



(86) Date de dépôt PCT/PCT Filing Date: 2013/02/22

(87) Date publication PCT/PCT Publication Date: 2013/08/29

(45) Date de délivrance/Issue Date: 2022/07/05

(85) Entrée phase nationale/National Entry: 2014/08/18

(86) N° demande PCT/PCT Application No.: KR 2013/001417

(87) N° publication PCT/PCT Publication No.: 2013/125891

(30) Priorité/Priority: 2012/02/24 (KR10-2012-0019221)

(51) Cl.Int./Int.Cl. *C07K 19/00* (2006.01),  
*C07K 16/00* (2006.01), *C07K 16/18* (2006.01),  
*C07K 16/28* (2006.01), *C07K 16/46* (2006.01)

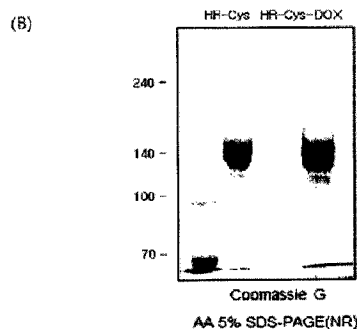
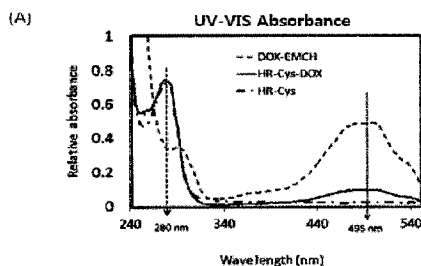
(72) Inventeurs/Inventors:  
PARK, SOON JAE, KR;  
CHUNG, HYE-SHIN, KR;  
KWON, SEONHUN, KR;  
LEE, SUNBAE, KR;  
YOU, SUN-AH, KR;  
KIM, YONG MO, KR

(73) Propriétaire/Owner:  
ALTEOGEN INC., KR

(74) Agent: GOWLING WLG (CANADA) LLP

(54) Titre : ANTICORPS MODIFIÉ DANS LEQUEL UN MOTIF CONTENANT UN RESIDU DE CYSTEINE EST LIÉ

(54) Title: MODIFIED ANTIBODY IN WHICH MOTIF COMPRISING CYSTEINE RESIDUE IS BOUND



(57) **Abrégé/Abstract:**

The present invention relates to an antibody in which a motif composed of an amino acid or peptide sequence including one or more cysteine residues is bound to the terminus of a parent antibody, particularly the terminus of the heavy chain of the parent antibody. Also, the present invention relates to a modified antibody-drug conjugate (mADC) comprising a drug bound to the antibody, and a method for producing the antibody or the modified antibody-drug conjugate. The modified antibody-drug conjugate according to the invention can accurately deliver the drug to a target cell due its high specificity to antigen, and thus can increase the therapeutic effect of the drug. Also, it can increase the usability of drugs, particularly anticancer drugs, the use of which is restricted due to their toxicity, despite their high efficacy. Moreover, the invention relates to a composition for treatment of diseases, particularly cancers, which comprise the modified antibody-drug conjugate.

**ABSTRACT**

The present invention relates to an antibody in which a motif composed of an amino acid or peptide sequence including one or more cysteine residues is bound to the terminus of a parent antibody, particularly the terminus of the heavy chain of the parent antibody. Also, the present invention relates to a modified antibody-drug conjugate (mADC) comprising a drug bound to the antibody, and a method for producing the antibody or the modified antibody-drug conjugate. The modified antibody-drug conjugate according to the invention can accurately deliver the drug to a target cell due its high specificity to antigen, and thus can increase the therapeutic effect of the drug. Also, it can increase the usability of drugs, particularly anticancer drugs, the use of which is restricted due to their toxicity, despite their high efficacy. Moreover, the invention relates to a composition for treatment of diseases, particularly cancers, which comprise the modified antibody-drug conjugate.

**MODIFIED ANTIBODY IN WHICH MOTIF COMPRISING CYSTEINE  
RESIDUE IS BOUND**

5

**TECHNICAL FIELD**

The present invention relates to a modified antibody comprising a cysteine (Cys)-containing motif bound to a parent antibody, preferably the terminus of the parent antibody, more preferably the terminus of the heavy or light chain of the parent antibody, and most preferably the C-terminus of the heavy or light chain of the parent antibody, a modified antibody-drug conjugate (mADC) comprising a drug for diagnosis or treatment bound to the modified antibody, and a method for producing the modified antibody or the modified antibody-drug conjugate.

The modified antibody-drug conjugate according to the present invention can accurately deliver the drug to a target cell due to the high antigen specificity of the parent antibody included in the conjugate, and thus can increase the therapeutic effect of the drug. Also, it can increase the utility of drugs, particularly anticancer drugs with high anticancer efficacy, the use of which is restricted due to their toxicity.

Moreover, the present invention relates to a composition,

including the modified antibody-drug conjugates, for the treatment of diseases, particularly cancer and to a method of treating diseases using the modified antibody-drug conjugate.

The modified antibody-drug conjugate according to the present invention may contain a number of cysteine residues that can be used for conjugating to drugs, because the motif containing one or more cysteine residues is bound to the parent antibody. Thus, it can contain a large amount of drugs bound thereto, and the large amount of bound drug can be effectively delivered to a target cell or tissue. Moreover, because the number of cysteine residues that are bound to the parent antibody can be easily controlled, the amount of the drug contained in the modified antibody-drug conjugate (mADC) can be easily controlled to a desired level.

In addition, because the modified antibody-drug conjugate according to the present invention can accurately deliver the drug to a target cell due to its high specificity to an antigen, it can increase the therapeutic effect of the drug and can also improve the utility of drugs, particularly anticancer drugs, which are restricted in use due to their toxicity, despite their high anticancer effect.

#### **BACKGROUND**

Among biopharmaceuticals, therapeutic agents using antibody

are being most actively studied, which bind specifically to targets (i.e., antigens) that are expressed specifically in certain diseases. Particularly, identification of tumor-related antigens that are expressed on the surface of cancer cells is being actively performed, and methods of diagnosing and treating tumors using antibodies (i.e., anticancer antibodies) that bind to the antigens to inhibit cell growth or induce cell death are being widely used, and the field of these methods also has a very good prospect.

10        Such anticancer antibodies have very high target specificity, but their cytotoxic effects on the cancer cells are usually lower than those of existing cytotoxic drugs (i.e., anticancer drugs or agents). So in many cases, these anticancer antibodies are subject to a combination therapy  
15 with cytotoxic drugs or other cell proliferation inhibitory drugs.

      In connection with the combination therapy as described above, there have been active studies on a modified antibody-drug conjugate (mADC), in which the therapeutic effect of the bound-cytotoxic drugs increases while the toxicity decreases.  
20 It is recognized that when the modified antibody-drug conjugate is used, the systemic toxicity of the drug can be reduced and the cytotoxicity of the drug can be enhanced specifically in cells (particularly cancer cells) having a

target overexpressed therein, thereby increasing the therapeutic effect of the drug.

Indeed, modified antibody-drug conjugates comprising a cytotoxic drug or radioisotope conjugated to an antibody, such as ZEVALIN<sup>TM</sup> [Witzig et al., J. Clin. Oncol, 2002, 20(15): 3262-3269]) or MYLOTARG<sup>TM</sup> [Drugs of the Future, 2000, 25(7):686]), have been successfully developed for the treatment of non-Hodgkin lymphoma or acute myeloid leukemia. In addition, many attempts have been made to conjugate highly toxic mertansine (such as cantuzumab mertansine (Immunogen, Inc. [Xie et al., J. of Pharm. and Exp. Ther. 2004, 308 (3):1073-1082]) or trastuzumab mertansine (Roche [Isakoff et al., J. Clin. Oncol. 2011, 29(4): 351-4])) to an antibody or conjugate other cytotoxic drugs, for example, dolastatin derivatives such as auristatin peptides, auristatin E (AE), monomethyl auristatin (MMAE) or MMAF to antibodies like cBR96 (specific to Lewis Y on carcinomas), cAC10 which is specific to CD30 on hematological malignancies, or anti-CD20 antibody for treatment of CD20-expressing cancer, or Rituxan for immune disorders, anti-EphB2R antibody for treatment of colorectal cancer, 2H9, anti-IL-8, or E-selectin antibody ([Klussman, et al., Bioconjugate Chemistry, 2004, 15(4):765-773]; [Doronina et al., Nature Biotechnology, 2003, 21(7):778-784]; [Francisco et al., Blood, 2003, 102(4):1458-

1465]) US 2004/0018194 A1) WO 04/032828 A3; [Mao et al., Cancer Research, 2004, 64(3):781-788]; [Bhaskar et al., Cancer Res, 2003, 63:6387-6394]).

In addition, an attempt has also been made to develop  
5 modified antibody-drug conjugates using daunomycin, doxorubicin, methotrexate or vindesine. It is known that bacterial toxins such as diphtheria toxin, plant toxins such as ricin, or small molecules such as geldanamycin ([Mandler et al., J. of the Nat. Cancer Inst, 2000, 92 (19):1573-1581];  
10 [Mandler et al., Bioorganic & Med. Chem. Letters, 2000, 10: 1025-1028]; [Mandler et al., Bioconjugate Chem, 2002, 13:786-791]), maytansinoid ([EP 1391213 A1]; [Liu et al., Proc. Natl. Acad. Sci. USA, 1996, 93: 8618-8623]) or calicheamicin ([Lode et al., Cancer Res, 1998, 58: 2928]; [Hinman et al., Cancer  
15 Res, 1993, 53: 3336-3342]) may be used as drugs in antibody-drug conjugates. These cytotoxic drugs exhibit cytotoxic and cell proliferation inhibitory effects by mechanisms such as tubulin binding, DNA binding or topoisomerase inhibition.

When a conventional process that was used to induce  
20 covalent linkage between a drug and an antibody is employed to prepare the antibody-drug conjugate as described above, the drug will be bound to a number of sites in the antibody, producing a heterogeneous mixture. For example, a cytotoxic drug is likely to be bound to an antibody through a number of

lysine residues contained in the antibody to produce a heterogeneous antibody-drug conjugate mixture. Also, the heterogeneous mixture might have different drug binding distribution ranging from 0 to about 8 depending on reaction  
5 conditions, which means that the number of drug molecules bound per unit antibody varies.

Also for the antibody drug conjugate having the defined number of drug binding, there might be another potential heterogeneity, depending on the various conjugation sites.  
10 Homogeneous purification from this heterogeneous mixture is not suitable for use in the mass production of medicines ([Hamblett et al., Clin. Cancer Res, 2003, 10, 7063-7070], [Wang et al., Protein Sci. 2005, 14, 2436-2446]).

Another method for conjugating a drug to an antibody is  
15 reducingdisulfide bonds between cysteine residues in antibody by reducing agents, and then conjugating drugs to free thiol groups of the reduced cysteine residues. This method also has disadvantages in that the inherent characteristics of the antibody can be lost and a heterogeneous mixture is produced  
20 in large amounts. Specifically, immunoglobulin M is an example of a disulfide-linked pentamer, while immunoglobulin G is an example of a protein with internal disulfide bridges bonding the subunits together. In such proteins, reduction of the disulfide bonds with a reagent such as dithiothreitol



(DTT) or selenol generates reactive free thiols ([Singh et al., Anal. Biochem. 2002, 304:147-156]). This approach may result in loss of antibody tertiary structure and antigen binding specificity ([Jagath et al., Nature Biotechnology, 5 2008, 26(8):925-32]).

The representative disadvantage of the conventional antibody-drug conjugation method as described above is that it is difficult to precisely control the drug conjugation sites in antibody and the number of conjugated drugs. In an attempt to overcome this problem and to introduce a free 10 thiol group, a specific amino acid was substituted with cysteine where the substituted cysteine did not impair the function of an antibody. For this purpose, a screening method for optimum cysteine mutant was developed after predicting 15 the reactivity of thiol group in each possible mutating site in antibody (Korean Patent Laid-Open Publication No. 2007-0054682, 'ThioFab technology'). An antibody-drug conjugate of trastuzumab mertansine produced by this method is in clinical trials for treatment of metastatic breast cancer ([Burris III 20 et al., J. Clin. Oncol, 2011, 29(4):398-405]). The above-described ThioFab technology has an advantage in that damage to a disulfide bond in a parent antibody can be minimized by introducing a new cysteine into the antibody, but the concern about modification of the structure and the function of the

parent antibody still remains, because some amino acids in the parent antibody are mutated by cysteine.

Accordingly, there is an urgent need for the development of a novel antibody-drug conjugate and a production method thereof, in which the number and position of drug molecules conjugated to a parent antibody can be accurately controlled while retaining the structural and functional characteristics of the parent antibody.

10

#### DISCLOSURE OF INVENTION

The present invention has been made in order to solve the above-described problems occurring in the prior art, and it is an object of the present invention to provide a novel antibody (hereinafter referred to as "modified antibody"), which comprises a motif containing one or more cysteine residues bound to a parent antibody to provide a plurality of drug-binding sites. Another object of the present invention is to provide a modified antibody-drug conjugate comprising a drug bound to the modified antibody.

20

The modified antibody according to the present invention can efficiently be conjugated with various drugs while retaining the characteristics of a parent antibody, and thus it can have a high target specificity and increase the therapeutic effect of the conjugated drugs.

The modified antibody of the present invention has advantages in that it comprises a motif containing one or more cysteine residues, and thus a drug can be conjugated to the cysteine residues and the conjugated drug can be effectively delivered to a target tissue. In addition, the number of cysteine residues that can bind to the drug can be accurately controlled, and thus the number or amount of drug molecules in the modified antibody-drug conjugate can be controlled. Accordingly, the modified antibody-drug conjugate of the present invention can be used as an excellent drug delivery system that overcomes the problem of conventional antibody-drug conjugates and can be efficiently used for the treatment of diseases such as cancer.

#### **BRIEF DESCRIPTION OF THE DRAWINGS**

FIG. 1 is a vector map of a modified-antibody expression vector comprising cysteine-containing trastuzumab.

FIG. 2 shows the westernblot image of HR-Cys after purification of trastuzumab (two right lanes) and a cysteine-containing trastuzumab variant (HR-Cys, two left lanes).

FIG. 3 shows the results of UV-VIS spectrometry (FIG. 3A) and SDS-PAGE (FIG. 3B) performed to confirm whether doxorubicin was conjugated to HR-Cys.

FIG. 4 shows the results of SDS-PAGE gel electrophoresis

(FIG. 4A) and the analysis of relative fluorescence intensity of Alexa488 dye bound to each of the heavy and light chains (FIG. 4B) for a Her-M2(Cys)-Alexa488 that is a conjugate of a Her-M2(Cys) antibody variant and Alexa Fluor® 488.

5        FIG. 5 shows the results of an anti-proliferation MTS assay performed to examine the cell growth inhibitory effect of HR-Cys-DOX in HER2-expressing BT-474 cells in comparison with those of Herceptin, a 1:2 mixture of Herceptin and doxorubicin, and doxorubicin.

10       FIG. 6 shows the results of an anti-proliferation assay performed to examine the cell growth inhibitory effect of HER-M(Cys)-MMAE that is a Herceptin-MMAE antibody-drug conjugate in HER2-expressing SK-BR3 cells in comparison with that of Herceptin.

