



HU000025488T2

(19) **HU**(11) Lajstromszám: **E 025 488**(13) **T2****MAGYARORSZÁG**  
Szellemi Tulajdon Nemzeti Hivatala**EURÓPAI SZABADALOM**  
**SZÖVEGÉNEK FORDÍTÁSA**

- (21) Magyar ügyszám: **E 10 723936**
- (22) A bejelentés napja: **2010. 06. 02.**
- (96) Az európai bejelentés bejelentési száma:  
**EP 20100723936**
- (97) Az európai bejelentés közzétételi adatai:  
**EP 2437874 A1**                      **2010. 12. 09.**
- (97) Az európai szabadalom megadásának meghirdetési adatai:  
**EP 2437874 B1**                      **2015. 05. 06.**
- (51) Int. Cl.: **B01F 7/00**                      (2006.01)  
**C12M 1/02**                      (2006.01)  
**C12M 1/06**                      (2006.01)
- (86) A nemzetközi (PCT) bejelentési szám:  
**PCT/EP 10/003356**
- (87) A nemzetközi közzétételi szám:  
**WO 10139470**

(30) Elsőbbségi adatok: <b>09007457</b> <b>2009. 06. 05.</b> <b>EP</b>	(73) Jogosult(ak): <b>F.HOFFMANN-LA ROCHE AG, 4070 Basel (CH)</b>
(72) Feltaláló(k): <b>JENZSCH, Marco, 82377 Penzberg (DE)</b> <b>LECHNER, Max, 82377 Penzberg (DE)</b> <b>POHLSCHIEDT, Michael, KR (KR)</b> <b>REESE, Christoph, 81375 München (DE)</b> <b>SCHOLZ, Alexander, 76187 Karlsruhe (DE)</b> <b>TEBBE, Hermann, 82444 Schlehdorf (DE)</b>	(74) Képviselő: <b>dr. Miskolczi Mária, Budapest</b>

(54) **Berendezés sejtenyésztésre**

Az európai szabadalom ellen, megadásának az Európai Szabadalmi Közlönyben való meghirdetésétől számított kilenc hónapon belül, felszólalást lehet benyújtani az Európai Szabadalmi Hivatalnál. (Európai Szabadalmi Egyezmény 99. cikk(1))

A fordítást a szabadalmas az 1995. évi XXXIII. törvény 84/H. §-a szerint nyújtotta be. A fordítás tartalmi helyességét a Szellemi Tulajdon Nemzeti Hivatala nem vizsgálta.



(11) **EP 2 437 874 B1**

(12) **EUROPEAN PATENT SPECIFICATION**

(45) Date of publication and mention of the grant of the patent:  
**06.05.2015 Bulletin 2015/19**

(51) Int Cl.:  
**B01F 7/00** <sup>(2006.01)</sup> **C12M 1/02** <sup>(2006.01)</sup>  
**C12M 1/06** <sup>(2006.01)</sup>

(21) Application number: **10723936.0**

(86) International application number:  
**PCT/EP2010/003356**

(22) Date of filing: **02.06.2010**

(87) International publication number:  
**WO 2010/139470 (09.12.2010 Gazette 2010/49)**

(54) **Device for cultivating cells**

Vorrichtung zur Kultivierung von Zellen

Dispositif pour cultiver des cellules

(84) Designated Contracting States:  
**AL AT BE BG CH CY CZ DE DK EE ES FI FR GB GR HR HU IE IS IT LI LT LU LV MC MK MT NL NO PL PT RO SE SI SK SM TR**

(74) Representative: **Burger, Alexander et al**  
**Roche Diagnostics GmbH**  
**Patent Department (LPP.....6164)**  
**P.O.Box 11 52**  
**82372 Penzberg (DE)**

(30) Priority: **05.06.2009 EP 09007457**

(43) Date of publication of application:  
**11.04.2012 Bulletin 2012/15**

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**EP-A2- 0 224 800 WO-A1-01/41919**  
**US-A- 4 438 074 US-A- 5 633 165**

(73) Proprietor: **F.Hoffmann-La Roche AG**  
**4070 Basel (CH)**

(72) Inventors:  
• **JENZSCH, Marco**  
**82377 Penzberg (DE)**  
• **LECHNER, Max**  
**82377 Penzberg (DE)**  
• **POHLSCHIEDT, Michael**  
**KR (KR)**  
• **REESE, Christoph**  
**81375 München (DE)**  
• **SCHOLZ, Alexander**  
**76187 Karlsruhe (DE)**  
• **TEBBE, Hermann**  
**82444 Schlehdorf (DE)**

- **NIENOW A W: "AGITATORS FOR MYCELIAL FERMENTATIONS" TRENDS IN BIOTECHNOLOGY, ELSEVIER PUBLICATIONS, CAMBRIDGE, GB LNKD- DOI:10.1016/0167-7799 (90)90180-6, vol. 8, no. 8, 1 August 1990 (1990-08-01), pages 224-233, XP000142634 ISSN: 0167-7799**
- **TOSHIO NOMURA ET AL: "DEVELOPMENT AND MIXING CHARACTERISTICS OF FOLDING ANCHOR IMPELLER FOR ROUND-BOTTOMED FLASK" JOURNAL OF CHEMICAL ENGINEERING OF JAPAN, SOCIETY OF CHEMICAL ENGINEERS. TOKYO, JP, vol. 29, no. 1, 1 February 1996 (1996-02-01), pages 134-138, XP000583013 ISSN: 0021-9592**

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**EP 2 437 874 B1**

**Description**

**[0001]** Herein is reported an agitator, a device comprising the agitator and the use of the agitator for the cultivation of cells. The agitator comprises at least one axially-conveying element and at least one radially-conveying element, such as the combination of an anchor impeller as a radially-conveying element and one or more inclined-blade impeller(s) as an axially-conveying element. The use of the agitator results in an improved mixing and in particular a lower biofouling in dialysis processes as well as a higher mass transfer rate.

**Technological background**

**[0002]** The production of recombinant proteins, vaccines and antibodies with the aid of prokaryotic and eukaryotic cells plays an essential role in modern pharmaceutical production. To produce complex post-translationally modified proteins and antibodies animal derived cells are primarily used. However, the use of animal derived cells sets high demands for the fermentation process due to the specific characteristics of these cells such as e.g. the culture medium, the sensitivity towards limitation and inhibitions (for example by lactate, CO<sub>2</sub>, ammonium etc.), the sensitive outer membrane (shear stress), the low specific rates and the sensitivity towards variations in the culture conditions (e.g. due to local inhomogeneities, pH variations, pO<sub>2</sub> variations etc.). These properties have to be taken into consideration when designing bioreactors and for process control.

**[0003]** In recent years various types of reactors for culturing cells have been developed. Irrespective of the type, the reactor must be able to fulfill the following basic technical functions: adequate suspension as well as homogenization, adequate material and heat transport as well as a minimal shear stress on the cells. The stirred-tank reactor is especially suitable for industrial use. In this reactor the necessary energy for fulfilling the basic functions is introduced by mechanical stirring.

**[0004]** In order to achieve high product titer and specification compliant product quality, the operating mode of the reactor in particular plays an important role in addition to cell line development, media composition and design of the reactor. Generally the following operating modes are employed: batch processes, fed batch processes, continuous processes with or without cell retention (for example perfusion or chemostat) as well as semi-continuous processes such as e.g. internal or external dialysis.

**[0005]** To prevent nutrient limitations feed solutions are added to reactor in the form of concentrated solutions (so-called fed batch processes). To avoid inhibition by end products of the cell's metabolism the products of the cell's metabolism have to be removed from the reactor or from the culture medium in the reactor. This can be carried out for example by perfusion or dialysis. In the case of dialysis one differentiates between external or internal dialysis.

**[0006]** EP 1 474 223 reports a dynamic mixer. An animal cell culturing device comprising a container for the uptake of a cell suspension, a device for feeding- and discharge of gas into the cell suspension, and a device for production of a current in the cell suspension is reported in DE 10 2005 053 333. Method and apparatus for cell culture is reported in EP 0 224 800. The apparatus comprises a stirrer and a semipermeable membrane separating a first chamber and a second chamber inside a device for cultivating cells. Nomura, T., et al., J. Chem. Eng. Jap. 29 (1996) 134-138 report development and mixing characteristics of folding anchor impeller for round bottom flask. An impeller draft tube agitation system for gas-liquid mixing in a stirred tank reactor is reported in WO 01/41919. In US 4,438,074 a continuous polymerization reactor is reported. An agitation system and method for gas transfer into liquids is reported in EP-A 0 200 886. In EP-B 0 049 229 a cooking apparatus having a stirring device is reported.

**[0007]** Nienow et al. (Trends. Biotechnol. 8 (1990) 224-233) report agitators for mycelial fermentations. In US 5,633,165 a fermentor with vertical shaft is reported.

**Summary of the invention**

**[0008]** The agitator as reported herein at least i) allows for rapid mixing of culture media compared to other stirrers e.g. for introducing correction agents such as acids or bases via the liquid or cultivation medium surface, ii) reduces foam formation, iii) reduces biofouling in dialysis processes, and iv) increase mass transfer in dialysis processes due to the direct orthogonal flow against the dialysis module as a result of the combination of differently conveying elements.

**[0009]** Herein is reported an agitator (combination stirrer) comprising at least one axially-conveying element and at least one radially-conveying element relative to the shaft of the agitator wherein the largest diameter of the at least one axially-conveying element is equal to or less than the inner diameter  $d_i$  of the radially-conveying element.

**[0010]** Reported herein is an agitator, comprising

- one radially-conveying element comprising at least two stirrer blades, and
- one or more axially-conveying elements each comprising at least two stirrer blades,

wherein the stirrer blades of the radially-conveying element are parallel to each other and to the shaft axis of the agitator, and

wherein the outer diameter of all axially-conveying elements is equal to or less than the inner diameter of the radially-conveying element, and

5 wherein all axially-conveying elements are individually connected to the radially-conveying element, and

wherein all axially-conveying elements are located within the radially-conveying element, and

wherein all conveying elements have a fixed spatial orientation relative to each other and to the shaft axis of the agitator.

**[0011]** In one embodiment the agitator comprises 1 to 5 axially-conveying elements, or in another embodiment 1 to 3 axially-conveying elements, or in also an embodiment 1 or 2 axially-conveying elements. In one embodiment the at least  
10 one axially-conveying element is situated in the upper four fifths of the agitator determined from the head of the agitator and is at a maximum distance of  $h_{4/5}$  to the head of the agitator. In also an embodiment one axially-conveying element is located at a maximum distance of 80 % from the top of the blades of the radially-conveying element (i.e. at a maximum distance of 0.8 h) and/or one axially-conveying element is located at a maximum distance of 20 % from the top of the blades of the radially-conveying element (i.e. at a maximum distance of 0.2 h). In a further embodiment the at least one  
15 axially-conveying element and the at least one radially-conveying element form together a single element. In another embodiment opposing stirrer blades of the radially-conveying element are linked to each other by two opposite stirrer blades of an axially-conveying element. In one embodiment the diameter of all axially-conveying elements is identical. In another embodiment the axially-conveying elements is selected independently of each other from a propeller agitator, pitched-blade agitator, or inclined-blade agitator.

