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(54) RECOMBINANT NARBONOLIDE POLYKETIDE SYNTHASE

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## ABSTRACT

Recombinant DNA compounds that encode all or a portion of the narbonolide polyketide synthase are used to express recombinant polyketide synthase genes in host cells for the production of narbonolide, narbonolide derivatives, and polyketides that are useful as antibiotics and as intermediates in the synthesis of compounds with pharmaceutical value.





Figure 1


Figure 2


Figure 3


A(b)

$\stackrel{5}{4}$
$\begin{aligned} & \text { S. venezuelae } \\ & \text { ATCC15439 }\end{aligned}$
pKOS023-27 pKOS023-27
S. venezuelae
K:039-03

$$
\xrightarrow{\text { picAlli }} \text { picAIV picB picCIII }
$$



Figure 5


Figure 6

## RECOMBINANT NARBONOLIDE POLYKETIDE SYNTHASE

## REFERENCE TO GOVERNMENT FUNDING

[0001] This invention was supported in part by SBIR grant 1R43-CA75792-01. The U.S. government has certain rights in this invention.

## FIELD OF THE INVENTION

[0002] The present invention provides recombinant methods and materials for producing polyketides by recombinant DNA technology. The invention relates to the fields of agriculture, animal husbandry, chemistry, medicinal chemistry, medicine, molecular biology, pharmacology, and veterinary technology.

## BACKGROUND OF THE INVENTION

[0003] Polyketides represent a large family of diverse compounds synthesized from 2-carbon units through a series of condensations and subsequent modifications. Polyketides occur in many types of organisms, including fungi and mycelial bacteria, in particular, the actinomycetes. There are a wide variety of polyketide structures, and the class of polyketides encompasses numerous compounds with diverse activities. Tetracycline, erythromycin, FK506, FK520, narbomycin, picromycin, rapamycin, spinocyn, and tylosin, are examples of such compounds. Given the difficulty in producing polyketide compounds by traditional chemical methodology, and the typically low production of polyketides in wild-type cells, there has been considerable interest in finding improved or alternate means to produce polyketide compounds. See PCT publication Nos. WO 93/13663; WO 95/08548; WO 96/40968; 97/02358; and 98/27203; U.S. Pat. Nos. $4,874,748 ; 5,063,155 ; 5,098,837$; 5,149,639; 5,672,491; and 5,712,146; Fu et al., 1994, Biochemistry 33: 9321-9326; McDaniel et al., 1993, Science 262: 1546-1550; and Rohr, 1995, Angew. Chem. Int. Ed. Engl. 34(8): 881-888, each of which is incorporated herein by reference.
[0004] Polyketides are synthesized in nature by polyketide synthase (PKS) enzymes. These enzymes, which are complexes of multiple large proteins, are similar to the synthases that catalyze condensation of 2 -carbon units in the biosynthesis of fatty acids. PKS enzymes are encoded by PKS genes that usually consist of three or more open reading frames (ORFs). Each ORF typically comprises two or more "modules" of ketosynthase activity, each module of which consists of at least two (if a loading module) and more typically three or more enzymatic activities or "domains." Two major types of PKS enzymes are known; these differ in their composition and mode of synthesis. These two major types of PKS enzymes are commonly referred to as Type I or "modular" and Type II "iterative" PKS enzymes.
[0005] Modular PKSs are responsible for producing a large number of 12,14 , and 16 -membered macrolide antibiotics including methymycin, erythromycin, narbomycin, picromycin, and tylosin. These large multifunctional enzymes ( $>300,000 \mathrm{kDa}$ ) catalyze the biosynthesis of polyketide macrolactones through multistep pathways involving decarboxylative condensations between acyl thioesters followed by cycles of varying $\beta$-carbon process-
ing activities (see O'Hagan, D. The polyketide metabolites; E. Horwood: New York, 1991, incorporated herein by reference).
[0006] During the past half decade, the study of modular PKS function and specificity has been greatly facilitated by the plasmid-based Streptomyces coelicolor expression system developed with the 6-deoxyerythronolide B (6-dEB) synthase (DEBS) genes (see Kao et al., 1994, Science, 265: 509-512, McDaniel et al., 1993, Science 262: 1546-1557, and U.S. Pat. Nos. 5,672,491 and 5,712,146, each of which is incorporated herein by reference). The advantages to this plasmid-based genetic system for DEBS were that it overcame the tedious and limited techniques for manipulating the natural DEBS host organism, Saccharopolyspora erythaea, allowed more facile construction of recombinant PKSs, and reduced the complexity of PKS analysis by providing a "clean" host background. This system also expedited construction of the first combinatorial modular polyketide library in Streptomyces (see PCT publication No. WO 98/49315, incorporated herein by reference).
[0007] The ability to control aspects of polyketide biosynthesis, such as monomer selection and degree of $\beta$-carbon processing, by genetic manipulation of PKSs has stimulated great interest in the combinatorial engineering of novel antibiotics (see Hutchinson, 1998, Curr. Opin. Microbiol. 1: 319-329; Carreras and Santi, 1998, Curr. Opin. Biotech. 9: 403-411; and U.S. Pat. Nos. 5,712,146 and 5,672,491, each of which is incorporated herein by reference). This interest has resulted in the cloning, analysis, and manipulation by recombinant DNA technology of genes that encode PKS enzymes. The resulting technology allows one to manipulate a known PKS gene cluster either to produce the polyketide synthesized by that PKS at higher levels than occur in nature or in hosts that otherwise do not produce the polyketide. The technology also allows one to produce molecules that are structurally related to, but distinct from, the polyketides produced from known PKS gene clusters.
[0008] The present invention provides methods and reagents relating to the PKS gene cluster for the polyketide antibiotics known as narbomycin and picromycin. Narbomycin is produced in Streptomyces narbonensis, and both narbomycin and picromycin are produced in $S$. venezuelae. These species are unique among macrolide producing organisms in that they produce, in addition to the 14 -membered macrolides narbomycin and picromycin (picromycin is shown in FIG. 1, compound 1), the 12 -membered macrolides neomethymycin and methymycin (methymycin is shown in FIG. 1, compound 2). Based on the structural similarities between picromycin and methymycin, it was speculated that methymycin would result from premature cyclization of a hexaketide intermediate in the picromycin pathway.
[0009] Glycosylation of the C5 hydroxyl group of the polyketide precursor, narbonolide, is achieved through an endogenous desosaminyl transferase to produce narbomycin. In Streptomyces venezuelae, narbomycin is then converted to picromycin by the endogenously produced narbomycin hydroxylase. Thus, as in the case of other macrolide antibiotics, the macrolide product of the narbonolide PKS is further modified by hydroxylation and glycosylation.
[0010] Picromycin (FIG. 1, compound 1) is of particular interest because of its close structural relationship to
ketolide compounds (e.g. HMR 3004, FIG. 1, compound 3). The ketolides are a new class of semi-synthetic macrolides with activity against pathogens resistant to erythromycin (see Agouridas et al., 1998, J. Med. Chem. 41: 4080-4100, incorporated herein by reference). Thus, genetic systems that allow rapid engineering of the narbonolide PKS would be valuable for creating novel ketolide analogs for pharmaceutical applications. Furthermore, the production of picromycin as well as novel compounds with useful activity could be accomplished if the heterologous expression of the narbonolide PKS in Streptomyces lividans and other host cells were possible. The present invention meets these and other needs.

## SUMMARY OF THE INVENTION

[0011] The present invention provides recombinant methods and materials for expressing PKSs derived in whole and in part from the narbonolide PKS and other genes involved in narbomycin and picromycin biosynthesis in recombinant host cells. The invention also provides the polyketides derived from the narbonolide PKS. The invention provides the complete PKS gene cluster that ultimately results, in Streptomyces venezuelae, in the production of picromycin. The ketolide product of this PKS is narbonolide. Narbonolide is glycosylated to obtain narbomycin and then hydroxylated at C12 to obtain picromycin. The enzymes responsible for the glycosylation and hydroxylation are also provided in recombinant form by the invention.
[0012] Thus, in one embodiment, the invention is directed to recombinant materials that contain nucleotide sequences encoding at least one domain, module, or protein encoded by a narbonolide PKS gene. The invention also provides recombinant materials useful for conversion of ketolides to antibiotics. These materials include recombinant DNA compounds that encode the C12 hydroxylase (the picK gene), the desosamine biosynthesis and desosaminyl transferase enzymes, and the beta-glucosidase enzyme involved in picromycin biosynthesis in $S$. venezuelae and the recombinant proteins that can be produced from these nucleic acids in the recombinant host cells of the invention.
[0013] In one embodiment, the invention provides a recombinant expression vector that comprises a heterologous promoter positioned to drive expression of the narbonolide PKS. In a preferred embodiment, the promoter is derived from a PKS gene. In a related embodiment, the invention provides recombinant host cells comprising the vector that produces narbonolide. In a preferred embodiment, the host cell is Streptomyces lividans or $S$. coelicolor.
[0014] In another embodiment, the invention provides a recombinant expression vector that comprises the desosamine biosynthetic genes as well as the desosaminyl transferase gene. In a related embodiment, the invention provides recombinant host cells comprising the vector that produces the desosamine biosynthetic gene products and desosaminyl transferase gene product. In a preferred embodiment, the host cell is Streptomyces lividans or $S$. coelicolor.
[0015] In another embodiment, the invention provides a method for desosaminylating polyketide compounds in recombinant host cells, which method comprises expressing the PKS for the polyketide and the desosaminyl transferase and desosamine biosynthetic genes in a host cell. In a preferred embodiment, the host cell expresses a beta-glucosidase gene as well. This preferred method is especially advantageous when producing desosaminylated polyketides
in Streptomyces host cells, because such host cells typically glucosylate desosamine residues of polyketides, which can decrease desired activity, such as antibiotic activity. By coexpression of beta-glucosidase, the glucose residue is removed from the polyketide.
[0016] In another embodiment, the invention provides the picK hydroxylase gene in recombinant form and methods for hydroxylating polyketides with the recombinant gene product. The invention also provides polyketides thus produced and the antibiotics or other useful compounds derived therefrom.
[0017] In another embodiment, the invention provides a recombinant expression vector that comprises a promoter positioned to drive expression of a hybrid PKS comprising all or part of the narbonolide PKS and at least a part of a second PKS. In a related embodiment, the invention provides recombinant host cells comprising the vector that produces the hybrid PKS and its corresponding polyketide. In a preferred embodiment, the host cell is Streptomyces lividans or S. coelicolor.
[0018] In a related embodiment, the invention provides recombinant materials for the production of libraries of polyketides wherein the polyketide members of the library are synthesized by hybrid PKS enzymes of the invention. The resulting polyketides can be further modified to convert them to other useful compounds, such as antibiotics, typically through hydroxylation and/or glycosylation. Modified macrolides provided by the invention that are useful intermediates in the preparation of antibiotics are of particular benefit.
[0019] In another related embodiment, the invention provides a method to prepare a nucleic acid that encodes a modified PKS, which method comprises using the narbonolide PKS encoding sequence as a scaffold and modifying the portions of the nucleotide sequence that encode enzymatic activities, either by mutagenesis, inactivation, insertion, or replacement. The thus modified narbonolide PKS encoding nucleotide sequence can then be expressed in a suitable host cell and the cell employed to produce a polyketide different from that produced by the narbonolide PKS. In addition, portions of the narbonolide PKS coding sequence can be inserted into other PKS coding sequences to modify the products thereof. The narbonolide PKS can itself be manipulated, for example, by fusing two or more of its open reading frames, particularly those for extender modules 5 and 6, to make more efficient the production of 14 -membered as opposed to 12 -membered macrolides.
[0020] In another related embodiment, the invention is directed to a multiplicity of cell colonies, constituting a library of colonies, wherein each colony of the library contains an expression vector for the production of a modular PKS derived in whole or in part from the narbonolide PKS. Thus, at least a portion of the modular PKS is identical to that found in the PKS that produces narbonolide and is identifiable as such. The derived portion can be prepared synthetically or directly from DNA derived from organisms that produce narbonolide. In addition, the invention provides methods to screen the resulting polyketide and antibiotic libraries.
[0021] The invention also provides novel polyketides and antibiotics or other useful compounds derived therefrom. The compounds of the invention can be used in the manufacture of another compound. In a preferred embodiment, the antibiotic compounds of the invention are formulated in a mixture or solution for administration to an animal or human.
[0022] These and other embodiments of the invention are described in more detail in the following description, the examples, and claims set forth below.

## BRIEF DESCRIPTION OF THE FIGURES

[0023] FIG. 1 shows the structures of picromycin (compound 1), methymycin (compound 2), and the ketolide HMR 3004 (compound 3).
[0024] FIG. 2 shows a restriction site and function map of cosmid pKOS023-27.
[0025] FIG. 3 shows a restriction site and function map of cosmid pKOS023-26.
[0026] FIG. 4 has three parts. In Part A, the structures of picromycin (A(a)) and methymycin (A(b)) are shown, as well as the related structures of narbomycin, narbonolide, and methynolide. In the structures, the bolded lines indicate the two or three carbon chains produced by each module (loading and extender) of the narbonolide PKS. Part B shows the organization of the narbonolide PKS genes on the chromosome of Streptomyces venezuelae, including the location of the various module encoding sequences (the loading module domains are identified as $\mathrm{sKS}{ }^{*}$, sAT, and sACP ), as well as the picB thioesterase gene and two desosamine biosynthesis genes (picCII and picCIII). Part C shows the engineering of the $S$. venezuelae host of the invention in which the picAI gene has been deleted. In the Figure, ACP is acyl carrier protein; AT is acyltransferase; DH is dehydratase; ER is enoylreductase; $K R$ is ketoreductase; KS is ketosynthase; and TE is thioesterase.
[0027] FIG. 5 shows the narbonolide PKS genes encoded by plasmid $\mathrm{pKOS039-86}$, the compounds synthesized by each module of that PKS and the narbonolide (compound 4) and 10-deoxymethynolide (compound 5) products produced in heterologous host cells transformed with the plasmid. The Figure also shows a hybrid PKS of the invention produced by plasmid pKOS038-18, which encodes a hybrid of DEBS and the narbonolide PKS. The Figure also shows the compound, 3,6-dideoxy-3-oxo-erythronolide B (compound 6), produced in heterologous host cells comprising the plasmid.
[0028] FIG. 6 shows a restriction site and function map of plasmid pKOS039-104, which contains the desosamine biosynthetic, beta-glucosidase, and desosaminyl transferase genes under transcriptional control of actII-4.

## DETAILED DESCRIPTION OF THE INVENTION

[0029] The present invention provides useful compounds and methods for producing polyketides in recombinant host cells. As used herein, the term recombinant refers to a compound or composition produced by human intervention. The invention provides recombinant DNA compounds encoding all or a portion of the narbonolide PKS. The invention also provides recombinant DNA compounds encoding the enzymes that catalyze the further modification of the ketolides produced by the narbonolide PKS. The invention provides recombinant expression vectors useful in producing the narbonolide PKS and hybrid PKSs composed of a portion of the narbonolide PKS in recombinant host cells. Thus, the invention also provides the narbonolide PKS, hybrid PKSs, and polyketide modification enzymes in recombinant form. The invention provides the polyketides produced by the recombinant PKS and polyketide modification enzymes. In particular, the invention provides methods for producing the polyketides 10 -deoxymethynolide,
narbonolide, YC17, narbomycin, methymycin, neomethymycin, and picromycin in recombinant host cells.
[0030] To appreciate the many and diverse benefits and applications of the invention, the description of the invention below is organized as follows. First, a general description of polyketide biosynthesis and an overview of the synthesis of narbonolide and compounds derived therefrom in Streptomyces venezuelae are provided. This general description and overview are followed by a detailed description of the invention in six sections. In Section I, the recombinant narbonolide PKS provided by the invention is described. In Section II, the recombinant desosamine biosynthesis genes, the desosaminyl transferase gene, and the beta-glucosidase gene provided by the invention are described. In Section III, the recombinant picK hydroxylase gene provided by the invention is described. In Section IV, methods for heterologous expression of the narbonolide PKS and narbonolide modification enzymes provided by the invention are described. In Section V, the hybrid PKS genes provided by the invention and the polyketides produced thereby are described. In Section VI, the polyketide compounds provided by use invention and pharmaceutical compositions of those compounds are described. The detailed description is followed by a variety of working examples illustrating the invention.
[0031] The narbonolide synthase gene, like other PKS genes, is composed of coding sequences organized in a loading module, a number of extender modules, and a thioesterase domain. As described more fully below, each of these domains and modules is a polypeptide with one or more specific functions. Generally, the loading module is responsible for binding the first building block used to synthesize the polyketide and transferring it to the first extender module. The building blocks used to form complex polyketides are typically acylthioesters, most commonly acetyl, propionyl, malonyl, methylmalonyl, and ethylmalonyl CoA. Other building blocks include amino acid like acylthioesters. PKSs catalyze the biosynthesis of polyketides through repeated, decarboxylative Claisen condensations between the acylthioester building blocks. Each module is responsible for binding a building block, performing one or more functions on that building block, and transferring the resulting compound to the next module. The next module, in turn, is responsible for attaching the next building block and transferring the growing compound to the next module until synthesis is complete. At that point, an enzymatic thioesterase activity cleaves the polyketide from the PKS.
[0032] Such modular organization is characteristic of the class of PKS enzymes that synthesize complex polyketides and is well known in the art. The polyketide known as 6 -deoxyerythronolide B is a classic example of this type of complex polyketide. The genes, known as eryAI, eryAII, and eryAIII (also referred to herein as the DEBS genes, for the proteins, known as DEBS1, DEBS2, and DEBS3, that comprise the 6 -dEB synthase), that code for the multisubunit protein known as DEBS that synthesizes 6 -dEB are described in U.S. Pat. No. 5,824,513, incorporated herein by reference. Recombinant methods for manipulating modular PKS genes are described in U.S. Pat. Nos. 5,672,491; $5,843,718 ; 5,830,750$; and $5,712,146$; and in PCT publication Nos. 98/49315 and 97/02358, each of which is incorporated herein by reference.
[0033] The loading module of DEBS consists of two domains, an acyl-transferase (AT) domain and an acyl
carrier protein (ACP) domain. Each extender module of DEBS, like those of other modular PKS enzymes, contains a ketosynthase (KS), AT, and ACP domains, and zero, one, two, or three domains for enzymatic activities that modify the beta-carbon of the growing polyketide chain. A module can also contain domains for other enzymatic activities, such as, for example, a methyltransferase or dimethyltransferase activity. Finally, the releasing domain contains a thioesterase and, often, a cyclase activity.
[0034] The AT domain of the loading module recognizes a particular acyl-CoA (usually acetyl or propionyl but sometimes butyryl) and transfers it as a thiol ester to the ACP of the loading module. Concurrently, the AT on each of the extender modules recognizes a particular extender-CoA (malonyl or alpha-substituted malonyl, i.e., methylmalonyl, ethylmalonyl, and carboxylglycolyl) and transfers it to the ACP of that module to form a thioester. Once the PKS is primed with acyl- and malonyl-ACPs, the acyl group of the loading module migrates to form a thiol ester (trans-esterification) at the KS of the first extender module; at this stage, extender module 1 possesses an acyl-KS adjacent to a malonyl (or substituted malonyl) ACP. The acyl group derived from the loading module is then covalently attached to the alpha-carbon of the malonyl group to form a carboncarbon bond, driven by concomitant decarboxylation, and generating a new acyl-ACP that has a backbone two carbons longer than the loading unit (elongation or extension). The growing polyketide chain is transferred from the ACP to the KS of the next module, and the process continues.
[0035] The polyketide chain, growing by two carbons each module, is sequentially passed as covalently bound thiol esters from module to module, in an assembly line-like process. The carbon chain produced by this process alone would possess a ketone at every other carbon atom, producing a polyketone, from which the name polyketide arises. Most commonly, however, additional enzymatic activities modify the beta keto group of each two-carbon unit just after it has been added to the growing polyketide chain but before it is transferred to the next module. Thus, in addition to the minimal module containing KS, AT, and ACP domains necessary to form the carbon-carbon bond, modules may contain a ketoreductase (ICR) that reduces the keto group to an alcohol. Modules may also contain a KR plus a dehydratase (DH) that dehydrates the alcohol to a double bond. Modules may also contain a KR, a DH, and an enoylreductase (ER) that converts the double bond to a saturated single bond using the beta carbon as a methylene function. As noted above, modules may contain additional enzymatic activities as well.
[0036] Once a polyketide chain traverses the final extender module of a PKS, it encounters the releasing domain or thioesterase found at the carboxyl end of most PKSs. Here, the polyketide is cleaved from the enzyme and cyclyzed. The resulting polyketide can be modified further by tailoring enzymes; these enzymes add carbohydrate groups or methyl groups, or make other modifications, i.e., oxidation or reduction, on the polyketide core molecule.
[0037] While the above description applies generally to modular PKS enzymes, there are a number of variations that exist in nature. For example, some polyketides, such as epothilone, incorporate a building block that is derived from an amino acid. PKS enzymes for such polyketides include an
activity that functions as an amino acid ligase or as a non-ribosomal peptide synthetase (NRPS). Another example of a variation, which is actually found more often than the two domain loading module construct found in DEBS, occurs when the loading module of the PKS is not composed of an AT and an ACP but instead utilizes an inactivated KS , an AT, and an ACP. This inactivated KS is in most instances called KSQ, where the superscript letter is the abbreviation for the amino acid, glutamine, that is present instead of the active site cysteine required for activity. For example, the narbonolide PKS loading module contains aKS ${ }^{\text {. }}$. Yet another example of a variation has been mentioned above in the context of modules that include a methyltransferase or dimethyltransferase activity; modules can also include an epimerase activity. These variations will be described further below in specific reference to the narbonolide PKS and the various recombinant and hybrid PKSs provided by the invention.
[0038] With this general description of polyketide biosynthesis, one can better appreciate the biosynthesis of narbonolide related polyketides in Streptomyces venezuelae and $S$. narbonensis. The narbonolide PKS produces two polyketide products, narbonolide and 10-deoxymethynolide. Narbonolide is the polyketide product of all six extender modules of the narbonolide PKS. 10-deoxymethynolide is the polyketide product of only the first five extender modules of the narbonolide PKS. These two polyketides are desosaminylated to yield narbomycin and YC17, respectively. These two glycosylated polyketides are the final products produced in $S$. narbonensis. In S. venezuelae, these products are hydroxylated by the picK gene product to yield picromycin and either methymycin (hydroxylation at the C10 position of YC7) or neomethymycin (hydroxylation at the C12 position of YC17). The present invention provides the genes required for the biosynthesis of all of these polyketides in recombinant form.

## [0039] Section 1: The Narbonolide PKS

[0040] The narbonolide PKS is composed of a loading module, six extender modules, and a thioesterase domain. FIG. 4, part B, shows the organization of the narbonolide PKS genes on the Streptomyces venezuelae chromosome, as well as the location of the module encoding sequences in those genes, and the various domains within those modules. In the Figure, the loading module is not numbered, and its domains are indicated as sKS *, sAT , and ACP. Also shown in the Figure, part A, are the structures of picromycin and methymycin.
[0041] The loading and six extender modules and the thioesterase domain of the narbonolide PKS reside on four proteins, designated PICAI, PICAII, PICAII, and PICAIV. PICAI includes the loading module and extender modules 1 and 2 of the PKS. PICAII includes extender modules 3 and 4. PICAIII includes extender module 5. PICAIV includes extender module 6 and a thioesterase domain. There is a second thioesterase domain (TEII) on a separate protein, designated PICB. The amino acid sequences of these proteins are shown below.


#### Abstract

Amino acid sequence of narbonolide synthase subunit 1, PICAI 1 MSTVSKSESE EFVSVSNDAG SAHGTAEPVA VVGISCRVPG ARDPPEFWEL LAAGGQAVTD 61 VPADRWNAGD FYDPDRSAPG RSNSRWGGFI EDVDRFDAAF FGISPREAAE MDPQQRLALE 121 LGWEALERAG IDPSSLTGTR TGVFAGAIWD DYATLKHRQG GAAITPHTVT GLHRGIIANR 181 LSYTLGLRGP SMVVDSGQSS SLVAVHLACE SLRRGESELA LAGGVSLNLV PDSIIGASKF 241 GGLSPDGRAY TFDARANGYV RGEGGGFVVL KRLSRAVADG DPVLAVIRGS AVNNGGAAQG 301 MTTPDAQAQE AVLREAHERA GTAPADVRYV ELHGTGTPVG DPIEAAALGA ALGTGRPAGQ 361 PLLVGSVKTN IGHLEGAAGI AGLIKAVLAV RGRALPASLN YETPNPAIPF EELNLRVNTE 421 YLPWEPEHDG QRMVVGVSSF GMGGTNAHVV LEEAPGVVEG ASVVESTVGG SAVGGGVVPW 481 VVSAKSAAAL DAQIERLAAF ASRDRTDGVD AGAVDAGAVD AGAVARVLAG GRAQFEHRAV 541 VVGSGPDDLA AALAAPEGLV RGVASGVGRV AFVFPGQGTQ WAGMGAELLD SSAVFAAAMA 601 ECEAALSPYV DWSLEAVVRQ APGAPTLERV DVVQPVTFAV MVSLARVWQH HGVTPQAVVG 661 HSQGEIAAAY VAGALSLDDA ARVVTLRSKS IAAHLAGKGG MLSLALSEDA VLERLAGFDG 721 LSVAAVNGPT ATVVSGDPVQ IEELARACEA DGVRARVIPV DYASHSRQVE IIESELAEVL 781 AGLSPQAPRV PFFSTLEGAW ITEPVLDGGY WYRNLRHRVG FAPAVETLAT DEGFTHFVEV 841 SAHPVLTMAL PGTVTGLATL RRDNGGQDRL VASLAEAWAN GLAVDWSPLL PSATGHHSDL 901 PTYAFQTERH WLGEIEALAP AGEPAVQPAV LRTEAAEPAE LDRDEQLRVI LDKVRAQTAQ 961 VLGYATGGQI EVDRTFREAG CTSLTGVDLR NRINAAFGVR MAPSMIFDFP TPEALAEQLL 1021 LVVHGEAAAN PAGAEPAPVA AAGAVDEPVA IVGMACRLPG GVASPEDLWR LVAGGGDAIS 1081 EFPQDRGWDV EGLYHPDPEH PGTSYVRQGG FIENVAGFDA AFFGISPREA LAMDPQQRLL 1141 LETSWEAVED AGIDPTSLRG RQVGVFTGAM THEYGPSLRD GGEGLDGYLL TGNTASVMSG 1201 RVSYTLGLEG PALTVDTACS SSLVALHLAV QALRKGEVDM ALAGGVAVMP TPGMFVEF'SR 1261 QRGLAGDGRS KAFAASADGT SWSEGVGVLL VERLSDARRN GHQVLAVVRG SAVNQDGASN 1321 GLTAPNGPSQ ORVIRRALAD ARLTTSDVDV VEAHGTGTRL GDPIEAQALI ATYGQGRDDE 1381 QPLRLGSLKS NIGHTQAAAG VSGVIKMVQA MRHGLLPKTL HVDEPSDQID WSAGAVELLT 1441 EAVDWPEKQD GGLRRAAVSS FGISGTNAHV VLEEAPVVVE GASVVEPSVG GSAVGGGVTP 1501 WVVSAKSAAA LDAQIERLAA FASRDRTDDA DAGAVDAGAV AHVLADGRAQ FEHRAVALGA 1561 GADDLVQALA DPDGLIRGTA SGVGRVAFVF PGQGTQWAGM GAELLDSSAV FAAAMAECEA 1621 ALSPYVDWSL EAVVRQAPGA PTLERVDVVQ PVTFAVMVSL ARVWQHHGVT PQAVVGHSQG 1681 EIAAAYVAGA LPLDDAARVV TLRSKSIAAH LAGKGGMLSL ALNEDAVLER LSDFDGLSVA 1741 AVNGPTATVV SGDPVQIEEL AQACKADGFR ARIIPVDYAS HSRQVEIIES ELAQVLAGLS 1801 PQAPRVPFFS TLEGTWITEP VLDGTYWYRN LRHRVGFAPA IETLAVDEGF THFVEVSAHP 1861 VLTMTLPETV TGLGTLRREQ GGQERLVTSL AEAWVNGLPV AWTSLLPATA SRPGLPTYAF 1921 QAERYWLENT PAALATGDDW RYRIDWKRLP AAEGSERTGL SGRWLAVTPE DHSAQAAAVL 1981 TALVDAGAKV EVLTAGADDD REALAARLTA LTTGDGFTGV VSLLDGLVPQ VAWVQALGDA 2041 GIKAPLWSVT QGAVSVGRLD TPADPDRAML WGLGRVVALE HPERWAGLVD LPAQPDAAAL 2101 AHLVTALSGA TGEDQIAIRT TGLHARRLAR APLHGRRPTR DWQPHGTVLI TGGTGALGSH 2161 AARWMAHHGA EHLLLVSRSG EQAPGATQLT AELTASGARV TIAACDVADP HAMRTLLDAI 2221 PAETPLTAVV HTAGALDDGI VDTLTAEQVR RAHRAKAVGA SVLDELTRDL DLDAFVLFSS


## 2281 VSSTLGIPGQ GNYAPHNAYL DALAARRRAT GRSAVSVAWG PWDGGGMAAG DGVAERLRNH

 2341 GVPGMDPELA LAALESALGR DETAITVADI DWDRFYLAYS SGRPQPLVEE LPEVRRIIDA 2401 RDSATSGQGG SSAQGANPLA RRLAAAAPGE RTEILLGLVR AQAAAVLRMR SPEDVAADRA 2461 FKDIGFDSLA GVELRNRLTR ATGLQLPATL VFDHPTPLAL VSLLRSEFLG DEETADARRS 2521 AALPATVGAG AGAGAGTDAD DDPIAIVAMS CRYPGDIRSP EDLWRMLSEG GEGITPFPTD 2581 RGWDLDGLYD ADPDALGRAY VREGGFLHDA AEFDAEFFGV SPREALAMDP QQRMLLTTSW 2641 EAFERAGIEP ASLRGSSTGV FIGLSYQDYA ARVPNAPRGV EGYLLTGSTP SVASGRIAYT 2701 FGLEGPATTV DTACSSSLTA LHLAVRALRS GECTMALAGG VAMMATPHMF VEFSRQRALA 2761 PDGRSKAFSA DADGFGAAEG VGLLLVERLS DARRNGHPVL AVVRGTAVNQ DGASNGLTAP 2821 NGPSQQRVIR QALADARLAP GDIDAVETHG TGTSLGDPIE AQGLQATYGK ERPAERPLAI 2881 GSVKSNIGHT QAAAGAAGII KMVLAMRHGT LPKTLHADEP SPHVDWANSG LALVTEPIDW 2941 PAGTGPRRAA VSSFGISGTN AHVVLEQAPD AAGEVLGADE VPEVSETVAM AGTAGTSEVA 3001 EGSEASEAPA APGSREASLP GHLPWVLSAK DEQSLRGQAA ALHAWLSEPA ADLSDADGPA 3061 RLRDVGYTLA TSRTAFAHRA AVTAADRDGF LDGLATLAQG GTSAHVHLDT ARDGTTAFLF 3121 TGQGSQRPGA GRELYDRHPV FARALDEICA HLDGHLELPL LDVMFAAEGS AEAALLDETR 3181 YTQCALFALE VALFRLVESW GMRPAALLGH SVGEIAAAHV AGVFSLADAA RLVAARGRLM 3241 QELPAGGAML AVQAAEDEIR VWLETEERYA GRLDVAAVNG PEAAVLSGDA DAAREAEAYW 3301 SGLGRRTRAL RVSHAFHSAH MDGMLDGFRA VLETVEFRRP SLTVVSNVTG LAAGPDDLCD 3361 PEYWVRHVRG TVRFLDGVRV LRDLGVRTCL ELGPDGVLTA MAADGLADTP ADSAAGSPVG 3421 SPAGSPADSA AGALRPRPLL VALLRRKRSE TETVADALGR AHAHGTGPDW HAWFAGSGAH 3481 RVDLPTYSFR RDRYWLDAPA ADTAVDTAGL GLGTADHPL工 GAVVSLPDRD GLLLTGRLSL 3541 RTHPWLADHA VLGSVLLPGA AMVELAAHAA ESAGLRDVRE LTLLEPLVLP EHGGVELRVT 3601 VGAPAGEPGG ESAGDGARPV SLHSRLADAP AGTAWSCHAT GLLATDRPEL PVAPDRAAMW 3661 PPQGAEEVPL DGLYERLDGN GLAFGPLFQG LNAVWRYEGE VFADIALPAT TNATAPATAN 3721 GGGSAAAAPY GIHPALLDAS LHAIAVGGLV DEPELVRVPF HWSGVTVHAA GAAAARVRLA 3781 SAGTDAVSLS LTDGEGRPLV SVERLTLRPV TADQAAASRV GGLMHRVAWR PYALASSGEQ 3841 DPHATSYGPT AVLGKDELKV AAALESAGVE VGLYPDLAAL SQDVAAGAPA PRTVLAPLPA 3901 GPADGGAEGV RGTVARTLEL LQAWLADEHL AGTRLLLVTR GAVRDPEGSG ADDGGEDLSH 3961 AAAWGLVRTA QTENPGRFGL LDLADDASSY RTLPSVLSDA GLRDEPQLAL HDGTIRLARL 4021 ASVRPETGTA APALAPEGTV LLTGGTGGLG GLVARHVVGE WGVRRLLLVS RRGTDAPGAD 4081 ELVHELEALG ADVSVAACDV ADREALTAVL DAIPAEHPLT AVVHTAGVLS DGTLPSMTTE 4141 DVEHVLRPKV DAAFLLDELT STPAYDLAAF VMFSSAAAVF GGAGQGAYAA ANATLDALAW 4201 RRRAAGLPAL SLGWGLWAET SGMTGELGQA DLRRMSRAGI GGISDAEGIA LLDAALRDDR 4261 HPVLLPLRLD AAGLRDAAGN DPAGIPALFR DVVGARTVRA RPSAASASTT AGTAGTPGTA 4321 DGAAETAAVT LADRAATVDG PARQRLLLEF VVGEVAEVLG HARGHRIDAE RGFLDLGFDS 4381 LTAVELRNRL NSAGGLALPA TLVFDHPSPA ALASHLDAEL PRGASDQDGA GNRNGNENGT 4441 TASRSTAETD ALLAQLTRLE GALVLTGLSD APGSEEVLEH LRSLRSMVTG ETGTGTASGA 4501 PDGAGSGAED RPWAAGDGAG GGSEDGAGVP DFMNASAEEL FGLLDQDPST D
#### Abstract

-continued 1 VSTVNEEKYL DYLRRATADL HEARGRLREL EAKAGEPVAI VGMACRLPGG VASPEDLWRL 61 VAGGEDAISE FPQDRGWDVE GLYDPNPEAT GKSYAREAGF LYEAGEFDAD FFGISPREAL 121 AMDPQQRLLL EASWEAFEHA GIPAATARGT SVGVFTGVMY HDYATRLTDV PEGIEGYLGT 181 GNSGSVASGR VAYTLGLEGP AVTVDTACSS SLVALHLAVQ ALRKGEVDMA LAGGVTVMST 241 PSTFVEFSRQ RGLAPDGRSK SFSSTADGTS WSEGVGVLLV ERLSDARRKG HRILAVVRGT 301 AVNODGASSG LTAPNGPSQQ RVIRRALADA RLTTSDVDVV EAHGTGTRLG DPIEAQAVIA 361 TYGQGRDGEQ PLRLGSLKSN IGHTQAAAGV SGVIKMVQAM RHGVLPKTLH VEKPTDQVDW 421 SAGAVELLTE AMDWPDKGDG GLRRAAVSSF GVSGTNAHVV LEEAPAAEET PASEATPAVE 481 PSVGAGLVPW LVSAKTPAAL DAQIGRLAAF ASQGRTDAAD PGAVARVLAG GRAEFEHRAV 541 VLGTGQDDFA QALTAPEGLI RGTPSDVGRV AFVFPGQGTQ WAGMGAELLD VSKEFAAAMA 601 ECESALSRYV DWSLEAVVRQ APGAPTLERV DVVQPVTFAV MVSLAKVWQH HGVTPQAVVG 661 HSQGEIAAAY VAGALTLDDA ARVVTLRSKS IAAHLAGKGG MISLALSEEA TRQRIENLHG 721 LSIAAVNGPT ATVVSGDPTQ IQELAQACEA DGVRARIIPV DYASHSAHVE TIESELAEVL 781 AGLSPRTPEV PFFSTLEGAW ITEPVLDGTY WYRNLRHRVG FAPAVETLAT DEGFTHFIEV 841 SAHPVLTMTL PETVTGLGTL RREQGGQERL VTSLAEAWTN GLTIDWAPVL PTATGHHPEL 901 PTYAFQRRHY WLHDSPAVQG SVQDSWRYRI DWKRLAVADA SERAGLSGRW LVVVPEDRSA 961 EAAPVLAALS GAGADPVQLD VSPLGDRQRL AATLGEALAA AGGAVDGVLS LLANDESAHP 1021 GHPAPFTRGT GATLTLVQAL EDAGVAAPLW CVTHGAVSVG RADHVTSPAQ AMVWGMGRVA 1081 ALEHPERWGG LIDLPSDADR AALDRMTTVL AGGTGEDQVA VRASGLLARR LVRASLPAHG 1141 TASPWWQADG TVLVTGAEEP AAAEAARRLA RDGAGHLLLH TTPSGSEGAE GTSGAAEDSG 1201 LAGLVAELAD LGATATVVTC DLTDAEAAAR LLAGVSDAHP LSAVLHLPPT VDSEPLAATD 1261 ADALARVVTA KATAALHLDR LLREAAAAGG RPPVLVLFSS VAAIWGGAGQ GAYAAGTAFL 1321 DALAGQHRAD GPTVTSVAWS PWEGSRVTEG ATGERLRRLG LRPLAPATAL TALDTALGHG 1381 DTAVTIADVD WSSFAPGFTT ARPGTLLADL PEARRALDEQ QSTTAADDTV LSRELGALTG 1441 AEQQRRMQEL VREHLAVVLN HPSPEAVDTG RAFRDLGFDS LTAVELRNRL KNATGLALPA 1501 TLVFDYPTPR TLAEFLLAEI LGEQAGAGEQ LPVDGGVDDE PVAIVGMACR LPGGVASPED 1561 LWRLVAGGED AISGFPQDRG WDVEGLYDPD PDASGRTYCR AGGFLDEAGE FDADFFGISP 1621 REALAMDPQQ RLLLETSWEA VEDAGIDPTS LQGQQVGVFA GTNGPHYEPL LRNTAEDLEG 1681 YVGTGNAASI MSGRVSYTLG LEGPAVTVDT ACSSSLVALH LAVQALRKGE CGLALAGGVT 1741 VMSTPTTFVE FSRQRGLAED GRSKAFAASA DGFGPAEGVG MLLVERLSDA RRNGHRVLAV 1801 VRGSAVNQDG ASNGLTAPNG PSQQRVIRRA LADARLTTAD VDVVEAHGTG TRLGDPIEAQ 1861 ALIATYGQGR DTEQPLRLGS LKSNIGHTQA AAGVSGIIKM VQAMRHGVLP KTLHVDRPSD 1921 QIDWSAGTVE LLTEAMDWPR KQEGGLRRAA VSSFGISGTN AHIVLEEAPV DEDAPADEPS 1981 VGGVVPWLVS AKTPAALDAQ IGRLAAFASQ GRTDAADPGA VARVLAGGRA QFEHRAVALG 2041 TGQDDLAAAL AAPEGLVRGV ASGVGRVAFV FPGQGTQWAG MGAELLDVSK EFAAAMAECE 2101 AALAPYVDWS LEAVVRQAPG APTLERVDVV QPVTFAVMVS LAKVWQHHGV TPQAVVGHSQ 2161 GEIAAAYVAG ALSLDDAARV VTLRSKSIGA HLAGQGGMLS LALSEAAVVE RLAGFDGLSV 2221 AAVNGPTATV VSGDPTQIQE LAQACEADGV RARIIPVDYA SHSAHVETIE SELADVLAGL 2281 SPQTPQVPFF STLEGAWITE PALDGGYWYR NLRHRVGFAP AVETLATDEG FTHFVEVSAH


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2341 PVLTMALPET VTGLGTLRRD NGGQHRLTTS LAEAWANGLT VDWASLLPTT TTHPDLPTYA 2401 FQTERYWPQP DLSAAGDITS AGLGAAEHPL LGAAVALADS DGCLLTGSLS LRTHPWLADH 2461 AVAGTVLLPG TAFVELAFRA GDQVGCDLVE ELTLDAPLVL PRRGAVRVQL SVGASDESGR 2521 RTFGLYAHPE DAPGEAEWTR HATGVLAARA DRTAPVADPE AWPPPGAEPV DVDGLYERFA 2581 ANGYGYGPLF QGVRGVWRRG DEVFADVALP AEVAGAEGAR FGLHPALLDA AVQAAGAGGA 2641 FGAGTRLPFA WSGISLYAVG ATALRVRLAP AGPDTVSVSA ADSSGOPVFA ADSLTVLPVD 2701 PAQLAAFSDP TLDALHLLEW TAWDGAAQAL PGAVVLGGDA DGLAAALRAG GTEVLSFPDL 2761 TDLVEAVDRG ETPAPATVLV ACPAAGPGGP EHVREALHGS LALMOAWLAD ERFTDGRLVL 2821 VTRDAVAARS GDGLRSTGQA AVWGLGRSAQ TESPGRFVLL DLAGEARTAG DATAGDGLTT 2881 GDATVGGTSG DAALGSALAT ALGSGEPQLA LRDGALLVPR LARAAAPAAA DGLAAADGLA 2941 ALPLPAAPAL WRLEPGTDGS LESLTAAPGD AETLAPEPLG PGQVRIAIRA TGLNFRDVLI 3001 ALGMYPDPAL MGTEGAGVVT ATGPGVTHLA PGDRVMGLLS GAYAPVVVAD ARTVARMPEG 3061 WTFAQGASVP VVFLTAVYAL RDLADVKPGE RLLVHSAAGG VGMAAVQLAR HWGVEVHGTA 3121 SHGKWDALRA LGLDDAHIAS SRTLDFESAF RAASGGAGMD VVLNSLAREF VDASLRLLGP 3181 GGRFVEMGKT DVRDAERVAA DHPGVGYRAF DLGEAGPERI GEMLAEVIAL FEDGVLRHLP 3241 VTTWDVRRAR DAFRHVSQAR HTGKVVLTMP SGLDPEGTVL LTGGTGALGG IVARHVVGEW 3301 GVRRLLLVSR RGTDAPGAGE LVHELEALGA DVSVAACDVA DREALTAVLD SIPAEHPLTA 3361 VVHTAGVLSD GTLPSMTAED VEHVLRPKVD AAFLLDELTS TPGYDLAAFV MFSSAAAVFG 3421 GAGQGAYAAA NATLDALAWR RRTAGLPALS LGWGLWAETS GMTGGLSDTD RSRLARSGAT 3481 PMDSELTLSL LDAAMRRDDP ALVPIALDVA ALRAQQRDGM LAPLLSGLTR GSRVGGAPVN 3541 QRRAAAGGAG EADTDLGGRL AAMTPDDRVA HLRDLVRTHV ATVLGHGTPS RVDLERAFRD 3601 TGFDSLTAVE LRNRLNAATG LRLPATLVFD HPTPGELAGH LLDELATAAG GSWAEGTGSG 3661 DTASATDRQT TAALAELDRL EGVLASLAPA AGGRPELAAR LRALAAALGD DGDDATDLDE 3721 ASDDDLFSFI DKELGDSDF

