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(54) POLYNUCLEOTIDES FOR PRODUCTION OF FARNESYL DIBENZODIAZEPINONES

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(60) Division of application No. 11/330,123, filed on Jan. 12, 2006, now Pat. No. 7,297,524, which is a continuation-in-part of application No. 10/762,107, filed on Jan. 21, 2004, now Pat. No. 7,101,872. (60) Provisional application No. 60/441,126, filed on Jan.
21, 2003, provisional application No. 60/492,997, filed on Aug. 7, 2003, provisional application No. 60/518,286, filed on Nov. 10, 2003.

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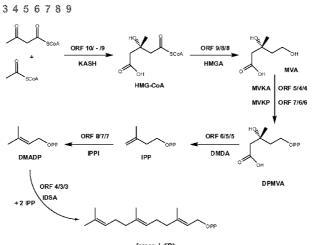
(52) **U.S. Cl.** **435/121**; 536/23.2; 435/193; 435/320.1; 435/252.3

(57) **ABSTRACT**

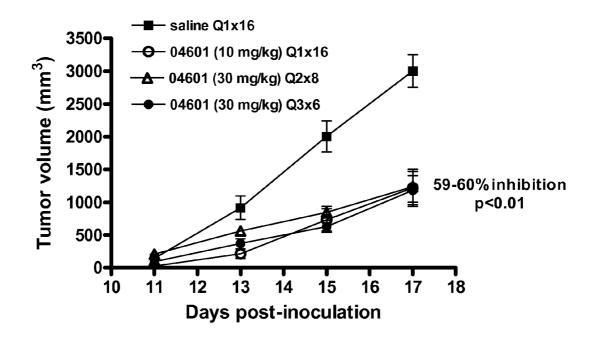
This invention provides genes and their encoded proteins, involved in the biosynthesis of farnesyl dibenzodiazepinones, including ECO-04601. The invention relates to expression vectors comprising the genes and to host cells transformed with these vectors. The invention further relates to methods of producing farnesyl dibenzodiazepinone compounds using the genes and proteins of the invention, for example, involving expression of biosynthetic pathway genes in transformed host cells.

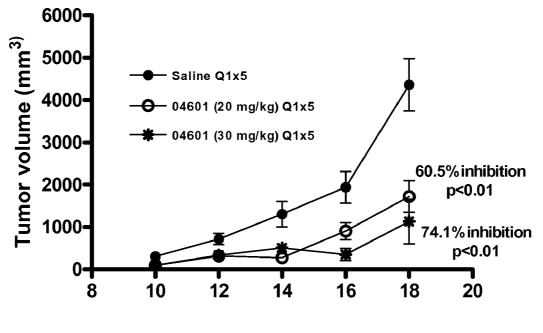
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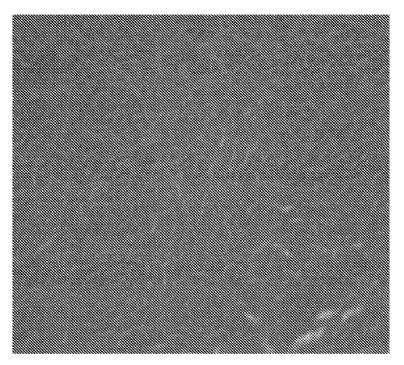


farnesyl-diPh

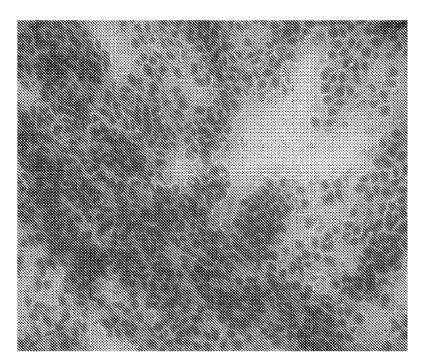




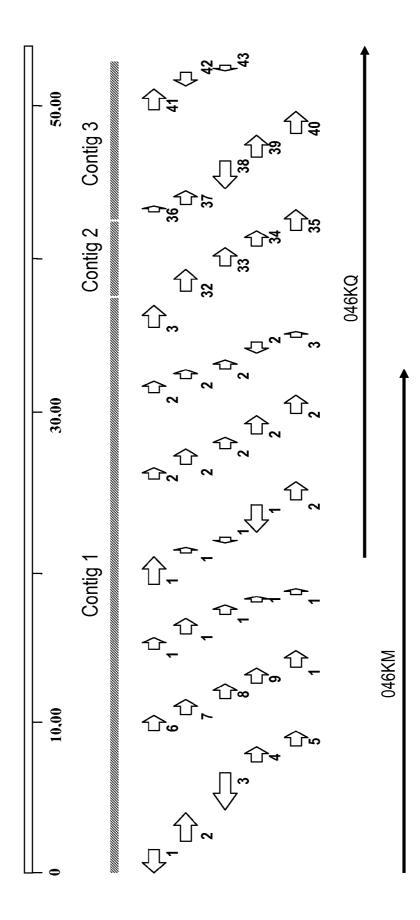
Days post-inoculation



Saline

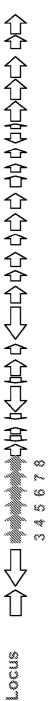


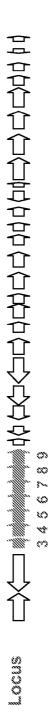
ECO-04601 (20 mg/kg)

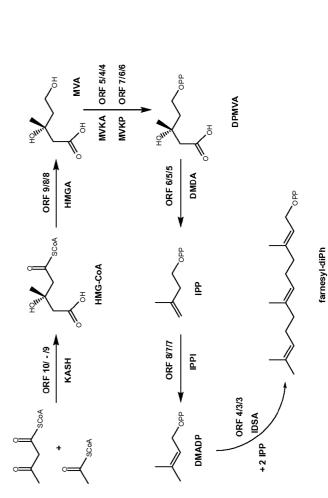


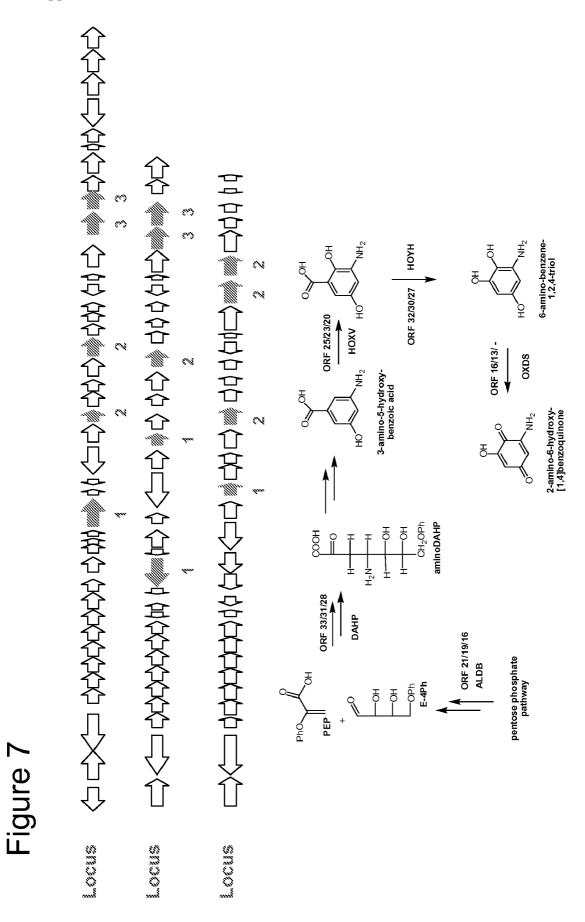




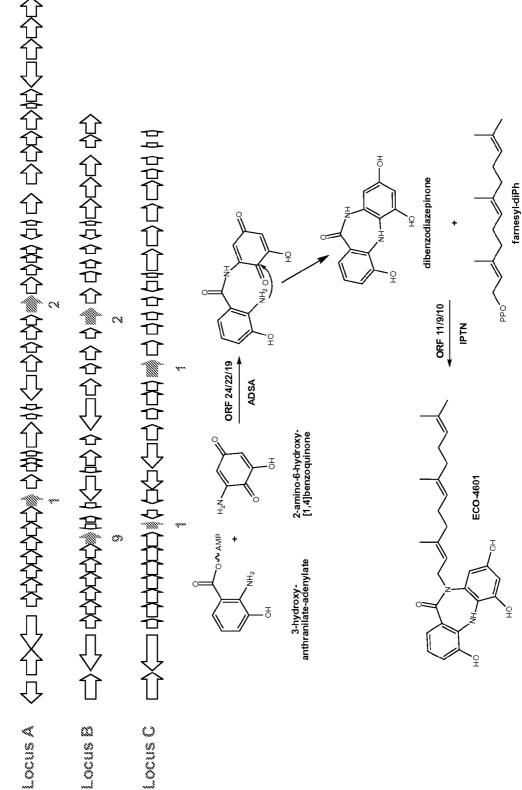












Locus A Locus B Locus C	VAELYSTIEESARQLDVPCSRDRVWPILSAYGDAFAHPEAVVAFRVATALRHAG MPGTSEAVELCSTIEESARLLNVACSRDRVWSLLSAYGDAFAHPGAVVAFRVATAMRHVG MFATAGAAELHAVVEDSARLLGVTCSPDTVAPILSTYGDTFEHDATVVAFRVATGKRHIG ** :.:*:*** *.*.* * .:**:*** * :********
Locus A Locus B Locus C	ELDCRFRTHPDDRDPYASALARGLTPRTDHPVGALLSEVHRRCPVESHGIDFGVVGGFKK ELDCRFTTHPDDRDPYARALSRGLTPETDHPVGTLLSEVQGRCPVESHGIDFGVVGGFKK ELDCRFTTHPTHRDPYALALSNGLTPKTGHPVGSLLSALQERLPIDSYGIDFGVVGGFKK ****** *** .**** **:.****.*.****:*** :: * *::*:********
Locus A Locus B Locus C	IYAAFAPDELQVATSLAGIPAMPRSLAANADFFTRHGLDDRVGVLGFDYPARTVNVYFND IYAFFTPDDLQETSKLAEIPAMPRSLAGNVEFFARHGLDDRVGVFGIDYPSRTVNVYFND IYSFFTPDALQEVAALAGIPSMPRSLAG-RDFFERYGCTTGR-VIGIDYPH **: *:** ** .: ** **:***** ::** *:* *:*
Locus A Locus B Locus C	VPRECFEPETIRSTLRRTGMAEPSEQMLRLGTGAFGLYVTLGWDSPEIERICYAAATTDL VPAESFHSETIRSTLREIGMAEPSERMLKLGEKAFGLYVTLGWDSSRIERICYAAATTDL
Locus A Locus B Locus C	TTLPVPVEPEIEKFVKSVPYGGGDRKFVYGVALTPKGEYYKLESHYKWKPGAVNFI TTLPVPVEPEIEKFVRSVPYGGEDRKFVYGVALTPHGEYYKLESHYRWKPGAMDFI

Locus A Locus B Locus C	VSEPSSSLPRLGQWHGLEDLRRLQEKQLAETFTWAARSPFYRARLASGAPPVTPADLADL VNDPRPSLPQLGQWHGPEDLQRLQEKQLSQTVTWATRSPFYRDRLDPGALPATAADLADL VNPTRSSLPRLGQWNGPEDLRLLQEKQLQQTVGWASRSPFYRGRLDTAALPTTIDDLASL ****:****:* ***: ****** :*. **:****** *** *.*
Locus A Locus B Locus C	PLTTKQDLRDNYPFGMLAVPRERLATYHESSGTAGKPTPSYYTAEDWTDLAERFARKWIG PLTTKQDLRDNYPFGMLAVPKERLATYHESSGTAGRPTPSYYTAEDWTDLAERFARKWIG PLTTKQDLRDNYPFGMLAVPKERLATYHESSGTAGRPTPSYYTADDWIDLAERFARKWIG ************************************
Locus A Locus B Locus C	MSADDVFLVRTPYALLLTGHLAHAAARLRGATVVPGDNRSLAMPYARVVRVMHDLDVTLT MSAEDVFLVRTPYALLLTGHLAHAAGRLRGATVVPGDNRSLAMPYARVVRVMHDLGVTLT ITAEDVFLVRTPYALLLTGHLAHAAGRLHGATVVPGDNRSLAMPYARVVRVMHDLGVTLT ::*:********************************
Locus A Locus B Locus C	WSVPTECLIWAAAAIAAGHRPDIDFPALRALFVGGEPMTDARRRRISRLWGVPVIEEYGS WSVPTECLIWAAAATAAGHRPDVDFPALRALFVGGEPLTDARRRRISRLWGVPVIEEYGS WSVPTECLIWAAAATAAGHRPSEDFPALRALFVGGEPLTTARRDRISRLWGVPVIEEYGS ************************************
Locus A Locus B Locus C	TETGSLAGECPEGRLHLWADRALFEVYDPDTGAVRADGDGQLVVTPLFREAMPLLRYNLE TETGSLAGECPNGRMHLWADRALFEVYDPRTGTVSADGDGQLVVTPLFREAMPLLRYNLE TETGSLAGECPHGRMHLWADRALFEVYDPQTGTVRAEGEGQLVVTPLYREAMPLLRYNLE *****************
Locus A Locus B Locus C	DNVSVSYDDCGCGWKLPTVRVLGRSAFGYRVGGTTITQHQLEELVFSLPEAHRVMFWRAK DDVTVSYDDCACGWNLPTVRVLGRAAFGYRVGAATITQHRLEEVVFSLPESHGVVFWRAK DNVSVAYDDCACGWKLPTVQVLGRAAFGHRVGATTVTQHRLEELVFSLPDAYQVVFWRAR *:*:*:****.***:****:****:****:***
Locus A Locus B Locus C	AEPALLRVEIEVAAAHRVAAEAELTAAIRAAFGVDSEVTGLAPGTLIPLDALTSMPDVVK AEPTVLRIEIEVAEEHRTAAQAELTASVRATFGIDSEVTGLTPGTLVPREALTSMPDVVK AEPAALRIEIEVPEEHRAAAEAELVHSVRTAFGVDSTVTGLPPGTLIPHGALTAMPDVVK ***: **:****. **.**:***. ::*::**:** ****.********
Locus A Locus B Locus C	PRSLFGPDEDWSKALLYY PRSLFGPDEDWGKALLYY PRSLFGPDEDWGKALLYY **********

POLYNUCLEOTIDES FOR PRODUCTION OF FARNESYL DIBENZODIAZEPINONES

RELATED APPLICATIONS

[0001] This application is a divisional of U.S. patent application Ser. No. 11/330,123, filed Jan. 12, 2006, which is a continuation-in-part of U.S. patent application Ser. No. 10/762,107, filed Jan. 21, 2004, now issued as U.S. Pat. No. 7,101,872, which claims priority to each of U.S. Provisional Application No. 60/441,126, filed Jan. 21, 2003, U.S. Provisional Application No. 60/492,997, filed Aug. 7, 2003, and U.S. Provisional Application No. 60/518,286, filed Nov. 10, 2003. The entire disclosures of each of these applications are herein incorporated by reference.

SEQUENCE LISTING ON COMPACT DISK

[0002] The content of the following submissions on compact discs are incorporated herein by reference in its entirety: A compact disc copy of the Sequence Listing (COPY 1) (file name: 3005-5US-50US.ST25.txt, date recorded Jan. 10, 2006, size: 298 KB) and a duplicate compact disc copy of the Sequence Listing (COPY 2) (file name: 3005-5US-50US. ST25.txt, date recorded Jan. 10, 2006, size: 298 KB).

FIELD OF THE INVENTION

[0003] The invention relates to novel polynucleotide sequences and their encoded proteins, which are involved in the biosynthesis of a farnesyl dibenzodiazepinone compound and analogs. The invention relates to the use of such polynucleotides and proteins to produce farnesyl dibenzodiazepinone compounds and analogs. One method of obtaining the compound is by cultivation of a novel modified strain of *Micromonospora* sp., i.e., 046-ECO11 or [S01]046; another method involves expression of biosynthetic pathway genes in transformed host cells. The present invention further relates to cosmids 046KM and 046KQ and their methods of use.

BACKGROUND OF THE INVENTION

[0004] The euactinomycetes are a subset of a large and complex group of Gram-positive bacteria known as actinomycetes. Over the past few decades these organisms, which are abundant in soil, have generated significant commercial and scientific interest as a result of the large number of therapeutically useful compounds produced as secondary metabolites. The intensive search for strains able to produce new secondary metabolites having potential therapeutic applications has led to the identification of hundreds of new species. Many of the euactinomycetes, particularly *Streptomyces* and the closely related *Saccharopolyspora* genera, have been extensively studied. Both of these genera produce a notable diversity of biologically active metabolites. Because of the commercial significance of these compounds, much is known about the genetics and physiology of these organisms.

[0005] Microbial genomic information is unique in that, unlike the organization of genomic information in higher life forms, microbial secondary metabolic biosynthetic genes are known to cluster together within the genome. This information allows identification of the gene locus encoding the enzymes responsible for the biosynthesis of a specific molecule. Equally, the identification of the genes present within a cluster allows prediction of the structure of the secondary metabolite. The identification of the genes and proteins responsible for the production of active molecules allows for example, generation of structural analogs or improvement of the production process.

[0006] U.S. patent application Ser. No. 10/762,107 describes a dibenzodiazepinone secondary metabolite, specifically 10-farnesyl-4,6,8-trihydroxy-dibenzodiazepin-11-one (named ECO-04601) produced by a known euactino-mycetes strain, *Micromonospora* sp. (IDAC 231203-01). Likewise, U.S. Pat. No. 5,541,181 (Ohkuma et al.) also discloses a dibenzodiazepinone secondary metabolite, specifically 5-farnesyl-4,7,9-trihydroxy-dibenzodiazepin-11-one (named "BU-4664L"), produced by a known euactino-mycetes strain, *Micromonospora* sp. M990-6 (ATCC 55378). Both these dibenzodiazepinones have been reported to have anti-tumor activity.

[0007] Although many biologically active compounds have been identified from bacteria, there remains the need to obtain novel naturally occurring compounds with enhanced properties. Current methods of obtaining such compounds include screening of natural isolates and chemical modification of existing compounds, both of which are costly and time consuming. Current screening methods are based on general biological properties of the compound, which require prior knowledge of the structure of the molecules. Methods for chemically modifying known active compounds exist, but still suffer from practical limitations as to the type of compounds obtainable.

[0008] Thus, there exists a considerable need to obtain pharmaceutically active compounds in a cost-effective manner and with high yield. The present invention solves these problems by providing polynucleotides, polypeptides, vectors comprising the polynucleotides and host cells comprising the vectors for production of dibenzodiazepinones, as well as methods to generate farnesyl dibenzodiazepinones by de novo biosynthesis (heterologous or homologous expression of biosynthetic genes) or semi-synthesis rather than by chemical synthesis.

SUMMARY OF THE INVENTION

[0009] The invention further encompasses an isolated polynucleotide comprising one or more of SEQ ID NOs. 1, 64 and 73, wherein the polynucleotide encodes a polypeptide that participates in a biosynthetic pathway for a farnesyl dibenzodiazepinone.

[0010] The invention further encompasses an isolated polynucleotide comprising SEQ ID NOs. 1, 64 and 73, wherein the polynucleotide encodes a polypeptide that participates in a biosynthetic pathway for a farnesyl dibenzodiazepinone.

[0011] The invention further encompasses an isolated polynucleotide that encodes a polypeptide selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 71, 74, 76, 78, 80, 82, 84, 86, 88, 90, 92, 94 and 96.

[0012] The invention further provides an isolated nucleic acid comprising a nucleotide sequence identical or complementary to a polynucleotide encoding a polypeptide having at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95% or at least 99% identity to a sequence selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 71, 74, 76, 78, 80, 82, 84, 86, 88, 90, 92, 94 and 96 said polypeptide having the same biological function as its corresponding protein.

[0013] The invention further provides an isolated nucleic acid comprising a nucleotide sequence hybridizing under low, moderate, high or very high stringency conditions to the complement of a polynucleotide encoding a sequence selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 71, 74, 76, 78, 80, 82, 84, 86, 88, 90, 92, 94 and 96, said polypeptide having the same biological function as its corresponding protein.

[0014] The invention provides an isolated, purified or enriched nucleic acid comprising a polynucleotide, or a nucleotide sequence complementary thereto, said polynucleotide encoding a polypeptide selected from an adenylating amide synthetase (ADSA) having at least 80%, at least 90%, or at least 95% identity to the adenylating amide synthetase of SEQ ID NO: 48; and an isoprenyl transferase (IPTN) having at least 80%, at least 90%, or at least 95% identity to the isoprenyl transferase of SEQ ID NO: 22. In one embodiment, the invention provides an expression vector comprising said ADSA or IPTN-encoding nucleic acid. In another embodiment, the invention provides host cells transformed which such vector.

[0015] The invention further provides a polypeptide selected from an adenylating amide synthetase (ADSA) having at least 80%, at least 90%, or at least 95% identity to the adenylating amide synthetase of SEQ ID NO: 48; and an isoprenyl transferase (IPTN) having at least 80%, at least 90%, or at least 95% identity to the isoprenyl transferase of SEQ ID NO: 22.

[0016] In one embodiment, the isolated polynucleotide comprising SEQ ID No. 1 encodes a polypeptide selected from the group consisting of SEQ ID Nos. 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60 and 62.

[0017] In another embodiment, the isolated polynucleotide comprising SEQ ID No. 64 encodes a polypeptide selected from the group consisting of SEQ ID NOS: 65, 67, 69 and 71.

[0018] In another embodiment, the isolated polynucleotide comprising SEQ ID No. 73, encodes a polypeptide selected from the group consisting of SEQ ID NOS: 74, 76, 78, 80, 82, 84, 86 and 88.

[0019] The invention further encompasses an isolated polypeptide of SEQ ID NO. 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 71, 74, 76, 78, 80, 82, 84, 86, 88, 90, 92, 94 and 96.

[0020] The invention further provides an isolated polypeptide having at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95% or at least 99% identity to a sequence selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 71, 74, 76, 78, 80, 82, 84, 86, 88, 90, 92, 94 and 96, said polypeptide having the same biological function as its corresponding protein.

[0021] In one embodiment, the polypeptide participates in a biosynthetic pathway for a farnesyl dibenzodiazepinone.

[0022] The invention further encompasses an expression vector comprising one or more of the polynucleotides described herein.

[0023] The invention further encompasses a recombinant prokaryotic organism comprising one or more such expression vectors.

[0024] In one embodiment, the organism is an actinomycete.

[0025] In another embodiment, the organism requires the expression vector to synthesize a farnesyl dibenzodiazepinone. That is, the organism is deficient in the ability to synthesize a farnesyl dibenzodiazepinone before transformation with a polynucleotide as described herein.

[0026] The invention further encompasses a method of making a farnesyl dibenzodiazepinone de novo in a prokaryote, comprising the steps of: (a) providing a prokaryote that is incapable of synthesizing a farnesyl dibenzodiazepinone; (b) transforming the prokaryote with an expression vector as described herein; and (c) culturing the prokaryote under conditions such that a polypeptide of the invention is expressed and catalyses the synthesis of a farnesyl dibenzodiazepinone compound or analog.

[0027] In one embodiment, the prokaryote is an actinomycete.

[0028] In another embodiment, the vector expresses a polypeptide of SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 71, 74, 76, 78, 80, 82, 84, 86, 88, 90, 92, 94 and 96.

BRIEF DESCRIPTION OF THE FIGURES

[0029] FIG. 1: shows inhibition of tumor growth resulting from administration of 10 to 30 mg/kg of ECO-04601 to glioblastoma-bearing mice beginning one day after tumor cell inoculation.

[0030] FIG. **2**: shows inhibition of tumor growth resulting from administration of 20-30 mg/kg of ECO-04601 to glioblastoma-bearing mice beginning ten days after tumor cell inoculation.

[0031] FIG. **3**: shows micrographs of tumor sections from mice bearing glioblastoma tumors and treated with saline or ECO-04601. The cell density of tumor treated with ECO-04601 appears decreased and nuclei from ECO-04601-treated tumor cells are larger and pynotic suggesting a cytotoxic effect.

[0032] FIG. **4**: shows the biosynthetic locus of ECO-04601, isolated from *Micromonospora* sp. strain 046-ECO11, including the positions of cosmids 046KM and 046KQ.

[0033] FIGS. 5 to 8: show the different steps involved in the biosynthetic pathway of ECO-04601. Each of FIGS. 5 to 8 shows the three biosynthetic loci A, B and C where ORFs are represented by arrows. Highlighted ORFs are involved in the steps described in the schematic diagram. The biosynthetic enzymes involved in the steps depicted in schematic diagrams are indicated by their family designation and the respective ORF number in each of Loci A, B and C (e.g., 8/7/7).

[0034] FIG. **5**: shows a schematic diagram of the biosynthetic pathway for the production of farnesyl-diphosphate, providing the farnesyl group of ECO-04601.

[0035] FIG. **6**: shows a schematic diagram of the biosynthetic pathway for the production of 3-hydroxy-anthranilateadenylate precursor of the dibenzodiazepinone group.

[0036] FIG. 7: shows a schematic diagram of the biosynthetic pathway for the production of 2-amino-6-hydroxy-[1, 4]benzoquinone precursor of the core dibenzodiazepinone.

[0037] FIG. **8**: shows a schematic diagram of the biosynthetic pathway for the assembly of the ECO-04601 precursors, farnesyl-diphosphate, 3-hydroxy-anthranilate-adeny-late and 2-amino-6-hydroxy-[1,4]benzoquinone.

[0038] FIGS. 9 and 10: show clustal alignments respectively of isoprenyl transferase and adenylating amide synthetase enzymes of locus A with the corresponding enzymes present in loci B and C. In each of the clustal alignments: (i) an asterisk "*" indicates positions which have a single, fully conserved residues; (ii) a colon ":" indicates that one of the following strong groups is fully conserved in a specific position: (S, T or A); (N, E, Q or K); (N, H, Q or K); (N, D, E or Q); (Q, H, R or K); (M, I, L or V); (M, I, L or F); (H or Y); and (F, Y or W); and (iii) a period "." indicates that one of the following weaker groups is fully conserved: (C, S or A); (A, T or V); (S, A or G); (S, T, N or K); (S, T, P or A); (S, G, N or D); (S, N, D, E, Q or K); (N, D, E, Q, H or K); (N, E, Q, H, R or K); (F, V, L, I or M): and (H, F or Y). The number at the end of each line indicates the position of the last amino acid of the line within the specific domain.

