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(54) **Title:** USE OF PLANT CELLS EXPRESSING A TNF α POLYPEPTIDE INHIBITOR IN THERAPY

(57) **Abstract:** A method of treating a TNF Alpha associated medical condition selected from the group consisting of obesity, metabolic syndrome, diabetes and a liver disease or disorder is provided. The method comprising enterally administering to a subject in need thereof a therapeutically effective amount of plant cells expressing a TNF Alpha polypeptide inhibitor, thereby treating the TNF Alpha associated medical condition.



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USE OF PLANT CELLS EXPRESSING A TNFalpha POLYPEPTIDE INHIBITOR IN THERAPY

5 FIELD AND BACKGROUND OF THE INVENTION

The present invention, in some embodiments thereof, relates to the use of plant cells expressing a TNFalpha polypeptide inhibitor in therapy.

Tumor necrosis factor alpha (TNF α) is an important, pro-inflammatory cytokine mediating the regulation of diverse inflammatory, infectious and immune-related processes and diseases, TNF α being considered the most important mediator responsible for inflammatory pathology.

TNF-alpha is a 17 kD molecular weight protein, initially synthesized as a transmembrane protein arranged in stable trimers, then cleaved by metalloprotease-TNF alpha converting enzyme (TACE) to form the homotrimeric soluble TNF (sTNF) which engages to its cognate receptors (TNFR1, p55 and TNFR2, p75), expressed ubiquitously. The ubiquitous TNF receptors provides the basis for the wide variety of TNF-alpha mediated cellular responses.

TNF-alpha induces a wide variety of cellular responses, many of which result in deleterious consequences, such as cachexia (loss of fat and whole body protein depletion, leading to anorexia, common in cancer and AIDS patients) and septic shock. Elevated secretion of TNF-alpha has been implicated in a variety of human diseases including diabetes, allograft rejection, sepsis, inflammatory bowel diseases, osteoporosis, in many autoimmune diseases such as multiple sclerosis, rheumatoid arthritis, psoriasis, psoriatic arthritis, hypersensitivity, immune complex diseases, and even in malaria, cancer and lung fibrosis.

The biological effect of TNF α is mediated by the two distinct receptors. TNF-alpha receptors, when shed from mononuclear cells, lower the TNF-alpha levels by "mopping up" and acting as natural inhibitors. Neutralization of TNF α by specific antibodies and decoy receptors has become a common strategy for regulation of TNF α mediated toxicity.

To date, five protein-based TNF α antagonists have been approved by the US FDA for clinical use: Cimzia (Certolizumab pegol), a TNFmAb Fab' fragment-PEG conjugate; Remicade (Infliximab), a TNF rmAB; Humira (Adalimumab, a TNF rmAB,

Simponi™ (Golimumab), an anti-TNF and etanercept, a fusion protein of soluble 75 kDa TNF α receptors fused to the Fc fragment of human IgG (registered as Enbrel™).

Etanercept is indicated for rheumatoid arthritis (RA) and other arthritic indications such as juvenile idiopathic arthritis (JIA), psoriasis and Ankylosing Spondylitis (AS). Rheumatoid arthritis (RA) is a chronic disease that affects approximately five million people World Wide. Nearly 500,000 patients worldwide across indications are treated with Enbrel. Enbrel sales in 2010 were 7.8 billion dollars and the total anti- TNF market amounted to 24.04 Billion dollars. Clinical trials of Enbrel therapy, current or completed, include such diverse indications as adult respiratory distress syndrome, pemphigus, Alzheimer's disease, Behcet's syndrome, HIV, myocardial infarct, knee joint synovitis, lupus nephritis, lichen planus, systemic amyloidosis, sciatica, vitiligo, chronic fatigue syndrome, anorexia, TMJ, asthma, bronchitis, diabetes, myelodysplastic disease and others.

Biopharmaceuticals typically pose a number of challenges, however, that drug developers must overcome in order to successfully develop these compounds into safe and effective therapeutics. For example, proteins and peptides tend to be destroyed by proteolytic enzymes or, in the case of the higher molecular weight proteins, may generate neutralizing antibodies. Moreover, large complex molecules can exhibit low solubility or poor stability, leading to short shelf lives. As a result, biopharmaceutical therapeutics often quickly lose their effectiveness or require frequent dosing. These factors impact not only cost of therapy, but also patient acceptance and compliance, thus affecting their therapeutic efficacy.

Oral Administration:

The most common mode of protein and peptide-based administration is by invasive methods of drug delivery, such as injections and infusions. Although these are the primary modes for administering macromolecular drugs for systemic diseases, they are also the least desirable for patients and practitioners. The obvious downside of this delivery method is patient acceptance and compliance, limiting most macromolecule development to indications in which the need to use invasive administration routes are not outweighed by associated expenses or inconvenience. As a simple, non-invasive method for systemically delivering drugs, oral administration provides many advantages: ease and convenience of use, access to extensive volume of absorptive

surface, high degree of vascularization, relatively lengthy retention time, natural disposal of inactive, non-metabolized ingredients, and more.

Nonetheless, investigations of oral administration of macromolecular pharmaceuticals have not indicated satisfactory levels of efficiency to match the potential of this route. Some of the obstacles are difficulties of ingestion of pills and other solid formulations, lability of biologically active macromolecules in the GI tract, concentration of the biologically active agents at the mucosa, and permeability of GI membranes to biologically active macromolecules.

The oral route of administration of biologically active substances is complicated by both high acidity and enzymatic degradation in the stomach, which can inactivate or destroy biologically active macromolecules before they reach their intended target tissue. Further, effective concentrations of a biologically active macromolecule are difficult to achieve in the large volumes encountered in the GI tract. Thus, to be effective, most drugs must be protected from absorption and/or the environment in the upper GI tract, and then be abruptly released into the intestine or colon. Various strategies are being developed in the pharmaceutical industry to overcome the problems associated with oral or enteral administration of therapeutic macromolecules such as proteins. These strategies include covalent linkage with a carrier, coatings and formulations (pH sensitive coatings, polymers and multi-layered coatings, encapsulation, timed release formulations, bioadhesives systems, osmotic controlled delivery systems, etc) designed to slow or prevent release of active ingredients in harsh conditions such as the stomach and upper GI tract. However, preparation of biologically active agents in such formulations requires complex and costly processes. Also employed are mucosal adhesives and penetration enhancers (salicylates, lipid-bile salt-mixed micelles, glycerides, acylcarnitines, etc) for increasing uptake at the mucosa. However, some of these can cause serious local toxicity problems, such as local irritation, abrasion of the epithelial layer and inflammation of tissue. Other strategies to improve oral delivery include mixing the biologically active agent with protease inhibitors, such as aprotinin, soybean trypsin inhibitor, and amastatin; however, enzyme inhibitors are not selective, and also inhibit endogenous macromolecules, causing undesirable side effects. Thus, present methods of oral administration of biologically active biopharmaceuticals cannot ensure efficient delivery of desired biological activity at the target tissue.

Attempts at orally administering TNFR2:Fc (Enbrel) have failed to due to the high acidity and enzymatic degradation in the stomach that inactivates or destroys the molecule before reaching the circulation. Elaborate, complicated mechanisms, including devices for automatic parenteral administration have evolved to ensure compliance with dosage regimens.

Additional background art includes: US Patent NO. 7,915,225 to Finck et al, US Patent Applications Nos. 13/021,545 and 10/853,479 to Finck et al, US Patent Application No. 11/906,600 to Li et al, US Patent Application No. 10/115,625 to Warren et al and US Patent Application No. 11/784,538 to Gombotz et al.

SUMMARY OF THE INVENTION

According to an aspect of some embodiments of the present invention there is provided a method of treating a TNF α associated medical condition selected from the group consisting of obesity, metabolic syndrome, diabetes and a liver disease or disorder, the method comprising enterally administering to a subject in need thereof a therapeutically effective amount of plant cells expressing a TNF α polypeptide inhibitor, thereby treating the TNF α associated medical condition.

According to an aspect of some embodiments of the present invention there is provided a use of plant cells expressing a TNF α polypeptide inhibitor for the enteral treatment of a TNF α associated medical condition selected from the group consisting of obesity, metabolic syndrome, diabetes and a liver disease or disorder.

According to some embodiments of the invention, the enteral is oral administration.

According to some embodiments of the invention, the TNF α polypeptide inhibitor is an anti-TNF α antibody.

According to some embodiments of the invention, the anti-TNF α antibody is infliximab, adalimumab or golimumab.

According to some embodiments of the invention, the TNF α polypeptide inhibitor is a chimeric polypeptide comprising:

(i) a first domain which comprises a TNF α binding domain of a TNF receptor; and

(ii) a second domain which comprises an Fc domain of an immunoglobulin, wherein the first domain and the second domain are N-terminally to C-terminally respectively sequentially translationally fused and wherein the chimeric polypeptide specifically binds TNF α .

5 According to some embodiments of the invention, the chimeric polypeptide further comprises a third domain which comprises an endoplasmic reticulum retention signal, wherein the first domain, second domain and third domain are N-terminally to C-terminally respectively sequentially translationally fused.

10 According to some embodiments of the invention, the method or use comprising an additional domain encoding an endoplasmic reticulum signal peptide translationally fused N-terminally to the first domain.

According to some embodiments of the invention, the signal peptide is a plant signal peptide.

15 According to some embodiments of the invention, the plant signal peptide is as set forth in SEQ ID NO: 4.

According to some embodiments of the invention, the first domain is 200-250 amino acids long.

According to some embodiments of the invention, the first domain comprises the amino acid sequence LCAP (SEQ ID NO: 11) and VFCT (SEQ ID NO: 12).

20 According to some embodiments of the invention, the first domain further comprises the amino acid sequence LPAQVAFXPYAPEPGSTC (SEQ ID NO: 13).

According to some embodiments of the invention, the first domain is as set forth in SEQ ID NO: 2.

According to some embodiments of the invention, the immunoglobulin is IgG₁.

25 According to some embodiments of the invention, the second domain is as set forth in SEQ ID NO: 9.

According to some embodiments of the invention, the chimeric polypeptide is as set forth in SEQ ID NO: 6.

30 According to some embodiments of the invention, the chimeric polypeptide is as set forth in SEQ ID NO: 7, 204 or 205.

According to some embodiments of the invention, the chimeric polypeptide is capable of inhibiting TNF α -induced apoptosis.

According to some embodiments of the invention, the TNF α polypeptide inhibitor comprises a plant-specific glycan.

According to some embodiments of the invention, the plant-specific glycan is selected from the group consisting of a core xylose and a core α -(1,3) fucose.

5 According to some embodiments of the invention, the plant cells are *Nicotiana tabacum* plant cells.

According to some embodiments of the invention, the *Nicotiana tabacum* plant cell is a Bright Yellow (BY-2) cell.

According to some embodiments of the invention, the plant cells are lyophilized.

10 According to some embodiments of the invention, the plant cells are grown in suspension.

According to some embodiments of the invention, the liver disease or disorder is selected from the group consisting of hepatitis, liver cirrhosis, liver cancer, hepatotoxicity, chronic liver disease, fatty liver disease and non-alcoholic steatohepatitis
15 (NASH).

According to some embodiments of the invention, the hepatotoxicity is induced by a chemical agent selected from the group consisting of acetaminophen, NTHES, glucocorticoid, isniazed, arsenic, carbon tetrachloride and vinyl chloride.

20 According to some embodiments of the invention, the diabetes is selected from the group consisting of type I diabetes, type II diabetes and LADA disease.

According to some embodiments of the invention, the plant cells are provided in an oral nutritional form.

According to some embodiments of the invention, the oral nutritional form is a complete meal, a powder for dissolution, a bar, a baked product, a cereal bar, a dairy bar,
25 a snack-food, a breakfast cereal, muesli, candies, tabs, cookies, biscuits, crackers, chocolate, and dairy products.

Unless otherwise defined, all technical and/or scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which
30 the invention pertains. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of embodiments of the invention, exemplary methods and/or materials are described below. In case of conflict, the patent

specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and are not intended to be necessarily limiting.

BRIEF DESCRIPTION OF THE DRAWINGS

Some embodiments of the invention are herein described, by way of example only, with reference to the accompanying drawings. With specific reference now to the drawings in detail, it is stressed that the particulars shown are by way of example and for purposes of illustrative discussion of embodiments of the invention. In this regard, the description taken with the drawings makes apparent to those skilled in the art how embodiments of the invention may be practiced.

In the drawings:

FIG. 1 is a schematic illustration of the amino acid sequence of plant expressed recombinant human (prh) TNFR2:Fc (also termed herein PRX-106, SEQ ID NO:6). prh TNFR2:Fc cDNA for expression in BY2 cells was assembled with a signal peptide for targeting the fusion polypeptide composed of the TNF-binding moiety of the TNF receptor and FC protein to the secretory pathway. Colour code for the amino acid sequence: Yellow: signal peptide; Black: TNF receptor portion; Blue: Fc portion of IgG1; Red: ER retention signal;

FIGs. 2A-B show a comparison of PRH TNFR2:FC and commercial Enbrel® by Western-blot. Prh TNFR2:Fc (lane 1) and commercial Enbrel (lane 2) were analyzed under reducing conditions (FIG. 2A) and non-reducing conditions (FIG. 2B) by 12 % and 8 % Tris-Glycine SDS-PAGE, respectively. Membranes were blotted with an anti human IgG (antiFC) antibody (upper panel) and with an anti TNFR2 antibody (lower panel). Molecular weight marker is shown in right lanes. Lane 1:PRH TNFR2:FC; Lane 2:commercial Enbrel®;

FIG. 3 is a graph showing TNF α binding by prh TNFR2:Fc and commercial Enbrel® by quantitative non radioactive assay for prh TNFR2:Fc binding activity and molecular integrity. An ELISA plate pre-coated with antibodies against TNF α , was incubated with TNF α followed by exposure to commercial Enbrel® and supernatant from BY2 cells expressing prh TNFR2:Fc. Serial dilutions of both preparations are shown in the X axis. Fc portion of the molecule was detected with Goat anti human IgG Fc HRP;

FIG. 4 is an image showing screening of individual plant cell lines for expression of prh TNFR2:Fc by Western blot analysis with anti-IgG (anti-Fc) antibody;

FIGs. 5A-F are images showing TNF α cytotoxicity in A375 cells in the presence of prh TNFR2:Fc or commercial Enbrel® by MTT viability assay. FIG. 5A-untreated cultured A375 cells ; FIG. 5B-treated with TNF α ; FIG. 5C- TNF α exposed cells treated with prh TNFR2:Fc (3.125 ng/ml); FIG. 5D- TNF α exposed cells treated with commercial Enbrel® (3.125 ng/ml); FIG. 5E- TNF α exposed cells treated with prh TNFR2:Fc (100 ng/ml) ; FIG. 5F- TNF α exposed cells treated with commercial Enbrel® (100 ng/ml);

FIG. 5G is a bar graph showing TNF α cytotoxicity in A375 cells in the presence of prh TNFR2:Fc or commercial Enbrel by MTT viability assay;

FIGs. 6A-F are images showing TNF α cytotoxicity in L929 cells in the presence of prh TNFR2:Fc or commercial Enbrel® by MTT viability assay. FIG. 6A-untreated cultured L929 cells; FIG. 6B-treated with TNF α ; FIG. 6C- TNF α exposed cells treated with prh TNFR2:Fc (3.125 ng/ml) ; FIG. 6D- TNF α exposed cells treated with commercial Enbrel® (3.125 ng/ml); FIG. 6E- TNF α exposed cells treated with prh TNFR2:Fc (100 ng/ml) ; FIG. 6F- TNF α exposed cells treated with commercial Enbrel® (100 ng/ml);

FIG. 6G is a bar graph showing are images showing TNF α cytotoxicity in L929 cells in the presence of prh TNFR2:Fc or commercial Enbrel® by MTT viability assay;

FIGs. 7A-C are bar graphs illustrating the effective anti-inflammatory activity of plant cells expressing recombinant TNFR2:Fc on serum markers of hepatotoxicity in the concanavalin A (Con A) mouse immune mediated hepatitis model. Mice received plant cells expressing recombinant TNFR2:Fc (plant TNFR2:Fc), steroid anti-inflammatory treatment (Dexamethasone), host plant control cells (BY2) or no treatment (Saline) 6 hours prior to i.v. administration of concanavalin A (Con A). 14 hours after con A administration serum liver enzymes (alanine aminotransferase ALT and aspartate aminotransferase AST) were assayed to assess extent of liver parenchymal damage. FIGs. 7A and 7C- column 1-saline control; column 2-Dexamethasone; column 3- plant cells expressing recombinant TNFR2:Fc equivalent to 5 μ g TNFR2:Fc protein; column 4- equivalent volume host plant control cells (BY2). Figure 7B- column 1-saline control; column 2-Dexamethasone; column 3- plant cells expressing recombinant

TNFR2:Fc equivalent to 0.5 μ g TNFR2:Fc protein; column 4- plant cells expressing recombinant TNFR2:Fc equivalent to 5 μ g TNFR2:Fc protein; column 5-host plant control cells (BY2) equivalent volume to column 3; column 6- host plant control cells (BY2) equivalent volume to column 4. FIG. 7A- n=6, * p<0.01; ** p<0.0005 ; *** p<0.00005, relative to saline & p< 0.05, relative to negative control. FIG. 7B-n=6, * p<0.02; relative to saline, & p< 0.0005, relative to negative control, # p< 0.03, relative to negative control. FIG. 7C- n=6, * p<0.01; ** p<0.0005 ; *** p<0.00005, relative to saline, & p< 0.00005, relative to negative control;

FIGs 8A-8C are bar graphs illustrating the effective anti-inflammatory activity of oral administration of plant cells expressing recombinant TNFR2:Fc on serum IFN-gamma levels in the concanavalin A (Con A) mouse immune-mediated hepatitis model. Mice received oral administration of plant cells expressing recombinant TNFR2:Fc, host plant control cells (BY2), steroid or saline prior to administration of Con A as described in FIGs. 7A-7C. 14 hours after con A administration serum INF-gamma was assayed by ELISA. FIGs. 8A and 8C- column 1-saline control; column 2-Dexamethasone; column 3- plant cells expressing recombinant TNFR2:Fc equivalent to 5 μ g TNFR2:Fc protein; column 4- host plant control cells (BY2) equivalent volume. FIG. 8B- column 1-saline control; column 2-Dexamethasone; column 3- plant cells expressing recombinant TNFR2:Fc equivalent to 0.5 μ g TNFR2:Fc protein; column 4- plant cells expressing recombinant TNFR2:Fc equivalent to 5 μ g TNFR2:Fc protein; column 5- host plant control cells (BY2) equivalent to column 3; column 6- host plant control cells (BY2) equivalent to column 4. FIG. 8A-n=6, * p<0.05; ** p<0.00001, relative to saline, & p< 0.0004, relative to negative control. FIG. 8B- n=6, * p<0.05; ** p<0.00001, relative to saline, & p< 0.004, relative to negative control, # p< 0.02, relative to negative control. FIG. 8C- n=6, * p<0.05, relative to saline, & p< 0.09, relative to negative control;

FIGs. 9A-9C are photomicrographs of exemplary liver slices illustrating prevention of hepatotoxicity by oral administration of plant cells expressing recombinant TNFR2:Fc in the mouse concanavalin A (Con A) immune-mediated hepatitis model. Mice received plant cells expressing recombinant TNFR2:Fc, host plant control cells (BY2), or saline prior to administration of Con A as described in FIGs. 6A-6C. 14 hours after con A administration livers were excised, fixed in formaldehyde, sectioned and stained with hematoxylin and evaluated by light microscopy. FIG. 9A-Con A + saline

(control). FIG. 9B- Con A + plant cells expressing recombinant TNFR2:Fc equivalent to 0.5µg TNFR2:Fc protein. FIG. 9C- Con A + mass of host plant control BY2 cells (BY2) equivalent to FIG. 9B; and

FIGs. 10A and 10B are bar graphs illustrating the effective anti-inflammatory activity of orally administered plant cells expressing recombinant TNFR2:Fc on serum markers of hepatotoxicity in the concanavalin A (Con A) mouse immune-mediated hepatitis model, as compared to that of mammalian recombinant cell-produced TNFR2:Fc. Mice received plant cells expressing recombinant TNFR2:Fc (plant TNFR2:Fc) equivalent to 5µg TNFR2:Fc protein, administered orally (FIG. 10A, column 2), 0.1 mg mammalian recombinant TNFR2:Fc (Etanercept), administered intraperitoneally (FIG. 9B, column 2) or control treatment (FIG. 10A and 10B, column 1), 6 hours prior to i.v. administration of concanavalin A (Con A). 14 hours after con A administration serum liver biochemistry marker alanine aminotransferase (ALT) was assayed to assess extent of liver parenchymal damage. FIG. 10A- n=6, * p<0.01; ** p<0.0005 ; *** p<0.00005, relative to saline & p< 0.05, relative to negative control. Note the equivalent anti-inflammatory effect of the orally administered plant cells expressing recombinant TNFR2:Fc to that of 0.1 mg mammalian recombinant TNFR2:Fc fusion protein (Etanercept) administered i.p.

FIG. 11 is a bar graph showing the effect of oral administration of recombinant TNFR2:Fc in plant cells on serum levels in high fat diet mouse model.

FIGs. 12A-B are bar graphs showing the effect of oral administration of recombinant TNFR2:Fc in plant cells on serum TGs in high fat diet mouse model. * p<0.0001, compared to saline; &, p<0.002, compared to mock.

FIG. 13 is a graph showing weight gain in HFD mice.

FIG. 14 is a bar graph showing the effect of oral administration of recombinant TNFR2:Fc in plant cells on hepatic Tregs in HFD mice. * p<0.05, compared to saline.

FIG. 15 is a bar graph showing the effect of oral administration of recombinant TNFR2:Fc in plant cells on hepatic NK cells in HFD mice. * p<0.05, compared to saline.

FIG. 16 is a bar graph showing the effect of oral administration of recombinant TNFR2:Fc in plant cells on splenic/hepatic CD4+CD25+FOXP3+ Ratio.

FIG. 17 is a bar graph showing the effect of oral administration of recombinant TNFR2:Fc in plant cells on splenic/hepatic CD8+CD25+FOXP3+ Ratio.

DESCRIPTION OF SPECIFIC EMBODIMENTS OF THE INVENTION

5 The present invention, in some embodiments thereof, relates to the use of plant cells expressing a TNF-alpha inhibitor in therapy.

Before explaining at least one embodiment of the invention in detail, it is to be understood that the invention is not necessarily limited in its application to the details set forth in the following description or exemplified by the Examples. The invention is
10 capable of other embodiments or of being practiced or carried out in various ways.

Accurate delivery of biopharmaceuticals to their target tissues poses challenges impacting not only cost of therapy, but also patient acceptance and compliance, thus affecting their therapeutic efficacy. Oral administration of macromolecular biopharmaceuticals must overcome obstacles such as ingestion of pills and other solid
15 formulations, lability of biologically active macromolecules in the GI tract, concentration of the biologically active agents at the mucosa, and low permeability of GI membranes to biologically active macromolecules.

Previous attempts at orally administering TNFR2:Fc (Enbrel®) have failed due to acidity and enzymatic degradation in the stomach. The present inventors have
20 surprisingly shown that a biologically active TNF-binding protein (TNFR2:Fc) can be effectively orally administered by feeding plant cells expressing the recombinant TNFR2:Fc, and that oral administration of the plant cells expressing recombinant TNFR2:Fc provides significant protection from immune-mediated inflammatory disease.

When tobacco BY2 cells were transformed with a nucleic acid construct
25 encoding recombinant TNFR2:Fc and cultured, the resulting TNF-binding protein was shown to be accurately expressed (see Example 1), having similar electrophoretic mobility, immunological cross reactivity and TNF alpha binding characteristics to those of commercial, mammalian cell expressed recombinant TNFR2:Fc (Enbrel®)(see Figures 2-3). In vitro assay of biological function of the plant cell expressed
30 recombinant TNFR2:Fc provided further evidence of protection of cells from TNF-mediated apoptosis, using two distinct types of target cells, (see Figures 5A-F and 6A-6G) comparable to that of Enbrel®.

Surprisingly, when cultured plant cells expressing the recombinant TNFR2:Fc were fed to mice prior to induction of concanavalin A immune-mediated hepatotoxicity, a significant and dose-dependent reduction in liver damage and serum levels of cytokine markers of inflammation was observed (Example 3, figures 7-9). Comparison of oral
5 administration of plant cells expressing the recombinant TNFR2:Fc and conventional intraperitoneal administration of Enbrel® revealed nearly identical reduction of serum liver enzyme levels, indicating effective protection from the immune-related inflammatory injury characteristic of the con A hepatotoxicity model.

While further reducing some embodiment of the present invention to practice, the
10 present inventors have uncovered that oral administration of plant cells expressing the recombinant TNFR2:Fc causes ameliorates certain clinical manifestation of fatty acid disease modeled by high fat diet mice (see Figures 11-17). Thus, oral administration of plant cells expressing the recombinant TNFR2:Fc caused a decrease in serum enzymes and triglycerides in the animal model of fatty liver disease. The drug also altered the
15 splenic and hepatic distribution of various populations of T cells and NK cells, indicating that the drug also functions as an immunomodulator of NAFLD and in metabolic syndrome.

Thus, according to an aspect of the invention there is provided a method of treating a TNF α associated medical condition selected from the group consisting of
20 obesity, metabolic syndrome, diabetes, hyperlipidemia and a liver disease or disorder, the method comprising enterally administering to a subject in need thereof a therapeutically effective amount of plant cells expressing a TNF α polypeptide inhibitor, thereby treating the TNF α associated medical condition.

Alternatively or additionally there is provided a use of plant cells expressing a
25 TNF α polypeptide inhibitor for the treatment of a TNF α associated medical condition directly associated with obesity, metabolic syndrome, diabetes and a liver disease or disorder.

The term “treating” refers to inhibiting, preventing or arresting the development of a pathology (disease, disorder or condition) and/or causing the reduction, remission,
30 or regression of a pathology. Those of skill in the art will understand that various methodologies and assays can be used to assess the development of a pathology, and

similarly, various methodologies and assays may be used to assess the reduction, remission or regression of a pathology.

As used herein, the term “preventing” refers to keeping a disease, disorder or condition from occurring in a subject who may be at risk for the disease, but has not yet
5 been diagnosed as having the disease.

As used herein, the term “subject” includes mammals, e.g., human beings at any age which suffer from the pathology. According to a specific embodiment, this term encompasses individuals who are at risk to develop the pathology.

The present teachings are thus directed at treating or preventing medical
10 conditions which are directly associated with obesity, metabolic syndrome, diabetes and a liver disease or disorder. According to some embodiments of some aspects of the present invention, the compositions of the present invention comprising plant cells expressing a recombinant TNF α polypeptide inhibitor can be used to prevent, treat and control diseases and conditions including obesity, metabolic syndrome and diabetes. In
15 general, the terms `prevent`, `control` and `treat` encompass the prevention of the development of a disease or a symptom from a patient who may have a predisposition of the disease or the symptom but has yet been diagnosed to have the disease or the symptom; the inhibition of the symptoms of a disease, namely, inhibition or retardation of the progression thereof; and the alleviation of the symptoms of a disease, namely,
20 regression of the disease or the symptoms, or inversion of the progression of the symptoms.

All types of obesity may be controlled or treated in accordance with the invention, including endogenous obesity, exogenous obesity, hyperinsular obesity, hyperplastic-hypertrophic obesity, hypertrophic obesity, hypothyroid obesity and
25 morbid obesity. However, inflammation-mediated obesity may be treated particularly effectively in accordance with the invention. By `prevent` or `control` or `treat` it is meant that body weight gain, specifically body fat gain, is slowed down, stopped or reversed, resulting in a maintenance or decrease in body weight. A decrease in weight or body fat may protect against cardiovascular disease by lowering blood pressure, total
30 cholesterol, LDL cholesterol and triglycerides, and may alleviate symptoms associated with chronic conditions such as hypertension, coronary heart disease, type 2 diabetes, hyperlipidemia, osteoarthritis, sleep apnea and degenerative joint disease.

Metabolic syndrome, or Syndrome X, is a complex multi-factorial condition accompanied by an assortment of abnormalities including hypertension, hypertriglyceridemia, hyperglycemia, low levels of HDL-C, and abdominal obesity. Individuals with these characteristics typically manifest a prothrombotic and pro-inflammatory state. Available data suggest that metabolic syndrome is truly a syndrome (a grouping of risk factors).

According to the World Health Organization (WHO) Guideline, metabolic syndrome is present if an individual manifests: a) hypertension (>140 mm Hg systolic or >90 mm Hg diastolic); (b) dyslipidemia, defined as elevated plasma triglycerides (150 mg/dL), and/or low high-density lipoprotein (HDL) cholesterol (<35 mg/dL in men, <39 mg/dL in women); 3) visceral obesity, defined as a high body mass index (BMI) (30 kg/m^2) and/or a high waist-to-hip ratio (>0.90 in men, >0.85 in women); and 4) microalbuminuria (urinary albumin excretion rate of 20 g/min). See WHO-International Society of Hypertension Guidelines for the Management of Hypertension. Guidelines Subcommittee. *J. Hypertens.* 17:151-183, 1999.

According to the National Cholesterol Education Program (NCEP ATP III study) metabolic syndrome is diagnosed if three (3) or more of the following five (5) risk factors are present: (1) a waist circumference >102 cm (40 in) for men or >88 cm (37 in) for women; (2) a triglyceride level of 150 mg/dL; (3) an HDL cholesterol level <40 mg/dL for men or <50 mg/dL for women; (4) blood pressure $>130/85$ mm Hg; or (5) a fasting glucose >110 mg/dL. *JAMA* 285: 2486-2497, 2001.