Amino acid sequence of narbonolide synthase subunit 3, PICAIII 1 MANNEDKLRD YLKRVTAELQ QNTRRLREIE GRTHEPVAIV GMACRLPGGV ASPEDLWQLV 61 AGDGDAISEF PQDRGWDVEG LYDPDPDASG RTYCRSGGFL HDAGEFDADF FGISPREALA 121 MDPQQRLSLT TAWEAIESAG IDPTALKGSG LGVFVGGWHT GYTSGQTTAV QSPELEGHLV 181 SGAALGFLSG RIAYVLGTDG PALTVDTACS SSLVALHLAV QALRKGECDM ALAGGVTVMP 241 NADLFVQFSR QRGLAADGRS KAFATSADGF GPAEGAGVLL VERLSDARRN GHRILAVVRG 301 SAVNQDGASN GLTAPHGPSQ QRVIRRALAD ARLAPGDVDV VEAHGTGTRL GDPIEAQALI 361 ATYGQEKSSE QPLRLGALKS NIGHTQAAAG VAGVIKMVQA MRHGLLPKTL HVDEPSDQID 421 WSAGTVELLT EAVDWPEKQD GGLRRAAVSS FGISGTNAHV VLEEAPAVED SPAVEPPAGG 481 GVVPWPVSAK TPAALDAQIG QLAAYADGRT DVDPAVAARA LVDSRTAMEH RAVAVGDSRE 541 ALRDALRMPE GLVRGTSSDV GRVAFVFPGQ GTQWAGMGAE LLDSSPEFAA SMAECETALS 601 RYVDWSLEAV VRQEPGAPTL DRVDVVQPVT FAVMVSLAKV WQHHGITPQA VVGHSQGEIA 661 AAYVAGALTL DDAARVVTLR SKSIAAHLAG KGGMISLALD EAAVLKRLSD FDGLSVAAVN 721 GPTATVVSGD PTQIEELART CEADGVRARI IPVDYASHSR QVEIIEKELA EVLAGLAPQA 781 PHVPFFSTLE GTWITEPVLD GTYWYRNLRH RVGFAPAVET LAVDGFTHFI EVSAHPVLTM


#### Abstract

-continued 841 TLPETVTGLG TLRREQGGQE RLVTSLAEAW ANGLTIDWAP ILPTATGHHP ELPTYAFQTE 901 RFWLQSSAPT SAADDWRYRV EWKPLTASGQ ADLSGRWIVA VGSEPEAELL GALKAAGAEV 961 DVLEAGADDD REALAARLTA LTTGDGFTGV VSLLDDLVPQ VAWVQALGDA GIKAPLWSVT 1021 QGAVSVGRLD TPADPDRAML WGLGRVVALE HPERWAGLVD LPAQPDAAAL AHLVTALSGA 1081 TGEDQIAIRT TGLHARRLAR APLHGRRPTR DWQPHGTVLI TGGTGALGSH AARWMAHHGA 1141 EHLLLVSRSG EQAPGATQLT AELTASGARV TIAACDVADP HAMRTLLDAI PAETPLTAVV 1201 HTAGAPGGDP LDVTGPEDIA RILGAKTSGA EVLDDLLRGT PLDAFVLYSS NAGVWGSGSQ 1261 GVYAAANAHL DALAARRRAR GETATSVAWG LWAGDGMGRG ADDAYWQRRG IRPMSPDRAL 1321 DELAKALSHD ETFVAVADVD WERFAPAFTV SRPSLLLDGV PEARQALAAP VGAPAPGDAA 1381 VAPTGQSSAL AAITALPEPE RRPALLTLVR THAAAVLGHS SPDRVAPGRA FTELGFDSLT 1441 AVQLRNQLST VVGNRLPATT VFDHPTPAAL AAHLHEAYLA PAEPAPTDWE GRVRRALAEL 1501 PLDRLRDAGV LDTVLRLTGI EPEPGSGGSD GGAADPGAEP EASIDDLDAE ALIRMALGPR 1561 NT

Amino acid sequence of narbonolide synthase subunit 4, PICAIV 1 MTSSNEQLVD ALRASLKENE ELRKESRRRA DRRQEPMAIV GMSCRFAGGI RSPEDLWDAV 61 AAGKDLVSEV PEERGWDIDS LYDPVPGRKG TTYVRNAAFL DDAAGFDAAF FGISPREALA 121 MDPQQRQLLE ASWEVFERAG IDPASVRGTD VGVYVGCGYQ DYAPDIRVAP EGTGGYVVTG 181 NSSAVASGRI AYSLGLEGPA VTVDTACSSS LVALHLALKG LRNGDCSTAL VGGVAVLATP 241 GAFIEFSSQQ AMAADGRTKG FASAADGLAW GEGVAVLLLE RLSDARRKGH RVLAVVRGSA 301 INQDGASNGL TAPHGPSQQR LIRQALADAR LTSSDVDVVE GHGTGTRLGD PIEAQALLAT 361 YGQGRAPGQP LRLGTLKSNI GHTQAASGVA GVIKMVQALR HGVLPKTLHV DEPTDQVDWS 421 AGSVELLTEA VDWPERPGRL RRAGVSAFGV GGTNAHVVLE EAPAVEESPA VEPPAGGGVV 481 PWPVSAKTSA ALDAQIGQLA AYAEDRTDVD PAVAARALVD SRTAMEHRAV AVGDSREALR 541 DALRMPEGLV RGTVTDPGRV AFVFPGQGTQ WAGMGAELLD SSPEFAAAMA ECETALSPYV 601 DWSLEAVVRQ APSAPTLDRV DVVQPVTFAV MVSLAKVWQH HGITPEAVIG HSQGEIAAAY 661 VAGALTLDDA ARVVTLRSKS IAAHLAGKGG MISLALSEEA TRQRIENLHG LSIAAVNGPT 721 ATVVSGDPTQ IQELAQACEA DGIRARIIPV DYASHSAHVE TIENELADVL AGLSPQTPQV 781 PFFSTLEGTW ITEPALDGGY WYRNLRHRVG FAPAVETLAT DEGFTHFIEV SAHPVLTMTL 841 PDKVTGLATL RREDGGQHRL TTSLAEAWAN GLALDWASLL PATGALSPAV PDLPTYAFQH 901 RSYWISPAGP GEAPAHTASG REAVAETGLA WGPGAEDLDE EGRRSAVLAM VMRQAASVLR 961 CDSPEEVPVD RPLREIGFDS LTAVDFRNRV NRLTGLQLPP TVVFEHPTPV ALAERISDEL 1021 AERNWAVAEP SDHEQAEEEK AAAPAGARSG ADTGAGAGMF RALFRQAVED DRYGEFLDVL 1081 AEASAFRPQF ASPEACSERL DPVLLAGGPT DRAEGRAVLV GCTGTAANGG PHEFLRLSTS 1141 FQEERDFLAV PLPGYGTGTG TGTALLPADL DTALDAQARA ILRAAGDAPV VLLGHSGGAL 1201 LAHELAFRLE RAHGAPPAGI VLVDPYPPGH QEPIEVWSRQ LGEGLFAGEL EPMSDARLLA 1261 MGRYARFLAG PRPGRSSAPV LLVRASEPLG DWQEERGDWR AHWDLPHTVA DVPGDHFTMM 1321 RDHAPAVAEA VLSWLDAIEG IEGAGK


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61 RQDRRAEPCL ESVEELAEHV VAATEPWWQE GRLAFFGHSL GASVAFETAR ILEQRHGVRP

121 EGLYVSGRRA PSLAPDRLVH QLDDRAFLAE IRRLSGTDER FLQDDELLRL VLPALRSDYK
181 AAETYLHRPS AKLTCPVMAL AGDRDPKAPL NEVAEWRRHT SGPFCLRAYS GGHFYLNDQW

241 HEICNDISDH LLVTRGAPDA RVVQPPTSLI EGAAKRWQNP R
[0042] The DNA encoding the above proteins can be isolated in recombinant form from the recombinant cosmid pKOS023-27 of the invention, which was deposited with the American Type Culture Collection under the terms of the Budapest Treaty on 20 Aug. 1998 and is available under accession number ATCC 203141. Cosmid pKOS023-27 contains an insert of Streptomyces venezuelae DNA of - 38506 nucleotides. The complete sequence of the insert from cosmid pKOS023-27 is shown below. The location of the various ORFs in the insert, as well as the boundaries of the sequences that encode the various domains of the multiple modules of the PKS, are summarized in the Table below. FIG. 2 shows a restriction site and function map of pKOSO23-27, which contains the complete coding sequence for the four proteins that constitute narbonolide PKS and four additional ORFs. One of these additional ORFs encodes the picB gene product, the type II thioesterase mentioned above. PICB shows a high degree of similarity to other type II thioesterases, with an identity of $51 \%, 49 \%, 45 \%$ and $40 \%$ as compared to those of Amycolatopsis mediterranae, $S$. griseus, $S$. fradiae and Saccharopolyspora erythraea, respectively. The three additional ORFs in the cosmid pKOS023-27 insert DNA sequence, from the picCII, picCIII, and picCVI, genes, are involved in desosamine biosynthesis and transfer and described in the following section.

| From Nucleotide | To Nucleotide | Description |
| :---: | ---: | :--- |
| 70 | 13725 | picAI |
| 70 | 13725 | narbonolide synthase 1 (PICAI) |
| 148 | 3141 | loading module |
| 148 | 1434 | KS loading module |
| 1780 | 2802 | AT loading module |
| 2869 | 3141 | ACP loading module |
| 3208 | 7593 | extender module 1 |
| 3208 | 4497 | KS1 |
| 4828 | 5847 | AT1 |
| 6499 | 7257 | KR1 |
| 7336 | 7593 | ACP1 |
| 7693 | 13332 | extender module 2 |
| 7693 | 8974 | KS2 |

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| From Nucleotide | To Nucleotide | Description |
| :---: | :---: | :--- |
| 9418 | 10554 | AT2 |
| 10594 | 11160 | DH2 |
| 12175 | 12960 | KR2 |
| 13063 | 13332 | ACP2 |
| 13830 | 25049 | picAII |
| 13830 | 25049 | narbonolide synthase 2 (PICAII) |
| 13935 | 18392 | extender module 3 |
| 13935 | 15224 | KS3 |
| 15540 | 16562 | AT3 |
| 17271 | 18071 | KR3 (inactive) |
| 18123 | 18392 | ACP3 |
| 18447 | 24767 | extender module 4 |
| 18447 | 19736 | KS4 |
| 20031 | 21050 | AT4 |
| 21093 | 21626 | DH4 |
| 22620 | 23588 | ER4 |
| 23652 | 24423 | KR4 |
| 24498 | 24765 | ACP4 |
| 25133 | 29821 | picAIII |
| 25133 | 29821 | narbonolide synthase 3 (PICAIII) |
| 25235 | 29567 | extender module 5 |
| 25235 | 36530 | KS5 |
| 26822 | 37439 | 36011 |

[0043]
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361 GCCGCCTTCT TCGGCATCTC GCCCCGCGAG GCCGCGGAGA TGGACCCGCA GCAGCGGCTC 421 GCCCTGGAGC TGGGCTGGGA GGCCCTGGAG CGCGCCGGGA TCGACCCGTC CTCGCTCACC 481 GGCACCCGCA CCGGCGTCTT CGCCGGCGCC ATCTGGGACG ACTACGCCAC CCTGAAGCAC 541 CGCCAGGGCG GCGCCGCGAT CACCCCGCAC ACCGTCACCG GCCTCCACCG CGGCATCATC 601 GCGAACCGAC TCTCGTACAC GCTCGGGCTC CGCGGCCCCA GCATGGTCGT CGACTCCGGC 661 CAGTCCTCGT CGCTCGTCGC CGTCCACCTC GCGTGCGAGA GCCTGCGGCG CGGCGAGTCC 721 GAGCTCGCCC TCGCCGGCGG CGTCTCGCTC AACCTGGTGC CGGACAGCAT CATCGGGGCG 781 AGCAAGTTCG GCGGCCTCTC CCCCGACGGC CGCGCCTACA CCTTCGACGC GCGCGCCAAC 841 GGCTACGTAC GCGGCGAGGG CGGCGGTTTC GTCGTCCTGA AGCGCCTCTC CCGGGCCGTC 901 GCCGACGGCG ACCCGGTGCT CGCCGTGATC CGGGGCAGCG CCGTCAACAA CGGCGGCGCC 961 GCCCAGGGCA TGACGACCCC CGACGCGCAG GCGCAGGAGG CCGTGCTCCG CGAGGCCCAC 1021 GAGCGGGCCG GGACCGCGCC GGCCGACGTG CGGTACGTCG AGCTGCACGG CACCGGCACC 1081 CCCGTGGGCG ACCCGATCGA GGCCGCTGCG CTCGGCGCCG CCCTCGGCAC CGGCCGCCCG 1141 GCCGGACAGC CGCTCCTGGT CGGCTCGGTC AAGACGAACA TCGGCCACCT GGAGGGCGCG 1201 GCCGGCATCG CCGGCCTCAT CAAGGCCGTC CTGGCGGTCC GCGGTCGCGC GCTGCCCGCC 1261 AGCCTGAACT ACGAGACCCC GAACCCGGCG ATCCCGTTCG AGGAACTGAA CCTCCGGGTG 1321 AACACGGAGT ACCTGCCGTG GGAGCCGGAG CACGACGGGC AGCGGATGGT CGTCGGCGTG 1381 TCCTCGTTCG GCATGGGCGG CACGAACGCG CATGTCGTGC TCGAAGAGGC CCCGGGGGTT 1441 GTCGAGGGTG CTTCGGTCGT GGAGTCGACG GTCGGCGGGT CGGCGGTCGG CGGCGGTGTG 1501 GTGCCGTGGG TGGTGTCGGC GAAGTCCGCT GCCGCGCTGG ACGCGCAGAT CGAGCGGCTT 1561 GCCGCGTTCG CCTCGCGGGA TCGTACGGAT GGTGTCGACG CGGGCGCTGT CGATGCGGGT 1621 GCTGTCGATG CGGGTGCTGT CGCTCGCGTA CTGGCCGGCG GGCGTGCTCA GTTCGAGCAC 1681 CGGGCCGTCG TCGTCGGCAG CGGGCCGGAC GATCTGGCGG CAGCGCTGGC CGCGCCTGAG 1741 GGTCTGGTCC GGGGCGTGGC TTCCGGTGTC GGGCGAGTGG CGTTCGTGTT CCCCGGGCAG 1801 GGCACGCAGT GGGCCGGCAT GGGTGCCGAA CTGCTGGACT CTTCCGCGGT GTTCGCGGCG 1861 GCCATGGCCG AATGCGAGGC CGCACTCTCC CCGTACGTCG ACTGGTCGCT GGAGGCCGTC 1921 GTACGGCAGG CCCCCGGTGC GCCCACGCTG GAGCGGGTCG ATGTCGTGCA GCCTGTGACG 1981 TTCGCCGTCA TGGTCTCGCT GGCTCGCGTG TGGCAGCACC ACGGGGTGAC GCCCCAGGCG 2041 GTCGTCGGCC ACTCGCAGGG CGAGATCGCC GCCGCGTACG TCGCCGGTGC CCTGAGCCTG 2101 GACGACGCCG CTCGTGTCGT GACCCTGCGC AGCAAGTCCA TCGCCGCCCA CCTCGCCGGC 2161 AAGGGCGGCA TGCTGTCCCT CGCGCTGAGC GAGGACGCCG TCCTGGAGCG ACTGGCCGGG 2221 TTCGACGGGC TGTCCGTCGC CGCTGTGAAC GGGCCCACCG CCACCGTGGT CTCCGGTGAC 2281 CCCGTACAGA TCGAAGAGCT TGCTCGGGCG TGTGAGGCCG ATGGGGTCCG TGCGCGGGTC 2341 ATTCCCGTCG ACTACGCGTC CCACAGCCGG CAGGTCGAGA TCATCGAGAG CGAGCTCGCC 2401 GAGGTCCTCG CCGGGCTCAG CCCGCAGGCT CCGCGCGTGC CGTTCTTCTC GACACTCGAA 2461 GGCGCCTGGA TCACCGAGCC CGTGCTCGAC GGCGGCTACT GGTACCGCAA CCTGCGCCAT 2521 CGTGTGGGCT TCGCCCCGGC CGTCGAGACC CTGGCCACCG ACGAGGGCTT CACCCACTTC 2581 GTCGAGGTCA CCGCCCACCC CGTCCTCACC ATGGCCCTCC CCGGGACCGT CACCGGTCTG

## 2641 GCGACCCTGC GTCGCGACAA CGGCGGTCAG GACCGCCTCG TCGCCTCCCT CGCCGAAGCA

 2701 TGGGCCAACG GACTCGCGGT CGACTGGAGC CCGCTCCTCC CCTCCGCGAC CGGCCACCAC 2761 TCCGACCTCC CCACCTACGC GTTCCAGACC GAGCGCCACT GGCTGGGCGA GATCGAGGCG 2821 CTCGCCCCGG CGGGCGAGCC GGCGGTGCAG CCCGCCGTCC TCCGCACGGA GGCGGCCGAG 2881 CCGGCGGAGC TCGACCGGGA CGAGCAGCTG CGCGTGATCC TGGACAAGGT CCGGGCGCAG 2941 ACGGCCCAGG TGCTGGGGTA CGCGACAGGC GGGCAGATCG AGGTCGACCG GACCTTCCGT 3001 GAGGCCGGTT GCACCTCCCT GACCGGCGTG GACCTGCGCA ACCGGATCAA CGCCGCCTTC 3061 GGCGTACGGA TGGCGCCGTC CATGATCTTC GACTTCCCCA CCCCCGAGGC TCTCGCGGAG 3121 CAGCTGCTCC TCGTCGTGCA CGGGGAGGCG GCGGCGAACC CGGCCGGTGC GGAGCCGGCT 3181 CCGGTGGCGG CGGCCGGTGC CGTCGACGAG CCGGTGGCGA TCGTCGGCAT GGCCTGCCGC 3241 CTGCCCGGTG GGGTCGCCTC GCCGGAGGAC CTGTGGCGGC TGGTGGCCGG CGGCGGGGAC 3301 GCGATCTCGG AGTTCCCGCA GGACCGCGGC TGGGACGTGG AGGGGCTGTA CCACCCGGAT 3361 CCCGAGCACC CCGGCACGTC GTACGTCCGC CAGGGCGGTT TCATCGAGAA CGTCGCCGGC 3421 TTCGACGCGG CCTTCTTCGG GATCTCGCCG CGCGAGGCCC TCGCCATGGA CCCGCAGCAG 3481 CGGCTCCTCC TCGAAACCTC CTGGGAGGCC GTCGAGGACG CCGGGATCGA CCCGACCTCC 3541 CTGCGGGGAC GGCAGGTCGG CGTCTTCACT GGGGCGATGA CCCACGAGTA CGGGCCGAGC 3601 CTGCGGGACG GCGGGGAAGG CCTCGACGGC TACCTGCTGA CCGGCAACAC GGCCAGCGTG 3661 ATGTCGGGCC GCGTCTCGTA CACACTCGGC CTTGAGGGCC CCGCCCTGAC GGTGGACACG 3721 GCCTGCTCGT CGTCGCTGGT CGCCCTGCAC CTCGCCGTGC AGGCCCTGCG CAAGGGCGAG 3781 GTCGACATGG CGCTCGCCGG CGGCGTGGCC GTGATGCCCA CGCCCGGGAT GTTCGTCGAG 3841 TTCAGCCGGC AGCGCGGGCT GGCCGGGGAC GGCCGGTCGA AGGCGTTCGC CGCGTCGGCG 3901 GACCGGACCA GCTGGTCCGA GGGCGTCGGC GTCCTCCTCG TCGAGCGCCT GTCGGACGCC 3961 CGCCGCAACG GACACCAGGT CCTCGCGGTC GTCCGCGGCA GCGCCGTGAA CCAGGACGGC 4021 GCGAGCAACG GCCTCACGGC TCCGAACGGG CCCTCGCAGC AGCGCGTCAT CCGGCGCGCG 4081 CTGGCGGACG CCCGGCTGAC GACCTCCGAC GTGGACGTCG TCGAGGCACA CGGCACGGGC 4141 ACGCGACTCG GCGACCCGAT CGAGGCGCAG GCCCTGATCG CCACCTACGG CCAGGGCCGT 4201 GACGACGAAC AGCCGCTGCG CCTCGGGTCG TTGAAGTCCA ACATCGGGCA CACCCAGGCC 4261 GCGGCCGGCG TCTCCGGTGT CATCAAGATG GTCCAGGCGA TGCGCCACGG ACTGCTGCCG 4321 AAGACGCTGC ACGTCGACGA GCCCTCGGAC CAGATCGACT GGTCGGCTGG CGCCGTGGAA 4381 CTCCTCACCG AGGCCGTCGA CTGGCCGGAG AAGCAGGACG GCGGGCTGCG CCGGGCCGCC 4441 GTCTCCTCCT TCGGGATCAG CGGCACCAAT GCGCATGTGG TGCTCGAAGA GGCCCCGGTG 4501 GTTGTCGAGG GTGCTTCGGT CGTCGAGCCG TCGGTTGGCG GGTCGGCGGT CGGCGGCGGT 4561 GTGACGCCTT GGGTGGTGTC GGCGAAGTCC GCTGCCGCGC TCGACGCGCA GATCGAGCGG 4621 CTTGCCGCAT TCGCCTCGCG GGATCGTACG GATGACGCCG ACGCCGGTGC TGTCGACGCG 4681 GGCGCTGTCG CTCACGTACT GGCTGACGGG CGTGCTCAGT TCGAGCACCG GGCCGTCGCG 4741 CTCGGCGCCG GGGCGGACGA CCTCGTACAG GCGCTGGCCG ATCCGGACGG GCTGATACGC 4801 GGAACGGCTT CCGGTGTCGG GCGAGTGGCG TTCGTGTTCC CCGGTCAGGG CACGCAGTGG 4861 GCTGGCATGG GTGCCGAACT GCTGGACTCT TCCGCGGTGT TCGCGGCGGC CATGGCCGAG 4921 TGTGAGGCCG CGCTGTCCCC GTACGTCGAC TGGTCGCTGG AGGCCGTCGT ACGGCAGGCC
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4981 CCCGGTGCGC CCACGCTGGA GCGGGTCGAT GTCGTGCAGC CTGTGACGTT CGCCGTCATG 5041 GTCTCGCTGG CTCGCGTGTG GCAGCACCAC GGTGTGACGC CCCAGGCGGT CGTCGGCCAC 5101 TCGCAGGGCG AGATCGCCGC CGCGTACGTC GCCGGAGCCC TGCCCCTGGA CGACGCCGCC 5161 CGCGTCGTCA CCCTGCGCAG CAAGTCCATC GCCGCCCACC TCGCCGGCAA GGGCGGCATG 5221 CTGTCCCTCG CGCTGAACGA GGACGCCGTC CTGGAGCGAC TGAGTGACTT CGACGGGCTG 5281 TCCGTCGCCG CCGTCAACGG GCCCACCGCC ACTGTCGTGT CGGGTGACCC CGTACAGATC 5341 GAAGAGCTTG CTCAGGCGTG CAAGGCGGAC GGATTCCGCG CGCGGATCAT TCCCGTCGAC 5401 TACGCGTCCC ACAGCCGGCA GGTCGAGATC ATCGAGAGCG AGCTCGCCCA GGTCCTCGCC 5461 GGTCTCAGCC CGCAGGCCCC GCGCGTGCCG TTCTTCTCGA CGCTCGAAGG CACCTGGATC 5521 ACCGAGCCCG TCCTCGACGG CACCTACTGG TACCGCAACC TCCGTCACCG CGTCGGCTTC 5581 GCCCCCGCCA TCGAGACCCT GGCCGTCGAC GAGGGCTTCA CGCACTTCGT CGAGGTCAGC 5641 GCCCACCCCG TCCTCACCAT GACCCTCCCC GAGACCGTCA CCGGCCTCGG CACCCTCCGT 5701 CGCGAACAGG GAGGCCAAGA GCGTCTGGTC ACCTCGCTCG CCGAGGCGTG GGTCAACGGG 5761 CTTCCCGTGG CATGGACTTC GCTCCTGCCC GCCACGGCCT CCCGCCCCGG TCTGCCCACC 5821 TACGCCTTCC AGGCCGAGCG CTACTGGCTC GAGAACACTC CCGCCGCCCT GGCCACCGGC 5881 GACGACTGGC GCTACCGCAT CGACTGGAAG CGCCTCCCGG CCGCCGAGGG GTCCGAGCGC 5941 ACCGGCCTGT CCGGCCGCTG GCTCGCCGTC ACGCCGGAGG ACCACTCCGC GCAGGCCGCC 6001 GCCGTGCTCA CCGCGCTGGT CGACGCCGGG GCGAAGGTCG AGGTGCTGAC GGCCGGGGCG 6061 GACGACGACC GTGAGGCCCT CGCCGCCCGG CTCACCGCAC TGACGACCGG TGACGGCTTC 6121 ACCGGCGTGG TCTCGCTCCT CGACGGACTC GTACCGCAGG TCGCCTGGGT CCAGGCGCTC 6181 GGCGACGCCG GAATCAAGGC GCCCCTGTGG TCCGTCACCC AGGGCGCGGT CTCCGTCGGA 6241 CGTCTCGACA CCCCCGCCGA CCCCGACCGG GCCATGCTCT GGGGCCTCGG CCGCGTCGTC 6301 GCCCTTGAGC ACCCCGAACG CTGGGCCGGC CTCGTCGACC TCCCCGCCCA GCCCGATGCC 6361 GCCGCCCTCG CCCACCTCGT CACCGCACTC TCCGGCGCCA CCGGCGAGGA CCAGATCGCC 6421 ATCCGCACCA CCGGACTCCA CGCCCGCCGC CTCGCCCGCG CACCCCTCCA CGGACGTCGG 6481 CCCACCCGCG ACTGGCAGCC CCACGGCACC GTCCTCATCA CCGGCGGCAC CGGAGCCCTC 6541 GGCAGCCACG CCGCACGCTG GATGGCCCAC CACGGAGCCG AACACCTCCT CCTCGTCAGC 6601 CGCAGCGGCG AACAAGCCCC CGGAGCCACC CAACTCACCG CCGAACTCAC CGCATCGGGC 6661 GCCCGCGTCA CCATCGCCGC CTGCGACGTC GCCGACCCCC ACGCCATGCG CACCCTCCTC 6721 GACGCCATCC CCGCCGAGAC GCCCGTCACC GCCGTCGTCC ACACCGCCGG CGCGCTCGAC 6781 GACGGCATCG TGGACACGCT GACCGCCGAG CAGGTCCGGC GGGCCCACCG TGCGAAGGCC 6841 GTCGGCGCCT CGGTGCTCGA CGAGCTGACC CGGGACCTCG ACCTCGACGC GTTCGTGCTC 6901 TTCTCGTCCG TGTCGAGCAC TCTGGGCATC CCCGGTCAGG GCAACTACGC CCCGCACAAC 6961 GCCTACCTCG ACGCCCTCGC GGCTCGCCGC CGGGCCACCG GCCGGTCCGC CGTCTCGGTG 7021 GCCTGGGGAC CGTGGGACGG TGGCGGCATG GCCGCCGGTG ACGGCGTGGC CGAGCGGCTG 7081 CGCAACCACG GCGTGCCCGG CATGGACCCG GAACTCGCCC TGGCCGCACT GGAGTCCGCG 7141 CTCGGCCGGG ACGAGACCGC GATCACCGTC GCGGACATCG ACTGGGACCG CTTCTACCTC 7201 GCGTACTCCT CCGGTCGCCC GCAGCCCCTC GTCGAGGAGC TGCCCGAGGT GCGGCGCATC

## 7261 ATCGACGCAC GGGACAGCGC CACGTCCGGA CAGGGCGGGA GCTCCGCCCA GGGCGCCAAC

 7321 CCCCTGGCCG AGCGGCTGGC CGCCGCGGCT CCCGGCGAGC GTACGGAGAT CCTCCTCGGT 7381 CTCGTACGGG CGCAGGCCGC CGCCGTGCTC CGGATGCGTT CGCCGGAGGA CGTCGCCGCC 7441 GACCGCGCCT TCAAGGACAT CGGCTTCGAC TCGCTCGCCG GTGTCGAGCT GCGCAACAGG 7501 CTGACCCGGG CGACCGGGCT CCAGCTGCCC GCGACGCTCG TCTTCGACCA CCCGACGCCG 7561 CTGGCCCTCG TGTCGCTGCT CCGCAGCGAG TTCCTCGGTG ACGAGGAGAC GGCGGACGCC 7621 CGGCGGTCCG CGGCGCTGCC CGCGACTGTC GGTGCCGGTG CCGGCGCCGG CGCCGGCACC 7681 GATGCCGACG ACGATCCGAT CGCGATCGTC GCGATGAGCT GCCGCTACCC CGGTGACATC 7741 CGCAGCCCGG AGGACCTGTG GCGGATGCTG TCCGAGGGCG GCGAGGGCAT CACGCCGTTC 7801 CCCACCGACC GCGGCTGGGA CCTCGACGGC CTGTACGACG CCGACCCGGA CGCGCTCGGC 7861 AGGGCGTACG TCCGCGAGGG CGGGTTCCTG CACGACGCGG CCGAGTTCGA CGCGGAGTTC 7921 TTCGGCGTCT CGCCGCGCGA GGCGCTGGCC ATGGACCCGC AGCAGCGGAT GCTCCTGACG 7981 ACGTCCTGGG AGGCCTTCGA GCGGGCCGGC ATCGAGCCGG CATCGCTGCG CGGCAGCAGC 8041 ACCGGTGTCT TCATCGGCCT CTCCTACCAG GACTACGCGG CCCGCGTCCC GAACGCCCCG 8101 CGTGGCGTGG AGGGTTACCT GCTGACCGGC AGCACGCCGA GCGTCGCGTC GGGCCGTATC 8161 GCGTACACCT TCGGTCTCGA AGGGCCCGCG ACGACCGTCG ACACCGCCTG CTCGTCGTCG 8221 CTGACCGCCC TGCACCTGGC GGTGCGGGCG CTGCGCAGCG GCGAGTGCAC GATGGCGCTC 8281 GCCGGTGGCG TGGCGATGAT GGCGACCCCG CACATGTTCG TGGAGTTCAG CCGTCAGCGG 8341 GCGCTCGCCC CGGACGGCCG CAGCAAGGCC TTCTCGGCGG ACGCCGACGG GTTCGGCGCC 8401 GCGGAGGGCG TCGGCCTGCT GCTCGTGGAG CGGCTCTCGG ACGCGCGGCG CAACGGTCAC 8461 CCGGTGCTCG CCGTGGTCCG CGGTACCGCC GTCAACCAGG ACGGCGCCAG CAACGGGCTG 8521 ACCGCGCCCA ACGGACCCTC GCAGCAGCGG GTGATCCGGC AGGCGCTCGC CGACGCCCGG 8581 CTGGCACCCG GCGACATCGA CGCCGTCGAG ACGCACGGCA CGGGAACCTC GCTGGGCGAC 8641 CCCATCGAGG CCCAGGGCCT CCAGGCCACG TACGGCAAGG AGCGGCCCGC GGAACGGCCG 8701 CTCGCCATCG GCTCCGTGAA GTCCAACATC GGACACACCC AGGCCGCGGC CGGTGCGGCG 8761 GGCATCATCA AGATGGTCCT CGCGATGCGC CACGGCACCC TGCCGAAGAC CCTCCACGCC 8821 GACGAGCCGA GCCCGCACGT CGACTGGGCG AACAGCGGCC TGGCCCTCGT CACCGAGCCG 8881 ATCGACTGGC CGGCCGGCAC CGGTCCGCGC CGCGCCGCCG TCTCCTCCTT CGGCATCAGC 8941 GGGACGAACG CGCACGTCGT GCTGGAGCAG GCGCCGGATG CTGCTGGTGA GGTGCTTGGG 9001 GCCGATGAGG TGCCTGAGGT GTCTGAGACG GTAGCGATGG CTGGGACGGC TGGGACCTCC 9061 GAGGTCGCTG AGGGCTCTGA GGCCTCCGAG GCCCCCGCGG CCCCCGGCAG CCGTGAGGCG 9121 TCCCTCCCCG GGCACCTGCC CTGGGTGCTG TCCGCCAAGG ACGAGCAGTC GCTGCGCGGC 9181 CAGGCCGCCG CCCTGCACGC GTGGCTGTCC GAGCCCGCCG CCGACCTGTC GGACGCGGAC 9241 GGACCGGCCC GCCTGGCGGGA CGTCGGGTAC ACGCTCGCCA CGAGCCGTAC CGCCTTCGCG 9301 CACCGCGCCG CCGTGACCGC CGCCGACCGG GACGGGTTCC TGGACGGGCT GGCCACGCTG 9361 GCCCAGGGCG GCACCTCGGC CCACGTCCAC CTGGACACCG CCCGGGACGG CACCACCGCG 9421 TTCCTCTTCA CCGGCCAGGG CAGTCAGCGC CCCGGCGCCG GCCGTGAGCT GTACGACCGG 9481 CACCCCGTCT TCGCCCGGGC GCTCGACGAG ATCTGCGCCC ACCTCGACGG TCACCTCGAA 9541 CTGCCCCTGC TCGACGTGAT GTTCGCGGCC GAGGGCAGCG CGGAGGCCGC GCTGCTCGAC
#### Abstract

-continued 9601 GAGACGCGGT ACACGCAGTG CGCGCTGTTC GCCCTGGAGG TCGCGCTCTT CCGGCTCGTC 9661 GAGAGCTGGG GCATGCGGCC GGCCGCACTG CTCGGTCACT CGGTCGGCGA GATCGCCGCC 9721 GCGCACGTCG CCGGTGTGTT CTCGCTCGCC GACGCCGCCC GCCTGGTCGC CGCGCGCGGC 9781 CGGCTCATGC AGGAGCTGCC CGCCGGTGGC GCGATGCTCG CCGTCCAGGC CGCGGAGGAC 9841 GAGATCCGCG TGTGGCTGGA GACGGAGGAG CGGTACGCGG GACGTCTGGA CGTCGCCGCC 9901 GTCAACGGCC CCGAGGCCGC CGTCCTGTCC GGCGACGCGG ACGCGGCGCG GGAGGCGGAG 9961 GCGTACTGGT CCGGGCTCGG CCGCAGGACC CGCGCGCTGC GGGTCAGCCA CGCCTTCCAC 10021 TCCGCGCACA TGGACGGCAT GCTCGACGGG TTCCGCGCCG TCCTGGAGAC GGTGGAGTTC 10081 CGGCGCCCCT CCCTGACCGT GGTCTCGAAC GTCACCGGCC TGGCCGCCGG CCCGGACGAC 10141 CTGTGCGACC CCGAGTACTG GGTCCGGCAC GTCCGCGGCA CCGTCCGCTT CCTCGACGGC 10201 GTCCGTGTCC TGCGCGACCT CGGCGTGCGG ACCTGCCTGG AGCTGGGCCC CGACGGGGTC 10261 CTCACCGCCA TGGCGGCCGA CGGCCTCGCG GACACCCCCG CGGATTCCGC TGCCGGCTCC 10321 CCCGTCGGCT CTCCCGCCGG CTCTCCCGCC GACTCCGCCG CCGGCGCGCT CCGGCCCCGG 10381 CCGCTGCTCG TGGCGCTGCT GCGCCGCAAG CGGTCGGAGA CCGAGACCGT CGCGGACGCC 10441 CTCGGCAGGG CGCACGCCCA CGGCACCGGA CCCGACTGGC ACGCCTGGTT CGCCGGCTCC 10501 GGGGCGCACC GCGTGGACCT GCCCACGTAC TCCTTCCGGC GCGACCGCTA CTGGCTGGAC 10561 GCCCCGGCGG CCGACACCGC GGTGGACACC GCCGGCCTCG GTCTCGGCAC CGCCGACCAC 10621 CCGCTGCTCG GCGCCGTGGT CAGCCTTCCG GACCGGGACG GCCTGCTGCT CACCGGCCGC 10681 CTCTCCCTGC GCACCCACCC GTGGCTCGCG GACCACGCCG TCCTGGGGAG CGTCCTGCTC 10741 CCCGGCGCCG CGATGGTCGA ACTCGCCGCG CACGCTGCGG AGTCCGCCGG TCTGCGTGAC 10801 GTGCGGGAGC TGACCCTCCT TGAACCGCTG GTACTGCCCG AGCACGGTGG CGTCGAGCTG 10861 CGCGTGACGG TCGGGGCGCC GGCCGGAGAG CCCGGTGGCG AGTCGGCCGG GGACGGCGCA 10921 CGGCCCGTCT CCCTCCACTC GCGGCTCGCC GACGCGCCCG CCGGTACCGC CTGGTCCTGC 10981 CACGCGACCG GTCTGCTGGC CACCGACCGG CCCGAGCTTC CCGTCGCGCC CGACCGTGCG 11041 GCCATGTGGC CGCCGCAGGG CGCCGAGGAG GTGCCGCTCG ACGGTCTCTA CGAGCGGCTC 11101 GACGGGAACG GCCTCGCCTT CGGTCCGCTG TTCCAGGGGC TGAACGCGGT GTGGCGGTAC 11161 GAGGGTGAGG TCTTCGCCGA CATCGCGCTC CCCGCCACCA CGAATGCGAC CGCGCCCGCG 11221 ACCGCGAACG GCGGCGGGAG TGCGGCGGCG GCCCCCTACG GCATCCACCC CGCCCTGCTC 11281 GACGCTTCGC TGCACGCCAT CGCGGTCGGC GGTCTCGTCG ACGAGCCCGA GCTCGTCCGC 11341 GTCCCCTTCC ACTGGAGCGG TGTCACCGTG CACGCGGCCG GTGCCGCGGC GGCCCGGGTC 11401 CGTCTCGCCT CCGCGGGGAC GGACGCCGTC TCGCTGTCCC TGACGGACGG CGAGGGACGC 11461 CCGCTGGTCT CCGTGGAACG GCTCACGCTG CGCCCGGTCA CCGCCGATCA GGCGGCGGCG 11521 AGCCGCGTCG GCGGGCTGAT GCACCGGGTG GCCTGGCGTC CGTACGCCCT CGCCTCGTCC 11581 GGCGAACAGG ACCCGCACGC CACTTCGTAC GGGCCGACCG CCGTCCTCGG CAAGGACGAG 11641 CTGAAGGTCG CCGCCGCCCT GGAGTCCGCG GGCGTCGAAG TCGGGCTCTA CCCCGACCTG 11701 GCCGCGCTGT CCCAGGACGT GGCGGCCGGC GCCCCGGCGC CCCGTACCGT CCTTGCGCCG 11761 CTGCCCGCGG GTCCCGCCGA CGGCGGCGCG GAGGGTGTAC GGGGCACGGT GGCCCGGACG 11821 CTGGAGCTGC TCCAGGCCTG GCTGGCCGAC GAGCACCTCG CGGGCACCCG CCTGCTCCTG


