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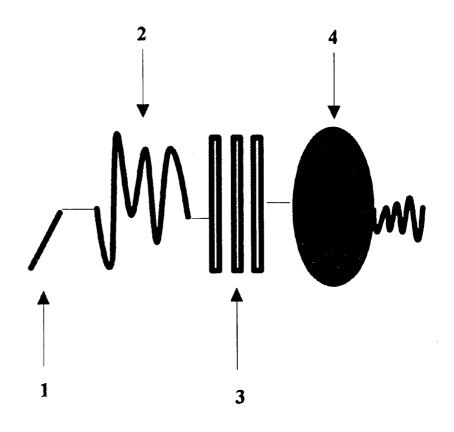
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## (54) Title: HYBRID PROTEINS HAVING CROSS-LINKING AND TISSUE-BINDING ACTIVITIES

#### (57) Abstract

(30) Priority Data:

Hybrid proteins having crosslinking and tissue-binding activities, DNA molecules encoding such proteins and methods for producing the hybrid proteins from recombinant host cells are disclosed. The hybrid proteins disclosed herein are useful in tissue sealant and wound healing formulations.



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### Description

Hybrid Proteins Having Cross-Linking and Tissue-Binding Activities

### 5 Technical Field

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The present invention relates generally toward methods for producing recombinant hybrid proteins, and more specifically, to methods for producing hybrid proteins from host cells through the use of recombinant DNA techniques.

### Background of the Invention

The utilization of tissue sealants to replace or augment the use of mechanical wound closure devices has expanded in recent years in many surgical and trauma applications. Tissue sealants include adhesives (e.g. fibrin-based adhesives) and synthetic preparations (e.g. cyanoacrylates). Ιt is widely acknowledged that the use of synthetic preparations of tissue sealants is limited due to their toxicity and limited applications. Biological tissue adhesives have demonstrated utility in cases where the use of mechanical devices to close wounds is insufficient, such joining blood vessels, closing holes in the dura, and in surgery on small or delicate tissues such as in the eye or ear.

Fibrin-based biological tissue adhesives generally contain fibrinogen, factor XIII and thrombin as principal ingredients, although in practice biological tissue adhesives are derived from whole blood and contain additional blood proteins. The fibrinogen and factor XIII components of these adhesives are prepared from pooled human plasma by cryoprecipitation (e.g. U.S. Patents No. 4,377,572; 4,362,567; 4,909,251), by ethanol precipitation (e.g. U.S. Patent No. 4,442,655) or from single donor plasma (e.g. U.S. Patent No. 4,627,879; Spotnitz et al., 166-168, 1989). The resultant Am. Surq. <u>55</u>:

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fibrinogen/factor XIII preparation is mixed with bovine thrombin immediately before use to convert the fibrinogen to fibrin and activate the factor XIII, thus initiating coagulation of the adhesive.

Fibrin-based tissue adhesives, in their current significant drawbacks that form, have include standardization, lack of quality control from batch to batch and the possibility of transmission of immunodeficiency virus (HIV), hepatitis virus and other etiologic agents. While recombinant production thrombin and factor XIII have been reported, and while proteins might be used in biological tissue adhesives, the biological tissue adhesives still rely on large amounts of fibrinogen that is obtained from pooled human blood. At present, current fibrin(ogen)-based tissue adhesives are not approved for use in the United

There is therefore a need in the art for tissue adhesive components, particularly components that facilitate cross-linking to improve clot strength, that are prepared at high levels with reproducible activity levels and which do not carry the possibility of transmission of viral or other etiologic agents. The present invention addresses these needs by providing recombinant hybrid proteins that provide cross-linking and tissue-adhesive properties and that may be prepared at high levels.

### Disclosure of the Invention

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Briefly stated, the present invention provides hybrid proteins having cross-linking and tissue-binding activities, DNA molecules encoding such hybrid proteins and methods for producing hybrid proteins by recombinant means. In one aspect, In one aspect of the invention, the hybrid proteins comprise a tissue-binding domain from a first protein covalently linked to a cross-linking domain from a second protein. Within a related aspect of the

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invention, the tissue-binding domain of the first protein is a heparin binding domain of thrombospondin, a heparin binding domain of fibronectin, a collagen binding domain of fibronectin or a cell binding domain of fibronectin. Within a preferred embodiment, the tissue-binding domain of the first protein comprises the amino acid sequence of Sequence ID No. 6 from Alanine, amino acid 2 to Glutamic acid, amino acid number 926. Within another related aspect of the invention, the cross-linking domain of the second protein comprises the carboxy-terminal 103 amino acids of loricrin, the ten amino acid repeat beginning with glutamine amino acid number 496 of involucrin or the 400 amino-terminal amino acids of the fibrinogen  $\alpha$  chain. Within a preferred embodiment of the invention, tissue-binding domain of the second protein comprises the amino acid sequence of Sequence ID No. 6 from Glycine, amino acid number 928 to Proline, amino acid number 1336. Within a particularly preferred embodiment, the hybrid protein comprises the amino acid sequence of Sequence ID No. 6 from alanine, amino acid number 2 to proline, amino acid number 1336.

The present invention provides DNA molecules encoding hybrid proteins of the present invention comprising a first DNA segment encoding a tissue-binding domain from a first protein joined to a second DNA segment encoding a cross-linking domain from a second protein. one embodiment, the first DNA segment comprises the nucleotide sequence of Sequence ID No. 5 from nucleotide 3 to nucleotide 2780. In another embodiment, the second DNA segment comprises the nucleotide sequence of Sequence ID No. 5 from nucleotide 2784 to nucleotide 4013. preferred embodiment, the DNA molecule comprises nucleotide sequence of Sequence ID Number from nucleotide 3 to nucleotide 4013.

In related embodiments of the invention, DNA constructs are provided which comprise a DNA molecule encoding a hybrid protein, whereins said DNA molecule

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comprises a first DNA segment encoding a tissue-binding domain from a first protein joined to a second DNA segment encoding a cross-linking domain from a second protein and wherein said DNA molecule is operably linked to other DNA segments required for the expression of the DNA molecule. Other embodiments of the invention concern host cells containing the DNA constructs of the present invention and methods of producing hybrid proteins.

### 10 Brief Description of the Drawings

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Figure 1 discloses a representative hybrid protein containing (1) an N-terminal end-to-end interchain cross-linking domain, (2) a domain that promotes inter-chain cross-linking; (3) a domain that confers tissue binding activity; and (4) a carboxy-terminal domain that promotes end-to-end inter-chain cross-linking.

Figures 2-5 disclose absorbance time courses of representative cross-linking assays carried out in the presence of varying levels of factor XIII (activated to factor XIIIa via thrombin during the assay) or factor XIIIa.

### Detailed Description of the Invention

present invention provides novel hybrid proteins having cross-linking and tissue adhesive The hybrid proteins comprise a cross-linking activities. domain from a first protein covalently linked to a tissuebinding domain from a second protein. The hybrid proteins of the present invention are capable of cross-linking to themselves and to other proteins such as fibrin and fibrinogen and are capable of adhering to cell surfaces and/or extracellular matrix components. While not wishing to be bound by a graphical representation, Figure 1 shows a representative hybrid protein containing an N-terminal end-to-end inter-chain cross-linking domain; a domain that promotes inter-chain cross-linking; a domain that confers tissue binding activity; and a carboxy-terminal domain

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that promotes end-to-end inter-chain cross-linking. As used herein, cross-linking refers to the formation of covalent bonds between polypeptides.

The hybrid proteins of the present invention are useful as components of tissue sealant formulations to provide matrix material and to improve clot strength over a wound site, and as components in formulations that promote wound healing. The proteins of the present invention may contain native (i.e. wild-type) protein domains as well as domains that are allelic variants and genetically engineered or synthetic variants of the respective naturally occurring domains. Such variants are characterized by the presence of conservative amino acid substitutions and/or other minor additions, substitutions or deletions of amino acids.

As used within the context of the invention, tissue-binding domains include protein domains containing amino acid sequences that facilitate adherence to cell surfaces and/or to extracellular matrix components fibronectin, hyaluronic acid as collagen, Fibronectin, for example, contains glycosaminoglycans. the sequence Gly-Arg-Gly-Asp-Ser (from amino acid 1614 through amino acid 1618 of Sequence I.D. No. 3) that has been shown to be central to cell recognition by the fibronectin receptor (for review see Yamada, Opinion in Cell Biology 1: 956-963, 1989). The heparin binding domains of fibronectin (Sekiguchi et al., Proc. Natl. Acad. Sci. USA <u>77</u>: 2661-2665, 1980), thrombospondin (Zardi et al., EMBO J. 6: 2337-3342, 1987 and Gutman and Kornblihtt, Proc. Natl. Acad. Sci. USA 84: 7179-7182, 1987) contain sequences that recognize heparin sulfate-containing glycosaminoglycans which are extracellular matrix components. The collagen binding domain of fibronectin (Sekiguchi et al. ibid., contains amino acid sequences that bind to the extracellular matrix component collagen.

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Particularly preferred tissue-binding are the heparin binding domain of fibronectin, comprising the sequence of amino acids of Sequence I.D. No. 2 from alanine, amino acid number 1812 to valine, amino acid number 2171; the collagen binding domain of fibronectin, comprising the sequence of amino acids of Sequence I.D. No. 2 from glycine, amino acid number 282 to serine, amino acid number 608; and the amino terminal 229 amino acids of In this regard, a particularly preferred thrombospondin. tissue-binding domain is the cell-binding domain fibronectin, comprising the sequence of amino acids of Sequence I.D. No. 3 from alanine, amino acid number 1357 to glutamic acid, amino acid number 1903. It will be evident to one skilled in the art that smaller portions of the cell-binding domain of fibronectin may be used within the hybrid proteins of the present invention, particularly the sequence of amino acids of Sequence I.D. No. 3 from isoleucine, number 1532 through threonine, amino acid number 1631. As noted above, it is generally accepted that the sequence Gly-Arg-Gly-Asp-Ser (Amino acids 1614 to 1618 of Sequence I.D. No. 3) is central to cell recognition by fibronectin.

Cross-linking domains suitable for use in the hybrid proteins of the present invention are domains which contain amino acid sequences required for the formation of specific covalent bonds between peptide chains. In a preferred embodiment the inter-chain crosslinks are covalent bonds formed by the action of transglutaminase such as factor XIII, transglutaminase, prostate transglutaminase, keratinocyte transglutaminase, epidermal transglutaminase or placental Transglutaminases transglutaminase. catalyze the formation of  $\epsilon$ -( $\gamma$ -glutamyl)lysine bonds between specific glutamine and lysine residues. However, other inter-chain cross-links, such as those formed by disulfide bonds, are also suitable cross-links. Suitable cross-linking domains include domains from the fibrinogen chain, the

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glutamine/lysine rich domains of loricrin that involved in isodipeptide cross-link formation (Hohl et al., J. Biol. Chem. 266: 6626-6636, 1991), and at least one of the 10 amino acid-long repeats of involucrin (Cell 46: 583-589, 1986 and Etoh et al., <u>Biochem. Biophys. Res.</u> Comm. 136: 51-56, 1986). Preferred cross-linking domains are the carboxy-terminal 103 amino acids of loricrin (Hohl et al., ibid.) and the ten-amino acid repeat beginning with glutamine, amino acid number 496 of involucrin (Simon (J. Biol. Chem. 263: 18093-18098, 1988). particularly preferred cross-linking domain comprises the 400 amino-terminal amino acids of the fibrinogen  $\alpha$  chain (Doolittle et al., <u>Nature</u> 280: 464-468, 1979; Rixon et 3250-3256, Biochemistry 22: 1983). particularly, the amino acid sequence of Sequence ID No. 6 from Glycine, amino acid number 928 to Proline, amino acid number 1336 is preferred.

Although the hybrid proteins of the present invention may consist essentially of covalently linked cross-linking and tissue binding domains, they may further contain domains that facilitate end-to-end covalent crosslinking. The  $\gamma$  chain of fibrinogen contains a domain that facilitates end-to-end cross-linking to another  $\gamma$  chain via  $\epsilon$ -( $\gamma$ -glutamyl)lysine bonds. This domain includes at 19 carboxy-terminal amino acids and least the preferably includes the amino-terminal 275 amino acids of the fibrinogen  $\gamma$  chain. The  $\alpha$  chain of fibrinogen contains an amino-terminal domain that is involved in interchain disulfide bond formation between  $\alpha$  chains. includes the amino-terminal portion of the  $\alpha$  chain of fibrinogen from glycine, amino acid 36 to glycine, amino acid 67 of Sequence ID Number 4.

As will be evident to one skilled in the art, the hybrid proteins of the present invention may contain domains of human and other animal proteins. Proteins containing domains suitable for use in the present invention from human and other animals and the DNA

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molecules encoding such proteins have been reported. Involucrin, loricrin, fibrinogen and fibronectin, example, have been studied in a variety of animals. DNA sequences encoding primate, canine and porcine involucrin have been reported (Djian and Green, Mol. Biol. Evol. 9: 417-432, 1992; Djian and Green, Proc. Natl. Acad. Sci. USA 88: 5321-5325, 1991 and Tseng and Green, Mol. Biol. Evol. Mehrel et al. (Cell 61: 1103-1112, 7: 293-302, 1990). reported a DNA sequence encoding 1990) have DNA sequences encoding rat and frog fibrinogen gamma chain have been reported (Haidaris and Courtney, Blood 79: 1218-1224, 1992 and Bhattacharya et al., Mol. Cell. Endocrinol. 72: 213-220, 1990; respectively). sequences encoding chicken and lamprey fibrinogen  $\alpha$  chains have been reported by Weissbach and Greininger (Proc. Natl. Acad. Sci. USA 87: 5198-5202, 1990) and Pan and Doolittle (Proc. Natl. Acad. Sci. USA 89: 1992), respectively. DNA sequences encoding bovine and rat fibronectin have been reported by Petersen et al. (Proc. Natl. Acad. Sci. USA 80: 137-141, 1983) and Schwarzbauer et al., (Cell 35: 421-431, 1983). general, it is preferred to prepare proteins that contain component domains from a single species to minimize the immunogenicity. Thus, possibility of the present invention provides hybrid proteins that can be used in human and veterinary medicine.

the present According to invention hybrid cross-linking proteins having and tissue adhesive activities are produced recombinantly from host cells transformed with a DNA construct comprising a DNA segment encoding a cross-linking domain from a first protein joined to a DNA segment encoding a tissue-binding domain from a second protein. As used within the context of the present invention, two or more DNA coding sequences are said to be joined when, as a result of in-frame fusions between the DNA coding sequences or as a result of the removal of intervening sequences by normal cellular

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processing, the DNA coding sequences can be translated into a polypeptide fusion. Unless otherwise noted, the DNA segments may be joined in any order to result in a DNA coding sequence that can be translated into a polypeptide chain. Thus, the DNA segment encoding the tissue-binding domain may be joined to the 5' or the 3' end of the DNA segment encoding the cross-linking domain. However, as will be evident to one skilled in the art, the production of hybrid proteins that additionally include domains that facilitate end-to-end cross-linking will require that the DNA segments encoding such domains be positioned at the 5' and 3' termini of the molecules.

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invention Thus the present also isolated DNA molecules encoding hybrid proteins comprising a cross-linking domain from a first protein covalently linked to a tissue-binding domain from a second protein. In general, cDNA sequences are preferred for carrying out the present invention due to their lack of intervening sequences which can lead to aberrant RNA processing and reduced expression levels. DNA molecules encoding human fibronectin (Dufour et al., Exper. Cell Res. 193: 331-338, 1991) and a human fibrinogen  $\alpha$  chain (Rixon et al., Biochemistry 22: 3250-3256, 1983) may be obtained from libraries prepared from liver cells according to standard It will be understood however, laboratory procedures. that suitable DNA sequences can also be obtained from genomic clones or can be synthesized de novo according to conventional procedures. If partial clones are obtained, it is necessary to join them in proper reading frame to produce a full length clone, using such techniques as endonuclease cleavage, ligation, and loop-out mutagenesis.

DNA sequences encoding hybrid proteins of the present invention may be prepared from cloned DNAs using conventional procedures of endonuclease cleavage, exonuclease digestion, ligation and <u>in vitro</u> mutagenesis. Alternatively, DNA sequences encoding the cross-linking

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and tissue-binding domains, such as those mentioned above, may be synthesized using standard laboratory techniques.

An exemplary DNA molecule encoding a hybrid protein having cross-linking and tissue-binding activities may be prepared by joining a DNA segment encoding at least the cell-binding domain of fibronectin and a DNA segment encoding at least an inter-chain cross-linking domain of fibrinogen at a convenient restriction site synthetic adapters to facilitate in-frame joining of the Alternatively, such DNA segments encoding DNA segments. hybrid proteins of the present invention may be prepared by joining the two domains at a convenient restriction site followed by loop-out mutagenesis to precisely remove unnecessary sequences and directly join the DNA segment encoding the cell-binding domain of fibronectin with the DNA segment encoding the cross-linking domain fibrinogen.