15       FIG. 7 shows the results of observing caspase activation at various treatment concentrations in order to examine the cell death effect (apoptosis) of HR-Cys2-MMAE in HER2-expressing SK-BR-3 cells in comparison with that of Herceptin.

20       FIG. 8 shows the results of observing caspase activation at various treatment concentrations in order to examine the apoptosis effect of MMAE conjugated to a herceptin antibody variant in HER2-expressing SK-BR3 cells in comparison with that of Herceptin.

**BEST MODE FOR CARRYING OUT THE INVENTION**

Hereinafter, the present invention will be described in further detail.

The present invention provides a novel modified antibody, 5 which comprises a motif containing one or more cysteine residues bound to a parent antibody to provide a plurality of antibody drug sites, and a modified antibody-drug conjugate comprising the modified antibody. Such a modified antibody can be conjugated to various drugs while retaining the 10 characteristics of the parent antibody, and thus can be efficiently used as a modified antibody-drug conjugate.

As used herein, the term "parent antibody" means a conventional "antibody" having no cysteine-containing motif. Any parent antibody having binding affinity and specificity 15 to a specific antigen may be used in the present invention without limitation. Examples of parent antibodies that may be used in the present invention include monoclonal or polyclonal antibodies, such as antibodies derived from animals such as mice, chimeric antibodies, humanized 20 antibodies, and human antibodies developed using transgenic mice or phage-display technology. In addition, it will be obvious to those skilled in the art that modified antibodies such as bispecific antibodies, or fragments of the antibodies, may also be used in the present invention.

As used herein, the term "fragment of the antibody" refers to a fragment that at least retains a binding affinity to an antigen. Examples of the antibody fragment include single-chain antibodies, diabodies, triabodies, tetrabodies, Fab fragments, F(ab')<sub>2</sub> fragments, Fd, scFv, domain antibodies, minibodies, single-chain antibodies (scAb), derivatives of antibody constant regions, and artificial antibodies based on protein scaffolds.

In addition, examples of parent antibodies that may be used in the present invention include all types of immunoglobulin molecules (e.g., IgG, IgE, IgM, IgD, and IgA) and subtypes thereof (e.g., IgG1, IgG2, IgG3, IgG4, IgA1 and IgA2). In addition, the parent antibody may be one derived from any species.

The parent antibody in the present invention has binding affinity and specificity to cancer-specific antigens such as tumor-associated antigens (TAAs), cell surface receptor proteins, cell surface proteins and molecules other than receptor, transmembrane proteins, signaling proteins, cell survival regulators, cell proliferation regulators, molecules associated with (for e.g., known or suspected to contribute functionally to) tissue development or differentiation, lymphokines, cytokines, molecules involved in cell cycle regulation, molecules involved in

vasculogenesis and molecules associated with (for e.g., known or suspected to contribute functionally to) angiogenesis.

Specifically, antigens to which the parent antibody in the present invention can bind, but are not limited to:

- 5 (1) BMPRIIB (bone morphogenetic protein receptor-type IB; Genbank Accession No. NM\_001203);
- (2) E16 (LAT1, SLC7A5; Genbank Accession No. NM\_003486);
- (3) STEAP1 (six transmembrane epithelial antigen of prostate; Genbank Accession No. NM\_012449);
- 10 (4) 0772P (CA125, MUC16, Genbank Accession No. AF361486);
- (5) MPF (MPF, MSLN, SMR, megakaryocyte potentiating factor, mesothelin; Genbank Accession No. NM\_005823);
- (6) Napi3b (NAPI-3B, NPTIIB, SLC34A2, solute carrier
- 15 family 34 (sodium phosphate), member 2, type II sodium-dependent phosphate transporter 3b; Genbank Accession No. NM\_006424);
- (7) Sema 5b (FLJ10372, KIAA1445, Mm.42015, SEMA5B, SEMAG, Semaphorin 5b Hlog, sema domain, seven thrombospondin repeats
- 20 (type 1 and type 1-like), transmembrane domain (TM) and short cytoplasmic domain, (semaphorin) 5B; Genbank Accession No. AB040878);
- (8) PSCA hlg (2700050C12Rik, C530008O16Rik, RIKEN cDNA 2700050C12, RIKEN cDNA 2700050C12; Genbank Accession No.

AY358628);

(9) ETBR (Endothelin type B receptor; Genbank Accession No. AY275463);

(10) MSG783 (RNF124, hypothetical protein FLJ20315, 5 Genbank Accession No. NM\_017763);

(11) STEAP2 (HGNC\_8639, IPCA-1, PCANAP1, STAMP1, STEAP2, STMP, prostate cancer associated gene 1, prostate cancer associated protein 1, six-transmembrane epithelial antigen of prostate 2, six-transmembrane prostate protein, Genbank 10 Accession No. AF455138);

(12) TrpM4 (BR22450, FLJ20041, TRPM4, TRPM4B, transient receptor potential cation channel, subfamily M, member 4; Genbank Accession No. NM\_017636);

(13) CRIPTO (CR, CR1, CRGF, CRIPTO, TDGF1, 15 teratocarcinoma-derived growth factor; Genbank Accession No. NP\_003203 or NM\_003212);

(14) CD21 (CR2 (Complement receptor 2) or C3DR (C3d/Epstein Barr virus receptor) or Hs.73792; Genbank Accession No. M26004);

20 (15) CD79b (CD79B, CD79 $\beta$ , Igb (immunoglobulin-associated beta), B29, Genbank Accession No. NM\_000626);

(16) FcRH2 (IFGP4, IRTA4, SPAP1A (SH2 domain containing phosphatase anchor protein 1a), SPAP1B, SPAP1C; Genbank Accession No. NM\_030764);



- (17) HER2 (Genbank Accession No. M11730);
- (18) NCA (Genbank Accession No. M18728);
- (19) MDP (Genbank Accession No. BC017023);
- (20) IL20R $\alpha$  (Genbank Accession No. AF184971);
- 5 (21) Brevican (Genbank Accession No. AF229053);
- (22) EphB2R (Genbank Accession No. NM\_004442);
- (23) ASLG659 (Genbank Accession No. AX092328);
- (24) PSCA (Genbank Accession No. AJ297436);
- (25) GEDA (Genbank Accession No. AY260763);
- 10 (26) BAFF-R (B cell-activating factor receptor, BLyS receptor, BR3, NP\_443177.1);
- (27) CD22 (B-cell receptor CD22-B isoform; NP-001762.1);
- (28) CD79a (CD79A, CD79.alpha., immunoglobulin-associated alpha, a B cell-specific protein that covalently
- 15 interacts with Ig beta (CD79B) and forms a complex on the surface with IgM molecules, transduces a signal involved in B-cell differentiation; Genbank Accession No. NP\_001774.1);
- (29) CXCR5 (Burkitt's lymphoma receptor 1, a G protein-coupled receptor that is activated by the CXCL 13 chemokine,
- 20 functions in lymphocyte migration and humoral defense, plays a role in HIV-2 infection and regarded for development of AIDS, lymphoma, myeloma, and leukemia; Genbank Accession No. NP\_001707.1);
- (30) HLA-DOB (Beta subunit of MHC class II molecule (la

antigen) that binds peptides and presents them to CD4+ T lymphocytes; Genbank Accession No. NP\_002111.1);

(31) P2X5 (Purinergic receptor P2X ligand-gated ion channel 5, an ion channel gated by extracellular ATP, may be involved in synaptic transmission and neurogenesis, deficiency may contribute to the pathophysiology of idiopathic detrusor instability; Genbank Accession No. NP\_002552.2);

(32) CD72 (B-cell differentiation antigen CD72, Lyb-2; Genbank Accession No. NP\_001773.1);

(33) LY64 (Lymphocyte antigen 64 (RP105), type I membrane protein of the leucine rich repeat (LRR) family, regulates B-cell activation and apoptosis, loss of function is associated with increased disease activity in patients with systemic lupus erythematosus; Genbank Accession No. NP\_005573.1);

(34) FcRH1 (Fc receptor-like protein 1, a putative receptor for the immunoglobulin Fc domain that contains C2 type Ig-like and ITAM domains, may have a role in B-lymphocyte differentiation; Genbank Accession No. NP\_443170.1);

(35) IRTA2 (Immunoglobulin superfamily receptor translocation associated 2, a putative immunoreceptor with possible roles in B cell development and lymphomagenesis;

deregulation of the gene by translocation occurs in some B cell malignancies; Genbank Accession No. NP\_112571.1); and

(36) TENB2 (putative transmembrane proteoglycan, related to the EGF/heregulin family of growth factors and  
5 follistatin; Genbank Accession No. AF179274). In addition, examples of the antigens include all antigens that may be used for treatment and diagnosis.

In a preferred embodiment, the parent antibody in the present invention has binding affinity and specificity to an  
10 ErbB receptor selected from among EGFR, HER2, HER3 and HER4, and other cancer antigens.

Particularly, the parent antibody that is used in the present invention comprises one or more selected from among trastuzumab (trade name: Herceptin), rituximab (trade name:  
15 Rituxan), bevacizumab (trade name: Avastin), cetuximab (trade name: Erbitux), cBR96, cAC10, anti-CD20 antibody, anti-EphB2 antibody, anti-IL-8, E-selectin antibody, anti-MUC16 antibody, and anti-CD30 antibody, but is not limited thereto.

HER2 means a member of the epidermal growth factor  
20 receptor (EGFR) family, an important signaling cascade that is implicated in the proliferation and survival of breast cancer cells. It is known that the receptor tyrosine kinases of the EGFR family are composed of erb1, erb2/HER2, erb3 and erb4 and are involved in the regulation of the adhesion,

migration and differentiation of cells in addition to cell proliferation and proliferation.

There is no ligand that binds to erb2/HER2 of the four members of the erb family, but erb2/HER2 is known as the most potent oncogene in breast cancer. If the level of HER2 is normal, it is involved in the growth and development of normal mammary tissue, but if HER2 is abnormally overexpressed or amplified, normal cell regulation will be broken so that aggressive cancer cells will be formed in mammary tissue. In other words, if HER2 is activated by oligomerization with other members of the EGFR family, it will phosphorylate many downstream molecules, which leads to activation of a variety of signaling cascades. The SOS-Ras-Raf-MEK-MAPK pathway that is involved in cell proliferation and the PI-3K/Akt pathway that inhibits cell death are representative mechanisms associated with cancer cell proliferation.

The results of preclinical and clinical trials indicate that the overexpression of HER2 is an important biomarker that appears from the initial state of cancer development and plays an important role in the growth and progression of cancer. It is known that the overexpression of HER2 appears in about 20-30% of invasive breast cancers and is also associated with poor prognosis of breast cancer with higher

aggressiveness and malignancy.

According to the present invention, a motif containing one or more cysteine residues is bound to a parent antibody that has specificity to a growth factor receptor selected from the group consisting of the HER2 receptors or EGF  
5 receptors. And a conventional anticancer drug is conjugated to the motif to obtain a modified antibody-drug conjugate of the present invention. When the modified antibody-drug conjugate is administered to a patient in an amount effective  
10 to inhibit the growth of tumor cells in the patient, it can exhibit an excellent effect of treating cancer by inhibiting the growth of tumor cells that overexpress the growth factor receptor while inducing the cell death.

In addition, the parent antibody in the present  
15 invention is preferably trastuzumab. Although trastuzumab is prepared so as to be substantially free of C-terminal lysine (Lys), the modified antibody-drug conjugate of the present invention could be applied to trastuzumab that contains lysine (Lys) or trastuzumab that does not contain lysine  
20 (Lys) in the C-terminal region thereof, where the cysteine (Cys)-containing motif may be bound thereto.

Amino acids in the present invention are expressed by their known 3-letter or 1-letter abbreviations. Nucleotides present in various nucleic acid fragments are designated by

the standard single-letter designation used routinely in the art.

The cysteine-containing motif in the present invention has 1-100 amino acid residues, preferably 1-50 amino acid residues, more preferably 1-30 amino acid residues, and most preferably 1-10 amino acid residues, and contains one or more cysteine residues. Particularly, the cysteine-containing motif in the present invention preferably contains 1-20 cysteine residues, more preferably 1-10 cysteine residues, and even more preferably 1-5 cysteine residues.

The cysteine-containing motif in the present invention may be a simple peptide motif having no specific functionality or secondary or tertiary structure and is preferably a motif having a specific functionality or a secondary or tertiary structure. The specific functionality is preferably the property capable of maintaining/protecting the chemical conjugation ability of cysteine residues, but is not limited thereto. Particularly, the cysteine-containing motif can more effectively maintain the functionality of cysteine residues by preventing or retarding the oxidation of the cysteine residues due to binding to a specific ligand or the secondary or tertiary structure of the peptide motif itself.

The cysteine-containing motif in the present invention

has a structure represented by the following formula 1:

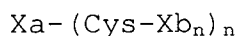


wherein  $(\text{M}_{\text{Cys}})_n$  represents a simple cysteine residue or a peptide motif that contains a cysteine residue and has a specific functionality or a secondary or tertiary structure;  
 5 Xa and  $\text{Xb}_n$  each independently represents a peptide comprising 0 to 20 amino acid residues other than cysteine; and n is an integer ranging from 1 to 20.

In formula (1),  $(\text{M}_{\text{Cys}})_n$ , that is,  $(\text{M}_{\text{Cys}})_1$ ,  $(\text{M}_{\text{Cys}})_2$  ....  $(\text{M}_{\text{Cys}})_n$ ,  
 10 may be the same as or different from each other. Also,  $\text{Xb}_n$ , that is,  $\text{Xb}_1$ ,  $\text{Xb}_2$  ....  $\text{Xb}_n$ , may be the same as or different from each other.

In addition, if  $(\text{M}_{\text{Cys}})_n$  in formula (1) is a simple cysteine residue, the cysteine-containing motif according to  
 15 the present invention has a structure represented by the following formula (2):

Formula (2)



wherein Xa,  $\text{Xb}_n$  and n are the same as defined in formula  
 20 1.

Also, if  $(\text{M}_{\text{Cys}})_n$  in formula (1) is a peptide motif that contains a cysteine residue and has a specific functionality or a secondary or tertiary structure,  $(\text{M}_{\text{Cys}})_n$  may preferably be a metal ion binding motif containing a cysteine residue. It

is known that the metal ion binding motif containing a cysteine residue can bind to a metal ion to inhibit oxidation of the cysteine residue, thereby effectively retaining the alkylation reactivity of the cysteine residue (Van Horn et al. 5 (2003) J. Biol. Inorg. Chem. 8: 601-610).

Examples of a metal ion binding motif containing a cysteine residue, which may be used in the present invention, include, but are not limited to, metal ion chelators (Zhang et al. (2012) Biochem. Genet. 50(7-8): 585-599) that are used 10 to control the concentration of metal ions *in vivo*, chaperones (Ansbacher and Shurki, J. Phys. Chem. B (2012) 116(15): 4425-4432; Allen et al. (2012) Biochemistry 51(7): 1439-48; Click et al. (2012) H. Comput. Chem. 33(11): 1142-51) that function to deliver metal ions to a specific 15 position outside or inside cells, transcriptional regulators (Gunther et al. (2012) Biochim. Biophys. Acta. 1823(2): 476-483; Sitthisak et al. (2012) FEMS Microbiol. Lett. 327(2): 126-133) that regulate transcription depending on the concentration of metal ions, zinc finger motifs (MacPherson 20 et al. (2006) Microbiol. Mol. Bio. Rev. 70(3):583-604; Schaeffer et al. (2012) Nucleic Acids Res. 40(18): 9298-9307) that are widely present in many proteins and are involved in protein-protein interactions or protein-DNA interactions, and motifs (Zielazinski et al. (2012) Biochemistry 51(40):7891-



7900; Zhou et al. (2012) FEBS J. 279(2):285-298; Cochran et al. (2011) Nat. Struct. Mol. Biol. 19(1):122-127) derived from various enzymes. The metal ion binding motifs contains a cysteine residue as an essential binding group thereof, and  
 5 such metal ion binding motifs may be used in the production of the modified antibody-drug conjugate of the present invention.