11881 GTCACCCGCG GTGCGGTGCG GGACCCCGAG GGGTCCGGCG CCGACGATGG CGGCGAGGAC 11941 CTGTCGCACG CGGCCGCCTG GGGTCTCGTA CGGACCGCGC AGACCGAGAA CCCCGGCCGC 12001 TTCGGCCTTC TCGACCTGGC CGACGACGCC TCGTCGTACC GGACCCTGCC GTCGGTGCTC 12061 TCCGACGCGG GCCTGCGCGA CGAACCGCAG CTCGCCCTGC ACGACGGCAC CATCAGGCTG 12121 GCCCGCCTGG CCTCCGTCCG GCCCGAGACC GGCACCGCCG CACCGGCGCT CGCCCCGGAG 12181 GGCACGGTCC TGCTGACCGG CGGCACCGGC GGCCTGGGCG GACTGGTCGC CCGGCACGTG 12241 GTGGGCGAGT GGGGCGTACG ACGCCTGCTG CTGGTGAGCC GGCGGGGCAC GGACGCCCCG 12301 GGCGCCGACG AGCTCGTGCA CGAGCTGGAG GCCCTGGGAG CCGACGTCTC GGTGGCCGCG 12361 TGCGACGTCG CCGACCGCGA AGCCCTCACC GCCGTACTCG ACGCCATCCC CGCCGAACAC 12421 CCGCTCACCG CGGTCGTCCA CACGGCAGGC GTCCTCTCCG ACGGCACCCT CCCGTCCATG 12481 ACGACGGAGG ACGTGGAACA CGTACTGCGG CCCAAGGTCG ACGCCGCGTT CCTCCTCGAC 12541 GAACTCACCT CGACGCCCGC ATACGACCTG GCAGCGTTCG TCATGTTCTC CTCCGCCGCC 12601 GCCGTCTTCG GTGGCGCGGG GCAGGGCGCC TACGCCGCCG CCAACGCCAC CCTCGACGCC 12661 CTCGCCTGGC GCCGCCGGGC AGCCGGACTC CCCGCCCTCT CCCTCGGCTG GGGCCTCTGG 12721 GCCGAGACCA GCGGCATGAC CGGCGAGCTC GGCCAGGCGG ACCTGCGCCG GATGAGCCGC 12781 GCGGGCATCG GCGGGATCAG CGACGCCGAG GGCATCGCGC TCCTCGACGC CGCCCTCCGC 12841 GACGACCGCC ACCCGGTCCT GCTGCCCCTG CGGCTCGACG CCGCCGGGCT GCGGGACGCG 12901 GCCGGGAACG ACCCGGCCGG AATCCCGGCG CTCTTCCGGG ACGTCGTCGG CGCCAGGACC 12961 GTCCGGGCCC GGCCGTCCGC GGCCTCCGCC TCGACGACAG CCGGGACGGC CGGCACGCCG 13021 GGGACGGCGG ACGGCGCGGC GGAAACGGCG GCGGTCACGC TCGCCGACCG GGCCGCCACC 13081 GTGGACGGGC CCGCACGGCA GCGCCTGCTG CTCGAGTTCG TCGTCGGCGA GGTCGCCGAA 13141 GTACTCGGCC ACGCCCGCGG TCACCGGATC GACGCCGAAC GGGGCTTCCT CGACCTCGGC 13201 TTCGACTCCC TGACCGCCGT CGAACTCCGC AACCGGCTCA ACTCCGCCGG TGGCCTCGCC 13261 CTCCCGGCGA CCCTGGTCTT CGACCACCCA AGCCCGGCGG CACTCGCCTC CCACCTGGAC 13321 GCCGAGCTGC CGCGCGGCGC CTCGGACCAG GACGGAGCCG GGAACCGGAA CGGGAACGAG 13381 AACGGGACGA CGGCGTCCCG GAGCACCGCC GAGACGGACG CGCTGCTGGC ACAACTGACC 13441 CGCCTGGAAG GCGCCTTGGT GCTGACGGGC CTCTCGGACG CCCCCGGGAG CGAAGAAGTC 13501 CTGGAGCACC TGCGGTCCCT GCGCTCGATG GTCACGGGCG AGACCGGGAC CGGGACCGCG 13561 TCCGGAGCCC CGGACGGCGC CGGGTCCGGC GCCGAGGACC GGCCCTGGGC GGCCGGGGAC 13621 GGAGCCGGGG GCGGGAGTGA GGACGGCGCG GGAGTGCCGG ACTTCATGAA CGCCTCGGCC 13681 GAGGAACTCT TCGGCCTCCT CGACCAGGAC CCCAGCACGG ACTGATCCCT GCCGCACGGT 13741 CGCCTCCCGC CCCGGACCCC GTCCCGGGCA CCTCGACTCG AATCACTTCA TGCGCGCCTC 13801 GGGCGCCTCC AGGAACTCAA GGGGACAGCG TGTCCACGGT GAACGAAGAG AAGTACCTCG 13861 ACTACCTGCG TCGTGCCACG GCGGACCTCC ACGAGGCCCG TGGCCGCCTC CGCGAGCTGG 13921 AGGCGAAGGC GGGCGAGCCG GTGGCGATCG TCGGCATGGC CTGCCGCCTG CCCGGCGGCG 13981 TCGCCTCGCC CGAGGACCTG TGGCGGCTGG TGGCCGGCGG CGAGGACGCG ATCTCGGAGT 14041 TCCCCCAGGA CCGCGGCTGG GACGTGGAGG GCCTGTACGA CCCGAACCCG GAGGCCACGG 14101 GCAAGAGTTA CGCCCGCGAG GCCGGATTCC TGTACGAGGC GGGCGAGTTC GACGCCGACT 14161 TCTTCGGGAT CTCGCCGCGC GAGGCCCTCG CCATGGACCC GCAGCAGCGT CTCCTCCTGG
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14221 AGGCCTCCTG GGAGGCGTTC GAGCACGCCG GGATCCCGGC GGCCACCGCG CGCGGCACCT 14281 CGGTCGGCGT CTTCACCGGC GTGATGTACC ACGACTACGC CACCCGTCTC ACCGATGTCC 14341 CGGAGGGCAT CGAGGGCTAC CTGGGCACCG GCAACTCCGG CAGTGTCGCC TCGGGCCGCG 14401 TCGCGTACAC GCTTGGCCTG GAGGGGCCGG CCGTCACGGT CGACACCGCC TGCTCGTCCT 14461 CGCTGGTCGC CCTGCACCTC GCCGTGCAGG CCCTGCGCAA GGGCGAGGTC GACATGGCGC 14521 TCGCCGGCGG CGTGACGGTC ATGTCGACGC CCAGCACCTT CGTCGAGTTC AGCCGTCAGC 14581 GCGGGCTGGC GCCGGACGGC CGGTCGAAGT CCTTCTCGTC GACGGCCGAC GGCACCAGCT 14641 GGTCCGAGGG CGTCGGCGTC CTCCTCGTCG AGCGCCTGTC CGACGCGCGT CGCAAGGGCC 14701 ATCGGATCCT CGCCGTGGTC CGGGGCACCG CCGTCAACCA GGACGGCGCC AGCAGCGGCC 14761 TCACGGCTCC GAACGGGCCG TCGCAGCAGC GCGTCATCCG ACGTGCCCTG GCGGACGCCC 14821 GGCTCACGAC CTCCGACGTG GACGTCGTCG AGGCCCACGG CACGGGTACG CGACTCGGCG 14881 ACCCGATCGA GGCGCAGGCC GTCATCGCCA CGTACGGGCA GGGCCGTGAC GGCGAACAGC 14941 CGCTGCGCCT CGGGTCGTTG AAGTCCAACA TCGGACACAC CCAGGCCGCC GCCGGTGTCT 15001 CCGGCGTGAT CAAGATGGTC CAGGCGATGC GCCACGGCGT CCTGCCGAAG ACGCTCCACG 15061 TGGAGAAGCC GACGGACCAG GTGGACTGGT CCGCGGGCGC GGTCGAGCTG CTCACCGAGG 15121 CCATGGACTG GCCGGACAAG GGCGACGGCG GACTGCGCAG GGCCGCGGTC TCCTCCTTCG 15181 GCGTCAGCGG GACGAACGCG CACGTCGTGC TCGAAGAGGC CCCGGCGGCC GAGGAGACCC 15241 CTGCCTCCGA GGCGACCCCG GCCGTCGAGC CGTCGGTCGG CGCCGGCCTG GTGCCGTGGC 15301 TGGTGTCGGC GAAGACTCCG GCCGCGCTGG ACGCCCAGAT CGGACGCCTC GCCGCGTTCG 15361 CCTCGCAGGG CCGTACGGAC GCCGCCGATC CGGGCGCGGT CGCTCGCGTA CTGGCCGGCG 15421 GGCGCGCCGA GTTCGAGCAC CGGGCCGTCG TGCTCGGCAC CGGACAGGAC GATTTCGCGC 15481 AGGCGCTGAC CGCTCCGGAA GGACTGATAC GCGGCACGCC CTCGGACGTG GGCCGGGTGG 15541 CGTTCGTGTT CCCCGGTCAG GGCACGCAGT GGGCCGGGAT GGGCGCCGAA CTCCTCGACG 15601 TGTCGAAGGA GTTCGCGGCG GCCATGGCCG AGTGCGAGAG CGCGCTCTCC CGCTATGTCG 15661 ACTGGTCGCT GGAGGCCGTC GTCCGGCAGG CGCCGGGCGC GCCCACGCTG GAGCGGGTCG 15721 ACGTCGTCCA GCCCGTGACC TTCGCTGTCA TGGTTTCGCT GGCGAAGGTC TGGCAGCACC 15781 ACGGCGTGAC GCCGCAGGCC GTCGTCGGCC ACTCGCAGGG CGAGATCGCC GCCGCGTACG 15841 TCGCCGGTGC CCTCACCCTC GACGACGCCG CCCGCGTCGT CACCCTGCGC AGCAAGTCCA 15901 TCGCCGCCCA CCTCGCCGGC AAGGGCGGCA TGATCTCCCT CGCCCTCAGC GAGGAAGCCA 15961 CCCGGCAGCG CATCGAGAAC CTCCACGGAC TGTCGATCGC CGCCGTCAAC GGCCCCACCG 16021 CCACCGTGGT TTCGGGCGAC CCCACCCAGA TCCAAGAGCT CGCTCAGGCG TGTGAGGCCG 16081 ACGGGGTCCG CGCACGGATC ATCCCCGTCG ACTACGCCTC CCACAGCGCC CACGTCGAGA 16141 CCATCGAGAG CGAACTCGCC GAGGTCCTCG CCGGGCTCAG CCCGCGGACA CCTGAGGTGC 16201 CGTTCTTCTC GACACTCGAA GGCGCCTGGA TCACCGAGCC GGTGCTCGAC GGCACCTACT 16261 GGTACCGCAA CCTCCGCCAC CGCGTCGGCT TCGCCCCCGC CGTCGAGACC CTCGCCACCG 16321 ACGAAGGCTT CACCCACTTC ATCGAGGTCA GCGCCCACCC CGTCCTCACC ATGACCCTCC 16381 CCGAGACCGT CACCGGCCTC GGCACCCTCC GCCGCGAACA GGGAGGCCAG GAGCGTCTGG 16441 TCACCTCACT CGCCGAAGCC TGGACCAACG GCCTCACCAT CGACTCGGCG CCCGTCCTCC

16501 CCACCGCAAC CGGCCACCAC CCCGAGCTCC CCACCTACGC CTTCCAGCGC CGTCACTACT 16561 GGCTCCACGA CTCCCCCGCC GTCCAGGGCT CCGTGCAGGA CTCCTGGCGC TACCGCATCG 16621 ACTGGAAGCG CCTCGCGGTC GCCGACGCGT CCGAGCGCGC CGGGCTGTCC GGGCGCTGGC 16681 TCGTCGTCGT CCCCGAGGAC CGTTCCGCCG AGGCCGCCCC GGTGCTCGCC GCGCTGTCCG 16741 GCGCCGGCGC CGACCCCGTA CAGCTGGACG TGTCCCCGCT GGGCGACCGG CAGCGGCTCG 16801 CCGCGACGCT GGGCGAGGCC CTGGCGGCGG CCGGTGGAGC CGTCGACGGC GTCCTCTCGC 16861 TGCTCGCGTG GGACGAGAGC GCGCACCCCG GCCACCCCGC CCCCTTCACC CGGGGCACCG 16921 GCGCCACCCT CACCCTGGTG CAGGCGCTGG AGGACGCCGG CGTCGCCGCC CCGCTGTGGT 16981 GCGTGACCCA CGGCGCGGTG TCCGTCGGCC GGGCCGACCA CGTCACCTCC CCCGCCCAGG 17041 CCATGGTGTG GGGCATGGGC CGGGTCGCCG CCCTGGAGCA CCCCGAGCGG TGGGGCGGCC 17101 TGATCGACCT GCCCTCGGAC GCCGACCGGG CGGCCCTGGA CCGCATGACC ACGGTCCTCG 17161 CCGGCGGTAC GGGTGAGGAC CAGGTCGCGG TACGCGCCTC CGGGCTGCTC GCCCGCCGCC 17221 TCGTCCGCGC CTCCCTCCCG GCGCACGGCA CGGCTTCGCC GTGGTGGCAG GCCGACGGCA 17281 CGGTGCTCGT CACCGGTGCC GAGGAGCCTG CGGCCGCCGA GGCCGCACGC CGGCTGGCCC 17341 GCGACGGCGC CGGACACCTC CTCCTCCACA CCACCCCCTC CGGCAGCGAA GGCGCCGAAG 17401 GCACCTCCGG TGCCGCCGAG GACTCCGGCC TCGCCGGGCT CGTCGCCGAA CTCGCGGACC 17461 TGGGCGCGAC GGCCACCGTC GTGACCTGCG ACCTCACGGA CGCGGAGGCG GCCGCCCGGC 17521 TGCTCGCCGG CGTCTCCGAC GCGCACCCGC TCAGCGCCGT CCTCCACCTG CCGCCCACCG 17581 TCGACTCCGA GCCGCTCGCC GCGACCGACG CGGACGCGCT CGCCCGTGTC GTGACCGCGA 17641 AGGCCACCGC CGCGCTCCAC CTGGACCGCC TCCTGCGGGA GGCCGCGGCT GCCGGAGGCC 17701 GTCCGCCCGT CCTGGTCCTC TTCTCCTCGG TCGCCGCGAT CTGGGGCGGC GCCGGTCAGG 17761 GCGCGTACGC CGCCGGTACG GCCTTCCTCG ACGCCCTCGC CGGTCAGCAC CGGGCCGACG 17821 GCCCCACCGT GACCTCGGTG GCCTGGAGCC CCTGGGAGGG CAGCCGCGTC ACCGAGGGTG 17881 CGACCGGGGA GCGGCTGCGC CGCCTCGGCC TGCGCCCCCT CGCCCCCGCG ACGGCGCTCA 17941 CCGCCCTGGA CACCGCGCTC GGCCACGGCG ACACCGCCGT CACGATCGCC GACGTCGACT 18001 GGTCGAGCTT CGCCCCCGGC TTCACCACGG CCCGGCCGGG CACCCTCCTC GCCGATCTGC 18061 CCGAGGCGCG CCGCGCGCTC GACGAGCAGC AGTCGACGAC GGCCGCCGAC GACACCGTCC 18121 TGAGCCGCGA GCTCGGTGCG CTCACCGGCG CCGAACAGCA GCGCCGTATG CAGGAGTTGG 18181 TCCGCGAGCA CCTCGCCGTG GTCCTCAACC ACCCCTCCCC CGAGGCCGTC GACACGGGGC 18241 GGGCCTTCCG TGACCTCGGA TTCGACTCGC TGACGGCGGT CGAGCTCCGC AACCGCCTCA 18301 AGAACGCCAC CGGCCTGGCC CTCCCGGCCA CTCTGGTCTT CGACTACCCG ACCCCCCGGA 18361 CGCTGGCGGA GTTCCTCCTC GCGGAGATCC TGGGCGAGCA GGCCGGTGCC GGCGAGCAGC 18421 TTCCGGTGGA CGGCGGGGTC GACGACGAGC CCGTCGCGAT CGTCGGCATG GCGTGCCGCC 18481 TGCCGGGCGG TGTCGCCTCG CCGGAGGACC TGTGGCGGCT GGTGGCCGGC GGCGAGGACG 18541 CGATCTCCGG CTTCCCGCAG GACCGCGGCT GGGACGTGGA GGGGCTGTAC GACCCGGACC 18601 CGGACGCGTC CGGGCGGACG TACTGCCGTG CCGGTGGCTT CCTCGACGAG GCGGGCGAGT 18661 TCGACGCCGA CTTCTTCGGG ATCTCGCCGC GCGAGGCCCT CGCCATGGAC CCGCAGCAGC 18721 GGCTCCTCCT GGAGACCTCC TGGGAGGCCG TCGAGGACGC CGGGATCGAC CCGACCTCCC 18781 TTCAGGGGCA GCAGGTCGGC GTGTTCGCGG GCACCAACGG CCCCCACTAC GAGCCGCTGC
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18841 TCCGCAACAC CGCCGAGGAT CTTGAGGGTT ACGTCGGGAC GGGCAACGCC GCCAGCATCA 18901 TGTCGGGCCG TGTCTCGTAC ACCCTCGGCC TGGAGGGCCC GGCCGTCACG GTCGACACCG 18961 CCTGCTCCTC CTCGCTGGTC GCCCTGCACC TCGCCGTGCA GGCCCTGCGC AAGGGCGAAT 19021 GCGGACTGGC GCTCGCGGGC GGTGTGACGG TCATGTCGAC GCCCACGACG TTCGTGGAGT 19081 TCAGCCGGCA GCGCGGGCTC GCGGAGGACG GCCGGTCGAA GGCGTTCGCC GCGTCGGCGG 19141 ACGGCTTCGG CCCGGCGGAG GGCGTCGGCA TGCTCCTCGT CGAGCGCCTG TCGGACGCCC 19201 GCCGCAACGG ACACCGTGTG CTGGCGGTCG TGCGCGGCAG CGCGGTCAAC CAGGACGGCG 19261 CGAGCAACGG CCTGACCGCC CCGAACGGGC CCTCGCAGCA GCGCGTCATC CGGCGCGCGC 19321 TCGCGGACGC CCGACTGACG ACCGCCGACG TGGACGTCGT CGAGGCCCAC GGCACGGGCA 19381 CGCGACTCGG CGACCCGATC GAGGCACAGG CCCTCATCGC CACCTACGGC CAGGGGCGCG 19441 ACACCGAACA GCCGCTGCGC CTGGGGTCGT TGAAGTCCAA CATCGGACAC ACCCAGGCCG 19501 CCGCCGGTGT CTCCGGCATC ATCAAGATGG TCCAGGCGAT GCGCCACGGC GTCCTGCCGA 19561 AGACGCTCCA CGTGGACCGG CCGTCGGACC AGATCGACTG GTCGCCGGGC ACGGTCGAGC 19621 TGCTCACCGA GGCCATGGAC TGGCCGAGGA AGCAGGAGGG CGGGCTGCGC CGCGCGGCCG 19681 TCTCCTCCTT CGGCATCAGC GGCACGAACG CGCACATCGT GCTCGAAGAA GCCCCGGTCG 19741 ACGAGGACGC CCCGGCGGAC GAGCCGTCGG TCGGCGGTGT GGTGCCGTGG CTCGTGTCCG 19801 CGAAGACTCC GGCCGCGCTG GACGCCCAGA TCGGACGCCT CGCCGCGTTC GCCTCGCAGG 19861 GCCGTACGGA CGCCGCCGAT CCGGGCGCGG TCGCTCGCGT ACTGGCCGGC GGGCGTGCGC 19921 AGTTCGAGCA CCGGGCCGTC GCGCTCGGCA CCGGACAGGA CGACCTGGCG GCCGCACTGG 19981 CCGCGCCTGA GGGTCTGGTC CGGGGTGTGG CCTCCGGTGT GGGTCGAGTG GCGTTCGTGT 20041 TCCCGGGACA GGGCACGCAG TGGGCCGGGA TGGGTGCCGA ACTCCTCGAC GTGTCGAAGG 20101 AGTTCGCGGC GGCCATGGCC GAGTGCGAGG CCGCGCTCGC TCCGTACGTG GACTGGTCGC 20161 TGGAGGCCGT CGTCCGACAG GCCCCCGGCG CGCCCACGCT GGAGCGGGTC GATGTCGTCC 20221 AGCCCGTGAC GTTCGCCGTC ATGGTCTCGC TGGCGAAGGT CTGGCAGCAC CACGGGGTGA 20281 CCCCGCAAGC CGTCGTCGGC CACTCGCAGG GCGAGATCGC CGCCGCGTAC GTCGCCGGTG 20341 CCCTGAGCCT GGACGACGCC GCTCGTGTCG TGACCCTGCG CAGCAAGTCC ATCGGCGCCC 20401 ACCTCGCGGG CCAGGGCGGC ATGCTGTCCC TCGCGCTGAG CGAGGCGGCC GTTGTGGAGC 20461 GACTGGCCGG GTTCGACGGG CTGTCCGTCG CCGCCGTCAA CGGGCCTACC GCCACCGTGG 20521 TTTCGGGCGA CCCGACCCAG ATCCAAGAGC TCGCTCAGGC GTGTGAGGCC GACGGGGTCC 20581 GCGCACGGAT CATCCCCGTC GACTACGCCT CCCACAGCGC CCACGTCGAG ACCATCGAGA 20641 GCGAACTCGC CGACGTCCTG GCGGGGTTGT CCCCCCAGAC ACCCCAGGTC CCCTTCTTCT 20701 CCACCCTCGA AGGCGCCTGG ATCACCGAAC CCGCCCTCGA CGGCGGCTAC TGGTACCGCA 20761 ACCTCCGCCA TCGTGTGGGC TTCGCCCCGG CCGTCGAAAC CCTGGCCACC GACGAAGGCT 20821 TCACCCACTT CGTCGAGGTC AGCGCCCACC CCGTCCTCAC CATGGCCCTG CCCGAGACCG 20881 TCACCGGCCT CGGCACCCTC CGCCGTGACA ACGGCGGACA GCACCGCCTC ACCACCTCCC 20941 TCGCCGAGGC CTGGGCCAAC GGCCTCACCG TCGACTGGGC CTCTCTCCTC CCCACCACGA 21001 CCACCCACCC CGATCTGCCC ACCTACGCCT TCCAGACCGA GCGCTACTGG CCGCAGCCCG 21061 ACCTCTCCGC CGCCGGTGAC ATCACCTCCG CCGGTCTCGG GGCGGCCGAG CACCCGCTGC

21121 TCGGCGCGGC CGTGGCGCTC GCGGACTCCG ACGGCTGCCT GCTCACGGGG AGCCTCTCCC 21181 TCCGTACGCA CCCCTGGCTG GCGGACCACG CGGTGGCCGG CACCGTGCTG CTGCCGGGAA 21241 CGGCGTTCGT GGAGCTGGCG TTCCGAGCCG GGGACCAGGT CGGTTGCGAT CTGGTCGAGG 21301 AGCTCACCCT CGACGCGCCG CTCGTGCTGC CCCGTCGTGG CGCGGTCCGT GTGCAGCTGT 21361 CCGTCGGCGC GAGCGACGAG TCCGGGCGTC GTACCTTCGG GCTCTACGCG CACCCGGAGG 21421 ACGCGCCGGG CGAGGCGGAG TGGACGCGGC ACGCCACCGG TGTGCTGGCC GCCCGTGCGG 21481 ACCGCACCGC CCCCGTCGCC GACCCGGAGG CCTGGCCGCC GCCGGGCGCC GAGCCGGTGG 21541 ACGTGGACGG TCTGTACGAG CGCTTCGCGG CGAACGGCTA CGGCTACGGC CCCCTCTTCC 21601 AGGGCGTCCG TGGTGTCTGG CGGCGTGGCG ACGAGGTGTT CGCCGACGTG GCCCTGCCGG 21661 CCGAGGTCGC CGGTGCCGAG GGCGCGCGGT TCGGCCTTCA CCCGGCGCTG CTCGACGCCG 21721 CCGTGCAGGC GGCCGGTGCG GGCGGGGCGT TCGGCGCGGG CACGCGGCTG CCGTTCGCCT 21781 GGAGCGGGAT CTCCCTGTAC GCGGTCGGCG CCACCGCCCT CCGCGTGCGG CTGGCCCCCG 21841 CCGGCCCGGA CACGGTGTCC GTGAGCGCCG CCGACTCCTC CGGGCAGCCG GTGTTCGCCG 21901 CGGACTCCCT CACGGTGCTG CCCGTCGACC CCGCGCAGCT GGCGGCCTTC AGCGACCCGA 21961 CTCTGGACGC GCTGCACCTG CTGGAGTGGA CCGCCTGGGA CGGTGCCGCG CACGCCCTGC 22021 CCGGCGCGGT CGTGCTGGGC GGCGACGCCG ACGGTCTCGC CGCGGCGCTG CGCGCCGGTG 22081 GCACCGAGGT CCTGTCCTTC CCGGACCTTA CGGACCTGGT GGAGGCCGTC GACCGGGGCG 22141 AGACCCCGGC CCCGGCGACC GTCCTGGTGG CCTGCCCCGC CGCCGGCCCC GGTGGGCCGG 22201 AGCATGTCCG CGAGGCCCTG CACGGGTCGC TCGCGCTGAT GCAGGCCTGG CTGGCCGACG 22261 AGCGGTTCAC CGATGGGCGC CTGGTGCTCG TGACCCGCGA CGCGGTCGCC GCCCGTTCCG 22321 GCGACGGCCT GCGGTCCACG GGACAGGCCG CCGTCTGGGG CCTCGGCCGG TCCGCGCAGA 22381 CGGAGAGCCC GGGCCGGTTC GTCCTGCTCG ACCTCGCCGG GGAAGCCCGG ACGGCCGGGG 22441 ACGCCACCGC CGGGGACGGC CTGACGACCG GGGACGCCAC CGTCGGCGGC ACCTCTGGAG 22501 ACGCCGCCCT CGGCAGCGCC CTCGCGACCG CCCTCGGCTC GGGCGAGCCG CAGCTCGCCC 22561 TCCGGGACGG GGCGCTCCTC GTACCCCGCC TGGCGCGGGC CGCCGCGCCC GCCGCGGCCG 22621 ACGGCCTCGC CGCGGCCGAC GGCCTCGCCG CTCTGCCGCT GCCCGCCGCT CCGGCCCTCT 22681 GGCGTCTGGA GCCCGGTACG GACGGCAGCC TGGAGAGCCT CACGGCGGCG CCCGGCGACG 22741 CCGAGACCCT CGCCCCGGAG CCGCTCGGCC CGGGACAGGT CCGCATCGCG ATCCGGGCCA 22801 CCGGTCTCAA CTTCCGCGAC GTCCTGATCG CCCTCGGCAT GTACCCCGAT CCGGCGCTGA 22861 TGGGCACCGA GGGAGCCGGC GTGGTCACCG CGACCGGCCC CGGCGTCACG CACCTCGCCC 22921 CCGGCGACCG GGTCATGGGC CTGCTCTCCG GCGCGTACGC CCCGGTCGTC GTGGCGGACG 22981 CGCGGACCGT CGCGCGGATG CCCGAGGGGT GGACGTTCGC CCAGGGCGCC TCCGTGCCGG 23041 TGGTGTTCCT GACGGCCGTC TACGCCCTGC GCGACCTGGC GGACGTCAAG CCCGGCGAGC 23101 GCCTCCTGGT CCACTCCGCC GCCGGTGGCG TGGGCATGGC CGCCGTGCAG CTCGCCCGGC 23161 ACTGGGGCGT GGAGGTCCAC GGCACGGCGA GTCACGGGAA GTGGGACGCC CTGCGCGCGC 23221 TCGGCCTGGA CGACGCGCAC ATCGCCTCCT CCCGCACCCT GGACTTCGAG TCCGCGTTCC 23281 GTGCCGCTTC CGGCGGGGCG GGCATGGACG TCGTACTGAA CTCGCTCGCC CGCGAGTTCG 23341 TCGACGCCTC GCTGCGCCTG CTCGGGCCGG GCGGCCGGTT CGTGGAGATG GGGAAGACCG 23401 ACGTCCGCGA CGCGGAGCGG GTCGCCGCCG ACCACCCCGG TGTCGGCTAC CGCGCCTTCG


#### Abstract

23461 ACCTGGGCGA GGCCGGGCCG GAGCGGATCG GCGAGATGCT CGCCGAGGTC ATCGCCCTCT 23521 TCGAGGACGG GGTGCTCCGG CACCTGCCCG TCACGACCTG GGACGTGCGC CGGGCCCGCG 23581 ACGCCTTCCG GCACGTCAGC CAGGCCCGCC ACACGGGCAA GGTCGTCCTC ACGATGCCGT 23641 CGGGCCTCGA CCCGGAGGGT ACGGTCCTGC TGACCGGCGG CACCGGTGCG CTGGGGGGCA 23701 TCGTGGCCCG GCACGTGGTG GGCGAGTGGG GCGTACGACG CCTGCTGCTC GTGAGCCGGC 23761 GGgGCACGGA CGCCCCGGGC GCCGGCGAGC TCGTGCACGA GCTGGAGGCC CTGGGAGCCG 23821 ACGTCTCGGT GGCCGCGTGC GACGTCGCCG ACCGCGAAGC CCTCACCGCC GTACTCGACT 23881 CGATCCCCGC CGAACACCCG CTCACCGCGG TCGTCCACAC GGCAGGCGTC CTCTCCGACG 23941 GCACCCTCCC CTCGATGACA GCGGAGGATG TGGAACACGT ACTGCGTCCC AAGGTCGACG 24001 CCGCGTTCCT CCTCGACGAA CTCACCTCGA CGCCCGGCTA CGACCTGGCA GCGTTCGTCA 24061 TGTTCTCCTC CGCCGCCGCC GTCTTCGGTG GCGCGGGGCA GGGCGCCTAC GCCGCCGCCA 24121 ACGCCACCCT CGACGCCCTC GCCTGGCGCC GCCGGACAGC CGGACTCCCC GCCCTCTCCC 24181 TCGGCTGGGG CCTCTGGGCC GAGACCAGCG GCATGACCGG CGGACTCAGC GACACCGACC 24241 GCTCGCGGCT GGCCCGTTCC GGGGCGACGC CCATGGACAG CGAGCTGACC CTGTCCCTCC 24301 TGGACGCGGC CATGCGCCGC GACGACCCGG CGCTCGTCCC GATCGCCCTG GACGTCGCCG 24361 CGCTCCGCGC CCAGCAGCGC GACGGCATGC TGGCGCCGCT GCTCAGCGGG CTCACCCGCG 24421 GATCGCGGGT CGGCGGCGCG CCGGTCAACC AGCGCAGGGC AGCCGCCGGA GGCGCGGGCG 24481 AGGCGGACAC GGACCTCGGC GGGCGGCTCG CCGCGATGAC ACCGGACGAC CGGGTCGCGC 24541 ACCTGCGGGA CCTCGTCCGT ACGCACGTGG CGACCGTCCT GGGACACGGC ACCCCGAGCC 24601 GGGTGGACCT GGAGCGGGCC TTCCGCGACA CCGGTTTCGA CTCGCTCACC GCCGTCGAAC 24661 TCCGCAACCG TCTCAACGCC GCGACCGGGC TGCGGCTGCC GGCCACGCTG GTCTTCGACC 24721 ACCCCACCCC GGGGGAGCTC GCCGGGCACC TGCTCGACGA ACTCGCCACG GCCGCGGGCG 24781 GGTCCTGGGC GGAAGGCACC GGGTCCGGAG ACACGGCCTC GGCGACCGAT CGGCAGACCA 24841 CGGCGGCCCT CGCCGAACTC GACCGGCTGG AAGGCGTGCT CGCCTCCCTC GCGCCCGCCG 24901 CCGGCGGCCG TCCGGAGCTC GCCGCCCGGC TCAGGGCGCT GGCCGCGGCC CTGGGGGACG 24961 ACGGCGACGA CGCCACCGAC CTGGACGAGG CGTCCGACGA CGACCTCTTC TCCTTCATCG 25021 ACAAGGAGCT GGGCGACTCC GACTTCTGAC CTGCCCGACA CCACCGGCAC CACCGGCACC 25081 ACCAGCCCCC CTCACACACG GAACACGGAA CGGACAGGCG AGAACGGGAG CCATGGCGAA 25141 CAACGAAGAC AAGCTCCGCG ACTACCTCAA GCGCGTCACC GCCGAGCTGC AGCAGAACAC 25201 CAGGCGTCTG CGCGAGATCG AGGGACGCAC GCACGAGCCG GTGGCGATCG TGGGCATGGC 25261 CTGCCGCCTG CCGGGCGGTG TCGCCTCGCC CGAGGACCTG TGGCAGCTGG TGGCCGGGGA 25321 CGGGGACGCG ATCTCGGAGT TCCCGCAGGA CCGCGGCTGG GACGTGGAGG GGCTGTACGA 25381 CCCCGACCCG GACGCGTCCG GCAGGACGTA CTGCCGGTCC GGCGGATTCC TGCACGACGC 25441 CGGCGAGTTC GACGCCGACT TCTTCGGGAT CTCGCCGCGC GAGGCCCTCG CCATGGACCC 25501 GCAGCAGCGA CTGTCCCTCA CCACCGCGTG GGAGGCGATC GAGAGCGCGG GCATCGACCC 25561 GACGGCCCTG AAGGGCAGCG GCCTCGGCGT CTTCGTCGGC GGCTGGCACA CCGGCTACAC 25621 CTCGGGGCAG ACCACCGCCG TGCAGTCGCC CGAGCTGGAG GGCCACCTGG TCAGCGGCGC 25681 GGCGCTGGGC TTCCTGTCCG GCCGTATCGC GTACGTCCTC GGTACGGACG GACCGGCCCT


25741 GACCGTGGAC ACGGCCTGCT CGTCCTCGCT GGTCGCCCTG CACCTCGCCG TGCAGGCCCT 25801 CCGCAAGGGC GAGTGCGACA TGGCCCTCGC CGGTGGTGTC ACGGTCATGC CCAACGCGGA 25861 CCTGTTCGTG CAGTTCAGCC GGCAGCGCGG GCTGGCCGCG GACGGCCGGT CGAAGGCGTT 25921 CGCCACCTCG GCGGACGGCT TCGGCCCCGC GGAGGGCGCC GGAGTCCTGC TGGTGGAGCG 25981 CCTGTCGGAC GCCCGCCGCA ACGGACACCG GATCCTCGCG GTCGTCCGCG GCAGCGCGGT 26041 CAACCAGGAC GGCGCCAGCA ACGGCCTCAC GGCTCCGCAC GGGCCCTCCC AGCAGCGCGT 26101 CATCCGACGG GCCCTGGCGG ACGCCCGGCT CGCGCCGGGT GACGTGGACG TCGTCGAGGC 26161 GCACGGCACG GGCACGCGGC TCGGCGACCC GATCGAGGCG CAGGCCCTCA TCGCCACCTA 26221 CGGCCAGGAG AAGAGCAGCG AACAGCCGCT GAGGCTGGGC GCGTTGAAGT CGAACATCGG 26281 GCACACGCAG GCCGCGGCCG GTGTCGCAGG TGTCATCAAG ATGGTCCAGG CGATGCGCCA 26341 CGGACTGCTG CCGAAGACGC TGCACGTCGA CGAGCCCTCG GACCAGATCG ACTGGTCGGC 26401 GGGCACGGTG GAACTCCTCA CCGAGGCCGT CGACTGGCCG GAGAAGCAGG ACGGCGGGCT 26461 GCGCCGCGCG GCTGTCTCCT CCTTCGGCAT CAGCGGGACG AACGCGCACG TCGTCCTGGA 26521 GGAGGCCCCG GCGGTCGAGG ACTCCCCGGC CGTCGAGCCG CCGGCCGGTG GCGGTGTGGT 26581 GCCGTGGCCG GTGTCCGCGA AGACTCCGGC CGCGCTGGAC GCCCAGATCG GGCAGCTCGC 26641 CGCGTACGCG GACGGTCGTA CGGACGTGGA TCCGGCGGTG GCCGCCCGCG CCCTGGTCGA 26701 CAGCCGTACG GCGATGGAGC ACCGCGCGGT CGCGGTCGGC GACAGCCGGG AGGCACTGCG 26761 GGACGCCCTG CGGATGCCGG AAGGACTGGT ACGCGGCACG TCCTCGGACG TGGGCCGGGT 26821 GGCGTTCGTC TTCCCCGGCC AGGGCACGCA GTGGGCCGGC ATGGGCGCCG AACTCCTTGA 26881 CAGCTCACCG GAGTTCGCTG CCTCGATGGC CGAATGCGAG ACCGCGCTCT CCCGCTACGT 26941 CGACTGGTCT CTTGAAGCCG TCGTCCGACA GGAACCCGGC GCACCCACGC TCGACCGCGT 27001 CGACGTCGTC CAGCCCGTGA CCTTCGCTGT CATGGTCTCG CTGGCGAAGG TCTGGCAGCA 27061 CCACGGCATC ACCCCCCAGG CCGTCGTCGG CCACTCGCAG GGCGAGATCG CCGCCGCGTA 27121 CGTCGCCGGT GCACTCACCC TCGACGACGC CGCCCGCGTC GTCACCCTGC GCAGCAAGTC 27181 CATCGCCGCC CACCTCGCCG GCAAGGGCGG CATGATCTCC CTCGCCCTCG ACGAGGCGGC 27241 CGTCCTGAAG CGACTGAGCG ACTTCGACGG ACTCTCCGTC GCCGCCGTCA ACGGCCCCAC 27301 CGCCACCGTC GTCTCCGGCG ACCCGACCCA GATCGAGGAA CTCGCCCGCA CCTGCGAGGC 27361 CGACGGCGTC CGTGCGCGGA TCATCCCGGT CGACTACGCC TCCCACAGCC GGCAGGTCGA 27421 GATCATCGAG AAGGAGCTGG CCGAGGTCCT CGCCGGACTC GCCCCGCAGG CTCCGCACGT 27481 GCCGTTCTTC TCCACCCTCG AAGGCACCTG GATCACCGAG CCGGTGCTCG ACGGCACCTA 27541 CTGGTACCGC AACCTGCGCC ATCGCGTGGG CTTCGCCCCC GCCGTGGAGA CCTTGGCGGT 27601 TGACGGCTTC ACCCACTTCA TCGAGGTCAG CGCCCACCCC GTCCTCACCA TGACCCTCCC 27661 CGAGACCGTC ACCGGCCTCG GCACCCTCCG CCGCGAACAG GGAGGCCAGG AGCGTCTGGT 27721 CACCTCACTC GCGGAAGCCT GGGCCAACGG CCTCACCATC GACTGGGCGC CCATCCTCCC 27781 CACCGCAACC GGCCACCACC CCGAGCTCCC CACCTACGCC TTCCAGACCG AGCGCTTCTG 27841 GCTGCAGAGC TCCGCGCCCA CCAGCGCCGC CGACGACTGG CGTTACCGCG TCGAGTGGAA 27901 GCCGCTGACG GCCTCCGGCC AGGCGGACCT GTCCGGGCGG TGGATCGTCG CCGTCGGGAG 27961 CGAGCCAGAA GCCGAGCTGC TGGGCGCGCT GAAGGCCGCG GGAGCGGAGG TCGACGTACT 28021 GGAAGCCGGG GCGGACGACG ACCGTGAGGC CCTCGCCGCC CGGCTCACCG CACTGACGAC


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30361 CCGCGGCACC GACGTCGGCG TGTACGTGGG CTGTGGCTAC CAGGACTACG CGCCGGACAT 30421 CCGGGTCGCC CCCGAAGGCA CCGGCGGTTA CGTCGTCACC GGCAACTCCT CCGCCGTGGC 30481 CTCCGGGCGC ATCGCGTACT CCCTCGGCCT GGAGGGACCC GCCGTGACCG TGGACACGGC 30541 GTGCTCCTCT TCGCTCGTCG CCCTGCACCT CGCCCTGAAG GGCCTGCGGA ACGGCGACTG 30601 CTCGACGGCA CTCGTGGGCG GCGTGGCCGT CCTCGCGACG CCGGGCGCGT TCATCGAGTT 30661 CAGCAGCCAG CAGGCCATGG CCGCCGACGG CCGGACCAAG GGCTTCGCCT CGGCGGCGGA 30721 CGGCCTCGCC TGGGGCGAGG GCGTCGCCGT ACTCCTCCTC GAACGGCTCT CCGACGCGCG 30781 GCGCAAGGGC CACCGGGTCC TGGCCGTCGT GCGCGGCAGC GCCATCAACC AGGACGGCGC 30841 GAGCAACGGC CTCACGGCTC CGCACGGGCC CTCCCAGCAG CGCCTGATCC GCCAGGCCCT 30901 GGCCGACGCG CGGCTCACGT CGAGCGACGT GGACGTCGTG GAGGGCCACG GCACGGGGAC 30961 CCGTCTCGGC GACCCGATCG AGGCGCAGGC GCTGCTCGCC ACGTACGGGC AGGGGCGCGC 31021 CCCGGGGCAG CCGCTGCGGC TGGGGACGCT GAAGTCGAAC ATCGGGCACA CGCAGGCCGC 31081 TTCGGGTGTC GCCGGTGTCA TCAAGATGGT GCAGGCGCTG CGCCACGGGG TGCTGCCGAA 31141 GACCCTGCAC GTGGACGAGC CGACGGACCA GGTCGACTGG TCGGCCGGTT CGGTCGAGCT 31201 GCTCACCGAG GCCGTGGACT GGCCGGAGCG GCCGGGCCGG CTCCGCCGGG CGGGCGTCTC 31261 CGCGTTCGGC GTGGGCGGGA CGAACGCGCA CGTCGTCCTG GAGGAGGCCC CGGCGGTCGA 31321 GGAGTCCCCT GCCGTCGAGC CGCCGGCCGG TGGCGGCGTG GTGCCGTGGC CGGTGTCCGC 31381 GAAGACCTCG GCCGCACTGG ACGCCCAGAT CGGGCAGCTC GCCGCATACG CGGAAGACCG 31441 CACGGACGTG GATCCGGCGG TGGCCGCCCG CGCCCTGGTC GACAGCCGTA CGGCGATGGA 31501 GCACCGCGCG GTCGCGGTCG GCGACAGCCG GGAGGCACTG CGGGACGCCC TGCGGATGCC 31561 GGAAGGACTG GTACGGGGCA CGGTCACCGA TCCGGGCCGG GTGGCGTTCG TCTTCCCCGG 31621 CCAGGGCACG CAGTGGGCCG GCATGGGCGC CGAACTCCTC GACAGCTCAC CCGAATTCGC 31681 CGCCGCCATG GCCGAATGCG AGACCGCACT CTCCCCGTAC GTCGACTGGT CTCTCGAAGC 31741 CGTCGTCCGA CAGGCTCCCA GCGCACCGAC ACTCGACCGC GTCGACGTCG TCCAGCCCGT 31801 CACCTTCGCC GTCATGGTCT CCCTCGCCAA GGTCTGGCAG CACCACGGCA TCACCCCCGA 31861 GGCCGTCATC GGCCACTCCC AGGGCGAGAT CGCCGCCGCG TACGTCGCCG GTGCCCTCAC 31921 CCTCGACGAC GCCGCTCGTG TCGTGACCCT CCGCAGCAAG TCCATCGCCG CCCACCTCGC 31981 CGGCAAGGGC GGCATGATCT CCCTCGCCCT CAGCGAGGAA GCCACCCGGC AGCGCATCGA 32041 GAACCTCCAC GGACTGTCGA TCGCCGCCGT CAACGGGCCT ACCGCCACCG TGGTTTCGGG 32101 CGACCCCACC CAGATCCAAG AACTTGCTCA GGCGTGTGAG GCCGACGGCA TCCGCGCACG 32161 GATCATCCCC GTCGACTACG CCTCCCACAG CGCCCACGTC GAGACCATCG AGAACGAACT 32221 CGCCGACGTC CTGGCGGGGT TGTCCCCCCA GACACCCCAG GTCCCCTTCT TCTCCACCCT 32281 CGAAGGCACC TGGATCACCG AACCCGCCCT CGACGGCGGC TACTGGTACC GCAACCTCCG 32341 CCATCGTGTG GGCTTCGCCC CGGCCGTCGA GACCCTCGCC ACCGACGAAG GCTTCACCCA 32401 CTTCATCGAG GTCAGCGCCC ACCCCGTCCT CACCATGACC CTCCCCGACA AGGTCACCGG 32461 CCTGGCCACC CTCCGACGCG AGGACGGCGG ACAGCACCGC CTCACCACCT CCCTTGCCGA 32521 GGCCTGGGCC AACGGCCTCG CCCTCGACTG GGCCTCCCTC CTGCCCGCCA CGGGCGCCCT 32581 CAGCCCCGCC GTCCCCGACC TCCCGACGTA CGCCTTCCAG CACCGCTCGT ACTGGATCAG 32641 CCCCGCGGGT CCCGGCGAGG CGCCCGCGCA CACCGCTTCC GGGCGCGAGG CCGTCGCCGA

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32701 GACGGGGCTC GCGTGGGGCC CGGCTGCCGA GGACCTCGAC GAGGAGGGCC GGCGCAGCGC 32761 CGTACTCGCG ATGGTGATGC GGCAGGCGGC CTCCGTGCTC CGGTGCGACT CGCCCGAAGA 32821 GGTCCCCGTC GACCGCCCGC TGCGGGAGAT CGGCTTCGAC TCGCTGACCG CCGTCGACTT 32881 CCGCAACCGC GTCAACCGGC TGACCGGTCT CCAGCTGCCG CCCACCGTCG TGTTCGAGCA 32941 CCCGACGCCC GTCGCGCTCG CCGAGCGCAT CAGCGACGAG CTGGCCGAGC GGAACTGGGC 33001 CGTCGCCGAG CCGTCGGATC ACGAGCAGGC GGAGGAGGAG AAGGCCGCCG CTCCGGCGGG 33061 GGCCCGCTCC GGGGCCGACA CCGGCGCCGG CGCCGGGATG TTCCGCGCCC TGTTCCGGCA 33121 GGCCGTGGAG GACGACCGGT ACGGCGAGTT CCTCGACGTC CTCGCCGAAG CCTCCGCGTT 33181 CCGCCCGCAG TTCGCCTCGC CCGAGGCCTG CTCGGAGCGG CTCGACCCGG TGCTGCTCGC 33241 CGGCGGTCCG ACGGACCGGG CGGAAGGCCG TGCCGTTCTC GTCGGCTGCA CCGGCACCGC 33301 GGCGAACGGC GGCCCGCACG AGTTCCTGCG GCTCAGCACC TCCTTCCAGG AGGAGCGGGA 33361 CTTCCTCGCC GTACCTCTCC CCGGCTACGG CACGGGTACG GGCACCGGCA CGGCCCTCCT 33421 CCCGGCCGAT CTCGACACCG CGCTCGACGC CCAGGCCCGG GCGATCCTCC GGGCCGCCGG 33481 GGACGCCCCG GTCGTCCTGC TCGGGCACTC CGGCGGCGCC CTGCTCGCGC ACGAGCTGGC 33541 CTTCCGCCTG GAGCGGGCGC ACGGCGCGCC GCCGGCCGGG ATCGTCCTGG TCGACCCCTA 33601 TCCGCCGGGC CATCAGCAGC CCATCGAGGT GTGGAGCAGG CAGCTGGGCG AGGGCCTGTT 33661 CGCGGGCGAG CTGGAGCCGA TGTCCGATGC GCGGCTGCTG GCCATGGGCC GGTACGCGCG 33721 GTTCCTCGCC GGCCCGCGGC CGGGCCGCAG CAGCGCGCCC GTGCTTCTGG TCCGTGCCTC 33781 CGAACCGCTG GGCGACTGGC AGGAGGAGCG GGGCGACTGG CGTGCCCACT GGGACCTTCC 33841 GCACACCGTC GCGGACGTGC CGGGCGACCA CTTCACGATG ATGCGGGACC ACGCGCCGGC 33901 CGTCGCCGAG GCCGTCCTCT CCTGGCTCGA CGCCATCGAG GGCATCGAGG GGGCGGGCAA 33961 GTGACCGACA GACCTCTGAA CGTGGACAGC GGACTGTGGA TCCGGCGCTT CCACCCCGCG 34021 CCGAACAGCG CGGTGCGGCT GGTCTGCCTG CCGCACGCCG GCGGCTCCGC CAGCTACTTC 34081 TTCCGCTTCT CGGAGGAGCT GCACCCCTCC GTCGAGGCCC TGTCGGTGCA GTATCCGGGC 34141 CGCCAGGACC GGCGTGCCGA GCCGTGTCTG GAGAGCGTCG AGGAGCTCGC CGAGCATGTG 34201 GTCGCGGCCA CCGAACCCTG GTGGCAGGAG GGCCGGCTGG CCTTCTTCGG GCACAGCCTC 34261 GGCGCCTCCG TCGCCTTCGA GACGGCCCGC ATCCTGGAAC AGCGGCACGG GGTACGGCCC 34321 GAGGGCCTGT ACGTCTCCGG TCGGCGCGCC CCGTCGCTGG CGCCGGACCG GCTCGTCCAC 34381 CAGCTGGACG ACCGGGCGTT CCTGGCCGAG ATCCGGCGGC TCAGCGGCAC CGACGAGCGG 34441 TTCCTCCAGG ACGACGAGCT GCTGCGGCTG GTGCTGCCCG CGCTGCGCAG CGACTACAAG 34501 GCGGCCGAGA CGTACCTGCA CCGGCCGTCC GCCAAGCTCA CCTGCCCGGT GATGGCCCTG 34561 GCCGGCCACC GTGACCCGAA GGCGCCGCTG AACGAGGTGG CCGAGTGGCG TCGGCACACC 34621 AGCGGGCCGT TCTGCCTCCG GGCGTACTCC GGCGGCCACT TCTACCTCAA CGACCAGTGG 34681 CACGAGATCT GCAACGACAT CTCCGACCAC CTGCTCGTCA CCCGCGGCGC GCCCGATGCC 34741 CGCGTCGTGC AGCCCCCGAC CAGCCTTATC GAAGGAGCGG CGAAGAGATG GCAGAACCCA 34801 CGGTGACCGA CGACCTGACG GGGGCCCTCA CGCAGCCCCC GCTGGGCCGC ACCGTCCGCG 34861 CGGTGGCCGA CCGTGAACTC GGCACCCACC TCCTGGAGAC CCGCGGCATC CACTGGATCC 34921 ACGCCGCGAA CGGCGACCCG TACGCCACCG TGCTGCGCGG CCAGGCGGAC GACCCGTATC

