



US008202706B2

(12) **United States Patent**
Bathe et al.

(10) **Patent No.:** **US 8,202,706 B2**
(45) **Date of Patent:** **Jun. 19, 2012**

(54) **METHOD OF PRODUCTION OF L-AMINO ACIDS**

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(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 0 days.

(21) Appl. No.: **12/750,968**

(22) Filed: **Mar. 31, 2010**

(65) **Prior Publication Data**

US 2010/0261257 A1 Oct. 14, 2010

Related U.S. Application Data

(62) Division of application No. 11/777,423, filed on Jul. 13, 2007, now Pat. No. 7,785,840.

(60) Provisional application No. 60/830,331, filed on Jul. 13, 2006.

(30) **Foreign Application Priority Data**

Jul. 13, 2006 (DE) 10 2006 032 634

(51) **Int. Cl.**

C12P 19/34 (2006.01)

C12N 15/09 (2006.01)

C12N 1/20 (2006.01)

C12N 15/00 (2006.01)

C12N 15/77 (2006.01)

(52) **U.S. Cl.** **435/91.1**; 435/320.1; 435/252.3; 435/252.32; 435/252.33

(58) **Field of Classification Search** 435/91.1, 435/320.1, 252.3, 252.32, 252.33

See application file for complete search history.

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(57) **ABSTRACT**

An isolated polynucleotide encodes a polypeptide comprising the amino acid sequence of SEQ ID NO: 2, with the L-aspartic acid at position 5 of the amino acid sequence replaced by another proteinogenic amino acid, and possesses citrate synthase activity. In addition, a vector comprises the polynucleotide and a bacterium comprises the vector. An isolated polynucleotide comprises a nucleotide sequence comprising, from position 1 to 39, the nucleotide sequence corresponding to position 1 to 39 of SEQ ID NO: 11, from position 40 to 105, a nucleotide sequence encoding the amino acid sequence of SEQ ID NO: 12, with each proteinogenic amino acid except L-aspartic acid being present at position 5. A method of producing an L-amino acids is also described.

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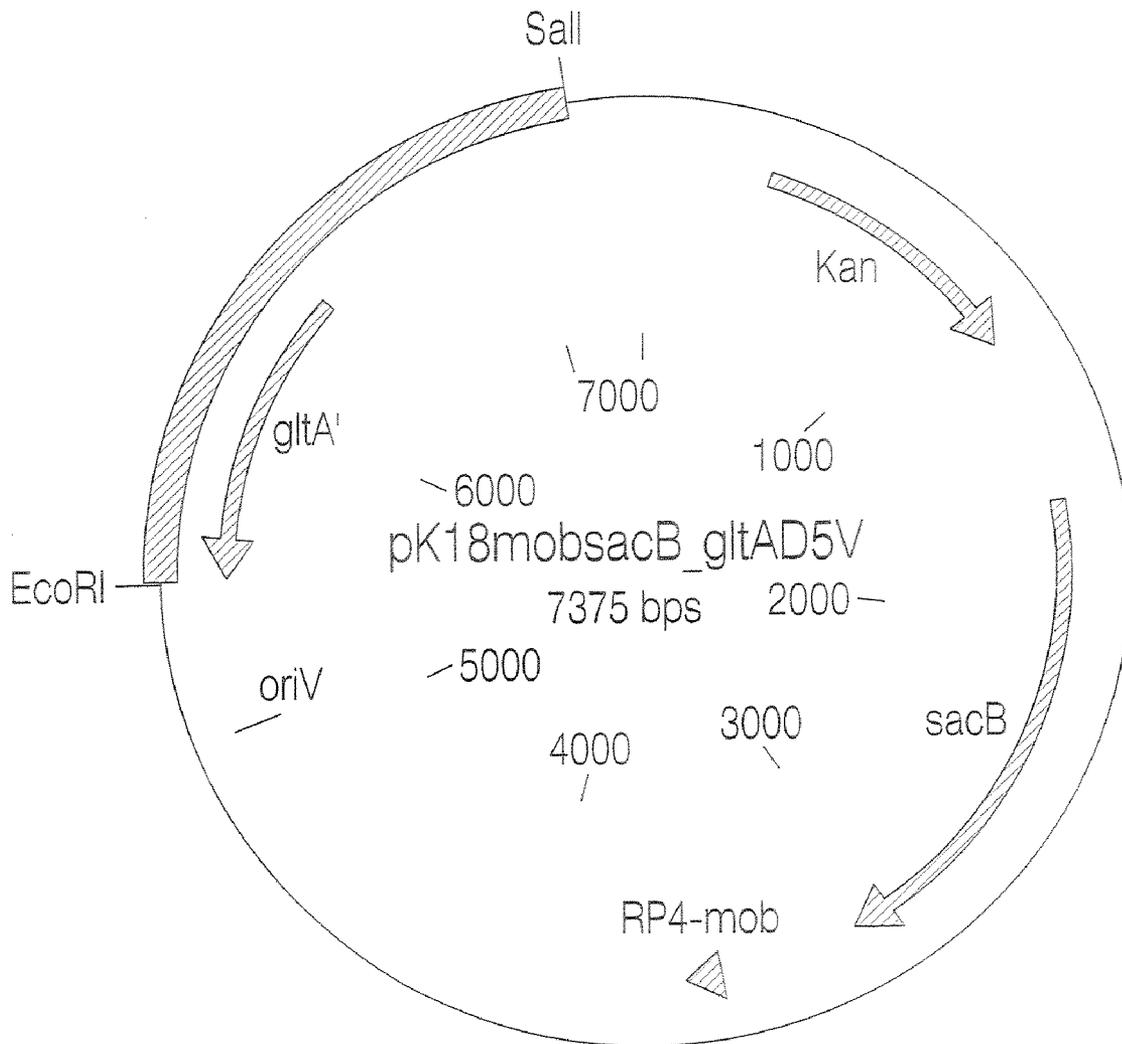
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METHOD OF PRODUCTION OF L-AMINO ACIDS

CROSS REFERENCE TO RELATED APPLICATIONS

This application is a divisional application of U.S. patent application Ser. No. 11/777,423, filed Jul. 13, 2007, now U.S. Pat. No. 7,785,840, issued Aug. 31, 2010, which claims priority to U.S. provisional application 60/830,331, filed Jul. 13, 2006.

BACKGROUND OF THE INVENTION

1. Field of the Invention

The invention relates to novel polynucleotides coding for a polypeptide with citrate synthase activity, bacteria containing the polynucleotides and polypeptides and methods of production of amino acids using these bacteria.

2. Discussion of the Background

All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. Further, the materials, methods, and examples are illustrative only and are not intended to be limiting, unless otherwise specified.

Amino acids are used in human medicine, in the pharmaceutical industry, in the food industry and quite particularly in animal nutrition.

Amino acids may be produced by fermentation of strains of coryneform bacteria, preferably *Corynebacterium glutamicum*. Owing to their great importance, work is constantly in progress for improving the production processes. Process improvements may relate to the fermentation technology, for example, stirring and supply of oxygen, or to the composition of the nutrient media, for example, the sugar concentration during fermentation, or processing to the product form by, for example, ion-exchange chromatography, or to the intrinsic performance characteristics of the microorganism itself.

Methods of mutagenesis, selection and mutant screening are employed for improving the performance characteristics of these microorganisms. In this way we obtain strains that are resistant to antimetabolites or are auxotrophic for metabolites of regulatory importance, and which produce amino acids. A known antimetabolite is the lysine analog S-(2-aminoethyl)-L-cysteine (AEC).

Methods of recombinant DNA technology have also been used for some years now for strain improvement of L-amino acid-producing strains of the genus *Corynebacterium*, preferably *Corynebacterium glutamicum*, by amplifying individual amino acid biosynthesis genes and investigating the effect on amino acid production.

Synoptic descriptions of the biology, genetics and biotechnology of *Corynebacterium glutamicum* are given in "Handbook of *Corynebacterium glutamicum*" (Eds.: L. Eggeling and M. Bott, CRC Press, Taylor & Francis, 2005), in the special issue of the Journal of Biotechnology (Chief Editor: A. Pühler) with the title "A new era in *Corynebacterium glutamicum* biotechnology" (Journal of Biotechnology 104/1-3, (2003)) and in the book by T. Scheper (Managing Editor) "Microbial Production of L-Amino Acids" (Advances in Biochemical Engineering/Biotechnology 79, Springer Verlag, Berlin, Germany, 2003).

The nucleotide sequence of the genome of *Corynebacterium glutamicum* is described in Ikeda and Nakagawa (Ap-

plied Microbiology and Biotechnology 62, 99-109 (2003)), in EP 1 108 790 and in Kalinowski et al. (Journal of Biotechnology 104/1-3, 2003)).

The nucleotide sequence of the genome of *Corynebacterium efficiens* is described in Nishio et al. (Genome Research, 13 (7), 1572-1579 (2003)).

The nucleotide sequences of the genome of *Corynebacterium glutamicum* and *Corynebacterium efficiens* are also available in the database of the National Center for Biotechnology Information (NCBI) of the National Library of Medicine (Bethesda, Md., USA), in the DNA Data Bank of Japan (DDBJ, Mishima, Japan) or in the nucleotide sequence database of the European Molecular Biology Laboratories (EMBL, Heidelberg, Germany and Cambridge, UK).

The wild-type sequence of the coding region of the *gltA* gene of *Corynebacterium glutamicum* is presented in SEQ ID NO: 1 in the specification of the present application. In addition, the sequences located upstream and downstream of the coding region are shown in SEQ ID NO: 3 and 25. The amino acid sequence of the encoded GltA polypeptide (citrate synthase) is accordingly given in SEQ ID NOs: 2, 4 and 26.

SUMMARY OF THE INVENTION

It was an object of the present invention to provide novel measures for the improved production of L-amino acids, preferably L-lysine, L-valine and L-isoleucine, and more preferably L-lysine.

This and other objects have been achieved by the present invention the first embodiment of which includes an isolated polynucleotide, encoding a polypeptide comprising the amino acid sequence of SEQ ID NO: 2, wherein the L-aspartic acid at position 5 of the amino acid sequence is replaced by another proteinogenic amino acid and wherein the polypeptide possesses citrate synthase activity.

The invention further provides a vector comprising the isolated polynucleotide and a bacterium that has been transformed with the vector.

The invention also provides a method of production of an L-amino acid.

BRIEF DESCRIPTION OF THE DRAWING

The FIGURE represents a map of the plasmid pK18mobsacB_gltAD5V.

DETAILED DESCRIPTION OF THE INVENTION

The invention relates to an isolated polynucleotide that codes for a polypeptide which comprises the amino acid sequence of SEQ ID NO: 2, wherein the L-aspartic acid at position 5 of the amino acid sequence is replaced by another proteinogenic amino acid, preferably L-valine, L-leucine and L-isoleucine, and more preferably L-valine, and wherein the polypeptide possesses citrate synthase activity (EC No. 4.1.3.7). Optionally, the polypeptide comprises at least one conservative amino acid substitution, with the citrate synthase activity of the polypeptide being essentially unchanged by the conservative amino acid substitutions.

Proteinogenic amino acids are understood as meaning the amino acids that occur in natural proteins, i.e. in proteins of microorganisms, plants, animals and humans. These include in particular L-amino acids, selected from the group L-aspartic acid, L-asparagine, L-threonine, L-serine, L-glutamic acid, L-glutamine, glycine, L-alanine, L-cysteine, L-valine,

L-methionine, L-isoleucine, L-leucine, L-tyrosine, L-phenylalanine, L-histidine, L-lysine, L-tryptophan, L-proline and L-arginine.

The terms polypeptide and protein are used as synonyms.

The invention further relates to vectors and bacteria, preferably of the genus *Corynebacterium* and *Escherichia*, and more preferably of the species *Corynebacterium glutamicum* and *Escherichia coli*, which contain the stated polynucleotide or were produced using the stated polynucleotide.

The invention also relates finally to bacteria preferably of the genus *Corynebacterium* and *Escherichia*, and more preferably of the species *Corynebacterium glutamicum* and *Escherichia coli*, which have been transformed with the stated vector.

The term transformation comprises all methods for transferring polynucleotides, preferably DNA, into a desired bacterium. Among other things these include the use of isolated DNA in transformation, electrotransformation or electroporation, transfer by cellular contact as in conjugation or the transfer of DNA by particle bombardment.

A further aspect of the invention relates to a bacterium, that may be a recombinant bacterium, of the genus *Corynebacterium*, which comprises a polynucleotide that codes for a polypeptide with citrate synthase activity, which comprises the amino acid sequence of SEQ ID NO: 2, wherein each proteinogenic amino acid except L-aspartic acid, preferably L-valine, L-leucine and L-isoleucine, and preferably L-valine, is contained at position 5 of the amino acid sequence. Optionally, the polypeptide may contain one or more conservative amino acid substitution(s), with the citrate synthase activity of the polypeptide being essentially unchanged by the conservative amino acid substitutions.

A further aspect of the invention relates to a method of production of L-amino acids, preferably L-lysine, L-valine and L-isoleucine, and more preferably L-lysine, comprising the following steps:

a) fermentation of the recombinant bacteria of the genus *Corynebacterium* according to the invention in a suitable nutrient medium, and

b) accumulation of the L-amino acid in the nutrient medium or in the cells of the bacteria.

"L-amino acids" means the proteinogenic amino acids.

If L-lysine or lysine is mentioned hereinafter, this is intended to mean not only the bases, but also the salts, for example L-lysine monohydrochloride or L-lysine sulfate.

With regard to the bacteria of the genus *Corynebacterium*, L-amino acid-excreting strains are preferred, based on the following species:

Corynebacterium efficiens, for example the strain DSM44549,

Corynebacterium glutamicum, for example the strain ATCC13032,

Corynebacterium thermoaminogenes, for example the strain FERM BP-1539, and

Corynebacterium ammoniagenes, for example the strain ATCC6871, the species *Corynebacterium glutamicum* being preferred.

Some representatives of the species *Corynebacterium glutamicum* are also known under different designations. Examples include:

Corynebacterium acetoacidophilum ATCC13870,

Corynebacterium lilium DSM20137,

Corynebacterium melassecola ATCC17965,

Brevibacterium flavum ATCC14067,

Brevibacterium lactofermentum ATCC13869, and

Brevibacterium divaricatum ATCC14020.

Known representatives of amino acid-excreting strains of the genus *Corynebacterium* are, for example, the L-lysine producing strains:

Corynebacterium glutamicum DM58-1/pDM6 (=DSM4697) described in EP 0 358 940,

Corynebacterium glutamicum MH20-22B (=DSM16835) described in Menkel et al. (Applied and Environmental Microbiology 55(3), 684-688 (1989)),

Corynebacterium glutamicum AHP-3 (=FERM BP-7382) described in EP 1 108 790,

Corynebacterium glutamicum DSM16834 described in (PCT/EP2005/012417),

Corynebacterium glutamicum DSM17119 described in (PCT/EP2006/060851),

Corynebacterium glutamicum DSM17223 described in (PCT/EP2006/062010),

Corynebacterium glutamicum DSM16937 described in (PCT/EP2005/057216), and

Corynebacterium thermoaminogenes AJ12521 (=FERM BP-3304) described in U.S. Pat. No. 5,250,423;

or the L-valine producing strains:

Brevibacterium lactofermentum FERM BP-1763 described in U.S. Pat. No. 5,188,948,

Brevibacterium lactofermentum FERM BP-3007 described in U.S. Pat. No. 5,521,074,

Corynebacterium glutamicum FERM BP-3006

described in U.S. Pat. No. 5,521,074, and

Corynebacterium glutamicum FERM BP-1764

described in U.S. Pat. No. 5,188,948;

or the L-isoleucine producing strains:

Brevibacterium flavum FERM-BP 759

described in U.S. Pat. No. 4,656,135,

Corynebacterium glutamicum FERM-BP 757

described in U.S. Pat. No. 4,656,135,

Brevibacterium flavum FERM-BP 760

described in U.S. Pat. No. 4,656,135,

Corynebacterium glutamicum FERM-BP 758

described in U.S. Pat. No. 4,656,135,

Brevibacterium flavum FERM BP-2215

described in U.S. Pat. No. 5,705,370, and

Brevibacterium flavum FERM BP-2433

described in U.S. Pat. No. 5,705,370.

Information on the taxonomic classification of strains of this group of bacteria can be found inter alia in Seiler (Journal of General Microbiology 129, 1433-1477 (1983)), Kampf and Kroppenstedt (Canadian Journal of Microbiology 42, 989-1005 (1996)), Liebl et al. (International Journal of Systematic Bacteriology 41, 255-260 (1991)) and in U.S. Pat. No. 5,250,434.

Strains with the designation "ATCC" can be obtained from the American Type Culture Collection (Manassas, Va., USA). Strains with the designation "DSM" can be obtained from the Deutsche Sammlung von Mikroorganismen und Zellkulturen (DSMZ, Braunschweig, Germany). Strains with the designation "FERM" can be obtained from the National Institute of Advanced Industrial Science and Technology (AIST Tsukuba Central 6, 1-1-1 Higashi, Tsukuba Ibaraki, Japan). The strain *Corynebacterium thermoaminogenes* (FERM BP-1539) is described in U.S. Pat. No. 5,250,434.

For production of the polynucleotides, it is possible to use classical in-vivo mutagenesis techniques with cell populations of bacteria of the genus *Corynebacterium* using mutagenic substances such as N-methyl-N'-nitro-N-nitrosoguanidine (MNNG) or using ultraviolet light. Mutagenesis techniques are described for example in the Manual of Methods for General Bacteriology (Gerhard et al. (Eds.), American Society for Microbiology, Washington, D.C.,

USA, 1981) or in Tosaka et al. (Agricultural and Biological Chemistry 42(4), 745-752 (1978)) or in Konicek et al. (Folia Microbiologica 33, 337-343 (1988)).

From the mutagenized cell population, those mutants are taken and multiplied which require L-glutamic acid or citric acid in order to be able to grow on a minimal agar or whose growth on the minimal agar is improved by adding L-glutamic acid or citric acid. It is also possible, starting from mutants requiring L-glutamic acid or citric acid, to isolate so-called revertants, which do not require L-glutamic acid or citric acid for their growth. These L-glutamic acid—auxotrophic or citric acid—auxotrophic mutants or their respective revertants are then investigated. Technical details on the isolation of mutants with defective citrate synthase activity can be found for example in Shiio et al. (Agricultural and Biological Chemistry 46(1), 101-107 (1982)).

Next, DNA is prepared or isolated from the mutants and by means of, for example, the polymerase chain reaction (PCR) using primer pairs which allow the amplification of the *gltA* gene or *gltA* allele, the corresponding polynucleotide is synthesized and isolated.

For this, it is possible to select any primer pairs from the nucleotide sequence located upstream and downstream of the coding region and the nucleotide sequence that is complementary to it (see SEQ ID NOs: 3 and 25). A primer of a primer pair then comprises preferably at least 15, at least 18, at least 20, at least 21 or at least 24 consecutive nucleotides selected from the nucleotide sequence between position 1 and 1000 of SEQ ID NO: 25. The associated second primer of a primer pair comprises at least 15, at least 18, at least 20, at least 21 or at least 24 consecutive nucleotides selected from the complementary nucleotide sequence between position 3314 and 2312 of SEQ ID NO: 25.

A person skilled in the art will find instructions and information on PCR for example in the handbook "PCR Strategies" (Innis, Felfand and Sninsky, Academic Press, Inc., 1995), in the handbook by Diefenbach and Dveksler "PCR Primer—a laboratory manual" (Cold Spring Harbor Laboratory Press, 1995), in Gait's handbook "Oligonucleotide Synthesis: a Practical Approach" (IRL Press, Oxford, UK, 1984) and in Newton and Graham "PCR" (Spektrum Akademischer Verlag, Heidelberg, Germany, 1994).

Further instructions on PCR can be found for example in WO 06/100177 on pages 15 to 17.

In a further step, the nucleotide sequence of the polynucleotide is then determined. This can for example be determined by the chain-terminating technique of Sanger et al. (Proceedings of the National Academies of Sciences, USA, 74, 5463-5467 (1977)) with the modifications stated by Zimmermann et al. (Nucleic Acids Research 18, 1067 pp (1990)).

The polypeptide encoded by this nucleotide sequence can then be analyzed with respect to the amino acid sequence. For this, the nucleotide sequence is input in a program for translating a DNA sequence into an amino acid sequence. Suitable programs are for example the "Patentin" program, which is obtainable from patent offices, for example the US Patent Office (USPTO), or the "Translate Tool", which is available on the ExPASy Proteomics Server on the World Wide Web (Gasteiger et al., Nucleic Acids Research 31, 3784-3788 (2003)).

It is also possible for the polynucleotide, which is also designated hereinafter as *gltA* allele, to be produced by methods of in-vitro genetics.

Suitable methods for in-vitro mutagenesis including among others treatment with hydroxylamine according to Miller (Miller, J. H.: A Short Course in Bacterial Genetics. A Laboratory Manual and Handbook for *Escherichia coli* and

Related Bacteria, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, 1992) or the use of a polymerase chain reaction using a DNA polymerase, which has a high error rate. Such a DNA polymerase is for example the Mutazyme DNA Polymerase (GeneMorph PCR Mutagenesis Kit, No. 600550) of the company Stratagene (La Jolla, Calif., USA). It is also possible to use mutagenic oligonucleotides, as described by T. A. Brown (Gentechnologie für Einsteiger, Spektrum Akademischer Verlag, Heidelberg, 1993) and R. M. Horton (PCR-mediated recombination and mutagenesis, Molecular Biotechnology 3, 93-99 (1995)). The method using the "Quik Change Site-directed Mutagenesis Kit" of the company Stratagene (La Jolla, Calif., USA) described by Papworth et al. (Strategies 9(3), 3-4 (1996)) can also be used.

Methods for the determination of citrate synthase activity can be found in Eikmanns et al. (Microbiology 140, 1817-1828 (1994)) and in Shiio et al. (Agricultural and Biological Chemistry 46(1), 101-107 (1982)).

It is moreover possible to overexpress the citrate synthase allele according to the invention in *Corynebacterium glutamicum* or *Escherichia coli*, and it can then be prepared in purified or isolated form.

A polynucleotide with the nucleotide sequence shown in SEQ ID NO: 5 was isolated in this way. The polypeptide encoded by this polynucleotide is shown in SEQ ID NO: 6 and 8. It contains L-valine instead of L-aspartic acid at position 5 of the amino acid sequence.

It was found that when the strain ATCC13032 is provided, instead of the wild-type *gltA* gene, with the *gltA* allele according to the invention, which codes for the citrate synthase according to SEQ ID NO: 6 (ATCC13032::*gltA* D5V), in comparison with the wild-type strain ATCC13032, which contains the citrate synthase according to SEQ ID NO: 2, with enzyme activity reduced by approx. 40% up to a maximum of approx. 90%, preferably with enzyme activity reduced by approx. 70% up to a maximum of approx. 90%.

It is known that conservative amino acid substitutions only change the enzyme activity insignificantly. Accordingly, the invention also relates to polynucleotides that code for polypeptides with citrate synthase activity, which in addition to the amino acid substitutions at position 5 of the amino acid sequence contain one (1) or more conservative amino acid substitution(s), which does not alter the enzyme activity substantially. That is, it remains essentially unchanged. The term "not altered substantially," "essentially unchanged," or "substantially unchanged" means in this context that the citrate synthase activity of the polypeptide is altered by at most 20%, preferably at most 10% and more preferably at most 5% to at most 2% in comparison with the citrate synthase activity of the polypeptide according to SEQ ID NO: 10 or SEQ ID NO: 6, preferably SEQ ID NO: 6.

For an experimental test, the *gltA* gene of strain ATCC13032 is substituted for the *gltA* allele, which codes for a polypeptide containing the amino acid substitution at position 5 and at least one conservative amino acid substitution. Then the strain is cultivated, a cellular extract is produced and the citrate synthase activity is determined. As a reference, the citrate synthase activity in strain ATCC13032::*gltA* D5V is determined.

Instead of strain ATCC13032, it is also possible to use L-lysine-excreting strains of *Corynebacterium glutamicum* which comprises the coding region of the *gltA* gene of the wild type including the nucleotide sequences located upstream, corresponding to the nucleotide sequence between position 1 and 2064 of SEQ ID NO: 3, preferably SEQ ID NO: 3. Suitable strains are, for example, DSM16833 described in PCT/EP2005/012417, DSM13994 described in EP 1 239 040

A2 or DSM17576 described in DE 102005045301. In these strains, the appropriate mutation(s) can be inserted in the coding region of the *gltA* gene by, for example, allelic substitution.

It is also possible to purify the polypeptides and to conduct the comparative tests on the purified polypeptides.

The enzyme citrate synthase (EC No. 4.1.3.7) catalyzes the condensation reaction of oxaloacetate and acetyl-CoA, with formation of citric acid and coenzyme A (CoA) as reaction products. The enzyme is assigned the number EC 2.3.3.1 in the Kyoto Encyclopedia of Genes and Genomes (KEGG, Kanehisa Laboratory, Bioinformatics Center, Institute for Chemical Research, Kyoto University, Japan).

In the case of aromatic L-amino acids, we talk of conservative substitutions when L-phenylalanine, L-tryptophan and L-tyrosine are substituted for one another. In the case of hydrophobic L-amino acids, we talk of conservative substitutions when L-leucine, L-isoleucine and L-valine are substituted for one another. In the case of polar L-amino acids, we talk of conservative substitutions when L-glutamine and L-asparagine are substituted for one another. In the case of basic L-amino acids, we talk of conservative substitutions when L-arginine, L-lysine and L-histidine are substituted for one another. In the case of acidic L-amino acids, we talk of conservative substitutions when L-aspartic acid and L-glutamic acid are substituted for one another. In the case of L-amino acids containing hydroxyl groups, we talk of conservative substitutions when L-serine and L-threonine are substituted for one another.

