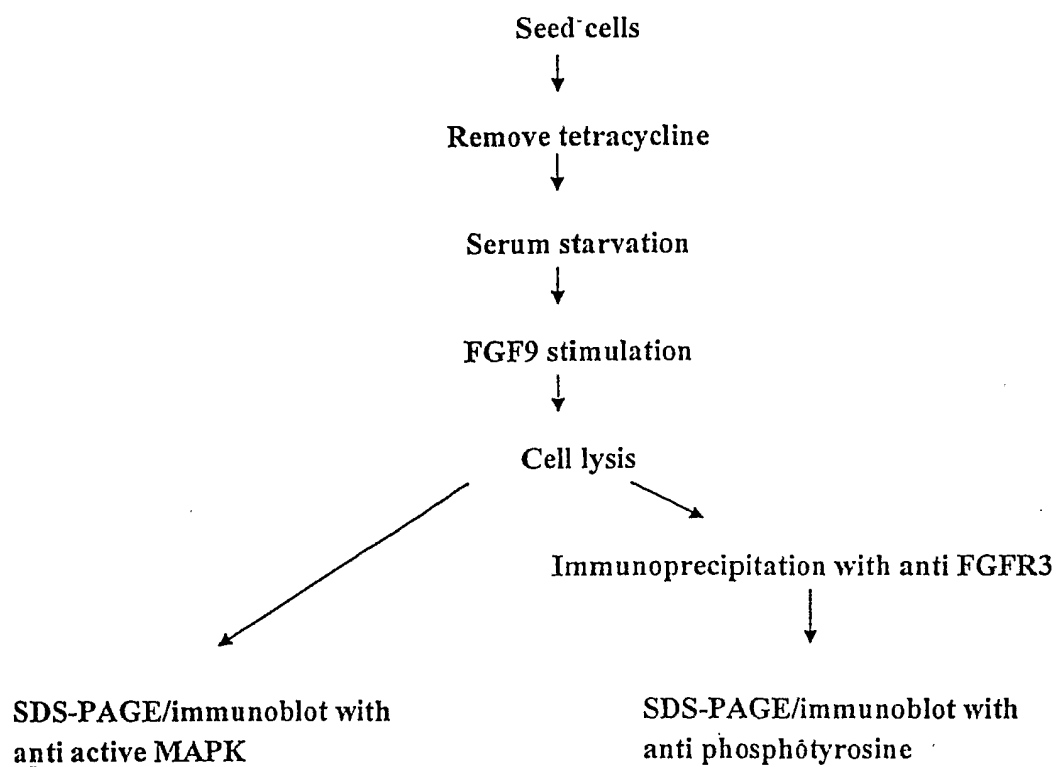


FIG. 21

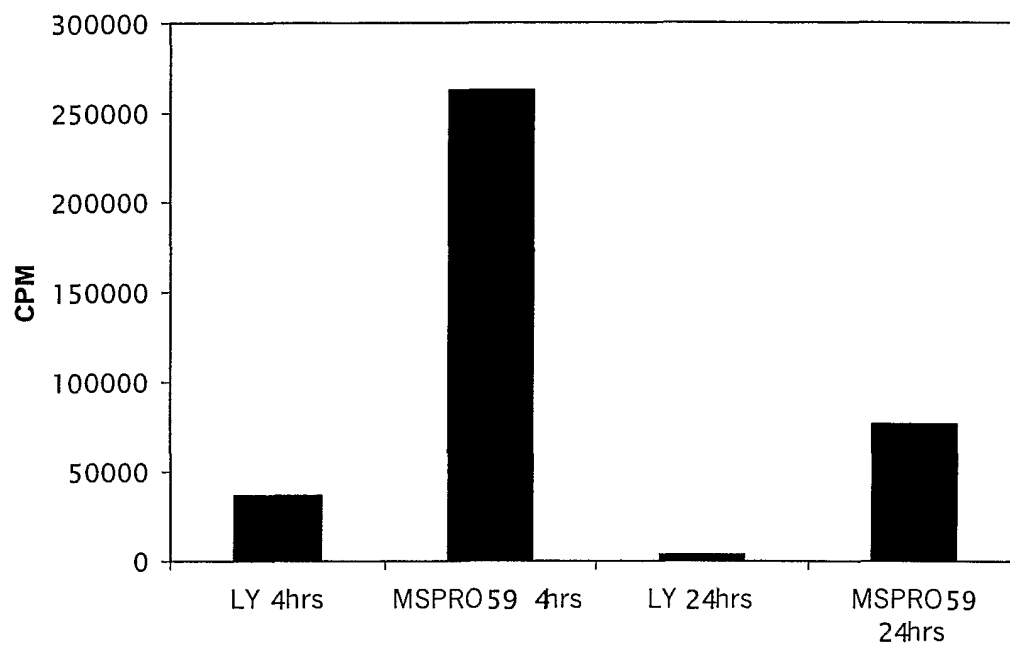
Figure 22

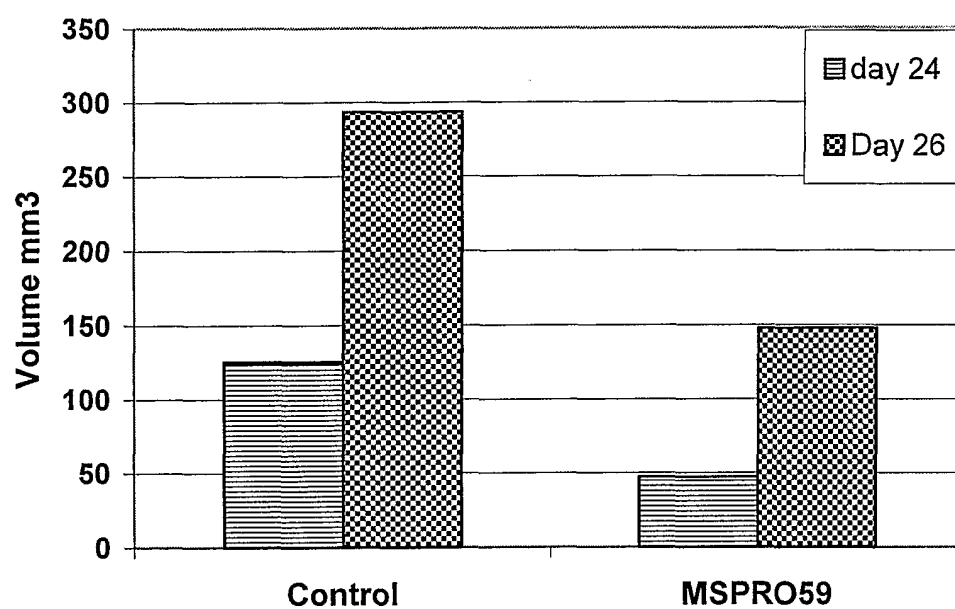
Figure 23

Figure 24

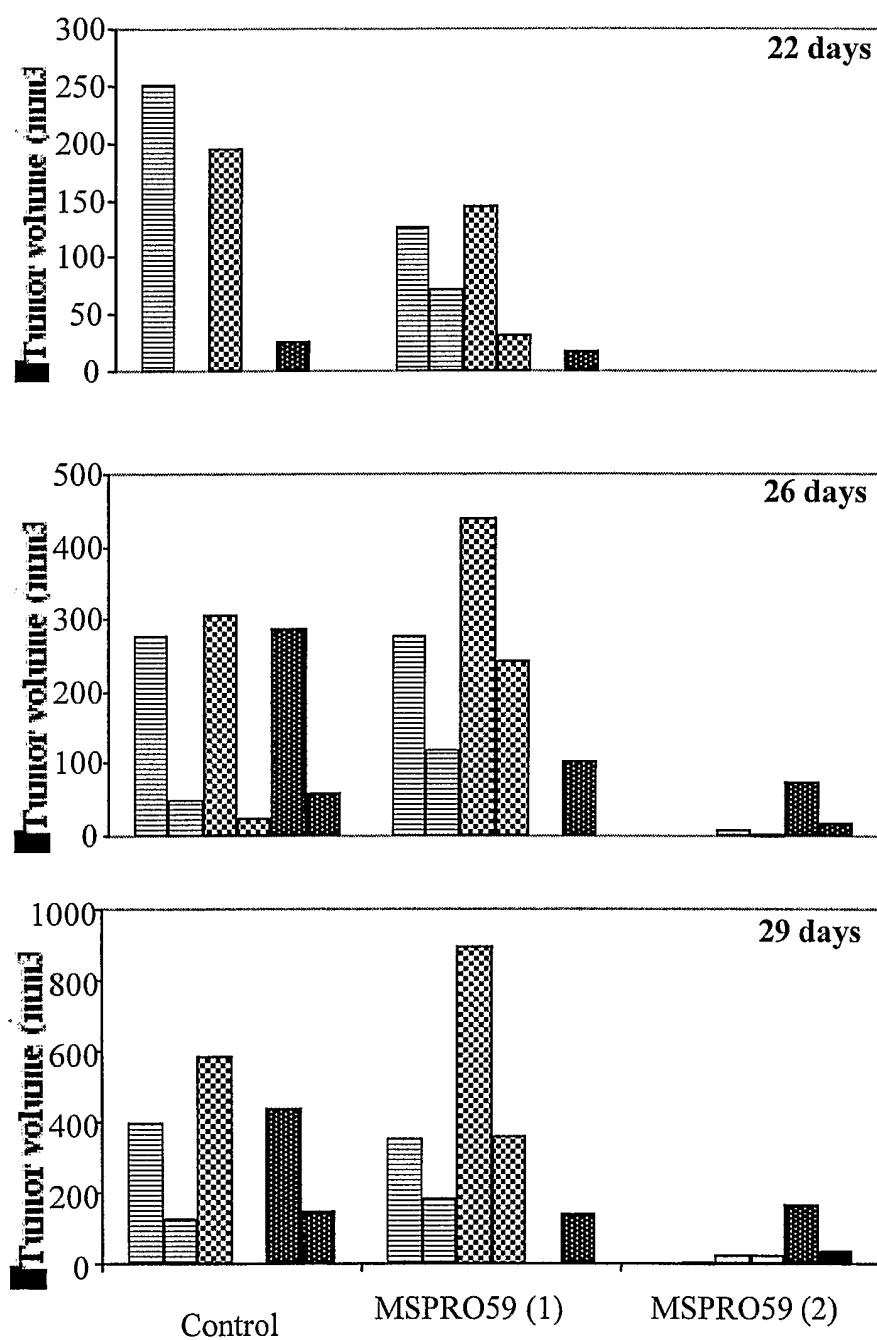


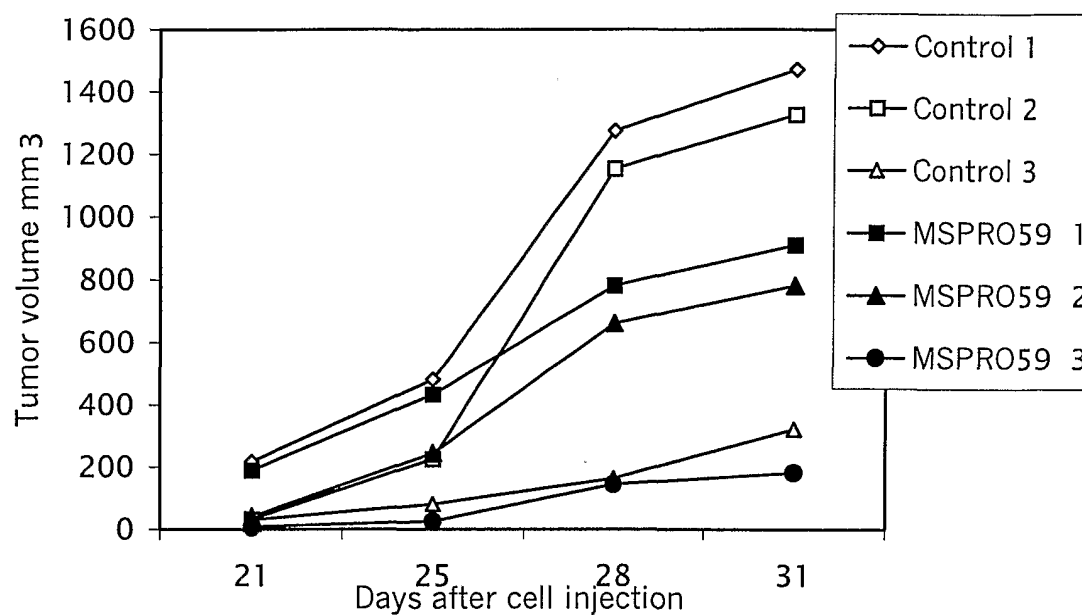
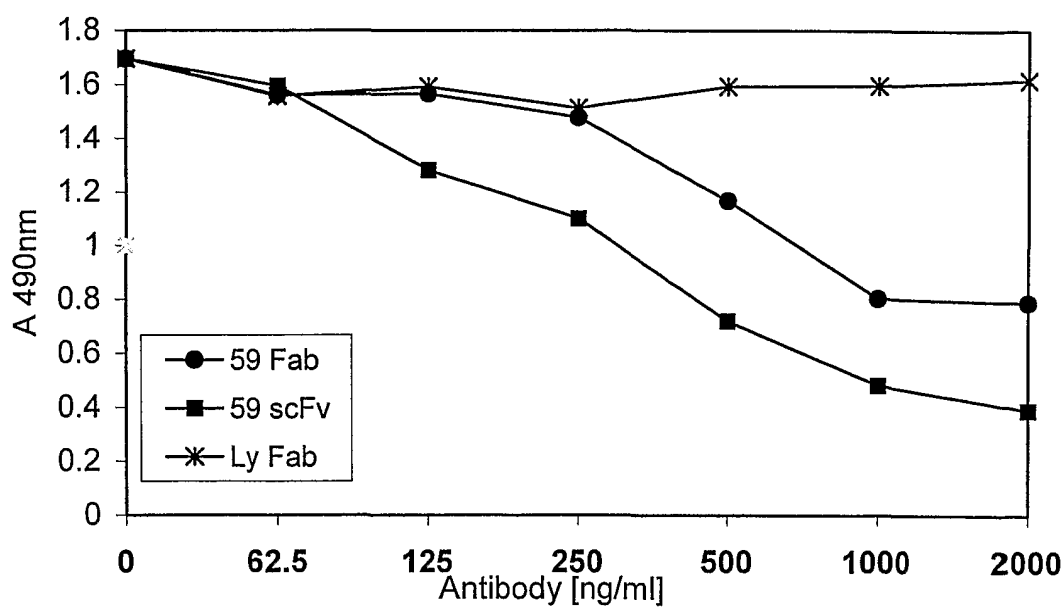
Figure 25A**Figure 25B**

