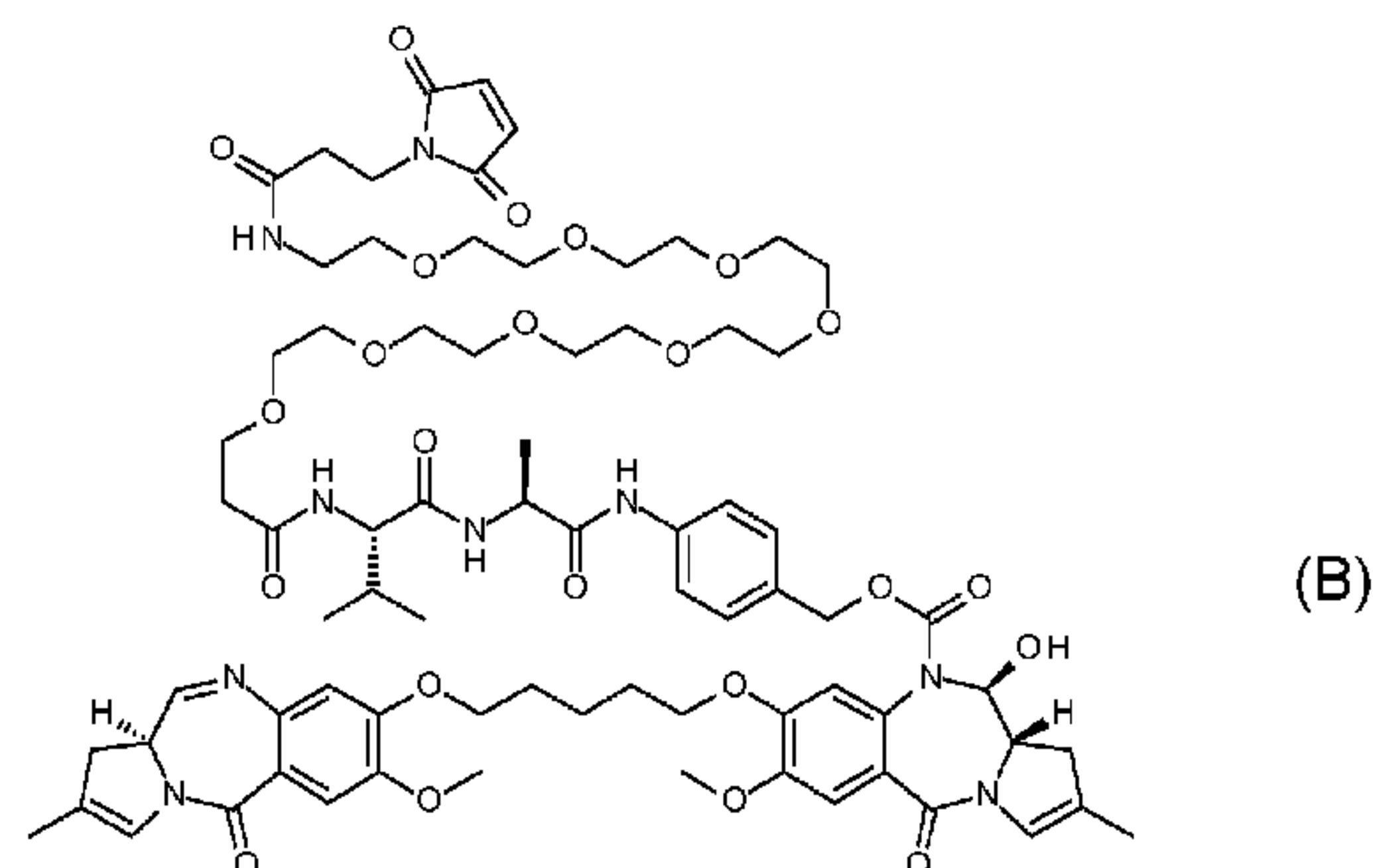
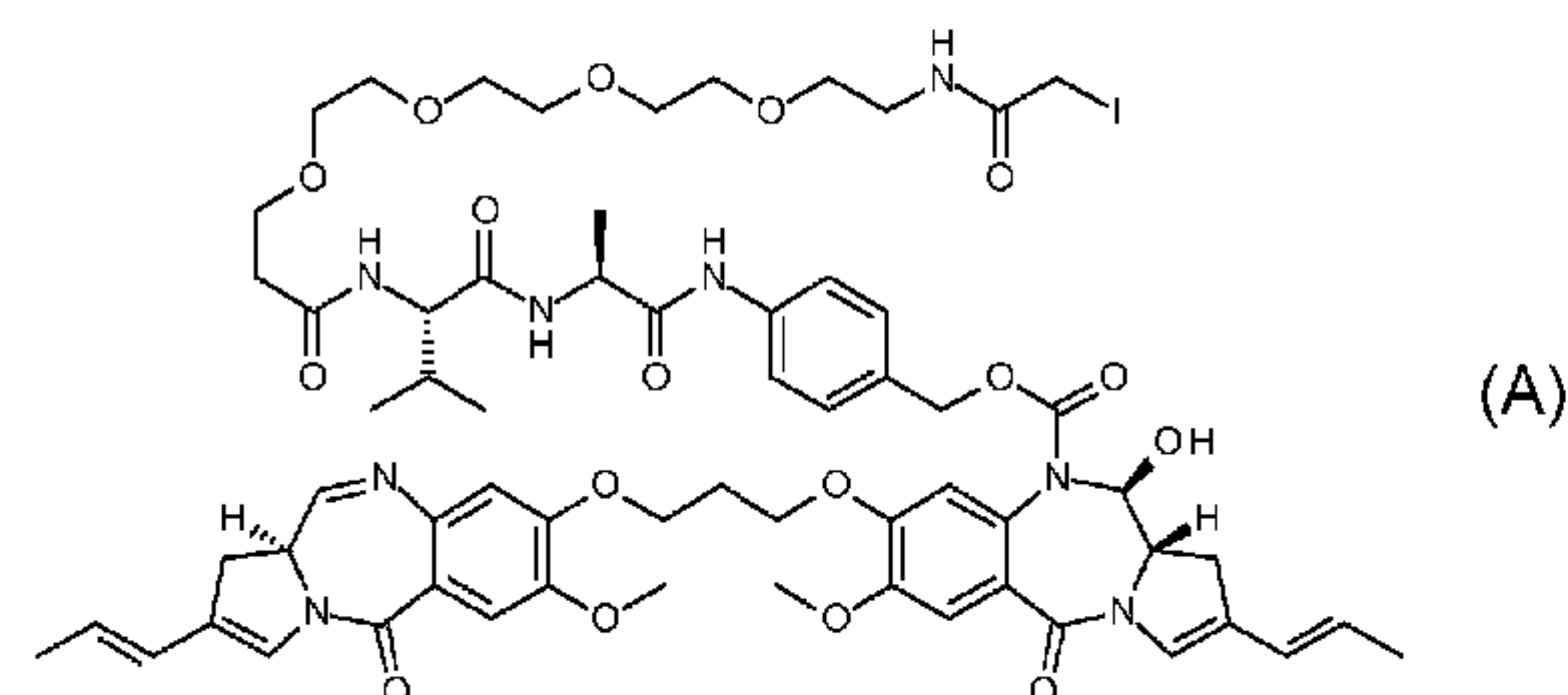
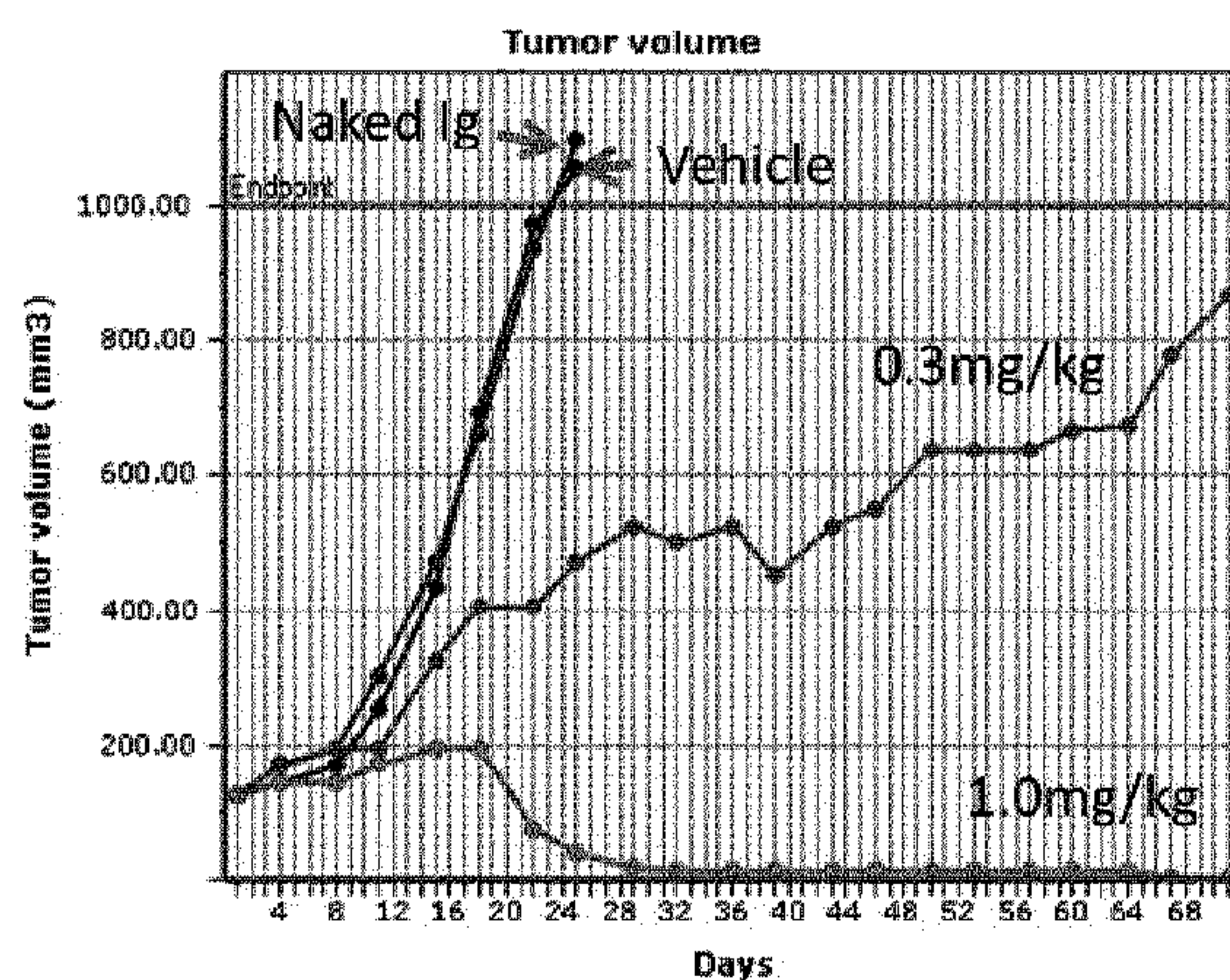




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(54) **Titre : PYRROLOBENZODIAZEPINES ET LEURS CONJUGUES**
(54) **Title: PYRROLOBENZODIAZEPINES AND CONJUGATES THEREOF**



(57) **Abstrégé/Abstract:**

A compound which is either A or B and salts and solvates thereof, as well as conjugates thereof with cell binding agent.



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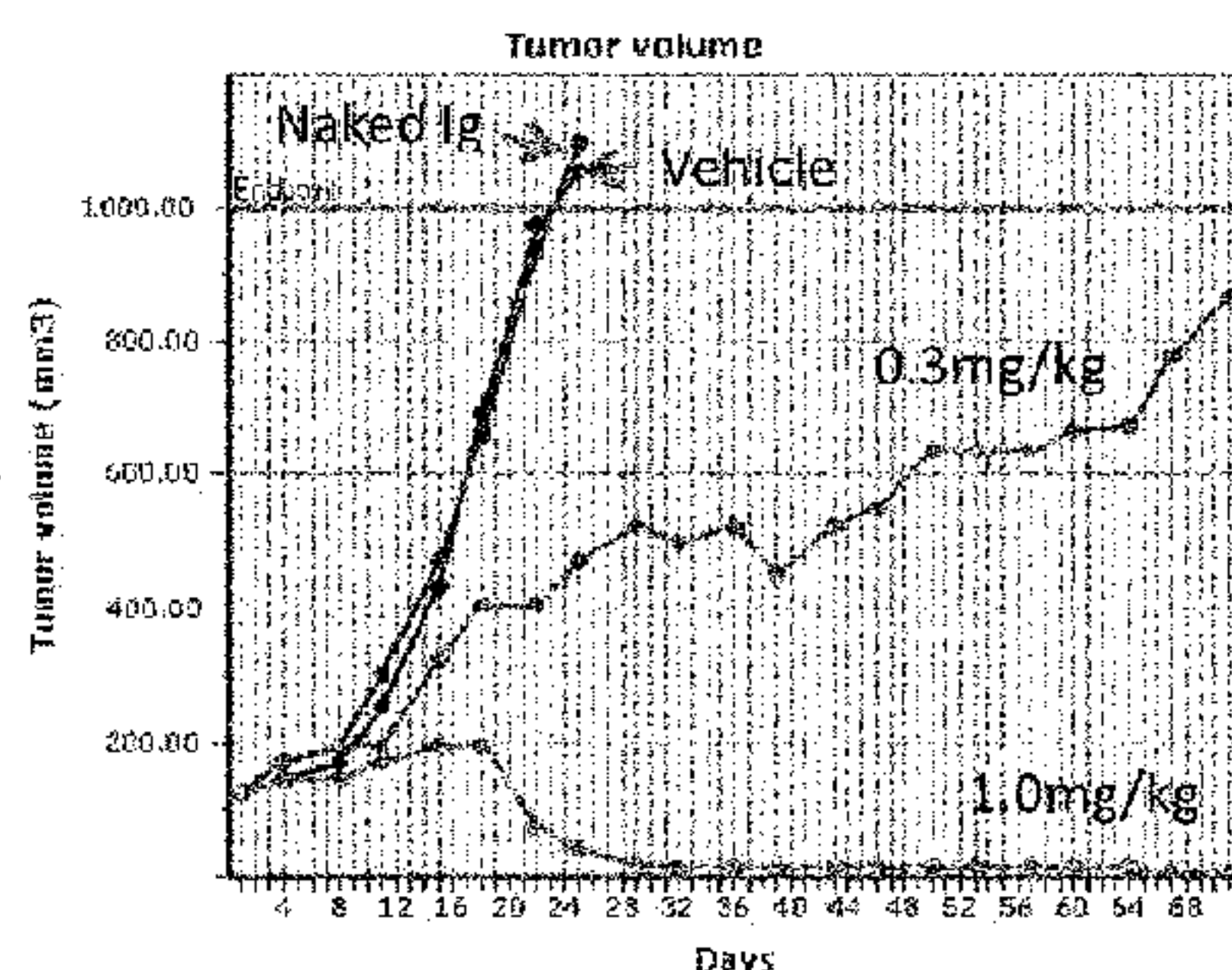
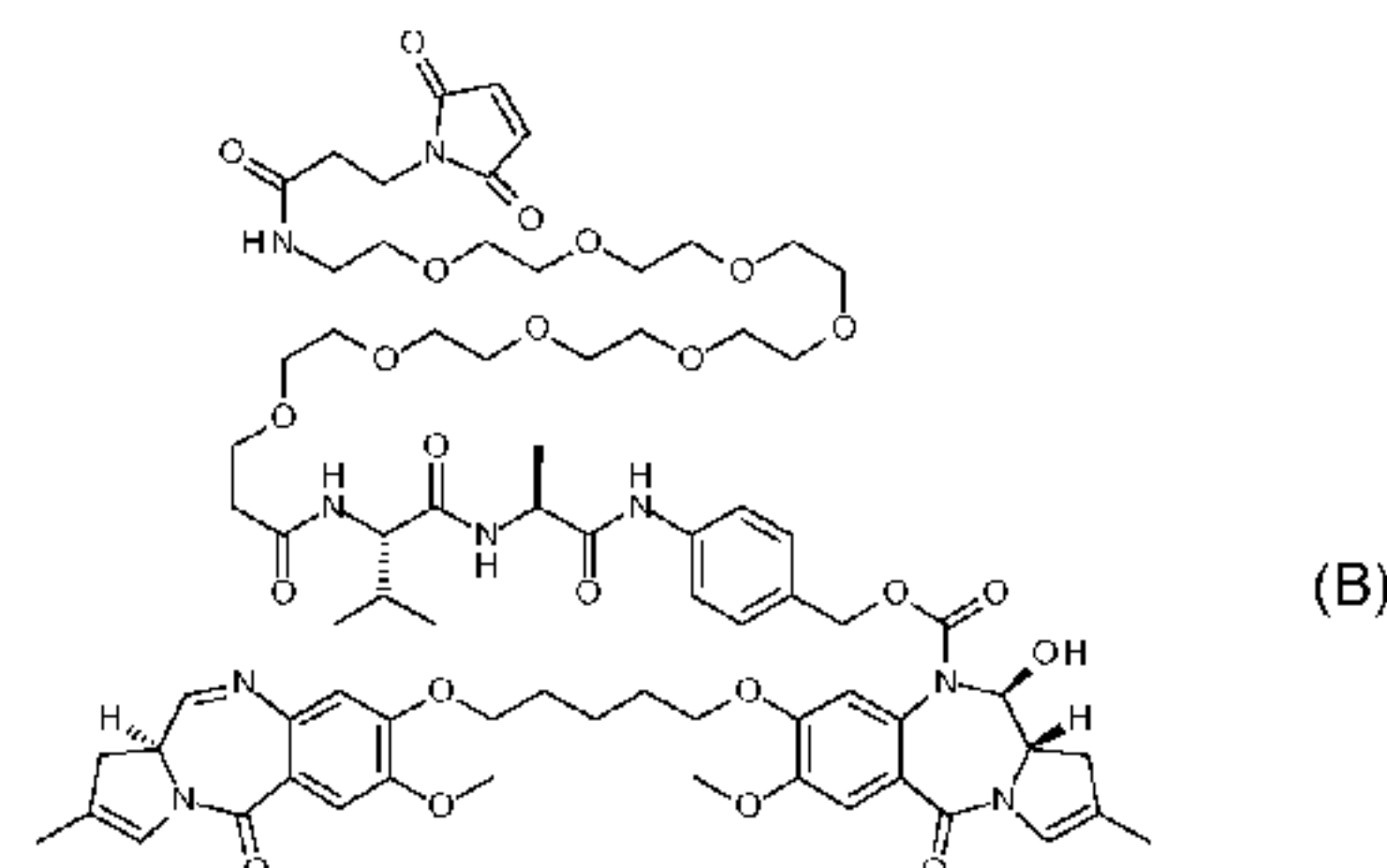
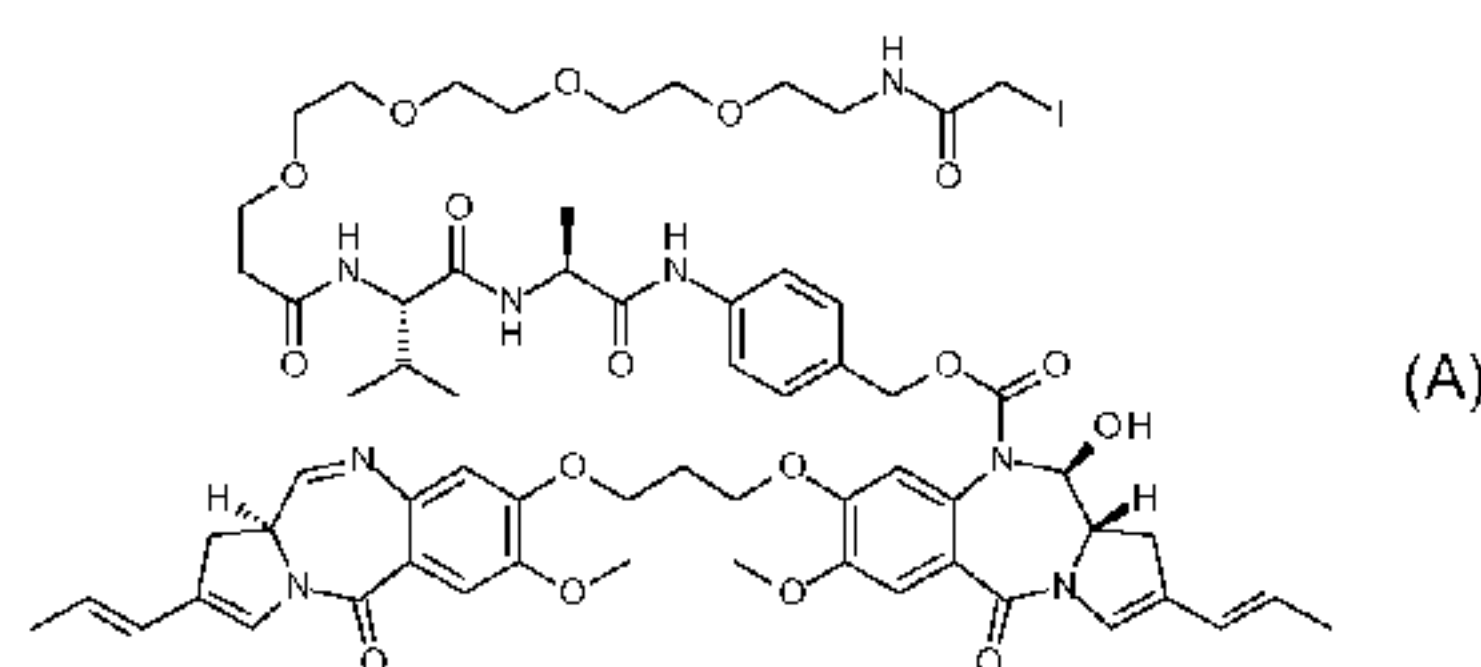


Figure 1



(57) Abstract: A compound which is either A or B and salts and solvates thereof, as well as conjugates thereof with cell binding agent.

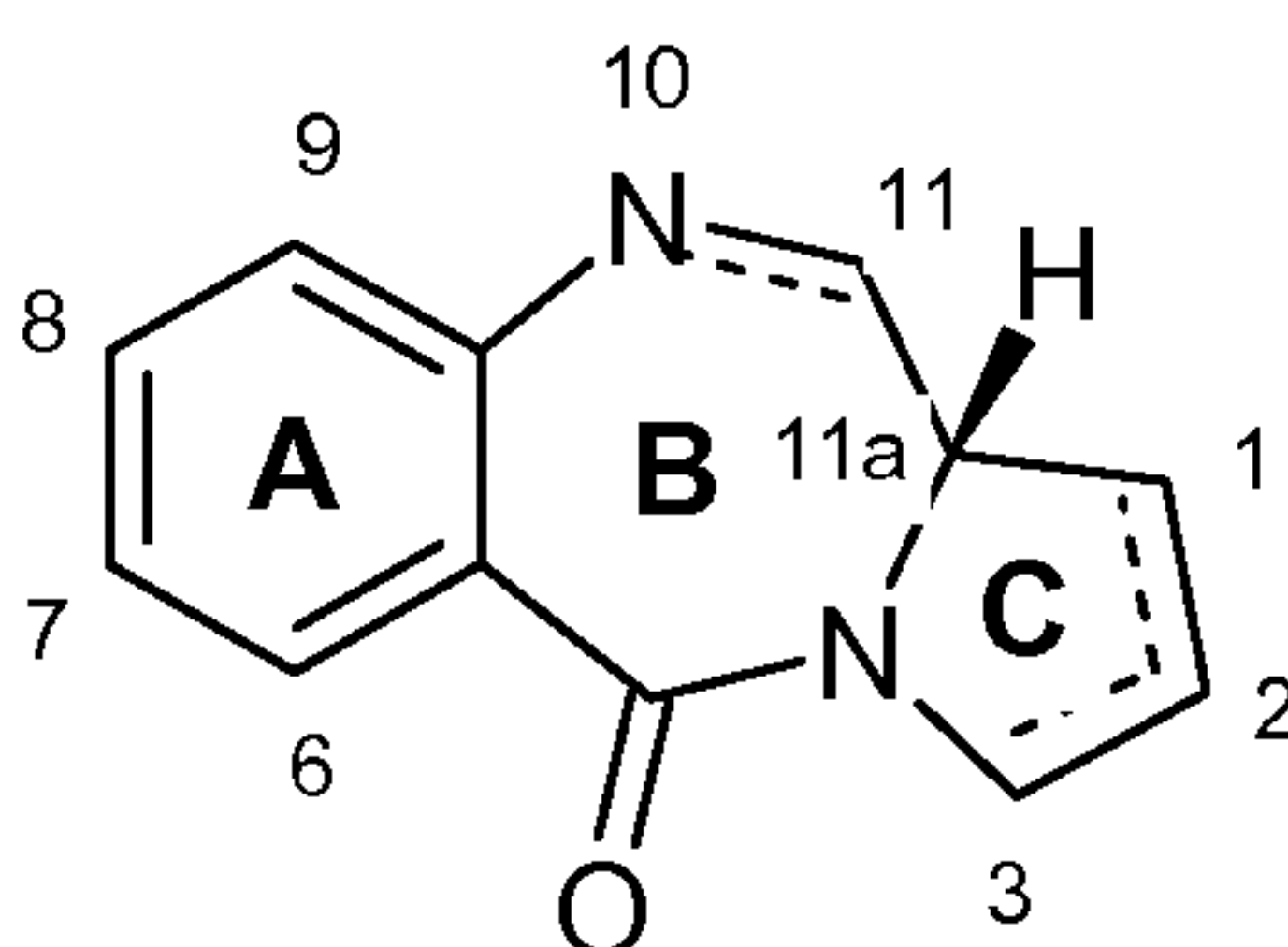
PYRROLOBENZODIAZEPINES AND CONJUGATES THEREOF

The present invention relates to pyrrolobenzodiazepines (PBDs), in particular pyrrolobenzodiazepines having a labile N10 protecting group, in the form of a linker to a cell
5 binding agent.

Background to the invention

Pyrrolobenzodiazepines

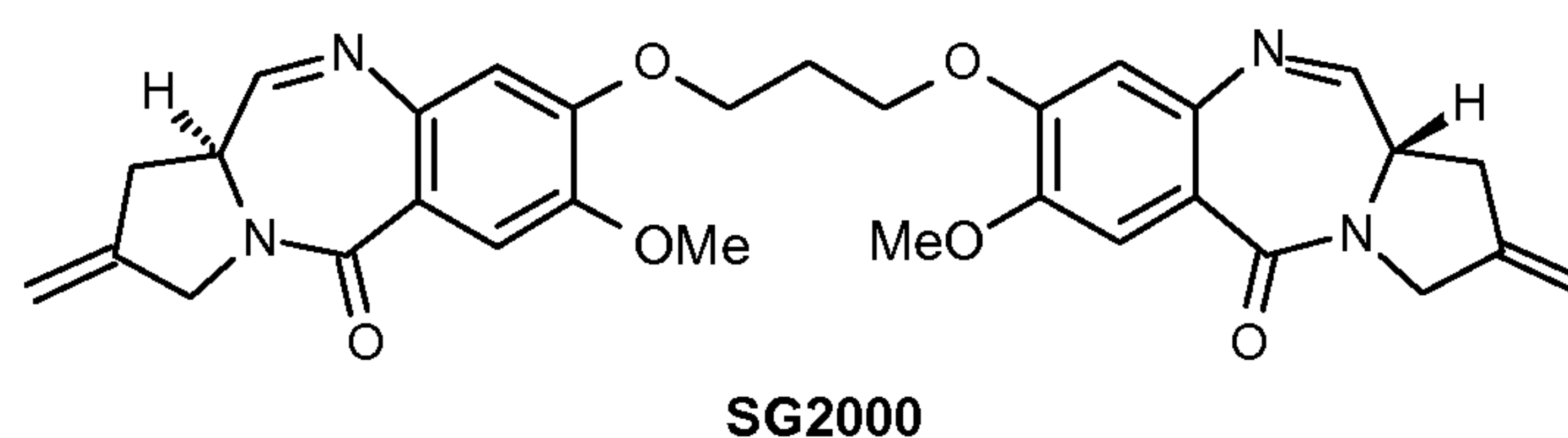
Some pyrrolobenzodiazepines (PBDs) have the ability to recognise and bond to specific
10 sequences of DNA; the preferred sequence is PuGPu. The first PBD antitumour antibiotic, anthramycin, was discovered in 1965 (Leimgruber, *et al.*, *J. Am. Chem. Soc.*, **87**, 5793-5795 (1965); Leimgruber, *et al.*, *J. Am. Chem. Soc.*, **87**, 5791-5793 (1965)). Since then, a number of naturally occurring PBDs have been reported, and over 10 synthetic routes have been developed to a variety of analogues (Thurston, *et al.*, *Chem. Rev.* **1994**, 433-465 (1994);
15 Antonow, D. and Thurston, D.E., *Chem. Rev.* **2011** 111 (4), 2815-2864). Family members include abbeymycin (Hochlowski, *et al.*, *J. Antibiotics*, **40**, 145-148 (1987)), chicamycin (Konishi, *et al.*, *J. Antibiotics*, **37**, 200-206 (1984)), DC-81 (Japanese Patent 58-180 487; Thurston, *et al.*, *Chem. Brit.*, **26**, 767-772 (1990); Bose, *et al.*, *Tetrahedron*, **48**, 751-758 (1992)), mazethramycin (Kuminoto, *et al.*, *J. Antibiotics*, **33**, 665-667 (1980)), neothramycins
20 A and B (Takeuchi, *et al.*, *J. Antibiotics*, **29**, 93-96 (1976)), porothramycin (Tsunakawa, *et al.*, *J. Antibiotics*, **41**, 1366-1373 (1988)), prothracarcin (Shimizu, *et al.*, *J. Antibiotics*, **29**, 2492-2503 (1982); Langley and Thurston, *J. Org. Chem.*, **52**, 91-97 (1987)), sibanomycin (DC-102)(Hara, *et al.*, *J. Antibiotics*, **41**, 702-704 (1988); Itoh, *et al.*, *J. Antibiotics*, **41**, 1281-1284 (1988)), sibiromycin (Leber, *et al.*, *J. Am. Chem. Soc.*, **110**, 2992-2993 (1988)) and
25 tomamycin (Arima, *et al.*, *J. Antibiotics*, **25**, 437-444 (1972)). PBDs are of the general structure:



They differ in the number, type and position of substituents, in both their aromatic A rings
30 and pyrrolo C rings, and in the degree of saturation of the C ring. In the B-ring there is either an imine (N=C), a carbinolamine(NH-CH(OH)), or a carbinolamine methyl ether (NH-

CH(OMe)) at the N10-C11 position which is the electrophilic centre responsible for alkylating DNA. All of the known natural products have an (S)-configuration at the chiral C11a position which provides them with a right-handed twist when viewed from the C ring towards the A ring. This gives them the appropriate three-dimensional shape for isohelicity with the minor groove of B-form DNA, leading to a snug fit at the binding site (Kohn, In *Antibiotics III*. Springer-Verlag, New York, pp. 3-11 (1975); Hurley and Needham-VanDevanter, *Acc. Chem. Res.*, **19**, 230-237 (1986)). Their ability to form an adduct in the minor groove, enables them to interfere with DNA processing, hence their use as antitumour agents.

A particularly advantageous pyrrolobenzodiazepine compound is described by Gregson *et al.* (*Chem. Commun.* **1999**, 797-798) as compound **1**, and by Gregson *et al.* (*J. Med. Chem.* **2001**, *44*, 1161-1174) as compound **4a**. This compound, also known as SG2000, is shown below:



WO 2007/085930 describes the preparation of dimer PBD compounds having linker groups for connection to a cell binding agent, such as an antibody. The linker is present in the bridge linking the monomer PBD units of the dimer.

The present inventors have described dimer PBD compounds having linker groups for connection to a cell binding agent, such as an antibody, in WO 2011/130598. The linker in these compounds is attached to one of the available N10 positions, and are generally cleaved by action of an enzyme on the linker group.

Antibody-drug conjugates

Antibody therapy has been established for the targeted treatment of patients with cancer, immunological and angiogenic disorders (Carter, P. (2006) *Nature Reviews Immunology* 6:343-357). The use of antibody-drug conjugates (ADC), i.e. immunoconjugates, for the local delivery of cytotoxic or cytostatic agents, i.e. drugs to kill or inhibit tumor cells in the treatment of cancer, targets delivery of the drug moiety to tumors, and intracellular accumulation therein, whereas systemic administration of these unconjugated drug agents may result in unacceptable levels of toxicity to normal cells (Xie *et al* (2006) *Expert. Opin. Biol. Ther.* 6(3):281-291; Kovtun *et al* (2006) *Cancer Res.* 66(6):3214-3121; Law *et al* (2006)

Cancer Res. 66(4):2328-2337; Wu *et al* (2005) *Nature Biotech.* 23(9):1137-1145; Lambert J.
 (2005) *Current Opin. in Pharmacol.* 5:543-549; Hamann P. (2005) *Expert Opin. Ther.*
Patents 15(9):1087-1103; Payne, G. (2003) *Cancer Cell* 3:207-212; Trail *et al* (2003) *Cancer*
Immunol. Immunother. 52:328-337; Syrigos and Epenetos (1999) *Anticancer Research*
 5 19:605-614).

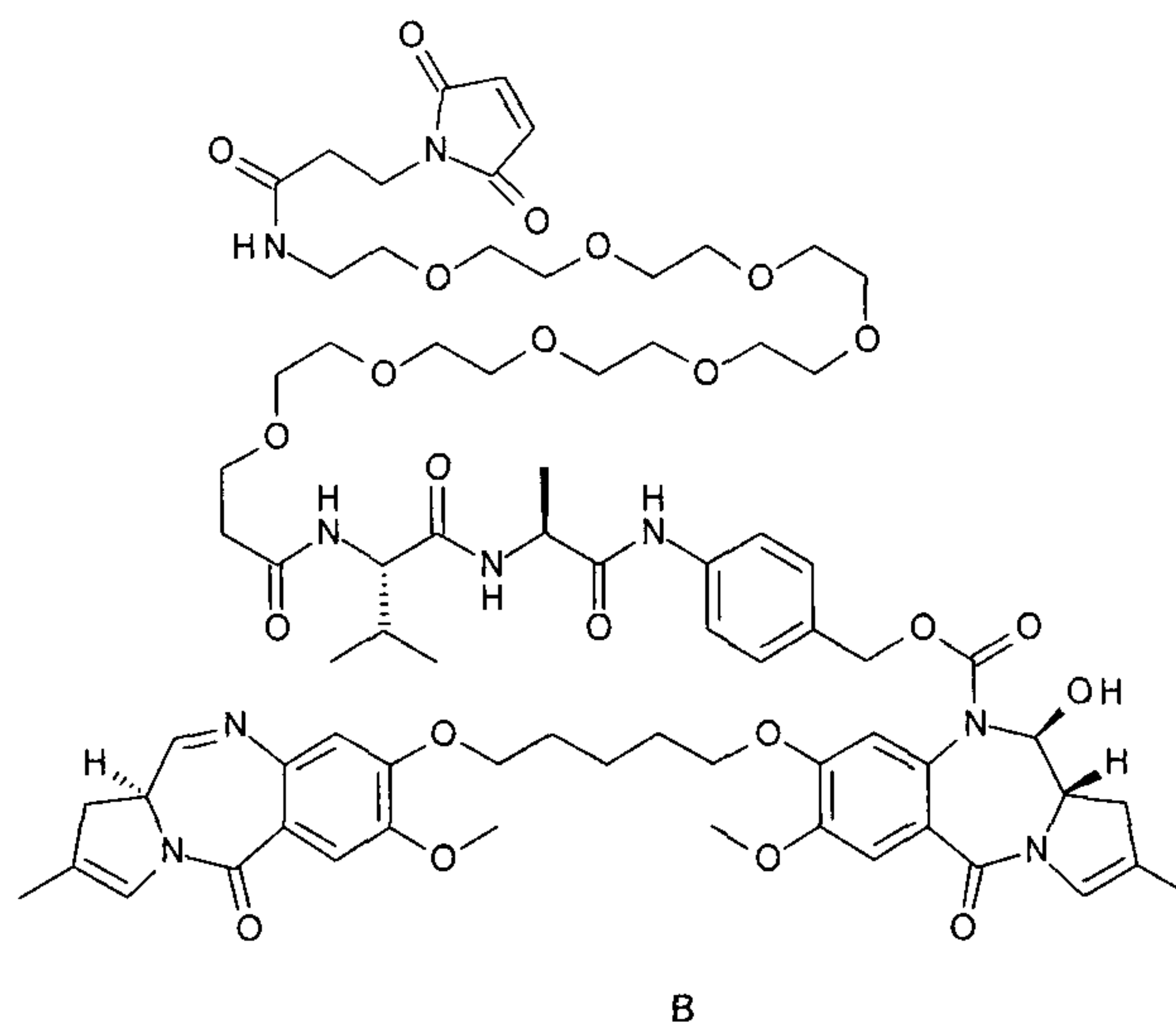
Maximal efficacy with minimal toxicity is sought thereby. Efforts to design and refine ADC
 have focused on the selectivity of monoclonal antibodies (mAbs) as well as drug mechanism
 of action, drug-linking, drug/antibody ratio (loading), and drug-releasing properties (Junutula,
 10 *et al.*, 2008b *Nature Biotech.*, 26(8):925-932; Dornan *et al* (2009) *Blood* 114(13):2721-2729;
 US 7521541; US 7723485; WO2009/052249; McDonagh (2006) *Protein Eng. Design & Sel.*
 19(7): 299-307; Doronina *et al* (2006) *Bioconj. Chem.* 17:114-124; Erickson *et al* (2006)
Cancer Res. 66(8):1-8; Sanderson *et al* (2005) *Clin. Cancer Res.* 11:843-852; Jeffrey *et al*
 (2005) *J. Med. Chem.* 48:1344-1358; Hamblett *et al* (2004) *Clin. Cancer Res.* 10:7063-
 15 7070). Drug moieties may impart their cytotoxic and cytostatic effects by mechanisms
 including tubulin binding, DNA binding, proteasome and/or topoisomerase inhibition. Some
 cytotoxic drugs tend to be inactive or less active when conjugated to large antibodies or
 protein receptor ligands.

20 The present inventors have developed particular PBD dimers with linking groups for the
 formation of PBD conjugates with cell binding agents, and in particular PBD antibody
 conjugates.

3a

Summary of the Invention

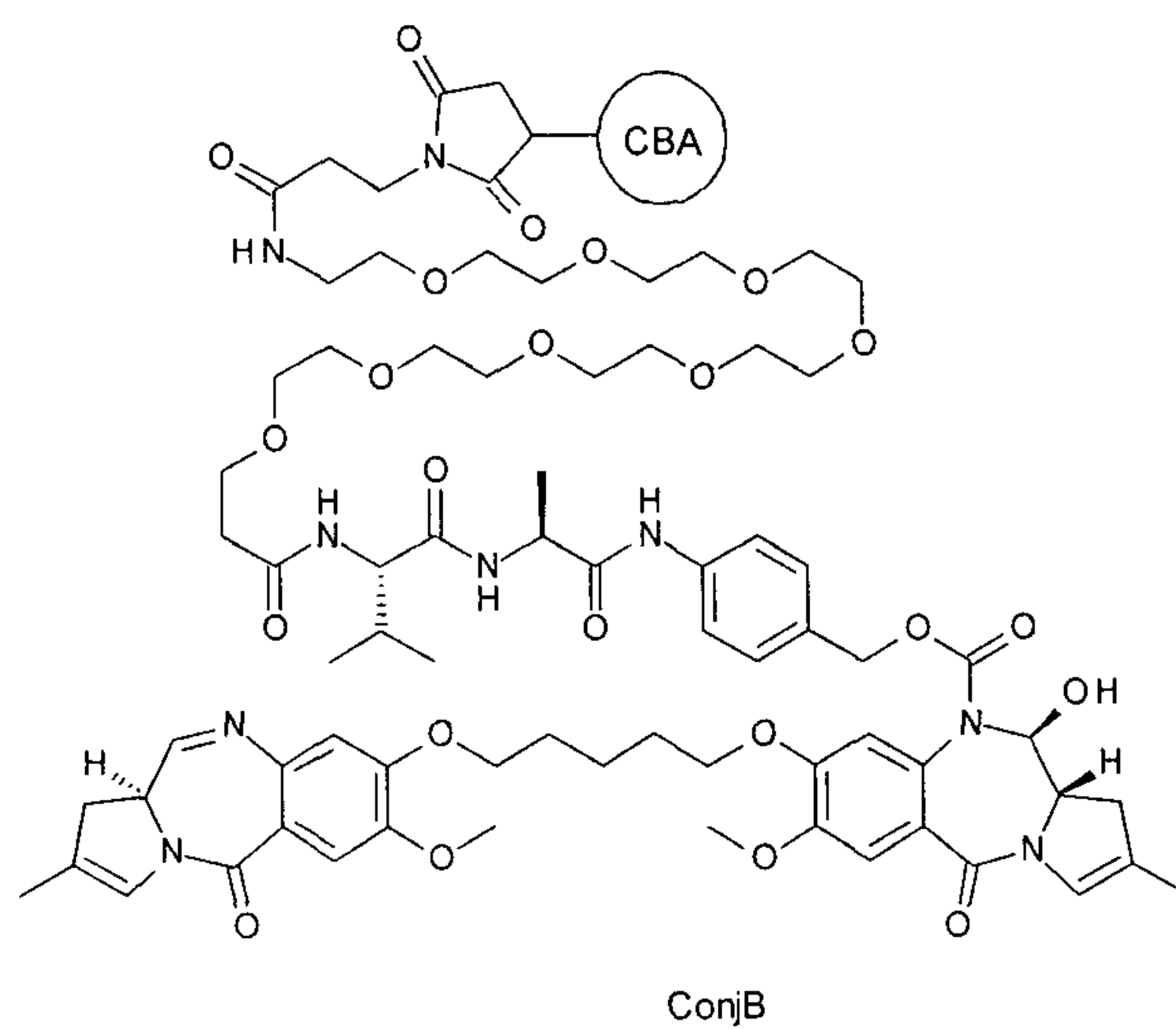
Certain exemplary embodiments provide a compound B:



and salts and solvates thereof.

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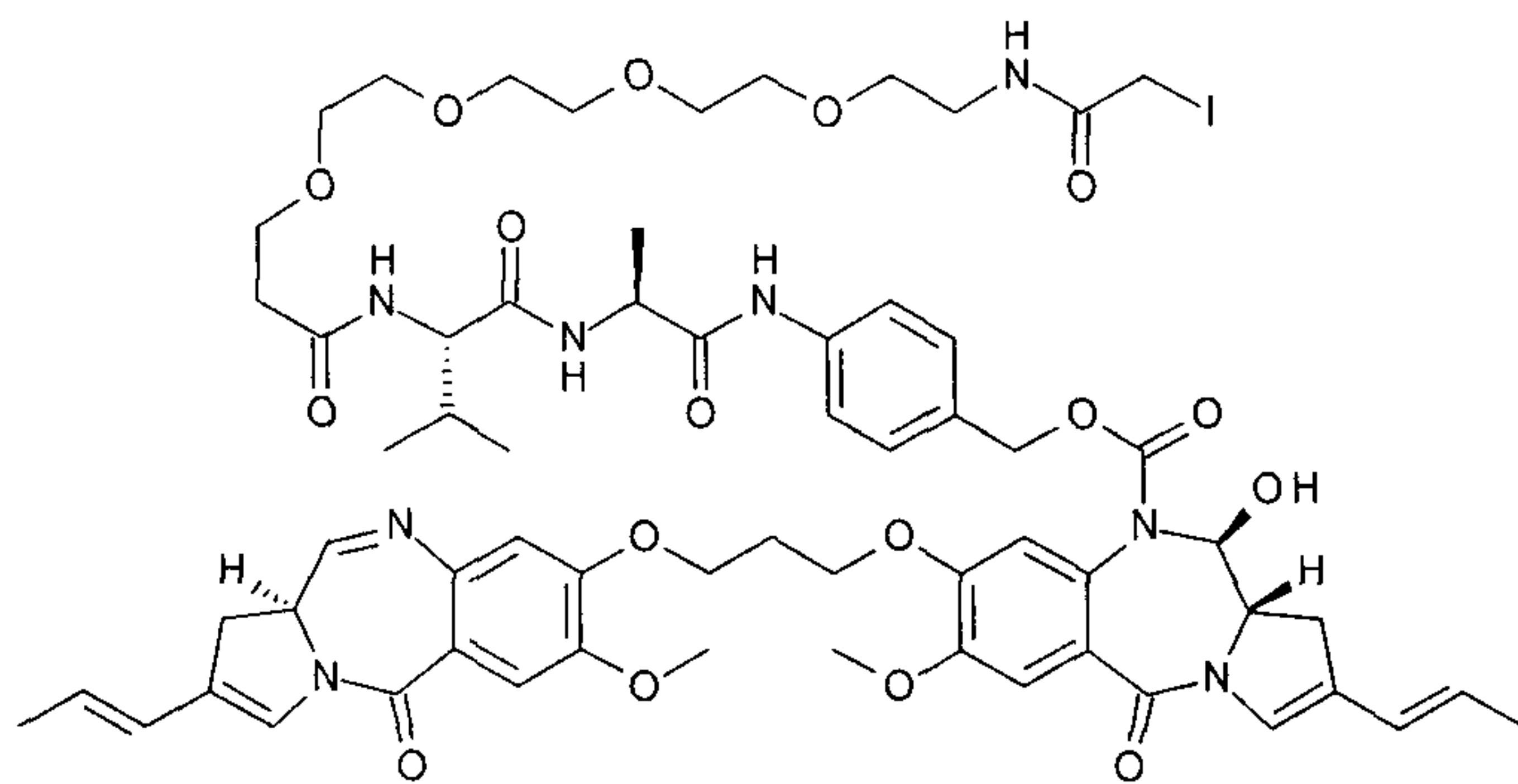
Other exemplary embodiments provide a conjugate of formula ConjB:



where CBA represents a cell binding agent.

3b

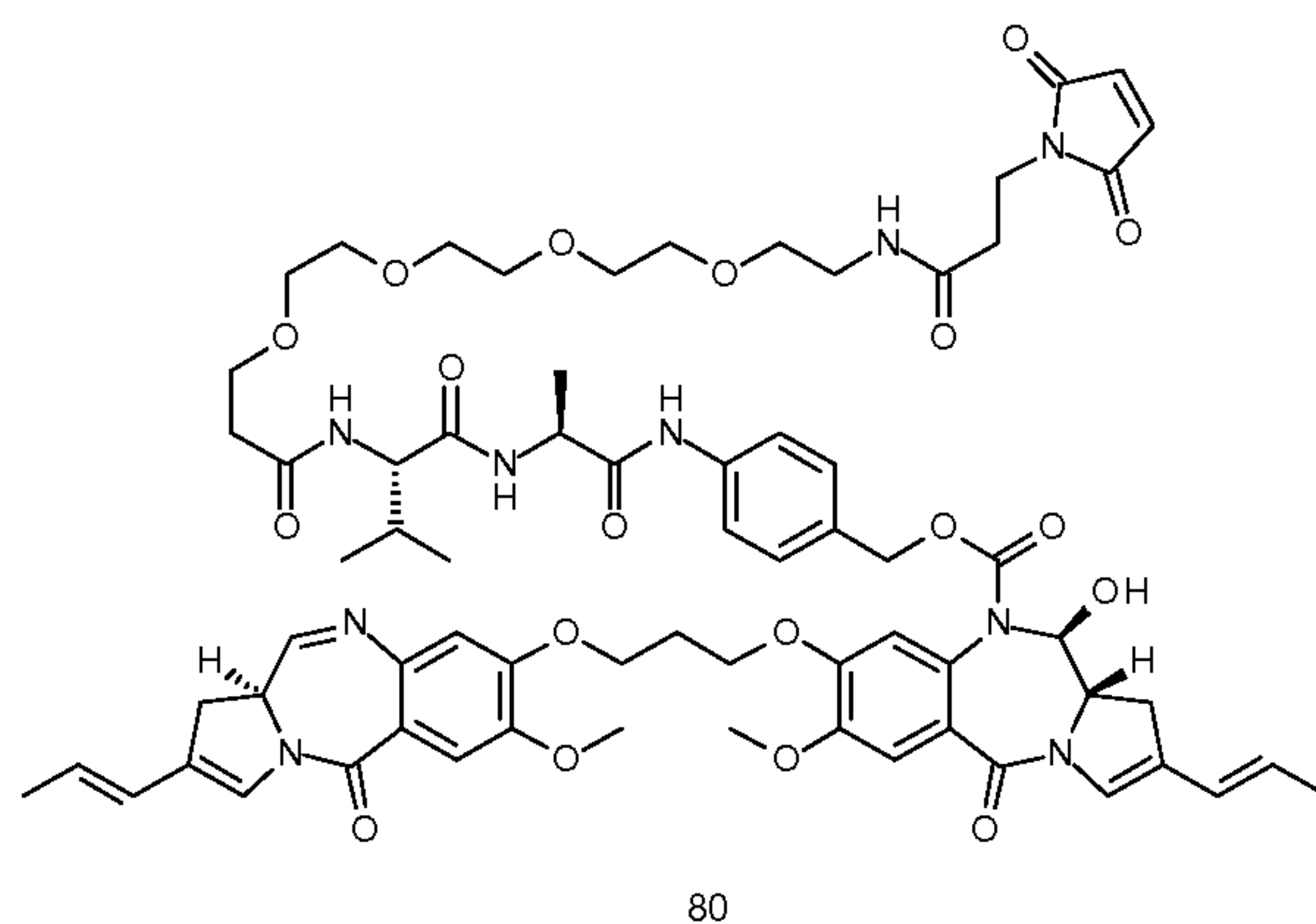
In a first aspect, the present invention provides compound A:



A

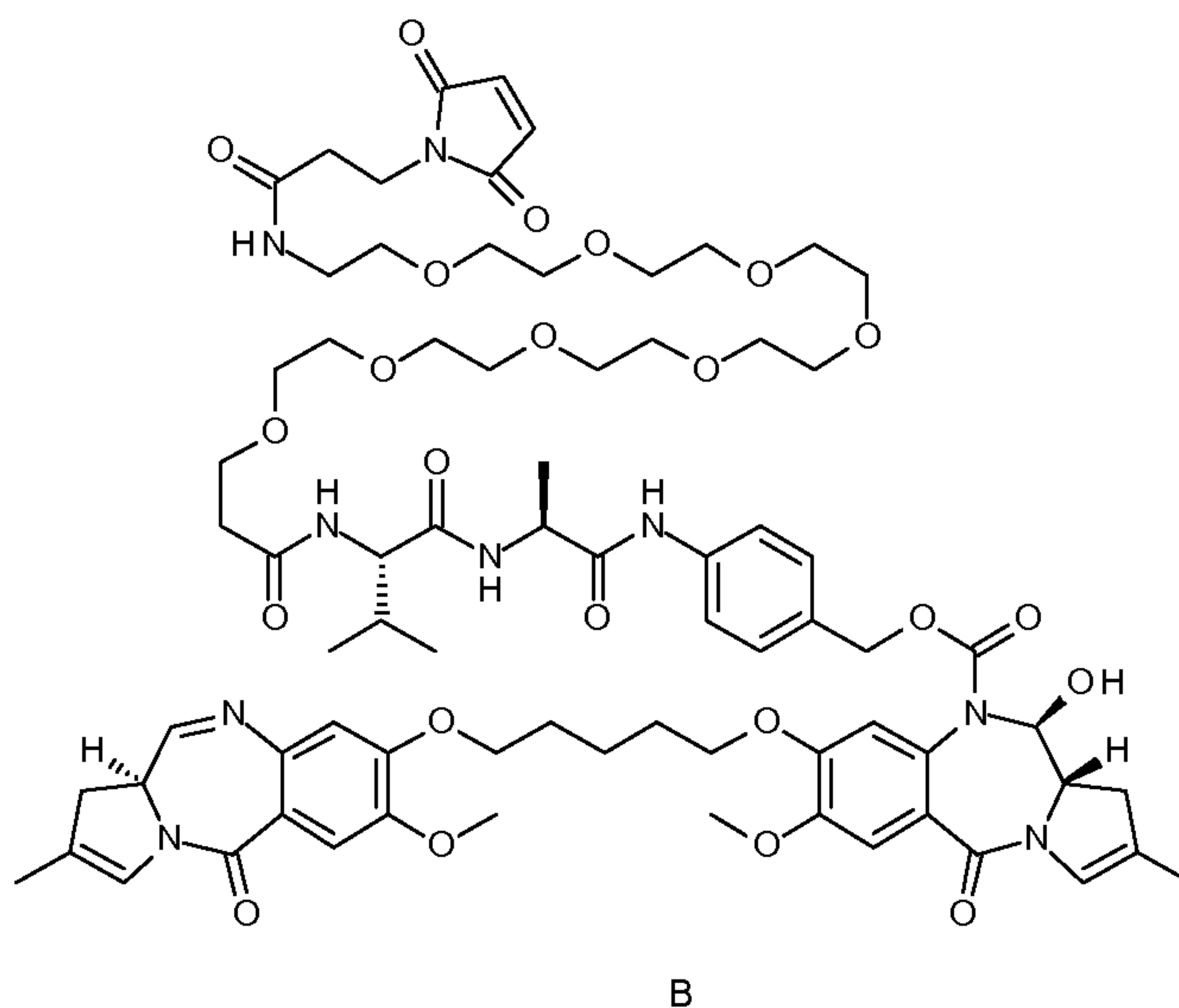
and salts of solvates thereof.

WO 2011/130598 discloses compound 80:



Compound A differs from this by comprising an iodoacetamide group for linking to the cell binding agent. This group may offer advantages over compound 80 with regards to its stability when bound to a cell binding agent (see below). The maleimide group in compound 80 can undergo a retro-Michael reaction, becoming unconjugated from the cell binding agent, and thus vulnerable to scavenging by other thiol containing biological molecules, such as albumin and glutathione. Such unconjugation cannot occur with compound A. Also, the iodoacetamide group may avoid other unwanted side reactions.

In a second aspect, the present invention provides compound B:



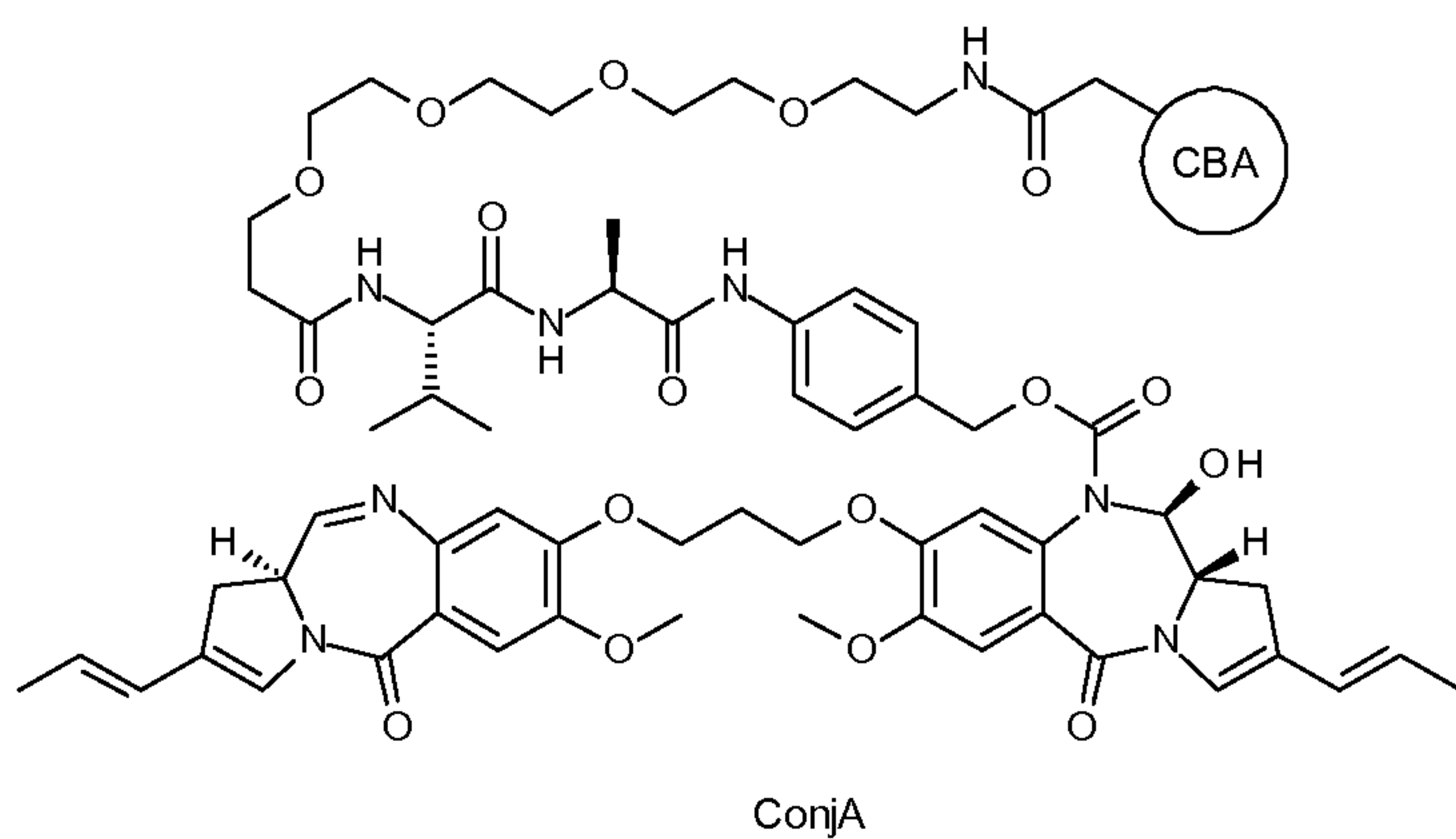
and salts and solvates thereof.

Compound B differs from previously disclosed PBD dimers with a drug linker having a C2-3 endo-double bond, by having a smaller, less lipophilic C2 substituent, e.g. 4F-phenyl, propylene. As such, the conjugates of compound B (see below) are less likely to aggregate

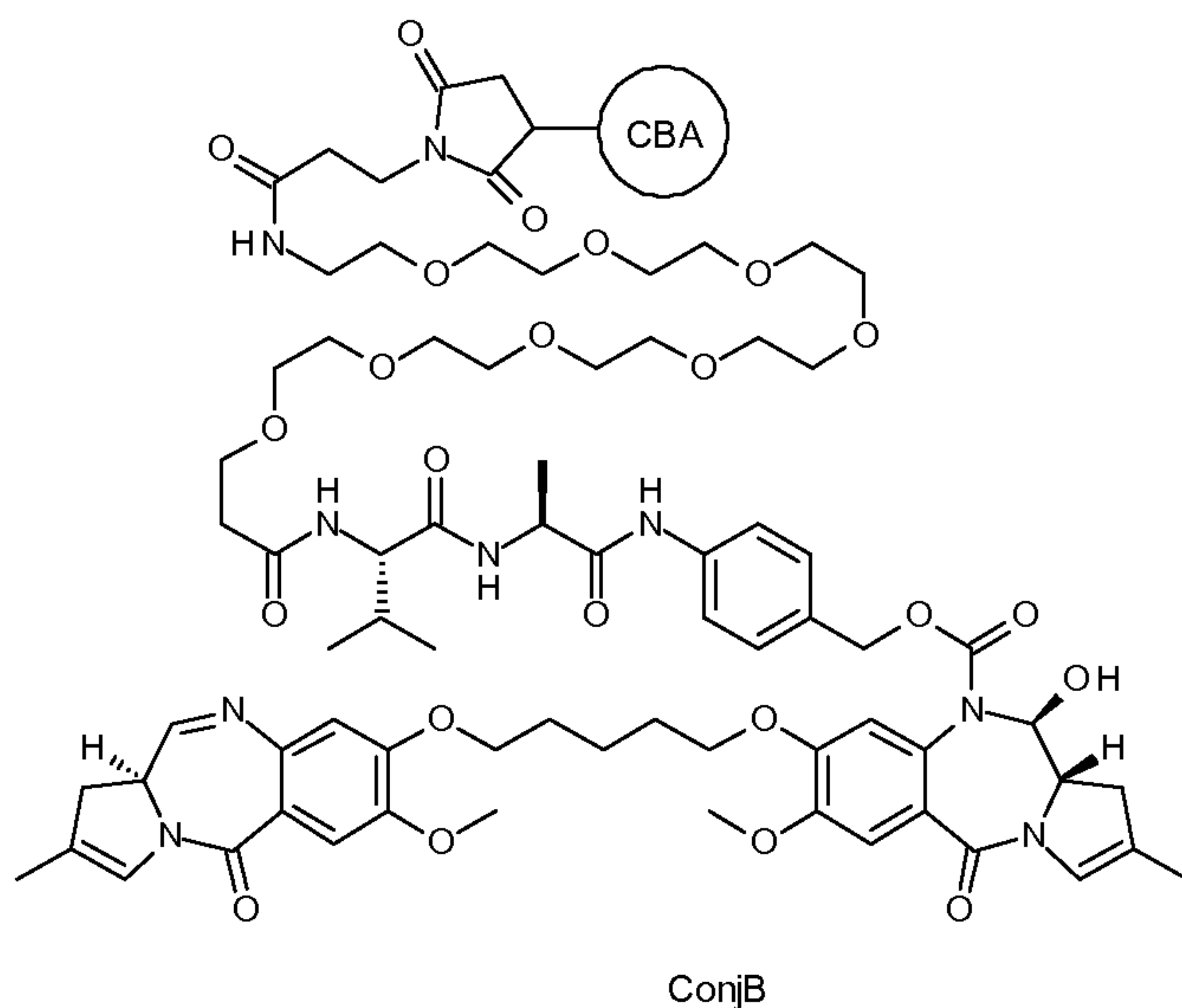
once synthesised. Such aggregation of conjugates can be measured by Size exclusion chromatography (SEC).

Both compound A and B have two sp^2 centres in each C-ring, which may allow for stronger binding in the minor groove of DNA, than for compounds with only one sp^2 centre in each C-ring.

A third aspect of the present invention provides a conjugate of formula ConjA:



10 or ConjB:



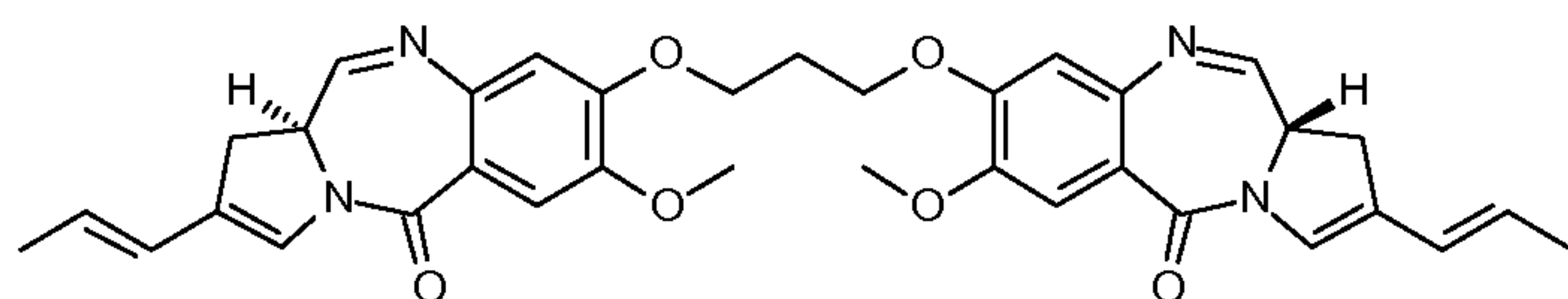
where CBA represents a cell binding agent. The link to the moiety shown is via a free S (active thiol) on the cell binding agent.

Detailed Description of the Invention

The present invention provides a PBD dimer with a linker connected through the N10 position on one of the PBD moieties suitable for forming a PBD dimer conjugated via the linker to a cell binding agent.

5

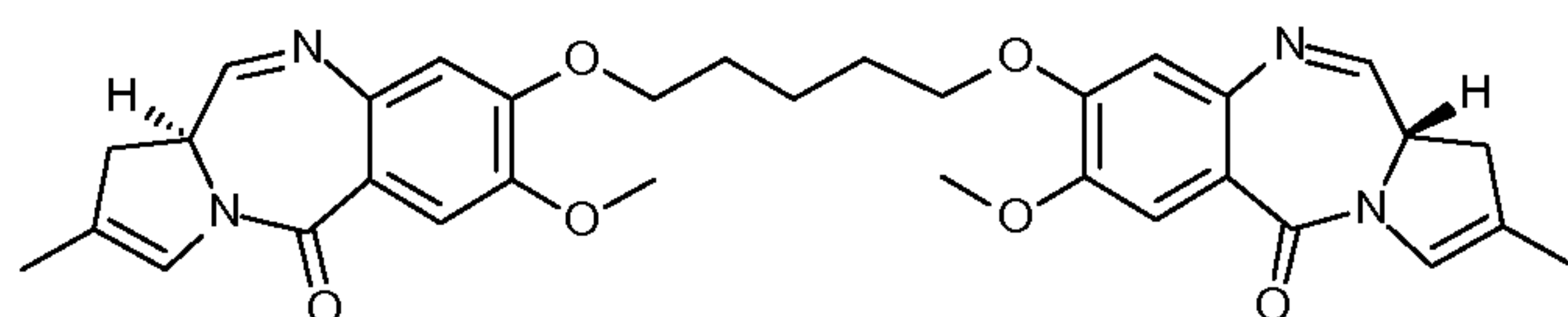
The present invention is suitable for use in providing a PBD compound to a preferred site in a subject. The conjugate allows the release of an active PBD compound that does not retain any part of the linker. There is no stub present that could affect the reactivity of the PBD compound. Thus ConjA would release the compound RelA:



RelA

10

and ConjB would release the compound RelB:



RelB

A further aspect of the present invention is the compound RelB, and salts and solvates thereof.

15

The specified link between the PBD dimer and the cell binding agent, e.g. antibody, in the present invention is preferably stable extracellularly. Before transport or delivery into a cell, the antibody-drug conjugate (ADC) is preferably stable and remains intact, i.e. the antibody remains linked to the drug moiety. The linkers are stable outside the target cell and may be cleaved at some efficacious rate inside the cell. An effective linker will: (i) maintain the specific binding properties of the antibody; (ii) allow intracellular delivery of the conjugate or drug moiety; (iii) remain stable and intact, i.e. not cleaved, until the conjugate has been delivered or transported to its targetted site; and (iv) maintain a cytotoxic, cell-killing effect or a cytostatic effect of the PBD drug moiety. Stability of the ADC may be measured by standard analytical techniques such as mass spectroscopy, HPLC, and the separation/analysis technique LC/MS.

20

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Delivery of the compounds of formulae RelA or RelB is achieved at the desired activation site of the conjugates of formulae ConjA or ConjB by the action of an enzyme, such as cathepsin, on the linking group, and in particular on the valine-alanine dipeptide moiety.

Cell Binding Agent

A cell binding agent may be of any kind, and include peptides and non-peptides. These can include antibodies or a fragment of an antibody that contains at least one binding site, lymphokines, hormones, hormone mimetics, vitamins, growth factors, nutrient-transport
5 molecules, or any other cell binding molecule or substance.

Peptides

In one embodiment, the cell binding agent is a linear or cyclic peptide comprising 4-30, preferably 6-20, contiguous amino acid residues. In this embodiment, it is preferred that one
10 cell binding agent is linked to one monomer or dimer pyrrolobenzodiazepine compound.

In one embodiment the cell binding agent comprises a peptide that binds integrin $\alpha_v\beta_6$. The peptide may be selective for $\alpha_v\beta_6$ over $\alpha_5\beta_1$.

15 In one embodiment the cell binding agent comprises the A20FMDV-Cys polypeptide. The A20FMDV-Cys has the sequence: NAVPNLRGDLQVLAQKVARTC. Alternatively, a variant of the A20FMDV-Cys sequence may be used wherein one, two, three, four, five, six, seven, eight, nine or ten amino acid residues are substituted with another amino acid residue. Furthermore, the polypeptide may have the sequence NAVXXXXXXXXXXXXXXXXXRTC.

Antibodies

The term "antibody" herein is used in the broadest sense and specifically covers monoclonal antibodies, polyclonal antibodies, dimers, multimers, multispecific antibodies (e.g., bispecific antibodies), and antibody fragments, so long as they exhibit the desired biological activity
25 (Miller *et al* (2003) *Jour. of Immunology* 170:4854-4861). Antibodies may be murine, human, humanized, chimeric, or derived from other species. An antibody is a protein generated by the immune system that is capable of recognizing and binding to a specific antigen. (Janeway, C., Travers, P., Walport, M., Shlomchik (2001) *Immuno Biology*, 5th Ed., Garland Publishing, New York). A target antigen generally has numerous binding sites, also called
30 epitopes, recognized by CDRs on multiple antibodies. Each antibody that specifically binds to a different epitope has a different structure. Thus, one antigen may have more than one corresponding antibody. An antibody includes a full-length immunoglobulin molecule or an immunologically active portion of a full-length immunoglobulin molecule, *i.e.*, a molecule that contains an antigen binding site that immunospecifically binds an antigen of a target of
35 interest or part thereof, such targets including but not limited to, cancer cell or cells that produce autoimmune antibodies associated with an autoimmune disease. The

immunoglobulin can be of any type (e.g. IgG, IgE, IgM, IgD, and IgA), class (e.g. IgG1, IgG2, IgG3, IgG4, IgA1 and IgA2) or subclass of immunoglobulin molecule. The immunoglobulins can be derived from any species, including human, murine, or rabbit origin.

- 5 "Antibody fragments" comprise a portion of a full length antibody, generally the antigen binding or variable region thereof. Examples of antibody fragments include Fab, Fab', F(ab')₂, and scFv fragments; diabodies; linear antibodies; fragments produced by a Fab expression library, anti-idiotypic (anti-Id) antibodies, CDR (complementary determining region), and epitope-binding fragments of any of the above which immunospecifically bind to
10 cancer cell antigens, viral antigens or microbial antigens, single-chain antibody molecules; and multispecific antibodies formed from antibody fragments.

The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, i.e. the individual antibodies comprising
15 the population are identical except for possible naturally occurring mutations that may be present in minor amounts. Monoclonal antibodies are highly specific, being directed against a single antigenic site. Furthermore, in contrast to polyclonal antibody preparations which include different antibodies directed against different determinants (epitopes), each monoclonal antibody is directed against a single determinant on the antigen. In addition to
20 their specificity, the monoclonal antibodies are advantageous in that they may be synthesized uncontaminated by other antibodies. The modifier "monoclonal" indicates the character of the antibody as being obtained from a substantially homogeneous population of antibodies, and is not to be construed as requiring production of the antibody by any particular method. For example, the monoclonal antibodies to be used in accordance with
25 the present invention may be made by the hybridoma method first described by Kohler *et al* (1975) *Nature* 256:495, or may be made by recombinant DNA methods (see, US 4816567). The monoclonal antibodies may also be isolated from phage antibody libraries using the techniques described in Clackson *et al* (1991) *Nature*, 352:624-628; Marks *et al* (1991) *J. Mol. Biol.*, 222:581-597 or from transgenic mice carrying a fully human immunoglobulin
30 system (Lonberg (2008) *Curr. Opinion* 20(4):450-459).

The monoclonal antibodies herein specifically include "chimeric" antibodies in which a portion of the heavy and/or light chain is identical with or homologous to corresponding sequences in antibodies derived from a particular species or belonging to a particular
35 antibody class or subclass, while the remainder of the chain(s) is identical with or homologous to corresponding sequences in antibodies derived from another species or

belonging to another antibody class or subclass, as well as fragments of such antibodies, so long as they exhibit the desired biological activity (US 4816567; and Morrison *et al* (1984) *Proc. Natl. Acad. Sci. USA*, 81:6851-6855). Chimeric antibodies include "primatized" antibodies comprising variable domain antigen-binding sequences derived from a non-
 5 human primate (e.g. Old World Monkey or Ape) and human constant region sequences.

An "intact antibody" herein is one comprising a VL and VH domains, as well as a light chain constant domain (CL) and heavy chain constant domains, CH1, CH2 and CH3. The constant domains may be native sequence constant domains (e.g. human native sequence
 10 constant domains) or amino acid sequence variant thereof. The intact antibody may have one or more "effector functions" which refer to those biological activities attributable to the Fc region (a native sequence Fc region or amino acid sequence variant Fc region) of an antibody. Examples of antibody effector functions include C1q binding; complement dependent cytotoxicity; Fc receptor binding; antibody-dependent cell-mediated cytotoxicity
 15 (ADCC); phagocytosis; and down regulation of cell surface receptors such as B cell receptor and BCR.

