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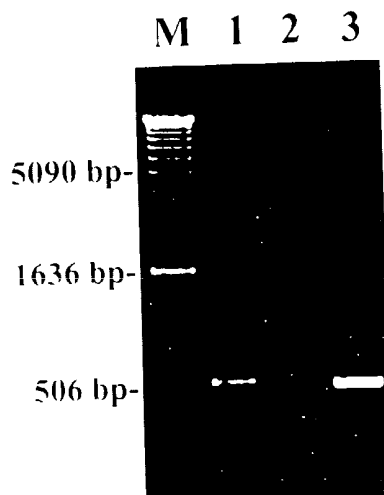
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(54) Title: METHODS TO CULTURE CIRCOVIRUS

(57) Abstract: The present invention relates to methods for culturing circovirus and in particular, porcine circovirus. The present invention provides compositions and methods for culturing porcine circovirus in mammalian cells expressing mammalian adenovirus E1 function.



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METHODS TO CULTURE CIRCOVIRUS

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TECHNICAL FIELD

The present invention relates to the field of circovirus and provides compositions and methods for culturing circovirus, in particular porcine circovirus. In particular, the present invention relates to methods for culturing porcine circovirus in mammalian cells expressing a mammalian adenovirus E1 gene function.

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BACKGROUND ART

A family of viruses, named *Circoviridae*, found in a range of plant and animal species and commonly referred to as circoviruses, are characterized as round, non-enveloped virions with mean diameters from 17 to 23.5 nm containing circular, single-stranded deoxyribonucleic acid (ssDNA). The ssDNA genome of the circoviruses represent the smallest viral DNA replicons known. As disclosed in WO 99/45956, at least six viruses have been identified as members of the family according to The Sixth Report of the International Committee for the Taxonomy of Viruses (Lukert, P.D. et al. 1995, *The Circoviridae*, pp. 166-168. In F.A. Murphy, et al. (eds.) Virus Taxonomy, Sixth Report of the International Committee on Taxonomy of Viruses, Arch. Virol. 10 Suppl.).

Animal viruses included in the family are chicken anemia virus (CAV); beak and feather disease virus (BFDV); porcine circovirus (PCV); and pigeon circovirus. PCV was originally isolated in porcine kidney cell cultures. PCV replicates in the cell nucleus and produces large intranuclear inclusion bodies. See Murphy et al. (1999, *Circoviridae* p. 357-361, Veterinary Virology, 3rd ed. Academic Press, San Diego). There are currently two recognized types of PCV, PCV type 1 (PCV1) and PCV type 2 (PCV2). PCV1, isolated as a persistent contaminant of the continuous porcine kidney cell line PK-15 (ATCC CCL31), does not cause detectable cytopathic effects in cell culture and fails to produce

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clinical disease in pigs after experimental infection (see Allan G., 1995, *Vet. Microbiol.* 44: 49-64; Tischer, I. et al., 1982, *Nature* 295:64-66; and Tischer, I. et al., 1986, *Arch. Virol.* 91:271-276). PCV2, in contrast to PCV1, is closely associated with post weaning multisystemic wasting syndrome (PMWS) in weanling pigs (see Allan G. et al., 1998, *Europe. J. Vet. Diagn. Investig.* 10:3-10; Ellis, J. et al., 1998, *Can. Vet. J.* 39:44-51 and Morozov, I. et al., 1998, *J. Clin. Microbiol.* 36:2535-2541). The nucleotide sequences for PCV1 are disclosed in Mankertz, A., et al. (1997, *J. Virol.* 71:2562-2566) and Meehan, B.M., et al. (1997, *J. Gen. Virol.* 78:221-227) and the nucleotide sequences for PCV2 are disclosed in Hamel, A.L. et al. (1998, *J. Virol.* 72:5262-5267); Mankertz, A. et al. (2000, *Virus Res.* 66:65-77) and Meehan, B.M. et al. (1998, *J. Gen. Virol.* 79:2171-2179). Strains of PCV2 are disclosed in WO 00/01409 and have been deposited at the European Collection of Cell Cultures, Centre for Applied Microbiology & Research, Porton Down, Salisbury, Wiltshire SP4 0JG, United Kingdom and include: accession No. V97100219; accession No. V9700218; accession No. V97100217; accession No. V98011608; and accession No. V98011609. WO 00/77216 also discloses PCV2.

Published studies to date on PCV2 used either tissue homogenate or cultured virus derived from field isolates. Tischer et al. (1987, *Arch Virol.* 96:39-57) report that porcine kidney cells are stimulated to entry to the S phase in the cell cycle by D-glucosamine treatment. However, the treatment must be performed with caution because D-glucosamine is toxic for cell culture (see, Allan et al., (2000). *J. Vet. Diagn. Investigation.* 12:3-14). There remains a need for methods for culturing circovirus, such as for example, PCV1 and PCV2, and other circoviruses, such that pure circovirus is obtained. Such methods would be advantageous, in particular for preparation of PCV2 antigens as vaccines directed against PMWS. The present invention addresses that need.

All patents and publications are hereby incorporated herein in their entirety.

DISCLOSURE OF THE INVENTION

The present invention provides methods for culturing mammalian circovirus comprising: a) obtaining mammalian cells expressing a mammalian adenovirus E1 function, wherein said cells are permissive for mammalian circovirus replication; b) introducing said mammalian circovirus genome, or a portion thereof capable of replication, into said mammalian cells; and c) culturing said mammalian cells under conditions suitable for replication of said mammalian circovirus. In some embodiments, the method further comprises recovering said circovirus from said cultured cells.