Figure 26

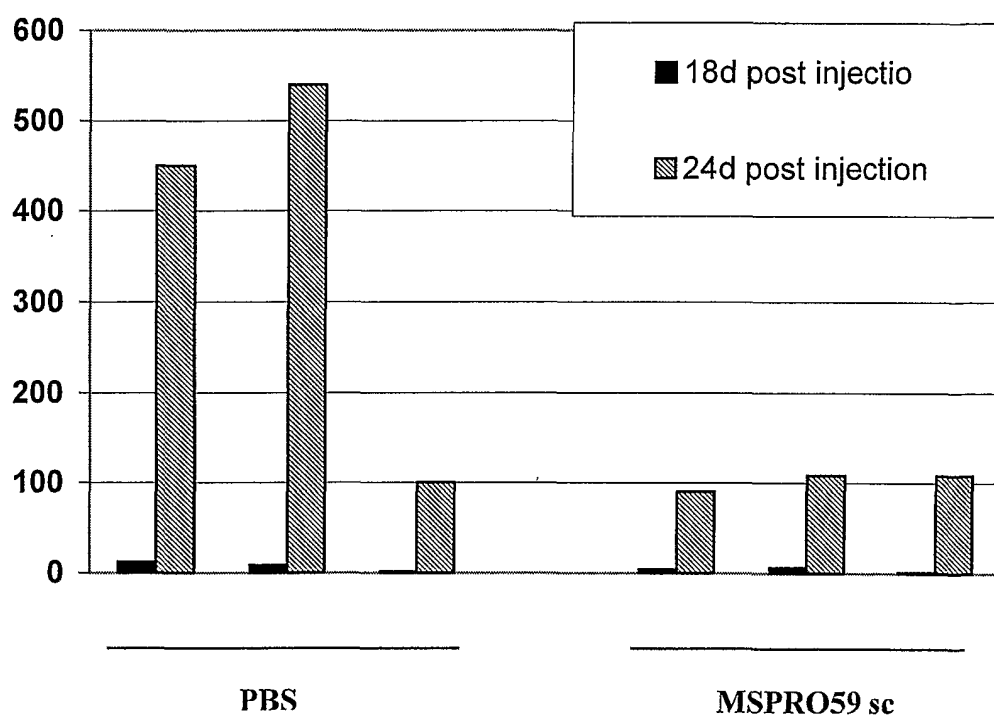


Figure 27

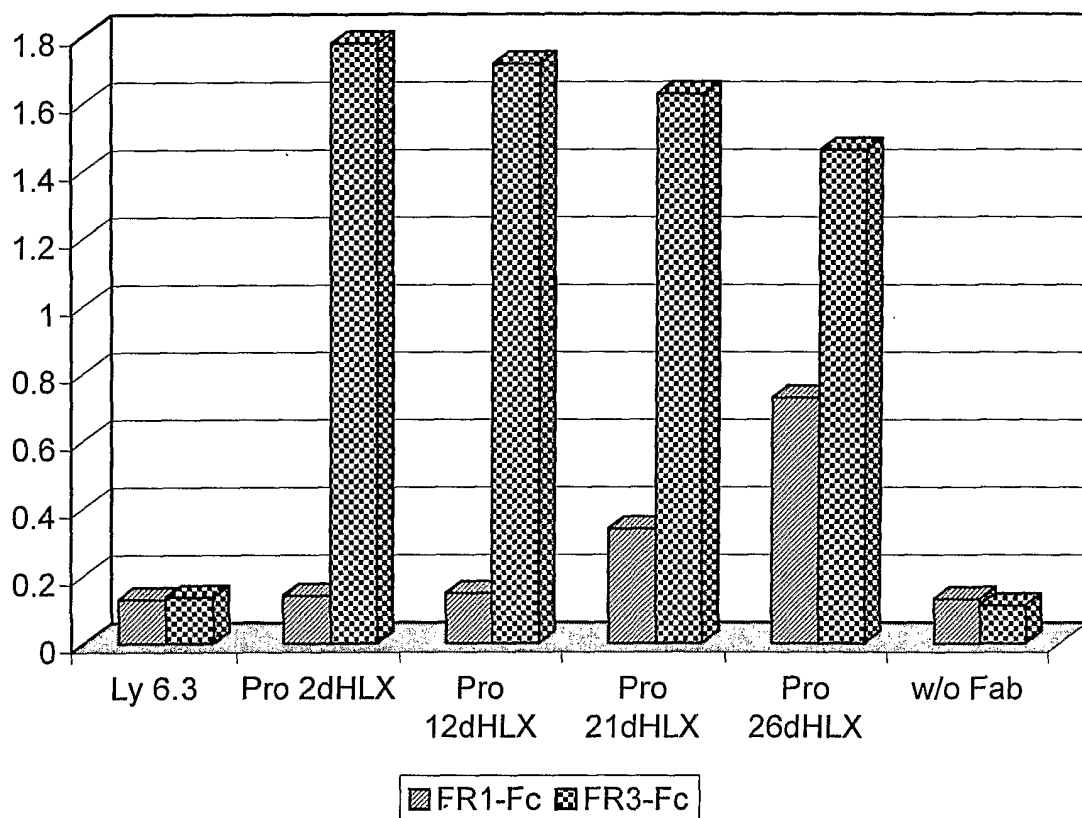


Figure 28A

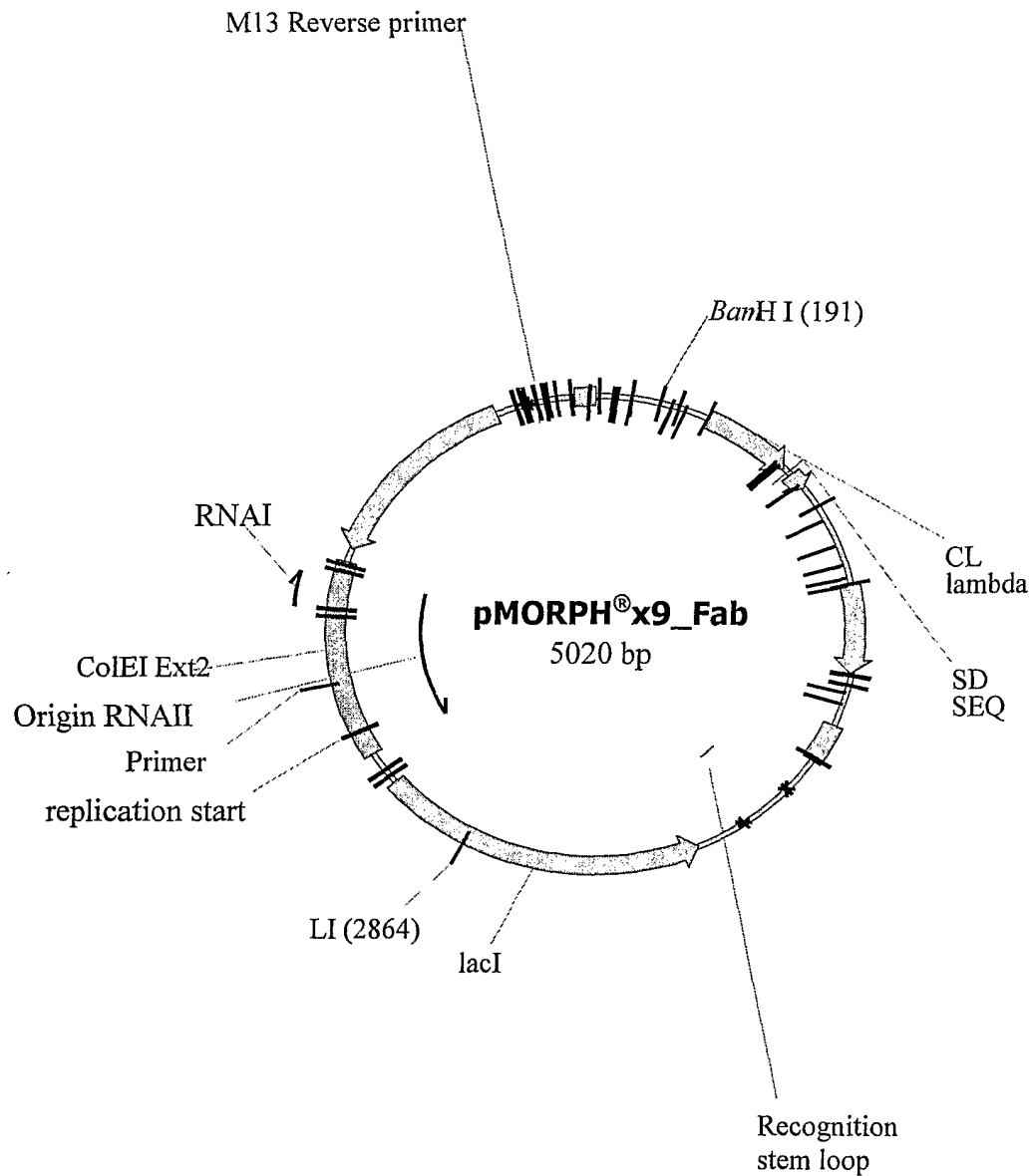


Figure 28B

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 CTGGTAGAGC ACATCGCCGT CGTCGTCGTT GTAACCGTCG TTGATACACT  
  
 10 XmaI  
  
 KpnI              SmaI  
 ~~~~~  
 15 Acc65I Aval
 ~~~~~  
 101 GCTGGTACCA GCAGTTGCCC GGGACGGCGC CGAAACTGCT GATTTATGAT  
 CGACCATGGT CGTCAACGGG CCCTGCCGCG GCTTTGACGA CTAATACTA  
  
 20 Bsu36I              BamHI  
 ~~~~~  
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 25 BpuAI
 ~~~~~  
 BbsI  
 ~~~~~  
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 30 Bsu36I
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 35 MscI              DraIII  
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 StuI SphI
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BspEI
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AvaI
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SfuI
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NspV  
~~~~~

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CelII
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2351 ATATAACATG AGCTGTCCTC GGTATCGTCG TATCCCCTA CCGAGATGTC
TATATTGTAC TCGACAGGAG CCATAGCAGC ATAGGGTGAT GGCTCTACAG

25 2401 CGCACCAACG CGCAGCCCGG ACTCGGTAAT GGCACGCATT GCGCCCAGCG
GCGTGGTTGC GCGTCGGGCC TGAGCCATTA CCGTGCGTAA CGCGGGTCGC

2451 CCATCTGATC GTTGGCAACC AGCATCGCAG TGGGAACGAT GCCCTCATTC
GGTAGACTAG CAACCGTTGG TCGTAGCGTC ACCCTTGCTA CGGGAGTAAG

30 2501 AGCATTTGCA TGGTTTGTG AAAACCGGAC ATGGCACTCC AGTCGCCTTC
TCGTAAACGT ACCAAACAAC TTTTGGCCTG TACCGTGAGG TCAGCGGAAG

2551 CCGTTCCGCT ATCGGCTGAA TTTGATTGCG AGTGAGATAT TTATGCCAGC
35 GGCAAGGCGA TAGCCGACTT AAATAACGC TCACTCTATA AATACGGTGC

2601 CAGCCAGACG CAGACGCGCC GAGACAGAAC TTAATGGGCC AGCTAACAGC
GTCGGTCTGC GTCTGCGCGG CTCTGTCTTG AATTACCCGG TCGATTGTCG

40 2651 GCGATTTGCT GGTGGCCCAA TGCGACCAGA TGCTCCACGC CCAGTCGCGT
CGCTAAACGA CCACCGGGTT ACGCTGGTCT ACGAGGTGCG GGTCAGCGCA

2701 ACCGTCCTCA TGGGAGAAAA TAATACTGTT GATGGGTGTC TGGTCAGAGA
TGGCAGGAGT ACCCTCTTTT ATTATGACAA CTACCCACAG ACCAGTCTCT