## 34981 CCGCGTACGA GCGGGTGCGT GCCCGCGGCG CGCTCTCCTT CAGCCCGACG GGCAGCTGGG

 35041 TCACCGCCGA TCACGCCCTG GCGGCGAGCA TCCTCTGCTC GACGGACTTC GGGGTCTCCG 35101 GCGCCGACGG CGTCCCGGTG CCGCAGCAGG TCCTCTCGTA CGGGGAGGGC TGTCCGCTGG 35161 AGCGCGAGCA GGTGCTGCCG GCGGCCGGTG ACGTGCCGGA GGGCGGGCAG CGTGCCGTGG 35221 TCGAGGGGAT CCACCGGGAG ACGCTGGAGG GTCTCGCGCC GGACCCGTCG GCGTCGTACG 35281 CCTTCGAGCT GCTGGGCGGT TTCGTCCGCC CGGCGGTGAC GGCCGCTGCC GCCGCCGTGC 35341 TGGGTGTTCC CGCGGACCGG CGCGCGGACT TCGCGGATCT GCTGGAGCGG CTCCGGCCGC 35401 TGTCCGACAG CCTGCTGGCC CCGCAGTCCC TGCGGACGGT ACGGGCGGCG GACGGCGCGC 35461 TGGCCGAGCT CACGGCGCTG CTCGCCGATT CGGACGACTC CCCCGGGGCC CTGCTGTCGG 35521 CGCTCGGGGT CACCGCAGCC GTCCAGCTCA CCGGGAACGC GGTGCTCGCG CTCCTCGCGC 35581 ATCCCGAGCA GTGGCGGGAG CTGTGCGACC GGCCCGGGCT CGCGGCGGCC GCGGTGGAGG 35641 AGACCCTCCG CTACGACCCG CCGGTGCAGC TCGACGCCCG GGTGGTCCGC GGGGAGACGG 35701 AGCTGGCGGG CCGGCGGCTG CCGGCCGGGG CGCATGTCGT CGTCCTGACC GCCGCGACCG 35761 GCCGGGACCC GGAGGTCTTC ACGGACCCGG AGCGCTTCGA CCTCGCGCGC CCCGACGCCG 35821 CCGCGCACCT CGCGCTGCAC CCCGCCGGTC CGTACGGCCC GGTGGCGTCC CTGGTCCGGC 35881 TTCAGGCGGA GGTCGCGCTG CGGACCCTGG CCGGGCGTTT CCCCGGGCTG CGGCAGGCGG 35941 GGGACGTGCT CCGCCCCCGC CGCGCGCCTG TCGGCCGCGG GCCGCTGAGC GTCCCGGTCA 36001 GCAGCTCCTG AGACACCGGG GCCCCGGTCC GCCCGGCCCC CCTTCGGACG GACCGGACGG 36061 CTCGGACCAC GGGGACGGCT CAGACCGTCC CGTGTGTCCC CGTCCGGCTC CCGTCCGCCC 36121 CATCCCGCCC CTCCACCGGC AAGGAAGGAC ACGACGCCAT GCGCGTCCTG CTGACCTCGT 36181 TCGCACATCA CACGCACTAC TACGGCCTGG TGCCCCTGGC CTGGGCGCTG CTCGCCGCCG 36241 GGCACGAGGT GCGGGTCGCC AGCCAGCCCG CGCTCACGGA CACCATCACC GGGTCCGGGC 36301 TCGCCGCGGT GCCGGTCGGC ACCGACCACC TCATCCACGA GTACCGGGTG CGGATGGCGG 36361 GCGAGCCGCG CCCGAACCAT CCGGCGATCG CCTTCGACGA GGCCCGTCCC GAGCCGCTGG 36421 ACTGGGACCA CGCCCTCGGC ATCGAGGCGA TCCTCGCCCC GTACTTCTAT CTGCTCGCCA 36481 ACAACGACTC GATGGTCGAC GACCTCGTCG ACTTCGCCCG GTCCTGGCAG CCGGACCTGG 36541 TGCTGTGGGA GCCGACCACC TACGCGGGCG CCGTCGCCGC CCAGGTCACC GGTGCCGCGC 36601 ACGCCCGGGT CCTGTGGGGG CCCGACGTGA TGGGCAGCGC CCGCCGCAAG TTCGTCGCGC 36661 TGCGGGACCG GCAGCCGCCC GAGCACCGCG AGGACCCCAC CGCGGAGTGG CTGACGTGGA 36721 CGCTCGACCG GTACGGCGCC TCCTTCGAAG AGGAGCTGCT CACCGGCCAG TTCACGATCG 36781 ACCCGACCCC GCCGAGCCTG CGCCTCGACA CGGGCCTGCC GACCGTCGGG ATGCGTTATG 36841 TTCCGTACAA CGGCACGTCG GTCGTGCCGG ACTGGCTGAG TGAGCCGCCC GCGCGGCCCC 36901 GGGTCTGCCT GACCCTCGGC GTCTCCGCGC GTGAGGTCCT CGGCGGCGAC GGCGTCTCGC 36961 AGGGCGACAT CCTGGAGGCG CTCGCCGACC TCGACATCGA GCTCGTCGCC ACGCTCGACG 37021 CGAGTCAGCG CGCCGAGATC CGCAACTACC CGAAGCACAC CCGGTTCACG GACTTCGTGC 37081 CGATGCACGC GCTCCTGGCCG AGCTGCTCGG CGATCATCCA CCACGGCGGG GCGGGCACCT 37141 ACGCGACCGC CGTGATCAAC GCGGTGCCGC AGGTCATGCT CGCCGAGCTG TGGGACGCGC 37201 CGGTCAAGGC GCGGGCCGTC GCCGAGCAGG GGGCGGGGTT CTTCCTGCCG CCGGCCGAGC 37261 TCACGCCGCA GGCCGTGCGG GACGCCGTCG TCCGCATCCT CGACGACCCC TCGGTCGCCA
#### Abstract

-continued 37321 CCGCCGCGCA CCGGCTGCGC GAGGAGACCT TCGGCGACCC CACCCCGGCC GGGATCGTCC 37381 CCGAGCTGGA GCGGCTCGCC GCGCAGCACC GCCGCCCGCC GGCCGACGCC CGGCACTGAG 37441 CCGCACCCCT CGCCCCAGGC CTCACCCCTG TATCTGCGCC GGGGGACGCC CCCGGCCCAC 37501 CCTCCGAAAG ACCGAAAGCA GGAGCACCGT GTACGAAGTC GACCACGCCG ACGTCTACGA 37561 CCTCTTCTAC CTGGGTCGCG GCAAGGACTA CGCCGCCGAG GCCTCCGACA TCGCCGACCT 37621 GGTGCGCTCC CGTACCCCCG AGGCCTCCTC GCTCCTGGAC GTGGCCTGCG GTACGGGCAC 37681 GCATCTGGAG CACTTCACCA AGGAGTTCGG CGACACCGCC GGCCTGGAGC TGTCCGAGGA 37741 CATGCTCACC CACGCCCGCA AGCGGCTGCC CGACGCCACG CTCCACCAGG GCGACATGCG 37801 GGACTTCCGG CTCGGCCGGA AGTTCTCCGC CGTGGTCAGC ATGTTCAGCT CCGTCGGCTA 37861 CCTGAAGACG ACCGAGGAAC TCGGCGCGGC CGTCGCCTCG TTCGCGGAGC ACCTGGAGCC 37921 CGGTGGCGTC GTCGTCGTCG AGCCGTGGTG GTTCCCGGAG ACCTTCGCCG ACGGCTGGGT 37981 CAGCGCCGAC GTCGTCCGCC GTGACGGGCG CACCGTGGCC CGTGTCTCGC ACTCGGTGCG 38041 GGAGGGGAAC GCGACGCGCA TGGAGGTCCA CTTCACCGTG GCCGACCCGG GCAAGGGCGT 38101 GCGGCACTTC TCCGACGTCC ATCTCATCAC CCTGTTCCAC CAGGCCGAGT ACGAGGCCGC 38161 GTTCACGGCC GCCGGGCTGC GCGTCGAGTA CCTGGAGGGC GGCCCGTCGG GCCGTGGCCT 38221 CTTCGTCGGC GTCCCCGCCT GAGCACCGCC CAAGACCCCC CGGGGCGGGA CGTCCCGGGT 38281 GCACCAAGCA AAGAGAGAGA AACGAACCGT GACAGGTAAG ACCCGAATAC CGCGTGTCCG 38341 CCGCGGCCGC ACCACGCCCA GGGCCTTCAC CCTGGCCGTC GTCGGCACCC TGCTGGCGGG 38401 CACCACCGTG GCGGCCGCCG CTCCCGGCGC CGCCGACACG GCCAATGTTC AGTACACGAG 38461 CCGGGCGGCG GAGCTCGTCG CCCAGATGAC GCTCGACGAG AAGATC


[0044] Those of skill in the art will recognize that, due to the degenerate nature of the genetic code, a variety of DNA compounds differing in their nucleotide sequences can be used to encode a given amino acid sequence of the invention. The native DNA sequence encoding the narbonolide PKS of Streptomyces venezuelae is shown herein merely to illustrate a preferred embodiment of the invention, and the invention includes DNA compounds of any sequence that encode the amino acid sequences of the polypeptides and proteins of the invention. In similar fashion, a polypeptide can typically tolerate one or more amino acid substitutions, deletions, and insertions in its amino acid sequence without loss or significant loss of a desired activity. The present invention includes such polypeptides with alternate amino acid sequences, and the amino acid sequences shown merely illustrate preferred embodiments of the invention.
[0045] The recombinant nucleic acids, proteins, and peptides of the invention are many and diverse. To facilitate an understanding of the invention and the diverse compounds and methods provided thereby, the following description of the various regions of the narbonolide PKS and corresponding coding sequences is provided.
[0046] The loading module of the narbonolide PKS contains an inactivated KS domain, an AT domain, and an ACP domain. The AT domain of the loading module binds propionyl CoA. Sequence analysis of the DNA encoding the KS domain indicates that this domain is enzymatically inactivated, as a critical cysteine residue in the motif TVDAC-

SSSL, which is highly conserved among KS domains, is replaced by a glutamine and so is referred to as a KSQ domain. Such inactivated KS domains are also found in the PKS enzymes that synthesize the 16 -membered macrolides carbomycin, spiromycin, tylosin, and niddamycin. While the KS domain is inactive for its usual function in extender modules, it is believed to serve as a decarboxylase in the loading module.
[0047] The present invention provides recombinant DNA compounds that encode the loading module of the narbonolide PKS and useful portions thereof. These recombinant DNA compounds are useful in the construction of PKS coding sequences that encode all or a portion of the narbonolide PKS and in the construction of hybrid PKS encoding DNA compounds of the invention, as described in the section concerning hybrid PKSs below. To facilitate description of the invention, reference to a PKS, protein, module, or domain herein can also refer to DNA compounds comprising coding sequences therefor and vice versa. Also, reference to a heterologous PKS refers to a PKS or DNA compounds comprising coding sequences therefor from an organism other than Streptomyces venezuelae. In addition, reference to a PKS or its coding sequence includes reference to any portion thereof.
[0048] The present invention provides recombinant DNA compounds that encode one or more of the domains of each of the six extender modules (modules 1-6, inclusive) of the narbonolide PKS. Modules 1 and 5 of the narbonolide PKS
are functionally similar. Each of these extender modules contains a KS domain, an AT domain specific for methylmalonyl CoA, a KR domain, and an ACP domain. Module 2 of the narbonolide PKS contains a KS domain, an AT domain specific for malonyl CoA, a KR domain, a DH domain, and an ACP domain. Module 3 differs from extender modules f and 5 only in that it contains an inactive ketoreductase domain. Module 4 of the narbonolide PKS contains a KS domain, an AT domain specific for methylmalonyl CoA, a IR domain, a DH domain, an ER domain, and an ACP domain. Module 6 of the narbonolide PKS contains a KS domain, an AT domain specific for methylmalonyl CoA, and an ACP domain.
[0049] In one important embodiment, the invention provides a recombinant narbonolide PKS that can be used to express only narbonolide (as opposed to the mixture of narbonolide and 10-deoxymethynolide that would otherwise be produced) in recombinant host cells. This recombinant narbonolide PKS results from a fusion of the coding sequences of the picAIII and picAIV genes so that extender modules 5 and 6 are present on a single protein. This recombinant PKS can be constructed on the Streptomyces venezuelae or $S$. narbonensis chromosome by homologous recombination. Alternatively, the recombinant PKS can be constructed on an expression vector and introduced into a heterologous host cell. This recombinant PKS is preferred for the expression of narbonolide and its glycosylated and/or hydroxylated derivatives, because a lesser amount or no 10 -deoxymethynolide is produced from the recombinant PKS as compared to the native PKS. In a related embodiment, the invention provides a recombinant narbonolide PKS in which the picAIV gene has been rendered inactive by an insertion, deletion, or replacement. This recombinant PKS of the invention is useful in the production of 10 -deoxymethynolide and its derivatives without production of narbonolide.
[0050] In similar fashion, the invention provides recombinant narbonolide PKS in which any of the domains of the native PKS have been deleted or rendered inactive to make the corresponding narbonolide or 10 -deoxymethynolide derivative. Thus, the invention also provides recombinant narbonolide PKS genes that differ from the narbonolide PKS gene by one or more deletions. The deletions can encompass one or more modules and/or can be limited to a partial deletion within one or more modules. When a deletion encompasses an entire module, the resulting narbonolide derivative is at least two carbons shorter than the polyketide produced from the PKS encoded by the gene from which deleted PKS gene and corresponding polyketide were derived. When a deletion is within a module, the deletion typically encompasses a KR, DH, or ER domain, or both DH and ER domains, or both KR and DH domains, or all three $K R, D H$, and $E R$ domains.
[0051] This aspect of the invention is illustrated in FIG. 4, parts B and C, which shows how a vector of the invention, plasmid pKOS039-16 (not shown), was used to delete or "knock out" the picAI gene from the Streptomyces venezuelae chromosome. Plasmid pKOS039-16 comprises two segments (shown as cross-hatched boxes in FIG. 4, part B) of DNA flanking the picAI gene and isolated from cosmid pKOS023-27 (shown as a linear segment in the Figure) of the invention. When plasmid pKOS039-16 was used to transform $S$. venezuelae and a double crossover homologous
recombination event occurred, the picAI gene was deleted. The resulting host cell, designated K039-03 in the Figure, does not produce picromycin unless a functional picAI gene is introduced.
[0052] This Streptomyces venezuelae K039-03 host cell and corresponding host cells of the invention are especially useful for the production of polyketides produced from hybrid PKS or narbonolide PKS derivatives. Especially preferred for production in this host cell are narbonolide derivatives produced by PKS enzymes that differ from the narbonolide PKS only in the loading module and/or extender modules 1 and/or 2. These are especially preferred, because one need only introduce into the host cell the modified picAI gene or other corresponding gene to produce the desired PKS and corresponding polyketide. These host cells are also preferred for desosaminylating polyketides in accordance with the method of the invention in which a polyketide is provided to an $S$. venezuelae cell and desosaminylated by the endogenous desosamine biosynthesis and desosaminyl transferase gene products.
[0053] The recombinant DNA compounds of the invention that encode each of the domains of each of the modules of the narbonolide PKS are also useful in the construction of expression vectors for the heterologous expression of the narbonolide PKS and for the construction of hybrid PKS expression vectors, as described further below.
[0054] Section II: The Genes for Desosamine Biosynthesis and Transfer and for Beta-glucosidase
[0055] Narbonolide and 10-deoxymethynolide are desosaminylated in Streptomyces venezuelae and S. narbonensis to yield narbomycin and YC-17, respectively. This conversion requires the biosynthesis of desosamine and the transfer of the desosamine to the substrate polyketides by the enzyme desosaminyl transferase. Like other Streptomyces, S. venezuelae and $S$. narbonensis produce glucose and a glucosyl transferase enzyme that glucosylates desosamine at the 2 ' position. However, $S$. venezuelae and $S$. narbonensis also produce an enzyme called beta-glucosidase, which removes the glucose residue from the desosamine. The present invention provides recombinant DNA compounds and expression vectors for each of the desosamine biosynthesis enzymes, desosaminyl transferase, and beta-glucosidase.
[0056] As noted above, cosmid pKOS023-27 contains three ORFs that encode proteins involved in desosamine biosynthesis and transfer. The first ORF is from the picCII gene, also known as des VIII, a homologue of enyCII, believed to encode a 4-keto-6-deoxyglucose isomerase. The second ORF is from the picCIII gene, also known as des VII, a homologue of eryCIII, which encodes a desosaminyl transferase. The third ORF is from the picCVI gene, also known as desVI, a homologue of eryCVI, which encodes a 3-amino dimethyltransferase.
[0057] The three genes above and the remaining desosamine biosynthetic genes can be isolated from cosmid pKOS023-26, which was deposited with the American Type Culture Collection on 20 Aug. 1998 under the Budapest Treaty and is available under the accession number ATCC 203141. FIG. 3 shows a restriction site and function map of cosmid pKOS023-26. This cosmid contains a region of overlap with cosmid pKOS02327 representing nucleotides 14252 to nucleotides 38506 of pKOS 023 -27.
[0058] The remaining desosamine biosynthesis genes on cosmid pKOS023-26 include the following genes. ORF11, also known as desR, encodes beta-glucosidase and has no ery gene homologue. The picCI gene, also known as desV, is a homologue of eryCI. ORF14, also known as desIV, has no known ery gene homologue and encodes an NDP glucose 4,6-dehydratase. ORF13, also known as desIII, has no known ery gene homologue and encodes an NDP glucose synthase. The picCV gene, also known as desII, a homologue of eryCV is required for desosamine biosynthesis. The picCIV gene also known as desI, is a homologue of eryCIV, and its product is believed to be a 3,4 -dehydratase. Other ORFs on cosmid pKOS02326 include ORF12, believed to
be a regulatory gene; ORF15, which encodes an S-adenosyl methionine synthase; and ORF16, which is a homolog of the M. tuberculosis cbhK gene. Cosmid pKOS023-26 also encodes the picK gene, which encodes the cytochrome P450 hydroxylase that hydroxylates the C12 of narbomycin and the C10 and C12 positions of YC-17. This gene is described in more detail in the following section.
[0059] Below, the amino acid sequences or partial amino acid sequences of the gene products of the desosamine biosynthesis and transfer and beta-glucosidase genes are shown. These amino acid sequences are followed by the DNA sequences that encode them.


#### Abstract

Amino acid sequence of PICCI 1VSSRAETPRV PFLDLKAAYE ELRAETDAAI ARVLDSGRYL LGPELEGFEA EFAAYCETDH 61 AVGVNSGMDA LQLALRGLGI GPGDEVIVPS HTYIASWLAV SATGATPVPV EPHEDHPTLD 121 PLLVEKAITP RTRALLPVHL YGHPADMDAL RELADRHGLH IVEDAAQAHG ARYRGRRIGA 181 GSSVAAFSFY PGKNLGCFGD GGAVVTGDPE LAERLRMLRN YGSRQKYSHE TKGTNSRLDE 241 MQAAVLRIRL XHLDSWNGRR SALAAEYLSG LAGLPGIGLP VTAPDTDPVW HLFTVRTERR 301 DELRSHLDAR GIDTLTHYPV PVHLSPAYAG EAPPEGSLPR AESFARQVLS LPIGPHLERP 361 QALRVIDAVR EWAERVDQA

Amino acid sequence of 3-keto-6-deoxyglucose isomerase, PICCII 1 VADRELGTHL LETRGIHWIH AANGDPYATV LRGQADDPYP AYERVRARGA LSFSPTGSWV 61 TADHALAASI LCSTDFGVSG ADGVPVPQQV LSYGEGCPLE REQVLPAAGD VPEGGQRAVV 121 EGIHRETLEG LAPDPSASYA FELLGGFVRP AVTAAAAAVL GVPADRRADF ADLLERLRPL 181 SDSLLAPQSL RTVRAADGAL AELTALLADS DDSPGALLSA LGVTAAVQLT GNAVLALLAH 241 PEQWRELCDR PGLAAAAVEE TLRYDPPVQL DARVVRGETE LAGRRLPAGA HVVVLTAATG 301 RDPEVFTDPE RFDLARPDAA AHLALHPAGP YGPVASLVRL QAEVALRTLA GRFPGLRQAG 361 DVLRPRRAPV GRGPLSVPVS SS

Amino acid sequence of desosaminyl transferase, PICCIII 1 MRVLLTSFAH HTHYYGLVPL AWALLAAGHE VRVASQPALT DTITGSGLAA VPVGTDHLIH 61 EYRVRMAGEP RPNHPAIAFD EARPEPLDWD HALGIEAILA PYFYLLANND SMVDDLVDFA 121 RSWQPDLVLW EPITYAGAVA AQVTGAAHAR VLWGPDVMGS ARRKFVALRD RQPPEHREDP 181 TAEWLTWTLD RYGASFEEEL LTGQFTIDPT PPSLRLDTGL PTVGMRYVPY NGTSVVPDWL 241 SEPPARPRVC LTLGVSAREV LGGDGVSQGD ILEALADLDI ELVATLDASQ RAEIRNYPKH 301 TRFTDFVPMH ALLPSCSAII HHGGAGTYAT AVINAVPQVM LAELWDAPVK ARAVAEQGAG 361 FFLPPAELTP QAVRDAVVRI LDDPSVATAA HRLREETFGD PTPAGIVPEL ERLAAQHRRP 421 PADARH

Partial amino acid sequence of aminotransferase-dehydrase, PICCIV 1 VKSALSDLAF FGGPAAFDQP LLVGRPNRID RARLYERLDR ALDSQWLSNG GPLVREFEER 61 VAGLAGVRHA VATCNATAGL QLLAHAAGLT GEVIMPSMTF AATPHALRWI GLTPVFADID 121 PDTGNLDPDQ VAAAVTPRTS AVVGVHLWGR PCAADQLRKV ADEHGLRLYF DAAHALGCAV 181 DGRPAGSLGD AEVFSFHATK AVNAFEGGAV VTDDADLAAR IRALHNFGFD LPGGSPAGGT 241 NAKMSEAAAA MGLTSLDAFP EVIDRNRRNH AXYREHLADL PGVLVADHDR HGLNNHQYVI 301 VEIDEATTGI HRDLVMEVLK AEGVHTRAYF S


#### Abstract

-continued Amino acid sequence of $P I C C V$ 1 MTAPALSATA PAERCAHPGA DLGAAVHAVG QTLAAGGLVP PDEAGTTARH LVRLAVRYGN 61 SPFTPLEEAR HDLGVDRDAF RRLLALFGQV PELRTAVETG PAGAYWKNTL LPLEQRGVFD 121 AALARKPVFP YSVGLYPGPT CMFRCHFCVR VTGARYDPSA LDAGNAMFRS VIDEIPAGNP 181 SAMYFSGGLE PLTNPGLGSL AAHATDHGLR PTVYTNSFAL TERTLERQPG LWGLHAIRTS 241 LYGLNDEEYE QTTGKKAAFR RVRENLRRFQ QLRAERESPI NLGFAYIVLP GRASRLLDLV 301 DFIADLNDAG QGRTIDFVNI REDYSGRDDG KLPQEERAEL QEALNAFEER VRERTPGLHI 361DYGYALNSLR TGADAELLRI KPATMRPTAH PQVAVQVDLL GDVYLYREAG FPDLDGATRY 421 IAGRVTPDTS LTEVVRDFVE RGGEVAAVDG DEYFMDGFDQ VVTARLNQLE RDAADGWEEA 481 RGFLR

Amino acid sequence of 3 -amino dimethyl transferase, PICCVI 1 VYEVDHADVY DLFYLGRGKD YAAEASDIAD LVRSRTPEAS SLLDVACGTG THLEHFTKEF 61 GDTAGLELSE DMLTHARKRL PDATLHOGDM RDFRLGRKFS AVVSMFSSVG YLKTTEELGA 121 AVASFAEHLE PGGVVVVEPW WFPETFADGW VSADVVRRDG RTVARVSHSV REGNATRMEV 181 HFTVADPGKG VRHFSDVHLI TLFHOAEYEA AFTAAGLRVE YLEGGPSGRG LFVGVPA

Partial amino acid sequence of beta-glucosidase, ORF11 1 MTLDEKISFV HWALDPDRQN VGYLPGVPRL GIPELRAADG PNGIRLVGQT ATALPAPVAL 61 ASTFDDTMAD SYGKVMGRDG RALNQDMVLG PMMNNIRVPH GGRNYETFSE DPLVSSRTAV 121 AQIKGIQGAG LMTTAKHFAA NNQENNRFSV NANVDEQTLR EIEFPAFEAS SKAGAGSFMC 181AYNGLNGKPS CGNDELLNNV LRTQWGFQGW VMSDWLATPG TDAITKGLDQ EMGVELPGDV 241 PKGEPSPPAK FFGEALKTAV LNGTVPEAAV TRSAERIVGQ MEKFGLLLAT PAPRPERDKA 01GAQAVSRKVA ENGAVLLRNE GQALPLAGDA GKSIAVIGPT AVDPKVTGLG SAHVVPDSA 361 APLDTIKARA GAGATVTYET GEETFGTQIP AGNLSPAFNQ GHOLEPGKAG ALYDGTLTVP 421 ADGEYRIAVR ATGGYATVQL GSHTIEAGQV YGKVSSPLLK LTKGTHKLTI SGFAMSATPL 481 SLELGWVTPA AADATIAKAV ESARKARTAV VFAYDDGTEG VDRPNLSLPG TQDKLISAVA 541 DANPNTIVVL NTGSSVLMPW LSKTRAVLDM WYPGQAGAEA TAALLYGDVN PSGKLTQSFP 601 AAENQHAVAG DPTSYPGVDN QQTYREGIHV GYRWFDKENV KPLFPFGHGL SYTSFTQSAP 661 TVVRTSTGGL KVTVTVRNSG KRAGQEVVQA YLGASPNVTA POAKKKLVGY TKVSLAAGEA 721 KTVTVNVDRR QLQFWDAATD NWKTGTGNRL LQTGSSSADL RGSATVNVW

Amino acid sequence of transcriptional activator, ORF12 1 MNLVERDGEI AHLRAVLDAS AAGDGTLLLV SGPAGSGKTE LLRSLRRLAA ERETPVWSVR 61 ALPGDRDIPL GVLCQLLRSA EOHGADTSAV RDLLDAASRR AGTSPPPPTR RSASTRHTAC 121 TTGCSPSPAG TPFLVAVDDL THADTASLRF LLYCAAHHDQ GGIGFVMTER ASQRAGYRVF 181 RAELLROPHC RNMWLSGLPP SGVRQLLAHY YGPEAAERRA PAYHATTGGN PLLLRALTOD 241 RQASHTTLGA AGGDEPVHGD AFAQAVLDCL HRSAEGTLET ARWLAVLEQS DPLLVERLTG 301 TTAAAVERHI QELAAIGLLD EDGTLGQPAI REAALQDLPA GERTELHRRA AEQLHRDGAD 361 EDTVARHLLV GGAPDAPWAL PLLERGAQQA LFDDRLDDAF RILEFAVRSS TDNTQLARLA 421 PHLVAASWRM NPHMTTRALA LFDRLLSGEL PPSHPVMALI RCLVWYGRTP FAADALSRTR 481PSSDNDALEL SLTRMWLAAL CPPLLESLPA TPEPERGPVP VRLAPRTTAL QAQAGVFQRG 541 PDNASVAQAE QILQGCRLSE ETYEALETAL LVLVHADRLD RALFWSDAL工 AEAVERRSLG 601 WEAVFAATRA MIAIRCGDLP TARERAELAL SHAAPESWGL AVGMPLSALL LACTEAGEYE


-continued


#### Abstract

661 QAERVLRQPV PDAMFDSRHG MEYMHARGRY WLAXGRLHAA LGEFMLCGEI LGSWNLDQPS 721 IVPWRTSAAE VYLRLGNRQK ARALAEAQLA LVRPGRSRTR GLTLRVLAAA VDGQQAERLH 781 AEAVDMLHDS GDRLEHARAL AGMSRHQQAQ GDNYRARMTA RLAGDMAWAC GAYPLAEEIV 841 PGRGGRRAKA VSTELELPGG PDVGLLSEAE RRVAALAARG LTNRQIARRL CVTASTVEQH 901LTRVYRKLNV TRRADLPISL AQDKSVTA

Amino acid sequence of dNDP-glucose synthase (glucose-1-phosphate thymidyl transferase), ORF13 1MKGIVLAGGS GTRLHPATSV ISKQILPVYN KPMIYYPLSV LMLGGIREIQ IISTPQHIEL 61 FQSLLGNGRH LGIELDYAVQ KEPAGIADAL LVGAEHIGDD TCALILGDNI FHGPGLYTLL 121 RDSIARLDGC VLFGYPVKDP ERYGVAEVDA TGRLTDLVEK PVKPRSNLAV TGLYLYDNDV 181 VDIAKNIRPS PRGELEITDV NRVYLERGRA ELVNLGRGFA WLDTGTHDSL LRAAQYVQVL


 241 EERQGVWIAG LEEIAFRMGF IDAEACHGLG EGLSRTEYGS YLMEIAGREG APAmino acid sequence of dNDP-glucose 4,6-dehydratase, ORF14 1VRLLVTGGAG FIGSHFVRQL LAGAYPDVPA DEVIVLDSLT YAGNRANLAP VDADPRLRFV 61 HGDIRDAGLL ARELRGVDAI VHFAAESHVD RSIAGASVFT ETNVQGTQTL LQCAVDAGVG 121 RVVHVSTDEV YGSIDSGSWT ESSPLEPNSP YAASKAGSDL VARAYHRTYG LDVRITRCCN 181 NYGPYQHPEK LIPLFVTNLL DGGTLPLYGD GANVREWVHT DDHCRGIALV LAGGRAGEIY 241 HIGGGLELTN RELTGILLDS LGADWSSVRK VADRKGHDLR YSLDGGKIER ELGYRPQVSF 301 ADGLARTVRW YRENRGWWEP LKATAPQLPA TAVEVSA

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                                    Partial amino acid sequence of S-adenosylmethionine
            synthase, ORF15
    1 IGYDSSKKGF DGASCGVSVS IGSQSPDIAQ GVDTAYEKRV EGASQRDEGD ELDKQGAGDQ
    61GLMFGYASDE TPELMPLPIH LAHRLSRRLT EVRKNGTIPY LRPDGKTQVT IEYDGDRAVR
121LDTVVVSSQH ASDIDLESLL APDVRKFVVE HVLAQLVEDG IKLDTDGYRL LVNPTGRFEI
181GGPMGDAGLT GRKIIIDTYG GMARHGGGAF SGKDPSKVDR SAAYAMRWVA KNVVAAGLAS
241RCEVQVAYAI GKAEPVGLFV ETFGTHKIET EKIENAIGEV FDLRPAAIIR DLDLLRPIYS
301QTAAYGHFGR ELPDFTWERT DRVDALKKAA GL
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                Partial amino acid sequence of ORF16
    (homologous to $M$. tuberculosis cbhK)
1 MRIAVTGSIA TDHLMTFPGR FAEOILPDQL AHVSLSFLVD TLDIRHGGVA ANIAYGLGLL
61 GRRPVLVGAV GKDFDGYGQL LRAAGVDTDS VRVSDRQHTA RFMCTTDEDG NQLASFYAGA
121 MAEARDIDLG ETAGRPGGID LVLVGADDPE AMVRHTRVCR ELGLRPAADP SQQLARLEGD
181 SVRELVDGAE LLFTNAYERA LLLSKTGWTE QEVLARVGTW ITTLGAKGCR
[0060] While not all of the insert DNA of cosmid pKOS02326 has been sequenced, five large contigs shown of FIG. 3 have been assembled and provide sufficient sequence information to manipulate the genes therein in accordance with the methods of the invention. The sequences of each of these five contigs are shown below.
[0061] Contig 001 from cosmid pKOS023-26 contains 2401 nucleotides, the first 100 bases of which correspond to 100 bases of the insert sequence of cosmid pKOS023-27. Nucleotides 80-2389 constitute ORF11, which encodes 1 beta glucosidase.

[^0]
## -continued


#### Abstract

181 GCTGCGTGCC GCCGACGGCC CGAACGGCAT CCGCCTGGTG GGGCAGACCG CCACCGCGCT 241 GCCCGCGCCG GTCGCCCTGG CCAGCACCTT CGACGACACC ATGGCCGACA GCTACGGCAA 301 GGTCATGGGC CGCGACGGTC GCGCGCTCAA CCAGGACATG GTCCTGGGCC CGATGATGAA 361 CAACATCCGG GTGCCGCACG GCGGCCGGAA CTACGAGACC TTCAGCGAGG ACCCCCTGGT 421 CTCCTCGCGC ACCGCGGTCG CCCAGATCAA GGGCATCCAG GGTGCGGGTC TGATGACCAC 481 GGCCAAGCAC TTCGCGGCCA ACAACCAGGA GAACAACCGC TTCTCCGTGA ACGCCAATGT 541 CGACGAGCAG ACGCTCCGCG AGATCGAGTT CCCGGCGTTC GAGGCGTCCT CCAAGGCCGG 601 CGCGGGCTCC TTCATGTGTG CCTACAACGG CCTCAACGGG AAGCCGTCCT GCGGCAACGA 661 CGAGCTCCTC AACAACGTGC TGCGCACGCA GTGGGGCTTC CAGGGCTGGG TGATGTCCGA 721 CTGGCTCGCC ACCCCGGGCA CCGACGCCAT CACCAAGGGC CTCGACCAGG AGATGGGCGT 781 CGAGCTCCCC GGCGACGTCC CGAAGGGCGA GCCCTCGCCG CCGGCCAAGT TCTTCGGCGA 841 GGCGCTGAAG ACGGCCGTCC TGAACGGCAC GGTCCCCGAG GCGGCCGTGA CGCGGTCGGC 901 GGAGCGGATC GTCGGCCAGA TGGAGAAGTT CGGTCTGCTC CTCGCCACTC CGGCCCCGCG 961 GCCCGAGCGC GACAAGGCGG GTGCCCAGGC GGTGTCCCGC AAGGTCGCCG AGAACGGCGC 1021 GGTGCTCCTG CGCAACGAGG GCCAGGCCCT GCCGCTCGCC GGTGACGCCG GCAAGAGCAT 1081 CGCGGTCATC GGCCCGACGG CCGTCGACCC CAAGGTCACC GGCCTGGGCA GCGCCCACGT 1141 CGTCCCGGAC TCGGCGGCGG CGCCACTCGA CACCATCAAG GCCCGCGCGG GTGCGGGTGC 1201 GACGGTGACG TACGAGACGG GTGAGGAGAC CTTCGGGACG CAGATCCCGG CGGGGAACCT 1261 CAGCCCGGCG TTCAACCAGG GCCACCAGCT CGAGCCGGGC AAGGCGGGGG CGCTGTACGA 1321 CGGCACGCTG ACCGTGCCCG CCGACGGCGA GTACCGCATC GCGGTCCGTG CCACCGGTGG 1381 TTACGCCACG GTGCAGCTCG GCAGCCACAC CATCGAGGCC GGTCAGGTCT ACGGCAAGGT 1441 GAGCAGCCCG CTCCTCAAGC TGACCAAGGG CACGCACAAG CTCACGATCT CGGGCTTCGC 1501 GATGAGTGCC ACCCCGCTCT CCCTGGAGCT GGGCTGGGTN ACGCCGGCGG CGGCCGACGC 1561 GACGATCGCG AAGGCCGTGG AGTCGGCGCG GAAGGCCCGT ACGGCGGTCG TCTTCGCCTA 1621 CGACGACGGC ACCGAGGGCG TCGACCGTCC GAACCTGTCG CTGCCGGGTA CGCAGGACAA 1681 GCTGATCTCG GCTGTCGCGG ACGCCAACCC GAACACGATC GTGGTCCTCA ACACCGGTTC 1741 GTCGGTGCTG ATGCCGTGGC TGTCCAAGAC CCGCGCGGTC CTGGACATGT GGTACCCGGG 1801 CCAGGCGGGC GCCGAGGCCA CCGCCGCGCT GCTCTACGGT GACGTCAACC CGAGCGGCAA 1861 GCTCACGCAG AGCTTCCCGG CCGCCGAGAA CCAGCACGCG GTCGCCGGCG ACCCGACCAG 1921 CTACCCGGGC GTCGACAACC AGCAGACGTA CCGCGAGGGC ATCCACGTCG GGTACCGCTG 1981 GTTCGACAAG GAGAACGTCA AGCCGCTGTT CCCGTTCGGG CACGGCCTGT CGTACACCTC 2041 GTTCACGCAG AGCGCCCCGA CCGTCGTGCG TACGTCCACG GGTGGTCTGA AGGTCACGGT 2101 CACGGTCCGC AACAGCGGGA AGCGCGCCGG CCAGGAGGTC GTCCAGGCGT ACCTCGGTGC 2161 CAGCCCGAAC GTGACGGCTC CGCAGGCGAA GAAGAAGCTC GTGGGCTACA CGAAGGTCTC 2221 GCTCGCCGCG GGCGAGGCGA AGACGGTGAC GGTGAACGTC GACCGCCGTC AGCTGCAGTT 2281 CTGGGATGCC GCCACGGACA ACTGGAAGAC GGGAACGGGC AACCGCCTCC TGCAGACCGG 2341 TTCGTCCTCC GCCGACCTGC GGGGCAGCGC CACGGTCAAC GTCTGGTGAC GTGACGCCGT 2401 G


[0062] Contig 002 from cosmid pKOS023-26 contains 5970 nucleotides and the following ORFs: from nucleotide 995 to 1 is an ORF of picCIV that encodes a partial sequence of an amino transferase-dehydrase; from nucleotides 1356 to

2606 is an ORF of picK that encodes a cytochrome P450 hydroxylase; and from nucleotides 2739 to 5525 is ORF12, which encodes a transcriptional activator.


