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(54) Title: DIAGNOSTIC MICROSPHERES

(57) Abstract: The invention provides a diagnostic composition for detecting both aspiration and gastroesophageal reflux comprising bio-degradable microspheres having a diameter of about 0.1-10 microns.



WO 2004/011035 A2

DIAGNOSTIC MICROSPHERES

The present invention relates to a diagnostic marker for the diagnosis of aspiration and gastroesophageal reflux in children and adults, to the manufacture of such markers and the use thereof.

Aspiration into the airways is one of the major causes of lung disease in infants and young children. Unfortunately the diagnosis of aspiration is often delayed due to unawareness and to the low sensitivity and specificity of existing diagnostic tests.

Aspiration of food or gastric material into the tracheobronchial tree can result in a variety of disease states such as laryngeal and tracheo-bronchial inflammation leading to stridor, cough, wheezing, recurrent pneumonia, bronchiectasis, pulmonary fibrosis and even suffocation and death due to massive aspiration (Mendelson CL. The aspiration of stomach contents into the lungs during obstetric anesthesia. *Am J Obstet Gynecol* 1946;52:191-205; Bauer ML, Figueroa-Colon, Georgeson K, Young DW. Chronic pulmonary aspiration in children. *South Med J* 1993; 86:789-795; Moran TJ. Experimental aspiration pneumonia. *Arch Pathol* 1955;60:122-129)

The common conditions that predispose to aspiration in infants and young children include prematurity due to developmental delay of the coordination between swallowing and breathing (Harding R, Johnson P, McClelland ME. Liquid sensitive laryngeal receptors in the developing ship cat and monkey. *J Physiol Lond* 1978;277:409-422), abnormal communication between airways and esophagus such as laryngo-tracheal cleft or tracheo-esophageal fistula, massive gastro-esophageal reflux, and neurological abnormalities such as cerebral palsy, vocal cord paralysis and familial dysautonomia.

The available tests for the diagnosis of aspiration such as barium swallow during videofluoroscopy and gastroesophageal scintigraphy are not sensitive enough (McVeagh P, Howman-Giles R, Kemp A. Pulmonary aspiration studied by radionuclide milk scanning and barium swallow roentgenography. *Am J Dis Child* 1987;141:917-921). In 1985, Corwin and Irwin (Corwin RW, Irwin RS. The lipid-laden alveolar macrophage as a marker of aspiration in parenchymal lung disease. *Am Rev Respir Dis* 1985;132:576-581) introduced the Oil-Red-O staining of lipid laden alveolar macrophages (LLAM) as a marker for aspiration. More recently, others have noticed that this test is not specific for aspiration and that other conditions can also

result in an increase in fat vacuoles inside alveolar macrophages (Kajetanowicz A, Stinson D, Laybolt KS, Resch L. Lipid-laden macrophages in tracheal aspirate of ventilated neonates receiving intralipid. *Pediatr Pulmonol* 1999; 28:101-108; Knauer-Fisher S, Ratjen F. Lipid-laden macrophages in bronchoalveolar lavage fluid as a marker for pulmonary aspiration. *Pediatr Pulmonol* 1999;27:419-422; Vichinsky E, Williams R, Das M, Earles AN, Lewis N, Adler A, McQuintty J. Pulmonary fat embolism: a distinct cause of severe acute chest syndrome in sickle cell anemia. *Blood* 1994;11:3107-3122).

Elidemir and his colleagues (Elidemir O, Fan LL, Colasurdo N. A novel diagnostic method for pulmonary aspiration in a murine model. Immunocytochemical staining of milk proteins in alveolar macrophages. *Am J Respir Crit Care Med* 2000;161:622-626) have recently shown that aspiration of milk in mice could be diagnosed by specific immunocytochemical staining of milk proteins in bronchoalveolar macrophages. Although this method seems to be specific for the detection of milk, aspiration of other food substances cannot be detected by this method. Furthermore they stained milk proteins (α -lactalbumin and β -lactoglobulin) in alveolar macrophages by an immunocytochemical method. They found that their method is very sensitive and very specific as compared to Oil-Red O staining, but the stainings were positive only for the 3-4 days following induced aspiration. Aspiration can occur from above as in children with neurologic impairment, but may be secondary to gastro-esophageal reflux and may occur only during the massive episodes and not necessarily every day.