Preferably the polypeptide contains at most two (2), at most three (3), at most four (4) or at most five (5) conservative amino acid substitutions in addition to the substitution at position 5 of SEQ ID NO: 2.

It is known that the terminal methionine may be removed during protein synthesis by enzymes that are intrinsic to the host, so-called aminopeptidases.

The isolated polynucleotides, which code for the citrate synthase variant, or portions thereof, can be used for producing recombinant strains of the genus *Corynebacterium*, preferably *Corynebacterium glutamicum*, which comprises the amino acid substitution at position 5 of the amino acid sequence of the citrate synthase polypeptide and which provide improved release of L-amino acids into the surrounding medium or accumulation of them inside the cell, compared with the starting or parent strain.

The initial strains preferably used are those which already possess the capacity to excrete at least 1 g/l, preferably at least 5 g/l, and more preferably at least 10 g/l of the desired L-amino acid into the surrounding nutrient medium.

A widely used method for incorporating mutations in genes of bacteria of the genus *Corynebacterium*, preferably of the species *Corynebacterium glutamicum*, is allelic substitution, which is also known as "gene replacement". In this technique, a DNA fragment that contains the mutation of interest is transferred into the desired strain and the mutation is incorporated in the chromosome of the desired strain by at least two recombination events or cross-over events or a gene sequence present in the strain in question is replaced by the mutated sequence.

In this method, the DNA fragment containing the mutation of interest may be located in a vector, preferably a plasmid, which preferably is not replicated by the strain that is to be provided with the mutation, or such replication is limited, i.e. occurs under selected culture conditions. A bacterium of the genus *Escherichia*, preferably of the species *Escherichia coli*, may be used as auxiliary or intermediate host, in which the vector can be replicated.

Examples of such plasmid vectors are the pK**mob* and pK**mobsacB* vectors, for example pK18*mobsacB*, described by Schäfer et al. (Gene 145, 69-73 (1994)), and the vectors described in WO 02/070685 and WO 03/014362. These vectors can replicate in *Escherichia coli* but not in *Corynebacterium*. Preferably, suitable vectors are those which contain a gene with conditionally negative dominant action for example the *sacB* gene (levansucrase gene) or for example *Bacillus* or the *galK* gene (galactose kinase gene) of for example *Escherichia coli*. "Gene with conditionally negative dominant action" means a gene which under certain conditions is disadvantageous, for example toxic to the host, but in other conditions does not have adverse effects on the host carrying the gene. These make it possible to select for recombination events in which the vector is eliminated from the chromosome.

Furthermore, Nakamura et al. (U.S. Pat. No. 6,303,383) described a temperature-sensitive plasmid for *Corynebacterium*, which can only replicate at temperatures below 31° C. It can also be used for the purposes of the invention.

The vector is then transferred into the *Corynebacterium* by conjugation, for example, by Schäfer's method (Journal of Bacteriology 172, 1663-1666 (1990)) or transformation, for example, by Dunican and Shivnan's method (Bio/Technology 7, 1067-1070 (1989)) or the method of Thierbach et al. (Applied Microbiology and Biotechnology 29, 356-362 (1988)). Optionally, the transfer of the DNA can also be achieved by ballistic methods (e.g. particle bombardment).

After homologous recombination by means of a first cross-over event producing integration and a suitable second cross-over event causing an excision in the target gene or in the target sequence, incorporation of the mutation is achieved and a recombinant bacterium is obtained. "Target gene" means the gene in which the desired substitution is to take place.

The strains obtained can be identified and characterized using, among others, the methods of Southern blotting hybridization, polymerase chain reaction and sequencing, the method of fluorescence resonance energy transfer (FRET) (Lay et al. Clinical Chemistry 43, 2262-2267 (1997)) or methods of enzymology.

This method was used by Schwarzer and Pühler (Bio/Technology 9, 84-87 (1991)) for incorporating a *lysA* allele carrying a deletion, and a *lysA* allele carrying an insertion, into the chromosome of *C. glutamicum* instead of the wild-type gene.

This method was used by Nakagawa et al. (EP 1108790) and Ohnishi et al. (Applied Microbiology and Biotechnology 58(2), 217-223 (2002)) for incorporating various mutations into the chromosome of *C. glutamicum* starting from the isolated alleles or polynucleotides. In this way, Nakagawa et al. succeeded in incorporating a mutation designated Val59Ala into the homoserine dehydrogenase gene (*hom*), a mutation designated Thr311Ile into the aspartate kinase gene (*lysC* or *ask*), a mutation designated Pro458Ser into the pyruvate carboxylase gene (*pyc*) and a mutation designated Ala213Thr into the glucose-6-phosphate-dehydrogenase gene (*zwf*) of *C. glutamicum* strains.

For inserting the mutation in the *gltA* gene into the chromosome by means of allelic substitution, it is possible to use a polynucleotide that codes for an amino acid sequence which has the amino acid substitution at position 5 of SEQ ID NO: 2, as shown in SEQ ID NO: 10, and possesses, upstream and downstream thereof, a nucleotide sequence with a length in each case of at least approx. 51 (cf. SEQ ID NO: 11 and 12) preferably in each case at least approx. 101 or 102 (cf. SEQ ID NO: 13 and 14), preferably in each case at least approx. 201 nucleobases (cf. SEQ ID NO: 15 and 16) and more preferably

in each case at least approx. 500 or 498 nucleobases (cf. SEQ ID NO: 17 and 18) selected from SEQ ID NO: 9. The maximum length of the nucleotide sequence located upstream and downstream of the mutation is generally approx. 500, approx. 750, approx. 1000, approx. 1500, approx. 2000 to 2100 nucleobases. The nucleotide sequence located upstream of the mutation comprises, for example, the sequence between position 1 to 762 of SEQ ID NO: 9 or the sequence between position 1 to 1012 of SEQ ID NO: 25. The nucleotide sequence located downstream of the mutation comprises for example the sequence between position 766 to 2814 of SEQ ID NO: 9 or the sequence between position 1016 to 3314 of SEQ ID NO: 25. The total length of the polynucleotide used for the allelic substitution is accordingly at most approx. 1500, at most approx. 2000, at most approx. 3000 or at most approx. 4000 to 4200 nucleobases.

Accordingly, the invention relates to a polynucleotide that comprises a nucleotide sequence which contains, from position 1 to 39, the nucleotide sequence corresponding to position 1 to 39 of SEQ ID NO: 11 and, from position 40 to 105, a nucleotide sequence that codes for the amino acid sequence according to SEQ ID NO: 12, having every proteinogenic amino acid except L-aspartic acid being contained at position 5.

In this context, the stated positions 1 to 39 of SEQ ID NO: 11 correspond to the stated positions 712 to 750 of SEQ ID NO: 9. The stated positions 40 to 105 of SEQ ID NO: 11 correspond to the stated positions 751 to 816 of SEQ ID NO: 9.

In one embodiment, the polynucleotide comprises the nucleotide sequence of SEQ ID NO: 11, with the codon corresponding to position 52 to 54 coding for every proteinogenic amino acid except L-aspartic acid.

In another embodiment the polynucleotide comprises the nucleotide sequence from position 712 to 816 of SEQ ID NO: 7.

Accordingly, the invention also relates to a polynucleotide that comprises a nucleotide sequence which comprises, from position 1 to 89, the nucleotide sequence corresponding to position 1 to 89 of SEQ ID NO: 13 and, from position 90 to 206, a nucleotide sequence which codes for the amino acid sequence according to SEQ ID NO: 14, having every proteinogenic amino acid except L-aspartic acid being contained at position 5.

In this context, the stated positions 1 to 89 of SEQ ID NO: 13 correspond to the stated positions 662 to 750 of SEQ ID NO: 9. The stated positions 90 to 206 of SEQ ID NO: 13 correspond to the stated positions 751 to 867 of SEQ ID NO: 9.

In one embodiment, the polynucleotide comprises the nucleotide sequence of SEQ ID NO: 13, with the codon corresponding to position 102 to 104 coding for every proteinogenic amino acid except L-aspartic acid.

In another embodiment, the polynucleotide comprises the nucleotide sequence from position 662 to 867 of SEQ ID NO: 7.

Accordingly, the invention also relates to a polynucleotide that comprises a nucleotide sequence which comprises, from position 1 to 189, the nucleotide sequence corresponding to position 1 to 189 of SEQ ID NO: 15 and, from position 190 to 405, a nucleotide sequence which codes for the amino acid sequence according to SEQ ID NO: 16, having every proteinogenic amino acid except L-aspartic acid being contained at position 5.

In this context, the stated positions 1 to 189 of SEQ ID NO: 15 correspond to the stated positions 562 to 750 of SEQ ID

NO: 9. The stated positions 190 to 405 of SEQ ID NO: 15 correspond to the stated positions 751 to 966 of SEQ ID NO: 9.

In one embodiment, the polynucleotide comprises the nucleotide sequence of SEQ ID NO: 15, with the codon corresponding to position 202 to 204 coding for every proteinogenic amino acid except L-aspartic acid.

In another embodiment, the polynucleotide comprises the nucleotide sequence from position 562 to 966 of SEQ ID NO: 7.

Accordingly, the invention also relates to a polynucleotide that comprises a nucleotide sequence which comprises, from position 1 to 488, the nucleotide sequence corresponding to position 1 to 488 of SEQ ID NO: 17 and, from position 489 to 1001, a nucleotide sequence which codes for the amino acid sequence according to SEQ ID NO: 18, with every proteinogenic amino acid except L-aspartic acid being contained at position 5.

In this context, the stated positions 1 to 488 of SEQ ID NO: 17 correspond to the stated positions 263 to 750 of SEQ ID NO: 9. The stated positions 489 to 1001 of SEQ ID NO: 15 correspond to the stated positions 751 to 1263 of SEQ ID NO: 9.

In one embodiment, the polynucleotide comprises the nucleotide sequence of SEQ ID NO: 17, with the codon corresponding to position 501 to 503 coding for every proteinogenic amino acid except L-aspartic acid.

In another embodiment, the polynucleotide comprises the nucleotide sequence from position 263 to 1263 of SEQ ID NO: 7.

In another embodiment, the polynucleotide comprises the nucleotide sequence from position 9 to 1687 of SEQ ID NO: 31.

The invention also relates to vectors, preferably plasmids, comprising the stated polynucleotides.

The invention also relates to bacteria preferably of the genus *Escherichia*, more preferably of the species *Escherichia coli*, and *Corynebacterium*, more preferably of the species *Corynebacterium glutamicum*, comprising the stated vectors.

The invention also relates to strains of the genus *Corynebacterium*, preferably of the species *Corynebacterium glutamicum*, which have been produced using the polynucleotides or vectors comprising the polynucleotides.

It is also possible to insert the *gltA* allele at another site in the chromosome of *Corynebacterium glutamicum*. Possible examples are the sites or genes *aecD*, *ccpA1*, *ccpA2*, *citA*, *citB*, *citE*, *fda*, *gluA*, *gluB*, *gluC*, *gluD*, *luxR*, *luxS*, *lysR1*, *lysR2*, *lysR3*, *menE*, *mqo*, *pck*, *pgi* and *poxB*, as described in WO 03/04037. Other possibilities are, for example, intergenic regions, DNA of prophages, defective phages and phage components, as described in WO 04/069996.

The obtained recombinant strains display, relative to the initial strain or parent strain used, increased excretion or production of the desired amino acid in a fermentation process.

The L-lysine-excreting starting strains that can be used for the purposes of the invention possess, in addition to other properties, in particular a lysine-insensitive aspartate kinase.

“Lysine-insensitive aspartate kinase” means a polypeptide or protein with aspartate kinase activity (EC No. 2.7.2.4), which in comparison with the wild form, have lower sensitivity to inhibition by mixtures of lysine and threonine or mixtures of AEC (aminoethylcysteine) and threonine or lysine alone or AEC alone. Aspartate kinases of this kind are also called feedback-resistant or desensitized aspartate kinases. The nucleotide sequences coding for these desensi-

tized aspartate kinases or aspartate kinase variants are also designated as *lysC^{FBR}* alleles. Information on numerous *lysC^{FBR}* alleles is available in public databases. The *lysC*-gene is also designated as the ask-gene by some authors.

The coding region of the wild-type *lysC* gene of *Corynebacterium glutamicum* corresponding to access number AX756575 of the NCBI database is shown in SEQ ID NO: 19 and the polypeptide encoded by this gene is shown in SEQ ID NO: 20. The nucleotide sequences located upstream of the 5' end and downstream of the 3' end of the coding region are also shown in SEQ ID NO: 21. SEQ ID NO: 20 corresponds to SEQ ID NO: 22.

The L-lysine-excreting bacteria preferably have a *lysC* allele, which codes for an aspartate kinase variant possessing the amino acid sequence of SEQ ID NO: 20, comprising one or more of the amino acid substitutions selected from the group:

a) *LysC* A279T (substitution of L-alanine at position 279 of the encoded aspartate kinase protein according to SEQ ID NO: 20 for L-threonine; see U.S. Pat. No. 5,688,671 and access numbers E06825, E06826, E08178 and I74588 to I74597),

b) *LysC* A279V (substitution of L-alanine at position 279 of the encoded aspartate kinase protein according to SEQ ID NO: 20 for L-valine, see JP 6-261766 and access number E08179),

c) *LysC* L297Q (substitution of L-leucine at position 297 of the encoded aspartate kinase protein according to SEQ ID NO: 20 for another proteinogenic amino acid, preferably L-glutamine; see DE 102006026328),

d) *LysC* S301F (substitution of L-serine at position 301 of the encoded aspartate kinase protein according to SEQ ID NO: 20 for L-phenylalanine; see U.S. Pat. No. 6,844,176 and access number E08180),

e) *LysC* S301Y (substitution of L-serine at position 301 of the encoded aspartate kinase protein according to SEQ ID NO: 20 for L-tyrosine, see Kalinowski et al. (Molecular and General Genetics 224, 317-324 (1990)) and access number λ 57226),

f) *LysC* T308I (substitution of L-threonine at position 308 of the encoded aspartate kinase protein according to SEQ ID NO: 20 for L-isoleucine; see JP 6-261766 and access number E08181),

g) *LysC* T311I (substitution of L-threonine at position 311 of the encoded aspartate kinase protein according to SEQ ID NO: 20 for L-isoleucine; see WO 00/63388 and U.S. Pat. No. 6,893,848),

h) *LysC* S317A (substitution of L-serine at position 317 of the encoded aspartate kinase protein according to SEQ ID NO: 20 for L-alanine; see U.S. Pat. No. 5,688,671 and access number I74589),

i) *LysC* R320G (substitution of L-arginine at position 320 of the encoded aspartate kinase protein according to SEQ ID NO: 20 for glycine; see Jetten et al. (Applied Microbiology and Biotechnology 43, 76-82 (1995)) and access number L27125),

j) *LysC* G345D (substitution of glycine at position 345 of the encoded aspartate kinase protein according to SEQ ID NO: 20 for L-aspartic acid; see Jetten et al. (Applied Microbiology and Biotechnology 43, 76-82 (1995)) and access number L16848),

k) *LysC* T380I (substitution of L-threonine at position 380 of the encoded aspartate kinase protein according to SEQ ID NO: 20 for L-isoleucine; see WO 01/49854 and access number AX192358), and

l) *LysC* S381F (substitution of L-serine at position 381 of the encoded aspartate kinase protein according to SEQ ID NO: 20 for L-phenylalanine; see EP 0435132).

Strains comprise aspartate kinase variants comprising the amino acid substitution *LysC* T311I or at least one amino acid substitution selected from the group *LysC* A279T, *LysC* L297Q, *LysC* S317A, *LysC* T380I and *LysC* S381F.

Naturally it is also possible for insertion of the mutation in the *gltA* gene of the chromosome of a bacterium of the genus *Corynebacterium* to be carried out first, followed by insertion of one or more of the desired mutation(s) in the *lysC* gene of the strain in question.

In one embodiment, the described aspartate kinases are overexpressed in the *Corynebacterium* which comprises the amino acid substitution according to the invention in the *gltA* gene.

Overexpression means an increase in the intracellular concentration or activity of a ribonucleic acid, a protein or an enzyme compared with the initial strain (parent strain) or wild-type strain. Initial strain (parent strain) means the strain on which the measure leading to overexpression was carried out.

The increase in concentration or activity can be achieved, for example, by increasing the copy number of the corresponding polynucleotides chromosomally or extrachromosomally by at least one copy.

A widely used method of increasing the copy number comprises inserting the corresponding polynucleotide in a vector, preferably a plasmid, which is replicated by a coryneform bacterium. Suitable plasmid vectors are, for example, pZ1 (Merkel et al., Applied and Environmental Microbiology (1989) 64: 549-554) or the pSELF vectors described by Tauch et al. (Journal of Biotechnology 99, 79-91 (2002)). A review article on the subject of plasmids in *Corynebacterium glutamicum* can be found in Tauch et al. (Journal of Biotechnology 104, 27-40 (2003)).

Transposons, insertion elements (IS elements) or phages can also be used as vectors. Such genetic systems are stated for example in patent specifications U.S. Pat. No. 4,822,738, U.S. Pat. No. 5,804,414 and U.S. Pat. No. 5,804,414. Similarly, it is possible to use the IS element ISaB1 described in WO 92/02627 or the transposon Tn45 of plasmid pXZ10142 (cited in "Handbook of *Corynebacterium glutamicum*" (Publisher: L. Eggeling and M. Bott)).

Another widely used method for achieving overexpression is the technique of chromosomal gene amplification. In this method, at least one additional copy of the polynucleotide of interest is inserted in the chromosome of a coryneform bacterium. Such amplification techniques are described for example in WO 03/014330 or WO 03/040373.

Another method of achieving overexpression comprises operably linking the corresponding gene or allele with a promoter or an expression cassette. Suitable promoters for *Corynebacterium glutamicum* are described for example in FIG. 1 of the review article by Patek et al. (Journal of Biotechnology 104(1-3), 311-323 (2003)). The variants of the *dapA* promoter described by Vasicova et al. (Journal of Bacteriology 181, 6188-6191 (1999)), for example the promoter A25, can be used similarly. It is also possible to use the *gap*-promoter of *Corynebacterium glutamicum* (EP 06007373). Finally it is possible to use the sufficiently well-known promoters T3, T7, SP6, M13, lac, tac and trc described by Amann et al. (Gene 69(2), 301-315 (1988)) and Amann and Brosius (Gene 40(2-3), 183-190 (1985)). Such a promoter can for example be inserted upstream of the gene in question, typically at a distance of about 1-500 nucleobases from the start codon.

As a result of the measures for overexpression, the activity or concentration of the corresponding polypeptide is increased by at least 10%, 25%, 50%, 75%, 100%, 150%, 200%, 300%, 400% or 500%, at most up to 1000% or 2000% relative to the activity or concentration of the polypeptide in the strain prior to the measure leading to overexpression.

In a further embodiment, the bacteria of the genus *Corynebacterium*, which preferably in addition comprises a polynucleotide that codes for a lysine-insensitive aspartate kinase variant, possess one or more of the characters selected from the group

a) overexpressed polynucleotide (dapA gene), which codes for a dihydrodipicolinate synthase (DapA, EC No. 4.2.1.52),

b) overexpressed polynucleotide (asd gene), which codes for an aspartate semialdehyde dehydrogenase (Asd, EC No. 1.2.1.11),

c) overexpressed polynucleotide (lysA gene), which codes for a diaminopimelate decarboxylase (LysA, EC No. 4.1.1.20),

d) overexpressed polynucleotide (aat gene), which codes for an aspartate aminotransferase (Aat, EC No. 2.6.1.1),

e) overexpressed polynucleotide (lysE gene), which codes for a polypeptide with L-lysine exporting activity (LysE, Lysin Efflux Permease),

f) switched-off or attenuated activity of malate dehydrogenase (Mdh, EC No. 1.1.1.37),

g) switched-off or attenuated activity of malate-quinone oxidoreductase (Mqo, EC No. 1.1.99.16),

h) overexpressed polynucleotide, which codes for a pyruvate carboxylase (Pyc, EC No. 6.4.1.1), and

i) switched-off or attenuated activity of the E1p subunit of the pyruvate dehydrogenase complex (AceE, EC No. 1.2.4.1).

Characters a) to g) are preferred.

The known genes, for example, the wild-type genes, of *Escherichia coli* (Blattner et al., *Science* 277(5), 1453-1462 (1997)), *Bacillus subtilis* (Kunst et al., *Nature* 390 (6657), 249-256 (1997)), *Bacillus licheniformis* (Veith et al., *Journal of Molecular Microbiology and Biotechnology* 7 (4), 204-211 (2004)), *Mycobacterium tuberculosis* (Fleischmann et al., *Journal of Bacteriology* 184(1), 5479-5490 (2004)), *Mycobacterium bovis* (Garnier et al., *Proceedings of the National Academy of Sciences USA* 100 (13), 7877-7882 (2003)), *Streptomyces coelicolor* (Redenbach et al., *Molecular Microbiology* 21 (1), 77-96 (1996)), *Lactobacillus acidophilus* (Alternann et al., *Proceedings of the National Academy of Sciences USA* 102 (11), 3906-3912 (2005)), *Lactobacillus johnsonii* (Pridmore et al., *Proceedings of National Academy of Sciences USA* 101 (8), 2512-2517 (2004)), *Bifidobacterium longum* (Schell et al., *Proceedings of National Academy of Sciences USA* 99 (22), 14422-14427 (2002)), and *Saccharomyces cerevisiae* can be used for overexpression of the listed genes or polynucleotides. The genomes of the wild-type forms of these bacteria are available in sequenced or annotated form. Preferably the endogenous genes or polynucleotides of the genus *Corynebacterium*, more preferably of the species *Corynebacterium glutamicum*, are used.

"Endogenous genes or polynucleotides" means the open reading frames (ORF), genes or alleles or their polynucleotides present in the population of a species.

The dapA gene of *Corynebacterium glutamicum* strain ATCC13032 is described for example in EP 0 197 335. The MC20 and MA16 mutations of the dapA promoter, as described in U.S. Pat. No. 6,861,246, can also be used, among others, for overexpression of the dapA gene of *Corynebacterium glutamicum*.

The asd gene of *Corynebacterium glutamicum* strain ATCC21529 is described for example in U.S. Pat. No. 6,927,046.

The lysA gene of *Corynebacterium glutamicum* ATCC13869 (*Brevibacterium lactofermentum*) is described for example in U.S. Pat. No. 6,090,597.

The aat gene of *Corynebacterium glutamicum* ATCC13032 is described for example in Kalinowski et al. (*Journal of Biotechnology* 104 (1-3), 5-25 (2003)); see also access number NC_006958). There it is designated aspB gene. In U.S. Pat. No. 6,004,773 a gene coding for an aspartate aminotransferase is designated aspC. Marienhagen et al. (*Journal of Bacteriology* 187 (22), 7639-7646 (2005)) denote the aat gene as aspT gene.

The lysE gene of *Corynebacterium glutamicum* R127 is described for example in U.S. Pat. No. 6,858,406. Strain R127 is a restriction-defective mutant of ATCC13032 (Liebl et al., *FEMS Microbiology Letters* 65, 299-304 (1989)). The lysE gene of strain ATCC13032 used in U.S. Pat. No. 6,861,246 can be used similarly.

The pyc gene of *Corynebacterium glutamicum* of strain ATCC 13032 is described for example in WO 99/18228 and WO 00/39305. Furthermore, alleles of the pyc gene can be used, such as are described in U.S. Pat. No. 6,965,021. The pyruvate carboxylases described in this patent specification possess one or more of the amino acid substitutions selected from the group: Pyc E153D (substitution of L-glutamic acid at position 153 for L-aspartic acid), Pyc A1825 (substitution of L-alanine at position 182 for L-serine), Pyc A2065 (substitution of L-alanine at position 206 for L-serine), Pyc H227R (substitution of L-histidine at position 227 for L-arginine), Pyc A455G (substitution of L-alanine at position 455 for glycine), and Pyc D1120E (substitution of L-aspartic acid at position 1120 for L-glutamic acid). Similarly, it is possible to use the pyc allele described in EP 1 108 790, which codes for a pyruvate carboxylase containing the amino acid substitution Pyc P458S (substitution of L-proline at position 458 for L-serine).

"Switched-off or attenuated activity" means reduction or switching-off of the intracellular activity or concentration of one or more enzymes or proteins in a microorganism, which is encoded by the corresponding polynucleotide or DNA.

For production of a strain in which the intracellular activity of a desired polypeptide is switched off, a deletion or insertion of at least one (1) nucleobase, preferably of one (1) or of two (2) nucleobases, is inserted in the coding region of the corresponding gene. It is also possible to delete at least one (1) or more codon(s) within the coding region. These measures lead to a shift of the reading frame (frame shift mutations) and therefore typically to the synthesis of a nonfunctional polypeptide. The introduction of a nonsense mutation by transversion or transition of at least one (1) nucleobase within the coding region has a similar effect. Owing to the stop codon that forms, there is premature termination of translation. The stated measures are preferably carried out in the region between the start codon and the penultimate coding codon, more preferably in the 5'-terminal portion of the coding region, which codes for the N-terminus of the polypeptide. If the total length of a polypeptide (measured as the number of chemically bound L-amino acids) is designated as 100%, then the portion of the amino acid sequence which, reckoned from the start amino acid L-formyl methionine, contains 80% of the subsequent L-amino acids, belongs to the N-terminus of the polypeptide.

Genetic measures for switching off malate-quinone oxidoreductase (Mqo) or reducing its expression are described for example in U.S. Pat. No. 7,094,106. U.S. Pat. No. 7,094,

106 describes switching off the *mqo* gene by incorporating deletions or insertions of at least one base pair or substitutions generating a stop codon into the *mqo* gene, wherein reduction of expression was achieved by placing the expression of the *mqo* gene under the control of the *E. coli* *trc* promoter/LacI^q repressor system.