#### Abstract

1 GGCGAGAAGT AGGCGCGGGT GTGCACGCCT TCGGCCTTCA GGACCTCCAT GACGAGGTCG 61 CGGTGGATGC CGGTGGTGGC CTCGTCGATC TCGACGATCA CGTACTGGTG GTTGTTGAGG 121 CCGTGGCGGT CGTGGTCGGC GACGAGGACG CCGGGGAGGT CCGCGAGGTG CTCGCGGTAG 181 sCGGCGTGGT TGCGCCGGTT CCGGTCGATG ACCTCGGGAA ACGCGTCGAG GGAGGTGAGG 241 CCCATGGCGG CGGCGGCCTC GCTCATCTTG GCGTTGGTCC CGCCGGCGGG GCTGCCGCCG 301 GGCAGGTCGA AGCCGAAGTT GTGGAGGGCG CGGATCCGGG CGGCGAGGTC GGCGTCGTCG 361 GTGACGACGG CGCCGCCCTC GAAGGCGTTG ACGGCCTTGG TGGCGTGGAA GCTGAAGACC 421 TCGGCGTCGC CGAGGCTGCC GGCGGGCCGG CCGTCGACCG CGCAGCCGAG GGCGTGCGCG 481 GCGTCGAAGT ACAGCCGCAG GCCGTGCTCG TCGGCGACCT TCCGCAGCTG GTCGGCGGCG 541 CAGGGGCGGC CCCAGAGGTG GACGCCGACG ACGGCCGAGG TGCGGGGTGT GACCGCGGCG 601 GCCACCTGGT CCGGGTCGAG GTTGCCGGTG TCCGGGTCGA TGTCGGCGAA GACCGGGGTG 661 AGGCCGATCC AGCGCAGTGC GTGCGGGGTG GCGGCGAACG TCATCGACGG CATGATCACT 721 TCGCCGGTGA GGCCGGCGGC GTGCGCGAGG AGCTGGAGCC CGGCCGTGGC GTTGCAGGTG 781 GCCACGGCAT GCCGGACCCC GGCGAGCCCG GCGACGCGCT CCTCGAACTC GCGGACGAGC 841 GGGCCGCCGT TGGACAGCCA CTGGCTGTCG AGGGCCCCGT CGAGCCGCTC GTACAGCCTG 901 GCGCGGTCGA TGCGGTTGGG CCGCCCCACG AGGAGCGGCT GGTCGAAAGC GGCGGGGCCG 961 CCGAAGAATG CGAGGTCGGA TAAGGCGCTT TTCACGGATG TTCCCTCCGG GCCACCGTCA 1021 CGAAATGATT CGCCGATCCG GGAATCCCGA ACGAGGTCGC CGCGCTCCAC CGTGACGTAC 1081 GACGAGATGG TCGATTGTGG TGGTCGATTT CGGGGGGACT CTAATCCGCG CGGAACGGGA 1141 CCGACAAGAG CACGCTATGC GCTCTCGATG TGCTTCGGAT CACATCCGCC TCCGGGGTAT 1201 TCCATCGGCG GCCCGAATGT GATGATCCTT GACAGGATCC GGGAATCAGC CGAGCCGCCG 1261 GGAGGGCCGG GGCGCGCTCC GCGGAAGAGT ACGTGTGAGA AGTCCCGTTC CTCTTCCCGT 1321 TTCCGTTCCG CTTCCGGCCC GGTCTGGAGT TCTCCGTGCG CCGTACCCAG CAGGGAACGA 1381 CCGCTCCTCC CCCGGTACTC GACCTCGGGG CCCTGGGGCA GGATTTCGCG GCCGATCCGT 1441 ATCCGACGTA CGCGAGACTG CGTGCCGAGG GTCCGGCCCA CCGGGTGCGC ACCCCCGAGG 1501 GGGACGAGGT GTGGCTGGTC GTCGGCTACG ACCGGGCGCG GGCGGTCCTC GCCGATCCCC 1561 GGTTCAGCAA GGACTGGCGC AACTCCACGA CTCCCCTGAC CGAGGCCGAG GCCGCGCTCA 1621 ACCACAACAT GCTGGAGTCC GACCCGCCGC GGCACACCCG GCTGCGCAAG CTGGTGGCCC 1681 GTGAGTTCAC CATGCGCCGG GTCGAGTTGC TGCGGCCCCG GGTCCAGGAG ATCGTCGACG 1741 GGCTCGTGGA CGCCATGCTG GCGGCGCCCG ACGGCCGCGC CGATCTGATG GAGTCCCTGG 1801 CCTGGCCGCT GCCGATCACC GTGATCTCCG AACTCCTCGG CGTGCCCGAG CCGGACCGCG 1861 CCGCCTTCCG CGTCTGGACC GACGCCTTCG TCTTCCCGGA CGATCCCGCC CAGGCCCAGA 1921 CCGCCATGGC CGAGATGAGC GGCTATCTCT CCCGGCTCAT CGACTCCAAG CGCGGGCAGG 1981 ACGGCGAGGA CCTGCTCAGC GCGCTCGTGC GGACCAGCGA CGAGGACGGC TCCCGGCTGA 2041 CCTCCGAGGA GCTGCTCGGT ATGGCCCACA TCCTGCTCGT CGCGGGGCAC GAGACCACGG 2101 TCAATCTGAT CGCCAACGGC ATGTACGCGC TGCTCTCGCA CCCCGACCAG CTGGCCGCCC


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2161 TGCGGGCCGA CATGACGCTC TTGGACGGCG CGGTGGAGGA GATGTTGCGC TACGAGGGCC 221 CGGTGGAATC CGCGACCTAC CGCTTCCCGG TCGAGCCCGT CGACCTGGAC GGCACGGTCA 2281 TCCCGGCCGG TGACACGGTC CTCGTCGTCC TGGCCGACGC CCACCGCACC CCCGAGCGCT 2341 TCCCGGACCC GCACCGCTTC GACATCCGCC GGGACACCGC CGGCCATCTC GCCTTCGGCC 2401 ACGGCATCCA CTTCTGCATC GGCGCCCCCT TGGCCCGGTT GGAGGCCCGG ATCGCCGTCC 2461 GCGCCCTTCT CGAACGCTGC CCGGACCTCG CCCTGGACGT CTCCCCCGGC GAACTCGTGT 2521 GGTATCCGAA CCCGATGATC CGCGGGCTCA AGGCCCTGCC GATCCGGTGG CGGCGAGGAC 2581 GGGAGGCGGG CCGCCGTACC GGTTGAACCC GCACGTCACC CATTACGACT CCTTGTCACG 2641 GAAGCCCCGG ATCGGTCCCC CCTCGCCGTA ACAAGACCTG GTTAGAGTGA TGGAGGACGA 2701 CGAAGGGTTC GGCGCCCGGA CGAGGGGGGA CTTCCGCGAT GAATCTGGTG GAACGCGACG 2761 GGGAGATAGC CCATCTCAGG GCCGTTCTTG ACGCATCCGC CGCAGGTGAC GGGACGCTCT 2821 TACTCGTCTC CGGACCGGCC GGCAGCGGGA AGACGGAGCT GCTGCGGTCG CTCCGCCGGC 2881 TGGCCGCCGA GCGGGAGACC CCCGTCTGGT CGGTCCGGGC GCTGCCGGGT GACCGCGACA 2941 TCCCCCTGGG CGTCCTCTGC CAGTTACTCC GCAGCGCCGA ACAACACGGT GCCGACACCT 3001 CCGCCGTCCG CGACCTGCTG GACGCCGCCT CGCGGCGGGC CGGAACCTCA CCTCCCCCGC 3061 CGACGCGCCG CTCCGCGTCG ACGAGACACA CCGCCTGCAC GACTGGCTGC TCTCCGTCTC 3121 CCGCCGGCAC CCCGTTCCTC GTCGCCGTCG ACGACCTGAC CCACGCCGAC ACCGCGTCCC 3181 TGAGGTTCCT CCTGTACTGC GCCGCCCACC ACGACCAGGG CGGCATCGGC TTCGTCATGA 3241 CCGAGCGGGC CTCGCAGCGC GCCGGATACC GGGTGTTCCG CGCCGAGCTC CTCCGCCAGC 3301 CGCACTGCCG CAACATGTGG CTCTCCGGGC TTCCCCCCAG CGGGGTACGC CAGTTACTCG 3361 CCCACTACTA CGGCCCCGAG GCCGCCGAGC GGCGGGCCCC CGCGTACCAC GCGACGACCG 3421 GCGGGAACCC GCTGCTCCTG CGGGCGCTGA CCCAGGACCG GCAGGCCTCC CACACCACCC 3481 TCGGCGCGGC CGGCGGCGAC GAGCCCGTCC ACGGCGACGC CTTCGCCCAG GCCGTCCTCG 3541 ACTGCCTGCA CCGCAGCGCC GAGGGCACAC TGGAGACCGC CCGCTGGCTC GCGGTCCTCG 3601 AACAGTCCGA CCCGCTCCTG GTGGAGCGGC TCACGGGAAC GACCGCCGCC GCCGTCGAGC 3661 GCCACATCCA GGAGCTCGCC GCCATCGGCC TCCTGGACGA GGACGGCACC CTGGGACAGC 3721 CCGCGATCCG CGAGGCCGCC CTCCAGGACC TGCCGGCCGG CGAGCGCACC GAACTGCACC 3781 GGCGCGCCGC GGAGCAGCTG CACCGGGACG GCGCCGACGA GGACACCGTG GCCCGCCACC 3841 TGCTGGTCGG CGGCGCCCCC GACGCTCCCT GGGCGCTGCC CCTGCTCGAA CGGGGCGCGC 3901 AGCAGGCCCT GTTCGACGAC CGACTCGACG ACGCCTTCCG GATCCTCGAG TTCGCCGTGC 3961 GGTCGAGCAC CGACAACACC CAGCTGGCCC GCCTCGCCCC ACACCTGGTC GCGGCCTCCT 4021 GGCGGATGAA CCCGCACATG ACGACCCGGG CCCTCGCACT CTTCGACCGG CTCCTGAGCG 4081 GTGAACTGCC GCCCAGCCAC CCGGTCATGG CCCTGATCCG CTGCCTCGTC TGGTACGGNC 4141 GGCTGCCCGA GGCCGCCGAC GCGCTGTCCC GGCTGCGGCC CAGCTCCGAC AACGATGCCT 4201 TGGAGCTGTC GCTCACCCGG ATGTGGCTCG CGGCGCTGTG CCCGCCGCTC CTGGAGTCCC 321 TGCCGGCCAC GCCGGAGCCG GAGCGGGGTC CCGTCCCCGT ACGGCTCGCG CCGCGGACGA 4321 CCGCGCTCCA GGCCCAGGCC GGCGTCTTCC AGCGGGGCCC GGACAACGCC TCGGTCGCGC 4381 AGGCCGAACA GATCCTGCAG GGCTGCCGGC TGTCGGAGGA GACGTACGAG GCCCTGGAGA


#### Abstract

4441 CGGCCCTCTT GCTCCTCGTC CACGCCGACC GGCTCGACCG GGCGCTGTTC TGGTCGGACG 4501 CCCTGCTCGC CGAGGCCGTG GAGCGGCGGT CGCTCGGCTG GGAGGCGGTC TTCGCCGCGA 4561 CCCGGGCGAT GATCGCGATC CGCTGCGGCG ACCTCCCGAC GGCGCGGGAG CGGGCCGAGC 4621 TGGCGCTCTC CCACGCGGCG CCGGAGAGCT GGGGCCTCGC CGTGGGCATG CCCCTCTCCG 4681 CGCTGCTGCT CGCCTGCACG GAGGCCGGCG AGTACGAACA GGCGGAGCGG GTCCTGCGGC 4741 AGCCGGTGCC GGACGCGATG TTCGACTCGC GGCACGGCAT GGAGTACATG CACGCCCGGG 4801 GCCGCTACTG GCTGGCGANC GGCCGGCTGC ACGCGGCGCT GGGCGAGTTC ATGCTCTGCG 4861 GGGAGATCCT GGGCAGCTGG AACCTCGACC AGCCCTCGAT CGTGCCCTGG CGGACCTCCG 4921 CCGCCGAGGT GTACCTGCGG CTCGGCAACC GCCAGAAGGC CAGGGCGCTG GCCGAGGCCC 4981 AGCTCGCCCT GGTGCGGCCC GGGCGCTCCC GCACCCGGGG TCTCACCCTG CGGGTCCTGG 5041 CGGCGGCGGT GGACGGCCAG CAGGCGGAGC GGCTGCACGC CGAGGCGGTC GACATGCTGC 5101 ACGACAGCGG CGACCGGCTC GAACACGCCC GCGCGCTCGC CGGGATGAGC CGCCACCAGC 5161 AGGCCCAGGG GGACAACTAC CGGGCGAGGA TGACGGCGCG GCTCGCCGGC GACATGGCGT 5221 GGGCCTGCGG CGCGTACCCG CTGGCCGAGG AGATCGTGCC GGGCCGCGGC GGCCGCCGGG 5281 CGAAGGCGGT GAGCACGGAG CTGGAACTGC CGGGCGGCCC GGACGTCGGC CTGCTCTCGG 5341 AGGCCGAACG CCGGGTGGCG GCCCTGGCAG CCCGAGGATT GACGAACCGC CAGATAGCGC 5401 GCCGGCTCTG CGTCACCGCG AGCACGGTCG AACAGCACCT GACGCGCGTC TACCGCAAAC 5461 TGAACGTGAC CCGCCGAGCA GACCTCCCGA TCAGCCTCGC CCAGGACAAG TCCGTCACGG 5521 CCTGAGCCAC CCCCGGTGTC CCCGTGCGAC GACCCGCCGC ACGGGCCACC GGGCCCGCCG 5581 GGACACGCCG GTGCGACACG GGGGCGCGCC AGGTGCCATG GGGACCTCCG TGACCGCCCG 5641 AGGCGCCCGA GGCGCCCGGT GCGGCACCCG GAGACGCCAG GACCGCCGGG ACCACCGGAG 5701 ACGCCAGGGA CCGCTGGGGA CACCGGGACC TCAGGGACCG CCGGGACCGC CCGAGTTGCA 5761 CCCGGTGCGC CCGGGGACAC CAGACCGCCG GGACCACCCG AGGGTGCCCG GTGTGGCCCC 5821 GGCGGCCGGG GTGTCCTTCA TCGGTGGGCC TTCATCGGCA GGAGGAAGCG ACCGTGAGAC 5881 CCGTCGTGCC GTCGGCGATC AGCCGCCTGT ACGGGCGTCG GACTCCCTGG CGGTCCCGGA 5941 CCCGTCGTAC GGGCTCGCGG GACCCGGTGC


[0063] Contig 003 from cosmid pKOS023-26 contains 3292 nucleotides and the following ORFs: from nucleotide 104 to 982 is ORF13, which encodes dNDP glucose synthase (glucose-1-phosphate thymidyl transferase); from
nucleotide 1114 to 2127 is ORF14, which encodes dNDPglucose 4,6-dehydratase; and from nucleotide 2124 to 3263 is the picCI ORF

1 ACCCCCCAAA GGGGTGGTGA CACTCCCCCT GCGCAGCCCC TAGCGCCCCC CTAACTCGCC
61 ACGCCGACCG TTATCACCGG CGCCCTGCTG CTAGTTTCCG ACAATGAAGG GAATAGTCCT 121 GGCCGGCGGG AGCGGAACTC GGCTGCATCC GGCGACCTCG GTCATTTCGA AGCAGATTCT 181 TCCGGTCTAC AACAAACCGA TGATCTACTA TCCGCTGTCG GTTCTCATGC TCGGCGGTAT 241 TCGCGAGATT CAAATCATCT CGACCCCCCA GCACATCGAA CTCTTCCAGT CGCTTCTCGG 301 AAACGGCAGG CACCTGGGAA TAGAACTCGA CTATGCGGTC CAGAAAGAGC CCGCAGGAAT 361 CGCGGACGCA CTTCTCGTCG GAGCCGAGCA CATCGGCGAC GACACCTGCG CCCTGATCCT 421 GGGCGACAAC ATCTTCCACG GGCCCGGCCT CTACACGCTC CTGCGGGACA GCATCGCGCG

## 481 CCTCGACGGC TGCGTGCTCT TCGGCTACCC GGTCAAGGAC CCCGAGCGGT ACGGCGTCGC

 541 CGAGGTGGAC GCGACGGGCC GGCTGACCGA CCTCGTCGAG AAGCCCGTCA AGCCGCGCTC 601 CAACCTCGCC GTCACCGGCC TCTACCTCTA CGACAACGAC GTCGTCGACA TCGCCAAGAA 661 CATCCGGCCC TCGCCGCGCG GCGAGCTGGA GATCACCGAC GTCAACCGCG TCTACCTGGA 721 GCGGGGCCGG GCCGAACTCG TCAACCTGGG CCGCGGCTTC GCCTGGCTGG ACACCGGCAC 781 CCACGACTCG CTCCTGCGGG CCGCCCAGTA CGTCCAGGTC CTGGAGGAGC GGCAGGGCGT 841 CTGGATCGCG GGCCTTGAGG AGATCGCCTT CCGCATGGGC TTCATCGACG CCGAGGCCTG 901 TCACGGCCTG GGAGAAGGCC TCTCCCGCAC CGAGTACGGC AGCTATCTGA TGGAGATCGC 961 CGGCCGCGAG GGAGCCCCGT GAGGGCACCT CGCGGCCGAC GCGTTCCCAC GACCGACAGC 1021 GCCACCGACA GTGCGACCCA CACCGCGACC CGCACCGCCA CCGACAGTGC GACCCACACC 1081 GCGACCTACA GCGCGACCGA AAGGAAGACG GCAGTGCGGC TTCTGGTGAC CGGAGGTGCG 1141 GGCTTCATCG GCTCGCACTT CGTGCGGCAG CTCCTCGCCG GGGCGTACCC CGACGTGCCC 1201 GCCGATGAGG TGATCGTCCT GGACAGCCTC ACCTACGCGG GCAACCGCGC CAACCTCGCC 1261 CCGGTGGACG CGGACCCGCG ACTGCGCTTC GTCCACGGCG ACATCCGCGA CGCCGGCCTC 1321 CTCGCCCGGG AACTGCGCGG CGTGGACGCC ATCGTCCACT TCGCGGCCGA GAGCCACGTG 1381 GACCGCTCCA TCGCGGGCGC GTCCGTGTTC ACCGAGACCA ACGTGCAGGG CACGCAGACG 1441 CTGCTCCAGT GCGCCGTCGA CGCCGGCGTC GGCCGGGTCG TGCACGTCTC CACCGACGAG 1501 GTGTACGGGT CGATCGACTC CGGCTCCTGG ACCGAGAGCA GCCCGCTGGA GCCCAACTCG 1561 CCCTACGCGG CGTCCAAGGC CGGCTCCGAC CTCGTTGCCC GCGCCTACCA CCGGACGTAC 1621 GGCCTCGACG TACGGATCAC CCGCTGCTGC AACAACTACG GGCCGTACCA GCACCCCGAG 1681 AAGCTCATCC CCCTCTTCGT GACGAACCTC CTCGACGGCG GGACGCTCCC GCTGTACGGC 1741 GACGGCGCGA ACGTCCGCGA GTGGGTGCAC ACCGACGACC ACTGCCGGGG CATCGCGCTC 1801 GTCCTCGCGG GCGGCCGGGC CGGCGAGATC TACCACATCG GCGGCGGCCT GGAGCTGACC 1861 AACCGCGAAC TCACCGGCAT CCTCCTGGAC TCGCTCGGCG CCGACTGGTC CTCGGTCCGG 1921 AAGGTCGCCG ACCGCAAGGG CCACGACCTG CGCTACTCCC TCGACGGCGG CAAGATCGAG 1981 CGCGAGCTCG GCTACCGCCC GCAGGTCTCC TTCGCGGACG GCCTCGCGCG GACCGTCCGC 2041 TGGTACCGGG AGAACCGCGG CTGGTGGGAG CCGCTCAAGG CGACCGCCCC GCAGCTGCCC 2101 GCCACCGCCG TGGAGGTGTC CGCGTGAGCA GCCGCGCCGA GACCCCCCGC GTCCCCTTCC 2161 TCGACCTCAA GGCCGCCTAC GACGAGCTCC GCGCGGAGAC CGACGCCGCG ATCGCCCGCG 2221 TCCTCGACTC GGGGCGCTAC CTCCTCGGAC CCGAACTCGA AGGATTCGAG GCGGAGTTCG 2281 CCGCGTACTG CGAGACGGAC CACGCCGTCG GCGTGAACAG CGGGATGGAC GCCCTCCAGC 2341 TCGCCCTCCG CGGCCTCGGC ATCGGACCCG GGGACGAGGT GATCGTCCCC TCGCACACGT 2401 ACATCGCCAG CTGGCTCGCG GTGTCCGCCA CCGGCGCGAC CCCCGTGCCC GTCGAGCCGC 2461 ACGAGGACCA CCCCACCCTG GACCCGCTGC TCGTCGAGAA GGCGATCACC CCCCGCACCC 2521 GGGCGCTCCT CCCCGTCCAC CTCTACGGGC ACCCCGCCGA CATGGACGCC CTCCGCGAGC 2581 TCGCGGACCG GCACGGCCTG CACATCGTCG AGGACGCCGC GCAGGCCCAC GGCGCCCGCT 2641 ACCGGGGCCG GCGGATCGGC GCCGGGTCGT CGGTGGCCGC GTTCAGCTTC TACCCGGGCA 2701 AGAACCTCGG CTGCTTCGGC GACGGCGGCG CCGTCGTCAC CGGCGACCCC GAGCTCGCCG 2761 AACGGCTCCG GATGCTCCGC AACTACGGCT CGCGGCAGAA GTACAGCCAC GAGACGAAGG-continued
2821 GCACCAACTC CCGCCTGGAC GAGATGCAGG CCGCCGTGCT GCGGATCCGG CTCGNCCACC 2881 TGGACAGCTG GAACGGCCGC AGGTCGGCGC TGGCCGCGGA GTACCTCTCC GGGCTCGCCG 2941 GACTGCCCGG CATCGGCCTG CCGGTGACCG CGCCCGACAC CGACCCGGTC TGGCACCTCT 3001 TCACCGTGCG CACCGAGCGC CGCGACGAGC TGCGCAGCCA CCTCGACGCC CGCGGCATCG 3061 ACACCCTCAC GCACTACCCG GTACCCGTGC ACCTCTCGCC CGCCTACGCG GGCGAGGCAC 3121 CGCCGGAAGG CTCGCTCCCG CGGGCCGAGA GCTTCGCGCG GCAGGTCCTC AGCCTGCCGA 3181 TCGGCCCGCA CCTGGAGCGC CCGCAGGCGC TGCGGGTGAT CGACGCCGTG CGCGAATGGG 3241 CCGAGCGGGT CGACCAGGCC TAGTCAGGTG GTCCGGTAGA CCCAGCAGGC CG
[0064] Contig 004 from cosmid pKOS023-26 contains 1693 nucleotides and the following ORFs: from nucleotide 1692 to 694 is ORF15, which encodes a part of S-adenos-
ylmethionine synthetase; and from nucleotide 692 to 1 is ORF16, which encodes a part of a protein homologous to the M. tuberculosis cbhK gene.

> 1 ATGCGGCACC CCTTGGCGCC GAGCGTGGTG ATCCAGGTGC CGACCCGGGC GAGCACCTCC
-continued
1561 GGAGGCACCC TCGACCCGCT TCTCGTACGC GGTGTCGACA CCCTGGGCGA TGTCCGGGGA 1621 CTGCGACCCG ATGGACACCG ACACGCCGCA GGAGGCGCCG TCGAAGCCCT TCTTCGAGGA 1681 GTCGTACCCG ATC
[0065] Contig 005 from cosmid pKOS023-26 contains 1565 nucleotides and contains the ORF of the picCV gene that encodes PICCV, involved in desosamine biosynthesis.


#### Abstract

1 CCCCGCTCGC GGCCCCCCAG ACATCCACGC CCACGATTGG ACGCTCCCGA TGACCGCCCC 61 CGCCCTCTCC GCCACCGCCC CGGCCGAACG CTGCGCGCAC CCCGGAGCCG ATCTGGGGGC 121 GGCGGTCCAC GCCGTCGGCC AGACCCTCGC CGCCGGCGGC CTCGTGCCGC CCGACGAGGC 181 CGGAACGACC GCCCGCCACC TCGTCCGGCT CGCCGTGCGC TACGGCAACA GCCCCTTCAC 241 CCCGCTGGAG GAGGCCCGCC ACGACCTGGG CGTCGACCGG GACGCCTTCC GGCGCCTCCT 301 CGCCCTGTTC GGGCAGGTCC CCGAGCTCCG CACCGCGGTC GAGACCGGCC CCGCCGGGGC 361 GTACTGGAAC AACACCCTGC TCCCGCTCGA ACAGCGCGGC GTCTTCGACG CGGCGCTCGC 421 CAGGAAGCCC GTCTTCCCGT ACAGCGTCGG CCTCTACCCC GGCCCGACCT GCATGTTCCG 481 CTGCCACTTC TGCGTCCGTG TGACCGGCGC CCGCTACGAC CCGTCCGCCC TCGACGCCGG 541 CAACGCCATG TTCCGGTCGG TCATCCACGA GATACCCGCG GGCAACCCCT CGGCGATGTA 601 CTTCTCCGGC GGCCTGGAGC CGCTCACCAA CCCCGGCCTC GGGAGCCTGG CCGCGCACGC 661 CACCGACCAC GGCCTGCGGC CCACCGTCTA CACGAACTCC TTCGCGCTCA CCGAGCGCAC 721 CCTGGAGCGC CAGCCCGGCC TCTGGGGCCT GCACGCCATC CCCACCTCGC TCTACGGCCT 781 CAACGACGAG GAGTACGAGC AGACCACCGG CAAGAAGGCC GCCTTCCGCC GCGTCCGCGA 841 GAACCTGCGC CGCTTCCAGC AGCTGCGCGC CGAGCGCGAG TCGCCGATCA ACCTCGGCTT 901 CGCCTACATC GTGCTCCCGG GCCGTGCCTC CCGCCTGCTC GACCTGGTCG ACTTCATCGC 961 CGACCTCAAC GACGCCGGGC AGGGCAGGAC GATCGACTTC GTCAACATTC GCGAGGACTA 1021 CAGCGGCCGT GACGACGGCA AGCTGCCGCA GGAGGAGCGG GCCGAGCTCC AGGAGGCCCT 1081 CAACGCCTTC GAGGAGCGGG TCCGCGAGCG CACCCCCGGA CTCCACATCG ACTACGGCTA 1141 CGCCCTGAAC AGCCTGCGCA CCGGGGCCGA CGCCGAACTG CTGCCGATCA AGCCCGCCAC 1201 CATGCGGCCC ACCGCGCACC CGCAGGTCGC GGTGCAGGTC GATCTCCTCG GCGACGTGTA 1261 CCTGTACCGC GAGGCCGGCT TCCCCGACCT GGACGGCGCG ACCCGCTACA TCGCGGGCCG 1321 CGTGACCCCC GACACCTCCC TCACCGAGGT CGTCAGGGAC TTCGTCGAGC GCGGCGGCGA 1381 GGTGGCGGCC GTCGACGGCG ACGAGTACTT CATGGACGGC TTCGATCAGG TCGTCACCGC 1441 CCGCCTGAAC CAGCTGGAGC GCGACGCCGC GGACGGCTGG GAGGAGGCCC GCGGCTTCCT 1501 GCGCTGACCC GCACCCGCCC CGATCCCCC GATCCCCCCC CCACGATCCC CCCACCTGAG 1561 GGCCC


[0066] The recombinant desosamine biosynthesis and transfer and beta-glucosidase genes and proteins provided by the invention are useful in the production of glycosylated polyketides in a variety of host cells, as described in Section IV below.
[0067] Section III. The picK Hydroxylase Gene
[0068] The present invention provides the picK gene in recombinant form as well as recombinant PicK protein. The availability of the hydroxylase encoded by the picK gene in
recombinant form is of significant benefit in that the enzyme can convert narbomycin into picromycin and accepts in addition a variety of polyketide substrates, particularly those related to narbomycin in structure. The present invention also provides methods of hydroxylating polyketides, which method comprises contacting the polyketide with the recombinant PicK enzyme under conditions such that hydroxylation occurs. This methodology is applicable to large numbers of polyketides.
[0069] DNA encoding the picK gene can be isolated from cosmid pKOS023-26 of the invention. The DNA sequence of the picK gene is shown in the preceding section. This DNA sequence encodes one of the recombinant forms of the enzyme provided by the invention. The amino acid sequence of this form of the picK gene is shown below. The present invention also provides a recombinant picK gene that encodes a picK gene product in which the PicK protein is fused to a number of consecutive histidine residues, which facilitates purification from recombinant host cells.
tems. These systems contain the coding sequences operably linked to promoters, enhancers, and/or termination sequences that operate to effect expression of the coding sequence in compatible host cells. The host cells are modified by transformation with the recombinant DNA expression vectors of the invention to contain these sequences either as extrachromosomal elements or integrated into the chromosome. The invention also provides methods to produce PKS and post-PKS tailoring enzymes as well as polyketides and antibiotics using these modified host cells.
[0073] As used herein, the term expression vector refers to a nucleic acid that can be introduced into a host cell or cell-free transcription and translation medium. An expression vector can be maintained stably or transiently in a cell, whether as part of the chromosomal or other DNA in the cell or in any cellular compartment, such as a replicating vector in the cytoplasm. An expression vector also comprises a gene that serves to produce RNA, which typically is translated into a polypeptide in the cell or cell extract. To drive

[^1][0070] The recombinant PicK enzyme of the invention hydroxylates narbomycin at the C12 position and YC-17 at either the C10 or C12 position. Hydroxylation of these compounds at the respective positions increases the antibiotic activity of the compound relative to the unhydroxylated compound. Hydroxylation can be achieved by a number of methods. First, the hydroxylation may be performed in vitro using purified hydroxylase, or the relevant hydroxylase can be produced recombinantly and utilized directly in the cell that produces it. Thus, hydroxylation may be effected by supplying the nonhydroxylated precursor to a cell that expresses the hydroxylase. These and other details of this embodiment of the invention are described in additional detail below in Section IV and the examples.
[0071] Section IV: Heterologous Expression of the Narbonolide PKS; the Desosamine Biosynthetic and transferase Genes; the Beta-Glucosidase Gene; and the picK Hydroxylase Gene
[0072] In one important embodiment, the invention provides methods for the heterologous expression of one or more of the genes involved in picromycin biosynthesis and recombinant DNA expression vectors useful in the method. Thus, included within the scope of the invention in addition to isolated nucleic acids encoding domains, modules, or proteins of the narbonolide PKS, glycosylation, and/or hydroxylation enzymes, are recombinant expression sys-
production of the RNA, the expression vector typically comprises one or more promoter elements. Furthermore, expression vectors typically contain additional functional elements, such as, for example, a resistance-conferring gene that acts as a selectable marker.
[0074] The various components of an expression vector can vary widely, depending on the intended use of the vector. In particular, the components depend on the host cell(s) in which the vector will be introduced or in which it is intended to function. Components for expression and maintenance of vectors in E. coli are widely known and commercially available, as are components for other commonly used organisms, such as yeast cells and Streptomyces cells.
[0075] One important component is the promoter, which can be referred to as, or can be included within, a control sequence or control element, which drives expression of the desired gene product in the heterologous host cell. Suitable promoters include those that function in eucaryotic or procaryotic host cells. In addition to a promoter, a control element can include, optionally, operator sequences, and other elements, such as ribosome binding sites, depending on the nature of the host. Regulatory sequences that allow for regulation of expression of the heterologous gene relative to the growth of the host cell may also be included. Examples of such regulatory sequences known to those of skill in the art are those that cause the expression of a gene
to be turned on or off in response to a chemical or physical stimulus. Preferred host cells for purposes of selecting vector components include fungal host cells such as yeast and procaryotic, especially E. coli and Streptomyces, host cells, but single cell cultures of, for example, mammalian cells can also be used. In hosts such as yeasts, plants, or mammalian cells that ordinarily do not produce polyketides, it may be necessary to provide, also typically by recombinant means, suitable holo-ACP synthases to convert the recombinantly produced PKS to functionality. Provision of such enzymes is described, for example, in PCI publication Nos. WO 97/13845 and 98/27203, each of which is incorporated herein by reference. Control systems for expression in yeast, including controls that effect secretion are widely available and can be routinely used. For $E$. coli or other bacterial host cells, promoters such as those derived from sugar metabolizing enzymes, such as galactose, lactose (lac), and maltose, can be used. Additional examples include promoters derived from genes encoding biosynthetic enzymes, and the tryptophan (trp), the beta-lactamase (bla), bacteriophage lambda PL, and T5 promoters. In addition, synthetic promoters, such as the tac promoter (U.S. Pat. No. $4,551,433$ ), can also be used.
[0076] Particularly preferred are control sequences compatible with Streptomyces spp. Particularly useful promoters for Streptomyces host cells include those from PKS gene clusters that result in the production of polyketides as secondary metabolites, including promoters from aromatic (Type II) PKS gene clusters. Examples of Type II PKS gene cluster promoters are act gene promoters and tem gene promoters; an example of a Type I PKS gene cluster promoter is the spiramycin PKS gene promoter.
[0077] If a Streptomyces or other host ordinarily produces polyketides, it may be desirable to modify the host so as to prevent the production of endogenous polyketides prior to its use to express a recombinant PKS of the invention. Such hosts have been described, for example, in U.S. Pat. No. $5,672,491$, incorporated herein by reference. In such hosts, it may not be necessary to provide enzymatic activities for all of the desired post-translational modifications of the enzymes that make up the recombinantly produced PKS, because the host naturally expresses such enzymes. In particular, these hosts generally contain holo-ACP synthases that provide the pantotheinyl residue needed for functionality of the PKS.
[0078] Thus, in one important embodiment, the vectors of the invention are used to transform Streptomyces host cells to provide the recombinant Streptomyces host cells of the invention. Streptomyces is a convenient host for expressing narbonolide or 10 -deoxymethynolide or derivatives of those compounds, because narbonolide and 10 -deoxymethynolide are naturally produced in certain Streptomyces species, and Streptomyces generally produce the precursors needed to form the desired polyketide. The present invention also provides the narbonolide PKS gene promoter in recombinant form, located upstream of the picAI gene on cosmid pKOS023-27. This promoter can be used to drive expression of the narbonolide PKS or any other coding sequence of interest in host cells in which the promoter functions, particularly $S$. venezuelae and generally any Streptomyces species. As described below, however, promoters other than the promoter of the narbonolide PKS genes will typically be used for heterologous expression.
[0079] For purposes of the invention, any host cell other than Streptomyces venezuelae is a heterologous host cell. Thus, $S$. narbonensis, which produces narbomycin but not picromycin is a heterologous host cell of the invention, although other host cells are generally preferred for purposes of heterologous expression. Those of skill in the art will recognize that, if a Streptomyces host that produces a picromycin or methymycin precursor is used as the host cell, the recombinant vector need drive expression of only a portion of the genes constituting the picromycin gene cluster. As used herein, the picromycin gene cluster includes the narbonolide PKS, the desosamine biosynthetic and transferase genes, the beta-glucosidase gene, and the picK hydroxylase gene. Thus, such a vector may comprise only a single ORF, with the desired remainder of the polypeptides encoded by the picromycin gene cluster provided by the genes, on the host cell chromosomal DNA.
[0080] The present invention also provides compounds and recombinant DNA vectors useful for disrupting any gene in the picromycin gene cluster (as described above and illustrated in the examples below). Thus, the invention provides a variety of modified host cells (particularly, $S$. narbonensis and $S$. venezuelae) in which one or more of the genes in the picromycin gene cluster have been disrupted. These cells are especially useful when it is desired to replace the disrupted function with a gene product expressed by a recombinant DNA vector. Thus, the invention provides such Streptomyces host cells, which are preferred host cells for expressing narbonolide derivatives of the invention. Particularly preferred host cells of this type include those in which the coding sequence for the loading module has been disrupted, those in which one or more of any of the PKS gene ORFs has been disrupted, and/or those in which the picK gene has been disrupted.
[0081] In a preferred embodiment, the expression vectors of the invention are used to construct a heterologous recombinant Streptomyces host cell that expresses a recombinant PKS of the invention. As noted above, a heterologous host cell for purposes of the present invention is any host cell other than $S$. venezuelae, and in most cases other than $S$. narbonensis as well. Particularly preferred heterologous host cells are those which lack endogenous functional PKS genes. Illustrative host cells of this type include the modified Streptomyces coelicolor CH999 and similarly modified S. lividans described in PCT publication No. WO 96/40968.
[0082] The invention provides a wide variety of expression vectors for use in Streptomyces. For replicating vectors, the origin of replication can be, for example and without limitation, a low copy number vector, such as SCP2* (see Hopwood et al., Genetic Manipulation of Streptomyces: A Laboratory manual (The John Innes Foundation, Norwich, U.K., 1985); Lydiate et al., 1985, Gene 35: 223-235; and Kieser and Melton, 1988, Gene 65: 83-91, each of which is incorporated herein by reference), SLP1.2 (Thompson et al., 1982, Gene 20: 51-62, incorporated herein by reference), and pSG5(ts) (Muth et al., 1989, Mol. Gen. Genet. 219: 341-348, and Bierman et al., 1992, Gene 116: 43-49, each of which is incorporated herein by reference), or a high copy number vector, such as pIJ101 and pJV1 (see Katz et al., 1983, J. Gen. Microbiol. 129: 2703-2714; Vara et al., 1989, J. Bacteriol. 171: 5782-5781; and Servin-Gonzalez, 1993, Plasmid 30: 131-140, each of which is incorporated herein by reference). High copy number vectors are generally, however, not preferred for expression of large genes or multiple genes. For non-replicating and integrating vectors and generally for any vector, it is useful to include at least
an $E$. coli origin of replication, such as from pUC, $\mathrm{p} 1 \mathrm{P}, \mathrm{p} 1 \mathrm{I}$, and pBR . For phage based vectors, the phage phiC31 and its derivative KC515 can be employed (see Hopwood et al., supra). Also, plasmid pSET152, plasmid pSAM, plasmids pSE101 and pSE211, all of which integrate site-specifically in the chromosomal DNA of S. lividans, can be employed.
[0083] Preferred Streptomyces host cell/vector combinations of the invention include $S$. coelicolor CH 999 and, $S$. lividans K4-114 host cells, which do not produce actinorhodin, and expression vectors derived from the pRM1 and pRM5 vectors, as described in U.S. Pat. No. 5,830,750 and U.S. patent application Ser. No. 08/828,898, filed 31 Mar. 1997, and Ser. No. 09/181,833, filed 28 Oct. 1998, each of which is incorporated herein by reference.
[0084] As described above, particularly useful control sequences are those that alone or together with suitable regulatory systems activate expression during transition from growth to stationary phase in the vegetative mycelium. The system contained in the illustrative plasmid pRM5, i.e., the actI/actIII promoter pair and the actII-ORF4 activator gene, is particularly preferred. Other useful Streptomyces promoters include without limitation those from the ermE gene and the melC1 gene, which act constitutively, and the tipA gene and the merA gene, which can be induced at any growth stage. In addition, the T7 RNA polymerase system has been transferred to Streptomyces and can be employed in the vectors and host cells of the invention. In this system, the coding sequence for the T7 RNA polymerase is inserted into a neutral site of the chromosome or in a vector under the control of the inducible merA promoter, and the gene of interest is placed under the control of the T7 promoter. As noted above, one or more activator genes can also be employed to enhance the activity of a promoter. Activator genes in addition to the actII-ORF4 gene described above include dnrI, redD, and ptpA genes (see U.S. patent application Ser. No. 09/181,833, supra).
[0085] Typically, the expression vector will comprise one or more marker genes by which host cells containing the vector can be identified and/or selected. Selectable markers are often preferred for recombinant expression vectors. A variety of markers are known that are useful in selecting for transformed cell lines and generally comprise a gene that confers a selectable phenotype on transformed cells when the cells are grown in an appropriate selective medium. Such markers include, for example, genes that confer antibiotic resistance or sensitivity to the plasmid. Alternatively, several polyketides are naturally colored, and this characteristic can provide a built-in marker for identifying cells. Preferred selectable markers include antibiotic resistance conferring genes. Preferred for use in Streptomyces host cells are the ermE (confers resistance to erythromycin and lincomycin), tsr (confers resistance to thiostrepton), aadA (confers resistance to spectinomycin and streptomycin), aacC4 (confers resistance to apramycin, kanamycin, gentamicin, geneticin (G418), and neomycin), hyg (confers resistance to hygromycin), and vph (confers resistance to viomycin) resistance conferring genes.
[0086] To provide a preferred host cell and vector for purposes of the invention, the narbonolide PKS genes were placed on a recombinant expression vector that was transferred to the non-macrolide producing host Streptomyces lividans K4-114, as described in Example 3. Transformation of $S$. lividans K4-114 with this expression vector resulted in a strain which produced two compounds in similar yield ( $5-10 \mathrm{mg} / \mathrm{L}$ each). Analysis of extracts by LC/MS followed
by ${ }^{1} \mathrm{H}$-NMR spectroscopy of the purified compounds established their identity as narbonolide (FIG. 5, compound 4) and 10 -deoxymethynolide (FIG. 5, compound 5), the respective 14 and 12 -membered polyketide precursors of narbomycin and YC17.
[0087] To provide a host cell of the invention that produces the narbonolide PKS as well as an additional narbonolide biosynthetic gene and to investigate the possible role of the Pik TEII in picromycin biosynthesis, the picB gene was integrated into the chromosome to provide the host cell of the invention Streptomyces lividans K39-18. The picB gene was cloned into the Streptomyces genome integrating vector pSET152 (see Bierman et al., 1992, Gene 116:43, incorporated herein by reference) under control of the same promoter (PactI) as the PKS on plasmid pKOS03986.
[0088] A comparison of strains Streptomyces lividans K39-18/pKOS039-86 and KS 114/pKOS039-86 grown under identical conditions indicated that the strain containing TEII produced 47 times more total polyketide. This increased production indicates that the enzyme is functional in this strain and is consistent with the observation that yields fall to below $5 \%$ for both picromycin and methymycin when picB is disrupted in S. venezuelae. Because the production levels of compound 4 and 5 from K39-18/ pKOS03986 increased by the same relative amounts, TEII does not appear to influence the ratio of 12 and 14 -membered lactone ring formation. Thus, the invention provides methods of coexpressing the picB gene product or any other type II thioesterase with the narbonolide PKS or any other PKS in heterologous host cells to increase polyketide production.
[0089] In accordance with the methods of the invention, picromycin biosynthetic genes in addition to the genes encoding the PKS and Pik TEII can be introduced into heterologous host cells. In particular, the picK gene, desosamine biosynthetic genes, and the desosaminyl transferase gene can be expressed in the recombinant host cells of the invention to produce any and all of the polyketides in the picromycin biosynthetic pathway (or derivatives thereof). Those of skill will recognize that the present invention enables one to select whether only the 12 -membered polyketides, or only the 14 -membered polyketides, or both 12 - and 14 -membered polyketides will be produced. To produce only the 12 -membered polyketides, the invention provides expression vectors in which the last module is deleted or the KS domain of that module is deleted or rendered inactive. To produce only the 14 -membered polyketides, the invention provides expression vectors in which the coding sequences of extender modules 5 and 6 are fused to provide only a single polypeptide.
[0090] In one important embodiment, the invention provides methods for desosaminylating polyketides or other compounds. In this method, a host cell other than Streptomyces venezuelae is transformed with one or more recombinant vectors of the invention comprising the desosamine biosynthetic and desosaminyl transferase genes and control sequences positioned to express those genes. The host cells so transformed can either produce the polyketide to be desosaminylated naturally or can be transformed with expression vectors encoding the PKS that produces the desired polyketide. Alternatively, the polyketide can be supplied to the host cell containing those genes. Upon production of the polyketide and expression of the desosamine biosynthetic and desosaminyl transferase genes, the
desired desosaminylated polyketide is produced. This method is especially useful in the production of polyketides to be used as antibiotics, because the presence of the desosamine residue is known to increase, relative to their undesosaminylated counterparts, the antibiotic activity of many polyketides significantly. The present invention also provides a method for desosaminylating a polyketide by transforming an $S$. venezuelae or $S$. narbonensis host cell with a recombinant vector that encodes a PKS that produces the polyketide and culturing the transformed cell under conditions such that said polyketide is produced and desosaminylated. In this method, use of an $S$. venezuelae or $S$. narbonensis host cell of the invention that does not produce a functional endogenous narbonolide PKS is preferred.
[0091] In a related aspect, the invention provides a method for improving the yield of a desired desosaminylated polyketide in a host cell, which method comprises transforming the host cell with a beta-glucosidase gene. This method is not limited to host cells that have been transformed with expression vectors of the invention encoding the desosamine biosynthetic and desosaminyl transferase genes of the invention but instead can be applied to any host cell that desosaminylates polyketides or other compounds. Moreover, while the beta-glucosidase gene from Streptomyces venezuelae provided by the invention is preferred for use in the method, any beta-glucosidase gene may be employed. In another embodiment, the beta-glucosidase treatment is conducted in a cell free extract.
[0092] Thus, the invention provides methods not only for producing narbonolide and 10 -deoxymethynolide in heterologous host cells but also for producing narbomycin and YC-17 in heterologous host cells. In addition, the invention provides methods for expressing the picK gene product in heterologous host cells, thus providing a means to produce picromycin, methymycin, and neomethymycin in heterologous host cells. Moreover, because the recombinant expression vectors provided by the invention enable the artisan to provide for desosamine biosynthesis and transfer and/or C10 or C12 hydroxylation in any host cell, the invention provides methods and reagents for producing a very wide variety of glycosylated and/or hydroxylated polyketides. This variety of polyketides provided by the invention can be better appreciated upon consideration of the following section relating to the production of polyketides from heterologous or hybrid PKS enzymes provided by the invention.