Each of the disorders associated with metabolic syndrome are risk factors in their own right, and can promote atherosclerosis, cardiovascular disease, stroke, and other adverse health consequences. However, when present together, these factors are predictive of increased risk of cardiovascular disease and stroke.

By `control` or `treat` it is meant that the symptoms of the metabolic syndrome shown in an individual are reduced in severity and/or in number. Such symptoms may include elevated blood glucose, glucose intolerance, insulin resistance, elevated triglycerides, elevated LDL-cholesterol, low high-density lipoprotein (HDL) cholesterol, elevated blood pressure, abdominal obesity, pro-inflammatory states, and pro-thrombotic states. By `prevent` or `control` or `treat` it is additionally or alternatively meant that the risk of developing associated diseases is reduced and/or the

onset of such diseases is delayed. Such associated diseases include cardiovascular disease, coronary heart disease and other diseases related to plaquing of the artery walls and diabetic conditions.

Diabetic conditions include, for example, type 1 diabetes, type 2 diabetes, gestational diabetes, pre-diabetes, slow onset autoimmune diabetes type 1 (LADA), hyperglycemia, and metabolic syndrome. For the purposes of treatment, the diabetes may be overt, diagnosed diabetes, e.g., type 2 diabetes, or a pre-diabetic condition.

Diabetes mellitus (generally referred to herein as "diabetes") is a disease that is characterized by impaired glucose regulation. Diabetes is a chronic disease that occurs when the pancreas fails to produce enough insulin or when the body cannot effectively use the insulin that is produced, resulting in an increased concentration of glucose in the blood (hyperglycemia). Diabetes may be classified as type 1 diabetes (insulin-dependent, juvenile, or childhood-onset diabetes), type 2 diabetes (non-insulin-dependent or adult-onset diabetes), LADA diabetes (late autoimmune diabetes of adulthood) or gestational diabetes. Additionally, intermediate conditions such as impaired glucose tolerance and impaired fasting glycemia are recognized as conditions that indicate a high risk of progressing to type 2 diabetes.

In type 1 diabetes, insulin production is absent due to autoimmune destruction of pancreatic beta-cells. There are several markers of this autoimmune destruction, detectable in body fluids and tissues, including islet cell autoantibodies, insulin autoantibodies, glutamic acid decarboxylase autoantibodies, and tyrosine phosphatase ICA512/IA-2 autoantibodies. In type 2 diabetes, comprising 90% of diabetics worldwide, insulin secretion may be inadequate, but peripheral insulin resistance is believed to be the primary defect. Type 2 diabetes is commonly, although not always, associated with obesity, a cause of insulin resistance.

Type 2 diabetes is often preceded by pre-diabetes, in which blood glucose levels are higher than normal but not yet high enough to be diagnosed as diabetes. The term "pre-diabetes," as used herein, is interchangeable with the terms "Impaired Glucose Tolerance" or "Impaired Fasting Glucose," which are terms that refer to tests used to measure blood glucose levels.

Chronic hyperglycemia in diabetes is associated with multiple, primarily vascular complications affecting microvasculature and/or macrovasculature. These

long-term complications include retinopathy (leading to focal blurring, retinal detachment, and partial or total loss of vision), nephropathy (leading to renal failure), neuropathy (leading to pain, numbness, and loss of sensation in limbs, and potentially resulting in foot ulceration and/or amputation), cardiomyopathy (leading to heart failure), and increased risk of infection. Type 2, or noninsulin-dependent diabetes mellitus (NIDDM), is associated with resistance of glucose-utilizing tissues like adipose tissue, muscle, and liver, to the physiological actions of insulin. Chronically elevated blood glucose associated with NIDDM can lead to debilitating complications including nephropathy, often necessitating dialysis or renal transplant; peripheral neuropathy; retinopathy leading to blindness; ulceration and necrosis of the lower limbs, leading to amputation; fatty liver disease, which may progress to cirrhosis; and susceptibility to coronary artery disease and myocardial infarction. By `prevent` it is meant that the risk of developing of diabetes is reduced or the onset of the disease is delayed. By `control` or `treat` it is meant that the risk of developing associated complications is reduced and/or the onset of such complications is delayed.

Diabetic conditions that are subject to treatment with plant cells expressing a recombinant TNF α polypeptide inhibitor according to the methods of the present invention can be diagnosed or monitored using any of a number of assays known in the field. Examples of assays for diagnosing or categorizing an individual as diabetic or pre-diabetic or monitoring said individual include, but are not limited to, a glycosylated hemoglobin (HbA1c) test, a connecting peptide (C-peptide) test, a fasting plasma glucose (FPG) test, an oral glucose tolerance test (OGTT), and a casual plasma glucose test.

HbA1c is a biomarker that measures the amount of glycosylated hemoglobin in the blood. HbA1c designates a stable minor glycated sub fraction of hemoglobin. It is a reflection of the mean blood glucose levels during the last 6-8 weeks, and is expressed in percent (%) of total hemoglobin. Alternatively, diabetes or pre-diabetes can be diagnosed by measuring blood glucose levels using any of several known tests in the field, including a fasting plasma glucose test or an oral glucose tolerance test. Using the fasting plasma glucose (FPG) test, a patient is classified as diabetic and is subject to treatment according to the methods of the present invention if the patient has a threshold FPG greater than 125 mg/dl, and a patient is classified as pre-diabetic and is subject to

treatment according to the methods of the present invention if the patient has a threshold FPG greater than 100 mg/dl but less than or equal to 125 mg/dl. Using the oral glucose tolerance test (OGTT), a patient is classified as diabetic and is subject to treatment according to the methods of the present invention if the patient has a threshold 2-hour OGTT glucose level greater than 200 mg/dl. A patient is classified as pre-diabetic and is subject to treatment according to the methods of the present invention if the patient has a threshold 2-hour OGTT glucose level greater than 140 mg/dl but less than 200 mg/dl.

C-peptide, produced from proinsulin molecules, is secreted from islet cells into the bloodstream in equimolar proportion as insulin, and is used a biomarker for beta-cell function and insulin secretion. A fasting C-peptide measurement greater than 2.0 ng/ml is indicative of high levels of insulin, while a fasting C-peptide measurement less than 0.5 ng/ml indicates insufficient insulin production.

A subject who has been classified as having a diabetic condition, and who is subject to treatment with plant cells expressing a recombinant TNF α polypeptide inhibitor according to the methods of the present invention, may be monitored for efficacy of treatment by measuring any of the biomarkers and/or blood glucose indicators described herein, including but not limited to, glycosylated hemoglobin levels, C-peptide levels, fasting plasma glucose levels, and oral glucose tolerance test (OGTT) levels. For the biomarkers and/or blood glucose indicators described herein, efficacy of treatment can be determined by quantitating the level of a biomarker or blood glucose indicator in a sample from a subject and determining whether the level of the biomarker or blood glucose indicator has reached or is approaching a threshold level. In some embodiments, a threshold level may correspond to a level of biomarker or blood glucose indicator that is a "normal" (i.e., non-diabetic) value according to standards known in the art, or a threshold level may correspond to a level of biomarker or blood glucose indicator that is a pre-diabetic or diabetic value according to standards known in the art.

In some embodiments, efficacy of treatment is determined by taking a first measurement of one or more of the biomarkers and/or blood glucose indicators in a subject prior to the start of treatment, and comparing the first measurement with secondary measurements of the same biomarker and/or blood glucose indicator in the subject at one or more time points after the onset of treatment, wherein a second

measurement that has reached or exceeded a threshold value (either above or below, depending on the biomarker being measured), or is closer to the threshold value than the first measurement is to the threshold value, indicates that the treatment is efficacious.

Alternatively or additionally, efficacy of treatment may be monitored by determining whether there has been an amelioration of the secondary conditions and symptoms that are associated with the diabetic condition. For example, a subject being treated by the methods of the present invention can be monitored for improvement or reduction in symptoms of retinopathy (e.g., improvement in vision), nephropathy (e.g., improvement in kidney structure or function), neuropathy (e.g., improvement in nerve function), and/or cardiovascular disease (e.g., decreased blood pressure or lower lipid levels).

Hyperlipidemia:

According to some embodiments of some aspects of the present invention, the compositions of the present invention comprising plant cells expressing a recombinant TNF α polypeptide inhibitor can be used to prevent, treat and control hyperlipidemia (also referred to as hyperlipoproteinemia, or hyperlipidaemia) which involves abnormally elevated levels of any or all lipids and/or lipoproteins in the blood.^[1] It is the most common form of dyslipidemia (which includes any abnormal lipid levels). Hyperlipidemias are also classified according to which types of lipids are elevated, that is hypercholesterolemia, hypertriglyceridemia or both in combined hyperlipidemia. Elevated levels of Lipoprotein(a) are also classified as a form of hyperlipidemia. Under the terms include are also, hyperlipoproteinemia Type I, hyperlipoproteinemia Type II, hyperlipoproteinemia Type III, hyperlipoproteinemia Type IV and hyperlipoproteinemia Type V. As well as unclassified familial forms and acquired forms of hyperlipidemia.

Liver Disease:

According to some embodiments of some aspects of the present invention, the compositions of the present invention comprising plant cells expressing a recombinant TNF α polypeptide inhibitor can be used to prevent, treat and control liver diseases and disorders including hepatitis, cirrhosis, non-alcoholic steatohepatitis (NASH) (also known as non-alcoholic fatty liver disease-NAFLD), hepatotoxicity and chronic liver disease. In general, the terms `prevent`, `control` and `treat` encompass the prevention

of the development of a disease or a symptom from a patient who may have a predisposition of the disease or the symptom but has yet been diagnosed to have the disease or the symptom; the inhibition of the symptoms of a disease, namely, inhibition or retardation of the progression thereof; and the alleviation of the symptoms of a disease, namely, regression of the disease or the symptoms, or inversion of the progression of the symptoms.

The term "liver disease" applies to many diseases and disorders that cause the liver to function improperly or to cease functioning, and this loss of liver function is indicative of liver disease. Thus, liver function tests are frequently used to diagnose liver disease. Examples of such tests include, but are not limited to, the following;

(1) Assays to determine the levels of serum enzymes such as lactate dehydrogenase (LDH), alkaline phosphatase (ALP), aspartate aminotransferase (AST), and alanine aminotransferase (ALT), where an increase in enzyme levels indicates liver disease. One of skill in the art will reasonably understand that these enzyme assays indicate only that the liver has been damaged. They do not assess the liver's ability to function. Other tests can be used to assay a liver's ability to function;

(2) Assays to determine serum bilirubin levels. Serum bilirubin levels are reported as total bilirubin and direct bilirubin. Normal values of total serum bilirubin are 0.1-1.0 mg/dl (e.g., about 2-18 mmol/L). Normal values of direct bilirubin are 0.0-0.2 mg/dl (0-4 mmol/L). Increases in serum bilirubin are indicative of liver disease.

(3) Assays to determine serum protein levels, for example, albumin and the globulins (e.g., alpha, beta, gamma). Normal values for total serum proteins are 6.0-8.0 g/dl (60-80 g/L). A decrease in serum albumin is indicative of liver disease. An increase in globulin is indicative of liver disease.

Other tests include prothrombin time, international normalized ratio, activated clotting time (ACT), partial thromboplastin time (PTT), prothrombin consumption time (PCT), fibrinogen, coagulation factors; alpha-fetoprotein, and alpha-fetoprotein-L3 (percent).

One clinically important type of liver disease is hepatitis. Hepatitis is an inflammation of the liver that can be caused by viruses (e.g., hepatitis virus A, B and C (HAV, HBV, and HCV, respectively)), chemicals, drugs, alcohol, inherited diseases, or the patient's own immune system (autoimmune hepatitis). This inflammation can be

acute and resolve within a few weeks to months, or chronic, and persist over many years. Chronic hepatitis can persist for decades before causing significant symptoms, such as cirrhosis (scarring and loss of function), liver cancer, or death. Other important examples of the different diseases and disorders encompassed by the term "liver disease" and suitable for treatment or prevention or control using the compositions and methods of the present invention include, but are not limited to amebic liver abscess, biliary atresia, fibrosis, cirrhosis, coccidioidomycosis, delta agent, hepatocellular carcinoma (HCC), alcoholic liver disease, primary biliary cirrhosis, pyogenic liver abscess, Reye's syndrome, sclerosing cholangitis, and Wilson's disease. In some embodiments, the compositions and methods described herein are suitable for the treatment of liver disease characterized by the loss or damage of parenchymal liver cells. In some aspects, the etiology of this can be a local or systemic inflammatory response.

Liver failure occurs when large parts of the liver become damaged and the liver is no longer able to perform its normal physiological function. In some aspects, liver failure can be diagnosed using the above described assays of liver function or by a subject's symptoms. Symptoms that are associated with liver failure include, for example, one or more of the following, nausea, loss of appetite, fatigue, diarrhea, jaundice, abnormal/excessive bleeding (e.g., coagulopathy), swollen abdomen, mental disorientation or confusion (e.g., hepatic encephalopathy), sleepiness, and coma.

Chronic liver failure occurs over months to years and is most commonly caused by viruses (e.g., HBV and HCV), long-term/excessive alcohol consumption, cirrhosis, hemochromatosis, and malnutrition. Acute liver failure is the appearance of severe complications after the first signs of liver disease (e.g., jaundice) and includes a number of conditions, all of which involve severe hepatocyte injury or necrosis. In some embodiments, the compositions and methods described herein are particularly suitable for the treatment of hyperacute, acute, and subacute liver failure, fulminant hepatic failure and late onset fulminant hepatic failure, all of which are referred to herein as "acute liver failure." Common causes for acute liver failure include, for example, viral hepatitis, exposure to certain drugs and toxins (e.g., fluorinated hydrocarbons (e.g., trichloroethylene and tetrachloroethane), amanita phalloides (e.g., commonly found in the "death-cap mushroom"), acetaminophen (paracetamol), haloethanes, sulfonamides,

henytoins), cardiac-related hepatic ischemia (e.g., myocardial infarction, cardiac arrest, cardiomyopathy, and pulmonary embolism), renal failure, occlusion of hepatic venous outflow (e.g., Budd-Chiari syndrome), Wilson's disease, acute fatty liver of pregnancy, amebic abscesses, and disseminated tuberculosis.

5 The term "hepatitis" is used to describe a liver condition which implies injury to the liver characterized by the presence of inflammatory cells in the tissue of the organ. The condition can be self-limiting, healing on its own, or can progress to scarring of the liver. Hepatitis is acute when it lasts less than six months and chronic when it persists longer than six months. A group of viruses known as the hepatitis viruses cause most
10 cases of liver damage worldwide. Hepatitis can also be due to toxins (notably alcohol), other infections or from autoimmune process. Hepatitis includes hepatitis from viral infections, including Hepatitis A through E (A, B, C, D and E--more than 95% of viral cause), Herpes simplex, Cytomegalovirus, Epstein-Barr virus, yellow fever virus, adenoviruses; non-viral infections, including toxoplasma, Leptospira, Q fever, rocky
15 mountain spotted fever, alcohol, toxins, including amanita toxin in mushrooms, carbon tetrachloride, asafetida, among others, drugs, including paracetamol, amoxycillin, antituberculosis medicines, minocycline and numerous others as described herein; ischemic hepatitis (circulatory insufficiency); pregnancy; autoimmune conditions, including Systemic Lupus Erythematosus (SLE); and non-alcoholic steatohepatitis.

20 "Sterile inflammation" is used to describe inflammation of the liver which is triggered by intracellular molecules released from dying cells that have lost integrity of their plasma membrane. This inflammation occurs in the absence of causative agents such as viruses or bacteria and alcohol. A number of intracellular molecules have been identified that can stimulate other cells to produce proinflammatory cytokines and
25 chemokines. Such proinflammatory cellular molecules are thought to function by engaging receptors on cytokine-producing cells. If left untreated, sterile inflammation may progress to non-alcoholic fatty liver disease (NAFLD), non-alcoholic steatohepatitis (NASH) or cyrrhosis.

30 "Non-alcoholic steatohepatitis" or "NASH" is a condition of the liver in which inflammation is caused by a buildup of fat in the liver. NASH is part of a group of liver diseases, known as nonalcoholic fatty liver disease, in which fat builds up in the liver and sometimes causes liver damage that gets worse over time (progressive liver

damage). "Non-alcoholic fatty liver disease" (NAFLD) is fatty inflammation of the liver which is not due to excessive alcohol use. It is related to insulin resistance and the metabolic syndrome, obesity, high cholesterol and triglycerides, and diabetes., and may respond to treatments originally developed for other insulin resistant states (e.g. diabetes mellitus type 2), such as weight loss, metformin and thiazolidinediones. Non-alcoholic steatohepatitis (NASH) is the most extreme form of NAFLD, which is regarded as a major cause of cirrhosis of the liver of unknown cause.

Other factors that have been known to contribute to NASH include: surgery that shorten the intestines, the stomach, or both, such as jejunal bypass operation or biliopancreatic diversion; prolonged use of feeding tube or other method of receiving nutrition; certain drugs, including amiodarone, glucocorticoids, synthetic estrogens, and tamoxifen.

NASH is a condition that may get worse over time (called a progressive condition) and can cause scarring (fibrosis) of the liver, which leads to cirrhosis. "Cirrhosis" describes a condition in which liver cells have been replaced by scar tissue. The term "cirrhosis of the liver" or "cirrhosis" is used to describe a chronic liver disease characterized by replacement of liver tissue by fibrous scar tissue as well as regenerative nodules, leading to progressive loss of liver function. Cirrhosis is most commonly caused by fatty liver disease, including NASH, as well as alcoholism and hepatitis B and C, but also may be of unknown cause. Potentially life-threatening complications of cirrhosis are hepatic encephalopathy (confusion and coma) and bleeding from esophageal varices. Cirrhosis has historically been thought to be generally irreversible once it occurs, and historical treatment focused on preventing progression and complications. In advanced stages of cirrhosis, the only option is a liver transplant. The plant cells expressing a recombinant TNF α polypeptide inhibitor and methods of the present invention may be used to limit, inhibit, reduce the likelihood or treat cirrhosis of the liver without regard to its etiology.

The plant cells expressing a recombinant TNF α polypeptide inhibitor and methods of the present invention can be used to treat, prevent or control chemical liver trauma and hepatotoxicity. "Chemical trauma" or "acute chemical trauma" refers to serious injury which occurs to a patient over a short duration as a consequence of chemical toxicity, including drug-induced toxicity or trauma. Drug-induced acute liver

trauma, including acetaminophen-induced acute liver trauma, is acute liver injury which occurs as a result or consequence of exposure to a drug (e.g., drug overdose), especially acetaminophen toxicity. Compounds according to the present invention are useful for reducing the injury to the liver which occurs from physical and chemical trauma, especially including drug-induced (drug overdose) and acetaminophen-induced acute liver trauma.

Hepatotoxicity is chemical liver trauma resulting from a hepatotoxic agent, or hepatotoxicity-inducing bioactive agent. The terms "hepatotoxic agent" and "a hepatotoxicity inducing bioactive agent" are used synonymously in context to describe compounds which often produce hepatotoxicity in patients administered such agents. Examples of hepatotoxicity agents include, for example, anaesthetic agents, antiviral agents, anti-retroviral agents (nucleoside reverse transcriptase inhibitors and non-nucleoside reverse transcriptase inhibitors), especially anti-HIV agents, anticancer agents, organ transplant drugs (cyclosporin, tacrolimus, OKT3), antimicrobial agents (anti-TB, anti-fungal, antibiotics), anti-diabetes drugs, vitamin A derivatives, steroidal agents, especially including oral contraceptives, anabolic steroids, androgens, non-steroidal anti-inflammatory agents, anti-depressants (especially tricyclic antidepressants) glucocorticoids, natural products and herbal and alternative remedies, especially including St. John's wort.

Hepatotoxicity may manifest as triglyceride accumulation which leads to either small droplet (microvesicular) or large droplet (macrovesicular) fatty liver. There is a separate type of steatosis where phospholipid accumulation leads to a pattern similar to the diseases with inherited phospholipid metabolism defects (e.g. Tay-Sachs disease).

According to a specific embodiment, the liver disease is a fatty liver disease (e.g., non-alcoholic). In this case and according to some embodiments, the TNF α inhibitor (e.g., orally administered plant cells expressing recombinant TNFR2:Fc) causes a reduction in serum enzymes (e.g., AST or ALT or both) and/or triglycerides and can alternatively or additionally alter the distribution of T cells and NK cells in the liver and spleen, as compared to that of an untreated subject in the same disease stage.

The plant cells expressing a recombinant TNF α polypeptide inhibitor and methods of the present invention can be used to treat, prevent or control chronic liver disease. Chronic liver disease is marked by the gradual destruction of liver tissue over

time. Several liver diseases fall under this category, including cirrhosis and fibrosis, the latter of which is often the precursor to cirrhosis. Cirrhosis is the result of acute and chronic liver disease and is characterized by the replacement of liver tissue by fibrotic scar tissue and regenerative nodules leading to a progressive loss of liver function.

5 Fibrosis and nodular regeneration results in the loss of the normal microscopic lobular architecture of the liver. Fibrosis represents the growth of scar tissue resulting from, for example, infection, inflammation, injury, and even healing. Over time, the fibrotic scar tissue slowly replaces the normal functioning liver tissue resulting in a decreasing amount of blood flow to the liver leaving the liver incapable of fully processing
10 nutrients, hormones, drugs, and poisons that are found in the bloodstream. More common causes of cirrhosis include alcoholism, hepatitis C viral infections, ingestion of toxins, and fatty liver, but many other possible causes also exist. Chronic hepatitis C virus (HCV) infection and non-alcoholic steatohepatitis (NASH) are the two major causes of chronic liver disease in the United States estimated to affect between 3-5
15 million people. A rising concern is the continuously increasing number of U.S. citizens, currently numbering over 30 million, with obesity and metabolic syndrome that have non-alcoholic fatty liver disease (NAFLD) with approximately 10% who will eventually develop NASH. Other bodily complications are a consequence of a loss of liver function. The most common complication of cirrhosis is a condition known as ascites,
20 an accumulation of fluid in the peritoneal cavity, which can lead to an increased risk of spontaneous bacterial peritonitis possibly resulting in the premature death of the patient.

The plant cells expressing a recombinant TNF α polypeptide inhibitor and methods of the present invention may be used to limit, inhibit, reduce the likelihood or treat cancer of the liver. Risk factors for liver cancer include type 2 diabetes
25 (exacerbated by obesity) and metabolic syndrome. The risk of liver cancer in type 2 diabetics is greater (about 3 to 7 times the non-diabetic risk) depending on the duration of diabetes and treatment protocol. Metabolic syndrome results in inflammation, steatosis, fibrosis, cirrhosis, apoptosis, altered gene expression and eventually even liver cancer. In addition, lipid metabolism abnormality, hypertension, hyperglycemia and
30 metabolic syndrome, exacerbating hepatitis and the progress of hepatitis to cirrhosis, which can further lead to liver cancer, for example, by the activation of stellate cells.

Various methodologies can be used in the screening and diagnosis of liver cancer and are well known in the art. Indicators for liver cancer include tumor markers such as elevated alpha-fetoprotein (AFP) or des-gamma carboxyprothrombin (DCP). Scanning and imaging techniques are also helpful, including ultrasound, CT scans and
5 MRI. Macroscopically, liver cancer may be nodular, or an infiltrative tumor which is diffuse and poorly circumscribed.

As used the term "TNF α " refers to Tumor necrosis factor-alpha (TNF, cachexin, or cachectin) that is a cytokine involved in systemic inflammation and a member of a group of cytokines that stimulate the acute phase reaction. TNF α is produced primarily
10 by activated macrophages (M1), although it can be produced by many other cell types as CD4+ lymphocytes, NK cells and neurons. The protein is encoded by TNFA gene and has the Ref_seq number: NP_000585. The protein is known to stimulate an inflammatory response (pro-inflammatory cytokine).

A "TNF α polypeptide inhibitor" as used herein refers to a polypeptide that binds
15 TNF α and inhibits and/or hinders TNF α activity as reflected in TNF α binding to a TNF α -receptor (TNFR) including any of the following: (a) TNFR, preferably endogenous (i.e., native to the individual or host), cell membrane bound TNFR; (b) the extracellular domain(s) of TNFR; and/or (c) the TNF α binding domains of TNFR (which may be a portion of the extracellular domain). According to a specific embodiment,
20 inhibition of TNF α binding to the receptor is by at least 50 %, e.g., 50-100 %. 50-95 %, 60-90 % or even 70-90 %.

TNF α inhibitors include, but are not limited to, TNF α receptors (or appropriate portions thereof, as described herein) and anti-TNF α antibodies.

As used herein, the "biological activity" of a TNF α inhibitor is to bind to TNF α
25 and inhibit and/or hinder TNF α from binding to any of the following: (a) TNFR, preferably endogenous, cell membrane bound TNFR; (b) the extracellular domain(s) of TNFR; and (c) the TNF α binding domains of TNFR (which may be a portion of the extracellular domain). A TNF α inhibitor can be shown to exhibit biological activity using assays known in the art to measure TNF α activity and its inhibition, an example of
30 which is provided herein.

As used herein, the terms "TNF receptor polypeptide" and "TNFR" refer to polypeptides derived from TNFR (from any species e.g., human) which are capable of

binding TNF α . Two distinct cell-surface TNFRs have been described: Type II TNFR (or p75 TNFR or TNFR_{II}) and Type I TNFR (or p55 TNFR or TNFR_I). The mature full-length human p75 TNFR is a glycoprotein having a molecular weight of about 75-80 kilodaltons (kD). The mature full-length human p55 TNFR is a glycoprotein having a molecular weight of about 55-60 kD. The preferred TNFR polypeptides of this invention are derived from TNFR Type I and/or TNFR type II. Exemplary accession numbers are provided hereinbelow. According to a specific embodiment, the TNFR is capable of binding TNF α in a specific manner e.g., K_d below 10⁻⁵ M.

According to a specific embodiment, the TNF α inhibitor is a chimeric polypeptide.

A "chimeric polypeptide" or "fusion polypeptide" is a polypeptide comprising regions in a different position than occurs in nature. The regions may normally exist in separate proteins and are brought together in the chimeric or fusion polypeptide, or they may normally exist in the same protein but are placed in a new arrangement in the chimeric or fusion polypeptide. A chimeric or fusion polypeptide may also arise from polymeric forms, whether linear or branched, of TNFR polypeptide(s).

As used herein, an "extracellular domain" of TNFR refers to a portion of TNFR that is found between the amino-terminus of TNFR and the amino-terminal end of the TNFR transmembrane region. The extracellular domain of TNFR binds TNF α .

The terms "polypeptide", "peptide" and "protein" are used interchangeably herein to refer to polymers of amino acids of any length. The terms also encompass an amino acid polymer that has been modified; for example, disulfide bond formation, glycosylation, lipidation, or conjugation with a labeling component.

Specific examples of TNF α polypeptide inhibitors include, but are not limited to, infliximab (RemicadeTM) and adalimumab (HumiraTM), which consist of, chimeric human-mouse anti-TNF α monoclonal antibodies and fully human anti-TNF-alpha monoclonal antibodies, respectively. Another example of an anti-TNF α antibody which can be used in accordance with the present teachings include golimumab (SimponiTM).

Also included under this definition are chimeric polypeptides which include the commercial etanercept (further described hereinbelow) and Lenercept (a chimeric polypeptide consisting of p55sTNF-RI-IgG1) under their scope. Such chimeric polypeptides are further described hereinbelow.

Thus, according to a specific embodiment the TNF α polypeptide inhibitor is a chimeric polypeptide comprising:

- (i) a first domain which comprises a TNF α binding domain of a TNF receptor; and
 - (ii) a second domain which comprises an Fc domain of an immunoglobulin, wherein
- 5 the first domain and the second domain are N-terminally to C-terminally respectively sequentially translationally fused and wherein the chimeric polypeptide specifically binds TNF α .

The first domain is thus composed of at least the TNF binding domain of a TNF receptor (TNFR). The first domain is a soluble protein. Thus according to a specific

10 embodiment, the first domain and even the entire chimeric polypeptide are soluble proteins which are not membrane anchored.

Soluble forms of TNFRs may include monomers, fusion proteins (also called "chimeric proteins), dimers, trimers or higher order multimers. In certain embodiments of the invention, the soluble TNFR derivative is one that mimics the 75 kDa TNFR or

15 the 55 kDa TNFR and that binds to TNF α . *in vivo*. The soluble TNFR mimics of the present invention may be derived from TNFRs p55 or p75 or fragments thereof. TNFRs other than p55 and p75 also are useful for deriving soluble TNFR for treating the various medical disorders described herein, such for example the TNFR that is described in WO 99/04001. Soluble TNFR molecules used to construct TNFR mimics include, for

20 example, analogs or fragments of native TNFRs having at least 20 amino acids, that lack the transmembrane region of the native TNFR, and that are capable of binding TNF α . Such soluble forms of TNFR compete for TNF α with the receptors on the cell surface, thus inhibiting TNF α from binding to cells, thereby preventing it from manifesting its biological activities. Binding of soluble TNFRs to TNF α can be assayed using ELISA or

25 any other convenient assay.

According to a specific embodiment, the first domain is derived from TNFR2. (e.g., AAA36755).

According to an embodiment of the invention, the first domain is 200-250 amino acids long.

30 According to a specific embodiment, the first domain comprises the amino acid sequence LCAP (SEQ ID NO: 11) and VFCT (SEQ ID NO: 12).