20 **[0012]** If the diameter of the axially-conveying element is less than inner diameter of the radially-conveying element the spatial distance between the tip of the stirrer blade of the axially-conveying element and the inner edge of the radially-conveying element is bridged by a connector. The connector is in one embodiment selected from wire, rod, sheet plate and disc.

**[0013]** In a further embodiment all conveying elements, i.e. the axially-conveying as well as the radially-conveying  
25 elements, rotate with the same number of rotations per time unit around the shaft axis of the agitator when the agitator is operated in a cultivation vessel. In one embodiment the conveying elements are permanently joined together and the agitator consists of one part, i.e. both elements are driven by the same rotary shaft and have the same number of rotations per time unit around the shaft axis of the agitator.

**[0014]** The ratio  $d/D$  of agitator diameter ( $d$ ) to cultivation vessel diameter ( $D$ ) is in one embodiment of from 0.2 to 0.8,  
30 in another embodiment of from 0.3 to 0.6, and in also an embodiment of from 0.33 to 0.5. In another embodiment the ratio  $h/d$  of blade height ( $h$ ) to agitator diameter ( $d$ ) is of from 0.5 to 5, in another embodiment of from 1 to 4, and in also an embodiment of from 1 to 3. In yet another embodiment the ratio  $b/d$  of agitator blade width ( $b$ ) to agitator diameter ( $d$ ) is of from 0.05 to 0.3, in another embodiment of form 0.1 to 0.25. The term "of from ... to" denotes a range including the listed boundary values. In another embodiment the stirrer diameter ( $d$ ) is selected from 500 mm, 600 mm, 700 mm,  
35 800 mm, 1000 mm, 1200 mm, 1400 mm, 1600 mm, 1800 mm and 2000 mm. In a further embodiment the agitator blade width ( $b$ ) is selected from, 42 mm, 60 mm, 89 mm, 108 mm, 133 mm.

**[0015]** In one embodiment the one radially-conveying element is an anchor impeller. In a further embodiment the elements connecting the individual stirrer blades of the radially-conveying element are the stirrer blades of the axially-conveying element (connecting stirrer blades). In an embodiment the connecting stirrer blades act of from 20 % to 100  
40 % as an axially-conveying element, in another embodiment of from 50 % to 100 % as an axially-conveying element, and in also an embodiment of approximately 100 % as an axially-conveying element. The term "approximately" denotes that the given value is the center point of a range spanning plus/minus 10 % around the value. If this value is a percentage value then "approximately" denotes also means plus/minus 10 %, but the value 100 % cannot be exceeded.

**[0016]** In a further embodiment the axially-conveying element is an inclined-blade stirrer. In another embodiment the  
45 radially-conveying element is an anchor impeller and the axially-conveying element is an inclined-blade stirrer where the inclined-blade stirrer is designed to act as a connecting element of blades of the anchor impeller. In another embodiment all opposing blades of the radially-conveying element are connected to each other via blades of an axially-conveying element. In one embodiment the agitators consists of a combination of one or two axially-conveying elements and one radially-conveying element relative to the shaft axis of the agitator. In another embodiment the radially-conveying element  
50 is an anchor impeller and the axially-conveying elements are two inclined-blade stirrers wherein blades of the inclined-blade stirrers are designed to act as connecting elements of opposing anchor impeller blades.

**[0017]** The ratio  $h_{SB}/b$  of the height of the axially-conveying element and the width of the blades of the radially-conveying element is of from 0.5 to 4, in another embodiment of from 0.8 to 3, and also in an embodiment of from 1 to 2. In another  
55 embodiment the pitch of the stirrer blades of the inclined-blade stirrer is of from 10° to 80°, in a further embodiment of from 24° to 60°, and in also an embodiment of from 40° to 50° relative to the shaft axis of the agitator.

**[0018]** In one embodiment the radially-conveying element has of from 1 to 8 stirrer blades, in another embodiment of from 1 to 4 stirrer blades, and also in an embodiment 4 stirrer blades. The axially-conveying element has in one embodi-  
ment of from 1 to 10 stirrer blades, in another embodiment of from 2 to 6 stirrer blades, and also in an embodiment 4

stirrer blades. In another embodiment the radially-conveying and axially-conveying elements have the same number of stirrer blades.

**[0019]** In one embodiment the locking device on the shaft of the agitator is in the lower third seen from the head of the stirrer and has a ratio  $h_u/h_m$  of the height of the reduction part ( $h_u$ ) and the height of the tightening lid ( $h_m$ ) of from 0.05 to 1, in another embodiment of from 0.2 to 0.8, and in also an embodiment of from 0.3 to 0.5.

**[0020]** In one embodiment the agitator has a height of at least 200 mm, in another embodiment of from 200 mm to 5000 mm. In another embodiment the height of the agitator is the height  $h$  of the blades of the radially-conveying element.

**[0021]** Herein is reported as an aspect a device comprising the agitator as reported herein, a dialysis module and a cultivation vessel. In one embodiment the agitator and the cultivation vessel form a functional unit, i.e. the agitator is within the cultivation vessel and can rotate within the cultivation vessel without any spatial limitation. In one embodiment the cultivation vessel is a stirred tank reactor. In a further embodiment the cultivation vessel is an aerated or submersed gassed stirred tank reactor. In one embodiment the cultivation vessel comprises of from 2 to 4 baffles. In another embodiment the baffles are spaced equidistally around the circumference of the inside surface of the cultivation vessel.

**[0022]** The ratio  $d/D$  of agitator diameter ( $d$ ) to cultivation vessel diameter ( $D$ ) is in one embodiment of from 0.2 to 0.8, in another embodiment of from 0.3 to 0.6, and in also an embodiment of from 0.33 to 0.5. In another embodiment the ratio  $H/D$  of filling height of the reaction vessel ( $H$ ) to cultivation vessel diameter ( $D$ ) is of from 1.0 to 2.5, in a further embodiment of from 1.1 to 2.0, and also in an embodiment of from 1.4 to 1.8. In one embodiment the cultivation vessel has a working volume of from 5l to 25,000 l.

**[0023]** Herein is also reported as an aspect the use of the device as reported herein. In one embodiment the use is for the cultivation of cells for the production of a polypeptide, i.e. for the cultivating of cells expressing a polypeptide encoded by a heterologous nucleic acid. In one embodiment the cultivating is a dialysis. In a further embodiment the cultivating is carried out in a submersed gassed stirred tank reactor. In another embodiment the cell is a eukaryotic cell. In also an embodiment the cell is a mammalian cell. In yet a further embodiment the cell is selected from a CHO cell, a BHK cell, an NS0 cell, a COS cell, a PER.C6 cell, a Sp2/0 cell, and a HEK 293 cell. In one embodiment the polypeptide is an antibody. In a further embodiment the antibody is an antibody against CD20, CD22, HLA-DR, CD33, CD52, EGFR, G250, GD3, HER2, PSMA, CD56, VEGF, VEGF2, CEA, Lewis Y antigen, the IL-6 receptor or the IGF-1 receptor.

**[0024]** Herein is further reported as an aspect a method for the production of a polypeptide comprising

- a) providing a cell comprising a nucleic acid encoding the polypeptide,
- b) providing a device as reported herein,
- c) cultivating the cell in the device in a cultivation medium wherein the agitator provides a turbulent flow within the cultivation vessel, and
- d) recovering the polypeptide from the cells or the cultivation medium and thereby producing a polypeptide.

**[0025]** In one embodiment the method is for the cultivating of a cell expressing a polypeptide encoded by a heterologous nucleic acid. In one embodiment the cultivating is a dialysis. In a further embodiment the cultivating is carried out in a submersed gassed stirred tank reactor. In another embodiment the cell is a eukaryotic cell. In also an embodiment the cell is a mammalian cell. In yet a further embodiment the cell is selected from a CHO cell, a BHK cell, an NS0 cell, a COS cell, a PER.C6 cell, a Sp2/0 cell, and a HEK 293 cell. In one embodiment the polypeptide is an antibody. In another embodiment the antibody is an antibody against CD20, CD22, HLA-DR, CD33, CD52, EGFR, G250, GD3, HER2, PSMA, CD56, VEGF, VEGF2, CEA, Lewis Y antigen, the IL-6 receptor or the IGF-1 receptor.

### **Detailed description of the invention**

**[0026]** Herein reported is an agitator, which is a combination stirrer, comprising a radially-conveying element with one or more defined and integrated axially-conveying elements. The term "element" denotes a (functional) unit of stirrer blades which are in a fixed spatial configuration relative to one another with regard to distance and angle. A radially-conveying element denotes a stirrer which stirrer blades have no pitch with respect to the shaft axis. An axially-conveying element denotes a stirrer which stirrer blades have a pitch with respect to the shaft axis. The stirrer blades of the conveying elements are in one embodiment rectangular plates although other geometric forms can be used. The conveying direction of an element is denoted with respect to the shaft axis of the agitator. The axially-conveying elements, especially the individual blades thereof, are arranged as connecting pieces between the blades of the radially-conveying element. Also reported herein is a device comprising the agitator and a cultivation vessel. Further reported is the use of the agitator and the device in the cultivating of cells, in particular in a semi-continuous process such as e.g. internal or external dialysis. Exemplary embodiments of the agitators as reported herein are shown in Figure 1.

**[0027]** Each of the conveying elements consists of a defined number of stirrer blades. Each blade is either directly connected to the rotary shaft or is connected to the rotary shaft via a hub. Each stirrer blade independent of the conveying element has an outer edge and an inner edge. The part of each stirrer blade that has the maximum distance to the shaft

axis is denoted as tip of the blade. Each conveying element has an outer diameter and an inner diameter. For example, the outer diameter of an axially-conveying element is the maximum distance between the tips of opposing stirrer blades and the inner diameter of a radially-conveying element is the minimum distance between inner edges of opposing stirrer blades. In the agitator as reported herein the axially-conveying element(s) is(are) located within the radially-conveying element. Therefore, the outer diameter of the (all) axially-conveying element(s) is (are) equal to or less than the inner diameter of the radially-conveying element allowing the axially-conveying element(s) to be placed inside of the radially-conveying element. Some stirrer blades of (each of) the axially-conveying element(s) connect opposing stirrer blades of the radially-conveying element. As the radially-conveying elements are not directly connected to each other by this connection (each of) the axially-conveying element is individually, i.e. by itself, connected to the radially-conveying element. The connecting stirrer blades of the axially-conveying element do not need to be formed as stirrer blades for the entire distance between the stirrer blades of the radially-conveying element. Thus, the connecting blades of the axially-conveying element do not need to have the function of an axially-conveying over the entire distance, i.e. they do not have to be formed as a stirrer blade over the entire distance. That is the stirrer blade of the axially-conveying element may be shorter than the spatial distance to the inner edge of the stirrer blade of the radially-conveying element. This spatial gap is filled by a connector that has no special form but provides for the connection of the tip of the blade of the axially-conveying element to the inner edge of the blade of the radially-conveying element.

**[0028]** The height  $h$  of the radially-conveying element is more than 10 times the height  $h_{SB}$  of the axially-conveying elements. Thus, each of the axially-conveying elements can be placed at different positions relative to the height of the radially-conveying element. In one embodiment an axially-conveying element is located, i.e. it is placed, within a maximum distance of 80 % of the total height  $h$  of the radially-conveying element relative to the head  $K$  of the agitator. This denotes that the axially-conveying element has a maximum distance from the head of the agitator of 0.8  $h$ .

