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(54) Title: NUCLEOSIDE DERIVATIVE, MODIFIED OLIGONUCLEOTIDE, METHOD FOR THEIR SYNTHESIS AND AP-
PLICATIONS THEREOF

(57) Abstract: New class of metallated nucleoside and nucleotide conjugates containing metallacarborane cage is provided, also
oligonucleotides containing metallacarborane modification are disclosed. Nucleosides, nucleotides and oligonucleotides containing
metallacarborane cage can be used among others as modified primers in amplification of RNA and DNA, antisense drugs, boron
carriers for BNCT, radiopharmaceuticals bearing a range of isotopes useful in different types of radiotherapy, molecular probes,
elements of biosensors, materials for nanotechnology and others.



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Nucleoside derivative, modified oligonucleotide, method for their synthesis and applications thereof

FIELD OF THE INVENTION

This invention is in the area of synthetic organic chemistry, and in particular conjugates of nucleosides and oligonucleotides with boron clusters containing metals and other elements and their isotopes, their molecular structure and method of preparation and use.

The invention concerns also application of carboranes and metallacarboranes as a new type of electrochemically active label for biological compounds. The derivatives labelled with carborane or metallacarborane can be detected by electrochemical methods and may find several other practical applications, specifically as materials for nanoconstruction, in DNA array technology or for the construction of biosensors, especially electrochemical biosensors.

BACKGROUND OF THE INVENTION

Nucleosides and nucleotides, components of nucleic acids, and oligonucleotides (short stretches of nucleic acids) find broad application among others in biology, medicine and nanotechnology [Agrawal, 1996; Cotter, 1996; Crooke, 1995; Ells, 1996; Jefferies and De Clercq, 1995; Keller and Manak, 1993; Niemeyer, 1997; Seeman et al, 1998; Wiedbrauk and Farkas, 1995].

Due to the different requirements dependent upon specific applications of oligonucleotides and their components, natural nucleosides, nucleotides and oligonucleotides are modified in order to obtain molecules with designed and needed properties. Essentially all parts of nucleoside, nucleotide and oligonucleotide can be modified, namely sugar residue, nucleic base and phosphate group (Agrawal, 1993; Micklefield, 2001; Shabarova and Bogdanov, 1994; Sanghvi and Cook, 1994; Uhlmann, E., Peyman, A., 1990).

One of the modifications is based on the use of boron clusters such as carboranes, as modifying entity. The carborane group can be attached to the nucleoside unit directly or through a linker. Most nucleosides modified with carborane group belong to pyrimidine nucleoside family. (Leśnikowski i Schinazi, 1995; Tjarks, W., 2000).

Three types of carboranyl group ($-C_2B_{10}H_{12}$) containing DNA-oligonucleotides have been described so far: 1) (*o*-carboran-1-yl)methylphosphonate-oligonucleotides (CBMP-oligonucleotides), consisting carborane cage within internucleotide linkage (Lesnikowski and Schinazi, 1993; Lesnikowski et al, 1997; Lesnikowski, 2003), 2) 5-(*o*-carboran-1-yl)-2'-deoxyuridine-oligonucleotides (CDU-oligonucleotides) containing carborane cage attached to nucleobase

(Fulcrand-El Kattan et al, 1994; Lesnikowski et al, 1996; Lesnikowski, 2003), and 3) 2'-O-(*o*-carboran-1-yl)methyl-oligonucleotides (2'-CBM-oligonucleotides) with carborane cage linked to sugar residue at 2' position (Olejniczak et al, 2002; Olejniczak and Lesnikowski, 2002; Lesnikowski, 2003). Other type of modified nucleosides, nucleotides and oligonucleotides are derivatives of these compounds consisting metal complexes (Bigey et al., 1997; Dervan et al., 1988; Dougan et al., 1997; Dubey et al., 2000; Hurley and Tor, 1998; Ossipov et al, 2001; Strobel et al, 1988; Yu et al, 2000). Nucleoside conjugates containing metal complexes with ligands other than carborane cage or nucleosides containing carborane cage without metal ion have been described. For example, U.S. Patent No. 6,180,766 discloses nucleosides and oligonucleotides containing boron clusters. Application of these derivatives as boron carriers for BNCT of tumors, antisense biotherapeutics and molecular probes in medical diagnostics are proposed.

Application WO 2002053571 describes new derivatives of polycyclic hydrocarbons and naphthaleneimides type of ferrocene and methods of their synthesis, their intercalating properties, method for their electrochemical detection and several examples of practical applications. The disclosed compounds are easy for synthesis and purification, and are efficient intercalators towards double stranded DNA and RNA. However, as electrochemical label it binds to nucleic acid molecule through a weak intercalating interactions only what limits substantially practical utility of the proposed compounds.

The subject of JP 2001064298 is preparation of oligonucleotide/ferrocene conjugates and their application for electrochemical detection of DNA. Disclosed is a method for synthesis of conductively or electrochemically active oligonucleotides. Oligonucleotides are transformed into organic solvent soluble form *via* ion pair formation with suitable lipids and next are treated with a compound type of ferrocene monocarboxylic acid N-hydroxy succinimide ester. Though, in the above invention the method for the synthesis of oligonucleotide conjugates containing metal ion is described, it is limited to the incorporation into the oligonucleotide molecule iron only.

In the application US 5599796 the method and compounds for treating urogenital tumors, and particular, cancer of the prostate, bladder, and kidney with BCNT, are disclosed. Preferred boron carriers include 5-carboranyl-2'-deoxyuridine (CDU) and 5-*o*-carboranyl-1-(2-deoxy-2-fluoro- β -D-arabinofuranosyl)uracil (CFAU). Nucleosides and oligonucleotides bearing an -O-[(carboran-1-yl)alkyl]phosphate, S-[(carboran-1-yl)alkyl]phosphorothioate, or Se-[(carboran-1-yl)alkyl]phosphoroselenoate in place of the (carboran-1-yl)phosphonate moiety can be used.

Oligonucleotides of specific gene sequences that include one or more 3',5'-linking-(carboran-1-yl)phosphonate moieties can also be used in antisense therapy in the selective modification of gene expression. The therapy is accomplished by administering the boron-contg. compd. By any appropriate route, including by i.v. injection, oral delivery or by catheter or other direct means, in such a manner that the compd. accumulates in the target tumor. After desired accumulation of the compd. in the tumor, the site is irradiated with an effective amt. of low energy neutrons.

In the application JP 2002080491 preparation of electrophilic boron-containing nucleotide analogs and their intermediates is disclosed. Described nucleotide analogues of general structure $(\text{HO})_2\text{BX}[\text{P}(\text{O})(\text{O}-)\text{O}]_n\text{Nu}$ (I; Nu = nucleoside residue; X = CF_2 , CH_2 ; n = 2, 3) are prepared by difluoromethylation of $\text{HP}(\text{O})(\text{OR}_1)_2$ (R_1 = alkyl), reaction with $(\text{R}_2\text{O})_3\text{B}$ (R_2 = lower alkyl), deprotection, protection with 1,2-diols, coupling with nucleotides, and deprotection.

Application US5130302 is focused on boronated nucleoside, nucleotide and oligonucleotide compounds, compositions and methods for using same. A novel class of pharmaceutically active boronated nucleosides are provided. The nucleosides are boronated at a nitrogen of the purine or pyrimidine ring or analogues thereof. Also provided are phosphate esters of these nucleosides and oligomers thereof. Methods of making and using of the boronated nucleosides are also disclosed.

Both above applications provide method for the synthesis of some boron containing nucleoside derivatives, but does not allow incorporation of more than one boron atom per modification and does not permit incorporation into modified molecule other than boron metals.

Patent US5466679 discloses carboranyl uridines and their use in BNCT. The invention relates to novel boron-containing nucleosides and amino acids which can utilize the enzymatic systems in tumor cells for incorporating such boron-containing structures into nucleic acids and proteins. Subsequent use of boron neutron capture therapy provides a method for treatment of tumor cells. The subject of US5171849 is 2' and 3' carboranyl uridines and their diethyl ether adducts. There is disclosed a process for preparing carboranyl uridine nucleoside compounds and their diethyl ether adducts which exhibit a tenfold increase in boron content over prior art boron containing nucleoside compounds. Said carboranyl uridine nucleoside compounds exhibit enhanced lipophilicity and hydrophilic properties adequate to enable solvation in aqueous media for subsequent incorporation of said compounds in methods for boron neutron capture therapy in mammalian tumor cells. In the application US5405598 new sensitizing agents for use in boron neutron capture therapy are proposed. In spite that the compound proposed in the last three applications contain tenfold more

boron atoms than described earlier derivatives consisting boric acid residue, but still twofold less than proposed in the present invention. It should be also pointed out that any of the proposed so far methods does not allow incorporation into designed biomolecule such as nucleoside, nucleic acid or other a broad spectrum of different metal ions.

Nucleic acid hybridization technology essentially began with the work of Hall and Spiegelman (Hall and Spiegelman, 1961). Originally, probe and target were hybridized in solution and hybrids were isolated by equilibrium-density gradient centrifugation. This procedure was slow, labor-intensive and inaccurate. The substantial progress was achieved due to development the first solid phase hybridization method (Denhardt, 1966). Extensive studies during next 20 years yielded new, automated techniques for hybridization technology. The advancement was possible mainly due improved methods of DNA/RNA or DNA-oligonucleotide probes attachment to solid support and better methods of their labeling and detection (Keller and Manak, 1993). The most frequently used labels were radioactive isotopes such as ^{32}P , ^3H , ^{14}C and ^{35}S , fluorescent compounds and dyes absorbing in visible light, and the most often used test format was ELISA type immunoenzymatic assay. Comparison of selected methods are shown in Table 1.

Table 1. Comparison of sensitivity of selected methods used for DNA detection.

Enzyme	Label	Method of detection	Sensitivity
Alkaline phosphatase	4-nitrophenyl phosphate	Colorimetric	5 fmol
Alkaline phosphatase	NBT/BCIP	Colorimetric	0.5 fmol
Horse-radish peroxidase	o-Phenylenediamine	Colorimetric	0.1 fmol
Horse-radish peroxidase	Luminol	Colorimetric	1.0 fmol
Horse-radish peroxidase	ENH/LUM	Chemiluminescence	0.05 fmol
-	^{32}P	Scintillation	0.05 fmol
-	Fluorescein	Fluorimetry	500 fmol
-	Texas red	Fluorimetry	100 fmol
-	Rodamine	Fluorimetry	100 fmol
-	Isoluminol	Chemiluminescence	100 fmol
metal ions		Voltammetry	1 fmol ^b

^aUrdea MS., Warner BD., Running JA., Stempien M., Clyne J., Horn T., Nucl. Acids Res., 16, 4937 (1988); ^bThara T., Maruo Y., Takenaka S., Takagi M., Nucl. Acids Res., 24, 4273 (1996).

At the present stage of development of labeling and detection of oligonucleotide probes, and application of nucleic acids hybridization technology in research and medical diagnostics, an important role plays DNA chip technology and its application in genomics (Hoheisel, 1997; Anthony et al., 2001; Lipshutz et al., 1995).

Labelling of DNA-oligonucleotide probes with electrochemical labels and their application in electrochemical detection of nucleic acids was developed only recently. Earlier attempts to electrochemical detection of nucleic acids was based on detection of natural, unlabeled nucleic acid molecules (Wang et al., 1997; Steel et al., 1998; Singhal and Kuhr, 1997). Electrochemical detection of natural nucleic acids is however unspecific and characterized by high background. Because chemical similarities and alike nucleoside composition of oligonucleotide probes with different base sequences, selective detection of these probes without labelling is very difficult or impossible. In addition, detection of unlabeled oligonucleotide probes is hindered by the electrochemical activity of water. The ferrocene is one of a very few electrochemical labels proposed for labelling of oligonucleotide probes (Ihara et al., 1996; Ihara et al., 1997). In this case however, the label is limited to the iron and its redox characteristics.

Electrochemical detection of nucleic acids is disclosed in US 6391558 B1 patent dated May 21 2002. An electrochemical detection system which specifically detects selected nucleic acid segments is described. The system utilizes biological probes such as nucleic acid or peptide nucleic acid probes which are complementary and specifically hybridize with selected nucleic acid segments in order to generate a measurable current when an amperometric potential is applied. The electrochemical signal can be quantified. In the above invention electrochemical detection of only unlabeled nucleic acids and using only amperometric method is proposed.

Nucleic acid detection method and apparatus, and vessel for detecting nucleic acid are subject of US 20020064795 A1 patent dated May 30 2002. There is disclosed a nucleic acid detection app. including a nucleic acid immobilized electrode constituted by immobilizing a nucleic acid probe to a conductor, a plurality of vessels for bringing the nucleic acid probe into contact with a subject substance, a counter electrode disposed on a bottom surface or a inside surface of the vessel, and an elec. circuit for applying a voltage between the nucleic acid immobilized electrode and the counter electrode. A nucleic acid is detected by

inserting the nucleic acid immobilized electrode into each vessel contg. the subject substance, and using the counter electrode disposed on the bottom surface or inside surface of the vessel to elec. control reaction.

Quantitative detection of nucleic acids by differential hybridization using electrochemical labelling of samples is described in application JP 2002000299 A2 dated February 8 2002. A method for quant. detection of nucleic acids by differential hybridization using immobilized oligonucleotide probes complementary to nucleic acid samples labelled with conductive material, is disclosed. An elec. potential is applied and current is measured by differential pulse voltammog. DNA or PNA can be used as probes. Detection of ferrocene labelled oligonucleotides (20mer of adenine, A20) using thymine 20mer probe modified with mercapto hexyl group, or PNA thymine 10mer modified with 1,2-bis(vinylsulfonyl acetamide) ethane, is described. Hybridization-based gene detection method using intercalator and electrochemical detection of immobilized probe is described in application WO2002/057488 dated January 21, 2002. The gene detection method according to the above invention is characterized in that an intercalator is introduced into the double strand, and the double strand is electrochem. detected. According to present method, a probe and a sample gene are hybridized in a uniform soln. or in a soln. in the vicinity of an electrode surface, an intercalator is introduced thereinto for labeling, the probe is then immobilized on the electrode, and the amt. of immobilized probe and the amt. of double strand are sep. detected at the same time, making it possible to accurately det. the amt. in which the double strand is produced per unit amt. of immobilized probe. An advantage of the present method is that reactions can be conducted with higher efficiency because the hybridization and intercalation are carried out in a soln. without immobilizing the probe on the electrode.