According to a specific embodiment, the first domain comprises the amino acid sequence LPAQVAFXPYAPEPGSTC (SEQ ID NO: 13) or LPAQVAFTPYAPEPGSTC (SEQ ID NO: 17)

According to a specific embodiment, the first domain is as set forth in SEQ ID NO: 2 (encoded by SEQ ID NO: 1).

As used herein “an Fc domain of an immunoglobulin” refers to a region of a heavy chain of an antibody, typically comprising at least 2 constant domains (e.g., CH2 and CH3 domains, as these terms are defined in the art) of the heavy chain. The Fc domain may be obtained, for example, in the form of a dimer, by digestion of an antibody by papain. A dimer of Fc domain polypeptides, connected by disulfide bonds, forms the “tail” region of an antibody. As is known in the art, Fc domains of some classes of antibodies may be in the form of multimers. Thus, the Fc domain is optionally monomeric, optionally dimeric and optionally multimeric. Optionally, the polypeptide described herein is in the form of a dimer, the polypeptide comprising an Fc dimer, or in the form of a multimer, the polypeptide comprising an Fc multimer.

The Fc domain may encompass modified forms of a native Fc domain (i.e., a domain which occurs naturally in an antibody), for example, polypeptides having at least 90 % homology, optionally at least 95 % homology, and optionally at least 98 % homology, to a native Fc domain. Modified Fc domains are described, for example, in International Patent Applications WO 97/34631 and WO 96/32478.

Optionally, a native Fc is modified so as to remove sites which provide structural features or biological activity that are not required for embodiments of the present invention. Examples of such sites include residues that affect or are involved in disulfide bond formation, incompatibility with a selected host cell, N-terminal heterogeneity upon expression in a selected host cell, glycosylation, interaction with complement, binding to an Fc receptor (other than a neonatal Fc receptor), and/or antibody-dependent cellular cytotoxicity.

The polypeptide according to embodiments of the present invention may also comprise a fragment of an Fc domain. Optionally, the fragment comprises at least 20 %, optionally at least 50 %, and optionally at least 80 % of an Fc domain, as defined hereinabove.

The Fc domain or fragment thereof optionally includes a binding site for a neonatal Fc receptor (FcRn). This is of particular significance when administering the chimeric polypeptide via an enteral route.

According to one embodiment, attachment of an Fc domain or a fragment thereof to the first domain results in a polypeptide having a longer half-life *in vivo* than the first domain *per se*. This may be due to the long serum half-life of the Fc domain (which may be due to salvage of the Fc via binding to FcRn) and/or due to the greater size of the polypeptide in comparison to the first domain *per se*, which reduces clearance from the bloodstream by glomerular filtration. According to another embodiment, the resulting polypeptides have reduced immunogenicity as compared to the first domain *per se*.

According to optional embodiments, the Fc domain or fragment thereof is a human Fc domain (e.g., derived from a human antibody) or fragment thereof.

According to exemplary embodiments, the Fc domain (or fragment thereof) is an IgG (e.g., IgG1) Fc domain (or fragment thereof).

According to a specific embodiment, the second domain is as set forth in SEQ ID NO: 9 (encoded by SEQ ID NO: 8).

Thus, the second domain of the chimeric polypeptide comprises at least a portion of a constant immunoglobulin domain, e.g. a constant heavy immunoglobulin domain or a constant light immunoglobulin domain. Preferably, the second domain comprises at least a portion of a constant heavy immunoglobulin domain. The constant heavy immunoglobulin domain is preferably an Fc fragment comprising the CH2 and CH3 domain and, optionally, at least a part of the hinge region. The immunoglobulin domain may be an IgG, IgM, IgD or IgE immunoglobulin domain or a modified immunoglobulin domain derived, therefrom. Preferably, the second domain comprises at least a portion of a constant IgG immunoglobulin domain. The IgG immunoglobulin domain may be selected from IgG1, IgG2, IgG3 or IgG4 domains or from modified domains such as are described in U.S. Pat. No. 5,925,734. The immunoglobulin domain may exhibit effector functions. In some embodiments, however, modified immunoglobulin domains having modified, e.g. at least partially deleted, effector functions may be used. Thus for example, the receptor.

According to an embodiment of the invention, the chimeric fusion of the first domain and the second domain forms Etanercept (Immunex) having SEQ ID NO: 10.

It will be appreciated that the species origin of the first domain and the second domain is selected according to the treated subject. Thus, according to a specific embodiment, the first domain and the second domain are of human origin or modified such that they don't incur immunogenic reaction when administered to human subjects.

As used herein "Etanercept" and "EnbrelTM" are interchangeably used to designate the commercially available TNFR2:Fc by Immunex Corporation. Etanercept is a dimeric fusion polypeptide consisting of the extracellular ligand-binding portion of the human 75 kilodalton (p75) tumor necrosis factor receptor (TNFR) linked to the Fc portion of human IgG1. The Fc component of etanercept contains the constant heavy 2 (CH2) domain, the constant heavy 3 (CH3) domain and hinge region, but not the constant heavy 1 (CH1) domain of human IgG1. Plant cells expressing TNFR2:Fc are also termed PRX-106.

According to another embodiment, the chimeric polypeptide comprises:

- (i) a first domain which comprises a TNF α binding domain of a TNF receptor;
- (ii) a second domain which comprises an Fc domain of an immunoglobulin; and
- (iii) a third domain comprising an endoplasmic reticulum retention signal;

wherein the first domain, second domain and third domain are N-terminally to C-terminally respectively sequentially translationally fused and wherein the chimeric polypeptide specifically binds TNF α .

Thus, according to this aspect of the invention, the chimeric protein is expressed such that it is retained in the endoplasmic reticulum. According to a specific embodiment, at least a portion of the TNFR2:Fc molecules (e.g., at least 20 %) in the cell are retained in the ER.

As used herein, the term "endoplasmic reticulum retention signal peptide" refers to a peptide sequence which, when present at the N- or C- terminus of a polypeptide, causes the polypeptide to be retrieved from the Golgi apparatus, and retained in the endoplasmic reticulum (see Rayon et al. Journal of Experimental Botany, Vol. 49, No. 326, pp. 1463–1472, 1998; and Neumann, et al Annals of Botany, 2003;92:167-180). In one embodiment, the endoplasmic reticulum retention signal peptide is HDEL (SEQ ID NO: 14), KDEL (SEQ ID NO: 15) or SEKDEL (SEQ ID NO: 16).

As mentioned, the first domain and second domain (and third domain when present) are N-terminally to C-terminally respectively sequentially translationally fused. This means that the first domain is located N-terminally to the second domain (the carboxy terminus of the first domain is translationally fused to the N-terminus of the second domain), and the second domain is located N-terminally of the third domain (the carboxy terminus of the second domain is translationally fused to the N-terminus of the third domain). Thus, the second domain is practically sandwiched by the first domain at the N-terminus and the third domain at the C-terminus. Schematic presentation is as follows: first domain>second domain(>third domain) are orderly oriented from the N-terminus to the C-terminus (see Figure 1). The linkage between the domains may be direct or indirect by the use of linkers such as peptide linkers.

The molecule may further comprise an additional domain which encodes for an endoplasmic reticulum signal sequence which is oriented upstream (N-terminally) of the first domain and translationally fused thereto.

As used herein "an endoplasmic reticulum (ER) signal peptide" refers to a signal sequence, leader sequence or leader peptide that is a short (e.g., 5-30 amino acids long) peptide present at the N-terminus of the majority of newly synthesized proteins that are destined towards the secretory pathway.

According to a specific embodiment, the ER signal peptide is derived (taken, truncated) from a plant protein.

According to a specific embodiment, the endoplasmic reticulum signal peptide is from *N. plumbaginifolia* Calreticulin protein.

According to a further specific embodiment, the signal peptide from *N. plumbaginifolia* Calreticulin protein is as set forth in SEQ ID NO: 4 and encoded by the nucleic acid sequence of SEQ ID NO: 3.

As used herein the term "translationally fused at the N-terminal" or "translationally fused at the C-terminal" refers to covalent attachment of the indicated peptide via a peptide bond to the N-terminal or C-terminal amino acid of the respective domain typically as a result of recombinant expression.

According to a specific embodiment, the chimeric polypeptide is as set forth in SEQ ID NO: 6.

According to a specific embodiment, the chimeric polypeptide is as set forth in SEQ ID NO: 7, 204 or 205.

As mentioned the recombinant chimeric proteins of the invention are produced in plant cells.

5 In order to express the polypeptide, an isolated polynucleotide comprising a nucleic acid sequence encoding the chimeric polypeptide as described herein is ligated into a "plant nucleic acid expression construct".

As used herein the term "plant nucleic acid expression construct" refers to a nucleic acid construct which includes the nucleic acid of some embodiments of the invention and at least one promoter for directing transcription of nucleic acid in a host
10 plant cell. Further details of suitable transformation approaches are provided hereinbelow.

According to some embodiments of the invention, there is provided a nucleic acid expression construct comprising the nucleic acid sequence of the invention, and a
15 promoter for directing transcription of the nucleic acid sequence in a plant host cell.

As used herein the term "nucleic acid sequence" refers to a single or double stranded nucleic acid sequence which is isolated and provided in the form of an RNA sequence, a complementary polynucleotide sequence (cDNA), a genomic polynucleotide sequence and/or a composite polynucleotide sequences (e.g., a combination of the
20 above).

As used herein the phrase "complementary polynucleotide sequence" refers to a sequence, which results from reverse transcription of messenger RNA using a reverse transcriptase or any other RNA dependent DNA polymerase. Such a sequence can be subsequently amplified *in vivo* or *in vitro* using a DNA dependent DNA polymerase.

25 As used herein the phrase "genomic polynucleotide sequence" refers to a sequence derived (isolated) from a chromosome and thus it represents a contiguous portion of a chromosome.

As used herein the phrase "composite polynucleotide sequence" refers to a sequence, which is at least partially complementary and at least partially genomic. A
30 composite sequence can include some exonal sequences required to encode the polypeptide of the present invention, as well as some intronic sequences interposing therebetween. The intronic sequences can be of any source, including of other genes, and

typically will include conserved splicing signal sequences. Such intronic sequences may further include cis acting expression regulatory elements.

According to some embodiments of the present invention, the nucleic acid sequences encoding the polypeptides of the present invention are optimized for expression in plants. Examples of such sequence modifications include, but are not limited to, an altered G/C content to more closely approach that typically found in the plant species of interest, and the removal of codons atypically found in the plant species commonly referred to as codon optimization. In one embodiment, the codon usage of the nucleic acid sequence encoding the chimeric polypeptide is optimized for *Nicotiana tabacum* or *Nicotiana benthamiana*.

The phrase "codon optimization" refers to the selection of appropriate DNA nucleotides for use within a structural gene or fragment thereof that approaches codon usage within the plant of interest. Therefore, an optimized gene or nucleic acid sequence refers to a gene in which the nucleotide sequence of a native or naturally occurring gene has been modified in order to utilize statistically-preferred or statistically-favored codons within the plant. The nucleotide sequence typically is examined at the DNA level and the coding region optimized for expression in the plant species determined using any suitable procedure, for example as described in Sardana et al. (1996, Plant Cell Reports 15:677-681). In this method, the standard deviation of codon usage, a measure of codon usage bias, may be calculated by first finding the squared proportional deviation of usage of each codon of the native gene relative to that of highly expressed plant genes, followed by a calculation of the average squared deviation. The formula used is: $SDCU = \sqrt{\frac{1}{N} \sum_{n=1}^N \left(\frac{X_n - Y_n}{Y_n} \right)^2}$, where X_n refers to the frequency of usage of codon n in highly expressed plant genes, where Y_n to the frequency of usage of codon n in the gene of interest and N refers to the total number of codons in the gene of interest. A table of codon usage from highly expressed genes of dicotyledonous plants has been compiled using the data of Murray et al. (1989, Nuc Acids Res. 17:477-498).

One method of optimizing the nucleic acid sequence in accordance with the preferred codon usage for a particular plant cell type is based on the direct use, without performing any extra statistical calculations, of codon optimization tables such as those provided on-line at the Codon Usage Database through the NIAS (National Institute of Agrobiological Sciences) DNA bank in Japan (Hypertext Transfer Protocol://World

Wide Web (dot) kazusa (dot) or (dot) jp/codon/). The Codon Usage Database contains codon usage tables for a number of different species, with each codon usage table having been statistically determined based on the data present in Genbank.

By using such codon optimization tables to determine the most preferred or most favored codons for each amino acid in a particular species (for example, rice), a naturally-occurring nucleotide sequence encoding a protein of interest can be codon optimized for that particular plant species. This is effected by replacing codons that may have a low statistical incidence in the particular species genome with corresponding codons, in regard to an amino acid, that are statistically more favored. However, one or more less-favored codons may be selected to delete existing restriction sites, to create new ones at potentially useful junctions (5' and 3' ends to add signal peptide or termination cassettes, internal sites that might be used to cut and splice segments together to produce a correct full-length sequence), or to eliminate nucleotide sequences that may negatively affect mRNA stability or expression.

The desired encoding nucleotide sequence may already, in advance of any modification, contain a number of codons that correspond to a statistically-favored codon in a particular plant species. Therefore, codon optimization of the native nucleotide sequence may comprise determining which codons, within the desired nucleotide sequence, are not statistically-favored with regards to a particular plant, and modifying these codons in accordance with a codon usage table of the particular plant to produce a codon optimized derivative. A modified nucleotide sequence may be fully or partially optimized for plant codon usage provided that the protein encoded by the modified nucleotide sequence is produced at a level higher than the protein encoded by the corresponding naturally occurring or native gene. Construction of synthetic genes by altering the codon usage is described in for example PCT Patent Application 93/07278.

Thus according to a specific embodiment, there is provided a *Nicotinia tobaccum* optimized sequence as set forth in SEQ ID NO: 5.

According to some embodiments of the invention, the nucleic acid sequence coding for the cimeric polypeptide is operably linked to a cis-acting regulatory sequence active in plant cells, such as a plant promoter sequence.

A coding nucleic acid sequence is “operably linked” to a regulatory sequence (e.g., promoter) if the regulatory sequence is capable of exerting a regulatory effect on (e.g. effect on the expression of) the coding sequence linked thereto.

Any suitable promoter sequence can be used by the nucleic acid construct of the present invention. Preferably the promoter is a constitutive promoter, a tissue-specific, or an inducible promoter.

As used herein the phrase "plant-expressible" refers to a promoter sequence, including any additional regulatory elements added thereto or contained therein, is at least capable of inducing, conferring, activating or enhancing expression in a plant cell, tissue or organ, preferably a monocotyledonous or dicotyledonous plant cell, tissue, or organ. Such a promoter can be constitutive, i.e., capable of directing high level of gene expression in a plurality of tissues, tissue specific, i.e., capable of directing gene expression in a particular tissue or tissues, inducible, i.e., capable of directing gene expression under a stimulus, or chimeric, i.e., formed of portions of at least two different promoters.

Examples of preferred promoters useful for the methods of some embodiments of the invention are presented in Table I, II, III and IV.

Table I
Exemplary constitutive promoters for use in the performance of some embodiments of the invention

<i>Gene Source</i>	<i>Expression Pattern</i>	<i>Reference</i>
Actin	constitutive	McElroy et al, Plant Cell, 2: 163-171, 1990
CAMV 35S	constitutive	Odell et al, Nature, 313: 810-812, 1985
CaMV 19S	constitutive	Nilsson et al., Physiol. Plant 100:456-462, 1997
GOS2	constitutive	de Pater et al, Plant J Nov;2(6):837-44, 1992
ubiquitin	constitutive	Christensen et al, Plant Mol. Biol. 18: 675-689, 1992
Rice cyclophilin	constitutive	Bucholz et al, Plant Mol Biol. 25(5):837-43, 1994
Maize H3 histone	constitutive	Lepetit et al, Mol. Gen. Genet. 231: 276-285, 1992
Actin 2	constitutive	An et al, Plant J. 10(1);107-121, 1996

Table II
Exemplary seed-preferred promoters for use in the performance of some
embodiments of the invention

<i>Gene Source</i>	<i>Expression Pattern</i>	<i>Reference</i>
Seed specific genes	seed	Simon, et al., Plant Mol. Biol. 5. 191, 1985; Scofield, et al., J. Biol. Chem. 262: 12202, 1987.; Baszczynski, et al., Plant Mol. Biol. 14: 633, 1990.
Brazil Nut albumin	seed	Pearson' et al., Plant Mol. Biol. 18: 235- 245, 1992.
legumin	seed	Ellis, et al.Plant Mol. Biol. 10: 203-214, 1988
Glutelin (rice)	seed	Takaiwa, et al., Mol. Gen. Genet. 208: 15-22, 1986; Takaiwa, et al., FEBS Letts. 221: 43-47, 1987
Zein	seed	Matzke et al Plant Mol Biol, 143).323-32 1990
napA	seed	Stalberg, et al, Planta 199: 515-519, 1996
wheat LMW and HMW, glutenin-1	endosperm	Mol Gen Genet 216:81-90, 1989; NAR 17:461-2,
Wheat SPA	seed	Albanietal, Plant Cell, 9: 171-184, 1997
wheat a, b and g gliadins	endosperm	EMBO3:1409-15, 1984
Barley ltr1 promoter	endosperm	
barley B1, C, D hordein	endosperm	Theor Appl Gen 98:1253-62, 1999; Plant J 4:343-55, 1993; Mol Gen Genet 250:750- 60, 1996
Barley DOF	endosperm	Mena et al, The Plant Journal, 116(1): 53- 62, 1998
Biz2	endosperm	EP99106056.7
Synthetic promoter	endosperm	Vicente-Carbajosa et al., Plant J. 13: 629-640, 1998
rice prolamin NRP33	endosperm	Wu et al, Plant Cell Physiology 39(8) 885- 889, 1998
rice -globulin Glb-1	endosperm	Wu et al, Plant Cell Physiology 39(8) 885-889, 1998
rice OSH1	embryo	Sato et al, Proc. Nati. Acad. Sci. USA, 93: 8117-8122
rice alpha-globulin REB/OHP-1	endosperm	Nakase et al. Plant Mol. Biol. 33: 513-S22, 1997
rice ADP-glucose PP	endosperm	Trans Res 6:157-68, 1997
maize ESR gene family	endosperm	Plant J 12:235-46, 1997
sorghum gamma- kafirin	endosperm	PMB 32:1029-35, 1996
KNOX	embryo	Postma-Haarsma ef al, Plant Mol. Biol. 39:257-71, 1999
rice oleosin	Embryo and aleuton	Wu et at, J. Biochem., 123:386, 1998
sunflower oleosin	Seed (embryo and dry seed)	Cummins, etal., Plant Mol. Biol. 19: 873- 876, 1992

Table III***Exemplary flower-specific promoters for use in the performance of the invention***

Gene Source	Expression Pattern	Reference
AtPRP4	flowers	www.dotsalusdotmediumdotedu/mg/tierney/html
chalcone synthase (chsA)	flowers	Van der Meer, et al., Plant Mol. Biol. 15, 95-109, 1990.
LAT52	anther	Twiss et al Mol. Gen. Genet. 217:240-245 (1989)
apetala- 3	flowers	

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Table IV***Alternative rice promoters for use in the performance of the invention***

PRO #	gene	expression
PR00001	Metallothionein Mte	transfer layer of embryo + calli
PR00005	putative beta-amylase	transfer layer of embryo
PR00009	Putative cellulose synthase	Weak in roots
PR00012	lipase (putative)	
PR00014	Transferase (putative)	
PR00016	peptidyl prolyl cis-trans isomerase (putative)	
PR00019	unknown	
PR00020	prp protein (putative)	
PR00029	noduline (putative)	
PR00058	Proteinase inhibitor Rgpi9	seed
PR00061	beta expansin EXPB9	Weak in young flowers
PR00063	Structural protein	young tissues+calli+embryo
PR00069	xylosidase (putative)	
PR00075	Prolamine 10Kda	strong in endosperm
PR00076	allergen RA2	strong in endosperm
PR00077	prolamine RP7	strong in endosperm
PR00078	CBP80	
PR00079	starch branching enzyme I	
PR00080	Metallothionein-like ML2	transfer layer of embryo + calli
PR00081	putative caffeoyl- CoA 3-O methyltransferase	shoot
PR00087	prolamine RM9	strong in endosperm
PR00090	prolamine RP6	strong in endosperm
PR00091	prolamine RP5	strong in endosperm
PR00092	allergen RA5	
PR00095	putative methionine aminopeptidase	embryo
PR00098	ras-related GTP binding protein	
PR00104	beta expansin EXPB1	
PR00105	Glycine rich protein	
PR00108	metallothionein like protein (putative)	
PR00110	RCc3 strong root	
PR00111	uclacyanin 3-like protein	weak discrimination center / shoot meristem
PR00116	26S proteasome regulatory particle non-ATPase subunit 11	very weak meristem specific
PR00117	putative 40S ribosomal protein	weak in endosperm

PR00122	chlorophyll a/lb-binding protein precursor (Cab27)	very weak in shoot
PR00123	putative protochlorophyllide reductase	Strong leaves
PR00126	metallothionein RiCMT	strong discrimination center shoot meristem
PR00129	GOS2	Strong constitutive
PR00131	GOS9	
PR00133	chitinase Cht-3	very weak meristem specific
PR00135	alpha- globulin	Strong in endosperm
PR00136	alanine aminotransferase	Weak in endosperm
PR00138	Cyclin A2	
PR00139	Cyclin D2	
PR00140	Cyclin D3	
PR00141	Cyclophyllin 2	Shoot and seed
PR00146	sucrose synthase SS1 (barley)	medium constitutive
PR00147	trypsin inhibitor ITR1 (barley)	weak in endosperm
PR00149	ubiquitine 2 with intron	strong constitutive
PR00151	WS118	Embryo and stress
PR00156	HVA22 homologue (putative)	
PR00157	EL2	
PR00169	aquaporine	medium constitutive in young plants
PR00170	High mobility group protein	Strong constitutive
PR00171	reversibly glycosylated protein RGP1	weak constitutive
PR00173	cytosolic MDH	shoot
PR00175	RAB21	Embryo and stress
PR00176	CDPK7	
PR00177	Cdc2-1	very weak in meristem
PR00197	sucrose synthase 3	
PRO0198	OsVP1	
PRO0200	OSH1	very weak in young plant meristem
PRO0208	putative chlorophyllase	
PRO0210	OsNRT1	
PRO0211	EXP3	
PRO0216	phosphate transporter OjPT1	
PRO0218	oleosin 18kd	aleurone + embryo
PRO0219	ubiquitine 2 without intron	
PRO0220	RFL	
PRO0221	maize UBI delta intron	not detected
PRO0223	glutelin-1	
PRO0224	fragment of prolamin RP6 promoter	
PRO0225	4xABRE	
PRO0226	glutelin OSGLUA3	
PRO0227	BLZ-2_short (barley)	
PRO0228	BLZ-2_long (barley)	

The nucleic acid construct of some embodiments of the invention can further include an appropriate selectable marker and/or an origin of replication. According to some embodiments of the invention, the nucleic acid construct utilized is a shuttle

vector, which can propagate both in *E. coli* (wherein the construct comprises an appropriate selectable marker and origin of replication) and be compatible with propagation in cells. The construct according to the present invention can be, for example, a plasmid, a bacmid, a phagemid, a cosmid, a phage, a virus or an artificial chromosome.

The nucleic acid construct of some embodiments of the invention can be utilized to stably or transiently transform plant cells. In stable transformation, the nucleic acid is integrated into the plant genome and as such it represents a stable and inherited trait. In transient transformation, the exogenous polynucleotide is expressed by the cell transformed but it is not integrated into the genome and as such it represents a transient trait.

Thus, according to some aspects of the present invention, there is provided an isolated cell comprising the nucleic acid construct of the invention.

As used herein, the term "isolated cell" refers to a cell at least partially separated from the natural environment e.g., from a plant. In some embodiments, the isolated cell is a plant cell of a whole plant. In some embodiments, the isolated cell is a plant cell, for example, a plant cell in culture.

The term "plant" as used herein encompasses whole plants, ancestors and progeny of the plants and plant parts, including seeds, shoots, stems, roots (including tubers), and plant cells, tissues and organs. The plant may be in any form including suspension cultures, embryos, meristematic regions, callus tissue, leaves, gametophytes, sporophytes, pollen, and microspores. Plants that are particularly useful in the methods of the invention include all plants which belong to the superfamily Viridiplantae, in particular monocotyledonous and dicotyledonous plants including a fodder or forage legume, ornamental plant, food crop, tree, or shrub selected from the list comprising *Acacia* spp., *Acer* spp., *Actinidia* spp., *Aesculus* spp., *Agathis australis*, *Albizia amara*, *Alsophila tricolor*, *Andropogon* spp., *Arachis* spp., *Areca catechu*, *Astelia fragrans*, *Astragalus cicer*, *Baikiaea plurijuga*, *Betula* spp., *Brassica* spp., *Bruguiera gymnorhiza*, *Burkea africana*, *Butea frondosa*, *Cadaba farinosa*, *Calliandra* spp., *Camellia sinensis*, *Canna indica*, *Capsicum* spp., *Cassia* spp., *Centroema pubescens*, *Chacoomeles* spp., *Cinnamomum cassia*, *Coffea arabica*, *Colophospermum mopane*, *Coronilla varia*, *Cotoneaster serotina*, *Crataegus* spp., *Cucumis* spp., *Cupressus* spp., *Cyathea dealbata*,

Cydonia oblonga, Cryptomeria japonica, Cymbopogon spp., Cynthea dealbata, Cydonia
 oblonga, Dalbergia monetaria, Davallia divaricata, Desmodium spp., Dicksonia
 squarosa, Dibeteropogon amplexans, Dioclea spp, Dolichos spp., Dorycnium rectum,
 Echinochloa pyramidalis, Ehraffia spp., Eleusine coracana, Eragrestis spp., Erythrina
 5 spp., Eucalyptus spp., Euclea schimperii, Eulalia villosa, Pagopyrum spp., Feijoa
 sellowiana, Fragaria spp., Flemingia spp, Freycinetia banksii, Geranium thunbergii,
 GynAndro biloba, Glycine javanica, Gliricidia spp, Gossypium hirsutum, Grevillea spp.,
 Guibourtia coleosperma, Hedysarum spp., Hemaphysalis altissima, Heteropogon contortus,
 Hordeum vulgare, Hyparrhenia rufa, Hypericum erectum, Hypochaeris dissoluta, Indigo
 10 incanata, Iris spp., Leptarrhena pyrrolifolia, Lespedeza spp., Lettuce spp., Leucaena
 leucocephala, Loudetia simplex, Lotonus bainesii, Lotus spp., Macrotyloma axillare,
 Malus spp., Manihot esculenta, Medicago sativa, Metasequoia glyptostroboides, Musa
 sapientum, Nicotiana spp., Onobrychis spp., Ornithopus spp., Oryza spp.,
 Peltophorum africanum, Pennisetum spp., Persea gratissima, Petunia spp., Phaseolus
 15 spp., Phoenix canariensis, Phormium cookianum, Photinia spp., Picea glauca, Pinus spp.,
 Pisum sativum, Podocarpus totara, Pogonarthria fleckii, Pogonaffria squarrosa,
 Populus spp., Prosopis cineraria, Pseudotsuga menziesii, Pterolobium stellatum, Pyrus
 communis, Quercus spp., Rhamphiolepis umbellata, Rhopalostylis sapida, Rhus
 natalensis, Ribes grossularia, Ribes spp., Robinia pseudoacacia, Rosa spp., Rubus spp.,
 20 Salix spp., Schyzachyrium sanguineum, Sciadopitys verticillata, Sequoia sempervirens,
 Sequoiadendron giganteum, Sorghum bicolor, Spinacia spp., Sporobolus fimbriatus,
 Stiburus alopecuroides, Stylosanthes humilis, Tadehagi spp, Taxodium distichum,
 Themeda triandra, Trifolium spp., Triticum spp., Tsuga heterophylla, Vaccinium spp.,
 Vicia spp., Vitis vinifera, Watsonia pyramidata, Zantedeschia aethiopica, Zea mays,
 25 amaranth, artichoke, asparagus, broccoli, Brussels sprouts, cabbage, canola, carrot,
 cauliflower, celery, collard greens, flax, kale, lentil, oilseed rape, okra, onion, potato,
 rice, soybean, straw, sugar beet, sugar cane, sunflower, tomato, squash, tea, maize,
 wheat, barley, rye, oat, peanut, pea, lentil and alfalfa, cotton, rapeseed, canola, pepper,
 sunflower, tobacco, eggplant, eucalyptus, a tree, an ornamental plant, a perennial grass
 30 and a forage crop. Alternatively algae and other non-Viridiplantae can be used for the
 methods of the present invention.

According to some embodiments of the invention, the plant or plant cell is a duckweed plant, cell or nodule. Duckweed (members of the monocotyledonous family Lemnaceae, or Lemna) plant or duckweed nodule cultures can be efficiently transformed with an expression cassette containing a nucleotide sequence of interest by any one of a number of methods including *Agrobacterium*-mediated gene transfer, ballistic bombardment, or electroporation. Methods for molecular engineering of duckweed cells and detailed description of duckweed expression systems useful for commercial production of polypeptides are known in the art (see, for example, US Patent Nos. 6,040,498 and 6,815,184 to Stomp, et al, and 8,022,270 to Dickey et al, all of which are incorporated fully by reference herein).

According to some embodiments of the invention, the plant or plant cell used by the method of the invention is a crop plant or cell of a crop plant such as rice, maize, wheat, barley, peanut, potato, sesame, olive tree, palm oil, banana, soybean, sunflower, canola, sugarcane, alfalfa, millet, leguminosae (bean, pea), flax, lupinus, rapeseed, tobacco, poplar and cotton.

