

(51) International Patent Classification:
C12N 15/82 (2006.01) C12N 15/01 (2006.01)

(21) International Application Number:
PCT/IB2015/054641

(22) International Filing Date:
19 June 2015 (19.06.2015)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
62/014,808 20 June 2014 (20.06.2014) US

(71) Applicant: CELLECTIS [FR/FR]; 8 rue de la Croix Jarry, F-75013 Paris (FR).

(72) Inventors: CLASEN, Benjamin; 2255 Jewell Lane, South St. Paul, 55075 (US). VOYTAS, Daniel F.; 2197 Folwell Avenue, Falcon Heights, Minnesota 55108 (US). ZHANG, Feng; 4530 Juneau Lane N., Plymouth, Minnesota 55446 (US).

KZ, LA, LC, LK, LR, LS, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PA, PE, PG, PH, PL, PT, QA, RO, RS, RU, RW, SA, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

(84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LR, LS, MW, MZ, NA, RW, SD, SL, ST, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, RU, TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, KM, ML, MR, NE, SN, TD, TG).

Declarations under Rule 4.17:

— as to applicant's entitlement to apply for and be granted a patent (Rule 4.17(ii))

— as to the applicant's entitlement to claim the priority of the earlier application (Rule 4.17(iii))

Published:

— with international search report (Art. 21(3))

— with sequence listing part of description (Rule 5.2(a))

(54) Title: POTATOES WITH REDUCED GRANULE-BOUND STARCH SYNTHASE

FIG. 1

GBSS_T1
ac~~t~~ctgactcacaatgggtttaagggcgtttaacaagcttgatggcctccaatcaagaactaataactaaggttaacacccaag
atggcatccagaactgagaccaagagacctggatgctcagctaccattgtttgtggaagggaatgaacttgatctttgtg
ggtactgaggttggtccttgagcaaaactggtgcactagtgatcttcttggtggactaccaccagcccttgacgta
{SEQ ID NO:1}

(57) Abstract: Materials and methods for making plants (e.g., *Solanum* varieties) with decreased levels of amylose are provided herein. The methods can include making mutations in the gene encoding granule bound starch synthase (GBSS), where the mutations are induced using a rare-cutting endonuclease.

WO 2015/193858 A1

POTATOES WITH REDUCED GRANULE-BOUND STARCH SYNTHASE

CROSS-REFERENCE TO RELATED APPLICATIONS

This application is claims benefit of priority from U.S. Provisional Application
5 No. 62/014,808, filed on June 20, 2014.

TECHNICAL FIELD

This document provides materials and methods for creating potato varieties
with reduced granule bound starch synthase.

10

BACKGROUND

The world starch market was estimated to be 48.5 million tons in 2000, with
an output value of 20 billion dollars per year. Only about 5% of the global starch
supply (2.6 million tons) is obtained from potatoes (European Commission – DG
15 Agriculture, *Evaluation of the Community Policy for Starch and Starch Product*,
2002, LMC INTERNATIONAL, Oxford, England, pp. 1-12). Potato tuber starch is
mainly composed of two polymers – amylopectin and amylose. The different
physiochemical properties of these two molecules can require costly processing
before the starch is amenable to industrial processes.

20

SUMMARY

This document provides materials and methods for creating potato varieties
that have reduced granule-bound starch synthase (GBSS). GBSS is an enzyme
involved in the synthesis of amylose, and reduced levels of GBSS therefore can result
25 in starch that contains a reduced proportion of amylose and a comparatively higher
proportion of amylopectin. Potato varieties having such modified starch are also
provided.

The disclosure herein is based at least in part on the discovery that potatoes
having reduced GBSS can be obtained, without the use of transgenesis, using a
30 sequence-specific nuclease to make a targeted mutation or knockout in the GBSS
gene. The modified potatoes can have improved starch characteristics for particular
industrial purposes, as compared to non-modified potatoes. Further, the potatoes do

not carry any foreign DNA and therefore may not be considered by regulatory agencies as a transgenic or genetically modified (GM) crop. This document also is based at least in part on the development of potato cultivars with loss-of-function GBSS mutations that are created by sequence-specific nucleases.

5 In one aspect, this document features a *Solanum* plant, plant part, or plant cell containing a mutation in at least two (e.g., at least three) granule-bound starch synthase (GBSS) alleles endogenous to the plant, plant part, or plant cell, such that the plant, plant part, or plant cell has reduced expression of GBSS as compared to a control *Solanum* plant, plant part, or plant cell that lacks the mutation. Each mutation
10 can be a deletion of more than one nucleotide base pair, and can be at a target sequence as set forth in SEQ ID NO:4, at a target sequence having at least 95 percent identity to SEQ ID NO:4, at a target sequence as set forth in SEQ ID NO:1, or at a target sequence having at least 95 percent identity to SEQ ID NO:1.

The plant, plant part, or plant cell can be made using a rare-cutting
15 endonuclease (e.g., a transcription activator-like effector endonuclease (TALE-nuclease)). The TALE-nuclease can bind to a sequence as set forth in SEQ ID NO:2 or SEQ ID NO:3, for example. Each of the at least two GBSS alleles can exhibit removal of an endogenous nucleic acid, without including any exogenous nucleic acid. Every endogenous GBSS allele may be mutated, and may exhibit removal of an
20 endogenous nucleic acid without including any exogenous nucleic acid. The plant, plant part, or plant cell may have no detectable expression of GBSS.

The *Solanum* plant, plant part, or plant cell can be, for example a *S. tuberosum* plant, plant part, or plant cell. The plant, plant part, or plant cell can have decreased levels of amylose as compared to a control plant, plant part, or plant cell that lacks the
25 mutation.

In another aspect, this document features a method for making a *Solanum* plant. The method can include (a) contacting a population of *Solanum* plant cells containing a functional GBSS allele with a rare-cutting endonuclease targeted to an endogenous GBSS sequence, (b) selecting from the population a cell in which at least
30 two (e.g., at least three) GBSS alleles have been inactivated, and (c) growing the selected plant cell into a *Solanum* plant, wherein the *Solanum* plant has reduced levels of amylose as compared to a control *Solanum* plant in which the at least two GBSS

alleles have not been inactivated. The *Solanum* plant cells can be, for example, protoplasts. The method can include transforming the population of plant cells with a nucleic acid (e.g., an mRNA or a vector) encoding the rare-cutting endonuclease, or introducing into the population of plant cells a rare-cutting endonuclease protein.

5 In some embodiments, the rare-cutting endonuclease can be a TALE-nuclease (e.g., a TALE-nuclease targeted to a sequence as set forth in SEQ ID NO:4, to a sequence having at least 95 percent identity to the sequence set forth in SEQ ID NO:4, to a sequence as set forth in SEQ ID NO:1, or to a sequence having at least 95 percent identity to the sequence set forth in SEQ ID NO:1. The TALE-nuclease can bind to a
10 sequence as set forth in SEQ ID NO:2 or SEQ ID NO:3.

The method can further include culturing the selected plant cells to generate plant lines, and/or isolating genomic DNA containing at least a portion of the GBSS locus from the plant cells. In some cases, the *Solanum* plant cells are *S. tuberosum* plant cells.

15 In still another aspect, this document features a method for producing a starch product. The method can include (a) providing a *Solanum* plant or plant part that contains a mutation in at least two (e.g., at least three) GBSS alleles endogenous to the plant or plant part, such that the plant, plant part, or plant cell has reduced expression of GBSS and reduced levels of amylose as compared to a control *Solanum*
20 plant or plant part that lacks the mutation, and (b) producing a starch product from the plant or plant part. Each said mutation can be at a target sequence as set forth in SEQ ID NO:4, at a target sequence having at least 95 percent identity to SEQ ID NO:4, at a target sequence as set forth in SEQ ID NO:1, or at a target sequence having at least 95 percent identity to SEQ ID NO:1. Each mutation can be made using a rare-cutting
25 endonuclease (e.g., a TALE-nuclease, such as a TALE-nuclease that binds to a sequence as set forth in SEQ ID NO:2 or SEQ ID NO:3). The *Solanum* plant or plant part can be a *S. tuberosum* plant or plant part, and may have no detectable expression of granule-bound starch synthase.

This document also features a starch product produced from a *Solanum* plant
30 or plant part that contains a mutation in each GBSS allele endogenous to the plant or plant part, such that the plant, plant part, or plant cell has no functional GBSS allele. Each mutation can be at a target sequence as set forth in SEQ ID NO:4, at a target

sequence having at least 95 percent identity to SEQ ID NO:4, at a target sequence as set forth in SEQ ID NO:1, or at a target sequence having at least 95 percent identity to SEQ ID NO:1. Each mutation can be made using a rare-cutting endonuclease (e.g., a TALE-nuclease). The TALE-nuclease can bind to a sequence as set forth in SEQ ID NO:2 or SEQ ID NO:3. The starch product can have decreased levels of amylose as compared to a starch product made from a control plant or plant part that lacks the mutation. The *Solanum* plant or plant part can be a *S. tuberosum* plant or plant part, and the starch product can be a reduced-amylose starch.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention pertains. Although methods and materials similar or equivalent to those described herein can be used to practice the invention, suitable methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

The details of one or more embodiments of the invention are set forth in the accompanying drawings and the description below. Other features, objects, and advantages of the invention will be apparent from the description and drawings, and from the claims.

