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(54) **HOST CELL FOR MAKING ANTIBODY
Fc-HETERODIMERIC MOLECULES USING
ELECTROSTATIC STEERING EFFECTS**

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(57) **ABSTRACT**

The invention relates to methods of making Fc-heterodimeric proteins or polypeptides. The invention also relates to the Fc-heterodimeric proteins or polypeptides themselves, including the individual polypeptide components that comprise the heterodimer Nucleic acids encoding such polypeptides, expression vectors, and host cells. Moreover, the invention relates to pharmaceutical compositions comprising one or more Fc-heterodimeric proteins or polypeptides.

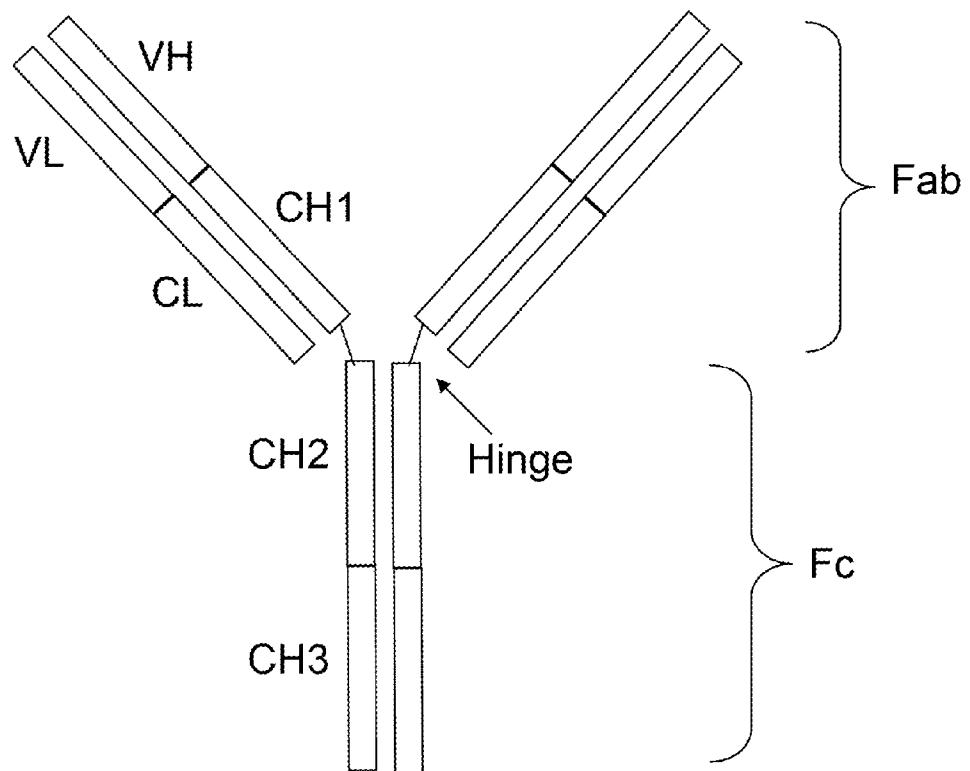


Figure 1

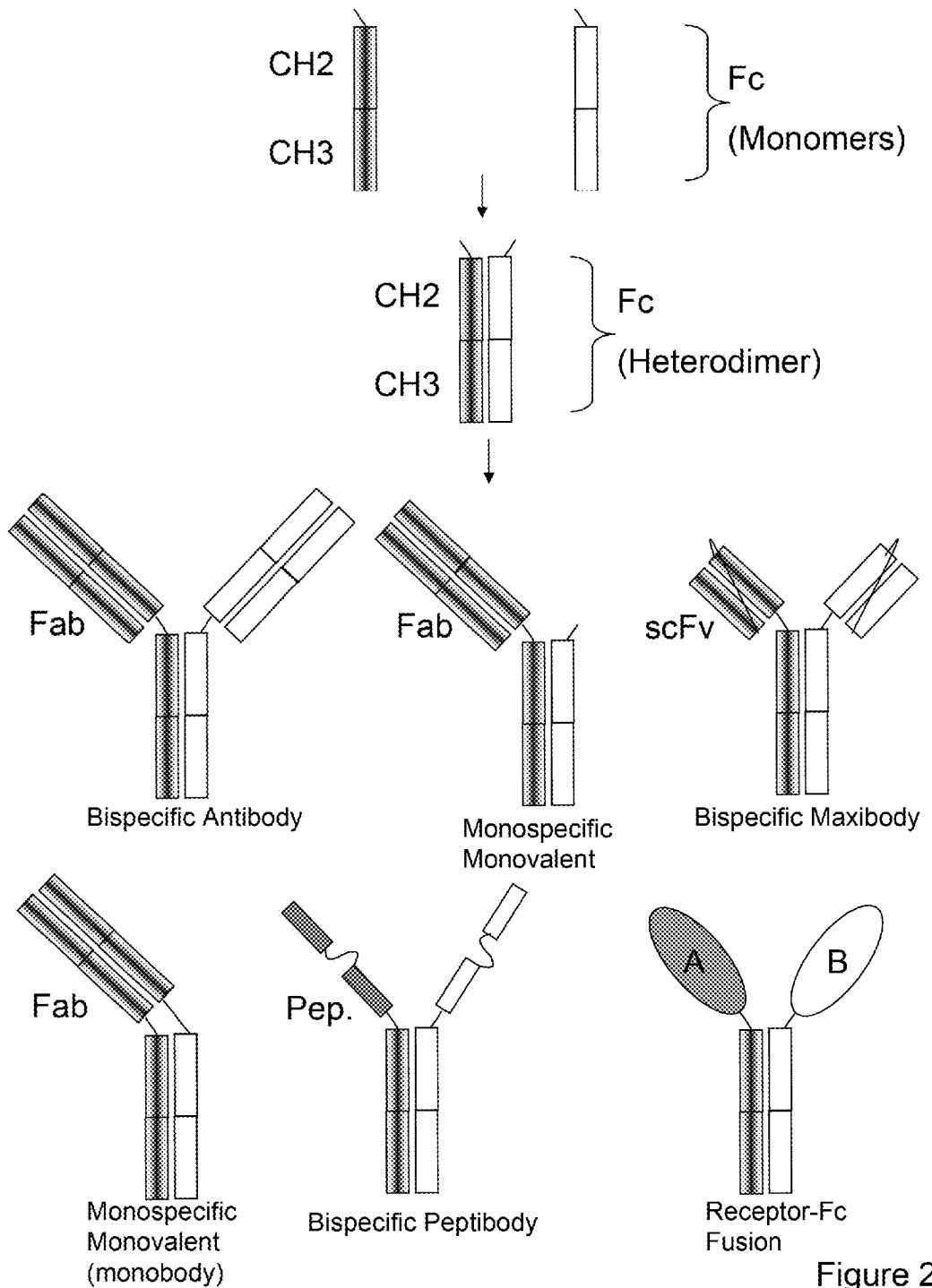


Figure 2

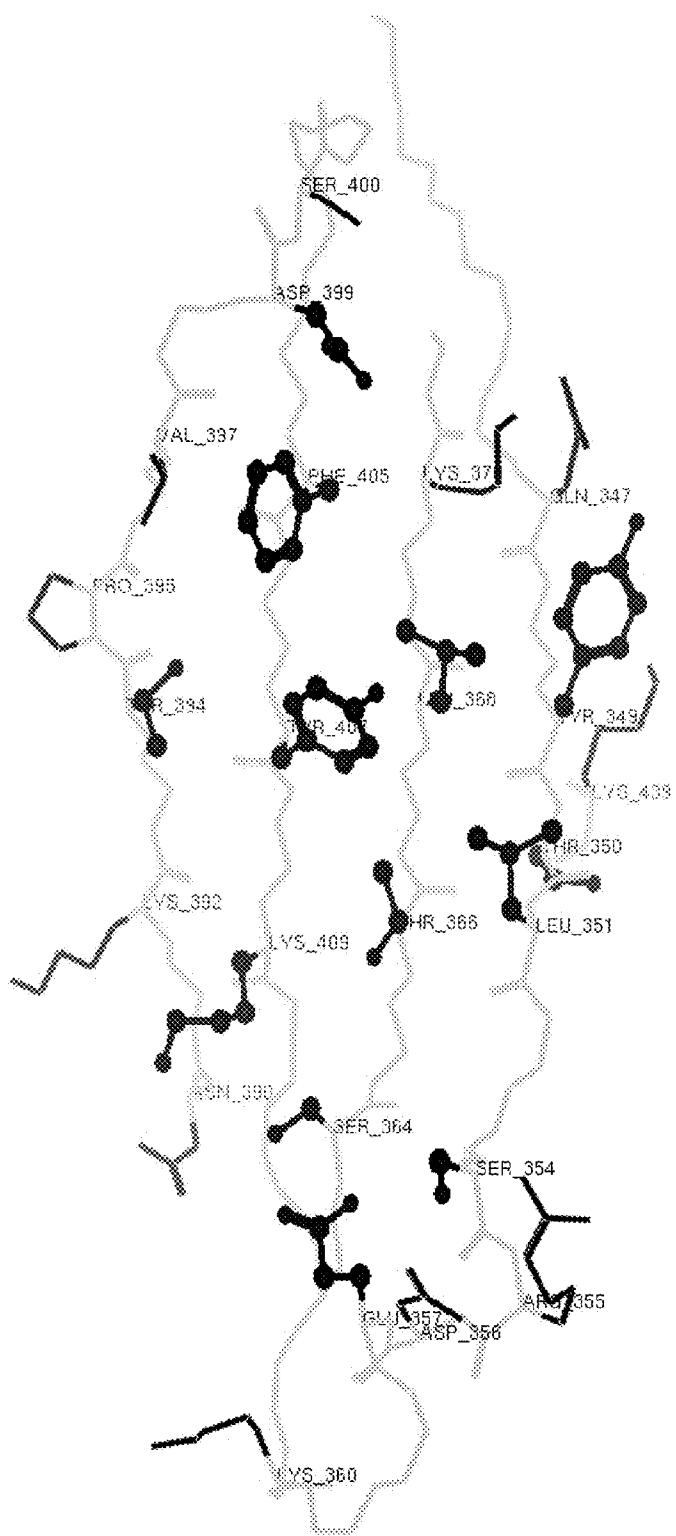


Figure 3

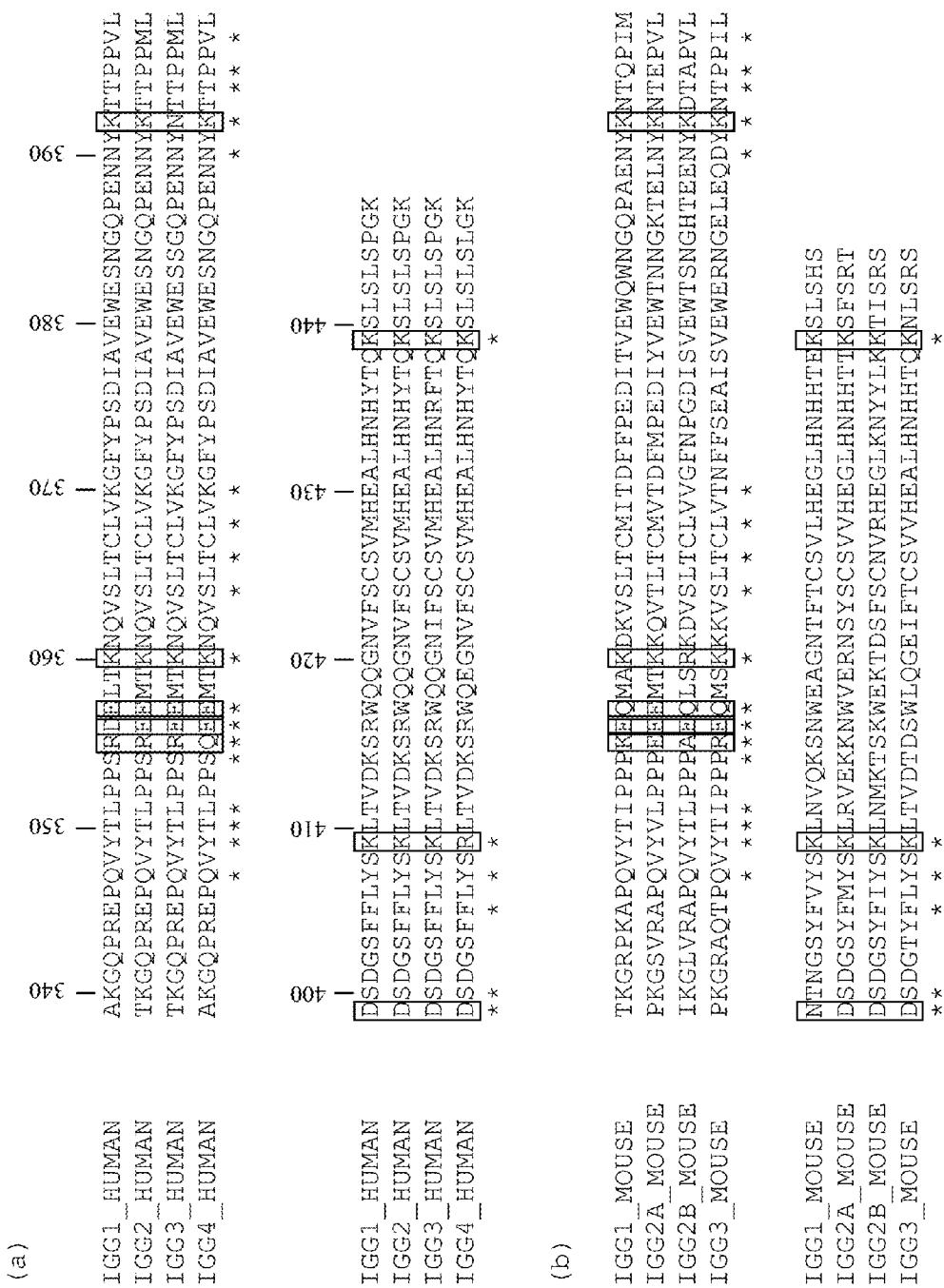


Figure 4

(c)

IGA_HUMAN
IGE_HUMAN
IGD_HUMAN
IGM_HUMAN

SGNT-FRPEVHILLPPSSEELALNELVTLLCLARGEFSPKDVLVRWLQGSQELPREKYLTW
TSGPR-AAPEVYAFATPEWPGSRDK-RTLACLIQNEFMPEDI SVQWLHNEVQLFDARHSTT
REPAA-QAPVKLSLNLIASSDPPPEAAASWLLICEVSGEFSPPNILLMWLEDQREVNTSGEAPA
PKGVALHRPDVYILLPPAREQQLNLRESATITCLVTGFSPADVFVQNMQRGQPLSPEKYVTS
* * * * *