Genetic measures for switching off malate dehydrogenase (Mdh) are described for example in WO 02/02778 (equivalent to U.S. Pat. No. 6,995,002). In WO 02/02778, the *mdh* gene was switched off by the insertion of a plasmid unable to replicate in *Corynebacterium glutamicum* comprising a central part of the coding region of the *mdh* gene into the host *mdh* gene by homologous recombination.

Genetic measures for switching off the E1p subunit (AceE) of the pyruvate dehydrogenase complex are described for example in EP 06119615 and in Schreiner et al. (Journal of Bacteriology 187(17), 6005-6018 (2005)). EP 06119615 and Schreiner et al. describe switching off the *aceE* gene by deleting a central part of the coding region of the *aceE* gene.

It is also possible, by suitable amino acid substitutions, to lower the catalytic property of the polypeptide in question.

In the case of malate-quinone oxidoreductase (Mqo) this can be achieved, as described in WO 06/077004, by preparing or using alleles of the *mqo* gene of SEQ ID NO: 23, which code for an Mqo variant that possesses the amino acid sequence of SEQ ID NO: 24 and contains one or more amino acid substitutions selected from the group

a) substitution of the L-serine at position 111 for another proteinogenic amino acid, preferably L-phenylalanine or L-alanine, and

b) substitution of the L-alanine at position 201 for another proteinogenic amino acid, preferably L-serine.

WO 06/077004 (equivalent to U.S. Pat. No. 7,214,526) describes an isolated coryneform bacterium mutant which comprises a gene encoding a polypeptide possessing malate quinone oxidoreductase enzyme activity, wherein the polypeptide comprises an amino acid sequence in which any proteinogenic amino acid except L-serine is present at position 111 or a comparable position.

Strains that comprise an *mqo* allele that codes for an Mqo variant which comprises the amino acid sequence of SEQ ID NO: 24, and contains L-phenylalanine at position 111, are preferred.

By the measures of switching-off or attenuation, the activity or concentration of the corresponding protein is generally lowered to 0 to 75%, 0 to 50%, 0 to 25%, 0 to 10% or 0 to 5% of the activity or concentration of the wild-type protein, or of the activity or concentration of the protein in the initial strain or parent strain.

The performance of the produced bacteria of the genus *Corynebacterium* or of the fermentation process using the produced bacteria with respect to one or more parameters selected from L-amino acid concentration (L-amino acid formed per volume), L-amino acid yield (L-amino acid formed per carbon source consumed), L-amino acid formation (L-amino acid formed per volume and time) and the specific L-amino acid formation (L-amino acid formed per cell dry mass or dry biomass and time or L-amino acid formed per cell protein and time) or other process variables and combinations thereof, is increased by at least 0.5%, at least 1%, at least 1.5% or at least 2% relative to the initial strain or parent strain or the fermentation process using them.

The produced bacteria of the genus *Corynebacterium* can be cultivated continuously, as described for example in PCT/EP2004/008882, or discontinuously in a batch process, a fed batch process or a repeated fed batch process, for the purpose of production of the desired L-amino acids. A summary of a

general nature covering known culture methods is given in Chmiel's textbook (Bioprozesstechnik 1 Einführung in die Bioverfahrenstechnik (Gustav Fischer Verlag, Stuttgart, 1991)) or in the textbook by Storhas (Bioreaktoren and periphere Einrichtungen (Vieweg Verlag, Braunschweig/Wiesbaden, 1994)). PCT/EP2004/008882 (equivalent to WO 05/021772 and DE 10339847) describes a fermentation process comprising incubating and culturing in at least first nutrient medium a coryneform bacterium producing L-lysine, feeding continuously further nutrient media to the culture in one or several streams and removing at the same time culturing broth with a removal stream or streams corresponding to the feed streams, wherein over the entire period of time of feeding and removing concentration of the source of carbon is not more than 10 g/L and L-lysine is formed.

The culture medium or fermentation medium to be used matches the requirements of the particular strains. Descriptions of culture media for various microorganisms are given in "Manual of Methods for General Bacteriology" of the American Society for Bacteriology (Washington D.C., USA, 1981). The terms culture medium and fermentation medium or medium are interchangeable.

Sugars and carbohydrates, for example glucose, sucrose, lactose, fructose, maltose, molasses, sucrose-containing solutions from sugar beet or sugar cane production, starch, hydrolyzed starch and cellulose, oils and fats, for example soya oil, sunflower oil, peanut oil and coconut oil, fatty acids, for example palmitic acid, stearic acid and linoleic acid, alcohols, for example glycerol, methanol and ethanol and organic acids, for example acetic acid or lactic acid can be used as the carbon source. These materials can be used individually or as a mixture.

Organic nitrogen-containing compounds such as peptones, yeast extract, meat extract, malt extract, corn-steep liquor, soybean flour and urea or inorganic compounds such as ammonium sulfate, ammonium chloride, ammonium phosphate, ammonium carbonate and ammonium nitrate can be used as the nitrogen source. The nitrogen sources can be used individually or as a mixture.

Phosphoric acid, potassium dihydrogen phosphate or dipotassium hydrogen phosphate or the corresponding sodium-containing salts can be used as the phosphorus source.

The culture medium in addition contains salts, for example, in the form of chlorides or sulfates of metals such as sodium, potassium, magnesium, calcium and iron, for example magnesium sulfate or iron sulfate, which are necessary for growth. Finally, essential growth substances can be used, such as amino acids for example homoserine and vitamins for example thiamine, biotin or pantothenic acid, in addition to the aforementioned substances.

The aforementioned ingredients can be added to the culture as a single charge, or can be supplied in a suitable manner during cultivation.

Basic compounds such as sodium hydroxide, potassium hydroxide, ammonia or ammonia water, or acidic compounds such as phosphoric acid or sulfuric acid can be used in a suitable manner for pH control of the culture. The pH is generally adjusted to a value of 6.0 to 9.0, preferably 6.5 to 8. Antifoaming agents, for example polyglycol esters of fatty acids, can be used for controlling foaming. To maintain the stability of plasmids, suitable substances with selective action, for example antibiotics, can be added to the medium. To maintain aerobic conditions, oxygen or oxygen-containing gas mixtures, for example air, are fed into the culture. The use of liquids enriched with hydrogen peroxide is also possible. Optionally, the fermentation is carried out at excess pressure, for example at a pressure of 0.03 to 0.2 MPa. The

temperature of the culture is normally in the range from 20° C. to 45° C. and preferably from 25° C. to 40° C. In batch processes, cultivation is continued until a maximum of the desired L-amino acid has formed at given conditions. This goal is normally reached within 10 hours to 160 hours. Longer cultivation times are possible with continuous processes. The activity of the bacteria leads to enrichment (accumulation) of the L-amino acid in the fermentation medium and/or in the bacterial cells.

Examples of suitable fermentation media are given inter alia in patent specifications U.S. Pat. No. 5,770,409, U.S. Pat. No. 5,840,551 and U.S. Pat. No. 5,990,350 or U.S. Pat. No. 5,275,940.

Methods for the determination of L-amino acids are known. The analysis can be carried out for example as described in Spackman et al. (Analytical Chemistry, 30, (1958), 1190) by anion-exchange chromatography followed by ninhydrin derivatization, or it can be carried out by reversed phase HPLC, as described by Lindroth et al. (Analytical Chemistry (1979) 51: 1167-1174).

Accordingly, the invention also relates to a method of production of an L-amino acid, wherein the following steps are carried out:

- a) fermentation of the bacteria according to the invention in a suitable nutrient medium,
- b) accumulation of the L-amino acid in the nutrient medium or in the cells of said bacteria.

These steps may be followed by collecting of the L-amino acid that accumulated in the nutrient medium, in the fermentation broth or in the cells of the bacteria, in order to obtain a solid or a liquid product.

A fermentation broth means a fermentation medium or nutrient medium in which a microorganism has been cultivated for a certain time and at a certain temperature. The fermentation medium or the media used during fermentation contain(s) all substances or components for ensuring multiplication of the microorganism and formation of the desired amino acid.

At the end of fermentation, the resulting fermentation broth accordingly contains a) the biomass (cell mass) of the microorganism, formed as a result of multiplication of the cells of the microorganism, b) the desired L-amino acid that formed in the course of fermentation, such as L-lysine, L-valine or L-isoleucine, c) the organic by-products that formed in the course of fermentation, and d) the constituents of the fermentation medium or of the ingredients for example vitamins such as biotin, amino acids such as homoserine or salts such as magnesium sulfate, that were not consumed in the fermentation.

The organic by-products include substances which may be produced and may be excreted by the microorganisms used in the fermentation in addition to the particular desired organic compound. These also include sugars, for example trehalose.

In the case of the amino acids L-valine and L-isoleucine, isolation and purification, for example using one or more methods selected from the group comprising chromatographic techniques, crystallization techniques and the use of activated charcoal, is preferred, so that pure products are largely obtained, for example products with purity of ≥ 90 wt. % or ≥ 95 wt. %.

In the case of the amino acid L-lysine, essentially four different product forms are known.

One group of L-lysine-containing products comprises concentrated, aqueous, alkaline solutions of purified L-lysine (EP-B-0534865). Another group, as described for example in U.S. Pat. No. 6,340,486 and U.S. Pat. No. 6,465,025, comprises aqueous, acidic, biomass-containing concentrates of

L-lysine-containing fermentation broths. Another group of solid products comprises powder or crystalline forms of purified or pure L-lysine, which may be in the form of a salt, for example L-lysine monohydrochloride. Yet another group of solid product forms is described for example in EP-B-0533039. The product form described there contains, in addition to L-lysine, most of the ingredients employed during fermentation but not consumed, and possibly the biomass of the microorganism used at a proportion of $>0\%$ - 100% .

In accordance with the various product forms, a great variety of methods is known for collecting, isolating or purifying the L-lysine from the fermentation broth, in order to produce an L-lysine-containing product or purified L-lysine.

Solid, pure L-lysine may be produced using methods of ion-exchange chromatography possibly with the use of activated charcoal and crystallization techniques. In this way we obtain the corresponding base or a corresponding salt, for example the monohydrochloride (Lys-HCl) or lysine sulfate (Lys₂-H₂SO₄).

A method of production of aqueous, basic L-lysine-containing solutions from fermentation broths is described in EP-B-0534865. In the method described there, the biomass is separated from the fermentation broth and discarded. A pH between 9 and 11 is established by means of a base, for example, sodium, potassium or ammonium hydroxide. The mineral constituents (inorganic salts) are separated from the broth after concentration and cooling, and either used as fertilizers or discarded.

In the case of methods for production of lysine using the bacteria, methods are preferred in which products are obtained that contain the constituents of the fermentation broth. These are used in particular as animal feed additives.

Depending on what is required, the biomass can be removed from the fermentation broth completely or partially by separation techniques such as centrifugation, filtration, decanting or a combination thereof, or it can be left in it completely. Optionally, the biomass or the fermentation broth containing the biomass is inactivated during a suitable process step, for example, by thermal treatment (heating) or by adding acid.

In one embodiment, the biomass is removed completely or almost completely, so that the finished product has a biomass content of zero (0%) or max. 30%, max. 20%, max. 10%, max. 5%, max. 1% or max. 0.1%. In another embodiment, the biomass is not removed or only a small proportion is removed, so that the finished product contains all the biomass (100%) or more than 70%, 80%, 90%, 95%, 99% or 99.9% biomass. In a method according to the invention, the biomass is accordingly removed in proportions from $\geq 0\%$ to $\leq 100\%$.

Finally, the fermentation broth obtained after the fermentation can be adjusted to an acid pH with an inorganic acid such as hydrochloric acid, sulfuric acid or phosphoric acid or organic acid such as propionic acid, before or after complete or partial removal of the biomass (GB 1,439,728 or EP 1 331 220). It is also possible to acidify the fermentation broth still containing all the biomass. Finally, the broth can also be stabilized by adding sodium bisulfite (NaHSO₃, GB 1,439,728) or another salt for example ammonium, alkali or alkaline-earth salt of sulfurous acid.

In separating the biomass, any organic or inorganic solids contained in the fermentation broth are removed partially or completely. The organic by-products dissolved in the fermentation broth and the dissolved, unconsumed constituents of the fermentation medium (ingredients) remain in the product at least partially ($>0\%$), preferably to at least 25%, preferably to at least 50% and more preferably to at least 75%. Optionally, these also remain in the product completely (100%) or

almost completely, i.e. >95% or >98% or over 99%. If a product in this sense contains at least a proportion of the constituents of the fermentation broth, it is also described with the term "product based on fermentation broth".

Then the broth is dewatered or thickened or concentrated using known methods, e.g. by means of a rotary evaporator, thin-film evaporator, falling-film evaporator, by reverse osmosis or by nanofiltration. This concentrated fermentation broth can then be processed by techniques of freeze drying, spray drying, spray granulation or by other methods, for example the circulating fluidized bed as described in PCT/EP2004/006655, to pourable products and preferably to a fine powder or preferably coarse granules. If required, a desired product can be isolated from the granules thus obtained by sieving or dust separation.

It is also possible for the fermentation broth to be dried directly, i.e. without previous concentration, by spray drying or spray granulation.

"Pourable" means powders which, from a series of glass discharge vessels with outlet openings of different sizes, are discharged freely from the vessel with the 5 mm (millimeter) opening (Klein: Seifen, Öle, Fette, Wachse 94, 12 1968)).

"Fine" means a powder having mainly (>50%) a grain size of 20 to 200 μm diameter. "Coarse" means a product having mainly (>50%) a grain size from 200 to 2000 μm diameter.

Grain size can be determined using methods of laser diffraction spectrometry. The relevant methods are described in the textbook on "Particle Size Measurement in Laboratory Practice" by R. H. Müller and R. Schuhmann, Wissenschaftliche Verlagsgesellschaft Stuttgart (1996) or in the textbook "Introduction to Particle Technology" by M. Rhodes, Publ. Wiley & Sons (1998).

The pourable, fine powder can be converted by suitable compaction or granulation techniques to a coarse, storable and largely dust-free product with good pourability.

The term "dust-free" means that the product only contains a small proportion (<5%) of particles with grain size under 100 μm diameter.

"Storable", in the sense of this invention, means a product that can be stored in cool, dry conditions for at least one (1) year or longer, preferably at least 1.5 years or longer, more preferably two (2) years or longer, without any substantial loss (max. 5%) of the particular amino acid.

The invention further relates to a method of manufacturing an L-lysine comprising product, which is described in broad outline in DE 102006016158, and in which the fermentation broth obtained using the microorganisms according to the invention, from which the biomass has been optionally separated completely or partially, is further processed, by carrying out a process that comprises at least the following steps:

a) the pH is lowered to 4.0-5.2, preferably 4.9-5.1, (by adding sulfuric acid, and a sulfate/L-lysine molar ratio of 0.85-1.2, preferably 0.9-1.0, more preferably >0.9 to <0.95, is established in the broth, if necessary by adding one or more additional sulfate-containing compound(s) and

b) the mixture thus obtained is concentrated by dewatering, and optionally granulated, optionally with one or both of the following measures being carried out before step a):

c) measurement of the sulfate/L-lysine molar ratio for determining the required amount of sulfate-containing compound(s)

d) addition of a sulfate-containing compound selected from the group comprising ammonium sulfate, ammonium hydrogensulfate and sulfuric acid in suitable proportions.

Optionally, also prior to step b), a salt of sulfurous acid, preferably an alkali metal hydrogensulfite, and more prefer-

ably sodium hydrogensulfite, at a concentration of 0.01-0.5 wt. %, preferably 0.1-0.3 wt. %, more preferably 0.1-0.2 wt. % relative to the fermentation broth is used.

DE 102006016158 (equivalent to US2007082031 and WO 07/042,363) describes relatively light and thermally stable granulated fermentation-broth-based animal feed additives having a high content of L-lysine and low-loss methods for production the additives from broths obtained by fermentation.

As preferred sulfate-containing compounds in the sense of the aforementioned process steps we may mention in particular ammonium sulfate and/or ammonium hydrogensulfate or corresponding mixtures of ammonia and sulfuric acid and sulfuric acid itself.

The sulfate/L-lysine molar ratio V is calculated from the formula: $V=2 \times [\text{SO}_4^{2-}]/[\text{L-lysine}]$. This formula takes account of the fact that the SO_4^{2-} anion, or sulfuric acid, is divalent. A ratio $V=1$ means that the $\text{Lys}_2\text{-H}_2\text{SO}_4$ is of stoichiometric composition, whereas at a ratio of $V=0.9$ there is a 10% sulfate deficit and at a ratio of $V=1.1$ there is a 10% sulfate excess.

During granulating or compacting the usual organic or inorganic auxiliaries, or carriers such as starch, gelatin, cellulose derivatives or similar substances, as are usually employed in the processing of foodstuffs or animal feed as binders, gelling agents or thickeners, or other substances for example silicic acids, silicates (EP0743016A) or stearates may be used.

Treatment the surface of the obtained granules with oils, as described in WO 04/054381 may be used. The oils used can be mineral oils, vegetable oils or mixtures of vegetable oils. Examples of such oils are soya oil, olive oil, soya oil/lecithin mixtures. Similarly, silicone oils, polyethylene glycols or hydroxyethyl cellulose are also suitable. Treatment of the surfaces with the aforesaid oils gives increased abrasion resistance of the product and a reduction in the proportion of dust. The content of oil in the product is 0.02-2.0 wt. %, preferably 0.02-1.0 wt. %, and more preferably 0.2-1.0 wt. % relative to the total amount of the feed additive.

Products are preferred having a proportion of ≥ 97 wt. % of a grain size from 100 to 1800 μm or a proportion of ≥ 95 wt. % of a grain size from 300 to 1800 μm diameter. The proportion of dust, i.e. particles with a grain size <100 μm , is preferably >0 to 1 wt. %, max. 0.5 wt. % being more preferred.

Alternatively, the product can also be coated with an organic or inorganic carrier that is known and usual in animal feed processing, for example silicic acids, silicates, grits, bran, meal or flour, starch, sugars or other substances and/or mixed and stabilized with usual thickeners or binders. Examples of applications and the methods employed are described in the literature (Die Mühle+Mischfüttertechnik 132 (1995) 49, page 817).

Finally, using coating processes with film-forming agents such as metal carbonates, silicic acids, silicates, alginates, stearates, starches, gums and cellulose ethers, as described in DE-C-4100920, the product can be brought to a state in which it is stable against digestion in animal stomachs preferably the stomach of ruminants. DE4100920 (equivalent to U.S. Pat. No. 5,279,832 and EP 0495349) describes an active-substance preparation for oral administration, preferably for ruminants, comprising an active-substance core comprising at least one biologically active substance and a coating around this core which delays the release of the core after oral administration due to its geometrical shape as well as a method of preparing an accordingly shaped core pellet by coating.

For establishing a desired L-lysine concentration in the product, depending on requirements, the L-lysine can be

added during the process in the form of a concentrate or optionally a substantially pure substance or salt thereof in liquid or solid form. These can be added individually or as mixtures to the fermentation broth obtained or concentrated, or alternatively during the drying or granulation process.

The invention relates further to a method of production of a solid lysine-containing product, as described in broad outline in US 20050220933, and which comprises the processing of the fermentation broth obtained using the microorganisms according to the invention, in the following steps:

a) filtration of the fermentation broth, preferably with a membrane filter, so that a biomass-containing sludge and a filtrate are obtained,

b) concentration of the filtrate, preferably so that a solids content of 48-52 wt. % is obtained,

c) granulation of the concentrate obtained in step b), preferably at a temperature from 50° C. to 62° C., and

d) coating of the granules obtained in c) with one or more coating agent(s)

For the coating in step d), it is preferable to use coating agents that are selected from the group comprising

d1) the biomass obtained in step a)

d2) a compound containing L-lysine, preferably selected from the group comprising L-lysine hydrochloride or L-lysine sulfate,

d3) an essentially L-lysine-free material with L-lysine content <1 wt. %, preferably <0.5 wt. %, preferably selected from the group consisting of starch, carrageenan, agar, silicic acids, silicates, grits, bran and meal, and

d4) a water-repellent substance, preferably selected from the group consisting of oils, polyethylene glycols and liquid paraffins.

The content of L-lysine is adjusted to a desired value by the measures corresponding to steps d1) to d4), preferably d1) to d3).

In the production of L-lysine-containing products, the ratio of the ions is preferably adjusted so that the molar ionic ratio according to the following formula

$$\frac{2 \times [\text{SO}_4^{2-}] + [\text{Cl}^-] - [\text{NH}_4^+] - [\text{Na}^+] - [\text{K}^+] - 2 \times [\text{Mg}^{2+}]}{2 \times [\text{Ca}^{2+}] / [\text{L-Lys}]} \text{ is } 0.68-0.95,$$

preferably 0.68-0.90, as described by Kushiki et al. in US 20030152633.

In the case of lysine, the solid product based on fermentation broth produced in this way has a lysine content (lysine base) from 10 wt. % to 70 wt. % or 20 wt. % to 70 wt. %, preferably 30 wt. % to 70 wt. % and more preferably from 40 wt. % to 70 wt. % based on the dry mass of the product. Maximum contents of lysine base of 71 wt. %, 72 wt. %, 73 wt. % are also possible. The water content of the L-lysine-containing, solid product is up to 5 wt. %, preferably up to 4 wt. %, and more preferably less than 3 wt. %. Having generally described this invention, a further understanding can be obtained by reference to certain specific examples which are provided herein for purposes of illustration only, and are not intended to be limiting unless otherwise specified.

EXAMPLES

Example 1

Sequencing of the *gltA* Gene of the Strain DM678

The strain *Corynebacterium glutamicum* DM678 (U.S. Pat. No. 6,861,246) is a lysine-producing strain developed by mutagenesis and screening. It is auxotrophic for L-threonine and L-methionine-sensitive. The strain was deposited at the

Deutsche Sammlung für Mikroorganismen and Zellkulturen (DSMZ, Braunschweig, Germany) as DSM12866.

The method of Eikmanns et al. (Microbiology 140: 1817-1828 (1994)) was used to isolate chromosomal DNA from the strain DM678. The polymerase chain reaction was used to amplify a DNA segment which harbors the *gltA* gene. The following oligonucleotides were used as primers for this:

gltA_XL-A1 (SEQ ID NO: 27):
5' tgagttctatgtggcgtgacc 3'

gltA_XL-E1 (SEQ ID NO: 28):
5' ttgccaaacgatgatgtcag 3'

The depicted primers were synthesized by MWG Biotech (Ebersberg, Germany). They make it possible to amplify a DNA segment which was about 1.8 kb long and harbors the *gltA* gene. The primer *gltA_XL-A1* binds to the region corresponding to position 490 to 509 of the strand complementary to SEQ ID NO: 3 (and SEQ ID NO: 7). The primer *gltA_XL-E1* binds to the region corresponding to position 2266 to 2247 of the strand shown in SEQ ID NO: 3 (and SEQ ID NO: 7).

The PCR reaction was carried out using the Phusion high fidelity DNA polymerase (New England Biolabs, Frankfurt, Germany). The reaction mixture was made up as specified by the manufacturer and contained 10 µl of the 5× Phusion HF buffer supplied, deoxynucleoside triphosphates each in a concentration of 200 µM, primers in a concentration of 0.5 µM, approximately 50 ng of template DNA and 2 units of Phusion polymerase in a total volume of 50 µl. The volume was adjusted to 50 µl by adding H₂O.

The PCR mixture was first subjected to an initial denaturation at 98° C. for 30 seconds. This was followed by a denaturation step at 98° C. for 20 seconds, repeated 35×, a step for binding the primers to the introduced DNA at 60° C. for 20 seconds and the extension step to extend the primers at 72° C. for 60 seconds. After the final extension step at 72° C. for 5 minutes, the PCR mixture was subjected to an agarose gel electrophoresis (0.85% agarose). A DNA fragment about 1.8 kb long was identified, isolated from the gel and purified using the QIAquick gel extraction kit from Qiagen (Hilden, Germany).

The nucleotide sequence of the amplified DNA fragment or PCR product was determined by Agowa (Berlin, Germany).

The nucleotide sequence of the coding region of the *gltA* allele from the strain DM678 contains thymine as nucleobase at position 14. The wild-type gene (see SEQ ID NO: 1) contains adenine as nucleobase at this position. This adenine-thymine transversion leads to an amino acid exchange from aspartate to valine at position 5 of the resulting amino acid sequence. This mutation is referred to hereinafter as *gltAD5V*. The allele *gltAD5V* is depicted in SEQ ID NO: 5, and the amino acid sequence of the protein which was revealed with the aid of the Patent program is depicted in SEQ ID NO: 6.

Example 2

Construction of the Exchange Vector pK18mobsacB_ *gltAD5V*

The polymerase chain reaction was used to amplify a DNA fragment which harbors part of the upstream region of the *gltA* gene and part of the coding region which contains the *gltAD5V* mutation. The chromosomal DNA obtained in

example 1 from DM678 was used as template. The following oligonucleotides were selected as primers for the PCR:

```
gltA_1.p (SEQ ID NO: 29):
5' CCGTCGACAATAGCCTGAA 3'

gltA_2.p (SEQ ID NO: 30):
5' CC-GAATTC-TTCGAGCATCTCCAGAAC 3'
```

They were synthesized by MWG Biotech (Ebersberg, Germany) and make it possible to amplify a DNA segment about 1.7 kb long comprising 832 bp of the upstream region and nucleotides 1-855 bp of the coding region of the *gltA* gene from DM678.

The primer *gltA_1.p* binds to the region corresponding to position 169 to 187 of the strand complementary to SEQ ID NO: 25. Nucleotides 9 to 26 of the primer *gltA_2.p* bind to the region corresponding to position 1855 to 1838 of the strand shown in SEQ ID NO: 25. In addition, the primer *gltA_1.p* contains the native cleavage site of the restriction enzyme *Sall*, and the primer *gltA_2.p* contains the sequence of the cleavage site of the restriction enzyme *EcoRI*, which are each marked by underlining in the nucleotide sequence depicted above.