According to further embodiments the plant cells includes tobacco cells, *Agrobacterium rhizogenes* transformed root cell, celery cell, ginger cell, horseradish cell and carrot cells. In one embodiment the tobacco cells are from a tobacco cell line, such as, but not limited to *Nicotiana tabacum* L. cv Bright Yellow (BY-2) cells. The plant cells may be grown according to any type of suitable culturing method, including but not limited to, culture on a solid surface (such as a plastic culturing vessel or plate for example) or in suspension. It will be noted that some cells, such as the BY-2 and carrot cells can be cultured and grown in suspension. Suitable devices and methods for culturing plant cells in suspension are known in the art, for example, as described in International Patent Application PCT IL2008/000614. In yet another embodiment the cells are cells of whole tobacco plants or plant tissues, including, but not limited to *Nicotiana benthamiana*. According to yet another embodiment, the plant cells are carrot cells.

There are various methods of introducing foreign genes into both monocotyledonous and dicotyledonous plants (Potrykus, I., Annu. Rev. Plant. Physiol., Plant. Mol. Biol. (1991) 42:205-225; Shimamoto et al., Nature (1989) 338:274-276).

The principle methods of causing stable integration of exogenous DNA into plant genomic DNA include two main approaches:

(i) Agrobacterium-mediated gene transfer: Klee et al. (1987) *Annu. Rev. Plant Physiol.* 38:467-486; Klee and Rogers in *Cell Culture and Somatic Cell Genetics of Plants*, Vol. 6, Molecular Biology of Plant Nuclear Genes, eds. Schell, J., and Vasil, L. K., Academic Publishers, San Diego, Calif. (1989) p. 2-25; Gatenby, in *Plant Biotechnology*, eds. Kung, S. and Arntzen, C. J., Butterworth Publishers, Boston, Mass. (1989) p. 93-112.

(ii) Direct DNA uptake: Paszkowski et al., in *Cell Culture and Somatic Cell Genetics of Plants*, Vol. 6, Molecular Biology of Plant Nuclear Genes eds. Schell, J., and Vasil, L. K., Academic Publishers, San Diego, Calif. (1989) p. 52-68; including methods for direct uptake of DNA into protoplasts, Toriyama, K. et al. (1988) *Bio/Technology* 6:1072-1074. DNA uptake induced by brief electric shock of plant cells: Zhang et al. *Plant Cell Rep.* (1988) 7:379-384. Fromm et al. *Nature* (1986) 319:791-793. DNA injection into plant cells or tissues by particle bombardment, Klein et al. *Bio/Technology* (1988) 6:559-563; McCabe et al. *Bio/Technology* (1988) 6:923-926; Sanford, *Physiol. Plant.* (1990) 79:206-209; by the use of micropipette systems: Neuhaus et al., *Theor. Appl. Genet.* (1987) 75:30-36; Neuhaus and Spangenberg, *Physiol. Plant.* (1990) 79:213-217; glass fibers or silicon carbide whisker transformation of cell cultures, embryos or callus tissue, U.S. Pat. No. 5,464,765 or by the direct incubation of DNA with germinating pollen, DeWet et al. in *Experimental Manipulation of Ovule Tissue*, eds. Chapman, G. P. and Mantell, S. H. and Daniels, W. Longman, London, (1985) p. 197-209; and Ohta, *Proc. Natl. Acad. Sci. USA* (1986) 83:715-719.

The Agrobacterium system includes the use of plasmid vectors that contain defined DNA segments that integrate into the plant genomic DNA. Methods of inoculation of the plant tissue vary depending upon the plant species and the Agrobacterium delivery system. A widely used approach is the leaf disc procedure which can be performed with any tissue explant that provides a good source for initiation of whole plant differentiation. See, e.g., Horsch et al. in *Plant Molecular Biology Manual A5*, Kluwer Academic Publishers, Dordrecht (1988) p. 1-9. A supplementary approach employs the Agrobacterium delivery system in combination

with vacuum infiltration. The *Agrobacterium* system is especially viable in the creation of transgenic dicotyledonous plants.

There are various methods of direct DNA transfer into plant cells. In electroporation, the protoplasts are briefly exposed to a strong electric field. In microinjection, the DNA is mechanically injected directly into the cells using very small micropipettes. In microparticle bombardment, the DNA is adsorbed on microprojectiles such as magnesium sulfate crystals or tungsten particles, and the microprojectiles are physically accelerated into cells or plant tissues.

Following stable transformation plant propagation is exercised. The most common method of plant propagation is by seed. Regeneration by seed propagation, however, has the deficiency that due to heterozygosity there is a lack of uniformity in the crop, since seeds are produced by plants according to the genetic variances governed by Mendelian rules. Basically, each seed is genetically different and each will grow with its own specific traits. Therefore, it is preferred that the transformed plant be produced such that the regenerated plant has the identical traits and characteristics of the parent transgenic plant. Therefore, it is preferred that the transformed plant be regenerated by micropropagation which provides a rapid, consistent reproduction of the transformed plants.

Micropropagation is a process of growing new generation plants from a single piece of tissue that has been excised from a selected parent plant or cultivar. This process permits the mass reproduction of plants having the preferred tissue expressing the fusion protein. The new generation plants which are produced are genetically identical to, and have all of the characteristics of, the original plant. Micropropagation allows mass production of quality plant material in a short period of time and offers a rapid multiplication of selected cultivars in the preservation of the characteristics of the original transgenic or transformed plant. The advantages of cloning plants are the speed of plant multiplication and the quality and uniformity of plants produced.

Micropropagation is a multi-stage procedure that requires alteration of culture medium or growth conditions between stages. Thus, the micropropagation process involves four basic stages: Stage one, initial tissue culturing; stage two, tissue culture multiplication; stage three, differentiation and plant formation; and stage four, greenhouse culturing and hardening. During stage one, initial tissue culturing, the tissue

culture is established and certified contaminant-free. During stage two, the initial tissue culture is multiplied until a sufficient number of tissue samples are produced to meet production goals. During stage three, the tissue samples grown in stage two are divided and grown into individual plantlets. At stage four, the transformed plantlets are transferred to a greenhouse for hardening where the plants' tolerance to light is gradually increased so that it can be grown in the natural environment.

According to some embodiments of the invention, the transgenic plants are generated by transient transformation of leaf cells, meristematic cells or the whole plant.

Transient transformation can be effected by any of the direct DNA transfer methods described above or by viral infection using modified plant viruses.

Viruses that have been shown to be useful for the transformation of plant hosts include CaMV, Tobacco mosaic virus (TMV), brome mosaic virus (BMV) and Bean Common Mosaic Virus (BV or BCMV). Transformation of plants using plant viruses is described in U.S. Pat. No. 4,855,237 (bean golden mosaic virus; BGV), EP-A 67,553 (TMV), Japanese Published Application No. 63-14693 (TMV), EPA 194,809 (BV), EPA 278,667 (BV); and Gluzman, Y. et al., Communications in Molecular Biology: Viral Vectors, Cold Spring Harbor Laboratory, New York, pp. 172-189 (1988). Pseudovirus particles for use in expressing foreign DNA in many hosts, including plants are described in WO 87/06261.

According to some embodiments of the invention, the virus used for transient transformations is avirulent and thus is incapable of causing severe symptoms such as reduced growth rate, mosaic, ring spots, leaf roll, yellowing, streaking, pox formation, tumor formation and pitting. A suitable avirulent virus may be a naturally occurring avirulent virus or an artificially attenuated virus. Virus attenuation may be effected by using methods well known in the art including, but not limited to, sub-lethal heating, chemical treatment or by directed mutagenesis techniques such as described, for example, by Kurihara and Watanabe (Molecular Plant Pathology 4:259-269, 2003), Galon et al. (1992), Atreya et al. (1992) and Huet et al. (1994).

Suitable virus strains can be obtained from available sources such as, for example, the American Type Culture Collection (ATCC) or by isolation from infected plants. Isolation of viruses from infected plant tissues can be effected by techniques well known in the art such as described, for example by Foster and Tatlor, Eds. "Plant

Virology Protocols: From Virus Isolation to Transgenic Resistance (Methods in Molecular Biology (Humana Pr), Vol 81)”, Humana Press, 1998. Briefly, tissues of an infected plant believed to contain a high concentration of a suitable virus, preferably young leaves and flower petals, are ground in a buffer solution (e.g., phosphate buffer solution) to produce a virus infected sap which can be used in subsequent inoculations.

Construction of plant RNA viruses for the introduction and expression of non-viral nucleic acid sequences in plants is demonstrated by the above references as well as by Dawson, W. O. et al., Virology (1989) 172:285-292; Takamatsu et al. EMBO J. (1987) 6:307-311; French et al. Science (1986) 231:1294-1297; Takamatsu et al. FEBS Letters (1990) 269:73-76; and U.S. Pat. No. 5,316,931.

When the virus is a DNA virus, suitable modifications can be made to the virus itself. Alternatively, the virus can first be cloned into a bacterial plasmid for ease of constructing the desired viral vector with the foreign DNA. The virus can then be excised from the plasmid. If the virus is a DNA virus, a bacterial origin of replication can be attached to the viral DNA, which is then replicated by the bacteria. Transcription and translation of this DNA will produce the coat protein which will encapsidate the viral DNA. If the virus is an RNA virus, the virus is generally cloned as a cDNA and inserted into a plasmid. The plasmid is then used to make all of the constructions. The RNA virus is then produced by transcribing the viral sequence of the plasmid and translation of the viral genes to produce the coat protein(s) which encapsidate the viral RNA.

In one embodiment, a plant viral polynucleotide is provided in which the native coat protein coding sequence has been deleted from a viral polynucleotide, a non-native plant viral coat protein coding sequence and a non-native promoter, preferably the subgenomic promoter of the non-native coat protein coding sequence, capable of expression in the plant host, packaging of the recombinant plant viral polynucleotide, and ensuring a systemic infection of the host by the recombinant plant viral polynucleotide, has been inserted. Alternatively, the coat protein gene may be inactivated by insertion of the non-native polynucleotide sequence within it, such that a protein is produced. The recombinant plant viral polynucleotide may contain one or more additional non-native subgenomic promoters. Each non-native subgenomic promoter is capable of transcribing or expressing adjacent genes or polynucleotide

sequences in the plant host and incapable of recombination with each other and with native subgenomic promoters. Non-native (foreign) polynucleotide sequences may be inserted adjacent the native plant viral subgenomic promoter or the native and a non-native plant viral subgenomic promoters if more than one polynucleotide sequence is included. The non-native polynucleotide sequences are transcribed or expressed in the host plant under control of the subgenomic promoter to produce the desired products.

In a second embodiment, a recombinant plant viral polynucleotide is provided as in the first embodiment except that the native coat protein coding sequence is placed adjacent one of the non-native coat protein subgenomic promoters instead of a non-native coat protein coding sequence.

In a third embodiment, a recombinant plant viral polynucleotide is provided in which the native coat protein gene is adjacent its subgenomic promoter and one or more non-native subgenomic promoters have been inserted into the viral polynucleotide. The inserted non-native subgenomic promoters are capable of transcribing or expressing adjacent genes in a plant host and are incapable of recombination with each other and with native subgenomic promoters. Non-native polynucleotide sequences may be inserted adjacent the non-native subgenomic plant viral promoters such that the sequences are transcribed or expressed in the host plant under control of the subgenomic promoters to produce the desired product.

In a fourth embodiment, a recombinant plant viral polynucleotide is provided as in the third embodiment except that the native coat protein coding sequence is replaced by a non-native coat protein coding sequence.

The viral vectors are encapsidated by the coat proteins encoded by the recombinant plant viral polynucleotide to produce a recombinant plant virus. The recombinant plant viral polynucleotide or recombinant plant virus is used to infect appropriate host plants. The recombinant plant viral polynucleotide is capable of replication in the host, systemic spread in the host, and transcription or expression of foreign gene(s) (exogenous polynucleotide) in the host to produce the desired protein.

Techniques for inoculation of viruses to plants may be found in Foster and Taylor, eds. "Plant Virology Protocols: From Virus Isolation to Transgenic Resistance (Methods in Molecular Biology (Humana Pr), Vol 81)", Humana Press, 1998; Maramorosh and Koprowski, eds. "Methods in Virology" 7 vols, Academic Press, New

York 1967-1984; Hill, S.A. "Methods in Plant Virology", Blackwell, Oxford, 1984; Walkey, D.G.A. "Applied Plant Virology", Wiley, New York, 1985; and Kado and Agrawa, eds. "Principles and Techniques in Plant Virology", Van Nostrand-Reinhold, New York.

5 In addition to the above, the polynucleotide of the present invention can also be introduced into a chloroplast genome thereby enabling chloroplast expression.

A technique for introducing exogenous nucleic acid sequences to the genome of the chloroplasts is known. This technique involves the following procedures. First, plant cells are chemically treated so as to reduce the number of chloroplasts per cell to about
10 one. Then, the exogenous polynucleotide is introduced via particle bombardment into the cells with the aim of introducing at least one exogenous polynucleotide molecule into the chloroplasts. The exogenous polynucleotides selected such that it is integratable into the chloroplast's genome via homologous recombination which is readily effected by enzymes inherent to the chloroplast. To this end, the nucleic acid sequence includes,
15 in addition to a gene of interest, at least one polynucleotide stretch which is derived from the chloroplast's genome. In addition, the exogenous polynucleotide includes a selectable marker, which serves by sequential selection procedures to ascertain that all or substantially all of the copies of the chloroplast genomes following such selection will include the exogenous polynucleotide. Further details relating to this technique are
20 found in U.S. Pat. Nos. 4,945,050; and 5,693,507 which are incorporated herein by reference. A polypeptide can thus be produced by the protein expression system of the chloroplast and become integrated into the chloroplast's inner membrane.

According to some embodiments of the invention, the method further comprises growing the plant cell expressing the nucleic acid. The plant cells can be any plant cells
25 desired. The plant cells can be cultured cells, cells in cultured tissue or cultured organs, or cells in a plant. In some embodiments, the plant cells are cultured cells, or cells in cultured tissue or cultured organs. In yet further embodiments, the plant cells are any type of plant that is used in gene transference. The plant cell can be grown as part of a whole plant, or, alternatively, in plant cell culture.

30 According to some aspects of the invention, the plant cells are grown in a plant cell suspension culture. As used herein, the term "suspension culture" refers to the growth of cells separate from the organism. Suspension culture can be facilitated via use

of a liquid medium (a "suspension medium"). Suspension culture can refer to the growth of cells in liquid nutrient media. Methods and devices suitable for growing plant cells of the invention in plant cell suspension culture are described in detail in, for example, PCT WO2008/135991, US Patent No. 6,391,683, US Patent Application No. 10/784,295; 5 International Patent Publications PCT Nos. WO2004/091475, WO2005/080544 and WO 2006/040761, all of which are hereby incorporated by reference as if fully set forth herein.

Thus, the invention encompasses plants or plant cultures expressing the nucleic acid sequences, so as to produce the TNF α polypeptide inhibitor of the invention. Once 10 expressed within the plant cell or the entire plant, the level of the TNF α inhibitor encoded by the nucleic acid sequence can be determined by methods well known in the art such as, activity assays, Western blots using antibodies capable of specifically binding the TNF α inhibitor e.g., chimeric polypeptide (anti TNFR2, and anti Fc, See Examples section which follows), Enzyme-Linked Immuno Sorbent Assay (ELISA), 15 radio-immuno-assays (RIA), immunohistochemistry, immunocytochemistry, immunofluorescence and the like.

Methods of determining the level in the plant of the RNA transcribed from the nucleic acid sequence are well known in the art and include, for example, Northern blot analysis, reverse transcription polymerase chain reaction (RT-PCR) analysis (including 20 quantitative, semi-quantitative or real-time RT-PCR) and RNA-*in situ* hybridization.

According to some embodiments of the invention, the expressed recombinant chimeric polypeptide of the present invention is glycosylated in the plant cell, resulting in a chimeric polypeptide having one, or two or three or more glycan structures having plant specific glycan residues. Thus, according to some embodiments of the invention, 25 the cells expressing the expression vector of the invention produce a chimeric polypeptide having various amounts of glycan structures arranged in one, two, three or more antennae. All structures may contain a core structure of two GlcNAcs and one mannose, and variations of different amounts of mannose, in addition to core alpha (1,3) fucose, beta (1,2) xylose, and/or GlcNAc residues. Structures can be of the high 30 mannose type, having at least one, optionally at least two, optionally at least three or optionally at least four or more mannose residues in addition to the core structure ; or complex type having both mannose and other glycan types on each glycan, or of the

hybrid type having both high mannose and complex antennae. In other embodiments the cells expressing the expression vector of the invention produce a TNF α inhibitor having at least one, optionally at least two, optionally at least three or optionally at least four or more core xylose residues. In yet other embodiments the cells expressing the expression vector of the invention produce a TNF α inhibitor having at least one, optionally at least two, optionally at least three or optionally at least four or more core α -(1,3) fucose residues. In one embodiment the cells expressing the expression vector of the invention produce a TNF α inhibitor protein having at least one exposed mannose residue, at least one core xylose residue and at least one α -(1,3) fucose residue. In yet further embodiments, the cells expressing the expression vector of the invention produce a TNF α inhibitor having at least one, at least two, at least 3 or more terminal N-acetyl glucosamine substitutions on the outer mannose sugars.

According to a specific embodiment the TNF α inhibitor e.g., chimeric polypeptide, lacks sialic acid residues. Yet further according to a specific embodiment, the TNF α inhibitor e.g., chimeric polypeptide, comprises at least 40 %, 45 %, 50 %, 55 %, 60 %, 65 %, 70 % or more complex glycans. According to a specific embodiment, the chimeric polypeptide comprises 40-70 % complex glycans.

Plant cells expressing the TNF α polypeptide inhibitor of the invention is utilized for the treatment of TNF α -associated medical conditions.

It has been shown in Example 2 of the Examples section that plant cells expressing TNF α polypeptide inhibitor (e.g., chimeric polypeptide) can be used as an effective systemic delivery system, when provided for enteral administration to the subject (see WO2007/010533). Thus, in some embodiments, the TNF α polypeptide inhibitor can be formulated in a pharmaceutical composition for oral or enteral delivery comprising transformed plant cell expressing the chimeric polypeptide and a pharmaceutically acceptable carrier. In some embodiments, the transformed plant cells of the pharmaceutical composition are lyophilized plant cells, although the use of fresh (non-lyophilized cells), plant tissues, plant parts or whole plants is also contemplated herein.

Prior to lyophilization the cells may be washed to remove any cell debris that may be present in the growth medium.

As the cells are being prepared for lyophilization, it is sometimes desirable to incubate the cells in a maintenance medium to reduce the metabolic processes of the cells.

Pretreatment (although not necessary) can be performed at room temperature or at temperatures in which the plant cells are typically cultured. Pretreatment is performed at about room temperature (20 °C) for ease of handling and as most plant cells are fairly stable at room temperature. Stabilizers can be added directly to the medium and replenished as necessary during the pretreatment process.

Pretreatments may also involve incubating cells in the presence of one or more osmotic agents. Examples of useful osmotic agents include sugars such as saccharides and saccharide derivatives, amino or imino acids such as proline and proline derivatives, or combinations of these agents. Some of the more useful sugars and sugar derivatives are fructose, glucose, maltose, mannitol, sorbitol, sucrose and trehalose. Osmotic agents are utilized at a concentration that prepares cells for subsequent lyophilization.

Lyophilization is directed at reducing the water content of the cells by vacuum evaporation. Vacuum evaporation involves placing the cells in an environment with reduced air pressure. Depending on the rate of water removal desired, the reduced ambient pressure operating at temperatures of between about -30 °C to -50° C may be at 100 torr, 1 torr, 0.01 torr or less. According to a specific embodiment, the cells are lyophilized by freezing to -40 °C and then applying a vacuum to a pressure of 0.1 mbar for overnight. The cells are then heated to -10 °C so all the ice content will be sublimated and evaporated. Under conditions of reduced pressure, the rate of water evaporation is increased such that up to 60-95 % of the water in a cell can be removed.

According to a specific embodiment, lyophilization removes over 60 %, 70 %, 80% or specifically over 90 %, 91 %, 92 %, 93 %, 94 %, 95 % or 98 % of the water from the cells. According to a specific embodiment, the final water content is about 5-10 %, 5-8 % or 6-7 %.

As used herein the phrase “enteral administration” refers to administration through any part of the gastro-intestinal tract, such as rectal administration, colonic administration, intestinal administration (proximal or distal) and gastric administration. In some embodiments, enteral administration refers to oral administration. It will be appreciated that the present teachings also aim at mucosal administration.

The cells may be formulated as a solid, formulated as a liquid or formulated as a powder. In some embodiments, the cells are resuspended, lyophilized cells.

Thus, the oral dosage form may be provided as an oral nutritional form (e.g., as long as the protein is not exposed to denaturing conditions which include heating above
5 37 °C and compression), as a complete meal, as a powder for dissolution, e.g. health drinks, as a solution, as a ready-made drink, optionally low calorie, such as a soft drink, including juices, milk-shake, yoghurt drink, smoothie or soy-based drink, in a bar, or dispersed in foods of any sort, such as baked products, cereal bars, dairy bars, snack-foods, breakfast cereals, muesli, candies, tabs, cookies, biscuits, crackers (such as a rice
10 crackers), chocolate, and dairy products.

The cells can be administered to the subject *per se*, or alternatively, the cells of the present invention can be administered to the subject in a pharmaceutical composition where they are mixed with suitable carriers or excipients.

As used herein, a "pharmaceutical composition" refers to a preparation of cells
15 expressing TNFalpha inhibitor with other chemical components such as physiologically suitable carriers and excipients. The purpose of a pharmaceutical composition is to facilitate administration of a compound to an organism.

As used herein, the term "active ingredient" refers to the cells expressing TNFalpha inhibitor accountable for the intended biological effect.

Hereinafter, the phrases "physiologically acceptable carrier" and
20 "pharmaceutically acceptable carrier," which may be used interchangeably, refer to a carrier or a diluent that does not cause significant irritation to an organism and does not abrogate the biological activity and properties of the administered compound. An adjuvant is included under these phrases. Preferably the carrier used is a non-
25 immunogenic carrier and further preferably does not stimulate the gut associated lymphatic tissue.

Herein, the term "excipient" refers to an inert substance added to a pharmaceutical composition to further facilitate administration of an active ingredient. Examples, without limitation, of excipients include calcium carbonate, calcium
30 phosphate, various sugars and types of starch, cellulose derivatives, gelatin, vegetable oils, and polyethylene glycols.

Techniques for formulation and administration of drugs may be found in the latest edition of "Remington's Pharmaceutical Sciences," Mack Publishing Co., Easton, PA, which is herein fully incorporated by reference.

Pharmaceutical compositions for use in accordance with the present invention
5 thus may be formulated in conventional manner using one or more physiologically acceptable carriers comprising excipients and auxiliaries, which facilitate processing of the active ingredients into preparations that can be used pharmaceutically.

For oral administration, the pharmaceutical composition can be formulated readily by combining the active compounds with pharmaceutically acceptable carriers
10 well known in the art. Such carriers enable the pharmaceutical composition to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions, and the like, for oral ingestion by a patient. Pharmacological preparations for oral use can be made using a solid excipient, optionally grinding the resulting mixture, and processing the mixture of granules, after adding suitable auxiliaries as desired, to obtain
15 tablets or dragee cores. Suitable excipients are, in particular, fillers such as sugars, including lactose, sucrose, mannitol, or sorbitol; cellulose preparations such as, for example, maize starch, wheat starch, rice starch, potato starch, gelatin, gum tragacanth, methyl cellulose, hydroxypropylmethyl-cellulose, and sodium carbomethylcellulose; and/or physiologically acceptable polymers such as polyvinylpyrrolidone (PVP). If
20 desired, disintegrating agents, such as cross-linked polyvinyl pyrrolidone, agar, or alginic acid or a salt thereof, such as sodium alginate, may be added.

Dragee cores are provided with suitable coatings. For this purpose, concentrated sugar solutions may be used which may optionally contain gum arabic, talc, polyvinyl pyrrolidone, carbopol gel, polyethylene glycol, titanium dioxide, lacquer solutions, and
25 suitable organic solvents or solvent mixtures. Dyestuffs or pigments may be added to the tablets or dragee coatings for identification or to characterize different combinations of active compound doses.

Pharmaceutical compositions that can be used orally include push-fit capsules made of gelatin, as well as soft, sealed capsules made of gelatin and a plasticizer, such
30 as glycerol or sorbitol. The push-fit capsules may contain the active ingredients in admixture with filler such as lactose, binders such as starches, lubricants such as talc or magnesium stearate, and, optionally, stabilizers. In soft capsules, the active ingredients

may be dissolved or suspended in suitable liquids, such as fatty oils, liquid paraffin, or liquid polyethylene glycols. In addition, stabilizers may be added.

The dosage forms may include additives such as one or more of calcium, magnesium, iron, zinc, phosphorus, vitamin D and vitamin K. A suitable daily amount
5 is 0.1 mg to 3.6 g calcium, preferably 320 to 530 mg. In general, the daily dosage of vitamins and minerals in the nutritional formulation or medicament of the invention is 25-100% by weight of the dosages recommended by the health authorities. Dietary fiber may also be a component of the compositions of the invention. Further components of the supplement may include any bioactive compounds or extracts which are known to
10 have health benefits, especially for improving physical performance.

Generally the unit dosage form may further comprise an antioxidant (exemplary embodiments are provided above-. In another embodiment, the antioxidant is a pharmaceutically acceptable antioxidant. In another embodiment, the antioxidant is selected from the group consisting of vitamin E, superoxide dismutase (SOD), omega-3,
15 and beta-carotene.

In another embodiment, the unit dosage form further comprises an enhancer of the biologically active protein or peptide. In another embodiment, the unit dosage form further comprises a cofactor of the biologically active protein or peptide.

In another embodiment, a unit dosage form of the present invention further
20 comprises pharmaceutical-grade surfactant. Surfactants are well known in the art, and are described, inter alia, in the Handbook of Pharmaceutical Excipients (eds. Raymond C Rowe, Paul J Sheskey, and Sian C Owen, copyright Pharmaceutical Press, 2005). In another embodiment, the surfactant is any other surfactant known in the art.

In another embodiment, a unit dosage form of the present invention further
25 comprises pharmaceutical-grade emulsifier or emulgator (emollient). Emulsifiers and emulgators are well known in the art, and are described, inter alia, in the Handbook of Pharmaceutical Excipients (ibid). Non-limiting examples of emulsifiers and emulgators are eumulgin, Eumulgin B1 PH, Eumulgin B2 PH, hydrogenated castor oil cetostearyl alcohol, and cetyl alcohol. In another embodiment, the emulsifier or emulgator is any
30 other emulsifier or emulgator known in the art.

In another embodiment, a unit dosage form of the present invention further comprises pharmaceutical-grade stabilizer. Stabilizers are well known in the art, and are

described, inter alia, in the Handbook of Pharmaceutical Excipients (ibid). In another embodiment, the stabilizer is any other stabilizer known in the art.

In another embodiment, a unit dosage form of the present invention further comprises an amino acid selected from the group consisting of arginine, lysine, aspartate, glutamate, and histidine. In another embodiment, analogues and modified versions of arginine, lysine, aspartate, glutamate and histidine are included in the terms "arginine," "lysine," "aspartate", "glutamate" and "histidine," respectively. In another embodiment, the amino acid provides additional protection of ribonuclease or other active molecules. In another embodiment, the amino acid promotes interaction of biologically active protein or peptide with a target cell. In another embodiment, the amino acid is contained in an oil component of the unit dosage form.

In another embodiment, a unit dosage form of the present invention further comprises one or more pharmaceutically acceptable excipients, into which the matrix carrier unit dosage form is mixed. In another embodiment, the excipients include one or more additional polysaccharides. In another embodiment, the excipients include one or more waxes. In another embodiment, the excipients provide a desired taste to the unit dosage form. In another embodiment, the excipients influence the drug consistency, and the final dosage form such as a gel capsule or a hard gelatin capsule.