IGA_HUMAN
IGE_HUMAN
IGD_HUMAN
IGM_HUMAN

ASRQEPEPSQGTTTFAVTSILRVAEDWKKGDTFSCMWGHEAL-PLAFTQKTIIDLAGK
QPRKT---KGSGFFVFSRLEVTRAWEQKDEEICRAVHEAASPSQTVQRAVSVNGK
RPPPQP---GSTTIFWAWSVLRVPAPPSPQPATYTCVVSHEDSRILLNASRSLEVSYVT
APMPEP-QAPGRYFAHSILTVSEEWNTGETYTCVVVAHEAL-PNRVTERTVDKSTGK
* * * * *

Figure 4

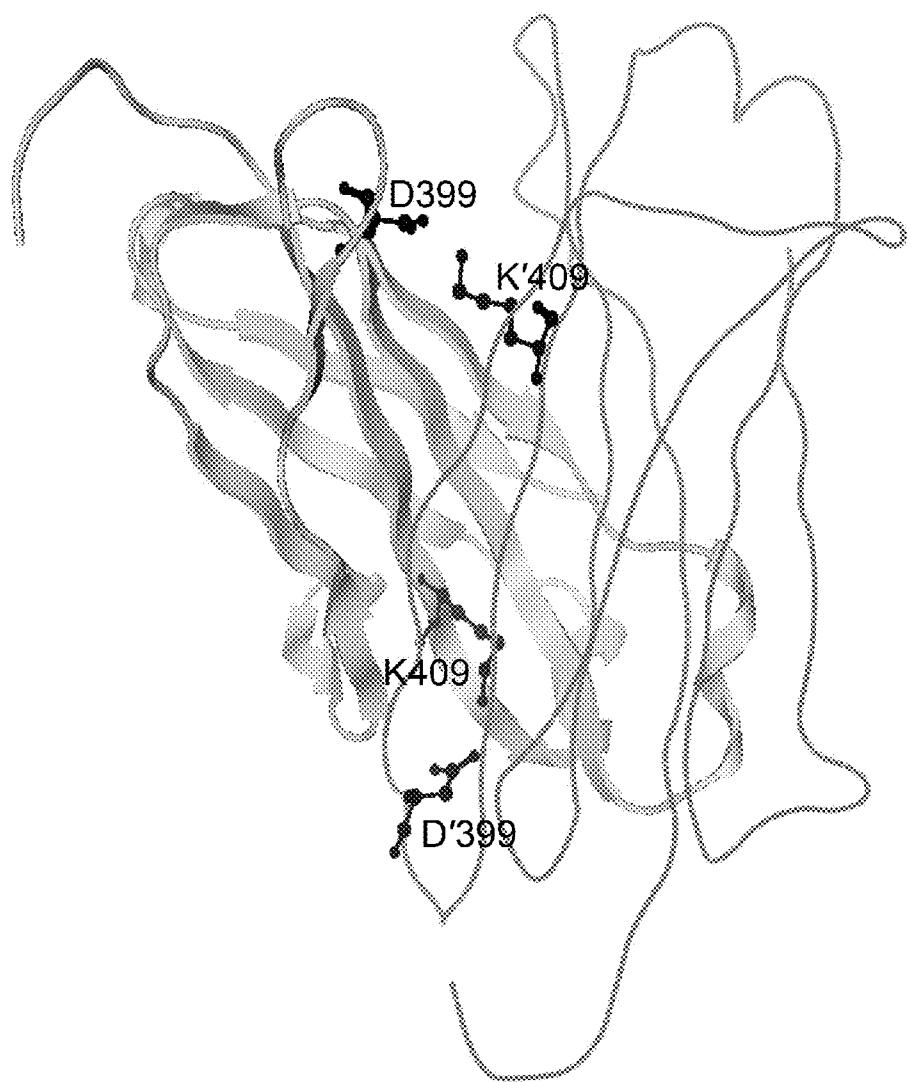


Figure 5

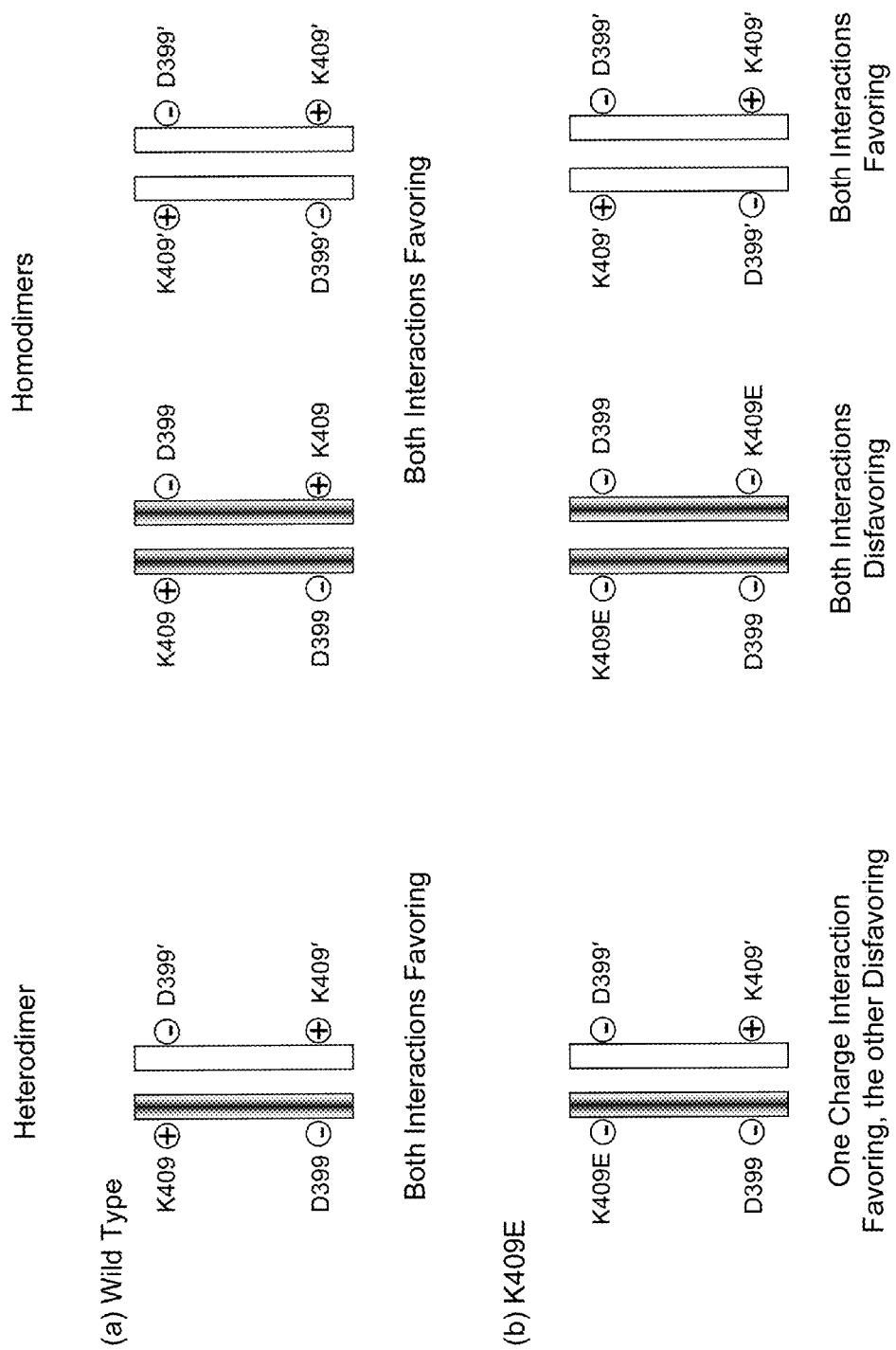
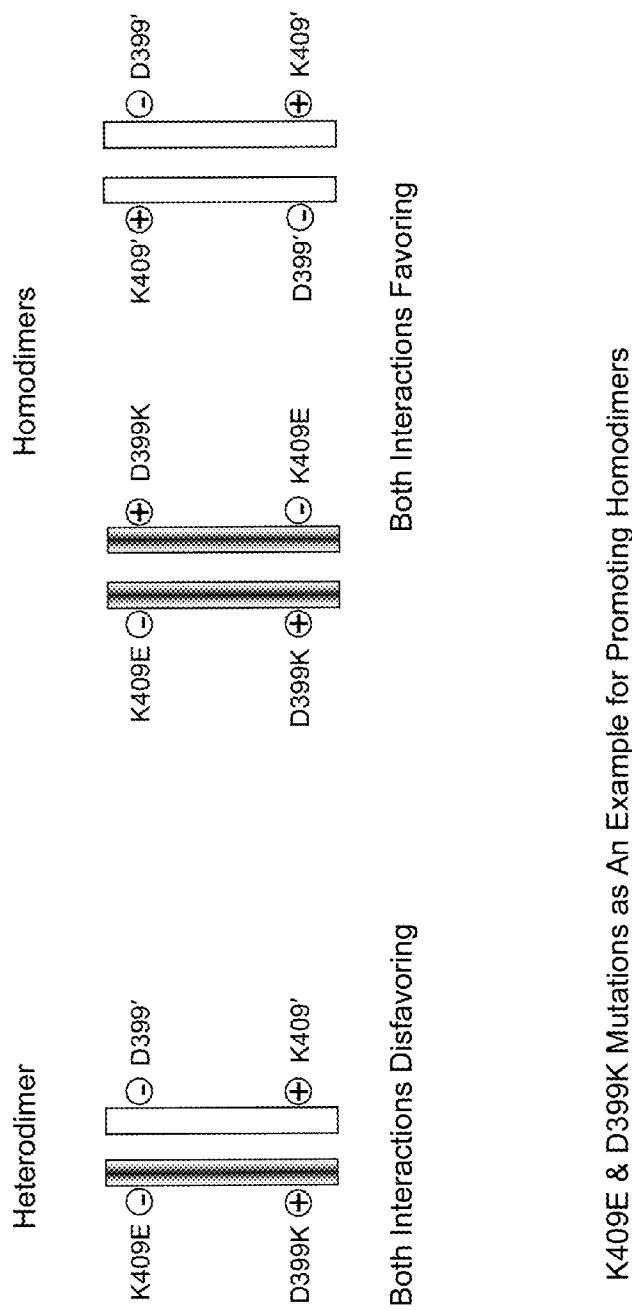