It is therefore an object of the present invention to provide a sensitive and specific marker that could help diagnose aspiration that may have occurred days or weeks previously. As will be described and explained hereinafter the marker of the present invention is one that is safe and bio-degradable, can be given orally with food, can easily be "swallowed" by alveolar macrophages and can be identified in bronchoalveolar lavage (BAL) alveolar macrophages for a substantial period of time.

In a recent study (Avital A, Sherman Y, Springer C. Charcoal is a sensitive, specific and stable marker for the diagnosis of aspiration in hamsters. *Pediatr Res* 51:1-5, 2002) the present inventors used activated charcoal particles mixed with milk or normal saline and performed tracheal instillation in hamsters. Charcoal particles were easily identified in BAL for the three months following a single

instillation, although hypercellularity was found in BAL after three months and it was attributed to the fact that some of the particles were large and may have induced airway inflammation.

As will be realized, charcoal particles were not suitable and would not be acceptable as a diagnostic tool for humans due to the danger inherent in introducing charcoal particles into the lungs.

With the above state of the art in mind, it is clear that there is a need for a safe diagnostic marker to detect both aspiration and gastroesophageal reflux for populations at risk such as the young pediatric and the geriatric population as well as those who suffer from neurological deficit leading to swallowing disturbances. As will be realized these groups are prone to aspiration of food to the lungs, which can result in severe lung damage and even death.

Thus according to the present invention there is now provided a diagnostic composition for detecting both aspiration and gastroesophageal reflux comprising bio-degradable microspheres having a diameter of about 0.1 - 10 microns.

In preferred embodiments of the present invention said microspheres have a diameter of about 1 - 4 microns.

In especially preferred embodiments of the present invention there are provided diagnostic composition for detecting both aspiration and gastroesophageal reflux comprising bio-degradable polymeric microspheres having a diameter of about 0.1 - 10 microns.

Any polymer, which is degradable via a hydrolytic and /or enzymatic mechanism, may be suitable. Polymers which undergo degradation under these conditions are, among others, polyesters, polyphosphate esters, polyphosphazenes, polyorthoesters, polyanhydrides, polycarbonates, polyamides and proteins (polypeptides). Typical polyesters are e.g. homopolymers or different copolymers of lactic acid (or lactide), glycolic acid (or glycolide), mandelic acid, caprolactone, α -hydroxy acids (e.g. hydroxy butyric acid). Polymers containing optically active monomers, may consist of only d- or l- monomers, or any combination of the two. By varying the chemical composition and/or the molecular weight of the polymer, it is possible to control its degradation rate.

For any specific polymer or copolymer, different solvents, non-solvents and surfactants may be suitable in the preparation process thereof.

In another aspect of the present invention there is provided a food product containing a diagnostic composition for detecting both aspiration and gastroesophageal reflux said composition comprising bio-degradable microspheres having a diameter of about 0.1 - 10 microns.

Especially preferred for use in the present invention are microspheres of polylactic acid.

As will be described hereinafter, according to the present invention there was performed tracheal instillation of small (2.1 μ) and uniform polylactic acid microspheres suspended in a saline solution in hamsters, compared it to tracheal instillation of normal saline alone and followed for BAL cytology, microsphere index and lung histology during the following three months.

While the invention will now be described in connection with certain preferred embodiments in the following examples so that aspects thereof may be more fully understood and appreciated, it is not intended to limit the invention to these particular embodiments. On the contrary, it is intended to cover all alternatives, modifications and equivalents as may be included within the scope of the invention as defined by the appended claims. Thus, the following examples which include preferred embodiments will serve to illustrate the practice of this invention, it being understood that the particulars shown are by way of example and for purposes of illustrative discussion of preferred embodiments of the present invention only and are presented in the cause of providing what is believed to be the most useful and readily understood description of formulation procedures as well as of the principles and conceptual aspects of the invention.