DNA segments encoding the hybrid proteins of the instant invention are inserted into DNA constructs. used within the context of the present invention, a DNA construct is understood to refer to a DNA molecule, or a clone of such a molecule, either single- or doublestranded, which has been modified through intervention to contain segments of DNA combined and juxtaposed in a manner that would not otherwise exist in nature. DNA constructs of the present invention comprise a first DNA segment encoding a hybrid protein operably linked to additional DNA segments required for expression of the first DNA segment. Within the context of the present invention, additional DNA segments will generally include promoters and transcription terminators, and may further include enhancers and other elements.

DNA constructs may also contain DNA segments necessary to direct the secretion of a polypeptide or protein of interest. Such DNA segments may include at least one secretory signal sequence. Secretory signal sequences, also called leader sequences, prepro sequences

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and/or pre sequences, are amino acid sequences that act to direct the secretion of mature polypeptides or proteins from a cell. Such sequences are characterized by a core of hydrophobic amino acids and are typically (but not exclusively) found at the amino termini synthesized proteins. DNA segments encoding secretory signal sequences are placed in-frame and in the correct spatial relationship to the DNA segment encoding the protein of interest in order to direct the secretion of the protein. Very often the secretory peptide is cleaved from the mature protein during secretion. Such secretory peptides contain processing sites that allow cleavage of the secretory peptides from the mature proteins as they secretory pathway. through the Α preferred processing site is a dibasic cleavage site, such as that recognized by the Saccharomyces cerevisiae KEX2 gene. particularly preferred processing site is a Lys-Arg processing site. Processing sites may be encoded within the secretory peptide or may be added to the peptide by, for example, in vitro mutagenesis.

Preferred secretory signals include the α factor signal sequence (pre-pro sequence: Kurjan and Herskowitz, Cell 30: 933-943, 1982; Kurjan et al., U.S. Patent No. 4,546,082; Brake, U.S. Patent No. 4,870,008), the PHO5 signal sequence (Beck et al., WO 86/00637), the BAR1 secretory signal sequence (MacKay et al., U.S. Patent No. 4,613,572; MacKay, WO 87/002670), the SUC2 signal sequence (Carlsen et al., Molecular and Cellular Biology 3: 439-447, 1983). Alternately, a secretory signal sequence may be synthesized according to the rules established, for example, by von Heinje (European Journal of Biochemistry 133: 17-21, 1983; Journal of Molecular Biology 184: 99-105, 1985; Nucleic Acids Research 14: 4683-4690, 1986).

Secretory signal sequences may be used singly or may be combined. For example, a DNA segment encoding a first secretory signal sequence may be used in combination with a DNA segment encoding the third domain of barrier

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(described in U.S. Patent No. 5,037,243, which is incorporated by reference herein in its entirety). The DNA segment encoding the third domain of barrier may be positioned in proper reading frame 3' of the DNA segment of interest or 5' to the DNA segment and in proper reading frame with both the DNA segment encoding the secretory signal sequence and the DNA segment of interest.

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The choice of suitable promoters, terminators and secretory signals is well within the level of ordinary skill in the art. Methods for expressing cloned genes in Saccharomyces cerevisiae are generally known in the art (see, "Gene Expression Technology," Methods in Enzymology, Vol. 185, Goeddel (ed.), Academic Press, San Diego, CA, 1990 and "Guide to Yeast Genetics and Molecular Biology," Methods in Enzymology, Guthrie and Fink (eds.), Academic Press, San Diego, CA, 1991; which are incorporated herein Transformation systems for other yeasts, by reference). including Hansenula polymorpha, Schizosaccharomyces pombe, Kluyveromyces lactis, Kluyveromyces fragilis, <u>Ustilaqo</u> maydis, Pichia pastoris, Pichia quillermondil and Candida maltosa are known in the art. See, for example, Gleeson et al., J. Gen. Microbiol. 132:3459-3465, 1986 and Cregg, U.S. Patent No. 4,882,279.

Proteins of the present invention can also be expressed in filamentous fungi, for example, strains of the fungi Aspergillus (McKnight et al., U.S. Patent No. 4,935,349, which is incorporated herein by reference). Methods for transforming <u>Acremonium chrysogenum</u> are disclosed by Sumino et al., U.S. Patent No. 5,162,228, which is incorporated herein by reference.

Other higher eukaryotic cells may also be used as hosts, including insect cells, plant cells and avian cells. Transformation of insect cells and production of foreign proteins therein is disclosed by Guarino et al., U.S. Patent No. 5,162,222 and Bang et al., U.S. Patent No. 4,775,624, which are incorporated herein by reference. The use of Agrobacterium rhizogenes as a vector for

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expressing genes in plant cells has been reviewed by Sinkar et al., <u>J. Biosci. (Bangalore)</u> 11:47-58, 1987.

Expression of cloned genes in cultured mammalian cells and in <u>E</u>. <u>coli</u>, for example, is discussed in detail in Sambrook et al. (<u>Molecular Cloning: A Laboratory Manual</u>, Second Edition, Cold Spring Harbor, NY, 1989; which is incorporated herein by reference). In addition to <u>E</u>. <u>coli</u>, <u>Bacillus</u> and other genera are useful prokaryotic hosts for expressing foreign proteins. As would be evident to one skilled in the art, one could express the proteins of the instant invention in other host cells such as avian, insect and plant cells using regulatory sequences, vectors and methods well established in the literature.

In yeast, suitable vectors for use in the 15 present invention include YRp7 (Struhl et al., Proc. Natl. Acad. Sci. USA 76: 1035-1039, 1978), YEp13 (Broach et al., Gene 8: 121-133, 1979), POT vectors (Kawasaki et al, U.S. Patent No. 4,931,373, which is incorporated by reference 20 herein), pJDB249 and pJDB219 (Beggs, Nature 275:104-108, 1978) and derivatives thereof. Preferred promoters for use in yeast include promoters from yeast glycolytic genes (Hitzeman et al., <u>J. Biol. Chem.</u> <u>255</u>: 12073-12080, 1980; Alber and Kawasaki, <u>J. Mol. Appl. Genet.</u> 1: 419-434, 1982; 4,599,311) U.S. Patent No. or 25 Kawasaki, dehydrogenase genes (Young et al., in Genetic Engineering of Microorganisms for Chemicals, Hollaender (eds.), p. 355, Plenum, New York, 1982; Ammerer, Meth. 192-201, 1983). In this Enzymol. <u> 101</u>: regard, particularly preferred promoters are the TPI1 promoter 30 (Kawasaki, U.S. Patent No. 4,599,311, 1986) and the ADH2-<u>4<sup>C</sup></u> promoter (Russell et al., <u>Nature</u> <u>304</u>: 652-654, 1983; Irani and Kilgore, U.S. Patent Application Serial No. 07/631,763, CA 1,304,020 and EP 284 044, which are incorporated herein by reference). The expression units 35 may also include a transcriptional terminator.

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preferred transcriptional terminator is the <u>TPI1</u> terminator (Alber and Kawasaki, ibid.).

Host cells containing DNA constructs of the present invention are then cultured to produce the hybrid The cells are cultured according to standard proteins. methods in a culture medium containing nutrients required for growth of the particular host cells. A variety of suitable media are known in the art and generally include a carbon source, a nitrogen source, essential amino acids, vitamins, minerals and growth factors. The growth medium generally select for cells containing the construct by, for example, drug selection or deficiency in essential nutrient which is complemented selectable marker on the DNA construct or co-transfected with the DNA construct.

Selection of a medium appropriate for particular host cell used is within the level of ordinary Yeast cells, for example, skill in the art. preferably cultured in a chemically defined medium, comprising a non-amino acid nitrogen source, inorganic salts, vitamins and essential amino acid supplements. pH of the medium is preferably maintained at a pH greater than 2 and less than 8, preferably at pH 6.5. Methods for maintaining a stable pH include buffering and constant pH control, preferably through the addition of hydroxide or ammonium hydroxide. Preferred buffering agents include succinic acid and Bis-Tris (Sigma Chemical Co., St. Louis, MO). Yeast cells having a defect in a gene required for asparagine-linked glycosylation are preferably grown in a medium containing an osmotic stabilizer. A preferred osmotic stabilizer is sorbitol supplemented into the medium at a concentration between 0.1 M and 1.5 M, preferably at 0.5 M or 1.0 M. Cultured mammalian cells are generally cultured in commercially available serum-containing or serum-free media.

The recombinant hybrid proteins expressed using the methods described herein are isolated and purified by

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conventional procedures, including separating the cells the medium by centrifugation or filtration. proteinaceous precipitating the components of supernatant or filtrate by means of a salt, e.g. ammonium sulfate, purification by a variety of chromatographic procedures, e.g. ion exchange chromatography or affinity chromatography, or the like. Methods protein purification are known in the art (see generally, Scopes, <u>Protein Purification</u>, Springer-Verlag, NY which is incorporated herein by reference) and may be applied to the purification of the recombinant proteins of the present invention.

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The hybrid proteins of the present invention may be used as components of tissue adhesives. is preferred that the tissue adhesives be formulated provide a concentration of the hybrid proteins of the present invention of between about 5 mg/ml to 100 mg/ml, with concentrations in the range of 35 to 50 mg/ml being particularly preferred. disclosed above, tissue As adhesives generally contain factor XIII and thrombin. Additional components may also be included in the tissue adhesive formulations. These additional components include growth factors such as PDGF, bFGF, TGFa, or EGF and protease inhibitors, such as aprotinin, transexamic acid, alpha-2 plasmin inhibitor, alpha-1-antitrypsin or the Pittsburgh mutant of alpha-1-antitrypsin (Arg-358 alpha-1-antitrypsin). The tissue adhesives may contain salts, buffering agents, reducing agents, bulking agents, and solubility enhancers. Albumin, NaCl, CaCl2, citrate and phosphate buffers, for example, may included. Preferably, the tissue adhesives of the present invention are prepared as lyophilized powders, concentrates of ready-to-use liquids. Lyophilized powders are preferred for ease of handling and storage.

The following examples are offered by way of illustration and not by way of limitation.

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#### **EXAMPLES**

Example 1 -Subcloning and Modification of ADH2 Promoters

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An <u>ADH2-4</u><sup>C</sup> promoter was constructed as described in co-pending U.S. Patent Application 07/631,763, CA 1,304,020 and EP 284 044, which are incorporated herein by reference. A DNA construct comprising the complete <u>ADH2-4</u><sup>C</sup> promoter mutagenized at the 3' end to place an Eco RI site in place of the translation start codon, designated p410-4<sup>C</sup> (deposited with the American Type Culture Collection (12301 Parklawn Dr., Rockville, MD 20852) under accession number 68861) was used as the source of the ADH2-4<sup>C</sup> promoter.

A PAP-I cDNA (U.S. Patent No. 4,937,324) was joined with the <u>ADH2-4<sup>C</sup></u> promoter. Plasmid pAP1.7, comprising the 1.7 kb cDNA in pUC18, was cut with Nco I and Bam HI, and the linearized plasmid was isolated through two rounds of gel purification. The <u>ADH2-4<sup>C</sup></u> promoter from p410-4<sup>C</sup> was joined to the 5' end of the PAP-I cDNA via an Eco RI-Nco I adapter. The 1.2 kb Bam HI-Eco RI promoter fragment from p410-4<sup>C</sup>, Eco RI-Nco I adapter and the Nco I-Bam HI linearized pAP1.7 plasmid were ligated. The resultant plasmid was designed pPR1. The presence of the correct promoter fusion was confirmed by DNA sequencing.

A yeast expression vector comprising the  $\underline{ADH2-4^C}$  promoter, the PAP-I cDNA and the  $\underline{TPI1}$  terminator was constructed. Plasmid pZUC13 (comprising the  $\underline{S.}$  cerevisiae chromosomal  $\underline{LEU2}$  gene and the origin of replication from  $\underline{S.}$  cerevisiae 2 micron plasmid inserted into pUC13 and constructed in a manner analogous to pZUC12, described in published EP 195,691, using the plasmid pMT212, which is described in published EP 163 529) was cut with Bam HI. Plasmid pPR1 was digested completely digested with Bam HI and partially digested with Sac I to isolate the 2.1 kb  $\underline{ADH2-4^C}$  promoter-PAP-I cDNA fragment. Plasmid pTT1 (described in detail below) was digested with Sac I and

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Bam HI to isolate the 0.69 bp <u>TPI1</u> terminator fragment. The Bam HI-Sac I fragment from pPR1 and the Sac I-Bam HI fragment from pTT1 were ligated with the Bam HI-linearized pZUC13. A plasmid containing the expression unit was designated pZ3.

### Example 2 - Subcloning of the TPI1 terminator

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The yeast <u>TPI1</u> terminator fragment was obtained from plasmid p270 described by Murray and Kelly (U.S. Patent 4,766,073, which is incorporated by reference herein in its entirety). Plasmid p270 contains the <u>TPI1</u> terminator inserted as and Xba I-Bam HI fragment into YEp13. Alternatively, the <u>TPI1</u> terminator may be obtained from plasmid pM220 (deposited with American Type Culture Collection as an <u>E</u>. <u>coli</u> RR1 transformant under accession number 39853) by digesting the plasmid with Xba I, and Bam HI and purifying the <u>TPI1</u> terminator fragment (~700 bp).

The <u>TPI1</u> terminator was removed from plasmid p270 as a Xba I-Bam HI fragment. This fragment was cloned into pUC19 along with another fragment containing the <u>TPI1</u> promoter fused to the CAT (chloramphenicol acetyl transferase) gene to obtain a <u>TPI1</u> terminator fragment with an Eco RV end. The resultant plasmid was designated pCAT. The <u>TPI1</u> terminator was then cut from pCAT as an Eco RV-Bam HI fragment and cloned into pIC19H (Marsh et al., <u>Gene 32</u>:481-486, 1984) which had been cut with the same enzymes, to obtain pTT1 (disclosed in U.S. Patent No. 4,937,324, which is incorporated herein by reference).

# 30 Example 3 - <u>Construction of Yeast Vectors pDPOT and pRPOT</u>

Plasmid pDPOT was derived from plasmid pCPOT (ATCC No. 39685) by replacing the 750 bp Sph I-Bam HI fragment of pCPOT containing 2 micron and pBR322 sequences with a 186 bp Sph I-Bam HI fragment derived from the pBR322 tetracycline resistance gene.

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Plasmid pRPOT was derived from plasmid pDPOT by replacing the Sph I-Bam HI fragment with a polylinker. Plasmid pDPOT was digested with Sph I and Bam HI to isolate the 10.8 kb fragment. Oligonucleotides ZC1551 and ZC1552 (Sequence ID Nos. 7 and 8) were designed to form an adapter with a Bam HI adhesive end and an Sph I adhesive end flanking Sma I, Sst I and Xho I restriction sites. Oligonucleotides ZC1551 and ZC1552 (Sequence ID Nos. 7 and 8) were kinased and annealed to form the Bam HI-Sph I adapter. The 10.8 kb pDPOT fragment was circularized by ligation with the ZC1551/ZC1552 adapter (Sequence ID Nos. 7 and 8). The resultant plasmid was termed pRPOT.

# Example 4 - <u>Construction of a Fibrinogen: Fibronectin</u> Hybrid cDNA Expression Vector

### A. Construction of pFN14A

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Α DNA construct containing а DNA segment encoding the fibronectin cell-binding domain operably linked to the ADH2-4° promoter in plasmid pUC19 was The fibronectin coding sequence was obtained constructed. from plasmid pFH103 (Dufour et al., Exper. Cell Res. 193: 331-338, 1991). Plasmid pFH103 was digested with Nco I and I to isolate the 4 kb fragment containing the fibronectin coding sequence. Oligonucleotides ZC2052 and ZC2053 (Sequence ID Nos. 9 and 10) were designed to provide, upon annealing, an adapter containing a 5' Eco RI end, an internal Nco I site, a DNA segment encoding a methionine and amino acids 979 through 981 of Sequence ID Number 2 and a 3' Nco I adhesive end that destroys the Nco I site. Oligonucleotides ZC2052 and ZC2053 (Sequence ID Nos. 9 and 10) were annealed and ligated with the 4 kb Nco I-Xba I fibronectin fragment into Eco RI-Xba I linearized pUC19. The resultant plasmid was designated pFN4.

Plasmid pFN4 was digested with Hind III and Apa I to isolate the 3.3 kb fibronectin fragment. Oligonucleotides ZC2493 and ZC2491 (Sequence ID Nos. 12

and 11) were designed to provide, when annealed, an Apa I-Xba I adapter encoding the amino acids Pro and Phe followed by a stop codon. The oligonucleotides were annealed and combined with the 3.3 kb Hind III-Apa I fragment and Hind III-Xba I linearized pUC19 to form plasmid pFN7. Plasmid pFN7 comprises a DNA segment encoding amino acids 1273-2186 of Sequence ID Number 2 followed by an in-frame stop codon.

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The <u>ADH2-4<sup>C</sup></u> promoter was joined to the 5' end of the fibronectin cDNA in plasmid pFN5. Plasmid pFN4 was digested with Nco I and Hind III to isolate the 0.89 kb fibronectin coding sequence. Plasmid pZ3 (described in detail above) was digested with Bam HI and Nco I to isolate the 1.25 kb <u>ADH2-4<sup>C</sup></u> promoter fragment. The 1.25 kb Bam HI-Nco I promoter fragment and the Nco I-Hind III fibronectin coding sequence fragment were ligated to Bam HI-Hind III linearized pUC19 to form plasmid pFN5.