**[0029]** The rotary shaft (also termed drive shaft) extends through the longitudinal axis of the cultivation vessel in which the agitator is used.

**[0030]** The modification with respect to a primarily radially-conveying impeller, e.g. to an anchor impeller, is the (axially) downwards-conveying inclined-blade stirrer which is integrated in the upper and/or middle and/or lower region of the radially-conveying element. The agitator as reported herein provides among other things for an improved mixing of liquid solutions, for example in order to introduce correcting agents such as e.g. an acid or a base, nutrient solutions, anti-foaming agents or also  $CO_2$  or  $O_2$  via the liquid surface. Furthermore, the agitator reduces foam formation and in a dialysis process reduces biofouling and increases mass transfer by a direct orthogonal flow against the dialysis module. It has been found that the additional axially-conveying element(s) surprisingly does(do) not increase shear stress while at the same time improving the mixing and mass transport in the dialysis module.

**[0031]** In semi-continuous culturing processes, such as e.g. external or internal dialysis, substrates are fed to the reactor across a membrane and at the same time inhibiting components / metabolic products of the cultured cells are lead away. This exchange of material is by diffusion. The main influence factors therefore are the prevailing concentration difference, the membrane material, the membrane surface, the diffusion coefficients of the respective compounds inside the membrane material and the thickness of the phase interface which is determined by the flow against the membrane.

**[0032]** When dialysis is employed in high cell density fermentation it is a perfusion-like, semi-continuous process in which a (hollow fiber) dialysis module attached in the reactor provides the exchange area between the culture medium and fresh nutrient medium. The nutrient medium is pumped from a storage container through the dialysis module and thereafter returned again into the storage container (for a schematic diagram see Figure 2). The dialysis module can be located outside the reactor (external dialysis) or within the reactor (internal dialysis). The same physical laws apply to both operating modes.

**[0033]** Thus, reported herein is a device comprising an agitator as reported herein and a cultivation vessel. In one embodiment the device also comprises a dialysis module. The components of the device are dimensioned in a way that they can exert their intended function, i.e. the cultivation vessel can take up the cultivation medium, the agitator can mix the medium and disperse added compound therein and the dialysis module can provide fresh medium and lead away metabolic compounds secreted by the cultivated cell. Thus, the agitator has a diameter that allows for an unhampered rotation within the vessel in the presence and absence of the dialysis module. It has been found that with the device as reported herein a high cell density cultivation as perfusion cultivation or as dialysis cultivation can be carried out advantageously. The cultivating is carried out in one embodiment at a rotation speed of the agitator at which a Reynolds-number independent constant power input to the cultivation medium can be achieved, i.e. during the cultivating a turbulent cultivation medium flow in the cultivation vessel is provided. It is possible with a device as reported herein to cultivate shear sensitive mammalian cells at a lower rotation speed of the agitator but at the same power input compared to known stirrers.

**[0034]** The form of the cultivation vessel is not limited. In one embodiment the cultivation vessel is a cylindrical vessel. In another embodiment the cultivation vessel is a stirred tank reactor. The cultivation vessel may have any dimension. In one embodiment the cultivation vessel has a working volume of from 5 l to 25,000 l.

**[0035]** Components from the fresh nutrient medium diffuse from the interior of the dialysis module through the semi-

permeable hollow fiber membrane into the reactor and at the same time metabolites of the cultivated cells diffuse in the opposite direction from the reactor into the nutrient medium according to the concentration difference. The aim is to keep the absolute concentration of inhibiting metabolites in the reactor as low as possible (dilution) and at the same time to maintain the concentration of essential nutrients as long as possible at an optimal level for the culture. This results in improved culture conditions compared to a process without dialysis enabling to achieve higher maximum cell density or product titer.

**[0036]** The transport processes in the dialysis module can be described in an equivalent manner to mass transfer on a gas bubble by the two-film theory in conjunction with the first Fick's law. Thus, assuming linear gradients based on the exchange area  $A_H$  of the hollow fiber dialysis module, the effective transferred diffusion flux  $J_{eff}$  is according to equation 1:

$$J_{eff,i} = -D_{eff,i} \cdot A_H \cdot \frac{c_{2,i} - c_{1,i}}{x_2 - x_1} = -D_{eff,i} \cdot A_H \cdot \frac{\Delta c_i}{z_{eff}} \quad (\text{Equation 1}).$$

**[0037]** The driving force of diffusion is the concentration difference  $\Delta c_i$  between the inside and outside of the dialysis module relative to the effective diffusion path  $z_{eff}$ . This effective diffusion path is composed of the individual paths through the inner laminar boundary layer on the inner side of the hollow fiber membrane of the dialysis module  $\delta_{BI}$ , through the hollow fiber membrane itself  $\delta_M$  and through the outer laminar boundary layer on the outer side of the hollow fiber membrane in the reactor  $\delta_{HI}$  (see Figure 3). They generally depend on the size and shape of the diffusing molecule, the properties of the surrounding medium and the temperature. Separate mass transfer coefficients can be defined for the respective individual sections and from the summation of their reciprocals the total mass transfer resistance  $1/k$  can be given according to equation 2:

$$\frac{1}{k} = \frac{1}{k_{HI}} + \frac{1}{k_M} + \frac{1}{k_{BI}} \quad (\text{Equation 2}).$$

**[0038]** The transport resistances in the laminar boundary layers on the inner and outer side of the hollow fiber membrane also depend on the flow against the hollow fiber membrane. The better, i.e. the more perpendicular, the flow towards the membrane is the narrower the laminar boundary layers become and the lower are the corresponding transport resistances.

**[0039]** For a dialysis module in a reactor a direct dependency of the transport resistance of the outer laminar boundary layer on among others from the following factors exists:

- the speed of rotation of the stirrer,
- the type of stirrer,
- the flow against the membranes,
- the primary flow profile generated by the stirrer.

**[0040]** The resistance of the laminar boundary layer on the inner side of the hollow fiber membrane can be neglected due to the low inner diameter and the concomitant high flow velocities. Within this application the term "inner side of the hollow fiber membrane" denotes the side of the hollow fiber membrane which faces the storage container. The term "outer side of the hollow fiber membrane" denotes the side of the hollow fiber membrane which faces the reactor. The total mass transfer resistance is thus a series resistance to which mainly the resistance within the membrane and the resistance of the outer laminar boundary layer contribute (Rehm, et. al., Biotechnology - volume 3: Bioprocessing, VCH Weinheim, 1993). The total mass transfer coefficient  $k$  results from the reciprocal total mass transfer resistance and can be related to the surface area by multiplication with the volume-specific surface  $a$  of the hollow fiber dialysis module ( $ka$  value).

**[0041]** The following equation 3 can be used to describe the concentration time courses for the balance spaces reactor and storage container:

$$\frac{dc_R}{dt} = ka \cdot (c^* - c_R) \quad (\text{Equation 3}).$$

[0042] The typical concentration time courses in the reactor  $c_R$  and the storage container  $c_V$  are shown in Figure 4.

[0043] In general submersed gased reactors are used in cell culture. In these cases a one-stage or two-stage axially-conveying stirrer system is mainly used. This generates a flow profile which is essentially parallel to the rotary shaft of the employed stirrer. Thus, in the arrangement as shown in Figure 2 with the dialysis module parallel to the rotary shaft a direct flow against the dialysis module is not achieved. This has a disadvantageous effect on the mass transport in the dialysis module (wider outer laminar boundary layer on the fiber surface).

[0044] A direct tangential or radial flow against the dialysis membrane has an advantageous effect which can for example be achieved by a standard anchor impeller. This impeller generates a flow which is directed directly onto the dialysis module or modules in the reactor and thus reduces the laminar boundary layer on the surface of the dialysis module(s). This simple radial flow is, however, disadvantageous for the other basic technical process functions in particular with regard to mixing the reactor and mass transfer especially in submersed gased reactors. The gas can be introduced into the cultivation vessel e.g. via a pipe sparger or a ring sparger.

[0045] It has been found that the agitator (combination stirrer) as reported herein provides for a direct orthogonal flow against or towards the dialysis module and at the same time is suitable for mixing in liquids at/from the surface of the culture medium and for rapid total mixing of the culture medium whereby the shear stress for the cells in the reactor is almost not increased when compared to other stirring systems. It has turned out that the axial flow generated by the axially-conveying element(s), i.e. for example by inclined-blade stirrers, ensures a mixing of the culture medium in a short time. In addition shear sensitive cells such as mammalian cells can be cultivated at the same shear stress with increased mixing efficiency by using the agitator as reported herein.

[0046] The agitator as reported herein combines or integrates the properties of a radially-conveying element, i.e. for example of an anchor impeller, which are important for an application in dialysis processes with those of an axially-conveying element, such as e.g. an inclined-blade stirrer, which are important to fulfill the basic technical process requirements or functions. Exemplary embodiments of the agitator as reported herein are shown in Figures 1a) to 1f) in which one or more of the connecting elements of a radially-conveying element (anchor impeller) are designed as axially-conveying element (inclined-blade stirrer). Specific embodiments are shown in Figures 1a) to 1f).

[0047] The rotary speed of the stirrer  $n$  is used as a characteristic velocity and the stirrer width  $d$  is used as a characteristic length.

[0048] In fluid dynamics the Reynolds's number describes the ratio of inertial force to inner frictional force in a hydrodynamic system. Therefrom also statements can be made about the degree of turbulence of the moved medium. For stirred liquids the stirrer Reynolds's number is defined in equation 4 to be:

$$Re = \frac{n \cdot d^2}{\nu} = \frac{n \cdot d^2 \cdot \rho}{\eta} \quad . \quad \text{(Equation 4)}$$

[0049] The Newton number (also referred to as power number) describes the ratio of resistance force to flow force and is thus a measure for the flow resistance of a stirrer in a stirred material and is described in equation 5:

$$Ne = \frac{P}{\rho \cdot n^3 \cdot d^5} \quad . \quad \text{(Equation 5)}$$

[0050] Agitators with a low Newton number, such as propeller or inclined blade stirrers, convert the power input more efficiently in hydrodynamic output, i.e. fluid motion, than those with a high Newton number, such as rushton turbines.

[0051] A criterion for assessing the stirring processes in culture or cultivation processes is the mixing time. The "mixing time" in an inhomogeneous liquid-liquid mixture denotes the time which is required to achieve a defined homogeneity in the culture medium. Factors influencing the mixing time are the degree of mixing and the site of observation. The degree of mixing in turn depends on the reactor geometry, the stirrer geometry, the rotary frequency of the stirrer, and the substances of the stirred materials.

[0052] It is important for culture or cultivation processes that, as far as possible, all cells are optimally and uniformly supplied with the necessary substrates (such as nutrient medium,  $O_2$ ) and that metabolites (such as overflow products,  $CO_2$ ) are concomitantly led away. This means that repositories and sinks that may occur spatially as well as temporarily in the reactor have to be avoided or minimized in order to avoid damage to the cells. This can be achieved e.g. by using an adapted stirring system for mixing the reactor's contents. The mixing processes can be divided into the sub-processes micro-mixing and macro-mixing. Micro-mixing is defined as the molecular concentration adjustment due to diffusion or microturbulences; in contrast macro-mixing is defined as the convective coarse mixing caused by the stirrer (see e.g.



obtained and thus this is the system with the lowest shear at a constant power input. Thus, at the same shear forces for example for the cultured cells it is possible to achieve a higher power input which, due to a higher number of revolutions, leads to higher turbulences and a higher flow against the dialysis module as well as to a better complete mixing.