## [0093] Section V: Hybrid PKS Genes

[0094] The present invention provides recombinant DNA compounds encoding each of the domains of each of the modules of the narbonolide PKS, the proteins involved in desosamine biosynthesis and transfer to narbonolide, and the PicK protein. The availability of these compounds permits their use in recombinant procedures for production of desired portions of the narbonolide PKS fused to or expressed in conjunction with all or a portion of a heterologous PKS. The resulting hybrid PKS can then be expressed in a host cell, optionally with the desosamine biosynthesis and transfer genes and/or the picK hydroxylase gene to produce a desired polyketide.
[0095] Thus, in accordance with the methods of the invention, a portion of the narbonolide PKS coding sequence that encodes a particular activity can be isolated and manipulated, for example, to replace the corresponding region in a different modular PKS. In addition, coding sequences for
individual modules of the PKS can be ligated into suitable expression systems and used to produce the portion of the protein encoded. The resulting protein can be isolated and purified or can may be employed in situ to effect polyketide synthesis. Depending on the host for the recombinant production of the domain, module, protein, or combination of proteins, suitable control sequences such as promoters, termination sequences, enhancers, and the like are ligated to the nucleotide sequence encoding the desired protein in the construction of the expression vector.
[0096] In one important embodiment, the invention thus provides a hybrid PKS and the corresponding recombinant DNA compounds that encode those hybrid PKS enzymes. For purposes of the invention, a hybrid PKS is a recombinant PKS that comprises all or part of one or more extender modules, loading module, and/or thioesterase/cyclase domain of a first PKS and all or part of one or more extender modules, loading module, and/or thioesterase/cyclase domain of a second PKS. In one preferred embodiment, the first PKS is most but not all of the narbonolide PKS, and the second PKS is only a portion or all of a non-narbonolide PKS. An illustrative example of such a hybrid PKS includes a narbonolide PKS in which the natural loading module has been replaced with a loading module of another PKS. Another example of such a hybrid PKS is a narbonolide PKS in which the AT domain of extender module 3 is replaced with an AT domain that binds only malonyl CoA.
[0097] In another preferred embodiment, the first PKS is most but not all of a non-narbonolide PKS, and the second PKS is only a portion or all of the narbonolide PKS. An illustrative example of such a hybrid PKS includes a DEBS PKS in which an AT specific for methylmalonyl CoA is replaced with the AT from the narbonolide PKS specific for malonyl CoA.
[0098] Those of skill in the art will recognize that all or part of either the first or second PKS in a hybrid PKS of the invention need not be isolated from a naturally occurring source. For example, only a small portion of an AT domain determines its specificity. See U.S. provisional patent application Ser. No. 60/091,526, and Lau et al., infra, incorporated herein by reference. The state of the art in DNA synthesis allows the artisan to construct de novo DNA compounds of size sufficient to construct a useful portion of a PKS module or domain. Thus, the desired derivative coding sequences can be synthesized using standard solid phase synthesis methods such as those described by Jaye et al., 1984, J. Biol. Chem. 259: 6331, and instruments for automated synthesis are available commercially from, for example, Applied Biosystems, Inc. For purposes of the invention, such synthetic DNA compounds are deemed to be a portion of a PKS.
[0099] With this general background regarding hybrid PKSs of the invention, one can better appreciate the benefit provided by the DNA compounds of the invention that encode the individual domains, modules, and proteins that comprise the narbonolide PKS. As described above, the narbonolide PKS is comprised of a loading module, six extender modules composed of a KS, AT, ACP, and zero, one, two, or three KR, DH, and ER domains, and a thioesterase domain. The DNA compounds of the invention that encode these domains individually or in combination are useful in the construction of the hybrid PKS encoding DNA compounds of the invention.
[0100] The recombinant DNA compounds of the invention that encode the loading module of the narbonolide PKS and the corresponding polypeptides encoded thereby are useful for a variety of applications. In one embodiment, a DNA compound comprising a sequence that encodes the narbonolide PKS loading module is inserted into a DNA compound that comprises the coding sequence for a heterologous PKS. The resulting construct, in which the coding sequence for the loading module of the heterologous PKS is replaced by that for the coding sequence of the narbonolide PKS loading module provides a novel PKS. Examples include the 6-deoxyerythronolide B, rapamycin, FK506, FK520, rifamycin, and avermectin PKS coding sequences. In another embodiment, a DNA compound comprising a sequence that encodes the narbonolide PKS loading module is inserted into a DNA compound that comprises the coding sequence for the narbonolide PKS or a recombinant narbonolide PKS that produces a narbonolide derivative.
[0101] In another embodiment, a portion of the loading module coding sequence is utilized in conjunction with a heterologous coding sequence. In this embodiment, the invention provides, for example, replacing the propionyl CoA specific AT with an acetyl CoA, butyryl CoA, or other CoA specific AT. In addition, the $\mathrm{KS}^{\mathrm{Q}}$ and/or ACP can be replaced by another inactivated KS and/or another ACP. Alternatively, the $\mathrm{KS}^{\mathrm{Q}}, \mathrm{AT}$, and ACP of the loading module can be replaced by an AT and ACP of a loading module such as that of DEBS. The resulting heterologous loading module coding sequence can be utilized in conjunction with a coding sequence for a PKS that synthesizes narbonolide, a narbonolide derivative, or another polyketide.
[0102] The recombinant DNA compounds of the invention that encode the first extender module of the narbonolide PKS and the corresponding polypeptides encoded thereby are useful for a variety of applications. In one embodiment, a DNA compound comprising a sequence that encodes the narbonolide PKS first extender module is inserted into a DNA compound that comprises the coding sequence for a heterologous PKS. The resulting construct, in which the coding sequence for a module of the heterologous PKS is either replaced by that for the first extender module of the narbonolide PKS or the latter is merely added to coding sequences for modules of the heterologous PKS, provides a novel PKS coding sequence. In another embodiment, a DNA compound comprising a sequence that encodes the first extender module of the narbonolide PKS is inserted into a DNA compound that comprises coding sequences for the narbonolide PKS or a recombinant narbonolide PKS that produces a narbonolide derivative.
[0103] In another embodiment, a portion or all of the first extender module coding sequence is utilized in conjunction with other PKS coding sequences to create a hybrid module. In this embodiment, the invention provides, for example, replacing the methylmalonyl $\mathrm{Co} A$ specific AT with a malonyl CoA, ethylmalonyl CoA, or carboxyglycolyl CoA specific $A T$; deleting (which includes inactivating) the KR; inserting a DH or a DH and ER; and/or replacing the KR with another KR, a DH and KR, or a DH, KR, and ER. In addition, the KS and/or ACP can be replaced with another KS and/or ACP. In each of these replacements or insertions, the heterologous MS, AT, DH, KR, ER, or ACP coding sequence can originate from a coding sequence for another module of the narbonolide PKS, from a gene for a PKS that
produces a polyketide other than narbonolide, or from chemical synthesis. The resulting heterologous first extender module coding sequence can be utilized in conjunction with a coding sequence for a PKS that synthesizes narbonolide, a narbonolide derivative, or another polyketide.
[0104] In an illustrative embodiment of this aspect of the invention, the invention provides recombinant PKSs and recombinant DNA compounds and vectors that encode such PKSs in which the KS domain of the first extender module has been inactivated. Such constructs are especially useful when placed in translational reading frame with the remaining modules and domains of a narbonolide PKS or narbonolide derivative PKS. The utility of these constructs is that host cells expressing, or cell free extracts containing, the PKS encoded thereby can be fed or supplied with N -acetylcysteamine thioesters of novel precursor molecules to prepare narbonolide derivatives. See U.S. patent application Ser. No. 60/117,384, filed 27 Jan. 1999, and PCT publication Nos. WO 99/03986 and 97/02358, each of which is incorporated herein by reference.
[0105] The recombinant DNA compounds of the invention that encode the second extender module of the narbonolide PKS and the corresponding polypeptides encoded thereby are useful for a variety of applications. In one embodiment, a DNA compound comprising a sequence that encodes the narbonolide PKS second extender module is inserted into a DNA compound that comprises the coding sequence for a heterologous PKS. The resulting construct, in which the coding sequence for a module of the heterologous PKS is either replaced by that for the second extender module of the narbonolide PKS or the latter is merely added to coding sequences for the modules of the heterologous PKS, provides a novel PKS. In another embodiment, a DNA compound comprising a sequence that encodes the second extender module of the narbonolide PKS is inserted into a DNA compound that comprises the coding sequences for the narbonolide PKS or a recombinant narbonolide PKS that produces a narbonolide derivative.
[0106] In another embodiment, a portion or all of the second extender module coding sequence is utilized in conjunction with other PKS coding sequences to create a hybrid module. In this embodiment, the invention provides, for example, replacing the malonyl CoA specific AT with a methylmalonyl CoA, ethylmalonyl CoA, or carboxyglycolyl CoA specific AT, deleting (or inactivating) the KR, the DH, or both the DH and KR; replacing the KR or the KR and DH with a KR, a KR and a DH, or a KR, DH, and ER; and/or inserting an ER. In addition, the KS and/or ACP can be replaced with another KS and/or ACP. In each of these replacements or insertions, the heterologous KS, AT, DH, KR, ER, or ACP coding sequence can originate from a coding sequence for another module of the narbonolide PKS, from a coding sequence for a PKS that produces a polyketide other than narbonolide, or from chemical synthesis. The resulting heterologous second extender module coding sequence can be utilized in conjunction with a coding sequence from a PKS that synthesizes narbonolide, a narbonolide derivative, or another polyketide.
[0107] The recombinant DNA compounds of the invention that encode the third extender module of the narbonolide PKS and the corresponding polypeptides encoded thereby are useful for a variety of applications. In one embodiment,
a DNA compound comprising a sequence that encodes the narbonolide PKS third extender module is inserted into a DNA compound that comprises the coding sequence for a heterologous PKS. The resulting construct, in which the coding sequence for a module of the heterologous PKS is either replaced by that for the third extender module of the narbonolide PKS or the latter is merely added to coding sequences for the modules of the heterologous PKS, provides a novel PKS. In another embodiment, a DNA compound comprising a sequence that encodes the third extender module of the narbonolide PKS is inserted into a DNA compound that comprises coding sequences for the narbonolide PKS or a recombinant narbonolide PKS that produces a narbonolide derivative.
[0108] In another embodiment, a portion or all of the third extender module coding sequence is utilized in conjunction with other PKS coding sequences to create a hybrid module. In this embodiment, the invention provides, for example, replacing the methylmalonyl $\mathrm{Co} A$ specific AT with a malonyl CoA, ethylmalonyl CoA, or carboxyglycolyl CoA specific AT; deleting the inactive KR; and/or inserting a KR, or a KR and DH, or a KR, DH, and ER. In addition, the KS and/or ACP can be replaced with another KS and/or ACP. In each of these replacements or insertions, the heterologous KS, AT, DH, KR, ER, or ACP coding sequence can originate from a coding sequence for another module of the narbonolide PKS, from a gene for a PKS that produces a polyketide other than narbonolide, or from chemical synthesis. The resulting heterologous third extender module coding sequence can be utilized in conjunction with a coding sequence for a PKS that synthesizes narbonolide, a narbonolide derivative, or another polyketide.
[0109] The recombinant DNA compounds of the invention that encode the fourth extender module of the narbonolide PKS and the corresponding polypeptides encoded thereby are useful for a variety of applications. In one embodiment, a DNA compound comprising a sequence that encodes the narbonolide PKS fourth extender module is inserted into a DNA compound that comprises the coding sequence for a heterologous PKS. The resulting construct, in which the coding sequence for a module of the heterologous PKS is either replaced by that for the fourth extender module of the narbonolide PKS or the latter is merely added to coding sequences for the modules of the heterologous PKS, provides a novel PKS. In another embodiment, a DNA compound comprising a sequence that encodes the fourth extender module of the narbonolide. PKS is inserted into a DNA compound that comprises coding sequences for the narbonolide PKS or a recombinant narbonolide PKS that produces a narbonolide derivative.
[0110] In another embodiment, a portion of the fourth extender module coding sequence is utilized in conjunction with other PKS coding sequences to create a hybrid module. In this embodiment, the invention provides, for example, replacing the methylmalonyl CoA specific AT with a malonyl CoA, ethylmalonyl CoA, or carboxyglycolyl CoA specific AT; deleting any one, two, or all three of the ER, DH, and KR; and/or replacing any one, two, or all three of the ER, DH, and KR with either a KR, a DH and KR, or a KR, DH, and ER. In addition, the KS and/or ACP can be replaced with another KS and/or ACP. In each of these replacements or insertions, the heterologous KS, AT, DH, KR, ER, or ACP coding sequence can originate from a coding sequence for
another module of the narbonolide PKS, from a coding sequence for a PKS that produces a polyketide other than narbonolide, or from chemical synthesis. The resulting heterologous fourth extender module coding sequence can be utilized in conjunction with a coding sequence for a PKS that synthesizes narbonolide, a narbonolide derivative, or another polyketide.
[0111] The recombinant DNA compounds of the invention that encode the fifth extender module of the narbonolide PKS and the corresponding polypeptides encoded thereby are useful for a variety of applications. In one embodiment, a DNA compound comprising a sequence that encodes the narbonolide PKS fifth extender module is inserted into a DNA compound that comprises the coding sequence for a heterologous PKS. The resulting construct, in which the coding sequence for a module of the heterologous PKS is either replaced by that for the fifth extender module of the narbonolide PKS or the latter is merely added to coding sequences for the modules of the heterologous PKS, provides a novel PKS. In another embodiment, a DNA compound comprising a sequence that encodes the fifth extender module of the narbonolide PKS is inserted into a DNA compound that comprises the coding sequence for the narbonolide PKS or a recombinant narbonolide PKS that produces a narbonolide derivative.
[0112] In another embodiment, a portion or all of the fifth extender module coding sequence is utilized in conjunction with other PKS coding sequences to create a hybrid module. In this embodiment, the invention provides, for example, replacing the methylmalonyl CoA specific AT with a malonyl CoA, ethylmalonyl CoA, or carboxyglycolyl CoA specific AT; deleting (or inactivating) the KR; inserting a DH or a DH and ER; and/or replacing the KR with another KR, a DH and KR, or a DH, KR, and ER. In addition, the KS and/or ACP can be replaced with another KS and/or ACP. In each of these replacements or insertions, the heterologous $\mathrm{KS}, \mathrm{AT}, \mathrm{DH}, \mathrm{KR}, \mathrm{ER}$, or ACP coding sequence can originate from a coding sequence for another module of the narbonolide PKS, from a coding sequence for a PKS that produces a polyketide other than narbonolide, or from chemical synthesis. The restyling heterologous fifth extender module coding sequence can be utilized in conjunction with a coding sequence for a PKS that synthesizes narbonolide, a narbonolide derivative, or another polyketide.
[0113] The recombinant DNA compounds of the invention that encode the sixth extender module of the narbonolide PKS and the corresponding polypeptides encoded thereby are useful for a variety of applications. In one embodiment, a DNA compound comprising a sequence that encodes the narbonolide PKS sixth extender module is inserted into a DNA compound that comprises the coding sequence for a heterologous PKS. The resulting construct, in which the coding sequence for a module of the heterologous PKS is either replaced by that for the sixth extender module of the narbonolide PKS or the latter is merely added to coding sequences for the modules of the heterologous PKS, provides a novel PKS. In another embodiment, a DNA compound comprising a sequence that encodes the sixth extender module of the narbonolide PKS is inserted into a DNA compound that comprises the coding sequences for the narbonolide PKS or a recombinant narbonolide PKS that produces a narbonolide derivative.
[0114] In another embodiment, a portion or all of the sixth extender module coding sequence is utilized in conjunction with other PKS coding sequences to create a hybrid module. In this embodiment, the invention provides, for example, replacing the methylmalonyl CoA specific AT with a malonyl CoA, ethylmalonyl CoA, or carboxyglycolyl CoA specific AT; and/or inserting a KR, a KR and DH, or a KR, DH, and an ER. In addition, the KS and/or ACP can be replaced with another KS and/or ACP. In each of these replacements or insertions, the heterologous KS, AT, DH, KR, ER, or ACP coding sequence can originate from a coding sequence for another module of the narbonolide PKS, from a coding sequence for a PKS that produces a polyketide other than narbonolide, or from chemical synthesis. The resulting heterologous sixth extender module coding sequence can be utilized in conjunction with a coding sequence for a PKS that synthesizes narbonolide, a narbonolide derivative, or another polyketide.
[0115] The sixth extender module of the narbonolide PKS is followed by a thioesterase domain. This domain is important in the cyclization of the polyketide and its cleavage from the PKS. The present invention provides recombinant DNA compounds that encode hybrid PKS enzymes in which the narbonolide PKS is fused to a heterologous thioesterase or a heterologous PKS is fused to the narbonolide synthase thioesterase. Thus, for example, a thioesterase domain coding sequence from another PKS gene can be inserted at the end of the sixth extender module coding sequence in recombinant DNA compounds of the invention. Recombinant DNA compounds encoding this thioesterase domain are therefore useful in constructing DNA compounds that encode the narbonolide PKS, a PKS that produces a narbonolide derivative, and a PKS that produces a polyketide other than narbonolide or a narbonolide derivative. The following Table lists references describing illustrative PKS genes and corresponding enzymes that can be utilized in the construction of the recombinant hybrid PKSs and the corresponding DNA compounds that encode them of the invention. Also presented are various references describing tailoring enzymes and corresponding genes that can be employed in accordance with the methods of the invention.
[0116] Avermectin
[0117] U.S. Pat. No. 5,252,474 to Merck.
[0118] MacNeil et al., 1993, Industrial Microorganisms: Basic and Applied Molecular Genetics, Baltz, Hegeman, \& Skatrud, eds. (ASM), pp. 245-256, A Comparison of the Genes Encoding the Polyketide Synthases for Avermectin, Erythromycin, and Nemadectin.
[0119] MacNeil et al., 1992, Gene 115: 119-125, Complex Organization of the Streptomyces avermitilis genes encoding the avermectin polyketide synthase.
[0120] Candicidin (FR008)
[0121] Hu et al., 1994, Mol. Microbiol. 14 163-172.
[0122] Epothilone
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[0171] Merson-Davies and Cundliffe, 1994, Mol. Microbiol. 13: 349-355. Analysis of five tylosin biosynthetic genes from the tylBA region of the Streptomyces fradiae genome.
[0172] As the above Table illustrates, there are a wide variety of PKS genes that serve as readily available sources of DNA and sequence information for use in constructing the hybrid PKS-encoding DNA compounds of the invention. Methods for constructing hybrid PKS-encoding DNA compounds are described without reference to the narbonolide PKS in U.S. Pat. Nos. 5,672,491 and 5,712,146 and PCT publication No. 98/49315, each of which is incorporated herein by reference.
[0173] In constructing hybrid PKSs of the invention, certain general methods may be helpful. For example, it is often beneficial to retain the framework of the module to be altered to make the hybrid PKS. Thus, if one desires to add DH and ER functionalities to a module, it is often preferred to replace the KR domain of the original module with a KR, DH, and ER domain-containing segment from another module, instead of merely inserting DH and ER domains. One can alter the stereochemical specificity of a module by replacement of the KS domain with a KS domain from a module that specifies a different stereochemistry. See Lau et a1., 1999, Dissecting the role of acyltransferase domains of modular polyketide synthases in the choice and stereochemical fate of extender units"Biochemistry 38(5):1643-1651, incorporated herein by reference. One can alter the specificity of an AT domain by changing only a small segment of the domain. See Lau et al., supra. One can also take advantage of known linker regions in PKS proteins to link modules from two different PKSs to create a hybrid PKS. See Gokhale et al., 16 Apr. 1999, Dissecting and Exploiting Intermodular Communication in Polyketide Synthases", Science 284: 482-485, incorporated herein by reference.
[0174] The hybrid PKS-encoding DNA compounds of the invention can be and often are hybrids of more than two PKS genes. Even where only two genes are used, there are often two or more modules in the hybrid gene in which all or part of the module is derived from a second (or third) PKS gene. Thus, as one illustrative example, the invention provides a hybrid narbonolide PKS that contains the naturally occurring loading module and thioesterase domain as well as extender modules one, two, four, and six of the narbonolide PKS and further contains hybrid or heterologous extender modules three and five. Hybrid or heterologous extender modules three and five contain AT domains specific for malonyl CoA and derived from, for example, the rapamycin PKS genes.
[0175] To construct a hybrid PKS or narbonolide derivative PKS of the invention, one can employ a technique, described in PCT Pub. No. 98/27203, which is incorporated herein by reference, in which the large PKS gene cluster is divided into two or more, typically three, segments, and each segment is placed on a separate expression vector. In this manner, each of the segments of the gene can be altered, and various altered segments can be combined in a single host cell to provide a recombinant PKS gene of the invention. This technique makes more efficient the construction of large libraries of recombinant PKS genes, vectors for expressing those genes, and host cells comprising those vectors.
[0176] The invention also provides libraries of PKS genes, PKS proteins, and ultimately, of polyketides, that are constructed by generating modifications in the narbonolide PKS so that the protein complexes produced have altered activities in one or more respects and thus produce polyketides other than the natural product of the PKS. Novel polyketides may thus be prepared, or polyketides in general prepared more readily, using this method. By providing a large number of different genes or gene clusters derived from a naturally occurring PKS gene cluster, each of which has been modified in a different way from the native cluster, an effectively combinatorial library of polyketides can be produced as a result of the multiple variations in these activities. As will be further described below, the metes and bounds of
this embodiment of the invention can be described on both the protein level and the encoding nucleotide sequence level.
[0177] As described above, a modular PKS "derived from" the narbonolide or other naturally occurring PKS includes a modular PKS (or its corresponding encoding gene(s)) that retains the scaffolding of the utilized portion of the naturally occurring gene. Not all modules need be included in the constructs. On the constant scaffold, at least one enzymatic activity is mutated, deleted, replaced, or inserted so as to alter the activity of the resulting PKS relative to the original PKS. Alteration results when these activities are deleted or are replaced by a different version of the activity, or simply mutated in such a way that a polyketide other then the natural product results from these collective activities. This occurs because there has been a resulting alteration of the starter unit and/or extender unit, and/or stereochemistry, and/or chain length or cyclization, and/or reductive or dehydration cycle outcome at a corresponding position in the product polyketide. Where a deleted activity is replaced, the origin of the replacement activity may come from a corresponding activity in a different naturally occurring PKS or from a different region of the narbonolide PKS. Any or all of the narbonolide PKS genes may be included in the derivative or portions of any of these may be included, but the scaffolding of the PKS protein is retained in whatever derivative is constructed. The derivative preferably contains a thioesterase activity from the narbonolide or another PKS.
[0178] In summary, a PKS derived from the narbonolide PKS includes a PKS that contains the scaffolding of all or a portion of the narbonolide PKS. The derived PKS also contains at least two extender modules that are functional, preferably three extender modules, and more preferably four or more extender modules, and most preferably six extender modules. The derived PKS also contains mutations, deletions, insertions, or replacements of one or more of the activities of the functional modules of the narbonolide PKS so that the nature of the resulting polyketide is altered. This definition applies both at the protein and DNA sequence levels. Particular preferred embodiments include those wherein a KS, AT, KR, DH, or ER has been deleted or replaced by a version of the activity from a different PKS or from another location within the same PKS. Also preferred are derivatives where at least one non-condensation cycle enzymatic activity ( $\mathrm{KR}, \mathrm{DH}$, or ER ) has been deleted or added or wherein any of these activities has been mutated so as to change the structure of the polyketide synthesized by the PKS.
[0179] Conversely, also included within the definition of a PKS derived from the narbonolide PKS are functional PKS modules or their encoding genes wherein at least one portion, preferably two portions, of the narbonolide PKS activities have bean inserted. Exemplary is the use of the narbonolide AT for extender module 2 which accepts a malonyl CoA extender unit rather than methylmalonyl CoA to replace a methylmalonyl specific AT in a PKS. Other examples include insertion of portions of non-condensation cycle enzymatic activities or other regions of narbonolide synthase activity into a heterologous PKS. Again, the derived from definition applies to the PKS at both the genetic and protein levels.
[0180] Thus, there are at least five degrees of freedom for constructing a hybrid PKS in terms of the polyketide that
will be produced. First, the polyketide chain length is determined by the number of modules in the PKS. Second, the nature of the carbon skeleton of the PKS is determined by the specificities of the acyl transferases that determine the nature of the extender units at each position, e.g., malonyl, methylmalonyl, ethylmalonyl, or other substituted malonyl. Third, the loading module specificity also has an effect on the resulting carbon skeleton of the polyketide. The loading module may use a different starter unit, such as acetyl, butyryl, and the like. As noted above and in the examples below, another method for varying loading module specificity involves inactivating the KS activity in extender module 1 (KS1) and providing alternative substrates, called diketides that are chemically synthesized analogs of extender module 1 diketide products, for extender module 2. This approach was illustrated in PCT publication Nos. 97/02358 and 99/03986, incorporated herein by reference, wherein the KS1 activity was inactivated through mutation. Fourth, the oxidation state at various positions of the polyketide will be determined by the dehydratase and reductase portions of the modules. This will determine the presence and location of ketone and alcohol moieties and $\mathrm{C}-\mathrm{C}$ double bonds or C-C single bonds in the polyketide. Finally, the stereochemistry of the resulting polyketide is a function of three aspects of the synthase. The first aspect is related to the AT/KS specificity associated with substituted malonyls as extender units, which affects stereochemistry only when the reductive cycle is missing or when it contains only a ketoreductase, as the dehydratase would abolish chirality. Second, the specificity of the ketoreductace may determine the chirality of any beta-OH. Finally, the enoylreductase specificity for substituted malonyls as extender units may influence the result when there is a complete KR/DH/ER available.
[0181] Thus, the modular PKS systems, and in particular, the narbonolide PKS system, permit a wide range of polyketides to be synthesized. As compared to the aromatic PKS systems, a wider range of starter units including aliphatic monomers (acetyl, propionyl, butyryl, isovaleryl, etc.), aromatics (aminohydroxybenzoyl), alicyclics (cyclohexanoyl), and heterocyclics (thiazolyl) are found in various macrocyclic polyketides. Recent studies have shown that modular PKSs have relaxed specificity for their starter units (Kao et al., 1994, Science, supra). Modular PKSs also exhibit considerable variety with regard to the choice of extender units in each condensation cycle. The degree of beta-ketoreduction following a condensation reaction has also been shown to be altered by genetic manipulation (Donadio et al., 1991, Science, supra; Donadio et al., 1993, Proc. Natl. Acad. Sci. USA 90: 7119-7123). Likewise, the size of the polyketide product can be varied by designing mutants with the appropriate number of modules (Kao et al., 1994, J. Am. Chem. Soc. 116:1612-11613). Lastly, these enzymes are particularly well known for generating an impressive range of asymmetric centers in their products in a highly controlled manner. The polyketides and antibiotics produced by the methods of the invention are typically single stereoisomeric forms. Although the compounds of the invention can occur as mixtures of stereoisomers, it may be beneficial in some instances to generate individual stereoisomers. Thus, the combinatorial potential within modular PKS pathways based on any naturally occurring modular, such as the narbonolide, PKS scaffold is virtually unlimited.
[0182] The combinatorial potential is increased even further when one considers that mutations in DNA encoding a polypeptide can be used to introduce, alter, or delete an activity in the encoded polypeptide. Mutations can be made to the native sequences using conventional techniques. The substrates for mutation can be an entire cluster of genes or only one or two of them; the substrate for mutation may also be portions of one or more of these genes. Techniques for mutation include preparing synthetic oligonucleotides including the mutations and inserting the mutated sequence into the gene encoding a PKS subunit using restriction endonuclease digestion. See, e.g., Kunkel, 1985, Proc. Natl. Acad. Sci. USA 82: 448; Geisselsoder et al., 1987, BioTechniques 5:786. Alternatively, the mutations can be effected using a mismatched primer (generally 10-20 nucleotides in length) that hybridizes to the native nucleotide sequence, at a temperature below the melting temperature of the mismatched duplex. The primer can be made specific by keeping primer length and base composition within relatively narrow limits and by keeping the mutant base centrally located, See Zoller and Smith, 1983, Methods Enzymol. 100:468. Primer extension is effected using DNA polymerase, the product cloned, and clones containing the mutated DNA, derived by segregation of the primer extended strand, selected. Identification can be accomplished using the mutant primer as a hybridization probe. The technique is also applicable for generating multiple point mutations. See, e.g., Dalbie-McFarland et al., 1982, Proc. Natl. Acad. Sci. USA 79: 6409. PCR mutagenesis can also be used to effect the desired mutations. Random mutagenesis of selected portions of the nucleotide sequences encoding enzymatic activities can also be accomplished by several different techniques known in the art, e.g., by inserting an oligonucleotide linker randomly into a plasmid, by irradiation with X -rays or ultraviolet light, by incorporating incorrect nucleotides during in vitro DNA synthesis, by error-prone PCR mutagenesis, by preparing synthetic mutants, or by damaging plasmid DNA in vitro with chemicals. Chemical mutagens include, for example, sodium bisulfite, nitrous acid, nitrosoguanidine, hydroxylamine, agents which damage or remove bases thereby preventing normal base-pairing such as hydrazine or formic acid, analogues of nucleotide precursors such as 5 -bromouracil, 2 -aminopurine, or acrdine intercaculating agents such as proflavine, acriflavine, quinacrine, and the like. Generally, plasmid DNA or DNA fragments are treated with chemicals, transformed into E. coli and propagated as a pool or library of mutant plasmids.
[0183] In constructing a hybrid PKS of the invention, regions encoding enzymatic activity, i.e., regions encoding corresponding activities from different PKS synthases or from different locations in the same PKS, can be recovered, for example, using PCR techniques with appropriate primers. By "corresponding" activity encoding regions is meant those regions encoding the same general type of activity. For example, a KR activity encoded at one location of a gene cluster "corresponds" to a KR encoding activity in another location in the gene cluster or in a different gene cluster. Similarly, a complete reductase cycle could be considered corresponding. For example, KR/DH/ER corresponds to KR alone.
[0184] If replacement of a particular target region in a host PKS is to be made, this replacement can be conducted in vitro using suitable restriction enzymes. The replacement
can also be effected in vivo using recombinant techniques involving homologous sequences framing the replacement gene in a donor plasmid and a receptor region in a recipient plasmid. Such systems, advantageously involving plasmids of differing temperature sensitivities are described, for example, in PCT publication No. WO 96/40968, incorporated herein by reference. The vectors used to perform the various operations to replace the enzymatic activity in the host PKS genes or to support mutations in these regions of the host PKS genes can be chosen to contain control sequences operably linked to the resulting coding sequences in a manner such that expression of the coding sequences can be effected in an appropriate host.
[0185] However, simple cloning vectors may be used as well. If the cloning vectors employed to obtain PKS genes encoding derived PKS lack control sequences for expression operably linked to the encoding nucleotide sequences, the nucleotide sequences are inserted into appropriate expression vectors. This need not be done individually, but a pool of isolated encoding nucleotide sequences can be inserted into expression vectors, the resulting vectors transformed or transfected into host cells, and the resulting cells plated out into individual colonies.
[0186] The various PKS nucleotide sequences can be cloned into one or more recombinant vectors as individual cassettes, with separate control elements, or under the control of, e.g., a single promoter. The PKS subunit encoding regions can include flanking restriction sites to allow for the easy deletion and insertion of other PKS subunit encoding sequences so that hybrid PKSs can be generated. The design of such unique restriction sites is known to those of skill in the art and can be accomplished using the techniques described above, such as site-directed mutagenesis and PCR.
[0187] The expression vectors containing nucleotide sequences encoding a variety of PKS enzymes for the production of different polyketides are then transformed into the appropriate host cells to construct the library. In one straightforward approach, a mixture of such vectors is transformed into the selected host cells and the resulting cells plated into individual colonies and selected to identify successful transformants. Each individual colony has the ability to produce a particular PKS synthase and ultimately a particular polyketide. Typically, there will be duplications in some, most, or all of the colonies; the subset of the transformed colonies that contains a different PKS in each member colony can be considered the library. Alternatively, the expression vectors can be used individually to transform hosts, which transformed hosts are then assembled into a library. A variety of strategies are available to obtain a multiplicity of colonies each containing a PKS gene cluster derived from the naturally occurring host gene cluster so that each colony in the library produces a different PKS and ultimately a different polyketide. The number of different polyketides that are produced by the library is typically at least four, more typically at least ten, and preferably at least 20 , and more preferably at least 50 , reflecting similar numbers of different altered PKS gene clusters and PKS gene products. The number of members in the library is arbitrarily chosen; however, the degrees of freedom outlined above with respect to the variation of starter, extender units, stereochemistry, oxidation state, and chain length is quite large.
[0188] Methods for introducing the recombinant vectors of the invention into suitable hosts are known to those of skill in the art and typically include the use of $\mathrm{CaCl}_{2}$ or agents such as other divalent cations, lipofection, DMSO, protoplast transformation, infection, transfection, and electroporation. The polyketide producing colonies can be identified and isolated using known techniques and the produced polyketides further characterized. The polyketides produced by these colonies can be used collectively in a panel to represent a library or may be assessed individually for activity.
[0189] The libraries of the invention can thus be considered at four levels: (1) a multiplicity of colonies each with a different PKS encoding sequence; (2) colonies that contain the proteins that are members of the PKS library produced by the coding sequences; (3) the polyketides produced; and (4) antibiotics or compounds with other desired activities derived from the polyketides. Of course, combination libraries can also be constructed wherein-members of a library derived, for example, from the narbonolide PKS can be considered as a part of the same library as those derived from, for example, the rapamycin PKS or DEBS.
[0190] Colonies in the library are induced to produce the relevant synthases and thus to produce the relevant polyketides to obtain a library of polyketides. The polyketides secreted into the media can be screened for binding to desired targets, such as receptors, signaling proteins, and the like. The supernatants per se can be used for screening, or partial or complete purification of the polyketides can first be effected. Typically, such screening methods involve detecting the binding of each member of the library to receptor or other target ligand. Binding can be detected either directly or through a competiton assay. Means to screen such libraries for binding are well known in the art. Alternatively, individual polyketide members of the library can be tested against a desired target. In this event, screens wherein the biological response of the target is measured can more readily be included. Antibiotic activity can be verified using typical screening assays such as those set forth in Lehrer et al., 1991, J. Immunol. Meth. 137:167173 , incorporated herein by reference, and in the examples below.
[0191] The invention provides methods for the preparation of a large number of polyketides. These polyketides are useful intermediates in formation of compounds with antibiotic or other activity through hydroxylation and glycosylation reactions as described above. In general, the polyketide products of the PKS must be further modified, typically by hydroxylation and glycosylation, to exhibit antibiotic activity. Hydroxylation results in the novel polyketides of the invention that contain hydroxyl groups at C6, which can be accomplished using the hydroxylase encoded by the erF gene, and/or C12, which can be accomplished using the hydroxylase encoded by the picK or eryK gene. The presence of hydroxyl groups at these positions can enhance the antibiotic activity of the resulting compound relative to its unhydroxylated counterpart.
[0192] Gycosylation is important in conferring antibiotic activity to a polyketide as well. Methods for glycosylating the polyketides are generally known in the art; the glycosylation may be effected intracellularly by providing the appropriate glycosylation enzymes or may be effected in vitro using chemical synthetic means as described herein and in PCT publication No. WO 98/49315, incorporated herein by reference. Preferably, glycosylation with desos-
amine is effected in accordance with the methods of the invention in recombinant host cells provided by the invention. In general, the approaches to effecting glycosylation mirror those described above with respect to hydroxylation. The purified enzymes, isolated from native sources or recombinantly produced may be used in vitro. Alternatively and as noted, glycosylation may be effected intracellularly using endogenous or recombinantly produced intracellular glycosylases. In addition, synthetic chemical methods may be employed.
[0193] The antibiotic modular polyketides may contain any of a number of different sugars, although D-desosamine, or a close analog thereof, is most common. Erythromycin, picromycin, narbomycin and methymycin contain desosamine. Erythromycin also contains L-cladinose (34-methyl mycarose). Tylosin contains mycaminose (4-hydroxy desosamine), mycarose and 6-deoxy-D-allose. 2-acetyl-1-bromodesosamine has been used as a donor to glycosylate polyketides by Masamune et al., 1975, J. Am. Chem. Soc. 97 : 3512-3513. Other, apparently more stable donors include glycosyl fluorides, thioglycosides, and trichloroacetimidates; see Woodward et al., 1981, J. Am. Chem. Soc. 103: 3215; Martinet al., 1997, J. Am. Chem. Soc. 119: 3193; Toshima et al., 1995, J. Am. Chem. Soc. 117: 3717; Matsumoto et al., 1988, Tetrahedron Lett. 29: 3575. Glycosylation can also be effected using the polyketide aglycones as starting materials and using Saccharopolyspora erythraea or Streptomyces venezuelae to make the conversion, preferably using mutants unable to synthesize macrolides.
[0194] To provide an illustrative hybrid PKS of the invention as well as an expression vector for that hybrid PKS and host cells comprising the vector and producing the hybrid polyketide, a portion of the narbonolide PKS gene was fused to the DEBS genes. This construct also allowed the examination of whether the TE domain of the narbonolide PKS (pikTE) could promote formation of 12 -membered lactones in the context of a different PKS. A construct was generated, plasmid pKOS039-18, in which the pikTE ORF was fused with the DEBS genes in place of the DEBS TE ORF (see FIG. 5). To allow the TE to distinguish between substrates most closely resembling those generated by the narbonolide PKS, the fusion junction was chosen between the AT and ACP to eliminate ketoreductase activity in DEBS extender module 6 (KR6). This results in a hybrid PKS that presents the TE with a $\beta$-ketone heptaketide intermediate and a $\beta$-(S)-hydroxy hexaketide intermediate to cyclize, as in narbonolide and 10 -deoxymethynolide biosynthesis.
[0195] Analysis of this construct indicated the production of the 14-membered ketolide 3,6-dideoxy-3-oxo-erythronolide B (FIG. 5, compound 6). Extracts were analyzed by LC/MS. The identity of compound 6 was verified by comparison to a previously authenticated sample (see PCT publication No. 98/49315, incorporated herein by reference). The predicted 12 -membered macrolactone, ( $8 \mathrm{R}, 9 \mathrm{~S}$ )-8,9-dihydromethyl-9-hydroxy-10-deoxymethynolide (see Kao et al, 1995, J. Am. Chem. Soc. 127, incorporated herein by reference) was not detected. This result, along with others reported herein, suggests that protein interactions between the narbonolide PKS modules play a role in formation of the 12 and 14-membered macrolides.
[0196] The above example illustrates also how engineered PKSs can be improved for production of novel compounds. Compound 6 was originally produced by deletion of the KR6 domain in DEBS to create a 3-ketolide producing PKS (see U.S. patent application Ser. No. 09/073,538, filed 6 May

1998, and PCT publication No. WO 98/49315, each of which is incorporated herein by reference). Although the desired molecule was made, purification of compound 6 from this strain was hampered by the presence of 2-desmethyl ketolides that could not be easily separated. Extracts from Streptomyces lividans KS 114/pKOS039-18, however, do not contain the 2 -desmethyl compounds, greatly simplifying purification. Thus, the invention provides a useful method of producing such compounds. The ability to combine the narbonolide PKS with DEBS and other modular PKSs provides a significant advantage in the production of macrolide antibiotics.
[0197] Two other hybrid PKSs of the invention were constructed that yield this same compound. These constructs also illustrate the method of the invention in which hybrid PKSs are constructed at the protein, as opposed to the module, level. Thus, the invention provides a method for constructing a hybrid PKS which comprises the coexpression of at least one gene from a first modular PKS gene cluster in a host cell that also expresses at least one gene from a second PKS gene cluster. The invention also provides novel hybrid PKS enzymes prepared in accordance with the method. This method is not limited to hybrid PKS enzymes composed of at least one narbonolide PKS gene, although such constructs are illustrative and preferred. Moreover, the hybrid PKS enzymes are not limited to hybrids composed of unmodified proteins; as illustrated below, at least one of the genes can optionally be a hybrid PKS gene.
[0198] In the first construct, the eryAI and eryAI genes were coexpressed with picAIV and a gene encoding a hybrid extender module 5 composed of the KS and AT domains of extender module 5 of DEBS3 and the KR and ACP domains of extender module 5 of the narbonolide PKS. In the second construct, the picAIV coding sequence was fused to the hybrid extender module 5 coding sequence used in the first construct to yield a single protein. Each of these constructs produced 3-deoxy-3-oxo-6-deoxyerythronolide B. In a third construct, the coding sequence for extender module 5 of DEBS3 was fused to the picAIV coding sequence, but the levels of product produced were below the detection limits of the assay.
[0199] A variant of the first construct hybrid PKS was constructed that contained an inactivated DEBS1 extender module 1 KS domain. When host cells containing the resultant hybrid PKS were supplied the appropriate diketide precursor, the desired 13-desethyl-13-propyl compounds were obtained, as described in the examples below.
[0200] Other illustrative hybrid PKSs of the invention were made by coexpressing the picAI and picAII genes with genes encoding DEBS3 or DEBS3 variants. These constructs illustrate the method of the invention in which a hybrid PKS is produced from coexpression of PKS genes unmodified at the modular or domain level. In the first construct, the enjAIII gene was coexpressed with the picAI and picAII genes, and the hybrid PKS produced 10-desm-ethyl-10,11-anhydro-6-deoxyerythronolide B in Streptomyces lividans. Such a hybrid PKS could also be constructed in accordance with the method of the invention by transformation of $S$. venzuelae with an expression vector that produces the enyAIII gene product, DEBS3. In a preferred embodiment, the $S$. venezuelae host cell has been modified to inactivate the picAIII gene.
[0201] In the second construct, the DEBS3 gene was a variant that had an inactive KR in extender module 5. The hybrid PKS produced 5,6-dideoxy-5-oxo-10-desmethyl-10, 11-anhydroerythronolide B in Streptomyces lividans.
[0202] In the third construct, the DEBS3 gene was a variant in which the KR domain of extender module 5 was replaced by the DH and KR domains of extender module 4 of the rapamycin PKS. This construct produced 5,6-dideoxy-5-oxo-10-desmethyl-10,11-anhydroerythronolide B and 5,6-dideoxy-4,5-anhydro-10-desmethyl-10,11-anhydroerythronolide B in Streptomyces lividans, indicating that the rapamycin DH and KR domains functioned only inefficiently in this construct.
[0203] In the fourth construct, the DEBS3 gene was a variant in which the KR domain of extender module 5 was replaced by the DH, KR, and ER domains of extender module 1 of the rapamycin PKS. This construct produced 5,6-dideoxy-5-oxo-10-desmethyl-10,11-anhydroerythronolide B as well as 5,6 -dideoxy-10-desmethyl-10,11-anhydroerythronolide B in Streptomyces lividans, indicating that the rapamycin DH, KR, and ER domains functioned only inefficiently in this construct.
[0204] In the fifth construct, the DEBS3 gene was a variant in which the KR domain of extender module 6 was replaced by the DH and KR domains of extender module 4 of the rapamycin PKS. This construct produced 3,6-dideoxy-2,3-anhydro-10-desmethyl-10,11-anhydroerythronolide B in Streptomyces lividans.
[0205] In the sixth construct, the DEBS3 gene was a variant in which the AT domain of extender module 6 was replaced by the AT domain of extender module 2 of the rapamycin PKS. This construct produced 2,10 -didesmethyl-10,11-anhydro-6-deoxyerythronolide B in Streptomyces lividans.
[0206] These hybrid PKSs illustrate the wide variety of polyketides that can be produced by the methods and compounds of the invention. These polyketides are useful as antibiotics and as intermediates in the synthesis of other useful compounds, as described in the following section.