Example 1. Procedure of PLA microspheres preparation by precipitation with a non-solvent:

0.5 g of d,l- PLA (polylactic acid – Boehringer), of a molecular weight of either 2000 or 30000, were dissolved in 42 ml of acetone (Frutarom). 100 ml of distilled water were added dropwise, over a period of 60 minutes, under mild magnetic stirring. Spherical particles of PLA, 1–2 microns in diameter, are formed at an early stage in the process by precipitation. The acetone and part of the distilled water, were then evaporated by using a rotary evaporator (Buchi), at 40°C. At this stage, a surfactant (Pluronic F-68) is added to the suspension, at a concentration of 1.5%, in order to prevent aggregation and caking of particles following precipitation. It is

worth mentioning that different surfactants and surfactant concentrations may be suitable for this purpose. Also, the method is tolerant to relatively large changes in the amount of both the acetone and distilled water.

Although the microspheres are easily visible by light microscopy, both with and without a polarizer, the possibility of coloring the microspheres was explored. The colorants, which are added to the acetone solution, should dissolve in acetone and be relatively hydrophobic, in order to be included inside the particles during the precipitation process. Alternatively, another indicator, such as a fluorescent material may be added.

Methods:

Animals: give number female Syrian hamsters (*Mesocricetus auratus*) 9-12 weeks old, weighing 100-140 g and obtained from Harlan Sprague Dawley (Indianapolis, USA), were used for the experiments. The study was approved by the local Institutional Animal Care and Use Committee.

give number hamsters were used as naive controls (without tracheal instillation).

give number hamsters had 0.1 ml tracheal instillation of either sterile normal saline (n=18) or white polylactic acid microspheres (n=24) of uniform size (2.1 μ). On days 1, 3, 10, 32, 60 and 90 after instillation, the animals were sacrificed (saline, n=3 and microspheres, n=4) and BAL was performed.

All microspheres were efficiently suspended (10mg/ml) within sterile normal saline by sonification as shown by light microscopy.

Anesthesia: A mixture of ketamine HCl (50 mg/ml, 10 ml) and dehydrobenzperidol (2.5 mg/ml, 2 ml) was administered by intraperitoneal injection, 0.25 ml before tracheal instillation, and 0.4 ml before exsanguination and BAL.

Tracheal instillation: Direct intubation was performed with a blunted metal needle. A few ventilations with a small ambu bag were given to ascertain intratracheal position of the needle by confirming chest movement. Normal saline or microsphere suspension (0.1 ml) was slowly instilled into the trachea, followed by a few ventilations with the ambu bag to disperse the fluid and to prevent apnea.

Bronchoalveolar lavage: On the planned BAL day, under anesthesia, exsanguination was performed by transection of the abdominal aorta. The trachea was exposed and a blunted needle covered with a polyethylene cannula was

inserted into the trachea. Three aliquots of 5 ml sterile 0.9% saline were injected and withdrawn with a total recovery of 85.3 ± 0.8 %. The fluid was examined for total cell counts and slides for differential counts were prepared on a Shandon Cytospin 3 (Cheshire, England) using approximately 100 μ L of BAL fluid.

Microsphere index: The first 100 consecutive intact macrophages viewed were evaluated and the total number of white microspheres within the 100 macrophages was defined as the microsphere index on each BAL day.

Histology: Lungs from all animals were preserved in formaldehyde for further evaluation and four slides from both lungs were prepared. Other internal organs (kidneys, liver, spleen, adrenals, pancreas, heart and brain) from 8 hamsters after 90 days of tracheal instillation (5 following white microsphere instillation and 3 following saline instillation) were preserved in formaldehyde for further evaluation. Animals instilled black microspheres had all extrapulmonary organs, including perihilar lymph glands, preserved in formaldehyde for histological evaluation. Two slides from each extrapulmonary organ were prepared. Hematoxylin-eosin was used for staining. Slides were thoroughly evaluated by two senior pathologists (V.D and Y.S.).