Depending on the amino acid sequence of the constant domain of their heavy chains, intact antibodies can be assigned to different "classes." There are five major classes of intact
 20 antibodies: IgA, IgD, IgE, IgG, and IgM, and several of these may be further divided into "subclasses" (isotypes), e.g., IgG1, IgG2, IgG3, IgG4, IgA, and IgA2. The heavy-chain constant domains that correspond to the different classes of antibodies are called α , δ , ϵ , γ , and μ , respectively. The subunit structures and three-dimensional configurations of different classes of immunoglobulins are well known.

25

Humanisation

Techniques to reduce the *in vivo* immunogenicity of a non-human antibody or antibody fragment include those termed "humanisation".

30 A "humanized antibody" refers to a polypeptide comprising at least a portion of a modified variable region of a human antibody wherein a portion of the variable region, preferably a portion substantially less than the intact human variable domain, has been substituted by the corresponding sequence from a non-human species and wherein the modified variable region is linked to at least another part of another protein, preferably the constant region of a
 35 human antibody. The expression "humanized antibodies" includes human antibodies in which one or more complementarity determining region ("CDR") amino acid residues and/or

one or more framework region ("FW" or "FR") amino acid residues are substituted by amino acid residues from analogous sites in rodent or other non-human antibodies. The expression "humanized antibody" also includes an immunoglobulin amino acid sequence variant or fragment thereof that comprises an FR having substantially the amino acid sequence of a human immunoglobulin and a CDR having substantially the amino acid sequence of a non-human immunoglobulin.

"Humanized" forms of non-human (e.g., murine) antibodies are chimeric antibodies that contain minimal sequence derived from non-human immunoglobulin. Or, looked at another way, a humanized antibody is a human antibody that also contains selected sequences from non-human (e.g. murine) antibodies in place of the human sequences. A humanized antibody can include conservative amino acid substitutions or non-natural residues from the same or different species that do not significantly alter its binding and/or biologic activity. Such antibodies are chimeric antibodies that contain minimal sequence derived from non-human immunoglobulins.

There are a range of humanisation techniques, including 'CDR grafting', 'guided selection', 'deimmunization', 'resurfacing' (also known as 'veneering'), 'composite antibodies', 'Human String Content Optimisation' and framework shuffling.

CDR grafting

In this technique, the humanized antibodies are human immunoglobulins (recipient antibody) in which residues from a complementary-determining region (CDR) of the recipient antibody are replaced by residues from a CDR of a non-human species (donor antibody) such as mouse, rat, camel, bovine, goat, or rabbit having the desired properties (in effect, the non-human CDRs are 'grafted' onto the human framework). In some instances, framework region (FR) residues of the human immunoglobulin are replaced by corresponding non-human residues (this may happen when, for example, a particular FR residue has significant effect on antigen binding).

Furthermore, humanized antibodies can comprise residues that are found neither in the recipient antibody nor in the imported CDR or framework sequences. These modifications are made to further refine and maximize antibody performance. Thus, in general, a humanized antibody will comprise all of at least one, and in one aspect two, variable domains, in which all or all of the hypervariable loops correspond to those of a non-human immunoglobulin and all or substantially all of the FR regions are those of a human

immunoglobulin sequence. The humanized antibody optionally also will comprise at least a portion of an immunoglobulin constant region (Fc), or that of a human immunoglobulin.

Guided selection

- 5 The method consists of combining the V_H or V_L domain of a given non-human antibody specific for a particular epitope with a human V_H or V_L library and specific human V domains are selected against the antigen of interest. This selected human V_H is then combined with a V_L library to generate a completely human $V_H V_L$ combination. The method is described in Nature Biotechnology (N.Y.) 12, (1994) 899-903.

10

Composite antibodies

- In this method, two or more segments of amino acid sequence from a human antibody are combined within the final antibody molecule. They are constructed by combining multiple human V_H and V_L sequence segments in combinations which limit or avoid human T cell epitopes in the final composite antibody V regions. Where required, T cell epitopes are limited or avoided by, exchanging V region segments contributing to or encoding a T cell epitope with alternative segments which avoid T cell epitopes. This method is described in US 2008/0206239 A1.

Deimmunization

- 20 This method involves the removal of human (or other second species) T-cell epitopes from the V regions of the therapeutic antibody (or other molecule). The therapeutic antibodies V-region sequence is analysed for the presence of MHC class II- binding motifs by, for example, comparison with databases of MHC-binding motifs (such as the "motifs" database hosted at www.wehi.edu.au). Alternatively, MHC class II- binding motifs may be identified using computational threading methods such as those devised by Altuvia et al. (J. Mol. Biol. 249 244-250 (1995)); in these methods, consecutive overlapping peptides from the V-region sequences are testing for their binding energies to MHC class II proteins. This data can then be combined with information on other sequence features which relate to successfully presented peptides, such as amphipathicity, Rothbard motifs, and cleavage sites for cathepsin B and other processing enzymes.

30

- Once potential second species (e.g. human) T-cell epitopes have been identified, they are eliminated by the alteration of one or more amino acids. The modified amino acids are usually within the T-cell epitope itself, but may also be adjacent to the epitope in terms of the primary or secondary structure of the protein (and therefore, may not be adjacent in the

35

primary structure). Most typically, the alteration is by way of substitution but, in some circumstances amino acid addition or deletion will be more appropriate.

All alterations can be accomplished by recombinant DNA technology, so that the final molecule may be prepared by expression from a recombinant host using well established methods such as Site Directed Mutagenesis. However, the use of protein chemistry or any other means of molecular alteration is also possible.

Resurfacing

10 This method involves:

(a) determining the conformational structure of the variable region of the non-human (e.g. rodent) antibody (or fragment thereof) by constructing a three-dimensional model of the non-human antibody variable region;

15 (b) generating sequence alignments using relative accessibility distributions from x-ray crystallographic structures of a sufficient number of non-human and human antibody variable region heavy and light chains to give a set of heavy and light chain framework positions wherein the alignment positions are identical in 98% of the sufficient number of non-human antibody heavy and light chains;

20 (c) defining for the non-human antibody to be humanized, a set of heavy and light chain surface exposed amino acid residues using the set of framework positions generated in step (b);

25 (d) identifying from human antibody amino acid sequences a set of heavy and light chain surface exposed amino acid residues that is most closely identical to the set of surface exposed amino acid residues defined in step (c), wherein the heavy and light chain from the human antibody are or are not naturally paired;

(e) substituting, in the amino acid sequence of the non-human antibody to be humanized, the set of heavy and light chain surface exposed amino acid residues defined in step (c) with the set of heavy and light chain surface exposed amino acid residues identified in step (d);

30 (f) constructing a three-dimensional model of the variable region of the non-human antibody resulting from the substituting specified in step (e);

(g) identifying, by comparing the three-dimensional models constructed in steps (a) and (f), any amino acid residues from the sets identified in steps (c) or (d), that are within 5 Angstroms of any atom of any residue of the complementarity determining regions of the non-human antibody to be humanized; and

35

(h) changing any residues identified in step (g) from the human to the original non-human amino acid residue to thereby define a non-human antibody humanizing set of surface exposed amino acid residues; with the proviso that step (a) need not be conducted first, but must be conducted prior to step (g).

5

Superhumanization

The method compares the non-human sequence with the functional human germline gene repertoire. Those human genes encoding canonical structures identical or closely related to the non-human sequences are selected. Those selected human genes with highest
10 homology within the CDRs are chosen as FR donors. Finally, the non-human CDRs are grafted onto these human FRs. This method is described in patent WO 2005/079479 A2.

Human String Content Optimization

This method compares the non-human (e.g. mouse) sequence with the repertoire of human
15 germline genes and the differences are scored as Human String Content (HSC) that quantifies a sequence at the level of potential MHC/T-cell epitopes. The target sequence is then humanized by maximizing its HSC rather than using a global identity measure to generate multiple diverse humanized variants (described in Molecular Immunology, 44, (2007) 1986–1998).

20

Framework Shuffling

The CDRs of the non-human antibody are fused in-frame to cDNA pools encompassing all known heavy and light chain human germline gene frameworks. Humanised antibodies are then selected by e.g. panning of the phage displayed antibody library. This is described in
25 *Methods* **36**, 43-60 (2005).

Examples of cell binding agents include those agents described for use in WO 2007/085930.

Tumour-associate antigens and cognate antibodies for use in embodiments of the present
30 invention are listed below.

TUMOR-ASSOCIATED ANTIGENS AND COGNATE ANTIBODIES

(1) *BMPR1B* (bone morphogenetic protein receptor-type 1B)

Nucleotide

35 Genbank accession no. NM_001203

Genbank version no. NM_001203.2 GI:169790809

Genbank record update date: Sep 23, 2012 02:06 PM

Polypeptide

5 Genbank accession no. NP_001194

Genbank version no. NP_001194.1 GI:4502431

Genbank record update date: Sep 23, 2012 02:06 PM

Cross-references

- 10 ten Dijke, P., *et al Science* 264 (5155): 101-104 (1994), *Oncogene* 14 10 (11):1377-1382 (1997)); WO2004/063362 (Claim 2); WO2003/042661 (Claim 12); US2003/134790-A1 (Page 38-39); WO2002/102235 (Claim 13; Page 296); WO2003/055443 (Page 91-92); WO2002/99122 (Example 2; Page 528-530); WO2003/029421 (Claim 6); WO2003/024392 (Claim 2; Fig 112); WO2002/98358 (Claim 1; Page 183); WO2002/54940 (Page 100-101); WO2002/59377 (Page 349-350); WO2002/30268 (Claim 27; Page 376);
- 15 WO2001/48204 (Example; Fig 4); NP_001194 bone morphogenetic protein receptor, type IB /pid=NP_001194.1.; MIM:603248; AY065994

(2) E16 (LAT1, SLC7A5)

20 Nucleotide

Genbank accession no. NM_003486

Genbank version no. NM_003486.5 GI:71979931

Genbank record update date: Jun 27, 2012 12:06 PM

25 Polypeptide

Genbank accession no. NP_003477

Genbank version no. NP_003477.4 GI:71979932

Genbank record update date: Jun 27, 2012 12:06 PM

30 Cross references

Biochem. Biophys. Res.

Commun. 255 (2), 283-288 (1999), *Nature* 395 (6699):288-291 (1998), Gaugitsch, H.W., *et al* (1992) *J. Biol. Chem.* 267 (16):11267-11273); WO2004/048938 (Example 2);

WO2004/032842 (Example IV); WO2003/042661 (Claim 12); WO2003/016475 (Claim 1);

- 35 WO2002/78524 (Example 2); WO2002/99074 (Claim 19; Page 127-129); WO2002/86443 (Claim 27; Pages 222, 393); WO2003/003906 (Claim 10; Page 293); WO2002/64798 (Claim

33; Page 93-95); WO2000/14228 (Claim 5; Page 133-136); US2003/224454 (Fig 3);
25 WO2003/025138 (Claim 12; Page 150); NP_003477 solute carrier family 7 (cationic
amino

acid transporter, y⁺system), member 5 /pid=NP_003477.3 - Homo sapiens;

5 MIM:600182;; NM_015923.

(3) STEAP1 (*six transmembrane epithelial antigen of prostate*)

Nucleotide

Genbank accession no. NM_012449

10 Genbank version no. NM_012449.2 GI:22027487

Genbank record update date: Sep 9, 2012 02:57 PM

Polypeptide

Genbank accession no. NP_036581

15 Genbank version no. NP_036581.1 GI:9558759

Genbank record update date: Sep 9, 2012 02:57 PM

Cross references

Cancer Res. 61 (15), 5857-5860 (2001), Hubert, R.S., *et al* (1999) *Proc. Natl.*

20 *Acad. Sci. U.S.A.* 96 (25):14523-14528); WO2004/065577 (Claim 6); WO2004/027049 (Fig
1L); EP1394274 (Example 11); WO2004/016225 (Claim 2); WO2003/042661 (Claim 12);
US2003/157089 (Example 5); US2003/185830 (Example 5); US2003/064397 (Fig 2);
WO2002/89747 (Example 5; Page 618-619); WO2003/022995 (Example 9; Fig 13A,
35 Example 53; Page 173, Example 2; Fig 2A); six transmembrane epithelial
25 antigen of the prostate; MIM:604415.

(4) 0772P (CA125, MUC16)

Nucleotide

Genbank accession no. AF361486

30 Genbank version no. AF361486.3 GI:34501466

Genbank record update date: Mar 11, 2010 07:56 AM

Polypeptide

Genbank accession no. AAK74120

35 Genbank version no. AAK74120.3 GI:34501467

Genbank record update date: Mar 11, 2010 07:56 AM

Cross references

J. Biol. Chem. 276 (29):27371-27375 (2001)); WO2004/045553 (Claim 14); WO2002/92836 (Claim 6; Fig 12); WO2002/83866 (Claim 15; Page 116-121); US2003/124140 (Example 16);
 5 GI:34501467;

(5) MPF (MPF, MSLN, SMR, megakaryocyte potentiating factor, mesothelin)

Nucleotide

Genbank accession no. NM_005823

10 Genbank version no. NM_005823.5 GI:293651528

Genbank record update date: Sep 2, 2012 01:47 PM

Polypeptide

Genbank accession no. NP_005814

15 Genbank version no. NP_005814.2 GI:53988378

Genbank record update date: Sep 2, 2012 01:47 PM

Cross references

Yamaguchi, N., *et al Biol. Chem.* 269 (2), 805-808 (1994), *Proc. Natl. Acad. Sci. U.S.A.* 96
 20 (20):11531-11536 (1999), *Proc. Natl. Acad. Sci. U.S.A.* 93 10 (1):136-140 (1996), *J. Biol. Chem.* 270 (37):21984-21990 (1995)); WO2003/101283 (Claim 14); (WO2002/102235 (Claim 13; Page 287-288); WO2002/101075 (Claim 4; Page 308- 309); WO2002/71928 (Page 320-321); WO94/10312 (Page 52-57); IM:601051.

25 *(6) Napi3b (NAPI-3B, NPTIIb, SLC34A2, solute carrier family 34 (sodium phosphate), member 2, type II sodium-dependent phosphate transporter 3b)*

Nucleotide

Genbank accession no. NM_006424

Genbank version no. NM_006424.2 GI:110611905

30 Genbank record update date: Jul 22, 2012 03:39 PM

Polypeptide

Genbank accession no. NP_006415

Genbank version no. NP_006415.2 GI:110611906

35 Genbank record update date: Jul 22, 2012 03:39 PM

Cross references

J. Biol. Chem. 277 (22):19665-19672 (2002), *Genomics* 62 (2):281-284 (1999), Feild, J.A., *et al* (1999) *Biochem. Biophys. Res. Commun.* 258 (3):578-582; WO2004/022778 (Claim 2); EP1394274 (Example 11); WO2002/102235 (Claim 13; Page 20 326); EP0875569 (Claim 1; Page 17-19); WO2001/57188 (Claim 20; Page 329); WO2004/032842 (Example IV); WO2001/75177 (Claim 24; Page 139-140); MIM:604217.

(7) *Sema 5b* (FLJ10372, KIAA1445, Mm.42015, SEMA5B, SEMAG, Semaphorin 5b Hlog, 25 sema domain, seven thrombospondin repeats (type 1 and type 1-like), transmembrane domain (TM) and short cytoplasmic domain, (semaphorin) 5B)

Nucleotide

Genbank accession no. AB040878

Genbank version no. AB040878.1 GI:7959148

Genbank record update date: Aug 2, 2006 05:40 PM

Polypeptide

Genbank accession no. BAA95969

Genbank version no. BAA95969.1 GI:7959149

Genbank record update date: Aug 2, 2006 05:40 PM

Cross references

Nagase T., *et al* (2000) *DNA Res.* 7 (2):143-150; WO2004/000997 (Claim 1); WO2003/003984 (Claim 1); WO2002/06339 (Claim 1; Page 50); WO2001/88133 (Claim 1; Page 41-43, 48-58); WO2003/054152 (Claim 20); WO2003/101400 (Claim 11); Accession: 30 Q9P283; Genew; HGNC:10737

(8) *PSCA* hlg (2700050C12Rik, C530008O16Rik, RIKEN cDNA 2700050C12, RIKEN cDNA 2700050C12 gene)

Nucleotide

30 Genbank accession no. AY358628

Genbank version no. AY358628.1 GI:37182377

Genbank record update date: Dec 1, 2009 04:15 AM

Polypeptide

35 Genbank accession no. AAQ88991

Genbank version no. AAQ88991.1 GI:37182378

Genbank record update date: Dec 1, 2009 04:15 AM

Cross references

Ross *et al* (2002) *Cancer Res.* 62:2546-2553; US2003/129192 (Claim 2); US2004/044180
 5 (Claim 12); US2004/044179 35 (Claim 11); US2003/096961 (Claim 11); US2003/232056
 (Example 5); WO2003/105758 16 (Claim 12); US2003/206918 (Example 5); EP1347046
 (Claim 1); WO2003/025148 (Claim 20); GI:37182378.

(9) ETBR (Endothelin type B receptor)

10 Nucleotide

Genbank accession no. AY275463

Genbank version no. AY275463.1 GI:30526094

Genbank record update date: Mar 11, 2010 02:26 AM

15 Polypeptide

Genbank accession no. AAP32295

Genbank version no. AAP32295.1 GI:30526095

Genbank record update date: Mar 11, 2010 02:26 AM

20 Cross references

Nakamuta M., *et al Biochem. Biophys. Res. Commun.* 177, 34-39, 1991; Ogawa Y., *et al*
Biochem. Biophys. Res. Commun. 178, 248-255, 1991; Arai H., *et al Jpn. Circ. J.* 56, 1303-
 1307, 1992; Arai H., *et al J. Biol. Chem.* 268, 3463-3470, 1993; Sakamoto A., Yanagisawa
 M., *et al Biochem. Biophys. Res. Commun.* 178, 656-663, 1991; Elshourbagy N.A., *et al J.*
 25 *Biol. Chem.* 268, 3873-3879, 1993; Haendler B., *et al J. Cardiovasc. Pharmacol.* 20, s1-S4,
 1992; Tsutsumi M., *et al Gene* 228, 43-49, 1999; Strausberg R.L., *et al Proc. Natl. Acad. Sci.*
U.S.A. 99, 16899-16903, 2002; Bourgeois C., *et al J. Clin. Endocrinol. Metab.* 82, 3116-
 3123, 1997;

Okamoto Y., *et al Biol. Chem.* 272, 21589-21596, 1997; Verheij J.B., *et al Am. J. Med.*

30 *Genet.* 108, 223-225, 2002; Hofstra R.M.W., *et al Eur. J. Hum. Genet.* 5, 180-185, 1997;

Puffenberger E.G., *et al Cell* 79, 1257-1266, 1994; Attie T., *et al, Hum. Mol. Genet.* 4, 2407-
 15 2409, 1995; Auricchio A., *et al Hum. Mol. Genet.* 5:351-354, 1996; Amiel J., *et al Hum.*
Mol.

Genet. 5, 355-357, 1996; Hofstra R.M.W., *et al Nat. Genet.* 12, 445-447, 1996; Svensson

35 P.J., *et al Hum. Genet.* 103, 145-148, 1998; Fuchs S., *et al Mol. Med.* 7, 115-124, 2001;

Pingault V., *et al (2002) Hum. Genet.* 111, 198-206; WO2004/045516 (Claim 1);

WO2004/048938 (Example 2); WO2004/040000 (Claim 151); WO2003/087768 (Claim 1);
 20 WO2003/016475 (Claim 1); WO2003/016475 (Claim 1); WO2002/61087 (Fig 1);
 WO2003/016494 (Fig 6); WO2003/025138 (Claim 12; Page 144); WO2001/98351 (Claim 1;
 Page 124-125); EP0522868 (Claim 8; Fig 2); WO2001/77172 (Claim 1; Page 297-299);
 5 US2003/109676; US6518404 (Fig 3); US5773223 (Claim 1a; Col 31-34); WO2004/001004.

(10) MSG783 (RNF124, hypothetical protein FLJ20315)

Nucleotide

Genbank accession no. NM_017763

10 Genbank version no. NM_017763.4 GI:167830482

Genbank record update date: Jul 22, 2012 12:34 AM

Polypeptide

Genbank accession no. NP_060233

15 Genbank version no. NP_060233.3 GI:56711322

Genbank record update date: Jul 22, 2012 12:34 AM

Cross references

WO2003/104275 (Claim 1); WO2004/046342 (Example 2); WO2003/042661 (Claim 12);
 20 WO2003/083074 (Claim 14; Page 61); WO2003/018621 (Claim 1); WO2003/024392 (Claim
 2; Fig 93); WO2001/66689 (Example 6); LocusID:54894.

**(11) STEAP2 (HGNC_8639, IPCA-1, PCANAP1, STAMP1, STEAP2, STMP, prostate cancer
 associated gene 1, prostate cancer associated protein 1, six transmembrane epithelial
 25 antigen of prostate 2, six transmembrane prostate protein)**

Nucleotide

Genbank accession no. AF455138

Genbank version no. AF455138.1 GI:22655487

Genbank record update date: Mar 11, 2010 01:54 AM

30

Polypeptide

Genbank accession no. AAN04080

Genbank version no. AAN04080.1 GI:22655488

Genbank record update date: Mar 11, 2010 01:54 AM

35

Cross references

Lab. Invest. 82 (11):1573-1582 (2002)); WO2003/087306; US2003/064397 (Claim 1; Fig 1); WO2002/72596 (Claim 13; Page 54-55); WO2001/72962 (Claim 1; Fig 4B); 35 WO2003/104270 (Claim 11); WO2003/104270 (Claim 16); US2004/005598 (Claim 22); 5 WO2003/042661 (Claim 12); US2003/060612 (Claim 12; Fig 10); WO2002/26822 (Claim 23; Fig 2); WO2002/16429 (Claim 12; Fig 10); GI:22655488.

(12) TrpM4 (BR22450, FLJ20041, TRPM4, TRPM4B, transient receptor potential cation 5 channel, subfamily M, member 4)

10 Nucleotide

Genbank accession no. NM_017636
Genbank version no. NM_017636.3 GI:304766649
Genbank record update date: Jun 29, 2012 11:27 AM

15 Polypeptide

Genbank accession no. NP_060106
Genbank version no. NP_060106.2 GI:21314671
Genbank record update date: Jun 29, 2012 11:27 AM

20 Cross references

Xu, X.Z., et al *Proc. Natl. Acad. Sci. U.S.A.* 98 (19):10692-10697 (2001), *Cell* 109 (3):397-407 (2002), *J. Biol. Chem.* 278 (33):30813-30820 (2003)); US2003/143557 (Claim 4); WO2000/40614 (Claim 14; Page 100-103); WO2002/10382 (Claim 1; Fig 9A); WO2003/042661 (Claim 12); WO2002/30268 (Claim 27; Page 391); US2003/219806 (Claim 25 4); WO2001/62794 (Claim 10 14; Fig 1A-D); MIM:606936.

(13) CRIPTO (CR, CR1, CRGF, CRIPTO, TDGF1, teratocarcinoma-derived growth factor)

Nucleotide

30 Genbank accession no. NM_003212
Genbank version no. NM_003212.3 GI:292494881
Genbank record update date: Sep 23, 2012 02:27 PM

Polypeptide

35 Genbank accession no. NP_003203
Genbank version no. NP_003203.1 GI:4507425

Genbank record update date: Sep 23, 2012 02:27 PM

Cross references

Ciccodicola, A., et al *EMBO J.* 8 (7):1987-1991 (1989), *Am. J. Hum. Genet.* 49 (3):555-565
 5 (1991)); US2003/224411 (Claim 1); WO2003/083041 (Example 1); WO2003/034984 (Claim
 12); WO2002/88170 (Claim 2; Page 52-53); WO2003/024392 (Claim 2; Fig 58);
 WO2002/16413 (Claim 1; Page 94-95, 105); WO2002/22808 (Claim 2; Fig 1); US5854399
 (Example 2; Col 17-18); US5792616 (Fig 2); MIM:187395.

10 **(14)** *CD21 (CR2 (Complement receptor 2) or C3DR (C3d/Epstein Barr virus receptor) or
 Hs.73792)*

Nucleotide

Genbank accession no M26004

Genbank version no. M26004.1 GI:181939

15 Genbank record update date: Jun 23, 2010 08:47 AM

Polypeptide

Genbank accession no. AAA35786

Genbank version no. AAA35786.1 GI:181940

20 Genbank record update date: Jun 23, 2010 08:47 AM

Cross references

Fujisaku et al (1989) *J. Biol. Chem.* 264 (4):2118-2125); Weis J.J., et al *J. Exp. Med.* 167,
 1047-1066, 1988; Moore M., et al *Proc. Natl. Acad. Sci. U.S.A.* 84, 9194-9198, 1987; Barel
 25 M., et al *Mol. Immunol.* 35, 1025-1031, 1998; Weis J.J., et al *Proc. Natl. Acad. Sci. U.S.A.*
 83, 5639-5643, 1986; Sinha S.K., et al (1993) *J. Immunol.* 150, 5311-5320; WO2004/045520
 (Example 4); US2004/005538 (Example 1); WO2003/062401 (Claim 9); WO2004/045520
 (Example 4); WO91/02536 (Fig 9.1-9.9); WO2004/020595 (Claim 1); Accession: P20023;
 Q13866; Q14212; EMBL; M26004; AAA35786.1.

30

(15) *CD79b (CD79B, CD79 β , Igb (immunoglobulin-associated beta), B29)*

Nucleotide

Genbank accession no NM_000626

Genbank version no. NM_000626.2 GI:90193589

35 Genbank record update date: Jun 26, 2012 01:53 PM

Polypeptide

Genbank accession no. NP_000617

Genbank version no. NP_000617.1 GI:11038674

Genbank record update date: Jun 26, 2012 01:53 PM

5

Cross references

Proc. Natl. Acad. Sci. U.S.A. (2003) 100 (7):4126-

4131, *Blood* (2002) 100 (9):3068-3076, Muller *et al* (1992) *Eur. J. Immunol.* 22 (6):1621-

1625); WO2004/016225 (claim 2, Fig 140); WO2003/087768, US2004/101874 (claim 1,

10 page 102); WO2003/062401 (claim 9); WO2002/78524 (Example 2); US2002/150573 (claim

35 5, page 15); US5644033; WO2003/048202 (claim 1, pages 306 and 309); WO 99/58658,

US6534482 (claim 13, Fig 17A/B); WO2000/55351 (claim 11, pages 1145-1146);

MIM:147245

15 **(16)** *FcRH2 (IFGP4, IRTA4, SPAP1A (SH2 domain containing phosphatase anchor protein*
5 1a), SPAP1B, SPAP1C)

Nucleotide

Genbank accession no NM_030764

Genbank version no. NM_030764.3 GI:227430280

20 Genbank record update date: Jun 30, 2012 12:30 AM

Polypeptide

Genbank accession no. NP_110391

Genbank version no. NP_110391.2 GI:19923629

25 Genbank record update date: Jun 30, 2012 12:30 AM

Cross references

AY358130); *Genome Res.* 13 (10):2265-2270 (2003), *Immunogenetics* 54 (2):87-95 (2002),

Blood 99 (8):2662-2669 (2002), *Proc. Natl. Acad. Sci. U.S.A.* 98 (17):9772-9777 (2001), Xu,

30 M.J., *et al* (2001) *Biochem. Biophys. Res. Commun.* 280 (3):768-775; WO2004/016225

(Claim 2); WO2003/077836; WO2001/38490 (Claim 5; Fig 18D-1-18D-2); WO2003/097803

(Claim 12);

10 WO2003/089624 (Claim 25);: MIM:606509.

(17) HER2 (ErbB2)Nucleotide

Genbank accession no M11730

Genbank version no. M11730.1 GI:183986

5 Genbank record update date: Jun 23, 2010 08:47 AM

Polypeptide

Genbank accession no. AAA75493

Genbank version no. AAA75493.1 GI:306840

10 Genbank record update date: Jun 23, 2010 08:47 AM

Cross references

Coussens L., *et al Science* (1985) 230(4730):1132-1139); Yamamoto T., *et al Nature* 319, 230-234, 1986; Semba K., *et al Proc. Natl. Acad. Sci. U.S.A.* 82, 6497-6501, 1985; Swiercz
 15 J.M., *et al J. Cell Biol.* 165, 869- 15 880, 2004; Kuhns J.J., *et al J. Biol. Chem.* 274, 36422-
 36427, 1999; Cho H.-S., *et al Nature* 421, 756-760, 2003; Ehsani A., *et al (1993) Genomics*
 15, 426-429; WO2004/048938 (Example 2); WO2004/027049 (Fig 1I); WO2004/009622;
 WO2003/081210;
 WO2003/089904 (Claim 9); WO2003/016475 (Claim 1); US2003/118592; WO2003/008537
 20 (Claim 1); WO2003/055439 (Claim 29; Fig 1A-B); WO2003/025228 (Claim 37; Fig 5C);
 20 WO2002/22636 (Example 13; Page 95-107); WO2002/12341 (Claim 68; Fig 7);
 WO2002/13847 (Page 71-74); WO2002/14503 (Page 114-117); WO2001/53463 (Claim 2;
 Page 41-46); WO2001/41787 (Page 15); WO2000/44899 (Claim 52; Fig 7); WO2000/20579
 (Claim 3; Fig 2); US5869445 (Claim 3; Col 31-38); WO9630514 (Claim 2; Page 56-61);
 25 EP1439393 (Claim 7); WO2004/043361 (Claim 7); WO2004/022709; WO2001/00244
 25 (Example 3; Fig 4); Accession: P04626; EMBL; M11767; AAA35808.1. EMBL; M11761;
 AAA35808.1

ANTIBODIES

30 Abbott: US20110177095

For example, an antibody comprising CDRs having overall at least 80% sequence
 identity to CDRs having amino acid sequences of SEQ ID NO:3 (CDR-H1), SEQ ID
 NO:4 (CDR-H2), SEQ ID NO:5 (CDR-H3), SEQ ID NO:104 and/or SEQ ID NO:6
 (CDR-L1), SEQ ID NO:7 (CDR-L2), and SEQ ID NO:8 (CDR-L3), wherein the anti-
 35 HER2 antibody or anti-HER2 binding fragment has reduced immunogenicity as
 compared to an antibody having a VH of SEQ ID NO:1 and a VL of SEQ ID NO:2.

Biogen: US20100119511

For example, ATCC accession numbers: PTA-10355, PTA-10356, PTA-10357, PTA-10358

- 5 For example, a purified antibody molecule that binds to HER2 comprising a all six CDR's from an antibody selected from the group consisting of BIIB71F10 (SEQ ID NOs:11, 13), BIIB69A09 (SEQ ID NOs:15, 17); BIIB67F10 (SEQ ID NOs:19, 21); BIIB67F11 (SEQ ID NOs:23, 25), BIIB66A12 (SEQ ID NOs:27, 29), BIIB66C01 (SEQ ID NOs:31, 33), BIIB65C10 (SEQ ID NOs:35, 37), BIIB65H09 (SEQ ID NOs:39, 41) and BIIB65B03 (SEQ ID NOs:43, 45), or CDRs which are identical or which have no more than two alterations from said CDRs.
- 10

Herceptin (Genentech) - US6,054,297; ATCC accession no. CRL-10463 (Genentech)

15 Pertuzumab (Genentech)

US20110117097

for example, see SEQ IDs No. 15&16, SEQ IDs No. 17&18, SEQ IDs No. 23&24 & ATCC accession numbers HB-12215, HB-12216, CRL 10463, HB-12697.

20 US20090285837

US20090202546

for example, ATCC accession numbers: HB-12215, HB-12216, CRL 10463, HB-12698.

US20060088523

- 25
- for example, ATCC accession numbers: HB-12215, HB-12216
 - for example, an antibody comprising the variable light and variable heavy amino acid sequences in SEQ ID Nos. 3 and 4, respectively.
 - for example, an antibody comprising a light chain amino acid sequence selected from SEQ ID No. 15 and 23, and a heavy chain amino acid sequence selected from SEQ ID No. 16 and 24
- 30

US20060018899

- for example, ATCC accession numbers: (7C2) HB-12215, (7F3) HB-12216, (4D5) CRL-10463, (2C4) HB-12697.
 - for example, an antibody comprising the amino acid sequence in SEQ ID No. 23, or a deamidated and/or oxidized variant thereof.
- 35

US2011/0159014

- for example, an antibody having a light chain variable domain comprising the hypervariable regions of SEQ ID NO: 1".
- For example, an antibody having a heavy chain variable domain comprising the hypervariable regions of SEQ ID NO: 2.

US20090187007

Glycotope: TrasGEX™ antibody

- 10 For example, see International Joint Cancer Institute and Changhai Hospital Cancer Cent: HMTI-Fc Ab - Gao J., et al *BMB Rep.* 2009 Oct 31;42(10):636-41.

Symphogen: US20110217305

- 15 Union Stem Cell & Gene Engineering, China - Liu HQ., et al *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi.* 2010 May;26(5):456-8.

(18) NCA (CEACAM6)

20 Nucleotide

Genbank accession no M18728

Genbank version no. M18728.1 GI:189084

Genbank record update date: Jun 23, 2010 08:48 AM

25 Polypeptide

Genbank accession no. AAA59907

Genbank version no. AAA59907.1 GI:189085

Genbank record update date: Jun 23, 2010 08:48 AM

30 Cross references

- Barnett T., et al *Genomics* 3, 59-66, 1988; Tawaragi Y., et al *Biochem. Biophys. Res. Commun.* 150, 89-96, 1988; Strausberg R.L., et al *Proc. Natl. Acad. Sci. U.S.A.* 99:16899-16903, 2002; WO2004/063709; EP1439393 (Claim 7); WO2004/044178 (Example 4); WO2004/031238; WO2003/042661 (Claim 12); WO2002/78524 (Example 2);
- 35 WO2002/86443 (Claim 27; Page 427); WO2002/60317 (Claim 2); Accession: P40199; Q14920; EMBL; M29541; AAA59915.1.

EMBL; M18728.

(19) MDP (DPEP1)

Nucleotide

- 5 Genbank accession no BC017023
Genbank version no. BC017023.1 GI:16877538
Genbank record update date: Mar 6, 2012 01:00 PM

Polypeptide

- 10 Genbank accession no. AAH17023
Genbank version no. AAH17023.1 GI:16877539
Genbank record update date: Mar 6, 2012 01:00 PM

Cross references

- 15 *Proc. Natl. Acad. Sci. U.S.A.* 99 (26):16899-16903 (2002)); WO2003/016475 (Claim 1);
WO2002/64798 (Claim 33; Page 85- 87); JP05003790 (Fig 6-8); WO99/46284 (Fig 9);
MIM:179780.

(20) IL20R-alpha (IL20Ra, ZCYTOR7)

20 Nucleotide

Genbank accession no AF184971
Genbank version no. AF184971.1 GI:6013324
Genbank record update date: Mar 10, 2010 10:00 PM

25 Polypeptide

Genbank accession no. AAF01320
Genbank version no. AAF01320.1 GI:6013325
Genbank record update date: Mar 10, 2010 10:00 PM

30 Cross references

- Clark H.F., *et al Genome Res.* 13, 2265-2270, 2003; Mungall A.J., *et al Nature* 425, 805-811, 2003; Blumberg H., *et al Cell* 104, 9-19, 2001; Dumoutier L., *et al J. Immunol.* 167, 3545-3549,
2001; Parrish-Novak J., *et al J. Biol. Chem.* 277, 47517-47523, 2002; Pletnev S., *et al* (2003)
35 10 *Biochemistry* 42:12617-12624; Sheikh F., *et al* (2004) *J. Immunol.* 172, 2006-2010;
EP1394274 (Example 11); US2004/005320 (Example 5); WO2003/029262 (Page 74-75);

WO2003/002717 (Claim 2; Page 63); WO2002/22153 (Page 45-47); US2002/042366 (Page 20-21); WO2001/46261 (Page 57-59); WO2001/46232 (Page 63-65); WO98/37193 (Claim 1; Page 55-59); Accession: Q9UHF4; Q6UWA9; Q96SH8; EMBL; AF184971; AAF01320.1.

5 **(21) *Brevican (BCAN, BEHAB)***

Nucleotide

Genbank accession no AF229053

Genbank version no. AF229053.1 GI:10798902

Genbank record update date: Mar 11, 2010 12:58 AM

10

Polypeptide

Genbank accession no. AAG23135

Genbank version no. AAG23135.1 GI:10798903

Genbank record update date: Mar 11, 2010 12:58 AM

15

Cross references

Gary S.C., *et al Gene* 256, 139-147, 2000; Clark H.F., *et al Genome Res.* 13, 2265-2270, 2003; Strausberg R.L., *et al Proc. Natl. Acad. Sci. U.S.A.* 99, 16899-16903, 2002;

US2003/186372 (Claim 11); US2003/186373 (Claim 11); US2003/119131 (Claim 1; Fig 52);

20 US2003/119122 (Claim 1; 20 Fig 52); US2003/119126 (Claim 1); US2003/119121 (Claim 1; Fig 52); US2003/119129 (Claim 1); US2003/119130 (Claim 1); US2003/119128 (Claim 1; Fig 52); US2003/119125 (Claim 1); WO2003/016475 (Claim 1); WO2002/02634 (Claim 1)

25 **(22) *EphB2R (DRT, ERK, Hek5, EPHT3, Tyro5)***

Nucleotide

Genbank accession no NM_004442

Genbank version no. NM_004442.6 GI:111118979

Genbank record update date: Sep 8, 2012 04:43 PM

30

Polypeptide

Genbank accession no. NP_004433

Genbank version no. NP_004433.2 GI:21396504

Genbank record update date: Sep 8, 2012 04:43 PM

35

Cross references

Chan, J. and Watt, V.M., Oncogene 6 (6), 1057-1061 (1991) Oncogene 10 (5):897-905 (1995), Annu. Rev. Neurosci. 21:309-345 (1998), Int. Rev. Cytol. 196:177-244 (2000)); WO2003042661 (Claim 12); WO200053216 (Claim 1; Page 41); WO2004065576 (Claim 1);
 5 WO2004020583 (Claim 9); WO2003004529 (Page 128-132); WO200053216 (Claim 1; Page 42); MIM:600997.

(23) ASLG659 (B7h)Nucleotide

10 Genbank accession no. AX092328
 Genbank version no. AX092328.1 GI:13444478
 Genbank record update date: Jan 26, 2011 07:37 AM

Cross references

15 US2004/0101899 (Claim 2); WO2003104399 (Claim 11); WO2004000221 (Fig 3);
 US2003/165504 (Claim 1); US2003/124140 (Example 2); US2003/065143 (Fig 60);
 WO2002/102235 (Claim 13; Page 299); US2003/091580 (Example 2); WO2002/10187
 (Claim 6; Fig 10); WO2001/94641 (Claim 12; Fig 7b); WO2002/02624 (Claim 13; Fig 1A-1B);
 US2002/034749 (Claim 54; Page 45-46); WO2002/06317 (Example 2; Page 320-321, Claim
 20 34; Page 321-322); WO2002/71928 (Page 468-469); WO2002/02587 (Example 1; Fig 1);
 WO2001/40269 (Example 3; Pages 190-192); WO2000/36107 (Example 2; Page 205-207);
 WO2004/053079 (Claim 12); WO2003/004989 (Claim 1); WO2002/71928 (Page 233-234,
 452-453); WO 01/16318.

25 (24) PSCA (Prostate stem cell antigen precursor)Nucleotide

Genbank accession no. AJ297436
 Genbank version no. AJ297436.1 GI:9367211
 Genbank record update date: Feb 1, 2011 11:25 AM

30

Polypeptide

Genbank accession no. CAB97347
 Genbank version no. CAB97347.1 GI:9367212
 Genbank record update date: Feb 1, 2011 11:25 AM

35

Cross references

Reiter R.E., *et al Proc. Natl. Acad. Sci. U.S.A.* 95, 1735-1740, 1998; Gu Z., *et al Oncogene* 19,

1288-1296, 2000; *Biochem. Biophys. Res. Commun.* (2000) 275(3):783-788;

- 5 WO2004/022709; EP1394274 (Example 11); US2004/018553 (Claim 17); WO2003/008537 (Claim 1); WO2002/81646 (Claim 1; Page 164); WO2003/003906 (Claim 10; Page 288); WO2001/40309 (Example 1; Fig 17); US2001/055751 (Example 1; Fig 1b); WO2000/32752 (Claim 18; Fig 1); WO98/51805 (Claim 17; Page 97); WO98/51824 (Claim 10; Page 94); WO98/40403 (Claim 2; Fig 1B); Accession: O43653; EMBL; AF043498; AAC39607.1

10

(25) GEDANucleotide

Genbank accession no AY260763

Genbank version no. AY260763.1 GI:30102448

- 15 Genbank record update date: Mar 11, 2010 02:24 AM

Polypeptide

Genbank accession no. AAP14954

Genbank version no. AAP14954.1 GI:30102449

- 20 Genbank record update date: Mar 11, 2010 02:24 AM

Cross references

AP14954 lipoma HMGIC fusion-partnerlike protein /pid=AAP14954.1 - Homo sapiens (human); WO2003/054152 (Claim 20); WO2003/000842 (Claim 1); WO2003/023013

- 25 (Example 3, Claim 20); US2003/194704 (Claim 45); GI:30102449;

(26) BAFF-R (B cell -activating factor receptor, BLyS receptor 3, BR3)Nucleotide

Genbank accession no AF116456

- 30 Genbank version no. AF116456.1 GI:4585274

Genbank record update date: Mar 10, 2010 09:44 PM

Polypeptide

Genbank accession no. AAD25356

- 35 Genbank version no. AAD25356.1 GI:4585275

Genbank record update date: Mar 10, 2010 09:44 PM

Cross references

BAFF receptor /pid=NP_443177.1 - Homo sapiens: Thompson, J.S., *et al Science* 293 (5537), 2108-2111 (2001); WO2004/058309; WO2004/011611; WO2003/045422 (Example; 5 Page 32-33); WO2003/014294 (Claim 35; Fig 6B); WO2003/035846 (Claim 70; Page 615-616); WO2002/94852 (Col 136-137); WO2002/38766 25 (Claim 3; Page 133); WO2002/24909 (Example 3; Fig 3); MIM:606269; NP_443177.1; NM_052945_1; AF132600

(27) CD22 (B-cell receptor CD22-B isoform, BL-CAM, Lyb-8, Lyb8, SIGLEC-2, FLJ22814)

10 Nucleotide

Genbank accession no AK026467

Genbank version no. AK026467.1 GI:10439337

Genbank record update date: Sep 11, 2006 11:24 PM

15 Polypeptide

Genbank accession no. BAB15489

Genbank version no. BAB15489.1 GI:10439338

Genbank record update date: Sep 11, 2006 11:24 PM

20 Cross references

Wilson *et al* (1991) *J. Exp. Med.* 173:137-146; 30 WO2003/072036 (Claim 1; Fig 1); IM:107266; NP_001762.1; NM_001771_1.

(27a) CD22 (CD22 molecule)

25 Nucleotide

Genbank accession no X52785

Genbank version no. X52785.1 GI:29778

Genbank record update date: Feb 2, 2011 10:09 AM

30 Polypeptide

Genbank accession no. CAA36988

Genbank version no. CAA36988.1 GI:29779

Genbank record update date: Feb 2, 2011 10:09 AM

35 Cross references

Stamenkovic I. *et al.*, *Nature* 345 (6270), 74-77 (1990)??

Other information

Official Symbol: CD22

Other Aliases: SIGLEC-2, SIGLEC2

- 5 Other Designations: B-cell receptor CD22; B-lymphocyte cell adhesion molecule; BL-CAM; CD22 antigen; T-cell surface antigen Leu-14; sialic acid binding Ig-like lectin 2; sialic acid-binding Ig-like lectin 2

ANTIBODIES

- 10 G5/44 (Inotuzumab): DiJoseph JF., et al *Cancer Immunol Immunother.* 2005 Jan;54(1):11-24.

Epratuzumab- Goldenberg DM., et al *Expert Rev Anticancer Ther.* 6(10): 1341-53, 2006.

15

(28) *CD79a (CD79A, CD79alpha), immunoglobulin-associated alpha, a B cell-specific protein that covalently interacts with Ig beta (CD79B) and forms a complex on the surface with Ig M*

35 molecules, transduces a signal involved in B-cell differentiation), pI: 4.84, MW: 25028 TM:

20 2

[P] Gene Chromosome: 19q13.2).

Nucleotide

Genbank accession no NM_001783

Genbank version no. NM_001783.3 GI:90193587

- 25 Genbank record update date: Jun 26, 2012 01:48 PM

Polypeptide

Genbank accession no. NP_001774

Genbank version no. NP_001774.1 GI:4502685

- 30 Genbank record update date: Jun 26, 2012 01:48 PM

Cross references

WO2003/088808, US2003/0228319; WO2003/062401 (claim 9); US2002/150573 (claim 4, pages 13-14); WO99/58658 (claim 13, Fig 16); WO92/07574 (Fig 1); US5644033; Ha et al

- 35 (1992) *J. Immunol.* 148(5):1526-1531; Müller et al (1992) *Eur. J. Immunol.* 22:1621-1625; Hashimoto et al (1994) *Immunogenetics* 40(4):287-295; Preud'homme et al (1992) *Clin. Exp.*

5 *Immunol.* 90(1):141-146; Yu *et al* (1992) *J. Immunol.* 148(2) 633-637; Sakaguchi *et al* (1988)

EMBO J. 7(11):3457-3464

- 5 **(29)** *CXCR5 (Burkitt's lymphoma receptor 1, a G protein-coupled receptor that is activated by the CXCL13 chemokine, functions in lymphocyte migration and humoral defense, plays a role in HIV-2 infection and perhaps development of AIDS, lymphoma, myeloma, and leukemia); 372 aa, pI: 8.54 MW: 41959 TM: 7 [P] Gene Chromosome: 11q23.3,*

Nucleotide

- 10 Genbank accession no NM_001716
Genbank version no. NM_001716.4 GI:342307092
Genbank record update date: Sep 30, 2012 01:49 PM

Polypeptide

- 15 Genbank accession no. NP_001707
Genbank version no. NP_001707.1 GI:4502415
Genbank record update date: Sep 30, 2012 01:49 PM

Cross references

- 20 WO2004/040000; WO2004/015426; US2003/105292 (Example 2); US6555339 (Example 2);
WO2002/61087 (Fig 1); WO2001/57188 (Claim 20, page 269); WO2001/72830 (pages 12-13);
WO2000/22129 (Example 1, pages 152-153, 15 Example 2, pages 254-256);
WO99/28468 (claim 1, page 38); US5440021 (Example 2, col 49-52); WO94/28931 (pages 56-58);
WO92/17497 (claim 7, Fig 5); Dobner *et al* (1992) *Eur. J. Immunol.* 22:2795-2799;
25 Barella *et al* (1995) *Biochem. J.* 309:773-779

(30) *HLA-DOB (Beta subunit of MHC class II molecule (Ia antigen) that binds peptides and presents them to CD4+ T lymphocytes); 273 aa, pI: 6.56, MW: 30820.TM: 1 [P] Gene Chromosome: 6p21.3)*

30 Nucleotide

Genbank accession no NM_002120
Genbank version no. NM_002120.3 GI:118402587
Genbank record update date: Sep 8, 2012 04:46 PM

35 Polypeptide

Genbank accession no. NP_002111

Genbank version no. NP_002111.1 GI:4504403

Genbank record update date: Sep 8, 2012 04:46 PM

Cross references

- 5 Tonnelle *et al* (1985) *EMBO J.* 4(11):2839-2847; Jonsson *et al* (1989) *Immunogenetics* 29(6):411-413; Beck *et al* (1992) *J. Mol. Biol.* 228:433-441; Strausberg *et al* (2002) *Proc. Natl. Acad. Sci USA* 99:16899- 16903; Servenius *et al* (1987) *J. Biol. Chem.* 262:8759-8766; Beck *et al* (1996) *J. Mol. Biol.* 25 255:1-13; Naruse *et al* (2002) *Tissue Antigens* 59:512-519; WO99/58658 (claim 13, Fig 15); US6153408 (Col 35-38); US5976551 (col 168-170);
- 10 US6011146 (col 145-146); Kasahara *et al* (1989) *Immunogenetics* 30(1):66-68; Larhammar *et al* (1985) *J. Biol. Chem.* 260(26):14111-14119

- (31) P2X5 (Purinergic receptor P2X ligand-gated ion channel 5, an ion channel gated by extracellular ATP, may be involved in synaptic transmission and neurogenesis, deficiency may contribute to the pathophysiology of idiopathic detrusor instability); 422 aa), pI: 7.63, MW: 47206 TM: 1 [P] Gene Chromosome: 17p13.3).
- 15

Nucleotide

Genbank accession no NM_002561

Genbank version no. NM_002561.3 GI:325197202

- 20 Genbank record update date: Jun 27, 2012 12:41 AM

Polypeptide

Genbank accession no. NP_002552

Genbank version no. NP_002552.2 GI:28416933

- 25 Genbank record update date: Jun 27, 2012 12:41 AM

Cross references

- Le *et al* (1997) *FEBS Lett.* 418(1-2):195-199; WO2004/047749; WO2003/072035 (claim 10); Touchman *et al* (2000) *Genome Res.* 10:165-173; WO2002/22660 (claim 20);
- 30 WO2003/093444 (claim 1); WO2003/087768 (claim 1); WO2003/029277 (page 82)

(32) CD72 (B-cell differentiation antigen CD72, Lyb-2); 359 aa, pI: 8.66, MW: 40225, TM: 1 5 [P] Gene Chromosome: 9p13.3).

Nucleotide

- 35 Genbank accession no NM_001782

Genbank version no. NM_001782.2 GI:194018444

Genbank record update date: Jun 26, 2012 01:43 PM

Polypeptide

Genbank accession no. NP_001773

5 Genbank version no. NP_001773.1 GI:4502683

Genbank record update date: Jun 26, 2012 01:43 PM

Cross references

10 WO2004042346 (claim 65); WO2003/026493 (pages 51-52, 57-58); WO2000/75655 (pages 105-106); Von Hoegen *et al* (1990) *J. Immunol.* 144(12):4870-4877; Strausberg *et al* (2002) *Proc. Natl. Acad. Sci USA* 99:16899-16903.

15 **(33)** *LY64 (Lymphocyte antigen 64 (RP105), type I membrane protein of the leucine rich repeat (LRR) family, regulates B-cell activation and apoptosis, loss of function is associated with increased disease activity in patients with systemic lupus erythematosus); 661 aa, pI: 6.20, MW: 74147 TM: 1 [P] Gene Chromosome: 5q12).*

Nucleotide

Genbank accession no NM_005582

Genbank version no. NM_005582.2 GI:167555126

20 Genbank record update date: Sep 2, 2012 01:50 PM

Polypeptide

Genbank accession no. NP_005573

Genbank version no. NP_005573.2 GI:167555127

25 Genbank record update date: Sep 2, 2012 01:50 PM

Cross references

30 US2002/193567; WO97/07198 (claim 11, pages 39-42); Miura *et al* (1996) 15 *Genomics* 38(3):299-304; Miura *et al* (1998) *Blood* 92:2815-2822; WO2003/083047; WO97/44452 (claim 8, pages 57-61); WO2000/12130 (pages 24-26).

(34) *FcRH1 (Fc receptor-like protein 1, a putative receptor for the immunoglobulin Fc domain that contains C2 type Ig-like and ITAM domains, may have a role in B-lymphocyte 20 differentiation); 429 aa, pI: 5.28, MW: 46925 TM: 1 [P] Gene Chromosome: 1q21-1q22)*

Nucleotide

35 Genbank accession no NM_052938

Genbank version no. NM_052938.4 GI:226958543

Genbank record update date: Sep 2, 2012 01:43 PM

Polypeptide

5 Genbank accession no. NP_443170

Genbank version no. NP_443170.1 GI:16418419

Genbank record update date: Sep 2, 2012 01:43 PM

Cross references

10 WO2003/077836; WO2001/38490 (claim 6, Fig 18E-1-18-E-2); Davis *et al* (2001) *Proc. Natl. Acad. Sci USA* 98(17):9772-9777; WO2003/089624 (claim 8); EP1347046 (claim 1); WO2003/089624 (claim 7).

15 **(35)** *IRTA2 (Immunoglobulin superfamily receptor translocation associated 2, a putative immunoreceptor with possible roles in B cell development and lymphomagenesis; deregulation of the gene by translocation occurs in some B cell malignancies); 977 aa, pI: 6.88, MW: 106468, TM: 1 [P] Gene Chromosome: 1q21)*

Nucleotide

Genbank accession no AF343662

20 Genbank version no. AF343662.1 GI:13591709

Genbank record update date: Mar 11, 2010 01:16 AM

Polypeptide

Genbank accession no. AAK31325

25 Genbank version no. AAK31325.1 GI:13591710

Genbank record update date: Mar 11, 2010 01:16 AM

Cross references

30 AF343663, AF343664, AF343665, AF369794, AF397453, AK090423, AK090475, AL834187, AY358085; Mouse:AK089756, AY158090, AY506558; NP_112571.1; WO2003/024392 (claim 2, Fig 97); Nakayama *et al* (2000) *Biochem. Biophys. Res. Commun.* 277(1):124-127; WO2003/077836; WO2001/38490 (claim 3, Fig 18B-1-18B-2).

35 **(36)** *TENB2 (TMEFF2, tomoregulin, TPEF, HPP1, TR, putative transmembrane 35 proteoglycan, related to the EGF/heregulin family of growth factors and follistatin); 374 aa)*

Nucleotide

Genbank accession no AF179274

Genbank version no. AF179274.2 GI:12280939

5 Genbank record update date: Mar 11, 2010 01:05 AM

Polypeptide

Genbank accession no. AAD55776

Genbank version no. AAD55776.2 GI:12280940

10 Genbank record update date: Mar 11, 2010 01:05 AM

Cross references

NCBI Accession: AAD55776, AAF91397, AAG49451, NCBI RefSeq: NP_057276; NCBI

Gene: 23671; OMIM: 605734; SwissProt Q9UIK5; AY358907, CAF85723, CQ782436;

15 WO2004/074320; JP2004113151; WO2003/042661; WO2003/009814; EP1295944 (pages

69-70); WO2002/30268 (page 329); WO2001/90304; US2004/249130; US2004/022727;

WO2004/063355; US2004/197325; US2003/232350; 5 US2004/005563; US2003/124579;

Horie *et al* (2000) *Genomics* 67:146-152; Uchida *et al* (1999) *Biochem. Biophys. Res.*

Commun. 266:593-602; Liang *et al* (2000) *Cancer Res.* 60:4907-12; Glynne-Jones *et al*

20 (2001) *Int J Cancer.* Oct 15; 94(2):178-84.

(37) PSMA – FOLH1 (Folate hydrolase (prostate-specific membrane antigen) 1)

Nucleotide

25 Genbank accession no M99487

Genbank version no. M99487.1 GI:190663

Genbank record update date: Jun 23, 2010 08:48 AM

Polypeptide

30 Genbank accession no. AAA60209

Genbank version no. AAA60209.1 GI:190664

Genbank record update date: Jun 23, 2010 08:48 AM

Cross references

35 Israeli R.S., et al *Cancer Res.* 53 (2), 227-230 (1993)

Other information

Official Symbol: FOLH1

Other Aliases: GIG27, FGCP, FOLH, GCP2, GCPII, NAALAD1, NAALAdase, PSM, PSMA, mGCP

- 5 Other Designations: N-acetylated alpha-linked acidic dipeptidase 1; N-acetylated-alpha-linked acidic dipeptidase I; NAALADase I; cell growth-inhibiting gene 27 protein; folylpoly-gamma-glutamate carboxypeptidase; glutamate carboxylase II; glutamate carboxypeptidase 2; glutamate carboxypeptidase II; membrane glutamate carboxypeptidase; prostate specific membrane antigen variant F; pteroylpoly-gamma-glutamate carboxypeptidase

10

ANTIBODIES

US 7,666,425:

Antibodies produces by Hybridomas having the following ATCC references: ATCC accession No. HB-12101, ATCC accession No. HB-12109, ATCC accession No. HB-12127 and ATCC
15 accession No. HB-12126.

Proscan: a monoclonal antibody selected from the group consisting of 8H12, 3E11, 17G1, 29B4, 30C1 and 20F2 (US 7,811,564; Moffett S., et al *Hybridoma (Larchmt)*. 2007 Dec;26(6):363-72).

20

Cytogen: monoclonal antibodies 7E11-C5 (ATCC accession No. HB 10494) and 9H10-A4 (ATCC accession No. HB11430) – US 5,763,202

GlycoMimetics: NUH2 - ATCC accession No. HB 9762 (US 7,135,301)

25

Human Genome Science: HPRAJ70 - ATCC accession No. 97131 (US 6,824,993); Amino acid sequence encoded by the cDNA clone (HPRAJ70) deposited as American Type Culture Collection ("ATCC") Deposit No. 97131

30

Medarex: Anti-PSMA antibodies that lack fucosyl residues - US 7,875,278

Mouse anti-PSMA antibodies include the 3F5.4G6, 3D7.1.1, 4E10-1.14, 3E11, 4D8, 3E6, 3C9, 2C7, 1G3, 3C4, 3C6, 4D4, 1G9, 5C8B9, 3G6, 4C8B9, and monoclonal antibodies. Hybridomas secreting 3F5.4G6, 3D7.1.1, 4E10-1.14, 3E11, 4D8, 3E6, 3C9, 2C7, 1G3, 3C4,
35 3C6, 4D4, 1G9, 5C8B9, 3G6 or 4C8B9 have been publicly deposited and are described in U.S. Pat. No. 6,159,508. Relevant hybridomas have been publicly deposited and are

described in U.S. Pat. No. 6,107,090. Moreover, humanized anti-PSMA antibodies, including a humanized version of J591, are described in further detail in PCT Publication WO 02/098897.

- 5 Other mouse anti-human PSMA antibodies have been described in the art, such as mAb 107-1A4 (Wang, S. et al. (2001) Int. J. Cancer 92:871-876) and mAb 2C9 (Kato, K. et al. (2003) Int. J. Urol. 10:439-444).

10 Examples of human anti-PSMA monoclonal antibodies include the 4A3, 7F12, 8C12, 8A11, 16F9, 2A10, 2C6, 2F5 and 1C3 antibodies, isolated and structurally characterized as originally described in PCT Publications WO 01/09192 and WO 03/064606 and in U.S. Provisional Application Ser. No. 60/654,125, entitled "Human Monoclonal Antibodies to Prostate Specific Membrane Antigen (PSMA)", filed on Feb. 18, 2005. The V.sub.H amino acid sequences of 4A3, 7F12, 8C12, 8A11, 16F9, 2A10, 2C6, 2F5 and 1C3 are shown in
15 SEQ ID NOs: 1-9, respectively. The V.sub.L amino acid sequences of 4A3, 7F12, 8C12, 8A11, 16F9, 2A10, 2C6, 2F5 and 1C3 are shown in SEQ ID NOs: 10-18, respectively.

Other human anti-PSMA antibodies include the antibodies disclosed in PCT Publication WO 03/034903 and US Application No. 2004/0033229.

20

NW Biotherapeutics: A hybridoma cell line selected from the group consisting of 3F5.4G6 having ATCC accession number HB12060, 3D7-1.I. having ATCC accession number HB12309, 4E10-1.14 having ATCC accession number HB12310, 3E11 (ATCC HB12488), 4D8 (ATCC HB12487), 3E6 (ATCC HB12486), 3C9 (ATCC HB12484), 2C7 (ATCC
25 HB12490), 1G3 (ATCC HB12489), 3C4 (ATCC HB12494), 3C6 (ATCC HB12491), 4D4 (ATCC HB12493), 1G9 (ATCC HB12495), 5C8B9 (ATCC HB12492) and 3G6 (ATCC HB12485) – see US 6,150,508

30 PSMA Development Company / Progenics / Cytogen – Seattle Genetics: mAb 3.9, produced by the hybridoma deposited under ATCC Accession No. PTA-3258 or mAb 10.3, produced by the hybridoma deposited under ATCC Accession No. PTA-3347 - US 7,850,971

PSMA Development Company– Compositions of PSMA antibodies (US 20080286284, Table 1)

35 This application is a divisional of U.S. patent application Ser. No. 10/395,894, filed on Mar. 21, 2003 (US 7,850,971)

University Hospital Freiburg, Germany - mAbs 3/A12, 3/E7, and 3/F11 (Wolf P., et al *Prostate*. 2010 Apr 1;70(5):562-9).

5 **(38) SST (Somatostatin Receptor; note that there are 5 subtypes)**

(38.1) SSTR2 (Somatostatin receptor 2)

Nucleotide

Genbank accession no NM_001050

Genbank version no. NM_001050.2 GI:44890054

10 Genbank record update date: Aug 19, 2012 01:37 PM

Polypeptide

Genbank accession no. NP_001041

Genbank version no. NP_001041.1 GI:4557859

15 Genbank record update date: Aug 19, 2012 01:37 PM

Cross references

Yamada Y., et al Proc. Natl. Acad. Sci. U.S.A. 89 (1), 251-255 (1992); Susini C., et al Ann Oncol. 2006 Dec;17(12):1733-42

20

Other information

Official Symbol: SSTR2

Other Designations: SRIF-1; SS2R; somatostatin receptor type 2

25 **(38.2) SSTR5 (Somatostatin receptor 5)**

Nucleotide

Genbank accession no D16827

Genbank version no. D16827.1 GI:487683

Genbank record update date: Aug 1, 2006 12:45 PM

30

Polypeptide

Genbank accession no. BAA04107

Genbank version no. BAA04107.1 GI:487684

Genbank record update date: Aug 1, 2006 12:45 PM

35

Cross references

Yamada, Y., et al *Biochem. Biophys. Res. Commun.* 195 (2), 844-852 (1993)

Other information

5 Official Symbol: SSTR5

Other Aliases: SS-5-R

Other Designations: Somatostatin receptor subtype 5; somatostatin receptor type 5

(38.3) SSTR1

10 (38.4) SSTR3

(38.5) SSTR4

AvB6 – Both subunits (39+40)

15 ***(39) ITGAV (Integrin, alpha V;***

Nucleotide

Genbank accession no M14648 J02826 M18365

Genbank version no. M14648.1 GI:340306

Genbank record update date: Jun 23, 2010 08:56 AM

20

Polypeptide

Genbank accession no. AAA36808

Genbank version no. AAA36808.1 GI:340307

Genbank record update date: Jun 23, 2010 08:56 AM

25

Cross references

Suzuki S., et al *Proc. Natl. Acad. Sci. U.S.A.* 83 (22), 8614-8618 (1986)

Other information

30 Official Symbol: ITGAV

Other Aliases: CD51, MSK8, VNRA, VTNR

Other Designations: antigen identified by monoclonal antibody L230; integrin alpha-V;
integrin alphaVbeta3; integrin, alpha V (vitronectin receptor, alpha polypeptide, antigen
CD51); vitronectin receptor subunit alpha

35

(40) ITGB6 (Integrin, beta 6)Nucleotide

Genbank accession no NM_000888

Genbank version no. NM_000888.3 GI:9966771

5 Genbank record update date: Jun 27, 2012 12:46 AM

Polypeptide

Genbank accession no. NP_000879

Genbank version no. NP_000879.2 GI:9625002

10 Genbank record update date: Jun 27, 2012 12:46 AM

Cross referencesSheppard D.J., et al *Biol. Chem.* 265 (20), 11502-11507 (1990)15 Other information

Official Symbol: ITGB6

Other Designations: integrin beta-6

ANTIBODIES

20 Biogen: US 7,943,742 - Hybridoma clones 6.3G9 and 6.8G6 were deposited with the ATCC, accession numbers ATCC PTA-3649 and -3645, respectively.

Biogen: US7,465,449 - In some embodiments, the antibody comprises the same heavy and light chain polypeptide sequences as an antibody produced by hybridoma 6.1A8, 6.3G9,
25 6.8G6, 6.2B1, 6.2B10, 6.2A1, 6.2E5, 7.1G10, 7.7G5, or 7.1C5.

Centocor (J&J): US7,550,142; US7,163,681

For example in US 7,550,142 - an antibody having human heavy chain and human light chain variable regions comprising the amino acid sequences shown in SEQ ID
30 NO: 7 and SEQ ID NO: 8.

Seattle Genetics: 15H3 (Ryan MC., et al Cancer Res April 15, 2012; 72(8 Supplement): 4630)

(41) CEACAM5 (Carcinoembryonic antigen-related cell adhesion molecule 5)Nucleotide

Genbank accession no M17303

Genbank version no. M17303.1 GI:178676

5 Genbank record update date: Jun 23, 2010 08:47 AM

Polypeptide

Genbank accession no. AAB59513

Genbank version no. AAB59513.1 GI:178677

10 Genbank record update date: Jun 23, 2010 08:47 AM

Cross referencesBeauchemin N., et al *Mol. Cell. Biol.* 7 (9), 3221-3230 (1987)15 Other information

Official Symbol: CEACAM5

Other Aliases: CD66e, CEA

Other Designations: meconium antigen 100

20 ANTIBODIESAstraZeneca-MedImmune:US 20100330103; US20080057063;
US20020142359

- for example an antibody having complementarity determining regions (CDRs) with the following sequences: heavy chain; CDR1 - DNYMH, CDR2 - WIDPENGDT E YAPKFRG, CDR3 - LIYAGYLAMD Y; and light chain CDR1 - SASSSVTYMH, CDR2 - STSNLAS, CDR3 - QQRSTYPLT.
- Hybridoma 806.077 deposited as European Collection of Cell Cultures (ECACC) deposit no. 96022936.

30 Research Corporation Technologies, Inc.:US5,047,507

Bayer Corporation: US6,013,772

BioAlliance: US7,982,017; US7,674,605

35 • US 7,674,605

- an antibody comprising the heavy chain variable region sequence from the amino acid sequence of SEQ ID NO: 1, and the light chain variable region sequence from the amino acid sequence of SEQ ID NO:2.
- an antibody comprising the heavy chain variable region sequence from the amino acid sequence of SEQ ID NO:5, and the light chain variable region sequence from the amino acid sequence of SEQ ID NO:6.

Celltech Therapeutics Limited: US5,877,293

10 The Dow Chemical Company: US5,472,693; US6,417,337; US6,333,405

US5,472,693 – for example, ATCC No. CRL-11215

US6,417,337 – for example, ATCC CRL-12208

US6,333,405 – for example, ATCC CRL-12208

15 Immunomedics, Inc: US7,534,431; US7,230,084; US7,300,644; US6,730,300;

US20110189085

- an antibody having CDRs of the light chain variable region comprise: CDR1 comprises KASQDVGTSVA (SEQ ID NO: 20); CDR2 comprises WTSTRHT (SEQ ID NO: 21); and CDR3 comprises QQYSLYRS (SEQ ID NO: 22);

and the CDRs of the heavy chain variable region of said anti-CEA antibody comprise: CDR1 comprises TYWMS (SEQ ID NO: 23); CDR2 comprises EIHPDSSTINYAPSLKD (SEQ ID NO: 24); and CDR3 comprises LYFGFPWFAY (SEQ ID NO: 25).

25 US20100221175; US20090092598; US20070202044; US20110064653;

US20090185974; US20080069775.

(42) MET (met proto-oncogene; hepatocyte growth factor receptor)

Nucleotide

30 Genbank accession no M35073

Genbank version no. M35073.1 GI:187553

Genbank record update date: Mar 6, 2012 11:12 AM

Polypeptide

35 Genbank accession no. AAA59589

Genbank version no. AAA59589.1 GI:553531

Genbank record update date: Mar 6, 2012 11:12 AM

Cross references

Dean M., et al *Nature* 318 (6044), 385-388 (1985)

5

Other information

Official Symbol: MET

Other Aliases: AUTS9, HGFR, RCCP2, c-Met

10 Other Designations: HGF receptor; HGF/SF receptor; SF receptor; hepatocyte growth factor receptor; met proto-oncogene tyrosine kinase; proto-oncogene c-Met; scatter factor receptor; tyrosine-protein kinase Met

ANTIBODIES

Abgenix/Pfizer: US20100040629

15 for example, the antibody produced by hybridoma 13.3.2 having American Type Culture Collection (ATCC) accession number PTA-5026; the antibody produced by hybridoma 9.1.2 having ATCC accession number PTA-5027; the antibody produced by hybridoma 8.70.2 having ATCC accession number PTA-5028; or the antibody produced by hybridoma 6.90.3 having ATCC accession number PTA-5029.

20

Amgen/Pfizer: US20050054019

25 for example, an antibody comprising a heavy chain having the amino acid sequences set forth in SEQ ID NO: 2 where X2 is glutamate and X4 is serine and a light chain having the amino acid sequence set forth in SEQ ID NO: 4 where X8 is alanine, without the signal sequences; an antibody comprising a heavy chain having the amino acid sequences set forth in SEQ ID NO: 6 and a light chain having the amino acid sequence set forth in SEQ ID NO: 8, without the signal sequences; an antibody comprising a heavy chain having the amino acid sequences set forth in SEQ ID NO: 10 and a light chain having the amino acid sequence set forth in SEQ ID NO: 12, without the signal sequences; or an antibody comprising a heavy chain having the amino acid sequences set forth in SEQ ID NO: 14 and a light chain having the amino acid sequence set forth in SEQ ID NO: 16, without the signal sequences.

30

Agouron Pharmaceuticals (Now Pfizer): US20060035907

35

Eli Lilly: US20100129369

Genentech: US5,686,292; US20100028337; US20100016241; US20070129301;
US20070098707; US20070092520, US20060270594; US20060134104; US20060035278;
US20050233960; US20050037431

- 5 US 5,686,292 – for example, ATCC HB-11894 and ATCC HB-11895
 US 20100016241 – for example, ATCC HB-11894 (hybridoma 1A3.3.13) or HB-
 11895 (hybridoma 5D5.11.6)

- National Defense Medical Center, Taiwan: Lu RM., et al *Biomaterials*. 2011
10 Apr;32(12):3265-74.

Novartis: US20090175860

- for example, an antibody comprising the sequences of CDR1, CDR2 and
 CDR3 of heavy chain 4687, wherein the sequences of CDR1, CDR2, and
15 CDR3 of heavy chain 4687 are residues 26-35, 50-65, and 98-102,
 respectively, of SEQ ID NO: 58; and the sequences of CDR1, CDR2, and
 CDR3 of light chain 5097, wherein the sequences of CDR1, CDR2, and
 CDR3 of light chain 5097 are residues 24-39, 55-61, and 94-100 of SEQ ID
 NO: 37.

20

Pharmacia Corporation: US20040166544

Pierre Fabre: US20110239316, US20110097262, US20100115639

- 25 Sumsung: US 20110129481 – for example a monoclonal antibody produced from a
 hybridoma cell having accession number KCLRF-BP-00219 or accession number of KCLRF-
 BP-00223.

- Samsung: US 20110104176 – for example an antibody produced by a hybridoma cell having
30 Accession Number: KCLRF-BP-00220.

University of Turin Medical School: DN-30 Pacchiana G., et al *J Biol Chem*. 2010 Nov
12;285(46):36149-57

- 35 Van Andel Research Institute: Jiao Y., et al *Mol Biotechnol*. 2005 Sep;31(1):41-54.

(43) MUC1 (*Mucin 1, cell surface associated*)

Nucleotide

Genbank accession no J05581

Genbank version no. J05581.1 GI:188869

5 Genbank record update date: Jun 23, 2010 08:48 AM

Polypeptide

Genbank accession no. AAA59876

Genbank version no. AAA59876.1 GI:188870

10 Genbank record update date: Jun 23, 2010 08:48 AM

Cross references

Gendler S.J., et al *J. Biol. Chem.* 265 (25), 15286-15293 (1990)

15 Other information

Official Symbol: MUC1

Other Aliases: RP11-263K19.2, CD227, EMA, H23AG, KL-6, MAM6, MUC-1, MUC-1/SEC, MUC-1/X, MUC1/ZD, PEM, PEMT, PUM

Other Designations: DF3 antigen; H23 antigen; breast carcinoma-associated antigen DF3;

20 carcinoma-associated mucin; episialin; krebs von den Lungen-6; mucin 1, transmembrane; mucin-1; peanut-reactive urinary mucin; polymorphic epithelial mucin; tumor associated epithelial mucin; tumor-associated epithelial membrane antigen; tumor-associated mucin

ANTIBODIES

25 AltaRex- Quest Pharma Tech: US 6,716,966 – for example an Alt-1 antibody produced by the hybridoma ATCC No PTA-975.

AltaRex- Quest Pharma Tech: US7,147,850

30 CRT: 5E5 - Sørensen AL., et al *Glycobiology* vol. 16 no. 2 pp. 96–107, 2006; HMFG2 – Burchell J., et al *Cancer Res.*, 47, 5476–5482 (1987)

Glycotope GT-MAB: GT-MAB 2.5-GEX

35 Immunogen: US7,202,346

- for example, antibody MJ-170: hybridoma cell line MJ-170 ATCC accession no. PTA-5286; monoclonal antibody MJ-171: hybridoma cell line MJ-171 ATCC accession no. PTA-5287; monoclonal antibody MJ-172: hybridoma cell line MJ-172 ATCC accession no. PTA-5288; or monoclonal antibody MJ-173: hybridoma cell line MJ-173 ATCC accession no. PTA-5302

Immunomedics: US 6,653,104

10 Ramot Tel Aviv Uni: US7,897,351

Regents Uni. CA: US 7,183,388; US20040005647; US20030077676.