Electrochemical detection of nucleic acid hybridization is a subject of application US6361951. A method of detecting a nucleic acid (e.g., DNA, RNA) that contains at least one preselected base (e.g., adenine, guanine, 6-mercaptoguanine, 8-oxo-guanine, and 8-oxo-adenine) comprises (a) reacting the nucleic acid with a transition metal complex capable of oxidizing the preselected base in an oxidation-reduction reaction; (b) detecting the oxidation-reduction reaction; and (c) determining the presence or absence of the nucleic acid from the detected oxidation-reduction reaction at the preselected base. The method may be used in a variety of applications, including DNA sequencing, diagnostic assays, and quantitative analysis.

The requirement of the presence of one or more preselected nucleic bases in the detected DNA or RNA is an significant disadvantage of the above method. If the nucleic bases are present in the probe in nature (e.g adenine, guanine) the specificity of the detection is decreased, if they are incorporated into nucleic acids artificially – the additional procedure is required making the process of detection more complicated. The range of the redox potentials is also limited due to the preselection of nucleic bases, this makes impossible detection of several probes in the same mixture.

Determination of sequence variations in nucleic acids by electrochemical detection of hybrids using probes labelled with redox groups is described in application WO 2001/007665 A2. The present invention is directed to methods and compns. for the use of self-assembled monolayers to electronically detect nucleic acids, particularly alterations such as nucleotide substitutions (mismatches) and single nucleotide polymorphisms (SNPs). The method uses arrays of probes immobilized in self-assembling monolayers on an electrode surface. Probes and target sequences are labelled with redox groups and hybridization of the probe results in a change in redox potential. Prepn. of chips with probes immobilized *via* disulfide bridges to thiolated DNA is demonstrated and the effects of variables such as temp of hybridization are studied. Methods of using competimers, perfectly matching probes that will replace weak or unstable hybrids, to improve the specificity of the hybridization are described.

Electrodes coated with metal complex-containing film and its use for electrochemical detection of nucleic acid bases is described in patent US 6180346 B1. A modified electrode prepd. by electropolymg. a film on the conductive working surface of an electrode is disclosed. The coated electrode is used for electrochem. detection of nucleic acid bases. The electrode is modified by reductive electropolymn. of a thin film of poly[Ru(vbpy)₃2+] or poly[Ru(vbpy)₃2+/vba] (vbpy=4-vinyl-4'methyl-2,2'-bipyridine and vba=p-vinylbenzoic acid) and the electrode is used for the electrochem. detection of aq. GMP, poly[G], and surface-immobilized single-stranded DNA probes. A DNA probe is attached covalently to the carboxylate group *via* a carbodiimide reaction followed by amidation of an amino-linked single-stranded DNA. In the presence of these guanine contg. moieties, a dramatic enhancement in the oxidative current for the Ru^{3+/2+} couple (present in the polymeric thin film) due to the catalytic oxidn. of guanine is obsd. This invention shows example some metal complexes for detection of DNA proves their practical importance.

Electrochemical detection of the hybridization process of a probe and target nucleic with the application of the catalyst of the redox reaction attached to electrode surface is described in the

application WO 2001/021635 A2. In the disclosed method DNA- or PNA-oligonucleotide probe containing catalyst of the redox reaction attached to the one end is linked to the electrode surface from the opposite side. Signal is generated as a result of hybridization of the target nucleic acid to the probe and formation of the double stranded structure allowing flow of the electric current from the electrode to the catalyst on the opposite end and initiation of the redox reaction. In this format hybridization process can be detected using voltametric, amperometric, potentiometric or conductometric methods. Electrochemical detection of the nucleic acid hybridization process with the application of the probes coupled with electrochemical label is a subject of the application WO 2000/031101 A1. This method can be successfully adapted for the application with the electrochemical labels being a subject of the present invention.

Method for detecting/quantitating sample nucleic acid fragment by scanning electrochemical microscopy is described in application JP 2001013103 A2 dated January 19 2001. A highly sensitive method is provided for detecting/quantitating a sample nucleic acid fragment complementary to DNA or PNA fragment immobilized on the anal. element surface by scanning electrochem. microscopy using DNA or PNA anal. element. The sample nucleic acid is contacted with the DNA or PNA anal. element in the presence of a hybrid DNA (or PNA)-binding electrochem. active mol. (e.g, ferrocene-modified intercalate). Then, the sample nucleic acid fragment complementary to the DNA or PNA fragment immobilized on the anal. element as well as the electrochem. active mol. are bound to the anal. element. The complementary sample nucleic acid fragment is detected/quantitated by measuring the elec. current generated in the electrochem. active mol.-binding region on the anal. element surface upon applying an elec. potential to the anal. element surface by a scanning electrochem. microscope. This method can be successfully adapted for the application with the electrochemical labels being a subject of the present invention.

An electrochemical method for detecting DNA using DNA sensor and intercalator is described in application JP 2000125865 A2 dated May 9 2000. The DNA sensor comprises more than two electrodes equipped with the resp. terminals for output. On the surface of the electrodes, DNA probes possessing the base sequences differing from each other are immobilized. On this DNA sensor, the sample DNA disscod. into single chains are bound with the probe DNA in the presence of the electrochem. activity-embedded-type intercalator, or the intercalator is bound with the DNA hybrid formed beforehand. A DNA gene is detected by measuring the current generated through the intercalator trapped into DNA hybrid. The above method is similar to several methods mentioned

earlier utilizing electrochemically labeled intercalating molecule. As a label for the intercalator molecule the electrochemical labels being a subject of the present invention can be also applied.

The aim of the present invention is providing new and versatile electrochemical label being a boron cluster, favourably in the form of metal complex, typically metallacarborane and modified nucleosides or their derivatives comprising thereof. Application of the metallacarborane group is especially desirable because it is characterized by unusually high boron contents, and it can accommodate in its structure a broad variety of other elements, specifically metals, their ions and isotopes. This increases substantially potential of the proposed new class of molecules beyond their use as electrochemical label and expand a range of their practical applications.

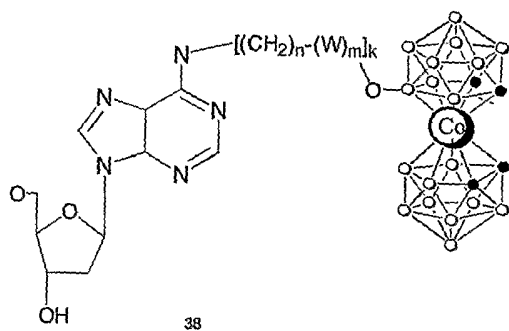
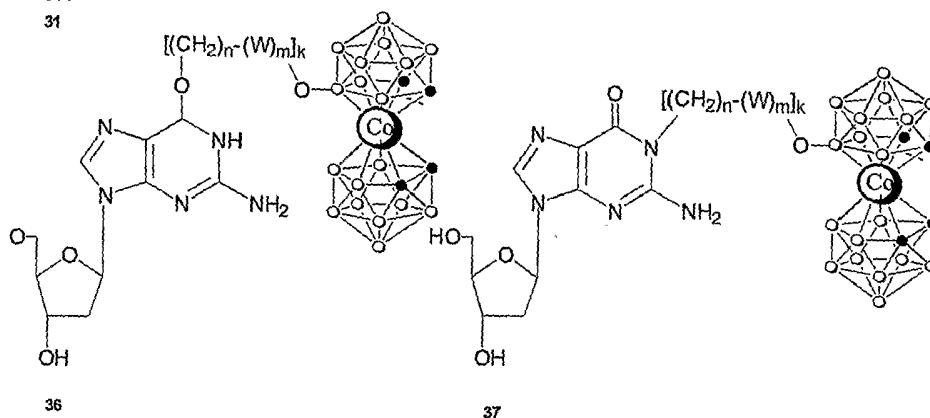
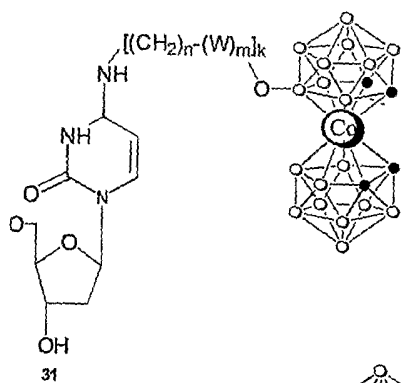
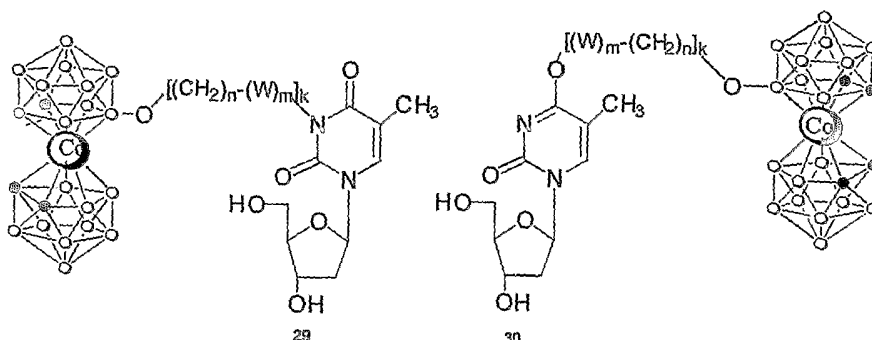
This aims have been unexpectedly achieved in the present invention.

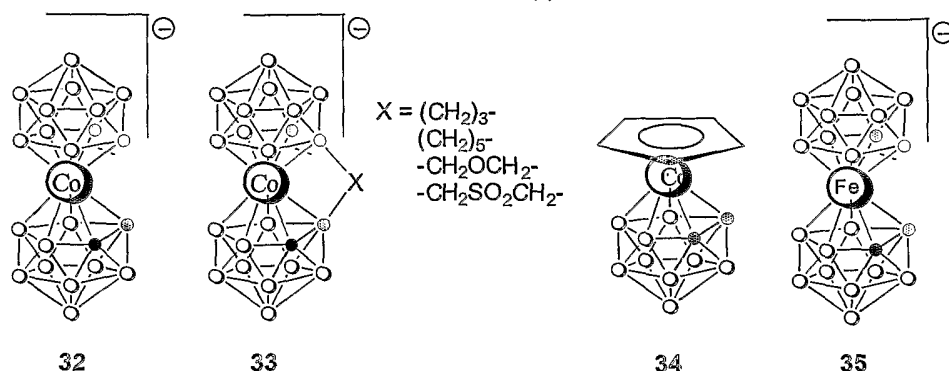
SUMMARY OF THE INVENTION

According to the invention, a nucleoside derivative is provided that contains metallacarborane complex attached to at least one of the following constituents of the nucleoside molecule: nucleic base, sugar residue, or analogue of sugar residue. Nonlimiting examples are the nucleosides of formulas 29-38, derivatives of all four key nucleosides: thymidine, 2'-deoxycytidine, 2'-deoxyguanosine and 2'-deoxyadenosine, illustrated below.

The base is preferably a purine or pyrimidine base or its derivative, and preferably is thymine, uracil, 5-halouracil, cytosine, 5-halocytosine, adenine, guanine, 2,6-diaminopurine, 2-amino-6-chloropurine, 2-aminopurine, 5-alkyluracil, 5-alkylocytosine, 2-thiouracil, 2,4-dithiouracil or 4-thiouracil. Preferably the metallacarborane group contains a metal atom (Me) selected among Sn, Mn, Tc, Re, Fe, Ru, Os, Co, Rh, Ir, Ni, Pd, Pt, Cu, Ag, Au, Sc, Cr, Mg, Zr, Mo, Sm, Yb, Hf, W, Hg, Gd, U or Y or nonmetal selected among As, S, Si, Se, Te, P, Sb, Bi, Ge or N, or theirs isotopes. Preferably the metallacarborane group is attached to the nucleoside directly or through a linker according to the formula $-(CH_2)_n-(W)_m]_k-$ wherein n is 0-5, m is 0 or 1, k is 1-6, and W is O, S, S(O), S(O)₂, Se, NR (wherein R = H, alkyl, haloalkyl, alkoxyalkyl or aryl), X-P(Z)(Y)O (wherein X = O, S, Se; Z = O, S, Se; Y = OH, SH, SeH or alkyl, haloalkyl, alkoxyalkyl, aryl or halogen, specifically fluorine, also CH=CH, CC, N=N, CHOH and CHN₃

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In one embodiment, a nucleoside derivative is provided that contains $-\{8-[(1,2\text{-dikaiba-}closo\text{-undekaborane})\text{-}3,3'\text{-Me-(1',2'\text{-dikaiba-}closo\text{-undekaborane})ate}]\}$ or $-\{8-[(1,2\text{-dikaiba-}closo\text{-undekaborano})\text{-}3\text{-Me-cyclopentadiene}ate]\}$ group. Preferably the metallocarborane group is a $\{(1,2\text{-dikaiba-}closo\text{-undekaborane})\text{-}3,3'\text{-cobalt-(1',2'\text{-dikaiba-}closo\text{-undekaborane})]ate\}$ (COSAN) (32) or $\{[(1,2\text{-dikaiba-}closo\text{-undekaborane})\text{-}3\text{-cobalt-cyklopentadiene}ate]\}$ (34). Nonlimiting examples are the metallocarborane groups of 32-35 illustrated above.

In one embodiment the pyrimidine nucleoside derivative provided in this invention contains $-\{diethylenoxy-\{8-[(1,2\text{-dikaiba-}closo\text{-undekaborane})\text{-}3,3'\text{-cobalt-(1',2'\text{-dikaiba-}closo\text{-undekaborane})an}]\}\}$ group preferably at position 2O, 4O or 3N of thymine or 2O or 4N of cytosine. In another embodiment the purine nucleoside derivative provided in this invention contains $-\{diethylenoxy-\{8-[(1,2\text{-dikaiba-}closo\text{-undekaborane})\text{-}3,3'\text{-cobalt-(1',2'\text{-dikaiba-}closo\text{-undekaborane})an}]\}\}$ group preferably at position 1N, 2N or 6O of guanine or 6N of adenine. In another embodiment one or more hydroxyl functions (-OH) of the sugar residue is replaced by $-\text{OP}(\text{Z})(\text{Y})\text{X}$ group, wherein: Z = O, S, Se; Y = OH, SH, SeH or alkyl, haloalkyl, alkoxyalkyl, aryl or halogene; X = OH, SH, SeH or alkyl, haloalkyl, alkoxyalkyl, aryl or halogene. In another embodiment one or more hydroxyl functions (-OH) of the sugar residue is replaced by $-\text{OP}(\text{Y})\text{X}$ group, wherein: Y = alkyl, haloalkyl, alkoxyalkyl (-O-alkyl), aryl, aryloxyaryl (-O-aryl), NR_2 or halogene; X = alkyl, haloalkyl, alkoxyalkyl (-O-alkyl), aryl, aryloxyaryl (-O-aryl), NR_2 or halogene. Also preferable the nucleoside derivative provided in this invention is nucleotide, preferably nucleoside mono-, di- or triphosphate. Preferably the free hydroxyl functions of sugar residue of natural or modified nucleoside as well as amino groups of nucleic base are protected as known to those skilled in the art, allowing use of the nucleoside for the synthesis of DNA- or RNA-oligonucleotides.