[0039] FIG. **9**: shows an amino acid alignment comparing the isoprenyl transferase (IPTN) enzyme of locus A (SEQ ID NO: 22), isolated from *Micronospora* sp. strain 046-ECO11, with the isoprenyl transferase enzyme of locus B (SEQ ID NO 90) isolated from *Micromonospora echinospora challisensis* NRRL 12255, and the partial isoprenyl transferase enzyme of locus C (SEQ ID NO: 94) isolated from *Streptomyces carzinostaticus neocarzinostaticus* ATCC 15944.

[0040] FIG. **10**: shows an amino acid alignment comparing the adenylating amide synthetase (ADSA) enzyme of locus A (SEQ ID NO: 48), isolated from *Micronospora* sp. strain 046-ECO11, with the adenylating amide synthetase of locus B (SEQ ID NO 92) isolated from *Micromonospora echinospora challisensis* NRRL 12255, and locus C (SEQ ID NO: 96) isolated from *Streptomyces carzinostaticus neocarzinostaticus* ATCC 15944.

DETAILED DESCRIPTION OF THE INVENTION

[0041] The present invention provides isolated and purified polynucleotides that encode farnesyl dibenzodiazepinoneproducing enzymes, i.e., polypeptides from farnesyl dibenzodiazepinone-producing microorganisms, fragments thereof, vectors containing those polynucleotides, and host cells transformed with those vectors. These polynucleotides, fragments thereof, and vectors comprising the polynucleotides can be used as reagents in the production of farnesyl dibenzodiazepinones. The invention also relates to a method for producing new farnesyl dibenzodiazepinones, by selectively altering the genetic information of an organism or by feeding the proteins or a host cell transformed with vectors comprising nucleic acids encoding them, with close analogs of the key intermediates. Portions of the polynucleotide sequences disclosed herein are also useful as primers for the amplification of DNA or as probes to identify related domains from other farnesyl dibenzodiazepinone producing microorganisms.

I. Definitions

[0042] For convenience, the meaning of certain terms and phrases used in the specification, examples, and appended claims, are provided below.

[0043] As used herein, the term "farnesyl dibenzodiazepinone" refers to a class of dibenzodiazepinone compounds containing a farnesyl moiety. The term includes, but is not limited to, the exemplified compound of the present invention, 10-farnesyl-4,6,8-trihydroxy-dibenzodiazepin-11-one, which is referred to herein as "ECO-04601."

[0044] The terms "farnesyl dibenzodiazepinone-producing microorganism" and "producer of farnesyl dibenzodiazepinone," as used herein, refer to a microorganism that carries genetic information necessary to produce a farnesyl dibenzodiazepinone compound, whether or not the organism naturally produces the compound. The terms apply equally to organisms in which the genetic information to produce the farnesyl dibenzodiazepinone compound is found in the organism as it exists in its natural environment, and to organisms (host cells) in which the genetic information is introduced by recombinant techniques.

[0045] Specific organisms contemplated herein include, without limitation, organisms of the family Micromonosporaceae, of which preferred genera include *Micromonospora*, Actinoplanes and Dactylosporangium; the family Streptomycetaceae, of which preferred genera include Streptomyces and Kitasatospora; the family Pseudonocardiaceae, of which preferred genera are Amycolatopsis and Saccharopolyspora; and the family Actinosynnemataceae, of which preferred genera include Saccharothrix and Actinosynnema; however the terms are intended to encompass all organisms containing genetic information necessary to produce a farnesyl dibenzodiazepinone compound. A preferred producer of a farnesyl dibenzodiazepinone compound includes microbial strain 046-ECO11, a deposit of which was made on Mar. 7, 2003, with the International Depository Authority of Canada (IDAC), Bureau of Microbiology, Health Canada, 1015 Arlington Street, Winnipeg, Manitoba, Canada R3E 3R2, under Accession No. IDAC 070303-01.

[0046] The term "gene" means the segment of DNA involved in producing a polypeptide chain; it includes regions preceding and following the coding region (leader and trailer) as well as, where applicable, intervening regions (introns) between individual coding segments (exons).

[0047] The terms "gene locus, "gene cluster," and "biosynthetic locus" refer to a group of genes or variants thereof involved in the biosynthesis of a farnesyl dibenzodiazepinone compound. For example, the biosynthetic locus in strain 046-ECO11 that directs the production of ECO-04601 referred to herein as "046D" or "locus A", the biosynthetic locus in Micromonospora echinospora challisensis NRRL 12255 referred to herein as "052E" or "locus B", the biosynthetic locus in Streptomyces carzinostaticus neocarzinostaticus ATCC 15944 referred to herein as "237C" or "locus C", or the corresponding biosynthetic locus from a farnesyl dibenzodiazepinone-producing microorganism. Genetic modification of gene locus, gene cluster or biosynthetic locus refers to any genetic recombinant techniques known in the art including mutagenesis, inactivation, or replacement of nucleic acids that can be applied to generate variants of ECO-04601.

[0048] A DNA or nucleotide "coding sequence" or "sequence encoding" a particular polypeptide or protein, is a DNA sequence which is transcribed and translated into a polypeptide or protein when placed under the control of an appropriate regulatory sequence.

[0049] "Oligonucleotide" refers to a nucleic acid, generally of at least 10, preferably 15 and more preferably at least 20 nucleotides in length, preferably no more than 100 nucleotides in length, that are hybridizable to a genomic DNA molecule, a cDNA molecule, or an mRNA molecule encoding a gene, mRNA, cDNA or other nucleic acid of interest.

[0050] A promoter sequence is "operably linked to" a coding sequence recognized by RNA polymerase which initiates transcription at the promoter and transcribes the coding sequence into mRNA.

[0051] The term "repl icon" as used herein means any genetic element, such as a plasmid, cosmid, chromosome or virus, that behaves as an autonomous unit of polynucleotide replication within a cell. An "expression vector" or "vector" is a replicon in which another polynucleotide fragment is attached, such as to bring about the replication and/or expression of the attached fragment. "Plasmids" are designated herein by a lower case "p" preceded or followed by capital letters and/or numbers. The starting plasmids disclosed herein are commercially available, publicly available on an unrestricted basis, or can be constructed from available plasmids in accordance with published procedures. In addition, equivalent plasmids to those described herein are known in the art and will be apparent to the skilled artisan.

[0052] The terms "express" and "expression" means allowing or causing the information in a gene or DNA sequence to become manifest, for example producing a protein by activating the cellular functions involved in transcription and translation of a corresponding gene or DNA sequence. A DNA sequence is expressed in or by a cell to form an "expression product" such as a protein. The expression product itself, e.g. the resulting protein, may also be said to be "expressed" by the cell. An expression product can be characterized as intracellular, extracellular or secreted.

[0053] "Digestion" of DNA refers to enzymatic cleavage of the DNA with a restriction enzyme that acts only at certain sequences in the DNA. The various restriction enzymes used herein are commercially available and their reaction conditions, cofactors and other requirements were used as would be known to the ordinary skilled artisan. For analytical purposes, typically 1 µg of plasmid or DNA fragment is used with about 2 units of enzyme in about 20 μ l of buffer solution. For the purpose of isolating DNA fragments for plasmid construction, typically 5 to 50 µg of DNA are digested with 20 to 250 units of enzyme in a larger volume. Appropriate buffers and substrate amounts for particular enzymes are specified by the manufacturer. Incubation times of about 1 hour at 37° C. are ordinarily used, but may vary in accordance with the supplier's instructions. After digestion the gel electrophoresis may be performed to isolate the desired fragment.

[0054] The term "isolated" as used herein means that the material is removed from its original environment (e.g. the natural environment where the material is naturally occurring). For example, a naturally occurring polynucleotide or polypeptide present in a living animal is not isolated, but the same polynucleotide or polypeptide, which is separated from some or all of the coexisting materials in the natural system, is isolated. Such polynucleotides could be part of a vector and/or such polynucleotides or polypeptides could be part of a composition, and still be isolated in that the vector or composition is not part of the natural environment.

[0055] The term "restriction fragment" as used herein refers to any linear DNA generated by the action of one or more restriction enzymes.

[0056] The term "transformation" means the introduction of a foreign gene, foreign nucleic acid, DNA or RNA sequence to a host cell, so that the host cell will express the introduced gene or sequence to produce a desired substance, typically a protein or enzyme coded by the introduced gene or sequence. The introduced gene or sequence may also be called a "cloned" or "foreign" gene or sequence, may include regulatory or control sequences, such as start, stop, promoter, signal, secretion, or other sequences used by a cell's genetic machinery. The gene or sequence may include nonfunctional sequences or sequences with no known function. A host cell that receives and expresses introduced DNA or RNA has been "transformed" and is a "transformant" or a "clone" or "recombinant". The DNA or RNA introduced to a host cell can come from any source, including cells of the same genus or species as the host cell, or cells of a different genus or species.

[0057] The terms "recombinant polynucleotide" and "recombinant polypeptide" as used herein mean a polynucleotide or polypeptide which by virtue of its origin or manipulation is not associated with all or a portion of the polynucleotide or polypeptide with which it is associated in nature and/or is linked to a polynucleotide or polypeptide other than that to which it is linked in nature.

[0058] The term "host cell" as used herein, refer to both prokaryotic and eukaryotic cells which are used as recipients of the recombinant polynucleotides and vectors provided herein. In one embodiment, the host cell is a prokaryote.

[0059] The terms "open reading frame" and "ORF" as used herein refers to a region of a polynucleotide sequence which encodes a polypeptide; this region may represent a portion of a coding sequence or a total coding sequence.

[0060] As used herein and as known in the art, the term "identity" is the relationship between two or more polynucleotide sequences, as determined by comparing the sequences. Identity also means the degree of sequence relatedness between polynucleotide sequences, as determined by the match between strings of such sequences. Identity can be readily calculated (see, e.g., Computation Molecular Biology, Lesk, A. M., eds., Oxford University Press, New York (1998), and Biocomputing: Informatics and Genome Projects, Smith, D. W., ed., Academic Press, New York (1993), both of which are incorporated by reference herein). While there exist a number of methods to measure identity between two polynucleotide sequences, the term is well known to skilled artisans (see, e.g., Sequence Analysis in Molecular Biology, von Heinje, G., Academic Press (1987); and Sequence Analysis Primer, Gribskov., M. and Devereux, J., eds., M. Stockton Press, New York (1991)). Methods commonly employed to determine identity between sequences include, for example, those disclosed in Carillo, H., and Lipman, D., SIAM J. Applied Math. (1988) 48:1073. "Substantially identical," as used herein, means there is a very high degree of homology (preferably 100% sequence identity) between subject polynucleotide sequences. However, polynucleotides having greater than 90%, or 95% sequence identity may be used in the present invention, and thus sequence variations that might be expected due to genetic mutation, strain polymorphism, or evolutionary divergence can be tolerated.

II. Method of Making a Farnesyl Dibenzodiazepinone by Fermentation

[0061] The farnesyl dibenzodiazepinone compounds of the present invention may be biosynthesized by various microorganisms. Microorganisms that may synthesize the compounds of the present invention include but are not limited to bacteria of the order Actinomycetales, also referred to as actinomycetes. Non-limiting examples of members belonging to the genera of Actinomycetes include *Nocardia, Geo*

dermatophilus, Actinoplanes, Micromonospora, Nocardioides, Saccharothrix, Amycolatopsis, Kutzneria, Saccharomonospora, Saccharopolyspora, Kitasatospora, Streptomyces, Microbispora, Streptosporangium, and Actinomadura. The taxonomy of actinomycetes is complex and reference is made to Goodfellow, Suprageneric Classification of Actinomycetes (1989); Bergey's Manual of Systematic Bacteriology, Vol. 4 (Williams and Wilkins, Baltimore, pp. 2322-2339); and to Embley and Stackebrandt, "The molecular phylogeny and systematics of the actinomycetes," Annu. Rev. Microbiol. (1994) 48:257-289, each of which is hereby incorporated by reference in its entirety, for genera that may synthesize the compounds of the invention.

[0062] Farnesyl dibenzodiazepinone-producing microorganisms are cultivated in culture medium containing known nutritional sources for actinomycetes. Such media having assimilable sources of carbon, nitrogen, plus optional inorganic salts and other known growth factors at a pH of about 6 to about 9. Suitable media include, without limitation, the growth media provided in Table 1. Microorganisms are cultivated at incubation temperatures of about 18° C. to about 40° C. for about 3 to about 40 days. dibenzodiazepinone compounds can be extracted and isolated from the cultivated culture media by techniques known to a skilled person in the art and/or disclosed herein, including for example centrifugation, chromatography, adsorption, filtration. For example, the cultivated culture media can be mixed with a suitable organic solvent such as n-butanol, n-butyl acetate or 4-methyl-2-pentanone, the organic layer can be separated for example, by centrifugation followed by the removal of the solvent, by evaporation to dryness or by evaporation to dryness under vacuum. The resulting residue can optionally be reconstituted with for example water, ethanol, ethyl acetate, methanol or a mixture thereof, and re-extracted with a suitable organic solvent such as hexane, carbon tetrachloride, methylene chloride or a mixture thereof. Following removal of the solvent, the compounds may be further purified by the use of standard techniques, such as chromatography.

III. Method of Making a Farnesyl Dibenzodiazepinone by Recombinant Technology

[0063] In another embodiment, the present invention relates to nucleic acid molecules that encode proteins useful

TABLE 1

Examples of Fermentation Media								
Component	QB	MA	КН	RM	JA	FA	HI	CL
pH *1	7.2	7.5	7	6.85	7.3	7.0	7.0	7.0
Glucose	12		10	10		10		
Sucrose				100				
Cane molasses						15		
Corn starch					30			
Soluble starch	10	25						
Potato dextrin			20			40	20	20
Corn steep solid	5							
Corn steep liquor	5				15			
Dried yeast		2						
Yeast extract			5				8.34	
Malt extract					35			
Pharmamedia ™	10				15			
Glycerol							30	20
NZ-Amine A			5			10		
Soybean powder		15						
Fish meal								10
Bacto-peptone							2.5	5
MgSO ₄ •7H ₂ O						1		
CaCO ₃		4	1		2	2	3	2
NaCl		5						
$(NH_4)_2SO_4$		2						2
$K_2 SO_4$				0.25				
MgCl ₂ •6H ₂ O				10				
Na ₂ HPO ₄						3		
Casamino acid				0.1		-		
Proflo oil ™ (mL/L)	4							
MOPS	•			21				
Trace element				2				
solution * ² ml/L				-				

Unless otherwise indicated all the ingredients are in g/L.

*1 The pH is to adjusted as marked prior to the addition of CaCO3.

*² Trace elements solution contains: ZnCl₂ 40 mg; Fe Cl₃ 6H₂O (200 mg); CuCl₂ 2H₂O (10

mg); MnCl₂•4H₂O; Na₂B₄O₇•10H₂O (10 mg); (NH₄)₆ MO₇O₂₄•4H₂O (10 mg) per litre.

The culture media inoculated with the farnesyl dibenzodiazepinone-producing microorganisms may be aerated by incubating the inoculated culture media with agitation, for example, shaking on a rotary shaker, or a shaking water bath. Aeration may also be achieved by the injection of air, oxygen or an appropriate gaseous mixture to the inoculated culture media during incubation. Following cultivation, the farnesyl in the production of farnesyl benzodiazepinones. Specifically, the present invention provides recombinant DNA vectors and nucleic acid molecules that encode all or part of the biosynthetic locus in strain 046-ECO11, which directs the production of ECO-04601, and is referred to herein as "046D." The invention further includes genetic modification of 046D using conventional genetic recombinant techniques, such as mutagenesis, inactivation, or replacement of nucleic acids, to produce chemical variants of ECO-04601.

[0064] The invention thus provides a method for making a farnesyl benzodiazepinone compound using a transformed host cell comprising a recombinant DNA vector that encodes one or more of the polypeptides of the present invention, and culturing the host cell under conditions such that farnesyl benzodiazepinone is produced. In one embodiment, the host cell is an actinomycete. In another embodiment, the host cell is a *Streptomyces* host cell. In a further embodiment, the host cell is a non-*Streptomyces* actinomycete such as a *Rhodococcus*, a *Mycobaterium*, or an *Amycolatopsis* specie.

[0065] The invention provides recombinant nucleic acids that produce a variety of farnesyl dibenzodiazepinone compounds that cannot be readily synthesized by chemical methodology alone. The invention allows direct manipulation of 046D biosynthetic locus via genetic engineering of the enzymes involved in the biosynthesis of a farnesyl dibenzodiazepinone according to the invention. The 046D biosynthetic locus is described in Example 5.

[0066] Farnesyl dibenzodazepinones and analogs are also produced by feeding one or more key intermediates or biosynthetic precursors (as defined in FIGS. **5-8**) or close structural analogs, to a host cell comprising a recombinant DNA vector that encodes one or more of the polypeptides of the present invention, and culturing the host cell under conditions such that the farnesyl benzodiazepinone or analog is produced. Key intermediates are contacted directly with an isolated protein of the invention to perform the necessary steps for the production of a farnesyl dibenzodiazepinone (e.g., the farnesyl diphopshate and dibenzodiazepinone precursors can be coupled using an IPTN protein of the invention).

[0067] Key intermediates may be commercially available or may be prepared using standard chemical procedures or using the proteins of this invention. For example, farnesyl diphosphate and 3-hydroxyanthranilic acid are commercially available (e.g., Fluka F6892 and Aldrich 148776). 3-Amino-5-hydroxybenzoic acid, a precursor of the 2-amino-6-hydroxybenzoquinone, is prepared as described in Herlt et al (1981), *Aust. J. Chem.*, vol 34, 1319-1324.

[0068] Recombinant DNA Vectors

[0069] Vectors of the invention typically comprise the DNA of a transmissible agent, into which foreign DNA is inserted. A common way to insert one segment of DNA into another segment of DNA involves the use of specific enzymes called restriction enzymes that cleave DNA at specific sites (specific groups of nucleotides) called restriction sites. A "cassette" refers to a DNA coding sequence or segment of DNA that codes for an expression product that can be inserted into a vector at defined restriction sites. The cassette restriction sites are designed to ensure insertion of the cassette in the proper reading frame. Generally, a nucleic acid molecule that encodes a protein useful in the production of a farnesyl dibenzodiazepinone is inserted at one or more restriction sites of the vector DNA, and then is carried by the vector into a prokaryote e.g. actinomycte, by transformation (see below). A segment or sequence of DNA having inserted or added DNA, such as an expression vector, can also be called a "DNA construct". A common type of vector is a "plasmid" which generally is a self-contained molecule of double-stranded DNA, usually of bacterial origin, that can readily accept additional (foreign) DNA and which can be readily introduced into a suitable host cell. A plasmid vector often contains coding DNA and promoter DNA and has one or more restriction sites suitable for inserting foreign DNA. Coding DNA is a DNA sequence that encodes a particular amino acid sequence for a particular protein or enzyme. In one embodiment of the invention, the coding DNA encodes for polypeptides of SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86, 88, 90, 92, 94, 96 or 98 that may be useful for the biosynthesis of a farnesyl dibenzodiazepinone.

[0070] Promoter DNA of a recombinant vector is a DNA sequence that initiates, regulates, or otherwise mediates or controls the expression of the coding DNA. Promoter DNA and coding may be from the same or different organisms. Recombinant cloning vectors will often include one or more replication systems for cloning or expression, one or more markers for selection in the host, e.g. antibiotic resistance, and one or more expression cassettes. Vector constructs may be produced using conventional molecular biology and recombinant DNA techniques within the skill of the art. Such techniques are explained fully in the literature. See, e.g., Sambrook, Fritsch & Maniatis, Molecular Cloning: A Laboratory Manual, Second Edition (1989) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (herein "Sambrook et al., 1989"); DNA Cloning: A Practical Approach, Volumes I and II (D. N. Glover ed. 1985); F. M. Ausubel et al. (eds.), Current Protocols in Molecular Biology, John Wiley & Sons, Inc. (1994).

[0071] Examples of promoters that function in actinomycetes, e.g. *Streptomyces*, are taught in U.S. Pat. Nos. 5,830,695 and 5,466,590. Another example of a transcription promoter useful in Actinomycetes expression vectors is tipA, a promoter inducible by the antibiotic thiostrepton [c.f. Murakami, T., et al., (1989), J. Bacteriol, 171, 1459].

[0072] Transformation of Actinomycetes

[0073] A suitable transformation method for use with an actinomycete comprises forming the actinomycete culture into spheroplasts using lysozyme. A buffer solution containing recombinant DNA vectors and polyethylene glycol is then added, in order to introduce the vector into the host cells, by using either of the methods of Thompson or Keiser [c. f. Thompson, C. J., et al., (1982), J. Bacteriol., 151, 668-677 or Keiser, T. et al. (2000), "Practical *Streptomyces* Genetics", The John Innes Foundation, Norwich], for example. A thiostrepton-resistance gene is frequently used as a selective marker in the transformation plasmid [c.f. Hopwood, D. A., et al., (1987), "Methods in Enzymology" 153, 116, Academic Press, New York], but the present invention is not limited thereto. Additional methods for the transformation of actinomycetes are taught in U.S. Pat. No. 5,393,665.

[0074] Assay for Farnesyl Dibenzodiazepinone or Biosynthetic Intermediates

[0075] Actinomycetes defective in farnesyl dibenzodiazepinone biosynthesis are transformed with one or more expression vectors encoding one or more proteins in the farnesyl benzodiazepinone biosynthetic pathway, thus restoring farnesyl benzodiazepinone biosynthesis by genetic complementation of the specific defect.

[0076] The presence or absence of farnesyl dibenzodiazepinone or intermediates in the biosynthetic pathway (see FIGS. **5** to **8**) in a recombinant actinomycete can be determined using methodologies that are well known to persons of skill in the art. For example, ethyl acetate extracts of fermentation media used for the culture of a recombinant actinomycete are processed as described in Example 2 and fractions containing farnesyl dibenzodiazepinone or intermediates detected by TLC on commercial Kieselgel 60 F_{254} plates. Farnesyl dibenzodiazepinone and intermediate compounds are visualized by inspection of dried plates under UV light or by spraying the plates with a spray containing vanillin (0.75%) and concentrated sulfuric acid (1.5%, v/v) in ethanol and subsequently heating the plate. The exact identity of the compounds separated by TLC is then determined using gas chromatography-mass spectroscopy. Methods of mass spectroscopy are taught in the published U.S. Patent Application No. US2003/0052268.

[0077] Mutagenesis

[0078] The invention allows direct manipulation of 046D biosynthetic locus via genetic engineering of the enzymes involved in the biosynthesis of a farnesyl benzodiazepinone according to the invention.

[0079] A number of methods are known in the art that permit the random as well as targeted mutation of the DNA sequences of the invention (see for example, Ausubel et. al. Short Protocols in Molecular Biology (1995) 3rd Ed. John Wiley & Sons, Inc.). In addition, there are a number of commercially available kits for site-directed mutagenesis, including both conventional and PCR-based methods. Examples include the EXSITETM PCR-Based Site-directed Mutagenesis Kit available from Stratagene (Catalog No. 200502) and the QUIKCHANGETM Site-directed mutagenesis Kit from Stratagene (Catalog No. 200518), and the CHAMELEON® double-stranded Site-directed mutagenesis kit, also from Stratagene (Catalog No. 200509).

[0080] In addition the nucleotides of the invention may be generated by insertional mutation or truncation (N-terminal, internal or C-terminal) according to methodology known to a person skilled in the art.

[0081] Older methods of site-directed mutagenesis known in the art rely on sub-cloning of the sequence to be mutated into a vector, such as an M13 bacteriophage vector, that allows the isolation of single-stranded DNA template. In these methods, one anneals a mutagenic primer (i.e., a primer capable of annealing to the site to be mutated but bearing one or more mismatched nucleotides at the site to be mutated) to the single-stranded template and then polymerizes the complement of the template starting from the 3' end of the mutagenic primer. The resulting duplexes are then transformed into host bacteria and plaques are screened for the desired mutation.

[0082] More recently, site-directed mutagenesis has employed PCR methodologies, which have the advantage of not requiring a single-stranded template. In addition, methods have been developed that do not require sub-cloning. Several issues must be considered when PCR-based site-directed mutagenesis is performed. First, in these methods it is desirable to reduce the number of PCR cycles to prevent expansion of undesired mutations introduced by the polymerase. Second, a selection must be employed in order to reduce the number of non-mutated parental molecules persisting in the reaction. Third, an extended-length PCR method is preferred in order to allow the use of a single PCR primer set. And fourth, because of the non-template-dependent terminal extension activity of some thermostable polymerases it is often necessary to incorporate an end-polishing step into the procedure prior to blunt-end ligation of the PCR-generated mutant product.

[0083] The protocol described below accommodates these considerations through the following steps. First, the template concentration used is approximately 1000-fold higher than that used in conventional PCR reactions, allowing a reduction in the number of cycles from 25-30 down to 5-10 without dramatically reducing product yield. Second, the restriction endonuclease Dpn I (recognition target sequence: 5-Gm6ATC-3, where the A residue is methylated) is used to select against parental DNA, since most common strains of *E. coli* Dam methylate their DNA at the sequence 5-GATC-3. Third, Taq Extender is used in the PCR mix in order to increase the proportion of long (i.e., full plasmid length) PCR products. Finally, Pfu DNA polymerase is used to polish the ends of the PCR product prior to intramolecular ligation using T4 DNA ligase.