Preferred examples of the metal ion binding motif in the present invention include, but are not limited to, C<sub>2</sub>H<sub>2</sub> group  
 10 (Cys<sub>2</sub>His<sub>2</sub> class: Cys-X<sub>2-4</sub>-Cys-X<sub>12</sub>-His-X<sub>3-5</sub>-His), C<sub>4</sub> group (C<sub>4</sub> class: Cys-X<sub>2</sub>-Cys-X<sub>n</sub>-Cys-X<sub>2</sub>-Cys-X<sub>m</sub>-Cys-X<sub>2</sub>-Cys-X<sub>n</sub>-Cys-X<sub>2</sub>-Cys) or C<sub>6</sub> group (C<sub>6</sub> class: Cys-X<sub>2</sub>-Cys-X<sub>6</sub>-Cys-X<sub>5-12</sub>-Cys-X<sub>2</sub>-Cys-X<sub>6-8</sub>-Cys) of zinc finger protein, a Cys-X<sub>m</sub>-Cys motif such as Cys-X-X-Cys or Cys-X-Cys, a Met-X-Cys-X-X-Cys motif or a C-Q-C-Q-C-A-C  
 15 motif, which is frequently found in transcription regulatory proteins, metal chaperone proteins, metal ion transporters, superoxide dismutase or the like, a Ser-Pro-Cys motif of membrane protein ATPase, etc. In the metal binding peptide motif described above, X represents amino acid residues other  
 20 than Cys; m is an integer ranging from 1 to 10, and preferably from 1 to 5; and X<sub>m</sub> or X<sub>m-q</sub> represents amino acid residues other than Cys, the number of which is indicated by m or m to q.

More specifically, examples of the metal ion binding

motif of zinc finger protein, which can be used in the present invention, include, but are not limited to,

YKCKQCGKAFGCPSNLRHRGRTH (SEQ ID NO:1),  
 YQCNICGGKCFSCNSNLHRHQORTH (SEQ ID NO:2),  
 5 YSCGICGKSFSDSAKRRHCILH (SEQ ID NO:3),  
 YTCSDCGKAFRDKSCLNRHRRTH (SEQ ID NO:4),  
 YRCKYCDRSFSDSSNLQRHVRNIH (SEQ ID NO:5),  
 YKCKECGKA FNHSSNFNKHHRTH (SEQ ID NO:6),  
 FKCPVCGKA FRHSSSLVRHQORTH (SEQ ID NO:7),  
 10 YRCKYCCDRSFSISSNLQRHVRNIH (SEQ ID NO:8),  
 YECDHCGKA FSGSNLNVHRRTH (SEQ ID NO:9),  
 YGCHLCCKA FSKSSNLRRHEMTH (SEQ ID NO:10),  
 YKCKECGQA FRQRAHLIRHKLH (SEQ ID NO:11),  
 YKCHQCGKA FIQSFNLRRHERTH (SEQ ID NO:12),  
 15 FQCNQCGASFTQKGNLNRHIKLH (SEQ ID NO:13),  
 YTCSYCGKSFTQSNTLKQHTRTH (SEQ ID NO:14),  
 YACHLCGKAFTQSSHRRHEKTH (SEQ ID NO:15),  
 YKCGQCGKFYSQVSHLTRHQKTH (SEQ ID NO:16),  
 YACHLCGKAFTQCSHLRRHEKTH (SEQ ID NO:17),  
 20 YACHLCAKA FIQCSHLRRHEKTH (SEQ ID NO:18),  
 YVCRECGRGFRQHSHLVRHKRTH (SEQ ID NO:19),  
 YKCEECEGKA FRQSSHLTTHKTH (SEQ ID NO:20),  
 YECDHCGKSFSQSSHLNVHKRTH (SEQ ID NO:21),  
 YMCSECGRGFSQKSNLTIHQORTH (SEQ ID NO:22),

YKCEECGKAFTQSSNLTKHKKIH (SEQ ID NO:23),  
 FECKDCGKAFIQKSNLIRHQORTH (SEQ ID NO:24),  
 YVCRECRRGFSQKSNLIRHQORTH (SEQ ID NO:25),  
 YECEKCGKAFNQSSNLTRHKKSH (SEQ ID NO:26),  
 5 YECVQCGKSYSQSSNLFRHQRRH (SEQ ID NO:27),  
 YECVQCGKGFTQSSNLITHQRVH (SEQ ID NO:28),  
 YECNTRKRTFSQKSNLIVHQORTH (SEQ ID NO:29),  
 YVCSKCGKAFTQSSNLTVHQKIH (SEQ ID NO:30),  
 YKCDECGKNFTQSSNLIVHKRIH (SEQ ID NO:31),  
 10 YECDVCGKTFTQKSNLGVHQORTH (SEQ ID NO:32),  
 YKCPDCGKSFSQSSSLIRHQORTH (SEQ ID NO:33),  
 YECQDCGRAFNQNSSLGRHKRTH (SEQ ID NO:34),  
 YECNECGKFFSQSSSLIRHRRSH (SEQ ID NO:35),  
 YKCEECGKAFNQSSTLTRHKIVH (SEQ ID NO:36),  
 15 YECNECGKAFAQNSTLRVHQRIH (SEQ ID NO:37),  
 YEVDHDCGKSFRQSTHTLTQHRRIH (SEQ ID NO:38),  
 YECHDCGKSFRQSTHLTRHRRRIH (SEQ ID NO:39),  
 HKCLECGKCFSQNTHLTRHQORTH (SEQ ID NO:40),  
 YVCDVEGCTWKFARSDELNRHKKRH (SEQ ID NO:41),  
 20 YHCDWDGCGWKFARSDELTRHYRKH (SEQ ID NO:42),  
 YRCSWEGCEWRFARSDELTRHFRKH (SEQ ID NO:43),  
 FSCSWKGCERRFARSDELSRHRRTTH (SEQ ID NO:44),  
 FACSWQDCNKKFARSDELARHYRTH (SEQ ID NO:45),  
 YHCNWDGCGWKFARSDELTRHYRKH (SEQ ID NO:46),

FLCQYCAQRFRGRKDHLTRHMKHSH (SEQ ID NO:47),  
 CRCNECGKSFSRRDHLVRHQORTH (SEQ ID NO:48),  
 FQCKTCQRKFSRSDHLKTHTRTH (SEQ ID NO:49),  
 FACEVCGVRFTNRNDKLIHMRKH (SEQ ID NO:50),  
 5 YVCDVEGCTWKFARSDKLNRRHKKRH (SEQ ID NO:51),  
 YKMECGKAFNRRSHLTRHQRIH (SEQ ID NO:52),  
 YICRKCGRGFSRKSNLIRHQORTH (SEQ ID NO:53),  
 YECKECGKAFSSGSNFTRHQRIH (SEQ ID NO:54),  
 FHCGYCEKSFSVKDYLTKHIRTH (SEQ ID NO:55),  
 10 YECDHCGKAFSVSSNLNVHRRRIH (SEQ ID NO:56),  
 YTCKQCGKAFSVSSSLRRHETTH (SEQ ID NO:57),  
 YECNYCGKTFSVSSTLIRHQRIH (SEQ ID NO:58),  
 YRCEECGKAFRWPSNLTRHKRIH (SEQ ID NO:59),  
 FACDICGRKFARSDERKRHTKIH (SEQ ID NO:60),  
 15 CPVESCRRFRSRSDELTRHIRIH (SEQ ID NO:61),  
 CDICGRKFARSDERKRHTKIH (SEQ ID NO:62), etc. The metal ion  
 binding motif of zinc finger protein may be any one selected  
 from the metal ion binding motifs of zinc finger proteins  
 described in Macpherson et al. (2006) Microbiol. Mol. Biol.  
 20 Rev. 70(3), 583-604, which is consistent with the technical  
 characteristics of the present

invention.

Examples of the metal ion binding motif of transcription regulatory proteins, metal chaperone proteins or the like, which can be used in the present invention, include, but are

5 not limited to, the followings:

CadC protein:

CEIFCYDEEKVNRIQGDLQTVDISGVSQILKAIADENRAKITYALCQDEELCVC (SEQ ID NO:63),

AztR

protein:

10 DTHLVHLDNVRSSQAQILPTDKAQQMAEIFGVLADTNRIRLLSALASSELCVC (SEQ ID NO:64),

ZiaR protein:

CDQPLVHLEQVRQVQPEVMSLDQAQQMAEFFSALADPSRLRLMSALARQELCVC (SEQ ID NO:65),

15 BxmR protein:

CDRAHLVDCSRVGDIQTQVLNTAKAQRMAEFFSLLGDANRLRVVSVLAKQELCVC (SEQ ID NO:66),

ArsR protein: LSDETRLGIVLLLREMGELCVCDLCM (SEQ ID NO:67),

CCTLATGPLSSDESEHYADLFKVLGDPVRLRILSQLAAGGC (SEQ ID NO:68),

20 YRAAMPVVRLVAYLTENCCHGTRDC (SEQ ID NO:69),

CmtR protein:

CLRGCGLVVATYEGRQVRYALADSHLARALGELVQVVLAVDTDQPC (SEQ ID NO:70),

LRDCGLVVTVDPDGRRSRYELADERLGHALDDLRAAVVAVDADRTCPDADELECC (SEQ ID

NO:71), etc.

Zinc (Zn) binding proteins found in membrane proteins that are involved in bacterial signaling transduction have an inherent metal ion binding motif composed of one cysteine  
 5 residue and three histidine residues (Draper et al., J. Bacteriol. 2011, 193 (17), 4338-4345). Herein, the metal ion binding motif has a structure of HXXWFYLY<sub>21-28</sub>CXLFMVIGXWFLVIX<sub>18-27</sub>HXXH, wherein X represents any amino acid, and X<sub>m-q</sub> represents amino acid residues other than Cys, the number of  
 10 which is indicated by m to q. In addition, 150 or more zinc binding proteins reported to date may be used as the metal ion binding motif in the present invention. Known metal ion transporters include cation diffusion facilitators, Zrt, Irk-like proteins, cation exchangers,  
 15 copper transporters, heavy metal P-type ATPase, ATP-binding cassette transporters, etc. (Hanikenne et al. Plant Physiology 2005, 137, 428-446; Hall and Williams, J. Experimental Botany, 2003, 54(393) 2601-2613), and the following M-X-C-X-X-C motif may preferably be used in the  
 20 present invention, but is not limited thereto:

*E.coli* ZntA : VSGMDCAACARKVENAVRQLAGVNQVQVLFA (SEQ ID NO:72)

Tn501 MerP : VPGMTCSACPITVKKAISEVEGVSKVDVTFE (SEQ ID NO:73)

Tn501 MerA : ITGMTCDSCAAHVKEALEKVPGVQSALVSY (SEQ ID NO:74)

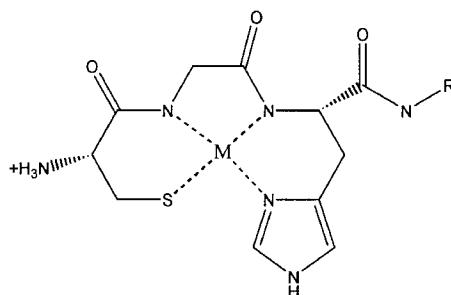
- S.aureus* CadA : VQ**GFTCANC**AGKFEKNVKKIPGVQDAKVNFG (SEQ ID NO:75)
- Human Menkes : VE**GMTCN**SCVWTIEQQIGKVNGEHHIKVSLE (SEQ ID NO:76)
- Yeast Atx1 : V**VMTC**SGCSGAVNKVLTLEPDVSKIDIS (SEQ ID NO:77)
- 5 Rat Wilsons : **GMTCA**SCVANIERNLRREEGIYSV (SEQ ID NO:78)
- Hum Wilsons : YE**GMTCQ**SCVSSIEGKYRKLQGVVRYKVSL (SEQ ID NO:79)
- Rice Cu ATPase : **GMSCQ**GCAGAVRRVLTKMEGVETFDIDME (SEQ ID NO:80)
- H.pylori* Cu ATPase : VP**SITCS**HCVDKIEKFVGEIEGVSFIDANVE (SEQ ID NO:81)
- 10 Ran1 : VTGMTCAACSNSVEAALMNVNGVDVGGMTCGGCSASVKKLLESQPCVASASV (SEQ ID NO:82)
- Cpx89 : VSGMVCAACSTAVENALLSCSGV (SEQ ID NO:83)
- 15 Paa1 : DVGGMTCGGCSASVKKILESQP (SEQ ID NO:84)
- Cpx1184 : DVGGMKCGGCVEHVKKILEEQFGVTSAS (SEQ ID NO:85)

In addition, various metal ion binding motifs artificially designed based on wild-type metal ion binding motifs found *in vivo* may be used in the modified antibody-

20 drug conjugate according to the present invention. Examples for these metal binding motifs include, but are not limited to, a CGH motif (Van Horn et al. (2003) J. Biol. Inorg. Chem. 8: 601-610) designed based on a GGH motif from Zinc finger

protein which does not contain cysteine residue and metal ion binding motifs (Jancso et al. (2011) Metallomics 3(12):1331-1339) obtained by substituting one or more cysteine residues in a peptide motif with other amino acid residue such as threonine, serine or histidine. The CGH motif has a structure represented by the following chemical formula 1, and preferably has a structure of ACGHA having alanines both at the C-and the N-termini.

Chemical Formula 1



[wherein M represents a metal ion, and R represent an amino acid residue other than cysteine, preferably alanine.]

The CGH motif in the present invention still has the metal ion binding property even when the positions of the N-terminus and the C-terminus are reversed. Thus, it will be obvious to those skilled in this field of art that an HGC motif having the reversed positions of the N-terminus and the C-terminus falls within the scope of the CGH motif in the present invention.

Metal ion binding motifs artificially designed based on



zinc finger motifs, which may be used in the present invention, include the following metal binding peptide motifs (see Roehm and Berg, J. Am. Chem. Soc. 1998, 120. 13083-13087) obtained by substituting a cysteine residue (which  
 5 plays the most essential role) in PYKCPECGKSFSQKSALVKHQRTHTH with methyl-cysteine (Me-Cys) or substituting a histidine residue in this sequence with cysteine:

PYKCPECGKSFSQKSALVKHQRTHTC (SEQ ID NO:86),  
 PYKCPECGKSFSQKSALVKHQRTHTM (SEQ ID NO:87),  
 10 PYKCPECGKSFSQKSALVKHQRTHT (Me-C) (SEQ ID NO:88),  
 PYK (Me-C) PEGKSFSQKSALVKHQRTHTH (SEQ ID NO:89),  
 PYKCPE (Me-C) GKSFSQKSALVKHQRTHTH (SEQ ID NO:90), and  
 PYKCPE (Me-C) GKSFSQKSALVKHQ (SEQ ID NO:91)

15 [In the metal binding peptide motif described above, Me-C represents a methylated cysteine residue.]