The PCR reaction was carried out using the Phusion high fidelity DNA polymerase (New England Biolabs, Frankfurt, Germany). The reaction mixture had the composition described above. The PCR was carried out as described above. The nucleotide sequence of the amplicon about 1.7 kb long is depicted in SEQ ID NO: 31.

The amplicon was treated with the restriction endonucleases *Sall* and *EcoRI* and identified by electrophoresis in a 0.8% agarose gel. It was subsequently isolated from the gel and purified using the QIAquick gel extraction kit from Qiagen.

The DNA fragment purified in this way contains the described *gltAD5V* mutation and has ends compatible with DNA cut with *Sall* and *EcoRI* (respectively *gltAD5V* fragment and *gltA'* in the FIGURE). It was subsequently cloned into the mobilizable vector *pK18mobsacB* described by Schäfer et al. (Gene, 145, 69-73 (1994)) in order to make an allelic or mutation substitution possible. For this purpose, *pK18mobsacB* was digested with the restriction enzymes *EcoRI* and *Sall*, and the ends were dephosphorylated with alkaline phosphatase (alkaline phosphatase, Boehringer Mannheim, Germany). The vector prepared in this way was mixed with the *gltAD5V* fragment, and the mixture was treated with the ready-to-go T4 DNA ligase kit (Amersham-Pharmacia, Freiburg, Germany).

Subsequently, the *E. coli* strain S17-1 (Simon et al., Bio/Technologie 1: 784-791, 1993) was transformed with the ligation mixture (Hanahan, In. DNA cloning. A practical approach. Vol. 1. ILR-Press, Cold Spring Harbor, N.Y., 1989). Selection of plasmid-harboring cells took place by plating out the transformation mixture on LB agar (Sambrook et al., Molecular Cloning: a laboratory manual. 2nd Ed. Cold Spring Harbor, N.Y., 1989) which had been supplemented with 25 mg/l kanamycin.

Plasmid DNA was isolated from a transformant using the QIAprep spin miniprep kit from Qiagen and checked by restriction cleavage with the enzymes *Sall* and *EcoRI* and subsequent agarose gel electrophoresis. The plasmid was called *pK18mobsacB_gltAD5V* and is depicted in the FIGURE. The abbreviations and designations used have the following meaning. The stated numbers of base pairs are approximations obtained within the scope of the reproducibility of measurements.

Kan:	kanamycin-resistance gene
<i>Sall</i> :	cleavage site of the restriction enzyme <i>Sall</i>
<i>EcoRI</i> :	cleavage site of the restriction enzyme <i>EcoRI</i>
<i>gltA'</i> :	cloned DNA fragment containing the <i>gltAD5V</i> mutation
<i>sacB</i> :	<i>sacB</i> gene
RP4-mob:	mob region with the origin of replication for transfer (<i>oriT</i>)
<i>oriV</i> :	origin of replication V

Example 3

Incorporation of the *gltAD5V* Mutation into the Strain *Corynebacterium glutamicum* DM1797.

The intention was to introduce the *gltAD5V* mutation into the strain *Corynebacterium glutamicum* DM1797. The strain DM1797 was an aminoethylcysteine-resistant mutant of *Corynebacterium glutamicum* ATCC13032 and described in PCT/EP/2005/012417. It was deposited under the number DSM16833 at the Deutsche Sammlung für Mikroorganismen und Zellkulturen (DSMZ, Braunschweig, Germany).

The vector *pK18mobsacB_gltAD5V* described in example 2 was transferred by conjugation according to the protocol of Schäfer et al. (Journal of Microbiology 172: 1663-1666 (1990)) into the *C. glutamicum* strain DM1797. The vector was incapable of independent replication in DM1797 and was retained in the cell only if it was integrated into the chromosome as the result of a recombination event. Selection of transconjugants, i.e. of clones having integrated *pK18mobsacB_gltAD5V*, took place by plating out the conjugation mixture on LB agar which had been supplemented with 25 mg/l kanamycin and 50 mg/l of nalidixic acid. Kanamycin-resistant transconjugants were subsequently streaked onto LB agar plates supplemented with kanamycin (25 mg/l) and incubated at 33° C. for 24 hours. Mutants in which, as a result of a second recombination event, excision of the plasmid had taken place were selected by culturing the clones nonselectively in LB liquid medium for 30 hours, then streaking onto LB agar, which had been supplemented with 10% sucrose, and incubating at 33° C. for 24 hours.

The plasmid *pK18mobsacB_gltAD5V* contains, just like the initial plasmid *pK18mobsacB*, besides the kanamycin-resistance gene a copy of the *sacB* gene which codes for the levan sucrose from *Bacillus subtilis*. The sucrose-inducible expression of the *sacB* gene leads to the formation of levan sucrose which catalyzes the synthesis of the product levan which is toxic for *C. glutamicum*. Thus, the only clones to grow on sucrose-supplemented LB agar were those in which the integrated *pK18mobsacB_gltAD5V* has been excised as the result of a second recombination event. Depending on the position of the second recombination event in relation to the site of mutation, the excision is associated with allelic substitution or incorporation of the mutation instead, or the original copy remains in the host's chromosome.

A clone in which the desired exchange, i.e. the incorporation of the *gltAD5V* mutation, had taken place was then sought. For this purpose, the sequence of the *gltA* gene was determined for 10 clones with the phenotype "growth in the presence of sucrose" and "non-growth in the presence of kanamycin". In this way, a clone harboring the *gltAD5V* mutation was identified. This strain was designated *C. glutamicum* DM1797_ *gltAD5V*.

Example 4

Production of L-Lysine

The strain DM1797_gltAD5V obtained in example 3 and the initial strain DM1797 were cultured in a nutrient medium suitable for producing lysine, and the lysine content in the culture supernatant was determined.

For this purpose, the clones were initially grown on brain-heart agar plates (Merck, Darmstadt, Germany) at 33° C. for 24 hours. These agar plate cultures were each used for inoculation of a preculture (10 ml of medium in a 100 ml Erlenmeyer flask). The medium MM was used as medium for the precultures. The precultures were incubated at 33° C. and 240 rpm on a shaker for 24 hours. Each of these precultures was used to inoculate a main culture, so that the initial OD (660 nm) of the main culture was 0.1 OD. The medium MM was likewise used for the main cultures.

Medium MM	
CSL	5 g/l
MOPS	20 g/l
Glucose (autoclaved separately)	50 g/l
Salts:	
(NH ₄) ₂ SO ₄	25 g/l
KH ₂ PO ₄	0.1 g/l
MgSO ₄ * 7 H ₂ O	1.0 g/l
CaCl ₂ * 2 H ₂ O	10 mg/l
FeSO ₄ * 7 H ₂ O	10 mg/l
MnSO ₄ * H ₂ O	5.0 mg/l
Biotin (sterilized by filtration)	0.3 mg/l
Thiamine * HCl (sterilized by filtration)	0.2 mg/l
CaCO ₃	25 g/l

CSL (corn steep liquor), MOPS (morpholinopropane-sulfonic acid) and the salt solution were adjusted to pH 7 with aqueous ammonia and autoclaved. The sterile substrate and vitamin solutions, and the dry autoclaved CaCO₃, were then added.

Culturing took place in volumes of 10 ml which were present in 100 ml Erlenmeyer flasks with baffles. The temperature was at 33° C., the number of revolutions was 250 rpm and the humidity was 80%.

After 48 hours, the optical density (OD) was determined at a measurement wavelength of 660 nm using a Biomek 1000 (Beckmann Instruments GmbH, Munich). The amount of lysine formed was determined using an amino acid analyzer from Eppendorf-BioTronik (Hamburg, Germany) by ion-exchange chromatography and post-column derivatization with ninhydrin detection.

TABLE 1

Strain	OD (660)	Lysine HCl (g/l)
DM1797	11.9	4.8
DM1797_gltAD5V	11.2	5.3

Example 5

Incorporation of the gltAD5V Mutation into the Strain *Brevibacterium Lactofermentum* FERM BP-1763

It was intended to introduce the gltAD5V mutation into the strain *Brevibacterium lactofermentum* FERM BP-1763. The strain FERM BP-1763 is a mycophenolic acid-resistant

valine producer (U.S. Pat. No. 5,188,948). It is auxotrophic for L-isoleucine and L-methionine.

The vector pK18mobsacB_gltAD5V described in example 2 was transferred by conjugation according to the protocol of Schäfer et al. (Journal of Microbiology 172: 1663-1666 (1990)) into the strain FERM-BP-1763. The vector was incapable of independent replication in FERM BP-1763 and was retained in the cell only if it was integrated into the chromosome as the result of a recombination event. Selection of transconjugants, i.e. of clones having integrated pK18mobsacB_gltAD5V, took place by plating out the conjugation mixture on LB agar which had been supplemented with 25 mg/l kanamycin and 50 mg/l of nalidixic acid. Kanamycin-resistant transconjugants were subsequently streaked onto LB agar plates supplemented with kanamycin (25 mg/l) and incubated at 33° C. for 24 hours. Mutants in which, as a result of a second recombination event, excision of the plasmid had taken place were selected by culturing the clones nonselectively in LB liquid medium for 30 hours, then streaking onto LB agar, which had been supplemented with 10% sucrose, and incubating at 33° C. for 24 hours.

The plasmid pK18mobsacB_gltAD5V contained, just like the initial plasmid pK18mobsacB, besides the kanamycin-resistance gene a copy of the sacB gene which coded for the levan sucrose from *Bacillus subtilis*. The sucrose-inducible expression of the sacB gene led to the formation of levan sucrose which catalyzes the synthesis of the product levan which was toxic for *C. glutamicum*. Thus, the only clones to grow on sucrose-supplemented LB agar were those in which the integrated pK18mobsacB_gltAD5V has been excised as the result of a second recombination event. Depending on the position of the second recombination event in relation to the site of mutation, the excision was associated with allelic substitution or incorporation of the mutation instead, or the original copy remains in the host's chromosome.

A clone in which the desired exchange, i.e. the incorporation of the gltAD5V mutation, had taken place was then sought. For this purpose, the sequence of the gltA gene was determined for 10 clones with the phenotype "growth in the presence of sucrose" and "non-growth in the presence of kanamycin". In this way, a clone harboring the gltAD5V mutation was identified. This strain was designated *C. glutamicum* FERM BP-1763_gltAD5V.

Example 6

Production of L-Valine

The strain FERM BP-1763_gltAD5V obtained in example 5 and the initial strain FERM BP-1763 were cultured in a nutrient medium suitable for producing valine, and the valine content in the culture supernatant was determined.

For this purpose, the clones were initially grown on brain-heart agar plates (Merck, Darmstadt, Germany) at 33° C. for 24 hours. These agar plate cultures were each used for inoculation of a preculture (10 ml of medium in a 100 ml Erlenmeyer flask).

The medium CgIII (2.5 g/l NaCl, 10 g/l Bacto peptone, 10 g/l Bacto yeast extract, pH 7.4, 20 g/l glucose (autoclaved separately) was used for the precultures. The precultures were incubated at 33° C. and 240 rpm on a shaker for 24 hours. Each of these precultures was used to inoculate a main culture, so that the initial OD (660 nm) of the main culture was 0.1 OD. The medium MM was likewise used for the main cultures.

Medium MM	
CSL	5 g/l
MOPS	20 g/l
Glucose (autoclaved separately)	50 g/l
Salts:	
(NH ₄) ₂ SO ₄	25 g/l
KH ₂ PO ₄	0.1 g/l
MgSO ₄ * 7 H ₂ O	1.0 g/l
CaCl ₂ * 2 H ₂ O	10 mg/l
FeSO ₄ * 7 H ₂ O	10 mg/l
MnSO ₄ * H ₂ O	5.0 mg/l
Isoleucine (sterilized by filtration)	0.1 g/l
Methionine (sterilized by filtration)	0.1 g/l
Leucine (sterilized by filtration)	0.1 g/l
Thiamine * HCl (sterilized by filtration)	0.2 mg/l
CaCO ₃	25 g/l

CSL (corn steep liquor), MOPS (morpholinopropane-sulfonic acid) and the salt solution were adjusted to pH 7 with aqueous ammonia and autoclaved. The sterile substrate and vitamin solutions, and the dry autoclaved CaCO₃, were then added.

Culturing took place in volumes of 10 ml which were present in 100 ml Erlenmeyer flasks with baffles. The temperature was at 33° C., the number of revolutions was 250 rpm and the humidity was 80%.

After 48 hours, the optical density (OD) was determined at a measurement wavelength of 660 nm using the Biomek 1000 (Beckmann Instruments GmbH, Munich). The amount of valine formed was determined using an amino acid analyzer from Eppendorf-BioTronik (Hamburg, Germany) by ion-exchange chromatography and post-column derivatization with ninhydrin detection.

TABLE 2

Strain	OD (660)	Valine (g/l)
FERM BP-1763	8.4	11.9
FERM BP-1763_dltAD5V	7.8	12.6

German patent application 102006032634.2, filed Jul. 13, 2006, and U.S. provisional application 60/830,331, filed Jul. 13, 2006, are incorporated herein by reference.

Numerous modification and variations on the present invention are possible in light of the above teachings. It is therefore to be understood that within the scope of the appended claims, the invention may be practiced otherwise than as specifically described herein.

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 32

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 <222> LOCATION: (1)..(1311)
 <223> OTHER INFORMATION: gltA-Wildtype-Gene
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (13)..(15)

<400> SEQUENCE: 1

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1           5           10           15

cac tac ccc ggt ggc gag ttc gaa atg gac atc atc gag gct tct gag      96
His Tyr Pro Gly Gly Glu Phe Glu Met Asp Ile Ile Glu Ala Ser Glu
                20           25           30

ggt aac aac ggt gtt gtc ctg ggc aag atg ctg tct gag act gga ctg      144
Gly Asn Asn Gly Val Val Leu Gly Lys Met Leu Ser Glu Thr Gly Leu
                35           40           45

atc act ttt gac cca ggt tat gtg agc act ggc tcc acc gag tcg aag      192
Ile Thr Phe Asp Pro Gly Tyr Val Ser Thr Gly Ser Thr Glu Ser Lys
                50           55           60

atc acc tac atc gat ggc gat gcg gga atc ctg cgt tac cgc ggc tat      240
Ile Thr Tyr Ile Asp Gly Asp Ala Gly Ile Leu Arg Tyr Arg Gly Tyr
        65           70           75           80

gac atc gct gat ctg gct gag aat gcc acc ttc aac gag gtt tct tac      288
Asp Ile Ala Asp Leu Ala Glu Asn Ala Thr Phe Asn Glu Val Ser Tyr
                85           90           95

cta ctt atc aac ggt gag cta cca acc cca gat gag ctt cac aag ttt      336
Leu Leu Ile Asn Gly Glu Leu Pro Thr Pro Asp Glu Leu His Lys Phe
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aac gac gag att cgc cac cac acc ctt ctg gac gag gac ttc aag tcc Asn Asp Glu Ile Arg His His Thr Leu Leu Asp Glu Asp Phe Lys Ser 115 120 125	384
cag ttc aac gtg ttc cca cgc gac gct cac cca atg gca acc ttg gct Gln Phe Asn Val Phe Pro Arg Asp Ala His Pro Met Ala Thr Leu Ala 130 135 140	432
tcc tcg gtt aac att ttg tct acc tac tac cag gac cag ctg aac cca Ser Ser Val Asn Ile Leu Ser Thr Tyr Tyr Gln Asp Gln Leu Asn Pro 145 150 155 160	480
ctc gat gag gca cag ctt gat aag gca acc gtt cgc ctc atg gca aag Leu Asp Glu Ala Gln Leu Asp Lys Ala Thr Val Arg Leu Met Ala Lys 165 170 175	528
gtt cca atg ctg gct gcg tac gca cac cgc gca cgc aag ggt gct cct Val Pro Met Leu Ala Ala Tyr Ala His Arg Ala Arg Lys Gly Ala Pro 180 185 190	576
tac atg tac cca gac aac tcc ctc aat gcg cgt gag aac ttc ctg cgc Tyr Met Tyr Pro Asp Asn Ser Leu Asn Ala Arg Glu Asn Phe Leu Arg 195 200 205	624
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gtc aag gct ctg gac aag ctg ctc atc ctg cac gct gac cac gag cag Val Lys Ala Leu Asp Lys Leu Leu Ile Leu His Ala Asp His Glu Gln 225 230 235 240	720
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cac ggt ggc gca aac cag gct gtt ctg gag atg ctc gaa gac atc aag His Gly Gly Ala Asn Gln Ala Val Leu Glu Met Leu Glu Asp Ile Lys 275 280 285	864
agc aac cac ggt ggc gac gca acc gag ttc atg aac aag gtc aag aac Ser Asn His Gly Gly Asp Ala Thr Glu Phe Met Asn Lys Val Lys Asn 290 295 300	912
aag gaa gac ggc gtc cgc ctc atg ggc ttc gga cac cgc gtt tac aag Lys Glu Asp Gly Val Arg Leu Met Gly Phe Gly His Arg Val Tyr Lys 305 310 315 320	960
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cca act gac ttc ttc acc gta ttg ttc gca atc ggt cgt ctg cca gga Pro Thr Asp Phe Phe Thr Val Leu Phe Ala Ile Gly Arg Leu Pro Gly 385 390 395 400	1200
tgg atc gct cac tac cgc gag cag ctc ggt gca gca ggc aac aag atc Trp Ile Ala His Tyr Arg Glu Gln Leu Gly Ala Ala Gly Asn Lys Ile 405 410 415	1248
aac cgc cca cgc cag gtc tac acc ggc aac gaa tcc cgc aag ttg gtt Asn Arg Pro Arg Gln Val Tyr Thr Gly Asn Glu Ser Arg Lys Leu Val 420 425 430	1296

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 35 40 45
 Ile Thr Phe Asp Pro Gly Tyr Val Ser Thr Gly Ser Thr Glu Ser Lys
 50 55 60
 Ile Thr Tyr Ile Asp Gly Asp Ala Gly Ile Leu Arg Tyr Arg Gly Tyr
 65 70 75 80
 Asp Ile Ala Asp Leu Ala Glu Asn Ala Thr Phe Asn Glu Val Ser Tyr
 85 90 95
 Leu Leu Ile Asn Gly Glu Leu Pro Thr Pro Asp Glu Leu His Lys Phe
 100 105 110
 Asn Asp Glu Ile Arg His His Thr Leu Leu Asp Glu Asp Phe Lys Ser
 115 120 125
 Gln Phe Asn Val Phe Pro Arg Asp Ala His Pro Met Ala Thr Leu Ala
 130 135 140
 Ser Ser Val Asn Ile Leu Ser Thr Tyr Tyr Gln Asp Gln Leu Asn Pro
 145 150 155 160
 Leu Asp Glu Ala Gln Leu Asp Lys Ala Thr Val Arg Leu Met Ala Lys
 165 170 175
 Val Pro Met Leu Ala Ala Tyr Ala His Arg Ala Arg Lys Gly Ala Pro
 180 185 190
 Tyr Met Tyr Pro Asp Asn Ser Leu Asn Ala Arg Glu Asn Phe Leu Arg
 195 200 205
 Met Met Phe Gly Tyr Pro Thr Glu Pro Tyr Glu Ile Asp Pro Ile Met
 210 215 220
 Val Lys Ala Leu Asp Lys Leu Leu Ile Leu His Ala Asp His Glu Gln
 225 230 235 240
 Asn Cys Ser Thr Ser Thr Val Arg Met Ile Gly Ser Ala Gln Ala Asn
 245 250 255
 Met Phe Val Ser Ile Ala Gly Gly Ile Asn Ala Leu Ser Gly Pro Leu
 260 265 270
 His Gly Gly Ala Asn Gln Ala Val Leu Glu Met Leu Glu Asp Ile Lys
 275 280 285
 Ser Asn His Gly Gly Asp Ala Thr Glu Phe Met Asn Lys Val Lys Asn
 290 295 300
 Lys Glu Asp Gly Val Arg Leu Met Gly Phe Gly His Arg Val Tyr Lys
 305 310 315 320
 Asn Tyr Asp Pro Arg Ala Ala Ile Val Lys Glu Thr Ala His Glu Ile
 325 330 335
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 340 345 350
 Glu Glu Ile Ala Leu Ala Asp Asp Tyr Phe Ile Ser Arg Lys Leu Tyr
 355 360 365

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Pro Asn Val Asp Phe Tyr Thr Gly Leu Ile Tyr Arg Ala Met Gly Phe
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Pro Thr Asp Phe Phe Thr Val Leu Phe Ala Ile Gly Arg Leu Pro Gly
 385 390 395 400

Trp Ile Ala His Tyr Arg Glu Gln Leu Gly Ala Ala Gly Asn Lys Ile
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 <223> OTHER INFORMATION: sequence upatream of the coding region
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 <222> LOCATION: (751)..(2061)
 <223> OTHER INFORMATION: gltA-Wild-Type-Gene
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 <223> OTHER INFORMATION: sequence downstream of the coding region

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 Thr Asp Asn Asn Lys Ala Val Leu His Tyr Pro Gly Gly Glu Phe Glu
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 atg gac atc atc gag gct tct gag ggt aac aac ggt gtt gtc ctg ggc 870
 Met Asp Ile Ile Glu Ala Ser Glu Gly Asn Asn Gly Val Val Leu Gly
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 aag atg ctg tct gag act gga ctg atc act ttt gac cca ggt tat gtg 918
 Lys Met Leu Ser Glu Thr Gly Leu Ile Thr Phe Asp Pro Gly Tyr Val
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 agc act ggc tcc acc gag tgc aag atc acc tac atc gat ggc gat gcg 966

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      395                               400                               405

ctc ggt gca gca ggc aac aag atc aac cgc cca cgc cag gtc tac acc   2022
Leu Gly Ala Ala Gly Asn Lys Ile Asn Arg Pro Arg Gln Val Tyr Thr
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gaaaagccac agattgagct accggtcggg ccagcaccgg aagatctcgt aatctctgac   2251

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ggcggacgtc gtcagctgac cattccgcca gaggtgctt acggccctga gggttccggc   2491

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<212> TYPE: PRT
<213> ORGANISM: Corynebacterium glutamicum

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<400> SEQUENCE: 4

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Gly Asn Asn Gly Val Val Leu Gly Lys Met Leu Ser Glu Thr Gly Leu
      35           40           45

Ile Thr Phe Asp Pro Gly Tyr Val Ser Thr Gly Ser Thr Glu Ser Lys
      50           55           60

Ile Thr Tyr Ile Asp Gly Asp Ala Gly Ile Leu Arg Tyr Arg Gly Tyr
      65           70           75           80

Asp Ile Ala Asp Leu Ala Glu Asn Ala Thr Phe Asn Glu Val Ser Tyr
      85           90           95

Leu Leu Ile Asn Gly Glu Leu Pro Thr Pro Asp Glu Leu His Lys Phe
      100          105          110

Asn Asp Glu Ile Arg His His Thr Leu Leu Asp Glu Asp Phe Lys Ser
      115          120          125

Gln Phe Asn Val Phe Pro Arg Asp Ala His Pro Met Ala Thr Leu Ala
      130          135          140

Ser Ser Val Asn Ile Leu Ser Thr Tyr Tyr Gln Asp Gln Leu Asn Pro
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atc act ttt gac cca ggt tat gtg agc act ggc tcc acc gag tcg aag	192
Ile Thr Phe Asp Pro Gly Tyr Val Ser Thr Gly Ser Thr Glu Ser Lys	
50 55 60	
atc acc tac atc gat ggc gat gcg gga atc ctg cgt tac cgc ggc tat	240
Ile Thr Tyr Ile Asp Gly Asp Ala Gly Ile Leu Arg Tyr Arg Gly Tyr	
65 70 75 80	
gac atc gct gat ctg gct gag aat gcc acc ttc aac gag gtt tct tac	288
Asp Ile Ala Asp Leu Ala Glu Asn Ala Thr Phe Asn Glu Val Ser Tyr	
85 90 95	
cta ctt atc aac ggt gag cta cca acc cca gat gag ctt cac aag ttt	336
Leu Leu Ile Asn Gly Glu Leu Pro Thr Pro Asp Glu Leu His Lys Phe	
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Asn Asp Glu Ile Arg His His Thr Leu Leu Asp Glu Asp Phe Lys Ser	
115 120 125	
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Gln Phe Asn Val Phe Pro Arg Asp Ala His Pro Met Ala Thr Leu Ala	
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Ser Ser Val Asn Ile Leu Ser Thr Tyr Tyr Gln Asp Gln Leu Asn Pro	
145 150 155 160	
ctc gat gag gca cag ctt gat aag gca acc gtt cgc ctc atg gca aag	528
Leu Asp Glu Ala Gln Leu Asp Lys Ala Thr Val Arg Leu Met Ala Lys	
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gtt cca atg ctg gct gcg tac gca cac cgc gca cgc aag ggt gct cct	576
Val Pro Met Leu Ala Ala Tyr Ala His Arg Ala Arg Lys Gly Ala Pro	
180 185 190	
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Tyr Met Tyr Pro Asp Asn Ser Leu Asn Ala Arg Glu Asn Phe Leu Arg	
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Met Met Phe Gly Tyr Pro Thr Glu Pro Tyr Glu Ile Asp Pro Ile Met	
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Val Lys Ala Leu Asp Lys Leu Leu Ile Leu His Ala Asp His Glu Gln	
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245 250 255	
atg ttt gtc tcc atc gct ggt ggc atc aac gct ctg tcc ggc cca ctg	816
Met Phe Val Ser Ile Ala Gly Gly Ile Asn Ala Leu Ser Gly Pro Leu	
260 265 270	
cac ggt ggc gca aac cag gct gtt ctg gag atg ctc gaa gac atc aag	864
His Gly Gly Ala Asn Gln Ala Val Leu Glu Met Leu Glu Asp Ile Lys	
275 280 285	
agc aac cac ggt ggc gac gca acc gag ttc atg aac aag gtc aag aac	912
Ser Asn His Gly Gly Asp Ala Thr Glu Phe Met Asn Lys Val Lys Asn	
290 295 300	
aag gaa gac ggc gtc cgc ctc atg ggc ttc gga cac cgc gtt tac aag	960
Lys Glu Asp Gly Val Arg Leu Met Gly Phe Gly His Arg Val Tyr Lys	
305 310 315 320	
aac tac gat cca cgt gca gca atc gtc aag gag acc gca cac gag atc	1008
Asn Tyr Asp Pro Arg Ala Ala Ile Val Lys Glu Thr Ala His Glu Ile	
325 330 335	
ctc gag cac ctc ggt ggc gac gat ctt ctg gat ctg gca atc aag ctg	1056
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340 345 350	