45 2751 CATCAAGAAA TAACGCCGGA ACATTAGTGC AGGCAGCTTC CACAGCAATA
GTAGTTCTTT ATTGCGGCCT TGTAATCACG TCCGTCGAAG GTGTCGTTAT

2801 GCATCCTGGT CATCCAGCGG ATAGTTAATA ATCAGCCAC TGACACGTTG
50 CGTAGGACCA GTAGGTCGCC TATCAATTAT TAGTCGGGTG ACTGTGCAAC

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2851 CGCGAGAAGA TTGTGCACCG CCGCTTTACA GGCTTCGACG CCGCTTCGTT  
55 GCGCTCTTCT AACACGTGGC GGCGAAATGT CCGAAGCTGC GGCGAAGCAA

2901 CTACCATCGA CACGACCACG CTGGCACCCA GTTGATCGGC GCGAGATTTA

GATGGTAGCT GTGCTGGTGC GACCGTGGGT CAACTAGCCG CGCTCTAAAT

2951 ATCGCCGCGA CAATTTGCGA CGGCGCGTGC AGGGCCAGAC TGGAGGTGGC  
TAGCGGCGCT GTTAAACGCT GCCGCGCACG TCCCGGTCTG ACCTCCACCG

5 3001 AACGCCAATC AGCAACGACT GTTTGCCCGC CAGTTGTTGT GCCACGCGGT  
TTGCGGTAG TCGTTGCTGA CAAACGGGCG GTCAACAACA CGGTGCGCCA

10 3051 TAGGAATGTA ATTCAGCTCC GCCATCGCCG CTTCCACTTT TTCCCGCGTT  
ATCCTTACAT TAAGTCGAGG CGGTAGCGGC GAAGGTGAAA AAGGGCGCAA

15 3101 TTCGCAGAAA CGTGGCTGGC CTGGTTCACC ACGCGGGAAA CGGTCTGATA  
AAGCGTCTTT GCACCGACCG GACCAAGTGG TGCGCCCTTT GCCAGACTAT

20 3151 AGAGACACCG GCATACTCTG CGACATCGTA TAACGTTACT GGTTTCACAT  
TCTCTGTGGC CGTATGAGAC GCTGTAGCAT ATTGCAATGA CCAAAGTGTA

25 3201 TCACCACCCT GAATTGACTC TCTTCGGGC GCTATCATGC CATACCGCGA  
AGTGGTGGGA CTTAACTGAG AGAAGGCCCG CGATAGTACG GTATGGCGCT

30 3251 AAGGTTTTGC GCCATTCGAT GCTAGCCATG TGAGCAAAAG GCCAGCAAAA  
TTCCAAAACG CGGTAAGCTA CGATCGGTAC ACTCGTTTTC CGGTGTTTT

35 3301 GGCCAGGAAC CGTAAAAAGG CCGCGTTGCT GCGTTTTTC CATAGGCTCC  
CCGGTCCTTG GCATTTTTCC GGCACAACGA CCGCAAAAAG GTATCCGAGG

40 3351 GCCCCCTGA CGAGCATCAC AAAAATCGAC GCTCAAGTCA GAGGTGGCGA  
CGGGGGGACT GCTCGTAGTG TTTTAGCTG CGAGTTCAGT CTCCACCGCT

45 3401 AACCCGACAG GACTATAAAG ATACCAGGCG TTTCCCCCTG GAAGCTCCCT  
TTGGGCTGTC CTGATATTTT TATGGTCCGC AAAGGGGGAC CTTCGAGGGA

50 3451 CGTGCGCTCT CCTGTTCCGA CCCTGCCGCT TACCGGATAC CTGTCCGCCT  
GCACGCGAGA GGACAAGGCT GGGACGGCGA ATGGCCTATG GACAGGCGGA

55 3501 TTCTCCCTTC GGGAAGCGTG GCGCTTTCTC ATAGCTCACG CTGTAGGTAT  
AAGAGGGAAG CCCTTCGCAC CGCGAAAGAG TATCGAGTGC GACATCCATA

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3551 CTCAGTTCGG TGTAGGTCGT TCGCTCCAAG CTGGGCTGTG TGCACGAACC
GAGTCAAGCC ACATCCAGCA AGCGAGGTTC GACCCGACAC ACGTGCTTGG