**[0060]** In addition to NaCl, glucose was also used as a tracer substance to experimentally determine mass transfer coefficients. In general it could be shown that the mass transfer of the dialysis module is, on the one hand, influenced by the power input and, on the other hand, by the stirrer geometry, i.e. the primary generated flow mode. Tangential or radial flow profiles have proven to be particularly suitable with regard to reducing the outer laminar boundary layers around the hollow fiber dialysis membranes (higher mass transfer rates). The highest mass transfer coefficients can be achieved in each case by the agitator as reported herein.

**[0061]** Figure 7 shows a comparison of the Newton numbers. As can be seen the agitator as reported herein provides for a considerably higher Newton number. At the same time it can be seen that the power number is independent of the Reynolds number. Thus, the agitator provides for an increased power number independent of the Reynolds number when operated to produce a turbulent flow within the reaction vessel.

**[0062]** Figure 8 shows that the highest mass transfer coefficients of the dialysis module at a constant volume-specific power input can be achieved with the agitator as reported herein. This is due to the radial flow generated by the stirrer as reported herein which, in comparison to the standard stirrer systems, allows a better flow against the hollow fiber dialysis module.

**[0063]** Compared to standard stirrer systems the agitator as reported herein provides for considerable advantages in the mixing (Figure 5, Table 1), the generated shear stress (Figure 6), as well as in the mass transfer coefficients in dialysis (Figure 8). It is particularly remarkable that, despite the introduced modifications, no significantly higher shear stresses can be measured (see e.g. Pohlscheidt, M., et. al., Chem. Ing. Tec. 80 (2008) 821-830).

**[0064]** The abbreviations used in this application have the following meanings (see also Figure 1a):

b: width of the blades of the radially-conveying element

d: agitator total outer diameter

$d_w$ : diameter of the shaft

h: height of the stirrer blades of the radially-conveying element

$h_m$ : height of the fastening sleeve

$h_{SB}$ : height of an axially-conveying element

$h_u$ : height of the reducer

$\Delta h$ : height difference of two axially-conveying elements

l: length of the stirrer blades of an axially-conveying element

$\alpha$ : blade pitch of the blades of an axially-conveying element

z: number of stirrer blades per stirrer

$d_j$ : inner distance between the stirrer blades of the radially-conveying element

$h_{4/5}$ : 4/5 height from above of h

K: stirrer head, i.e. the highest point of the stirrer when it is not attached to a rotary shaft

D: cultivation vessel inner diameter

H: filling height of the cultivation vessel.

**[0065]** In one embodiment the ratio of the height difference ( $\Delta h$ ) of two axially-conveying elements to the cultivation vessel diameter (D) is at least 0.75.

**[0066]** Herein is reported the use of the agitator as reported herein for the culture or cultivation of cells for the recombinant production of proteins or antibodies as an aspect. In one embodiment the culture is a dialysis. In a further embodiment the culture is carried out in a submerged gassed stirred tank reactor. In another embodiment the cell is a eukaryotic cell, in another embodiment a mammalian cell. In yet a further embodiment the cell is a CHO cell, a BHK cell, an NS0 cell, a COS cell, a PER.C6 cell, a Sp2/0 cell or a HEK 293 cell. In one embodiment the cell is selected from *Arthrobacter protophormiae*, *Aspergillus niger*, *Aspergillus oryzae*, *Bacillus amyloliquefaciens*, *Bacillus subtilis*, BHK cells, *Candida boidinii*, *Cellulomonas cellulans*, *Corynebacterium lilium*, *Corynebacterium glutamicum*, CHO cells, *E.coli*, *Geobacillus stearothermophilus*, *H. polymorpha*, HEK cells, HeLa cells, *Lactobacillus delbrueckii*, *Leuconostoc mesenteroides*, *Micrococcus luteus*, MDCK cells, *Paenebacillus macerans*, *P. pastoris*, *Pseudomonas* species, *S. cerevisiae*, *Rhodobacter* species, *Rhodococcus erythropolis*, *Streptomyces* species, *Streptomyces anulatus*, *Streptomyces hygroscopicus*, Sf-9 cells, and *Xantomonas campestris*. In yet a further embodiment the antibody is an antibody against CD20, CD22, HLA-DR, CD33, CD52, EGFR, G250, GD3, HER2, PSMA, CD56, VEGF, VEGF2, CEA, Lewis Y antigen, IL-6 receptor or IGF-1 receptor.

**[0067]** The following example and figures are provided to illustrate the subject matter of the invention. The protective scope is defined by the attached patent claims

**Description of the figures****[0068]**

- 5 **Figure 1** Schematic diagram of various embodiments of the combination stirrer as reported herein; b: width of the stirrer blade; d: stirrer diameter;  $d_w$ : diameter of the rotary shaft; h: height of the stirrer blade of the radially-conveying stirrer;  $h_m$ : height of the fastening sleeve;  $h_{SB}$ : height of the axially-conveying stirrer;  $h_u$ : height of the reducer; l: length of the stirrer blade of the axially-conveying stirrer;  $\alpha$ : blade pitch of the axially-conveying stirrer; z: number of stirrer blades per stirrer;  $d_i$ : inner distance between the stirrer blades of the radially-conveying stirrer;  $h_{4/5}$ : 4/5 height from above of h; K: stirrer head; a) general scheme of the agitator as reported herein; b) to f) embodiments of the agitator as reported herein; g) scheme of a radially conveying element seen from the top along the shaft axis; h) scheme of an axially conveying element seen from the top along the shaft axis; i) scheme of an embodiment of the agitator as reported herein seen from the top along the shaft axis showing the connection of opposing stirrer blades of the radially-conveying element via connecting stirrer blades of the axially-conveying element; j) scheme of an embodiment of the agitator as reported herein seen from the top along the shaft axis showing the connection of opposing stirrer blades of the radially-conveying element via connecting stirrer blades of the axially-conveying element wherein the diameter of the axially-conveying element is less than the inner diameter of the radially-conveying element and the spatial distance is bridged by a connector.
- 10
- 15
- 20 **Figure 2** Schematic diagram of a device for dialysis cultivation.  
**Figure 3** Schematic diagram of the concentration gradients on the hollow fibers of the dialysis module.  
**Figure 4** Typical concentration time course in the reactor ( $C_R$ ) and storage container ( $C_V$ ).  
**Figure 5** Comparison of the mixing coefficients  $C_H$  for different stirrer as a function of the Reynolds's number (Re); KR = combination stirrer as reported herein; SSR = standard disk stirrer; SBR = inclined-blade stirrer.
- 25 **Figure 6** Reference flake diameter  $d_{VF}$  as a function of the volume-specific power input and stirrer configuration; KR = combination stirrer as reported herein; SSR = standard disk stirrer; SBR = inclined-blade stirrer; Pohlsc-heidt, M., et al. = Chem. Ing. Tec. 80 (2008) 821-830.
- Figure 7** Diagram of the power coefficient Ne of different stirrer as a function of the Reynolds's number (Re); KR = combination stirrer as reported herein; SSR = standard disk stirrer; SBR = inclined-blade stirrer.
- 30 **Figure 8** Mass transfer coefficient  $k_a$  in the dialysis as a function of the specific power input; KR = combination stirrer as reported herein; SBR = inclined-blade stirrer.

**Example 1****Cultivation vessel**

**[0069]** All investigations were carried out in a 100 l Plexiglas® model container (referred to as DN 440 in the following).

**Example 2**

40

**Power input**

**[0070]** The power input of different stirrer was determined by measuring the torque on the rotary shaft. A data processing system model GMV2 together with the torque sensor model DRFL-II-5-A (both from the "ETH Messtechnik" Company, Gschwend, Germany) were used to record the torque. For each stirrer system the torque was firstly recorded at various revolution speeds in the unfilled state ( $M_{empty}$ ) and subsequently by means of a triplicate determination in the filled state ( $M_{load}$ ) according to equation 9:

$$M = M_{load} - M_{empty} \quad . \quad \text{(Equation 9).}$$

**[0071]** Afterwards the corresponding Newton number (Ne number) and the Reynolds's number (Re number) were calculated for each point. Since the Newton numbers for a stirrer system become constant in the turbulent flow region, the calculated Newton numbers were subsequently averaged in this region (Uhl, V.W. and Gray, J.B., Mixing Theory and Practice, Academic Press, 1966). This mean represents the total Newton number of the respective stirrer.

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**Example 3****Homogenization**

5 **[0072]** The homogenization was determined using the color change method as well as using the conductivity method.  
**[0073]** The color change method is based on the decolorization of a starch solution stained with iodine-potassium iodide by addition of sodium thiosulfate (I, KI, starch, Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> obtained from the Carl Roth GmbH & Co KG Company, Karlsruhe, Germany). A one molar sodium thiosulfate solution and a one molar iodine-potassium iodide solution (Lugol's solution) as well as a starch solution at a concentration of 10 g/l were used as the starting solutions. In correspondence  
10 with the conductivity experiments, at least four speed steps were examined per stirrer (quadruplicate determinations per speed step) in which a maximum of four experiments were carried out per container filling amount. In each case the starch solution was added once per filling of the container. For each individual measurement the corresponding volume of the iodine-potassium iodide solution was firstly added and subsequently the sodium thiosulfate was added. The mixing time has been determined manually from the time point at which the sodium thiosulfate was added and one second was  
15 subtracted in each case in order to take the addition time into consideration. After completion of the measurement the container filling volume was titrated (neutralized) with iodine-potassium iodide in order to compensate for the excess of the previously added sodium thiosulfate.  
**[0074]** In the conductivity method the mixing time is defined as the time from addition of an electrolyte solution to the time at which the measured conductivity fluctuations for the last time exceed a tolerance range of  $\pm 5\%$  around the  
20 conductivity values which are reached in a stationary state. If several probes are used, the longest detected mixing time in each case is regarded as representative for the entire system.  
**[0075]** A 30 % (w/v) NaCl solution (NaCl crystalline, Merck KGaA Company, Darmstadt, Germany) was used as an electrolyte solution to determine the mixing time by the conductivity method. This was added in pulses onto the liquid surface at the rotary shaft of the stirrer and the volume per addition was selected such that the jumps in conductivity  
25 which resulted in a stationary state did not exceed 200 mS/cm.  
**[0076]** For each stirrer at least four speed steps were examined. The mixing time was determined at least eight times per speed step and these eight values were averaged. The mixing coefficient of the respective stirrer systems is given as the mean of the mixing coefficients averaged per speed step. The conductivity was in each case measured by three  
30 4-pole conductivity probes (TetraCon, WTW Company, Weilheim) at various radial or axial positions in the container. The conductivity signals were read out online via the measuring amplifier that was used (Cond813, Knick "Elektronische Messgeräte GmbH & Co, KG" Company, Berlin, Germany). The measured values were stored online and simultaneously for all probes by means of the software Paraly SW 109 (Knick "Elektronische Messgeräte GmbH & Co, KG" Company, Berlin, Germany) at a sampling rate of 5 seconds. After the series of measurements was completed the data were  
35 evaluated separately for each probe.