Non limiting examples of excipients include: Antifoaming agents (dimethicone, simethicone); Antimicrobial preservatives (benzalkonium chloride, benzethonium chloride, butylparaben, cetylpyridinium chloride, chlorobutanol, chlorocresol, cresol, ethylparaben, methylparaben, methylparaben sodium, phenol, phenylethyl alcohol, phenylmercuric acetate, phenylmercuric nitrate, potassium benzoate, potassium sorbate, propylparaben, propylparaben sodium, sodium benzoate, sodium dehydroacetate, sodium propionate, sorbic acid, thimerosal, thymol); Chelating agents (edetate disodium, ethylenediaminetetraacetic acid and salts, edetic acid); Coating agents (sodium carboxymethyl-cellulose, cellulose acetate, cellulose acetate phthalate, ethylcellulose, gelatin, pharmaceutical glaze, hydroxypropyl cellulose, hydroxypropyl methylcellulose, hydroxypropyl methylcellulose phthalate, methacrylic acid copolymer, methylcellulose, polyethylene glycol, polyvinyl acetate phthalate, shellac, sucrose, titanium dioxide, carnauba wax, microcrystalline wax, zein); Colorants (caramel, red, yellow, black or blends, ferric oxide); Complexing agents (ethylenediaminetetraacetic

acid and salts (EDTA), edetic acid, gentisic acid ethanolmaide, oxyquinoline sulfate); Desiccants (calcium chloride, calcium sulfate, silicon dioxide); Emulsifying and/or solubilizing agents (acacia, cholesterol, diethanolamine (adjunct), glyceryl monostearate, lanolin alcohols, lecithin, mono- and di-glycerides, monoethanolamine (adjunct), oleic acid (adjunct), oleyl alcohol (stabilizer), poloxamer, polyoxyethylene 50 stearate, polyoxyl 35 castor oil, polyoxyl 40 hydrogenated castor oil, polyoxyl 10 oleyl ether, polyoxyl 20 cetostearyl ether, polyoxyl 40 stearate, polysorbate 20, polysorbate 40, polysorbate 60, polysorbate 80, propylene glycol diacetate, propylene glycol monostearate, sodium lauryl sulfate, sodium stearate, sorbitan monolaurate, sorbitan monooleate, sorbitan monopalmitate, sorbitan monostearate, stearic acid, trolamine, emulsifying wax); Flavors and perfumes (anethole, benzaldehyde, ethyl vanillin, menthol, methyl salicylate, monosodium glutamate, orange flower oil, peppermint, peppermint oil, peppermint spirit, rose oil, stronger rose water, thymol, tolu balsam tincture, vanilla, vanilla tincture, vanillin); Humectants (glycerin, hexylene glycol, propylene glycol, sorbitol); Polymers (e.g., cellulose acetate, alkyl celluloses, hydroxyalkylcelluloses, acrylic polymers and copolymers); Suspending and/or viscosity-increasing agents (acacia, agar, alginic acid, aluminum monostearate, bentonite, purified bentonite, magma bentonite, carbomer 934p, carboxymethylcellulose calcium, carboxymethylcellulose sodium, carboxymethylcellulose sodium 12, carrageenan, microcrystalline and carboxymethylcellulose sodium cellulose, dextrin, gelatin, guar gum, hydroxyethyl cellulose, hydroxypropyl cellulose, hydroxypropyl methylcellulose, magnesium aluminum silicate, methylcellulose, pectin, polyethylene oxide, polyvinyl alcohol, povidone, propylene glycol alginate, silicon dioxide, colloidal silicon dioxide, sodium alginate, tragacanth, xanthan gum); Sweetening agents (aspartame, dextrates, dextrose, excipient dextrose, fructose, mannitol, saccharin, calcium saccharin, sodium saccharin, sorbitol, solution sorbitol, sucrose, compressible sugar, confectioner's sugar, syrup); This list is not meant to be exclusive, but instead merely representative of the classes of excipients and the particular excipients which may be used in oral dosage unit dosage forms of the present invention.

Conventional additives may be included in the compositions of the invention, including any of those selected from preservatives, chelating agents, effervescing agents, natural or artificial sweeteners, flavoring agents, coloring agents, taste masking

agents, acidulants, emulsifiers, thickening agents, suspending agents, dispersing or wetting agents, antioxidants, and the like. Flavoring agents can be added to the compositions of the invention to aid in compliance with a dosing regimen. Typical flavoring agents include, but are not limited to natural or synthetic essences, oils and/or extracts of pineapple, orange, lemon, mint, berry, chocolate, vanilla and melon.

The amount of a composition to be administered will, of course, be dependent on the subject being treated, the severity of the affliction, the manner of administration, the judgment of the prescribing physician, etc.

In another embodiment the effective chimeric polypeptide amount per adult dose range is about 0.0002 mg/kg to 2 mg/kg, about 0.002-2 mg/kg, about 0.02-2 mg/kg, about 0.2-2 mg/kg, about 0.002-0.2 mg/kg, about 0.0002-1 mg/kg, about 0.002-0.1 mg/kg, about 0.002-0.02 mg/kg, about 0.002-0.01 mg/kg, about 0.002-0.008 mg/kg, about 0.02-0.1 mg/kg, about 0.001-0.05 mg/kg, about 0.001-0.01 mg/kg, about 0.01-1 mg/kg, about 0.01-15 mg/kg, about 0.005 -1 mg/kg, about 0.01-5 mg/kg, about 0.005-0.01 mg/kg or about 0.05-0.1 mg/kg. According to a specific embodiment, the effective chimeric polypeptide amount per adult dose ranges about 0.002-0.2 mg/kg.

According to a specific embodiment, a flat dose of 0.01-100 mg, 0.1-100 mg, 0.1-50 mg, 0.1-20 mg, 0.1-10 mg, 0.1-5 mg is administered.

According to a specific embodiment the flat dose is about 0.1-10 mg.

According to a specific embodiment, the oral dose is administered daily. The dose may be divided for a number of administrations during the day (say 2-4 times a day). The dose can also be administered every two days, two times a week, three times a week, biweekly, weekly doses, or separated by several weeks (for example 2 to 8).

As used herein the term "about" refers to $\pm 10\%$.

The terms "comprises", "comprising", "includes", "including", "having" and their conjugates mean "including but not limited to".

The term "consisting of" means "including and limited to".

The term "consisting essentially of" means that the composition, method or structure may include additional ingredients, steps and/or parts, but only if the additional ingredients, steps and/or parts do not materially alter the basic and novel characteristics of the claimed composition, method or structure.

As used herein, the singular form "a", "an" and "the" include plural references unless the context clearly dictates otherwise. For example, the term "a compound" or "at least one compound" may include a plurality of compounds, including mixtures thereof.

Throughout this application, various embodiments of this invention may be presented in a range format. It should be understood that the description in range format is merely for convenience and brevity and should not be construed as an inflexible limitation on the scope of the invention. Accordingly, the description of a range should be considered to have specifically disclosed all the possible subranges as well as individual numerical values within that range. For example, description of a range such as from 1 to 6 should be considered to have specifically disclosed subranges such as from 1 to 3, from 1 to 4, from 1 to 5, from 2 to 4, from 2 to 6, from 3 to 6 etc., as well as individual numbers within that range, for example, 1, 2, 3, 4, 5, and 6. This applies regardless of the breadth of the range.

Whenever a numerical range is indicated herein, it is meant to include any cited numeral (fractional or integral) within the indicated range. The phrases "ranging/ranges between" a first indicate number and a second indicate number and "ranging/ranges from" a first indicate number "to" a second indicate number are used herein interchangeably and are meant to include the first and second indicated numbers and all the fractional and integral numerals therebetween.

As used herein the term "method" refers to manners, means, techniques and procedures for accomplishing a given task including, but not limited to, those manners, means, techniques and procedures either known to, or readily developed from known manners, means, techniques and procedures by practitioners of the chemical, pharmacological, biological, biochemical and medical arts.

As used herein, the term "treating" includes abrogating, substantially inhibiting, slowing or reversing the progression of a condition, substantially ameliorating clinical or aesthetical symptoms of a condition or substantially preventing the appearance of clinical or aesthetical symptoms of a condition.

It is appreciated that certain features of the invention, which are, for clarity, described in the context of separate embodiments, may also be provided in combination in a single embodiment. Conversely, various features of the invention, which are, for

brevity, described in the context of a single embodiment, may also be provided separately or in any suitable subcombination or as suitable in any other described embodiment of the invention. Certain features described in the context of various embodiments are not to be considered essential features of those embodiments, unless
5 the embodiment is inoperative without those elements.

Various embodiments and aspects of the present invention as delineated hereinabove and as claimed in the claims section below find experimental support in the following examples.

10 EXAMPLES

Reference is now made to the following examples, which together with the above descriptions illustrate some embodiments of the invention in a non limiting fashion.

Generally, the nomenclature used herein and the laboratory procedures utilized in the present invention include molecular, biochemical, microbiological and recombinant
15 DNA techniques. Such techniques are thoroughly explained in the literature. See, for example, "Molecular Cloning: A laboratory Manual" Sambrook et al., (1989); "Current Protocols in Molecular Biology" Volumes I-III Ausubel, R. M., ed. (1994); Ausubel et al., "Current Protocols in Molecular Biology", John Wiley and Sons, Baltimore, Maryland (1989); Perbal, "A Practical Guide to Molecular Cloning", John Wiley &
20 Sons, New York (1988); Watson et al., "Recombinant DNA", Scientific American Books, New York; Birren et al. (eds) "Genome Analysis: A Laboratory Manual Series", Vols. 1-4, Cold Spring Harbor Laboratory Press, New York (1998); methodologies as set forth in U.S. Pat. Nos. 4,666,828; 4,683,202; 4,801,531; 5,192,659 and 5,272,057; "Cell Biology: A Laboratory Handbook", Volumes I-III Cellis, J. E., ed. (1994);
25 "Culture of Animal Cells - A Manual of Basic Technique" by Freshney, Wiley-Liss, N. Y. (1994), Third Edition; "Current Protocols in Immunology" Volumes I-III Coligan J. E., ed. (1994); Stites et al. (eds), "Basic and Clinical Immunology" (8th Edition), Appleton & Lange, Norwalk, CT (1994); Mishell and Shiigi (eds), "Selected Methods in Cellular Immunology", W. H. Freeman and Co., New York (1980); available
30 immunoassays are extensively described in the patent and scientific literature, see, for example, U.S. Pat. Nos. 3,791,932; 3,839,153; 3,850,752; 3,850,578; 3,853,987; 3,867,517; 3,879,262; 3,901,654; 3,935,074; 3,984,533; 3,996,345; 4,034,074;

4,098,876; 4,879,219; 5,011,771 and 5,281,521; "Oligonucleotide Synthesis" Gait, M. J., ed. (1984); "Nucleic Acid Hybridization" Hames, B. D., and Higgins S. J., eds. (1985); "Transcription and Translation" Hames, B. D., and Higgins S. J., eds. (1984); "Animal Cell Culture" Freshney, R. I., ed. (1986); "Immobilized Cells and Enzymes" IRL Press, (1986); "A Practical Guide to Molecular Cloning" Perbal, B., (1984) and "Methods in Enzymology" Vol. 1, 2, 317, Academic Press; "PCR Protocols: A Guide To Methods And Applications", Academic Press, San Diego, CA (1990); Marshak et al., "Strategies for Protein Purification and Characterization - A Laboratory Course Manual" CSHL Press (1996); all of which are incorporated by reference as if fully set forth herein.

Other general references are provided throughout this document. The procedures therein are believed to be well known in the art and are provided for the convenience of the reader. All the information contained therein is incorporated herein by reference.

EXAMPLE 1

MATERIALS AND EXPERIMENTAL PROCEDURES

Expression constructs and expression

cDNA encoding prh TNFR2:Fc was optimized and synthesized by GENEART AG (Regensburg, Germany). The codon usage was adapted to the codon bias of *Nicotiana tabacum* genes. The IgG1 portion was cloned from Fc IgG1 heavy chain constant region [Homo sapiens] ACCESSION AEV43323.

During the optimization process the following cis-acting sequence motifs were avoided: Internal TATA-boxes, chi-sites and ribosomal entry sites, AT-rich or GC-rich sequence stretches, RNA instability elements ("Killer motifs"), Repeat sequences and RNA secondary structures, splice donor (cryptic) and acceptor sites, branch points. In addition, regions of very high (>80%) or very low (<30%) GC content were avoided. The resultant DNA sequence is as set forth in SEQ ID NO: 1. The encoded polypeptide is as set forth in SEQ ID NO: 2. To the native cDNA sequence, a signal peptide (e.g. endoplasmic reticulum target signal peptide) from *N. plumbaginifolia* Calreticulin protein was added to the N' terminus of the gene, allowing efficient targeting of Prh TNFR2:Fc to the secretory pathway and is then cleaved from the polypeptide, by signal peptidase, once the protein has been translocated into the endoplasmic reticulum (SEQ ID NO: 3, SEQ ID NO: 4, representing the DNA and peptide sequences of the ER

signal peptide, respectively). Additionally, an ER retention signal SEKDEL was added to the C' terminus of the gene. This signal allows protein retrieval from the Golgi apparatus to the ER, and localization in the ER. The entire coding sequence (signal peptide- prh TNFR2:Fc-SEKDEL) is encoded by SEQ ID NO: 5 and the encoded polypeptide is as set forth in SEQ ID NO: 6. The resultant protein following cleavage of the N-terminal signal peptide is as set forth in SEQ ID NO: 7, 204 or 205 (prh TNFR2:Fc-SEKDEL).

Stable expression in N. tabacum BY2 cells

Agrobacterium mediated transformation is widely used to introduce foreign genes into a plant cell genome. Using this approach, a T-DNA molecule consisting of a foreign gene and its regulatory elements is randomly introduced into the plant genome. Since the site of integration, as well as the copy number of the gene insertions cannot be controlled, the transformation process results in a highly heterogeneous transgenic 'pool' composed of cells with various levels of transgene expression. The transgenic 'pool' is subsequently used for clone isolation. The transformation process, results in establishment of numerous single cell lines, each representing an individual transformation event, from which the clone with the highest expression level of the foreign gene is selected. For prh TNFR2:Fc (PRH TNFR2:FC) the transformation was conducted with a plasmid carrying the prh TNFR2:Fc cassette (Figure 1 SEQ ID NOS: 7 and 8). As a result, the recombinant protein is targeted to the Endoplasmic reticulum (ER) of the cells. The transformations of the BY2 cells with the PRH TNFR2:FC-ER expression vector were performed by the Agrobacterium tumefaciens mediated plant transformation procedure as follow: BY2 (Bright Yellow 2) suspension culture was co-cultivated, for 48 hours, with the Agrobacterium tumefaciens strain carrying the vector harboring the prhTNFR2:FC- gene and the neomycin phosphotransferase (NPTII) selection gene. Subsequently, cells were kept in media supplemented with 50mg/L of Kanamycin and 250mg/L Cefotaxime. The NPTII gene confers resistance to Kanamycin, thus only NPTII positive BY2 cells survive in this selection media. The Cefotaxime was used to selectively kill the agrobacterium, the plant cells being resistant to this antibiotic.

Screening for the optimal expressing clone

In order to select individual cell lines, aliquots of highly diluted cell suspension were spread on solid BY-2 medium (Toshiyuki Nagata & Fumi Kumagai *Methods in Cell Science* **21**: 123–127, 1999). The cells were then grown until small calli developed. Each callus was then re-suspended in liquid culture. Cells were then sampled and evaluated for PRH TNFR2:FC. About 500 cell line were screened by Western blot under denaturing conditions (Figure 4). The lines with high expression levels were further re-analyzed by the same method to select the highest expressing clone of prh TNFR2:FC producing clone.

Gel electrophoresis:

Sodium Dodecyl Sulfate-Polyacrylamide Gel Electrophoresis (SDS-PAGE) separates proteins on an electrical field according to their size. Proteins in the presence of the detergent SDS migrate as a linear function of the logarithm of their molecular weight. Migration pattern and identification of PRH TNFR2:FC on SDS-PAGE was compared to commercial molecular weight standard proteins (New England BioLabs; cat No. P7708S) and to the commercially available, mammalian-cell derived Enbrel® expressed in CHO cells (Entanercept; Wyeth). PRH TNFR2:FC was extracted from cells either by reducing sample buffer containing β -mercaptoethanol or by native extraction buffer. The native extraction supernatant was mixed with non-reducing sample buffer prior to analysis. Electrophoresis was performed using Criterion™ cell vertical electrophoresis apparatus (Bio-Rad Lab.) with premixed electrophoresis Tris-Glycine-SDS running buffer (Bio-Rad Laboratories). Following electrophoresis, the proteins were transferred from the Polyacrylamide gel to a protein binding nitrocellulose membrane (iBlot™). Membranes were blocked for 1hr at RT with 5% milk buffer containing 0.1% Tween 20. For identification of the Fc portion of the molecule, Goat anti human IgG conjugated to HRP (cat # 109-035-098, Jackson.) was used. For TNFR2 detection, a Rabbit Anti-TNFR2 (ID: ab109853, Abcam) followed by Goat anti Rabbit HRP (cat # 111-035-003, Jackson) were employed. Detection was carried out with ECL detection kit (Pierce). The immunoreactivity of PRH TNFR2:FC was compared to that of commercial Enbrel® (Entanercept; Wyeth). Bands were detected using the Molecular Imager Gel Doc XR System (Bio-Rad Laboratories).

Amino acid sequencing by Mass-spectrometry

prhTNFR2:FC is sent for sequencing analysis at the Smoler Proteomics Center at the Technion - Israel Institute of Technology (Haifa, Israel). The protein is extracted from the gel, reduced with 2.8mM DTT (60°C for 30 min), modified with 8.8mM iodoacetamide in 100mM ammonium bicarbonate (in the dark, room temperature for 30 min) and digested in 10% ACN and 10mM ammonium bicarbonate with modified Trypsin (Promega) or with ChymoTrypsin overnight at 37°C in a 1:50 enzyme-to-substrate ratio. 3% of the resulting peptides are resolved by reverse-phase chromatography on 0.075 X 200-mm fused silica capillaries (J&W) packed with Reprosil reversed phase material (Dr Maisch GmbH, Germany). The peptides are eluted with linear 60 minutes gradients of 5 to 45 % and 15 minutes at 95 % acetonitrile with 0.1 % formic acid in water at flow rates of 0.25 µl/min. On line mass spectrometry is performed by an ion-trap mass spectrometer (Orbitrap, Thermo) in a positive mode using repetitively full MS scan followed by collision induces dissociation (CID) of the 7 most dominant ion selected from the first MS scan.

The mass spectrometry data is analyzed using the Sequest 3.31 software (J. Eng and J.Yates, University of Washington and Finnigan, San Jose) vs a specific sequence.

Glycosylation analysis

The major difference between glycoproteins produced in Chinese Hamster Ovary (CHO) cell and plant cell systems is the glycosylation profile and glycan structure. Preliminary analysis has been performed to characterize the various N-linked glycan structures attached to the protein. These results are compared to results of the N-glycosylation profile found in commercial Enbrel®. The presence of O-linked glycans, and glycan site analysis is determined.

Samples of PRH TNFR2:FC and commercial Enbrel are reduced, alkylated and separated on SDS-PAGE. The protein bands at ~75 KDa (a total of about 200 µg protein) are taken for glycan analysis using trypsin digestion followed by either PNGase A or PNGase F digestion (~80% and ~20% of the total protein, respectively) for PRH TNFR2:FC and PNGase F digestion only for commercial Enbrel. Digestion with Trypsin, followed by PNGase A releases all the N-linked glycans and digestion with PNGase F releases all glycans except those containing alpha 1-3 core fucose (found in plants). The released glycans are extracted, cleaned and then labeled with the

fluorescent reagent anthranilamide (2-aminobenzamide, 2AB) followed by removal of excess 2AB. The analytical method includes separation of the glycans on a Waters HPLC system with a normal phase amide-based column (Tosoh TSK Amide-80 column), coupled with a fluorescence detector (330 nm excitation, 420 nm emission).

5 Sequencing of the labeled glycan pool is achieved by sequential digestion with various exoglycosidases followed by additional HPLC analysis. Using sequential digestion with various exoglycosidases provides additional information on the profile of the glycans structures and their relative amounts. The exoglycosidase digestions that are carried out for the glycans released from PRH TNFR2:FC are with JBH (Jack bean
10 beta-N-Acetylhexosaminidase) that removes beta 1-2, 3, 4 and 6 *N*-acetylglucosamine (GlcNAc), with JBM (Jack bean mannosidase) that removes mannose alpha 1-2, 6 > 3 mannose and with BKF (Bovine testis fucosidase) that removes alpha 1-6 and alpha 1-3 core fucose. The fluorescence labeling enables a semi-quantitative analysis of the distribution of the various glycan structures in the total digested glycan pool. The
15 glycans are then separated according to unique glycan linkages and in order of increasing size using a gradient solvent flow consisting of ammonium formate and acetonitrile. Retention time of individual glycans is compared to the retention times of a standard mix of partially hydrolysed dextran fragments, giving a ladder of glucose units (GU). The glycans are assigned to peaks according to their GU values, based on
20 standards and a comparison to an external data base (glycobase website 8080). The final assignment and relative peak areas are calculated from the chromatogram of the PNGase A digestion.

Enzyme-linked immunosorbent assay (ELISA)

Binding ELISA: TNF α binding ELISA is a combination of a commercial
25 TNF α detection ELISA kit (Human TNF- α ; Hycult Biotech Inc.#HK307) and a commercial anti human IgG antibody (Goat anti human IgG FC specific HRP; Sigma). The assay is a quantitative non radioactive assay for prhTNFR2:FC binding activity. This binding ELISA enables to detect functional (capable of binding TNF α) molecules comprising both the TNFR and IgG domains.

30 An ELISA plate pre-coated with antibodies against TNF α was incubated with TNF α (60ng/ml, Sigma) for 1 hour at room temperature. Between each ELISA step the plate was washed three times with commercial wash buffer. Commercial Enbrel and

supernatant from BY2 cells expressing PRH TNFR2:Fc (serial dilutions) were incubated on ELISA plate for 2hr at RT. Goat anti human IgG Fc HRP was diluted 1:10,000 and incubated on plate for 1hr at RT. TMB was used as substrate for HRP. The colorimetric reaction was stopped with 10% HCL and absorbance determined at 450nm.

Prevention of TNF α induced apoptosis in A375 cells

A375 cells (human melanoma cells) were grown in suspension in culture medium (ATCC, # 30-2002, supplemented with 10% FBS). 10^4 /well cells were plated in 96-well assay plates and incubated overnight in assay medium (ATCC, # 30-2002, supplemented with 5% FBS). Recombinant TNF α (2ng/ml, ProSpec, Rehovot, Israel) was incubated for 2 hr at 37 °C in the presence of different concentrations (1.562-100ng/ml) of prhTNFR2:Fc or commercial Enbrel (Entanercept; Wyeth). Following incubation, the mixed solution was added to A375 cells in the presence of actinomycin-D (0.8 μ g/ml), incubated for further 24hr at 37°C, 5% CO₂ in a humidified incubator and quantification of apoptosis was determined by MTT assay (Sigma Cat. No. M5655). The plate was read at 570-650nm and the inhibition of TNF- α induced cytotoxicity (%) was calculated.

EXAMPLE 2

PROTEIN ANALYSIS

prhTNFR2:Fc was analyzed under reducing (Figure 2A) and non-reducing conditions (native extraction in the Figure 2B). prhTNFR2:Fc (Lane 1) and commercial Enbrel (lane 2) were detected using anti Fc antibody (upper panel) and anti TNFR2 antibody (lower panel). The two proteins demonstrate a slight difference in migration characteristics, presumably due to differences in glycosylation patterns between the plant and mammalian cell-expressed enzymes.

TNF α binding by both commercial Enbrel and prh TNFR2:Fc was examined by comparing serial dilutions of BY2 cells expressing prh TNFR2:Fc (PRX- 106) lysates to commercial Enbrel. prh TNFR2:Fc serial dilutions demonstrate a dose response binding pattern similar to the commercial protein (see Figure 3). The selection of transgenic cell lines according to protein expression was done by Western blotting. Thus, to allow for the selection of individual cell lines, aliquots of highly diluted cell suspension were spread on solid BY-2 medium. The cells were then grown until small

calli developed. Each callus was then re-suspended in liquid culture. Cells were then sampled and evaluated for prh TNFR2:Fc expression levels by extraction under reducing conditions followed by Western Blot identification (anti FC antibody) of the produced target protein (Figure 4). The functionality of the expressed protein was established by its ability to prevent TNF α induced apoptosis. Specifically, TNF α activity can be measured by its ability to induce cell death of certain cell lines in the presence of the transcriptional inhibitor, actinomycin D. Pre-incubation with a neutralizing protein of TNF α prevents binding to the receptors (TNF-R1 and TNF-R2), thereby inhibiting the cytokine effect and preventing TNF α induced cell death. Quantification of cell viability by MTT assay provides an in-cell activity assay for TNF α cytotoxicity. The results are shown in Figures 5A-G on melanoma cells A375 and in Figures 6A-G on L929 fibroblasts.

EXAMPLE 3

ORAL ADMINISTRATION OF PLANT CELLS EXPRESSING RECOMBINANT TNFR2:Fc EFFECTIVELY REDUCES HEPATOTOXICITY IN THE CON A IMMUNE-MEDIATED HEPATITIS MODEL

The Concanavalin A (Con A) model is well established animal model for investigating T-cell, Natural killer (NK) T cells (NKT) and macrophage dependent liver injury, which closely mimics the pathogenesis and pathological changes characteristic to Immune-Mediated Hepatitis. Amelioration of hepatotoxicity by oral administration of plant cells expressing recombinant TNFR2:Fc in this model of immune-mediated hepatitis provides evidence for effective anti-inflammatory capabilities of the plant cells expressing a recombinant TNFR2:Fc.

Materials and Methods

Animals: Male, C57Bl/6 mice, 11-12 weeks old were used in all experiments. Each experimental group included 5 to 8 mice.

Con A model: Concanavalin A (MP Biomedicals, OH, USA), dissolved in 50 mM Tris (pH 7), 150 mM sodium chloride, 4 mM CaCl₂ was administered intravenously into the tail vein at a dose of 20mg/Kg body weight. Mice were sacrificed 14 hours after Con A administration and blood samples were collected by cardiac puncture, allowed to coagulate and serum removed for determination of serum

liver enzymes (alanine aminotransferase, ALT and aspartate aminotransferase, AST) and cytokine (IFN-gamma) levels. Livers were excised and prepared for histopathological evaluation (see below).

Oral administration of recombinant plant cells: Oral administration of plant cells expressing recombinant TNFR2:Fc was initiated 6 hours before administration of Con A. Mice received plant cells expressing recombinant TNFR2:Fc, equivalent to 0.5µg (X1) or 5µg (X10) TNFR2:Fc protein, freshly prepared by emulsification in saline. Negative controls received the same orally administered volumes of host BY2(-) plant cells, in place of the plant cells expressing recombinant TNFR:Fc. Oral administration was performed by gavage in a volume of 350 µl.

Steroid Controls: Steroid treatment was provided by oral administration of 0.35 mg Dexamethasone (Teva, Israel) per mouse, 6 hours prior to the administration of Con A.

Hepatotoxicity: Liver enzymes (alanine aminotransferase, ALT and aspartate aminotransferase, AST), markers of damage to the liver parenchyma, were evaluated in serum using a Reflovet Plus clinical chemistry analyzer (Roche Diagnostics, Mannheim Germany). Cytokine (IFN-gamma) levels were evaluated in the serum of the treated and control mice by ELISA, using the Quantikine Colorimetric Sandwich ELISA kit (R&D Systems, Minneapolis MN, USA).

Pathology: Histopathology was determined in individual livers after fixation of the tissue in 10% formaldehyde and storage at room temperature, embedding in paraffin, sectioning and staining with hematoxylin and eosin (H&E) for morphological and histological examination by light microscopy.

Results:

In three separate series of experiments, oral administration of plant cells expressing recombinant TNFR2:Fc, at both low doses (equivalent to 0.5µg TNFR2:Fc protein “X1”) and higher doses (equivalent to 5µg TNFR2:Fc protein “X10”) significantly reduced the hepatotoxic effects of Con A. Elevation of serum enzyme markers of liver damage (AST, ALT) was largely prevented in all three experiments (see Figures 7A, 7B and 7C), with efficacy approaching that of oral steroid treatment (Figures 8A, 8B and 8C, Dex.).

Determination of the cytokine IFN-gamma in serum of the mice 14 hours after Con A administration (see Figures 8A, 8B and 8C) also revealed a significant reduction in serum IFN-gamma in the groups receiving oral administration of plant cells expressing recombinant TNFR2:Fc, at both low doses (equivalent to 0.5µg TNFR2:Fc protein “X1”) and higher doses (equivalent to 5µg TNFR2:Fc protein “X10”).

Histopathological evaluation (hematoxylin and eosin) of the livers of treated and control mice (Figures 9A, 9B and 9C) revealed severe hepatic necrosis in the control livers (Figure 9A), but preservation of liver architecture and normal liver histology in the livers of the mice treated with plant cells expressing recombinant TNFR2:Fc (Figure 9B).

Oral administration of plant cells expressing recombinant TNFR2:Fc (low dose, equivalent to 0.5µg TNFR2:Fc protein) and intraperitoneal administration of 100 µg of the commercial mammalian cell expressed TNFR2:Fc Etanercept (ENBREL®, Wyeth) were compared for their effect on hepatotoxicity in the Con A immune-mediated hepatitis model. Comparing levels of serum liver damage markers AST and ALT in the treated mice, relative to untreated controls revealed that even a low dose of the plant cells expressing recombinant TNFR2:Fc, orally administered, was as effective (87-85%, Figures 10A and 10B) as 100 µg of Etanercept administered intraperitoneally in preventing elevation of liver damage markers in response to Con A-induced immune-mediated hepatitis.

EXAMPLE 4

ORAL ADMINISTRATION OF PLANT CELLS EXPRESSING RECOMBINANT TNFR2:Fc EFFECTIVELY AMELIORATES IMMUNO-PATHOGENESIS IN FATTY LIVER DISEASE MODELED BY HIGH FAT DIET (HFD)

A well accepted animal model of fatty liver disease is induced by High Fat Diet (HFD). Mice with diet-induced obesity are characterized by elevated serum lipid profile, increased hepatic triglycerides and immune system alterations.

The effect of orally administered plant cells expressing recombinant TNFR2:Fc on mice fed with HFD was determined. Analysis included the effect of the treatment on clinical manifestations of the disease and as an immunomodulator.

Materials and methods

Animals: Male C57bl/6 mice, 6-7 weeks old were used in all experiments. Each experimental group included 10 mice. The mice were purchased from Harlan Laboratories, Jerusalem, Israel. All mice were fed with HFD (Harlan, TD88137 in which 42% of calories are from fat) from day 0 until their sacrifice, after 24 weeks.