Roche GlycArt: US8,021,856

15

Russian National Cancer Research Center: Imuteran- Ivanov PK., et al *Biotechnol J.* 2007 Jul;2(7):863-70

Technische Univ Braunschweig: (IIB6, HT186-B7, HT186-D11, HT186-G2, HT200-3A-C1, HT220-M-D1, HT220-M-G8) - Thie H., et al *PLoS One.* 2011 Jan 14;6(1):e15921

(44) CA9 (Carbonic anhydrase IX)

Nucleotide

Genbank accession no . X66839

25 Genbank version no. X66839.1 GI:1000701

Genbank record update date: Feb 2, 2011 10:15 AM

Polypeptide

Genbank accession no. CAA47315

30 Genbank version no. CAA47315.1 GI:1000702

Genbank record update date: Feb 2, 2011 10:15 AM

Cross references

Pastorek J., et al *Oncogene* 9 (10), 2877-2888 (1994)

35

Other information

Official Symbol: CA9

Other Aliases: CAIX, MN

Other Designations: CA-IX; P54/58N; RCC-associated antigen G250; RCC-associated
 5 protein G250; carbonate dehydratase IX; carbonic anhydrase 9; carbonic dehydratase;
 membrane antigen MN; pMW1; renal cell carcinoma-associated antigen G250

ANTIBODIES

Abgenix/Amgen: US20040018198

10

Affibody: Anti-CAIX Affibody molecules

Bayer: US7,462,696

15 Bayer/Morphosys: 3ee9 mAb - Petrul HM., et al *Mol Cancer Ther.* 2012 Feb;11(2):340-9

Harvard Medical School: Antibodies G10, G36, G37, G39, G45, G57, G106, G119, G6, G27,
 G40 and G125. Xu C., et al *PLoS One.* 2010 Mar 10;5(3):e9625

20 Institute of Virology, Slovak Academy of Sciences (Bayer) - US5,955,075

- for example, M75- ATCC Accession No. HB 11128 or MN12 – ATCC
 Accession No. HB 11647

Institute of Virology, Slovak Academy of Sciences: US7,816,493

25

- for example the M75 monoclonal antibody that is secreted from the
 hybridoma VU-M75, which was deposited at the American Type Culture
 Collection under ATCC No. HB 11128; or the V/10 monoclonal antibody
 secreted from the hybridoma V/10-VU, which was deposited at the
 International Depository Authority of the Belgian Coordinated Collection of
 30 Microorganisms (BCCM) at the Laboratorium voor Moleculaire Biologie-
 Plasmidencollectie (LMBP) at the Universeit Gent in Gent, Belgium, under
 Accession No. LMBP 6009CB.

Institute of Virology, Slovak Academy of Sciences US20080177046; US20080176310;
 35 US20080176258; US20050031623

Novartis: US20090252738

Wilex: US7,691,375 – for example the antibody produced by the hybridoma cell line DSM
5 ASC 2526.

Wilex: US20110123537; Rencarex: Kennett RH., et al *Curr Opin Mol Ther.* 2003
Feb;5(1):70-5

10 Xencor: US20090162382

(45) EGFRvIII (Epidermal growth factor receptor (EGFR), transcript variant 3,
Nucleotide

Genbank accession no. NM_201283

15 Genbank version no. NM_201283.1 GI:41327733

Genbank record update date: Sep 30, 2012 01:47 PM

Polypeptide

Genbank accession no. NP_958440

20 Genbank version no. NP_958440.1 GI:41327734

Genbank record update date: Sep 30, 2012 01:47 PM

Cross-references

Batra SK., et al *Cell Growth Differ* 1995;6:1251–1259.

25

ANTIBODIES:

US7,628,986 and US7,736,644 (Amgen)

For example, a heavy chain variable region amino acid sequence selected from the
group consisting of SEQ ID NO: 142 and variants & a light chain variable region
30 amino acid sequence selected from the group consisting of: SEQ ID NO: 144 and
variants.

US20100111979 (Amgen)

For example, an antibody comprising a heavy chain amino acid sequence
35 comprising:

CDR1 consisting of a sequence selected from the group consisting of the amino acid sequences for the CDR1 region of antibodies 13.1.2 (SEQ ID NO: 138), 131 (SEQ ID NO: 2), 170 (SEQ ID NO: 4), 150 (SEQ ID NO: 5), 095 (SEQ ID NO: 7), 250 (SEQ ID NO: 9), 139 (SEQ ID NO: 10), 211 (SEQ ID NO: 12), 124 (SEQ ID NO: 13), 318

5 (SEQ ID NO: 15), 342 (SEQ ID NO: 16), and 333 (SEQ ID NO: 17);

CDR2 consisting of a sequence selected from the group consisting of the amino acid sequences for the CDR2 region of antibodies 13.1.2 (SEQ ID NO: 138), 131 (SEQ ID NO: 2), 170 (SEQ ID NO: 4), 150 (SEQ ID NO: 5), 095 (SEQ ID NO: 7), 250 (SEQ ID NO: 9), 139 (SEQ ID NO: 10), 211 (SEQ ID NO: 12), 124 (SEQ ID NO: 13), 318

10 (SEQ ID NO: 15), 342 (SEQ ID NO: 16), and 333 (SEQ ID NO: 17); and

CDR3 consisting of a sequence selected from the group consisting of the amino acid sequences for the CDR3 region of antibodies 13.1.2 (SEQ ID NO: 138), 131 (SEQ ID NO: 2), 170 (SEQ ID NO: 4), 150 (SEQ ID NO: 5), 095 (SEQ ID NO: 7), 250 (SEQ ID NO: 9), 139 (SEQ ID NO: 10), 211 (SEQ ID NO: 12), 124 (SEQ ID NO: 13), 318

15 (SEQ ID NO: 15), 342 (SEQ ID NO: 16), and 333 (SEQ ID NO: 17).

US20090240038 (Amgen)

For example, an antibody having at least one of the heavy or light chain polypeptides comprises an amino acid sequence that is at least 90% identical to the amino acid sequence selected from the group consisting of: SEQ ID NO: 2, SEQ ID NO: 19, SEQ ID NO: 142, SEQ ID NO: 144, and any combination thereof.

20

US20090175887 (Amgen)

For example, an antibody having a heavy chain amino acid sequence selected from the group consisting of the heavy chain amino acid sequence of antibody 13.1.2 (SEQ ID NO: 138), 131 (SEQ ID NO: 2), 170 (SEQ ID NO: 4), 150 (SEQ ID NO: 5), 095 (SEQ ID NO: 7), 250 (SEQ ID NO: 9), 139 (SEQ ID NO: 10), 211 (SEQ ID NO: 12), 124 (SEQ ID NO: 13), 318 (SEQ ID NO: 15), 342 (SEQ ID NO: 16), and 333 (SEQ ID NO: 17).

25

30

US20090156790 (Amgen)

For example, antibody having heavy chain polypeptide and a light chain polypeptide, wherein at least one of the heavy or light chain polypeptides comprises an amino acid sequence that is at least 90% identical to the amino acid sequence selected from the group consisting of: SEQ ID NO: 2, SEQ ID NO: 19, SEQ ID NO: 142, SEQ ID NO: 144, and any combination thereof.

35

US20090155282, US20050059087 and US20050053608 (Amgen)

For example, an antibody heavy chain amino acid sequence selected from the group consisting of the heavy chain amino acid sequence of antibody 13.1.2 (SEQ ID NO: 138), 131 (SEQ ID NO: 2), 170 (SEQ ID NO: 4), 150 (SEQ ID NO: 5), 095 (SEQ ID NO: 7), 250 (SEQ ID NO: 9), 139 (SEQ ID NO: 10), 211 (SEQ ID NO: 12), 124 (SEQ ID NO: 13), 318 (SEQ ID NO: 15), 342 (SEQ ID NO: 16), and 333 (SEQ ID NO: 17).

MR1-1 (US7,129,332; Duke)

For example, a variant antibody having the sequence of SEQ ID NO.18 with the substitutions S98P-T99Y in the CDR3 VH, and F92W in CDR3 VL.

L8A4, H10, Y10 (Wikstrand CJ., et al *Cancer Res.* 1995 Jul 15;55(14):3140-8; Duke)

US20090311803 (Harvard University)

For example, SEQ ID NO:9 for antibody heavy chain variable region, and SEQ ID NO: 3 for light chain variable region amino acid sequences

US20070274991 (EMD72000, also known as matuzumab; Harvard University)

For example, SEQ ID NOs: 3 & 9 for light chain and heavy chain respectively

US6,129,915 (Schering)

For example, SEQ. ID NOs: 1, 2, 3, 4, 5 and 6.

mAb CH12 - Wang H., et al *FASEB J.* 2012 Jan;26(1):73-80 (Shanghai Cancer Institute).

RAbDMvIII - Gupta P., et al *BMC Biotechnol.* 2010 Oct 7;10:72 (Stanford University Medical Center).

mAb Ua30 - Ohman L., et al *Tumour Biol.* 2002 Mar-Apr;23(2):61-9 (Uppsala University).

Han DG., et al *Nan Fang Yi Ke Da Xue Xue Bao.* 2010 Jan;30(1):25-9 (Xi'an Jiaotong University).

(46) CD33 (CD33 molecule)Nucleotide

Genbank accession no. M_23197

Genbank version no. NM_23197.1 GI:180097

5 Genbank record update date: Jun 23, 2010 08:47 AM

Polypeptide

Genbank accession no. AAA51948

Genbank version no. AAA51948.1 GI:188098

10 Genbank record update date: Jun 23, 2010 08:47 AM

Cross-referencesSimmons D., et al *J. Immunol.* 141 (8), 2797-2800 (1988)15 Other information

Official Symbol: CD33

Other Aliases: SIGLEC-3, SIGLEC3, p67

Other Designations: CD33 antigen (gp67); gp67; myeloid cell surface antigen CD33; sialic acid binding Ig-like lectin 3; sialic acid-binding Ig-like lectin

20

ANTIBODIESH195 (Lintuzumab)- Raza A., et al *Leuk Lymphoma*. 2009 Aug;50(8):1336-44; US6,759,045 (Seattle Genetics/Immunomedics)25 mAb OKT9: Sutherland, D.R. et al. *Proc Natl Acad Sci USA* 78(7): 4515-4519 1981, Schneider, C., et al *J Biol Chem* 257, 8516-8522 (1982)mAb E6: Hoogenboom, H.R., et al *J Immunol* 144, 3211-3217 (1990)

30 US6,590,088 (Human Genome Sciences)

For example, SEQ ID NOs: 1 and 2 and ATCC accession no. 97521

US7,557,189 (Immunogen)

35 For example, an antibody or fragment thereof comprising a heavy chain variable region which comprises three CDRs having the amino acid sequences of SEQ ID

NOs:1-3 and a light chain variable region comprising three CDRs having the amino acid sequences of SEQ ID NOs:4-6.

(47) CD19 (CD19 molecule)

5 Nucleotide

Genbank accession no. NM_001178098

Genbank version no. NM_001178098.1 GI:296010920

Genbank record update date: Sep 10, 2012 12:43 AM

10 Polypeptide

Genbank accession no. NP_001171569

Genbank version no. NP_001171569.1 GI:296010921

Genbank record update date: Sep 10, 2012 12:43 AM

15 Cross-references

Tedder TF., et al J. Immunol. 143 (2): 712-7 (1989)

Other information

Official Symbol: CD19

20 Other Aliases: B4, CVID3

Other Designations: B-lymphocyte antigen CD19; B-lymphocyte surface antigen B4; T-cell surface antigen Leu-12; differentiation antigen CD19

ANTIBODIES

25 Immunogen: HuB4 - Al-Katib AM., et al *Clin Cancer Res.* 2009 Jun 15;15(12):4038-45.

4G7: Kügler M., et al *Protein Eng Des Sel.* 2009 Mar;22(3):135-47

For example, sequences in Fig. 3 of Knappik, A. et al. J Mol Biol 2000 Feb;296(1):57-86

30

AstraZeneca /MedImmune: MEDI-551 - Herbst R., et al *J Pharmacol Exp Ther.* 2010 Oct;335(1):213-22

Glenmark Pharmaceuticals: GBR-401 - Hou S., et al *Mol Cancer Ther* November 2011 10

35 (Meeting Abstract Supplement) C164

US7,109,304 (Immunomedics)

For example, an antibody comprising the sequence of hA19Vk (SEQ ID NO:7) and the sequence of hA19VH (SEQ ID NO:10)

5 US7,902,338 (Immunomedics)

For example, an antibody or antigen-binding fragment thereof that comprises the light chain complementarity determining region CDR sequences CDR1 of SEQ ID NO: 16 (KASQSVDDYDGDSYLN); CDR2 of SEQ ID NO: 17 (DASNLVS); and CDR3 of SEQ ID NO: 18 (QQSTEDPWT) and the heavy chain CDR sequences CDR1 of SEQ ID NO: 19 (SYWMN); CDR2 of SEQ ID NO: 20 (QIWPGDGDNTNYNGKFKG) and CDR3 of SEQ ID NO: 21 (RETTTVGRYYYAMDY) and also comprises human antibody framework (FR) and constant region sequences with one or more framework region amino acid residues substituted from the corresponding framework region sequences of the parent murine antibody, and wherein said substituted FR residues comprise the substitution of serine for phenylalanine at Kabat residue 91 of the heavy chain variable region.

Medarex: MDX-1342 – Cardarelli PM., et al *Cancer Immunol Immunother.* 2010 Feb;59(2):257-65.

20

MorphoSys /Xencor: MOR-208/XmAb-5574 - Zalevsky J., et al *Blood.* 2009 Apr 16;113(16):3735-43

US7,968,687 (Seattle Genetics)

25 An antibody or antigen-binding fragment comprising a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:9 and a light chain variable domain comprising the amino acid sequence of SEQ ID NO: 24.

4G7 chim - Lang P., et al *Blood.* 2004 May 15;103(10):3982-5 (University of Tübingen)

30

For example, fig. 6 and SEQ ID No: 80 of US20120082664

Zhejiang University School of Medicine: 2E8 - Zhang J., et al *J Drug Target.* 2010 Nov;18(9):675-8

35

(48) *IL2RA* (Interleukin 2 receptor, alpha); NCBI Reference Sequence: NM_000417.2);

Nucleotide

Genbank accession no. NM_000417

Genbank version no. NM_000417.2 GI:269973860

5 Genbank record update date: Sep 09, 2012 04:59 PM

Polypeptide

Genbank accession no. NP_000408

Genbank version no. NP_000408.1 GI:4557667

10 Genbank record update date: Sep 09, 2012 04:59 PM

Cross-references

Kuziel W.A., et al *J. Invest. Dermatol.* 94 (6 SUPPL), 27S-32S (1990)

15 Other information

Official Symbol: IL2RA

Other Aliases: RP11-536K7.1, CD25, IDDM10, IL2R, TCGFR

Other Designations: FIL-2 receptor subunit alpha; IL-2-RA; IL-2R subunit alpha; IL2-RA; TAC antigen; interleukin-2 receptor subunit alpha; p55

20

ANTIBODIES

US6,383,487 (Novartis/UCL: Baxilisimab [Simulect™])

US6,521,230 (Novartis/UCL: Baxilisimab [Simulect])

25 For example, an antibody having an antigen binding site comprises at least one domain which comprises CDR1 having the amino acid sequence in SEQ. ID. NO: 7, CDR2 having the amino acid sequence in SEQ. ID. NO: 8, and CDR3 having the amino acid sequence in SEQ. ID. NO: 9; or said CDR1, CDR2 and CDR3 taken in sequence as a whole comprise an amino acid sequence which is at least 90% identical to SEQ. ID. NOs: 7, 8 and 9 taken in sequence as a whole.

30

Daclizumab – Rech A.J., et al *Ann N Y Acad Sci.* 2009 Sep;1174:99-106 (Roche)

(49) *AXL* (AXL receptor tyrosine kinase)

35 Nucleotide

Genbank accession no. M76125

Genbank version no. M76125.1 GI:292869

Genbank record update date: Jun 23, 2010 08:53 AM

Polypeptide

5 Genbank accession no. AAA61243

Genbank version no. AAA61243.1 GI:29870

Genbank record update date: Jun 23, 2010 08:53 AM

Cross-references

10 O'Bryan J.P., et al *Mol. Cell. Biol.* 11 (10), 5016-5031 (1991); Bergsagel P.L., et al *J. Immunol.* 148 (2), 590-596 (1992)

Other information

Official Symbol: AXL

15 Other Aliases: JTK11, UFO

Other Designations: AXL oncogene; AXL transforming sequence/gene; oncogene AXL; tyrosine-protein kinase receptor UFO

ANTIBODIES

20 YW327.6S2 - Ye X., et al *Oncogene*. 2010 Sep 23;29(38):5254-64. (Genentech)

BergenBio: BGB324

(50) *CD30 - TNFRSF8 (Tumor necrosis factor receptor superfamily, member 8)*

25 Nucleotide

Genbank accession no. M83554

Genbank version no. M83554.1 GI:180095

Genbank record update date: Jun 23, 2010 08:53 AM

30 Polypeptide

Genbank accession no. AAA51947

Genbank version no. AAA51947.1 GI:180096

Genbank record update date: Jun 23, 2010 08:53 AM

35 Cross-references

Durkop H., et al *Cell* 68 (3), 421-427 (1992)

Other information

Official Symbol: TNFRSF8

Other Aliases: CD30, D1S166E, Ki-1

- 5 Other Designations: CD30L receptor; Ki-1 antigen; cytokine receptor CD30; lymphocyte activation antigen CD30; tumor necrosis factor receptor superfamily member 8

(51) BCMA (B-cell maturation antigen) - TNFRSF17 (Tumor necrosis factor receptor superfamily, member 17)

10 Nucleotide

Genbank accession no. Z29574

Genbank version no. Z29574.1 GI:471244

Genbank record update date: Feb 02, 2011 10:40 AM

15 Polypeptide

Genbank accession no. CAA82690

Genbank version no. CAA82690.1 GI:471245

Genbank record update date: Feb 02, 2011 10:40 AM

20 Cross-references

Laabi Y., et al Nucleic Acids Res. 22 (7), 1147-1154 (1994)

Other information

Official Symbol: TNFRSF17

- 25 Other Aliases: BCM, BCMA, CD269

Other Designations: B cell maturation antigen; B-cell maturation factor; B-cell maturation protein; tumor necrosis factor receptor superfamily member 17

- 30 *(52) CT Ags – CTA (Cancer Testis Antigens)*

Cross-references

Fratta E., et al. *Mol Oncol.* 2011 Apr;5(2):164-82; Lim SH., et al *Am J Blood Res.* 2012;2(1):29-35.

(53) CD174 (Lewis Y) - FUT3 (fucosyltransferase 3 (galactoside 3(4)-L-fucosyltransferase, Lewis blood group)

Nucleotide

Genbank accession no. NM000149

5 Genbank version no. NM000149.3 GI:148277008

Genbank record update date: Jun 26, 2012 04:49 PM

Polypeptide

Genbank accession no. NP_000140

10 Genbank version no. NP_000140.1 GI:4503809

Genbank record update date: Jun 26, 2012 04:49 PM

Cross-references

Kukowska-Latallo, J.F., et al *Genes Dev.* 4 (8), 1288-1303 (1990)

15

Other information

Official Symbol: FUT3

Other Aliases: CD174, FT3B, FucT-III, LE, Les

Other Designations: Lewis FT; alpha-(1,3/1,4)-fucosyltransferase; blood group Lewis alpha-

20 4-fucosyltransferase; fucosyltransferase III; galactoside 3(4)-L-fucosyltransferase

(54) CLEC14A (C-type lectin domain family 14, member A; Genbank accession no. NM175060)

Nucleotide

25 Genbank accession no. NM175060

Genbank version no. NM175060.2 GI:371123930

Genbank record update date: Apr 01, 2012 03:34 PM

Polypeptide

30 Genbank accession no. NP_778230

Genbank version no. NP_778230.1 GI:28269707

Genbank record update date: Apr 01, 2012 03:34 PM

Other information

35 Official Symbol: CLEC14A

Other Aliases: UNQ236/PRO269, C14orf27, CEG1, EGFR-5

Other Designations: C-type lectin domain family 14 member A; CLECT and EGF-like domain containing protein; epidermal growth factor receptor 5

(55) *GRP78 – HSPA5 (heat shock 70kDa protein 5 (glucose-regulated protein, 78kDa)*

5 Nucleotide

Genbank accession no. NM005347

Genbank version no. NM005347.4 GI:305855105

Genbank record update date: Sep 30, 2012 01:42 PM

10 Polypeptide

Genbank accession no. NP_005338

Genbank version no. NP_005338.1 GI:16507237

Genbank record update date: Sep 30, 2012 01:42 PM

15 Cross-references

Ting J., et al *DNA* 7 (4), 275-286 (1988)

Other information

Official Symbol: HSPA5

20 Other Aliases: BIP, GRP78, MIF2

Other Designations: 78 kDa glucose-regulated protein; endoplasmic reticulum luminal Ca(2+)-binding protein grp78; immunoglobulin heavy chain-binding protein

(56) *CD70 (CD70 molecule) L08096*

25 Nucleotide

Genbank accession no. L08096

Genbank version no. L08096.1 GI:307127

Genbank record update date: Jun 23, 2012 08:54 AM

30 Polypeptide

Genbank accession no. AAA36175

Genbank version no. AAA36175.1 GI:307128

Genbank record update date: Jun 23, 2012 08:54 AM

35 Cross-references

Goodwin R.G., et al *Cell* 73 (3), 447-456 (1993)

Other information

Official Symbol: CD70

Other Aliases: CD27L, CD27LG, TNFSF7

- 5 Other Designations: CD27 ligand; CD27-L; CD70 antigen; Ki-24 antigen; surface antigen CD70; tumor necrosis factor (ligand) superfamily, member 7; tumor necrosis factor ligand superfamily member 7

ANTIBODIES

- 10 MDX-1411 against CD70 (Medarex)

h1F6 (Ofiazoglu, E., et al, Clin Cancer Res. 2008 Oct 1;14(19):6171-80; Seattle Genetics)

For example, see US20060083736 SEQ ID NOs: 1, 2, 11 and 12 and Fig. 1.

- 15 *(57) Stem Cell specific antigens. For example:*

- 5T4 (see entry (63) below)
- CD25 (see entry (48) above)
- CD32

- Polypeptide

- 20
- Genbank accession no. ABK42161
 - Genbank version no. ABK42161.1 GI:117616286
 - Genbank record update date: Jul 25, 2007 03:00 PM

- LGR5/GPR49

- Nucleotide

- 25
- Genbank accession no. NM_003667
 - Genbank version no. NM_003667.2 GI:24475886
 - Genbank record update date: Jul 22, 2012 03:38 PM

- Polypeptide

- 30
- Genbank accession no. NP_003658
 - Genbank version no. NP_003658.1 GI:4504379
 - Genbank record update date: Jul 22, 2012 03:38 PM

- Prominin/CD133

- Nucleotide

- 35
- Genbank accession no. NM_006017
 - Genbank version no. NM_006017.2 GI:224994187

- Genbank record update date: Sep 30, 2012 01:47 PM
- Polypeptide
 - Genbank accession no. NP_006008
 - Genbank version no. NP_006008.1 GI:5174387
 - Genbank record update date: Sep 30, 2012 01:47 PM

(58) ASG-5Cross-references

(Smith L.M., et.al *AACR 2010 Annual Meeting* (abstract #2590); Gudas J.M., et.al. *AACR 2010 Annual Meeting* (abstract #4393))

ANTIBODIES

Anti- AGS-5 Antibody: M6.131 (Smith, L.M., et.al *AACR 2010 Annual Meeting* (abstract #2590))

(59) ENPP3 (Ectonucleotide pyrophosphatase/phosphodiesterase 3)Nucleotide

Genbank accession no. AF005632

Genbank version no. AF005632.2 GI:4432589

Genbank record update date: Mar 10, 2010 09:41 PM

Polypeptide

Genbank accession no. AAC51813

Genbank version no. AAC51813.1 GI:2465540

Genbank record update date: Mar 10, 2010 09:41 PM

Cross-references

Jin-Hua P., et al *Genomics* 45 (2), 412-415 (1997)

Other information

Official Symbol: ENPP3

Other Aliases: RP5-988G15.3, B10, CD203c, NPP3, PD-IBETA, PDNP3

Other Designations: E-NPP 3; dJ1005H11.3 (phosphodiesterase I/nucleotide pyrophosphatase 3); dJ914N13.3 (phosphodiesterase I/nucleotide pyrophosphatase 3);

ectonucleotide pyrophosphatase/phosphodiesterase family member 3; gp130RB13-6;

phosphodiesterase I beta; phosphodiesterase I/nucleotide pyrophosphatase 3;
phosphodiesterase-I beta

(60) *PRR4 (Proline rich 4 (lacrima))*

5 Nucleotide

Genbank accession no. NM_007244

Genbank version no. NM_007244.2 GI:154448885

Genbank record update date: Jun 28, 2012 12:39 PM

10 Polypeptide

Genbank accession no. NP_009175

Genbank version no. NP_009175.2 GI:154448886

Genbank record update date: Jun 28, 2012 12:39 PM

15 Cross-references

Dickinson D.P., et al *Invest. Ophthalmol. Vis. Sci.* 36 (10), 2020-2031 (1995)

Other information

Official Symbol: PRR4

20 Other Aliases: LPRP, PROL4

Other Designations: lacrimal proline-rich protein; nasopharyngeal carcinoma-associated proline-rich protein 4; proline-rich polypeptide 4; proline-rich protein 4

(61) *GCC – GUCY2C (guanylate cyclase 2C (heat stable enterotoxin receptor))*

25 Nucleotide

Genbank accession no. NM_004963

Genbank version no. NM_004963.3 GI:222080082

Genbank record update date: Sep 02, 2012 01:50 PM

30 Polypeptide

Genbank accession no. NP_004954

Genbank version no. NP_004954.2 GI:222080083

Genbank record update date: Sep 02, 2012 01:50 PM

Cross-references

De Sauvage F.J., et al *J. Biol. Chem.* 266 (27), 17912-17918 (1991); Singh S., et al *Biochem. Biophys. Res. Commun.* 179 (3), 1455-1463 (1991)

5 Other information

Official Symbol: GUCY2C

Other Aliases: DIAR6, GUC2C, MUCIL, STAR

Other Designations: GC-C; STA receptor; guanylyl cyclase C; hSTAR; heat-stable enterotoxin receptor; intestinal guanylate cyclase

10

(62) *Liv-1 – SLC39A6 (Solute carrier family 39 (zinc transporter), member 6)*

Nucleotide

Genbank accession no. U41060

Genbank version no. U41060.2 GI:12711792

15 Genbank record update date: Nov 30, 2009 04:35 PM

Polypeptide

Genbank accession no. AAA96258

Genbank version no. AAA96258.2 GI:12711793

20 Genbank record update date: Nov 30, 2009 04:35 PM

Cross-references

Taylor KM., et al *Biochim Biophys Acta.* 2003 Apr 1;1611(1-2):16-30

25 Other information

Official Symbol: SLC39A6

Other Aliases: LIV-1

Other Designations: LIV-1 protein, estrogen regulated; ZIP-6; estrogen-regulated protein LIV-1; solute carrier family 39 (metal ion transporter), member 6; solute carrier family 39 member 6; zinc transporter ZIP6; zrt- and Irt-like protein 6

30

(63) *5T4, Trophoblast glycoprotein, TPBG – TPBG (trophoblast glycoprotein)*

Nucleotide

Genbank accession no. AJ012159

35 Genbank version no. AJ012159.1 GI:3805946

Genbank record update date: Feb 01, 2011 10:27 AM

Polypeptide

Genbank accession no. CAA09930

Genbank version no. CAA09930.1 GI:3805947

5 Genbank record update date: Feb 01, 2011 10:27 AM

Cross-references

King K.W., et al *Biochim. Biophys. Acta* 1445 (3), 257-270 (1999)

10 Other information

- Official Symbol: TPBG
- Other Aliases: 5T4, 5T4AG, M6P1
- Other Designations: 5T4 oncofetal antigen; 5T4 oncofetal trophoblast glycoprotein; 5T4 oncotrophoblast glycoprotein

15

(64) CD56 – NCMA1 (Neural cell adhesion molecule 1)

Nucleotide

Genbank accession no. NM_000615

Genbank version no. NM_000615.6 GI:336285433

20 Genbank record update date: Sep 23, 2012 02:32 PM

Polypeptide

Genbank accession no. NP_000606

Genbank version no. NP_000606.3 GI:94420689

25 Genbank record update date: Sep 23, 2012 02:32 PM

Cross-references

Dickson, G., et al, *Cell* 50 (7), 1119-1130 (1987)

30 Other information

Official Symbol: NCAM1

Other Aliases: CD56, MSK39, NCAM

Other Designations: antigen recognized by monoclonal antibody 5.1H11; neural cell adhesion molecule, NCAM

35

ANTIBODIES

Immunogen: HuN901 (Smith SV., et al *Curr Opin Mol Ther.* 2005 Aug;7(4):394-401)

For example, see humanized from murine N901 antibody. See Fig. 1b and 1e of Roguska, M.A., et al. *Proc Natl Acad Sci USA* Feb 1994;91:969-973.

5

(65) CanAg (Tumor associated antigen CA242)

Cross-references

Haglund C., et al *Br J Cancer* 60:845-851, 1989; Baeckstrom D., et al *J Biol Chem* 266:21537-21547, 1991

10

ANTIBODIES

huC242 (Tolcher AW et al., *J Clin Oncol.* 2003 Jan 15;21(2):211-22; Immunogen)

For example, see US20080138898A1 SEQ ID NO: 1 and 2

15

(66) FOLR1 (Folate Receptor 1)

Nucleotide

Genbank accession no. J05013

Genbank version no. J05013.1 GI:182417

20

Genbank record update date: Jun 23, 2010 08:47 AM

Polypeptide

Genbank accession no. AAA35823

Genbank version no. AAA35823.1 GI:182418

25

Genbank record update date: Jun 23, 2010 08:47 AM

Cross-references

Elwood P.C., et al *J. Biol. Chem.* 264 (25), 14893-14901 (1989)

30

Other information

Official Symbol: FOLR1

Other Aliases: FBP, FOLR

Other Designations: FR-alpha; KB cells FBP; adult folate-binding protein; folate binding protein; folate receptor alpha; folate receptor, adult; ovarian tumor-associated antigen

35

MOv18

ANTIBODIES

M9346A - Whiteman KR., et al *Cancer Res* April 15, 2012; 72(8 Supplement): 4628
(Immunogen)

5 **(67) GPNMB (Glycoprotein (transmembrane) nmb)**Nucleotide

Genbank accession no. X76534

Genbank version no. X76534.1 GI:666042

Genbank record update date: Feb 02, 2011 10:10 AM

10

Polypeptide

Genbank accession no. CAA54044

Genbank version no. CAA54044.1 GI:666043

Genbank record update date: Feb 02, 2011 10:10 AM

15

Cross-references

Weterman M.A., et al *Int. J. Cancer* 60 (1), 73-81 (1995)

Other information

20 Official Symbol: GPNMB

Other Aliases: UNQ1725/PRO9925, HGFIN, NMB

Other Designations: glycoprotein NMB; glycoprotein nmb-like protein; osteoactivin;
transmembrane glycoprotein HGFIN; transmembrane glycoprotein NMB

25 **ANTIBODIES**

Celldex Therapeutics: CR011 (Tse KF., et al *Clin Cancer Res.* 2006 Feb 15;12(4):1373-82)

For example, see EP1827492B1 SEQ ID NO: 22, 24, 26, 31, 33 and 35

(68) TIM-1 – HAVCR1 (Hepatitis A virus cellular receptor 1)30 Nucleotide

Genbank accession no. AF043724

Genbank version no. AF043724.1 GI:2827453

Genbank record update date: Mar 10, 2010 06:24 PM

35 Polypeptide

Genbank accession no. AAC39862

Genbank version no. AAC39862.1 GI:2827454

Genbank record update date: Mar 10, 2010 06:24 PM

Cross-references

- 5 Feigelsstock D., et al *J. Virol.* 72 (8), 6621-6628 (1998)

Other information

Official Symbol: HAVCR1

Other Aliases: HAVCR, HAVCR-1, KIM-1, KIM1, TIM, TIM-1, TIM1, TIMD-1, TIMD1

- 10 Other Designations: T cell immunoglobulin domain and mucin domain protein 1; T-cell membrane protein 1; kidney injury molecule 1

(69) *RG-1/Prostate tumor target Mindin – Mindin/RG-1*

Cross-references

- 15 Parry R., et al *Cancer Res.* 2005 Sep 15;65(18):8397-405

(70) *B7-H4 – VTCN1 (V-set domain containing T cell activation inhibitor 1*

Nucleotide

Genbank accession no. BX648021

- 20 Genbank version no. BX648021.1 GI:34367180
Genbank record update date: Feb 02, 2011 08:40 AM

Cross-references

Sica GL., et al *Immunity.* 2003 Jun;18(6):849-61

25

Other information

Official Symbol: VTCN1

Other Aliases: RP11-229A19.4, B7-H4, B7H4, B7S1, B7X, B7h.5, PRO1291, VCTN1

- 30 Other Designations: B7 family member, H4; B7 superfamily member 1; T cell costimulatory molecule B7x; T-cell costimulatory molecule B7x; V-set domain-containing T-cell activation inhibitor 1; immune costimulatory protein B7-H4

(71) *PTK7 (PTK7 protein tyrosine kinase 7)*

- 35 Nucleotide

Genbank accession no. AF447176

Genbank version no. AF447176.1 GI:17432420

Genbank record update date: Nov 28, 2008 01:51 PM

Polypeptide

5 Genbank accession no. AAL39062

Genbank version no. AAL39062.1 GI:17432421

Genbank record update date: Nov 28, 2008 01:51 PM

Cross-references

10 Park S.K., et al *J. Biochem.* 119 (2), 235-239 (1996)

Other information

Official Symbol: PTK7

Other Aliases: CCK-4, CCK4

15 Other Designations: colon carcinoma kinase 4; inactive tyrosine-protein kinase 7; pseudo tyrosine kinase receptor 7; tyrosine-protein kinase-like 7

(72) CD37 (*CD37 molecule*)

Nucleotide

20 Genbank accession no. NM_001040031

Genbank version no. NM_001040031.1 GI:91807109

Genbank record update date: Jul 29, 2012 02:08 PM

Polypeptide

25 Genbank accession no. NP_001035120

Genbank version no. NP_001035120.1 GI:91807110

Genbank record update date: Jul 29, 2012 02:08 PM

Cross-references

30 Schwartz-Albiez R., et al *J. Immunol.* 140 (3), 905-914 (1988)

Other information

Official Symbol: CD37

Other Aliases: GP52-40, TSPAN26

35 Other Designations: CD37 antigen; cell differentiation antigen 37; leukocyte antigen CD37; leukocyte surface antigen CD37; tetraspanin-26; tspan-26

ANTIBODIES

Boehringer Ingelheim: mAb 37.1 (Heider KH., et al *Blood*. 2011 Oct 13;118(15):4159-68)

- 5 Trubion: CD37-SMIP (G28-1 scFv-Ig) ((Zhao X., et al *Blood*. 2007;110: 2569-2577)

For example, see US20110171208A1 SEQ ID NO: 253

Immunogen: K7153A (Deckert J., et al *Cancer Res* April 15, 2012; 72(8 Supplement): 4625)

- 10 **(73)** *CD138 – SDC1 (syndecan 1)*

Nucleotide

Genbank accession no. AJ551176

Genbank version no. AJ551176.1 GI:29243141

Genbank record update date: Feb 01, 2011 12:09 PM

15

Polypeptide

Genbank accession no. CAD80245

Genbank version no. CAD80245.1 GI:29243142

Genbank record update date: Feb 01, 2011 12:09 PM

20

Cross-references

O'Connell FP., et al *Am J Clin Pathol*. 2004 Feb;121(2):254-63

Other information

- 25 Official Symbol: SDC1

Other Aliases: CD138, SDC, SYND1, syndecan

Other Designations: CD138 antigen; heparan sulfate proteoglycan fibroblast growth factor receptor; syndecan proteoglycan 1; syndecan-1

- 30 *ANTIBODIES*

Biotest: chimerized MAb (nBT062) - (Jagannath S., et al Poster *ASH* #3060, 2010; WIPO Patent Application WO/2010/128087)

For example, see US20090232810 SEQ ID NO: 1 and 2

- 35 Immunogen: B-B4 (Tassone P., et al *Blood* 104_3688-3696)

For example, see US20090175863A1 SEQ ID NO: 1 and 2

(74) CD74 (CD74 molecule, major histocompatibility complex, class II invariant chain)

Nucleotide

Genbank accession no. NM_004355

5 Genbank version no. NM_004355.1 GI:343403784

Genbank record update date: Sep 23, 2012 02:30 PM

Polypeptide

Genbank accession no. NP_004346

10 Genbank version no. NP_004346.1 GI:10835071

Genbank record update date: Sep 23, 2012 02:30 PM

Cross-references

Kudo, J., et al *Nucleic Acids Res.* 13 (24), 8827-8841 (1985)

15

Other information

Official Symbol: CD74

Other Aliases: DHLA, HLADG, II, Ia-GAMMA

20 Other Designations: CD74 antigen (invariant polypeptide of major histocompatibility complex, class II antigen-associated); HLA class II histocompatibility antigen gamma chain; HLA-DR antigens-associated invariant chain; HLA-DR-gamma; Ia-associated invariant chain; MHC HLA-DR gamma chain; gamma chain of class II antigens; p33

ANTIBODIES

25 Immunomedics: hLL1 (Milatuzumab,) - Berkova Z., et al *Expert Opin Investig Drugs.* 2010 Jan;19(1):141-9)

For example, see US20040115193 SEQ ID NOs: 19, 20, 21, 22, 23 and 24

Genmab: HuMax-CD74 (see website)

30

(75) Claudins – CLs (Claudins)

Cross-references

Offner S., et al *Cancer Immunol Immunother.* 2005 May; 54(5):431-45, Suzuki H., et al *Ann N Y Acad Sci.* 2012 Jul;1258:65-70)

35

In humans, 24 members of the family have been described – see literature reference.

(76) EGFR (Epidermal growth factor receptor)Nucleotide

Genbank accession no. NM_005228

5 Genbank version no. NM_005228.3 GI:41927737

Genbank record update date: Sep 30, 2012 01:47 PM

Polypeptide

Genbank accession no. NP_005219

10 Genbank version no. NP_005219.2 GI:29725609

Genbank record update date: Sep 30, 2012 01:47 PM

Cross-references

Dhomen NS., et al *Crit Rev Oncog.* 2012;17(1):31-50

15

Other information

Official Symbol: EGFR

Other Aliases: ERBB, ERBB1, HER1, PIG61, mENA

Other Designations: avian erythroblastic leukemia viral (v-erb-b) oncogene homolog; cell

20 growth inhibiting protein 40; cell proliferation-inducing protein 61; proto-oncogene c-ErbB-1;
receptor tyrosine-protein kinase erbB-1

ANTIBODIES

BMS: Cetuximab (ErbixTM) - Broadbridge VT., et al *Expert Rev Anticancer Ther.* 2012

25 May;12(5):555-65.

For example, see US6217866 – ATTC deposit No. 9764.

Amgen: Panitumumab (VectibixTM) - Argiles G., et al *Future Oncol.* 2012 Apr;8(4):373-89

For example, see US6235883 SEQ ID NOs: 23-38.

30

Genmab: Zalutumumab - Rivera F., et al *Expert Opin Biol Ther.* 2009 May;9(5):667-74.

YM Biosciences: Nimotuzumab - Ramakrishnan MS., et al *MAbs.* 2009 Jan-Feb;1(1):41-8.

For example, see US5891996 SEQ ID NOs: 27-34.

35

(77) *Her3 (ErbB3) – ERBB3 (v-erb-b2 erythroblastic leukemia viral oncogene homolog 3 (avian))*

Nucleotide

Genbank accession no. M34309

5 Genbank version no. M34309.1 GI:183990

Genbank record update date: Jun 23, 2010 08:47 PM

Polypeptide

Genbank accession no. AAA35979

10 Genbank version no. AAA35979.1 GI:306841

Genbank record update date: Jun 23, 2010 08:47 PM

Cross-references

Plowman, G.D., et al., *Proc. Natl. Acad. Sci. U.S.A.* 87 (13), 4905-4909 (1990)

15

Other information

Official Symbol: ERBB3

Other Aliases: ErbB-3, HER3, LCCS2, MDA-BF-1, c-erbB-3, c-erbB3, erbB3-S, p180-ErbB3, p45-sErbB3, p85-sErbB3

20 Other Designations: proto-oncogene-like protein c-ErbB-3; receptor tyrosine-protein kinase erbB-3; tyrosine kinase-type cell surface receptor HER3

ANTIBODIES

Merimack Pharma : MM-121 (Schoeberl B., et al Cancer Res. 2010 Mar 15;70(6):2485-2494)

25

For example, see US2011028129 SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7 and 8.

(78) *RON - MST1R (macrophage stimulating 1 receptor (c-met-related tyrosine kinase))*

Nucleotide

30 Genbank accession no. X70040

Genbank version no. X70040.1 GI:36109

Genbank record update date: Feb 02, 2011 10:17 PM

Polypeptide

35 Genbank accession no. CCA49634

Genbank version no. CCA49634.1 GI:36110

Genbank record update date: Feb 02, 2011 10:17 PM

Cross-references

Ronsin C., et al *Oncogene* 8 (5), 1195-1202 (1993)

5

Other information

Official Symbol: MST1R

Other Aliases: CD136, CDw136, PTK8, RON

Other Designations: MSP receptor; MST1R variant RON30; MST1R variant RON62; PTK8
 10 protein tyrosine kinase 8; RON variant E2E3; c-met-related tyrosine kinase; macrophage-
 stimulating protein receptor; p185-Ron; soluble RON variant 1; soluble RON variant 2;
 soluble RON variant 3; soluble RONvariant 4

(79) *EPHA2 (EPH receptor A2)*

15 Nucleotide

Genbank accession no. BC037166

Genbank version no. BC037166.2 GI:33879863

Genbank record update date: Mar 06, 2012 01:59 PM

20 Polypeptide

Genbank accession no. AAH37166

Genbank version no. AAH37166.1 GI:22713539

Genbank record update date: Mar 06, 2012 01:59 PM

25 Cross-references

Strausberg R.L., et al *Proc. Natl. Acad. Sci. U.S.A.* 99 (26), 16899-16903 (2002)

Other information

Official Symbol: EPHA2

30 Other Aliases: ARCC2, CTPA, CTPP1, ECK

Other Designations: ephrin type-A receptor 2; epithelial cell receptor protein tyrosine kinase;
 soluble EPHA2 variant 1; tyrosine-protein kinase receptor ECK

ANTIBODIES

35 Medimmune: 1C1 (Lee JW., et al *Clin Cancer Res.* 2010 May 1;16(9):2562-2570)

For example, see US20090304721A1 Fig. 7 and 8.

(80) CD20 – MS4A1 (membrane-spanning 4-domains, subfamily A, member 1)

Nucleotide

Genbank accession no. M27394

5 Genbank version no. M27394.1 GI:179307

Genbank record update date: Nov 30, 2009 11:16 AM

Polypeptide

Genbank accession no. AAA35581

10 Genbank version no. AAA35581.1 GI:179308

Genbank record update date: Nov 30, 2009 11:16 AM

Cross-references

Tedder T.F., et al *Proc. Natl. Acad. Sci. U.S.A.* 85 (1), 208-212 (1988)

15

Other information

Official Symbol: MS4A1

Other Aliases: B1, Bp35, CD20, CVID5, LEU-16, MS4A2, S7

Other Designations: B-lymphocyte antigen CD20; B-lymphocyte cell-surface antigen B1;

20 CD20 antigen; CD20 receptor; leukocyte surface antigen Leu-16

ANTIBODIES

Genentech/Roche: Rituximab - Abdulla NE., et al *BioDrugs*. 2012 Apr 1;26(2):71-82.

For example, see US5736137, ATCC deposit No. HB-69119.

25

GSK/Genmab: Ofatumumab - Nightingale G., et al *Ann Pharmacother*. 2011

Oct;45(10):1248-55.

For example, see US20090169550A1 SEQ ID NOs: 2, 4 and 5.

30 Immunomedics: Veltuzumab - Goldenberg DM., et al *Leuk Lymphoma*. 2010 May;51(5):747-55.

For example, see US7919273B2 SEQ ID NOs: 1, 2, 3, 4, 5 and 6.

(81) Tenascin C – TNC (Tenascin C)

35 Nucleotide

Genbank accession no. NM_002160

Genbank version no. NM_002160.3 GI:340745336

Genbank record update date: Sep 23, 2012 02:33 PM

Polypeptide

5 Genbank accession no. NP_002151

Genbank version no. NP_002151.2 GI:153946395

Genbank record update date: Sep 23, 2012 02:33 PM

Cross-references

10 Nies D.E., et al J. Biol. Chem. 266 (5), 2818-2823 (1991); Siri A., et al Nucleic Acids Res. 19 (3), 525-531 (1991)

Other information

Official Symbol: TNC

15 Other Aliases: 150-225, GMEM, GP, HXB, JI, TN, TN-C

Other Designations: GP 150-225; cytotactin; glioma-associated-extracellular matrix antigen; hexabrachion (tenascin); myotendinous antigen; neuronectin; tenascin; tenascin-C isoform 14/AD1/16

20 *ANTIBODIES*

Philogen : G11 (von Lukowicz T., et al *J Nucl Med.* 2007 Apr;48(4):582-7) and F16 (Pedretti M., et al Lung Cancer. 2009 Apr;64(1):28-33)

For example, see US7968685 SEQ ID NOs: 29, 35, 45 and 47.

25 **(82)** *FAP (Fibroblast activation protein, alpha)*

Nucleotide

Genbank accession no. U09278

Genbank version no. U09278.1 GI:1888315

Genbank record update date: Jun 23, 2010 09:22 AM

30

Polypeptide

Genbank accession no. AAB49652

Genbank version no. AAB49652.1 GI:1888316

Genbank record update date: Jun 23, 2010 09:22 AM

35

Cross-references

Scanlan, M.J., et al *Proc. Natl. Acad. Sci. U.S.A.* 91 (12), 5657-5661 (1994)

Other information

5 Official Symbol: FAP

Other Aliases: DPPIV, FAPA

Other Designations: 170 kDa melanoma membrane-bound gelatinase; integral membrane serine protease; seprase

10 **(83)** *DKK-1 (Dickkopf 1 homolog (Xenopus laevis))*

Nucleotide

Genbank accession no. NM_012242

Genbank version no. NM_012242.2 GI:61676924

Genbank record update date: Sep 30, 2012 01:48 PM

15

Polypeptide

Genbank accession no. NP_036374

Genbank version no. NP_036374.1 GI:7110719

Genbank record update date: Sep 30, 2012 01:48 PM

20

Cross-references

Fedi P. et al *J. Biol. Chem.* 274 (27), 19465-19472 (1999)

Other information

25 Official Symbol: DKK1

Other Aliases: UNQ492/PRO1008, DKK-1, SK

Other Designations: dickkopf related protein-1; dickkopf-1 like; dickkopf-like protein 1; dickkopf-related protein 1; hDkk-1

30 **ANTIBODIES**

Novartis: BHQ880 (Fulciniti M., et al *Blood*. 2009 Jul 9;114(2):371-379)

For example, see US20120052070A1 SEQ ID NOs: 100 and 108.

(84) *CD52 (CD52 molecule)*

35 Nucleotide

Genbank accession no. NM_001803

Genbank version no. NM_001803.2 GI:68342029

Genbank record update date: Sep 30, 2012 01:48 PM

Polypeptide

5 Genbank accession no. NP_001794

Genbank version no. NP_001794.2 GI:68342030

Genbank record update date: Sep 30, 2012 01:48 PM

Cross-references

10 Xia M.Q., et al *Eur. J. Immunol.* 21 (7), 1677-1684 (1991)

Other information

Official Symbol: CD52

Other Aliases: CDW52

15 Other Designations: CAMPATH-1™ antigen; CD52 antigen (CAMPATH-1 antigen); CDW52 antigen (CAMPATH-1 antigen); cambridge pathology 1 antigen; epididymal secretory protein E5; he5; human epididymis-specific protein 5

ANTIBODIES

20 Alemtuzumab (Campath™) - Skoetz N., et al *Cochrane Database Syst Rev.* 2012 Feb 15;2:CD008078.

For example, see Drugbank Acc. No. DB00087 (BIOD00109, BTD00109)

(85) CS1 - SLAMF7 (SLAM family member 7)

25 Nucleotide

Genbank accession no. NM_021181

Genbank version no. NM_021181.3 GI:1993571

Genbank record update date: Jun 29, 2012 11:24 AM

30 Polypeptide

Genbank accession no. NP_067004

Genbank version no. NP_067004.3 GI:19923572

Genbank record update date: Jun 29, 2012 11:24 AM

35 Cross-references

Boles K.S., et al *Immunogenetics* 52 (3-4), 302-307 (2001)

Other information

Official Symbol: SLAMF7

Other Aliases: UNQ576/PRO1138, 19A, CD319, CRACC, CS1

- 5 Other Designations: 19A24 protein; CD2 subset 1; CD2-like receptor activating cytotoxic cells; CD2-like receptor-activating cytotoxic cells; membrane protein FOAP-12; novel LY9 (lymphocyte antigen 9) like protein; protein 19A

ANTIBODIES

- 10 BMS: elotuzumab/HuLuc63 (Benson DM., et al *J Clin Oncol.* 2012 Jun 1;30(16):2013-2015)
For example, see US20110206701 SEQ ID NOs: 9, 10, 11, 12, 13, 14, 15 and 16.

(86) Endoglin – ENG (Endoglin)Nucleotide

- 15 Genbank accession no. AF035753
Genbank version no. AF035753.1 GI:3452260
Genbank record update date: Mar 10, 2010 06:36 PM

Polypeptide

- 20 Genbank accession no. AAC32802
Genbank version no. AAC32802.1 GI:3452261
Genbank record update date: Mar 10, 2010 06:36 PM

Cross-references

- 25 Rius C., et al *Blood* 92 (12), 4677-4690 (1998)
Official Symbol: ENG

Other information

Other Aliases: RP11-228B15.2, CD105, END, HHT1, ORW, ORW1

- 30 Other Designations: CD105 antigen

(87) Annexin A1 – ANXA1 (Annexin A1)Nucleotide

- Genbank accession no. X05908
35 Genbank version no. X05908.1 GI:34387
Genbank record update date: Feb 02, 2011 10:02 AM

Polypeptide

Genbank accession no. CCA29338

Genbank version no. CCA29338.1 GI:34388

5 Genbank record update date: Feb 02, 2011 10:02 AM

Cross-references

Wallner B.P., et al *Nature* 320 (6057), 77-81 (1986)

10 Other information

Official Symbol: ANXA1

Other Aliases: RP11-71A24.1, ANX1, LPC1

Other Designations: annexin I (lipocortin I); annexin-1; calpactin II; calpactin-2; chromobindin-9; lipocortin I; p35; phospholipase A2 inhibitory protein

15

(88) V-CAM (CD106) - VCAM1 (*Vascular cell adhesion molecule 1*)

Nucleotide

Genbank accession no. M60335

Genbank version no. M60335.1 GI:340193

20 Genbank record update date: Jun 23, 2010 08:56 AM

Polypeptide

Genbank accession no. AAA61269

Genbank version no. AAA61269.1 GI:340194

25 Genbank record update date: Jun 23, 2010 08:56 AM

Cross-references

Hession C., et al *J. Biol. Chem.* 266 (11), 6682-6685 (1991)

30 Other information

Official Symbol VCAM1

Other Aliases: CD106, INCAM-100

Other Designations: CD106 antigen; vascular cell adhesion protein 1

35

Antibody Sequences*Anti-Integrin $\alpha v\beta 6$* RHAB6.2

QVQLVQSGSELKKPGASVKISCKASGFAFTDSYMHWRQAPGQGLEWMGWIDPENGDT
 5 YAPKFQGRFVFSLDTSVSTAYLQISSLKAEDTAVYYCTRGTPPTAVPNLRGDLQVLAQKVAG
 PYPFDYWGQGTLVTVSS

RHCB6.2

QVQLVQSGAEVKKPGASVKVSCKASGYTFIDSYMHWRQAPGQRLEWMGWIDPENGDT
 10 YAPKFQGRVTITTDTSASTAYMELSSLRSEDVAVYYCARGTPPTAVPNLRGDLQVLAQKVAG
 PYPFDYWGQGTLVTVSS

RHF

QVQLVQSGAEVKKPGASVKVSCKASGFNFIDSYMHWRQAPGQRLEWMGWIDPENGDT
 15 EYAPKFQGRVTFTTDTASTAYMELSSLRSEDVAVYYCNEGTPPTGPYYFDYWGQGTLVTV
 SS

RHFB6

QVQLVQSGAEVKKPGASVKVSCKASGFNFIDSYMHWRQAPGQRLEWMGWIDPENGDT
 20 EYAPKFQGRVTFTTDTASTAYMELSSLRSEDVAVYYCNEGTPPTAVPNLRGDLQVLAQKVA
 GPYYFDYWGQGTLVTVSS

RHAY100bP

QVQLVQSGSELKKPGASVKISCKASGFAFTDSYMHWRQAPGQGLEWMGWIDPENGDT
 25 YAPKFQGRFVFSLDTSVSTAYLQISSLKAEDTAVYYCTRGTPPTGPYPFDYWGQGTLVTVSS

RKF

ENVLTQSPGTLSSLSPGERATLSCSASSSVSYMHWFQQKPGQAPRLLIYSTSNLASGIPDRF
 SGSGSGTDFTLTISRLEPEDFAVYYCQQRSSYPLTFGGGGTKVEIK
 30

RKFL36L50

ENVLTQSPGTLSSLSPGERATLSCSASSSVSYMHWLQQKPGQAPRLLIYLTSNLASGIPDRF
 SGSGSGTDFTLTISRLEPEDFAVYYCQQRSSYPLTFGGGGTKVEIK

RKC

EIVLTQSPGTLSSLSPGERATLSCSASSSVSYMHWFFQQKPGQAPRLLIYSTSNLASGIPDRFS
GSGSGTDFTLTISRLEPEDFAVYYCQQRSSYPLTFGGGGTKVEIK

5 *Anti-CD33*CD33 Hum195 VH

QVQLVQSGAEVKKPGSSVKVSCKASGYTFTDYNMHWVRQAPGQGLEWIGYIYPYNGGTG
YNQKFKSKATITADESTNTAYMELSSLRSEDVAVYYCARGRPAMDYWGQGTLVTVSS

10 CD33 Hum195 VK

DIQMTQSPSSLSASVGDRVTITCRASESDNYGISFMNWFQQKPGKAPKLLIYAASNQGS
VPSRFSGSGSGTDFTLTISLQPDFFATYYCQQSKEVPWTFGGGGTKVEIK

*Anti-CD19*15 CD19 B4 resurfaced VH

QVQLVQPGAEEVVKPGASVKLSCKTSGYTFTSNWMHWVKQRPGQGLEWIGEIDPSDSYTN
YNQNFKGKAKLTVDKSTSTAYMEVSSLRSDDTAVYYCARGSNPYYYAMDYWGQGTSVTV
SS

20 CD19 B4 resurfaced VK

EIVLTQSPAISASPGERVMTTCSASSGVNYMHWYQQKPGTSPRRWIYDTSKLASGVPAR
FSGSGSGTSYSLTISSMEDPAATYYCHQRGSYTFGGGGTKLEIK

*Anti-Her2*25 Herceptin VH chain

EVQLVESGGGLVQPGGSLRLSCAASGFMNIKDTYIHWVRQAPGKGLEWVARIYPTNGYTRY
ADSVKGRFTISADTSKNTAYLQMNSLRADDTAVYYCSRWGGDGFYAMDYWGQGTLVTVS
S

30 Herceptin VL chain

DIQMTQSPSSLSASVGDRVTITCRASQDVNTAVAWYQQKPGKAPKLLIYSASFLYSGVPSR
FSGSRSGTDFTLTISLQPEDFATYYCQQHYTTPPTFGGQGTKVEIK

*Anti-CD25*Simulect VK (also known as Basiliximab)

QIVSTQSPAIMASAPGEKVTMTCSASSRSYMQWYQQKPGTSPKRWIYDTSKLASGVPAR
FSGSGSGTSYSLTISSMEAEDAATYYCHQRSSYTFGGGGTKLEIK

5

Simulect VH

QLQQSGTVLARPGASVKMSCKASGYSFTRYWMHWIKQRPQGGLWIGAIYPGNSDTSYN
QKFEGKAKLTAVTSASTAYMELSSLTHEDSAVYYCSRDYGYDFDFWGQGTTTLTVSS

10 *Anti-PSMA*Deimmunised VH '1

EVQLVQSGPEVKKPGATVKISCKTSGYTFTEYTIHWVKQAPGKGLEWIGNINPNNGGTTYN
QKFEDKATLTVDKSTDAYMELSSLRSEDVAVYYCAAGWNFDYWGQGTTLLTVSS

15 Deimmunised VK '1

DIQMTQSPSSLSTSVGDRVTLTCKASQDVGTAVDWYQQKPGPSPKLLIYWASTRHTGIPSR
FSGSGSGTDFTLTISLQPEDFADYYCQQYNSYPLTFGPGTKVDIK

Deimmunised VH1 '5

20 EVKLVESGGGLVQPGGSMKLSCVASGFTFSNYWMNWVRQAPGKGLEWVAEIRSQSNNF
ATHYAESVKGRVTISRDDSKSIVYLQMNNLRAEDTGVYYCTRRWNNFWGQGTTTVTVSS

Deimmunised VH2 '5

EVKLVESGGGLVQPGGSLKLSCVASGFTFSNYWMNWVRQAPGKGLEWVAEIRSQSNNFA
25 THYAESVKGRVTISRDDSKSIVYLQMNNLRAEDTAVYYCTRRWNNFWGQGTTTVTVSS

Deimmunised VH3 '5

EVKLVESGGGLVQPGGSLKLSCVASGFTFSNYWMNWVRQAPGKGLEWVAEIRSQSNNFA
THYAESVKGRVTISRDDSKSIVYLQMNNLRAEDTAVYYCTRRWNNFWGQGTTTVTVSS

30

Deimmunised VH4 '5

EVKLVESGGGLVQPGGSLKLSCVASGFTFSNYWMNWVRQAPGKGLEWVAEIRSQSNNFA
THYAESVKGRFTISRDDSKSIVYLQMNNLRAEDTAVYYCTRRWNNFWGQGTTTVTVSS

35

Deimmunised VK1 '5

NIVMTQFPSSMSASVGDRVITITCKASENVGTYVSWYQQKPDQSPKMLIYGASNRFTGVDP
RFTGSGSATDFTLTISLQTEDLADYYCGQSYTFPYTFGQGGTKLEMK

5 Deimmunised VK2 '5

NIVMTQFPSSMSASVGDRVITITCKASENVGTYVSWYQQKPDQSPKMLIYGASNRFTGVDP
RFSGSGSGTDFTLTISLQAEDLADYYCGQSYTFPYTFGQGGTKLEIK

Deimmunised VK3 '5

10 NIQMTQFPSAMSASVGDRVITITCKASENVGTYVSWYQQKPDQSPKMLIYGASNRFTGVDP
RFSGSGSGTDFTLTISLQAEDLADYYCGQSYTFPYTFGQGGTKLEIK

Deimmunised VK4 '5

NIQMTQFPSAMSASVGDRVITITCKASENVGTYVSWYQQKPDQSPKMLIYGASNRFTGVDP
15 RFSGSGSGTDFTLTISLQAEDLADYYCGQSYTFPYTFGQGGTKLEIK

Deimmunised VK DI '5

NIVMTQFPKSMSASAGERMTLTCKASENVGTYVSWYQQKPTQSPKMLIYGASNRFTGVDP
RFSGSGSGTDFILTISVQAEDLVDYYCGQSYTFPYTFGGGTKLEMK
20

Deimmunised VH DI '5

EVKLEESGGGLVQPGGSMKISCVASGFTFSNYWMNWVRQSPEKGLEWVAEIRSQSNNFA
THYAESVKGRVIISRDDSKSSVYLQMNSLRAEDTAVYYCTRRWNNFWGQGTTVTVSS

25 Humanised RHA '5

EVQLVESGGGLVQPGGSLKLSCAASGFTFSNYWMNWVRQASGKGLEWVGEIRSQSNNFA
THYAESVKGRFTISRDDSKNTAYLQMNSLKTEDTAVYYCTRRWNNFWGQGTTVTVSS

Humanised RHB '5

30 EVKLVESGGGLVQPGGSLKLSCAASGFTFSNYWMNWVRQASGKGLEWVAEIRSQSNNFA
THYAESVKGRVIISRDDSKNTVYLQMNSLRTEDTAVYYCTRRWNNFWGQGTTVTVSS

Humanised RHC '5

EVQLVESGGGLVQPGGSLKLSCAASGFTFSNYWMNWVRQASGKGLEWVAEIRSQSNNFA
35 THYAESVKGRVIISRDDSKNTVYLQMNSLRTEDTAVYYCTRRWNNFWGQGTTVTVSS

Humanised RHD '5

EVKLVESGGGLVQPGGSLKLSCAASGFTFSNYWMNWVRQASGKGLEWVGEIRSQSNNFA
 THYAESVKGRVIISRDDSKNTVYLQMNSLRTEDTAVYYCTRRWNNFWGQGTTVTVSS

5 Humanised RHE '5

EVKLVESGGGLVQPGGSLKLSCAASGFTFSNYWMNWVRQASGKGLEWVAEIRSQSNNFA
 THYAESVKGRFTISRDDSKNTVYLQMNSLRTEDTAVYYCTRRWNNFWGQGTTVTVSS

Humanised RHF '5

10 EVKLVESGGGLVQPGGSLKLSCAASGFTFSNYWMNWVRQASGKGLEWVAEIRSQSNNFA
 THYAESVKGRVIISRDDSKNTAYLQMNSLRTEDTAVYYCTRRWNNFWGQGTTVTVSS

Humanised RHG '5

EVKLVESGGGLVQPGGSLKLSCAASGFTFSNYWMNWVRQASGKGLEWVAEIRSQSNNFA
 15 THYAESVKGRVIISRDDSKNTAYLQMNSLRTEDTAVYYCTRRWNNFWGQGTTVTVSS

Humanised RKA '5

DIQMTQSPSSVSASVGDRVITITCKASENVGTYVSWYQQKPGTAPKLLIYGASNRFTGVPSR
 FSGSGSATDFTLTINNLQPEDFATYYCGQSYTFPYTFGQGGTKVEIK
 20

Humanised RKB '5

DIQMTQSPSSVSASVGDRVITITCKASENVGTYVSWYQQKPGTAPKLLIYGASNRFTGVPSR
 FSGSGSATDFTLTINNLQPEDFATYYCGQSYTFPYTFGQGGTKVEIK

25 Humanised RKC '5

DIQMTQSPSSVSASVGDRVITITCKASENVGTYVSWYQQKPGTAPKMLIYGASNRFTGVPS
 RFSGSGSATDFTLTINNLQPEDFATYYCGQSYTFPYTFGQGGTKVEIK

Humanised RKD '5

30 DIQMTQSPSSVSASVGDRVITITCKASENVGTYVSWYQQKPGTAPKMLIYGASNRFTGVPS
 RFSGSGSATDFTLTINNLQPEDFATYYCGQSYTFPYTFGQGGTKVEIK

Humanised RKE '5

NIVMTQSPSSVSASVGDRVITITCKASENVGTYVSWYQQKPGTAPKLLIYGASNRFTGVPDR
 35 FTGSGSATDFILTINNLQPEDFATYYCGQSYTFPYTFGQGGTKVEIK

Humanised RKF '5

NIVMTQSPSSVSASVGDRVITITCKASENVGTYVSWYQQKPGTAPKMLIYGASNRFTGVPSR
FSGSGSATDFILTINNLPEDFATYYCGQSYTFPYTFGQGGTKVEIK

5 Humanised RKG '5

NIVMTQSPSSVSASVGDRVITITCKASENVGTYVSWYQQKPGTAPKMLIYGASNRFTGVPDR
FTGSGSATDFTLTINNLPEDFATYYCGQSYTFPYTFGQGGTKVEIK

- 10 The parent antibody may also be a fusion protein comprising an albumin-binding peptide (ABP) sequence (Dennis *et al.* (2002) "Albumin Binding As A General Strategy For Improving The Pharmacokinetics Of Proteins" *J Biol Chem.* 277:35035-35043; WO 01/45746). Antibodies of the invention include fusion proteins with ABP sequences taught by: (i) Dennis *et al* (2002) *J Biol Chem.* 277:35035-35043 at Tables III and IV, page 35038;
15 (ii) US 2004/0001827 at [0076]; and (iii) WO 01/45746 at pages 12-13.

In one embodiment, the antibody has been raised to target specific the tumour related antigen $\alpha_v\beta_6$.

- 20 The cell binding agent may be labelled, for example to aid detection or purification of the agent either prior to incorporation as a conjugate, or as part of the conjugate. The label may be a biotin label. In another embodiment, the cell binding agent may be labelled with a radioisotope.
- 25 Embodiments of the present invention include ConjA wherein the cell binding agent is selected from an antibody to any of the antigens discussed above.

Embodiments of the present invention include ConjB wherein the cell binding agent is selected from an antibody to any of the antigens discussed above.

30

Embodiments of the present invention include ConjA wherein the cell binding agent is selected from any of the antibodies discussed above.

- Embodiments of the present invention include ConjB wherein the cell binding agent is
35 selected from any of the antibodies discussed above.

The present invention may also relate to conjugates where the cell binding agent is selected from an antibody to any of the antigens discussed above and any of the antibodies discussed above linked to different drugs.

5

Drug loading

The drug loading is the average number of PBD drugs per cell binding agent, e.g. antibody. Where the compounds of the invention are bound to cysteines, drug loading may range from 1 to 8 drugs (D) per cell binding agent, i.e. where 1, 2, 3, 4, 5, 6, 7, and 8 drug moieties are covalently attached to the cell binding agent. Compositions of conjugates include collections of cell binding agents, e.g. antibodies, conjugated with a range of drugs, from 1 to 8. Where the compounds of the invention are bound to lysines, drug loading may range from 1 to 80 drugs (D) per cell binding agent, although an upper limit of 40, 20, 10 or 8 may be preferred. Compositions of conjugates include collections of cell binding agents, e.g. antibodies, conjugated with a range of drugs, from 1 to 80, 1 to 40, 1 to 20, 1 to 10 or 1 to 8.

The average number of drugs per antibody in preparations of ADC from conjugation reactions may be characterized by conventional means such as UV, reverse phase HPLC, HIC, mass spectroscopy, ELISA assay, and electrophoresis. The quantitative distribution of ADC in terms of p may also be determined. By ELISA, the averaged value of p in a particular preparation of ADC may be determined (Hamblett et al (2004) Clin. Cancer Res. 10:7063-7070; Sanderson et al (2005) Clin. Cancer Res. 11:843-852). However, the distribution of p (drug) values is not discernible by the antibody-antigen binding and detection limitation of ELISA. Also, ELISA assay for detection of antibody-drug conjugates does not determine where the drug moieties are attached to the antibody, such as the heavy chain or light chain fragments, or the particular amino acid residues. In some instances, separation, purification, and characterization of homogeneous ADC where p is a certain value from ADC with other drug loadings may be achieved by means such as reverse phase HPLC or electrophoresis. Such techniques are also applicable to other types of conjugates.

30

For some antibody-drug conjugates, p may be limited by the number of attachment sites on the antibody. For example, an antibody may have only one or several cysteine thiol groups, or may have only one or several sufficiently reactive thiol groups through which a linker may be attached. Higher drug loading, e.g. $p > 5$, may cause aggregation, insolubility, toxicity, or loss of cellular permeability of certain antibody-drug conjugates.

35

Typically, fewer than the theoretical maximum of drug moieties are conjugated to an antibody during a conjugation reaction. An antibody may contain, for example, many lysine residues that do not react with the drug-linker intermediate (D-L) or linker reagent. Only the most reactive lysine groups may react with an amine-reactive linker reagent. Also, only the most reactive cysteine thiol groups may react with a thiol-reactive linker reagent. Generally, antibodies do not contain many, if any, free and reactive cysteine thiol groups which may be linked to a drug moiety. Most cysteine thiol residues in the antibodies of the compounds exist as disulfide bridges and must be reduced with a reducing agent such as dithiothreitol (DTT) or TCEP, under partial or total reducing conditions. The loading (drug/antibody ratio) of an ADC may be controlled in several different manners, including: (i) limiting the molar excess of drug-linker intermediate (D-L) or linker reagent relative to antibody, (ii) limiting the conjugation reaction time or temperature, and (iii) partial or limiting reductive conditions for cysteine thiol modification.

Certain antibodies have reducible interchain disulfides, i.e. cysteine bridges. Antibodies may be made reactive for conjugation with linker reagents by treatment with a reducing agent such as DTT (dithiothreitol). Each cysteine bridge will thus form, theoretically, two reactive thiol nucleophiles. Additional nucleophilic groups can be introduced into antibodies through the reaction of lysines with 2-iminothiolane (Traut's reagent) resulting in conversion of an amine into a thiol. Reactive thiol groups may be introduced into the antibody (or fragment thereof) by engineering one, two, three, four, or more cysteine residues (e.g., preparing mutant antibodies comprising one or more non-native cysteine amino acid residues). US 7521541 teaches engineering antibodies by introduction of reactive cysteine amino acids.

Cysteine amino acids may be engineered at reactive sites in an antibody and which do not form intrachain or intermolecular disulfide linkages (Junutula, et al., 2008b Nature Biotech., 26(8):925-932; Dornan et al (2009) Blood 114(13):2721-2729; US 7521541; US 7723485; WO2009/052249). The engineered cysteine thiols may react with linker reagents or the drug-linker reagents of the present invention which have thiol-reactive, electrophilic groups such as maleimide or alpha-halo amides to form ADC with cysteine engineered antibodies and the PBD drug moieties. The location of the drug moiety can thus be designed, controlled, and known. The drug loading can be controlled since the engineered cysteine thiol groups typically react with thiol-reactive linker reagents or drug-linker reagents in high yield. Engineering an IgG antibody to introduce a cysteine amino acid by substitution at a single site on the heavy or light chain gives two new cysteines on the symmetrical antibody.

A drug loading near 2 can be achieved with near homogeneity of the conjugation product ADC.

Where more than one nucleophilic or electrophilic group of the antibody reacts with a drug-linker intermediate, or linker reagent followed by drug moiety reagent, then the resulting product is a mixture of ADC compounds with a distribution of drug moieties attached to an antibody, e.g. 1, 2, 3, etc. Liquid chromatography methods such as polymeric reverse phase (PLRP) and hydrophobic interaction (HIC) may separate compounds in the mixture by drug loading value. Preparations of ADC with a single drug loading value (p) may be isolated, however, these single loading value ADCs may still be heterogeneous mixtures because the drug moieties may be attached, via the linker, at different sites on the antibody.

Thus the antibody-drug conjugate compositions of the invention include mixtures of antibody-drug conjugate compounds where the antibody has one or more PBD drug moieties and where the drug moieties may be attached to the antibody at various amino acid residues.

In one embodiment, the average number of dimer pyrrolobenzodiazepine groups per cell binding agent is in the range 1 to 20. In some embodiments the range is selected from 1 to 8, 2 to 8, 2 to 6, 2 to 4, and 4 to 8.

In some embodiments, there is one dimer pyrrolobenzodiazepine group per cell binding agent.

25 *Includes Other Forms*

Unless otherwise specified, included in the above are the well known ionic, salt, solvate, and protected forms of these substituents. For example, a reference to carboxylic acid (-COOH) also includes the anionic (carboxylate) form (-COO⁻), a salt or solvate thereof, as well as conventional protected forms. Similarly, a reference to an amino group includes the protonated form (-N⁺HR¹R²), a salt or solvate of the amino group, for example, a hydrochloride salt, as well as conventional protected forms of an amino group. Similarly, a reference to a hydroxyl group also includes the anionic form (-O⁻), a salt or solvate thereof, as well as conventional protected forms.

Salts

It may be convenient or desirable to prepare, purify, and/or handle a corresponding salt of the active compound, for example, a pharmaceutically-acceptable salt. Examples of pharmaceutically acceptable salts are discussed in Berge, *et al.*, *J. Pharm. Sci.*, **66**, 1-19 (1977).