In some embodiments, the mammalian circovirus is porcine circovirus, such as for example, porcine circovirus 1 (PCV1) or porcine circovirus 2 (PCV2). In yet additional embodiments, the porcine circovirus comprises a chimeric nucleotide sequence. In other embodiments, the mammalian cells are of porcine origin. In yet other embodiments, the mammalian cells are porcine retina cells.

In other embodiments, the mammalian adenovirus E1 function is human adenovirus E1 function. In yet other embodiments, the mammalian adenovirus E1 function is porcine adenovirus E1 function. In further embodiments, the E1 function is E1A and/or E1B function. In yet further embodiments, the mammalian cell expressing the mammalian E1 function is stably transformed with mammalian E1 gene sequences. In other embodiments, the mammalian E1 gene sequence is heterologous to said mammalian cell.

The present invention also provides recombinant mammalian cells that express a mammalian adenovirus E1 function and comprise a mammalian circovirus genome, or a portion thereof capable of replication, and wherein said cells are permissive for the replication of said mammalian circovirus. In some embodiments, the mammalian circovirus is porcine circovirus, such as for example, porcine circovirus 1 (PCV1) or porcine circovirus 2 (PCV2). In yet additional embodiments, the porcine circovirus comprises a chimeric nucleotide sequence. In some embodiments, the adenovirus E1 function is human adenovirus E1 function. In other embodiments, the E1 function is porcine adenovirus E1 function. In other embodiments, the mammalian cell is of porcine origin. In further embodiments, the mammalian cell is a porcine retinal cell. In

yet further embodiments, the mammalian cell expressing the mammalian E1 function is stably transformed with mammalian adenovirus E1 gene sequences. In other embodiments, the mammalian E1 gene sequence is heterologous to said mammalian cell.

5 The present invention also provides methods of preparing a recombinant mammalian cell expressing a mammalian adenovirus E1 function and comprising a mammalian circovirus genome comprising the steps of, a) obtaining a mammalian cell expressing a mammalian adenovirus E1 function; and b) introducing said mammalian circovirus genome, or a portion thereof capable of
10 replication, into said mammalian cell. In additional embodiments, the method comprises the additional step of culturing the recombinant mammalian cell under conditions suitable for the replication of said mammalian circovirus. In further embodiments, the method comprises recovering said circovirus from said cultured cells. In some embodiments, the mammalian circovirus is porcine circovirus,
15 such as for example, porcine circovirus 1 (PCV1) or porcine circovirus 2 (PCV2). In yet additional embodiments, the porcine circovirus comprises a chimeric nucleotide sequence. In further embodiments, the mammalian cells are of porcine origin. In yet further embodiments, the mammalian cells are porcine retina cells. In additional embodiments, the adenovirus E1 function is human adenovirus E1
20 function or porcine adenovirus E1 function. In yet further embodiments, the mammalian cell expressing the mammalian adenovirus E1 function is stably transformed with mammalian adenovirus E1 gene sequences. In other embodiments, the mammalian E1 gene sequence is heterologous to said mammalian cell.

BRIEF DESCRIPTION OF THE DRAWINGS

25 Figures 1A-1B provide the characterization and titration of PCV2 virus generated by DNA transfection and extraction from infected VIDO R1 cells by Hirt's method. (A) PCR using PCV2-specific primers and DNA from PCV2-
30 infected (lane 1) and mock-infected (lane 2) cells. A plasmid containing PCV2 genome was used as a control (lane 3). The 1-kb DNA ladder from GIBCO BRL was loaded in lane M. (B) Viral DNA from PCV2-infected (lanes 1, 3, and 5) and

mock-infected (lanes 2, 4, and 6) cells were digested with *NcoI* and *StuI* (lanes 1 and 2), *EcoRI* and *StuI* (lanes 3 and 4), and *EcoRI* and *EcoRV* (lanes 5 and 6). The 1-kb-plus DNA ladder from GIBCO BRL was loaded in lane M.

5 Figures 2A-2B depict titration of PCV2 by immunoperoxidase staining. At 72 h.p.i., mock- (A) or PCV2- (B) infected VIDO R1 cells were incubated with rabbit anti-ORF2 polyclonal antibody and biotinylated secondary antibody. After application of an avidin and biotinylated horseradish peroxidase complex, the monolayer was developed by diaminobenzidine tetrahydrochloride (DAB). One dark cell resulted from one virus particle infection.

10 Figures 3A-3C show the nucleotide sequence (A) and amino acid sequence for ORF 1 (B) and ORF 2 (C) of porcine circovirus 2 (PCV2) as described in Genbank accession number AF086834.

BEST MODE FOR CARRYING OUT THE INVENTION

15 The present invention relates to compositions and methods for culturing mammalian circovirus, in particular porcine circovirus. The present invention is based on the finding that a porcine cell expressing human E1 function was able to be transfected with a PCV2 virus genome and generated PCV2 virus with a high virus titer. VIDO R1 cell line, deposited with the ATCC and having ATCC accession number PTA-155, is a porcine retina cell line transformed with
20 human adenovirus-5 (HAV5) E1, that has been shown to induce the S phase of the cell cycle and transactivate transcription. See, Shenk, T. (1996). "Adenoviridae: the viruses and their replication" In Fields Virology, 3rd ed. B.N. Fields, D.M. Knipe and P.M. Howley (ed.) Lippincott-Raven Publishers, Philadelphia, New York, pp. 2111-2148. As described herein in Example 3, VIDO R1 cells, were
25 transfected with a PCV2 genome and generated virus at 2×10^7 IU/ml.