Plasmid pFN5 was digested with Bam HI and Hind III to isolate the 2.1 kb promoter-fibronectin fragment. Plasmid pFN7 was digested with Hind III and Xba I to isolate the 2.8 kb fibronectin fragment that was modified to encode a stop codon following the Pro-Phe sequence. The TPI1 terminator sequence was obtained from pTT1 as a The 2.1 kb Bam HI-Hind III 0.7 kb Xba I-Sal I fragment. promoter-fibronectin fragment, the 2.8 kb Hind III-Xba I fibronectin fragment and the 0.7 kb TPI1 terminator fragment were joined in a four-part ligation with Bam HIlinearized pRPOT. A plasmid containing the fibronectin expression unit in the pRPOT vector was designated pR1.

The original clone pFH103 contained a frameshift mutation in the EIIIB region of the fibronectin cDNA. The mutation was corrected by the replacement of the region with an analogous region from the plasmid pFHA3 (obtained from Jean Paul Thiery, Laboratoire de Physiopathologie du Developpement, CNRS URA 1337, Ecole Normale Superiure, 46 rue d'Ulm, 75230 Paris Cedex 05,

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Plasmid pFHA3 was derived from pFH103 France). by excising the 3211 pd Xba I-Asp 718I fragment of fibronectin, blunting of the resultant adhesive ends and religating. Plasmid pFHA3 contains a DNA segment encoding the signal and propeptides, the first three and one half type I repeats, and the carboxy-terminal half of human fibronectin from the middle of the EIIIB segment.

Plasmid pR1 was digested with Bam HI and Kpn I to isolate the 2.2 kb promoter-fibronectin fragment. Plasmid pFHA3 was digested with Kpn I and Apa I to isolate the internal fibronectin fragment that corrects the frame-shift mutation present in the parent cDNA from pFH103. Plasmid pR1 was digested with Apa I and Bam HI to isolate the TPI1 terminator fragment. The 2.2 kb Bam HI-Kpn I promoter-fibronectin fragment, the 2.75 kb Kpn I-Apa I internal fibronectin fragment and the 0.69 kb Apa I-Bam HI TPI1 terminator fragment were joined in a four-part ligation with Bam HI-linearized pDPOT. The resulting construction was designated pD32.

A DNA segment encoding the <u>ADH2-4</u><sup>C</sup> promoter and initiation methionine from plasmid pD32 was subcloned into pIC19H (Marsh et al., <u>Gene 32</u>:481-486, 1984) as a 1.25 kb Bam HI-Nco I fragment. Plasmid pD32 was also digested with Nco I and Bgl II to isolate the 3 kb fibronectin cDNA fragment encoding amino acids 979-1972 of Sequence ID Number 2. The 1.25 kb Bam HI-Nco I fragment and the Nco I-Bgl II fragment were ligated with Bam HI-linearized pIC19H. A plasmid containing a Bam HI site proximal to the <u>ADH2-4</u><sup>C</sup> promoter was designated pFN14A.

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### B. - Construction of Plasmid pD38

An expression vector comprising a DNA segment encoding a fibronectin-fibrinogen hybrid protein operably linked to the  $\underline{ADH2-4}^{C}$  promoter and the TPI1 terminator was constructed. To assemble the DNA sequence encoding the hybrid protein, a DNA segment encoding approximately the

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carboxy-terminal 409 amino acids of the  $\alpha$  chain of fibrinogen was first subcloned.

A fibrinogen  $\alpha$  chain cDNA was obtained from Dominic W. Chung (Department of Biochemistry, University of Washington, Seattle, WA) in plasmid pHI $\alpha$ 3 (Rixon et al., Biochemistry 22: 3250-3256, 1983). Sequence analysis of the cDNA insert in plasmid pHI $\alpha$ -3 revealed a deletion of codons 1348-1350 of the published sequence resulting in the deletion of Serine, amino acid 417.

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The DNA segment encoding the carboxy-terminus of the fibrinogen  $\alpha$  chain was subcloned into plasmid pUC19. Plasmid pHI $\alpha$ -3 was digested with Asp 718 and Ssp I to isolate the approximately 2 kb fragment encoding the carboxy-terminus of the fibrinogen  $\alpha$  chain from amino acid 244 to amino acid 643 and some 3' untranslated sequence of Sequence ID Number 4. Plasmid pTT1 was digested with Eco RV and Sal I to isolate the approximately 700 bp TPI1 terminator fragment. The 2 kb fibrinogen  $\alpha$  chain sequence and the TPI1 terminator sequence were ligated with pUC19 that had been linearized with Asp 718 and Sal I. ligation mixture was transformed into E. coli, and plasmid DNA was prepared and analyzed by restriction endonuclease and DNA sequence analysis. DNA sequence analysis of a candidate clone revealed that the Sal I site joining the TPI1 terminator sequence and the pUC19 polylinker site was Plasmid DNA from the candidate clone was not present. 718 and Bam HI to liberate the digested with Asp approximately 1.9 kb fibrinogen-TPI1 terminator fragment.

To join the fibronectin coding sequence with the fibrinogen  $\alpha$  chain sequence, synthetic oligonucleotides were synthesized to provide, when annealed, a Sal I-Asp 718 adapter encoding an internal Afl II restriction site, and a sequence encoding amino acids 1886 through 1903 of fibronectin (Sequence ID Number 2), a glycine residue and amino acids 235 through 243 of the fibrinogen  $\alpha$  chain (Sequence ID Number 4). Oligonucleotides ZC3521 and ZC3522 (Sequence ID Nos. 13 and 14) were annealed. The

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1.9 kb Asp 718-Bam HI fibrinogen-TPI1 terminator fragment and the Sal I-Asp 718 ZC3521/ZC3522 adapter (Sequence ID Nos. 13 and 14) were ligated with pUC19 that had been linearized with Sal I and Bam HI. The resultant plasmid was designated pFG4.

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The DNA segment encoding the fibronectinfibrinogen  $\alpha$  chain sequence in plasmid pFG4 was joined with the DNA segment encoding the amino-terminal fibronectin sequence (from amino acid 989 to amino acid Sequence ID Number 2) in plasmid pFN14A to construct plasmid pD37. Plasmid pFN14A was digested with Bgl II and Afl II to isolate the approximately 3.9 kb <u>ADH2-4</u><sup>C</sup> promoter-fibronectin fragment. Plasmid pFG4 was Afl digested with ΙI and Bam HΙ to isolate approximately 2 kb fibronectin-fibrinogen-TPI1 terminator The 3.9 kb Bgl II-Afl II fragment and the 2 kb fragment. Afl II-Bam HI fragment were ligated with Bam HI-linearized A plasmid with the expression unit inserted with the direction of transcription in the same direction as the POT1 gene in the pDPOT vector was designated pD37.

To place the expression unit present in pD37 in the opposite orientation, such that the direction of transcription of the expression unit was in the opposite direction to that of the POT1 gene, plasmid pD37 was digested with Nco I and Xba I to isolate the approximately 4 kb fibronectin-fibrinogen  $\alpha$  chain fragment. pFN14A was digested with Bam HI and Nco I to isolate the approximately 1.3 kb ADH2-4<sup>C</sup> promoter fragment. pTT1 was digested with Bam HI and Xba I to isolate the approximately 700 bp TPI1 terminator fragment. HI-Nco I ADH2-4<sup>C</sup> promoter fragment, the Nco I-Xba I fibronectin-fibrinogen  $\alpha$  chain fragment and the Xba I-Bam HI TPI1 terminator fragment were ligated with Bam HIlinearized pDPOT that had been treated with calf alkaline phosphatase to prevent recircularization. containing the expression unit in the opposite orientation relative to the POT1 gene was designated pD38.

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nucleotide sequence and deduced amino acid sequence of the DNA segment encoding the fibronectin-fibrinogen hybrid of plasmid pD38 is shown in Sequence ID Number 5. Plasmid pD38 was deposited on December 15, 1992 with the American Type Culture Collection (12301 Parklawn Drive, Rockville, MD) as an <u>E. coli</u> transformant.

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# Example 5 - <u>Expression of a Fibronectin-Fibrinogen</u> Hybrid Protein in Yeast

Plasmid pD38 was transformed into the Saccharomyces cerevisiae host strain ZM118 (MATa/MATa ura3/ura3 <u>Δtpi1::URA3/Δtpi1::URA3</u> <u>leu2-3,112/leu2-3,112</u> bar1/bar1 pep4::URA3/pep4::URA3 [cir<sup>0</sup>]) using essentially the method described by Hinnen et al. (Proc. Natl. Acad. 75: 1929-1933, 1978). Transformants Sci. USA selected for their ability to grow on medium containing glucose as the sole carbon source.

The ZM118[pD38] transformant was scaled up in a liter fermenter to facilitate purification of the hybrid protein. A single ZM118[pD38] colony was selected from a YEPD + Ade + Leu plate (Table 1) and inoculated into -LeuTrpThrD medium (Table 1). The culture was incubated for approximately 52 hours after which the cells The cells were washed in T.E. buffer were harvested. (Sambrook et al., ibid.), resuspended in T.E. buffer + 30% glycerol, and aliquotted into 1 ml seed vials. seed vials were stored at -80°C. One seed vial was used to inoculate 100 ml of YEPD + Ade + Leu (Table 1). culture was grown for approximately 28 hours to a final The 100 ml culture of ZM118[pD38] was  $A_{660}$  of 7.7. inoculated into a 10 liter fermenter with a final volume of 6.0 liters of medium containing 10 g/L (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 5 g/L KH<sub>2</sub>PO<sub>4</sub>, 5 g/L MgSO<sub>4</sub>·7H<sub>2</sub>O, 1 g/L NaCl, 0.5 g/L CaCl<sub>2</sub>·2H<sub>2</sub>O, 3.68 g/L A.A.I. (Table 1), 4.2 g/L citric acid, 60 g/L glucose, 10 ml/L Trace Metal Solution (Table 1), 0.4 ml/L PPG-2025 (Polypropylene glycol, MW 2025, Union Carbide Corp, Danbury, CT) that had been pH adjusted to pH 5.0

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with NaOH. In addition to the inoculation culture, 30 ml of Vitamin solution was added (Table 1). The culture was grown for 23 hours at  $30^{\circ}$ C with the addition of 2 M NaOH to maintain pH of approximately 5.

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# <u>Table</u> 1 <u>Media Recipes</u>

5 -LeuThrTrp Amino Acid Mixture	
	4 g adenine
	3 g L-arginine
	5 g L-aspartic acid
	2 g L-histidine free base
10	6 g L-isoleucine
	4 g L-lysine-mono hydrochloride
	2 g L-methionine
	6 g L-phenylalanine
	5 g L-serine
15	5 g L-tyrosine
	4 g uracil
	6 g L-valine
	Mix all the ingredients and grind with
20	a mortar and pestle until the mixture is finely
	ground.
	-LeuTrpThrD
	20 g glucose
25	6.7 g Yeast Nitrogen Base without amino
	acids (DIFCO Laboratories, Detroit,
	MI)
	0.6 g -LeuThrTrp Amino Acid Mixture
	18 g Agar
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	Mix all the ingredients in distilled
water. Add distilled water to a final vo	
	1 liter. Autoclave 15 minutes. Pour plates and
	allow to solidify.

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## Table 1 continued

	Table 1 Continued		
	YEPD + Ade + Leu Plates		
	20 g	glucose	
	20 g	Bacto Peptone (DIFCO Laboratories)	
5	10 g	Bacto Yeast Extract (DIFCO	
		Laboratories)	
	18 g	agar	
	4 ml	1% adenine	
	8 ml	1% L-leucine	
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		Mix all ingredients in distilled	
	water, an	nd bring to a final volume of 1 liter.	
	Autoclave	25 minutes and pour plates.	
15	YEPD + Ad	e + Leu Medium	
	20 g	glucose	
	20 g	Bacto Peptone (DIFCO Laboratories)	
	10 g	Bacto Yeast Extract (DIFCO	
		Laboratories)	
20	4 ml	1% adenine	
	8 ml	1% L-leucine	
		Mix all ingredients in distilled	
	water, an	nd bring to a final volume of 1 liter.	

Autoclave 25 minutes.

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A.A.I.

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## Table 1 continued

	4.0 g	adenine
	5.0 g	L-alanine
5	2.0 g	L-arginine
	5.0 g	L-asparagine
	5.0 g	L-aspartic acid
	5.0 g	L-cysteine
	5.0 g	L-glutamine
10	5.0 g	L-glutamic acid
	5.0 g	L-glycine
	8.0 g	L-histidine
	5.0 g	L-isoleucine
	3.0 g	L-lysine-mono hydrochloride
15	2.0 g	L-methionine
	5.0 g	L-phenylalanine
	5.0 g	L-proline
	5.0 g	L-serine
	5.0 g	L-threonine
20	2.0 g	L-tryptophan
	3.0 g	L-tyrosine
	3.0 g	uracil
	5.0 g	L-valine
25		Mix all the ingredients and grind with
	a mortar	and pestle until the mixture is finely
	ground.	Store at room temperature.

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### Table 1 continued

	Trace Metal Solution	
	0.68 g	ZnCl <sub>2</sub>
	5.4 g	FeCl <sub>3</sub> .6H <sub>2</sub> O
5	1.91 g	$MnCl_2 \cdot 4H_2O$
	0.22 g	CuSO <sub>4</sub> • 5H <sub>2</sub> O
	0.258 g	CoCl <sub>2</sub>
	0.062 g	$H_3BO_3$
	0.002 g	$(NH_4)_6Mo_2O_2$
10	0.002 g	KI
	10.0 ml 37	7% HCl

Dissolve solids in water and bring to a final volume of 1 liter.

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### <u>Vitamin Solution</u>

	25 mg	d-biotin
	400 mg	thiamine
	400 mg	pyridoxine
20	7.5 g	meso-inositol
	7.5 g	Ca pantothenate
	300 mg	niacinamide
	50 mg	folic acid
	100 mg	riboflavin
25	500 mg	choline

Dissolve solids in water and bring to a final volume of 1 liter.

A 60 liter fermenter with a final volume of 50 liters of medium containing 60 g/L yeast extract (Universal Foods, Milwaukee, WI), 2.5 g/L MgSO<sub>4</sub>·7H<sub>2</sub>O (Mallinkrodt Inc., St. Louis, MO), 1 g/L CaCl<sub>2</sub>·2H<sub>2</sub>O (Mallinkrodt, Inc.), 1 g/L KCl (Mallinkrodt, Inc.), 10 ml/L of Trace Metal Solution (Table 1), 0.5 ml/L PPG-2025 (Union Carbide) that had been adjusted to a pH of 5.0 with

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H<sub>3</sub>PO<sub>4</sub> was prepared, and the medium was sterilized. After sterilization, 5.0 liters of the 23 hour fermentation culture and 500 ml of Vitamin Solution (Table 1) were inoculated into the medium. During the fermentation, a solution of 50% glucose, 5% (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 0.05% citric acid was fed into the fermenter at a rate of 150 ml/hour, and the pH was maintained at approximately pH 5 by the addition of 2 M NH<sub>4</sub>OH. PPG-2025 was added as needed to control foaming. At approximately 49 hours post inoculation, an ethanol feed was begun by the addition of ethanol to the fermenter at a rate of 150 ml/min. The culture was grown for a total of 67.25 hours at 30°C.

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At the end of the fermentation, 50 liters of the culture was diluted to 100 liters with water. were removed from the spent medium by centrifuging 50 liters at a time through a Westfalia CSA 19 centrifuge (Westfalia, Oelde, Germany) at a flow rate liters/min. The cells were rinsed with water. From the centrifugation, approximately 20 liters of cell slurry containing approximately 35% cells was obtained. were added to the slurry to achieve a final concentration of the following salts: 50 mM NaCl, 10 mM Na2HPO4, 5 mM The cell slurry was passed through a Dynomill bead mill using 0.5 mm lead-free glass beads (Willy A Bachofen AG MashinenFabrik, Basle, Switzerland) at a rate of 4 liters per minute. The Dynomill was rinsed with Lysis buffer (50 mM NaCl, 10 mM Na2HPO4, 5 mM EDTA, pH 7.2) to a final volume of 80 liters. The final slurry had a pH of temperature of approximately 10°C and а a conductivity of 5 ms/cm.

The cell slurry was subjected to centrifugation as described above, and the cell pellet was rinsed with lysis buffer. After centrifugation approximately 20 liters of cell slurry was obtained. The cell slurry was extracted by first adjusting the concentration of the cell debris to approximately 40-50% with lysis buffer. Solid urea, NaCl and EDTA were added to the cell slurry to

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achieve a final concentration of approximately 8 M urea, 0.3 M NaCl and 10 mM EDTA. The approximate salt concentrations were obtained by the addition of 450 g/L of urea, 18 g/L of NaCl and 4.2 g/L of EDTA. The cell slurry was adjusted to pH 7.8 with 0.5 M NaOH. The solids were dissolved into the slurry and the pellets were extracted for a total of 50 minutes. Following extraction, the mixture was diluted 1 to 4 with water, adjusted to a conductivity of 12.5 ms/cm with NaCl and adjusted to a pH of 9.5 with 0.5 M NaOH.