**Example 4****Shear stress**

40 **[0077]** A model particle system, the blue clay polymer flake system, was used to determine shear stress. This is a model particle system consisting of a cationic polymer (Praestol BC 650) and a clay mineral (blue clay) which is placed in the vessel. A flocculation reaction is started by adding Praestol BC 650 which generates flakes of a defined size. These flakes are subsequently broken up by the mechanical and hydrodynamic stress of the stirrer system. In the case  
45 of bubble-gassed systems they are additionally broken up by the energy dissipation when the bubbles are formed and burst. The average particle diameter of the model particle system was used as a measured variable to characterize the shear stress. In this case the change in the particle size was measured in situ by a Focused Beam Reflectance Measurement Probe from the Mettler Toledo Company (referred to as FBRM® in the following). The rates of change in particle  
50 size that were determined are a measure for the shear stress prevailing in the model system. The gradient of the rate of change of particle size becomes smaller during the course of the experiment but no equilibrium state is formed (particle comminution down to a diameter of the blue clay primary particles of  $\approx 15 \mu\text{m}$ ). For this reason an end flake diameter  $d_{P50}$  for the blue clay polymer flake system was determined according to the following criterion (equation 10):

$$55 \quad \frac{d(d_{P50})}{dt} \leq 0.0055 [\mu\text{m} / \text{s}] \quad \rightarrow \quad d_{P50} = d_{P50}' \quad (\text{Equation 10}).$$

**[0078]** In order to ensure comparability of the end flake diameters at different power inputs and between different

stirrers, the reference flake diameter was calculated as follows (equations 11 to 13):

$$d_{VF} = m \cdot d_{P50}^1 - b \quad \text{(Equation 11)}$$

$$m = 1,3 \cdot 10^{-6} \cdot \left(\frac{P}{V}\right)^2 + 1,37 \cdot 10^{-3} \cdot \left(\frac{P}{V}\right) + 2,46 \quad \text{(Equation 12)}$$

$$b = 8,12 \cdot 10^{-5} \cdot \left(\frac{P}{V}\right)^2 + 6,48 \cdot 10^{-3} \cdot \left(\frac{P}{V}\right) + 76,9 \quad \text{(Equation 13)}$$

**Table 2:** Substances used to determine the particle stress (concentrations are based on the container fill volume).

Ingredient	Concentration	Manufacturer
Wittschlicker blue clay	5 g/l	Braun Tonbergbau Co., Germany
Praestol 650 BC (solution 2 g/l)	5 ml/l	Stockhaus GmbH & Co. KG, Krefeld, Germany
NaCl	1 g/l	Merck KGaA, Darmstadt, Germany
CaCl <sub>2</sub> (solution 30 g/l)	--	Carl Roth GmbH & Co.KG, Karlsruhe, Germany

**[0079]** Firstly the 100 l model container was filled with a corresponding volume (H/D ratio) of completely demineralized water (VE water) and maintained at a temperature of 20°C. Subsequently the conductivity was adjusted to a value of 1000 µS/cm by titration with a CaCl<sub>2</sub> solution. The conductivity was measured by a 4-pole conductivity probe (probe: TetraCon, WTW Co. Weilheim; measuring amplifier: Cond813, Knick "Elektronische Messgeräte GmbH & Co, KG" Company, Berlin, Germany). Afterwards the blue clay and the NaCl were added in appropriate amounts to the solution. Subsequently a homogenization phase took place at the highest speed with a duration of at least 20 minutes. The FBRM® probe (FBRM® Lasentec® D600L, Mettler-Toledo GmbH Co., Giessen, Germany) was mounted in the container perpendicular from above (immersion depth 300 mm) at a radial distance of 70 mm to the wall. The flocculation reaction was subsequently started by adding Praestol 650 BS at a defined speed. The measured values were recorded online by means of the program data acquisition control interface version 6.7.0 (Mettler-Toledo GmbH, Giessen, Germany). The reference flake diameter was determined from the measurement data. At least three power inputs were measured for each stirrer. In each case three measurements were carried out per power input.

#### **Example 5**

##### **Dialysis (mass transfer liquid - liquid)**

**[0080]** A NaCl solution (NaCl crystalline, Merck KGaA Company, Darmstadt, Germany) was used as a tracer substance to determine the concentration half-life of the module (DIADYN-DP 070 F1 OL; MICRODYN-NADIR GmbH Company, Wiesbaden, Germany) in relation to the stirrer system that was used and the volume-specific power input. The tracer substance was adjusted in the storage container at the start of each experimental run to a base-line conductivity of 1500 µS/cm. The reactor was filled with completely demineralized water for each experimental run. The filling volume of the reactor was 100 l (H/D = 1.6) and that of the storage container was 400 l (H/D = 2.0) and both containers were maintained at a temperature of 20°C at the start of each experiment. The conductivity in both containers was measured by a 4-pole conductivity probe (probe: TetraCon, WTW Co. Weilheim, Germany; measuring amplifier: Cond813, Knick "Elektronische Messgeräte GmbH & Co, KG" Company, Berlin, Germany). The sampling rate of the measurement amplifiers that were used was 5 seconds and the measurement values were stored online and simultaneously for all probes by means of

the software Paraly SW 109 (Knick "Elektronische Messgeräte GmbH & Co, KG" Company, Berlin, Germany). The NaCl solution was circulated by means of a peristaltic pump between supply container and dialysis module (housing pump 520 U, Watson-Marlow GmbH, Company, Rommerskirchen, Germany) at a constant flow rate of 2.1 l/min. For the evaluation, probe 1 was used as a reference probe for the reactor and probe 3 was used as a reference probe for the storage container. The data of these two probes were evaluated by an evaluation routine. In each case at least six different power inputs in the reactor were investigated per stirrer.

**[0081]** In order to compare the mass transfer characteristics determined by means of the NaCl solution, additional measurements were carried out with a glucose solution as tracer substance. The experimental setup was not changed for this. A defined glucose concentration (glucose solid, Merck KGaA Company, Darmstadt, Germany) at a concentration of 3 g/l was provided in the storage container. The glucose concentration was determined manually and simultaneously for the storage container and reactor at a time interval of 10 minutes by means of a blood sugar measuring instrument (ACCU-CHEK® Aviva, Roche Diagnostics GmbH Company, Mannheim, Germany).

## Claims

1. Device comprising an agitator and a dialysis module within a cultivation vessel, **characterized in that** the agitator comprises
  - one radially-conveying element comprising at least two stirrer blades, and
  - one or more axially-conveying elements each comprising at least two stirrer blades, wherein the stirrer blades of the radially-conveying element are parallel to each other, wherein the outer diameter of all axially-conveying elements is equal to or less than the inner diameter of the radially-conveying element,
  - wherein all axially-conveying elements are individually connected to the radially-conveying element, wherein all axially-conveying elements are located within the radially-conveying element, and wherein all conveying elements have a fixed spatial orientation relative to each other.
2. Device according to claim 1, **characterized in that** the number of axially-conveying elements is 1 or 2 or 3.
3. Device according to any one of the previous claims, **characterized in that** opposing stirrer blades of the radially-conveying element are linked to each other by two opposite stirrer blades of an axially-conveying element.
4. Device according to any one of the previous claims, **characterized in that** the radially-conveying element is an anchor impeller.
5. Device according to any one of the previous claims, **characterized in that** the axially-conveying element is an inclined-blade stirrer.
6. Device according to claim 5, **characterized in that** the pitch of the stirrer blades of the inclined-blade stirrer is between 10° and 80° relative to the shaft axis of the agitator.
7. Device according to any one of the preceding claims, **characterized in that** the radially-conveying element has a height of at least 200 mm.
8. Device according to any one of claims 1 to 7, **characterized in that** the ratio d/D of agitator diameter (d) to cultivation vessel diameter (D) is of from 0.2 to 0.8.
9. Device according to any one of claims 1 to 8, **characterized in that** the ratio H/D of filling height of the cultivation vessel (H) to cultivation vessel width (D) is of from 1.0 to 2.5.
10. Use of a device according to any one of claims 1 to 9 for cultivating a cell expressing a polypeptide.
11. Use according to claim 10, characterized that the cell is a mammalian cell.
12. Use according to claim 11, **characterized in that** the mammalian cell is selected from a CHO cell, a BHK cell, an NS0 cell, a COS cell, a PER.C6 cell, a Sp2/0 cell, or a HEK 293 cell.

13. Use according to any one of claims 10 to 12, **characterized in that** the polypeptide is an antibody.
14. Method for the production of a polypeptide comprising
- 5 a) providing a cell comprising a nucleic acid encoding the polypeptide,  
b) providing a device as reported in any one of claims 1 to 9,  
c) cultivating the cell in the device in a cultivation medium wherein the agitator provides a turbulent flow within  
the cultivation vessel, and  
10 d) recovering the polypeptide from the cells or the cultivation medium and thereby producing a polypeptide.
15. Method according to claim 14, characterized that the cell is a mammalian cell.
16. Method according to claim 15, **characterized in that** the mammalian cell is selected from a CHO cell, a BHK cell,  
15 an NS0 cell, a COS cell, a PER.C6 cell, a Sp2/0 cell, or a HEK 293 cell.
17. Method according to any one of claim 14 to 16, **characterized in that** the polypeptide is an antibody.

### Patentansprüche

- 20 1. Vorrichtung, umfassend ein Rührwerk und ein Dialysem modul innerhalb eines Kultivierungsgefäßes, **dadurch gekennzeichnet, dass** das Rührwerk
- 25 - ein Radialförderelement, umfassend wenigstens zwei Rührschaufeln, und  
- ein oder mehr Axialförderelemente, die jeweils wenigstens zwei Rührschaufeln umfassen,  
umfasst,  
wobei die Rührschaufeln des Radialförderelements zueinander parallel sind,  
wobei der Außendurchmesser aller Axialförderelemente gleich dem oder kleiner als der Innendurchmesser des  
30 Radialförderelements sind,  
wobei alle Axialförderelemente einzeln mit dem Radialförderelement verbunden sind,  
wobei alle Axialförderelemente innerhalb des Radialförderelements angeordnet sind und  
wobei alle Förderelemente eine unveränderliche räumliche Orientierung relativ zueinander aufweisen.
- 35 2. Vorrichtung gemäß Anspruch 1, **dadurch gekennzeichnet, dass** die Anzahl der Axialförderelemente 1 oder 2 oder 3 beträgt.
- 40 3. Vorrichtung gemäß einem der vorstehenden Ansprüche, **dadurch gekennzeichnet, dass** gegenüberliegende Rührschaufeln des Radialförderelements durch zwei gegenüberliegende Rührschaufeln eines Axialförderelements miteinander verbunden sind.
- 45 4. Vorrichtung gemäß einem der vorstehenden Ansprüche, **dadurch gekennzeichnet, dass** das Radialförderelement ein Anker-Rührer ist.
5. Vorrichtung gemäß einem der vorstehenden Ansprüche, **dadurch gekennzeichnet, dass** das Axialförderelement ein Rührwerk mit geneigten Schaufeln ist.
6. Vorrichtung gemäß Anspruch 5, **dadurch gekennzeichnet, dass** die Neigung der Rührschaufeln des Rührwerks mit geneigten Schaufeln zwischen 10° und 80° bezogen auf die Wellenachse des Rührwerks beträgt.
- 50 7. Vorrichtung gemäß einem der vorstehenden Ansprüche, **dadurch gekennzeichnet, dass** das Radialförderelement eine Höhe von wenigstens 200 mm aufweist.
8. Vorrichtung gemäß einem der Ansprüche 1 bis 7, **dadurch gekennzeichnet, dass** das Verhältnis  $d/D$  des Durchmessers des Rührwerks ( $d$ ) zu dem Durchmesser des Kultivierungsgefäßes ( $D$ ) von 0,2 bis 0,8 beträgt.
- 55 9. Vorrichtung gemäß einem der Ansprüche 1 bis 8, **dadurch gekennzeichnet, dass** das Verhältnis  $H/D$  der Füllhöhe des Kultivierungsgefäßes ( $H$ ) zu der Breite des Kultivierungsgefäßes ( $D$ ) von 1,0 bis 2,5 beträgt.