In addition, peptides artificially designed based on many metal ion binding protein motifs reported to date may also be used in the present invention.

20 Metal ion binding motifs based on the secondary or tertiary structure of protein include beta-sheets, mixed alpha/beta motifs, and alpha-helix structures which are most frequently found. The alpha-helix structures include peptides having single-stranded, double-stranded, triple

stranded or quadruple-stranded alpha-helix structures.

With respect to these multiple-stranded alpha-helices, the TRI family, for example, has a structure of G(LKALEEK)<sub>4</sub>G having four repeats of the peptide sequence LKALEEK.

5 Artificially designed metal ion-binding peptide motifs obtained by substituting a specific amino acid in this TRI family with cysteine have been reported (Peakcock et al. 2009. Dalton Trans. 7(13). 2271-2280), and artificially designed peptide motifs having the following structures may be used in  
10 the present invention, but are not limited thereto:

GLKALEEK**C**KALEEKLKALEEKLKALEEKG (SEQ ID NO:92)

GLKALEEKLKALEEKLKACEEKLKALEEKG (SEQ ID NO:93)

GLKALEEK**C**KALEEKLKACEEKLKALEEKG (SEQ ID NO:94)

GLKALEEKLKALEEK**C**KALEEKLKALEEKG (SEQ ID NO:95)

15 GLKALEEKLKALEEKLKALEEK**C**KALEEKG (SEQ ID NO:96)

GLKALEEKLKALEEKLKALEEKLKAAEEK**C**KALEEKG (SEQ ID NO:97)

GLKALEEKLKALEEK**C**KALEEKLKAAEEK**C**KALEEKG (SEQ ID NO:98)

ELYALEKELGALEKELACLEKELGALEKELYALEK (SEQ ID NO:99)

KLYALKEKLGALKEKLACLEKELGALKEKLYALKE (SEQ ID NO:100)

20 ELYALEKELGALEKELACLEKELGALKEKLYALKE (SEQ ID NO:101)

KLYALKEKLGALKEKLACLEKELGALEKELYALEK (SEQ ID NO:102)

In addition, many metal ion binding peptide motifs (Nivorozhkin et al. 2000. Inorg. Chem. 39(11) 2306-2313)

having a circular structure, such as Cyclo[K 1,12] (QCGVCGKCIACK), may also be used in the present invention.

Cysteine-containing metal ion binding motifs that may be used in the modified antibody of the present invention are summarized in Table 1 below.

Table 1: Cysteine-containing metal ion binding motif structures that may be used in the present invention

|                                          |                                                             |                                     | Motif structures                                                                                                                                |
|------------------------------------------|-------------------------------------------------------------|-------------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------|
| Natural protein-derived metal ion motif  | Zinc finger                                                 | C <sub>2</sub> H <sub>2</sub> group | Cys-X <sub>2-4</sub> -Cys-X <sub>12</sub> -His-X <sub>3-5</sub> -His                                                                            |
|                                          |                                                             | C <sub>4</sub> group                | Cys-X <sub>2</sub> -Cys-X <sub>n</sub> -Cys-X <sub>2</sub> -Cys-X <sub>n</sub> -Cys-X <sub>2</sub> -Cys-X <sub>n</sub> -Cys-X <sub>2</sub> -Cys |
|                                          |                                                             | C <sub>6</sub> group                | Cys-X <sub>2</sub> -Cys-X <sub>6</sub> -Cys-X <sub>5-12</sub> -Cys-X <sub>2</sub> -Cys-X <sub>6-8</sub> -Cys                                    |
|                                          | transcription regulatory proteins, metal chaperone proteins |                                     | Cys-X <sub>n</sub> -Cys                                                                                                                         |
|                                          | membrane proteins                                           |                                     | HXXWFYLYX <sub>21</sub> - <sub>28</sub> CXLFMVIGXWFLVIX <sub>18-27</sub> HXXH (x is any amino acid)                                             |
|                                          | metal ion transporters                                      |                                     | M-X-C-X-X-C                                                                                                                                     |
| Artificially synthesized metal ion motif | zinc finger motif                                           |                                     | PYKCPECGKSFSQKSALVKHQRTTH                                                                                                                       |
|                                          |                                                             |                                     | modified peptide<br>ACGHA                                                                                                                       |

|  |                           |            |                                                                       |
|--|---------------------------|------------|-----------------------------------------------------------------------|
|  | multiple-stranded-helices | TRI family | Modified peptide of (LKALEEK) <sub>n</sub> (n=1 to 5)                 |
|  |                           | Others     | Cyclo(K 1,12)-QCGVCGKCIACK<br>ELYALEKELGALEKELACLEKELGALE<br>KELYALEK |

Such cysteine-containing motifs may be bound to the N-terminus or C-terminus of the light or heavy chain of a parent antibody without limitation, so long as a target drug  
5 can be conjugated thereto while maintaining the specificity of the parent antibody. The motif is preferably bound to the C-terminus of the heavy or light chain. Particularly, it is bound to the C-terminus of the heavy chain, that is, the terminus of the constant region of the parent antibody. If  
10 an antibody fragment is used, the motif is preferably bound to the C-terminus of the heavy chain of the fragment.

However, as long as the specificity of the parent antibody is maintained, the cysteine-containing motif in the present invention, specifically cysteine motif having a  
15 metal ion-binding functionality or certain specific functionality or a secondary or tertiary structure, could be linked to any position other than the heavy and light chain of the parent antibody and may be introduced into the parent antibody by a long-chain peptide linker. It is well known

that, when an antibody-drug conjugate is produced by conjugating drugs to site-specifically mutated cysteine or lysine residues in either heavy or light chains (except CDR region) of antibody through a long-chain hydrocarbon linker, it still can retain the structural characteristics and specificity of the parent antibody. Thus, a cysteine-containing metal ion binding peptide motif linked to this position of the parent antibody by a long-chain peptide linker can provide an antibody-drug conjugate that has high homogeneity while retaining the specificity of the parent antibody.

In the modified antibody according to the present invention, the cysteine-containing motif can be fused directly to the parent antibody by an amide bond. Alternatively, the terminal functional group of the cysteine-containing motif can be chemically bound to the terminal functional group of the parent antibody. Alternatively, the motif can also be bound to the parent antibody in a linker mediated manner by using a first linker.

A drug can be conjugated directly to the cysteine residue and the thiol group (-SH) of the cysteine in the cysteine-containing motif bound to the parent antibody and may also be linked to the cysteine residue through a second linker.

Binding of the second linker that links a drug to the cysteine residue in the cysteine-containing motif may be performed by using alkylation, disulfide exchange or trans-thioesterification. The second linker may be one or more  
5 selected from among alkyl halide derivatives containing a haloacetyl functional group, derivatives containing a maleimide group, aziridine derivatives, acryloyl derivatives, and aryl halide derivatives containing fluorobenzen or the like, but is not limited thereto. The derivatives may be  
10 bound to the thiol group of the cysteine residue in the motif by an alkylating reactive group, an arylating reactive group, a maleimide group, an aziridine group, an acryloyl group, or a disulfide exchange reactive group containing pyridyl disulfide and thionitrobenzoic acid (Bioconjugate techniques,  
15 2nd edition, pp 182 ~ 192, Gerg T.Hermanson, ELSVIER).

For example, a maleimide group that is generally used for linking thiol and a linker is used to conjugate a drug to cysteine, because the nucleophilic reactivity of the thiol group of a cysteine residue for the maleimide group is about  
20 1000 times higher than that of the amine group or N-terminal amine group of other amino acid residue, such as lysine residues. Thus in the case of a modified antibody-drug conjugate comprising maleimide or iodoacetamide as the second linker, cysteine is bound to the drug by a thioether bond.

Generally, the second linker has a reactive site having an electrophilic group that reacts with nucleophilic cysteine present on the antibody.

A drug that is bound to the modified antibody of the present invention may be any drug having disease therapeutic effects. Particularly, it is preferably a cancer therapeutic drug having the effect of inhibiting the proliferation of tumor cells.

Specifically, a drug that may be used in the modified antibody-drug conjugate of the present invention comprises any compound, moiety or group that has the effect of cytotoxicity or inhibiting cell proliferation, and examples thereof include:

(i) chemotherapeutic agents capable of functioning as microtubulin inhibitors, mitotic inhibitors, topoisomerase inhibitors, or DNA intercalators;

(ii) protein toxins capable of functioning as enzymes;

(iii) micro-RNA (miRNA), siRNA, or shRNA, which can inhibit the expression of a specific oncogene; and

(iv) radioisotopes.

Examples of such drugs include, but are not limited to, maytansinoid, auristatin, dolastatin, trichothecene, CC1065 (cytotoxic compound), calicheamicin and other enediyne antibiotics, taxane, anthracycline, methotrexate, adriamycin,

vindesine, vinca alkaloids (vincristine, vinblastine, etoposide), doxorubicin, melphalan, mitomycin C, chlorambucil, daunorubicin, daunomycin, and stereoisomers, isosteres, analogs or derivatives thereof, other intercalating agents, 5 enzymes and fragments thereof such as nucleolytic enzymes, antibiotics, and toxins such as small molecule toxins or enzymatically active toxins of bacterial, fungal, plant or animal origin, and various antitumor or anticancer agents such as cisplatin, CPT-11, doxorubicin, paclitaxel and 10 docetaxel.

In addition, a nucleophilic group on the drug moiety in the present invention includes one or more selected from amine, thiol, hydroxyl, hydrazide, oxime, hydrazine, thiosemicarbazone, hydrazine carboxylate and aryl hydrazide 15 groups, which can react with the second linker moiety and an electrophilic group on the second linker reagent to form a covalent bond, but is not limited thereto.

As described above, the number of drug molecules that can be conjugated to the antibody by the second linker moiety 20 can increase as the number of cysteine residues in the motif bound to the parent antibody increases.

As described above, the modified antibody according to the present invention can comprise a number of cysteine residues, because the motif containing one or more cysteine



residues is bound to the parent antibody, preferably the C-terminus of the parent antibody. Thus, a large amount of a drug can be conjugated to the modified antibody in the present invention, and a large amount of the conjugated drug  
5 can be effectively delivered to a target cell or tissue.

Also, because the number of cysteine residues that are bound to the parent antibody is easily controlled, the amount of a drug that is contained in the modified antibody-drug conjugate (mADC) can be easily controlled to a desired level.  
10 This characteristic feature overcomes the shortcomings of conventional modified antibody-drug conjugates, and the novel modified antibody-drug conjugate according to the present invention can be used as an excellent drug delivery system and can be used efficiently for the treatment of diseases  
15 such as cancer.

The present invention also provides a method for producing a modified antibody, which comprises a motif containing one or more cysteine residues bound to a parent antibody.

20 The parent antibody can be produced by constructing a suitable expression vector containing a nucleotide sequence encoding the heavy-chain or light-chain constant domain of the parent antibody, transforming a prokaryotic or eukaryotic cell with the constructed expression vector to express an

antibody protein, isolating the antibody protein, and then purifying the isolated antibody protein to a pharmaceutically acceptable purity.

A suitable expression vector in the present invention  
5 may include expression regulatory elements, such as a promoter, an initiation codon, a stop codon, a polyadenylation signal, and an enhancer, as well as signal sequences for membrane targeting or secretion. Examples of promoters that are generally available in prokaryotic cells  
10 include lac, tac, T3 and T7 promoters. Examples of promoters available in eukaryotic cells include simian virus 40 (SV40) promoter, mouse mammary tumor virus (MMTV) promoter, human immunodeficiency virus (HIV) promoter such as the HIV Long Terminal Repeat (LTR) promoter, moloney virus promoter,  
15 cytomegalovirus (CMV) promoter, epstein barr virus (EBV) promoter, rous sarcoma virus (RSV) promoter, as well as  $\beta$ -actin promoter, promoters from human genes such as human, human hemoglobin, human muscle creatine and human metallothionein.

20 An expression vector may include a selectable marker that allows selection of host cells containing the vector. Markers that confer selectable phenotypes, such as resistance to drugs, nutrient requirement, resistance to cytotoxic agents or expression of surface proteins, are used as

selectable markers. Since only cells expressing a selectable marker survive in the environment treated with a selective agent, transformed cells can be selected. Also, a replicable expression vector may include a replication origin with a specific nucleic acid sequence which initiates replication.

As recombinant expression vectors for expressing exogenous genes, various types of vectors such as plasmid, virus or cosmid vectors may be used. The type of recombinant vector is not specifically limited, as long as it functions to express a desired gene in various host cells such as prokaryotic and eukaryotic cells and to produce a desired protein. However, it is preferable to use a vector that includes a promoter exhibiting strong activity and can produce exogenous protein, which retains similarity to a naturally occurring protein, with high yield.

A variety of expression host/vector combinations may be used to express a parent antibody or a modified antibody comprising a motif containing one or more cysteine residues. Useful expression vectors for eukaryotic hosts include, but are not limited to, vectors comprising expression control sequences from SV40, bovine papilloma virus, adenovirus, adeno-associated virus, cytomegalovirus and retrovirus. Useful expression vectors for bacterial hosts include bacterial plasmids, such as plasmids from *E. coli*, including

pET, pRSET, pBluescript, pGEX2T, pUC, col E1, pCR1, pBR322, pMB9 and their derivatives, wider host range plasmids, such as RP4, phage DNAs, e.g., the numerous derivatives of phage lambda, e.g.  $\lambda$ gt10,  $\lambda$ gt11, NM989, and other DNA phages, such as M13 and filamentous single stranded DNA phages. Useful expression vectors for yeast cells include the 2  $\mu$ m plasmid and derivatives thereof. Useful vectors for insect cells include pVL 941.

In another aspect, the present invention provides a host cell transformed with the above-described recombinant vector. The recombinant vector is inserted into a host cell to form a transformant. A suitable host cell to be transformed with the vector may be a prokaryotic cell such as *E. coli*, *Bacillus subtilis*, *Streptomyces* sp., *Pseudomonas* sp., *Proteus mirabilis* or *Staphylococcus* sp. Also, the host cells may be fungal cells such as *Aspergillus* sp., yeast cells such as *Pichia pastoris*, *Saccharomyces cerevisiae*, *Schizosaccharomyces* sp. and *Neurospora crassa*, lower eukaryotic cells, and higher eukaryotic cells such as insect cells. Also, the host cells may be derived from plants and/or mammals. Preferred examples of the host cells include, but are not limited to, PER.C6 cells, monkey kidney cells 7 (COS7; particularly simian COS cells), NSO cells, SP2/0, Chinese hamster ovary (CHO) cells, W138, baby hamster kidney

(BHK) cells, Madin-Darby canine kidney (MDCK) cells, myeloma cell lines, HuT 78 cells, HEK293 cells, and other mammalian host cells that produces the antibody protein according to the present invention.

5        Particularly, to maximize expression efficiency, the host cell in the present invention is preferably one or more selected from among *E. coli* cells, simian COS cells, Chinese hamster ovary (CHO) cells, and other mammalian host cells that produces the antibody protein according to the present  
10 invention. More preferably, the host cell in the present invention is the hamster ovary cell CHO-K1.