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The extracted slurry was centrifuged as described above with the lysis buffer rinse. The the supernatant was adjusted to pH 9.5 with 0.5 M NaOH. The supernatant was analyzed by SDS polyacrylamide gel electrophoresis (SDS-PAGE) using the PHAST Separation and Control Unit (Pharmacia LKB Biotechnology Inc., Piscataway, NJ), and the protein was visualized using Coomassie Blue staining. A 2 liter Q-sepharose column (Pharmacia) was equilibrated at 5 liters/hour with successive washes of the following solutions: 8 liters of 3 M urea, 1 M NaCl, 50 mM glycine, pH 11.5; 5 liters of 0.5 M NaOH; 1.5 liters of water; 5 liters of 0.1 M HCl; and 6.0 liters of Wash buffer (50 mM glycine, 90 mM NaCl, with a conductivity of 12.5 ms/cm). supernatant (110 liters) was then applied to the column at 5 liters per hour.

The column ran dry after loading the The gel was resuspended in Wash buffer and supernatant. The repacked column was washed with 4 liters of 50 mM glycine, 90 mM NaCl, 5 mM EDTA, pH 10.0. material was eluted with elution buffer (50 mM glycine, 5 mM EDTA (pH 9.9) with a final concentration of NaCl giving a conductivity of 30.2 cm/ms (approximately 270 mM NaCl)). at 100 ml per minute. The approximately 600 ml fractions were collected after the conductivity of the eluant reached the conductivity of the elution buffer. Fractions

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were analyzed by SDS-PAGE analysis as described above and fractions 1 through 10 were pooled.

The pooled fractions were then applied to a 2 liter phenyl Sepharose column (Pharmacia) that had been equilibrated by successive washes at 5 liters per hour with the following solutions: 3 liters of 0.5 M NaOH; 3 liters of water; 3 liters of 2 M urea, 50 mM glycine, pH 10.5; 1.5 liters of water; 3 liters of 0.1 M HCl; and 3 liters of Equilibration buffer (50 mM glycine, 2.5 M NaCl, 2 mM EDTA (pH 10.0) with a conductivity of 180 ms/cm). The pooled peak fractions, which had been adjusted to a conductivity of 180 ms/cm with NaCl and a pH of 10.0 with 0.5 M NaOH, were loaded onto the phenyl sepharose column. Following the loading of the peak fractions, the column was washed with Equilibration buffer. The column was eluted with 6 liters of 50 mM glycine, 2 mM EDTA (pH 10.25) with a NaCl concentration giving the solution a conductivity of 96 ms/cm. The conductivity of the eluant was measured throughout the elution. The conductivity of the eluant upon starting the elution was 180 ms/cm. the third fraction, the conductivity of the eluant dropped At this point, the elution buffer was to 96 ms/cm. changed to a buffer having the conductivity of 42 ms/cm. The eluant was collected through fraction number 8.

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## Example 6 - <u>Cross-Linking Assay Using the Hybrid</u> Fibrinogen-Fibronectin Protein

The ability of the purified fibrinogen-fibronectin hybrid protein to form transglutminase-catalyzed interchain cross links was assessed. The transglutaminase activity was provided by the addition of recombinant factor XIII and thrombin or by the addition of recombinant factor XIIIa.

### 10 A. Preparation of Factor XIII

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Recombinant factor XIII was prepared essentially as described in co-pending U.S. Patent Application No. 07/927,196, which is incorporated by reference herein in its entirety. Briefly, factor XIII was isolated from a strain of the yeast Saccharomyces cerevisiae that had been transformed with an expression vector capable of directing the expression of factor XIII. The factor XIII-producing cells were harvested and lysed, and a cleared lysate was The lysate was fractionated by anion exchange prepared. chromatography at neutral to slightly alkaline pH using a column of derivatized agarose, such as DEAE FAST-FLOW SEPHAROSE (Pharmacia LKB Biotechnology, Piscataway, NJ) or Factor XIII was then precipitated from the the like. column eluate by concentrating the eluate and adjusting the pH to between 5.2 and 5.5, such as by diafiltration against ammonium succinate buffer. The precipitate was then dissolved and further purified using conventional chromatographic techniques, such as gel filtration and hydrophobic interaction chromatography. The purified factor XIII was dialyzed, filtered, aliquotted The factor XIIIa content was determined lyophilized. (Bishop et al., <u>Biochemistry</u> 29: 1861-1869, 1990, which is incorporated by reference herein in its entirety) fluorometric assay of the dissolved, thrombin-activated material.

Factor XIII was activated to factor XIIIa by adding 2 U of thrombin per 100 mg of factor XIII. The

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factor XIII was dissolved in buffer (20 mM sodium borate (pH 8.3), 1 mM  $CaCl_2$ ). The thrombin was added, and the reaction was incubated at room temperature for twenty minutes.

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### B. Cross-Linking Assays

The level of cross-linking between the hybrid proteins was measured as a rise in the absorbance at 350 nm over time in reaction mixtures containing the hybrid protein, factor XIII and thrombin or the hybrid protein Control reactions were prepared factor XIIIa. containing factor XIII and thrombin or factor XIIIa alone. Cross-linking reactions were carried out in 1 ml cuvettes. For cross-linking reactions containing factor XIII and thrombin, each reaction mixture was set up by placing 110 μl containing 40 Units of factor XIII, 36.7 μl containing 13 Units of factor XIII or 12.2  $\mu$ l containing 4 Units of factor XIII (described above) in one corner of the cuvette and 20 µl containing 4 Units of thrombin (Sigma) in the opposite corner such that the solutions were not mixed. The reaction was initiated by the addition of 1 ml of 2 mg/ml hybrid protein in buffer (10 mM Tris (pH 7.6), 20 mM sodium borate, 140 mM NaCl, 10 mM CaCl<sub>2</sub>). The absorbance of each reaction was read at 350 nm with the addition of protein being the first absorbance point. linking reactions containing factor XIIIa, each reaction was set up by placing 110  $\mu$ l containing 40 Units of factor XIIIa, 36.7  $\mu$ l containing 13 Units of factor XIIIa or 12.2 ul containing 4 Units of factor XIIIa in the cuvette and adding 1 ml of 2 mg/ml hybrid in buffer (10 mM Tris (pH 7.6), 140 mM NaCl, 10 mM CaCl<sub>2</sub>). The absorbance of the solution was read at 350 nm as described above. of the data generated from the absorbance time courses showed a sharp increase in absorbance in the presence of hybrid protein and the active transglutaminase relative to the rise in absorbance in the absence of hybrid protein 5

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(Figures 2-5). The results indicated that the hybrid protein is capable of transglutaminase-induced cross-linking.

From the foregoing it will be appreciated that, although specific embodiments of the invention have been described herein for the purpose of illustration, various modifications may be made without deviation from the spirit and scope of the invention. Accordingly, the invention is not to be limited except as by the following claims.

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#### SEQUENCE LISTING

- (1) GENERAL INFORMATION:
  - (i) APPLICANT: Irani, Meher H.
  - (ii) TITLE OF INVENTION: HYBRID CROSS-LINKING PROTEINS
  - (iii) NUMBER OF SEQUENCES: 14
    - (iv) CORRESPONDENCE ADDRESS:
      - (A) ADDRESSEE: ZymoGenetics, Inc.
      - (B) STREET: 4225 Roosevelt Way, N.E.
      - (C) CITY: Seattle
      - (D) STATE: WA
      - (E) COUNTRY: USA
      - (F) ZIP: 98105
    - (v) COMPUTER READABLE FORM:
      - (A) MEDIUM TYPE: Floppy disk
      - (B) COMPUTER: IBM PC compatible
      - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
      - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
    - (vi) CURRENT APPLICATION DATA:
      - (A) APPLICATION NUMBER: WO
      - (B) FILING DATE:
      - (C) CLASSIFICATION:
  - (vii) PRIOR APPLICATION DATA:
    - (A) APPLICATION NUMBER: US 07/998,271
    - (B) FILING DATE: 31-DEC-1992
  - (viii) ATTORNEY/AGENT INFORMATION:
    - (A) NAME: Parker, Gary E
    - (B) REGISTRATION NUMBER: 31-648
    - (C) REFERENCE/DOCKET NUMBER: 92-26PC
    - (ix) TELECOMMUNICATION INFORMATION:
      - (A) TELEPHONE: 206-547-8080 ext 322
      - (B) TELEFAX: 206-548-2329
- (2) INFORMATION FOR SEQ ID NO:1:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 7803 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ix) FEATURE:
    - (A) NAME/KEY: CDS

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(B) LOCATION: 6..7346

37

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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			GGG Gly													g	95
			CAG Gln													14	43
			CCC Pro 50													19	91
CAA Gln	CAG Gln	TGG Trp 65	GAG Glu	CGG Arg	ACC Thr	TAC Tyr	CTA Leu 70	GGT Gly	AAT Asn	GTG Val	TTG Leu	GTT Val 75	TGT Cys	ACT Thr	TGT Cys	23	39
			AGC Ser													28	87
			TTT Phe													33	35
			CGT Arg													38	83
			CGA Arg 130													43	31
GAA Glu	GGG Gly	GGT Gly 145	CAG Gln	TCC Ser	TAC Tyr	AAG Lys	ATT Ile 150	GGT Gly	GAC Asp	ACC Thr	TGG Trp	AGG Arg 155	AGA Arg	CCA Pro	CAT His	47	79
			GGT Gly													57	27
			ACC Thr													57	75
GCT Ala	GGG Gly	ACT Thr	TCC Ser	TAT Tyr 195	GTG Val	GTC Val	GGA Gly	GAA Glu	ACG Thr 200	TGG Trp	GAG Glu	AAG Lys	CCC Pro	TAC Tyr 205	CAA Gln	6	23

		GAT Asp						671
		AGA Arg						719
		GAC Asp						767
		TGC Cys 260						815
		GTG Val						863
		GCT Ala						911
		TGT Cys						959
		AAG Lys						1007
		GTC Val 340						1055
		AAT Asn						1103
		TAC Tyr						1151
		ACA Thr						1199
		CAC His						1247
		TGC Cys 420						1295

					TCT Ser											1343
GGG Gly	ACC Thr	ACA Thr	CAG Gln 450	AAC Asn	TAT Tyr	GAT Asp	GCC Ala	GAC Asp 455	CAG Gln	AAG Lys	TTT Phe	GGG Gly	TTC Phe 460	TGC Cys	CCC Pro	1391
ATG Met	GCT Ala	GCC Ala <b>465</b>	CAC His	GAG Glu	GAA Glu	ATC Ile	TGC Cys <b>470</b>	ACA Thr	ACC Thr	AAT Asn	GAA Glu	GGG Gly 475	GTC Val	ATG Met	TAC Tyr	1439
CGC Arg	ATT Ile <b>4</b> 80	GGA Gly	GAT Asp	CAG Gln	TGG Trp	GAT Asp 485	AAG Lys	CAG Gln	CAT His	GAC Asp	ATG Met 490	GGT Gly	CAC His	ATG Met	ATG Met	1487
					GGG Gly 500											1535
TAC Tyr	TCG Ser	CAA Gln	CTT Leu	CGA Arg 515	GAT Asp	CAG Gln	TGC Cys	ATT Ile	GTT Val 520	GAT Asp	GAC Asp	ATC Ile	ACT Thr	TAC Tyr <b>525</b>	AAT Asn	1583
GTG Val	AAC Asn	GAC Asp	ACA Thr 530	TTC Phe	CAC His	AAG Lys	CGT Arg	CAT His 535	GAA Glu	GAG Glu	GGG Gly	CAC His	ATG Met 540	CTG Leu	AAC Asn	1631
					CAG Gln											1679
					TCA Ser											1727
TCA Ser 575	TGG Trp	GAG Glu	AAG Lys	TAT Tyr	GTG Val 580	CAT His	GGT Gly	GTC Val	AGA Arg	TAC Tyr 585	CAG Gln	TGC Cys	TAC Tyr	TGC Cys	TAT Tyr 590	1775
GGC Gly	CGT Arg	GGC Gly	ATT Ile	GGG Gly 595	GAG Glu	TGG Trp	CAT His	TGC Cys	CAA Gln 600	CCT Pro	TTA Leu	CAG Gln	ACC Thr	TAT Tyr 605	CCA Pro	1823
				Pro	GTC Val											1871
					ATC Ile			Asn					Ser			1919

			TGG Trp 645					1967
			GGC Gly					2015
			TAC Tyr					2063
			ACT Thr					2111
			AAC Asn					2159
			TCT Ser 725					2207
			GTC Val					2255
			AGT Ser					2303
			ACT Thr					2351
			AAT Asn					2399
			ACT Thr 805					2447
			CAA Gln					2495
			CCC Pro					2543
			AGC Ser					2591

AAC TCC GTC Asn Ser Val 865												2639
ACT ATC TAT Thr Ile Tyr 880												2687
CAA CAA GAA Gln Gln Glu 895												2735
AGG GAC CTG Arg Asp Leu												2783
TGG ACA CCG Trp Thr Pro			Val									2831
CCC GTC AAC Pro Val Asn 945												2879
AAC ACC TTT Asn Thr Phe 960												2927
TTC AAA GTC Phe Lys Val 975												2975
GCT CAA CAG Ala Gln Gln						Thr					Val	3023
AAT GAA ACT Asn Glu Thr					Arg					Arg		3071
CAG ATA ACA Gln Ile Thr 102	Gly Tyr			Val					Arg			3119
CCC AGG CAG Pro Arg Gln 1040	TAC AAT Tyr Asn	GTG GGT Val Gly 104	Pro	TCT Ser	GTC Val	TCC Ser	AAG Lys 1050	Tyr	CCC Pro	CTG Leu	AGG Arg	3167
AAT CTG CAG Asn Leu Gln 1055	CCT GCA Pro Ala	TCT GAG Ser Glu 1060	TAC Tyr	ACC Thr	GTA Val	TCC Ser 106	Leu	GTG Val	GCC Ala	ATA Ile	AAG Lys 1070	3215

AAC Asn				Pro					Val					Gln	3263
GGG Gly			Пe					Thr					Thr		3311
GTG Val		Thr					Pro					Lys			3359
CGA Arg 1120	Pro					Glu					Val				3407
GGA Gly					Ser					Gly					3455
ACC Thr				Leu					Glu					Ile	3503
AAC Asn			Val					Pro					His		3551
GCA Ala		Pro					Leu					Glu			3599
ACC Thr 1200	Pro					Tyr					Thr				3647
CAG Gln					Leu					His					3695
TGC Cys				Asn					Leu					Ser	3743
TAC Tyr			Lys					Ser					Asp		3791
ATC Ile		Glu					Thr					Val			3839
GAT Asp 1280	Ser					Arg					Asn				3887

ATT ATT GGG TAC ( Ile Ile Gly Tyr A 1295	CGC ATC ACA GTA Arg Ile Thr Val 1300	GTT GCG GCA GG/ Val Ala Ala Gl 1305	y Glu Gly Ile	CCT 3935 Pro 1310
ATT TTT GAA GAT I Ile Phe Glu Asp I	TTT GTG TAC TCC Phe Val Tyr Ser 1315	TCA GTA GGA TA Ser Val Gly Ty 1320	C TAC ACA GTC r Tyr Thr Val 1325	Thr
GGG CTG GAG CCG ( Gly Leu Glu Pro ( 1330	Gly Ile Asp Tyr	GAT ATC AGC GT Asp Ile Ser Va 1335	T ATC ACT CTC 1 Ile Thr Leu 1340	ATT 4031 Ile
AAT GGC GGC GAG A Asn Gly Gly Glu S 1345	AGT GCC CCT ACT Ser Ala Pro Thr 1350	Thr Leu Thr Gl	A CAA ACG GCT n Gln Thr Ala 1355	GTT 4079 Val
CCT CCT CCC ACT ( Pro Pro Pro Thr / 1360	GAC CTG CGA TTC Asp Leu Arg Phe 1365	ACC AAC ATT GG Thr Asn Ile Gl 13	y Pro Asp Thr	ATG 4127 Met
CGT GTC ACC TGG ( Arg Val Thr Trp / 1375	GCT CCA CCC CCA Ala Pro Pro Pro 1380	TCC ATT GAT TT. Ser Ile Asp Le 1385	u Thr Asn Phe	CTG 4175 Leu 1390
GTG CGT TAC TCA ( Val Arg Tyr Ser				Ser
ATT TCT CCT TCA ( Ile Ser Pro Ser / 1410	Asp Asn Ala Val	GTC TTA ACA AA Val Leu Thr As 1415	T CTC CTG CCT n Leu Leu Pro 1420	GGT 4271 Gly
ACA GAA TAT GTA ( Thr Glu Tyr Val 1 1425	GTG AGT GTC TCC Val Ser Val Ser 1430	Ser Val Tyr Gl	A CAA CAT GAG u Gln His Glu 1435	AGC 4319 Ser
ACA CCT CTT AGA Thr Pro Leu Arg 1440	GGA AGA CAG AAA Gly Arg Gln Lys 1445	Thr Gly Leu As	T TCC CCA ACT p Ser Pro Thr 50	GGC 4367 Gly
ATT GAC TTT TCT Ile Asp Phe Ser 1455	GAT ATT ACT GCC Asp Ile Thr Ala 1460	AAC TCT TTT AC Asn Ser Phe Th 1465	T GTG CAC TGG r Val His Trp	ATT 4415 Ile 1470
GCT CCT CGA GCC Ala Pro Arg Ala	ACC ATC ACT GGC Thr Ile Thr Gly 1475	TAC AGG ATC CG Tyr Arg Ile Ar 1480	CC CAT CAT CCC g His His Pro 1485	Glu
CAC TTC AGT GGG His Phe Ser Gly 1490	Arg Pro Arg Glu	GAT CGG GTG CC Asp Arg Val Pr 1495	CC CAC TCT CGG TO His Ser Arg 1500	AAT 4511 Asn

