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DESCRIPTION

Field of the invention:

[0001] The present invention relates to modified nucleic acid sequences coding for coagulation factor VIII (FVIII) and for non-modified von Willebrand factor (VWF) as well as complexes thereof and their derivatives, recombinant expression vectors containing such nucleic acid sequences, host cells transformed with such recombinant expression vectors, recombinant polypeptides and derivatives coded for by said nucleic acid sequences which recombinant polypeptides and derivatives do have biological activities together with prolonged in vivo half-life and/or improved in vivo recovery compared to the unmodified wild-type protein. The invention also relates to corresponding FVIII sequences that result in improved expression yield. The present invention further relates to processes for the manufacture of such recombinant proteins and their derivatives. The invention also relates to a transfer vector for use in human gene therapy, which comprises such modified nucleic acid sequences.

Background of the invention:

[0002] There are various bleeding disorders caused by deficiencies of blood coagulation factors. The most common disorders are hemophilia A and B, resulting from deficiencies of blood coagulation factor VIII and IX, respectively. Another known bleeding disorder is von Willebrand's disease.

[0003] In plasma FVIII exists mostly as a noncovalent complex with VWF and its coagulant function is to accelerate factor IXa dependent conversion of factor X to Xa. Due to the complex formation of FVIII and VWF it was assumed for a long time that FVIII and VWF functions are two functions of the same molecule. Only in the seventies it became clear that FVIII and VWF are separate molecules that form a complex under physiologic conditions. In the eighties then the dissociation constant of about 0.2 nmol/L was determined (Leyte et al., Biochem J 1989, 257: 679-683) and the DNA sequence of both molecules was studied.

[0004] Classic hemophilia or hemophilia A is an inherited bleeding disorder. It results from a chromosome X-linked deficiency of blood coagulation FVIII, and affects almost exclusively males with an incidence of between one and two individuals per 10.000. The X-chromosome defect is transmitted by female carriers who are not themselves hemophiliacs. The clinical manifestation of hemophilia A is an increased bleeding tendency. Before treatment with FVIII concentrates was introduced the mean life span for a person with severe hemophilia was less than 20 years. The use of concentrates of FVIII from plasma has considerably improved the situation for the hemophilia A patients increasing the mean life span extensively, giving most of them the possibility to live a more or less normal life. However, there have been certain problems with the plasma derived concentrates and their use, the most serious of which have

been the transmission of viruses. So far, viruses causing hepatitis B, non-A non-B hepatitis and AIDS have hit the population seriously. Since then different virus inactivation methods and new highly purified FVIII concentrates have recently been developed which established a very high safety standard also for plasma derived FVIII.

[0005] The cloning of the cDNA for FVIII (Wood et al. 1984. Nature 312:330-336; Vehar et al. 1984. Nature 312:337-342) made it possible to express FVIII recombinantly leading to the development of several recombinant FVIII products, which were approved by the regulatory authorities between 1992 and 2003. The fact that the central B domain of the FVIII polypeptide chain residing between amino acids Arg-740 and Glu-1649 does not seem to be necessary for full biological activity has also led to the development of a B domain deleted FVIII.

[0006] The mature FVIII molecule consists of 2332 amino acids which can be grouped into three homologous A domains, two homologous C domains and a B Domain which are arranged in the order: A1-A2-B-A3-C1-C2. The complete amino acid sequence of mature human FVIII is shown in SEQ ID NO:15. During its secretion into plasma FVIII is processed intracellularly into a series of metal-ion linked heterodimers as single chain FVIII is cleaved at the B-A3 boundary and at different sites within the B-domain. This processing leads to heterogeneous heavy chain molecules consisting of the A1, the A2 and various parts of the B-domain which have a molecular size ranging from 90 kDa to 200 kDa. The heavy chains are bound via a metal ion to the light chains, which consist of the A3, the C1 and the C2 domain (Saenko et al. 2002. Vox Sang. 83:89-96). In plasma this heterodimeric FVIII binds with high affinity to von Willebrand Factor (VWF), which protects it from premature catabolism. The half-life of non-activated FVIII bound to VWF is about 12 hours in plasma.

[0007] Coagulation FVIII is activated via proteolytic cleavage by FXa and thrombin at amino acids Arg372 and Arg740 within the heavy chain and at Arg1689 in the light chain resulting in the release of von Willebrand Factor and generating the activated FVIII heterotrimer which will form the tenase complex on phospholipid surfaces with FIXa and FX provided that Ca^{2+} is present. The heterotrimer consists of the A1 domain, a 50 kDa fragment, the A2 domain, a 43 kDa fragment and the light chain (A3-C1-C2), a 73 kDa fragment. Thus the active form of FVIII (FVIIIa) consists of an A1-subunit associated through the divalent metal ion linkage to a thrombin-cleaved A3-C1-C2 light chain and a free A2 subunit relatively loosely associated with the A1 and the A3 domain.

[0008] To avoid excessive coagulation, FVIIIa must be inactivated soon after activation. The inactivation of FVIIIa via activated Protein C (APC) by cleavage at Arg336 and Arg562 is not considered to be the major rate-limiting step. It is rather the dissociation of the non covalently attached A2 subunit from the heterotrimer which is thought to be the rate limiting step in FVIIIa inactivation after thrombin activation (Fay et al. 1991. J. Biol. Chem. 266 8957, Fay & Smudzin 1992. J. Biol. Chem. 267:13246-50). This is a rapid process, which explains the short half-life of FVIIIa in plasma, which is only 2.1 minutes (Saenko et al. 2002. Vox Sang. 83:89-96).

[0009] In severe hemophilia A patients undergoing prophylactic treatment FVIII has to be

administered intravenously (i.v.) about 3 times per week due to the short plasma half-life of FVIII of about 12 to 14 hours. Each i.v. administration is cumbersome, associated with pain and entails the risk of an infection especially as this is mostly done at home by the patients themselves or by the parents of children being diagnosed for hemophilia A.

[0010] It would thus be highly desirable to create a FVIII with increased functional half-life allowing the manufacturing of pharmaceutical compositions containing FVIII, which have to be administered less frequently.

[0011] Several attempts have been made to prolong the half-life of non-activated FVIII either by reducing its interaction with cellular receptors (WO 03/093313A2, WO 02/060951A2), by covalently attaching polymers to FVIII (WO 94/15625, WO 97/11957 and US 4970300), by encapsulation of FVIII (WO 99/55306), by introduction of novel metal binding sites (WO 97/03193), by covalently attaching the A2 domain to the A3 domain either by peptidic (WO 97/40145 and WO 03/087355) or disulfide linkage (WO 02/103024A2) or by covalently attaching the A1 domain to the A2 domain (WO2006/108590).

[0012] Another approach to enhance the functional half-life of FVIII or VWF is by PEGylation of FVIII (WO 2007/126808, WO 2006/053299, WO 2004/075923) or by PEGylation of VWF (WO 2006/071801) which pegylated VWF by having an increased half-life would indirectly also enhance the half-life of FVIII present in plasma.

[0013] As none of the above described approaches has yet resulted in an approved FVIII pharmaceutical and as introducing mutations into the FVIII wild-type sequence or introducing chemical modifications entails at least a theoretical risk of creating immunogenic FVIII variants there is an ongoing need to develop modified coagulation factor VIII molecules which exhibit prolonged half-life.

[0014] In view of a potential thrombogenic risk it is more desirable to prolong the half-life of the non-activated form of FVIII than that of FVIIIa.

[0015] VWF, which is missing, functionally defect or only available in reduced quantity in different forms of von Willebrand disease (VWD), is a multimeric adhesive glycoprotein present in the plasma of mammals, which has multiple physiological functions. During primary hemostasis VWF acts as a mediator between specific receptors on the platelet surface and components of the extracellular matrix such as collagen. Moreover, VWF serves as a carrier and stabilizing protein for procoagulant FVIII. VWF is synthesized in endothelial cells and megakaryocytes as a 2813 amino acid precursor molecule. The amino acid sequence and the cDNA sequence of wild-type VWF are disclosed in Collins et al. 1987, Proc Natl. Acad. Sci. USA 84:4393-4397. The precursor polypeptide, pre-pro-VWF, consists of a 22-residue signal peptide, a 741- residue pro-peptide and the 2050-residue polypeptide found in mature plasma VWF (Fischer et al., FEBS Lett. 351: 345-348, 1994). After cleavage of the signal peptide in the endoplasmic reticulum a C-terminal disulfide bridge is formed between two monomers of VWF. During further transport through the secretory pathway 12 N-linked and 10 O-linked

carbohydrate side chains are added. More important, VWF dimers are multimerized via N-terminal disulfide bridges and the propeptide of 741 amino acids length is cleaved off by the enzyme PACE/furin in the late Golgi apparatus. The propeptide as well as the high-molecular-weight multimers of VWF (VWF-HMWM) are stored in the Weibel-Pallade bodies of endothelial cells or in the α -Granules of platelets.

[0016] Once secreted into plasma the protease ADAMTS13 cleaves VWF within the A1 domain of VWF. Plasma VWF therefore consists of a whole range of multimers ranging from single dimers of 500 kDa to multimers consisting of up to more than 20 dimers of a molecular weight of over 10,000 kDa. The VWF-HMWM hereby having the strongest hemostatic activity, which can be measured in ristocetin cofactor activity (VWF:RCo). The higher the ratio of VWF:RCo/VWF antigen, the higher the relative amount of high molecular weight multimers.

[0017] Defects in VWF are causal to von Willebrand disease (VWD), which is characterized by a more or less pronounced bleeding phenotype. VWD type 3 is the most severe form in which VWF is completely missing, VWD type 1 relates to a quantitative loss of VWF and its phenotype can be very mild. VWD type 2 relates to qualitative defects of VWF and can be as severe as VWD type 3. VWD type 2 has many sub forms some of them being associated with the loss or the decrease of high molecular weight multimers. Von VWD type 2a is characterized by a loss of both intermediate and large multimers. VWD type 2B is characterized by a loss of highest-molecular-weight multimers.

[0018] VWD is the most frequent inherited bleeding disorder in humans and can be treated by replacement therapy with concentrates containing VWF of plasmatic or recombinant origin. VWF can be prepared from human plasma as for example described in EP 05503991. EP 0784632 describes a method for isolating recombinant VWF.

[0019] In plasma FVIII binds with high affinity to VWF, which protects it from premature catabolism and thus, plays in addition to its role in primary hemostasis a crucial role to regulate plasma levels of FVIII and as a consequence is also a central factor to control secondary hemostasis. The half-life of non-activated FVIII bound to VWF is about 12 to 14 hours in plasma. In von Willebrand disease type 3, where no or almost no VWF is present, the half-life of FVIII is only about 6 hours, leading to symptoms of mild to moderate hemophilia A in such patients due to decreased concentrations of FVIII. The stabilizing effect of VWF on FVIII has also been used to aid recombinant expression of FVIII in CHO cells (Kaufman et al. 1989, Mol Cell Biol).

[0020] Until today the standard treatment of Hemophilia A and VWD involves frequent intravenous infusions of preparations of FVIII and VWF concentrates or of concentrates comprising a complex of FVIII and VWF derived from the plasmas of human donors or in case of FVIII that of pharmaceutical preparations based on recombinant FVIII. While these replacement therapies are generally effective, e.g. in severe hemophilia A patients undergoing prophylactic treatment FVIII has to be administered intravenously (i.v.) about 3 times per week due to the short plasma half life of FVIII of about 12 hours. Already above levels of 1% of the

FVIII activity in non-hemophiliacs, e.g. by a raise of FVIII levels by 0.01 U/ml, severe hemophilia A is turned into moderate hemophilia A. In prophylactic therapy dosing regimes are designed such that the trough levels of FVIII activity do not fall below levels of 2-3% of the FVIII activity in non-hemophiliacs. Each i.v. administration is cumbersome, associated with pain and entails the risk of an infection especially as this is mostly done in home treatment by the patients themselves or by the parents of children being diagnosed for hemophilia A. In addition the frequent i.v. injections inevitably result in scar formation, interfering with future infusions. As prophylactic treatment in severe hemophilia is started early in life, with children often being less than 2 years old, it is even more difficult to inject FVIII 3 times per week into the veins of such small patients. For a limited period, implantation of port systems may offer an alternative. Despite the fact that repeated infections may occur and ports can cause inconvenience during physical exercise, they are nevertheless typically considered as favorable as compared to intravenous injections.

[0021] In the prior art fusions of coagulation factors to albumin (WO 01/79271), alpha-fetoprotein (WO 2005/024044) and immunoglobulin (WO 2004/101740) as half-life enhancing polypeptides have been described. These were taught to be attached to the carboxy- or the amino-terminus or to both termini of the respective therapeutic protein moiety, occasionally linked by peptidic linkers, preferably by linkers consisting of glycine and serine.

[0022] Ballance et al. (WO 01/79271) described N- or C-terminal fusion polypeptides of a multitude of different therapeutic polypeptides fused to human serum albumin. Long lists of potential fusion partners are described without disclosing experimental data for almost any of these polypeptides whether or not the respective albumin fusion proteins actually retain biological activity and have improved properties. Among said list of therapeutic polypeptides also FVIII and VWF are mentioned.

[0023] A C-terminal fusion would not have been seriously considered by the man skilled in the art as the C2 domain of FVIII at the very C-terminal part of FVIII between amino acid 2303 and 2332 of FVIII comprises a platelet membrane binding site which is essential for FVIII function. This is why there are many amino acid mutations known in this region which lead to hemophilia A. It was thus surprising that a relatively large heterologous polypeptide like albumin can be fused to the C-terminal part of FVIII without preventing FVIII function by preventing platelet binding. In addition, the C2 domain also contains a binding site for VWF. This site together with the amino acid sequence 1649-1689 is responsible for the high affinity binding of FVIII to VWF. Therefore, a man skilled in the art would also not have expected that a FVIII with a C-terminal albumin fusion would retain its binding to VWF.

[0024] It was surprisingly found that in contrast to the prediction by Ballance et al. an albumin fusion to the N-terminus of FVIII was not secreted into the culture medium. Therefore and because of the reasons detailed above it was now even more surprisingly found that a FVIII fused at its C-terminal part to albumin is secreted into the culture medium and retains its biological function including binding to membranes of activated platelets and to VWF.

[0025] It was also surprising to find that the modified FVIII of the invention shows an increase of in vivo recovery by about 20% compared to the wild type FVIII.

Description of the invention

[0026] It is an objective of this invention to provide a modified FVIII or complexes of modified FVIII with non-modified VWF, with enhanced in vivo half-life.

[0027] The term "modified FVIII" in the sense of the invention means FVIII polypeptides which are fused to half-life enhancing polypeptides, encompassing also natural alleles, variants, deletions and insertions of FVIII.

[0028] It is another objective of this invention to provide a modified FVIII as well as complexes of modified FVIII with non-modified VWF with improved in vivo recovery.

[0029] Another objective of the invention is that this modified FVIII or complexes of modified FVIII with non-modified VWF can be expressed by mammalian cells and retain their respective biological activities.

[0030] In summary, surprisingly the modified FVIII or complexes of modified FVIII with non-modified VWF of the invention have retained biological activity, increased in vivo half-life and in vivo recovery.

[0031] An additional potential benefit of those embodiments of the present invention in which the FVIII is modified and in which the A2 domain remains only non covalently attached to the A3 domain after activation is that only the half-life of the non-activated form of FVIII is increased, whereas the half-life of the activated form of FVIII remains essentially the same, which might result in a decreased risk of thrombogenicity as compared to FVIII variants which lead to a stabilization of the activated form of FVIII.

[0032] The modified FVIII or complexes of modified FVIII with non-modified VWF of the invention can be generated by fusing a half-life enhancing protein (HLEP) moiety to the C-terminal part of FVIII.

[0033] HLEPs in the sense of the present invention are selected from a group consisting of members of human albumin.

[0034] The present invention therefore relates to a modified FVIII or of modified FVIII with non-modified VWF having at the C-terminal part of the modified FVIII a fusion to a HLEP, characterized in that the modified FVIII or the complex of modified FVIII with non-modified VWF has prolonged functional half-life compared to the functional half-life of the wild-type FVIII or the complex of wild-type VWF and wild-type FVIII.

[0035] The present invention also relates to C-terminal fusions to more than one HLEP wherein the HLEP, which is fused several times.

[0036] The present invention also relates to a modified FVIII having at the C-terminal part a fusion to a HLEP, characterized in that the modified FVIII or the complex of modified FVIII with non-modified VWF has improved in vivo recovery compared to the in vivo recovery of the wild-type FVIII or the complex of wild-type VWF and wild-type FVIII.

[0037] Another embodiment of the invention are modified FVIII polypeptides having at the C-terminal part a fusion to a HLEP, characterized in that the modified FVIII is secreted into a fermentation medium at a higher yield as a wild-type FVIII.

[0038] Another aspect of the invention are polynucleotides or combinations of polynucleotides encoding the modified FVIII.

[0039] The invention further relates to plasmids or vectors comprising a polynucleotide described herein, to host cells comprising a polynucleotide or a plasmid or vector described herein.

[0040] Another aspect of the invention is a method of producing a modified FVIII or a complex of modified FVIII with non-modified VWF, comprising:

1. (a) culturing host cells of the invention under conditions such that the modified coagulation factor is expressed; and
2. (b) optionally recovering the modified coagulation factor from the host cells or from the culture medium.

[0041] The invention further pertains to pharmaceutical compositions comprising a modified FVIII or a complex of modified FVIII with non-modified VWF, a polynucleotide, or a plasmid or vector described herein.

Yet another aspect of the invention is the use of a modified FVIII or a complex of modified FVIII with non-modified VWF, one or more polynucleotides, or one or more plasmids or vectors, or of host cells according to this invention for the manufacture of a medicament for the treatment or prevention of a blood coagulation disorder.

Detailed description of the invention

[0042] The invention pertains to a complex comprising FVIII and VWF or one of its individual polypeptidic components wherein the FVIII of said complex is fused at the C-terminal part of its primary translation product to the N-terminal part of a half-life enhancing polypeptide (HLEP).

[0043] The invention also pertains to a modified FVIII or a complex comprising modified FVIII and non-modified wherein the modified FVIII is fused at a C-terminal part of the primary translation polypeptide of FVIII to the N-terminal part of a HLEP.

[0044] In preferred embodiments the invention pertains to a modified FVIII or a complex comprising modified FVIII and non-modified, wherein

1. (a) the modified FVIII has a prolonged functional half-life compared to the functional half-life of wild-type FVIII or
2. (b) the complex comprising modified FVIII and non-modified VWF has a prolonged functional half-life compared to the functional half-life of the corresponding complex comprising wild-type FVIII and wild-type VWF.

[0045] A preferred embodiment of the invention is a modified polypeptide or a complex comprising said modified polypeptide or a complex comprising said modified polypeptides as described above, wherein the modified polypeptide has a functional half-life increased by at least 25% as compared to the functional half-life of the corresponding wild-type polypeptide or the complex comprising said modified polypeptide or a complex comprising said modified polypeptides has a functional half-life increased by at least 25% as compared to the corresponding complex of wild-type FVIII and wild-type VWF.

[0046] Another embodiment of the invention is a modified FVIII or a complex comprising modified FVIII and non-modified VWF, wherein

1. (a) the modified FVIII has a prolonged antigen half-life compared to the antigen half-life of wild-type FVIII or
2. (b) the complex comprising modified FVIII and non-modified VWF has a prolonged antigen half-life compared to the antigen half-life of the corresponding complex comprising wild-type FVIII and wild-type VWF.

[0047] A preferred embodiment of the invention is a modified polypeptide or a complex comprising said modified polypeptide or a complex comprising said modified polypeptides as described above, wherein the modified polypeptide has an antigen half-life increased by at least 25% as compared to the antigen half-life of the corresponding wild-type polypeptide or the complex comprising said modified polypeptide or a complex comprising said modified polypeptides has an antigen half-life increased by at least 25% as compared to the corresponding complex of wild-type FVIII and wild-type VWF.

[0048] Still another embodiment of the invention is a modified FVIII or a complex comprising modified FVIII and non-modified VWF, wherein

1. (a) the modified FVIII has an increased in vivo recovery compared to the in vivo recovery

of wild-type FVIII or

2. (b) the complex comprising modified FVIII and non-modified VWF has an increased in vivo recovery compared to the in vivo recovery of the corresponding complex comprising wild-type FVIII and wild-type VWF .

[0049] Another preferred embodiment of the invention is a modified polypeptide or a complex comprising said modified polypeptide or a complex comprising said modified polypeptides as described above, wherein the modified polypeptide has an in vivo recovery increased by at least 10% as compared to the in vivo recovery of the corresponding wild-type polypeptide or the complex comprising said modified polypeptide or a complex comprising said modified polypeptides has an in vivo recovery increased by at least 10% as compared to the corresponding complex of wild-type FVIII and wild-type VWF.

Another preferred embodiment of the invention is

1. (a) a modified polypeptide or a complex comprising said modified polypeptide or a complex comprising said modified polypeptides as described above, wherein at least one polypeptidic component of said complex is fused at the C-terminal amino acid of its primary translation product to the N-terminal part of a HLEP or
2. (b) a modified polypeptide or a complex comprising said modified polypeptide or a complex comprising said modified polypeptides as described above, wherein at least one polypeptidic component of said complex is fused at the C-terminal part of its primary translation product to the N-terminal amino acid of a HLEP or
3. (c) a modified polypeptide or a complex comprising said modified polypeptide or a complex comprising said modified polypeptides as described above, wherein at least one polypeptidic component of said complex is fused at the C-terminal amino acid of its primary translation product to the N-terminal amino acid of a HLEP.