 The practice of the present invention employs, unless otherwise indicated, conventional microbiology, immunology, virology, molecular biology, and recombinant DNA techniques which are within the skill of the art. These techniques are fully explained in the literature. See, e.g., Maniatis et al.,
30 *Molecular Cloning: A Laboratory Manual* (1982); *DNA Cloning: A Practical Approach*, vols. I & II (D. Glover, ed.); *Oligonucleotide Synthesis* (N. Gait, ed.

(1984)); *Nucleic Acid Hybridization* (B. Hames & S. Higgins, eds. (1985));
Transcription and Translation (B. Hames & S. Higgins, eds. (1984)); *Animal*
Cell Culture (R. Freshney, ed. (1986)); Perbal, *A Practical Guide to Molecular*
Cloning (1984); Ausubel, *et al.*, *Current Protocols In Molecular Biology*, John
5 Wiley & Sons (1987, 1988, 1989, 1990, 1991, 1992, 1993, 1994, 1995, 1996);
and Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual* (2nd Edition);
vols. I, II & III (1989).

Circoviridae, a family of viruses having round, non-enveloped virions
with mean diameters from 17 to 23.5 nm containing circular, single-stranded
10 DNA (ssDNA), are described in The Sixth Report of the International Committee
for the Taxonomy of Viruses, *supra*. Members of the group include the porcine
circoviruses, PCV1 and PCV2. Some of the PCVs are known to be pathogenic,
such as PCV2, associated with PMWS.

Nucleotide sequences for PCV1 are provided in Mankertz, A., et al., 1997,
15 *J. Virol.* 71:2562-2566 and Meehan, B.M. et al., 1997, *J. Gen. Virol.* 78:221-227.
Nucleotide sequences for PCV2 are provided in Hamel, A.L. et al., 1998, *J. Virol.*
72:5262-5267; Mankertz, A. et al., 2000, *Virus Res.* 66:65-77 and Meehan, B.M.
et al., 1998, *J. Gen. Virol.* 79:2171-2179. Representative strains of PCV2 have
been deposited with the European Collection of Cell Cultures, Centre for Applied
20 Microbiology & Research, Porton Down, Salisbury, Wiltshire SP4 0JG, United
Kingdom and include accession No. V97100219; accession No. V9700218;
accession No. V97100217; accession No. V98011608; and accession No.
V98011609. WO 00/77216 also discloses PCV2. PCV2 nucleotide sequences
have also been published in Hamel et al., (1998), *J. Virol.* vol. 72, 6:5262-5267
25 (GenBank AF027217) and in Morozov et al., (1998), *J. Clinical Microb.* vol. 36,
9:2535-2541, as well as GenBank AF086834; AF086835; and AF086836.
Comparison of the published nucleotide sequences for PCV1 and PCV2 reveals a
< 80% identity, although the genomic organization is similar, especially in the
arrangement of the two largest open reading frames (ORFs) with a putative origin
30 of DNA replication.

The present invention encompasses methods of culturing mammalian
circovirus and in particular, porcine circovirus (PCV). The present invention

encompasses methods of culturing PCV comprising the PCV nucleotide sequences disclosed herein or known in the art, or ORFs thereof, or portions thereof that are capable of replication. The present invention also encompasses methods of culturing PCV having PCV nucleotide sequences differing through the degeneracy of the genetic code to those disclosed herein or known in the art, or ORFs thereof, or portions thereof capable of replication. The present invention further encompasses methods of culturing PCV comprising PCV nucleotide sequence variations which do not change the functionality or strain specificity of the nucleotide sequence, or ORFs thereof, or portions thereof capable of replication. The present invention also encompasses methods of culturing PCV comprising PCV nucleotide sequences capable of hybridizing to those sequences disclosed herein under conditions of intermediate to high stringency, and methods of culturing PCV comprising mutations of the PCV nucleotide sequence disclosed herein or known in the art, such as deletions or point mutations, or ORFs thereof, or portions thereof, capable of replication. The present invention also encompasses methods of culturing PCV comprising heterologous nucleotide sequences. The present invention encompasses methods of culturing PCV that comprise chimeric circovirus nucleotide sequences, such as, for example, nucleotide sequences from porcine circovirus in fusion with nucleotide sequences from other pathogenic viruses, such as a pathogenic porcine virus, including parvovirus.

As used herein, a heterologous nucleotide sequence, with respect to a circovirus or mammalian cell, is one which is not normally associated with the circovirus sequences as part of the circovirus genome or one which is not normally associated with the mammalian cell, respectively. Heterologous nucleotide sequences include synthetic sequences. Hybridization reactions can be performed under conditions of different "stringency". Conditions that increase stringency of a hybridization reaction are widely known and published in the art. See, for example, Sambrook et al. (1989) at page 7.52. Examples of relevant conditions include (in order of increasing stringency): incubation temperatures of 25°C, 37°C, 50°C and 68°C; buffer concentrations of 10 X SSC, 6 X SSC, 1 X SSC, 0.1 X SSC (where SSC is 0.15 M NaCl and 15 mM citrate buffer) and their

equivalents using other buffer systems; formamide concentrations of 0%, 25%, 50%, and 75%; incubation times from 5 minutes to 24 hours; 1, 2, or more washing steps; wash incubation times of 1, 2, or 15 minutes; and wash solutions of 6 X SSC, 1 X SSC, 0.1 X SSC, or deionized water. An exemplary set of
5 stringent hybridization conditions is 68°C and 0.1 X SSC.