[0084] A non-limiting example for the isolation of mutant polynucleotides is described in detail as follows:

[0085] Plasmid template DNA (approximately 0.5 pmole) is added to a PCR cocktail containing: 1× mutagenesis buffer (20 mM Tris HCl, pH 7.5; 8 mM MgCl₂; 40 µg/ml BSA); 12-20 pmole of each primer (one of skill in the art may design a mutagenic primer as necessary, giving consideration to those factors such as base composition, primer length and intended buffer salt concentrations that affect the annealing characteristics of oligonucleotide primers; one primer must contain the desired mutation, and one (the same or the other) must contain a 5' phosphate to facilitate later ligation), 250 μM each dNTP, 2.5 U Taq DNA polymerase, and 2.5 U of Taq Extender (Available from Stratagene; See Nielson et al. (1994) Strategies 7: 27, and U.S. Pat. No. 5,556,772). Primers can be prepared using the triester method of Matteucci et al., 1981, J. Am. Chem. Soc. 103:3185-3191, incorporated herein by reference. Alternatively automated synthesis may be preferred, for example, on a Biosearch 8700 DNA Synthesizer using cyanoethyl phosphoramidite chemistry.

[0086] The PCR cycling is performed as follows: 1 cycle of 4 min at 94° C., 2 min at 50° C. and 2 min at 72° C.; followed by 5-10 cycles of 1 min at 94° C., 2 min at 54° C. and 1 min at 72° C. The parental template DNA and the linear, PCRgenerated DNA incorporating the mutagenic primer are treated with DpnI (10 U) and Pfu DNA polymerase (2.5 U). This results in the DpnI digestion of the in vivo methylated parental template and hybrid DNA and the removal, by Pfu DNA polymerase, of the non-template-directed Taq DNA polymerase-extended base(s) on the linear PCR product. The reaction is incubated at 37° C. for 30 min and then transferred to 72° C. for an additional 30 min. Mutagenesis buffer (115 ul of 1x) containing 0.5 mM ATP is added to the DpnI-digested, Pfu DNA polymerase-polished PCR products. The solution is mixed and 10 ul are removed to a new microfuge tube and T4 DNA ligase (2-4 U) is added. The ligation is incubated for greater than 60 min at 37° C. Finally, the treated solution is transformed into competent E. coli according to standard methods.

[0087] Methods of random mutagenesis, which will result in a panel of mutants bearing one or more randomly situated mutations, exist in the art. Such a panel of mutants may then be screened for those exhibiting reduced uracil detection activity relative to the wild-type polymerase (e.g., by measuring the incorporation of 10 nmoles of dNTPs into polymeric form in 30 minutes in the presence of 200 μ M dUTP and at the optimal temperature for a given DNA polymerase). An example of a method for random mutagenesis is the socalled "error-prone PCR method". As the name implies, the method amplifies a given sequence under conditions in which the DNA polymerase does not support high fidelity incorporation. The conditions encouraging error-prone incorporation for different DNA polymerases vary, however one skilled in the art may determine such conditions for a given enzyme. A key variable for many DNA polymerases in the fidelity of amplification is, for example, the type and concentration of divalent metal ion in the buffer. The use of manganese ion and/or variation of the magnesium or manganese ion concentration may therefore be applied to influence the error rate of the polymerase.

[0088] Genes for desired mutant polypeptides generated by mutagenesis may be sequenced to identify the sites and number of mutations. For those mutants comprising more than one mutation, the effect of a given mutation may be evaluated by introduction of the identified mutation to the wild-type gene by site-directed mutagenesis in isolation from the other mutations borne by the particular mutant. Screening assays of the single mutant thus produced will then allow the determination of the effect of that mutation alone.

IV. Genes and Proteins for the Production of ECO-04601

[0089] As discussed in more detail below, the isolated, purified or enriched nucleic acids of one of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 66, 68, 70, 72, 75, 77, 79, 81, 83, 85, 87 and 89 may be used to prepare one of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88, respectively, or fragments comprising at least 50, 75, 100, 200, 300, 500 or more consecutive amino acids of one of the polypeptides of SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88, respectively.

[0090] Accordingly, another aspect of the present invention is an isolated, purified or enriched nucleic acid which encodes one of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 or fragments comprising at least 50, 75, 100, 150, 200, 300 or more consecutive amino acids of one of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88. The coding sequences of these nucleic acids may be identical to one of the coding sequences of one of the nucleic acids of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 66, 68, 70, 72, 75, 77, 79, 81, 83, 85, 87 and 89 or a fragment thereof, or may be different coding sequences which encode one of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 or fragments comprising at least 50, 75, 100, 150, 200, 300 consecutive amino acids of one of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 as a result of the redundancy or degeneracy of the genetic code. The genetic code is well known to those of skill in the art and can be obtained, for example, from Stryer, Biochemistry, 3rd edition, W.H. Freeman & Co., New York.

[0091] The isolated, purified or enriched nucleic acid which encodes one of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 may include, but is not limited to: (1) only the coding sequences of one of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 66, 68, 70, 72, 75, 77, 79, 81, 83, 85, 87 and 89; (2) the coding sequences of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 66, 68, 70, 72, 75, 77, 79, 81, 83, 85, 87 and 89 and additional coding sequences, such as leader sequences or proprotein; and (3) the coding sequences of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 66, 68, 70, 72, 75, 77, 79, 81, 83, 85, 87 and 89 and non-coding sequences, such as noncoding sequences 5' and/or 3' of the coding sequence. Thus, as used herein, the term "polynucleotide encoding a polypeptide" encompasses a polynucleotide that includes only coding sequence for the polypeptide as well as a polynucleotide that includes additional coding and/or non-coding sequence.

[0092] The invention relates to polynucleotides based on SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 66, 68, 70, 72, 75, 77, 79, 81, 83, 85, 87 and 89 but having polynucleotide changes that are "silent", for example changes which do not alter the amino acid sequence encoded by the polynucleotides of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 66, 68, 70, 72, 75, 77, 79, 81, 83, 85, 87 and 89. The invention also relates to polynucleotides which have nucleotide changes which result in amino acid substitutions, additions, deletions, fusions and truncations of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88. Such nucleotide changes may be introduced using techniques such as site directed mutagenesis, random chemical mutagenesis, exonuclease III deletion, and other recombinant DNA techniques.

[0093] The isolated, purified or enriched nucleic acids of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 66, 68, 70, 72, 75, 77, 79, 81, 83, 85, 87 and 89, the sequences complementary thereto, or a fragment comprising at least 100, 150, 200, 300, 400 or more consecutive bases of one of the sequence of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 66, 68, 70, 72, 75, 77, 79, 81, 83, 85, 87 and 89, or the sequences complementary thereto may be used as probes to identify and isolate DNAs encoding the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 respectively. In such procedures, a genomic DNA library is constructed from a sample microorganism or a sample containing a microorganism capable of producing a farnesyl dibenzodiazepinone. The genomic DNA library is then contacted with a probe comprising a coding sequence or a fragment of the coding sequence, encoding one of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88, or

a fragment thereof under conditions which permit the probe to specifically hybridize to sequences complementary thereto. In a preferred embodiment, the probe is an oligonucleotide of about 10 to about 30 nucleotides in length designed based on a nucleic acid of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 66, 68, 70, 72, 75, 77, 79, 81, 83, 85, 87 and 89. Genomic DNA clones which hybridize to the probe are then detected and isolated. Procedures for preparing and identifying DNA clones of interest are disclosed in Ausubel et al., Current Protocols in Molecular Biology, John Wiley 503 Sons, Inc. 1997; and Sambrook et al., Molecular Cloning: A Laboratory Manual 2d Ed., Cold Spring Harbor Laboratory Press, 1989. In another embodiment, the probe is a restriction fragment or a PCR amplified nucleic acid derived from SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 66, 68, 70, 72, 75, 77, 79, 81, 83, 85, 87 and 89.

[0094] The isolated, purified or enriched nucleic acids of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 66, 68, 70, 72, 75, 77, 79, 81, 83, 85, 87 and 89, the sequences complementary thereto, or a fragment comprising at least 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, 150, 200, 300, 400 or 500 consecutive bases of one of the sequences of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 66, 68, 70, 72, 75, 77, 79, 81, 83, 85, 87 and 89 or the sequences complementary thereto may be used as probes to identify and isolate related nucleic acids. In some embodiments, the related nucleic acids may be genomic DNAs (or cDNAs) from potential farnesyl dibenzodiazepinone producers. In such procedures, a nucleic acid sample containing nucleic acids from a potential farnesyl dibenzodiazepinone producer is contacted with the probe under conditions that permit the probe to specifically hybridize to related sequences. The nucleic acid sample may be a genomic DNA (or cDNA) library from the potential farnesyl dibenzodiazepinone-producer. Hybridization of the probe to nucleic acids is then detected using any of the methods described above.

[0095] Hybridization may be carried out under conditions of low stringency, moderate stringency or high stringency. As an example of nucleic acid hybridization, a polymer membrane containing immobilized denatured nucleic acids is first prehybridized for 30 minutes at 45° C. in a solution consisting of 0.9 M NaCl, 50 mM NaH₂PO₄, pH 7.0, 5.0 mM Na₂EDTA, 0.5% SDS, 10×Denhardt's, and 0.5 mg/ml polyriboadenylic acid. Approximately 2×10^7 cpm (specific activity $4-9 \times 10^8$ cpm/ug) of ³²P end-labeled oligonucleotide probe are then added to the solution. After 12-16 hours of incubation, the membrane is washed for 30 minutes at room temperature in 1×SET (150 mM NaCl, 20 mM Tris hydrochloride, pH 7.8, 1 mM Na₂EDTA) containing 0.5% SDS, followed by a 30 minute wash in fresh 1×SET at Tm-10° C. for the oligonucleotide probe where Tm is the melting temperature. The membrane is then exposed to autoradiographic film for detection of hybridization signals.

[0096] By varying the stringency of the hybridization conditions used to identify nucleic acids, such as genomic DNAs or cDNAs, which hybridize to the detectable probe, nucleic acids having different levels of homology to the probe can be identified and isolated. Stringency may be varied by conducting the hybridization at varying temperatures below the melting temperatures of the probes. The melting temperature of the probe may be calculated using the following formulas:

[0097] For oligonucleotide probes between 14 and 70 nucleotides in length the melting temperature (Tm) in degrees Celcius may be calculated using the formula: Tm=81.5+16. $6(\log [Na+])+0.41$ (fraction G+C)-(600/N) where N is the length of the oligonucleotide.

[0098] If the hybridization is carried out in a solution containing formamide, the melting temperature may be calculated using the equation $Tm=81.5+16.6(\log [Na+])+0.41$ (fraction G+C)-(0.63% formamide)-(600/N) where N is the length of the probe.

[0099] Prehybridization may be carried out in 6×SSC, 5×Denhardt's reagent, 0.5% SDS, 0.1 mg/ml denatured fragmented salmon sperm DNA or 6×SSC, 5×Denhardt's reagent, 0.5% SDS, 0.1 mg/ml denatured fragmented salmon sperm DNA, 50% formamide. The composition of the SSC and Denhardt's solutions are listed in Sambrook et al., supra.

[0100] Hybridization is conducted by adding the detectable probe to the hybridization solutions listed above. Where the probe comprises double stranded DNA, it is denatured by incubating at elevated temperatures and quickly cooling before addition to the hybridization solution. It may also be desirable to similarly denature single stranded probes to eliminate or diminish formation of secondary structures or oligomerization. The filter is contacted with the hybridization solution for a sufficient period of time to allow the probe to hybridize to cDNAs or genomic DNAs containing sequences complementary thereto or homologous thereto. For probes over 200 nucleotides in length, the hybridization may be carried out at 15-25° C. below the Tm. For shorter probes, such as oligonucleotide probes, the hybridization may be conducted at 5-10° C. below the Tm. Preferably, the hybridization is conducted in 6×SSC, for shorter probes. Preferably, the hybridization is conducted in 50% formamide containing solutions, for longer probes. All the foregoing hybridizations would be considered to be examples of hybridization performed under conditions of high stringency.

[0101] Following hybridization, the filter is washed for at least 15 minutes in $2\times$ SSC, 0.1% SDS at room temperature or higher, depending on the desired stringency. The filter is then washed with 0.1×SSC, 0.5% SDS at room temperature (again) for 30 minutes to 1 hour. Nucleic acids which have hybridized to the probe are identified by conventional autoradiography and non-radioactive detection methods.

[0102] The above procedure may be modified to identify nucleic acids having decreasing levels of homology to the probe sequence. For example, to obtain nucleic acids of decreasing homology to the detectable probe, less stringent conditions may be used. For example, the hybridization temperature may be decreased in increments of 5° C. from 68° C. to 42° C. in a hybridization buffer having a Na+ concentration of approximately 1 M. Following hybridization, the filter may be washed with 2×SSC, 0.5% SDS at the temperature of hybridization. These conditions are considered to be "moderate stringency" conditions above 50° C. and "low stringency" conditions below 50° C. A specific example of "moderate stringency" hybridization conditions is when the above hybridization is conducted at 55° C. A specific example of "low stringency" hybridization conditions is when the above hybridization is conducted at 45° C.

[0103] Alternatively, the hybridization may be carried out in buffers, such as $6\times$ SSC, containing formamide at a temperature of 42° C. In this case, the concentration of forma-

mide in the hybridization buffer may be reduced in 5% increments from 50% to 0% to identify clones having decreasing levels of homology to the probe. Following hybridization, the filter may be washed with 6×SSC, 0.5% SDS at 50° C. These conditions are considered to be "moderate stringency" conditions above 25% formamide and "low stringency" conditions below 25% formamide. A specific example of "moderate stringency" hybridization conditions is when the above hybridization is conducted at 30% formamide. A specific example of "low stringency" hybridization conditions is when the above hybridization is conducted at 10% formamide. Nucleic acids which have hybridized to the probe are identified by conventional autoradiography and non-radioactive detection methods. Examples of conditions of different stringency are also provided in Table 2.

TABLE 2

	(det		Very High ences sharii	Stringency ng at least 90% identity)		
Hybridization	in	5x	SCC at	65° C.	16 hours	
Wash twice	in	2x	SCC at	room temeprature	15 mnutes each	
Wash twice	in	0.5x	SCC at	65° C.	20 minutes each	
			High Str	ingency		
	(dete	ects seque	ences sharii	ng at least 80% identity)		
Hybridization	in	5x	SCC at	65° C.	16 hours	
Wash twice	in	2x	SCC at	room temeprature	20 mnutes each	
Wash once	in	1x	SCC at	55° Ĉ.	30 minutes each	
			Low Str	ingency		
(detects sequences sharing at least 50% identity)						
Hybridization	in	6x	SCC at	room temperature	16 hours	
Wash twice	in	3x	SCC at	room temeprature	20 minutes each	

[0104] The preceding methods may be used to isolate nucleic acids having at least 97%, at least 95%, at least 90%, at least 85%, at least 80%, or at least 70% sequence identity to a nucleic acid sequence selected from the group consisting of the sequences of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 66, 68, 70, 72, 75, 77, 79, 81, 83, 85, 87 and 89. The isolated nucleic acid may have a coding sequence that is a naturally occurring allelic variant of one of the coding sequences described herein. Such allelic variant may have a substitution, deletion or addition of one or more nucleotides when compared to the nucleic acids of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 66, 68, 70, 72, 75, 77, 79, 81, 83, 85, 87 and 89, or the sequences complementary thereto.

[0105] Additionally, the above procedures may be used to isolate nucleic acids which encode polypeptides having at least 99%, at least 95%, at least 90%, at least 85%, at least 80%, or at least 70% identity to a polypeptide having the sequence of one of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 or fragments comprising at least 50, 75, 100, 150, 200, 300 consecutive amino acids thereof.

[0106] Another aspect of the present invention is an isolated or purified polypeptide comprising the sequence of one of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 or

fragments comprising at least 50, 75, 100, 150, 200 or 300 consecutive amino acids thereof. As discussed herein, such polypeptides may be obtained by inserting a nucleic acid encoding the polypeptide into a vector such that the coding sequence is operably linked to a sequence capable of driving the expression of the encoded polypeptide in a suitable host cell. For example, the expression vector may comprise a promoter, a ribosome binding site for translation initiation and a transcription terminator. The vector may also include appropriate sequences for modulating expression levels, an origin of replication and a selectable marker.

[0107] Promoters suitable for expressing the polypeptide or fragment thereof in bacteria include the *E. coli* lac or trp promoters, the lad promoter, the lacZ promoter, the T3 promoter, the T7 promoter, the gpt promoter, the lambda P_{R}

promoter, the lambda P_L promoter, promoters from operons encoding glycolytic enzymes such as 3-phosphoglycerate kinase (PGK), and the acid phosphatase promoter. Fungal promoters include the a factor promoter. Eukaryotic promoters include the CMV immediate early promoter, the HSV thymidine kinase promoter, heat shock promoters, the early and late SV40 promoter, LTRs from retroviruses, and the mouse metallothionein-I promoter. Other promoters known to control expression of genes in prokaryotic or eukaryotic cells or their viruses may also be used.

[0108] Mammalian expression vectors may also comprise an origin of replication, any necessary ribosome binding sites, a polyadenylation site, splice donors and acceptor sites, transcriptional termination sequences, and 5' flanking nontranscribed sequences. In some embodiments, DNA sequences derived from the SV40 splice and polyadenylation sites may be used to provide the required nontranscribed genetic elements.

[0109] Vectors for expressing the polypeptide or fragment thereof in eukaryotic cells may also contain enhancers to increase expression levels. Enhancers are cis-acting elements of DNA, usually from about 10 to about 300 bp in length that act on a promoter to increase its transcription. Examples include the SV40 enhancer on the late side of the replication origin bp 100 to 270, the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and the adenovirus enhancers.

[0110] In addition, the expression vectors preferably contain one or more selectable marker genes to permit selection of host cells containing the vector. Examples of selectable markers that may be used include genes encoding dihydrofolate reductase or genes conferring neomycin resistance for eukaryotic cell culture, genes conferring tetracycline or ampicillin resistance in E. coli, and the S. cerevisiae TRP1 gene. [0111] The appropriate DNA sequence may be inserted into the vector by a variety of procedures. In general, the DNA sequence is ligated to the desired position in the vector following digestion of the insert and the vector with appropriate restriction endonucleases. Alternatively, appropriate restriction enzyme sites can be engineered into a DNA sequence by PCR. A variety of cloning techniques are disclosed in Ausbel et al. Current Protocols in Molecular Biology, John Wiley 503 Sons, Inc. 1997 and Sambrook et al., Molecular Cloning: A Laboratory Manual 2d Ed., Cold Spring Harbour Laboratory Press, 1989. Such procedures and others are deemed to be within the scope of those skilled in the art.

[0112] The vector may be, for example, in the form of a plasmid, a viral particle, or a phage. Other vectors include derivatives of chromosomal, nonchromosomal and synthetic DNA sequences, viruses, bacterial plasmids, phage DNA, baculovirus, yeast plasmids, vectors derived from combinations of plasmids and phage DNA, viral DNA such as vaccinia, adenovirus, fowl pox virus, and pseudorabies. A variety of cloning and expression vectors for use with prokaryotic and eukaryotic hosts are described by Sambrook et al., Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor, N.Y., (1989), incorporated by reference in its entirety for all purposes.

[0113] Particular bacterial vectors which may be used include the commercially available plasmids comprising genetic elements of the well known cloning vector pBR322 (ATCC 37017), pKK223-3 (Pharmacia Fine Chemicals, Uppsala, Sweden), pGEM1 (Promega Biotec, Madison, Wis., USA) pQE70, pQE60, pQE-9 (Qiagen), pD10, phiX174, pBluescript[™] II KS, pNH8A, pNH16a, pNH18A, pNH46A (Stratagene), ptrc99a, pKK223-3, pKK223-3, pDR540, pRIT5 (Pharmacia), pKK223-8 and pCM7. Particular eukaryotic vectors include pSV2CAT, pOG44, pXT1, pSG (Stratagene), pSVK3, PBPV, PMSG, and PSVL (Pharmacia). However, any other vector may be used as long as it is replicable and stable in the host cell.

[0114] The host cell may be any of the host cells familiar to those skilled in the art, including prokaryotic cells or eukaryotic cells. As representative examples of appropriate hosts, there may be mentioned: bacteria cells, such as E. coli, Streptomyces lividans, Streptomyces griseofuscus, Streptomyces ambofaciens, Rhodococcus, Amycolatopsis, Bacillus subtilis, Salmonella typhimurium and various species within the genera Pseudomonas, Streptomyces, Bacillus, and Staphylococcus, fungal cells, such as yeast, insect cells such as Drosophila S2 and Spodoptera Sf9, animal cells such as CHO, COS or Bowes melanoma, and adenoviruses. The selection of an appropriate host is within the abilities of those skilled in the art, see for example Manual of Industrial Microbiology and Biotechnology, 2nd Edition, ASM Press, Washington D.C., incorporated by reference in its entirety, and more particularly Sections IV, V and VII.

[0115] The vector may be introduced into the host cells using any of a variety of techniques, including electroporation transformation, transfection, transduction, viral infection, gene guns, or Ti-mediated gene transfer. Where appropriate, the engineered host cells can be cultured in conventional nutrient media modified as appropriate for activating promoters, selecting transformants or amplifying the

genes of the present invention. Following transformation of a suitable host strain and growth of the host strain to an appropriate cell density, the selected promoter may be induced by appropriate means (e.g., temperature shift or chemical induction) and the cells may be cultured for an additional period to allow them to produce the desired polypeptide or fragment thereof.

[0116] Cells are typically harvested by centrifugation, disrupted by physical or chemical means, and the resulting crude extract is retained for further purification. Microbial cells employed for expression of proteins can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing agents. Such methods are well known to those skilled in the art. The expressed polypeptide or fragment thereof can be recovered and purified from recombinant cell cultures by methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography. Protein refolding steps can be used, as necessary, in completing configuration of the polypeptide. If desired, high performance liquid chromatography (HPLC) can be employed for final purification steps.

[0117] Various mammalian cell culture systems can also be employed to express recombinant protein. Examples of mammalian expression systems include the COS-7 lines of monkey kidney fibroblasts (described by Gluzman, Cell, 23:175 (1981)), and other cell lines capable of expressing proteins from a compatible vector, such as the C127, 3T3, CHO, HeLa and BHK cell lines. The constructs in host cells can be used in a conventional manner to produce the gene product encoded by the recombinant sequence. Polypeptides of the invention may or may not also include an initial methionine amino acid residue.

[0118] Alternatively, the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 or fragments comprising at least 50, 75, 100, 150, 200 or 300 consecutive amino acids thereof can be synthetically produced by conventional peptide synthesizers. In other embodiments, fragments or portions of the polynucleotides may be employed for producing the corresponding full-length polypeptide by peptide synthesis; therefore, the fragments may be employed as intermediates for producing the full-length polypeptides.

[0119] Cell-free translation systems can also be employed to produce one of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 or fragments comprising at least 50, 75, 100, 150, 200 or 300 consecutive amino acids thereof using mRNAs transcribed from a DNA construct comprising a promoter operably linked to a nucleic acid encoding the polypeptide or fragment thereof. In some embodiments, the DNA construct may be linearized prior to conducting an in vitro transcription reaction. The transcribed mRNA is then incubated with an appropriate cell-free translation extract, such as a rabbit reticulocyte extract, to produce the desired polypeptide or fragment thereof.

[0120] The present invention also relates to variants of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54,

56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 or fragments comprising at least 50, 75, 100, 150, 200 or 300 consecutive amino acids thereof. The term "variant" includes derivatives or analogs of these polypeptides. In particular, the variants may differ in amino acid sequence from the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 by one or more substitutions, additions, deletions, fusions and truncations, which may be present in any combination.

[0121] The variants may be naturally occurring or created in vitro. In particular, such variants may be created using genetic engineering techniques such as site directed mutagenesis, random chemical mutagenesis, exonuclease III deletion procedures, and standard cloning techniques. Alternatively, such variants, fragments, analogs, or derivatives may be created using chemical synthesis or modification procedures.

[0122] Other methods of making variants are also familiar to those skilled in the art. These include procedures in which nucleic acid sequences obtained from natural isolates are modified to generate nucleic acids that encode polypeptides having characteristics which enhance their value in industrial or laboratory applications. In such procedures, a large number of variant sequences having one or more nucleotide differences with respect to the sequence obtained from the natural isolate are generated and characterized. Preferably, these nucleotide differences result in amino acid changes with respect to the polypeptides encoded by the nucleic acids from the natural isolates.

[0123] The variants of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 may be variants in which one or more of the amino acid residues of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 are substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue) and such substituted amino acid residue may or may not be one encoded by the genetic code.

[0124] Conservative substitutions are those that substitute a given amino acid in a polypeptide by another amino acid of like characteristics. Typically seen as conservative substitutions are the following replacements: replacements of an aliphatic amino acid such as Ala, Val, Leu and Ile with another aliphatic amino acid; replacement of a Ser with a Thr or vice versa; replacement of an acidic residue such as Asp or Glu with another acidic residue; replacement of a residue bearing an amide group, such as Asn or Gln, with another residue bearing an amide group; exchange of a basic residue such as Lys or Arg with another basic residue; and replacement of an aromatic residue such as Phe or Tyr with another aromatic residue.

[0125] Other variants are those in which one or more of the amino acid residues of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 include a substituent group. Still other variants are those in which the polypeptide is associated with another compound, such as a compound to increase the half-life of the polypeptide (for example, polyethylene glycol). Additional variants are those in which addi-

tional amino acids are fused to the polypeptide, such as leader sequence, a secretory sequence, a proprotein sequence or a sequence that facilitates purification, enrichment, or stabilization of the polypeptide.

[0126] In some embodiments, the fragments, derivatives and analogs retain the same biological function or activity as the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88. In other embodiments, the fragment, derivative or analogue includes a fused heterologous sequence that facilitates purification, enrichment, detection, stabilization or secretion of the polypeptide that can be enzymatically cleaved, in whole or in part, away from the fragment, derivative tive or analogue.

[0127] Another aspect of the present invention are polypeptides or fragments thereof which have at least 70%, at least 80%, at least 85%, at least 90%, or more than 95% identity to one of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 or a fragment comprising at least 50, 75, 100, 150, 200 or 300 consecutive amino acids thereof. It will be appreciated that amino acid "substantially identity" includes conservative substitutions such as those described above.

[0128] The polypeptides or fragments having homology to one of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 or a fragment comprising at least 50, 75, 100, 150, 200 or 300 consecutive amino acids thereof may be obtained by isolating the nucleic acids encoding them using the techniques described above.