For example, if the compound is anionic, or has a functional group which may be anionic (e.g. -COOH may be -COO⁻), then a salt may be formed with a suitable cation. Examples of suitable inorganic cations include, but are not limited to, alkali metal ions such as Na⁺ and K⁺, alkaline earth cations such as Ca²⁺ and Mg²⁺, and other cations such as Al³⁺. Examples of suitable organic cations include, but are not limited to, ammonium ion (i.e. NH₄⁺) and substituted ammonium ions (e.g. NH₃R⁺, NH₂R₂⁺, NHR₃⁺, NR₄⁺). Examples of some suitable substituted ammonium ions are those derived from: ethylamine, diethylamine, dicyclohexylamine, triethylamine, butylamine, ethylenediamine, ethanolamine, diethanolamine, piperazine, benzylamine, phenylbenzylamine, choline, meglumine, and tromethamine, as well as amino acids, such as lysine and arginine. An example of a common quaternary ammonium ion is N(CH₃)₄⁺.

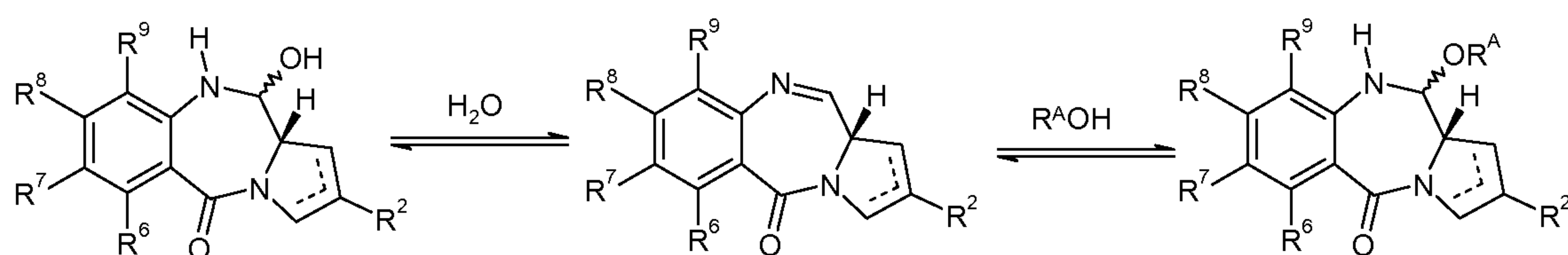
If the compound is cationic, or has a functional group which may be cationic (e.g. -NH₂ may be -NH₃⁺), then a salt may be formed with a suitable anion. Examples of suitable inorganic anions include, but are not limited to, those derived from the following inorganic acids: hydrochloric, hydrobromic, hydroiodic, sulfuric, sulfurous, nitric, nitrous, phosphoric, and phosphorous.

Examples of suitable organic anions include, but are not limited to, those derived from the following organic acids: 2-acetyoxybenzoic, acetic, ascorbic, aspartic, benzoic, camphorsulfonic, cinnamic, citric, edetic, ethanedisulfonic, ethanesulfonic, fumaric, glucheptonic, gluconic, glutamic, glycolic, hydroxymaleic, hydroxynaphthalene carboxylic, isethionic, lactic, lactobionic, lauric, maleic, malic, methanesulfonic, mucic, oleic, oxalic, palmitic, pantoic, pantothenic, phenylacetic, phenylsulfonic, propionic, pyruvic, salicylic, stearic, succinic, sulfanilic, tartaric, toluenesulfonic, trifluoroacetic acid and valeric. Examples of suitable polymeric organic anions include, but are not limited to, those derived from the following polymeric acids: tannic acid, carboxymethyl cellulose.

Solvates

It may be convenient or desirable to prepare, purify, and/or handle a corresponding solvate of the active compound. The term “solvate” is used herein in the conventional sense to refer to a complex of solute (e.g. active compound, salt of active compound) and solvent. If the solvent is water, the solvate may be conveniently referred to as a hydrate, for example, a mono-hydrate, a di-hydrate, a tri-hydrate, etc.

The invention includes compounds where a solvent adds across the imine bond of the PBD moiety, which is illustrated below where the solvent is water or an alcohol ($R^A\text{OH}$, where R^A is C_{1-4} alkyl):



These forms can be called the carbinolamine and carbinolamine ether forms of the PBD (as described in the section relating to R^{10} above). The balance of these equilibria depend on the conditions in which the compounds are found, as well as the nature of the moiety itself.

These particular compounds may be isolated in solid form, for example, by lyophilisation.

Isomers

Certain compounds of the invention may exist in one or more particular geometric, optical, enantiomeric, diastereomeric, epimeric, atropic, stereoisomeric, tautomeric, conformational, or anomeric forms, including but not limited to, cis- and trans-forms; E- and Z-forms; c-, t-, and r- forms; endo- and exo-forms; R-, S-, and meso-forms; D- and L-forms; d- and l-forms; (+) and (-) forms; keto-, enol-, and enolate-forms; syn- and anti-forms; synclinal- and anticlinal-forms; α - and β -forms; axial and equatorial forms; boat-, chair-, twist-, envelope-, and halfchair-forms; and combinations thereof, hereinafter collectively referred to as “isomers” (or “isomeric forms”).

The term “chiral” refers to molecules which have the property of non-superimposability of the mirror image partner, while the term “achiral” refers to molecules which are superimposable on their mirror image partner.

The term “stereoisomers” refers to compounds which have identical chemical constitution, but differ with regard to the arrangement of the atoms or groups in space.

“Diastereomer” refers to a stereoisomer with two or more centers of chirality and whose molecules are not mirror images of one another. Diastereomers have different physical properties, e.g. melting points, boiling points, spectral properties, and reactivities. Mixtures of diastereomers may separate under high resolution analytical procedures such as electrophoresis and chromatography.

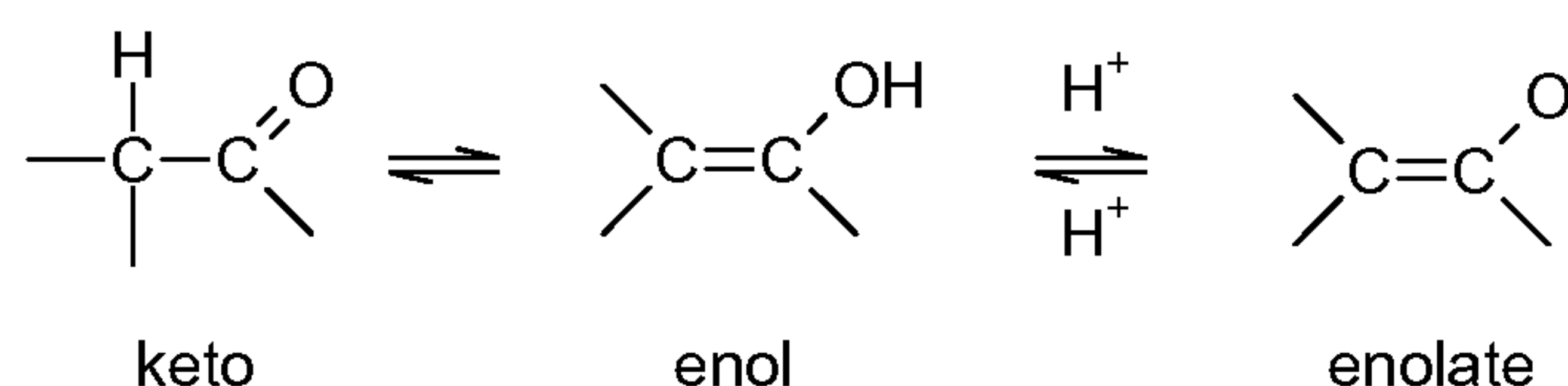
“Enantiomers” refer to two stereoisomers of a compound which are non-superimposable mirror images of one another.

Stereochemical definitions and conventions used herein generally follow S. P. Parker, Ed., *McGraw-Hill Dictionary of Chemical Terms* (1984) McGraw-Hill Book Company, New York; and Eliel, E. and Wilen, S., “Stereochemistry of Organic Compounds”, John Wiley & Sons, Inc., New York, 1994. The compounds of the invention may contain asymmetric or chiral centers, and therefore exist in different stereoisomeric forms. It is intended that all stereoisomeric forms of the compounds of the invention, including but not limited to, diastereomers, enantiomers and atropisomers, as well as mixtures thereof such as racemic mixtures, form part of the present invention. Many organic compounds exist in optically active forms, i.e., they have the ability to rotate the plane of plane-polarized light. In describing an optically active compound, the prefixes D and L, or R and S, are used to denote the absolute configuration of the molecule about its chiral center(s). The prefixes d and l or (+) and (-) are employed to designate the sign of rotation of plane-polarized light by the compound, with (-) or l meaning that the compound is levorotatory. A compound prefixed with (+) or d is dextrorotatory. For a given chemical structure, these stereoisomers are identical except that they are mirror images of one another. A specific stereoisomer may also be referred to as an enantiomer, and a mixture of such isomers is often called an enantiomeric mixture. A 50:50 mixture of enantiomers is referred to as a racemic mixture or a racemate, which may occur where there has been no stereoselection or stereospecificity in a chemical reaction or process. The terms “racemic mixture” and “racemate” refer to an equimolar mixture of two enantiomeric species, devoid of optical activity.

Note that, except as discussed below for tautomeric forms, specifically excluded from the term “isomers”, as used herein, are structural (or constitutional) isomers (i.e. isomers which differ in the connections between atoms rather than merely by the position of atoms in space). For example, a reference to a methoxy group, -OCH₃, is not to be construed as a reference to its structural isomer, a hydroxymethyl group, -CH₂OH. Similarly, a reference to

ortho-chlorophenyl is not to be construed as a reference to its structural isomer, meta-chlorophenyl. However, a reference to a class of structures may well include structurally isomeric forms falling within that class (e.g. C₁₋₇ alkyl includes n-propyl and iso-propyl; butyl includes n-, iso-, sec-, and tert-butyl; methoxyphenyl includes ortho-, meta-, and para-methoxyphenyl).

The above exclusion does not pertain to tautomeric forms, for example, keto-, enol-, and enolate-forms, as in, for example, the following tautomeric pairs: keto/enol (illustrated below), imine/enamine, amide/imino alcohol, amidine/amidine, nitroso/oxime, thioketone/enethiol, N-nitroso/hydroxyazo, and nitro/aci-nitro.



The term “tautomer” or “tautomeric form” refers to structural isomers of different energies which are interconvertible via a low energy barrier. For example, proton tautomers (also known as prototropic tautomers) include interconversions via migration of a proton, such as keto-enol and imine-enamine isomerizations. Valence tautomers include interconversions by reorganization of some of the bonding electrons.

Note that specifically included in the term “isomer” are compounds with one or more isotopic substitutions. For example, H may be in any isotopic form, including ¹H, ²H (D), and ³H (T); C may be in any isotopic form, including ¹²C, ¹³C, and ¹⁴C; O may be in any isotopic form, including ¹⁶O and ¹⁸O; and the like.

Examples of isotopes that can be incorporated into compounds of the invention include isotopes of hydrogen, carbon, nitrogen, oxygen, phosphorous, fluorine, and chlorine, such as, but not limited to ²H (deuterium, D), ³H (tritium), ¹¹C, ¹³C, ¹⁴C, ¹⁵N, ¹⁸F, ³¹P, ³²P, ³⁵S, ³⁶Cl, and ¹²⁵I. Various isotopically labeled compounds of the present invention, for example those into which radioactive isotopes such as ³H, ¹³C, and ¹⁴C are incorporated. Such isotopically labelled compounds may be useful in metabolic studies, reaction kinetic studies, detection or imaging techniques, such as positron emission tomography (PET) or single-photon emission computed tomography (SPECT) including drug or substrate tissue distribution assays, or in radioactive treatment of patients. Deuterium labelled or substituted therapeutic compounds of the invention may have improved DMPK (drug metabolism and

pharmacokinetics) properties, relating to distribution, metabolism, and excretion (ADME). Substitution with heavier isotopes such as deuterium may afford certain therapeutic advantages resulting from greater metabolic stability, for example increased in vivo half-life or reduced dosage requirements. An ^{18}F labeled compound may be useful for PET or
5 SPECT studies. Isotopically labeled compounds of this invention and prodrugs thereof can generally be prepared by carrying out the procedures disclosed in the schemes or in the examples and preparations described below by substituting a readily available isotopically labeled reagent for a non-isotopically labeled reagent. Further, substitution with heavier isotopes, particularly deuterium (i.e., ^2H or D) may afford certain therapeutic advantages
10 resulting from greater metabolic stability, for example increased in vivo half-life or reduced dosage requirements or an improvement in therapeutic index. It is understood that deuterium in this context is regarded as a substituent. The concentration of such a heavier isotope, specifically deuterium, may be defined by an isotopic enrichment factor. In the compounds of this invention any atom not specifically designated as a particular isotope is meant to
15 represent any stable isotope of that atom.

Unless otherwise specified, a reference to a particular compound includes all such isomeric forms, including (wholly or partially) racemic and other mixtures thereof. Methods for the preparation (e.g. asymmetric synthesis) and separation (e.g. fractional crystallisation and
20 chromatographic means) of such isomeric forms are either known in the art or are readily obtained by adapting the methods taught herein, or known methods, in a known manner.

Biological Activity

In vitro cell proliferation assays

25 Generally, the cytotoxic or cytostatic activity of an antibody-drug conjugate (ADC) is measured by: exposing mammalian cells having receptor proteins, e.g. HER2, to the antibody of the ADC in a cell culture medium; culturing the cells for a period from about 6 hours to about 5 days; and measuring cell viability. Cell-based *in vitro* assays are used to measure viability (proliferation), cytotoxicity, and induction of apoptosis (caspase activation)
30 of an ADC of the invention.

The *in vitro* potency of antibody-drug conjugates can be measured by a cell proliferation assay. The CellTiter-Glo[®] Luminescent Cell Viability Assay is a commercially available (Promega Corp., Madison, WI), homogeneous assay method based on the recombinant
35 expression of *Coleoptera* luciferase (US Patent Nos. 5583024; 5674713 and 5700670). This cell proliferation assay determines the number of viable cells in culture based on quantitation

of the ATP present, an indicator of metabolically active cells (Crouch *et al* (1993) *J. Immunol. Meth.* 160:81-88; US 6602677). The CellTiter-Glo[®] Assay is conducted in 96 well format, making it amenable to automated high-throughput screening (HTS) (Cree *et al* (1995) *AntiCancer Drugs* 6:398-404). The homogeneous assay procedure involves adding the
5 single reagent (CellTiter-Glo[®] Reagent) directly to cells cultured in serum-supplemented medium. Cell washing, removal of medium and multiple pipetting steps are not required. The system detects as few as 15 cells/well in a 384-well format in 10 minutes after adding reagent and mixing. The cells may be treated continuously with ADC, or they may be treated and separated from ADC. Generally, cells treated briefly, i.e. 3 hours, showed the
10 same potency effects as continuously treated cells.

The homogeneous “add-mix-measure” format results in cell lysis and generation of a luminescent signal proportional to the amount of ATP present. The amount of ATP is directly proportional to the number of cells present in culture. The CellTiter-Glo[®] Assay generates a
15 “glow-type” luminescent signal, produced by the luciferase reaction, which has a half-life generally greater than five hours, depending on cell type and medium used. Viable cells are reflected in relative luminescence units (RLU). The substrate, Beetle Luciferin, is oxidatively decarboxylated by recombinant firefly luciferase with concomitant conversion of ATP to AMP and generation of photons.

20 The *in vitro* potency of antibody-drug conjugates can also be measured by a cytotoxicity assay. Cultured adherent cells are washed with PBS, detached with trypsin, diluted in complete medium, containing 10% FCS, centrifuged, re-suspended in fresh medium and counted with a haemocytometer. Suspension cultures are counted directly. Monodisperse
25 cell suspensions suitable for counting may require agitation of the suspension by repeated aspiration to break up cell clumps.

The cell suspension is diluted to the desired seeding density and dispensed (100µl per well) into black 96 well plates. Plates of adherent cell lines are incubated overnight to allow
30 adherence. Suspension cell cultures can be used on the day of seeding.

A stock solution (1ml) of ADC (20µg/ml) is made in the appropriate cell culture medium. Serial 10-fold dilutions of stock ADC are made in 15ml centrifuge tubes by serially transferring 100µl to 900µl of cell culture medium.

Four replicate wells of each ADC dilution (100µl) are dispensed in 96-well black plates, previously plated with cell suspension (100µl), resulting in a final volume of 200 µl. Control wells receive cell culture medium (100µl).

If the doubling time of the cell line is greater than 30 hours, ADC incubation is for 5 days,
5 otherwise a four day incubation is done.

At the end of the incubation period, cell viability is assessed with the Alamar Blue™ assay. AlamarBlue (Invitrogen) is dispensed over the whole plate (20µl per well) and incubated for 4 hours. Alamar Blue fluorescence is measured at excitation 570nm, emission 585nm on the Varioskan™ flash plate reader. Percentage cell survival is calculated from the mean
10 fluorescence in the ADC treated wells compared to the mean fluorescence in the control wells.

In vivo efficacy

The *in vivo* efficacy of antibody-drug conjugates (ADC) of the invention can be measured by
15 tumor xenograft studies in mice. For example, the *in vivo* efficacy of an anti-HER2 ADC of the invention can be measured by a high expressing HER2 transgenic explant mouse model. An allograft is propagated from the Fo5 mmtv transgenic mouse which does not respond to, or responds poorly to, HERCEPTIN® therapy. Subjects are treated once with ADC at certain dose levels (mg/kg) and PBD drug exposure (µg/m²); and placebo buffer control
20 (Vehicle) and monitored over two weeks or more to measure the time to tumor doubling, log cell kill, and tumor shrinkage.

Use

The conjugates of the invention may be used to provide a PBD compound at a target
25 location.

The target location is preferably a proliferative cell population. The antibody is an antibody for an antigen present on a proliferative cell population.

30 In one embodiment the antigen is absent or present at a reduced level in a non-proliferative cell population compared to the amount of antigen present in the proliferative cell population, for example a tumour cell population.

At the target location the linker may be cleaved so as to release a compound RelA or RelB. Thus, the conjugate may be used to selectively provide a compound RelA or RelB to the target location.

- 5 The linker may be cleaved by an enzyme present at the target location.

The target location may be *in vitro*, *in vivo* or *ex vivo*.

- 10 The antibody-drug conjugate (ADC) compounds of the invention include those with utility for anticancer activity. In particular, the compounds include an antibody conjugated, i.e. covalently attached by a linker, to a PBD drug moiety, i.e. toxin. When the drug is not conjugated to an antibody, the PBD drug has a cytotoxic effect. The biological activity of the PBD drug moiety is thus modulated by conjugation to an antibody. The antibody-drug conjugates (ADC) of the invention selectively deliver an effective dose of a cytotoxic agent to
15 tumor tissue whereby greater selectivity, i.e. a lower efficacious dose, may be achieved.

Thus, in one aspect, the present invention provides a conjugate compound as described herein for use in therapy.

- 20 In a further aspect there is also provides a conjugate compound as described herein for use in the treatment of a proliferative disease. A second aspect of the present invention provides the use of a conjugate compound in the manufacture of a medicament for treating a proliferative disease.

- 25 One of ordinary skill in the art is readily able to determine whether or not a candidate conjugate treats a proliferative condition for any particular cell type. For example, assays which may conveniently be used to assess the activity offered by a particular compound are described in the examples below.

- 30 The term "proliferative disease" pertains to an unwanted or uncontrolled cellular proliferation of excessive or abnormal cells which is undesired, such as, neoplastic or hyperplastic growth, whether *in vitro* or *in vivo*.

- Examples of proliferative conditions include, but are not limited to, benign, pre-malignant,
35 and malignant cellular proliferation, including but not limited to, neoplasms and tumours (e.g. histocytoma, glioma, astrocytoma, osteoma), cancers (e.g. lung cancer, small cell lung

cancer, gastrointestinal cancer, bowel cancer, colon cancer, breast carcinoma, ovarian carcinoma, prostate cancer, testicular cancer, liver cancer, kidney cancer, bladder cancer, pancreas cancer, brain cancer, sarcoma, osteosarcoma, Kaposi's sarcoma, melanoma), lymphomas, leukemias, psoriasis, bone diseases, fibroproliferative disorders (e.g. of
5 connective tissues), and atherosclerosis. Cancers of particular interest include, but are not limited to, leukemias and ovarian cancers.

Any type of cell may be treated, including but not limited to, lung, gastrointestinal (including, e.g. bowel, colon), breast (mammary), ovarian, prostate, liver (hepatic), kidney (renal),
10 bladder, pancreas, brain, and skin.

In one embodiment, the treatment is of a pancreatic cancer.

In one embodiment, the treatment is of a tumour having $\alpha_v\beta_6$ integrin on the surface of the cell.
15

It is contemplated that the antibody-drug conjugates (ADC) of the present invention may be used to treat various diseases or disorders, e.g. characterized by the overexpression of a tumor antigen. Exemplary conditions or hyperproliferative disorders include benign or malignant tumors; leukemia, haematological, and lymphoid malignancies. Others include
20 neuronal, glial, astrocytal, hypothalamic, glandular, macrophagal, epithelial, stromal, blastocoelic, inflammatory, angiogenic and immunologic, including autoimmune, disorders.

Generally, the disease or disorder to be treated is a hyperproliferative disease such as cancer. Examples of cancer to be treated herein include, but are not limited to, carcinoma, lymphoma, blastoma, sarcoma, and leukemia or lymphoid malignancies. More particular
25 examples of such cancers include squamous cell cancer (e.g. epithelial squamous cell cancer), lung cancer including small-cell lung cancer, non-small cell lung cancer, adenocarcinoma of the lung and squamous carcinoma of the lung, cancer of the peritoneum, hepatocellular cancer, gastric or stomach cancer including gastrointestinal cancer, pancreatic cancer, glioblastoma, cervical cancer, ovarian cancer, liver cancer, bladder
30 cancer, hepatoma, breast cancer, colon cancer, rectal cancer, colorectal cancer, endometrial or uterine carcinoma, salivary gland carcinoma, kidney or renal cancer, prostate cancer, vulval cancer, thyroid cancer, hepatic carcinoma, anal carcinoma, penile carcinoma, as well as head and neck cancer.

Autoimmune diseases for which the ADC compounds may be used in treatment include rheumatologic disorders (such as, for example, rheumatoid arthritis, Sjögren's syndrome, scleroderma, lupus such as SLE and lupus nephritis, polymyositis/dermatomyositis, cryoglobulinemia, anti-phospholipid antibody syndrome, and psoriatic arthritis), osteoarthritis, autoimmune gastrointestinal and liver disorders (such as, for example, inflammatory bowel diseases (e.g. ulcerative colitis and Crohn's disease), autoimmune gastritis and pernicious anemia, autoimmune hepatitis, primary biliary cirrhosis, primary sclerosing cholangitis, and celiac disease), vasculitis (such as, for example, ANCA-associated vasculitis, including Churg-Strauss vasculitis, Wegener's granulomatosis, and polyarteritis), autoimmune neurological disorders (such as, for example, multiple sclerosis, opsoclonus myoclonus syndrome, myasthenia gravis, neuromyelitis optica, Parkinson's disease, Alzheimer's disease, and autoimmune polyneuropathies), renal disorders (such as, for example, glomerulonephritis, Goodpasture's syndrome, and Berger's disease), autoimmune dermatologic disorders (such as, for example, psoriasis, urticaria, hives, pemphigus vulgaris, bullous pemphigoid, and cutaneous lupus erythematosus), hematologic disorders (such as, for example, thrombocytopenic purpura, thrombotic thrombocytopenic purpura, post-transfusion purpura, and autoimmune hemolytic anemia), atherosclerosis, uveitis, autoimmune hearing diseases (such as, for example, inner ear disease and hearing loss), Behcet's disease, Raynaud's syndrome, organ transplant, and autoimmune endocrine disorders (such as, for example, diabetic-related autoimmune diseases such as insulin-dependent diabetes mellitus (IDDM), Addison's disease, and autoimmune thyroid disease (e.g. Graves' disease and thyroiditis)). More preferred such diseases include, for example, rheumatoid arthritis, ulcerative colitis, ANCA-associated vasculitis, lupus, multiple sclerosis, Sjögren's syndrome, Graves' disease, IDDM, pernicious anemia, thyroiditis, and glomerulonephritis.

Methods of Treatment

The conjugates of the present invention may be used in a method of therapy. Also provided is a method of treatment, comprising administering to a subject in need of treatment a therapeutically-effective amount of a conjugate compound of the invention. The term "therapeutically effective amount" is an amount sufficient to show benefit to a patient. Such benefit may be at least amelioration of at least one symptom. The actual amount administered, and rate and time-course of administration, will depend on the nature and severity of what is being treated. Prescription of treatment, e.g. decisions on dosage, is within the responsibility of general practitioners and other medical doctors.

A compound of the invention may be administered alone or in combination with other treatments, either simultaneously or sequentially dependent upon the condition to be treated. Examples of treatments and therapies include, but are not limited to, chemotherapy (the administration of active agents, including, e.g. drugs, such as chemotherapeutics); surgery;
 5 and radiation therapy.

A “chemotherapeutic agent” is a chemical compound useful in the treatment of cancer, regardless of mechanism of action. Classes of chemotherapeutic agents include, but are not limited to: alkylating agents, antimetabolites, spindle poison plant alkaloids,
 10 cytotoxic/antitumor antibiotics, topoisomerase inhibitors, antibodies, photosensitizers, and kinase inhibitors. Chemotherapeutic agents include compounds used in “targeted therapy” and conventional chemotherapy.

Examples of chemotherapeutic agents include: erlotinib (TARCEVA®, Genentech/OSI
 15 Pharm.), docetaxel (TAXOTERE®, Sanofi-Aventis), 5-FU (fluorouracil, 5-fluorouracil, CAS No. 51-21-8), gemcitabine (GEMZAR®, Lilly), PD-0325901 (CAS No. 391210-10-9, Pfizer), cisplatin (cis-diamine, dichloroplatinum(II), CAS No. 15663-27-1), carboplatin (CAS No. 41575-94-4), paclitaxel (TAXOL®, Bristol-Myers Squibb Oncology, Princeton, N.J.), trastuzumab (HERCEPTIN®, Genentech), temozolomide (4-methyl-5-oxo- 2,3,4,6,8-
 20 pentazabicyclo [4.3.0] nona-2,7,9-triene- 9-carboxamide, CAS No. 85622-93-1, TEMODAR®, TEMODAL®, Schering Plough), tamoxifen ((Z)-2-[4-(1,2-diphenylbut-1-enyl)phenoxy]-N,N-dimethylethanamine, NOLVADEX®, ISTUBAL®, VALODEX®), and doxorubicin (ADRIAMYCIN®), Akti-1/2, HPPD, and rapamycin.

More examples of chemotherapeutic agents include: oxaliplatin (ELOXATIN®, Sanofi),
 25 bortezomib (VELCADE®, Millennium Pharm.), sunitinib (SUNITINIB®, SU11248, Pfizer), letrozole (FEMARA®, Novartis), imatinib mesylate (GLEEVEC®, Novartis), XL-518 (Mek inhibitor, Exelixis, WO 2007/044515), ARRY-886 (Mek inhibitor, AZD6244, Array BioPharma, Astra Zeneca), SF-1126 (PI3K inhibitor, Semafore Pharmaceuticals), BEZ-235 (PI3K inhibitor, Novartis), XL-147 (PI3K inhibitor, Exelixis), PTK787/ZK 222584 (Novartis),
 30 fulvestrant (FASLODEX®, AstraZeneca), leucovorin (folinic acid), rapamycin (sirolimus, RAPAMUNE®, Wyeth), lapatinib (TYKERB®, GSK572016, Glaxo Smith Kline), lonafarnib (SARASAR™, SCH 66336, Schering Plough), sorafenib (NEXAVAR®, BAY43-9006, Bayer Labs), gefitinib (IRESSA®, AstraZeneca), irinotecan (CAMPTOSAR®, CPT-11, Pfizer), tipifarnib (ZARNESTRA™, Johnson & Johnson), ABRAXANE™ (Cremophor-free), albumin-
 35 engineered nanoparticle formulations of paclitaxel (American Pharmaceutical Partners, Schaumburg, IL), vandetanib (rINN, ZD6474, ZACTIMA®, AstraZeneca), chloranmbucil,

AG1478, AG1571 (SU 5271; Sugen), temsirolimus (TORISEL®, Wyeth), pazopanib (GlaxoSmithKline), canfosfamide (TELCYTA®, Telik), thiotepa and cyclophosphamide (CYTOXAN®, NEOSAR®); alkyl sulfonates such as busulfan, improsulfan and piposulfan; aziridines such as benzodopa, carboquone, meturedopa, and uredopa; ethylenimines and

5 methylamelamines including altretamine, triethylenemelamine, triethylenephosphoramidate, triethylenethiophosphoramidate and trimethylmelamine; acetogenins (especially bullatacin and bullatacinone); a camptothecin (including the synthetic analog topotecan); bryostatin; callystatin; CC-1065 (including its adozelesin, carzelesin and bizelesin synthetic analogs); cryptophycins (particularly cryptophycin 1 and cryptophycin 8); dolastatin; duocarmycin

10 (including the synthetic analogs, KW-2189 and CB1-TM1); eleutherobin; pancratistatin; a sarcodictyin; spongistatin; nitrogen mustards such as chlorambucil, chlornaphazine, chlorophosphamide, estramustine, ifosfamide, mechlorethamine, mechlorethamine oxide hydrochloride, melphalan, novembichin, phenesterine, prednimustine, trofosfamide, uracil mustard; nitrosoureas such as carmustine, chlorozotocin, fotemustine, lomustine, nimustine,

15 and ranimustine; antibiotics such as the enediyne antibiotics (e.g. calicheamicin, calicheamicin gamma11, calicheamicin omega1 (*Angew Chem. Intl. Ed. Engl.* (1994) 33:183-186); dynemicin, dynemicin A; bisphosphonates, such as clodronate; an esperamicin; as well as neocarzinostatin chromophore and related chromoprotein enediyne antibiotic chromophores), aclacinomysins, actinomycin, authramycin, azaserine, bleomycins,

20 cactinomycin, carabacin, carminomycin, carzinophilin, chromomycinis, dactinomycin, daunorubicin, detorubicin, 6-diazo-5-oxo-L-norleucine, morpholino-doxorubicin, cyanomorpholino-doxorubicin, 2-pyrrolino-doxorubicin and deoxydoxorubicin), epirubicin, esorubicin, idarubicin, nemorubicin, marcellomycin, mitomycins such as mitomycin C, mycophenolic acid, nogalamycin, olivomycins, peplomycin, porfiromycin, puromycin,

25 quelamycin, rodorubicin, streptonigrin, streptozocin, tubercidin, ubenimex, zinostatin, zorubicin; anti-metabolites such as methotrexate and 5-fluorouracil (5-FU); folic acid analogs such as denopterin, methotrexate, pteropterin, trimetrexate; purine analogs such as fludarabine, 6-mercaptopurine, thiamiprine, thioguanine; pyrimidine analogs such as ancitabine, azacitidine, 6-azauridine, carmofur, cytarabine, dideoxyuridine, doxifluridine,

30 enocitabine, floxuridine; androgens such as calusterone, dromostanolone propionate, epitostanol, mepitostane, testolactone; anti-adrenals such as aminoglutethimide, mitotane, trilostane; folic acid replenisher such as frolinic acid; aceglatone; aldophosphamide glycoside; aminolevulinic acid; eniluracil; amsacrine; bestrabucil; bisantrene; edatraxate; defofamine; demecolcine; diaziquone; elfornithine; elliptinium acetate; an epothilone;

35 etoglucid; gallium nitrate; hydroxyurea; lentinan; lonidainine; maytansinoids such as maytansine and ansamitocins; mitoguazone; mitoxantrone; mopidanmol; nitraerine;

pentostatin; phenamet; pirarubicin; losoxantrone; podophyllinic acid; 2-ethylhydrazide; procarbazine; PSK® polysaccharide complex (JHS Natural Products, Eugene, OR); razoxane; rhizoxin; sizofiran; spirogermanium; tenuazonic acid; triaziquone; 2,2',2"-trichlorotriethylamine; trichothecenes (especially T-2 toxin, verracurin A, roridin A and anguidine); urethan; vindesine; dacarbazine; mannomustine; mitobronitol; mitolactol; pipobroman; gacytosine; arabinoside ("Ara-C"); cyclophosphamide; thiotepa; 6-thioguanine; mercaptopurine; methotrexate; platinum analogs such as cisplatin and carboplatin; vinblastine; etoposide (VP-16); ifosfamide; mitoxantrone; vincristine; vinorelbine (NAVELBINE®); novantrone; teniposide; edatrexate; daunomycin; aminopterin; capecitabine (XELODA®, Roche); ibandronate; CPT-11; topoisomerase inhibitor RFS 2000; difluoromethylornithine (DMFO); retinoids such as retinoic acid; and pharmaceutically acceptable salts, acids and derivatives of any of the above.

Also included in the definition of "chemotherapeutic agent" are: (i) anti-hormonal agents that act to regulate or inhibit hormone action on tumors such as anti-estrogens and selective estrogen receptor modulators (SERMs), including, for example, tamoxifen (including NOLVADEX®; tamoxifen citrate), raloxifene, droloxifene, 4-hydroxytamoxifen, trioxifene, keoxifene, LY117018, onapristone, and FARESTON® (toremifine citrate); (ii) aromatase inhibitors that inhibit the enzyme aromatase, which regulates estrogen production in the adrenal glands, such as, for example, 4(5)-imidazoles, aminoglutethimide, MEGASE® (megestrol acetate), AROMASIN® (exemestane; Pfizer), formestanie, fadrozole, RIVISOR® (vorozole), FEMARA® (letrozole; Novartis), and ARIMIDEX® (anastrozole; AstraZeneca); (iii) anti-androgens such as flutamide, nilutamide, bicalutamide, leuprolide, and goserelin; as well as troxacitabine (a 1,3-dioxolane nucleoside cytosine analog); (iv) protein kinase inhibitors such as MEK inhibitors (WO 2007/044515); (v) lipid kinase inhibitors; (vi) antisense oligonucleotides, particularly those which inhibit expression of genes in signaling pathways implicated in aberrant cell proliferation, for example, PKC-alpha, Raf and H-Ras, such as oblimersen (GENASENSE®, Genta Inc.); (vii) ribozymes such as VEGF expression inhibitors (e.g., ANGIOZYME®) and HER2 expression inhibitors; (viii) vaccines such as gene therapy vaccines, for example, ALLOVECTIN®, LEUVECTIN®, and VAXID®; PROLEUKIN® rIL-2; topoisomerase 1 inhibitors such as LURTOTECAN®; ABARELIX® rmRH; (ix) anti-angiogenic agents such as bevacizumab (AVASTIN®, Genentech); and pharmaceutically acceptable salts, acids and derivatives of any of the above.

Also included in the definition of "chemotherapeutic agent" are therapeutic antibodies such as alemtuzumab (Campath), bevacizumab (AVASTIN®, Genentech); cetuximab (ERBITUX®, Imclone); panitumumab (VECTIBIX®, Amgen), rituximab (RITUXAN®,

Genentech/Biogen Idec), ofatumumab (ARZERRA®, GSK), pertuzumab (PERJETA™, OMNITARG™, 2C4, Genentech), trastuzumab (HERCEPTIN®, Genentech), tositumomab (Bexxar™, Corixa), and the antibody drug conjugate, gemtuzumab ozogamicin (MYLOTARG®, Wyeth).

5

Humanized monoclonal antibodies with therapeutic potential as chemotherapeutic agents in combination with the conjugates of the invention include: alemtuzumab, apolizumab, aselizumab, atlizumab, bapineuzumab, bevacizumab, bivatuzumab mertansine, cantuzumab mertansine, cedelizumab, certolizumab pegol, cidfusituzumab, cidtuzumab, daclizumab, 10 eculizumab, efalizumab, epratuzumab, erlizumab, felvizumab, fontolizumab, gemtuzumab ozogamicin, inotuzumab ozogamicin, ipilimumab, labetuzumab, lintuzumab, matuzumab, mepolizumab, motavizumab, motovizumab, natalizumab, nimotuzumab, nolovizumab, numavizumab, ocrelizumab, omalizumab, palivizumab, pascolizumab, pecfusituzumab, pectuzumab, pertuzumab, pexelizumab, ralivizumab, ranibizumab, reslivizumab, reslizumab, 15 resyvizumab, rovelizumab, ruplizumab, sibrotuzumab, siplizumab, sontuzumab, tacatuzumab tetraxetan, tadocizumab, talizumab, tefibazumab, tocilizumab, toralizumab, trastuzumab, tucotuzumab celmoleukin, tucosituzumab, umavizumab, urtoxazumab, and visilizumab.

20 Pharmaceutical compositions according to the present invention, and for use in accordance with the present invention, may comprise, in addition to the active ingredient, i.e. a conjugate compound, a pharmaceutically acceptable excipient, carrier, buffer, stabiliser or other materials well known to those skilled in the art. Such materials should be non-toxic and should not interfere with the efficacy of the active ingredient. The precise nature of the 25 carrier or other material will depend on the route of administration, which may be oral, or by injection, e.g. cutaneous, subcutaneous, or intravenous.

Pharmaceutical compositions for oral administration may be in tablet, capsule, powder or liquid form. A tablet may comprise a solid carrier or an adjuvant. Liquid pharmaceutical 30 compositions generally comprise a liquid carrier such as water, petroleum, animal or vegetable oils, mineral oil or synthetic oil. Physiological saline solution, dextrose or other saccharide solution or glycols such as ethylene glycol, propylene glycol or polyethylene glycol may be included. A capsule may comprise a solid carrier such a gelatin.

35 For intravenous, cutaneous or subcutaneous injection, or injection at the site of affliction, the active ingredient will be in the form of a parenterally acceptable aqueous solution which is

pyrogen-free and has suitable pH, isotonicity and stability. Those of relevant skill in the art are well able to prepare suitable solutions using, for example, isotonic vehicles such as Sodium Chloride Injection, Ringer's Injection, Lactated Ringer's Injection. Preservatives, stabilisers, buffers, antioxidants and/or other additives may be included, as required.

5

Formulations

While it is possible for the conjugate compound to be used (e.g., administered) alone, it is often preferable to present it as a composition or formulation.

10 In one embodiment, the composition is a pharmaceutical composition (e.g., formulation, preparation, medicament) comprising a conjugate compound, as described herein, and a pharmaceutically acceptable carrier, diluent, or excipient.

15 In one embodiment, the composition is a pharmaceutical composition comprising at least one conjugate compound, as described herein, together with one or more other pharmaceutically acceptable ingredients well known to those skilled in the art, including, but not limited to, pharmaceutically acceptable carriers, diluents, excipients, adjuvants, fillers, buffers, preservatives, anti-oxidants, lubricants, stabilisers, solubilisers, surfactants (e.g., wetting agents), masking agents, colouring agents, flavouring agents, and sweetening
20 agents.

In one embodiment, the composition further comprises other active agents, for example, other therapeutic or prophylactic agents.

25 Suitable carriers, diluents, excipients, etc. can be found in standard pharmaceutical texts. See, for example, Handbook of Pharmaceutical Additives, 2nd Edition (eds. M. Ash and I. Ash), 2001 (Synapse Information Resources, Inc., Endicott, New York, USA), Remington's Pharmaceutical Sciences, 20th edition, pub. Lippincott, Williams & Wilkins, 2000; and Handbook of Pharmaceutical Excipients, 2nd edition, 1994.

30

Another aspect of the present invention pertains to methods of making a pharmaceutical composition comprising admixing at least one [¹¹C]-radiolabelled conjugate or conjugate-like compound, as defined herein, together with one or more other pharmaceutically acceptable ingredients well known to those skilled in the art, e.g., carriers, diluents, excipients, etc. If
35 formulated as discrete units (e.g., tablets, etc.), each unit contains a predetermined amount (dosage) of the active compound.

The term “pharmaceutically acceptable,” as used herein, pertains to compounds, ingredients, materials, compositions, dosage forms, etc., which are, within the scope of sound medical judgment, suitable for use in contact with the tissues of the subject in question (e.g., human) without excessive toxicity, irritation, allergic response, or other problem or complication, commensurate with a reasonable benefit/risk ratio. Each carrier, diluent, excipient, etc. must also be “acceptable” in the sense of being compatible with the other ingredients of the formulation.

- 10 The formulations may be prepared by any methods well known in the art of pharmacy. Such methods include the step of bringing into association the active compound with a carrier which constitutes one or more accessory ingredients. In general, the formulations are prepared by uniformly and intimately bringing into association the active compound with carriers (e.g., liquid carriers, finely divided solid carrier, etc.), and then shaping the product, if necessary.

The formulation may be prepared to provide for rapid or slow release; immediate, delayed, timed, or sustained release; or a combination thereof.

- 20 Formulations suitable for parenteral administration (e.g., by injection), include aqueous or non-aqueous, isotonic, pyrogen-free, sterile liquids (e.g., solutions, suspensions), in which the active ingredient is dissolved, suspended, or otherwise provided (e.g., in a liposome or other microparticulate). Such liquids may additionally contain other pharmaceutically acceptable ingredients, such as anti-oxidants, buffers, preservatives, stabilisers, bacteriostats, suspending agents, thickening agents, and solutes which render the formulation isotonic with the blood (or other relevant bodily fluid) of the intended recipient. Examples of excipients include, for example, water, alcohols, polyols, glycerol, vegetable oils, and the like. Examples of suitable isotonic carriers for use in such formulations include Sodium Chloride Injection, Ringer's Solution, or Lactated Ringer's Injection. Typically, the concentration of the active ingredient in the liquid is from about 1 ng/ml to about 10 µg/ml, for example from about 10 ng/ml to about 1 µg/ml. The formulations may be presented in unit-dose or multi-dose sealed containers, for example, ampoules and vials, and may be stored in a freeze-dried (lyophilised) condition requiring only the addition of the sterile liquid carrier, for example water for injections, immediately prior to use. Extemporaneous injection solutions and suspensions may be prepared from sterile powders, granules, and tablets.

Dosage

It will be appreciated by one of skill in the art that appropriate dosages of the conjugate compound, and compositions comprising the conjugate compound, can vary from patient to patient. Determining the optimal dosage will generally involve the balancing of the level of therapeutic benefit against any risk or deleterious side effects. The selected dosage level will depend on a variety of factors including, but not limited to, the activity of the particular compound, the route of administration, the time of administration, the rate of excretion of the compound, the duration of the treatment, other drugs, compounds, and/or materials used in combination, the severity of the condition, and the species, sex, age, weight, condition, general health, and prior medical history of the patient. The amount of compound and route of administration will ultimately be at the discretion of the physician, veterinarian, or clinician, although generally the dosage will be selected to achieve local concentrations at the site of action which achieve the desired effect without causing substantial harmful or deleterious side-effects.

Administration can be effected in one dose, continuously or intermittently (e.g., in divided doses at appropriate intervals) throughout the course of treatment. Methods of determining the most effective means and dosage of administration are well known to those of skill in the art and will vary with the formulation used for therapy, the purpose of the therapy, the target cell(s) being treated, and the subject being treated. Single or multiple administrations can be carried out with the dose level and pattern being selected by the treating physician, veterinarian, or clinician.

In general, a suitable dose of the active compound is in the range of about 100 ng to about 25 mg (more typically about 1 µg to about 10 mg) per kilogram body weight of the subject per day. Where the active compound is a salt, an ester, an amide, a prodrug, or the like, the amount administered is calculated on the basis of the parent compound and so the actual weight to be used is increased proportionately.

In one embodiment, the active compound is administered to a human patient according to the following dosage regime: about 100 mg, 3 times daily.

In one embodiment, the active compound is administered to a human patient according to the following dosage regime: about 150 mg, 2 times daily.

In one embodiment, the active compound is administered to a human patient according to the following dosage regime: about 200 mg, 2 times daily.

However in one embodiment, the conjugate compound is administered to a human patient
5 according to the following dosage regime: about 50 or about 75 mg, 3 or 4 times daily.

In one embodiment, the conjugate compound is administered to a human patient according to the following dosage regime: about 100 or about 125 mg, 2 times daily.

10 The dosage amounts described above may apply to the conjugate (including the PBD moiety and the linker to the antibody) or to the effective amount of PBD compound provided, for example the amount of compound that is releasable after cleavage of the linker.

For the prevention or treatment of disease, the appropriate dosage of an ADC of the
15 invention will depend on the type of disease to be treated, as defined above, the severity and course of the disease, whether the molecule is administered for preventive or therapeutic purposes, previous therapy, the patient's clinical history and response to the antibody, and the discretion of the attending physician. The molecule is suitably administered to the patient at one time or over a series of treatments. Depending on the type
20 and severity of the disease, about 1 µg/kg to 15 mg/kg (e.g. 0.1-20 mg/kg) of molecule is an initial candidate dosage for administration to the patient, whether, for example, by one or more separate administrations, or by continuous infusion. A typical daily dosage might range from about 1 µg/kg to 100 mg/kg or more, depending on the factors mentioned above. An exemplary dosage of ADC to be administered to a patient is in the range of about 0.1 to
25 about 10 mg/kg of patient weight. For repeated administrations over several days or longer, depending on the condition, the treatment is sustained until a desired suppression of disease symptoms occurs. An exemplary dosing regimen comprises a course of administering an initial loading dose of about 4 mg/kg, followed by additional doses every week, two weeks, or three weeks of an ADC. Other dosage regimens may be useful. The progress of this
30 therapy is easily monitored by conventional techniques and assays.

Treatment

The term "treatment," as used herein in the context of treating a condition, pertains generally to treatment and therapy, whether of a human or an animal (e.g., in veterinary applications),
35 in which some desired therapeutic effect is achieved, for example, the inhibition of the progress of the condition, and includes a reduction in the rate of progress, a halt in the rate

of progress, regression of the condition, amelioration of the condition, and cure of the condition. Treatment as a prophylactic measure (i.e., prophylaxis, prevention) is also included.

- 5 The term “therapeutically-effective amount,” as used herein, pertains to that amount of an active compound, or a material, composition or dosage from comprising an active compound, which is effective for producing some desired therapeutic effect, commensurate with a reasonable benefit/risk ratio, when administered in accordance with a desired treatment regimen.

10

Similarly, the term “prophylactically-effective amount,” as used herein, pertains to that amount of an active compound, or a material, composition or dosage from comprising an active compound, which is effective for producing some desired prophylactic effect, commensurate with a reasonable benefit/risk ratio, when administered in accordance with a
15 desired treatment regimen.

Preparation of Drug conjugates

Antibody drug conjugates, as well as conjugates with other cell binding agents, may be prepared by several routes, employing organic chemistry reactions, conditions, and reagents
20 known to those skilled in the art, including reaction of a nucleophilic group of an antibody or cell binding agent with a drug-linker reagent. This method may be employed with a variety of antibodies and cell binding agents to prepare the antibody-drug conjugates of the invention.

- 25 Nucleophilic groups on antibodies include, but are not limited to side chain thiol groups, e.g. cysteine. Thiol groups are nucleophilic and capable of reacting to form covalent bonds with electrophilic groups on linker moieties such as those of the present invention. Certain antibodies have reducible interchain disulfides, i.e. cysteine bridges. Antibodies may be made reactive for conjugation with linker reagents by treatment with a reducing agent such
30 as DTT (Cleland's reagent, dithiothreitol) or TCEP (tris(2-carboxyethyl)phosphine hydrochloride; Getz et al (1999) Anal. Biochem. Vol 273:73-80; Soltec Ventures, Beverly, MA). Each cysteine disulfide bridge will thus form, theoretically, two reactive thiol nucleophiles. Additional nucleophilic groups can be introduced into antibodies through the reaction of lysines with 2-iminothiolane (Traut's reagent) resulting in conversion of an amine
35 into a thiol.

The Subject/Patient

The subject/patient may be an animal, mammal, a placental mammal, a marsupial (e.g., kangaroo, wombat), a monotreme (e.g., duckbilled platypus), a rodent (e.g., a guinea pig, a hamster, a rat, a mouse), murine (e.g., a mouse), a lagomorph (e.g., a rabbit), avian (e.g., a bird), canine (e.g., a dog), feline (e.g., a cat), equine (e.g., a horse), porcine (e.g., a pig), ovine (e.g., a sheep), bovine (e.g., a cow), a primate, simian (e.g., a monkey or ape), a monkey (e.g., marmoset, baboon), an ape (e.g., gorilla, chimpanzee, orangutang, gibbon), or a human.

- 10 Furthermore, the subject/patient may be any of its forms of development, for example, a fetus. In one preferred embodiment, the subject/patient is a human.

In one embodiment, the patient is a population where each patient has a tumour having $\alpha_v\beta_6$ integrin on the surface of the cell.

15

Examples**General Experimental Methods for Examples 1 and 2**

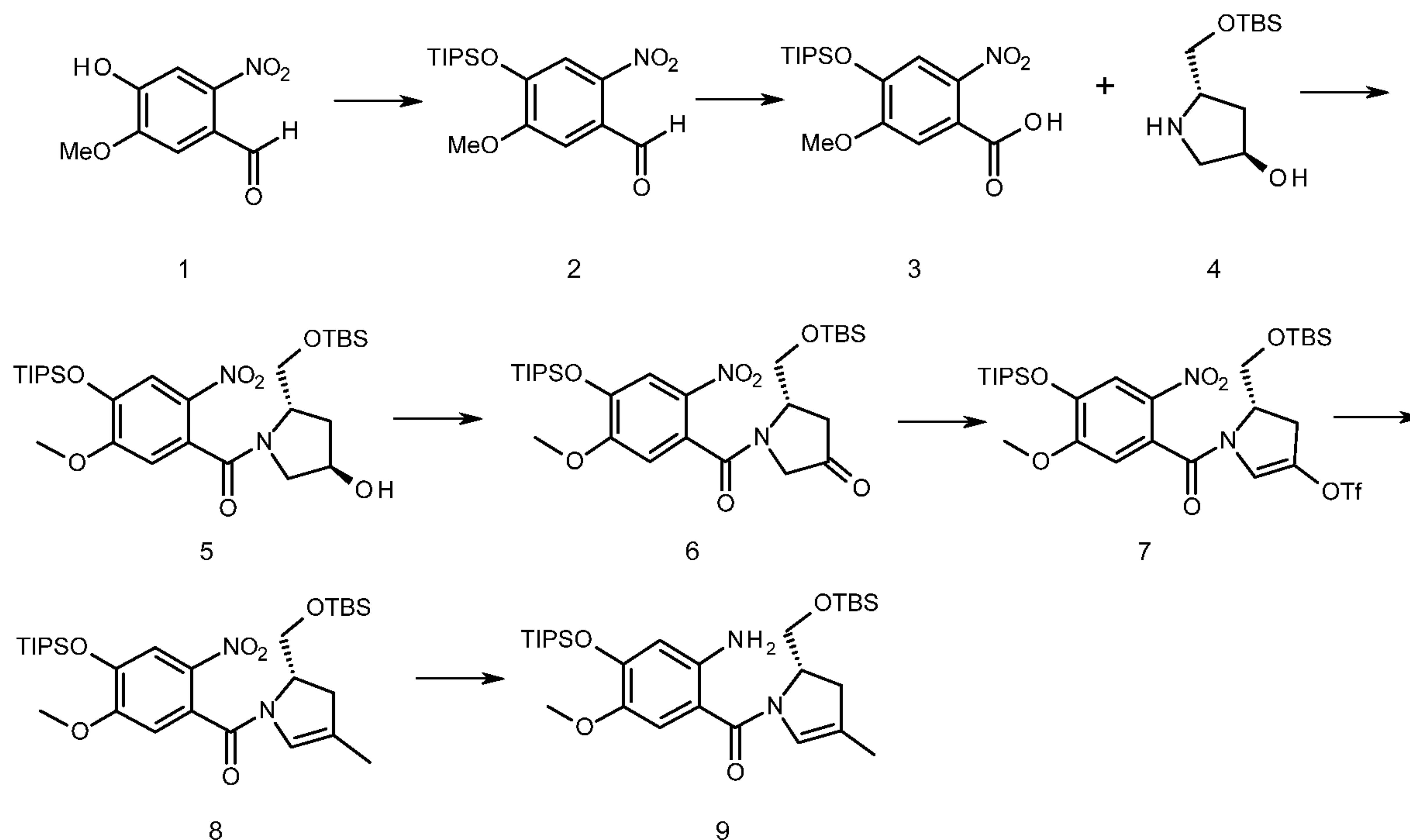
- Optical rotations were measured on an ADP 220 polarimeter (Bellingham Stanley Ltd.) and concentrations (*c*) are given in g/100mL. Melting points were measured using a digital melting point apparatus (Electrothermal). IR spectra were recorded on a Perkin-Elmer Spectrum 1000 FT IR Spectrometer. ^1H and ^{13}C NMR spectra were acquired at 300 K using a Bruker AvanceTM NMR spectrometer at 400 and 100 MHz, respectively. Chemical shifts are reported relative to TMS ($\delta = 0.0$ ppm), and signals are designated as s (singlet), d (doublet), t (triplet), dt (double triplet), dd (doublet of doublets), ddd (double doublet of doublets) or m (multiplet), with coupling constants given in Hertz (Hz). Mass spectroscopy (MS) data were collected using a Waters MicromassTM ZQ instrument coupled to a Waters 2695 HPLC with a Waters 2996 PDA. Waters Micromass ZQ parameters used were: Capillary (kV), 3.38; Cone (V), 35; Extractor (V), 3.0; Source temperature ($^{\circ}\text{C}$), 100; Desolvation Temperature ($^{\circ}\text{C}$), 200; Cone flow rate (L/h), 50; De-solvation flow rate (L/h), 250. High-resolution mass spectroscopy (HRMS) data were recorded on a Waters Micromass QTOF Global in positive W-mode using metal-coated borosilicate glass tips to introduce the samples into the instrument. Thin Layer Chromatography (TLC) was performed on silica gel aluminium plates (Merck 60, F₂₅₄), and flash chromatography utilised silica gel (Merck 60, 230-400 mesh ASTM). Except for the HOBt (NovaBiochem) and solid-supported reagents (Argonaut), all other chemicals and solvents were purchased from Sigma-Aldrich and were used as supplied without further purification. Anhydrous solvents

were prepared by distillation under a dry nitrogen atmosphere in the presence of an appropriate drying agent, and were stored over 4Å molecular sieves or sodium wire. Petroleum ether refers to the fraction boiling at 40-60°C.

- 5 General LC/MS conditions: The HPLC (Waters Alliance™ 2695) was run using a mobile phase of water (A) (formic acid 0.1%) and acetonitrile (B) (formic acid 0.1%). Gradient: initial composition 5% B over 1.0 min then 5% B to 95% B over 2.5 min. The composition was held for 0.5 min at 95% B, and then returned to 5% B in 0.1 minutes and held there for 0.9 min. Total gradient run time equals 5 min. Flow rate 3.0 mL/min, 400µL was split *via* a zero
- 10 dead volume tee piece which passes into the mass spectrometer. Wavelength detection range: 220 to 400 nm. Function type: diode array (535 scans). Column: Phenomenex® Onyx Monolithic C18 50 x 4.60 mm.

Example 1

(i) (S)-(2-amino-5-methoxy-4-((triisopropylsilyl)oxy)phenyl)(2-(((tert-butyl)dimethylsilyl)oxy)methyl)-4-methyl-2,3-dihydro-1H-pyrrol-1-yl)methanone (9)



5 (a) *5-methoxy-2-nitro-4-((triisopropylsilyl)oxy)benzaldehyde (2)*

Neat triisopropylsilylchloride (56.4 mL, 262 mmol) was added to a mixture of imidazole (48.7 g, 715.23 mmol) and 4-hydroxy-5-methoxy-2-nitrobenzaldehyde **1** (47 g, 238 mmol) (ground together). The mixture was heated until the phenol and imidazole melted and went into solution (100 °C). The reaction mixture was allowed to stir for 15 minutes and was then
 10 allowed to cool, whereupon a solid was observed to form at the bottom of the flask (imidazole chloride). The reaction mixture was diluted with 5% EtOAc/ hexanes and loaded directly onto silica gel and the pad was eluted with 5% EtOAc/ hexanes , followed by 10% EtOAc/hexanes (due to the low excess, very little unreacted TIPSCl was found in the product). The desired product was eluted with 5 % ethyl acetate in hexane. Excess eluent
 15 was removed by rotary evaporation under reduced pressure, followed by drying under high vacuum to afford a crystalline light sensitive solid (74.4 g, 88 %). Purity satisfactory by LC/MS (4.22 min (ES+) *m/z* (relative intensity) 353.88 ($[M + H]^+$, 100)); ^1H NMR (400 MHz, CDCl_3) δ 10.43 (s, 1H), 7.60 (s, 1H), 7.40 (s, 1H), 3.96 (s, 3H), 1.35 – 1.24 (m, 3H), 1.10 (m, 18H).

(b) 5-methoxy-2-nitro-4-((triisopropylsilyl)oxy)benzoic acid (3)

A solution of sodium chlorite (47.3 g, 523 mmol, 80 % technical grade) and sodium dihydrogenphosphate monobasic (35.2 g, 293 mmol) (NaH_2PO_4) in water (800 mL) was added to a solution of compound **2** (74 g, 209 mmol) in tetrahydrofuran (500 mL) at room temperature. Hydrogen peroxide (60 % w/w, 140 mL, 2.93 mol) was immediately added to the vigorously stirred biphasic mixture. The reaction mixture evolved gas (oxygen), the starting material dissolved and the temperature of the reaction mixture rose to 45°C. After 30 minutes LC/MS revealed that the reaction was complete. The reaction mixture was cooled in an ice bath and hydrochloric acid (1 M) was added to lower the pH to 3 (this step was found unnecessary in many instances, as the pH at the end of the reaction is already acidic; please check the pH before extraction). The reaction mixture was then extracted with ethyl acetate (1 L) and the organic phases washed with brine (2 x 100 mL) and dried over magnesium sulphate. The organic phase was filtered and excess solvent removed by rotary evaporation under reduced pressure to afford the product **6** in quantitative yield as a yellow solid. LC/MS (3.93 min (ES-) m/z (relative intensity) 367.74 ($[M - H]^-$, 100)); ^1H NMR (400 MHz, CDCl_3) δ 7.36 (s, 1H), 7.24 (s, 1H), 3.93 (s, 3H), 1.34 – 1.22 (m, 3H), 1.10 (m, 18H).

(c) ((2S,4R)-2-(((tert-butyldimethylsilyl)oxy)methyl)-4-hydroxypyrrolidin-1-yl)(5-methoxy-2-nitro-4-((triisopropylsilyl)oxy)phenyl)methanone (5)

DCC (29.2 g, 141 mmol, 1.2 eq) was added to a solution of acid **3** (43.5 g, 117.8 mmol, 1eq), and hydroxybenzotriazole hydrate (19.8 g, 129.6 mmol, 1.1 eq) in dichloromethane (200 mL) at 0 °C. The cold bath was removed and the reaction was allowed to proceed for 30 mins at room temperature, at which time a solution of (2S,4R)-2-*t*-butyldimethylsilyloxymethyl-4-hydroxypyrrolidine **4** (30 g, 129.6 mmol, 1.1 eq) and triethylamine (24.66 mL, 176 mmol, 1.5 eq) in dichloromethane (100 mL) was added rapidly at -10 °C under argon (on large scale, the addition time could be shortened by cooling the reaction mixture even further. The reaction mixture was allowed to stir at room temperature for 40 minutes to 1 hour and monitored by LC/MS and TLC (EtOAc). The solids were removed by filtration over CeliteTM and the organic phase was washed with cold aqueous 0.1 M HCl until the pH was measured at 4 or 5. The organic phase was then washed with water, followed by saturated aqueous sodium bicarbonate and brine. The organic layer was dried over magnesium sulphate, filtered and excess solvent removed by rotary evaporation under reduced pressure. The residue was subjected to column flash chromatography (silica gel; gradient 40/60 ethyl acetate/hexane to 80/20 ethyl acetate/ hexane). Excess solvent was removed by rotary evaporation under reduced pressure afforded the pure product **13**, (45.5 g of pure product 66%, and 17 g of slightly impure product, 90% in total). LC/MS 4.43 min (ES+) m/z (relative

intensity) 582.92 ($[M + H]^+$, 100); ^1H NMR (400 MHz, CDCl_3) δ 7.66 (s, 1H), 6.74 (s, 1H), 4.54 (s, 1H), 4.40 (s, 1H), 4.13 (s, 1H), 3.86 (s, 3H), 3.77 (d, $J = 9.2$ Hz, 1H), 3.36 (dd, $J = 11.3, 4.5$ Hz, 1H), 3.14 – 3.02 (m, 1H), 2.38 – 2.28 (m, 1H), 2.10 (ddd, $J = 13.3, 8.4, 2.2$ Hz, 1H), 1.36 – 1.19 (m, 3H), 1.15 – 1.05 (m, 18H), 0.91 (s, 9H), 0.17 – 0.05 (m, 6H), (presence of rotamers).

(d) (S)-5-(((*tert*-butyldimethylsilyl)oxy)methyl)-1-(5-methoxy-2-nitro-4-((*triisopropylsilyl*)oxy)benzoyl)pyrrolidin-3-one (**6**)

TCCA (8.82 g, 40 mmol, 0.7 eq) was added to a stirred solution of **5** (31.7 g, 54 mmol, 1 eq) and TEMPO (0.85 g, 5.4 mmol, 0.1 eq) in dry dichloromethane (250 mL) at 0 °C. The reaction mixture was vigorously stirred for 20 minutes, at which point TLC (50/50 ethyl acetate/hexane) revealed complete consumption of the starting material. The reaction mixture was filtered through Celite and the filtrate washed with aqueous saturated sodium bicarbonate (100 mL), sodium thiosulphate (9 g in 300 mL), brine (100 mL) and dried over magnesium sulphate. Rotary evaporation under reduced pressure afforded product **6** in quantitative yield. LC/MS 4.52 min (ES+) m/z (relative intensity) 581.08 ($[M + H]^+$, 100); ^1H NMR (400 MHz, CDCl_3) δ 7.78 – 7.60 (m, 1H), 6.85 – 6.62 (m, 1H), 4.94 (dd, $J = 30.8, 7.8$ Hz, 1H), 4.50 – 4.16 (m, 1H), 3.99 – 3.82 (m, 3H), 3.80 – 3.34 (m, 3H), 2.92 – 2.17 (m, 2H), 1.40 – 1.18 (m, 3H), 1.11 (t, $J = 6.2$ Hz, 18H), 0.97 – 0.75 (m, 9H), 0.15 – -0.06 (m, 6H), (presence of rotamers).

(e) (S)-5-(((*tert*-butyldimethylsilyl)oxy)methyl)-1-(5-methoxy-2-nitro-4-((*triisopropylsilyl*)oxy)benzoyl)-4,5-dihydro-1H-pyrrol-3-yl trifluoromethanesulfonate (**7**)

Triflic anhydride (27.7 mL, 46.4 g, 165 mmol, 3 eq) was injected (temperature controlled) to a vigorously stirred suspension of ketone **6** (31.9 g, 55 mmol, 1 eq) in dry dichloromethane (900 mL) in the presence of 2,6-lutidine (25.6 mL, 23.5 g, 220 mmol, 4 eq, dried over sieves) at -50 °C (acetone/dry ice bath). The reaction mixture was allowed to stir for 1.5 hours when LC/MS, following a mini work-up (water/dichloromethane), revealed the reaction to be complete. Water was added to the still cold reaction mixture and the organic layer was separated and washed with saturated sodium bicarbonate, brine and magnesium sulphate. The organic phase was filtered and excess solvent was removed by rotary evaporation under reduced pressure. The residue was subjected to column flash chromatography (silica gel; 10/90 v/v ethyl acetate/hexane), removal of excess eluent afforded the product **7** (37.6 g, 96 %) LC/MS, method 2, 4.32 min (ES+) m/z (relative intensity) 712.89 ($[M + H]^+$, 100); ^1H NMR (400 MHz, CDCl_3) δ 7.71 (s, 1H), 6.75 (s, 1H), 6.05 (d, $J = 1.8$ Hz, 1H), 4.78 (dd, $J = 9.8, 5.5$ Hz, 1H), 4.15 – 3.75 (m, 5H), 3.17 (ddd, $J = 16.2, 10.4, 2.3$ Hz, 1H), 2.99 (ddd, $J =$

16.3, 4.0, 1.6 Hz, 1H), 1.45 – 1.19 (m, 3H), 1.15 – 1.08 (m, 18H), 1.05 (s, 6H), 0.95 – 0.87 (m, 9H), 0.15 – 0.08 (m, 6H).

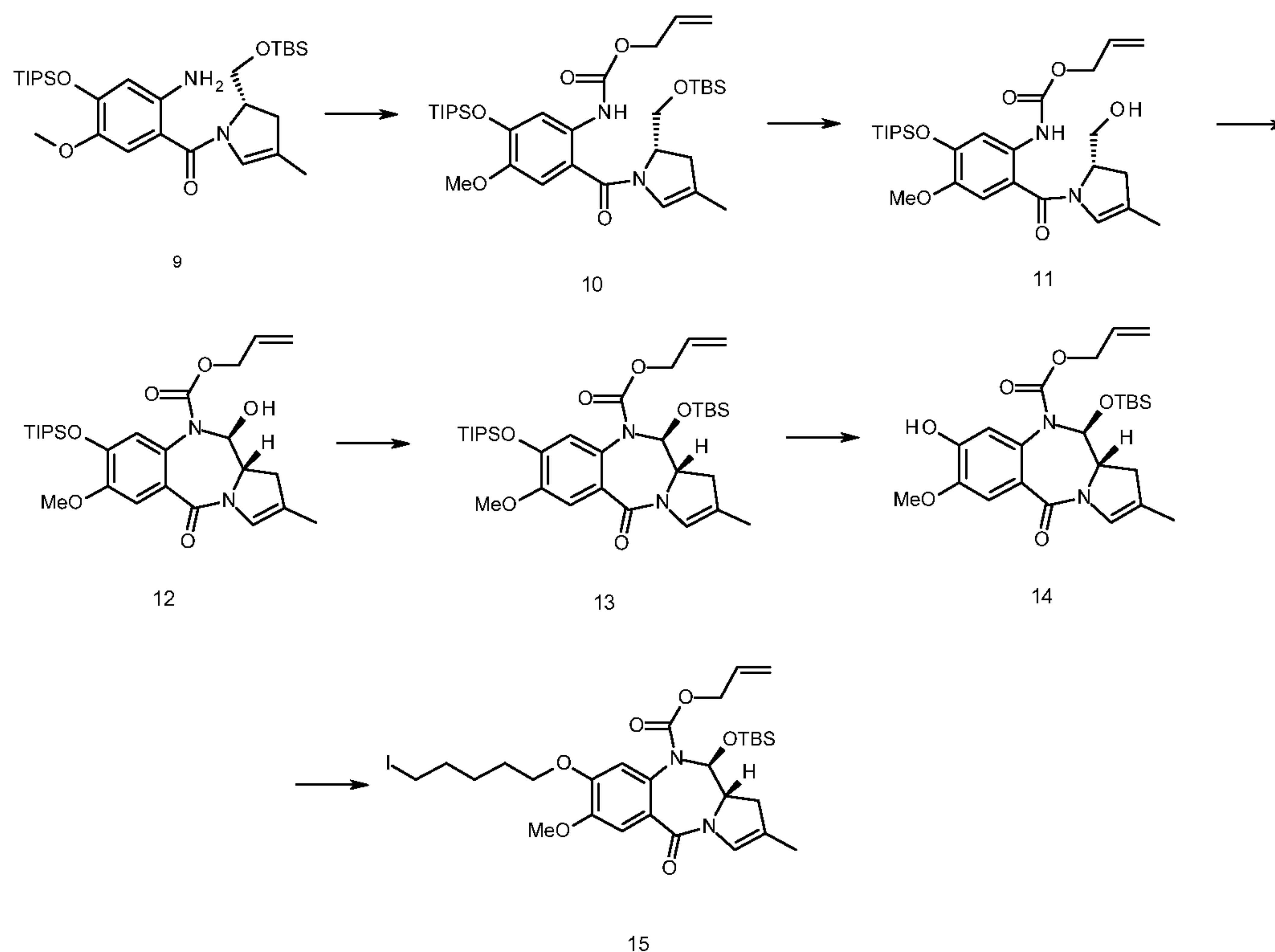
(f) (S)-(2-(((tert-butyldimethylsilyl)oxy)methyl)-4-methyl-2,3-dihydro-1H-pyrrol-1-yl)(5-methoxy-2-nitro-4-((triisopropylsilyl)oxy)phenyl)methanone (**8**)

Triphenylarsine (1.71 g, 5.60 mmol, 0.4 eq) was added to a mixture of triflate **7** (10.00 g, 14 mmol, 1eq), methylboronic acid (2.94 g, 49.1 mmol, 3.5 eq), silver oxide (13 g, 56 mmol, 4 eq) and potassium phosphate tribasic (17.8 g, 84 mmol, 6 eq) in dry dioxane (80 mL) under an argon atmosphere. The reaction was flushed with argon 3 times and
 10 bis(benzonitrile)palladium(II) chloride (540 mg, 1.40 mmol, 0.1 eq) was added. The reaction was flushed with argon 3 more times before being warmed instantaneously to 110°C (the driesyn heating block was previously warmed to 110°C prior addition of the flask). After 10 mins the reaction was cooled to room temperature and filtered through a pad Celite. The solvent was removed by rotary evaporation under reduced pressure. The resulting residue
 15 was subjected to column flash chromatography (silica gel; 10 % ethyl acetate / hexane). Pure fractions were collected and combined, and excess eluent was removed by rotary evaporation under reduced pressure afforded the product **8** (4.5 g, 55 %). LC/MS, 4.27 min (ES+) *m/z* (relative intensity) 579.18 ($[M + H]^+$, 100); ^1H NMR (400 MHz, CDCl_3) δ 7.70 (s, 1H), 6.77 (s, 1H), 5.51 (d, $J = 1.7$ Hz, 1H), 4.77 – 4.59 (m, 1H), 3.89 (s, 3H), 2.92 – 2.65 (m, 1H), 2.55 (d, $J = 14.8$ Hz, 1H), 1.62 (d, $J = 1.1$ Hz, 3H), 1.40 – 1.18 (m, 3H), 1.11 (s, 9H),
 20 1.10 (s, 9H), 0.90 (s, 9H), 0.11 (d, $J = 2.3$ Hz, 6H).

(g) (S)-(2-amino-5-methoxy-4-((triisopropylsilyl)oxy)phenyl)(2-(((tert-butyldimethylsilyl)oxy)methyl)-4-methyl-2,3-dihydro-1H-pyrrol-1-yl)methanone (**9**)
 25 Zinc powder (28 g, 430 mmol, 37 eq) was added to a solution of compound **8** (6.7 g, 11.58 mmol) in 5% formic acid in ethanol v/v (70 mL) at around 15°C. The resulting exotherm was controlled using an ice bath to maintain the temperature of the reaction mixture below 30°C. After 30 minutes the reaction mixture was filtered through a pad of Celite. The filtrate was diluted with ethyl acetate and the organic phase was washed with water, saturated aqueous
 30 sodium bicarbonate and brine. The organic phase was dried over magnesium sulphate, filtered and excess solvent removed by rotary evaporation under reduced pressure. The resulting residue was subjected to flash column chromatography (silica gel; 10 % ethyl acetate in hexane). The pure fractions were collected and combined and excess solvent was removed by rotary evaporation under reduced pressure to afford the product **9** (5.1 g, 80 %).
 35 LC/MS, 4.23 min (ES+) *m/z* (relative intensity) 550.21 ($[M + H]^+$, 100); ^1H NMR (400 MHz, CDCl_3) δ 7.28 (s, 1H), 6.67 (s, 1H), 6.19 (s, 1H), 4.64 – 4.53 (m, $J = 4.1$ Hz, 1H), 4.17 (s,

1H), 3.87 (s, 1H), 3.77 – 3.69 (m, 1H), 3.66 (s, 3H), 2.71 – 2.60 (m, 1H), 2.53 – 2.43 (m, 1H), 2.04 – 1.97 (m, $J = 11.9$ Hz, 1H), 1.62 (s, 3H), 1.26 – 1.13 (m, 3H), 1.08 – 0.99 (m, 18H), 0.82 (s, 9H), 0.03 – -0.03 (m, $J = 6.2$ Hz, 6H).

5 (ii) (11S,11aS)-allyl 11-((*tert*-butyldimethylsilyl)oxy)-8-((5-iodopentyl)oxy)-7-methoxy-2-methyl-5-oxo-11,11a-dihydro-1H-benzo[e]pyrrolo[1,2-a][1,4]diazepine-10(5H)-carboxylate



(a) (*S*)-allyl (2-(2-(((*tert*-butyldimethylsilyl)oxy)methyl)-4-methyl-2,3-dihydro-1H-pyrrole-1-carbonyl)-4-methoxy-5-((*triisopropylsilyl*)oxy)phenyl)carbamate (**10**)

- 10 Allyl chloroformate (0.30 mL, 3.00 mmol, 1.1 eq) was added to a solution of amine **9** (1.5 g, 2.73 mmol) in the presence of dry pyridine (0.48 mL, 6.00 mmol, 2.2 eq) in dry dichloromethane (20 mL) at -78°C (acetone/dry ice bath). After 30 minutes, the bath was removed and the reaction mixture was allowed to warm to room temperature. The reaction mixture was diluted with dichloromethane and saturated aqueous copper sulphate was
- 15 added. The organic layer was then washed sequentially with saturated aqueous sodium bicarbonate and brine. The organic phase was dried over magnesium sulphate, filtered and excess solvent removed by rotary evaporation under reduced pressure to afford the product **10** which was used directly in the next reaction. LC/MS, 4.45 min (ES+) m/z (relative intensity) 632.91 ($[M + H]^+$, 100)

(b) *(S)*-allyl (2-(2-(hydroxymethyl)-4-methyl-2,3-dihydro-1*H*-pyrrole-1-carbonyl)-4-methoxy-5-((triisopropylsilyl)oxy)phenyl)carbamate (**11**)

The crude **10** was dissolved in a 7:1:1:2 mixture of acetic acid/methanol/tetrahydrofuran/water (28:4:4:8 mL) and allowed to stir at room temperature.