Statistics: Group data are expressed as mean \pm standard error (SE) of the mean. Differences between groups were compared using ANOVA with Bonferroni correction. Differences were taken as significant when $p < 0.05$.

Results:

The results of this experiment are shown in Figures 1 and 2 appended hereto in which Figure 1 shows polylactic acid instilled directly into the trachea of hamsters and detected inside alveolar macrophages obtained after whole lung lavage and Figure 2 shows alveolar macrophages of the saline instilled animals.

As can be seen, the white microspheres of polylactic acid are clearly identified within the alveolar macrophages and therefore can be used as a specific sensitive and stable marker for the diagnosis of aspiration.

As will be realized, in addition to polylactic acid other polymers and copolymers as well as different solvents, non-solvents and surfactants are suitable in the preparation process of the microspheres for use in the present invention.

Examples of proteins, which may be used for this purpose are human albumin, hemoglobin, gelatin and soybean proteins.

Example 2. Precipitation with a non-solvent:

The procedure of Example 1 as described for PLA is repeated, except that the solvent is either water or an aqueous solution and the non-solvent is a water-soluble volatile solvent, such as acetone.

Example 3. Interfacial crosslinking process:

2.5 g of protein were dissolved in 35 ml of 0.1N NaOH solution (aqueous phase), under magnetic stirring. The protein solution was emulsified in 75 ml of cyclohexane(organic phase-Baker), containing 0.5% phosphatidylcholine (surfactant – Sigma), under 350 rpm mechanical stirring. A solution of 168.2 mg of succinylchloride (SC) (Sigma) in 38 ml cyclohexane was added dropwise to the emulsion, for a period of 45 minutes. SC reacts predominantly with the proteins amine groups via a condensation reaction mechanism, at room temperature. The microspheres were then separated by decantation, washed three times with acetone (Frutarom) and air dried.

The crosslinking agent can be any diacylchloride, or any di- or poly functional molecule able to react with the proteins chemically active groups. Using this method, either microspheres or microcapsules may be obtained. The rigidity of the microspheres may be controlled by changing the nature and concentration of the crosslinking agent, as well as the nature of the organic phase and rinsing solvents.

Example 4. Denaturation by heating:

The protein is either precipitated as in Example 2, or emulsified as in Example 3, after which, the suspension is heated sufficiently to induce denaturation of the protein, thus stabilizing the particles.

It will be evident to those skilled in the art that the invention is not limited to the details of the foregoing illustrative examples and that the present invention may be embodied in other specific forms without departing from the essential attributes thereof, and it is therefore desired that the present embodiments and examples be considered in all respects as illustrative and not restrictive, reference being made to the appended claims, rather than to the foregoing description, and all changes which come within the meaning and range of equivalency of the claims are therefore intended to be embraced therein.

WHAT IS CLAIMED IS:

1. A diagnostic composition for detecting both aspiration and gastroesophageal reflux comprising bio-degradable microspheres having a diameter of about 0.1 - 10 microns.

2. A diagnostic composition according to claim 1 wherein said microspheres have a diameter of about 1 - 4 microns.

3. A diagnostic composition according to claim 1 for detecting both aspiration and gastroesophageal reflux comprising bio-degradable polymeric microspheres having a diameter of about 0.1 - 10 microns.

4. A diagnostic composition according to claim 3 wherein said polymeric microspheres are formed from polymeric materials selected from the group consisting of polyesters, polyphosphate esters, polyphosphazenes, polyorthoesters, polyanhydrides, polycarbonates, polyamides and proteins.

5. A diagnostic composition according to claim 5 wherein said polyesters are selected from the group consisting of homopolymers and copolymers of lactic acid, glycolic acid, mandelic acid, caprolactone, α -hydroxy acids, lactides and glycolides.