[0129] Alternatively, the homologous polypeptides or fragments may be obtained through biochemical enrichment or purification procedures. The sequence of potentially homologous polypeptides or fragments may be determined by proteolytic digestion, gel electrophoresis and/or microsequencing. The sequence of the prospective homologous polypeptide or fragment can be compared to one of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 or a fragment comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids thereof.

[0130] The polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88 or fragments, derivatives or analogs thereof comprising at least 40, 50, 75, 100, 150, 200 or 300 consecutive amino acids thereof invention may be used in a variety of applications. For example, the polypeptides or fragments, derivatives or analogs thereof may be used to catalyze biochemical reactions as described elsewhere in the specification.

EXAMPLES

[0131] Unless otherwise indicated, all numbers expressing quantities of ingredients, properties such as molecular weight, reaction conditions, IC_{50} and so forth used in the specification and claims are to be understood as being modified in all instances by the term "about". Accordingly, unless indicated to the contrary, the numerical parameters set forth in the present specification and attached claims are approxima-

tions. At the very least, and not as an attempt to limit the application of the doctrine of equivalents to the scope of the claims, each numerical parameter should at least be construed in light of the number of significant figures and by applying ordinary rounding techniques. Notwithstanding that the numerical ranges and parameters setting forth the broad scope of the invention are approximations, the numerical values set in the examples, Tables and Figures are reported as precisely as possible. Any numerical values may inherently contain certain errors resulting from variations in experiments, testing measurements, statistical analyses and such.

[0132] Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, suitable methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

Example 1

Preparation of Production Culture

[0133] Unless otherwise noted, all reagents were purchased from Sigma Chemical Co. (St. Louis, Mo.), (Aldrich). Micromonospora spp. (deposit accession number IDAC 070303-01) was maintained on agar plates of ISP2 agar (Difco Laboratories, Detroit, Mich.). An inoculum for the production phase was prepared by transferring the surface growth of the Micromonospora spp. from the agar plates to 125-mL flasks containing 25 mL of sterile medium comprised of 24 g potato dextrin, 3 g beef extract, 5 g Bactocasitone, 5 g glucose, 5 g yeast extract, and 4 g CaCO₃ made up to one liter with distilled water (pH 7.0). The culture was incubated at about 28° C. for approximately 60 hours on a rotary shaker set at 250 rpm. Following incubation, 10 mL of culture was transferred to a 2 L baffled flask containing 500 mL of sterile production medium containing 20 g/L potato dextrin, 20 g/L glycerol, 10 g/L Fish meal, 5 g/L Bactopeptone, 2 g/L CaCO₃, and 2 g/L (NH₄)₂SO₄, pH 7.0. Fermentation broth was prepared by incubating the production culture at 28° C. in a rotary shaker set at 250 rpm for one week.

Example 2

Isolation

[0134] 500 mL ethyl acetate was added to 500 mL of fermentation broth prepared as described in Example 1 above. The mixture was agitated for 30 minutes on an orbital shaker at 200 rpm to create an emulsion. The phases were separated by centrifugation and decantation. Between 4 and 5 g of anhydrous $MgSO_4$ was added to the organic phase, which was then filtered and the solvents removed in vacuo.

[0135] An ethyl acetate extract from 2 L fermentation was mixed with HP-20 resin (100 mL; Mitsubishi Casei Corp., Tokyo, Japan) in water (300 mL). Ethyl acetate was removed in vacuo, the resin was filtered on a Büchner funnel and the filtrate was discarded. The adsorbed HP-20 resin was then

washed successively with 2×125 mL of 50% acetonitrile in water, 2×125 mL of 75% acetonitrile in water and 2×125 mL of acetonitrile.

[0136] Fractions containing ECO-04601 were evaporated to dryness and 100 mg was digested in the 5 mL of the upper phase of a mixture prepared from chloroform, cyclohexane, methanol, and water in the ratios, by volume, of 5:2:10:5. The sample was subjected to centrifugal partition chromatography using a High Speed Countercurrent (HSCC) system (Kromaton Technologies, Angers, France) fitted with a 200 mL cartridge and prepacked with the upper phase of this two-phase system. The HSCC was run with the lower phase mobile and ECO-04601 was eluted at approximately one-half column volume. Fractions were collected and ECO-04601 was detected by TLC of aliquots of the fractions on commercial Kieselgel $60\mathrm{F}_{254}$ plates. Compound could be visualized by inspection of dried plates under UV light or by spraying the plates with a spray containing vanillin (0.75%) and concentrated sulfuric acid (1.5%, v/v) in ethanol and subsequently heating the plate. Fractions contained substantially pure ECO-04601, although highly colored. A buff-colored sample could be obtained by chromatography on HPLC as follows.

[0137] 6 mg of sample was dissolved in acetonitrile and injected onto a preparative HPLC column (XTerra ODS (10 μ m), 19×150 mm, Waters Co., Milford, Mass.), with a 9 mL/min flow rate and UV peak detection at 300 nm. The column was eluted with acetonitrile/buffer (20 mM of NH₄HCO₃) according to the following gradient shown in Table 3.

TABLE 3

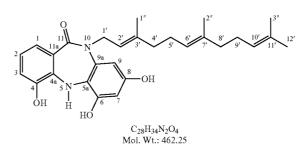
Time (min)	Water (%)	Acetonitrile (%)
0	70	30
10	5	95
15	5	95
20	70	30

[0138] Fractions containing ECO-04061 eluted at approximately 11:0 min and were combined, concentrated and lyophilized to give a yield of 3.8 mg compound.

Example 3

Elucidation of the Structure of ECO-04601

[0139]



[0140] The structure of ECO-04601 above was derived from spectroscopic data, including mass, UV, and NMR spectroscopy. Mass was determined by electrospray mass spectrometry to be 462.6, UVmax 230 nm with a shoulder at 290

nm. NMR data were collected dissolved in MeOH-d4 including proton, and multidimensional pulse sequences. Proton and carbon NMR data are detailed in Table 4 below.

TABLE 4

¹ H and ¹³ C NM	IR (δ _H , ppm) of	ECO-04601 in	n MeOH-D ₄
Assignment	$^{1}\mathrm{H}$	¹³ C	Group
1	7.15	122.3	СН
2 3	6.74	121.0	CH
	6.83	116.9	CH
4		146.0	C—OH
4a		142.0	С
5a	_	126.0	С
6		148.2	C—OH
7	6.20	100.0	CH
8		153.0	C—OH
9	6.25	101.0	CH
9a		135.0	С
11		170.0	C(O)
11a		125.0	С
1'	4.52	48.7	CH ₂
2'	5.35	121.1	CH
3'		138.5	С
4'	2.03	39.5	CH ₂
5'	2.08	26.7	CH ₂
6'	5.09	124.1	CH
7'		135.0	С
8'	1.95	39.6	CH ₂
9'	2.02	26.3	CH ₂
10'	5.06	124.4	CH
11'	_	130.9	С
12'	1.64	24.8	CH ₃
1"	1.72	15.5	CH3
2"	1.59	14.9	CH3
3"	1.55	16.5	CH ₃

[0141] A number of cross peaks in the 2D spectra of ECO-04601 are key in the structural determination. For example, the farnesyl chain is placed on the amide nitrogen by a strong cross peak between the proton signal of the terminal methylene of that chain at 4.52 ppm and the amide carbonyl carbon at 170 ppm in the gHMBC experiment. This conclusion is confirmed by a cross peak in the NOESY spectrum between the same methylene signals at 4.52 ppm and the aromatic proton signal at 6.25 ppm from one of the two protons of the tetra substituted benzenoid ring.

[0142] Based on the mass, UV and NMR spectroscopy data, the structure of the compound was determined to be the structure of ECO-04601.

Example 4

In Vivo Efficacy in a Glioma Model

[0143] The aim of this study was to test whether ECO-04601 when administered by the i.p. route prevents or delays tumor growth in C6 glioblastoma cell-bearing mice, and to determine an effective dosage regimen.

[0144] Animals: A total of 60 six-week-old female mice (*Mus musculus* nude mice), ranging between 18 to 25 g in weight, were observed for 7 days before treatment. Animal experiments were performed according to ethical guidelines of animal experimentation (*Charté du comite d'éthique du CNRS, juillet* 2003) and the English guidelines for the welfare of animals in experimental neoplasia (WORKMAN, P., TWENTYMAN, P., BALKWILL, F., et al. (1998). United Kingdom Coordinating Committee on Cancer Research (UKCCCR) Guidelines for the welfare of animals in experi-

mental neoplasia (Second Edition, July 1997; British Journal of Cancer, 77:1-10). Any dead or apparently sick mice were promptly removed and replaced with healthy mice. Sick mice were euthanized upon removal from the cage. Animals were maintained in rooms under controlled conditions of temperature (23+2° C.), humidity (45±5%), photoperiodicity (12 hrs light/12 hrs dark) and air exchange. Animals were housed in polycarbonate cages (5/single cage) that were equipped to provide food and water. Animal bedding consisted of sterile wood shavings that were replaced every other day. Food was provided ad libitum, being placed in the metal lid on the top of the cage. Autoclaved tap water was provided ad libitum. Water bottles were equipped with rubber stoppers and sipper tubes. Water bottles were cleaned, sterilized and replaced once a week. Two different numbers engraved on two earrings identified the animals. Each cage was labeled with a specific code.

[0145] Tumor Cell Line: The C6 cell line was cloned from a rat glial tumor induced by N-nitrosomethyurea (NMU) according to Premont et al. (Premont J, Benda P, Jard S., [3H] norepinephrine binding by rat glial cells in culture. Lack of correlation between binding and adenylate cyclase activation. Biochim Biophys Acta. 1975 Feb. 13; 381(2):368-76.) after series of alternate culture and animal passages. Cells were grown as adherent monolayers at 37° C. in a humidified atmosphere (5% CO₂, 95% air). The culture medium was DMEM supplemented with 2 mM L-glutamine and 10% fetal bovine serum. For experimental use, tumor cells were detached from the culture flask by a 10 min treatment with trypsin-versen. The cells were counted in a hemocytometer and their viability assessed by 0.25% trypan blue exclusion. [0146] Preparation of the Test Article: for the Test Article, the Following Procedure was followed for reconstitution (performed immediately preceding injection). The vehicle consisted of a mixture of benzyl alcohol (1.5%), ethanol (8.5%), propylene glycol (27%), PEG 400 (27%), dimethylacetamide (6%) and water (30%). The vehicle solution was first vortexed in order to obtain a homogeneous liquid. 0.6 mL of the vortexed vehicle solution was added to each vial containing the test article (ECO-04601). Vials were mixed thoroughly by vortexing for 1 minute and inverted and shaken vigorously. Vials were mixed again prior to injection into each animal.

[0147] Animal Inoculation with tumor cells: Experiment started at day 0 (D_0). On D_0 , mice received a superficial intramuscular injection of C6 tumor cells (5×10^5 cells) in 0.1 mL of DMEM complete medium into the upper right posterior leg.

[0148] Treatment Regimen and Results:

[0149] First series of experiments: In a first series of experiments, treatment started 24 hrs following inoculation of C6 cells. On the day of the treatment, each mouse was slowly injected with $100 \,\mu$ L of test or control articles by the i.p. route. For all groups, treatment was performed until the tumor volume of the saline-treated mice (group 1) reached approximately 3 cm³ (around day 16). Mice of group 1 were treated daily with a saline isosmotic solution for 16 days. Mice of group 2 were treated daily with the vehicle solution for 16 days. Mice of group 3 were treated daily with 10 mg/kg of ECO-04601 for 16 days. Mice of group 4 were treated every two days with 30 mg/kg of ECO-04601 and received 8 treatments. Mice of group 5 were treated every three days with 30 mg/kg of ECO-04601 and received 6 treatments. Measurement of tumor volume started as soon as tumors became

palpable (>100 mm³; day 11 post-inoculation) and was evaluated every second day until the end of the treatment using callipers. As shown in Table 5 and FIG. 1, the mean value of the tumor volume of all ECO-4061-treated groups (6 mice/ group) was significantly reduced as demonstrated by the oneway analysis of variance (Anova) test followed by the nonparametric Dunnett's multiple comparison test comparing treated groups to the saline group. An asterisk in the P value column of Table 5 indicates a statistically significant value, while "ns" signifies not significant.

TABLE 5

ECO-04601 in vivo antitumor efficacy against C6 glioblastoma							
Treatment	Treatment regimen	Tumor volume (mm ³) (mean ± SEM)	% Inhibition	P value			
Saline	Q1 × 16	3,004.1 ± 249.64	_	_			
Vehicle solution	Q1 × 16	$2,162.0 \pm 350.0$	28.0%	>0.05 ns			
ECO-04601 (10 mg/kg)	Q1 × 16	1,220.4 ± 283.46	59.4%	<0.01 *			
ECO-04601 (30 mg/kg)	Q2 × 8	1,236.9 ± 233.99	58.8%	<0.01 *			
(30 mg/kg) ECO-04601 (30 mg/kg)	Q3 × 6	1,184.1 ± 221.45	60.6%	<0.01 *			

[0150] Second series experiments: In a second series of experiments, treatment started at day 10 following inoculation of C6 cells when tumors became palpable (around 100 to 200 mm³). Treatment was repeated daily for 5 consecutive days. On the day of the treatment, each mouse was slowly injected with 100 µL of ECO-04601 by i.p. route. Mice of group 1 were treated daily with saline isosmotic solution. Mice of group 2 were treated daily with the vehicle solution. Mice of group 3 were treated daily with 20 mg/kg of ECO-04601. Mice of group 4 were treated daily with 30 mg/kg of ECO-04601. Mice were treated until the tumor volume of the saline-treated control mice (group 1) reached around 4 cm^3 . Tumor volume was measured every second day until the end of the treatment using callipers. As shown in Table 6 and FIG. 2, the mean value of the tumor volume of all ECO-04601treated groups (6 mice/group) was significantly reduced as demonstrated by the one-way analysis of variance (Anova) test followed by the non-parametric Dunnett's multiple comparison test comparing treated groups to the saline group. An asterisk in the P value column of Table 6 indicates a statistically significant value, while "ns" signifies not significant.

[0151] Histological analysis of tumor sections showed pronounced morphological changes between tumors from ECO-04601-treated mice and those from mice in the control groups. In tumors from mice treated with ECO-04601 (20-30 mg/kg), cell density was decreased and the nuclei of remaining tumor cells appeared larger and pycnotic while no such changes were observed for tumors from vehicle-treated mice (FIG. 3).

TABLE 6

ECO-04601 in vivo antitumor efficacy against C6 glioblastoma							
Treatment	Treatment regimen	Tumor volume (mm ³) (mean ± SEM)	% Inhibition	P value			
Saline Vehicle solution	Q1 × 5 Q1 × 5	4,363.1 ± 614.31 3,205.0 ± 632.37	26.5%				

TABLE 6-continued

ECO-046	501 in vivo a	ntitumor efficacy ag	ainst C6 gliobla	istoma
Treatment	Treatment regimen	Tumor volume (mm ³) (mean ± SEM)	% Inhibition	P value
ECO-04601 (20 mg/kg)	Q1 × 5	1,721.5 ± 374.79	60.5%	<0.01 *
(20 mg/kg) ECO-04601 (30 mg/kg)	Q1 × 5	1,131.6 ± 525.21	74.1%	<0.01 *

Example 5

Genes and Proteins for the Production of Farnesyl Dibenzodiazepinones

[0152] *Micromonospora* sp. strain 046-ECO11 is a representative microorganism useful in the production of the compound of the invention. Strain 046-ECO11 has been deposited with the International Depositary Authority of Canada (IDAC), Bureau of Microbiology, Health Canada, 1015 Arlington Street, Winnipeg, Manitoba, Canada R3E 3R2 on Mar. 7, 2003 and was assigned IDAC accession no. 070303-01. The biosynthetic locus for the production of ECO-04601 was identified in the genome of *Micromonospora* sp. strain 046-ECO11 using the genome scanning method described in U.S. Ser. No. 10/232,370, CA 2, 352, 451 and Zazopoulos et. al., *Nature Biotechnol.*, 21, 187-190 (2003).

[0153] The biosynthetic locus spans approximately 52,400 base pairs of DNA and encodes 43 proteins. More than 10 kilobases of DNA sequence were analyzed on each side of the locus and these regions were deemed to contain primary genes or genes unrelated to the synthesis of ECO-04601. As illustrated in FIG. 4, the locus is contained within three sequences of contiguous base pairs, namely Contig 1 having the 36,602 contiguous base pairs of SEQ ID NO: 1 and comprising ORFs 1 to 31 (SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61 and 63), Contig 2 having the 5,960 contiguous base pairs of SEQ ID NO: 64 and comprising ORFs 32 to 35 (SEQ ID NOS: 66, 68, 70 and 72), and Contig 3 having the 9,762 base pairs of SEQ ID NO: 73 and comprising ORFs 36 to 43 (SEQ ID NOS: 75, 77, 79, 81, 83, 85, 87 and 89). The order, relative position and orientation of the 43 open reading frames representing the proteins of the biosynthetic locus are illustrated schematically in FIG. 4. The top line in FIG. 4 provides a scale in base pairs. The gray bars depict the three DNA contigs (SEQ ID NOS: 1, 64 and 73) that cover the locus. The empty arrows represent the 43 open reading frames of this biosynthetic locus. The black arrows represent the two deposited cosmid clones covering the locus. [0154] The biosynthetic locus will be further understood with reference to the sequence listing which provides contiguous nucleotide sequences and deduced amino acid sequences of the locus from Micromonospora sp. strain 046-ECO11. The contiguous nucleotide sequences are arranged such that, as found within the biosynthetic locus, Contig 1 (SEQ ID NO: 1) is adjacent to the 5' end of Contig 2 (SEQ ID NO: 64), which in turn is adjacent to Contig 3 (SEQ ID NO: 73). The ORFs illustrated in FIG. 4 and provided in the sequence listing represent open reading frames deduced from the nucleotide sequences of Contigs 1, 2 and 3 (SEQ ID NOS: 1,64 and 73). Referring to the Sequence Listing, ORF 1 (SEQ ID NO: 3) is the polynucleotide drawn from residues 2139 to 424 of SEQ ID NO: 1, and SEQ ID NO: 2 represents that polypeptide deduced from SEQ ID NO: 3. ORF 2 (SEQ ID NO: 5) is the polynucleotide drawn from residues 2890 to 4959 of SEO ID NO: 1, and SEO ID NO: 4 represents the polypeptide deduced from SEQ ID NO: 5. ORF 3 (SEQ ID NO: 7) is the polynucleotide drawn from residues 7701 to 5014 of SEQ ID NO: 1, and SEQ ID NO: 6 represents the polypeptide deduced from SEQ ID NO: 7. ORF 4 (SEQ ID NO: 9) is the polynucleotide drawn from residues 8104 to 9192 of SEQ ID NO: 1, and SEQ ID NO: 8 represents the polypeptide deduced from SEQ ID NO: 9. ORF 5 (SEQ ID NO: 11) is the polynucleotide drawn from residues 9192 to 10256 of SEQ ID NO: 1, and SEQ ID NO: 10 represents the polypeptide deduced from SEO ID NO: 11. ORF 6 (SEO ID NO: 13) is the polynucleotide drawn from residues 10246 to 11286 of SEQ ID NO: 1, and SEQ ID NO: 12 represents the polypeptide deduced from SEQ ID NO: 13. ORF 7 (SEQ ID NO: 15) is the polynucleotide drawn from residues 11283 to 12392 of SEQ ID NO: 1, and SEQ ID NO: 14 represents the polypeptide deduced from SEQ ID NO: 15. ORF 8 (SEQ ID NO: 17) is the polynucleotide drawn from residues 12389 to 13471 of SEQ ID NO: 1, and SEQ ID NO: 16 represents the polypeptide deduced from SEQ ID NO: 17. ORF 9 (SEQ ID NO: 19) is the polynucleotide drawn from residues 13468 to 14523 of SEQ ID NO: 1, and SEQ ID NO: 18 represents the polypeptide deduced from SEQ ID NO: 19. ORF 10 (SEQ ID NO: 21) is the polynucleotide drawn from residues 14526 to 15701 of SEQ ID NO: 1, and SEQ ID NO: 20 represents the polypeptide deduced from SEQ ID NO: 21. ORF 11 (SEQ ID NO: 23) is the polynucleotide drawn from residues 15770 to 16642 of SEQ ID NO: 1, and SEQ ID NO: 22 represents the polypeptide deduced from SEQ ID NO: 23. ORF 12 (SEQ ID NO: 25) is the polynucleotide drawn from residues 16756 to 17868 of SEQ ID NO: 1, and SEQ ID NO: 24 represents the polypeptide deduced from SEQ ID NO: 25. ORF 13 (SEQ ID NO: 27) is the polynucleotide drawn from residues 17865 to 18527 of SEQ ID NO: 1, and SEQ ID NO: 26 represents the polypeptide deduced from SEQ ID NO: 27. ORF 14 (SEQ ID NO: 29) is the polynucleotide drawn from residues 18724 to 19119 of SEQ ID NO: 1, and SEQ ID NO: 28 represents the polypeptide deduced from SEQ ID NO: 29. ORF 15 (SEQ ID NO: 31) is the polynucleotide drawn from residues 19175 to 19639 of SEQ ID NO: 1, and SEQ ID NO: 30 represents the polypeptide deduced from SEQ ID NO: 31. ORF 16 (SEQ ID NO: 33) is the polynucleotide drawn from residues 19636 to 21621 of SEQ ID NO: 1, and SEQ ID NO: 32 represents the polypeptide deduced from SEQ ID NO: 33. ORF 17 (SEQ ID NO: 35) is the polynucleotide drawn from residues 21632 to 22021 of SEQ ID NO: 1, and SEQ ID NO: 34 represents the polypeptide deduced from SEQ ID NO: 35. ORF 18 (SEQ ID NO: 37) is the polynucleotide drawn from residues 22658 to 22122 of SEQ ID NO: 1, and SEQ ID NO: 36 represents the polypeptide deduced from SEQ ID NO: 37. ORF 19 (SEQ ID NO: 39) is the polynucleotide drawn from residues 24665 to 22680 of SEQ ID NO: 1, and SEQ ID NO: 38 represents the polypeptide deduced from SEQ ID NO: 39. ORF 20 (SEQ ID NO: 41) is the polynucleotide drawn from residues 24880 to 26163 of SEQ ID NO: 1, and SEQ ID NO: 40 represents the polypeptide deduced from SEQ ID NO: 41. ORF 21 (SEQ ID NO: 43) is the polynucleotide drawn from residues 26179 to 27003 of SEQ ID NO: 1, and SEQ ID NO: 42 represents the polypeptide deduced from SEQ ID NO: 43. ORF 22 (SEQ ID NO: 45) is the polynucleotide drawn from residues 27035 to 28138 of SEQ ID NO: 1, and SEQ ID NO: 44 represents the polypeptide deduced from SEQ ID NO: 45. ORF 23 (SEQ ID NO: 47) is the polynucleotide drawn from residues 28164 to 28925 of SEQ ID NO: 1, and SEQ ID NO: 46 represents the polypeptide deduced from SEQ ID NO: 47. ORF 24 (SEQ ID NO: 49) is the polynucleotide drawn from residues 28922 to 30238 of SEO ID NO: 1, and SEO ID NO: 48 represents the polypeptide deduced from SEQ ID NO: 49. ORF 25 (SEQ ID NO: 51) is the polynucleotide drawn from residues 30249 to 31439 of SEQ ID NO: 1, and SEQ ID NO: 50 represents the polypeptide deduced from SEQ ID NO: 51. ORF 26 (SEQ ID NO: 53) is the polynucleotide drawn from residues 31439 to 32224 of SEQ ID NO: 1, and SEQ ID NO: 52 represents the polypeptide deduced from SEQ ID NO: 53. ORF 27 (SEQ ID NO: 55) is the polynucleotide drawn from residues 32257 to 32931 of SEQ ID NO: 1, and SEQ ID NO: 54 represents the polypeptide deduced from SEQ ID NO: 55. ORF 28 (SEQ ID NO: 57) is the polynucleotide drawn from residues 32943 to 33644 of SEQ ID NO: 1, and SEQ ID NO: 56 represents the polypeptide deduced from SEQ ID NO: 57. ORF 29 (SEQ ID NO: 59) is the polynucleotide drawn from residues 34377 to 33637 of SEQ ID NO: 1, and SEQ ID NO: 58 represents the polypeptide deduced from SEQ ID NO: 59. ORF 30 (SEQ ID NO: 61) is the polynucleotide drawn from residues 34572 to 34907 of SEQ ID NO: 1, and SEQ ID NO: 60 represents the polypeptide deduced from SEQ ID NO: 61. ORF 31 (SEQ ID NO: 63) is the polynucleotide drawn from residues 34904 to 36583 of SEQ ID NO: 1, and SEQ ID NO: 62 represents the polypeptide deduced from SEQ ID NO: 63. ORF 32 (SEQ ID NO: 66) is the polynucleotide drawn from residues 23 to 1621 of SEQ ID NO: 64, and SEQ ID NO: 65 represents the polypeptide deduced from SEQ ID NO: 66. ORF 33 (SEQ ID NO: 68) is the polynucleotide drawn from residues 1702 to 2973 of SEQ ID NO: 64, and SEQ ID NO: 67 represents the polypeptide deduced from SEQ ID NO: 68. ORF 34 (SEQ ID NO: 70) is the polynucleotide drawn from residues 3248 to 4270 of SEQ ID NO: 64, and SEQ ID NO: 69 represents the polypeptide deduced from SEQ ID NO: 70. ORF 35 (SEQ ID NO: 72) is the polynucleotide drawn from residues 4452 to 5933 of SEQ ID NO: 64, and SEQ ID NO: 71 represents the polypeptide deduced from SEQ ID NO: 72. ORF 36 (SEQ ID NO: 75) is the polynucleotide drawn from residues 30 to 398 of SEQ ID NO: 73, and SEQ ID NO: 74 represents the polypeptide deduced from SEQ ID NO: 75. ORF 37 (SEQ ID NO: 77) is the polynucleotide drawn from residues 395 to 1372 of SEQ ID NO: 73, and SEQ ID NO: 76 represents the polypeptide deduced from SEQ ID NO: 77. ORF 38 (SEQ ID NO: 79) is the polynucleotide drawn from residues 3388 to 1397 of SEQ ID NO: 73, and SEQ ID NO: 78 represents the polypeptide deduced from SEQ ID NO: 79. ORF 39 (SEQ ID NO: 81) is the polynucleotide drawn from residues 3565 to 5286 of SEQ ID NO: 73, and SEQ ID NO: 80 represents the polypeptide deduced from SEQ ID NO: 81. ORF 40 (SEQ ID NO: 83) is the polynucleotide drawn from residues 5283 to 7073 of SEQ ID NO: 73, and SEQ ID NO: 82 represents the polypeptide deduced from SEQ ID NO: 83. ORF 41 (SEQ ID NO: 85) is the polynucleotide drawn from residues 7108 to 8631 of SEQ ID NO: 73, and SEQ ID NO: 84 represents the polypeptide deduced from SEQ ID NO: 85. ORF 42 (SEQ ID NO: 87) is the polynucleotide drawn from residues 9371 to 8673 of SEQ ID NO: 73, and SEQ ID NO: 86 represents the polypeptide deduced from SEQ ID NO: 87. ORF 43 (SEQ ID NO: 89) is the polynucleotide drawn from residues 9762 to 9364 of SEQ ID NO: 73, and SEQ ID NO: 88 represents the polypeptide deduced from SEQ ID NO: 89.