TCC ATC ACC CTC Ser Ile Thr Leu 1505						4559
ATC GTT GCT CTT Ile Val Ala Leu 1520		Glu Glu Se		Leu Ile (		4607
CAA TCA ACA GTT Gln Ser Thr Val 1535						4655
ACC CCC ACC AGC Thr Pro Thr Ser			p Ala Pro	Ala Val		4703
AGA TAT TAC AGG Arg Tyr Tyr Arg 157	Ile Thr Tyr					4751
CAG GAG TTC ACT Gln Glu Phe Thr 1585						4799
CTT AAA CCT GGA Leu Lys Pro Gly 1600		Thr Ile Th		Ala Val		4847
CGT GGA GAC AGC Arg Gly Asp Ser 1615						4895
ACA GAA ATT GAC Thr Glu Ile Asp			n Val Thr	Asp Val (		4943
AAC AGC ATT AGT Asn Ser Ile Ser 165	Val Lys Trp				Thr Gly	4991
TAC AGA GTA ACC Tyr Arg Val Thr 1665						5039
AAA ACT GCA GGT Lys Thr Ala Gly 1680		Thr Glu Me		Glu Gly I		5087
CCC ACA GTG GAG Pro Thr Val Glu 1695						5135
GAG AGT CAG CCT Glu Ser Gln Pro			1 Thr Asn	Ile Asp A		5183

AAA GGA Lys Gly	CTG GCA Leu Ala 173	Phe Thr	GAT GTG Asp Val	GAT GTC Asp Val 1735	GAT TO Asp Se	CC ATC AAA er Ile Lys 1740	Ile .	GCT Ala	5231
TGG GAA Trp Glu	AGC CCA Ser Pro 17 <b>4</b> 5	CAG GGG Gln Gly	CAA GTT Gln Val 175	Ser Arg	TAC AC Tyr Ai	GG GTG ACC rg Val Thr 1755	TAC Tyr	TCG Ser	5279
AGC CCT Ser Pro 176	Glu Asp	GGA ATC Gly Ile	CAT GAG His Glu 1765	CTA TTC Leu Phe	Pro A	CA CCT GAT la Pro Asp 770	GGT Gly	GAA Glu	5327
GAA GAC Glu Asp 1775	ACT GCA Thr Ala	GAG CTG Glu Leu 178	Gln Gly	CTC AGA Leu Arg	CCG GG Pro G 1785	GT TCT GAG ly Ser Glu	Tyr	ACA Thr 1790	5375
					Glu Se	GC CAG CCC er Gln Pro		Ile	5423
GGA ACC Gly Thr	CAG TCC Gln Ser 181	Thr Ala	ATT CCT Ile Pro	GCA CCA Ala Pro 1815	ACT GA	AC CTG AAG sp Leu Lys 182	Phe	ACT Thr	5471
CAG GTC Gln Val	ACA CCC Thr Pro 1825	ACA AGC Thr Ser	CTG AGC Leu Ser 183	Ala Gln	TGG AG	CA CCA CCC hr Pro Pro 1835	AAT Asn	GTT Val	5519
CAG CTC Gln Leu 184	Thr Gly	TAT CGA Tyr Arg	GTG CGG Val Arg 1845	GTG ACC Val Thr	Pro L	AG GAG AAG ys Glu Lys 850	ACC Thr	GGA Gly	5567
			Leu Ala			CA TCC GTG er Ser Val			5615
TCA GGA Ser Gly	CTT ATG Leu Met	GTG GCC Val Ala 1875	ACC AAA Thr Lys	TAT GAA Tyr Glu 188	Val S	GT GTC TAT Ser Val Tyr	GCT Ala 1885	Leu	5663
AAG GAC Lys Asp	ACT TTG Thr Leu 189	Thr Ser	AGA CCA Arg Pro	GCT CAG Ala Gln 1895	GGT G Gly V	GTT GTC ACC Val Val Thr 190	Thr	CTG Leu	5711
GAG AAT Glu Asr	GTC AGC Val Ser 1905	CCA CCA Pro Pro	AGA AGG Arg Arg 191	Ala Arg	GTG A	ACA GAT GCT Thr Asp Ala 1915	ACT Thr	GAG Glu	5759
ACC ACC Thr Thr 192	· Ile Thr	ATT AGC Ile Ser	TGG AGA Trp Arg 1925	ACC AAG Thr Lys	: Thr G	GAG ACG ATC Glu Thr Ile 1930	ACT Thr	GGC Gly	5807

TTC CAA GTT GAT GC Phe Gln Val Asp Al 1935			Thr Pro Ile Gln	
ACC ATC AAG CCA GA Thr Ile Lys Pro As				Pro
GGC ACT GAC TAC AA Gly Thr Asp Tyr Ly 1970				
AGC TCC CCT GTG GT Ser Ser Pro Val Va 1985		Ser Thr Ala		
AAC CTG CGT TTC CT Asn Leu Arg Phe Le 2000				
CAG CCG CCA CGT GC Gln Pro Pro Arg Al 2015			Ile Lys Tyr Glu	
CCT GGG TCT CCT CC Pro Gly Ser Pro Pr 20				Val
ACA GAG GCT ACT AT Thr Glu Ala Thr Il 2050				
TAT GTC ATT GCC CT Tyr Val Ile Ala Le 2065		Gln Lys Ser		
AGG AAA AAG ACA GA Arg Lys Lys Thr As 2080	C GAG CTT CCC p Glu Leu Pro 2085	CAA CTG GTA Gln Leu Val	ACC CTT CCA CAC Thr Leu Pro His 2090	CCC 6287 Pro
AAT CTT CAT GGA CC Asn Leu His Gly Pr 2095			Ser Thr Val Gln	
ACC CCT TTC GTC AC Thr Pro Phe Val Th 21				Gln
CTT CCT GGC ACT TO Leu Pro Gly Thr Se 2130				
TTT GAG GAA CAT GG Phe Glu Glu His Gl 2145		Thr Thr Pro		

CCC ATA AGG CAT Pro Ile Arg His 2160				
GCT CTC TCT CAG Ala Leu Ser Gln 2175			Phe Gln Asp	
GAG TAC ATC ATT Glu Tyr Ile Ile			Asp Glu Glu	
CAG TTC AGG GTT Gln Phe Arg Val 221	Pro Gly Thr Se			
ACC AGA GGT GCC Thr Arg Gly Ala 2225	Thr Tyr Asn Il			
CAG AGG CAT AAG Gln Arg His Lys 2240				
AAC GAA GGC TTG Asn Glu Gly Leu 2255			Cys Phe Asp	
ACA GTT TCC CAT Thr Val Ser His			Glu Arg Met	
TCA GGC TTT AAA Ser Gly Phe Lys 229	Leu Leu Cys G	G TGC TTA GGC n Cys Leu Gly 2295	TTT GGA AGT Phe Gly Ser 2300	GGT CAT 6911 Gly His
TTC AGA TGT GAT Phe Arg Cys Asp 2305	Ser Ser Arg Ti	G TGC CAT GAC p Cys His Asp 10	AAT GGT GTG Asn Gly Val 2315	AAC TAC 6959 Asn Tyr
AAG ATT GGA GAG Lys Ile Gly Glu 2320				
AGC TGC ACA TGT Ser Cys Thr Cys 2335			Phe Lys Cys	
CAT GAG GCA ACG His Glu Ala Thr	TGT TAC GAT GA Cys Tyr Asp A 2355	AT GGG AAG ACA op Gly Lys Thr 2360	Tyr His Val	GGA GAA 7103 Gly Glu 2365

CAG TGG Gln Trp			Glu					Ile					Cys		7151
GGA GGC Gly Gly		Arg					Asp					Pro			7199
GAA CCC Glu Pro 2400	Ser					Thr					Asn				7247
CAG AGA Gln Arg 2415					Thr					Asn					7295
TGC TTC Cys Phe		Pro		Asp					Arg					Glu	7343
TAAATCAT	rct 1	TTCCA	ATCC	A GA	\GGA <i>P</i>	CAAG	CAT	GTCT	СТС	TGCC	:AAG <i>A</i>	ATC C	CATCT	TAAACT	7403
GGAGTGAT	rgt 1	TAGCA	GACC	C AG	CTTA	\GAGT	ТСТ	тстт	TCT	TTCT	TAAG	acc c	CTTTG	CTCTG	7463
GAGGAAGT	TC T	ГССАG	CTTC	A GC	TCAA	CTCA	CAG	CTTC	TCC	AAGC	ATCA	CC C	TGGG	AGTTT	7523
CCTGAGGG	T T	гтстс	ATAA	A TO	AGGG	CTGC	ACA	TTGC	CTG	TTCT	GCTT	CG A	AGTA	TTCAA	7583
TACCGCTC	AG T	TATTT	TAAA	T GA	AGTO	ATTO	TAA	GATT	TGG	TTTG	IGGAT	CA A	TAGG	AAAGC	7643
ATATGCAG	icc A	AACCA	AGAT	G CA	AATG	TTTT	GAA	ATGA	TAT	GACC	AAAA	TT T	TAAG	TAGGA	7703
AAGTCACC	CA A	ACAC	TTCT	G CT	TTCA	CTTA	AGT	GTCT	GGC	CCGC	AATA	CT G	TAGG	AACAA	7763
GCATGATO	CTT G	STTAC	TGTG	A TA	TTTT	TAAAT	ATC	CACA	GTA						7803

## (2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2446 amino acids
  - (B) TYPE: amino acid
    (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Leu Arg Gly Pro Gly Pro Gly Leu Leu Leu Leu Ala Val Leu Cys 1 5 10 15

Leu Gly Thr Ala Val Pro Ser Thr Gly Ala Ser Lys Ser Lys Arg Gln  $20 \hspace{1cm} 25 \hspace{1cm} 30$ 

320

Ala Gln Gln Met Val Gln Pro Gln Ser Pro Val Ala Val Ser Gln Ser 40 Lys Pro Gly Cys Tyr Asp Asn Gly Lys His Tyr Gln Ile Asn Gln Gln Trp Glu Arg Thr Tyr Leu Gly Asn Val Leu Val Cys Thr Cys Tyr Gly Gly Ser Arg Gly Phe Asn Cys Glu Ser Lys Pro Glu Ala Glu Glu Thr Cys Phe Asp Lys Tyr Thr Gly Asn Thr Tyr Arg Val Gly Asp Thr Tyr 105 Glu Arg Pro Lys Asp Ser Met Ile Trp Asp Cys Thr Cys Ile Gly Ala Gly Arg Gly Arg Ile Ser Cys Thr Ile Ala Asn Arg Cys His Glu Gly Gly Gln Ser Tyr Lys Ile Gly Asp Thr Trp Arg Arg Pro His Glu Thr Gly Gly Tyr Met Leu Glu Cys Val Cys Leu Gly Asn Gly Lys Gly Glu Trp Thr Cys Lys Pro Ile Ala Glu Lys Cys Phe Asp His Ala Ala Gly Thr Ser Tyr Val Val Gly Glu Thr Trp Glu Lys Pro Tyr Gln Gly Trp Met Met Val Asp Cys Thr Cys Leu Gly Glu Gly Ser Gly Arg Ile Thr Cys Thr Ser Arg Asn Arg Cys Asn Asp Gln Asp Thr Arg Thr Ser Tyr 225 Arg Ile Gly Asp Thr Trp Ser Lys Lys Asp Asn Arg Gly Asn Leu Leu 250 Gln Cys Ile Cys Thr Gly Asn Gly Arg Gly Glu Trp Lys Cys Glu Arg His Thr Ser Val Gln Thr Thr Ser Ser Gly Ser Gly Pro Phe Thr Asp 280 Val Arg Ala Ala Val Tyr Gln Pro Gln Pro His Pro Gln Pro Pro 300 Tyr Gly His Cys Val Thr Asp Ser Gly Val Val Tyr Ser Val Gly Met

Gln	Trp	Leu	Lys	Thr 325	Gln	Gly	Asn	Lys	Gln 330	Met	Leu	Cys	Thr	Cys <b>335</b>	Leu
Gly	Asn	Gly	Va1 340	Ser	Cys	Gln	Glu	Thr 345	Ala	Val	Thr	Gln	Thr 350	Tyr	Gly
Gly	Asn	Leu 355	Asn	Gly	Glu	Pro	Cys 360	Val	Leu	Pro	Phe	Thr 365	Tyr	Asn	Gly
Arg	Thr 370	Phe	Tyr	Ser	Cys	Thr 375	Thr	Glu	Gly	Arg	Gln 380	Asp	Gly	His	Leu
Trp 385	Cys	Ser	Thr	Thr	Ser 390	Asn	Tyr	Glu	Gln	Asp 395	Gln	Lys	Tyr	Ser	Phe 400
Cys	Thr	Asp	His	Thr 405	Val	Leu	Val	Gln	Thr 410	Gln	Gly	Gly	Asn	Ser 415	Asn
Gly	Ala	Leu	Cys <b>420</b>	His	Phe	Pro	Phe	Leu 425	Tyr	Asn	Asn	His	Asn 430	Tyr	Thr
Asp	Cys	Thr 435	Ser	Glu	Gly	Arg	Arg <b>44</b> 0	Asp	Asn	Met	Lys	Trp 445	Cys	Gly	Thr
Thr	G1n <b>45</b> 0	Asn	Tyr	Asp	Ala	Asp <b>45</b> 5	Gln	Lys	Phe	Gly	Phe 460	Cys	Pro	Met	Ala
A1a 465	His	Glu	Glu	Ile	Cys 470	Thr	Thr	Asn	Glu	Gly 475	Val	Met	Tyr	Arg	Ile 480
Gly	Asp	Gln	Trp	Asp 485	Lys	Gln	His	Asp	Met 490	Gly	His	Met	Met	Arg 495	Cys
Thr	Cys	Val	Gly 500	Asn	Gly	Arg	Gly	G1u 505	Trp	Thr	Cys	Ile	Ala 510	Tyr	Ser
Gln	Leu	Arg 515	Asp	Gln	Cys	Ile	Val 520	Asp	Asp	Ile	Thr	Tyr 525	Asn	Val	Asn
Asp	Thr 530	Phe	His	Lys	Arg	His <b>535</b>	G1u	G1u	Gly	His	Met 540	Leu	Asn	Cys	Thr
Cys <b>54</b> 5	Phe	Gly	Gln	Gly	Arg 550	Gly	Arg	Trp	Lys	Cys <b>555</b>	Asp	Pro	Val	Asp	Gln 560
Cys	Gln	Asp	Ser	G1u 565	Thr	Gly	Thr	Phe	Tyr 570	Gln	Ile	Gly	Asp	Ser 575	Trp
Glu	Lys	Tyr	Val 580	His	Gly	Val	Arg	Tyr 585	Gln	Cys	Tyr	Cys	Tyr <b>59</b> 0	Gly	Arg
Gly	Ile	Gly 595	Glu	Trp	His	Cys	Gln 600	Pro	Leu	Gln	Thr	Tyr 605	Pro	Ser	Ser

Ser Gly Pro Val Glu Val Phe Ile Thr Glu Thr Pro Ser Gln Pro Asn 620 Ser His Pro Ile Gln Trp Asn Ala Pro Gln Pro Ser His Ile Ser Lys 630 Tyr Ile Leu Arg Trp Arg Pro Lys Asn Ser Val Gly Arg Trp Lys Glu 650 655 Ala Thr Ile Pro Gly His Leu Asn Ser Tyr Thr Ile Lys Gly Leu Lys Pro Gly Val Val Tyr Glu Gly Gln Leu Ile Ser Ile Gln Gln Tyr Gly 680 His Gln Glu Val Thr Arg Phe Asp Phe Thr Thr Ser Thr Ser Thr Pro Val Thr Ser Asn Thr Val Thr Gly Glu Thr Thr Pro Phe Ser Pro 715 710 Leu Val Ala Thr Ser Glu Ser Val Thr Glu Ile Thr Ala Ser Ser Phe Val Val Ser Trp Val Ser Ala Ser Asp Thr Val Ser Gly Phe Arg Val Glu Tyr Glu Leu Ser Glu Glu Gly Asp Glu Pro Gln Tyr Leu Asp Leu 765 Pro Ser Thr Ala Thr Ser Val Asn Ile Pro Asp Leu Leu Pro Gly Arg Lys Tyr Ile Val Asn Val Tyr Gln Ile Ser Glu Asp Gly Glu Gln Ser Leu Ile Leu Ser Thr Ser Gln Thr Thr Ala Pro Asp Ala Pro Pro Asp 805 810 Pro Thr Val Asp Gln Val Asp Asp Thr Ser Ile Val Val Arg Trp Ser 825 Arg Pro Gln Ala Pro Ile Thr Gly Tyr Arg Ile Val Tyr Ser Pro Ser 840 Val Glu Gly Ser Ser Thr Glu Leu Asn Leu Pro Glu Thr Ala Asn Ser 855 Val Thr Leu Ser Asp Leu Gln Pro Gly Val Gln Tyr Asn Ile Thr Ile 875 880 Tyr Ala Val Glu Glu Asn Gln Glu Ser Thr Pro Val Val Ile Gln Gln 890 885