3601 CCCCGTTCAG CCCGACCGCT GCGCCTTATC CGGTAAGTAT CGTCTTGAGT
GGGGCAAGTC GGGCTGGCGA CGCGGAATAG GCCATTGATA GCAGAACTCA

3651 CCAACCCGGT AAGACACGAC TTATCGCCAC TGGCAGCAGC CACTGGTAAC
GGTTGGGCCA TTCTGTGCTG AATAGCGGTG ACCGTCGTCG GTGACCATTG

3701 AGGATTAGCA GAGCGAGGTA TGTAGGCGGT GCTACAGAGT TCTTGAAGTG
TCCTAATCGT CTCGCTCCAT ACATCCGCCA CGATGTCTCA AGAACTTCA

3751 GTGGCCTAAC TACGGCTACA CTAGAAGAAC AGTATTTGGT ATCTGCGCTC
CACCGGATTG ATGCCGATGT GATCTTCTTG TCATAAACCA TAGACGCGAG

3801 TGCTGTAGCC AGTTACCTTC GGAAAAAGAG TTGGTAGCTC TTGATCCGGC
ACGACATCGG TCAATGGAAG CCTTTTTCTC AACCATCGAG AACTAGGCCG

3851 AAACAAACCA CCGCTGGTAG CGGTGGTTTT TTTGTTTGCA AGCAGCAGAT
TTTGTGTTGGT GGCGACCATC GCCACCAAAA AAACAAACGT TCGTCGTCTA

5 3901 TACGCGCAGA AAAAAAGGAT CTCAAGAAGA TCCTTTGATC TTTTCTACGG
ATGCGCGTCT TTTTTTCCTA GAGTTCTTCT AGGAAACTAG AAAAGATGCC

3951 GGTCTGACGC TCAGTGGAAC GAAAACTCAC GTTAAGGGAT TTTGGTCAGA
CCAGACTGCG AGTCACCTTG CTTTTGAGTG CAATTCCCTA AAACCACTCT

10 4001 TCTAGCACCA GGCGTTTAAG GGCACCAATA ACTGCCTTAA AAAAATTACG
AGATCGTGGT CCGCAAATC CCGTGGTTAT TGACGGAATT TTTTAATGC

4051 CCCC GCCCTG CCACTCATCG CAGTACTGTT GTAATTCATT AAGCATTCTG
15 GGGGCGGGAC GGTGAGTAGC GTCATGACAA CATTAAAGTAA TTCGTAAGAC

4101 CCGACATGGA AGCCATCACA AACGGCATGA TGAACCTGAA TCGCCAGCGG
GGCTGTACCT TCGGTAGTGT TTGCCGTACT ACTTGGA CT AGCGGTGCGC

20 4151 CATCAGCACC TTGTCGCCTT GCGTATAATA TTTGCCCATTA GTGAAAACGG
GTAGTCGTGG AACAGCGGAA CGCATATTAT AAACGGGTAT CACTTTTGCC

4201 GGGCGAAGAA GTTGTCCATA TTGGCTACGT TTAAATCAAA ACTGGTGAAA
CCCGCTTCTT CAACAGGTAT AACCGATGCA AATTTAGTTT TGACCACTTT

25 4251 CTCACCCAGG GATTGGCTGA GACGAAAAAC ATATTCTCAA TAAACCCCTT
GAGTGGGTCC CTAACCGACT CTGCTTTTTG TATAAGAGTT ATTTGGGAAA

4301 AGGGAAATAG GCCAGGTTTT CACCGTAACA CGCCACATCT TGCGAATATA
30 TCCCTTTATC CGGTCCAAAA GTGGCATTGT GCGGTGTAGA ACGCTTATAT

4351 TGTGTAGAAA CTGCCGGAAA TCGTCGTGGT ATTCACTCCA GAGCGATGAA
ACACATCTTT GACGGCCTTT AGCAGCACCA TAAGTGAGGT CTCGCTACTT

35 4401 AACGTTTCAG TTTGCTCATG GAAAACGGTG TAACAAGGGT GAACACTATC
TTGCAAAGTC AAACGAGTAC CTTTGGCCAC ATTGTTCCCA CTTGTGATAG

4451 CCATATCACC AGCTCACCGT CTTTCATTGC CATA CGGAAC TCCGGGTGAG
GGTATAGTGG TCGAGTGGCA GAAAGTAACG GTATGCCTTG AGGCCCACTC

40 4501 CATT CATCAG GCGGGCAAGA ATGTGAATAA AGGCCGGATA AA ACTTGTGC
GTAAGTAGTC CGCCCGTTCT TACACTTATT TCCGGCCTAT TTTGAACACG

4551 TTATTTTTCT TTACGGTCTT TAAAAAGGCC GTAATATCCA GCTGAACGGT
45 AATAAAAAGA AATGCCAGAA ATTTTCCGG CATTATAGGT CGACTTGCCA

4601 CTGGTTATAG GTACATTGAG CAACTGACTG AAATGCCTCA AAATGTTCTT
GACCAATATC CATGTA ACTC GTTGACTGAC TTTACGGAGT TTTACAAGAA

50 4651 TACGATGCCA TTGGGATATA TCAACGGTGG TATATCCAGT GATTTTTTTC
ATGCTACGGT AACCTATAT AGTTGCCACC ATATAGGTCA CTAAAAAAG

4701 TCCATTTTAG CTTCTTAGC TCCTGAAAAT CTCGATAACT CAAAAAATAC
AGGTAAAATC GAAGGAATCG AGGACTTTTA GAGCTATTGA GTTTTTTATG

55 4751 GCCCGGTAGT GATCTTATTT CATTATGGTG AAAGTTGGAA CCTCACCCGA
CGGGCCATCA CTAGAATAAA GTAATACCAC TTTCAACCTT GGAGTGGGCT

4801 CGTCTAATGT GAGTTAGCTC ACTCATTAGG CACCCCAGGC TTTACACTTT
GCAGATTACA CTCAATCGAG TGAGTAATCC GTGGGGTCCG AAATGTGAAA

5 4851 ATGCTTCCGG CTCGTATGTT GTGTGGAATT GTGAGCGGAT AACAAATTCA
TACGAAGGCC GAGCATACAA CACACCTTAA CACTCGCCTA TTGTTAAAGT

M13 Reverse primer 100.0% XbaI

10 4901 CACAGGAAAC AGCTATGACC ATGATTACGA ATTTCTAGAT AACGAGGGCA
GTGTCCTTTG TCGATACTGG TACTAATGCT TAAAGATCTA TTGCTCCCGT

15 4951 AAAAATGAAA AAGACAGCTA TCGCGATTGC AGTGGCACTG GCTGGTTTCG