- 5 After 3 hours, complete disappearance of starting material was observed by LC/MS. The reaction mixture was diluted with ethyl acetate and washed sequentially with water (2 x 500 mL), saturated aqueous sodium bicarbonate (200 mL) and brine. The organic phase was dried over magnesium sulphate filtered and excess ethyl acetate removed by rotary
- 10 chromatography (silica gel, 25% ethyl acetate in hexane). Pure fractions were collected and combined and excess eluent was removed by rotary evaporation under reduced pressure to afford the desired product **11** (1 g, 71 %). LC/MS, 3.70 min (ES+) *m/z* (relative intensity) 519.13 ($[M + H]^+$, 95); ^1H NMR (400 MHz, CDCl_3) δ 8.34 (s, 1H), 7.69 (s, 1H), 6.78 (s, 1H), 6.15 (s, 1H), 5.95 (ddt, $J = 17.2, 10.5, 5.7$ Hz, 1H), 5.33 (dq, $J = 17.2, 1.5$ Hz, 1H), 5.23 (ddd,
- 15 $J = 10.4, 2.6, 1.3$ Hz, 1H), 4.73 (tt, $J = 7.8, 4.8$ Hz, 1H), 4.63 (dt, $J = 5.7, 1.4$ Hz, 2H), 4.54 (s, 1H), 3.89 – 3.70 (m, 5H), 2.87 (dd, $J = 16.5, 10.5$ Hz, 1H), 2.19 (dd, $J = 16.8, 4.6$ Hz, 1H), 1.70 (d, $J = 1.3$ Hz, 3H), 1.38 – 1.23 (m, 3H), 1.12 (s, 10H), 1.10 (s, 8H).

(c) *(11S,11aS)*-allyl 11-hydroxy-7-methoxy-2-methyl-5-oxo-8-((triisopropylsilyl)oxy)-11,11a-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**12**)

- Dimethyl sulphoxide (0.35 mL, 4.83 mmol, 2.5 eq) was added dropwise to a solution of oxalyl chloride (0.2 mL, 2.32 mmol, 1.2 eq) in dry dichloromethane (10 mL) at -78°C (dry ice /acetone bath) under an atmosphere of argon. After 10 minutes a solution of **11** (1 g, 1.93 mmol) in dry dichloromethane (8 mL) was added slowly with the temperature still at -78°C .
- 25 After 15 min triethylamine (1.35 mL, dried over 4Å molecular sieves, 9.65 mmol, 5 eq) was added dropwise and the dry ice/acetone bath was removed. The reaction mixture was allowed to reach room temperature and was extracted with cold hydrochloric acid (0.1 M), saturated aqueous sodium bicarbonate and brine. The organic phase was dried over magnesium sulphate, filtered and excess dichloromethane was removed by rotary
- 30 evaporation under reduced pressure to afford product **12** (658 mg, 66%).

Larger scale synthesis

- Dimethyl sulphoxide (3.49 mL, 49.1 mmol, 2.5 eq) was added dropwise to a solution of oxalyl chloride (2 mL, 23.5 mmol, 1.2 eq) in dry dichloromethane (80 mL) at -78°C (dry ice
- 35 /acetone bath) under an atmosphere of argon. After 15 minutes a solution of **11** (10.2 g, 19.66 mmol) in dry dichloromethane (100 mL) was added slowly with the temperature still at

-78°C. After 20 minutes triethylamine (13.7 mL, dried over 4Å molecular sieves, 98.3 mmol, 5 eq) was added dropwise and the dry ice/acetone bath was removed. The reaction mixture was allowed to reach room temperature for 2 hours and was extracted with cold hydrochloric acid (0.1 M, 400 mL), saturated aqueous sodium bicarbonate (300 mL) and brine (400 mL).

- 5 The organic phase was dried over magnesium sulphate, filtered and excess dichloromethane was removed by rotary evaporation under reduced pressure to afford product **12** as crude.

10 LC/MS, 3.52 min (ES+) *m/z* (relative intensity) 517.14 ($[M + H]^+$, 100); ^1H NMR (400 MHz, CDCl_3) δ 7.20 (s, 1H), 6.75 – 6.63 (m, $J = 8.8, 4.0$ Hz, 2H), 5.89 – 5.64 (m, $J = 9.6, 4.1$ Hz, 2H), 5.23 – 5.03 (m, 2H), 4.68 – 4.38 (m, 2H), 3.84 (s, 3H), 3.83 – 3.77 (m, 1H), 3.40 (s, 1H), 3.05 – 2.83 (m, 1H), 2.59 (d, $J = 17.1$ Hz, 1H), 1.78 (d, $J = 1.3$ Hz, 3H), 1.33 – 1.16 (m, 3H), 1.09 (d, $J = 2.2$ Hz, 9H), 1.07 (d, $J = 2.1$ Hz, 9H).

- 15 (d) (11*S*,11*aS*)-allyl 11-((*tert*-butyldimethylsilyl)oxy)-7-methoxy-2-methyl-5-oxo-8-((*triisopropylsilyl*)oxy)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**13**)

20 *Tert*-butyldimethylsilyltriflate (0.70 mL, 3.00 mmol, 3 eq) was added to a solution of compound **12** (520 mg, 1.00 mmol) and 2,6-lutidine (0.46 mL, 4.00 mmol, 4 eq) in dry dichloromethane (40 mL) at 0°C under argon. After 10 min, the cold bath was removed and the reaction mixture was stirred at room temperature for 1 hour. The reaction mixture was extracted with water, saturated aqueous sodium bicarbonate and brine. The organic phase was dried over magnesium sulphate, filtered and excess was removed by rotary evaporation under reduced pressure. The resulting residue was subjected to flash column

25 chromatography (silica gel; gradient, 10 % ethyl acetate in hexane to 20 % ethyl acetate in hexane). Pure fractions were collected and combined and excess eluent was removed by rotary evaporation under reduced pressure to give the product **13** (540 mg, 85 %).

Large scale synthesis

30 *Tert*-butyldimethylsilyltriflate (12.7 mL, 55.3 mmol, 3 eq) was added to a solution of compound **12** (9.40 g, 18.46 mmol) and 2,6-lutidine (8.61 mL, 72.2 mmol, 4 eq) in dry dichloromethane (180 mL) at 0°C under argon. After 10 minutes, the cold bath was removed and the reaction mixture was stirred at room temperature for 1 hour. The reaction mixture was extracted with citric acid (200 mL), saturated aqueous sodium bicarbonate (200 mL) and

35 brine (200 mL). The organic phase was dried over magnesium sulphate, filtered and excess was removed by rotary evaporation under reduced pressure. The resulting residue was

subjected to column chromatography (biotage 340 g with 34 g sample). Pure fractions were collected and combined and excess eluent was removed by rotary evaporation under reduced pressure to give the product **13** (7.00 g, 56 % over 2 steps).

5 LC/MS, 4.42 min (ES+) m/z (relative intensity) 653.14 ($[M + Na]^+$, 100); ^1H NMR (400 MHz, $CDCl_3$) δ 7.20 (s, 1H), 6.71 – 6.64 (m, $J = 5.5$ Hz, 2H), 5.83 (d, $J = 9.0$ Hz, 1H), 5.80 – 5.68 (m, $J = 5.9$ Hz, 1H), 5.14 – 5.06 (m, 2H), 4.58 (dd, $J = 13.2, 5.2$ Hz, 1H), 4.36 (dd, $J = 13.3, 5.5$ Hz, 1H), 3.84 (s, 3H), 3.71 (td, $J = 10.1, 3.8$ Hz, 1H), 2.91 (dd, $J = 16.9, 10.3$ Hz, 1H),
10 2.36 (d, $J = 16.8$ Hz, 1H), 1.75 (s, 3H), 1.31 – 1.16 (m, 3H), 1.12 – 1.01 (m, $J = 7.4, 2.1$ Hz, 18H), 0.89 – 0.81 (m, 9H), 0.25 (s, 3H), 0.19 (s, 3H).

(e) (11*S*,11*aS*)-allyl 11-((*tert*-butyldimethylsilyl)oxy)-8-hydroxy-7-methoxy-2-methyl-5-oxo-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**14**)

Lithium acetate (87 mg, 0.85 mmol) was added to a solution of compound **13** (540 mg, 0.85
15 mmol) in wet dimethylformamide (6 mL, 50:1 DMF/water). After 4 hours, the reaction was complete and the reaction mixture was diluted with ethyl acetate (25 mL) and washed with aqueous citric acid solution (pH ~ 3), water and brine. The organic layer was dried over magnesium sulphate filtered and excess ethyl acetate was removed by rotary evaporation under reduced pressure. The resulting residue was subjected to flash column
20 chromatography (silica gel; gradient, 25% to 75% ethyl acetate in hexane). Pure fractions were collected and combined and excess eluent was removed by rotary evaporation under reduced pressure to give the product **14** (400 mg, quantitative).

Larger scale synthesis

25 Lithium acetate (1.13 g, 11.09 mmol) was added to a solution of compound **13** (7 g, 11.09 mmol) in wet dimethylformamide (80 mL, 50:1 DMF/water). After 4 hours, the reaction was complete and the reaction mixture was concentrated to remove majority of DMF. The reaction mixture was then diluted with ethyl acetate (200 mL) and washed with aqueous citric acid solution (pH ~ 3, 0.1 M, 100 mL), water (200 mL) and brine (100 mL). The organic
30 layer was dried over magnesium sulphate filtered and excess ethyl acetate was removed by rotary evaporation under reduced pressure to afford product **14** as crude.

LC/MS, (3.33 min (ES+) m/z (relative intensity) 475.26 ($[M+H]^+$, 100).

(f) (11*S*,11*aS*)-allyl 11-((*tert*-butyldimethylsilyl)oxy)-8-((5-iodopentyl)oxy)-7-methoxy-2-methyl-5-oxo-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**15**)

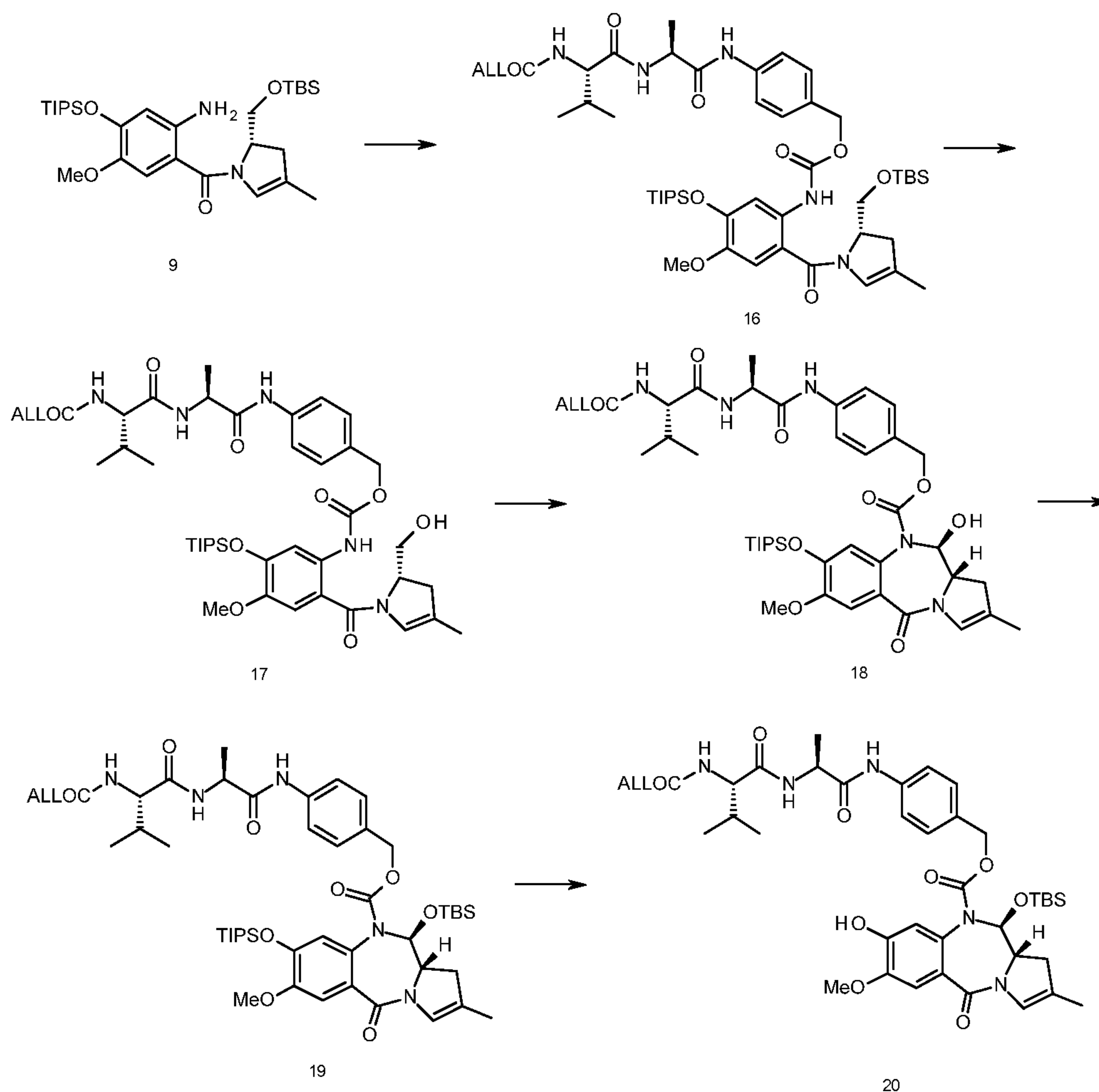
Diiodopentane (0.63 mL, 4.21 mmol, 5 eq) and potassium carbonate (116 mg, 0.84 mmol, 1 eq) were added to a solution of phenol **14** (400 mg, 0.84 mmol) in acetone (4 mL, dried over molecular sieves). The reaction mixture was then warmed to 60°C and stirred for 6 hours. Acetone was removed by rotary evaporation under reduced pressure. The resulting residue was subjected to flash column chromatography (silica gel; 50/50, v/v, hexane/ethyl acetate,). Pure fractions were collected and combined and excess eluent was removed to provide **15** in 90% yield.

Larger scale synthesis

Diiodopentane (8.26 mL, 55.46 mmol, 5 eq) and potassium carbonate (1.5 g, 11.09 mmol, 1 eq) were added to a solution of phenol **14** (5.26 g, 11.09 mmol) in 2-butanone (50 mL, dried over molecular sieves). The reaction mixture was then warmed to 70°C and stirred for 5 hours. 2-butanone was removed by rotary evaporation under reduced pressure. The resulting residue was dissolved in ethyl acetate (250 mL) and washed with water (250 mL) and brine (250 mL). The organic layer was dried over magnesium sulphate filtered and excess ethyl acetate was removed by rotary evaporation under reduced pressure. The residue was subjected to flash column chromatography (Biotage™ 750 g with 80 g silica for dry loading). Pure fractions were collected and combined and excess eluent was removed to provide **15** (5.75 g, 77% over 2 steps).

LC/MS, 3.90 min (ES+) *m/z* (relative intensity) 670.91 ($[M]^+$, 100). ¹H NMR (400 MHz, CDCl₃) δ 7.23 (s, 1H), 6.69 (s, 1H), 6.60 (s, 1H), 5.87 (d, *J* = 8.8 Hz, 1H), 5.83 – 5.68 (m, *J* = 5.6 Hz, 1H), 5.15 – 5.01 (m, 2H), 4.67 – 4.58 (m, 1H), 4.45 – 4.35 (m, 1H), 4.04 – 3.93 (m, 2H), 3.91 (s, 3H), 3.73 (td, *J* = 10.0, 3.8 Hz, 1H), 3.25 – 3.14 (m, *J* = 8.5, 7.0 Hz, 2H), 2.92 (dd, *J* = 16.8, 10.3 Hz, 1H), 2.38 (d, *J* = 16.8 Hz, 1H), 1.95 – 1.81 (m, 4H), 1.77 (s, 3H), 1.64 – 1.49 (m, 2H), 0.88 (s, 9H), 0.25 (s, 3H), 0.23 (s, 3H).

(iii) (11S,11aS)-4-(2-(1-((1-(allyloxy)-4-methyl-1,2-dioxopentan-3-yl)amino)-1-oxopropan-2-yl)hydrazinyl)benzyl 11-((tert-butyldimethylsilyl)oxy)-8-hydroxy-7-methoxy-2-methyl-5-oxo-11,11a-dihydro-1H-benzo[e]pyrrolo[1,2-a][1,4]diazepine-10(5H)-carboxylate (20)



- 5 (a) *Allyl 3-(2-(2-(4-(((2-((S)-2-((tert-butyldimethylsilyl)oxy)methyl)-4-methyl-2,3-dihydro-1H-pyrrole-1-carbonyl)-4-methoxy-5-((triisopropylsilyl)oxy)phenyl)carbamoyl)oxy)methyl)phenyl)hydrazinyl)propanamido)-4-methyl-2-oxopentanoate (16)*

Triethylamine (2.23 mL, 18.04 mmol, 2.2 eq) was added to a stirred solution of the amine **9** (4 g, 8.20 mmol) and triphosgene (778 mg, 2.95 mmol, 0.36 eq) in dry tetrahydrofuran (40 mL) at 5 °C (ice bath). The progress of the isocyanate reaction was monitored by periodically removing aliquots from the reaction mixture and quenching with methanol and performing LC/MS analysis. Once the isocyanate formation was complete a solution of the alloc-Val-Ala-PABOH (4.12 g, 12.30 mmol, 1.5 eq) and triethylamine (1.52 mL, 12.30 mmol, 1.5 eq) in dry tetrahydrofuran (40 mL) was rapidly added by injection to the freshly prepared isocyanate. The reaction mixture was allowed to stir at 40 °C for 4 hours. Excess solvent was removed by rotary evaporation under reduced pressure. The resulting residue was

subjected to flash column chromatography (silica gel; gradient, 1 % methanol to 5% methanol in dichloromethane). (Alternative chromatography conditions using EtOAc and Hexane have also been successful). Pure fractions were collected and combined and excess eluent was removed by rotary evaporation under reduced pressure to give the product **16** (3.9 g, 50%). LC/MS, 4.23 min (ES+) m/z (relative intensity) 952.36 ($[M + H]^+$, 100); ^1H NMR (400 MHz, CDCl_3) δ 8.62 (br s, 1H), 8.46 (s, 1H), 7.77 (br s, 1H), 7.53 (d, $J = 8.4$ Hz, 2H), 7.32 (d, $J = 8.5$ Hz, 2H), 6.76 (s, 1H), 6.57 (d, $J = 7.6$ Hz, 1H), 6.17 (s, 1H), 6.03 – 5.83 (m, 1H), 5.26 (dd, $J = 33.8, 13.5$ Hz, 3H), 5.10 (s, 2H), 4.70 – 4.60 (m, 2H), 4.58 (dd, $J = 5.7, 1.3$ Hz, 2H), 4.06 – 3.99 (m, 1H), 3.92 (s, 1H), 3.82 – 3.71 (m, 1H), 3.75 (s, 3H), 2.79 – 2.64 (m, 1H), 2.54 (d, $J = 12.9$ Hz, 1H), 2.16 (dq, $J = 13.5, 6.7$ Hz, 1H), 1.67 (s, 3H), 1.46 (d, $J = 7.0$ Hz, 3H), 1.35 – 1.24 (m, 3H), 1.12 (s, 9H), 1.10 (s, 9H), 0.97 (d, $J = 6.8$ Hz, 3H), 0.94 (d, $J = 6.8$ Hz, 3H), 0.87 (s, 9H), 0.07 – -0.02 (m, 6H).

(b) *Allyl 3-(2-(2-(4-(((2-((S)-2-(hydroxymethyl)-4-methyl-2,3-dihydro-1H-pyrrole-1-carbonyl)-4-methoxy-5-((triisopropylsilyl)oxy)phenyl)carbamoyl)oxy)methyl)phenyl)hydrazinyl)propanamido)-4-methyl-2-oxopentanoate (17)*

The TBS ether **16** (1.32 g, 1.38 mmol) was dissolved in a 7:1:1:2 mixture of acetic acid/methanol/tetrahydrofuran/water (14:2:2:4 mL) and allowed to stir at room temperature. After 3 hours no more starting material was observed by LC/MS. The reaction mixture was diluted with ethyl acetate (25 mL) and washed sequentially with water, saturated aqueous sodium bicarbonate and brine. The organic phase was dried over magnesium sulphate filtered and excess ethyl acetate removed by rotary evaporation under reduced pressure. The resulting residue was subjected to flash column chromatography (silica gel, 2% methanol in dichloromethane). Pure fractions were collected and combined and excess eluent was removed by rotary evaporation under reduced pressure to afford the desired product **17** (920 mg, 80%). LC/MS, 3.60 min (ES+) m/z (relative intensity) 838.18 ($[M+H]^+$, 100). ^1H NMR (400 MHz, CDCl_3) δ 8.55 (s, 1H), 8.35 (s, 1H), 7.68 (s, 1H), 7.52 (d, $J = 8.1$ Hz, 2H), 7.31 (d, $J = 8.4$ Hz, 2H), 6.77 (s, 1H), 6.71 (d, $J = 7.5$ Hz, 1H), 6.13 (s, 1H), 5.97 – 5.82 (m, $J = 5.7$ Hz, 1H), 5.41 – 5.15 (m, 3H), 5.10 (d, $J = 3.5$ Hz, 2H), 4.76 – 4.42 (m, 5H), 4.03 (t, $J = 6.6$ Hz, 1H), 3.77 (s, 5H), 2.84 (dd, $J = 16.7, 10.4$ Hz, 1H), 2.26 – 2.08 (m, 2H), 1.68 (s, 3H), 1.44 (d, $J = 7.0$ Hz, 3H), 1.30 (dt, $J = 14.7, 7.4$ Hz, 3H), 1.12 (s, 9H), 1.10 (s, 9H), 0.96 (d, $J = 6.8$ Hz, 3H), 0.93 (d, $J = 6.8$ Hz, 3H).

(c) (11*S*,11*aS*)-4-(2-(1-((1-(allyloxy)-4-methyl-1,2-dioxopentan-3-yl)amino)-1-oxopropan-2-yl)hydrazinyl)benzyl 11-hydroxy-7-methoxy-2-methyl-5-oxo-8-((triisopropylsilyl)oxy)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**18**)

Dimethyl sulphoxide (0.2 mL, 2.75 mmol, 2.5 eq) was added dropwise to a solution of oxalyl chloride (0.11 mL, 1.32 mmol, 1.2 eq) in dry dichloromethane (7 mL) at -78°C (dry ice /acetone bath) under an atmosphere of argon. After 10 minutes a solution of **17** (920 mg, 1.10 mmol) in dry dichloromethane (5 mL) was added slowly with the temperature still at -78°C. After 15 min triethylamine (0.77 mL, dried over 4Å molecular sieves, 5.50 mmol, 5 eq) was added dropwise and the dry ice/acetone bath was removed. The reaction mixture was allowed to reach room temperature and was extracted with cold hydrochloric acid (0.1 M), saturated aqueous sodium bicarbonate and brine. The organic phase was dried over magnesium sulphate, filtered and excess dichloromethane was removed by rotary evaporation under reduced pressure. The resulting residue was subjected to column flash chromatography (silica gel; gradient 2% methanol to 5 % methanol in dichloromethane). Pure fractions were collected and combined and removal of excess eluent by rotary evaporation under reduced pressure afforded the product **18** (550 mg, 60%).

Larger scale synthesis

Dimethyl sulphoxide (3.25 mL, 45.71 mmol, 2.5 eq) was added dropwise (7 minutes) to a solution of oxalyl chloride (1.86 mL, 21.94 mmol, 1.2 eq) in dry dichloromethane (120 mL) at -78°C (dry ice /acetone bath) under an atmosphere of argon. After 15 minutes a solution of **17** (15.34 g, 18.28 mmol) in dry dichloromethane (80 mL) was added slowly (20 minutes) with the temperature still at -78°C. After 15 min triethylamine (12.7 mL, dried over 4Å molecular sieves, 91.40 mmol, 5 eq) was added dropwise (6 minutes) and the dry ice/acetone bath was removed. The reaction mixture was allowed to reach room temperature (2 hours) and was extracted with cold hydrochloric acid (0.1 M, 500 mL), saturated aqueous sodium bicarbonate (400 mL) and brine (400 mL). The organic phase was dried over magnesium sulphate, filtered and excess dichloromethane was removed by rotary evaporation under reduced pressure. The resulting residue was subjected to column flash chromatography (Biotage 340 g, sample 34 g). Pure fractions were collected and combined and removal of excess eluent by rotary evaporation under reduced pressure afforded the product **18** (12.8 g, 84%) and recovered starting material (1.57 g, 10.2% recovery).

Further large scale synthesis

To a solution of oxalyl chloride (0.77 mL, 9.2 mmol) in dichloromethane (49.5 mL) under Nitrogen at -78°C was added dimethyl sulfoxide (1.41 mL, 20.0 mmol) drop wise while maintaining the temperature < -65 °C, and the mixture was allowed to stir at -65 to -78 °C for 10-15 minutes. A solution of **17** (6.5 g, 7.8 mmol) in dichloromethane (35.3 mL) was added while maintaining the reaction temperature at -65 to -78°C, and the mixture was allowed to stir at -65 to -78 °C for 15-20 minutes. Triethylamine (5.4 mL) was added drop wise while maintaining the reaction temperature at -68 to -78°C. The reaction mixture was then allowed to warm to room temperature and stir until the reaction was judged complete by HPLC. The organics were washed with 1N HCl (35.3 mL), saturated sodium bicarbonate (35.3 mL), and brine (35.3 mL). The combined organics were dried with magnesium sulfate (2.15 g; 17.9 mmol), filtered, and the cake rinsed with dichloromethane (35.3 mL). The filtrate was then concentrated to dryness with a bath temperature not exceeding 23°C. The residue was purified by Biotage Chromatography (Si, 40 + m, 2 column volumes in each fraction, 10 fractions of 2% methanol in dichloromethane and 5 fractions of 5% methanol in dichloromethane). The positive fractions were combined and concentrated to dryness with a bath temperature not exceeding 22 °C to give **18** as a sticky solid (5.0 g, 77%, 90% AUC).

LC/MS, 3.43 min (ES+) *m/z* (relative intensity) 836.01 ([M]⁺, 100). ¹H NMR (400 MHz, CDCl₃) δ 8.39 (s, 1H), 7.52 – 7.40 (m, 2H), 7.21 – 7.08 (m, *J* = 11.5 Hz, 2H), 6.67 (s, 1H), 6.60 – 6.47 (m, *J* = 7.4 Hz, 1H), 5.97 – 5.83 (m, 1H), 5.79 – 5.66 (m, 1H), 5.38 – 4.90 (m, 6H), 4.68 – 4.52 (m, *J* = 18.4, 5.5 Hz, 4H), 4.04 – 3.94 (m, *J* = 6.5 Hz, 1H), 3.87 – 3.76 (m, 5H), 3.00 – 2.88 (m, 1H), 2.66 – 2.49 (m, 2H), 2.21 – 2.08 (m, 2H), 1.76 (s, 3H), 1.45 (d, *J* = 7.0 Hz, 3H), 1.09 – 0.98 (m, *J* = 8.9 Hz, 18H), 0.96 (d, *J* = 6.7 Hz, 3H), 0.93 (d, *J* = 6.9 Hz, 3H).

25

(*d*) (11*S*,11*aS*)-4-(2-(1-((1-(*Allyloxy*)-4-methyl-1,2-dioxopentan-3-yl)amino)-1-oxopropan-2-yl)hydrazinyl)benzyl 11-((*tert*-butyldimethylsilyl)oxy)-7-methoxy-2-methyl-5-oxo-8-((*triisopropylsilyl*)oxy)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**19**)

30

Tert-butyldimethylsilyltriflate (0.38 mL, 1.62 mmol, 3 eq) was added to a solution of compound **18** (450 mg, 0.54 mmol) and 2,6-lutidine (0.25 mL, 2.16 mmol, 4 eq) in dry dichloromethane (5 mL) at 0°C under argon. After 10 min, the cold bath was removed and the reaction mixture was stirred at room temperature for 1 hour. The reaction mixture was extracted with water, saturated aqueous sodium bicarbonate and brine. The organic phase was dried over magnesium sulphate, filtered and excess solvent was removed by rotary evaporation under reduced pressure. The resulting residue was subjected to column flash

35

chromatography (silica gel; 50/50 v/v hexane/ethyl acetate). Pure fractions were collected and combined and excess eluent was removed by rotary evaporation under reduced pressure to give the product **19** (334 mg, 65%).

5 *Larger scale synthesis*

Tert-butyldimethylsilyltriflate (10.5 mL, 45.91 mmol, 3 eq) was added to a solution of compound **18** (12.8 g, 15.30 mmol) and 2,6-lutidine (7.12 mL, 61.21 mmol, 4 eq) in dry dichloromethane (140 mL) at 0°C under argon. After 10 minutes, the cold bath was removed and the reaction mixture was stirred at room temperature for 2 hour. The reaction mixture was washed with citric acid (0.1 M, 200 mL), saturated aqueous sodium bicarbonate (200 mL) and brine (200 mL). The organic phase was dried over magnesium sulphate, filtered and excess solvent was removed by rotary evaporation under reduced pressure to give the product **19** as crude.

15 *Further synthesis*

To a solution of **18** (5.0 g, 6.0 mmol) in dichloromethane (55.6 mL) under Nitrogen at 0-5°C was added 2,6-lutidine (2.8 mL, 23.9 mmol) followed by *tert*-butyldimethylsilyl trifluoromethanesulfonate (4.94 mL, 21.5 mmol) drop wise while maintaining the temperature 0-5°C, and the mixture was allowed to stir at 0-5°C for 15-30 minutes. The reaction mixture was then allowed to warm to room temperature and stir for 2 hours until the reaction was judged complete by HPLC. The organics were washed with water (55.6 mL), saturated sodium bicarbonate (55.6 mL), and brine (55.6 mL). The combined organics were dried with magnesium sulfate (0.89 g; 7.4 mmol), filtered, and the cake rinsed with dichloromethane (55.6 mL). The filtrate was then concentrated to dryness with a bath temperature not exceeding 22°C. The residue was purified by Biotage Chromatography (Si, 40 + m, 2 column volumes in each fraction, 15 fractions of 50% EtOAc in heptane. The positive fractions were combined and concentrated to dryness with a bath temperature not exceeding 22°C to give **19** as a sticky solid (3.1 g, 54%, 93% AUC).

30 LC/MS, 4.18 min (ES+) *m/z* (relative intensity) 950.50 ($[M]^+$, 100). ^1H NMR (400 MHz, CDCl_3) δ 8.53 (s, 1H), 8.02 (s, 1H), 7.44 (d, J = 7.6 Hz, 2H), 7.21 (s, 1H), 7.08 (d, J = 8.2 Hz, 2H), 6.72 – 6.61 (m, J = 8.9 Hz, 2H), 6.16 (s, 1H), 5.97 – 5.79 (m, J = 24.4, 7.5 Hz, 2H), 5.41 – 5.08 (m, 5H), 4.86 (d, J = 12.5 Hz, 1H), 4.69 – 4.60 (m, 1H), 4.57 (s, 1H), 4.03 (t, J = 6.7 Hz, 1H), 3.87 (s, 3H), 3.74 (td, J = 9.6, 3.6 Hz, 1H), 2.43 – 2.09 (m, J = 34.8, 19.4, 11.7 Hz, 3H), 1.76 (s, 3H), 1.43 (d, J = 6.9 Hz, 3H), 1.30 – 1.21 (m, 3H), 0.97 (d, J = 6.7 Hz, 3H), 0.92 (t, J = 8.4 Hz, 3H), 0.84 (s, 9H), 0.23 (s, 3H), 0.12 (s, 3H).

(e) (11*S*,11*aS*)-4-(2-(1-((1-(Allyloxy)-4-methyl-1,2-dioxopentan-3-yl)amino)-1-oxopropan-2-yl)hydrazinyl)benzyl 11-((*tert*-butyldimethylsilyl)oxy)-8-hydroxy-7-methoxy-2-methyl-5-oxo-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**20**)

5 Lithium acetate (50 mg, 0.49 mmol) was added to a solution of compound **19** (470 mg, 0.49 mmol) in wet dimethylformamide (4 mL, 50:1 DMF/water). After 4 hours, the reaction was complete and the reaction mixture was diluted with ethyl acetate and washed with citric acid (pH ~ 3), water and brine. The organic layer was dried over magnesium sulphate filtered and excess ethyl acetate was removed by rotary evaporation under reduced pressure. The
10 resulting residue was subjected to column flash chromatography (silica gel; gradient, 50/50 to 25/75 v/v hexane/ethyl acetate). Pure fractions were collected and combined and excess eluent was removed by rotary evaporation under reduced pressure to give the product **20** (400 mg, quantitative).

15 *Larger scale synthesis*

Lithium acetate (1.56 g, 15.31 mmol) was added to a solution of compound **19** (15.31 mmol) in wet dimethylformamide (120 mL, 50:1 DMF/water). After 5 hours, the reaction was complete and the reaction mixture was concentrated to have about 20 mL of DMF. The reaction mixture was then diluted with ethyl acetate (200 mL) and washed with citric acid
20 (0.1M, 200 mL), brine (200 mL). The organic layer was dried over magnesium sulphate filtered and excess ethyl acetate was removed by rotary evaporation under reduced pressure. The resulting residue was subjected to column flash chromatography (Biotage 750 g, 80 g of silica for loading). Pure fractions were collected and combined and excess eluent was removed by rotary evaporation under reduced pressure to give the product **20** (7.7 g,
25 63% over 2 steps).

Further synthesis

To a solution of **19** (3.1 g, 3.3 mmol) in DMF (26.4 mL) and water (0.53 mL) under Nitrogen at 20-25°C was added lithium acetate (0.33 g, 5.0 mmol), and the mixture was allowed to stir
30 until the reaction was judged complete by HPLC (10 hours). Ethyl acetate (33.0 mL) and aqueous citric acid (33.0 mL pH 3) were added, and the layers were separated. The organics were washed with water (33.0 mL) and brine (33.0 mL). The combined organics were dried with magnesium sulfate (1.69 g; 14.0 mmol;), filtered, and the cake rinsed with ethyl acetate (33.0 mL). The filtrate was then concentrated to dryness with a bath
35 temperature not exceeding 22°C. The residue was purified by Biotage chromatography (Si, 45 g, 3-4 column volumes in each fraction, product loaded with a minimal amount of

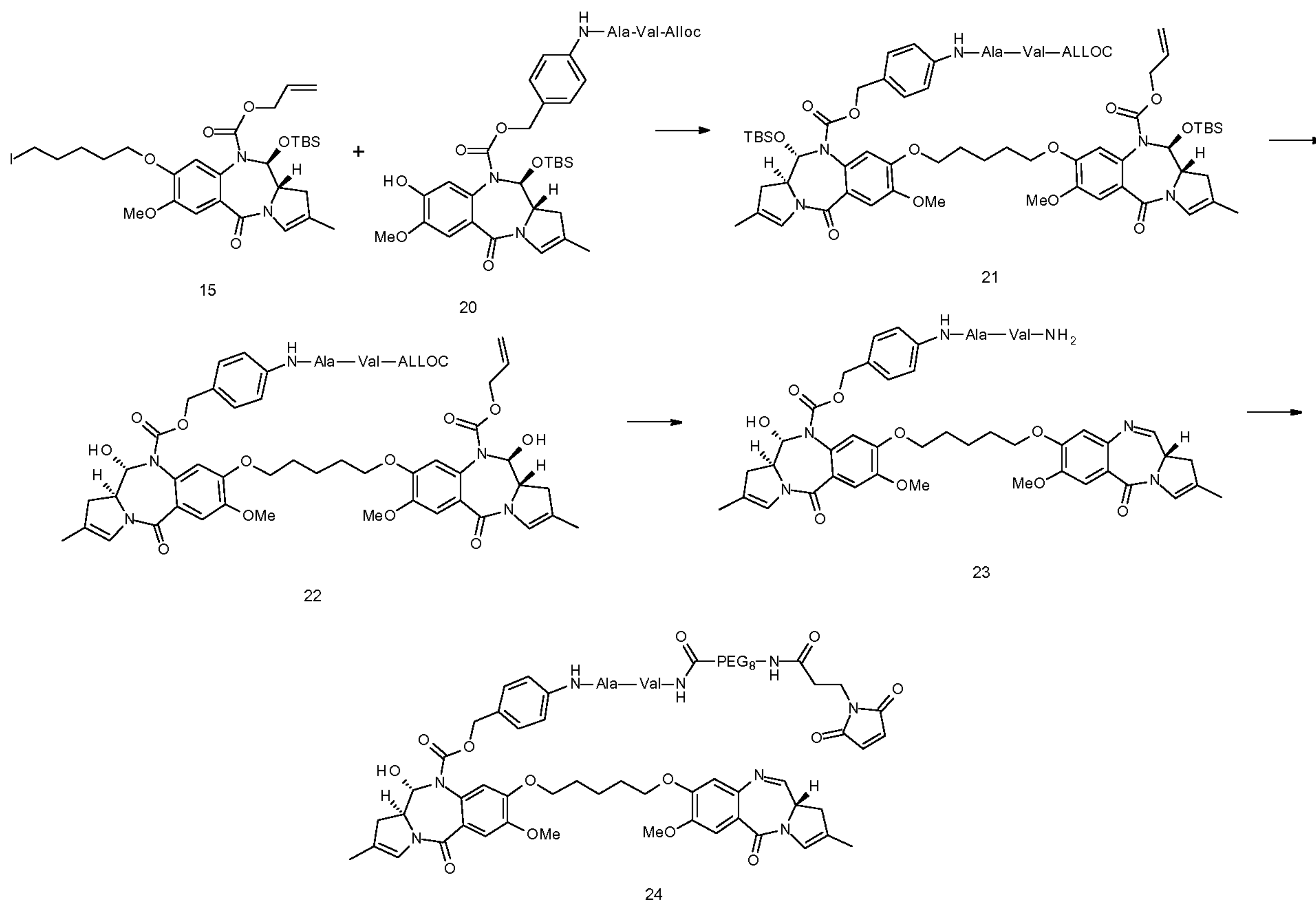
dichloromethane, 3 fractions of 50% ethyl acetate in heptane followed by 75% ethyl acetate in heptane until the product is no longer observed eluting from the column. The positive fractions were combined and concentrated to dryness with a bath temperature not exceeding 22°C to give **20** as a sticky solid (1.9 g, 73%, 94% AUC).

5

LC/MS, 3.32 min (ES+) m/z (relative intensity) 794.18 ($[M+H]^+$, 100). ^1H NMR (400 MHz, $CDCl_3$) δ 8.53 (s, 1H), 8.02 (s, 1H), 7.44 (d, $J = 7.6$ Hz, 2H), 7.21 (s, 1H), 7.08 (d, $J = 8.2$ Hz, 2H), 6.72 – 6.61 (m, $J = 8.9$ Hz, 2H), 6.16 (s, 1H), 5.97 – 5.79 (m, $J = 24.4, 7.5$ Hz, 2H), 5.41 – 5.08 (m, 5H), 4.86 (d, $J = 12.5$ Hz, 1H), 4.69 – 4.60 (m, 1H), 4.57 (s, 1H), 4.03 (t, $J = 6.7$ Hz, 1H), 3.87 (s, 3H), 3.74 (td, $J = 9.6, 3.6$ Hz, 1H), 2.43 – 2.09 (m, $J = 34.8, 19.4, 11.7$ Hz, 3H), 1.76 (s, 3H), 1.43 (d, $J = 6.9$ Hz, 3H), 1.30 – 1.21 (m, 3H), 0.97 (d, $J = 6.7$ Hz, 3H), 0.92 (t, $J = 8.4$ Hz, 3H), 0.84 (s, 9H), 0.23 (s, 3H), 0.12 (s, 3H).

10

- (iv) (11*S*,11*aS*)-4-((2*S*,5*S*)-37-(2,5-dioxo-2,5-dihydro-1*H*-pyrrol-1-yl)-5-isopropyl-2-methyl-4,7,35-trioxo-10,13,16,19,22,25,28,31-octaoxa-3,6,34-triazaheptatriacontanamido)benzyl 11-hydroxy-7-methoxy-8-(((5-(((*S*)-7-methoxy-2-methyl-5-oxo-5,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepin-8-yl)oxy)pentyl)oxy)-2-methyl-5-oxo-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**24**)



- (a) (11*S*)-allyl 8-((5-(((11*S*)-10-(((4-(2-(1-((1-(allyloxy)-4-methyl-1,2-dioxopentan-3-yl)amino)-1-oxopropan-2-yl)hydrazinyl)benzyl)oxy)carbonyl)-11-((*tert*-butyldimethylsilyl)oxy)-7-methoxy-2-methyl-5-oxo-5,10,11,11*a*-tetrahydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepin-8-yl)oxy)pentyl)oxy)-11-((*tert*-butyldimethylsilyl)oxy)-7-methoxy-2-methyl-5-oxo-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**21**)

Potassium carbonate (70 mg, 0.504 mmol, 1 eq) was added to a solution of **15** (370 mg, 0.552 mmol, 1.2 eq) and phenol **20** (400 mg, 0.504 mmol) in dry acetone (25 mL). The reaction was stirred 8 hours at 70°C. The LC/MS showed that all the starting material was not consumed, so the reaction was allowed to stir overnight at room temperature and stirred for an additional 2 hours the next day. Acetone was removed by rotary evaporation under reduced pressure. The resulting residue was subjected to flash column chromatography (silica gel; 80% ethyl acetate in hexane to 100% ethyl acetate). Pure fractions were collected and combined and excess eluent was removed by rotary evaporation under reduced pressure to give the product **21** (385 mg, 57%).

Larger scale synthesis

Potassium carbonate (790 mg, 5.71 mmol, 1 eq) was added to a solution of **15** (5.75 g, 8.57 mmol, 1.5 eq) and phenol **20** (4.54 g, 5.71 mmol) in dry 2-butanone (90 mL). The reaction was stirred 40 hours at 75°C. The reaction mixture was filtered and rinsed with ethyl acetate (90 mL). Solvent was removed by rotary evaporation under reduced pressure. The resulting residue was subjected to flash column chromatography (biotage 750g). Pure fractions were collected and combined and excess eluent was removed by rotary evaporation under reduced pressure to give the product **21** (6.1 g, 80%). Starting material **15** was also recovered.

Further synthesis

To a solution of **15** (1.8 g, 2.3 mmol) and **20** (2.27 g, 3.5 mmol) in acetone (144.0 mL) under Nitrogen at 20-25°C was added potassium carbonate (0.31 g, 2.3 mmol), and the mixture was heated to reflux and allowed to stir until the reaction was judged complete by HPLC (43 hours). After cooling to room temperature, the mixture was filtered and cake rinsed with acetone (36.0 mL). The filtrate was then concentrated to dryness with a bath temperature not exceeding 22°C. The residue was purified by Biotage chromatography (2x, 1st the crude product followed by the mixed fractions, 45 g Si, 2 column volumes in each fraction, 8 fractions of 80% EtOAc in heptane followed by 5 fractions of ethyl acetate. The pure fractions were combined and concentrated to dryness with a bath temperature not exceeding 22°C to give **21** as a sticky solid (3.0 g, 99%, 87% AUC).

LC/MS, 4.07 min (ES+) *m/z* (relative intensity) 1336.55 ([M+H]⁺, 50).

(b) (11*S*)-allyl 8-((5-(((11*S*)-10-(((4-(2-(1-((1-(allyloxy)-4-methyl-1,2-dioxopentan-3-yl)amino)-1-oxopropan-2-yl)hydrazinyl)benzyl)oxy)carbonyl)-11-hydroxy-7-methoxy-2-methyl-5-oxo-5,10,11,11*a*-tetrahydro-1*H*-benzo[e]pyrrolo[1,2-*a*][1,4]diazepin-8-yl)oxy)pentyl)oxy)-11-hydroxy-7-methoxy-2-methyl-5-oxo-11,11*a*-dihydro-1*H*-benzo[e]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**22**)

Tetra-*n*-butylammonium fluoride (1M, 0.34 mL, 0.34 mmol, 2 eq) was added to a solution of **21** (230 mg, 0.172 mmol) in dry tetrahydrofuran (3 mL). The starting material was totally consumed after 10 minutes. The reaction mixture was diluted with ethyl acetate (30 mL) and washed sequentially with water and brine. The organic phase was dried over magnesium sulphate filtered and excess ethyl acetate removed by rotary evaporation under reduced pressure. The resulting residue **22** was used as a crude mixture for the next reaction.

Larger scale synthesis

A buffered solution of tetra-*n*-butylammonium fluoride (1M, 18.2 mL, 18.2 mmol, 4 eq) and acetic acid (1.04 mL, 18.2 mmol, 4 eq) was added to a solution of **21** (6.07 g, 4.54 mmol, 1 eq) in dry tetrahydrofuran (40 mL). The starting material was totally consumed after 24 hour.

- 5 The reaction mixture was diluted with ethyl acetate (140 mL) and washed sequentially with water (100 mL), aqueous sodium hydrogen carbonate (100 mL), and brine (100 mL). The organic phase was dried over magnesium sulphate (12 g) filtered and excess ethyl acetate removed by rotary evaporation under reduced pressure. The resulting residue was subjected to flash column chromatography (Biotage system, column: SNAP 340 g, loading: 34 g
- 10 samplet, flow rate: 100 mL/min, Methanol / Ethyl Acetate, equilibration 100% Ethyl Acetate (6 CV), gradient 0% to 8% in 14 CV). Pure fractions were collected and combined and excess eluent was removed by rotary evaporation under reduced pressure to give the product **22** (3.16 g, 63%).

15 *Further synthesis*

- To a solution of **21** (1.50g. 1.12 mmol) in tetrahydrofuran (11.5 mL) under Nitrogen at 20-25°C was added a mixture of TBAF (1 M in THF; 4.5 mL) and acetic acid (0.51 mL), and the mixture was allowed to stir until the reaction was judged complete by HPLC (89 h). Note: Two additional charges of a mixture of TBAF (1 M in THF; 2.5 mL) and acetic acid (0.25 mL)
- 20 were made at 42 and 60 hours. Ethyl acetate (34.6 mL) was added. The organics were washed with water (11.5 mL), saturated sodium bicarbonate (11.5 mL) and brine (11.5 mL). The combined organics were dried with magnesium sulfate (1.15 g; 9.6 mmol), filtered, and the cake rinsed with ethyl acetate (5.8 mL). The filtrate was then concentrated to dryness with a bath temperature not exceeding 22°C. The residue was purified by Biotage
- 25 chromatography (Si, 2-3 column volumes in each fraction, product loaded with a minimal amount of dichloromethane, and eluted with 1% MeOH in EtOAc until the product is no longer observed eluting from the column. The positive fractions were combined and concentrated to dryness with a bath temperature not exceeding 22°C to give **22** as a sticky solid (1.17 g, 94%, 90% AUC).

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LC/MS, 2.87 min (ES+) *m/z* (relative intensity) 1108.11 ([M+H]⁺, 100).

- (c) (11*S*)-4-(2-(1-((1-amino-3-methyl-1-oxobutan-2-yl)amino)-1-oxopropan-2-yl)hydrazinyl)benzyl 11-hydroxy-7-methoxy-8-((5-((7-methoxy-2-methyl-5-oxo-5,11*a*-dihydro-
- 35 1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepin-8-yl)oxy)pentyl)oxy)-2-methyl-5-oxo-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**23**)

Tetrakis(triphenylphosphine)palladium(0) (12 mg, 0.01 mmol, 0.06 eq) was added to a solution of crude **22** (0.172 mmol) and pyrrolidine (36 μ L, 0.43 mmol, 2.5 eq) in dry dichloromethane (10 mL). The reaction mixture was stirred 20 minutes and diluted with dichloromethane and washed sequentially with saturated aqueous ammonium chloride and
 5 brine. The organic phase was dried over magnesium sulphate filtered and excess dichloromethane removed by rotary evaporation under reduced pressure. The resulting residue **23** was used as a crude mixture for the next reaction.

Larger scale synthesis

10 *Tetrakis*(triphenylphosphine)palladium(0) (166 mg, 0.143 mmol, 0.05 eq) was added to a solution of **22** (3.16 g, 2.85 mmol) and pyrrolidine (0.59 mL, 7.18 mmol, 2.5 eq) in dry dichloromethane (150 mL). The reaction was flushed with Argon three times and stirred 27 minutes at room temperature. Then the reaction was diluted with dichloromethane (90 mL) and washed sequentially with saturated aqueous ammonium chloride (95 mL) and brine (90
 15 mL). The organic phase was dried over magnesium sulphate (5.4 g) filtered and excess dichloromethane removed by rotary evaporation under reduced pressure. The resulting crude residue **23** (4.1 g) was dissolved in dichloromethane (310 mL) and Deloxan Metal Scavenging Resin (16.7 g) was added and the reaction was stirred for 20 min. The mixture was filtered and excess dichloromethane removed by rotary evaporation under reduced
 20 pressure. The resulting residue **23** was used as a crude mixture for the next reaction.

Further synthesis

To a solution of **22** (1.15 g, 1.04 mmol) and pyrrolidine (0.21 mL, 2.5 mmol) in dichloromethane (65.8 mL) under Nitrogen at 20-25°C was added
 25 *tetrakis*(triphenylphosphine)palladium (0) (0.072 g; 0.06 mmol), and the mixture was allowed to stir until the reaction was judged complete by TLC (< 1 hour). Dichloromethane (32.9 mL) was charged to the reactor. The organics were washed with saturated ammonium chloride (32.9 mL) and brine (32.9 mL). The combined organics were dried with magnesium sulfate (1.31 g; 10.9 mmol), filtered, and the cake rinsed with dichloromethane (13.2 mL). The
 30 filtrate was then concentrated to dryness with a bath temperature not exceeding 22°C to give **23** as a sticky solid that was used directly in the next step.

LC/MS, 2.38 min (ES+) *m/z* (relative intensity) 922.16 ([M+H]⁺; 40).

35 (d) (11*S*,11*aS*)-4-((2*S*,5*S*)-37-(2,5-dioxo-2,5-dihydro-1*H*-pyrrol-1-yl)-5-isopropyl-2-methyl-4,7,35-trioxo-10,13,16,19,22,25,28,31-octaoxa-3,6,34-triazaheptatriacontanamido)benzyl 11-

hydroxy-7-methoxy-8-((5-(((S)-7-methoxy-2-methyl-5-oxo-5,11a-dihydro-1H-benzo[e]pyrrolo[1,2-a][1,4]diazepin-8-yl)oxy)pentyl)oxy)-2-methyl-5-oxo-11,11a-dihydro-1H-benzo[e]pyrrolo[1,2-a][1,4]diazepine-10(5H)-carboxylate (**24**)

1-ethyl-3-(3'-dimethylaminopropyl)carbodiimide (EDCI, 33 mg, 0.172 mmol) was added to a solution of crude **23** (0.172 mmol) and Mal-(PEG)₈-acid (100 mg, 0.172 mmol) in dry dichloromethane (10 mL). The reaction was stirred for 2 hours and the presence of starting material was no longer observed by LC/MS. The reaction was diluted with dichloromethane and washed sequentially with water and brine. The organic phase was dried over magnesium sulphate filtered and excess dichloromethane removed by rotary evaporation under reduced pressure. The resulting residue was subjected to flash column chromatography (silica gel; 100% chloroform to 10% methanol in chloroform). Pure fractions were collected and combined and excess eluent was removed by rotary evaporation under reduced pressure to give **24 (B)** (60 mg, 25% over 3 steps).

15 *Larger scale synthesis*

1-ethyl-3-(3'-dimethylaminopropyl)carbodiimide (EDCI, 550 mg, 2.85 mmol) was added to a solution of crude **23** (2.85 mmol) and Mal-(PEG)₈-acid (1.69 g, 2.85 mmol) in dry dichloromethane (120 mL). The reaction was degassed three times with Argon and stirred for 3 hours and the presence of starting material was still observed by LC/MS (72 % conversion). An additional amount of EDCI (150 mg) and Mal-(PEG)₈-acid (475 mg) was added and the reaction was stirred for 13 hours. The reaction was diluted with dichloromethane (80 mL) and washed sequentially with water (142 mL) and brine (80 mL). The organic phase was dried over magnesium sulphate (10 g) filtered and excess dichloromethane removed by rotary evaporation under reduced pressure. The resulting residue was subjected to flash column chromatography (Biotage system, column: SNAP 340 g, loading: dissolved in 20 mL of Dichloromethane and injected, flow rate: 100 mL/min, Methanol / Chloroform, equilibration 2% (6 CV), gradient 2% to 20% in 10 CV). Pure fractions were collected and combined and excess eluent was removed by rotary evaporation under reduced pressure to give **24 (B)** (2.75 g, 64% over 2 steps, 96% purity). However one impurity was over 2% so another column chromatography was performed to give **24 (B)** (1.75 g, 41% over 2 steps, 98% purity).

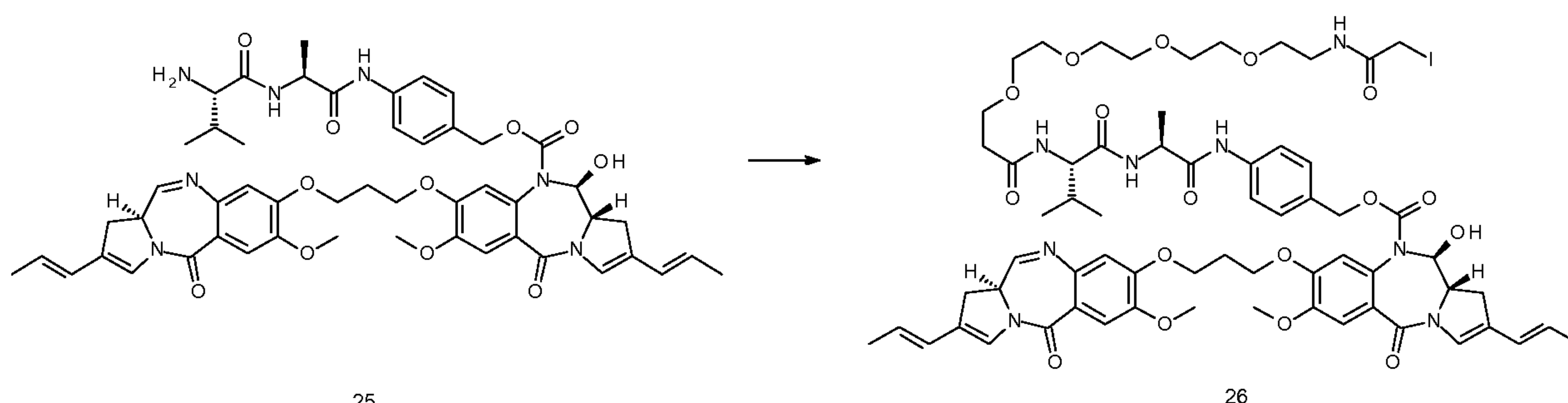
Further synthesis

To a suspension of **23** (1.04 mmol), Mal-(PEG)₈ Acid (0.616 g; 1.04 mmol; ChemPep; Lot 282808) and dichloromethane (65.9 mL) under Nitrogen at 20-25°C was added EDCI (0.199 g; 1.04 mmol; Sigma Aldrich) and the mixture was allowed to stir until the reaction was

judged complete by HPLC (16 hours). Dichloromethane (26.3 mL) was charged to the reactor. The organics were washed with water (51.7 mL) and brine (26.3 mL). The combined organics were dried with magnesium sulfate (1.32 g; 10.9 mmol), filtered, and the cake rinsed with dichloromethane (13.2 mL). The filtrate was then concentrated to dryness with a bath temperature not exceeding 22°C. The residue was purified by Biotage chromatography (Si, 40 + m, 2 column volumes in each fraction, product loaded with a minimal amount of dichloromethane (Sigma Aldrich), and eluted with 6-30% MeOH (Sigma Aldrich) in dichloromethane (Sigma Aldrich). Fractions 1-15: 6% MeOH in DCM. Fractions 16-19: 7.5% MeOH in DCM. Fractions 20-23: 10% MeOH in DCM. Fraction mapping was utilized to determine which fractions to combine. The combined fractions were concentrated to dryness with a bath temperature not exceeding 22°C. Methanol (10.0 mL) was used to dissolve the residue, which was passed through a 0.2 µ filter, which was rinsed with additional methanol (10.0 mL). The combined filtrates concentrated to dryness with a bath temperature not exceeding 22°C to give **24 (B)** as a sticky solid (0.60 g, 39%, 91% AUC).

LC/MS, method 2, 2.65 min (ES+) m/z (relative intensity) 1496.78 ($[M+H]^+$, 20). $[\alpha]_D^{24} = +262^\circ$ ($c = 0.056$, $CHCl_3$).

Example 2



Compound 25 is compound 79 of WO 2011/130598

(11*S*)-4-(1-iodo-20-isopropyl-23-methyl-2,18,21-trioxo-6,9,12,15-tetraoxa-3,19,22-triazatetracosanamido)benzyl 11-hydroxy-7-methoxy-8-(3-((7-methoxy-5-oxo-2-((*E*)-prop-1-en-1-yl)-5,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepin-8-yl)oxy)propoxy)-5-oxo-2-((*E*)-prop-1-en-1-yl)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**26**)

N,N'-diisopropylcarbodiimide (DIC, 4.71 µL, 0.0304 mmol) was added to a solution of **amine 25** (0.0276 mmol) and Iodo-(PEG)₄-acid (13.1 mg, 0.0304 mmol) in dry dichloromethane (0.8 mL). The reaction was stirred for 3 hours and the presence of starting material was no longer

observed by LC/MS. The reaction mixture was directly loaded onto a thin-layer chromatography (TLC) plate and purified by prep-TLC (10% methanol in chloroform). Pure bands were scraped off the TLC plate, taken up in 10% methanol in chloroform, filtered and excess eluent removed by rotary evaporation under reduced pressure to give **26 (A)** (20.9 mg, 56%). LC/MS, method 2, 3.08 min (ES+) *m/z* (relative intensity) 1361.16 ([M+H]⁺, 100).

General Experimental Methods for Example 3

LCMS data were obtained using an Agilent 1200 series LC/MS with an Agilent 6110 quadrupole MS, with Electrospray ionisation. Mobile phase A - 0.1% Acetic acid in water. Mobile Phase B - 0.1% in acetonitrile. Flow rate of 1.00ml/min. Gradient from 5% B rising up to 95% B over 3 minutes, remaining at 95% B for 1 minute and then back down to 5% B over 6 seconds. The total run time is 5 minutes. Column: Phenomenex Gemini-NX 3µm C18, 30 x 2.00mm. Chromatograms based on UV detection at 254nm. Mass Spectra were achieved using the MS in positive mode. Proton NMR chemical shift values were measured on the delta scale at 400 MHz using a Bruker AV400. The following abbreviations have been used: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; br, broad. Coupling constants are reported in Hz. Unless otherwise stated, column chromatography (by the flash procedure) were performed on Merck KieselgelTM silica (Art. 9385). Mass spectroscopy (MS) data were collected using a Waters Micromass LCT instrument coupled to a Waters 2795 HPLC separations module. Thin Layer Chromatography (TLC) was performed on silica gel aluminium plates (Merck 60, F₂₅₄). All other chemicals and solvents were purchased from Sigma-Aldrich or Fisher Scientific and were used as supplied without further purification.

Optical rotations were measured on an ADP 220 polarimeter (Bellingham Stanley Ltd.) and concentrations (*c*) are given in g/100mL. Melting points were measured using a digital melting point apparatus (Electrothermal). IR spectra were recorded on a Perkin-Elmer Spectrum 1000 FT IR Spectrometer. ¹H and ¹³C NMR spectra were acquired at 300 K using a Bruker Avance NMR spectrometer at 400 and 100 MHz, respectively. Chemical shifts are reported relative to TMS (δ = 0.0 ppm), and signals are designated as s (singlet), d (doublet), t (triplet), dt (double triplet), dd (doublet of doublets), ddd (double doublet of doublets) or m (multiplet), with coupling constants given in Hertz (Hz). Mass spectroscopy (MS) data were collected using a Waters Micromass ZQ instrument coupled to a Waters 2695 HPLC with a Waters 2996 PDA. Waters Micromass ZQ parameters used were: Capillary (kV), 3.38; Cone (V), 35; Extractor (V), 3.0; Source temperature (°C), 100; Desolvation Temperature (°C), 200; Cone flow rate (L/h), 50; De-solvation flow rate (L/h), 250. High-resolution mass spectroscopy (HRMS) data were recorded on a Waters Micromass QTOF Global in positive

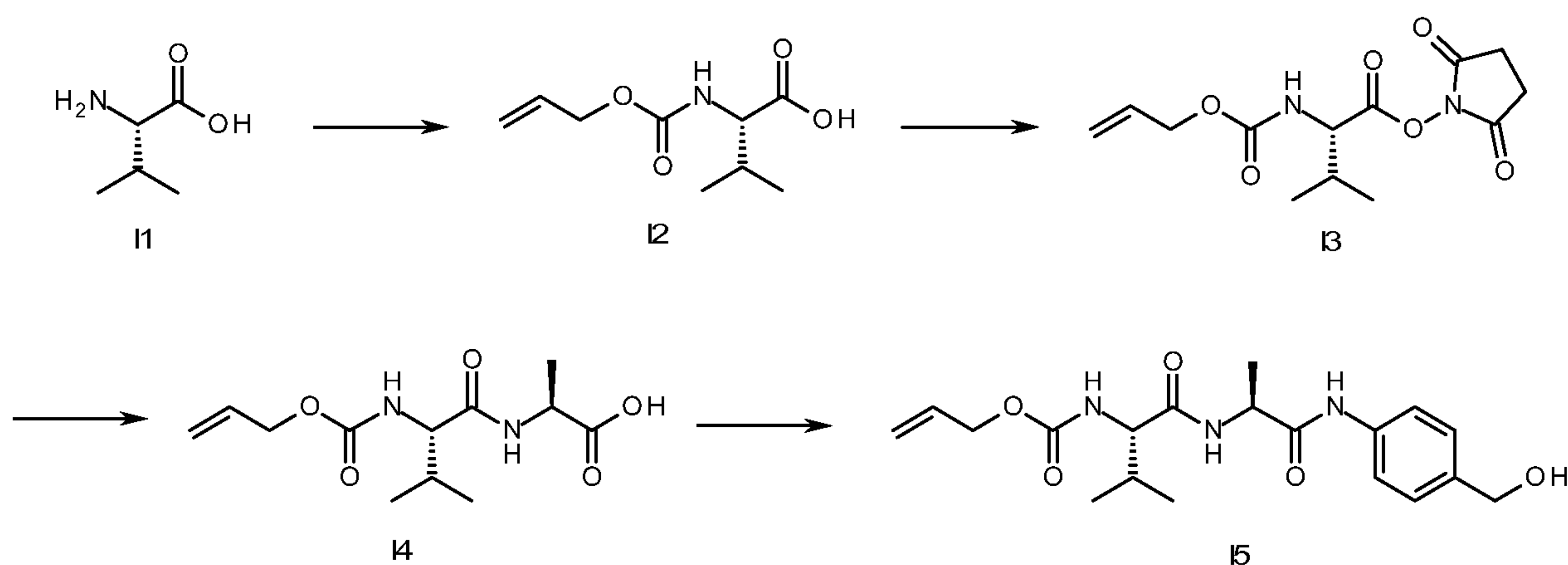
W-mode using metal-coated borosilicate glass tips to introduce the samples into the instrument. Thin Layer Chromatography (TLC) was performed on silica gel aluminium plates (Merck 60, F₂₅₄), and flash chromatography utilised silica gel (Merck 60, 230-400 mesh ASTM). Except for the HOBt (NovaBiochem) and solid-supported reagents (Argonaut), all other chemicals and solvents were purchased from Sigma-Aldrich and were used as supplied without further purification. Anhydrous solvents were prepared by distillation under a dry nitrogen atmosphere in the presence of an appropriate drying agent, and were stored over 4Å molecular sieves or sodium wire. Petroleum ether refers to the fraction boiling at 40-60°C.

General LC/MS conditions: The HPLC (Waters Alliance 2695) was run using a mobile phase of water (A) (formic acid 0.1%) and acetonitrile (B) (formic acid 0.1%). Gradient: initial composition 5% B over 1.0 min then 5% B to 95% B within 3 min. The composition was held for 0.5 min at 95% B, and then returned to 5% B in 0.3 minutes. Total gradient run time equals 5 min. Flow rate 3.0 mL/min, 400µL was split *via* a zero dead volume tee piece which passes into the mass spectrometer. Wavelength detection range: 220 to 400 nm. Function type: diode array (535 scans). Column: Phenomenex® Onyx Monolithic C18 50 x 4.60 mm

Example 3

(i) Key Intermediates

(a)



(a-i) (S)-2-(allyloxycarbonylamino)-3-methylbutanoic acid (12)

Allyl chloroformate (36.2 ml, 340.59 mmol, 1.2 eq) was added dropwise to a stirred solution of L-valine (11) (33.25 g, 283.82 mmol, 1.0 eq) and potassium carbonate (59.27 g, 425.74 mmol, 1.5 eq) in water (650 mL) and THF (650 mL). The reaction mixture was stirred at room temperature for 18 hours, then the solvent was concentrated under reduced pressure

and the remaining solution extracted with diethyl ether (3 x 100 mL). The aqueous portion was acidified to pH 2 with conc. HCl and extracted with DCM (3 x 100 mL). The combined organics were washed with brine, dried over MgSO₄, filtered and concentrated under reduced pressure to afford the product as a colourless oil (57.1 g, assumed 100% yield).

5 LC/MS (1.966 min (ES⁺)), *m/z*: 202.1 [M+H]⁺. ¹H NMR (400 MHz, DMSO-d₆) δ 12.57 (br s, 1H), 7.43 (d, 1H, *J* = 8.6 Hz), 5.96 – 5.86 (m, 1H), 5.30 (ddd, 1H, *J* = 17.2, 3.4, 1.7 Hz), 5.18 (ddd, 1H, *J* = 10.4, 2.9, 1.6 Hz), 4.48 (dt, 2H, *J* = 5.3, 1.5 Hz), 3.85 (dd, 1H, *J* = 8.6, 6.0 Hz), 2.03 (oct, 1H, *J* = 6.6 Hz), 0.89 (d, 3H, *J* = 6.4 Hz), 0.87 (d, 3H, *J* = 6.5 Hz).

10 (a-ii) (S)-2,5-dioxopyrrolidin-1-yl 2-(allyloxycarbonylamino)-3-methylbutanoate (**13**)

To a stirred solution of the protected acid **12** (60.6 g, 301.16 mmol, 1.0 eq) and N-hydroxysuccinimide (34.66 g, 301.16 mmol, 1.0 eq) in dry THF (800 mL) was added dicyclohexylcarbodiimide (62.14 g, 301.16 mmol, 1 eq). The reaction was stirred for 18 hours at room temperature. The reaction mixture was then filtered, the solid washed with THF and

15 the combined filtrate was concentrated under reduced pressure. The residue was re-dissolved in DCM and left to stand at 0°C for 30 minutes. The suspension was filtered and washed with cold DCM. Concentration of the filtrate under reduced pressure afforded the product as a viscous colourless oil (84.7 g, assumed 100% yield) which was used in the next step without further purification. LC/MS (2.194 min (ES⁺)), *m/z*: 321.0 [M+Na]⁺. ¹H NMR (400
20 MHz, DMSO-d₆) δ 8.0 (d, 1H, *J* = 8.3 Hz), 5.97 – 5.87 (m, 1H), 5.30 (ddd, 1H, *J* = 17.2, 3.0, 1.7 Hz), 5.19 (ddd, 1H, *J* = 10.4, 2.7, 1.4 Hz), 4.52 (dt, 2H, *J* = 5.3, 1.4 Hz), 4.32 (dd, 1H, *J* = 8.3, 6.6 Hz), 2.81 (m, 4H), 2.18 (oct, 1H, *J* = 6.7 Hz), 1.00 (d, 6H, *J* = 6.8 Hz),

(a-iii) (S)-2-((S)-2-(allyloxycarbonylamino)-3-methylbutanamido)propanoic acid (**14**)

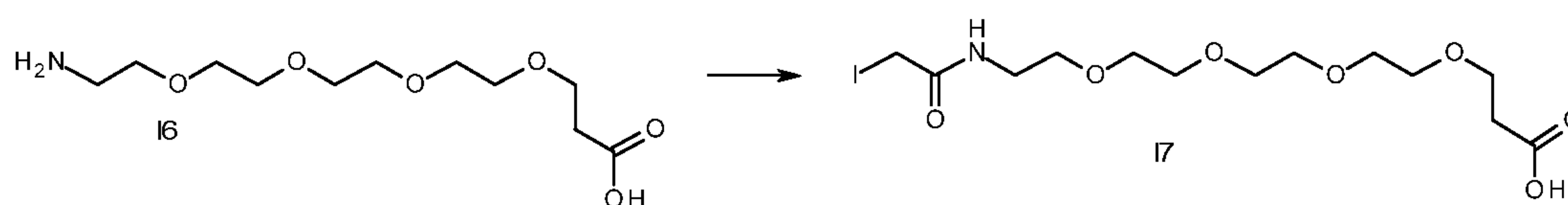
25 A solution of succinimide ester **13** (12.99 g, 43.55 mmol, 1.0 eq) in THF (50 mL) was added to a solution of L-alanine (4.07 g, 45.73 mmol, 1.05 eq) and NaHCO₃ (4.02 g, 47.90 mmol, 1.1 eq) in THF (100 mL) and H₂O (100 mL). The mixture was stirred at room temperature for 72 hours when the THF was removed under reduced pressure. The pH was adjusted to 3-4 with citric acid to precipitate a white gum. After extraction with ethyl acetate (6 x 150 mL), the
30 combined organics were washed with H₂O (200 mL), dried over MgSO₄, filtered and concentrated under reduced pressure. Trituration with diethyl ether afforded the product as a white powder which was collected by filtration and washed with diethyl ether (5.78 g, 49%).
LC/MS (1.925 min (ES⁺)), *m/z*: 273.1 [M+H]⁺. ¹H NMR (400 MHz, DMSO-d₆) δ 12.47 (br s, 1H), 8.17 (d, 1H, *J* = 6.8 Hz), 7.16 (d, 1H, *J* = 9.0 Hz), 5.95 – 5.85 (m, 1H), 5.29 (dd, 1H, *J* =
35 17.2, 1.7 Hz), 5.17 (dd, 1H, *J* = 10.4, 1.5 Hz), 4.46 (m, 2H), 4.18 (quin, 1H, *J* = 7.2 Hz), 3.87

(dd, 1H, $J = 9.0, 7.1$ Hz), 1.95 (oct, 1H, $J = 6.8$ Hz), 1.26 (d, 3H, $J = 7.3$ Hz), 0.88 (d, 3H, $J = 6.8$ Hz), 0.83 (d, 3H, $J = 6.8$ Hz).

(a-iv) *Allyl (S)-1-((S)-1-(4-(hydroxymethyl)phenylamino)-1-oxopropan-2-ylamino)-3-methyl-1-oxobutan-2-ylcarbamate (15)*

EEDQ (5.51 g, 22.29 mmol, 1.05 eq) was added to a solution of *p*-aminobenzyl alcohol (2.74 g, 22.29 mmol, 1.05 eq) and acid **14** (5.78 g, 21.23 mmol, 1 eq) in dry THF (100 mL). and stirred at room temperature for 72 hours. The reaction mixture was then concentrated under reduced pressure and the resulting brown solid was triturated with diethyl ether and filtered with subsequent washing with an excess of diethyl ether to afford the product as an off-white solid (7.1 g, 88 %). LC/MS (1.980 min (ES⁺)), m/z : 378.0 [M+H]⁺. ¹H NMR (400 MHz, DMSO-d₆) δ 9.89 (br s, 1H), 8.13 (d, 1H, $J = 7.0$ Hz), 7.52 (d, 2H, $J = 8.5$ Hz), 7.26 (m, 1H), 7.23 (d, 2H, $J = 8.5$ Hz), 5.91 (m, 1H), 5.30 (m, 1H), 5.17 (m, 1H), 4.46 (m, 2H), 5.09 (t, 1H, $J = 5.6$ Hz), 4.48 (m, 2H), 4.42 (m, 3H), 3.89 (dd, 1H, $J = 8.6, 6.8$ Hz), 1.97 (m, 1H), 1.30 (d, 3H, $J = 7.1$ Hz), 0.88 (d, 3H, $J = 6.8$ Hz), 0.83 (d, 3H, $J = 6.7$ Hz).