Table 1 (Experimental design):

Group	N	High fat diet	Treatment
A	10	+	PO 35µl Saline 3 days a week
B	10	+	PO, 28.8mg BY- (mock cells), 3 days a week
C	10	+	PO, 2.88mg (0.5µg TNF) BY+ 3 days a week
D	10	+	PO, 2.88mg (10 µg anti TNF) BY+ 3 days a week

Oral administration of recombinant plant cells: Oral administration of plant cells expressing recombinant TNFR:Fc (batch **Ly013+**) was initiated 3 times a week. Negative controls received the same orally administered dose of mock cells (BY-). All oral administrations to mice were in a total volume of 35 µl. Fresh preparations were made before each administration.

Endpoint measured on a weekly basis:

1. Body weight

Endpoints measured on a once-a month basis:

1. Fasting blood glucose levels
2. Serum ALT, AST levels*
3. Serum triglycerides levels*

* Monitoring of serum liver enzymes and triglycerides was by measuring the Reflovet Plus clinical chemistry analyzer (Roche Diagnostics, GmbH, Mannheim, Germany).

Additional Endpoints:

1. Fasting serum Insulin levels on day 1 and on week 24 (ELISA).
2. Glucose tolerance test (GTT) on week 8 and on week 24.
3. Liver fat content (triglycerides); after sacrifice
4. Liver histology, after sacrifice
5. Serum cytokine levels (TNF- α), after sacrifice (ELISA).

6. Flow cytometry (FACS) for subsets of T cells and Tregs (spleen and liver).
7. CD8- APC/CD4-FITC/CD25-PE/ Foxp3-PE-Cy7
8. CD3- FITC/NK 1.1-APC

5

Following sacrifice of the mice (at week 24):

Cytokine secretion: Cytokine (TNF- α) levels were measured in the serum of treated and control mice by ELISA, using the Quantikine Sandwich ELISA Kit (R&D Systems, Minneapolis, MN, USA).

- 10 ***Histopathology:*** Livers were excised and then fixed in 10 % formaldehyde, embedded in paraffin, sectioned and stained with H&E and with Mason trichome (for fibrosis). H&E tissues were examined and scored by light microscopy for morphological and histo-pathological changes characteristic for NASH by a blinded pathologist.

- 15 ***Triglyceride determination:*** Accumulation of intracellular triglycerides (TG) within the liver was quantified using a modification of the Folch method. TG were extracted from aliquots of snap-frozen livers and then assayed spectrophotometrically using the GPO-Trinder kit (Sigma, Rehovot, Israel) and normalized to the protein content in the homogenate.

FACS analysis was performed for subsets of T cells and Tregs taken from spleen.

20 ***Results***

- The effect of orally administered plant cells expressing recombinant TNFR2:Fc on serum enzymes was tested. As can be seen in Figure 11, oral administration of the cells expressing the inhibitor, caused a decrease in AST levels in the treated mice, as measured on sacrifice day (week 24). A trend of decrease in ALT levels was also
25 evident (data not shown).

- The effect of orally administered plant cells expressing recombinant TNFR2:Fc on serum triglycerides (TG) was tested. As can be seen in Figure 12, oral administration of the cells expressing the inhibitor, caused a significant decrease in TG levels in the treated mice, on sacrifice day (week 24). Importantly, the results obtained
30 support therapeutic efficacy of the TNFR2:Fc, since the effect on serum enzymes and TGs was evident despite persistent gain weight in all groups tested (Figure 13).

Next the distribution of T cell subpopulation in the liver and spleen of the model mice was tested by histopathology and FACS. Figure 14 shows the results of hepatic Tregs. As can be seen, intra hepatic Tregs significantly decreased in the high dose treated mice.

Figure 15 shows the results of hepatic NK cells. As can be seen, intra hepatic NK cells significantly increased in the high dose treated mice.

Figure 16 shows the results of the effect of oral administration of plant cells expressing recombinant TNFR2:Fc on splenic/hepatic CD4+CD25+FOXP3+ ratio. As can be seen, an increase in the ratio of spleen to liver for Tregs (CD4+CD25+FOXP3+) was noted.

0.5µg of PRX-106 increased this ratio by 10% and 10µg of PRX-106 increased this ratio by 22%, compared to saline-treated mice.

Figure 17 shows the results of the effect of oral administration of recombinant TNFR2:Fc in plant cells on splenic/hepatic CD8+CD25+FOXP3+ ratio. As can be seen, a considerable increase in the ratio of spleen to liver was noted for another subset of cells: CD8+CD25+FOXP3+ cells. Low dose of 0.5 µg of the drug increased this ratio by 74% compared to saline-treated mice.

These results suggest that oral administration of recombinant TNFR2:Fc in plant cells alters the T cells distribution affecting the intrahepatic to periphery (splenic) T cell functions in HFD mice modeling a fatty liver disease.

EXAMPLE 5

TOXICOLOGY STUDIES IN MICE

Methods

Animals

Male and female SD Rats (Harlan Laboratories, Israel) 8 weeks at study initiation were housed under standard laboratory conditions. Mean weight at study initiation was approximately 6.8 gr for males and 6.3 gr for females. Animals were fed with commercial rodent diet (Teklad Certified Global 18% Protein Diet cat #: 2018SC) and had free access to autoclaved and acidified drinking water (pH between 2.5 and 3.5).

Study design

Four groups, 3 dosing groups comprising 12 rats per group (6 males and 6 females) and a control group comprising 6 rats per group (3 males and 3 females), were assigned. In each gender, the control group received dilution buffer (0.2 M mannitol) and three treated groups received cells expressing TNFR2:Fc at dose levels of 0.1, 0.5 and 1 mg TNFR2:Fc/Kg body weight. Cells were aliquoted in accordance with requested expressed protein amount. Each aliquot was mixed with 30 grams powder of commercial rodent diet and dilution buffer, to create a pellet. The control pellet was made with dilution buffer and commercial rodent diet powder alone. All animals were daily orally fed with the pellets for 14 days. During the study, mortality and general clinical observation were performed, bodyweight was monitored daily. At study termination (Day 15) after light anesthesia with carbon dioxide inhalation, three blood samples were drawn from all animals from the retro orbital sinus gross, after which, animals were sacrificed, pathology was executed and selected organs were harvested .

Results

No adverse clinical symptoms were recorded throughout the 14-day safety study. All blood parameters were within the normal range with no significant deviations. Body weight gain was persistent and normal with no significant difference between the groups (treated or Control). Cells expressing were found to be safe and well tolerated with no adverse effects. No effect on biochemical parameters or clinical symptoms was found. Gross necropsy observation did not reveal pathological findings. No animal was found in a moribund state or under severe distress conditions. There were no observations of animals presenting severe pain or decreased body weight.

EXAMPLE 6

SEQUENCING OF PRX-106

N terminus sequencing by Edman degradation

Analysis was performed at Alphalyse (Denmark) using an ABI Procise 494 sequencer. The procedure determines the N-terminal amino acid sequence of proteins and peptides by the Edman degradation chemistry. The Edman degradation is a cyclic procedure where amino acid residues are cleaved off one at a time and identified by chromatography. Here are 3 steps in the cyclic procedure. In step 1, the PITC reagent

is coupled to the N-terminal amino group under alkaline conditions. In step 2, the N-terminal residue is cleaved in acidic media. In step 3, the PITC coupled residue is transferred to a flask, converted to a PTH-residue and identified by HPLC chromatography. The next cycle is then started for identification of the next N-terminal residue.

Results:

The sequence was determined to be **LPAQV (SEQ ID NO: 18)**.

Amino Acid Sequence verification by reverse phase HPLC coupled to a Mass Spectrometry detector.

Sequencing was performed at the Smoler Proteomics Center (Technion- Israel Institute of Technology, Haifa, Israel). Analyses were carried out using reverse-phase HPLC coupled to a mass spectrometry detector.

Method

Proteolysis

The analyzed samples were resuspended in 8 M Urea, 100 mM ammonium bicarbonate (ABC) followed by reduction with 2.8 mM DTT (60°C for 30 min) and modified with 8.8 mM iodoacetamide in 100 mM ABC in the dark, at ambient temperature for an additional 30 min. The proteins were digested overnight at 37 °C using modified trypsin (Promega) at a 1:50 enzyme-to-substrate ratio in 2 M Urea, 25 mM ABC.

Mass spectrometry analysis

The tryptic or chymotryptic peptides were desalted using stage tips (home-made C18), the residual buffer was evaporated and the pellet was resuspended in 0.1 % (v/v) formic acid. Twenty nanogram of the resulting peptides were resolved by reversed-phase liquid chromatography on a 0.075X200-mm fused silica capillaries (J and W) packed with Reprosil reversed phase material (Dr Maisch GmbH, Germany). Peptides were eluted with a linear 60 minutes gradient of 5 to 45 % followed by 15 minutes at 95 % acetonitrile with 0.1 % formic acid in water at flow rates of 0.25 µL/min. On-line mass spectrometry was performed on an ion-trap mass spectrometer (Orbitrap, Thermo) in a positive mode using repetitively full MS scan followed by collision induced dissociation (CID) of the 7 most dominant ions selected from the first MS scan. The

mass spectrometry data was analyzed using the Discoverer software version 1.3 software using a specific protein derived database.

Results

The sequence was compared to the peptide sequence of the Etanercept sequence.

- 5 The identified sequences are presented in Table V, below. Presented is 84.8% coverage of the reference sequence (see green color, Figure 20).

Table V- Peptides Identified Following Digestion with Trypsin (SEQ ID NO: 19-203, ordered)

WQQGnVFScSVMHEALHnHYTQK
 WQQGNVFScSVMHEALHNHYTqK
 GFYPSDIAVEWESNGqPENnYKT
 qYNSTYRVVSVLTVLHqDWLNGK
 WQqGNVFScSVMHEALHNHYTqKS
 VVSVLTVLHQDWLNGKEYKc
 VVSVLTVLHqDWLnGKEYK
 SqHTqPTPEPSTAPSTSfLLPmGPSPPAEGSTGDEPK
 WQQGnVFScSVMHEALHNHY
 ScDKTHTcPPcPAPELLGGPSVFLFPPKPKD
 GQPREPqVYTLPPSREEMTK
 GFYPSDIAVEWESNGQPEnNYKT
 LPAqVAFTPYAPEPGSTcR
 EALHnHYTqK
 qNRITcRPGWYcALSKQEGcR
 WQQGNVFScSVMHEALHnHYTQK
 SqHTQPTPEPSTAPSTSfLLPMGPSPPAEGSTGDEPK
 GQPREPqVYTLPPSREEmTK
 GFYPSDIAVEWESnGQPENNYK
 SqHTQPTPEPSTAPSTSfLLPmGPSPPAEGSTGDEPK
 VVSVLTVLHQDWLnGK
 TYTqLWNWVPEcLSGSRCSSDqVETQAcTR
 WQQGNVFScSVMHEALHNHYTQK
 GFYPSDIAVEWESnGQPEnnYKT
 VVDVSHEDPEVK
 PSTSFLLPMGPSPPAEGSTGDEPK
 LPAQVAFTPYAPEPGSTcR
 TTPPVLDSDGSFFL
 LSLSPGK
 EPQVYTLPPSREEMTKN
 SmAPGAVHLPQ
 TTPPVLDSDGSFFLYSK
 WQQGNVFScSVMHEALHNHYTQK
 SMAPGAVH
 SVMHEALHNHYTQK
 VVSVLTVLH
 SQHTQPTPEPSTAPSTSfLLPMGPSPPAEGSTGDEPK
 GQPREPQVY
 AQVAFTPYAPEPGSTcR
 cAPLRK

EPQVYTLPPSREEmTKnQVSLTcLVK
SmAPGAVH
VSVLTVLHQD
LFPPKPK
GSFFLYSK
IcTcRPGWY
SQHTQPTPEPS
SVLTVLHQDWLnGKEYK
QVETQAcTR
SLSLSPGK
SDGSFFLYSK
KALPAPIEK
ALPAPIEK
AVcTSTSPTR
SQHTQPTPEPSTAPSTF
QVSLTcLVK
LREYYDQTAqmccSKcSPGQHAK
WQQGNVFSsVMHEALH
DTLmISR
PmGPSPPAEGSTGDEPK
THTcPPcPAPELLGGPSVF
DTLMISR
SDQVETQAcTR
KcRPGFGVAR
WYVDGVEVHNAK
YVDGVEVHNAK
TTPPVLDSDGSFF
THTcPPcPAPELLGGPSVFLFPPKPK
PSPPAEGSTGDEPK
SLSLSPGKSEK
MAPGAVHLPQPVSTR
VDGVEVHNAK
ScDKTHTcPPcPAPELLGGPSVF
VSVLTVLHQDWLNGK
SLSLSPGKSEK
PPcPAPELLGGPSVFLFPPKPK
SFFLYSK
FNWYVDGVEVHNAK
FLLPMGPSPPAEGSTGDEPK
DAVcTSTSPTR
NQVSLTcLVK
NqVSLTcLVKG
SLSPGKSEK
TPEVTcVVVDVSHEDPEVK
LREYYDQTAQM
GFYPSDIAVEWESNGQPENNYK
FNWYVDGVEVHN
VSVLTVLHQDWLN
SQHTQPTPEPSTAPST
RTPEVTcVVVDVSHEDPEVK
SLSLSPGKS
LSPGKSEKDEL

LPQPVSTR
TTPPVLDSDGSFFLY
TSDTVcDScEDSTYTQLWN
ALPAQVAFTPYAPEPGSTcR
EEQYNSTYR
ScDKTHTcPPcPAPELLGGPSVFLFPPKPK
cSPGQHAKVFcTK
TPEVTcVVVDVSHED
SMAPGAVHLPQPV
TcRPGWYcALSK
TcPPcPAPELLGGPSVFLFPPKPK
TSDTVcDScEDSTYTQLWNWVPEcLScGSR
LcAPLRK
SPPAEGSTGDEPK
WVPEcLScGSR
GPSPPAEGSTGDEPK
SSDQVETQAcTR
EEQYnSTYR
VAFTPYAPEPGSTcR
PGWYcALSK
cRPGFGVAR
ScSVmHEALHnHYTqK
VSVLTVLHQDWLNGKEYK
LcAPLR
EPQVYTLPPSREEMTKnQVSLTcLVK
LLPMGPSPPAEGSTGDEPK
SQHTQPTPEPSTAPSTSFLLPmGPSPPAEGSTGDEPK
SLSLSPGKSE
EEMTKNqV
SVMHEALHNHYTQK
SQHTQPTPEPSTAPSTSFLLPmGPSPPAEGSTGDEPKScDK
EEmTKnQVSLTcLVKG
LREYDQTAQmccSK
cSSDqVETQAcTR
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THTcPPcPAELLG
VSVLTVLHQDWLn
EPQVYTLPPSR
SmAPGAVHLPQPVSTR
GQPREPQVYTLPPSREEmTK
TPYAPEPGSTcR
EVTcVVVDVSHEDPEVK
TKPREEQYnSTYR

VS_nKALPAPIEK
 LREYYDQTAQM_{cc}SK
 FTPYAPEPGST_cR
 SMAPGAVHLPQPVSTR
 GPSVFLFPPKPK
 VVSVLTVLHQDWLnGKEYK
 SQHTQPTPEPSTAPS
 SMAPGAVHLPQPV_S
 AVHLPQPVSTR
 GQPREPQVYTLPPSR
 PGAVHLPQPVSTR
 TLMISR
 KNqVSLT_cLVKGFYPSDIAVEWESNGqPEN_nYK
 LREYYDQTAQM_c
 SmAPGAVHLPQPV
 LPAPIEK
 EYYDQTAQM_{cc}SK
 NWVPE_cLScGSR
 SLSPGKSEKDEL
 IcT_cRPGWY_cALSK
 SMAPGAVHLPQPVST
 EYYDQTAQM_{cc}SK
 ASMDAV_cTSTSPTR
 SQHTQPTPEPSTAPSTS
 TLPPSREEMTK
 SQHTQPTPEPSTAPSTSFL
 TLMISR
 EPQVYTLPPSREEmTK
 GQPREPQVYTLPPSREEMTK
 TPEVT_cVVDVSHEDPEVKFN
 ScDKTHT_cPPcPAPELLG
 GFYPSDIAVEWESNGqPEN_nYK
 AKGQPREPQVYTLPPSR
 LREYYDQTAQM_{cc}
 LPmGPSPPAEGSTGDEPK
 ScSVMHEALHNHYTQK
 FNWYVDGVEVHnAK
 PMGPSPPAEGSTGDEPK
 SMAPGAVHLPqPVSTR
 SMAPGAVHLPQ
 LPMGPSPPAEGSTGDEPK

Although the invention has been described in conjunction with specific
 embodiments thereof, it is evident that many alternatives, modifications and variations
 will be apparent to those skilled in the art. Accordingly, it is intended to embrace all
 5 such alternatives, modifications and variations that fall within the spirit and broad scope
 of the appended claims.

All publications, patents and patent applications mentioned in this specification are herein incorporated in their entirety by reference into the specification, to the same extent as if each individual publication, patent or patent application was specifically and individually indicated to be incorporated herein by reference. In addition, citation or
5 identification of any reference in this application shall not be construed as an admission that such reference is available as prior art to the present invention. To the extent that section headings are used, they should not be construed as necessarily limiting.

WHAT IS CLAIMED IS:

1. A method of treating a TNF α associated medical condition selected from the group consisting of obesity, metabolic syndrome, diabetes, hyperlipidemia and a liver disease or disorder, the method comprising enterally administering to a subject in need thereof a therapeutically effective amount of plant cells expressing a TNF α polypeptide inhibitor, thereby treating the TNF α associated medical condition.
2. Use of plant cells expressing a TNF α polypeptide inhibitor for the enteral treatment of a TNF α associated medical condition selected from the group consisting of obesity, metabolic syndrome, diabetes, hyperlipidemia and a liver disease or disorder.
3. The method of claim 1 or the use of claim 2, wherein said enteral is oral administration.
4. The method of claim 1 or the use of claim 2, wherein said TNF α polypeptide inhibitor is an anti-TNF α antibody.
5. The method of use of claim 4, wherein said anti-TNF α antibody is infliximab, adalimumab or golimumab.
6. The method of claim 1 or the use of claim 2, wherein said TNF α polypeptide inhibitor is a chimeric polypeptide comprising:
 - (i) a first domain which comprises a TNF α binding domain of a TNF receptor; and
 - (ii) a second domain which comprises an Fc domain of an immunoglobulin, wherein said first domain and said second domain are N-terminally to C-terminally respectively sequentially translationally fused and wherein the chimeric polypeptide specifically binds TNF α .
7. The method or use of claim 6, wherein said chimeric polypeptide further comprises a third domain which comprises an endoplasmic reticulum retention signal,

wherein said first domain, second domain and third domain are N-terminally to C-terminally respectively sequentially translationally fused.

8. The method or use of any one of claims 6-7, comprising an additional domain encoding an endoplasmic reticulum signal peptide translationally fused N-terminally to said first domain.

9. The method or use of claim 8, wherein said signal peptide is a plant signal peptide.

10. The method or use of claim 9, wherein said plant signal peptide is as set forth in SEQ ID NO: 4.

11. The method or use of any one of claims 6-7, wherein said first domain is 200-250 amino acids long.

12. The method or use of claim 11, wherein said first domain comprises the amino acid sequence LCAP (SEQ ID NO: 11) and VFCT (SEQ ID NO: 12).

13. The method or use of claim 12, wherein said first domain further comprises the amino acid sequence LPAQVAFXPYAPEPGSTC (SEQ ID NO: 13).

14. The method or use of claim 13, wherein said first domain is as set forth in SEQ ID NO: 2.

15. The method or use of any one of claims 6-7, wherein said immunoglobulin is IgG₁.

16. The method or use of any one of claims 6-7, wherein said second domain is as set forth in SEQ ID NO: 9.

17. The method or use of any one of claims 6-7, wherein said chimeric polypeptide is as set forth in SEQ ID NO: 6.

18. The method or use of claim 7, wherein said chimeric polypeptide is as set forth in SEQ ID NO: 7, 204 or 205.

19. The method or use of any one of claims 6-7, wherein said chimeric polypeptide is capable of inhibiting TNF α -induced apoptosis.

20. The method or use of any one of claims 1-2, wherein said TNF α polypeptide inhibitor comprises a plant-specific glycan.

21. The method or use of claim 20, wherein said plant-specific glycan is selected from the group consisting of a core xylose and a core α -(1,3) fucose.

22. The method or use of any one of claims 1-2, wherein said plant cells are *Nicotiana tabacum* plant cells.

23. The method or use of claim 22, wherein said *Nicotiana tabacum* plant cell is a Bright Yellow (BY-2) cell.

24. The method or use of any one of claims 1-2, wherein said plant cells are lyophilized.

25. The method or use of any one of claims 1-2, wherein said plant cells are grown in suspension.

26. The method or use of any one of claims 1-2, wherein said liver disease or disorder is selected from the group consisting of hepatitis, liver cirrhosis, liver cancer, hepatotoxicity, chronic liver disease, fatty liver disease and non-alcoholic steatohepatitis (NASH).

27. The method or use of claim 26, wherein said hepatotoxicity is induced by a chemical agent selected from the group consisting of acetaminophen, NSAIDS, glucocorticoid, isniazed, arsenic, carbon tetrachloride and vinyl chloride.

28. The method or use of claim 26, wherein said diabetes is selected from the group consisting of type I diabetes, type II diabetes and LADA disease.

29. The method or use of any one of claims 1-2, wherein said plant cells are provided in an oral nutritional form.

30. The method or use of claim 29, wherein said oral nutritional form is a complete meal, a powder for dissolution, a bar, a baked product, a cereal bar, a dairy bar, a snack-food, a breakfast cereal, muesli, candies, tabs, cookies, biscuits, crackers, chocolate, and dairy products.

31. The method or use of any one of claims 1-27, 29 or 30, wherein said liver disease or disorder is a fatty liver disease.

32. The method of use of claim 30, wherein said TNF α polypeptide inhibitor alters hepatic and splenic T cell distribution.

33. The method of use of claim 30, wherein said TNF α polypeptide inhibitor reduces a serum enzyme or a metabolite.

34. The method of use of claim 30, wherein said serum enzyme is aspartate aminotransferase (AST) or alanine aminotransferase (ALT).