The PCV genomes encode several polypeptide sequences, ranging in approximate size from 8 to 35kD. It is deemed routine to determine open reading frames (ORFs) for porcine circoviruses using standard software such as for example, MacVector® (Oxford Molecular Group Inc., MD 21030). The largest
10 ORF, ORF1, of the two types of PCV shows only minor variation with an identity of 85% (as measured by the clustal program) and has been demonstrated to be the Rep protein in PCV1 (Mankertz, A., et al., 1998, *J. Gen. Virol.* 79:381-384). Without wanting to be bound by theory, a higher rate of variation displayed in the ORF2 sequences of PCV1 and PCV2 (identity about 65%) would suggest that
15 type-specific features of PCV might be determined by the respective ORF2 protein. Several PCV type-specific epitopes have been mapped on PCV2 ORF2 sequences. See Mahe, D. et al., 2000, *J. Gen. Virol.* 81:1815-24. In another recent study, PCV2 ORF2 has been identified as a major structural protein that can form viral capsid-like particles in insect cells infected with ORF2 expressing
20 recombinant baculovirus. See Nawagitgul, P. et al., 2000, *J. Gen. Virol.* 81:2281-2287.

In some illustrative embodiments of the invention disclosed herein, a recombinant vector comprising a PCV genome or an ORF thereof, or a portion thereof, such as an antigenic region, is constructed by *in vitro* recombination
25 between a plasmid and a PCV genome. In some embodiments, the PCV genome is a PCV2 genome. In other embodiments, a recombinant vector comprising a PCV genome or an ORF thereof, or a portion thereof, such as an antigenic region, is constructed by *in vivo* recombination. Methods for *in vivo* recombination are known in the art and include, for example, the methods disclosed in Chartier, et al.
30 (1996, *J. Virol.* 70:4805-4810). Vectors for constructing circovirus genomes include for example, bacterial plasmids which allow multiple copies of the cloned circovirus nucleotide sequence to be produced. In some embodiments, the

plasmid is co-transfected into a suitable host cell for recombination. Suitable host cells for recombination include any cell that will support recombination between a PCV genome and a plasmid containing PCV sequences, or between two or more plasmids, each containing PCV sequences. Recombination is generally performed in procaryotic cells, such as *E. coli* for example, while generation of circovirus is preferably performed in mammalian cells permissive for PCV replication, such as for example porcine cells and in particular, porcine cells capable of expressing mammalian adenovirus E1 function.

The present invention encompasses the use of any mammalian host cell permissive for circovirus replication, and in particular, permissive for replication of PCV, such as PCV1 and PCV2. Allan et al. (1995, *Veterinary Microbiology* 44: 49-64) report that PCV replicate in porcine and bovine monocyte/macrophage cultures. Tischer et al. (1987, *Arch. Virol.* 96:39-57) report that PCV is known to require actively dividing cells for replication in cell culture. Examples of cells or cell lines useful for replication of PCV include mammalian cells comprising E1 function and permissive for PCV replication, including porcine cells, such as porcine monocyte/macrophage cells and porcine retinal cells, expressing adenovirus E1 function. In an illustrative embodiment disclosed herein, porcine retina cells expressing human adenovirus E1 function are shown to be permissive for replication of PCV2 and shown to generate virus at 2×10^7 IU/ml. Porcine cell lines are available from public sources such as for example, the American Type Tissue Collection (ATCC). The growth of bacterial cell cultures, as well as culture and maintenance of eukaryotic cells and mammalian cell lines are procedures well-known to those of skill in the art.

The present invention encompasses methods of culturing mammalian circovirus, in particular, porcine circovirus, in mammalian host cells transfected with mammalian adenovirus E1 gene sequences. In some embodiments, the mammalian cell is stably transformed with adenovirus E1 gene sequences. In some embodiments, the E1 gene sequences are integrated into the genome of the mammalian cell. In other embodiments, the E1 gene sequences are present on a replicating plasmid. In yet other embodiments, the E1 gene sequence is heterologous to the mammalian cell. In an illustrative embodiment disclosed