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10. Verwendung einer Vorrichtung gemäß einem der Ansprüche 1 bis 9 zum Kultivieren einer Zelle, die ein Polypeptid exprimiert.

11. Verwendung gemäß Anspruch 10, **dadurch gekennzeichnet, dass** die Zelle eine Säugerzelle ist.

12. Verwendung gemäß Anspruch 11, **dadurch gekennzeichnet, dass** die Säugerzelle ausgewählt ist aus einer CHO-Zelle, einer BHK-Zelle, einer NSO-Zelle, einer COS-Zelle, einer PER.C6-Zelle, einer Sp2/0-Zelle und einer HEK-293-Zelle.

13. Verwendung gemäß einem der Ansprüche 10 bis 12, **dadurch gekennzeichnet, dass** das Polypeptid ein Antikörper ist.

14. Verfahren für die Herstellung eines Polypeptids, umfassend

a) Bereitstellen einer Zelle, die eine Nukleinsäure umfasst, die das Polypeptid kodiert,

b) Bereitstellen einer Vorrichtung gemäß einem der Ansprüche 1 bis 9,

c) Kultivieren der Zelle in der Vorrichtung in einem Kulturmedium, wobei das Rührwerk eine turbulente Strömung innerhalb des Kulturgefäßes erbringt, und

d) Gewinnung des Polypeptids aus den Zellen oder dem Kulturmedium, um so ein Polypeptid herzustellen.

15. Verfahren gemäß Anspruch 14, **dadurch gekennzeichnet, dass** die Zelle eine Säugerzelle ist.

16. Verfahren gemäß Anspruch 15, **dadurch gekennzeichnet, dass** die Säugerzelle ausgewählt ist aus einer CHO-Zelle, einer BHK-Zelle, einer NSO-Zelle, einer COS-Zelle, einer PER.C6-Zelle, einer Sp2/0-Zelle und einer HEK-293-Zelle.

17. Verfahren gemäß einem der Ansprüche 14 bis 16, **dadurch gekennzeichnet, dass** das Polypeptid ein Antikörper ist.

### Revendications

1. Dispositif comprenant un agitateur et un module de dialyse dans un récipient de culture, **caractérisé en ce que** l'agitateur comprend :

- un élément à transport radial comprenant au moins deux pales d'agitateur, et

- un ou plusieurs éléments à transport axial comprenant chacun au moins deux pales d'agitateur,

dans lequel les pales d'agitateur de l'élément à transport radial sont parallèles les unes aux autres,

dans lequel le diamètre extérieur de tous les éléments à transport axial est égal ou inférieur au diamètre intérieur de l'élément à transport radial,

dans lequel tous les éléments à transport axial sont connectés individuellement à l'élément à transport radial,

dans lequel tous les éléments à transport axial sont situés dans l'élément à transport radial, et

dans lequel tous les éléments de transport ont une orientation spatiale fixe les uns par rapport aux autres.

2. Dispositif suivant la revendication 1, **caractérisé en ce que** le nombre d'éléments à transport axial est égal à 1 ou 2 ou 3.

3. Dispositif suivant l'une quelconque des revendications précédentes, **caractérisé en ce que** les pales d'agitateur opposées de l'élément à transport radial sont reliées les unes aux autres par deux pales d'agitateur opposées de l'élément à transport axial.

4. Dispositif suivant l'une quelconque des revendications précédentes, **caractérisé en ce que** l'élément à transport radial est un agitateur de type ancre.

5. Dispositif suivant l'une quelconque des revendications précédentes, **caractérisé en ce que** l'élément à transport axial est un agitateur à pales inclinées.

6. Dispositif suivant la revendication 5, **caractérisé en ce que** le pas des pales d'agitateur de l'agitateur à pales inclinées est compris entre 10° et 80° par rapport à l'axe d'arbre de l'agitateur.

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7. Dispositif suivant l'une quelconque des revendications précédentes, **caractérisé en ce que** l'élément à transport radial a une hauteur d'au moins 200 mm.
- 5 8. Dispositif suivant l'une quelconque des revendications 1 à 7, **caractérisé en ce que** le rapport  $d/D$  du diamètre de l'agitateur ( $d$ ) au diamètre du récipient de culture ( $D$ ) va de 0,2 à 0,8.
9. Dispositif suivant l'une quelconque des revendications 1 à 8, **caractérisé en ce que** le rapport  $H/D$  de la hauteur de remplissage du récipient de culture ( $H$ ) à la largeur du récipient de culture ( $D$ ) va de 1,0 à 2,5.
- 10 10. Utilisation d'un dispositif suivant l'une quelconque des revendications 1 à 9, pour la culture d'une cellule exprimant un polypeptide.
11. Utilisation suivant la revendication 10, **caractérisée en ce que** la cellule est une cellule de mammifère.
- 15 12. Utilisation suivant la revendication 11, **caractérisée en ce que** la cellule de mammifère est choisie entre une cellule CHO, une cellule BHK, une cellule NS0, une cellule COS, une cellule PER.C6, une cellule Sp2/0 et une cellule HEK 293.
- 20 13. Utilisation suivant l'une quelconque des revendications 10 à 12, **caractérisée en ce que** le polypeptide est un anticorps.
14. Procédé pour la production de polypeptide, comprenant :
- 25 a) la fourniture d'une cellule comprenant un acide nucléique codant pour le polypeptide,  
b) la fourniture d'un dispositif tel que décrit dans l'une quelconque des revendications 1 à 9,  
c) la culture de la cellule dans le dispositif dans un milieu de culture, dans lequel l'agitateur engendre un courant turbulent à l'intérieur du récipient de culture, et  
d) la récupération du polypeptide à partir des cellules ou du milieu de culture et, ainsi, la production d'un polypeptide.
- 30 15. Procédé suivant la revendication 14, **caractérisé en ce que** la cellule est une cellule de mammifère.
16. Procédé suivant la revendication 15, **caractérisé en ce que** la cellule de mammifère est choisie entre une cellule CHO, une cellule BHK, une cellule NS0, une cellule COS, une cellule PER.C6, une cellule Sp2/0 et une cellule HEK 293.
- 35 17. Procédé suivant l'une quelconque des revendications 14 à 16, **caractérisé en ce que** le polypeptide est un anticorps.

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Figure 1

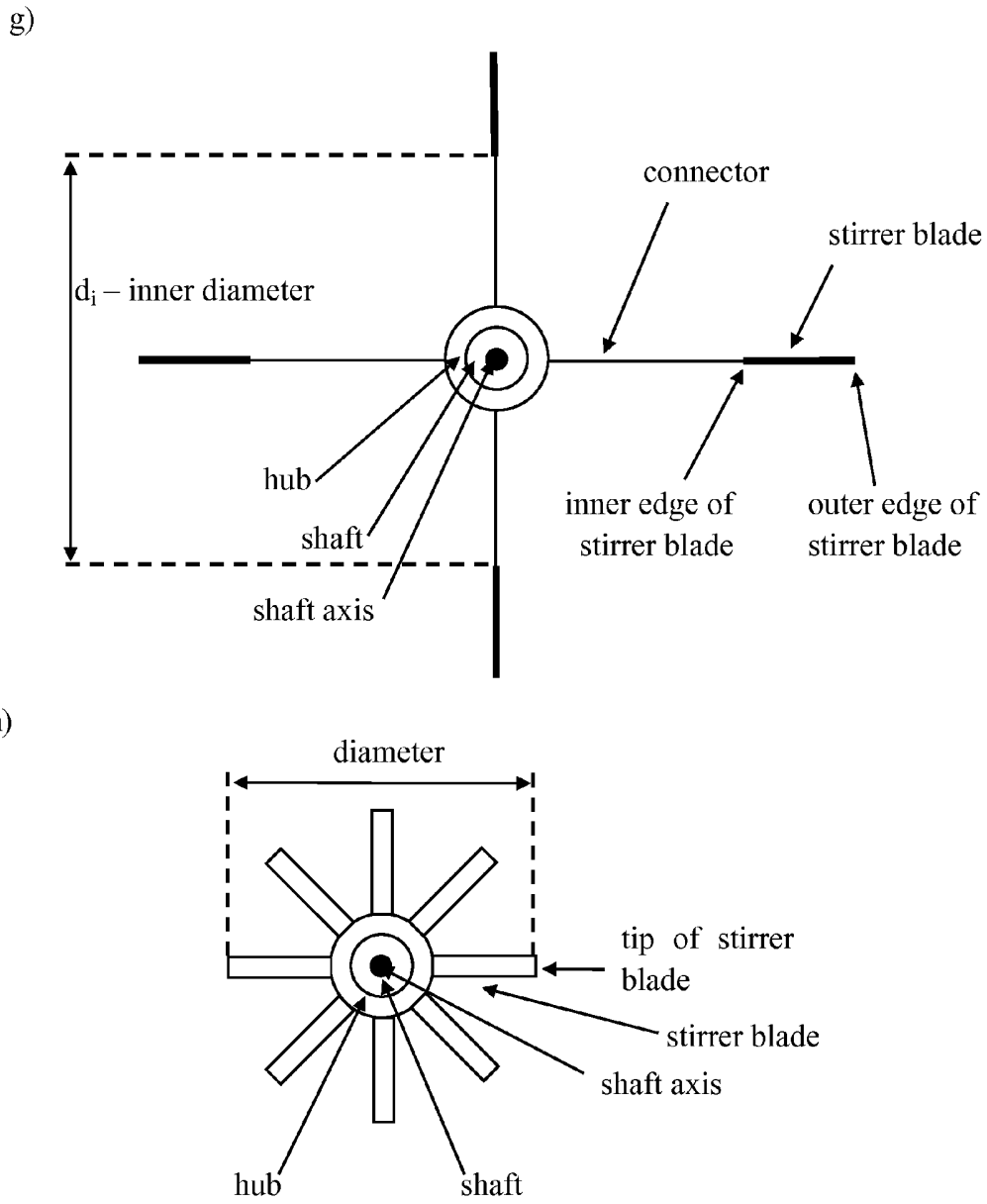
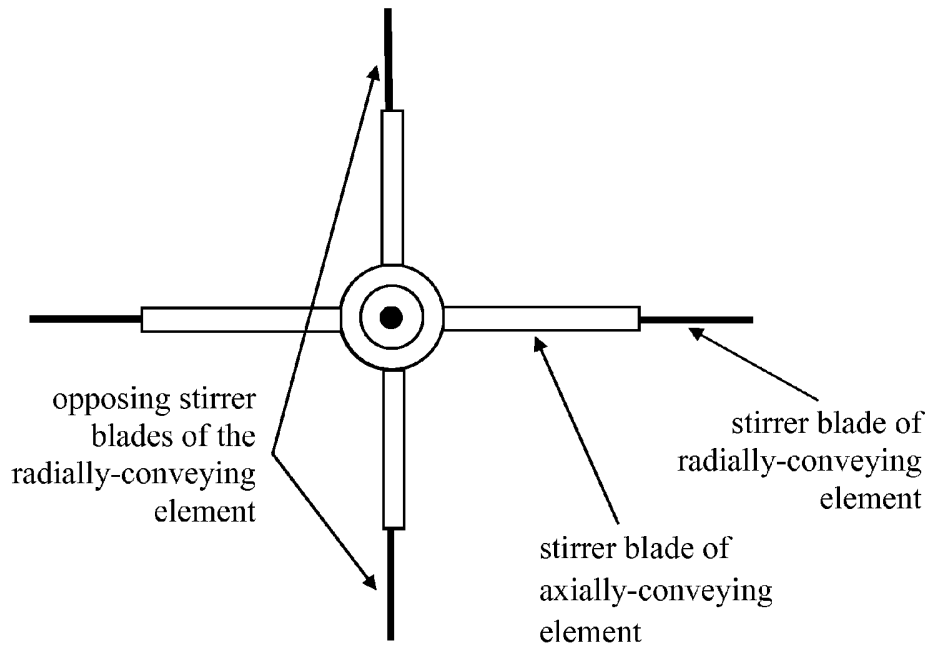