[0155] Some open reading frames provided in the Sequence Listing, namely ORF 2 (SEQ ID NO: 5), ORF 5 (SEQ ID NO: 11), ORF 12 (SEQ ID NO: 25), ORF 13 (SEQ ID NO: 27), ORF 15 (SEQ ID NO: 31), ORF 17 (SEQ ID NO: 35), ORF 19 (SEQ ID NO: 39), ORF 20 (SEQ ID NO: 41), ORF 22 (SEQ ID NO: 45), ORF 24 (SEQ ID NO: 49), ORF 26 (SEQ ID NO: 53) and ORF 27 (SEQ ID NO: 55) initiate with non-standard initiation codons (eg. GTG-Valine, or CTG-Leucine) rather than standard initiation codon ATG methionine. All ORFs are listed with the appropriate M, V or L amino acids at the amino-terminal position to indicate the specificity of the first codon of the ORF. It is expected, however, that in all cases the biosynthesized protein will contain a methionine residue, and more specifically a formylmethionine residue, at the amino terminal position, in keeping with the widely accepted principle that protein synthesis in bacteria initiate with methionine (formylmethionine) even when the encoding gene specifies a non-standard initiation codon (e.g. Stryer BioChemistry 3rd edition, 1998, W.H. Freeman and Co., New York, pp. 752-754).

[0156] ORF 32 (SEQ ID NO: 65) is incomplete and contains a truncation of 10 to 20 amino acids from its carboxy terminus. This is due to incomplete sequence information between Contigs 2 and 3 (SEQ ID NOS: 64 and 73, respectively).

[0157] Deposits of *E. coli* DH10B vectors, each harbouring a cosmid clone (designated in FIG. **4** as 046KM and 046KQ respectively) of a partial biosynthetic locus for the farnesyl dibenzodiazepinone from *Micromonospora* sp. strain 046-ECO11 and together spanning the full biosynthetic locus for production of ECO-04601 have been deposited with the International Depositary Authority of Canada, Bureau of Microbiology, Health Canada, 1015 Arlington Street, Winnipeg, Manitoba, Canada R3E 3R2 on Feb. 25, 2003. The cosmid clone designated 046KM was assigned deposit accession numbers IDAC 250203-06, and the cosmid clone designated 046KQ was assigned deposit accession numbers IDAC 250203-07. Cosmid 046KM covers residue 1 to residue 32,250 of Contig 1 (SEQ ID NO: 1). Cosmid 046KQ covers residue 21,700 of Contig 1 (SEQ ID NO: 1) to residue 9,762 of Contig 3 (SEQ ID NO: 73). The sequence of the polynucleotides comprised in the deposited strains, as well as the amino acid sequence of any polypeptide encoded thereby are controlling in the event of any conflict with any description of sequences herein.

[0158] The deposit of the deposited strains has been made under the terms of the Budapest Treaty on the International Recognition of the Deposit of Micro-organisms for Purposes of Patent Procedure. The deposited strains will be irrevocably and without restriction or condition released to the public upon the issuance of a patent. The deposited strains are provided merely as convenience to those skilled in the art and are not an admission that a deposit is required for enablement, such as that required under 35 U.S.C. §112. A license may be required to make, use or sell the deposited strains, and compounds derived therefrom, and no such license is hereby granted.

[0159] In order to identify the function of the proteins coded by the genes forming the biosynthetic locus for the production of ECO-04601 the gene products of ORFs 1 to 43, namely SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 71, 74, 76, 78, 80, 82, 84, 86 and 88 were compared, using the BLASTP version 2.2.10 algorithm with the default parameters, to sequences in the National Center for Biotechnology Information (NCBI) nonredundant protein database and the DECIPHER® database of microbial genes, pathways and natural products (Ecopia BioSciences Inc. St.-Laurent, QC, Canada).

[0160] The accession numbers of the top GenBankTM hits of this BLAST analysis are presented in Table 7 along with the corresponding E values. The E value relates the expected number of chance alignments with an alignment score at least equal to the observed alignment score. An E value of 0.00 indicates a perfect homolog. The E values are calculated as described in Altschul et al. *J. Mol. Biol.*, 215, 403-410 (1990). The E value assists in the determination of whether two sequences display sufficient similarity to justify an inference of homology.

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TA	BL	Æ	7

	Sequence comparison and ORF correlation						
ORF	SEQ ID	Family	# aa	GenBank homology	Probability	% Identity (% Similarity)	Proposed function of GenBank match
1	2	ABCC	571	NP_736627.1	1E-107	45% (56%)	ABC transporter Corynebacterium efficiens
				590aa NP_600638.1 510aa	5E-80	37% (52%)	ABC transporter Corynebacterium efficiens
				NP_600638.1	3E-12	30% (43%)	ABC transporter Corynebacterium efficiens
2	4	RECH	689	510aa CAC93719.1 923aa	3E-17	36% (55%)	regulator[Lechevalieria aerocolonigenes]
				BAC55205.1 943aa	3E-12	30% (48%)	transcriptional activator [Streptomyces sp.
				945aa NP_631154.1 932aa	3E-07	46% (63%)	regulator. [Streptomyces coelicolor A3(2)
3	6	REGD	895	CAC93719.1 923aa	3E-20	28% (43%)	regulator [Lechevalieria aerocolonigenes]
				923aa BAC55205.1 943aa	1E-15	29% (36%)	activator [Streptomyces sp. TP-A0274]
				NP_733725.1 908aa	3E-12	28% (41%)	regulator [Streptomyces coelicolor A3(2)]

SEQ

GenBank

% Identity

18

ORF	ID SEQ	Family	# aa	homology	Probability	(% Similarity)	Proposed function of GenBank match
4	8	IDSA	362	NP_601376.2	2E-80	49% (65%)	GGPP synthase [Corynebacterium glutamicum
				371aa NP_738677.1	3E-79	48% (62%)	polyprenyl synthase, Corynebacterium efficiens
				366aa NP_216689.1	2E-78	46% (61%)	idsA2 [Mycobacterium tuberculosis H37Rv]
5	10	MVKA	354	352aa BAB07790.1	2E-71	46% (59%)	mevalonate kinase [Streptomyces sp. CL190]
				345aa BAB07817.1 334aa	5E-66	45% (57%)	mevalonate kinase [Kitasatospora griseola]
				NP_720650.1 332aa	3E-36	29% (48%)	mevalonate kinase [Streptococcus mutans
6	12	DMDA	346	BAB07791.1 350aa	2E-88	58% (65%)	diphosphomevalonate decarboxylase [Streptomyces sp.
				BAB07818.1 300aa	2E-69	53% (61%)	mevalonate diPH decaroboxylase [<i>Kitasatospora griseola</i>]
				NP_785307.1 325aa	3E-44	34% (46%)	diphosphomevalonate decarboxylase [Lactobacillus plantarum
7	14	MVKP	369	BAB07792.1 374aa	4E-93	50% (60%)	phosphomevalonate kinase [Streptomyces sp. CL190]
				BAB07819.1 360aa	6E-77	48% (56%)	phosphomevalonate kinase [Kitasatospora griseola]
				AAG02442.1 368aa	2E-31	29% (42%)	3 phosphomevalonate kinase [Enterococcus faecalis]
8	16	IPPI	360	Q9KWF6 364aa	1E-128	66% (74%)	Isopentenyl-diphosphate delta-isomerase
				Q9KWG2 363aa	1E-128	66% (77%)	Isopentenyl-diphosphate delta-isomerase
				NP_814639.1 347aa	5E-73	44% (61%)	isopentenyl diphosphate isomerase [Enterococcus faecalis
9	18	HMGA	351	BAA70975.1 353aa	1E-165	82% (91%)	3-hydroxy-3-methylglutaryl coenzyme A reductase [<i>Streptomyces</i> sp.]
				BAA74565.1 353aa	1E-160	81% (89%)	3-hydroxy-3-methylglutaryl coenzyme A reductase [<i>Kitasatospora griseola</i>]
				BAA74566.1 353aa	1E-155	80% (86%)	3-hydroxy-3-methylglutaryl coenzyme A reductase [<i>Streptomyces</i> sp.]
10	20	KASH	391	BAB07795.1 389aa	1E-148	67% (78%)	3-hydroxy-3-methylglutaryl CoA synthase [Streptomyces sp. CL190]
				BAB07822.1 346aa	1E-136	70% (78%)	HMG-CoA synthase [Kitasatospora griseola]
				CAD24420.1 388aa	6E-79	43% (54%)	HMG-CoA synthase [Paracoccus zeaxanthinifaciens]
11	22	IPTN	290	NP_631248.1 295aa	5E-22	28% (44%)	hypothetical protein [<i>Streptomyces</i> <i>coelicolor</i> A3(2)]
				AAN65239.1 324aa	5E-06	25% (40%)	cloQ [Streptomyces roseochromogenes subsp. oscitans]
12	24	SPKG	370	AAM78435.1 344aa	5E-48	54% (63%)	two-component sensor [Streptomyces coelicolor A3(2)]
				NP_630507.1 382aa	5E-48	54% (63%)	sensor kinase [Streptomyces coelicolor A3(2)]
				ZP_00058991.1 407aa	9E-34	44% (58%)	Signal transduction histidine kinase [Thermobifida fusca]
13	26	RREB	220	NP_630508.1 224aa	3E-79	67% (81%)	regulatory protein [<i>Streptomyces coelicolor</i> A3(2)]
				ZP_00058992.1 221aa	4E-67	59% (75%)	Response regulator [Thermobifida fusca]
				NP_625364.1 221aa	6E-66	60% (74%)	response regulator [<i>Streptomyces</i> <i>coelicolor</i> A3(2)]
14 15	28 30	UNES UNEZ		No hit NP_649459.2	7.6E-02	38% (60%)	— CG1090-PB [Drosophila melanogaster]
				628aa NP_730819.1	7.6E-02	38% (60%)	CG1090-PA [Drosophila melanogaster]
				473aa AAM11079.1 428aa	7.6E-02	38% (60%)	GH23040p [Drosophila melanogaster]
16	32	OXDS	661	428aa NP_242948.1	1E-52	30% (46%)	unknown conserved protein [Bacillus
				500aa ZP_00091617.1 480aa	3E-32	29% (41%)	halodurans] Putative multicopper oxidases [Azotobacter vinelandii]
				480aa NP_252457.1 463aa	1E-31	28% (42%)	metallo-oxidoreductase [<i>Pseudomonas</i> aeruginosa PA01]
							101115111001 1701

TABLE 7-continued Sequence comparison and ORF correlation

				Sequ	ience compariso	on and ORF corre	elation
ORF	SEQ ID	Family	# aa	GenBank homology	Probability	% Identity (% Similarity)	Proposed function of GenBank match
17	34	UNFD	129	NP_437360.1 127aa	7E-33	60% (72%)	bleomycin resistance protein family [Sinorhizobium meliloti]
				AAO91879.1 123aa	1E-31	58% (74%)	unknown [uncultured bacterium]
				NP_103287.1 131aa	1E-23	48% (62%)	unknown protein [Mesorhizobium loti]
18 19	36 38	UNFA CSMB	178 661	ZP_00137697.1	1E-166	51% (66%)	Anthranilate/para-aminobenzoate synthase
				769aa NP_250594.1	1E-166	51% (66%)	[Pseudomonas aeruginosa phenazine biosynthesis protein PhzE [Pseudomonas gomerinosa PA01]
				627aa ZP_00137701.1 687aa	1E-166	51% (66%)	[Pseudomonas aeruginosa PA01] Anthranilate/para-aminobenzoate synthas [Pseudomonas aeruginosa
20	40	AAKD	427	P41403 421aa	1E-64	38% (51%)	Aspartokinase (Aspartate kinase)
				ZP_00057166.1 445aa	2E-64	37% (52%)	Aspartokinases [Thermobifida fusca]
				AAD49567.1 421aa	6E-64	37% (52%)	aspartokinase subunit A [<i>Amycolatopsis mediterranei</i>]
21	42	ALDB	274	NP_275722.1 266aa	2E-53	45% (64%)	conserved protein [Methanothermobacter thermautotrophicus]
				NP_614692.1 270aa	2E-52	43% (61%)	Fructose-1,6-bisphosphate aldolase [Methanopyrus kandleri AV19]
22		INFO	2/7	NP_615406.1 267aa	2E-50	43% (61%)	fructose-bisphosphate aldolase [Methanosarcina acetivorans str. C2A]
22	44	UNFC	367	NP_275723.1 378aa NP_614691.1	4E-46	38% (56%)	conserved protein [<i>Methanothermobacter</i> <i>thermautotrophicus</i>] alternative 3-dehydroquinate synthase
				NP_614691.1 402aa NP_248244.1	2E-45 2E-43	39% (55%) 40% (59%)	[Methanopyrus kandleri conserved hypothetical protein
23	46	HYDK	253	NF248244.1 361aa NP577771.1	2E-43	40% (39%) 31% (49%)	[Methanococcus jannaschii metal-dependent hydrolase [Pyrococcus
			200	247aa NP_142108.1	1E-12	33% (52%)	<i>furiosus</i> DSM 3638] hypothetical protein PH0093 [<i>Pyrococcus</i>
				247aa NP_125791.1	1E-11	28% (50%)	horikoshii] hypothetical protein [Pyrococcus abyssi]
24	48	ADSA	438	248aa NP_070499.1	2E-41	35% (49%)	coenzyme F390 synthetase
				433aa NP_618724.1 434aa	5E-41	34% (50%)	[Archaeoglobus fulgidus coenzyme F390 synthetase [Methanosarcina acetivorans
				NP_632700.1 437aa	7E-41	35% (50%)	Coenzyme F390 synthetase [<i>Methanosarcina mazei</i> Goe1]
25	50	HOXV	396	ZP_00027430.1 442aa	8E-76	42% (59%)	2-polyprenyl-6-methoxyphenol hydroxylase [Burkholderia fungorum]
				NP627457.1 420aa	1E-71	38% (51%)	salicylate hydroxylase [<i>Streptomyces coelicolor</i> A3(2)]
				ZP_00033877.1 403aa	2E-68	37% (51%)	2-polyprenyl-6-methoxyphenol hydroxylase [Burkholderia fungorum]
26	52	SDRA	261	NP_391080.1 261aa	6E-58	46% (57%)	2,3-dihydro-2,3-dihydroxybenzoate dehydrogenase [<i>Bacillus subtilis</i>]
				ZP_00059512.1 260aa	1E-55	45% (56%)	Dehydrogenase [<i>Thermobifida fusca</i>]
27	54	DHBS	<u>2</u> 24	AAG31126.1 257aa Q51790	9E-55 7E-60	46% (56%)	MxcC [<i>Stigmatella aurantiaca</i>] isochorismatase
21	54	ривэ	224	Q51790 207aa Q51518	1E-58	56% (72%) 56% (71%)	isochorismatase
				207aa NP_391077.1	2E-58	52% (69%)	isochorismatase [Bacillus subtilis]
28	56	SDRA	233	312aa NP_103491.1	9E-21	32% (49%)	acyl-carrier protein reductase
				242aa AAL14912.1	1E-15	28% (44%)	[Mesorhizobium loti] short-chain dehydrogenase [Rhizobium
				245aa NP_902480.1	7E-15	29% (44%)	leguminosarum bv. trifolii] oxidoreductase [Chromobacterium
29	58	UNIQ	246	235aa S18541 281aa	4.5E-02	29% (43%)	violaceum hypothetical protein 3 - Streptomyces coelicolor
				281aa NP_629228.1 281aa	5.9E-02	29% (43%)	coelicolor hypothetical protein [<i>Streptomyces</i> <i>coelicolor</i> A3(2)]

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TABLE 7-continued

	Sequence comparison and ORF correlation							
ORF	SEQ ID	Family	# aa	GenBank homology	Probability	% Identity (% Similarity)	Proposed function of GenBank match	
30	60	UNFE	111	ZP_00058149.1	1E-10	36% (48%)	membrane protein [Thermobifida fusca]	
				130aa NP_737701.1 120aa	1E-09	33% (46%)	hypothetical protein [Corynebacterium efficiens	
				NP_827629.1 118aa	7E-09	33% (49%)	hypothetical protein [<i>Streptomyces</i> avermitilis MA-4680]	
31	62	EFFT	559	ZP_00058148.1 537aa	2E-67	32% (49%)	Predicted symporter [<i>Thermobifida fusca</i>]	
				NP_626090.1 544aa	4E-66	31% (49%)	transport protein [<i>Streptomyces coelicolor</i> A3(2)]	
				NP_827630.1 549aa	7E-63	31% (49%)	sodium-dependent symporter [<i>Streptomyces avermitilis</i>	
32	65	НОҮН	532	AAM96655.1 544aa	2E-92	39% (53%)	2,4-dihydroxybenzoate monooxygenase [Sphingobium chlorophenolicum]	
				ZP_00029353.1 543aa	1E-73	35% (49%)	2-polyprenyl-6-methoxyphenol hydroxylase [Burkholderia fungorum]	
				NP_769326.1 569aa	5E-62	33% (48%)	blr2686 [Bradyrhizobium japonicum] dbj	
33	67	DAHP	423	T03226 391aa	1E-111	54% (68%)	hypothetical protein - Streptomyces hygroscopicus	
				ZP_00137693.1 405aa	3E-87	45% (61%)	DAHP synthase [Pseudomonas aeruginosa UCBPP-PA14]	
				NP_250592.1 405aa	1E-86	45% (61%)	phenazine biosynthesis protein PhzC [Pseudomonas aeruginosa	
34	69	REGG	340	BAC53615.1 346aa	1E-67	46% (62%)	regulator protein [Streptomyces kasugaensis]	
				S44506 424aa	3E-66	46% (60%)	regulator protein - Streptomyces glaucescens	
				AAK81822.1 348aa	1E-65	44% (59%)	transcriptional regulator [Streptomyces lavendulae]	
35	71	UNFJ	493	ZP_00073237.1 678aa	7E-35	27% (43%)	RTX toxins [Trichodesmium erythraeum IMS101]	
				NP_484716.1 433aa	3E-05	23% (37%)	similar to vanadium chloroperoxidase [Nostoc sp.	
				ZP_00067005.1 667aa	7.4E-02	27% (37%)	hypothetical protein [Microbulbifer degradans 2-40]	
36	74	RECI	112	NP_627088.1 125aa	3E-17	48% (59%)	hypothetical protein. [<i>Streptomyces</i> <i>coelicolor</i> A3(2)]	
				NP_846017.1 109aa	7E-15	40% (59%)	hypothetical protein [Bacillus anthracis str. Ames]	
				NP_241272.1 174aa	9E-15	37% (58%)	unknown conserved protein [Bacillus halodurans]	
37	76	UNIQ	325	NP_422203.1 187aa	1E-03	39% (59%)	hypothetical protein [Caulobacter crescentus CB15]	
38	78	OXAH	663	ZP_00058724.1 659aa	0E+00	57% (67%)	Acyl-CoA dehydrogenases [Thermobifida fusca]	
				AAB97825.1 433aa	5E-93	46% (56%)	acyl-CoA oxidase [Myxococcus xanthus]	
				AAF14635.1, 694aa	5E-85	37% (52%)	1 acyl-CoA oxidase [Petroselinum crispum]	
39	80	ABCA	537	T14162 574aa	9E-62	37% (47%)	hABC transport protein - Mycobacterium smegmatis	
				NP_624808.1	4E-60	35% (46%)	ABC transporter [<i>Streptomyces coelicolor</i> A3(2)]	
				NP_822745.1	8E-32	31% (42%)	ABC transportert [<i>Streptomyces avermitilis</i> MA-4680]	
40	82	ABCA	596	T14180 1122aa	1E-107	40% (51%)	exiT protein - Mycobacterium smegmatis	
				AAC82548.1 589aa	1E-107	40% (51%)	unknown [Mycobacterium smegmatis]	
				NP_624810.1 601aa	3E-97	37% (48%)	ABC-transporter [<i>Streptomyces coelicolor</i> A3(2)]	
41	84	UNIQ	507	NP_831570.1 676aa	8E-07	24% (44%)	methyltransferases [Bacillus cereus	
				NP_655735.1 676aa	2E-06	23% (44%)	ubiE/COQ5 methyltransferase family [<i>Bacillus anthracis</i>	
				NP_844290.1 681aa	2E-06	23% (44%)	hypothetical protein [Bacillus anthracis str. Ames]	

	Sequence comparison and ORF correlation							
ORF	SEQ ID	Family	# aa	GenBank homology	Probability	% Identity (% Similarity)	Proposed function of GenBank match	
42	86		232	NP_830809.1 208aa	8E-08	22% (35%)	Transporter, LysE family [Bacillus cereus]	
				NP_844737.1 210aa	2E-07	22% (35%)	homoserine/threonine efflux protein[Bacillus anthracis	
				NP_655752.1 208aa	1E-06	22% (36%)	LysE, LysE type translocator [<i>Bacillus anthracis</i>	
43	88		132	NP_827272.1 127aa	4E-09	36% (49%)	hypothetical protein [<i>Streptomyces</i> avermitilis MA-4680]	
				NP_246491.1, 112aa	5E-02	22% (47%)	unknown [Pasteurella multocida]	

[0161] The ORFs encoding proteins involved in the biosynthesis of farnesyl dibenzodiazepinones are assigned a putative function and grouped together in families based on sequence similarity to known proteins. To correlate structure and function, the protein families are given a four-letter designation used throughout the description and figures as indicated in Table 8. The meaning of the four letter designations is as follows: AAKD designates an amino acid kinase; ABCA and ABCC designate ABC transporters; ADSA designates an amide synthetase; ALDB designates an aldolase function; CSMB designates a chorismate transaminase; DAHP designates a 3,4-dideoxy-4-amino-D-arabino-heptulosonic acid 7-phosphate synthase activity; DHBS designates a 2,3-dihydro-2,3-dihydroxybenzoate synthase activity; DMDA designates a diphosphomevalonate decarboxylase; EFFT designates an efflux protein; HMGA designates a 3-hydroxy-3methylglutaryl-CoA reductase; HOXV designates a monooxygenase activity; HOYH designates a hydroxylase/ decarboxylase activity; HYDK designates a hydrolase activity; IDSA designates an isopentenyl diphosphate synthase; IPPI designates an isopentenyl diphosphate isomerase; IPTN designates an isoprenyltransferase; KASH designates 3-hydroxy-3-methylglutaryl-CoA synthase; MVKA designates a mevalonate kinase; MVPK designates a phosphomevalonate kinase; OXAH designates an acylCoA oxidase; OXDS designates an oxidoreductase; RECH, RECI, REGD, REGG and RREB designate regulators; SDRA designates a dehydrogenase/ketoreductase, SPKG designates a sensory protein kinase; UNES, UNEZ, UNFA, UNFC, UNFD, UNFE, UNFJ and UNIQ designate proteins of unknown function.

TABLE 8

		_ M
FAMILY	FUNCTION:	
AAKD	amino acid kinase; strong homology to	
	primary aspartate kinases, converting L-	Ož
	aspartate to 4-phospho-L-aspartate	O
ABCA	ABC transporter	RJ
ABCC	ABC transporter	RJ
ADSA	adenylating amide synthetase	
ALDB	aldolase; similarity to fructose-1,6-	
	biphosphate aldolase that generates D-	RI
	glyceraldehyde-3Ph, precursor of D-	K
	erythrose-4Ph involved in the shikimate	
	pathway	
CSMB	chorismate transaminase, similarity to	
	anthranilate synthase	RJ
DAHP	DAHP synthase, class II; involved in	RI
	formation of aminoDAHP from PEP and	
	erythrose-4-phosphate	

TABLE 8-continued

FAMILY	FUNCTION:
DHBS	2,3-dihydro-2,3-dihydroxybenzoate
	synthase (isochorismatase)
DMDA	diphosphomevalonate decarboxylase
	(mevalonate pyrophosphate decarboxylase)
EFFT	efflux protein
HMGA	HMG-CoA reductase; converts 3-hydroxy-3-
	methylglutaryl-CoA to mevalonate plus
	CoA in isoprenoid biosynthesis
HOXV	FAD monooxygenase; shows homology to a
	variety of monooxygenases including salicylate hydroxylases, zeaxanthin
	epoxidases
ноун	hydroxylase/decarboxylase; FAD-
110111	dependent monooxygenase
HYDK	hydrolase
IDSA	isoprenyl diphosphate synthase, catalyzes
	the addition of 2 molecules of isopentenyl
	pyrophosphate to dimethylallyl pyrophos-
	phate to generate GGPP
IPPI	isopentenyl diphosphate isomerase, catalyzes
	the isomerization of IPP to produce
	dimethylallyl diphosphate
IPTN	isoprenyltransferase; catalyzes covalent N-
	terminal attachment of isoprenyl units to
	amide groups of nitrogen-containing
KASH	heterocycle rings HMG-CoA synthase; condenses acetyl-CoA
казн	with acetoacetyl-CoA to form 3-
	hydroxy-3-methylglutaryl-CoA
MEBI	membrane protein
MVKA	mevalonate kinase; converts mevalonate
IVI V KA	to 5-phosphomevalonate in the mevalonate
	pathway of isoprenoid biosynthesis
MVKP	phosphomevalonate kinase; converts 5-
	phosphomevalonate to 5-diphosphomevalonate
	in the mevalonate pathway of isoprenoid
	biosynyhesis
OXAH	acyl CoA oxidase
OXDS	oxidoreductase
RECH	regulator
RECI	regulator; similarity to PadR transcriptional
	regulators involved in repression of phenolic
	acid metabolism
REGD	transcriptional regulator; relatively large
	regulators with an N-terminal ATP-binding
	domain containing Walker A and B motifs and
	a C-terminal LuxR type DNA-binding domain
REGG	regulator
RREB	response regulator; similar to response
	regulators that are known to bind DNA and
	act as transcriptional activators

TABLE 8-continued

FAMILY	FUNCTION:
SDRA	dehydrogenase/ketoreductase, NAD-dependent
SPKD	sensory protein kinase, two component system
SPKG	sensory protein kinase, two component system
UNES	unknown function
UNEZ	unknown function
UNFA	unknown function
UNFC	unknown function
UNFD	unknown function
UNFE	putative membrane protein
UNFJ	unknown function
UNIQ	unknown function

[0162] Biosynthesis of ECO-04601 involves the action of various enzymes that synthesize the three building blocks of the compound, namely the farnesyl-diphosphate component (FIG. **5**), the 3-hydroxy-anthranilate-adenylate component (FIG. **6**) and the 2-amino-6-hydroxy-benzoquinone component (FIG. **7**) that are subsequently condensed to form the final compound (FIG. **8**).