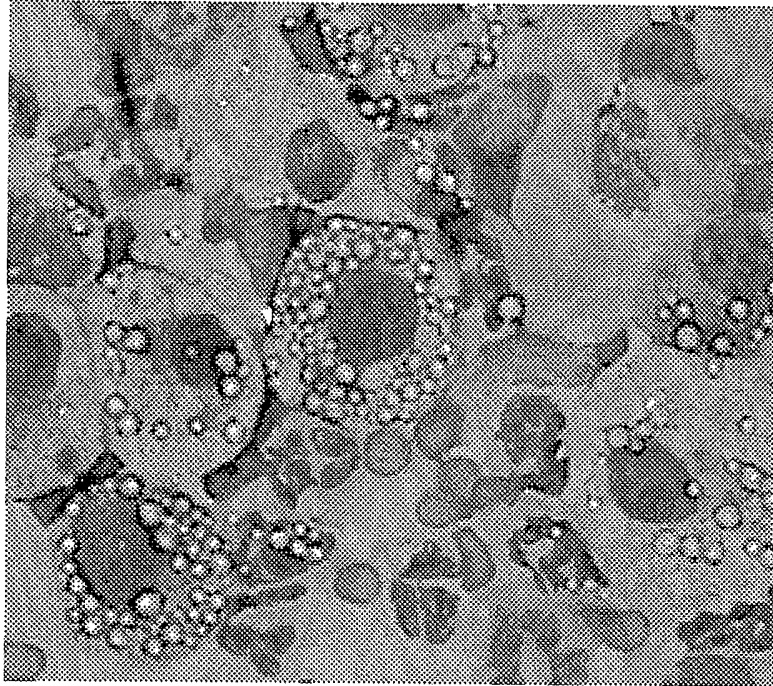
6. A diagnostic composition according to claim 1 comprising bio-degradable microspheres of polylactic acid.

7. A food product containing a diagnostic composition for detecting both aspiration and gastroesophageal reflux said composition comprising bio-degradable microspheres having a diameter of about 0.1 - 10 microns.

8. The use of bio-degradable microspheres having a diameter of about 0.1 - 10 microns for the manufacture of a diagnostic composition for detecting both aspiration and gastroesophageal reflux.

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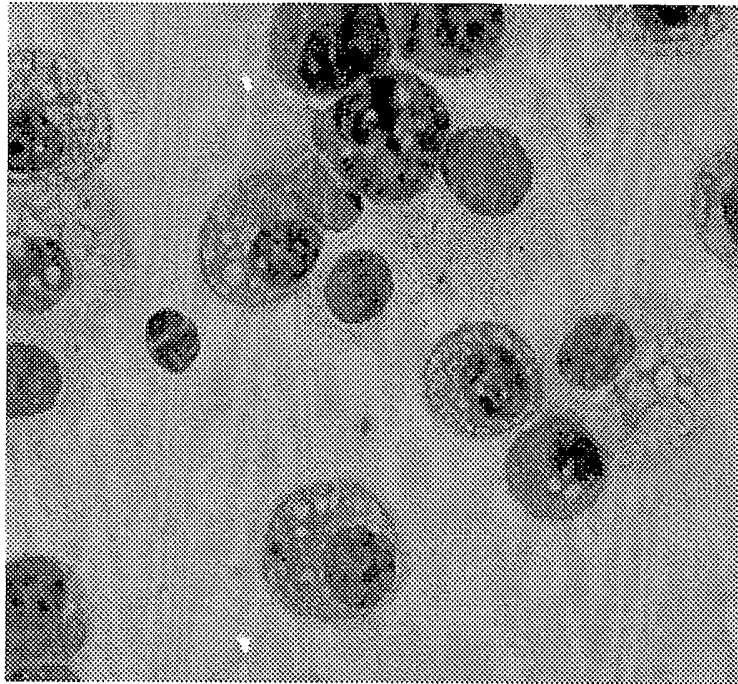
Fig. 1



Poly(lactic acid) instilled animal

2/2

Fig. 2



Saline instilled animal