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(54) Title: NUCLEIC ACID AND CORRESPONDING PROTEIN ENTITLED 85P1B3 USEFUL IN TREATMENT AND DETECTION OF CANCER

(57) Abstract: A novel gene (designated 85P1B3) and its encoded protein are described. While 85P1B3 exhibits tissue specific expression in normal adult tissue, it is aberrantly expressed in multiple cancers including set forth in Table 1. Consequently, 85P1B3 provides a diagnostic and/or therapeutic target for cancers. The 85P1B3 gene or fragment thereof, or its encoded protein or a fragment thereof, can be used to elicit an immune response.

NUCLEIC ACID AND CORRESPONDING PROTEIN ENTITLED 85P1B3 USEFUL IN TREATMENT AND DETECTION OF CANCER

This application claims the benefit of United States provisional patent application serial number 60/228,432, filed August 28, 2000, the entire contents of which are hereby incorporated herein by reference.

FIELD OF THE INVENTION

The invention described herein relates to a novel gene and its encoded protein, termed 85P1B3, and to diagnostic and therapeutic methods and compositions useful in the management of various cancers that express 85P1B3.

BACKGROUND OF THE INVENTION

Cancer is the second leading cause of human death next to coronary disease. Worldwide, millions of people die from cancer every year. In the United States alone, as reported by the American Cancer Society, cancer causes the death of well over a half-million people annually, with over 1.2 million new cases diagnosed per year. While deaths from heart disease have been declining significantly, those resulting from cancer generally are on the rise. In the early part of the next century, cancer is predicted to become the leading cause of death.

Worldwide, several cancers stand out as the leading killers. In particular, carcinomas of the lung, prostate, breast, colon, pancreas, and ovary represent the primary causes of cancer death. These and virtually all other carcinomas share a common lethal feature. With very few exceptions, metastatic disease from a carcinoma is fatal. Moreover, even for those cancer patients who initially survive their primary cancers, common experience has shown that their lives are dramatically altered. Many cancer patients experience strong anxieties driven by the awareness of the potential for recurrence or treatment failure. Many cancer patients experience physical debilitations following treatment. Furthermore, many cancer patients experience a recurrence.

Worldwide, prostate cancer is the fourth most prevalent cancer in men. In North America and Northern Europe, it is by far the most common cancer in males and is the second leading cause of cancer death in men. In the United States alone, well over 30,000 men die annually of this disease - second only to lung cancer. Despite the magnitude of these figures, there is still no effective treatment for metastatic prostate cancer. Surgical prostatectomy, radiation therapy, hormone ablation therapy, surgical castration

and chemotherapy continue to be the main treatment modalities. Unfortunately, these treatments are ineffective for many and are often associated with undesirable consequences.

On the diagnostic front, the lack of a prostate tumor marker that can accurately detect early-stage, localized tumors remains a significant limitation in the diagnosis and management of this disease. Although the serum prostate specific antigen (PSA) assay has been a very useful tool, however its specificity and general utility is widely regarded as lacking in several important respects.

Progress in identifying additional specific markers for prostate cancer has been improved by the generation of prostate cancer xenografts that can recapitulate different stages of the disease in mice. The LAPC (Los Angeles Prostate Cancer) xenografts are prostate cancer xenografts that have survived passage in severe combined immune deficient (SCID) mice and have exhibited the capacity to mimic the transition from androgen dependence to androgen independence (Klein et al., 1997, Nat. Med. 3:402). More recently identified prostate cancer markers include PCTA-1 (Su et al., 1996, Proc. Natl. Acad. Sci. USA 93: 7252), prostate-specific membrane (PSM) antigen (Pinto et al., Clin Cancer Res 1996 Sep 2 (9): 1445-51), STEAP (Hubert, et al., Proc Natl Acad Sci U S A. 1999 Dec 7; 96(25): 14523-8) and prostate stem cell antigen (PSCA) (Reiter et al., 1998, Proc. Natl. Acad. Sci. USA 95: 1735).

While previously identified markers such as PSA, PSM, PCTA and PSCA have facilitated efforts to diagnose and treat prostate cancer, there is need for the identification of additional markers and therapeutic targets for prostate and related cancers in order to further improve diagnosis and therapy.

Renal cell carcinoma (RCC) accounts for approximately 3 percent of adult malignancies. Once adenomas reach a diameter of 2 to 3 cm, malignant potential exists. In the adult, the two principal malignant renal tumors are renal cell adenocarcinoma and transitional cell carcinoma of the renal pelvis or ureter. The incidence of renal cell adenocarcinoma is estimated at more than 29,000 cases in the United States, and more than 11,600 patients died of this disease in 1998. Transitional cell carcinoma is less frequent, with an incidence of approximately 500 cases per year in the United States.

Surgery has been the primary therapy for renal cell adenocarcinoma for many decades. Until recently, metastatic disease has been refractory to any systemic therapy. With recent developments in systemic therapies, particularly immunotherapies, metastatic renal cell carcinoma may be approached aggressively in appropriate patients with a possibility of durable responses. Nevertheless, there is a remaining need for effective therapies for these patients.

Of all new cases of cancer in the United States, bladder cancer represents approximately 5 percent in men (fifth most common neoplasm) and 3 percent in women (eighth most common neoplasm). The incidence is increasing slowly, concurrent with an increasing older population. In 1998, there was an estimated 54,500 cases, including 39,500 in men and 15,000 in women. The age-adjusted incidence in the United States is 32 per 100,000 for men and 8 per 100,000 in women. The historic male/female ratio of 3:1

may be decreasing related to smoking patterns in women. There were an estimated 11,000 deaths from bladder cancer in 1998 (7,800 in men and 3,900 in women). Bladder cancer incidence and mortality strongly increase with age and will be an increasing problem as the population becomes more elderly.

Most bladder cancers recur in the bladder. Bladder cancer is managed with a combination of transurethral resection of the bladder (TUR) and intravesical chemotherapy or immunotherapy. The multifocal and recurrent nature of bladder cancer points out the limitations of TUR. Most muscle-invasive cancers are not cured by TUR alone. Radical cystectomy and urinary diversion is the most effective means to eliminate the cancer but carry an undeniable impact on urinary and sexual function. There continues to be a significant need for treatment modalities that are beneficial for bladder cancer patients.

An estimated 130,200 cases of colorectal cancer occurred in 2000 in the United States, including 93,800 cases of colon cancer and 36,400 of rectal cancer. Colorectal cancers are the third most common cancers in men and women. Incidence rates declined significantly during 1992-1996 (-2.1% per year). Research suggests that these declines have been due to increased screening and polyp removal, preventing progression of polyps to invasive cancers. There were an estimated 56,300 deaths (47,700 from colon cancer, 8,600 from rectal cancer) in 2000, accounting for about 11% of all U.S. cancer deaths.

At present, surgery is the most common form of therapy for colorectal cancer, and for cancers that have not spread, it is frequently curative. Chemotherapy, or chemotherapy plus radiation is given before or after surgery to most patients whose cancer has deeply perforated the bowel wall or has spread to the lymph nodes. A permanent colostomy (creation of an abdominal opening for elimination of body wastes) is occasionally needed for colon cancer and is infrequently required for rectal cancer. There continues to be a need for effective diagnostic and treatment modalities for colorectal cancer.

There were an estimated 164,100 new cases of lung and bronchial cancer in 2000, accounting for 14% of all U.S. cancer diagnoses. The incidence rate of lung and bronchial cancer is declining significantly in men, from a high of 86.5 per 100,000 in 1984 to 70.0 in 1996. In the 1990s, the rate of increase among women began to slow. In 1996, the incidence rate in women was 42.3 per 100,000.

Lung and bronchial cancer caused an estimated 156,900 deaths in 2000, accounting for 28% of all cancer deaths. During 1992–1996, mortality from lung cancer declined significantly among men (-1.7% per year) while rates for women were still significantly increasing (0.9% per year). Since 1987, more women have died each year of lung cancer than breast cancer, which, for over 40 years, was the major cause of cancer death in women. Decreasing lung cancer incidence and mortality rates most likely resulted from decreased smoking rates over the previous 30 years; however, decreasing smoking patterns among women lag behind those of men. Of concern, although the declines in adult tobacco use have slowed, tobacco use in youth is increasing again.

Treatment options for lung and bronchial cancer are determined by the type and stage of the cancer and include surgery, radiation therapy, and chemotherapy. For many localized cancers, surgery is usually the treatment of choice. Because the disease has usually spread by the time it is discovered, radiation therapy and chemotherapy are often needed in combination with surgery. Chemotherapy alone or combined with radiation is the treatment of choice for small cell lung cancer; on this regimen, a large percentage of patients experience remission, which in some cases is long lasting. There is however, an ongoing need for effective treatment and diagnostic approaches for lung and bronchial cancers.

An estimated 182,800 new invasive cases of breast cancer were expected to occur among women in the United States during 2000. Additionally, about 1,400 new cases of breast cancer were expected to be diagnosed in men in 2000. After increasing about 4% per year in the 1980s, breast cancer incidence rates in women have leveled off in the 1990s to about 110.6 cases per 100,000.

In the U.S. alone, there were an estimated 41,200 deaths (40,800 women, 400 men) in 2000 due to breast cancer. Breast cancer ranks second among cancer deaths in women. According to the most recent data, mortality rates declined significantly during 1992–1996 with the largest decreases in younger women, both white and black. These decreases were probably the result of earlier detection and improved treatment.

Taking into account the medical circumstances and the patient's preferences, treatment of breast cancer may involve lumpectomy (local removal of the tumor) and removal of the lymph nodes under the arm; mastectomy (surgical removal of the breast) and removal of the lymph nodes under the arm; radiation therapy; chemotherapy; or hormone therapy. Often, two or more methods are used in combination. Numerous studies have shown that, for early stage disease, long-term survival rates after lumpectomy plus radiotherapy are similar to survival rates after modified radical mastectomy. Significant advances in reconstruction techniques provide several options for breast reconstruction after mastectomy. Recently, such reconstruction has been done at the same time as the mastectomy.

Local excision of ductal carcinoma in situ (DCIS) with adequate amounts of surrounding normal breast tissue may prevent the local recurrence of the DCIS. Radiation to the breast and/or tamoxifen may reduce the chance of DCIS occurring in the remaining breast tissue. This is important because DCIS, if left untreated, may develop into invasive breast cancer. Nevertheless, there are serious side effects or sequelae to these treatments. There is, therefore, a need for efficacious breast cancer treatments.

There were an estimated 23,100 new cases of ovarian cancer in the United States in 2000. It accounts for 4% of all cancers among women and ranks second among gynecologic cancers. During 1992–1996, ovarian cancer incidence rates were significantly declining. Consequent to ovarian cancer, there were an estimated 14,000 deaths in 2000. Ovarian cancer causes more deaths than any other cancer of the female reproductive system.

Surgery, radiation therapy, and chemotherapy are treatment options for ovarian cancer. Surgery usually includes the removal of one or both ovaries, the fallopian tubes (salpingo-oophorectomy), and the uterus (hysterectomy). In some very early tumors, only the involved ovary will be removed, especially in young women who wish to have children. In advanced disease, an attempt is made to remove all intra-abdominal disease to enhance the effect of chemotherapy. There continues to be an important need for effective treatment options for ovarian cancer.

There were an estimated 28,300 new cases of pancreatic cancer in the United States in 2000. Over the past 20 years, rates of pancreatic cancer have declined in men. Rates among women have remained approximately constant but may be beginning to decline. Pancreatic cancer caused an estimated 28,200 deaths in 2000 in the United States. Over the past 20 years, there has been a slight but significant decrease in mortality rates among men (about –0.9% per year) while rates have increased slightly among women.

Surgery, radiation therapy, and chemotherapy are treatment options for pancreatic cancer. These treatment options can extend survival and/or relieve symptoms in many patients but are not likely to produce a cure for most. There is a significant need for additional therapeutic and diagnostic options for pancreatic cancer.

SUMMARY OF THE INVENTION

The present invention relates to a novel gene, designated 85P1B3, that is over-expressed in multiple cancers listed in Table I. Northern blot expression analysis of 85P1B3 gene expression in normal tissues shows a restricted expression pattern in adult tissues. The nucleotide (Figure 2) and amino acid (Figure 2, and Figure 3) sequences of 85P1B3 are provided. The tissue-related profile of 85P1B3 in normal adult tissues, combined with the over-expression observed in prostate and other tumors, shows that 85P1B3 is aberrantly over-expressed in at least some cancers, and thus serves as a useful diagnostic and/or therapeutic target for cancers of the tissues such as those listed in Table I.

The invention provides polynucleotides corresponding or complementary to all or part of the 85P1B3 genes, mRNAs, and/or coding sequences, preferably in isolated form, including polynucleotides encoding 85P1B3-related proteins and fragments of 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, or more than 25 contiguous amino acids; at least 30, 35, 40, 45, 50, 55, 60, 65, 70, 80, 85, 90, 95, 100 or more than 100 contiguous amino acids of a 85P1B3-related protein, as well as the peptides/proteins themselves; DNA, RNA, DNA/RNA hybrids, and related molecules, polynucleotides or oligonucleotides complementary or having at least a 90% homology to the 85P1B3 genes or mRNA sequences or parts thereof, and polynucleotides or oligonucleotides that hybridize to the 85P1B3 genes, mRNAs, or to 85P1B3-encoding polynucleotides. Also provided are means for isolating cDNAs and the genes encoding 85P1B3. Recombinant DNA molecules containing 85P1B3 polynucleotides, cells transformed

or transduced with such molecules, and host-vector systems for the expression of 85P1B3 gene products are also provided. The invention further provides antibodies that bind to 85P1B3 proteins and polypeptide fragments thereof, including polyclonal and monoclonal antibodies, murine and other mammalian antibodies, chimeric antibodies, humanized and fully human antibodies, and antibodies labeled with a detectable marker.

The invention further provides methods for detecting the presence and status of 85P1B3 polynucleotides and proteins in various biological samples, as well as methods for identifying cells that express 85P1B3. A typical embodiment of this invention provides methods for monitoring 85P1B3 gene products in a tissue or hematology sample having or suspected of having some form of growth dysregulation such as cancer.

The invention further provides various immunogenic or therapeutic compositions and strategies for treating cancers that express 85P1B3 such as prostate cancers, including therapies aimed at inhibiting the transcription, translation, processing or function of 85P1B3 as well as cancer vaccines.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1. 85P1B3 SSH sequence. The 85P1B3 SSH sequence contains 259 bp. (SEQ ID. NO. : _____)

Figure 2. The cDNA (SEQ ID. NO. :____) and amino acid sequence (SEQ ID. NO. :____) of 85P1B3. The start methionine is underlined. The open reading frame extends from nucleic acid 13 to 702 including the stop codon.

Figure 3. Amino acid sequence of 85P1B3 (SEQ ID. NO. :____). The 85P1B3 protein has 229 amino acids.

Figure 4. Sequence alignment of 85P1B3 with GenBank accession number AAC39561.1 (AF025441), Opa-interacting protein OIP5 (SEQ ID. NO. :_____).

Figure 5. Hydrophilicity amino acid profile of 85P1B3 determined by computer algorithm sequence analysis using the method of Hopp and Woods (Hopp T.P., Woods K.R., 1981. Proc. Natl. Acad. Sci. U.S.A. 78:3824-3828) accessed on the Protscale website (www.expasy.ch/cgi-bin/protscale.pl) through the ExPasy molecular biology server.

Figure 6. Hydropathicity amino acid profile of 85P1B3 determined by computer algorithm sequence analysis using the method of Kyte and Doolittle (Kyte J., Doolittle R.F., 1982. J. Mol. Biol. 157:105-132) accessed on the ProtScale website (www.expasy.ch/cgi-bin/protscale.pl) through the ExPasy molecular biology server.

Figure 7. Percent accessible residues amino acid profile of 85P1B3 determined by computer algorithm sequence analysis using the method of Janin (Janin J., 1979 Nature 277:491-492) accessed on the ProtScale website (www.expasy.ch/cgi-bin/protscale.pl) through the ExPasy molecular biology server.

Figure 8. Average flexibility amino acid profile of 85P1B3 determined by computer algorithm sequence analysis using the method of Bhaskaran and Ponnuswamy (Bhaskaran R., and Ponnuswamy P.K., 1988. Int. J. Pept. Protein Res. 32:242-255) accessed on the ProtScale website (www.expasy.ch/cgi-bin/protscale.pl) through the ExPasy molecular biology server.

Figure 9. Beta-turn amino acid profile of 85P1B3 determined by computer algorithm sequence analysis using the method of Deleage and Roux (Deleage, G., Roux B. 1987 Protein Engineering 1:289-294) accessed on the ProtScale website (www.expasy.ch/cgi-bin/protscale.pl) through the ExPasy molecular biology server.

Figure 10. RT-PCR analysis of 85P1B3 expression. First strand cDNA was prepared from vital pool 1 (VP1: liver, lung and kidney), vital pool 2 (VP2, pancreas, spleen and stomach), prostate xenograft pool (LAPC-4AD, LAPC-4AI, LAPC-9AD, LAPC-9AI), prostate cancer pool, bladder cancer pool, kidney cancer pool, colon cancer pool, lung cancer pool, ovary cancer pool, breast cancer pool, and cancer metastasis pool. Normalization was performed by PCR using primers to actin and GAPDH. Semi-quantitative PCR, using primers to 85P1B3, was performed at 26 and 30 cycles of amplification.

Figure 11. Expression of 85P1B3 in normal human tissues. Two multiple tissue northern blots (Clontech) with 2 μg of mRNA/lane, were probed with the 85P1B3 SSH sequence. Size standards in kilobases (kb) are indicated on the side. The results show exclusive expression of an approximately 1.4 kb 85P1B3 transcript in testis but not in any other normal tissues.

Figure 12. Expression of 85P1B3 in human cancer cell lines. RNA was extracted from a panel of human cancer cell lines. Northern blots with 10 μg of total RNA/lane were probed with the 85P1B3 SSH sequence. Size standards in kilobases (kb) are indicated on the side.

Figure 13. Expression of 85P1B3 in human patient cancer specimens and cancer cell lines. Expression of 85P1B3 was assayed in a panel of human cancers (T) and their respective matched normal tissues (N) on RNA dot blots. 85P1B3 expression was detected in the cancers of the breast, prostate, uterus, cervix, stomach and lung. 85P1B3 was also found to be highly expressed in all human cancer cell lines tested.

Figure 14. Expression of 85P1B3 in colon cancer patient specimens. RNA was extracted from colon cancer cell lines (CL), normal colon (N), colon tumors (T) and their normal adjacent tissues (Nat) derived from colon cancer patients. Northern blots with 10 μg of total RNA/lane were probed with the 85P1B3 SSH sequence. Size standards in kilobases (kb) are indicated on the side. Results show expression

of 85P1B3 in 2 colon tumor specimens but not in the corresponding normal adjacent tissue. Expression is also seen in all 4 colon cancer cell lines (Colo 205, LoVo, T84, Caco-2). P1 – Stage III, T2N1MX; P2 – Stage III, T3N1MX.

Figure 15. Expression of 85P1B3 in bladder cancer patient specimens. RNA was extracted from bladder cancer cell lines (CL), normal bladder (N), bladder tumors (T) and their normal adjacent tissue (Nat) derived from bladder cancer patients. Northern blot with 10 μg of total RNA/lane were probed with the 85P1B3 SSH sequence. Size standards in kilobases (kb) are indicated on the side. Results show expression of 85P1B3 in 3 of 5 bladder tumor specimens. Expression is also seen in all three bladder cancer cell lines, UM-UC-3, J82, and SCABER.

Figure 16. Expression of 85P1B3 in lung cancer patient specimens. RNA was extracted from lung cancer cell lines (CL), normal lung (N), lung tumors (T) and their normal adjacent tissue (NAT) derived from lung cancer patients. Northern blot with 10 μg of total RNA/lane was probed with the 85P1B3 SSH sequence. Size standards in kilobases (kb) are indicated on the side. Results show expression of 85P1B3 in three lung tumor specimens. Expression is also seen in all lung cancer cell lines.

Figure 17. Expression of 85P1B3 in Prostate Cancer Xenografts Following Castration. Male mice were injected with LAPC-9AD tumor cells. When tumor reached a palpable size (0.3-0.5cm in diameter), mice were castrated and tumors harvested at different time points following castration. RNA was isolated from the xenograft tissues. Northern blots with 10 μg of total RNA/lane were probed with the 85P1B3 SSH fragment. Size standards in kilobases (kb) are indicated on the side. Results show expression of 85P1B3 is maintained following castration. A picture of the ethidium-bromide staining of the RNA gel is also presented. Hybridization of the same northern blot with the androgen-dependent gene TMPRSS2 confirms the quality of the the androgen deprivation following castration.

Figure 18. Expression of 85P1B3 in PC3 Cells Following Retroviral-Mediated Gene Delivery. PC3 cells were transduced with the pSRα retroviral vector encoding the 85P1B3 gene. Following selection with neomycin, the cells were expanded and RNA was extracted. Northern blot with 10 μg of total RNA/lane was probed with the 85P1B3 SSH sequence. Size standards in kilobases (kb) are indicated on the side. Results show expression of the 85P1B3 transcript driven from the retroviral LTR, which migrates slower than the endogenous 1.4 kb 85P1B3 transcript. LAPC-9AI shows only expression of the endogenous 85P1B3, but not the pSRα transcript.

Figure 19. Schematic diagram of the alignment of 85P1B3 with its splice variant. The region of homology between 85P1B3 and its splice variant 1 is marked with a hatched box. Regions specific for 85P1B3 are marked in white boxes, and the ones specific for the splice variant 1 as black boxes. The SSH sequence of 85P1B3 is also indicated by a white box.

Figure 20. Western analysis of 85P1B3 expression with an anti-85P1B3 polyclonal antibody. Panel A. Detection of GST-85P1B3 with anti-GST-85P1B3 rabbit serum. 200 ng of GST-85P1B3 (amino acids 1-229) and 200 ng of GST alone were separated by SDS-PAGE and transferred to nitrocellulose. The blot was then incubated with indicated dilutions of anti-85P1B3 serum. Immunoreactive bands were detected by incubation with anti-rabbit IgG HRP-secondary antibody and visualized by enhanced chemiluminescence and exposure to autoradiography film. Shown with arrows is detection of the GST-85P1B3 protein and minimal detection of GST alone. Panel B. 293T cells were transiently transfected with either empty pCDNA 3.1 vector or pCDNA 3.1 carrying the 85P1B3 cDNA. Lysates of the cells were separated by SDS-PAGE and subjected to Western analysis as performed for the data in Panel A, with 2 μg/ml of purified anti-85P1B3 polyclonal antibody. Panel C. Western analysis was carried out as for the data in Panel B, but using an anti-His polyclonal antibody (Santa Cruz Biotechnology, Santa Cruz, CA). Arrows indicate the immunoreactive bands corresponding to His-tagged 85P1B3 protein.

Figure 21. Secondary structure and transmembrane prediction for 85P1B3. Panel A. The secondary structure of 85P1B3 protein was predicted using the HNN - Hierarchical Neural Network method (Guermeur, 1997, http://pbil.ibcp.fr/cgi-bin/npsa_automat.pl?page=npsa_nn.html), accessed from the ExPasy molecular biology server (http://www.expasy.ch/tools/). This method indicates the presence and location of alpha helices (h), extended strands (e), and random coils (c) from the primary protein sequence. The percent of the protein in a given secondary structure is also given. Panel B. Schematic representation of the probability of existence of transmembrane regions of 85P1B3 based on the TMpred algorithm of Hofmann and Stoffel which utilizes TMBASE (K. Hofmann, W. Stoffel. TMBASE - A database of membrane spanning protein segments Biol. Chem. Hoppe-Seyler 374:166, 1993). Stretches of amino acids approximately 17-33 amino acids in length with a value greater than 0 are potential transmembrane helices. This program indicates the presence of one helix in 85P1B3. Panel C. Schematic representation of the probability of the existence of transmembrane regions and the extracellular and intracellular orientation of 85P1B3 based on the algorithm of Sonnhammer, von Heijne, and Krogh (Erik, L.L., et al., A hidden Markov model for predicting transmembrane helices in protein sequences. In Proc. of Sixth Int. Conf. on Intelligent Systems for Molecular Biology, p 175-182 Ed J. Glasgow, et al., Menlo Park, CA: AAAI Press, 1998). This program indicates 85P1B3 to be an intracellular protein without transmembrane domains. These transmembrane prediction results are also summarized in Table XXV.

DETAILED DESCRIPTION OF THE INVENTION

Outline of Sections

I.) **Definitions**

II.) **85P1B3 Polynucleotides**

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X.C.4. Vaccine Compositions Comprising DC Pulsed with CTL and/or HTL Peptides
X.D.) Adoptive Immunotherapy
X.E.) Administration of Vaccines for Therapeutic or Prophylactic Purposes
XI.) Diagnostic and Prognostic Embodiments of 85P1B3.
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XII.A.) Inhibition of 85P1B3 With Intracellular Antibodies
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XIII.) KITS

I.) Definitions:

Unless otherwise defined, all terms of art, notations and other scientific terms or terminology used herein are intended to have the meanings commonly understood by those of skill in the art to which this invention pertains. In some cases, terms with commonly understood meanings are defined herein for clarity and/or for ready reference, and the inclusion of such definitions herein should not necessarily be construed to represent a substantial difference over what is generally understood in the art. Many of the techniques and procedures described or referenced herein are well understood and commonly employed using conventional methodology by those skilled in the art, such as, for example, the widely utilized molecular cloning methodologies described in Sambrook et al., Molecular Cloning: A Laboratory Manual 2nd. edition (1989) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. As appropriate, procedures involving the use of commercially available kits and reagents are generally carried out in accordance with manufacturer defined protocols and/or parameters unless otherwise noted.

The terms "advanced prostate cancer", "locally advanced prostate cancer", "advanced disease" and "locally advanced disease" mean prostate cancers that have extended through the prostate capsule, and are meant to include stage C disease under the American Urological Association (AUA) system, stage C1 - C2 disease under the Whitmore-Jewett system, and stage T3 - T4 and N+ disease under the TNM (tumor, node, metastasis) system. In general, surgery is not recommended for patients with locally advanced disease, and these patients have substantially less favorable outcomes compared to patients having clinically localized (organ-confined) prostate cancer. Locally advanced disease is clinically identified by palpable evidence of induration beyond the lateral border of the prostate, or asymmetry or induration above the prostate base. Locally advanced prostate cancer is presently diagnosed pathologically following radical prostatectomy if the tumor invades or penetrates the prostatic capsule, extends into the surgical margin, or invades the seminal vesicles.

"Altering the native glycosylation pattern" is intended for purposes herein to mean deleting one or more carbohydrate moieties found in native sequence 85P1B3 (either by removing the underlying glycosylation site or by deleting the glycosylation by chemical and/or enzymatic means), and/or adding one or more glycosylation sites that are not present in the native sequence 85P1B3. In addition, the phrase includes qualitative changes in the glycosylation of the native proteins, involving a change in the nature and proportions of the various carbohydrate moieties present.

The term "analog" refers to a molecule which is structurally similar or shares similar or corresponding attributes with another molecule (e.g. a 85P1B3-related protein). For example an analog of the 85P1B3 protein can be specifically bound by an antibody or T cell that specifically binds to 85P1B3.

The term "antibody" is used in the broadest sense. Therefore an "antibody" can be naturally occurring or man-made such as monoclonal antibodies produced by conventional hybridoma technology. Anti-85P1B3 antibodies comprise monoclonal and polyclonal antibodies as well as fragments containing the antigen-binding domain and/or one or more complementarity determining regions of these antibodies.

An "antibody fragment" is defined as at least a portion of the variable region of the immunoglobulin molecule that binds to its target, i.e., the antigen-binding region. In one embodiment it specifically covers single anti-85P1B3 antibodies and clones thereof (including agonist, antagonist and neutralizing antibodies) and anti-85P1B3 antibody compositions with polyepitopic specificity.

The term "codon optimized sequences" refers to nucleotide sequences that have been optimized for a particular host species by replacing any codons having a usage frequency of less than about 20%. Nucleotide sequences that have been optimized for expression in a given host species by elimination of spurious polyadenylation sequences, elimination of exon/intron splicing signals, elimination of transposon-like repeats and/or optimization of GC content in addition to codon optimization are referred to herein as an "expression enhanced sequences."

The term "cytotoxic agent" refers to a substance that inhibits or prevents the function of cells and/or causes destruction of cells. The term is intended to include radioactive isotopes chemotherapeutic agents, and toxins such as small molecule toxins or enzymatically active toxins of bacterial, fungal, plant or animal origin, including fragments and/or variants thereof. Examples of cytotoxic agents include, but are not limited to maytansinoids, yttrium, bismuth, ricin, ricin A-chain, doxorubicin, daunorubicin, taxol, ethidium bromide, mitomycin, etoposide, tenoposide, vincristine, vinblastine, colchicine, dihydroxy anthracin dione, actinomycin, diphtheria toxin, Pseudomonas exotoxin (PE) A, PE40, abrin, abrin A chain, modeccin A chain, alpha-sarcin, gelonin, mitogellin, retstrictocin, phenomycin, enomycin, curicin, crotin, calicheamicin, sapaonaria officinalis inhibitor, and glucocorticoid and other chemotherapeutic agents, as well as radioisotopes such as At^{211}, I^{131}, I^{125}, Y^{90}, Re^{186}, Re^{188}, Sm^{153}, Bi^{212}, P^{32} and radioactive isotopes of Lu. Antibodies may also be conjugated to an anti-cancer pro-drug activating enzyme capable of converting the pro-drug to its active form.

The term "homolog" refers to a molecule which exhibits homology to another molecule, by for example, having sequences of chemical residues that are the same or similar at corresponding positions.

"Human Leukocyte Antigen" or "HLA" is a human class I or class II Major Histocompatibility Complex (MHC) protein (*see, e.g.*, Stites, *et al.*, IMMUNOLOGY, 8TH ED., Lange Publishing, Los Altos, CA (1994).

The terms "hybridize", "hybridizing", "hybridizes" and the like, used in the context of polynucleotides, are meant to refer to conventional hybridization conditions, preferably such as hybridization in 50% formamide/6XSSC/0.1% SDS/100 μg/ml ssDNA, in which temperatures for

hybridization are above 37 degrees C and temperatures for washing in 0.1XSSC/0.1% SDS are above 55 degrees C.

The phrases "isolated" or "biologically pure" refer to material which is substantially or essentially free from components which normally accompany the material as it is found in its native state. Thus, isolated peptides in accordance with the invention preferably do not contain materials normally associated with the peptides in their *in situ* environment. For example, a polynucleotide is said to be "isolated" when it is substantially separated from contaminant polynucleotides that correspond or are complementary to genes other than the 85P1B3 gene or that encode polypeptides other than 85P1B3 gene product or fragments thereof. A skilled artisan can readily employ nucleic acid isolation procedures to obtain an isolated 85P1B3 polynucleotide. A protein is said to be "isolated," for example, when physical, mechanical or chemical methods are employed to remove the 85P1B3 protein from cellular constituents that are normally associated with the protein. A skilled artisan can readily employ standard purification methods to obtain an isolated 85P1B3 protein. Alternatively, an isolated protein can be prepared by chemical means.

The term "mammal" refers to any organism classified as a mammal, including mice, rats, rabbits, dogs, cats, cows, horses and humans. In one embodiment of the invention, the mammal is a mouse. In another embodiment of the invention, the mammal is a human.

The terms "metastatic prostate cancer" and "metastatic disease" mean prostate cancers that have spread to regional lymph nodes or to distant sites, and are meant to include stage D disease under the AUA system and stage TxNxM+ under the TNM system. As is the case with locally advanced prostate cancer, surgery is generally not indicated for patients with metastatic disease, and hormonal (androgen ablation) therapy is a preferred treatment modality. Patients with metastatic prostate cancer eventually develop an androgen-refractory state within 12 to 18 months of treatment initiation. Approximately half of these androgen-refractory patients die within 6 months after developing that status. The most common site for prostate cancer metastasis is bone. Prostate cancer bone metastases are often osteoblastic rather than osteolytic (i.e., resulting in net bone formation). Bone metastases are found most frequently in the spine, followed by the femur, pelvis, rib cage, skull and humerus. Other common sites for metastasis include lymph nodes, lung, liver and brain. Metastatic prostate cancer is typically diagnosed by open or laparoscopic pelvic lymphadenectomy, whole body radionuclide scans, skeletal radiography, and/or bone lesion biopsy.

The term "monoclonal antibody" refers to an antibody obtained from a population of substantially homogeneous antibodies, i.e., the antibodies comprising the population are identical except for possible naturally occurring mutations that are present in minor amounts.

A "motif", as in biological motif of an 85P1B3-related protein, refers to any pattern of amino acids forming part of the primary sequence of a protein, that is associated with a particular function (e.g. protein-

protein interaction, protein-DNA interaction, etc) or modification (e.g. that is phosphorylated, glycosylated or amidated), or localization (e.g. secretory sequence, nuclear localization sequence, etc.) or a sequence that is correlated with being immunogenic, either humorally or cellularly. A motif can be either contiguous or capable of being aligned to certain positions that are generally correlated with a certain function or property. In the context of HLA motifs, "motif" refers to the pattern of residues in a peptide of defined length, usually a peptide of from about 8 to about 13 amino acids for a class I HLA motif and from about 6 to about 25 amino acids for a class II HLA motif, which is recognized by a particular HLA molecule. Peptide motifs for HLA binding are typically different for each protein encoded by each human HLA allele and differ in the pattern of the primary and secondary anchor residues.

A "pharmaceutical excipient" comprises a material such as an adjuvant, a carrier, pH-adjusting and buffering agents, tonicity adjusting agents, wetting agents, preservative, and the like.

"Pharmaceutically acceptable" refers to a non-toxic, inert, and/or composition that is physiologically compatible with humans or other mammals.

The term "polynucleotide" means a polymeric form of nucleotides of at least 10 bases or base pairs in length, either ribonucleotides or deoxynucleotides or a modified form of either type of nucleotide, and is meant to include single and double stranded forms of DNA and/or RNA. In the art, this term if often used interchangeably with "oligonucleotide". A polynucleotide can comprise a nucleotide sequence disclosed herein wherein thymidine (T) (as shown for example in SEQ ID NO: 702) can also be uracil (U); this definition pertains to the differences between the chemical structures of DNA and RNA, in particular the observation that one of the four major bases in RNA is uracil (U) instead of thymidine (T).

The term "polypeptide" means a polymer of at least about 4, 5, 6, 7, or 8 amino acids. Throughout the specification, standard three letter or single letter designations for amino acids are used. In the art, this term is often used interchangeably with "peptide" or "protein".

An HLA "primary anchor residue" is an amino acid at a specific position along a peptide sequence which is understood to provide a contact point between the immunogenic peptide and the HLA molecule. One to three, usually two, primary anchor residues within a peptide of defined length generally defines a "motif" for an immunogenic peptide. These residues are understood to fit in close contact with peptide binding groove of an HLA molecule, with their side chains buried in specific pockets of the binding groove. In one embodiment, for example, the primary anchor residues for an HLA class I molecule are located at position 2 (from the amino terminal position) and at the carboxyl terminal position of a 8, 9, 10, 11, or 12 residue peptide epitope in accordance with the invention. In another embodiment, for example, the primary anchor residues of a peptide that will bind an HLA class II molecule are spaced relative to each other, rather than to the termini of a peptide, where the peptide is generally of at least 9 amino acids in length. The primary anchor positions for each motif and supermotif are set forth in Table IV. For example,

analog peptides can be created by altering the presence or absence of particular residues in the primary and/or secondary anchor positions shown in Table IV. Such analogs are used to modulate the binding affinity and/or population coverage of a peptide comprising a particular HLA motif or supermotif.

A "recombinant" DNA or RNA molecule is a DNA or RNA molecule that has been subjected to molecular manipulation *in vitro*.

"Stringency" of hybridization reactions is readily determinable by one of ordinary skill in the art, and generally is an empirical calculation dependent upon probe length, washing temperature, and salt concentration. In general, longer probes require higher temperatures for proper annealing, while shorter probes need lower temperatures. Hybridization generally depends on the ability of denatured nucleic acid sequences to reanneal when complementary strands are present in an environment below their melting temperature. The higher the degree of desired homology between the probe and hybridizable sequence, the higher the relative temperature that can be used. As a result, it follows that higher relative temperatures would tend to make the reaction conditions more stringent, while lower temperatures less so. For additional details and explanation of stringency of hybridization reactions, see Ausubel et al., Current Protocols in Molecular Biology, Wiley Interscience Publishers, (1995).

"Stringent conditions" or "high stringency conditions", as defined herein, are identified by, but not limited to, those that: (1) employ low ionic strength and high temperature for washing, for example 0.015 M sodium chloride/0.0015 M sodium citrate/0.1% sodium dodecyl sulfate at 50°C; (2) employ during hybridization a denaturing agent, such as formamide, for example, 50% (v/v) formamide with 0.1% bovine serum albumin/0.1% Ficoll/0.1% polyvinylpyrrolidone/50 mM sodium phosphate buffer at pH 6.5 with 750 mM sodium chloride, 75 mM sodium citrate at 42 °C; or (3) employ 50% formamide, 5 x SSC (0.75 M NaCl, 0.075 M sodium citrate), 50 mM sodium phosphate (pH 6.8), 0.1% sodium pyrophosphate, 5 x Denhardt's solution, sonicated salmon sperm DNA (50 μg/ml), 0.1% SDS, and 10% dextran sulfate at 42 °C, with washes at 42°C in 0.2 x SSC (sodium chloride/sodium. citrate) and 50% formamide at 55 °C, followed by a high-stringency wash consisting of 0.1 x SSC containing EDTA at 55 °C. "Moderately stringent conditions" are described by, but not limited to, those in Sambrook et al., Molecular Cloning: A Laboratory Manual, New York: Cold Spring Harbor Press, 1989, and include the use of washing solution and hybridization conditions (e.g., temperature, ionic strength and %SDS) less stringent than those described above. An example of moderately stringent conditions is overnight incubation at 37°C in a solution comprising: 20% formamide, 5 x SSC (150 mM NaCl, 15 mM trisodium citrate), 50 mM sodium phosphate (pH 7.6), 5 x Denhardt's solution, 10% dextran sulfate, and 20 mg/mL denatured sheared salmon sperm DNA, followed by washing the filters in 1 x SSC at about 37-50°C. The skilled artisan will recognize how to adjust the temperature, ionic strength, etc. as necessary to accommodate factors such as probe length and the like.

An HLA "supermotif" is a peptide binding specificity shared by HLA molecules encoded by two or more HLA alleles.

As used herein "to treat" or "therapeutic" and grammatically related terms, refrer to any improvement of any consequence of disease, such as prolonged survival, less morbidity, and/or a lessening of side effects which are the byproducts of an alternative therapeutic modality; full erdaication of disease is not required.

A "transgenic animal" (e.g., a mouse or rat) is an animal having cells that contain a transgene, which transgene was introduced into the animal or an ancestor of the animal at a prenatal, e.g., an embryonic stage. A "transgene" is a DNA that is integrated into the genome of a cell from which a transgenic animal develops.

As used herein, an HLA or cellular immune response "vaccine" is a composition that contains or encodes one or more peptides of the invention. There are numerous embodiments of such vaccines, such as a cocktail of one or more individual peptides; one or more peptides of the invention comprised by a polyepitopic peptide; or nucleic acids that encode such individual peptides or polypeptides, *e.g.*, a minigene that encodes a polyepitopic peptide. The "one or more peptides" can include any whole unit integer from 1-150 or more, e.g., at least 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, or 150 or more peptides of the invention. The peptides or polypeptides can optionally be modified, such as by lipidation, addition of targeting or other sequences. HLA class I peptides of the invention can be admixed with, or linked to, HLA class II peptides, to facilitate activation of both cytotoxic T lymphocytes and helper T lymphocytes. HLA vaccines can also comprise peptide-pulsed antigen presenting cells, *e.g.*, dendritic cells.

The term "variant" refers to a molecule that exhibits a variation from a described type or norm, such as a protein that has one or more different amino acid residues in the corresponding position(s) of a specifically described protein (e.g. the 85P1B3 protein shown in Figure 2 or Figure 3). An analog is an example of a variant protein.

The 85P1B3-related proteins of the invention include those specifically identified herein, as well as allelic variants, conservative substitution variants, analogs and homologs that can be isolated/generated and characterized without undue experimentation following the methods outlined herein or readily available in the art. Fusion proteins that combine parts of different 85P1B3 proteins or fragments thereof, as well as fusion proteins of a 85P1B3 protein and a heterologous polypeptide are also included. Such 85P1B3 proteins are collectively referred to as the 85P1B3-related proteins, the proteins of the invention, or 85P1B3. The term "85P1B3-related protein" refers to a polypeptide fragment or an 85P1B3 protein sequence of 4, 5, 6, 7, 8, 9, 10,

11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, or more than 25 amino acids; or, at least 30, 35, 40, 45, 50, 55, 60, 65, 70, 80, 85, 90, 95, 100 or more than 100 amino acids.

II.) 85P1B3 Polynucleotides

One aspect of the invention provides polynucleotides corresponding or complementary to all or part of an 85P1B3 gene, mRNA, and/or coding sequence, preferably in isolated form, including polynucleotides encoding an 85P1B3-related protein and fragments thereof, DNA, RNA, DNA/RNA hybrid, and related molecules, polynucleotides or oligonucleotides complementary to an 85P1B3 gene or mRNA sequence or a part thereof, and polynucleotides or oligonucleotides that hybridize to an 85P1B3 gene, mRNA, or to an 85P1B3 encoding polynucleotide (collectively, "85P1B3 polynucleotides"). In all instances when referred to in this section, T can also be U in Figure 2.

Embodiments of a 85P1B3 polynucleotide include: a 85P1B3 polynucleotide having the sequence shown in Figure 2, the nucleotide sequence of 85P1B3 as shown in Figure 2, wherein T is U; at least 10 contiguous nucleotides of a polynucleotide having the sequence as shown in Figure 2; or, at least 10 contiguous nucleotides of a polynucleotide having the sequence as shown in Figure 2 where T is U. For example, embodiments of 85P1B3 nucleotides comprise, without limitation:

(a) a polynucleotide comprising or consisting of the sequence as shown in Figure 2 (SEQ ID NO.: ____), wherein T can also be U;

(b) a polynucleotide comprising or consisting of the sequence as shown in Figure 2 (SEQ ID NO.: ____), from nucleotide residue number 13 through nucleotide residue number 699, wherein T can also be U;

(c) a polynucleotide that encodes a 85P1B3-related protein whose sequence is encoded by the cDNAs contained in the plasmid designated _____ deposited with American Type Culture Collection as Accession No. _____;

(d) a polynucleotide that encodes an 85P1B3-related protein that is at least 90% homologous to the entire amino acid sequence shown in SEQ ID NO.: ____;

(e) a polynucleotide that encodes an 85P1B3-related protein that is at least 90% identical to the entire amino acid sequence shown in SEQ ID NO: ____;

(f) a polynucleotide that encodes at least one peptide set forth in Tables V-XVIII;

(g) a polynucleotide that encodes a peptide region of at least 5 amino acids of Figure 3 in any whole number increment up to 229 that includes an amino acid position having a value greater than 0.5 in the Hydrophilicity profile of Figure 5;

(h) a polynucleotide that encodes a peptide region of at least 5 amino acids of Figure 3 in any whole number increment up to 229 that includes an amino acid position having a value less than 0.5 in the Hydropathicity profile of Figure 6;

(i) a polynucleotide that encodes a peptide region of at least 5 amino acids of Figure 3 in any whole number increment up to 229 that includes an amino acid position having a value greater than 0.5 in the Percent Accessible Residues profile of Figure 7;

(j) a polynucleotide that encodes a peptide region of at least 5 amino acids of Figure 3 in any whole number increment up to 229 that includes an amino acid position having a value greater than 0.5 in the Average Flexibility profile on Figure 8;

(k) a polynucleotide that encodes a peptide region of at least 5 amino acids of Figure 3 in any whole number increment up to 229 that includes an amino acid position having a value greater than 0.5 in the Beta-turn profile of Figure 9;

(l) a polynucleotide that is fully complementary to a polynucleotide of any one of (a)-(k);

(m) a polynucleotide that selectively hybridizes under stringent conditions to a polynucleotide of (a)-(l); and

(n) a polynucleotide of any of (a)-(m)or peptide of (o) (see immediately below) together with a pharmaceutical excipient and/or in a human unit dose form.

Regarding item (n) immediately above, examples of embodiments of 85P1B3 polypeptides comprise, without limitation:

(o) a peptide that is encoded by any of (a)–(k).

As used herein, a range is understood to specifically disclose all whole unit positions thereof.

Typical embodiments of the invention disclosed herein include 85P1B3 polynucleotides that encode specific portions of the 85P1B3 mRNA sequence (and those which are complementary to such sequences) such as those that encode the protein and fragments thereof, for example of 4, 5, 6, 7, 8, 9, 10,

11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, 150, 155, 160, 165, 170, 175, 180, 185, 190, 195, 200, 205, 210, 215, 220, 225, or 229 contiguous amino acids.

For example, representative embodiments of the invention disclosed herein include: polynucleotides and their encoded peptides themselves encoding about amino acid 1 to about amino acid 10 of the 85P1B3 protein shown in Figure 2 or Figure 3, polynucleotides encoding about amino acid 10 to about amino acid 20 of the 85P1B3 protein shown in Figure 2, or Figure 3, polynucleotides encoding about amino acid 20 to about amino acid 30 of the 85P1B3 protein shown in Figure 2 or Figure 3, polynucleotides encoding about amino acid 30 to about amino acid 40 of the 85P1B3 protein shown in Figure 2 or Figure 3, polynucleotides encoding about amino acid 40 to about amino acid 50 of the 85P1B3 protein shown in Figure 2 or Figure 3, polynucleotides encoding about amino acid 50 to about amino acid 60 of the 85P1B3 protein shown in Figure 2 or Figure 3, polynucleotides encoding about amino acid 60 to about amino acid 70 of the 85P1B3 protein shown in Figure 2 or Figure 3, polynucleotides encoding about amino acid 70 to about amino acid 80 of the 85P1B3 protein shown in Figure 2 or Figure 3, polynucleotides encoding about amino acid 80 to about amino acid 90 of the 85P1B3 protein shown in Figure 2 or Figure 3, polynucleotides encoding about amino acid 90 to about amino acid 100 of the 85P1B3 protein shown in Figure 2 or Figure 3, in increments of about 10 amino acids, ending at the carboxyl terminal amino acid set forth in Figure 2 or Figure 3. Accordingly polynucleotides encoding portions of the amino acid sequence (of about 10 amino acids), of amino acids 100 through the carboxyl terminal amino acid of the 85P1B3 protein are embodiments of the invention. Wherein it is understood that each particular amino acid position discloses that position plus or minus five amino acid residues.

Polynucleotides encoding relatively long portions of the 85P1B3 protein are also within the scope of the invention. For example, polynucleotides encoding from about amino acid 1 (or 20 or 30 or 40 etc.) to about amino acid 20, (or 30, or 40 or 50 etc.) of the 85P1B3 protein shown in Figure 2 or Figure 3 can be generated by a variety of techniques well known in the art. These polynucleotide fragments can include any portion of the 85P1B3 sequence as shown in Figure 2 or Figure 3.

Additional illustrative embodiments of the invention disclosed herein include 85P1B3 polynucleotide fragments encoding one or more of the biological motifs contained within the 85P1B3 protein sequence, including one or more of the motif-bearing subsequences of the 85P1B3 protein set forth in Tables V-XVIII. In another embodiment, typical polynucleotide fragments of the invention encode one or more of the regions of 85P1B3 that exhibit homology to a known molecule. In another embodiment of the invention, typical polynucleotide fragments can encode one or more of the 85P1B3 N-glycosylation sites, cAMP and cGMP-dependent protein kinase phosphorylation sites, casein kinase II phosphorylation sites or N-myristoylation site and amidation sites.

II.A.) Uses of 85P1B3 Polynucleotides

II.A.1.) Monitoring of Genetic Abnormalities

The polynucleotides of the preceding paragraphs have a number of different specific uses. The human 85P1B3 gene maps to the chromosomal location set forth in Example 3. For example, because the 85P1B3 gene maps to this chromosome, polynucleotides that encode different regions of the 85P1B3 protein are used to characterize cytogenetic abnormalities of this chromosomal locale, such as abnormalities that are identified as being associated with various cancers. In certain genes, a variety of chromosomal abnormalities including rearrangements have been identified as frequent cytogenetic abnormalities in a number of different cancers (see e.g. Krajinovic et al., Mutat. Res. 382(3-4): 81-83 (1998); Johansson et al., Blood 86(10): 3905-3914 (1995) and Finger et al., P.N.A.S. 85(23): 9158-9162 (1988)). Thus, polynucleotides encoding specific regions of the 85P1B3 protein provide new tools that can be used to delineate, with greater precision than previously possible, cytogenetic abnormalities in the chromosomal region that encodes 85P1B3 that may contribute to the malignant phenotype. In this context, these polynucleotides satisfy a need in the art for expanding the sensitivity of chromosomal screening in order to identify more subtle and less common chromosomal abnormalities (see e.g. Evans et al., Am. J. Obstet. Gynecol 171(4): 1055-1057 (1994)).

Furthermore, as 85P1B3 was shown to be highly expressed in prostate and other cancers, 85P1B3 polynucleotides are used in methods assessing the status of 85P1B3 gene products in normal versus cancerous tissues. Typically, polynucleotides that encode specific regions of the 85P1B3 protein are used to assess the presence of perturbations (such as deletions, insertions, point mutations, or alterations resulting in a loss of an antigen etc.) in specific regions of the 85P1B3 gene, such as such regions containing one or more motifs. Exemplary assays include both RT-PCR assays as well as single-strand conformation polymorphism (SSCP) analysis (see, e.g., Marrogi et al., J. Cutan. Pathol. 26(8): 369-378 (1999), both of which utilize polynucleotides encoding specific regions of a protein to examine these regions within the protein.

II.A.2.) Antisense Embodiments

Other specifically contemplated nucleic acid related embodiments of the invention disclosed herein are genomic DNA, cDNAs, ribozymes, and antisense molecules, as well as nucleic acid molecules based on an alternative backbone, or including alternative bases, whether derived from natural sources or synthesized, and include molecules capable of inhibiting the RNA or protein expression of 85P1B3. For example, antisense molecules can be RNAs or other molecules, including peptide nucleic acids (PNAs) or non-nucleic acid molecules such as phosphorothioate derivatives, that specifically bind DNA or RNA in a base pair-

dependent manner. A skilled artisan can readily obtain these classes of nucleic acid molecules using the 85P1B3 polynucleotides and polynucleotide sequences disclosed herein.

Antisense technology entails the administration of exogenous oligonucleotides that bind to a target polynucleotide located within the cells. The term "antisense" refers to the fact that such oligonucleotides are complementary to their intracellular targets, e.g., 85P1B3. See for example, Jack Cohen, Oligodeoxynucleotides, Antisense Inhibitors of Gene Expression, CRC Press, 1989; and Synthesis 1:1-5 (1988). The 85P1B3 antisense oligonucleotides of the present invention include derivatives such as S-oligonucleotides (phosphorothioate derivatives or S-oligos, see, Jack Cohen, supra), which exhibit enhanced cancer cell growth inhibitory action. S-oligos (nucleoside phosphorothioates) are isoelectronic analogs of an oligonucleotide (O-oligo) in which a nonbridging oxygen atom of the phosphate group is replaced by a sulfur atom. The S-oligos of the present invention can be prepared by treatment of the corresponding O-oligos with 3H-1,2-benzodithiol-3-one-1,1-dioxide, which is a sulfur transfer reagent. See Iyer, R. P. et al, J. Org. Chem. 55:4693-4698 (1990); and Iyer, R. P. et al., J. Am. Chem. Soc. 112:1253-1254 (1990). Additional 85P1B3 antisense oligonucleotides of the present invention include morpholino antisense oligonucleotides known in the art (see, e.g., Partridge et al., 1996, Antisense & Nucleic Acid Drug Development 6: 169-175).

The 85P1B3 antisense oligonucleotides of the present invention typically can be RNA or DNA that is complementary to and stably hybridizes with the first 100 5' codons or last 100 3' codons of the 85P1B3 genomic sequence or the corresponding mRNA. Absolute complementarity is not required, although high degrees of complementarity are preferred. Use of an oligonucleotide complementary to this region allows for the selective hybridization to 85P1B3 mRNA and not to mRNA specifying other regulatory subunits of protein kinase. In one embodiment, 85P1B3 antisense oligonucleotides of the present invention are 15 to 30-mer fragments of the antisense DNA molecule that have a sequence that hybridizes to 85P1B3 mRNA. Optionally, 85P1B3 antisense oligonucleotide is a 30-mer oligonucleotide that is complementary to a region in the first 10 5' codons or last 10 3' codons of 85P1B3. Alternatively, the antisense molecules are modified to employ ribozymes in the inhibition of 85P1B3 expression, see, e.g., L. A. Couture & D. T. Stinchcomb; *Trends Genet* 12: 510-515 (1996).

II.A.3.) Primers and Primer Pairs

Further specific embodiments of this nucleotides of the invention include primers and primer pairs, which allow the specific amplification of polynucleotides of the invention or of any specific parts thereof, and probes that selectively or specifically hybridize to nucleic acid molecules of the invention or to any part thereof. Probes can be labeled with a detectable marker, such as, for example, a radioisotope, fluorescent compound, bioluminescent compound, a chemiluminescent compound, metal chelator or

enzyme. Such probes and primers are used to detect the presence of a 85P1B3 polynucleotide in a sample and as a means for detecting a cell expressing a 85P1B3 protein.

Examples of such probes include polypeptides comprising all or part of the human 85P1B3 cDNA sequence shown in Figure 2. Examples of primer pairs capable of specifically amplifying 85P1B3 mRNAs are also described in the Examples. As will be understood by the skilled artisan, a great many different primers and probes can be prepared based on the sequences provided herein and used effectively to amplify and/or detect a 85P1B3 mRNA.

The 85P1B3 polynucleotides of the invention are useful for a variety of purposes, including but not limited to their use as probes and primers for the amplification and/or detection of the 85P1B3 gene(s), mRNA(s), or fragments thereof; as reagents for the diagnosis and/or prognosis of prostate cancer and other cancers; as coding sequences capable of directing the expression of 85P1B3 polypeptides; as tools for modulating or inhibiting the expression of the 85P1B3 gene(s) and/or translation of the 85P1B3 transcript(s); and as therapeutic agents.

The present invention includes the use of any probe as described herein to identify and isolate a 85P1B3 or 85P1B3 related nucleic acid sequence from a naturally occurring source, such as humans or other mammals, as well as the isolated nucleic acid sequence *per se*, which would comprise all or most of the sequences found in the probe used.

II.A.4.) Isolation of 85P1B3-Encoding Nucleic Acid Molecules

The 85P1B3 cDNA sequences described herein enable the isolation of other polynucleotides encoding 85P1B3 gene product(s), as well as the isolation of polynucleotides encoding 85P1B3 gene product homologs, alternatively spliced isoforms, allelic variants, and mutant forms of the 85P1B3 gene product as well as polynucleotides that encode analogs of 85P1B3-related proteins. Various molecular cloning methods that can be employed to isolate full length cDNAs encoding an 85P1B3 gene are well known (see, for example, Sambrook, J. et al., Molecular Cloning: A Laboratory Manual, 2d edition, Cold Spring Harbor Press, New York, 1989; Current Protocols in Molecular Biology. Ausubel et al.; Eds., Wiley and Sons, 1995). For example, lambda phage cloning methodologies can be conveniently employed, using commercially available cloning systems (e.g., Lambda ZAP Express, Stratagene). Phage clones containing 85P1B3 gene cDNAs can be identified by probing with a labeled 85P1B3 cDNA or a fragment thereof. For example, in one embodiment, the 85P1B3 cDNA (Figure 2) or a portion thereof can be synthesized and used as a probe to retrieve overlapping and full-length cDNAs corresponding to a 85P1B3 gene. The 85P1B3 gene itself can be isolated by screening genomic DNA libraries, bacterial artificial chromosome libraries (BACs), yeast artificial chromosome libraries (YACs), and the like, with 85P1B3 DNA probes or primers.

II.A.5.) Recombinant Nucleic Acid Molecules and Host-Vector Systems

The invention also provides recombinant DNA or RNA molecules containing an 85P1B3 polynucleotide, a fragment, analog or homologue thereof, including but not limited to phages, plasmids, phagemids, cosmids, YACs, BACs, as well as various viral and non-viral vectors well known in the art, and cells transformed or transfected with such recombinant DNA or RNA molecules. Methods for generating such molecules are well known (see, for example, Sambrook et al, 1989, supra).

The invention further provides a host-vector system comprising a recombinant DNA molecule containing a 85P1B3 polynucleotide, fragment, analog or homologue thereof within a suitable prokaryotic or eukaryotic host cell. Examples of suitable eukaryotic host cells include a yeast cell, a plant cell, or an animal cell, such as a mammalian cell or an insect cell (e.g., a baculovirus-infectible cell such as an Sf9 or HighFive cell). Examples of suitable mammalian cells include various prostate cancer cell lines such as DU145 and TsuPr1, other transfectable or transducible prostate cancer cell lines, primary cells (PrEC), as well as a number of mammalian cells routinely used for the expression of recombinant proteins (e.g., COS, CHO, 293, 293T cells). More particularly, a polynucleotide comprising the coding sequence of 85P1B3 or a fragment, analog or homolog thereof can be used to generate 85P1B3 proteins or fragments thereof using any number of host-vector systems routinely used and widely known in the art.

A wide range of host-vector systems suitable for the expression of 85P1B3 proteins or fragments thereof are available, see for example, Sambrook et al., 1989, supra; Current Protocols in Molecular Biology, 1995, supra). Preferred vectors for mammalian expression include but are not limited to pcDNA 3.1 myc-His-tag (Invitrogen) and the retroviral vector pSRαtkneo (Muller et al., 1991, MCB 11:1785). Using these expression vectors, 85P1B3 can be expressed in several prostate cancer and non-prostate cell lines, including for example 293, 293T, rat-1, NIH 3T3 and TsuPr1. The host-vector systems of the invention are useful for the production of a 85P1B3 protein or fragment thereof. Such host-vector systems can be employed to study the functional properties of 85P1B3 and 85P1B3 mutations or analogs.

Recombinant human 85P1B3 protein or an analog or homolog or fragment thereof can be produced by mammalian cells transfected with a construct encoding a 85P1B3-related nucleotide. For example, 293T cells can be transfected with an expression plasmid encoding 85P1B3 or fragment, analog or homolog thereof, the 85P1B3 or related protein is expressed in the 293T cells, and the recombinant 85P1B3 protein is isolated using standard purification methods (e.g., affinity purification using anti-85P1B3 antibodies). In another embodiment, a 85P1B3 coding sequence is subcloned into the retroviral vector pSRαMSVtkneo and used to infect various mammalian cell lines, such as NIH 3T3, TsuPr1, 293 and rat-1 in order to establish 85P1B3 expressing cell lines. Various other expression systems well known in the art can also be employed. Expression constructs encoding a leader peptide joined in frame to the 85P1B3 coding sequence can be used for the generation of a secreted form of recombinant 85P1B3 protein.

As discussed herein, redundancy in the genetic code permits variation in 85P1B3 gene sequences. In particular, it is known in the art that specific host species often have specific codon preferences, and thus one can adapt the disclosed sequence as preferred for a desired host. For example, preferred analog codon sequences typically have rare codons (i.e., codons having a usage frequency of less than about 20% in known sequences of the desired host) replaced with higher frequency codons. Codon preferences for a specific species are calculated, for example, by utilizing codon usage tables available on the INTERNET such as at URL www.dna.affrc.go.jp/~nakamura/codon.html.

Additional sequence modifications are known to enhance protein expression in a cellular host. These include elimination of sequences encoding spurious polyadenylation signals, exon/intron splice site signals, transposon-like repeats, and/or other such well-characterized sequences that are deleterious to gene expression. The GC content of the sequence is adjusted to levels average for a given cellular host, as calculated by reference to known genes expressed in the host cell. Where possible, the sequence is modified to avoid predicted hairpin secondary mRNA structures. Other useful modifications include the addition of a translational initiation consensus sequence at the start of the open reading frame, as described in Kozak, *Mol. Cell Biol.*, 9:5073-5080 (1989). Skilled artisans understand that the general rule that eukaryotic ribosomes initiate translation exclusively at the 5' proximal AUG codon is abrogated only under rare conditions (see, e.g., Kozak PNAS 92(7): 2662-2666, (1995) and Kozak NAR 15(20): 8125-8148 (1987)).