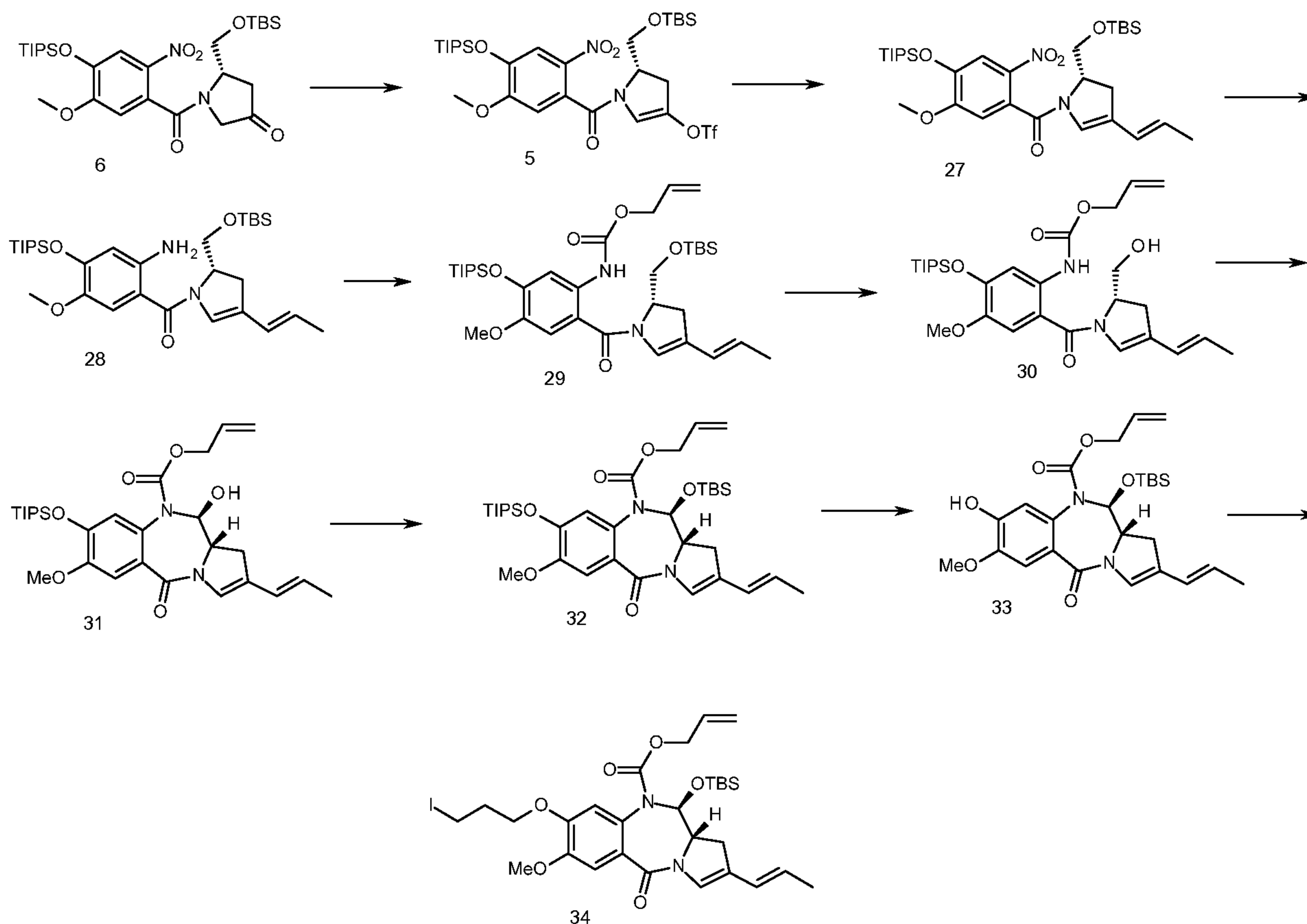
(b)



1-iodo-2-oxo-6,9,12,15-tetraoxa-3-azaocetadecan-18-oic acid (17)

A solution of iodoacetic anhydride (0.250 g, 0.706 mmol, 1.1 eq) in dry DCM (1 mL) was added to amino-PEG₍₄₎-acid **16** (0.170 g, 0.642 mmol, 1.0 eq) in DCM (1 mL). The mixture was stirred in the dark at room temperature overnight. The reaction mixture was washed with 0.1 M HCl, water, dried over MgSO₄, filtered and concentrated under reduced pressure. The residue was purified by flash chromatography (silica gel, 3% MeOH and 0.1% formic acid in chloroform to 10% MeOH and 0.1% formic acid in chloroform) to afford the product as an orange oil (0.118 g, 42%). LC/MS (1.623 min (ES⁺)), m/z : 433..98 [M+H]⁺. ¹H NMR (400 MHz, CDCl₃) δ 8.069 (s, 1H), 7.22 (br s, 1H), 3.79 (t, 2H, $J = 5.8$ Hz), 3.74 (s, 2H), 3.72 – 3.58 (m, 14H), 3.50 – 3.46 (m, 2H), 2.62 (t, 2H, $J = 5.8$ Hz).

(ii) (11S,11aS)-allyl 11-(*tert*-butyldimethylsilyloxy)-8-(3-iodopropoxy)-7-methoxy-5-oxo-2-((*E*)-prop-1-enyl)-11,11a-dihydro-1H-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5H)-carboxylate (34)



5

(a) (*S*)-5-((*tert*-butyldimethylsilyloxy)methyl)-1-(5-methoxy-2-nitro-4-(*triisopropylsilyloxy*)benzoyl)-4,5-dihydro-1H-pyrrol-3-yl trifluoromethanesulfonate (7)

Triflic anhydride (28.4 g, 100.0 mmol, 3.0 eq) was added dropwise, over 25 mins, to a vigorously stirred solution of the ketone **6** (19.5 g, 30.0 mmol, 1.0 eq) in DCM (550 mL) containing 2,6-lutidine (14.4 g, 130.0 mmol, 4.0 eq) at -50°C. The reaction mixture was stirred for 1.5 hours when LC/MS indicated complete reaction. The organic phase was washed successively with water (100 mL), saturated sodium bicarbonate (150 mL), brine (50 mL), and the organic phase was dried over MgSO₄, filtered and concentrated under reduced pressure. The residue was purified by flash chromatography (silica gel, 90/10 v/v *n*-hexane/EtOAc) to afford the product as a pale yellow oil (19.5 g, 82 %). LC/MS (4.391 min (ES⁺)), *m/z*: 713.25 [M+H]⁺. ¹H NMR (400 MHz, CDCl₃) δ 7.68 (s, 1H), 6.72 (s, 1H), 6.02 (t, 1H, *J* = 1.9 Hz), 4.75 (m, 1H), 4.05 (m, 2H), 3.87 (s, 3H), 3.15 (ddd, 1H, *J* = 16.2, 10.3, 2.3 Hz), 2.96 (ddd, 1H, *J* = 16.2, 4.0, 1.6 Hz), 1.28 – 1.21 (m, 3H), 1.07 (d, 18H, *J* = 7.2 Hz), 0.88 (s, 9H), 0.09 (s, 3H), 0.08 (s, 3H).

(b) *(S,E)*-(2-((*tert*-butyldimethylsilyloxy)methyl)-4-(prop-1-enyl)-2,3-dihydro-1*H*-pyrrol-1-yl)(5-methoxy-2-nitro-4-(triisopropylsilyloxy)phenyl)methanone (**27**)

Tetrakis(triphenylphosphine)palladium(0) (0.41 g, 0.35 mmol, 0.03 eq) was added to a mixture of the triflate **7** (8.4 g, 11.8 mmol, 1.0 eq), *E*-1-propene-1-ylboronic acid (1.42 g, 16.5 mmol, 1.4 eq) and potassium phosphate (5.0 g, 23.6 mmol, 2.0 eq) in dry dioxane (60 mL) under a nitrogen atmosphere. The mixture was stirred at 25°C for 120 mins when LC/MS indicated complete reaction. Ethyl acetate (120 mL) and water (120 mL) were added, the organic phase was removed, washed with brine (20 mL), dried over MgSO₄, filtered and concentrated under reduced pressure. The residue was purified by flash chromatography (silica gel, 95/5 v/v *n*-hexane/EtOAc to 90/10 v/v *n*-hexane/EtOAc) to afford the product as a yellow foam (4.96 g, 70 %). LC/MS (4.477 min (ES⁺)), *m/z*: 605.0 [M+H]⁺. ¹H NMR (400 MHz, CDCl₃) δ 7.67 (s, 1H), 6.74 (s, 1H), 5.93 (d, 1H, *J* = 15.4 Hz), 5.67 (s, 1H), 4.65 (m, 1H), 4.04 (m, 2H), 3.86 (s, 3H), 2.85 (m, 1H), 2.71 (m, 1H), 1.72 (dd, 3H, *J* = 6.8, 1.0 Hz), 1.30 – 1.22 (m, 3H), 1.07 (d, 18H, *J* = 7.2 Hz), 0.87 (s, 9H), 0.08 (s, 3H), 0.07 (s, 3H).

15

(c) *(S,E)*-(2-amino-5-methoxy-4-(triisopropylsilyloxy)phenyl)(2-((*tert*-butyldimethylsilyloxy)methyl)-4-(prop-1-enyl)-2,3-dihydro-1*H*-pyrrol-1-yl)methanone (**28**)

Zinc dust (22.0 g, 0.33 mol, 37 eq) was added, in portions over 20 mins, to a solution of the propenyl intermediate **27** (5.5 g, 9.1 mmol, 1.0 eq) in 5% v/v formic acid / ethanol (55 mL), using an ice bath to maintain the temperature between 25-30°C. After 30 mins, the reaction mixture was filtered through a short bed of Celite®. The Celite® was washed with ethyl acetate (65 mL) and the combined organics were washed successively with water (35 mL), saturated sodium bicarbonate (35 mL) and brine (10 mL). The organic phase was dried over MgSO₄, filtered and concentrated under reduced pressure. The residue was purified by flash chromatography (silica gel, 90/10 v/v *n*-hexane/EtOAc) to afford the product as a pale yellow oil (3.6 g, 69.0 %). LC/MS (4.439 min (ES⁺)), *m/z*: 575.2 [M+H]⁺. ¹H NMR (400 MHz, CDCl₃) δ 6.75 (m, 1H), 6.40 (br s, 1H), 6.28 (m, 1H), 6.11 (d, 1H, *J* = 15.4 Hz), 5.53 (m, 1H), 4.67 (m, 1H), 4.36 (m, 2H), 3.93 (br s, 1H), 3.84 (br s, 1H), 3.73 (s, 3H), 2.86 (dd, 1H, *J* = 15.7, 10.4 Hz), 2.73 (dd, 1H, *J* = 15.9, 4.5 Hz), 1.80 (dd, 3H, *J* = 6.8, 1.3 Hz), 1.35 – 1.23 (m, 3H), 1.12 (d, 18H, *J* = 7.3 Hz), 0.89 (s, 9H), 0.08 (s, 3H), 0.07 (s, 3H).

30

(d) *(S,E)*-allyl 2-(2-((*tert*-butyldimethylsilyloxy)methyl)-4-(prop-1-enyl)-2,3-dihydro-1*H*-pyrrole-1-carbonyl)-4-methoxy-5-(triisopropylsilyloxy)phenylcarbamate (**29**)

Allyl chloroformate (0.83 g, 6.88 mmol, 1.1 eq) was added to a solution of the amine **28** (3.6 g, 6.26 mmol, 1.0 eq) in dry DCM (80 mL) containing dry pyridine (1.09 g, 13.77 mmol, 2.2 eq) at -78°C. The dry ice was removed and the reaction mixture allowed to warm to room

35

temperature. After stirring for a further 15 minutes, LC/MS indicated complete reaction. The organic phase was washed successively with 0.01N HCl (50 mL), saturated sodium bicarbonate (50 mL), brine (10 mL), dried over MgSO₄, filtered and concentrated under reduced pressure to leave a pale yellow oil which was used in the next step without further purification (4.12g, assumed 100% yield). LC/MS (4.862 min (ES⁺)), *m/z*: 659.2 [M+H]⁺.

(e)(S,E)-allyl 2-(2-(hydroxymethyl)-4-(prop-1-enyl)-2,3-dihydro-1H-pyrrole-1-carbonyl)-4-methoxy-5-(triisopropylsilyloxy)phenylcarbamate (30)

The crude intermediate **29** (assumed 100% yield, 4.12 g, 6.25 mmol, 1.0 eq) was dissolved in a mixture of acetic acid (70 mL), methanol (10 mL), THF (10 mL) and water (20 mL) and allowed to stir at room temperature. After 6 hours the reaction mixture was diluted with ethyl acetate (500 mL) and washed successively with water (2 x 500 mL), saturated sodium bicarbonate (300 mL) and brine (50 mL). The organic phase was dried over MgSO₄, filtered and concentrated under reduced pressure. The residue was purified by flash chromatography (silica gel, 1/99 v/v methanol/DCM to 5/95 v/v methanol/DCM) to afford the product as a yellow oil and a further 1 g of unreacted starting material was recovered. This material was subjected to the same reaction conditions as above, but was left stirring for 16 h. After work up and purification, additional product was isolated (2.7 g, 79%, 2 steps) LC/MS (3.742 min (ES⁺)), *m/z*: 545.2 [M+H]⁺. ¹H NMR (400 MHz, CDCl₃) δ 8.38 (m, 1H), 7.72 (m, 1H), 6.81 (s, 1H), 6.37 (m, 1H), 6.10 (d, 1H, *J* = 15.8 Hz), 5.97 (m, 1H), 5.53 (m, 1H), 5.36 (ddd, 1H, *J* = 17.2, 3.1, 1.5 Hz), 5.25 (ddd, 1H, *J* = 10.4, 2.5, 1.3 Hz), 4.78 (m, 1H), 4.65 (dt, 2H, *J* = 5.7, 1.3 Hz), 3.84 (m, 3H), 3.79 (s, 3H), 3.04 (dd, 1H, *J* = 16.7, 10.5 Hz), 2.40 (dd, 1H, *J* = 16.0, 4.5 Hz), 1.82 (dd, 3H, *J* = 6.8, 1.0 Hz), 1.36 – 1.26 (m, 3H), 1.14 (d, 18H, *J* = 7.3 Hz).

(f) (11S,11aS)-allyl 11-hydroxy-7-methoxy-5-oxo-2-((E)-prop-1-enyl)-8-(triisopropylsilyloxy)-11,11a-dihydro-1H-benzo[e]pyrrolo[1,2-a][1,4]diazepine-10(5H)-carboxylate (31)

Dry dimethyl sulfoxide (1.16 g, 14.87 mmol, 3.0 eq) was added dropwise to a solution of oxalyl chloride (0.94 g, 7.43 mmol, 1.5 eq) in DCM (25 mL) at -78°C under an atmosphere of nitrogen. Maintaining the temperature at -78°C, after 10 mins a solution of the primary alcohol **30** (2.7 g, 4.96 mmol, 1.0 eq) in DCM (20 mL) was added dropwise. After a further 15 mins, dry triethylamine (2.5g, 24.78 mmol, 5.0 eq) was added, and the reaction mixture allowed to warm to room temperature. The reaction mixture was washed successively with cold 0.1N HCl (50 mL), saturated sodium hydrogen carbonate (50 mL) and brine (10 mL) and the organic layer was dried over MgSO₄, filtered and concentrated under reduced

pressure to afford the product as a yellow oil which was used in the next step without further purification (2.68g, assumed 100% yield). LC/MS (3.548 min (ES⁺)), *m/z*: 543.2 [M+H]⁺.

(g) (11*S*,11*aS*)-allyl 11-(*tert*-butyldimethylsilyloxy)-7-methoxy-5-oxo-2-((*E*)-prop-1-enyl)-8-(triisopropylsilyloxy)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**32**)

Tert-butyldimethylsilyltrifluoromethane sulfonate (3.93 g, 14.87 mmol, 3.0 eq) was added to a solution of the carbinolamine **31** (assumed 100% yield, 2.68 g, 4.96 mmol, 1.0 eq) and 2,6-lutidine (2.12 g, 19.83 mmol, 4.0 eq) in dry DCM (40 mL) at 0°C under an atmosphere of nitrogen. After 10 minutes, the reaction mixture was allowed to warm to room temperature and stirred for a further 60 minutes. The organic phase was washed successively with water (10 mL), saturated sodium bicarbonate (10 mL) and brine (5 mL), dried over MgSO₄, filtered and concentrated under reduced pressure. The residue was purified by flash chromatography (silica gel, chloroform to 2/98 v/v Methanol/chloroform) to afford the product as a yellow oil (2.0g, 63%, 2 steps). LC/MS (4.748 min (ES⁺)), *m/z*: 657.2 [M+H]⁺. ¹H NMR (400 MHz, CDCl₃) δ 7.19 (s, 1H), 6.86 (m, 1H), 6.66 (s, 1H), 6.22 (d, 1H, *J* = 15.4 Hz), 5.81 (d, 1H, *J* = 8.8 Hz), 5.78 (m, 1H), 5.48 (m, 1H), 5.11 (d, 1H, *J* = 5.0 Hz), 5.08 (m, 1H), 4.58 (dd, 1H, *J* = 13.4, 5.4 Hz), 4.35 (dd, 1H, *J* = 13.2, 5.7 Hz), 3.83 (s, 3H), 3.76 (s, 1H), 3.00 (dd, 1H, *J* = 15.6, 11.0 Hz), 2.53 (m, 1H), 1.81 (dd, 3H, *J* = 6.8, 0.9 Hz), 1.30 – 1.18 (m, 3H), 1.08 (d, 9H, *J* = 2.3 Hz), 1.06 (d, 9H, *J* = 2.3 Hz), 0.86 (s, 9H), 0.25 (s, 3H), 0.18 (s, 3H).

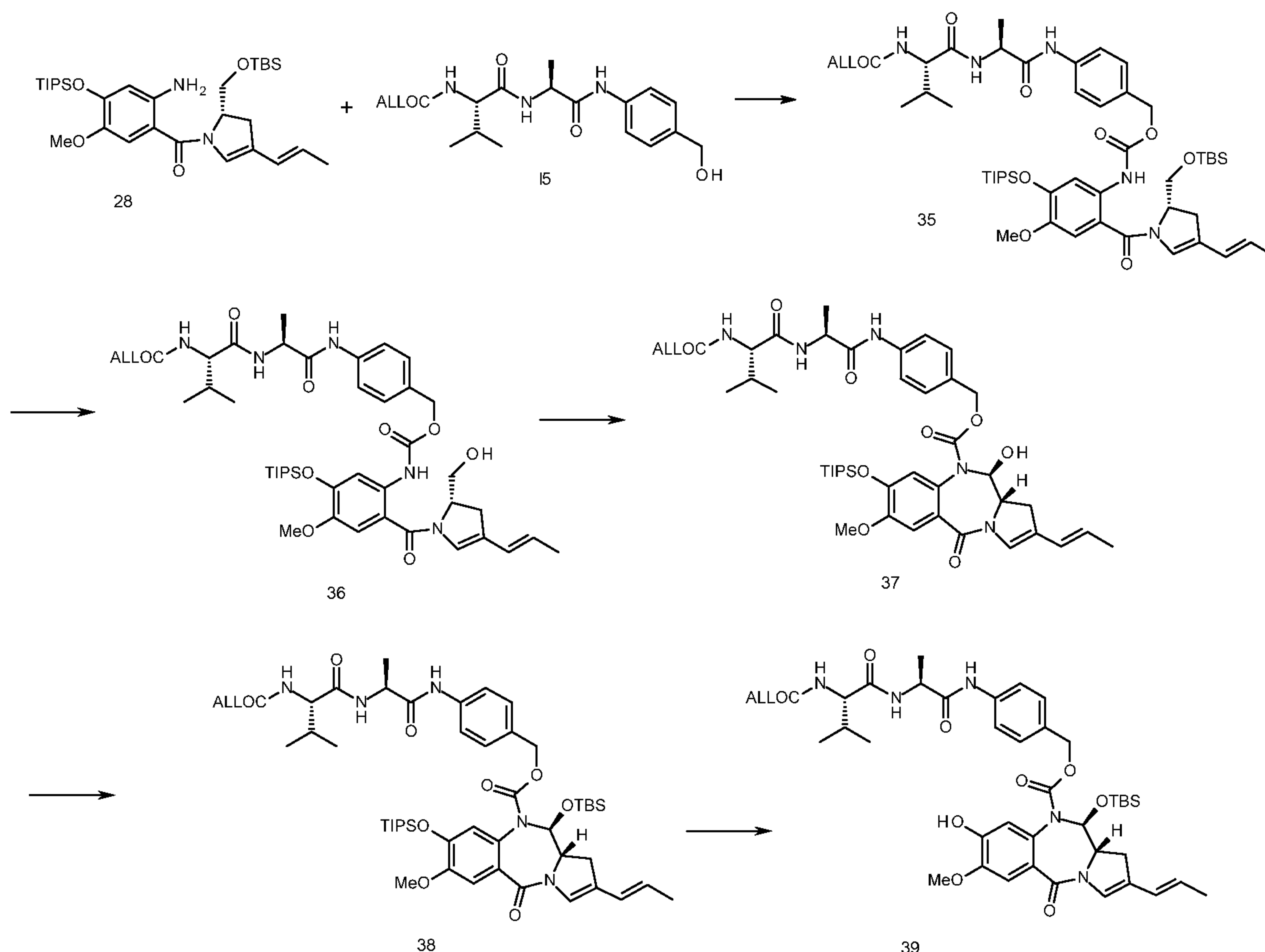
(h) (11*S*,11*aS*)-allyl 11-(*tert*-butyldimethylsilyloxy)-8-hydroxy-7-methoxy-5-oxo-2-((*E*)-prop-1-enyl)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**33**)

Lithium acetate dihydrate (0.31 g, 3.04 mmol, 1.0 eq) was added to a solution of the diazepine **32** (2.0 g, 3.04 mmol, 1.0 eq) in wet DMF (20 mL) at 25°C and stirred for 4 hours. The reaction mixture was diluted with ethyl acetate (200 mL) and washed successively with 0.1M citric acid (50 mL, pH 3), water (50 mL) and brine (10 mL), dried over MgSO₄, filtered and concentrated under reduced pressure. The residue was purified by flash chromatography (silica gel, 50/50 v/v *n*-hexane/EtOAc to 25/75 v/v *n*-hexane/EtOAc) to afford the product as a pale yellow solid (0.68g, 45 %). LC/MS (3.352 min (ES⁺)), *m/z*: 501.1 [M+H]⁺. ¹H NMR (400 MHz, CDCl₃) δ 7.02 (s, 1H), 6.66 (m, 1H), 6.53 (s, 1H), 6.03 (d, 1H, *J* = 15.5 Hz), 5.80 (s, 1H), 5.63 (d, 1H, *J* = 8.9 Hz), 5.55 (m, 1H), 5.29 (m, 1H), 4.87 (m, 2H), 4.39 (dd, 1H, *J* = 13.5, 4.2 Hz), 4.20 (dd, 1H, *J* = 13.2, 5.7 Hz), 3.73 (s, 3H), 3.59 (m, 1H), 2.81 (dd, 1H, *J* = 16.1, 10.5 Hz), 2.35 (d, 1H, *J* = 15.7 Hz), 1.61 (d, 3H, *J* = 6.4 Hz), 0.67 (s, 9H), 0.05 (s, 3H), 0.00 (s, 3H).

(i) (11*S*,11*aS*)-allyl 11-(*tert*-butyldimethylsilyloxy)-8-(3-iodopropoxy)-7-methoxy-5-oxo-2-((*E*)-prop-1-enyl)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**34**)

5 Diiodopropane (0.295 g, 1.00 mmol, 5.0 eq) and potassium carbonate (0.028 g, 0.20 mmol, 1.0 eq) were added to a solution of the phenol **33** (0.100 g, 0.020 mmol, 1.0 eq) in dry acetone (5 mL). The reaction mixture was heated at 60°C for 6 hours when LC/MS showed complete reaction. The reaction mixture was concentrated to dryness under reduced pressure and the residue was purified by flash chromatography (silica gel, 75/25 v/v *n*-hexane/EtOAc to 50/50 v/v *n*-hexane/EtOAc) to afford the product as a colourless oil (0.074 g, 56%). LC/MS (3.853 min (ES⁺)), *m/z*: 669.0 [M+H]⁺. ¹H NMR (400 MHz, CDCl₃) δ 7.26 (s, 1H), 6.90 (s, 1H), 6.68 (s, 1H), 6.24 (d, 1H, *J* = 15.3 Hz), 5.87 (d, 1H, *J* = 8.9 Hz), 5.78 (m, 1H), 5.53 (m, 1H), 5.12 (m, 2H), 4.65 (m, 2H), 4.41 (m, 1H), 4.11 (m, 1H), 3.93 (s, 3H), 3.81 (m, 1H), 3.40 (t, 2H, *J* = 6.7 Hz), 3.05 (dd, 1H, *J* = 16.3, 10.1 Hz), 2.57 (m, 1H), 2.34 (m, 2H), 1.84 (d, 3H, *J* = 6.6 Hz), 0.92 (s, 9H), 0.28 (s, 3H), 0.26 (s, 3H).

(iii) (11*S*,11*aS*)-4-((*S*)-2-((*S*)-2-(allyloxycarbonylamino)-3-methylbutanamido)propanamido)benzyl 11-(*tert*-butyldimethylsilyloxy)-8-hydroxy-7-methoxy-5-oxo-2-((*E*)-prop-1-enyl)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**39**)



5

(a) Allyl ((*S*)-1-(((*S*)-1-((4-(((2-((*S*)-2-(((*tert*-butyldimethylsilyl)oxy)methyl)-4-((*E*)-prop-1-en-1-yl)-2,3-dihydro-1*H*-pyrrole-1-carbonyl)-4-methoxy-5-((triisopropylsilyl)oxy)phenyl)carbonyl)oxy)methyl)phenyl)amino)-1-oxopropan-2-yl)amino)-3-methyl-1-oxobutan-2-yl)carbamate (**35**)

- 10 Triethylamine (0.256 mL, 1.84 mmol, 2.2 eq) was added to a stirred solution of the amine **28** (0.480 g, 0.835 mmol, 1.0 eq) and triphosgene (0.089 g, 0.301 mmol, 0.36 eq) in dry THF (15 mL) at 5°C (ice bath). The progress of the isocyanate reaction was monitored by periodically removing aliquots from the reaction mixture and quenching with methanol and performing LCMS analysis. Once the isocyanate reaction was complete a solution of Alloc-
- 15 Val-Ala-PABOH **15** (0.473 g, 1.25 mmol, 1.5 eq) and triethylamine (0.174 mL, 1.25 mmol, 1.5 eq) in dry THF (10 mL) was rapidly added by injection to the freshly prepared isocyanate. The reaction was allowed to stir at 40°C for 4 hours followed by stirring at room temperature overnight. The mixture was concentrated under reduced pressure, and purified by flash chromatography (silica gel, 20/80 v/v *n*-hexane/EtOAc to 50/50 v/v *n*-hexane/EtOAc, then

1/99 v/v DCM/MeOH to 5/95 v/v DCM/MeOH) to afford the product as a yellow solid (0.579 g, 71%). LC/MS (4.468 min (ES⁺)), *m/z*: 978.55 [M+H]⁺. ¹H NMR (400 MHz, CDCl₃) δ 8.63 (br s, 1H), 8.42 (s, 1H), 7.78 (br s, 1H), 7.53 (d, 2H, *J* = 8.1 Hz), 7.31 (d, 2H, *J* = 8.6 Hz), 6.76 (s, 1H), 6.59 (d, 1H, *J* = 7.6 Hz), 6.36 (br s, 1H), 6.04 (d, 1H, *J* = 15.9 Hz), 5.90 (m, 1H), 5.55 (m, 1H), 5.33 – 5.21 (m, 3H), 5.10 (s, 2H), 4.66 (m, 2H), 4.57 (dd, 2H, *J* = 5.6, 1.0 Hz), 3.98 (dd, 1H, *J* = 7.3, 6.8 Hz), 3.90 (m, 1H), 3.81 (m, 1H), 3.78 (s, 3H), 2.82 (dd, 1H, *J* = 15.4, 9.6 Hz), 2.72 (dd, 1H, *J* = 15.9, 3.5 Hz), 2.17 (m, 1H), 1.78 (dd, 3H, *J* = 6.5, 0.8 Hz), 1.46 (d, 3H, *J* = 7.1 Hz), 1.29 (m, 3H), 1.11 (d, 18H, *J* = 7.1 Hz), 0.97 (d, 3H, *J* = 6.8 Hz), 0.92 (d, 3H, *J* = 6.8 Hz), 0.83 (s, 9H), 0.04 (s, 3H), 0.01 (s, 3H).

(b) *Allyl ((S)-1-(((S)-1-((4-(((2-((S)-2-(hydroxymethyl)-4-((E)-prop-1-en-1-yl)-2,3-dihydro-1H-pyrrole-1-carbonyl)-4-methoxy-5-((triisopropylsilyl)oxy)phenyl)carbonyl)oxy)methyl)phenyl)amino)-1-oxopropan-2-yl)amino)-3-methyl-1-oxobutan-2-yl)carbamate (36)*

The silyl ether **35** (1.49 g, 1.52 mmol, 1.0 eq) was dissolved in a 7:1:1:2 mixture of acetic acid/ methanol/ tetrahydrofuran/ water (14:2:2:4 mL) and allowed to stir at room temperature. After 2 hours the reaction was diluted with EtOAc (100 mL), washed sequentially with water, aq. sodium bicarbonate then brine. The organic phase was then dried over MgSO₄, filtered and concentrated under reduced pressure. The residue was purified by flash chromatography (silica gel, 100/0 then 99/1 to 92/8 v/v DCM/ MeOH) to afford the product as an orange solid (1.2 g, 92%). LC/MS (3.649 min (ES⁺)), *m/z*: 865.44 [M+H]⁺. ¹H NMR (400 MHz, CDCl₃) δ 8.44 (s, 1H), 8.35 (s, 1H), 7.69 (br s, 1H), 7.53 (d, 2H, *J* = 8.7 Hz), 7.32 (d, 2H, *J* = 8.3 Hz), 6.78 (s, 1H), 6.56 (m, 2H), 6.32 (br s, 1H), 6.05 (d, 1H, *J* = 14.9 Hz), 5.90 (m, 1H), 5.56 (m, 1H), 5.30 (m, 2H), 5.22 (m, 1H), 5.10 (d, 2H, *J* = 3.1 Hz), 4.73 (m, 1H), 4.64 (m, 1H), 4.57 (d, 2H, *J* = 5.8 Hz), 4.01 (m, 1H), 3.79 (m, 2H), 3.76 (s, 3H), 2.98 (dd, 1H, *J* = 16.3, 10.2 Hz), 2.38 (dd, 1H, *J* = 16.6, 4.1 Hz), 2.16 (m, 1H), 1.78 (dd, 3H, *J* = 6.8, 0.9 Hz), 1.46 (d, 3H, *J* = 7.1 Hz), 1.29 (m, 3H), 1.11 (d, 18H, *J* = 7.4 Hz), 0.97 (d, 3H, *J* = 6.7 Hz), 0.92 (d, 3H, *J* = 6.8 Hz).

(c) *((11S,11aS)-4-((S)-2-((S)-2-(allyloxycarbonylamino)-3-methylbutanamido)propanamido)benzyl 11-hydroxy-7-methoxy-5-oxo-2-((E)-prop-1-enyl)-8-(triisopropylsilyloxy)-11,11a-dihydro-1H-benzo[e]pyrrolo[1,2-a][1,4]diazepine-10(5H)-carboxylate (37)*

Dry dimethyl sulfoxide (0.180 g, 2.3 mmol, 3.0 eq) was added dropwise to a solution of oxalyl chloride (0.147 g, 1.1 mmol, 1.5 eq) in DCM (10 mL) at -78°C under an atmosphere of nitrogen. Maintaining the temperature at -78°C, after 20 minutes, a solution of the primary

alcohol **36** (0.666 g, 0.77 mmol, 1.0 eq) in DCM (10 mL) was added dropwise. After a further 15 minutes, dry triethylamine (0.390 g, 3.85 mmol, 5.0 eq) was added, and the reaction mixture allowed to warm to room temperature. The reaction mixture was washed successively with cold 0.1N HCl (10 mL), saturated sodium hydrogen carbonate (10 mL) and brine (5 mL). The organic layer was then dried over MgSO₄, filtered and concentrated under reduced pressure. The residue was then purified by flash chromatography (silica gel, 50/50 v/v *n*-hexane/EtOAc to 25/75 v/v *n*-hexane/EtOAc) to afford the product as a white solid (0.356g, 54 %). LC/MS (3.487 min (ES⁺)), *m/z*: 862.2 [M+H]⁺. ¹H NMR (400 MHz, CDCl₃) δ 8.34 (br s, 1H), 7.47 (d, 2H, *J* = 7.6 Hz), 7.17 (s, 1H), 7.14 (d, 2H, *J* = 7.5 Hz), 6.86 (br s, 1H), 6.65 (br s, 1H), 6.42 (d, 1H, *J* = 7.6 Hz), 6.22 (d, 1H, *J* = 14.4 Hz), 5.80 (m, 1H), 5.40 (m, 1H), 5.53 (m, 1H), 5.32 (m, 1H), 5.21 (d, 2H, *J* = 9.6 Hz), 5.06 (d, 1H, *J* = 12.3 Hz), 4.90 (m, 1H), 4.58 (m, 3H), 3.98 (m, 1H), 3.84 (m, 1H), 3.81 (s, 3H), 3.50 (m, 1H), 3.05 (dd, 1H, *J* = 16.0, 10.3 Hz), 2.76 (m, 1H), 2.15 (m, 1H), 1.80 (dd, 3H, *J* = 6.7, 0.8 Hz), 1.44 (d, 3H, *J* = 7.1 Hz), 1.16 (m, 3H), 1.01 (d, 18H, *J* = 6.6 Hz), 0.96 (d, 3H, *J* = 6.8 Hz), 0.92 (d, 3H, *J* = 6.8 Hz).

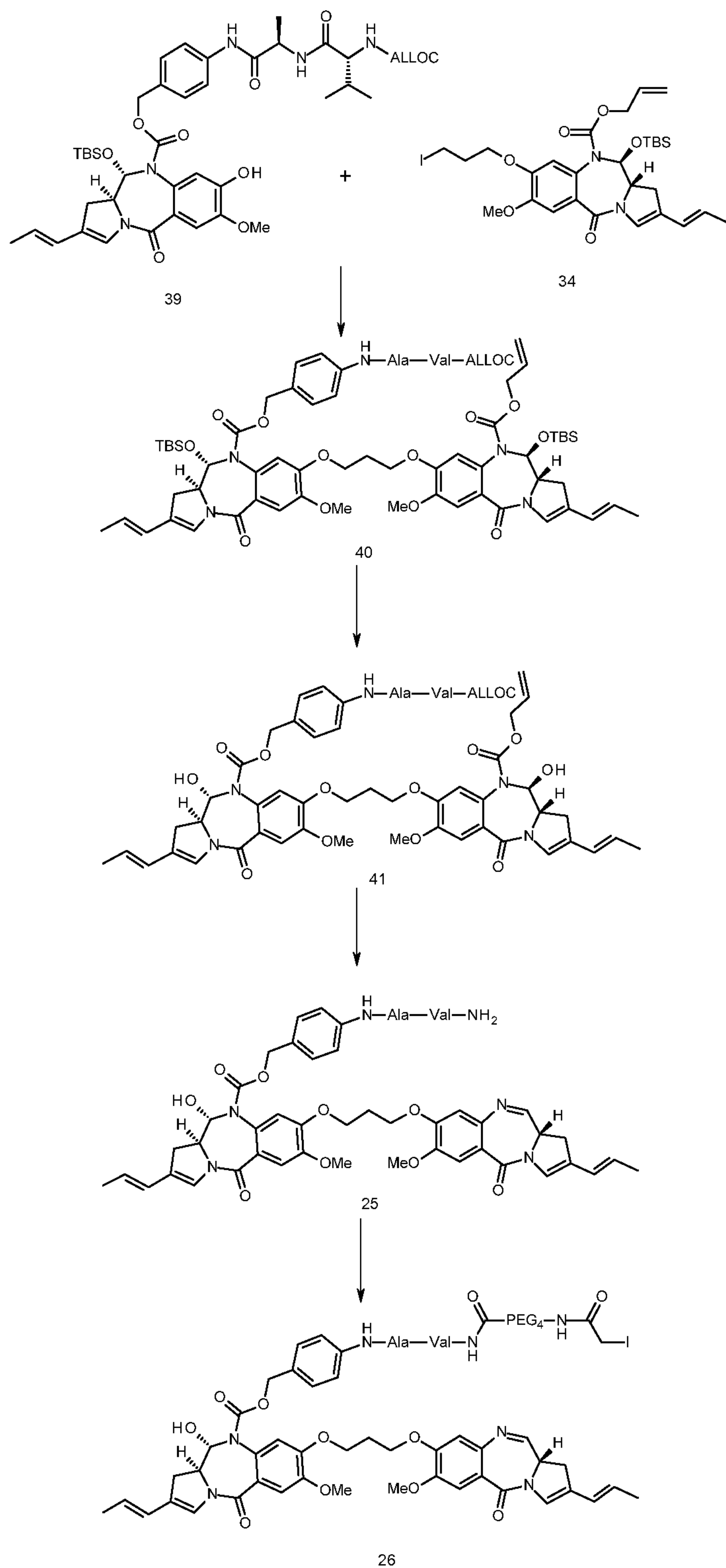
(*d*) (11*S*,11*aS*)-4-((*S*)-2-((*S*)-2-(allyloxycarbonylamino)-3-methylbutanamido)propanamido)benzyl 11-(*tert*-butyldimethylsilyloxy)-7-methoxy-5-oxo-2-((*E*)-prop-1-enyl)-8-(triisopropylsilyloxy)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**38**)

Tert-butyldimethylsilyltrifluoromethane sulfonate (0.46 g, 1.74mmol, 3.0 eq) was added to a solution of secondary alcohol **37** (0.5 g, 0.58 mmol, 1.0 eq) and 2,6-lutidine (0.25 g, 2.32 mmol, 4.0 eq) in dry DCM (10 mL) at 0°C under an atmosphere of nitrogen. After 10 minutes, the reaction mixture was allowed to warm to room temperature and stirred for a further 120 mins. The organic phase was then washed successively with water (10 mL), saturated sodium bicarbonate (10 mL) and brine (5 mL), dried over MgSO₄, filtered and concentrated under reduced pressure. The residue was purified by flash chromatography (silica gel, 50/50 v/v *n*-hexane/EtOAc) to afford the product as a white solid (0.320 g, 57 %). LC/MS (4.415 min (ES⁺)), *m/z*: 976.52 [M+H]⁺. ¹H NMR (400 MHz, CDCl₃) δ 8.31 (br s, 1H), 7.48 (d, 2H, *J* = 8.0 Hz), 7.21 (s, 1H), 7.14 (d, 2H, *J* = 8.3 Hz), 6.89 (s, 1H), 6.65 (s, 1H), 6.38 (d, 1H, *J* = 7.3 Hz), 6.25 (d, 1H, *J* = 14.6 Hz), 5.93 (m, 1H), 5.85 (d, 1H, *J* = 8.8 Hz), 5.50 (m, 1H), 5.34 (m, 1H), 5.24 (m, 2H), 5.15 (d, 1H, *J* = 12.5 Hz), 4.86 (d, 1H, *J* = 12.2 Hz), 4.62 (m, 3H), 4.01 (m, 1H), 3.86 (s, 3H), 3.78 (m, 1H), 3.04 (m, 1H), 2.56 (m, 1H), 2.20 (m, 1H), 1.84 (dd, 3H, *J* = 6.6, 0.7 Hz), 1.48 (d, 3H, *J* = 6.8 Hz), 1.20 (m, 3H), 1.05 (d, 9H, *J* = 2.9 Hz), 1.03 (d, 9H, *J* = 2.9 Hz), 0.99 (d, 3H, *J* = 6.8 Hz), 0.95 (d, 3H, *J* = 6.8 Hz), 0.88 (s, 9H), 0.27 (s, 3H), 0.14 (s, 3H).

(e) (11*S*,11*aS*)-4-((*S*)-2-((*S*)-2-(allyloxycarbonylamino)-3-methylbutanamido)propanamido)benzyl 11-(*tert*-butyldimethylsilyloxy)-8-hydroxy-7-methoxy-5-oxo-2-((*E*)-prop-1-enyl)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**39**)

Lithium acetate dihydrate (0.010 g, 0.10 mmol, 1.0 eq) was added to a solution of the silyl ether **38** (0.100 g, 0.10 mmol, 1.0 eq) in wet DMF (2 mL) at 25°C for 3 hours. The reaction mixture was then diluted with ethyl acetate (20 mL) and washed successively with 0.1M citric acid (20 mL, pH 3), water (20 mL) and brine (5 mL), dried over MgSO₄, filtered and concentrated under reduced pressure. The residue was purified by flash chromatography (silica gel, 5/95 v/v methanol/DCM) to afford the product as a pale yellow oil (0.070 g, 83 %). LC/MS (3.362 min (ES⁺)), *m/z*: 820.2 [M+H]⁺. ¹H NMR (400 MHz, CDCl₃) δ 8.39 (s, 1H), 7.48 (d, 2H, *J* = 8.2 Hz), 7.25 (s, 1H), 7.12 (d, 2H, *J* = 8.1 Hz), 6.88 (s, 1H), 6.68 (s, 1H), 6.47 (d, 1H, *J* = 7.6 Hz), 6.24 (d, 1H, *J* = 15.2 Hz), 6.03 (s, 1H), 5.92 (m, 1H), 5.84 (d, 1H, *J* = 8.9 Hz), 5.50 (m, 1H), 5.34 (m, 1H), 5.26 (m, 2H), 5.18 (d, 1H, *J* = 12.3 Hz), 4.80 (d, 1H, *J* = 12.4 Hz), 4.66 – 4.60 (m, 3H), 4.02 (m, 1H), 3.95 (s, 3H), 3.81 (m, 1H), 3.03 (m, 1H), 2.57 (m, 1H), 2.19 (m, 1H), 1.84 (dd, 3H, *J* = 6.8, 0.8 Hz), 1.48 (d, 3H, *J* = 7.1 Hz), 1.00 (d, 3H, *J* = 6.8 Hz), 0.95 (d, 3H, *J* = 6.8 Hz), 0.87 (s, 9H), 0.26 (s, 3H), 0.12 (s, 3H).

(iv) (11*S*,11*aS*)-4-((20*S*,23*S*)-1-iodo-20-isopropyl-23-methyl-2,18,21-trioxo-6,9,12,15-tetraoxa-3,19,22-triazatetracosanamido)benzyl 11-hydroxy-7-methoxy-8-(3-((*S*)-7-methoxy-5-oxo-2-((*E*)-prop-1-enyl)-5,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepin-8-yloxy)propoxy)-5-oxo-2-((*E*)-prop-1-enyl)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**26, A**)



(a) (11*S*,11*aS*)-allyl 8-(3-((11*S*,11*aS*)-10-((4-((*R*)-2-((*R*)-2-(allyloxycarbonylamino)-3-methylbutanamido)propanamido)benzyloxy)carbonyl)-11-(*tert*-butyldimethylsilyloxy)-7-methoxy-5-oxo-2-((*E*)-prop-1-enyl)-5,10,11,11*a*-tetrahydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepin-8-yloxy)propoxy)-11-(*tert*-butyldimethylsilyloxy)-7-methoxy-5-oxo-2-((*E*)-prop-1-enyl)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**40**)

Potassium carbonate (0.030 g, 0.21 mmol, 1.0 eq) was added to a solution of the phenol **39** (0.175 g, 0.21 mmol, 1.0 eq) and the iodo linker **34** (0.214 g, 0.32 mmol, 1.5 eq) in acetone (10 mL). The reaction mixture was heated under a nitrogen atmosphere at 75°C in a sealed flask for 17 hours. The reaction mixture was concentrated to dryness under reduced pressure and purified by flash chromatography (silica gel, 2/98 v/v methanol/DCM to 5/95 v/v methanol/DCM) to afford the product as a pale yellow solid (0.100 g, 35%). LC/MS (4.293 min (ES⁺)), *m/z*: 1359.13 [M]⁺.

(b) (11*S*,11*aS*)-allyl 8-(3-((11*S*,11*aS*)-10-((4-((*R*)-2-((*R*)-2-(allyloxycarbonylamino)-3-methylbutanamido)propanamido)benzyloxy)carbonyl)-11-hydroxy-7-methoxy-5-oxo-2-((*E*)-prop-1-enyl)-5,10,11,11*a*-tetrahydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepin-8-yloxy)propoxy)-11-hydroxy-7-methoxy-5-oxo-2-((*E*)-prop-1-enyl)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**41**)

Tetra-*n*-butylammonium fluoride (1M, 0.22 mL, 0.22 mmol, 2.0 eq) was added to a solution of silyl ether **40** (0.150 g, 0.11 mmol, 1.0 eq) in dry THF (2 mL). The reaction mixture was stirred at room temperature for 20 minutes, after which LC/MS indicated complete reaction. The reaction mixture was diluted with ethyl acetate (10 mL) and washed sequentially with water (5 mL) and brine (5 mL). The organic phase was dried over MgSO₄, filtered and concentrated under reduced pressure to leave a yellow solid. Purification by flash chromatography (silica gel, 6/94 v/v methanol/DCM to 10/90 v/v methanol/DCM) afforded the product as a pale yellow solid (0.090 g, 73%). LC/MS (2.947 min (ES⁺)), *m/z*: 1154.0 [M+Na]⁺. ¹H NMR (400 MHz, CDCl₃) δ 8.39 (br s, 1H), 7.39 (d, 2H, *J* = 7.6 Hz), 7.18 (d, 2H, *J* = 10.6 Hz), 7.10 (m, 3H), 6.86 (d, 2H, *J* = 10.0 Hz), 6.74 (s, 1H), 6.55 (s, 1H), 6.22 (dd, 2H, *J* = 15.3, 6.6 Hz), 5.85 (m, 2H), 5.74 (m, 3H), 5.52 (m, 2H), 5.22 (m, 1H), 5.00 (m, 2H), 4.57 (m, 6H), 4.41 (m, 2H), 4.09 (m, 4H), 3.85 (m, 11H), 3.06 (m, 2H), 2.76 (m, 2H), 2.20 (m, 2H), 2.08 (m, 1H), 1.79 (d, 6H, *J* = 6.4 Hz), 1.40 (d, 3H, *J* = 6.1 Hz), 0.90 (m, 6H).

(c) (11*S*,11*aS*)-4-((*R*)-2-((*R*)-2-amino-3-methylbutanamido)propanamido)benzyl 11-hydroxy-7-methoxy-8-(3-((*S*)-7-methoxy-5-oxo-2-((*E*)-prop-1-enyl)-5,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepin-8-yloxy)propoxy)-5-oxo-2-((*E*)-prop-1-enyl)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**25**)

5 Tetrakis(triphenylphosphene)palladium(0) (0.005 g, 0.005 mmol, 0.06 eq) was added to a solution of the bis-carbinolamine **41** (0.090 g, 0.08 mmol, 1.0 eq) and pyrrolidine (16 μ L, 0.20 mmol, 2.5 eq) in dry DCM (5 mL). After 20 minutes, the reaction mixture was diluted with DCM (10 mL) and washed sequentially with saturated ammonium chloride (5 mL) and brine (5 mL), dried over MgSO₄, filtered and the solvent was removed under reduced
10 pressure to leave the crude product as a pale yellow solid which was used in the next step without further purification (0.075 g, assumed 100% yield). LC/MS (2.060 min (ES⁺)), *m/z*: 947.2 [M+H]⁺.

(d) (11*S*,11*aS*)-4-((20*S*,23*S*)-1-iodo-20-isopropyl-23-methyl-2,18,21-trioxo-6,9,12,15-
15 tetraoxa-3,19,22-triazatetracosanamido)benzyl 11-hydroxy-7-methoxy-8-(3-((*S*)-7-methoxy-5-oxo-2-((*E*)-prop-1-enyl)-5,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepin-8-yloxy)propoxy)-5-oxo-2-((*E*)-prop-1-enyl)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**26, A**)

EDCI (0.015 g, 0.08 mmol, 1.0 eq) was added to a solution of amine **25** (assumed 100%
20 yield 0.075 g, 0.08 mmol, 1.0 eq) and iodoacetamide-PEG₄-acid **17** (0.034 g, 0.08 mmol, 1.0 eq) in dry dichloromethane (5 mL) and the reaction was stirred in the dark. After 50 minutes, a further amount of iodoacetamide-PEG₄-acid **17** (0.007 g, 0.016 mmol, 0.2 eq) was added along with a further amount of EDCI (0.003 g, 0.016 mmol, 0.2 eq). After a total of 2.5 hours, the reaction mixture was diluted with dichloromethane (15 mL) and washed sequentially with
25 water (10 mL) and brine (10 mL). The organic phase was dried over MgSO₄, filtered and concentrated under reduced pressure. The resulting residue was purified by flash chromatography (silica gel, Chloroform 100% to 90:10 v/v Chloroform:Methanol). Pure fractions were combined to afford the product (0.0254 g, 23%, 2 steps). The crude fractions were collected and purified by preparative TLC (silica gel, 90:10 v/v Chloroform:Methanol) to
30 afford a second batch of product (0.0036 g, 3%, 2 steps). LC/MS (2.689 min (ES⁺)), *m/z*: 681.0 1/2[M+2H]⁺.

General Experimental Methods for Example 4

Reaction progress was monitored by thin-layer chromatography (TLC) using Merck Kieselgel 60 F254 silica gel, with fluorescent indicator on aluminium plates. Visualisation of TLC was achieved with UV light or iodine vapour unless otherwise stated. Flash chromatography was performed using Merck Kieselgel 60 F254 silica gel. Extraction and chromatography solvents were bought and used without further purification from Fisher Scientific, U.K. All chemicals were purchased from Aldrich, Lancaster or BDH.

Proton NMR chemical shift values were measured on the delta scale at 400 MHz using a Bruker AV400. The following abbreviations have been used: s, singlet; d, doublet; t, triplet; q, quartet; quin, quintet; m, multiplet; br, broad. Coupling constants are reported in Hz.

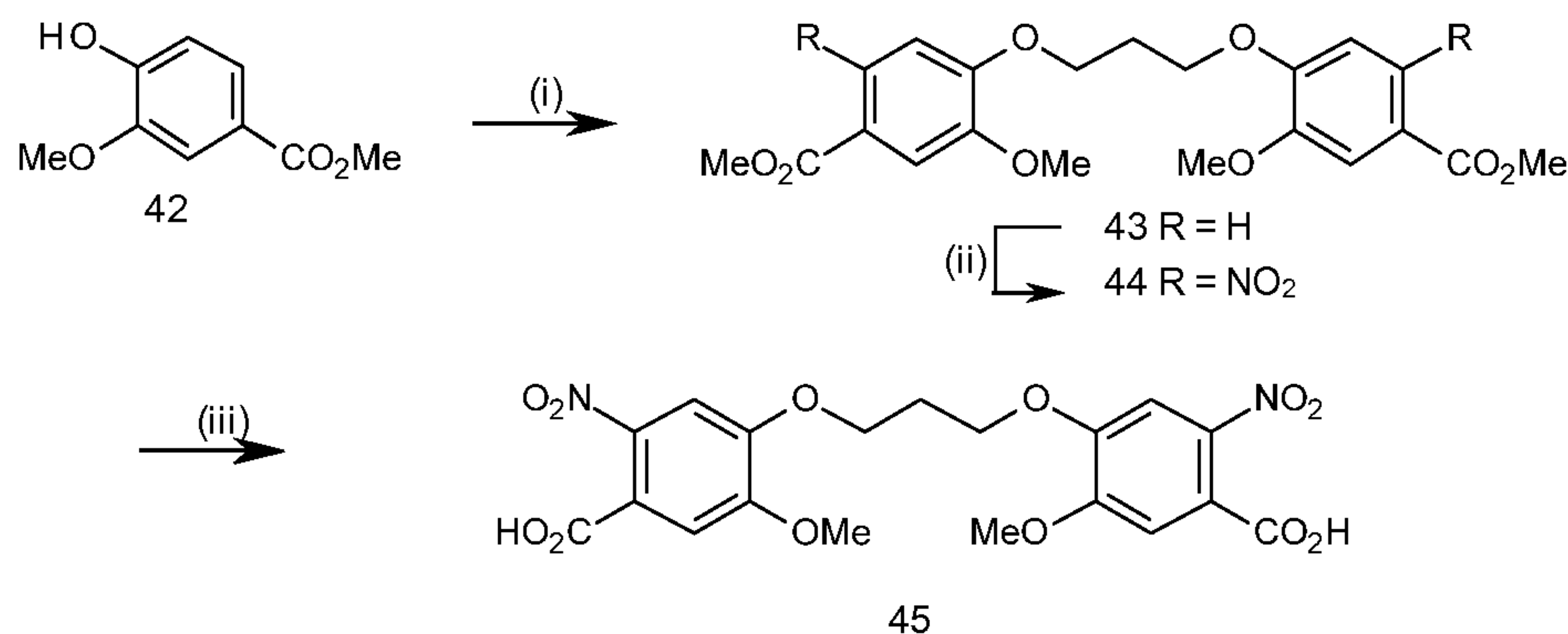
Unless otherwise stated, column chromatography (by the flash procedure) were performed on Merck Kieselgel silica (Art. 9385). Mass spectroscopy (MS) data were collected using a Waters Micromass LCT instrument coupled to a Waters 2795 HPLC separations module.

Thin Layer Chromatography (TLC) was performed on silica gel aluminium plates (Merck 60, F₂₅₄). All other chemicals and solvents were used as supplied without further purification.

LCMS data were obtained using a Shimadzu Nexera™ series LC/MS with a Shimadzu LCMS-2020 quadrupole MS, with Electrospray ionisation. Mobile phase A - 0.1% formic acid in water. Mobile phase B - 0.1% formic acid in acetonitrile. Flow rate of 0.80ml/min. Gradient from 5% B rising up to 100% B over 2.00 min, remaining at 100% B for 0.50 min and then back down to 5% B over 0.05 min (held for 0.45 min). The total run time is 3 min. Column: Waters Aquity UPLC BEH Shield RP18 1.7 µm, 2.1 x 50mm; (System 1).

Or, gradient from 5% B rising up to 100% B over 10.00 min, remaining at 100% B for 2.00 min and then back down to 5% B over 0.10 minutes (held for 2.90 min). The total run time is 15 minutes. Column: Gemini-NX 3u C18 110A, 100 x 2.00mm; (System 2).

Chromatograms based on UV detection at 254nm. Mass Spectra were achieved using the MS in positive mode.

Example 4**(a) 1',3'-Bis(4-carboxy-2-methoxy-5-nitrophenoxy) propane (45)**

5

(i) 1',3'-Bis[2-methoxy-4-(methoxycarbonyl)phenoxy]propane (43)

Diisopropyl azodicarboxylate (71.3 mL, 73.2 g, 362 mmol) was added drop-wise over a period of 60 minutes to an overhead stirred solution of methyl vanillate **42** (60.0 g, 329 mmol) and Ph₃P (129.4 g, 494 mmol) in anhydrous THF (800 mL) at 0-5 °C (ice/acetone) under a nitrogen atmosphere. The reaction mixture was allowed to stir at 0-5 °C for an additional 1 hour after which time a solution of 1,3-propanediol (11.4 mL, 12.0 g, 158 mmol) in THF (12 mL) was added drop-wise over a period of 20 minutes. The reaction mixture was allowed to warm to room temperature and stirred for 5 days. The resulting white precipitate **43** was collected by vacuum filtration, washed with THF and dried in a vacuum desiccator to constant weight. Yield = 54.7 g (84% based on 1,3-propanediol). Purity satisfactory by LC/MS (3.20 min (ES+) *m/z* (relative intensity) 427 ([*M* + Na]⁺, 10); ¹H NMR (400 MHz, CDCl₃) δ 7.64 (dd, 2H, *J* = 1.8, 8.3 Hz), 7.54 (d, 2H, *J* = 1.8 Hz), 6.93 (d, 2H, *J* = 8.5 Hz), 4.30 (t, 4H, *J* = 6.1 Hz), 3.90 (s, 6H), 3.89 (s, 6H), 2.40 (p, 2H, *J* = 6.0 Hz).

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(ii) 1',3'-Bis[2-methoxy-4-(methoxycarbonyl)-5-nitrophenoxy]propane (44)

Solid Cu(NO₃)₂·3H₂O (81.5 g, 337.5 mmol) was added slowly to an overhead stirred slurry of the bis-ester **43** (54.7 g, 135 mmol) in acetic anhydride (650 mL) at 0-5 °C (ice/acetone). The reaction mixture was allowed to stir for 1 hour at 0-5 °C and then allowed to warm to room temperature. A mild exotherm (ca. 40-50 °C), accompanied by thickening of the mixture and evolution of NO₂ was observed at this stage. Additional acetic anhydride (300 mL) was added and the reaction mixture was allowed to stir for 16 hours at room temperature. The reaction mixture was poured on to ice (~ 1.5 L), stirred and allowed to return to room temperature. The resulting yellow precipitate was collected by vacuum filtration and dried in a desiccator to afford the desired *bis*-nitro compound **44** as a yellow

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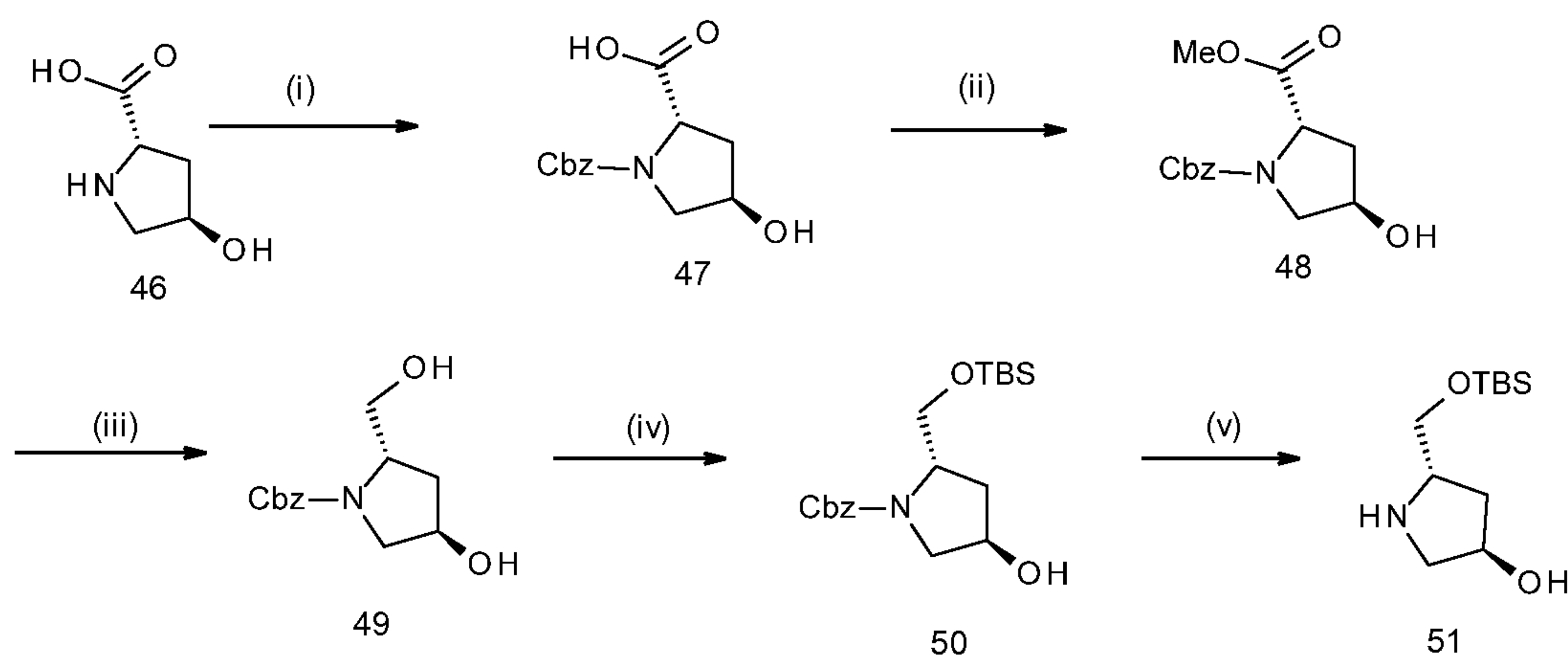
solid. Yield = 66.7 g (100%). Purity satisfactory by LC/MS (3.25 min (ES+) m/z (relative intensity) 517 ($[M + Na]^+$, 40); ^1H NMR (400 MHz, $CDCl_3$) δ 7.49 (s, 2H), 7.06 (s, 2H), 4.32 (t, 4H, $J = 6.0$ Hz), 3.95 (s, 6H), 3.90 (s, 6H), 2.45–2.40 (m, 2H).

5 (iii) *1',3'-Bis(4-carboxy-2-methoxy-5-nitrophenoxy) propane (45)*

A slurry of the methyl ester **44** (66.7 g, 135 mmol) in THF (700 mL) was treated with 1N NaOH (700 mL) and the reaction mixture was allowed to stir vigorously at room temperature. After 4 days stirring, the slurry became a dark coloured solution which was subjected to rotary evaporation under reduced pressure to remove THF. The resulting aqueous residue
10 was acidified to pH 1 with concentrated HCl and the colourless precipitate **45** was collected and dried thoroughly in a vacuum oven (50°C). Yield = 54.5 g (87%). Purity satisfactory by LC/MS (2.65 min (ES+) m/z (relative intensity) 489 ($[M + Na]^+$, 30)); ^1H NMR (400 MHz, DMSO- d_6) δ 7.62 (s, 2H), 7.30 (s, 2H), 4.29 (t, 4H, $J = 6.0$ Hz), 3.85 (s, 6H), 2.30–2.26 (m, 2H).

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(b) *(2S,4R)-2-t-Butyldimethylsilyloxymethyl-4-hydroxypyrrolidine (51)*



(i) *(2S,4R)-N-(Benzyloxycarbonyl)-2-carboxy-4-hydroxypyrrolidine (47)*

A solution of benzyl chloroformate (12.5 mL, 14.9 g, 87.5 mmol) in toluene (40 mL) was added to a solution of *trans*-4-hydroxy-L-proline **46** (10 g, 76.3 mmol) and $NaHCO_3$ (16 g, 190 mmol) in H_2O (165 mL) over a period of 15 minutes. After stirring at room temperature for 12 hours, the two phases were allowed to separate. The aqueous phase was washed with diethyl ether (4 x 50 mL), cooled in an ice bath, and then acidified to pH 2 with conc. HCl. The aqueous layer was extracted with ethyl acetate (5 x 50 mL) and the combined
20 organic extracts were dried ($MgSO_4$) and the excess solvent evaporated *in vacuo* to afford colourless viscous oil **47** (20.30 g, 100%). $[\alpha]_D^{27} = -565^\circ$ ($c = 0.1$, MeOH);

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(ii) (2S,4R)-N-(Benzyloxycarbonyl)-2-methoxycarbonyl-4-hydroxypyrrolidine (48)

A catalytic amount of concentrated H₂SO₄ (1.1 mL) was added to a solution of the (2S,4R)-N-(benzyloxycarbonyl)-2-carboxy-4-hydroxypyrrolidine **47** (80 g, 302 mmol) in anhydrous MeOH (462 mL) at 10 °C. The reaction mixture was refluxed for 3 hours, allowed to cool to room temperature, and then treated with Et₃N (66 mL) and stirred for 1 hour. Following removal of excess MeOH by rotary evaporation under reduced pressure the residue was dissolved in EtOAc (460 mL), washed with saturated brine, and dried (MgSO₄). Filtration and removal of excess solvent *in vacuo* afforded ester **48** as a viscous oil. Yield 84.4 g (100%).

[α]_D²⁰ = -59.4° (c = 0.014, CHCl₃);

(iii) (2S,4R)-N-(Benzyloxycarbonyl)-2-hydroxymethyl-4-hydroxypyrrolidine (49)

Solid LiBH₄ (9.89 g, 454 mmol) was added portion wise to a solution of the ester **48** (84.4 g, 302 mmol) in anhydrous THF (600 mL) at 0 °C. After the sample was stirred for 30 min at 0 °C, the reaction mixture was allowed to warm to room temperature and then stirred for a further 2 hours at room temperature. During this time, a thick suspension formed despite the addition of further THF (150 mL); TLC (EtOAc) at this point revealed the complete disappearance of starting material. The suspension was cooled to 0 °C, diluted with water (389 mL), and treated dropwise with aqueous HCl (2 M, 400 mL) provoking vigorous effervescence. Excess THF was removed by rotary evaporation under reduced pressure, and the aqueous residue was extracted with EtOAc (4 x 250 mL). The combined organic phase was washed with brine (300 mL) and dried over MgSO₄. Filtration and solvent removal afforded the product **49** as a colorless oil. Yield 73.2 g (96%). [α]_D²⁶ = -42.5° (c = 1, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.45 – 7.29 (m, 5H), 5.26 – 5.01 (m, 2H), 4.72 (d, J = 7.4 Hz, 1H), 4.50 – 4.30 (m, 1H), 4.27 – 4.03 (m, 1H), 3.84 – 3.68 (m, 1H), 3.60 (d, J = 9.2 Hz, 2H), 3.47 (dd, J = 12.1, 3.8 Hz, 1H), 2.56 (s, 1H), 2.14 – 1.93 (m, 1H), 1.80 – 1.62 (m, 1H).

(iv) (2S,4R)-N-(Benzyloxycarbonyl)-2-t-butyldimethylsilyloxymethyl-4-hydroxypyrrolidine (50)

t-Butyldimethylsilyl chloride (35.9 g, 238 mmol) was added to a solution of the diol **49** (77.8 g, 310 mmol), Et₃N (43.7 mL, 31.7 g, 313 mmol), and DBU (9.17 mL, 9.34 g, 61.3 mmol) in freshly distilled anhydrous CH₂Cl₂ (650 mL). The reaction mixture stirred overnight at room temperature under a N₂ atmosphere. The reaction mixture was washed with saturated aqueous NH₄Cl (300 mL), brine (300 mL), and then dried (MgSO₄). Filtration and removal of excess solvent furnished the crude product, which was subjected to flash column chromatography (80/20 v/v hexane/ ethyl acetate to remove the *bis* protected material,

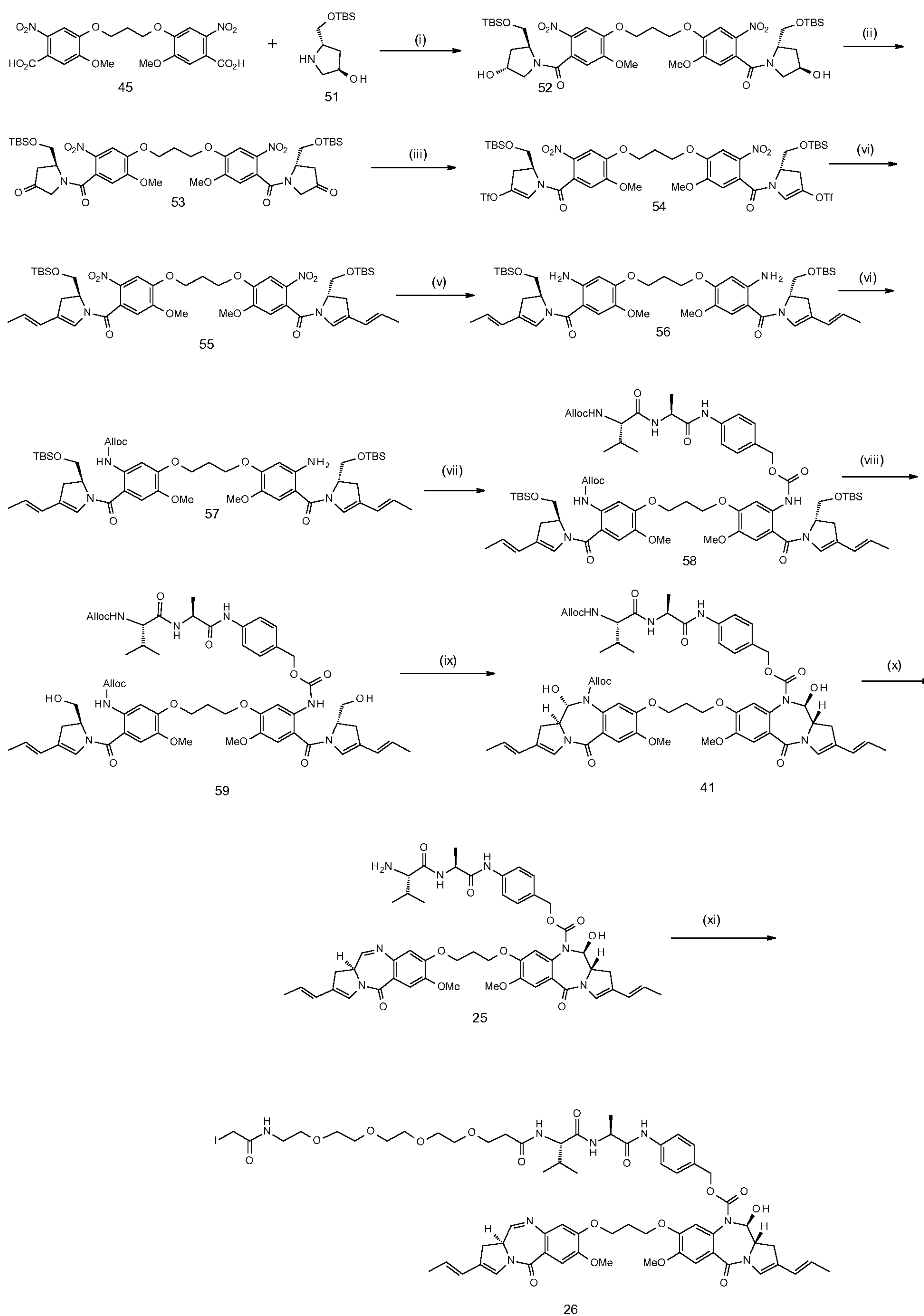
followed by 50/50 v/v hexane/ ethyl acetate to elute the desired product, and then pure EtOAc to recover the starting material) to isolate the silyl ether **50**. Yield: 67.9 g (78% based on *t*-butyldimethylsilylchloride). $[\alpha]_D^{25} = -47.5^\circ$ ($c = 1$, CHCl_3);

5 (v) (2*S*,4*R*)-2-*t*-Butyldimethylsilyloxymethyl-4-hydroxypyrrolidine (**51**)

A solution of the silyl ether **50** (100 g, 273 mmol) in isopropanol (4 x 200 mL) over 10% Pd/C (5% w/w, 5 g) was hydrogenated under pressure (45 psi) on a Parr apparatus for 30 minutes. The reaction mixture was filtered through Celite to remove the Pd/C, and the filter pad was rinsed with isopropanol. Excess solvent was removed by rotary evaporation under reduced pressure. The residue was re-dissolved in ethyl acetate and the solvent removed under reduced pressure. This ethyl acetate cycle was repeated once again, followed by drying under high vacuum. The product, **51**, crystallised as a white solid 57 g (90%). $[\alpha]_D^{22} = +35.6^\circ$ ($c = 0.042$, CHCl_3).