In another embodiment the metallacarborane group is attached to phosphorus atom of the nucleotide presented in this invention. Preferably the metallacarborane group is attached to the phosphorus directly or through a linker.

The subject of this invention are also compounds selected among following nonlimiting examples: 5'-*O*-monomethoxytrityl-3'-*O*-acetyl-3-*N*-(diethyleneoxy-8-COSAN)-thymidine (24a), 5'-*O*-monomethoxytrityl-3'-*O*-acetyl-4-*O*-(diethyleneoxy-8-COSAN)-thymidine (24b), 4-*O*-(diethyleneoxy-8-COSAN)-thymidine (28b), 8-dioxane-*O*-COSAN (23), 5'-*O*-monomethoxytrityl-4-*O*-(diethyleneoxy-8-COSAN)-thymidine (25a), 5'-*O*-monomethoxytrityl-3'-*O*-(*N,N*-diisopropyl-beta-cyanoethyl)-4-*O*-(diethyleneoxy-8-COSAN)thymidine amidophosphonate (26). The subject of the invention are also compounds 36-38 being nucleosides containing metallacarborane including cytosine, adenine or guanine.

The nucleosides containing metallacarborane group presented in this invention are natural nucleosides such as thymidine, deoxycytidine, deoxyadenosine, deoxyguanosine or theirs counterparts in ribo-series, or synthetic analogues of nucleosides modified within nucleic base or sugar residue such as L-nucleosides, carbocyclic nucleosides, acyclic nucleosides, and others. The above mentioned nucleoside analogues are examples only and do not limit the subject of the present invention. The metallacarboranes group can contain at least one or more metal ions or playing its function nonmetal atom.

According to other aspect of the invention, an oligonucleotide is provided that contains at least one metallacarborane complex attached to nucleic base, sugar residue or internucleotide linkage or analogue of sugar residue. Preferably the metallacarborane modification is attached to the 3'-terminal nucleoside unit, also preferably it is attached to the 5'-terminal unit, at 5'-end of the oligonucleotide chain or its middle location.

Preferably the metallacarborane group contains a metal atom (Me) selected among Sn, Mn, Tc, Re, Fe, Ru, Os, Co, Rh, Ir, Ni, Pd, Pt, Cu, Ag, Au, Sc, Cr, Mg, Zr, Mo, Sm, Yb, Hf, W, Hg, Gd, U or Y or nonmetal selected among As, S, Si, Se, Te, P, Sb, Bi, Ge or N, or theirs isotopes. Preferably the metallacarborane group is attached to the nucleoside directly or through a linker according to the formula $-(CH_2)_n-(W)_m)_k-$ wherein *n* is 0-5, *m* is 0 or 1, *k* is 1-6, and *W* is O, S, S(O), S(O)₂, Se, NR (wherein *R* = H, alkyl, haloalkyl, alkoxyalkyl or aryl), X-P(Z)(Y)O (wherein X = O, S, Se; Z = O, S, Se; Y = OH, SH, SeH or alkyl, haloalkyl, alkoxyalkyl, aryl or halogen, specifically fluorine, also CH=CH, CC, N=N, CHOH and CHN₃).

Preferably the metallacarborane modified oligonucleotide can hybridize *in vitro* and/or *in vivo* to complementary DNA or RNA forming double stranded structure, it can also preferably hybridize *in vivo* to the double stranded DNA.

In one embodiment, a oligonucleotide is provided that contains -{8-[(1,2-dikarba-closo-undekaborane)-3,3'-Me-(1',2'-dikarba-closo-undekaborane)ate]} or -{8-[(1,2-dikarba-closo-undekaborano)-3-Me-cyclopentadiene]ate} group. Preferably the metallacarborane group is a -{8-[(1,2-dikarba-closo-undekaborane)-3,3'-cobalt-(1',2'-dikarba-closo-undekaborane)ate]} or -{8-[(1,2-dikarba-closo-undekaborane)-3-cobalt-cyklopentadiene]ate}.

Preferably the modified oligonucleotide provided in this invention contains -{diethyleneoxy-{8-[(1,2-dikarba-closo-undekaborane)-3,3'-cobalt-(1',2'-dikarba-closo-undekaborane)ate]} group preferably at position 2O, 4O or 3N. Preferably the modifying metallacarborane moiety is attached at position 3N- or 4O- of thymidine (^{BCCo}T).

In one embodiment the modified oligonucleotide is 5'-d(^{BCCo}TGCTGGTTTGGCTG)-3'. This example is merely illustrative only, and not intended to limit the scope of the invention.

The oligonucleotides containing metallacarborane group presented in this invention are natural nucleic acids type of DNA i RNA as well as oligonucleotides modified within nucleic base, sugar residue and internucleotide linkage. The unlimiting examples of these oligonucleotides are phosphorothioate, methylphosphonate and peptide (PNA) oligomers containing at least one modification within the oligonucleotide chain.

According to other aspect of the present invention, a novel method is provided for the preparation of nucleoside derivative, particularly oligonucleotide, containing metallacarborane group. The disclosed method for the synthesis of nucleoside/metallacarborane conjugate is based on the use of suitably protected nucleoside or its analogue containing at least one carbonyl group able to participate in keto-imine equilibrium and/or amine or hydroxyl group in nucleic base. The nucleoside is then activated in the reaction with a proper activator type of inorganic base such as sodium hydride, sodium aluminum hydride, potassium tert-butoxide or organic base and treated with 8-dioxane-{8-[(1,2-dikarba-closo-undekaborane)-3,3'-Me-(1',2'-dikarba-closo-undekaborane)ate]} or 8-tetrahydrofurane-, or 8-tetrahydropyran-{8-[(1,2-dikarba-closo-undekaborane)-3,3'-Me-(1',2'-dikarba-closo-undekaborane)ate]}. The reaction medium is a hydrocarbon or its derivative characterized by the melting point from minus 100 up to plus 150 Celsius degree such as xylenes,

mesitylene, hexamethylbenzene or naphthalene. The desired product is isolated then 2', 3' and 5'-hydroxyl groups are deprotected into free hydroxyl (-OH) functions as desired (Example 1,2).

The subject of this invention is application of the carboranes and metallacarborane according to invention, as defined above, as a label for nucleosides, nucleotides, oligonucleotides, proteins, antibodies and other biological compounds and their derivatives.

Preferably the carborane and metallacarborane labels provided in this invention are detected by electrochemical methods. Preferably the compounds labelled with the carborane or metallacarborane group are oligonucleotides or their derivatives.

Preferably the oligonucleotides labelled as described in this invention will be used as hybridization probes.

According to other aspect of the present invention, oligonucleotides containing carborane or metallacarborane groups will be used as a material for nanoconstruction, construction of DNA microarrays and biosensors, especially electrochemical biosensors. Preferably compounds described in this invention are used as detecting element immobilized on the electrode surface.

The subject of this invention is also application of modified nucleotide according to the invention containing carborane and/or metallacarborane group, or its derivative, as defined above, in the methods of electrochemical detection. Preferably the derivative of the nucleotide is oligonucleotide.

In one embodiment the compounds described in the present invention are nucleosides, therefore also nucleotides and oligonucleotides labelled with carborane group. Compounds of this kind, and method of their synthesis are described in the documents referred to in the description of the background of the invention, namely US 6180766.

In the present invention an entirely new class of metallated nucleoside and nucleotide conjugates containing metallacarborane cage is provided, also oligonucleotides containing metallacarborane modification are proposed. The nucleoside can contain purine as well as pyrimidine nucleic base, and be natural or modified entity. The provided nucleoside conjugates contain metal in the form of metallacarborane complex and consist from two up to eighteen times more boron than it was available so far. Because of unique properties of boron clusters they form complexes with a broad range of metals such as Sn, Mn, Tc, Re, Fe, Ru, Os, Co, Rh, Ir, Ni, Pd, Pt, Cu, Ag, Au, Sc, Cr, Mg, Zr, Mo, Sm, Yb, Hf, W, Hg, Gd, U or Y and others and can include several nonmetals such as As, S, Si, Se, Te, P, Sb, Bi, Ge or N and their isotopes. (Grimes, 1995; Hall i inni, 2000; Hosmane i Maguire, 1987; Housecroft, 1990; Saxena i Hosmane, 1993).

Provided in the present invention nucleosides, nucleotides and oligonucleotides containing metallacarborane cage can be applied as among others as: modified primers in amplification of RNA and DNA, drugs, boron carriers for BNCT, radiopharmaceuticals bearing a range of isotopes useful in different types of radiotherapy, molecular probes, elements of biosensors, materials for nanotechnology and others.

In contrast to the sandwich type compounds such as ferrocene, the number of metallacarborane of different structures, containing various metals is very high. The electrochemical properties of many metallacarborane complexes are known. The chemical nature of the new class of label provided in this invention makes possible incorporation into the label molecule of a range of different metals such as Sn, Mn, Tc, Re, Fe, Ru, Os, Co, Rh, Ir, Ni, Pd, Pt, Cu, Ag, Au, Y and others. This property increases dramatically the range of potential practical applications of the oligonucleotides containing metallacarborane label. Because of a great diversity of metals which can be included into the metallacarborane structure and a broad range of redox potentials characteristic for these metals, it may be expected that the oligonucleotide probes labelled with different metallacarborane can be detected selectively in the mixture. This in turn, may be of importance for potential applications of metallacarborane labeled oligonucleotides in such technologies as nanotechnology, DNA-chips or DNA sensors.

BRIEF DESCRIPTION OF THE FIGURES

For the better presentation of the substance of this invention and enable its implementation the text of the application is supplemented with figures.

Figure 1. Synthesis of 5'-*O*-monomethoxytrityl-3'-*O*-acetyl-3-*N*-(diethylenoxy-8-COSAN)-thymidine (24a), 5'-*O*-monomethoxytrityl-3'-*O*-acetyl-4-*O*-(diethylenoxy-8-COSAN)-thymidine (24b) and 4-*O*-(diethylenoxy-8-COSAN)-thymidine (28b).

Figure 2. Synthesis of modified monomer for oligonucleotide preparation: 5'-*O*-monomethoxytrityl-4-*O*-(diethylenoxy-8-COSAN)thymidine 3'-*O*-(*N,N*-diisopropyl-beta-cyanoethyl)amidophosphonate (26).

Figure 3. UV spektrum of 4-*O*-(diethylenoxy-8-COSAN)thymidine (28b).

Figure 4. RP- HPLC trace of 4-*O*-(diethylenoxy-8-COSAN)thymidine (28b).

Figure 5. Mass spektrum (FAB) of 4-*O*-(diethylenoxy-8-COSAN)thymidine (28b).

Figure 6. Mass spectrum (FAB) of 5'-*O*-monomethoxytrityl-4-*O*-(diethylenoxy-8-COSAN)thymidine 3'-*O*-(*N,N*-diisopropyl-beta-cyanoethyl)amidophosphonate (26).

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Figure 7. Synthesis of tetradecanucleotide 5'-d(^{BEC}TGCTGGTTTGGCTG)-3' (27) complementary to the fragment of cytomegalovirus (HCMV) genome containing 4-*O*-(diethylenoxy-8-COSAN)thymidine (28b).

Figure 8. RP-HPLC trace for 2'-*O*-[*nido*-(*o*-carboran-1-yl)]methyluridine (31) ($R_t = 4, 6$ min), electrochemical detection at + 600 mV.

Figure 9. RP-HPLC trace for unmodified uridine 1 ($R_t = 2,4$ min), electrochemical detection at + 600 mV.

Figure 10. RP-HPLC trace for carborane containing oligonucleotide 5'-d(CGCTGGTTTGGC(U₂-CBM)G)-3' (19) ($R_t = 2, 05$ min), electrochemical detection at + 900 mV.

Figure 11. RP-HPLC trace for unmodified oligonucleotide 15 ($R_t = 2, 08$ min), electrochemical detection at + 900 mV.

Figure 12. RP-HPLC trace for 4-*O*-(diethylenoxy-8-COSAN)thymidine (28b) ($R_t = 4,86$ min), electrochemical detection at + 1700 mV.

Figure 13. RP-HPLC trace for unmodified thymidine ($R_t = 2,66$ min), electrochemical detection at + 1700 mV.

DETAILED DESCRIPTION OF THE INVENTION

The term alkyl, as used herein, unless otherwise specified, refers to a saturated straight, branched, or cyclic, primary, secondary, or tertiary hydrocarbon of C1 to C10, and specifically includes methyl, ethyl, propyl, isopropyl, butyl, isobutyl, *t*-butyl, pentyl, cyclopentyl, isopentyl, neopentyl, hexyl, isohexyl, cyclohexyl, cyclohexylmethyl, 3-methylpentyl, 2,2-dimethylbutyl, and 2,3-dimethylbutyl. The alkyl group can be optionally substituted with one or more moieties selected from the group consisting of hydroxyl, amino, alkylamino, arylamino, alkoxy, aryloxy, nitro, cyano, sulfonic acid, sulfate, phosphonic acid, phosphate, or phosphonate, either unprotected, or protected as necessary, as known to those skilled in the art, for example, as taught in Greene, et al., "Protective Groups in Organic Synthesis," John Wiley and Sons, Second Edition, 1991. The term lower alkyl, as used herein, and unless otherwise specified, refers to a C1 to C4 saturated straight or branched alkyl group.

The term alkylamino or arylamino refers to an amino group that has one or two alkyl or aryl substituents, respectively.

The term "protected" as used herein and unless otherwise defined refers to a group that is added to an oxygen, nitrogen, or phosphorus atom to prevent its further reaction or for other purposes. A wide

variety of oxygen and nitrogen protecting groups are known to those skilled in the art of organic synthesis.

The term aryl, as used herein, and unless otherwise specified, refers to phenyl, biphenyl, or naphthyl, and preferably phenyl. The aryl group can be optionally substituted with one or more moieties selected from the group consisting of hydroxyl, amino, alkylamino, arylamino, alkoxy, aryloxy, nitro, cyano, sulfonic acid, sulfate, phosphonic acid, phosphate, or phosphonate, either unprotected, or protected as necessary, as known to those skilled in the art, for example, as taught in Greene, et al., "Protective Groups in Organic Synthesis," John Wiley and Sons, Second Edition, 1991.