TTTTTACTTT TTCTGTCGAT AGCGCTAACG TCACCGTGAC CGACCAAAGC

EcoRV

20 5001 CTACCGTAGC GCAGGCCGAT
GATGGCATCG CGTCCGGCTA

5

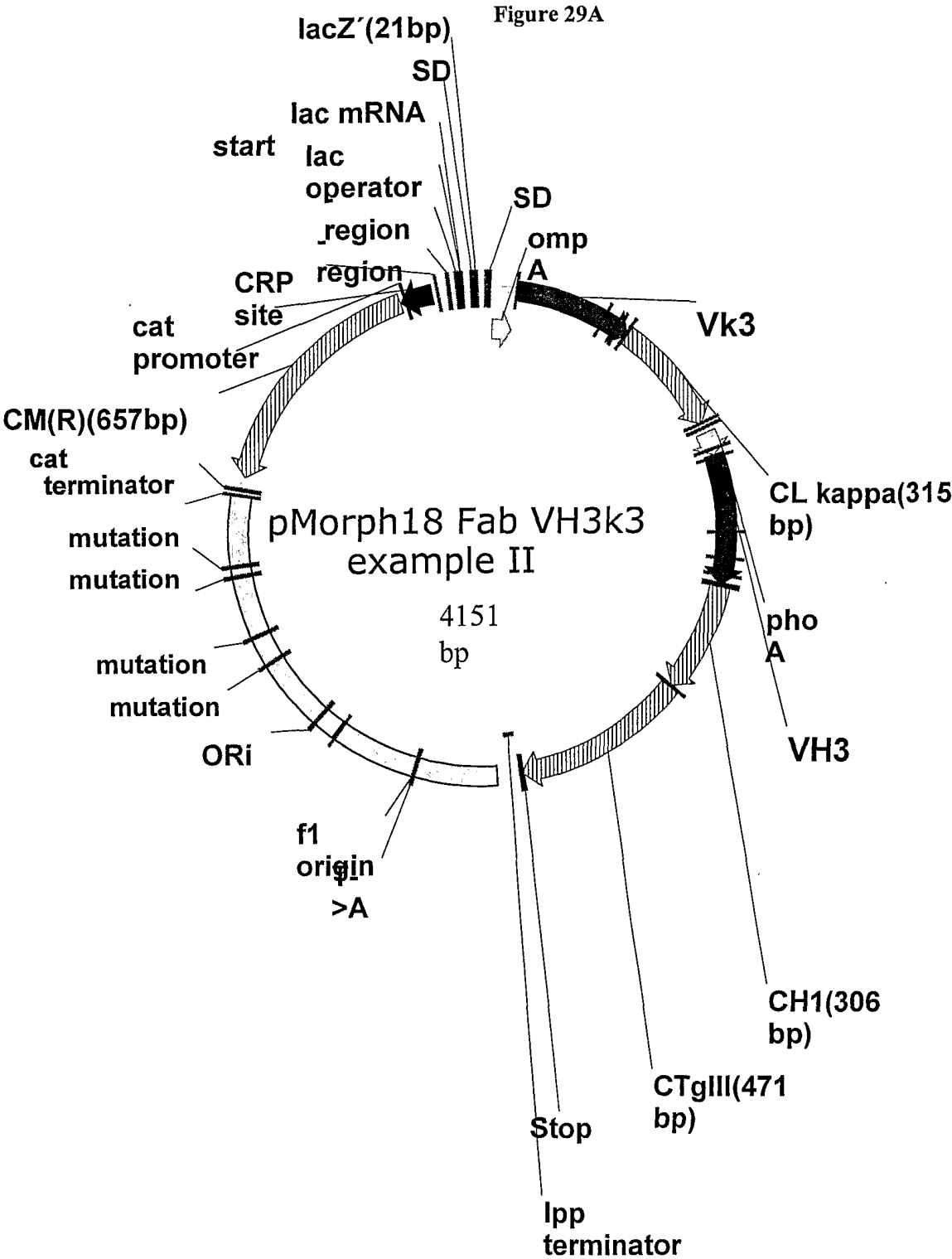


Figure 29B

lacZ' SD ompA
 ~~~~~  
 XbaI  
 ~~~~~  
 5 M K K T A I A I A V ·
 SEQ ID NO:53 TCTAGATAAC GAGGGCAAAA AATGAAAAAG ACAGCTATCG CGATTGCAGT
 AGATCTATTG CTCCCGTTTT TTACTTTTTC TGTCGATAGC GCTAACGTCA
 ~~~~~  
 10 Vk3  
 ompA  
 ~~~~~  
 EcoRV
 ~~~~~  
 15 A L A G F A T V A Q A D I V L T Q ·  
 GGCAGTGGCT GGTTCGCTA CCGTAGCGCA GGCCGATATC GTGCTGACCC  
 CCGTGACCGA CCAAAGCGAT GGCATCGCGT CCGGCTATAG CACGACTGGG  
 ~~~~~  
 20 101 · S P A T L S L S P G E R A T L S
 AGAGCCCGGC GACCCTGAGC CTGTCTCCGG GCGAACGTGC GACCCTGAGC
 TCTCGGGCCG CTGGGACTCG GACAGAGGCC CGCTTGACAG CTGGGACTCG
 ~~~~~  
 25 151 C R A S Q S V S S S Y L A W Y Q Q ·  
 TGCAGAGCGA GCCAGAGCGT GAGCAGCAGC TATCTGGCGT GGTACCAGCA  
 ACGTCTCGCT CGGTCTCGCA CTCGTCTCG ATAGACCGCA CCATGGTCGT  
 ~~~~~  
 30 201 · K P G Q A P R L L I Y G A S S R A ·
 GAAACCAGGT CAAGCACCGC GTCTATTAAT TTATGGCGCG AGCAGCCGTG
 CTTTGGTCCA GTTCGTGGCG CAGATAATTA AATACCGCGC TCGTCGGCAC
 ~~~~~  
 35 251 · T G V P A R F S G S G S G T D F  
 CAACTGGGGT CCCGGCGCGT TTAGCGGCT CTGGATCCGG CACGGATTTT  
 GTTGACCCCA GGGCCGCGCA AAATCGCCGA GACCTAGGCC GTGCCTAAAA  
 ~~~~~  
 40 BbsI
 ~~~~~  
 301 T L T I S S L E P E D F A V Y Y C ·  
 ACCCTGACCA TTAGCAGCCT GGAACCTGAA GACTTTGCGG TGTATTATTG  
 TGGGACTGGT AATCGTCGGA CCTTGGACTT CTGAAACGCC ACATAATAAC  
 ~~~~~  
 45 MscI
 ~~~~~  
 50 351 · Q Q H Y T T P P T F G Q G T K V E ·  
 CCAGCAGCAT TATACCACCC CGCCGACCTT TGGCCAGGGT ACGAAAGTTG  
 GGTCTGTCGTA ATATGGTGGG GCGGCTGGAA ACCGGTCCCA TGCTTTCAAC  
 ~~~~~  
 55 CL kappa
 ~~~~~  
 401 BsiWI  
 ~~~~~  
 60 · I K R T V A A P S V F I F P P S
 AAATTAACG TACGGTGGCT GCTCCGAGCG TGTTTATTTT TCCGCCGAGC
 TTTAATTGTC ATGCCACCGA CGAGGCTCGC ACAAATAAAA AGGCGGCTCG

CL kappa

451 DEQLKSGTASVVCLLNN·
GATGAACAAC TGAAAAGCGG CACGGCGAGC GTGGTGTGCC TGCTGAACAA
5 CTACTTGTG ACTTTTCGCC GTGCCGCTCG CACCACACGG ACGACTTGT

CL kappa

501 ·FYPREAKVQWKVDNALQ·
CTTTTATCCG CGTGAAGCGA AAGTTCAGTG GAAAGTAGAC AACGCGCTGC
10 GAAAATAGGC GCACTTCGCT TTCAAGTCAC CTTTCATCTG TTGCGCGACG

CL kappa

551 ·SGNSQESVTEQDSKDS
AAAGCGCAA CAGCCAGGAA AGCGTGACCG AACAGGATAG CAAAGATAGC
15 TTTCGCCGTT GTCGGTCCTT TCGCACTGGC TTGTCCTATC GTTCTATCG

CL kappa

601 TYSLSSTLTLSKADYEK·
ACCTATTCTC TGAGCAGCAC CCTGACCCTG AGCAAAGCGG ATTATGAAAA
20 TGGATAAGAG ACTCGTCGTG GCACTGGGAC TCGTTTCGCC TAATACTTTT

CL kappa

651 ·HKVYACEVTHQGLSSPV·
ACATAAAGTG TATGCGTGCG AAGTGACCCA TCAAGGTCTG AGCAGCCCGG
25 TGTATTTTAC ATACGCACGC TTCACTGGGT AGTTCAGAC TCGTCGGGCC

CL kappa

StuI SphI

701 ·TKSFNRGEA
TGACTAAATC TTTTAATCGT GGCGAGGCCT GATAAGCATG CGTAGGAGAA
30 ACTGATTTAG AAAATTAGCA CCGCTCCGGA CTATTCGTAC GCATCCTCT

phoA

SapI

751 MKQSTIALALLPLLF·
AATAAAATGA AACAAAGCAC TATTGCACTG GCACTCTTAC CGTTGCTCTT
40 TTATTTTACT TTGTTTCGTG ATAACGTGAC CGTGAGAATG GCAACGAGAA

VH3

phoA

SapI MfeI

801 ·TPVTKAQVQLVESGGGL·
CACCCCTGTT ACCAAAGCCG AAGTGCAATT GGTGGAAAGC GGCGGCGGCC
GTGGGGACAA TGGTTTCGGC TTCACGTAA CCACCTTTCG CCGCCGCCG

VH3

851 ·VQPGGSLRLSCAASGF
TGGTGCAACC GGGCGGCAGC CTGCGTCTGA GCTGCGCGGC CTCCGGATT
ACCACGTTGG CCCGCCGTCG GACGCAGACT CGACGCGCCG GAGGCCTAAA

VH3

901 TFS SYAMSWVRQAPGKG·
ACCTTTAGCA GCTATGCGAT GAGCTGGGTG CGCCAAGCCC CTGGGAAGGG
TGGAAATCGT CGATACGCTA CTCGACCCAC GCGGTTTCGGG GACCCTTCCC