DESCRIPTION OF DRAWINGS

FIG. 1 shows a representative DNA sequence (SEQ ID NO:1) from a GBSS gene. The underlined sequences (SEQ ID NOS:2 and 3) represent target sites for TALE-nucleases that recognize the GBSS gene.

FIG. 2 shows the DNA sequence of a naturally occurring *S. tuberosum* GBSS nucleotide sequence (SEQ ID NO:4).

FIG. 3 shows examples of TALE-nuclease-induced mutations in the GBSS gene. The top line of each panel shows the DNA sequence of the recognition site for the GBSS TALE-nucleases (underlined). The other sequences show representative

mutations that were induced by imprecise non-homologous end-joining (NHEJ). Deletion sizes are given on the right.

FIGS. 4A and 4B show examples of six individual plant genotypes that contain TALE-nuclease-induced mutations in the GBSS gene. St226 refers to the experiment number, and the -# refers to the specific plant that was regenerated from experiment St226. Underlined nucleotides correspond to the TALE-nuclease binding site. The size of the deletion, the frequency of the sequenced alleles, and the SEQ ID NOS are shown to the right of the sequences.

FIG. 5 shows the genotype of plant St226-8, which contains TALE-nuclease-induced mutations in the GBSS gene. Underlined nucleotides correspond to the TALE-nuclease target site. The size of the deletion, the frequency of the sequenced alleles, and the SEQ ID NO are shown to the right of the sequences.

FIG. 6 is a graph plotting results from an amylose/amylopectin assay of tubers from plant lines St226-1, St226-2, St226-4, St226-5, St226-6, St226-9, and wild type (WT). The percent amylose for each line is shown above the corresponding bar.

FIG. 7 is a graph plotting results from an amylose/amylopectin assay of tubers from plant lines St226-8 and WT. Five different tubers, labeled Tuber 1 – Tuber 5, were sampled from line St226-8. The percent amylose for each line is shown above the corresponding bar.

DETAILED DESCRIPTION

The main component of potato tuber starch is a mixture of two glucose polymers, amylopectin and amylose. Amylopectin, which is the major component of starch, is a soluble branched glucose structure composed of chains of 20-25 α -1,4 D-glucose residues with α -1,6 branching linkages (Aiyer, *African J. Biotech.* 4(13):1525-1529, 2005). Amylose is an insoluble linear helical polymer with minimal branching that constitutes 20-30% of the starch in plant-storage organs (Denyer et al., *Biochem. J.* 340:183-191, 1999). The synthesis of amylose is due to the GBSS mechanism of transferring glucosyl residues from ADP-glucose to glucan substrates, such as malto-oligosaccharides, by α -1,4 linkages (Aiyer, *African J. Biotech.* 4(13):1525-1529, 2005; and Zeeman et al., *Annu. Rev. Plant Biol.* 61:209-234, 2010). In many industrial applications, only the thickening amylopectin component is

desired, but separating amylopectin from amylose typically is energy intensive and not economically viable.

This document provides potato plant varieties, particularly of the species *Solanum tuberosum*, that have reduced or even lack GBSS activity. Methods for generating such plant varieties, methods for using such plant varieties to produce industrial starch products, and industrial starch products produced from such plant varieties also are provided.

As used herein, the terms “plant” and “plant part” refer to cells, tissues, organs, seeds, and severed parts (e.g., roots, leaves, and flowers) that retain the distinguishing characteristics of the parent plant. “Seed” refers to any plant structure that is formed by continued differentiation of the ovule of the plant, following its normal maturation point at flower opening, irrespective of whether it is formed in the presence or absence of fertilization and irrespective of whether or not the seed structure is fertile or infertile.

The term “allele(s)” means any of one or more alternative forms of a gene at a particular locus. In a diploid (or amphidiploid) cell of an organism, alleles of a given gene are located at a specific location or locus on a chromosome, with one allele being present on each chromosome of the pair of homologous chromosomes. Similarly, in a tetraploid cell of an organism, one allele is present on each chromosome of the group of four homologous chromosomes. “Heterozygous” alleles are different alleles residing at a specific locus, positioned individually on corresponding homologous chromosomes. “Homozygous” alleles are identical alleles residing at a specific locus, positioned individually on corresponding homologous chromosomes in the cell.

“Wild type” as used herein refers to a typical form of a plant or a gene as it most commonly occurs in nature. A “wild type GBSS allele” is a naturally occurring GBSS allele (e.g., as found within naturally occurring *S. tuberosum* plants) that encodes a functional GBSS protein, while a “non-functional mutant GBSS allele” is a GBSS allele that does not encode a functional GBSS protein. Such a “non-functional mutant GBSS allele” can include one or more mutations in its nucleic acid sequence, where the mutation(s) result in a reduced or even no detectable amount of functional GBSS protein in the plant or plant cell *in vivo*.

“Mutagenesis” as used herein refers to processes in which mutations are introduced into a selected DNA sequence. Mutations induced by endonucleases generally are obtained by a double strand break, which results in insertion/deletion mutations (“indels”) that can be detected by deep-sequencing analysis. Such mutations typically are deletions of several base pairs, and have the effect of inactivating the mutated allele. In the methods described herein, for example, mutagenesis occurs via double-stranded DNA breaks made by TALE-nucleases targeted to selected DNA sequences in a plant cell. Such mutagenesis results in “TALE-nuclease-induced mutations” (e.g., TALE-nuclease-induced knockouts) and reduced expression of the targeted gene. Following mutagenesis, plants can be regenerated from the treated cells using known techniques (e.g., planting seeds in accordance with conventional growing procedures, followed by self-pollination).

The term “expression” as used herein refers to the transcription of a particular nucleic acid sequence to produce sense or antisense RNA or mRNA, and/or the translation of an mRNA molecule to produce a polypeptide, with or without subsequent post-translational events.

“Reducing the expression” of a gene or polypeptide in a plant or a plant cell includes inhibiting, interrupting, knocking-out, or knocking-down the gene or polypeptide, such that transcription of the gene and/or translation of the encoded polypeptide is reduced as compared to a corresponding control plant, plant cell, or population of plants or plant cells in which expression of the gene or polypeptide is not inhibited, interrupted, knocked-out, or knocked-down. “Reduced expression” encompasses any decrease in expression level (e.g., a decrease of 10% or more, 20% or more, 30% or more, 40% or more, 50% or more, 60% or more, 70% or more, 80% or more, 90% or more, or even 100%) as compared to the corresponding control plant, plant cell, or population of plants or plant cells. In some embodiments, reducing expression by 50% or more may be particularly useful. Expression levels can be measured using methods such as, for example, reverse transcription-polymerase chain reaction (RT-PCR), Northern blotting, dot-blot hybridization, *in situ* hybridization, nuclear run-on and/or nuclear run-off, RNase protection, or immunological and enzymatic methods such as ELISA, radioimmunoassay, and western blotting.

The potato genome usually contains only one GBSS gene, but because cultivated potato is a tetraploid, multiple alleles of GBSS are present in each variety. The methods provided herein can be used to inactivate at least one (e.g., at least two, at least three, or all four) functional alleles of GBSS, thereby removing at least some full-length RNA transcripts and functional GBSS protein from potato cells, and in some cases completely removing all full-length RNA transcripts and functional GBSS protein.

A representative example of a naturally occurring *S. tuberosum* GBSS nucleotide sequence is shown in FIG. 2 (SEQ ID NO:4). In some embodiments, the *S. tuberosum* plants, cells, plant parts, seeds, and progeny thereof that are provided herein can have a mutation in each endogenous GBSS allele, such that expression of the gene is reduced or completely inhibited. Thus, in some cases, the plants, cells, plant parts, seeds, and progeny do not exhibit detectable levels of granule-bound starch synthase expressed from the GBSS gene.

The plants, plant cells, plant parts, seeds, and progeny provided herein can be generated using a rare-cutting endonuclease (e.g., a transcription activator-like effector nuclease (TALE-nuclease)) system to make a targeted knockout in one or more alleles of the GBSS gene. Thus, this document provides materials and methods for using rare-cutting endonucleases (e.g., TALE-nucleases) to generate potato plants and related products (e.g., seeds and plant parts) that are particularly suitable for providing reduced-amylose starch, due to targeted knockouts in the GBSS gene. Other sequence-specific nucleases also may be used to generate the desired plant material, including engineered homing endonucleases or zinc finger nucleases (ZFNs).