- Glu Thr Thr Gly Thr Pro Arg Ser Asp Thr Val Pro Ser Pro Arg Asp 900 905 910
- Leu Gln Phe Val Glu Val Thr Asp Val Lys Val Thr Ile Met Trp Thr 915 920 925
- Pro Pro Glu Ser Ala Val Thr Gly Tyr Arg Val Asp Val Ile Pro Val 930 935 940
- Asn Leu Pro Gly Glu His Gly Gln Arg Leu Pro Ile Ser Arg Asn Thr 945 950 955 960
- Phe Ala Glu Val Thr Gly Leu Ser Pro Gly Val Thr Tyr Tyr Phe Lys 965 970 975
- Val Phe Ala Val Ser His Gly Arg Glu Ser Lys Pro Leu Thr Ala Gln 980 985 990
- Gln Thr Thr Lys Leu Asp Ala Pro Thr Asn Leu Gln Phe Val Asn Glu 995 1000 1005
- Thr Asp Ser Thr Val Leu Val Arg Trp Thr Pro Pro Arg Ala Gln Ile 1010 1015 1020
- Thr Gly Tyr Arg Leu Thr Val Gly Leu Thr Arg Arg Gly Gln Pro Arg 1025 1030 1035 1040
- Gln Tyr Asn Val Gly Pro Ser Val Ser Lys Tyr Pro Leu Arg Asn Leu 1045 1050 1055
- Gln Pro Ala Ser Glu Tyr Thr Val Ser Leu Val Ala Ile Lys Gly Asn 1060 1065 1070
- Gln Glu Ser Pro Lys Ala Thr Gly Val Phe Thr Thr Leu Gln Pro Gly 1075 1080 1085
- Ser Ser Ile Pro Pro Tyr Asn Thr Glu Val Thr Glu Thr Thr Ile Val 1090 1095 1100
- Ile Thr Trp Thr Pro Ala Pro Arg Ile Gly Phe Lys Leu Gly Val Arg 1105 1110 1115 1120
- Pro Ser Gln Gly Glu Ala Pro Arg Glu Val Thr Ser Asp Ser Gly 1125 1130 1135
- Ser Ile Val Val Ser Gly Leu Thr Pro Gly Val Glu Tyr Val Tyr Thr 1140 1145 1150
- Ile Gln Val Leu Arg Asp Gly Gln Glu Arg Asp Ala Pro Ile Val Asn 1155 1160 1165
- Lys Val Val Thr Pro Leu Ser Pro Pro Thr Asn Leu His Leu Glu Ala 1170 1175 1180

Asn Pr 1185	o Asp	Thr	Gly	Val 1190	Leu )	Thr	Va1	Ser				Ser		Thr 1200
Pro As	p Ile	Thr		Tyr 5								Asn	Gly 121	
Gln Gl	y Asn	Ser 1220	Leu )	Glu	Glu	Val	Val 1225	His 5	Ala	Asp	Gln	Ser 1230		Cys
Thr Ph		Asn 5	Leu	Ser	Pro	Gly 1240		G1 u	Tyr	Asn	Val 124		Val	Tyr

Thr Val Lys Asp Asp Lys Glu Ser Val Pro Ile Ser Asp Thr Ile Ile 1250 1255 1260

Pro Glu Val Pro Gln Leu Thr Asp Leu Ser Phe Val Asp Ile Thr Asp 1265 1270 1275 1280

Ser Ser Ile Gly Leu Arg Trp Thr Pro Leu Asn Ser Ser Thr Ile Ile 1285 1290 1295

Gly Tyr Arg Ile Thr Val Val Ala Ala Gly Glu Gly Ile Pro Ile Phe 1300 1305 1310

Glu Asp Phe Val Tyr Ser Ser Val Gly Tyr Tyr Thr Val Thr Gly Leu 1315 1320 1325

Glu Pro Gly Ile Asp Tyr Asp Ile Ser Val Ile Thr Leu Ile Asn Gly 1330 1335 1340

Gly Glu Ser Ala Pro Thr Thr Leu Thr Gln Gln Thr Ala Val Pro Pro 1345 1350 1355 1360

Pro Thr Asp Leu Arg Phe Thr Asn Ile Gly Pro Asp Thr Met Arg Val 1365 1370 1375

Thr Trp Ala Pro Pro Pro Ser Ile Asp Leu Thr Asn Phe Leu Val Arg 1380 1385 1390

Tyr Ser Pro Val Lys Asn Glu Glu Asp Val Ala Glu Leu Ser Ile Ser 1395 1400 1405

Pro Ser Asp Asn Ala Val Val Leu Thr Asn Leu Leu Pro Gly Thr Glu 1410 1415 1420

Tyr Val Val Ser Val Ser Ser Val Tyr Glu Gln His Glu Ser Thr Pro 1425 1430 1435 1440

Leu Arg Gly Arg Gln Lys Thr Gly Leu Asp Ser Pro Thr Gly Ile Asp 1445 1450 1455

Phe Ser Asp Ile Thr Ala Asn Ser Phe Thr Val His Trp Ile Ala Pro 1460 1465 1470

- Arg Ala Thr Ile Thr Gly Tyr Arg Ile Arg His His Pro Glu His Phe 1475 1480 1485
- Ser Gly Arg Pro Arg Glu Asp Arg Val Pro His Ser Arg Asn Ser Ile 1490 1495 1500
- Thr Leu Thr Asn Leu Thr Pro Gly Thr Glu Tyr Val Val Ser Ile Val 1505 1510 1515 1520
- Ala Leu Asn Gly Arg Glu Glu Ser Pro Leu Leu Ile Gly Gln Gln Ser 1525 1530 1535
- Thr Val Ser Asp Val Pro Arg Asp Leu Glu Val Val Ala Ala Thr Pro
  1540 1545 1550
- Thr Ser Leu Leu Ile Ser Trp Asp Ala Pro Ala Val Thr Val Arg Tyr 1555 1560 1565
- Tyr Arg Ile Thr Tyr Gly Glu Thr Gly Gly Asn Ser Pro Val Gln Glu 1570 1575 1580
- Phe Thr Val Pro Gly Ser Lys Ser Thr Ala Thr Ile Ser Gly Leu Lys 1585 1590 1595 1600
- Pro Gly Val Asp Tyr Thr Ile Thr Val Tyr Ala Val Thr Gly Arg Gly 1605 1610 1615
- Asp Ser Pro Ala Ser Ser Lys Pro Ile Ser Ile Asn Tyr Arg Thr Glu 1620 1625 1630
- Ile Asp Lys Pro Ser Gln Met Gln Val Thr Asp Val Gln Asp Asn Ser 1635 1640 1645
- Ile Ser Val Lys Trp Leu Pro Ser Ser Ser Pro Val Thr Gly Tyr Arg 1650 1655 1660
- Val Thr Thr Thr Pro Lys Asn Gly Pro Gly Pro Thr Lys Thr Lys Thr 1665 1670 1675 1680
- Ala Gly Pro Asp Gln Thr Glu Met Thr Ile Glu Gly Leu Gln Pro Thr 1685 1690 1695
- Val Glu Tyr Val Val Ser Val Tyr Ala Gln Asn Pro Ser Gly Glu Ser 1700 1705 1710
- Gln Pro Leu Val Gln Thr Ala Val Thr Asn Ile Asp Arg Pro Lys Gly
  1715 1720 1725
- Leu Ala Phe Thr Asp Val Asp Val Asp Ser Ile Lys Ile Ala Trp Glu 1730 1735 1740
- Ser Pro Gln Gly Gln Val Ser Arg Tyr Arg Val Thr Tyr Ser Ser Pro 1745 1750 1755 1760

- Glu Asp Gly Ile His Glu Leu Phe Pro Ala Pro Asp Gly Glu Glu Asp 1765 1770 1775
- Thr Ala Glu Leu Gln Gly Leu Arg Pro Gly Ser Glu Tyr Thr Val Ser 1780 1785 1790
- Val Val Ala Leu His Asp Asp Met Glu Ser Gln Pro Leu Ile Gly Thr 1795 1800 1805
- Gln Ser Thr Ala Ile Pro Ala Pro Thr Asp Leu Lys Phe Thr Gln Val 1810 1815 1820
- Thr Pro Thr Ser Leu Ser Ala Gln Trp Thr Pro Pro Asn Val Gln Leu 1825 1830 1835 1840
- Thr Gly Tyr Arg Val Arg Val Thr Pro Lys Glu Lys Thr Gly Pro Met 1845 1850 1855
- Lys Glu Ile Asn Leu Ala Pro Asp Ser Ser Ser Val Val Ser Gly 1860 1865 1870
- Leu Met Val Ala Thr Lys Tyr Glu Val Ser Val Tyr Ala Leu Lys Asp 1875 1880 1885
- Thr Leu Thr Ser Arg Pro Ala Gln Gly Val Val Thr Thr Leu Glu Asn 1890 1895 1900
- Val Ser Pro Pro Arg Arg Ala Arg Val Thr Asp Ala Thr Glu Thr Thr 1905 1910 1915 1920
- Ile Thr Ile Ser Trp Arg Thr Lys Thr Glu Thr Ile Thr Gly Phe Gln
  1925 1930 1935
- Val Asp Ala Val Pro Ala Asn Gly Gln Thr Pro Ile Gln Arg Thr Ile 1940 1945 1950
- Lys Pro Asp Val Arg Ser Tyr Thr Ile Thr Gly Leu Gln Pro Gly Thr 1955 1960 1965
- Asp Tyr Lys Ile Tyr Leu Tyr Thr Leu Asn Asp Asn Ala Arg Ser Ser 1970 1975 1980
- Pro Val Val Ile Asp Ala Ser Thr Ala Ile Asp Ala Pro Ser Asn Leu 1985 1990 1995 2000
- Arg Phe Leu Ala Thr Thr Pro Asn Ser Leu Leu Val Ser Trp Gln Pro 2005 2010 2015
- Pro Arg Ala Arg Ile Thr Gly Tyr Ile Ile Lys Tyr Glu Lys Pro Gly 2020 2025 2030
- Ser Pro Pro Arg Glu Val Val Pro Arg Pro Arg Pro Gly Val Thr Glu 2035 2040 2045

- Ala Thr Ile Thr Gly Leu Glu Pro Gly Thr Glu Tyr Thr Ile Tyr Val 2050 2055 2060
- Ile Ala Leu Lys Asn Asn Gln Lys Ser Glu Pro Leu Ile Gly Arg Lys 2065 2070 2075 2080
- Lys Thr Asp Glu Leu Pro Gln Leu Val Thr Leu Pro His Pro Asn Leu 2085 2090 2095
- His Gly Pro Glu Ile Leu Asp Val Pro Ser Thr Val Gln Lys Thr Pro 2100 2105 2110
- Phe Val Thr His Pro Gly Tyr Asp Thr Gly Asn Gly Ile Gln Leu Pro 2115 2120 2125
- Gly Thr Ser Gly Gln Gln Pro Ser Val Gly Gln Gln Met Ile Phe Glu 2130 2135 2140
- Glu His Gly Phe Arg Arg Thr Thr Pro Pro Thr Thr Ala Thr Pro Ile 2145 2150 2155 2160
- Arg His Arg Pro Arg Pro Tyr Pro Pro Asn Val Gly Gln Glu Ala Leu 2165 2170 2175
- Ser Gln Thr Thr Ile Ser Trp Ala Pro Phe Gln Asp Thr Ser Glu Tyr 2180 2185 2190
- Ile Ile Ser Cys His Pro Val Gly Thr Asp Glu Glu Pro Leu Gln Phe 2195 2200 2205
- Arg Val Pro Gly Thr Ser Thr Ser Ala Thr Leu Thr Gly Leu Thr Arg 2210 2215 2220
- Gly Ala Thr Tyr Asn Ile Ile Val Glu Ala Leu Lys Asp Gln Gln Arg 2225 2230 2235 2240
- His Lys Val Arg Glu Glu Val Val Thr Val Gly Asn Ser Val Asn Glu 2245 2250 2255
- Gly Leu Asn Gln Pro Thr Asp Asp Ser Cys Phe Asp Pro Tyr Thr Val 2260 2265 2270
- Ser His Tyr Ala Val Gly Asp Glu Trp Glu Arg Met Ser Glu Ser Gly 2275 2280 2285
- Phe Lys Leu Cys Gln Cys Leu Gly Phe Gly Ser Gly His Phe Arg 2290 2295 2300
- Cys Asp Ser Ser Arg Trp Cys His Asp Asn Gly Val Asn Tyr Lys Ile 2305 2310 2315 2320
- Gly Glu Lys Trp Asp Arg Gln Gly Glu Asn Gly Gln Met Met Ser Cys 2325 2330 2335

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7	ſhr	Cys	Leu	Gly 2340		Gly	Lys	Gly	G1u 2345		Lys	Cys	Asp	Pro 2350	His O	G1u		
ļ	\la	Thr	Cys 235!		Asp	Asp	Gly	Lys 2360		Tyr	His	Val	Gly 236		Gln	Trp		
(	aln	Lys 2370		Tyr	Leu	Gly	Ala 237		Cys	Ser	Cys	Thr 2380		Phe	Gly	Gly		
	31 n 2385		Gly	Trp	Arg	Cys 2390		Asn	Cys	Arg	Arg 2395		Gly	Gly	Glu	Pro 2400		
S	Ser	Pro	Glu	Gly	Thr 240		Gly	Gln	Ser	Tyr 2410		Gln	Tyr	Ser	Gln 241			
7	yr	His	Gln	Arg 2420		Asn	Thr	Asn	Val 2425		Cys	Pro	Ile	G1u 2430	Cys O	Phe		
M	1et	Pro	Leu 243		Val	Gln	Ala	Asp 2440	Arg )	G1u	Asp	Ser	Arg 2445					
(	(2)	INFO	ORMAT	ΓΙΟΝ	FOR	SEQ	ID N	<b>10:3</b> :	:									
		(i)	4) E) O)	(i) LE 3) T' (c) ST	ENGTI (PE: [RANI	HARACH: 2] nucl DEDNE DGY:	l79 l leic ESS:	ase acio sino	pair 1	<b>`</b> S							•	
		(ix)	(/		AME/H	KEY: ION:		. 1962	2									
		(xi)	SEC	QUENC	CE DE	ESCRI	PTIO	ON: S	SEQ 1	D NO	):3:							
(	TC7	TAGG/	AGC (	CAGC	CCA	CC CT	TAGA	)AAA		Phe						C TGC Cys		54
															GGT Gly		•	102
															GTT Val			150
															TGC Cys 55			198

		TAC Tyr						246
		AAT Asn						294
		GAA Glu						342
		ATG Met 110						390
		ACC Thr						438
		AAG Lys						486
		AAT Asn						534
		GAT Asp						582
		CGT Arg 190						630
		CAG Gln						678
		CCA Pro						726
Pro		AAG Lys						774
		ATG Met						822
		ACT Thr 270						870

					CCC Pro											918
					CCT Pro											966
					ACT Thr											1014
					TGG Trp											1062
					GGA Gly 350											1110
					GAA Glu											1158
					AGT Ser											1206
					CCA Pro											1254
					GAA Glu											1302
					ACA Thr 430											1350
					GAG Glu											1398
					AAA Lys											1446
GGT Gly	CAC His	AAA Lys 475	GAA Glu	GTT Val	ACC Thr	AAA Lys	GAA Glu 480	GTG Val	GTG Val	ACC Thr	TCC Ser	GAA Glu 485	GAT Asp	GGT Gly	TCT Ser	1494

									ACA Thr							1542
									GAT Asp							1590
									GGT G1 <i>y</i> 530							1638
									AGG Arg							1686
									CAT His							1734
									TAC Tyr							1782
									ACA Thr							1830
									GCC Ala 610							1878
		Lys							CGC Arg							1926
	Ser								TCC Ser		TAGA	CTAA	GT T	TAAAT	ATTTC	1979
TGCA	CAGT	GT T	CCCA	TGGC	c cc	TTGC	ATTT	ССТ	тстт	AAC	тстс	TGTT	AC A	CGTC	ATTGA	2039
AACT	ACAC	T T	TTTG	GTCT	G TT	TTTG	TGCT	AGA	CTGT	AAG	TTCC	TTGG	igg g	CAGG	GCCTT	2099
TGTC	TGTC	TC A	тстс	TGTA	T TC	CCAA	ATGC	CTA	ACAG	TAC	AGAG	CCAT	GA C	TCAA	TAAAT	2159
ACAT	GTTA	AA T	GGAT	GAAT	G											2179