III.) 85P1B3-related Proteins

Another aspect of the present invention provides 85P1B3-related proteins. Specific embodiments of 85P1B3 proteins comprise a polypeptide having all or part of the amino acid sequence of human 85P1B3 as shown in Figure 2 or Figure 3. Alternatively, embodiments of 85P1B3 proteins comprise variant, homolog or analog polypeptides that have alterations in the amino acid sequence of 85P1B3 shown in Figure 2 or Figure 3.

In general, naturally occurring allelic variants of human 85P1B3 share a high degree of structural identity and homology (e.g., 90% or more homology). Typically, allelic variants of the 85P1B3 protein contain conservative amino acid substitutions within the 85P1B3 sequences described herein or contain a substitution of an amino acid from a corresponding position in a homologue of 85P1B3. One class of 85P1B3 allelic variants are proteins that share a high degree of homology with at least a small region of a particular 85P1B3 amino acid sequence, but further contain a radical departure from the sequence, such as a non-conservative substitution, truncation, insertion or frame shift. In comparisons of protein sequences, the terms, similarity, identity, and homology each have a distinct meaning as appreciated in the field of genetics. Moreover,

orthology and paralogy can be important concepts describing the relationship of members of a given protein family in one organism to the members of the same family in other organisms.

Amino acid abbreviations are provided in Table II. Conservative amino acid substitutions can frequently be made in a protein without altering either the conformation or the function of the protein. Proteins of the invention can comprise 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15 conservative substitutions. Such changes include substituting any of isoleucine (I), valine (V), and leucine (L) for any other of these hydrophobic amino acids; aspartic acid (D) for glutamic acid (E) and vice versa; glutamine (Q) for asparagine (N) and vice versa; and serine (S) for threonine (T) and vice versa. Other substitutions can also be considered conservative, depending on the environment of the particular amino acid and its role in the three-dimensional structure of the protein. For example, glycine (G) and alanine (A) can frequently be interchangeable, as can alanine (A) and valine (V). Methionine (M), which is relatively hydrophobic, can frequently be interchanged with leucine and isoleucine, and sometimes with valine. Lysine (K) and arginine (R) are frequently interchangeable in locations in which the significant feature of the amino acid residue is its charge and the differing pK's of these two amino acid residues are not significant. Still other changes can be considered "conservative" in particular environments (see, e.g. Table III herein; pages 13-15 "Biochemistry" 2nd ED. Lubert Stryer ed (Stanford University); Henikoff et al., PNAS 1992 Vol 89 10915-10919; Lei et al., J Biol Chem 1995 May 19; 270(20):11882-6).

Embodiments of the invention disclosed herein include a wide variety of art-accepted variants or analogs of 85P1B3 proteins such as polypeptides having amino acid insertions, deletions and substitutions. 85P1B3 variants can be made using methods known in the art such as site-directed mutagenesis, alanine scanning, and PCR mutagenesis. Site-directed mutagenesis (Carter et al., *Nucl. Acids Res., 13*:4331 (1986); Zoller et al., *Nucl. Acids Res., 10*:6487 (1987)), cassette mutagenesis (Wells et al., Gene, 34:315 (1985)), restriction selection mutagenesis (Wells et al., *Philos. Trans. R. Soc. London SerA*, 317:415 (1986)) or other known techniques can be performed on the cloned DNA to produce the 85P1B3 variant DNA.

Scanning amino acid analysis can also be employed to identify one or more amino acids along a contiguous sequence that is involved in a specific biological activity such as a protein-protein interaction. Among the preferred scanning amino acids are relatively small, neutral amino acids. Such amino acids include alanine, glycine, serine, and cysteine. Alanine is typically a preferred scanning amino acid among this group because it eliminates the side-chain beyond the beta-carbon and is less likely to alter the main-chain conformation of the variant. Alanine is also typically preferred because it is the most common amino acid. Further, it is frequently found in both buried and exposed positions (Creighton, *The Proteins*, (W.H. Freeman & Co., N.Y.); Chothia, J. Mol. Biol., 150:1 (1976)). If alanine substitution does not yield adequate amounts of variant, an isosteric amino acid can be used.

As defined herein, 85P1B3 variants, analogs or homologs, have the distinguishing attribute of having at least one epitope that is "cross reactive" with a 85P1B3 protein having the amino acid sequence of SEQ ID NO: 703. As used in this sentence, "cross reactive" means that an antibody or T cell that specifically binds to an 85P1B3 variant also specifically binds to the 85P1B3 protein having the amino acid sequence of SEQ ID NO: 703. A polypeptide ceases to be a variant of the protein shown in SEQ ID NO: 703 when it no longer contains any epitope capable of being recognized by an antibody or T cell that specifically binds to the 85P1B3 protein. Those skilled in the art understand that antibodies that recognize proteins bind to epitopes of varying size, and a grouping of the order of about four or five amino acids, contiguous or not, is regarded as a typical number of amino acids in a minimal epitope. See, e.g., Nair et al., J. Immunol 2000 165(12): 6949-6955; Hebbes et al., Mol Immunol (1989) 26(9):865-73; Schwartz et al., J Immunol (1985) 135(4):2598-608.

Another class of 85P1B3-related protein variants share 70%, 75%, 80%, 85% or 90% or more similarity with the amino acid sequence of SEQ ID NO: 703 or a fragment thereof. Another specific class of 85P1B3 protein variants or analogs comprise one or more of the 85P1B3 biological motifs described herein or presently known in the art. Thus, encompassed by the present invention are analogs of 85P1B3 fragments (nucleic or amino acid) that have altered functional (e.g. immunogenic) properties relative to the starting fragment. It is to be appreciated that motifs now or which become part of the art are to be applied to the nucleic or amino acid sequences of Figure 2 or Figure 3.

As discussed herein, embodiments of the claimed invention include polypeptides containing less than the full amino acid sequence of the 85P1B3 protein shown in Figure 2 or Figure 3. For example, representative embodiments of the invention comprise peptides/proteins having any 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15 or more contiguous amino acids of the 85P1B3 protein shown in Figure 2 or Figure 3.

Moreover, representative embodiments of the invention disclosed herein include polypeptides consisting of about amino acid 1 to about amino acid 10 of the 85P1B3 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 10 to about amino acid 20 of the 85P1B3 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 20 to about amino acid 30 of the 85P1B3 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 30 to about amino acid 40 of the 85P1B3 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 40 to about amino acid 50 of the 85P1B3 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 50 to about amino acid 60 of the 85P1B3 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 60 to about amino acid 70 of the 85P1B3 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 70 to about amino acid 80 of the 85P1B3 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 80 to about amino acid 90 of the 85P1B3 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 90 to about

amino acid 100 of the 85P1B3 protein shown in Figure 2 or Figure 3, etc. throughout the entirety of the 85P1B3 amino acid sequence. Moreover, polypeptides consisting of about amino acid 1 (or 20 or 30 or 40 etc.) to about amino acid 20, (or 130, or 140 or 150 etc.) of the 85P1B3 protein shown in Figure 2 or Figure 3 are embodiments of the invention. It is to be appreciated that the starting and stopping positions in this paragraph refer to the specified position as well as that position plus or minus 5 residues.

85P1B3-related proteins are generated using standard peptide synthesis technology or using chemical cleavage methods well known in the art. Alternatively, recombinant methods can be used to generate nucleic acid molecules that encode a 85P1B3-related protein. In one embodiment, nucleic acid molecules provide a means to generate defined fragments of the 85P1B3 protein (or variants, homologs or analogs thereof).

III.A.) Motif-bearing Protein Embodiments

Additional illustrative embodiments of the invention disclosed herein include 85P1B3 polypeptides comprising the amino acid residues of one or more of the biological motifs contained within the 85P1B3 polypeptide sequence set forth in Figure 2 or Figure 3. Various motifs are known in the art, and a protein can be evaluated for the presence of such motifs by a number of publicly available Internet sites (see, e.g., URL addresses: pfam.wustl.edu/; searchlauncher.bcm.tmc.edu/seq-search/struc-predict.html psort.ims.u-tokyo.ac.jp/; www.cbs.dtu.dk/; www.ebi.ac.uk/interpro/scan.html; www.expasy.ch/tools/scnpsit1.html; Epimatrix™ and Epimer™, Brown University, www.brown.edu/Research/TB-HIV_Lab/epimatrix/epimatrix.html; and BIMAS, bimas.dcrt.nih.gov/.).

Motif bearing subsequences of the 85P1B3 protein are set forth and identified in Table XIX.

Table XX sets forth several frequently occurring motifs based on pfam searches (see URL address pfam.wustl.edu/). The columns of Table XX list (1) motif name abbreviation, (2) percent identity found amongst the different member of the motif family, (3) motif name or description and (4) most common function; location information is included if the motif is relevant for location.

Polypeptides comprising one or more of the 85P1B3 motifs discussed above are useful in elucidating the specific characteristics of a malignant phenotype in view of the observation that the 85P1B3 motifs discussed above are associated with growth dysregulation and because 85P1B3 is overexpressed in certain cancers (See, e.g., Table I). Casein kinase II, cAMP and camp-dependent protein kinase, and Protein Kinase C, for example, are enzymes known to be associated with the development of the malignant phenotype (see e.g. Chen et al., Lab Invest., 78(2): 165-174 (1998); Gaiddon et al., Endocrinology 136(10): 4331-4338 (1995); Hall et al., Nucleic Acids Research 24(6): 1119-1126 (1996); Peterziel et al., Oncogene 18(46): 6322-6329 (1999) and O'Brian, Oncol. Rep. 5(2): 305-309 (1998)). Moreover, both glycosylation and myristoylation are protein modifications also associated with cancer and cancer progression (see e.g. Dennis et al., Biochem. Biophys. Acta 1473(1):21-34 (1999); Raju et al., Exp. Cell Res. 235(1): 145-154

(1997)). Amidation is another protein modification also associated with cancer and cancer progression (see e.g. Treston et al., J. Natl. Cancer Inst. Monogr. (13): 169-175 (1992)).

In another embodiment, proteins of the invention comprise one or more of the immunoreactive epitopes identified in accordance with art-accepted methods, such as the peptides set forth in Tables V-XVIII. CTL epitopes can be determined using specific algorithms to identify peptides within an 85P1B3 protein that are capable of optimally binding to specified HLA alleles (e.g., Table IV; Epimatrix™ and Epimer™, Brown University, URL www.brown.edu/Research/TB-HIV_Lab/epimatrix/epimatrix.html; and BIMAS, URL bimas.dcrt.nih.gov/.) Moreover, processes for identifying peptides that have sufficient binding affinity for HLA molecules and which are correlated with being immunogenic epitopes, are well known in the art, and are carried out without undue experimentation. In addition, processes for identifying peptides that are immunogenic epitopes, are well known in the art, and are carried out without undue experimentation either *in vitro* or *in vivo*.

Also known in the art are principles for creating analogs of such epitopes in order to modulate immunogenicity. For example, one begins with an epitope that bears a CTL or HTL motif (see, e.g., the HLA Class I and HLA Class II motifs/supermotifs of Table IV). The epitope is analoged by substituting out an amino acid at one of the specified positions, and replacing it with another amino acid specified for that position. For example, one can substitute out a deleterious residue in favor of any other residue, such as a preferred residue as defined in Table IV; substitute a less-preferred residue with a preferred residue as defined in Table IV; or substitute an originally-occurring preferred residue with another preferred residue as defined in Table IV. Substitutions can occur at primary anchor positions or at other positions in a peptide; see, e.g., Table IV.

A variety of references reflect the art regarding the identification and generation of epitopes in a protein of interest as well as analogs thereof. See, for example, WO 9733602 to Chesnut et al.; Sette, Immunogenetics 1999 50(3-4): 201-212; Sette et al., J. Immunol. 2001 166(2): 1389-1397; Sidney et al., Hum. Immunol. 1997 58(1): 12-20; Kondo et al., Immunogenetics 1997 45(4): 249-258; Sidney et al., J. Immunol. 1996 157(8): 3480-90; and Falk et al., Nature 351: 290-6 (1991); Hunt et al., Science 255:1261-3 (1992); Parker et al., J. Immunol. 149:3580-7 (1992); Parker et al., J. Immunol. 152:163-75 (1994)); Kast et al., 1994 152(8): 3904-12; Borras-Cuesta et al., Hum. Immunol. 2000 61(3): 266-278; Alexander et al., J. Immunol. 2000 164(3); 164(3): 1625-1633; Alexander et al., PMID: 7895164, UI: 95202582; O'Sullivan et al., J. Immunol. 1991 147(8): 2663-2669; Alexander et al., Immunity 1994 1(9): 751-761 and Alexander et al., Immunol. Res. 1998 18(2): 79-92.

Related embodiments of the inventions include polypeptides comprising combinations of the different motifs set forth in Table XIX, and/or, one or more of the predicted CTL epitopes of Table V through Table XVIII, and/or, one or more of the T cell binding motifs known in the art. Preferred

embodiments contain no insertions, deletions or substitutions either within the motifs or the intervening sequences of the polypeptides. In addition, embodiments which include a number of either N-terminal and/or C-terminal amino acid residues on either side of these motifs may be desirable (to, for example, include a greater portion of the polypeptide architecture in which the motif is located). Typically the number of N-terminal and/or C-terminal amino acid residues on either side of a motif is between about 1 to about 100 amino acid residues, preferably 5 to about 50 amino acid residues.

85P1B3-related proteins are embodied in many forms, preferably in isolated form. A purified 85P1B3 protein molecule will be substantially free of other proteins or molecules that impair the binding of 85P1B3 to antibody, T cell or other ligand. The nature and degree of isolation and purification will depend on the intended use. Embodiments of a 85P1B3-related proteins include purified 85P1B3-related proteins and functional, soluble 85P1B3-related proteins. In one embodiment, a functional, soluble 85P1B3 protein or fragment thereof retains the ability to be bound by antibody, T cell or other ligand.

The invention also provides 85P1B3 proteins comprising biologically active fragments of the 85P1B3 amino acid sequence shown in Figure 2 or Figure 3. Such proteins exhibit properties of the 85P1B3 protein, such as the ability to elicit the generation of antibodies that specifically bind an epitope associated with the 85P1B3 protein; to be bound by such antibodies; to elicit the activation of HTL or CTL; and/or, to be recognized by HTL or CTL.

85P1B3-related polypeptides that contain particularly interesting structures can be predicted and/or identified using various analytical techniques well known in the art, including, for example, the methods of Chou-Fasman, Garnier-Robson, Kyte-Doolittle, Eisenberg, Karplus-Schultz or Jameson-Wolf analysis, or on the basis of immunogenicity. Fragments that contain such structures are particularly useful in generating subunit-specific anti-85P1B3 antibodies, or T cells or in identifying cellular factors that bind to 85P1B3.

CTL epitopes can be determined using specific algorithms to identify peptides within an 85P1B3 protein that are capable of optimally binding to specified HLA alleles (e.g., by using the SYFPEITHI site at World Wide Web URL syfpeithi.bmi-heidelberg.com/; the listings in Table IV(A)-(E); Epimatrix™ and Epimer™, Brown University, URL (www.brown.edu/Research/TB-HIV_Lab/epimatrix/epimatrix.html); and BIMAS, URL bimas.dcrt.nih.gov/). Illustrating this, peptide epitopes from 85P1B3 that are presented in the context of human MHC class I molecules HLA-A1, A2, A3, A11, A24, B7 and B35 were predicted (Tables V-XVIII). Specifically, the complete amino acid sequence of the 85P1B3 protein was entered into the HLA Peptide Motif Search algorithm found in the Bioinformatics and Molecular Analysis Section (BIMAS) web site listed above. The HLA peptide motif search algorithm was developed by Dr. Ken Parker based on binding of specific peptide sequences in the groove of HLA Class I molecules, in particular HLA-A2 (see, e.g., Falk et al., Nature 351: 290-6 (1991); Hunt et al., Science 255:1261-3 (1992); Parker et al., J. Immunol. 149:3580-7 (1992); Parker et al., J. Immunol. 152:163-75 (1994)). This algorithm allows

location and ranking of 8-mer, 9-mer, and 10-mer peptides from a complete protein sequence for predicted binding to HLA-A2 as well as numerous other HLA Class I molecules. Many HLA class I binding peptides are 8-, 9-, 10 or 11-mers. For example, for class I HLA-A2, the epitopes preferably contain a leucine (L) or methionine (M) at position 2 and a valine (V) or leucine (L) at the C-terminus (see, e.g., Parker et al., J. Immunol. 149:3580-7 (1992)). Selected results of 85P1B3 predicted binding peptides are shown in Tables V-XVIII herein. In Tables V-XVIII, the top 50 ranking candidates, 9-mers and 10-mers, for each family member are shown along with their location, the amino acid sequence of each specific peptide, and an estimated binding score. The binding score corresponds to the estimated half time of dissociation of complexes containing the peptide at 37°C at pH 6.5. Peptides with the highest binding score are predicted to be the most tightly bound to HLA Class I on the cell surface for the greatest period of time and thus represent the best immunogenic targets for T-cell recognition.

Actual binding of peptides to an HLA allele can be evaluated by stabilization of HLA expression on the antigen-processing defective cell line T2 (see, e.g., Xue et al., Prostate 30:73-8 (1997) and Peshwa et al., Prostate 36:129-38 (1998)). Immunogenicity of specific peptides can be evaluated *in vitro* by stimulation of CD8+ cytotoxic T lymphocytes (CTL) in the presence of antigen presenting cells such as dendritic cells.

It is to be appreciated that every epitope predicted by the BIMAS site, Epimer™ and Epimatrix™ sites, or specified by the HLA class I or class II motifs available in the art or which become part of the art such as set forth in Table IV (or determined using World Wide Web site URL syfpeithi.bmi-heidelberg.com/) are to be "applied" to the 85P1B3 protein. As used in this context "applied" means that the 85P1B3 protein is evaluated, e.g., visually or by computer-based patterns finding methods, as appreciated by those of skill in the relevant art. Every subsequence of the 85P1B3 of 8, 9, 10, or 11 amino acid residues that bears an HLA Class I motif, or a subsequence of 9 or more amino acid residues that bear an HLA Class II motif are within the scope of the invention.

III.B.) Expression of 85P1B3-related Proteins

In an embodiment described in the examples that follow, 85P1B3 can be conveniently expressed in cells (such as 293T cells) transfected with a commercially available expression vector such as a CMV-driven expression vector encoding 85P1B3 with a C-terminal 6XHis and MYC tag (pcDNA3.1/mycHIS, Invitrogen or Tag5, GenHunter Corporation, Nashville TN). The Tag5 vector provides an IgGK secretion signal that can be used to facilitate the production of a secreted 85P1B3 protein in transfected cells. The secreted HIS-tagged 85P1B3 in the culture media can be purified, e.g., using a nickel column using standard techniques.

III.C.) Modifications of 85P1B3-related Proteins

Modifications of 85P1B3-related proteins such as covalent modifications are included within the scope of this invention. One type of covalent modification includes reacting targeted amino acid residues of a 85P1B3 polypeptide with an organic derivatizing agent that is capable of reacting with selected side chains or the N- or C- terminal residues of the 85P1B3. Another type of covalent modification of the 85P1B3 polypeptide included within the scope of this invention comprises altering the native glycosylation pattern of a protein of the invention. Another type of covalent modification of 85P1B3 comprises linking the 85P1B3 polypeptide to one of a variety of nonproteinaceous polymers, e.g., polyethylene glycol (PEG), polypropylene glycol, or polyoxyalkylenes, in the manner set forth in U.S. Patent Nos. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192 or 4,179,337.

The 85P1B3-related proteins of the present invention can also be modified to form a chimeric molecule comprising 85P1B3 fused to another, heterologous polypeptide or amino acid sequence. Such a chimeric molecule can be synthesized chemically or recombinantly. A chimeric molecule can have a protein of the invention fused to another tumor-associated antigen or fragment thereof. Alternatively, a protein in accordance with the invention can comprise a fusion of fragments of the 85P1B3 sequence (amino or nucleic acid) such that a molecule is created that is not, through its length, directly homologous to the amino or nucleic acid sequences shown in Figure 2 or Figure 3. Such a chimeric molecule can comprise multiples of the same subsequence of 85P1B3. A chimeric molecule can comprise a fusion of a 85P1B3-related protein with a polyhistidine epitope tag, which provides an epitope to which immobilized nickel can selectively bind, with cytokines or with growth factors. The epitope tag is generally placed at the amino- or carboxyl- terminus of the 85P1B3. In an alternative embodiment, the chimeric molecule can comprise a fusion of a 85P1B3-related protein with an immunoglobulin or a particular region of an immunoglobulin. For a bivalent form of the chimeric molecule (also referred to as an "immunoadhesin"), such a fusion could be to the Fc region of an IgG molecule. The Ig fusions preferably include the substitution of a soluble (transmembrane domain deleted or inactivated) form of a 85P1B3 polypeptide in place of at least one variable region within an Ig molecule. In a preferred embodiment, the immunoglobulin fusion includes the hinge, CH2 and CH3, or the hinge, CHI, CH2 and CH3 regions of an IgGI molecule. For the production of immunoglobulin fusions see, e.g., U.S. Patent No. 5,428,130 issued June 27, 1995.

III.D.) Uses of 85P1B3-related Proteins

The proteins of the invention have a number of different specific uses. As 85P1B3 is highly expressed in prostate and other cancers, 85P1B3-related proteins are used in methods that assess the status of 85P1B3 gene products in normal versus cancerous tissues, thereby elucidating the malignant phenotype. Typically, polypeptides from specific regions of the 85P1B3 protein are used to assess the presence of perturbations (such as deletions, insertions, point mutations etc.) in those regions (such as regions

containing one or more motifs). Exemplary assays utilize antibodies or T cells targeting 85P1B3-related proteins comprising the amino acid residues of one or more of the biological motifs contained within the 85P1B3 polypeptide sequence in order to evaluate the characteristics of this region in normal versus cancerous tissues or to elicit an immune response to the epitope. Alternatively, 85P1B3-related proteins that contain the amino acid residues of one or more of the biological motifs in the 85P1B3 protein are used to screen for factors that interact with that region of 85P1B3.

85P1B3 protein fragments/subsequences are particularly useful in generating and characterizing domain-specific antibodies (e.g., antibodies recognizing an extracellular or intracellular epitope of an 85P1B3 protein), for identifying agents or cellular factors that bind to 85P1B3 or a particular structural domain thereof, and in various therapeutic and diagnostic contexts, including but not limited to diagnostic assays, cancer vaccines and methods of preparing such vaccines.

Proteins encoded by the 85P1B3 genes, or by analogs, homologs or fragments thereof, have a variety of uses, including but not limited to generating antibodies and in methods for identifying ligands and other agents and cellular constituents that bind to an 85P1B3 gene product. Antibodies raised against an 85P1B3 protein or fragment thereof are useful in diagnostic and prognostic assays, and imaging methodologies in the management of human cancers characterized by expression of 85P1B3 protein, such as those listed in Table I. Such antibodies can be expressed intracellularly and used in methods of treating patients with such cancers. 85P1B3-related nucleic acids or proteins are also used in generating HTL or CTL responses.

Various immunological assays useful for the detection of 85P1B3 proteins are used, including but not limited to various types of radioimmunoassays, enzyme-linked immunosorbent assays (ELISA), enzyme-linked immunofluorescent assays (ELIFA), immunocytochemical methods, and the like. Antibodies can be labeled and used as immunological imaging reagents capable of detecting 85P1B3-expressing cells (e.g., in radioscintigraphic imaging methods). 85P1B3 proteins are also particularly useful in generating cancer vaccines, as further described herein.

IV.) 85P1B3 Antibodies

Another aspect of the invention provides antibodies that bind to 85P1B3-related proteins. Preferred antibodies specifically bind to a 85P1B3-related protein and do not bind (or bind weakly) to peptides or proteins that are not 85P1B3-related proteins. For example, antibodies bind 85P1B3 can bind 85P1B3-related proteins such as the homologs or analogs thereof.

85P1B3 antibodies of the invention are particularly useful in prostate cancer diagnostic and prognostic assays, and imaging methodologies. Similarly, such antibodies are useful in the treatment, diagnosis, and/or prognosis of other cancers, to the extent 85P1B3 is also expressed or overexpressed in these other cancers. Moreover, intracellularly expressed antibodies (e.g., single chain antibodies) are

therapeutically useful in treating cancers in which the expression of 85P1B3 is involved, such as advanced or metastatic prostate cancers.

The invention also provides various immunological assays useful for the detection and quantification of 85P1B3 and mutant 85P1B3-related proteins. Such assays can comprise one or more 85P1B3 antibodies capable of recognizing and binding a 85P1B3-related protein, as appropriate. These assays are performed within various immunological assay formats well known in the art, including but not limited to various types of radioimmunoassays, enzyme-linked immunosorbent assays (ELISA), enzyme-linked immunofluorescent assays (ELIFA), and the like.

Immunological non-antibody assays of the invention also comprise T cell immunogenicity assays (inhibitory or stimulatory) as well as major histocompatibility complex (MHC) binding assays.

In addition, immunological imaging methods capable of detecting prostate cancer and other cancers expressing 85P1B3 are also provided by the invention, including but not limited to radioscintigraphic imaging methods using labeled 85P1B3 antibodies. Such assays are clinically useful in the detection, monitoring, and prognosis of 85P1B3 expressing cancers such as prostate cancer.

85P1B3 antibodies are also used in methods for purifying a 85P1B3-related protein and for isolating 85P1B3 homologues and related molecules. For example, a method of purifying a 85P1B3-related protein comprises incubating an 85P1B3 antibody, which has been coupled to a solid matrix, with a lysate or other solution containing a 85P1B3-related protein under conditions that permit the 85P1B3 antibody to bind to the 85P1B3-related protein; washing the solid matrix to eliminate impurities; and eluting the 85P1B3-related protein from the coupled antibody. Other uses of the 85P1B3 antibodies of the invention include generating anti-idiotypic antibodies that mimic the 85P1B3 protein.

Various methods for the preparation of antibodies are well known in the art. For example, antibodies can be prepared by immunizing a suitable mammalian host using a 85P1B3-related protein, peptide, or fragment, in isolated or immunoconjugated form (Antibodies: A Laboratory Manual, CSH Press, Eds., Harlow, and Lane (1988); Harlow, Antibodies, Cold Spring Harbor Press, NY (1989)). In addition, fusion proteins of 85P1B3 can also be used, such as a 85P1B3 GST-fusion protein. In a particular embodiment, a GST fusion protein comprising all or most of the amino acid sequence of Figure 2 or Figure 3 is produced, then used as an immunogen to generate appropriate antibodies. In another embodiment, a 85P1B3-related protein is synthesized and used as an immunogen.

In addition, naked DNA immunization techniques known in the art are used (with or without purified 85P1B3-related protein or 85P1B3 expressing cells) to generate an immune response to the encoded immunogen (for review, see Donnelly et al., 1997, Ann. Rev. Immunol. 15: 617-648).

The amino acid sequence of 85P1B3 as shown in Figure 2 or Figure 3 can be analyzed to select specific regions of the 85P1B3 protein for generating antibodies. For example, hydrophobicity and

hydrophilicity analyses of the 85P1B3 amino acid sequence are used to identify hydrophilic regions in the 85P1B3 structure. Regions of the 85P1B3 protein that show immunogenic structure, as well as other regions and domains, can readily be identified using various other methods known in the art, such as Chou-Fasman, Garnier-Robson, Kyte-Doolittle, Eisenberg, Karplus-Schultz or Jameson-Wolf analysis. Thus, each region identified by any of these programs or methods is within the scope of the present invention. Methods for the generation of 85P1B3 antibodies are further illustrated by way of the examples provided herein. Methods for preparing a protein or polypeptide for use as an immunogen are well known in the art. Also well known in the art are methods for preparing immunogenic conjugates of a protein with a carrier, such as BSA, KLH or other carrier protein. In some circumstances, direct conjugation using, for example, carbodiimide reagents are used; in other instances linking reagents such as those supplied by Pierce Chemical Co., Rockford, IL, are effective. Administration of a 85P1B3 immunogen is often conducted by injection over a suitable time period and with use of a suitable adjuvant, as is understood in the art. During the immunization schedule, titers of antibodies can be taken to determine adequacy of antibody formation.

85P1B3 monoclonal antibodies can be produced by various means well known in the art. For example, immortalized cell lines that secrete a desired monoclonal antibody are prepared using the standard hybridoma technology of Kohler and Milstein or modifications that immortalize antibody-producing B cells, as is generally known. Immortalized cell lines that secrete the desired antibodies are screened by immunoassay in which the antigen is a 85P1B3-related protein. When the appropriate immortalized cell culture is identified, the cells can be expanded and antibodies produced either from *in vitro* cultures or from ascites fluid.

The antibodies or fragments of the invention can also be produced, by recombinant means. Regions that bind specifically to the desired regions of the 85P1B3 protein can also be produced in the context of chimeric or complementarity determining region (CDR) grafted antibodies of multiple species origin. Humanized or human 85P1B3 antibodies can also be produced, and are preferred for use in therapeutic contexts. Methods for humanizing murine and other non-human antibodies, by substituting one or more of the non-human antibody CDRs for corresponding human antibody sequences, are well known (see for example, Jones et al., 1986, Nature 321: 522-525; Riechmann et al., 1988, Nature 332: 323-327; Verhoeyen et al., 1988, Science 239: 1534-1536). See also, Carter et al., 1993, Proc. Natl. Acad. Sci. USA 89: 4285 and Sims et al., 1993, J. Immunol. 151: 2296.

Methods for producing fully human monoclonal antibodies include phage display and transgenic methods (for review, see Vaughan et al., 1998, Nature Biotechnology 16: 535-539). Fully human 85P1B3 monoclonal antibodies can be generated using cloning technologies employing large human Ig gene combinatorial libraries (i.e., phage display) (Griffiths and Hoogenboom, Building an *in vitro* immune system: human antibodies from phage display libraries. In: Protein Engineering of Antibody Molecules for Prophylactic and Therapeutic Applications in Man, Clark, M. (Ed.), Nottingham Academic, pp 45-64 (1993);

Burton and Barbas, Human Antibodies from combinatorial libraries. Id., pp 65-82). Fully human 85P1B3 monoclonal antibodies can also be produced using transgenic mice engineered to contain human immunoglobulin gene loci as described in PCT Patent Application WO98/24893, Kucherlapati and Jakobovits et al., published December 3, 1997 (see also, Jakobovits, 1998, Exp. Opin. Invest. Drugs 7(4): 607-614; U.S. patents 6,162,963 issued 19 December 2000; 6,150,584 issued 12 November 2000; and, 6,114598 issued 5 September 2000). This method avoids the *in vitro* manipulation required with phage display technology and efficiently produces high affinity authentic human antibodies.

Reactivity of 85P1B3 antibodies with an 85P1B3-related protein can be established by a number of well known means, including Western blot, immunoprecipitation, ELISA, and FACS analyses using, as appropriate, 85P1B3-related proteins, 85P1B3-expressing cells or extracts thereof. A 85P1B3 antibody or fragment thereof can be labeled with a detectable marker or conjugated to a second molecule. Suitable detectable markers include, but are not limited to, a radioisotope, a fluorescent compound, a bioluminescent compound, chemiluminescent compound, a metal chelator or an enzyme. Further, bi-specific antibodies specific for two or more 85P1B3 epitopes are generated using methods generally known in the art. Homodimeric antibodies can also be generated by cross-linking techniques known in the art (e.g., Wolff et al., Cancer Res. 53: 2560-2565).

V.) 85P1B3 Cellular Immune Responses

The mechanism by which T cells recognize antigens has been delineated. Efficacious peptide epitope vaccine compositions of the invention induce a therapeutic or prophylactic immune responses in very broad segments of the world-wide population. For an understanding of the value and efficacy of compositions of the invention that induce cellular immune responses, a brief review of immunology-related technology is provided.

A complex of an HLA molecule and a peptidic antigen acts as the ligand recognized by HLA-restricted T cells (Buus, S. *et al.*, *Cell* 47:1071, 1986; Babbitt, B. P. *et al.*, *Nature* 317:359, 1985; Townsend, A. and Bodmer, H., *Annu. Rev. Immunol.* 7:601, 1989; Germain, R. N., *Annu. Rev. Immunol.* 11:403, 1993). Through the study of single amino acid substituted antigen analogs and the sequencing of endogenously bound, naturally processed peptides, critical residues that correspond to motifs required for specific binding to HLA antigen molecules have been identified and are set forth in Table IV (*see* also, *e.g.*, Southwood, *et al.*, *J. Immunol.* 160:3363, 1998; Rammensee, *et al.*, *Immunogenetics* 41:178, 1995; Rammensee *et al.*, SYFPEITHI, access via World Wide Web at URL syfpeithi.bmi-heidelberg.com/; Sette, A. and Sidney, J. *Curr. Opin. Immunol.* 10:478, 1998; Engelhard, V. H., *Curr. Opin. Immunol.* 6:13, 1994; Sette, A. and Grey, H. M., *Curr. Opin. Immunol.* 4:79, 1992; Sinigaglia, F. and Hammer, J. *Curr. Biol.* 6:52, 1994; Ruppert *et al.*, *Cell* 74:929-937, 1993; Kondo *et al.*, *J. Immunol.* 155:4307-4312, 1995; Sidney

et al., *J. Immunol.* 157:3480-3490, 1996; Sidney *et al.*, *Human Immunol.* 45:79-93, 1996; Sette, A. and Sidney, J. *Immunogenetics 1999 Nov; 50(3-4):201-12,* Review).

Furthermore, x-ray crystallographic analyses of HLA-peptide complexes have revealed pockets within the peptide binding cleft/groove of HLA molecules which accommodate, in an allele-specific mode, residues borne by peptide ligands; these residues in turn determine the HLA binding capacity of the peptides in which they are present. (*See, e.g.*, Madden, D.R. *Annu. Rev. Immunol.* 13:587, 1995; Smith, *et al.*, *Immunity* 4:203, 1996; Fremont *et al.*, *Immunity* 8:305, 1998; Stern *et al.*, *Structure* 2:245, 1994; Jones, E.Y. *Curr. Opin. Immunol.* 9:75, 1997; Brown, J. H. *et al.*, *Nature* 364:33, 1993; Guo, H. C. *et al.*, *Proc. Natl. Acad. Sci. USA* 90:8053, 1993; Guo, H. C. *et al.*, *Nature* 360:364, 1992; Silver, M. L. *et al.*, *Nature* 360:367, 1992; Matsumura, M. *et al.*, *Science* 257:927, 1992; Madden *et al.*, *Cell* 70:1035, 1992; Fremont, D. H. *et al.*, *Science* 257:919, 1992; Saper, M. A. , Bjorkman, P. J. and Wiley, D. C., *J. Mol. Biol.* 219:277, 1991.)

Accordingly, the definition of class I and class II allele-specific HLA binding motifs, or class I or class II supermotifs allows identification of regions within a protein that are correlated with binding to particular HLA antigen(s).

Thus, by a process of HLA motif identification, candidates for epitope-based vaccines have been identified; such candidates can be further evaluated by HLA-peptide binding assays to determine binding affinity and/or the time period of association of the epitope and its corresponding HLA molecule. Additional confirmatory work can be performed to select, amongst these vaccine candidates, epitopes with preferred characteristics in terms of population coverage, and/or immunogenicity.

Various strategies can be utilized to evaluate cellular immunogenicity, including:

1) Evaluation of primary T cell cultures from normal individuals (*see, e.g.*, Wentworth, P. A. *et al.*, *Mol. Immunol.* 32:603, 1995; Celis, E. *et al.*, *Proc. Natl. Acad. Sci. USA* 91:2105, 1994; Tsai, V. *et al.*, *J. Immunol.* 158:1796, 1997; Kawashima, I. *et al.*, *Human Immunol.* 59:1, 1998). This procedure involves the stimulation of peripheral blood lymphocytes (PBL) from normal subjects with a test peptide in the presence of antigen presenting cells *in vitro* over a period of several weeks. T cells specific for the peptide become activated during this time and are detected using, *e.g.*, a lymphokine- or ^{51}Cr-release assay involving peptide sensitized target cells.

2) Immunization of HLA transgenic mice (*see, e.g.*, Wentworth, P. A. *et al.*, *J. Immunol.* 26:97, 1996; Wentworth, P. A. *et al.*, *Int. Immunol.* 8:651, 1996; Alexander, J. *et al.*, *J. Immunol.* 159:4753, 1997). For example, in such methods peptides in incomplete Freund's adjuvant are administered subcutaneously to HLA transgenic mice. Several weeks following immunization, splenocytes are removed and cultured *in vitro* in the presence of test peptide for approximately one week. Peptide-specific T cells are detected

using, *e.g.*, a ^{51}Cr-release assay involving peptide sensitized target cells and target cells expressing endogenously generated antigen.

3) Demonstration of recall T cell responses from immune individuals who have been either effectively vaccinated and/or from chronically ill patients (*see, e.g.*, Rehermann, B. *et al.*, *J. Exp. Med.* 181:1047, 1995; Doolan, D. L. *et al.*, *Immunity* 7:97, 1997; Bertoni, R. *et al.*, *J. Clin. Invest.* 100:503, 1997; Threlkeld, S. C. *et al.*, *J. Immunol.* 159:1648, 1997; Diepolder, H. M. *et al.*, *J. Virol.* 71:6011, 1997). Accordingly, recall responses are detected by culturing PBL from subjects that have been exposed to the antigen due to disease and thus have generated an immune response "naturally", or from patients who were vaccinated against the antigen. PBL from subjects are cultured *in vitro* for 1-2 weeks in the presence of test peptide plus antigen presenting cells (APC) to allow activation of "memory" T cells, as compared to "naive" T cells. At the end of the culture period, T cell activity is detected using assays including ^{51}Cr release involving peptide-sensitized targets, T cell proliferation, or lymphokine release.

VI.) 85P1B3 Transgenic Animals

Nucleic acids that encode a 85P1B3-related protein can also be used to generate either transgenic animals or "knock out" animals which, in turn, are useful in the development and screening of therapeutically useful reagents. In accordance with established techniques, cDNA encoding 85P1B3 can be used to clone genomic DNA that encodes 85P1B3. The cloned genomic sequences can then be used to generate transgenic animals containing cells that express DNA that encode 85P1B3. Methods for generating transgenic animals, particularly animals such as mice or rats, have become conventional in the art and are described, for example, in U.S. Patent Nos. 4,736,866 issued 12 April 1988, and 4,870,009 issued 26 September 1989. Typically, particular cells would be targeted for 85P1B3 transgene incorporation with tissue-specific enhancers.

Transgenic animals that include a copy of a transgene encoding 85P1B3 can be used to examine the effect of increased expression of DNA that encodes 85P1B3. Such animals can be used as tester animals for reagents thought to confer protection from, for example, pathological conditions associated with its overexpression. In accordance with this aspect of the invention, an animal is treated with a reagent and a reduced incidence of a pathological condition, compared to untreated animals that bear the transgene, would indicate a potential therapeutic intervention for the pathological condition.

Alternatively, non-human homologues of 85P1B3 can be used to construct a 85P1B3 "knock out" animal that has a defective or altered gene encoding 85P1B3 as a result of homologous recombination between the endogenous gene encoding 85P1B3 and altered genomic DNA encoding 85P1B3 introduced into an embryonic cell of the animal. For example, cDNA that encodes 85P1B3 can be used to clone

genomic DNA encoding 85P1B3 in accordance with established techniques. A portion of the genomic DNA encoding 85P1B3 can be deleted or replaced with another gene, such as a gene encoding a selectable marker that can be used to monitor integration. Typically, several kilobases of unaltered flanking DNA (both at the 5' and 3' ends) are included in the vector (see, e.g., Thomas and Capecchi, Cell, 51:503 (1987) for a description of homologous recombination vectors). The vector is introduced into an embryonic stem cell line (e.g., by electroporation) and cells in which the introduced DNA has homologously recombined with the endogenous DNA are selected (see, e.g.,, Li et al., Cell, 69:915 (1992)). The selected cells are then injected into a blastocyst of an animal (e.g., a mouse or rat) to form aggregation chimeras (see, e.g.,, Bradley, in *Teratocarcinomas and Embryonic Stem Cells: A Practical Approach*, E. J. Robertson, ed. (IRL, Oxford, 1987), pp. 113-152). A chimeric embryo can then be implanted into a suitable pseudopregnant female foster animal, and the embryo brought to term to create a "knock out" animal. Progeny harboring the homologously recombined DNA in their germ cells can be identified by standard techniques and used to breed animals in which all cells of the animal contain the homologously recombined DNA. Knock out animals can be characterized, for example, for their ability to defend against certain pathological conditions or for their development of pathological conditions due to absence of the 85P1B3 polypeptide.

VII.) Methods for the Detection of 85P1B3

Another aspect of the present invention relates to methods for detecting 85P1B3 polynucleotides and 85P1B3-related proteins, as well as methods for identifying a cell that expresses 85P1B3. The expression profile of 85P1B3 makes it a diagnostic marker for metastasized disease. Accordingly, the status of 85P1B3 gene products provides information useful for predicting a variety of factors including susceptibility to advanced stage disease, rate of progression, and/or tumor aggressiveness. As discussed in detail herein, the status of 85P1B3 gene products in patient samples can be analyzed by a variety protocols that are well known in the art including immunohistochemical analysis, the variety of Northern blotting techniques including *in situ* hybridization, RT-PCR analysis (for example on laser capture micro-dissected samples), Western blot analysis and tissue array analysis.

More particularly, the invention provides assays for the detection of 85P1B3 polynucleotides in a biological sample, such as serum, bone, prostate, and other tissues, urine, semen, cell preparations, and the like. Detectable 85P1B3 polynucleotides include, for example, a 85P1B3 gene or fragment thereof, 85P1B3 mRNA, alternative splice variant 85P1B3 mRNAs, and recombinant DNA or RNA molecules that contain a 85P1B3 polynucleotide. A number of methods for amplifying and/or detecting the presence of 85P1B3 polynucleotides are well known in the art and can be employed in the practice of this aspect of the invention.

In one embodiment, a method for detecting an 85P1B3 mRNA in a biological sample comprises producing cDNA from the sample by reverse transcription using at least one primer; amplifying the cDNA

so produced using an 85P1B3 polynucleotides as sense and antisense primers to amplify 85P1B3 cDNAs therein; and detecting the presence of the amplified 85P1B3 cDNA. Optionally, the sequence of the amplified 85P1B3 cDNA can be determined.

In another embodiment, a method of detecting a 85P1B3 gene in a biological sample comprises first isolating genomic DNA from the sample; amplifying the isolated genomic DNA using 85P1B3 polynucleotides as sense and antisense primers; and detecting the presence of the amplified 85P1B3 gene. Any number of appropriate sense and antisense probe combinations can be designed from the nucleotide sequence provided for the 85P1B3 (Figure 2) and used for this purpose.

The invention also provides assays for detecting the presence of an 85P1B3 protein in a tissue or other biological sample such as serum, semen, bone, prostate, urine, cell preparations, and the like. Methods for detecting a 85P1B3-related protein are also well known and include, for example, immunoprecipitation, immunohistochemical analysis, Western blot analysis, molecular binding assays, ELISA, ELIFA and the like. For example, a method of detecting the presence of a 85P1B3-related protein in a biological sample comprises first contacting the sample with a 85P1B3 antibody, a 85P1B3-reactive fragment thereof, or a recombinant protein containing an antigen binding region of a 85P1B3 antibody; and then detecting the binding of 85P1B3-related protein in the sample.

Methods for identifying a cell that expresses 85P1B3 are also within the scope of the invention. In one embodiment, an assay for identifying a cell that expresses a 85P1B3 gene comprises detecting the presence of 85P1B3 mRNA in the cell. Methods for the detection of particular mRNAs in cells are well known and include, for example, hybridization assays using complementary DNA probes (such as *in situ* hybridization using labeled 85P1B3 riboprobes, Northern blot and related techniques) and various nucleic acid amplification assays (such as RT-PCR using complementary primers specific for 85P1B3, and other amplification type detection methods, such as, for example, branched DNA, SISBA, TMA and the like). Alternatively, an assay for identifying a cell that expresses a 85P1B3 gene comprises detecting the presence of 85P1B3-related protein in the cell or secreted by the cell. Various methods for the detection of proteins are well known in the art and are employed for the detection of 85P1B3-related proteins and cells that express 85P1B3-related proteins.

85P1B3 expression analysis is also useful as a tool for identifying and evaluating agents that modulate 85P1B3 gene expression. For example, 85P1B3 expression is significantly upregulated in prostate cancer, and is expressed in cancers of the tissues listed in Table I. Identification of a molecule or biological agent that inhibits 85P1B3 expression or over-expression in cancer cells is of therapeutic value. For example, such an agent can be identified by using a screen that quantifies 85P1B3 expression by RT-PCR, nucleic acid hybridization or antibody binding.

VIII.) Methods for Monitoring the Status of 85P1B3-related Genes and Their Products

Oncogenesis is known to be a multistep process where cellular growth becomes progressively dysregulated and cells progress from a normal physiological state to precancerous and then cancerous states (see, e.g., Alers et al., Lab Invest. 77(5): 437-438 (1997) and Isaacs et al., Cancer Surv. 23: 19-32 (1995)). In this context, examining a biological sample for evidence of dysregulated cell growth (such as aberrant 85P1B3 expression in cancers) allows for early detection of such aberrant physiology, before a pathologic state such as cancer has progressed to a stage that therapeutic options are more limited and or the prognosis is worse. In such examinations, the status of 85P1B3 in a biological sample of interest can be compared, for example, to the status of 85P1B3 in a corresponding normal sample (e.g. a sample from that individual or alternatively another individual that is not affected by a pathology). An alteration in the status of 85P1B3 in the biological sample (as compared to the normal sample) provides evidence of dysregulated cellular growth. In addition to using a biological sample that is not affected by a pathology as a normal sample, one can also use a predetermined normative value such as a predetermined normal level of mRNA expression (see, e.g., Grever et al., J. Comp. Neurol. 1996 Dec 9;376(2):306-14 and U.S. Patent No. 5,837,501) to compare 85P1B3 status in a sample.

The term "status" in this context is used according to its art accepted meaning and refers to the condition or state of a gene and its products. Typically, skilled artisans use a number of parameters to evaluate the condition or state of a gene and its products. These include, but are not limited to the location of expressed gene products (including the location of 85P1B3 expressing cells) as well as the level, and biological activity of expressed gene products (such as 85P1B3 mRNA, polynucleotides and polypeptides). Typically, an alteration in the status of 85P1B3 comprises a change in the location of 85P1B3 and/or 85P1B3 expressing cells and/or an increase in 85P1B3 mRNA and/or protein expression.

85P1B3 status in a sample can be analyzed by a number of means well known in the art, including without limitation, immunohistochemical analysis, *in situ* hybridization, RT-PCR analysis on laser capture micro-dissected samples, Western blot analysis, and tissue array analysis. Typical protocols for evaluating the status of the 85P1B3 gene and gene products are found, for example in Ausubel et al. eds., 1995, Current Protocols In Molecular Biology, Units 2 (Northern Blotting), 4 (Southern Blotting), 15 (Immunoblotting) and 18 (PCR Analysis). Thus, the status of 85P1B3 in a biological sample is evaluated by various methods utilized by skilled artisans including, but not limited to genomic Southern analysis (to examine, for example perturbations in the 85P1B3 gene), Northern analysis and/or PCR analysis of 85P1B3 mRNA (to examine, for example alterations in the polynucleotide sequences or expression levels of 85P1B3 mRNAs), and, Western and/or immunohistochemical analysis (to examine, for example alterations in polypeptide sequences, alterations in polypeptide localization within a sample, alterations in expression levels of 85P1B3 proteins and/or associations of 85P1B3 proteins with polypeptide binding partners). Detectable 85P1B3 polynucleotides include, for example, a 85P1B3 gene or fragment thereof, 85P1B3 mRNA, alternative

splice variants, 85P1B3 mRNAs, and recombinant DNA or RNA molecules containing a 85P1B3 polynucleotide.

The expression profile of 85P1B3 makes it a diagnostic marker for local and/or metastasized disease, and provides information on the growth or oncogenic potential of a biological sample. In particular, the status of 85P1B3 provides information useful for predicting susceptibility to particular disease stages, progression, and/or tumor aggressiveness. The invention provides methods and assays for determining 85P1B3 status and diagnosing cancers that express 85P1B3, such as cancers of the tissues listed in Table I. For example, because 85P1B3 mRNA is so highly expressed in prostate and other cancers relative to normal prostate tissue, assays that evaluate the levels of 85P1B3 mRNA transcripts or proteins in a biological sample can be used to diagnose a disease associated with 85P1B3 dysregulation, and can provide prognostic information useful in defining appropriate therapeutic options.

The expression status of 85P1B3 provides information including the presence, stage and location of dysplastic, precancerous and cancerous cells, predicting susceptibility to various stages of disease, and/or for gauging tumor aggressiveness. Moreover, the expression profile makes it useful as an imaging reagent for metastasized disease. Consequently, an aspect of the invention is directed to the various molecular prognostic and diagnostic methods for examining the status of 85P1B3 in biological samples such as those from individuals suffering from, or suspected of suffering from a pathology characterized by dysregulated cellular growth, such as cancer.

As described above, the status of 85P1B3 in a biological sample can be examined by a number of well-known procedures in the art. For example, the status of 85P1B3 in a biological sample taken from a specific location in the body can be examined by evaluating the sample for the presence or absence of 85P1B3 expressing cells (e.g. those that express 85P1B3 mRNAs or proteins). This examination can provide evidence of dysregulated cellular growth, for example, when 85P1B3-expressing cells are found in a biological sample that does not normally contain such cells (such as a lymph node), because such alterations in the status of 85P1B3 in a biological sample are often associated with dysregulated cellular growth. Specifically, one indicator of dysregulated cellular growth is the metastases of cancer cells from an organ of origin (such as the prostate) to a different area of the body (such as a lymph node). In this context, evidence of dysregulated cellular growth is important for example because occult lymph node metastases can be detected in a substantial proportion of patients with prostate cancer, and such metastases are associated with known predictors of disease progression (see, e.g., Murphy et al., Prostate 42(4): 315-317 (2000);Su et al., Semin. Surg. Oncol. 18(1): 17-28 (2000) and Freeman et al., J Urol 1995 Aug 154(2 Pt 1):474-8).

In one aspect, the invention provides methods for monitoring 85P1B3 gene products by determining the status of 85P1B3 gene products expressed by cells from an individual suspected of having

a disease associated with dysregulated cell growth (such as hyperplasia or cancer) and then comparing the status so determined to the status of 85P1B3 gene products in a corresponding normal sample. The presence of aberrant 85P1B3 gene products in the test sample relative to the normal sample provides an indication of the presence of dysregulated cell growth within the cells of the individual.

In another aspect, the invention provides assays useful in determining the presence of cancer in an individual, comprising detecting a significant increase in 85P1B3 mRNA or protein expression in a test cell or tissue sample relative to expression levels in the corresponding normal cell or tissue. The presence of 85P1B3 mRNA can, for example, be evaluated in tissue samples including but not limited to those listed in Table I. The presence of significant 85P1B3 expression in any of these tissues is useful to indicate the emergence, presence and/or severity of a cancer, since the corresponding normal tissues do not express 85P1B3 mRNA or express it at lower levels.

In a related embodiment, 85P1B3 status is determined at the protein level rather than at the nucleic acid level. For example, such a method comprises determining the level of 85P1B3 protein expressed by cells in a test tissue sample and comparing the level so determined to the level of 85P1B3 expressed in a corresponding normal sample. In one embodiment, the presence of 85P1B3 protein is evaluated, for example, using immunohistochemical methods. 85P1B3 antibodies or binding partners capable of detecting 85P1B3 protein expression are used in a variety of assay formats well known in the art for this purpose.

In a further embodiment, one can evaluate the status of 85P1B3 nucleotide and amino acid sequences in a biological sample in order to identify perturbations in the structure of these molecules. These perturbations can include insertions, deletions, substitutions and the like. Such evaluations are useful because perturbations in the nucleotide and amino acid sequences are observed in a large number of proteins associated with a growth dysregulated phenotype (see, e.g., Marrogi et al., 1999, J. Cutan. Pathol. 26(8):369-378). For example, a mutation in the sequence of 85P1B3 may be indicative of the presence or promotion of a tumor. Such assays therefore have diagnostic and predictive value where a mutation in 85P1B3 indicates a potential loss of function or increase in tumor growth.

A wide variety of assays for observing perturbations in nucleotide and amino acid sequences are well known in the art. For example, the size and structure of nucleic acid or amino acid sequences of 85P1B3 gene products are observed by the Northern, Southern, Western, PCR and DNA sequencing protocols discussed herein. In addition, other methods for observing perturbations in nucleotide and amino acid sequences such as single strand conformation polymorphism analysis are well known in the art (see, e.g., U.S. Patent Nos. 5,382,510 issued 7 September 1999, and 5,952,170 issued 17 January 1995).

Additionally, one can examine the methylation status of the 85P1B3 gene in a biological sample. Aberrant demethylation and/or hypermethylation of CpG islands in gene 5' regulatory regions frequently occurs in immortalized and transformed cells, and can result in altered expression of various genes. For

example, promoter hypermethylation of the pi-class glutathione S-transferase (a protein expressed in normal prostate but not expressed in >90% of prostate carcinomas) appears to permanently silence transcription of this gene and is the most frequently detected genomic alteration in prostate carcinomas (De Marzo et al., Am. J. Pathol. 155(6): 1985-1992 (1999)). In addition, this alteration is present in at least 70% of cases of high-grade prostatic intraepithelial neoplasia (PIN) (Brooks et al, Cancer Epidemiol. Biomarkers Prev., 1998, 7:531-536). In another example, expression of the LAGE-I tumor specific gene (which is not expressed in normal prostate but is expressed in 25-50% of prostate cancers) is induced by deoxy-azacytidine in lymphoblastoid cells, suggesting that tumoral expression is due to demethylation (Lethe et al., Int. J. Cancer 76(6): 903-908 (1998)). A variety of assays for examining methylation status of a gene are well known in the art. For example, one can utilize, in Southern hybridization approaches, methylation-sensitive restriction enzymes which cannot cleave sequences that contain methylated CpG sites to assess the methylation status of CpG islands. In addition, MSP (methylation specific PCR) can rapidly profile the methylation status of all the CpG sites present in a CpG island of a given gene. This procedure involves initial modification of DNA by sodium bisulfite (which will convert all unmethylated cytosines to uracil) followed by amplification using primers specific for methylated versus unmethylated DNA. Protocols involving methylation interference can also be found for example in Current Protocols In Molecular Biology, Unit 12, Frederick M. Ausubel et al. eds., 1995.

Gene amplification is an additional method for assessing the status of 85P1B3. Gene amplification is measured in a sample directly, for example, by conventional Southern blotting or Northern blotting to quantitate the transcription of mRNA (Thomas, 1980, Proc. Natl. Acad. Sci. USA, 77:5201-5205), dot blotting (DNA analysis), or *in situ* hybridization, using an appropriately labeled probe, based on the sequences provided herein. Alternatively, antibodies are employed that recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. The antibodies in turn are labeled and the assay carried out where the duplex is bound to a surface, so that upon the formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Biopsied tissue or peripheral blood can be conveniently assayed for the presence of cancer cells using for example, Northern, dot blot or RT-PCR analysis to detect 85P1B3 expression. The presence of RT-PCR amplifiable 85P1B3 mRNA provides an indication of the presence of cancer. RT-PCR assays are well known in the art. RT-PCR detection assays for tumor cells in peripheral blood are currently being evaluated for use in the diagnosis and management of a number of human solid tumors. In the prostate cancer field, these include RT-PCR assays for the detection of cells expressing PSA and PSM (Verkaik et al., 1997, Urol. Res. 25:373-384; Ghossein et al., 1995, J. Clin. Oncol. 13:1195-2000; Heston et al., 1995, Clin. Chem. 41:1687-1688).

A further aspect of the invention is an assessment of the susceptibility that an individual has for developing cancer. In one embodiment, a method for predicting susceptibility to cancer comprises detecting

85P1B3 mRNA or 85P1B3 protein in a tissue sample, its presence indicating susceptibility to cancer, wherein the degree of 85P1B3 mRNA expression correlates to the degree of susceptibility. In a specific embodiment, the presence of 85P1B3 in prostate or other tissue is examined, with the presence of 85P1B3 in the sample providing an indication of prostate cancer susceptibility (or the emergence or existence of a prostate tumor). Similarly, one can evaluate the integrity 85P1B3 nucleotide and amino acid sequences in a biological sample, in order to identify perturbations in the structure of these molecules such as insertions, deletions, substitutions and the like. The presence of one or more perturbations in 85P1B3 gene products in the sample is an indication of cancer susceptibility (or the emergence or existence of a tumor).

The invention also comprises methods for gauging tumor aggressiveness. In one embodiment, a method for gauging aggressiveness of a tumor comprises determining the level of 85P1B3 mRNA or 85P1B3 protein expressed by tumor cells, comparing the level so determined to the level of 85P1B3 mRNA or 85P1B3 protein expressed in a corresponding normal tissue taken from the same individual or a normal tissue reference sample, wherein the degree of 85P1B3 mRNA or 85P1B3 protein expression in the tumor sample relative to the normal sample indicates the degree of aggressiveness. In a specific embodiment, aggressiveness of a tumor is evaluated by determining the extent to which 85P1B3 is expressed in the tumor cells, with higher expression levels indicating more aggressive tumors. Another embodiment is the evaluation of the integrity of 85P1B3 nucleotide and amino acid sequences in a biological sample, in order to identify perturbations in the structure of these molecules such as insertions, deletions, substitutions and the like. The presence of one or more perturbations indicates more aggressive tumors.

Another embodiment of the invention is directed to methods for observing the progression of a malignancy in an individual over time. In one embodiment, methods for observing the progression of a malignancy in an individual over time comprise determining the level of 85P1B3 mRNA or 85P1B3 protein expressed by cells in a sample of the tumor, comparing the level so determined to the level of 85P1B3 mRNA or 85P1B3 protein expressed in an equivalent tissue sample taken from the same individual at a different time, wherein the degree of 85P1B3 mRNA or 85P1B3 protein expression in the tumor sample over time provides information on the progression of the cancer. In a specific embodiment, the progression of a cancer is evaluated by determining 85P1B3 expression in the tumor cells over time, where increased expression over time indicates a progression of the cancer. Also, one can evaluate the integrity 85P1B3 nucleotide and amino acid sequences in a biological sample in order to identify perturbations in the structure of these molecules such as insertions, deletions, substitutions and the like, where the presence of one or more perturbations indicates a progression of the cancer.

The above diagnostic approaches can be combined with any one of a wide variety of prognostic and diagnostic protocols known in the art. For example, another embodiment of the invention is directed to methods for observing a coincidence between the expression of 85P1B3 gene and 85P1B3 gene products (or

perturbations in 85P1B3 gene and 85P1B3 gene products) and a factor that is associated with malignancy, as a means for diagnosing and prognosticating the status of a tissue sample. A wide variety of factors associated with malignancy can be utilized, such as the expression of genes associated with malignancy (e.g. PSA, PSCA and PSM expression for prostate cancer etc.) as well as gross cytological observations (see, e.g., Bocking et al., 1984, Anal. Quant. Cytol. 6(2):74-88; Epstein, 1995, Hum. Pathol. 26(2):223-9; Thorson et al., 1998, Mod. Pathol. 11(6):543-51; Baisden et al., 1999, Am. J. Surg. Pathol. 23(8):918-24). Methods for observing a coincidence between the expression of 85P1B3 gene and 85P1B3 gene products (or perturbations in 85P1B3 gene and 85P1B3 gene products) and another factor that is associated with malignancy are useful, for example, because the presence of a set of specific factors that coincide with disease provides information crucial for diagnosing and prognosticating the status of a tissue sample.