[0163] The farnesyl-diphosphate biosynthesis involves the concerted action of seven enzymes (FIG. 5). ORF 10 (KASH) (SEQ ID NO: 20) encodes a hydroxymethylglutaryl-CoA synthase that catalyzes an aldol addition of acetyl-CoA onto acetoacyl-CoA to yield 3-hydroxy-3-methylglutaryl-CoA (HMG-CoA). This product is subsequently reduced through the action of ORF 9 (HMGA) (SEQ ID NO: 18) to form mevalonic acid (MVA). ORF 5 (MVKA) (SEQ ID NO: 10) phosphorylates mevalonate to 5'-phosphomevalonate using ATP as the phosphate donor. The next step in the farnesyldiphosphate biosynthesis is the phosphorylation reaction of the 5'-phosphomevalonate to 5'-pyrophosphomevalonate (DPMVA) that is catalyzed by ORF 7 (MVKP) (SEQ ID NO: 14). Subsequent decarboxylation of 5'-pyrophosphomevalonate catalyzed by ORF 6 (DMDA) (SEQ ID NO: 12) yields isopentenyl diphosphate (IPP) which is then converted to dimethylallyldiphosphate (DMADP) through the action of ORF 8 (IPPI) (SEQ ID NO: 16) that has isomerase enzymatic activity. The final step in the biosynthesis of farnesyl-diphosphate is the condensation of one molecule of dimethylallyldiphosphate with two molecules of isopentenyl diphosphate catalyzed by the isoprenyl diphosphate synthase ORF 4 (IDSA) (SEO ID NO: 8). The described pathway involved in synthesis of farnesyl-diphosphate is entirely consistent with related mevalonate pathways described in other actinomycete species (Takagi et al., J. Bacteriol. 182, 4153-4157, (2000)). [0164] Biosynthesis of the 3-hydroxy-anthranilate component involves the use of precursors derived from the shikimate pathway (FIG. 6). Chorismic acid is transaminated through the action of ORF 19 (CSMB) (SEQ ID NO: 38) to form aminodeoxyisochorismic acid. This enzyme resembles anthranilate synthases and is likely to catalyze specifically the transfer of the amino group using glutamine as the amino donor. The next step involves isochorismatase activity and is mediated by ORF 27 (DHBS) (SEQ ID NO: 54). This reaction consists in the removal of the pyruvate side chain from aminodeoxyisochorismic acid to form 6-amino-5-hydroxycyclohexa-1,3-dienecarboxylic acid. This compound is subsequently oxidized through the action of ORF 26 (SDRA) (SEQ ID NO: 52) yielding 3-hydroxy-anthranilic acid. ORF 24 (ADSA) (SEQ ID NO: 48) catalyzes the activation of 3-hydroxy-anthranilic acid through adenylation generating the 3-hydroxy-anthranilate-adenylate component (FIG. 6).

[0165] Biosynthesis of the 2-amino-6-hydroxy-benzoquinone component of the farnesyl dibenzodiazepinone, requires components derived from the aminoshikimate pathway. FIG. 7 depicts the series of enzymatic reactions involved in the biosynthesis of this constituent. ORF 21 (ALDB) (SEQ ID NO: 42) resembles aldolases involved in the generation of precursors of D-erythrose-4-phosphate which is part of the aminoshikimate pathway used for the generation of 2-amino-6-hydroxy-[1,4]-benzoquinone. ORF 33 (DAHP) (SEQ ID NO: 67) catalyzes the initial step in the aminoshikimate pathway that corresponds to the formation of 3,4-dideoxy-4amino-D-arabino-heptulosonic acid 7-phosphate (amino DAHP) from phosphoenolpyruvate (PEP) and erythrose 4-phosphate (E-4Ph). Subsequent reactions leading to 3-amino-5-hydroxy-benzoic acid are catalyzed by enzymes provided by primary metabolism biosynthetic pathways present in Micromonospora sp. strain 046-ECO11. ORF 25 (HOXV) (SEQ ID NO: 50) hydroxylates 3-amino-5-hydroxy-benzoic acid at position 2, generating 3-amino-2,5dihydroxy-benzoic acid. This intermediate is further modified by ORF 32 (HOYH) (SEQ ID NO: 65) that catalyzes a decarboxylative oxidation reaction yielding 6-amino-benzene-1,2,4-triol. A final oxidation reaction is performed by ORF 16 (OXDS) (SEQ ID NO: 32) yielding 2-amino-6-hydroxy-[1,4]-benzoquinone (FIG. 7).

[0166] Assembly of the three components resulting in the farnesyl dibenzodiazepinone is catalyzed by ORFs 24 and 11 (FIG. 8). ORF 24 (ADSA) (SEQ ID NO: 48) catalyzes the condensation of the adenylated 3-hydroxy-anthranilate with the 2-amino-6-hydroxy-[1,4]-benzoquinone component. A spontaneous condensation between the free amino group of the 3-hydroxy-anthranilate and one of the carbonyl groups present on the 2-amino-6-hydroxy-[1,4]-benzoquinone component occurs yielding a dibenzodiazepinone intermediate. This compound is further modified through transfer of the farnesyl group of the farnesyl-diphosphate intermediate onto the nitrogen of the amide of the dibenzodiazepinone catalyzed by ORF 11 (IPTN) (SEQ ID NO: 22) and resulting in the formation of the farnesyl dibenzodiazepinone (FIG. 8). [0167] Additional ORFs, namely ORF 2 (RECH) (SEQ ID NO: 4), ORF 3 (REGD) (SEQ ID NO: 6), ORF 12 (SPKG) (SEQ ID NO: 24), ORF 13 (RREB) (SEQ ID NO: 26), ORF 34 (REGG) (SEQ ID NO: 69) and ORF 36 (RECI) (SEQ ID NO: 74) are involved in the regulation of the biosynthetic locus encoding the farnesyl dibenzodiazepinone. Other ORFs, namely ORF 1 (ABCC) (SEQ ID NO: 2), ORF 31 (EFFT) (SEQ ID NO: 62), ORFs 39 and 40 (ABCA) (SEQ ID NOS: 80 and 82, respectively) and ORF 42 (SEQ ID NO: 86) are involved in transport. Other ORFs involved in the biosynthesis of the farnesyl dibenzodiazepinone include ORF 20 (AAKD) (SEQ ID NO: 40), ORF 23 (HYDK) (SEQ ID NO: 46), ORF 38 (OXAH) (SEQ ID NO: 78) as well as ORFs 14, 15, 17, 18, 22, 29, 30, 35, 37, 41 and 43 (SEQ ID NOS: 28, 30, 34, 34, 44, 58, 60, 71, 76, 84 and 88, respectively) of unknown function.

Example 6

Farnesyl Dibenzodiazepinone Loci from Actinomycetes Species

[0168] A. Correlation of Loci A, B and C

[0169] Loci related to the biosynthetic locus present in *Micromonospora* sp. strain 046ECO-11 as described in Example 5 (referred to herein as locus A) and directing the biosynthesis of farnesyl diabenodiazepinones related to ECO-04601 were detected in the genome of two actinomycetes using the genome scanning method described in U.S.

Ser. No. 10/232,370, CA 2,352,451 and Zazopoulos et. al., *Nature Biotechnol.*, 21, 187-190 (2003).

[0170] Locus B (052E) was detected in *Micromonospora* echinospora challisensis NRRL 12255. The locus spans approximately 38,000 base pairs of DNA and encodes 33 proteins. Locus C (237C) was detected in *Streptomyces carzinostaticus neocarzinostaticus* ATCC 15944. This locus spans approximately 37,000 base pairs of DNA and encodes 33 proteins. More than 10 kilobases of DNA sequence were analyzed on each side of the two loci and these regions were deemed to contain primary genes.

[0171] In order to identify the function of the proteins coded by the genes forming the biosynthetic loci B an C the gene products of their ORFs 1 to 33, were compared, using the BLASTP version 2.2.10 algorithm with the default parameters, to sequences in the National Center for Biotechnology Information (NCBI) nonredundant protein database and the DECIPHER® database of microbial genes, pathways and natural products (Ecopia BioSciences Inc. St.-Laurent, QC, Canada).

[0172] The ORFs encoding proteins present in loci A, B, and C are assigned a putative function and grouped together in families based on sequence similarity to known proteins. To correlate structure and function, the protein families are given a four-letter designation used throughout the description and figures as indicated in Table 8 of Example 5.

[0173] Comparison of loci A, B and C clearly indicates that all three loci are related and encode similar enzymatic functions. Therefore, the compounds produced by the enzymes encoded by loci B and C are structurally closely related to ECO-04601. Table 9 correlates the protein families of loci B and C to those of locus A. All 33 ORFs found in locus B have counterparts in locus A. Similarly, all 33 ORFs present in locus C have counterpart proteins in locus A, with the exception of ORFs 30, 31, and 32 that encode a sensory protein kinase protein, a response regulator and a membrane protein. These observations suggest that the compounds produced by loci B and C encoded proteins share a high degree of similarity with ECO-04601.

TABLE 9

	3 and C ORFs fi		
	А	В	С
ABCC	1	_	_
RECH	2	1	1
REGD	3	2	2
IDSA	4	3	3
MVKA	5	4	4
DMDA	6	5	5
MVKP	7	6	6
IPPI	8	7	7
HMGA	9	8	8
KASH	10	_	9
IPTN	11	9	10
SPKG	12	15	12
RREB	13	16	11
UNES	14	10	33
UNEZ	15	14	_
OXDS	16	13	_
UNFD	17	12	_
UNFA	18	11	
CSMB	19	17	14
AAKD	20	18	15
ALDB	21	19	16
UNFC	22	20	17
HYDK	23	21	18

TABLE 9-continued

Loci A, B and C ORFs function and correlation						
	А	В	С			
ADSA	24	22	19			
HOXV	25	23	20			
SDRA	26	24	21			
DHBS	27	25	22			
SDRA	28	26	23			
UNGA	29	27	24			
UNFE	30	28	25			
EFFT	31	29	26			
HOYH	32	30	27			
DAHP	33	31	28			
REGG	34	32	_			
UNFJ	35	33	13/29			
RECI	36					
UNIQ	37					
OXAH	38					
ABCA	39					
ABCA	40					
UNIQ	41					
SPKD			30			
RREB			31			
MEBI	_		32			

[0174] FIG. 5 depicts the three biosynthetic loci A, B and C. All ORFs are represented by arrows and their orientation indicate the direction of the transcription of each ORF; highlighted ORFs are involved in the biosynthesis of the farnesyl unit. ORFs 4, 5, 6, 7, 8, 9, and 10 in locus A participate in the synthesis of the farnesyl unit present in the farnesyl dibenzodiazepinone. Counterparts of these ORFs are found in locus B (ORFs 3, 4, 5, 6, 7 and 8) as well as in locus C(ORFs 3, 4, 5, 6, 7, 8 and 9). As shown in FIG. 5, proteins encoded by these ORFs participate in an orderly fashion in the biosynthesis of farnesyl-diphosphate component starting with acetoacetyl-CoA and acetyl-CoA. All enzymes necessary for the synthesis of farnesyl-diphosphate are present in all three loci with the exception of a hydroxymethylglutaryl-CoA synthase (KASH) which is absent from locus B. The product of this enzymatic reaction, 3-hydroxy-3-methylglutaryl-CoA is provided by an alternative biosynthetic pathway of the primary metabolism of the microorganism or by a hydroxymethylglutaryl-CoA synthase located elsewhere in the genome. The described pathway involved in synthesis of farnesyl-diphosphate is entirely consistent with related mevalonate pathways described in other actinomycete species (Takagi et al., J. Bacteriol. 182, 4153-4157, (2000) and FIG. 5).

[0175] FIG. **6** depicts ORFs 19, 21, 24, 26 and 27 in locus A involved in the biosynthesis of the 3-hydroxy-anthranilate component of the farnesyl dibenzodiazepinone. Counterparts of these ORFs are found in locus B (ORFs 17, 19, 22, 24 and 25) as well as in locus C(ORFs 14, 16, 19, 21 and 22). As shown in FIG. **6**, proteins encoded by these ORFs participate in an orderly fashion to the biosynthesis of the 3-hydroxy-anthranilate-adenylate component starting with precursors from the pentose phosphate pathway and chorismic acid. In particular, the enzyme responsible for the adenylation of 3-hydroxy-anthranilic acid (ADSA) that corresponds to ORFs 24, 22 and 19 in loci A, B and C respectively is present in all three loci as well as the remaining enzymes that participate in the biosynthesis of 3-hydroxy-anthranilate component present in dibenzodiazepinones.

[0176] FIG. **7** highlights ORFs 16, 24, 25, 32 and 33 in locus A involved in the biosynthesis of the 2-amino-6-hy-

droxy-[1,4]benzoquinone component of the farnesyl dibenzodiazepinone. Counterparts of these ORFs are found in locus B (ORFs 13, 19, 23, 30 and 31) as well as in locus C(ORFs 16, 20, 27 and 28) with the exception of ORF corresponding to the oxidoreductase (OXDS) present in loci A and B. As shown in FIG. 7, proteins encoded by these ORFs participate in an orderly fashion in the biosynthesis of the 2-amino-6-hydroxy-[1,4]benzoquinone component starting with precursors from the pentose phosphate pathway and 3,4-dideoxy-4-amino-D-arabino-heptulosonic acid 7-phosphate (amino DAHP).

[0177] FIG. 8 highlights ORFs 11 (SEQ ID NO: 22) and 24 (SEQ ID NO: 48) in locus A involved in the assembly of all three components, 3-hydroxy-anthranilate, 2-amino-6-hydroxy-[1,4]benzoquinone and farnesyl-diphosphate to form the farnesyl dibenzodiazepinone. Counterparts of these ORFs are found in locus B (ORFs 9 (SEQ ID NO: 90) and 22 (SEQ ID NO: 92)) as well as in locus C(ORFs 10 (SEQ ID NO: 94) and 19 (SEQ ID NO: 96)). The isoprenyltransferase ORF 10 of locus C (SEQ ID NO: 96) is partial and represents the N-terminal part of the protein. IPTN ORFs 11 (SEQ ID NO: 22), 9 (SEQ ID NO: 90) and 10 (SEQ ID NO: 94) in loci A, B and C respectively catalyze the transfer of the farnesyl unit onto the core element of the farnesyl dibenzodiazepinone and related compounds produced by loci B and C. ADSA ORFs 24 (SEQ ID NO: 48), 22 (SEQ ID NO: 92) and 19 (SEO ID NO: 96) in loci A, B and C respectively catalyze the condensation of 3-hydroxy-anthranilate and 2-amino-6-hydroxy-[1, 4]benzoquinone and farnesyl-diphosphate to form the dibenzodiazepinone core element of ECO-04601 and related compounds produced by loci B and C.

[0178] B. Clustal[™] Alignments

[0179] Alignements of isoprenyl transferases (IPTN) and adenylating amide synthetases (ADSA) of loci A, B and C, respectively presented in FIGS. **9** and **10**, were generated by the ClustalTM alignment method.

[0180] FIG. **9** shows an alignment of ORFs 11 (SEQ ID NO: 22), 9 (SEQ ID NO: 90, which represents the polypeptide deduced from SEQ ID NO:91) and 10 (SEQ ID NO: 94, which represents the polypeptide deduced from SEQ ID NO:95) in loci A, B and C respectively, highlighting the phylogenetic relatedness of these three proteins. The amino acid sequence of all three proteins is extremely conserved as shown by the codes on the fourth line, suggesting that these proteins share a well-conserved and related isoprenyltransferase enzymatic function. The following consensus amino acid sequence (also as SEQ ID NO: 98) that represents all three sequences was generated using the hmmemit algorithm (HMMER, Washington University in St-Louis, School of Medicine, MO, USA, http://hmmer.wustl.edu):

[0181] "AaELysviEesARILdvaCsrDrvwpiL-

saYGDaFaHpaavvAFRvAtalRHvGELD CRFttHPddRD-PYAIALsrGLtPktdH-

PvGsLLsevqeRIPvesyGiDFGvvGGFKKiYafFtPDe

LqevaaLAgiPamPRsLAgnadFFeR-

yGlddrvGvlGiDYPartvnvyfndvpaesfesetirstlreiGma epsermI kIGekafGlyvtlGwdsseiericyaaattdIttIpvpvepeiekfvksvpyGGedrkfvyGvaltpkGey ykleshykwkpGavdfi"

[0182] FIG. **10** shows an alignment of ORFs 24 (SEQ ID NO: 48), 22 (SEQ ID NO: 92, which represents the polypeptide deduced from SEQ ID NO: 93) and 19 (SEQ ID NO: 96, which represents the polypeptide deduced from SEQ ID NO: 97) in loci A, B and C respectively, highlighting the phyloge-

netic relatedness of these three proteins. The amino acid sequence of all three proteins is extremely conserved as shown by the codes on the fourth line, suggesting that these proteins share a well-conserved and related adenylating amide synthetase enzymatic function. The following consensus amino acid sequence (also as SEQ ID NO: 99) that represents all three sequences was generated using the hmmemit algorithm:

"VneprssLPrLGqWhGpEDLrrLqEKqLaqtvtWAaRsPFYRdRLds gAlPvtaaDLAdLPLttKqDLRDnYPFGmLAvPkERLAtYHEssGtAGr PtPsYYtAeDWtDLAERFARKWiGmsAeDvFLvRtPYALLLtGHLAH AAgRLrGAtvvPGDnRsLAmPYARvvRvmHDLgvtLtWsvPtECLiW AAAAtAAGHRPdvDFPALRALFvGGEPltdARRrRisRLWGvPviEE YGstEtGsLAGECPeGRIHLWADRALFEvYDPdtGtvrAdGdGqLvv tPLfREAmPLLRYnLEDnvsvsYDDCaCGWkLPtvrvLGRaAFGyRv GattitqHrLEElvFsLPeahrvvFWRAkAEPavLRiEiEvaeeHRv AAeAELtasvRaaFGvDsevtGLaPGtLiPreALtsmPDvvKPRsLF GPDEDWgKALLYY"

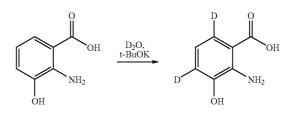
[0183] The amino acid shown for the consensus sequences (SEQ ID NOs: 98 and 99) are the highest probability amino acid at that position according to the HMM (hidden Markov model). Highly conserved residues (those with a probability of >0.5) are shown by capital letters while other residues (lowercase letters) are deduced by the program from the most common amino acid found at the specific position in the aligned proteins (*HMMER User's Guide*, Sean Eddy, October 2003, Washington University of Medicine, MO, USA, p 23-24).

Example 7

Labeled 3-Hydroxyanthranilic Acid Feeding

[0184] This experiment was designed to confirm the farnesyl dibenzodiazepinone biosynthetic pathway involves a 3-hydroxyanthranilate intermediate. First, labeled 4,6-dideuterio-3-hydroxyanthranilic acid was prepared. Then the labeled intermediate was fed to the *Micromonospora* sp. strain, the product was purified (see Example 2) and the results were analyzed. The following is an exemplary procedure to accomplish the feeding experiment:

[0185] A. Preparation of 4,6-dideuterio-3-hydroxyanthranilic acid



[0186] 3-Hydroxyanthranilic acid (108 mg, Sigma-Aldrich) was suspended in D_2O (2 mL). Potassium t-butoxide (154 mg) was added to give a brown solution. The solution was stirred at 100° C. under nitrogen for about 6 days. The reaction mixture was cooled to room temperature. The solution was acidified to pH 6 with 10N hydrochloric acid and white solid precipitated. The solid was filtered and dried in vacuo (93 mg). The ¹H NMR of the isolated product showed about 92-96% reduction of the proton signals (doublets) at the 4 and 6 positions. The ¹H NMR signal of the unchanged proton (5 position) also reflected the incorporation of the two deuterium; coupling to the 4 and 6 protons was nearly lost (triplet changed to a singlet having two very small side peaks).

[0187] B. 4,6-dideuterio-3-hydroxyanthranilic Acid Feeding

B.1. Culture Conditions:

[0188] To prepare a vegetative culture, *Micromonospora* sp. 046-Eco11 was grown on ISP2 agar (Difco) for 10 to 15 days, and the surface growth from the agar plate was homogenized and transferred to a 125 ml flask containing three glass beads (5 mm diameter), and 25 ml of sterile medium KH composed of 10 g glucose, 20 g potato dextrin, 5 g yeast extract, 5 g NZ-Amine A, and 1 g $CaCO_3$ made up to one liter with tap water and adjusted to pH 7 with 1 M NaOH.). This vegetative culture was incubated at 28° C. for about 70 hours on a shaker at 250 rpm with a 1-inch throw.

[0189] Following incubation, 18 ml was used to inoculate 2 L baffled flasks each containing 600 ml of sterile Hi production medium consisting of 20 g potato dextrin, 30 g glycerol, 2.5 g Bacto-peptone, 8.34 g yeast extract, and 3 g $CaCO_3$ made to one liter with distilled water and adjusted at pH 7.0 with 1 MNaOH. The culture was incubated at 28° C. for about 96 hours on a shaker at 250 rpm with 1-inch throw.

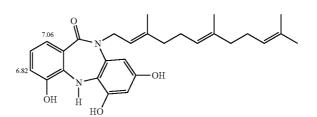
B.2. Feeding Experiment:

[0190] Vegetative cultures of *Micromonospora* sp. 046-Ecol 1 prepared in medium KH as explained above were used to inoculate Hi medium (four 125-mL flasks containing 25 mL). The medium was fed with 4,6-D₂-3-hydroxyanthranilic acid at 0.5 mg/mL before inoculation with the vegetative culture at 2% level. Control cultures without adding the labeled compound were prepared for each medium in the same way mentioned above. Effect of adding 4,6-D₂-3-hydroxyanthranilic acid on the production titre and growth was measured by adding the unlabeled compound to each medium in the same fashion explained above. The purified compound obtained from each experiment was tested by ¹H-NMR for incorporation ratio of the labeled substrate.

[0191] C. Results:

[0192] The purified farnesyl dibenzodiazepinone from the feeding experiment was analyzed both by ¹H NMR and mass spectrum. The ¹H NMR (in DMSO-d₆) was compared to the unlabelled standard. About 31% reduction in the intensity of the signals at 6.82 and 7.06 ppm in DMSO-d₆ (correspond to protons signals at 6.83 and 7.14 ppm in MeOH-d₄) was observed, which reflected a 31% incorporation of the deuterium at these positions. Mass spectral analysis gave about 47% incorporation of the deuterium labeled precursor.





[0193] The result indicated a direct incorporation of 3-hydroxyanthranilate as a precursor in the biosynthesis of ECO-04601.

Example 8

Methods of Using the Deposited Cosmids

[0194] Two deposits of *E. coli* DH10B vectors (046KM and 046KQ), having deposit accession numbers IDAC 250203-06 and IDAC 250203-07 respectively, each contain a cosmid clone and together span the whole biosynthetic locus of ECO-04601. The coverage of the locus by each deposited cosmid is described in Example 5 and shown on FIG. **4**.

[0195] Culture conditions to be employed for growing the deposited cosmid-containing DH10BTM E. coli are understood by a person of skill in the art (Sambrook et al. (1989) Molecular Cloning: A Laboratory Manual, 2nd ed., Cold Spring Harbor Laboratory Press). As a non-limiting example, upon receiving a sample of the deposited strain, either as a frozen glycerol stock or as an agar stab or in a liquid media, a small aliquot of the strain is gathered using a sterile metal loop and thereafter streaked onto a selective media agar on freshly prepared growth plates (e.g. disposable plastic Petri® plates). The aliquot is streaked so that single bacterial colonies can be isolated. A number of different growth media can be used, provided that the media contain an appropriate amount of a selective agent, for example an antibiotic. Standard growth media are known in the art, such as standard Luria Bertani (LB) media (10 grams of NaCl, 10 grams of tryptone, 5 grams of yeast extract, 20 grams of agar, with pH adjusted to 7.0 with 5.0 N NaOH add deionized water to a final volume of 1.0 liters, autoclaved then cooled to 55° C. followed by addition of 10 mL of 10-mg/mL filter-sterilized ampicilin or 5 ml of 10-mg/mL filter-sterilized kanamycin). Plates with streaked bacteria are incubated overnight (approximately 16 hours) at 37° C. to allow for growth of the bacterial colonies.