herein a porcine mammalian cell is transformed with a human adenovirus 5 E1 gene sequence. The present invention encompasses the use of any mammalian cell or mammalian cell line expressing E1 function as long as the mammalian cell or cell line expressing E1 function is permissive for the replication of circovirus, in particular porcine circovirus, such as for example, porcine circovirus 1 or porcine circovirus 2. In preferred embodiments, the mammalian cell is a porcine cell or cell line. The present invention encompasses the use of any mammalian E1 function as long as the mammalian host cell expressing the mammalian E1 function is permissive for replication of circovirus, in particular, PCV, such as for example, porcine circovirus 1 or porcine circovirus 2. Mammalian adenovirus genomes are known in the art and are disclosed in, for example, Reddy et al. (1998, *Journal of Virology*, 72:1394) which discloses nucleotide sequence, genome organization, and transcription map of bovine adenovirus 3 (BAV3); and Kleiboeker (1995, *Virus Res.* 36:259-268), which discloses the E1 region of PAV-4. The present invention encompasses E1 function from any of the various serotypes of human adenovirus, such as Ad2, Ad5, Ad12, and Ad40. In an illustrative embodiment disclosed herein in Example 1, E1 function is human Ad5 E1 function. The human E1A gene is expressed immediately after viral infection (0-2 hours) and before any other viral genes. Flint (1982) *Biochem. Biophys. Acta* 651:175-208; Flint (1986) *Advances Virus Research* 31:169-228; Grand (1987) *Biochem. J.* 241:25-38. The transcription start site of Ad5 E1A is at nucleotide 498 and the ATG start site of the E1A protein is at nucleotide 560 in the virus genome. The E1B protein functions in *trans* and is necessary for transport of late mRNA from the nucleus to the cytoplasm. The E1B promoter of Ad5 consists of a single high-affinity recognition site for Spl and a TATA box. In particular, human adenovirus 5 E1A and E1B gene sequences are located at nucleotides 505-4034 of the nucleotide sequence provided in Chroboczek, J. et al. (1992, *Virology*. 186:280-285). In an illustrative embodiment disclosed herein in the Examples, the mammalian host cell is a porcine host cell transfected with human adenovirus 5 E1 gene sequences.

The PCV genome can be isolated from PCV virions, or can comprise a PCV genome that has been inserted into a plasmid, using standard techniques of

molecular biology and biotechnology. Cloning of the full-length PCV2 genome into vector pBluescript II KS(+) from Strategene by PCR is described in Liu, et al. (2000, *J. Clin. Microbiol.* vol 38:3474-3477). The full-length PCV2 genome DNA can be released from the resulting plasmid upon *SacII* digestion.

5 Introduction of circovirus nucleotide sequences into permissive mammalian host cells can be achieved by any method known in the art, including, but not limited to, transfection and transformation including, but not limited to, microinjection, electroporation, CaPO_4 precipitation, DEAE-dextran, liposomes, particle bombardment, *etc.* An illustrative method for transfecting PCV2
10 nucleotide sequences into VIDO R1 cells is described herein in Example 3.

Methods for culturing procaryotic cells, such as bacterial cells, and eukaryotic cells, such as mammalian host cells expressing adenovirus E1 function are deemed routine to those of skill in the art.

The following examples are provided to illustrate but not limit the
15 invention. All references and patent publications disclosed herein are hereby incorporated in their entirety by reference.

EXAMPLES

Example 1: Preparation of porcine retinal cells transfected with human adenovirus E1 gene sequences (VIDO R1 cells)

20 Primary cultures of porcine embryonic retina cells were transfected with 10 μg of plasmid pTG 4671 (Transgene, Strasbourg, France) by the calcium phosphate technique. The pTG 4671 plasmid contains the entire E1A and E1B sequences (nts 505-4034) of HAV-5, along with the puromycin acetyltransferase gene as a selectable marker. In this plasmid, the E1 region is under the control of
25 the constitutive promoter from the mouse phosphoglycerate kinase gene, and the puromycin acetyltransferase gene is controlled by the constitutive SV40 early promoter. Transformed cells were selected by three passages in medium containing 7 $\mu\text{g}/\text{ml}$ puromycin, identified based on change in their morphology from single foci (*i.e.*, loss of contact inhibition), and subjected to single cell
30 cloning. The established cell line was first tested for its ability to support the growth of E1 deletion mutants of HAV-5. Subsequently the cell line was further investigated for the presence of E1 sequences in the genome by PCR, expression

of the E1A and E1B proteins by Western blot, and doubling time under cell culture conditions. E1 sequences were detected, and production of E1A and E1B proteins was demonstrated by immunoprecipitation. Doubling time was shorter, when compared to that of the parent cell line.

5 To assess the stability of E1 expression, VIDO R1 cells were cultured through more than 50 passages (split 1:3 twice weekly) and tested for their ability to support the replication of E1-deleted HAV-5. Expression of the E1A and E1B proteins at regular intervals was also monitored by Western blot. The results indicated that the VIDO R1 line retained the ability to support the growth of
10 E1-deleted virus and expressed similar levels of E1 proteins during more than 50 passages in culture. Therefore, VIDO R1 can be considered to be an established cell line. VIDO R1 cell line has been deposited with the American Type Culture collection (ATCC) and has ATCC accession number PTA-155.

Example 2

15 Example 2 provides a description of the molecular cloning of full-length PCV2 genome.

Initially, PCV2 DNA was amplified by PCR from total DNA extracted from a piglet with PMWS. The cloning of the full-length PCV2 genome DNA into vector pBluescript II KS(+) (Stratagene) by polymerase chain reaction (PCR)
20 was described in Liu et al. (2000). *J. Clin. Microbiol.* 38:3474-3477). The PCV2 sequence was submitted to GenBank (Accession no. AF086834). The full-length PCV2 genome DNA is released from the resulting plasmid upon *SacII* digestion.

Example 3

Example 3 describes the transfection of VIDO R1 cells, as described in
25 Example 1, with a plasmid containing the PCV2 genome as constructed in Example 2.

Material and methods

Cell culture

Fetal porcine retina cell line, VIDO R1, as described in Example 1 and
30 Vero cells (ATCC) were maintained at 37°C with 5% CO₂ in Eagle's based MEM media supplemented with 10% or 5% heat-inactivated fetal bovine serum (FBS), respectively.