## [0207] Section VI: Compounds

[0208] The methods and recombinant DNA compounds of the invention are useful in the production of polyketides. In one important aspect, the invention provides methods for making ketolides, polyketide compounds with significant antibiotic activity. See Griesgraber et al., 1996, J. Antibiot. 49: 465-477, incorporated herein by reference. Most if not all of the ketolides prepared to date are synthesized using erythromycin A , a derivative of $6-\mathrm{dEB}$, as an intermediate. While the invention provides hybrid PKSs that produce a polyketide different in structure from $6-\mathrm{dEB}$, the invention also provides methods for making intermediates useful in preparing traditional, 6 -dEB-derived ketolide compounds.
[0209] Because 6-dEB in part differs from narbonolide in that it comprises a 10 -methyl group, the novel hybrid PKS genes of the invention based on the narbonolide PKS provide many novel ketolides that differ from the known ketolides only in that they lack a 10 -methyl group. Thus, the invention provides the 10 -desmethyl analogues of the ketolides and intermediates and precursor compounds described in, for example, Griesgraber et al., supra; Agouridas et al., 1998, J. Med. Chem. 41: 4080-4100, U.S. Pat. Nos. 5,770,579; 5,760,233; 5,750,510; 5,747,467; 5,747, 466; 5,656,607; 5,635,485; 5,614,614; 5,556,118; 5,543,

400; 5,527,780; 5,444,051; 5,439,890; 5,439,889; and PCT publication Nos. WO 98/09978 and 98/28316, each of which is incorporated herein by reference. Because the invention also provides hybrid PKS genes that include a methylma-lonyl-specific AT domain in extender module 2 of the narbonolide PKS, the invention also provides hybrid PKS that can be used to produce the 10 -methyl-containing ketolides known in the art.
[0210] Thus, a hybrid PKS of the invention that produces 10 -methyl narbonolide is constructed by substituting the malonyl-specific AT domain of the narbonolide PKS extender module 2 with a methylmalonyl specific AT domain from a heterologous PKS. A hybrid narbonolide PKS in which the AT of extender module 2 was replaced with the AT from DEBS extender module 2 was constructed using boundaries described in PCT publication No. 98/49315, incorporated herein by reference. However, when the hybrid PKS expression vector was introduced into Streptomyces venezuelae, detectable quantities of 10-methyl picromycin were not produced. Thus, to construct such a hybrid PKS of the invention, an AT domain from a module other than DEBS extender module 2 is preferred. One could also employ DEBS extender module 2 or another methylmalonyl specific AT but utilize instead different boundaries than those used for the substitution described above. In addition, one can construct such a hybrid PKS by substituting, in addition to the AT domain, additional extender module 2 domains, including the KS, the KR, and the DH, and/or additional extender module 3 domains.
[0211] Although modification of extender module 2 of the narbonolide PKS is required, the extent of hybrid modules engineered need not be limited to module 2 to make $10-\mathrm{me}-$ thyl narbonolide. For example, substitution of the KS domain of extender module 3 of the narbonolide PKS with a heterologous domain or module can result in more efficient processing of the intermediate generated by the hybrid extender module 2. Likewise, a heterologous TE domain may be more efficient in cyclizing 10 -methyl narbonolide.
[0212] Substitution of the entire extender module 2 of the narbonolide PKS with a module encoding the correct enzymatic activities, i.e., a KS, a methylmalonyl specific AT, a KR, a DH, and an ACP, can also be used to create a hybrid PKS of the invention that produces a 10 -methyl ketolide. Modules useful for such whole module replacements include extender modules 4 and 10 from the rapamycin PKS, extender modules 1 and 5 from the FK506 PKS, extender module 2 of the tylosin PKS, and extender module 4 of the rifamycin PKS. Thus, the invention provides many different hybrid PKSs that can be constructed starting from the narbonolide PKS that can be used to produce 10-methyl narbonolide. While 10 -methyl narbonolide is referred to in describing these hybrid PKSs, those of skill recognize that the invention also therefore provides the corresponding derivatives produces by glycosylation and hydroxylation. For example, if the hybrid PKS is expressed in Streptomyces narbonensis or $S$. venezuelae, the compounds produced are 10 -methyl narbomycin and picromycin, respectively. Alternatively, the PKS can be expressed in a host cell transformed with the vectors of the invention that encode the desosamine biosynthesis and desosaminyl transferase and picK hydroxylase genes.
[0213] Other important compounds provided by the invention are the 6 -hydroxy ketolides. These compounds include

3-deoxy-3-oxo erythronolide B, 6-hydroxy narbonolide, and 6 -hydroxy-10-methyl narbonolide. In the examples below, the invention provides a method for utilizing EryF to hydroxylate 3-ketolides that is applicable for the production of any 6 -hydroxy-3-ketolide.
[0214] Thus, the hybrid PKS genes of the invention can be expressed in a host cell that contains the desosamine biosynthetic genes and desosaminyl transferase gene as well as the required hydroxylase gene(s), which may be either picK (for the C12 position) or eryK (for the C12 position) and/or eryF (for the C6 position). The resulting compounds have antibiotic activity but can be further modified, as described in the patent publications referenced above, to yield a desired compound with improved or otherwise desired properties. Alternatively, the aglycone compounds can be produced in the recombinant host cell, and the desired glycosylation and hydroxylation steps carried out in vitro or in vivo, in the latter case by supplying the converting cell with the aglycone.
[0215] The compounds of the invention are thus optionally glycosylated forms of the polyketide set forth in formula (2) below which are hydroxylated at either the C6 or the C12 or both. The compounds of formula (2) can be prepared using the loading and the six extender modules of a modular PKS, modified or prepared in hybrid form as herein described. These polyketides have the formula:

[0216] including the glycosylated and isolated stereoisomeric forms thereof;
[0217] wherein $\mathrm{R}^{*}$ is a straight chain, branched or cyclic, saturated or unsaturated substituted or unsubstituted hydrocarbyl of 1-4C;
[0218] each of $\mathrm{R}^{1}-\mathrm{R}^{6}$ is independently H or alkyl (1-4C) wherein any alkyl at $\mathrm{R}^{1}$ may optionally be substituted;
[0219] each of $\mathrm{X}^{1}-\mathrm{X}^{5}$ is independently two $\mathrm{H}, \mathrm{H}$ and OH , or $=\mathrm{O}$; or
[0220] each of $\mathrm{X}^{1}-\mathrm{X}^{5}$ is independently H and the compound of formula (2) contains a double-bond in the ring adjacent to the position of said X at 2-3, 4-5, 6-7,8-9 and/or 10-11;
[0221] with the proviso that:
[0222] at least two of $\mathrm{R}^{1}-\mathrm{R}^{6}$ are alkyl (1-4C).
[0223] Preferred compounds comprising formula 2 are those wherein at least three of $R^{1}-R^{5}$ are alkyl (1-4C),
preferably methyl or ethyl; more preferably wherein at least four of $\mathrm{R}^{1}-\mathrm{R}^{5}$ are alkyl (1-4C), preferably methyl or ethyl. Also preferred are those wherein $\mathrm{X}^{2}$ is two $\mathrm{H},=\mathrm{O}$, or H and OH , and/or $\mathrm{X}^{3}$ is H , and/or $\mathrm{X}^{1}$ is OH and/or $\mathrm{X}^{5}$ is OH and/or $\mathrm{X}^{5}$ is OH . Also preferred are compounds with variable $\mathrm{R}^{*}$ when $\mathrm{R}^{1}-\mathrm{R}^{5}$ is methyl, $\mathrm{X}^{2}$ is $=\mathrm{O}$, and $\mathrm{X}^{1}, \mathrm{X}^{4}$ and $\mathrm{X}^{5}$ are OH . The glycosylated forms of the foregoing are also preferred.
[0224] The invention also provides the 12 -membered macrolides corresponding to the compounds above but produced from a narbonolide-derived PKS lacking extender modules 5 and 6 of the narbonolide PKS.
[0225] The compounds of the invention can be produced by growing and fermenting the host cells of the invention under conditions known in the art for the production of other polyketides. The compounds of the invention can be isolated from the fermentation broths of these cultured cells and purified by standard procedures. The compounds can be readily formulated to provide the pharmaceutical compositions of the invention. The pharmaceutical compositions of the invention can be used in the form of a pharmaceutical preparation, for example, in solid, semisolid, or liquid form. This preparation will contain one or more of the compounds of the invention as an active ingredient in admixture with an organic or inorganic carrier or excipient suitable for external, enteral, or parenteral application. The active ingredient may be compounded, for example, with the usual non-toxic, pharmaceutically acceptable carriers for tablets, pellets, capsules, suppositories, solutions, emulsions, suspensions, and any other form suitable for use.
[0226] The carriers which can be used include water, glucose, lactose, gum acacia, gelatin, mannitol, starch paste, magnesium trisilicate, talc, corn starch, keratin, colloidal silica, potato starch, urea, and other carriers suitable for use in manufacturing preparations, in solid, semi-solid, or liquefied form. In addition, auxiliary stabilizing, thickening, and coloring agents and perfumes may be used. For example, the compounds of the invention may be utilized with hydroxypropyl methylcellulose essentially as described in U.S. Pat. No. 4,916,138, incorporated herein by reference, or with a surfactant essentially as described in EPO patent publication No. 428,169 , incorporated herein by reference.
[0227] Oral dosage forms may be prepared essentially as described by Hondo et al., 1987, Transplantation Proceedings $X I X$, Supp. 6: 17-22, incorporated herein by reference. Dosage forms for external application may be prepared essentially as described in EPO patent publication No. 423,714 , incorporated herein by reference. The active compound is included in the pharmaceutical composition in an amount sufficient to produce the desired effect upon the disease process or condition.
[0228] For the treatment of conditions and diseases caused by infection, a compound of the invention may be administered orally, topically, parenterally, by inhalation spray, or rectally in dosage unit formulations containing conventional non-toxic pharmaceutically acceptable carriers, adjuvant, and vehicles. The term parenteral, as used herein, includes subcutaneous injections, and intravenous, intramuscular, and intrasternal injection or infusion techniques.
[0229] Dosage levels of the compounds of the invention are of the order from about 0.01 mg to about 50 mg per kilogram of body weight per day, preferably from about 0.1
mg to about 10 mg per kilogram of body weight per day. The dosage levels are useful in the treatment of the aboveindicated conditions (from about 0.7 mg to about 3.5 mg per patient per day, assuming a 70 kg patient). In addition, the compounds of the invention may be administered on an intermittent basis, i.e., at semi-weekly, weekly, semimonthly, or monthly intervals.
[0230] The amount of active ingredient that may be combined with the carrier materials to produce a single dosage form will vary depending upon the host treated and the particular mode of administration. For example, a formulation intended for oral administration to humans may contain from 0.5 mg to 5 gm of active agent compounded with an appropriate and convenient amount of carrier material, which may vary from about 5 percent to about 95 percent of the total composition. Dosage unit forms will generally contain from about 0.5 mg to about 500 mg of active ingredient. For external administration, the compounds of the invention may be formulated within the range of, for example, $0.00001 \%$ to $60 \%$ by weight, preferably from $0.001 \%$ to $10 \%$ by weight, and most preferably from about $0.005 \%$ to $0.8 \%$ by weight.
[0231] It will be understood, however, that the specific dose level for any particular patient will depend on a variety of factors. These factors include the activity of the specific compound employed; the age, body weight, general health, sex, and diet of the subject; the time and route of administration and the rate of excretion of the drug; whether a drug combination is employed in the treatment; and the severity of the particular disease or condition for which therapy is sought.
[0232] A detailed description of the invention having been provided above, the following examples are given for the purpose of illustrating the invention and shall not be construed as being a limitation on the scope of the invention or claims.

## EXAMPLE 1

## General Methodology

[0233] Bacterial strains, plasmids, and culture conditions. Streptomyces coelicolor CH999 described in WO 95/08548, published 30 Mar. 1995, or S. lividans K4-114, described in Ziermann and Betlach, Jan. 99, Recombinant Polyketide Synthesis in Streptomyces: Engineering of Improved Host Strains, BioTechniques 26:106-110, incorporated herein by reference, was used as an expression host. DNA manipulations were performed in Escherichia coli XL1-Blue, available from Stratagene. E. coli MC1061 is also suitable for use as a host for plasmid manipulation. Plasmids were passaged through E. coli ET12567 (dam dcm hsdS Cm ${ }^{\text {r }}$ ) (MacNeil, 1988, J. Bacteriol. 170: 5607, incorporated herein by reference) to generate unmethylated DNA prior to transformation of $S$. coelicolor. $E$. coli strains were grown under standard conditions. S. coelicolor strains were grown on R2YE agar plates (Hopwood et al., Genetic manipulation of Streptomyces. A laboratory manual. The John Innes Foundation: Norwich, 1985, incorporated herein by reference).
[0234] Many of the expression vectors of the invention illustrated in the examples are derived from plasmid pRM5, described in WO 95/08548, incorporated herein by reference. This plasmid includes a colEI replicon, an appropri-
ately truncated SCP2* Streptomyces replicon, two act-promoters to allow for bidirectional cloning, the gene encoding the actII-ORF4 activator which induces transcription from act promoters during the transition from growth phase to stationary phase, and appropriate marker genes. Engineered restriction sites in the plasmid facilitate the combinatorial construction of PKS gene clusters starting from cassettes encoding individual domains of naturally occurring PKSs. When plasmid pRM5 is used for expression of a PKS, all relevant biosynthetic genes can be plasmid-borne and therefore amenable to facile manipulation and mutagenesis in $E$. coli. This plasmid is also suitable for use in Streptomyces host cells. Streptomyces is genetically and physiologically well-characterized and expresses the ancillary activities required for in vivo production of most polyketides. Plasmid pRM5 utilizes the act promoter for PKS gene expression, so polyketides are produced in a secondary metabolite-like manner, thereby alleviating the toxic effects of synthesizing potentially bioactive compounds in vivo.
[0235] Manipulation of DNA and organisms. Polymerase chain reaction (PCR) was performed using Pfu polymerase (Stratagene; Taq polymerase from Perkin Elmer Cetus can also be used) under conditions recommended by the enzyme manufacturer. Standard in vitro techniques were used for DNA manipulations (Sambrook et al. Molecular Cloning: A Laboratory Manual (Current Edition)). E. coli was transformed using standard calcium chloride-based methods; a Bio-Rad E. coli pulsing apparatus and protocols provided by Bio-Rad could also be used. S. coelicolor was transformed by standard procedures (Hopwood et al. Genetic manipulation of Streptomyces. A laboratory manual. The John Innes Foundation: Norwich, 1985), and depending on what selectable marker was employed, transformants were selected using 1 mL of a $1.5 \mathrm{mg} / \mathrm{mL}$ thiostrepton overlay, 1 mL of a $2 \mathrm{mg} / \mathrm{mL}$ apramycin overlay, or both.

## EXAMPLE 2

## Cloning of the Picromycin Biosynthetic Gene Cluster from Streptomyices venezuelae

[0236] Genomic DNA ( $100 \mu \mathrm{~g}$ ) isolated from Streptomyces venezuelae ATCC15439 using standard procedures was partially digested with Sau3AI endonuclease to generate fragments -40 kbp in length. SuperCosI (Stratagene) DNA cosmid arms were prepared as directed by the manufacturer. A cosmid library was prepared by ligating $2.5 \mu \mathrm{~g}$ of the digested genomic DNA with $1.5 \mu \mathrm{~g}$ of cosmid arms in a 20 $\mu \mathrm{L}$ reaction. One microliter of the ligation mixture was propagated in E. coli XL1-Blue MR (Stratagene) using a GigapackIII XL packaging extract kit (Stratagene). The resulting library of $\sim 3000$ colonies was plated on a $10 \times 150$ mm agar plate and replicated to a nylon membrane.
[0237] The library was initially screened by direct colony hybridization with a DNA probe specific for ketosynthase domain coding sequences of PKS genes. Colonies were alkaline lysed, and the DNA was crosslinked to the membrane using UV irradiation. After overnight incubation with the probe at $42^{\circ} \mathrm{C}$., the membrane was washed twice at $25^{\circ}$ C. in $2 \times$ SSC buffer $+0.1 \%$ SDS for 15 minutes, followed by two 15 minute washes with $2 \times$ SSC buffer at $55^{\circ} \mathrm{C}$. Approximately 30 colonies gave positive hybridization signals with the degenerate probe. Several cosmids were selected and divided into two classes based on restriction digestion
patterns. A representative cosmid was selected from each class for further analysis. The representative cosmids were designated pKOS023-26 and pKOS023-27. These cosmids were determined by DNA sequencing to comprise the narbonolide PKS genes, the desosamine biosynthesis and transferase genes, the beta-glucosidase gene and the picK hydroxylase gene.
[0238] These cosmids were deposited with the American Type Culture Collection in accordance with the terms of the Budapest Treaty. Cosmid pKOS023-26 was assigned accession number ATCC 203141, and cosmid pKOS023-27 was assigned accession number ATCC 203142.
[0239] To demonstrate that the narbonolide PKS genes had been cloned and to illustrate how the invention provides methods and reagents for constructing deletion variants of narbonolide PKS genes, a narbonolide PKS gene was deleted from the chromosome of Streptomyces venezuelae. This deletion is shown schematically in FIG. 4, parts B and C. A $\sim 2.4 \mathrm{~kb}$ EcoRI-KpnI fragment and a $\sim 2.1 \mathrm{~kb}$ KpnIXhoI fragment, which together comprise both ends of the picAI gene (but lack a large portion of the coding sequence), were isolated from cosmid pKOSO23-27 and ligated together into the commercially available vector pLitmus 28 (digested with restriction enzymes EcoRI and XhoI) to give plasmid pKOS039-07. The $\sim 4.5 \mathrm{~kb}$ HindIII-SpeI fragment from plasmid pKOS039-07 was ligated with the 2.5 kb HindIII-NheI fragment of integrating vector pSET152, available from the NRRL, which contains an E. coli origin of replication and an apramycin resistance-conferring gene to create plasmid pKOS039-16. This vector was used to transform S. venezuelae, and apramycin-resistant transformants were selected.
[0240] Then, to select for double-crossover mutants, the selected transformants were grown in TSB liquid medium without antibiotics for three transfers and then plated onto non-selective media to provide single colony isolates. The isolated colonies were tested for sensitivity to apramycin, and the apramycin-sensitive colonies were then tested to determine if they produced picromycin. The tests performed included a bioassay and LC/MS analysis of the fermentation media. Colonies determined not to produce picromycin (or methymycin or neomethymycin) were then analyzed using PCR to detect an amplification product diagnostic of the deletion. A colony designated K3903 was identified, providing confirmation that the narbonolide PKS genes had been cloned. Transformation of strain K39-03 with plasmid pKOS039-27 comprising an intact picA gene under the control of the ermE* promoter from plasmid pWHM3 (see Vara et al., 1989, J. Bact. 171: 5872-5881, incorporated herein by reference) was able to restore picromycin production.
[0241] To determine that the cosmids also contained the picK hydroxylase gene, each cosmid was probed by Southern hybridization using a labeled DNA fragment amplified by PCR from the Saccharopolyspora erythraea C12-hydroxylase gene, eryK. The cosmids were digested with BamHI endonuclease and electrophoresed on a $1 \%$ agarose gel, and the resulting fragments were transferred to a nylon membrane. The membrane was incubated with the enyK probe overnight at $42^{\circ} \mathrm{C}$., washed twice at $25^{\circ} \mathrm{C}$. in $2 \times \mathrm{SSC}$ buffer with $0.1 \%$ SDS for 15 minutes, followed by two 15 minute washes with $2 \times \operatorname{SSC}$ buffer at $50^{\circ} \mathrm{C}$. Cosmid
pKOS023-26 produced an -3 kb fragment that hybridized with the probe under these conditions. This fragment was subcloned into the PCRscript ${ }^{\mathrm{TM}}$ (Stratagene) cloning vector to yield plasmid pKOS023-28 and sequenced. The $\sim 1.2 \mathrm{~kb}$ gene designated picK above was thus identified. The picK gene product is homologous to eryK and other known macrolide cytochrome P450 hydroxylases.
[0242] By such methodology, the complete set of picromycin biosynthetic genes were isolated and identified. DNA sequencing of the cloned DNA provided further confirmation that the correct genes had been cloned. In addition, and as described in the following example, the identity of the genes was confirmed by expression of narbomycin in heterologous host cells.

## EXAMPLE 3

Heterologous Expression of the Narbonolide PKS and the Picromycin Biosynthetic Gene Cluster
[0243] To provide a preferred host cell and vector for purposes of the invention, the narbonolide PKS was transferred to the non-macrolide producing host Streptomyces lividans K4-114 (see Ziermann and Betlach, 1999, Biotechniques 26, 106-110, and U.S. patent application Ser. No. $09 / 181,833$, filed 28 Oct. 1998, each of which is incorporated herein by reference). This was accomplished by replacing the three DEBS ORFs on a modified version of pCK 7 (see Kao et al., 1994, Science 265, 509-512, and U.S. Pat. No. 5,672,491, each of which is incorporated herein by reference) with all four narbonolide PKS ORFs to generate plasmid pKOS039-86 (see FIG. 5). The pCK7 derivative employed, designated pCK7‘Kan', differs from pCK7 only in that it contains a kanamycin resistance conferring gene inserted at its HindIII restriction enzyme recognition site. Because the plasmid contains two selectable markers, one can select for both markers and so minimize contamination with cells containing rearranged, undesired vectors.
[0244] Protoplasts were transformed using standard procedures and transformants selected using overlays containing antibiotics. The strains were grown in liquid R5 medium for growth/seed and production cultures at $30^{\circ} \mathrm{C}$. Transformed strains produced two compounds in similar yield ( $\sim 5-10 \mathrm{mg} / \mathrm{L}$ each). Polyketides produced in the host cells were analyzed by bioassay against Bacillus subtilis and by LC/MS analysis. Analysis of extracts by LC/MS followed by ${ }^{1} \mathrm{H}-\mathrm{NMR}$ spectroscopy of the purified compounds established their identity as narbonolide (FIG. 5, compound 4; see Kaiho et al., 1982, J. Org. Chem. 47: 1612-1614, incorporated herein by reference) and 10 -deoxymethynolide (FIG. 5, compound 5; see Lambalot et al., 1992, J. Antibiotics $45,1981-1982$, incorporated herein by reference), the respective 14 and 12-membered polyketide aglycones of YC17, narbomycin, picromycin, and methymycin.
[0245] The production of narbonolide in Streptomyces lividans represents the expression of an entire modular polyketide pathway in a heterologous host. The combined yields of compounds 4 and 5 are similar to those obtained with expression of DEBS from pCK7 (see Kao et al., 1994, Science 265: 509-512, incorporated herein by reference). Furthermore, based on the relative ratios ( $\sim 1: 1$ ) of compounds 4 And 5 produced, it is apparent that the narbonolide PKS itself possesses an inherent ability to produce both 12
and 14-membered macrolactones without the requirement of additional activities unique to $S$. venezuelae. Although the existence of a complementary enzyme present in S. lividans that provides this function is possible, it would be unusual to find such a specific enzyme in an organism that does not produce any known macrolide.
[0246] To provide a heterologous host cell of the invention that produces the narbonolide PKS and the picB gene, the picB gene was integrated into the chromosome of Streptomyces lividans harboring plasmid $\mathrm{pKOS} 039-86$ to yield $S$. lividans K39-18/pKOS039-86. To provide the integrating vector utilized, the picB gene was cloned into the Streptomyces genome integrating vector pSET152 (see Bierman et al., 1992, Gene 116, 43, incorporated herein by reference) under control of the same promoter (Pact1) as the PKS on plasmid pKOS039-86.
[0247] A comparison of strains K39-18/pKOS39-86 and K4-114/pKOS039-86 grown under identical conditions indicated that the strain containing TEII produced 47 times more total polyketide. Each strain was grown in 30 mL of R 5 (see Hopwood et al., Genetic Manipulation of Streptomyces: A Laboratory Manual; John Innes Foundation: Norwich, UK, 1985, incorporated herein by reference) liquid (with 20 $\mu \mathrm{g} / \mathrm{mL}$ thiostrepton) at $30^{\circ} \mathrm{C}$. for 9 days. The fermentation broth was analyzed directly by reverse phase HPLC. Absorbance at 235 nm was used to monitor compounds and measure relative abundance. This increased production indicates that the enzyme is functional in this strain. As noted above, because the production levels of compound 4 and 5 from K39-18/pKOS03986 increased by the same relative amounts, TEII does not appear to influence the ratio of 12 and 14 -membered lactone ring formation.
[0248] To express the glycosylated counterparts of narbonolide (narbomycin) and 10 -deoxymethynolide (YC17) in heterologous host cells, the desosamine biosynthetic genes and desosaminyl transferase gene were transformed into the host cells harboring plasmid pKOS039-86 (and, optionally, the picB gene, which can be integrated into the chromosome as described above).
[0249] Plasmid pKOS039-104, see FIG. 6, comprises the desosamine biosynthetic genes, the beta-glucosidase gene, and the desosaminyl transferase gene. This plasmid was constructed by first inserting a polylinker oligonucleotide, containing a restriction enzyme recognition site for Pacl, a Shine-Dalgarno sequence, and restriction enzyme recognition sites for NdeI, BgIII, and HindIII, into a pUC19 derivative, called pKOS2447, to yield plasmid pKOS03998.
[0250] An $\sim 0.3 \mathrm{~kb}$ PCR fragment comprising the coding sequence for the N -terminus of the desI gene product and an $\sim 0.12 \mathrm{~kb} \mathrm{PCR}$ fragment comprising the coding sequence for the C-terminus of the desR gene product were amplified from cosmid pKOS23-26 (ATCC 203141) and inserted together into pLitmus 28 treated with restriction enzymes NsiI and EcoRI to produce plasmid pKOS039-101. The $\sim 6$ kb SphI-PstI restriction fragment of pKOS23-26 containing the desI, desII, desIII, desIV, and desV genes was inserted into plasmid pUC19 (Stratagene) to yield plasmid pKOS039-102. The $\sim 6 \mathrm{~kb}$ SphI-EcoRI restriction fragment from plasmid $\mathrm{pKOS} 039-102$ was inserted into pKOS 039 101 to produce plasmid pKOS039-103. The $\sim 6 \mathrm{~kb}$ BgIII-PstI fragment from pKOS23-26 that contains the desR, desVI,
desVII, and desVIII genes was inserted into pKOS39-98 to yield pKOS39-100. The $\sim 6 \mathrm{~kb}$ PacI-PstI restriction fragment of pKOS39-100 and the 6.4 kb NsiI-EcoRI fragment of pKOS39-103 were cloned into pKOS39-44 to yield pKOS39-104.
[0251] When introduced into Streptomyces lividans host cells comprising the recombinant narbonolide PKS of the invention, plasmid pKOS39-104 drives expression of the desosamine biosynthetic genes, the beta-glucosidase gene, and the desosaminyl transferase gene. The glycosylated antibiotic narbomycin was produced in these host cells, and it is believed that YC17 was produced as well. When these host cells are transformed with vectors that drive expression of the picK gene, the antibiotics methymycin, neomethymycin, and picromycin are produced.
[0252] In similar fashion, when plasmid pKOS039-18, which encodes a hybrid PKS of the invention that produces 3-deoxy-3-oxo-6-deoxyerythronolide B was expressed in Streptomyces lividans host cells transformed with plasmid pKOS39-104, the 5-desosaminylated analog was produced. Likewise, when plasmid pCK7, which encodes DEBS, which produces 6 -deoxyerythronolide $B$, was expressed in Streptomyces lividans host cells transformed with plasmid pKC639-104, the 5-desosaminylated analog was produced. These compounds have antibiotic activity and are useful as intermediates in the synthesis of other antibiotics.

## EXAMPLE 4

## Expression Vector for Desosaminyl Transferase

[0253] While the invention provides expression vectors comprising all of the genes required for desosamine biosynthesis and transfer to a polyketide, the invention also provides expression vectors that encode any subset of those genes or any single gene. As one illustrative example, the invention provides an expression vector for desosaminyl transferase. This vector is useful to desosaminylate polyketides in host cells that produce NDP-desosamine but lack a desosaminyl transferase gene or express a desosaminyl transferase that does not function as efficiently on the polyketide of interest as does the desosaminyl transferase of Streptomyces venezuelae. This expression vector was constructed by first amplifying the desosaminyl transferase coding sequence from $\mathrm{pKOSO} 03-27$ using the primers:

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N3917: 5'-CCCTGCAGCGGCAAGGAAGGACACGACGCCA-3';
and
N3918: 5'-AGGTCTAGAGCTCAGTGCCGGGCGTCGGCCGG-3',
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[0254] to give a 1.5 kb product. This product was then treated with restriction enzymes PstI and XbaI and ligated with HindIII and XbaI digested plasmid pKOS039-06 together with the 7.6 kb PstI-HindIII restriction fragment of plasmid pWHM1104 to provide plasmid pKOS039-14. Plasmid pWHM1104, described in Tang et al., 1996, Molec. Microbiol. 22(5): 801-813, incorporated herein by reference, encodes the ermE* promoter. Plasmid pKOS039-14 is constructed so that the desosaminyl transferase gene is placed under the control of the ermE* promoter and is suitable for expression of the desosaminyl transferase in Streptomyces, Saccharopolyspora erythraea, and other host cells in which the ermE* promoter functions.

## EXAMPLE 5

> Heterologous Expression of the picK Gene Product in E. coli
[0255] The picK gene was PCR amplified from plasmid pKOS023-28 using the oligonucleotide primers:

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N024-36B (forward):
5'-TTGCATGCATATGCGCCGTACCCAGCAGGGAACGACC;
and
N024-37B (reverse):
5'-TTGAATTCTCAACTAGTACGGCGGCCCGCCTCCCGTCC.
```

[0256] These primers alter the Streptomyces GTG start codon to ATG and introduce a SpeI site at the C-terminal end of the gene, resulting in the substitution of a serine for the terminal glycine amino acid residue. The blunt-ended PCR product was subcloned into the commercially available vector pCRscript at the SrfI site to yield plasmid $\mathrm{pKOS} 023-$ 60 . An $\sim 1.3 \mathrm{~kb}$ NdeI-Xhol fragment was then inserted into the NdeI/XhoI sites of the T7 expression vector pET22b (Novagen, Madison, Wis.) to generate pKOS023-61. Plasmid pKOS023-61 was digested with restriction enzymes SpeI and EcoRI, and a short linker fragment encoding 6 histidine residues and a stop codon (composed of oligonucleotides 30-85a: 5'-CTAGTATGCATCATCATCATCAT-CATTAA-3'; and 30-85b: 5'-AATTTTAATGATGATGAT-GATGATGCATA-3') was inserted to obtain plasmid pKOS023-68. Both plasmid pKOS023-61 and pKOS023-68 produced active PicK enzyme in recombinant $E$. coli host cells.
[0257] Plasmid pKOS023-61 was transformed into E. coli BL21-DE3. Successful transformants were grown in LBcontaining carbenicillin $(100 \mu \mathrm{~g} / \mathrm{ml})$ at $37^{\circ} \mathrm{C}$. to an $\mathrm{OD}_{600}$ of 0.6. Isopropyl-beta-D-thiogalactopyranoside (G) was added to a final concentration of 1 mM , and the cells were grown for an additional 3 hours before harvesting. The cells were collected by centrifugation and frozen at $-80^{\circ} \mathrm{C}$. A control culture of BL21-DE3 containing the vector plasmid pET21c (Invitrogen) was prepared in parallel.
[0258] The frozen BL21-DE3/pKOS023-61 cells were thawed, suspended in $2 \mu \mathrm{~L}$ of cold cell disruption buffer ( 5 mM imidazole, $500 \mathrm{mM} \mathrm{NaCl}, 20 \mathrm{mM}$ Tris/ $\mathrm{HCl}, \mathrm{pH} 8.0$ ) and sonicated to facilitate lysis. Cellular debris and supernatant were separated by centrifugation and subjected to SDSPAGE on $10-15 \%$ gradient gels, with Coomassie Blue staining, using a Pharmacia Phast Gel Electrophoresis system. The soluble crude extract from BL21-DE3/pKOS02361 contained a Coomassie stained band of $\mathrm{M}_{\mathrm{r}} \sim 46 \mathrm{kDa}$, which was absent in the control strain BL21-DE3/pET21c.
[0259] The hydroxylase activity of the picK protein was assayed as follows. The crude supernatant ( $20 \mu \mathrm{~L}$ ) was added to a reaction mixture ( $100 \mu \mathrm{~L}$ total volume) containing 50 mM Tris $/ \mathrm{HCl}(\mathrm{pH} 7.5$ ), $20 \mu \mathrm{M}$ spinach ferredoxin, 0.025 Unit of spinach ferredoxin: $\mathrm{NADP}^{+}$oxidoreductase, 0.8 Unit of glucose-6-phosphate dehydrogenase, $1.4 \mathrm{mM} \mathrm{NADP}{ }^{+}$, 7.6 mM glucose- 6 phosphate, and 20 mmol of narbomycin. The narbomycin was purified from a culture of Streptomyces narbonensis, and upon LC/MS analysis gave a single peak of $[\mathrm{M}+\mathrm{H}]^{+}=510$. The reaction was allowed to proceed for 105 minutes at $30^{\circ} \mathrm{C}$. Half of the reaction mixture was
loaded onto an HPLC, and the effluent was analyzed by evaporative light scattering (ELSD) and mass spectrometry. The control extract (BL21-DE3/pET21c) was processed identically. The BL21-DE3/pKOS023-61 reaction contained a compound not present in the control having the same retention time, molecular weight and mass fragmentation pattern as picromycin $\left([\mathrm{M}+\mathrm{H}]^{+}=526\right)$. The conversion of narbomycin to picromycin under these conditions was estimated to be greater than $90 \%$ by ELSD peak area.
[0260] The poly-histidine-linked PicK hydroxylase was prepared from pKOS023-68 transformed into E. coli BL21 (DE3) and cultured as described above. The cells were harvested and the PicK protein purified as follows. All purification steps were performed at $4^{\circ}$ C. E. coli cell pellets were suspended in $32 \mu \mathrm{~L}$ of cold binding buffer ( 20 mM Tris/ $\mathrm{HCl}, \mathrm{pH} 8.0,5 \mathrm{mM}$ imidazole, 500 mM NaCl ) per mL of culture and lysed by sonication. For analysis of $E$. coli cell-free extracts, the cellular debris was removed by lowspeed centrifugation, and the supernatant was used directly in assays. For purification of PicK/6-His, the supernatant was loaded ( $0.5 \mathrm{~mL} / \mathrm{min}$.) onto a 5 mL HiTrap Chelating column (Pharmacia, Piscataway, N.J.), equilibrated with binding buffer. The column was washed with $25 \mu \mathrm{~L}$ of binding buffer and the protein was eluted with a $35 \mu \mathrm{~L}$ linear gradient ( $5-500 \mathrm{mM}$ imidazole in binding buffer). Column effluent was monitored at 280 nm and 416 nm . Fractions corresponding to the 416 nm absorbance peak were pooled and dialyzed against storage buffer ( 45 mM Tris/ $/ \mathrm{HCl}, \mathrm{pH}$ $7.5,0.1 \mathrm{mM}$ EDTA, 0.2 mM DTT, $10 \%$ glycerol). The purified 46 kDa protein was analyzed by SDSPAGE using Coomassie blue staining, and enzyme concentration and yield were determined.
[0261] Narbomycin was purified as described above from a culture of Streptomyces narbonensis ATCC19790. Reactions for kinetic assays ( $100 \mu \mathrm{~L}$ ) consisted of 50 mM Tris $/ \mathrm{HCl}(\mathrm{pH} 7.5), 100 \mu \mathrm{M}$ spinach ferredoxin, 0.025 Unit of spinach ferredoxin:NADP ${ }^{+}$oxidoreductase, 0.8 U glucose-6-phosphate dehydrogenase, 1.4 mM NADP +7.6 mM glucose-6-phosphate, $20-500 \mu \mathrm{M}$ narbomycin substrate, and $50-500 \mathrm{nM}$ of PicK enzyme. The reaction proceeded at $30^{\circ}$ C., and samples were withdrawn for analysis at $5,10,15$, and 90 minutes.
[0262] Reactions were stopped by heating to $100^{\circ} \mathrm{C}$., for 1 minute, and denatured protein was removed by centrifugation. Depletion of narbomycin and formation of picromycin were determined by high performance liquid chromatography (HPLC, Beckman C-18 $0.46 \times 15 \mathrm{~cm}$ column) coupled to atmospheric pressure chemical ionization (APCI) mass spectroscopic detection (Perkin Elmer/Sciex API 100) and evaporative light scattering detection (Alltech 500 ELSD).

## EXAMPLE 6

Expression of the picK Gene Encoding the Hydroxalase in Streptomyces narbonensis
[0263] To produce picromycin in Streptomyces narbonensis, a host that produces narbomycin but not picromycin, the methods and vectors of the invention were used to express the picK gene in this host.
[0264] The picK gene was amplified from cosmid pKOS023-26 using the primers:

```
N3903: 5'-TCCTCTAGACGTTTCCGT-3';
and
N3904: 5'-TGAAGCTTGAATTCAACCGGT-3'
```

[0265] to obtain an $\sim 1.3 \mathrm{~kb}$ product. The product was treated with restriction enzymes XbaI and HindIII and ligated with the $7.6 \mathrm{~kb} \mathrm{XbaI}-\mathrm{HindIII}$ restriction fragment of plasmid pWHM1104 to provide plasmid pKOS039-01, placing the picK gene under the control of the ermE* promoter The resulting plasmid was transformed into purified stocks of $S$. narbonensis by protoplast fusion and electroporation. The transformants were grown in suitable media and shown to convert narbomycin to picromycin at a yield of over $95 \%$.

## EXAMPLE 7

## Construction of a Hybrid DEBS/Narbonolide PKS

[0266] This example describes the construction of illustrative hybrid PKS expression vectors of the invention. The hybrid PKS contains portions of the narbonolide PKS and portions of rapamycin and/or DEBS PKS. In the first constructs, pKOS039-18 and pKOS039-19 the hybrid PKS comprises the narbonolide PKS extender module 6 ACP and thioesterase domains and the DEBS loading module and extender modules $1-5$ as well as the KS and AT domains of DEBS extender module 6 (but not the KR domain of extender module 6). In pKOS039-19, the hybrid PKS is identical except that the KS1 domain is inactivated, i.e., the ketosynthase in extender module 1 is disabled. The inactive DEBS KS1 domain and its construction are described in detail in PCT publication Nos. WO 97/02358 and 99/03986, each of which is incorporated herein by reference. To construct pKOS039-18, the 2.33 kb BamHI-EcoRI fragment of pKOS023-27, which contains the desired sequence, was amplified by PCR and subcloned into plasmid pUC9. The primers used in the PCR were:

$$
\begin{aligned}
& \text { N3905: } \\
& \text { and } \\
& \text { N3906: }
\end{aligned} \text { (5TTATGCATCCCGCGGGTCCCGGCGAG-3'; } \quad \text { (TCAGAATTCTGTCGGTCACTTGCCCGC-3'. }
$$

[0267] The 1.6 kb PCR product was digested with PstI and EcoRI and cloned into the corresponding sites of plasmid pKOS015-52 (this plasmid contains the relevant portions of the coding sequence for the DEBS extender module 6) and commercially available plasmid pLitmus 28 to provide plasmids pKOS039-12 and pKOS039-13, respectively. The BgIII-EcoRI fragment of plasmid pKOS039-12 was cloned into plasmid pKOS011-77, which contains the functional DEBS gene cluster and into plasmid pJRJ2, which contains the mutated DEBS gene that produces a DEBS PKS in which the KS domain of extender module I has been rendered inactive. Plasmid pJRJ2 is described in PCI publication Nos. 99/03986 and 97/02358, incorporated herein by reference.
[0268] Plasmids pKOS039-18 and pKOS039-19, respectively, were obtained. These two plasmids were transformed into Streptomyces coelicolor CH999 by protoplast fusion.

The resulting cells were cultured under conditions such that expression of the PKS occurred. Cells transformed with plasmid pKOS039-18 produced the expected product 3-deoxy-3-oxo-6-deoxyerythronolide B. When cells transformed with plasmid pKOS039-19 were provided (2S,3R)-2-methyl-3-hydroxyhexanoate NACS, 13-desethyl-13-pro-pyl-3-deoxy-3-oxo-6-deoxyerythronolide B was produced.

## EXAMPLE 8

## 6-Hydroxylation of 3,6-dideoxy-3-oxoerythronolide B Using the eryF Hydroxylase

[0269] Certain compounds of the invention can be hydroxylated at the C6 position in a host cell that expresses the eryF gene. These compounds can also be hydroxylated in vitro, as illustrated by this example.
[0270] The 6-hydroxylase encoded by eryF was expressed in E. coli, and partially purified. The hydroxylase ( 100 pmol in $10 \mu \mathrm{~L}$ ) was added to a reaction mixture ( $100 \mu \mathrm{l}$ total volume) containing 50 mM Tris $/ \mathrm{HCl}(\mathrm{pH} 7.5), 20 \mu \mathrm{M}$ spinach ferredoxin, 0.025 Unit of spinach ferredoxin:NADP ${ }^{+}$oxidoreductase, 0.8 Unit of glucose-6-phosphate dehydrogenase, $1.4 \mathrm{mM} \mathrm{NADP}^{+}$, 7.6 mM glucose-6-phosphate, and 10 nmol 6 -deoxyerythronolide $B$. The reaction was allowed to proceed for 90 minutes at $30^{\circ} \mathrm{C}$. Half of the reaction mixture was loaded onto an HPLC, and the effluent was analyzed by mass spectrometry. The production of erythronolide B as evidenced by a new peak eluting earlier in the gradient and showing $[\mathrm{M}+\mathrm{H}]^{+}=401$. Conversion was estimated at $50 \%$ based on relative total ion counts.
[0271] Those of skill in the art will recognize the potential for hemiketal formation in the above compound and compounds of similar structure. To reduce the amount of hemiketal formed, one can use more basic (as opposed to acidic) conditions or employ sterically hindered derivative compounds, such as 5 -desosaminylated compounds.

## EXAMPLE 9

## Measurement of Antibacterial Activity

[0272] Antibacterial activity was determined using either disk diffusion assays with Bacillus cereus as the test organism or by measurement of minimum inhibitory concentrations (MIC) in liquid culture against sensitive and resistant strains of Staphylococcus pneumoniae.

## Example 10

## Construction of Desosamine Containing Polyketide <br> Libraries Using a Glycosyltransferase with Broad Substrate Specificity

[0273] Desosamine is an important deoxyaminosugar present on a number of structurally related macrolide antibiotics such as erythromycin and is the only glycoside present on picromycin, methymycin, and the highly potent semisynthetic ketolides. In this example, a set of nine deoxysugar biosynthetic and auxiliary genes from the picromycin/methymycin (pik) cluster was integrated in the chromosome of Streptomyces lividans to create a host that synthesizes TDP-D-desosamine and can be used in combination with PKS expression plasmids to generate libraries of desosaminylated polyketides. The versatility of the DesVII
desosaminyltransferase is demonstrated by formation of desosaminylated macrolides from more than twenty different 14 -membered lactones. The attachment of desosamine is sufficient to confer antibiotic activity to each of the otherwise inactive aglycones, reinforcing the belief that this sugar plays a critical role in the molecular binding properties of erythromycin and related macrolides. This host and others that can be engineered to produce deoxysugar and polyketide tailoring pathways in accordance with the methods of the invention are valuable tools for expanding the size and diversity of polyketides that can be generated by combinatorial biosynthesis. References cited in this example are indicated by a reference number; the numbered list of references is located at the end of this example. All references cited are incorporated herein by reference.
[0274] Much of the structural diversity and complexity among polyketides can be attributed to the chemistry performed by PKSs (1), and the modular architecture of catalytic domains within PKSs has been exploited by different rational and combinatorial engineering approaches to create polyketide diversity (24). However, structural variability among polyketides can also result from post-PKS biosynthetic steps, including oxidation and/or glycosylation with unique deoxy and amino sugars. Such modifications are often necessary to impart or enhance the specific biological activity of the molecule. For example, erythromycin A contains two deoxysugar moieties, L-cladinose and D-desosamine, that are required for antibacterial activity and the absence of either carbohydrate results in loss of potency. Although some chemical modifications to erythromycin have been discovered that can ameliorate the loss of the cladinose residue (5-7), there has been no substitution found for desosamine. This important deoxyaminosugar is also present in other macrolide antibiotics, such as oleandomycin and megalomicin, and is the only glycoside necessary to confer antibacterial activity to picromycin, methymycin, and the semisynthetic ketolide pharmacophores.
[0275] Polyketide libraries generated by genetic modification of macrolide PKSs in which enzymatic domains and entire protein subunits were removed, added, or exchanged in various combinations have been produced ( $3,4,8$ ). Because these libraries were constructed in heterologous hosts lacking glycosylation pathways, only the corresponding aglycones were produced. The methods and reagents of the present invention can be used to expand the capabilities of the combinatorial biosynthesis strategies described to incorporate post-PKS tailoring steps, in particular the addition of deoxysugar components.
[0276] Some experiments have been performed in which structurally modified macrolactones are subsequently glycosylated in their native hosts (9-13), and also in bioconversion experiments in which a modified aglycone is fed to a PKS blocked mutant strain (14). These experiments indicate that glycosyltransferases are able to accept polyketide substrates with some amount of structural alteration. However, neither of these approaches is well-suited for the production and biological screening of large numbers of compounds, because most polyketide host organisms are difficult to manipulate genetically and the bioconversion of aglycones requires a tedious initial purification step.
[0277] A more practical approach is the heterologous expression of deoxysugar biosynthetic pathways in hosts
that have been developed for library expression. Although the effort to clone entire deoxysugar biosynthetic pathways in a heterologous organism can be a significant initial investment (most deoxysugars require six or more enzymatic steps whose genes are typically scattered within a polyketide gene cluster), these expression vectors, once made, can be easily combined with those containing PKSs to engineer glycosylated libraries rapidly. Olano et al. recently utilized a two-plasmid system to produce L-daunosamine, the deoxyaminosugar of daunorubicin and doxorubicin, in Streptomyces lividans (15).
[0278] Here we report the development of a single expression vector for the production of desosaminylated macrolides in Streptomyces. Desosamine was selected as the sugar constituent, because it was believed that addition of this single deoxysugar would be sufficient to confer antibacterial activity upon macrolactones to which it was attached. The expression vector was combined with a library of existing PMS expression plasmids to produce several novel glycosylated macrolide compounds in S. lividans, providing the first examples in which both polyketide and deoxysugar pathways have been placed in a single heterologous host.

## [0279] A. Material and Methods

[0280] (i) Strains, Culture Conditions, and DNA Manipulation
[0281] DNA manipulation was performed in Escherichia coli XL1-Blue (Stratagene) using standard protocols (16). Bacillis subtilis was grown in LB at $37^{\circ} \mathrm{C}$. PCR was performed with Pfu polymerase (Stratagene) under conditions recommended by the manufacturer. S. lividans K4-114 (17) was used as the host for expression of engineered PKS and desosamine genes. $S$. lividans strains were maintained on R2YE agar plates (18) with appropriate antibiotic selection. S. lividans protoplasts were transformed by the standard procedure (18) and transformants were selected using 1 ml of a $1 \mathrm{mg} / \mathrm{ml}$ thiostrepton and/or 1 ml of a $2 \mathrm{mg} / \mathrm{ml}$ apramycin overlay on R2YE regeneration plates.
[0282] (ii) Construction of Expression Plasmids
[0283] Expression plasmid pKOS39-104 was constructed as follows. The $6.0 \mathrm{~kb} \mathrm{Bgl} \mathrm{II}-\mathrm{Pst}$ I fragment containing the picromycin des VIII, des VII, desVI and desR (partial) genes from cosmid pKOS23-26 (19) was subcloned into the Bg 1 II-Pst I sites of pKOS39-98, a pUC19 derivative with a redesigned multiple cloning site. The resulting plasmid, pKOS39-100, contains a Pac I site upstream of the Bgl II site which is used in a later cloning step. The 6 kb Sph I-Pst I fragment containing the desI (partial), desII, desIII, desIV and des V genes from pKOS23-26 was subcloned into the Sph I-Pst I of pUC19 to make pKOS39-102. The remaining $3^{\prime}$-end of the desR gene and $5^{\prime}$-end of the desI gene were PCR amplified from cosmid $\mathrm{pKOS} 23-26$ with the following oligonucleotides (restriction sites shown in italics):
desR gene:

forward | '-AGATGCATTTCTGGGATGCCGCCACGGA; |
| :--- |
| and |
| reverse |
| 5'-CGTCTAGACGTCACCAGACGTTGACCGTG; |
| desI gene: |