The term “rare-cutting endonuclease” as used herein refers to a natural or engineered protein having endonuclease activity directed to a nucleic acid sequence with a recognition sequence (target sequence) about 12-40 bp in length (e.g., 14-40, 15-36, or 16-32 bp in length; see, e.g., Baker, *Nature Methods* 9:23-26, 2012). Typical rare-cutting endonucleases cause cleavage inside their recognition site, leaving 4 nt staggered cuts with 3'OH or 5'OH overhangs. In some embodiments, a rare-cutting endonuclease can be a meganuclease, such as a wild type or variant homing endonuclease (e.g., a homing endonuclease belonging to the dodecapeptide

family (LAGLIDADG; SEQ ID NO:18); *see*, WO 2004/067736). In some embodiments, a rare-cutting endonuclease can be a fusion protein that contains a DNA binding domain and a catalytic domain with cleavage activity. TALE-nucleases and ZFNs are examples of fusions of DNA binding domains with the catalytic domain of the endonuclease *FokI*. Customized TALE-nucleases are commercially available under the trade name TALEN™ (Cellestis, Paris, France).

Transcription activator-like (TAL) effectors are found in plant pathogenic bacteria in the genus *Xanthomonas*. These proteins play important roles in disease, or trigger defense, by binding host DNA and activating effector-specific host genes (*see*, e.g., Gu et al., *Nature* 435:1122-1125, 2005; Yang et al., *Proc. Natl. Acad. Sci. USA* 103:10503-10508, 2006; Kay et al. *Science* 318:648-651, 2007; Sugio et al., *Proc. Natl. Acad. Sci. USA* 104:10720-10725, 2007; and Römer et al. *Science* 318:645-648, 2007). Specificity depends on an effector-variable number of imperfect, typically 34 amino acid repeats (Schornack et al., *J. Plant Physiol.* 163:256-272, 2006; and WO 2011/072246). Polymorphisms are present primarily at repeat positions 12 and 13, which are referred to herein as the repeat variable-diresidue (RVD).

The RVDs of TAL effectors correspond to the nucleotides in their target sites in a direct, linear fashion, one RVD to one nucleotide, with some degeneracy and no apparent context dependence. This mechanism for protein-DNA recognition enables target site prediction for new target specific TAL effectors, as well as target site selection and engineering of new TAL effectors with binding specificity for the selected sites.

TAL effector DNA binding domains can be fused to other sequences, such as endonuclease sequences, resulting in chimeric endonucleases targeted to specific, selected DNA sequences, and leading to subsequent cutting of the DNA at or near the targeted sequences. Such cuts (double-stranded breaks) in DNA can induce mutations into the wild type DNA sequence via NHEJ or homologous recombination, for example. In some cases, TALE-nucleases can be used to facilitate site directed mutagenesis in complex genomes, knocking out or otherwise altering gene function with great precision and high efficiency. As described in the Examples below, TALE-nucleases targeted to the *S. tuberosum* GBSS gene can be used to mutagenize the endogenous gene, resulting in plants without detectable expression of GBSS. The

fact that some endonucleases (e.g., *FokI*) function as dimers can be used to enhance the target specificity of the TALE-nuclease. For example, in some cases a pair of TALE-nuclease monomers targeted to different DNA sequences (e.g., the underlined target sequences shown in FIG. 1; SEQ ID NO:2 and 3) can be used. When the two
 5 TALE-nuclease recognition sites are in close proximity, as depicted in FIG. 1, the inactive monomers can come together to create a functional enzyme that cleaves the DNA. By requiring DNA binding to activate the nuclease, a highly site-specific restriction enzyme can be created.

In some embodiments, a mutated *Solanum* plant, plant part, or plant cell as
 10 provided herein can have its expression of GBSS reduced by about 50 percent or more (e.g., by 60 percent or more, 70 percent or more, 80 percent or more, or 90 percent or more) as compared to a control *Solanum* plant or to a population of control *Solanum* plants that lacks the mutation(s). The control *Solanum* plant can be, for example, a corresponding wild-type version of the *Solanum* plant in which the GBSS gene was
 15 mutated.

Further, the amylose level in a mutated *Solanum* plant as provide herein can be decreased by about 1 percent or more (e.g., 1 to 5%, 5 to 10%, 10 to 20%, 20 to 30%, 30 to 40%, 40 to 50%, 50 to 60%, 60 to 70%, 70 to 80%, 80 to 90%, or 90 to 100%) as compared to the control *Solanum* plant or population of *Solanum* plants. The term
 20 “reduced amylose” or “decreased amylose” refers to a decrease in amylose levels relative to amylopectin levels within potato tuber starch. For example, starch from tubers of a wild type potato plant contains an approximate amylose/amylopectin ratio of 25/75. Starch from tubers of a potato plant with reduced amylose can have amylose/amylopectin ratios lower than 25/75 (e.g., 20/80, 15/85, 10/90, 5/95, or
 25 0/100). The amylose/amylopectin ratio can also be defined as a percentage. For example, an amylose/amylopectin ratio of 25/75, 20/80, 15/85, 10/90, 5/95, or 0/100 can also be represented as 25% amylose, 20% amylose, 15% amylose, 10% amylose, 5% amylose or 0% amylose, respectively. Thus, in some embodiments, a plant as
 30 provided herein can have 20-25%, 20-23%, or 23-25% amylose (e.g., when only one or two GBSS alleles are mutated). In some embodiments, such as when three or all four of the GBSS alleles are mutated, a plant as provided herein can have 15-20%, 10-15%, 5-10%, or 0-5% amylose. Methods for measuring amylose levels in plants are

known in the art. See, e.g., Hovenkamp-Hermelink et al., *Potato Research* 31:241-246, 1988.

In some cases, a *Solanum* plant, plant part, or plant cell as provided herein can contain a GBSS nucleotide sequence with at least about 75 percent sequence identity to a representative GBSS nucleotide sequence. For example, a nucleotide sequence
 5 can have at least 75 percent, at least 80 percent, at least 85 percent, at least 90 percent, at least 91 percent, at least 92 percent, at least 93 percent, at least 94 percent, at least 95 percent, at least 96 percent, at least 97 percent, at least 98 percent, or at least 99 percent sequence identity to a representative, naturally occurring GBSS nucleotide
 10 sequence (e.g., SEQ ID NO:4).

In some cases, a mutation can be at a target sequence as set forth in a GBSS sequence as set forth herein (e.g., SEQ ID NO:1 or SEQ ID NO:4), or at a target sequence that is at least 95 percent (e.g., at least 96 percent, at least 97 percent, at least 98 percent, or at least 99 percent) identical to the sequence set forth in a GBSS
 15 sequence as set forth herein (e.g., SEQ ID NO:1 or SEQ ID NO:4).

The percent sequence identity between a particular nucleic acid or amino acid sequence and a sequence referenced by a particular sequence identification number is determined as follows. First, a nucleic acid or amino acid sequence is compared to the sequence set forth in a particular sequence identification number using the BLAST
 20 2 Sequences (Bl2seq) program from the stand-alone version of BLASTZ containing BLASTN version 2.0.14 and BLASTP version 2.0.14. This stand-alone version of BLASTZ can be obtained online at fr.com/blast or at ncbi.nlm.nih.gov. Instructions explaining how to use the Bl2seq program can be found in the readme file accompanying BLASTZ. Bl2seq performs a comparison between two sequences
 25 using either the BLASTN or BLASTP algorithm. BLASTN is used to compare nucleic acid sequences, while BLASTP is used to compare amino acid sequences. To compare two nucleic acid sequences, the options are set as follows: -i is set to a file containing the first nucleic acid sequence to be compared (e.g., C:\seq1.txt); -j is set to a file containing the second nucleic acid sequence to be compared (e.g., C:\seq2.txt); -
 30 p is set to blastn; -o is set to any desired file name (e.g., C:\output.txt); -q is set to -l; -r is set to 2; and all other options are left at their default setting. For example, the following command can be used to generate an output file containing a comparison

between two sequences: C:\Bl2seq -i c:\seq1.txt -j c:\seq2.txt -p blastn -o c:\output.txt
 -q -1 -r 2. To compare two amino acid sequences, the options of Bl2seq are set as
 follows: -i is set to a file containing the first amino acid sequence to be compared
 (e.g., C:\seq1.txt); -j is set to a file containing the second amino acid sequence to be
 compared (e.g., C:\seq2.txt); -p is set to blastp; -o is set to any desired file name (e.g.,
 C:\output.txt); and all other options are left at their default setting. For example, the
 following command can be used to generate an output file containing a comparison
 between two amino acid sequences: C:\Bl2seq -i c:\seq1.txt -j c:\seq2.txt -p blastp -o
 c:\output.txt. If the two compared sequences share homology, then the designated
 output file will present those regions of homology as aligned sequences. If the two
 compared sequences do not share homology, then the designated output file will not
 present aligned sequences.