[0196] Cosmid DNA containing insert DNA are prepared from the above-noted strains by methods that are known in the art. As a non-limiting example, a single bacterial colony is selected from an agar plate (as referred to above) and restreaked onto a fresh agar plate, containing the appropriate selective agent as noted above, and allowed to grow overnight at 37° C. From this second agar plate, a single bacterial colony is selected and inoculated into 2.0 to 5.0 ml of liquid broth containing the appropriate amount of a selective agent, for example LB broth (prepared as per LB media, but lacking agar) containing ampicillin or kanamycin in a concentration as noted in the preceding paragraph, in order to generate a liquid starter culture of the single bacterial colony. This starter culture is grown to late logarithmic stage (approximately 8 hours), at which time an aliquot of the starter culture is withdrawn and diluted, by a factor of 500 to 1000, into a volume of broth containing the selective agent and grown with vigorous shaking (approximately 300 revolutions per minute) to late logarithmic/stationary phase (approximately 10 to 12 hours) to achieve a cell density of approximately 3 to 4×10^9 cells per ml. Cell density is estimated by taking an aliquot of the liquid culture and obtaining an OD₆₀₀ reading using a spectrophotometer, or by centrifuging the liquid culture and thereafter measuring the weight of the resulting bacterial pellet. Typically, 1.0 liter volume of an liquid culture of E. coli that is grown overnight at 37° C., 300 rpm with a cell density of approximately 3 to 4×10^9 cells per ml will correspond to a pellet weight of approximately 3 g/l. Depending on the desired amount of insert-bearing cosmid DNA that is required, a person skilled in the art would understand that either a liquid "mini-culture" of 2.0 to 5.0 ml or a liquid "maxi-culture" of 500 ml may be required to be grown to result in the desired amount of cosmid DNA to be isolated. [0197] Cosmid DNA, bearing the insert DNA of interest, is isolated from the bacteria grown in liquid cultures, as described in the preceding paragraph, using procedures that are known in the art. Non-limiting examples include the use of commercially available kits, for example the QIAGEN® Large-Construct Kit (QIAGEN Inc., Catalogue No. 12462) or Perfectprep® BAC 96 Kit (catalogue order number 955150431) available from Eppendorf North America (Westbury, N.Y.). Alternatively, the insert-bearing cosmid DNA is isolated by following procedures detailed for a traditional alkaline lysis method as described in Birnboim and Doly (1979) Nucleic Acids Research 7(6): 1513-1523, or in a cosmid-specific manual (e.g. the SuperCos™ 1 Cosmid Vector Kit Instruction Manual published online at www.stratagene.com). As an example of an alkaline lysis procedure, insert-bearing cosmid-containing bacterial cells from a 5.0 ml culture are collected by centrifugation (using an appropriate, sterile centrifuge tube) for 2 minutes followed by aspiration of the supernatant and resuspension of the pellet by vortexing in 200 µl of an ice cold solution of 50 mM glucose, 10 mM EDTA, 25 Mm Tris-HCl (pH 8.0). Following resuspension of the bacteria, 400 µl of a freshly prepared solution of 0.2 N NaOH, 1% SDS is added and the contents gently mixed by inversion (vortexing must be avoided), followed by incubation on ice for 5 minutes. Following incubation on ice, 300 ul of ice-cold potassium acetate (approximate pH 4.8) is added, and the tube gently inverted twice and incubated on ice for a further 5 minutes. The tube is then centrifuged for 5 minutes at 4° C. and 500 µl of the supernatant is transferred to a fresh (sterile) tube. The transferred supernatant is deproteinated by extraction with phenol-chloroform, keeping the upper phase to which is then added 1.0 ml of ethanol. The tube is left standing at room temperature for 5 minutes, and thereafter microfuged for 30 minutes, followed by aspiration of the liquid from the tube. The remaining DNA pellet is washed in 70% ethanol, centrifuged (in a microfuge), and after aspiration of the liquid and drying (avoiding complete dryness) of the pellet, the DNA is resuspended in 50 µl of Tris-EDTA (TE). DNA concentration is estimated by taking an OD_{600} reading on a 1/100 diluted aliquot of the purified insertbearing cosmid DNA. The insert-bearing cosmid DNA is thereafter used in any number of downstream applications that would be appreciated by a person skilled in the art.

[0198] Segments or regions of the insert DNA can be generated by performing a restriction digestion on the insertbearing cosmid DNA using protocols that are known to those of skill in the art. The segments or regions of the insert DNA may be of interest to the person of skill in the art as the particular nucleotide may be that for a gene(s) that is to be manipulated for a downstream application. As well, the segments or regions of the insert DNA may be of interest to the person of skill in the art as the particular nucleotide may be that for an entire biosynthetic locus, or a portion thereof, that encodes for the production of a natural product. It is possible that the nucleotide sequence of the insert DNA encodes one or more modules, which may be comprised of one or more domains, of a nonribosomal peptide synthetase or a polyketide synthase locus encodes for the production of a bioactive natural product.

[0199] As an example that is not intended to be limiting, if the sequence of the insert DNA is known, the presence of particular restriction enzyme sites within the insert DNA are determined and the region (i.e. the fragment) of DNA situated between two restriction enzyme sites cut or digested from the cosmid DNA. Generally, it is preferred in the art to use a restriction enzyme that recognizes a six base pair (bp) DNA recognition sequence as opposed to a four base pair recognition site, as there will be fewer restriction sites in a given stretch of DNA for six bp restriction enzyme, thereby offering less chance of digesting the cosmid (i.e. the vector) DNA per se. Selection of a given restriction enzyme may also be dependent upon whether the ends of the generated DNA fragment are to be blunt or are to possess overhangs so as to facilitate sub-cloning of the DNA fragment. Restriction digestion conditions are known to those skilled in the art. While not intending to be limiting, a digestion is usually performed using a minimum of 0.2 µg of DNA. If the DNA fragment to be generated is to be used as a probe, for example in Southern blotting, then an amount of DNA of at least 10 µg will be required for digestion. A restriction digestion can usually be performed in a range of reaction volume between 10 µl to 50 µl, using a requisite number of units of the given restriction endonuclease plus the particular buffer for the restriction enzyme and a necessary amount of sterile water to give the desired reaction volume. One unit of a restriction endonuclease will digest 1 µg of DNA in one hour, and it is common to use a ten-fold excess of the restriction enzyme to ensure complete digestion, provided that the volume of the restriction enzyme used does not exceed 10% of the final reaction volume. Upon addition of the restriction enzyme as the last component of the reaction mixture, the tube containing the mixture should be gently flicked with a finger to ensure proper mixing of the tube contents, followed by a brief centrifugation and incubation of the tube at 37° C., or at an elevated temperature 50-65° C. if the restriction enzyme is one isolated from a thermophilic bacteria, for a time span ranging from one to four hours. The reaction time may be extended beyond for greater lengths of time if it is desired. Reaction and deproteination may be accomplished by heat inactivating the restriction enzyme followed by phenol-chloroform extraction of the reaction (as described above), or by using a commercially available kit such as the MinElute® Reaction Cleanup Kit from QIAGEN.

[0200] Downstream uses of the insert DNA are discussed in Section V11 above and include: Labeling and use of the fragments as probes to detect the presence of the given gene or the expression of the given gene in a different organism; Use of the fragment in hybridization experiments; PCR amplification of the insert DNA or regions of interest of the insert DNA; Mutagenesis of the particular DNA segment of interest in order to produce substitutions, additions, deletions, fusions or truncations in the expressed polypeptide, which can be accomplished by random chemical mutagenesis, site directed mutagenesis, error-prone PCR, exonuclease II deletion, oligonucleotide mutagenesis for PCR; Generation of variant forms of the peptide of interest with conservative vs. non-conservative changes in the amino acid sequence to result in the production of novel end-product compounds; Cloning and use of the DNA sequence of interest in a heterologous expression system (yeast, mammalian, insect, plant expression vectors) for the production of the peptide of interest, and the creation of tagged (e.g. His, c-myc, Ni-tagged, etc.) fusion proteins; Use of the peptide that is produced to raise polyclonal or monoclonal antibodies (via the production of hybridomas). **[0201]** Antibodies (Ab's) are also used as probes to isolate interacting proteins—Ab's are generated against the peptides resulting from the heterologous expression of the DNA sequence of interest. Proteins that may potentially interact with that encoded by the DNA sequence of interest may also be identified by yeast two-hybird screening as described in U.S. Pat. No. 5,283,173.

[0202] All patents, patent applications, and published references cited herein are hereby incorporated by reference in their entirety. While this invention has been particularly shown and described with references to preferred embodiments thereof, it will be understood by those skilled in the art that various changes in form and details may be made therein without departing from the scope of the invention encompassed by the appended claims.

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Ile 865	Asn	Ser	Arg	Val	Glu 870	Leu		Arg	Cys	Phe 875		Ala	His	Glu	Ala 880

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Thr Arg Asp Ala Val Glu Leu Leu Arg Gly Ala Phe Glu Arg Ser Pro225230235240
Arg Thr Arg Asp Glu Phe Val Ser Arg Val Thr Ser Leu Thr Glu Ala 245 250 255
Ala Ala His Asp Leu Leu Gln Gly Arg Val Ala Asp Phe Gly Ala Arg 260 265 270
Leu Thr Glu Asn His Arg Leu Leu Arg Glu Val Gly Ile Ser Thr Glu 275 280 285
Arg Ile Asp Arg Met Val Asp Ala Ala Leu Ala Ala Gly Ser Pro Gly290295300
Ala Lys Ile Ser Gly Gly Gly Gly Leu Gly Gly Cys Met Ile Ala Leu Ala305310315320
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gtcgtgtacg gcgctccggc gctcgccgtc ccggtgccgc aactgaccgc cgtggccaag	180
gcgcggcggg ccggcggcga cggcggcgac gaggteteet tegecatege egggetggag	240
agcccggagg tgacgtcgct tccgaccgac ggcctgcaac atctggtgac ggagttccgg	300
cagegggeeg eegteacega geegatgege gtegaegtge tegtggaetg egeeateeeg	360
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gacgcgttcg accgccgcct cgacgccgcc acggtgttcg atctggtgca gacctcggag	480
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cgggaccgcc aggaatccgc ggcggtggtg cggagcgtcc agcaggccgg cgccgtccgc	1020
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Gly Ala Ala Asp Glu Val Val Leu Asp Gly Ser Pro Ala Asp Gly 65 70 75 80	
Glu Arg Arg Gln Arg Val Val Thr Phe Leu Asp Leu Val Arg Lys Leu 85 90 95	
Ala Gly Arg Thr Glu Arg Ala Cys Val Asp Thr Arg Asn Ser Val Pro 100 105 110	
Thr Gly Ala Gly Leu Ala Ser Ser Ala Ser Gly Phe Ala Ala Leu Ala 115 120 125	
Leu Ala Gly Ala Ala Ala Tyr Gly Leu Asp Leu Asp Thr Thr Ala Leu 130 135 140	

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Gly Gly Phe Ala Met Cys His Ala Gly Pro Gly Ala Gly Thr Ala Ala 165 170 175	
Asp Leu Gly Ser Tyr Ala Glu Pro Val Pro Val Ala Pro Leu Asp Val 180 185 190	
Ala Leu Val Ile Ala Ile Val Asp Ala Gly Pro Lys Ala Val Ser Ser 195 200 205	
Arg Glu Gly Met Arg Arg Thr Val Arg Thr Ser Pro Leu Tyr Gln Ser 210 215 220	
Trp Val Ala Ser Gly Arg Ala Asp Leu Ala Glu Met Arg Ala Ala Leu 225 230 235 240	
Leu Gln Gly Asp Leu Asp Ala Val Gly Glu Ile Ala Glu Arg Asn Ala 245 250 250 255	
Leu Gly Met His Ala Thr Met Leu Ala Ala Arg Pro Ala Val Arg Tyr	
260 265 270 Leu Ala Pro Val Thr Val Ala Val Leu Asp Ser Val Leu Arg Leu Arg	
275 280 285 Ala Asp Gly Val Ser Ala Tyr Ala Thr Met Asp Ala Gly Pro Asn Val	
290 295 300 Jys Val Leu Cys Arg Arg Ala Asp Ala Asp Arg Val Ala Asp Thr Leu	
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atecegtaeg cegacageet gtegatgaeg etegaegtet teeegaecae caecaecegte	180 240
ggategaea geggegegge ggeegaegag gtegteeteg aeggetegee egeegaegge	300
aacgggget gegtegaeae eegeaaetee gtgeeeaeeg gegeeggeet gegteeteg	360
gegageggat tegeegeeet egeeetegee ggegeegeeg egtaeggeet egaeeteggae	420
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geogggeoga aggoggtgte gageogogag gggatgogge gaacogteog gaeeteeeg	660
tetateagt egtgggtege etceggeege geegaeetgg eegagatgeg ggeegegetg	720
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ccaccatge tggeegeeeg geeggeggtg egetaeetgg egeeggteae tgtegeegtg	840
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Val	Ala	Val 35	Asp	Arg	Gly	Val	Asp 40	Val	Thr	Val	Ser	Gly 45	Ala	Asp	Ala	
His	Leu 50	Val	Val	Asp	Ser	Asp 55	Leu	Суз	Pro	Glu	Gln 60	Ala	Суз	Leu	Arg	
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Asp	Ala	Leu	Gly	Ala 85	Val	Val	Ser	Ala	Ile 90	Glu	Val	Val	Gly	Glu 95	Leu	
Leu	Thr	Gly	Arg 100	-	Leu	Arg	Pro	Leu 105	Pro	Met	Arg	Val	Ala 110	Ile	Thr	
Ser	Arg	Leu 115	His	Arg	Asp	Gly	Thr 120	Lys	Phe	Gly	Leu	Gly 125	Ser	Ser	Gly	
Ala	Val 130	Thr	Val	Ala	Thr	Val 135		Ala	Val	Ala	Ala 140	Tyr	His	Gly	Val	
Glu 145	Leu	Ser	Leu	Glu	Ser 150	-	Phe	Arg	Leu	Ala 155	Met	Leu	Ala	Thr	Val 160	
Arg	Asp	Gly	Ala	Asp 165		Ser	Gly	Gly	Asp 170	Leu	Ala	Ala	Ser	Val 175	Trp	
Gly	Gly	Trp	Ile 180	Ala	Tyr	Gln	Ala	Pro 185	Asp	Arg	Ala	Ala	Val 190	Arg	Glu	
Met	Ala	Arg 195	Arg	Arg	Gly	Val	Glu 200	Glu	Thr	Met	Arg	Ala 205	Pro	Trp	Pro	
Gly	Leu 210	Arg	Val	Arg	Arg	Leu 215	Pro	Pro	Pro	Arg	Gly 220	Leu	Ala	Leu	Glu	
Val 225	Gly	Trp	Thr	Gly	Glu 230		Ala	Ser	Ser	Ser 235	Ser	Leu	Thr	Gly	Arg 240	
Leu	Ala	Ala	Ser	Arg 245		Arg	Gly	Ser	Pro 250	Ala	Arg	Trp	Ser	Phe 255	Thr	
Ser	Arg	Ser	Gln 260	Glu	Суз	Val	Arg	Thr 265	Ala	Ile	Asp	Ala	Leu 270	Glu	Arg	
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Leu 305	Thr	Ala	Leu	СЛа	Asp 310		Ala	Glu	Thr	Val 315	Gly	Gly	Ala	Ala	Lys 320	
Pro	Ser	Gly	Ala	Gly	Gly	Gly	Asp	Суз	Gly	Ile	Ala	Leu	Leu	Asp	Ala	

325 330 335 Thr Ala Ala Thr Arg Thr Ala Arg Leu Arg Glu Gln Trp Ala Ala Ala 340 345 350 Gly Val Leu Pro Met Pro Ile Gln Val His Gln Thr Asn Gly Ser Ala 355 360 365 Arq <210> SEQ ID NO 15 <211> LENGTH: 1110 <212> TYPE: DNA <213> ORGANISM: Micromonospora sp. strain 046-EC011 <400> SEQUENCE: 15 gtgaccggcc cgggcgccgt gcgccgccac gcgccgggca agctgttcgt cgccggtgag 60 tacgcggtgc tggagccggg ccacccggcg ctgctggtgg cggtcgacag gggagtggac 120 gtcaccgtct ccggcgccga cgcccacctc gttgtcgact ccgacctctg cccggagcag 180 gcgtgcctgc ggtggcagga cggccggctc gtcggcgcgg gcgacgggca gccggcgccc 240 gacgccctcg gcgccgtggt ctcggcgatc gaggtggtcg gcgaactcct gaccggacga 300 gggctgcgcc cgctgcccat gcgggtggcg atcaccagcc ggctgcaccg cgacggcacg 360 aagtteggee tegggtegag eggggeggtg acagtegeea eggtgaeege agtggeegeg 420 taccacgggg tggagctgtc gctcgaatcg cggttccggc tggcgatgct ggcgacggtg 480 cgtgacggcg ccgacgcctc cggcggtgat ctggccgcga gcgtctgggg cggctggatc 540 gcctaccagg cgcccgaccg cgcggccgtg cgcgagatgg cgcggcggcg cggcgtcgag 600 660 gagacgatge gegegeeetg geegggeetg egggteegge ggetgeeaee acegegtgge 720 ctcgcgctgg aggtgggctg gaccggcgag ccggcgagca gcagctcgtt gaccgggcgg 780 ctggccgcct cccggtggcg gggcagcccg gcgcggtgga gcttcaccag ccgtagccag 840 caggtccggc gggcccggca cgtgcttgcc gagctggacg acgaggtccg gctcgggatc 900 ttcacccccc ggctgacggc gctgtgcgac gccgccgaga ccgtcggcgg cgcggccaaa 960 ccgtccggcg ccggtggcgg ggactgcggc atcgcgttgc tggacgccac cgccgcgacg 1020 cggaccgcgc ggctgcgcga gcagtgggcc gccgccgggg tgctccccat gccgatccag 1080 gtccatcaga cgaacgggag cgcgcgatga 1110 <210> SEQ ID NO 16 <211> LENGTH: 360 <212> TYPE: PRT <213> ORGANISM: Micromonospora sp. strain 046-ECO11 <400> SEQUENCE: 16 Met Ile Ala Asn Arg Lys Asp Asp His Val Arg Leu Ala Ala Glu Gln 1 5 10 15 Gln Gly Arg Leu Gly Gly His His Glu Phe Asp Asp Val Ser Phe Val 20 25 30 His His Ala Leu Ala Gly Ile Asp Arg Ser Asp Val Ser Leu Ala Thr 40 45 35 Ser Phe Gly Gly Ile Asp Trp Pro Val Pro Leu Cys Ile Asn Ala Met 50 55 60

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Ala Ala Arg Glu Thr Gly Val Pro Ile Ala Thr Gly Ser Met Ser Ala 85 90 95									
Tyr Phe Ala Asp Glu Ser Val Ala Glu Ser Phe Ser Val Met Arg Arg 100 105 110									
Glu Asn Pro Asp Gly Phe Ile Met Ala Asn Val Asn Ala Thr Ala Ser 115 120 125									
Val Glu Arg Ala Arg Ala Val Asp Leu Met Arg Ala Asp Ala Leu 130 135 140									
Gln Ile His Leu Asn Thr Ile Gln Glu Thr Val Met Pro Glu Gly Asp 145 150 155 160									
Arg Ser Phe Ala Ala Trp Gly Pro Arg Ile Glu Gln Ile Val Ala Gly 165 170 175									
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Glu Thr Leu Arg Leu Arg Asp Met Gly Val Arg Val Ala Asp Val 195 200 205									
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Ile Glu Ala Leu Met Thr Ala Leu Gly Ala Arg Thr Pro Ala Asp Leu 305 310 315 320									
Thr Arg Cys Asp Leu Leu Ile Gln Gly Arg Leu Ser Ala Phe Cys Ala 325 330 335									
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atcaacgega tgaeeggegg eageaceaag aceggeetga teaaceggga eetggegate 240									
geggeeeggg agaeeggegt acegategee acegggtega tgagegeeta ettegeegae 300									
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Pro	Ala 50	Pro	Asp	Glu	Asp	Ile 55	Val	Thr	Met	Ala	Ala 60	Ala	Ala	Ala	Ala
Pro 65	Val	Val	Ala	Arg	His 70	Gly	Thr	Asp	Arg	Ile 75	Arg	Thr	Val	Val	Phe 80
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Asp	Pro	Ser	Gly 180	Met	Phe	Thr	Ala	Asp 185	Val	Met	Aab	Phe	Trp 190	Arg	Pro
Asn	Tyr	Arg 195	Thr	Thr	Ala	Leu	Val 200	Asp	Gly	His	Glu	Ser 205	Ile	Ser	Ala
Tyr	Leu 210	Gln	Ala	Leu	Glu	Gly 215	Ser	Trp	Lys	Asp	Tyr 220	Thr	Glu	Arg	Gly
Gly 225	Arg	Thr	Leu	Asp	Glu 230	Phe	Gly	Ala	Phe	Сув 235	Tyr	His	Gln	Pro	Phe 240
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	-	275				Gly	280		-			285		-	
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305	-				310	Gly		-		315					320
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Arg	Ala	Ala	Ile 340	Asp	Arg	Arg	Gln	Glu 345	Ile	Asp	Tyr	Ala	Thr 350	Tyr	Arg
		355				Phe	360					365			
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Lys Lys Ile Tyr Ala Ala Phe Ala Pro Asp Glu Leu Gln Val Ala Thr 120 115 125 Ser Leu Ala Gly Ile Pro Ala Met Pro Arg Ser Leu Ala Ala Asn Ala 130 135 140 Asp Phe Phe Thr Arg His Gly Leu Asp Asp Arg Val Gly Val Leu Gly 145 150 155 160 Phe Asp Tyr Pro Ala Arg Thr Val Asn Val Tyr Phe Asn Asp Val Pro 165 170 175 Arg Glu Cys Phe Glu Pro Glu Thr Ile Arg Ser Thr Leu Arg Arg Thr 185 180 190 Gly Met Ala Glu Pro Ser Glu Gln Met Leu Arg Leu Gly Thr Gly Ala 200 195 2.05 Phe Gly Leu Tyr Val Thr Leu Gly Trp Asp Ser Pro Glu Ile Glu Arg 215 210 220 Ile Cys Tyr Ala Ala Ala Thr Thr Asp Leu Thr Thr Leu Pro Val Pro 235 225 230 240 Val Glu Pro Glu Ile Glu Lys Phe Val Lys Ser Val Pro Tyr Gly Gly 245 250 255 Gly Asp Arg Lys Phe Val Tyr Gly Val Ala Leu Thr Pro Lys Gly Glu 260 265 Tyr Tyr Lys Leu Glu Ser His Tyr Lys Trp Lys Pro Gly Ala Val Asn 275 280 285 Phe Ile 290 <210> SEQ ID NO 23 <211> LENGTH: 873 <212> TYPE: DNA <213> ORGANISM: Micromonospora sp. strain 046-EC011 <400> SEOUENCE: 23 gtqgccqaqc tctactcqac catcqaqqaa tcqqcccqqc aactqqacqt gccqtqttcq 60 egegaceggg tetggeceat cetgteegeg taeggegaeg egttegecea teeegaggeg 120 gtggtegeet teegggtgge gaeegegetg egteaegegg gegagetgga etgeeggtte 180 cggacgcatc cggacgaccg ggacccgtac gcctcggcgc tcgcccgggg cctcaccccg 240 cgcacggacc accccgtcgg cgcgctgctc tccgaggtcc accggcgctg cccggtggag 300 agccacggca tcgacttcgg ggtggtcggc ggcttcaaga agatctacgc ggccttcgcc 360 ccggacgagc tgcaggtggc cacgtcgctc gccggcattc cggcgatgcc ccgcagcctc 420 gccgcgaacg ccgacttctt cacccggcac ggcctcgacg accgggtcgg cgtgctggga 480 ttcgactacc cggcccggac cgtgaacgtc tacttcaacg acgtgccgcg tgagtgcttc 540 gageeggaga ceateeggte gaegetgege eggaeeggga tggeegagee gagegageag 600 atgeteegge teggeacegg ggegtteggg etetacgtea egetgggetg ggaeteeeeg 660 gagategage ggatetgeta egeegeggeg accaeggace tgaceaeget teeggtacee 720 gtggaaccgg agatcgagaa gttcgtgaaa agcgttccgt acggcggcgg ggaccggaag 780 ttcgtctacg gcgtggcgct gacccccaag ggggagtact acaaactcga gtcgcactac 840 873 aaatqqaaqc cqqqcqcqqt qaacttcatt tqa

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COILC	TITO	cu

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Leu	Gly	Leu 35	Leu	Gly	Phe	Ala	Leu 40	Val	Ala	Ala	Ser	Gly 45	Leu	Ala	Leu
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Arg	Glu	Glu	Thr	Ala 165	Arg	Leu	Arg	Ala	Thr 170	Gln	Glu	Arg	Leu	His 175	Ile
Ala	Arg	Glu	Leu 180	His	Asp	Ser	Leu	Thr 185	His	Gln	Ile	Ser	Ile 190	Ile	Lys
Val	Gln	Ala 195	Glu	Val	Ala	Val	His 200	Leu	Ala	Arg	ГЛа	Arg 205	Gly	Glu	Gln
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Arg 225	Glu	Leu	Arg	Ala	Thr 230	Leu	Glu	Thr	Leu	Arg 235	Asp	Leu	Thr	Гла	Ser 240
Pro	Ser	His	Gly	Leu 245	Asp	His	Leu	Pro	Glu 250	Leu	Leu	Ala	Gly	Ala 255	Glu
Lys	Ile	Gly	Leu 260	Ala	Thr	Thr	Leu	Thr 265	Ile	Glu	Gly	Asp	Gln 270	Arg	Asp
Val	Pro	Glu 275	Ala	Val	Gly	Arg	Thr 280	Ala	Tyr	Arg	Ile	Val 285	Gln	Glu	Ser
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Ile 305	Asp	Tyr	Arg	Pro	Asp 310	Ala	Leu	Ser	Ile	Arg 315	Ile	Asp	Asp	Asp	Gly 320
Thr	Ala	Arg	Pro	Gly 325	Ala	Ala	Pro	Val	Pro 330	Gly	Val	Gly	Leu	Leu 335	Gly
Met	His	Glu	Arg 340	Val	Leu	Ala	Leu	Gly 345	Gly	Arg	Leu	Arg	Ala 350	Glu	Pro
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Val Pro 370