In one embodiment, methods for observing a coincidence between the expression of 85P1B3 gene and 85P1B3 gene products (or perturbations in 85P1B3 gene and 85P1B3 gene products) and another factor associated with malignancy entails detecting the overexpression of 85P1B3 mRNA or protein in a tissue sample, detecting the overexpression of PSA mRNA or protein in a tissue sample (or PSCA or PSM expression), and observing a coincidence of 85P1B3 mRNA or protein and PSA mRNA or protein overexpression (or PSCA or PSM expression). In a specific embodiment, the expression of 85P1B3 and PSA mRNA in prostate tissue is examined, where the coincidence of 85P1B3 and PSA mRNA overexpression in the sample indicates the existence of prostate cancer, prostate cancer susceptibility or the emergence or status of a prostate tumor.

Methods for detecting and quantifying the expression of 85P1B3 mRNA or protein are described herein, and standard nucleic acid and protein detection and quantification technologies are well known in the art. Standard methods for the detection and quantification of 85P1B3 mRNA include *in situ* hybridization using labeled 85P1B3 riboprobes, Northern blot and related techniques using 85P1B3 polynucleotide probes, RT-PCR analysis using primers specific for 85P1B3, and other amplification type detection methods, such as, for example, branched DNA, SISBA, TMA and the like. In a specific embodiment, semi-quantitative RT-PCR is used to detect and quantify 85P1B3 mRNA expression. Any number of primers capable of amplifying 85P1B3 can be used for this purpose, including but not limited to the various primer sets specifically described herein. In a specific embodiment, polyclonal or monoclonal antibodies specifically reactive with the wild-type 85P1B3 protein can be used in an immunohistochemical assay of biopsied tissue.

IX.) Identification of Molecules That Interact With 85P1B3

The 85P1B3 protein and nucleic acid sequences disclosed herein allow a skilled artisan to identify proteins, small molecules and other agents that interact with 85P1B3, as well as pathways activated by 85P1B3 via any one of a variety of art accepted protocols. For example, one can utilize one of the so-

called interaction trap systems (also referred to as the "two-hybrid assay"). In such systems, molecules interact and reconstitute a transcription factor which directs expression of a reporter gene, whereupon the expression of the reporter gene is assayed. Other systems identify protein-protein interactions *in vivo* through reconstitution of a eukaryotic transcriptional activator, see, e.g., U.S. Patent Nos. 5,955,280 issued 21 September 1999, 5,925,523 issued 20 July 1999, 5,846,722 issued 8 December 1998 and 6,004,746 issued 21 December 1999. Algorithms are also available in the art for genome-based predictions of protein function (see, e.g., Marcotte, et al., Nature 402: 4 November 1999, 83-86).

Alternatively one can screen peptide libraries to identify molecules that interact with 85P1B3 protein sequences. In such methods, peptides that bind to 85P1B3 are identified by screening libraries that encode a random or controlled collection of amino acids. Peptides encoded by the libraries are expressed as fusion proteins of bacteriophage coat proteins, the bacteriophage particles are then screened against the 85P1B3 protein.

Accordingly, peptides having a wide variety of uses, such as therapeutic, prognostic or diagnostic reagents, are thus identified without any prior information on the structure of the expected ligand or receptor molecule. Typical peptide libraries and screening methods that can be used to identify molecules that interact with 85P1B3 protein sequences are disclosed for example in U.S. Patent Nos. 5,723,286 issued 3 March 1998 and 5,733,731 issued 31 March 1998.

Alternatively, cell lines that express 85P1B3 are used to identify protein-protein interactions mediated by 85P1B3. Such interactions can be examined using immunoprecipitation techniques (see, e.g., Hamilton BJ, et al. Biochem. Biophys. Res. Commun. 1999, 261:646-51). 85P1B3 protein can be immunoprecipitated from 85P1B3-expressing cell lines using anti-85P1B3 antibodies. Alternatively, antibodies against His-tag can be used in a cell line engineered to express fusions of 85P1B3 and a His-tag (vectors mentioned above). The immunoprecipitated complex can be examined for protein association by procedures such as Western blotting, ^{35}S-methionine labeling of proteins, protein microsequencing, silver staining and two-dimensional gel electrophoresis.

Small molecules and ligands that interact with 85P1B3 can be identified through related embodiments of such screening assays. For example, small molecules can be identified that interfere with protein function, including molecules that interfere with 85P1B3's ability to mediate phosphorylation and de-phosphorylation, interaction with DNA or RNA molecules as an indication of regulation of cell cycles, second messenger signaling or tumorigenesis. Similarly, small molecules that modulate 85P1B3-related ion channel, protein pump, or cell communication functions are identified and used to treat patients that have a cancer that expresses 85P1B3 (see, e.g., Hille, B., Ionic Channels of Excitable Membranes 2nd Ed., Sinauer Assoc., Sunderland, MA, 1992). Moreover, ligands that regulate 85P1B3 function can be identified based on their ability to bind 85P1B3 and activate a reporter construct. Typical methods are

discussed for example in U.S. Patent No. 5,928,868 issued 27 July 1999, and include methods for forming hybrid ligands in which at least one ligand is a small molecule. In an illustrative embodiment, cells engineered to express a fusion protein of 85P1B3 and a DNA-binding protein are used to co-express a fusion protein of a hybrid ligand/small molecule and a cDNA library transcriptional activator protein. The cells further contain a reporter gene, the expression of which is conditioned on the proximity of the first and second fusion proteins to each other, an event that occurs only if the hybrid ligand binds to target sites on both hybrid proteins. Those cells that express the reporter gene are selected and the unknown small molecule or the unknown ligand is identified. This method provides a means of identifying modulators which activate or inhibit 85P1B3.

An embodiment of this invention comprises a method of screening for a molecule that interacts with an 85P1B3 amino acid sequence shown in Figure 2 or Figure 3, comprising the steps of contacting a population of molecules with the 85P1B3 amino acid sequence, allowing the population of molecules and the 85P1B3 amino acid sequence to interact under conditions that facilitate an interaction, determining the presence of a molecule that interacts with the 85P1B3 amino acid sequence, and then separating molecules that do not interact with the 85P1B3 amino acid sequence from molecules that do. In a specific embodiment, the method further comprises purifying, characterizing and identifying a molecule that interacts with the 85P1B3 amino acid sequence. The identified molecule can be used to modulate a function performed by 85P1B3. In a preferred embodiment, the 85P1B3 amino acid sequence is contacted with a library of peptides.

X.) Therapeutic Methods and Compositions

The identification of 85P1B3 as a protein that is normally expressed in a restricted set of tissues, but which is also expressed in prostate and other cancers, opens a number of therapeutic approaches to the treatment of such cancers. As contemplated herein, 85P1B3 functions as a transcription factor involved in activating tumor-promoting genes or repressing genes that block tumorigenesis.

Accordingly, therapeutic approaches that inhibit the activity of the 85P1B3 protein are useful for patients suffering from a cancer that expresses 85P1B3. These therapeutic approaches generally fall into two classes. One class comprises various methods for inhibiting the binding or association of the 85P1B3 protein with its binding partner or with other proteins. Another class comprises a variety of methods for inhibiting the transcription of the 85P1B3 gene or translation of 85P1B3 mRNA.

X.A.) Anti-Cancer Vaccines

The invention provides cancer vaccines comprising a 85P1B3-related protein or 85P1B3-related nucleic acid. In view of the expression of 85P1B3, cancer vaccines prevent and/or treat 85P1B3-expressing cancers with minimal or no effects on non-target tissues. The use of a tumor antigen in a vaccine that generates

humoral and/or cell-mediated immune responses as anti-cancer therapy is well known in the art and has been employed in prostate cancer using human PSMA and rodent PAP immunogens (Hodge et al., 1995, Int. J. Cancer 63:231-237; Fong et al., 1997, J. Immunol. 159:3113-3117).

Such methods can be readily practiced by employing a 85P1B3-related protein, or an 85P1B3-encoding nucleic acid molecule and recombinant vectors capable of expressing and presenting the 85P1B3 immunogen (which typically comprises a number of antibody or T cell epitopes). Skilled artisans understand that a wide variety of vaccine systems for delivery of immunoreactive epitopes are known in the art (see, e.g., Heryln et al., Ann Med 1999 Feb 31(1):66-78; Maruyama et al., Cancer Immunol Immunother 2000 Jun 49(3):123-32) Briefly, such methods of generating an immune response (e.g. humoral and/or cell-mediated) in a mammal, comprise the steps of: exposing the mammal's immune system to an immunoreactive epitope (e.g. an epitope present in the 85P1B3 protein shown in SEQ ID NO: 703 or analog or homolog thereof) so that the mammal generates an immune response that is specific for that epitope (e.g. generates antibodies that specifically recognize that epitope). In a preferred method, the 85P1B3 immunogen contains a biological motif, see e.g., Tables V-XVIII, or a peptide of a size range from 85P1B3 indicated in Figure 5, Figure 6, Figure 7, Figure 8, and Figure 9.

The entire 85P1B3 protein, immunogenic regions or epitopes thereof can be combined and delivered by various means. Such vaccine compositions can include, for example, lipopeptides (*e.g.*,Vitiello, A. *et al.*, *J. Clin. Invest.* 95:341, 1995), peptide compositions encapsulated in poly(DL-lactide-co-glycolide) ("PLG") microspheres (*see, e.g.*, Eldridge, *et al.*, *Molec. Immunol.* 28:287-294, 1991: Alonso *et al.*, *Vaccine* 12:299-306, 1994; Jones *et al.*, *Vaccine* 13:675-681, 1995), peptide compositions contained in immune stimulating complexes (ISCOMS) (*see, e.g.*, Takahashi *et al.*, *Nature* 344:873-875, 1990; Hu *et al.*, *Clin Exp Immunol.* 113:235-243, 1998), multiple antigen peptide systems (MAPs) (*see e.g.*, Tam, J. P., *Proc. Natl. Acad. Sci. U.S.A.* 85:5409-5413, 1988; Tam, J.P., *J. Immunol. Methods* 196:17-32, 1996), peptides formulated as multivalent peptides; peptides for use in ballistic delivery systems, typically crystallized peptides, viral delivery vectors (Perkus, M. E. *et al.*, In: *Concepts in vaccine development*, Kaufmann, S. H. E., ed., p. 379, 1996; Chakrabarti, S. *et al.*, *Nature* 320:535, 1986; Hu, S. L. *et al.*, *Nature* 320:537, 1986; Kieny, M.-P. *et al.*, *AIDS Bio/Technology* 4:790, 1986; Top, F. H. *et al.*, *J. Infect. Dis.* 124:148, 1971; Chanda, P. K. *et al.*, *Virology* 175:535, 1990), particles of viral or synthetic origin (*e.g.*, Kofler, N. *et al.*, *J. Immunol. Methods.* 192:25, 1996; Eldridge, J. H. *et al.*, *Sem. Hematol.* 30:16, 1993; Falo, L. D., Jr. *et al.*, *Nature Med.* 7:649, 1995), adjuvants (Warren, H. S., Vogel, F. R., and Chedid, L. A. *Annu. Rev. Immunol.* 4:369, 1986; Gupta, R. K. *et al.*, *Vaccine* 11:293, 1993), liposomes (Reddy, R. *et al.*, *J. Immunol.* 148:1585, 1992; Rock, K. L., *Immunol. Today* 17:131, 1996), or, naked or particle absorbed cDNA (Ulmer, J. B. *et al.*, *Science* 259:1745, 1993; Robinson, H. L., Hunt, L. A., and Webster, R. G., *Vaccine* 11:957, 1993; Shiver, J. W. *et al.*, In: *Concepts in vaccine development*, Kaufmann, S. H. E., ed.,

p. 423, 1996; Cease, K. B., and Berzofsky, J. A., *Annu. Rev. Immunol.* 12:923, 1994 and Eldridge, J. H. *et al.*, *Sem. Hematol.* 30:16, 1993). Toxin-targeted delivery technologies, also known as receptor mediated targeting, such as those of Avant Immunotherapeutics, Inc. (Needham, Massachusetts) may also be used.

In patients with 85P1B3-associated cancer, the vaccine compositions of the invention can also be used in conjunction with other treatments used for cancer, *e.g.*, surgery, chemotherapy, drug therapies, radiation therapies, *etc.* including use in combination with immune adjuvants such as IL-2, IL-12, GM-CSF, and the like.

Cellular Vaccines:

CTL epitopes can be determined using specific algorithms to identify peptides within 85P1B3 protein that bind corresponding HLA alleles (see e.g., Table IV; Epimer™ and Epimatrix™, Brown University (URL www.brown.edu/Research/TB-HIV_Lab/epimatrix/epimatrix.html); and, BIMAS, (URL bimas.dcrt.nih.gov/; SYFPEITHI at URL syfpeithi.bmi-heidelberg.com/). In a preferred embodiment, the 85P1B3 immunogen contains one or more amino acid sequences identified using techniques well known in the art, such as the sequences shown in Tables V-XVIII or a peptide of 8, 9, 10 or 11 amino acids specified by an HLA Class I motif/supermotif (e.g., Table IV (A), Table IV (D), or Table IV (E)) and/or a peptide of at least 9 amino acids that comprises an HLA Class II motif/supermotif (e.g., Table IV (B) or Table IV (C)). As is appreciated in the art, the HLA Class I binding groove is essentially closed ended so that peptides of only a particular size range can fit into the groove and be bound, generally HLA Class I epitopes are 8, 9, 10, or 11 amino acids long. In contrast, the HLA Class II binding groove is essentially open ended; therefore a peptide of about 9 or more amino acids can be bound by an HLA Class II molecule. Due to the binding groove differences between HLA Class I and II, HLA Class I motifs are length specific, i.e., position two of a Class I motif is the second amino acid in an amino to carboxyl direction of the peptide. The amino acid positions in a Class II motif are relative only to each other, not the overall peptide, i.e., additional amino acids can be attached to the amino and/or carboxyl termini of a motif-bearing sequence. HLA Class II epitopes are often 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 amino acids long, or longer than 25 amino acids.

Antibody-based Vaccines

A wide variety of methods for generating an immune response in a mammal are known in the art (for example as the first step in the generation of hybridomas). Methods of generating an immune response in a mammal comprise exposing the mammal's immune system to an immunogenic epitope on a protein (e.g. the 85P1B3 protein) so that an immune response is generated. A typical embodiment consists of a method for generating an immune response to 85P1B3 in a host, by contacting the host with a sufficient amount of at least one 85P1B3 B cell or cytotoxic T-cell epitope or analog thereof; and at least one periodic interval thereafter re-contacting the host with the 85P1B3 B cell or cytotoxic T-cell epitope or analog

thereof. A specific embodiment consists of a method of generating an immune response against a 85P1B3-related protein or a man-made multiepitopic peptide comprising: administering 85P1B3 immunogen (e.g. the 85P1B3 protein or a peptide fragment thereof, an 85P1B3 fusion protein or analog etc.) in a vaccine preparation to a human or another mammal. Typically, such vaccine preparations further contain a suitable adjuvant (see, e.g., U.S. Patent No. 6,146,635) or a universal helper epitope such as a PADRE™ peptide (Epimmune Inc., San Diego, CA; see, e.g., Alexander et al., J. Immunol. 2000 164(3); 164(3): 1625-1633; Alexander et al., Immunity 1994 1(9): 751-761 and Alexander et al., Immunol. Res. 1998 18(2): 79-92). An alternative method comprises generating an immune response in an individual against a 85P1B3 immunogen by: administering *in vivo* to muscle or skin of the individual's body a DNA molecule that comprises a DNA sequence that encodes an 85P1B3 immunogen, the DNA sequence operatively linked to regulatory sequences which control the expression of the DNA sequence; wherein the DNA molecule is taken up by cells, the DNA sequence is expressed in the cells and an immune response is generated against the immunogen (see, e.g., U.S. Patent No. 5,962,428). Optionally a genetic vaccine facilitator such as anionic lipids; saponins; lectins; estrogenic compounds; hydroxylated lower alkyls; dimethyl sulfoxide; and urea is also administered.

Nucleic Acid Vaccines:

Vaccine compositions of the invention include nucleic acid-mediated modalities. DNA or RNA that encode protein(s) of the invention can be administered to a patient. Genetic immunization methods can be employed to generate prophylactic or therapeutic humoral and cellular immune responses directed against cancer cells expressing 85P1B3. Constructs comprising DNA encoding a 85P1B3-related protein/immunogen and appropriate regulatory sequences can be injected directly into muscle or skin of an individual, such that the cells of the muscle or skin take-up the construct and express the encoded 85P1B3 protein/immunogen. Alternatively, a vaccine comprises a 85P1B3-related protein. Expression of the 85P1B3-related protein immunogen results in the generation of prophylactic or therapeutic humoral and cellular immunity against cells that bear 85P1B3 protein. Various prophylactic and therapeutic genetic immunization techniques known in the art can be used (for review, see information and references published at Internet address www.genweb.com). Nucleic acid-based delivery is described, for instance, in Wolff *et. al.*, *Science* 247:1465 (1990) as well as U.S. Patent Nos. 5,580,859; 5,589,466; 5,804,566; 5,739,118; 5,736,524; 5,679,647; WO 98/04720. Examples of DNA-based delivery technologies include "naked DNA", facilitated (bupivicaine, polymers, peptide-mediated) delivery, cationic lipid complexes, and particle-mediated ("gene gun") or pressure-mediated delivery (*see, e.g.*, U.S. Patent No. 5,922,687).

For therapeutic or prophylactic immunization purposes, proteins of the invention can be expressed via viral or bacterial vectors. Various viral gene delivery systems that can be used in the practice of the invention include, but are not limited to, vaccinia, fowlpox, canarypox, adenovirus, influenza, poliovirus,

adeno-associated virus, lentivirus, and sindbis virus (see, e.g., Restifo, 1996, Curr. Opin. Immunol. 8:658-663; Tsang et al. J. Natl. Cancer Inst. 87:982-990 (1995)). Non-viral delivery systems can also be employed by introducing naked DNA encoding a 85P1B3-related protein into the patient (e.g., intramuscularly or intradermally) to induce an anti-tumor response.

Vaccinia virus is used, for example, as a vector to express nucleotide sequences that encode the peptides of the invention. Upon introduction into a host, the recombinant vaccinia virus expresses the protein immunogenic peptide, and thereby elicits a host immune response. Vaccinia vectors and methods useful in immunization protocols are described in, *e.g.*, U.S. Patent No. 4,722,848. Another vector is BCG (Bacille Calmette Guerin). BCG vectors are described in Stover *et al.*, *Nature* 351:456-460 (1991). A wide variety of other vectors useful for therapeutic administration or immunization of the peptides of the invention, *e.g.* adeno and adeno-associated virus vectors, retroviral vectors, *Salmonella typhi* vectors, detoxified anthrax toxin vectors, and the like, will be apparent to those skilled in the art from the description herein.

Thus, gene delivery systems are used to deliver a 85P1B3-related nucleic acid molecule. In one embodiment, the full-length human 85P1B3 cDNA is employed. In another embodiment, 85P1B3 nucleic acid molecules encoding specific cytotoxic T lymphocyte (CTL) and/or antibody epitopes are employed.

Ex Vivo Vaccines

Various *ex vivo* strategies can also be employed to generate an immune response. One approach involves the use of antigen presenting cells (APCs) such as dendritic cells (DC) to present 85P1B3 antigen to a patient's immune system. Dendritic cells express MHC class I and II molecules, B7 co-stimulator, and IL-12, and are thus highly specialized antigen presenting cells. In prostate cancer, autologous dendritic cells pulsed with peptides of the prostate-specific membrane antigen (PSMA) are being used in a Phase I clinical trial to stimulate prostate cancer patients' immune systems (Tjoa et al., 1996, Prostate 28:65-69; Murphy et al., 1996, Prostate 29:371-380). Thus, dendritic cells can be used to present 85P1B3 peptides to T cells in the context of MHC class I or II molecules. In one embodiment, autologous dendritic cells are pulsed with 85P1B3 peptides capable of binding to MHC class I and/or class II molecules. In another embodiment, dendritic cells are pulsed with the complete 85P1B3 protein. Yet another embodiment involves engineering the overexpression of the 85P1B3 gene in dendritic cells using various implementing vectors known in the art, such as adenovirus (Arthur et al., 1997, Cancer Gene Ther. 4:17-25), retrovirus (Henderson et al., 1996, Cancer Res. 56:3763-3770), lentivirus, adeno-associated virus, DNA transfection (Ribas et al., 1997, Cancer Res. 57:2865-2869), or tumor-derived RNA transfection (Ashley et al., 1997, J. Exp. Med. 186:1177-1182). Cells that express 85P1B3 can also be engineered to express immune modulators, such as GM-CSF, and used as immunizing agents.

X.B.) 85P1B3 as a Target for Antibody-based Therapy

85P1B3 is an attractive target for antibody-based therapeutic strategies. A number of antibody strategies are known in the art for targeting both extracellular and intracellular molecules (see, e.g., complement and ADCC mediated killing as well as the use of intrabodies). Because 85P1B3 is expressed by cancer cells of various lineages relative to corresponding normal cells, systemic administration of 85P1B3-immunoreactive compositions are prepared that exhibit excellent sensitivity without toxic, non-specific and/or non-target effects caused by binding of the immunoreactive composition to non-target organs and tissues. Antibodies specifically reactive with domains of 85P1B3 are useful to treat 85P1B3-expressing cancers systemically, either as conjugates with a toxin or therapeutic agent, or as naked antibodies capable of inhibiting cell proliferation or function.

85P1B3 antibodies can be introduced into a patient such that the antibody binds to 85P1B3 and modulates a function, such as an interaction with a binding partner, and consequently mediates destruction of the tumor cells and/or inhibits the growth of the tumor cells. Mechanisms by which such antibodies exert a therapeutic effect can include complement-mediated cytolysis, antibody-dependent cellular cytotoxicity, modulation of the physiological function of 85P1B3, inhibition of ligand binding or signal transduction pathways, modulation of tumor cell differentiation, alteration of tumor angiogenesis factor profiles, and/or apoptosis.

Those skilled in the art understand that antibodies can be used to specifically target and bind immunogenic molecules such as an immunogenic region of the 85P1B3 sequence shown in Figure 2 or Figure 3. In addition, skilled artisans understand that it is routine to conjugate antibodies to cytotoxic agents (see, e.g., Slevers et al. Blood 93:11 3678-3684 (June 1, 1999)). When cytotoxic and/or therapeutic agents are delivered directly to cells, such as by conjugating them to antibodies specific for a molecule expressed by that cell (e.g. 85P1B3), the cytotoxic agent will exert its known biological effect (i.e. cytotoxicity) on those cells.

A wide variety of compositions and methods for using antibody-cytotoxic agent conjugates to kill cells are known in the art. In the context of cancers, typical methods entail administering to an animal having a tumor a biologically effective amount of a conjugate comprising a selected cytotoxic and/or therapeutic agent linked to a targeting agent (e.g. an anti-85P1B3 antibody) that binds to a marker (e.g. 85P1B3) expressed, accessible to binding or localized on the cell surfaces. A typical embodiment is a method of delivering a cytotoxic and/or therapeutic agent to a cell expressing 85P1B3, comprising conjugating the cytotoxic agent to an antibody that immunospecifically binds to a 85P1B3 epitope, and, exposing the cell to the antibody-agent conjugate. Another illustrative embodiment is a method of treating an individual suspected of suffering from metastasized cancer, comprising a step of administering parenterally to said individual a pharmaceutical composition comprising a therapeutically effective amount of an antibody conjugated to a cytotoxic and/or therapeutic agent.

Cancer immunotherapy using anti-85P1B3 antibodies can be done in accordance with various approaches that have been successfully employed in the treatment of other types of cancer, including but not limited to colon cancer (Arlen et al., 1998, Crit. Rev. Immunol. 18:133-138), multiple myeloma (Ozaki et al., 1997, Blood 90:3179-3186, Tsunenari et al., 1997, Blood 90:2437-2444), gastric cancer (Kasprzyk et al., 1992, Cancer Res. 52:2771-2776), B-cell lymphoma (Funakoshi et al., 1996, J. Immunother. Emphasis Tumor Immunol. 19:93-101), leukemia (Zhong et al., 1996, Leuk. Res. 20:581-589), colorectal cancer (Moun et al., 1994, Cancer Res. 54:6160-6166; Velders et al., 1995, Cancer Res. 55:4398-4403), and breast cancer (Shepard et al., 1991, J. Clin. Immunol. 11:117-127). Some therapeutic approaches involve conjugation of naked antibody to a toxin, such as the conjugation of Y^{91} or I^{131} to anti-CD20 antibodies (e.g., Zevalin™, IDEC Pharmaceuticals Corp. or Bexxar™, Coulter Pharmaceuticals), while others involve co-administration of antibodies and other therapeutic agents, such as Herceptin™ (trastuzumab) with paclitaxel (Genentech, Inc.). To treat prostate cancer, for example, 85P1B3 antibodies can be administered in conjunction with radiation, chemotherapy or hormone ablation.

Although 85P1B3 antibody therapy is useful for all stages of cancer, antibody therapy can be particularly appropriate in advanced or metastatic cancers. Treatment with the antibody therapy of the invention is indicated for patients who have received one or more rounds of chemotherapy. Alternatively, antibody therapy of the invention is combined with a chemotherapeutic or radiation regimen for patients who have not received chemotherapeutic treatment. Additionally, antibody therapy can enable the use of reduced dosages of concomitant chemotherapy, particularly for patients who do not tolerate the toxicity of the chemotherapeutic agent very well.

Cancer patients can be evaluated for the presence and level of 85P1B3 expression, preferably using immunohistochemical assessments of tumor tissue, quantitative 85P1B3 imaging, or other techniques that reliably indicate the presence and degree of 85P1B3 expression. Immunohistochemical analysis of tumor biopsies or surgical specimens is preferred for this purpose. Methods for immunohistochemical analysis of tumor tissues are well known in the art.

Anti-85P1B3 monoclonal antibodies that treat prostate and other cancers include those that initiate a potent immune response against the tumor or those that are directly cytotoxic. In this regard, anti-85P1B3 monoclonal antibodies (mAbs) can elicit tumor cell lysis by either complement-mediated or antibody-dependent cell cytotoxicity (ADCC) mechanisms, both of which require an intact Fc portion of the immunoglobulin molecule for interaction with effector cell Fc receptor sites on complement proteins. In addition, anti-85P1B3 mAbs that exert a direct biological effect on tumor growth are useful to treat cancers that express 85P1B3. Mechanisms by which directly cytotoxic mAbs act include: inhibition of cell growth, modulation of cellular differentiation, modulation of tumor angiogenesis factor profiles, and the induction of apoptosis. The mechanism(s) by which a particular anti-85P1B3 mAb exerts an anti-tumor

effect is evaluated using any number of *in vitro* assays that evaluate cell death such as ADCC, ADMMC, complement-mediated cell lysis, and so forth, as is generally known in the art.

In some patients, the use of murine or other non-human monoclonal antibodies, or human/mouse chimeric mAbs can induce moderate to strong immune responses against the non-human antibody. This can result in clearance of the antibody from circulation and reduced efficacy. In the most severe cases, such an immune response can lead to the extensive formation of immune complexes which, potentially, can cause renal failure. Accordingly, preferred monoclonal antibodies used in the therapeutic methods of the invention are those that are either fully human or humanized and that bind specifically to the target 85P1B3 antigen with high affinity but exhibit low or no antigenicity in the patient.

Therapeutic methods of the invention contemplate the administration of single anti-85P1B3 mAbs as well as combinations, or cocktails, of different mAbs. Such mAb cocktails can have certain advantages inasmuch as they contain mAbs that target different epitopes, exploit different effector mechanisms or combine directly cytotoxic mAbs with mAbs that rely on immune effector functionality. Such mAbs in combination can exhibit synergistic therapeutic effects. In addition, anti-85P1B3 mAbs can be administered concomitantly with other therapeutic modalities, including but not limited to various chemotherapeutic agents, androgen-blockers, immune modulators (e.g., IL-2, GM-CSF), surgery or radiation. The anti-85P1B3 mAbs are administered in their "naked" or unconjugated form, or can have a therapeutic agent(s) conjugated to them.

Anti-85P1B3 antibody formulations are administered via any route capable of delivering the antibodies to a tumor cell. Routes of administration include, but are not limited to, intravenous, intraperitoneal, intramuscular, intratumor, intradermal, and the like. Treatment generally involves repeated administration of the anti-85P1B3 antibody preparation, via an acceptable route of administration such as intravenous injection (IV), typically at a dose in the range of about 0.1 to about 10 mg/kg body weight. In general, doses in the range of 10-500 mg mAb per week are effective and well tolerated.

Based on clinical experience with the Herceptin mAb in the treatment of metastatic breast cancer, an initial loading dose of approximately 4 mg/kg patient body weight IV, followed by weekly doses of about 2 mg/kg IV of the anti- 85P1B3 mAb preparation represents an acceptable dosing regimen. Preferably, the initial loading dose is administered as a 90 minute or longer infusion. The periodic maintenance dose is administered as a 30 minute or longer infusion, provided the initial dose was well tolerated. As appreciated by those of skill in the art, various factors can influence the ideal dose regimen in a particular case. Such factors include, for example, the binding affinity and half life of the Ab or mAbs used, the degree of 85P1B3 expression in the patient, the extent of circulating shed 85P1B3 antigen, the desired steady-state antibody concentration level, frequency of treatment, and the influence of

chemotherapeutic or other agents used in combination with the treatment method of the invention, as well as the health status of a particular patient.

Optionally, patients should be evaluated for the levels of 85P1B3 in a given sample (e.g. the levels of circulating 85P1B3 antigen and/or 85P1B3 expressing cells) in order to assist in the determination of the most effective dosing regimen, etc. Such evaluations are also used for monitoring purposes throughout therapy, and are useful to gauge therapeutic success in combination with the evaluation of other parameters (for example, urine cytology and/or ImmunoCyt levels in bladder cancer therapy, or by analogy, serum PSA levels in prostate cancer therapy).

Anti-idiotypic anti-85P1B3 antibodies can also be used in anti-cancer therapy as a vaccine for inducing an immune response to cells expressing a 85P1B3-related protein. In particular, the generation of anti-idiotypic antibodies is well known in the art; this methodology can readily be adapted to generate anti-idiotypic anti-85P1B3 antibodies that mimic an epitope on a 85P1B3-related protein (see, for example, Wagner et al., 1997, Hybridoma 16: 33-40; Foon et al., 1995, J. Clin. Invest. 96:334-342; Herlyn et al., 1996, Cancer Immunol. Immunother. 43:65-76). Such an anti-idiotypic antibody can be used in cancer vaccine strategies.

X.C.) 85P1B3 as a Target for Cellular Immune Responses

Vaccines and methods of preparing vaccines that contain an immunogenically effective amount of one or more HLA-binding peptides as described herein are further embodiments of the invention. Furthermore, vaccines in accordance with the invention encompass compositions of one or more of the claimed peptides. A peptide can be present in a vaccine individually. Alternatively, the peptide can exist as a homopolymer comprising multiple copies of the same peptide, or as a heteropolymer of various peptides. Polymers have the advantage of increased immunological reaction and, where different peptide epitopes are used to make up the polymer, the additional ability to induce antibodies and/or CTLs that react with different antigenic determinants of the pathogenic organism or tumor-related peptide targeted for an immune response. The composition can be a naturally occurring region of an antigen or can be prepared, *e.g.*, recombinantly or by chemical synthesis.

Carriers that can be used with vaccines of the invention are well known in the art, and include, *e.g.*, thyroglobulin, albumins such as human serum albumin, tetanus toxoid, polyamino acids such as poly L-lysine, poly L-glutamic acid, influenza, hepatitis B virus core protein, and the like. The vaccines can contain a physiologically tolerable (*i.e.*, acceptable) diluent such as water, or saline, preferably phosphate buffered saline. The vaccines also typically include an adjuvant. Adjuvants such as incomplete Freund's adjuvant, aluminum phosphate, aluminum hydroxide, or alum are examples of materials well known in the art. Additionally, as disclosed herein, CTL responses can be primed by conjugating peptides of the invention to lipids, such as tripalmitoyl-S-glycerylcysteinlyseryl- serine (P_3CSS). Moreover, an adjuvant

such as a synthetic cytosine-phosphorothiolated-guanine-containing (CpG) oligonucleotides has been found to increase CTL responses 10- to 100-fold. (see, e.g. Davila and Celis J. Immunol. 165:539-547 (2000))

Upon immunization with a peptide composition in accordance with the invention, via injection, aerosol, oral, transdermal, transmucosal, intrapleural, intrathecal, or other suitable routes, the immune system of the host responds to the vaccine by producing large amounts of CTLs and/or HTLs specific for the desired antigen. Consequently, the host becomes at least partially immune to later development of cells that express or overexpress 85P1B3 antigen, or derives at least some therapeutic benefit when the antigen was tumor-associated.

In some embodiments, it may be desirable to combine the class I peptide components with components that induce or facilitate neutralizing antibody and or helper T cell responses directed to the target antigen. A preferred embodiment of such a composition comprises class I and class II epitopes in accordance with the invention. An alternative embodiment of such a composition comprises a class I and/or class II epitope in accordance with the invention, along with a cross reactive HTL epitope such as PADRE™ (Epimmune, San Diego, CA) molecule (described *e.g.*, in U.S. Patent Number 5,736,142).

A vaccine of the invention can also include antigen-presenting cells (APC), such as dendritic cells (DC), as a vehicle to present peptides of the invention. Vaccine compositions can be created *in vitro*, following dendritic cell mobilization and harvesting, whereby loading of dendritic cells occurs *in vitro*. For example, dendritic cells are transfected, *e.g.*, with a minigene in accordance with the invention, or are pulsed with peptides. The dendritic cell can then be administered to a patient to elicit immune responses *in vivo*. Vaccine compositions, either DNA- or peptide-based, can also be administered *in vivo* in combination with dendritic cell mobilization whereby loading of dendritic cells occurs *in vivo*.

Preferably, the following principles are utilized when selecting an array of epitopes for inclusion in a polyepitopic composition for use in a vaccine, or for selecting discrete epitopes to be included in a vaccine and/or to be encoded by nucleic acids such as a minigene. It is preferred that each of the following principles be balanced in order to make the selection. The multiple epitopes to be incorporated in a given vaccine composition may be, but need not be, contiguous in sequence in the native antigen from which the epitopes are derived.

1.) Epitopes are selected which, upon administration, mimic immune responses that have been observed to be correlated with tumor clearance. For HLA Class I this includes 3-4 epitopes that come from at least one tumor associated antigen (TAA). For HLA Class II a similar rationale is employed; again 3-4 epitopes are selected from at least one TAA (*see, e.g.*, Rosenberg *et al.*, *Science* 278:1447-1450). Epitopes from one TAA may be used in combination with epitopes from one or more additional TAAs to produce a vaccine that targets tumors with varying expression patterns of frequently-expressed TAAs.

2.) Epitopes are selected that have the requisite binding affinity established to be correlated with immunogenicity: for HLA Class I an IC_{50} of 500 nM or less, often 200 nM or less; and for Class II an IC_{50} of 1000 nM or less.

3.) Sufficient supermotif bearing-peptides, or a sufficient array of allele-specific motif-bearing peptides, are selected to give broad population coverage. For example, it is preferable to have at least 80% population coverage. A Monte Carlo analysis, a statistical evaluation known in the art, can be employed to assess the breadth, or redundancy of, population coverage.

4.) When selecting epitopes from cancer-related antigens it is often useful to select analogs because the patient may have developed tolerance to the native epitope.

5.) Of particular relevance are epitopes referred to as "nested epitopes." Nested epitopes occur where at least two epitopes overlap in a given peptide sequence. A nested peptide sequence can comprise B cell, HLA class I and/or HLA class II epitopes. When providing nested epitopes, a general objective is to provide the greatest number of epitopes per sequence. Thus, an aspect is to avoid providing a peptide that is any longer than the amino terminus of the amino terminal epitope and the carboxyl terminus of the carboxyl terminal epitope in the peptide. When providing a multi-epitopic sequence, such as a sequence comprising nested epitopes, it is generally important to screen the sequence in order to insure that it does not have pathological or other deleterious biological properties.

6.) If a polyepitopic protein is created, or when creating a minigene, an objective is to generate the smallest peptide that encompasses the epitopes of interest. This principle is similar, if not the same as that employed when selecting a peptide comprising nested epitopes. However, with an artificial polyepitopic peptide, the size minimization objective is balanced against the need to integrate any spacer sequences between epitopes in the polyepitopic protein. Spacer amino acid residues can, for example, be introduced to avoid junctional epitopes (an epitope recognized by the immune system, not present in the target antigen, and only created by the man-made juxtaposition of epitopes), or to facilitate cleavage between epitopes and thereby enhance epitope presentation. Junctional epitopes are generally to be avoided because the recipient may generate an immune response to that non-native epitope. Of particular concern is a junctional epitope that is a "dominant epitope." A dominant epitope may lead to such a zealous response that immune responses to other epitopes are diminished or suppressed.

7.) Where the sequences of multiple variants of the same target protein are present, potential peptide epitopes can also be selected on the basis of their conservancy. For example, a criterion for conservancy may define that the entire sequence of an HLA class I binding peptide or the entire 9-mer core of a class II binding peptide be conserved in a designated percentage of the sequences evaluated for a specific protein antigen.

X.C.1. Minigene Vaccines

A number of different approaches are available which allow simultaneous delivery of multiple epitopes. Nucleic acids encoding the peptides of the invention are a particularly useful embodiment of the invention. Epitopes for inclusion in a minigene are preferably selected according to the guidelines set forth in the previous section. A preferred means of administering nucleic acids encoding the peptides of the invention uses minigene constructs encoding a peptide comprising one or multiple epitopes of the invention.

The use of multi-epitope minigenes is described below and in, Ishioka *et al.*, *J. Immunol.* 162:3915-3925, 1999; An, L. and Whitton, J. L., *J. Virol.* 71:2292, 1997; Thomson, S. A. *et al.*, *J. Immunol.* 157:822, 1996; Whitton, J. L. *et al.*, *J. Virol.* 67:348, 1993; Hanke, R. *et al.*, *Vaccine* 16:426, 1998. For example, a multi-epitope DNA plasmid encoding supermotif- and/or motif-bearing epitopes derived 85P1B3, the PADRE® universal helper T cell epitope (or multiple HTL epitopes from 85P1B3), and an endoplasmic reticulum-translocating signal sequence can be engineered. A vaccine may also comprise epitopes that are derived from other TAAs.

The immunogenicity of a multi-epitopic minigene can be confirmed in transgenic mice to evaluate the magnitude of CTL induction responses against the epitopes tested. Further, the immunogenicity of DNA-encoded epitopes *in vivo* can be correlated with the *in vitro* responses of specific CTL lines against target cells transfected with the DNA plasmid. Thus, these experiments can show that the minigene serves to both: 1.) generate a CTL response and 2.) that the induced CTLs recognized cells expressing the encoded epitopes.

For example, to create a DNA sequence encoding the selected epitopes (minigene) for expression in human cells, the amino acid sequences of the epitopes may be reverse translated. A human codon usage table can be used to guide the codon choice for each amino acid. These epitope-encoding DNA sequences may be directly adjoined, so that when translated, a continuous polypeptide sequence is created. To optimize expression and/or immunogenicity, additional elements can be incorporated into the minigene design. Examples of amino acid sequences that can be reverse translated and included in the minigene sequence include: HLA class I epitopes, HLA class II epitopes, antibody epitopes, a ubiquitination signal sequence, and/or an endoplasmic reticulum targeting signal. In addition, HLA presentation of CTL and HTL epitopes may be improved by including synthetic (*e.g.* poly-alanine) or naturally-occurring flanking sequences adjacent to the CTL or HTL epitopes; these larger peptides comprising the epitope(s) are within the scope of the invention.

The minigene sequence may be converted to DNA by assembling oligonucleotides that encode the plus and minus strands of the minigene. Overlapping oligonucleotides (30-100 bases long) may be synthesized, phosphorylated, purified and annealed under appropriate conditions using well known

techniques. The ends of the oligonucleotides can be joined, for example, using T4 DNA ligase. This synthetic minigene, encoding the epitope polypeptide, can then be cloned into a desired expression vector.

Standard regulatory sequences well known to those of skill in the art are preferably included in the vector to ensure expression in the target cells. Several vector elements are desirable: a promoter with a down-stream cloning site for minigene insertion; a polyadenylation signal for efficient transcription termination; an *E. coli* origin of replication; and an *E. coli* selectable marker (*e.g.* ampicillin or kanamycin resistance). Numerous promoters can be used for this purpose, *e.g.*, the human cytomegalovirus (hCMV) promoter. See, *e.g.*, U.S. Patent Nos. 5,580,859 and 5,589,466 for other suitable promoter sequences.

Additional vector modifications may be desired to optimize minigene expression and immunogenicity. In some cases, introns are required for efficient gene expression, and one or more synthetic or naturally-occurring introns could be incorporated into the transcribed region of the minigene. The inclusion of mRNA stabilization sequences and sequences for replication in mammalian cells may also be considered for increasing minigene expression.

Once an expression vector is selected, the minigene is cloned into the polylinker region downstream of the promoter. This plasmid is transformed into an appropriate *E. coli* strain, and DNA is prepared using standard techniques. The orientation and DNA sequence of the minigene, as well as all other elements included in the vector, are confirmed using restriction mapping and DNA sequence analysis. Bacterial cells harboring the correct plasmid can be stored as a master cell bank and a working cell bank.

In addition, immunostimulatory sequences (ISSs or CpGs) appear to play a role in the immunogenicity of DNA vaccines. These sequences may be included in the vector, outside the minigene coding sequence, if desired to enhance immunogenicity.

In some embodiments, a bi-cistronic expression vector which allows production of both the minigene-encoded epitopes and a second protein (included to enhance or decrease immunogenicity) can be used. Examples of proteins or polypeptides that could beneficially enhance the immune response if co-expressed include cytokines (*e.g.*, IL-2, IL-12, GM-CSF), cytokine-inducing molecules (*e.g.*, LeIF), costimulatory molecules, or for HTL responses, pan-DR binding proteins (PADRE™, Epimmune, San Diego, CA). Helper (HTL) epitopes can be joined to intracellular targeting signals and expressed separately from expressed CTL epitopes; this allows direction of the HTL epitopes to a cell compartment different than that of the CTL epitopes. If required, this could facilitate more efficient entry of HTL epitopes into the HLA class II pathway, thereby improving HTL induction. In contrast to HTL or CTL induction, specifically decreasing the immune response by co-expression of immunosuppressive molecules (*e.g.* TGF-β) may be beneficial in certain diseases.

Therapeutic quantities of plasmid DNA can be produced for example, by fermentation in *E. coli*, followed by purification. Aliquots from the working cell bank are used to inoculate growth medium, and

grown to saturation in shaker flasks or a bioreactor according to well-known techniques. Plasmid DNA can be purified using standard bioseparation technologies such as solid phase anion-exchange resins supplied by QIAGEN, Inc. (Valencia, California). If required, supercoiled DNA can be isolated from the open circular and linear forms using gel electrophoresis or other methods.

Purified plasmid DNA can be prepared for injection using a variety of formulations. The simplest of these is reconstitution of lyophilized DNA in sterile phosphate-buffer saline (PBS). This approach, known as "naked DNA," is currently being used for intramuscular (IM) administration in clinical trials. To maximize the immunotherapeutic effects of minigene DNA vaccines, an alternative method for formulating purified plasmid DNA may be desirable. A variety of methods have been described, and new techniques may become available. Cationic lipids, glycolipids, and fusogenic liposomes can also be used in the formulation (see, *e.g.*, as described by WO 93/24640; Mannino & Gould-Fogerite, *BioTechniques* 6(7): 682 (1988); U.S. Pat No. 5,279,833; WO 91/06309; and Felgner, *et al., Proc. Nat'l Acad. Sci. USA* 84:7413 (1987). In addition, peptides and compounds referred to collectively as protective, interactive, non-condensing compounds (PINC) could also be complexed to purified plasmid DNA to influence variables such as stability, intramuscular dispersion, or trafficking to specific organs or cell types.

Target cell sensitization can be used as a functional assay for expression and HLA class I presentation of minigene-encoded CTL epitopes. For example, the plasmid DNA is introduced into a mammalian cell line that is suitable as a target for standard CTL chromium release assays. The transfection method used will be dependent on the final formulation. Electroporation can be used for "naked" DNA, whereas cationic lipids allow direct *in vitro* transfection. A plasmid expressing green fluorescent protein (GFP) can be co-transfected to allow enrichment of transfected cells using fluorescence activated cell sorting (FACS). These cells are then chromium-51 (^{51}Cr) labeled and used as target cells for epitope-specific CTL lines; cytolysis, detected by ^{51}Cr release, indicates both production of, and HLA presentation of, minigene-encoded CTL epitopes. Expression of HTL epitopes may be evaluated in an analogous manner using assays to assess HTL activity.

In vivo immunogenicity is a second approach for functional testing of minigene DNA formulations. Transgenic mice expressing appropriate human HLA proteins are immunized with the DNA product. The dose and route of administration are formulation dependent (*e.g.*, IM for DNA in PBS, intraperitoneal (i.p.) for lipid-complexed DNA). Twenty-one days after immunization, splenocytes are harvested and restimulated for one week in the presence of peptides encoding each epitope being tested. Thereafter, for CTL effector cells, assays are conducted for cytolysis of peptide-loaded, ^{51}Cr-labeled target cells using standard techniques. Lysis of target cells that were sensitized by HLA loaded with peptide epitopes, corresponding to minigene-encoded epitopes, demonstrates DNA vaccine function for *in vivo*

induction of CTLs. Immunogenicity of HTL epitopes is confirmed in transgenic mice in an analogous manner.

Alternatively, the nucleic acids can be administered using ballistic delivery as described, for instance, in U.S. Patent No. 5,204,253. Using this technique, particles comprised solely of DNA are administered. In a further alternative embodiment, DNA can be adhered to particles, such as gold particles.

Minigenes can also be delivered using other bacterial or viral delivery systems well known in the art, *e.g.*, an expression construct encoding epitopes of the invention can be incorporated into a viral vector such as vaccinia.

X.C.2. Combinations of CTL Peptides with Helper Peptides

Vaccine compositions comprising CTL peptides of the invention can be modified, e.g., analoged, to provide desired attributes, such as improved serum half life, broadened population coverage or enhanced immunogenicity.

For instance, the ability of a peptide to induce CTL activity can be enhanced by linking the peptide to a sequence which contains at least one epitope that is capable of inducing a T helper cell response. Although a CTL peptide can be directly linked to a T helper peptide, often CTL epitope/HTL epitope conjugates are linked by a spacer molecule. The spacer is typically comprised of relatively small, neutral molecules, such as amino acids or amino acid mimetics, which are substantially uncharged under physiological conditions. The spacers are typically selected from, *e.g.*, Ala, Gly, or other neutral spacers of nonpolar amino acids or neutral polar amino acids. It will be understood that the optionally present spacer need not be comprised of the same residues and thus may be a hetero- or homo-oligomer. When present, the spacer will usually be at least one or two residues, more usually three to six residues and sometimes 10 or more residues. The CTL peptide epitope can be linked to the T helper peptide epitope either directly or via a spacer either at the amino or carboxy terminus of the CTL peptide. The amino terminus of either the immunogenic peptide or the T helper peptide may be acylated.

In certain embodiments, the T helper peptide is one that is recognized by T helper cells present in a majority of a genetically diverse population. This can be accomplished by selecting peptides that bind to many, most, or all of the HLA class II molecules. Examples of such amino acid bind many HLA Class II molecules include sequences from antigens such as tetanus toxoid at positions 830-843 (QYIKANSKFIGITE; SEQ ID NO: 710), *Plasmodium falciparum* circumsporozoite (CS) protein at positions 378-398 (DIEKKIAKMEKASSVFNVVNS; SEQ ID NO: 711), and *Streptococcus* 18kD protein at positions 116-131 (GAVDSILGGVATYGAA; SEQ ID NO: 712). Other examples include peptides bearing a DR 1-4-7 supermotif, or either of the DR3 motifs.

Alternatively, it is possible to prepare synthetic peptides capable of stimulating T helper lymphocytes, in a loosely HLA-restricted fashion, using amino acid sequences not found in nature (*see*,

e.g., PCT publication WO 95/07707). These synthetic compounds called Pan-DR-binding epitopes (*e.g.*, PADRE™, Epimmune, Inc., San Diego, CA) are designed to most preferably bind most HLA-DR (human HLA class II) molecules. For instance, a pan-DR-binding epitope peptide having the formula: aKXVAAWTLKAAa (SEQ ID NO: 713), where "X" is either cyclohexylalanine, phenylalanine, or tyrosine, and a is either D-alanine or L-alanine, has been found to bind to most HLA-DR alleles, and to stimulate the response of T helper lymphocytes from most individuals, regardless of their HLA type. An alternative of a pan-DR binding epitope comprises all "L" natural amino acids and can be provided in the form of nucleic acids that encode the epitope.

HTL peptide epitopes can also be modified to alter their biological properties. For example, they can be modified to include D-amino acids to increase their resistance to proteases and thus extend their serum half life, or they can be conjugated to other molecules such as lipids, proteins, carbohydrates, and the like to increase their biological activity. For example, a T helper peptide can be conjugated to one or more palmitic acid chains at either the amino or carboxyl termini.

X.C.3. Combinations of CTL Peptides with T Cell Priming Agents

In some embodiments it may be desirable to include in the pharmaceutical compositions of the invention at least one component which primes B lymphocytes or T lymphocytes. Lipids have been identified as agents capable of priming CTL *in vivo*. For example, palmitic acid residues can be attached to the ε-and α- amino groups of a lysine residue and then linked, *e.g.*, via one or more linking residues such as Gly, Gly-Gly-, Ser, Ser-Ser, or the like, to an immunogenic peptide. The lipidated peptide can then be administered either directly in a micelle or particle, incorporated into a liposome, or emulsified in an adjuvant, *e.g.*, incomplete Freund's adjuvant. In a preferred embodiment, a particularly effective immunogenic composition comprises palmitic acid attached to ε- and α- amino groups of Lys, which is attached via linkage, *e.g.*, Ser-Ser, to the amino terminus of the immunogenic peptide.

As another example of lipid priming of CTL responses, *E. coli* lipoproteins, such as tripalmitoyl-S-glycerylcysteinlyseryl- serine (P_3CSS) can be used to prime virus specific CTL when covalently attached to an appropriate peptide (*see, e.g.*, Deres, *et al., Nature* 342:561, 1989). Peptides of the invention can be coupled to P_3CSS, for example, and the lipopeptide administered to an individual to specifically prime an immune response to the target antigen. Moreover, because the induction of neutralizing antibodies can also be primed with P_3CSS-conjugated epitopes, two such compositions can be combined to more effectively elicit both humoral and cell-mediated responses.

X.C.4. Vaccine Compositions Comprising DC Pulsed with CTL and/or HTL Peptides

An embodiment of a vaccine composition in accordance with the invention comprises *ex vivo* administration of a cocktail of epitope-bearing peptides to PBMC, or isolated DC therefrom, from the patient's blood. A pharmaceutical to facilitate harvesting of DC can be used, such as Progenipoietin™ (Pharmacia-Monsanto, St. Louis, MO) or GM-CSF/IL-4. After pulsing the DC with peptides and prior to reinfusion into patients, the DC are washed to remove unbound peptides. In this embodiment, a vaccine comprises peptide-pulsed DCs which present the pulsed peptide epitopes complexed with HLA molecules on their surfaces.

The DC can be pulsed *ex vivo* with a cocktail of peptides, some of which stimulate CTL responses to 85P1B3. Optionally, a helper T cell (HTL) peptide, such as a natural or artificial loosely restricted HLA Class II peptide, can be included to facilitate the CTL response. Thus, a vaccine in accordance with the invention is used to treat a cancer which expresses or overexpresses 85P1B3.

X.D. Adoptive Immunotherapy

Antigenic 85P1B3-related peptides are used to elicit a CTL and/or HTL response *ex vivo*, as well. The resulting CTL or HTL cells, can be used to treat tumors in patients that do not respond to other conventional forms of therapy, or will not respond to a therapeutic vaccine peptide or nucleic acid in accordance with the invention. *Ex vivo* CTL or HTL responses to a particular antigen are induced by incubating in tissue culture the patient's, or genetically compatible, CTL or HTL precursor cells together with a source of antigen-presenting cells (APC), such as dendritic cells, and the appropriate immunogenic peptide. After an appropriate incubation time (typically about 7-28 days), in which the precursor cells are activated and expanded into effector cells, the cells are infused back into the patient, where they will destroy (CTL) or facilitate destruction (HTL) of their specific target cell (e.g., a tumor cell). Transfected dendritic cells may also be used as antigen presenting cells.

X.E. Administration of Vaccines for Therapeutic or Prophylactic Purposes

Pharmaceutical and vaccine compositions of the invention are typically used to treat and/or prevent a cancer that expresses or overexpresses 85P1B3. In therapeutic applications, peptide and/or nucleic acid compositions are administered to a patient in an amount sufficient to elicit an effective B cell, CTL and/or HTL response to the antigen and to cure or at least partially arrest or slow symptoms and/or complications. An amount adequate to accomplish this is defined as "therapeutically effective dose." Amounts effective for this use will depend on, *e.g.*, the particular composition administered, the manner of administration, the stage and severity of the disease being treated, the weight and general state of health of the patient, and the judgment of the prescribing physician.

For pharmaceutical compositions, the immunogenic peptides of the invention, or DNA encoding them, are generally administered to an individual already bearing a tumor that expresses 85P1B3. The peptides or DNA encoding them can be administered individually or as fusions of one or more peptide

sequences. Patients can be treated with the immunogenic peptides separately or in conjunction with other treatments, such as surgery, as appropriate.

For therapeutic use, administration should generally begin at the first diagnosis of 85P1B3-associated cancer. This is followed by boosting doses until at least symptoms are substantially abated and for a period thereafter. The embodiment of the vaccine composition (*i.e.*, including, but not limited to embodiments such as peptide cocktails, polyepitopic polypeptides, minigenes, or TAA-specific CTLs or pulsed dendritic cells) delivered to the patient may vary according to the stage of the disease or the patient's health status. For example, in a patient with a tumor that expresses 85P1B3, a vaccine comprising 85P1B3-specific CTL may be more efficacious in killing tumor cells in patient with advanced disease than alternative embodiments.

It is generally important to provide an amount of the peptide epitope delivered by a mode of administration sufficient to effectively stimulate a cytotoxic T cell response; compositions which stimulate helper T cell responses can also be given in accordance with this embodiment of the invention.

The dosage for an initial therapeutic immunization generally occurs in a unit dosage range where the lower value is about 1, 5, 50, 500, or 1,000 μg and the higher value is about 10,000; 20,000; 30,000; or 50,000 μg. Dosage values for a human typically range from about 500 μg to about 50,000 μg per 70 kilogram patient. Boosting dosages of between about 1.0 μg to about 50,000 μg of peptide pursuant to a boosting regimen over weeks to months may be administered depending upon the patient's response and condition as determined by measuring the specific activity of CTL and HTL obtained from the patient's blood. Administration should continue until at least clinical symptoms or laboratory tests indicate that the neoplasia, has been eliminated or reduced and for a period thereafter. The dosages, routes of administration, and dose schedules are adjusted in accordance with methodologies known in the art.

In certain embodiments, the peptides and compositions of the present invention are employed in serious disease states, that is, life-threatening or potentially life threatening situations. In such cases, as a result of the minimal amounts of extraneous substances and the relative nontoxic nature of the peptides in preferred compositions of the invention, it is possible and may be felt desirable by the treating physician to administer substantial excesses of these peptide compositions relative to these stated dosage amounts.

The vaccine compositions of the invention can also be used purely as prophylactic agents. Generally the dosage for an initial prophylactic immunization generally occurs in a unit dosage range where the lower value is about 1, 5, 50, 500, or 1000 μg and the higher value is about 10,000; 20,000; 30,000; or 50,000 μg. Dosage values for a human typically range from about 500 μg to about 50,000 μg per 70 kilogram patient. This is followed by boosting dosages of between about 1.0 μg to about 50,000 μg of peptide administered at defined intervals from about four weeks to six months after the initial

administration of vaccine. The immunogenicity of the vaccine can be assessed by measuring the specific activity of CTL and HTL obtained from a sample of the patient's blood.

The pharmaceutical compositions for therapeutic treatment are intended for parenteral, topical, oral, nasal, intrathecal, or local (*e.g.* as a cream or topical ointment) administration. Preferably, the pharmaceutical compositions are administered parentally, *e.g.*, intravenously, subcutaneously, intradermally, or intramuscularly. Thus, the invention provides compositions for parenteral administration which comprise a solution of the immunogenic peptides dissolved or suspended in an acceptable carrier, preferably an aqueous carrier.

A variety of aqueous carriers may be used, *e.g.*, water, buffered water, 0.8% saline, 0.3% glycine, hyaluronic acid and the like. These compositions may be sterilized by conventional, well-known sterilization techniques, or may be sterile filtered. The resulting aqueous solutions may be packaged for use as is, or lyophilized, the lyophilized preparation being combined with a sterile solution prior to administration.

The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions, such as pH-adjusting and buffering agents, tonicity adjusting agents, wetting agents, preservatives, and the like, for example, sodium acetate, sodium lactate, sodium chloride, potassium chloride, calcium chloride, sorbitan monolaurate, triethanolamine oleate, *etc.*

The concentration of peptides of the invention in the pharmaceutical formulations can vary widely, *i.e.*, from less than about 0.1%, usually at or at least about 2% to as much as 20% to 50% or more by weight, and will be selected primarily by fluid volumes, viscosities, *etc.*, in accordance with the particular mode of administration selected.

A human unit dose form of a composition is typically included in a pharmaceutical composition that comprises a human unit dose of an acceptable carrier, in one embodiment an aqueous carrier, and is administered in a volume/quantity that is known by those of skill in the art to be used for administration of such compositions to humans (*see*, *e.g.*, Remington's Pharmaceutical Sciences, 17th Edition, A. Gennaro, Editor, Mack Publishing Co., Easton, Pennsylvania, 1985). For example a peptide dose for initial immunization can be from about 1 to about 50,000 μg, generally 100-5,000 μg, for a 70 kg patient. For example, for nucleic acids an initial immunization may be performed using an expression vector in the form of naked nucleic acid administered IM (or SC or ID) in the amounts of 0.5-5 mg at multiple sites. The nucleic acid (0.1 to 1000 μg) can also be administered using a gene gun. Following an incubation period of 3-4 weeks, a booster dose is then administered. The booster can be recombinant fowlpox virus administered at a dose of 5-10^7 to 5x10^9 pfu. For antibodies, a reatment generally involves repeated administration of the anti-85P1B3 antibody preparation, via an acceptable route of administration such as intravenous injection (IV), typically at a dose in the range of about 0.1 to about 10 mg/kg body weight. In

general, doses in the range of 10-500 mg mAb per week are effective and well tolerated. Moreover, an initial loading dose of approximately 4 mg/kg patient body weight IV, followed by weekly doses of about 2 mg/kg IV of the anti- 85P1B3 mAb preparation represents an acceptable dosing regimen. As appreciated by those of skill in the art, various factors can influence the ideal dose in a particular case. Such factors include, for example, half life of a composition, the binding affinity of an Ab, the immunogenicity of a substacne, the degree of 85P1B3 expression in the patient, the extent of circulating shed 85P1B3 antigen, the desired steady-state concentration level, frequency of treatment, and the influence of chemotherapeutic or other agents used in combination with the treatment method of the invention, as well as the health status of a particular patient.

In one embodiment, human unit dose forms of polynucleotides comprise a suitable dosage range or effective amount that provides any therapeutic effect. As appreciated by one of ordinary skill in the art a therapeutic effect depends on a number of factors, including the sequence of the polynucleotide, molecular weight of the polynucleotide and route of administration. Dosages are generally selected by the physician or other health care professional in accordance with a variety of parameters known in the art, such as severity of symptoms, history of the patient and the like. Generally, for a polynucleotide of about 20 bases, a dosage range may be selected from, for example, an independently selected lower limit such as about 0.1, 0.25, 0.5, 1, 2, 5, 10, 20, 30, 40, 50, 60, 80, 100, 200, 300, 400 or 500 mg/kg up to an independently selected upper limit, greater than the lower limit, of about 60, 80, 100, 200, 300, 400, 500, 750, 1000, 1500, 2000, 3000, 4000, 5000, 6000, 7000, 8000, 9000 or 10,000 mg/kg. For example, a dose may be about any of the following: 0.1 to 100 mg/kg, 0.1 to 50 mg/kg, 0.1 to 25 mg/kg, 0.1 to 10 mg/kg, 1 to 500 mg/kg, 100 to 400 mg/kg, 200 to 300 mg/kg, 1 to 100 mg/kg, 100 to 200 mg/kg, 300 to 400 mg/kg, 400 to 500 mg/kg, 500 to 1000 mg/kg, 500 to 5000 mg/kg, or 500 to 10,000 mg/kg. Generally, parenteral routes of administration may require higher doses of polynucleotide compared to more direct application to the nucleotide to diseased tissue, as do polynucleotides of increasing length.