The term alkaryl or alkylaryl refers to an alkyl group with an aryl substituent.

The term aralkyl or arylalkyl refers to an aryl group with an alkyl substituent.

The term halo, as used herein, includes chloro, bromo, iodo, and fluoro.

The term purine or pyrimidine base includes, but is not limited to, adenine, N6 -alkylpurines, N6 -acylpurines (wherein acyl is C(O) (alkyl, aryl, alkylaryl, or arylalkyl), N6 -benzylpurine, N6 -halopurine, N6 -vinylpurine, N6 -acetylenic purine, N6 -acyl purine, N6 -hydroxyalkyl purine, N6 -thioalkyl purine, N2 -alkylpurines, N2 -alkyl-6-thiopurines, thymine, cytosine, 6-azapyrimidine, 2- and/or 4-thiopyrimidine, uracil, C5 -alkylpyrimidines, C5 -benzylpyrimidines, C5 -halopyrimidines, C5 -vinylpyrimidine, C5 -acetylenic pyrimidine, C5 -acyl pyrimidine, C5 -hydroxyalkyl purine, C5 -amidopyrimidine, C5 -cyanopyrimidine, C5 -nitropyrimidine, C5 -aminopyrimidine, N2 -alkylpurines, N2 -alkyl-6-thiopurines, 5-azacytidinyl, 5-azauracilyl, triazolopyridinyl, imidazolopyridinyl, pyrrolopyrimidinyl, pyrazolopyrimidinyl. Functional oxygen and nitrogen groups on the base can be protected as necessary or desired. Suitable protecting groups are well known to those skilled in the art, and include trimethylsilyl, dimethylhexylsilyl, t-butyl dimethylsilyl, and t-butyl diphenylsilyl, trityl, alkyl groups, acyl groups such as acetyl and propionyl, methylsulfonyl, and p-tolylsulfonyl.

The term heteroaryl or heteroaromatic, as used herein, refers to an aromatic moiety that includes at least one sulfur, oxygen, or nitrogen in the aromatic ring. Nonlimiting examples are furyl, pyridyl, pyrimidyl, thienyl, isothiazolyl, imidazolyl, tetrazolyl, pyrazinyl, benzofuranyl, benzothiophenyl, quinolyl, isoquinolyl, benzothienyl, isobenzofuryl, pyrazolyl, indolyl, isoindolyl, benzimidazolyl, purinyl, carbazolyl, oxazolyl, thiazolyl, isothiazolyl, 1,2,4-thiadiazolyl, isooxazolyl, pyrrolyl, quinazolinyl, pyridazinyl, pyrazinyl, cinnolinyl, phthalazinyl, quinoxalinyl, xanthinyl,

hypoxanthinyl, and pteridinyl. Functional oxygen and nitrogen groups on the heterocyclic base can be protected as necessary or desired. Suitable protecting groups are well known to those skilled in the art, and include trimethylsilyl, dimethylhexylsilyl, t-butyl dimethylsilyl, and t-butyl diphenylsilyl, trityl or substituted trityl, alkyl groups, acyl groups such as acetyl and propionyl, methylsulfonyl, and p-toluylsulfonyl.

The term alkenyl, as referred to herein, and unless otherwise specified, refers to a straight, branched, hydrocarbon of C2 to C10 with at least one double bond.

The term acyl refers to moiety of the formula $-C(O)R'$, wherein R' is alkyl; alkoxyalkyl including methoxymethyl; arylalkyl including benzyl; aryloxyalkyl such as phenoxymethyl; aryl including phenyl optionally substituted with halogen, C1 to C4 alkyl or C1 to C4 alkoxy, or the residue of an amino acid.

The term oligonucleotide refers to an oligomer of thirty-five or less nucleotides linked through their 3' and 5'-hydroxyl or 2'- and 5'-hydroxyl groups.

The term "metallacarboranes", as used herein, refers to compounds of metallocene type complexes consisting at least one carborane cage ligand and one or more metal atoms. For a review of metallacarborane chemistry see Grimes, R. N., 2000. Metallacarboranes in the new millennium, *Coord. Chem. Rev.*, 200-202, 773-811; Saxena, A.K., Hosmane, N.S., 1993. Recent advances in the chemistry of carborane metal complexes incorporating d- and f-block elements. *Chem. Rev.*, 93, 1081-1124; Grimes, R. N., Transition Metal Metallacarboranes in *Comprehensive Organometallic Chemistry*, 1995, vol. 1, Housecroft C. E., ed., Pergamon, New York, 373-430.

Derivative (of the nucleoside, nucleotide, oligonucleotide, protein, antibodies and other biological molecules) is a chemically or physically changed form of the mother molecule (substrate).

DNA hybridization technology - the technologies and their applications based on specific formation of double stranded nucleic acid structure due to specific nucleic bases recognition according to Watson-Crick rule.

DNA microarrays - terminologies that have been used in the literature to describe this technology include, but not limited to: biochip, DNA chip, DNA microarray, and gene array. *Affymetrix, Inc.* owns a registered trademark, *GeneChip®*, which refers to its high density, oligonucleotide-based DNA arrays. However, in some articles appeared in professional journals, popular magazines, and the WWW the term "gene chip(s)" has been used as a general terminology that refers to the microarray technology. Base-pairing (i.e., A-T and G-C for DNA; A-U and G-C for RNA) or

hybridization is the underlining principle of DNA microarray. An array is an orderly arrangement of samples. It provides a medium for matching known and unknown DNA samples based on base-pairing rules and automating the process of identifying the unknowns. In general, arrays are described as *macroarrays* or *microarrays*, the difference being the size of the sample spots. Macroarrays contain sample spot sizes of about 300 microns or larger and can be easily imaged by existing gel and blot scanners. The sample spot sizes in microarray are typically less than 200 microns in diameter and these arrays usually contains thousands of spots.

DNA sensor, is an immobilized DNA recognition element (receptor) coupled to a transducer unit and an electronic amplifier. Dependent on the kind of interaction between the analyte (S) and the receptor (R) one differentiates between: Affinity sensors (affinity to the substrate; $S + R \rightleftharpoons RS$), metabolism sensors (use of the substrate; $S + R \rightleftharpoons RS \rightleftharpoons P + R$).

Electrochemical DNA sensors, the device detecting hybridization process between target nucleic acid and nucleic acid probe. The DNA sensor comprises a nucleic acid recognition layer and a signal transducer. The signal transducer determine that that hybridization occurred on the base of the electrochemical process triggered by the hybridization and converts this into electronic signal

Nanotechnology, the development and use of devices that have a size of only a few nanometres. Research has been carried out into very small components, many of which depend on quantum effects and may involve movement of a very small number of electrons in their action. Such devices would act faster than larger components. Considerable interest has been shown in the production of structures on a molecular level by suitable sequences of chemical reactions or lithographic techniques. It is also possible to manipulate individual atoms on surfaces using a variant of the atomic force microscope to make, for example, high density data storage devices.

Nucleoside is defined as a compound consisting a five-carbon sugar molecule (a pentose, ribose or deoxyribose) and an organic base, purine or pyrimidine.

Nucleotide is defined as a nucleoside consisting phosphate group.

Example 1. *5'-O-Monomethoxytrityl-3'-O-acetyl-3-N-(diethyleneoxy-8-COSAN)thymidine* (24a) and *5'-O-monomethoxytrityl-3'-O-acetyl-4-O-(diethyleneoxy-8-COSAN)thymidine* (24b) (Figure 1). The procedure was performed under positive pressure of argon. *5'-O-monomethoxytrityl-3'-O-acetylthymidine* (22) (0.9 g, 1.6 mmol) and 8-dioxane-COSAN (2, 1.4 g, 3.3 mmol) were mixed together then dried under high vacuum over P_2O_5 for 24 h, next NaH (60% suspension in mineral oil, 80 mg, 3.3 mmole) was added followed by anhydrous toluene (18 mL). The reaction mixture

was stirred at 70°C in oil bath. After 8 h an excess of NaH was removed by centrifugation and supernatant added dropwise into hexane (135 mL). The resultant precipitate was separated by centrifugation and the sediment was dried under vacuum yielding 2.4 g of crude 24 as a mixture of 3-*N* and 4-*O* isomers (24a and 24b, respectively). Crude 24 was purified by silica gel column chromatography (70g silica gel, 230-400 mesh) using 12% CH₃OH in CHCl₃, containing 1% Et₃N as eluting solvent system. The yield of 24a was 221 mg and 24b was 450 mg. 24a: TLC (CH₃CN/CHCl₃, 1:2): R_f 0.14; UV (CH₃CN): λ_{min} 227.46, 249.18, 287.30 nm, λ_{max} 234.43, 261.89, 313.93 nm; ¹H-NMR (C₆D₆): 1.8-3.5 (bm, 21H, BH-COSAN), 1.58 (s, 3H, CH₃CO), 1.71 (s, 3H, CH₃-5), 2.54 (m, 2H-2'), 2.95-3.08 (m, 4H, 4 x CH-COSAN), 3.28- 3.41 (bs, 4H, 2 x OCH₂), 3.43 (s, 3H, CH₃O), 3.46-3.52 (q, 2H-5',5''), 3.92 (bs, 2H, OCH₂), 3.96 (s, 1H-4'), 4.48-4.59 (m, 2H, OCH₂), 5.36 (s, 1H-3'), 6.31 (s, -H arom. in 4-CH1H-1'), 6.87 (d, 2H, ₃OPh), 7.28-7.60 (m, 12H, H arom. in MMTr), 8.11 (s, 1H-6); ¹³C-NMR (C₆D₆): 13.62 (CH₃-5), 20.99 (CH₃ in CH₃CO), 39.76 (C-2'), 48.10, 51.94 (C-COSAN), 55.72 (CH₃O), 64.61 (C-5'), 68.73, 73.48 (OCH₂), 75.67 (C-3'), 86.41 (C-4'), 87.46 (C-1'), 88.74 (C-methylidene in MMTr), 114.65, 129.23, 131.40, 144.85, 145.06 (MMTr), 135.92 -C in 4-CH(C-6), 156.91 (C-2), 160.24 (₃OPh), 170.30 (CO in CH₃CO), 176.38 (C-4); ¹¹B-NMR (C₆D₆): 30.50-32.00 (bs, 18B); FAB-MS (-VE, NBA) 966.9 [M-1]. 24b: TLC (CH₃CN/CHCl₃, 1:2): R_f 0.41; UV (CH₃CN): λ_{min} 252.87 nm, λ_{max} 283.91 nm, 311.84 nm; ¹H-NMR (C₆D₆): 1.50-4.00 (bm, 21H, BH-COSAN), 1.48 (s, 3H, CH₃-5), 1.62 (s, 3H, CH₃CO), 2.55 (m, 2H-2'), 2.77-3.07 (m, 4H, 4 x CH-COSAN), 3.21 (bs, 4H, 2 x OCH₂), 3.37 (s, 3H, CH₃O), 3.45-3.51 (q, 2H-5',5''), J_{5',5''}=9.04), 3.88 (bs, 2H, OCH₂), 4.08 (s, 1H-4'), 4.37-4.43 (m, 2H, OCH₂), 5.43 (s, 1H-3'), 6.64 (s, -H arom. in CH1H-1'), 6.82 (d, 2H, ₃OPh), 7.06-7.52 (m, 12H, H arom. in MMTr), 8.20 (s, 1H-6); ¹³C-NMR (C₆D₆): 12.33 (CH₃-5), 21.09 (CH₃-acetyl), 40.22 (C-2'), 48.12, 51.85, 51.95 (C-COSAN), 55.60 (CH₃O), 64.43 (C-5'), 68.27, 71.68, 73.26 (OCH₂), 75.53 (C-3'), 85.97 (C-4'), 87.75 (C-1'), 88.56 (C-methylidene in MMTr), 114.55, 129.14, 129.23, 129.55, 131.41, 142.33, 144.94, 145.09 (MMTr), 135.95 (C-6), 158.93 (C-2), 160.21 (C-4 in 4-CH₃OPh), 170.58 (C-4), 172.52 (CH₃-acetyl); ¹¹B-NMR (C₆D₆): 30.50-32.00 (bs, 18B); FAB-MS (-VE, NBA) 966.9 [M-1].

Analogously other metallacarboranes can be incorporated into purine or pyrimidine nucleic base and sugar residue of the nucleoside unit.

Example 2. 4-*O*-(diethylenoxy-8-COSAN)thymidine (28) (Figure 1). 5'-*O*-Monomethoxytrityl-4-*O*-(diethylenoxy-8-COSAN)thymidine (25a) (0.1 g, 0.11 mmol) was dissolved in CH₃CN (5 mL) then

to the resultant solution acetic acid was added (80% CH₃COOH, 10 mL). After 2.5 h at room temperature the reaction went to the completion (TLC control, solvent system CHCl₃/CH₃OH, 8:2) and the solvents were evaporated under reduced pressure yielding crude 28 (0.18 g). Crude 28 was purified by silica gel column chromatography (2 g of silica gel, 230-400 mesh) using 30% CH₃OH in CHCl₃ as eluting solvent system. The yield of 28 was 40 mg (58%). TLC (CHCl₃/CH₃OH, 8:2): R_f 0,18; UV (CH₃CN): λ_{min} 237,93, λ_{max} 282,76, λ_{max} 310,67; ¹H-NMR (CD₃OD): 1.2-3.2 (bm, 21H, BH-COSAN), 2.02 (s, 3H, CH₃-5), 2.16-2.21 (m 1H-2'), 2.40-2.44 (m, 1H-2''), 3.60-3.65 (m, 4H, 2 × OCH₂), 3.73-3.76 (dd, 1H-5'), 3.82-3.85 (m, 3H, 1H-5'' and OCH₂), 3.96 (q, 1H-4'), 4.13 (s, 2H, 2 × CH-COSAN), 4.38-4.39 (m, 1H-3'), 4.42-4.46 (m, 1H, OCH₂), 4.52-4.56 (m, 1H, OCH₂), 6.26 (t, 1H-1', J_{1,2}=6.36), 8.13 (s, 1H-6); ¹³C-NMR (CD₃OD): 12.40 (CH₃-5), 42.24 (C-2'), 48.04 (C-COSAN), 55.15 (C-COSAN), 62.54 (C-5'), 68.13 (OCH₂), 69.83 (OCH₂), 69.98 (OCH₂), 71.69 (C-3'), 73.07 (OCH₂), 87.97 (C-1'), 89.11 (C-4'), 142.08 (C-6), 158.20 (C-2), 172.16 (C-4); ¹¹B-NMR (CD₃OD, H-decoupled): 28.54 (s), 10.30 (s), 6.33 (s), 3.48 (s), 1.32 (s), from -1,59 to -2,19 (d), -11,47 (s), -14,68 (s); FAB-MS (-VE, NBA) 652,6 [M-1].