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ccg aac gta gac ttc tac acc ggc ctg atc tac cgc gca atg ggc ttc    1152
Pro Asn Val Asp Phe Tyr Thr Val Gly Leu Ile Tyr Arg Ala Met Gly Phe
      370                      375                      380

cca act gac ttc ttc acc gta ttg ttc gca atc ggt cgt ctg cca gga    1200
Pro Thr Asp Phe Phe Thr Val Leu Phe Ala Ile Gly Arg Leu Pro Gly
      385                      390                      395                      400

tgg atc gct cac tac cgc gag cag ctc ggt gca gca ggc aac aag atc    1248
Trp Ile Ala His Tyr Arg Glu Gln Leu Gly Ala Ala Gly Asn Lys Ile
      405                      410                      415

aac cgc cca cgc cag gtc tac acc ggc aac gaa tcc cgc aag ttg gtt    1296
Asn Arg Pro Arg Gln Val Tyr Thr Gly Asn Glu Ser Arg Lys Leu Val
      420                      425                      430

cct cgc gag gag cgc taa                                          1314
Pro Arg Glu Glu Arg
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<210> SEQ ID NO 6
<211> LENGTH: 437
<212> TYPE: PRT
<213> ORGANISM: Corynebacterium glutamicum

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<400> SEQUENCE: 6

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 20     25     30

Gly Asn Asn Gly Val Val Leu Gly Lys Met Leu Ser Glu Thr Gly Leu
 35     40     45

Ile Thr Phe Asp Pro Gly Tyr Val Ser Thr Gly Ser Thr Glu Ser Lys
 50     55     60

Ile Thr Tyr Ile Asp Gly Asp Ala Gly Ile Leu Arg Tyr Arg Gly Tyr
 65     70     75     80

Asp Ile Ala Asp Leu Ala Glu Asn Ala Thr Phe Asn Glu Val Ser Tyr
 85     90     95

Leu Leu Ile Asn Gly Glu Leu Pro Thr Pro Asp Glu Leu His Lys Phe
100    105    110

Asn Asp Glu Ile Arg His His Thr Leu Leu Asp Glu Asp Phe Lys Ser
115    120    125

Gln Phe Asn Val Phe Pro Arg Asp Ala His Pro Met Ala Thr Leu Ala
130    135    140

Ser Ser Val Asn Ile Leu Ser Thr Tyr Tyr Gln Asp Gln Leu Asn Pro
145    150    155    160

Leu Asp Glu Ala Gln Leu Asp Lys Ala Thr Val Arg Leu Met Ala Lys
165    170    175

Val Pro Met Leu Ala Ala Tyr Ala His Arg Ala Arg Lys Gly Ala Pro
180    185    190

Tyr Met Tyr Pro Asp Asn Ser Leu Asn Ala Arg Glu Asn Phe Leu Arg
195    200    205

Met Met Phe Gly Tyr Pro Thr Glu Pro Tyr Glu Ile Asp Pro Ile Met
210    215    220

Val Lys Ala Leu Asp Lys Leu Leu Ile Leu His Ala Asp His Glu Gln
225    230    235    240

Asn Cys Ser Thr Ser Thr Val Arg Met Ile Gly Ser Ala Gln Ala Asn
245    250    255

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Met Phe Val Ser Ile Ala Gly Gly Ile Asn Ala Leu Ser Gly Pro Leu
 260 265 270

His Gly Gly Ala Asn Gln Ala Val Leu Glu Met Leu Glu Asp Ile Lys
 275 280 285

Ser Asn His Gly Gly Asp Ala Thr Glu Phe Met Asn Lys Val Lys Asn
 290 295 300

Lys Glu Asp Gly Val Arg Leu Met Gly Phe Gly His Arg Val Tyr Lys
 305 310 315 320

Asn Tyr Asp Pro Arg Ala Ala Ile Val Lys Glu Thr Ala His Glu Ile
 325 330 335

Leu Glu His Leu Gly Gly Asp Asp Leu Leu Asp Leu Ala Ile Lys Leu
 340 345 350

Glu Glu Ile Ala Leu Ala Asp Asp Tyr Phe Ile Ser Arg Lys Leu Tyr
 355 360 365

Pro Asn Val Asp Phe Tyr Thr Gly Leu Ile Tyr Arg Ala Met Gly Phe
 370 375 380

Pro Thr Asp Phe Phe Thr Val Leu Phe Ala Ile Gly Arg Leu Pro Gly
 385 390 395 400

Trp Ile Ala His Tyr Arg Glu Gln Leu Gly Ala Ala Gly Asn Lys Ile
 405 410 415

Asn Arg Pro Arg Gln Val Tyr Thr Gly Asn Glu Ser Arg Lys Leu Val
 420 425 430

Pro Arg Glu Glu Arg
 435

<210> SEQ ID NO 7
 <211> LENGTH: 2814
 <212> TYPE: DNA
 <213> ORGANISM: Corynebacterium glutamicum
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (1)..(750)
 <223> OTHER INFORMATION: sequence upstream of the coding region
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (751)..(2061)
 <223> OTHER INFORMATION: gltA-Allele
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (763)..(765)
 <220> FEATURE:
 <221> NAME/KEY: mutation
 <222> LOCATION: (764)..(764)
 <223> OTHER INFORMATION: Transversion A > T
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (2062)..(2814)
 <223> OTHER INFORMATION: sequence downstream of the coding region

<400> SEQUENCE: 7

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ggccagggat cgtaaacgat ctgacccaac aactataacc ctgaagctgt cagttcctag 120
cacctagat tcttcacgca gtctcccaaa cgatgaaaaa cgcccaaac tggcgacacc 180
gaactattga aaacgcgggg attagttgac cagccaccaa tttgggggta gctcaaagtt 240
ttgcaaagtt ttcaatttct aggttgtaa tatcccctga ggttgcgta tagggtggcg 300
aattgcatgg gaaaagctac tcggcaccca tccttgtcgc gtgcatcaca aactttgcta 360
aactgtgcac cagtccactt attgtgggat ttttaatgcc ttaaaggcca gcattttcac 420
cctctagcgg ggttgaatgc tggccttgag ggtgcagaac taaatagcag cacatcgcca 480
caattgatct gaggttctatt ggcgtgaccg tggtactga ttacggtggc tgtgggtggt 540

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cgggaatgat gtaaccaacg tgattgtggg ggaattggct ctcacttcgg atatggctaa	600
accgcattta tcggtatagc gtgtaaccg gaccagattg gaaagaat gtgtcgagta	660
acaaaaactg acatgcgctt ggcgcatccc agttggtaag aataaacggg actacttcg	720
taatccggaa gagttttttt ccgaacaaat atg ttt gaa agg gtt atc gtg gct	774
Met Phe Glu Arg Val Ile Val Ala	
1 5	
act gat aac aac aag gct gtc ctg cac tac ccc ggt ggc gag ttc gaa	822
Thr Asp Asn Asn Lys Ala Val Leu His Tyr Pro Gly Gly Glu Phe Glu	
10 15 20	
atg gac atc atc gag gct tct gag ggt aac aac ggt gtt gtc ctg ggc	870
Met Asp Ile Ile Glu Ala Ser Glu Gly Asn Asn Gly Val Val Leu Gly	
25 30 35 40	
aag atg ctg tct gag act gga ctg atc act ttt gac cca ggt tat gtg	918
Lys Met Leu Ser Glu Thr Gly Leu Ile Thr Phe Asp Pro Gly Tyr Val	
45 50 55	
agc act ggc tcc acc gag tgc aag atc acc tac atc gat ggc gat gcg	966
Ser Thr Gly Ser Thr Glu Ser Lys Ile Thr Tyr Ile Asp Gly Asp Ala	
60 65 70	
gga atc ctg cgt tac cgc ggc tat gac atc gct gat ctg gct gag aat	1014
Gly Ile Leu Arg Tyr Arg Gly Tyr Asp Ile Ala Asp Leu Ala Glu Asn	
75 80 85	
gcc acc ttc aac gag gtt tct tac cta ctt atc aac ggt gag cta cca	1062
Ala Thr Phe Asn Glu Val Ser Tyr Leu Leu Ile Asn Gly Glu Leu Pro	
90 95 100	
acc cca gat gag ctt cac aag ttt aac gac gag att cgc cac cac acc	1110
Thr Pro Asp Glu Leu His Lys Phe Asn Asp Glu Ile Arg His His Thr	
105 110 115 120	
ctt ctg gac gag gac ttc aag tcc cag ttc aac gtg ttc cca cgc gac	1158
Leu Leu Asp Glu Asp Phe Lys Ser Gln Phe Asn Val Phe Pro Arg Asp	
125 130 135	
gct cac cca atg gca acc ttg gct tcc tgc gtt aac att ttg tct acc	1206
Ala His Pro Met Ala Thr Leu Ala Ser Ser Val Asn Ile Leu Ser Thr	
140 145 150	
tac tac cag gac cag ctg aac cca ctg gat gag gca cag ctt gat aag	1254
Tyr Tyr Gln Asp Gln Leu Asn Pro Leu Asp Glu Ala Gln Leu Asp Lys	
155 160 165	
gca acc gtt cgc ctc atg gca aag gtt cca atg ctg gct gcg tac gca	1302
Ala Thr Val Arg Leu Met Ala Lys Val Pro Met Leu Ala Ala Tyr Ala	
170 175 180	
cac cgc gca cgc aag ggt gct cct tac atg tac cca gac aac tcc ctc	1350
His Arg Ala Arg Lys Gly Ala Pro Tyr Met Tyr Pro Asp Asn Ser Leu	
185 190 195 200	
aat gcg cgt gag aac ttc ctg cgc atg atg ttc ggt tac cca acc gag	1398
Asn Ala Arg Glu Asn Phe Leu Arg Met Met Phe Gly Tyr Pro Thr Glu	
205 210 215	
cca tac gag atc gac cca atc atg gtc aag gct ctg gac aag ctg ctc	1446
Pro Tyr Glu Ile Asp Pro Ile Met Val Lys Ala Leu Asp Lys Leu Leu	
220 225 230	
atc ctg cac gct gac cac gag cag aac tgc tcc acc tcc acc gtt cgt	1494
Ile Leu His Ala Asp His Glu Gln Asn Cys Ser Thr Ser Thr Val Arg	
235 240 245	
atg atc ggt tcc gca cag gcc aac atg ttt gtc tcc atc gct ggt ggc	1542
Met Ile Gly Ser Ala Gln Ala Asn Met Phe Val Ser Ile Ala Gly Gly	
250 255 260	
atc aac gct ctg tcc ggc cca ctg cac ggt ggc gca aac cag gct gtt	1590
Ile Asn Ala Leu Ser Gly Pro Leu His Gly Gly Ala Asn Gln Ala Val	
265 270 275 280	
ctg gag atg ctc gaa gac atc aag agc aac cac ggt ggc gac gca acc	1638

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Leu	Glu	Met	Leu	Glu	Asp	Ile	Lys	Ser	Asn	His	Gly	Gly	Asp	Ala	Thr	
				285					290					295		
gag	ttc	atg	aac	aag	gtc	aag	aac	aag	gaa	gac	ggc	gtc	cgc	ctc	atg	1686
Glu	Phe	Met	Asn	Lys	Val	Lys	Asn	Lys	Glu	Asp	Gly	Val	Arg	Leu	Met	
			300					305					310			
ggc	ttc	gga	cac	cgc	ggt	tac	aag	aac	tac	gat	cca	cgt	gca	gca	atc	1734
Gly	Phe	Gly	His	Arg	Val	Tyr	Lys	Asn	Tyr	Asp	Pro	Arg	Ala	Ala	Ile	
		315					320					325				
gtc	aag	gag	acc	gca	cac	gag	atc	ctc	gag	cac	ctc	ggt	ggc	gac	gat	1782
Val	Lys	Glu	Thr	Ala	His	Glu	Ile	Leu	Glu	His	Leu	Gly	Gly	Asp	Asp	
		330				335					340					
ctt	ctg	gat	ctg	gca	atc	aag	ctg	gaa	gaa	att	gca	ctg	gct	gat	gat	1830
Leu	Leu	Asp	Leu	Ala	Ile	Lys	Leu	Glu	Glu	Ile	Ala	Leu	Ala	Asp	Asp	
		345			350					355					360	
tac	ttc	atc	tcc	cgc	aag	ctc	tac	ccg	aac	gta	gac	ttc	tac	acc	ggc	1878
Tyr	Phe	Ile	Ser	Arg	Lys	Leu	Tyr	Pro	Asn	Val	Asp	Phe	Tyr	Thr	Gly	
				365					370					375		
ctg	atc	tac	cgc	gca	atg	ggc	ttc	cca	act	gac	ttc	ttc	acc	gta	ttg	1926
Leu	Ile	Tyr	Arg	Ala	Met	Gly	Phe	Pro	Thr	Asp	Phe	Phe	Thr	Val	Leu	
			380					385					390			
ttc	gca	atc	ggt	cgt	ctg	cca	gga	tgg	atc	gct	cac	tac	cgc	gag	cag	1974
Phe	Ala	Ile	Gly	Arg	Leu	Pro	Gly	Trp	Ile	Ala	His	Tyr	Arg	Glu	Gln	
		395					400					405				
ctc	ggt	gca	gca	ggc	aac	aag	atc	aac	cgc	cca	cgc	cag	gtc	tac	acc	2022
Leu	Gly	Ala	Ala	Gly	Asn	Lys	Ile	Asn	Arg	Pro	Arg	Gln	Val	Tyr	Thr	
	410					415						420				
ggc	aac	gaa	tcc	cgc	aag	ttg	ggt	cct	cgc	gag	gag	cgc	taaatttagc			2071
Gly	Asn	Glu	Ser	Arg	Lys	Leu	Val	Pro	Arg	Glu	Glu	Arg				
	425				430				435							
ggatgattct	cggtcaactt	cgccgaagc	cacttcgtct	gtcataatga	cagggatggt											2131
ttcggcogtt	tttgcataaa	acccaaaaat	acgattttca	aggagcatgt	acagcacatg											2191
gaaaagccac	agattgagct	accggtcggc	ccagcaccgg	aagatctcgt	aatctctgac											2251
atcategttg	gcaaggagc	agaagcccgc	ccaggtggag	aagttgaggt	ccactatgtg											2311
ggcggtgact	ttgaaaccgg	cgaggagttt	gactcttctc	gggatcgtgg	acagaccagc											2371
cagttcccac	tcaacggcct	cattgcaggt	tggcaagagg	gaattccagg	catgaaggtc											2431
ggcggaacg	gtcagctgac	cattccgcca	gaggctgctt	acggccctga	gggttccggc											2491
caccactgt	ctggccgtac	cctggtgttc	atcatcgatt	tgatcagcgc	ataattttct											2551
ttactgcgct	aaacgctcaa	atcgtgtgaa	gagactgtcg	cgtcccgcgc	tctccggatt											2611
gttatccaat	tggagagagg	cggttctgat	tgtgccgaga	atttctcaa	caaagtgtc											2671
ggtttcggcg	acgatcccgt	cgataagccc	ttggcttaaa	agtgcggtgc	cctgcacgcc											2731
ttgtcgtct	atgatttccg	cgccgtggtt	ggtgtcgcgg	aagaggatgg	ccgagcgccc											2791
ctctggtggc	aatgcccaga	gcc														2814

<210> SEQ ID NO 8
 <211> LENGTH: 437
 <212> TYPE: PRT
 <213> ORGANISM: Corynebacterium glutamicum

<400> SEQUENCE: 8

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His	Tyr	Pro	Gly	Gly	Glu	Phe	Glu	Met	Asp	Ile	Ile	Glu	Ala	Ser	Glu
			20					25					30		
Gly	Asn	Asn	Gly	Val	Val	Leu	Gly	Lys	Met	Leu	Ser	Glu	Thr	Gly	Leu

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35					40					45					
Ile	Thr	Phe	Asp	Pro	Gly	Tyr	Val	Ser	Thr	Gly	Ser	Thr	Glu	Ser	Lys
50					55					60					
Ile	Thr	Tyr	Ile	Asp	Gly	Asp	Ala	Gly	Ile	Leu	Arg	Tyr	Arg	Gly	Tyr
65					70					75					80
Asp	Ile	Ala	Asp	Leu	Ala	Glu	Asn	Ala	Thr	Phe	Asn	Glu	Val	Ser	Tyr
				85					90					95	
Leu	Leu	Ile	Asn	Gly	Glu	Leu	Pro	Thr	Pro	Asp	Glu	Leu	His	Lys	Phe
			100					105					110		
Asn	Asp	Glu	Ile	Arg	His	His	Thr	Leu	Leu	Asp	Glu	Asp	Phe	Lys	Ser
		115					120					125			
Gln	Phe	Asn	Val	Phe	Pro	Arg	Asp	Ala	His	Pro	Met	Ala	Thr	Leu	Ala
		130					135					140			
Ser	Ser	Val	Asn	Ile	Leu	Ser	Thr	Tyr	Tyr	Gln	Asp	Gln	Leu	Asn	Pro
145				150						155				160	
Leu	Asp	Glu	Ala	Gln	Leu	Asp	Lys	Ala	Thr	Val	Arg	Leu	Met	Ala	Lys
				165					170					175	
Val	Pro	Met	Leu	Ala	Ala	Tyr	Ala	His	Arg	Ala	Arg	Lys	Gly	Ala	Pro
			180					185					190		
Tyr	Met	Tyr	Pro	Asp	Asn	Ser	Leu	Asn	Ala	Arg	Glu	Asn	Phe	Leu	Arg
		195					200					205			
Met	Met	Phe	Gly	Tyr	Pro	Thr	Glu	Pro	Tyr	Glu	Ile	Asp	Pro	Ile	Met
		210					215					220			
Val	Lys	Ala	Leu	Asp	Lys	Leu	Leu	Ile	Leu	His	Ala	Asp	His	Glu	Gln
225				230						235				240	
Asn	Cys	Ser	Thr	Ser	Thr	Val	Arg	Met	Ile	Gly	Ser	Ala	Gln	Ala	Asn
				245					250					255	
Met	Phe	Val	Ser	Ile	Ala	Gly	Gly	Ile	Asn	Ala	Leu	Ser	Gly	Pro	Leu
			260					265					270		
His	Gly	Gly	Ala	Asn	Gln	Ala	Val	Leu	Glu	Met	Leu	Glu	Asp	Ile	Lys
		275					280					285			
Ser	Asn	His	Gly	Gly	Asp	Ala	Thr	Glu	Phe	Met	Asn	Lys	Val	Lys	Asn
		290					295					300			
Lys	Glu	Asp	Gly	Val	Arg	Leu	Met	Gly	Phe	Gly	His	Arg	Val	Tyr	Lys
305				310						315				320	
Asn	Tyr	Asp	Pro	Arg	Ala	Ala	Ile	Val	Lys	Glu	Thr	Ala	His	Glu	Ile
				325					330					335	
Leu	Glu	His	Leu	Gly	Gly	Asp	Asp	Leu	Leu	Asp	Leu	Ala	Ile	Lys	Leu
			340					345					350		
Glu	Glu	Ile	Ala	Leu	Ala	Asp	Asp	Tyr	Phe	Ile	Ser	Arg	Lys	Leu	Tyr
		355					360					365			
Pro	Asn	Val	Asp	Phe	Tyr	Thr	Gly	Leu	Ile	Tyr	Arg	Ala	Met	Gly	Phe
		370					375					380			
Pro	Thr	Asp	Phe	Phe	Thr	Val	Leu	Phe	Ala	Ile	Gly	Arg	Leu	Pro	Gly
385				390								395		400	
Trp	Ile	Ala	His	Tyr	Arg	Glu	Gln	Leu	Gly	Ala	Ala	Gly	Asn	Lys	Ile
			405						410					415	
Asn	Arg	Pro	Arg	Gln	Val	Tyr	Thr	Gly	Asn	Glu	Ser	Arg	Lys	Leu	Val
			420					425					430		
Pro	Arg	Glu	Glu	Arg											
		435													

<210> SEQ ID NO 9
 <211> LENGTH: 2814

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<212> TYPE: DNA
<213> ORGANISM: Corynebacterium glutamicum
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(750)
<223> OTHER INFORMATION: sequence upstream of the coding region
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (751)..(2061)
<223> OTHER INFORMATION: gltA-Allele
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (763)..(765)
<223> OTHER INFORMATION: n is a, c, g, or t
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (2062)..(2814)
<223> OTHER INFORMATION: sequence downstream of the coding region

<400> SEQUENCE: 9

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cacctagat tcttcacgca gtctcccaaa cgatgaaaaa cgcccaaac tggcgacacc      180
gaactattga aaacgcgggg attagttgac cagccaccaa tttgggggta gctcaaagtt      240
ttgcaaagtt ttcaatttct aggttgtaa tatcccctga ggttgcgcta tagggtggcg      300
aattgcatgg gaaaagctac tcggcaccca tccttgcgcg gtgcatacaca aactttgcta      360
aactgtgcac cagtccactt attgtgggat ttttaatgcc ttaaaggcca gcattttcac      420
cctctagcgg ggttgaatgc tggccttgag ggtgcagaac taaatagcag cacatcggca      480
caattgatct gagttctatt ggcgtgaccg tggctactga ttacggtggc tgtgggtggg      540
cgggaatgat gtaaccaacg tgattgtggg ggaattggct ctcaactcgg atatggctaa      600
accgcattta tcggtatagc gtgttaaccg gaccagattg ggaaagaaat gtgtcgagta      660
acaaaaactg acatgcgcctt ggcgcacccc agttggtaag aataaacggg actacttccg      720
taatccggaa gagttttttt ccgaacaaat atg ttt gaa agg nnn atc gtg gct      774
                               Met Phe Glu Arg Xaa Ile Val Ala
                               1                               5

act gat aac aac aag gct gtc ctg cac tac ccc ggt ggc gag ttc gaa      822
Thr Asp Asn Asn Lys Ala Val Leu His Tyr Pro Gly Gly Glu Phe Glu
    10                               15                               20

atg gac atc atc gag gct tct gag ggt aac aac ggt gtt gtc ctg ggc      870
Met Asp Ile Ile Glu Ala Ser Glu Gly Asn Asn Gly Val Val Leu Gly
    25                               30                               35                               40

aag atg ctg tct gag act gga ctg atc act ttt gac cca ggt tat gtg      918
Lys Met Leu Ser Glu Thr Gly Leu Ile Thr Phe Asp Pro Gly Tyr Val
    45                               50                               55

agc act ggc tcc acc gag tgc aag atc acc tac atc gat ggc gat gcg      966
Ser Thr Gly Ser Thr Glu Ser Lys Ile Thr Tyr Ile Asp Gly Asp Ala
    60                               65                               70

gga atc ctg cgt tac cgc ggc tat gac atc gct gat ctg gct gag aat      1014
Gly Ile Leu Arg Tyr Arg Gly Tyr Asp Ile Ala Asp Leu Ala Glu Asn
    75                               80                               85

gcc acc ttc aac gag gtt tct tac cta ctt atc aac ggt gag cta cca      1062
Ala Thr Phe Asn Glu Val Ser Tyr Leu Leu Ile Asn Gly Glu Leu Pro
    90                               95                               100

acc cca gat gag ctt cac aag ttt aac gac gag att cgc cac cac acc      1110
Thr Pro Asp Glu Leu His Lys Phe Asn Asp Glu Ile Arg His His Thr
    105                               110                               115                               120

ctt ctg gac gag gac ttc aag tcc cag ttc aac gtg ttc cca cgc gac      1158
Leu Leu Asp Glu Asp Phe Lys Ser Gln Phe Asn Val Phe Pro Arg Asp
    125                               130                               135