In one embodiment, human unit dose forms of T-cells comprise a suitable dosage range or effective amount that provides any therapeutic effect. As appreciated by one of ordinary skill in the art, a therapeutic effect depends on a number of factors. Dosages are generally selected by the physician or other health care professional in accordance with a variety of parameters known in the art, such as severity of symptoms, history of the patient and the like. A dose may be about 10^4 cells to about 10^6 cells, about 10^6 cells to about 10^8 cells, about 10^8 to about 10^{11} cells, or about 10^8 to about 5×10^{10} cells. A dose may also about 10^6 cells/m^2 to about 10^{10} cells/m^2 , or about 10^6 cells/m^2 to about 10^8 cells/m^2 .

Proteins(s) of the invention, and/or nucleic acids encoding the protein(s), can also be administered via liposomes, which may also serve to: 1) target the proteins(s) to a particular tissue, such as lymphoid tissue; 2) to target selectively to diseases cells; or, 3) to increase the half-life of the peptide composition.

Liposomes include emulsions, foams, micelles, insoluble monolayers, liquid crystals, phospholipid dispersions, lamellar layers and the like. In these preparations, the peptide to be delivered is incorporated as part of a liposome, alone or in conjunction with a molecule which binds to a receptor prevalent among lymphoid cells, such as monoclonal antibodies which bind to the CD45 antigen, or with other therapeutic or immunogenic compositions. Thus, liposomes either filled or decorated with a desired peptide of the invention can be directed to the site of lymphoid cells, where the liposomes then deliver the peptide compositions. Liposomes for use in accordance with the invention are formed from standard vesicle-forming lipids, which generally include neutral and negatively charged phospholipids and a sterol, such as cholesterol. The selection of lipids is generally guided by consideration of, *e.g.*, liposome size, acid lability and stability of the liposomes in the blood stream. A variety of methods are available for preparing liposomes, as described in, *e.g.*, Szoka, *et al.*, *Ann. Rev. Biophys. Bioeng.* 9:467 (1980), and U.S. Patent Nos. 4,235,871, 4,501,728, 4,837,028, and 5,019,369.

For targeting cells of the immune system, a ligand to be incorporated into the liposome can include, *e.g.*, antibodies or fragments thereof specific for cell surface determinants of the desired immune system cells. A liposome suspension containing a peptide may be administered intravenously, locally, topically, *etc.* in a dose which varies according to, *inter alia*, the manner of administration, the peptide being delivered, and the stage of the disease being treated.

For solid compositions, conventional nontoxic solid carriers may be used which include, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharin, talcum, cellulose, glucose, sucrose, magnesium carbonate, and the like. For oral administration, a pharmaceutically acceptable nontoxic composition is formed by incorporating any of the normally employed excipients, such as those carriers previously listed, and generally 10-95% of active ingredient, that is, one or more peptides of the invention, and more preferably at a concentration of 25%-75%.

For aerosol administration, immunogenic peptides are preferably supplied in finely divided form along with a surfactant and propellant. Typical percentages of peptides are about 0.01%-20% by weight, preferably about 1%-10%. The surfactant must, of course, be nontoxic, and preferably soluble in the propellant. Representative of such agents are the esters or partial esters of fatty acids containing from about 6 to 22 carbon atoms, such as caproic, octanoic, lauric, palmitic, stearic, linoleic, linolenic, olesteric and oleic acids with an aliphatic polyhydric alcohol or its cyclic anhydride. Mixed esters, such as mixed or natural glycerides may be employed. The surfactant may constitute about 0.1%-20% by weight of the composition, preferably about 0.25-5%. The balance of the composition is ordinarily propellant. A carrier can also be included, as desired, as with, *e.g.*, lecithin for intranasal delivery.

XI.) Diagnostic and Prognostic Embodiments of 85P1B3.

As disclosed herein, 85P1B3 polynucleotides, polypeptides, reactive cytotoxic T cells (CTL), reactive helper T cells (HTL) and anti-polypeptide antibodies are used in well known diagnostic, prognostic and therapeutic assays that examine conditions associated with dysregulated cell growth such as cancer, in particular the cancers listed in Table I (see, e.g., both its specific pattern of tissue expression as well as its overexpression in certain cancers as described for example in Example 4).

85P1B3 can be analogized to a prostate associated antigen PSA, the archetypal marker that has been used by medical practitioners for years to identify and monitor the presence of prostate cancer (see, e.g., Merrill et al., J. Urol. 163(2): 503-5120 (2000); Polascik et al., J. Urol. Aug; 162(2):293-306 (1999) and Fortier et al., J. Nat. Cancer Inst. 91(19): 1635-1640(1999)). A variety of other diagnostic markers are also used in similar contexts including p53 and K-ras (see, e.g., Tulchinsky et al., Int J Mol Med 1999 Jul 4(1):99-102 and Minimoto et al., Cancer Detect Prev 2000;24(1):1-12). Therefore, this disclosure of the 85P1B3 polynucleotides and polypeptides (as well as the 85P1B3 polynucleotide probes and anti-85P1B3 antibodies used to identify the presence of these molecules) and their properties allows skilled artisans to utilize these molecules in methods that are analogous to those used, for example, in a variety of diagnostic assays directed to examining conditions associated with cancer.

Typical embodiments of diagnostic methods which utilize the 85P1B3 polynucleotides, polypeptides, reactive T cells and antibodies are analogous to those methods from well-established diagnostic assays which employ, e.g., PSA polynucleotides, polypeptides, reactive T cells and antibodies. For example, just as PSA polynucleotides are used as probes (for example in Northern analysis, see, e.g., Sharief et al., Biochem. Mol. Biol. Int. 33(3):567-74(1994)) and primers (for example in PCR analysis, see, e.g., Okegawa et al., J. Urol. 163(4): 1189-1190 (2000)) to observe the presence and/or the level of PSA mRNAs in methods of monitoring PSA overexpression or the metastasis of prostate cancers, the 85P1B3 polynucleotides described herein can be utilized in the same way to detect 85P1B3 overexpression or the metastasis of prostate and other cancers expressing this gene. Alternatively, just as PSA polypeptides are used to generate antibodies specific for PSA which can then be used to observe the presence and/or the level of PSA proteins in methods to monitor PSA protein overexpression (see, e.g., Stephan et al., Urology 55(4):560-3 (2000)) or the metastasis of prostate cells (see, e.g., Alanen et al., Pathol. Res. Pract. 192(3):233-7 (1996)), the 85P1B3 polypeptides described herein can be utilized to generate antibodies for use in detecting 85P1B3 overexpression or the metastasis of prostate cells and cells of other cancers expressing this gene.

Specifically, because metastases involves the movement of cancer cells from an organ of origin (such as the lung or prostate gland etc.) to a different area of the body (such as a lymph node), assays which examine a biological sample for the presence of cells expressing 85P1B3 polynucleotides and/or polypeptides can be used to provide evidence of metastasis. For example, when a biological sample from

tissue that does not normally contain 85P1B3-expressing cells (lymph node) is found to contain 85P1B3-expressing cells such as the 85P1B3 expression seen in LAPC4 and LAPC9, xenografts isolated from lymph node and bone metastasis, respectively, this finding is indicative of metastasis.

Alternatively 85P1B3 polynucleotides and/or polypeptides can be used to provide evidence of cancer, for example, when cells in a biological sample that do not normally express 85P1B3 or express 85P1B3 at a different level are found to express 85P1B3 or have an increased expression of 85P1B3 (see, e.g., the 85P1B3 expression in the cancers listed in Table I and in patient samples etc. shown in the accompanying Figures). In such assays, artisans may further wish to generate supplementary evidence of metastasis by testing the biological sample for the presence of a second tissue restricted marker (in addition to 85P1B3) such as PSA, PSCA etc. (see, e.g., Alanen et al., Pathol. Res. Pract. 192(3): 233-237 (1996)).

Just as PSA polynucleotide fragments and polynucleotide variants are employed by skilled artisans for use in methods of monitoring PSA, 85P1B3 polynucleotide fragments and polynucleotide variants are used in an analogous manner. In particular, typical PSA polynucleotides used in methods of monitoring PSA are probes or primers which consist of fragments of the PSA cDNA sequence. Illustrating this, primers used to PCR amplify a PSA polynucleotide must include less than the whole PSA sequence to function in the polymerase chain reaction. In the context of such PCR reactions, skilled artisans generally create a variety of different polynucleotide fragments that can be used as primers in order to amplify different portions of a polynucleotide of interest or to optimize amplification reactions (see, e.g., Caetano-Anolles, G. Biotechniques 25(3): 472-476, 478-480 (1998); Robertson et al., Methods Mol. Biol. 98:121-154 (1998)). An additional illustration of the use of such fragments is provided in Example 4, where a 85P1B3 polynucleotide fragment is used as a probe to show the expression of 85P1B3 RNAs in cancer cells. In addition, variant polynucleotide sequences are typically used as primers and probes for the corresponding mRNAs in PCR and Northern analyses (see, e.g., Sawai et al., Fetal Diagn. Ther. 1996 Nov-Dec 11(6):407-13 and Current Protocols In Molecular Biology, Volume 2, Unit 2, Frederick M. Ausubel et al. eds., 1995)). Polynucleotide fragments and variants are useful in this context where they are capable of binding to a target polynucleotide sequence (e.g. the 85P1B3 polynucleotide shown in SEQ ID NO: 701) under conditions of high stringency.

Furthermore, PSA polypeptides which contain an epitope that can be recognized by an antibody or T cell that specifically binds to that epitope are used in methods of monitoring PSA. 85P1B3 polypeptide fragments and polypeptide analogs or variants can also be used in an analogous manner. This practice of using polypeptide fragments or polypeptide variants to generate antibodies (such as anti-PSA antibodies or T cells) is typical in the art with a wide variety of systems such as fusion proteins being used by practitioners (see, e.g., Current Protocols In Molecular Biology, Volume 2, Unit 16, Frederick M. Ausubel et al. eds., 1995). In this context, each epitope(s) functions to provide the architecture with which an

antibody or T cell is reactive. Typically, skilled artisans create a variety of different polypeptide fragments that can be used in order to generate immune responses specific for different portions of a polypeptide of interest (see, e.g., U.S. Patent No. 5,840,501 and U.S. Patent No. 5,939,533). For example it may be preferable to utilize a polypeptide comprising one of the 85P1B3 biological motifs discussed herein or a motif-bearing subsequence which is readily identified by one of skill in the art based on motifs available in the art. Polypeptide fragments, variants or analogs are typically useful in this context as long as they comprise an epitope capable of generating an antibody or T cell specific for a target polypeptide sequence (e.g. the 85P1B3 polypeptide shown in SEQ ID NO: 703).

As shown herein, the 85P1B3 polynucleotides and polypeptides (as well as the 85P1B3 polynucleotide probes and anti-85P1B3 antibodies or T cells used to identify the presence of these molecules) exhibit specific properties that make them useful in diagnosing cancers such as those listed in Table I. Diagnostic assays that measure the presence of 85P1B3 gene products, in order to evaluate the presence or onset of a disease condition described herein, such as prostate cancer, are used to identify patients for preventive measures or further monitoring, as has been done so successfully with PSA. Moreover, these materials satisfy a need in the art for molecules having similar or complementary characteristics to PSA in situations where, for example, a definite diagnosis of metastasis of prostatic origin cannot be made on the basis of a test for PSA alone (see, e.g., Alanen et al., Pathol. Res. Pract. 192(3): 233-237 (1996)), and consequently, materials such as 85P1B3 polynucleotides and polypeptides (as well as the 85P1B3 polynucleotide probes and anti-85P1B3 antibodies used to identify the presence of these molecules) must be employed to confirm metastases of prostatic origin.

Finally, in addition to their use in diagnostic assays, the 85P1B3 polynucleotides disclosed herein have a number of other utilities such as their use in the identification of oncogenetic associated chromosomal abnormalities in the chromosomal region to which the 85P1B3 gene maps (see Example 3 below). Moreover, in addition to their use in diagnostic assays, the 85P1B3-related proteins and polynucleotides disclosed herein have other utilities such as their use in the forensic analysis of tissues of unknown origin (see, e.g., Takahama K Forensic Sci Int 1996 Jun 28;80(1-2): 63-9).

Additionally, 85P1B3-related proteins or polynucleotides of the invention can be used to treat a pathologic condition characterized by the over-expression of 85P1B3. For example, the amino acid or nucleic acid sequence of Figure 2 or Figure 3, or fragments of either, can be used to generate an immune response to the 85P1B3 antigen. Antibodies or other molecules that react with 85P1B3 can be used to modulate the function of this molecule, and thereby provide a therapeutic benefit.

XII.) Inhibition of 85P1B3 Protein Function

The invention includes various methods and compositions for inhibiting the binding of 85P1B3 to its binding partner or its association with other protein(s) as well as methods for inhibiting 85P1B3 function.

XII.A.) Inhibition of 85P1B3 With Intracellular Antibodies

In one approach, a recombinant vector that encodes single chain antibodies that specifically bind to 85P1B3 are introduced into 85P1B3 expressing cells via gene transfer technologies. Accordingly, the encoded single chain anti-85P1B3 antibody is expressed intracellularly, binds to 85P1B3 protein, and thereby inhibits its function. Methods for engineering such intracellular single chain antibodies are well known. Such intracellular antibodies, also known as "intrabodies", are specifically targeted to a particular compartment within the cell, providing control over where the inhibitory activity of the treatment is focused. This technology has been successfully applied in the art (for review, see Richardson and Marasco, 1995, TIBTECH vol. 13). Intrabodies have been shown to virtually eliminate the expression of otherwise abundant cell surface receptors (see, e.g., Richardson et al., 1995, Proc. Natl. Acad. Sci. USA 92: 3137-3141; Beerli et al., 1994, J. Biol. Chem. 289: 23931-23936; Deshane et al., 1994, Gene Ther. 1: 332-337).

Single chain antibodies comprise the variable domains of the heavy and light chain joined by a flexible linker polypeptide, and are expressed as a single polypeptide. Optionally, single chain antibodies are expressed as a single chain variable region fragment joined to the light chain constant region. Well-known intracellular trafficking signals are engineered into recombinant polynucleotide vectors encoding such single chain antibodies in order to precisely target the intrabody to the desired intracellular compartment. For example, intrabodies targeted to the endoplasmic reticulum (ER) are engineered to incorporate a leader peptide and, optionally, a C-terminal ER retention signal, such as the KDEL amino acid motif. Intrabodies intended to exert activity in the nucleus are engineered to include a nuclear localization signal. Lipid moieties are joined to intrabodies in order to tether the intrabody to the cytosolic side of the plasma membrane. Intrabodies can also be targeted to exert function in the cytosol. For example, cytosolic intrabodies are used to sequester factors within the cytosol, thereby preventing them from being transported to their natural cellular destination.

In one embodiment, intrabodies are used to capture 85P1B3 in the nucleus, thereby preventing its activity within the nucleus. Nuclear targeting signals are engineered into such 85P1B3 intrabodies in order to achieve the desired targeting. Such 85P1B3 intrabodies are designed to bind specifically to a particular 85P1B3 domain. In another embodiment, cytosolic intrabodies that specifically bind to the 85P1B3 protein are used to prevent 85P1B3 from gaining access to the nucleus, thereby preventing it from exerting any biological activity within the nucleus (e.g., preventing 85P1B3 from forming transcription complexes with other factors).

In order to specifically direct the expression of such intrabodies to particular cells, the transcription of the intrabody is placed under the regulatory control of an appropriate tumor-specific promoter and/or enhancer. In order to target intrabody expression specifically to prostate, for example, the PSA promoter and/or promoter/enhancer can be utilized (See, for example, U.S. Patent No. 5,919,652 issued 6 July 1999).

XII.B.) Inhibition of 85P1B3 with Recombinant Proteins

In another approach, recombinant molecules bind to 85P1B3 and thereby inhibit 85P1B3 function. For example, these recombinant molecules prevent or inhibit 85P1B3 from accessing/binding to its binding partner(s) or associating with other protein(s). Such recombinant molecules can, for example, contain the reactive part(s) of a 85P1B3 specific antibody molecule. In a particular embodiment, the 85P1B3 binding domain of a 85P1B3 binding partner is engineered into a dimeric fusion protein, whereby the fusion protein comprises two 85P1B3 ligand binding domains linked to the Fc portion of a human IgG, such as human IgG1. Such IgG portion can contain, for example, the C_H2 and C_H3 domains and the hinge region, but not the C_H1 domain. Such dimeric fusion proteins are administered in soluble form to patients suffering from a cancer associated with the expression of 85P1B3, whereby the dimeric fusion protein specifically binds to 85P1B3 and blocks 85P1B3 interaction with a binding partner. Such dimeric fusion proteins are further combined into multimeric proteins using known antibody linking technologies.

XII.C.) Inhibition of 85P1B3 Transcription or Translation

The present invention also comprises various methods and compositions for inhibiting the transcription of the 85P1B3 gene. Similarly, the invention also provides methods and compositions for inhibiting the translation of 85P1B3 mRNA into protein.

In one approach, a method of inhibiting the transcription of the 85P1B3 gene comprises contacting the 85P1B3 gene with a 85P1B3 antisense polynucleotide. In another approach, a method of inhibiting 85P1B3 mRNA translation comprises contacting the 85P1B3 mRNA with an antisense polynucleotide. In another approach, a 85P1B3 specific ribozyme is used to cleave the 85P1B3 message, thereby inhibiting translation. Such antisense and ribozyme based methods can also be directed to the regulatory regions of the 85P1B3 gene, such as the 85P1B3 promoter and/or enhancer elements. Similarly, proteins capable of inhibiting a 85P1B3 gene transcription factor are used to inhibit 85P1B3 mRNA transcription. The various polynucleotides and compositions useful in the aforementioned methods have been described above. The use of antisense and ribozyme molecules to inhibit transcription and translation is well known in the art.

Other factors that inhibit the transcription of 85P1B3 by interfering with 85P1B3 transcriptional activation are also useful to treat cancers expressing 85P1B3. Similarly, factors that interfere with 85P1B3 processing are useful to treat cancers that express 85P1B3. Cancer treatment methods utilizing such factors are also within the scope of the invention.

XII.D.) General Considerations for Therapeutic Strategies

Gene transfer and gene therapy technologies can be used to deliver therapeutic polynucleotide molecules to tumor cells synthesizing 85P1B3 (i.e., antisense, ribozyme, polynucleotides encoding intrabodies and other 85P1B3 inhibitory molecules). A number of gene therapy approaches are known in the art. Recombinant vectors encoding 85P1B3 antisense polynucleotides, ribozymes, factors capable of interfering with 85P1B3 transcription, and so forth, can be delivered to target tumor cells using such gene therapy approaches.

The above therapeutic approaches can be combined with any one of a wide variety of surgical, chemotherapy or radiation therapy regimens. The therapeutic approaches of the invention can enable the use of reduced dosages of chemotherapy (or other therapies) and/or less frequent administration, an advantage for all patients and particularly for those that do not tolerate the toxicity of the chemotherapeutic agent well.

The anti-tumor activity of a particular composition (e.g., antisense, ribozyme, intrabody), or a combination of such compositions, can be evaluated using various *in vitro* and *in vivo* assay systems. *In vitro* assays that evaluate therapeutic activity include cell growth assays, soft agar assays and other assays indicative of tumor promoting activity, binding assays capable of determining the extent to which a therapeutic composition will inhibit the binding of 85P1B3 to a binding partner, etc.

In vivo, the effect of a 85P1B3 therapeutic composition can be evaluated in a suitable animal model. For example, xenogenic prostate cancer models can be used, wherein human prostate cancer explants or passaged xenograft tissues are introduced into immune compromised animals, such as nude or SCID mice (Klein et al., 1997, Nature Medicine 3: 402-408). For example, PCT Patent Application WO98/16628, Sawyers et al., published April 23, 1998, describes various xenograft models of human prostate cancer capable of recapitulating the development of primary tumors, micrometastasis, and the formation of osteoblastic metastases characteristic of late stage disease. Efficacy can be predicted using assays that measure inhibition of tumor formation, tumor regression or metastasis, and the like. *In vivo* assays that evaluate the promotion of apoptosis are useful in evaluating therapeutic compositions. In one embodiment, xenografts from tumor bearing mice treated with the therapeutic composition can be examined for the presence of apoptotic foci and compared to untreated control xenograft-bearing mice. The extent to which apoptotic foci are found in the tumors of the treated mice provides an indication of the therapeutic efficacy of the composition.

The therapeutic compositions used in the practice of the foregoing methods can be formulated into pharmaceutical compositions comprising a carrier suitable for the desired delivery method. Suitable carriers include any material that when combined with the therapeutic composition retains the anti-tumor function of the therapeutic composition and is generally non-reactive with the patient's immune system. Examples include, but are not limited to, any of a number of standard pharmaceutical carriers such as sterile

phosphate buffered saline solutions, bacteriostatic water, and the like (see, generally, Remington's Pharmaceutical Sciences 16th Edition, A. Osal., Ed., 1980).

Therapeutic formulations can be solubilized and administered via any route capable of delivering the therapeutic composition to the tumor site. Potentially effective routes of administration include, but are not limited to, intravenous, parenteral, intraperitoneal, intramuscular, intratumor, intradermal, intraorgan, orthotopic, and the like. A preferred formulation for intravenous injection comprises the therapeutic composition in a solution of preserved bacteriostatic water, sterile unpreserved water, and/or diluted in polyvinylchloride or polyethylene bags containing 0.9% sterile Sodium Chloride for Injection, USP. Therapeutic protein preparations can be lyophilized and stored as sterile powders, preferably under vacuum, and then reconstituted in bacteriostatic water (containing for example, benzyl alcohol preservative) or in sterile water prior to injection.

Dosages and administration protocols for the treatment of cancers using the foregoing methods will vary with the method and the target cancer, and will generally depend on a number of other factors appreciated in the art.

XIII.) Kits

For use in the diagnostic and therapeutic applications described herein, kits are also within the scope of the invention. Such kits can comprise a carrier, package or container that is compartmentalized to receive one or more containers such as vials, tubes, and the like, each of the container(s) comprising one of the separate elements to be used in the method. For example, the container(s) can comprise a probe that is or can be detectably labeled. Such probe can be an antibody or polynucleotide specific for a 85P1B3-related protein or a 85P1B3 gene or message, respectively. Where the method utilizes nucleic acid hybridization to detect the target nucleic acid, the kit can also have containers containing nucleotide(s) for amplification of the target nucleic acid sequence and/or a container comprising a reporter-means, such as a biotin-binding protein, such as avidin or streptavidin, bound to a reporter molecule, such as an enzymatic, florescent, or radioisotope label. The kit can include all or part of the amino acid sequence of Figure 2 or Figure 3 or analogs thereof, or a nucleic acid molecules that encodes such amino acid sequences.

The kit of the invention will typically comprise the container described above and one or more other containers comprising materials desirable from a commercial and user standpoint, including buffers, diluents, filters, needles, syringes, and package inserts with instructions for use.

A label can be present on the container to indicate that the composition is used for a specific therapy or non-therapeutic application, and can also indicate directions for either *in vivo* or *in vitro* use, such as those described above. Directions and or other information can also be included on an insert which is included with the kit.

EXAMPLES

Various aspects of the invention are further described and illustrated by way of the several examples that follow, none of which are intended to limit the scope of the invention.

Example 1: SSH-Generated Isolation of a cDNA Fragment of the 85P1B3 Gene

To isolate genes that are involved in the progression of androgen dependent (AD) prostate cancer to androgen independent (AI) cancer, we conducted an experiment with the LAPC-4 AD xenograft in male SCID mice. Mice that harbored LAPC-4 AD xenografts were castrated when the tumors reached a size of 1 cm in diameter. The tumors regressed in size and temporarily stopped producing the androgen dependent protein PSA. Seven to fourteen days post-castration, PSA levels were detectable again in the blood of the mice. Eventually the tumors develop an AI phenotype and start growing again in the castrated males. Tumors were harvested at different time points after castration to identify genes that are turned on or off during the transition to androgen independence.

The gene 85P1B3 was derived from an LAPC-4 AD (3 days post-castration) minus LAPC-4 AD subtraction. The SSH DNA sequence of 319 bp (Figure 1) is a fragment of the Opa-Interacting Protein 5 gene (OIP-5).

Materials and Methods

LAPC Xenografts and Human Tissues:

LAPC xenografts were obtained from Dr. Charles Sawyers (UCLA) and generated as described (Klein et al, 1997, Nature Med. 3: 402-408; Craft et al., 1999, Cancer Res. 59: 5030-5036). Androgen dependent and independent LAPC-4 AD and AI xenografts were grown in male SCID mice and were passaged as small tissue chunks in recipient males. LAPC-4 AI xenografts were derived from LAPC-4 AD tumors, respectively. To generate the AI xenografts, male mice bearing AD tumors were castrated and maintained for 2-3 months. After the tumors re-grew, the tumors were harvested and passaged in castrated males or in female SCID mice.

Cell Lines:

Human cell lines (e.g., HeLa) were obtained from the ATCC and were maintained in DMEM with 5% fetal calf serum.

RNA Isolation:

Tumor tissue and cell lines were homogenized in Trizol reagent (Life Technologies, Gibco BRL) using 10 ml/ g tissue or 10 ml/ 10^8 cells to isolate total RNA. Poly A RNA was purified from total RNA using Qiagen's Oligotex mRNA Mini and Midi kits. Total and mRNA were quantified by spectrophotometric analysis (O.D. 260/280 nm) and analyzed by gel electrophoresis.

Oligonucleotides:

The following HPLC purified oligonucleotides were used.

DPNCDN (cDNA synthesis primer):

5'TTTT**GATC**AAGCTT$_{30}$3' (SEQ ID NO: 714)

Adaptor 1:

5'CTAATACGACTCACTATAGGGCTCGAGCGGCCGCCCGGGCAG3' (SEQ ID NO: 715)

3'GGCCCGTC**CTAG**5' (SEQ ID NO: 716)

Adaptor 2:

5'GTAATACGACTCACTATAGGGCAGCGTGGTCGCGGCCGAG3' (SEQ ID NO:717)

3'CGGCTC**CTAG**5' (SEQ ID NO: 718)

PCR primer 1:

5'CTAATACGACTCACTATAGGGC3' (SEQ ID NO: 719)

Nested primer (NP)1:

5'TCGAGCGGCCGCCCGGGCAG**GA**3' (SEQ ID NO: 720)

Nested primer (NP)2:

5'AGCGTGGTCGCGGCCGAG**GA**3' (SEQ ID NO: 721)

Suppression Subtractive Hybridization:

Suppression Subtractive Hybridization (SSH) was used to identify cDNAs corresponding to genes that may be differentially expressed in prostate cancer. The SSH reaction utilized cDNA from two LAPC-4 AD xenografts. Specifically, to isolate genes that are involved in the progression of androgen dependent (AD) prostate cancer to androgen independent (AI) cancer, an experiment was conducted with the LAPC-4 AD xenograft in male SCID mice. Mice that harbored LAPC-4 AD xenografts were castrated when the tumors reached a size of 1 cm in diameter. The tumors regressed in size and temporarily stopped producing the androgen dependent protein PSA. Seven to fourteen days post-castration, PSA levels were detectable again in the blood of the mice. Eventually the tumors develop an AI phenotype and start growing again in the castrated males. Tumors were harvested at different time points after castration to identify genes that are turned on or off during the transition to androgen independence.

The gene 85P1B3 was derived from an LAPC-4 AD (3 days post-castration) minus LAPC-4 AD subtraction. The SSH DNA sequence (Figure 1) was identified.

The cDNA derived from an LAPC-4 AD tumor (grown in intact male mouse) was used as the source of the "driver" cDNA, while the cDNA from the LAPC-4 AD tumor (3 days post-castration) was used as the source of the "tester" cDNA. Double stranded cDNAs corresponding to tester and driver cDNAs were synthesized from 2 µg of poly(A)$^+$ RNA isolated from the relevant xenograft tissue, as described above, using CLONTECH's PCR-Select cDNA Subtraction Kit and 1 ng of oligonucleotide DPNCDN as primer. First- and second-strand synthesis were carried out as described in the Kit's user manual protocol (CLONTECH Protocol No. PT1117-1, Catalog No. K1804-1). The resulting cDNA was digested with Dpn II for 3 hrs at 37°C. Digested cDNA was extracted with phenol/chloroform (1:1) and ethanol precipitated.

Driver cDNA was generated by combining in a 1:1 ratio Dpn II digested cDNA from the relevant xenograft source (see above) with a mix of digested cDNAs derived from the human cell lines HeLa, 293, A431, Colo205, and mouse liver.

Tester cDNA was generated by diluting 1 µl of Dpn II digested cDNA from the relevant xenograft source (see above) (400 ng) in 5 µl of water. The diluted cDNA (2 µl, 160 ng) was then ligated to 2 µl of Adaptor 1 and Adaptor 2 (10 µM), in separate ligation reactions, in a total volume of 10 µl at 16°C overnight, using 400 u of T4 DNA ligase (CLONTECH). Ligation was terminated with 1 µl of 0.2 M EDTA and heating at 72°C for 5 min.

The first hybridization was performed by adding 1.5 µl (600 ng) of driver cDNA to each of two tubes containing 1.5 µl (20 ng) Adaptor 1- and Adaptor 2- ligated tester cDNA. In a final volume of 4 µl, the samples were overlaid with mineral oil, denatured in an MJ Research thermal cycler at 98°C for 1.5 minutes, and then were allowed to hybridize for 8 hrs at 68°C. The two hybridizations were then mixed together with an additional 1 µl of fresh denatured driver cDNA and were allowed to hybridize overnight at 68°C. The second hybridization was then diluted in 200 µl of 20 mM Hepes, pH 8.3, 50 mM NaCl, 0.2 mM EDTA, heated at 70°C for 7 min. and stored at -20°C.

PCR Amplification, Cloning and Sequencing of Gene Fragments Generated from SSH:

To amplify gene fragments resulting from SSH reactions, two PCR amplifications were performed. In the primary PCR reaction 1 µl of the diluted final hybridization mix was added to 1 µl of PCR primer 1 (10 µM), 0.5 µl dNTP mix (10 µM), 2.5 µl 10 x reaction buffer (CLONTECH) and 0.5 µl 50 x Advantage cDNA polymerase Mix (CLONTECH) in a final volume of 25 µl. PCR 1 was conducted using the following conditions: 75°C for 5 min., 94°C for 25 sec., then 27 cycles of 94°C for 10 sec, 66°C for 30 sec, 72°C for 1.5 min. Five separate primary PCR reactions were performed for each experiment. The

products were pooled and diluted 1:10 with water. For the secondary PCR reaction, 1 µl from the pooled and diluted primary PCR reaction was added to the same reaction mix as used for PCR 1, except that primers NP1 and NP2 (10 µM) were used instead of PCR primer 1. PCR 2 was performed using 10-12 cycles of 94°C for 10 sec, 68°C for 30 sec, and 72°C for 1.5 minutes. The PCR products were analyzed using 2% agarose gel electrophoresis.

The PCR products were inserted into pCR2.1 using the T/A vector cloning kit (Invitrogen). Transformed *E. coli* were subjected to blue/white and ampicillin selection. White colonies were picked and arrayed into 96 well plates and were grown in liquid culture overnight. To identify inserts, PCR amplification was performed on 1 ml of bacterial culture using the conditions of PCR1 and NP1 and NP2 as primers. PCR products were analyzed using 2% agarose gel electrophoresis.

Bacterial clones were stored in 20% glycerol in a 96 well format. Plasmid DNA was prepared, sequenced, and subjected to nucleic acid homology searches of the GenBank, dBest, and NCI-CGAP databases.

RT-PCR Expression Analysis:

First strand cDNAs can be generated from 1 µg of mRNA with oligo (dT)12-18 priming using the Gibco-BRL Superscript Preamplification system. The manufacturer's protocol was used which included an incubation for 50 min at 42°C with reverse transcriptase followed by RNAse H treatment at 37°C for 20 min. After completing the reaction, the volume can be increased to 200 µl with water prior to normalization. First strand cDNAs from 16 different normal human tissues can be obtained from Clontech.

Normalization of the first strand cDNAs from multiple tissues was performed by using the primers 5'atatcgccgcgctcgtcgtcgacaa3' (SEQ ID NO: 722) and 5'agccacacgcagctcattgtagaagg 3' (SEQ ID NO: 723) to amplify β-actin. First strand cDNA (5 µl) were amplified in a total volume of 50 µl containing 0.4 µM primers, 0.2 µM each dNTPs, 1XPCR buffer (Clontech, 10 mM Tris-HCL, 1.5 mM $MgCl_2$, 50 mM KCl, pH8.3) and 1X Klentaq DNA polymerase (Clontech). Five µl of the PCR reaction can be removed at 18, 20, and 22 cycles and used for agarose gel electrophoresis. PCR was performed using an MJ Research thermal cycler under the following conditions: Initial denaturation can be at 94°C for 15 sec, followed by a 18, 20, and 22 cycles of 94°C for 15, 65°C for 2 min, 72°C for 5 sec. A final extension at 72°C was carried out for 2 min. After agarose gel electrophoresis, the band intensities of the 283 b.p. β-actin bands from multiple tissues were compared by visual inspection. Dilution factors for the first strand cDNAs were calculated to result in equal β-actin band intensities in all tissues after 22 cycles of PCR. Three rounds of normalization can be required to achieve equal band intensities in all tissues after 22 cycles of PCR.

To determine expression levels of the 85P1B3 gene, 5 µl of normalized first strand cDNA were analyzed by PCR using 26, and 30 cycles of amplification. Semi-quantitative expression analysis can be achieved by comparing the PCR products at cycle numbers that give light band intensities.

A typical RT-PCR expression analysis is shown in Figure 10. RT-PCR expression analysis was performed on first strand cDNAs generated using pools of tissues from multiple samples. The cDNAs were shown to be normalized using beta-actin PCR. Strong expression of 85P1B3 was observed in xenograft pool, bladder cancer pool, kidney cancer pool, colon cancer pool, lung cancer pool, breast cancer pool, ovary cancer pool, and cancer metastasis pool. Lower levels of expression were observed in VP1, VP2, and prostate cancer pool.

Example 2: Full Length Cloning of 85P1B3

To isolate genes that are involved in the progression of androgen dependent (AD) prostate cancer to androgen independent (AI) cancer, an experiment was conducted with the LAPC-4 AD xenograft in male SCID mice. Mice that harbored LAPC-4 AD xenografts were castrated when the tumors reached a size of 1 cm in diameter. The tumors regressed in size and temporarily stopped producing the androgen dependent protein PSA. Seven to fourteen days post-castration, PSA levels were detectable again in the blood of the mice. Eventually the tumors develop an AI phenotype and start growing again in the castrated males. Tumors were harvested at different time points after castration to identify genes that are turned on or off during the transition to androgen independence.

The gene 85P1B3 was derived from an LAPC-4 AD (3 days post-castration minus LAPC-4 AD) (no castration) subtraction. The SSH DNA sequence (Figure 1) was designated 85P1B3. cDNA clone 85P1B3-clone A (Figure 2) was identified by screening a human testis library (Display Target, Pangene) using the 85P1B3 SSH DNA sequence. The cDNA (clone A) of 1,262 bp revealed an ORF encoding 229 amino acids (Figure 2 and Figure 3). The nucleotide and protein sequence of 85P1B3 corresponds to the OIP-5 gene (Figure 4). The 85P1B3 protein is predicted to be cytoplasmic using the PSORT program (URL psort.nibb.ac.jp:8800/ form.html).

Example 3: Chromosomal Localization

Chromosomal localization can implicate genes in disease pathogenesis. Several chromosome mapping approaches are available in the art, including fluorescent *in situ* hybridization (FISH), human/hamster radiation hybrid (RH) panels (Walter et al., 1994; Nature Genetics 7:22; Research Genetics, Huntsville Al), human-rodent somatic cell hybrid panels such as is available from the Coriell Institute (Camden, New Jersey), and genomic viewers utilizing BLAST homologies to sequenced and mapped genomic clones (NCBI, Bethesda, Maryland).

85P1B3 maps to chromosme 15q14, using 85P1B3 sequence and the NCBI BLAST tool: (http://www.ncbi.nlm.nih.gov/genome/seq/page.cgi?F=HsBlast.html&&ORG=Hs).

The chromosomal localization of 85P1B3 was also determined using the GeneBridge4 Human/Hamster radiation hybrid (RH) panel (Walter et al., 1994; Nature Genetics 7:22)(Research Genetics, Huntsville Al).

The following PCR primers were used:

85P1B3.1	5' catgggactctgcatcttaattcc 3'
85P1B3.2	5' caggttcaggctttattgctgtct 3'

The resulting 85P1B3 mapping vector for the 93 radiation hybrid panel DNAs (10010001010100010100000000000011010000001210110000101110010000101110001001010110011011010110101), and the mapping program available at the internet address http://www-genome.wi.mit.edu/cgi-bin/contig/rhmapper.pl, localize the 85P1B3 gene to chromosome 15q13.2- q14.

Of note, chromosome 15q13.2- q14 is a region implicated in cancers (Tomlinson et al., Gastroenterology 1999 Apr;116(4):789-95).

Example 4: Expression analysis of 85P1B3 in normal tissues and patient specimens

Analysis of 85P1B3 by RT-PCR is shown in Figure 10. Strong expression of 85P1B3 is observed in xenograft pool, bladder cancer pool, kidney cancer pool, colon cancer pool, lung cancer pool, breast cancer pool, ovary cancer pool, and cancer metastasis pool. Lower levels of expression are observed in VP1, VP2, and prostate cancer pool.

Extensive Northern blot analysis of 85P1B3 in 16 human normal tissues demonstrated that 85P1B3 expression is reminiscent of a cancer-testis gene (Figure 11). A 1.4 kb transcript was detected in testis but not in any other normal tissues. 85P1B3 expression was also shown in prostate cancer xenografts and in all cancer cell lines tested, such as in the cancers of the prostate (LAPC 4AD, LAPC 4AI, LAPC 9AD, LAPC 9AI, LNCaP, PC-3, DU145, Tsu-Pr1 and LAPC-4 CL), bladder (HT1197, SCaBER, UM-UC-3, TCCSUP, J82, 5637), 293T cell line, Ewing's sarcoma (EWS), brain (PFSK-1, T98G), bone (SK-ES-1, HOS, U-2 OS, RD-ES), lung (CALU-1, A427, NCI-H82, NCI-H146), kidney (769-P, A498, CAKI-1, SW839), breast (CAMA-1, DU4475, MCF-7, MDA-MB-435s), testicular (NTERRA-2, NCCIT, TERA-1, TERA-2), ovarian (OV-1063, PA-1, SW 626), pancreas (PANC-1, Bx PC-3, HPAC, Capan-1), colon

(Caco-2, LoVo, T84, Colo205), and cervical (A431) (Figure 12). These results indicate that 85P1B3 is a testis-specific gene that is upregulated in multiple cancers.

Expression of 85P1B3 was assayed in a panel of human patient tumors (T) and their respective matched normal tissues (N) on RNA dot blots (Figure 13). 85P1B3 expression was seen in the cancers of the breast, prostate, uterus, ovary, cervix, stomach and lung. The expression detected in normal adjacent tissues (isolated from diseased tissues) but not in normal tissues (isolated from healthy donors) may indicate that these tissues are not fully normal and that 85P1B3 may be expressed in early stage tumors. 85P1B3 was also found to be highly expressed in all human cancer cell lines tested, HeLa (cervical carcinoma), Daudi (Burkitt's lymphoma), K562 (CML), HL-60 (PML), G361 (melanoma), A549 (lung carcinoma), MOLT-4 (lymphoblastic leukemia), SW480 (colorectal carcinoma), and Raji (Burkitt's lymphoma).

Northern blot analysis on individual patient tumor specimens showed expression of 85P1B3 in two colon tumor tissues tested, and in the colon cancer cell lines Colo 205, LoVo, T84 and Caco-2, but not in normal colon (Figure 14).

Expression of 85P1B3 was also detected in the tumors of 4 out of 5 bladder cancer patients, and in all three bladder cancer cell lines tested, but not in normal bladder (Figure 15).

In lung cancer samples, 85P1B3 expression was observed in three lung tumor specimens, all three lung cancer cell lines tested, but not in normal lung (Figure 16).

In order to assay for androgen regulation of 85P1B3 expression, LAPC-9AD tumor cells were injected in male mice (Figure 17). When tumors reached a palpable size (0.3-0.5cm in diameter), mice were castrated and tumors harvested at different time points. RNA was isolated from the xenograft tissues and Northern blots with 10 μg of total RNA/lane were probed with the 85P1B3 SSH fragment. Results showed that expression of 85P1B3 is not affected by androgen deprivation, and therefore, is not androgen-regulated.

The restricted expression of 85P1B3 in normal tissues and the expression detected in bladder cancer, kidney cancer, colon cancer, lung cancer, prostate cancer, ovarian cancer, and breast cancer indicate that 85P1B3 is a therapeutic and/or prophylactic target and a prognostic and/or diagnostic marker for human cancers.

Example 5: Production of Recombinant 85P1B3 in Prokaryotic Systems

To express recombinant 85P1B3 in prokaryotic cells, the full or partial length 85P1B3 cDNA sequences can be cloned into any one of a variety of expression vectors known in the art. One or more of the following regions of 85P1B3 are expressed in these contructs, amino acids 1 to 229; or any 8, 9, 10, 11,

12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30 or more contiguous amino acids from 85P1B3, variants, or analogs thereof.

A. *In vitro* transcription and translation constructs:

pCRII: To generate 85P1B3 sense and anti-sense RNA probes for RNA *in situ* investigations, pCRII constructs (Invitrogen, Carlsbad CA) are generated encoding either all or fragments of the 85P1B3 cDNA. The pCRII vector has Sp6 and T7 promoters flanking the insert to drive the transcription of 85P1B3 RNA for use as probes in RNA *in situ* hybridization experiments. These probes are used to analyze the cell and tissue expression of 85P1B3 at the RNA level. Transcribed 85P1B3 RNA representing the cDNA amino acid coding region of the 85P1B3 gene is used in *in vitro* translation systems such as the TnT™ Coupled Reticulolysate Sytem (Promega, Corp., Madison, WI) to synthesize 85P1B3 protein.

B. Bacterial Constructs:

pGEX Constructs: To generate recombinant 85P1B3 proteins in bacteria that are fused to the Glutathione S-transferase (GST) protein, all or parts of the 85P1B3 cDNA protein coding sequence are fused to the GST gene by cloning into pGEX-6P-1 or any other GST- fusion vector of the pGEX family (Amersham Pharmacia Biotech, Piscataway, NJ). These constructs allow controlled expression of recombinant 85P1B3 protein sequences with GST fused at the amino-terminus and a six histidine epitope (6X His) at the carboxyl-terminus. The GST and 6X His tags permit purification of the recombinant fusion protein from induced bacteria with the appropriate affinity matrix and allow recognition of the fusion protein with anti-GST and anti-His antibodies. The 6X His tag is generated by adding 6 histidine codons to the cloning primer at the 3' end, e.g., of the open reading frame (ORF). A proteolytic cleavage site, such as the PreScission™ recognition site in pGEX-6P-1, may be employed such that it permits cleavage of the GST tag from 85P1B3-related protein. The ampicillin resistance gene and pBR322 origin permits selection and maintenance of the pGEX plasmids in *E. coli.*

In one embodiment, a GST-fusion protein encoding the full length 85P1B3 protein sequence (amino acids 1-229) was constructed and purified from induced bacteria. This preparation was then used as immunogen to generate a rabbit anti-85P1B3 polyclonal antibody (see the section entitled "Generation of 85P1B3 Polyclonal Antibodies". As can be seen in Figure 20A, the pAb strongly recognizes the original GST-fusion immunogen as well as 85P1B3 protein expressed in 293T cells (Figure 20B and Figure 20C).

pMAL Constructs: To generate, in bacteria, recombinant 85P1B3 proteins that are fused to maltose-binding protein (MBP), all or parts of the 85P1B3 cDNA protein coding sequence are fused to the MBP gene by cloning into the pMAL-c2X and pMAL-p2X vectors (New England Biolabs, Beverly, MA). These constructs allow controlled expression of recombinant 85P1B3 protein sequences with MBP fused at the amino-terminus and a 6X His epitope tag at the carboxyl-terminus. The MBP and 6X His tags permit purification of the recombinant protein from induced bacteria with the appropriate affinity matrix and allow

recognition of the fusion protein with anti-MBP and anti-His antibodies. The 6X His epitope tag is generated by adding 6 histidine codons to the 3' cloning primer. A Factor Xa recognition site permits cleavage of the pMAL tag from 85P1B3. The pMAL-c2X and pMAL-p2X vectors are optimized to express the recombinant protein in the cytoplasm or periplasm respectively. Periplasm expression enhances folding of proteins with disulfide bonds.

pET Constructs: To express 85P1B3 in bacterial cells, all or parts of the 85P1B3 cDNA protein coding sequence are cloned into the pET family of vectors (Novagen, Madison, WI). These vectors allow tightly controlled expression of recombinant 85P1B3 protein in bacteria with and without fusion to proteins that enhance solubility, such as NusA and thioredoxin (Trx), and epitope tags, such as 6X His and S-Tag ™ that aid purification and detection of the recombinant protein. For example, constructs are made utilizing pET NusA fusion system 43.1 such that regions of the 85P1B3 protein are expressed as amino-terminal fusions to NusA.

C. Yeast Constructs:

pESC Constructs: To express 85P1B3 in the yeast species *Saccharomyces cerevisiae* for generation of recombinant protein and functional studies, all or parts of the 85P1B3 cDNA protein coding sequence are cloned into the pESC family of vectors each of which contain 1 of 4 selectable markers, HIS3, TRP1, LEU2, and URA3 (Stratagene, La Jolla, CA). These vectors allow controlled expression from the same plasmid of up to 2 different genes or cloned sequences containing either Flag™ or Myc epitope tags in the same yeast cell. This system is useful to confirm protein-protein interactions of 85P1B3. In addition, expression in yeast yields similar post-translational modifications, such as glycosylations and phosphorylations, that are found when expressed in eukaryotic cells.

pESP Constructs: To express 85P1B3 in the yeast species *Saccharomyces pombe*, all or parts of the 85P1B3 cDNA protein coding sequence are cloned into the pESP family of vectors. These vectors allow controlled high level of expression of a 85P1B3 protein sequence that is fused at either the amino terminus or at the carboxyl terminus to GST which aids purification of the recombinant protein. A Flag™ epitope tag allows detection of the recombinant protein with anti- Flag™ antibody.

Example 6: Production of Recombinant 85P1B3 in Eukaryotic Systems

A. Mammalian Constructs:

To express recombinant 85P1B3 in eukaryotic cells, the full or partial length 85P1B3 cDNA sequences can be cloned into any one of a variety of expression vectors known in the art. One or more of the following regions of 85P1B3 are expressed in these contructs, amino acids 1 to 229; or any 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30 or more contiguous amino acids from 85P1B3, variants, or analogs thereof.

The constructs can be transfected into any one of a wide variety of mammalian cells such as 293T cells. Transfected 293T cell lysates can be probed with the anti-85P1B3 polyclonal serum, described herein.

pcDNA4/HisMax Constructs: To express 85P1B3 in mammalian cells, the 85P1B3 ORF, or portions thereof, of 85P1B3 are cloned into pcDNA4/HisMax Version A (Invitrogen, Carlsbad, CA). Protein expression is driven from the cytomegalovirus (CMV) promoter and the SP16 translational enhancer. The recombinant protein has Xpress™ and six histidine (6X His) epitopes fused to the amino-terminus. The pcDNA4/HisMax vector also contains the bovine growth hormone (BGH) polyadenylation signal and transcription termination sequence to enhance mRNA stability along with the SV40 origin for episomal replication and simple vector rescue in cell lines expressing the large T antigen. The Zeocin resistance gene allows for selection of mammalian cells expressing the protein and the ampicillin resistance gene and ColE1 origin permits selection and maintenance of the plasmid in *E. coli.*

pcDNA3.1/MycHis Constructs: To express 85P1B3 in mammalian cells, the 85P1B3 ORF, or portions thereof, of 85P1B3 with a consensus Kozak translation initiation site are cloned into pcDNA3.1/MycHis Version A (Invitrogen, Carlsbad, CA). Protein expression is driven from the cytomegalovirus (CMV) promoter. The recombinant proteins have the myc epitope and 6X His epitope fused to the carboxyl-terminus. The pcDNA3.1/MycHis vector also contains the bovine growth hormone (BGH) polyadenylation signal and transcription termination sequence to enhance mRNA stability, along with the SV40 origin for episomal replication and simple vector rescue in cell lines expressing the large T antigen. The Neomycin resistance gene can be used, as it allows for selection of mammalian cells expressing the protein and the ampicillin resistance gene and ColE1 origin permits selection and maintenance of the plasmid in *E. coli.*

pcDNA3.1/CT-GFP-TOPO Construct: To express 85P1B3 in mammalian cells and to allow detection of the recombinant proteins using fluorescence, the 85P1B3 ORF, or portions thereof, of 85P1B3 with a consensus Kozak translation initiation site are cloned into pcDNA3.1/CT-GFP-TOPO (Invitrogen, CA). Protein expression is driven from the cytomegalovirus (CMV) promoter. The recombinant proteins have the Green Fluorescent Protein (GFP) fused to the carboxyl-terminus facilitating non-invasive, in vivo detection and cell biology studies. The pcDNA3.1CT-GFP-TOPO vector also contains the bovine growth hormone (BGH) polyadenylation signal and transcription termination sequence to enhance mRNA stability along with the SV40 origin for episomal replication and simple vector rescue in cell lines expressing the large T antigen. The Neomycin resistance gene allows for selection of mammalian cells that express the protein, and the ampicillin resistance gene and ColE1 origin permits selection and maintenance of the plasmid in *E. coli.* Additional constructs with an amino-terminal GFP fusion are made in pcDNA3.1/NT-GFP-TOPO spanning the entire length of the 85P1B3 proteins.

PAPtag: The 85P1B3 ORF, or portions thereof, of 85P1B3 are cloned into pAPtag-5 (GenHunter Corp. Nashville, TN). This construct generates an alkaline phosphatase fusion at the carboxyl-terminus of the 85P1B3 proteins while fusing the IgGκ signal sequence to the amino-terminus. Constructs are also generated in which alkaline phosphatase with an amino-terminal IgGκ signal sequence is fused to the amino-terminus of 85P1B3 proteins. The resulting recombinant 85P1B3 proteins are optimized for secretion into the media of transfected mammalian cells and can be used to identify proteins such as ligands or receptors that interact with the 85P1B3 proteins. Protein expression is driven from the CMV promoter and the recombinant proteins also contain myc and 6X His epitopes fused at the carboxyl-terminus that facilitates detection and purification. The Zeocin resistance gene present in the vector allows for selection of mammalian cells expressing the recombinant protein and the ampicillin resistance gene permits selection of the plasmid in *E. coli.*

ptag5: The 85P1B3 ORF, or portions thereof, of 85P1B3 was cloned into pTag-5. This vector is similar to pAPtag but without the alkaline phosphatase fusion. This construct generated 85P1B3 protein with an amino-terminal IgGκ signal sequence and myc and 6X His epitope tags at the carboxyl-terminus that facilitate detection and affinity purification. The resulting recombinant 85P1B3 protein was optimized for secretion into the media of transfected mammalian cells, and was used as immunogen or ligand to identify proteins such as ligands or receptors that interact with the 85P1B3 proteins. Protein expression is driven from the CMV promoter. The Zeocin resistance gene present in the vector allows for selection of mammalian cells expressing the protein, and the ampicillin resistance gene permits selection of the plasmid in *E. coli.*

PsecFc: The 85P1B3 ORF, or portions thereof, of 85P1B3 are also cloned into psecFc. The psecFc vector was assembled by cloning the human immunoglobulin G1 (IgG) Fc (hinge, CH2, CH3 regions) into pSecTag2 (Invitrogen, California). This construct generates an IgG1 Fc fusion at the carboxyl-terminus of the 85P1B3 proteins, while fusing the IgGK signal sequence to N-terminus. 85P1B3 fusions utilizing the murine IgG1 Fc region are also used. The resulting recombinant 85P1B3 proteins are optimized for secretion into the media of transfected mammalian cells, and can be used as immunogens or to identify proteins such as ligands or receptors that interact with the 85P1B3 protein. Protein expression is driven from the CMV promoter. The hygromycin resistance gene present in the vector allows for selection of mammalian cells that express the recombinant protein, and the ampicillin resistance gene permits selection of the plasmid in *E. coli.*

pSRα Constructs: To generate mammalian cell lines that express 85P1B3, or portions thereof, constitutively, the ORF of 85P1B3 was cloned into pSRα constructs. Amphotropic and ecotropic retroviruses were generated by transfection of pSRα constructs into the 293T-10A1 packaging line or co-

transfection of pSRα and a helper plasmid (containing deleted packaging sequences) into the 293 cells, respectively. The retrovirus was used to infect a variety of mammalian cell lines, resulting in the integration of the cloned gene, 85P1B3, into the host cell-lines. Protein expression is driven from a long terminal repeat (LTR). The Neomycin resistance gene present in the vector allows for selection of mammalian cells that express the protein, and the ampicillin resistance gene and ColE1 origin permit selection and maintenance of the plasmid in *E. coli*. Figure18 shows expression of 85P1B3 using the pSRα retroviral vector in the prostate cancer cell line PC3. The retroviral vectors can thereafter be used for infection and generation of various cell lines using, for example, SCaBER, NIH 3T3, TsuPr1, 293 or rat-1 cells.

Additional pSRα constructs are made that fuse an epitope tag such as the FLAG™ tag to the carboxyl-terminus of 85P1B3 sequences to allow detection using anti-Flag antibodies. For example, the FLAG™ sequence 5' gat tac aag gat gac gac gat aag 3' is added to cloning primer at the 3' end of the ORF. Additional pSRα constructs are made to produce both amino-terminal and carboxyl-terminal GFP and myc/6X His fusion proteins of the full-length 85P1B3 proteins.

Additional Viral Vectors: Additional constructs are made for viral-mediated delivery and expression of 85P1B3. High virus titer leading to high level expression of 85P1B3 is achieved in viral delivery systems such as adenoviral vectors and herpes amplicon vectors. The 85P1B3 coding sequences or fragments thereof are amplified by PCR and subcloned into the AdEasy shuttle vector (Stratagene). Recombination and virus packaging are performed according to the manufacturer's instructions to generate adenoviral vectors. Alternatively, 85P1B3 coding sequences or fragments thereof are cloned into the HSV-1 vector (Imgenex) to generate herpes viral vectors. The viral vectors are thereafter used for infection of various cell lines such as SCaBER, NIH 3T3, 293 or rat-1 cells.

Regulated Expression Systems: To control expression of 85P1B3 in mammalian cells, coding sequences of 85P1B3, or portions thereof, are cloned into regulated mammalian expression systems such as the T-Rex System (Invitrogen), the GeneSwitch System (Invitrogen) and the tightly-regulated Ecdysone System (Sratagene). These systems allow the study of the temporal and concentration dependent effects of recombinant 85P1B3. These vectors are thereafter used to control expression of 85P1B3 in various cell lines such as SCaBER, NIH 3T3, 293 or rat-1 cells.

B. Baculovirus Expression Systems

To generate recombinant 85P1B3 proteins in a baculovirus expression system, 85P1B3 ORF, or portions thereof, are cloned into the baculovirus transfer vector pBlueBac 4.5 (Invitrogen), which provides a His-tag at the N-terminus. Specifically, pBlueBac-85P1B3 is co-transfected with helper plasmid pBac-N-Blue (Invitrogen) into SF9 (*Spodoptera frugiperda*) insect cells to generate recombinant baculovirus (see

Invitrogen instruction manual for details). Baculovirus is then collected from cell supernatant and purified by plaque assay.

Recombinant 85P1B3 protein is then generated by infection of HighFive insect cells (Invitrogen) with purified baculovirus. Recombinant 85P1B3 protein can be detected using anti-85P1B3 or anti-His-tag antibody. 85P1B3 protein can be purified and used in various cell-based assays or as immunogen to generate polyclonal and monoclonal antibodies specific for 85P1B3.

Example 7 Antigenicity Profiles and Secondary Structure

Figure 5, Figure 6, Figure 7, Figure 8, and Figure 9 depict graphically five amino acid profiles of the 85P1B3 amino acid sequence, each assessment available by accessing the ProtScale website (URL www.expasy.ch/cgi-bin/protscale.pl) on the ExPasy molecular biology server.

These profiles: Figure 5, Hydrophilicity, (Hopp T.P., Woods K.R., 1981. Proc. Natl. Acad. Sci. U.S.A. 78:3824-3828); Figure 6, Hydropathicity, (Kyte J., Doolittle R.F., 1982. J. Mol. Biol. 157:105-132); Figure 7, Percentage Accessible Residues (Janin J., 1979 Nature 277:491-492); Figure 8, Average Flexibility, (Bhaskaran R., and Ponnuswamy P.K., 1988. Int. J. Pept. Protein Res. 32:242-255); Figure 9, Beta-turn (Deleage, G., Roux B. 1987 Protein Engineering 1:289-294); and optionally others available in the art, such as on the ProtScale website, were used to identify antigenic regions of the 85P1B3 protein. Each of the above amino acid profiles of 85P1B3 were generated using the following ProtScale parameters for analysis: 1) A window size of 9; 2) 100% weight of the window edges compared to the window center; and, 3) amino acid profile values normalized to lie between 0 and 1.

Hydrophilicity (Figure 5), Hydropathicity (Figure 6) and Percentage Accessible Residues (Figure 7) profiles were used to determine stretches of hydrophilic amino acids (i.e., values greater than 0.5 on the Hydrophilicity and Percentage Accessible Residues profile, and values less than 0.5 on the Hydropathicity profile). Such regions are likely to be exposed to the aqueous environment, be present on the surface of the protein, and thus available for immune recognition, such as by antibodies.

Average Flexibility (Figure 8) and Beta-turn (Figure 9) profiles determine stretches of amino acids (i.e., values greater than 0.5 on the Beta-turn profile and the Average Flexibility profile) that are not constrained in secondary structures such as beta sheets and alpha helices. Such regions are also more likely to be exposed on the protein and thus accessible to immune recognition, such as by antibodies.

Antigenic sequences of the 85P1B3 protein indicated, e.g., by the profiles set forth in Figure 5, Figure 6, Figure 7, Figure 8, and/or Figure 9 are used to prepare immunogens, either peptides or nucleic acids that encode them, to generate therapeutic and diagnostic anti-85P1B3 antibodies. The immunogen can be any 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 30, 35, 40, 45, 50 or more than 50 contiguous amino acids, or the corresponding nucleic acids that encode them, from the

85P1B3 protein. In particular, peptide immunogens of the invention can comprise, a peptide region of at least 5 amino acids of Figure 2 in any whole number increment up to 229 that includes an amino acid position having a value greater than 0.5 in the Hydrophilicity profile of Figure 5; a peptide region of at least 5 amino acids of Figure 2 in any whole number increment up to 229 that includes an amino acid position having a value less than 0.5 in the Hydropathicity profile of Figure 6; a peptide region of at least 5 amino acids of Figure 2 in any whole number increment up to 229 that includes an amino acid position having a value greater than 0.5 in the Percent Accessible Residues profile of Figure 7; a peptide region of at least 5 amino acids of Figure 2 in any whole number increment up to 229 that includes an amino acid position having a value greater than 0.5 in the Average Flexibility profile on Figure 8; and, a peptide region of at least 5 amino acids of Figure 2 in any whole number increment up to 229 that includes an amino acid position having a value greater than 0.5 in the Beta-turn profile of Figure 9. Peptide immunogens of the invention can also comprise nucleic acids that encode any of the forgoing.

All immunogens of the invention, peptide or nucleic acid, can be embodied in human unit dose form, or comprised by a composition that includes a pharmaceutical excipient compatible with human physiology.