Once aligned, the number of matches is determined by counting the number of
 positions where an identical nucleotide or amino acid residue is presented in both
 sequences. The percent sequence identity is determined by dividing the number of
 matches either by the length of the sequence set forth in the identified sequence (e.g.,
 SEQ ID NO:1), or by an articulated length (e.g., 100 consecutive nucleotides or
 amino acid residues from a sequence set forth in an identified sequence), followed by
 multiplying the resulting value by 100. For example, a nucleic acid sequence that has
 200 matches when aligned with the sequence set forth in SEQ ID NO:1 is 83.3
 percent identical to the sequence set forth in SEQ ID NO:1 (i.e., $200 \div 240 \times 100 =$
 83.3). It is noted that the percent sequence identity value is rounded to the nearest
 tenth. For example, 75.11, 75.12, 75.13, and 75.14 are rounded down to 75.1, while
 75.15, 75.16, 75.17, 75.18, and 75.19 are rounded up to 75.2. It also is noted that the
 length value will always be an integer.

Methods for selecting endogenous target sequences and generating TALE-
 nucleases targeted to such sequences can be performed as described elsewhere. *See*,
 for example, PCT Publication No. WO 2011/072246, which is incorporated herein by
 reference in its entirety. In some embodiments, software that specifically identifies
 TALE-nuclease recognition sites, such as TALE-NT 2.0 (Doyle et al., *Nucleic Acids*
Res 40:W117-122, 2012) can be used.

Methods for using rare-cutting endonucleases (e.g., TALE-nucleases) to generate potato plants, plant cells, or plant parts having mutations in endogenous genes include, for example, those described in the Examples herein. For example, one or more nucleic acids encoding TALE-nucleases targeted to selected GBSS sequences (e.g., the GBSS sequence shown in FIG. 1) can be transformed into plant cells (e.g., protoplasts), where they can be expressed. In some cases, one or more TALE-nuclease proteins can be introduced into plant cells (e.g., protoplasts). The cells, or a plant cell line or plant part generated from the cells, can subsequently be analyzed to determine whether mutations have been introduced at the target site(s), through nucleic acid-based assays or protein-based assays to detect expression levels as described above, for example, or using nucleic acid-based assays (e.g., PCR and DNA sequencing, or PCR followed by a T7E1 assay; Mussolino et al., *Nucleic Acids Res.* 39:9283-9293, 2011) to detect mutations at the genomic loci. In a T7E1 assay, genomic DNA can be isolated from pooled calli, and sequences flanking TALE-nuclease recognition sites for GBSS can be PCR-amplified. Amplification products then can be denatured and re-annealed. If the re-annealed fragments form a heteroduplex, T7 endonuclease I cuts at the site of mismatch. The digested products can be visualized by gel electrophoresis to quantify mutagenesis activity of the TALE-nuclease.

In some embodiments, a method as provided herein can include contacting a population of *Solanum* plant cells (e.g., protoplasts) having a functional GBSS allele with a rare-cutting endonuclease that is targeted to an endogenous GBSS sequence, selecting from the population a cell in which at least one (e.g., one, two, three, or four) GBSS alleles have been inactivated, and growing the selected cell into a *Solanum* plant. The plant may have reduced amylose levels, as compared to a control *Solanum* plant that does not contain the inactivated GBSS alleles. The rare-cutting endonuclease can be introduced into the population of cells via a nucleic acid (e.g., a vector or a mRNA) that encodes the rare-cutting endonuclease, or as a protein. In some cases, a method as provided herein can include a step of culturing a plant cell containing the inactivated GBSS allele(s) to generate one or more plant lines. In addition or alternatively, a method as provided herein can include a step of isolating genomic DNA containing at least a portion of the GBSS locus from the plant cells.

In some embodiments, methods for delivering sequence-specific nucleases to a *Solanum* plant can include Agrobacterium-mediated transformation of plant parts or plant cells (e.g., leaves, stems, petiole, internode explants, callus, or protoplasts) with T-DNA encoding the sequence-specific nucleases (see, for example, Beaujean et al.,
5 *J. Exp. Biol.* 49:1589-1595, 1998), biolistic transformation of plant parts or plant cells with one or more nucleic acids encoding the sequence-specific nucleases (see, for example, Craig et al., *Plant Cell Reports* 24:603-611, 2005), and/or cell-penetrating peptide-mediated transformation of plant parts or plant cells with purified sequence-specific nucleases or nucleic acids (RNA or DNA) encoding the sequence-specific
10 nucleases (see, for example, Chen et al., *FEBS Lett.* 581:1891-1897, 2007).

In some embodiments, *Solanum* lines having mutations in one or more GBSS alleles can be generated by polyethylene glycol- (PEG-) mediated transformation. For example, protoplasts can be isolated from surface sterilized leaves, and transformed in the presence of PEG with plasmids encoding one or more sequence specific
15 nucleases. Transformation efficiencies can be monitored by delivery of a detectable marker such as a YFP plasmid, which can be visualized using fluorescence microscopy or flow cytometry. After PEG-mediated transformation, protoplasts can be cultured using methods and media described elsewhere (see, e.g., Gamborg et al., in: Plant Tissue Culture Methods and Applications in Agriculture, Thorpe (Ed.),
20 Academic Press, Inc., New York, NY, 1981, pp.115-153). After a suitable length of time in culture, protoplast-derived calli identified as mutants can be grown, transferred to shoot-inducing medium, and then (once roots form) transferred to soil and grown to maturity for tuber production (see, e.g., Haberlach et al., *supra*; and Gamborg et al., *supra*).

25 In some embodiments, delivery of one or more sequence-specific nucleases to a *Solanum* plant can be achieved through transient delivery or stable integration into the host genome. To transiently deliver sequence-specific nucleases, transformed potato plant parts or plant cells (using the above-described methods) can be placed on regeneration medium containing no selective agent, and potato plants can be
30 regenerated. Regenerated plants then can be screened to identify those containing nuclease-induced mutations. To stably integrate the genome engineering reagents into the host genome, nucleic acids encoding the sequence-specific nucleases can be co-

delivered with nucleic acid encoding a plant selectable marker (e.g., kanamycin [nptII], hygromycin [hpt], methotrexate [dhfr], phosphinothricin [bar], or phleomycin [ble]). The selectable marker can be harbored on the same vector as the sequence-specific nuclease(s), or can be delivered as a separate vector. After transformation,
5 potato plant parts or plant cells can be placed on regeneration medium containing the appropriate selectable agent, and transgenic potato plants can be regenerated.

In some embodiments, a nuclease can be co-delivered to a plant cell with a plasmid encoding one or more exonuclease proteins to increase sequence specific nuclease induced mutagenesis efficiency. Such exonucleases include, without
10 limitation, members of the TREX (Therapeutic red cell exchange exonucleases) family of exonucleases, such as TREX2 (Shevelev et al., *Scientific World Journal* 2:275-281, 2002). Co-delivery of an exonuclease such as TREX with one or more rare-cutting endonucleases (e.g., TALE-nucleases) may increase the frequency of NHEJ events, as compared with the frequency of NHEJ events after delivery of the
15 rare-cutting endonuclease(s) alone. It is to be noted that other exonucleases also can be used in the methods provided herein.

Another genome engineering tool that can be used in the methods provided herein is based on the RNA-guided Cas9 nuclease from the type II prokaryotic CRISPR (Clustered Regularly Interspaced Short palindromic Repeats) adaptive
20 immune system (see, e.g., Belahj et al., *Plant Methods* 9:39, 2013). This system allows for cleavage of DNA sequences that are flanked by a short sequence motif, referred as proto-spacer adjacent motif (PAM). Cleavage is achieved by engineering a specific crRNA that is complementary to the target sequence. The crRNA associates into a living cell with a heterologously expressed Cas9 endonuclease from
25 *Streptococcus pyogenes*. In the crRNA/Cas9 complex, a dual tracrRNA:crRNA structure acts as a guide RNA that directs the Cas9 endonuclease to the cognate target sequence. Since several PAM motifs are present in the nucleotide sequence of the GBSS gene, crRNA specific to GBSS gene can be designed to introduce mutations or to inactivate one or more GBSS alleles within *Solanum* plant cells into which the
30 Cas9 endonuclease and the crRNA are transfected and then expressed. In some embodiments, therefore, this approach can be used to obtain GBSS mutant plants as described herein.

In some embodiments, the plants provided herein can contain further mutations introduced into other *Solanum* genes. Such mutations can, for example:

- provide acrylamide reduction by modifying the expression of genes involved in asparagine synthesis;

- 5 - prevent black spot bruise by reducing polyphenol oxidase-5 expression;
- prevent Potato Virus Y by reducing eIF4E gene expression;
- prevent late blight; and/or
- improve nematode, herbicide, or insect resistance.

Thus, the methods provided herein can be used to obtain gene stacking in a *Solanum* trait.