#### (2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 643 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Phe Ser Met Arg Ile Val Cys Leu Val Leu Ser Val Val Gly Thr 1 5 10 15

Ala Trp Thr Ala Asp Ser Gly Glu Gly Asp Phe Leu Ala Glu Gly Gly
20 25 30

Gly Val Arg Gly Pro Arg Val Val Glu Arg His Gln Ser Ala Cys Lys 35 40 45

Asp Ser Asp Trp Pro Phe Cys Ser Asp Glu Asp Trp Asn Tyr Lys Cys 50 55 60

Pro Ser Gly Cys Arg Met Lys Gly Leu Ile Asp Glu Val Asn Gln Asp 65 70 75 80

Phe Thr Asn Arg Ile Asn Lys Leu Lys Asn Ser Leu Phe Glu Tyr Gln
85 90 95

Lys Asn Asn Lys Asp Ser His Ser Leu Thr Thr Asn Ile Met Glu Ile 100 105 110

Leu Arg Gly Asp Phe Ser Ser Ala Asn Asn Arg Asp Asn Thr Tyr Asn 115 120 125

Arg Val Ser Glu Asp Leu Arg Ser Arg Ile Glu Val Leu Lys Arg Lys 130 135 140

Val Ile Glu Lys Val Gln His Ile Gln Leu Leu Gln Lys Asn Val Arg 145 150 155 160

Ala Gln Leu Val Asp Met Lys Arg Leu Glu Val Asp Ile Asp Ile Lys 165 170 175

Ile Arg Ser Cys Arg Gly Ser Trp Ser Arg Ala Leu Ala Arg Glu Val 180 185 190

Asp Leu Lys Asp Tyr Glu Asp Gln Gln Lys Gln Leu Glu Gln Val Ile 195 200 205

Ala Lys Asp Leu Leu Pro Ser Arg Asp Arg Gln His Leu Pro Leu Ile 210 215 220

Lys Met Lys Pro Val Pro Asp Leu Val Pro Gly Asn Phe Lys Ser Gln 225 230 235 240

Leu Gln Lys Val Pro Pro Glu Trp Lys Ala Leu Thr Asp Met Pro Gln Met Arg Met Glu Leu Glu Arg Pro Gly Gly Asn Glu Ile Thr Arg Gly Gly Ser Thr Ser Tyr Gly Thr Gly Ser Glu Thr Glu Ser Pro Arg Asn Pro Ser Ser Ala Gly Ser Trp Asn Ser Gly Ser Ser Gly Pro Gly Ser Thr Gly Asn Arg Asn Pro Gly Ser Ser Gly Thr Gly Gly Thr Ala Thr Trp Lys Pro Gly Ser Ser Gly Pro Gly Ser Ala Gly Ser Trp Asn Ser 330 Gly Ser Ser Gly Thr Gly Ser Thr Gly Asn Gln Asn Pro Gly Ser Pro Arg Pro Gly Ser Thr Gly Thr Trp Asn Pro Gly Ser Ser Glu Arg Gly 360 Ser Ala Gly His Trp Thr Ser Glu Ser Ser Val Ser Gly Ser Thr Gly Gln Trp His Ser Glu Ser Gly Ser Phe Arg Pro Asp Ser Pro Gly Ser Gly Asn Ala Arg Pro Asn Asn Pro Asp Trp Gly Thr Phe Glu Glu Val 410 Ser Gly Asn Val Ser Pro Gly Thr Arg Arg Glu Tyr His Thr Glu Lys Leu Val Thr Lys Gly Asp Lys Glu Leu Arg Thr Gly Lys Glu Lys Val Thr Ser Gly Ser Thr Thr Thr Arg Arg Ser Cys Ser Lys Thr Val Thr Lys Thr Val Ile Gly Pro Asp Gly His Lys Glu Val Thr Lys Glu 470 Val Val Thr Ser Glu Asp Gly Ser Asp Cys Pro Glu Ala Met Asp Leu Gly Thr Leu Ser Gly Ile Gly Thr Leu Asp Gly Phe Arg His Arg His Pro Asp Glu Ala Ala Phe Phe Asp Thr Ala Ser Thr Gly Lys Thr Phe 520 525

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Pro	Gly <b>5</b> 30	Phe	Phe	Ser	Pro	Met 535	Leu	Gly	Glu	Phe	Val 540	Ser	Glu	Thr	Glu
Ser 545	Arg	Gly	Ser	Glu	Ser 550	Gly	Ile	Phe	Thr	Asn 555	Thr	Lys	Glu	Ser	Ser 560
Ser	His	His	Pro	Gly 565	Ile	Ala	Glu	Phe	Pro 570	Ser	Arg	Gly	Lys	Ser <b>5</b> 75	Ser
Ser	Tyr	Ser	Lys <b>58</b> 0	Gln	Phe	Thr	Ser	Ser <b>5</b> 85	Thr	Ser	Tyr	Asn	Arg <b>59</b> 0	Gly	Asp
Ser	Thr	Phe 595	Glu	Ser	Lys	Ser	Tyr 600	Lys	Met	Ala	Asp	G1u 605	Ala	Gly	Ser
Glu	Ala 610	Asp	His	Glu	Gly	Thr 615	His	Ser	Thr	Lys	Arg 620	Gly	His	Ala	Lys
Ser 625	Arg	Pro	Val	Arg	Gly 630	Ile	His	Thr	Ser	Pro 635	Leu	Gly	Lys	Pro	Ser 640
Leu	Ser	Pro													

## (2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 4027 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

## (ix) FEATURE:

(A) NAME/KEY: CDS (B) LOCATION: 3..4013

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

		AGT Ser							47
					Leu			GAA Glu	95
		· Val						ATA i Ile	143

	CGA Arg								191
	GTG Val								239
	TCT Ser								287
	CCC Pro								335
	CCA Pro 115								383
	ACG Thr							,	431
	GGA Gly								479
	GTG Val							!	527
	CTG Leu							!	575
	ACA Thr 195							(	623
	ACT Thr							(	671
	ACT Thr							7	719
	TCT Ser							7	767
	AAC Asn							8	315

ACT Thr	GTC <b>V</b> al	AAG Lys	GAT Asp 275	GAC Asp	AAG Lys	GAA Glu	AGT Ser	GTC Val 280	CCT Pro	ATC Ile	TCT Ser	GAT Asp	ACC Thr 285	ATC Ile	ATC Ile	863
													ATA Ile			911
													ACC Thr			959
													CCT Pro			1007
													ACA Thr			1055
													ATT Ile 365			1103
													GTT Val			1151
CCC Pro	ACT Thr 385	GAC Asp	CTG Leu	CGA Arg	TTC Phe	ACC Thr 390	AAC Asn	ATT Ile	GGT Gly	CCA Pro	GAC Asp 395	ACC Thr	ATG Met	CGT Arg	GTC Val	1199
													CTG Leu			1247
													TCA Ser			1295
													GGT Gly 445			1343
													AGC Ser			1391
CTT Leu	AGA Arg 465	GGA Gly	AGA Arg	CAG Gln	AAA Lys	ACA Thr 470	GGT Gly	CTT Leu	GAT Asp	TCC Ser	CCA Pro 475	ACT Thr	GGC Gly	ATT Ile	GAC Asp	1439

				AAC Asn						1487
				TAC Tyr						1535
				GAT Asp						1583
				CCA Pro						1631
				GAA Glu 550						1679
				AGG Arg						1727
				TGG Trp						1775
				GAA Glu						1823
				AAG Lys						1871
_	_			ATC Ile 630			_	_		1919
				AAG Lys						1967
				ATG Met						2015
				CCT Pro						2063
				AAT Asn						2111

													CAG Gln			2159
													GGA Gly			2207
													CCT Pro			2255
													GCT Ala 765			2303
													TCG Ser			2351
													GAA Glu			2399
													ACA Thr			2447
													ATT Ile			2495
													ACT Thr 845			2543
ACA Thr	CCC Pro	ACA Thr 850	AGC Ser	CTG Leu	AGC Ser	GCC Ala	CAG Gln 855	TGG Trp	ACA Thr	CCA Pro	CCC Pro	AAT Asn 860	GTT Val	CAG Gln	CTC Leu	2591
													GGA Gly			2639
													GTA Val			2687
CTT Leu	ATG Met	GTG Val	GCC Ala	ACC Thr 900	AAA Lys	TAT Tyr	GAA Glu	GTG Val	AGT Ser 905	GTC Val	TAT Tyr	GCT Ala	CTT Leu	AAG Lys 910	GAC Asp	2735

TTG Leu															2783
AAT Asn															2831
ACA Thr 945															2879
GAG Glu															2927
GAA Glu															2975
TCT Ser								Arg					Ser		3023
GGA Gly		Thr					Pro					Pro			3071
GGA Gly 1025	Ser					Ser					Ser				3119
AAC Asn )					Arg					Gly					3167
AGC Ser				Gly					Trp					Ser	3215
TCT Ser			Thr					Ser					Phe		3263
GAT Asp		Pro					Ala					Pro			3311
ACA Thr 1105	Phe					Gly					Gly				3359
TAC Tyr					Leu					Asp					3407

		AAG GTC Lys Val 1140				Thr					Arg	3455
		ACC GTT Thr Val 5			Val					Gly		3503
AAA GAA Lys Glu	GTT ACC Val Thr 1170	AAA GAA Lys Glu	GTG GT Val Va 11	Thr	TCC Ser	GAA Glu	GAT Asp	GGT Gly 1180	Ser	GAC Asp	TGT Cys	3551
	Ala Met	GAT TTA Asp Leu						Gly				3599
		AGG CAC Arg His 120	Pro As				Phe					3647
		ACA TTC Thr Phe 1220				Ser					Glu	3695
		ACT GAG Thr Glu 5			Ser					Phe		3743
AAT ACA Asn Thr	AAG GAA Lys Glu 1250	TCC AGT Ser Ser	TCT CA Ser Hi 12	s His	CCT Pro	GGG Gly	ATA Ile	GCT Ala 126	Glu	TTC Phe	CCT Pro	3791
	Gly Lys	TCT TCA Ser Ser						Thr				3839
AGT TAC Ser Tyr 1280	AAC AGA Asn Arg	GGA GAC Gly Asp 128	Ser Th	A TTT r Phe	GAA Glu	AGC Ser 129	Lys	AGC Ser	TAT Tyr	AAA Lys	ATG Met 1295	3887
		GGA AGT Gly Ser 1300				Glu					Thr	3935
		GCT AAA Ala Lys 5			Val					Thr		3983
		CCT TCC Pro Ser		r Pro		ACTA	AGT	TAAA	TAT			4027

- (2) INFORMATION FOR SEQ ID NO:6:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 1336 amino acids
    - (B) TYPE: amino acid (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:
- Met Ala Val Ser His Gly Arg Glu Ser Lys Pro Leu Thr Ala Gln Gln 1 5 10 15
- Thr Thr Lys Leu Asp Ala Pro Thr Asn Leu Gln Phe Val Asn Glu Thr 20 25 30
- Asp Ser Thr Val Leu Val Arg Trp Thr Pro Pro Arg Ala Gln Ile Thr 35 40 45
- Gly Tyr Arg Leu Thr Val Gly Leu Thr Arg Arg Gly Gln Pro Arg Gln 50 55 60
- Tyr Asn Val Gly Pro Ser Val Ser Lys Tyr Pro Leu Arg Asn Leu Gln
  65 70 75 80
- Pro Ala Ser Glu Tyr Thr Val Ser Leu Val Ala Ile Lys Gly Asn Gln 85 90 95
- Glu Ser Pro Lys Ala Thr Gly Val Phe Thr Thr Leu Gln Pro Gly Ser 100 105 110
- Ser Ile Pro Pro Tyr Asn Thr Glu Val Thr Glu Thr Thr Ile Val Ile 115 120 125
- Thr Trp Thr Pro Ala Pro Arg Ile Gly Phe Lys Leu Gly Val Arg Pro 130 135 140
- Ser Gln Gly Gly Glu Ala Pro Arg Glu Val Thr Ser Asp Ser Gly Ser 145 150 155 160
- Ile Val Val Ser Gly Leu Thr Pro Gly Val Glu Tyr Val Tyr Thr Ile 165 170 175
- Gln Val Leu Arg Asp Gly Gln Glu Arg Asp Ala Pro Ile Val Asn Lys 180 185 190
- Val Val Thr Pro Leu Ser Pro Pro Thr Asn Leu His Leu Glu Ala Asn 195 200 205
- Pro Asp Thr Gly Val Leu Thr Val Ser Trp Glu Arg Ser Thr Thr Pro 210 215 220
- Asp Ile Thr Gly Tyr Arg Ile Thr Thr Thr Pro Thr Asn Gly Gln Gln 225 230 235 240

Gly	Asn	Ser	Leu	G1u 245	Glu	Val	Val	His	Ala 250	Asp	Gln	Ser	Ser	Cys <b>255</b>	Thr
Phe	Asp	Asn	Leu 260	Ser	Pro	Gly	Leu	G1u 265	Tyr	Asn	Val	Ser	Val 270	Tyr	Thr
Val	Lys	Asp 275	Asp	Lys	Glu	Ser	Val 280	Pro	Ile	Ser	Asp	Thr 285	Ile	Ile	Pro
Glu	Va1 290	Pro	Gln	Leu	Thr	Asp 295	Leu	Ser	Phe	Val	Asp 300	Ile	Thr	Asp	Ser
Ser 305	Ile	Gly	Leu	Arg	Trp 310	Thr	Pro	Leu	Asn	Ser 315	Ser	Thr	Ile	Ile	Gly 320
Tyr	Arg	Ile	Thr	Val 325	Val	Ala	Ala	Gly	G1u 330	Gly	Ile	Pro	Ile	Phe 335	Glu
Asp	Phe	Va1	Tyr 340	Ser	Ser	Val	Gly	Tyr 345	Tyr	Thr	Val	Thr	Gly 350	Leu	Glu
Pro	Gly	Ile 355	Asp	Tyr	Asp	Ile	Ser 360	Val	Ile	Thr	Leu	Ile 365	Asn	Gly	Gly
Glu	Ser 370	Ala	Pro	Thr	Thr	Leu 375	Thr	Gln	Gln	Thr	Ala 380	Val	Pro	Pro	Pro
Thr 385	Asp	Leu	Arg	Phe	Thr 390	Asn	Ile	Gly	Pro	Asp 395	Thr	Met	Arg	Val	Thr 400
Trp	Ala	Pro	Pro	Pro 405	Ser	Ile	Asp	Leu	Thr 410	Asn	Phe	Leu	Val	Arg 415	Tyr
Ser	Pro	Val	Lys 420	Asn	Glu	Glu	Asp	Val 425	Ala	Glu	Leu	Ser	Ile 430	Ser	Pro
Ser	Asp	Asn 435	Ala	Val	Val	Leu	Thr 440	Asn	Leu	Leu	Pro	Gly 445	Thr	Glu	Tyr
Val	Val 450	Ser	Val	Ser	Ser	Val 455	Tyr	Glu	Gln	His	G1u 460	Ser	Thr	Pro	Leu
Arg <b>46</b> 5	Gly	Arg	Gln	Lys	Thr 470	Gly	Leu	Asp	Ser	Pro 475	Thr	Gly	Ile	Asp	Phe 480
Ser	Asp	Ile	Thr	A1 a 485	Asn	Ser	Phe	Thr	Val 490	His	Trp	Ile	Ala	Pro 495	Arg
Ala	Thr	Ile	Thr 500	Gly	Tyr	Arg	Ile	Arg 505	His	His	Pro	Glu	His 510	Phe	Ser
Gly	Arg	Pro 515	Arg	Glu	Asp	Arg	Val 520	Pro	His	Ser	Arg	Asn 525	Ser	Ile	Thr