Method for the automated production of oligonucleotides are described below (Example 3 and 4). Given the disclosure herein, one of ordinary skill in the art will know how to prepare a wide variety of oligonucleotides with metallacarborane-containing nucleotide monomers for a diverse range of applications, all of which are intended to fall within the scope of this invention.

Example 3. 5'-O-Monomethoxytrityl-4-O-(diethyleneoxy-8-COSAN)thymidine 3'-O-(N,N-diisopropyl-2-cyanoethyl)phosphoramidite (26) (Figure 2). All procedures were performed under positive pressure of argon. 5'-O-Monomethoxytrityl-4-O-(bisethoxy-8-COSAN)thymidine (25a) (0.1 g, 0.11 mmol) was dried under high vacuum over P₂O₅ for not less than 12 h, then was dissolved in CH₂Cl₂ (fresh distilled over CaH₂, 1.3 mL) and next N,N-diisopropylethylamine (0.075 mL) was added. To the resultant solution 2-cyanoethyl-N,N-diisopropylchlorophosphoramidite was added dropwise (0.072 mL, 0.32 mmole). After 4 h the reaction mixture was washed with H₂O (3 × 5 mL) then the organic layer was dried over MgSO₄ and solvent evaporated. The crude product 7 was obtained as colorless oil (0.11 g). Crude 7 was purified by silica gel column chromatography (6 g of silica gel, 230-400 mesh) using CH₃CN/CH₂Cl₂ (1:3) as eluting solvent system. The yield of 7 was 78 mg (65%). TLC (CH₃CN/CH₂Cl₂, 1:3): R_f 0.25, UV (anhydrous CH₃CN): λ_{min} 255.2 nm, λ_{max} 281.6 nm, λ_{max} 311.3 nm; ³¹P-NMR (C₆D₆): 149,00 and 149,82 (1:1), FAB-MS [-VE, NBA] 1177 [M + 2×Na]

Example 4. 4-*O*-(diethyleneoxy-8-COSAN)thymidine (^{B^{EC}}T)-containing tetradecanucleotides 5'-d(^{B^{EC}}T GCTGGTTTGGCTG)-3' (8) and unmodified oligonucleotide 5'-d(CGCTGGTTTGGCTG)-3' (9) (Figure 7). The natural oligonucleotide 9 and the modified oligonucleotide 8 were synthesized using a Beckman Oligo 1000 DNA synthesizer. Column loaded with controlled pore glass functionalized with 5'-*O*-dimethoxytrityl 2'-*O*-deoxyguanosine mol) were used as solid support. Suitable 5'-*O*-dimethoxytrityl-2'-*O*-deoxynucleoside 3'-(*N,N*-diisopropyl-beta-cyanoethyl)phosphoramidites (9, B =Gua, Cyt, Thy) were prepared as a 0.5 g/10 mL solution in anhydrous acetonitrile. Elongation of the oligomers with natural nucleotide was performed using a standard 0.2 μmol DNA synthesis beta-cyanoethyl cycle without changes in condensation time. Coupling of the 5'-terminal modified monomer 5'-*O*-monomethoxytrityl-4-*O*-(bisethoxy-8-COSAN)thymidine 3'-*O*-(*N,N*-diisopropyl-2-cyanoethyl)phosphoramidite (26) as well as oxidation step were performed manually (capping step was omitted). After thirteen coupling cycles, detritylation and washing with acetonitrile the column was detached from the DNA synthesizer and dried under high vacuum (5 min). Monomer 26 (20 mg, 0.02 mmol) was dissolved in anhydrous acetonitrile (120 μL) followed by addition of tetrazole (0.09 mL, 0.5 M, 0.045 mmol). A solution of activated 26 was applied to the column and the coupling reaction was performed for 30 min. The column was washed with anhydrous acetonitrile (2 × 5 mL) followed by drying under high vacuum. The oxidation step was performed using a *tert*-butyl hydroperoxide solution (0.5 M, 1 mL) for 2 min. followed by washing with acetonitrile (1 × 5 mL) and drying under high vacuum. Oligonucleotides were then cleaved from the support by 1 h incubation with concentrated aqueous ammonia solution (30%, 1 mL) at room temperature then base deprotection was achieved by incubation of resultant solution at 50°C for 2 h. The solution of crude 5'-*O*-monomethoxytrityl protected oligonucleotide 8 and 5'-*O*-dimethoxytrityl protected oligomer 9 was degassed with a stream of argon and evaporated to dryness under vacuum, then redissolved in water. Resultant solution of crude 5'-*O*-protected oligonucleotides 8 and 9 having 33 and 64 A₂₆₀ optical density units, respectively, were purified using HPLC C₁₈ reverse phase column (RP-HPLC) using conditions as follows: 20 min from 0 % B to 100 % B, 5 min 100 % B, 5 min from 100 % B to 0 % B. Fractions containing the desired product were collected, and the buffer was evaporated under vacuum. The residue was co-evaporated with 96% ethyl alcohol to remove triethylammonium bicarbonate (TEAB), then detritylation was performed using a 80% acetic acid (1.0 mL) at room temperature for 20 min. Next the acetic acid solution was evaporated to dryness under vacuum and the totally deprotected oligonucleotides were

purified by RP-HPLC using conditions as above. Buffer A contained 0.1 M TEAB (pH 7.0) in a mixture of acetonitrile and water (2:98, v/v), buffer B contained 0.1 M TEAB (pH 7.0) in a mixture of acetonitrile and water 60:40 (v/v) for modified oligonucleotide 8 (condition I) and 40:60 (v/v) for unmodified oligonucleotide 9 (condition II). Flow rate 1 mL/min. $\lambda = 266$ nm. Fractions containing the desired product were collected, and the buffer was evaporated under vacuum. The residue was co-evaporated with 96% ethyl alcohol to remove TEAB. Both oligonucleotides were stored as dry solid at -20°C . When needed, they were redissolved in water, stored as frozen solution, and relyophilized as soon as possible. Yield of purified modified oligonucleotides 8 was 9.1 A_{260} optical density units, and oligonucleotide 9 36.9 A_{260} optical density units, respectively. 8: UV (H_2O): $\lambda_{\min}=233.6$ nm, $\lambda_{\max}=263.1$ nm; RP-HPLC (condition I) $R_f=18.65$ min; 9: UV (H_2O): $\lambda_{\min}=230.0$ nm, $\lambda_{\max}=257.5$; RP-HPLC (condition II) $R_f=12.32$ min; MALDI-MS 4301 [M].

Example procedure for the detection of the nucleoside or oligonucleotide labelled with carborane or metallacarborane group.

Example 5. Electrochemical detection of 2'-O-*nido*-(*o*-carboran-1-yl)methyluridine (31).

The stock solution of 31 was prepared by dissolving of compound 31 (7,9 mg, 0,02 mmole) in the mixture of acetonitrile/water (40:60 v/v, 1,5 mL) containing ammonium acetate (0,05 M, pH 5,5), then an aliquot (1 μL) was taken from this solution and was diluted 10^3 -fold with the same as above mixture of solvents up to 1 mL, yielding solution containing 10 ng of compound 31 in 1,9 μL of solution. From the diluted as above solution an aliquot (1,9 μL) was taken and further diluted 10-folds up to 20 μL with a mixture acetonitrile/water (40:60 v/v) containing ammonium acetate (0,05 M, pH 5,5). A final concentration of 31 was 1,3 μM corresponding to 0,5 ng in 1 μL of solution. This was analyzed by RP-HPLC (Econosil RP C_{18} , column, 5 μm , 4,7 x 250 mm) using an electrochemical detection in the range of potentials from +100 mV to +1400 mV and from -100 mV to -600 mV. As eluent a mixture of acetonitrile/water (40:60 v/v) containing ammonium acetate (0,05 M, pH 5,5) was used. The elution time was 5 min at flow rate 1 mL/min. The analysis was performed at ambient temperature.

Example 6. Control. An approach to electrochemical detection unmodified uridine 1.

The stock solution of uridine (1) was prepared by dissolving 1 (1 mg, 4 μmole) in a mixture of acetonitrile/water (40:60 v/v, 1 mL) containing ammonium acetate (0,05 M, pH 5,5). From this solution an aliquot was withdrawn (1 μL) then was diluted 10^3 -fold with the same mixture of solvents up to 1 mL yielding solution containing 10 ng of compounds 1 in 10 μL . From that solution

an aliquot was taken (10 μ L) then was diluted 2-fold up to 20 μ L by addition of a mixture of acetonitrile/water (40:60 v/v) containing ammonium acetate (0,05 M, pH 5,5). The final solution containing 2,05 μ M of nucleoside 1 was analyzed by RP-HPLC (Econosil RP C₁₈ column, 5 μ m, 4,7 x 250 mm) using electrochemical detection, the potential range was from +100 mV to +1400 mV and from -100 mV to -600 mV. As an eluent a mixture of acetonitrile/water (40:60 v/v) containing ammonium acetate (0,05 M, pH 5,5) was used. Elution time was 5 min, and flow rate was 1 mL/min. The elution was performed at ambient temperature.

Example 7. Electrochemical detection of oligonucleotide 5'-d(CGCTGGTTTGGCU_{2'}-CBM_G)-3' (19) labelled with 2'-O-*nido*-(o-carboran-1-yl)methyl group.

The stock solution of oligonucleotide 19 was prepared by dissolving 0,05 ODU_{A260} (ca. 1,65 μ g, 0,37 nmole) of the oligomer in a mixture of acetonitrile/water (40:60 v/v) (0,5 mL) containing ammonium acetate (0,05 M, pH 5,5) yielding solution containing 3×10^{-4} ODU_{A260} (ca. 10 ng) of 19 in 3 μ L of solution. From this solution an aliquot (3 μ L) was taken and was diluted 7-fold up to 20 μ L by addition of a mixture acetonitrile/water (40:60 v/v) containing ammonium acetate (0,05 M, pH 5,5). The final solution containing 0,12 μ M of 19 was analyzed by RP-HPLC (Econosil RP C₁₈ column, 5 μ m, 4,7 x 250 mm) using electrochemical detection in the potential ranging from +100 mV to +1400 mV and from -100 mV to -600 mV. As eluent a mixture of acetonitrile/water (40:60 v/v) containing ammonium acetate (0,05 M, pH 5,5) was used. The eluting time was 5 min, flow rate was 1 mL/min. The analysis was performed at room temperature.

Example 8. Control. An approach to electrochemical detection of unlabeled oligonucleotide 5'-d(CGCTGGTTTGGCTG)-3' (15).

The stock solution of 15 was prepared by dissolving 0,05 ODU_{A260} (ca. 1,65 μ g, 0,38 nmole) of the oligomers 15 in a mixture of acetonitrile/water (40:60 v/v, 0,5 mL) containing ammonium acetate (0,05 M, pH 5,5) yielding solution containing 3×10^{-4} ODU_{A260} (ca. 10 ng, 2,3 pmole) of compound 15 in 3 μ L of the solution. From this solution an aliquot (3 μ L) was taken and was diluted 7-fold up to 20 μ L with a mixture of acetonitrile/water (40:60 v/v) containing ammonium acetate (0,05 M, pH 5,5). The final solution 0,12 μ M of 15 was analyzed by RP-HPLC (Econosil RP C₁₈ column, 5 μ m, 4,7 x 250 mm) using electrochemical detection in the potential range from +100 mV to +1400 mV and from -100 mV to -600 mV. As eluent a mixture of acetonitrile/water (40:60 v/v) containing ammonium acetate (0,05 M, pH 5,5) was used. The elution time was 5 min, the flow rate was 1 mL/min. The analysis was performed at room temperature.

Example 9. Electrochemical detection of the thymidine labelled with metallacarborane, 4-O-(bisethoxy-8-COSAN)thymidine (28).

The stock solution of compound 28 was obtained by dissolving 28 (0,88 mg, 0,001 mmole) in a mixture of acetonitrile/water (60:40 v/v, 1 mL) containing ammonium acetate (0,05 M, pH 5,5) yielding solution containing 880 ng of 28 in 1 mL of solution. From this solution an aliquot (1 μ L) was taken and was diluted 20-fold up 20 μ L with a mixture of acetonitrile/water (60:40 v/v) containing ammonium acetate (0,05 M, pH 5,5). The final solution containing 67,5 μ M of 28 was analyzed by RP-HPLC system (Econosil RP C₁₈ column, 5 μ m, 4,7 x 250 mm) equipped with electrochemical detector. The potential range was from +100 mV to +2000 mV and from -100 mV to -2000 mV. As eluent a mixture of acetonitrile/water (60:40 v/v) containing ammonium acetate (0,05 M, pH 5,5) was used. The eluting time was 10 min, flow rate 1 mL/min. The analysis was performed at room temperature.

Example 10. Control. An approach to electrochemical detection of unlabeled thymidine

The stock solution of thymidine was prepared by dissolving thymidine (1 mg, 4,4 μ mola) in a mixture of acetonitrile/water (60:40 v/v, 1 mL) containing ammonium acetate (0,05 M, pH 5,5) yielding a solution containing 1 μ g of thymidine in 1 μ L of solution. From this solution an aliquot was taken (1 μ L) and was diluted up to 1 mL with the same mixture of solvents yielding solution containing 10 ng of thymidine in 10 μ L of solution, next this solution was further diluted 2-fold up to 20 μ L with same mixture of solvents. The final solution containing 2,2 μ M of thymidine was analyzed RP-HPLC (Econosil RP C₁₈ column, 5 μ m, 4,7 x 250 mm) using electrochemical detection in the range of the potentials from +100 mV to +2000 mV and from -100 mV to -2000 mV. As eluent a mixture of acetonitrile/water (60:40 v/v) containing ammonium acetate (0,05 M, pH 5,5) was used. Elution time was 10 min, flow rate was 1 mL/min. The analysis was performed at room temperature.