[0050] Another preferred embodiment of the invention is a modified polypeptide or a complex comprising said modified polypeptide or a complex comprising said modified polypeptides as described above, wherein the modified polypeptide has at least 10% of the biological activity of wild-type polypeptide or the complex comprising the modified polypeptide or a complex comprising said modified polypeptides has at least 10% of the biological activity of the corresponding complex of wild-type FVIII and wild-type VWF.

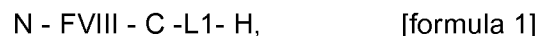
[0051] Also comprised in the present invention is a method of preparing a modified having increased functional half-life, comprising fusing the N-terminal part of a half-life-enhancing polypeptide to a C-terminal part of the primary translation polypeptide of the FVIII as well as a method of preparing a complex comprising modified FVIII and non-modified VWF by mixing a modified FVIII prepared by the method described above with wild-type VWF prepared by the method described above.

[0052] Also encompassed in the invention is the use of a modified FVIII as prepared by the method described above and wild-type VWF for the manufacture of a combined pharmaceutical preparation for simultaneous, separate or sequential use in the therapy of bleeding disorders, preferentially in the therapy of hemophilia A and/or von Willebrand disease.

[0053] The "functional half-life" according to the present invention is the half-life of the biological activity of the modified FVIII or a complex of modified FVIII with non-modified VWF once it has been administered to a mammal and can be measured in vitro in blood samples taken at different time intervals from said mammal after the modified FVIII or the complex of modified FVIII with non-modified VWF has been administered.

[0054] The phrases "fusing" or "fused" refer to the addition of amino acids to the C-terminal part of FVIII. When referring herein to a "fusion to the C-terminal amino acid of FVIII" this means a fusion exactly to the C-terminal amino acid of FVIII at amino acid 2332 of the mature wild-type FVIII cDNA sequence. Mature FVIII meaning the respective polypeptide after cleavage of the propeptide. However the invention also encompasses a "fusion to the C-terminal part of FVIII" in the sense of this invention may also include a fusion to a FVIII molecule respectively in which one or more amino acid position up to n amino acids from the C-terminal amino acid of FVIII are deleted. The figure n is an integer that should not be greater than 5%, preferably not greater than 1% of the total number of amino acids of the FVIII. Usually, n is 20, preferably 15, more preferably 10, still more preferably 5 or less (e.g. 1, 2, 3, 4 or 5).

[0055] In one embodiment, the modified FVIII has the following structure:



wherein

N is an N-terminal part of FVIII,

L1 is a chemical bond or a linker sequence

H is a HLEP, and

C is a C-terminal part of FVIII

[0056] L1 may be a chemical bond or a linker sequence consisting of one or more amino acids, e.g. of 1 to 20, 1 to 15, 1 to 10, 1 to 5 or 1 to 3 (e.g. 1, 2 or 3) amino acids and which may be equal or different from each other. Usually, the linker sequences are not present at the corresponding position in the wild-type coagulation factor. Examples of suitable amino acids present in L1 include Gly and Ser.

[0057] Preferred HLEP sequences are described infra. Likewise encompassed by the

invention are fusions to the exact "N-terminal amino acid" of the respective HLEP, or fusions to the "N-terminal part" of the respective HLEP, which includes N-terminal deletions of one or more amino acids of the HLEP.

[0058] The modified FVIII or the complex of the modified FVIII with the non-modified VWF, of the invention may comprise more than one HLEP sequence, e.g. two or three HLEP sequences. These multiple HLEP sequences may be fused to the C-terminal part of FVIII in tandem, e.g. as successive repeats.

[0059] FVIII may be processed proteolytically at various stages. For example, as mentioned supra, during its secretion into plasma single chain FVIII is cleaved intracellularly at the B-A3 boundary and at different sites within the B-domain. The heavy chain is bound via a metal ion to the light chain having the domain structure A3-C1-C2. FVIII is activated via proteolytic cleavage at amino acids Arg372 and Arg740 within the heavy chain and at Arg1689 in the light chain generating the activated FVIII heterotrimer consisting of the A1 domain, the A2 domain, and the light chain (A3-C1-C2), a 73 kDa fragment. Thus the active form of FVIII (FVIIIa) consists of an A1-subunit associated through the divalent metal ion linkage to a thrombin-cleaved A3-C1-C2 light chain and a free A2 subunit relatively loosely associated with the A1 and the A3 domain.

[0060] Accordingly, the present invention encompasses also modified FVIII that is not present as a single chain polypeptide but consists of several polypeptides (e.g. one or two or three) that are associated with each other via non-covalent linkages.

[0061] Preferably N - FVIII - C comprises the full length sequence of FVIII. Also encompassed are N-terminal, C-terminal or internal deletions of FVIII as long as the biological activity of FVIII is retained. The biological activity is retained in the sense of the invention if the FVIII with deletions retains at least 10%, preferably at least 25%, more preferably at least 50%, most preferably at least 75% of the biological activity of wild-type FVIII. The biological activity of FVIII can be determined by the artisan as described below.

[0062] A suitable test to determine the biological activity of FVIII is for example the one stage or the two stage coagulation assay (Rizza et al. 1982. Coagulation assay of FVIII:C and FIXa in Bloom ed. The Hemophilias. NY Churchill Livingston 1992) or the chromogenic substrate FVIII:C assay (S. Rosen, 1984. Scand J Haematol 33: 139-145, suppl.). The content of these references is incorporated herein by reference.

[0063] The cDNA sequence and the amino acid sequence of the mature wild-type form of human blood coagulation FVIII are shown in SEQ ID NO:14 and SEQ ID NO:15, respectively. The reference to an amino acid position of a specific sequence means the position of said amino acid in the FVIII wild-type protein and does not exclude the presence of mutations, e.g. deletions, insertions and/or substitutions at other positions in the sequence referred to. For example, a mutation in "Glu2004" referring to SEQ ID NO:15 does not exclude that in the modified homologue one or more amino acids at positions 1 through 2332 of SEQ ID NO:15

are missing.

[0064] The terms "blood coagulation Factor VIII", "Factor VIII" and "FVIII" are used interchangeably herein. "Blood coagulation Factor VIII" includes wild-type blood coagulation FVIII as well as derivatives of wild-type blood coagulation FVIII having the procoagulant activity of wild-type blood coagulation FVIII. Derivatives may have deletions, insertions and/or additions compared with the amino acid sequence of wild-type FVIII. The term FVIII includes proteolytically processed forms of FVIII, e.g. the form before activation, comprising heavy chain and light chain.

[0065] The term "FVIII" includes any FVIII variants or mutants having at least 25%, more preferably at least 50%, most preferably at least 75% of the biological activity of wild-type factor VIII.

[0066] As non-limiting examples, FVIII molecules include FVIII mutants preventing or reducing APC cleavage (Amano 1998. *Thromb. Haemost.* 79:557-563), FVIII mutants further stabilizing the A2 domain (WO 97/40145), FVIII mutants resulting in increased expression (Swaroop et al. 1997. *JBC* 272:24121-24124), FVIII mutants reducing its immunogenicity (Lollar 1999. *Thromb. Haemost.* 82:505-508), FVIII reconstituted from differently expressed heavy and light chains (Oh et al. 1999. *Exp. Mol. Med.* 31:95-100), FVIII mutants reducing binding to receptors leading to catabolism of FVIII like HSPG (heparan sulfate proteoglycans) and/or LRP (low density lipoprotein receptor related protein) (Ananyeva et al. 2001. *TCM*, 11:251-257), disulfide bond-stabilized FVIII variants (Gale et al., 2006. *J. Thromb. Hemost.* 4:1315-1322), FVIII mutants with improved secretion properties (Miao et al., 2004. *Blood* 103:3412-3419), FVIII mutants with increased cofactor specific activity (Wakabayashi et al., 2005. *Biochemistry* 44:10298-304), FVIII mutants with improved biosynthesis and secretion, reduced ER chaperone interaction, improved ER-Golgi transport, increased activation or resistance to inactivation and improved half-life (summarized by Pipe 2004. *Sem. Thromb. Hemost.* 30:227-237). All of these FVIII mutants and variants are incorporated herein by reference in their entirety.

[0067] VWF may be processed proteolytically at various stages. For example, as mentioned supra, the protease ADAMTS13 cleaves VWF within the A2 domain of VWF. Accordingly, the present invention encompasses also modified VWF which has been cleaved proteolytically e.g. by ADAMTS13. Such cleavage would result in multimeric chains of VWF which comprise at their ends at least one or at most two monomers of VWF which have been cleaved by ADAMTS 13.

[0068] Preferably N - VWF - C comprises the full length sequence of VWF. Also encompassed are N-terminal, C-terminal or internal deletions of VWF as long as the biological activity of VWF is retained. The biological activity is retained in the sense of the invention if the VWF with deletions retains at least 10%, preferably at least 25%, more preferably at least 50%, most preferably at least 75% of the biological activity of wild-type VWF. The biological activity of wild-type VWF can be determined by the artisan using methods for ristocetin cofactor activity

(Federici AB et al. 2004. Haematologica 89:77-85), binding of VWF to GP Iba of the platelet glycoprotein complex Ib-V-IX (Sucker et al. 2006. Clin Appl Thromb Hemost. 12:305-310), or a collagen binding assay (Kallas & Talpsep. 2001. Annals of Hematology 80:466-471).

[0069] "FVIII" and/or "VWF" within the above definition also include natural allelic variations that may exist and occur from one individual to another. "FVIII" and/or "VWF" within the above definition further includes variants of FVIII and or VWF. Such variants differ in one or more amino acid residues from the wild-type sequence. Examples of such differences may include as conservative amino acid substitutions, i.e. substitutions within groups of amino acids with similar characteristics, e.g. (1) small amino acids, (2) acidic amino acids, (3) polar amino acids, (4) basic amino acids, (5) hydrophobic amino acids, and (6) aromatic amino acids. Examples of such conservative substitutions are shown in the following table.

Table 1:

(1)	Alanine	Glycine		
(2)	Aspartic acid	Glutamic acid		
(3)	Asparagine	Glutamine	Serine	Threonine
(4)	Arginine	Histidine	Lysine	
(5)	Isoleucine	Leucine	Methionine	Valine
(6)	Phenylalanine	Tyrosine	Tryptophane	

[0070] One or more HLEPs may be fused to the C-terminal part of FVIII preferably as not to interfere with the binding capabilities of FVIII for example to VWF, platelets or FIX.

[0071] Once FVIII is endogenously activated during coagulation in vivo, it may be no longer desirable to maintain the increased functional half-life of the now activated FVIII as this might lead to thrombotic complications what is already the case for a wild-type activated coagulation factor as FVIIa (Aledort 2004. J Thromb Haemost 2:1700-1708) and what may be more relevant if the activated factor would have an increased functional half-life. It is therefore another objective of the present invention to provide long-lived FVIII molecules, which after endogenous activation in vivo or after availability of a cofactor do have a functional half-life comparable to that of unmodified FVIII. This can by way of non-limiting example be achieved by introducing a cleavage site for example for a coagulation factor between the C-terminal part of FVIII and the HLEP. With such FVIII-HLEP connecting sequences the activation of the FVIII chimeric protein of the invention will lead to a concomitant complete separation of FVIIIa from the HLEP moiety. Accordingly, in one embodiment, the functional half-life of the endogenously activated modified FVIII is substantially the same as that of the activated wild-type FVIII (e.g. $\pm 15\%$, preferably $\pm 10\%$).

[0072] In yet another embodiment of the invention, however, one or more of the proteolytical cleavage sites, preferably the thrombin cleavage sites at Arg740 and/or Arg372, are mutated or deleted in order to prevent cleavage and result in an insertion protein which displays improved properties like enhanced functional half-life even as an activated molecule.

[0073] In another embodiment of the invention the FVIII proteins of the invention may be expressed as two separate chains (see *infra*).

[0074] The modified FVIII according to this invention may be a single chain polypeptide, or it may be composed of two or three polypeptide chains that are associated via non-covalent linkages, due to proteolytic processing.

[0075] In another embodiment of the invention, the amino acids at or near the PACE/Furin cleavage site (Arg1648) are mutated or deleted in order to prevent cleavage by PACE/Furin. This is thought to result in a one-chain FVIII/HLEP fusion molecule with improved half-life.

[0076] In one embodiment of the invention, the modified FVIII of the invention exhibits an increased functional half-life compared to the corresponding FVIII form containing no integrated HLEP and/or to the wild-type form FVIII. The functional half-life e.g. can be determined *in vivo* in animal models of hemophilia A, like FVIII knockout mice, in which one would expect a longer lasting hemostatic effect as compared to wild-type FVIII. The hemostatic effect could be tested for example by determining time to arrest of bleeding after a tail clip.

[0077] The functional half-life in one embodiment of the invention is the half-life of the biological activity of the FVIII once it has been administered to a mammal and is measured *in vitro*. The functional half-life of the modified FVIII according to the invention is greater than that of the FVIII lacking the modification as tested in the same species. The functional half-life is preferably increased by at least 10%, preferably 25%, more preferably by at least 50%, and even more preferably by at least 100% compared to the wild-type form of FVIII.

[0078] The functional half-life of a modified FVIII comprising a HLEP modification, can be determined by administering the respective modified FVIII (and in comparison wild-type FVIII) to rats, rabbits or other experimental animal species intravenously or subcutaneously and following the elimination of the biological activity of said modified or respectively non-modified coagulation factor in blood samples drawn at appropriate intervals after application. Suitable test methods are the activity tests described herein.

[0079] As a surrogate marker for the half-life of biological activity also the levels of antigen of the modified or respectively wild-type FVIII or the levels of wild-type VWF can be measured. Thus also encompassed by the invention are modified FVIII molecules having at the C-terminal part of FVIII a fusion to a HLEP, characterized in that the modified FVIII or the complex of modified FVIII with non-modified VWF has a prolonged half-life of the FVIII antigen compared to the half-life of the FVIII antigen lacking said insertion. The "half-life of the FVIII antigen" according to the present invention is the half-life of the antigen of the FVIII once it has been administered to a mammal and is measured *in vitro*. Antigen test methods based on specific antibodies in an enzyme immunoassay format as known to the artisan and commercially available (e.g. Dade Behring, Instrumentation Laboratory, Abbott Laboratories, Diagnostica Stago). Functional and antigen half-lives can be calculated using the time points of the beta

phase of elimination according to the formula $t_{1/2} = \ln 2 / k$, whereas k is the slope of the regression line.

[0080] In another embodiment, the functional half-life of the endogenously activated modified FVIII is prolonged compared to that of the activated wild-type FVIII. The increase may be more than 15%, for example at least 20% or at least 50%. Again, such functional half-life values can be measured and calculated as described for functional half-lives supra. Increased half-lives of the endogenously activated modified FVIII molecules may be beneficial in situations where only very low levels of FVIII are available that therefore are not thrombogenic. Such situations may occur e.g. upon gene therapy treatment where often only low expression rates can be achieved. Therefore, such stabilized FVIII molecules might be beneficial in e.g. gene therapy despite a thrombogenic risk connected to such FVIII molecules if administered as proteins in high or physiologic doses.

[0081] In another embodiment of the invention, the modified FVIII of the invention exhibits an improved in vivo recovery compared to the wild-type FVIII. The in vivo recovery can be determined in vivo for example in normal animals or in animal models of hemophilia A, like FVIII knockout mice, in which one would expect an increased percentage of the modified FVIII of the invention be found by antigen or activity assays in the circulation shortly (5 to 10 min.) after i.v. administration compared to the corresponding wild-type FVIII or wild-type VWF.

[0082] The in vivo recovery is preferably increased by at least 10%, more preferably by at least 20%, and even more preferably by at least 40% compared to wild-type form FVIII or to wild-type VWF.

[0083] In yet another embodiment of the invention immunoglobulin constant regions or portions thereof are used as HLEPs. Preferably the Fc region comprised of a CH2 and CH3 domain and a hinge region of an IgG, more preferably of an IgG1 or fragments or variants thereof are used, variants including mutations which enhance binding to the neonatal Fc receptor (FcRn).

[0084] It is another objective of the present invention to provide long-lived FVIII molecules, which after proteolytic processing in vivo do have a functional half-life comparable to that of an unmodified FVIII. This can be achieved by maintaining certain cleavage sites in the modified FVIII leading to a proteolytic cleavage for example when in contact with activated coagulation factors, which separates the FVIII from the HLEP. Accordingly, in one embodiment, the functional half-life of the proteolytically processed modified FVIII is substantially the same as that of the non-modified VWF lacking the modification, and/or it is substantially the same as that of the wild-type VWF (e.g. $\pm 15\%$, preferably $\pm 10\%$).

[0085] Still another embodiment of the invention are modified FVIII polypeptides which are fused to a HLEP for example albumin at the C-terminal part of the FVIII molecule which do have reduced binding to VWF or do not bind VWF at all.

Linker sequences

[0086] According to this invention, the therapeutic polypeptide moiety may be coupled to the HLEP moiety by a peptide linker. The linker should be non-immunogenic and may be a non-cleavable or cleavable linker.

[0087] Non-cleavable linkers may be comprised of alternating glycine and serine residues as exemplified in WO2007/090584.

[0088] In another embodiment of the invention the peptidic linker between the FVIII and/or the VWF moiety and the albumin moiety consists of peptide sequences, which serve as natural interdomain linkers in human proteins. Preferably such peptide sequences in their natural environment are located close to the protein surface and are accessible to the immune system so that one can assume a natural tolerance against this sequence. Examples are given in WO2007/090584.

[0089] Cleavable linkers should be flexible enough to allow cleavage by proteases. In a preferred embodiment the cleavage of the linker proceeds comparably fast as the activation of FVIII within the fusion protein, if the fusion protein is a modified FVIII.

[0090] The cleavable linker preferably comprises a sequence derived from

1. a) the therapeutic polypeptide to be administered itself if it contains proteolytic cleavage sites that are proteolytically cleaved during activation of the therapeutic polypeptide,
2. b) a substrate polypeptide cleaved by a protease which is activated or formed by the involvement of the therapeutic polypeptide.
3. c) a polypeptide involved in coagulation or fibrinolysis

[0091] The linker region in a more preferred embodiment comprises a sequence of FVIII and/or VWF, which should result in a decreased risk of neoantigenic properties of the expressed fusion protein. Also in case the therapeutic protein is FVIII which needs to be proteolytically activated, the kinetics of the peptide linker cleavage will more closely reflect the coagulation-related activation kinetics of the zymogen.

[0092] In a preferred embodiment, the therapeutic polypeptide is FVIII zymogen and the HLEP is albumin. In this case the linker sequence is either derived from the sequences of the activation regions of FVIII, from the cleavage region of any substrate of FIX like FX or FVII or from the cleavage region of any substrate polypeptide that is cleaved by a protease in whose activation FIXa is involved.

[0093] In a highly preferred embodiment the linker peptide is derived from FVIII itself and

comprises of sequences encompassing the thrombin cleavage sites at amino acid positions 372, 740 and 1689 of SEQ ID NO. 15, respectively. In another preferred embodiment the linker peptide is derived from FX, FIX, FVII or FXI.

[0094] The linker peptides are preferably cleavable by the proteases of the coagulation system, for example FIIa, FIXa, FXa, FXIa, FXIIa and FVIIa.

[0095] Said linker sequences can also be used in the modified VWF of the invention.

[0096] Exemplary combinations of therapeutic polypeptide, cleavable linker and HLEP include the constructs listed in WO2007/090584 (for example in table 2 and figure 4) and WO2007/144173 (for example in table 3a and 3b), but are not limited to these.

Half-life enhancing polypeptides (HLEPs)

[0097] A "half-life enhancing polypeptide" as used herein is selected from the group consisting of albumin., It may be a full-length half-life-enhancing protein described herein or one or more fragments thereof that are capable of stabilizing or prolonging the therapeutic activity or the biological activity of the coagulation factor. Such fragments may be of 10 or more amino acids in length or may include at least about 15, at least about 20, at least about 25, at least about 30, at least about 50, at least about 100, or more contiguous amino acids from the HLEP sequence or may include part or all of specific domains of the respective HLEP, as long as the HLEP fragment provides a functional half-life extension of at least 25% compared to a wild-type FVIII or wild-type VWF.

[0098] The HLEP portion of the proposed coagulation factor insertion constructs of the invention may be a variant of a normal HLEP. The term "variants" includes insertions, deletions and substitutions, either conservative or non-conservative, where such changes do not substantially alter the active site, or active domain which confers the biological activities of the modified FVIII or modified VWF.

[0099] In particular, the proposed FVIII HLEP or VWF HLEP fusion constructs of the invention may include naturally occurring polymorphic variants of HLEPs and fragments of HLEPs. The HLEP may be derived from any vertebrate, especially any mammal, for example human, monkey, cow, sheep, or pig. Non-mammalian HLEPs include, but are not limited to, hen and salmon.

Albumin as HLEP

[0100] The terms, "human serum albumin" (HSA) and "human albumin" (HA) and "albumin" (ALB) are used interchangeably in this application. The terms "albumin" and "serum albumin"

are broader, and encompass human serum albumin (and fragments and variants thereof) as well as albumin from other species (and fragments and variants thereof).

[0101] As used herein, "albumin" refers collectively to albumin polypeptide or amino acid sequence, or an albumin fragment or variant, having one or more functional activities (e.g., biological activities) of albumin. In particular, "albumin" refers to human albumin or fragments thereof, especially the mature form of human albumin as shown in SEQ ID NO:16 herein or albumin from other vertebrates or fragments thereof, or analogs or variants of these molecules or fragments thereof.