VH3

951 · L E W V S A I S G S G G S T Y Y A ·
 TCTCGAGTGG GTGAGCGCGA TTAGCGGTAG CGGCGGCAGC ACCTATTATG
 AGAGCTCACC CACTCGCGCT AATCGCCATC GCCGCCGTCG TGGATAATAC
 VH3
 5 ~~~~~
 PmlI
 ~~~~~  
 1001 · D S V K G R F T I S R D N S K N  
 CGGATAGCGT GAAAGGCCGT TTTACCATTT CACGTGATAA TTCGAAAAAC  
 GCCTATCGCA CTTTCCGGCA AAATGGTAAA GTGCACTATT AAGCTTTTGT  
 VH3  
 10 ~~~~~  
 1051 T L Y L Q M N S L R A E D T A V Y ·  
 ACCCTGTATC TGCAAATGAA CAGCCTGCGT GCGGAAGATA CGGCCGTGTA  
 TGGGACATAG ACGTTTACTT GTCGGACGCA CGCCTTCTAT GCCGGCACAT  
 VH3  
 15 ~~~~~  
 BssHII  
 ~~~~~  
 20 · Y C A R W G G D G F Y A M D Y W G ·
 1101 TTATTGCGCG CGTTGGGGCG GCGATGGCTT TTATGCGATG GATTATTGGG
 AATAACGCGC GCAACCCCGC CGCTACCGAA AATACGCTAC CTAATAACCC
 CH1
 25 ~~~~~
 VH3
 ~~~~~  
 SalI  
 ~~~~~  
 StyI BlnI
 ~~~~~                      ~~~~~  
 30 · Q G T L V T V S S A S T K G P S  
 1151 GCCAAGGCAC CCTGGTGACG GTTAGCTCAG CGTCGACCAA AGGTCCAAGC  
 CGGTTCCGTG GGACCACTGC CAATCGAGTC GCAGCTGGTT TCCAGGTTTC  
 CH1  
 35 ~~~~~  
 1201 V F P L A P S S K S T S G G T A A ·  
 GTGTTTCCGC TGGCTCCGAG CAGCAAAAGC ACCAGCGGCG GCACGGCTGC  
 CACAAAGGCG ACCGAGGCTC GTCGTTTTCG TGGTCGCCGC CGTGCCGACG  
 CH1  
 40 ~~~~~  
 1251 · L G C L V K D Y F P E P V T V S W ·  
 CCTGGGCTGC CTGGTTAAAG ATTATTTCCC GGAACCAGTC ACCGTGAGCT  
 GGACCCGACG GACCAATTTC TAATAAAGGG CCTTGGTCAG TGGCACTCGA  
 CH1  
 45 ~~~~~  
 1301 · N S G A L T S G V H T F P A V L  
 GGAACAGCGG GGCGCTGACC AGCGGCGTGC ATACCTTTCC GGCGGTGCTG  
 CCTTGTCGCC CCGCGACTGG TCGCCGCACG TATGGAAAGG CCGCCACGAC  
 CH1  
 50 ~~~~~  
 1351 Q S S G L Y S L S S V V T V P S S ·  
 CAAAGCAGCG GCCTGTATAG CCTGAGCAGC GTTGTGACCG TGCCGAGCAG  
 GTTTCGTCGC CGGACATATC GGAATCGTCG CAACACTGGC ACGGCTCGTC  
 CH1  
 55 ~~~~~  
 1401 · S L G T Q T Y I C N V N H K P S N ·  
 CAGCTTAGGC ACTCAGACCT ATATTTGCAA CGTGAACCAT AAACCGAGCA  
 GTCGAATCCG TGAGTCTGGA TATAAACGTT GCACTTGGTA TTTGGCTCGT  
 CH1                      CTgIII  
 60 ~~~~~

EcoRI  
~~~~~

5 1451 · T K V D K K V E P K S E F G G G
ACACCAAAGT GGATAAAAAA GTGGAACCGA AAAGCGAATT CGGGGGAGGG
TGTGGTTTCA CCTATTTTTT CACCTTGGCT TTTCGCTTAA GCCCCCTCCC
CTgIII
~~~~~

10 1501 S G S G D F D Y E K M A N A N K G ·  
AGCGGGAGCG GTGATTTTGA TTATGAAAAG ATGGCAAACG CTAATAAGGG  
TCGCCCTCGC CACTAAAACT AATACTTTTC TACCGTTTGC GATTATTCCC  
CTgIII  
~~~~~

15 1551 · A M T E N A D E N A L Q S D A K G ·
GGCTATGACC GAAAATGCCG ATGAAAACGC GCTACAGTCT GACGCTAAAG
CCGATACTGG CTTTACGGC TACTTTTGCG CGATGTCAGA CTGCGATTTC
CTgIII
~~~~~

20 1601 · K L D S V A T D Y G A A I D G F  
GCAAACCTGA TTCTGTCGCT ACTGATTACG GTGCTGCTAT CGATGGTTTC  
CGTTTGAAC T AAGACAGCGA TGACTAATGC CACGACGATA GCTACCAAAG  
CTgIII  
~~~~~

25 1651 I G D V S G L A N G N G A T G D F ·
ATTGGTGACG TTTCCGGCCT TGCTAATGGT AATGGTGCTA CTGGTGATT
TAACCACTGC AAAGGCCGGA ACGATTACCA TTACCACGAT GACCACTAAA
CTgIII
~~~~~

30 1701 · A G S N S Q M A Q V G D G D N S P ·  
TGCTGGCTCT AATTCCCAA TGGCTCAAGT CGGTGACGGT GATAATTCAC  
ACGACCGAGA TTAAGGGTTT ACCGAGTTCA GCCACTGCCA CTATTAAGTG  
CTgIII  
~~~~~

35 1751 · L M N N F R Q Y L P S L P Q S V
CTTTAATGAA TAATTCCGT CAATATTTAC CTTCCTCCC TCAATCGGT
GAAATTACTT ATTAAGGCA GTTATAAATG GAAGGGAGGG AGTTAGCCAA
CTgIII
~~~~~

40 1801 E C R P F V F G A G K P Y E F S I ·  
GAATGTCGCC CTTTTGTCTT TGGCGCTGGT AAACCATATG AATTTTCTAT  
CTTACAGCGG GAAAACAGAA ACCGCGACCA TTTGGTATAC TTAAAAGATA  
CTgIII  
~~~~~

45 1851 · D C D K I N L F R G V F A F L L Y ·
TGATTGTGAC AAAATAAACT TATTCCGTGG TGTCTTTGCG TTTCTTTTAT
ACTAACACTG TTTTATTGA ATAAGGCACC ACAGAAACGC AAAGAAAATA
CTgIII
~~~~~

50 1901 · V A T F M Y V F S T F A N I L R  
ATGTTGCCAC CTTTATGTAT GTATTTTCTA CGTTTGCTAA CATACTGCGT  
TACAACGGTG GAAATACATA CATAAAAGAT GCAAACGATT GTATGACGCA  
CTgIII  
~~~~~

55 Stop lpp terminator
~~~~~  
HindIII  
~~~~~

60 1951 N K E S
AATAAGGAGT CTTGATAAGC TTGACCTGTG AAGTGAAAAA TGGCGCAGAT
TTATTCCTCA GAACTATTCG AACTGGACAC TTCACTTTT ACCGCGTCTA
lpp terminator

~~~~~  
2001 TGTGCGACAT TTTTTTTGTC TGCCGTTTAA TGAAATTGTA AACGTTAATA  
ACACGCTGTA AAAAAAACAG ACGGCAAAATT ACTTTAACAT TTGCAATTAT  
~~~~~  
5 fl origin
2051 TTTTGTAAAA ATTTCGCGTTA AATTTTTGTT AAATCAGCTC ATTTTTTAAC
AAAACAATTT TAAGCGCAAT TAAAAACAA TTTAGTCGAG TAAAAAATTG
~~~~~  
10 fl origin  
2101 CAATAGGCCG AAATCGGCAA AATCCCTTAT AAATCAAAAG AATAGACCGA  
GTTATCCGGC TTTAGCCGTT TTAGGGAATA TTTAGTTTTC TTATCTGGCT  
~~~~~  
15 fl origin
2151 GATAGGGTTG AGTGTGTGTT CAGTTTGGAA CAAGAGTCCA CTATTAAAGA
CTATCCCAAC TCACAACAAG GTCAAACCTT GTTCTCAGGT GATAATTTCT
~~~~~  
20 fl origin  
2201 ACGTGGACTC CAACGTCAAA GGGCGAAAAA CCGTCTATCA GGGCGATGGC  
TGCACCTGAG GTTGCAGTTT CCCGCTTTTT GGCAGATAGT CCCGCTACCG  
~~~~~  
25 T->A
~
2251 CCACTACGAG AACCATCACC CTAATCAAGT TTTTGGGGT CGAGGTGCCG
GGTGATGCTC TTGGTAGTGG GATTAGTTCA AAAAACCCCA GCTCCACGGC
~~~~~  
30 fl origin  
2301 TAAAGCACTA AATCGGAACC CTAAAGGGAG CCCCCGATTT AGAGCTTGAC  
ATTTCGTGAT TTAGCCTTGG GATTTCCTC GGGGGCTAAA TCTCGAACTG  
~~~~~  
35 fl origin
2351 GGGGAAAGCC GGCGAACGTG GCGAGAAAGG AAGGGAAGAA AGCGAAAGGA
CCCCTTTCGG CCGCTTGCAC CGCTCTTTC TTCCCTTCTT TCGCTTTCCT
~~~~~  
40 fl origin  
2401 GCGGGCGCTA GGGCGCTGGC AAGTGTAAGC GTCACGCTGC GCGTAACCAC  
CGCCGCGAT CCCGCGACCG TTCACATCGC CAGTGCAGC CGCATTGGTG  
~~~~~  
45 fl origin
2451 CACACCCGCC GCGCTTAATG CGCCGCTACA GGGCGCGTGC TAGCCATGTG
GTGTGGGCGG CGCGAATTAC GCGGCGATGT CCCGCGCACG ATCGGTACAC
~~~~~  
50 fl origin ColEI  
2501 AGCAAAAGGC CAGCAAAAGG CCAGGAACCG TAAAAAGGCC GCGTTGCTGG  
TCGTTTTCCG GTCGTTTTC GGTCTTGGC ATTTTCCGG CGCAACGACC  
~~~~~  
55 ColEI
2551 CGTTTTTCCA TAGGCTCCGC CCCCTGACG AGCATCACAA AAATCGACGC
GCAAAAAGGT ATCCGAGGCG GGGGACTGC TCGTAGTGTT TTAGCTGCG
~~~~~  
60 ColEI  
2601 TCAAGTCAGA GGTGGCGAAA CCCGACAGGA CTATAAAGAT ACCAGGCGTT  
AGTTCAGTCT CCACCGCTTT GGGCTGTCT GATATTCTA TGGTCCGCAA  
~~~~~  
65 ColEI
2651 TCCCCCTGGA AGCTCCCTCG TGCCTCTCC TGTTCGACC CTGCCGCTTA