Figure 6



K409E & D399K Mutations as An Example for Promoting Homodimers

Figure 7

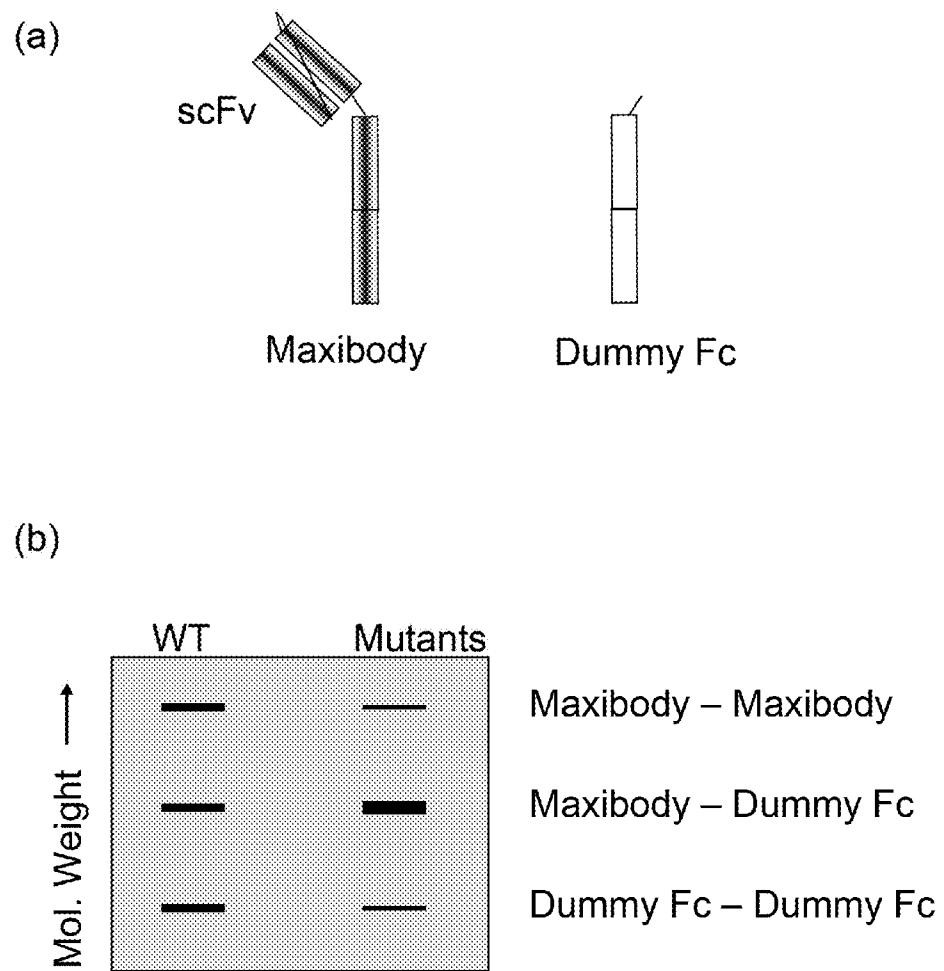


Figure 8

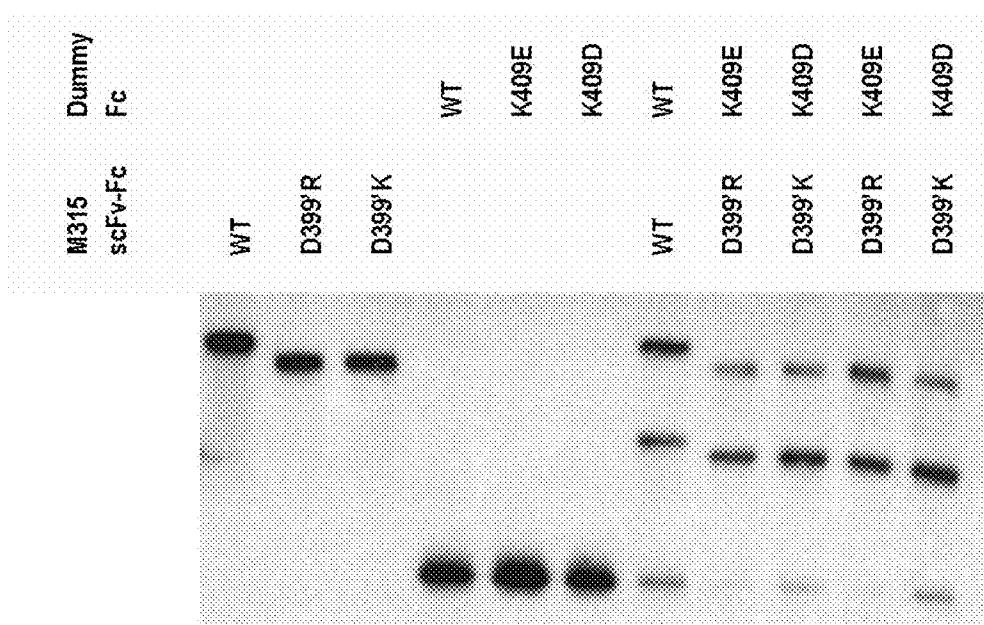


Figure 9

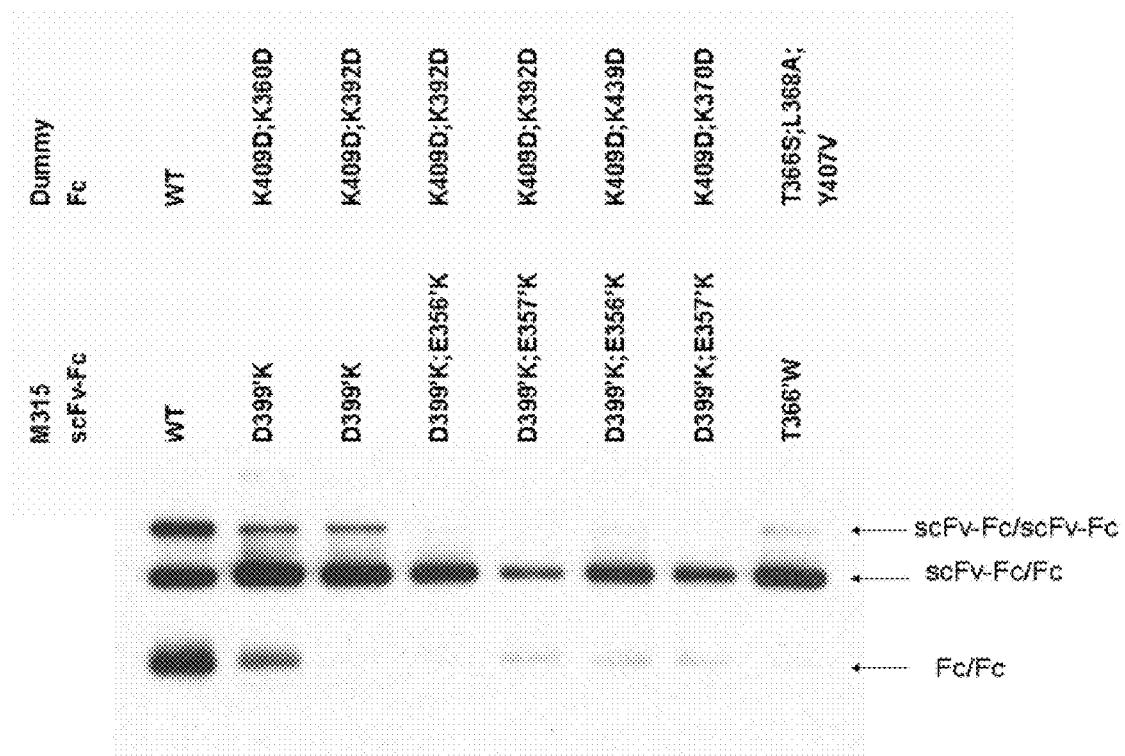


Figure 10

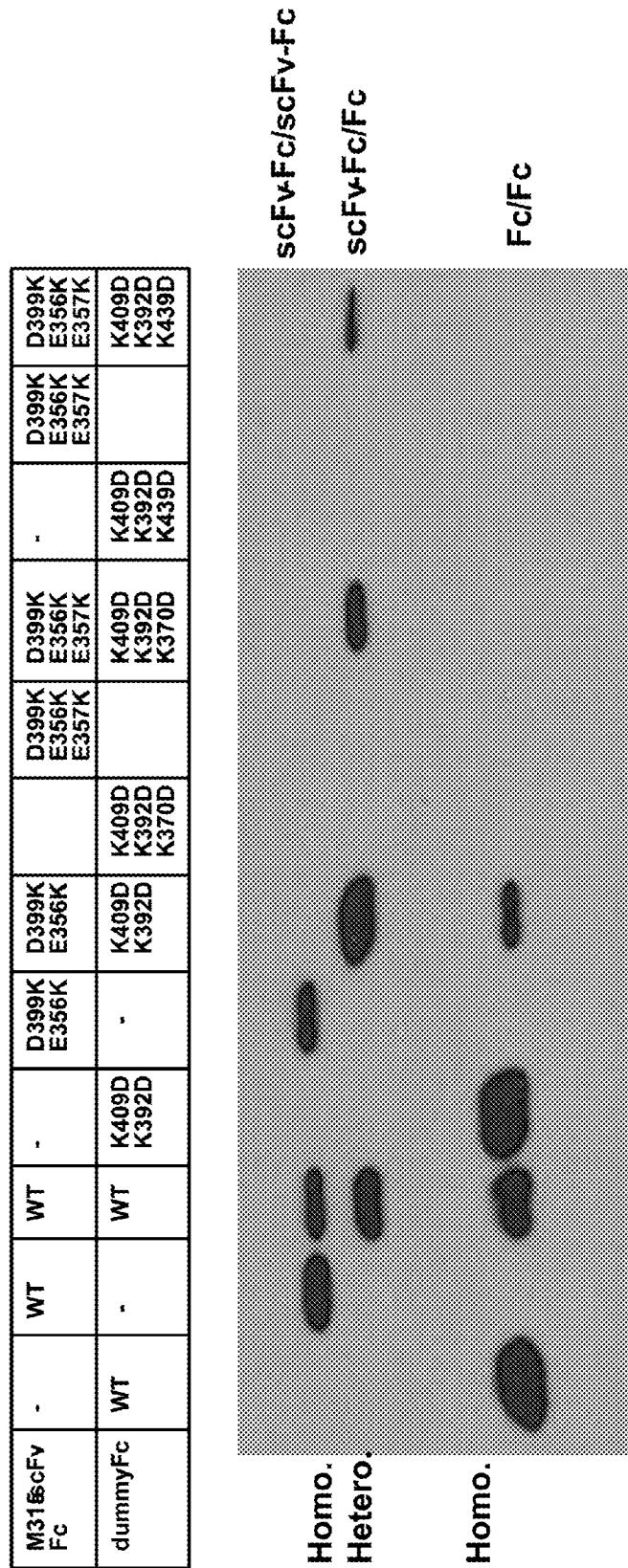


Figure 11

HOST CELL FOR MAKING ANTIBODY Fc-HETERODIMERIC MOLECULES USING ELECTROSTATIC STEERING EFFECTS

CROSS REFERENCE TO RELATED APPLICATIONS

[0001] This application is a Divisional of U.S. patent application Ser. No. 12/811,207 filed Jun. 29, 2010, which is a National Stage application under 35 U.S.C. §371 of International Application No. PCT/US2009/000071 (which designated the United States), having an international filing date of Jan. 6, 2009, which claims the priority benefit of U.S. Provisional Patent Application Ser. No. 61/019,569 filed Jan. 7, 2008 and U.S. Provisional Patent Application Ser. No. 61/120,305 filed Dec. 5, 2008, each of which is hereby incorporated by reference in its entirety.

REFERENCE TO THE SEQUENCE LISTING

[0002] The present application is being filed along with a Sequence Listing in electronic format. The Sequence Listing is provided as a file entitled A-1392-US-PCD_ST25.txt, created Sep. 25, 2013, which is 49,500 bytes in size. The information in the electronic format of the Sequence Listing is incorporated herein by reference in its entirety.

BACKGROUND

[0003] Antibodies have become the modality of choice within the biopharma industry because they possess several characteristics that are attractive to those developing therapeutic molecules. Along with the ability to target specific structures or cells, antibodies make its target susceptible to Fc-receptor cell-mediated phagocytosis and killing (Raghavan and Bjorkman 1996). Further, the antibody's ability to interact with neonatal Fc-receptor (FcRn) in a pH dependent manner confers it with extended serum half-life (Ghetie and Ward 2000). This unique feature of antibodies allows extending the half-life of therapeutic protein or peptide in the serum by engineering Fc-fusion molecules.

[0004] Antibodies belong to the immunoglobulin class of proteins which includes IgG, IgA, IgE, IgM, and IgD. The most abundant immunoglobulin class in human serum is IgG whose schematic structure is shown in the FIG. 1 (Deisenhofer 1981; Huber 1984; Roux 1999). The IgG structure has four chains, two light and two heavy chains; each light chain has two domains and each heavy chain has four domains. The antigen binding site is located in the Fab region (Fragment antigen binding) which contains a variable light (VL) and a variable heavy (VH) chain domain as well as constant light (LC) and constant heavy (CH1) chain domains. The CH2 and CH3 domain region of the heavy chain is called Fc (Fragment crystallizable). The IgG molecule can be considered as a heterotetramer having two heavy chains that are held together by disulfide bonds (-S-S-) at the hinge region and two light chains. The number of hinge disulfide bonds varies among the immunoglobulin subclasses (Papadea and Check 1989). The FcRn binding site is located in the Fc region of the antibody (Martin, West et al. 2001), and thus the extended serum half-life property of the antibody is retained in the Fc fragment. The Fc region alone can be thought of as a homodimer of heavy chains comprising CH2 and CH3 domains.

[0005] In certain instances, it is desirable to create a molecule that contains the Fc portion of an antibody but comprises a heterodimer. An important application of Fc het-

erodimeric molecules is the generation of bispecific antibodies (BsAbs). Bispecific antibodies refer to antibodies having specificities for at least two different antigens (Nolan and O'Kennedy 1990; de Leij, Molema et al. 1998; Carter 2001). Instead of having identical sequence in both the Fabs, bispecific antibodies bear different sequences in the two Fabs so that each arm of the Y-shaped molecule can bind to different antigens.

[0006] The use of bispecific antibodies for immunotherapy of cancer has been extensively reviewed in the literature (for example, see (Nolan and O'Kennedy 1990; de Leij, Molema et al. 1998; Carter 2001)). By having the ability to bind to two different epitopes or molecules, BsAbs provide means to both trigger an immune effector cell and bind a surface antigen on a tumor target cell. This helps to make use of the immune system to destroy cancer cells. Other applications of bispecific antibodies are extensively covered in U.S. Pat. Nos. 5,731,168 and 7,183,076.

[0007] The classical method of producing BsAbs by co-expressing two different IgGs in hybrid hybridomas leads to up to 10 possible combinations of heavy and light chains. This compromises the yield and imposes a purification challenge. Carter and co-workers engineered heavy chains for heterodimerization using a "knobs-into-holes" strategy (Ridgway, Presta et al. 1996; Atwell, Ridgway et al. 1997; Merchant, Zhu et al. 1998; Carter 2001). The knobs-into-holes concept was originally proposed by Crick as a model for packing of amino acid side chains between adjacent α -helices (Crick 1952). Carter and co-workers created a knob at the CH3 domain interface of the first chain by replacing a smaller amino acid side chain with a larger one (for example, T366Y); and a hole in the juxtaposed position at the CH3 interface of the second chain was created by replacing a larger amino acid side chain with a smaller one (for example, Y407T). The basis for creating knob and hole in the juxtaposed positions is that the knob and hole interaction will favor heterodimer formation, whereas the knob-knob and the hole-hole interaction will hinder homodimers formation due to the steric clash and deletion of favorable interactions, respectively. The knobs-into-holes mutations were also combined with inter-CH3 domain disulfide bond engineering to enhance heterodimer formation (Sowdhamini, Srinivasan et al. 1989; Atwell, Ridgway et al. 1997). In addition to these mutations, the input DNA ratio was also varied to maximize the yield (Merchant, Zhu et al. 1998). The "knobs-into-holes" technique is disclosed in U.S. Pat. Nos. 5,731,168 and 7,183,076.

SUMMARY

[0008] This application describes a strategy for altering the interaction of antibody domains, e.g., altering a CH3 domain to reduce the ability of the domain to interact with itself, i.e., form homodimers. In particular, one or more residues that make up the CH3-CH3 interface is replaced with a charged amino acid such that the interaction becomes electrostatically unfavorable. In preferred embodiments, a positive-charged amino acid in the interface, such as a lysine, arginine, or histidine, is replaced with a negative charged amino acid, such as aspartic acid or glutamic acid. In other embodiments, a negative-charged amino acid in the interface is replaced with a positive-charged amino acid. In certain embodiments, the amino acid is replaced with an unnatural amino acid having the desired charge characteristic.

[0009] Further described herein is a strategy for altering a pair of CH3 domains to reduce the ability of each domain to

interact with itself but to increase the ability of the domains to interact with each other, i.e., form heterodimers. This can be achieved by replacing one or more residues that make up the CH3-CH3 interface in both CH3 domains with a charged amino acid such that homodimer formation is electrostatically unfavorable but heterodimerization is electrostatically favorable. In certain embodiments, a charged amino acid in each CH3 domain is replaced with an amino acid with an opposite charge. For example, a positive-charged amino acid may be replaced with a negative charged amino acid in the first CH3 domain and a negative charged amino acid may be replaced with a positive-charged amino acid in the second CH3 domain. By reversing the charge of the amino acid, homodimer formation is reduced. When the replacements are coordinated properly, the reversed charges are electrostatically favorable, i.e., opposing charges in the interface, for heterodimerization formation.

[0010] In certain aspects, the invention provides a method of preparing a heterodimeric protein. The heterodimer may comprise a first CH3-containing polypeptide and a second CH3-containing polypeptide that meet together to form an interface engineered to promote heterodimer formation. The first CH3-containing polypeptide and second CH3-containing polypeptide are engineered to comprise one or more charged amino acids within the interface that are electrostatically unfavorable to homodimer formation but electrostatically favorable to heterodimer formation.

[0011] Such methods may include culturing a host cell comprising nucleic acids encoding the first and second CH3-containing polypeptides such that the polypeptides are co-expressed by the cell. In certain embodiments, the nucleic acids encoding the first and the second CH3-containing polypeptides are provided to the host cell at a ratio, for example 1:1, 1:2, 2:1, 1:3, 3:1, 1:4, 4:1, 1:5, 5:1, 1:6, 6:1, 1:7, 7:1, 1:8, 8:1, 1:9, 9:1, 1:10, 10:1. It is contemplated that altering the ratio of nucleic acids may increase the production of heterodimeric molecules versus homodimeric molecules.

[0012] The heterodimeric molecules may be purified from the host-cell culture using standard techniques. For example, when the heterodimeric protein comprises an Fc, the protein may be purified using a Protein A column. The purification techniques include but are not limited to chromatographic methods such as size exclusion, ion exchange and affinity-based chromatography and ultracentrifugation.

[0013] In certain embodiments, the CH3-containing polypeptide comprises an IgG Fc region, preferably derived from a wild-type human IgG Fc region. By “wild-type” human IgG Fc it is meant a sequence of amino acids that occurs naturally within the human population. Of course, just as the Fc sequence may vary slightly between individuals, one or more alterations may be made to a wild-type sequence and still remain within the scope of the invention. For example, the Fc region may contain additional alterations that are not related to the present invention, such as a mutation in a glycosylation site, inclusion of an unnatural amino acid, or a “knobs-into-holes” mutation.