```
            -continued
forward 5'-TTTCTAGACGGTGGCCCGGAGGGAACATC;
and
reverse 5'-CGGAATTCCGCAGCTGGTCGGCGGCGCA.
```

[0284] The two PCR fragments were digested with Nsi I-Xba I and Xba I-EcoR I, respectively, and ligated with Nsi I-EcoR I digested Litmus 28 (New England Biolabs) to obtain pKOS39-101B. The 6 kb Sph I-EcoR I fragment of pKOS39-102 was inserted into pKOS39-101B to make pKOS39-103. The 6.4 kb Nsi I-EcoR I fragment of pKOS39-103 and the 6 kb Pac I-Pst I fragment of pKOS39100 were then ligated together with the 8.5 kb Pac I-EcoR I fragment of pKOS39-44 (20), yielding the final expression plasmid pKOS39-104. A restriction site and function map of this plasmid is shown below.

[0285] (iii) Production and Analysis of Compounds
[0286] All stains were grown in 5 ml liquid R2YE medium at $30^{\circ} \mathrm{C}$. and analyzed following 5 days growth. For bioconversion experiments, aglycones ( $\sim 10 \mathrm{mg}$ /liter) were fed at the start of fermentation. Fermentation broth was analyzed directly by liquid chromatography/mass spectrometry (LC/MS) and evaporative light scattering detection (ELSD) as previously described (20). An authentic sample of narbomycin prepared from Streptomyces narbonensis (19) was used to validate production of this compound. For LC/MS analysis of strains containing PKS expression plasmids the cultures were extracted twice with 5 ml of ethyl acetate/triethylamine (99:1), concentrated to dryness and resuspended in 0.5 ml of acetonitrile.

## [0287] (iv) Antibacterial Assays

[0288] Extracts prepared from the culture broths as above were assayed for biological activity against. B. subtilis using an agar plate diffusion method (see Example 9). Samples (5 $\mu 1)$ from each of the extracts were pipetted to sterile filter disks, dried, and placed on an LB plate spread with $20 \mu$ of an overnight culture of $B$. subtilis. The plates were incubated overnight at $37^{\circ} \mathrm{C}$. to visualize zones of growth inhibition.
[0289] B. Results
[0290] (i) Construction and Validation of a Desosamine Expression System
[0291] The picromycin/methymycin (pik) gene cluster from Streptomyces venezuelae (21) was chosen as the source of desosamine biosynthetic genes rather than other available clusters (i.e. erythromycin, oleandomycin, or megalomicin) for several reasons. First, all of the genes required for biosynthesis of TDP desosamine from glucose-1-phosphate, a primary metabolite, as well as the desosaminyl transferase are present in the pik cluster whereas one or more of the genes are missing or not yet identified in each of the other clusters. Second, the genes from the pik cluster are comprised in a single contiguous segment of DNA (the des cluster), compared to those in other clusters which are dispersed among other genes, facilitating cloning and plasmid construction. The organization of these genes in the picromycin biosynthetic gene cluster is shown below, followed by the depiction of the biosynthetic pathway.
mycin, picromycin, and oleandomycin, causes inactivation and provides self-resistance to these compounds which are reactivated by a 13 -glucosidase upon export $(24,25)$. $S$. lividans is known to possess at least two such glucosyltransferases which inactivate erythromycin and picromycin by the same mechanism (26). Therefore, it was important to include this gene for expression in $S$. lividans to produce desosaminylated compounds without the glucose modification.
[0294] The expression system used here was adopted from the multi-vector system developed for separate expression of erythromycin PKS, or 6-deoxyerythronolide B synthase (DEBS), subunits in Streptomyces (4, 27; see also U.S. Pat. No. $6,033,883$ ). Plasmid pKOS39-104 contains the des genes cloned in a single orientation under control of the actI promoter and actII-44 activator. Since pKOS39-104 is a derivative of pSET152 (28), it contains the phiC31-int-attP loci for chromosomal integration in Streptomyces and can be used in conjunction with the pRM5-based PKS expression

[0292] Third, the natural substrates for the desosaminyl transferase from the pik gene cluster, narbonolide and 10 -deoxymethynolide, are themselves aglycones; in each of the other cases, desosamine is attached subsequent to addition of at least one other sugar. Furthermore, the difference in macrolactone ring sizes between narbonolide and 10 -deoxymethynolide ( 14 and 12 atoms, respectively) suggests that the desosaminyl transferase from this cluster is somewhat forgiving towards its polyketide substrate.
[0293] Seven genes in the des cluster, desI, desII, desIII, desIV, desV, desVI, and des VIII, are presumed to be responsible for the biosynthesis of TDP-D-desosamine (22). Also present is the des VII gene encoding the glycosyltransferase. In addition to catalyzing the transfer of desosamine to both 12- and 14 -membered macrolactones, it has been shown that DesVII is able to incorporate non-natural deoxysugar substrates $(22,23)$. The desR gene encodes a $\beta$-glucosidase that removes a glucose residue attached to the C-2' hydroxyl of desosamine (24). It is believed that the glucosylation of desosamine containing macrolides like methy-
plasmid library ( 3 ; see also U.S. Pat. No. 5,672,491). S. lividans K4114 was transformed with pKOS39-104 and designated K39-22. Confirmation that this strain produced TDP-D-desosamine was performed by feeding aglycones to the strain and looking for the presence of desosaminylated compounds by LC/MS analysis.
[0295] Four aglycones ( $10 \mathrm{mg} /$ liter each) were fed to liquid fermentations of $S$. lividans K39-22: narbonolide and 10 -deoxymethynolide, the natural substrates for DesVII, 3-keto-6-deoxyerythronolide B (dEB), and 6-dEB. Fermentation broth from all four aglycone fed strains displayed antibacterial activity against $B$. subtilis whereas $S$. lividans K39-22 alone produced no detectable activity. LC/MS analysis demonstrated that each of the corresponding desosaminylated compounds narbomycin, 10 -deoxymethymycin (YC17), 3-keto-5-O-desosaminyl-6-dEB, and 5-O-des-osaminyl- 6 -dEB were produced. In each case, the parent ion $\left(\mathrm{M}+\mathrm{H}^{+}\right)$of the expected compound was detected in addition to a characteristic ion at 158 amu produced by the desosamine fragment. Production of narbomycin in the narbono-
lide fed strain was further confirmed by comparison to authentic narbomycin obtained from S. narbonensis. LC/MS also revealed that a significant amount $(50-90 \%)$ of the aglycone remained unconverted in each of the samples.
[0296] These results established that the des expression vector was functional and that the DesVII glycosyltransferase was able to glycosylate non-natural macrolactone substrates. The bioassay results also confirmed that desosamine is sufficient to confer antibacterial activity to these macrolactones. There were no $2^{\prime}$-O-glucosyl derivatives detected, which indicates that the DesR glucosidase included in pKOS39-104 was also operational, although minor glucosylated products were putatively found in subsequent experiments with the strain (see below).
[0297] (ii) Co-Expression of Desosamine and Aglycone Pathways in $S$. lividans.
[0298] Although expression of both a modular polyketide pathway and a deoxysugar pathway together in a heterologous host has not been reported, the bioconversion results suggested that transformation of S. lividans K39-22 with plasmids encoding macrolide PKSs would lead to production of desosaminylated compounds. Plasmids encoding the PKSs that, in S. lividans, produce the same four aglycones used in the bioconversion studies were therefore transformed into $S$. lividans K39-22. Plasmid pKOS39-86 contains the picromycin/methymycin PKS and produces both narbonolide and 10 -deoxymethynolide (20). Plasmid pKAO127 contains DEBS and produces 6-dEB (17). Plasmid pKOS39-18 contains DEBS with a modified terminal module that produces 3-keto $\mathrm{dB}(20)$.
[0299] Culture broth from each of the transformed strains displayed activity against $B$. subtilis. LC/MS analysis as above confirmed the presence of each of the expected desosaminylated compounds as well as their aglycone precursors and minor amounts of the corresponding $2^{2}-\mathrm{O}$ glucosyl derivatives. The total yield of narbomycin and 10 -deoxymethymycin in S. lividans K39-22/pKOS3986 was approximately $1 \mathrm{mg} /$ liter each and represents about a $20 \%$ conversion of the total aglycone produced. Thus, although both PKS and deoxysugar pathways function as expected, complete glycosylation of even the natural substrates for DesVII did not occur under these conditions. S. lividans K39-22 contains a copy of the ermE macrolide resistance gene, and no obvious growth defects were observed with production of the biologically active compounds. These results suggest that a limiting amount of TDP-desosamine is being produced by the strain under these conditions.
[0300] (iii) Production and Biological Screening of a Glycosylated Macrolide Library
[0301] Over 50 PKS expression plasmids have been constructed and tested in using DEBS and other macrolide PKS genes ( $3,8,20$ ). These PKSs produce a variety of 14 -membered macrolactones in which single or multiple carbon centers have been altered. Each plasmid contains the same pRM5-based vector as above, providing a convenient opportunity to expand and diversify any existing aglycone library by routine transformation of S. lividans K39-22. Because a C-5 hydroxyl would be necessary for glycosylation, a subset of 19 additional plasmids encoding PKSs that produce compounds containing this functional group was selected and tested. The desired desosaminylated polyketides would theoretically possess antibiotic activity, and the transformed strains can therefore be readily analyzed in a simple bioassay for production of glycosylated macrolides.
[0302] All of the strains transformed and tested displayed antimicrobial activity against $B$. subtilis. The presumed structures of the desosamine containing compounds, based on the structures of the aglycones produced by the PKS on each plasmid, are shown below.


KOS39-86



KAO127


KOS39-18
-continued


KOS11-62


KOS11-62


KOS11-64


KOS11-66
-continued



KOS15-22





KOS11-82


KOS11-83





KOS39-20


KOS15-46



KOS24-15


KOS15-30


KOS15-34
[0303] Culture extracts from six of these stains (those containing plasmids pKOS15-22, pKOS15-106, pKOS3920, pKOS1142, pKOS15-30, and pKOS2415) were examined by LC/MS and, in each case, the expected parent ion was found along with the 158 amu desosamine fragment. Two compounds were detected in the strain containing pKOS15-106 with molecular weights corresponding to 3-hydroxy and 3-keto derivatives. This is consistent with both aglycones being produced by plasmid $\mathrm{pKOS} 15-109$ in S. lividans. Two compounds were also detected in the strain with pKOS112, the predicted molecule, 5-O-desosaminyl10 -desmethyl-6-dEB, and a putative dehydrated derivative at carbons $\mathrm{C}-10$ and $\mathrm{C}-11$. Both aglycones were also produced when the plasmid was originally analyzed in $S$. lividans K4-114 (3), although only the former was reported at that time. As with the first set of plasmids tested, small amounts of $2^{\prime}$-O-glucosylated derivatives could also be detected in some of the culture extracts. The yields of the desosamine containing compounds were too low to determine absolute titers ( $<1 \mathrm{mg} / \mathrm{L}$ ) and, therefore, the relative antibacterial activity of the compounds could not be determined from these assays.
[0304] C. Discussion
[0305] This example demonstrates that a minimal set of seven genes (desI, II, III, IV, V, VI, VIII) is sufficient for biosynthesis of TDP-desosamine from glucose-1-phosphate in $S$. lividans. The apparent low abundance of TDP-desosamine in the engineered host could be due either to the availability of glucose-1-phosphate in this host or to poor expression of the sugar biosynthesis and/or transferase genes. Alternatively, it is interesting to note that narbonolide and 10 -deoxymethynolide are present in the natural picromycin/methymycin producing organism, $S$. venezuelae, and could therefore reflect that one or more of the enzymes from the des cluster is relatively inefficient. One can increase the amount of TDP-desosamine either by increasing expression levels of these genes and/or by complementing one or more of the enzymes in the pathway with homologs from other clusters such as erythromycin or oleandomycin.
[0306] Expression of the minimal desosamine biosynthesis genes together with the DesVII desosaminyltransferase in S. lividans has enabled the production of more than 20 glycosylated macrolides with detectable antibacterial activity. The structures of the macrolides that were glycosylated highlight both the remarkable substrate tolerance of the DesVII glycosyltransferase as well as the ability of desosamine to impart biological activity to structurally diverse macrolactones. In addition to their antibacterial properties the desosamine containing compounds presented here may possess additional biological properties that are associated with erythromycin and other macrolides, including motilin antagonism and anti-inflammatory activities. Furthermore, the demonstration by others that DesVII and other glycosyltransferases can also tolerate modifications of the sugar substituent ( $22,23,29$ ) opens the door to manipulation of both polyketide and deoxysugar pathways for the production of 'unnatural' natural product libraries.

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[0336] The invention having now been described by way of written description and example, those of skill in the art will recognize that the invention can be practiced in a variety of embodiments and that the foregoing description and examples are for purposes of illustration and not limitation of the following claims.

## SEquence LISting



-continued









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## 4545

4550
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Thr Ala Asp Leu His Glu Ala Arg Gly Arg Leu Arg Glu Leu Glu Ala
Lys Ala Gly Glu Pro Val Ala Ile Val Gly Met Ala Cys Arg Leu Pro

Leu Leu Glu Ala Ser Trp Glu Ala Phe Glu His Ala Gly Ile Pro Ala
Ala Thr Ala Arg Gly Thr Ser Val Gly Val Phe Thr Gly Val Met Tyr
145
150
His Asp Tyr Ala Thr Arg Leu Thr Asp Val Pro Glu Gly Ile Glu Gly

| Tyr Leu Gly Thr Gly Asn Ser Gly Ser Val Ala Ser Gly Arg Val Ala |  |
| ---: | :--- |
| 180 | 185 |


| Tyr Thr Leu Gly Leu Glu Gly Pro Ala Val Thr Val Asp |
| ---: |
| 195 |
| 200 |

Ser Ser Ser Leu Val Ala Leu His Leu Ala Val gln Ala Leu Arg Lys
Gly Glu Val Asp Met Ala Leu Ala Gly Gly Val Thr Val Met Ser Thr
Pro Ser Thr Phe Val Glu Phe Ser Arg Gln Arg Gly Leu Ala Pro Asp


| Asp Gly Ala Ser Ser Gly Leu Thr Ala Pro Asn Gly Pro Ser Gln Gln |  |  |  |
| ---: | ---: | ---: | ---: |
| 305 | 310 | 315 | 320 |

Arg Val Ile Arg Arg Ala Leu Ala Asp Ala Arg Leu Thr Thr Ser Asp
Val Asp Val Val Glu Ala His Gly Thr Gly Thr Arg Leu Gly Asp Pro






-continued



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Ala Glu Leu Gln Gln Asn Thr Arg Arg Leu Arg Glu Ile Glu Gly Arg
Thr His Glu Pro Val Ala Ile Val Gly Met Ala Cys Arg Leu Pro Gly

| Gly Val Ala Ser Pro Glu Asp |  |
| ---: | :--- |
| 50 | Leu Trp Gln Leu Val Ala Gly Asp Gly |
| 60 |  |


| Asp |  |  |
| ---: | ---: | ---: |
| 65 | Ala Ile Ser Glu Phe Pro Gln Asp Arg Gly Trp Asp Val Glu Gly |  |
| 70 | 75 | 80 |

Leu Tyr Asp Pro Asp Pro Asp Ala Ser Gly Arg Thr Tyr Cys Arg Ser
Gly Gly Phe Leu His Asp Ala Gly Glu Phe Asp Ala Asp Phe Phe Gly
Ile Ser Pro Arg Glu Ala Leu Ala Met Asp Pro Gln Gln Arg Leu Ser
Leu Thr Thr Ala Trp Glu Ala Ile Glu Ser Ala Gly Ile Asp Pro Thr
Ala Leu Lys Gly Ser Gly Leu Gly Val Phe Val Gly Gly Trp His Thr
145
150
Gly Tyr Thr Ser Gly Gln Thr Thr Ala Val Gln Ser Pro Glu Leu Glu
Gly His Leu Val Ser Gly Ala Ala Leu Gly Phe Leu Ser Gly Arg Ile
Ala Tyr Val Leu Gly Thr Asp Gly Pro Ala Leu Thr Val Asp Thr Ala
195
-continued




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-continued



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$<212>$ TYPE: PRT
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1
Phe His Pro Ala Pro Asn Ser Ala Val Arg Leu Val Cys Leu Pro His
$20 \quad 25 \quad 30$

Ala Gly Gly Ser Ala Ser Tyr Phe Phe Arg Phe Ser Glu Glu Leu His

|  |  | 35 |  |  |  | 40 |  |  |  |  | 45 |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Pro | $\begin{array}{r} \text { Ser } \\ 50 \end{array}$ | Val | Glu Ala | Leu | $\begin{array}{r} \text { Ser } \\ 55 \end{array}$ | Val | Gln | Tyr | Pro | $\begin{array}{r} \text { Gly } \\ 60 \end{array}$ | Arg | Gln | Asp | Arg |
| $\begin{array}{r} \text { Arg } \\ 65 \end{array}$ | Ala | Glu | Pro Cys | $\begin{array}{r} \text { Leu } \\ 70 \end{array}$ | Glu |  | Val | Glu | $\begin{array}{r} \text { Glu } \\ 75 \end{array}$ | Leu | Ala |  | His | $\begin{array}{r} \mathrm{Val} \\ 80 \end{array}$ |
| Val | Ala | Ala | $\begin{array}{r} \text { Thr Glu } \\ 85 \end{array}$ | Pro | rp | $\operatorname{Trp}$ | $\mathrm{Gln}$ | $\begin{array}{r} \text { Glu } \\ 90 \end{array}$ | Gly | Arg | Leu | Ala | Phe 95 | Phe |
| Gly |  | Ser | $\begin{aligned} & \text { Leu Gly } \\ & 100 \end{aligned}$ | Ala | er | Val | $\begin{aligned} & \text { Ala } \\ & 105 \end{aligned}$ | Phe | Glu | Thr | Ala | $\begin{aligned} & \text { Arg } \\ & 110 \end{aligned}$ | Ile | Leu |
| Glu | $\mathrm{Gln}$ | $\begin{gathered} \text { Arg } \\ 115 \end{gathered}$ | His Gly | Val | Arg | $\begin{aligned} & \text { Pro } \\ & 120 \end{aligned}$ | Glu | Gly | Leu | Tyr | $\begin{aligned} & \mathrm{Val} \\ & 125 \end{aligned}$ | Ser | Gly | Arg |
| Arg | $\begin{gathered} \text { Ala } \\ 130 \end{gathered}$ | Pro | Ser Leu | Ala | $\begin{gathered} \text { Pro } \\ 135 \end{gathered}$ | Asp | Arg | Leu | $\mathrm{Val}$ | $\begin{aligned} & \text { His } \\ & 140 \end{aligned}$ | $\mathrm{Gln}$ | Leu | Asp | Asp |
| $\begin{aligned} & \text { Arg } \\ & 145 \end{aligned}$ | Ala | Phe | Leu Ala | $\begin{aligned} & \text { Glu } \\ & 150 \end{aligned}$ | Ile | Arg | Arg | Leu | $\begin{aligned} & \text { Ser } \\ & 155 \end{aligned}$ | Gly | Thr | Asp | Glu | $\begin{aligned} & \text { Arg } \\ & 160 \end{aligned}$ |
| Phe | Leu | Gln | $\begin{array}{r} \text { Asp Asp } \\ 165 \end{array}$ | Glu | Leu | Leu | Arg | $\begin{aligned} & \text { Leu } \\ & 170 \end{aligned}$ | Val | Leu | Pro | Ala | $\begin{gathered} \text { Leu } \\ 175 \end{gathered}$ | Arg |
| Ser | sp | Tyr | $\begin{aligned} & \text { Lys Ala } \\ & 180 \end{aligned}$ | Ala | Glu | Thr | $\begin{aligned} & \text { Tyr } \\ & 185 \end{aligned}$ | Leu | His | Arg | Pro | $\begin{aligned} & \text { Ser } \\ & 190 \end{aligned}$ | Ala | Lys |
| Leu | Thr | $\begin{aligned} & \text { Cys } \\ & 195 \end{aligned}$ | Pro Val | Met | Ala | $\begin{aligned} & \text { Leu } \\ & 200 \end{aligned}$ | Ala | Gly | Asp | Arg | $\begin{aligned} & \text { Asp } \\ & 205 \end{aligned}$ | Pro | Lys | Ala |
| Pro | $\begin{aligned} & \text { Leu } \\ & 210 \end{aligned}$ | Asn | Glu Val | Ala | $\begin{aligned} & \text { Glu } \\ & 215 \end{aligned}$ | $\operatorname{Trp}$ | Arg | Arg | His | $\begin{aligned} & \text { Thr } \\ & 220 \end{aligned}$ | Ser | Gly | Pro | Phe |
| $\begin{aligned} & \text { Cys } \\ & 225 \end{aligned}$ | Leu | Arg | Ala Tyr | $\begin{aligned} & \text { Ser } \\ & 230 \end{aligned}$ | Gly | Gly | His | Phe | $\begin{aligned} & \text { Tyr } \\ & 235 \end{aligned}$ | Leu | Asn | Asp | $\mathrm{Gln}$ | $\begin{aligned} & \text { Trp } \\ & 240 \end{aligned}$ |
| His | Glu | Ile | $\begin{array}{r} \text { Cys Asn } \\ 245 \end{array}$ | Asp | Ile | Ser | Asp | $\begin{aligned} & \text { His } \\ & 250 \end{aligned}$ | Leu | Leu | Val | Thr | Arg <br> 255 | Gly |
| Ala | Pro | Asp | $\begin{aligned} & \text { Ala Arg } \\ & 260 \end{aligned}$ | Val | Val | $\mathrm{Gln}$ | $\begin{aligned} & \text { Pro } \\ & 265 \end{aligned}$ | Pro | Thr | Ser | Leu | $\begin{aligned} & \text { Ile } \\ & 270 \end{aligned}$ | Glu | Gly |
| Ala | Ala | $\begin{aligned} & \text { Lys } \\ & 275 \end{aligned}$ | Arg Trp | $\mathrm{Gln}$ | Asn | $\begin{aligned} & \text { Pro } \\ & 280 \end{aligned}$ | Arg |  |  |  |  |  |  |  |

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Ala Ala Tyr Glu Glu Leu Arg Ala $\begin{array}{r}\text { Glu } \\ 20\end{array}$ Thr Asp Ala Ala Ile Ala Arg
Val Leu Asp Ser Gly Arg Tyr Leu Leu Gly Pro Glu Leu Glu Gly Phe
Glu Ala Glu Phe Ala Ala Tyr Cys Glu Thr Asp His Ala Val Gly Val
Asn Ser Gly Met Asp Ala Leu Gln Leu Ala Leu Arg Gly Leu Gly Ile
Gly Pro Gly Asp Glu Val Ile Val Pro Ser His Thr Tyr Ile Ala Ser

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| $\begin{aligned} \text { Val Ala As } \\ 1 \end{aligned}$ |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| His Trp Il |  |  |  |  |  |
| Gly Gln Al |  |  |  |  |  |
| Gly Ala Le |  |  |  |  |  |
| Ala Leu Al |  |  |  |  |  |

-continued


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Val Ala Ser Gln Pro Ala Leu Thr Asp Thr Ile Thr Gly Ser Gly Leu
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$<211>$ LENGTH: 485
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Streptomyces venezuelae


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$<212>$ TYPE: PRT
$<213>$ ORGANISM: Streptomyces venezuelae
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Arg Gly Lys Asp Tyr Ala Ala Glu Ala Ser Asp Ile Ala Asp Leu Val
Arg Ser Arg Thr Pro Glu Ala Ser Ser Leu Leu Asp Val Ala Cys Gly
Thr Gly Thr His Leu Glu His Phe Thr Lys Glu Phe Gly Asp Thr Ala

| Gly Leu Glu Leu Ser Glu Asp Met Leu Thr His Ala Arg Lys Arg Leu |  |
| ---: | :--- |
| 65 | 70 |

Pro Asp Ala Thr Leu His Gln Gly Asp Met Arg Asp Phe Arg Leu Gly
Arg Lys Phe Ser Ala Val Val Ser Met Phe Ser Ser Val Gly Tyr Leu
Lys Thr Thr Glu Glu Leu Gly Ala Ala Val Ala Ser Phe Ala Glu His
115
120
Leu Glu Pro Gly Gly Val Val Val Val Glu Pro Trp Trp Phe Pro Glu130135140
Thr Phe Ala Asp Gly Trp Val Ser Ala Asp Val Val Arg Arg Asp Gly
145
150
Arg Thr Val Ala Arg Val Ser His Ser Val Arg Glu Gly Asn Ala Thr
165 $\quad 170 \quad 175$

Arg Met Glu Val His Phe Thr Val Ala Asp Pro Gly Lys Gly Val Arg | 185 |
| ---: | :--- |
| 180 |

His Phe Ser Asp Val His Leu Ile Thr Leu Phe His Gln Ala Glu Tyr
Glu Ala Ala Phe Thr Ala Ala Gly Leu Arg Val Glu Tyr Leu Glu Gly
Gly Pro Ser Gly Arg Gly Leu Phe Val Gly Val Pro Ala
<212> TYPE: PRT
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| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Asp | Arg | Gln | $\begin{array}{r} \text { Asn } \\ 20 \end{array}$ | Val | Gly | $\mathrm{yr}$ | Leu | $\begin{array}{r} \text { Pro } \\ 25 \end{array}$ | Gly | Val | Pro A | Arg | $\begin{array}{r} \text { Leu } \\ 30 \end{array}$ | Gly | Ile |
| Pro | Glu | $\begin{array}{r} \text { Leu } \\ 35 \end{array}$ | Arg | Ala | la | sp | $\begin{array}{r} \text { Gly } \\ 40 \end{array}$ | ro | Asn | Gly | Ile A | $\begin{array}{r} \text { Arg } \\ 45 \end{array}$ | Leu | Val | Gly |
| Gln | $\begin{array}{r} \text { Thr } \\ 50 \end{array}$ | Ala | hr | Ala | eu | $\begin{array}{r} \text { Pro } \\ 55 \end{array}$ | Ala | ro | Val | Ala | $\begin{gathered} \text { Leu A } \\ 60 \end{gathered}$ | Ala | Ser |  | Phe |
| $\begin{array}{r} \text { Asp } \\ 65 \end{array}$ | Asp | Thr | Met | Ala | Asp 70 | Ser | Tyr | Gly | Lys | Val 75 | Met | Gly | Arg | Asp | $\begin{array}{r} \text { Gly } \\ 80 \end{array}$ |
| Arg | Ala | Leu | Asn | $\begin{array}{r} \mathrm{Gln} \\ 85 \end{array}$ | Asp | Met | Val | Leu | $\begin{array}{r} \text { Gly } \\ 90 \end{array}$ | Pro | Met M | Met | Asn | $\begin{array}{r} \text { Asn } \\ 95 \end{array}$ | Ile |
| Arg | Val | Pro | $\begin{aligned} & \text { His } \\ & 100 \end{aligned}$ | Gly | $1 y$ | rg | Asn | $\begin{aligned} & \text { Tyr } \\ & 105 \end{aligned}$ | Glu | r | he | er | $\begin{aligned} & \text { Glu } \\ & 110 \end{aligned}$ | Asp | Pro |
| Leu | Val | $\begin{aligned} & \text { Ser } \\ & 115 \end{aligned}$ | Ser | Arg | Chr | Ala | $\begin{aligned} & \text { Val } \\ & 120 \end{aligned}$ | Ala | $\mathrm{Gln}$ | Ile | Lys | $\begin{aligned} & \text { Gly } \\ & 125 \end{aligned}$ | Ile | Gln | Gly |
| Ala | $\begin{aligned} & \text { Gly } \\ & 130 \end{aligned}$ | Leu | Met | Thr | hr | $\begin{gathered} \text { Ala } \\ 135 \end{gathered}$ | Lys | His | Phe | Ala | $\begin{aligned} & \text { Ala A } \\ & 140 \end{aligned}$ | Asn | Asn | Gln | Glu |
| Asn | Asn | Arg | e | Ser | Val | sn | Ala | Asn | Val | Asp | Glu G | Gln | Thr | Leu | Arg |
| 145 |  |  |  |  | 150 |  |  |  |  | 155 |  |  |  |  | 160 |
| Glu | Ile | Glu | Pe | $\begin{aligned} & \text { Pro } \\ & 165 \end{aligned}$ | Ala | Phe | Glu | Ala | $\begin{aligned} & \text { Ser } \\ & 170 \end{aligned}$ | Ser | Lys | Ala | Gly | $\begin{gathered} \text { Ala } \\ 175 \end{gathered}$ | Gly |
| Ser | he | Met | $\begin{aligned} & \text { Cys } \\ & 180 \end{aligned}$ | Ala | yr | sn | Gly | Leu <br> 185 | Asn | Gly | Lys | ro | $\begin{aligned} & \text { Ser } \\ & 190 \end{aligned}$ | Cys | Gly |
| Asn | Asp | $\begin{gathered} \text { Glu } \\ 195 \end{gathered}$ | Leu | Leu | Asn | Asn | $\begin{aligned} & \text { Val } \\ & 200 \end{aligned}$ | Leu | Arg | Thr | $\mathrm{Gln} \begin{aligned} & \mathrm{T} \\ & 2 \end{aligned}$ | $\begin{aligned} & \text { Trp } \\ & 205 \end{aligned}$ | Gly | Phe | Gln |
| Gly | $\begin{aligned} & \text { Trp } \\ & 210 \end{aligned}$ | Val | Met | Ser | sp | $\begin{aligned} & \operatorname{Trp} \\ & 215 \end{aligned}$ | Leu | Ala | Thr | Pro | $\begin{gathered} \text { Gly } \\ 2 \supset 0 \end{gathered}$ | Thr | Asp | Ala | Ile |
| $\begin{aligned} & \text { Thr } \\ & 225 \end{aligned}$ | Lys | Gly | Leu | sp | $\begin{aligned} & \text { Gln } \\ & 230 \end{aligned}$ | flu | Met | Gly | al | $\begin{aligned} & \text { Glu } \\ & 235 \end{aligned}$ | Leu | Pro | Gly | Asp | $\begin{aligned} & \mathrm{Val} \\ & 240 \end{aligned}$ |
| Pro | Lys | Gly | Glu | $\begin{aligned} & \text { Pro } \\ & 245 \end{aligned}$ | Ser | Pro | Pro | Ala | $\begin{aligned} & \text { Lys } \\ & 250 \end{aligned}$ | Phe | Phe | Gly | Glu | $\begin{aligned} & \text { Ala } \\ & 255 \end{aligned}$ | Leu |
| Lys | Thr | Ala | $\begin{aligned} & \text { Val } \\ & 260 \end{aligned}$ | Leu | sn | Gly | $12$ | $\begin{aligned} & \text { Val } \\ & 265 \end{aligned}$ | P | Glu | Ala | Ala | $\begin{aligned} & \text { Val } \\ & 270 \end{aligned}$ | Thr | Arg |
| Ser | Ala | $\begin{aligned} & \text { Glu } \\ & 275 \end{aligned}$ | Arg | Ile | al | Gly | $\begin{aligned} & \mathrm{Gln} \\ & 280 \end{aligned}$ | Met | Glu | Lys | Phe G | Gly | Leu | Leu | Leu |
| Ala | $\begin{aligned} & \text { Thr } \\ & 290 \end{aligned}$ | Pro | Ala | Pro | Arg | $\begin{aligned} & \text { Pro } \\ & 295 \end{aligned}$ | Glu | Arg | Asp | Lys | $\begin{gathered} \text { Ala } \\ 300 \end{gathered}$ | Gly | Ala | Gln | Ala |
| $\begin{aligned} & \text { Val } \\ & 305 \end{aligned}$ | Ser | Arg | Lys | Val | $\begin{gathered} \text { Ala } \\ 310 \end{gathered}$ | Glu | Asn | Gly | Ala | $\begin{aligned} & \text { Val } \\ & 315 \end{aligned}$ | Leu | Leu | Arg | Asn | $\begin{aligned} & \text { Glu } \\ & 320 \end{aligned}$ |
| Gly | Gln | Ala | Leu | $\begin{aligned} & \text { Pro } \\ & 325 \end{aligned}$ | Leu | Ala | Gly | Asp | $\begin{gathered} \text { Ala } \\ 330 \end{gathered}$ | Gly | Lys S | Ser | Ile | $\begin{gathered} \text { Ala } \\ 335 \end{gathered}$ | Val |
| Ile | Gly | Pro | $\begin{aligned} & \text { Thr } \\ & 340 \end{aligned}$ | Ala | Val | Asp | Pro | $\begin{aligned} & \text { Lys } \\ & 345 \end{aligned}$ | Val | Thr | Gly L | Leu | $\begin{aligned} & \text { Gly } \\ & 350 \end{aligned}$ | Ser | Ala |
| His | Val | $\begin{aligned} & \text { Val } \\ & 355 \end{aligned}$ | Pro | Asp | Ser | Ala | $\begin{aligned} & \text { Ala } \\ & 360 \end{aligned}$ | Ala | Pro | Leu | $\begin{gathered} \text { Asp } \\ \\ 3 \end{gathered}$ | $\begin{aligned} & \text { Thr } \\ & 365 \end{aligned}$ | Ile | Lys | Ala |
| Arg | Ala $370$ | Gly | Ala | Gly | Ala | $\begin{aligned} & \text { Thr } \\ & 375 \end{aligned}$ | Val | Thr | Tyr | Glu | Thr <br> 380 | Gly | Glu | Glu | Thr |


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$<210>$ SEQ ID NO 14
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$<400>$ SEQUENCE $: 14$


$<210>$ SEQ ID NO 15
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$<400>$ SEQUENCE $: 15$



$<210>$ SEQ ID NO 17
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Val Ser Leu Ser Phe Leu Val Asp Thr Leu Asp Ile Arg His Gly Gly
35
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Val Leu Val Gly Ala Val Gly Lys Asp Phe Asp Gly Tyr Gly Gln Leu
Leu Arg Ala Ala Gly Val Asp Thr Asp Ser Val Arg Val Ser Asp Arg
Gln His Thr Ala Arg Phe Met Cys Thr Thr Asp Glu Asp Gly Asn Gln

Glu Leu Gly Leu Arg Arg Ala Ala Asp Pro Ser Gln Gln Leu Ala Arg
Leu Glu Gly Asp Ser Val Arg Glu Leu Val Asp Gly Ala Glu Leu Leu

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Gly Ala Lys Gly Cys Arg
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$<210>$ SEQ ID NO 18
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<220> FEATURE:

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<213> ORGANISM: Streptomyces venezuelae

<220> FEATURE:

<221> NAME/KEY: 2875

$<222>$ LOCATION: unsure

$<223>$ OTHER INFORMATION: unsure of nucleotide at this position

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1 5

1. An isolated recombinant DNA compound that comprises a coding sequence for a domain of a narbonolide PKS.
2. The isolated recombinant DNA compound of claim 1, wherein said domain is selected from the group consisting of a thioesterase domain, a KSQ domain, an AT domain, a KS domain, an ACP domain, a KR domain, a DH domain, and an ER domain.
3. The isolated recombinant DNA compound of claim 2 that comprises the coding sequence for a loading module, thioesterase domain, and all six extender modules of the narbonolide PKS.
4. An isolated recombinant DNA compound that comprises a coding sequence for a desosamine biosynthetic gene or a desosaminyl transferase gene or a beta-glucosidase gene of Streptomyces venezuelae.
5. An isolated recombinant DNA compound that comprises a coding sequence for a picK hydroxylase gene of Streptomyces venezuelae.
6. An isolated DNA compound of any of claim 1 that further comprises a promoter operably linked to said coding sequence.
7. The isolated recombinant DNA compound of claim 6, wherein said promoter is a promoter derived from a cell other than a Streptomyces venezuelae cell.
8. The isolated recombinant DNA compound of claim 7 that is a recombinant DNA expression vector.
9. The recombinant DNA expression vector of claim 8 that expresses a PKS in Streptomyces host cells.
10. The recombinant DNA expression vector of claim 9 that encodes a hybrid PKS comprising at least a portion of a narbonolide PKS gene and at least a portion of a second PKS gene for a macrolide aglycone other than narbonolide.
11. The recombinant DNA compound of claim 10 , wherein said second PKS gene is a DEBS gene.
12. The recombinant DNA compound of claim 11, wherein said hybrid PKS is composed of a loading module
and extender modules 1 through 6 of DEBS excluding a KR domain of extender module 6 of DEBS and an ACP of extender module 6 and a thioesterase domain of the narbonolide PKS.
13. A recombinant host cell, which in its untransformed state does not produce 10 -deoxymethynolide or narbonolide, that comprises a recombinant DNA expression vector of claim 9 that encodes a narbonolide PKS and said cell produces 10 -deoxymethynolide or narbonolide.
14. The recombinant host cell of claim 13 that further comprises a picB gene.
15. The recombinant host cell of claim 13 that further comprises desosamine biosynthetic genes and a gene for desosaminyl transferase and produces YC17 or narbomycin.
16. The recombinant host cell of claim 15 that further comprises a picK gene and produces methymycin, neomethymycin, or picromycin.
17. The recombinant host cell of any of claim 16 that is Streptomyces coelicolor or Streptomyces lividans.
18. A recombinant host cell other than a Streptomyces venezuelae cell that expresses a picK hydroxylase gene of $S$. venezuelae encoded by the DNA compound of claim 5.
19. A recombinant host cell other than a Streptomyces venezuelae host cell that expresses a desosamine biosynthetic gene or desosaminyl transferase gene of $S$. venezuelae encoded by the DNA compound of claim 4.
20. A method for increasing the yield of a desosaminylated polyketide in a cell, which method comprises transforming the cell with a recombinant expression vector that encodes a functional beta-glucosidase gene.

[^0]:    1 CGTGGCGGCC GCCGCTCCCG GCGCCGCCGA CACGGCCAAT GTTCAGTACA CGAGCCGGGC
    61 GGCGGAGCTC GTCGCCCAGA TGACGCTCGA CGAGAAGATC AGCTTCGTCC ACTGGGCGCT

    121 GGACCCCGAC CGGCAGAACG TCGGCTACCT TCCCGGCGTG CCGCGTCTGG GCATCCCGGA

[^1]:    Amino acid sequence of picromycin/methymycin
    cytochrome P450 hydroxylase, PicK
    1 VRRTQQGTTA SPPVLDLGAL GQDFAADPYP TYARLRAEGP AHRVRTPEGD EVWLVVGYDR

    61 ARAVLADPRF SKDWRNSTTP LTEAEAALNH NMLESDPPRH TRLRKLVARE FTMRRVELLR
    121 PRVQEIVDGL VDAMLAAPDG RADLMESLAW PLPITVISEL LGVPEPDRAA FRVWTDAFVF

    181 PDDPAQAQTA MAEMSGYLSR LIDSKRGQDG EDLLSALVRT SDEDGSRLTS EELLGMAHIL
    241 LVAGHETTVN LIANGMYALL SHPDQLAALR ADMTLLDGAV EEMLRYEGPV ESATYRFPVE

    301 PVDLDGTVIP AGDTVLVVLA DAHRTPERFP DPHRFDIRRD TAGHLAFGHG IHFCIGAPLA
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[^2]:    $<210>$ SEQ ID NO 9
    <211> LENGTH: 331
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