15 (c) (11*S*,11*aS*)-4-((20*S*,23*S*)-1-iodo-20-isopropyl-23-methyl-2,18,21-trioxo-6,9,12,15-tetraoxa-3,19,22-triazatetracosanamido)benzyl 11-hydroxy-7-methoxy-8-(3-(((*S*)-7-methoxy-5-oxo-2-((*E*)-prop-1-en-1-yl)-5,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepin-8-yl)oxy)propoxy)-5-oxo-2-((*E*)-prop-1-en-1-yl)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate, **26**

153



(i) (S,R)-(4,4'-(propane-1,3-diylbis(oxy))bis(5-methoxy-2-nitro-4,1-phenylene))bis(((2S,4R)-2-((tert-butyldimethylsilyloxy)methyl)-4-hydroxypyrrolidin-1-yl)methanone) (**52**)

5 Anhydrous DMF (approx. 0.5 mL, ~0.13equ.) was added dropwise to a stirred suspension of 4,4'-(propane-1,3-diylbis(oxy))bis(5-methoxy-2-nitrobenzoic acid) (**45**) (23.2g, 49.9mmol) and oxalyl chloride (12.7 mL, 150 mmol, 3.0 equ.) in anhydrous DCM (310 mL) until vigorous effervescence occurred. Addition of the same amount of anhydrous DMF was repeated 2 hours later and the reaction mixture was left to stir overnight. The reaction mixture was
10 evaporated to dryness, and triturated with diethyl ether. The resulting yellow precipitate was filtered from solution, washed with diethyl ether (100 mL) and immediately added to a solution of (3R,5S)-5-((tert-butyldimethylsilyloxy)methyl) pyrrolidin-3-ol **51** and anhydrous triethylamine (26.5 g, 115 mmol, 2.3 equ.) in anhydrous DCM (200 mL) at -40°C. The resulting thick suspension was diluted with anhydrous DCM (140 mL) and was allowed to
15 slowly warm to room temperature (over 90 minutes) after which, LCMS analysis indicated complete reaction. DCM (300 mL) was added and the mixture was transferred into a separating funnel. The organic layer was washed successively with 0.1M HCl (2 x 240 mL), saturated NaHCO₃ (240 mL) and brine 120 mL). After drying over MgSO₄ and filtration, evaporation of the solvent left the product, **52**, as a mustard coloured foam (42.6 g, 96%).
20 δ_H (CDCl₃): 7.64 (2H, s), 6.73 (2H, s), 4.47-4.52 (2H, m), 4.39 (2H, s (br)), 4.31-4.34 (2H, t, J=5.6), 4.12 (2H, m), 3.93-3.95 (2H, t, J=3.8), 3.92 (6H, s), 3.73-3.75 (2H, m), 3.27-3.31 (2H, m), 2.96-3.00 (2H, m), 2.55-2.56 (2H, m), 2.39-2.43 (4H, m), 2.27-2.33 (2H, m), 2.04-2.10 (2H, m), 0.89 (18H, s), 0.09 (6H, s), 0.07 (6H, s). LC/MS, System 1: 96% pure at 1.96 min (ES+) m/z (relative intensity) 893.35 ([M+H]⁺, 100).

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(ii) (S,R)-(4,4'-(propane-1,3-diylbis(oxy))bis(5-methoxy-2-nitro-4,1-phenylene))bis((S)-5-((tert-butyldimethylsilyloxy)methyl)-1-(4,5-dimethoxy-2-nitrobenzoyl)pyrrolidin-3-one) (**53**)

Trichloroisocyanuric acid (8.1 g, 0.035 mol, 2.2 eq) was added in one portion to a solution of diol **52** (14.2 g, 0.016 mol, 1.0 eq) and TEMPO (0.5 g, 0.035 mmol, 0.2 eq) in anhydrous
30 DCM (100 mL) under an atmosphere of nitrogen at 0°C. The reaction mixture was stirred at 0°C for 20 minutes after which, LCMS analysis of the reaction mixture showed complete reaction. The reaction mixture was diluted with DCM (100 mL) and the cold mixture was poured into saturated sodium bicarbonate (200 mL). The mixture was stirred vigorously for about 30 minutes, then transferred into a separating funnel. The organic layer was removed
35 and washed with 0.2M sodium thiosulfate (200 mL), brine (25 mL) and dried (MgSO₄).

Evaporation of the solvent gave the crude product **53** as a pale yellow solid (14.1 g, 100 %). The product was used without further purification in the next step. δ_{H} (CDCl_3): 7.76 (0.6h, s), 7.74 (2H, s), 7.74 (0.6H, s), 6.82 (0.6H, s), 6.74 (0.6H, s), 6.73 (2H, s), 4.95-4.98 (2H, d (br), J=8.9), 4.36 (1H, d, J=18.4) 4.27-4.35 (10H, m), 3.96 (1.8H, s), 3.94 (6H, s), 3.94 (1.8H, s),
 5 3.62-3.66 (1H, d, J=18.9), 3.70-3.73 (3H, dd, J=10.3, 1.9), 3.62-3.66 (3H, d, J=17), 3.42-3.46 (3.6H, d, J=17), 2.75-2.82 (4H, dd, J=17.8, 9.8), 2.52-2.57 (3H, d, J=17.8), 2.43-2.47 (4.6H, m), 0.86 (18H, s), 0.85 (8H, s), 0.10 (6H, s), 0.07 (6H, s), 0.02 (2H, s), 0.00 (2H, s). LC/MS, System 1: 100% pure at 2.05 min (ES+) m/z (relative intensity) 889.20 ($[\text{M}+\text{H}]^+$, 100).

10 (iii) (5*S*,5'*S*)-1,1'-(4,4'-(propane-1,3-diylbis(oxy))bis(5-methoxy-2-nitrobenzoyl))bis(5-(((tert-butyl)dimethylsilyl)oxy)methyl)-4,5-dihydro-1*H*-pyrrole-3,1-diyl) bis(trifluoromethanesulfonate), (**54**)

Triflic anhydride (22.84 g, 0.08 mol, 4.0 eq) was added dropwise to a vigorously stirred solution of crude bis-ketone **53**¹ (18.0 g, 0.02 mol, 1 eq) in anhydrous DCM (350 mL)
 15 containing 2,6-lutidine (13.0 g, 0.12 mol, 6.0 eq) at -50°C. The reaction mixture was stirred at -50°C for 1.5 hours after which, LCMS analysis indicated complete reaction. The organic phase was washed successively with 5% citric acid (300 mL), saturated sodium bicarbonate (400 mL) and brine (50 mL), dried over MgSO_4 and evaporated to leave a brown oil, **54** (23.0g, 99%) which was used in the next step without further purification. δ_{H} (CDCl_3): 7.76
 20 (2H, s), 6.77 (1H, s), 6.08 (2H, m), 4.78 (1H, m), 4.34-4.37 (4H, t, J=5.8), 3.92-3.97 (4H, m), 3.95 (6H, s), 3.14-3.19 (2H, ddd, J=16.5, 10.5, 2.5), 2.96-3.02 (2H, ddd, J=16.2, 4.0, 1.5), 2.45-2.48 (2H, quin=5.8), 0.91 (18H, s), 0.11 (6H, s), 0.11 (6H, s).

25 (iv) (*S,E*)-(4,4'-(propane-1,3-diylbis(oxy))bis(5-methoxy-2-nitro-4,1-phenylene))bis(((*S*)-2-(((tert-butyl)dimethylsilyloxy)methyl)-4-((*E*)-prop-1-enyl)-2,3-dihydro-1*H*-pyrrol-1-yl)methanone) (**55**)

Tetrakis(triphenylphosphine)palladium (2.3 g, 1.99 mmol, 0.1 equ.) was added to a mixture of crude bis-triflate **54** (23.0 g, 19.9mmol), potassium phosphate (16.9 g, 79.6 mmol, 4.0 equ.) and trans-1-propene-1-ylboronic acid (4.28 g, 49.8 mmol, 2.5 equ.) in dry dioxane (200
 30 mL) under nitrogen. The mixture was stirred at room temperature for 3 hours after which, LCMS analysis showed formation of the desired product along with mono-Suzuki coupled product. A further 10% catalyst was added and the reaction mixture was stirred overnight after which, LCMS analysis showed completion of the reaction. Water (100 mL) was added and the product was extracted with ethyl acetate (150 mL), washed with brine (50 mL) and
 35 dried over MgSO_4 . Evaporation of the solvent left a brown oil which was purified by flash chromatography (1% v/v methanol / dichloromethane) to afford product **55** as an orange

solid. (11.4 g, 54.3 % over 3 steps from diol **52**): δ_{H} (CDCl_3): 7.75 (2H, s), 6.79 (2H, s), 5.94-5.98 (2H, d (br), $J=15.6$), 5.73 (2H, s (br), 5.51-5.99 (2H, dq, $J=15.6, 6.6$), 4.69 (2H, m), 4.33-4.36 (4H, t, $J=5.9$), 3.76-4.15 (4H, m), 3.94 (6H, s), 2.86-2.92 (2H, m), 2.73-2.76 (2H, d (br), $J=16.0$), 2.43-2.49 (2H, quin, $J=5.9$), 1.74-1.76 (6H, dd, $J=6.6, 0.7$), 0.89 (18H, s), 0.11 (6H, s), 0.10 (6H, s). LC/MS, System 1: 100% pure at 2.36 min (ES+) m/z (relative intensity) 1896.85 ($[\text{2M}+\text{Na}]^+$, 100).

(v) *(S,E)-(4,4'-(propane-1,3-diylbis(oxy))bis(2-amino-5-methoxy-4,1-phenylene))bis(((S)-2-((tert-butyldimethylsilyloxy)methyl)-4-((E)-prop-1-enyl)-2,3-dihydro-1H-pyrrol-1-yl)methanone)* (**56**)

Zinc dust (39.9 g, 610 mmol, 36.0 eq) was added in one portion to a solution of bis-nitro compound **55** (15.9 g, 17.0 mmol, 1.0 eq) in 5% formic acid / methanol (350 mL) keeping the temperature between 25-30°C with the aid of a cold water bath. The reaction was stirred at 30°C for 20 minutes after which, LCMS showed complete reaction. The reaction mixture was filtered through a short bed of celite to remove the excess zinc, which was washed with ethyl acetate (750 mL). The organic fractions were washed with water (750 mL), saturated sodium bicarbonate (750 mL) and brine (50 mL), dried over MgSO_4 and evaporated to leave a brown oil. The product was purified by flash chromatography (DCM:MeOH 96%) to afford the bis-aniline **56** as a pale yellow solid (9.46g, 63.9 %). δ_{H} ($\text{DMSO}-d_6$): 6.64 (2H, s), 6.46 (2H, s (br)), 6.43 (2H, s), 6.17-6.21 (2H, d (br), $J=15$), 5.39-5.48 (2H, dq, $J=15, 6.5$), 5.14 (4H, s), 4.50-4.52 (2H, m), 4.05-4.08 (4H, t, $J=6.0$), 3.67-3.83 (4H, m), 3.62 (6H, s), 2.77-2.83 (2H, dd, $J=15.5, 10$), 2.54-2.59 (2H, dd, $J=15.5, 3.5$), 2.17-2.24 (2H, quin, $J=6.0$), 1.71-1.73 (6H, d, $J=6.6$), 0.83 (18H, s), 0.01 (6H, s), 0.00 (6H, s). LC/MS, System 1: 100% pure at 2.34 min (ES+) m/z (relative intensity) 877.20 ($[\text{M}+\text{H}]^+$, 100).

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(vi) *Allyl 5-(3-(5-amino-4-((S)-2-((tert-butyldimethylsilyloxy)methyl)-4-((E)-prop-1-enyl)-2,3-dihydro-1H-pyrrole-1-carbonyl)-2-methoxyphenoxy)propoxy)-2-((S)-2-((tert-butyldimethylsilyloxy)methyl)-4-((E)-prop-1-enyl)-2,3-dihydro-1H-pyrrole-1-carbonyl)-4-methoxyphenylcarbamate* (**57**)

Pyridine (1.53 mL, 18.89 mmol, 1.0 eq) was added to a solution of the bis-aniline **56** (16.57 g, 18.89 mmol, 1.0 eq) in anhydrous DCM (240 mL) at -78°C under an atmosphere of nitrogen. After 5 minutes, allyl chloroformate (1.50 mL, 14.17 mmol, 0.75 eq) in anhydrous DCM (10 mL) was added and the reaction mixture allowed to warm to room temperature. After 30 minutes, the organic layer was washed successively with 5% citric acid (50 mL), saturated sodium bicarbonate (50 mL) and brine (10 mL), dried over MgSO_4 and evaporated to leave a yellow foam. The product was purified by flash chromatography (Biotage SP4

purification system, Biotage Snap Cartridge HP-Sil 100g, gradient elution Hexane:EtOAc 70% - 0%) to afford 3 fractions:

Fraction one (bis alloc protected product) as an off-white solid (3.6 g)

Fraction two (desired product, **57**) as a yellow solid (7.1 g, 52.2 %)

5 Fraction three (unreacted starting material) as an orange solid (5.5 g)

δ_H (DMSO- d_6): 9.09 (1H, s (br)), 7.23 (1H, s (br)), 6.82 (1H, s (br)), 6.47 (1H, s (br)), 6.43 (1H, s), 6.10-6.28 (3H, m), 5.86-5.93 (1H, m), 5.40-5.48 (3H, m), 5.27-5.31 (1H, dd, J=17.2, 1.2), 5.15-5.19 (3H, m), 4.52 (4H, m), 4.12-4.15 (2H, t, J=6.0), 4.06-4.09 (2H, t, J=6.0), 3.66-3.87 (4H, m), 3.74 (3H, s), 3.62 (3H, s), 2.76-2.83 (2H, m), 2.54-2.58 (2H, d (br), J=16.0), 2.19-2.25 (2H, quin, J=6.0), 1.71-1.73 (6H, d, J=6.2), 0.85 (9H, s), 0.83 (9H, s), 0.05 (3H, s), 0.03 (3H, s), 0.01 (3H, s), 0.00 (3H, s). LC/MS, System 1: 100% pure at 2.41 min (ES+) m/z (relative intensity) 961.40 ($[M+H]^+$, 100).

15 The bis-alloc protected compound and unreacted starting material were recycled using the following procedure:

Tetrakis(triphenylphosphine)palladium (0.350 g, 0.30 mmol, 0.06 eq) was added to a solution of bis-alloc protected aniline (5.27 g, 5.04 mmol, 1 eq) and pyrrolidine (1.04 ml, 12.6 mmol, 2.5 eq) in anhydrous DCM (50 ml) under nitrogen and the mixture was stirred for 1 hour after which, LCMS analysis showed completion of the reaction. The mixture was diluted with DCM and transferred into a separating funnel. The product was washed with saturated NH_4Cl (100 ml), saturated $NaHCO_3$ (100 ml) and brine (60 ml). After drying over $MgSO_4$, evaporation of the solvent gave the product as an orange solid (4.5 g). A product from a separate reaction (1 g) was combined and the crude mixture was absorbed on silica, then purified by flash chromatography (Biotage SP4 purification system, Biotage Snap Cartridge HP-Sil 100g, gradient elution Hexane:EtOAc 70%-0%) to give the bis-aniline **56** as an orange solid (3.1 g), which was then combined with recovered unreacted **56** in the mono-alloc protection (using 0.7 equ. of allyl chloroformate).

(vii) Allyl 4-((S)-2-((S)-2-(((allyloxy)carbonyl)amino)-3-methylbutanamido)propanamido)benzyl ((S,E)-(propane-1,3-diylbis(oxy))bis(2-((S)-2-(((tert-butyl)dimethylsilyl)oxy)methyl)-4-((E)-prop-1-en-1-yl)-2,3-dihydro-1H-pyrrole-1-carbonyl)-4-methoxy-5,1-phenylene))dicarbamate (**58**)

Triethylamine (2.26 mL, 16.2 mmol, 2.2 eq) was added to a stirred solution of the aniline **57** (7.1 g, 7.38 mmol, 1.0 eq) and triphosgene (0.79 g, 2.65 mmol, 0.36 eq) in anhydrous THF (70 mL) under nitrogen at room temperature. The reaction mixture was stirred at room

temperature for 15 minutes. LCMS analysis of an aliquot quenched with methanol, showed formation of the isocyanate. A solution of Alloc-Val-Ala-PAB-OH (4.18 g, 11.08 mmol, 1.5 eq) and triethylamine (1.54 mL, 11.08 mmol, 1.5 eq) in dry THF (150 mL) was added in one portion using an addition funnel and the resulting mixture was stirred overnight at 40°C.

- 5 Evaporation of the reaction mixture left a brown oil which was purified by flash chromatography (DCM:MeOH 97.5%) to afford the product, **58**, as a pale yellow solid (6.7 g, 67.0 %). A further 1.2 g (12 %) was isolated from mixed fractions. δ_{H} (DMSO- d_6): 9.99 (1H, s (br)), 9.10 (2H, s (br)), 8.14-8.16 (1H, d, J=6.0), 7.56-7.58 (2H, d, J=8.5), 7.27-7.29 (2H, d, J=8.4), 7.22-7.24 (2H, d, J=8.8), 6.81 (2H, s (br)), 6.28 (2H, s (br)), 6.09-6.14 (2H, d (br), J=16.0), 5.86-5.94 (2H, m), 5.41-5.50 (2H, m), 5.27-5.32 (2H, ddd, J=17.0, 3.6, 1.6), 5.15-5.19 (2H, ddd, J=10.4, 3.1, 1.5), 4.96-5.04 (2H, m), 4.40-4.52 (7H, m), 4.12-4.15 (4H, m), 3.83-3.91 (3H, m), 3.74 (6H, s), 3.64-3.72 (2H, m), 2.73-2.79 (2H, m), 2.51-2.58 (2H, m), 2.19-2.25 (2H, m), 1.93-2.01 (1H, m), 1.71-1.73 (6H, d, J=6.0), 1.29-1.30 (3H, d, J=7.0), 0.82-0.89 (24H, m), -0.02-0.04 (12H, m). LC/MS, System 1: 100% pure at 2.38 min (ES+)
- 10 m/z (relative intensity) 1387.15 ($[M+Na]^+$, 100).
- 15

(viii) *Allyl 4-((S)-2-((S)-2-(((allyloxy)carbonyl)amino)-3-methylbutanamido)propanamido)benzyl ((S,E)-(propane-1,3-diylbis(oxy))bis(2-((S)-2-(hydroxymethyl)-4-((E)-prop-1-en-1-yl)-2,3-dihydro-1H-pyrrole-1-carbonyl)-4-methoxy-5,1-phenylene))dicarbamate (59)*

20

- (a) Method 1: Water (12 mL) was added to a solution of TBS ether **58** (1.73 g, 1.27 mmol) in AcOH (42 mL): MeOH (6 mL): THF (6 mL) and the mixture was stirred at room temperature. The reaction was monitored by LCMS, which showed formation of desired product along with small amounts of unreacted starting material and mono-deprotected products after 1
- 25 hour. 10mL of AcOH:MeOH:THF:H₂O (7:1:1:2) were added and the mixture was stirred for another hour. LCMS analysis showed near completion of the reaction, only small amounts of mono-deprotected products were observed. The mixture was diluted with EtOAc, transferred into a separating funnel, washed with water, then with sat. NaHCO₃ until basic pH. The organic layer was washed with brine, dried over MgSO₄ and concentrated under vacuum.
- 30 The crude product was absorbed on silica and purified by flash chromatography (gradient elution CHCl₃:MeOH 100%-95%) to give the product, **59**, as a cream-coloured solid (1.02g, 71%).

- (b) Method 2: Tetra-n-butylammonium fluoride (1M, 9.19 mL, 9.19 mmol, 2.0 eq) was added to a solution of the TBS protected compound **58** (6.25 g, 4.59 mmol, 1.0 eq) in anhydrous
- 35 THF (75 mL). The reaction mixture was stirred at room temperature for 20 minutes, after which LCMS indicated formation of the product, along with small amounts of mono-

deprotected compounds. The reaction mixture was diluted with ethyl acetate (500 mL) and washed sequentially with water (250 mL) and brine (50 mL). The organic phase was dried over MgSO_4 and evaporated to leave a yellow solid. Purification by flash chromatography (DCM:MeOH 94%) afforded the bis-alcohol **59** as a pale cream solid (3.43 g, 66.0 %). δ_{H} (DMSO- d_6): 9.99 (1H, s (br)), 9.08 (1H, s (br)), 9.05 (1H, s (br)), 8.14-8.16 (1H, d, $J=6.8$), 7.56-7.58 (2H, d, $J=8.5$), 7.28-7.30 (2H, d, $J=8.5$), 7.23-7.25 (2H, d, $J=8.7$), 7.19-7.24 (1H, m), 6.88 (2H, s (br)), 6.22 (2H, s (br)), 6.08-6.12 (2H, d, $J=16.3$), 5.86-5.94 (2H, m), 5.41-5.48 (2H, m), 5.26-5.32 (2H, m), 5.16-5.48 (2H, m), 5.00 (2H, s (br)), 4.84-4.88 (2H, m), 4.40-4.52 (7H, m), 4.11-4.15 (4H, m), 3.87-3.90 (1H, dd, $J=8.9, 7.2$), 3.75 (6H, s), 3.63-3.69 (2H, m), 3.48-3.54 (2H, m), 2.71-2.79 (2H, m), 2.55-2.61 (2H, m), 2.20-2.25 (2H, m), 1.93-2.01 (1H, m), 1.71-1.73 (6H, d, $J=6.0$), 1.29-1.31 (3H, d, $J=7.1$), 0.87-0.89 (3H, d, $J=6.8$), 0.82-0.84 (3H, d, $J=6.7$). LC/MS, System 1: 100% pure at 1.74 min (ES+) m/z (relative intensity) 1136.25 ($[\text{M}+\text{H}]^+$, 100).

(ix) (11*S*,11*aS*)-allyl 8-(3-((11*S*,11*aS*)-10-((4-((*S*)-2-((*S*)-2-(allyloxycarbonylamino)-3-methylbutanamido)propanamido)benzyloxy)carbonyl)-11-hydroxy-7-methoxy-5-oxo-2-((*E*)-prop-1-enyl)-5,10,11,11*a*-tetrahydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepin-8-yloxy)propoxy)-11-hydroxy-7-methoxy-5-oxo-2-((*E*)-prop-1-enyl)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (**41**)

A solution of oxalyl chloride (357 mg, 2.82 mmol, 3.2 equ.) in anhydrous THF¹ (~700 μL) was added dropwise to a pre-mixed solution of anhydrous DMSO (400 μL , 5.63 mmol, 6.4 equ.) in THF (12mL) at -78°C under nitrogen and the mixture was stirred for 10 minutes. A solution of diol **59** (1 g, 0.880 mmol) in THF (20mL) was added dropwise over 10 minutes and the mixture was stirred for a further 20 minutes. before the addition of triethylamine (1.23 mL, 8.80 mmol, 10 equ.). The mixture was then allowed to warm to room temperature and progression of the reaction was monitored by LCMS, which showed completion of the reaction after 85 minutes. The mixture was diluted with EtOAc and transferred into a separating funnel. The product was washed with 1N HCl and the aqueous layer was extracted with EtOAc. The combined organic extracts were washed with sat. NaHCO_3 , brine and dried over MgSO_4 . The crude product was dissolved in DCM:MeOH, absorbed on silica and purified by flash chromatography (DCM:MeOH 98%-97.5%-97%)³ to give the cyclised product **41** as a colourless solid (561mg, 56%). δ_{H} (CDCl_3): 8.67 (1H, s (br)), 7.42 (1H, s (br)), 7.40 (1H, s (br)), 7.19-7.22 (2H, d, $J=10.2$), 7.10-7.16 (2H, m), 6.87-6.90 (2H, m), 6.83 (1H, m), 6.76 (1H, s (br)), 6.54 (1H, s (br)), 6.25-6.27 (1H, d, $J=6.5$), 6.21-6.23 (1H, d, $J=7.0$), 5.84-5.94 (1H, m), 5.71-5.80 (3H, m), 5.44-5.59 (3H, m), 5.27-5.33 (2H, d, $J=17.7$), 5.19-5.29 (1H, d, $J=10.4$), 4.99-5.09 (2H, m), 4.68-4.76 (2H, m), 4.52-4.61 (4H, m), 4.39-

4.45 (1H, m), 4.10-4.16 (2H, m), 4.00-4.04 (2H, m), 3.95-3.96 (1H, m), 3.86-3.89 (3H, m), 3.86 (3H, s), 3.84 (3H, s), 3.04-3.12 (2H, m), 2.75-2.82 (2H, m), 2.10-2.28 (3H, m), 1.81-1.82 (6H, d, J=7.1), 1.39-1.45 (3H, m), 0.94-0.96 (3H, d, J=6.8), 0.91-0.93 (3H, d, J=7.0). LC/MS, System 1: 100% pure at 1.72 min (ES+) m/z (relative intensity) 1132.25 ($[M+H]^+$, 100).

5 HRMS: $C_{59}H_{69}N_8O_{16}^+$ requires 1132.4834, found 1132.2799 (MH^+), 1149.3333 ($M+NH_4^+$)

(x) (11*S*,11*aS*)-4-(2-(1-((1-amino-3-methyl-1-oxobutan-2-yl)amino)-1-oxopropan-2-yl)hydrazinyl)benzyl 11-hydroxy-7-methoxy-8-(3-(((*S*)-7-methoxy-5-oxo-2-((*E*)-prop-1-en-1-yl)-5,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepin-8-yl)oxy)propoxy)-5-oxo-2-((*E*)-prop-1-en-1-yl)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate (25)

Tetrakis(triphenylphosphine)palladium(0) (14 mg, 0.01 mmol, 0.06 eq) was added to a solution of **41** (0.208 mmol) and pyrrolidine (43 μ L, 0.52 mmol, 2.5 eq) in chloroform (5 mL). The reaction mixture was stirred 20 minutes and diluted with chloroform and washed sequentially with saturated aqueous ammonium chloride and brine. The organic phase was dried over magnesium sulphate filtered and excess chloroform removed by rotary evaporation under reduced pressure. The resulting residue **25** was used as a crude mixture for the next reaction. δ_H (DMSO- d_6): 10.26 (1H, s), 7.97 (1H, d, J=5.9), 7.59 (2H, d, J=8.4), 7.22 (3H, m), 7.08 (1H, s), 7.04 (1H, s), 7.00 (1H, m), 6.90 (2H, s), 6.77 (2H, m), 6.55 (1H, s), 6.30 (3H, m), 5.75 (2H, s), 5.55 (3H, m), 5.42 (1H, m), 5.23 (1H, d, J=12.4), 4.76 (1H, d, J=12.1), 4.33 (1H, m), 4.22 (2H, m), 4.10 (3H, m), 4.00 (3H, m), 3.80 (5H, m), 3.70 (3H, s), 3.17 (1H, d, J=5.1), 3.03 (3H, m), 2.87 (4H, m), 2.61 (3H, m), 2.26 (4H, m), 1.77 (10H, m), 1.19 (3H, d, J=7.0), 0.89 (3H, m), 0.79 (6H, t, J=6.4).

LCMS, System 2: 1.23 min.; m/z (ES $^+$): 946.35 ($M+H^+$)

25

(xi) (11*S*,11*aS*)-4-((20*S*,23*S*)-1-iodo-20-isopropyl-23-methyl-2,18,21-trioxo-6,9,12,15-tetraoxa-3,19,22-triazatetracosanamido)benzyl 11-hydroxy-7-methoxy-8-(3-(((*S*)-7-methoxy-5-oxo-2-((*E*)-prop-1-en-1-yl)-5,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepin-8-yl)oxy)propoxy)-5-oxo-2-((*E*)-prop-1-en-1-yl)-11,11*a*-dihydro-1*H*-benzo[*e*]pyrrolo[1,2-*a*][1,4]diazepine-10(5*H*)-carboxylate, (26)

N,N'-diisopropylcarbodiimide (DIC, 32 μ L, 0.208 mmol) was added to a solution of crude **25** (0.208 mmol) and iodoacetamide(PEG) $_4$ acid (90 mg, 0.208 mmol) in chloroform (8 mL). The reaction was stirred for 3 hours and the presence of starting material was still observed by LC/MS. Further *N,N'*-diisopropylcarbodiimide (0.104 mmol) and iodoacetamide(PEG) $_4$ acid (0.104 mmol) were added to the reaction mixture and allowed to react overnight at 4°C.

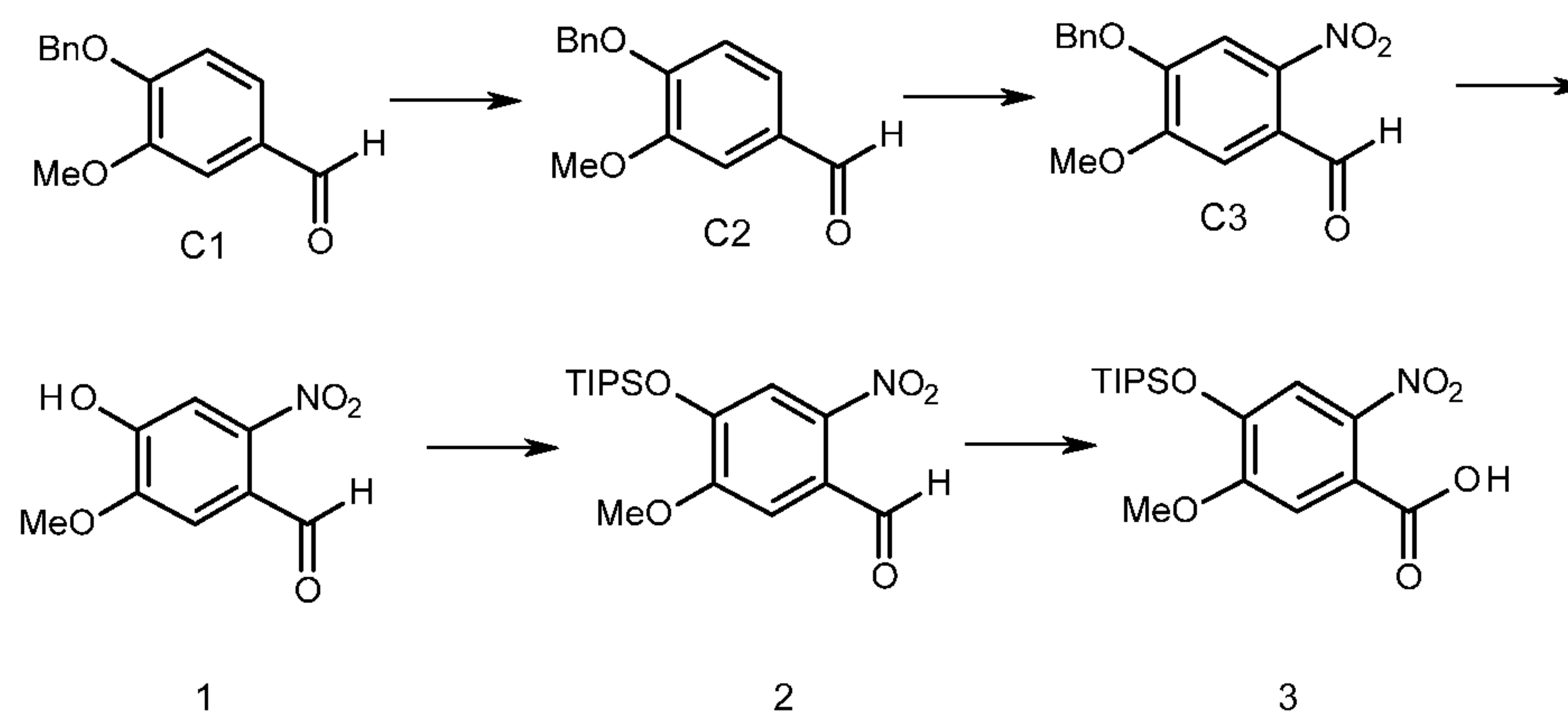
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The reaction mixture was loaded directly on to silica and subjected to flash column chromatography (silica gel; 100% dichloromethane to 5% methanol in dichloromethane).

Pure fractions were collected and combined and excess eluent was removed by rotary evaporation under reduced pressure to give **26** (149 mg, 53% over 2 steps). δ_{H} (DMSO- d_6):

5 9.92 (1H, s), 8.17 (1H, d, $J=6.5$), 7.97 (1H, d, $J=3.9$), 7.86 (1H, d, $J=8.6$), 7.54 (2H, m), 7.33 (1H, s), 7.17 (2H, m), 7.07 (1H, m), 6.88 (2H, br s), 6.78 (2H, m), 6.30 (2H, m), 5.56 (3H, m), 5.11 (1H, m), 4.86 (1H, m), 4.38 (2H, m), 4.20 (2H, m), 4.10 (2H, m), 4.06 (1H, s), 4.00 (1H, m), 3.80 (4H, m), 3.74 (2H, m), 3.64 (2H, s), 3.59 (2H, t, $J=6.4$), 3.49 (9H, m), 3.40 (4H, m), 3.25 (2H, m), 3.20 (2H, m), 3.00 (2H, m), 2.63 (2H, m), 2.40 (3H, m), 2.19 (3H, m), 1.95 (1H, m), 1.78 (5H, m), 1.28 (4H, m), 0.84 (6H, dd, $J=15.9, 6.7$).

LCMS, System 2: 1.54 min.; m/z (ES^+): 1361.40 ($\text{M}+\text{H}^+$); System 3: 6.14 min; m/z (ES^+): 1361.40 ($\text{M}+\text{H}^+$).

Example 5 – Further synthesis of 17**(i) 5-methoxy-2-nitro-4-((triisopropylsilyl)oxy)benzoic acid (3)****(a) C2**

- 5 1. Charge 500mL of anhydrous CH₃CN (Analytical reagent, 5V) to a 2 L flask.
2. Charge 99.0 g of **C1** (1.0eq) to the flask.
3. Charge 99.6g of powder K₂CO₃ (Analytical reagent, 1.1eq) to the flask
4. Heat to 40~50°C.
5. Drop-wise 123.2 g of Benzyl bromide (Chemical pure, 95%, 1.1eq) slowly.
- 10 6. Heat to 50~60°C.
7. Stir for 4 hours.
8. Sample for IPC (completion acceptance criteria: SM ≤2.0% by HPLC).
9. Cool to 20~25°C.
10. Pour the mixture to ice-water (1.5L, 15V).
- 15 11. Stir for about 1 hour.
12. Filter.
13. Wash the filter cake with water (50mL, 0.5V,).
14. Slurry the filter cake with hexane (200mL, 2V,).
15. Filter and collect the solid.
- 20 16. Dry the product using vacuum oven at 25-30°C for 20 hours (internal pressure around 20-50 mmHg).
17. Product information (145.0g, Purity: 99%; Yield: 92%).

(b) C3

- 25 1. Charge 2.5 L of conc HNO₃(70%, 5V) to a 5 L flask.
2. Cool to 2~7°C.
3. Charge 500g of **C2** (1.0eq) in portion while main the temperature at 2~7°C.
4. Stir for 8 hours at 2~7°C.

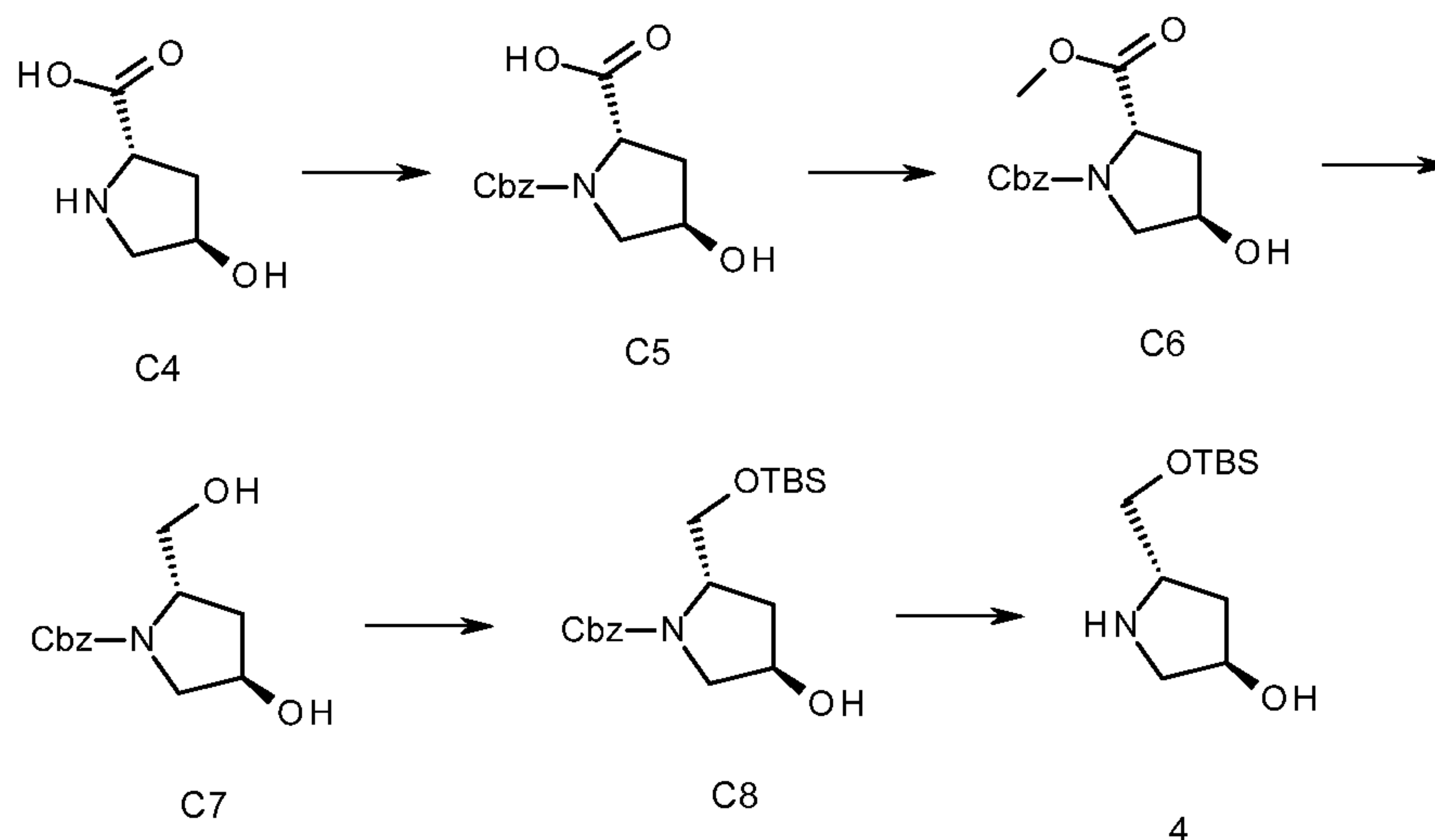
5. Sample for IPC (Completion acceptance Criteria: SM $\leq$ 2.0% by HPLC).
 6. Pour the mixture to ice-water (7.5L, 15V).
 7. Stir for about 1 hour.
 8. Filter.
 - 5 9. Wash the filter cake with water (500ML, 1V).
 10. Dry the product under vacuum.
 11. Recrystallize(stir at 70~80°C for 1 hour, partial dissolution) the solid product with EtOAc (1L, 2V).
 12. Filter.
 - 10 18. Dry the product Dry the product using vacuum oven at 20-30°C for 26 hours (internal pressure around 20-50 mmHg).
 13. Product information (420.0g, Purity: 98%; Yield: 71%).
- (c) 1
- 15 1. Charge 600mL of TFA (1.5V) to a 2 L flask.
 2. Heat to 40~50°C.
 3. Charge 400g of **C3** (1.0eq) slowly portion-wise to the flask.
 4. Heat to 75~85°C.
 5. Stir for 2 hours at 75~85°C.
 - 20 6. Sample for IPC (completion criteria: SM $\leq$ 2.0% by HPLC).
 7. Cool to the reaction mixture to 40~50°C.
 8. Concentrate the reaction mixture under vacuum at 40~50°C.
 9. Slurry the crude product with MTBE (400mL, 1V).
 10. Filter and collect the solid.
 - 25 11. Dry the product using vacuum oven at 20-30°C for 30 hours (internal pressure around 20-50 mmHg).
 12. Product information (247.0g, Purity: 98%; Yield: 90%).
- (d) 2
- 30 1. Charge 15.75L of CH₂Cl₂ (5V) to a 50 L reactor.
 2. Charge 3.15kg of **1** (1.0eq) to the reactor.
 3. Cool to 0~10°C.
 4. Charge 1.78kg of anhydrous Et₃N (Analytical reagent, 1.2eq) slowly to the reactor.
 - 35 5. Stir for 0.5 hour.
 6. Charge 3.08kg of TIPSCl (1.1eq) slowly to the reactor.

7. Heat to 5~15°C.
 8. Stir for 2 hours.
 9. Sample for IPC (completion criteria: SM $\leq$ 2.0% by HPLC).
 10. Concentrate the mixture under vacuum at 25~35°C.
 - 5 11. Purify the crude product by column chromatography eluting with 5% ethyl acetate in Petro ether.
 12. Slurry the crude product with hexane (1.6L, 0.5V).
 13. Filter and collect the solid.
 14. Dry the product using vacuum oven at 25-30°C for 18 hours (internal pressure around 20-50 mmHg).
 - 10 15. Product information (4.6kg, Purity: 98%; Yield: 81%).
- (e) 3
1. Charge 1.4L of H₂O (7V) to a 5 L flask.
 - 15 2. Charge 112.6g of NaClO₂ (Solid, analytical reagent, 82% active reagent, 2.2eq) to the flask.
 3. Charge 88.2g of NaH₂PO₄ (1.3eq) to the flask.
 4. Charge a solution of **2** (200.0g, 1.0eq) in THF (1.0L, 5V) to the flask.
 5. Charge 115.5mL of H₂O₂ (Analytical reagent, 30% w/w, 2.0eq) to the flask.
 - 20 6. Heat to 40~50°C.
 7. Stir for 1 hour.
 8. Sample for IPC (completion criteria: SM $\leq$ 2.0% by HPLC).
 9. Cool to 20~30°C.
 10. Acidify the mixture using 1M HCl to PH 2.
 - 25 11. Separate the organic phase.
 12. Extract the aqueous phase with ethyl acetate (1.4L, 7V).
 13. Separate the organic phase.
 14. Extract the aqueous phase with ethyl acetate (1.0L, 5V).
 15. Separate the organic phase.
 - 30 16. Combine the organic phase.
 17. Wash the organic phase with brine (1.0L, 5V).
 18. Dry over Na₂SO₄ (Na₂SO₄ is easy to filter).
 19. Filter and concentrate the mixture under vacuum at 30~40°C.
 20. Slurry the crude product with hexane (200mL, 1V).
 - 35 21. Filter and collect the solid.
 22. Dry the product using vacuum oven at 25-30°C for 16 hours (internal pressure

around 20-50 mmHg).

23. Product information (188g, Purity: 99%; Yield: 90%).

(ii) 4



(a) C5

1. Charge trans-4-hydroxy-L-proline **C4** (100g, 1.0eq) into the reactor.
 2. Charge ice water (1.2L, 6V/W) into the reactor with stirring at $15 \pm 5^\circ\text{C}$.
 3. Charge NaHCO_3 (160.3g, 2.5eq) as a solid into the solution,
 4. Add the solution of CbzCl (143.0g, 1.1eq) in toluene (400ml, 4V/W) drop-wise in 30 minutes.
 5. Stir the mixture at room temperature for 18 hours.
 6. Sample from aqueous layer showed $\text{SM} < 2.0\%$. Separate the two phases.
 7. Wash the aqueous phase with EtOAc (1 x 3V), cool in an ice bath, and then acidify to pH 2 with conc. HCl.
 8. Extract the aqueous layer with EtOAc (3x4V) and the combined organic extracts and dry with Na_2SO_4 .
 9. Remove the solvent under vacuum to afford colorless viscous oil.
- $[\alpha]_{\text{D}}^{22} 29.2^\circ, (c=0.1\text{g}/100\text{ml in Methanol})$

(b) C6

1. Charge **C5** (90.0g, 1.0eq) into the reactor.
2. Charge MeOH (540ml, 6V/W) into the reactor with stirring at $15 \pm 5^\circ\text{C}$.
3. Charge H_2SO_4 (4.5g, 0.05eq) into the solution. Heat the mixture to $65 \pm 5^\circ\text{C}$ and keep refluxing for 4 hours.

4. Sample for IPC by HPLC (completion criteria: C5<2.0%).
5. Cool the mixture to 25±5°C.
6. Charge Et₃N (18g, 0.2eq) into the reactor, stir for additional 30 minutes.
7. Then concentrate the mixture to minimum volume under vacuum at 35±5°C.
8. Dissolve the residue in EtOAc (5V/W) and wash it with water (1V/W).
9. Separate the organic phase and wash it with brine (1V/W).
10. Separate the organic phase and dry it with anhydrous Na₂SO₄.
11. Concentrate the solution to minimum volume under vacuum at 35±5°C to afford yellow viscous oil. $[\alpha]_D^{25} -66.9^\circ$, (c=0.016g/100ml in Chloroform)

10

(c) C7

1. Charge **C6** (90.0g, 1.0eq) into the reactor.
2. Charge THF (450ml, 5V/W) into the reactor with stirring at 0±5°C.
3. Charge LiBH₄ (11.0g, 1.5eq) into the solution.
4. Stir mixture for 30mins at 0±5°C, allow the mixture to warm to room temperature.
5. Stir the mixture at room temperature for 4 hours, sample for IPC by HPLC after 4 hours (completion criteria: C6<2.0%).
6. Cool to 0°C, dilute with water (5V), and treat dropwise with aq HCl (4V, 2M), in the progress, vigorous effervescence can be observed.
7. Concentrate under vacuum to remove THF, and extract the aqueous residue with iPrOAc (3V*4).
8. Then wash the organic phase with brine (3V) and dry over Na₂SO₄.
9. Concentrate under vacuum and get a colorless product. $[\alpha]_D^{23} -41.6^\circ$, (c=0.93g/100ml in Chloroform)

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20

25 (d) C8

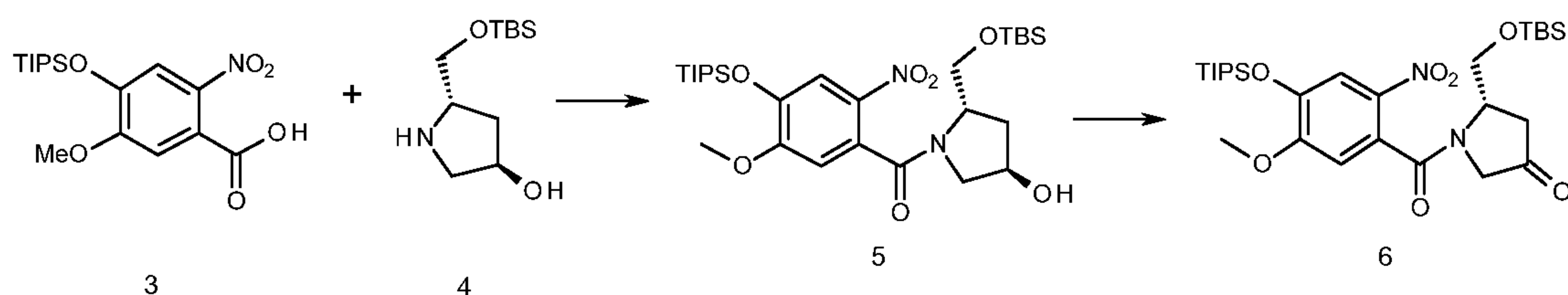
1. Charge **C7** (1.0eq) into the reactor.
2. Charge Toluene (5V/W) into the reactor with stirring.
3. Charge TEA (1.5eq) into the solution.
4. Rise to 50°C. Then charge TBSCl (1.1eq) into the reactor.
5. React for 18 hours at 60°C.
6. Sample for IPC by HPLC (completion criteria: starting material is less than 5.0%).
7. Cool to 30°C, quench with NH₄Cl (aq) (3V/W), separate the organic phase.
8. Then wash with brine (3V) and dry over Na₂SO₄. Concentrate under vacuum and get a brown product. $[\alpha]_D^{24} -52.4^\circ$, (c=0.94g/100ml in Chloroform)

30

(e) 4

1. Charge **C8** (1.0eq) into the reactor.
2. Charge IPA (8V/W) into the reactor with stirring.
3. Charge Pd/C (0.05W/W) into the solution.
- 5 4. Stir at 10°C-20°C. Then charge H₂ into the reactor.
5. React for 18 hours at 10°C-20°C.
6. Sample for IPC by HPLC (completion criteria: starting material is less than 1.0%).
7. Filter the reaction mixture.
8. Concentrate the filtrate at 35±5°C under vacuum.
- 10 9. Charge heptane into the residue and concentrate at 35±5°C under vacuum.
10. Charge heptane into the residue and concentrate at 35±5°C under vacuum once again.
11. Charge heptane into the residue.
12. Cool to -10±5°C and stir for 30 minutes.
- 15 13. Filtrate the mixture and collect the filtrate cake.
14. Dry the filtrate cake in the air at room temperature for 28 hours
15. Product information: 2.5kg (purity: 99.2%) [α]_D²⁴ +11.2.4°, (c=0.04g/100ml in Chloroform)

20 (iii) 6

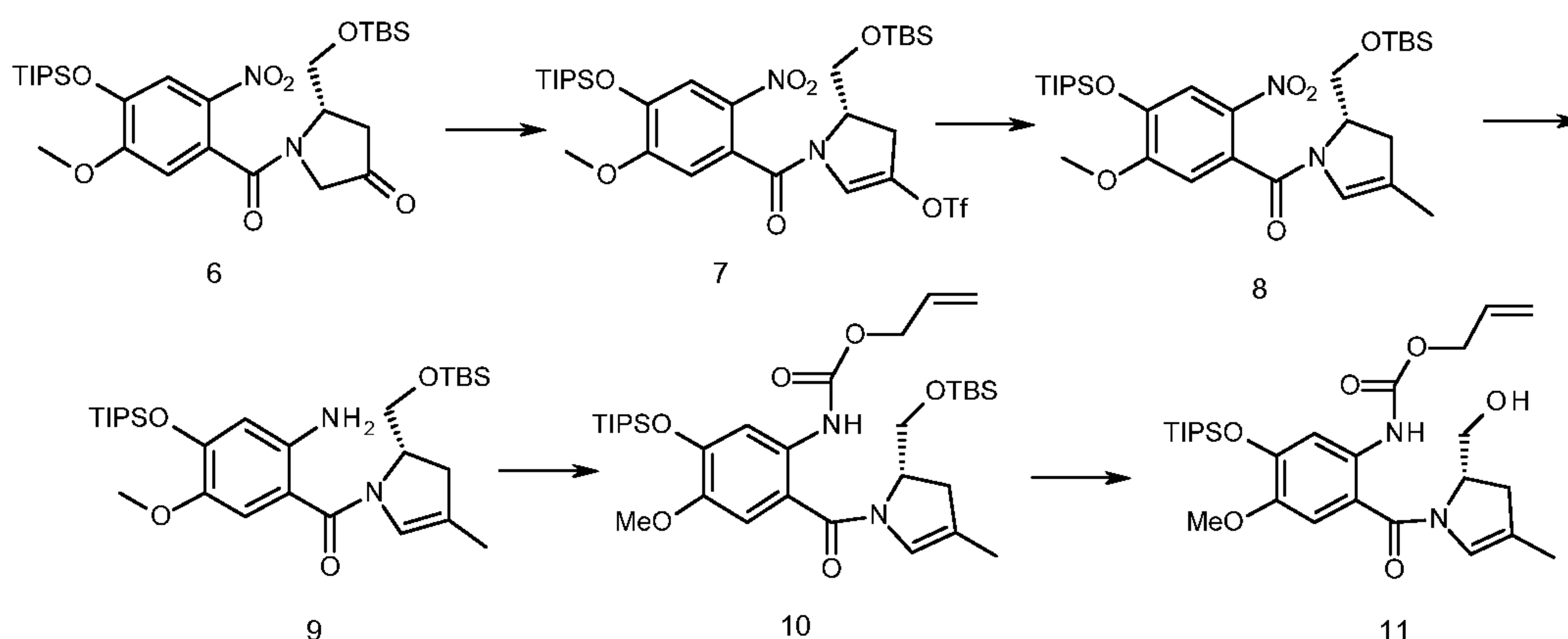


(a) 5

1. Charge 1.0L of anhydrous CH₂Cl₂(Analytical reagent, 8V) to a 3 L flask.
2. Charge 200g of **3** (1.0eq) to the flask.
- 25 3. Charge 80.4g of anhydrous HOBt (Analytical reagent, 1.1eq) to the flask.
4. Cool to 0~10°C.
5. Charge 124.5g of EDCI (Chemical pure, 95%, 1.2eq) to the flask.
6. Heat to 20~30°C.
7. Stir for 30 minutes.
- 30 8. Cool the mixture to -10~0°C.

9. Charge a solution of **4** (131.5g, 1.05eq) and TEA (68.5g, 1.25eq) in CH₂Cl₂ (600mL, 3V) rapidly to the flask.
 10. Heat to 20~30°C.
 11. Stir for 2 hours.
 - 5 12. Sample for IPC (completion criteria: 4 ≤2.0% by HPLC).
 13. Cool to 0~10°C.
 14. Wash the mixture with water (400mL, 2V)
 15. Separate the organic phase.
 16. Acidify the organic phase using 0.1M HCl to PH 4~5 (pH paper testing).
 - 10 17. Separate the organic phase.
 18. Wash the organic phase with water (400mL, 2V).
 19. Separate the organic phase.
 20. Wash the organic layer with saturated aqueous sodium bicarbonate (400mL, 2V).
 21. Separate the organic phase.
 - 15 22. Dry over Na₂SO₄.
 23. Filter and concentrate the mixture under vacuum at 25~35°C.
 24. Slurry the crude product with hexane (400mL, 2V).
 25. Filter and collect the solid.
 26. Dry the product using vacuum oven at 30-40°C for 20 hours (internal pressure around 20-50 mmHg).
 - 20 27. Product information (265g, Purity: 96%; Yield: 84%). $[\alpha]_D^{23} -100.1^\circ$, (c=0.35 g/100ml in Chloroform).
- (b) 6
1. Charge 600mL of CH₂Cl₂ (6V) to a 2 L flask.
 - 25 2. Charge 100g of **5** (1.0eq) to the flask.
 3. Charge 98.2g of DMP (1.35eq) to the flask.
 4. Heat to 25~35°C.
 5. Stir for 20 hours.
 6. Sample for IPC (completion criteria: 5 ≤2.0% by HPLC).
 - 30 7. Cool to 5~15°C.
 8. Quench the reaction with saturated aqueous sodium bicarbonate (600mL, 6V).
 9. Stir for 30 minutes.
 10. Filter and collect the filtrate.
 11. Wash the filter cake CH₂Cl₂ (200mL, 2V).
 - 35 12. Separate the organic phase.

13. Concentrate the organic phase under vacuum.
14. Dissolve the crude product with MTBE (400mL, 4V).
15. Filter and collect the filtrate.
16. Wash the filter cake MTBE (200mL, 2V).
- 5 17. Combine the organic phase.
18. Wash the organic phase with saturated aqueous sodium bicarbonate (300mL, 3V).
19. Separate the organic phase.
20. Extract the aqueous phase with MTBE (100mL, 1V).
- 10 21. Separate the organic phase.
22. Combine the organic phase.
23. Wash the organic layer with saturated aqueous sodium bicarbonate (300mL, 3V)
24. Separate the organic phase.
25. Wash the organic phase with water (300mL, 3V).
- 15 26. Separate the organic phase.
27. Dry over Na₂SO₄.
28. Filter and concentrate the mixture under vacuum at 25~35°C.
29. Dry the product in the air at room temperature for 48 hours
30. Product information (90g, Purity: 98%; Yield: 90%) [α]_D²⁶ -25.1° , (c=0.98
- 20 g/100ml in Chloroform).

(iv) 11(a) 7

- 25 1. Charge 30mL of anhydrous DCM(Analytical reagent)(6V) to a 100ml flask.
2. Charge 5g of **6** (1.0eq) to the flask under nitrogen.

3. Cool to -50°C.
4. Charge 2.8g of Lutidine (Chemically pure) (3.0eq) to the flask
5. Dropwise 6.1g of (Tf)₂O (Chemically pure) (2.5eq) slowly while main the temperature at -55~-45°C.
- 5 6. Stir for 2.0 hours at -50°C.
7. Sample for IPC (completion criteria: 6 ≤2.0% by LC-MS).
8. Pour the mixture to sat. aqueousNaHCO₃ (5V).
9. Stir for about 15 minutes.
10. Separate the organic phase and wash it with brine (5V).
- 10 11. Separate the organic phase and dry over anhydrous Na₂SO₄.
12. Filter and concentrate the organic phase under vacuum at 30~40°C to dryness.
13. The residue subject to flash column chromatography (silica gel: 15/1,v/v, hexane/ethyl acetate).
14. Collect pure fractionsand remove the solvent under vacuum at 30~40°C.
- 15 15. Drythe product in glass bottle using evaporator under vacuum at 30~40°C to constant amount (the internal pressure was around 20-50 mmHg)
16. Product information: brown oil (4.3g, Purity: 85.0%; Yield: 70.0%). $[\alpha]_D^{23}$ -52.97° , (c=0.202 g/100ml in Chloroform).
- 20 (b) 8
1. Charge 7.0 L of anhydrous toluene (Analytical reagent)(8V) to a 20 L flask.
2. Charge 880g of **7** (1.0eq),258.6g of MeB(OH)₂ (Chemically pure)(3.5eq)and 1.57Kg of K₃PO₄ (Chemically pure)(6.0eq) under nitrogen.
3. Charge 214.0g of Pd(PPh₃)₄ (Chemically pure) (0.15eq).
- 25 4. Stir for about 30 minutes at 60°C.
5. Sample for IPC (completion criteria: 7 ≤2.0% by LC-MS).
6. Filter and collect the filtrate.
7. Wash filtrate withwater (5V).
8. Separate the organic phase and wash it with sat. aqueous NaHCO₃ (5V).
- 30 9. Separate the organic phase and wash it with brine (5V).
10. Separate the organic phase and dry over anhydrous Na₂SO₄.
11. Filter and concentrate the organic phase under vacuum at 50~60°C to dryness.
12. Subject the residue to flash column chromatography (silica gel; 10/1, v/v, hexane/ethyl acetate).
- 35 13. Collect pure fractions and concentrate it under vacuum at 30~40°C.

14. Dry the product in glass bottle using evaporator under vacuum at 30~40°C to constant amount (the internal pressure was around 20-50 mmHg)
15. Product information: brown oil (470.0g, Purity: 93.4%; Yield: 66.0%).

5 (c) 9

1. Charge 2.60Kg of Zn (Chemical pure)(37.0eq) to a 20L flask.
2. Charge 6.3L of EtOH(Analytical reagent)/HCO₂H(10/0.05)(10V).
3. Cool to 5°C.
- 10 4. Add solution of **8** (623g, 1.0eq) in EtOH(1.5 L, 2.5 V) to the mixture drop-wise with stirring.
5. Main the temperature at 5±5°C and stir for 1 hour.
6. Sample for IPC (completion criteria: 8 ≤2.0% by LC-MS).
7. Filter and collect the filtrate.
8. Pour the filtrate to EtOAc (20V) and wash it with water (20V)
- 15 9. Separate the organic phase and wash it with sat. aqueous NaHCO₃ (20V).
10. Separate the organic phase and wash it with brine (20V).
11. Separate the organic phase and dry over anhydrous Na₂SO₄.
12. Filter and concentrate the organic phase under vacuum.
13. The residue subject to flash column chromatography (silica gel; 5/1, v/v,
- 20 hexane/ethyl acetate).
14. Collect pure fractions and concentrate it under vacuum at 30~40°C.
15. Dry the product in glass bottle using evaporator under vacuum at 30~40°C to constant amount (the internal pressure was around 20-50 mmHg).
16. Product information: brown oil (470.0g, Purity: 97.0% Yield: 79.6%). $[\alpha]_D^{23}$ -
- 25 176.47°, (c=0.187 g/100ml in Chloroform).

(d) 10

1. Charge 6.1L of DCM(Analytical reagent) (13V) and 470g of **9** (1.0eq) to a 10L flask.
2. Cool to -85°C.
- 30 3. Charge 149.0g of Pyridine (Analytical reagent) (2.2eq) to the flask.
4. Dropwise addition 113.5 g of Allylchloroformate (Chemically pure) (1.1eq) slowly while main the temperature at -85°C ~-75°C to the flask.
5. Stir for 30 minutes at -85°C ~-75°C.
6. Warm to room temperature.

172

7. Sample for IPC (completion criteria: $9 \leq 2.0\%$ by LC-MS).
8. Charge DCM (Analytical reagent) (10V) and saturated aqueous copper sulphate (20V).
9. Separate the organic phase and wash it with sat. aqueous NaHCO_3 (20V).
- 5 10. Separate the organic phase and wash it with brine (20V).
11. Separate the organic phase and dry over anhydrous Na_2SO_4 .
12. Filter and concentrate the organic phase under vacuum at $25 \sim 35^\circ\text{C}$.
13. Dry the product in glass bottle using evaporator under vacuum at $30 \sim 40^\circ\text{C}$ to constant amount (the internal pressure was around 20-50 mmHg).
- 10 14. Product information: brown oil (515.0g, Purity: 95.9% Yield: 95.0%). $[\alpha]_D^{23} -90.40^\circ$, ($c=0.177$ g/100ml in Chloroform).

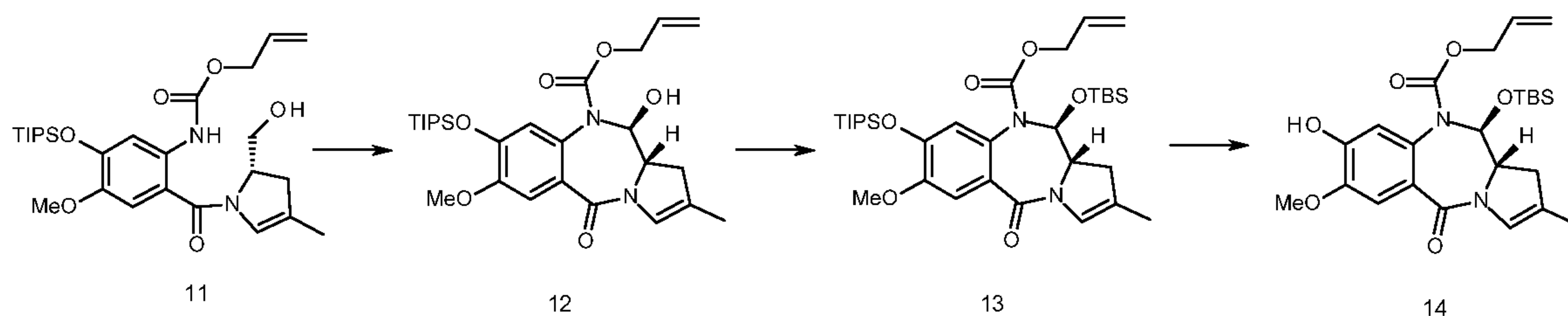
(e) 11

1. Charge 2.1L of AcOH/MeOH/THF/ H_2O (7:1:1:2) (4V) to a 5L flask.
- 15 2. Charge 515.0g of **10** to the flask.
3. Stir for 1h at $25^\circ\text{C} \sim 30^\circ\text{C}$.
4. Sample for IPC (completion criteria: $\text{SM} \leq 2.0\%$ by LC-MS).
5. Charge EtOAc (Analytical reagent) (30V) and water (20V).
6. Separate the organic phase and wash it with sat. aqueous NaHCO_3 (20V).
- 20 7. Separate the organic phase and wash it with brine (20V).
8. Separate the organic phase and dry over anhydrous Na_2SO_4 .
9. Filter and concentrate the organic phase under vacuum.
10. Subject the residue to flash column chromatography (silica gel; 4/1, v/v, hexane/ethyl acetate).
- 25 11. Collect pure fractions and concentrate it under vacuum at $30 \sim 40^\circ\text{C}$.
12. Dry the product in glass bottle using evaporator under vacuum at $30 \sim 40^\circ\text{C}$ to constant amount (the internal pressure was around 20-50 mmHg).
13. Product information (337.6g, Purity: 99.0%; Yield: 80.0%) $[\alpha]_D^{28} -80.95^\circ$, ($c=0.231$ g/100ml in Chloroform).

30

(v) 14

173



(a) 12

1. Charge 500mL of anhydrous DCM (Analytical reagent) (10V) to a 2 L flask.
2. Charge 14.7g of $(\text{COCl})_2$ (Chemical pure) (1.2eq) to the flask under nitrogen
3. Cool to -78°C .
4. Drop-wise 18.8 g of anhydrous DMSO (Analytical reagent) (2.5eq) slowly while main the temperature at $-78\sim-70^\circ\text{C}$.
5. Drop-wise 50.0 g of **11** (1.0eq) in anhydrous DCM (Analytical reagent) to the flask while main the temperature at $-78^\circ\text{C} \sim -70^\circ\text{C}$.
6. Stir for 15 minutes.
7. Drop-wise 50.0 g of TEA (Analytical reagent) (5.0eq) to the flask.
8. Stir for 2 hours.
9. Sample for IPC (completion acceptance criteria: $\text{SM} \leq 2.0\%$ by LC-MS).
10. Pour the mixture to 0.1mol ice-hydrochloric acid (5V).
11. Stir for about 15 minutes.
12. Separate the organic phase and wash it with sat. aqueous NaHCO_3 (5V).
13. Separate the organic phase and wash it with brine (5V).
14. Separate the organic phase and dry over anhydrous Na_2SO_4 .
15. Filter and concentrate the organic phase under vacuum at $30\sim40^\circ\text{C}$ to dryness.
16. Subject the residue to flash column chromatography (silica gel; 1/3, v/v, hexane/ethyl acetate).
17. Collect pure fractions and concentrate it under vacuum at $30\sim40^\circ\text{C}$ to dryness.
18. Product information (30.3g, Purity: 97.7%; Yield: 61.3%). $[\alpha]_{\text{D}}^{25} 101.95^\circ$, (c=0.205 g/100ml in Chloroform).

(b) 13

1. Charge 1.1 L of anhydrous DCM (Analytical reagent) (20V) to a 2 L flask.
2. Cool to 0°C .
3. Charge 55g of **12** (1.0eq) and lutidine (Analytical reagent) (4.0eq)
4. Drop-wise 84.4 g of TBSOTf (Chemical pure) (3.0eq) slowly while main the temperature at $0^\circ\text{C} \sim 5^\circ\text{C}$.

5. Stir for about 15 minutes at 0°C ~5°C.
6. Stir at room temperature for 1.5 hours.
7. Sample for IPC (Completion acceptance Criteria: $12 \leq 2.0\%$ by LC-MS).
8. Pour the mixture to water (1.1L, 20V).
- 5 9. Stir for about 30 minutes
10. Separate the organic phase and wash it with sat. Aqueous NaHCO₃ (5V).
11. Separate the organic phase and wash it with brine (5V).
12. Separate the organic phase and dry over anhydrous Na₂SO₄.
13. Filter and concentrate the organic phase under vacuum at 30~40°C to dryness.
- 10 14. Product information (62.0gcrude). $[\alpha]_D^{25} 54.62^\circ$ (c=0.216g/100ml in Chloroform).

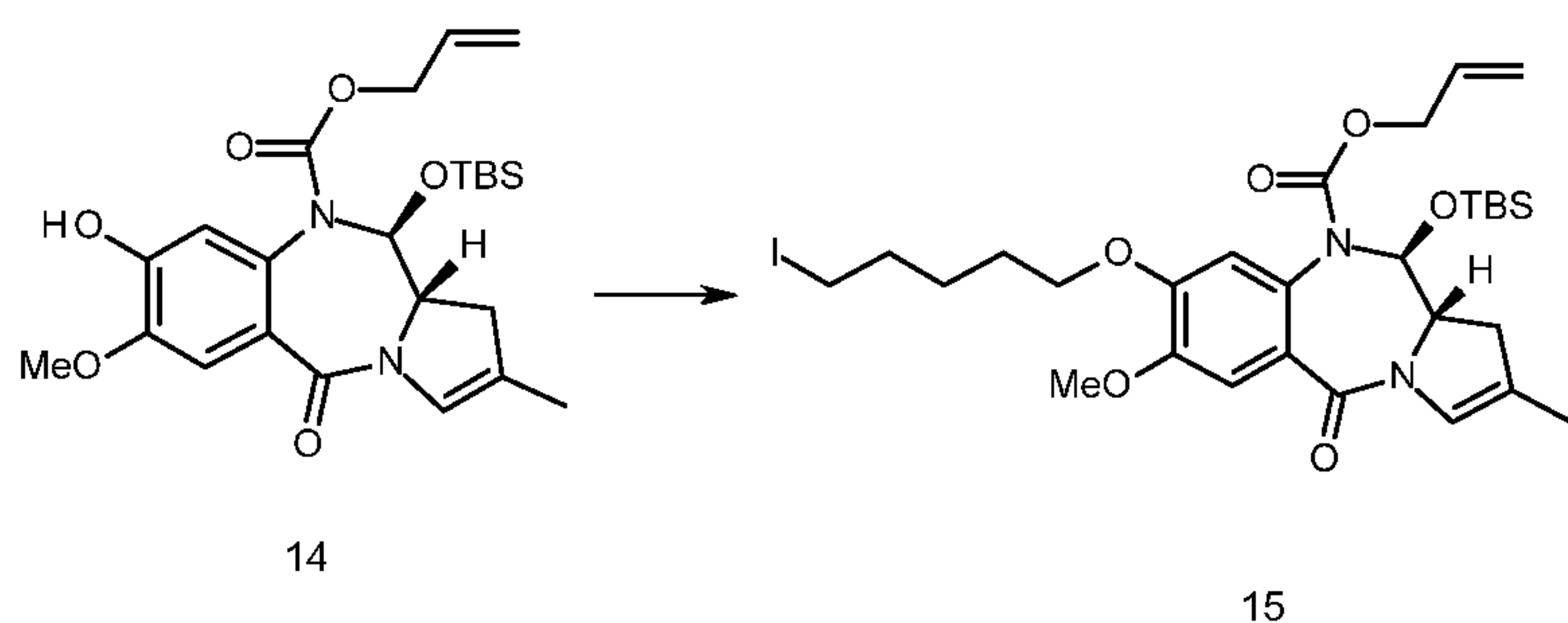
(c) 14

- 15 1. Charge 12mL of DMF (Analytical reagent)/H₂O(50/1) (12V) and 1g of **13** (1.0eq) to a 50ml flask.
2. Charge 0.16g of LiOAc(Chemical pure)(1.0eq) to the flask.
3. Stir for 4 hours at 25°C.
4. Sample for IPC (completion criteria: $13 \leq 2.0\%$ by LC-MS).
5. Pour the mixture to EtOAc (25ml, 25V).
- 20 6. Wash the organic phase with citrate solution (20V).
7. Separate the organic phase and wash it with water (20V)
8. Separate the organic phase and wash it with brine (20V).
9. Separate the organic phase and dry over anhydrous Na₂SO₄.
10. Filter and concentrate the organic phase under vacuum.
- 25 11. The residue subject to flash column chromatography (silica gel; 1/3, v/v, hexane/ethyl acetate).
12. Collect pure fractions and concentrate it under vacuum at 30~40°C to dryness.
13. Product information (0.6g, Purity: 99%; Yield: 80%). $[\alpha]_D^{28} 59.73^\circ$ (c=0.216 g/100ml in Chloroform).

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(vi) 15

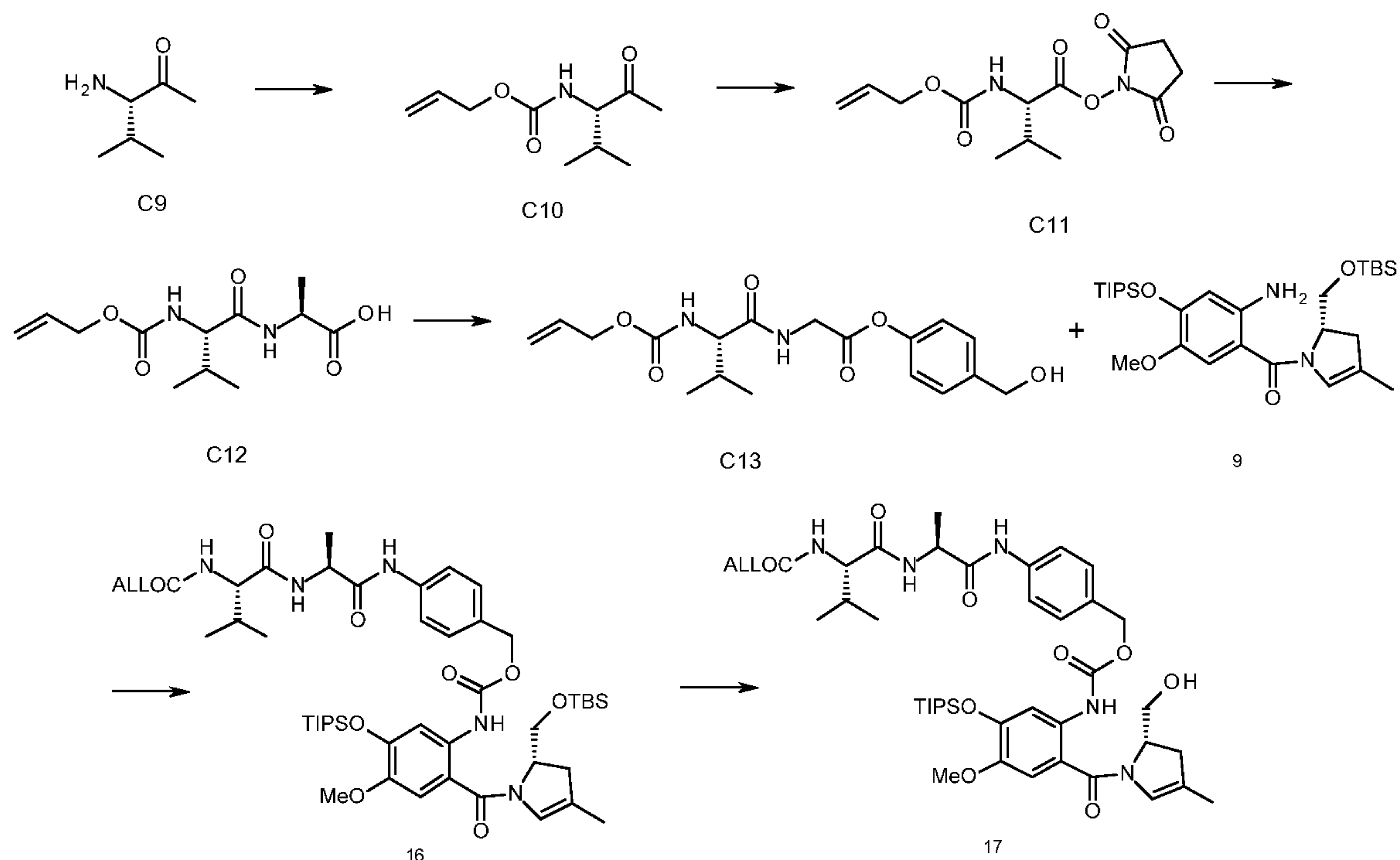
175



1. Charge 400mL of anhydrous Acetone (Analytical Reagent, 10V) and 40g of **14** (1.0eq) to a 1L flask.
2. Charge 136.5g of Diiodopentane (Analytical Reagent) (5.0eq) and particle K_2CO_3 (Analytical Reagent, 1eq) to the flask.
3. Warm to 60°C.
4. Stir for 10 hours.
5. Sample for IPC (completion acceptance criteria: $14 \leq 2.0\%$ by LC-MS).
6. Concentrate the mixture to remove the solvent under vacuum at 30~40°C.
7. The residue subject to flash column chromatography (silica gel; 1/1, v/v, hexane/ethyl acetate).
8. Collect pure fractions and concentrate to remove the solvent under vacuum at 30~40°C.
9. Dry the product in glass bottle using evaporator under vacuum at 30-40°C for 8 hours (internal pressure 20-40mmHg).
10. Product information (49.7g, Purity: 98.6%; Yield: 88.0%). $[\alpha]_D^{28} 60.21^\circ$, (c=0.204g/100ml in Chloroform).