Figure 1

i)



j)

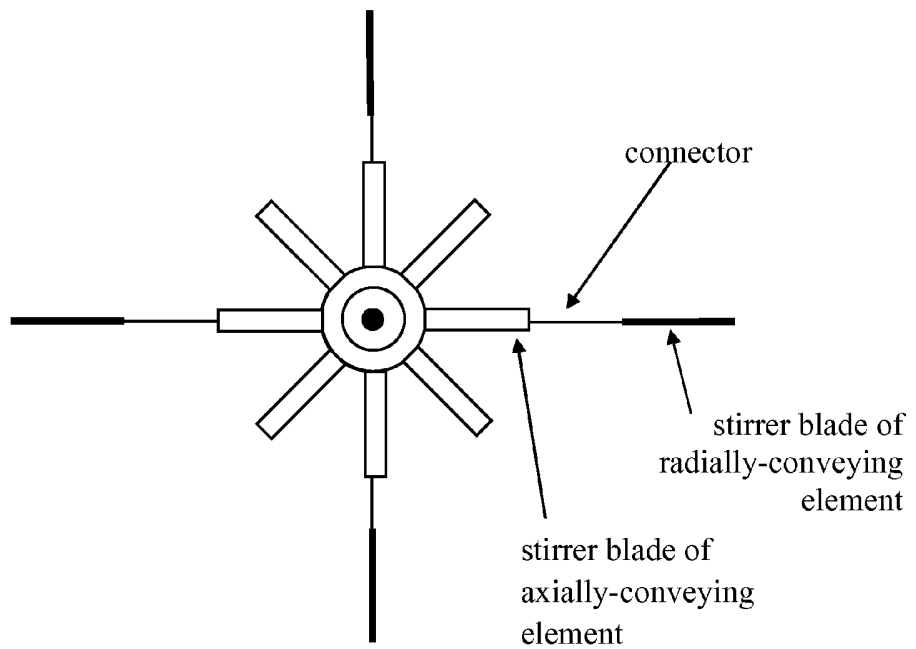


Figure 2

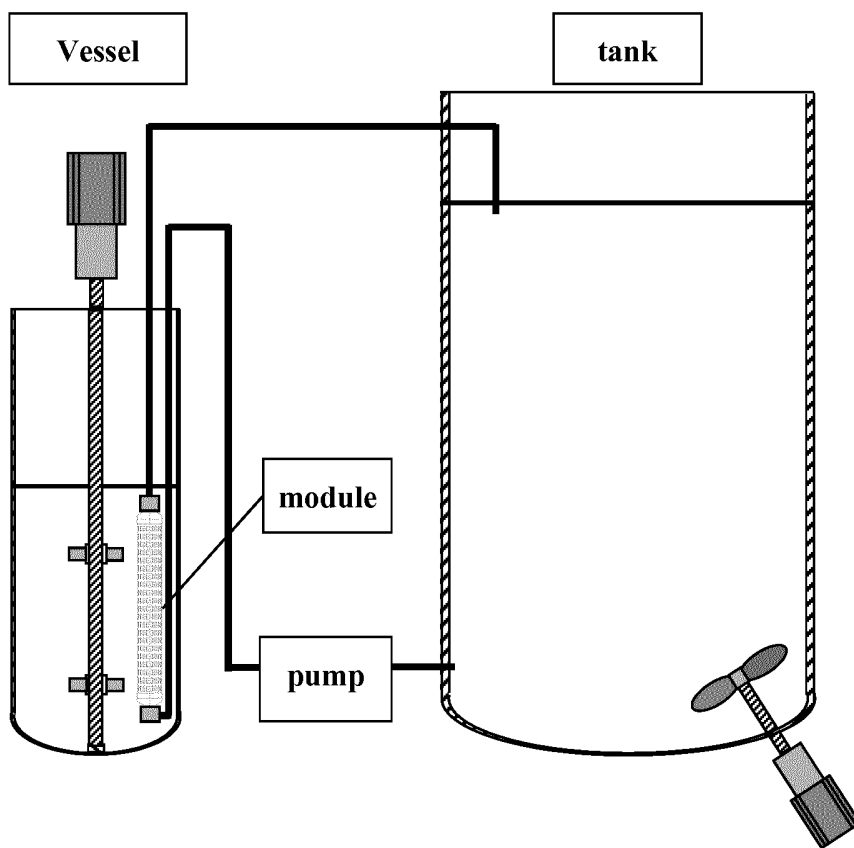


Figure 3

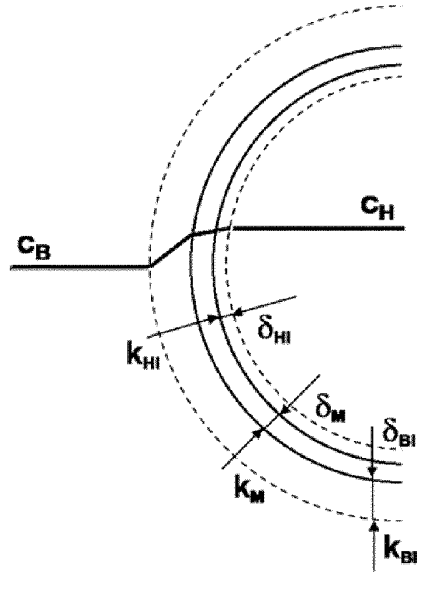


Figure 4

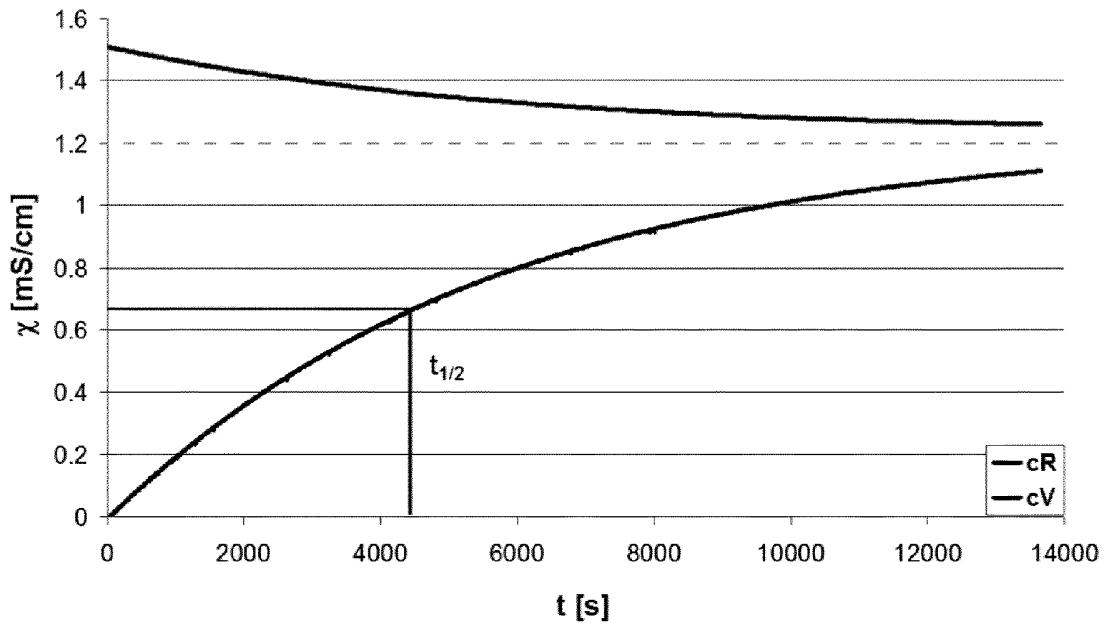


Figure 5

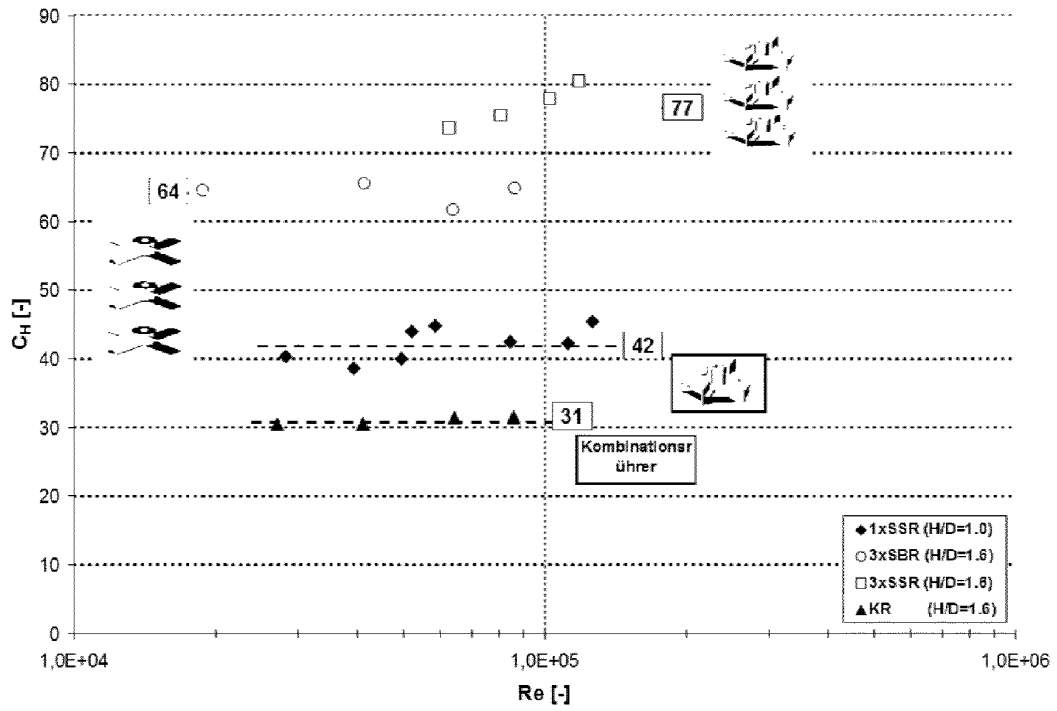


Figure 6

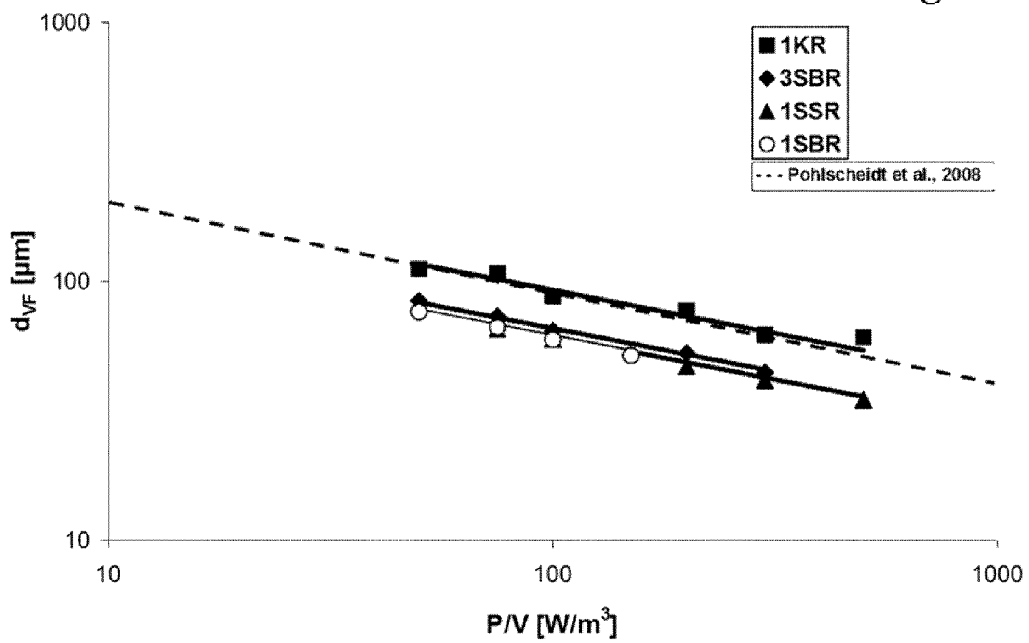


Figure 7

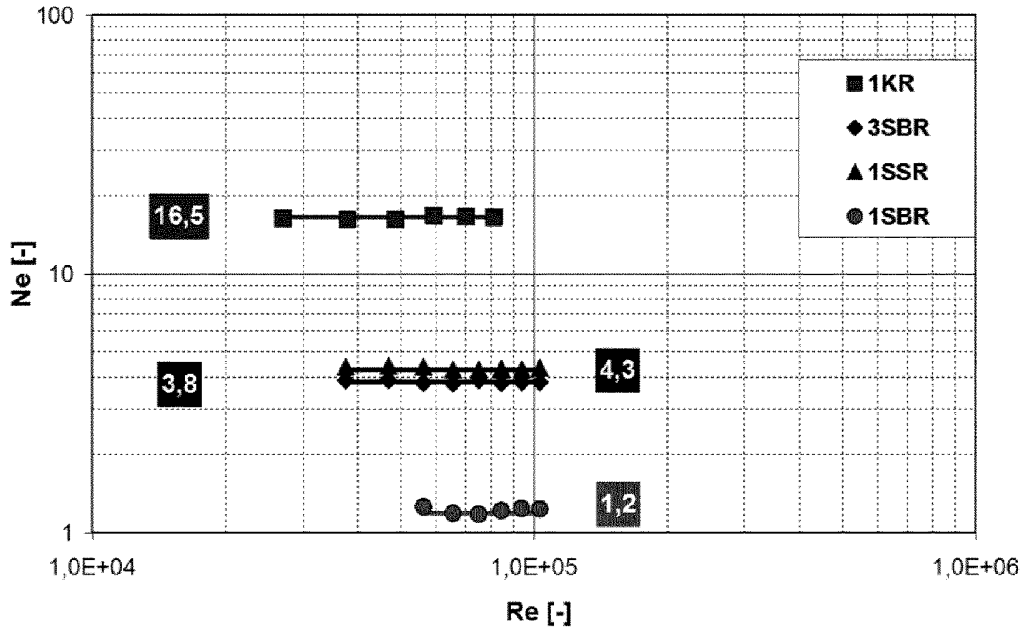
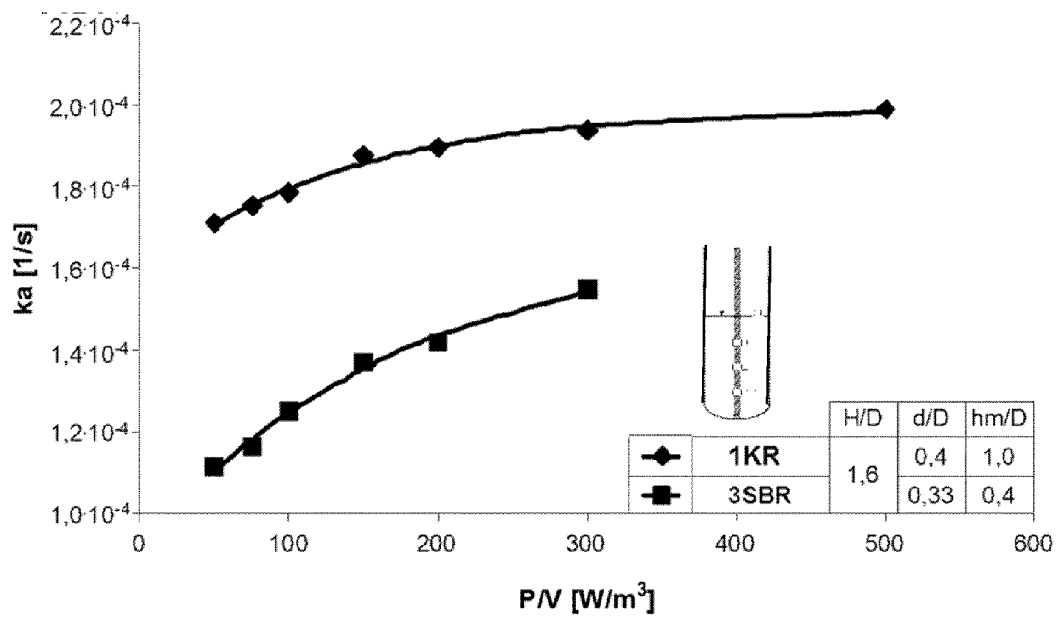


Figure 8



REFERENCES CITED IN THE DESCRIPTION

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**Az EP 2 437 874 lajstromszámú európai szabadalom igénypontjainak magyar fordítása:**

1. Berendezés, amely tenyésztőedényben keverőművet és dialízismodult tartalmaz, **azzal jellemezve, hogy a keverőmű tartalmaz**

- egy radiális szállítóelemet, amely legalább két keverőlapáttal rendelkezik, és

- egy vagy több axiális szállítóelemet, amelyek mindegyike legalább két keverőlapáttal rendelkezik,

ahol a radiális szállítóelem keverőlapátjai egymással párhuzamosak,

ahol az összes axiális szállítóelem külső átmérője egyenlő vagy kisebb, mint a radiális szállítóelem belső átmérője,

ahol az összes axiális szállítóelem egyenként össze van kötve a radiális szállítóelemmel,

ahol az összes axiális szállítóelem a radiális szállítóelemen belül van elhelyezve, és

ahol az összes szállítóelem egymáshoz viszonyítva rögzített térbeli orientációban van.

2. Az 1. igénypont szerinti berendezés, **azzal jellemezve, hogy az axiális szállítóelemek száma 1 vagy 2 vagy 3.**

3. Az előző igénypontok bármelyike szerinti berendezés, **azzal jellemezve, hogy a radiális szállítóelem átellenes keverőlapátjai egy axiális szállítóelem átellenes keverőlapátjain át egymással össze vannak kötve.**

4. Az előző igénypontok bármelyike szerinti berendezés, **azzal jellemezve, hogy a radiális szállítóelem egy horgonykeverő.**

5. Az előző igénypontok bármelyike szerinti berendezés, **azzal jellemezve, hogy az axiális szállítóelem egy ferde lapátú keverőmű.**