[0014] In certain embodiments, the polypeptide containing the CH3 region is an IgG molecule and further contains a CH1 and CH2 domain. Exemplary human IgG sequences comprise the constant regions of IgG1 (e.g., SEQ ID NO:3; CH1=amino acids 1-98, CH2=amino acids 111-223, CH3 =224-330), IgG2 (e.g., SEQ ID NO:4; CH1=amino acids 1-94, CH2=amino acids 111-219, CH3 =220-326), IgG3 (e.g., SEQ ID NO:5; CH1=amino acids 1-98, CH2=amino

acids 161-270, CH3 =271-377), and IgG4(e.g., SEQ ID NO:6; CH1=amino acids 1-98, CH2=amino acids 111-220, CH3=221-327). Those of skill in the art may differ in their understanding of the exact amino acids corresponding to the various domains of the IgG molecule. Thus, the N-terminus or C-terminus of the domains outlined above may extend or be shortened by 1, 2, 3, 4, 5, 6, 7, 8, 9, or even 10 amino acids. Also note that the numbering scheme used here to designate domains differ from the EU numbering scheme of Kabat that is used in the rest of this patent application. For example, IgG1 “CH3=224-330” corresponds to “CH3=341-447” in EU numbering scheme.

[0015] The Fc region also may be comprised within the constant region of an IgA (e.g., SEQ ID NO:7), IgD (e.g., SEQ ID NO:8), IgE (e.g., SEQ ID NO:9), and IgM (e.g., SEQ ID NO:10) heavy chain.

[0016] The polypeptide containing the CH3 region may be an antibody heavy chain and the host cell may further express one or more antibody light chains. In embodiments wherein more than one heavy chain and light chains are co-expressed (e.g., bivalent antibody), each heavy chain may comprise a mutation in the CH1 region and each light chain may comprise a mutation in the constant region to preferentially bind to each other but not bind to the other light or heavy chain, respectively. In preferred embodiments, such mutations involve altering the charge of one or more amino acids in the interface between the CH1 region and the constant region of a light chain.

[0017] Preferred embodiments of the invention include but are not limited to an antibody, a bispecific antibody, a monospecific monovalent antibody, a bispecific maxobody (maxobody refers to scFv-Fc), a monobody, a peptibody, a bispecific peptibody, a monovalent peptibody (a peptide fused to one arm of a heterodimeric Fc molecule), and a receptor-Fc fusion protein. See FIG. 2.

[0018] Examples of mammalian host cells that may be used include but are not limited to CHO, 293, and myeloma cell lines. The host cell may also be yeast or a prokaryote, such as *E. coli*.

[0019] The heterodimeric proteins may be particularly useful in therapeutic compositions. In certain embodiments, a heterodimeric protein may be formulated in a composition that includes one or more pharmaceutically acceptable buffer or excipient. Such therapeutic composition may be administered to a subject to treat a disease or may be given to prevent a disease or prevent the symptoms of a disease from progressing.

BRIEF DESCRIPTION OF THE DRAWINGS

[0020] FIG. 1. Schematic diagram of IgG1 antibody with the domains indicated. The IgG1 antibody is a Y-shaped tetramer with two heavy chains (longer length) and two light chains (shorter length). The two heavy chains are linked together by disulfide bonds (—S—S—) at the hinge region. Fab—fragment antigen binding, Fc—fragment crystallizable, VL—variable light chain domain, VH—variable heavy chain domain, CL—constant (no sequence variation) light chain domain, CH1—constant heavy chain domain 1, CH2—constant heavy chain domain 2, CH3—constant heavy chain domain 3.

[0021] FIG. 2. Figure depicts some of the embodiments that include Fc-heterodimeric molecules. These include bispecific antibodies (have specificity for two or more antigens) to

receptor-Fc fusion molecules. Preferably, the Fc retains its ability to interact with the FcRn receptor, even without the

[0022] Fab domains, leading to longer serum half-life for proteins/domains that are fused to the Fc heavy chains. scFv—single chain fragment variable, Pep.—peptibody, A and B stands for proteins or receptors or domains.

[0023] FIG. 3. CH3 domain interface structure with residues involved in the domain-domain interaction shown. The interface residues were identified using a distance cutoff method. Structurally conserved and buried (solvent accessible surface area <10%) residues are shown in the ball-and-stick model. Solvent exposed or structurally not conserved residues are shown in the stick representation. The analysis is based on the IgG1 crystal structure (PDB code: 1L6X) which is determined at high-resolution (1.65 Å) (Idusogie, Presta et al. 2000).

[0024] FIG. 4. Comparison of IgG subclass sequences from (a) human and (b) mouse. Only the heavy chain sequence corresponding to the CH3 domain is shown. The star (*) indicates residue positions involved in the CH3-CH3 domain interaction identified based on the IgG1 human Fc crystal structure (1L6X). Positions marked with rectangles are preferred residues for mutation to enhance heterodimer formation. It may be noted here that charged residues are highly conserved among the IgGs. (c) CH3 domain sequence comparison of other class of antibodies (IgA, IgE, IgD, and IgM). The interface residue positions (indicated by **) in (b) and (c) were identified based on sequence comparison with Hu IgG1 sequence that is also shown. In (a), the sequences derived from human IgG1, IgG2, IgG3, and IgG4 correspond to SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, and

[0025] SEQ ID NO:14, respectively. In (b), the sequences derived from human IgG1, mouse IgG1, mouse IgG2a, mouse IgG2b, and mouse IgG3 correspond to SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, and SEQ ID NO:19, respectively. In (c), the sequences derived from human IgG1, human IgA, human IgE, human IgD, and human IgM correspond to SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, and SEQ ID NO:24, respectively.

[0026] FIG. 5. Crystal structure of CH3 domain homodimer with one domain shown in ribbon representation and the other domain shown in wire model. The Lys409 (Lys409' in the second domain) and Asp399 (Asp399' in the second) residues are shown in ball-and-stick model in order to illustrate each pair-wise interaction is represented twice in the structure. This is due to the two-fold symmetry present in the CH3-CH3 domain interaction. The figure was created using the 1L6X co-ordinates deposited in the PDB.

[0027] FIG. 6. Schematics showing electrostatic interactions in the wild type and in the mutants designed as an example to enhance heterodimer formation and hinder homodimer formation. (a) In the case of WT, electrostatic interactions favor both heterodimer and homodimer formation giving them equal probability. (b) In the single mutant (K409E) case, one of the homodimer is discouraged by both the interactions and at the same time heterodimer is also discouraged by one of the interactions. In the double mutant case, both the electrostatic interactions favor heterodimer and disfavor homodimer formation. Additional mutations involving charge change (for example, K360E) could also be used to enhance the electrostatic steering effects on the formation of heterodimer and homodimer

[0028] FIG. 7. This figure shows that electrostatic interactions could also be used to favor homodimers and disfavor heterodimer formation, when two different chains are co-expressed.

[0029] FIG. 8. Figure (a) shows the schematic drawing of the constructs used in the Example. The first chain of the Fc has a maxibody (single chain fragment variable, scFv) covalently linked, and the second chain called dummy Fc does not have any domain or functionality attached to it. (b) Illustration of expected relative mobility on the SDS-PAGE. Because the Fc chain attached to the maxibody has a higher molecular weight than the dummy Fc, homodimers and heterodimer have different mobility on the SDS-PAGE. The thickness of the band on the SDS-PAGE can be used as a measure of fraction of heterodimer and homodimer yield. The wild type is included as a control and to monitor relative improvement on the heterodimer yield due to various mutations.

[0030] FIG. 9. SDS-PAGE analysis showing the effects of mutations on the D399'-K409 interaction pair. FIG. 10. SDS-PAGE analysis of charge residue mutations (listed in Table 6) in addition to D399'K-K409D pair mutations. Wild type (first lane) and knobs-into-holes mutations (last lane) are also shown for comparison. 1:2 input DNA ratio of dummy Fc and M315 maxibody was used here.

[0031] FIG. 11. Western blot demonstrating certain combinations of mutant achieve high selectivity for heterodimer formation. Fc molecules were detected using goat-anti-human Fc HRP conjugated at 1:10,000.

DETAILED DESCRIPTION OF PREFERRED EMBODIMENTS

[0032] A total of 48 antibody crystal structures which had co-ordinates corresponding to the Fc region were identified from the Protein Data Bank (PDB) (Bernstein, Koetzle et al. 1977) using a structure based search algorithm (Ye and Godzik 2004). Examination of the identified Fc crystal structures revealed that the structure determined at highest resolution corresponds to the Fc fragment of RITUXIMAB bound to a minimized version of the B-domain from protein A called Z34C (PDB code: 1L6X). The biological Fc homodimer structure for 1L6X was generated using the deposited Fc monomer co-ordinates and crystal symmetry. Two methods were used to identify the residues involved in the CH3-CH3 domain interaction: (i) contact as determined by distance limit criterion and (ii) solvent accessible surface area analysis.

[0033] According to the contact based method, interface residues are defined as residues whose side chain heavy atoms are positioned closer than a specified limit from the heavy atoms of any residues in the second chain. Though 4.5 Å distance limit is preferred, one could also use longer distance limit (for example, 5.5 Å) in order to identify the interface residues (Bahar and Jernigan 1997).

[0034] The second method involves calculating solvent accessible surface area (ASA) of the CH3 domain residues in the presence and absence of the second chain (Lee and Richards 1971). The residues that show difference (>1 Å²) in ASA between the two calculations are identified as interface residues. Both the methods identified similar set of interface residues. Further, they were consistent with the published work (Miller 1990).

[0035] Table 1 lists twenty four interface residues identified based on the contact criterion method, using the distance limit

of 4.5 Å. These residues were further examined for structural conservation. For this purpose, 48 Fc crystal structures identified from the PDB were superimposed and analyzed by calculating root mean square deviation for the side chain heavy atoms. The residue designations are based on the EU numbering scheme of Kabat, which also corresponds to the numbering in the Protein Data Bank (PDB).

[0036] FIG. 3 shows the CH3 domain interface along with the structurally conserved, buried (% ASA<10), and exposed (% ASA>10) positions (% ASA refers to ratio of observed ASA to the standard ASA of amino acids; (Lee and Richards 1971)). Conservation of interface residues among Human and Mouse IgG subclasses as well as among other Ig classes was also examined through sequence comparisons (FIG. 4).

TABLE 1

List of CH3 domain interface residues in the first chain (A) and their contacting residues in the second chain (B)^a

Interface Res. in Chain A	Contacting Residues in Chain B
GLN A 347	LYS B 360'
TYR A 349	SER B 354' ASP B 356' GLU B 357' LYS B 360'
THR A 350	SER B 354' ARG B 355'
LEU A 351	LEU B 351' PRO B 352' PRO B 353' SER B 354'
	THR B 366'
SER A 354	TYR B 349' THR B 350' LEU B 351'
ARG A 355 ^b	THR B 350'
ASP A 356	TYR B 349' LYS B 439'
GLU A 357	TYR B 349' LYS B 370'
<i>LYS A 360^b</i>	GLN B 347' TYR B 349'
SER A 364	LEU B 368' LYS B 370'
THR A 366	LEU B 351' TYR B 407'
LEU A 368	SER B 364' LYS B 409'
LYS A 370	GLU B 357' SER B 364'
ASN A 390	SER B 400'
LYS A 392	LEU B 398' ASP B 399' SER B 400' PHE B 405'
THR A 394	THR B 394' VAL B 397' PHE B 405' TYR B 407'
PRO A 395	VAL B 397'
VAL A 397	THR B 393' THR B 394' PRO B 395'
ASP A 399	LYS B 392' LYS B 409'
SER A 400	ASN B 390' LYS B 392'
PHE A 405	LYS B 392' THR B 394' LYS B 409'
TYR A 407	THR B 366' THR B 394' TYR B 407' SER B 408'
	LYS B 409'
LYS A 409	LEU B 368' ASP B 399' PHE B 405' TYR B 407'
LYS A 439	ASP B 356'

^aPositions involving interaction between oppositely charged residues are indicated in bold. Due to the 2-fold symmetry present in the CH3—CH3 domain interaction, each pair-wise interaction is represented twice in the structure (for example, Asp A 356 --- Lys B 439' & Lys A 439 --- Asp B 356; FIG. 5)

^bArg355 and Lys360 positions (shown in italics) could also be used for enhancing electrostatic steering effects though they are not involved in interaction with oppositely charged residues.

[0037] At neutral pH (~7.0), Asp and Glu residues are negatively charged and Lys, Arg and His are positively charged. These charged residues can be used to promote heterodimer formation and at the same time hinder homodimers. Attractive interaction takes place between opposite charges and repulsive interaction occurs between like charges. The method presented here makes use of the attractive and repulsive interactions for promoting heterodimer and hindering homodimer, respectively, by carrying out site directed mutagenesis of charged interface residues.

[0038] Examination of the identified CH3 domain interface residues (Table 1) reveals four unique charge residue pairs involved in the domain-domain interaction (Asp356-Lys439', Glu357-Lys370', Lys392-Asp399', Asp399-Lys409'; residue numbering in the second chain is indicated by prime'). These charge pairs are not necessarily involved in charge-charge

interaction in the crystal structure used here (1L6X), since crystal structure is an end product in the protein folding reaction pathway and it represents structure in the crystalline state. It is assumed here that in order to have electrostatic steering effects it is sufficient if the residues are close in space as defined by the distance limit criterion (4.5 Å). It must also be noted here that due to the 2-fold symmetry present in the CH3-CH3 domain interaction, each unique interaction will be represented twice in the structure (for example, Asp399-Lys409' & Lys409-Asp399'; FIG. 5).

[0039] The four pairs were ranked according to the extent of solvent accessibility (ASA analysis) (Lee and Richards 1971). In Lys409-Asp399' case, both the residues were structurally conserved as well as buried. In other three pairs case, at least one of the partner is solvent exposed (% ASA>10). Therefore, for the Example herein, the Lys409-Asp399' pair was chosen for site directed mutagenesis. The strategy is schematically shown in FIG. 6.

[0040] In the wild type, K409-D399' interaction favors both heterodimer and homodimer formation. A single mutation switching the charge polarity (K409E; positive to negative charge) in the first chain leads to unfavorable interactions for the formation of the first chain homodimer. The unfavorable interactions arise due to the repulsive interactions occurring between the same charges (negative-negative; D399-K409E & K409E-D399). A similar mutation switching the charge polarity (D399'K; negative to positive charge) in the second chain leads to unfavorable interactions (K409-D399'K & D399'K-K409') for the second chain homodimer formation. But, at the same time, these two mutations (K409E & D399'K) lead to favorable interactions (K409E-D399'K & D399-K409') for the heterodimer formation.

[0041] The electrostatic steering effects on heterodimer formation and homodimer discouragement can be further enhanced by mutation of additional charge residues which may or may not be paired with an oppositely charged residue in the second chain, such as Arg355 and Lys360, as shown in FIG. 6. The mutations shown in FIG. 6 are for the purpose of illustration only. Table 2 lists many possible mutations involving charge change, and the mutations can be combined to enhance the electrostatic effects.