[0102] In particular, the proposed FVIII fusion constructs of the invention may include naturally occurring polymorphic variants of human albumin and fragments of human albumin. Generally speaking, an albumin fragment or variant will be at least 10, preferably at least 40, most preferably more than 70 amino acids long. The albumin variant may preferentially consist of or alternatively comprise at least one whole domain of albumin or fragments of said domains, for example domains 1 (amino acids 1-194 of SEQ ID NO:16), 2 (amino acids 195-387 of SEQ ID NO: 16), 3 (amino acids 388-585 of SEQ ID NO: 16), 1 + 2 (1-387 of SEQ ID NO: 16), 2 + 3 (195-585 of SEQ ID NO: 16) or 1 + 3 (amino acids 1-194 of SEQ ID NO: 16 + amino acids 388-585 of SEQ ID NO: 16). Each domain is itself made up of two homologous subdomains namely 1-105, 120-194, 195-291, 316-387, 388-491 and 512-585, with flexible inter-subdomain linker regions comprising residues Lys106 to Glu119, Glu292 to Val315 and Glu492 to Ala511.

[0103] The albumin portion of the proposed FVIII fusion constructs of the invention may comprise at least one subdomain or domain of HA or conservative modifications thereof.

Polynucleotides

[0104] The invention further relates to a polynucleotide encoding a modified coagulation factor, preferably a modified FVIII variant as described in this application. The term "polynucleotide(s)" generally refers to any polyribonucleotide or polydeoxyribonucleotide that may be unmodified RNA or DNA or modified RNA or DNA. The polynucleotide may be single- or double-stranded DNA, single or double-stranded RNA. As used herein, the term "polynucleotide(s)" also includes DNAs or RNAs that comprise one or more modified bases and/or unusual bases, such as inosine. It will be appreciated that a variety of modifications may be made to DNA and RNA that serve many useful purposes known to those of skill in the art. The term "polynucleotide(s)" as it is employed herein embraces such chemically, enzymatically or metabolically modified forms of polynucleotides, as well as the chemical forms of DNA and RNA characteristic of viruses and cells, including, for example, simple and complex cells.

[0105] The skilled person will understand that, due to the degeneracy of the genetic code, a given polypeptide can be encoded by different polynucleotides. These "variants" are encompassed by this invention.

[0106] Preferably, the polynucleotide of the invention is an isolated polynucleotide. The term "isolated" polynucleotide refers to a polynucleotide that is substantially free from other nucleic acid sequences, such as and not limited to other chromosomal and extrachromosomal DNA and RNA. Isolated polynucleotides may be purified from a host cell. Conventional nucleic acid purification methods known to skilled artisans may be used to obtain isolated polynucleotides. The term also includes recombinant polynucleotides and chemically synthesized polynucleotides.

[0107] The invention further relates to a group of polynucleotides which together encode the modified FVIII and/or the modified VWF of the invention. A first polynucleotide in the group may encode the N-terminal part of the modified FVIII, and a second polynucleotide may encode the C-terminal part of the modified FVIII.

[0108] Yet another aspect of the invention is a plasmid or vector comprising a polynucleotide according to the invention. Preferably, the plasmid or vector is an expression vector. In a particular embodiment, the vector is a transfer vector for use in human gene therapy.

[0109] The invention also relates to a group of plasmids or vectors that comprise the above group of polynucleotides. A first plasmid or vector may contain said first polynucleotide, and a second plasmid or vector may contain said second polynucleotide. By way of example, and with reference to coagulation factor FVIII, the coding sequences of the signal peptide, the A1 and A2 domains, the B domain sequence remainder and the HLEP may be cloned into the first expression vector and the coding sequences of A3, C1 and C2 with an appropriate signal peptide sequence may be cloned into the second expression vector. Both expression vectors are cotransfected into a suitable host cell, which will lead to the expression of the light and heavy chains of the FVIII molecule of the invention and the formation of a functional protein.

[0110] Alternatively, the coding sequence of the FVIII signal peptide, the A1 and A2 domains are cloned into the first expression vector and the coding sequences of the HLEP, FVIII A3, C1 and C2 with an appropriate signal peptide sequence are cloned into the second expression vector. Both expression vectors are cotransfected into a suitable host cell, which will lead to the expression of the light and heavy chains of the FVIII molecule of the invention and the formation of a functional protein.

[0111] Alternatively, both coding sequences are cloned into one expression vector either using two separate promoter sequences or one promoter and an internal ribosome entry site (IRES) element to direct the expression of both FVIII chains.

[0112] Still another aspect of the invention is a host cell comprising a polynucleotide, a plasmid or vector of the invention, or a group of polynucleotides or a group of plasmids or vectors as described herein.

[0113] The host cells of the invention may be employed in a method of producing a modified

coagulation factor, preferably a modified FVIII molecule, which is part of this invention. The method comprises:

1. (a) culturing host cells of the invention under conditions such that the desired insertion protein is expressed; and
2. (b) optionally recovering the desired insertion protein from the host cells or from the culture medium.

[0114] It is preferred to purify the modified FVIII of the present invention to $\geq 80\%$ purity, more preferably $\geq 95\%$ purity, and particularly preferred is a pharmaceutically pure state that is greater than 99.9% pure with respect to contaminating macromolecules, particularly other proteins and nucleic acids, and free of infectious and pyrogenic agents. Preferably, an isolated or purified modified FVIII of the invention is substantially free of other, non-related polypeptides.

[0115] The various products of the invention are useful as medicaments. Accordingly, the invention relates to a pharmaceutical composition comprising a modified FVIII as described herein, a polynucleotide of the invention, or a plasmid or vector of the invention.

[0116] The invention also concerns a method of treating an individual suffering from a blood coagulation disorder such as hemophilia A or B. The method comprises administering to said individual an efficient amount of the FVIII or the complex of modified FVIII with non-modified VWF as described herein. In another embodiment, the method comprises administering to the individual an efficient amount of a polynucleotide of the invention or of a plasmid or vector of the invention. Alternatively, the method may comprise administering to the individual an efficient amount of the host cells of the invention described herein.

Expression of the proposed mutants

[0117] The production of recombinant mutant proteins at high levels in suitable host cells requires the assembly of the above-mentioned modified cDNAs into efficient transcriptional units together with suitable regulatory elements in a recombinant expression vector that can be propagated in various expression systems according to methods known to those skilled in the art. Efficient transcriptional regulatory elements could be derived from viruses having animal cells as their natural hosts or from the chromosomal DNA of animal cells. Preferably, promoter-enhancer combinations derived from the Simian Virus 40, adenovirus, BK polyoma virus, human cytomegalovirus, or the long terminal repeat of Rous sarcoma virus, or promoter-enhancer combinations including strongly constitutively transcribed genes in animal cells like beta-actin or GRP78 can be used. In order to achieve stable high levels of mRNA transcribed from the cDNAs, the transcriptional unit should contain in its 3'-proximal part a DNA region encoding a transcriptional termination-polyadenylation sequence. Preferably, this sequence is

derived from the Simian Virus 40 early transcriptional region, the rabbit beta-globin gene, or the human tissue plasminogen activator gene.

[0118] The cDNAs are then integrated into the genome of a suitable host cell line for expression of the modified FVIII and/or VWF proteins. Preferably this cell line should be an animal cell-line of vertebrate origin in order to ensure correct folding, disulfide bond formation, asparagine-linked glycosylation and other post-translational modifications as well as secretion into the cultivation medium. Examples on other post-translational modifications are tyrosine O-sulfation and proteolytic processing of the nascent polypeptide chain. Examples of cell lines that can be used are monkey COS-cells, mouse L-cells, mouse C127-cells, hamster BHK-21 cells, human embryonic kidney 293 cells, and hamster CHO-cells.

[0119] The recombinant expression vector encoding the corresponding cDNAs can be introduced into an animal cell line in several different ways. For instance, recombinant expression vectors can be created from vectors based on different animal viruses. Examples of these are vectors based on baculovirus, vaccinia virus, adenovirus, and preferably bovine papilloma virus.

[0120] The transcription units encoding the corresponding DNA's can also be introduced into animal cells together with another recombinant gene which may function as a dominant selectable marker in these cells in order to facilitate the isolation of specific cell clones which have integrated the recombinant DNA into their genome. Examples of this type of dominant selectable marker genes are Tn5 amino glycoside phosphotransferase, conferring resistance to geneticin (G418), hygromycin phosphotransferase, conferring resistance to hygromycin, and puromycin acetyl transferase, conferring resistance to puromycin. The recombinant expression vector encoding such a selectable marker can reside either on the same vector as the one encoding the cDNA of the desired protein, or it can be encoded on a separate vector which is simultaneously introduced and integrated to the genome of the host cell, frequently resulting in a tight physical linkage between the different transcription units.

[0121] Other types of selectable marker genes which can be used together with the cDNA of the desired protein are based on various transcription units encoding dihydrofolate reductase (dhfr). After introduction of this type of gene into cells lacking endogenous dhfr-activity, preferentially CHO-cells (DUKX-B11, DG-44), it will enable these to grow in media lacking nucleosides. An example of such a medium is Ham's F12 without hypoxanthine, thymidin, and glycine. These dhfr-genes can be introduced together with the FVIII cDNA transcriptional units into CHO-cells of the above type, either linked on the same vector or on different vectors, thus creating dhfr-positive cell lines producing recombinant protein.

[0122] If the above cell lines are grown in the presence of the cytotoxic dhfr-inhibitor methotrexate, new cell lines resistant to methotrexate will emerge. These cell lines may produce recombinant protein at an increased rate due to the amplified number of linked dhfr and the desired protein's transcriptional units. When propagating these cell lines in increasing concentrations of methotrexate (1-10000 nM), new cell lines can be obtained which produce

the desired protein at very high rate.

[0123] The above cell lines producing the desired protein can be grown on a large scale, either in suspension culture or on various solid supports. Examples of these supports are micro carriers based on dextran or collagen matrices, or solid supports in the form of hollow fibres or various ceramic materials. When grown in cell suspension culture or on micro carriers the culture of the above cell lines can be performed either as a bath culture or as a perfusion culture with continuous production of conditioned medium over extended periods of time. Thus, according to the present invention, the above cell lines are well suited for the development of an industrial process for the production of the desired recombinant mutant proteins

Purification and Formulation

[0124] The recombinant modified FVIII, which accumulates in the medium of secreting cells of the above types, can be concentrated and purified by a variety of biochemical and chromatographic methods, including methods utilizing differences in size, charge, hydrophobicity, solubility, specific affinity, etc. between the desired protein and other substances in the cell cultivation medium.

[0125] An example of such purification is the adsorption of the recombinant mutant protein to a monoclonal antibody, directed to e.g. a help human albumin, or directed to the respective coagulation factor, which is immobilised on a solid support. After adsorption of the modified FVIII to the support, washing and desorption, the protein can be further purified by a variety of chromatographic techniques based on the above properties. The order of the purification steps is chosen e.g. according to capacity and selectivity of the steps, stability of the support or other aspects. Preferred purification steps e.g. are but are not limited to ion exchange chromatography steps, immune affinity chromatography steps, affinity chromatography steps, hydrophobic interaction chromatography steps, dye chromatography steps, hydroxyapatite chromatography steps, multimodal chromatography steps, and size exclusion chromatography steps.

[0126] In order to minimize the theoretical risk of virus contaminations, additional steps may be included in the process that allow effective inactivation or elimination of viruses. Such steps e.g. are heat treatment in the liquid or solid state, treatment with solvents and/or detergents, radiation in the visible or UV spectrum, gamma-radiation or nanofiltration.

[0127] The modified polynucleotides (e.g. DNA) of this invention may also be integrated into a transfer vector for use in the human gene therapy.

[0128] The various embodiments described herein may be combined with each other. The present invention will be further described in more detail in the following examples thereof. This description of specific embodiments of the invention will be made in conjunction with the appended figures.

[0129] The modified FVIII as described in this invention can be formulated into pharmaceutical preparations for therapeutic use. The purified protein may be dissolved in conventional physiologically compatible aqueous buffer solutions to which there may be added, optionally, pharmaceutical excipients to provide pharmaceutical preparations.

[0130] Such pharmaceutical carriers and excipients as well as suitable pharmaceutical formulations are well known in the art (see for example "Pharmaceutical Formulation Development of Peptides and Proteins", Frokjaer et al., Taylor & Francis (2000) or "Handbook of Pharmaceutical Excipients", 3rd edition, Kibbe et al., Pharmaceutical Press (2000)). In particular, the pharmaceutical composition comprising the polypeptide variant of the invention may be formulated in lyophilized or stable liquid form. The polypeptide variant may be lyophilized by a variety of procedures known in the art. Lyophilized formulations are reconstituted prior to use by the addition of one or more pharmaceutically acceptable diluents such as sterile water for injection or sterile physiological saline solution.

[0131] Formulations of the composition are delivered to the individual by any pharmaceutically suitable means of administration. Various delivery systems are known and can be used to administer the composition by any convenient route. Preferentially, the compositions of the invention are administered systemically. For systemic use, insertion proteins of the invention are formulated for parenteral (e.g. intravenous, subcutaneous, intramuscular, intraperitoneal, intracerebral, intrapulmonar, intranasal or transdermal) or enteral (e.g., oral, vaginal or rectal) delivery according to conventional methods. The most preferential routes of administration are intravenous and subcutaneous administration. The formulations can be administered continuously by infusion or by bolus injection. Some formulations encompass slow release systems.

[0132] The insertion proteins of the present invention are administered to patients in a therapeutically effective dose, meaning a dose that is sufficient to produce the desired effects, preventing or lessening the severity or spread of the condition or indication being treated without reaching a dose which produces intolerable adverse side effects. The exact dose depends on many factors as e.g. the indication, formulation, mode of administration and has to be determined in preclinical and clinical trials for each respective indication.

[0133] The pharmaceutical composition of the invention may be administered alone or in conjunction with other therapeutic agents. These agents may be incorporated as part of the same pharmaceutical. One example of such an agent is the combination of modified FVIII with non-modified VWF.

Figures

[0134]

Figure 1: Antigen and activity levels of wild-type FVIII and FVIII-C-terminal albumin fusion polypeptides

Figure 2: Comparison of human FVIII:Ag pharmacokinetics in VWF ko mice following i.v. injection of 100 U (FVIII:Ag)/kg FVIII wildtype and FVIII-FP 1656 VWF (mean; n=4/timepoint)

Figure 3: VWF:RCo/VWF:Ag ratios of cell culture supernatants containing wt rVWF (1570/1212), rVWF-FP (1572/1212) containing C-terminally linked albumin, or a mixed expression cell culture containing a mixture of wt rVWF (1570/1212) and rVWF-FP (1572/1212) transfected in a ratio of 5:1. Values of about 0,8 were obtained in every case that are close to 1 which is the theoretical ratio of NHP according to the unit definitions.

Figure 4: SDS-Agarose gel electrophoresis of wild-type rVWF (1570/1212) expressed in HEK cells (B) and rVWF-FP (1572/1212) expressed also in HEK cells (A). Bands were detected using either antibodies to VWF or to albumin (HSA).

Figure 5: Comparison of human rVWF wildtype and rVWF-FP pharmacokinetics following i.v. injection of 100 IU VWF:Ag in rats (mean, n=2-3 /timepoint)

Examples:

Example 1: Generation of expression vectors for FVIII molecules with C-terminal albumin fusion

[0135] An expression plasmid based on pIRESpuro3 (BD Biosciences) containing the full length FVIII cDNA sequence in its multiple cloning site (pF8-FL) was first used to create a B domain deleted FVIII. For that oligonucleotides F8-1 and F8-2 (SEQ ID NO 1 and 2) were used in a site-directed mutagenesis experiment according to standard protocols (QuickChange XL Site Directed Mutagenesis Kit, Stratagene, La Jolla, CA, USA) using pF8-FL as a template to delete the B domain. In a second step a sequence encoding the amino acid sequence RRGR was introduced to connect R740 of the A2 domain with R1648 of the a3 domain. This was performed in another round of site-directed mutagenesis using primers F8-3 and F8-4 (SEQ ID NO 3 and 4). The resulting plasmid was called pF8-457.

[0136] A FVIII albumin fusion construct was generated stepwise. First, a PinAI cleavage site was introduced at the FVIII 3'terminus. For that a PCR fragment was generated using pF8-457 as template, using PCR primers We2827 and We2828 (SEQ ID NO 5 and 6), which was subsequently gel-purified, cut by restriction endonucleases BspE1 and NotI and ligated into pF8-457 previously digested with BspE1 and NotI. The resulting plasmid (pF8-1433) was then cut with enzymes PinAI and NotI and a fragment obtained by PCR on a human albumin cDNA containing plasmid using primers We 2829 and We 2830 (SEQ ID NO 7 and 8) and

subsequently digested with enzymes *PinAI* and *NotI* was inserted. The resulting expression plasmid (pF8-1434) contained the coding sequences for a B domain deleted FVIII followed by a *PinAI* site to insert linkers (encoding the amino acid sequence ThrGly) and the coding sequence for human albumin. The amino acid sequence encoded by pF8-1434 is depicted as SEQ ID NO 9.

[0137] Linker sequences separating the FVIII and albumin moieties could then easily be inserted into the newly created *PinAI* site described above. The insertion of two linker sequences is described in the following. In addition, based on pF8-1434, the TG linker might be deleted in completion and even deletions into the C-terminus of FVIII or the N-terminus of albumin can be performed using site directed mutagenesis.

[0138] Insertion of a cleavable linker, derived from the FVIII thrombin cleavage site: First a PCR fragment containing the sequence encoding the thrombin cleavage site at position 372 was generated by PCR using primers We2979 and We2980 (SEQ ID NO 10 and 11) and pF8-457 as template. This fragment was purified, digested with *PinAI* and ligated into *PinAI* digested pF8-1434. Sequencing verified insertion of correct orientation of the fragment, the resulting plasmid was called pF8-1563.

[0139] Insertion of a flexible glycine/serine linker: A PCR fragment containing the coding sequence for a 31 amino acid glycine/serine linker was amplified by PCR from pFVII-937 described in WO2007/090584 using primers We2991 and We2992 (SEQ ID NO 12 and 13). This fragment was then purified, digested by restriction endonuclease *PinAI* and ligated into *PinAI* digested pF8-1434. Sequencing verified insertion of correct orientation of the fragment, the resulting plasmid was called pF8-1568.

[0140] Using the protocols and plasmids described above and by applying molecular biology techniques known to those skilled in the art (and as described e.g. in Current Protocols in Molecular Biology, Ausubel FM et al. (eds.) John Wiley & Sons, Inc.; <http://www.currentprotocols.com/WileyCDA/>) other constructs can be made by the artisan to replace albumin by another HLEP or insert any other linker into the described *PinAI* site. Transfer of the FVIII/albumin cDNA into suitable vectors like pIRESneo3 (Invitrogen) and pEE12.4 (Lonza) permitted expression and selection of clones expressing the respective FVIII albumin fusion protein in CHO cells.

Example 2: Transfection and expression of FVIII and VWF proteins

[0141] Expression plasmids were grown up in *E.coli* TOP10 (Invitrogen, Carlsbad, CA, USA) and purified using standard protocols (Qiagen, Hilden, Germany). HEK-293 (Invitrogen) cells were transfected using the Lipofectamine 2000 reagent (Invitrogen) and grown up in serum-free medium (Invitrogen 293 Express) in the presence of 4 µg/ml Puromycin and optionally 0.5 IU/ml VWF. CHO cells (CHO-S, Invitrogen; CHOK1SV, Lonza) were transfected using the Lipofectamine 2000 reagent (Invitrogen) and grown up in serum-free medium (Invitrogen CD

CHO, 6 mM glutamine for CHO-S and CD-CHO for CHOK1SV) in the presence of 500-1000 µg/ml Geneticin (CHO-S only). For FVIII expression optionally 0.5 IU/ml VWF were added. For vWF expression an expression plasmid encoding PACE/furin (pFu-797) as described in WO2007/144173 was cotransfected. In another experiment two plasmids encoding VWF wild-type and VWF fused at the C-terminus to albumin were cotransfected with pFu-797 resulting in VWF multimeres with wild-type VWF monomers and albumin-fused VWF monomers (see figure 3). Transfected cell populations were spread through T-flasks into roller bottles or small scale fermenters from which supernatants were harvested for purification.

[0142] Table 2 lists HEK-293 expression data of the constructs described in example 1.

Table 2:

Construct	Activity [U/mL]
pF8-457	1.54
pF8-457 + 0.5 U/ml VWF	1.66
pF8-1434	1.59
pF8-1434 + 0.5 U/ml VWF	1.82
pF8-1563 + 0.5 U/ml VWF	2.04
pF8-1568 + 0.5 U/ml VWF	1.21

Example 3: Increased expression rate of FVIII albumin fusion protein

[0143] Figure 1 summarizes the results of an expression study of a FVIII albumin fusion protein in serum-free cell culture. HEK-293 cells were transfected in triplicate with pF8-1434 (FVIII C-terminal albumin fusion) and pF8-457 (FVIII wild-type), respectively, seeded into T80 flasks with equal cell numbers and grown in the absence of stabilizing VWF. Culture supernatant was then harvested after 96, 120 and 144 hours and tested for FVIII activity.