      In the present invention, "transformation" into host cells include any method for introducing nucleic acids into organisms, cells, tissues, or organs and may be performed  
15 using a standard technique selected depending on the type of host cell as known in the art. Examples of this method include, but are not limited to, electroporation, protoplast fusion, calcium phosphate ( $\text{CaPO}_4$ ) precipitation, calcium chloride ( $\text{CaCl}_2$ ) precipitation, agitation using silicon  
20 carbide fiber, agrobacterium-mediated transformation, and PEG-, dextran sulfate-, lipofectamine- or desiccation/inhibition-mediated transformation.

      The antibody protein expressed as described above can be recovered from the host cell, the supernatant of the culture,

or the cells after lysis, and purified by using a conventional protein purification technique, to yield the modified antibody comprising a motif containing one or more cysteine residues.

5       The antibody can be isolated from the culture medium by a conventional immunoglobulin purification procedure, for example, protein A Sepharose™, hydroxyl-apatite chromatography, gel electrophoresis, dialysis, or affinity chromatography.

      Nucleotide sequence encoding the cysteine containing  
10 motif can be fused to a nucleotide sequence encoding either heavy-chain constant domain or light-chain constant domain of parent antibody. Recombinant vector fused in this way can produce modified antibody with the similar method employed in parent antibody after transformed and expressed in  
15 prokaryotic or eukaryotic host cell.

      In addition, the modified antibody of the present invention can be produced by expressing each of a parent antibody and a motif containing one or more cysteine residues, and then chemically linking the terminal functional group of  
20 the parent antibody to the terminal functional group of the cysteine-containing motif or linking the parent antibody to the motif by a first linker. A drug may be bound to the produced modified antibody with the cysteine-containing motif, thereby producing a modified antibody-drug conjugate (mADC).

Specifically, a modified antibody-drug conjugate with multiple cysteines-containing motif can be produced by:

(a) conjugating a parent antibody containing a cysteine-containing motif with a linker reagent to form an covalently bound antibody-second linker intermediate, and then add an activated drug moiety to the intermediate; or

(b) conjugating the nucleophilic group of a drug moiety with a second linker reagent to form a covalently bound drug-second linker intermediate, and then add a modified antibody with a cysteine-containing motif.

In another aspect, the present invention provides a therapeutic composition comprising the modified antibody-drug conjugate as an active ingredient.

In the above mentioned composition, the drug conjugated to a modified antibody-drug conjugate may be a cytotoxic agent, a cell proliferation inhibitor, a chemotherapeutic agent, an immune inhibitor, an anti-inflammatory agent or the like, but is not limited thereto. In cancer therapy, the use of the antibody-drug conjugate for local delivery of a drug that kills or inhibits tumor cells allows the targeted delivery of the drug moiety into tumor cells by antibody-antigen interactions and the intracellular accumulation of the drug moiety. Administration of non-conjugated drug formulation may cause an unacceptable level of toxicity not

only in tumor cells, but also in normal cells. However, the modified antibody-drug conjugate according to the present invention can accurately deliver the drug due to the high antigen specificity of the antibody, increasing the therapeutic effect of the drug. In addition, it can expand the usability of drugs, particularly anticancer agents, the use of which is limited due to their toxicity, despite their high anticancer efficacy. According to the present invention, an antibody-drug conjugate that maximizes drug efficacy while minimizing drug toxicity is provided.

The present invention also provides a method of inhibiting the proliferation of target cells from cancer, autoimmune, inflammatory or infectious disease by the modified antibody-drug conjugate as an active ingredient .

Cancer that can be treated according to the present invention may be one or more selected from among liver cancer, stomach cancer, breast cancer, colon cancer, bone cancer, pancreatic cancer, head and neck cancer, uterine cancer, ovarian cancer, rectal cancer, esophageal cancer, small intestine cancer, anal cancer, fallopian tube cancer, endometrial cancer, cervical cancer, vaginal cancer, valve cancer, Hodgkin's disease, prostate cancer, bladder cancer, kidney cancer, ureter cancer, renal cell carcinoma, renal pelvis cancer, and cancer of the central nervous system. In



a specific example, proliferation of HER2-amplified breast cancer BT-474 cells can be inhibited by bringing the modified antibody-drug conjugate into contact with the cells *in vitro*. Therefore, it is evident that the inventive method of  
5 inhibiting the proliferation of target cells using the modified antibody-drug conjugate as an active ingredient has the effect of killing cells related to the above-described disease or reducing the proliferation rate of the cells.

Hereinafter, the present invention will be described in  
10 further detail with reference to examples. It will be obvious to a person having ordinary skill in the art that these examples are not to be construed to limit the scope of the present invention, and various modifications and changes can be made within the technical idea and scope of the  
15 present invention.

Unless otherwise defined, the technical or scientific terms as used herein have the same meanings as understood by those having ordinary knowledge in the technical field to which the present invention pertains. Also, detailed  
20 description of the same construction and effect as those of the prior art will be omitted herein.

## **1. Analytical method**

### **1.1: UV-VIS spectrophotometry**

In the present invention, an antibody-drug conjugate was produced by using a drug (DOX-EMCH) with a linker attached to the anticancer agent doxorubicin (6-maleimidocaproyl)hydrazine. The DOX-EMCH, when injected into  
5 a human body, binds to the thiol group of cysteine present in blood albumin to form an albumin-drug conjugate [Willner et al., Bioconjugate Chem. 1993 (4):521-7] and can be easily applied in describing the present invention.

In order to examine whether a drug is still bound to a  
10 protein after conjugation and purification steps, UV-VIS absorption spectroscopy is usually employed. Protein shows the maximum absorbance at a wavelength of 280 nm (UV wavelength range), and doxorubicin used in the present invention has the maximum absorbance at 495 nm (visible  
15 wavelength range). Both protein and doxorubicin have characteristic absorption coefficients, respectively, at specific wavelength region. Therefore when the absorbance is measured at 280 nm and 495 nm , the equivalent of drug bound per molecule of protein is determined (US patent No.  
20 7,528,234 B2).

### **1.2: In vitro cell proliferation inhibition assay**

The cytotoxicity or cell proliferation inhibitory activity of the modified antibody-drug conjugate (mADC) was

determined by exposing mammalian cells having receptor protein, for example, SK-BR-3 or BT-474 cells, to the modified antibody-drug conjugate (mADC) in cell culture medium. The mADC-treated cells were cultured for about 6  
5 hours ~ 5 days, and measured its viability.

### **1.3: Measurement of *in vitro* caspase 3/7 activity**

Herceptin does not directly induce cancer cell death, but does induce the death of Her2-positive cancer cells by  
10 ADCC (antibody-dependent cellular cytotoxicity). However, a drug therapy directly induces apoptotic cell death, therefore when the caspase activity is measured, it could be determined whether the modified antibody-drug conjugate (mADC) induces caspase-mediated apoptosis or not (Bayascas, et al. (2002),  
15 Cell Death and Differentiation. 9: 1078-1089; Preaudat, et al (2002), Journal of Biomolecular Screening. 7: 267-274; Phillips, et al. (2008), Cancer Research 68(22): 9280-9290).

In the present invention, in order to examine the mechanism of apoptosis by the modified antibody-drug  
20 conjugate (mADC), the activity of caspase 3/7 was measured. Generally, apoptosis by the modified antibody-drug conjugate (mADC) is determined by exposing mammalian cells having receptor protein, for example, SK-BR-3 cells or BT-474 cells, to the modified antibody-drug conjugate (mADC). The mADC-

treated cells were cultured in medium for ~ 2 days, and was measured its caspase activity.

## **2. Examples**

### 5      Example 1. Preparation of expression vector pAV4

Cloning of the expression vector required in the present invention was performed by using a pAV4 vector which was constructed and modified from the parent vector pSGHV0 (GenBank Accession No. AF285183), so as to be used for  
10 antibody production in the industrial field. Although human protein could be overexpressed in bacterial cells such as *E. coli*, it is difficult to obtain as an active substance. Therefore the parent vector was manufactured to express physiologically active protein in high concentration in  
15 animal cells in vitro, together with easy purification steps. Albeit its biggest merit of high protein expression level from this parent vector, there is a limitation for its use in the industrial field. Therefore the pAV4 vector was modified from its parent vector to be adaptable in the industrial  
20 field. In addition, the pAV4 vector is modified adequately for the purpose of expressing both the heavy-chain and light-chain domains of antibody.

### Example 2: Construction of vectors for the production of

trastuzumab and cysteine modified trastuzumab

To construct a trastuzumab (HHL002) vector, the cDNAs of the heavy chain and light chain of trastuzumab were synthesized into codon-optimized sequences so that the expression thereof in CHO cells is maximized. The genes were respectively cloned into the XhoI/NotI and ApaI/SmaI restriction sites of the pAV4 vector, thereby a trastuzumab vector (pHHL002) was prepared.

10        2.1: Construction of vectors for the modified  
trastuzumab antibodies HR-Cys and HR-Cys-Gly-Cys

In order to construct vectors for modified trastuzumab antibodies, HR-Cys (HHL002C), a single cysteine mutant in replace of terminal lysine at the C-terminus and another trastuzumab variant HR-Cys-Gly-Cys (HHL002C2) with a C-terminal peptide consisting of Cys-Gly-Cys (CGC), PCR amplification was performed using the constructed trastuzumab vector (pHHL002) as a template. Specifically, PCR amplification was performed using trastuzumab as a template and a XhoHH forward primer (5'-GGG GGG CTC GAG ACC ATG GGT TGG AGC TGT -3') and an HHNot reverse primer (5'-GCG GCC GGC CGC TCA ACA ACC CGG AGA CAG -3') for HHL002C or an HHNot reverse primer (5'-GCG GCC GGC CGC TCA ACA GCC ACA ACC CGG AGA CAG-3') for HHL002C2. The amplified nucleotide sequence

was cleaved with the restriction enzymes XhoI and NotI and ligated with the expression vector pHHL002 having XhoI/NotI cleavage sites, thereby constructing cysteine modified trastuzumab antibody vectors (pHHL002C; pHHL002C2). FIG. 1  
 5 is a schematic view of the vector. The cysteine-containing variant antibody produced in this Example is a cysteine-containing trastuzumab antibody containing a deletion of lysine at the C-terminus and an addition of single cysteine or a short peptide of cysteine, glycine and cysteine.

10

## 2.2: Construction of a vector for the modified trastuzumab antibody HR-M2(Cys)

In order to construct a vector encoding a modified trastuzumab antibody HR-M2(Cys) (HR-ACHGAACGHA; HHL002M2)  
 15 which has two metal ion binding motifs (CGH), PCR amplification was performed by using the trastuzumab vector (pHHL002) as a template and a XhoHH forward primer (5'-GGG GGG CTC GAG ACC ATG GGT TGG AGC TGT -3') and an M2 reverse primer (5' CCCCCG C GGC CGC CTA GGC ATG GCC ACA AGC AGC ATG GCC  
 20 ACA GGC GCC GGG AGA CAG AGA 3'). The amplified nucleotide sequence was cleaved with the restriction enzymes XhoI and NotI and ligated with the expression vector pHHL002 having XhoI/NotI cleavage sites, thereby constructing a cysteine modified trastuzumab antibody vector (pHHL002M2).

### 2.3: Production of modified trastuzumab antibody HR-M(Cys)

In order to construct a vector encoding modified  
 5 trastuzumab antibody HR-M(Cys) (HR-GGGACGHA; HHL002M) which  
 has a single metal ion binding motif (CGH), PCR amplification  
 was performed by using an EzChange Site-directed mutagenesis  
 kit (Enzymomics, Ez004S). The above-produced modified  
 trastuzumab antibody HR-M2(Cys) was used as a template with a  
 10 forward primer (5'-GGT GGA GGT GCT TGT GGC CAT TAA GC) and a  
 reverse primer (3'-GCC GGG AGA CAG AGA CAG TG). From the two  
 metal ion binding motifs in HR-M2(Cys), the one metal ion  
 binding motif from the C-terminus was removed and a glycine  
 linker(GGG) was added between the remaining motif and the  
 15 parent antibody. After EzChange pcr amplification, the  
 original template HR-M2(Cys) was cleaved with the restriction  
 enzyme DpnI , and the resulting 5-M(Cys) was ligated into a  
 double-stranded DNA vector by ligase, thereby constructing a  
 modified trastuzumab antibody vector (pHHL002M).

20

### 2.4: Construction of a vector for modified trastuzumab antibody HR-M2L(Cys)

In order to construct a vector encoding modified  
 trastuzumab antibody HR-M2L(Cys) (HR-ACGHAGGGACGHA,

HHL002M2L) with an three amino acid linker (GGG) inserted between metal ion binding motifs (CGH), PCR amplification was employed. The above-produced modified vector for trastuzumab antibody HR-M2(Cys) was used as a template with a forward  
 5 primer (5'-GGT GGA GGTGCT TGT GGC CAT GCC TAA GCG) and a reverse primer (3'-AGC ATG GCC ACA GGC GCC). After the original template HR-M2(Cys) was cleaved with the restriction enzyme DpnI, the resulting nucleotide encoding 5-M2L(Cys) was ligated into a double-stranded DNA vector by ligase, thereby  
 10 constructing a modified trastuzumab antibody vector (pHHL002M2L).

#### 2.5: Construction of a vetor for modified trastuzumab antibody HR-Z(Cys)

15 In order to construct a vector encoding HR-Z(Cys) (HR-CDICGRKFARSDERKRHTKIHLRQK, HHL002Z) which has a metal ion binding motif from Class I zinc finger protein, PCR amplification was performed using the trastuzumab vector (pHHL002) as a template with a XhoHH forward primer (5'-GGG  
 20 GGG CTC GAG ACC ATG GGT TGG AGC TGT -3', SEQ ID NO: 1) and a Z reverse primer (5'-GCA TGC GGC CGC CTT ACT TCT GCC GCA GGT GGA TCT TGG TAT GCC TTT TTC GCT CGT CGG ATC TAG CAA ATT TGC GTC CAC AAA TAT CGC ATT TGC CGG GAG ACA GAG A-3'). The amplified nucleotide sequence was cleaved with the



restriction enzymes XhoI and NotI and ligated with the expression vector pHHL002 having XhoI/NotI cleavage sites, thereby constructing a cysteine modified trastuzumab antibody (pHHL002Z).

5

Example 3: Expression and purification of trastuzumab and cysteine modified trastuzumab antibody

By using Chinese hamster ovary cells (CHO-K1), the expression of trastuzumab (HHL002) and its cysteine-modified antibodies (HHL002C, HHL002C2, HHL002M, HHL002M2, HHL002M2L, HHL002Z) were analyzed. Specifically, CHO-K1 cells were cultured in DMEM (Dulbecco's Modified Eagle Media) containing 10% FBS (fetal bovine serum) and antibiotic. The CHO-K1 cells were inoculated in a 100mm culture dish at a concentration of 5x 10<sup>6</sup> cells/ml and then cultured for 24 hour. 800 µl of FBS- and antibiotics-free DMEM was mixed with 10 µg vector and incubated at room temperature for 1 minutes, after which the mixture was mixed with 20 µg PEI (Polyethylenimine, linear, Polysciences Inc (Cat. no: 23966, MW~25,000)) and allowed to stand at room temperature for about 10-15 minutes. Meanwhile, the cells were washed with PBS and 6 ml of fresh DMEM was added thereto. The trastuzumab or cysteine-modified trastuzumab antibody vector maintained at room temperature for 10-15 minutes was added to the culture dish. On the next

day, the cells were washed with PBS, and FBS-free IMDM medium (Cat. No 12200-028, Gibco, Iscove's Modified Dulbecco's Medium) was added thereto to confirm the expression of the antibody protein.