Transfection and Infection

Monolayers of VIDO R1 cells grown in a six-well dish were transfected with cloned PCV2 DNA using Lipofectin according to the manufacturer's recommendations (GIBCO BRL). Prior to transfection, PCV2 full-length genome was released from the plasmid by digestion with *SacII* (Liu, Q., et al., 2000, *J. Clin. Microbiol.* 38:3474-3477). For infection, the transfected VIDO R1 cells were subjected to three cycles of freezing (-70°C) and thawing (37°C). The lysate was then clarified by centrifugation and used to infect fresh VIDO R1 cells. In published reports, a PCV1-free porcine kidney cell line is used to culture PCV2 virus. To stimulate the entry to the S phase in the cell cycle, the porcine kidney cells are always treated by D-glucosamine (see Tischler et al., (1987). *Arch Virol.* 96:39-57.). However, the treatment must be performed with caution because D-glucosamine is toxic for cell culture (see, Allan et al., (2000). *J. Vet. Diagn. Investigation.* 12:3-14). In contrast, since the VIDO R1 cell line used in this study has been transformed by HAV5-E1 that can induce the S phase, the D-glucosamine treatment was not necessary.

Example 4

Virus purification and titration

For the purification of PCV2 virus, PCV2-infected VIDO R1 cells were incubated with 0.5% Triton X-114 in phosphate-buffered saline (PBS) at 37°C for 45 min followed by Freon 113 (1,1,2-trichloro-trifluoroethane) extraction. The cell debris and membranes were clarified by centrifugation at 2000g for 15 min. The viruses in the supernatant were pelleted at 35000g for 3 h through a 20% sucrose cushion. The virus pellet was suspended in PBS and stored at -70°C. Virus titers were determined as infectious units (IU) by quantitative ORF2 protein immuno-peroxidase staining. For this purpose, the cell monolayers in 12-well dishes were infected with serial dilutions of virus. After adsorption of virus for 1 h, the cells were washed and overlaid with MEM containing 2% FBS and 0.7% agarose. On day 3 post infection (p.i.), the agarose overlay was removed and the cells were fixed and permeabilized with methanol/acetone (1:1 in volume) for 20 min at -20°C. After blocking with 1% bovine serum albumin for 1 h at room temperature, the cells were incubated with rabbit anti-ORF2 serum (Liu et al.,

2001, Protein Expression and Purification. 21:115-120). After 2 h incubation, the plates were washed with PBS and then processed using VECTASTAIN Elite ABC kit (Vector Laboratories). The reaction was developed with 3,3'-diaminobenzidine (DAB) tetrahydrochloride and observed under a microscope. By counting the positively stained cells, the virus titer was expressed as IU where 1 IU was defined as one positively stained cell/foci at 3 d.p.i.

Viral DNA extraction and characterization

Viral DNA was extracted from PCV2-infected VIDO RI cell monolayers by the method of Hirt (1967, *J. Mol. Biol.* 26:365-369). The viral DNA was then characterized by restriction analysis and polymerase chain reaction (PCR) as described in Liu et al., (2000). *J. Clin. Microbiol.* 38:3474-3477).

PCR using DNA extracted from the infected cells as template and PCV-2-specific primers amplified a product of specific size, while no DNA was amplified from control, uninfected cells. Consistent with the expected restriction patterns, digestions of viral DNA with NcoI and StuI resulted in two fragments of 1291 bp and 477 bp in size, respectively; digestion with EcoRI and StuI produced two fragments of 1492bp and 276bp in size, respectively; and digestion with EcoRI and EcoRV generated two fragments of 1094bp and 674bp in size, respectively. The data indicate that PCV2 virus was obtained. Using an immunostaining assay and by counting the positive stained cells, the virus titer of this preparation was determined to be 2×10^7 IU/ml.

CLAIMS

We claim:

1. A method for culturing a mammalian circovirus comprising:
 - a) obtaining mammalian cells expressing a mammalian adenovirus E1
5 function, wherein said cells are permissive for mammalian circovirus replication;
 - b) introducing the mammalian circovirus genome, or a portion thereof
capable of replication, into said mammalian cells; and
 - c) culturing said mammalian cells under conditions suitable for
replication of said mammalian circovirus.
- 10 2. The method of claim 1 further comprising recovering said circovirus
from said cultured cells.
3. The method of claim 1 wherein said mammalian circovirus is porcine
15 circovirus.
4. The method of claim 3 wherein said porcine circovirus is porcine
circovirus type 2.
- 20 5. The method of claim 3 wherein said porcine circovirus is porcine
circovirus type 1.
6. The method of claim 1 wherein said mammalian cells are of porcine
origin.
- 25 7. The method of claim 1 wherein said mammalian cells are porcine
retina cells.
8. The method of claim 1 wherein said mammalian adenovirus E1
30 function is human adenovirus E1 function.