The secondary structure of 85P1B3, namely the predicted presence and location of alpha helices, extended strands, and random coils, is predicted from the primary amino acid sequence using the HNN - Hierarchical Neural Network method (Guermeur, 1997, http://pbil.ibcp.fr/cgi-bin/npsa_automat.pl?page=npsa_nn.html), accessed from the ExPasy molecular biology server (http://www.expasy.ch/tools/). The analysis indicates that 85P1B3 is composed of 36.8% alpha helix, 13.97% extended strand, and 49.34% random coil (Figure 21A).

Analysis for the potential presence of transmembrane domains in 85P1B3 was carried out using a variety of transmembrane prediction algorithms accessed from the ExPasy molecular biology server (http://www.expasy.ch/tools/). A potential transmembrane domain composed of amino acids 129-149 is predicted by the TMpred program (Figure 21B). HMMTop predicts a transmembrane region from amino acids 134-158. The SOSUI and TMHMM (Figure 21C) programs predict that 85P1B3 is a soluble protein without transmembrane domains. The results of the transmembrane predictions are summarized in Table XXV.

Example 8: Generation of 85P1B3 Polyclonal Antibodies

Polyclonal antibodies can be raised in a mammal, for example, by one or more injections of an immunizing agent and, if desired, an adjuvant. Typically, the immunizing agent and/or adjuvant will be injected in the mammal by multiple subcutaneous or intraperitoneal injections. In addition to immunizing with the full length 85P1B3 protein, computer algorithms are employed in design of immunogens that,

based on amino acid sequence analysis contain characteristics of being antigenic and available for recognition by the immune system of the immunized host (see the Example entitled "Antigenicity Profiles"). Such regions would be predicted to be hydrophilic, flexible, in beta-turn conformations, and be exposed on the surface of the protein (see, e.g., Figure 5, Figure 6, Figure 7, Figure 8, or Figure 9 for amino acid profiles that indicate such regions of 85P1B3).

For example, 85P1B3 recombinant bacterial fusion proteins or peptides encoding hydrophilic, flexible, beta-turn regions of the 85P1B3 sequence, such as amino acids 1-77 and 190-229 are used as antigens to generate polyclonal antibodies in New Zealand White rabbits. It is useful to conjugate the immunizing agent to a protein known to be immunogenic in the mammal being immunized. Examples of such immunogenic proteins include, but are not limited to, keyhole limpet hemocyanin (KLH), serum albumin, bovine thyroglobulin, and soybean trypsin inhibitor. In one embodiment, a peptide encoding amino acids 190-206 of 85P1B3 is conjugated to KLH and used to immunize the rabbit. Alternatively the immunizing agent may include all or portions of the 85P1B3 protein, analogs or fusion proteins thereof. For example, the 85P1B3 amino acid sequence can be fused using recombinant DNA techniques to any one of a variety of fusion protein partners that are well known in the art, such as glutathione-S-transferase (GST) and HIS tagged fusion proteins. Such fusion proteins are purified from induced bacteria using the appropriate affinity matrix.

In one embodiment, a GST-fusion protein encoding the full length 85P1B3 protein sequence was produced and purified and used as immunogen (see the section entitled "Production of 85P1B3 in Prokaryotic Systems"). Shorter sequences are also fused to GST in order to direct antibody to specific regions of the protein such as amino acids 1-77 to generate amino-terminal specific antibodies. Other recombinant bacterial fusion proteins that may be employed include maltose binding protein, LacZ, thioredoxin, NusA, or an immunoglobulin constant region (see the section entitled "Production of 85P1B3 in Prokaryotic Systems" and Current Protocols In Molecular Biology, Volume 2, Unit 16, Frederick M. Ausubul et al. eds., 1995; Linsley, P.S., Brady, W., Urnes, M., Grosmaire, L., Damle, N., and Ledbetter, L.(1991) J.Exp. Med. 174, 561-566).

In addition to bacterial derived fusion proteins, mammalian expressed protein antigens are also used. These antigens are expressed from mammalian expression vectors such as the Tag5 and Fc-fusion vectors (see the section entitled "Production of Recombinant 85P1B3 in Eukaryotic Systems"), and retain post-translational modifications such as glycosylations found in native protein. In one embodiment, a predicted antigenic region of 85P1B3, amino acids 190-229, is cloned into the Tag5 mammalian secretion vector. The recombinant protein is purified by metal chelate chromatography from tissue culture supernatants of 293T cells stably expressing the recombinant vector. The purified Tag5 85P1B3 protein is then used as immunogen.

During the immunization protocol, it is useful to mix or emulsify the antigen in adjuvants that enhance the immune response of the host animal. Examples of adjuvants include, but are not limited to, complete Freund's adjuvant (CFA) and MPL-TDM adjuvant (monophosphoryl Lipid A, synthetic trehalose dicorynomycolate).

In a typical protocol, rabbits are initially immunized subcutaneously with up to 200 μg, typically 100-200 μg, of fusion protein or peptide conjugated to KLH mixed in complete Freund's adjuvant (CFA). Rabbits are then injected subcutaneously every two weeks with up to 200 μg, typically 100-200 μg, of the immunogen in incomplete Freund's adjuvant (IFA). Test bleeds are taken approximately 7-10 days following each immunization and used to monitor the titer of the antiserum by ELISA.

To test reactivity and specificity of immune serum, such as the rabbit serum raised from immunization with GST-85P1B3 full length fusion protein, the full-length 85P1B3 cDNA was cloned into pCDNA 3.1 myc-his expression vector (Invitrogen, see the Example entitled "Production of Recombinant 85P1B3 in Eukaryotic Systems"). After transfection of the constructs into 293T cells, cell lysates were probed with the anti-85P1B3 serum and with anti-His antibody (Santa Cruz Biotechnologies, Santa Cruz, CA) to determine specific reactivity to denatured 85P1B3 protein using the Western blot technique. As can be seen in Figure 20B, the anti-85P1B3 pAb specifically recognized 85P1B3 protein expressed in 293T cells that is the same molecular weight as that detected by the anti-His Ab (Figure 20C). Recognition of native protein by the antiserum is determined by immunoprecipitation and flow cytometric analyses of 293T and other recombinant 85P1B3-expressing cells. In addition, specificity of the antiserum is tested by Western blot, immunoprecipitation, fluorescent microscopy, and flow cytometric techniques using cells that endogenously express 85P1B3.

To purify the anti-serum derived from the GST-85P1B3 immunized rabbit, the serum was passed over an affinity column composed of GST to remove anti-GST reactive antibodies. The serum was then further purified by protein G affinity chromatography to isolate the IgG fraction. Serum from rabbits immunized with other fusion proteins, such as MBP fusion proteins, are purified by depletion of antibodies reactive to MBP, or other fusion partner sequence, by passage over an affinity column containing the fusion partner either alone or in the context of an irrelevant fusion protein. Sera from His-tagged protein and peptide immunized rabbits as well as fusion partner depleted sera are further purified by passage over an affinity column composed of the original protein immunogen or free peptide coupled to Affigel matrix (BioRad).

Example 9: Generation of 85P1B3 Monoclonal Antibodies (mAbs)

In one embodiment, therapeutic mAbs to 85P1B3 comprise those that react with epitopes of the protein that would disrupt or modulate the biological function of 85P1B3, for example those that would

disrupt its interaction with ligands or proteins that mediate or are involved in its biological activity. Therapeutic mAbs also comprise those which specifically bind epitopes of 85P1B3 exposed on the cell surface and thus are useful in targeting mAb-toxin conjugates. Immunogens for generation of such mAbs include those designed to encode or contain the entire 85P1B3 protein or regions of the 85P1B3 protein predicted to be antigenic from computer analysis of the amino acid sequence (see, e.g., Figure 5, Figure 6, Figure 7, Figure 8, or Figure 9, and the Example entitled "Antigenicity Profiles").

Immunogens include peptides, recombinant bacterial proteins, and mammalian expressed Tag 5 proteins and human and murine IgG FC fusion proteins. To generate mAbs to 85P1B3, mice are first immunized intraperitoneally (IP) with, typically, 10-50 μg of protein immunogen mixed in complete Freund's adjuvant. Mice are then subsequently immunized IP every 2-4 weeks with, typically, 10-50 μg of antigen mixed in incomplete Freund's adjuvant. Alternatively, MPL-TDM adjuvant is used in immunizations. In addition, a DNA-based immunization protocol is employed in which a mammalian expression vector encoding 85P1B3 sequence is used to immunize mice by direct injection of the plasmid DNA. For example, either pCDNA 3.1 encoding the full length 85P1B3 cDNA, amino acids 1-77, or 190-229 of 85P1B3 (predicted to be antigenic from sequence analysis, see, e.g., Figure 5, Figure 6, Figure 7, Figure 8 or Figure 9) fused at the amino-terminus to an IgK leader sequence and at the carboxyl-terminus to the coding sequence of murine or human IgG Fc region, is used. This protocol is used alone and in combination with protein immunogens. Test bleeds are taken 7-10 days following immunization to monitor titer and specificity of the immune response. Once appropriate reactivity and specificity is obtained as determined by ELISA, Western blotting, immunoprecipitation, fluorescence microscopy, and flow cytometric analyses, fusion and hybridoma generation is then carried out with established procedures well known in the art (see, e.g., Harlow and Lane, 1988).

In one embodiment for generating 85P1B3 monoclonal antibodies, a glutathione-S-transferase (GST) fusion protein encoding the full length 85P1B3 protein is expressed and purified. A cleavage fragment encoding 85P1B3 specific amino acids is then used as immunogen in which GST is removed by site-specific proteolysis. Balb C mice are initially immunized intraperitoneally with 25 μg of the 85P1B3 cleavage protein mixed in complete Freund's adjuvant. Mice are subsequently immunized every two weeks with 25 μg of 85P1B3 cleavage protein mixed in incomplete Freund's adjuvant for a total of three immunizations. The titer of serum from immunized mice is determined by ELISA using the full length GST-fusion protein and the cleaved immunogen. Reactivity and specificity of serum to full length 85P1B3 protein is monitored by Western blotting, immunoprecipitation and flow cytometry using 293T cells transfected with an expression vector encoding the 85P1B3 cDNA. Other recombinant 85P1B3-expressing cells (see e.g., the Example entitled "Production of 85P1B3 in Eukaryotic Systems") or cells endogenously

expressing 85P1B3 are also used. Mice showing the strongest reactivity are rested and given a final injection of 85P1B3 cleavage protein in PBS and then sacrificed four days later. The spleens of the sacrificed mice are harvested and fused to SPO/2 myeloma cells using standard procedures (Harlow and Lane, 1988). Supernatants from growth wells following HAT selection are screened by ELISA, Western blot, immunoprecipitation, fluorescent microscopy, and flow cytometry to identify 85P1B3 specific antibody-producing clones.

The binding affinity of a 85P1B3 monoclonal antibody is determined using standard technologies. Affinity measurements quantify the strength of antibody to epitope binding and are used to help define which 85P1B3 monoclonal antibodies preferred for diagnostic or therapeutic use, as appreciated by one of skill in the art. The BIAcore system (Uppsala, Sweden) is a preferred method for determining binding affinity. The BIAcore system uses surface plasmon resonance (SPR, Welford K. 1991, Opt. Quant. Elect. 23:1; Morton and Myszka, 1998, Methods in Enzymology 295: 268) to monitor biomolecular interactions in real time. BIAcore analysis conveniently generates association rate constants, dissociation rate constants, equilibrium dissociation constants, and affinity constants.

Example 10: HLA Class I and Class II Binding Assays

HLA class I and class II binding assays using purified HLA molecules are performed in accordance with disclosed protocols (*e.g.*, PCT publications WO 94/20127 and WO 94/03205; Sidney *et al.*, *Current Protocols in Immunology* 18.3.1 (1998); Sidney, *et al.*, *J. Immunol.* 154:247 (1995); Sette, *et al.*, *Mol. Immunol.* 31:813 (1994)). Briefly, purified MHC molecules (5 to 500 nM) are incubated with various unlabeled peptide inhibitors and 1-10 nM ^{125}I-radiolabeled probe peptides as described. Following incubation, MHC-peptide complexes are separated from free peptide by gel filtration and the fraction of peptide bound is determined. Typically, in preliminary experiments, each MHC preparation is titered in the presence of fixed amounts of radiolabeled peptides to determine the concentration of HLA molecules necessary to bind 10-20% of the total radioactivity. All subsequent inhibition and direct binding assays are performed using these HLA concentrations.

Since under these conditions [label]<[HLA] and $IC_{50} \geq$[HLA], the measured IC_{50} values are reasonable approximations of the true K_D values. Peptide inhibitors are typically tested at concentrations ranging from 120 μg/ml to 1.2 ng/ml, and are tested in two to four completely independent experiments. To allow comparison of the data obtained in different experiments, a relative binding figure is calculated for each peptide by dividing the IC_{50} of a positive control for inhibition by the IC_{50} for each tested peptide (typically unlabeled versions of the radiolabeled probe peptide). For database purposes, and inter-experiment comparisons, relative binding values are compiled. These values can subsequently be

converted back into IC_{50} nM values by dividing the IC_{50} nM of the positive controls for inhibition by the relative binding of the peptide of interest. This method of data compilation is accurate and consistent for comparing peptides that have been tested on different days, or with different lots of purified MHC.

Binding assays as outlined above may be used to analyze HLA supermotif and/or HLA motif-bearing peptides.

Example 11: Identification of HLA Supermotif- and Motif-Bearing CTL Candidate Epitopes

HLA vaccine compositions of the invention can include multiple epitopes. The multiple epitopes can comprise multiple HLA supermotifs or motifs to achieve broad population coverage. This example illustrates the identification and confirmation of supermotif- and motif-bearing epitopes for the inclusion in such a vaccine composition. Calculation of population coverage is performed using the strategy described below.

Computer searches and algorithms for identification of supermotif and/or motif-bearing epitopes

The searches performed to identify the motif-bearing peptide sequences in the Example entitled "Antigenicity Profiles" and Tables V-XVIII employ the protein sequence data from the gene product of 85P1B3 set forth in Figures 2 and 3.

Computer searches for epitopes bearing HLA Class I or Class II supermotifs or motifs are performed as follows. All translated 85P1B3 protein sequences are analyzed using a text string search software program to identify potential peptide sequences containing appropriate HLA binding motifs; such programs are readily produced in accordance with information in the art in view of known motif/supermotif disclosures. Furthermore, such calculations can be made mentally.

Identified A2-, A3-, and DR-supermotif sequences are scored using polynomial algorithms to predict their capacity to bind to specific HLA-Class I or Class II molecules. These polynomial algorithms account for the impact of different amino acids at different positions, and are essentially based on the premise that the overall affinity (or ΔG) of peptide-HLA molecule interactions can be approximated as a linear polynomial function of the type:

$$\text{“}\Delta G\text{”} = a_{1i} \times a_{2i} \times a_{3i} \ldots\ldots \times a_{ni}$$

where a_{ji} is a coefficient which represents the effect of the presence of a given amino acid (j) at a given position (i) along the sequence of a peptide of n amino acids. The crucial assumption of this method is that the effects at each position are essentially independent of each other (i.e., independent binding of individual side-chains). When residue j occurs at position i in the peptide, it is assumed to contribute a constant amount j_i to the free energy of binding of the peptide irrespective of the sequence of the rest of the peptide.

The method of derivation of specific algorithm coefficients has been described in Gulukota *et al.*, *J. Mol. Biol.* 267:1258-126, 1997; (see also Sidney *et al.*, *Human Immunol.* 45:79-93, 1996; and Southwood *et al.*, *J. Immunol.* 160:3363-3373, 1998). Briefly, for all *i* positions, anchor and non-anchor alike, the geometric mean of the average relative binding (ARB) of all peptides carrying *j* is calculated relative to the remainder of the group, and used as the estimate of f_i. For Class II peptides, if multiple alignments are possible, only the highest scoring alignment is utilized, following an iterative procedure. To calculate an algorithm score of a given peptide in a test set, the ARB values corresponding to the sequence of the peptide are multiplied. If this product exceeds a chosen threshold, the peptide is predicted to bind. Appropriate thresholds are chosen as a function of the degree of stringency of prediction desired.

Selection of HLA-A2 supertype cross-reactive peptides

Complete protein sequences from 85P1B3 are scanned utilizing motif identification software, to identify 8-, 9- 10- and 11-mer sequences containing the HLA-A2-supermotif main anchor specificity. Typically, these sequences are then scored using the protocol described above and the peptides corresponding to the positive-scoring sequences are synthesized and tested for their capacity to bind purified HLA-A*0201 molecules *in vitro* (HLA-A*0201 is considered a prototype A2 supertype molecule).

These peptides are then tested for the capacity to bind to additional A2-supertype molecules (A*0202, A*0203, A*0206, and A*6802). Peptides that bind to at least three of the five A2-supertype alleles tested are typically deemed A2-supertype cross-reactive binders. Preferred peptides bind at an affinity equal to or less than 500 nM to three or more HLA-A2 supertype molecules.

Selection of HLA-A3 supermotif-bearing epitopes

The 85P1B3 protein sequence scanned above is also examined for the presence of peptides with the HLA-A3-supermotif primary anchors. Peptides corresponding to the HLA A3 supermotif-bearing sequences are then synthesized and tested for binding to HLA-A*0301 and HLA-A*1101 molecules, the molecules encoded by the two most prevalent A3-supertype alleles. The peptides that bind at least one of the two alleles with binding affinities of ≤500 nM, often ≤ 200 nM, are then tested for binding cross-reactivity to the other common A3-supertype alleles (e.g., A*3101, A*3301, and A*6801) to identify those that can bind at least three of the five HLA-A3-supertype molecules tested.

Selection of HLA-B7 supermotif bearing epitopes

The 85P1B3 protein is also analyzed for the presence of 8-, 9- 10-, or 11-mer peptides with the HLA-B7-supermotif. Corresponding peptides are synthesized and tested for binding to HLA-B*0702, the molecule encoded by the most common B7-supertype allele (*i.e.*, the prototype B7 supertype allele). Peptides binding B*0702 with IC_{50} of ≤500 nM are identified using standard methods. These peptides are then tested for binding to other common B7-supertype molecules (e.g., B*3501, B*5101, B*5301, and

B*5401). Peptides capable of binding to three or more of the five B7-supertype alleles tested are thereby identified.

Selection of A1 and A24 motif-bearing epitopes

To further increase population coverage, HLA-A1 and -A24 epitopes can also be incorporated into vaccine compositions. An analysis of the 85P1B3 protein can also be performed to identify HLA-A1- and A24-motif-containing sequences.

High affinity and/or cross-reactive binding epitopes that bear other motif and/or supermotifs are identified using analogous methodology.

Example 12: Confirmation of Immunogenicity

Cross-reactive candidate CTL A2-supermotif-bearing peptides that are identified as described herein are selected to confirm *in vitro* immunogenicity. Confirmation is performed using the following methodology:

Target Cell Lines for Cellular Screening:

The .221A2.1 cell line, produced by transferring the HLA-A2.1 gene into the HLA-A, -B, -C null mutant human B-lymphoblastoid cell line 721.221, is used as the peptide-loaded target to measure activity of HLA-A2.1-restricted CTL. This cell line is grown in RPMI-1640 medium supplemented with antibiotics, sodium pyruvate, nonessential amino acids and 10% (v/v) heat inactivated FCS. Cells that express an antigen of interest, or transfectants comprising the gene encoding the antigen of interest, can be used as target cells to confirm the ability of peptide-specific CTLs to recognize endogenous antigen.

Primary CTL Induction Cultures:

Generation of Dendritic Cells (DC): PBMCs are thawed in RPMI with 30 μg/ml DNAse, washed twice and resuspended in complete medium (RPMI-1640 plus 5% AB human serum, non-essential amino acids, sodium pyruvate, L-glutamine and penicillin/streptomycin). The monocytes are purified by plating 10×10^6 PBMC/well in a 6-well plate. After 2 hours at 37°C, the non-adherent cells are removed by gently shaking the plates and aspirating the supernatants. The wells are washed a total of three times with 3 ml RPMI to remove most of the non-adherent and loosely adherent cells. Three ml of complete medium containing 50 ng/ml of GM-CSF and 1,000 U/ml of IL-4 are then added to each well. TNFα is added to the DCs on day 6 at 75 ng/ml and the cells are used for CTL induction cultures on day 7.

Induction of CTL with DC and Peptide: CD8+ T-cells are isolated by positive selection with Dynal immunomagnetic beads (Dynabeads® M-450) and the detacha-bead® reagent. Typically about 200-250x10^6 PBMC are processed to obtain 24x10^6 $CD8^+$ T-cells (enough for a 48-well plate culture). Briefly, the PBMCs are thawed in RPMI with 30μg/ml DNAse, washed once with PBS containing 1% human AB serum and resuspended in PBS/1% AB serum at a concentration of 20x10^6cells/ml. The magnetic beads

are washed 3 times with PBS/AB serum, added to the cells (140µl beads/20x10^6 cells) and incubated for 1 hour at 4°C with continuous mixing. The beads and cells are washed 4x with PBS/AB serum to remove the nonadherent cells and resuspended at 100x10^6 cells/ml (based on the original cell number) in PBS/AB serum containing 100µl/ml detacha-bead® reagent and 30 µg/ml DNAse. The mixture is incubated for 1 hour at room temperature with continuous mixing. The beads are washed again with PBS/AB/DNAse to collect the CD8+ T-cells. The DC are collected and centrifuged at 1300 rpm for 5-7 minutes, washed once with PBS with 1% BSA, counted and pulsed with 40µg/ml of peptide at a cell concentration of 1-2x10^6/ml in the presence of 3µg/ml $ß_2$- microglobulin for 4 hours at 20°C. The DC are then irradiated (4,200 rads), washed 1 time with medium and counted again.

Setting up induction cultures: 0.25 ml cytokine-generated DC (at 1x10^5 cells/ml) are co-cultured with 0.25ml of CD8+ T-cells (at 2x10^6 cell/ml) in each well of a 48-well plate in the presence of 10 ng/ml of IL-7. Recombinant human IL-10 is added the next day at a final concentration of 10 ng/ml and rhuman IL-2 is added 48 hours later at 10 IU/ml.

Restimulation of the induction cultures with peptide-pulsed adherent cells: Seven and fourteen days after the primary induction, the cells are restimulated with peptide-pulsed adherent cells. The PBMCs are thawed and washed twice with RPMI and DNAse. The cells are resuspended at 5x10^6 cells/ml and irradiated at ~4200 rads. The PBMCs are plated at 2x10^6 in 0.5 ml complete medium per well and incubated for 2 hours at 37°C. The plates are washed twice with RPMI by tapping the plate gently to remove the nonadherent cells and the adherent cells pulsed with 10µg/ml of peptide in the presence of 3 µg/ml $ß_2$ microglobulin in 0.25ml RPMI/5%AB per well for 2 hours at 37°C. Peptide solution from each well is aspirated and the wells are washed once with RPMI. Most of the media is aspirated from the induction cultures (CD8+ cells) and brought to 0.5 ml with fresh media. The cells are then transferred to the wells containing the peptide-pulsed adherent cells. Twenty four hours later recombinant human IL-10 is added at a final concentration of 10 ng/ml and recombinant human IL2 is added the next day and again 2-3 days later at 50IU/ml (Tsai *et al., Critical Reviews in Immunology* 18(1-2):65-75, 1998). Seven days later, the cultures are assayed for CTL activity in a ^{51}Cr release assay. In some experiments the cultures are assayed for peptide-specific recognition in the *in situ* IFNγ ELISA at the time of the second restimulation followed by assay of endogenous recognition 7 days later. After expansion, activity is measured in both assays for a side-by-side comparison.

Measurement of CTL lytic activity by ^{51}Cr release.

Seven days after the second restimulation, cytotoxicity is determined in a standard (5 hr) ^{51}Cr release assay by assaying individual wells at a single E:T. Peptide-pulsed targets are prepared by incubating the cells with 10µg/ml peptide overnight at 37°C.

Adherent target cells are removed from culture flasks with trypsin-EDTA. Target cells are labelled with 200μCi of ^{51}Cr sodium chromate (Dupont, Wilmington, DE) for 1 hour at 37°C. Labelled target cells are resuspended at 10^6 per ml and diluted 1:10 with K562 cells at a concentration of $3.3x10^6$/ml (an NK-sensitive erythroblastoma cell line used to reduce non-specific lysis). Target cells (100 μl) and effectors (100μl) are plated in 96 well round-bottom plates and incubated for 5 hours at 37°C. At that time, 100 μl of supernatant are collected from each well and percent lysis is determined according to the formula: [(cpm of the test sample- cpm of the spontaneous ^{51}Cr release sample)/(cpm of the maximal ^{51}Cr release sample- cpm of the spontaneous ^{51}Cr release sample)] x 100.

Maximum and spontaneous release are determined by incubating the labelled targets with 1% Trition X-100 and media alone, respectively. A positive culture is defined as one in which the specific lysis (sample- background) is 10% or higher in the case of individual wells and is 15% or more at the two highest E:T ratios when expanded cultures are assayed.

In situ Measurement of Human IFNγ Production as an Indicator of Peptide-specific and Endogenous Recognition

Immulon 2 plates are coated with mouse anti-human IFNγ monoclonal antibody (4 μg/ml 0.1M $NaHCO_3$, pH8.2) overnight at 4°C. The plates are washed with Ca^{2+}, Mg^{2+}-free PBS/0.05% Tween 20 and blocked with PBS/10% FCS for two hours, after which the CTLs (100 μl/well) and targets (100 μl/well) are added to each well, leaving empty wells for the standards and blanks (which received media only). The target cells, either peptide-pulsed or endogenous targets, are used at a concentration of $1x10^6$ cells/ml. The plates are incubated for 48 hours at 37°C with 5% CO_2.

Recombinant human IFN-gamma is added to the standard wells starting at 400 pg or 1200pg/100 microliter/well and the plate incubated for two hours at 37°C. The plates are washed and 100 μl of biotinylated mouse anti-human IFN-gamma monoclonal antibody (2 microgram/ml in PBS/3%FCS/0.05% Tween 20) are added and incubated for 2 hours at room temperature. After washing again, 100 microliter HRP-streptavidin (1:4000) are added and the plates incubated for one hour at room temperature. The plates are then washed 6x with wash buffer, 100 microliter/well developing solution (TMB 1:1) are added, and the plates allowed to develop for 5-15 minutes. The reaction is stopped with 50 microliter/well 1M H_3PO_4 and read at OD450. A culture is considered positive if it measured at least 50 pg of IFN-gamma/well above background and is twice the background level of expression.

CTL Expansion.

Those cultures that demonstrate specific lytic activity against peptide-pulsed targets and/or tumor targets are expanded over a two week period with anti-CD3. Briefly, $5x10^4$ CD8+ cells are added to a T25 flask containing the following: $1x10^6$ irradiated (4,200 rad) PBMC (autologous or allogeneic) per ml,

$2x10^5$ irradiated (8,000 rad) EBV- transformed cells per ml, and OKT3 (anti-CD3) at 30ng per ml in RPMI-1640 containing 10% (v/v) human AB serum, non-essential amino acids, sodium pyruvate, 25μM 2-mercaptoethanol, L-glutamine and penicillin/streptomycin. Recombinant human IL2 is added 24 hours later at a final concentration of 200IU/ml and every three days thereafter with fresh media at 50IU/ml. The cells are split if the cell concentration exceeds $1x10^6$/ml and the cultures are assayed between days 13 and 15 at E:T ratios of 30, 10, 3 and 1:1 in the ^{51}Cr release assay or at $1x10^6$/ml in the *in situ* IFNγ assay using the same targets as before the expansion.

Cultures are expanded in the absence of anti-$CD3^+$ as follows. Those cultures that demonstrate specific lytic activity against peptide and endogenous targets are selected and $5x10^4$ $CD8^+$ cells are added to a T25 flask containing the following: $1x10^6$ autologous PBMC per ml which have been peptide-pulsed with 10 μg/ml peptide for two hours at 37°C and irradiated (4,200 rad); $2x10^5$ irradiated (8,000 rad) EBV-transformed cells per ml RPMI-1640 containing 10%(v/v) human AB serum, non-essential AA, sodium pyruvate, 25mM 2-ME, L-glutamine and gentamicin.

Immunogenicity of A2 supermotif-bearing peptides

A2-supermotif cross-reactive binding peptides are tested in the cellular assay for the ability to induce peptide-specific CTL in normal individuals. In this analysis, a peptide is typically considered to be an epitope if it induces peptide-specific CTLs in at least individuals, and preferably, also recognizes the endogenously expressed peptide.

Immunogenicity can also be confirmed using PBMCs isolated from patients bearing a tumor that expresses 85P1B3. Briefly, PBMCs are isolated from patients, re-stimulated with peptide-pulsed monocytes and assayed for the ability to recognize peptide-pulsed target cells as well as transfected cells endogenously expressing the antigen.

Evaluation of A*03/A11 immunogenicity

HLA-A3 supermotif-bearing cross-reactive binding peptides are also evaluated for immunogenicity using methodology analogous for that used to evaluate the immunogenicity of the HLA-A2 supermotif peptides.

Evaluation of B7 immunogenicity

Immunogenicity screening of the B7-supertype cross-reactive binding peptides identified as set forth herein are confirmed in a manner analogous to the confirmation of A2-and A3-supermotif-bearing peptides.

Peptides bearing other supermotifs/motifs, *e.g.*, HLA-A1, HLA-A24 *etc.* are also confirmed using similar methodology

Example 13: Implementation of the Extended Supermotif to Improve the Binding Capacity of Native Epitopes by Creating Analogs

HLA motifs and supermotifs (comprising primary and/or secondary residues) are useful in the identification and preparation of highly cross-reactive native peptides, as demonstrated herein. Moreover, the definition of HLA motifs and supermotifs also allows one to engineer highly cross-reactive epitopes by identifying residues within a native peptide sequence which can be analoged to confer upon the peptide certain characteristics, *e.g.* greater cross-reactivity within the group of HLA molecules that comprise a supertype, and/or greater binding affinity for some or all of those HLA molecules. Examples of analoging peptides to exhibit modulated binding affinity are set forth in this example.

Analoging at Primary Anchor Residues

Peptide engineering strategies are implemented to further increase the cross-reactivity of the epitopes. For example, the main anchors of A2-supermotif-bearing peptides are altered, for example, to introduce a preferred L, I, V, or M at position 2, and I or V at the C-terminus.

To analyze the cross-reactivity of the analog peptides, each engineered analog is initially tested for binding to the prototype A2 supertype allele A*0201, then, if A*0201 binding capacity is maintained, for A2-supertype cross-reactivity.

Alternatively, a peptide is confirmed as binding one or all supertype members and then analogued to modulate binding affinity to any one (or more) of the supertype members to add population coverage.

The selection of analogs for immunogenicity in a cellular screening analysis is typically further restricted by the capacity of the parent wild type (WT) peptide to bind at least weakly, *i.e.*, bind at an IC_{50} of 5000nM or less, to three of more A2 supertype alleles. The rationale for this requirement is that the WT peptides must be present endogenously in sufficient quantity to be biologically relevant. Analoged peptides have been shown to have increased immunogenicity and cross-reactivity by T cells specific for the parent epitope (*see, e.g.*, Parkhurst *et al.*, *J. Immunol.* 157:2539, 1996; and Pogue *et al.*, *Proc. Natl. Acad. Sci. USA* 92:8166, 1995).

In the cellular screening of these peptide analogs, it is important to confirm that analog-specific CTLs are also able to recognize the wild-type peptide and, when possible, target cells that endogenously express the epitope.

Analoging of HLA-A3 and B7-supermotif-bearing peptides

Analogs of HLA-A3 supermotif-bearing epitopes are generated using strategies similar to those employed in analoging HLA-A2 supermotif-bearing peptides. For example, peptides binding to 3/5 of the A3-supertype molecules are engineered at primary anchor residues to possess a preferred residue (V, S, M, or A) at position 2.

The analog peptides are then tested for the ability to bind A*03 and A*11 (prototype A3 supertype alleles). Those peptides that demonstrate ≤ 500 nM binding capacity are then confirmed as having A3-supertype cross-reactivity.

Similarly to the A2- and A3- motif bearing peptides, peptides binding 3 or more B7-supertype alleles can be improved, where possible, to achieve increased cross-reactive binding or greater binding affinity or binding half life. B7 supermotif-bearing peptides are, for example, engineered to possess a preferred residue (V, I, L, or F) at the C-terminal primary anchor position, as demonstrated by Sidney *et al.* (*J. Immunol.* 157:3480-3490, 1996).

Analoging at primary anchor residues of other motif and/or supermotif-bearing epitopes is performed in a like manner.

The analog peptides are then be confirmed for immunogenicity, typically in a cellular screening assay. Again, it is generally important to demonstrate that analog-specific CTLs are also able to recognize the wild-type peptide and, when possible, targets that endogenously express the epitope.

Analoging at Secondary Anchor Residues

Moreover, HLA supermotifs are of value in engineering highly cross-reactive peptides and/or peptides that bind HLA molecules with increased affinity by identifying particular residues at secondary anchor positions that are associated with such properties. For example, the binding capacity of a B7 supermotif-bearing peptide with an F residue at position 1 is analyzed. The peptide is then analoged to, for example, substitute L for F at position 1. The analoged peptide is evaluated for increased binding affinity, binding half life and/or increased cross-reactivity. Such a procedure identifies analoged peptides with enhanced properties.

Engineered analogs with sufficiently improved binding capacity or cross-reactivity can also be tested for immunogenicity in HLA-B7-transgenic mice, following for example, IFA immunization or lipopeptide immunization. Analogued peptides are additionally tested for the ability to stimulate a recall response using PBMC from patients with 85P1B3-expressing tumors.

Other analoguing strategies

Another form of peptide analoguing, unrelated to anchor positions, involves the substitution of a cysteine with α-amino butyric acid. Due to its chemical nature, cysteine has the propensity to form disulfide bridges and sufficiently alter the peptide structurally so as to reduce binding capacity. Substitution of α-amino butyric acid for cysteine not only alleviates this problem, but has been shown to improve binding and crossbinding capabilities in some instances (*see, e.g.*, the review by Sette *et al.*, In: Persistent Viral Infections, Eds. R. Ahmed and I. Chen, John Wiley & Sons, England, 1999).

Thus, by the use of single amino acid substitutions, the binding properties and/or cross-reactivity of peptide ligands for HLA supertype molecules can be modulated.

Example 14. Identification and confirmation of 85P1B3-derived sequences with HLA-DR binding motifs

Peptide epitopes bearing an HLA class II supermotif or motif are identified and confirmed as outlined below using methodology similar to that described for HLA Class I peptides.

Selection of HLA-DR-supermotif-bearing epitopes.

To identify 85P1B3-derived, HLA class II HTL epitopes, the 85P1B3 antigen is analyzed for the presence of sequences bearing an HLA-DR-motif or supermotif. Specifically, 15-mer sequences are selected comprising a DR-supermotif, comprising a 9-mer core, and three-residue N- and C-terminal flanking regions (15 amino acids total).

Protocols for predicting peptide binding to DR molecules have been developed (Southwood *et al.*, *J. Immunol.* 160:3363-3373, 1998). These protocols, specific for individual DR molecules, allow the scoring, and ranking, of 9-mer core regions. Each protocol not only scores peptide sequences for the presence of DR-supermotif primary anchors (i.e., at position 1 and position 6) within a 9-mer core, but additionally evaluates sequences for the presence of secondary anchors. Using allele-specific selection tables (see, *e.g.*, Southwood *et al.*, *ibid.*), it has been found that these protocols efficiently select peptide sequences with a high probability of binding a particular DR molecule. Additionally, it has been found that performing these protocols in tandem, specifically those for DR1, DR4w4, and DR7, can efficiently select DR cross-reactive peptides.

The 85P1B3-derived peptides identified above are tested for their binding capacity for various common HLA-DR molecules. All peptides are initially tested for binding to the DR molecules in the primary panel: DR1, DR4w4, and DR7. Peptides binding at least two of these three DR molecules are then tested for binding to DR2w2 β1, DR2w2 β2, DR6w19, and DR9 molecules in secondary assays. Finally, peptides binding at least two of the four secondary panel DR molecules, and thus cumulatively at least four of seven different DR molecules, are screened for binding to DR4w15, DR5w11, and DR8w2 molecules in tertiary assays. Peptides binding at least seven of the ten DR molecules comprising the primary, secondary, and tertiary screening assays are considered cross-reactive DR binders. 85P1B3-derived peptides found to bind common HLA-DR alleles are of particular interest.

Selection of DR3 motif peptides

Because HLA-DR3 is an allele that is prevalent in Caucasian, Black, and Hispanic populations, DR3 binding capacity is a relevant criterion in the selection of HTL epitopes. Thus, peptides shown to be candidates may also be assayed for their DR3 binding capacity. However, in view of the binding

specificity of the DR3 motif, peptides binding only to DR3 can also be considered as candidates for inclusion in a vaccine formulation.

To efficiently identify peptides that bind DR3, target 85P1B3 antigens are analyzed for sequences carrying one of the two DR3-specific binding motifs reported by Geluk *et al.* (*J. Immunol.* 152:5742-5748, 1994). The corresponding peptides are then synthesized and confirmed as having the ability to bind DR3 with an affinity of 1μM or better, i.e., less than 1 μM. Peptides are found that meet this binding criterion and qualify as HLA class II high affinity binders.

DR3 binding epitopes identified in this manner are included in vaccine compositions with DR supermotif-bearing peptide epitopes.

Similarly to the case of HLA class I motif-bearing peptides, the class II motif-bearing peptides are analoged to improve affinity or cross-reactivity. For example, aspartic acid at position 4 of the 9-mer core sequence is an optimal residue for DR3 binding, and substitution for that residue often improves DR 3 binding.

Example 15: Immunogenicity of 85P1B3-derived HTL epitopes

This example determines immunogenic DR supermotif- and DR3 motif-bearing epitopes among those identified using the methodology set forth herein.

Immunogenicity of HTL epitopes are confirmed in a manner analogous to the determination of immunogenicity of CTL epitopes, by assessing the ability to stimulate HTL responses and/or by using appropriate transgenic mouse models. Immunogenicity is determined by screening for: 1.) *in vitro* primary induction using normal PBMC or 2.) recall responses from patients who have 85P1B3-expressing tumors.

Example 16: Calculation of phenotypic frequencies of HLA-supertypes in various ethnic backgrounds to determine breadth of population coverage

This example illustrates the assessment of the breadth of population coverage of a vaccine composition comprised of multiple epitopes comprising multiple supermotifs and/or motifs.

In order to analyze population coverage, gene frequencies of HLA alleles are determined. Gene frequencies for each HLA allele are calculated from antigen or allele frequencies utilizing the binomial distribution formulae gf=1-(SQRT(1-af)) (see, *e.g.*, Sidney *et al.*, *Human Immunol.* 45:79-93, 1996). To obtain overall phenotypic frequencies, cumulative gene frequencies are calculated, and the cumulative antigen frequencies derived by the use of the inverse formula $[af=1-(1-Cgf)^2]$.

Where frequency data is not available at the level of DNA typing, correspondence to the serologically defined antigen frequencies is assumed. To obtain total potential supertype population coverage no linkage disequilibrium is assumed, and only alleles confirmed to belong to each of the

supertypes are included (minimal estimates). Estimates of total potential coverage achieved by inter-loci combinations are made by adding to the A coverage the proportion of the non-A covered population that could be expected to be covered by the B alleles considered (e.g., total=A+B*(1-A)). Confirmed members of the A3-like supertype are A3, A11, A31, A*3301, and A*6801. Although the A3-like supertype may also include A34, A66, and A*7401, these alleles were not included in overall frequency calculations. Likewise, confirmed members of the A2-like supertype family are A*0201, A*0202, A*0203, A*0204, A*0205, A*0206, A*0207, A*6802, and A*6901. Finally, the B7-like supertype-confirmed alleles are: B7, B*3501-03, B51, B*5301, B*5401, B*5501-2, B*5601, B*6701, and B*7801 (potentially also B*1401, B*3504-06, B*4201, and B*5602).

Population coverage achieved by combining the A2-, A3- and B7-supertypes is approximately 86% in five major ethnic groups. Coverage may be extended by including peptides bearing the A1 and A24 motifs. On average, A1 is present in 12% and A24 in 29% of the population across five different major ethnic groups (Caucasian, North American Black, Chinese, Japanese, and Hispanic). Together, these alleles are represented with an average frequency of 39% in these same ethnic populations. The total coverage across the major ethnicities when A1 and A24 are combined with the coverage of the A2-, A3- and B7-supertype alleles is >95%. An analagous approach can be used to estimate population coverage achieved with combinations of class II motif-bearing epitopes.

Immunogenicity studies in humans (*e.g.*, Bertoni *et al.*, *J. Clin. Invest.* 100:503, 1997; Doolan *et al.*, *Immunity* 7:97, 1997; and Threlkeld *et al.*, *J. Immunol.* 159:1648, 1997) have shown that highly cross-reactive binding peptides are almost always recognized as epitopes. The use of highly cross-reactive binding peptides is an important selection criterion in identifying candidate epitopes for inclusion in a vaccine that is immunogenic in a diverse population.

With a sufficient number of epitopes (as disclosed herein and from the art), an average population coverage is predicted to be greater than 95% in each of five major ethnic populations. The game theory Monte Carlo simulation analysis, which is known in the art (see *e.g.*, Osborne, M.J. and Rubinstein, A. "A course in game theory" MIT Press, 1994), can be used to estimate what percentage of the individuals in a population comprised of the Caucasian, North American Black, Japanese, Chinese, and Hispanic ethnic groups would recognize the vaccine epitopes described herein. A preferred percentage is 90%. A more preferred percentage is 95%.

Example 17: CTL Recognition Of Endogenously Processed Antigens After Priming

This example confirms that CTL induced by native or analoged peptide epitopes identified and selected as described herein recognize endogenously synthesized, *i.e.*, native antigens.

Effector cells isolated from transgenic mice that are immunized with peptide epitopes, for example HLA-A2 supermotif-bearing epitopes, are re-stimulated *in vitro* using peptide-coated stimulator cells. Six days later, effector cells are assayed for cytotoxicity and the cell lines that contain peptide-specific cytotoxic activity are further re-stimulated. An additional six days later, these cell lines are tested for cytotoxic activity on ^{51}Cr labeled Jurkat-A2.1/K^b target cells in the absence or presence of peptide, and also tested on ^{51}Cr labeled target cells bearing the endogenously synthesized antigen, *i.e.* cells that are stably transfected with 85P1B3 expression vectors.

The results demonstrate that CTL lines obtained from animals primed with peptide epitope recognize endogenously synthesized 85P1B3 antigen. The choice of transgenic mouse model to be used for such an analysis depends upon the epitope(s) that are being evaluated. In addition to HLA-A*0201/K^b transgenic mice, several other transgenic mouse models including mice with human A11, which may also be used to evaluate A3 epitopes, and B7 alleles have been characterized and others (*e.g.*, transgenic mice for HLA-A1 and A24) are being developed. HLA-DR1 and HLA-DR3 mouse models have also been developed, which may be used to evaluate HTL epitopes.

Example 18: Activity Of CTL-HTL Conjugated Epitopes In Transgenic Mice

This example illustrates the induction of CTLs and HTLs in transgenic mice, by use of a 85P1B3-derived CTL and HTL peptide vaccine compositions. The vaccine composition used herein comprise peptides to be administered to a patient with a 85P1B3-expressing tumor. The peptide composition can comprise multiple CTL and/or HTL epitopes. The epitopes are identified using methodology as described herein. This example also illustrates that enhanced immunogenicity can be achieved by inclusion of one or more HTL epitopes in a CTL vaccine composition; such a peptide composition can comprise an HTL epitope conjugated to a CTL epitope. The CTL epitope can be one that binds to multiple HLA family members at an affinity of 500 nM or less, or analogs of that epitope. The peptides may be lipidated, if desired.

Immunization procedures: Immunization of transgenic mice is performed as described (Alexander *et al., J. Immunol.* 159:4753-4761, 1997). For example, A2/K^b mice, which are transgenic for the human HLA A2.1 allele and are used to confirm the immunogenicity of HLA-A*0201 motif- or HLA-A2 supermotif-bearing epitopes, and are primed subcutaneously (base of the tail) with a 0.1 ml of peptide in Incomplete Freund's Adjuvant, or if the peptide composition is a lipidated CTL/HTL conjugate, in DMSO/saline, or if the peptide composition is a polypeptide, in PBS or Incomplete Freund's Adjuvant. Seven days after priming, splenocytes obtained from these animals are restimulated with syngenic irradiated LPS-activated lymphoblasts coated with peptide.

Cell lines: Target cells for peptide-specific cytotoxicity assays are Jurkat cells transfected with the HLA-A2.1/K^b chimeric gene (*e.g.*, Vitiello *et al., J. Exp. Med.* 173:1007, 1991)

In vitro CTL activation: One week after priming, spleen cells ($30x10^6$ cells/flask) are co-cultured at 37°C with syngeneic, irradiated (3000 rads), peptide coated lymphoblasts ($10x10^6$ cells/flask) in 10 ml of culture medium/T25 flask. After six days, effector cells are harvested and assayed for cytotoxic activity.

Assay for cytotoxic activity: Target cells (1.0 to $1.5x10^6$) are incubated at 37°C in the presence of 200 μl of ^{51}Cr. After 60 minutes, cells are washed three times and resuspended in R10 medium. Peptide is added where required at a concentration of 1 μg/ml. For the assay, 10^4 ^{51}Cr-labeled target cells are added to different concentrations of effector cells (final volume of 200 μl) in U-bottom 96-well plates. After a six hour incubation period at 37°C, a 0.1 ml aliquot of supernatant is removed from each well and radioactivity is determined in a Micromedic automatic gamma counter. The percent specific lysis is determined by the formula: percent specific release = 100 x (experimental release - spontaneous release)/(maximum release - spontaneous release). To facilitate comparison between separate CTL assays run under the same conditions, % ^{51}Cr release data is expressed as lytic units/10^6 cells. One lytic unit is arbitrarily defined as the number of effector cells required to achieve 30% lysis of 10,000 target cells in a six hour ^{51}Cr release assay. To obtain specific lytic units/10^6, the lytic units/10^6 obtained in the absence of peptide is subtracted from the lytic units/10^6 obtained in the presence of peptide. For example, if 30% ^{51}Cr release is obtained at the effector (E): target (T) ratio of 50:1 (i.e., $5x10^5$ effector cells for 10,000 targets) in the absence of peptide and 5:1 (i.e., $5x10^4$ effector cells for 10,000 targets) in the presence of peptide, the specific lytic units would be: $[(1/50,000)-(1/500,000)] \times 10^6 = 18$ LU.

The results are analyzed to assess the magnitude of the CTL responses of animals injected with the immunogenic CTL/HTL conjugate vaccine preparation and are compared to the magnitude of the CTL response achieved using, for example, CTL epitopes as outlined above in the Example entitled "Confirmation of Immunogenicity". Analyses similar to this may be performed to confirm the immunogenicity of peptide conjugates containing multiple CTL epitopes and/or multiple HTL epitopes. In accordance with these procedures, it is found that a CTL response is induced, and concomitantly that an HTL response is induced upon administration of such compositions.

Example 19: Selection of CTL and HTL epitopes for inclusion in an 85P1B3-specific vaccine.

This example illustrates a procedure for selecting peptide epitopes for vaccine compositions of the invention. The peptides in the composition can be in the form of a nucleic acid sequence, either single or one or more sequences (*i.e.*, minigene) that encodes peptide(s), or can be single and/or polyepitopic peptides.

The following principles are utilized when selecting a plurality of epitopes for inclusion in a vaccine composition. Each of the following principles is balanced in order to make the selection.

Epitopes are selected which, upon administration, mimic immune responses that are correlated with 85P1B3 clearance. The number of epitopes used depends on observations of patients who spontaneously clear 85P1B3. For example, if it has been observed that patients who spontaneously clear 85P1B3 generate an immune response to at least three (3) from 85P1B3 antigen, then three or four (3-4) epitopes should be included for HLA class I. A similar rationale is used to determine HLA class II epitopes.

Epitopes are often selected that have a binding affinity of an IC_{50} of 500 nM or less for an HLA class I molecule, or for class II, an IC_{50} of 1000 nM or less; or HLA Class I peptides with high binding scores from the BIMAS web site, at URL bimas.dcrt.nih.gov/.

In order to achieve broad coverage of the vaccine through out a diverse population, sufficient supermotif bearing peptides, or a sufficient array of allele-specific motif bearing peptides, are selected to give broad population coverage. In one embodiment, epitopes are selected to provide at least 80% population coverage. A Monte Carlo analysis, a statistical evaluation known in the art, can be employed to assess breadth, or redundancy, of population coverage.

When creating polyepitopic compositions, or a minigene that encodes same, it is typically desirable to generate the smallest peptide possible that encompasses the epitopes of interest. The principles employed are similar, if not the same, as those employed when selecting a peptide comprising nested epitopes. For example, a protein sequence for the vaccine composition is selected because it has maximal number of epitopes contained within the sequence, *i.e.*, it has a high concentration of epitopes. Epitopes may be nested or overlapping (*i.e.*, frame shifted relative to one another). For example, with overlapping epitopes, two 9-mer epitopes and one 10-mer epitope can be present in a 10 amino acid peptide. Each epitope can be exposed and bound by an HLA molecule upon administration of such a peptide. A multi-epitopic, peptide can be generated synthetically, recombinantly, or via cleavage from the native source. Alternatively, an analog can be made of this native sequence, whereby one or more of the epitopes comprise substitutions that alter the cross-reactivity and/or binding affinity properties of the polyepitopic peptide. Such a vaccine composition is administered for therapeutic or prophylactic purposes. This embodiment provides for the possibility that an as yet undiscovered aspect of immune system processing will apply to the native nested sequence and thereby facilitate the production of therapeutic or prophylactic immune response-inducing vaccine compositions. Additionally such an embodiment provides for the possibility of motif-bearing epitopes for an HLA makeup that is presently unknown. Furthermore, this embodiment (absent the creating of any analogs) directs the immune response to multiple peptide sequences that are actually present in 85P1B3, thus avoiding the need to evaluate any junctional epitopes.

Lastly, the embodiment provides an economy of scale when producing nucleic acid vaccine compositions. Related to this embodiment, computer programs can be derived in accordance with principles in the art, which identify in a target sequence, the greatest number of epitopes per sequence length.

A vaccine composition comprised of selected peptides, when administered, is safe, efficacious, and elicits an immune response similar in magnitude to an immune response that controls or clears cells that bear or overexpress 85P1B3.

Example 20: Construction of "Minigene" Multi-Epitope DNA Plasmids

This example discusses the construction of a minigene expression plasmid. Minigene plasmids may, of course, contain various configurations of B cell, CTL and/or HTL epitopes or epitope analogs as described herein.

A minigene expression plasmid typically includes multiple CTL and HTL peptide epitopes. In the present example, HLA-A2, -A3, -B7 supermotif-bearing peptide epitopes and HLA-A1 and -A24 motif-bearing peptide epitopes are used in conjunction with DR supermotif-bearing epitopes and/or DR3 epitopes. HLA class I supermotif or motif-bearing peptide epitopes derived 85P1B3, are selected such that multiple supermotifs/motifs are represented to ensure broad population coverage. Similarly, HLA class II epitopes are selected from 85P1B3 to provide broad population coverage, *i.e.* both HLA DR-1-4-7 supermotif-bearing epitopes and HLA DR-3 motif-bearing epitopes are selected for inclusion in the minigene construct. The selected CTL and HTL epitopes are then incorporated into a minigene for expression in an expression vector.

Such a construct may additionally include sequences that direct the HTL epitopes to the endoplasmic reticulum. For example, the Ii protein may be fused to one or more HTL epitopes as described in the art, wherein the CLIP sequence of the Ii protein is removed and replaced with an HLA class II epitope sequence so that HLA class II epitope is directed to the endoplasmic reticulum, where the epitope binds to an HLA class II molecules.

This example illustrates the methods to be used for construction of a minigene-bearing expression plasmid. Other expression vectors that may be used for minigene compositions are available and known to those of skill in the art.

The minigene DNA plasmid of this example contains a consensus Kozak sequence and a consensus murine kappa Ig-light chain signal sequence followed by CTL and/or HTL epitopes selected in accordance with principles disclosed herein. The sequence encodes an open reading frame fused to the Myc and His antibody epitope tag coded for by the pcDNA 3.1 Myc-His vector.

Overlapping oligonucleotides that can, for example, average about 70 nucleotides in length with 15 nucleotide overlaps, are synthesized and HPLC-purified. The oligonucleotides encode the selected

peptide epitopes as well as appropriate linker nucleotides, Kozak sequence, and signal sequence. The final multiepitope minigene is assembled by extending the overlapping oligonucleotides in three sets of reactions using PCR. A Perkin/Elmer 9600 PCR machine is used and a total of 30 cycles are performed using the following conditions: 95°C for 15 sec, annealing temperature (5° below the lowest calculated Tm of each primer pair) for 30 sec, and 72°C for 1 min.

For example, a minigene is prepared as follows. For a first PCR reaction, 5 μg of each of two oligonucleotides are annealed and extended: In an example using eight oligonucleotides, i.e., four pairs of primers, oligonucleotides 1+2, 3+4, 5+6, and 7+8 are combined in 100 μl reactions containing *Pfu* polymerase buffer (1x= 10 mM KCL, 10 mM $(NH4)_2SO_4$, 20 mM Tris-chloride, pH 8.75, 2 mM $MgSO_4$, 0.1% Triton X-100, 100 μg/ml BSA), 0.25 mM each dNTP, and 2.5 U of *Pfu* polymerase. The full-length dimer products are gel-purified, and two reactions containing the product of 1+2 and 3+4, and the product of 5+6 and 7+8 are mixed, annealed, and extended for 10 cycles. Half of the two reactions are then mixed, and 5 cycles of annealing and extension carried out before flanking primers are added to amplify the full length product. The full-length product is gel-purified and cloned into pCR-blunt (Invitrogen) and individual clones are screened by sequencing.

Example 21: The Plasmid Construct and the Degree to Which It Induces Immunogenicity.

The degree to which a plasmid construct, for example a plasmid constructed in accordance with the previous Example, is able to induce immunogenicity is confirmed *in vitro* by determining epitope presentation by APC following transduction or transfection of the APC with an epitope-expressing nucleic acid construct. Such a study determines "antigenicity" and allows the use of human APC. The assay determines the ability of the epitope to be presented by the APC in a context that is recognized by a T cell by quantifying the density of epitope-HLA class I complexes on the cell surface. Quantitation can be performed by directly measuring the amount of peptide eluted from the APC (*see, e.g.*, Sijts *et al.*, *J. Immunol.* 156:683-692, 1996; Demotz *et al.*, *Nature* 342:682-684, 1989); or the number of peptide-HLA class I complexes can be estimated by measuring the amount of lysis or lymphokine release induced by diseased or transfected target cells, and then determining the concentration of peptide necessary to obtain equivalent levels of lysis or lymphokine release (*see, e.g.*, Kageyama *et al.*, *J. Immunol.* 154:567-576, 1995).

Alternatively, immunogenicity is confirmed through *in vivo* injections into mice and subsequent *in vitro* assessment of CTL and HTL activity, which are analyzed using cytotoxicity and proliferation assays, respectively, as detailed *e.g.,* in Alexander *et al.*, *Immunity* 1:751-761, 1994.

For example, to confirm the capacity of a DNA minigene construct containing at least one HLA-A2 supermotif peptide to induce CTLs *in vivo*, HLA-A2.1/K^b transgenic mice, for example, are immunized intramuscularly with 100 μg of naked cDNA. As a means of comparing the level of CTLs induced by cDNA immunization, a control group of animals is also immunized with an actual peptide composition that comprises multiple epitopes synthesized as a single polypeptide as they would be encoded by the minigene.

Splenocytes from immunized animals are stimulated twice with each of the respective compositions (peptide epitopes encoded in the minigene or the polyepitopic peptide), then assayed for peptide-specific cytotoxic activity in a ^{51}Cr release assay. The results indicate the magnitude of the CTL response directed against the A2-restricted epitope, thus indicating the *in vivo* immunogenicity of the minigene vaccine and polyepitopic vaccine.

It is, therefore, found that the minigene elicits immune responses directed toward the HLA-A2 supermotif peptide epitopes as does the polyepitopic peptide vaccine. A similar analysis is also performed using other HLA-A3 and HLA-B7 transgenic mouse models to assess CTL induction by HLA-A3 and HLA-B7 motif or supermotif epitopes, whereby it is also found that the minigene elicits appropriate immune responses directed toward the provided epitopes.

To confirm the capacity of a class II epitope-encoding minigene to induce HTLs *in vivo*, DR transgenic mice, or for those epitopes that cross react with the appropriate mouse MHC molecule, I-A^b-restricted mice, for example, are immunized intramuscularly with 100 μg of plasmid DNA. As a means of comparing the level of HTLs induced by DNA immunization, a group of control animals is also immunized with an actual peptide composition emulsified in complete Freund's adjuvant. CD4+ T cells, *i.e.* HTLs, are purified from splenocytes of immunized animals and stimulated with each of the respective compositions (peptides encoded in the minigene). The HTL response is measured using a ^3H-thymidine incorporation proliferation assay, (*see, e.g.*, Alexander et al. Immunity 1:751-761, 1994). The results indicate the magnitude of the HTL response, thus demonstrating the *in vivo* immunogenicity of the minigene.

DNA minigenes, constructed as described in the previous Example, can also be confirmed as a vaccine in combination with a boosting agent using a prime boost protocol. The boosting agent can consist of recombinant protein (*e.g.*, Barnett *et al.*, *Aids Res. and Human Retroviruses* 14, Supplement 3:S299-S309, 1998) or recombinant vaccinia, for example, expressing a minigene or DNA encoding the complete protein of interest (*see, e.g.*, Hanke *et al.*, *Vaccine* 16:439-445, 1998; Sedegah *et al.*, *Proc. Natl. Acad. Sci USA* 95:7648-53, 1998; Hanke and McMichael, *Immunol. Letters* 66:177-181, 1999; and Robinson *et al.*, *Nature Med.* 5:526-34, 1999).

For example, the efficacy of the DNA minigene used in a prime boost protocol is initially evaluated in transgenic mice. In this example, A2.1/K^b transgenic mice are immunized IM with 100 μg of

a DNA minigene encoding the immunogenic peptides including at least one HLA-A2 supermotif-bearing peptide. After an incubation period (ranging from 3-9 weeks), the mice are boosted IP with 10^7 pfu/mouse of a recombinant vaccinia virus expressing the same sequence encoded by the DNA minigene. Control mice are immunized with 100 μg of DNA or recombinant vaccinia without the minigene sequence, or with DNA encoding the minigene, but without the vaccinia boost. After an additional incubation period of two weeks, splenocytes from the mice are immediately assayed for peptide-specific activity in an ELISPOT assay. Additionally, splenocytes are stimulated *in vitro* with the A2-restricted peptide epitopes encoded in the minigene and recombinant vaccinia, then assayed for peptide-specific activity in an alpha, beta and/or gamma IFN ELISA.

It is found that the minigene utilized in a prime-boost protocol elicits greater immune responses toward the HLA-A2 supermotif peptides than with DNA alone. Such an analysis can also be performed using HLA-A11 or HLA-B7 transgenic mouse models to assess CTL induction by HLA-A3 or HLA-B7 motif or supermotif epitopes. The use of prime boost protocols in humans is described below in the Example entitled "Induction of CTL Responses Using a Prime Boost Protocol ."

Example 22: Peptide Composition for Prophylactic Uses

Vaccine compositions of the present invention can be used to prevent 85P1B3 expression in persons who are at risk for tumors that bear this antigen. For example, a polyepitopic peptide epitope composition (or a nucleic acid comprising the same) containing multiple CTL and HTL epitopes such as those selected in the above Examples, which are also selected to target greater than 80% of the population, is administered to individuals at risk for a 85P1B3-associated tumor.

For example, a peptide-based composition is provided as a single polypeptide that encompasses multiple epitopes. The vaccine is typically administered in a physiological solution that comprises an adjuvant, such as Incomplete Freunds Adjuvant. The dose of peptide for the initial immunization is from about 1 to about 50,000 μg, generally 100-5,000 μg, for a 70 kg patient. The initial administration of vaccine is followed by booster dosages at 4 weeks followed by evaluation of the magnitude of the immune response in the patient, by techniques that determine the presence of epitope-specific CTL populations in a PBMC sample. Additional booster doses are administered as required. The composition is found to be both safe and efficacious as a prophylaxis against 85P1B3-associated disease.