Leu	Thr 530	Asn	Leu	Thr	Pro	Gly <b>5</b> 35	Thr	Glu	Tyr	Val	Val 540	Ser	Ile	Val	Ala
Leu 545	Asn	Gly	Arg	Glu	G1u 550	Ser	Pro	Leu	Leu	Ile 555	Gly	Gln	Gln	Ser	Th: 560
Val	Ser	Asp	Val	Pro 565	Arg	Asp	Leu	Glu	Val 570	Val	Ala	Ala	Thr	Pro <b>5</b> 75	Thi
Ser	Leu	Leu	Ile 580	Ser	Trp	Asp	Ala	Pro 585	Ala	Val	Thr	Val	Arg 590	Tyr	Тун
Arg	Ile	Thr 595	Tyr	Gly	Glu	Thr	Gly 600	Gly	Asn	Ser	Pro	Val 605	Gln	Glu	Phe
Thr	Val 610	Pro	Gly	Ser	Lys	Ser 615	Thr	Ala	Thr	Ile	Ser 620	Gly	Leu	Lys	Pro
Gly 625	Val	Asp	Tyr	Thr	Ile 630	Thr	Val	Tyr	Ala	Val 635	Thr	Gly	Arg	Gly	Asp 640
Ser	Pro	Ala	Ser	Ser 645	Lys	Pro	Ile	Ser	Ile 650	Asn	Tyr	Arg	Thr	G1u 655	Πe
Asp	Lys	Pro	Ser 660	Gln	Met	Gln	Val	Thr 665	Asp	Val	Gln	Asp	Asn 670	Ser	Ιle
Ser	Val	Lys 675	Trp	Leu	Pro	Ser	Ser 680	Ser	Pro	Val	Thr	Gly 685	Tyr	Arg	Val
Thr	Thr 690	Thr	Pro	Lys	Asn	Gly 695	Pro	Gly	Pro	Thr	Lys <b>70</b> 0	Thr	Lys	Thr	Ala
Gly 705	Pro	Asp	Gln	Thr	Glu 710	Met	Thr	Ile	Glu	Gly 715	Leu	Gln	Pro	Thr	Val 720
Glu	Tyr	Val	Val	Ser 725	Val	Tyr	Ala	Gln	Asn 730	Pro	Ser	Gly	Glu	Ser 735	Gln
Pro	Leu	Val	Gln 740	Thr	Ala	Val	Thr	Asn 745	Ile	Asp	Arg	Pro	Lys 750	Gly	Leu
Ala	Phe	Thr 755	Asp	Val	Asp	Val	Asp 760	Ser	Ile	Lys	Ile	A1a 765	Trp	Glu	Ser
Pro	G1n 770	Gly	Gln	Val	Ser	Arg 775	Tyr	Arg	Val	Thr	Tyr 780	Ser	Ser	Pro	Glu
Asp 785	Gly	Ile	His	Glu	Leu 790	Phe	Pro	Ala	Pro	Asp 795	Gly	Glu	Glu	Asp	Thr 800
Ala	Glu	Leu	Gln	Gly 805	Leu	Arg	Pro	Gly	Ser 810	Glu	Tyr	Thr	Val	Ser 815	Val

- Val Ala Leu His Asp Asp Met Glu Ser Gln Pro Leu Ile Gly Thr Gln 820 825 830
- Ser Thr Ala Ile Pro Ala Pro Thr Asp Leu Lys Phe Thr Gln Val Thr 835 840 845
- Pro Thr Ser Leu Ser Ala Gln Trp Thr Pro Pro Asn Val Gln Leu Thr 850 855 860
- Gly Tyr Arg Val Arg Val Thr Pro Lys Glu Lys Thr Gly Pro Met Lys 865 870 875 880
- Glu Ile Asn Leu Ala Pro Asp Ser Ser Ser Val Val Ser Gly Leu 885 890 895
- Met Val Ala Thr Lys Tyr Glu Val Ser Val Tyr Ala Leu Lys Asp Thr 900 905 910
- Leu Thr Ser Arg Pro Ala Gln Gly Val Val Thr Thr Leu Glu Gly Gly 915 920 925
- Asn Phe Lys Ser Gln Leu Gln Lys Val Pro Pro Glu Trp Lys Ala Leu 930 935 940
- Thr Asp Met Pro Gln Met Arg Met Glu Leu Glu Arg Pro Gly Gly Asn 955 960
- Glu Ile Thr Arg Gly Gly Ser Thr Ser Tyr Gly Thr Gly Ser Glu Thr 965 970 975
- Glu Ser Pro Arg Asn Pro Ser Ser Ala Gly Ser Trp Asn Ser Gly Ser 980 985 990
- Ser Gly Pro Gly Ser Thr Gly Asn Arg Asn Pro Gly Ser Ser Gly Thr 995 1000 1005
- Gly Gly Thr Ala Thr Trp Lys Pro Gly Ser Ser Gly Pro Gly Ser Ala 1010 1015 1020
- Gly Ser Trp Asn Ser Gly Ser Ser Gly Thr Gly Ser Thr Gly Asn Gln 1025 1030 1035 1040
- Asn Pro Gly Ser Pro Arg Pro Gly Ser Thr Gly Thr Trp Asn Pro Gly 1045 1050 1055
- Ser Ser Glu Arg Gly Ser Ala Gly His Trp Thr Ser Glu Ser Ser Val 1060 1065 1070
- Ser Gly Ser Thr Gly Gln Trp His Ser Glu Ser Gly Ser Phe Arg Pro 1075 1080 1085
- Asp Ser Pro Gly Ser Gly Asn Ala Arg Pro Asn Asn Pro Asp Trp Gly 1090 1095 1100

- Thr Phe Glu Glu Val Ser Gly Asn Val Ser Pro Gly Thr Arg Arg Glu
- Tyr His Thr Glu Lys Leu Val Thr Lys Gly Asp Lys Glu Leu Arg Thr
- Gly Lys Glu Lys Val Thr Ser Gly Ser Thr Thr Thr Arg Arg Ser
- Cys Ser Lys Thr Val Thr Lys Thr Val Ile Gly Pro Asp Gly His Lys
- Glu Val Thr Lys Glu Val Val Thr Ser Glu Asp Gly Ser Asp Cys Pro
- Glu Ala Met Asp Leu Gly Thr Leu Ser Gly Ile Gly Thr Leu Asp Gly
- Phe Arg His Arg His Pro Asp Glu Ala Ala Phe Phe Asp Thr Ala Ser
- Thr Gly Lys Thr Phe Pro Gly Phe Phe Ser Pro Met Leu Gly Glu Phe
- Val Ser Glu Thr Glu Ser Arg Gly Ser Glu Ser Gly Ile Phe Thr Asn
- Thr Lys Glu Ser Ser Ser His His Pro Gly Ile Ala Glu Phe Pro Ser
- Arg Gly Lys Ser Ser Ser Tyr Ser Lys Gln Phe Thr Ser Ser Thr Ser
- Tyr Asn Arg Gly Asp Ser Thr Phe Glu Ser Lys Ser Tyr Lys Met Ala
- Asp Glu Ala Gly Ser Glu Ala Asp His Glu Gly Thr His Ser Thr Lys
- Arg Gly His Ala Lys Ser Arg Pro Val Arg Gly Ile His Thr Ser Pro
- Leu Gly Lys Pro Ser Leu Ser Pro

(2) INFORMATION FOR SEQ ID NO:7:

<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 27 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC1551	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:	
GATCCCCGGG GAGCTCCTCG AGGCATG	27
(2) INFORMATION FOR SEQ ID NO:8:	
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 19 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC1552	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:	
CCTCGAGGAG CTCCCCGGG	19
(2) INFORMATION FOR SEQ ID NO:9:	
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 20 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC2052	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:	
AATTCACCAT GGCAGTGAGT	20

(2) INFORMATION FOR SEQ ID NO:10:	
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 20 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC2053	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:	
CATGACTCAC TGCCATGGTG	20
(2) INFORMATION FOR SEQ ID NO:11:	
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 18 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC2491	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:	
CTAGATTAGA ATGGGGCC	18
(2) INFORMATION FOR SEQ ID NO:12:	
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 10 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC2493	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:	
CCATTCTAAT	10

(2) INFORMATION FOR SEQ ID NO:13:	
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 88 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC3521	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:	
TCGACTTAAG GACACTTTGA CAAGCAGACC AGCTCAGGGT GTTGTCACCA CTCTGGAGGG	60
AGGAAATTTT AAGAGCCAGC TTCAGAAG	88
(2) INFORMATION FOR SEQ ID NO:14:	
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 88 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
(vii) IMMEDIATE SOURCE: (B) CLONE: ZC3522	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:	
GTACCTTCTG AAGCTGGCTC TTAAAATTTC CTCCCTCCAG AGTGGTGACA ACACCCTGAG	60
CTGGTCTGCT TGTCAAAGTG TCCTTAAG	88

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WO 94/16085 PCT/US93/12687

#### I Claim:

1. A hybrid protein comprising a tissue-binding domain from a first protein covalently linked to a cross-linking domain from a second protein.

- 2. A hybrid protein according to claim 1 wherein the tissue-binding domain of the first protein is a heparin binding domain of thrombospondin, a heparin binding domain of fibronectin, a collagen binding domain of fibronectin or a cell binding domain of fibronectin.
- 3. A hybrid protein according to claim 1 wherein the tissue-binding domain of the first protein comprises the amino acid sequence of Sequence ID No. 6 from Alanine, amino acid 2 to Glutamic acid, amino acid number 926.
- 4. A hybrid protein according to claim 1 wherein the cross-linking domain of the second protein comprises the carboxy-terminal 103 amino acids of loricrin; the ten amino acid repeat beginning with glutamine, amino acid number 496 of involucrin; or the 400 amino-terminal amino acids of the fibrinogen  $\alpha$  chain.
- 5. A hybrid protein according to claim 1 wherein the cross-linking domain of the second protein comprises the amino acid sequence of Sequence ID No. 6 from Glycine, amino acid number 928 to Proline, amino acid number 1336.
- 6. A hybrid protein according to claim 1 comprising the amino acid sequence of Sequence ID Number 6 from alanine, amino acid number 2 to Proline, amino acid number 1336.
- 7. An isolated DNA molecule encoding a hybrid protein comprising a first DNA segment encoding a tissue-binding domain from a first protein joined to a second DNA segment encoding a cross-linking domain from a second protein.

8. A DNA molecule according to claim 7 wherein the first DNA segment encodes a heparin binding domain of thrombospondin, a heparin binding domain of fibronectin, a collagen binding domain of fibronectin, a collagen binding domain of fibronectin or a cell binding domain of fibronectin.

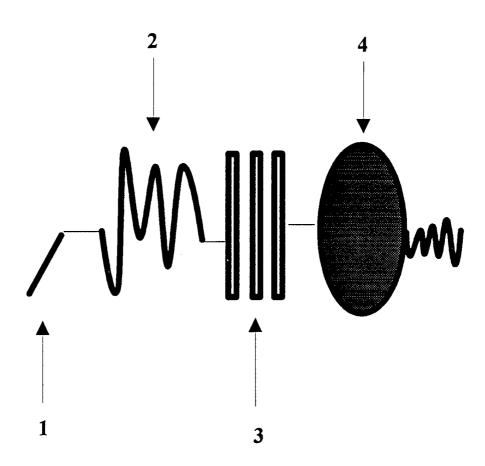
- 9. A DNA molecule according to claim 7 wherein the first DNA segment comprises the nucleotide sequence of Sequence ID No. 5 from nucleotide 3 to nucleotide 2780.
- 10. A DNA molecule according to claim 7 wherein the first DNA segment encodes the amino acid sequence of Sequence ID No. 6 from methionine, amino acid number 1 to glutamic acid, amino acid number 926.
- 11. A DNA molecule according to claim 7 wherein the second DNA segment encodes the carboxy-terminal 103 amino acids of loricrin; the ten amino acid repeat beginning with glutamine, amino acid number 496 of involucrin; or the 400 amino-terminal amino acids of the fibrinogen  $\alpha$  chain.
- 12. A DNA molecule according to claim 7 wherein the second DNA segment comprises the nucleotide sequence of Sequence ID No. 5 from nucleotide 2784 to nucleotide 4013.
- 13. A DNA molecule according to claim 7 wherein the second DNA segment encodes the amino acid sequence of Sequence ID No. 6 from glycine, amino acid number 928 to proline, amino acid number 1336.
- 14. A DNA molecule according to claim 7 wherein the DNA molecule encodes the amino acid sequence of Sequence ID Number 6 from Methionine, amino acid number 1 to Proline, amino acid number 1336.

15. A DNA molecule according to claim 7 wherein the DNA molecule comprises the nucleotide sequence of Sequence ID Number 5 from nucleotide 3 to nucleotide 4013.

- 16. A DNA construct comprising a DNA molecule encoding a hybrid protein, wherein said DNA molecule comprises a first DNA segment encoding a tissue-binding domain from a first protein joined to a second DNA segment encoding a cross-linking domain from a second protein, and wherein said DNA molecule is operably linked to other DNA segments required for the expression of the DNA molecule.
- 17. A DNA construct according to claim 16 wherein the first DNA segment encodes a heparin binding domain of thrombospondin, a heparin binding domain of fibronectin, a collagen binding domain of fibronectin or a cell binding domain of fibronectin.
- 18. A DNA construct according to claim 16 wherein the first DNA segment comprises the nucleotide sequence of Sequence ID No. 5 from nucleotide 3 to nucleotide 2780.
- 19. A DNA construct according to claim 16 wherein the first DNA segment encodes the amino acid sequence of Sequence ID No. 6 from methionine, amino acid 1 to Glutamic acid, amino acid number 926.
- 20. A DNA construct according to claim 16 wherein the second DNA segment encodes the carboxy-terminal 103 amino acids of loricrin; the ten amino acid repeat beginning with glutamine, amino acid number 496 of involucrin; or the 400 amino-terminal amino acids of the fibrinogen  $\alpha$  chain.
- 21. A DNA construct according to claim 16 wherein the second DNA segment comprises the nucleotide sequence of Sequence ID No. 5 from nucleotide 2784 to nucleotide 4013.

- 22. A DNA construct according to claim 16 wherein the second DNA segment encodes the amino acid sequence of Sequence ID No. 6 from glycine, amino acid number 928 to proline, amino acid number 1336.
- 23. A DNA construct according to claim 16 wherein the DNA molecule comprises the nucleotide sequence of Sequence ID Number 5 from nucleotide 1 to nucleotide 4013.
- 24. A DNA construct according to claim 16 wherein the DNA molecule encodes the amino acid sequence of Sequence ID Number 6 from Methionine, amino acid number 1 to Proline, amino acid number 1336.
- 25. A host cell containing a DNA construct according to claim 16.
- 26. A method for producing a hybrid protein comprising culturing a host cell according to claim 25 under conditions promoting the expression of the first DNA segment.

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# FIGURE 1

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Hybrid+FXIII+Thrombin

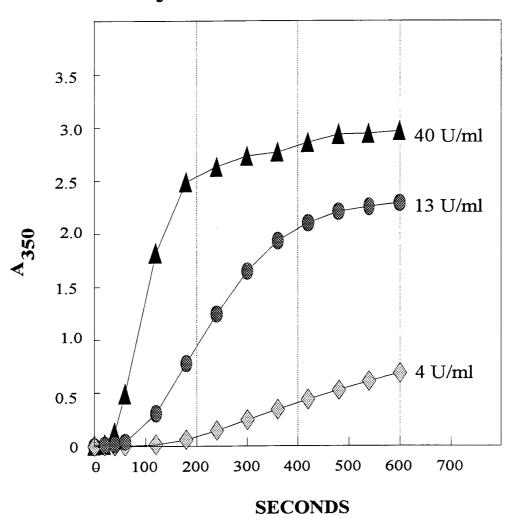


FIGURE 2

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## Hybrid+FXIIIa

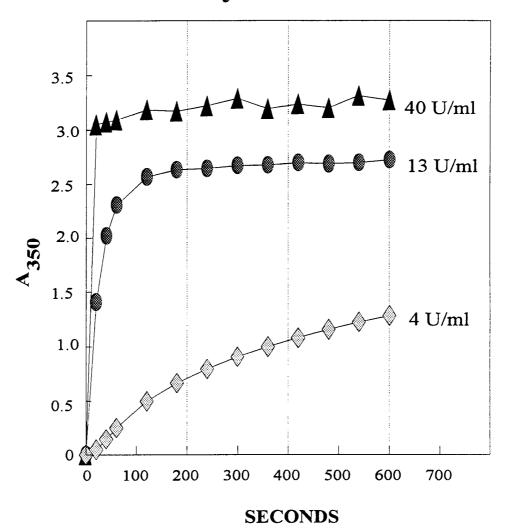


FIGURE 3

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### **FXIII+Thrombin**

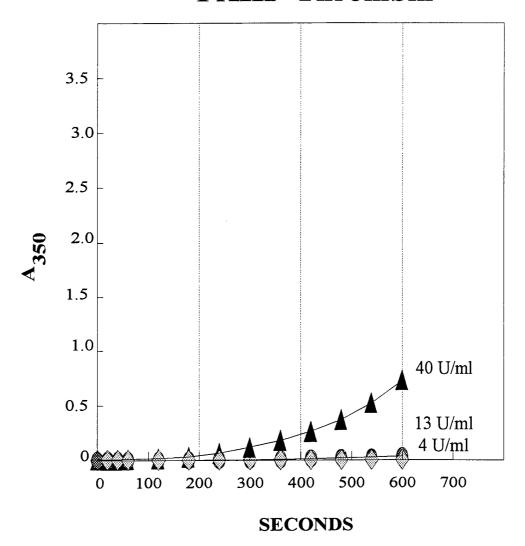
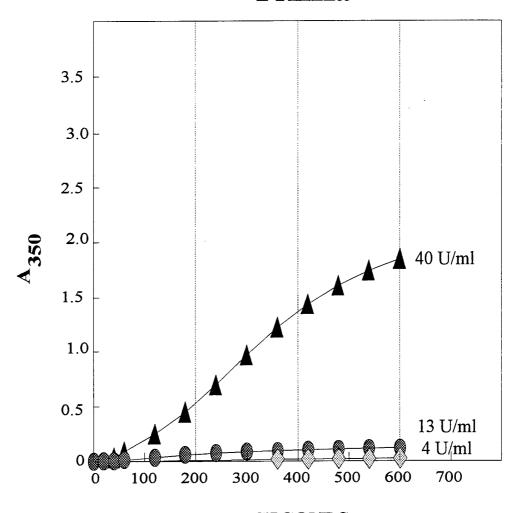


FIGURE 4

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**SECONDS** 

# FIGURE 5