[0144] The results demonstrated an expression enhancing effect of the albumin moiety when present as an integral part of the FVIII molecule in cell culture. Consequently, the productivity was clearly improved in the case of the fusion protein compared to wild-type FVIII (Figure 1).

Example 4: Purification of FVIII proteins

[0145] To the expression supernatant containing the FVIII molecule a sufficient amount of an immune affinity resin was added to bind the FVIII activity almost completely. The immune affinity resin had been prepared by binding an appropriate anti-FVIII MAb covalently to Sephacryl S1000 resin used as a support. After washing of the resin it was filled into a chromatography column and washed again. Elution was done using a buffer containing 250

mM CaCl₂ and 50% ethylene glycol.

[0146] The immune affinity chromatography (IAC) fractions containing FVIII:C activity were pooled, dialyzed against formulation buffer (excipients: sodium chloride, sucrose, histidine, calcium chloride, and Tween 80), and concentrated. Samples were either stored frozen or freeze-dried using an appropriate freeze-drying cycle.

[0147] Alternatively, the FVIII containing cell culture supernatant is concentrated/purified by a first ion exchange chromatography followed by further purification using immune affinity chromatography (IAC). In this case the eluate of the ion exchange chromatography is loaded onto an IAC column using the above mentioned resin.

Example 5: Analysis of FVIII activity and antigen

[0148] For activity determination of FVIII:C in vitro either a clotting assay (e.g. Pathromtin SL reagent and FVIII deficient plasma delivered by Dade Behring, Germany) or a chromogenic assay (e.g. Coamatic FVIII:C assay delivered by Haemochrom) were used. The assays were performed according to the manufacturers instructions.

[0149] FVIII antigen (FVIII:Ag) was determined by an ELISA whose performance is known to those skilled in the art. Briefly, microplates were incubated with 100 µL per well of the capture antibody (sheep anti-human FVIII IgG, Cedarlane CL20035K-C, diluted 1:200 in Buffer A [Sigma C3041]) for 2 hours at ambient temperature. After washing plates three times with buffer B (Sigma P3563), serial dilutions of the test sample in sample diluent buffer (Cedarlane) as well as serial dilutions of a FVIII preparation (CSL Behring; 200 - 2 mU/mL) in sample diluent buffer (volumes per well: 100 µL) were incubated for two hours at ambient temperature. After three wash steps with buffer B, 100 µL of a 1:2 dilution in buffer B of the detection antibody (sheep anti-human FVIII IgG, Cedarlane CL20035K-D, peroxidase labelled) were added to each well and incubated for another hour at ambient temperature. After three wash steps with buffer B, 100 µL of substrate solution (1:10 (v/v) TMB OUVF : TMB Buffer OUVG, Dade Behring) were added per well and incubated for 30 minutes at ambient temperature in the dark. Addition of 100 µL stop solution (Dade Behring, OSFA) prepared the samples for reading in a suitable microplate reader at 450 nm wavelength. Concentrations of test samples were then calculated using the standard curve with the FVIII preparation as reference.

Example 6: Assessment of Pharmacokinetics of FVIII-FP in VWF ko mice following a single i.v. injection

[0150] In order to compare the pharmacokinetics of FVIII wildtype (DNA 457) and a C-terminal FVIII-FP (DNA 1656), both FVIII variants were administered intravenously to mice. A VWF ko mouse strain (Denis C. et al, Proc. Natl. Acad. Sci. USA, 1998, Vol 95, 9524-9529) was chosen

because, amongst other functions, VWF serves as a carrier and stabilizing protein for FVIII, thereby protecting FVIII from premature degradation, e.g. by proteases, and from premature elimination from circulation. For unmodified FVIII an undisturbed interaction with VWF is essential as exemplified by hemophilia A cases, caused by mutation in the C terminal region resulting in decreasing binding to VWF. In the case of modified FVIII such binding may, however, be even unwanted, in order to examine or achieve improved pharmacokinetics. Accordingly both products were injected i.v. at a dose of 100 U (FVIII:Ag)/kg as bolus to two groups of mice (Tab. 3). Blood was sampled retroorbitally at appropriate intervals starting at 5 minutes after application of the test substances and up to 24 hours. One blood sample / mouse was taken, processed to plasma and stored frozen at -20°C until analysis. Human FVIII:Ag concentration was quantified using an ELISA assay specific for human FVIII or by a mixed ELISA specific for human albumin and FVIII, respectively. The mean plasma concentration of the, for each timepoint pooled, samples was used for calculation of pharmacokinetic parameters. Half-life was calculated using the time points of the beta phase of elimination according to the formula $t_{1/2} = \ln 2 / k$, whereas k is the slope of the regression line. The result is depicted in Figure 2. Surprisingly, FVIII-FP 1656 ($t_{1/2} = 3,06$ h, between 5 and 960 min) had an about 3-4 times longer terminal half-life as compared to FVIII wildtype ($t_{1/2} = 0,8$ h, between 5 and 240 min). In addition, the recovery of FVIII-FP 1656 was increased by about 20% as compared to wildtype FVIII (Tab. 4).

Table 3: Treatment groups for comparison of pharmacokinetics FVIII in VWF ko mice

Treatment	Dose (FVIII:C) / volume / schedule / route	N
FVIII wildtype	100 U (FVIII:Ag)/kg / 0.2 mL/20g b.w. / t=0 h /i.v..	24
FVIII-FP 1656	100 U(FVIII:Ag)/kg / 0.2 mL/20g b.w. / t=0 h /i.v..	24

Table 4: Bioavailability (%) of FVIII wildtype and modified FVIII, FVIII-FP 1656, upon i.v. injection into VWF ko mice

Treatment	Bioavailability (%)
FVIII wildtype	100
FVIII-FP 1656	120,4

Example 7: Generation of expression vectors for VWF wild-type and VWF albumin fusion proteins

[0151] An expression plasmid containing the full length VWF cDNA sequence in its multiple cloning site was generated first. For that the coding sequence of VWF was amplified by polymerase chain reaction (PCR) using primer set VWF+ and VWF- (SEQ ID NO. 17 and 18) under standard conditions known to those skilled in the art (and as described e.g. in Current Protocols in Molecular Biology, Ausubel FM et al. (eds.) John Wiley & Sons, Inc.;

<http://www.currentprotocols.com/WileyCDA/>) from a plasmid containing VWF cDNA (as obtainable commercially, e.g. pMT2-VWF from ATCC, No. 67122). The resulting PCR fragment was digested by restriction endonuclease EcoRI and ligated into expression vector pRESpuRO3 (BD Biosciences, Franklin Lakes, NJ, USA) which had been linearized by EcoRI. The resulting expression plasmid containing the wild-type cDNA of VWF downstream of the CMV promoter was called pVWF-1570.

[0152] A PCR fragment containing the coding sequence for a 31 amino acid glycine/serine linker and the human albumin cDNA was amplified from pFVII-937 described in WO2007/090584 using primers We2994 and We1335 (SEQ ID NO. 19 and 20). This PCR fragment was then digested by restriction endonuclease NotI and ligated into NotI digested pVWF-1570. The resulting plasmid containing the coding sequences of VWF wt, the linker sequence and human albumin was called pVWF-1574.

[0153] In order to achieve expression of a fusion protein several bases had to be deleted between VWF and the linker sequence. This was performed by site directed mutagenesis according to standard protocols (QuickChange XL Site Directed Mutagenesis Kit, Stratagene, La Jolla, CA, USA) using oligonucleotides We2995 and We2996 (SEQ ID NO 21 and 22). The resulting expression plasmid called pVWF-1572 contained the coding sequences of VWF in frame with that of a 31 amino acid glycin/serine linker and human albumin. The amino acid sequence of the expressed rVWF-FP is outlined as SEQ ID No. 25. The amino acid sequence of the human VWF preproprotein is outlined as SEQ ID NO. 24.

[0154] Using the protocols and plasmids described above and by applying molecular biology techniques known to those skilled in the art (and as described e.g. in Current Protocols in Molecular Biology, *ibid*) other constructs can be made by the artisan for replacement of the albumin sequence by another HLEP sequence or the linker sequence by another linker sequence.

Example 8: Purification of VWF and VWF albumin fusion proteins

[0155] Cell culture supernatants containing VWF wild-type (rVWF wt) or VWF albumin fusion protein (rVWF-FP) were sterile-filtered through a 0,2µm filter and dialysed against equilibration buffer (EB; 10mM Tris-HCl, 10mM CaCl₂, pH 7.0). This material was then applied to a Heparin Fractogel column equilibrated with EB. The column was washed with EB and VWF proteins were eluted with 500mM NaCl in EB. The elution peak was concentrated and dialysed against FB buffer (3g/L sodium chloride, 20 g/L glycine, 5.5 g/L trisodium citrate dihydrate, pH 7.0). Finally the material was sterile filtrated and frozen in aliquots. If needed, further purification steps were applied comprising anion and/or cation exchange chromatography, HIC and SEC.

Example 9: Analysis of VWF activity and antigen

[0156] Samples were analysed by immunoturbidimetric determination of VWF:Ag (OPAB03, Siemens Healthcare Diagnostics, Marburg, Germany) and for collagen binding (Technozym VWF:CBA ELISA, Ref. 5450301 with calibrator set 5450310 and control set 5450312, Technoclone, Vienna, Austria) as described by the manufacturer.

[0157] VWF:RCo testing was done using the BC VWF reagent of Siemens Healthcare Diagnostics, Marburg, Germany according to the manufacturers description. The International Concentrate Standard was used as a primary standard preparation to calibrate an in-house standard preparation for day to day use.

[0158] The ratios of VWF:RCo and VWF:Ag assays are calculated in order to compare this parameter for different constructs tested. As is shown in figure 3 the VWF:RCo/VWF:Ag ratio was comparable for wt rVWF and the C-terminal rVWF-albumin fusion protein.

[0159] For pharmacokinetic analyses VWF antigen was determined by an ELISA whose performance is known to those skilled in the art. Briefly, microplates were incubated with 100 µL per well of the capture antibody (rabbit anti human VWF-IgG, Dako A0082 [Dako, Hamburg, Germany], diluted 1:2000 in buffer A [Sigma C3041, Sigma-Aldrich, Munich, Germany]) overnight at ambient temperature. After washing plates three times with buffer B (Sigma P3563), each well was incubated with 200 µL buffer C (Sigma P3688) for 1.5 hours at ambient temperature (blocking). After another three wash steps with buffer B, serial dilutions of the test sample in buffer B as well as serial dilutions of standard human plasma (ORKL21; 20 - 0.2 mU/mL; Siemens Healthcare Diagnostics, Marburg, Germany) in buffer B (volumes per well: 100 µL) were incubated for 1.5 hours at ambient temperature. After three wash steps with buffer B, 100 µL of a 1:16000 dilution in buffer B of the detection antibody (rabbit anti human VWF-IgG, Dako P0226, peroxidase labelled) were added to each well and incubated for 1 hour at ambient temperature. After three wash steps with buffer B, 100 µL of substrate solution (OUVF, Siemens Healthcare Diagnostics) were added per well and incubated for 30 minutes at ambient temperature in the dark. Addition of 100 µL undiluted stop dilution (OSFA, Siemens Healthcare Diagnostics) prepared the samples for reading in a suitable microplate reader at 450 nm wavelength. Concentrations of the test samples were then calculated using the standard curve with standard human plasma as reference.

Example 10: Multimer analysis of VWF and VWF albumin fusion proteins

[0160] VWF Multimer analysis was performed by SDS-agarose gel electrophoresis as recently described (Tatewaki et al., Thromb. Res. 52: 23-32 (1988), and Metzner et al., Haemophilia 4 (Suppl. 3): 25-32 (1998)) with minor modifications. Briefly, after equilibration in running buffer ready to use 1% agarose mini gels (BioRad) were used to standardize the method as far as possible. Comparable amounts of VWF antigen were subjected to electrophoresis on the SDS-agarose gels. After Western blotting the VWF protein bands were detected using anti-VWF (DAKO, prod. No. 0854) or anti-albumin antibodies followed by alkaline phosphatase labelled

anti-IgG antibodies (SIGMA, prod. No. 1305) and colour reaction quantified by densitometry.

[0161] Using wild-type rVWF (1570/797) and rVWF-FP (1572/797) it could be demonstrated by Western blotting and detection using anti-albumin or anti VWF antibodies that rVWF-FP forms a regular multimer distribution detected both by anti-albumin and anti-VWF antibodies (Figure 4). This confirms that although every subunit of the multimeric VWF contains albumin, a regular VWF multimer pattern is formed. The albumin moiety obviously does neither inhibit the N-terminal dimerization nor the C-terminal multimerization of the VWF molecules.

Example 11: Assessment of pharmacokinetics of VWF and VWF albumin fusion protein in rats following a single i.v. injection

[0162] rVWF-FP and rVWF wt were administered intravenously to a total of 4 CD rats each. The dose was 100 U (VWF:Ag)/kg body weight, at an injection volume of 4 mL/kg. Blood samples were drawn retroorbitally at appropriate intervals starting at 5 minutes after application of the test substances, using an alternating sampling scheme, resulting in samples from 2 animals / timepoint (t=0, 5, 30, 90 min, 4h, 1d for subset Nr. 1 and 0, 15 min, 1, 2, 8 h and 2 d for subset Nr. 2). The scheme was designed to minimize potential effects of blood sampling on the plasma concentration to be quantified. Blood was processed to plasma and stored deep frozen until analysis. The VWF:Ag level in plasma was subsequently quantified by an ELISA as described in Example 9. The mean plasma concentration was used for calculation of pharmacokinetic parameters. Half-life was calculated using the time points of the beta phase of elimination according to the formula $t_{1/2} = \ln 2 / k$, whereas k is the slope of the regression line.

[0163] The result is depicted in Figure 5 (n=2/timepoint; mean). The terminal half-lives were calculated to be 32.4 min. for the rVWF-FP and 2.6 min. for rVWF wt. Recovery was also improved for the rVWF-FP with 42.1% compared to 16.1% for rVWF wt.

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His Pro 1250	Thr His Tyr Ser 1255	Ile Arg Ser Thr Leu 1260	Arg Met Glu Leu 1265
Met Gly 1265	Cys Asp Leu Asn 1270	Ser Cys Ser Met Pro 1275	Leu Gly Met Glu 1280
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Val Lys 1355	Glu Phe Leu Ile 1360	Ser Ser Gln Asp Gly 1365	His Gln Trp 1370
Thr Leu 1370	Phe Phe Gln Asn 1375	Gly Lys Val Lys Val 1380	Phe Gln Gly Asn 1385
Gln Asp 1385	Ser Phe Thr Pro 1390	Val Val Asn Ser Leu 1395	Asp Pro Pro Leu 1400
Leu Thr 1400	Arg Tyr Leu Arg 1405	Ile His Pro Gln Ser 1410	Trp Val His Gln 1415
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Tyr Thr 1430	Gly Asp Ala His 1435	Lys Ser Glu Val Ala 1440	His Arg Phe Lys 1445
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Val Gly 1865	Ser Lys Cys Cys Lys 1870	His Pro Glu Ala Lys 1875	Arg Met Pro
Cys Ala 1880	Glu Asp Tyr Leu Ser 1885	Val Val Leu Asn Gln 1890	Leu Cys Val
Leu His 1895	Glu Lys Thr Pro Val 1900	Ser Asp Arg Val Thr 1905	Lys Cys Cys
Thr Glu 1910	Ser Leu Val Asn Arg 1915	Arg Pro Cys Phe Ser 1920	Ala Leu Glu
Val Asp 1925	Glu Thr Tyr Val Pro 1930	Lys Glu Phe Asn Ala 1935	Glu Thr Phe
Thr Phe 1940	His Ala Asp Ile Cys 1945	Thr Leu Ser Glu Lys 1950	Glu Arg Gln
Ile Lys 1955	Lys Gln Thr Ala Leu 1960	Val Glu Leu Val Lys 1965	His Lys Pro
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515

520

525

Leu Thr Arg Tyr Tyr Ser Ser Phe Val Asn Met Glu Arg Asp Leu Ala
530 535 540

Ser Gly Leu Ile Gly Pro Leu Leu Ile Cys Tyr Lys Glu Ser Val Asp
545 550 555 560

Gln Arg Gly Asn Gln Ile Met Ser Asp Lys Arg Asn Val Ile Leu Phe
565 570 575

Ser Val Phe Asp Glu Asn Arg Ser Trp Tyr Leu Thr Glu Asn Ile Gln
580 585 590

Arg Phe Leu Pro Asn Pro Ala Gly Val Gln Leu Glu Asp Pro Glu Phe
595 600 605

Gln Ala Ser Asn Ile Met His Ser Ile Asn Gly Tyr Val Phe Asp Ser
610 615 620

Leu Gln Leu Ser Val Cys Leu His Glu Val Ala Tyr Trp Tyr Ile Leu
625 630 635 640

Ser Ile Gly Ala Gln Thr Asp Phe Leu Ser Val Phe Phe Ser Gly Tyr
645 650 655

Thr Phe Lys His Lys Met Val Tyr Glu Asp Thr Leu Thr Leu Phe Pro
660 665 670

Phe Ser Gly Glu Thr Val Phe Met Ser Met Glu Asn Pro Gly Leu Trp
675 680 685

Ile Leu Gly Cys His Asn Ser Asp Phe Arg Asn Arg Gly Met Thr Ala
690 695 700

Leu Leu Lys Val Ser Ser Cys Asp Lys Asn Thr Gly Asp Tyr Tyr Glu
705 710 715 720

Asp Ser Tyr Glu Asp Ile Ser Ala Tyr Leu Leu Ser Lys Asn Asn Ala
725 730 735

Ile Glu Pro Arg Ser Phe Ser Gln Asn Ser Arg His Arg Ser Thr Arg
740 745 750

Gln Lys Gln Phe Asn Ala Thr Thr Ile Pro Glu Asn Asp Ile Glu Lys
755 760 765

Thr Asp Pro Trp Phe Ala His Arg Thr Pro Met Pro Lys Ile Gln Asn
770 775 780

Val Ser Ser Ser Asp Leu Leu Met Leu Leu Arg Gln Ser Pro Thr Pro
785 790 795 800

His Gly Leu Ser Leu Ser Asp Leu Gln Glu Ala Lys Tyr Glu Thr Phe
 805 810 815
 Ser Asp Asp Pro Ser Pro Gly Ala Ile Asp Ser Asn Asn Ser Leu Ser
 820 825 830
 Glu Met Thr His Phe Arg Pro Gln Leu His His Ser Gly Asp Met Val
 835 840 845
 Phe Thr Pro Glu Ser Gly Leu Gln Leu Arg Leu Asn Glu Lys Leu Gly
 850 855 860
 Thr Thr Ala Ala Thr Glu Leu Lys Lys Leu Asp Phe Lys Val Ser Ser
 865 870 875 880
 Thr Ser Asn Asn Leu Ile Ser Thr Ile Pro Ser Asp Asn Leu Ala Ala
 885 890 895
 Gly Thr Asp Asn Thr Ser Ser Leu Gly Pro Pro Ser Met Pro Val His
 900 905 910
 Tyr Asp Ser Gln Leu Asp Thr Thr Leu Phe Gly Lys Lys Ser Ser Pro
 915 920 925
 Leu Thr Glu Ser Gly Gly Pro Leu Ser Leu Ser Glu Glu Asn Asn Asp
 930 935 940
 Ser Lys Leu Leu Glu Ser Gly Leu Met Asn Ser Gln Glu Ser Ser Trp
 945 950 955 960
 Gly Lys Asn Val Ser Ser Thr Glu Ser Gly Arg Leu Phe Lys Gly Lys
 965 970 975
 Arg Ala His Gly Pro Ala Leu Leu Thr Lys Asp Asn Ala Leu Phe Lys
 980 985 990
 Val Ser Ile Ser Leu Leu Lys Thr Asn Lys Thr Ser Asn Asn Ser Ala
 995 1000 1005
 Thr Asn Arg Lys Thr His Ile Asp Gly Pro Ser Leu Leu Ile Glu
 1010 1015 1020
 Asn Ser Pro Ser Val Trp Gln Asn Ile Leu Glu Ser Asp Thr Glu
 1025 1030 1035
 Phe Lys Lys Val Thr Pro Leu Ile His Asp Arg Met Leu Met Asp
 1040 1045 1050
 Lys Asn Ala Thr Ala Leu Arg Leu Asn His Met Ser Asn Lys Thr
 1055 1060 1065
 Thr Ser Ser Lys Asn Met Glu Met Val Gln Gln Lys Lys Glu Gly
 1070 1075 1080
 Pro Ile Pro Pro Asp Ala Gln Asn Pro Asp Met Ser Phe Phe Lys
 1085 1090 1095