Alternatively, a composition typically comprising transfecting agents is used for the administration of a nucleic acid-based vaccine in accordance with methodologies known in the art and disclosed herein.

Example 23: Polyepitopic Vaccine Compositions Derived from Native 85P1B3 Sequences

A native 85P1B3 polyprotein sequence is analyzed, preferably using computer algorithms defined for each class I and/or class II supermotif or motif, to identify "relatively short" regions of the polyprotein that comprise multiple epitopes. The "relatively short" regions are preferably less in length than an entire native antigen. This relatively short sequence that contains multiple distinct or overlapping, "nested" epitopes is selected; it can be used to generate a minigene construct. The construct is engineered to express the peptide, which corresponds to the native protein sequence. The "relatively short" peptide is generally less than 250 amino acids in length, often less than 100 amino acids in length, preferably less than 75 amino acids in length, and more preferably less than 50 amino acids in length. The protein sequence of the vaccine composition is selected because it has maximal number of epitopes contained within the sequence, *i.e.*, it has a high concentration of epitopes. As noted herein, epitope motifs may be nested or overlapping (*i.e.*, frame shifted relative to one another). For example, with overlapping epitopes, two 9-mer epitopes and one 10-mer epitope can be present in a 10 amino acid peptide. Such a vaccine composition is administered for therapeutic or prophylactic purposes.

The vaccine composition will include, for example, multiple CTL epitopes from 85P1B3 antigen and at least one HTL epitope. This polyepitopic native sequence is administered either as a peptide or as a nucleic acid sequence which encodes the peptide. Alternatively, an analog can be made of this native sequence, whereby one or more of the epitopes comprise substitutions that alter the cross-reactivity and/or binding affinity properties of the polyepitopic peptide.

The embodiment of this example provides for the possibility that an as yet undiscovered aspect of immune system processing will apply to the native nested sequence and thereby facilitate the production of therapeutic or prophylactic immune response-inducing vaccine compositions. Additionally such an embodiment provides for the possibility of motif-bearing epitopes for an HLA makeup that is presently unknown. Furthermore, this embodiment (excluding an analoged embodiment) directs the immune response to multiple peptide sequences that are actually present in native 85P1B3, thus avoiding the need to evaluate any junctional epitopes. Lastly, the embodiment provides an economy of scale when producing peptide or nucleic acid vaccine compositions.

Related to this embodiment, computer programs are available in the art which can be used to identify in a target sequence, the greatest number of epitopes per sequence length.

Example 24: Polyepitopic Vaccine Compositions From Multiple Antigens

The 85P1B3 peptide epitopes of the present invention are used in conjunction with epitopes from other target tumor-associated antigens, to create a vaccine composition that is useful for the prevention or treatment of cancer that expresses 85P1B3 and such other antigens. For example, a vaccine composition can be provided as a single polypeptide that incorporates multiple epitopes from 85P1B3 as well as tumor-

associated antigens that are often expressed with a target cancer associated with 85P1B3 expression, or can be administered as a composition comprising a cocktail of one or more discrete epitopes. Alternatively, the vaccine can be administered as a minigene construct or as dendritic cells which have been loaded with the peptide epitopes *in vitro*.

Example 25: Use of peptides to evaluate an immune response

Peptides of the invention may be used to analyze an immune response for the presence of specific antibodies, CTL or HTL directed to 85P1B3. Such an analysis can be performed in a manner described by Ogg *et al., Science* 279:2103-2106, 1998. In this Example, peptides in accordance with the invention are used as a reagent for diagnostic or prognostic purposes, not as an immunogen.

In this example highly sensitive human leukocyte antigen tetrameric complexes ("tetramers") are used for a cross-sectional analysis of, for example, 85P1B3 HLA-A*0201-specific CTL frequencies from HLA A*0201-positive individuals at different stages of disease or following immunization comprising an 85P1B3 peptide containing an A*0201 motif. Tetrameric complexes are synthesized as described (Musey *et al., N. Engl. J. Med.* 337:1267, 1997). Briefly, purified HLA heavy chain (A*0201 in this example) and β2-microglobulin are synthesized by means of a prokaryotic expression system. The heavy chain is modified by deletion of the transmembrane-cytosolic tail and COOH-terminal addition of a sequence containing a BirA enzymatic biotinylation site. The heavy chain, β2-microglobulin, and peptide are refolded by dilution. The 45-kD refolded product is isolated by fast protein liquid chromatography and then biotinylated by BirA in the presence of biotin (Sigma, St. Louis, Missouri), adenosine 5' triphosphate and magnesium. Streptavidin-phycoerythrin conjugate is added in a 1:4 molar ratio, and the tetrameric product is concentrated to 1 mg/ml. The resulting product is referred to as tetramer-phycoerythrin.

For the analysis of patient blood samples, approximately one million PBMCs are centrifuged at 300g for 5 minutes and resuspended in 50 µl of cold phosphate-buffered saline. Tri-color analysis is performed with the tetramer-phycoerythrin, along with anti-CD8-Tricolor, and anti-CD38. The PBMCs are incubated with tetramer and antibodies on ice for 30 to 60 min and then washed twice before formaldehyde fixation. Gates are applied to contain >99.98% of control samples. Controls for the tetramers include both A*0201-negative individuals and A*0201-positive non-diseased donors. The percentage of cells stained with the tetramer is then determined by flow cytometry. The results indicate the number of cells in the PBMC sample that contain epitope-restricted CTLs, thereby readily indicating the extent of immune response to the 85P1B3 epitope, and thus the status of exposure to 85P1B3, or exposure to a vaccine that elicits a protective or therapeutic response.

Example 26: Use of Peptide Epitopes to Evaluate Recall Responses

The peptide epitopes of the invention are used as reagents to evaluate T cell responses, such as acute or recall responses, in patients. Such an analysis may be performed on patients who have recovered from 85P1B3-associated disease or who have been vaccinated with an 85P1B3 vaccine.

For example, the class I restricted CTL response of persons who have been vaccinated may be analyzed. The vaccine may be any 85P1B3 vaccine. PBMC are collected from vaccinated individuals and HLA typed. Appropriate peptide epitopes of the invention that, optimally, bear supermotifs to provide cross-reactivity with multiple HLA supertype family members, are then used for analysis of samples derived from individuals who bear that HLA type.

PBMC from vaccinated individuals are separated on Ficoll-Histopaque density gradients (Sigma Chemical Co., St. Louis, MO), washed three times in HBSS (GIBCO Laboratories), resuspended in RPMI-1640 (GIBCO Laboratories) supplemented with L-glutamine (2mM), penicillin (50U/ml), streptomycin (50 µg/ml), and Hepes (10mM) containing 10% heat-inactivated human AB serum (complete RPMI) and plated using microculture formats. A synthetic peptide comprising an epitope of the invention is added at 10 µg/ml to each well and HBV core 128-140 epitope is added at 1 µg/ml to each well as a source of T cell help during the first week of stimulation.

In the microculture format, 4 x 10^5 PBMC are stimulated with peptide in 8 replicate cultures in 96-well round bottom plate in 100 µl/well of complete RPMI. On days 3 and 10, 100 ul of complete RPMI and 20 U/ml final concentration of rIL-2 are added to each well. On day 7 the cultures are transferred into a 96-well flat-bottom plate and restimulated with peptide, rIL-2 and 10^5 irradiated (3,000 rad) autologous feeder cells. The cultures are tested for cytotoxic activity on day 14. A positive CTL response requires two or more of the eight replicate cultures to display greater than 10% specific ^{51}Cr release, based on comparison with non-diseased control subjects as previously described (Rehermann, *et al., Nature Med.* 2:1104,1108, 1996; Rehermann *et al., J. Clin. Invest.* 97:1655-1665, 1996; and Rehermann *et al. J. Clin. Invest.* 98:1432-1440, 1996).

Target cell lines are autologous and allogeneic EBV-transformed B-LCL that are either purchased from the American Society for Histocompatibility and Immunogenetics (ASHI, Boston, MA) or established from the pool of patients as described (Guilhot, *et al. J. Virol.* 66:2670-2678, 1992).

Cytotoxicity assays are performed in the following manner. Target cells consist of either allogeneic HLA-matched or autologous EBV-transformed B lymphoblastoid cell line that are incubated overnight with the synthetic peptide epitope of the invention at 10 µM, and labeled with 100 µCi of ^{51}Cr (Amersham Corp., Arlington Heights, IL) for 1 hour after which they are washed four times with HBSS.

Cytolytic activity is determined in a standard 4-h, split well ^{51}Cr release assay using U-bottomed 96 well plates containing 3,000 targets/well. Stimulated PBMC are tested at effector/target (E/T) ratios of 20-50:1 on day 14. Percent cytotoxicity is determined from the formula: 100 x [(experimental release-spontaneous release)/maximum release-spontaneous release)]. Maximum release is determined by lysis of targets by detergent (2% Triton X-100; Sigma Chemical Co., St. Louis, MO). Spontaneous release is <25% of maximum release for all experiments.

The results of such an analysis indicate the extent to which HLA-restricted CTL populations have been stimulated by previous exposure to 85P1B3 or an 85P1B3 vaccine.

Similarly, Class II restricted HTL responses may also be analyzed. Purified PBMC are cultured in a 96-well flat bottom plate at a density of $1.5x10^5$ cells/well and are stimulated with 10 μg/ml synthetic peptide of the invention, whole 85P1B3 antigen, or PHA. Cells are routinely plated in replicates of 4-6 wells for each condition. After seven days of culture, the medium is removed and replaced with fresh medium containing 10U/ml IL-2. Two days later, 1 μCi ^3H-thymidine is added to each well and incubation is continued for an additional 18 hours. Cellular DNA is then harvested on glass fiber mats and analyzed for ^3H-thymidine incorporation. Antigen-specific T cell proliferation is calculated as the ratio of ^3H-thymidine incorporation in the presence of antigen divided by the ^3H-thymidine incorporation in the absence of antigen.

Example 27: Induction Of Specific CTL Response In Humans

A human clinical trial for an immunogenic composition comprising CTL and HTL epitopes of the invention is set up as an IND Phase I, dose escalation study and carried out as a randomized, double-blind, placebo-controlled trial. Such a trial is designed, for example, as follows:

A total of about 27 individuals are enrolled and divided into 3 groups:

Group I: 3 subjects are injected with placebo and 6 subjects are injected with 5 μg of peptide composition;

Group II: 3 subjects are injected with placebo and 6 subjects are injected with 50 μg peptide composition;

Group III: 3 subjects are injected with placebo and 6 subjects are injected with 500 μg of peptide composition.

After 4 weeks following the first injection, all subjects receive a booster inoculation at the same dosage.

The endpoints measured in this study relate to the safety and tolerability of the peptide composition as well as its immunogenicity. Cellular immune responses to the peptide composition are an

index of the intrinsic activity of this the peptide composition, and can therefore be viewed as a measure of biological efficacy. The following summarize the clinical and laboratory data that relate to safety and efficacy endpoints.

Safety: The incidence of adverse events is monitored in the placebo and drug treatment group and assessed in terms of degree and reversibility.

Evaluation of Vaccine Efficacy: For evaluation of vaccine efficacy, subjects are bled before and after injection. Peripheral blood mononuclear cells are isolated from fresh heparinized blood by Ficoll-Hypaque density gradient centrifugation, aliquoted in freezing media and stored frozen. Samples are assayed for CTL and HTL activity.

The vaccine is found to be both safe and efficacious.

Example 28: Phase II Trials In Patients Expressing 85P1B3

Phase II trials are performed to study the effect of administering the CTL-HTL peptide compositions to patients having cancer that expresses 85P1B3. The main objectives of the trial are to determine an effective dose and regimen for inducing CTLs in cancer patients that express 85P1B3, to establish the safety of inducing a CTL and HTL response in these patients, and to see to what extent activation of CTLs improves the clinical picture of these patients, as manifested, e.g., by the reduction and/or shrinking of lesions. Such a study is designed, for example, as follows:

The studies are performed in multiple centers. The trial design is an open-label, uncontrolled, dose escalation protocol wherein the peptide composition is administered as a single dose followed six weeks later by a single booster shot of the same dose. The dosages are 50, 500 and 5,000 micrograms per injection. Drug-associated adverse effects (severity and reversibility) are recorded.

There are three patient groupings. The first group is injected with 50 micrograms of the peptide composition and the second and third groups with 500 and 5,000 micrograms of peptide composition, respectively. The patients within each group range in age from 21-65 and represent diverse ethnic backgrounds. All of them have a tumor that expresses 85P1B3.

Clinical manifestations or antigen-specific T-cell responses are monitored to assess the effects of administering the peptide compositions. The vaccine composition is found to be both safe and efficacious in the treatment of 85P1B3-associated disease.

Example 29: Induction of CTL Responses Using a Prime Boost Protocol

A prime boost protocol similar in its underlying principle to that used to confirm the efficacy of a DNA vaccine in transgenic mice, such as described above in the Example entitled "The Plasmid Construct and the Degree to Which It Induces Immunogenicity," can also be used for the administration of the

vaccine to humans. Such a vaccine regimen can include an initial administration of, for example, naked DNA followed by a boost using recombinant virus encoding the vaccine, or recombinant protein/polypeptide or a peptide mixture administered in an adjuvant.

For example, the initial immunization may be performed using an expression vector, such as that constructed in the Example entitled "Construction of 'Minigene' Multi-Epitope DNA Plasmids" in the form of naked nucleic acid administered IM (or SC or ID) in the amounts of 0.5-5 mg at multiple sites. The nucleic acid (0.1 to 1000 μg) can also be administered using a gene gun. Following an incubation period of 3-4 weeks, a booster dose is then administered. The booster can be recombinant fowlpox virus administered at a dose of 5-10^7 to 5x10^9 pfu. An alternative recombinant virus, such as an MVA, canarypox, adenovirus, or adeno-associated virus, can also be used for the booster, or the polyepitopic protein or a mixture of the peptides can be administered. For evaluation of vaccine efficacy, patient blood samples are obtained before immunization as well as at intervals following administration of the initial vaccine and booster doses of the vaccine. Peripheral blood mononuclear cells are isolated from fresh heparinized blood by Ficoll-Hypaque density gradient centrifugation, aliquoted in freezing media and stored frozen. Samples are assayed for CTL and HTL activity.

Analysis of the results indicates that a magnitude of response sufficient to achieve a therapeutic or protective immunity against 85P1B3 is generated.

Example 30: Administration of Vaccine Compositions Using Dendritic Cells (DC)

Vaccines comprising peptide epitopes of the invention can be administered using APCs, or "professional" APCs such as DC. In this example, peptide-pulsed DC are administered to a patient to stimulate a CTL response *in vivo*. In this method, dendritic cells are isolated, expanded, and pulsed with a vaccine comprising peptide CTL and HTL epitopes of the invention. The dendritic cells are infused back into the patient to elicit CTL and HTL responses *in vivo*. The induced CTL and HTL then destroy or facilitate destruction, respectively, of the target cells that bear the 85P1B3 protein from which the epitopes in the vaccine are derived.

For example, a cocktail of epitope-comprising peptides is administered *ex vivo* to PBMC, or isolated DC therefrom. A pharmaceutical to facilitate harvesting of DC can be used, such as Progenipoietin™ (Monsanto, St. Louis, MO) or GM-CSF/IL-4. After pulsing the DC with peptides, and prior to reinfusion into patients, the DC are washed to remove unbound peptides.

As appreciated clinically, and readily determined by one of skill based on clinical outcomes, the number of DC reinfused into the patient can vary (*see, e.g., Nature Med.* 4:328, 1998; *Nature Med.* 2:52, 1996 and *Prostate* 32:272, 1997). Although 2-50 x 10^6 DC per patient are typically administered, larger

number of DC, such as 10^7 or 10^8 can also be provided. Such cell populations typically contain between 50-90% DC.

In some embodiments, peptide-loaded PBMC are injected into patients without purification of the DC. For example, PBMC generated after treatment with an agent such as Progenipoietin™ are injected into patients without purification of the DC. The total number of PBMC that are administered often ranges from 10^8 to 10^{10}. Generally, the cell doses injected into patients is based on the percentage of DC in the blood of each patient, as determined, for example, by immunofluorescence analysis with specific anti-DC antibodies. Thus, for example, if Progenipoietin™ mobilizes 2% DC in the peripheral blood of a given patient, and that patient is to receive 5 x 10^6 DC, then the patient will be injected with a total of 2.5 x 10^8 peptide-loaded PBMC. The percent DC mobilized by an agent such as Progenipoietin™ is typically estimated to be between 2-10%, but can vary as appreciated by one of skill in the art.

Ex vivo activation of CTL/HTL responses

Alternatively, *ex vivo* CTL or HTL responses to 85P1B3 antigens can be induced by incubating, in tissue culture, the patient's, or genetically compatible, CTL or HTL precursor cells together with a source of APC, such as DC, and immunogenic peptides. After an appropriate incubation time (typically about 7-28 days), in which the precursor cells are activated and expanded into effector cells, the cells are infused into the patient, where they will destroy (CTL) or facilitate destruction (HTL) of their specific target cells, *i.e.*, tumor cells.

Example 31: An Alternative Method of Identifying and Confirming Motif-Bearing Peptides

Another method of identifying and confirming motif-bearing peptides is to elute them from cells bearing defined MHC molecules. For example, EBV transformed B cell lines used for tissue typing have been extensively characterized to determine which HLA molecules they express. In certain cases these cells express only a single type of HLA molecule. These cells can be transfected with nucleic acids that express the antigen of interest, *e.g.* 85P1B3. Peptides produced by endogenous antigen processing of peptides produced as a result of transfection will then bind to HLA molecules within the cell and be transported and displayed on the cell's surface. Peptides are then eluted from the HLA molecules by exposure to mild acid conditions and their amino acid sequence determined, *e.g.*, by mass spectral analysis (*e.g.*, Kubo *et al.*, *J. Immunol.* 152:3913, 1994). Because the majority of peptides that bind a particular HLA molecule are motif-bearing, this is an alternative modality for obtaining the motif-bearing peptides correlated with the particular HLA molecule expressed on the cell.

Alternatively, cell lines that do not express endogenous HLA molecules can be transfected with an expression construct encoding a single HLA allele. These cells can then be used as described, *i.e.*, they can

then be transfected with nucleic acids that encode 85P1B3 to isolate peptides corresponding to 85P1B3 that have been presented on the cell surface. Peptides obtained from such an analysis will bear motif(s) that correspond to binding to the single HLA allele that is expressed in the cell.

As appreciated by one in the art, one can perform a similar analysis on a cell bearing more than one HLA allele and subsequently determine peptides specific for each HLA allele expressed. Moreover, one of skill would also recognize that means other than transfection, such as loading with a protein antigen, can be used to provide a source of antigen to the cell.

Example 32: Complementary Polynucleotides

Sequences complementary to the 85P1B3-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring 85P1B3. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using, e.g., OLIGO 4.06 software (National Biosciences) and the coding sequence of 85P1B3. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the 85P1B3-encoding transcript.

Example 33: Purification of Naturally-occurring or Recombinant 85P1B3 Using 85P1B3 Specific Antibodies

Naturally occurring or recombinant 85P1B3 is substantially purified by immunoaffinity chromatography using antibodies specific for 85P1B3. An immunoaffinity column is constructed by covalently coupling anti-85P1B3 antibody to an activated chromatographic resin, such as CNBr-activated SEPHAROSE (Amersham Pharmacia Biotech). After the coupling, the resin is blocked and washed according to the manufacturer's instructions.

Media containing 85P1B3 are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of 85P1B3 (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/85P1B3 binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and GCR.P is collected.

Example 34: Identification of Molecules Which Interact with 85P1B3

85P1B3, or biologically active fragments thereof, are labeled with 121 1 Bolton-Hunter reagent.

(See, e.g., Bolton et al. (1973) Biochem. J. 133:529.) Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled 85P1B3, washed, and any wells with labeled 85P1B3 complex are assayed. Data obtained using different concentrations of 85P1B3 are used to calculate values for the number, affinity, and association of 85P1B3 with the candidate molecules.

Throughout this application, various website data content, publications, applications and patents are referenced. (Websites are referenced by their Uniform Resource Locator, or URL, addresses on the World Wide Web.) The disclosures of each of these items of information are are hereby incorporated by reference herein in their entireties.

Example 35: *In Vivo* Assay for 85P1B3 Tumor Growth Promotion

The effect of the 85P1B3 protein on tumor cell growth can be confirmed *in vivo* by gene overexpression in a variety of cancer cells, including prostate, kidney and bladder. For example, SCID mice can be injected SQ on each flank with 1 x 10^6 prostate, kidney or bladder cancer cells (such as PC3, LNCaP, SCaBER, UM-UC-3, HT1376, RT4, T24, Caki, A-498 and SW839 cells) containing tkNeo empty vector or 85P1B3.

At least two strategies may be used: (1) Constitutive 85P1B3 expression under regulation of a promoter such as a constitutive promoter obtained from the genomes of viruses such as polyoma virus, fowlpox virus (UK 2,211,504 published 5 July 1989), adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis-B virus and Simian Virus 40 (SV40), or from heterologous mammalian promoters, *e.g.*, the actin promoter or an immunoglobulin promoter, provided such promoters are compatible with the host cell systems. (2) Regulated expression under control of an inducible vector system, such as ecdysone, tet, etc., can be used provided such promoters are compatible with the host cell systems. Tumor volume is then monitored at the appearance of palpable tumors and is followed over time to validate that 85P1B3-expressing cells grow at a faster rate and that tumors produced by 85P1B3-expressing cells demonstrate characteristics of altered aggressiveness (e.g. enhanced metastasis, vascularization, reduced responsiveness to chemotherapeutic drugs). Additionally, mice can be implanted with the same cells orthotopically in the prostate, bladder or kidney to determine if 85P1B3 has an effect on local growth in the prostate, bladder or kidney or on the ability of the cells to metastasize, specifically to lungs or lymph nodes (Fu, X., *et al.*, Int. J. Cancer, 1991. **49**: p. 938-939; Chang, S., *et al.*, Anticancer Res., 1997. **17**: p. 3239-3242; Peralta, E. A., *et al.*, J. Urol., 1999. **162**: p. 1806-1811).

Furthermore, this assay is useful to confirm the 85P1B3 inhibitory effect of candidate therapeutic compositions, such as for example, 85P1B3 antibodies or intrabodies, and 85P1B3 antisense molecules or ribozymes.

Example 36: 85P1B3 Monoclonal Antibody-mediated Inhibition of Tumors *In Vivo*

The significant expression of 85P1B3 in cancer tissues, together with its restricted expression in normal tissues, makes 85P1B3 an excellent target for antibody therapy. In cases where the monoclonal antibody target is a cell surface protein, antibodies have been shown to be efficacious at inhibiting tumor growth (See, e.g., (Saffran, D., *et al.*, PNAS 10:1073-1078 or www.pnas.org/cgi/doi/10.1073/pnas.051624698). In cases where the target is not on the cell surface, such as PSA and PAP in prostate cancer, antibodies have also been shown to recognize and inhibit growth of cells expressing those proteins (Saffran, D.C., *et al.*, Cancer and Metastasis Reviews, 1999. 18: p. 437-449). As with any cellular protein with a restricted expression profile, 85P1B3 is a target for T cell-based immunotherapy.

Accordingly, the therapeutic efficacy of anti-85P1B3 mAbs in human colon, kidney, bladder and prostate cancer mouse models is modeled in 85P1B3-expressing kidney, bladder or prostate cancer xenografts or cancer cell lines, such as those described in the Example entitled "*In Vivo* Assay for 85P1B3 Tumor Growth Promotion", that have been engineered to express 85P1B3.

Antibody efficacy on tumor growth and metastasis formation is confirmed, e.g., in a mouse orthotopic in the prostate, colon, bladder or kidney cancer xenograft model. The antibodies can be unconjugated, or can be conjugated to a therapeutic modality, as appreciated in the art. It is confirmed that anti-85P1B3 mAbs inhibit formation of 85P1B3-expressing kidney, bladder and prostate tumors. Anti-85P1B3 mAbs also retard the growth of established orthotopic tumors and prolong survival of tumor-bearing mice. These results indicate the utility of anti-85P1B3 mAbs in the treatment of local and advanced stages of cancer. (See, e.g., Saffran, D., et al., PNAS 10:1073-1078 or www.pnas.org/cgi/doi/10.1073/pnas.051624698)

Administration of anti-85P1B3 mAbs retard established orthotopic tumor growth and inhibit metastasis to distant sites, resulting in a significant prolongation in the survival of tumor-bearing mice. These studies indicate that 85P1B3 is an attractive target for immunotherapy and demonstrate the therapeutic potential of anti-85P1B3 mAbs for the treatment of local and metastatic kidney, colon, bladder and prostate cancer.

This example demonstrates that unconjugated 85P1B3 monoclonal antibodies effectively to inhibit the growth of human bladder tumors grown in SCID mice; accordingly a combination of such efficacious monoclonal antibodies is also effective.

Example 37: Induction Of Specific CTL Response In Humans

A human clinical trial for an immunogenic composition comprising CTL and HTL epitopes of the invention is set up as an IND Phase I, dose escalation study and carried out as a randomized, double-blind, placebo-controlled trial. Such a trial is designed, for example, as follows:

A total of about 27 individuals are enrolled and divided into 3 groups:

Group I: 3 subjects are injected with placebo and 6 subjects are injected with 5 μg of peptide composition;

Group II: 3 subjects are injected with placebo and 6 subjects are injected with 50 μg peptide composition;

Group III: 3 subjects are injected with placebo and 6 subjects are injected with 500 μg of peptide composition.

After 4 weeks following the first injection, all subjects receive a booster inoculation at the same dosage.

The endpoints measured in this study relate to the safety and tolerability of the peptide composition as well as its immunogenicity. Cellular immune responses to the peptide composition are an index of the intrinsic activity of this the peptide composition, and can therefore be viewed as a measure of biological efficacy. The following summarize the clinical and laboratory data that relate to safety and efficacy endpoints.

Safety: The incidence of adverse events is monitored in the placebo and drug treatment group and assessed in terms of degree and reversibility.

Evaluation of Vaccine Efficacy: For evaluation of vaccine efficacy, subjects are bled before and after injection. Peripheral blood mononuclear cells are isolated from fresh heparinized blood by Ficoll-Hypaque density gradient centrifugation, aliquoted in freezing media and stored frozen. Samples are assayed for CTL and HTL activity.

The vaccine is found to be both safe and efficacious.

Example 38: Splice Variants of 85P1B3

Splice variants are also called alternative transcripts. When a gene is transcribed from genomic DNA, the initial RNA is generally spliced to produce functional mRNA, which has only exons and is used for translation into an amino acid sequence. Accordingly, a given gene can have zero to many alternatively spliced mRNA products. Alternative transcripts each have a unique exon makeup, and can have different coding and/or non-coding (5' or 3' end) portions, from the original transcript. Alternative transcripts can code for similar proteins with the same or a similar function or may encode proteins with different

functions, and may be expressed in the same tissue at the same time, or at different tissue at different times, proteins encoded by alternative transcripts can have similar or different cellular or extracellular localizations, e.g., be secreted.

Splice variants are identified by a variety of art-accepted methods. For example, splice variants are identified by use of EST data. First, all human ESTs were grouped into clusters which show direct or indirect identity with each other. Second, ESTs in the same cluster were further grouped into sub-clusters and assembled into a consensus sequence. The starting gene is compared to the consensus sequence(s). Each consensus sequence is a potential splice variant for that gene (see, e.g., http://www.doubletwist.com/products/c11_agentsOverview.jhtml). Even when a variant is identified that is not a full-length clone, that portion of the variant is very useful for antigen generation and for further cloning of the full-length splice variant, using techniques known in the art.

Moreover, computer programs are available in the art that identify splice variants based on genomic sequences. Genomic-based variant identification programs include FgenesH (A. Salamov and V. Solovyev, "Ab initio gene finding in Drosophila genomic DNA," Genome Research. 2000 April;10(4):516-22); Grail (http://compbio.ornl.gov/Grail-bin/EmptyGrailForm) and GenScan (http://genes.mit.edu/GENSCAN.html). For a general discussion of splce variant identification protocols see., e.g., Southan C., "A genomic perspective on human proteases," FEBS Lett. 2001 Jun 8;498(2-3):214-8; de Souza SJ, et al., "Identification of human chromosome 22 transcribed sequences with ORF expressed sequence tags," Proc. Natl Acad Sci U S A. 2000 Nov 7;97(23):12690-3.

For variants identified by the EST-based method, Table XXI shows the nucleotide sequences of the splice variants. Figure Table XXII shows the alignment of the splice variant with the 85P1B3 nucleic acid sequence. Table XXIII displays the single longest alignment of an amino acid sequence encoded by a splice variant, out of all six potential reading frames with 85P1B3. Thus, for each splice variant, a variant's reading frame that encodes the longest single contiguous peptide homology between 85P1B3 and the variant is the proper reading frame orientation for the variant. Due to the possibility of sequencing errors in EST or genomic data, other peptides in the relevant reading frame orientation (5' to 3' or 3' to 5') can also be encoded by the variant. Table XXIV lays out all three frame shifted amino acid translations of the splice variant for the identified reading frame orientation.

For variants identified by any one of the genomic sequence-based methods, Table XXI shows the nucleotide sequences of the splice variant. Figure Table XXII shows the alignment of the splice variant with the 85P1B3 nucleic acid sequence. Table XXIII displays the alignment of amino acid sequence of the predicted transcripts with 85P1B3. The genomic-based computer programs predict a transcript from genomic sequence, and not only predict exons but also set open reading frame as the first forward open reading frame. The predicted transcript does not contain 5' or 3' untranslated region (UTR). It starts with

ATG and ends with a stop codon, TAG, TGA or TAA. In case the transcript is predicted on the reverse strand of the genomic sequence, the sequence of the transcript is reverse-complemented to the genomic sequence of the exons. Thus, the genomic-based programs provide the correct transcript sequence, with 5' to 3' orientation and +1 as the open reading frame. However, due to the possibility of inaccurate prediction of exons or the possibility of sequencing errors in genomic data, other peptides in other forward open reading frame can also be encoded by the variant. Table XXIV lays out all amino acid translations of the splice variant in each of the three forward reading frames.

To further confirm the parameters of a splice variant, a variety of techniques are available in the art, such as proteomic validation, PCR-based validation, and 5' RACE validation, etc. (see e.g., Proteomic Validation: Brennan SO, Fellowes AP, George PM.; "Albumin banks peninsula: a new termination variant characterised by electrospray mass spectrometry." Biochim Biophys Acta. 1999 Aug 17;1433(1-2):321-6; Ferranti P, et al., "Differential splicing of pre-messenger RNA produces multiple forms of mature caprine alpha(s1)-casein." Eur J Biochem. 1997 Oct 1;249(1):1-7; PCR-based Validation: Wellmann S, et al., "Specific reverse transcription-PCR quantification of vascular endothelial growth factor (VEGF) splice variants by LightCycler technology." Clin Chem. 2001 Apr;47(4):654-60; Jia HP, et al., Discovery of new human beta-defensins using a genomics-based approach," Gene. 2001 Jan 24;263(1-2):211-8; PCR-based and 5' RACE Validation: Brigle KE, et al., "Organization of the murine reduced folate carrier gene and identification of variant splice forms," Biochim Biophys Acta. 1997 Aug 7; 1353(2): 191-8.

It is known in the art that genomic regions are upregulated in cancers. When the genomic region to which 85P1B3 maps is upregulated in a particular cancer, the splice variants of 85P1B3 are upregulated as well. Disclosed herein is that 85P1B3 has a particular expression profile. Splice variants of 85P1B3 that are structurally and/or functionally similar to 85P1B3 share this expression pattern, thus serving as tumor-associated markers/antigens.

Using the EST assembly approach, we identified one splice variants designated splice variant 1.

Table XXIA. Nucleotide sequence of splice variant 1.

```
  1 TTTTTTTTTT CCTATCTAGC TATCTCTTAA AAACAAAAGC CATAGTAAAT GCATCAGAGA
 61 TGGATATTCA AAATGTTCCT CTATCAGAAA AGATTGCAGA GGTAAAATTT CATGATGGTT
121 GTATGCTTTT TTAAAATACA GACAACTCTT GATAACTTCT ACCAATGAAC TTGGGGATGA
181 TGAAATGGCA TGATGCTCAA TAATCCTTTT TACTTGATTT GACCTTCCCT ATTGAATTTG
241 TAATGAAAAA CAAAATACTA AAACCACACT GTAAGGTATA GTTCAGGAAG AAAGGAAAAG
301 CTGCTCAACT GCTGCACTCC TGCATTCTCC TTTGTGCTGG GAATGGATAT CATCATCTTG
361 CCATAGAGGT GTCTTCTTTG CAAATACCTT GTAATTGCTC AACTGTCTCA GACATAAGAG
421 TGATGAAACA GTTATTAAGA ATTCCTGGCC GGGCGTGGTG GCTCACGCCT GTAATCCCAG
481 CACTTTGGCC TCGTGC
```

Table XXIIA. Nucleotide sequence alignment of 85P1B3 with splice variant 1.

```
Score =  160 bits (83), Expect = 3e-36
Identities = 83/83 (100%)
 Strand = Plus / Plus

85P1B3: 524 gctatctcttaaaaacaaaagccatagtaaatgcatcagagatggatattcaaaatgttc
583 ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
gctatctcttaaaaacaaaagccatagtaaatgcatcagagatggatattcaaaatgttc 78
Vrnt 1: 19

85P1B3: 584 ctctatcagaaaagattgcagag 606
            ||||||||||||||||||||||
Vrnt 1: 79  ctctatcagaaaagattgcagag 101
```

Table XXIIIA. Amino acid sequence alignment of 85P1B3 and splice variant 1.

```
Score = 64.8 bits (135), Expect = 2e-08
Identities = 28/29 (96%)
 Frame = +1 / +3

85P1B3: 526 YLLKTKAIVNASEMDIQNVPLSEKIAELK 612
            YLLKTKAIVNASEMDIQNVPLSEKIAE+K
Vrnt 1: 21  YLLKTKAIVNASEMDIQNVPLSEKIAEVK 107
```

Table XXIVA. Peptide sequences from the translation of the nucleotide sequence of splice variant 1.

Open reading frame	Amino acid sequences
Frame 1	FFFSYLAIS*KQKP**MHQRWIFKMFLYQKRLQR*NFMMVVCFFKIQTTLDNFYQ*TWG* *NGMMLNNPFYLI*PSLLNL**KTKY*NHTVRYSSGRKEKLLNCCTPAFSFVLGMDIIIL P*RCLLCKYLVIAQLSQT*E**NSY*EFLAGRGGSRL*SQHFGLV
Frame 2	FFFPI*LSLKNKSHSKCIRDGYSKCSSIRKDCRGKIS*WLYAFLKYRQLLITSTNELGDD EMA*CSIILFT*FDLPY*ICNEKQNTKTTL*GIVQEERKSCSTAALLHSPLCWEWISSSC HRGVFFANTL*LLNCLRHKSDETVIKNSWPGVVAHACNPSTLASC
Frame 3	FFFLSSYLLKTKAIVNASEMDIQNVPLSEKIAEVKFHDGCMLF*NTDNS**LLPMNLGMM KWHDAQ*SFLLDLTFPIEFVMKNKILKPHCKV*FRKKGKAAQLLHSCILLCAGNGYHHLA IEVSSLQIPCNCSTVSDIRVMKQLLRIPGRAWWLTPVIPALWPR

Note: Frame 3 gives the longest subsequence that is identical with 85P1B3 amino acid sequence. In this Table each (*)indicates the product of a single codon, i.e., a single unknown amino acid or a stop codon.

Example 39: Expression analysis of 85P1B3 splice variants in normal tissues and patient tumor specimens

Expression of 85P1B3 described in Example 4 was performed using the 85P1B3 SSH sequence as a probe. This nucleic acid sequence spans region 701-1019 of the 85P1B3 gene, a region absent in the 85P1B3 splice variant 1 (Figure19). Therefore, the Northern blots described in Figure 11, Figure 12, Figure 13, Figure 14, Figure 15, Figure 16, Figure 17, and Figure 18 detected the transcript of 85P1B3 but not of splice variant 1.

A probe comprising region of homology between 85P1B3 and its splice variant 1 is generated (Probe 1). This region spans nucleotide positions 524-606 of 85P1B3 and 19-101 of splice variant 1. Normal tissue northern blots and patient cancer northern blots are probed with probe 1. The results have two bands, and show expression of the 1.2 kb transcript of 85P1B3 and the transcript of its splice variant 1.

In another study, a probe comprising a region present in the ORF of the splice variant but not in the ORF of 85P1B3 is generated (Probe 2). This region spans nucleotide positions 102-496 of the splice variant 1. Normal tissue northern blots and patient cancer Northern blots are probed with probe 2. The results have single bands, and show expression of splice variant 1 but not the transcript of 85P1B3.

When 85P1B3 splice variant 1 is expressed in patient cancer specimens, and shows restricted expression in normal tissues, 85P1B3 splice variant 1 is a suitable cancer target for cancer diagnosis and therapy.

Example 40: Splice Variant Protein Characteristics

The present variant protein is understood to be partial, and thus to comprise domains of the full protein. Amino acids 7-35 of the 85P1B3 variant 1 protein align with amino acids 172-200 of 85P1B3 with 96% identity, while the remaining downstream amino acids diverge from the 85P1B3. This pattern of high homology to one section of the parent protein coupled to a high divergence from the remaining portions of the parent protein form the hallmark of a splice variant.

Protein blast analysis of variant 1 shows that the 85P1B3 variant is homologous to OIP5, a human protein known to be involved in adhesion and invasion of epithelial cells (Brooks GF et al, Mol Microbiol. 1991, 5:3063; Weel JF et al, J Exp Med. 1991, 173:1395), with 96% identity over 28 amino acids. Analysis by pFam or prosite failed to identify any motifs. However motif homology was observed to Glyoxalase I at aa 114-153 of the variant protein. Glyoxalase is a glutathione-mediating detoxifying enzyme, that protects cells from advanced glycation endproducts (AGEs) (Thornalley PJ. Chem Biol Interact. 1998, 111:137). Glyoxalase is highly expressed in breast cancer cells (Rulli A et al, Breast Cancer Res Treat. 2001, 66:67).

Regarding localization, the 85P1B3 variant localizes to the cytoplasm (cytoplasmic 60.9% PSORT II) or the mitochondria (mitochondrial 0.519, PSORT).

Based on bioinformatic analysis (TMPred, Sosui) the 85P1B3 variant does not appear to contain transmembrane domains, but forms a soluble intracellular protein. Due to its homology to OIP5 and Glyoxalase I, 85P1B3 is involved in the adhesion and invasion of epithelial cells, and has a cancer-related expression pattern.

Example 41: Homology Comparison of 85P1B3 to Known Sequences

The 85P1B3 protein of Figure 3 has 229 amino acids with calculated molecular weight of 24.69 kDa, and pI of 7.02. 85P1B3 is predicted to be a mitochondrial (60.9%) or cytoplasmic (21.7) protein.

85P1B3 shows best homology to human Opa interacting protein 5 (gi 2815610, http://www.ncbi.nlm.nih.gov) sharing 100% identity with that protein. Opacity associated proteins (Opa) were identified in *Neisseria gonorrhoeae* as outer membrane proteins that are involved in mediating the adhesion of *Neisseria* to mammalian cells and the invasion of human epithelial cells (Brooks GF et al, Mol Microbiol. 1991, 5:3063; Weel JF et al, J Exp Med. 1991, 173:1395). OPA proteins bind to membrane proteins, such as CD66 and carcinoembryonic antigen related cellular molecule (CEACAM), on the surface of human epithelial and mononuclear cells, thereby facilitating entry of *Neisseria* into mammalian host cells (Muenzner P et al, J. Biol. Chem. 2001, 276:24331; Chen T et al, J. Exp. Med. 1997, 185:1557). In order to delineate the role of Opa in adherence and invasion of human cells, Williams et al used a two yeast hybrid system to identify Opa interacting proteins (Williams JM et al, Mol. Microbiol. 1998, 27:171). Screening a human cDNA library for Opa interacting partners, they identified Opa interacting protein 5 or OIP5. OIP5 is an intracellular, cytoplasmic protein with homology to thyroid hormone receptor interacting protein-6 (TRIP6) (Williams JM & Rest RF. Ann N Y Acad Sci. 1996, 797:288). TRIP6 is an intracellular signaling molecule that relays information to the nucleus thereby regulating gene expression (Zhao M et al, Gene Expr. 1999, :207; Wang Y et al, Gene. 1999, 234:403).

This information indicates that 85P1B3 can play a role in the adhesion and invasion of epithelial cells into adjacent tissues and basement membranes, and regulate transcription by transmitting cell surface signals to the nucleus.

Accordingly, when 85P1B3 functions as a regulator of cell adhesion and invasion, or as a modulator of transcription involved in activating genes associated with tumorigenesis or in blocking expression of genes that repress tumorigenesis, 85P1B3 is used for therapeutic, diagnostic, prognostic and/or preventative purposes.

Example 42: Identification and Confirmation of Potential Signal Transduction Pathways

Many mammalian proteins have been reported to interact with signaling molecules and to participate in regulating signaling pathways. (J Neurochem. 2001; 76:217-223). In particular, OPA has been reported to associate with a phosphatase and surface receptors (Hauck C et al, Infect. Immun. 1999, 67:5490; Muenzner P et al, J. Biol. Chem. 2001, 276:24331). Using immunoprecipitation and Western blotting techniques, proteins are identified that associate with 85P1B3 and mediate signaling events. Several pathways known to play a role in cancer biology can be regulated by 85P1B3, including

phospholipid pathways such as PI3K, AKT, etc, adhesion and migration pathways, including FAK, Rho, Rac-1, etc, as well as mitogenic/survival cascades such as ERK, p38, etc (Cell Growth Differ. 2000,11:279; J Biol Chem. 1999, 274:801; Oncogene. 2000, 19:3003, J. Cell Biol. 1997, 138:913.).

Using, e.g., Western blotting techniques the ability of 85P1B3 to regulate these pathways is confirmed. Cells expressing or lacking 85P1B3 are either left untreated or stimulated with cytokines, androgen and anti-integrin antibodies. Cell lysates are analyzed using anti-phospho-specific antibodies (Cell Signaling, Santa Cruz Biotechnology) in order to detect phosphorylation and regulation of ERK, p38, AKT, PI3K, PLC and other signaling molecules. When 85P1B3 plays a role in the regulation of signaling pathways, whether individually or communally, it is used as a target for diagnostic, prognostic, preventative and/or therapeutic purposes.

To confirm that 85P1B3 directly or indirectly activates known signal transduction pathways in cells, luciferase (luc) based transcriptional reporter assays are carried out in cells expressing individual genes. These transcriptional reporters contain consensus-binding sites for known transcription factors that lie downstream of well-characterized signal transduction pathways. The reporters and examples of these associated transcription factors, signal transduction pathways, and activation stimuli are listed below.

1. NFkB-luc, NFkB/Rel; Ik-kinase/SAPK; growth/apoptosis/stress
2. SRE-luc, SRF/TCF/ELK1; MAPK/SAPK; growth/differentiation
3. AP-1-luc, FOS/JUN; MAPK/SAPK/PKC; growth/apoptosis/stress
4. ARE-luc, androgen receptor; steroids/MAPK; growth/differentiation/apoptosis
5. p53-luc, p53; SAPK; growth/differentiation/apoptosis
6. CRE-luc, CREB/ATF2; PKA/p38; growth/apoptosis/stress

Gene-mediated effects can be assayed in cells showing mRNA expression. Luciferase reporter plasmids can be introduced by lipid-mediated transfection (TFX-50, Promega). Luciferase activity, an indicator of relative transcriptional activity, is measured by incubation of cell extracts with luciferin substrate and luminescence of the reaction is monitored in a luminometer.

Signaling pathways activated by 85P1B3 are mapped and used for the identification and validation of therapeutic targets. When 85P1B3 is involved in cell signaling, it is used as target for diagnostic, prognostic, preventative and/or therapeutic purposes.

Example 43: Involvement in Tumor Progression

The 85P1B3 gene can contribute to the growth of cancer cells. The role of 85P1B3 in tumor growth is confirmed in a variety of primary and transfected cell lines including prostate, colon, bladder and

kidney cell lines, as well as NIH 3T3 cells engineered to stably express 85P1B3. Parental cells lacking 85P1B3 and cells expressing 85P1B3 are evaluated for cell growth using a well-documented proliferation assay (Fraser SP, Grimes JA, Djamgoz MB. Prostate. 2000;44:61, Johnson DE, Ochieng J, Evans SL. Anticancer Drugs. 1996, 7:288).

To confirm the role of 85P1B3 in the transformation process, its effect in colony forming assays is investigated. Parental NIH3T3 cells lacking 85P1B3 are compared to NHI-3T3 cells expressing 85P1B3, using a soft agar assay under stringent and more permissive conditions (Song Z. et al. Cancer Res. 2000;60:6730).

To confirm the role of 85P1B3 in invasion and metastasis of cancer cells, a well-established assay is used, e.g., a Transwell Insert System assay (Becton Dickinson) (Cancer Res. 1999; 59:6010). Control cells, including prostate, colon, bladder and kidney cell lines lacking 85P1B3 are compared to cells expressing 85P1B3. Cells are loaded with the fluorescent dye, calcein, and plated in the top well of the Transwell insert coated with a basement membrane analog. Invasion is determined by fluorescence of cells in the lower chamber relative to the fluorescence of the entire cell population.

85P1B3 can also play a role in cell cycle and apoptosis. Parental cells and cells expressing 85P1B3 are compared for differences in cell cycle regulation using a well-established BrdU assay (Abdel-Malek ZA. J Cell Physiol. 1988, 136:247). In short, cells are grown under both optimal (full serum) and limiting (low serum) conditions are labeled with BrdU and stained with anti-BrdU Ab and propidium iodide. Cells are analyzed for entry into the G1, S, and G2M phases of the cell cycle. Alternatively, the effect of stress on apoptosis is evaluated in control parental cells and cells expressing 85P1B3, including normal and tumor prostate, colon and lung cells. Engineered and parental cells are treated with various chemotherapeutic agents, such as etoposide, flutamide, etc, and protein synthesis inhibitors, such as cycloheximide. Cells are stained with annexin V-FITC and cell death is measured by FACS analysis. The modulation of cell death by 85P1B3 can play a critical role in regulating tumor progression and tumor load.

When 85P1B3 plays a role in cell growth, transformation, invasion or apoptosis, it is used as a target for diagnostic, prognostic, preventative and/or therapeutic purposes.

Example 44: Involvement in Angiogenesis

Angiogenesis or new capillary blood vessel formation is necessary for tumor growth (Hanahan D, Folkman J. Cell. 1996, 86:353; Folkman J. Endocrinology. 1998 139:441). Several assays have been developed to measure angiogenesis *in vitro* and *in vivo*, such as the tissue culture assays endothelial cell tube formation and endothelial cell proliferation. Using these assays as well as *in vitro* neo-vascularization, the role of 85P1B3 in angiogenesis, enhancement or inhibition, is confirmed .

For example, endothelial cells engineered to express 85P1B3 are evaluated using tube formation and proliferation assays. The effect of 85P1B3 is also confirmed in animal models *in vivo*. For example, cells either expressing or lacking 85P1B3 are implanted subcutaneously in immunocompromised mice. Endothelial cell migration and angiogenesis are evaluated 5-15 days later using immunohistochemistry techniques. When 85P1B3 affects angiogenesis, it is used as a target for diagnostic, prognostic, preventative and/or therapeutic purposes

Example 45: Regulation of Transcription

The cytoplasmic localization of 85P1B3 and its similarity to TRIP5 support the use in accordance with the present invention of 85P1B3 as a modulator of the transcriptional regulation of eukaryotic genes. Regulation of gene expression is confirmed, e.g., by studying gene expression in cells expressing or lacking 85P1B3. For this purpose, two types of experiments are performed.

In the first set of experiments, RNA from parental and 85P1B3-expressing cells are extracted and hybridized to commercially available gene arrays (Clontech) (Smid-Koopman E et al. Br J Cancer. 2000. 83:246). Resting cells as well as cells treated with FBS or androgen are compared. Differentially expressed genes are identified in accordance with procedures known in the art. The differentially expressed genes are then mapped to biological pathways (Chen K et al. Thyroid. 2001. 11:41.).

In the second set of experiments, specific transcriptional pathway activation is evaluated using commercially available (Stratagene) luciferase reporter constructs including: NFkB-luc, SRE-luc, ELK1-luc, ARE-luc, p53-luc, and CRE-luc. These transcriptional reporters contain consensus binding sites for known transcription factors that lie downstream of well-characterized signal transduction pathways, and represent a good tool to ascertain pathway activation and screen for positive and negative modulators of pathway activation.

When 85P1B3 plays a role in gene regulation, it is used as a target for diagnostic, prognostic, preventative and/or therapeutic purposes.

Example 46: Involvement in Cell Adhesion

Cell adhesion plays a critical role in tissue colonization and metastasis. 85P1B3 can participate in cellular organization, and as a consequence cell adhesion and motility. This is supported by the presence of an RGD motif in the N-terminal portion of 85P1B3 (see Table XIX). To confirm that 85P1B3 regulates cell adhesion, control cells lacking 85P1B3 are compared to cells expressing 85P1B3, using techniques previously described (see, e.g., Haier et al, Br. J. Cancer. 1999, 80:1867; Lehr and Pienta, J. Natl. Cancer Inst. 1998, 90:118). Briefly, in one embodiment, cells labeled with a fluorescent indicator, such as calcein,

are incubated on tissue culture wells coated with media alone or with matrix proteins. Adherent cells are detected by fluorimetric analysis and percent adhesion is calculated. In another embodiment, cells lacking or expressing 85P1B3 are analyzed for their ability to mediate cell-cell adhesion using similar experimental techniques as described above. Both of these experimental systems are used to identify proteins, antibodies and/or small molecules that modulate cell adhesion to extracellular matrix and cell-cell interaction. Since cell adhesion plays a critical role in tumor growth, progression, and, colonization, when 85P1B3 is involved in these processes it serves as a diagnostic, prognostic, preventative and/or therapeutic modality.

Example 47: Protein-Protein Association

Two proteins with homology to 85P1B3, namely OIP5 and TRIP6, have been shown to interact with other proteins, thereby regulating signal transduction, gene transcription, and cell adhesion . Using immunoprecipitation techniques as well as two yeast hybrid systems, proteins are identified that associate with 85P1B3. Immunoprecipitates from cells expressing 85P1B3 and cells lacking 85P1B3 are compared for specific protein-protein associations.

Studies are performed to confirm the extent of association of 85P1B with effector molecules, such as receptors, adaptor proteins and SH2-containing proteins. Studies comparing 85P1B3 positive and 85P1B3 negative cells as well as studies comparing unstimulated/resting cells and cells treated with epithelial cell activators, such as cytokines, growth factors, androgen and anti-integrin Ab reveal unique interactions.

In addition, protein-protein interactions are confirmed using two yeast hybrid methodology (Curr Opin Chem Biol. 1999, 3:64). A vector carrying a library of proteins fused to the activation domain of a transcription factor is introduced into yeast expressing a 85P1B3-DNA-binding domain fusion protein and a reporter construct. Protein-protein interaction is detected by colorimetric reporter activity. Specific association with effector molecules and transcription factors directs one of skill to the mode of action of 85P1B3, and thus identifies therapeutic, prognostic, preventative and/or diagnostic targets for cancer. This and similar assays are also used to identify and screen for small molecules that interact with 85P1B3.

When 85P1B3 associates with proteins or small molecules it is used as a target for diagnostic, prognostic, preventative and/or therapeutic purposes.

Throughout this application, various website data content, publications, patent applications and patents are referenced. (Websites are referenced by their Uniform Resource Locator, or URL, addresses on the World Wide Web.) The disclosures of each of these references are hereby incorporated by reference herein in their entireties.

The present invention is not to be limited in scope by the embodiments disclosed herein, which are intended as single illustrations of individual aspects of the invention, and any that are functionally equivalent are within the scope of the invention. Various modifications to the models and methods of the invention, in addition to those described herein, will become apparent to those skilled in the art from the foregoing description and teachings, and are similarly intended to fall within the scope of the invention. Such modifications or other embodiments can be practiced without departing from the true scope and spirit of the invention.

TABLES

TABLE I: Tissues that Express 85P1B3 When Malignant

- Prostate
- Bladder
- Kidney
- Colon
- Lung
- Ovary
- Breast
- Stomach
- Uterus
- Cervix

TABLE II: AMINO ACID ABBREVIATIONS

SINGLE LETTER	THREE LETTER	FULL NAME
F	Phe	phenylalanine
L	Leu	leucine
S	Ser	serine
Y	Tyr	tyrosine
C	Cys	cysteine
W	Trp	tryptophan
P	Pro	proline
H	His	histidine
Q	Gln	glutamine
R	Arg	arginine
I	Ile	isoleucine
M	Met	methionine
T	Thr	threonine
N	Asn	asparagine
K	Lys	lysine
V	Val	valine

A	Ala	alanine
D	Asp	aspartic acid
E	Glu	glutamic acid
G	Gly	glycine

TABLE III: AMINO ACID SUBSTITUTION MATRIX

Adapted from the GCG Software 9.0 BLOSUM62 amino acid substitution matrix (block substitution matrix). The higher the value, the more likely a substitution is found in related, natural proteins. (See URL www.ikp.unibe.ch/manual/blosum62.html)

A	C	D	E	F	G	H	I	K	L	M	N	P	Q	R	S	T	V	W	Y	.
4	0	-2	-1	-2	0	-2	-1	-1	-1	-1	-2	-1	-1	-1	1	0	0	-3	-2	A
	9	-3	-4	-2	-3	-3	-1	-3	-1	-1	-3	-3	-3	-3	-1	-1	-1	-2	-2	C
		6	2	-3	-1	-1	-3	-1	-4	-3	1	-1	0	-2	0	-1	-3	-4	-3	D
			5	-3	-2	0	-3	1	-3	-2	0	-1	2	0	0	-1	-2	-3	-2	E
				6	-3	-1	0	-3	0	0	-3	-4	-3	-3	-2	-2	-1	1	3	F
					6	-2	-4	-2	-4	-3	0	-2	-2	-2	0	-2	-3	-2	-3	G
						8	-3	-1	-3	-2	1	-2	0	0	-1	-2	-3	-2	2	H
							4	-3	2	1	-3	-3	-3	-3	-2	-1	3	-3	-1	I
								5	-2	-1	0	-1	1	2	0	-1	-2	-3	-2	K
									4	2	-3	-3	-2	-2	-2	-1	1	-2	-1	L
										5	-2	-2	0	-1	-1	-1	1	-1	-1	M
											6	-2	0	0	1	0	-3	-4	-2	N
												7	-1	-2	-1	-1	-2	-4	-3	P
													5	1	0	-1	-2	-2	-1	Q
														5	-1	-1	-3	-3	-2	R
															4	1	-2	-3	-2	S
																5	0	-2	-2	T
																	4	-3	-1	V
																		11	2	W
																			7	Y

TABLE IV A

SUPERMOTIFS	POSITION 2 (Primary Anchor)	POSITION 3 (Primary Anchor)	POSITION C Terminus (Primary Anchor)
A1	**TIL***VMS*		**FWY**
A2	**LIVM***ATQ*		**IV***MATL*
A3	**VSMA***TLI*		**RK**
A24	**YF***WIVLMT*		**FI***YWLM*
B7	**P**		**VILF***MWYA*
B27	**RHK**		**FYL***WMIVA*
B44	***ED***		**FWYLIMVA**
B58	**ATS**		**FWY***LIVMA*
B62	**QL***IVMP*		**FWYMIVLA**
MOTIFS			
A1	**TSM**		**Y**
A1		**DE***AS*	**Y**
A2.1	**LM***VQIAT*		**V***LIMAT*
A3	**LMVISATF***CGD*		**KYR***HFA*
A11	**VTMLISAGN***CDF*		***KRYH***
A24	**YFW***M*		**FLIW**
A*3101	**MVT***ALIS*		**R***K*
A*3301	**MVALF***IST*		**RK**
A*6801	**AVT***MSLI*		**RK**
B*0702	**P**		**LMF***WYAIV*
B*3501	**P**		**LMFWY***IVA*
B51	**P**		**LIVF***WYAM*
B*5301	**P**		**IMFWY***ALV*
B*5401	**P**		**ATIV***LMFWY*

Bolded residues are preferred, italicized residues are less preferred: A peptide is considered motif-bearing if it has primary anchors at each primary anchor position for a motif or supermotif as specified in the above table.

TABLE IV (B): HLA CLASS II SUPERMOTIF

1	***6***	***9***
W, F, Y, V, .I, L	A, V, I, L, P, C, S, T	A, V, I, L, C, S, T, M, Y

TABLE IV C

IC		1° anchor 1	2	3	4	5	1° anchor 6	7	8	9
DR4	preferred	FMY*LIVW*	M	T		I	VST*CPALIM*	MH		MH
	deleterious				W			R		WDE
DR1	preferred	M*FLIVWY*			PAMQ		VMAT*SPLIC*	M		AVM
	deleterious		C	CH	FD	CWD		GDE	D	
DR7	preferred	M*FLIVWY*	M	W	A		IVMSA*CTPL*	M		IV
	deleterious		C		G			GRD	N	G
DR3	MOTIFS	1° anchor 1	2	3	1° anchor 4	5	1° anchor 6			
notif a referred		LIVMFY			D					
notif b referred		LIVMFAY			DNQEST		KRH			
DR permotif		M*FLIVWY*					VMSTA*CPLI*			

Italicized residues indicate less preferred or "tolerated" residues.

TABLE IV D

		POSITION								
		1	2	3	4	5	6	7	8	C-ter
SUPERMOTIFS										
A1			1° Anchor TI*LVMS*							1° A F
A2			1° Anchor LIVM*ATQ*							1° A LIV
A3	preferred		1° Anchor VSMA*TLI*	YFW(4/5)			YFW (3/5)	YFW (4/5)	P (4/5)	1° A I
	deleterious	DE (3/5); P (5/5)		DE (4/5)						
A24			1° Anchor YF*WIVLMT*							1° A FIY
B7	preferred	FWY (5/5) LIVM (3/5)	1° Anchor P	FWY(4/5)					FWY (3/5)	1°A VILF
	deleterious	DE (3/5); P(5/5); G(4/5); A(3/5); QN(3/5)				DE(3/5)	G(4/5)	QN(4/5)	DE(4/5)	
B27			1° Anchor RHK							1°A FYL

	1° Anchor E*D*	1° A FWY
	1° Anchor ATS	1° A FWY
B62	1° Anchor QL*IVMP*	1° A FWY

TABLE IV E

		1	2	3	4	5	6	7	8	9 or C-terminus	C-t
A1 9-mer	preferred	GFYW	1°Anchor STM	DEA	YFW		P	DEQN	YFW	1°Anchor Y	
	deleterious	DE		RHKLIVMP	A	G	A				
A1 9-mer	preferred	GRHK	ASTCLIVM	1°Anchor DE*AS*	GSTC		ASTC	LIVM	DE	1°Anchor Y	
	deleterious	A	RHKDEPY FW		DE	PQN	RHK	PG	GP		
A1 10-mer	preferred	YFW	1°Anchor STM	DEAQN	A	YFWQN		PASTC	GDE	P	1°/
	deleterious	GP		RHKGLIVM	DE	RHK	QNA	RHKYFW	RHK	A	
A1 10-mer	preferred	YFW	STCLIVM	1°Anchor DE*AS*	A	YFW		PG	G	YFW	1°/
	deleterious	RHK	RHKDEPY FW			P	G		PRHK	QN	
A2.1 9-mer	preferred	YFW	1°Anchor LM*IVQAT*	YFW	STC	YFW		A	P	1°Anchor V*LIMAT*	
	deleterious	DEP		DERKH			RKH	DERKH			

		1	2	3	4	5	6	7	8	9	C-t
A 1C	preferred	AYFW	1°Anchor LM*IVQAT*	LVIM	G		*G*		FYWL VIM		1° V
	deleterious	DEP		DE	RKHA	P		RKH	DERKH	RKH	
A3	preferred	RHK	1°Anchor LMVISATF CGD	YFW	PRHKYFW	A	YFW		P	1°Anchor KYR*HFA*	
	deleterious	DEP		DE							
A11	preferred	A	1°Anchor VTLMISAG N*CDF*	YFW	YFW	A	YFW	YFW	P	1°Anchor K*RYH*	
	deleterious	DEP						A	G		
A24 9-mer	preferred	YFWRHK	1°Anchor YFW*M*		STC			YFW	YFW	1°Anchor FLIW	
	deleterious	DEG		DE	G	QNP	DERHK	G	AQN		
A24 0-mer	preferred		1°Anchor YFW*M*		P	YFWP		P			1° F
	deleterious			GDE	QN	RHK	DE	A	QN	DEA	
A3101	preferred	RHK	1°Anchor MVT*ALIS*	YFW	P		YFW	YFW	AP	1°Anchor R*K*	

		1	2	3	4	5	6	7	8	9	C-te
	deleterious	DEP		DE		ADE	DE	DE	DE		
A0301	preferred		1°Anchor MVALF*IST*	YFW				AYFW		1°Anchor RK	
	deleterious	GP		DE							
A6801	preferred	YFWSTC	1°Anchor AVT*MSLI*			YFWLIVM		YFW	P	1°Anchor RK	
	deleterious	GP		DEG		RHK			A		
B0702	preferred	RHKFWY	1°Anchor P	RHK		RHK	RHK	RHK	PA	1°Anchor LMF*WYAIV*	
	deleterious	DEQNP		DEP	DE	DE	GDE	QN	DE		
B3501	preferred	FWYLIVM	1°Anchor P	FWY				FWY		1°Anchor LMFWY*IV A*	
	deleterious	AGP				G	G				
B51	preferred	LIVMFWY	1°Anchor P	FWY	STC	FWY		G	FWY	1°Anchor LIVF*WYAM*	
	deleterious	AGPDERH KSTC				DE	G	DEQN	GDE		

		1	2	3	4	5	6	7	8	9	C-te
35	preferred	LIVMFWY	1°Anchor P	FWY	STC	FWY		LIVMFWY	FWY	1°Anchor IMFWY*AL V*	
	deleterious	AGPQN					G	RHKQN	DE		
35401	preferred	FWY	1°Anchor P	FWYLIVM		LIVM		ALIVM	FWYAP	1°Anchor ATIV*LMF WY*	
	deleterious	GPQNDE		GDESTC		RHKDE	DE	QNDGE	DE		

Italicized residues indicate less preferred or "tolerated" residues. The information in this Table is specific for 9-mers unless otherwise specified.