This document also provides methods for producing starch products using potato plant varieties with reduced amylose, as well as industrial starch products made by such methods. These products include, without limitation, starches useful in the paper, textile, adhesive, and packing industries. Such starch products can be produced using various procedures and types of equipment, although all follow a similar process. For example, potatoes can be dropped into water flumes that clean the potatoes of stones and dirt. After further cleaning in a washer, the potatoes can be moved to a grinder or crusher to liberate the starch from the potato cells. The resulting slurry can be passed through a screen or rotary sieve to separate the fiber and potato skins, and the starch solution can then be further purified to remove soluble and insoluble impurities by alternate cycles of filtration and redispersion in water. The purified starch then can be dewatered, dried, and collected. *See*, for example, Robson, “U.S. Environmental Protection Agency. Starch Manufacturing: A Profile,” North Carolina: Center for Economics Research, March 1994 (RTI Project Number: 35U-5681-71 DR).

The invention will be further described in the following examples, which do not limit the scope of the invention described in the claims.

EXAMPLES

30 Example 1 – Engineering sequence-specific nucleases to mutagenize the GBSS gene

To completely inactivate or knock-out the alleles of the GBSS gene in *S. tuberosum*, sequence-specific nucleases were designed to target the protein coding

region in the first exon. In particular, a pair of TALE-nucleases (designated as GBSS_T1) was designed to target the GBSS gene family within the first 150 bp of the coding sequence, using software that specifically identifies TALE-nuclease recognition sites. The TALE-nuclease recognition sites for the GBSS genes are underlined in FIG. 1 and are listed in Table 1 (SEQ ID NOS:2 and 3). TALE-nucleases were synthesized using methods similar to those described elsewhere (Cermak et al., *Nucleic Acids Res.* 39:e82, 2011; Reyon et al., *Nat. Biotechnol.* 30:460-465, 2012; and Zhang et al., *Nat. Biotechnol.* 29:149-153, 2011).

10 Example 2 – Activity of GBSS TALE-nucleases at their endogenous target sites in *S. tuberosum*

GBSS_T1 activity at endogenous target sites in *S. tuberosum* was measured by expressing the TALE-nucleases in protoplasts and subsequently surveying the target sites for mutations introduced by NHEJ. Methods for protoplast preparation were performed as described elsewhere (Shepard, in: Genetic Improvement of Crops/Emergent Techniques, Rubenstein, Gengenbach, Philips, and Green (Eds.), Univ. of Minnesota Press, Minneapolis, MN, 1980, pp.185-219); and Shepard and Totten, *Plant Physiol.* 60:313-316, 1977). Briefly, *S. tuberosum* mini tubers were planted in moistened vermiculite and grown under low light conditions for 3-5 weeks. Young, fully expanded leaves were collected and surface sterilized, and protoplasts were isolated.

TALE-nuclease-encoding plasmids, together with a yellow fluorescent protein- (YFP-) encoding plasmid, were introduced into *S. tuberosum* protoplasts by polyethylene glycol- (PEG-) mediated transformation (Yoo et al., *Nature Protocols* 2:1565-1572, 2007). In some experiments, a plasmid encoding a TREX2 exonuclease was co-delivered with the TALE-nuclease-encoding plasmid. Twenty-four hours after treatment, transformation efficiency was measured using a fluorescent microscope to monitor YFP fluorescence in an aliquot of the transformed protoplasts. The remainder of the transformed protoplasts was harvested, and genomic DNA was prepared using a hexadecyltrimethylammonium bromide- (CTAB-) based method. Using genomic DNA prepared from the protoplasts as a template, a 491-bp fragment encompassing the TALE-nuclease recognition site was amplified by PCR. Allele

types were analyzed by individual clonal direct sequencing and 454 pyro-sequencing. Sequencing reads with indel mutations in the spacer region were considered to be derived from imprecise repair of a cleaved TALE-nuclease recognition site by NHEJ. Mutagenesis frequency was calculated as the number of sequencing reads with NHEJ mutations out of the total sequencing reads.

The activity of the GBSS TALE-nuclease pair, with or without TREX2, is summarized in Table 2. The TALE-nucleases induced NHEJ mutations in GBSS_T1 in 11% to 23% of the sequencing reads. Examples of TALE-nuclease-induced mutations in GBSS_T1 are shown in FIG. 3.

Example 3 – Regeneration of *S. tuberosum* lines with TALE-nuclease-induced GBSS mutations

Transgenic potato plant lines encoding the GBSS_T1 TALE-nuclease pair, were generated by Agrobacterium-mediated transformation of internodes. Briefly, the coding sequence for the GBSS_T1 TALE-nuclease pair was cloned into T-DNA. Methods for transforming potato using Agrobacterium-mediated transformation of internodes were followed as previously described (Beaujean et al., *supra*).

Example 4 – Verification of *S. tuberosum* lines with TALE-nuclease-induced GBSS mutations

Transgenic potato plants encoding the GBSS_T1 TALE-nuclease pair were assessed for mutations at the GBSS_T1 target site. To this end, genomic DNA was isolated from leaf tissue using a CTAB-based method (Murray and Thompson, *Nucl. Acids Res.* 8:4321-4326, 1980). The isolated genomic DNA was then used as a template in a PCR reaction with primers designed to amplify the TALE-nuclease target site. The resulting amplicons were cloned and sequenced. Seven plants were identified that contained mutations in at least one GBSS allele. A list of the GBSS-mutant plant genotypes are shown in (FIGS. 4A, 4B, and 5).

Example 5 – Determining whether mutant *S. tuberosum* lines have desired phenotypes

GBSS transcript quantification is determined using quantitative real-time PCR with cDNA generated from mutant and control tuber mRNA extracts (Bhaskar et al.,

Plant Physiol. 154(2):939-948, 2010). The level and any reduction of GBSS expression is quantified using a comparative cycle threshold method (Livak and Schmittgen, *Method. Methods* 25:402-408, 2001). Amylose levels are assessed using methods and media described elsewhere (Hovenkamp-Hermelink et al., *supra*), and/or
 5 using the amylose/amylopectin assay kit (K-AMYL; Megazyme, Ireland).

Tubers from candidate plants harboring mutations in the GBSS alleles were assessed for amylose levels using the K-AMYL amylose/amylopectin assay kit (Megazyme). Briefly, approximately 1 gram of potato tuber was ground to a paste using a mortar and pestle. About 50 mg of material was collected in a glass test tube
 10 and 1 mL of DMSO was added. The samples were vortexed and boiled for about 15 minutes. The samples were then incubated at room temperature for 5 minutes. Starch was precipitated from the samples by adding 6 mL of 95% ethanol. Samples were centrifuged at 2,000 g for 5 minutes and supernatant was removed. After allowing the ethanol to evaporate, the starch was dissolved in 2 mL of DMSO and the tubes were
 15 placed in a boiling water bath for 15 minutes. Following boiling, the samples were diluted in a concanavalin A (ConA) buffer to a total volume of 25 mL. The amylose and amylopectin content of this “total starch” solution was determined by adding 0.1 mL of an amyloglucosidase/alpha-amylase solution to 0.5 mL of the total starch solution and incubating the resulting solution for 10 minutes at 40°C. Next, 4 mL of a
 20 GOPOD reagent containing glucose oxidase plus peroxidase and 4-aminoantipyrine was added to the total starch solution. The total starch content was measured colorimetrically using a plate reader at 510 nm.

To determine the fraction of amylose within the total starch, 0.50 mL of the solution containing ConA was added to 1 mL of the total starch solution, which was
 25 then incubated at room temperature for 1 hour. The solution was boiled for 5 minutes to inactivate ConA, and the resulting sample was incubated at 40°C for 5 minutes, and then 0.1 mL of the amyloglucosidase/alpha-amylase solution was added. The solution was incubated at 40°C for 30 minutes. The sample was centrifuged at 2,000 g for 5 minutes and the supernatant was collected. To 1 mL of supernatant, 4 mL of the
 30 GOPOD reagent was added, and the sample was incubated at 40°C for 20 minutes. The amylose fraction was measured colorimetrically using a plate reader at 510 nm. The percentage of amylose was determined using the following equation: (510 nm

absorbance of the amylose supernatant)/(510 nm absorbance of the total starch supernatant) x 6.15/9.2 x 100/1.

The results of the amylose/amylopectin kit are shown in FIGS. 6 and 7. Tuber starch from plant lines St226-1, St226-2, St226-4, St226-5, St226-6 and St226-9 all contained amylose/amylopectin ratios similar to the tuber starch from wild type plants. Specifically, St226-1 contained 23.0% amylose and 77% amylopectin; St226-4 contained 27.3% amylose and 72.7% amylopectin; St226-5 contained 25.0% amylose and 75% amylopectin; St226-6 contained 27.9% amylose and 72.1% amylopectin; St226-9 contained 23.3% amylose and 76.7% amylopectin; wild type potato plants contained 28.7% amylose and 71.3% amylopectin. The genotype data presented in FIG. 4, along with the phenotype data presented in FIG. 6, suggest that potato plants containing one or more wild type GBSS alleles can produce tuber starch that has an amylose/amylopectin ratio similar to tuber starch from an unmodified, wild type plant.