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gct cac cca atg gca acc ttg gct tcc tcg gtt aac att ttg tct acc	1206
Ala His Pro Met Ala Thr Leu Ala Ser Ser Val Asn Ile Leu Ser Thr	
140 145 150	
tac tac cag gac cag ctg aac cca ctc gat gag gca cag ctt gat aag	1254
Tyr Tyr Gln Asp Gln Leu Asn Pro Leu Asp Glu Ala Gln Leu Asp Lys	
155 160 165	
gca acc gtt cgc ctc atg gca aag gtt cca atg ctg gct gcg tac gca	1302
Ala Thr Val Arg Leu Met Ala Lys Val Pro Met Leu Ala Ala Tyr Ala	
170 175 180	
cac cgc gca cgc aag ggt gct cct tac atg tac cca gac aac tcc ctc	1350
His Arg Ala Arg Lys Gly Ala Pro Tyr Met Tyr Pro Asp Asn Ser Leu	
185 190 195 200	
aat gcg cgt gag aac ttc ctg cgc atg atg ttc ggt tac cca acc gag	1398
Asn Ala Arg Glu Asn Phe Leu Arg Met Met Phe Gly Tyr Pro Thr Glu	
205 210 215	
cca tac gag atc gac cca atc atg gtc aag gct ctg gac aag ctg ctc	1446
Pro Tyr Glu Ile Asp Pro Ile Met Val Lys Ala Leu Asp Lys Leu Leu	
220 225 230	
atc ctg cac gct gac cac gag cag aac tgc tcc acc tcc acc gtt cgt	1494
Ile Leu His Ala Asp His Glu Gln Asn Cys Ser Thr Ser Thr Val Arg	
235 240 245	
atg atc ggt tcc gca cag gcc aac atg ttt gtc tcc atc gct ggt ggc	1542
Met Ile Gly Ser Ala Gln Ala Asn Met Phe Val Ser Ile Ala Gly Gly	
250 255 260	
atc aac gct ctg tcc ggc cca ctg cac ggt ggc gca aac cag gct gtt	1590
Ile Asn Ala Leu Ser Gly Pro Leu His Gly Gly Ala Asn Gln Ala Val	
265 270 275 280	
ctg gag atg ctc gaa gac atc aag agc aac cac ggt ggc gac gca acc	1638
Leu Glu Met Leu Glu Asp Ile Lys Ser Asn His Gly Gly Asp Ala Thr	
285 290 295	
gag ttc atg aac aag gtc aag aac aag gaa gac ggc gtc cgc ctc atg	1686
Glu Phe Met Asn Lys Val Lys Asn Lys Glu Asp Gly Val Arg Leu Met	
300 305 310	
ggc ttc gga cac cgc gtt tac aag aac tac gat cca cgt gca gca atc	1734
Gly Phe Gly His Arg Val Tyr Lys Asn Tyr Asp Pro Arg Ala Ala Ile	
315 320 325	
gtc aag gag acc gca cac gag atc ctc gag cac ctc ggt ggc gac gat	1782
Val Lys Glu Thr Ala His Glu Ile Leu Glu His Leu Gly Gly Asp Asp	
330 335 340	
ctt ctg gat ctg gca atc aag ctg gaa gaa att gca ctg gct gat gat	1830
Leu Leu Asp Leu Ala Ile Lys Leu Glu Glu Ile Ala Leu Ala Asp Asp	
345 350 355 360	
tac ttc atc tcc cgc aag ctc tac ccg aac gta gac ttc tac acc ggc	1878
Tyr Phe Ile Ser Arg Lys Leu Tyr Pro Asn Val Asp Phe Tyr Thr Gly	
365 370 375	
ctg atc tac cgc gca atg ggc ttc cca act gac ttc ttc acc gta ttg	1926
Leu Ile Tyr Arg Ala Met Gly Phe Pro Thr Asp Phe Phe Thr Val Leu	
380 385 390	
ttc gca atc ggt cgt ctg cca gga tgg atc gct cac tac cgc gag cag	1974
Phe Ala Ile Gly Arg Leu Pro Gly Trp Ile Ala His Tyr Arg Glu Gln	
395 400 405	
ctc ggt gca gca ggc aac aag atc aac cgc cca cgc cag gtc tac acc	2022
Leu Gly Ala Ala Gly Asn Lys Ile Asn Arg Pro Arg Gln Val Tyr Thr	
410 415 420	
ggc aac gaa tcc cgc aag ttg gtt cct cgc gag gag cgc taaatttagc	2071
Gly Asn Glu Ser Arg Lys Leu Val Pro Arg Glu Glu Arg	
425 430 435	
ggatgattct cgttcaactt cgccgaagc cacttcgtct gtcataatga cagggatggt	2131
ttcggccggt tttgcatgaa accaaaaaat acgattttca aggagcatgt acagcacatg	2191

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gaaaagccac agattgagct accggtcggc ccagcaccgg aagatctcgt aatctctgac 2251
atcatcgttg gcgaaggagc agaagcccgc ccaggtggag aagttgaggt ccactatgtg 2311
ggcggtgact ttgaaaccgg cgaggagttt gactcttctt gggatcgtgg acagaccagc 2371
cagttccacc tcaacggcct cattgcaggt tggcaagagg gaattccagg catgaaggtc 2431
ggcggacgtc gtcagctgac cattccgcca gaggtgctt acggccctga gggttccggc 2491
caccactgt ctggcgtac cctggtgttc atcatcgatt tgatcagcgc ataattttct 2551
ttactgcgct aaacgctcaa atcgtgtgaa gogactgtcg cgtcccggcc tctccggatt 2611
gttatccaat tcggagaggg cgttgctgat tgtgccgaga atttctcaa caaagtgtc 2671
ggtttcggcg acgateccgt cgataagccc ttggttaaa agtgcgtgcg cctgcacgcc 2731
ttgtcgtctc atgatttccg cggcgtggtt ggtgtcgcgg aagaggatgg ccgagcggcc 2791
ctctggtggc aatgcggaca gcc 2814

```

<210> SEQ ID NO 10

<211> LENGTH: 437

<212> TYPE: PRT

<213> ORGANISM: Corynebacterium glutamicum

<220> FEATURE:

<221> NAME/KEY: misc_feature

<222> LOCATION: (5)..(5)

<223> OTHER INFORMATION: The 'Xaa' at location 5 stands for Lys, Asn, Arg, Ser, Thr, Ile, Met, Glu, Asp, Gly, Ala, Val, Gln, His, Pro, Leu, Tyr, Trp, Cys, or Phe.

<400> SEQUENCE: 10

```

Met Phe Glu Arg Xaa Ile Val Ala Thr Asp Asn Asn Lys Ala Val Leu
1           5           10           15
His Tyr Pro Gly Gly Glu Phe Glu Met Asp Ile Ile Glu Ala Ser Glu
20          25          30
Gly Asn Asn Gly Val Val Leu Gly Lys Met Leu Ser Glu Thr Gly Leu
35          40          45
Ile Thr Phe Asp Pro Gly Tyr Val Ser Thr Gly Ser Thr Glu Ser Lys
50          55          60
Ile Thr Tyr Ile Asp Gly Asp Ala Gly Ile Leu Arg Tyr Arg Gly Tyr
65          70          75          80
Asp Ile Ala Asp Leu Ala Glu Asn Ala Thr Phe Asn Glu Val Ser Tyr
85          90          95
Leu Leu Ile Asn Gly Glu Leu Pro Thr Pro Asp Glu Leu His Lys Phe
100         105        110
Asn Asp Glu Ile Arg His His Thr Leu Leu Asp Glu Asp Phe Lys Ser
115        120        125
Gln Phe Asn Val Phe Pro Arg Asp Ala His Pro Met Ala Thr Leu Ala
130        135        140
Ser Ser Val Asn Ile Leu Ser Thr Tyr Tyr Gln Asp Gln Leu Asn Pro
145        150        155        160
Leu Asp Glu Ala Gln Leu Asp Lys Ala Thr Val Arg Leu Met Ala Lys
165        170        175
Val Pro Met Leu Ala Ala Tyr Ala His Arg Ala Arg Lys Gly Ala Pro
180        185        190
Tyr Met Tyr Pro Asp Asn Ser Leu Asn Ala Arg Glu Asn Phe Leu Arg
195        200        205
Met Met Phe Gly Tyr Pro Thr Glu Pro Tyr Glu Ile Asp Pro Ile Met
210        215        220
Val Lys Ala Leu Asp Lys Leu Leu Ile Leu His Ala Asp His Glu Gln

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225          230          235          240
Asn Cys Ser Thr Ser Thr Val Arg Met Ile Gly Ser Ala Gln Ala Asn
      245          250          255
Met Phe Val Ser Ile Ala Gly Gly Ile Asn Ala Leu Ser Gly Pro Leu
      260          265          270
His Gly Gly Ala Asn Gln Ala Val Leu Glu Met Leu Glu Asp Ile Lys
      275          280          285
Ser Asn His Gly Gly Asp Ala Thr Glu Phe Met Asn Lys Val Lys Asn
      290          295          300
Lys Glu Asp Gly Val Arg Leu Met Gly Phe Gly His Arg Val Tyr Lys
305          310          315          320
Asn Tyr Asp Pro Arg Ala Ala Ile Val Lys Glu Thr Ala His Glu Ile
      325          330          335
Leu Glu His Leu Gly Gly Asp Asp Leu Leu Asp Leu Ala Ile Lys Leu
      340          345          350
Glu Glu Ile Ala Leu Ala Asp Asp Tyr Phe Ile Ser Arg Lys Leu Tyr
      355          360          365
Pro Asn Val Asp Phe Tyr Thr Gly Leu Ile Tyr Arg Ala Met Gly Phe
      370          375          380
Pro Thr Asp Phe Phe Thr Val Leu Phe Ala Ile Gly Arg Leu Pro Gly
385          390          395          400
Trp Ile Ala His Tyr Arg Glu Gln Leu Gly Ala Ala Gly Asn Lys Ile
      405          410          415
Asn Arg Pro Arg Gln Val Tyr Thr Gly Asn Glu Ser Arg Lys Leu Val
      420          425          430
Pro Arg Glu Glu Arg
      435

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<210> SEQ ID NO 11
<211> LENGTH: 105
<212> TYPE: DNA
<213> ORGANISM: Corynebacterium glutamicum
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (40)..(105)
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (52)..(54)
<223> OTHER INFORMATION: n is a, c, g, or t
<400> SEQUENCE: 11

```

```

ctacttccgt aatccggaag agtttttttc cgaacaaat atg ttt gaa agg nnn      54
              Met Phe Glu Arg Xaa
              1              5
atc gtg gct act gat aac aac aag gct gtc ctg cac tac ccc ggt ggc      102
Ile Val Ala Thr Asp Asn Asn Lys Ala Val Leu His Tyr Pro Gly Gly
              10              15              20
gag
Glu

```

```

<210> SEQ ID NO 12
<211> LENGTH: 22
<212> TYPE: PRT
<213> ORGANISM: Corynebacterium glutamicum
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (5)..(5)
<223> OTHER INFORMATION: The 'Xaa' at location 5 stands for Lys, Asn,
Arg, Ser, Thr, Ile, Met, Glu, Asp, Gly, Ala, Val, Gln, His, Pro,
Leu, Tyr, Trp, Cys, or Phe.
<400> SEQUENCE: 12

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-continued

Met Phe Glu Arg Xaa Ile Val Ala Thr Asp Asn Asn Lys Ala Val Leu
 1 5 10 15

His Tyr Pro Gly Gly Glu
 20

<210> SEQ ID NO 13
 <211> LENGTH: 206
 <212> TYPE: DNA
 <213> ORGANISM: Corynebacterium glutamicum
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (90)..(206)
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (102)..(104)
 <223> OTHER INFORMATION: n is a, c, g, or t

<400> SEQUENCE: 13

caaaaactga catgcgcttg gcgcatccca gttgtaaga ataaacggga ctacttccgt 60
 aatccggaag agtttttttc cgaacaaat atg ttt gaa agg nnn atc gtg gct 113
 Met Phe Glu Arg Xaa Ile Val Ala
 1 5
 act gat aac aac aag gct gtc ctg cac tac ccc ggt ggc gag ttc gaa 161
 Thr Asp Asn Asn Lys Ala Val Leu His Tyr Pro Gly Gly Glu Phe Glu
 10 15 20
 atg gac atc atc gag gct tct gag ggt aac aac ggt gtt gtc ctg 206
 Met Asp Ile Ile Glu Ala Ser Glu Gly Asn Asn Gly Val Val Leu
 25 30 35

<210> SEQ ID NO 14
 <211> LENGTH: 39
 <212> TYPE: PRT
 <213> ORGANISM: Corynebacterium glutamicum
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (5)..(5)
 <223> OTHER INFORMATION: The 'Xaa' at location 5 stands for Lys, Asn,
 Arg, Ser, Thr, Ile, Met, Glu, Asp, Gly, Ala, Val, Gln, His, Pro,
 Leu, Tyr, Trp, Cys, or Phe.

<400> SEQUENCE: 14

Met Phe Glu Arg Xaa Ile Val Ala Thr Asp Asn Asn Lys Ala Val Leu
 1 5 10 15

His Tyr Pro Gly Gly Glu Phe Glu Met Asp Ile Ile Glu Ala Ser Glu
 20 25 30

Gly Asn Asn Gly Val Val Leu
 35

<210> SEQ ID NO 15
 <211> LENGTH: 405
 <212> TYPE: DNA
 <213> ORGANISM: Corynebacterium glutamicum
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (190)..(405)
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (202)..(204)
 <223> OTHER INFORMATION: n is a, c, g, or t

<400> SEQUENCE: 15

gattgtgggg gaattggctc tcaactcgga tatggctaaa ccgcatttat cggtatagcg 60
 tgtaaccgg accagattgg gaaagaaatg tgctcgagtaa caaaaactga catgcgcttg 120
 gcgcatccca gttgtaaga ataaacggga ctacttccgt aatccggaag agtttttttc 180

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cgaacaaat atg ttt gaa agg nnn atc gtg gct act gat aac aac aag gct 231
      Met Phe Glu Arg Xaa Ile Val Ala Thr Asp Asn Asn Lys Ala
      1             5             10

gtc ctg cac tac ccc ggt ggc gag ttc gaa atg gac atc atc gag gct 279
Val Leu His Tyr Pro Gly Gly Glu Phe Glu Met Asp Ile Ile Glu Ala
15             20             25             30

tct gag ggt aac aac ggt gtt gtc ctg ggc aag atg ctg tct gag act 327
Ser Glu Gly Asn Asn Gly Val Val Leu Gly Lys Met Leu Ser Glu Thr
      35             40             45

gga ctg atc act ttt gac cca ggt tat gtg agc act ggc tcc acc gag 375
Gly Leu Ile Thr Phe Asp Pro Gly Tyr Val Ser Thr Gly Ser Thr Glu
      50             55             60

tcg aag atc acc tac atc gat ggc gat gcg 405
Ser Lys Ile Thr Tyr Ile Asp Gly Asp Ala
      65             70

```

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<210> SEQ ID NO 16
<211> LENGTH: 72
<212> TYPE: PRT
<213> ORGANISM: Corynebacterium glutamicum
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (5)..(5)
<223> OTHER INFORMATION: The 'Xaa' at location 5 stands for Lys, Asn,
      Arg, Ser, Thr, Ile, Met, Glu, Asp, Gly, Ala, Val, Gln, His, Pro,
      Leu, Tyr, Trp, Cys, or Phe.

```

```

<400> SEQUENCE: 16

Met Phe Glu Arg Xaa Ile Val Ala Thr Asp Asn Asn Lys Ala Val Leu
1             5             10             15

His Tyr Pro Gly Gly Glu Phe Glu Met Asp Ile Ile Glu Ala Ser Glu
      20             25             30

Gly Asn Asn Gly Val Val Leu Gly Lys Met Leu Ser Glu Thr Gly Leu
      35             40             45

Ile Thr Phe Asp Pro Gly Tyr Val Ser Thr Gly Ser Thr Glu Ser Lys
      50             55             60

Ile Thr Tyr Ile Asp Gly Asp Ala
65             70

```

```

<210> SEQ ID NO 17
<211> LENGTH: 1001
<212> TYPE: DNA
<213> ORGANISM: Corynebacterium glutamicum
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (489)..(1001)
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (501)..(503)
<223> OTHER INFORMATION: n is a, c, g, or t

```

```

<400> SEQUENCE: 17

gttgtaata tcccctgagg ttgcgttata ggggtggcgaa ttgcatgggg aaagctactc 60
ggcaccatc cttgtgcgct gcatcacaaa ctttgctaaa ctgtgcacca gtccacttat 120
tgtgggattt ttaatgcctt aaaggccagc attttcaccc tctagcgggg ttgaatgctg 180
gccttgaggg tgcagaacta aatagcagca catcggcaca attgatctga gttctattgg 240
cgtgaccgtg gctactgatt acggtggctg tgggtggtcg ggaatgatgt aaccaacgtg 300
attgtggggg aattggctct cacttcggat atggctaaac cgcatttata ggtatagcgt 360
gttaaccgga ccagattggg aaagaaatgt gtcgagtaac aaaaactgac atgcgcttgg 420
cgcatcccag ttggtaagaa taaacgggac tacttccgta atccggaaga gtttttttcc 480

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gaacaaat atg ttt gaa agg nnn atc gtg gct act gat aac aac aag gct 530
      Met Phe Glu Arg Xaa Ile Val Ala Thr Asp Asn Asn Lys Ala
      1           5           10

gtc ctg cac tac ccc ggt ggc gag ttc gaa atg gac atc atc gag gct 578
Val Leu His Tyr Pro Gly Gly Glu Phe Glu Met Asp Ile Ile Glu Ala
15           20           25           30

tct gag ggt aac aac ggt gtt gtc ctg ggc aag atg ctg tct gag act 626
Ser Glu Gly Asn Asn Gly Val Val Leu Gly Lys Met Leu Ser Glu Thr
      35           40           45

gga ctg atc act ttt gac cca ggt tat gtg agc act ggc tcc acc gag 674
Gly Leu Ile Thr Phe Asp Pro Gly Tyr Val Ser Thr Gly Ser Thr Glu
      50           55           60

tcg aag atc acc tac atc gat ggc gat gcg gga atc ctg cgt tac cgc 722
Ser Lys Ile Thr Tyr Ile Asp Gly Asp Ala Gly Ile Leu Arg Tyr Arg
      65           70           75

ggc tat gac atc gct gat ctg gct gag aat gcc acc ttc aac gag gtt 770
Gly Tyr Asp Ile Ala Asp Leu Ala Glu Asn Ala Thr Phe Asn Glu Val
      80           85           90

tct tac cta ctt atc aac ggt gag cta cca acc cca gat gag ctt cac 818
Ser Tyr Leu Leu Ile Asn Gly Glu Leu Pro Thr Pro Asp Glu Leu His
95           100           105           110

aag ttt aac gac gag att cgc cac cac acc ctt ctg gac gag gac ttc 866
Lys Phe Asn Asp Glu Ile Arg His His Thr Leu Leu Asp Glu Asp Phe
      115           120           125

aag tcc cag ttc aac gtg ttc cca cgc gac gct cac cca atg gca acc 914
Lys Ser Gln Phe Asn Val Phe Pro Arg Asp Ala His Pro Met Ala Thr
      130           135           140

ttg gct tcc tcg gtt aac att ttg tct acc tac tac cag gac cag ctg 962
Leu Ala Ser Ser Val Asn Ile Leu Ser Thr Tyr Tyr Gln Asp Gln Leu
      145           150           155

aac cca ctc gat gag gca cag ctt gat aag gca acc gtt 1001
Asn Pro Leu Asp Glu Ala Gln Leu Asp Lys Ala Thr Val
      160           165           170

```

<210> SEQ ID NO 18

<211> LENGTH: 171

<212> TYPE: PRT

<213> ORGANISM: Corynebacterium glutamicum

<220> FEATURE:

<221> NAME/KEY: misc_feature

<222> LOCATION: (5)..(5)

<223> OTHER INFORMATION: The 'Xaa' at location 5 stands for Lys, Asn, Arg, Ser, Thr, Ile, Met, Glu, Asp, Gly, Ala, Val, Gln, His, Pro, Leu, Tyr, Trp, Cys, or Phe.