176

(vii) 17



(a) C10

- 5 1. Charge 24g of L-valine **C9** (1.0eq) to a 2 L four-necked, round-bottomed flask equipped with mechanical stirrer and pressure-equalizing addition funnel.
2. Charge 480 mL of THF (analytical reagent, 20V) to the flask.
3. Charge 480 mL of water (20V) to the flask.
4. Cool the solution to 5°C.
- 10 5. Charge 42.5g of powder K₂CO₃ (analytical reagent, 1.5eq) to the flask.
6. Add 29.7 g of Allyl chloroformate (Chemical pure, 1.2eq) dropwise to the flask with stirring at 5~10°C.
7. Remove the cooling bath after the addition is complete.
8. Stir for 4 hours at room temperature.
- 15 9. Sample for IPC (completion acceptance criteria: C9 ≤ 3.0% by HPLC).
10. Evaporate THF under reduced pressure
11. Extract the remaining solution with MTBE (3Vx 2).
12. Acidify the aqueous portion to pH 2 with conc. HCl
13. Extract aqueous solution with DCM (5Vx 3).
- 20 14. Wash the combined organic extracts with brine (4V),
15. Separate the organic phase.
16. Dry over Na₂SO₄.

17. Filter and concentrate the mixture under vacuum at 25~35°C to give a colorless oil (38 g, Purity: 92%, Yield: 88%), $[\alpha]_D^{23}$ -4.5°, (c=0.229 g/100ml in MeOH)

(b) C11

- 5 1. Charge 18g of **C10** (1.0eq) to a 500-mL flask equipped with mechanical stirrer and pressure-equalizing addition funnel.
2. Charge 180 mL of THF (analytical reagent, 10 V).
3. Charge 10.3 g of HOSu (1.0 eq).
4. Cool to 2~7°C.
- 10 5. Add solution of DCC (18.4 g, 1.0 eq) in THF (analytical reagent, 70 mL, 4V) to the mixture drop-wise with stirring.
6. Stir for 3 hours at 2~7°C.
7. Remove cooling bath.
8. Stir at room temperature for 15 hours.
- 15 9. Sample for IPC (Completion acceptance Criteria: C10 ≤2.0% by HPLC).
10. Filter off the precipitate.
11. Wash the filter cake with THF (20ml, 1V).
12. Concentrate the combined filtrate under reduced pressure.
13. Dissolved the residue in DCM (analytical reagent, 3V).
- 20 14. Stir at 0°C for 30 minutes.
15. Filter off the solid.
16. Wash the filter cake with DCM (20ml, 1V).
17. Concentrate the combined filtrate under reduced pressure to give a white solid
18. Product information (26g, Purity: 88%, Yield: 100%).

25 (c) C12

1. Charge 18g of L-alanine (1.0 eq) to a 1 L flask.
2. Charge 8.1g of NaHCO₃ (1.1eq) to the flask.
3. Charge 260 mL of THF (analytical reagent, 10 V).
4. Charge 260 mL of water (10 V).
- 30 5. Cool to 2~7°C.
6. Add solution of **C11** (26 g, 1.0 eq) in THF (100 mL, 4V) to the mixture drop-wise with stirring.
7. Stir for 1 hour at 2~7°C.
8. Remove cooling bath.
- 35 9. Stir for 18 hours at 20~30°C.

10. Sample for IPC (completion criteria: C11 $\leq$ 2.0% by HPLC).
11. Concentrate the reaction mixture under vacuum to 10V at 40~50°C.
12. Acidify the mixture with 20% HCl to pH 2~3 to precipitate the product;
13. Filter and collect the solid.
- 5 14. Wash with water.
15. Dry the product using vacuum oven at 45-50°C for 30 hours (internal pressure around 20-50 mmHg).
16. Product information (solid, 13g, Purity: 89%; Yield: 57%, over 3 steps). $[\alpha]_D^{23}$ -51.71° , (c=0.205 g/100ml in MeOH)

10

(d) C13

1. Charge **C12** (20g, 1.0 eq) to a 1L reactor.
2. Charge *p*-Aminobenzyl alcohol (9.6 g, 1.0eq) to the reactor.
3. Charge THF (400mL, 20V) to the reactor.
- 15 4. Charge EEDQ(19.2g, 1.0 eq) to the reactor
5. Stir for 60 hours at 20-30°C.
6. Sample for IPC (completion criteria: C12 $\leq$ 2.0% by HPLC).
7. Pour the reaction mixture to MTBE (1.2L, 60V) with stirring to precipitate the product.
- 20 8. Filter and collect the solid.
9. Wash the cake with MTBE (1.6L, 0.5V).
10. Dry the product using vacuum oven at 25-30°C for 18 hours to constant amount (internal pressure around 20-50 mmHg).
11. Product information (solid, 20.0g, Purity: 96.0%; Yield: 72%). $[\alpha]_D^{23}$ -83.01° ,
- 25 (c=0.206 g/100ml in MeOH)

(e) 16

1. Charge **9** (40g, 1.0eq) to a 3L flask.
2. Charge DCM (analytical reagent, 400mL, 10V) to the flask under N₂.
- 30 3. Cool to -10°C.
4. Charge triphosgene (analytical reagent, 7.8 g, 0.36 eq) to the mixture under N₂.
5. Add TEA (analytical reagent, 16.2g, 2.2eq) to the mixture drop-wise at -5±5°C.
6. Stir for 10 minutes at -5±5°C.
- 35 7. Sample for IPC to monitor the isocyanate formation (quench with methanol,

completion criteria: $9 \leq 10.0\%$ by LC-MS).

8. Charge solution of the C13 (41g, 1.5 eq) and triethylamine (analytical reagent, 1.5 eq) in dry THF(analytical reagent, 20 V) to the flask in one portion at $-5 \pm 5^\circ\text{C}$.
- 5 9. Stir for 10 minutes at $-5 \pm 5^\circ\text{C}$.
- 10 10. Remove the cooling bath.
11. Stir for 16 hours at $25 \pm 5^\circ\text{C}$.
12. Sample for IPC (quench with methanol, completion criteria: $9 \leq 10.0\%$ by LC-MS).
13. Filter and wash the cake with DCM (analytical reagent, 2V).
14. Concentrate the combined filtrate under vacuum.
15. Subject the residue to column chromatography (silica gel; 3/1, v/v, hexane/ethyl acetate).
16. Collect pure fractions and concentrate under vacuum at $30 \sim 40^\circ\text{C}$.
- 15 17. Dry the product in in vacuum oven at $35\text{-}40^\circ\text{C}$ for 10 hours to constant amount (the internal pressure was around 20-50 mmHg)
18. Product information: off-white solid (36g, Purity: 97.2%; Yield: 53%). $[\alpha]_{\text{D}}^{23} - 86.43^\circ$, (c=0.221g/100ml in Chloroform).
- (f) 17
- 20 1. Charge 6L of AcOH/MeOH/THF/H₂O(10V:1V:1V:2V) to a 10L flask.
2. Charge 416 g of **16** to the flask.
3. Stir for 2 hours at $25^\circ\text{C} \sim 30^\circ\text{C}$.
4. Sample for IPC (completion criteria: $17 \leq 2.0\%$ by LC-MS).
5. Charge EtOAc (Analytical reagent, 30V) and water (20V).
- 25 6. Separate the organic phase and wash with saturated aqueous NaHCO₃(20V).
7. Separate the organic phase and wash with brine (20V).
8. Separate the organic phase and dry over anhydrous Na₂SO₄.
9. Filter and concentrate the organic phase under vacuum.
10. Subject the residue to flash column chromatography (silica gel; hexane/ethyl acetate, 1/2, v/v).
- 30 11. Collect pure fractions and concentrate under vacuum at $30 \sim 40^\circ\text{C}$.
12. Dry the product in vacuum oven at $35\text{-}40^\circ\text{C}$ for 20 hours to constant amount (the internal pressure was around 20-30 mmHg)
13. Product information (308.8g, Purity: 98.1%; Yield: 85.1%).

General Experimental Methods for Example 6

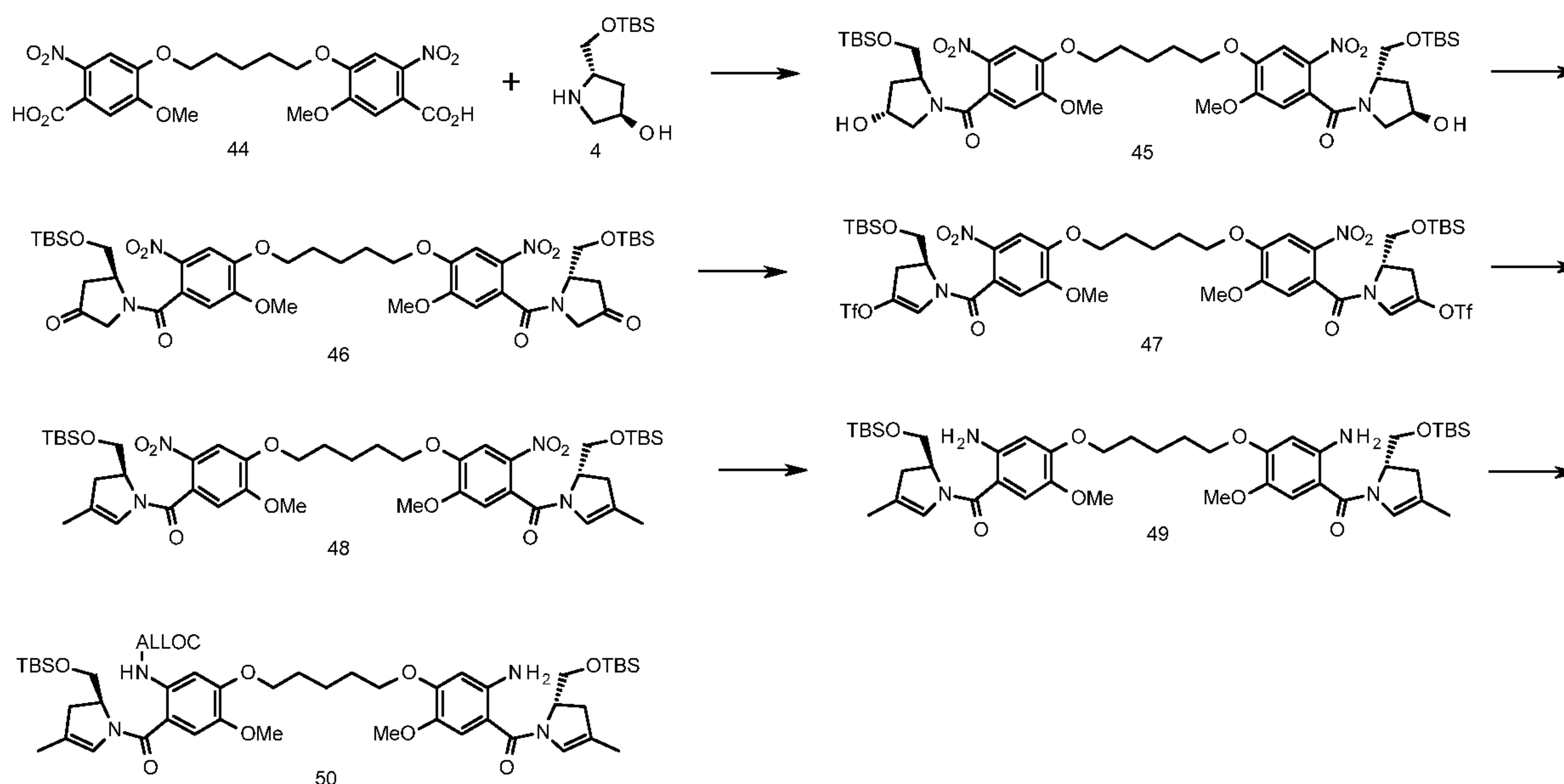
Optical rotations were measured on an ADP 220 polarimeter (Bellingham Stanley Ltd.) and concentrations (*c*) are given in g/100mL. Melting points were measured using a digital melting point apparatus (Electrothermal). IR spectra were recorded on a Perkin-Elmer

5 Spectrum 1000 FT IR Spectrometer. ¹H and ¹³C NMR spectra were acquired at 300 K using a Bruker Avance NMR spectrometer at 400 and 100 MHz, respectively. Chemical shifts are reported relative to TMS (δ = 0.0 ppm), and signals are designated as s (singlet), d (doublet), t (triplet), dt (double triplet), dd (doublet of doublets), ddd (double doublet of doublets) or m (multiplet), with coupling constants given in Hertz (Hz). Mass spectroscopy (MS) data were
10 collected using a Waters Micromass ZQ instrument coupled to a Waters 2695 HPLC with a Waters 2996 PDA. Waters Micromass ZQ parameters used were: Capillary (kV), 3.38; Cone (V), 35; Extractor (V), 3.0; Source temperature (°C), 100; Desolvation Temperature (°C), 200; Cone flow rate (L/h), 50; De-solvation flow rate (L/h), 250. High-resolution mass spectroscopy (HRMS) data were recorded on a Waters Micromass QTOF Global in positive
15 W-mode using metal-coated borosilicate glass tips to introduce the samples into the instrument. Thin Layer Chromatography (TLC) was performed on silica gel aluminium plates (Merck 60, F₂₅₄), and flash chromatography utilised silica gel (Merck 60, 230-400 mesh ASTM). Except for the HOBt (NovaBiochem) and solid-supported reagents (Argonaut), all other chemicals and solvents were purchased from Sigma-Aldrich and were used as
20 supplied without further purification. Anhydrous solvents were prepared by distillation under a dry nitrogen atmosphere in the presence of an appropriate drying agent, and were stored over 4Å molecular sieves or sodium wire. Petroleum ether refers to the fraction boiling at 40-60°C.

25 General LC/MS conditions: The HPLC (Waters Alliance 2695) was run using a mobile phase of water (A) (formic acid 0.1%) and acetonitrile (B) (formic acid 0.1%). Gradient: initial composition 5% B over 1.0 min then 5% B to 95% B over 2.5 min. The composition was held for 0.5 min at 95% B, and then returned to 5% B in 0.1 minutes and held there for 0.9 min. Total gradient run time equals 5 min. Flow rate 3.0 mL/min, 400µL was split *via* a zero
30 dead volume tee piece which passes into the mass spectrometer. Wavelength detection range: 220 to 400 nm. Function type: diode array (535 scans). Column: Phenomenex® Onyx Monolithic C18 50 x 4.60 mm

Example 6

(i) Allyl (5-((5-(5-amino-4-((*S*)-2-(((*tert*-butyldimethylsilyl)oxy)methyl)-4-methyl-2,3-dihydro-1H-pyrrole-1-carbonyl)-2-methoxyphenoxy)pentyl)oxy)-2-((*S*)-2-(((*tert*-butyldimethylsilyl)oxy)methyl)-4-methyl-2,3-dihydro-1H-pyrrole-1-carbonyl)-4-methoxyphenyl)carbamate (**50**)



(a) *(S,R)-((pentane-1,5-diylbis(oxy))bis(5-methoxy-2-nitro-4,1-phenylene))bis(((2*S*,4*R*)-2-(((*tert*-butyldimethylsilyl)oxy)methyl)-4-hydroxypyrrolidin-1-yl)methanone) (**45**)*

Anhydrous DMF (approx. 0.5 mL) was added dropwise to a stirred suspension of 4,4'-
 10 (pentane-1,5-diylbis(oxy))bis(5-methoxy-2-nitrobenzoic acid) (**44**) (36.64 g, 74.0 mmol) and
 oxalyl chloride (18.79 mL, 0.222 mol, 3.0 eq.) in anhydrous DCM (450 mL) until vigorous
 effervescence occurred and the reaction mixture was left to stir overnight. The reaction
 mixture was evaporated to dryness, and triturated with diethyl ether. The resulting yellow
 precipitate was filtered from solution, washed with diethyl ether (100 mL) and immediately
 15 added to a solution of (3*R*,5*S*)-5-((*tert*-butyldimethylsilyloxy)methyl) pyrrolidin-3-ol (**4**) (39.40
 g, 0.170 mol, 2.3 eq.) and anhydrous triethylamine (82.63 mL, 0.592 mol, 8 eq.) in
 anhydrous DCM (400 mL) at -40°C. The reaction mixture was allowed to slowly warm to
 room temperature (over 2.5 hours) after which, LCMS analysis indicated complete reaction.
 DCM (250 mL) was added and the mixture was transferred into a separating funnel. The
 20 organic layer was washed successively with 0.1M HCl (2 x 800 mL), saturated NaHCO₃ (500
 mL) and brine (300 mL). After drying over MgSO₄ and filtration, evaporation of the solvent
 left the product as a yellow foam (62.8 g, 92%). LC/MS, System 1: RT 1.96 min; MS (ES+)
m/z (relative intensity) 921.45 ([*M*+*H*]⁺, 100).

(b) (5S,5'S)-1,1'-(4,4'-(pentane-1,5-diylbis(oxy))bis(5-methoxy-2-nitrobenzoyl))bis(5-(((tert-butyl)dimethylsilyl)oxy)methyl)pyrrolidin-3-one) (**46**)

Trichloroisocyanuric acid (21.86 g, 94.07 mmol, 1.4 eq) was added in one portion to a solution of diol **45** (61.90 g, 67.20 mmol) and TEMPO (2.10 g, 13.44 mmol, 0.2 eq) in anhydrous DCM (500 mL) under an atmosphere of argon at 0°C. The reaction mixture was stirred at 0°C for 20 minutes after which, LCMS analysis of the reaction mixture showed complete reaction. The reaction mixture was diluted with DCM (400 mL) and washed with saturated sodium bicarbonate (500 mL), 0.2 M sodium thiosulfate solution (600 mL), brine (400 mL) and dried (MgSO₄). Evaporation of the solvent gave the crude product. Flash chromatography [gradient elution 80% n-hexane/20% ethyl acetate to 100% ethyl acetate] gave pure **46** as yellow solid (49.30 g, 80%). LC/MS: RT 2.03 min; MS (ES+) *m/z* (relative intensity) 917.55 ([M+H]⁺, 100).

(c) (5S,5'S)-1,1'-(4,4'-(pentane-1,5-diylbis(oxy))bis(5-methoxy-2-nitrobenzoyl))bis(5-(((tert-butyl)dimethylsilyl)oxy)methyl)-4,5-dihydro-1H-pyrrole-3,1-diyl) bis(trifluoromethanesulfonate), (**47**)

Triflic anhydride (24.19 mL, 0.144 mol, 6.0 eq) was added dropwise to a vigorously stirred solution of bis-ketone **46** (21.98 g, 23.96 mmol) in anhydrous DCM (400 mL) containing 2,6-lutidine (22.33 mL, 0.192 mol, 8.0 eq) at -40 °C. The reaction mixture was stirred at -40 °C for 30 min after which, LCMS analysis indicated complete reaction. Reaction mixture was rapidly diluted with DCM (500 mL) and washed with ice-cold water (600 mL), ice-cold saturated sodium bicarbonate (400 mL) and brine (500 mL), dried over MgSO₄, filtered and evaporated to leave a crude brown oil. Flash chromatography [gradient elution 80% n-hexane/20% ethyl acetate to 66% n-hexane/33% ethyl acetate] gave pure **47** as a brown foam (16.40 g, 58%). LC/MS: RT 2.28 min; MS (ES+) *m/z* (relative intensity) no data.

(d) (S)-((pentane-1,5-diylbis(oxy))bis(5-methoxy-2-nitro-4,1-phenylene))bis(((S)-2-(((tert-butyl)dimethylsilyl)oxy)methyl)-4-methyl-2,3-dihydro-1H-pyrrol-1-yl)methanone) (**48**)

Triflate **47** (5.06 g, 4.29 mmol), methyl boronic acid (1.80 g, 30.00 mmol, 7 eq) and triphenylarsine (1.05 g, 3.43 mmol, 0.8 eq) were dissolved in anhydrous dioxane and stirred under argon. Pd (II) bisbenzonitrile chloride was then added and the reaction mixture heated rapidly to 80°C for 20 minutes. Reaction mixture cooled, filtered through Celite (washed through with ethyl acetate), filtrate washed with water (500 mL), brine (500 mL), dried over MgSO₄, filtered and evaporated. Flash chromatography [gradient elution 50% n-hexane/50% ethyl acetate] gave pure **48** as a brown foam (4.31 g, 59%). LC/MS: RT 2.23 min; MS (ES+) *m/z* (relative intensity) 913.50 ([M+H]⁺, 100).

(e) (S)-((pentane-1,5-diylbis(oxy))bis(2-amino-5-methoxy-4,1-phenylene))bis(((S)-2-(((tert-butyl)dimethylsilyl)oxy)methyl)-4-methyl-2,3-dihydro-1H-pyrrol-1-yl)methanone) (**49**)

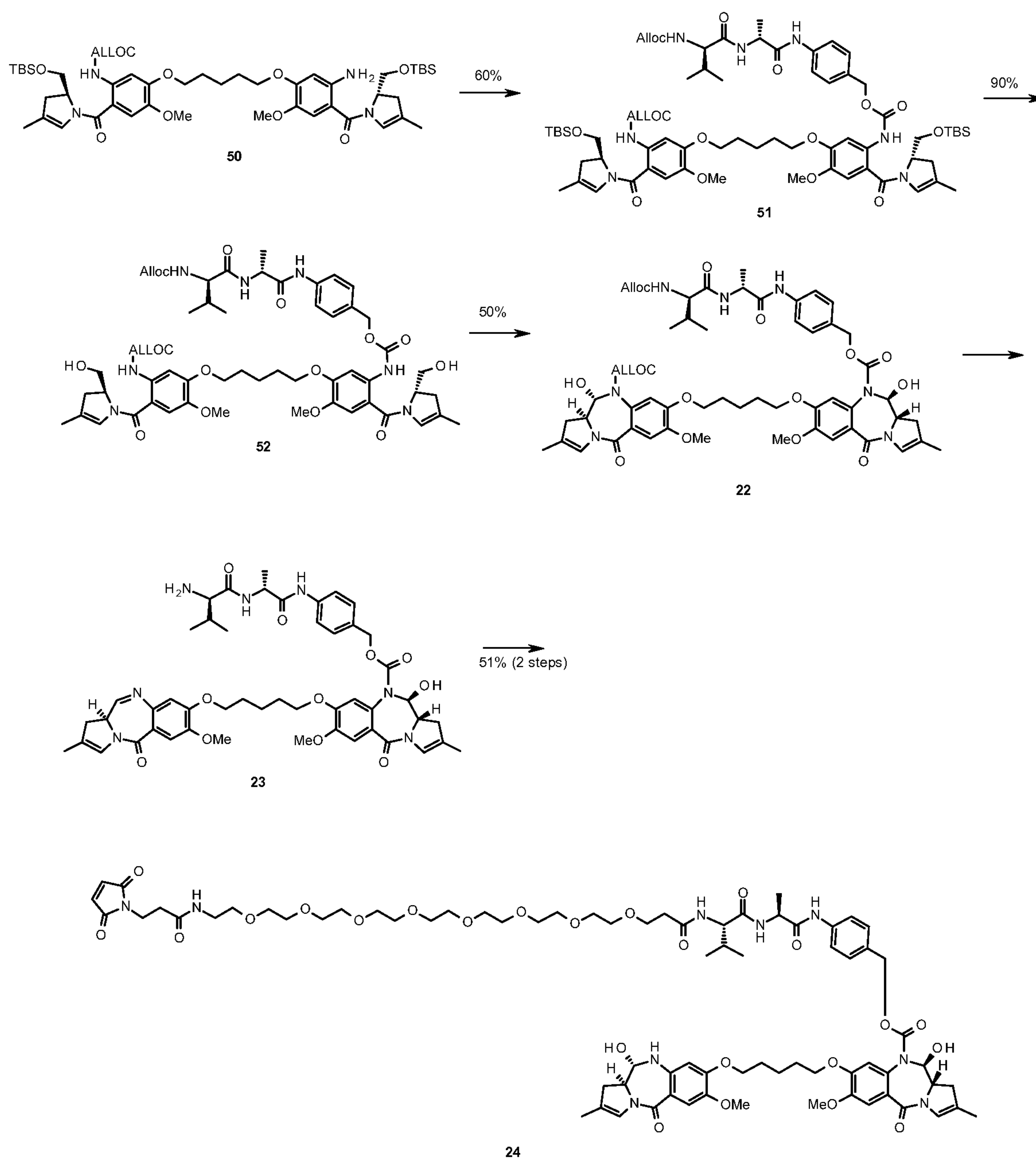
Zinc dust (26.48 g, 0.405 mol, 36.0 eq) was added in one portion to a solution of bis-nitro compound **48** (10.26 g, 11.24 mmol) in 5% formic acid / methanol (200 mL) keeping the temperature between 25-30°C with the aid of a cold water bath. The reaction was stirred at 30°C for 20 minutes after which, LCMS showed complete reaction. The reaction mixture was filtered through Celite to remove the excess zinc, which was washed with ethyl acetate (600 mL). The organic fractions were washed with water (500 mL), saturated sodium bicarbonate (500 mL) and brine (400 mL), dried over MgSO₄ and evaporated. Flash chromatography [gradient elution 100% chloroform to 99% chloroform/1% methanol] gave pure **49** as an orange foam (6.22 g, 65%). LC/MS: RT 2.20 min; MS (ES+) *m/z* (relative intensity) 853.50 ([M+H]⁺, 100).

(f) Allyl (5-((5-(5-amino-4-((S)-2-(((tert-butyl)dimethylsilyl)oxy)methyl)-4-methyl-2,3-dihydro-1H-pyrrole-1-carbonyl)-2-methoxyphenoxy)pentyl)oxy)-2-((S)-2-(((tert-butyl)dimethylsilyl)oxy)methyl)-4-methyl-2,3-dihydro-1H-pyrrole-1-carbonyl)-4-methoxyphenyl)carbamate (**50**)

Pyridine (1.156 mL, 14.30 mmol, 1.5 eq) was added to a solution of the bis-aniline **49** (8.14 g, 9.54 mmol) in anhydrous DCM (350 mL) at -78°C under an atmosphere of argon. After 5 minutes, allyl chloroformate (0.911 mL, 8.58 mmol, 0.9 eq) was added and the reaction mixture allowed to warm to room temperature. The reaction mixture was diluted with DCM (250 mL), washed with saturated CuSO₄ solution (400 mL), saturated sodium bicarbonate (400 mL) and brine (400 mL), dried over MgSO₄. Flash chromatography [gradient elution 66% n-hexane/33% ethyl acetate to 33% n-hexane/66% ethyl acetate] gave pure **50** as an orange foam (3.88 g, 43%). LC/MS: RT 2.27 min; MS (ES+) *m/z* (relative intensity) 937.55 ([M+H]⁺, 100).

(ii) Outline scheme to (11S,11aS)-4-((2S,5S)-37-(2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)-5-isopropyl-2-methyl-4,7,35-trioxo-10,13,16,19,22,25,28,31-octaoxa-3,6,34-triazaheptatriacontanamido)benzyl 11-hydroxy-7-methoxy-8-((5-(((S)-7-methoxy-2-methyl-5-oxo-5,11a-dihydro-1H-benzo[e]pyrrolo[1,2-a][1,4]diazepin-8-yl)oxy)pentyl)oxy)-2-methyl-5-oxo-11,11a-dihydro-1H-benzo[e]pyrrolo[1,2-a][1,4]diazepine-10(5H)-carboxylate (**24**)

184



Compound 24 can be synthesised according to the scheme shown above, using the methods described in earlier examples.

5

Example 7: Activity of released compounds

K562 assay

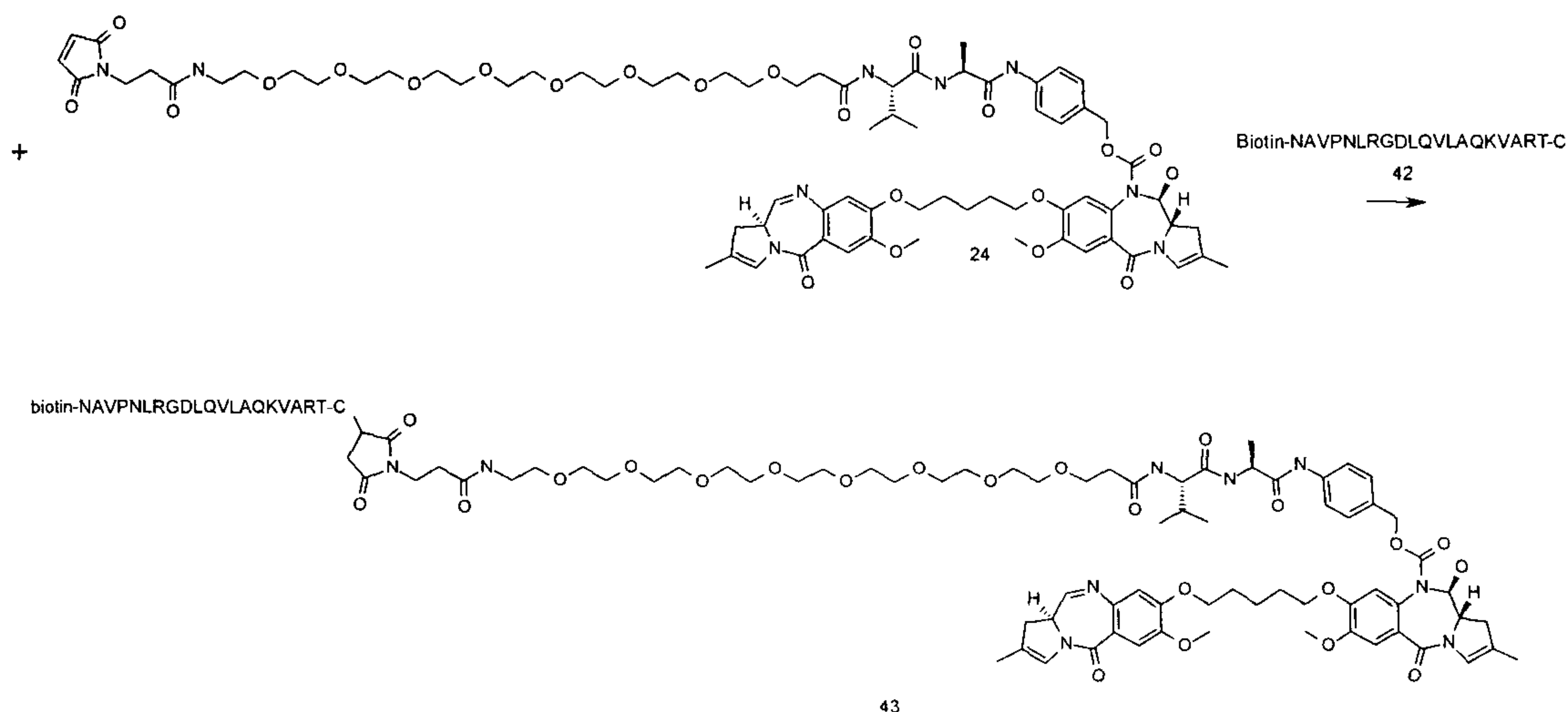
K562 human chronic myeloid leukaemia cells were maintained in RPM1 1640 medium supplemented with 10% fetal calf serum and 2 mM glutamine at 37°C in a humidified atmosphere containing 5% CO₂ and were incubated with a specified dose of drug for 1 hour or 96 hours at 37°C in the dark. The incubation was terminated by centrifugation (5 min, 300

g) and the cells were washed once with drug-free medium. Following the appropriate drug treatment, the cells were transferred to 96-well microtiter plates (10^4 cells per well, 8 wells per sample). Plates were then kept in the dark at 37°C in a humidified atmosphere containing 5% CO₂. The assay is based on the ability of viable cells to reduce a yellow soluble tetrazolium salt, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2*H*-tetrazolium bromide (MTT, Aldrich-Sigma), to an insoluble purple formazan precipitate. Following incubation of the plates for 4 days (to allow control cells to increase in number by approximately 10 fold), 20 µL of MTT solution (5 mg/mL in phosphate-buffered saline) was added to each well and the plates further incubated for 5 h. The plates were then centrifuged for 5 min at 300 g and the bulk of the medium pipetted from the cell pellet leaving 10–20 µL per well. DMSO (200 µL) was added to each well and the samples agitated to ensure complete mixing. The optical density was then read at a wavelength of 550 nm on a Titertek Multiskan™ ELISA plate reader, and a dose-response curve was constructed. For each curve, an IC₅₀ value was read as the dose required to reduce the final optical density to 50% of the control value.

15

Compound RelB has an IC₅₀ of 0.425 nM in this assay.

Example 8



20 Biotin-A20FMDV-Cys-2 (43)

A solution of the peptide **42** (12.06 mg, 4.8 µmol, 1.0 eq) in 1/1 acetonitrile/water (1 mL) was added to a solution of **24** (12.06 mg, 4.8 µmol, 1.0 eq) in 1/1 acetonitrile /water (1 mL). The solution was stirred at room temperature for 24 hours. The acetonitrile/water was removed by lyophilisation to give a yellow foam. Purification by preparative HPLC followed by lyophilisation gave the product as a white foam (7.4 mg, 38%)

25

Preparative HPLC method: Reverse-phase ultra-high-performance liquid chromatography (UPLC) was carried out on Phenomenex Gemini NX 5 μ C-18 columns of the following dimensions: 150 x 4.6 mm for analysis, and 150 x 21.20 mm for preparative work. All UPLC experiments were performed with gradient conditions: initial fixed composition 13% B to 75% B over 15 min, held for 2.0 min at 75% B, then 75% B to 13% B within 0.10 min held at 13% for 2.90 min. Total duration of gradient run was 20.00 min. Eluents used were solvent A (H₂O with 0.1% Formic acid) and solvent B (CH₃CN with 0.1% Formic acid). Flow rates used were 1.0 ml/min for analytical, and 20.0 ml/min for preparative HPLC. Detection was at 254 and 280 nm.

Analytical Data: LCMS 1.17 min (ES+) *m/z* (relative intensity) 1330([M + 2]⁺/3, 5%); 998([M + 3]⁺/4, 70); 798([M + 4]⁺/5, 100); 665([M + 5]⁺/6, 20).

Example 9: Formation of conjugates

General antibody conjugation procedure

Antibodies are diluted to 1-5 mg/mL in a reduction buffer (examples: phosphate buffered saline PBS, histidine buffer, sodium borate buffer, TRIS buffer). A freshly prepared solution of TCEP (tris(2-carboxyethyl)phosphine hydrochloride) is added to selectively reduce cysteine disulfide bridges. The amount of TCEP is proportional to the target level of reduction, within 1 to 4 molar equivalents per antibody, generating 2 to 8 reactive thiols. After reduction for several hours at 37°C, the mixture is cooled down to room temperature and excess drug-linker (**A**, **B**) added as a diluted DMSO solution (final DMSO content of up to 10% volume/volume of reaction mixture). The mixture is gently shaken at either 4°C or room temperature for the appropriate time, generally 1-3 hours for compound **B** and 10-30 hours for compound **A**. Excess reactive thiols can be reacted with a 'thiol capping reagent' like N-ethyl maleimide (NEM) at the end of the conjugation. Antibody-drug conjugates are concentrated using centrifugal spin-filters with a molecular weight cut-off of 10 kDa or higher, then purified by tangential flow filtration (TFF) or Fast Protein Liquid Chromatography (FPLC). Corresponding antibody-drug conjugates can be determined by analysis by High-Performance Liquid Chromatography (HPLC) or Ultra-High-Performance Liquid Chromatography (UHPLC) to assess drug-per-antibody ratio (DAR) using reverse-phase chromatography (RP) or Hydrophobic-Interaction Chromatography (HIC), coupled with UV-Visible, Fluorescence or Mass-Spectrometer detection; aggregate level and monomer purity can be analysed by HPLC or UHPLC using size-exclusion chromatography coupled with UV-Visible, Fluorescence or Mass-Spectrometer detection. Final conjugate concentration is determined by a combination of spectroscopic (absorbance at 280, 214 and 330 nm) and

biochemical assay (bicinchonic acid assay BCA; Smith, P.K., *et al.* (1985) *Anal. Biochem.* **150** (1): 76–85; using a known-concentration IgG antibody as reference). Antibody-drug conjugates are generally sterile filtered using 0.2 µm filters under aseptic conditions, and stored at +4°C, -20°C or -80°C.

5

Examples of particular conjugations are described below.

ADC1A

Antibody 1 (15 mg, 102 nanomoles) is diluted into 12.0 mL of a reduction buffer containing
10 10 mM sodium borate pH 8.4, 2.5 mM EDTA and a final antibody concentration of 1.25
mg/mL. A 10 mM solution of TCEP is added (2 molar equivalent/antibody, 204 nanomoles,
20 µL) and the reduction mixture is heated at 37°C for two hour in an orbital incubator. After
cooling down to room temperature, compound **A** is added as a DMSO solution (5 molar
equivalent/antibody, 510 nanomoles, in 1.2 mL DMSO). The solution is mixed 18 hour at
15 room temperature, then transferred into a 15 mL Amicon Ultracell™ 50kDa MWCO spin
filter, concentrated to ca. 2.0 mL and injected into a AKTA™FPLC using a GE Healthcare
XK16/70 column packed with Superdex™ 200 PG, eluting with 1.5 mL/min of sterile-filtered
Phosphate-buffered saline (PBS). Fractions corresponding to the desired ADC monomer
peak are pooled, analysed and sterile-filtered. BCA assay gives a concentration of final
20 **ADC1A** at 0.67 mg/mL in 12.5 mL, and the obtained mass is 8.4 mg (56% yield). HPLC
analysis on a Waters Alliance system using a Agilent PLRP-S 1000 A 8µm 150 x 2.1 mm
column eluting with a gradient of water and acetonitrile on a reduced sample of **ADC1A** at
280 nm shows a mixture of light and heavy chains attached to several molecules of **A**,
consistent with a drug-per-antibody ratio (DAR) of 2.2 molecules of **A** per antibody. SEC
25 analysis on an AKTA™FPLC using a GE Healthcare XK16/70 column packed with Superdex
200 PG, eluting with sterile-filtered Phosphate-buffered saline (PBS) on a sample of **ADC1A**
at 280 nm shows a monomer purity of 95.3% with 4.7% aggregates.

ADC1B

30 Antibody 1(15 mg, 102 nanomoles) is diluted into 12.0 mL of a reduction buffer containing 10
mM sodium borate pH 8.4, 2.5 mM EDTA and a final antibody concentration of 1.25 mg/mL.
A 10 mM solution of TCEP is added (2 molar equivalent/antibody, 204 nanomoles, 20 µL)
and the reduction mixture was heated at 37°C for two hour in an orbital incubator. After
cooling down to room temperature, Compound **B** is added as a DMSO solution (5 molar
35 equivalent/antibody, 510 nanomoles, in 1.2 mL DMSO). The solution is mixed 5 hour at room

temperature, then a second amount of Compound **B** is added as a DMSO solution (2 molar equivalent/antibody, 200 nanomoles, in 0.6 mL DMSO). The solution is mixed 15 hour at room temperature, then transferred into a 15 mL Amicon Ultracell 50 kDa MWCO spin filter, concentrated to ca. 2.0 mL and injected into an AKTA™FPLC using a GE Healthcare

5 XK16/70 column packed with Superdex 200 PG, eluting with 1.5 mL/min of sterile-filtered Phosphate-buffered saline (PBS). Fractions corresponding to **ADC1B** monomer peak are pooled, analysed and sterile-filtered. BCA assay gives a concentration of final **ADC1B** at 1.02 mg/mL in 10.5 mL, and the obtained mass is 10.7 mg (71% yield). HPLC analysis on a Waters Alliance system using a Agilent PLRP-S 1000 A 8um 150 x 2.1 mm column eluting

10 with a gradient of water and acetonitrile on a reduced sample of **ADC1B** at 280 nm and 330 nm (drug-linker specific) shows a mixture of light and heavy chains attached to several molecules of **B**, consistent with a drug-per-antibody ratio (DAR) of 2.2 molecules of **B** per antibody. SEC analysis on a AKTA™FPLC using a GE Healthcare XK16/70 column packed with Superdex 200 PG, eluting with sterile-filtered Phosphate-buffered saline (PBS) on a

15 sample of **ADC1B** at 280 nm shows a monomer purity of 94.9% with 5.1% aggregates.

As used herein, "Antibody 1" is an anti-Her2 antibody comprising a VH domain having the sequence according to SEQ ID NO. 1 and a VL domain having the sequence according to SEQ ID NO. 2.

20

Example 10: In vivo ADC efficacy studies

CB.17 SCID mice, aged 8-12 weeks, are injected with 1 mm³ tumour fragments derived from the BT-474 cell line sub cutaneously in the flank. When tumours reach an average size of 100 - 150 mm³, treatment is begun. Mice are weighed twice a week. Tumour size is

25 measured twice a week. Animals are monitored individually. The endpoint of the experiment is a tumour volume of 1000 mm³ or 60 days, whichever comes first. Responders can be followed longer.

Groups of 10 xenografted mice are injected i.v. with 0.2ml of antibody drug conjugate (ADC),

30 or naked antibody, in phosphate buffered saline (vehicle) or with 0.2ml of vehicle alone. The concentration of ADC is adjusted to give, for example, 0.3 or 1.0 mg ADC/ kg body weight in a single dose. Three identical doses may be given to each mouse at intervals of, for example, 1 week.

35 Figure 1 shows the effect on mean tumour volume in groups of 10 miced dosed with **ADC1A** at 0.3 (green) or 1.0 mg/kg (red) compared to vehicle (black) or naked Ig (blue) controls.

Figure 2 shows the effect on mean tumour volume in groups of 10 miced dosed with **ADC1B** at 0.3 (light blue) or 1.0 mg/kg (green) compared to vehicle (black) or naked Ig (dark blue) controls.

5

All documents and other references mentioned above are herein incorporated by reference.

Abbreviations

	Ac	acetyl
10	Acm	acetamidomethyl
	Alloc	allyloxycarbonyl
	Boc	di- <i>tert</i> -butyl dicarbonate
	t-Bu	tert-butyl
	Bzl	benzyl, where Bzl-OMe is methoxybenzyl and Bzl-Me is methylbenzene
15	Cbz or Z	benzyloxy-carbonyl, where Z-Cl and Z-Br are chloro- and bromobenzyloxy carbonyl respectively
	DMF	<i>N,N</i> -dimethylformamide
	Dnp	dinitrophenyl
	DTT	dithiothreitol
20	Fmoc	9 <i>H</i> -fluoren-9-ylmethoxycarbonyl
	imp	<i>N</i> -10 imine protecting group: 3-(2-methoxyethoxy)propanoate-Val-Ala-PAB
	MC-OSu	maleimidocaproyl- <i>O</i> - <i>N</i> -succinimide
	Moc	methoxycarbonyl
	MP	maleimidopropanamide
25	Mtr	4-methoxy-2,3,6-trimethylbenzenesulfonyl
	PAB	para-aminobenzyloxycarbonyl
	PEG	ethyleneoxy
	PNZ	<i>p</i> -nitrobenzyl carbamate
	Psec	2-(phenylsulfonyl)ethoxycarbonyl
30	TBDMS	tert-butyldimethylsilyl
	TBDPS	tert-butyldiphenylsilyl
	Teoc	2-(trimethylsilyl)ethoxycarbonyl
	Tos	tosyl
	Troc	2,2,2-trichlorethoxycarbonyl chloride
35	Trt	trityl
	Xan	xanthyl

SEQUENCES

SEQ ID NO. 1 (Her VH):

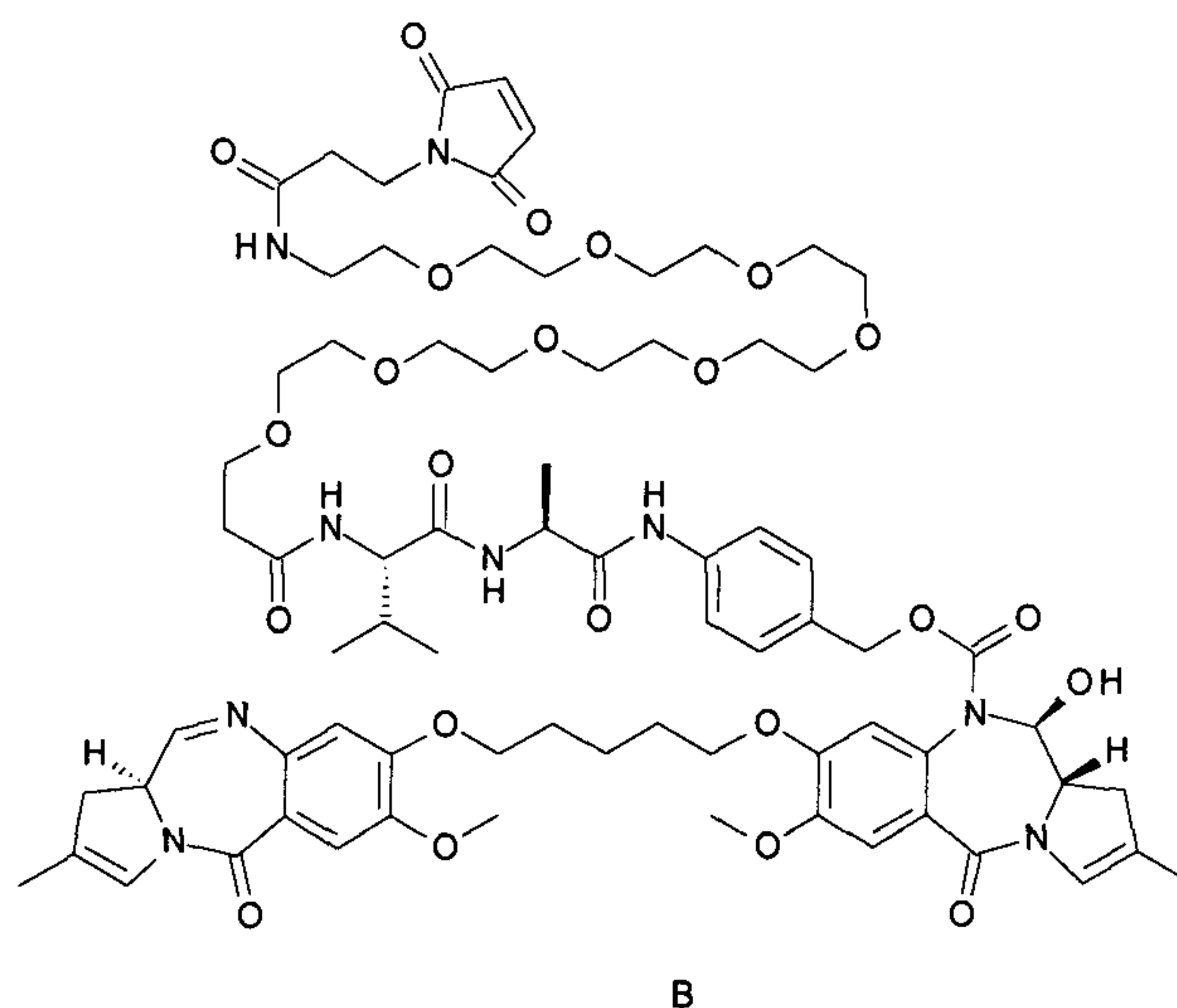
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5 ADSVKGRFTISADTSKNTAYLQMNSLRAEDTAVYYCSRWGGDGFYAMDYWGQGTLVTVS
S

SEQ ID NO. 2 (Her VL):

DIQMTQSPSSLSASVGDRVTITCRASQDVNTAVAWYQQKPGKAPKLLIYSASFLYSGVPSR
10 FSGSRSGTDFTLTISLQPEDFATYYCQQHYTTPPTFGQGGTKVEIK

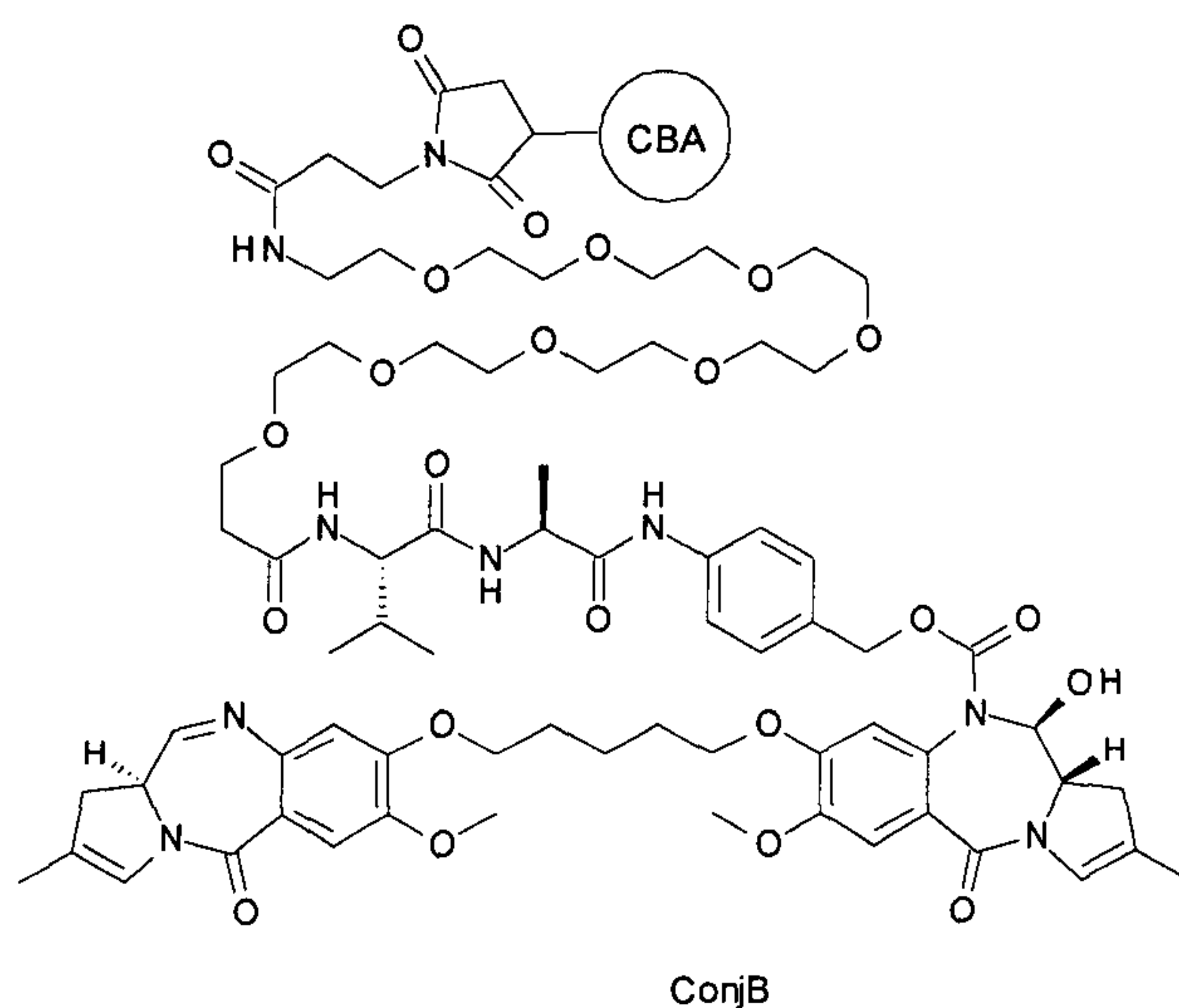
Claims

1. A compound B:



5 and salts and solvates thereof.

2. A conjugate of formula ConjB:



where CBA represents a cell binding agent.

10

3. The conjugate according to claim 2, wherein the cell binding agent is an antibody or an active fragment thereof.

4. The conjugate according to claim 3, wherein the antibody or antibody fragment is an
15 antibody or antibody fragment for a tumour-associated antigen.

15

5. The conjugate of claim 3, wherein the antibody or antibody fragment is an antibody which binds to one or more tumor-associated antigens or cell-surface receptors each independently being:

- (1) BMPR1B;
- 5 (2) E16;
- (3) STEAP1;
- (4) 0772P;
- (5) MPF;
- (6) Napi3b;
- 10 (7) Sema 5b;
- (8) PSCA hlg;
- (9) ETBR;
- (10) MSG783;
- (11) STEAP2;
- 15 (12) TrpM4;
- (13) CRIPTO;
- (14) CD21;
- (15) CD79b;
- (16) FcRH2;
- 20 (17) HER2;
- (18) NCA;
- (19) MDP;
- (20) IL20R-alpha;
- (21) Brevican;
- 25 (22) EphB2R;
- (23) ASLG659;
- (24) PSCA;
- (25) GEDA;
- (26) BAFF-R;
- 30 (27) CD22;
- (28) CD79a;
- (29) CXCR5;
- (30) HLA-DOB;
- (31) P2X5;
- 35 (32) CD72;
- (33) LY64;

- (34) FcRH1;
- (35) IRTA2;
- (36) TENB2;
- (37) PSMA – FOLH1;
- 5 (38) SST;
- (38.1) SSTR2;
- (38.2) SSTR5;
- (38.3) SSTR1;
- (38.4) SSTR3;
- 10 (38.5) SSTR4;
- (39) ITGAV;
- (40) ITGB6;
- (41) CEACAM5;
- (42) MET;
- 15 (43) MUC1;
- (44) CA9;
- (45) EGFRvIII;
- (46) CD33;
- (47) CD19;
- 20 (48) IL2RA;
- (49) AXL;
- (50) CD30 - TNFRSF8;
- (51) BCMA - TNFRSF17;
- (52) CT Ags – CTA;
- 25 (53) CD174 (Lewis Y) - FUT3;
- (54) CLEC14A;
- (55) GRP78 – HSPA5;
- (56) CD70;
- (57) Stem Cell specific antigens;
- 30 (58) ASG-5;
- (59) ENPP3;
- (60) PRR4;
- (61) GCC – GUCY2C;
- (62) Liv-1 – SLC39A6;
- 35 (63) 5T4;
- (64) CD56 – NCMA1;

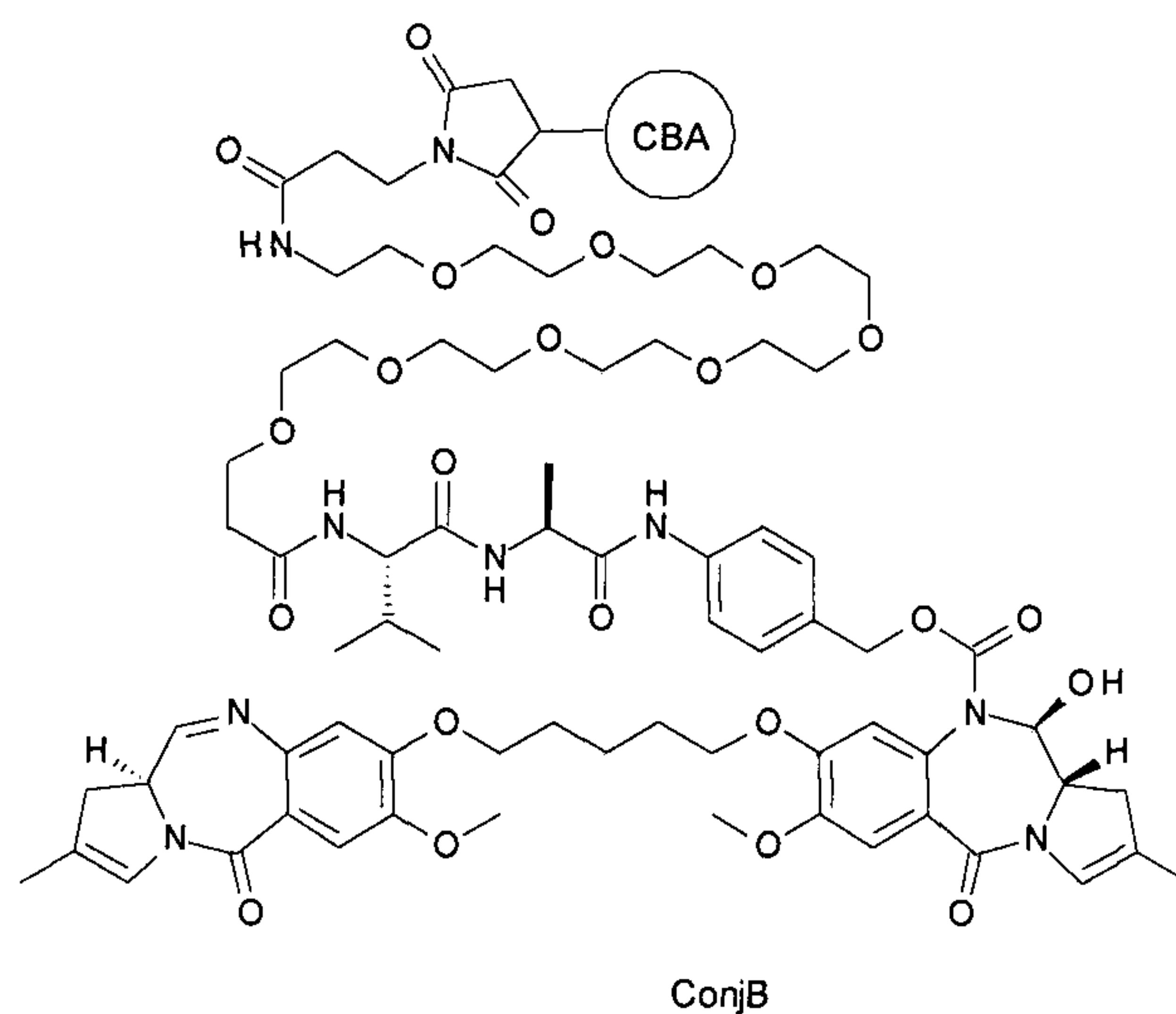
- (65) CanAg;
- (66) FOLR1;
- (67) GPNMB;
- (68) TIM-1 – HAVCR1;
- 5 (69) RG-1/Prostate tumor target Mindin – Mindin/RG-1;
- (70) B7-H4 – VTCN1;
- (71) PTK7;
- (72) CD37;
- (73) CD138 – SDC1;
- 10 (74) CD74;
- (75) Claudins – CLs;
- (76) EGFR;
- (77) Her3;
- (78) RON - MST1R;
- 15 (79) EPHA2;
- (80) CD20 – MS4A1;
- (81) Tenascin C – TNC;
- (82) FAP;
- (83) DKK-1;
- 20 (84) CD52;
- (85) CS1 - SLAMF7;
- (86) Endoglin – ENG;
- (87) Annexin A1 – ANXA1; or
- (88) V-CAM (CD106) - VCAM1.

25

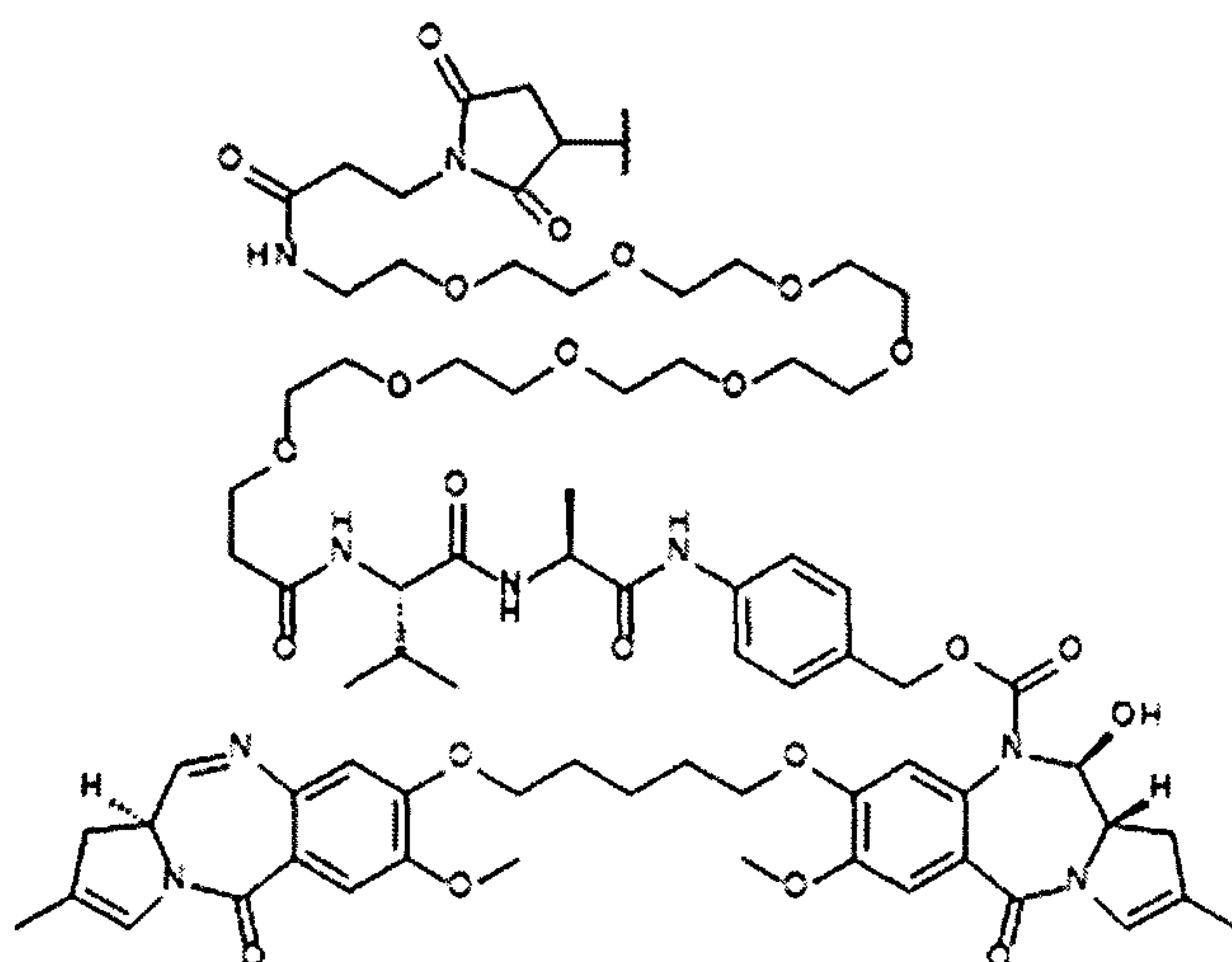
6. The conjugate of any one of claims 3 to 5, wherein the antibody or antibody fragment is a cysteine-engineered antibody.

195

7. A conjugate ConjB:



where CBA represents a cell binding agent, and where the drug loading of drugs D:



5

to cell binding agent is an integer from 1 to 8.

8. The conjugate according to claim 7, wherein the drug loading per cell binding agent
10 is 1, 2, 3, or 4.

9. A composition comprising a mixture of conjugates as defined in claim 7.

10. The conjugate according to any one of claims 2 to 8 for use in the treatment of a proliferative disease in a subject.
11. The composition according to claim 9 for use in the treatment of a proliferative
5 disease in a subject.
12. The conjugate according to claim 10, wherein the proliferative disease is cancer.
13. The composition according to claim 11, wherein the proliferative disease is cancer.
10
14. A pharmaceutical composition comprising the conjugate as defined in any one of claims 2 to 8 or the composition as defined in claim 9 and a pharmaceutically acceptable diluent, carrier or excipient.
- 15 15. The pharmaceutical composition of claim 14 further comprising a therapeutically effective amount of a chemotherapeutic agent.
16. A method of preparing a conjugate as defined in any one of claims 2 to 8, the method comprising the step of reacting a cell binding agent with compound B as defined in claim 1.

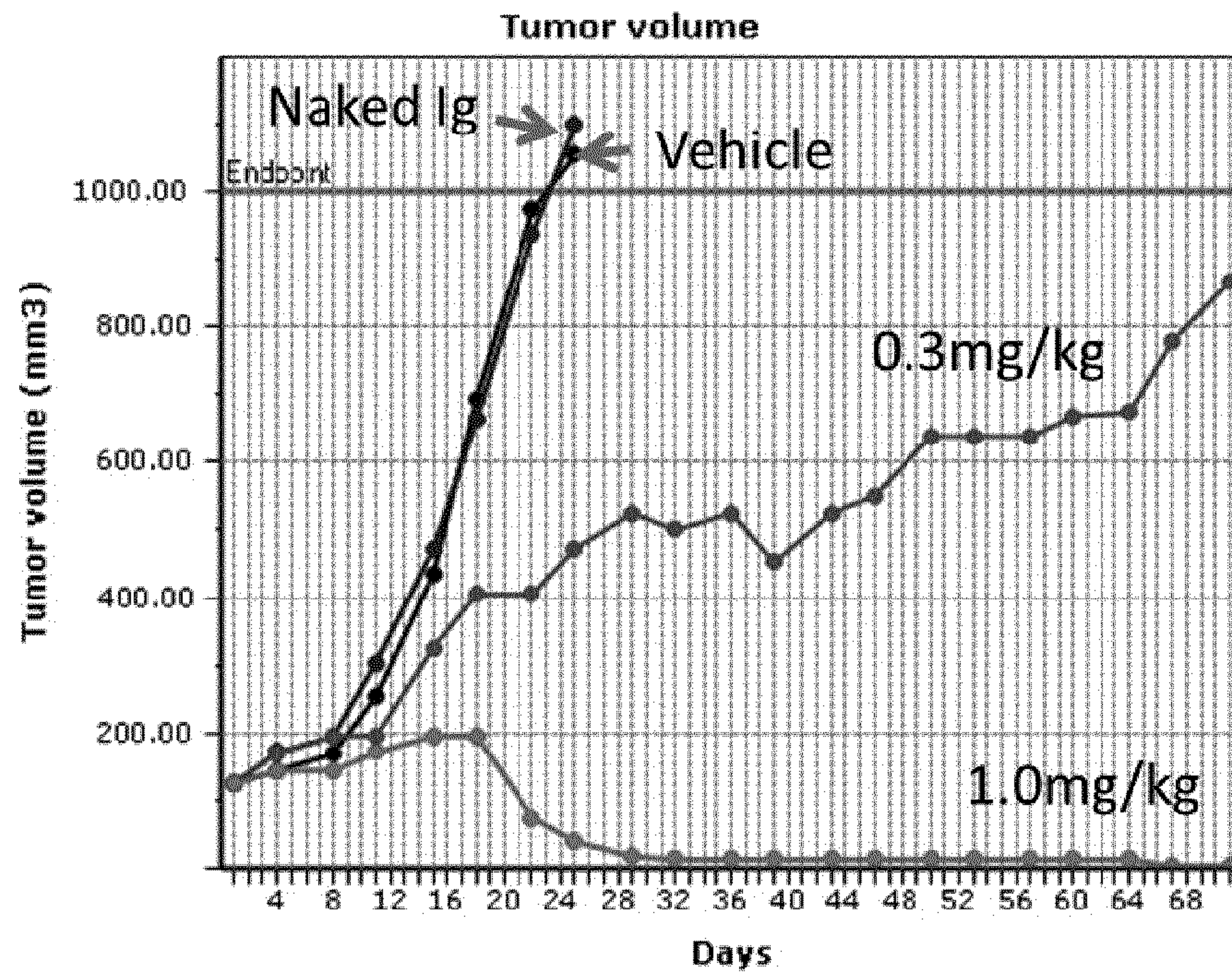


Figure 1

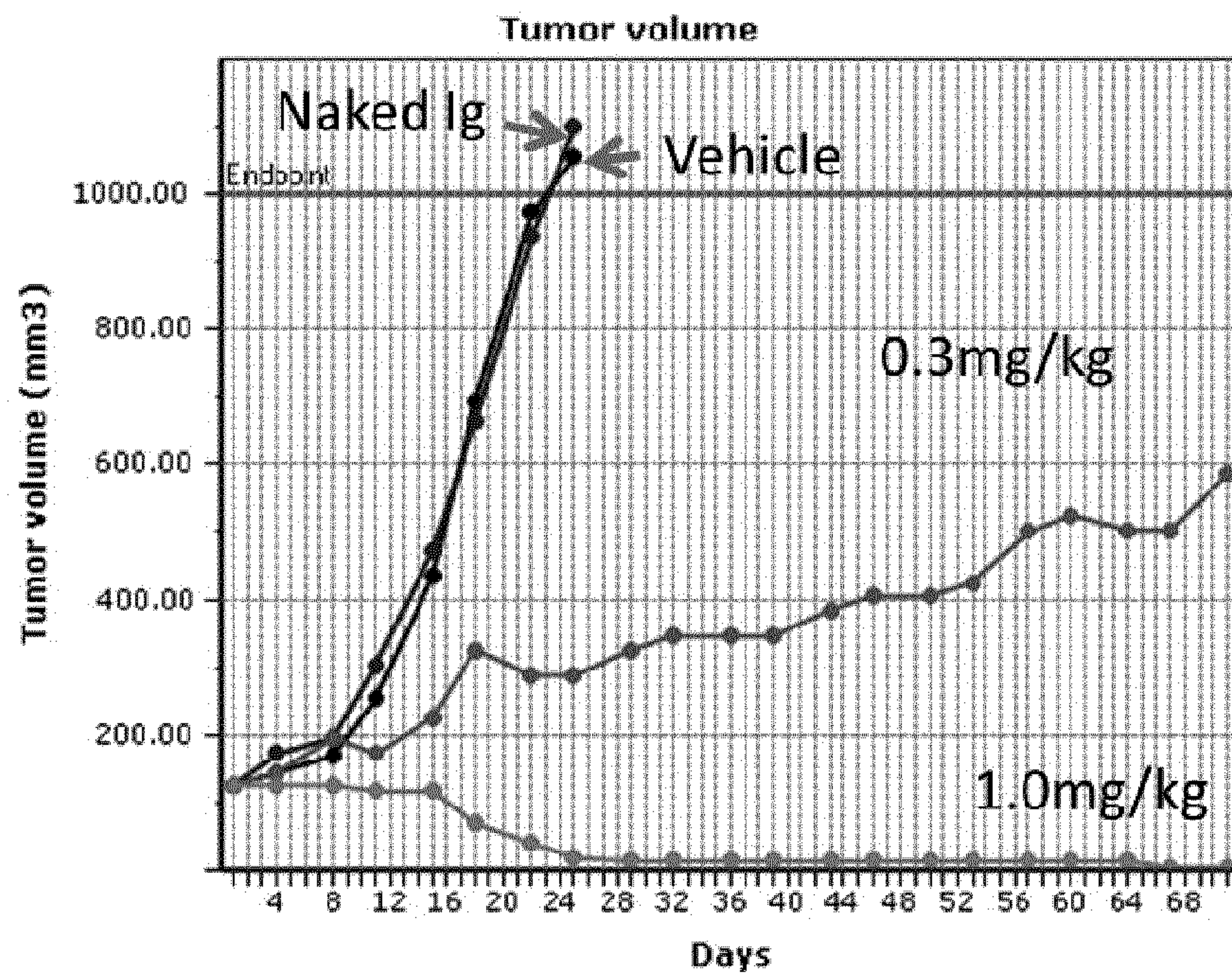


Figure 2