While tubers from plant lines St226-1, St226-2, St226-4, St226-5, St226-6 and St226-9 all contained amylose/amylopectin ratios similar to tubers from wild type plants, plant line St226-8 produced specific tubers that contained substantially less amylose than tubers from WT plants (FIG. 7). Specifically, St226-8 Tuber 1 contained 22.7% amylose and 77.3% amylopectin; St226-8 Tuber 2 contained 16.0% amylose and 84.0% amylopectin; St226-8 Tuber 3 contained 12.6% amylose and 87.4% amylopectin; St226-8 Tuber 4 contained 3.7% amylose and 96.3% amylopectin; St226-8 Tuber 5 contained 22.7% amylose and 77.3% amylopectin; wild type potato tubers contained 23.7% amylose and 76.3% amylopectin. These results indicate that TALE-nuclease-induced mutations within the GBSS gene can result in potato tubers with reduced amylose, as compared to potato tubers from unmodified plants. Further, these results indicate that at least three mutant GBSS alleles are necessary to obtain potato tubers with reduced amylose, as compared to potato tubers from unmodified plants.

Table 1*TALE-nuclease target sequences in GBSS_T1*

Target Sequence Left	SEQ ID NO:	Target Sequence Right	SEQ ID NO:
TCTGACTCACAATGGTT	2	AGCTTGATGGGCTCCAA	3

5

Table 2*Sequencing Data for GBSS TALE-nuclease*

Nucleases tested	Location of target site	NHEJ mutagenesis frequency
GBSS_T1	GBSS_T1	11%
GBSS_T1 + TREX2	GBSS_T1	23%

10

OTHER EMBODIMENTS

It is to be understood that while the invention has been described in conjunction with the detailed description thereof, the foregoing description is intended to illustrate and not limit the scope of the invention, which is defined by the scope of the appended claims. Other aspects, advantages, and modifications are within the scope of the following claims.

15

WHAT IS CLAIMED IS:

1. A *Solanum* plant, plant part, or plant cell comprising a mutation in at least three granule-bound starch synthase (GBSS) allele endogenous to said plant, plant part, or plant cell, such that said plant, plant part, or plant cell has reduced expression of GBSS and decreased levels of amylose as compared to a control *Solanum* plant, plant part, or plant cell that lacks said mutation.
2. The plant, plant part, or plant cell of claim 1, wherein each said mutation is a deletion of more than one nucleotide base pair.
3. The plant, plant part, or plant cell of claim 1, wherein each said mutation is at a target sequence as set forth in SEQ ID NO:4, or at a target sequence having at least 95 percent identity to SEQ ID NO:4.
4. The plant, plant part, or plant cell of claim 1, wherein each said mutation is at a target sequence as set forth in SEQ ID NO:1, or at a target sequence having at least 95 percent identity to SEQ ID NO:1.
5. The plant, plant part, or plant cell of claim 1, wherein said plant, plant part, or plant cell was made using a rare-cutting endonuclease.
6. The plant, plant part, or plant cell of claim 5, wherein said rare-cutting endonuclease is a transcription activator-like effector endonuclease (TALE-nuclease).
7. The plant, plant part, or plant cell of claim 6, wherein said TALE-nuclease binds to a sequence as set forth in SEQ ID NO:2 or SEQ ID NO:3.
8. The plant, plant part, or plant cell of claim 1, wherein each of said at least three GBSS alleles exhibits removal of an endogenous nucleic acid and does not include any exogenous nucleic acid.
9. The plant, plant part, or plant cell of claim 1, wherein every endogenous GBSS allele is mutated.

10. The plant, plant part, or plant cell of claim 9, wherein every endogenous GBSS allele exhibits removal of an endogenous nucleic acid and does not include any exogenous nucleic acid.
11. The plant, plant part, or plant cell of claim 9, wherein said plant, plant part, or plant cell has no detectable expression of GBSS.
12. The plant, plant part, or plant cell of claim 1, wherein said *Solanum* plant, plant part, or plant cell is a *S. tuberosum* plant, plant part, or plant cell.
13. A method for making a *Solanum* plant, comprising:
 - (a) contacting a population of *Solanum* plant cells comprising a functional GBSS allele with a rare-cutting endonuclease targeted to an endogenous GBSS sequence,
 - (b) selecting, from said population, a cell in which at least three GBSS alleles have been inactivated, and
 - (c) growing said selected plant cell into a *Solanum* plant, wherein said *Solanum* plant has reduced levels of amylose as compared to a control *Solanum* plant in which said at least three GBSS alleles have not been inactivated.
14. The method of claim 13, wherein said *Solanum* plant cells are protoplasts.
15. The method of claim 14, comprising transforming said protoplasts with a nucleic acid encoding said rare-cutting endonuclease.
16. The method of claim 15, wherein said nucleic acid is an mRNA.
17. The method of claim 15, wherein said nucleic acid is contained within a vector.
18. The method of claim 14, comprising introducing into said protoplasts a rare-cutting endonuclease protein.
19. The method of claim 13, wherein said rare-cutting endonuclease is a TALE-nuclease.

20. The method of claim 19, wherein said TALE-nuclease is targeted to a sequence as set forth in SEQ ID NO:4, or to a sequence having at least 95 percent identity to the sequence set forth in SEQ ID NO:4.
21. The method of claim 19, wherein said TALE-nuclease is targeted to a sequence as set forth in SEQ ID NO:1, or to a sequence having at least 95 percent identity to the sequence set forth in SEQ ID NO:1.
22. The method of claim 19, wherein said TALE-nuclease binds to a sequence as set forth in SEQ ID NO:2 or SEQ ID NO:3.
23. The method of claim 14, further comprising culturing said protoplasts to generate plant lines.
24. The method of claim 14, comprising isolating genomic DNA comprising at least a portion of the GBSS locus from said protoplasts.
25. The method of claim 13, wherein said *Solanum* plant cells are *S. tuberosum* plant cells.
26. A method for producing a starch product, comprising:
(a) providing a *Solanum* plant or plant part that comprises a mutation in at least three GBSS alleles endogenous to said plant or plant part, such that said plant, plant part, or plant cell has reduced expression of granule-bound starch synthase and decreased levels of amylose as compared to a control *Solanum* plant or plant part that lacks said mutation; and
(b) producing a starch product from said plant or plant part.
27. The method of claim 26, wherein each said mutation is at a target sequence as set forth in SEQ ID NO:4, or at a target sequence having at least 95 percent identity to SEQ ID NO:4.
28. The method of claim 26, wherein each said mutation is at a target sequence as set forth in SEQ ID NO:1, or at a target sequence having at least 95 percent identity to SEQ ID NO:1.

29. The method of claim 26, wherein each said mutation was made using a rare-cutting endonuclease.
30. The method of claim 29, wherein said rare-cutting endonuclease is a TALE-nuclease.
31. The method of claim 30, wherein said TALE-nuclease binds to a sequence as set forth in SEQ ID NO:2 or SEQ ID NO:3.
32. The method of claim 26, wherein said *Solanum* plant or plant part is a *S. tuberosum* plant or plant part.
33. The method of claim 26, wherein said *Solanum* plant or plant part has no detectable expression of granule-bound starch synthase.
34. A starch product produced from a *Solanum* plant or plant part that comprises a mutation in each GBSS allele endogenous to said plant or plant part, such that said plant, plant part, or plant cell has no functional GBSS allele, wherein said starch product has reduced levels of amylose as compared to a starch product made from a control plant or plant part that lacks said mutation.
35. The starch product of claim 34, wherein each said mutation is at a target sequence as set forth in SEQ ID NO:4, or at a target sequence having at least 95 percent identity to SEQ ID NO:4.
36. The starch product of claim 34, wherein each said mutation is at a target sequence as set forth in SEQ ID NO:1, or at a target sequence having at least 95 percent identity to SEQ ID NO:1.
37. The starch product of claim 34, wherein each said mutation was made using a rare-cutting endonuclease.
38. The starch product of claim 37, wherein said rare-cutting endonuclease is a TALE-nuclease.

39. The starch product of claim 38, wherein said TALE-nuclease binds to a sequence as set forth in SEQ ID NO:2 or SEQ ID NO:3.