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CLAIMS

1. An nucleoside that contains at least one metallacarborane group attached to at least one of nucleoside constituents: nucleic base, sugar residue or analogue of sugar residue.
2. The nucleoside of claim 1, wherein a nucleic base is purine, pyrimidine or its derivative.
3. The nucleoside of claim 2, wherein the nucleic base is thymine, uracil, 5-halouracil, cytosine, 5-halocytosine, adenine, guanine, 2,6-diaminopurine, 2-amino-6-chloropurine, 2-aminopurine, 5-alkyluracil, 5-alkylcytosine, 2-thiouracil, 2,4-dithiouracil or 4-thiouracil.
4. The nucleoside of claim 1, wherein metallacarborane group contains a metal atom (Me) selected from Sn, Mn, Tc, Re, Fe, Ru, Os, Co, Rh, Ir, Ni, Pd, Pt, Cu, Ag, Au, Sc, Cr, Mg, Zr, Mo, Sm, Yb, Hf, W, Hg, Gd, U and Y or nonmetal selected among As, S, Si, Se, Te, P, Sb, Bi, Ge oraz N, or theirs isotopes.
5. Nucleoside of claims 1-4, wherein the metallacarborane group is attached to nucleoside molecule directly or through a linker of structure preferably $-(CH_2)_n-(W)_m-k-$, wherein n is 0-5, m is 0 or 1, k is 1-6, while W is O, S, S(O), S(O)₂, Se, NR (wherein $R = H$, alkyl, haloalkyl, alkoxyalkyl, or aryl), X-P(Z)(Y)O (wherein $X = O, S, Se$; $Z = O, S, Se$; $Y = OH, SH, SeH$ or alkyl, haloalkyl, alkoxyalkyl, aryl, or haloegene preferably fluorin) or $CH=CH$, CC , $N=N$, $CHOH$ and CHN_3 .
6. Nucleoside of claim 2 wherein metallacarborane group is preferably $- \{8-[(1,2-dikarba-closo-undekaborane)-3,3'-Me-(1',2'-dikarba-closo-undekaborane)ate]\}$ or $- \{8-[(1,2-dikarba-closo-undekaborane)-3-Me-cyklopentadiene]ate\}$.
7. Nucleoside of claim 5, wherein metallacarborane group is $- \{8-[(1,2-dikarba-closo-undekaborane)-3,3'-cobalt-(1',2'-dikarba-closo-undekaborane)ate]\}$ or $- \{8-[(1,2-dikarba-closo-undekaborane)-3-kobalt-cyklopentadiene]ate\}$.
8. Nucleoside of claim 5 containing preferably 3N- or 4-O- $\{diethyleneoxy-\{8-[(1,2-dikarba-closo-undekaborane)-3,3'-cobalt-(1',2'-dikarba-closo-undekaborane)ate]\}$ group.
9. Nucleoside of claim 5 wherein, at least one hydroxyl function (-OH) of sugar residue is replaced by group -OP(Z)(Y)X ($Z = O, S, Se$; $Y = OH, SH, SeH$ or alkyl, haloalkyl, alkoxyalkyl, aryl or halogen; $X = OH, SH, SeH$ or alkyl, haloalkyl, alkoxyalkyl, aryl or halogen) or -OP(Y)X group [$Y = alkyl, haloalkyl, alkoxyalkyl (-Oalkyl), aryl, aryloxyl (-O-aryl), NR_2$ or halogen; $X = alkyl, haloalkyl, alkoxyalkyl (-O-alkyl), aryl, aryloxyl (-O-aryl), NR_2$ or halogen]

10. Nucleoside derivative of one of previous³¹ claims wherein it is nucleotide, preferably nucleoside mono-, di- or triphosphate.
11. Nucleotide of claim 10 wherein, the free hydroxyl functions (–OH) of sugar residue or its analogue and amine groups of nucleic base are protected with suitable protecting groups allowing use of this nucleotide in the synthesis of DNA- and RNA-oligonucleotides.
12. Nucleoside derivative of claim 10, wherein metallacarborane group is attached directly or through linker to phosphorus atom.
13. Compound selected from: 5'-O-monomethoxytrityl-3'-O-acetyl-3-N-(diethylenoxy-8-COSAN)-thymidine (24a), 5'-O-monomethoxytrityl-3'-O-acetyl-4-O-(diethylenoxy-8-COSAN)-thymidine (24b), 4-O-(diethylenoxy-8-COSAN)-thymidine (28b), 8-dioksane-O-COSAN (23), 5'-O-monomethoxytrityl-4-O-(diethylenoxy-8-COSAN)-thymidine (25a), 5'-O-monomethoxytrityl-4-O-(diethylenoxy-8-COSAN)thymidine 3'-O-(N,N-diisopropyl-beta-cyanoethyl)amidophosphate (26).
14. An oligonucleotide that contains at least one metallacarborane group attached to nucleic base, internucleotide linkage or sugar residue.
15. The oligonucleotide of claim 14 wherein, metallacarborane group is attached to 3'-terminal nucleotide or is located at 3'-fragment of the oligonucleotide chain.
16. The oligonucleotide of claim 14 wherein, metallacarborane group is attached to 5'-terminal nucleotide or is located within the 5'-fragment of the oligonucleotide chain.
17. The oligonucleotide of claim 14, wherein the metallacarborane group is located in the internal section of the oligonucleotide.
18. The oligonucleotide of claim 14 wherein, metallacarborane group contains a metal (Me) selected from Sn, Mn, Tc, Re, Fe, Ru, Os, Co, Rh, Ir, Ni, Pd, Pt, Cu, Ag, Au, Sc, Cr, Mg, Zr, Mo, Sm, Yb, Hf, W, Hg, Gd, U and Y or nonmetal selected from As, S, Si, Se, Te, P, Sb, Bi, Ge oraz N, or theirs isotopes.
19. The oligonucleotide of claim 14 wherein, metallacarborane group is attached to the nucleotide directly or through a linker preferably of formula $-\text{[(CH}_2\text{)}_n\text{-(W)}_m\text{]}_k-$ wherein n is 0-5, m is 0 or 1, k is 1-6, while W is O, S, S(O), S(O)₂, Se, NR (wherein R = H, alkyl, haloalkyl, aloxyalkyl, or aryl), X-P(Z)(Y)O (wherein X = O, S, Se; Z = O, S, Se; Y = OH, SH, SeH or alkyl, haloalkyl, aloxyalkyl, aryl, or haloegen preferably fluorin) or CH=CH, CC, N=N, CHOH and CHN₃.

20. The oligonucleotide of claim 14 capable³² of hybridization in vitro and/or in vivo to a complementary DNA or RNA strand.
21. The oligonucleotide of claim 14 that is capable of hybridization to a double stranded DNA.
22. The oligonucleotide of claim 14 wherein the metallacarborane group is $-\{8-[(1,2\text{-dika}\text{rba-}closo\text{-undekaborane})\text{-}3,3'\text{-Me-(1',2'-dika}\text{rba-}closo\text{-undekaborane})\text{ate}]\}$ or $-\{8-[(1,2\text{-dika}\text{rba-}closo\text{-undekaborane})\text{-}3\text{-Me-cyklopentadiene}]\text{ate}\}$.
23. The oligonucleotide of claim 14 wherein, the metallacarborane group is $-\{8-[(1,2\text{-dika}\text{rba-}closo\text{-undekaborane})\text{-}3,3'\text{-cobalt-(1',2'-dika}\text{rba-}closo\text{-undekaborane})\text{ate}]\}$ or $-\{8-[(1,2\text{-dika}\text{rba-}closo\text{-undekaborane})\text{-}3\text{-cobalt-cyklopentadiene}]\text{ate}\}$.
24. The oligonucleotide of claim 14 containing preferably $\{\text{diethyleneoxy-}\{8-[(1,2\text{-dika}\text{rba-}closo\text{-undekaborane})\text{-}3,3'\text{-cobalt-(1',2'-dika}\text{rba-}closo\text{-undekaborane})\text{ate}]\}\}$ group attached to 2O, 4O or 3N of pyrimidine.
25. The oligonucleotide of claim 14 containing 3N- lub 4-O-(dietylenoksy-8-COSAN)thymidine (TBCCo) wherein its sequence is 5'-d(TBCCoTGCTGGTTTGGCTG)-3'.
26. A method for the synthesis of nucleoside of claim 1-4 wherein, the nucleoside or its analogue with suitably protected hydroxyl functions and containing at least one carbonyl group of nucleic base capable to participation in keto-imine equilibrium and/or amine or hydroxyl group of nucleic base is treated with 8-dioxane- $\{8-[(1,2\text{-dika}\text{rba-}closo\text{-undekaborane})\text{-}3,3'\text{-Me-(1',2'-dika}\text{rba-}closo\text{-undekaborane})\text{ate}]\}$, or with z 8-tetrahydrofuran- $\{8-[(1,2\text{-dika}\text{rba-}closo\text{-undekaborane})\text{-}3,3'\text{-Me-(1',2'-dika}\text{rba-}closo\text{-undekaborane})\text{ate}]\}$ in anhydrous medium and is activated with a strong inorganic or organic base type of DBU, BDDDP. A reaction medium is selected from hydrocarbons or theirs derivatives characterized by the melting point in the range from -100 up to 150 degree Celsius and is performed at the temperature in the range from 0 up to 150 degree Celcius, the the resultant product is deprotected leaving free 3' or 5' or both 3' and 5' hydroxyl functions.
27. The bases of claim 26 wherein it is sodium hydride, sodium aluminum hydride or sodium tert-butoxide.
28. The method of claim 26 wherein the reaction medium is toluene, xylenes, mesitilene, hexamethylbenzene or naphthalene.
29. The method of claim 26 wherein, the metal (Me) of metallacarborane group is selected from Sn, Mn, Tc, Re, Fe, Ru, Os, Co, Rh, Ir, Ni, Pd, Pt, Cu, Ag, Au, Sc, Cr, Mg, Zr, Mo, Sm, Yb, Hf, W, Hg, Gd, U and Y or nonmetals As, S, Si, Se, Te, P, Sb, Bi, Ge oraz N, or theirs isotopes.

30. The method of claim 26 wherein the ³³nucleic base is purine, pyrimidine or its derivative.
31. The method of claim 26 wherein, the nucleic base is thymine, uracil, 5-halouracil, cytosine, 5-halocytosine, adenine, guanine, 2,6-diaminopurine, 2-amino-6-chloropurine, 2-aminopurine, 5-alkyluracil, 5-alkylcytosine, 2-thiouracil, 2,4-dithiouracil or 4-thiouracil.
32. The method of claim 26 wherein the nucleoside is phosphorylated or phosphitylated and/or it is incorporated into oligonucleotide.
33. The application of carboranes and metallacarboranes as labels for nucleosides, nucleotides, oligonucleotides, proteins, antibodies or other biological compounds and their derivatives.
34. The application of claim 33 wherein the labelled compounds are detected by electrochemical methods.
35. The application of claim 33 or 34 wherein the labelled compounds are oligonucleotides and their derivatives.
36. The application of claim 35 wherein the labelled oligonucleotides are used as molecular probes.
37. The application of claim 36 wherein that labelled oligonucleotides are used as a material for nanotechnology, construction of microarrays or DNA sensors, especially electrochemical sensors.
38. Application of claim from 32 to claim 37 wherein that the labelled compounds are immobilized on the electrode.
39. Application of modified nucleotide containing carborane or metallacarborane, or its derivative in electrochemical methods of detection.
40. Application of claim 39 wherein the derivative of modified nucleotide is oligonucleotide of claim from 14 to 25.
41. Application of claim 39 wherein the labelled nucleotide is derivative of nucleoside of claim from 1 to 13.