Met	Leu	Phe	Leu	Pro	Glu	Ser	Ala	Arg	Trp	Ile	Gln	Arg	Thr	His
1100						1105					1110			
Gly	Lys	Asn	Ser	Leu	Asn	Ser	Gly	Gln	Gly	Pro	Ser	Pro	Lys	Gln
1115						1120					1125			
Leu	Val	Ser	Leu	Gly	Pro	Glu	Lys	Ser	Val	Glu	Gly	Gln	Asn	Phe
1130						1135					1140			
Leu	Ser	Glu	Lys	Asn	Lys	Val	Val	Val	Gly	Lys	Gly	Glu	Phe	Thr
1145						1150					1155			
Lys	Asp	Val	Gly	Leu	Lys	Glu	Met	Val	Phe	Pro	Ser	Ser	Arg	Asn
1160						1165					1170			
Leu	Phe	Leu	Thr	Asn	Leu	Asp	Asn	Leu	His	Glu	Asn	Asn	Thr	His
1175						1180					1185			
Asn	Gln	Glu	Lys	Lys	Ile	Gln	Glu	Glu	Ile	Glu	Lys	Lys	Glu	Thr
1190						1195					1200			
Leu	Ile	Gln	Glu	Asn	Val	Val	Leu	Pro	Gln	Ile	His	Thr	Val	Thr
1205						1210					1215			
Gly	Thr	Lys	Asn	Phe	Met	Lys	Asn	Leu	Phe	Leu	Leu	Ser	Thr	Arg
1220						1225					1230			
Gln	Asn	Val	Glu	Gly	Ser	Tyr	Asp	Gly	Ala	Tyr	Ala	Pro	Val	Leu
1235						1240					1245			
Gln	Asp	Phe	Arg	Ser	Leu	Asn	Asp	Ser	Thr	Asn	Arg	Thr	Lys	Lys
1250						1255					1260			
His	Thr	Ala	His	Phe	Ser	Lys	Lys	Gly	Glu	Glu	Glu	Asn	Leu	Glu
1265						1270					1275			
Gly	Leu	Gly	Asn	Gln	Thr	Lys	Gln	Ile	Val	Glu	Lys	Tyr	Ala	Cys
1280						1285					1290			
Thr	Thr	Arg	Ile	Ser	Pro	Asn	Thr	Ser	Gln	Gln	Asn	Phe	Val	Thr
1295						1300					1305			
Gln	Arg	Ser	Lys	Arg	Ala	Leu	Lys	Gln	Phe	Arg	Leu	Pro	Leu	Glu
1310						1315					1320			
Glu	Thr	Glu	Leu	Glu	Lys	Arg	Ile	Ile	Val	Asp	Asp	Thr	Ser	Thr
1325						1330					1335			
Gln	Trp	Ser	Lys	Asn	Met	Lys	His	Leu	Thr	Pro	Ser	Thr	Leu	Thr
1340						1345					1350			
Gln	Ile	Asp	Tyr	Asn	Glu	Lys	Glu	Lys	Gly	Ala	Ile	Thr	Gln	Ser
1355						1360					1365			
Pro	Leu	Ser	Asp	Cys	Leu	Thr	Arg	Ser	His	Ser	Ile	Pro	Gln	Ala
1370						1375					1380			

Asn Arg	Ser Pro Leu Pro	Ile	Ala Lys Val Ser Ser	Phe Pro Ser
1385		1390		1395
Ile Arg	Pro Ile Tyr Leu Thr	Arg Val Leu Phe Gln	Asp Asn Ser	
1400		1405		1410
Ser His	Leu Pro Ala Ala Ser	Tyr Arg Lys Lys Asp	Ser Gly Val	
1415		1420		1425
Gln Glu	Ser Ser His Phe Leu	Gln Gly Ala Lys Lys	Asn Asn Leu	
1430		1435		1440
Ser Leu	Ala Ile Leu Thr Leu	Glu Met Thr Gly Asp	Gln Arg Glu	
1445		1450		1455
Val Gly	Ser Leu Gly Thr Ser	Ala Thr Asn Ser Val	Thr Tyr Lys	
1460		1465		1470
Lys Val	Glu Asn Thr Val Leu	Pro Lys Pro Asp Leu	Pro Lys Thr	
1475		1480		1485
Ser Gly	Lys Val Glu Leu Leu	Pro Lys Val His Ile	Tyr Gln Lys	
1490		1495		1500
Asp Leu	Phe Pro Thr Glu Thr	Ser Asn Gly Ser Pro	Gly His Leu	
1505		1510		1515
Asp Leu	Val Glu Gly Ser Leu	Leu Gln Gly Thr Glu	Gly Ala Ile	
1520		1525		1530
Lys Trp	Asn Glu Ala Asn Arg	Pro Gly Lys Val Pro	Phe Leu Arg	
1535		1540		1545
Val Ala	Thr Glu Ser Ser Ala	Lys Thr Pro Ser Lys	Leu Leu Asp	
1550		1555		1560
Pro Leu	Ala Trp Asp Asn His	Tyr Gly Thr Gln Ile	Pro Lys Glu	
1565		1570		1575
Glu Trp	Lys Ser Gln Glu Lys	Ser Pro Glu Lys Thr	Ala Phe Lys	
1580		1585		1590
Lys Lys	Asp Thr Ile Leu Ser	Leu Asn Ala Cys Glu	Ser Asn His	
1595		1600		1605
Ala Ile	Ala Ala Ile Asn Glu	Gly Gln Asn Lys Pro	Glu Ile Glu	
1610		1615		1620
Val Thr	Trp Ala Lys Gln Gly	Arg Thr Glu Arg Leu	Cys Ser Gln	
1625		1630		1635
Asn Pro	Pro Val Leu Lys Arg	His Gln Arg Glu Ile	Thr Arg Thr	
1640		1645		1650
Thr Leu	Gln Ser Asp Gln Glu	Glu Ile Asp Tyr Asp	Asp Thr Ile	
1655		1660		1665

1655	1660	1665
Ser Val 1670	Glu Met Lys Lys Glu 1675	Asp Phe Asp Ile Tyr 1680
Glu Asn 1685	Gln Ser Pro Arg Ser 1690	Phe Gln Lys Lys Thr 1695
Phe Ile 1700	Ala Ala Val Glu Arg 1705	Leu Trp Asp Tyr Gly 1710
Ser Pro 1715	His Val Leu Arg Asn 1720	Arg Ala Gln Ser Gly 1725
Gln Phe 1730	Lys Lys Val Val Phe 1735	Gln Glu Phe Thr Asp 1740
Thr Gln 1745	Pro Leu Tyr Arg Gly 1750	Glu Leu Asn Glu His 1755
Leu Gly 1760	Pro Tyr Ile Arg Ala 1765	Glu Val Glu Asp Asn 1770
Thr Phe 1775	Arg Asn Gln Ala Ser 1780	Arg Pro Tyr Ser Phe 1785
Leu Ile 1790	Ser Tyr Glu Glu Asp 1795	Gln Arg Gln Gly Ala 1800
Lys Asn 1805	Phe Val Lys Pro Asn 1810	Glu Thr Lys Thr Tyr 1815
Val Gln 1820	His His Met Ala Pro 1825	Thr Lys Asp Glu Phe 1830
Ala Trp 1835	Ala Tyr Phe Ser Asp 1840	Val Asp Leu Glu Lys 1845
Ser Gly 1850	Leu Ile Gly Pro Leu 1855	Leu Val Cys His Thr 1860
Asn Pro 1865	Ala His Gly Arg Gln 1870	Val Thr Val Gln Glu 1875
Phe Phe 1880	Thr Ile Phe Asp Glu 1885	Thr Lys Ser Trp Tyr 1890
Asn Met 1895	Glu Arg Asn Cys Arg 1900	Ala Pro Cys Asn Ile 1905
Asp Pro 1910	Thr Phe Lys Glu Asn 1915	Tyr Arg Phe His Ala 1920
Tyr Ile 1925	Met Asp Thr Leu Pro 1930	Gly Leu Val Met Ala 1935
Arg Ile 1940	Arg Trp Tyr Leu Leu 1945	Ser Met Gly Ser Asn 1950
		Glu Asn Ile

1940				1945				1950						
His	Ser	Ile	His	Phe	Ser	Gly	His	Val	Phe	Thr	Val	Arg	Lys	Lys
1955						1960					1965			
Glu	Glu	Tyr	Lys	Met	Ala	Leu	Tyr	Asn	Leu	Tyr	Pro	Gly	Val	Phe
1970						1975					1980			
Glu	Thr	Val	Glu	Met	Leu	Pro	Ser	Lys	Ala	Gly	Ile	Trp	Arg	Val
1985						1990					1995			
Glu	Cys	Leu	Ile	Gly	Glu	His	Leu	His	Ala	Gly	Met	Ser	Thr	Leu
2000						2005					2010			
Phe	Leu	Val	Tyr	Ser	Asn	Lys	Cys	Gln	Thr	Pro	Leu	Gly	Met	Ala
2015						2020					2025			
Ser	Gly	His	Ile	Arg	Asp	Phe	Gln	Ile	Thr	Ala	Ser	Gly	Gln	Tyr
2030						2035					2040			
Gly	Gln	Trp	Ala	Pro	Lys	Leu	Ala	Arg	Leu	His	Tyr	Ser	Gly	Ser
2045						2050					2055			
Ile	Asn	Ala	Trp	Ser	Thr	Lys	Glu	Pro	Phe	Ser	Trp	Ile	Lys	Val
2060						2065					2070			
Asp	Leu	Leu	Ala	Pro	Met	Ile	Ile	His	Gly	Ile	Lys	Thr	Gln	Gly
2075						2080					2085			
Ala	Arg	Gln	Lys	Phe	Ser	Ser	Leu	Tyr	Ile	Ser	Gln	Phe	Ile	Ile
2090						2095					2100			
Met	Tyr	Ser	Leu	Asp	Gly	Lys	Lys	Trp	Gln	Thr	Tyr	Arg	Gly	Asn
2105						2110					2115			
Ser	Thr	Gly	Thr	Leu	Met	Val	Phe	Phe	Gly	Asn	Val	Asp	Ser	Ser
2120						2125					2130			
Gly	Ile	Lys	His	Asn	Ile	Phe	Asn	Pro	Pro	Ile	Ile	Ala	Arg	Tyr
2135						2140					2145			
Ile	Arg	Leu	His	Pro	Thr	His	Tyr	Ser	Ile	Arg	Ser	Thr	Leu	Arg
2150						2155					2160			
Met	Glu	Leu	Met	Gly	Cys	Asp	Leu	Asn	Ser	Cys	Ser	Met	Pro	Leu
2165						2170					2175			
Gly	Met	Glu	Ser	Lys	Ala	Ile	Ser	Asp	Ala	Gln	Ile	Thr	Ala	Ser
2180						2185					2190			
Ser	Tyr	Phe	Thr	Asn	Met	Phe	Ala	Thr	Trp	Ser	Pro	Ser	Lys	Ala
2195						2200					2205			
Arg	Leu	His	Leu	Gln	Gly	Arg	Ser	Asn	Ala	Trp	Arg	Pro	Gln	Val
2210						2215					2220			
Asn	Asn	Pro	Lys	Glu	Trp	Leu	Gln	Val	Asp	Phe	Gln	Lys	Thr	Met
2225						2230					2235			

Lys Val Thr Gly Val Thr Thr Gln Gly Val Lys Ser Leu Leu Thr
2240 2245 2250

Ser Met Tyr Val Lys Glu Phe Leu Ile Ser Ser Ser Gln Asp Gly
2255 2260 2265

His Gln Trp Thr Leu Phe Phe Gln Asn Gly Lys Val Lys Val Phe
2270 2275 2280

Gln Gly Asn Gln Asp Ser Phe Thr Pro Val Val Asn Ser Leu Asp
2285 2290 2295

Pro Pro Leu Leu Thr Arg Tyr Leu Arg Ile His Pro Gln Ser Trp
2300 2305 2310

Val His Gln Ile Ala Leu Arg Met Glu Val Leu Gly Cys Glu Ala
2315 2320 2325

Gln Asp Leu Tyr
2330

<210> 16

<211> 585

<212> PRT

<213> Homo sapiens

<400> 16

Asp Ala His Lys Ser Glu Val Ala His Arg Phe Lys Asp Leu Gly Glu
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Glu Asn Phe Lys Ala Leu Val Leu Ile Ala Phe Ala Gln Tyr Leu Gln
20 25 30

Gln Cys Pro Phe Glu Asp His Val Lys Leu Val Asn Glu Val Thr Glu
35 40 45

Phe Ala Lys Thr Cys Val Ala Asp Glu Ser Ala Glu Asn Cys Asp Lys
50 55 60

Ser Leu His Thr Leu Phe Gly Asp Lys Leu Cys Thr Val Ala Thr Leu
65 70 75 80

Arg Glu Thr Tyr Gly Glu Met Ala Asp Cys Cys Ala Lys Gln Glu Pro
85 90 95

Glu Arg Asn Glu Cys Phe Leu Gln His Lys Asp Asp Asn Pro Asn Leu
100 105 110

Pro Arg Leu Val Arg Pro Glu Val Asp Val Met Cys Thr Ala Phe His
115 120 125

Asp Asn Glu Glu Thr Phe Leu Lys Lys Tyr Leu Tyr Glu Ile Ala Arg
130 135 140

Arg His Pro Tyr Phe Tyr Ala Pro Glu Leu Leu Phe Phe Ala Lys Arg
145 150 155 160

Tyr Lys Ala Ala Phe Thr Glu Cys Cys Gln Ala Ala Asp Lys Ala Ala
165 170 175

Cys Leu Leu Pro Lys Leu Asp Glu Leu Arg Asp Glu Gly Lys Ala Ser
180 185 190

Ser Ala Lys Gln Arg Leu Lys Cys Ala Ser Leu Gln Lys Phe Gly Glu
195 200 205

Arg Ala Phe Lys Ala Trp Ala Val Ala Arg Leu Ser Gln Arg Phe Pro
210 215 220

Lys Ala Glu Phe Ala Glu Val Ser Lys Leu Val Thr Asp Leu Thr Lys
225 230 235 240

Val His Thr Glu Cys Cys His Gly Asp Leu Leu Glu Cys Ala Asp Asp
245 250 255

Arg Ala Asp Leu Ala Lys Tyr Ile Cys Glu Asn Gln Asp Ser Ile Ser
260 265 270

Ser Lys Leu Lys Glu Cys Cys Glu Lys Pro Leu Leu Glu Lys Ser His
275 280 285

Cys Ile Ala Glu Val Glu Asn Asp Glu Met Pro Ala Asp Leu Pro Ser
290 295 300

Leu Ala Ala Asp Phe Val Glu Ser Lys Asp Val Cys Lys Asn Tyr Ala
305 310 315 320

Glu Ala Lys Asp Val Phe Leu Gly Met Phe Leu Tyr Glu Tyr Ala Arg
325 330 335

Arg His Pro Asp Tyr Ser Val Val Leu Leu Arg Leu Ala Lys Thr
340 345 350

Tyr Glu Thr Thr Leu Glu Lys Cys Cys Ala Ala Ala Asp Pro His Glu
355 360 365

Cys Tyr Ala Lys Val Phe Asp Glu Phe Lys Pro Leu Val Glu Glu Pro
370 375 380

Gln Asn Leu Ile Lys Gln Asn Cys Glu Leu Phe Glu Gln Leu Gly Glu
385 390 395 400

Tyr Lys Phe Gln Asn Ala Leu Leu Val Arg Tyr Thr Lys Lys Val Pro
405 410 415

Gln Val Ser Thr Pro Thr Leu Val Glu Val Ser Arg Asn Leu Gly Lys
420 425 430

Val Gly Ser Lys Cys Cys Lys His Pro Glu Ala Lys Arg Met Pro Cys

- 435 - - - 440 - 445 -
 Ala Glu Asp Tyr Leu Ser Val Val Leu Asn Gln Leu Cys Val Leu His
 450 455 460
 Glu Lys Thr Pro Val Ser Asp Arg Val Thr Lys Cys Cys Thr Glu Ser
 465 470 475 480
 Leu Val Asn Arg Arg Pro Cys Phe Ser Ala Leu Glu Val Asp Glu Thr
 485 490 495
 Tyr Val Pro Lys Glu Phe Asn Ala Glu Thr Phe Thr Phe His Ala Asp
 500 505 510
 Ile Cys Thr Leu Ser Glu Lys Glu Arg Gln Ile Lys Lys Gln Thr Ala
 515 520 525
 Leu Val Glu Leu Val Lys His Lys Pro Lys Ala Thr Lys Glu Gln Leu
 530 535 540
 Lys Ala Val Met Asp Asp Phe Ala Ala Phe Val Glu Lys Cys Cys Lys
 545 550 555 560
 Ala Asp Asp Lys Glu Thr Cys Phe Ala Glu Glu Gly Lys Lys Leu Val
 565 570 575
 Ala Ala Ser Gln Ala Ala Leu Gly Leu
 580 585

<210> 17

<211> 30

<212> DNA

<213> Artificial

<220>

<223> Primer

<400> 17

ttcgaattcc cgcagccctc atttcaggg 30

<210> 18

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<212> DNA

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<400> 18

tccgaattcc ggcagcagca ggcacccatg c 31

<210> 19

<211> 25

<212> DNA

<213> Artificial

<220>

<223> Primer

<400> 19

gcggcggccg cgagcccat ttccc 25

<210> 20

<211> 18

<212> DNA

<213> Artificial

<220>

<223> Primer

<400> 20

gagagggagt actcacc 18

<210> 21

<211> 27

<212> DNA

<213> Artificial

<220>

<223> Primer

<400> 21

ggaagtgcag caagtcgagc gggggat 27

<210> 22

<211> 27

<212> DNA

<213> Artificial

<220>

<223> Primer

<400> 22

atccccgct cgacttgctg cacttc 27

<210> 23

<211> 585

<212> PRT

<213> Homo sapiens

<400> 23

Asp Ala His Lys Ser Glu Val Ala His Arg Phe Lys Asp Leu Gly Glu

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Glu Asn Phe Lys Ala Leu Val Leu Ile Ala Phe Ala Gln Tyr Leu Gln
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Gln Cys Pro Phe Glu Asp His Val Lys Leu Val Asn Glu Val Thr Glu
35      40      45
Phe Ala Lys Thr Cys Val Ala Asp Glu Ser Ala Glu Asn Cys Asp Lys
50      55      60
Ser Leu His Thr Leu Phe Gly Asp Lys Leu Cys Thr Val Ala Thr Leu
65      70      75      80
Arg Glu Thr Tyr Gly Glu Met Ala Asp Cys Cys Ala Lys Gln Glu Pro
85      90      95
Glu Arg Asn Glu Cys Phe Leu Gln His Lys Asp Asp Asn Pro Asn Leu
100     105     110
Pro Arg Leu Val Arg Pro Glu Val Asp Val Met Cys Thr Ala Phe His
115     120     125
Asp Asn Glu Glu Thr Phe Leu Lys Lys Tyr Leu Tyr Glu Ile Ala Arg
130     135     140
Arg His Pro Tyr Phe Tyr Ala Pro Glu Leu Leu Phe Phe Ala Lys Arg
145     150     155     160
Tyr Lys Ala Ala Phe Thr Glu Cys Cys Gln Ala Ala Asp Lys Ala Ala
165     170     175
Cys Leu Leu Pro Lys Leu Asp Glu Leu Arg Asp Glu Gly Lys Ala Ser
180     185     190
Ser Ala Lys Gln Arg Leu Lys Cys Ala Ser Leu Gln Lys Phe Gly Glu
195     200     205
Arg Ala Phe Lys Ala Trp Ala Val Ala Arg Leu Ser Gln Arg Phe Pro
210     215     220
Lys Ala Glu Phe Ala Glu Val Ser Lys Leu Val Thr Asp Leu Thr Lys
225     230     235     240
Val His Thr Glu Cys Cys His Gly Asp Leu Leu Glu Cys Ala Asp Asp
245     250     255
Arg Ala Asp Leu Ala Lys Tyr Ile Cys Glu Asn Gln Asp Ser Ile Ser
260     265     270
Ser Lys Leu Lys Glu Cys Cys Glu Lys Pro Leu Leu Glu Lys Ser His
275     280     285
Cys Ile Ala Glu Val Glu Asn Asp Glu Met Pro Ala Asp Leu Pro Ser
290     295     300

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Leu Ala Ala Asp Phe Val Glu Ser Lys Asp Val Cys Lys Asn Tyr Ala
 305 310 315 320
 Glu Ala Lys Asp Val Phe Leu Gly Met Phe Leu Tyr Glu Tyr Ala Arg
 325 330 335
 Arg His Pro Asp Tyr Ser Val Val Leu Leu Leu Arg Leu Ala Lys Thr
 340 345 350
 Tyr Glu Thr Thr Leu Glu Lys Cys Cys Ala Ala Ala Asp Pro His Glu
 355 360 365
 Cys Tyr Ala Lys Val Phe Asp Glu Phe Lys Pro Leu Val Glu Glu Pro
 370 375 380
 Gln Asn Leu Ile Lys Gln Asn Cys Glu Leu Phe Glu Gln Leu Gly Glu
 385 390 395 400
 Tyr Lys Phe Gln Asn Ala Leu Leu Val Arg Tyr Thr Lys Lys Val Pro
 405 410 415
 Gln Val Ser Thr Pro Thr Leu Val Glu Val Ser Arg Asn Leu Gly Lys
 420 425 430
 Val Gly Ser Lys Cys Cys Lys His Pro Glu Ala Lys Arg Met Pro Cys
 435 440 445
 Ala Glu Asp Tyr Leu Ser Val Val Leu Asn Gln Leu Cys Val Leu His
 450 455 460
 Glu Lys Thr Pro Val Ser Asp Arg Val Thr Lys Cys Cys Thr Glu Ser
 465 470 475 480
 Leu Val Asn Arg Arg Pro Cys Phe Ser Ala Leu Glu Val Asp Glu Thr
 485 490 495
 Tyr Val Pro Lys Glu Phe Asn Ala Glu Thr Phe Thr Phe His Ala Asp
 500 505 510
 Ile Cys Thr Leu Ser Glu Lys Glu Arg Gln Ile Lys Lys Gln Thr Ala
 515 520 525
 Leu Val Glu Leu Val Lys His Lys Pro Lys Ala Thr Lys Glu Gln Leu
 530 535 540
 Lys Ala Val Met Asp Asp Phe Ala Ala Phe Val Glu Lys Cys Cys Lys
 545 550 555 560
 Ala Asp Asp Lys Glu Thr Cys Phe Ala Glu Glu Gly Lys Lys Leu Val
 565 570 575
 Ala Ala Ser Gln Ala Ala Leu Gly Leu
 580 585