9. The method of claim 1 wherein said mammalian adenovirus E1 function is porcine adenovirus E1 function.
- 5 10. The method of claim 1 wherein said mammalian cells expressing the mammalian adenovirus E1 function are stably transformed with a mammalian adenovirus E1 gene sequence.
- 10 11. The method of claim 10 wherein said E1 gene sequence is a human adenovirus E1 gene sequence.
12. The method of claim 10 wherein said mammalian adenovirus E1 gene sequence is heterologous to said mammalian cell.
- 15 13. The method of claim 1 wherein said E1 function is E1A and/or E1B function.
14. The method of claim 3 wherein said porcine circovirus comprises a chimeric nucleotide sequence.
- 20 15. A recombinant mammalian cell that expresses a mammalian adenovirus E1 function and comprises a porcine circovirus genome, or a portion thereof capable of replication, and wherein said cell is permissive for the replication of said porcine circovirus.
- 25 16. The recombinant mammalian cell of claim 15 wherein said adenovirus E1 function is human adenovirus E1 function.
17. The recombinant mammalian cell of claim 15 wherein said adenovirus E1 function is porcine adenovirus E1 function.
- 30

18. The recombinant mammalian cell of claim 15 wherein said cell is of porcine origin.
- 5 19. The recombinant mammalian cell of claim 18 wherein said cell is a porcine retina cell.
20. The recombinant mammalian cell of claim 15 wherein the mammalian cell expressing mammalian adenovirus E1 function is stably transformed with a mammalian adenovirus E1 gene sequence.
- 10 21. The recombinant mammalian cell of claim 20 wherein said E1 gene sequence is a human adenovirus E1 gene sequence.
22. The recombinant mammalian cell of claim 20 wherein said
- 15 mammalian adenovirus E1 gene sequence is heterologous to said mammalian cell.
23. A method of preparing a recombinant mammalian cell comprising a mammalian adenovirus E1 function and a porcine circovirus genome comprising the steps of, a) obtaining a mammalian cell expressing a
- 20 mammalian adenovirus E1 function; and b) introducing said porcine circovirus genome, or a portion thereof capable of replication, into the mammalian cell.
- 25 24. The method of claim 23 further comprising the step of culturing said recombinant mammalian cell under conditions suitable for the replication of said porcine circovirus.
- 30 25. The method of claim 24 further comprising recovering said circovirus from said cultured cells.

26. The method of claim 23 wherein said porcine circovirus is porcine circovirus type 2.
- 5 27. The method of claim 23 wherein said porcine circovirus is porcine circovirus type 1.
28. The method of claim 23 wherein said mammalian cells are of porcine origin.
- 10 29. The method of claim 28 wherein said mammalian cells are porcine retina cells.
30. The method of claim 23 wherein said adenovirus E1 function is human adenovirus E1 function.
- 15 31. The method of claim 23 wherein said adenovirus E1 function is porcine adenovirus E1 function.
32. The method of claim 23 wherein said porcine circovirus comprises a chimeric nucleotide sequence.
- 20 33. The method of claim 23 wherein said mammalian cell comprising a mammalian adenovirus E1 function is stably transformed with a mammalian adenovirus E1 gene sequence.
- 25 34. The method of claim 33 wherein said wherein said mammalian adenovirus E1 gene sequence is heterologous to said mammalian cell.