35. The method of use of claim 30, wherein said metabolite is a triglyceride.

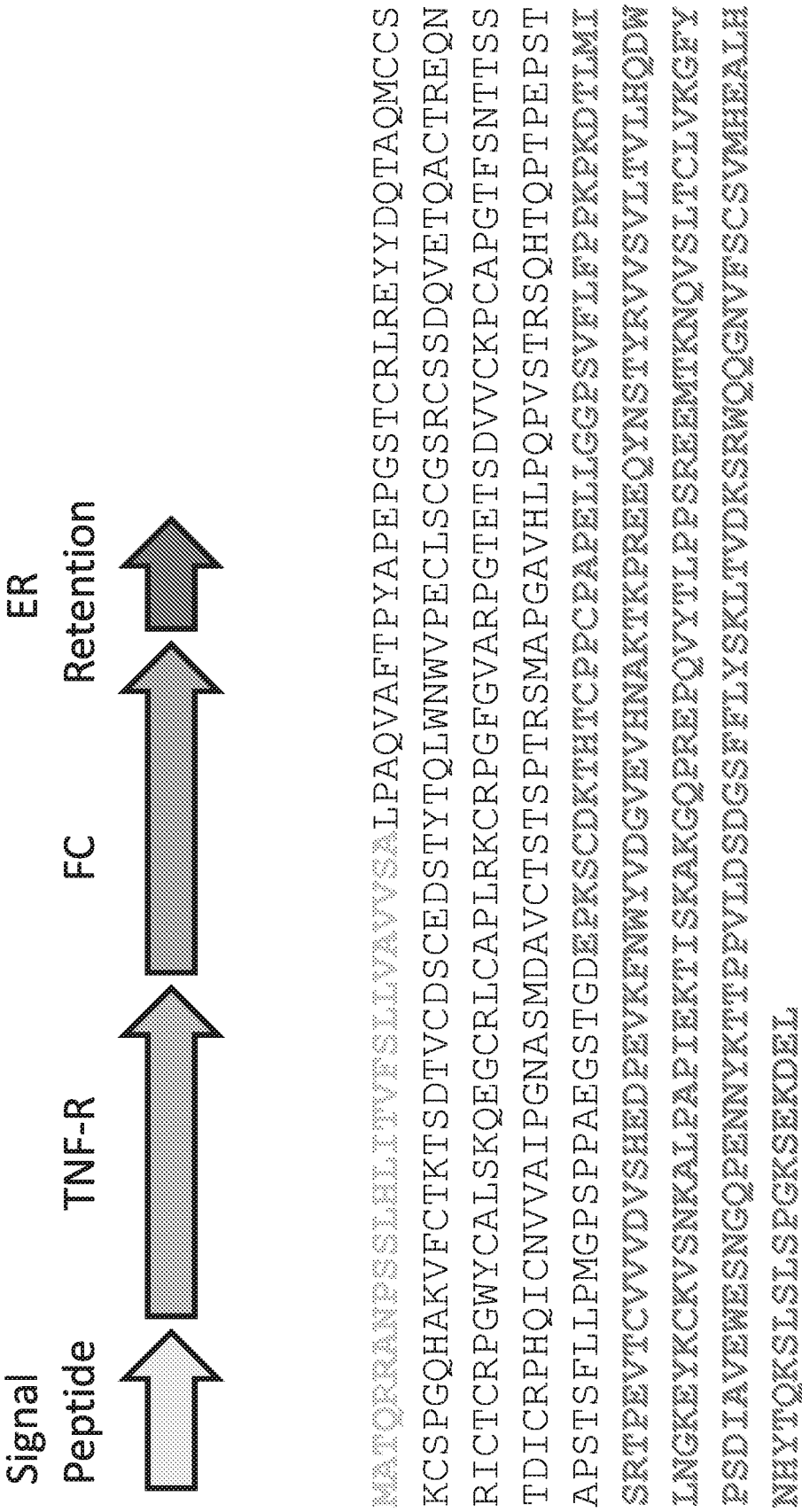


FIG. 1

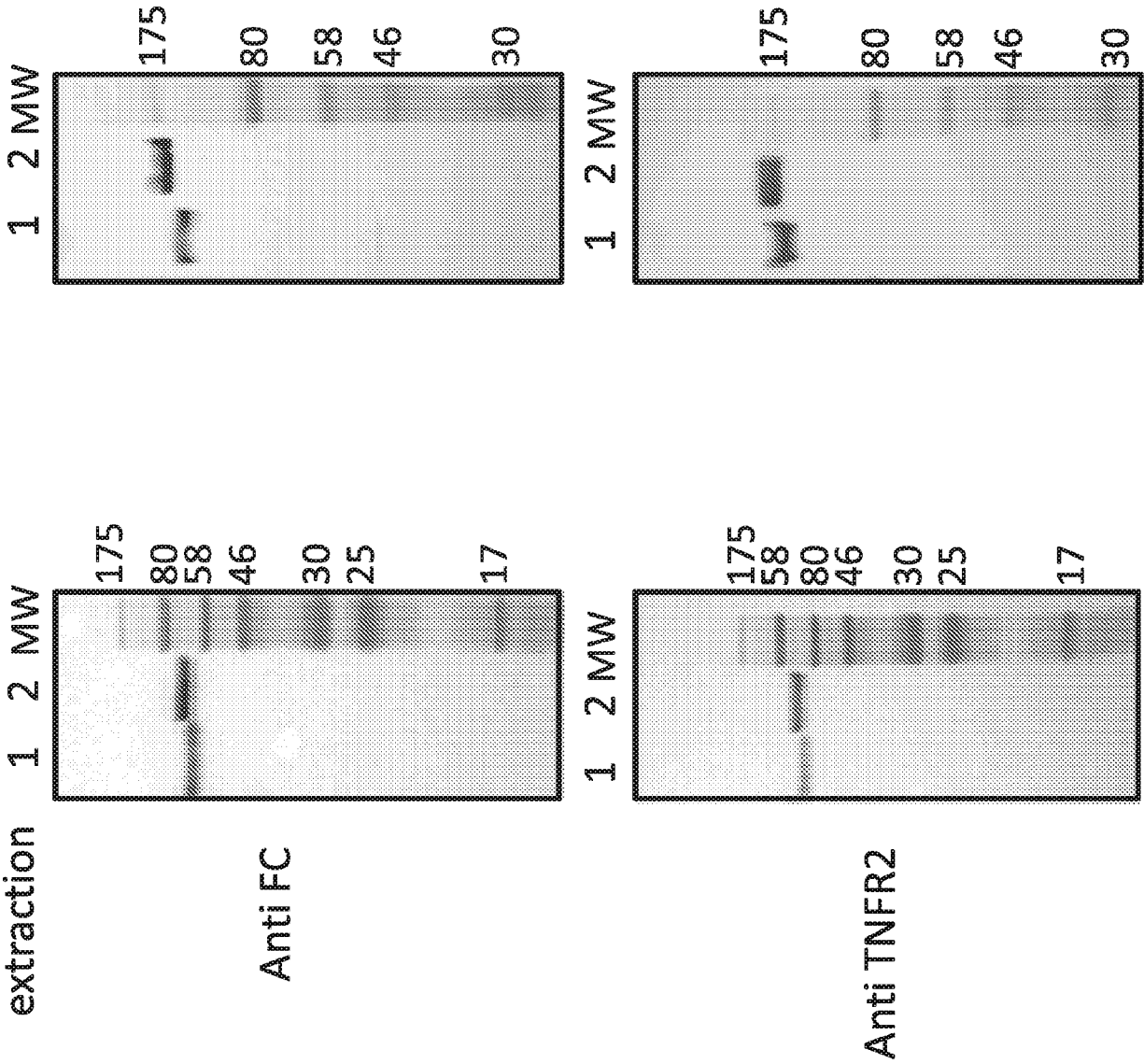


FIG. 2B

FIG. 2A

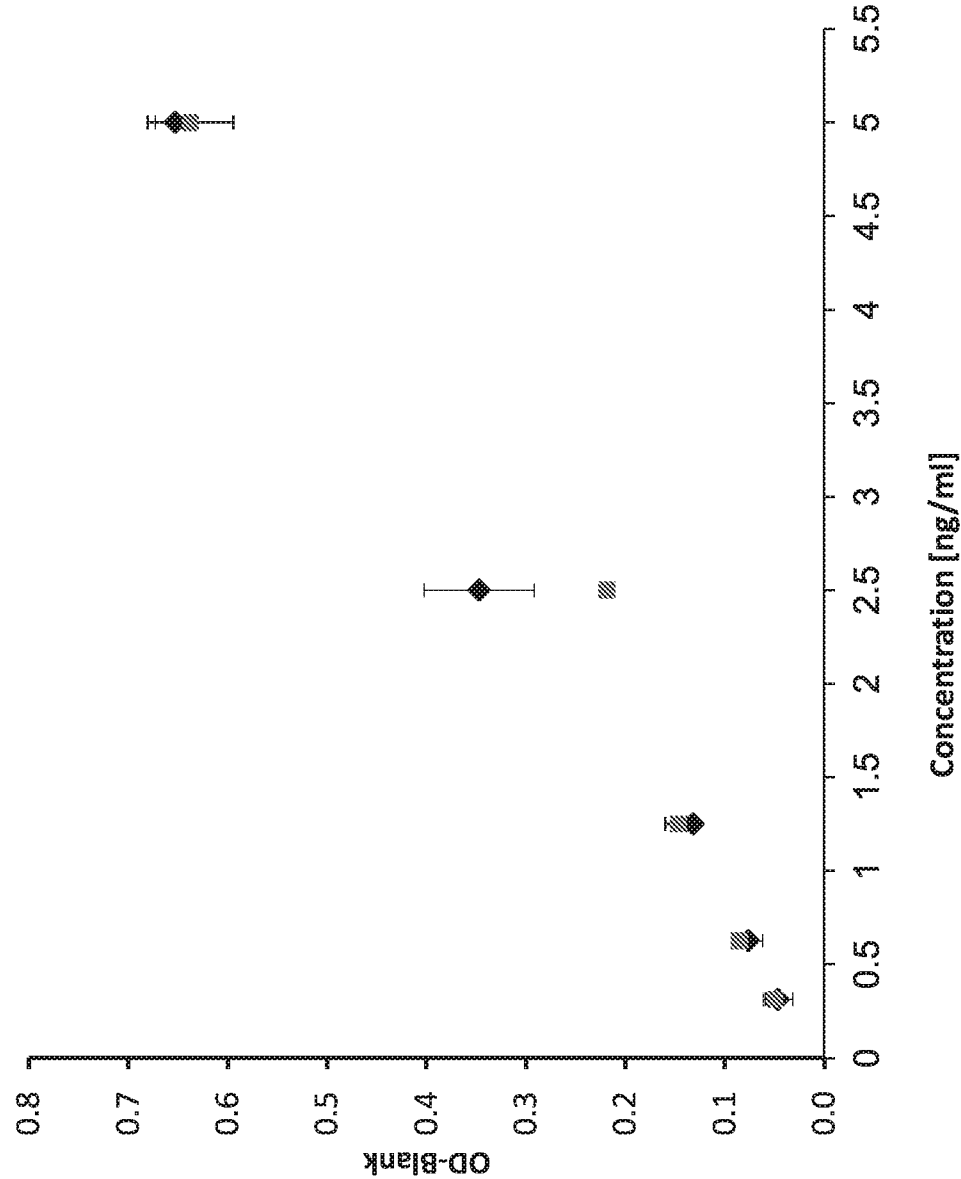


FIG. 3

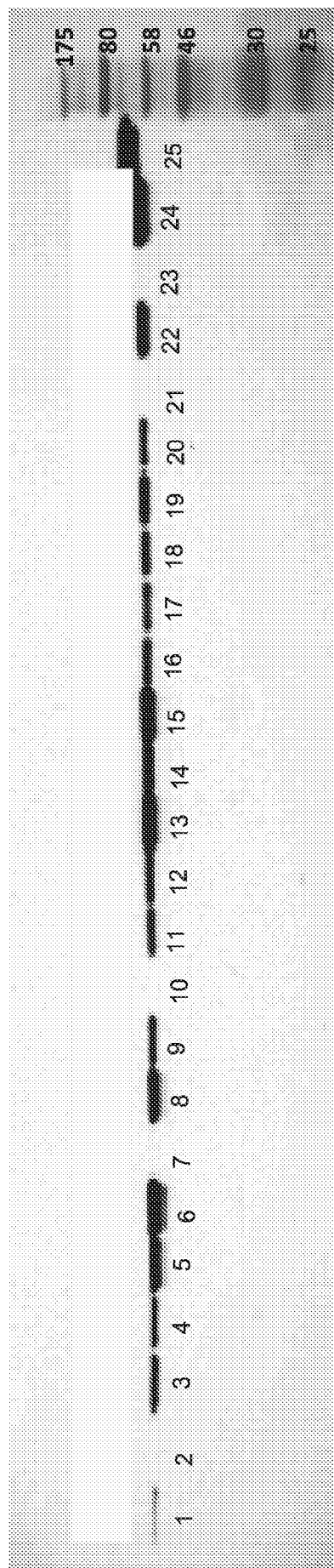


FIG. 4

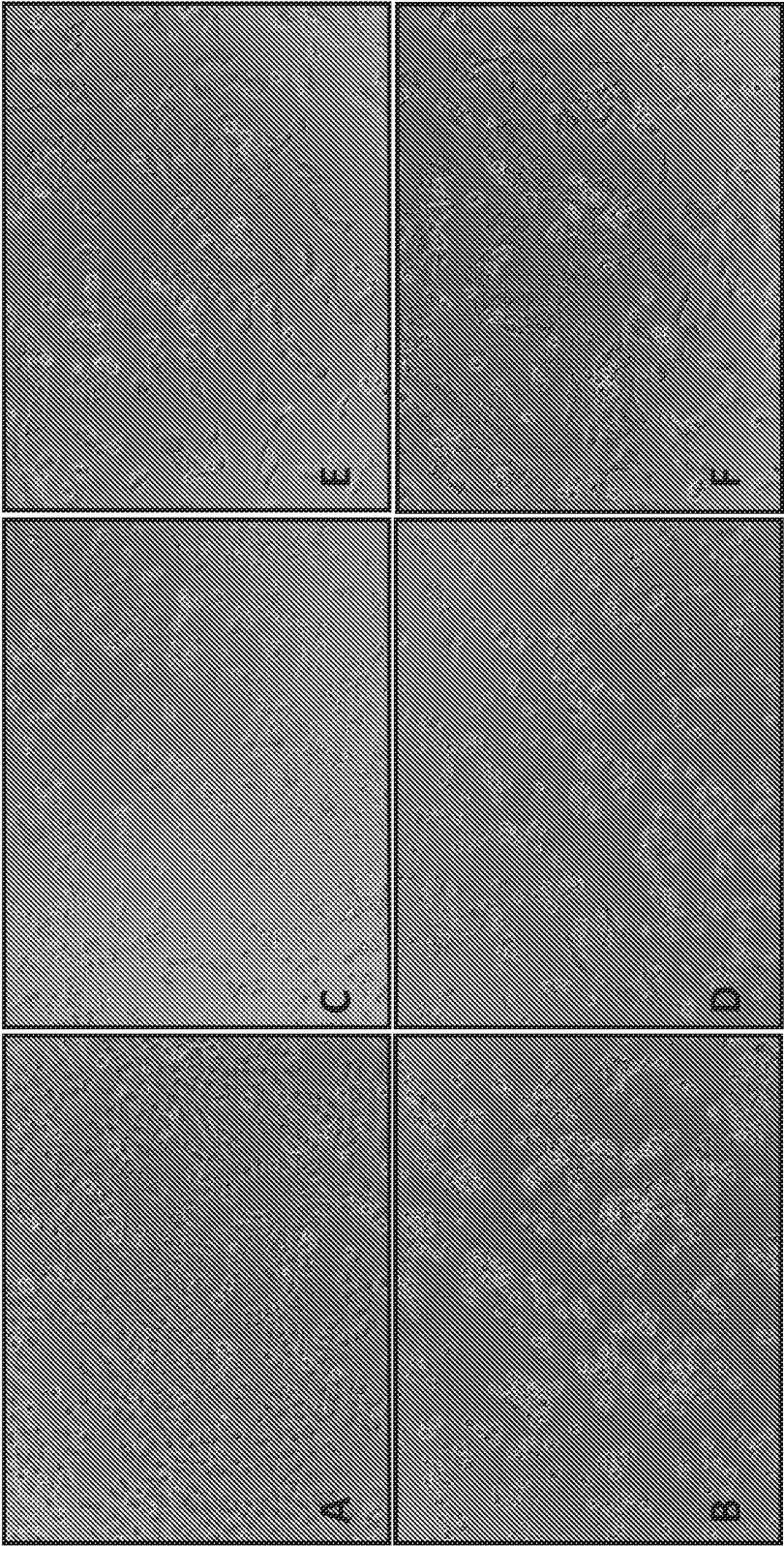


FIG. 5E

FIG. 5C

FIG. 5F

FIG. 5D

FIG. 5A

FIG. 5B

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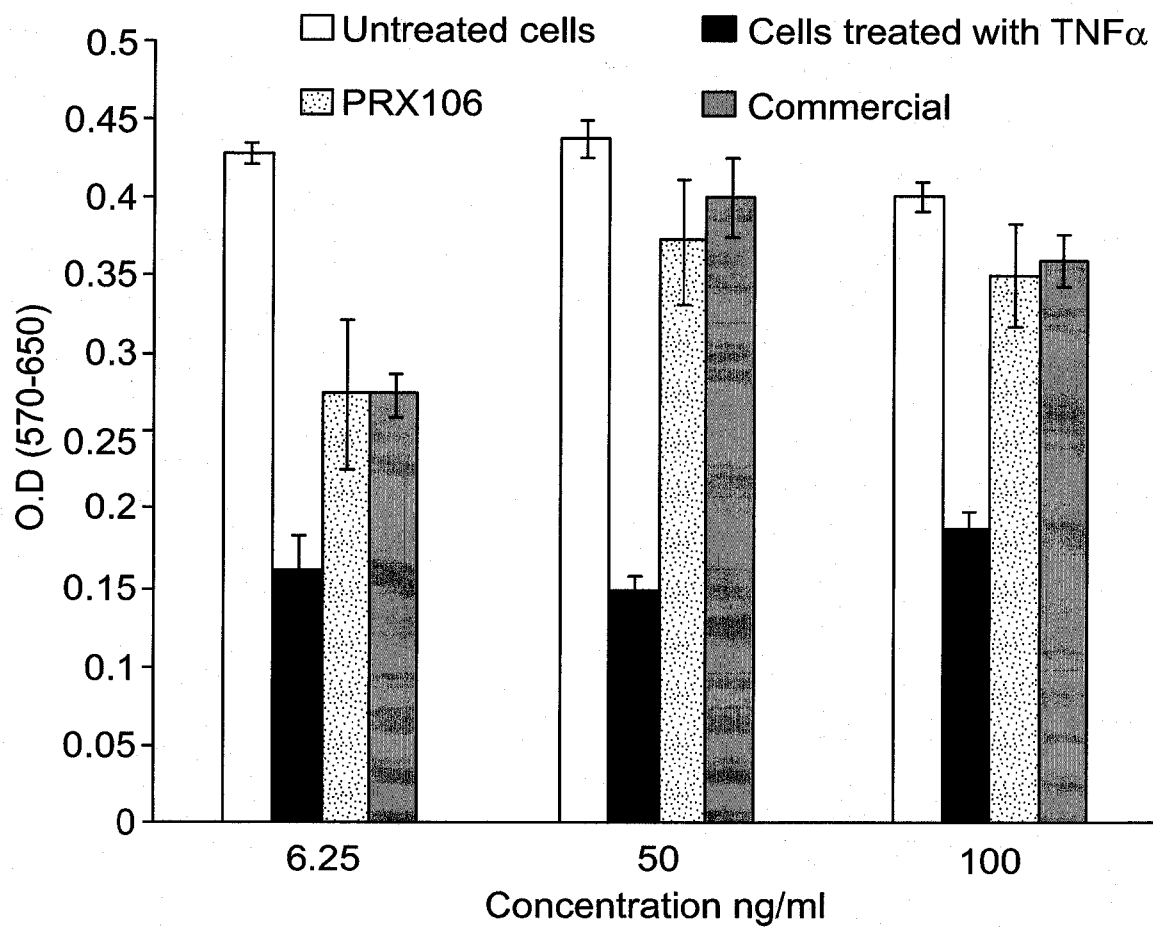


FIG. 5G

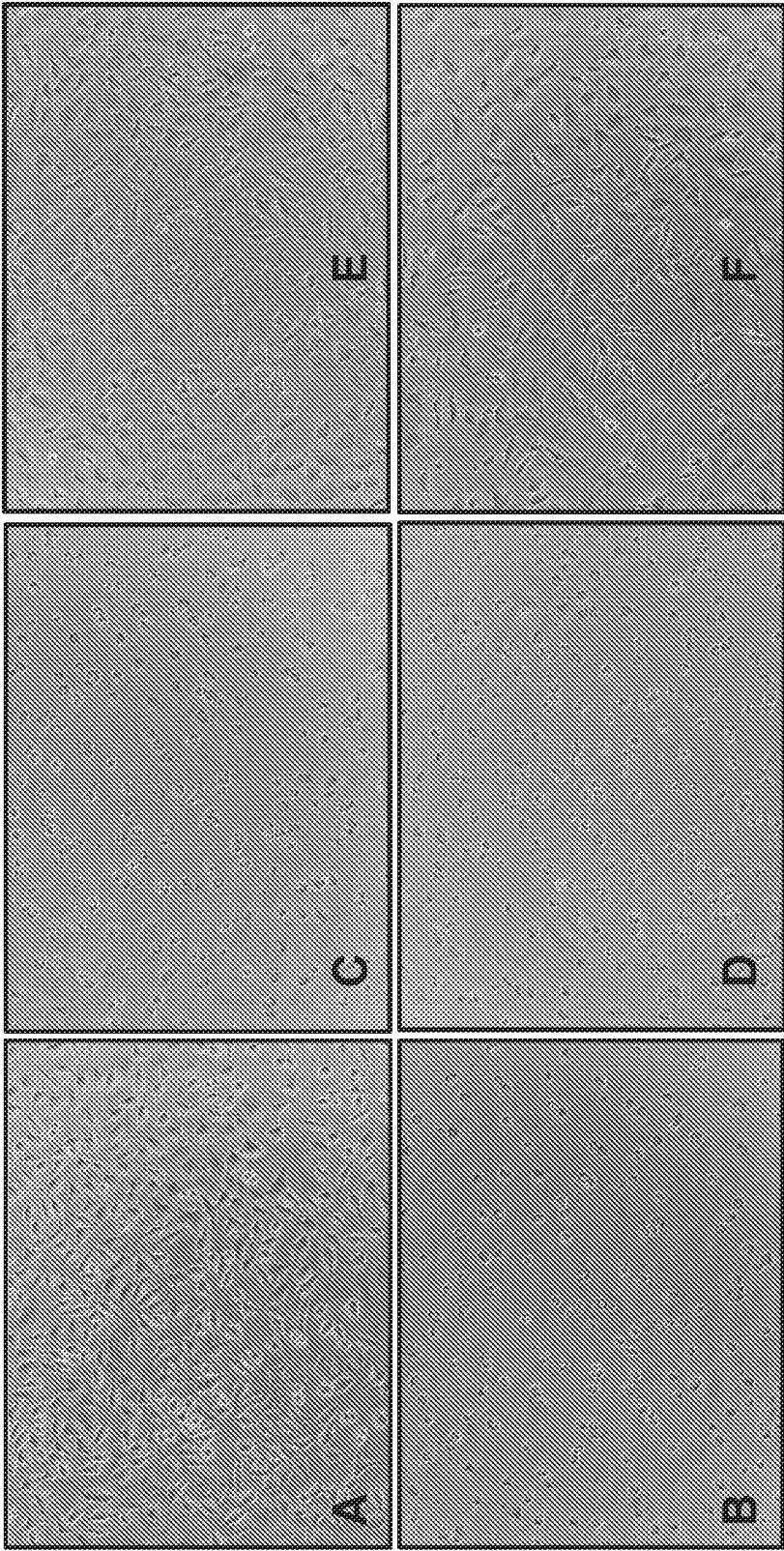


FIG. 6E

FIG. 6C

FIG. 6F

FIG. 6D

FIG. 6A

FIG. 6B

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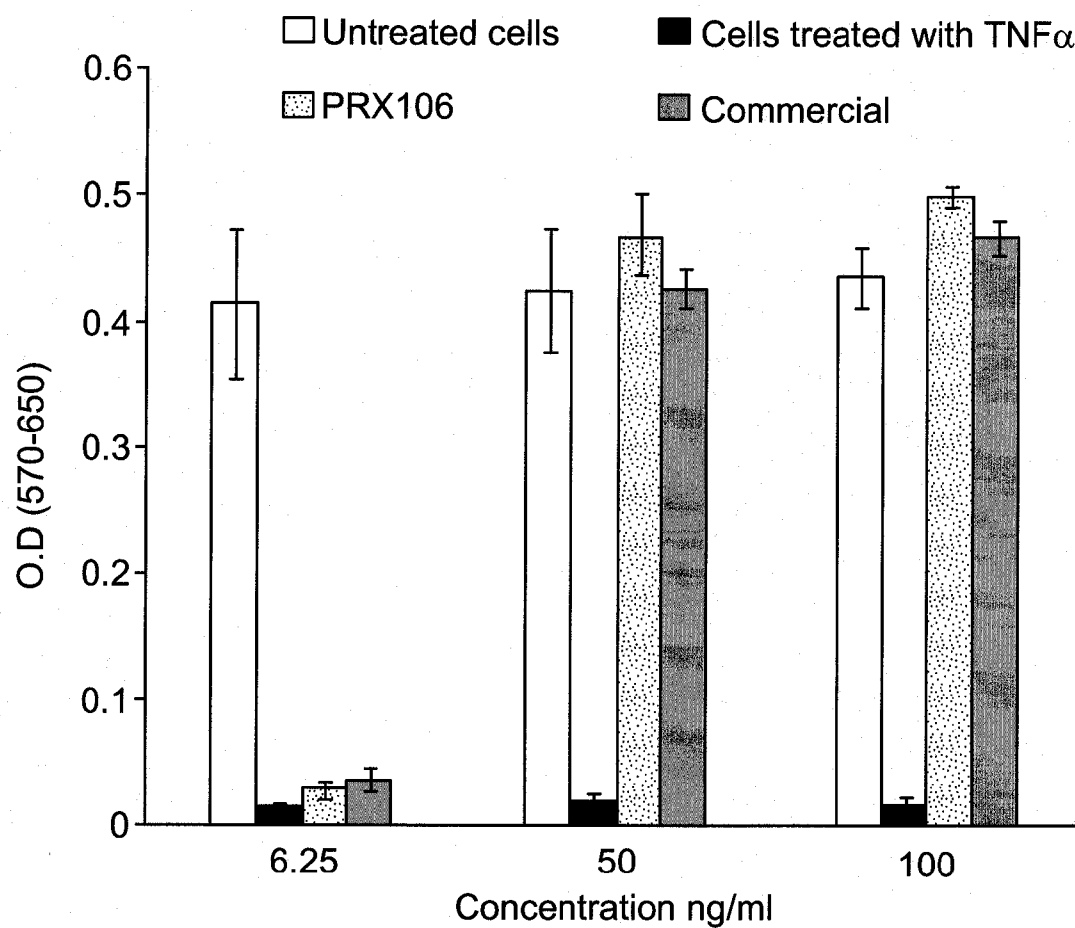


FIG. 6G

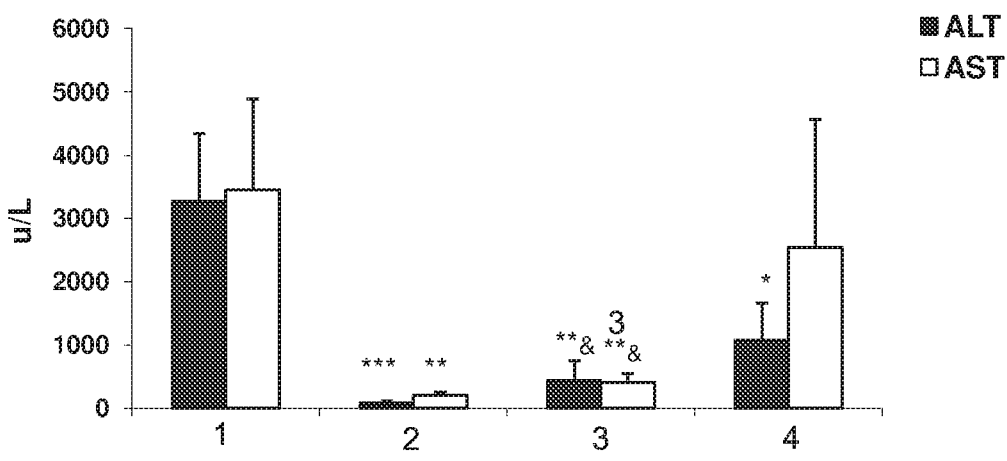


FIG. 7A

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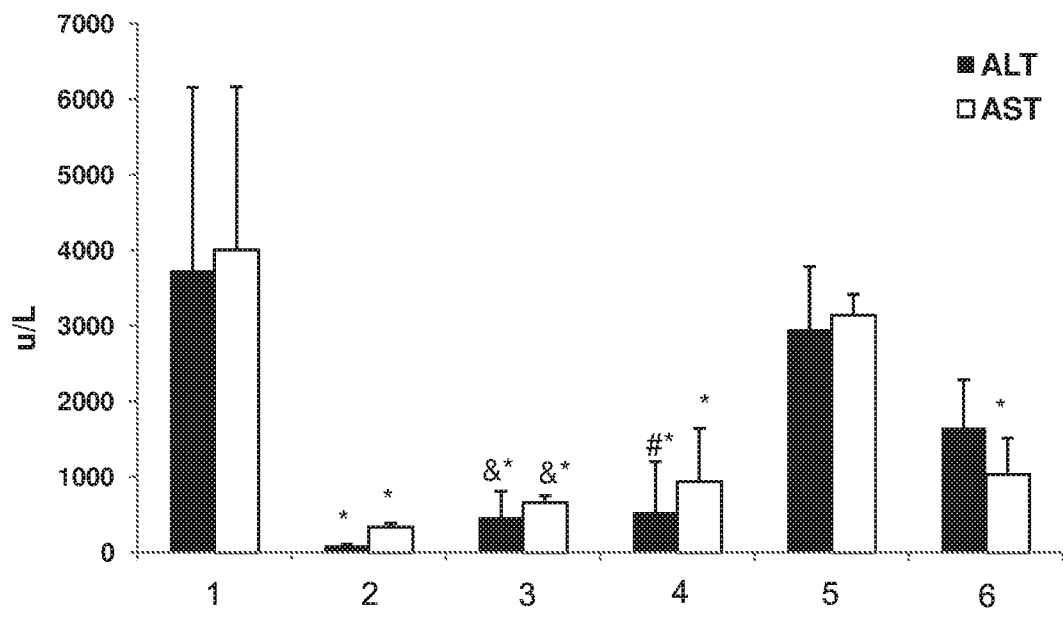


FIG. 7B

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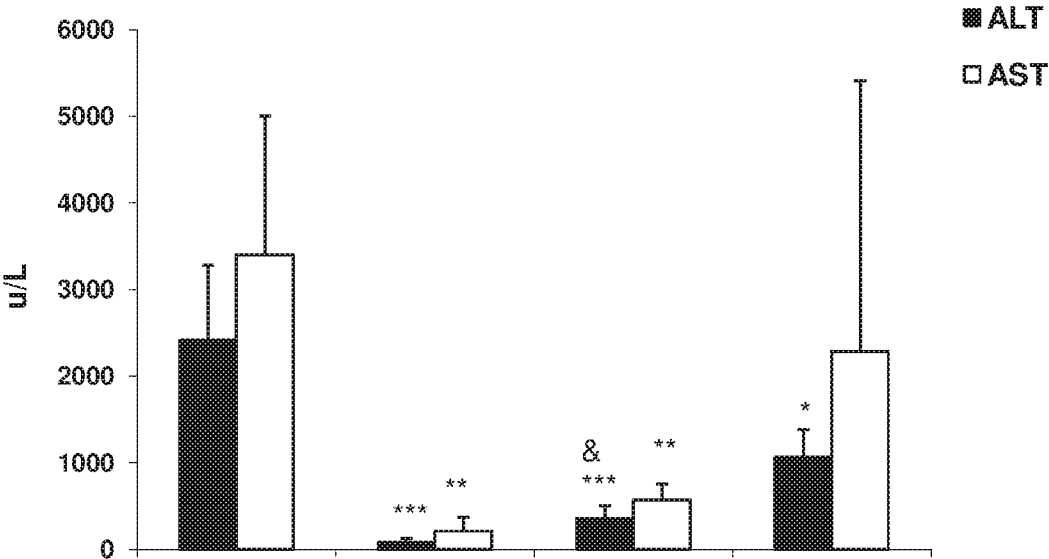


FIG. 7C

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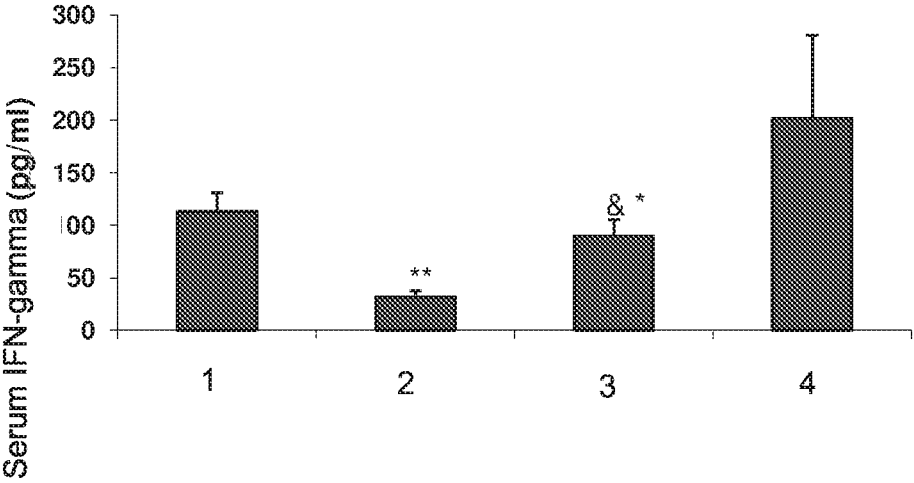


FIG. 8A

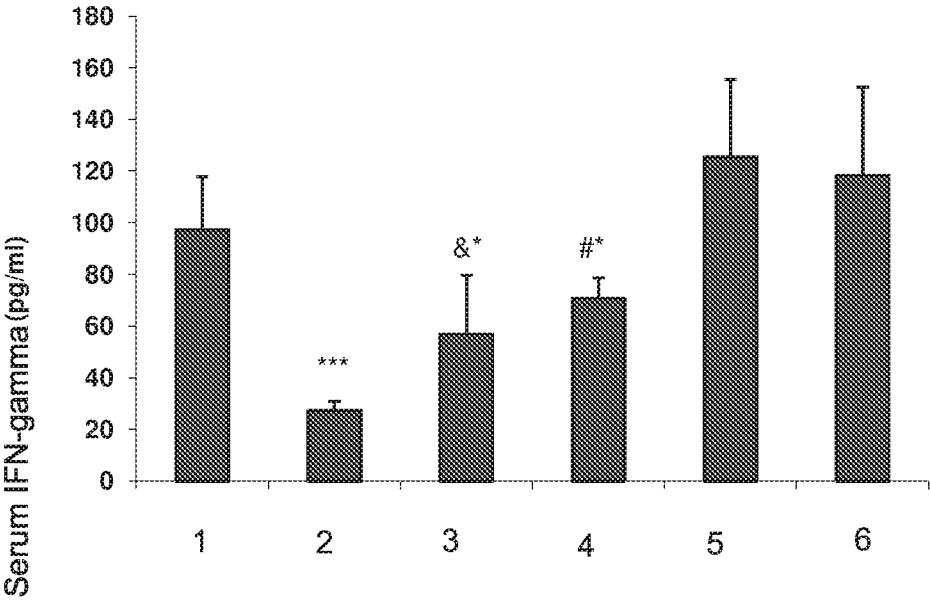


FIG. 8B

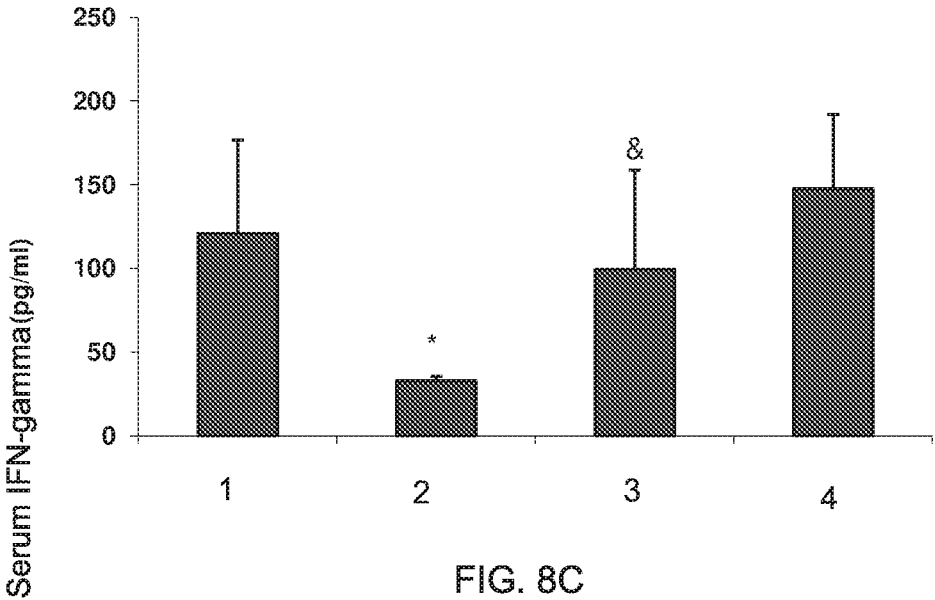


FIG. 9A

CON A + SALINE CONTROL



CON A + PLANT CELLS EXPRESSING RECOMBINANT TNFR:Fc 2.88 mg

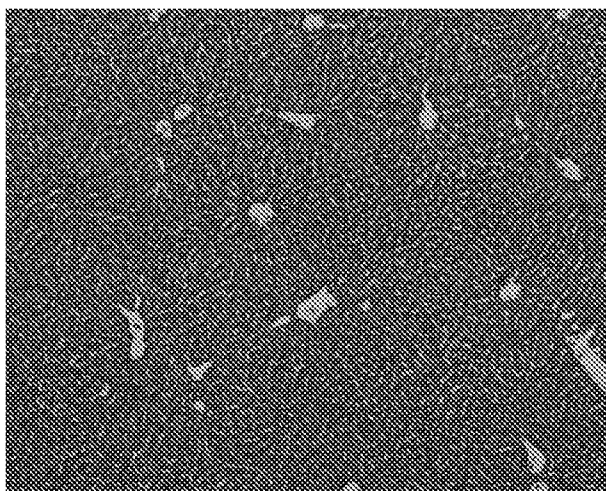


FIG. 9B

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CON A + BY2 (2.88 mg cells)