5        The trastuzumab and cysteine modified trastuzumab antibodies expressed as described above were purified in the following manner. Specifically, in order to purify the trastuzumab and cysteine modified trastuzumab antibodies secreted into the cell culture medium, the culture medium was  
10    centrifuged to remove cells, and the supernatant was injected into HiTrap Protein A HP column (GE Healthcare, USA) which was equilibrated with equilibration buffer. The column was sufficiently washed with equilibration buffer and then the protein was eluted by pH change with glycine buffer (100 mM  
15    Glycine, pH 2.8). The resulting solution was dialyzed against phosphate buffer, and then concentrated by using Vivaspin20 (Sartorius, USA).

20    Example 4: Production of a variant antibody and drug conjugate

4.1: Production of a trastuzumab-based variant antibody and drug (doxorubicin) conjugate

      In general, if the cysteine residues of proteins do not participate in a intramolecular disulfide bond, the proteins

usually form a dimer by intermolecular disulfide bond. However, as shown in FIG. 2, HR-Cys produced in the present invention was present as a whole monomeric antibody as determined by SDS-PAGE and Western blot analysis. In  
5 addition, a cysteine residue having a free thiol group in the protein can form a disulfide bond with intracellular glutathione or cysteine. For this reason, in order to break the disulfide bond, a step of treatment with a reducing agent, such as DTT (dithiothreitol) or TCEP (tris(2-carboxyethyl)  
10 phosphine), is required upon binding with a drug-second linker, but a sufficient binding reaction can also be achieved even by agitation at room temperature.

The second linker-conjugated drug used in this experiment is a (6-maleimidocaproyl)hydrazone derivative of  
15 doxorubicin known as DOXO-EMCH. A method of attaching a compound such as DOXO-EMCH, which has a maleimide group, to the thiol group of protein, is well described in the literature [Klussman, et al. (2004), Bioconjugate Chemistry  
15(4): 765-773, page 766; Emmanuel et al. (2010) Chemistry &  
20 Biology 2010(17):213-227].

HR-Cys purified in the present invention was mixed with DOXO-EMCH at a molar ratio of 1:10 and agitated at room temperature for 4 hours. Then, the remaining DOXO-EMCH was

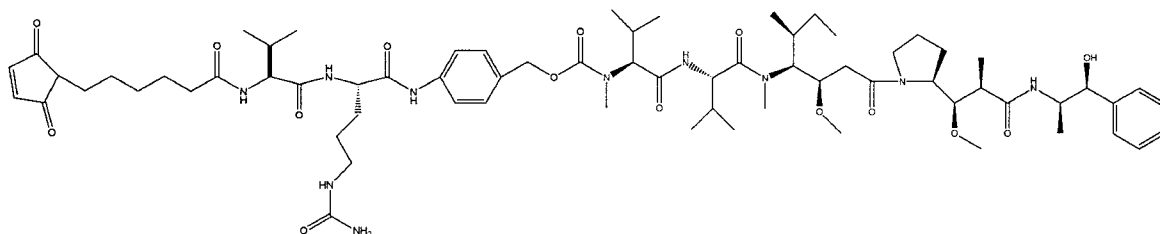
removed using a desalting column, and the remaining material was washed three times by ultrafiltration, thereby producing a HR-Cys-doxorubicin conjugate (HR-Cys-DOX). The produced conjugate was confirmed by UV-VIS spectrophotometry as shown in FIG. 3. From calculation based on the characteristic absorption coefficients of the protein and the drug, it could be seen that about two molecules of DOXO-EMCH were bound per HR-Cys molecule.

10        4.2: Production of a trastuzumab-based modified antibody and drug(MMAE) conjugate

         In the present invention, MMAE known to have cytotoxicity much higher than doxorubicin was conjugated to HR-Cys-Gly-Cys to make a Herceptin-MMAE conjugate (HR-Cys2-MMAE). Monomethyl Auristatin E (see Chemical Formula 2 below), a conjugatable derivative of Auristatin, is linked to the thiol-specific maleimide group through a valine-citurulline, which could be degraded by protease in cells, and a self-degrading spacer, para-aniline benzoic acid (PABA). This is known as MC (maleimido caproic acid)-VC(valine-citurulline)-PAB-MMAE, and a synthesis method thereof is already well known (US patent No. 6214345; US patent No. 7745394). Auristatin, a highly cytotoxic compound, is known to have an

IC50 value of 200-300 pM in a cell proliferation inhibition assay.

Chemical Formula 2: MC-vc-PAB-Methyl Auristatin E



5

In the present invention, 2-10 equivalents of the reducing agent TCEP was added to the modified trastuzumab antibody and reacted at 4 °C for 30 minutes to reduce the thiol group, after which 2-10 equivalents of MC-vc-PAB-MMAE was added thereto and reacted at room temperature for about 2-4 hours. The reaction was terminated by addition of an excess amount of cysteine, and unreacted MC-vc-PAB-MMAE and TCEP were removed by centrifugation-filtration and dialysis in phosphate buffered saline, thereby obtaining a purified trastuzumab variant-MC-vc-PAB-MMAE.

In order to confirm the selective binding specificity of the produced trastuzumab variant-MC-vc-PAB-MMAE, the peptide mapping of the produced HR-M2(Cys)-MC-vc-PAB-MMAE was performed (ProteinWorks Co., Ltd., Daejeon, Korea). The

20

results of the peptide mapping indicated that MC-vc-PAB-MMAE was introduced into the peptide cysteine residue that is a drug binding site in the C-terminus of heavy chain. Binding of the drug to the cysteine derived from the intrachain or interchain disulfide bond of the heavy or light chain was not observed. This suggests that the antibody variant designed in the present invention was effectively synthesized, which can significantly increase the homogeneity of the antibody-drug conjugate with the drug bound specifically to the C-terminus of the heavy chain.

4.3: Analysis of selective conjugation property of modified trastuzumab antibody by using a maleimide-containing fluorescent dye, Alexa Fluor® 488.

In order to examine whether a drug is selectively conjugated to a cysteine residue introduced into the C-terminus of Trastuzumab heavy chain, fluorescent dye Alexa Fluor® 488 substituted with a thiol-specific maleimide group was reacted with each of the modified trastuzumab antibodies. When the cysteine residues of proteins do not form an intramolecular disulfide bond, the proteins usually form a dimer by an intermolecular disulfide bond. However, as shown in FIG. 2, HR-Cys produced in the present invention was present as a whole monomeric antibody as determined by SDS-

PAGE and Western blot analysis. In addition, a cysteine residue having a free thiol group can form a disulfide bond with intracellular glutathione or cysteine. For this reason, in order to break the disulfide bond, a step of treatment  
5 with a reducing agent, such as DTT (dithiothreitol) or TCEP (tris(2-carboxyethyl) phosphine), is required upon binding with a drug-second linker, but a sufficient binding reaction can also be achieved even by agitation at room temperature. 2-4 equivalents of the reducing agent can reduce not only a  
10 cysteine residue introduced into the C-terminus of the heavy chain, but also the intrachain disulfide bond of each of the heavy and light chains and the interchain disulfide bond that links the heavy chain to the light chain. Thus, fluorescent dye can be bound to any cysteine residue reduced by the  
15 reducing agent. A drug that is bound to an interchain or intrachain disulfide can reduce the homogeneity of the antibody-drug conjugate, and the bound drug having a relatively large size is highly likely to reduce the structural stability and antigen specificity of the antibody  
20 protein itself, and for this reason, the efficacy thereof in the manufacture of biomedicines can be significantly reduced.

2-4 equivalents of TCEP is added to the modified trastuzumab antibody and reacted at 4 °C for about 30 minutes to reduce the thiol group. In this process, either the

antibody variant present as a dimer with a disulfide bond or the thiol functional group of cysteine which is present in an oxidant-bound form during an expression process is restored to a reduced form that can bind to a maleimide group. After  
5 the reduction reaction, the reducing agent such as DTT, which is reactive to the maleimide group, should be removed by, for example, centrifugation-filtration, but TCEP, which is not involved in the conjugation reaction between the maleimide group and the thiol group, is not necessarily removed. 2-10  
10 equivalents of Alexa Fluor® 488 is added to the modified trastuzumab antibody and reacted with stirring at room temperature for about 2-4 hours. The reaction is terminated by addition of an excess amount of cysteine, and unreacted dye and reducing agent are removed by centrifugation-filtration.  
15 The resulting material is dialyzed against phosphate buffered saline, thereby obtaining a modified trastuzumab antibody-Alexa488 dye conjugate.

The modified trastuzumab antibody-Alexa488 dye conjugate obtained as described above was separated by SDS-PAGE gel  
20 electrophoresis, and then the amount of the dye bound to each of the heavy and light chains was analyzed by a fluorescence image analyzer (Fluorescencen Image Ananlyzer, Typoon9410, Amersharm Bioscience Ltd.). The results of the fluorescence image analysis indicated that, in the case of the HR-Cys and



HR-Cys-Gly-Cys antibody variants, the amount of the dye bound to the heavy chain was larger than that of the dye bound to the light chain. Also in the case of HR-M(Cys), HR-M2(Cys), HR-M2L(Cys) and HR-Z(Cys), the amount of the dye bound to the heavy chain was larger than the amount of the dye bound to the light chain. As can be seen in FIG. 4, in the case of the HR-M2(Cys) antibody variant, the intensity of the fluorescent dye bound to the heavy chain was about 6 times higher than the intensity of the dye bound to the light chain. This difference in fluorescence intensity indicates that the amount of the dye bound to the heavy chain is much larger than the amount of the dye bound to the light chain, suggesting that the selectivity of the maleimide group to the heavy chain is significantly greater.

15

Example 5: Measurement of metal ion binding affinity of metal ion binding motif

As mentioned above, metal ions bind to a peptide having the metal ion binding motif CGH and inhibit or retard the oxidation of the cysteine residue, thereby preventing a sulfonation reaction caused by over-oxidation of the cysteine. Oxidation of the cysteine residue can occur through two reaction pathways. In one reaction pathway, a disulfide bond can be created by thiol-thiol bonding, and in the other

pathway, cysteine can also be oxidized by reaction with an excess of oxygen in air to produce sulfenic acid (R-S-OH) as an intermediate product. In this case, the disulfide formation is a reversible reaction in which the disulfide bond can be reduced to thiol, but the formation of sulfenic acid by reaction with oxygen is an irreversible one. When disulfide and sulfenic acid are exposed to oxygen for a long period of time, they are all oxidized to sulfonic acid (R-SO<sub>3</sub>H) by an irreversible reaction. Because this sulfenic acid or sulfonic acid have no reactivity to maleimide, oxidation to sulfenic or sulfonic acids reduce the reactivity of the antibody variant with maleimide, and thus can greatly influence the conjugation yield and homogeneity of the antibody-drug conjugate.

As described above, it has been reported that a peptide containing the synthetic motif CGH, derived from the metal ion binding motif GGH that is frequently found *in vivo*, binds metal ions and inhibits the sulfonation of cysteine (Van Horn et al. (2003) J. Biol. Inorg. Chem. 8: 601-610). After secreted to extracellular medium during expression process, antibody protein having the metal ion binding peptide motif can bind to a trace amount of metal ions present in the cell culture medium and effectively inhibits the sulfonation of cysteine. In order to measure the metal ion binding ability

of the antibody variant having this metal ion binding peptide motif, the binding affinity was measured by using a Fura-2 (Invitrogen, F-1200), a well-known metal ion chelator. From this measurement, HR-M2(Cys) had a dissociation constant ( $K_d$ ) of about 20 nM, suggesting that it formed a very strong bond with metal ions. In addition, to examine whether the antibody variant having a metal ion binding motif binds to metal ions and protects the cysteine residue, the alkylation rate was measured. It was observed that the antibody variant formed a very strong bond with zinc metal ions, and thus inhibited the cysteine alkylation for up to 24 hours compared to when there was no metal ion. Nickel ions did not bind to the CGH motif as strongly as zinc ions did, suggesting that the effect of inhibiting the cysteine alkylation is lower than that of zinc ions. However, when compared with the buffer without metal ions, nickel ions reduced the alkylation rate of cysteine. When a strong metal ion chelator EDTA was added to remove zinc ions from HR-M2(Cys), it was observed that the alkylation of cysteine occurred quickly, like the case in which there was no extraneously added metal ions.

#### Example 6: In vitro assay for cell proliferation inhibition

##### 6.1: Cell proliferation inhibition assay of a variant

antibody and doxorubicin conjugate

BT-474 cells were diluted with 10% FBS-containing DMEM/F12 medium, and 100  $\mu\text{l}$  of the diluted cell suspension was inoculated into each well of a 96-well plate at a density of 5  $1 \times 10^4$  cells/well. After inoculation, the well plate was incubated in a 5%  $\text{CO}_2$  incubator at 37  $^\circ\text{C}$  for 24 hours. Each of Herceptin, a 1:2 mixture of Herceptin and doxorubicin, HR-Cys-DOX prepared in the above Example, and doxorubicin, was diluted in medium, and then 100  $\mu\text{l}$  of each dilution was added 10 to each well of the plate at various concentrations of 1000 nM, 100 nM, 10 nM, 1 nM and 0.1 nM. Also medium (having no drug) was added to another wells for negative control. After incubation for about 5 days, 20  $\mu\text{l}$  of CellTiter 96-AQueous One Solution reagent [MTS-based assay; measuring cell 15 proliferation by the amount of purple formazan formed by MTS due to the dehydrogenase of viable cells] was added to each well, followed by incubation in an incubator at 37  $^\circ\text{C}$  for 2 hours. 20  $\mu\text{l}$  of 10% SDS solution was added to each well to stop the reaction, followed by sufficient mixing to induce 20 cell lysis. The absorbance of each well was measured by a spectrophotometer, and the viability (%) based on the absorbance is graphically shown in FIG. 5. As a result, it was shown that the cell growth inhibitory effect of HR-Cys-DOX used in low concentrations was similar to those of

Herceptin and the 1:2 mixtures of Herceptin and doxorubicin and HR-Cys-DOX at a high concentration showed cytotoxicity similar to that of doxorubicin. These results indicate that HR-Cys-DOX retains the characteristic function of Herceptin and sufficiently exhibits the characteristic cytotoxicity of doxorubicin, even though the doxorubicin is bound Herceptin by the second linker.