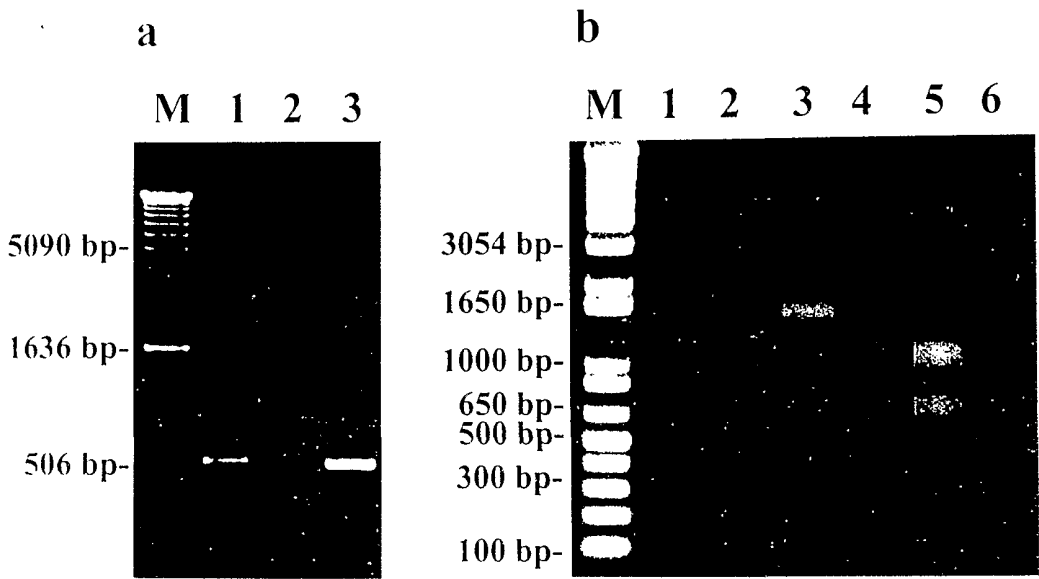


FIG. 1A-1B

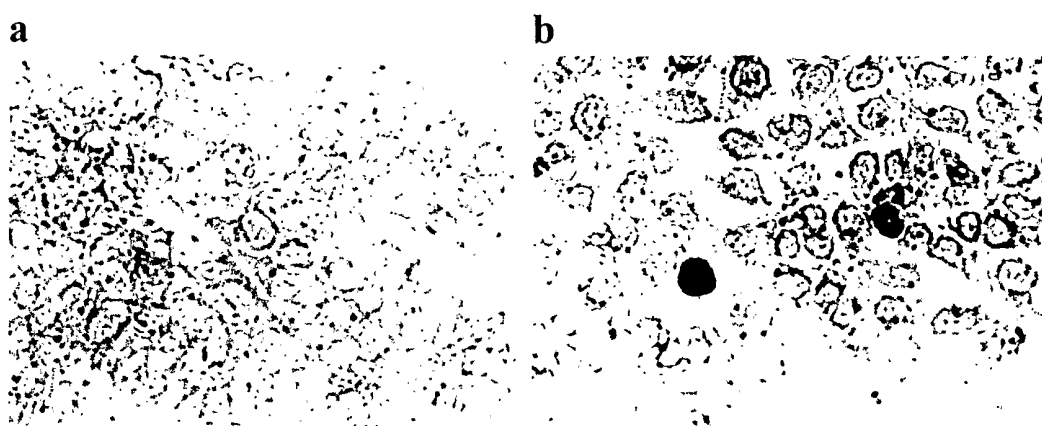


FIG. 2A-2B

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1 accagcgcac ttcggcagcg gcagcacctc ggcaacacct cagcagcaac atgccagca
61 agaagaatgg aagaagcggg ccccaaccac ataaaagggt ggtgttcacg ctgaataatc
121 cttccgaaga cgagcgcaag aaaatacggg agctcccaat ctccctattt gattatttta
181 ttgttggcga ggagggtaat gaggaaggac gaacacctca cctccagggg ttcgctaatt
241 ttgtgaagaa gcaaactttt aataaagtga agtggatatt ggtgcccgcg tgccacatcg
301 agaaagccaa aggaactgat cagcagaata aagaatattg tagtaaagaa ggcaacttac
361 ttattgaatg tggagctcct cgatctcaag gacaacggag tgacctgtct actgctgtga
421 gtaccttggt ggagagcggg attctgggtga ccgttgcaaa gcagcaccct gtaacgtttg
481 tcaaaaatth ccgcgggctg gctgaacttt tgaaagtga cgggaaaatg caaaagcgtg
541 attggaaaac caatgtacac ttcattgttg ggccacctgg gtgtggtaaa agcaaatggg
601 ctgctaattt tgcaaacccg gaaaccacat actggaaacc acctaaaaac aagtgggtggg
661 atgggttacca tgggtgaaaaa gtgggttgta ttgatgactt ttatggctgg ctgccgtggg
721 atgatctact gagactgtgt gatcgatatc cattgactgt aaaaactaaa ggtggaactg
781 tacctttttt ggcccgagc attctgatta ccagcaatca aaccccggtg gaatgggtact
841 cctcaactgc tgtcccagct gtagaagctc tctatcggag gattacttcc ttgggtattht
901 ggaagaatgt tacagaacaa tccacggagg aagggggcca gtttgtcacc ctttcccccc
961 catgccctga atthccatat gaaataaatt actgagtctt tttatcact togtaatggg
1021 ttttattatt catttagggg ttaagtggg ggtctthtaag attaaattct ctgaattgta
1081 catacatggg tacacggata ttgtagtctt ggtcgtattht actgtthtctg aacgcagtgc
1141 cgaggcctac gtgggtccaca tttctagagg tttgtagcct cagccaaagc tgattccttht
1201 tgttattthg ttggaagtaa tcaatagtgg agtcaagaac aggtthtggg gtgaagtaac
1261 gggagtggtg ggagaagggg tgggggattg tatggcggga ggagtagtht acatatgggt
1321 catagggttag ggctgtggcc tttgttacaa agttatcatc tagaataaca gcagtggagc
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1441 ttattctgta gtattcaaag ggtatagaga tttgttggt cccctctccc gggggaacaa
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1561 ccttgacagt atatccgaag gtgcgggaga ggcgggtgtt gaagatgcca ttttctctc
1621 tccaacggta gcggtggcgg ggggtggacga gccaggggcg gcggcgagg atctggccaa
1681 gatggctgcg ggggcggtgt cttcttctgc ggtaacgcct ccttggtatc gtcatagtcg
1741 aaaacgaaag aagtgcgctg taagtatt

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FIG. 3A

MPSKKNGRSGPQPHKRWVFTLNNPSEDERKKIRELPISLFDYFI
VGEEGNEEGRTPHLQGFANFVKKQTFNKVKWYLGARCHIEKAKGTDQQNKEYCSKEGN
LLIECGAPRSQGQRSDLSTAVSTLLESGILVTVAQHVPVTFVKNFRGLAELLKVSGKM
QKRDWKTNVHFIVGPPGCGKSKWAANFANPETTYWKPPKNKWWDGYHGEKVVIDDFY
GWL PWDDLRLCDRYPLTVKTKGGTVPF LARSILITSNQTPLEWYSSTAVPAVEALYR
RITSLVFWKNVTEQSTEEGGQFVTLSPPCPEFPYEINY"

FIG. 3B

MTYPRRRYRRRRHRPRSHLGQILRRRPWLHVHPRHRYRWRRKNGI
FNTRL SRTFGYTVKATTVTTPSWAVDMMRFNIDDFVPPGGGTNKISIPFEYYRIRKVK
VEFWPCSPITQGDRGVGSTAVILDDNFVTKATALTYDPYVNYSSRHTIPQPFSYHSRY
FTP KPVL DSTIDYFQPNKNRNQLWLRQLQTSRNV DHVGLGTAFENSKYDQDYNIRVTMY
VQFRE FN LKDPPLKP"

FIG. 3C