<400> SEQUENCE: 18

```

Met Phe Glu Arg Xaa Ile Val Ala Thr Asp Asn Asn Lys Ala Val Leu
1           5           10           15

His Tyr Pro Gly Gly Glu Phe Glu Met Asp Ile Ile Glu Ala Ser Glu
      20           25           30

Gly Asn Asn Gly Val Val Leu Gly Lys Met Leu Ser Glu Thr Gly Leu
      35           40           45

Ile Thr Phe Asp Pro Gly Tyr Val Ser Thr Gly Ser Thr Glu Ser Lys
      50           55           60

Ile Thr Tyr Ile Asp Gly Asp Ala Gly Ile Leu Arg Tyr Arg Gly Tyr
      65           70           75           80

Asp Ile Ala Asp Leu Ala Glu Asn Ala Thr Phe Asn Glu Val Ser Tyr
      85           90           95

Leu Leu Ile Asn Gly Glu Leu Pro Thr Pro Asp Glu Leu His Lys Phe
      100           105           110

Asn Asp Glu Ile Arg His His Thr Leu Leu Asp Glu Asp Phe Lys Ser

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115	120	125	
Gln Phe Asn Val Phe Pro Arg Asp Ala His Pro Met Ala Thr Leu Ala			
130	135	140	
Ser Ser Val Asn Ile Leu Ser Thr Tyr Tyr Gln Asp Gln Leu Asn Pro			
145	150	155	160
Leu Asp Glu Ala Gln Leu Asp Lys Ala Thr Val			
	165	170	
<210> SEQ ID NO 19			
<211> LENGTH: 1263			
<212> TYPE: DNA			
<213> ORGANISM: Corynebacterium glutamicum			
<220> FEATURE:			
<221> NAME/KEY: CDS			
<222> LOCATION: (1)..(1263)			
<223> OTHER INFORMATION: lysC-Wild-Type-Gene			
<400> SEQUENCE: 19			
gtg gcc ctg gtc gta cag aaa tat ggc ggt tcc tcg ctt gag agt gcg			48
Val Ala Leu Val Val Gln Lys Tyr Gly Ser Ser Leu Glu Ser Ala			
1	5	10	15
gaa cgc att aga aac gtc gct gaa cgg atc gtt gcc acc aag aag gct			96
Glu Arg Ile Arg Asn Val Ala Glu Arg Ile Val Ala Thr Lys Lys Ala			
	20	25	30
gga aat gat gtc gtg gtt gtc tgc tcc gca atg gga gac acc acg gat			144
Gly Asn Asp Val Val Val Val Cys Ser Ala Met Gly Asp Thr Thr Asp			
	35	40	45
gaa ctt cta gaa ctt gca gcg gca gtg aat ccc gtt ccg cca gct cgt			192
Glu Leu Leu Glu Leu Ala Ala Val Asn Pro Val Pro Pro Ala Arg			
	50	55	60
gaa atg gat atg ctc ctg act gct ggt gag cgt att tct aac gct ctc			240
Glu Met Asp Met Leu Leu Thr Ala Gly Glu Arg Ile Ser Asn Ala Leu			
	65	70	75
gtc gcc atg gct att gag tcc ctt ggc gca gaa gcc caa tct ttc acg			288
Val Ala Met Ala Ile Glu Ser Leu Gly Ala Glu Ala Gln Ser Phe Thr			
	85	90	95
ggc tct cag gct ggt gtg ctc acc acc cgc cac gga aac gca cgc			336
Gly Ser Gln Ala Gly Val Leu Thr Thr Glu Arg His Gly Asn Ala Arg			
	100	105	110
att gtt gat gtc act cca ggt cgt gtg cgt gaa gca ctc gat gag ggc			384
Ile Val Asp Val Thr Pro Gly Arg Val Arg Glu Ala Leu Asp Glu Gly			
	115	120	125
aag atc tgc att gtt gct ggt ttc cag ggt gtt aat aaa gaa acc cgc			432
Lys Ile Cys Ile Val Ala Gly Phe Gln Gly Val Asn Lys Glu Thr Arg			
	130	135	140
gat gtc acc acg ttg ggt cgt ggt ggt tct gac acc act gca gtt gcg			480
Asp Val Thr Thr Leu Gly Arg Gly Gly Ser Asp Thr Thr Ala Val Ala			
	145	150	155
ttg gca gct gct ttg aac gct gat gtg tgt gag att tac tcg gac gtt			528
Leu Ala Ala Ala Leu Asn Ala Asp Val Cys Glu Ile Tyr Ser Asp Val			
	165	170	175
gac ggt gtg tat acc gct gac ccg cgc atc gtt cct aat gca cag aag			576
Asp Gly Val Tyr Thr Ala Asp Pro Arg Ile Val Pro Asn Ala Gln Lys			
	180	185	190
ctg gaa aag ctc agc ttc gaa gaa atg ctg gaa ctt gct gct gtt ggc			624
Leu Glu Lys Leu Ser Phe Glu Glu Met Leu Glu Leu Ala Ala Val Gly			
	195	200	205
tcc aag att ttg gtg ctg cgc agt gtt gaa tac gct cgt gca ttc aat			672
Ser Lys Ile Leu Val Leu Arg Ser Val Glu Tyr Ala Arg Ala Phe Asn			
	210	215	220
gtg cca ctt cgc gta cgc tcg tct tat agt aat gat ccc gcc act ttg			720

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Val	Pro	Leu	Arg	Val	Arg	Ser	Ser	Tyr	Ser	Asn	Asp	Pro	Gly	Thr	Leu	
225					230					235					240	
att gcc ggc tct atg gag gat att cct gtg gaa gaa gca gtc ctt acc																768
Ile Ala Gly Ser Met Glu Asp Ile Pro Val Glu Glu Ala Val Leu Thr				245				250					255			
ggt gtc gca acc gac aag tcc gaa gcc aaa gta acc gtt ctg ggt att																816
Gly Val Ala Thr Asp Lys Ser Glu Ala Lys Val Thr Val Leu Gly Ile			260				265						270			
tcc gat aag cca ggc gag gct gcg aag gtt ttc cgt gcg ttg gct gat																864
Ser Asp Lys Pro Gly Glu Ala Ala Lys Val Phe Arg Ala Leu Ala Asp		275				280						285				
gca gaa atc aac att gac atg gtt ctg cag aac gtc tct tct gta gaa																912
Ala Glu Ile Asn Ile Asp Met Val Leu Gln Asn Val Ser Ser Val Glu		290				295				300						
gac ggc acc acc gac atc acc ttc acc tgc cct cgt tcc gac ggc cgc																960
Asp Gly Thr Thr Asp Ile Thr Phe Thr Cys Pro Arg Ser Asp Gly Arg		305			310				315						320	
cgc gcg atg gag atc ttg aag aag ctt cag gtt cag ggc aac tgg acc																1008
Arg Ala Met Glu Ile Leu Lys Lys Leu Gln Val Gln Gly Asn Trp Thr			325					330						335		
aat gtg ctt tac gac gac cag gtc ggc aaa gtc tcc ctc gtg ggt gct																1056
Asn Val Leu Tyr Asp Asp Gln Val Gly Lys Val Ser Leu Val Gly Ala			340				345							350		
ggc atg aag tct cac cca ggt gtt acc gca gag ttc atg gaa gct ctg																1104
Gly Met Lys Ser His Pro Gly Val Thr Ala Glu Phe Met Glu Ala Leu		355				360						365				
cgc gat gtc aac gtg aac atc gaa ttg att tcc acc tct gag att cgt																1152
Arg Asp Val Asn Val Asn Ile Glu Leu Ile Ser Thr Ser Glu Ile Arg		370				375						380				
att tcc gtg ctg atc cgt gaa gat gat ctg gat gct gct gca cgt gca																1200
Ile Ser Val Leu Ile Arg Glu Asp Asp Leu Asp Ala Ala Ala Arg Ala		385			390				395						400	
ttg cat gag cag ttc cag ctg ggc ggc gaa gac gaa gcc gtc gtt tat																1248
Leu His Glu Gln Phe Gln Leu Gly Gly Glu Asp Glu Ala Val Val Tyr			405					410						415		
gca ggc acc gga cgc																1263
Ala Gly Thr Gly Arg			420													

<210> SEQ ID NO 20

<211> LENGTH: 421

<212> TYPE: PRT

<213> ORGANISM: Corynebacterium glutamicum

<400> SEQUENCE: 20

Val	Ala	Leu	Val	Val	Gln	Lys	Tyr	Gly	Gly	Ser	Ser	Leu	Glu	Ser	Ala	
1				5					10					15		
Glu	Arg	Ile	Arg	Asn	Val	Ala	Glu	Arg	Ile	Val	Ala	Thr	Lys	Lys	Ala	
			20					25					30			
Gly	Asn	Asp	Val	Val	Val	Val	Cys	Ser	Ala	Met	Gly	Asp	Thr	Thr	Asp	
		35					40					45				
Glu	Leu	Leu	Glu	Leu	Ala	Ala	Ala	Val	Asn	Pro	Val	Pro	Pro	Ala	Arg	
		50				55					60					
Glu	Met	Asp	Met	Leu	Leu	Thr	Ala	Gly	Glu	Arg	Ile	Ser	Asn	Ala	Leu	
		65			70					75					80	
Val	Ala	Met	Ala	Ile	Glu	Ser	Leu	Gly	Ala	Glu	Ala	Gln	Ser	Phe	Thr	
				85					90					95		
Gly	Ser	Gln	Ala	Gly	Val	Leu	Thr	Thr	Glu	Arg	His	Gly	Asn	Ala	Arg	
			100					105						110		

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Ile Val Asp Val Thr Pro Gly Arg Val Arg Glu Ala Leu Asp Glu Gly
    115                                120                                125

Lys Ile Cys Ile Val Ala Gly Phe Gln Gly Val Asn Lys Glu Thr Arg
    130                                135                                140

Asp Val Thr Thr Leu Gly Arg Gly Gly Ser Asp Thr Thr Ala Val Ala
    145                                150                                155                                160

Leu Ala Ala Ala Leu Asn Ala Asp Val Cys Glu Ile Tyr Ser Asp Val
    165                                170                                175

Asp Gly Val Tyr Thr Ala Asp Pro Arg Ile Val Pro Asn Ala Gln Lys
    180                                185                                190

Leu Glu Lys Leu Ser Phe Glu Glu Met Leu Glu Leu Ala Ala Val Gly
    195                                200                                205

Ser Lys Ile Leu Val Leu Arg Ser Val Glu Tyr Ala Arg Ala Phe Asn
    210                                215                                220

Val Pro Leu Arg Val Arg Ser Ser Tyr Ser Asn Asp Pro Gly Thr Leu
    225                                230                                235                                240

Ile Ala Gly Ser Met Glu Asp Ile Pro Val Glu Glu Ala Val Leu Thr
    245                                250                                255

Gly Val Ala Thr Asp Lys Ser Glu Ala Lys Val Thr Val Leu Gly Ile
    260                                265                                270

Ser Asp Lys Pro Gly Glu Ala Ala Lys Val Phe Arg Ala Leu Ala Asp
    275                                280                                285

Ala Glu Ile Asn Ile Asp Met Val Leu Gln Asn Val Ser Ser Val Glu
    290                                295                                300

Asp Gly Thr Thr Asp Ile Thr Phe Thr Cys Pro Arg Ser Asp Gly Arg
    305                                310                                315                                320

Arg Ala Met Glu Ile Leu Lys Lys Leu Gln Val Gln Gly Asn Trp Thr
    325                                330                                335

Asn Val Leu Tyr Asp Asp Gln Val Gly Lys Val Ser Leu Val Gly Ala
    340                                345                                350

Gly Met Lys Ser His Pro Gly Val Thr Ala Glu Phe Met Glu Ala Leu
    355                                360                                365

Arg Asp Val Asn Val Asn Ile Glu Leu Ile Ser Thr Ser Glu Ile Arg
    370                                375                                380

Ile Ser Val Leu Ile Arg Glu Asp Asp Leu Asp Ala Ala Ala Arg Ala
    385                                390                                395                                400

Leu His Glu Gln Phe Gln Leu Gly Gly Glu Asp Glu Ala Val Val Tyr
    405                                410                                415

Ala Gly Thr Gly Arg
    420

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<210> SEQ ID NO 21
<211> LENGTH: 2266
<212> TYPE: DNA
<213> ORGANISM: Corynebacterium glutamicum
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(500)
<223> OTHER INFORMATION: sequence upstream of the coding region
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (501)..(1763)
<223> OTHER INFORMATION: coding region of lysC-Wild-Type-Gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1764)..(2266)
<223> OTHER INFORMATION: sequence downstream of the coding region

<400> SEQUENCE: 21

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cgacaggaca agcactgggt gcaactaccaa gaggggtgccg aaaccaagtg ctactgtttg	60
taagaaatat gccagcatcg cgtactcatg cctgcccacc acatcgggtgt catcagagca	120
ttgagtaaag gtgagctcct tagggagcca tcttttgggg tgcggagcgc gatccggtgt	180
ctgaccacgg tgccccatgc gattgttaat gccgatgcta gggcgaaaag cacggcgagc	240
agattgcttt gcaacttgatt cagggtagtt gactaaagag ttgctcgcga agtagcacct	300
gtcacttttg tctcaaatat taaatogaat atcaatatat ggtctgttta ttggaacgcg	360
tcccagtggc tgagacgcat ccgctaaagc cccaggaacc ctgtgcagaa agaaaacact	420
cctctggcta ggtagacaca gtttataaag gttagagttga gcgggtaact gtcagcacgt	480
agatcgaaag gtgcacaaag gtg gcc ctg gtc gta cag aaa tat ggc ggt tcc	533
Val Ala Leu Val Val Gln Lys Tyr Gly Gly Ser	
1 5 10	
tcg ctt gag agt gcg gaa cgc att aga aac gtc gct gaa cgg atc gtt	581
Ser Leu Glu Ser Ala Glu Arg Ile Arg Asn Val Ala Glu Arg Ile Val	
15 20 25	
gcc acc aag aag gct gga aat gat gtc gtg gtt gtc tgc tcc gca atg	629
Ala Thr Lys Lys Ala Gly Asn Asp Val Val Val Val Cys Ser Ala Met	
30 35 40	
gga gac acc acg gat gaa ctt cta gaa ctt gca gcg gca gtg aat ccc	677
Gly Asp Thr Thr Asp Glu Leu Leu Glu Leu Ala Ala Val Asn Pro	
45 50 55	
gtt ccg cca gct cgt gaa atg gat atg ctc ctg act gct ggt gag cgt	725
Val Pro Pro Ala Arg Glu Met Asp Met Leu Leu Thr Ala Gly Glu Arg	
60 65 70 75	
att tct aac gct ctc gtc gcc atg gct att gag tcc ctt gcc gca gaa	773
Ile Ser Asn Ala Leu Val Ala Met Ala Ile Glu Ser Leu Gly Ala Glu	
80 85 90	
gcc caa tct ttc acg ggc tct cag gct ggt gtg ctc acc acc gag cgc	821
Ala Gln Ser Phe Thr Gly Ser Gln Ala Gly Val Leu Thr Thr Glu Arg	
95 100 105	
cac gga aac gca cgc att gtt gat gtc act cca ggt cgt gtg cgt gaa	869
His Gly Asn Ala Arg Ile Val Asp Val Thr Pro Gly Arg Val Arg Glu	
110 115 120	
gca ctc gat gag ggc aag atc tgc att gtt gct ggt ttc cag ggt gtt	917
Ala Leu Asp Glu Gly Lys Ile Cys Ile Val Ala Gly Phe Gln Gly Val	
125 130 135	
aat aaa gaa acc cgc gat gtc acc acg ttg ggt cgt ggt ggt tct gac	965
Asn Lys Glu Thr Arg Asp Val Thr Thr Leu Gly Arg Gly Gly Ser Asp	
140 145 150 155	
acc act gca gtt gcg ttg gca gct gct ttg aac gct gat gtg tgt gag	1013
Thr Thr Ala Val Ala Leu Ala Ala Ala Leu Asn Ala Asp Val Cys Glu	
160 165 170	
att tac tcg gac gtt gac ggt gtg tat acc gct gac ccg cgc atc gtt	1061
Ile Tyr Ser Asp Val Asp Gly Val Tyr Thr Ala Asp Pro Arg Ile Val	
175 180 185	
cct aat gca cag aag ctg gaa aag ctc agc ttc gaa gaa atg ctg gaa	1109
Pro Asn Ala Gln Lys Leu Glu Lys Leu Ser Phe Glu Glu Met Leu Glu	
190 195 200	
ctt gct get gtt ggc tcc aag att ttg gtg ctg cgc agt gtt gaa tac	1157
Leu Ala Ala Val Gly Ser Lys Ile Leu Val Leu Arg Ser Val Glu Tyr	
205 210 215	
gct cgt gca ttc aat gtg cca ctt cgc gta cgc tcg tct tat agt aat	1205
Ala Arg Ala Phe Asn Val Pro Leu Arg Val Arg Ser Ser Tyr Ser Asn	
220 225 230 235	
gat ccc ggc act ttg att gcc ggc tct atg gag gat att cct gtg gaa	1253
Asp Pro Gly Thr Leu Ile Ala Gly Ser Met Glu Asp Ile Pro Val Glu	
240 245 250	

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gaa gca gtc ctt acc ggt gtc gca acc gac aag tcc gaa gcc aaa gta 1301
Glu Ala Val Leu Thr Gly Val Ala Thr Asp Lys Ser Glu Ala Lys Val
      255                      260                      265

acc gtt ctg ggt att tcc gat aag cca ggc gag gct gcg aag gtt ttc 1349
Thr Val Leu Gly Ile Ser Asp Lys Pro Gly Glu Ala Ala Lys Val Phe
      270                      275                      280

cgt gcg ttg gct gat gca gaa atc aac att gac atg gtt ctg cag aac 1397
Arg Ala Leu Ala Asp Ala Glu Ile Asn Ile Asp Met Val Leu Gln Asn
      285                      290                      295

gtc tct tct gta gaa gac ggc acc acc gac atc acc ttc acc tgc cct 1445
Val Ser Ser Val Glu Asp Gly Thr Thr Asp Ile Thr Phe Thr Cys Pro
      300                      305                      310                      315

cgt tcc gac ggc cgc cgc gcg atg gag atc ttg aag aag ctt cag gtt 1493
Arg Ser Asp Gly Arg Arg Ala Met Glu Ile Leu Lys Lys Leu Gln Val
      320                      325                      330

cag ggc aac tgg acc aat gtg ctt tac gac gac cag gtc ggc aaa gtc 1541
Gln Gly Asn Trp Thr Asn Val Leu Tyr Asp Asp Gln Val Gly Lys Val
      335                      340                      345

tcc ctc gtg ggt gct ggc atg aag tct cac cca ggt gtt acc gca gag 1589
Ser Leu Val Gly Ala Gly Met Lys Ser His Pro Gly Val Thr Ala Glu
      350                      355                      360

ttc atg gaa gct ctg cgc gat gtc aac gtg aac atc gaa ttg att tcc 1637
Phe Met Glu Ala Leu Arg Asp Val Asn Val Asn Ile Glu Leu Ile Ser
      365                      370                      375

acc tct gag att cgt att tcc gtg ctg atc cgt gaa gat gat ctg gat 1685
Thr Ser Glu Ile Arg Ile Ser Val Leu Ile Arg Glu Asp Asp Leu Asp
      380                      385                      390                      395

gct gct gca cgt gca ttg cat gag cag ttc cag ctg ggc ggc gaa gac 1733
Ala Ala Ala Arg Ala Leu His Glu Gln Phe Gln Leu Gly Gly Glu Asp
      400                      405                      410

gaa gcc gtc gtt tat gca ggc acc gga cgc taaagtttta aaggagtagt 1783
Glu Ala Val Val Tyr Ala Gly Thr Gly Arg
      415                      420

tttacaatga ccaccatcgc agttgttggt gcaaccggcc aggtcggcca ggttatgcgc 1843

accottttgg aagagcgcaa tttcccagct gacactgttc gtttctttgc ttccccacgt 1903

tccgcaggcc gtaagattga attccgtggc acggaatcg aggtagaaga cattaactcag 1963

gcaaccgagg agtccctcaa ggacatcgac gttgcgttgt tctccgctgg aggcaccgct 2023

tccaagcagt acgctccact gttcgtgct gcaggcgcga ctggttggga taactcttct 2083

gcttgccgca aggaacgacga ggttccacta atcgtctctg aggtgaaccc ttcgacaag 2143

gattccctgg tcaagggcat tattgcgaac cctaactgca ccaccatggc tgcgatgcca 2203

gtgctgaagc cacttcacga tgccgctggt cttgtaaagc ttcacgtttc ctcttaccag 2263

gct 2266

<210> SEQ ID NO 22
<211> LENGTH: 421
<212> TYPE: PRT
<213> ORGANISM: Corynebacterium glutamicum

<400> SEQUENCE: 22
Val Ala Leu Val Val Gln Lys Tyr Gly Gly Ser Ser Leu Glu Ser Ala
1      5      10      15
Glu Arg Ile Arg Asn Val Ala Glu Arg Ile Val Ala Thr Lys Lys Ala
20     25     30
Gly Asn Asp Val Val Val Val Cys Ser Ala Met Gly Asp Thr Thr Asp
35     40     45
Glu Leu Leu Glu Leu Ala Ala Ala Val Asn Pro Val Pro Pro Ala Arg

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50				55				60							
Glu	Met	Asp	Met	Leu	Leu	Thr	Ala	Gly	Glu	Arg	Ile	Ser	Asn	Ala	Leu
65				70						75					80
Val	Ala	Met	Ala	Ile	Glu	Ser	Leu	Gly	Ala	Glu	Ala	Gln	Ser	Phe	Thr
				85					90					95	
Gly	Ser	Gln	Ala	Gly	Val	Leu	Thr	Thr	Glu	Arg	His	Gly	Asn	Ala	Arg
			100					105					110		
Ile	Val	Asp	Val	Thr	Pro	Gly	Arg	Val	Arg	Glu	Ala	Leu	Asp	Glu	Gly
		115				120						125			
Lys	Ile	Cys	Ile	Val	Ala	Gly	Phe	Gln	Gly	Val	Asn	Lys	Glu	Thr	Arg
		130				135					140				
Asp	Val	Thr	Thr	Leu	Gly	Arg	Gly	Gly	Ser	Asp	Thr	Thr	Ala	Val	Ala
				145		150				155					160
Leu	Ala	Ala	Ala	Leu	Asn	Ala	Asp	Val	Cys	Glu	Ile	Tyr	Ser	Asp	Val
				165				170						175	
Asp	Gly	Val	Tyr	Thr	Ala	Asp	Pro	Arg	Ile	Val	Pro	Asn	Ala	Gln	Lys
			180					185						190	
Leu	Glu	Lys	Leu	Ser	Phe	Glu	Glu	Met	Leu	Glu	Leu	Ala	Ala	Val	Gly
		195				200						205			
Ser	Lys	Ile	Leu	Val	Leu	Arg	Ser	Val	Glu	Tyr	Ala	Arg	Ala	Phe	Asn
		210				215					220				
Val	Pro	Leu	Arg	Val	Arg	Ser	Ser	Tyr	Ser	Asn	Asp	Pro	Gly	Thr	Leu
		225			230					235					240
Ile	Ala	Gly	Ser	Met	Glu	Asp	Ile	Pro	Val	Glu	Glu	Ala	Val	Leu	Thr
				245				250						255	
Gly	Val	Ala	Thr	Asp	Lys	Ser	Glu	Ala	Lys	Val	Thr	Val	Leu	Gly	Ile
			260					265						270	
Ser	Asp	Lys	Pro	Gly	Glu	Ala	Ala	Lys	Val	Phe	Arg	Ala	Leu	Ala	Asp
		275				280					285				
Ala	Glu	Ile	Asn	Ile	Asp	Met	Val	Leu	Gln	Asn	Val	Ser	Ser	Val	Glu
		290				295					300				
Asp	Gly	Thr	Thr	Asp	Ile	Thr	Phe	Thr	Cys	Pro	Arg	Ser	Asp	Gly	Arg
		305			310					315					320
Arg	Ala	Met	Glu	Ile	Leu	Lys	Lys	Leu	Gln	Val	Gln	Gly	Asn	Trp	Thr
				325					330					335	
Asn	Val	Leu	Tyr	Asp	Asp	Gln	Val	Gly	Lys	Val	Ser	Leu	Val	Gly	Ala
			340					345					350		
Gly	Met	Lys	Ser	His	Pro	Gly	Val	Thr	Ala	Glu	Phe	Met	Glu	Ala	Leu
		355				360					365				
Arg	Asp	Val	Asn	Val	Asn	Ile	Glu	Leu	Ile	Ser	Thr	Ser	Glu	Ile	Arg
		370				375					380				
Ile	Ser	Val	Leu	Ile	Arg	Glu	Asp	Asp	Leu	Asp	Ala	Ala	Ala	Arg	Ala
		385			390					395					400
Leu	His	Glu	Gln	Phe	Gln	Leu	Gly	Gly	Glu	Asp	Glu	Ala	Val	Val	Tyr
				405					410					415	
Ala	Gly	Thr	Gly	Arg											
			420												

<210> SEQ ID NO 23
 <211> LENGTH: 1503
 <212> TYPE: DNA
 <213> ORGANISM: Corynebacterium glutamicum
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (1)..(1500)
 <223> OTHER INFORMATION: mqo-Wild-Type-Gene

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<400> SEQUENCE: 23

atg tca gat tcc ccg aag aac gca ccg agg att acc gat gag gca gat	48
Met Ser Asp Ser Pro Lys Asn Ala Pro Arg Ile Thr Asp Glu Ala Asp	
1 5 10 15	
gta gtt ctc att ggt gcc ggt atc atg agc tcc acg ctg ggt gca atg	96
Val Val Leu Ile Gly Ala Gly Ile Met Ser Ser Thr Leu Gly Ala Met	
20 25 30	
ctg cgt cag ctg gag cca agc tgg act cag atc gtc ttc gag cgt ttg	144
Leu Arg Gln Leu Glu Pro Ser Trp Thr Gln Ile Val Phe Glu Arg Leu	
35 40 45	
gat gga ccg gca caa gag tcg tcc tcc ccg tgg aac aat gca gga acc	192
Asp Gly Pro Ala Gln Glu Ser Ser Ser Pro Trp Asn Asn Ala Gly Thr	
50 55 60	
ggc cac tct gct cta tgc gag ctg aac tac acc cca gag gtt aag ggc	240
Gly His Ser Ala Leu Cys Glu Leu Asn Tyr Thr Pro Glu Val Lys Gly	
65 70 75 80	
aag gtt gaa att gcc aag gct gta gga atc aac gag aag ttc cag gtt	288
Lys Val Glu Ile Ala Lys Ala Val Gly Ile Asn Glu Lys Phe Gln Val	
85 90 95	
tcc cgt cag ttc tgg tct cac ctc gtt gaa gag gga gtg ctg tct gat	336
Ser Arg Gln Phe Trp Ser His Leu Val Glu Glu Gly Val Leu Ser Asp	
100 105 110	
cct aag gaa ttc atc aac cct gtt cct cac gta tct ttc ggc cag ggc	384
Pro Lys Glu Phe Ile Asn Pro Val Pro His Val Ser Phe Gly Gln Gly	
115 120 125	
gca gat cag gtt gca tac atc aag gct cgc tac gaa gct ttg aag gat	432
Ala Asp Gln Val Ala Tyr Ile Lys Ala Arg Tyr Glu Ala Leu Lys Asp	
130 135 140	
cac cca ctc ttc cag ggc atg acc tac gct gac gat gaa gct acc ttc	480
His Pro Leu Phe Gln Gly Met Thr Tyr Ala Asp Asp Glu Ala Thr Phe	
145 150 155 160	
acc gag aag ctg cct ttg atg gca aag ggc cgt gac ttc tct gat cca	528