6. Az 5. igénypont szerinti berendezés, **azzal jellemezve, hogy a ferde lapátú keverőmű keverőlapátjainak hajlásszöge  $10^\circ$  és  $80^\circ$  között van a keverőmű tengelyvonalára vonatkoztatva.**
7. Az előző igénypontok bármelyike szerinti berendezés, **azzal jellemezve, hogy a radiális szállítóelem magassága legalább 200 mm.**
8. Az 1-7. igénypontok bármelyike szerinti berendezés, **azzal jellemezve, hogy a keverőmű átmérője ( $d$ ) és a tenyésztőedény átmérője ( $D$ ) közötti  $d/D$  arány 0,2-0,8.**
9. Az 1-8. igénypontok bármelyike szerinti berendezés, **azzal jellemezve, hogy a tenyésztőedény töltési magassága ( $H$ ) és a tenyésztőedény szélessége ( $D$ ) közötti  $H/D$  arány 1,0-2,5.**
10. Az 1-9. igénypontok bármelyike szerinti berendezés használata olyan sejt tenyésztésére, amely polipeptidet expresszál.
11. A 10. igénypont szerinti használat, **azzal jellemezve, hogy a sejt emiős-sejt.**
12. A 11. igénypont szerinti használat, **azzal jellemezve, hogy az emiőssejt CHO-sejt, BHK-sejt, NSO-sejt, COS-sejt, PER.C6-sejt, Sp2/O-sejt vagy HEK293-sejt közül van kiválasztva.**

13. A 10-12. igénypontok bármelyike szerinti használat, **azzal jellemezve, hogy a polipeptid egy antitest.**

14. Eljárás polipeptid előállítására, amely eljárás a következőket tartalmazza:

a) egy sejt előkészítése, amely tartalmazza a polipeptidet kódoló nukleinsavat,

b) az 1-9. igénypontok egyike szerinti berendezés előkészítése,

c) a sejt tenyésztése a berendezésben tenyésztőközegben, ahol a keverőmű turbulens áramlást hoz létre a tenyésztőedényben, és

d) a polipeptid kinyerése a sejtekből vagy a tenyésztőközezből, és ezáltal egy polipeptid előállítása.

15. A 14. igénypont szerinti eljárás, **azzal jellemezve, hogy a sejt emlőssejt.**

16. A 15. igénypont szerinti eljárás, **azzal jellemezve, hogy az emlőssejt**

CHO-sejt, BHK-sejt, NSO-sejt, COS-sejt, PER.C6-sejt, Sp2/0-sejt vagy

HEK293-sejt közül van kiválasztva.

17. A 14-16. igénypontok bármelyike szerinti eljárás, **azzal jellemezve, hogy a polipeptid egy antitest.**

A meghatalmazott:



**Dr. Miskolczi Mária**

ügyvéd

1111 Budapest, Egy József u. 40.  
Tel.: 210-4148 Fax: 210-4608