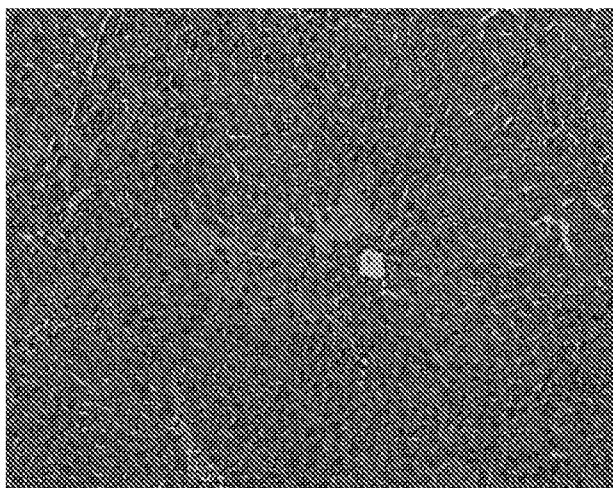


FIG. 9C

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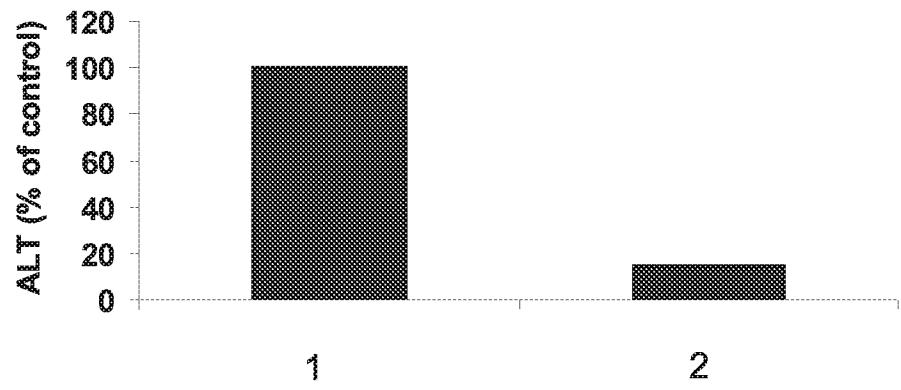


FIG. 10A

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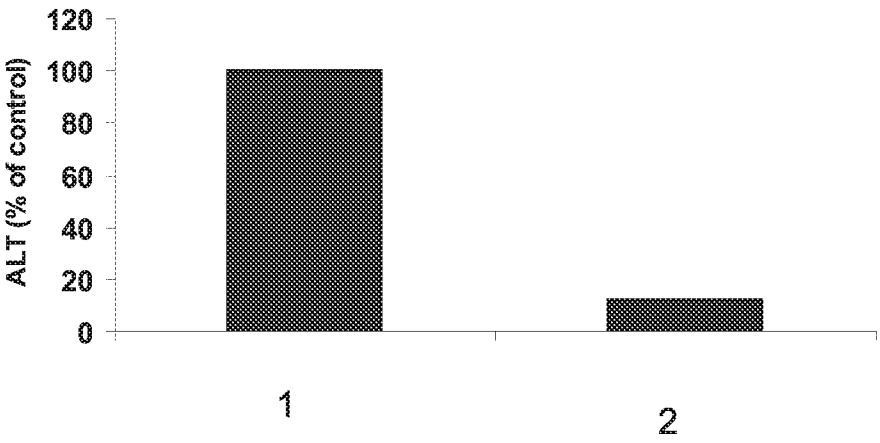


FIG. 10B

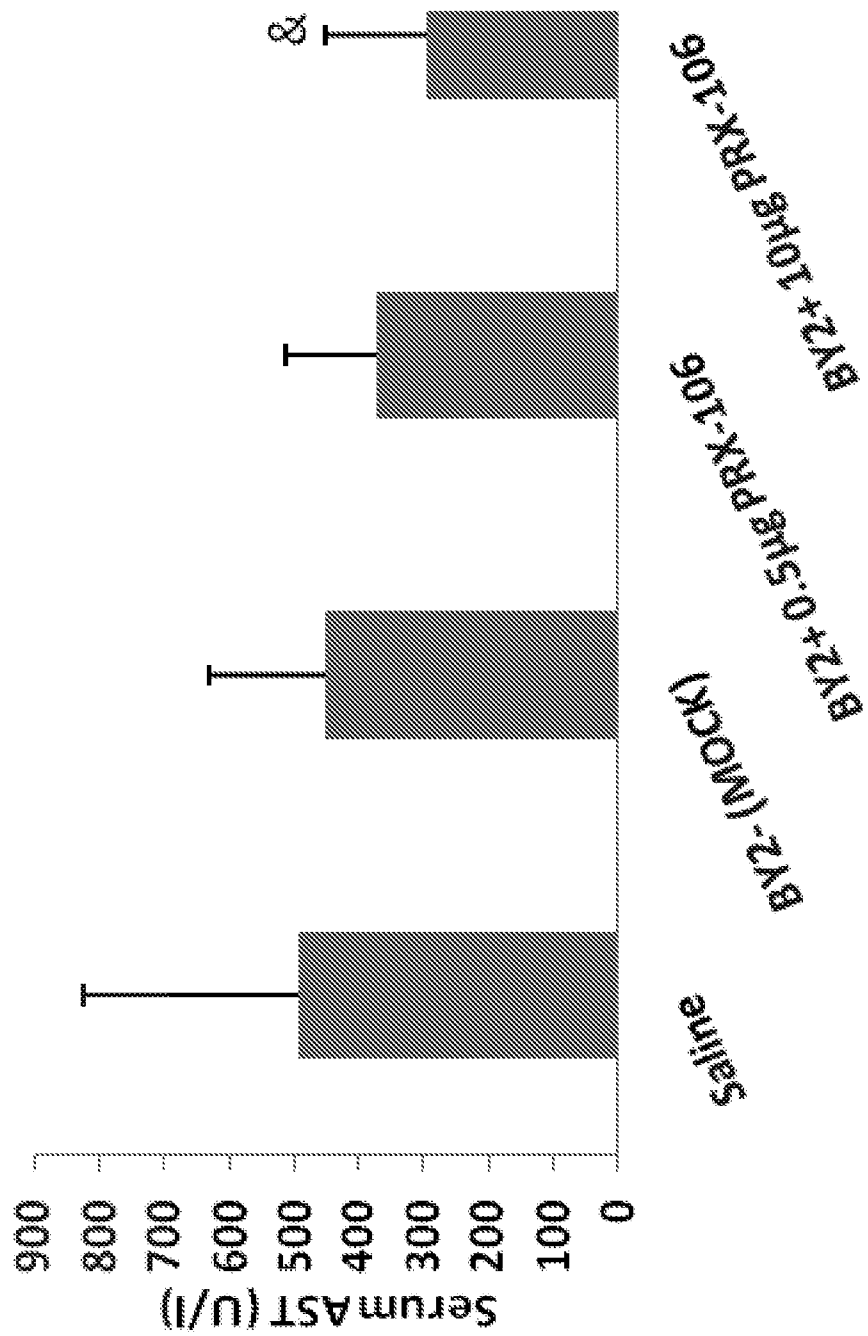
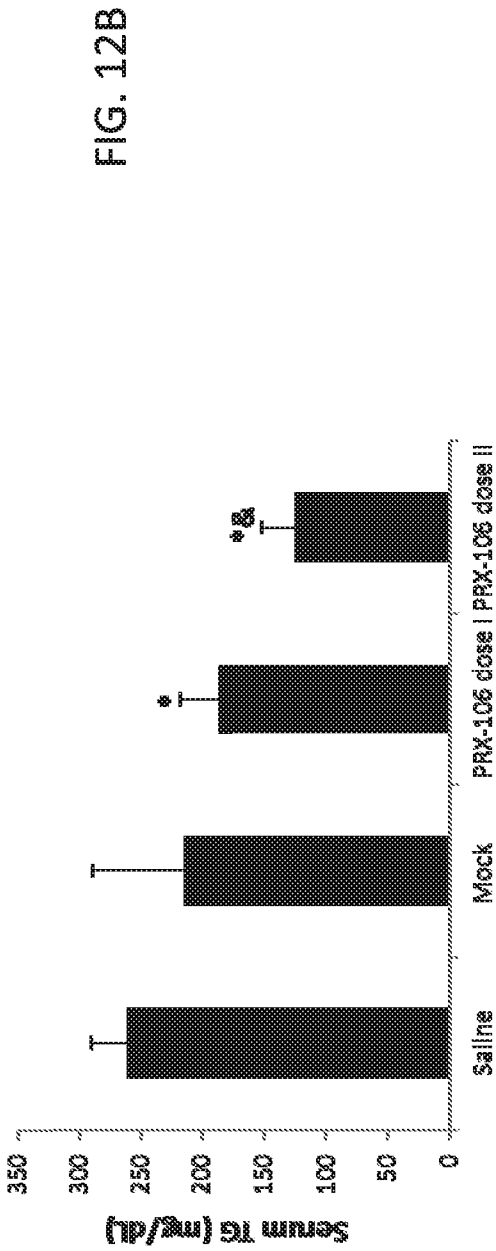
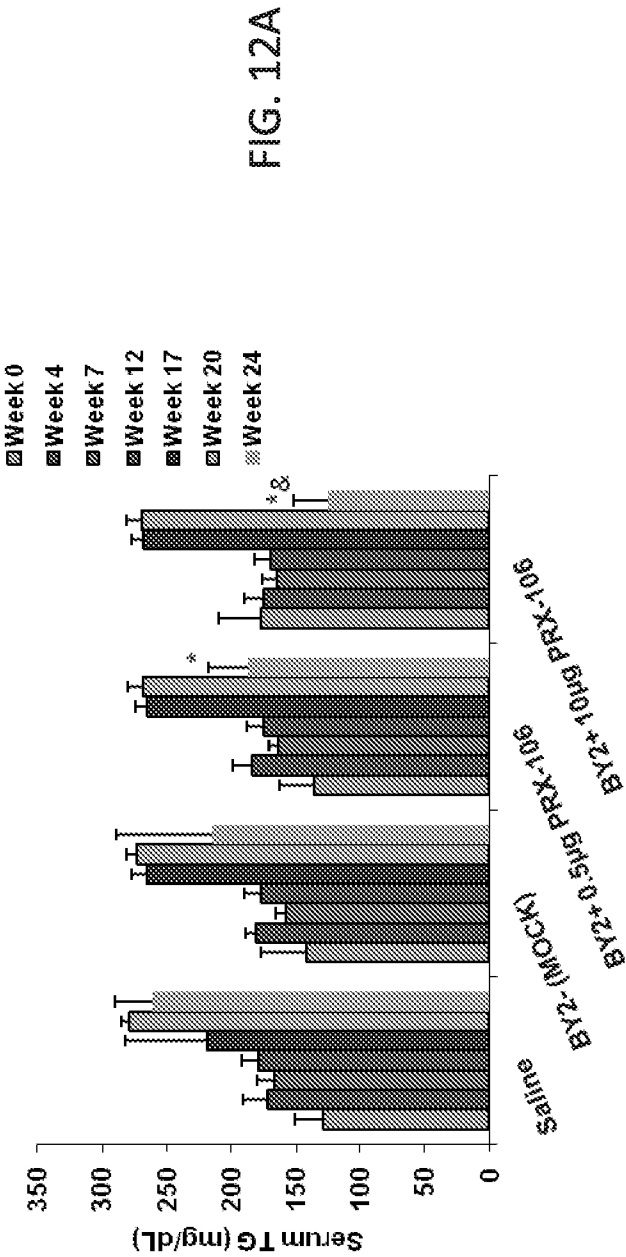


FIG. 11



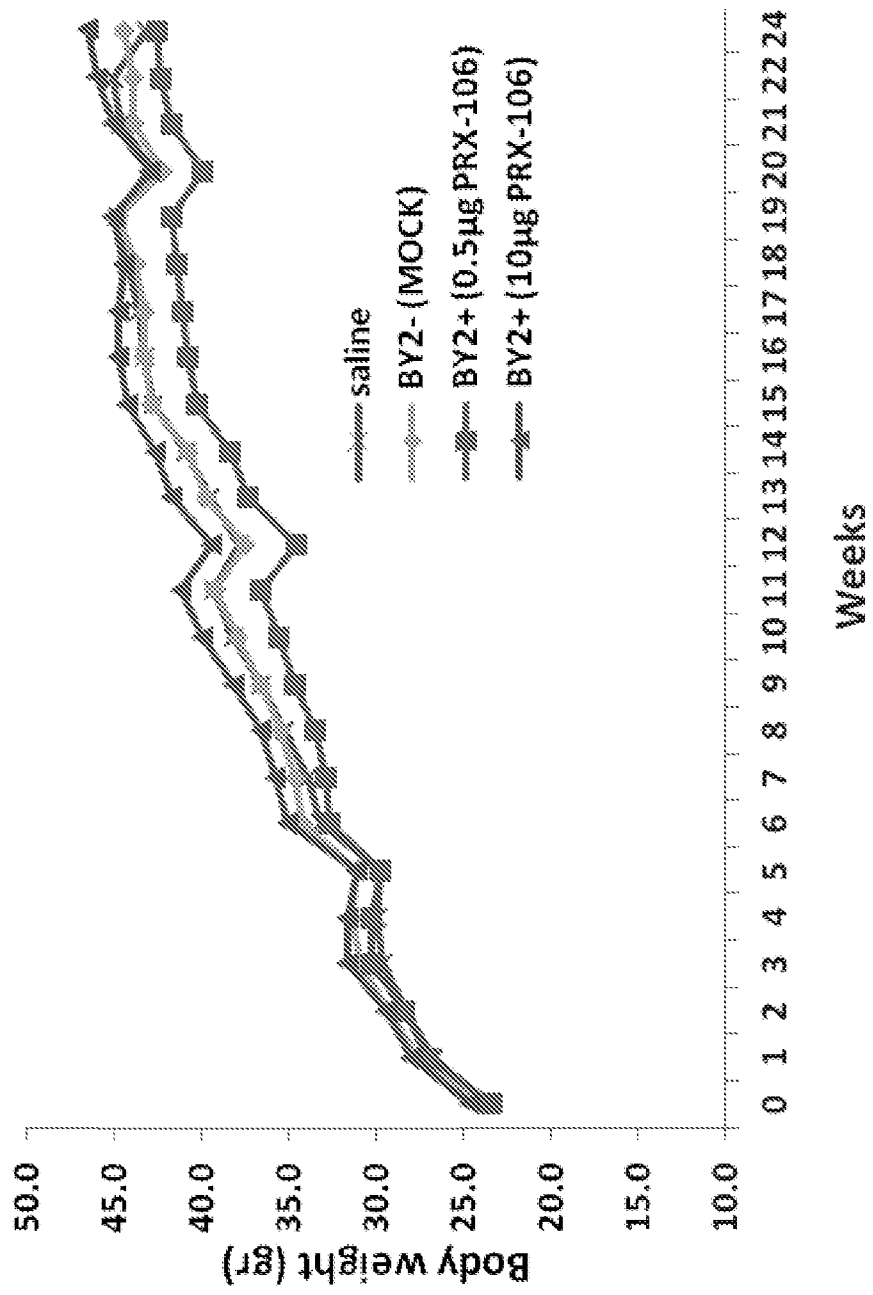


FIG. 13

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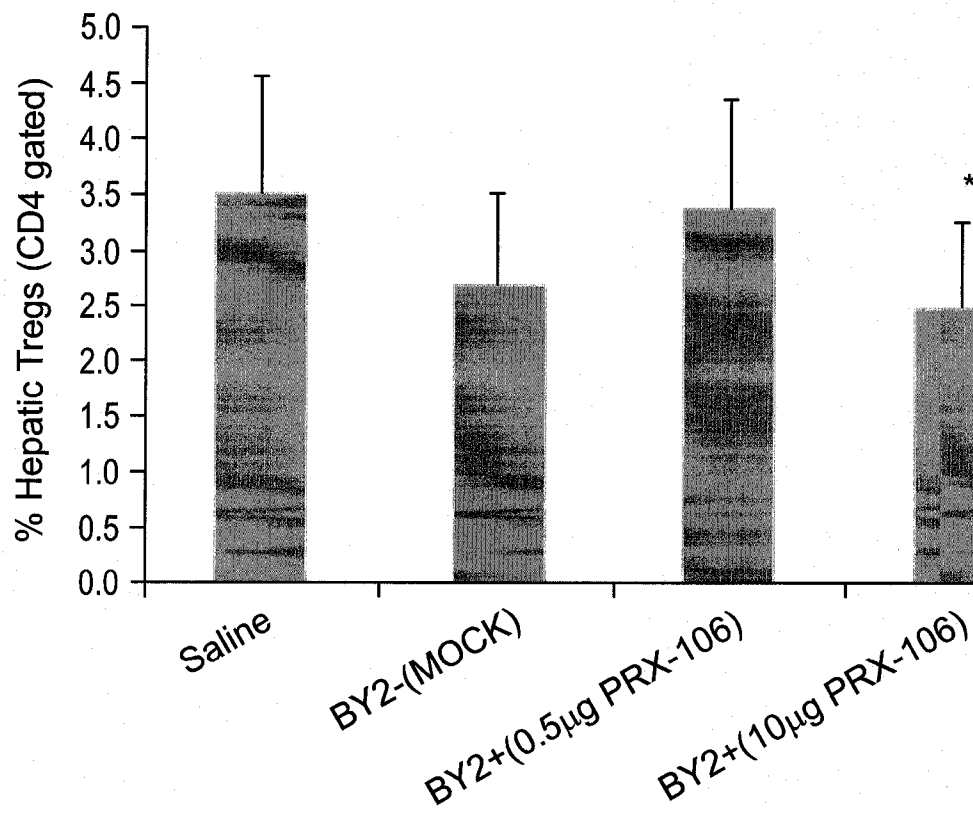


FIG. 14

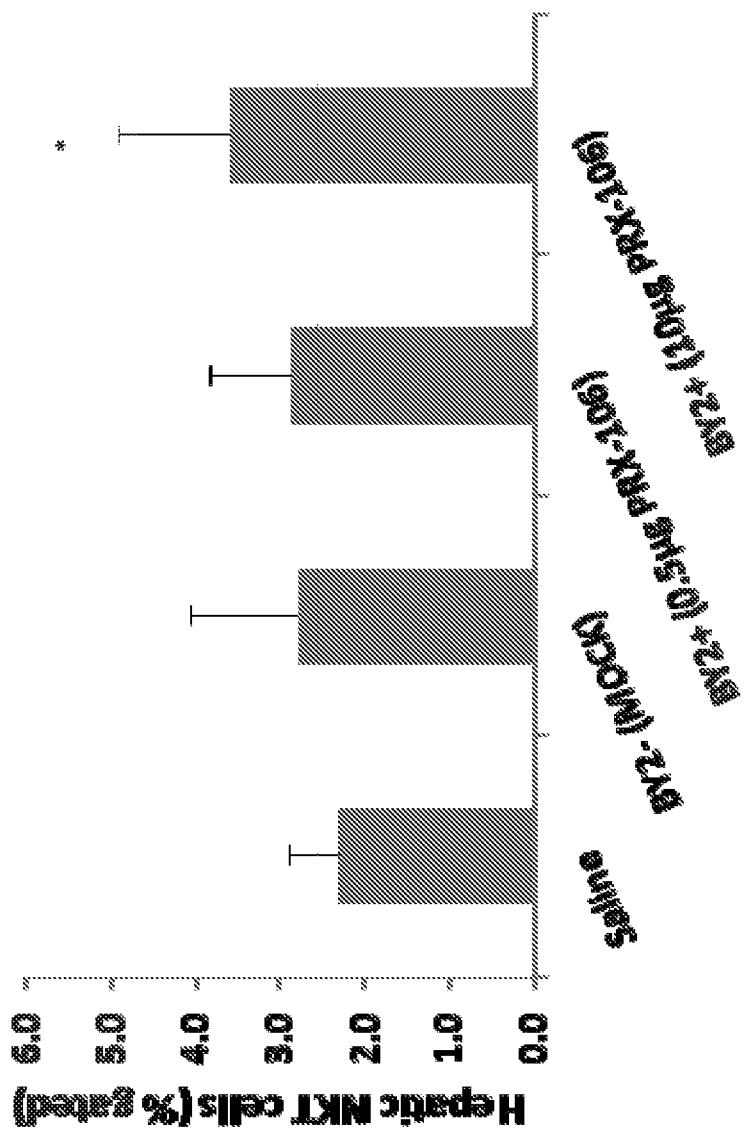


FIG. 15

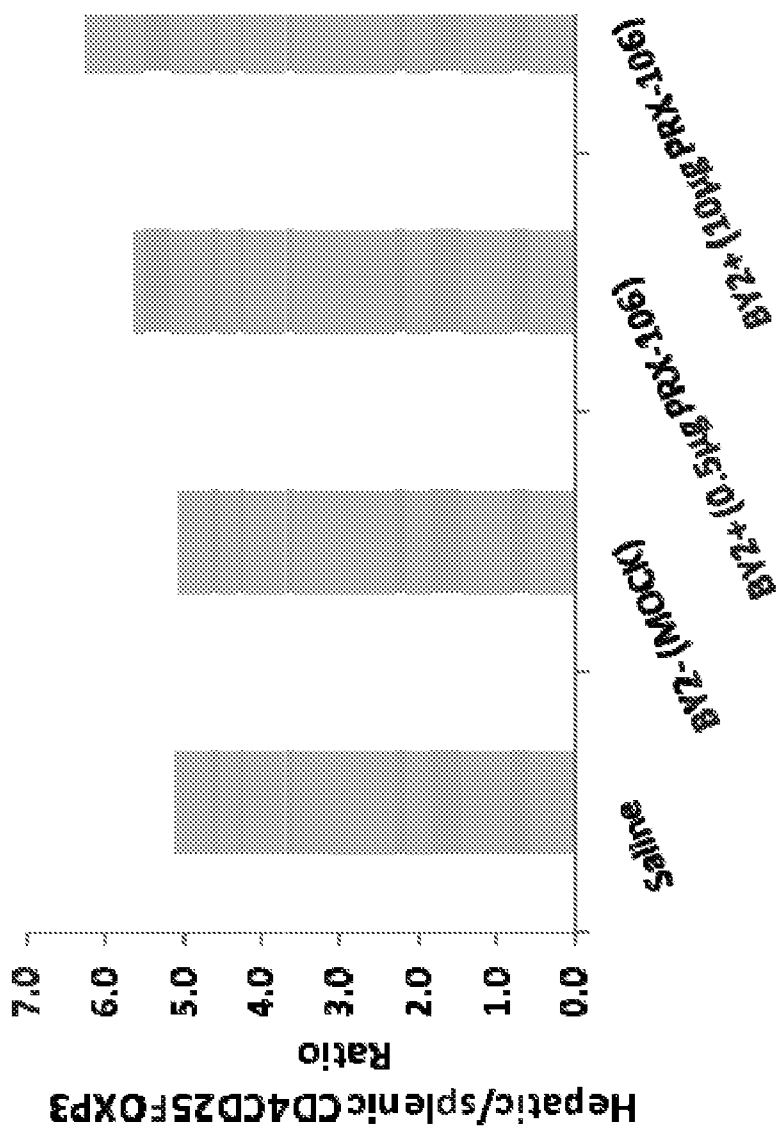


FIG. 16

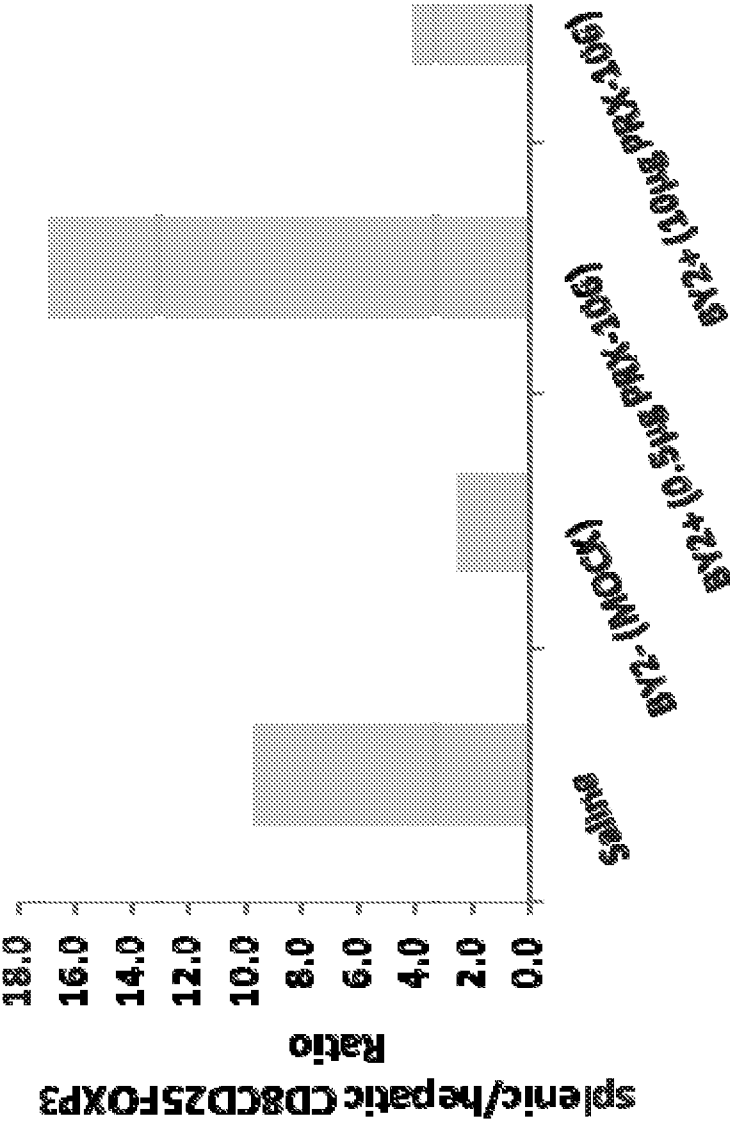


FIG. 17

INTERNATIONAL SEARCH REPORT

International application No
PCT/IL2014/050231

A. CLASSIFICATION OF SUBJECT MATTER
INV. C07K14/525 C12N15/82
ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
C07K C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

EPO-Internal, BIOSIS, Sequence Search, EMBASE, WPI Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	WO 2012/170938 A1 (ACCELERON PHARMA INC [US]; SEEHRA JASBIR [US]; KNOFF JOHN [US]; KUMAR) 13 December 2012 (2012-12-13) the whole document	1-35
Y	WINICHAYAKUL SOMRUTAI ET AL: "Head-to-tail fusions of camelid antibodies can be expressed in planta and bind in rumen fluid.", BIOTECHNOLOGY AND APPLIED BIOCHEMISTRY JUN 2009, vol. 53, no. Pt 2, June 2009 (2009-06), pages 111-122, XP002725894, ISSN: 1470-8744 the whole document	1-35



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

20 June 2014

Date of mailing of the international search report

04/07/2014

Name and mailing address of the ISA/

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040,
Fax: (+31-70) 340-3016

Authorized officer

Kania, Thomas

INTERNATIONAL SEARCH REPORT

International application No
PCT/IL2014/050231

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	WO 02/12502 A2 (CENTOCOR INC [US]) 14 February 2002 (2002-02-14) page 20, lines 9-28 page 48, lines 18-36 claims page 43, line 26 - page 49, line 12 -----	1-35
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