#### 6.2: Cell proliferation inhibition assay of a variant antibody and MMAE conjugate

SK-BR3 cells were diluted in 10% FBS-containing DMEM/F12 medium, and 100  $\mu$ l of the cell diluted suspension was added to each well of a 96-well plate at a density of  $1 \times 10^4$  cells/well. Then, the well plate was incubated in a 5% CO<sub>2</sub> incubator at 37 °C for 24 hours. Each of Herceptin and the Herceptin variant antibody-MC-vc-PAB-MMAE conjugate produced in the above Example was diluted in medium, and then added to each well of the plate at various concentrations of 66.7 nM, 33.3 nM, 6.7 nM, 3.3 nM, 0.67 nM, 0.33 nM, 0.067 nM and 0.0067 nM. In addition, medium (having no drug) was added to a control wells for negative control. After incubation for 5 days, 20  $\mu$ l of CellTiter 96-AQueous One Solution reagent [MTS-based assay; measuring cell proliferation by the amount of purple formazan formed by MTS due to the dehydrogenase of viable

cells] was added to each well, followed by incubation in an incubator at 37 °C for 2 hours. The absorbance of each well at 490 nm was measured by a spectrophotometer to determine the cell viability (%). As a result, all of the HR-Cys-MC-vc-PAB-MMAE, HR-Cys-Gly-Cys-MC-vc-PAB-MMAE, HR-M2(Cys)-MC-vc-PAB-MMAE, HR-M(Cys)-MC-vc-PAB-MMAE, HR-M2L(Cys)-MC-vc-PAB-MMAE and HR-Z(Cys)-MC-vc-PAB-MMAE showed excellent cell proliferation inhibitory ability compared to the parent antibody Herceptin. As shown in FIG. 6, the IC<sub>50</sub> value of HR-M2(Cys)-MC-vc-PAB-MMAE was at least 5 times lower than that of the parent antibody Herceptin (the median value between the highest and the lowest values was taken for the IC<sub>50</sub> for Herceptin because the viability in the case of Herceptin was not lowered to 50% or below). Also, the cell viability at high concentrations was reduced by about 85% for the antibody-drug conjugate, but was reduced only by about 40% for the parent antibody Herceptin. These results indicate that the antibody-drug conjugate has a very high cytotoxicity and a very low IC<sub>50</sub> value compared to the parent antibody.

#### Example 7: In vitro assay of caspase 3/7 activation

SK-BR3 cells were diluted with 10% FBS-containing RPMI 1640 medium, and 100  $\mu$ l of the cell diluted suspension was

added to each well of a 96-well plate at a density of  $1 \times 10^4$  cells/well. Then, the 96-well plate was incubated in a 5% CO<sub>2</sub> incubator at 37 °C for 24 hours. Each of Herceptin and the modified antibody-drug conjugates produced in the above

5 Example was diluted in medium, and then added to each well of the plate at various concentrations of 66.7 nM, 33.3 nM, 6.7 nM, 3.3 nM, 0.67 nM, 0.33 nM, 0.067 nM and 0.0067 nM. Also medium (having no drug) was added to a control well for negative control. After incubation for 48 hours, 100  $\mu$ l of

10 Caspase-Glo 3/7 reagent [Caspase-Glo 3/7 assay; measurement of luminescence resulting from the degradation of a caspase substrate by caspase 3/7 activity formed in cells in which apoptosis is induced by the caspase pathway] was added to each well, followed by incubation at room temperature for 30

15 minutes. Luminescence was measured by a luminometer.

As shown in FIG. 7, in the cells treated with the parent antibody Herceptin, little or no caspase activity was observed, but in the cells treated with HR-Cys2-MMAE, 3/7 activity increased as the concentration of HR-Cys2-MMAE

20 increased. These results suggest that the MMAE drug released from HR-Cys2-MMAE delivered into the cells induces apoptosis through the caspase pathway, unlike Herceptin.

In addition, as shown in FIG. 8, in the cells treated with Herceptin, little or no caspase activity appeared, but

in the cells treated with HR-M2(Cys)-MC-vc-PAB-MMAE, caspase activity increased as the concentration of HR-M2(Cys)-MC-vc-PAB-MMAE increased. These results indicate that the MMAE drug released from HR-M2(Cys)-MC-vc-PAB-MMAE delivered into  
5 the cells induce apoptosis through the caspase pathway, unlike Herceptin.

#### **INDUSTRIAL APPLICABILITY**

As described above, the modified antibody of the present  
10 invention and the modified antibody-drug conjugate comprising the same can accurately deliver a drug to a target cell due to its high specificity to antigen, and thus can increase the therapeutic effect of the drug. In addition, it can increase the usability of drugs, particularly anticancer agents, the  
15 use of which is limited due to their toxicity, despite their high efficacy.



**We Claim:**

1. An antibody-drug conjugate comprising a modified antibody comprising a cysteine-containing motif at the terminus of a parent antibody

wherein the cysteine-containing motif is a metal ion binding motif that contains a cysteine residue,

wherein the metal ion binding motif that contains a cysteine residue comprises one or two CGH or HGC motif(s) with alanine at C-terminus and N-terminus of the motif,

wherein the cysteine-containing motif is bound to the C-terminus of the heavy chain of the antibody, and

wherein the drug is conjugated to the thiol group of the cysteine residue in the metal ion binding motif.

2. The antibody-drug conjugate of claim 1, wherein the parent antibody is one or more of a monoclonal antibody, a bispecific antibody, a chimeric antibody, a human antibody, or a humanized antibody.

3. The antibody-drug conjugate of claim 1, wherein the parent antibody is one or more of IgA, IgD, IgE, IgG, or IgM.

4. The antibody-drug conjugate of claim 1, wherein the parent antibody has binding affinity and specificity to cancer-specific antigens, cell surface receptor proteins, cell surface proteins, transmembrane proteins, signaling proteins, cell survival regulators, cell proliferation regulators, molecules associated with tissue development or differentiation, lymphokines, cytokines, molecules involved in cell cycle regulation, molecules involved in vasculogenesis, or molecules associated with angiogenesis.

5. The antibody-drug conjugate of claim 4, wherein the parent antibody has binding affinity to one or more proteins of;

(1) BMPRIB (bone morphogenetic protein receptor-type IB; Genbank Accession No. NM\_001203);

- (2) E16 (LAT1, SLC7A5; Genbank Accession No. NM\_003486);
- (3) STEAP1 (six transmembrane epithelial antigen of prostate; Genbank Accession No. NM\_012449);
- (4) 0772P (CA125, MUC16; Genbank Accession No. AF361486);
- (5) MPF (MSLN, SMR, megakaryocyte potentiating factor, mesothelin; Genbank Accession No. NM\_005823);
- (6) Napi3b (NAPI-3B, NPTIIB, SLC34A2, solute carrier family 34 (sodium phosphate), member 2, type II sodium-dependent phosphate transporter 3b; Genbank Accession No. NM\_006424);
- (7) Sema 5b (FLJ10372, KIAA1445, Mm.42015, SEMA5B, SEMAG, Semaphorin 5b Hlog, sema domain, seven thrombospondin repeats (type 1 and type 1-like), transmembrane domain (TM) and short cytoplasmic domain, (semaphorin) 5B; Genbank Accession No. AB040878);
- (8) PSCA hlg (2700050C12Rik, C530008O16Rik, RIKEN cDNA 2700050C12, RIKEN cDNA 2700050C12; Genbank Accession No. AY358628);
- (9) ETBR (Endothelin type B receptor; Genbank Accession No. AY275463);
- (10) MSG783 (RNF124, hypothetical protein FLJ20315; Genbank Accession No. NM\_017763);
- (11) STEAP2 (HGNC\_8639, IPCA-1, PCANAP1, STAMP1, STEAP2, STMP, prostate cancer associated gene 1, prostate cancer associated protein 1, six-transmembrane epithelial antigen of prostate 2, six-transmembrane prostate protein; Genbank Accession No. AF455138);
- (12) TrpM4 (BR22450, FLJ20041, TRPM4, TRPM4B, transient receptor potential cation channel, subfamily M, member 4; Genbank Accession No. NM\_017636);
- (13) CRIPTO (CR, CR1, CRGF, CRIPTO, TDGF1, teratocarcinoma-derived growth factor; Genbank Accession No. NP\_003203 or NM\_003212);
- (14) CD21 (CR2 (Complement receptor 2) or C3DR (C3d/Epstein Barr virus receptor) or Hs.73792; Genbank Accession No. M26004);
- (15) CD79b (CD79B, CD79 $\beta$ , Igb (immunoglobulin-associated beta), B29; Genbank Accession No. NM\_000626);
- (16) FcRH2 (IFGP4, IRTA4, SPAP1A (SH2 domain containing phosphatase

- anchor protein 1a), SPAP1B, SPAP1C; Genbank Accession No. NM\_030764);
- (17) HER2 (Genbank Accession No. M11730);
  - (18) ErbB receptor selected from EGFR, HER3, and HER4;
  - (19) NCA (Genbank Accession No. M18728);
  - (20) MDP (Genbank Accession No. BC017023);
  - (21) IL20R $\alpha$  (Genbank Accession No. AF184971);
  - (22) Brevican (Genbank Accession No. AF229053);
  - (23) EphB2R (Genbank Accession No. NM\_004442);
  - (24) ASLG659 (Genbank Accession No. AX092328);
  - (25) PSCA (Genbank Accession No. AJ297436);
  - (26) GEDA (Genbank Accession No. AY260763);
  - (27) BAFF-R (B cell-activating factor receptor, BLyS receptor, BR3; Genbank Accession No. NP\_443177.1);
  - (28) CD22 (B-cell receptor CD22-B isoform; Genbank Accession No. NP-001762.1);
  - (29) CD79a (CD79A, CD79.alpha., immunoglobulin-associated alpha, a B cell-specific protein that covalently interacts with Ig beta (CD79B) and forms a complex on the surface with IgM molecules, transduces a signal involved in B-cell differentiation; Genbank Accession No. NP\_001774.1);
  - (30) CXCR5 (Burkitt's lymphoma receptor 1, a G protein-coupled receptor that is activated by the CXCL 13 chemokine, functions in lymphocyte migration and humoral defense, plays a role in HIV-2 infection and regarded for development of AIDS, lymphoma, myeloma, and leukemia; Genbank Accession No. NP\_001707.1);
  - (31) HLA-DOB (Beta subunit of MHC class II molecule (Ia antigen) that binds peptides and presents them to CD4<sup>+</sup> T lymphocytes; Genbank Accession No. NP\_002111.1);
  - (32) P2X5 (Purinergic receptor P2X ligand-gated ion channel 5, an ion channel gated by extracellular ATP, may be involved in synaptic transmission and neurogenesis, deficiency may contribute to the pathophysiology of idiopathic detrusor instability; Genbank Accession No. NP\_002552.2);
  - (33) CD72 (B-cell differentiation antigen CD72, Lyb-2; Genbank Accession No.

NP\_001773.1);

(34) LY64 (Lymphocyte antigen 64 (RP105), type I membrane protein of the leucine rich repeat (LRR) family, regulates B-cell activation and apoptosis, loss of function is associated with increased disease activity in patients with systemic lupus erythematosus; Genbank Accession No. NP\_005573.1);

(35) FcRH1 (Fc receptor-like protein 1, a putative receptor for the immunoglobulin Fc domain that contains C2 type Ig-like and ITAM domains, may have a role in B-lymphocyte differentiation; Genbank Accession No. NP\_443170.1);

(36) IRTA2 (Immunoglobulin superfamily receptor translocation associated 2, a putative immunoreceptor with possible roles in B cell development and lymphomagenesis, deregulation of the gene by translocation occurs in some B cell malignancies; Genbank Accession No. NP\_112571.1);

(37) TENB2 (putative transmembrane proteoglycan, related to the EGF/heregulin family of growth factors and follistatin; Genbank Accession No. AF179274);

(38) MAGE-C1/CT7 (protein overexpressed in testicular cancer);

(39) androgen receptor;

(40) CD20;

(41) CD30;

(42) CD33;

(43) CD52;

(44) EpCam;

(45) CEA;

(46) gpA33;

(47) Mucins;

(48) TAG-72;

(49) Carbonic anhydrase IX;

(50) PSMA;

(51) folate binding protein;

(52) gangliosides;

(53) hydrate/saccharide Lewis-Y;

(54) VEGF;

(55) VEGFR;  
 (56) aVb3;  
 (57) a5b1;  
 (58) ERB3;  
 (59) c-MET;  
 (60) EphA3;  
 (61) TRAIL-R1 or TRAIL-R2;  
 (62) RANKL;  
 (63) FAP;  
 (64) Tenascin;  
 (65) PTEN; or  
 (66) human kallikrein-related peptidase 3 (protein overexpressed in prostatic cancer).

6. The antibody-drug conjugate of claim 4, wherein the parent antibody comprises one or more antibodies which are trastuzumab, rituximab, bevacizumab, cetuximab, panitumumab, ipilimumab, alemtuzumab, ofatumumab, gemtuzumab, brentuximab, <sup>90</sup>Y-ibritumomab, <sup>131</sup>I-tositumomab, cBR96, cAC10, anti-CD20 antibody, anti-EphB2 antibody, anti-IL-8, E-selectin antibody, anti-MUC16 antibody, anti-CD30 antibody, anti-CD33 antibody, or anti-CD52 antibody.

7. The antibody-drug conjugate of claim 1, wherein the drug is conjugated to the cysteine residue in the cysteine-containing motif by a linker.

8. The antibody-drug conjugate of claim 7, wherein the linker is one or more of alkyl halide derivatives containing a haloacetyl group, derivatives containing a maleimide group, aryl halide derivatives containing fluorobenzene, aziridine derivatives or acryloyl derivatives, wherein the derivative comprises an alkylating reactive group, an arylating reactive group, a maleimide group, an aziridine group, an acryloyl group, or a disulfide exchange reactive group comprising pyridyl disulfide or thionitrobenzoic acid, and wherein the linker is covalently bound to the thiol group of the cysteine residue in the cysteine-containing motif.

9. The antibody-drug conjugate of claim 1, wherein the drug is one or more of microtubulin inhibitors, mitotic inhibitors, topoisomerase inhibitors, DNA intercalators, anticancer agents, protein toxins capable of functioning as enzymes, oncogene specific micro-RNA (miRNA), siRNA, shRNA, or radioisotopes.

10. The antibody-drug conjugate of claim 1, wherein the drug is one or more of maytansinoid, auristatin, aminopterin, actinomycin, bleomycin, talisomycin, camptothecin, N<sup>8</sup>- acetyl spermidine, 1-(2 chloroethyl)-1,2- dimethyl sulfonyl hydrazide, taxol, esperamicin, etoposide, 6-mercaptopurine, dolastatin, trichothecene, CC1065, calicheamicin and other enediyne antibiotics, taxane, anthracycline, methotrexate, adriamycin, vindesine, vinca alkaloids, vincristine, vinblastine, etoposide, doxorubicin, melphalan, mitomycin A, mitomycin C, chlorambucil, daunorubicin, daunomycin, duocamycin nucleolytic enzymes, antibiotics, toxins of bacterial, plant or animal origin, cisplatin, CPT-11, doxorubicin, paclitaxel, or docetaxel.

11. A method of producing a modified antibody comprising a cysteine-containing motif bound at the terminus of a parent antibody, the method comprising the steps of:

(a) constructing an expression vector comprising a polynucleotide sequence in which a polynucleotide sequence encoding the cysteine-containing motif and a polynucleotide sequence encoding the parent antibody are recombinantly linked to each other;

(b) expressing the constructed expression vector in a culture using host cells; and

(c) isolating and purifying the modified antibody from the culture,

wherein the cysteine-containing motif is a metal ion binding motif that contains a cysteine residue,

wherein the metal ion binding motif that contains a cysteine residue comprises one or two CGH or HGC motif(s) with alanine at a C-terminus and an N-terminus of the motif,

wherein the cysteine-containing motif is bound to the C-terminus of the heavy chain of the antibody, and

wherein a drug is conjugated to the thiol group of the cysteine residue in the metal ion binding motif.