<210> 24

<211> 2813

<212> PRT

<213> homo sapiens

<400> 24

Met Ile Pro Ala Arg Phe Ala Gly Val Leu Leu Ala Leu Ala Leu Ile
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Leu Pro Gly Thr Leu Cys Ala Glu Gly Thr Arg Gly Arg Ser Ser Thr
 20 25 30

Ala Arg Cys Ser Leu Phe Gly Ser Asp Phe Val Asn Thr Phe Asp Gly
 35 40 45

Ser Met Tyr Ser Phe Ala Gly Tyr Cys Ser Tyr Leu Leu Ala Gly Gly
 50 55 60

Cys Gln Lys Arg Ser Phe Ser Ile Ile Gly Asp Phe Gln Asn Gly Lys
 65 70 75 80

Arg Val Ser Leu Ser Val Tyr Leu Gly Glu Phe Phe Asp Ile His Leu
 85 90 95

Phe Val Asn Gly Thr Val Thr Gln Gly Asp Gln Arg Val Ser Met Pro
 100 105 110

Tyr Ala Ser Lys Gly Leu Tyr Leu Glu Thr Glu Ala Gly Tyr Tyr Lys
 115 120 125

Leu Ser Gly Glu Ala Tyr Gly Phe Val Ala Arg Ile Asp Gly Ser Gly
 130 135 140

Asn Phe Gln Val Leu Leu Ser Asp Arg Tyr Phe Asn Lys Thr Cys Gly
 145 150 155 160

Leu Cys Gly Asn Phe Asn Ile Phe Ala Glu Asp Asp Phe Met Thr Gln
 165 170 175

Glu Gly Thr Leu Thr Ser Asp Pro Tyr Asp Phe Ala Asn Ser Trp Ala
 180 185 190

Leu Ser Ser Gly Glu Gln Trp Cys Glu Arg Ala Ser Pro Pro Ser Ser
 195 200 205

Ser Cys Asn Ile Ser Ser Gly Glu Met Gln Lys Gly Leu Trp Glu Gln
 210 215 220

Cys Gln Leu Leu Lys Ser Thr Ser Val Phe Ala Arg Cys His Pro Leu
 225 230 235 240

Val Asp Pro Glu Pro Phe Val Ala Leu Cys Glu Lys Thr Leu Cys Glu
 245 250 255

Cys Ala Gly Gly Leu Glu Cys Ala Cys Pro Ala Leu Leu Glu Tyr Ala
 260 265 270

Arg Thr Cys Ala Gln Glu Gly Met Val Leu Tyr Gly Trp Thr Asp His

275					280					285					
Ser	Ala	Cys	Ser	Pro	Val	Cys	Pro	Ala	Gly	Met	Glu	Tyr	Arg	Gln	Cys
290						295					300				
Val	Ser	Pro	Cys	Ala	Arg	Thr	Cys	Gln	Ser	Leu	His	Ile	Asn	Glu	Met
305					310					315					320
Cys	Gln	Glu	Arg	Cys	Val	Asp	Gly	Cys	Ser	Cys	Pro	Glu	Gly	Gln	Leu
				325					330					335	
Leu	Asp	Glu	Gly	Leu	Cys	Val	Glu	Ser	Thr	Glu	Cys	Pro	Cys	Val	His
			340					345					350		
Ser	Gly	Lys	Arg	Tyr	Pro	Pro	Gly	Thr	Ser	Leu	Ser	Arg	Asp	Cys	Asn
		355					360					365			
Thr	Cys	Ile	Cys	Arg	Asn	Ser	Gln	Trp	Ile	Cys	Ser	Asn	Glu	Glu	Cys
	370					375					380				
Pro	Gly	Glu	Cys	Leu	Val	Thr	Gly	Gln	Ser	His	Phe	Lys	Ser	Phe	Asp
385					390					395					400
Asn	Arg	Tyr	Phe	Thr	Phe	Ser	Gly	Ile	Cys	Gln	Tyr	Leu	Leu	Ala	Arg
			405						410					415	
Asp	Cys	Gln	Asp	His	Ser	Phe	Ser	Ile	Val	Ile	Glu	Thr	Val	Gln	Cys
			420					425					430		
Ala	Asp	Asp	Arg	Asp	Ala	Val	Cys	Thr	Arg	Ser	Val	Thr	Val	Arg	Leu
		435					440					445			
Pro	Gly	Leu	His	Asn	Ser	Leu	Val	Lys	Leu	Lys	His	Gly	Ala	Gly	Val
	450					455					460				
Ala	Met	Asp	Gly	Gln	Asp	Ile	Gln	Leu	Pro	Leu	Leu	Lys	Gly	Asp	Leu
465					470					475					480
Arg	Ile	Gln	His	Thr	Val	Thr	Ala	Ser	Val	Arg	Leu	Ser	Tyr	Gly	Glu
			485						490					495	
Asp	Leu	Gln	Met	Asp	Trp	Asp	Gly	Arg	Gly	Arg	Leu	Leu	Val	Lys	Leu
			500					505					510		
Ser	Pro	Val	Tyr	Ala	Gly	Lys	Thr	Cys	Gly	Leu	Cys	Gly	Asn	Tyr	Asn
		515					520					525			
Gly	Asn	Gln	Gly	Asp	Asp	Phe	Leu	Thr	Pro	Ser	Gly	Leu	Ala	Glu	Pro
	530					535					540				
Arg	Val	Glu	Asp	Phe	Gly	Asn	Ala	Trp	Lys	Leu	His	Gly	Asp	Cys	Gln
545					550					555					560
Asp	Leu	Gln	Lys	Gln	His	Ser	Asp	Pro	Cys	Ala	Leu	Asn	Pro	Arg	Met
				565					570					575	

Thr Arg Phe Ser Glu Glu Ala Cys Ala Val Leu Thr Ser Pro Thr Phe
 580 585 590

Glu Ala Cys His Arg Ala Val Ser Pro Leu Pro Tyr Leu Arg Asn Cys
 595 600 605

Arg Tyr Asp Val Cys Ser Cys Ser Asp Gly Arg Glu Cys Leu Cys Gly
 610 615 620

Ala Leu Ala Ser Tyr Ala Ala Ala Cys Ala Gly Arg Gly Val Arg Val
 625 630 635 640

Ala Trp Arg Glu Pro Gly Arg Cys Glu Leu Asn Cys Pro Lys Gly Gln
 645 650 655

Val Tyr Leu Gln Cys Gly Thr Pro Cys Asn Leu Thr Cys Arg Ser Leu
 660 665 670

Ser Tyr Pro Asp Glu Glu Cys Asn Glu Ala Cys Leu Glu Gly Cys Phe
 675 680 685

Cys Pro Pro Gly Leu Tyr Met Asp Glu Arg Gly Asp Cys Val Pro Lys
 690 695 700

Ala Gln Cys Pro Cys Tyr Tyr Asp Gly Glu Ile Phe Gln Pro Glu Asp
 705 710 715 720

Ile Phe Ser Asp His His Thr Met Cys Tyr Cys Glu Asp Gly Phe Met
 725 730 735

His Cys Thr Met Ser Gly Val Pro Gly Ser Leu Leu Pro Asp Ala Val
 740 745 750

Leu Ser Ser Pro Leu Ser His Arg Ser Lys Arg Ser Leu Ser Cys Arg
 755 760 765

Pro Pro Met Val Lys Leu Val Cys Pro Ala Asp Asn Leu Arg Ala Glu
 770 775 780

Gly Leu Glu Cys Thr Lys Thr Cys Gln Asn Tyr Asp Leu Glu Cys Met
 785 790 795 800

Ser Met Gly Cys Val Ser Gly Cys Leu Cys Pro Pro Gly Met Val Arg
 805 810 815

His Glu Asn Arg Cys Val Ala Leu Glu Arg Cys Pro Cys Phe His Gln
 820 825 830

Gly Lys Glu Tyr Ala Pro Gly Glu Thr Val Lys Ile Gly Cys Asn Thr
 835 840 845

Cys Val Cys Arg Asp Arg Lys Trp Asn Cys Thr Asp His Val Cys Asp
 850 855 860

Ala Thr Cys Ser Thr Ile Gly Met Ala His Tyr Leu Thr Phe Asp Gly
 865 870 875 880

Leu Lys Tyr Leu Phe Pro Gly Glu Cys Gln Tyr Val Leu Val Gln Asp
 885 890 895
 Tyr Cys Gly Ser Asn Pro Gly Thr Phe Arg Ile Leu Val Gly Asn Lys
 900 905 910
 Gly Cys Ser His Pro Ser Val Lys Cys Lys Lys Arg Val Thr Ile Leu
 915 920 925
 Val Glu Gly Gly Glu Ile Glu Leu Phe Asp Gly Glu Val Asn Val Lys
 930 935 940
 Arg Pro Met Lys Asp Glu Thr His Phe Glu Val Val Glu Ser Gly Arg
 945 950 955 960
 Tyr Ile Ile Leu Leu Leu Gly Lys Ala Leu Ser Val Val Trp Asp Arg
 965 970 975
 His Leu Ser Ile Ser Val Val Leu Lys Gln Thr Tyr Gln Glu Lys Val
 980 985 990
 Cys Gly Leu Cys Gly Asn Phe Asp Gly Ile Gln Asn Asn Asp Leu Thr
 995 1000 1005
 Ser Ser Asn Leu Gln Val Glu Glu Asp Pro Val Asp Phe Gly Asn
 1010 1015 1020
 Ser Trp Lys Val Ser Ser Gln Cys Ala Asp Thr Arg Lys Val Pro
 1025 1030 1035
 Leu Asp Ser Ser Pro Ala Thr Cys His Asn Asn Ile Met Lys Gln
 1040 1045 1050
 Thr Met Val Asp Ser Ser Cys Arg Ile Leu Thr Ser Asp Val Phe
 1055 1060 1065
 Gln Asp Cys Asn Lys Leu Val Asp Pro Glu Pro Tyr Leu Asp Val
 1070 1075 1080
 Cys Ile Tyr Asp Thr Cys Ser Cys Glu Ser Ile Gly Asp Cys Ala
 1085 1090 1095
 Cys Phe Cys Asp Thr Ile Ala Ala Tyr Ala His Val Cys Ala Gln
 1100 1105 1110
 His Gly Lys Val Val Thr Trp Arg Thr Ala Thr Leu Cys Pro Gln
 1115 1120 1125
 Ser Cys Glu Glu Arg Asn Leu Arg Glu Asn Gly Tyr Glu Cys Glu
 1130 1135 1140
 Trp Arg Tyr Asn Ser Cys Ala Pro Ala Cys Gln Val Thr Cys Gln
 1145 1150 1155
 His Pro Glu Pro Leu Ala Cys Pro Val Gln Cys Val Glu Gly Cys
 1160 1165 1170

His Ala	His Cys Pro Pro Gly	Lys Ile Leu Asp Glu	Leu Leu Gln
1175	1180	1185	
Thr Cys	Val Asp Pro Glu Asp	Cys Pro Val Cys Glu	Val Ala Gly
1190	1195	1200	
Arg Arg	Phe Ala Ser Gly Lys	Lys Val Thr Leu Asn	Pro Ser Asp
1205	1210	1215	
Pro Glu	His Cys Gln Ile Cys	His Cys Asp Val Val	Asn Leu Thr
1220	1225	1230	
Cys Glu	Ala Cys Gln Glu Pro	Gly Gly Leu Val Val	Pro Pro Thr
1235	1240	1245	
Asp Ala	Pro Val Ser Pro Thr	Thr Leu Tyr Val Glu	Asp Ile Ser
1250	1255	1260	
Glu Pro	Pro Leu His Asp Phe	Tyr Cys Ser Arg Leu	Leu Asp Leu
1265	1270	1275	
Val Phe	Leu Leu Asp Gly Ser	Ser Arg Leu Ser Glu	Ala Glu Phe
1280	1285	1290	
Glu Val	Leu Lys Ala Phe Val	Val Asp Met Met Glu	Arg Leu Arg
1295	1300	1305	
Ile Ser	Gln Lys Trp Val Arg	Val Ala Val Val Glu	Tyr His Asp
1310	1315	1320	
Gly Ser	His Ala Tyr Ile Gly	Leu Lys Asp Arg Lys	Arg Pro Ser
1325	1330	1335	
Glu Leu	Arg Arg Ile Ala Ser	Gln Val Lys Tyr Ala	Gly Ser Gln
1340	1345	1350	
Val Ala	Ser Thr Ser Glu Val	Leu Lys Tyr Thr Leu	Phe Gln Ile
1355	1360	1365	
Phe Ser	Lys Ile Asp Arg Pro	Glu Ala Ser Arg Ile	Thr Leu Leu
1370	1375	1380	
Leu Met	Ala Ser Gln Glu Pro	Gln Arg Met Ser Arg	Asn Phe Val
1385	1390	1395	
Arg Tyr	Val Gln Gly Leu Lys	Lys Lys Lys Val Ile	Val Ile Pro
1400	1405	1410	
Val Gly	Ile Gly Pro His Ala	Asn Leu Lys Gln Ile	Arg Leu Ile
1415	1420	1425	
Glu Lys	Gln Ala Pro Glu Asn	Lys Ala Phe Val Leu	Ser Ser Val
1430	1435	1440	

Asp Glu	Leu Glu	Gln Gln	Arg	Asp Glu	Ile Val	Ser	Tyr Leu	Cys
1445			1450			1455		
Asp Leu	Ala Pro	Glu Ala	Pro	Pro Pro	Thr Leu	Pro	Pro Asp	Met
1460			1465			1470		
Ala Gln	Val Thr	Val Gly	Pro	Gly Leu	Leu Gly	Val	Ser Thr	Leu
1475			1480			1485		
Gly Pro	Lys Arg	Asn Ser	Met	Val Leu	Asp Val	Ala	Phe Val	Leu
1490			1495			1500		
Glu Gly	Ser Asp	Lys Ile	Gly	Glu Ala	Asp Phe	Asn	Arg Ser	Lys
1505			1510			1515		
Glu Phe	Met Glu	Glu Val	Ile	Gln Arg	Met Asp	Val	Gly Gln	Asp
1520			1525			1530		
Ser Ile	His Val	Thr Val	Leu	Gln Tyr	Ser Tyr	Met	Val Thr	Val
1535			1540			1545		
Glu Tyr	Pro Phe	Ser Glu	Ala	Gln Ser	Lys Gly	Asp	Ile Leu	Gln
1550			1555			1560		
Arg Val	Arg Glu	Ile Arg	Tyr	Gln Gly	Gly Asn	Arg	Thr Asn	Thr
1565			1570			1575		
Gly Leu	Ala Leu	Arg Tyr	Leu	Ser Asp	His Ser	Phe	Leu Val	Ser
1580			1585			1590		
Gln Gly	Asp Arg	Glu Gln	Ala	Pro Asn	Leu Val	Tyr	Met Val	Thr
1595			1600			1605		
Gly Asn	Pro Ala	Ser Asp	Glu	Ile Lys	Arg Leu	Pro	Gly Asp	Ile
1610			1615			1620		
Gln Val	Val Pro	Ile Gly	Val	Gly Pro	Asn Ala	Asn	Val Gln	Glu
1625			1630			1635		
Leu Glu	Arg Ile	Gly Trp	Pro	Asn Ala	Pro Ile	Leu	Ile Gln	Asp
1640			1645			1650		
Phe Glu	Thr Leu	Pro Arg	Glu	Ala Pro	Asp Leu	Val	Leu Gln	Arg
1655			1660			1665		
Cys Cys	Ser Gly	Glu Gly	Leu	Gln Ile	Pro Thr	Leu	Ser Pro	Ala
1670			1675			1680		
Pro Asp	Cys Ser	Gln Pro	Leu	Asp Val	Ile Leu	Leu	Leu Asp	Gly
1685			1690			1695		
Ser Ser	Ser Phe	Pro Ala	Ser	Tyr Phe	Asp Glu	Met	Lys Ser	Phe
1700			1705			1710		
Ala Lys	Ala Phe	Ile Ser	Lys	Ala Asn	Ile Gly	Pro	Arg Leu	Thr
1715			1720			1725		

Gln Val 1730	Ser Val	Leu Gln	Tyr 1735	Gly Ser	Ile Thr	Thr 1740	Ile Asp	Val
Pro Trp 1745	Asn Val	Val Pro	Glu 1750	Lys Ala	His Leu	Leu 1755	Ser Leu	Val
Asp Val 1760	Met Gln	Arg Glu	Gly 1765	Gly Pro	Ser Gln	Ile 1770	Gly Asp	Ala
Leu Gly 1775	Phe Ala	Val Arg	Tyr 1780	Leu Thr	Ser Glu	Met 1785	His Gly	Ala
Arg Pro 1790	Gly Ala	Ser Lys	Ala 1795	Val Val	Ile Leu	Val 1800	Thr Asp	Val
Ser Val 1805	Asp Ser	Val Asp	Ala 1810	Ala Ala	Asp Ala	Ala 1815	Arg Ser	Asn
Arg Val 1820	Thr Val	Phe Pro	Ile 1825	Gly Ile	Gly Asp	Arg 1830	Tyr Asp	Ala
Ala Gln 1835	Leu Arg	Ile Leu	Ala 1840	Gly Pro	Ala Gly	Asp 1845	Ser Asn	Val
Val Lys 1850	Leu Gln	Arg Ile	Glu 1855	Asp Leu	Pro Thr	Met 1860	Val Thr	Leu
Gly Asn 1865	Ser Phe	Leu His	Lys 1870	Leu Cys	Ser Gly	Phe 1875	Val Arg	Ile
Cys Met 1880	Asp Glu	Asp Gly	Asn 1885	Glu Lys	Arg Pro	Gly 1890	Asp Val	Trp
Thr Leu 1895	Pro Asp	Gln Cys	His 1900	Thr Val	Thr Cys	Gln 1905	Pro Asp	Gly
Gln Thr 1910	Leu Leu	Lys Ser	His 1915	Arg Val	Asn Cys	Asp 1920	Arg Gly	Leu
Arg Pro 1925	Ser Cys	Pro Asn	Ser 1930	Gln Ser	Pro Val	Lys 1935	Val Glu	Glu
Thr Cys 1940	Gly Cys	Arg Trp	Thr 1945	Cys Pro	Cys Val	Cys 1950	Thr Gly	Ser
Ser Thr 1955	Arg His	Ile Val	Thr 1960	Phe Asp	Gly Gln	Asn 1965	Phe Lys	Leu
Thr Gly 1970	Ser Cys	Ser Tyr	Val 1975	Leu Phe	Gln Asn	Lys 1980	Glu Gln	Asp
Leu Glu 1985	Val Ile	Leu His	Asn 1990	Gly Ala	Cys Ser	Pro 1995	Gly Ala	Arg
Gln Gly 2000	Cys Met	Lys Ser	Ile 2005	Glu Val	Lys His	Ser 2010	Ala Leu	Ser
Val Glu	Leu His	Ser Asp	Met	Glu Val	Thr Val	Asn	Gly Arg	Leu

2015	2020	2025
Val Ser 2030	Val Pro Tyr Val Gly 2035	Gly Asn Met Glu Val Asn Val Tyr 2040
Gly Ala 2045	Ile Met His Glu Val 2050	Arg Phe Asn His Leu Gly His Ile 2055
Phe Thr 2060	Phe Thr Pro Gln Asn 2065	Asn Glu Phe Gln Leu Gln Leu Ser 2070
Pro Lys 2075	Thr Phe Ala Ser Lys 2080	Thr Tyr Gly Leu Cys Gly Ile Cys 2085
Asp Glu 2090	Asn Gly Ala Asn Asp 2095	Phe Met Leu Arg Asp Gly Thr Val 2100
Thr Thr 2105	Asp Trp Lys Thr Leu 2110	Val Gln Glu Trp Thr Val Gln Arg 2115
Pro Gly 2120	Gln Thr Cys Gln Pro 2125	Ile Leu Glu Glu Gln Cys Leu Val 2130
Pro Asp 2135	Ser Ser His Cys Gln 2140	Val Leu Leu Leu Pro Leu Phe Ala 2145
Glu Cys 2150	His Lys Val Leu Ala 2155	Pro Ala Thr Phe Tyr Ala Ile Cys 2160
Gln Gln 2165	Asp Ser Cys His Gln 2170	Glu Gln Val Cys Glu Val Ile Ala 2175
Ser Tyr 2180	Ala His Leu Cys Arg 2185	Thr Asn Gly Val Cys Val Asp Trp 2190
Arg Thr 2195	Pro Asp Phe Cys Ala 2200	Met Ser Cys Pro Pro Ser Leu Val 2205
Tyr Asn 2210	His Cys Glu His Gly 2215	Cys Pro Arg His Cys Asp Gly Asn 2220
Val Ser 2225	Ser Cys Gly Asp His 2230	Pro Ser Glu Gly Cys Phe Cys Pro 2235
Pro Asp 2240	Lys Val Met Leu Glu 2245	Gly Ser Cys Val Pro Glu Glu Ala 2250
Cys Thr 2255	Gln Cys Ile Gly Glu 2260	Asp Gly Val Gln His Gln Phe Leu 2265
Glu Ala 2270	Trp Val Pro Asp His 2275	Gln Pro Cys Gln Ile Cys Thr Cys 2280
Leu Ser 2285	Gly Arg Lys Val Asn 2290	Cys Thr Thr Gln Pro Cys Pro Thr 2295