TABLE V

HLA Peptide Scoring Results – 85P1B3 – A1, 9-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
1	114	VLEAPFLVG	4.500	1.
2	192	LSEKIAELK	2.700	2.
3	87	LADSVHLAW	2.500	3.
4	27	AIDQASFTT	2.500	4.
5	164	LSSDKMVCY	1.500	5.
6	217	LSEVTPDQS	1.350	6.
7	182	ASEMDIQNV	1.350	7.
8	12	CATPPRGDF	1.000	8.
9	122	GIEGSLKGS	0.900	9.
10	196	IAELKEKIV	0.900	10.
11	141	GIPVGFHLY	0.500	11.
12	100	SLGAVVFSR	0.500	12.
13	184	EMDIQNVPL	0.500	13.
14	57	AEEPAAGPQ	0.450	14.
15	36	SMEWDTQVV	0.450	15.
16	46	GSSPLGPAG	0.300	16.
17	138	GSCGIPVGF	0.300	17.
18	13	ATPPRGDFC	0.250	18.
19	221	TPDQSKPEN	0.250	19.
20	23	GTERAIDQA	0.225	20.
21	61	AAGPQLPSW	0.200	21.
22	120	LVGIEGSLK	0.200	22.
23	169	MVCYLLKTK	0.200	23.
24	203	IVLTHNRLK	0.200	24.
25	56	GAEEPAAGP	0.180	25.
26	130	STYNLLFCG	0.125	26.
27	128	KGSTYNLLF	0.125	27.
28	140	CGIPVGFHL	0.125	28.
29	124	EGSLKGSTY	0.125	29.
30	109	VTNNVVLEA	0.125	30.
31	1	MAAQPLRHR	0.100	31.
32	2	AAQPLRHRS	0.100	32.

TABLE V

HLA Peptide Scoring Results – 85P1B3 – A1, 9-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
33	69	WLQPERCAV	0.100	33.
34	154	ALAALRGHF	0.100	34.
35	165	SSDKMVCYL	0.075	35.
36	31	ASFTTSMEW	0.075	36.
37	129	GSTYNLLFC	0.075	37.
38	149	YSTHAALAA	0.075	38.
39	66	LPSWLQPER	0.050	39.
40	136	FCGSCGIPV	0.050	40.
41	111	NNVVLEAPF	0.050	41.
42	150	STHAALAAL	0.050	42.
43	167	DKMVCYLLK	0.050	43.
44	49	PLGPAGLGA	0.050	44.
45	204	VLTHNRLKS	0.050	45.
46	163	CLSSDKMVC	0.050	46.
47	38	EWDTQVVKG	0.050	47.
48	152	HAALAALRG	0.050	48.
49	179	IVNASEMDI	0.050	49.
50	181	NASEMDIQN	0.050	50.

TABLE VI

HLA Peptide Scoring Results – 85P1B3 – A1, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
1	217	LSEVTPDQSK	27.000	51.
2	36	SMEWDTQVVK	18.000	52.
3	196	IAELKEKIVL	4.500	53.
4	69	WLQPERCAVF	2.000	54.
5	114	VLEAPFLVGI	1.800	55.
6	17	RGDFCGGTER	1.250	56.
7	140	CGIPVGFHLY	1.250	57.
8	13	ATPPRGDFCG	1.250	58.
9		CLSSDKMVCY	1.000	59.

TABLE VI

HLA Peptide Scoring Results – 85P1B3 – A1, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
10	2	AAQPLRHRSR	1.000	60.
11	56	GAEEPAAGPQ	0.900	61.
12	57	AEEPAAGPQL	0.900	62.
13	122	GIEGSLKGST	0.900	63.
14	99	RSLGAVVFSR	0.750	64.
15	27	AIDQASFTTS	0.500	65.
16	90	SVHLAWDLSR	0.500	66.
17	184	EMDIQNVPLS	0.500	67.
18	150	STHAALAALR	0.500	68.
19	46	GSSPLGPAGL	0.300	69.
20	23	GTERAIDQAS	0.225	70.
21	119	FLVGIEGSLK	0.200	71.
22	202	KIVLTHNRLK	0.200	72.
23	186	DIQNVPLSEK	0.200	73.
24	65	QLPSWLQPER	0.200	74.
25	165	SSDKMVCYLL	0.150	75.
26	182	ASEMDIQNVP	0.135	76.
27	94	AWDLSRSLGA	0.125	77.
28	71	QPERCAVFQC	0.113	78.
29	87	LADSVHLAWD	0.100	79.
30	12	CATPPRGDFC	0.100	80.
31	11	RCATPPRGDF	0.100	81.
32	153	AALAALRGHF	0.100	82.
33	61	AAGPQLPSWL	0.100	83.
34	168	KMVCYLLKTK	0.100	84.
35	129	GSTYNLLFCG	0.075	85.
36	192	LSEKIAELKE	0.068	86.
37	116	EAPFLVGIEG	0.050	87.
38	155	LAALRGHFCL	0.050	88.
39	203	IVLTHNRLKS	0.050	89.
40	112	NVVLEAPFLV	0.050	90.
41		SCGIPVGFHL	0.050	91.

TABLE VI

HLA Peptide Scoring Results – 85P1B3 – A1, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
42	178	AIVNASEMDI	0.050	92.
43	26	RAIDQASFTT	0.050	93.
44	159	RGHFCLSSDK	0.050	94.
45	110	TNNVVLEAPF	0.050	95.
46	108	RVTNNVVLEA	0.050	96.
47	30	QASFTTSMEW	0.050	97.
48	113	VVLEAPFLVG	0.050	98.
49	120	LVGIEGSLKG	0.050	99.
50	137	CGSCGIPVGF	0.050	100.

TABLE VII

HLA Peptide Scoring Results – 85P1B3 – A2, 9-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
1	113	VVLEAPFLV	910.291	101.
2	212	SLMKILSEV	591.888	102.
3	172	YLLKTKAIV	485.348	103.
4	69	WLQPERCAV	319.939	104.
5	86	VLADSVHLA	79.642	105.
6	134	LLFCGSCGI	65.622	106.
7	168	KMVCYLLKT	43.325	107.
8	78	FQCAQCHAV	32.438	108.
9	119	FLVGIEGSL	12.775	109.
10	112	NVVLEAPFL	10.281	110.
11	202	KIVLTHNRL	10.281	111.
12	195	KIAELKEKI	10.087	112.
13	162	FCLSSDKMV	7.727	113.
14	85	AVLADSVHL	6.916	114.
15	35	TSMEWDTQV	6.887	115.
16	156	AALRGHFCL	6.367	116.
17	54	GLGAEEPAA	4.968	117.
18		PLSEKIAEL	4.432	118.

TABLE VII

HLA Peptide Scoring Results – 85P1B3 – A2, 9-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
19	33	FTTSMEWDT	3.571	119.
20	93	LAWDLSRSL	3.433	120.
21	115	LEAPFLVGI	3.014	121.
22	27	AIDQASFTT	2.377	122.
23	26	RAIDQASFT	2.334	123.
24	147	HLYSTHAAL	2.324	124.
25	136	FCGSCGIPV	2.088	125.
26	163	CLSSDKMVC	2.037	126.
27	42	QVVKGSSPL	1.869	127.
28	96	DLSRSLGAV	1.560	128.
29	179	IVNASEMDI	1.552	129.
30	101	LGAVVFSRV	1.466	130.
31	36	SMEWDTQVV	1.318	131.
32	205	LTHNRLKSL	1.160	132.
33	140	CGIPVGFHL	0.809	133.
34	62	AGPQLPSWL	0.767	134.
35	126	SLKGSTYNL	0.748	135.
36	165	SSDKMVCYL	0.706	136.
37	209	RLKSLMKIL	0.705	137.
38	150	STHAALAAL	0.682	138.
39	155	LAALRGHFC	0.645	139.
40	197	AELKEKIVL	0.630	140.
41	184	EMDIQNVPL	0.463	141.
42	129	GSTYNLLFC	0.410	142.
43	133	NLLFCGSCG	0.276	143.
44	132	YNLLFCGSC	0.273	144.
45	109	VTNNVVLEA	0.270	145.
46	177	KAIVNASEM	0.242	146.
47	100	SLGAVVFSR	0.199	147.
48	13	ATPPRGDFC	0.186	148.
49	198	ELKEKIVLT	0.184	149.
50		NVPLSEKIA	0.178	150.

TABLE VIII

HLA Peptide Scoring Results – 85P1B3 – A2, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
1	100	SLGAVVFSRV	132.149	151.
2	204	VLTHNRLKSL	83.527	152.
3	195	KIAELKEKIV	56.266	153.
4	133	NLLFCGSCGI	38.601	154.
5	104	VVFSRVTNNV	38.280	155.
6	112	NVVLEAPFLV	35.298	156.
7	154	ALAALRGHFC	27.324	157.
8	211	KSLMKILSEV	13.523	158.
9	164	LSSDKMVCYL	12.295	159.
10	114	VLEAPFLVGI	9.921	160.
11	181	NASEMDIQNV	9.109	161.
12	183	SEMDIQNVPL	6.301	162.
13	212	SLMKILSEVT	5.539	163.
14	78	FQCAQCHAVL	4.085	164.
15	85	AVLADSVHLA	3.699	165.
16	155	LAALRGHFCL	2.925	166.
17	35	TSMEWDTQVV	2.824	167.
18	26	RAIDQASFTT	2.461	168.
19	41	TQVVKGSSPL	2.166	169.
20	96	DLSRSLGAVV	2.139	170.
21	190	VPLSEKIAEL	2.017	171.
22	61	AAGPQLPSWL	1.632	172.
23	76	AVFQCAQCHA	1.608	173.
24	149	YSTHAALAAL	1.475	174.
25	128	KGSTYNLLFC	1.436	175.
26	178	AIVNASEMDI	1.435	176.
27	197	AELKEKIVLT	1.233	177.
28	126	SLKGSTYNLL	1.122	178.
29	108	RVTNNVVLEA	1.000	179.
30	34	TTSMEWDTQV	0.966	180.
31		IQNVPLSEKI	0.881	181.

TABLE VIII

HLA Peptide Scoring Results – 85P1B3 – A2, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
32	139	SCGIPVGFHL	0.809	182.
33	111	NNVVLEAPFL	0.767	183.
34	169	MVCYLLKTKA	0.739	184.
35	173	LLKTKAIVNA	0.680	185.
36	147	HLYSTHAALA	0.541	186.
37	172	YLLKTKAIVN	0.520	187.
38	86	VLADSVHLAW	0.519	188.
39	125	GSLKGSTYNL	0.516	189.
40	3	AQPLRHRSRC	0.504	190.
41	95	WDLSRSLGAV	0.492	191.
42	170	VCYLLKTKAI	0.370	192.
43	216	ILSEVTPDQS	0.255	193.
44	146	FHLYSTHAAL	0.252	194.
45	162	FCLSSDKMVC	0.226	195.
46	88	ADSVHLAWDL	0.223	196.
47	82	QCHAVLADSV	0.222	197.
48	141	GIPVGFHLYS	0.214	198.
49	144	VGFHLYSTHA	0.204	199.
50	37	MEWDTQVVKG	0.193	200.

TABLE IX

HLA Peptide Scoring Results - 85P1B3 - A3, 9-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
1	100	SLGAVVFSR	54.000	201.
2	168	KMVCYLLKT	4.050	202.
3	141	GIPVGFHLY	3.600	203.
4	147	HLYSTHAAL	3.000	204.
5	134	LLFCGSCGI	3.000	205.
6	126	SLKGSTYNL	2.700	206.
7	120	LVGIEGSLK	2.000	207.
8		MVCYLLKTK	1.500	208.

TABLE IX

HLA Peptide Scoring Results - 85P1B3 - A3, 9-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
9	187	IQNVPLSEK	1.350	209.
10	212	SLMKILSEV	0.675	210.
11	119	FLVGIEGSL	0.608	211.
12	54	GLGAEEPAA	0.600	212.
13	154	ALAALRGHF	0.600	213.
14	86	VLADSVHLA	0.600	214.
15	209	RLKSLMKIL	0.450	215.
16	163	CLSSDKMVC	0.400	216.
17	160	GHFCLSSDK	0.300	217.
18	69	WLQPERCAV	0.300	218.
19	172	YLLKTKAIV	0.300	219.
20	37	MEWDTQVVK	0.300	220.
21	203	IVLTHNRLK	0.300	221.
22	202	KIVLTHNRL	0.270	222.
23	195	KIAELKEKI	0.270	223.
24	36	SMEWDTQVV	0.200	224.
25	184	EMDIQNVPL	0.180	225.
26	157	ALRGHFCLS	0.180	226.
27	114	VLEAPFLVG	0.180	227.
28	192	LSEKIAELK	0.150	228.
29	191	PLSEKIAEL	0.135	229.
30	113	VVLEAPFLV	0.135	230.
31	218	SEVTPDQSK	0.135	231.
32	179	IVNASEMDI	0.120	232.
33	76	AVFQCAQCH	0.100	233.
34	213	LMKILSEVT	0.100	234.
35	70	LQPERCAVF	0.090	235.
36	65	QLPSWLQPE	0.090	236.
37	85	AVLADSVHL	0.090	237.
38	42	QVVKGSSPL	0.090	238.
39	112	NVVLEAPFL	0.090	239.
40		VTNNVVLEA	0.090	240.

TABLE IX

HLA Peptide Scoring Results - 85P1B3 - A3, 9-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
41	207	HNRLKSLMK	0.080	241.
42	204	VLTHNRLKS	0.080	242.
43	138	GSCGIPVGF	0.068	243.
44	198	ELKEKIVLT	0.068	244.
45	18	GDFCGGTER	0.060	245.
46	92	HLAWDLSRS	0.060	246.
47	49	PLGPAGLGA	0.060	247.
48	216	ILSEVTPDQ	0.045	248.
49	23	GTERAIDQA	0.045	249.
50	150	STHAALAAL	0.045	250.

TABLE X

HLA Peptide Scoring Results – 85P1B3 – A3, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
1	168	KMVCYLLKTK	67.500	251.
2	119	FLVGIEGSLK	45.000	252.
3	36	SMEWDTQVVK	20.000	253.
4	163	CLSSDKMVCY	6.000	254.
5	191	PLSEKIAELK	4.500	255.
6	65	QLPSWLQPER	4.000	256.
7	69	WLQPERCAVF	3.000	257.
8	114	VLEAPFLVGI	2.700	258.
9	90	SVHLAWDLSR	2.400	259.
10	186	DIQNVPLSEK	1.350	260.
11	147	HLYSTHAALA	1.000	261.
12	100	SLGAVVFSRV	0.900	262.
13	202	KIVLTHNRLK	0.900	263.
14	126	SLKGSTYNLL	0.900	264.
15	133	NLLFCGSCGI	0.900	265.
16	198	ELKEKIVLTH	0.810	266.
17		RSLGAVVFSR	0.608	267.

TABLE X

HLA Peptide Scoring Results – 85P1B3 – A3, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
18	86	VLADSVHLAW	0.600	268.
19	204	VLTHNRLKSL	0.450	269.
20	157	ALRGHFCLSS	0.360	270.
21	173	LLKTKAIVNA	0.300	271.
22	154	ALAALRGHFC	0.200	272.
23	150	STHAALAALR	0.200	273.
24	108	RVTNNVVLEA	0.180	274.
25	178	AIVNASEMDI	0.180	275.
26	104	VVFSRVTNNV	0.150	276.
27	212	SLMKILSEVT	0.150	277.
28	217	LSEVTPDQSK	0.150	278.
29	76	AVFQCAQCHA	0.100	279.
30	85	AVLADSVHLA	0.090	280.
31	112	NVVLEAPFLV	0.090	281.
32	209	RLKSLMKILS	0.080	282.
33	141	GIPVGFHLYS	0.072	283.
34	172	YLLKTKAIVN	0.060	284.
35	96	DLSRSLGAVV	0.060	285.
36	54	GLGAEEPAAG	0.060	286.
37	92	HLAWDLSRSL	0.060	287.
38	216	ILSEVTPDQS	0.060	288.
39	195	KIAELKEKIV	0.045	289.
40	193	SEKIAELKEK	0.045	290.
41	113	VVLEAPFLVG	0.041	291.
42	125	GSLKGSTYNL	0.041	292.
43	166	SDKMVCYLLK	0.040	293.
44	206	THNRLKSLMK	0.040	294.
45	184	EMDIQNVPLS	0.036	295.
46	200	KEKIVLTHNR	0.036	296.
47	134	LLFCGSCGIP	0.030	297.
48	34	TTSMEWDTQV	0.030	298.
49		STYNLLFCGS	0.030	299.

TABLE X

HLA Peptide Scoring Results – 85P1B3 – A3, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
50	41	TQVVKGSSPL	0.027	300.

TABLE XI

HLA Peptide Scoring Results – 85P1B3 – A11, 9-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
1	120	LVGIEGSLK	2.000	301.
2	169	MVCYLLKTK	1.000	302.
3	187	IQNVPLSEK	0.600	303.
4	203	IVLTHNRLK	0.300	304.
5	100	SLGAVVFSR	0.240	305.
6	160	GHFCLSSDK	0.120	306.
7	37	MEWDTQVVK	0.120	307.
8	113	VVLEAPFLV	0.090	308.
9	218	SEVTPDQSK	0.090	309.
10	207	HNRLKSLMK	0.080	310.
11	76	AVFQCAQCH	0.040	311.
12	66	LPSWLQPER	0.040	312.
13	179	IVNASEMDI	0.040	313.
14	85	AVLADSVHL	0.030	314.
15	112	NVVLEAPFL	0.030	315.
16	42	QVVKGSSPL	0.030	316.
17	23	GTERAIDQA	0.030	317.
18	18	GDFCGGTER	0.024	318.
19	167	DKMVCYLLK	0.024	319.
20	109	VTNNVVLEA	0.020	320.
21	192	LSEKIAELK	0.020	321.
22	202	KIVLTHNRL	0.018	322.
23	9	RSRCATPPR	0.012	323.
24	91	VHLAWDLSR	0.012	324.
25	3	AQPLRHRSR	0.012	325.
26		GLGAEEPAA	0.012	326.

TABLE XI

HLA Peptide Scoring Results – 85P1B3 – A11, 9-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
27	141	GIPVGFHLY	0.012	327.
28	195	KIAELKEKI	0.012	328.
29	189	NVPLSEKIA	0.010	329.
30	150	STHAALAAL	0.010	330.
31	156	AALRGHFCL	0.009	331.
32	177	KAIVNASEM	0.009	332.
33	212	SLMKILSEV	0.008	333.
34	126	SLKGSTYNL	0.008	334.
35	147	HLYSTHAAL	0.008	335.
36	134	LLFCGSCGI	0.008	336.
37	172	YLLKTKAIV	0.006	337.
38	145	GFHLYSTHA	0.006	338.
39	70	LQPERCAVF	0.006	339.
40	130	STYNLLFCG	0.006	340.
41	108	RVTNNVVLE	0.006	341.
42	78	FQCAQCHAV	0.006	342.
43	209	RLKSLMKIL	0.006	343.
44	119	FLVGIEGSL	0.006	344.
45	205	LTHNRLKSL	0.005	345.
46	194	EKIAELKEK	0.005	346.
47	87	LADSVHLAW	0.004	347.
48	1	MAAQPLRHR	0.004	348.
49	154	ALAALRGHF	0.004	349.
50	151	THAALAALR	0.004	350.

TABLE XII

HLA Peptide Scoring Results – 85P1B3 – A11, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
1	168	KMVCYLLKTK	0.900	351.
2	90	SVHLAWDLSR	0.800	352.
3		FLVGIEGSLK	0.600	353.

TABLE XII

HLA Peptide Scoring Results – 85P1B3 – A11, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
4	36	SMEWDTQVVK	0.400	354.
5	150	STHAALAALR	0.200	355.
6	202	KIVLTHNRLK	0.180	356.
7	186	DIQNVPLSEK	0.120	357.
8	108	RVTNNVVLEA	0.120	358.
9	112	NVVLEAPFLV	0.090	359.
10	65	QLPSWLQPER	0.080	360.
11	159	RGHFCLSSDK	0.060	361.
12	99	RSLGAVVFSR	0.054	362.
13	166	SDKMVCYLLK	0.040	363.
14	76	AVFQCAQCHA	0.040	364.
15	191	PLSEKIAELK	0.040	365.
16	104	VVFSRVTNNV	0.040	366.
17	206	THNRLKSLMK	0.040	367.
18	200	KEKIVLTHNR	0.036	368.
19	85	AVLADSVHLA	0.030	369.
20	193	SEKIAELKEK	0.030	370.
21	217	LSEVTPDQSK	0.020	371.
22	169	MVCYLLKTKA	0.020	372.
23	17	RGDFCGGTER	0.012	373.
24	178	AIVNASEMDI	0.012	374.
25	205	LTHNRLKSLM	0.010	375.
26	34	TTSMEWDTQV	0.010	376.
27	41	TQVVKGSSPL	0.009	377.
28	86	VLADSVHLAW	0.008	378.
29	147	HLYSTHAALA	0.008	379.
30	148	LYSTHAALAA	0.008	380.
31	133	NLLFCGSCGI	0.006	381.
32	203	IVLTHNRLKS	0.006	382.
33	78	FQCAQCHAVL	0.006	383.
34	187	IQNVPLSEKI	0.006	384.
35		GFHLYSTHAA	0.006	385.

TABLE XII

HLA Peptide Scoring Results – 85P1B3 – A11, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
36	48	SPLGPAGLGA	0.006	386.
37	155	LAALRGHFCL	0.006	387.
38	11	RCATPPRGDF	0.006	388.
39	113	VVLEAPFLVG	0.006	389.
40	139	SCGIPVGFHL	0.006	390.
41	195	KIAELKEKIV	0.006	391.
42	171	CYLLKTKAIV	0.006	392.
43	43	VVKGSSPLGP	0.004	393.
44	163	CLSSDKMVCY	0.004	394.
45	100	SLGAVVFSRV	0.004	395.
46	135	LFCGSCGIPV	0.004	396.
47	196	IAELKEKIVL	0.004	397.
48	8	HRSRCATPPR	0.004	398.
49	189	NVPLSEKIAE	0.004	399.
50	126	SLKGSTYNLL	0.004	400.

TABLE XIII

HLA Peptide Scoring Results – 85P1B3 – A24, 9-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
1	171	CYLLKTKAI	75.000	401.
2	202	KIVLTHNRL	14.400	402.
3	131	TYNLLFCGS	10.800	403.
4	140	CGIPVGFHL	10.080	404.
5	209	RLKSLMKIL	9.600	405.
6	119	FLVGIEGSL	8.400	406.
7	62	AGPQLPSWL	7.200	407.
8	89	DSVHLAWDL	7.200	408.
9	112	NVVLEAPFL	6.000	409.
10	42	QVVKGSSPL	6.000	410.
11	47	SSPLGPAGL	6.000	411.
12		AVLADSVHL	6.000	412.

TABLE XIII

HLA Peptide Scoring Results – 85P1B3 – A24, 9-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
13	156	AALRGHFCL	6.000	413.
14	93	LAWDLSRSL	5.760	414.
15	148	LYSTHAALA	5.000	415.
16	126	SLKGSTYNL	4.000	416.
17	184	EMDIQNVPL	4.000	417.
18	147	HLYSTHAAL	4.000	418.
19	79	QCAQCHAVL	4.000	419.
20	165	SSDKMVCYL	4.000	420.
21	205	LTHNRLKSL	4.000	421.
22	150	STHAALAAL	4.000	422.
23	128	KGSTYNLLF	4.000	423.
24	70	LQPERCAVF	3.600	424.
25	111	NNVVLEAPF	3.600	425.
26	195	KIAELKEKI	3.168	426.
27	138	GSCGIPVGF	2.800	427.
28	161	HFCLSSDKM	2.750	428.
29	12	CATPPRGDF	2.400	429.
30	154	ALAALRGHF	2.400	430.
31	188	QNVPLSEKI	2.376	431.
32	177	KAIVNASEM	1.650	432.
33	179	IVNASEMDI	1.500	433.
34	134	LLFCGSCGI	1.000	434.
35	20	FCGGTERAI	1.000	435.
36	105	VFSRVTNNV	0.840	436.
37	77	VFQCAQCHA	0.750	437.
38	197	AELKEKIVL	0.600	438.
39	107	SRVTNNVVL	0.600	439.
40	58	EEPAAGPQL	0.600	440.
41	166	SDKMVCYLL	0.560	441.
42	191	PLSEKIAEL	0.528	442.
43	29	DQASFTTSM	0.500	443.
44		GFHLYSTHA	0.500	444.

TABLE XIII

HLA Peptide Scoring Results – 85P1B3 – A24, 9-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
45	19	DFCGGTERA	0.500	445.
46	127	LKGSTYNLL	0.480	446.
47	26	RAIDQASFT	0.360	447.
48	175	KTKAIVNAS	0.336	448.
49	168	KMVCYLLKT	0.330	449.
50	99	RSLGAVVFS	0.300	450.

TABLE XIV

HLA Peptide Scoring Results – 85P1B3 – A24, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
1	171	CYLLKTKAIV	7.500	451.
2	131	TYNLLFCGSC	7.500	452.
3	190	VPLSEKIAEL	6.600	453.
4	196	IAELKEKIVL	6.000	454.
5	41	TQVVKGSSPL	6.000	455.
6	125	GSLKGSTYNL	6.000	456.
7	111	NNVVLEAPFL	6.000	457.
8	84	HAVLADSVHL	6.000	458.
9	61	AAGPQLPSWL	5.760	459.
10	139	SCGIPVGFHL	5.600	460.
11	165	SSDKMVCYLL	5.600	461.
12	148	LYSTHAALAA	5.000	462.
13	19	DFCGGTERAI	5.000	463.
14	46	GSSPLGPAGL	4.800	464.
15	92	HLAWDLSRSL	4.800	465.
16	126	SLKGSTYNLL	4.800	466.
17	164	LSSDKMVCYL	4.800	467.
18	118	PFLVGIEGSL	4.200	468.
19	155	LAALRGHFCL	4.000	469.
20	11	RCATPPRGDF	4.000	470.
21		FQCAQCHAVL	4.000	471.

TABLE XIV

HLA Peptide Scoring Results – 85P1B3 – A24, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
22	149	YSTHAALAAL	4.000	472.
23	106	FSRVTNNVVL	4.000	473.
24	204	VLTHNRLKSL	4.000	474.
25	110	TNNVVLEAPF	3.600	475.
26	69	WLQPERCAVF	3.600	476.
27	153	AALAALRGHF	3.600	477.
28	137	CGSCGIPVGF	2.800	478.
29	97	LSRSLGAVVF	2.000	479.
30	187	IQNVPLSEKI	1.980	480.
31	178	AIVNASEMDI	1.500	481.
32	114	VLEAPFLVGI	1.500	482.
33	133	NLLFCGSCGI	1.500	483.
34	207	HNRLKSLMKI	1.100	484.
35	170	VCYLLKTKAI	1.000	485.
36	77	VFQCAQCHAV	0.750	486.
37	183	SEMDIQNVPL	0.720	487.
38	208	NRLKSLMKIL	0.720	488.
39	201	EKIVLTHNRL	0.720	489.
40	57	AEEPAAGPQL	0.720	490.
41	146	FHLYSTHAAL	0.600	491.
42	105	VFSRVTNNVV	0.600	492.
43	205	LTHNRLKSLM	0.600	493.
44	135	LFCGSCGIPV	0.500	494.
45	161	HFCLSSDKMV	0.500	495.
46	145	GFHLYSTHAA	0.500	496.
47	32	SFTTSMEWDT	0.500	497.
48	88	ADSVHLAWDL	0.480	498.
49	211	KSLMKILSEV	0.462	499.
50	26	RAIDQASFTT	0.360	500.

TABLE XV

HLA Peptide Scoring Results – 85P1B3 – B7, 9-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
1	85	AVLADSVHL	60.000	501.
2	156	AALRGHFCL	36.000	502.
3	112	NVVLEAPFL	20.000	503.
4	42	QVVKGSSPL	20.000	504.
5	62	AGPQLPSWL	12.000	505.
6	93	LAWDLSRSL	12.000	506.
7	150	STHAALAAL	4.000	507.
8	89	DSVHLAWDL	4.000	508.
9	202	KIVLTHNRL	4.000	509.
10	147	HLYSTHAAL	4.000	510.
11	126	SLKGSTYNL	4.000	511.
12	79	QCAQCHAVL	4.000	512.
13	205	LTHNRLKSL	4.000	513.
14	140	CGIPVGFHL	4.000	514.
15	119	FLVGIEGSL	4.000	515.
16	47	SSPLGPAGL	4.000	516.
17	209	RLKSLMKIL	4.000	517.
18	177	KAIVNASEM	3.000	518.
19	4	QPLRHRSRC	3.000	519.
20	179	IVNASEMDI	2.000	520.
21	97	LSRSLGAVV	2.000	521.
22	106	FSRVTNNVV	2.000	522.
23	184	EMDIQNVPL	1.200	523.
24	165	SSDKMVCYL	1.200	524.
25	197	AELKEKIVL	1.200	525.
26	113	VVLEAPFLV	1.000	526.
27	29	DQASFTTSM	1.000	527.
28	35	TSMEWDTQV	0.600	528.
29	117	APFLVGIEG	0.600	529.
30	212	SLMKILSEV	0.600	530.
31	157	ALRGHFCLS	0.600	531.
32		NVPLSEKIA	0.500	532.

TABLE XV

HLA Peptide Scoring Results – 85P1B3 – B7, 9-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
33	103	AVVFSRVTN	0.450	533.
34	20	FCGGTERAI	0.400	534.
35	134	LLFCGSCGI	0.400	535.
36	107	SRVTNNVVL	0.400	536.
37	188	QNVPLSEKI	0.400	537.
38	127	LKGSTYNLL	0.400	538.
39	191	PLSEKIAEL	0.400	539.
40	166	SDKMVCYLL	0.400	540.
41	58	EEPAAGPQL	0.400	541.
42	142	IPVGFHLYS	0.400	542.
43	195	KIAELKEKI	0.400	543.
44	75	CAVFQCAQC	0.300	544.
45	155	LAALRGHFC	0.300	545.
46	48	SPLGPAGLG	0.300	546.
47	53	AGLGAEEPA	0.300	547.
48	69	WLQPERCAV	0.300	548.
49	80	CAQCHAVLA	0.300	549.
50	26	RAIDQASFT	0.300	550.

TABLE XVI

HLA Peptide Scoring Results – 85P1B3 – B7, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
1	190	VPLSEKIAEL	80.000	551.
2	106	FSRVTNNVVL	40.000	552.
3	61	AAGPQLPSWL	36.000	553.
4	84	HAVLADSVHL	12.000	554.
5	155	LAALRGHFCL	12.000	555.
6	46	GSSPLGPAGL	4.000	556.
7	111	NNVVLEAPFL	4.000	557.
8	92	HLAWDLSRSL	4.000	558.
9		FQCAQCHAVL	4.000	559.

TABLE XVI

HLA Peptide Scoring Results – 85P1B3 – B7, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
10	204	VLTHNRLKSL	4.000	560.
11	139	SCGIPVGFHL	4.000	561.
12	149	YSTHAALAAL	4.000	562.
13	126	SLKGSTYNLL	4.000	563.
14	164	LSSDKMVCYL	4.000	564.
15	41	TQVVKGSSPL	4.000	565.
16	207	HNRLKSLMKI	4.000	566.
17	125	GSLKGSTYNL	4.000	567.
18	196	IAELKEKIVL	3.600	568.
19	15	PPRGDFCGGT	2.000	569.
20	48	SPLGPAGLGA	2.000	570.
21	4	QPLRHRSRCA	2.000	571.
22	142	IPVGFHLYST	2.000	572.
23	66	LPSWLQPERC	2.000	573.
24	85	AVLADSVHLA	1.500	574.
25	76	AVFQCAQCHA	1.500	575.
26	165	SSDKMVCYLL	1.200	576.
27	117	APFLVGIEGS	1.200	577.
28	88	ADSVHLAWDL	1.200	578.
29	183	SEMDIQNVPL	1.200	579.
30	178	AIVNASEMDI	1.200	580.
31	112	NVVLEAPFLV	1.000	581.
32	205	LTHNRLKSLM	1.000	582.
33	104	VVFSRVTNNV	1.000	583.
34	71	QPERCAVFQC	0.600	584.
35	157	ALRGHFCLSS	0.600	585.
36	181	NASEMDIQNV	0.600	586.
37	35	TSMEWDTQVV	0.600	587.
38	59	EPAAGPQLPS	0.600	588.
39	169	MVCYLLKTKA	0.500	589.
40	108	RVTNNVVLEA	0.500	590.
41		AQPLRHRSRC	0.450	591.

TABLE XVI

HLA Peptide Scoring Results – 85P1B3 – B7, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
42	201	EKIVLTHNRL	0.400	592.
43	170	VCYLLKTKAI	0.400	593.
44	146	FHLYSTHAAL	0.400	594.
45	133	NLLFCGSCGI	0.400	595.
46	187	IQNVPLSEKI	0.400	596.
47	208	NRLKSLMKIL	0.400	597.
48	57	AEEPAAGPQL	0.360	598.
49	53	AGLGAEEPAA	0.300	599.
50	154	ALAALRGHFC	0.300	600.

TABLE XVII

HLA Peptide Scoring Results – 85P1B3 – B35, 9-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
1	164	LSSDKMVCY	20.000	601.
2	177	KAIVNASEM	12.000	602.
3	209	RLKSLMKIL	6.000	603.
4	93	LAWDLSRSL	6.000	604.
5	89	DSVHLAWDL	5.000	605.
6	138	GSCGIPVGF	5.000	606.
7	47	SSPLGPAGL	5.000	607.
8	126	SLKGSTYNL	3.000	608.
9	12	CATPPRGDF	3.000	609.
10	156	AALRGHFCL	3.000	610.
11	106	FSRVTNNVV	3.000	611.
12	97	LSRSLGAVV	3.000	612.
13	31	ASFTTSMEW	2.500	613.
14	141	GIPVGFHLY	2.000	614.
15	128	KGSTYNLLF	2.000	615.
16	142	IPVGFHLYS	2.000	616.
17	35	TSMEWDTQV	2.000	617.
18		QPLRHRSRC	2.000	618.

TABLE XVII

HLA Peptide Scoring Results – 85P1B3 – B35, 9-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
19	70	LQPERCAVF	2.000	619.
20	202	KIVLTHNRL	2.000	620.
21	29	DQASFTTSM	2.000	621.
22	124	EGSLKGSTY	2.000	622.
23	195	KIAELKEKI	1.600	623.
24	165	SSDKMVCYL	1.500	624.
25	85	AVLADSVHL	1.500	625.
26	112	NVVLEAPFL	1.500	626.
27	61	AAGPQLPSW	1.500	627.
28	26	RAIDQASFT	1.200	628.
29	62	AGPQLPSWL	1.000	629.
30	119	FLVGIEGSL	1.000	630.
31	140	CGIPVGFHL	1.000	631.
32	154	ALAALRGHF	1.000	632.
33	42	QVVKGSSPL	1.000	633.
34	150	STHAALAAL	1.000	634.
35	205	LTHNRLKSL	1.000	635.
36	99	RSLGAVVFS	1.000	636.
37	79	QCAQCHAVL	1.000	637.
38	147	HLYSTHAAL	1.000	638.
39	111	NNVVLEAPF	1.000	639.
40	221	TPDQSKPEN	0.600	640.
41	198	ELKEKIVLT	0.600	641.
42	181	NASEMDIQN	0.600	642.
43	175	KTKAIVNAS	0.600	643.
44	125	GSLKGSTYN	0.500	644.
45	129	GSTYNLLFC	0.500	645.
46	149	YSTHAALAA	0.500	646.
47	182	ASEMDIQNV	0.450	647.
48	87	LADSVHLAW	0.450	648.
49	113	VVLEAPFLV	0.400	649.
50		FCGGTERAI	0.400	650.

TABLE XVIII

HLA Peptide Scoring Results – 85P1B3 – B35, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
1	190	VPLSEKIAEL	20.000	651.
2	106	FSRVTNNVVL	15.000	652.
3	97	LSRSLGAVVF	15.000	653.
4	164	LSSDKMVCYL	10.000	654.
5	46	GSSPLGPAGL	5.000	655.
6	149	YSTHAALAAL	5.000	656.
7	125	GSLKGSTYNL	5.000	657.
8	84	HAVLADSVHL	4.500	658.
9	153	AALAALRGHF	3.000	659.
10	61	AAGPQLPSWL	3.000	660.
11	35	TSMEWDTQVV	3.000	661.
12	155	LAALRGHFCL	3.000	662.
13	126	SLKGSTYNLL	3.000	663.
14	117	APFLVGIEGS	2.000	664.
15	140	CGIPVGFHLY	2.000	665.
16	59	EPAAGPQLPS	2.000	666.
17	205	LTHNRLKSLM	2.000	667.
18	4	QPLRHRSRCA	2.000	668.
19	66	LPSWLQPERC	2.000	669.
20	142	IPVGFHLYST	2.000	670.
21	163	CLSSDKMVCY	2.000	671.
22	211	KSLMKILSEV	2.000	672.
23	11	RCATPPRGDF	2.000	673.
24	48	SPLGPAGLGA	2.000	674.
25	181	NASEMDIQNV	1.800	675.
26	30	QASFTTSMEW	1.500	676.
27	165	SSDKMVCYLL	1.500	677.
28	111	NNVVLEAPFL	1.500	678.
29	196	IAELKEKIVL	1.350	679.
30	26	RAIDQASFTT	1.200	680.
31		HNRLKSLMKI	1.200	681.

TABLE XVIII

HLA Peptide Scoring Results – 85P1B3 – B35, 10-mers

Rank	Start Position	Subsequence Residue Listing	Score (Estimate of Half Time of Disassociation of a Molecule Containing This Subsequence)	Seq.ID#
32	204	VLTHNRLKSL	1.000	682.
33	137	CGSCGIPVGF	1.000	683.
34	86	VLADSVHLAW	1.000	684.
35	41	TQVVKGSSPL	1.000	685.
36	139	SCGIPVGFHL	1.000	686.
37	92	HLAWDLSRSL	1.000	687.
38	69	WLQPERCAVF	1.000	688.
39	78	FQCAQCHAVL	1.000	689.
40	110	TNNVVLEAPF	1.000	690.
41	195	KIAELKEKIV	0.800	691.
42	209	RLKSLMKILS	0.600	692.
43	15	PPRGDFCGGT	0.600	693.
44	71	QPERCAVFQC	0.600	694.
45	89	DSVHLAWDLS	0.500	695.
46	24	TERAIDQASF	0.450	696.
47	170	VCYLLKTKAI	0.400	697.
48	178	AIVNASEMDI	0.400	698.
49	133	NLLFCGSCGI	0.400	699.
50	187	IQNVPLSEKI	0.400	700.

Table XIX: Motifs and Post-translational modifications

N-glycosylation site
181-184 NASE

Protein kinase C phosphorylation site
Number of matches: 4
1 24-26 TER
2 126-128 SLK
3 166-168 SDK
4 193-195 SEK

Casein kinase II phosphorylation site
Number of matches: 3
1 35-38 TSME
2 183-186 SEMD
3 225-228 SKPE

N-myristoylation site
Number of matches: 5
1 23-28 GTERAI
2 122-127 GIEGSL
3 125-130 GSLKGS
4 129-134 GSTYNL
5 141-146 GIPVGF

RGD Cell attachment sequence
17-19 RGD

Cytochrome c family heme-binding site signature
80-85 CAQCHA

Table XX: Frequently Occurring Motifs			
Name	avrg. % identity	Description	Potential Function
zf-C2H2	34%	Zinc finger, C2H2 type	Nucleic acid-binding protein functions as transcription factor, nuclear location probable
cytochrome_b_N	68%	Cytochrome b(N-terminal)/b6/petB	membrane bound oxidase, generate superoxide
ig	19%	Immunoglobulin domain	domains are one hundred amino acids long and include a conserved intradomain disulfide bond.
WD40	18%	WD domain, G-beta repeat	tandem repeats of about 40 residues, each containing a Trp-Asp motif. Function in signal transduction and protein interaction
PDZ	23%	PDZ domain	may function in targeting signaling molecules to sub-membranous sites
LRR	28%	Leucine Rich Repeat	short sequence motifs involved in protein-protein interactions
pkinase	23%	Protein kinase domain	conserved catalytic core common to both serine/threonine and tyrosine protein kinases containing an ATP binding site and a catalytic site
PH	16%	PH domain	pleckstrin homology involved in intracellular signaling or as constituents of the cytoskeleton
EGF	34%	EGF-like domain	30-40 amino-acid long found in the extracellular domain of membrane-bound proteins or in secreted proteins
rvt	49%	Reverse transcriptase (RNA-dependent DNA polymerase)	
ank	25%	Ank repeat	Cytoplasmic protein, associates integral membrane proteins to the cytoskeleton

oxidored_q1	32%	NADH-Ubiquinone/plastoquinone (complex I), various chains	membrane associated. Involved in proton translocation across the membrane
efhand	24%	EF hand	calcium-binding domain, consists of a12 residue loop flanked on both sides by a 12 residue alpha-helical domain
rvp	79%	Retroviral aspartyl protease	Aspartyl or acid proteases, centered on a catalytic aspartyl residue
Collagen	42%	Collagen triple helix repeat (20 copies)	extracellular structural proteins involved in formation of connective tissue. The sequence consists of the G-X-Y and the polypeptide chains forms a triple helix.
fn3	20%	Fibronectin type III domain	Located in the extracellular ligand-binding region of receptors and is about 200 amino acid residues long with two pairs of cysteines involved in disulfide bonds
7tm_1	19%	7 transmembrane receptor (rhodopsin family)	seven hydrophobic transmembrane regions, with the N-terminus located extracellularly while the C-terminus is cytoplasmic. Signal through G proteins

TABLE XXV: Protein Properties

	Bioinformatic Program	URL	Outcome
ORF	ORF Finder	http://www.ncbi.nlm.gov/gorf	13-702 (includes stop)
Protein Length			229 amino acids
Transmembrane region	TM Pred	http://www.ch.embnet.org/	one TM at aa 129-149
	HMMTop	http://www.enzim.hu/hmmtop/	one TM at aa 134-158
	Sosui	http://www.genome.ad.jp/SOSui/	indicates no TM, soluble protein
	TMHMM	http://www.cbs.dtu.dk/services/TMHMM	indicates no TM
Signal Peptide	Signal P	http://www.cbs.dtu.dk/services/SignalP/	indicates no signal
pI	pI/MW tool	http://www.expasy.ch/tools/	pI 7.02
Molecular weight	pI/MW tool	http://www.expasy.ch/tools/	24.69 kDa
Localization	PSORT	http://psort.nibb.ac.jp/	Cytoplasmic 65 % Mitochondrial 10%
	PSORT II	http://psort.nibb.ac.jp/	Mitochondrial 60.9% Cytoplamic 21.7%
Motifs	Pfam	http://www.sanger.ac.uk/Pfam/	no motif detected
	Prints	http://www.biochem.ucl.ac.uk/	no significant motif
	Blocks	http://www.blocks.fhcrc.org/	Soybean trypsin inhibitor protease family, Cytochrome c
	Prosite	http://www.genome.ad.jp/	Cytochrome c family, heme binding signature

CLAIMS:

1. A method for monitoring 85P1B3 gene products in a biological sample from a patient who has or who is suspected of having cancer, the method comprising:

determining the status of 85P1B3 gene products expressed by cells in a tissue sample from an individual;

comparing the status so determined to the status of 85P1B3 gene products in a corresponding normal sample; and,

identifying the presence of aberrant 85P1B3 gene products in the sample relative to the normal sample.

2. A method of monitoring the presence of cancer in an individual comprising: performing the method of claim 1 whereby the presence of elevated 85P1B3 mRNA or protein expression in the test sample relative to the normal tissue sample indicates the presence or status of a cancer.

3. The method of claim 2, wherein the cancer occurs in a tissue set forth in Table I.

4. A composition comprising:

a substance that modulates the status of 85P1B3, or a molecule that is modulated by 85P1B3 whereby the status of a cell that expresses 85P1B3 is modulated.

5. The composition of claim 4, further comprising a pharmaceutically acceptable carrier.

6. A pharmaceutical composition that comprises the composition of claim 4 in a human unit dose form.

7. A composition of claim 4 wherein the substance comprises a 85P1B3-related protein.

8. The composition of claim 7, further comprising antigen presenting cells.

9. The composition of claim 7 comprising an analog peptide of eight, nine ten or eleven contiguous amino acids of Figure 2 (SEQ ID NO.: ____).

10. A composition of claim 7 comprising a CTL polypeptide epitope from Figure 2 (SEQ ID NO.: ____).

11. The composition of claim 10, wherein the CTL epitope comprises a polypeptide selected from

12. A composition comprising a peptide region of at least 5 amino acids of Figure 2 (SEQ ID NO.: ___) in any whole number increment up to 229 that includes an amino acid position selected from: an amino acid position having a value greater than 0.5 in the Hydrophilicity profile of Figure 5, an amino acid position having a value less than 0.5 in the Hydropathicity profile of Figure 6; an amino acid position having a value greater than 0.5 in the Percent Accessible Residues profile of Figure 7; an amino acid position having a value greater than 0.5 in the Average Flexibility profile on Figure 8; or an amino acid position having a value greater than 0.5 in the Beta-turn profile of Figure 9.

13. A polynucleotide that encodes an analog peptide of claim 9.

14. A composition of claim 4 wherein the substance comprises an antibody or fragment thereof that specifically binds to a 85P1B3-related protein.

15. The antibody or fragment thereof of claim 14, which is monoclonal.

16. A recombinant protein comprising the antigen-binding region of a monoclonal antibody of claim 15.

17. The antibody or fragment thereof of claim 14, which is labeled with a detectable marker.

18. The recombinant protein of claim 16, which is labeled with a detectable marker.

19. The antibody fragment of an antibody of claim 14, which is an Fab, F(ab')2, Fv or sFv fragment.

20. The antibody of claim 14, which is a human antibody.

21. The recombinant protein of claim 16, which comprises murine antigen binding region residues and human constant region residues.

22. A non-human transgenic animal that produces an antibody of claim 14.

23. A hybridoma that produces an antibody of claim 15.

24. A single chain monoclonal antibody that comprises the variable domains of the heavy and
l onoclonal antibody of claim 15.

25. A vector comprising a polynucleotide that encodes a single chain monoclonal antibody of claim 24 that immunospecifically binds to a 85P1B3-related protein.

26. A method of delivering a cytotoxic agent to a cell that expresses 85P1B3, said method comprising:

providing a cytotoxic agent conjugated to an antibody or fragment thereof of claim 14; and,

exposing the cell to the antibody-agent conjugate.

27. A composition of claim 4 wherein the substance comprises a polynucleotide that encodes a single chain monoclonal antibody that immunospecifically binds to an 85P1B3-related protein.

28. A composition of claim 4 wherein the substance comprises a polynucleotide that comprises an 85P1B3-related protein coding sequence.

29. A composition of claim 28 comprising a polynucleotide from position number 13 through number 702 of Figure 2 (SEQ ID NO.: ____).

30. The composition of claim 29, wherein T is substituted with U.

31. A composition of claim 28 comprising the polynucleotide of Figure 2 (SEQ ID NO.: ____) in a human unit dose form.

32. The composition of claim 31, wherein T is substituted with U.

33. A composition of claim 28 comprising a polynucleotide that encodes an 85P1B3-related protein that is at least 90% homologous to the entire amino acid sequence shown in Figure 2 (SEQ ID NO.: ____).

34. The composition of claim 33, wherein the polynucleotide encodes an 85P1B3-related protein that is at least 90% identical to the entire amino acid sequence shown in Figure 2 (SEQ ID NO: ____).

35. A composition of claim 28 comprising a polynucleotide that encodes at least one peptide set forth in Tables V-XVIII.

36. A composition of claim 28 comprising a polynucleotide that encodes a peptide region of at l[illegible]s of Figure 2 (SEQ ID NO.: ____) in any whole number increment up to 299 that includes an

amino acid position selected from: an amino acid position having a value greater than 0.5 in the Hydrophilicity profile of Figure 5, an amino acid position having a value less than 0.5 in the Hydropathicity profile of Figure 6; an amino acid position having a value greater than 0.5 in the Percent Accessible Residues profile of Figure 7; an amino acid position having a value greater than 0.5 in the Average Flexibility profile on Figure 8; or an amino acid position having a value greater than 0.5 in the Beta-turn profile of Figure 9.

37. A composition comprising a polynucleotide that is fully complementary to a polynucleotide of claim 29.

38. A composition comprising a polynucleotide that is fully complementary to a polynucleotide of claim 30.

39. A composition comprising a polynucleotide that is fully complementary to a polynucleotide of claim 31, in human unit dose form.

40. A composition comprising a polynucleotide that is fully complementary to a polynucleotide of claim 32.

41. A composition comprising a polynucleotide that is fully complementary to a polynucleotide of claim 33.

42. A composition comprising a polynucleotide that is fully complementary to a polynucleotide of claim 34.

43. A composition comprising a polynucleotide that is fully complementary to a polynucleotide of claim 35.

44. A pharmaceutical composition of claim 4 wherein the substance comprises a ribozyme that cleaves a polynucleotide having 85P1B3 coding sequence and a physiologically acceptable carrier.

45. A pharmaceutical composition of claim 4 wherein the substance comprises human T cells, wherein said T cells specifically recognize an 85P1B3 peptide sequence in the context of a particular HLA molecule.

46. A method of inhibiting growth of cancer cells that expresses 85P1B3, the method comprising: administering to the cells the composition of claim 4.

47. A method of claim 46 of inhibiting growth of cancer cells that express 85P1B3, the method comprising steps of:

administering to said cells an 85P1B3-related protein.

48. A method of claim 46 of inhibiting growth of cancer cells that express 85P1B3, the method comprising steps of:

administering to said cells an antibody or fragment thereof that specifically binds to a 85P1B3-related protein.

49. A method of claim 46 of inhibiting growth of cancer cells that express 85P1B3, the method comprising steps of:

administering to said cells a vector that encodes a single chain monoclonal antibody that immunospecifically binds to an 85P1B3-related protein.

50. A method of claim 46 of inhibiting growth of cancer cells that express 85P1B3, the method comprising steps of:

administering to said cells a vector that comprises a polynucleotide comprising a 85P1B3-related protein coding sequence.

51. A method of claim 46 of inhibiting growth of cancer cells that express 85P1B3, the method comprising steps of:

administering to said cells an antisense polynucleotide complementary to a polynucleotide having a 85P1B3 coding sequence.

52. A method of claim 46 of inhibiting growth of cancer cells that express 85P1B3, the method comprising steps of:

administering to said cells a ribozyme that cleaves a polynucleotide having 85P1B3 coding sequence.

53. A method of claim 46 of inhibiting growth of cancer cells that express 85P1B3 and a particular HLA molecule, the method comprising steps of:

administering to said cells human T cells, wherein said T cells specifically recognize an 85P1B3 peptide sequence in the context of the particular HLA molecule.

54. A method of treating a patient who bears cancer cells that expresses 85P1B3, the method comprising:

administering to the patient the composition of claim 4.

55. A method of claim 54 for treating a patient who bears cancer cells that expresses 85P1B3, the method comprising steps of:

administering to said patient an 85P1B3-related protein.

56. A method of claim 54 for treating a patient who bears cancer cells that expresses 85P1B3, the method comprising steps of:

administering to said patients an antibody or fragment thereof that specifically binds to a 85P1B3-related protein.

57. A method of claim 54 for treating a patient who bears cancer cells that expresses 85P1B3, the method comprising steps of:

administering to said patient a vector that encodes a single chain monoclonal antibody that immunospecifically binds to an 85P1B3-related protein.

58. A method of claim 57 for treating a patient with a cancer that expresses 85P1B3, the method comprising steps of:

administering to said patient a vector to cancer cells that express 85P1B3, whereby the vector delivers the single chain monoclonal antibody coding sequence to the cancer cells and the encoded single chain antibody is expressed intracellularly therein.

59. A method of claim 54 for treating a patient who bears cancer cells that expresses 85P1B3, the method comprising steps of:

administering to said patient a vector that comprises a polynucleotide comprising a 85P1B3-related protein coding sequence.

60. A method of claim 54 for treating a patient who bears cancer cells that expresses 85P1B3, the method comprising steps of:

administering to said patient an antisense polynucleotide complementary to a polynucleotide having a 85P1B3 coding sequence.

61. A method of claim 54 for treating a patient who bears cancer cells that expresses 85P1B3, the method comprising steps of:

administering to said patient a ribozyme that cleaves a polynucleotide having an 85P1B3 coding sequence.

62. A method of claim 54 for treating a patient who bears cancer cells that expresses 85P1B3, the

g steps of:

administering to said patient human T cells, wherein said T cells specifically recognize an 85P1B3 peptide sequence in the context of the particular HLA molecule.

63. A method of generating a mammalian immune response directed to 85P1B3, the method comprising:

exposing cells of the mammal's immune system to an immunogenic portion of an 85P1B3-related protein or a nucleotide sequence that encodes said protein, whereby an immune response is generated to 85P1B3.

64. A method of inducing an immune response of claim 63, said method comprising:

providing a 85P1B3-related protein that comprises at least one T cell or at least one B cell epitope;

contacting the epitope with a mammalian immune system T cell or B cell respectively, whereby the T cell or B cell is induced.

65. The method of claim 64, wherein the immune system cell is a B cell, whereby the induced B cell generates antibodies that specifically bind to the 85P1B3-related protein.

66. The method of claim 64, wherein the immune system cell is a T cell that is a cytotoxic T cell (CTL), whereby the activated CTL kills an autologous cell that expresses the 85P1B3 protein.

67. The method of claim 64, wherein the immune system cell is a T cell that is a helper T cell (HTL), whereby the activated HTL secretes cytokines that facilitate the cytotoxic activity of a CTL or the antibody producing activity of a B cell.

68. An assay for detecting the presence of a 85P1B3-related protein or polynucleotide in a biological sample from a patient who has or who is suspected of having cancer, comprising steps of:

contacting the sample a substance of claim 4 that specifically binds to the 85P1B3-related protein or polynucleotide, respectively; and,

determining that there is a complex of the substance and 85P1B3-related protein or the substance and 85P1B3-related polynucleotide, respectively.

69. An assay of claim 68 for detecting the presence of a 85P1B3-related protein in a biological sample from a patient who has or who is suspected of having cancer, comprising steps of:

contacting the sample with an antibody that specifically binds to the 85P1B3-related protein; and,

determining that there is a complex of the antibody and 85P1B3-related protein.

The assay in accordance with claim 68 further comprising the step of:

obtaining a sample from a patient who has or who is suspected of having cancer.

71. The assay of claim 68 for detecting the presence of an 85P1B3 polynucleotide in a biological sample, comprising:

contacting the sample with a polynucleotide probe that specifically hybridizes to a polynucleotide encoding an 85P1B3-related protein having the amino acid sequence of Figure 2 (SEQ ID NO.: ___); and,

detecting the presence of a hybridization complex formed by the hybridization of the probe with 85P1B3 polynucleotide in the sample, wherein the presence of the hybridization complex indicates the presence of 85P1B3 polynucleotide within the sample.

72. An assay in accordance with claim 68 for detecting the presence of 85P1B3 mRNA in a biological sample from a patient who has or who is suspected of having cancer, said method comprising:

(a) producing cDNA from the sample by reverse transcription using at least one primer;

(b) amplifying the cDNA so produced using 85P1B3 polynucleotides as sense and antisense primers, wherein the 85P1B3 polynucleotides used as the sense and antisense primers are capable of amplifying 85P1B3 cDNA; and

(c) detecting the presence of the amplified 85P1B3 cDNA.