AGGGGGACCT TCGAGGGAGC ACGCGAGAGG ACAAGGCTGG GACGGCGAAT
~~~~~  
ColEI  
mutation  
5  
2701 CCGGATACCT GTCCGCCTTT CTCCCTTCGG GAAGCGTGGC GCTTTCTCAT  
GGCCTATGGA CAGGCGGAAA GAGGGAAGCC CTCGCACCG CGAAAGAGTA  
~~~~~  
ColEI
10 mutation
2751 AGCTCACGCT GTAGGTATCT CAGTTCGGTG TAGGTCGTTT GCTCCAAGCT
TCGAGTGCGA CATCCATAGA GTCAAGCCAC ATCCAGCAAG CGAGGTTCTGA
~~~~~  
ColEI  
15 mutation  
2801 GGGCTGTGTG CACGAACCCC CCGTTCAGTC CGACCGCTGC GCCTTATCCG  
CCCGACACAC GTGCTTGGGG GGCAAGTCAG GCTGGCGACG CGGAATAGGC  
20 ~~~~~  
ColEI  
2851 GTAACATCG TCTTGAGTCC AACCCGGTAA GACACGACTT ATCGCCACTG  
CATTGATAGC AGAACTCAGG TTGGGCCATT CTGTGCTGAA TAGCGGTGAC  
~~~~~  
ColEI
25 2901 GCAGCAGCCA CTGGTAACAG GATTAGCAGA GCGAGGTATG TAGGCGGTGC
CGTCGTCGGT GACCATTGTC CTAATCGTCT CGCTCCATAC ATCCGCCACG
~~~~~  
ColEI  
30 mutation  
2951 TACAGAGTTC TTGAAGTGGT GGCCTAACTA CGGCTACACT AGAAGAACAG  
ATGTCTCAAG AACTTCACCA CCGGATTGAT GCCGATGTGA TCTTCTTGTC  
~~~~~  
ColEI
35 mutation
3001 TATTTGGTAT CTGCGCTCTG CTGTAGCCAG TTACCTTCGG AAAAAGAGTT
ATAAACCATA GACGCGAGAC GACATCGGTC AATGGAAGCC TTTTCTCAA
40 ~~~~~
ColEI
3051 GGTAAGCTCTT GATCCGGCAA ACAAACCACC GCTGGTAGCG GTGGTTTTTT
CCATCGAGAA CTAGGCCGTT TGTTTGGTGG CGACCATCGC CACCAAAAAA
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ColEI  
45 3101 TGTTTGCAAG CAGCAGATTA CGCGCAGAAA AAAAGGATCT CAAGAAGATC  
ACAAACGTTT GTCGTCTAAT GCGCGTCTTT TTTTCTAGA GTTCTTCTAG  
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ColEI
50 3151 CTTTGATCTT TTCTACGGGG TCTGACGCTC AGTGGAACGA AAATCAGCT
GAAACTAGAA AAGATGCCCC AGACTGCGAG TCACCTTGCT TTTGAGTGCA
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ColEI  
55 cat terminator  
BglII  
3201 TAAGGGATTT TGGTCAGATC TAGCACCAGG CGTTTAAGGG CACCAATAAC  
ATTCCCTAAA ACCAGTCTAG ATCGTGGTCC GCAAATTCCC GTGGTTATTG  
60 ~~~~~

ColEI  
cat terminator

5 3251 TGCCTTAAAA AAATTACGCC CCGCCCTGCC ACTCATCGCA GTACTGTTGT  
ACGGAATTTT TTTAATGCGG GGCGGGACGG TGAGTAGCGT CATGACAACA

10 3301 CM(R)  
AATTCATTAA GCATTCTGCC GACATGGAAG CCATCACAAA CGGCATGATG  
TTAAGTAATT CGTAAGACGG CTGTACCTTC GGTAAGTGTTC GCCGTACTAC

15 3351 CM(R)  
AACCTGAATC GCCAGCGGCA TCAGCACCTT GTCGCCTTGC GTATAATATT  
TTGGACTTAG CGGTCGCCGT AGTCGTGGAA CAGCGGAACG CATATTATAA

20 3401 CM(R)  
TGCCCATAGT GAAAACGGGG GCGAAGAAGT TGTCCATATT GGCTACGTTT  
ACGGGTATCA CTTTGGCCCC CGCTTCTTCA ACAGGTATAA CCGATGCAAA

25 3451 CM(R)  
AAATCAAAAC TGGTGAAACT CACCCAGGGA TTGGCTGAGA CGAAAAACAT  
TTAGTTTTGT ACCACTTTGA GTGGGTCCCT AACCGACTCT GCTTTTGTGA

30 3501 CM(R)  
ATTCTCAATA AACCTTTAG GGAAATAGGC CAGGTTTTCA CCGTAACACG  
TAAGAGTTAT TTGGGAAATC CCTTTATCCG GTCCAAAAGT GGCATTGTGC

35 3551 CM(R)  
CCACATCTTG CGAATATATG TGTAGAAACT GCCGGAAATC GTCGTGGTAT  
GGTGTAGAAC GCTTATATAC ACATCTTTGA CGGCCTTTAG CAGCACCATA

40 3601 CM(R)  
TCACTCCAGA GCGATGAAAA CGTTTCAGTT TGCTCATGGA AAACGGTGTA  
AGTGAGGTCT CGCTACTTTT GCAAAGTCAA ACGAGTACCT TTTGCCACAT

45 3651 CM(R)  
ACAAGGGTGA AACTATCCC ATATCACCAG CTCACCGTCT TTCATTGCCA  
TGTTCCCACT TGTGATAGGG TATAGTGGTC GAGTGGCAGA AAGTAACGGT

50 3701 CM(R)  
TACGGAATC CGGGTGAGCA TTCATCAGGC GGGCAAGAAT GTGAATAAAG  
ATGCCTTGAG GCCCACTCGT AAGTAGTCCG CCCGTTCTTA CACTATTTC

55 3751 CM(R)  
GCCGGATAAA ACTTGTGCTT ATTTTCTTT ACGGTCTTTA AAAAGGCCGT  
CGGCCTATTT TGAACACGAA TAAAAAGAAA TGCCAGAAAT TTTCCGGCA

60 3801 CM(R)  
AATATCCAGC TGAACGGTCT GGTATAGGT ACATTGAGCA ACTGACTGAA  
TTATAGGTCG ACTTGCCAGA CCAATATCCA TGTAACCTCGT TGACTGACTT

3851 CM(R)  
ATGCCTCAAA ATGTTCTTTA CGATGCCATT GGGATATATC AACGGTGGTA  
TACGGAGTTT TACAAGAAAT GCTACGGTAA CCCTATATAG TTGCCACCAT

3901 CM(R)  
TATCCAGTGA TTTTTTCTC CATTTTAGCT TCCTTAGCTC CTGAAAATCT  
ATAGGTCAC TAAAAAAGAG GTAAATCGA AGGAATCGAG GACTTTTAGA

CM(R) SD

cat promoter  
 3951 CGATAACTCA AAAAATACGC CCGGTAGTGA TCTTATTTCA TTATGGTGAA  
 GCTATTGAGT TTTTATGCG GGCCATCACT AGAATAAAGT AATACCACTT  
 ~~~~~  
 5 cat promoter
 CRP site
 ~~~~~  
 4001 AGTTGGAACC TCACCCGACG TCTAATGTGA GTTAGCTCAC TCATTAGGCA  
 TCAACCTTGG AGTGGGCTGC AGATTACACT CAATCGAGTG AGTAATCCGT  
 ~~~~~  
 10 cat promoter
 lac mRNA start
 ~
 lac operator
 ~~~~~  
 15 -35 region -10 region  
 ~~~~~ ~~~~~  
 4051 CCCAGGCTT TACACTTTAT GCTCCGGCT CGTATGTTGT GTGGAATTGT
 GGGGTCCGAA ATGTGAAATA CGAAGGCCGA GCATACAACA CACCTTAACA
 20 lac operator SD lacZ'
 ~~~~~  
 4101 GAGCGGATAA CAATTCACA CAGGAAACAG CTATGACCAT GATTACGAAT  
 CTCGCCTATT GTTAAAGTGT GTCCTTGTC GATACTGGTA CTAATGCTTA  
 lacZ'  
 25 ~  
 4151 T  
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## Framework 1

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**SUBSTITUTE SHEET (RULE 26)**