TABLE 2a

List of some possible pair-wise charge residue mutations to enhance heterodimer formation^a

Position in the First Chain	Mutation in the First Chain	Interacting Position in the Second Chain	Corresponding Mutation in the Second Chain
Lys409	Asp or Glu	Asp399'	Lys or Arg ^b
Lys392	Asp or Glu	Asp399'	Lys or Arg ^b
Lys439	Asp or Glu	Asp356'	Lys or Arg ^b
Lys370	Asp or Glu	Glu357'	Lys or Arg ^b
Asp399	Lys or Arg ^b	Lys409'	Asp or Glu
Asp399	Lys or Arg ^b	Lys392'	Asp or Glu
Asp356	Lys or Arg ^b	Lys439'	Asp or Glu
Glu357	Lys or Arg ^b	Lys370'	Asp or Glu

^aCombinations of the above pair-wise charge residue mutations could also be used. For example Lys409 --- Asp399' interaction pair mutations could be combined with Lys439 --- Asp356' pair mutations.

^bHistidine (His) could also be added to this list of positively charged residues, however, increase in side chain volume and pH dependency should be taken into account in the design.

TABLE 2b

Additional single charge residue mutations to enhance electrostatic steering effects ^a			
Position in Chain 1	Mutation	Position in Chain 2	Mutation
Arg355	Asp or Glu	Arg355'	Asp or Glu
Lys360	Asp or Glu	Lys360'	Asp or Glu

^aThese single residue mutations could be combined with the Table 2a pair-wise mutations to enhance the heterodimer formation (FIG. 6).

[0042] Each positively charged residue (Lys and Arg) can be mutated to two negatively charged residues (Asp or Glu) and vice versa, and as a result the method described here provides numerous combinations. It must be stated here that different combinations will have diverse effect on the quaternary (homodimer/heterodimer) structure formation depending on surrounding residues at the mutation site and role of water molecules. The amino acid Histidine (His) is positively charged at neutral pH and therefore mutation to His is also contemplated. However, mutating negatively charged residues (Asp or Glu) to His will lead to increase in side chain volume which may cause steric issues. Further, Histidine proton donor- and acceptor-form depends on the localized environment. These issues should be taken into consideration during the design strategy.

[0043] Because the interface residues are highly conserved in Human and Mouse IgG subclasses, electrostatic steering effects can be applied to Human or Mouse IgG1, IgG2, IgG3, or IgG4. This strategy can also be extended to modifying uncharged residues to charged residues at the CH3 domain interface. A similar strategy involving charge residue mutations can also be used to enhance homodimers and hinder heterodimer formation when two different heavy chains are co-expressed (FIG. 7).

[0044] In order to assess the stability of the charge residue mutants, EGAD software was used to estimate the CH3-CH3 domain binding free energy. By optimizing parameters used in the calculation, Pokala and Handel could predict the effects of nearly 400 mutations on protein-protein complex formation within 1.0kcal/mol error (Pokala and Handel 2005). EGAD was used to roughly compare the binding free energy of various mutations made at the CH3 domain interface.

[0045] Table 3 lists computed binding free energy (ΔG) for the interface charge residue mutants. The binding free energy of a mutant is defined as $\Delta G_{mut} = \mu(\Delta G_{mut} - \Delta G_{wt})$. Where, $\mu = 0.1$, in general) is the scaling factor used to normalize the predicted changes in binding affinity to have a slope of 1 when comparing with the experimental energies (Pokala and Handel 2005). The free energy of dissociation (ΔG) is defined as the energy difference between the complex (ΔG_{bound}) and free states (ΔG_{free}). The comparison shows that charged residue mutations affect the stability to a much lesser extent compared to the knobs-into-holes mutations. For comparison, melting temperatures reported for the wild type and knobs-into-holes mutants are given. The melting temperatures were measured by Carter and coworkers using only the CH3 domain construct (Atwell, Ridgway et al. 1997). For the knobs-into-holes mutants, decrease in enthalpy was also observed in the differential scanning calorimetry experiments.

TABLE 3

CH3-CH3 domain binding free energy for various mutants designed to enhance heterodimer formation, calculated using the EGAD program (Pokala and Handel 2005) ^a				
Protein	Description	ΔG (in kcal/mol)	ΔG_{mut} (in kcal/mol)	Melting Temp. T_m (in °C.)
WT	Wild Type	-30.69	0	80.4
T366W-Y407'A	Knob-Hole	-24.60	6.09	65.4
T366W-T366'S-L368'A-Y407'V	Knob-Hole	-28.57	2.12	69.4
K409E-D399'K	Charge-Charge	-29.56	1.13	ND
K409E-D399'R	Charge-Charge	-29.47	1.22	ND
K409D-D399'K	Charge-Charge	-28.16	2.53	ND
K409D-D399'R	Charge-Charge	-27.69	3.00	ND
K392E-D399'R	Charge-Charge	-29.27	1.42	ND
K392E-D399'K	Charge-Charge	-29.87	0.82	ND
K392D-D399'R	Charge-Charge	-28.82	1.87	ND
K392D-D399'R	Charge-Charge	-29.42	1.27	ND

^aNot all possible charge-charge pairs were considered for the binding free energy calculation. Wild type is listed for comparison. ΔG is defined as energy difference between the complex and free states. The binding free energy of a mutant (ΔG_{mut}) is defined as difference between the mutant (ΔG_{mut}) and wild type (ΔG_{wt}) free energies.

FIG. 2 depicts several embodiments comprising Fc heterodimeric molecules, from bispecific antibodies to heterodimeric receptor complexes. The two heavy chains of heterodimeric Fc molecules can be fused with proteins and/or domains that have different functionalities. For example, fusing Fabs that bind to different antigens will lead to bispecific antibodies (BsAbs). Fusing two different single-chain Fv (scFv; variable light and heavy chains joined by a flexible peptide linker) domains will lead to bispecific maxibodies. Further, domains or proteins that interact for functional reasons can also be fused with heterodimeric Fc for the purpose of developing functional assays or for therapeutic uses. For instance, in the hematopoietic receptor family gp130 is known to interact with other receptors such as Leukemia Inhibitory Factor Receptor (LIFR). The extra cellular domain (ECD) of gp130 can be fused to the first heavy chain of Fc and the ECD of LIFR can be fused to the second Fc heavy chain, which will lead to formation of gp130-LIFR complex that is likely to mimic the biological state. Since FcRn binding site is located in the Fc region, Fc fusion molecules are likely to have extended serum half-life - a feature that distinguishes Fc heterodimeric molecules from other heterodimeric molecules such as leucine zipper fusion proteins (Liu, Caderas et al. 2001). It is not essential to have different functionalities attached to the two heavy chains of the Fc heterodimer A monobody can also be created (FIG. 2).

[0046] In certain embodiments, e.g., when producing bispecific antibodies, multiple different light chains may be co-expressed with the multiple different heavy chains. To increase the fidelity of each light chain binding to the proper heavy chain thereby maintaining specificity of the antibody "arm," the CH1 domains of one or more of the heavy chains and the constant region of one or more of the light chains can be engineered to favor dimerization. Preferably, this is accomplished using an electrostatic steering technique similar to that described above for the CH3 domains.

[0047] The interaction of the kappa light chain sequence corresponding to the Protein Data Bank (PDB) deposition code 1NOX (SEQ ID NO:25) and the lambda light chain corresponding to (PDB) deposition code 7FAB (SEQ ID NO:26) with the heavy chain sequence corresponding to the

CH1 domain of IgG1 (SEQ ID NO:27) was analyzed. The lambda light chain-Heavy chain contacts within the interface are shown in Table 4.

TABLE 4

List of lambda light chain interface residues and their contacting residues in the heavy chain ^a	
Interface Res. in Lambda Light Chain	Contacting Residues in the Heavy Chain
THR L 112	ALA H 141
PHE L 114	LEU H 128 ALA H 129 ALA H 141 LEU H 142
	GLY H 143 VAL H 185
SER L 117	PHE H 126 PRO H 127
GLU L 119	VAL H 125 PHE H 126 PRO H 127 LYS H 213
GLU L 120	PHE H 126
LYS L 125	LYS H 147 ASP H 148
THR L 127	LEU H 145 LYS H 147
VAL L 129	LEU H 128 LEU H 145 SER H 183
LEU L 131	PHE H 170 SER H 183 VAL H 185
SER L 133	HIS H 168 PHE H 170
GLU L 156	VAL H 173 LEU H 174 GLN H 175 SER H 176
THR L 158	PRO H 171 ALA H 172 VAL H 173
SER L 161	PRO H 171
GLN L 163	HIS H 168
ALA L 169	HIS H 168 PHE H 170
SER L 171	PHE H 170 PRO H 171
TYR L 173	LEU H 145 VAL H 173 SER H 181 LEU H 182
	SER H 183

^aContacting residues were identified using 4.5 Å distance limit criterion. The light and heavy chain numbering scheme corresponds to that in the deposited co-ordinates file (PDB code: 7FAB).

[0048] The kappa light chain-heavy chain contacts within the interface are shown in Table 5.

TABLE 5

List of kappa light chain interface residues and their contacting residues in the heavy chain ^a	
Interface Res. in Kappa Light Chain	Contacting Residues in the Heavy Chain
PHE 116	THR H 139 ALA H 140 ALA H 141
PHE 118	LEU H 128 ALA H 129 PRO H 130 ALA H 141
	LEU H 142
SER 121	PHE H 126 PRO H 127
ASP 122	LYS H 218
GLU 123	VAL H 125 PHE H 126 LYS H 213
GLN 124	PHE H 126 LEU H 145 LYS H 147
SER 131	LEU H 145 LYS H 147
VAL 133	LEU H 128
LEU 135	ALA H 141 PHE H 170 VAL H 185
ASN 137	HIS H 168 THR H 187
ASN 138	HIS H 168
GLN 160	VAL H 173 LEU H 174 GLN H 175
SER 162	PHE H 170 PRO H 171 VAL H 173
THR 164	THR H 169 PHE H 170 PRO H 171
SER 174	HIS H 168 PHE H 170
SER 176	PHE H 170 SER H 183

^aContacting residues were identified using 4.5 Å distance limit criterion. The light chain numbering scheme corresponds to that in the deposited co-ordinates file (PDB code: 1NOX). The heavy chain numbering scheme corresponds to that in the Table 4.

[0049] In certain embodiments, Lys 125 of the lambda chain is mutated to a negatively charged amino acid and a corresponding mutation is made in a heavy chain at Asp148, changing the residue to a positively charged amino acid. Alternatively, or in addition, Glu 19 of the lambda chain is

mutated to a positively charged amino acid a corresponding mutation is made in a heavy chain at Lys213, changing the residue to a negatively charged amino acid.

[0050] The analysis of the light chain-heavy chain interaction revealed positions in which charge pairs could be introduced into the sequence to enhance binding of a specific light and heavy chain pair. These positions include Thr112 of lambda and A1a141 of the heavy chain, Glu156 of lambda and Ser176 of the heavy chain, and Ser171 of lambda and Ser183 of the heavy chain and other positions shown in Table 4 and 5 in bold face.

EXAMPLES

Example 1

[0051] This example demonstrates that CH3 domains can be engineered to favor heterodimerization while disfavoring homodimerization using electrostatic steering effects. A maxibody—dummy Fc construct as shown in FIG. 8(a) was made having charge residue mutations at the CH3 domain interface. The formation of homodimer and heterodimer yield was assessed through SDS polyacrylamide gel electrophoresis. Because the maxibody has a higher molecular weight compared to dummy Fc, the heterodimer (maxibody-dummy Fc) and homodimers (maxibody-maxibody & dummy Fc-dummy Fc) have different mobility on the SDS-PAGE facilitating the identification of the various pairings (FIG. 8(b)).

[0052] A rat anti-mouse NKG2D antibody, designated M315, was generated through conventional hybridoma fusions and the DNA sequences encoding the variable heavy chain (VH) and variable light chain (VL) were used to construct M315scFv-Fc using previously described method (Gilliland, Norris, et al. 1996).

[0053] The sequence of M315 scFv-Fc (SEQ ID NO:1) and huIgG1Fc (SEQ ID NO:2) were cloned into the pTT5 mammalian expression vector and the two constructs were used to co-transfect 293-6E cells to assess the formation Fc/scFv-Fc heterodimer relative to Fc homodimer and scFv-Fc homodimer.

SEQ ID NO: 1
M315scFv-huFc
HMAEVQLQQSGAELVKPGSSVKISCKASGYTFANNFMHWIKQQPGNGLEW
IGWIYPGDDTEYNQKFGKATLTADKSSSTAYMQLNSLTSEDAVYFCI
RLTEGTTWQGQGVMTVSSGGGGSGGGGGGGGSQFVLTQPNSVSTNLGS
TVKLSCKRSTGNIGSNYVNWYQQHEGRSPTTMIYRDDKRPDGVPDFSGS
IDGSSNSALLTINNVQTEDEADYPCQSYRSRGVSPVFGGTKLTVLXASPF
KSCDQKTHCPCPAPELPGPSVPLFPPKPKDILQIISRPPEVTCVWVDS
HEDPBYKPNWVWVWVWVWVWVWVWVWVWVWVWVWVWVWVWVWVWVWV
EYKCIWVSNKALPAPTEITISAKAKGPPERQWVTLPPSPEKNTINQVSLTC
LWKGKRYPSDIAVNEWESNGORENNKETTPPVLDSDGSEFLYSEKLTWIKSRN
GCGNVPSCGVWVWVWVWVWVWVWVWVWVWVWVWVWVWVWVWVWVWVWV

- continued

SEQ ID NO: 2

huIgG1Fc
 EPKSCDKTHTCPPCPAPELLGGPSVLFPPPKPKDTLMISRTPEVTCWWD
 VSHEDPEVKTIVWVDCGEVHNAKTKPREDQVNSTYRVSVLTIVHQDWLN
 GKEYKCKVSNKALPAPTEKTIKSKAKCOPPPPOVYTLPPSRREEMTQNOVSL
 TCLVKGEYFSEIATVWESNGCPENNYKTTPEVYLDSDGSFRFLYSKLTWIKS
 DNOGQWTFSCSYVMEAHBNRRTKQSLSSPCK

(Shading corresponds to the Fc region)

[0054] The charge residue pairs in the CH3 region identified through computational analysis were changed to amino acid of opposite charge polarity on either human IgG1Fc (dummy) or M315 scFv-Fc (mxb) constructs. The mutations, which are listed in Table 6, were generated using the QuikChange® mutagenesis kit from Stratagene and verified by DNA sequencing. The mutations are denoted by wild type residue followed by the position using the Kabat numbering system (Kabat et al., *Sequences of Proteins of Immunological Interest*, National Institutes of Health, Bethesda, Md., ed, 5, [1991]), which is consistent with the crystal structure (PDB code:1L6X) numbering scheme, and then the replacement residue in single letter code. The Fc sequence used in these two constructs was derived from human IgG1 non-(a) allotype, which has a Glu at position 356 and a Met at position 358. The CH3 sequences from the crystal structure are from a different IgG1 allotype, which has an Asp at position 356 and a Leu at position 368.