1 /13

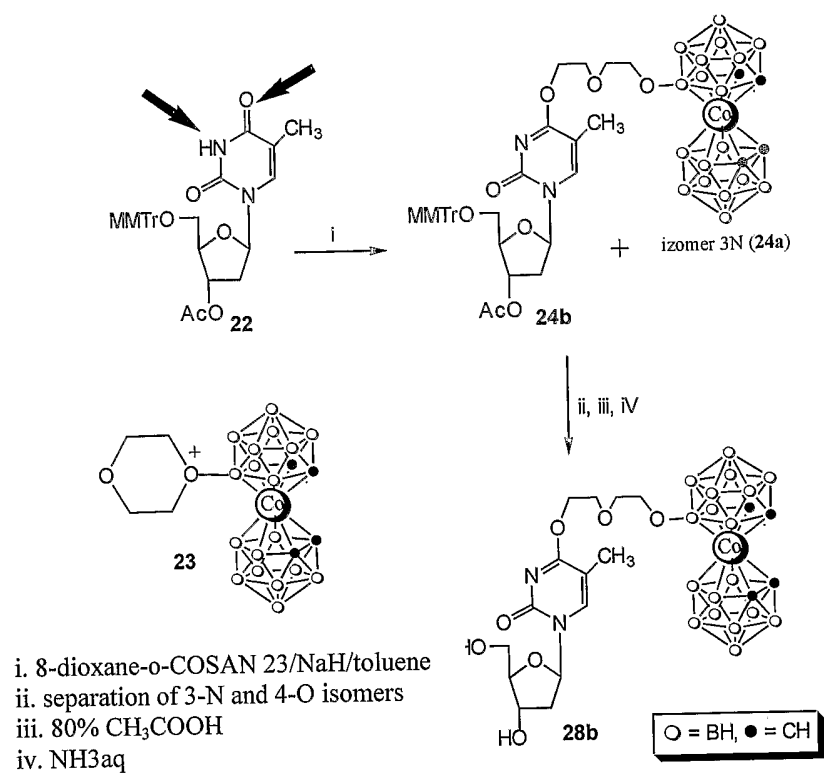


Fig. 1

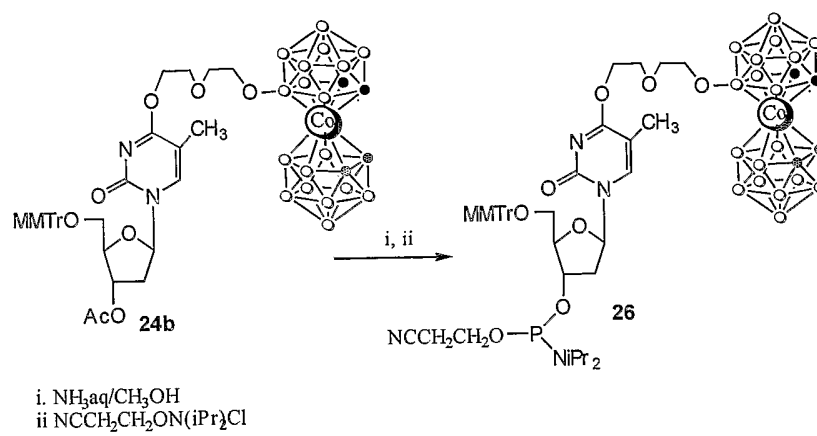


Fig. 2

3 /13

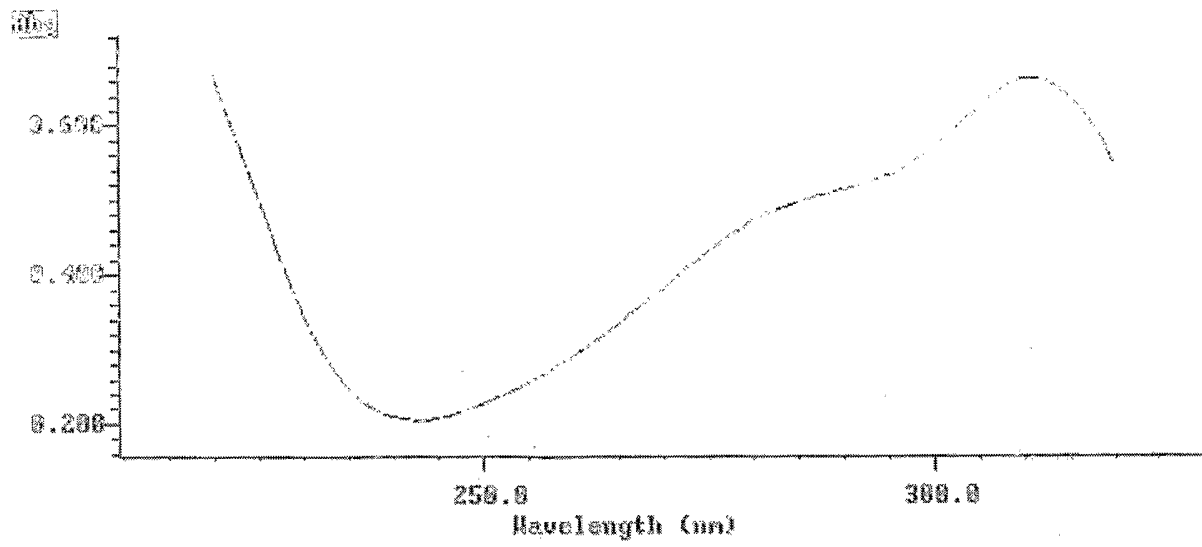


Fig. 3

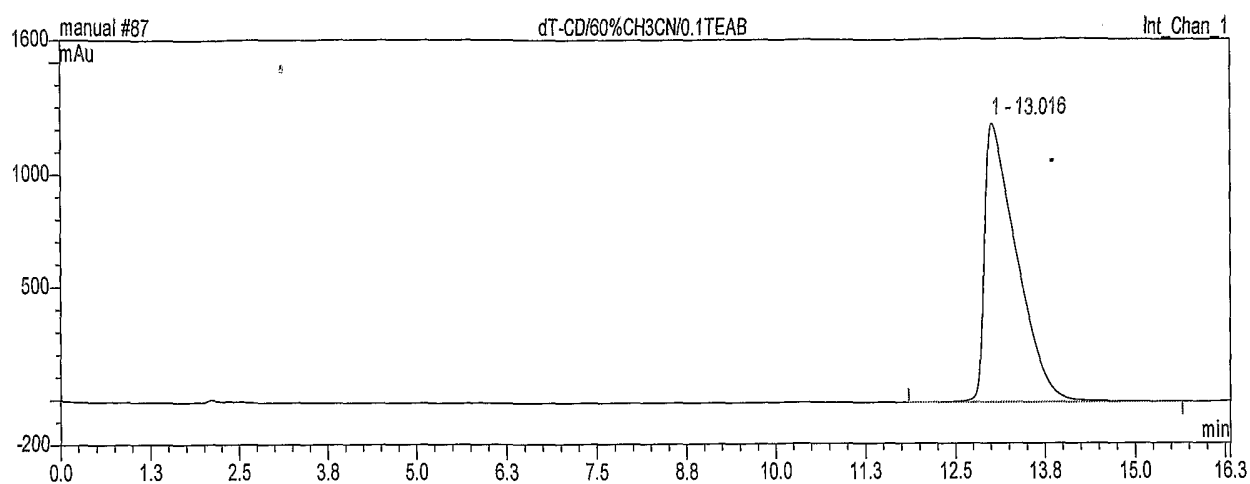


Fig. 4

5 /13

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Model: MAB -VE -LHR DECAY (EXP) OF LR MGI Study: MS CBM11 PAN Ledz
Oper: ja Client: CH10 PAN Ledz Inlet:
Pace: 133.0 Inten: 38403 Masses: 100 - 1000
Norm: 133.0 RJC: 548273 Peaks: 750
Data: 4/11/20

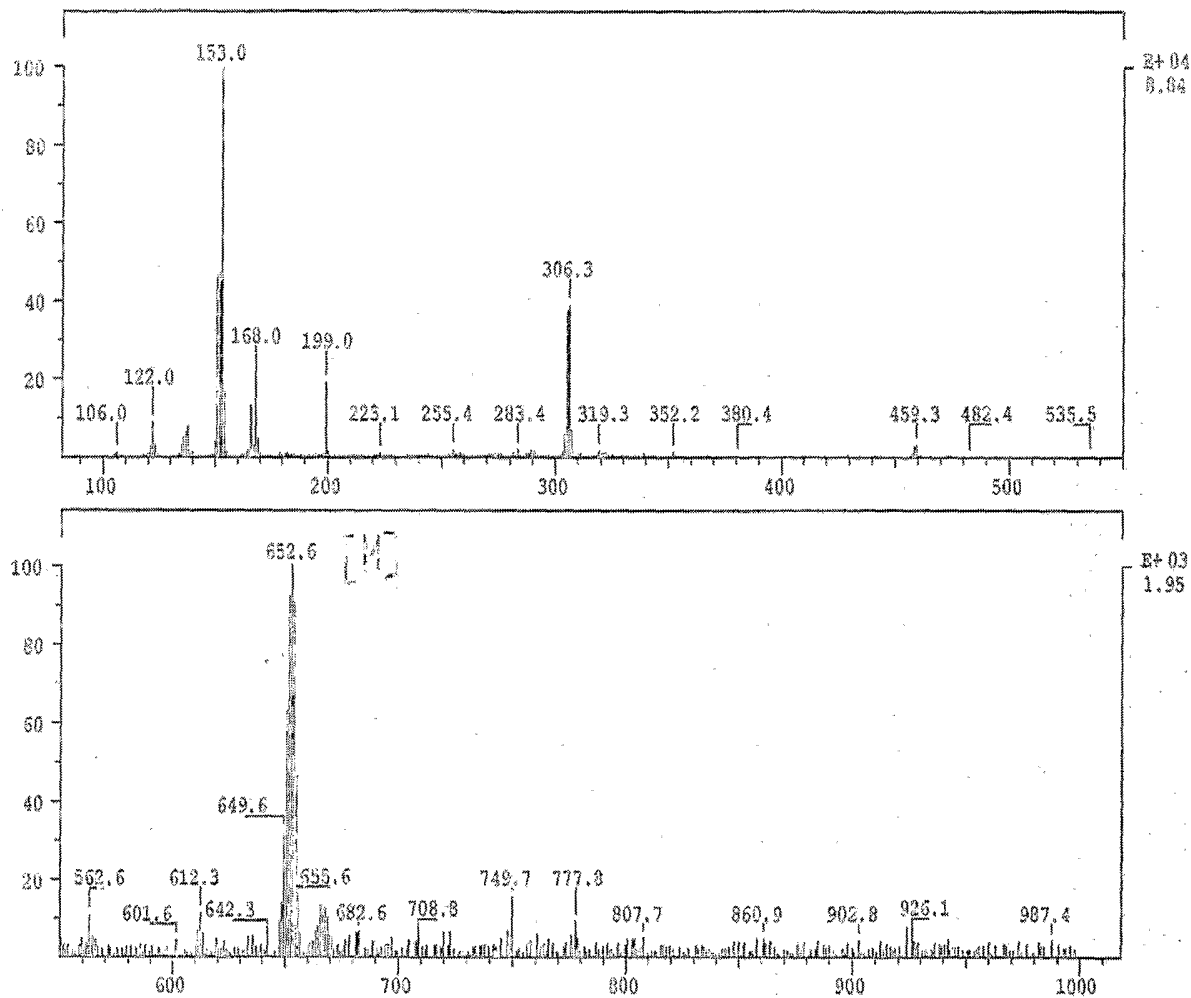


Fig. 5

6 /13

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Oper: jg Client: CRMIM PAN Lodz Inlet :
Base: 153.1 Inten : 190452 Masses: 100 > 1300
Norm: 153.1 RIC : 715502 Peaks: 139
Peak: 1000.00 amu
Data: 411:20

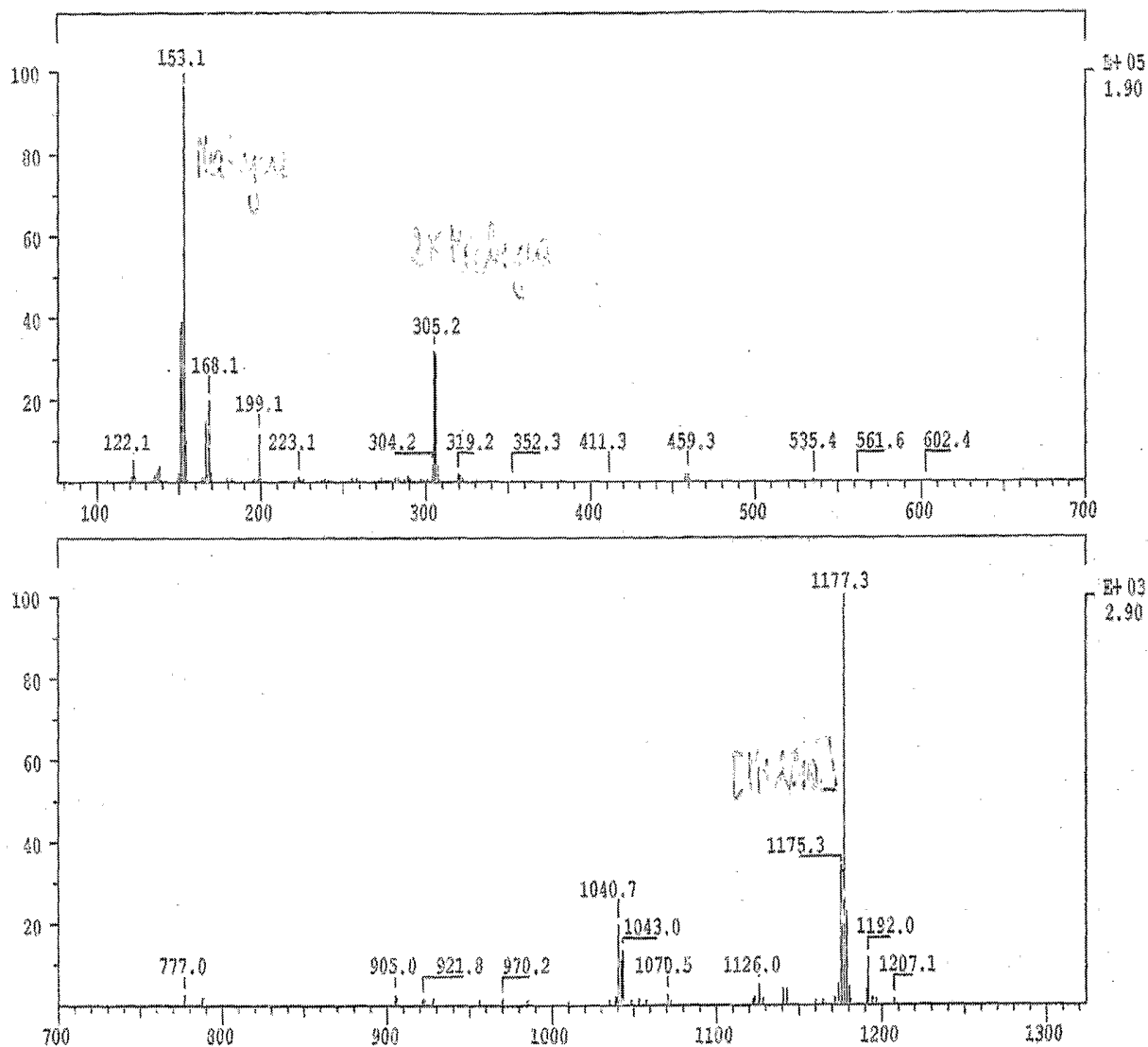
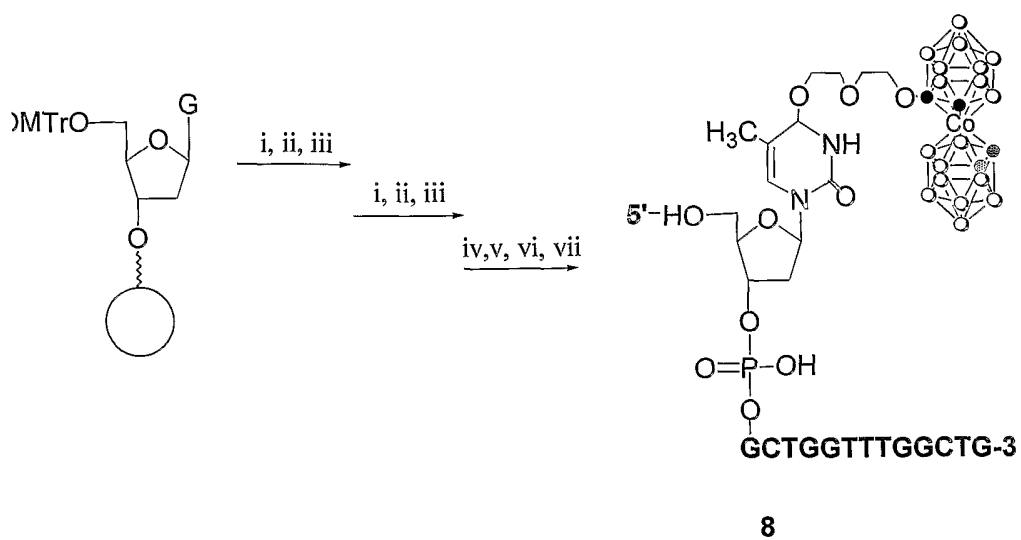


Fig. 6

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- . Detritilation: 3% DCA in CH_2Cl_2
- i. Coupling: unmodified monomers
- ii. Oxidation: 0.1 M I_2 in THF/pyridine/ H_2O (13:6:1), and capping: 1 M $(\text{CH}_3\text{CO})_2\text{O}$ in THF/pyridine (1:8) and 0.5 M DMAP in THF.
- v. Coupling: modified monomer 7.
- . Oxidation: 0.5 M t-BuOOH in dodecane
- i. Cleavage from the support: 30% NH_3 aq., 1 h, rt.
- ii. Base deprotection: 30% NH_3 aq., 2 h, 50°C .