40. The starch product of claim 34, wherein said *Solanum* plant or plant part is a *S. tuberosum* plant or plant part.

FIG. 1

GBSS_T1
actctgactcaccaatgggtttaagggctgttaaccaagcttgatgggctccaatcaagaactaataactaaggtaacacccaag
atggcatccagaactgagaccagagacctgggatgctcagctaccattgtttgtggaagggaatgaacttgatcttgtg
ggtactgagggttggtccttgagcaaaactgggtggactaggtgatgttcttgggtggactaccaccagcccttgtagta
(SEQ ID NO:1)

FIG. 2

S. tuberosum GBSS complete CDS; GenBank EU403426.2
ATGGCAAGCATCACAGCTTCACACCACTTTGTGTCAAGAAAGCCAAACTTCAC TAGACACCAAATCAACCTTGTCACAGATA
GGACTCAGGAACCATACTCTGACTCACAAATGGTTAAGGGCTGTTAACAAGCTTGATGGGCTCCAATCAAGAATAACTACT
AAGGTAACACCCCAAGATGGCATCCAGAACTGAGACCAAGAGACCTGGATGCTCAGCTACCATTTGTTGTGGAAGGGAATG
AACTTGATCTTTGTGGTACTGAGGTTGGTCCTTGGAGCAAAACTGGTGACTAGGTGATGTTCTTGGTGACTACCACCA
GCCCTTGAGCCCGGACATCGGGTAATGACAAATATCCCCCGTTATGACCAATAACAAAGATGCTTGGGATACTAGCGTT
GCGGTTGAGGTCAAAGTTGGAGACAGCATTGAAAATGTTTCGTTTCTTCACTGCTATAAAACGTGGGTTGATCGTGTTTTT
GTTGACCACCCAATGTTCTTGGAGAAAAGTTTGGGGCAAAACTGGTTCAAAAATCTATGGCCCCAAAGCTGGACTAGATTAT
CTGGACAAATGAACCTTAGGTTCAGCTTGTTGTCAAGCAGCCCTAGAGGCACCTAAAGTTTGTGAATTTGAACAGTAGCAAC
TACTTCTCAGGACCATATGGAGAGGATGTTCTCTTCATTGCCAATGATTGGCACACAGCTCTCATTCCTTGCTACTTGAAG
TCAAATGTACCAGTCCAGAGGAATCTATTGAATGCCAAGGTCGCTTTCCTGCATCCATAACATTGCCCTACCAAGGCCGATTT
TCTTTCTCTGACTTCCCCTCTTCTCAATCTTCCCTGATGAATTCAGGGTTCCTTTGATTTCAATTGATGGATATGAGAAGCCT
GTTAAGGGTAGGAATAACAACCTGGATGAAGGCTGGGATATTAGAATCACATAGGTGGTTACAGTGAGCCCATACTATGCC
CAAGAACTTGCTCTGCTGTTGACAAGGTGTTGAATTGGACAGTGTCCTTCGTAAGACTTGCAATAACTGGGATTGTGAAT
GGCATGGATACACAAGAGTGGAACCCAGCGACTGACAAATACACAGATGTCAAATACGATATAACCACTGTCATGGACGCA
AAACCTTTACTAAAGGAGGCTCTTCAAGCAGCAGTTGGCTTGCCTGTTGACAAGAAGATCCCCTTTGATTGGCTTCATCGGC
AGACTTGAGGAGCAGAAAAGGTTTCAAGATATTCTTGTGCTGCAATTACAAAGTTTCATCGGATTGGATGTTCAAATTTAGTCTC
CTTGGAACCTGGCAAAAAGGAGTTTGAGCAGGAGATTGAACAGCTCGAAGTGTGTACCCTAACAAAGCTAAAAGGAGTGGCA
AAATTCAATGTCCCCTTTGGCTCACATGATCACTGCTGGTGCTGATTTTATGTTGTTCCAAAGCAGATTTGAAACCTTGTGGT
CTCATTCAGTTACATGCTATGCCGATATGGAACAGTGCCAAATCTGTGCATCGACTGGTGACTTGTGACACTGTGAAAGAA
GGCTATACTGGATTCCATATGGGAGCCCTTCAATGTTGAATGCGATGTTGTTGACCCAGCTGATGTGCTTAAGATAGTAACA
ACAGTTGCTAGAGCTCTTGCAGTCTATGGCACCCCTCGCATTTGCTGAGATGATAAAAAATTGCATGTGAGAGAACTCTCC
TGGAAGGAACCTGCCAAGAAATGGGAGACATTGCTATTGGGCTTAGGAGCTTCTGGCAGTGAACCCGGTGTGAAGGGGAA
GAAATCGCTCCACTTGCCCAAGGAAAATGTAGCCACTCCCTAA (SEQ ID NO:4)

FIG. 3

GBSS_T1 2dpt	
Sequence	
<u>TCTGACTCACAAATGGTTTAAGGGCTGTTAACAAGCTTGATGGGCTCCAA</u>	Number of SEQ ID
TCTGACTCACAAATGGTTTAAGGT-----AACAAGCTTGATGGGCTCCAA	Deletions NO:
TCTGACTCACAAATGGTTTAATTA-----ACAAGCTTGATGGGCTCCAA	0 5
TCTGACTCACAAATGGTTTA-----TAACAAGCTTGATGGGCTCCAA	5 6
TCTGACTCACAAATGGTTTA-----ACAAGCTTGATGGGCTCCAA	6 7
TCTGACTCACAAATGGTTTA-----TAACAAGCTTGATGGGCTCCAA	8 8
TCTGACTCACAAATGGTTTA-----ACAAGCTTGATGGGCTCCAA	10 9
GBSS_T1 + TREX2 2dpt	
Sequence	
<u>TCTGACTCACAAATGGTTTAAGGGCTGTTAACAAGCTTGATGGGCTCCAA</u>	Number of SEQ ID
TCTGACTCACAAATGGTTTAAG-----TAACAAGCTTGATGGGCTCCAA	Deletions NO:
TCTGACTCACAAATGGTTTAAGGA-----CAAGCTTGATGGGCTCCAA	0 5
TCTGACTCACAAATGGTTT-----TAACAAGCTTGATGGGCTCCAA	6 10
TCTGACTCACAAATGGTTT-----AACGAGCGTGATGGGCTCCGA	7 11
	9 12
	10 13

FIG. 4A

Plant: St226-1					
<u>TCTGACTCACAAATGGTTTAAGGGCTGTTAACAAAGCTTGATGGGCTCCAA</u>	WT	3/7	5		
TCTGACTCACAAATGGTTTAAGGGCT---AACAAAGCTTGATGGGCTCCAA	-3	1/7	14		
TCTGACTCACAAATGGTTTAAGGGCT-----GTTTGATGGGCTCCAA	-8	1/7	15		
TCTGACTCACAAATGGTTTAAGGG-----AACAAAGCTTGATGGGCTCCAA	-5	2/7	16		
Plant: St226-2					
<u>TCTGACTCACAAATGGTTTAAGGGCTGTTAACAAAGCTTGATGGGCTCCAA</u>	WT	3/7	5		
TCTGACTCACAAATGGTTTAAGGGCT---AACAAAGCTTGATGGGCTCCAA	-3	2/7	14		
TCTGACTCACAAATGGTTTAAGGGCT-----GTTTGATGGGCTCCAA	-8	2/7	15		
Plant: St226-4					
<u>TCTGACTCACAAATGGTTTAAGGGCTGTTAACAAAGCTTGATGGGCTCCAA</u>	WT	4/8	5		
TCTGACTCACAAATGGTTTAAGGGCT-----GTTTGATGGGCTCCAA	-8	3/8	15		
TCTGACTCACAAATGGTTTAAGGG-----AACAAAGCTTGATGGGCTCCAA	-5	1/8	16		

FIG. 4B

Plant: St226-5				
TCTGACTCACAAATGGTTTAAAGGGCTGTTAAAGGCTTGATGGGCTCCAA	WT	Number of reads/total reads	SEQ ID NO:	5
<u>TCTGACTCACAAATGGTTTAAAGGGCT</u> ---AACAAAGCTTGATGGGCTCCAA	-3	1/8		
TCTGACTCACAAATGGTTTAAAGGGCT-----GTTTGATGGGCTCCAA	-8	1/8		14
TCTGACTCACAAATGGTTTAAAGGGCT-----GTTTGATGGGCTCCAA	-5	4/8		15
TCTGACTCACAAATGGTTTAAAGGG-----AACAAAGCTTGATGGGCTCCAA		2/8		16
Plant: St226-6				
TCTGACTCACAAATGGTTTAAAGGGCTGTTAAAGGCTTGATGGGCTCCAA	WT	Number of reads/total reads	SEQ ID NO:	5
<u>TCTGACTCACAAATGGTTTAAAGGGCT</u> ---AACAAAGCTTGATGGGCTCCAA	-3	3/8		
TCTGACTCACAAATGGTTTAAAGGGCT-----GTTTGATGGGCTCCAA	-8	2/8		14
TCTGACTCACAAATGGTTTAAAGGGCT-----GTTTGATGGGCTCCAA	-5	1/8		15
TCTGACTCACAAATGGTTTAAAGGG-----AACAAAGCTTGATGGGCTCCAA		2/8		16
Plant: St226-9				
TCTGACTCACAAATGGTTTAAAGGGCTGTTAAAGGCTTGATGGGCTCCAA	WT	Number of reads/total reads	SEQ ID NO:	5
<u>TCTGACTCACAAATGGTTTAAAGGGCT</u> ---AACAAAGCTTGATGGGCTCCAA	-3	1/8		
TCTGACTCACAAATGGTTTAAAGGGCT-----GTTTGATGGGCTCCAA	-8	4/8		14
TCTGACTCACAAATGGTTTAAAGGGCT-----GTTTGATGGGCTCCAA	-5	2/8		15
TCTGACTCACAAATGGTTTAAAGGG-----AACAAAGCTTGATGGGCTCCAA		1/8		16