TABLE 6

List of charge residue mutations

huIgG1Fc (dummy)	M315 scFv-Fc(mxb)
Fc-WT	M315 scFv-Fc(WT)
K409D	D399'K
K409E	D399'R
K409D&K360D	D399'K&E356'K
K409D&K370D	D399'K&E357'K
K409D&K392D	D399'K&E356'K&E357'K
K409D&K439D	

[0055] DNA was transfected into human embryonic kidney cell line 293-6E using Lipofectamine™ 2000 reagent (Invitrogen). The cell culture supernatant was harvested 3-4 days after transfection and analyzed on SDS-PAGE Gels under non-reduced condition. The gel was then transferred to nitrocellulose membrane and subject to western analysis using peroxidase-conjugated goat anti-human IgG antibody (Jackson ImmunoResearch Laboratories) and results are shown in FIG. 10.

[0056] Co-transfection of expression vector for M315 scFv-Fc (mxb) together with dummy Fc resulted in the formation of scFv-Fc/Fc heterodimer as well as scFv-Fc homodimer and Fc homodimer. The ratio of scFv-Fc/Fc heterodimer to scFv-Fc homodimer and Fc homodimer is close to 1:1:1 when the wild type CH3 sequence is used.

[0057] The introduction of one charge pair mutation K409D on dummy Fc and D399'K on M315 maxibody significantly increased the ratio of scFv-Fc/Fc heterodimer relative to scFv-Fc homodimer as well as Fc homodimer. Similar enhancement of heterodimer formation was also observed for other mutant variants such as K409D/D399'R, K409E/D399'K and K409E/D399'R (Fig.9), further underscore the importance of charge polarity complementation for the formation of Fc heterodimers. (The wild type M315 scFv-Fc construct used in this study has an extra tag at the carboxyl terminal of Fc, so it migrates slower on the SDS-PAGE gel.)

[0058] When additional mutations were introduced at charge residues that are located near K409 such as K360 and K392, a further increase of heterodimer formation was observed (FIG. 10). For example, the combination K409D; K392D on dummy Fc with D399'K on M315 maxibody showed increased ratio of heterodimer to homodimers, likely due to the disruption of Fc homodimer. A 25KD band correspond to the size of Fc monomer was detected on all transfections using K409D;K392D dummy Fc (data not shown). Adding another mutation such as D356'K or D357'K on top of D399'K variant of M315 maxibody showed additional improvement. The combination of K409D;K392D on dummy Fc with

[0059] D399'K;D356'K on M315 maxibody resulted almost exclusive formation of heterodimer. Other combinations such as K409D;K392D/D399'K;D357'K and K409D; K370D/D399'K;D357'K also offered significant improvement over the K409D/D399'K variant.

TABLE 7

Quantification of percentage of homodimer and heterodimer yields for the SDS-PAGE shown in FIG. 10.^a

Dummy Fc Homodimer	M315 scFv-Fc-		M315 scFv-Fc Homodimer	M315 scFv-Fc	Dummy Fc
	Dummy Fc	Heterodimer			
42.1	32.4	25.5	WT	WT	
28.1	55.1	16.8	D399'K	K409D; K360D	
ND	76.9	23.1	D399'K	K409D; K392D	
ND	100	ND	D399'K; E356'K	K409D; K392D	
20.9	79.1	ND	D399'K; E357'K	K409D; K392D	
7.7	92.3	ND	D399'K; E356'K	K409D; K439D	
14.8	85.2	ND	D399'K; E357'K	K409D; K370D	
ND	86.7	13.3	T366'W (Hole)	T366S; L368A; Y407V (Knob)	

^aND stands for Not Detectable in the density based analysis.

Example 2

[0060] This example demonstrates that CH3 domains containing certain triple charge-pair mutations were unable to form homodimers when expressed alone but were capable of forming heterodimers when co-expressed. Mutants were made and cells transfected as described in Example 1. When the constructs were co-transfected, a 1:1 ratio of plasmids were used. The results are shown in FIG. 11. Heterodimer and homodimers were detected by Western blot using goat-anti-human Fc HRP conjugated antibody. Interestingly, Fc-containing molecules having triple mutations wherein positive-charged residues were changed to negative-charged residues (K409D,K392D,K370D or K409D,K392D,K439D) were unable to be detected when expressed alone. Similarly, Fc-containing molecules having triple mutations wherein negative-charged residues were changed to positive-charged residues (D399K,E356K, E357K) were unable to be detected when expressed alone. When co-expressed with an Fc-containing molecule having mutations of opposite charge polarity, however, heterodimers only were detected.

[0061] Throughout this invention application, it is to be understood that use of a term in the singular may imply, where appropriate, use of respective term in the plural, and vice versa.

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His Met Ala Glu Val Gln Leu Gln Gln Ser Gly Ala Glu Leu Val Lys
1 5 10 15

Pro Gly Ser Ser Val Lys Ile Ser Cys Lys Ala Ser Gly Tyr Thr Phe
20 25 30

Ala Asn Asn Phe Met His Trp Ile Lys Gln Gln Pro Gly Asn Gly Leu
35 40 45

Glu Trp Ile Gly Trp Ile Tyr Pro Gly Asp Gly Asp Thr Glu Tyr Asn
50 55 60

Gln Lys Phe Ser Gly Lys Ala Thr Leu Thr Ala Asp Lys Ser Ser Ser
65 70 75 80

Thr Ala Tyr Met Gln Leu Asn Ser Leu Thr Ser Glu Asp Ser Ala Val
85 90 95

Tyr Phe Cys Ile Arg Leu Thr Glu Gly Thr Thr Tyr Trp Gly Gln Gly
100 105 110

Val Met Val Thr Val Ser Ser Gly Gly Ser Gly Gly Gly Gly
115 120 125

Ser Gly Gly Gly Ser Gln Phe Val Leu Thr Gln Pro Asn Ser Val
130 135 140

Ser Thr Asn Leu Gly Ser Thr Val Lys Leu Ser Cys Lys Arg Ser Thr
145 150 155 160

Gly Asn Ile Gly Ser Asn Tyr Val Asn Trp Tyr Gln Gln His Glu Gly
165 170 175

Arg Ser Pro Thr Thr Met Ile Tyr Arg Asp Asp Lys Arg Pro Asp Gly
180 185 190

Val Pro Asp Arg Phe Ser Gly Ser Ile Asp Gly Ser Ser Asn Ser Ala
195 200 205

Leu Leu Thr Ile Asn Asn Val Gln Thr Glu Asp Glu Ala Asp Tyr Phe
210 215 220

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Cys Gln Ser Tyr Ser Arg Gly Val Ser Pro Val Phe Gly Gly Gly Thr
225 230 235 240

Lys Leu Thr Val Leu Ala Ala Ala Glu Pro Lys Ser Cys Asp Lys Thr
245 250 255

His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser
260 265 270

Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg
275 280 285

Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro
290 295 300

Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala
305 310 315 320

Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val
325 330 335

Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr
340 345 350

Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr
355 360 365

Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu
370 375 380

Pro Pro Ser Arg Glu Glu Met Thr Lys Asn Gln Val Ser Leu Thr Cys
385 390 395 400

Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser
405 410 415

Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp
420 425 430

Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser
435 440 445

Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala
450 455 460

Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys
465 470 475 480

<210> SEQ ID NO 2
 <211> LENGTH: 232
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <400> SEQUENCE: 2

Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala
1 5 10 15

Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro
20 25 30

Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val
35 40 45

Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val
50 55 60

Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln
65 70 75 80

Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln
85 90 95

Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala
100 105 110

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Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro
 115 120 125

Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu Met Thr
 130 135 140

Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser
 145 150 155 160

Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr
 165 170 175

Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr
 180 185 190

Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe
 195 200 205

Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys
 210 215 220

Ser Leu Ser Leu Ser Pro Gly Lys
 225 230

<210> SEQ ID NO 3
 <211> LENGTH: 330
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens

<400> SEQUENCE: 3

Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Ser Ser Lys
 1 5 10 15

Ser Thr Ser Gly Gly Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr
 20 25 30

Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser
 35 40 45

Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser
 50 55 60

Leu Ser Ser Val Val Thr Val Pro Ser Ser Ser Leu Gly Thr Gln Thr
 65 70 75 80

Tyr Ile Cys Asn Val Asn His Lys Pro Ser Asn Thr Lys Val Asp Lys
 85 90 95

Lys Val Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys
 100 105 110

Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro
 115 120 125

Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys
 130 135 140

Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp
 145 150 155 160

Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu
 165 170 175

Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu
 180 185 190

His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn
 195 200 205

Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly
 210 215 220

Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu
 225 230 235 240

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Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr
 245 250 255

Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn
 260 265 270

Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe
 275 280 285

Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn
 290 295 300

Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr
 305 310 315 320

Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys
 325 330

<210> SEQ ID NO 4
 <211> LENGTH: 326
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens

<400> SEQUENCE: 4

Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Cys Ser Arg
 1 5 10 15

Ser Thr Ser Glu Ser Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr
 20 25 30

Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser
 35 40 45

Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser
 50 55 60

Leu Ser Ser Val Val Thr Val Pro Ser Ser Asn Phe Gly Thr Gln Thr
 65 70 75 80

Tyr Thr Cys Asn Val Asp His Lys Pro Ser Asn Thr Lys Val Asp Lys
 85 90 95

Thr Val Glu Arg Lys Cys Cys Val Glu Cys Pro Pro Cys Pro Ala Pro
 100 105 110

Pro Val Ala Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp
 115 120 125

Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp
 130 135 140

Val Ser His Glu Asp Pro Glu Val Gln Phe Asn Trp Tyr Val Asp Gly
 145 150 155 160

Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Phe Asn
 165 170 175

Ser Thr Phe Arg Val Val Ser Val Leu Thr Val Val His Gln Asp Trp
 180 185 190

Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Gly Leu Pro
 195 200 205

Ala Pro Ile Glu Lys Thr Ile Ser Lys Thr Lys Gly Gln Pro Arg Glu
 210 215 220

Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu Met Thr Lys Asn
 225 230 235 240

Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile
 245 250 255

Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr
 260 265 270

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Thr Pro Pro Met Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys
275 280 285

Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys
290 295 300

Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu
305 310 315 320

Ser Leu Ser Pro Gly Lys
325

<210> SEQ ID NO 5

<211> LENGTH: 377

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 5

Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Cys Ser Arg
1 5 10 15

Ser Thr Ser Gly Gly Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr
20 25 30

Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser
35 40 45

Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser
50 55 60

Leu Ser Ser Val Val Thr Val Pro Ser Ser Ser Leu Gly Thr Gln Thr
65 70 75 80

Tyr Thr Cys Asn Val Asn His Lys Pro Ser Asn Thr Lys Val Asp Lys
85 90 95

Arg Val Glu Leu Lys Thr Pro Leu Gly Asp Thr Thr His Thr Cys Pro
100 105 110

Arg Cys Pro Glu Pro Lys Ser Cys Asp Thr Pro Pro Pro Cys Pro Arg
115 120 125

Cys Pro Glu Pro Lys Ser Cys Asp Thr Pro Pro Pro Cys Pro Arg Cys
130 135 140

Pro Glu Pro Lys Ser Cys Asp Thr Pro Pro Pro Cys Pro Arg Cys Pro
145 150 155 160

Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys
165 170 175

Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val
180 185 190

Val Val Asp Val Ser His Glu Asp Pro Glu Val Gln Phe Lys Trp Tyr
195 200 205

Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu
210 215 220

Gln Tyr Asn Ser Thr Phe Arg Val Val Ser Val Leu Thr Val Leu His
225 230 235 240

Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys
245 250 255

Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Thr Lys Gly Gln
260 265 270

Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu Met
275 280 285

Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro
290 295 300

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Ser Asp Ile Ala Val Glu Trp Glu Ser Ser Gly Gln Pro Glu Asn Asn
 305 310 315 320
 Tyr Asn Thr Thr Pro Pro Met Leu Asp Ser Asp Gly Ser Phe Phe Leu
 325 330 335
 Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Ile
 340 345 350
 Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn Arg Phe Thr Gln
 355 360 365
 Lys Ser Leu Ser Leu Ser Pro Gly Lys
 370 375

<210> SEQ ID NO 6
 <211> LENGTH: 327
 <212> TYPE: PRT
 <213> ORGANISM: homo sapiens
 <400> SEQUENCE: 6

Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Cys Ser Arg
 1 5 10 15
 Ser Thr Ser Glu Ser Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr
 20 25 30
 Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser
 35 40 45
 Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser
 50 55 60
 Leu Ser Ser Val Val Thr Val Pro Ser Ser Ser Leu Gly Thr Lys Thr
 65 70 75 80
 Tyr Thr Cys Asn Val Asp His Lys Pro Ser Asn Thr Lys Val Asp Lys
 85 90 95
 Arg Val Glu Ser Lys Tyr Gly Pro Pro Cys Pro Ser Cys Pro Ala Pro
 100 105 110
 Glu Phe Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys
 115 120 125
 Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val
 130 135 140
 Asp Val Ser Gln Glu Asp Pro Glu Val Gln Phe Asn Trp Tyr Val Asp
 145 150 155 160
 Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Phe
 165 170 175
 Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp
 180 185 190
 Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Gly Leu
 195 200 205
 Pro Ser Ser Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg
 210 215 220
 Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Gln Glu Glu Met Thr Lys
 225 230 235 240
 Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp
 245 250 255
 Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys
 260 265 270
 Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser
 275 280 285

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Arg Leu Thr Val Asp Lys Ser Arg Trp Gln Glu Gly Asn Val Phe Ser
290 295 300

Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser
305 310 315 320

Leu Ser Leu Ser Leu Gly Lys
325

<210> SEQ ID NO 7

<211> LENGTH: 353

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 7

Ala Ser Pro Thr Ser Pro Lys Val Phe Pro Leu Ser Leu Cys Ser Thr
1 5 10 15

Gln Pro Asp Gly Asn Val Val Ile Ala Cys Leu Val Gln Gly Phe Phe
20 25 30

Pro Gln Glu Pro Leu Ser Val Thr Trp Ser Glu Ser Gly Gln Gly Val
35 40 45

Thr Ala Arg Asn Phe Pro Pro Ser Gln Asp Ala Ser Gly Asp Leu Tyr
50 55 60

Thr Thr Ser Ser Gln Leu Thr Leu Pro Ala Thr Gln Cys Leu Ala Gly
65 70 75 80

Lys Ser Val Thr Cys His Val Lys His Tyr Thr Asn Pro Ser Gln Asp
85 90 95

Val Thr Val Pro Cys Pro Val Pro Ser Thr Pro Pro Thr Pro Ser Pro
100 105 110

Ser Thr Pro Pro Thr Pro Ser Pro Ser Cys Cys His Pro Arg Leu Ser
115 120 125

Leu His Arg Pro Ala Leu Glu Asp Leu Leu Leu Gly Ser Glu Ala Asn
130 135 140

Leu Thr Cys Thr Leu Thr Gly Leu Arg Asp Ala Ser Gly Val Thr Phe
145 150 155 160

Thr Trp Thr Pro Ser Ser Gly Lys Ser Ala Val Gln Gly Pro Pro Glu
165 170 175

Arg Asp Leu Cys Gly Cys Tyr Ser Val Ser Ser Val Leu Pro Gly Cys
180 185 190

Ala Glu Pro Trp Asn His Gly Lys Thr Phe Thr Cys Thr Ala Ala Tyr
195 200 205

Pro Glu Ser Lys Thr Pro Leu Thr Ala Thr Leu Ser Lys Ser Gly Asn
210 215 220

Thr Phe Arg Pro Glu Val His Leu Leu Pro Pro Ser Glu Glu Leu
225 230 235 240

Ala Leu Asn Glu Leu Val Thr Leu Thr Cys Leu Ala Arg Gly Phe Ser
245 250 255

Pro Lys Asp Val Leu Val Arg Trp Leu Gln Gly Ser Gln Glu Leu Pro
260 265 270

Arg Glu Lys Tyr Leu Thr Trp Ala Ser Arg Gln Glu Pro Ser Gln Gly
275 280 285

Thr Thr Thr Phe Ala Val Thr Ser Ile Leu Arg Val Ala Ala Glu Asp
290 295 300

Trp Lys Lys Gly Asp Thr Phe Ser Cys Met Val Gly His Glu Ala Leu
305 310 315 320

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Pro Leu Ala Phe Thr Gln Lys Thr Ile Asp Arg Leu Ala Gly Lys Pro
325 330 335

Thr His Val Asn Val Ser Val Val Met Ala Glu Val Asp Gly Thr Cys
340 345 350

Tyr

<210> SEQ ID NO 8
<211> LENGTH: 384
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 8

Ala Pro Thr Lys Ala Pro Asp Val Phe Pro Ile Ile Ser Gly Cys Arg
1 5 10 15

His Pro Lys Asp Asn Ser Pro Val Val Leu Ala Cys Leu Ile Thr Gly
20 25 30

Tyr His Pro Thr Ser Val Thr Val Thr Trp Tyr Met Gly Thr Gln Ser
35 40 45

Gln Pro Gln Arg Thr Phe Pro Glu Ile Gln Arg Arg Asp Ser Tyr Tyr
50 55 60

Met Thr Ser Ser Gln Leu Ser Thr Pro Leu Gln Gln Trp Arg Gln Gly
65 70 75 80

Glu Tyr Lys Cys Val Val Gln His Thr Ala Ser Lys Ser Lys Lys Glu
85 90 95

Ile Phe Arg Trp Pro Glu Ser Pro Lys Ala Gln Ala Ser Ser Val Pro
100 105 110

Thr Ala Gln Pro Gln Ala Glu Gly Ser Leu Ala Lys Ala Thr Thr Ala
115 120 125

Pro Ala Thr Thr Arg Asn Thr Gly Arg Gly Glu Glu Lys Lys Lys
130 135 140

Glu Lys Glu Lys Glu Glu Gln Glu Glu Arg Glu Thr Lys Thr Pro Glu
145 150 155 160

Cys Pro Ser His Thr Gln Pro Leu Gly Val Tyr Leu Leu Thr Pro Ala
165 170 175

Val Gln Asp Leu Trp Leu Arg Asp Lys Ala Thr Phe Thr Cys Phe Val
180 185 190

Val Gly Ser Asp Leu Lys Asp Ala His Leu Thr Trp Glu Val Ala Gly
195 200 205

Lys Val Pro Thr Gly Gly Val Glu Glu Gly Leu Leu Glu Arg His Ser
210 215 220

Asn Gly Ser Gln Ser Gln His Ser Arg Leu Thr Leu Pro Arg Ser Leu
225 230 235 240

Trp Asn Ala Gly Thr Ser Val Thr Cys Thr Leu Asn His Pro Ser Leu
245 250 255

Pro Pro Gln Arg Leu Met Ala Leu Arg Glu Pro Ala Ala Gln Ala Pro
260 265 270

Val Lys Leu Ser Leu Asn Leu Leu Ala Ser Ser Asp Pro Pro Glu Ala
275 280 285

Ala Ser Trp Leu Leu Cys Glu Val Ser Gly Phe Ser Pro Pro Asn Ile
290 295 300

Leu Leu Met Trp Leu Glu Asp Gln Arg Glu Val Asn Thr Ser Gly Phe
305 310 315 320

Ala Pro Ala Arg Pro Pro Pro Gln Pro Gly Ser Thr Thr Phe Trp Ala

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325	330	335
Trp Ser Val Leu Arg Val Pro Ala Pro Pro Ser Pro Gln Pro Ala Thr		
340	345	350
Tyr Thr Cys Val Val Ser His Glu Asp Ser Arg Thr Leu Leu Asn Ala		
355	360	365
Ser Arg Ser Leu Glu Val Ser Tyr Val Thr Asp His Gly Pro Met Lys		
370	375	380
<210> SEQ ID NO 9		
<211> LENGTH: 428		
<212> TYPE: PRT		
<213> ORGANISM: Homo sapains		
<400> SEQUENCE: 9		
Ala Ser Thr Gln Ser Pro Ser Val Phe Pro Leu Thr Arg Cys Cys Lys		
1	5	10
15		
Asn Ile Pro Ser Asn Ala Thr Ser Val Thr Leu Gly Cys Leu Ala Thr		
20	25	30
Gly Tyr Phe Pro Glu Pro Val Met Val Thr Trp Asp Thr Gly Ser Leu		
35	40	45
Asn Gly Thr Thr Met Thr Leu Pro Ala Thr Thr Leu Thr Leu Ser Gly		
50	55	60
His Tyr Ala Thr Ile Ser Leu Leu Thr Val Ser Gly Ala Trp Ala Lys		
65	70	75
80		
Gln Met Phe Thr Cys Arg Val Ala His Thr Pro Ser Ser Thr Asp Trp		
85	90	95
Val Asp Asn Lys Thr Phe Ser Val Cys Ser Arg Asp Phe Thr Pro Pro		
100	105	110
Thr Val Lys Ile Leu Gln Ser Ser Cys Asp Gly Gly His Phe Pro		
115	120	125
Pro Thr Ile Gln Leu Leu Cys Leu Val Ser Gly Tyr Thr Pro Gly Thr		
130	135	140
Ile Asn Ile Thr Trp Leu Glu Asp Gly Gln Val Met Asp Val Asp Leu		
145	150	155
160		
Ser Thr Ala Ser Thr Thr Gln Glu Gly Glu Leu Ala Ser Thr Gln Ser		
165	170	175
Glu Leu Thr Leu Ser Gln Lys His Trp Leu Ser Asp Arg Thr Tyr Thr		
180	185	190
Cys Gln Val Thr Tyr Gln Gly His Thr Phe Glu Asp Ser Thr Lys Lys		
195	200	205
Cys Ala Asp Ser Asn Pro Arg Gly Val Ser Ala Tyr Leu Ser Arg Pro		
210	215	220
Ser Pro Phe Asp Leu Phe Ile Arg Lys Ser Pro Thr Ile Thr Cys Leu		
225	230	235
240		
Val Val Asp Leu Ala Pro Ser Lys Gly Thr Val Asn Leu Thr Trp Ser		
245	250	255
Arg Ala Ser Gly Lys Pro Val Asn His Ser Thr Arg Lys Glu Glu Lys		
260	265	270
Gln Arg Asn Gly Thr Leu Thr Val Thr Ser Thr Leu Pro Val Gly Thr		
275	280	285
Arg Asp Trp Ile Glu Gly Glu Thr Tyr Gln Cys Arg Val Thr His Pro		
290	295	300
His Leu Pro Arg Ala Leu Met Arg Ser Thr Thr Lys Thr Ser Gly Pro		

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305	310	315	320
Arg Ala Ala Pro Glu Val Tyr Ala Phe Ala Thr Pro Glu Trp Pro Gly			
325	330	335	
Ser Arg Asp Lys Arg Thr Leu Ala Cys Leu Ile Gln Asn Phe Met Pro			
340	345	350	
Glu Asp Ile Ser Val Gln Trp Leu His Asn Glu Val Gln Leu Pro Asp			
355	360	365	
Ala Arg His Ser Thr Thr Gln Pro Arg Lys Thr Lys Gly Ser Gly Phe			
370	375	380	
Phe Val Phe Ser Arg Leu Glu Val Thr Arg Ala Glu Trp Glu Gln Lys			
385	390	395	400
Asp Glu Phe Ile Cys Arg Ala Val His Glu Ala Ala Ser Pro Ser Gln			
405	410	415	
Thr Val Gln Arg Ala Val Ser Val Asn Pro Gly Lys			
420	425		

<210> SEQ ID NO 10

<211> LENGTH: 452

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 10

Gly Ser Ala Ser Ala Pro Thr Leu Phe Pro Leu Val Ser Cys Glu Asn			
1	5	10	15

Ser Pro Ser Asp Thr Ser Ser Val Ala Val Gly Cys Leu Ala Gln Asp			
20	25	30	

Phe Leu Pro Asp Ser Ile Thr Leu Ser Trp Lys Tyr Lys Asn Asn Ser			
35	40	45	

Asp Ile Ser Ser Thr Arg Gly Phe Pro Ser Val Leu Arg Gly Gly Lys			
50	55	60	

Tyr Ala Ala Thr Ser Gln Val Leu Pro Ser Lys Asp Val Met Gln			
65	70	75	80

Gly Thr Asp Glu His Val Val Cys Lys Val Gln His Pro Asn Gly Asn			
85	90	95	

Lys Glu Lys Asn Val Pro Leu Pro Val Ile Ala Glu Leu Pro Pro Lys			
100	105	110	

Val Ser Val Phe Val Pro Pro Arg Asp Gly Phe Phe Gly Asn Pro Arg			
115	120	125	

Lys Ser Lys Leu Ile Cys Gln Ala Thr Gly Phe Ser Pro Arg Gln Ile			
130	135	140	

Gln Val Ser Trp Leu Arg Glu Gly Lys Gln Val Gly Ser Gly Val Thr			
145	150	155	160

Thr Asp Gln Val Gln Ala Glu Ala Lys Glu Ser Gly Pro Thr Thr Tyr			
165	170	175	

Lys Val Thr Ser Thr Leu Thr Ile Lys Glu Ser Asp Trp Leu Gly Gln			
180	185	190	

Ser Met Phe Thr Cys Arg Val Asp His Arg Gly Leu Thr Phe Gln Gln			
195	200	205	

Asn Ala Ser Ser Met Cys Val Pro Asp Gln Asp Thr Ala Ile Arg Val			
210	215	220	

Phe Ala Ile Pro Pro Ser Phe Ala Ser Ile Phe Leu Thr Lys Ser Thr			
225	230	235	240

Lys Leu Thr Cys Leu Val Thr Asp Leu Thr Thr Tyr Asp Ser Val Thr

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245	250	255
Ile Ser Trp Thr Arg Gln Asn Gly Glu Ala Val Lys Thr His Thr Asn 260	265	270
Ile Ser Glu Ser His Pro Asn Ala Thr Phe Ser Ala Val Gly Glu Ala 275	280	285
Ser Ile Cys Glu Asp Asp Trp Asn Ser Gly Glu Arg Phe Thr Cys Thr 290	295	300
Val Thr His Thr Asp Leu Pro Ser Pro Leu Lys Gln Thr Ile Ser Arg 305	310	315
Pro Lys Gly Val Ala Leu His Arg Pro Asp Val Tyr Leu Leu Pro Pro 325	330	335
Ala Arg Glu Gln Leu Asn Leu Arg Glu Ser Ala Thr Ile Thr Cys Leu 340	345	350
Val Thr Gly Phe Ser Pro Ala Asp Val Phe Val Gln Trp Met Gln Arg 355	360	365
Gly Gln Pro Leu Ser Pro Glu Lys Tyr Val Thr Ser Ala Pro Met Pro 370	375	380
Glu Pro Gln Ala Pro Gly Arg Tyr Phe Ala His Ser Ile Leu Thr Val 385	390	395
Ser Glu Glu Glu Trp Asn Thr Gly Glu Thr Tyr Thr Cys Val Ala His 405	410	415
Glu Ala Leu Pro Asn Arg Val Thr Glu Arg Thr Val Asp Lys Ser Thr 420	425	430
Gly Lys Pro Thr Leu Tyr Asn Val Ser Leu Val Met Ser Asp Thr Ala 435	440	445
Gly Thr Cys Tyr 450		

<210> SEQ ID NO 11

<211> LENGTH: 109

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 11

Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser 1	5	10	15
Arg Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys 20	25	30	
Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln 35	40	45	
Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly 50	55	60	
Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln 65	70	75	80
Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn 85	90	95	
His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys 100	105		

<210> SEQ ID NO 12

<211> LENGTH: 109

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 12

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Thr Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser
 1 5 10 15
 Arg Glu Glu Met Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys
 20 25 30
 Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln
 35 40 45
 Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Met Leu Asp Ser Asp Gly
 50 55 60
 Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln
 65 70 75 80
 Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn
 85 90 95
 His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys
 100 105

<210> SEQ ID NO 13
 <211> LENGTH: 109
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens

<400> SEQUENCE: 13

Thr Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser
 1 5 10 15
 Arg Glu Glu Met Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys
 20 25 30
 Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Ser Gly Gln
 35 40 45
 Pro Glu Asn Asn Tyr Asn Thr Thr Pro Pro Met Leu Asp Ser Asp Gly
 50 55 60
 Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln
 65 70 75 80
 Gln Gly Asn Ile Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn
 85 90 95
 Arg Phe Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys
 100 105

<210> SEQ ID NO 14
 <211> LENGTH: 109
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens

<400> SEQUENCE: 14

Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser
 1 5 10 15
 Gln Glu Glu Met Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys
 20 25 30
 Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln
 35 40 45
 Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly
 50 55 60
 Ser Phe Phe Leu Tyr Ser Arg Leu Thr Val Asp Lys Ser Arg Trp Gln
 65 70 75 80
 Glu Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn
 85 90 95

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His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Leu Gly Lys
 100 105

<210> SEQ ID NO 15
 <211> LENGTH: 106
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <400> SEQUENCE: 15

Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser
 1 5 10 15

Arg Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys
 20 25 30

Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln
 35 40 45

Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly
 50 55 60

Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln
 65 70 75 80

Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn
 85 90 95

His Tyr Thr Gln Lys Ser Leu Ser Leu Ser
 100 105

<210> SEQ ID NO 16
 <211> LENGTH: 106
 <212> TYPE: PRT
 <213> ORGANISM: Mus musculus
 <400> SEQUENCE: 16