Thr Glu Lys Leu Pro Leu Met Ala Lys Gly Arg Asp Phe Ser Asp Pro	
165 170 175	
gta gca atc tct tgg atc gat gaa ggc acc gac atc aac tac ggt gct	576
Val Ala Ile Ser Trp Ile Asp Glu Gly Thr Asp Ile Asn Tyr Gly Ala	
180 185 190	
cag acc aag cag tac ctg gat gca gct gaa gtt gaa ggc act gaa atc	624
Gln Thr Lys Gln Tyr Leu Asp Ala Ala Glu Val Glu Gly Thr Glu Ile	
195 200 205	
cgc tat ggc cac gaa gtc aag agc atc aag gct gat ggc gca aag tgg	672
Arg Tyr Gly His Glu Val Lys Ser Ile Lys Ala Asp Gly Ala Lys Trp	
210 215 220	
atc gtg acc gtc aag aac gta cac act ggc gac acc aag acc atc aag	720
Ile Val Thr Val Lys Asn Val His Thr Gly Asp Thr Lys Thr Ile Lys	
225 230 235 240	
gca aac ttc gtg ttc gtc ggc gca ggc gga tac gca ctg gat ctg ctt	768
Ala Asn Phe Val Phe Val Gly Ala Gly Gly Tyr Ala Leu Asp Leu Leu	
245 250 255	
cgc agc gca ggc atc cca cag gtc aag ggc ttc gct gga ttc cca gta	816
Arg Ser Ala Gly Ile Pro Gln Val Lys Gly Phe Ala Gly Phe Pro Val	
260 265 270	
tcc ggc ctg tgg ctt cgt tgc acc aac gag gaa ctg atc gag cag cac	864
Ser Gly Leu Trp Leu Arg Cys Thr Asn Glu Glu Leu Ile Glu Gln His	
275 280 285	
gca gcc aag gta tat ggc aag gca tct gtt ggc gct cct cca atg tct	912
Ala Ala Lys Val Tyr Gly Lys Ala Ser Val Gly Ala Pro Pro Met Ser	
290 295 300	
gtt cct cac ctt gac acc cgc gtt atc gag ggt gaa aag ggt ctg ctc	960

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Pro Lys Glu Phe Ile Asn Pro Val Pro His Val Ser Phe Gly Gln Gly
 115 120 125
 Ala Asp Gln Val Ala Tyr Ile Lys Ala Arg Tyr Glu Ala Leu Lys Asp
 130 135 140
 His Pro Leu Phe Gln Gly Met Thr Tyr Ala Asp Asp Glu Ala Thr Phe
 145 150 155 160
 Thr Glu Lys Leu Pro Leu Met Ala Lys Gly Arg Asp Phe Ser Asp Pro
 165 170 175
 Val Ala Ile Ser Trp Ile Asp Glu Gly Thr Asp Ile Asn Tyr Gly Ala
 180 185 190
 Gln Thr Lys Gln Tyr Leu Asp Ala Ala Glu Val Glu Gly Thr Glu Ile
 195 200 205
 Arg Tyr Gly His Glu Val Lys Ser Ile Lys Ala Asp Gly Ala Lys Trp
 210 215 220
 Ile Val Thr Val Lys Asn Val His Thr Gly Asp Thr Lys Thr Ile Lys
 225 230 235 240
 Ala Asn Phe Val Phe Val Gly Ala Gly Gly Tyr Ala Leu Asp Leu Leu
 245 250 255
 Arg Ser Ala Gly Ile Pro Gln Val Lys Gly Phe Ala Gly Phe Pro Val
 260 265 270
 Ser Gly Leu Trp Leu Arg Cys Thr Asn Glu Glu Leu Ile Glu Gln His
 275 280 285
 Ala Ala Lys Val Tyr Gly Lys Ala Ser Val Gly Ala Pro Pro Met Ser
 290 295 300
 Val Pro His Leu Asp Thr Arg Val Ile Glu Gly Glu Lys Gly Leu Leu
 305 310 315
 Phe Gly Pro Tyr Gly Gly Trp Thr Pro Lys Phe Leu Lys Glu Gly Ser
 325 330 335
 Tyr Leu Asp Leu Phe Lys Ser Ile Arg Pro Asp Asn Ile Pro Ser Tyr
 340 345 350
 Leu Gly Val Ala Ala Gln Glu Phe Asp Leu Thr Lys Tyr Leu Val Thr
 355 360 365
 Glu Val Leu Lys Asp Gln Asp Lys Arg Met Asp Ala Leu Arg Glu Tyr
 370 375 380
 Met Pro Glu Ala Gln Asn Gly Asp Trp Glu Thr Ile Val Ala Gly Gln
 385 390 395 400
 Arg Val Gln Val Ile Lys Pro Ala Gly Phe Pro Lys Phe Gly Ser Leu
 405 410 415
 Glu Phe Gly Thr Thr Leu Ile Asn Asn Ser Glu Gly Thr Ile Ala Gly
 420 425 430
 Leu Leu Gly Ala Ser Pro Gly Ala Ser Ile Ala Pro Ser Ala Met Ile
 435 440 445
 Glu Leu Leu Glu Arg Cys Phe Gly Asp Arg Met Ile Glu Trp Gly Asp
 450 455 460
 Lys Leu Lys Asp Met Ile Pro Ser Tyr Gly Lys Lys Leu Ala Ser Glu
 465 470 475 480
 Pro Ala Leu Phe Glu Gln Gln Trp Ala Arg Thr Gln Lys Thr Leu Lys
 485 490 495
 Leu Glu Glu Ala
 500

<210> SEQ ID NO 25

<211> LENGTH: 3314

<212> TYPE: DNA

<213> ORGANISM: Corynebacterium glutamicum

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<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(1000)
<223> OTHER INFORMATION: sequence upstream of the coding region
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (1001)..(2311)
<223> OTHER INFORMATION: gltA-Wildtype-Gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1013)..(1015)
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (2312)..(3314)
<223> OTHER INFORMATION: sequence downstream of the coding region

<400> SEQUENCE: 25

atgagtccga aggttgctgc atcccagaat gcggtcgcac cacctagga aaggatgatc      60
tcgtagcctt ctggaagga gaagaggctg gagagtcct cgcggattga acccacgacg      120
tttttactg ccggctgacg gtgtgagta ccgatgacgg atcggatcc gtcgacaata      180
gcctgaatct gttctgctg aacctggaa ggtccgcagc cgaaacggcc gtcgccaggg      240
atgaactcag agggcagggt ggggaagtgc gtcattgtct cgggcaactt tctgcgcttg      300
gaagtaaaag gccacgggat cgttaacgat ctgaccaaac aactataacc ctgaagctgt      360
cagttcctag caccctagat tcttcacgca gtctcccaa cgatgaaaaa cgcccaaac      420
tggcgacacc gaactattga aaacgcgggg attagttgac cagccaccaa tttgggggta      480
gctcaaagt ttgcaaagt ttcaatttct aggttgtaa tatcccctga ggtgctgta      540
tagggtggcg aattgcatgg ggaaagctac tcggcaccca tccttgctgc gtgcatcaca      600
aactttgcta aactgtgcac cagtccactt attgtgggat ttttaatgcc ttaaaggcca      660
gcattttcac cctctagcgg ggttgaatgc tggccttgag ggtgcagaac taaatagcag      720
cacatcggca caattgatct gagttctatt ggcgtgaccg tggctactga ttacggtggc      780
tgtgggtggt cgggaatgat gtaaccaacg tgattgtggg ggaattggct ctcacttcgg      840
atatggctaa accgcattta tcggtatagc gtgttaaccg gaccagattg gaaagaaat      900
gtgtcgagta acaaaaactg acatgcgctt ggcgcacccc agttgtaag aataaacggg      960
actacttccg taatccggaa gagttttttt ccgaacaaat atg ttt gaa agg gat      1015
                                     Met Phe Glu Arg Asp
                                     1           5
atc gtg gct act gat aac aac aag gct gtc ctg cac tac ccc ggt ggc      1063
Ile Val Ala Thr Asp Asn Asn Lys Ala Val Leu His Tyr Pro Gly Gly
                                     10           15           20
gag ttc gaa atg gac atc atc gag gct tct gag ggt aac aac ggt gtt      1111
Glu Phe Glu Met Asp Ile Ile Glu Ala Ser Glu Gly Asn Asn Gly Val
                                     25           30           35
gtc ctg ggc aag atg ctg tct gag act gga ctg atc act ttt gac cca      1159
Val Leu Gly Lys Met Leu Ser Glu Thr Gly Leu Ile Thr Phe Asp Pro
                                     40           45           50
ggg tat gtg agc act ggc tcc acc gag tgc aag atc acc tac atc gat      1207
Gly Tyr Val Ser Thr Gly Ser Thr Glu Ser Lys Ile Thr Tyr Ile Asp
                                     55           60           65
ggc gat gcg gga atc ctg cgt tac cgc ggc tat gac atc gct gat ctg      1255
Gly Asp Ala Gly Ile Leu Arg Tyr Arg Gly Tyr Asp Ile Ala Asp Leu
                                     70           75           80           85
gct gag aat gcc acc ttc aac gag gtt tct tac cta ctt atc aac ggt      1303
Ala Glu Asn Ala Thr Phe Asn Glu Val Ser Tyr Leu Leu Ile Asn Gly
                                     90           95           100
gag cta cca acc cca gat gag ctt cac aag ttt aac gac gag att cgc      1351
Glu Leu Pro Thr Pro Asp Glu Leu His Lys Phe Asn Asp Glu Ile Arg

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105			110			115										
cac	cac	acc	ctt	ctg	gac	gag	gac	ttc	aag	tcc	cag	ttc	aac	gtg	ttc	1399
His	His	Thr	Leu	Leu	Asp	Glu	Asp	Phe	Lys	Ser	Gln	Phe	Asn	Val	Phe	
		120					125					130				
cca	cgc	gac	gct	cac	cca	atg	gca	acc	ttg	gct	tcc	tcg	ggt	aac	att	1447
Pro	Arg	Asp	Ala	His	Pro	Met	Ala	Thr	Leu	Ala	Ser	Ser	Val	Asn	Ile	
	135					140					145					
ttg	tct	acc	tac	tac	cag	gac	cag	ctg	aac	cca	ctc	gat	gag	gca	cag	1495
Leu	Ser	Thr	Tyr	Tyr	Gln	Asp	Gln	Leu	Asn	Pro	Leu	Asp	Glu	Ala	Gln	
	150				155					160					165	
ctt	gat	aag	gca	acc	ggt	cgc	ctc	atg	gca	aag	ggt	cca	atg	ctg	gct	1543
Leu	Asp	Lys	Ala	Thr	Val	Arg	Leu	Met	Ala	Lys	Val	Pro	Met	Leu	Ala	
			170					175						180		
gcg	tac	gca	cac	cgc	gca	cgc	aag	ggt	gct	cct	tac	atg	tac	cca	gac	1591
Ala	Tyr	Ala	His	Arg	Ala	Arg	Lys	Gly	Ala	Pro	Tyr	Met	Tyr	Pro	Asp	
		185						190						195		
aac	tcc	ctc	aat	gcg	cgt	gag	aac	ttc	ctg	cgc	atg	atg	ttc	ggt	tac	1639
Asn	Ser	Leu	Asn	Ala	Arg	Glu	Asn	Phe	Leu	Arg	Met	Met	Phe	Gly	Tyr	
		200					205						210			
cca	acc	gag	cca	tac	gag	atc	gac	cca	atc	atg	gtc	aag	gct	ctg	gac	1687
Pro	Thr	Glu	Pro	Tyr	Glu	Ile	Asp	Pro	Ile	Met	Val	Lys	Ala	Leu	Asp	
		215				220					225					
aag	ctg	ctc	atc	ctg	cac	gct	gac	cac	gag	cag	aac	tgc	tcc	acc	tcc	1735
Lys	Leu	Leu	Ile	Leu	His	Ala	Asp	His	Glu	Gln	Asn	Cys	Ser	Thr	Ser	
	230				235					240					245	
acc	ggt	cgt	atg	atc	ggt	tcc	gca	cag	gcc	aac	atg	ttt	gtc	tcc	atc	1783
Thr	Val	Arg	Met	Ile	Gly	Ser	Ala	Gln	Ala	Asn	Met	Phe	Val	Ser	Ile	
			250					255						260		
gct	ggt	ggc	atc	aac	gct	ctg	tcc	ggc	cca	ctg	cac	ggt	ggc	gca	aac	1831
Ala	Gly	Gly	Ile	Asn	Ala	Leu	Ser	Gly	Pro	Leu	His	Gly	Gly	Ala	Asn	
			265					270						275		
cag	gct	ggt	ctg	gag	atg	ctc	gaa	gac	atc	aag	agc	aac	cac	ggt	ggc	1879
Gln	Ala	Val	Leu	Glu	Met	Leu	Glu	Asp	Ile	Lys	Ser	Asn	His	Gly	Gly	
		280					285					290				
gac	gca	acc	gag	ttc	atg	aac	aag	gtc	aag	aac	aag	gaa	gac	ggc	gtc	1927
Asp	Ala	Thr	Glu	Phe	Met	Asn	Lys	Val	Lys	Asn	Lys	Glu	Asp	Gly	Val	
		295				300					305					
cgc	ctc	atg	ggc	ttc	gga	cac	cgc	ggt	tac	aag	aac	tac	gat	cca	cgt	1975
Arg	Leu	Met	Gly	Phe	Gly	His	Arg	Val	Tyr	Lys	Asn	Tyr	Asp	Pro	Arg	
		310			315					320				325		
gca	gca	atc	gtc	aag	gag	acc	gca	cac	gag	atc	ctc	gag	cac	ctc	ggt	2023
Ala	Ala	Ile	Val	Lys	Glu	Thr	Ala	His	Glu	Ile	Leu	Glu	His	Leu	Gly	
			330						335					340		
ggc	gac	gat	ctt	ctg	gat	ctg	gca	atc	aag	ctg	gaa	gaa	att	gca	ctg	2071
Gly	Asp	Asp	Leu	Leu	Asp	Leu	Ala	Ile	Lys	Leu	Glu	Glu	Ile	Ala	Leu	
			345					350						355		
gct	gat	gat	tac	ttc	atc	tcc	cgc	aag	ctc	tac	ccg	aac	gta	gac	ttc	2119
Ala	Asp	Asp	Tyr	Phe	Ile	Ser	Arg	Lys	Leu	Tyr	Pro	Asn	Val	Asp	Phe	
		360					365					370				
tac	acc	ggc	ctg	atc	tac	cgc	gca	atg	ggc	ttc	cca	act	gac	ttc	ttc	2167
Tyr	Thr	Gly	Leu	Ile	Tyr	Arg	Ala	Met	Gly	Phe	Pro	Thr	Asp	Phe	Phe	
		375				380					385					
acc	gta	ttg	ttc	gca	atc	ggt	cgt	ctg	cca	gga	tgg	atc	gct	cac	tac	2215
Thr	Val	Leu	Phe	Ala	Ile	Gly	Arg	Leu	Pro	Gly	Trp	Ile	Ala	His	Tyr	
		390			395					400				405		
cgc	gag	cag	ctc	ggt	gca	gca	ggc	aac	aag	atc	aac	cgc	cca	cgc	cag	2263
Arg	Glu	Gln	Leu	Gly	Ala	Ala	Gly	Asn	Lys	Ile	Asn	Arg	Pro	Arg	Gln	
			410					415						420		
gtc	tac	acc	ggc	aac	gaa	tcc	cgc	aag	ttg	ggt	cct	cgc	gag	gag	cgc	2311
Val	Tyr	Thr	Gly	Asn	Glu	Ser	Arg	Lys	Leu	Val	Pro	Arg	Glu	Glu	Arg	

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425	430	435	
taaatttagc	ggatgattct	cgttcaactt	cgccgaagc cacttcgtct gtcataatga 2371
cagggatggt	ttcgccggtt	tttgcataaa	acgattttca aggagcatgt 2431
acagcacatg	gaaaagccac	agattgagct	accggtcggt ccagcaccgg aagatctcgt 2491
aatctctgac	atcatcggtg	gcgaaggagc	agaagcccgc ccagggtggag aagttgaggt 2551
ccactatgtg	ggcgttgact	ttgaaaccgg	cgaggagttt gactcttctc gggatcgtgg 2611
acagaccagc	cagttcccac	tcaacggcct	cattgcaggt tggcaagagg gaattccagg 2671
catgaaggtc	ggcggacgtc	gtcagctgac	cattccgcca gaggtgctt acggccctga 2731
gggttccggc	cacccactgt	ctggccgtac	cctgggtgtc atcatcgatt tgatcagcgc 2791
ataattttct	ttactcgcgt	aaacgctcaa	atcgtgtgaa gcgactgtcg cgtcccgcgc 2851
tctccggatt	ggtatccaat	tccgagaggg	cggtgctgat tgtgccgaga atttcttcaa 2911
caaagtgtct	ggtttcggcg	acgatcccgt	cgataagccc ttggcttaaa agtgcgtgcg 2971
cctgcacgcc	ttgtcgtctc	atgatttccg	cgccgtgggt ggtgtcgcgg aagaggatgg 3031
ccgaggcgcc	ctctgggtgc	aatgcggaca	gccacgcggt ttcggccgcg tagaccagat 3091
cgccggggcag	catggccagc	gcgccaccgc	caacgccctg accaataatg accgaaacgg 3151
tggggagggg	agcgtcgata	agcctggaca	aggtgcgcgc aatcgagctt gcgatgccga 3211
gctcctcagc	cgctcgcgac	aattcggcgc	cgagggtgtc gatgatggac acgatcggca 3271
ggtttagctc	gcgcgccagc	gaaatgccac	gacgcgcaaa acg 3314

<210> SEQ ID NO 26

<211> LENGTH: 437

<212> TYPE: PRT

<213> ORGANISM: Corynebacterium glutamicum

<400> SEQUENCE: 26

Met	Phe	Glu	Arg	Asp	Ile	Val	Ala	Thr	Asp	Asn	Asn	Lys	Ala	Val	Leu
1				5					10					15	
His	Tyr	Pro	Gly	Gly	Glu	Phe	Glu	Met	Asp	Ile	Ile	Glu	Ala	Ser	Glu
			20					25					30		
Gly	Asn	Asn	Gly	Val	Val	Leu	Gly	Lys	Met	Leu	Ser	Glu	Thr	Gly	Leu
			35				40					45			
Ile	Thr	Phe	Asp	Pro	Gly	Tyr	Val	Ser	Thr	Gly	Ser	Thr	Glu	Ser	Lys
			50				55					60			
Ile	Thr	Tyr	Ile	Asp	Gly	Asp	Ala	Gly	Ile	Leu	Arg	Tyr	Arg	Gly	Tyr
			65			70				75				80	
Asp	Ile	Ala	Asp	Leu	Ala	Glu	Asn	Ala	Thr	Phe	Asn	Glu	Val	Ser	Tyr
			85						90					95	
Leu	Leu	Ile	Asn	Gly	Glu	Leu	Pro	Thr	Pro	Asp	Glu	Leu	His	Lys	Phe
			100					105					110		
Asn	Asp	Glu	Ile	Arg	His	His	Thr	Leu	Leu	Asp	Glu	Asp	Phe	Lys	Ser
			115				120					125			
Gln	Phe	Asn	Val	Phe	Pro	Arg	Asp	Ala	His	Pro	Met	Ala	Thr	Leu	Ala
			130				135				140				
Ser	Ser	Val	Asn	Ile	Leu	Ser	Thr	Tyr	Tyr	Gln	Asp	Gln	Leu	Asn	Pro
			145			150				155				160	
Leu	Asp	Glu	Ala	Gln	Leu	Asp	Lys	Ala	Thr	Val	Arg	Leu	Met	Ala	Lys
			165					170						175	
Val	Pro	Met	Leu	Ala	Ala	Tyr	Ala	His	Arg	Ala	Arg	Lys	Gly	Ala	Pro
			180					185					190		

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<400> SEQUENCE: 29

ccgtcgacaa tagcctgaa                               19

<210> SEQ ID NO 30
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer gltA_2.p

<400> SEQUENCE: 30

ccgaattcct cgagcatctc cagaac                               26

<210> SEQ ID NO 31
<211> LENGTH: 1695
<212> TYPE: DNA
<213> ORGANISM: Corynebacterium glutamicum
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(2)
<223> OTHER INFORMATION: sequence CC
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (3)..(8)
<223> OTHER INFORMATION: Sall restriction sequence
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (9)..(832)
<223> OTHER INFORMATION: sequence usptream of the coding region
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (833)..(1687)
<223> OTHER INFORMATION: N terminal portion of the coding region
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (847)..(847)
<223> OTHER INFORMATION: Transversion A > T
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1688)..(1693)
<223> OTHER INFORMATION: RcoRI restriction sequence
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1694)..(1695)
<223> OTHER INFORMATION: Sequence GG

<400> SEQUENCE: 31

ccgtcgacaa tagcctgaaat ctgttctggt cgaaccttgg aaggtccgca gccgaaacgg   60
ccgtcgccag g gatgaactc agagggcagg gtggggaagt cggtcatgtc ttcgggcaac   120
tttctgcgct t ggaagtaaa agggccaggg atcgttaacg atctgaccca acaactataa   180
ccctgaagct gtcagttcct agcaccttag attcttcacg cagtctccca aacgatgaaa   240
aacgcccaaa actggcgaca ccgaactatt gaaaacgcgg ggattagtgt accagccacc   300
aat ttggggg tagctcaaag ttttgcaaag ttttcaattt ctagggtgtt aatatcccct   360
gagggttgcg t ataggggtg cgaattgcat ggggaaagct actcggcacc catccttgtc   420
gcgtgcatca caaactttgc taaactgtgc accagtccac ttattgtggg atttttaatg   480
ccttaaaggc cagcattttc accctctagc ggggttgaat gctggccttg aggggtgcaga   540
actaaatagc agcacatcgg cacaattgat ctgagttcta ttggcgtgac cgtgggtact   600
gattacgggt gctgtgggtg gtcgggaatg atgtaaccaa cgtgattgtg ggggaattgg   660
ctctcacttc ggatattgct aaaccgcatt taccggtata gcgtgttaac cggaccagat   720
tgggaaagaa atgtgtcgag taacaaaaac tgacatgcgc ttggcgcac cagttggta   780
agaataaacg ggactacttc cgtaatccgg aagagttttt ttccgaacaa at atg ttt   838
Met Phe

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1

gaa agg gtt atc gtg gct act gat aac aac aag gct gtc ctg cac tac	886
Glu Arg Val Ile Val Ala Thr Asp Asn Asn Lys Ala Val Leu His Tyr	
5 10 15	
ccc ggt ggc gag ttc gaa atg gac atc atc gag gct tct gag ggt aac	934
Pro Gly Gly Glu Phe Glu Met Asp Ile Ile Glu Ala Ser Glu Gly Asn	
20 25 30	
aac ggt gtt gtc ctg ggc aag atg ctg tct gag act gga ctg atc act	982
Asn Gly Val Val Leu Gly Lys Met Leu Ser Glu Thr Gly Leu Ile Thr	
35 40 45 50	
ttt gac cca ggt tat gtg agc act ggc tcc acc gag tcc aag atc acc	1030
Phe Asp Pro Gly Tyr Val Ser Thr Gly Ser Thr Glu Ser Lys Ile Thr	
55 60 65	
tac atc gat ggc gat gcg gga atc ctg cgt tac cgc ggc tat gac atc	1078
Tyr Ile Asp Gly Asp Ala Gly Ile Leu Arg Tyr Arg Gly Tyr Asp Ile	
70 75 80	
gct gat ctg gct gag aat gcc acc ttc aac gag gtt tct tac cta ctt	1126
Ala Asp Leu Ala Glu Asn Ala Thr Phe Asn Glu Val Ser Tyr Leu Leu	
85 90 95	
atc aac ggt gag cta cca acc cca gat gag ctt cac aag ttt aac gac	1174
Ile Asn Gly Glu Leu Pro Thr Pro Asp Glu Leu His Lys Phe Asn Asp	
100 105 110	
gag att cgc cac cac acc ctt ctg gac gag gac ttc aag tcc cag ttc	1222
Glu Ile Arg His His Thr Leu Leu Asp Glu Asp Phe Lys Ser Gln Phe	
115 120 125 130	
aac gtg ttc cca cgc gac gct cac cca atg gca acc ttg gct tcc tcc	1270
Asn Val Phe Pro Arg Asp Ala His Pro Met Ala Thr Leu Ala Ser Ser	
135 140 145	
gtt aac att ttg tct acc tac tac cag gac cag ctg aac cca ctc gat	1318
Val Asn Ile Leu Ser Thr Tyr Tyr Gln Asp Gln Leu Asn Pro Leu Asp	
150 155 160	
gag gca cag ctt gat aag gca acc gtt cgc ctc atg gca aag gtt cca	1366
Glu Ala Gln Leu Asp Lys Ala Thr Val Arg Leu Met Ala Lys Val Pro	
165 170 175	
atg ctg gct gcg tac gca cac cgc gca cgc aag ggt gct cct tac atg	1414
Met Leu Ala Ala Tyr Ala His Arg Ala Arg Lys Gly Ala Pro Tyr Met	
180 185 190	
tac cca gac aac tcc ctc aat gcg cgt gag aac ttc ctg cgc atg atg	1462
Tyr Pro Asp Asn Ser Leu Asn Ala Arg Glu Asn Phe Leu Arg Met Met	
195 200 205 210	
ttc ggt tac cca acc gag cca tac gag atc gac cca atc atg gtc aag	1510
Phe Gly Tyr Pro Thr Glu Pro Tyr Glu Ile Asp Pro Ile Met Val Lys	
215 220 225	
gct ctg gac aag ctg ctc atc ctg cac gct gac cac gag cag aac tgc	1558
Ala Leu Asp Lys Leu Leu Ile Leu His Ala Asp His Glu Gln Asn Cys	
230 235 240	
tcc acc tcc acc gtt cgt atg atc ggt tcc gca cag gcc aac atg ttt	1606
Ser Thr Ser Thr Val Arg Met Ile Gly Ser Ala Gln Ala Asn Met Phe	
245 250 255	
gtc tcc atc gct ggt ggc atc aac gct ctg tcc ggc cca ctg cac ggt	1654
Val Ser Ile Ala Gly Gly Ile Asn Ala Leu Ser Gly Pro Leu His Gly	
260 265 270	
ggc gca aac cag gct gtt ctg gag atg ctc gaa gaattcgg	1695
Gly Ala Asn Gln Ala Val Leu Glu Met Leu Glu	
275 280 285	

<210> SEQ ID NO 32

<211> LENGTH: 285

<212> TYPE: PRT

<213> ORGANISM: Corynebacterium glutamicum

-continued

<400> SEQUENCE: 32

Met Phe Glu Arg Val Ile Val Ala Thr Asp Asn Asn Lys Ala Val Leu
 1 5 10 15
 His Tyr Pro Gly Gly Glu Phe Glu Met Asp Ile Ile Glu Ala Ser Glu
 20 25 30
 Gly Asn Asn Gly Val Val Leu Gly Lys Met Leu Ser Glu Thr Gly Leu
 35 40 45
 Ile Thr Phe Asp Pro Gly Tyr Val Ser Thr Gly Ser Thr Glu Ser Lys
 50 55 60
 Ile Thr Tyr Ile Asp Gly Asp Ala Gly Ile Leu Arg Tyr Arg Gly Tyr
 65 70 75 80
 Asp Ile Ala Asp Leu Ala Glu Asn Ala Thr Phe Asn Glu Val Ser Tyr
 85 90 95
 Leu Leu Ile Asn Gly Glu Leu Pro Thr Pro Asp Glu Leu His Lys Phe
 100 105 110
 Asn Asp Glu Ile Arg His His Thr Leu Leu Asp Glu Asp Phe Lys Ser
 115 120 125
 Gln Phe Asn Val Phe Pro Arg Asp Ala His Pro Met Ala Thr Leu Ala
 130 135 140
 Ser Ser Val Asn Ile Leu Ser Thr Tyr Tyr Gln Asp Gln Leu Asn Pro
 145 150 155 160
 Leu Asp Glu Ala Gln Leu Asp Lys Ala Thr Val Arg Leu Met Ala Lys
 165 170 175
 Val Pro Met Leu Ala Ala Tyr Ala His Arg Ala Arg Lys Gly Ala Pro
 180 185 190
 Tyr Met Tyr Pro Asp Asn Ser Leu Asn Ala Arg Glu Asn Phe Leu Arg
 195 200 205
 Met Met Phe Gly Tyr Pro Thr Glu Pro Tyr Glu Ile Asp Pro Ile Met
 210 215 220
 Val Lys Ala Leu Asp Lys Leu Leu Ile Leu His Ala Asp His Glu Gln
 225 230 235 240
 Asn Cys Ser Thr Ser Thr Val Arg Met Ile Gly Ser Ala Gln Ala Asn
 245 250 255
 Met Phe Val Ser Ile Ala Gly Gly Ile Asn Ala Leu Ser Gly Pro Leu
 260 265 270
 His Gly Gly Ala Asn Gln Ala Val Leu Glu Met Leu Glu
 275 280 285

The invention claimed is:

1. An isolated polynucleotide comprising a nucleotide sequence encoding the amino acid sequence of SEQ ID NO: 12, wherein the amino acid at position 5 is L-valine.

2. The isolated polynucleotide of claim 1, wherein said isolated polynucleotide comprises the nucleotide sequence from position 662 to 867 of SEQ ID NO: 7.

3. A vector comprising a polynucleotide as claimed in claim 1.

4. A bacterium that has been transformed with the vector as claimed in claim 3.

5. The bacterium of claim 4, wherein said bacterium is from the genus of *Corynebacterium* or *Escherichia*.

6. The bacterium of claim 5, wherein said bacterium is *Corynebacterium glutamicum* or *Escherichia coli*.

7. A bacterium of the genus *Corynebacterium*, which comprises a polynucleotide as in claim 1.

8. The bacterium of claim 7, wherein said bacterium is *Corynebacterium glutamicum*.

9. A recombinant bacterium of the genus *Corynebacterium* comprising an isolated polynucleotide comprising a nucleotide sequence encoding the amino acid sequence of SEQ ID NO: 12, wherein the amino acid at position 5 is L-valine.

* * * * *