Fig. 7

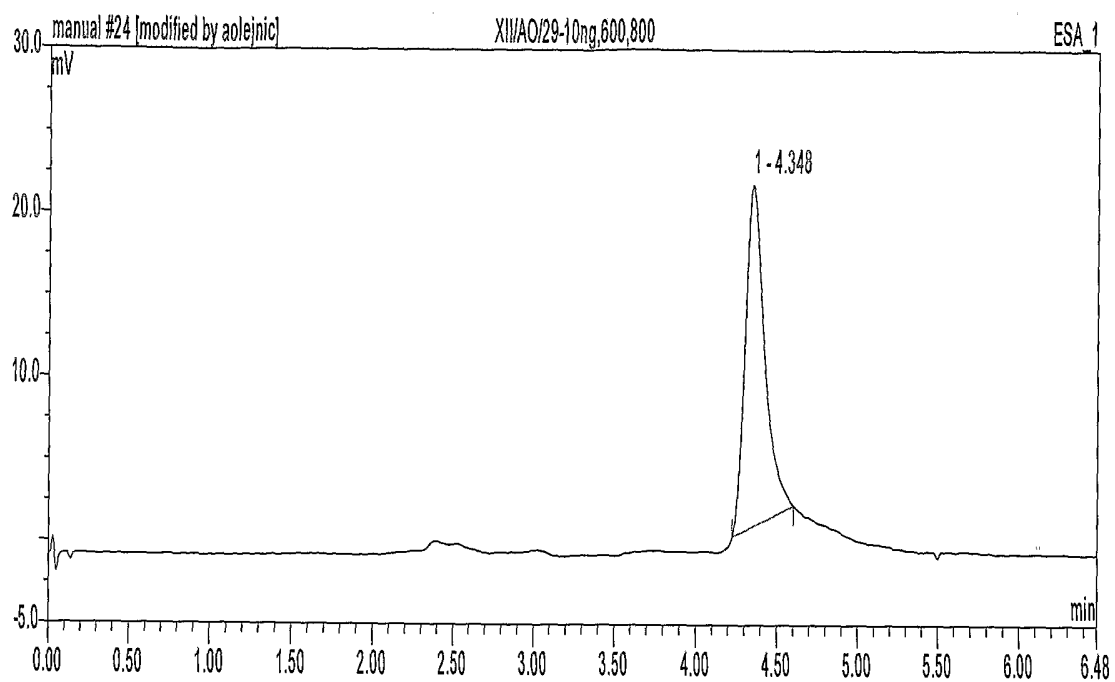


Fig. 8

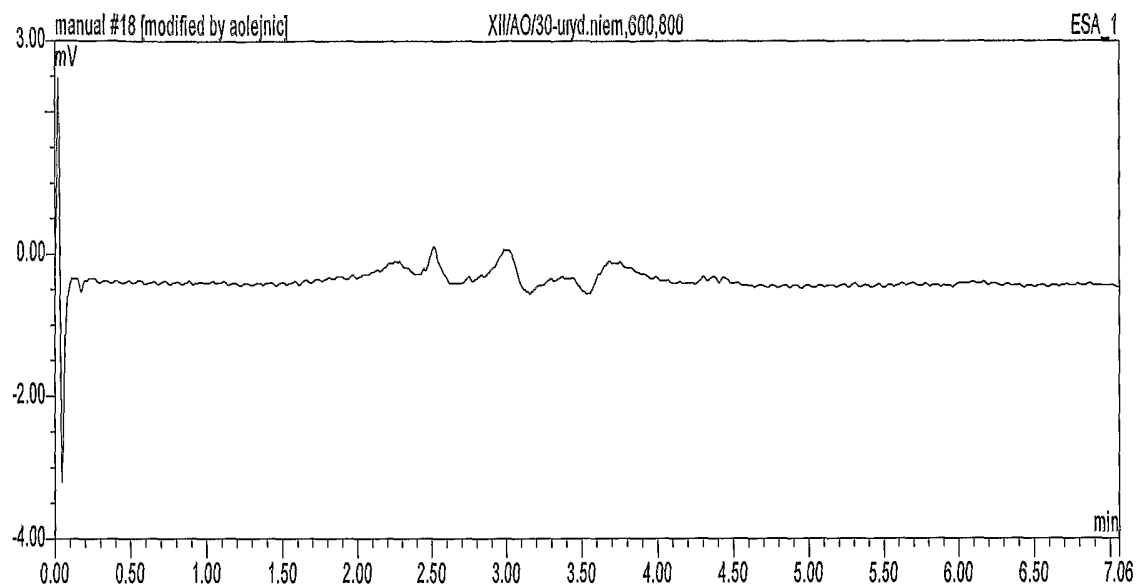


Fig. 9

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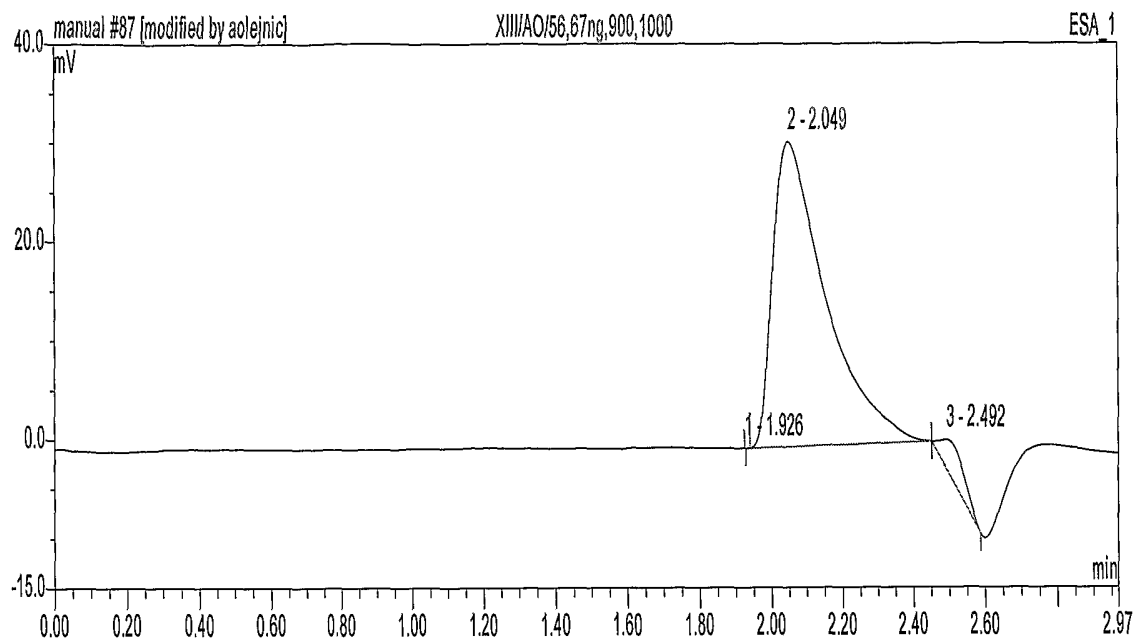


Fig. 10

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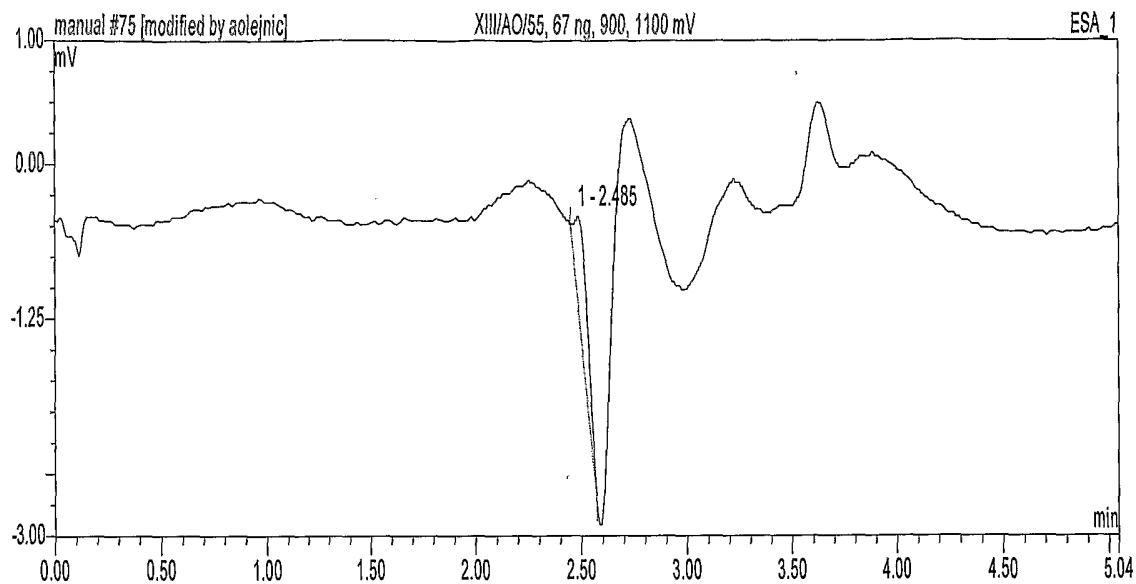


Fig. 11

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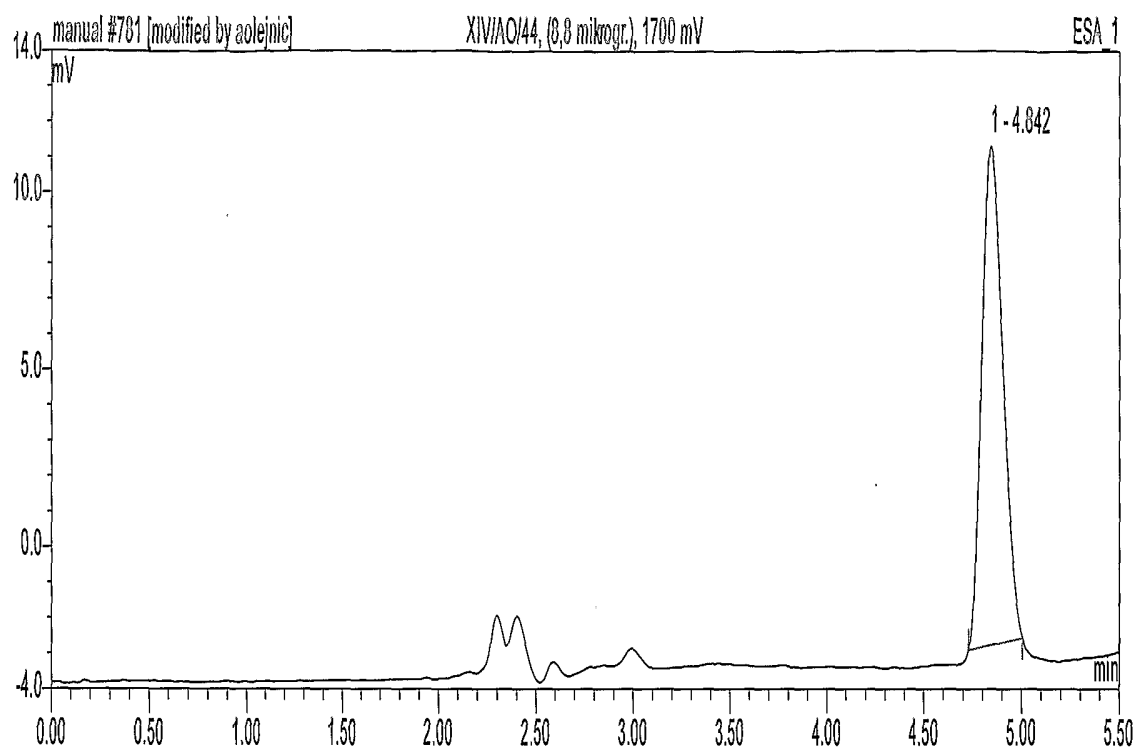


Fig. 12

13/13

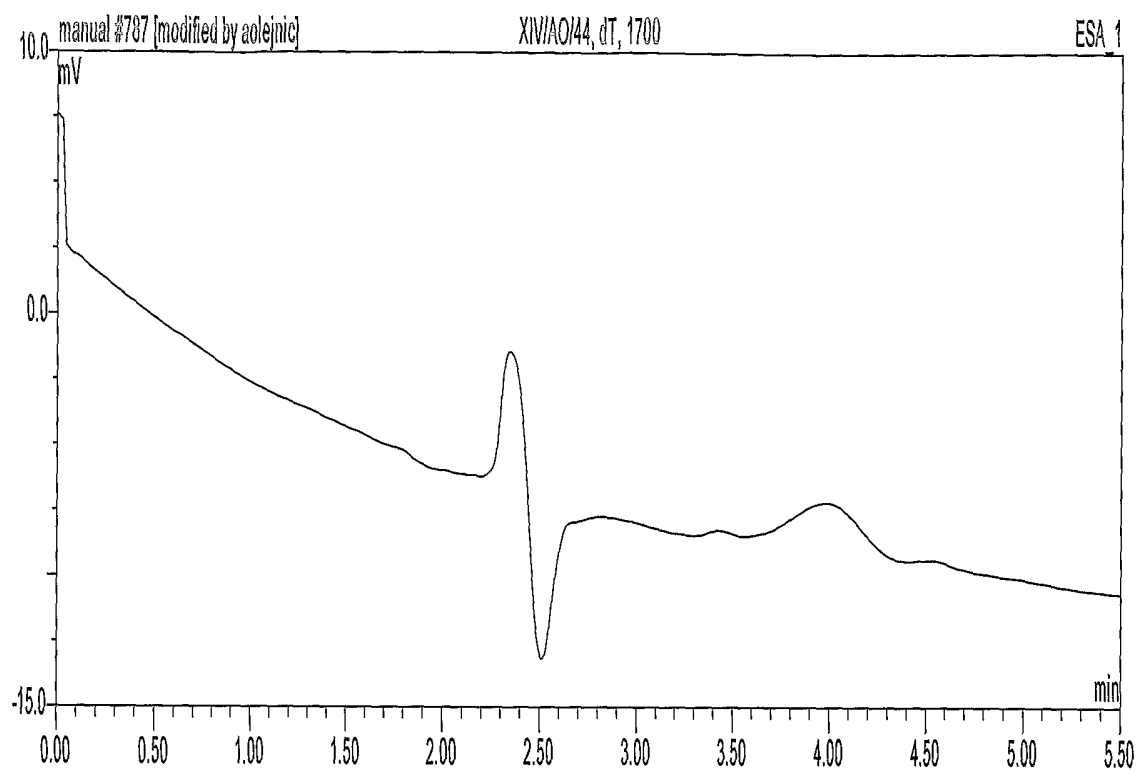


Fig. 13