12. The method of claim 11, wherein the host cells are monkey kidney cells 7 (COS7), NSO cells, SP2/0 cells, Chinese hamster ovary (CHO) cells, W138 cells, baby hamster kidney (BHK) cells, Madin-Darby canine kidney (MDCK) cells, myeloma cell lines, HuT 78 cells, or HEK293 cells.

13. A therapeutic composition comprising the antibody-drug conjugate according to claim 1 and a pharmaceutically acceptable carrier.

Drawings

1/6

FIG. 1

5

10

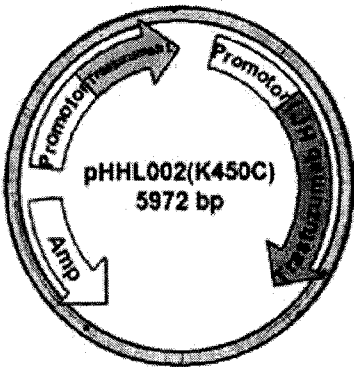


FIG. 2

15

20

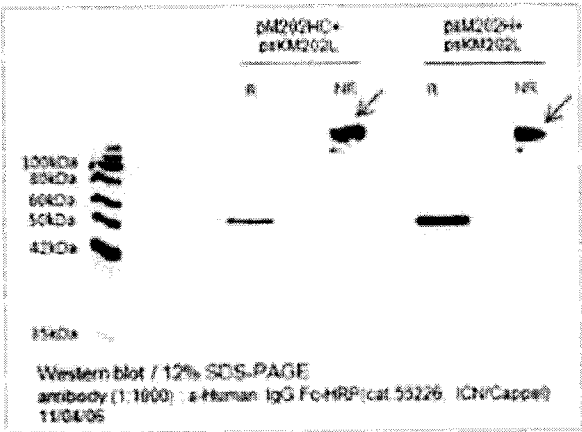
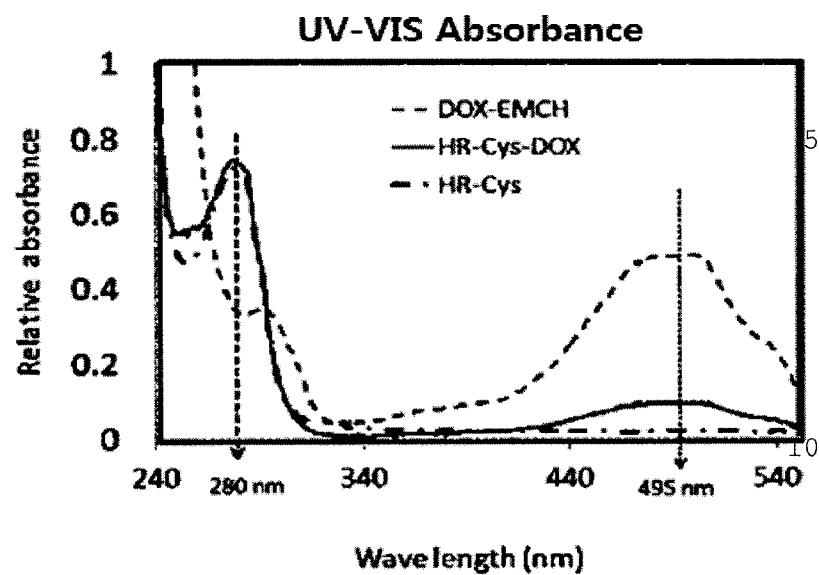




FIG. 3

(A)



(B)

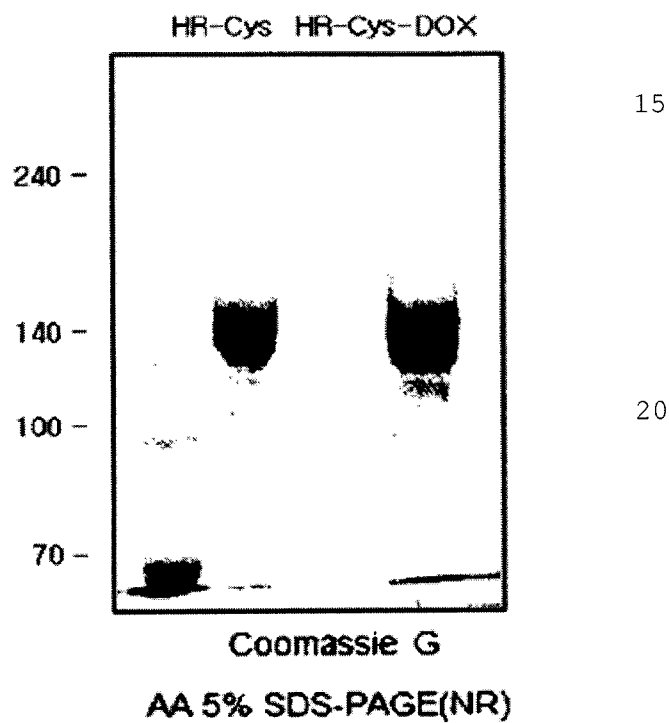


FIG. 4

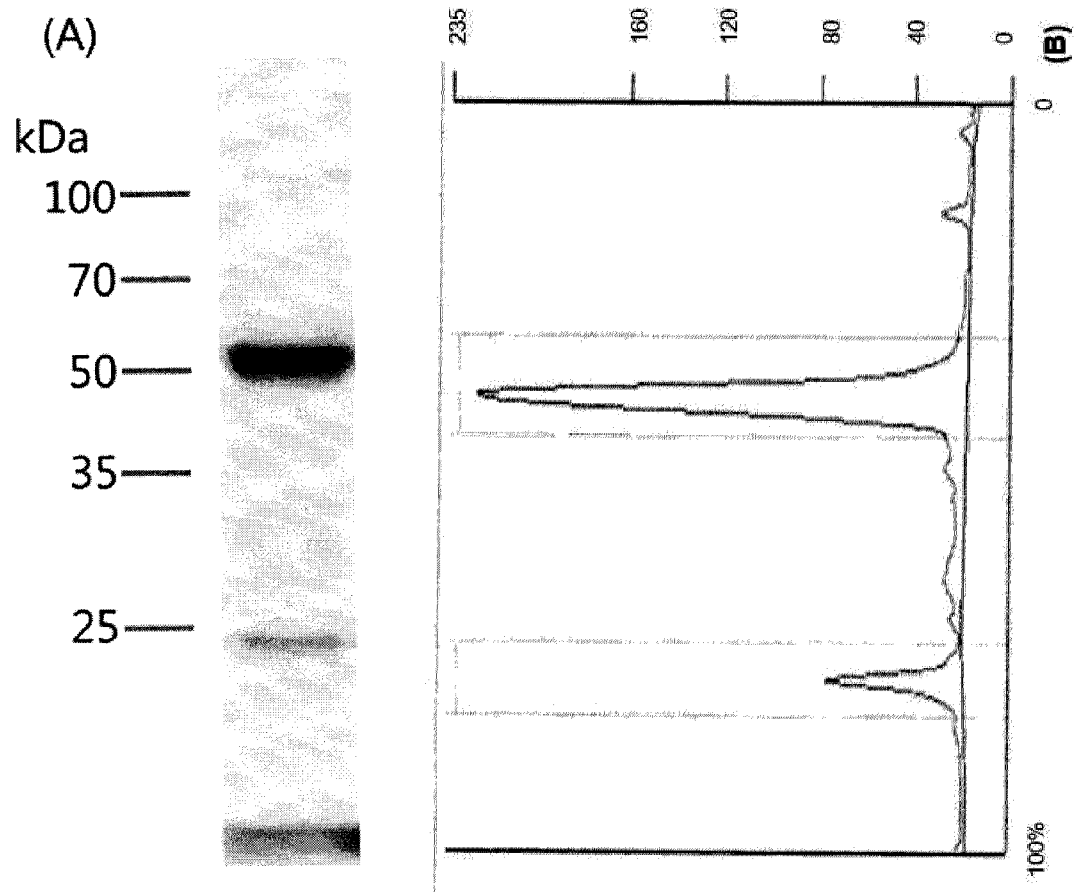


FIG. 5

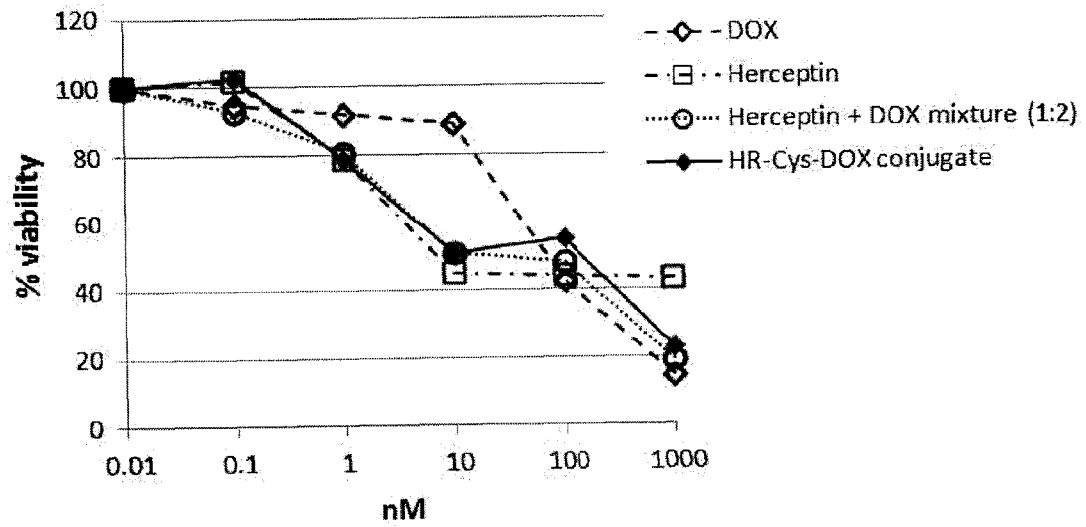


FIG. 6

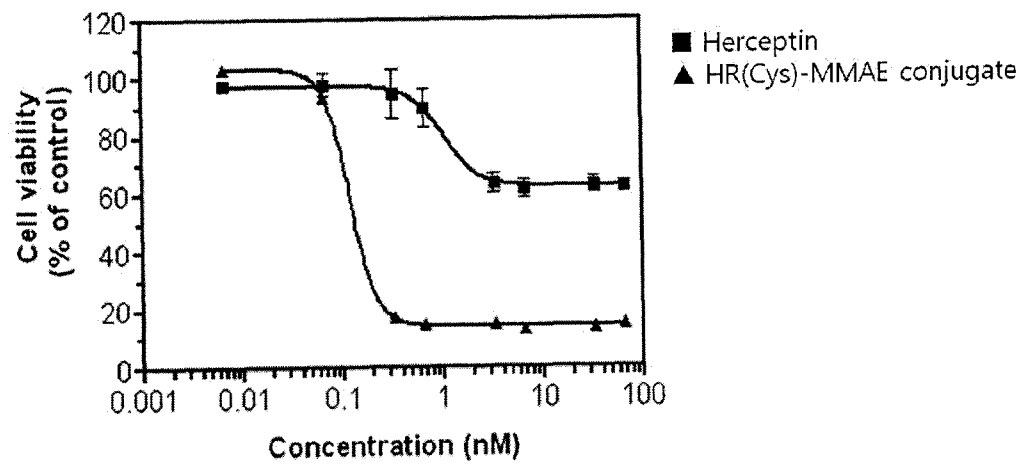


FIG. 7

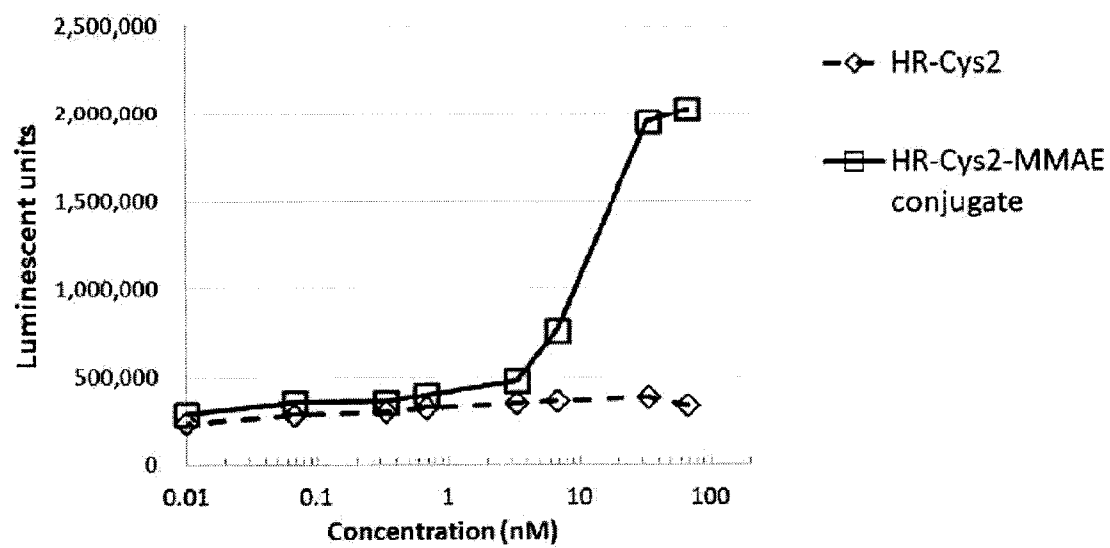
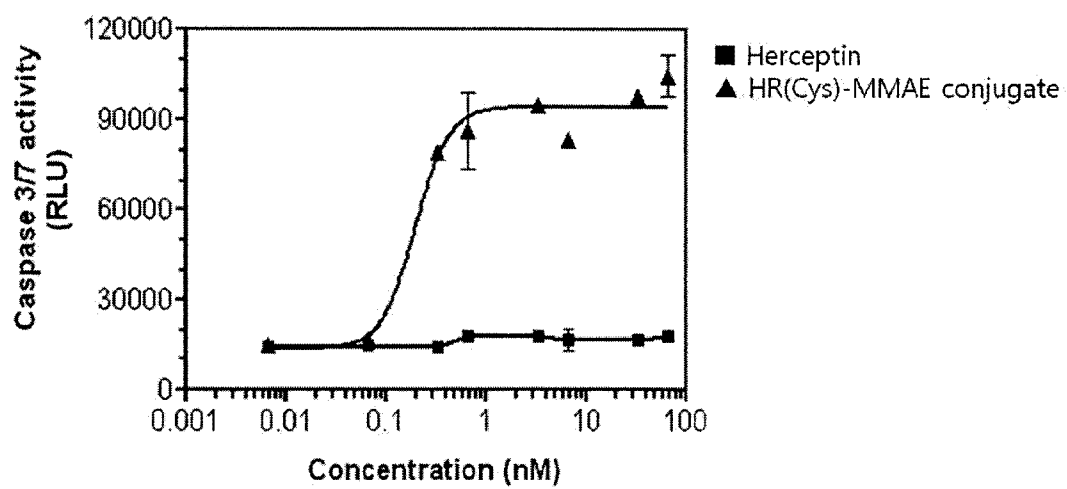
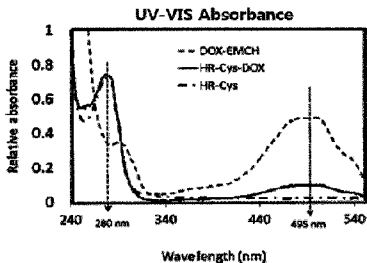


FIG. 8



(A)



(B)