Figure 1

85P1B3 SSH sequence and GenBank homology to OIP5

GATCAGAGGACACATGGGACTCTGCATCTTAATTCCTAAATTTACAGTCAAAGACATTTTCAG
AGATAAGTATTATGAATTCAATAAGAATCTAAAGTAAGTTCTTAAGGCAAATAGCTATAAAA
GAGAAGAATCCTTAGTCTCTCATCTTCTAAAAACAGCTTCACAAATAATTTGGAAAATCAGCC
TAAAGGTAAATAGAAACTGCATTTCCCCTCCATTCTTGAAGCCAATCTTTTTCAAGAAATGAC
TAAGCAGCACCTGTTGTTGAAGACAGCAATAAAGCCTGAACCTGACACTCAAGCTTTGGTACA
GGATC

gb|AF025441.1|AF025441 Homo sapiens Opa-interacting protein... 632 e-179
gb|AF158642.1|AF158642 Homo sapiens metalloproteinase-disin... 42 0.12
gb|AC005075.2|AC005075 Homo sapiens clone RG219E16, complet... 42 0.12
emb|AL096773.6|HS1000E10 Human DNA sequence from clone 1000... 40 0.48

>gb|AF025441.1|AF025441 Homo sapiens Opa-interacting protein OIP5 mRNA, partial cds
Length = 1197

Score = 632 bits (319), Expect = e-179
Identities = 319/319 (100%)
Strand = Plus / Minus

```
Query: 1    gatcagaggacacatgggactctgcatcttaattcctaaatttacagtcaaagacatttt 60
             ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct: 1013 gatcagaggacacatgggactctgcatcttaattcctaaatttacagtcaaagacatttt 954

Query: 61   cagagataagtattatgaattcaataagaatctaaagtaagttcttaaggcaaatagcta 120
             ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct: 953  cagagataagtattatgaattcaataagaatctaaagtaagttcttaaggcaaatagcta 894

Query: 121  taaaagagaagaatccttagtctctcatcttctaaaaacagcttcacaaataatttggaa 180
             ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct: 893  taaaagagaagaatccttagtctctcatcttctaaaaacagcttcacaaataatttggaa 834

Query: 181  aatcagcctaaaggtaaatagaaactgcatttcccctccattcttgaagccaatcttttt 240
             ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct: 833  aatcagcctaaaggtaaatagaaactgcatttcccctccattcttgaagccaatcttttt 774

Query: 241  caagaaatgactaagcagcacctgttgttgaagacagcaataaagcctgaacctgacact 300
             ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct: 773  caagaaatgactaagcagcacctgttgttgaagacagcaataaagcctgaacctgacact 714

Query: 301  caagctttggtacaggatc 319
             ||||||||||||||||||
Sbjct: 713  caagctttggtacaggatc 695
```

Figure 2

cDNA Sequence and ORF of 85P1B3/OIP5 clone A

```
                9          18          27          36          45          54
5'  GGC TGC GGG AAG ATG GCG GCT CAG CCG CTG CGG CAT CGC TCA CGT TGT GCA ACG
    --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
                    M   A   A   Q   P   L   R   H   R   S   R   C   A   T

           63          72          81          90          99          108
    CCG CCC CGG GGG GAC TTT TGT GGT GGC ACT GAG AGG GCG ATT GAC CAA GCT TCT
    --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
     P   P   R   G   D   F   C   G   G   T   E   R   A   I   D   Q   A   S

           117         126         135         144         153         162
    TTT ACG ACC TCC ATG GAG TGG GAT ACG CAG GTG GTG AAG GGG TCC TCG CCG CTC
    --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
     F   T   T   S   M   E   W   D   T   Q   V   V   K   G   S   S   P   L

           171         180         189         198         207         216
    GGC CCC GCA GGG CTG GGG GCT GAG GAG CCA GCC GCC GGC CCG CAG CTG CCG TCT
    --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
     G   P   A   G   L   G   A   E   E   P   A   A   G   P   Q   L   P   S

           225         234         243         252         261         270
    TGG CTG CAG CCT GAG AGG TGC GCT GTG TTC CAG TGC GCA CAG TGT CAC GCA GTG
    --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
     W   L   Q   P   E   R   C   A   V   F   Q   C   A   Q   C   H   A   V

           279         288         297         306         315         324
    CTC GCC GAC TCG GTG CAC CTC GCC TGG GAC CTG TCG CGG TCC CTC GGG GCC GTG
    --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
     L   A   D   S   V   H   L   A   W   D   L   S   R   S   L   G   A   V

           333         342         351         360         369         378
    GTC TTC TCC AGA GTT ACA AAT AAC GTC GTT TTG GAA GCG CCC TTC CTA GTT GGC
    --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
     V   F   S   R   V   T   N   N   V   V   L   E   A   P   F   L   V   G

           387         396         405         414         423         432
    ATT GAA GGT TCA CTC AAA GGC AGT ACT TAC AAC CTT TTA TTC TGT GGT TCT TGT
    --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
     I   E   G   S   L   K   G   S   T   Y   N   L   L   F   C   G   S   C

           441         450         459         468         477         486
    GGG ATT CCC GTT GGT TTC CAT CTG TAT TCT ACC CAT GCT GCC CTG GCT GCC TTG
    --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
     G   I   P   V   G   F   H   L   Y   S   T   H   A   A   L   A   A   L

           495         504         513         522         531         540
    AGA GGT CAC TTC TGC CTT TCC AGT GAC AAA ATG GTG TGC TAT CTC TTA AAA ACA
    --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
     R   G   H   F   C   L   S   S   D   K   M   V   C   Y   L   L   K   T

           549         558         567         576         585         594
    AAA GCC ATA GTA AAT GCA TCA GAG ATG GAT ATT CAA AAT GTT CCT CTA TCA GAA
    --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
     K   A   I   V   N   A   S   E   M   D   I   Q   N   V   P   L   S   E

           603         612         621         630         639         648
    AAG ATT GCA GAG CTG AAA GAG AAG ATA GTG CTA ACG CAC AAT CGC TTA AAA TCA
    --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
     K   I   A   E   L   K   E   K   I   V   L   T   H   N   R   L   K   S
```

```
        657         666         675         684         693         702
CTA ATG AAG ATT CTG AGT GAA GTG ACT CCT GAC CAG TCC AAG CCA GAA AAC TGA
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 L   M   K   I   L   S   E   V   T   P   D   Q   S   K   P   E   N   *

        711         720         729         738         747         756
TCC TGT ACC AAA GCT TGA GTG TCA GGT TCA GGC TTT ATT GCT GTC TTC AAC AAC
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
        765         774         783         792         801         810
AGG TGC TGC TTA GTC ATT TCT TGA AAA AGA TTG GCT TCA AGA ATG GAG GGG AAA
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
        819         828         837         846         855         864
TGC AGT TTC TAT TTA CCT TTA GGC TGA TTT TCC AAA TTA TTT GTG AAG CTG TTT
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
        873         882         891         900         909         918
TTA GAA GAT GAG AGA CTA AGG ATT CTT CTC TTT TAT AGC TAT TTG CCT TAA GAA
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
        927         936         945         954         963         972
CTT ACT TTA GAT TCT TAT TGA ATT CAT AAT ACT TAT CTC TGA AAA TGT CTT TGA
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
        981         990         999        1008        1017        1026
CTG TAA ATT TAG GAA TTA AGA TGC AGA GTC CCA TGT GTC CTC TGA TCT AAA GTT
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
       1035        1044        1053        1062        1071        1080
GCA TGG TTG GTC TGA AAA TAG AGT TGG GCT TAA TGT TGA CTT CTA TTA CTC CTG
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
       1089        1098        1107        1116        1125        1134
CAT GGA GCA GTT GTT ATG AAT ACT AAT ACA TCA CTT TTT AAC TTC TGT AAA ATA
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
       1143        1152        1161        1170        1179        1188
CAG ATC ATA ATA TTC TAT AGG TAA TGT TTA ATA AAT TGC CTG AAT AAT AAA AAA
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
       1197        1206        1215        1224        1233        1242
AAA AAA AAA AAA AAA AAA AAA AAA AAA AAA AAA AAA AAA AAA AAA AAA AAA AAA
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
       1251        1260
AAA AAA AAA AAA AAA AAA AA 3'
--- --- --- --- --- --- --
```

Figure 3

85P1B3/OIP5 protein sequence.

```
  1 MAAQPLRHRS RCATPPRGDF CGGTERAIDQ ASFTTSMEWD TQVVKGSSPL GPAGLGAEEP
 61 AAGPQLPSWL QPERCAVFQC AQCHAVLADS VHLAWDLSRS LGAVVFSRVT NNVVLEAPFL
121 VGIEGSLKGS TYNLLFCGSC GIPVGFHLYS THAALAALRG HFCLSSDKMV CYLLKTKAIV
181 NASEMDIQNV PLSEKIAELK EKIVLTHNRL KSLMKILSEV TPDQSKPEN*
```

Figure 4

Alignment of 85P1B3 with OIP5.

>gi|2815610|gb|AAC39561.1| (AF025441) Opa-interacting protein OIP5 [Homo sapiens]
Length = 231

Score = 462 bits (1189), Expect = e-130
Identities = 229/229 (100%), Positives = 229/229 (100%)

```
85P1B3: 1   MAAQPLRHRSRCATPPRGDFCGGTERAIDQASFTTSMEWDTQVVKGSSPLGPAGLGAEEP 60
            MAAQPLRHRSRCATPPRGDFCGGTERAIDQASFTTSMEWDTQVVKGSSPLGPAGLGAEEP
OIP5: 3     MAAQPLRHRSRCATPPRGDFCGGTERAIDQASFTTSMEWDTQVVKGSSPLGPAGLGAEEP 62

85P1B3: 61  AAGPQLPSWLQPERCAVFQCAQCHAVLADSVHLAWDLSRSLGAVVFSRVTNNVVLEAPFL 120
            AAGPQLPSWLQPERCAVFQCAQCHAVLADSVHLAWDLSRSLGAVVFSRVTNNVVLEAPFL
OIP5: 63    AAGPQLPSWLQPERCAVFQCAQCHAVLADSVHLAWDLSRSLGAVVFSRVTNNVVLEAPFL 122

85P1B3:121  VGIEGSLKGSTYNLLFCGSCGIPVGFHLYSTHAALAALRGHFCLSSDKMVCYLLKTKAIV 180
            VGIEGSLKGSTYNLLFCGSCGIPVGFHLYSTHAALAALRGHFCLSSDKMVCYLLKTKAIV
OIP5: 123   VGIEGSLKGSTYNLLFCGSCGIPVGFHLYSTHAALAALRGHFCLSSDKMVCYLLKTKAIV 182

85P1B3:181  NASEMDIQNVPLSEKIAELKEKIVLTHNRLKSLMKILSEVTPDQSKPEN 229
            NASEMDIQNVPLSEKIAELKEKIVLTHNRLKSLMKILSEVTPDQSKPEN
OIP5: 183   NASEMDIQNVPLSEKIAELKEKIVLTHNRLKSLMKILSEVTPDQSKPEN 231
```

Figure 5: 85P1B3 Hydrophilicity profile
(Hopp T.P., Woods K.R., 1981. Proc. Natl. Acad. Sci. U.S.A. 78:3824-3828)

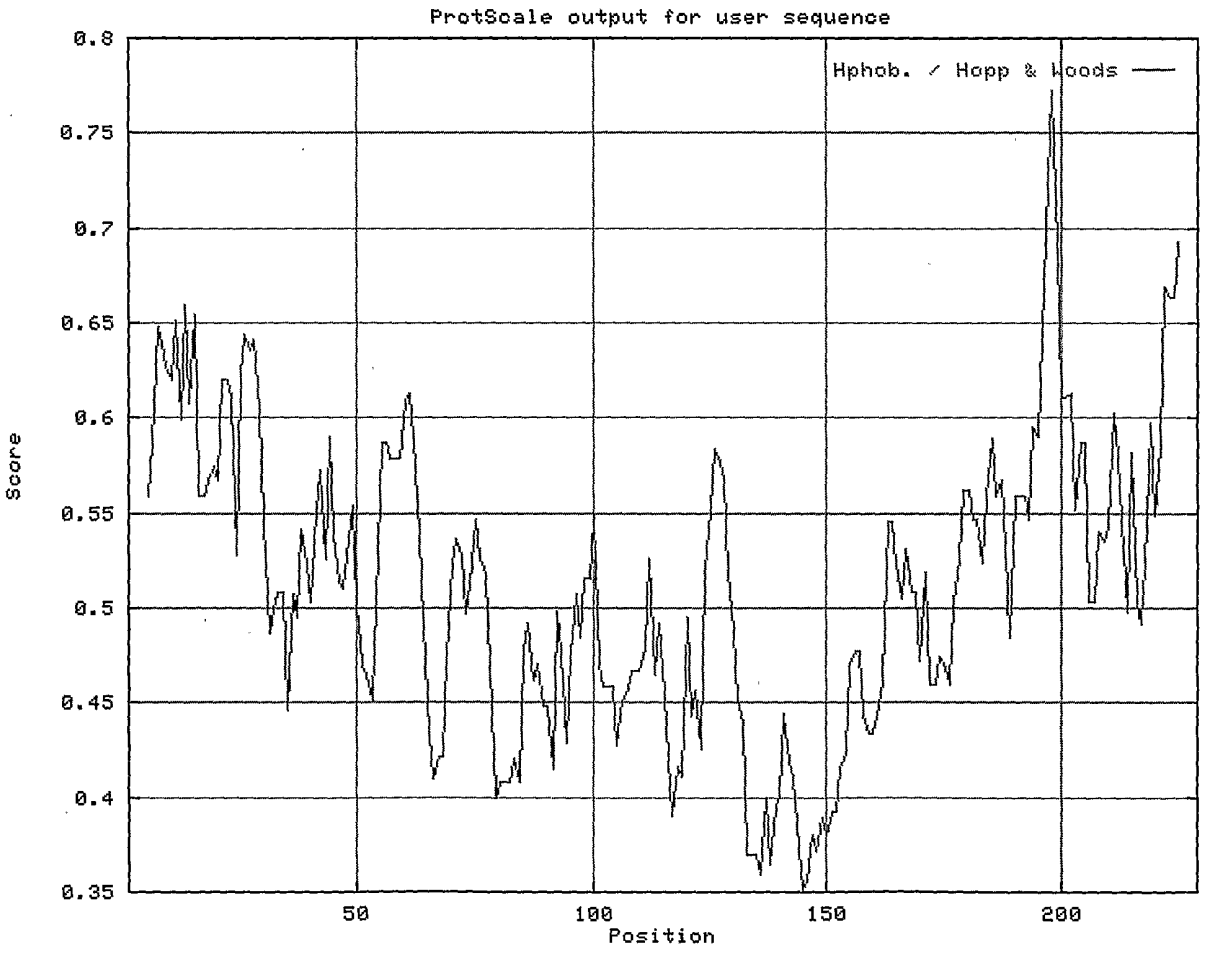


Figure 6: 85P1B3 Hydropathicity Profile
(Kyte J., Doolittle R.F., 1982. J. Mol. Biol. 157:105-132)

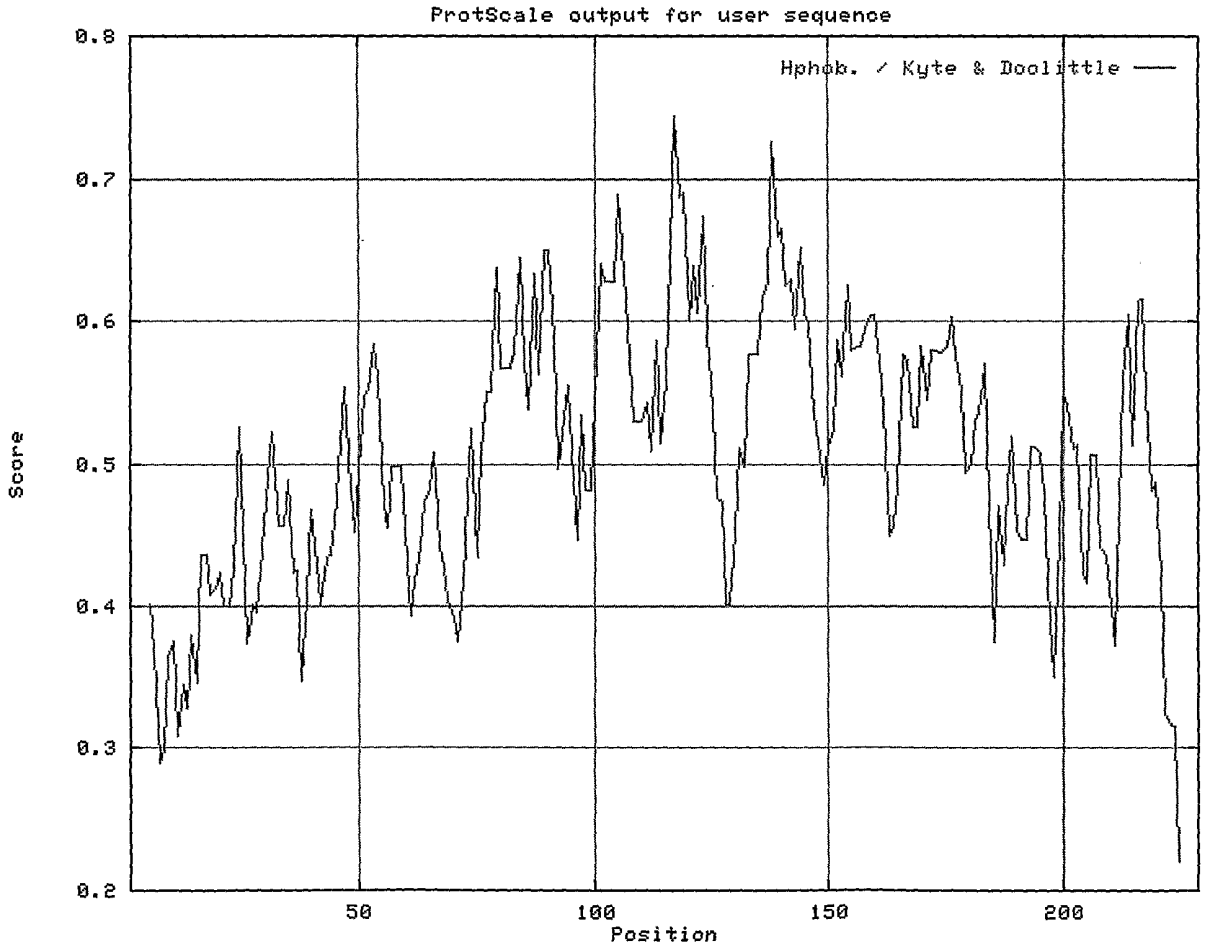


Figure 7: 85P1B3 % Accessible Residues Profile
(Janin J., 1979. Nature 277:491-492)

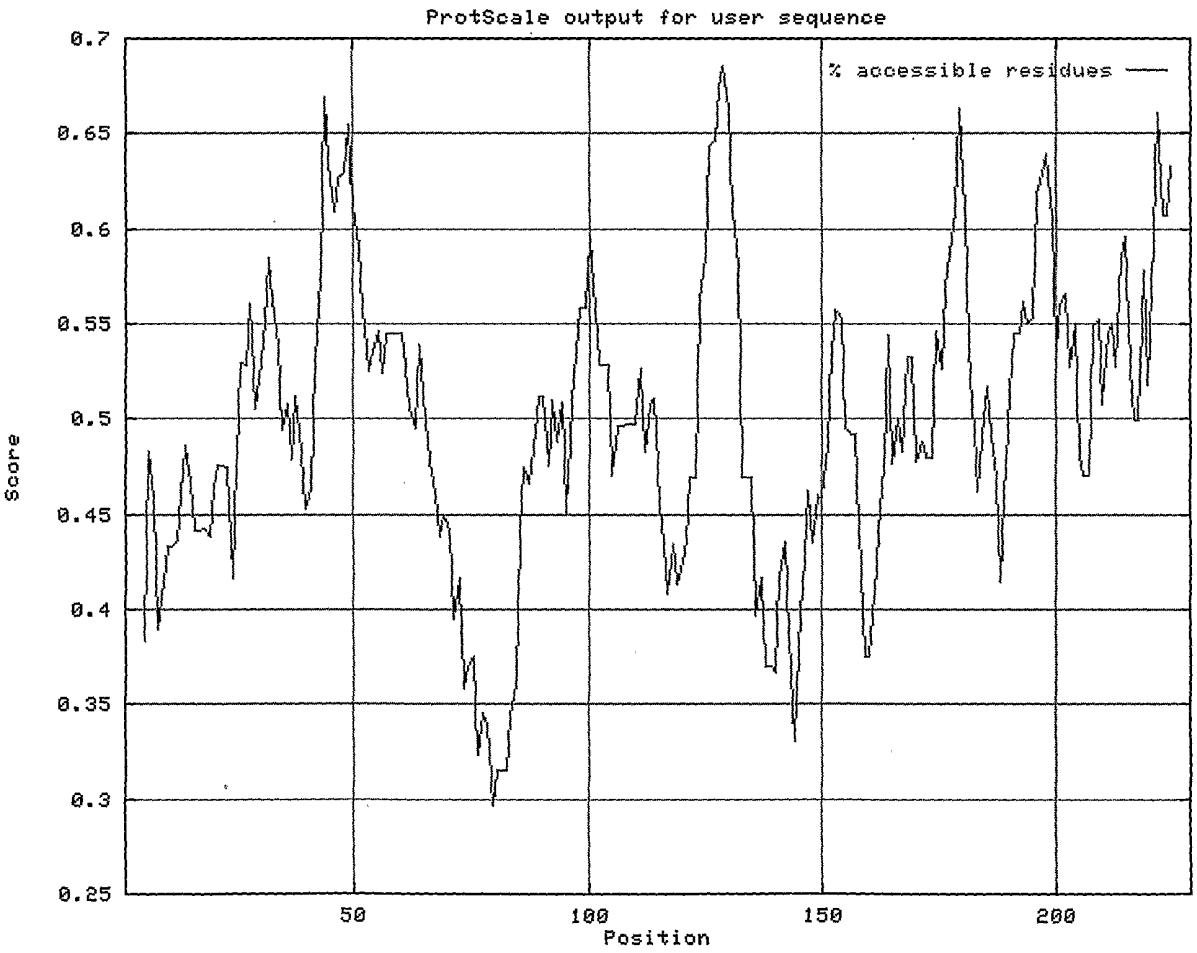


Figure 8: 85P1B3 Average Flexibility Profile

(Bhaskaran R., Ponnuswamy P.K., 1988.
Int. J. Pept. Protein Res. 32:242-255)

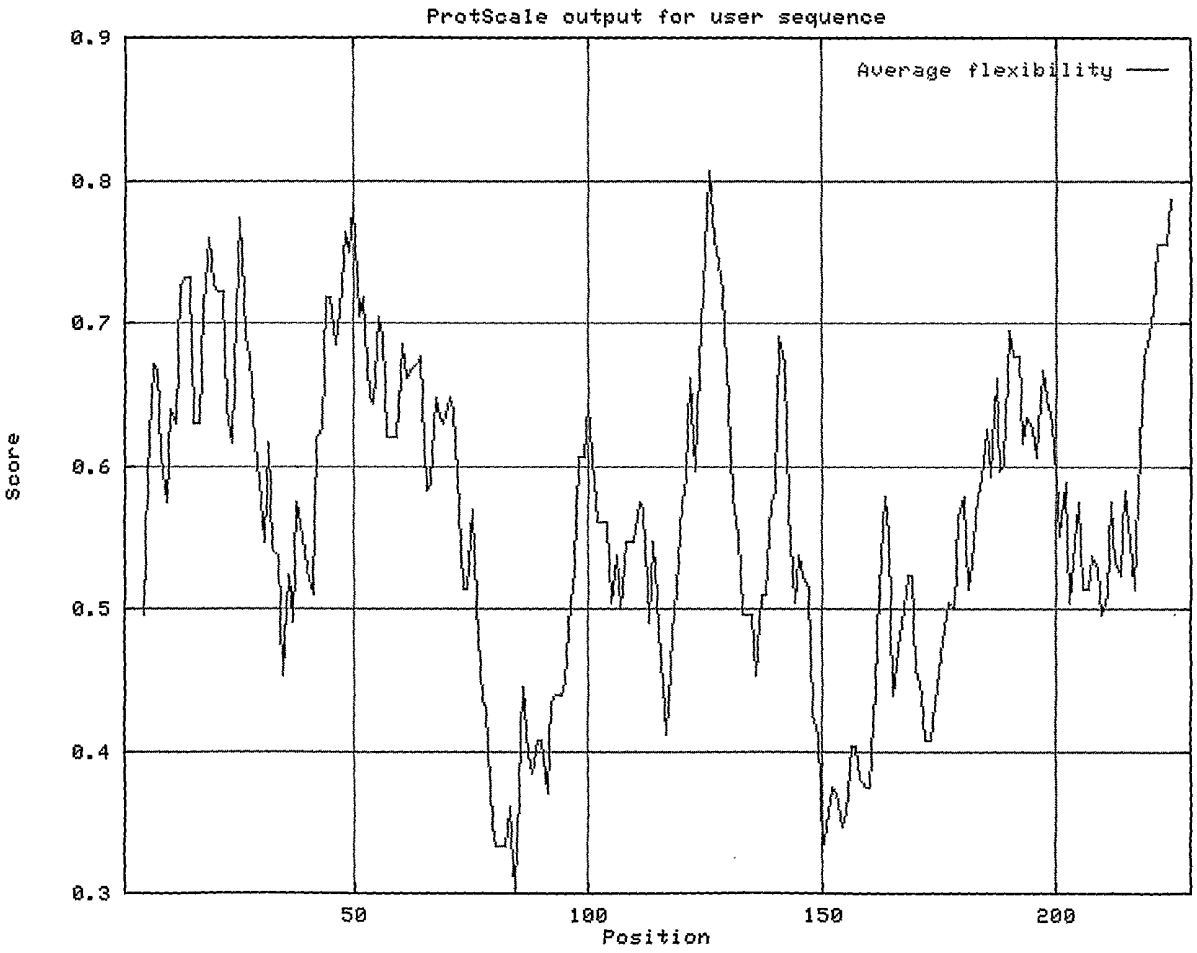


Figure 9: 85P1B3 Beta-turn Profile
(Deleage, G., Roux B. 1987. Protein Engineering 1:289-294)

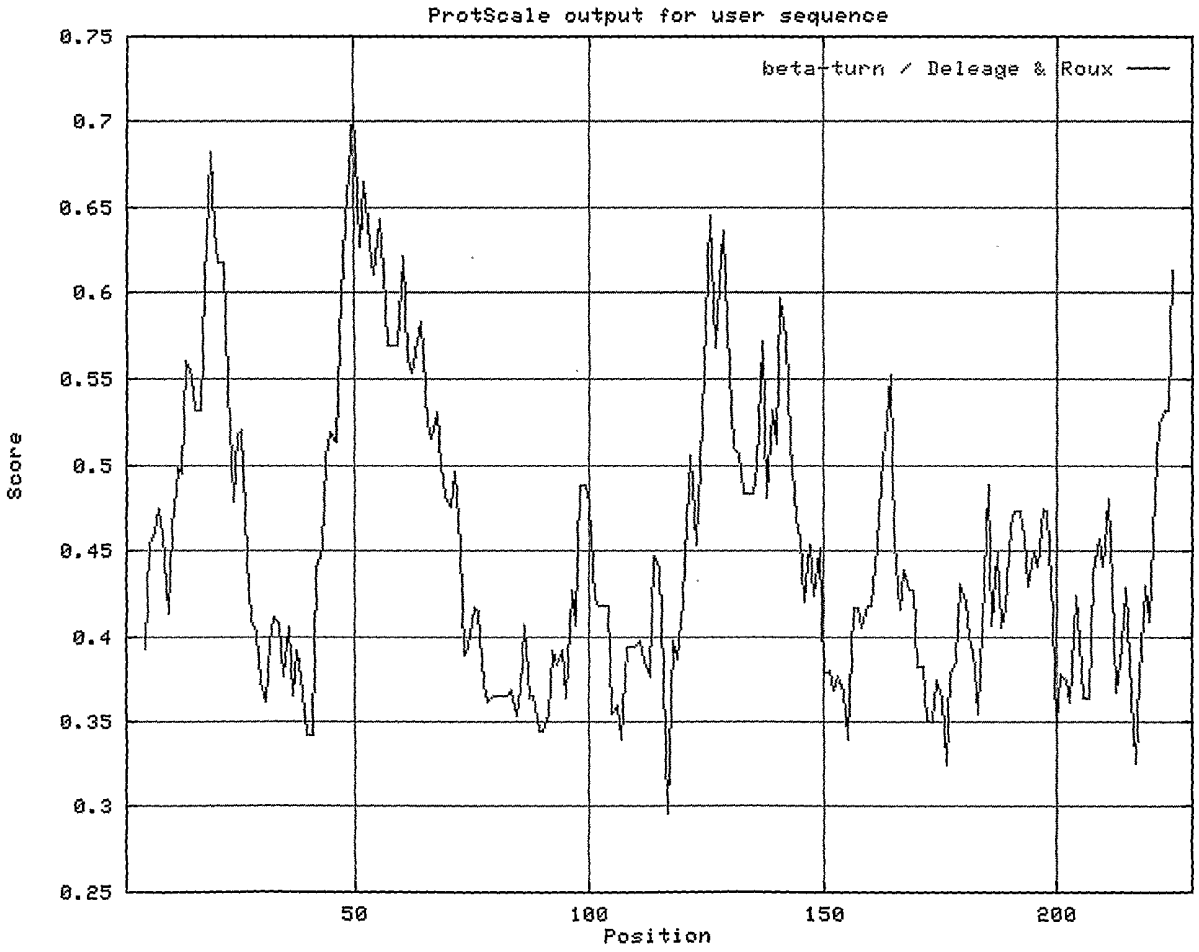


Figure 10 RT-PCR analysis of 85P1B3 expression.

1) Vital Pool 1
2) Vital Pool 2
3) Xenograft Pool
4) Prostate Cancer Pool
- Bladder Cancer Pool
- Kidney Cancer Pool
- Colon Cancer Pool
- Lung Cancer Pool
- Ovary Cancer Pool
- Breast Cancer Pool
- Metastasis Pool
- H2O

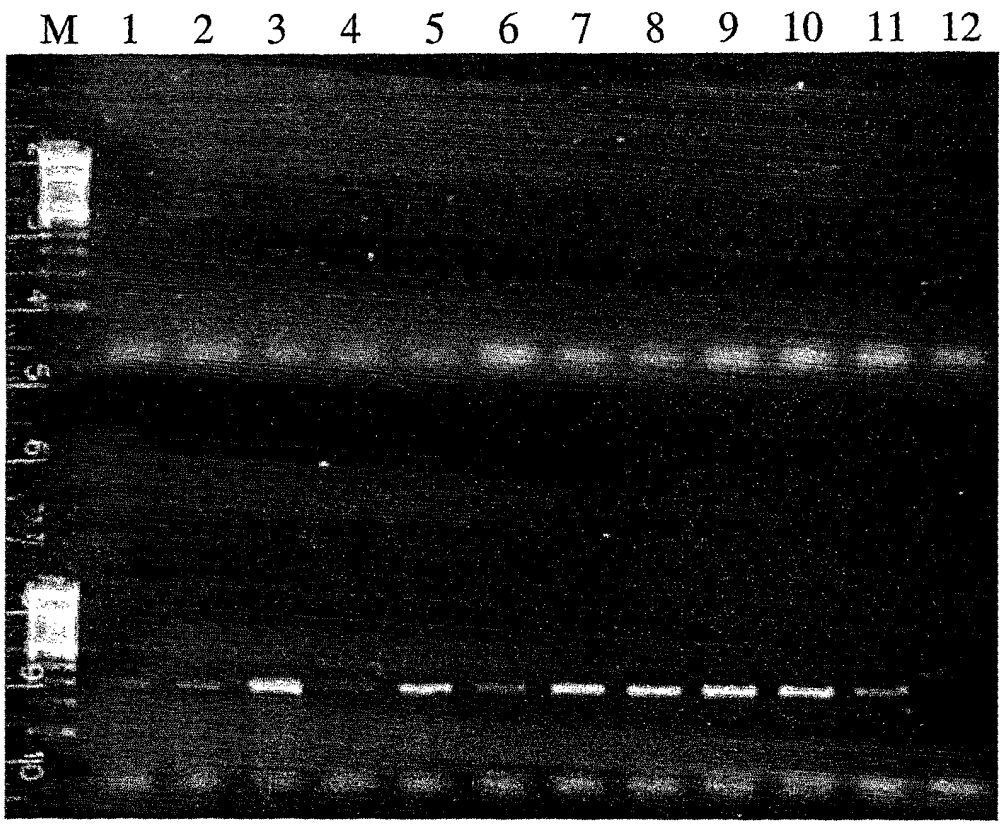
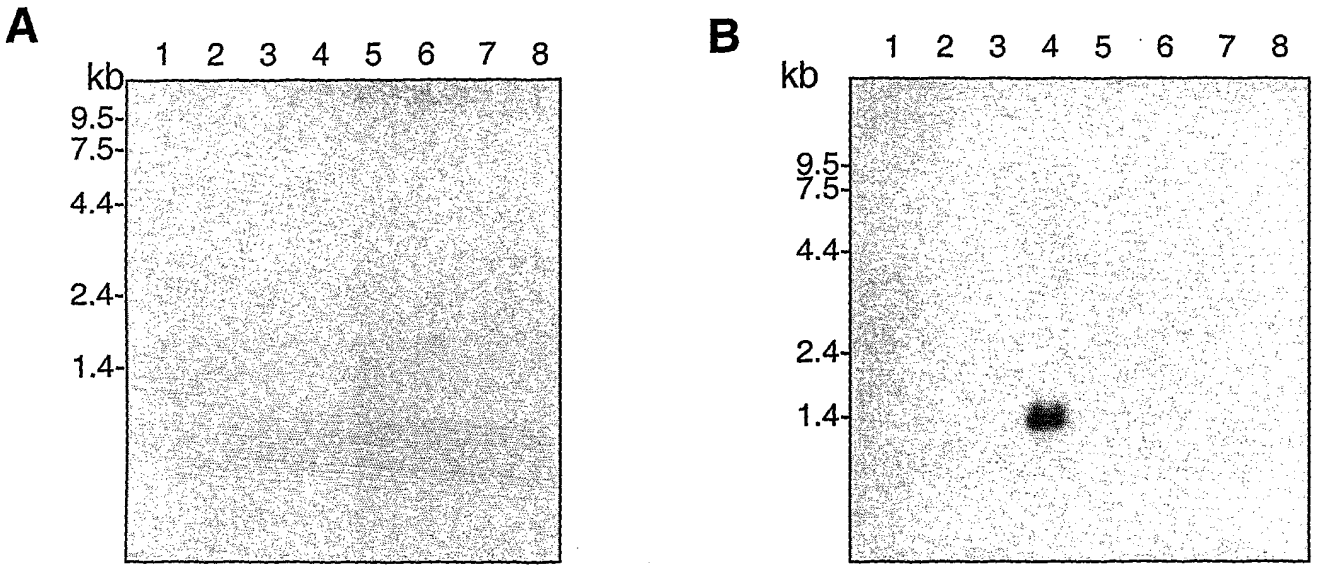


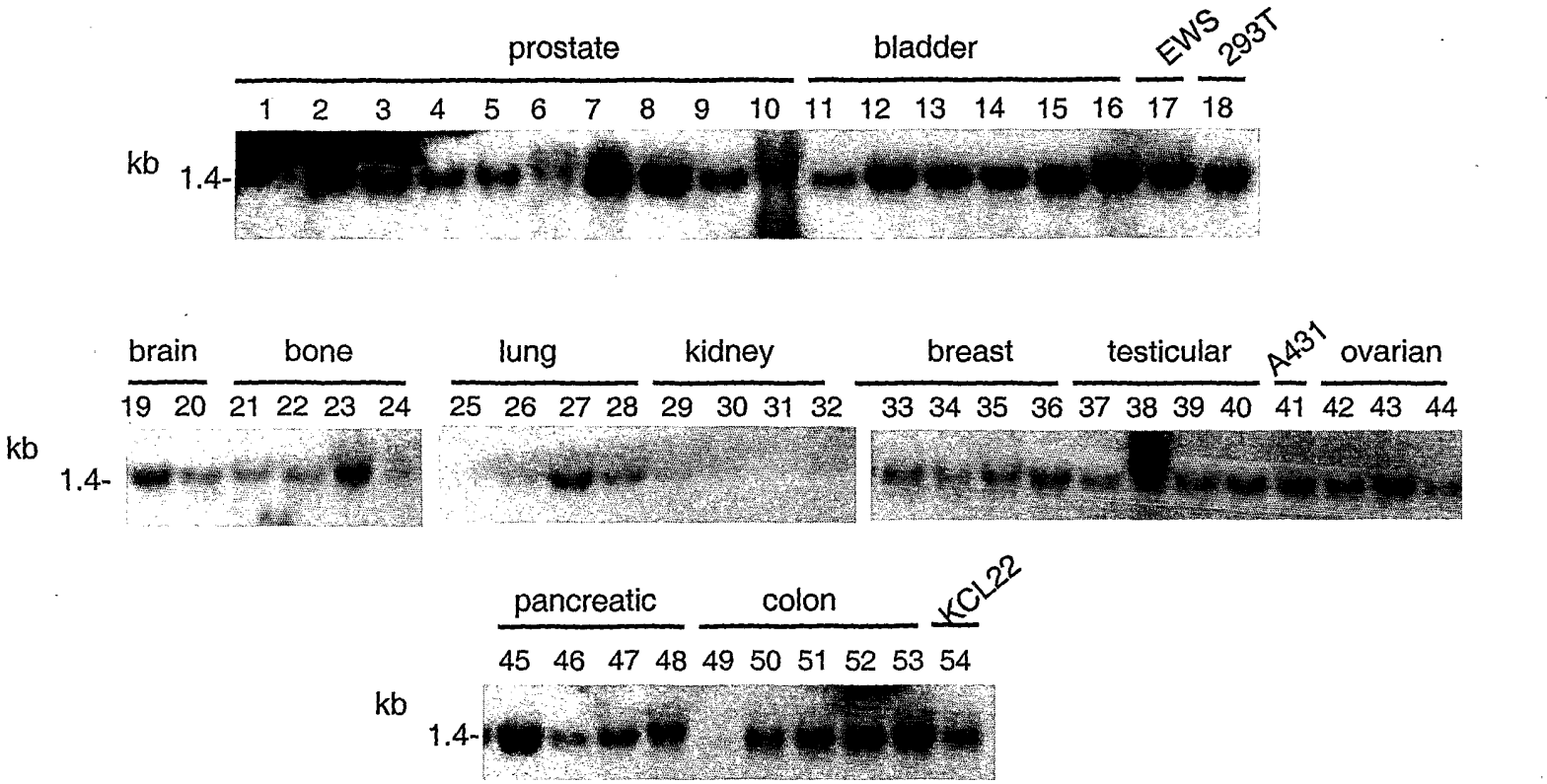
Figure 11 Expression of 85P1B3 in Normal Human Tissues



1. Heart
2. Brain
3. Placenta
4. Lung
5. Liver
6. Skeletal Muscle
7. Kidney
8. Pancreas

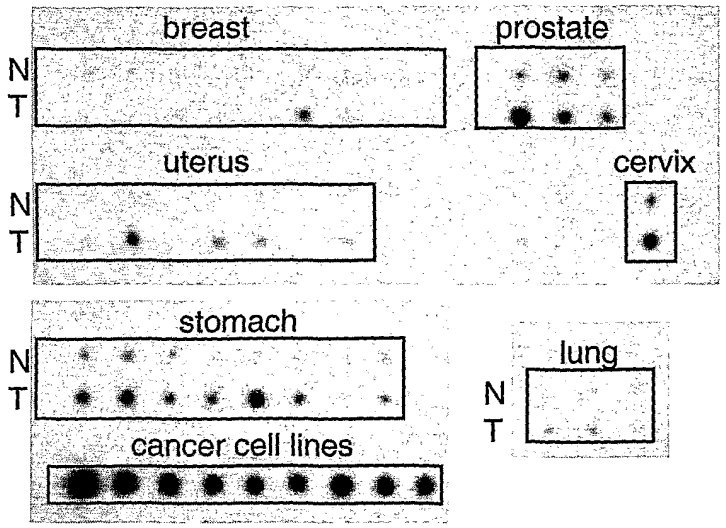
1. Spleen
2. Thymus
3. Prostate
4. Testis
5. Ovary
6. Small Intestine
7. Colon
8. Leukocytes

Figure 12 Expression of 85P1B3 in Human Cancer Cell lines



1. PrEC
2. LAPC-4 AD
3. LAPC-4 AI
4. LAPC-9 AD
5. LAPC-9 AI
6. LNCaP
7. PC-3
8. DU145
9. TsuPr1
10. LAPC-4 CL
11. HT1197
12. SCaBER
13. UM-UC-3
14. TCCSUP
15. J82
16. 5637
17. RD-ES
18. 293T
19. PFSK-1
20. T98G
21. SK-ES-1
22. HOS
23. U2-OS
24. RD-ES
25. CALU-1
26. A427
27. NCI-H82
28. NCI-H146
29. 769-P
30. A498
31. CAKI-1
32. SW839
33. CAMA-1
34. DU4475
35. MCF-7
36. MDA-MB-435s
37. NTERRA-2
38. NCCIT
39. TERA-1
40. TERA-2
41. A431
42. OV-1063
43. PA-1
44. SW626
45. PANC-1
46. Bx PC-3
47. HPAC
48. Capan-1
49. SK-CO-1
50. Caco-2
51. LoVo
52. T84
53. Colo205
54. KCL 22

Figure 13 Expression of 85P1B3 in Patient Cancer Specimens and Cancer Cell Lines

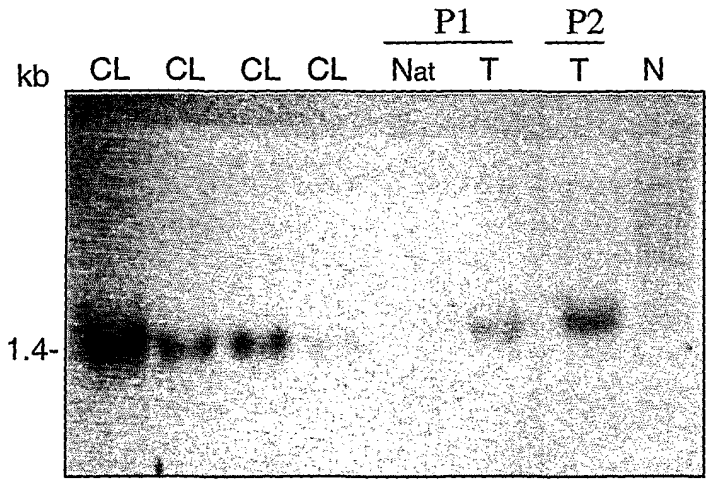


T = tumor RNA
N = normal adjacent tissue RNA

Cancer cell lines are:
(from left to right)

HeLa (cervical carcinoma)
Daudi (Burkitt's lymphoma)
K562 (CML)
HL-60 (PML)
G361 (melanoma)
A549 (lung carcinoma)
MOLT-4 (lymphoblastic leuk.)
SW480 (colorectal carcinoma)
Raji (Burkitt's lymphoma)

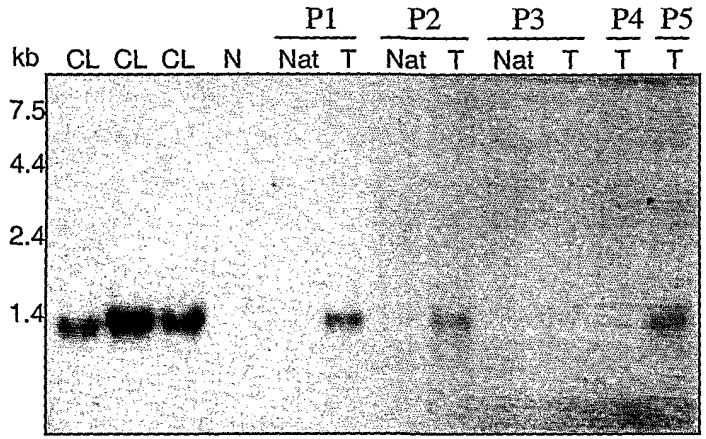
Figure 14 Expression of 85P1B3 in Colon Cancer Patient Specimens



P1 – Stage III, T2N1MX
P2 – Stage III, T3N1MX

Cell Lines (CL) - Colo 205, LoVo, T84, Caco-2
N = Normal Colon
Nat = Normal Adjacent tissue
T = Tumor

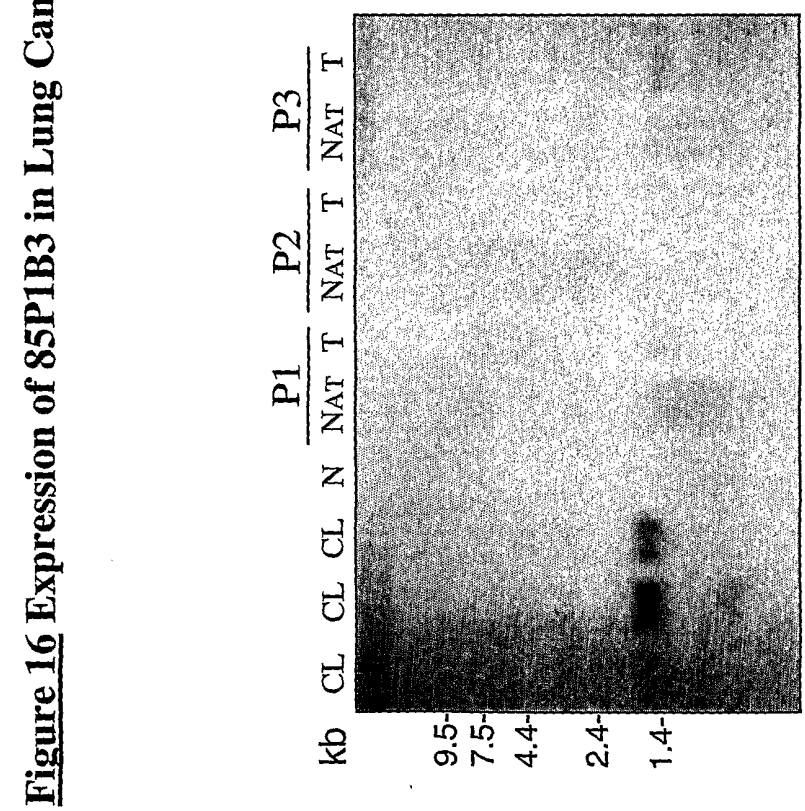
Figure 15 Expression of 85P1B3 in Bladder Cancer Patient Specimens



P1 - Transitional, grade 2
P2 - Transitional, grade 3/2
P3 - Transitional
P4 - Polypoid cystitis
P5 - Papillary, grade 3

Cl = cell lines listed in order: UM-UC-3, J82, SCABER
P = Patient
N = Normal Bladder
Nat = Normal adjacent tumor
T = Tumor

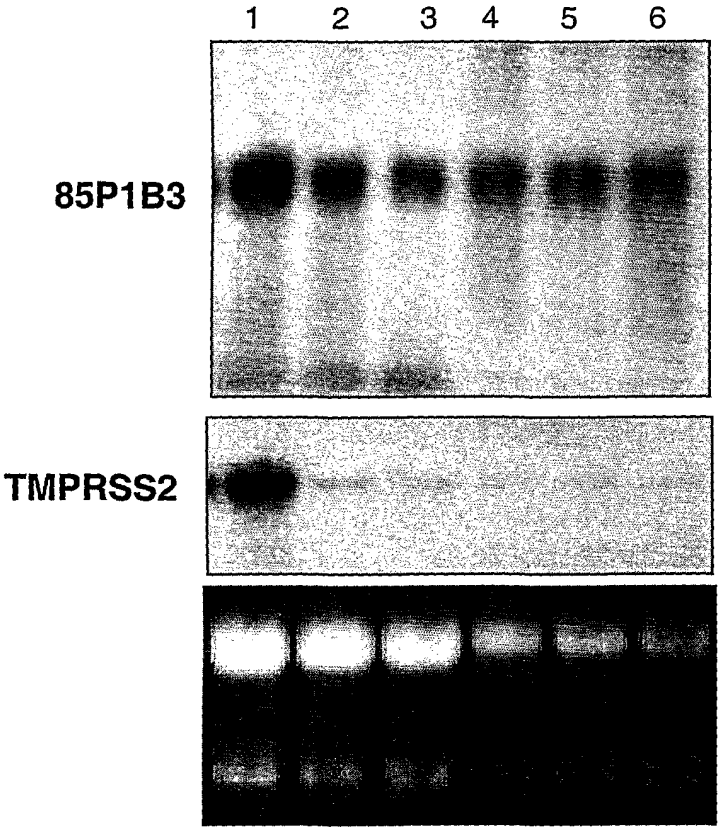
Figure 16 Expression of 85P1B3 in Lung Cancer Patient Specimens



P1 - Squamous, stage IB
P2 - Squamous, stage IIB
P3 - Squamous, stage IIIA

CL = cell lines (listed in order): A427, NCI-H82, NCI-H146
P = Patient
N = Normal Lung
NAT = Normal adjacent tissue
T = Tumor

Figure 17 Expression of 85P1B3 in Prostate Cancer Xenografts Following Castration



- LAPC-9AD Day 0 – Mouse #1
- LAPC-9AD Day 7 – Mouse #2
- LAPC-9AD Day 7 – Mouse #3
- LAPC-9AD Day 15 – Mouse #4
- LAPC-9AD Day 15 – Mouse #5
- LAPC-9AD Day 21 – Mouse #6

Figure 18 Expression of 85P1B3 in PC3 Cells Following Retroviral-Mediated Gene Delivery

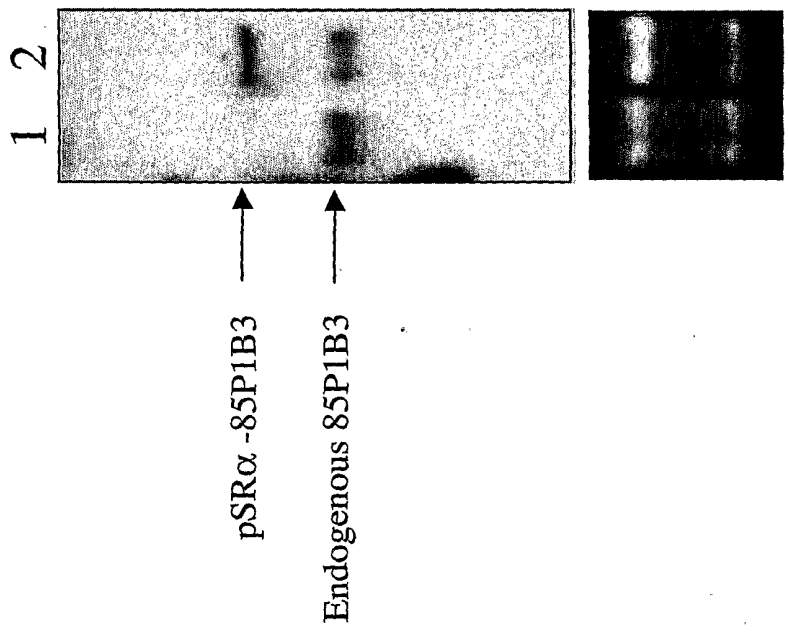
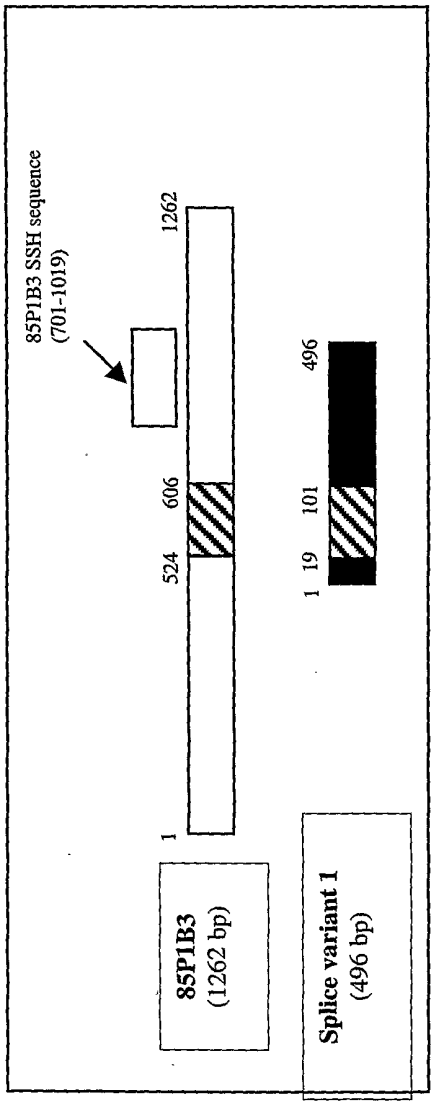


Figure 19.


85P1B3 SSH sequence
(701-1019)
1262
606
524
1
496
101
19
1
85P1B3
(1262 bp)
Splice variant 1
(496 bp)

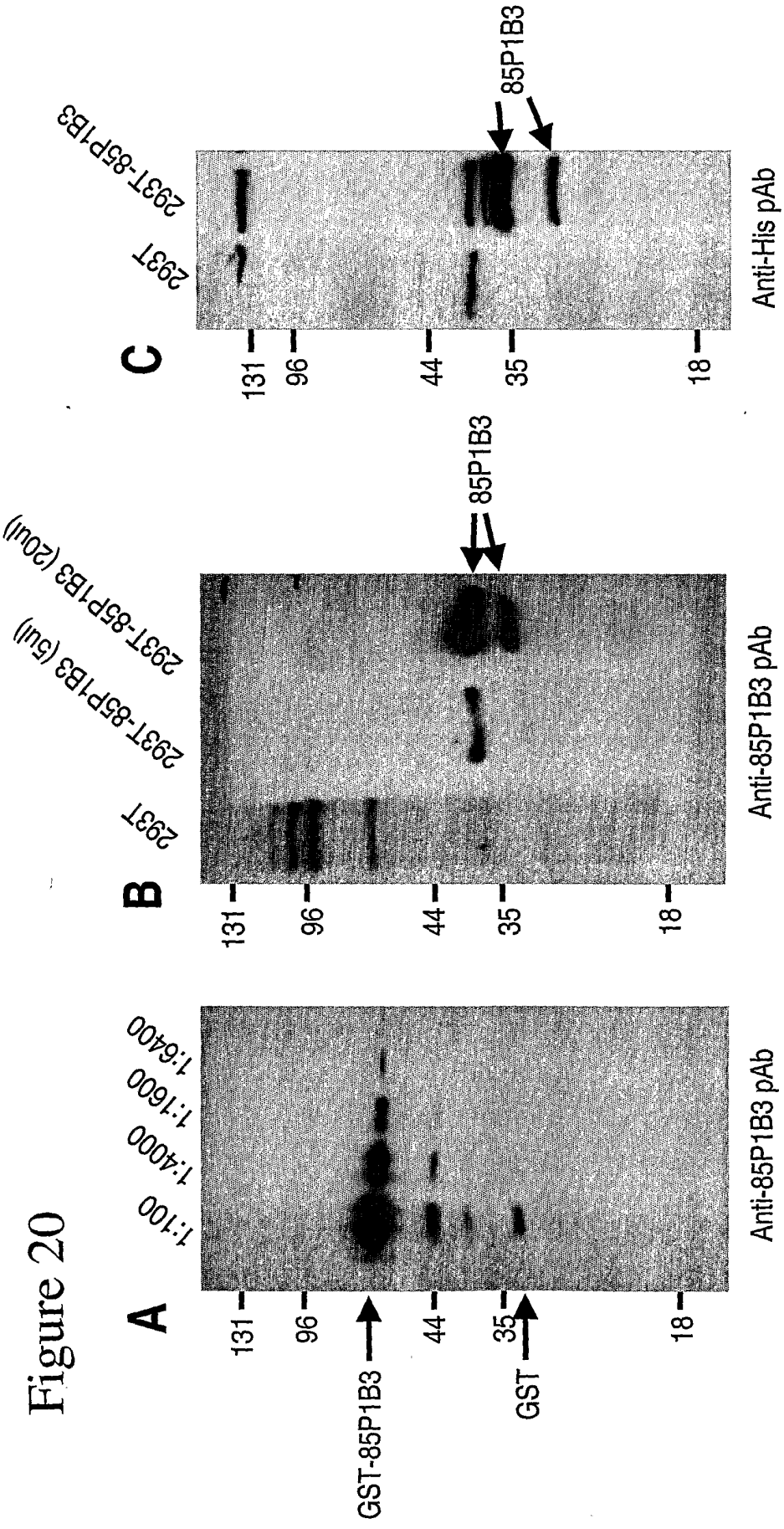

A
1:100
1:400
1:1600
1:6400
131
96
44
35
18
GST-85P1B3
GST
Anti-85P1B3 pAb
B
293T
293T-85P-1B3 (5ul)
293T-85P-1B3 (20ul)
85P1B3
131
96
44
35
18
Anti-85P1B3 pAb
C
293T
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85P1B3
131
96
44
35
18
Anti-His pAb

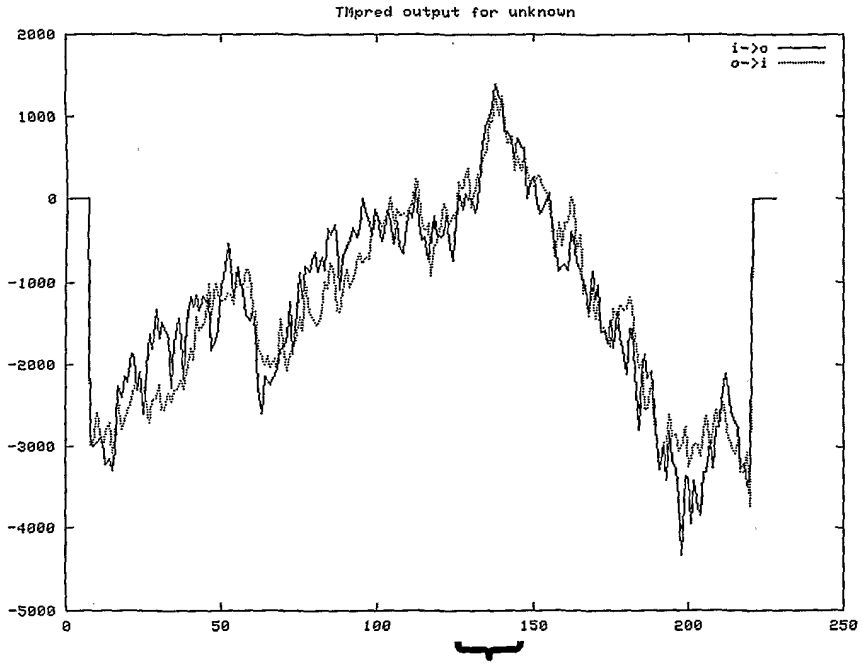
Figure 20

Fig. 21A

```
         10        20        30        40        50        60        70
          |         |         |         |         |         |         |
MAAQPLRHRSRCATPPRGDFCGGTERAIDQASFTTSMEWDTQVVKGSSPLGPAGLGAEEPAAGPQLPSWL
ccccccccccccccccccccccchhhhhccccceeccccccceeeeccccccccccccccccccccccccc
QPERCAVFQCAQCHAVLADSVHLAWDLSRSLGAVVFSRVTNNVVLEAPFLVGIEGSLKGSTYNLLFCGSC
chhhhhhhhhhhhhhhhhhhhhhhhhhhhccceeeeeeccccceeeeccceeeeeccccccccceeeeeeccc
GIPVGFHLYSTHAALAALRGHFCLSSDKMVCYLLKTKAIVNASEMDIQNVPLSEKIAELKEKIVLTHNRL
cccchhhhhhhhhhhhhhhcccecchhhhhhhhhhhceeeccccccccccchhhhhhhhhhhhhhhhhhh
KSLMKILSEVTPDQSKPEN
hhhhhhehccccccccccc
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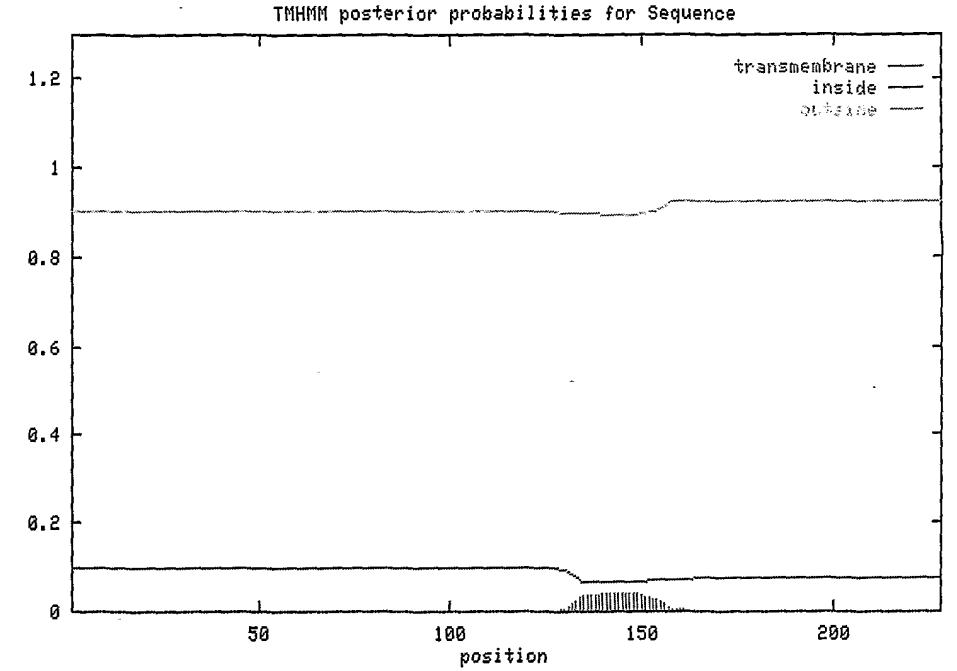
c: random coil (49.34%)
e: extended strand (13.97%)
h: alpha helix (36.68%)

Fig. 21B



1 transmembrane from amino acids 129-149

Fig. 21C



No transmembrane domains, soluble protein