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SexAI										AseI										SanDI									
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AAA	CCA	GGT	AAA	GCA	CCG	AAA	CTA	TTA	ATT	TAT	GCA	GCC	AGC	AGC	TTG	CAA	AGC	GGG	GTC	CCG	TCC	CGT	TTT	AGC	GGC				
AAA	CCA	GGT	AAA	GCA	CCG	AAA	CTA	TTA	ATT	TAT	GCA	GCC	AGC	AGC	TTG	CAA	AGC	GGG	GTC	CCG	TCC	CGT	TTT	AGC	GGC				
AAA	CCA	GGT	CAA	GCA	CCG	CGT	CTA	TTA	ATT	TAT	GGC	GCG	AGC	AGC	CGT	GCA	ACT	GGG	GTC	CCG	GCG	CGT	TTT	AGC	GGC				
AAA	CCA	GGT	CAA	GCA	CCG	CGT	CTA	TTA	ATT	TAT	GGC	GCG	AGC	AGC	CGT	GCA	ACT	GGG	GTC	CCG	GCG	CGT	TTT	AGC	GGC				
AAA	CCA	GGT	CAA	GCA	CCG	CGT	CTA	TTA	ATT	TAT	GGC	GCG	AGC	AGC	CGT	GCA	ACT	GGG	GTC	CCG	GCG	CGT	TTT	AGC	GGC				
AAA	CCA	GGT	CAG	CCG	CCG	AAA	CTA	TTA	ATT	TAT	TGG	GCA	TCC	ACC	CGT	GAA	AGC	GGG	GTC	CCG	GAT	CGT	TTT	AGC	GGC				
AAA	CCA	GGT	CAG	CCG	CCG	AAA	CTA	TTA	ATT	TAT	TGG	GCA	TCC	ACC	CGT	GAA	AGC	GGG	GTC	CCG	GAT	CGT	TTT	AGC	GGC				
CAT	CCC	GGG	AAG	GCG	CCG	AAA	CTG	ATG	ATT	TAT	GAT	GTG	AGC	AAC	CGT	CCC	TCA	GGC	GTG	AGC	AAC	CGT	TTT	AGC	GGA				
CAT	CCC	GGG	AAG	GCG	CCG	AAA	CTG	ATG	ATT	TAT	GAT	GTG	AGC	AAC	CGT	CCC	TCA	GGC	GTG	AGC	AAC	CGT	TTT	AGC	GGA				
CAT	CCC	GGG	AAG	GCG	CCG	AAA	CTG	ATG	ATT	TAT	GAT	GTG	AGC	AAC	CGT	CCC	TCA	GGC	GTG	AGC	AAC	CGT	TTT	AGC	GGA				
CAT	CCC	GGG	AAG	GCG	CCG	AAA	CTG	ATG	ATT	TAT	GAT	GTG	AGC	AAC	CGT	CCC	TCA	GGC	GTG	AGC	AAC	CGT	TTT	AGC	GGA				
AAA	CCC	GGG	CAG	GCG	CCA	GTT	CTG	GTG	ATT	TAT	GAT	GAT	TCT	GAC	CGT	CCC	TCA	GGC	ATC	CCG	GAA	CGC	TTT	AGC	GGA				
AAA	CCC	GGG	CAG	GCG	CCA	GTT	CTG	GTG	ATT	TAT	GAT	GAT	TCT	GAC	CGT	CCC	TCA	GGC	ATC	CCG	GAA	CGC	TTT	AGC	GGA				
AAA	CCC	GGG	CAG	GCG	CCA	GTT	CTG	GTG	ATT	TAT	GAT	GAT	TCT	GAC	CGT	CCC	TCA	GGC	ATC	CCG	GAA	CGC	TTT	AGC	GGA				
AAA	CCC	GGG	CAG	GCG	CCA	GTT	CTG	GTG	ATT	TAT	GAT	GAT	TCT	GAC	CGT	CCC	TCA	GGC	ATC	CCG	GAA	CGC	TTT	AGC	GGA				
AAA	CCC	GGG	CAG	GCG	CCA	GTT	CTG	GTG	ATT	TAT	GAT	GAT	TCT	GAC	CGT	CCC	TCA	GGC	ATC	CCG	GAA	CGC	TTT	AGC	GGA				

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XhoI																									
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CAG	GGT	CTC	GAG	TGG	ATG	GGC	GGC	ATT	ATT	CCG	-	-	ATT	TTT	GGC	ACG	GCG	AAC	TAC	GCG	CAG	AAG	TTT	CAG	GGC
CAG	GGT	CTC	GAG	TGG	ATG	GGC	GGC	ATT	ATT	CCG	-	-	ATT	TTT	GGC	ACG	GCG	AAC	TAC	GCG	CAG	AAG	TTT	CAG	GGC
CAG	GGT	CTC	GAG	TGG	ATG	GGC	GGC	ATT	ATT	CCG	-	-	ATT	TTT	GGC	ACG	GCG	AAC	TAC	GCG	CAG	AAG	TTT	CAG	GGC
CAG	GGT	CTC	GAG	TGG	ATG	GGC	GGC	ATT	ATT	CCG	-	-	ATT	TTT	GGC	ACG	GCG	AAC	TAC	GCG	CAG	AAG	TTT	CAG	GGC
CAG	GGT	CTC	GAG	TGG	ATG	GGC	TGG	ATT	AAC	CCG	-	-	AAT	AGC	GGC	GGC	ACG	AAC	TAC	GCG	CAG	AAG	TTT	CAG	GGC
CAG	GGT	CTC	GAG	TGG	ATG	GGC	TGG	ATT	AAC	CCG	-	-	AAT	AGC	GGC	GGC	ACG	AAC	TAC	GCG	CAG	AAG	TTT	CAG	GGC
CAG	GGT	CTC	GAG	TGG	ATG	GGC	TGG	ATT	AAC	CCG	-	-	AAT	AGC	GGC	GGC	ACG	AAC	TAC	GCG	CAG	AAG	TTT	CAG	GGC
CAG	GGT	CTC	GAG	TGG	ATG	GGC	TGG	ATT	AAC	CCG	-	-	AAT	AGC	GGC	GGC	ACG	AAC	TAC	GCG	CAG	AAG	TTT	CAG	GGC
CAG	GGT	CTC	GAG	TGG	ATG	GGC	TGG	ATT	AAC	CCG	-	-	AAT	AGC	GGC	GGC	ACG	AAC	TAC	GCG	CAG	AAG	TTT	CAG	GGC
CAG	GGT	CTC	GAG	TGG	ATG	GGC	TGG	ATT	AAC	CCG	-	-	AAT	AGC	GGC	GGC	ACG	AAC	TAC	GCG	CAG	AAG	TTT	CAG	GGC
AAA	GCC	CTC	GAG	TGG	ATG	GGC	TGG	ATT	AAC	CCG	-	-	AAT	AGC	GGC	GGC	ACG	AAC	TAT	AGC	ACC	AGC	CTG	AAA	ACG
AAA	GCC	CTC	GAG	TGG	CTG	GCT	CTG	ATT	GAT		-	-	TGG	GAT	GAT	GAT	AAG	TAT	TAT	AGC	ACC	AGC	CTG	AAA	ACG
CGT	GGC	CTC	GAG	TGG	CTG	GGC	CGT	ACC	TAT	TAT	CGT	-	AGC	AAA	TGG	TAT	AAC	GAT	TAT	GCG	GTG	AGC	GTG	AAA	ACG
CGT	GGC	CTC	GAG	TGG	CTG	GGC	CGT	ACC	TAT	TAT	CGT	-	AGC	AAA	TGG	TAT	AAC	GAT	TAT	GCG	GTG	AGC	GTG	AAA	ACG

[illegible]

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9	8	7	6	5	4	3	2	1	0	9	8	7	6	5	4	3	2	1	0	9	8	7	6	5	4	3	2	1	0
x	CAG	x	x	x	x	x	x	x	x	x	CAG	x	x	x	x	x	x	x	x	x	CAG	x	x	x	x	x	x	x	x
TTT	CAG	TAT	GGT	TCT	ATT	CCT	CCT			TTT	CAG	TAT	GGT	TCT	ATT	CCT	CCT			ACG	GCC	GTG	TAT	TAT	TAT	TAT	TAT	TAT	TAT
x	CAG	x	x	x	x	x	x	x	x	x	CAG	x	x	x	x	x	x	x	x	ACG	GCC	GTG	TAT	TAT	TAT	TAT	TAT	TAT	TAT
CAG	CAG	ATG	TCT	AAT	TAT	CCT	GAT			CAG	CAG	ATG	TCT	AAT	TAT	CCT	GAT			ACG	GCC	GTG	TAT	TAT	TAT	TAT	TAT	TAT	TAT
CAG	CAG	ACT	AAT	AAT	GCT	CCT	GTT			CAG	CAG	ACT	AAT	AAT	GCT	CCT	GTT			ACG	GCC	GTG	TAT	TAT	TAT	TAT	TAT	TAT	TAT
x	CAG	x	x	x	x	x	x	x	x	x	CAG	x	x	x	x	x	x	x	x	ACG	GCC	GTG	TAT	TAT	TAT	TAT	TAT	TAT	TAT
CAG	CAG	TAT	GAT	TCT	ATT	CCT	TAT			CAG	CAG	TAT	GAT	TCT	ATT	CCT	TAT			ACG	GCC	GTG	TAT	TAT	TAT	TAT	TAT	TAT	TAT
CAG	x	x	GAC	x	x	(x)	(x)	(x)	(x)	CAG	x	x	GAC	x	x	(x)	(x)	(x)	(x)	ACG	GCC	GTG	TAT	TAT	TAT	TAT	TAT	TAT	TAT
CAG	AGC	TAT	GAC	ATG	TAT	AAT	TAT	ATT		CAG	AGC	TAT	GAC	ATG	TAT	AAT	TAT	ATT		ACG	GCC	GTG	TAT	TAT	TAT	TAT	TAT	TAT	TAT
CAG	TCT	CAT		CAT	TTT	TAT	GAG			CAG	TCT	CAT		CAT	TTT	TAT	GAG			ACG	GCC	GTG	TAT	TAT	TAT	TAT	TAT	TAT	TAT
CAG	AGC	TAT	GAC	AAT	AAT	TCT	GAT	GTT		CAG	AGC	TAT	GAC	AAT	AAT	TCT	GAT	GTT		ACG	GCC	GTG	TAT	TAT	TAT	TAT	TAT	TAT	TAT
CAG	x	x	GAC	x	x	(x)	(x)	(x)	(x)	CAG	x	x	GAC	x	x	(x)	(x)	(x)	(x)	ACG	GCC	GTG	TAT	TAT	TAT	TAT	TAT	TAT	TAT
CAG	AGC	TAT	GAC	TAT	TTT	AAG	CTT	x		CAG	AGC	TAT	GAC	TAT	TTT	AAG	CTT	x		ACG	GCC	GTG	TAT	TAT	TAT	TAT	TAT	TAT	TAT
CAG	AGC	TAT	GAC	TAT	TCT	GCT	GAT	TAT		CAG	AGC	TAT	GAC	TAT	TCT	GCT	GAT	TAT		ACG	GCC	GTG	TAT	TAT	TAT	TAT	TAT	TAT	TAT
CAG	AGC	TAT	GAC	TTT	GAT	TTT	GCT			CAG	AGC	TAT	GAC	TTT	GAT	TTT	GCT			ACG	GCC	GTG	TAT	TAT	TAT	TAT	TAT	TAT	TAT
CAG	AGC	TAT	GAC	GGT	CCT	GAT	CTT	TGG		CAG	AGC	TAT	GAC	GGT	CCT	GAT	CTT	TGG		ACG	GCC	GTG	TAT	TAT	TAT	TAT	TAT	TAT	TAT

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x	x	x	GAT	x	TGG	GSC	styl			ACC	CTG	GTG	ACG	GTT	BlpI																																				
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