Ala Lys 2300	Ala Pro Thr Cys Gly 2305	Leu Cys Glu Val 2310	Ala Arg Leu Arg
Gln Asn 2315	Ala Asp Gln Cys Cys 2320	Pro Glu Tyr Glu Cys 2325	Val Cys Asp
Pro Val 2330	Ser Cys Asp Leu Pro 2335	Pro Val Pro His Cys 2340	Glu Arg Gly
Leu Gln 2345	Pro Thr Leu Thr Asn 2350	Pro Gly Glu Cys Arg 2355	Pro Asn Phe
Thr Cys 2360	Ala Cys Arg Lys Glu 2365	Glu Cys Lys Arg Val 2370	Ser Pro Pro
Ser Cys 2375	Pro Pro His Arg Leu 2380	Pro Thr Leu Arg Lys 2385	Thr Gln Cys
Cys Asp 2390	Glu Tyr Glu Cys Ala 2395	Cys Asn Cys Val Asn 2400	Ser Thr Val
Ser Cys 2405	Pro Leu Gly Tyr Leu 2410	Ala Ser Thr Ala Thr 2415	Asn Asp Cys
Gly Cys 2420	Thr Thr Thr Thr Cys 2425	Leu Pro Asp Lys Val 2430	Cys Val His
Arg Ser 2435	Thr Ile Tyr Pro Val 2440	Gly Gln Phe Trp Glu 2445	Glu Gly Cys
Asp Val 2450	Cys Thr Cys Thr Asp 2455	Met Glu Asp Ala Val 2460	Met Gly Leu
Arg Val 2465	Ala Gln Cys Ser Gln 2470	Lys Pro Cys Glu Asp 2475	Ser Cys Arg
Ser Gly 2480	Phe Thr Tyr Val Leu 2485	His Glu Gly Glu Cys 2490	Cys Gly Arg
Cys Leu 2495	Pro Ser Ala Cys Glu 2500	Val Val Thr Gly Ser 2505	Pro Arg Gly
Asp Ser 2510	Gln Ser Ser Trp Lys 2515	Ser Val Gly Ser Gln 2520	Trp Ala Ser
Pro Glu 2525	Asn Pro Cys Leu Ile 2530	Asn Glu Cys Val Arg 2535	Val Lys Glu
Glu Val 2540	Phe Ile Gln Gln Arg 2545	Asn Val Ser Cys Pro 2550	Gln Leu Glu
Val Pro 2555	Val Cys Pro Ser Gly 2560	Phe Gln Leu Ser Cys 2565	Lys Thr Ser
Ala Cys 2570	Cys Pro Ser Cys Arg 2575	Cys Glu Arg Met Glu 2580	Ala Cys Met

Leu Asn Gly Thr Val Ile Gly Pro Gly Lys Thr Val Met Ile Asp
 2585 2590 2595
 Val Cys Thr Thr Cys Arg Cys Met Val Gln Val Gly Val Ile Ser
 2600 2605 2610
 Gly Phe Lys Leu Glu Cys Arg Lys Thr Thr Cys Asn Pro Cys Pro
 2615 2620 2625
 Leu Gly Tyr Lys Glu Glu Asn Asn Thr Gly Glu Cys Cys Gly Arg
 2630 2635 2640
 Cys Leu Pro Thr Ala Cys Thr Ile Gln Leu Arg Gly Gly Gln Ile
 2645 2650 2655
 Met Thr Leu Lys Arg Asp Glu Thr Leu Gln Asp Gly Cys Asp Thr
 2660 2665 2670
 His Phe Cys Lys Val Asn Glu Arg Gly Glu Tyr Phe Trp Glu Lys
 2675 2680 2685
 Arg Val Thr Gly Cys Pro Pro Phe Asp Glu His Lys Cys Leu Ala
 2690 2695 2700
 Glu Gly Gly Lys Ile Met Lys Ile Pro Gly Thr Cys Cys Asp Thr
 2705 2710 2715
 Cys Glu Glu Pro Glu Cys Asn Asp Ile Thr Ala Arg Leu Gln Tyr
 2720 2725 2730
 Val Lys Val Gly Ser Cys Lys Ser Glu Val Glu Val Asp Ile His
 2735 2740 2745
 Tyr Cys Gln Gly Lys Cys Ala Ser Lys Ala Met Tyr Ser Ile Asp
 2750 2755 2760
 Ile Asn Asp Val Gln Asp Gln Cys Ser Cys Cys Ser Pro Thr Arg
 2765 2770 2775
 Thr Glu Pro Met Gln Val Ala Leu His Cys Thr Asn Gly Ser Val
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 Val Tyr His Glu Val Leu Asn Ala Met Glu Cys Lys Cys Ser Pro
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 Arg Lys Cys Ser Lys
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<213> Artificial

<220>

<223> Amino acid sequence of human VWF albumin fusion preproprotein

<400> 25

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Leu Pro Gly Thr Leu Cys Ala Glu Gly Thr Arg Gly Arg Ser Ser Thr
20 25 30

Ala Arg Cys Ser Leu Phe Gly Ser Asp Phe Val Asn Thr Phe Asp Gly
35 40 45

Ser Met Tyr Ser Phe Ala Gly Tyr Cys Ser Tyr Leu Leu Ala Gly Gly
50 55 60

Cys Gln Lys Arg Ser Phe Ser Ile Ile Gly Asp Phe Gln Asn Gly Lys
65 70 75 80

Arg Val Ser Leu Ser Val Tyr Leu Gly Glu Phe Phe Asp Ile His Leu
85 90 95

Phe Val Asn Gly Thr Val Thr Gln Gly Asp Gln Arg Val Ser Met Pro
100 105 110

Tyr Ala Ser Lys Gly Leu Tyr Leu Glu Thr Glu Ala Gly Tyr Tyr Lys
115 120 125

Leu Ser Gly Glu Ala Tyr Gly Phe Val Ala Arg Ile Asp Gly Ser Gly
130 135 140

Asn Phe Gln Val Leu Leu Ser Asp Arg Tyr Phe Asn Lys Thr Cys Gly
145 150 155 160

Leu Cys Gly Asn Phe Asn Ile Phe Ala Glu Asp Asp Phe Met Thr Gln
165 170 175

Glu Gly Thr Leu Thr Ser Asp Pro Tyr Asp Phe Ala Asn Ser Trp Ala
180 185 190

Leu Ser Ser Gly Glu Gln Trp Cys Glu Arg Ala Ser Pro Pro Ser Ser
195 200 205

Ser Cys Asn Ile Ser Ser Gly Glu Met Gln Lys Gly Leu Trp Glu Gln
210 215 220

Cys Gln Leu Leu Lys Ser Thr Ser Val Phe Ala Arg Cys His Pro Leu
225 230 235 240

Val Asp Pro Glu Pro Phe Val Ala Leu Cys Glu Lys Thr Leu Cys Glu
245 250 255

Cys Ala Gly Gly Leu Glu Cys Ala Cys Pro Ala Leu Leu Glu Tyr Ala
260 265 270

Arg Thr Cys Ala Gln Glu Gly Met Val Leu Tyr Gly Trp Thr Asp His
275 280 285

Ser Ala Cys Ser Pro Val Cys Pro Ala Gly Met Glu Tyr Arg Gln Cys
 290 295 300

Val Ser Pro Cys Ala Arg Thr Cys Gln Ser Leu His Ile Asn Glu Met
 305 310 315 320

Cys Gln Glu Arg Cys Val Asp Gly Cys Ser Cys Pro Glu Gly Gln Leu
 325 330 335

Leu Asp Glu Gly Leu Cys Val Glu Ser Thr Glu Cys Pro Cys Val His
 340 345 350

Ser Gly Lys Arg Tyr Pro Pro Gly Thr Ser Leu Ser Arg Asp Cys Asn
 355 360 365

Thr Cys Ile Cys Arg Asn Ser Gln Trp Ile Cys Ser Asn Glu Glu Cys
 370 375 380

Pro Gly Glu Cys Leu Val Thr Gly Gln Ser His Phe Lys Ser Phe Asp
 385 390 395 400

Asn Arg Tyr Phe Thr Phe Ser Gly Ile Cys Gln Tyr Leu Leu Ala Arg
 405 410 415

Asp Cys Gln Asp His Ser Phe Ser Ile Val Ile Glu Thr Val Gln Cys
 420 425 430

Ala Asp Asp Arg Asp Ala Val Cys Thr Arg Ser Val Thr Val Arg Leu
 435 440 445

Pro Gly Leu His Asn Ser Leu Val Lys Leu Lys His Gly Ala Gly Val
 450 455 460

Ala Met Asp Gly Gln Asp Ile Gln Leu Pro Leu Leu Lys Gly Asp Leu
 465 470 475 480

Arg Ile Gln His Thr Val Thr Ala Ser Val Arg Leu Ser Tyr Gly Glu
 485 490 495

Asp Leu Gln Met Asp Trp Asp Gly Arg Gly Arg Leu Leu Val Lys Leu
 500 505 510

Ser Pro Val Tyr Ala Gly Lys Thr Cys Gly Leu Cys Gly Asn Tyr Asn
 515 520 525

Gly Asn Gln Gly Asp Asp Phe Leu Thr Pro Ser Gly Leu Ala Glu Pro
 530 535 540

Arg Val Glu Asp Phe Gly Asn Ala Trp Lys Leu His Gly Asp Cys Gln
 545 550 555 560

Asp Leu Gln Lys Gln His Ser Asp Pro Cys Ala Leu Asn Pro Arg Met
 565 570 575

Thr Arg Phe Ser Glu Glu Ala Cys Ala Val Leu Thr Ser Pro Thr Phe
 580 585 590

Glu Ala Cys His Arg Ala Val Ser Pro Leu Pro Tyr Leu Arg Asn Cys
595 600 605

Arg Tyr Asp Val Cys Ser Cys Ser Asp Gly Arg Glu Cys Leu Cys Gly
610 615 620

Ala Leu Ala Ser Tyr Ala Ala Ala Cys Ala Gly Arg Gly Val Arg Val
625 630 635 640

Ala Trp Arg Glu Pro Gly Arg Cys Glu Leu Asn Cys Pro Lys Gly Gln
645 650 655

Val Tyr Leu Gln Cys Gly Thr Pro Cys Asn Leu Thr Cys Arg Ser Leu
660 665 670

Ser Tyr Pro Asp Glu Glu Cys Asn Glu Ala Cys Leu Glu Gly Cys Phe
675 680 685

Cys Pro Pro Gly Leu Tyr Met Asp Glu Arg Gly Asp Cys Val Pro Lys
690 695 700

Ala Gln Cys Pro Cys Tyr Tyr Asp Gly Glu Ile Phe Gln Pro Glu Asp
705 710 715 720

Ile Phe Ser Asp His His Thr Met Cys Tyr Cys Glu Asp Gly Phe Met
725 730 735

His Cys Thr Met Ser Gly Val Pro Gly Ser Leu Leu Pro Asp Ala Val
740 745 750

Leu Ser Ser Pro Leu Ser His Arg Ser Lys Arg Ser Leu Ser Cys Arg
755 760 765

Pro Pro Met Val Lys Leu Val Cys Pro Ala Asp Asn Leu Arg Ala Glu
770 775 780

Gly Leu Glu Cys Thr Lys Thr Cys Gln Asn Tyr Asp Leu Glu Cys Met
785 790 795 800

Ser Met Gly Cys Val Ser Gly Cys Leu Cys Pro Pro Gly Met Val Arg
805 810 815

His Glu Asn Arg Cys Val Ala Leu Glu Arg Cys Pro Cys Phe His Gln
820 825 830

Gly Lys Glu Tyr Ala Pro Gly Glu Thr Val Lys Ile Gly Cys Asn Thr
835 840 845

Cys Val Cys Arg Asp Arg Lys Trp Asn Cys Thr Asp His Val Cys Asp
850 855 860

Ala Thr Cys Ser Thr Ile Gly Met Ala His Tyr Leu Thr Phe Asp Gly
865 870 875 880

Leu Lys Tyr Leu Phe Pro Gly Glu Cys Gln Tyr Val Leu Val Gln Asp
885 890 895 900 905

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000          030          070

Tyr Cys Gly Ser Asn Pro Gly Thr Phe Arg Ile Leu Val Gly Asn Lys
    900                      905                      910

Gly Cys Ser His Pro Ser Val Lys Cys Lys Lys Arg Val Thr Ile Leu
    915                      920                      925

Val Glu Gly Gly Glu Ile Glu Leu Phe Asp Gly Glu Val Asn Val Lys
    930                      935                      940

Arg Pro Met Lys Asp Glu Thr His Phe Glu Val Val Glu Ser Gly Arg
    945                      950                      955                      960

Tyr Ile Ile Leu Leu Leu Gly Lys Ala Leu Ser Val Val Trp Asp Arg
    965                      970                      975

His Leu Ser Ile Ser Val Val Leu Lys Gln Thr Tyr Gln Glu Lys Val
    980                      985                      990

Cys Gly Leu Cys Gly Asn Phe Asp Gly Ile Gln Asn Asn Asp Leu Thr
    995                      1000                      1005

Ser Ser Asn Leu Gln Val Glu Glu Asp Pro Val Asp Phe Gly Asn
    1010                      1015                      1020

Ser Trp Lys Val Ser Ser Gln Cys Ala Asp Thr Arg Lys Val Pro
    1025                      1030                      1035

Leu Asp Ser Ser Pro Ala Thr Cys His Asn Asn Ile Met Lys Gln
    1040                      1045                      1050

Thr Met Val Asp Ser Ser Cys Arg Ile Leu Thr Ser Asp Val Phe
    1055                      1060                      1065

Gln Asp Cys Asn Lys Leu Val Asp Pro Glu Pro Tyr Leu Asp Val
    1070                      1075                      1080

Cys Ile Tyr Asp Thr Cys Ser Cys Glu Ser Ile Gly Asp Cys Ala
    1085                      1090                      1095

Cys Phe Cys Asp Thr Ile Ala Ala Tyr Ala His Val Cys Ala Gln
    1100                      1105                      1110

His Gly Lys Val Val Thr Trp Arg Thr Ala Thr Leu Cys Pro Gln
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Ser Cys Glu Glu Arg Asn Leu Arg Glu Asn Gly Tyr Glu Cys Glu
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Trp Arg Tyr Asn Ser Cys Ala Pro Ala Cys Gln Val Thr Cys Gln
    1145                      1150                      1155

His Pro Glu Pro Leu Ala Cys Pro Val Gln Cys Val Glu Gly Cys
    1160                      1165                      1170

His Ala His Cys Pro Pro Gly Lys Ile Leu Asp Glu Leu Leu Gln

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Thr Cys Val Asp Pro Glu Asp Cys Pro Val Cys Glu Val Ala Gly		
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Arg Arg Phe Ala Ser Gly Lys Lys Val Thr Leu Asn Pro Ser Asp		
1205	1210	1215
Pro Glu His Cys Gln Ile Cys His Cys Asp Val Val Asn Leu Thr		
1220	1225	1230
Cys Glu Ala Cys Gln Glu Pro Gly Gly Leu Val Val Pro Pro Thr		
1235	1240	1245
Asp Ala Pro Val Ser Pro Thr Thr Leu Tyr Val Glu Asp Ile Ser		
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Glu Pro Pro Leu His Asp Phe Tyr Cys Ser Arg Leu Leu Asp Leu		
1265	1270	1275
Val Phe Leu Leu Asp Gly Ser Ser Arg Leu Ser Glu Ala Glu Phe		
1280	1285	1290
Glu Val Leu Lys Ala Phe Val Val Asp Met Met Glu Arg Leu Arg		
1295	1300	1305
Ile Ser Gln Lys Trp Val Arg Val Ala Val Val Glu Tyr His Asp		
1310	1315	1320
Gly Ser His Ala Tyr Ile Gly Leu Lys Asp Arg Lys Arg Pro Ser		
1325	1330	1335
Glu Leu Arg Arg Ile Ala Ser Gln Val Lys Tyr Ala Gly Ser Gln		
1340	1345	1350
Val Ala Ser Thr Ser Glu Val Leu Lys Tyr Thr Leu Phe Gln Ile		
1355	1360	1365
Phe Ser Lys Ile Asp Arg Pro Glu Ala Ser Arg Ile Thr Leu Leu		
1370	1375	1380
Leu Met Ala Ser Gln Glu Pro Gln Arg Met Ser Arg Asn Phe Val		
1385	1390	1395
Arg Tyr Val Gln Gly Leu Lys Lys Lys Lys Val Ile Val Ile Pro		
1400	1405	1410
Val Gly Ile Gly Pro His Ala Asn Leu Lys Gln Ile Arg Leu Ile		
1415	1420	1425
Glu Lys Gln Ala Pro Glu Asn Lys Ala Phe Val Leu Ser Ser Val		
1430	1435	1440
Asp Glu Leu Glu Gln Gln Arg Asp Glu Ile Val Ser Tyr Leu Cys		
1445	1450	1455
Asp Leu Ala Pro Glu Ala Pro Pro Pro Thr Leu Pro Pro Asp Met		
1460	1465	1470

1460	1465	1470
Ala Gln Val Thr Val Gly 1475	Pro Gly Leu Leu Gly 1480	Val Ser Thr Leu 1485
Gly Pro Lys Arg Asn Ser 1490	Met Val Leu Asp Val 1495	Ala Phe Val Leu 1500
Glu Gly Ser Asp Lys Ile 1505	Gly Glu Ala Asp Phe 1510	Asn Arg Ser Lys 1515
Glu Phe Met Glu Glu Val 1520	Ile Gln Arg Met Asp 1525	Val Gly Gln Asp 1530
Ser Ile His Val Thr Val 1535	Leu Gln Tyr Ser Tyr 1540	Met Val Thr Val 1545
Glu Tyr Pro Phe Ser Glu 1550	Ala Gln Ser Lys Gly 1555	Asp Ile Leu Gln 1560
Arg Val Arg Glu Ile Arg 1565	Tyr Gln Gly Gly Asn 1570	Arg Thr Asn Thr 1575
Gly Leu Ala Leu Arg Tyr 1580	Leu Ser Asp His Ser 1585	Phe Leu Val Ser 1590
Gln Gly Asp Arg Glu Gln 1595	Ala Pro Asn Leu Val 1600	Tyr Met Val Thr 1605
Gly Asn Pro Ala Ser Asp 1610	Glu Ile Lys Arg Leu 1615	Pro Gly Asp Ile 1620
Gln Val Val Pro Ile Gly 1625	Val Gly Pro Asn Ala 1630	Asn Val Gln Glu 1635
Leu Glu Arg Ile Gly Trp 1640	Pro Asn Ala Pro Ile 1645	Leu Ile Gln Asp 1650
Phe Glu Thr Leu Pro Arg 1655	Glu Ala Pro Asp Leu 1660	Val Leu Gln Arg 1665
Cys Cys Ser Gly Glu Gly 1670	Leu Gln Ile Pro Thr 1675	Leu Ser Pro Ala 1680
Pro Asp Cys Ser Gln Pro 1685	Leu Asp Val Ile Leu 1690	Leu Leu Asp Gly 1695
Ser Ser Ser Phe Pro Ala 1700	Ser Tyr Phe Asp Glu 1705	Met Lys Ser Phe 1710
Ala Lys Ala Phe Ile Ser 1715	Lys Ala Asn Ile Gly 1720	Pro Arg Leu Thr 1725
Gln Val Ser Val Leu Gln 1730	Tyr Gly Ser Ile Thr 1735	Thr Ile Asp Val 1740
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Arg Val Thr Val Phe Pro Ile 1820	Gly Ile Gly Asp Arg Tyr Asp Ala 1825	
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Val Lys Leu Gln Arg Ile Glu 1850	Asp Leu Pro Thr Met Val Thr Leu 1855	
Gly Asn Ser Phe Leu His Lys 1865	Leu Cys Ser Gly Phe Val Arg Ile 1870	
Cys Met Asp Glu Asp Gly Asn 1880	Glu Lys Arg Pro Gly Asp Val Trp 1885	
Thr Leu Pro Asp Gln Cys His 1895	Thr Val Thr Cys Gln Pro Asp Gly 1900	
Gln Thr Leu Leu Lys Ser His 1910	Arg Val Asn Cys Asp Arg Gly Leu 1915	
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Gly 2045	Ala	Ile	Met	His	Glu	Val 2050	Arg	Phe	Asn	His	Leu 2055	Gly	His	Ile
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Pro 2075	Lys	Thr	Phe	Ala	Ser	Lys 2080	Thr	Tyr	Gly	Leu	Cys 2085	Gly	Ile	Cys
Asp 2090	Glu	Asn	Gly	Ala	Asn	Asp 2095	Phe	Met	Leu	Arg	Asp 2100	Gly	Thr	Val
Thr 2105	Thr	Asp	Trp	Lys	Thr	Leu 2110	Val	Gln	Glu	Trp	Thr 2115	Val	Gln	Arg
Pro 2120	Gly	Gln	Thr	Cys	Gln	Pro 2125	Ile	Leu	Glu	Glu	Gln 2130	Cys	Leu	Val
Pro 2135	Asp	Ser	Ser	His	Cys	Gln 2140	Val	Leu	Leu	Leu	Pro 2145	Leu	Phe	Ala
Glu 2150	Cys	His	Lys	Val	Leu	Ala 2155	Pro	Ala	Thr	Phe	Tyr 2160	Ala	Ile	Cys
Gln 2165	Gln	Asp	Ser	Cys	His	Gln 2170	Glu	Gln	Val	Cys	Glu 2175	Val	Ile	Ala
Ser 2180	Tyr	Ala	His	Leu	Cys	Arg 2185	Thr	Asn	Gly	Val	Cys 2190	Val	Asp	Trp
Arg 2195	Thr	Pro	Asp	Phe	Cys	Ala 2200	Met	Ser	Cys	Pro	Pro 2205	Ser	Leu	Val
Tyr 2210	Asn	His	Cys	Glu	His	Gly 2215	Cys	Pro	Arg	His	Cys 2220	Asp	Gly	Asn
Val 2225	Ser	Ser	Cys	Gly	Asp	His 2230	Pro	Ser	Glu	Gly	Cys 2235	Phe	Cys	Pro
Pro 2240	Asp	Lys	Val	Met	Leu	Glu 2245	Gly	Ser	Cys	Val	Pro 2250	Glu	Glu	Ala
Cys 2255	Thr	Gln	Cys	Ile	Gly	Glu 2260	Asp	Gly	Val	Gln	His 2265	Gln	Phe	Leu
Glu 2270	Ala	Trp	Val	Pro	Asp	His 2275	Gln	Pro	Cys	Gln	Ile 2280	Cys	Thr	Cys
Leu 2285	Ser	Gly	Arg	Lys	Val	Asn 2290	Cys	Thr	Thr	Gln	Pro 2295	Cys	Pro	Thr
Ala 2300	Lys	Ala	Pro	Thr	Cys	Gly 2305	Leu	Cys	Glu	Val	Ala 2310	Arg	Leu	Arg