FIG. 5

Plant: St226-8		Number of reads/total reads	SEQ ID NO:
<u>TCTGACTCACAAATGGTTTAAGGGCT</u> ----- <u>AACAAGCTTGATGGGCTCCAA</u>	-3	2/7	14
TCTGACTCACAAATGGTTTAAGGGCT-----GTTTGATGGGCTCCAA	-8	3/7	15
-----AAGCTTGATGGGCTCCAA	-31	2/7	17

FIG. 6

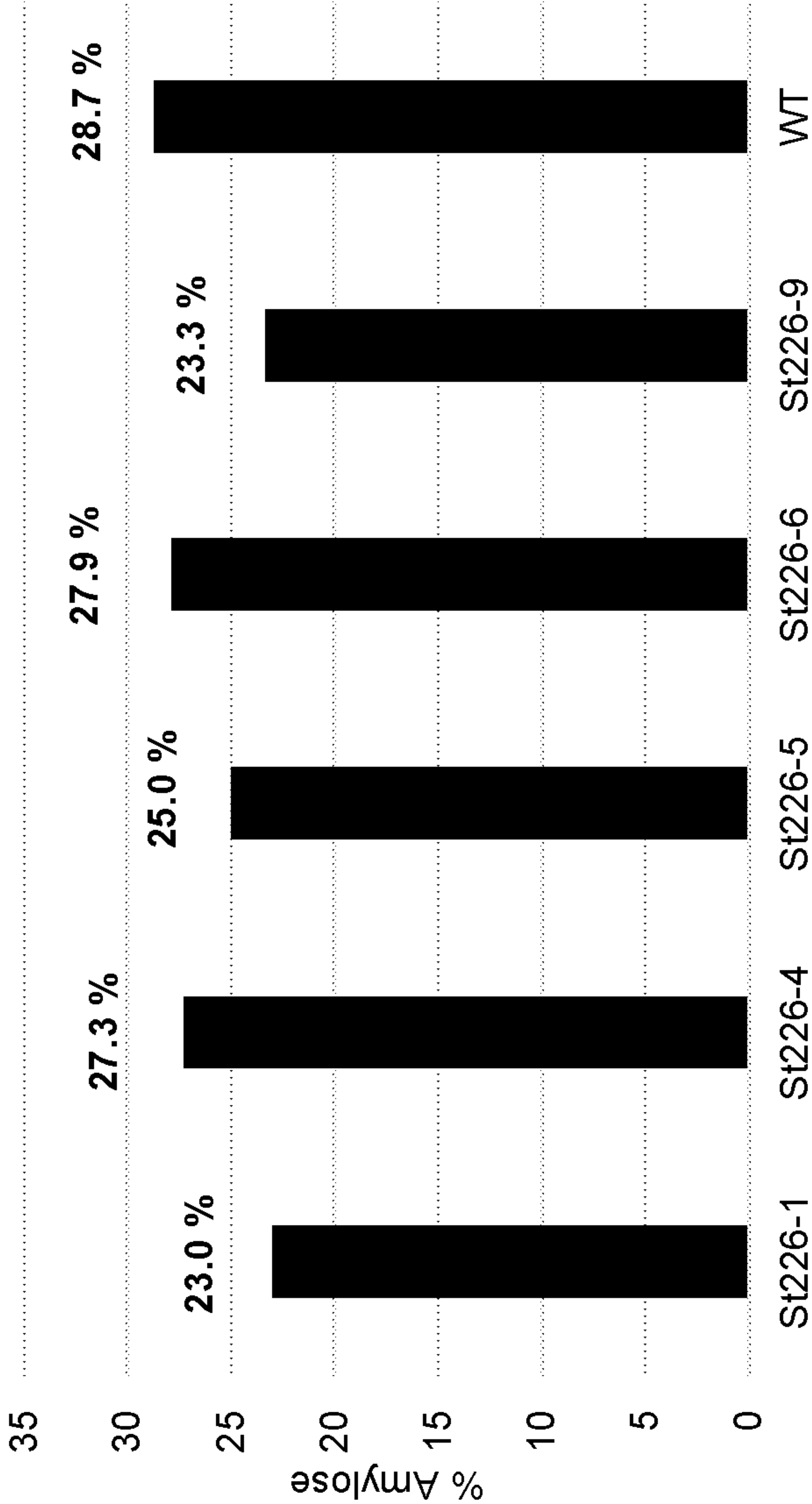


FIG. 7

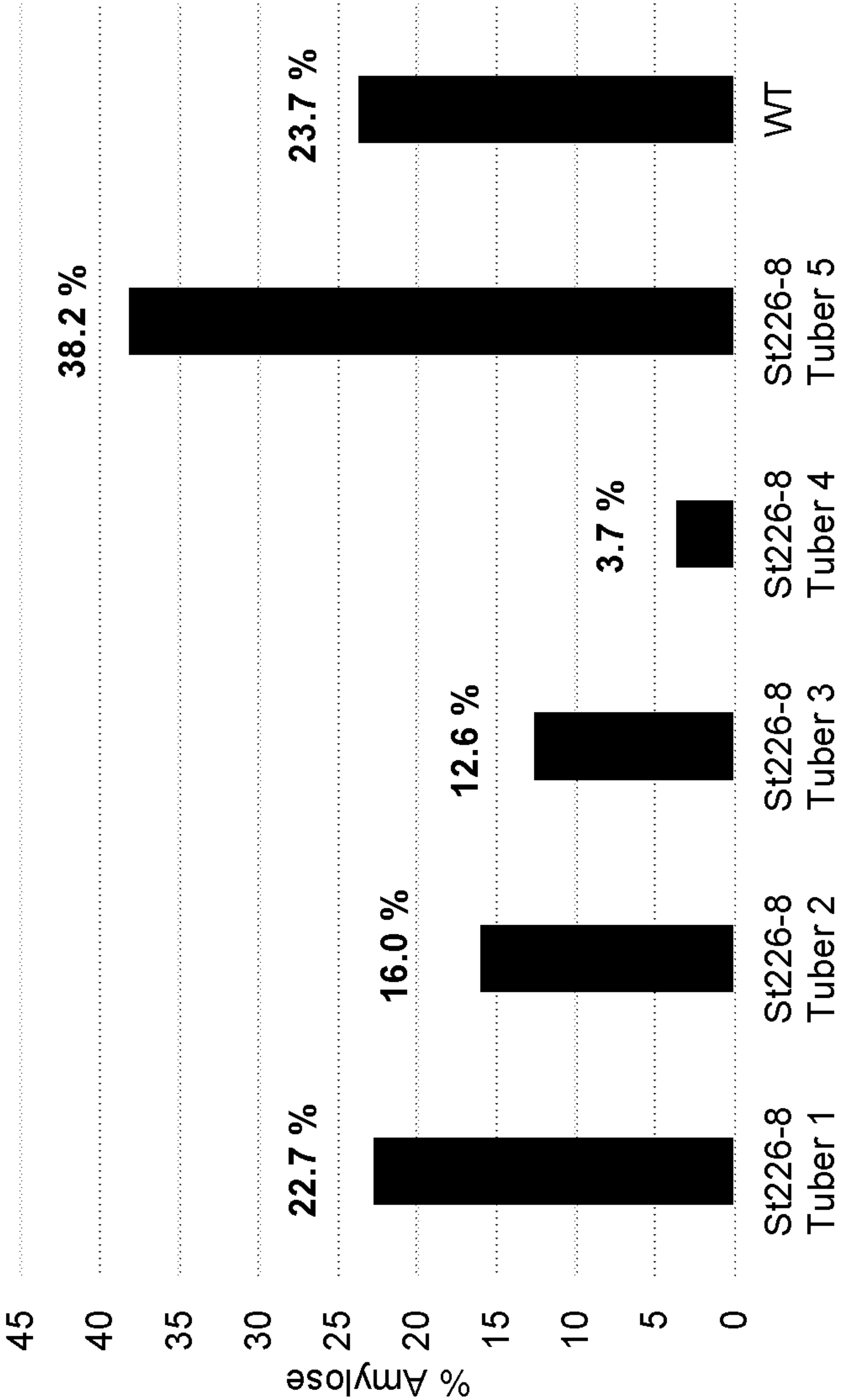


FIG. 1

GBSS_T1

actctgactcacaatggtttaagggctgttaacaagcttgatgggctccaatcaagaactaataactaaggtaacacccaag
atggcatccagaactgagaccaagagacctggatgctcagctaccattgtttgtggaaaggggaatgaacttgatctttgtg
ggtactgagggttggtccttggagcaaaactggtggactaggtgatgttcttgggtggactaccaccagcccttgcagta
(SEQ ID NO:1)