Thr Lys Gly Arg Pro Lys Ala Pro Gln Val Tyr Thr Ile Pro Pro Pro
 1 5 10 15

Lys Glu Gln Met Ala Lys Asp Lys Val Ser Leu Thr Cys Met Ile Thr
 20 25 30

Asp Phe Phe Pro Glu Asp Ile Thr Val Glu Trp Gln Trp Asn Gly Gln
 35 40 45

Pro Ala Glu Asn Tyr Lys Asn Thr Gln Pro Ile Met Asn Thr Asn Gly
 50 55 60

Ser Tyr Phe Val Tyr Ser Lys Leu Asn Val Gln Lys Ser Asn Trp Glu
 65 70 75 80

Ala Gly Asn Thr Phe Thr Cys Ser Val Leu His Glu Gly Leu His Asn
 85 90 95

His His Thr Glu Lys Ser Leu Ser His Ser
 100 105

<210> SEQ ID NO 17
 <211> LENGTH: 106
 <212> TYPE: PRT
 <213> ORGANISM: Mus musculus
 <400> SEQUENCE: 17

Pro Lys Gly Ser Val Arg Ala Pro Gln Val Tyr Val Leu Pro Pro Pro
 1 5 10 15

Glu Glu Glu Met Thr Lys Lys Gln Val Thr Leu Thr Cys Met Val Thr
 20 25 30

Asp Phe Met Pro Glu Asp Ile Tyr Val Glu Trp Thr Asn Asn Gly Lys
 35 40 45

-continued

Thr Glu Leu Asn Tyr Lys Asn Thr Glu Pro Val Leu Asp Ser Asp Gly
 50 55 60

Ser Tyr Phe Met Tyr Ser Lys Leu Arg Val Glu Lys Lys Asn Trp Val
 65 70 75 80

Glu Arg Asn Ser Tyr Ser Cys Ser Val Val His Glu Gly Leu His Asn
 85 90 95

His His Thr Thr Lys Ser Phe Ser Arg Thr
 100 105

<210> SEQ ID NO 18
 <211> LENGTH: 106
 <212> TYPE: PRT
 <213> ORGANISM: Mus musculus

<400> SEQUENCE: 18

Ile Lys Gly Leu Val Arg Ala Pro Gln Val Tyr Thr Leu Pro Pro Pro
 1 5 10 15

Ala Glu Gln Leu Ser Arg Lys Asp Val Ser Leu Thr Cys Leu Val Val
 20 25 30

Gly Phe Asn Pro Gly Asp Ile Ser Val Glu Trp Thr Ser Asn Gly His
 35 40 45

Thr Glu Glu Asn Tyr Lys Asp Thr Ala Pro Val Leu Asp Ser Asp Gly
 50 55 60

Ser Tyr Phe Ile Tyr Ser Lys Leu Asn Met Lys Thr Ser Lys Trp Glu
 65 70 75 80

Lys Thr Asp Ser Phe Ser Cys Asn Val Arg His Glu Gly Leu Lys Asn
 85 90 95

Tyr Tyr Leu Lys Lys Thr Ile Ser Arg Ser
 100 105

<210> SEQ ID NO 19
 <211> LENGTH: 106
 <212> TYPE: PRT
 <213> ORGANISM: Mus musculus

<400> SEQUENCE: 19

Pro Lys Gly Arg Ala Gln Thr Pro Gln Val Tyr Thr Ile Pro Pro Pro
 1 5 10 15

Arg Glu Gln Met Ser Lys Lys Val Ser Leu Thr Cys Leu Val Thr
 20 25 30

Asn Phe Phe Ser Glu Ala Ile Ser Val Glu Trp Glu Arg Asn Gly Glu
 35 40 45

Leu Glu Gln Asp Tyr Lys Asn Thr Pro Pro Ile Leu Asp Ser Asp Gly
 50 55 60

Thr Tyr Phe Leu Tyr Ser Lys Leu Thr Val Asp Thr Asp Ser Trp Leu
 65 70 75 80

Gln Gly Glu Ile Phe Thr Cys Ser Val Val His Glu Ala Leu His Asn
 85 90 95

His His Thr Gln Lys Asn Leu Ser Arg Ser
 100 105

<210> SEQ ID NO 20
 <211> LENGTH: 105
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens

-continued

<400> SEQUENCE: 20

Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser
 1 5 10 15

Arg Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys
 20 25 30

Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln
 35 40 45

Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly
 50 55 60

Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln
 65 70 75 80

Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn
 85 90 95

His Tyr Thr Gln Lys Ser Leu Ser Leu
 100 105

<210> SEQ ID NO 21

<211> LENGTH: 114

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 21

Ser Gly Asn Thr Phe Arg Pro Glu Val His Leu Leu Pro Pro Pro Ser
 1 5 10 15

Glu Glu Leu Ala Leu Asn Glu Leu Val Thr Leu Thr Cys Leu Ala Arg
 20 25 30

Gly Phe Ser Pro Lys Asp Val Leu Val Arg Trp Leu Gln Gly Ser Gln
 35 40 45

Glu Leu Pro Arg Glu Lys Tyr Leu Thr Trp Ala Ser Arg Gln Glu Pro
 50 55 60

Ser Gln Gly Thr Thr Phe Ala Val Thr Ser Ile Leu Arg Val Ala
 65 70 75 80

Ala Glu Asp Trp Lys Lys Gly Asp Thr Phe Ser Cys Met Val Gly His
 85 90 95

Glu Ala Leu Pro Leu Ala Phe Thr Gln Lys Thr Ile Asp Arg Leu Ala
 100 105 110

Gly Lys

<210> SEQ ID NO 22

<211> LENGTH: 112

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 22

Thr Ser Gly Pro Arg Ala Ala Pro Glu Val Tyr Ala Phe Ala Thr Pro
 1 5 10 15

Glu Trp Pro Gly Ser Arg Asp Lys Arg Thr Leu Ala Cys Leu Ile Gln
 20 25 30

Asn Phe Met Pro Glu Asp Ile Ser Val Gln Trp Leu His Asn Glu Val
 35 40 45

Gln Leu Pro Asp Ala Arg His Ser Thr Thr Gln Pro Arg Lys Thr Lys
 50 55 60

Gly Ser Gly Phe Phe Val Phe Ser Arg Leu Glu Val Thr Arg Ala Glu
 65 70 75 80

-continued

Trp Glu Gln Lys Asp Glu Phe Ile Cys Arg Ala Val His Glu Ala Ala
 85 90 95

Ser Pro Ser Gln Thr Val Gln Arg Ala Val Ser Val Asn Pro Gly Lys
 100 105 110

<210> SEQ ID NO 23

<211> LENGTH: 114

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 23

Arg Glu Pro Ala Ala Gln Ala Pro Val Lys Leu Ser Leu Asn Leu Leu
 1 5 10 15

Ala Ser Ser Asp Pro Pro Glu Ala Ala Ser Trp Leu Leu Cys Glu Val
 20 25 30

Ser Gly Phe Ser Pro Pro Asn Ile Leu Leu Met Trp Leu Glu Asp Gln
 35 40 45

Arg Glu Val Asn Thr Ser Gly Phe Ala Pro Ala Arg Pro Pro Pro Gln
 50 55 60

Pro Gly Ser Thr Thr Phe Trp Ala Trp Ser Val Leu Arg Val Pro Ala
 65 70 75 80

Pro Pro Ser Pro Gln Pro Ala Thr Tyr Thr Cys Val Val Ser His Glu
 85 90 95

Asp Ser Arg Thr Leu Leu Asn Ala Ser Arg Ser Leu Glu Val Ser Tyr
 100 105 110

Val Thr

<210> SEQ ID NO 24

<211> LENGTH: 115

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 24

Pro Lys Gly Val Ala Leu His Arg Pro Asp Val Tyr Leu Leu Pro Pro
 1 5 10 15

Ala Arg Glu Gln Leu Asn Leu Arg Glu Ser Ala Thr Ile Thr Cys Leu
 20 25 30

Val Thr Gly Phe Ser Pro Ala Asp Val Phe Val Gln Trp Met Gln Arg
 35 40 45

Gly Gln Pro Leu Ser Pro Glu Lys Tyr Val Thr Ser Ala Pro Met Pro
 50 55 60

Glu Pro Gln Ala Pro Gly Arg Tyr Phe Ala His Ser Ile Leu Thr Val
 65 70 75 80

Ser Glu Glu Glu Trp Asn Thr Gly Glu Thr Tyr Thr Cys Val Val Ala
 85 90 95

His Glu Ala Leu Pro Asn Arg Val Thr Glu Arg Thr Val Asp Lys Ser
 100 105 110

-continued

Thr Gly Lys
115

<210> SEQ ID NO 25
<211> LENGTH: 107
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 25

Arg Thr Val Ala Ala Pro Ser Val Phe Ile Phe Pro Pro Ser Asp Glu
1 5 10 15
Gln Leu Lys Ser Gly Thr Ala Ser Val Val Cys Leu Leu Asn Asn Phe
20 25 30
Tyr Pro Arg Glu Ala Lys Val Gln Trp Lys Val Asp Asn Ala Leu Gln
35 40 45
Ser Gly Asn Ser Gln Glu Ser Val Thr Glu Gln Asp Ser Lys Asp Ser
50 55 60
Thr Tyr Ser Leu Ser Ser Thr Leu Thr Leu Ser Lys Ala Asp Tyr Glu
65 70 75 80
Lys His Lys Val Tyr Ala Cys Glu Val Thr His Gln Gly Leu Arg Ser
85 90 95
Pro Val Thr Lys Ser Phe Asn Arg Gly Glu Cys
100 105

<210> SEQ ID NO 26
<211> LENGTH: 105
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 26

Gln Pro Lys Ala Ala Pro Ser Val Thr Leu Phe Pro Pro Ser Ser Glu
1 5 10 15
Glu Leu Gln Ala Asn Lys Ala Thr Leu Val Cys Leu Ile Ser Asp Phe
20 25 30
Tyr Pro Gly Ala Val Thr Val Ala Trp Lys Ala Asp Gly Ser Pro Val
35 40 45
Lys Ala Gly Val Glu Thr Thr Pro Ser Lys Gln Ser Asn Asn Lys
50 55 60
Tyr Ala Ala Ser Ser Tyr Leu Ser Leu Thr Pro Glu Gln Trp Lys Ser
65 70 75 80
His Lys Ser Tyr Ser Cys Gln Val Thr His Glu Gly Ser Thr Val Glu
85 90 95
Lys Thr Val Ala Pro Thr Glu Cys Ser
100 105

<210> SEQ ID NO 27
<211> LENGTH: 100
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 27

Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Ser Ser Lys
1 5 10 15
Ser Thr Ser Gly Gly Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr
20 25 30
Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser

-continued

35	40	45
Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser		
50	55	60
Leu Ser Ser Val Val Thr Val Pro Ser Ser Ser Leu Gly Thr Gln Thr		
65	70	75
Tyr Ile Cys Asn Val Asn His Lys Pro Ser Asn Thr Lys Val Asp Lys		
85	90	95
Lys Val Glu Pro		
	100	

1. A host cell for producing a heterodimeric protein, said host cell comprising a nucleic acid encoding a first CH3-containing polypeptide and a nucleic acid encoding a second CH3-containing polypeptide, wherein said first human CH3-containing polypeptide comprises a replacement of the amino acid at position 392 with a negative-charged amino acid and said second human IgG CH3-containing polypeptide comprises a replacement of Asp399, Glu356, Asp356, or Glu357 with a positive-charged amino acid.

2. The host cell of claim **1**, wherein Lys392 is replaced with a negative-charged amino acid.

3. The host cell of claim **1**, wherein Asn392 is replaced with a negative-charged amino acid.

4. The host cell of claim **1**, wherein said first human CH3-containing polypeptide further comprises Lys409 or Arg409 replaced with a negative-charged amino acid.

5. The host cell of claim **1**, wherein Lys392 or Asn392 is replaced with aspartic acid.

6. The host cell of claim **4**, wherein said Lys409 or Arg409 is replaced with aspartic acid.

7. The host cell of claim **1**, wherein said second human IgG CH3-containing polypeptide comprises a replacement of Asp399, Glu356, Asp356, or Glu357 with lysine.

8. The host cell of claim **1**, wherein said second human IgG CH3-containing polypeptide comprises a replacement of Asp399 and Glu356 with lysine.

9. The host cell of claim **1**, wherein the heterodimeric protein comprises a human IgG Fc region.

10. The host cell of claim **9**, wherein the human IgG Fc region comprises an IgG1 Fc region.

11. The host cell of claim **9**, wherein the IgG Fc region comprises an IgG2 Fc region.

12. The host cell of claim **9**, wherein the IgG Fc region comprises an IgG3 Fc region.

13. The host cell of claim **9**, wherein the IgG Fc region comprises an IgG4 Fc region.

14. The host cell of claim **1**, wherein the first CH3-containing polypeptide is an antibody heavy chain.

15. The host cell of claim **1**, wherein the second CH3-containing polypeptide is an antibody heavy chain.

16. The host cell of claim **1**, wherein the heterodimeric protein further comprises one or more antibody light chains.

17. The host cell of claim **1**, wherein the heterodimeric protein is selected from the group consisting of an antibody, a bispecific antibody, a monospecific monovalent antibody, a

bispecific maxibody, a monobody, a peptibody, a bispecific peptibody, a monovalent peptibody, and a receptor fusion protein.

18. The host cell of claim **1**, wherein the host cell is a mammalian host cell.

19. The host cell of claim **18**, wherein the mammalian host cell is a Chinese hamster ovary (CHO) cell line.

20. A host cell for producing a heterodimeric protein, said host cell comprising a nucleic acid encoding a first CH3-containing polypeptide and a nucleic acid encoding a second CH3-containing polypeptide, wherein said first CH3-containing polypeptide comprises replacement of the amino acids at positions 392 and 409 with a negative-charged amino acid and said second CH3-containing polypeptide comprises replacement of the amino acids at positions 356 and 399 with a positive-charged amino acid.

21. The host cell of claim **20**, wherein the negative charged amino acid is aspartic acid.

22. The host cell of claim **20**, wherein the positive charged amino acid is lysine.

23. The host cell of claim **21**, wherein the positive charged amino acid is lysine.

24. The host cell of claim **20**, wherein the mammalian host cell is a Chinese hamster ovary (CHO) cell line.

25. A host cell for producing a heterodimeric protein, said host cell comprising a nucleic acid encoding a first CH3-containing polypeptide and a nucleic acid encoding a second CH3-containing polypeptide, wherein the first CH3-containing polypeptide comprises replacement of the amino acids at positions 370, 392, and 409 with a negative-charged amino acid and said second CH3-containing polypeptide comprises replacement of the amino acids at positions 356, 357, and 399 with a positive-charged amino acid.

26. The host cell of claim **25**, wherein the negative charged amino acid is aspartic acid.

27. The host cell of claim **25**, wherein the positive charged amino acid is lysine.

28. The host cell of claim **26**, wherein the positive charged amino acid is lysine.

29. The host cell of claim **25**, wherein the mammalian host cell is a Chinese hamster ovary (CHO) cell line.

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