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Leu Gln	Pro Thr	Leu Thr	Asn	Pro Gly	Glu Cys	Arg	Pro Asn	Phe	
2345			2350			2355			
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2360			2365			2370			
Ser Cys	Pro Pro	His Arg	Leu	Pro Thr	Leu Arg	Lys	Thr Gln	Cys	
2375			2380			2385			
Cys Asp	Glu Tyr	Glu Cys	Ala	Cys Asn	Cys Val	Asn	Ser Thr	Val	
2390			2395			2400			
Ser Cys	Pro Leu	Gly Tyr	Leu	Ala Ser	Thr Ala	Thr	Asn Asp	Cys	
2405			2410			2415			
Gly Cys	Thr Thr	Thr Thr	Cys	Leu Pro	Asp Lys	Val	Cys Val	His	
2420			2425			2430			
Arg Ser	Thr Ile	Tyr Pro	Val	Gly Gln	Phe Trp	Glu	Glu Gly	Cys	
2435			2440			2445			
Asp Val	Cys Thr	Cys Thr	Asp	Met Glu	Asp Ala	Val	Met Gly	Leu	
2450			2455			2460			
Arg Val	Ala Gln	Cys Ser	Gln	Lys Pro	Cys Glu	Asp	Ser Cys	Arg	
2465			2470			2475			
Ser Gly	Phe Thr	Tyr Val	Leu	His Glu	Gly Glu	Cys	Cys Gly	Arg	
2480			2485			2490			
Cys Leu	Pro Ser	Ala Cys	Glu	Val Val	Thr Gly	Ser	Pro Arg	Gly	
2495			2500			2505			
Asp Ser	Gln Ser	Ser Trp	Lys	Ser Val	Gly Ser	Gln	Trp Ala	Ser	
2510			2515			2520			
Pro Glu	Asn Pro	Cys Leu	Ile	Asn Glu	Cys Val	Arg	Val Lys	Glu	
2525			2530			2535			
Glu Val	Phe Ile	Gln Gln	Arg	Asn Val	Ser Cys	Pro	Gln Leu	Glu	
2540			2545			2550			
Val Pro	Val Cys	Pro Ser	Gly	Phe Gln	Leu Ser	Cys	Lys Thr	Ser	
2555			2560			2565			
Ala Cys	Cys Pro	Ser Cys	Arg	Cys Glu	Arg Met	Glu	Ala Cys	Met	
2570			2575			2580			
Leu Asn	Gly Thr	Val Ile	Gly	Pro Gly	Lys Thr	Val	Met Ile	Asp	
2585			2590			2595			

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Val Phe	Leu Gly Met	Phe Leu	Tyr Glu Tyr	Ala Arg	Arg His Pro
3170		3175		3180	
Asp Tyr	Ser Val Val	Leu Leu	Leu Arg Leu	Ala Lys	Thr Tyr Glu
3185		3190		3195	
Thr Thr	Leu Glu Lys	Cys Cys	Ala Ala Ala	Asp Pro	His Glu Cys
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Tyr Ala	Lys Val Phe	Asp Glu	Phe Lys Pro	Leu Val	Glu Glu Pro
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Gln Asn	Leu Ile Lys	Gln Asn	Cys Glu Leu	Phe Glu	Gln Leu Gly
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Glu Tyr	Lys Phe Gln	Asn Ala	Leu Leu Val	Arg Tyr	Thr Lys Lys
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Val Pro	Gln Val Ser	Thr Pro	Thr Leu Val	Glu Val	Ser Arg Asn
3260		3265		3270	
Leu Gly	Lys Val Gly	Ser Lys	Cys Cys Lys	His Pro	Glu Ala Lys
3275		3280		3285	
Arg Met	Pro Cys Ala	Glu Asp	Tyr Leu Ser	Val Val	Leu Asn Gln
3290		3295		3300	
Leu Cys	Val Leu His	Glu Lys	Thr Pro Val	Ser Asp	Arg Val Thr
3305		3310		3315	
Lys Cys	Cys Thr Glu	Ser Leu	Val Asn Arg	Arg Pro	Cys Phe Ser
3320		3325		3330	
Ala Leu	Glu Val Asp	Glu Thr	Tyr Val Pro	Lys Glu	Phe Asn Ala
3335		3340		3345	
Glu Thr	Phe Thr Phe	His Ala	Asp Ile Cys	Thr Leu	Ser Glu Lys
3350		3355		3360	
Glu Arg	Gln Ile Lys	Lys Gln	Thr Ala Leu	Val Glu	Leu Val Lys
3365		3370		3375	
His Lys	Pro Lys Ala	Thr Lys	Glu Gln Leu	Lys Ala	Val Met Asp
3380		3385		3390	
Asp Phe	Ala Ala Phe	Val Glu	Lys Cys Cys	Lys Ala	Asp Asp Lys
3395		3400		3405	
Glu Thr	Cys Phe Ala	Glu Glu	Gly Lys Lys	Leu Val	Ala Ala Ser
3410		3415		3420	
Gln Ala	Ala Leu Gly	Leu			
3425					

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Patentkrav

1. En modificeret Faktor VIII eller et kompleks omfattende modificeret Faktor VIII og
5 ikke-modificeret VWF, hvor den modificerede Faktor VIII er fusioneret ved en C-terminal del af det primære translationspolypeptid fra Faktor VIII til den N-terminale del af et albumin.
2. Modificeret Faktor VIII eller et kompleks omfattende modificeret Faktor VIII og
10 ikke-modificeret VWF ifølge krav 1, hvor
 - a. den modificerede Faktor VIII har en forlænget funktionel halveringstid sammenlignet med den funktionelle halveringstid af vildtype Faktor VIII eller
 - b. komplekset omfattende modificeret Faktor VIII og ikke-modificeret VWF har
15 en forlænget funktionel halveringstid sammenlignet med den funktionelle halveringstid af det tilsvarende kompleks omfattende vildtype Faktor VIII og vildtype VWF.
3. Modificeret Faktor VIII eller et kompleks omfattende nævnte modificerede Faktor
20 VIII eller et kompleks omfattende modificeret Faktor VIII og ikke-modificeret VWF ifølge krav 2, hvor den modificerede Faktor VIII har en funktionel halveringstid øget med mindst 25% sammenlignet med den funktionelle halveringstid for den tilsvarende vildtype Faktor VIII eller et kompleks omfattende nævnte modificerede Faktor VIII og ikke-modificeret VWF har en funktionel halveringstid øget med mindst 25% sammenlignet med det tilsvarende kompleks af vildtype Faktor VIII og vildtype VWF.
25
4. Modificeret Faktor VIII eller et kompleks omfattende modificeret Faktor VIII og ikke-modificeret VWF ifølge krav 1, hvor
 - 30 a. den modificerede Faktor VIII har en forlænget antigen-halveringstid sammenlignet med antigen-halveringstiden for vildtype Faktor VIII eller
 - b. komplekset omfattende modificeret Faktor VIII og ikke-modificeret VWF har en forlænget antigen-halveringstid sammenlignet med antigen-halveringstiden

for det tilsvarende kompleks omfattende vildtype Faktor VIII og vildtype VWF.

5 5. Modificeret Faktor VIII eller et kompleks omfattende modificeret Faktor VIII og ikke-modificeret VWF ifølge krav 4, hvor den modificerede Faktor VIII har en antigen-halveringstid øget med mindst 25% sammenlignet med antigen-halveringstiden af den tilsvarende vildtype Faktor VIII eller et kompleks omfattende nævnte modificerede Faktor VIII og ikke-modificerede VWF har en antigen-halveringstid øget med mindst 25% sammenlignet med det tilsvarende kompleks af vildtype Faktor VIII og vildtype VWF.

6. Modificeret Faktor VIII eller et kompleks omfattende modificeret FVIII og ikke-modificeret VWF ifølge krav 1, hvor

- 15 a. den modificerede Faktor VIII har en øget in vivo genvinding sammenligning med in vivo genvindingen af vildtype Faktor VIII eller
- b. komplekset omfattende modificeret Faktor VIII og ikke-modificeret VWF har en øget in vivo genvinding sammenlignet med in vivo genvindingen af det tilsvarende kompleks omfattende vildtype Faktor VIII og vildtype VWF.

20

7. Modificeret Faktor VIII eller et kompleks omfattende modificeret Faktor VIII og ikke-modificeret VWF ifølge krav 6, hvor den modificerede Faktor VIII har en in vivo genvinding øget med mindst 10% sammenlignet med in vivo genvindingen af den tilsvarende vildtype Faktor VIII eller et kompleks omfattende nævnte modificerede Faktor VIII og ikke-modificerede VWF har et in vivo genvinding øget med mindst 10% sammenlignet med det tilsvarende kompleks af vildtype Faktor VIII og vildtype VWF.

8. Modificeret Faktor VIII eller et kompleks omfattende modificeret Faktor VIII og ikke-modificeret VWF ifølge et hvilket som helst af de foregående krav, hvor den modificerede Faktor VIII har mindst 10% af den biologiske aktivitet af vildtype Faktor VIII eller komplekset omfattende det modificerede polypeptid eller et kompleks omfattende nævnte modificerede polypeptider har mindst 10% af den biologiske aktivitet af det tilsvarende kompleks af vildtype FVIII og vildtype VWF.

30

9. Rekombinant modificeret FVIII ifølge et hvilket som helst af de foregående krav, hvor nævnte rekombinante modificerede FVIII er udskilt fra pattedyrsceller i højere udbytte som vildtype FVIII.

5 10. Polynukleotid eller en gruppe af polynukleotider, kodende for en modificeret Faktor VIII ifølge et hvilket som helst af kravene 1 til 9.

11. Plasmid eller vektor omfattende et polynukleotid ifølge krav 10, hvor nævnte plasmid eller vektor er en ekspressionsvektor eller en overføringsvektor til anvendelse
10 i human gentterapi.

12. Værtscelle omfattende et polynukleotid ifølge krav 10 eller et plasmid eller en vektor ifølge krav 11.

15 13. Fremgangsmåde til fremstilling af en modificeret Faktor VIII omfattende:

(a) dyrkning af værtsceller ifølge krav 12 under betingelser sådan at den modificerede Faktor VIII er eksprimeret; og

(b) eventuelt genvinding af den modificerede Faktor VIII fra værtscellerne eller fra dyrkningsmediet.
20

14. Farmaceutisk sammensætning omfattende et polypeptid eller et kompleks omfattende nævnte modificerede Faktor VIII ifølge et hvilket som helst af kravene 1 til 9, et polynukleotid ifølge krav 10 eller et plasmid eller en vektor ifølge krav 11.
25

15. Anvendelse af et polypeptid eller et kompleks omfattende nævnte modificerede Faktor VII ifølge et hvilket som helst af kravene 1 til 9, et polynukleotid ifølge krav 10 eller et plasmid eller en vektor ifølge krav 11 eller en værtscelle ifølge krav 12 til fremstilling af et medikament til behandling eller forebyggelse af en blodkoagulationsforstyrrelse.
30

16. Anvendelse ifølge krav 15, hvor blodkoagulationsforstyrrelsen er hæmofili A.

17. Anvendelse ifølge krav 15, hvor behandlingen omfatter human gentterapi.

DRAWINGS

Figure 1: Antigen and activity levels of wild-type FVIII (457) and FVIII-C-terminal (1434) albumin fusion polypeptides

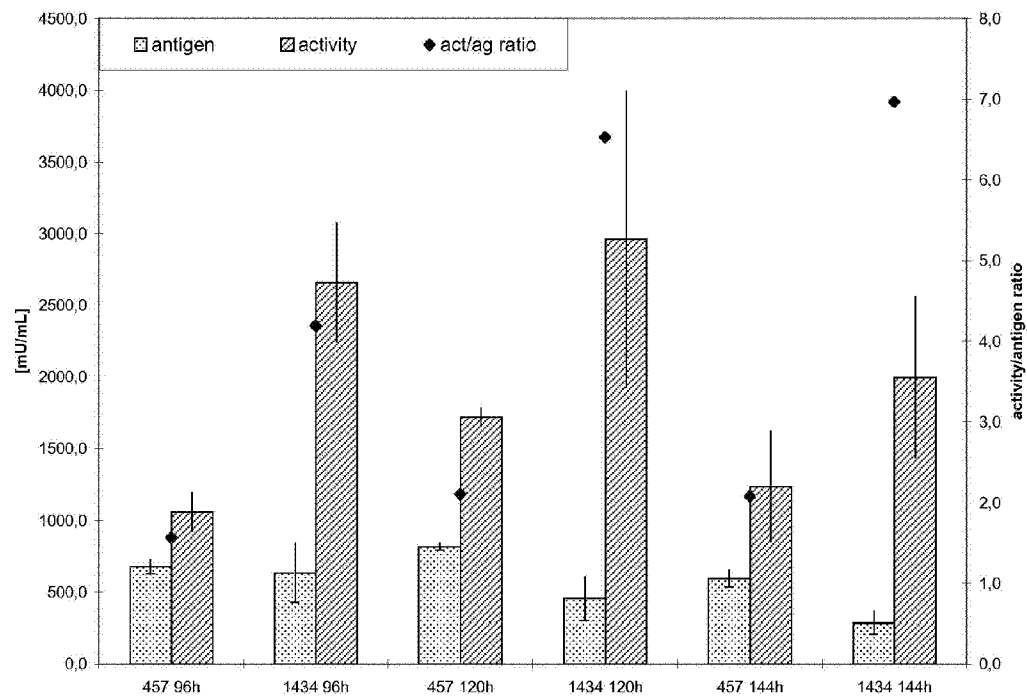


Figure 2: Comparison of human FVIII:Ag pharmacokinetics in VWF ko mice following i.v. injection of 100 U (FVIII:Ag)/kg FVIII wildtype and FVIII-FP 1656 VWF (mean; n=4/timepoint)

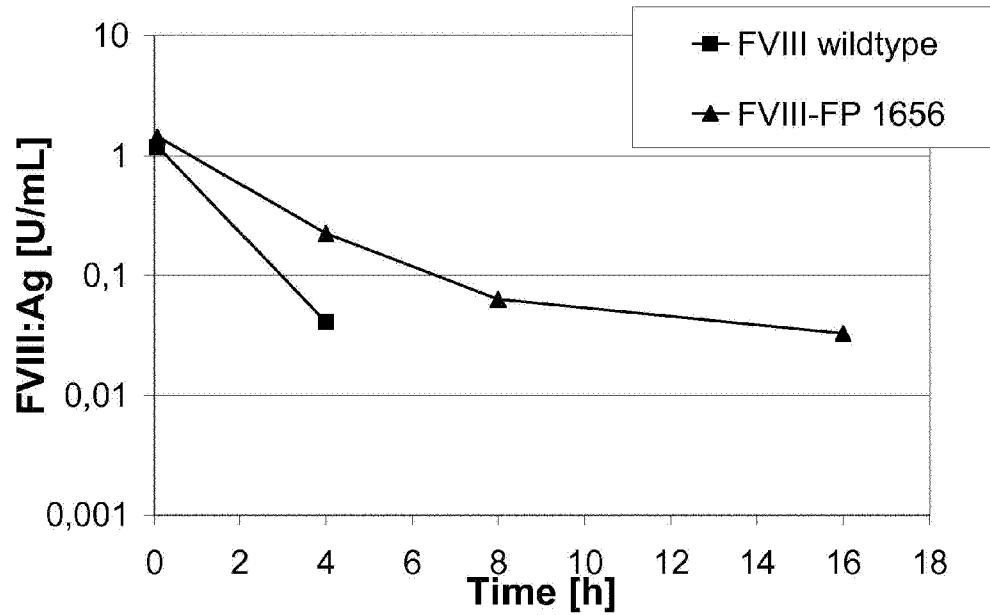


Figure 3: VWF:RCo/VWF:Ag ratios of cell culture supernatants containing wt rVWF (1570/797), rVWF-FP (1572/797) containing C-terminally linked albumin, or a mixed expression cell culture containing a mixture of wt rVWF (1570/797) and rVWF-FP (1572/797) transfected in a ratio of 5:1. Values of about 0,8 were obtained in every case that are close to 1 which is the theoretical ratio of NHP according to the unit definitions.

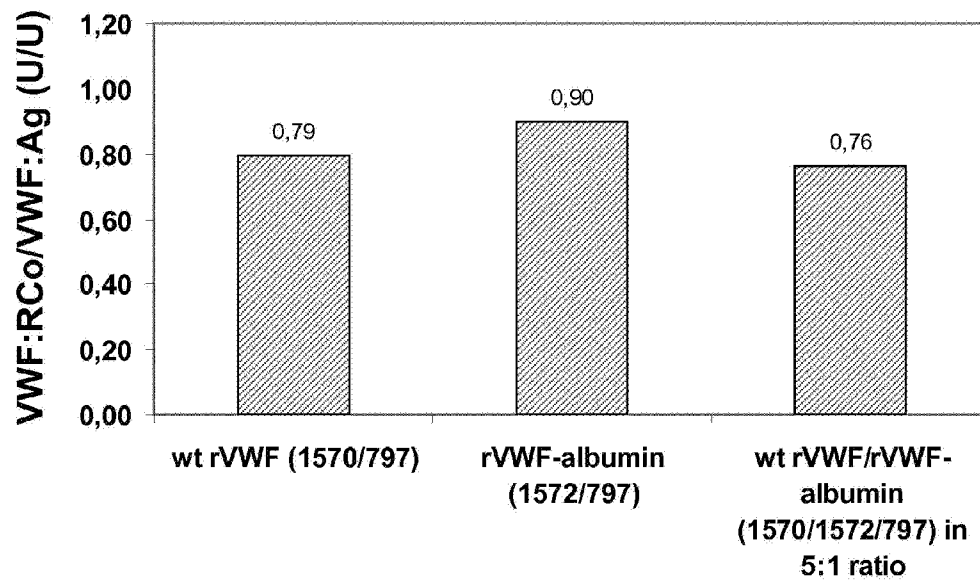
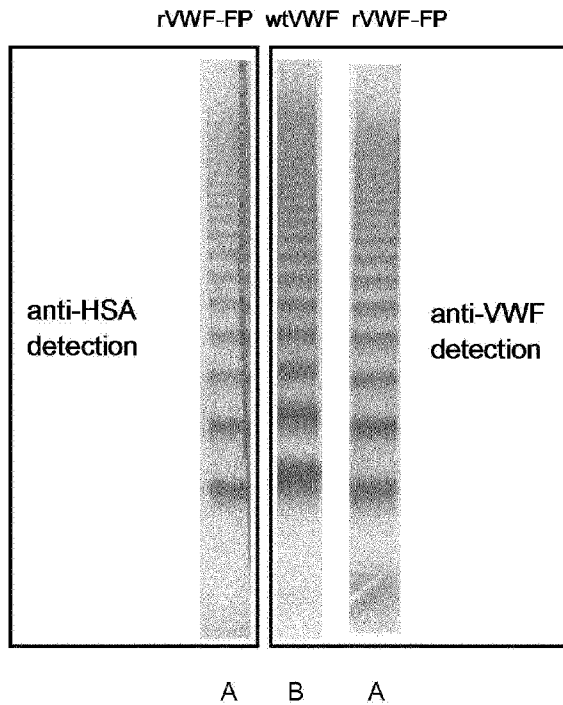


Figure 4: SDS-Agarose gel electrophoresis of wild-type rVWF (1570/797) (B) and rVWF-FP (1572/797), both expressed in HEK cells (A). Bands were detected using either antibodies to VWF or to albumin (HSA).



A = rVWF-FP (Expressed in presence of furin)

B = wt VWF (Expressed in presence of furin)

Figure 5: PK analysis of rVWF wt and rVWF-FP in rats based on VWF:Ag determination.