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90

Arg Ala Gly Ala Thr Gly Phe Leu Val Lys Asp Ile Glu Pro Asp Asp

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Pro Ser Ile Thr Arg Met Leu Ile Asn Arg Tyr Val Ser Glu Pro Leu 130 135 140	
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gegegeggtg acgegetget egegeegteg ateaceegga tgetgateaa caggtaegtg	420
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gaggeggteg ceetggeege eeggggeetg teeaaegaeg agategeega tegeatggtg	540
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Ala Leu Leu Gly Leu Val Leu Gly Gly Trp Ala Leu Arg Ser Ala Gly 50 55 60	
Arg Gly Gly Gly Arg Gly Asn Ala Ile Ala Ala Leu Val Leu Ala Val	

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1245 250 12 251 12 251 12 251 12 251 12 251 12 251 12 251 12 251 12 251
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275 280 285 Leu Arg Asn Asp 11e Glu Asp Gly Ala Frage Ass Asp Gly Gly Ala Asp Gly Ala Pro Gly Glu Asp Gly Ala Pro Gly Glu Pro Gly Ala Pro Gly Glu Pro Gly Ala Pro Glu Ala Pro Ala Asp Pro Glu Ala Pro Ala Pro Ala Pro Ala Pro Ala Pro Ala Pro
290 295 300 Tyr Asp Val Pro Cys Gly Glu Asp Gly Ala Pro Gly Ala Tr Gly Ala Tr Gly Ala Pro Gly Ala Tr
310 315 324 Ala Val Gln Pro Gly Gly Gly Glu Phe Val Tyr Arg Phe Gln Ala Arg 325 Ala Val Gln Pro Tyr Tyr Tyr His Thr His Gln Ala Ser His Pro Al. 345 Val Gly Thr Tyr Tyr Gly Thr Leu Val Val Thr Pro Arg Glu Ala Gly Gly Leu Tyr Gly Thr Leu Val Val Thr Pro Arg Glu Ala Glu Arg Gly Leu Arg Leu Arg Leu Thr Leu Pro Val His Thr 380 Arg Lys Gly Lau Tyr Gly Thr Leu Arg Leu Thr Leu Pro Val His Thr 380 Arg Arg Val Thr Ile Leu Gly Arg Gln Glu Gly Arg Ala Val Hir 395 Arg Pro Gly Gln Pro Val Arg Leu Arg Leu Ile Arg Thr Arg 415 Arg Pro His Trp Phe Ala Val Val Gly Ser Pro Phe Arg Val Val 425 Val Arg Gly Arg Arg Arg Leu Arg Gln Gly Gly Arg Ala Yal Hir 430 Val Arg Pro Gly Arg Arg Leu Arg Gln Gln Glu Gly Arg Ala Val 445 Val Arg Pro His Trp Phe Ala Val Val 425 Gly Ser Pro Phe Arg Val Val 445 Val Arg Leu Pro Ala Gly Gly Arg Tyr Arg Leu Thr Leu Ala Met 440 435 Thr Leu Leu Leu Arg Arg Arg Pro Gln Glu 445 Arg Pro Gly Val Thr Leu Leu Leu Arg Arg Arg Pro 40 Arg Pro Yal Pro Pro Glu Phe Arg Arg Arg His Phe Thr 10 500 Fro Sta Ser Ser Arg Gln Glu 747 411 Arg Pro Pro Glu Arg Arg Arg Arg Arg Arg His Pro 416 420 Thr Ala Arg Pro Glu Phe Arg Arg Arg His Phe Thr 10 510 Fro Arg Arg Arg Arg Pro Ser Val Pro Arg Ser Leu Gly 747 510 Fro Pro Arg Ara Arg Pro Ser V
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355 360 365 Pro Glu Ala Glu Arg Gly Gly Gly Asp Gln Glu Thr Leu Pro Val His Thr 370 375 Leu Asp Leu Thr Leu Pro Val His Thr 380 Asp Asp Val Thr Ile Leu Gly Asp Gln Glu Gly Arg Ala Val His 390 Asp Gly Gln Pro Val Arg Leu Arg Leu Ile Asn Thr Asp 405 Ang Leu Arg Leu Ile Asn Thr Asp 415 Asn Pro His Trp Phe Ala Val Val 425 Gly Ser Pro Phe Arg Val Val 420 Thr Phe Ala Val Val Gly Ser Pro Phe Arg Val Val 435 Val Asp Gly Arg Asp Leu Asn Gln Pro Gly Glu Val Arg Glu Val 435 Asp Gly Arg Asp Leu Asn Gln Pro Gly Glu Val Arg Glu Val 445 Leu Arg Leu Pro Ala Gly Gly Arg Tyr Asp Leu Thr Leu Ala Mer 450 Asp Ala Lys Val Thr Leu Leu Leu Asp Asn Asp Ser Asp Gln Gly 475 Asp Ala Lys Val Thr Pro Gly Val Gly Gly Gly Asp Arg Pro Leu Pro 485 Fro Gly Val Gly Gly Gly Asp Arg Pro Leu Pro 495 Leu Leu Arg Pro Pro Gly Val Gly Gly Gly Asp Arg Pro Leu Pro 485 Fro Son Thr Ala Asp Trp Pro Glu Phe Asp Leu Leu Gly Tyr Gly Glu Pro 500 Fro Son Pro Val Pro Phe Asp Ala Asp Asp Asp Ala Asp Arg His Pro Ala Tyr Al 500 Fro Val Asp Gly Arg Ala Leu Ala Mer Val Asp Cly Lys Pro Ala Tyr Al 550 Glu Gly Asp Yal Val Arg Phe Thr Val Asp Arg Pro Asp Gln Leu Val 550 Glu Gly Asp Yal Val Arg Phe Thr Val Asp Arg Arg Pro Leu Fro 560 Glu Gly Asp Pro Tyr Ser Gly Ser Pro Leu Trp Met Asp Arg Pro 500 Fro Trp His Leu His Gly His Pro Leu Trp Met Asp Asp Asp Pro 605
370 375 380 Asp Asp Val Thr 11e Leu Gly Asp Gln Glu Gly Arg Ala Val Hing Yal Arg Pro Gly Gln Pro Val Arg Leu Arg Leu Arg Leu Ile Asn Thr Asg Asn Pro His Trp Phe Ala Val Val Gly Ser Pro Phe Arg Val Val Asp Asp Cly Arg Arg Asp Leu Asn Gln Pro Cly Glu Val Arg Glu Val Arg Glu Val Thr Leu Asn Gln Pro Cly Glu Val Arg Glu Val Asp Ala Lys Val Thr Leu Leu Leu Asp Asn Asp Ser Asp Gln Glu Val Asp Ala Lys Val Thr Leu Leu Leu Leu Asp Asn Asp Ser Asp Gln Glu Val Asp Ala Lys Val Thr Ser Pro Glu Pro Glu Pro Gly Glu Asp Arg Pro Leu Pro 485 Pro Glu Pro 7400 Asp Asp Arg Pro 7500 Fre Val Pro Pro Son Trp Pro Glu Pro Son Pro Son Pro Son Pro 515 Pro Ala Try Asp Asp Arg Pro 7510 Pro 100 Fre Val Asp Arg Ala Leu Asp Asp Asp Arg Pro 500 Pro 495 Pro 495 Thr Ala Asp Trp Pro Pro Slu Pro 500 Pro 500 Pro 7510 Pro 710 Son Trp Pro 510 Pro 710 Pro 710 Pro 710 Pro 710 Son Trp Pro 710 Pro 710 Pro 710 Pro 710 Pro 710 Pro 710 Son Trp Pro 710 Son 710 Pro 710 Pro 710 Pro 710 Pro 71
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Arg Pro Gly Glu Val Trp Glu Val Ala Phe Arg Ala Asp Asn Pro 610 615 Val Trp Met Asn His Gys His Asn Leu Pro Pis Gln Glu Gln Gl
Val Trp Met Asn His Cys His Asn Leu Pro His Gln Glu Gln Gl 630 635 640 Met Leu Arg Leu Val Tyr Asp Gly Val Thr Thr Pro Phe Ala Se:
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Leu Val Gly Val Leu Ala Thr Leu Ala Gly Ala Val Ala Ala Glu Ala 55 60 50 Phe Thr Leu Ala Ala Arg Gly Phe Gly Val Pro Met Glu Ala Ala Gly 65 70 75 80 Val Trp Glu Glu Gln Ala Gln Ala Ile Pro Val Gly Ala Ile Ala Arg 85 90 95 Ser Val Val Leu Trp Ser Ile Gly Gly Ile Val Leu Ala Val Val Val 100 105 110 Ala Arg Arg Ala Arg Arg Pro Val Arg Ala Phe Val Ala Gly Thr Val 115 120 125 Ala Phe Thr Val Leu Ser Leu Ala Ala Pro Ala Phe Ala Arg Asp Thr 135 130 140 Pro Val Ser Thr Gln Leu Val Leu Ala Gly Thr His Val Ile Ala Gly 150 145 155 160 Ala Val Ile Ile Ser Ile Leu Ala Ala Arg Leu Ala Ala Pro Thr Pro 170 165 175 Pro Arg <210> SEQ ID NO 37 <211> LENGTH: 537 <212> TYPE: DNA <213> ORGANISM: Micromonospora sp. strain 046-EC011 <400> SEQUENCE: 37 atgttgactg ccgtcgtggc gtccccgcat tctcccgaga acacatcgag gcacccgacc 60 ggaggcgacg ccgtggatga ggccactccc cgaactcccg tcgcggcacg gcccacctgg 120 180 tegeeggeea cegeteeggt gtggetggte ggegtgetgg ceacectege eggggeegtg geogeggagg egtteacget egeogeoegg ggetteggeg tacegatgga ggeggeogge 240 gtctgggagg agcaggcgca ggcgatcccg gtggggggcca tcgcccgcag cgtcgtgctc 300 tggtcgatcg gcggaatcgt cctggcggtg gtcgtggcgc ggcgggcccg gcggcccgtg 360 cgtgccttcg tggccggcac cgtcgcgttc accgtgctgt ccctcgccgc gcccgccttc 420 gecegggaca ceceggtgte gaegeagete gteetegeeg geacecaegt gategeegge 480 gccgtgatca tctccatcct ggccgcgcgg ctcgccgcgc ccaccccgcc ccggtaa 537 <210> SEQ ID NO 38 <211> LENGTH: 661 <212> TYPE: PRT <213> ORGANISM: Micromonospora sp. strain 046-ECO11 <400> SEOUENCE: 38 Met Asp Gly Thr Glu Ser Asn Val Thr Gly Phe Pro Asp Leu Leu Ser 1 5 10 15 Gly Leu Gly Gly Asp Gly Arg Ala Phe Ala Leu Leu His Arg Pro Gly 20 25 30 Ala Ala Gly Cys Ala Tyr Val Glu Val Leu Thr Gly Glu Val Cys Asp 35 40 45 Val Asp Thr Leu Gly Glu Leu Pro Leu Pro Thr Glu Pro Ala Thr Gly 50 55 60 Ala Arg His Asp Leu Leu Val Ala Val Pro Tyr Arg Gln Val Thr Glu 65 70 75 80

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Pro	Asp	Gly	Pro	Asp 405	Gly	Gly	Arg	Thr	Thr 410	Ala	Ala	Arg	Ala	Arg 415	Ser
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Pro Tyr Glu Ser Glu Phe Arg Met Gly Phe Thr Ile Arg Arg Ser Gln 305 310 315 320						
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Arg Leu Ser Val Ile Val Ser Arg Asp Arg Thr Val Asp Ala Val Glu 385 390 395 400						
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Glu Gln Pro Gly Leu Met Ala Ala Ala Val Ala Arg Leu Val His Glu 255 245 250 Pro Arg His Val Pro Asp Arg Tyr Asp Val Asp Asp Arg Leu Ala Leu 260 265 270 Thr Ser <210> SEQ ID NO 43 <211> LENGTH: 825 <212> TYPE: DNA <213> ORGANISM: Micromonospora sp. strain 046-ECO11 <400> SEQUENCE: 43 gtggccgtac tcaacgcttc gttcgctcgt ggcctgcgtc tgcgccgact gttccgacgc 60 ggcgacggac gcctgctcgt cgtcccgctc gaccactccg tcaccgacgg gccgctgcgc 120 cgcggcgacc tgaactcgct gctcggtgag ctcgccggca ccggcgtgga cgccgtggtg 180 ctgcacaagg gcagcctgcg gcacgtcgac cacggctggt tcggcgacat gtcgctgatc 240 gtgcatctga gcgtgagcac ccggcacgcc ccggacccgg acgcgaagta cctggtcgcg 300 cacgtggagg aggcgctgcg gctgggcgcc gacgcggtca gcgtgcacgt caacctcggc 360 tcaccgcagg aggcgcggca gatcgccgac ctggcggcgg tggcgggggg gtgcgaccgc 420 480 tqqaacqtcc cqctqctqqc catqqtqtac qcccqcqqqc cqcaqatcac cqactcccqq 540 gcaccqqaqc tqqtqqcqca cqccqcqacq ctcqccqcqq acctcqqcqc cqacatcqtc 600 aagaccgact acgtgggcac gcccgagcag atggccgagg tggtgcgcgg ctgcccgatc cogotgateg tggccggcgg coogocteg googacacte cgacggtget egectacgte 660 720 teggaegege tgegeggegg egtggeeggg atggeeatgg geegeaacgt gtteeaggee gagcagcccg gcctgatggc cgccgccgtg gcacggctgg tgcacgagcc acggcacgtg 780 ccggaccggt acgacgtcga cgaccggctc gcccttacgt cctga 825 <210> SEO ID NO 44 <211> LENGTH: 367 <212> TYPE: PRT <213> ORGANISM: Micromonospora sp. strain 046-EC011 <400> SEOUENCE: 44 Val Lys Leu Cys Trp Leu Asp Ile Arg Asn Val Asn Gly Ala Lys Glu 1 5 10 15 Ala Ile Val Glu Glu Ala Val His Gln Arg Val Asp Ala Val Ala 20 25 30 Ala Asp Pro Ala Asp Leu Glu Thr Leu Pro Pro Thr Val Lys Lys Val 35 40 45 Leu Phe Pro Gln Gly Gly Pro Leu Pro Glu Lys Leu Glu Pro Ala Asp 50 55 60 Leu Val Ile Val Glu Pro Ala Arg His Gly Glu Pro Ala Glu Leu Ala 65 70 75 Ala Arg Tyr Pro Glu Val Glu Phe Gly Arg Phe Val Glu Ile Val Asp 90 85 Ala Asp Ser Leu Glu Asp Ala Cys Arg Ser Ala Arg His Asp Arg Trp 100 105 110 Ser Leu Leu Tyr Phe Arg Asp Pro Thr Lys Ile Pro Leu Glu Ile Val 120 115 125

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Asp	Gly	Val	Met	Leu 165	Ala	Pro	Arg	Ala	Val 170	Gly	Glu	Ala	Thr	Glu 175	Leu	
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His 225	Ser	Thr	Gly	Met	Ile 230	Leu	Сув	Сув	Ser	Glu 235	Thr	His	Pro	Leu	Pro 240	
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81

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Ala Arg	Ala 35	Val	Val	Суз	Ala	Gly 40	Gly	Thr	Ile	Asp	Leu 45	Asp	Arg	Leu		
Ser Arg 50	Gln	Leu	Val	Thr	Gly 55	Gly	His	Val	Glu	Thr 60	Asp	Ala	Asp	Asn		
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Phe Phe	Phe	Ala	Asn 85	Pro	His	Arg	Pro	Ala 90	Glu	Ala	Tyr	Arg	Ala 95	Arg		
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Ala Leu	Thr 115	Asp	Pro	Arg	Val	Ala 120	Asp	Leu	Val	Ala	Val 125	Ala	Ala	Glu		
Phe Asp 130		Pro	Val	Tyr	Val 135	Val	Сув	Leu	Asp	Arg 140	Pro	Gly	Ala	Gly		
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Leu Ile	Gln	Asp 180	Glu	Pro	Asn	Ile	Ser 185	Leu	Glu	Thr	Ser	Gly 190	Gly	Tyr		
Thr Cys	Val 195		Glu	Ala	Ala	Leu 200	Arg	Arg	Leu	Gly	Asp 205	Asp	Arg	Val		
Val Phe 210		Ser	Glu	Tyr	Pro 215	Leu	Gln	His	Pro	Ala 220	Val	Glu	Leu	Ala		
Lys Phe 225	Gln	Ala	Leu	Arg 230	Leu	Pro	Pro	Glu	Arg 235		Arg	Arg	Ile	Ala 240		
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ttettetteg ccaaccegea eeggeeggee gaggegtaee gggeeegege egeegagtte	300
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Ala Pro Pro Val Thr Pro Ala Asp Leu Ala Asp Leu Pro Leu Thr Thr 50 55 60	
Lys Gln Asp Leu Arg Asp Asn Tyr Pro Phe Gly Met Leu Ala Val Pro 65 70 75 80	
Arg Glu Arg Leu Ala Thr Tyr His Glu Ser Ser Gly Thr Ala Gly Lys 85 90 95	
Pro Thr Pro Ser Tyr Tyr Thr Ala Glu Asp Trp Thr Asp Leu Ala Glu 100 105 110	
Arg Phe Ala Arg Lys Trp Ile Gly Met Ser Ala Asp Asp Val Phe Leu 115 120 125	
Val Arg Thr Pro Tyr Ala Leu Leu Leu Thr Gly His Leu Ala His Ala 130 135 140	
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Thr	Glu	Thr	Gly	Ser 245	Leu	Ala	Gly	Glu	Cys 250	Pro	Glu	Gly	Arg	Leu 255	His	
Leu	Trp	Ala	Asp 260	Arg	Ala	Leu	Phe	Glu 265	Val	Tyr	Asp	Pro	Asp 270	Thr	Gly	
Ala	Val	Arg 275	Ala	Asp	Gly	Asp	Gly 280	Gln	Leu	Val	Val	Thr 285	Pro	Leu	Phe	
Arg	Glu 290	Ala	Met	Pro	Leu	Leu 295	Arg	Tyr	Asn	Leu	Glu 300	Asp	Asn	Val	Ser	
Val 305	Ser	Tyr	Asp	Asp	Cys 310	Gly	Сув	Gly	Trp	Lys 315	Leu	Pro	Thr	Val	Arg 320	
Val	Leu	Gly	Arg	Ser 325	Ala	Phe	Gly	Tyr	Arg 330	Val	Gly	Gly	Thr	Thr 335	Ile	
Thr	Gln	His	Gln 340	Leu	Glu	Glu	Leu	Val 345	Phe	Ser	Leu	Pro	Glu 350	Ala	His	
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Glu	Ile 370	Glu	Val	Ala	Ala	Ala 375	His	Arg	Val	Ala	Ala 380	Glu	Ala	Glu	Leu	
Thr 385	Ala	Ala	Ile	Arg	Ala 390	Ala	Phe	Gly	Val	Asp 395	Ser	Glu	Val	Thr	Gly 400	
Leu	Ala	Pro	Gly	Thr 405	Leu	Ile	Pro	Leu	Asp 410	Ala	Leu	Thr	Ser	Met 415	Pro	
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cccg	Jacat	ccg a	actto	cccg	gc go	ctgco	gegee	g ctę	gttco	gtcg	gcg	gega	gaa (gatga	accgac	660

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Gly Asp Ala Ala His P 290	Pro Met Leu Pro Phe Gln Ala Gln Gly Ala Asn 295 300
-	Ala Val Val Leu Ala Cys Leu Ala Gly Val 310 315 320
Glu Pro Ala Gly Leu G 325	Gly Ala Ala Leu Arg Arg Tyr Glu Arg Ile Arg 330 335
Leu Pro Arg Thr Thr A 340	Arg Ile Gln Arg Gln Ser Arg Ala Asn Ala Glu 345 350
Met Phe His Leu Ala A 355	Asp Gly Ala Asp Gln Arg Arg Asp Val Ala 360 365
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Pro Gly Met Ser Se 100	er Ala Glu Asp Asp Arg Gly 105	Ile Val Asp Asp Val 110	
Ala Pro Gln Pro Gl 115	y Asp Thr Val Leu Thr Lys 120	Trp Arg Tyr Ser Ala 125	
Phe Phe Arg Ser As 130	mp Leu Glu Glu Arg Leu Arg 135	Gly Ala Gly Arg Asp 140	
Gln Leu Val Val Cy 145	rs Gly Val Tyr Ala His Met 150 155		
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Thr Tyr Arg Ala Pro Thr Ser Glu Val Glu Phe His Val Asp Gly Arg 180 185 190 Leu Ala Tyr Trp Ala Arg Val Pro Val Pro Val Thr Gly Phe His Ala 195 200 205 Gly Met Ala Leu Phe Ser Ala Arg Asp Leu Ala Arg Tyr Pro Arg Glu 210 215 220 Gln Arg Glu His Gly Gln Gly Ala Thr Gly Trp Trp Gly Pro Trp Arg 230 225 235 240 Ile Ala Ser Gly Val Arg 245 <210> SEQ ID NO 59 <211> LENGTH: 741 <212> TYPE: DNA <213> ORGANISM: Micromonospora sp. strain 046-EC011 <400> SEQUENCE: 59 atgacgtcgg cactgagaac cagegegtgg acgtacgaeg acttcaccag cegegagetg 60 gaccccgccc gctgggcgat catgtcgatc gccggcggg acgggcagac ccacaggtac 120 caggaccgca acgcccaggt ccgcaccggc gacgggcggc tggagctgac cgtcgacccg 180 ttcacccgct tccacgacac cgatccccgg cagaacaacg ccaagcagat gtaccggtcg 240 gtgcggcgct tcgccgtgcc ggcggagggc tcgctgaccg tcgaggtgga gatgggcgtg 300 cqqacqtacc qqcaqatccc qcacqacctq ctqqacqcqt tcqqcacqqt qaacctqttc 360 420 gacetggaga eeggegtegt gtteaaegee geegeeaega aegaeaeegt gtaegegaeg gtcgagcgcc tggtgctgcc cggcgtgacc cagccgcacg agcactacat ccaccgggtg 480 gteetggaeg tgeegaegga geegggeegg gegeaeggat aegeeateae etaeegggeg 540 600 ccgacgtcgg aggtggagtt ccacgtcgac ggccggctcg cctactgggc gcgggtcccg gtgccggtga ccggattcca cgccggcatg gcgctcttct ccgcccgcga cctggcccgg 660 720 tacccccgcg agcagcggga gcacgggcag ggcgcgaccg ggtggtgggg gccgtggcgg atcgcctccg gcgtcagatg a 741 <210> SEQ ID NO 60 <211> LENGTH: 111 <212> TYPE: PRT <213> ORGANISM: Micromonospora sp. strain 046-ECO11 <400> SEQUENCE: 60 Met Asp Thr Ala Ala Pro Ala Thr Asp Gly Gly Arg Tyr Leu Ala Val 10 1 5 15 His His Ser Ala Glu Phe Arg Glu Leu Arg Arg Arg Ser Ser Thr Phe 20 25 30 Thr Leu Trp Ala Ser Val Ala Phe Phe Gly Trp Trp Phe Leu Gly Ser 40 35 45 Leu Leu Ala Thr Tyr Ala Pro Asp Phe Phe Arg Glu Lys Val Ala Gly 55 60 Pro Val Asn Val Gly Leu Leu Phe Val Phe Leu Ser Phe Ala Phe Val 65 70 75 Val Thr Leu Ala Ala Phe Tyr Leu Arg Tyr Ala Arg Thr His Leu Asp 90 95 85 Pro Leu Ser Glu Lys Ile Arg Ala Asp Leu Glu Gly Ala Ser Arg

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gcc	ggcto	ccg t	Egca	gacco	cg gt	tgggi	tggco	c cc	cgago	cagg	ggc	acgc	ccg g	gcggi	tacta	ac 240
gcg.	atcad	ccg a	acca	3333 (cd dá	gegea	accto	g cgé	ggtgt	ttcg	cgg	cggt	gtg 9	gcag	gagat	te 300
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	0> SE															
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Ile	Ala	Val	Cys 100	Val	Ser	Phe	Leu	Leu 105	Pro	Ser	Ala	Val	Pro 110	Val	Glu	
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Ala	Ala	Val	Leu	Thr 165	Met	Phe	Ala	Сув	Ala 170	Ala	Gly	Glu	Thr	Gly 175	Leu	
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n Thr Phe Asp Leu His Thr Pro His Clu Ala Pro Ala Ala Clu The Glu Asp Pro Pro Ala Ala Clu Pro Pro Ala Ala Ala Pro Pro Ala Ala Ala App Pro Ala Pro Ala Pro Ala Pro												-	con	tin	ued	
115 120 125 a Gly Arg Leu Phe Gly Cys Phe Ala Met Thr Glu Thr Gly His Gly 135 Thr Gly Val Thr Gly Gly Arg Met Thr Gly Val Tyr Asp Pro Gln Thr 160 r Asp Val Gln Gln Leu Arg Thr Pro His Glu Ala Ala Arg Lys Asp Tyr 165 Thr Gly Cly Asp Tyr 175 e Gly Asn Ala Ala Arg Asp Gly Arg Met Ala Val Val Phe Ala Gln 195 Thr Gly Gly Arg Arg His Gly Val His Ala Thr Leu Val Pro 205 e Arg Asp Glu His Gly Lys Pro Met Pro Gly Val Thr Ile Gly Asp 215 Thr Ala Ala Arg Asp Clu Val His Ala Thr Leu Val Pro 205 e Arg Asp Glu His Gly Lys Pro Met Pro Gly Val Thr Ile Gly Asp 215 Thr Ala Arg Asp Clu Var Arg Clu Var Asp Asn Gly Arg Leu Ser 235 a Gly Pro Lys Ala Cly Leu Clu Var 225 Thr Ala Ala Arg Asp Clu Var Arg Clu Asp Asn Glu Asp Ser 275 a Gly Pro Lys Ala Cly Leu Clu Thr Leu Val Arg Gly Arg Val Ser 265 a Asp His Val Arg Val Pro Arg Glu Met Leu Lau Asp Gli Arg Val Ser 275 g Arg Phe Pe Thr Met Leu Cly Thr Leu Val Arg Gly Asp 285 104 Ala Glu Asp Gli Thr Tyr Ser Ser Pro Ile Glu Asn Asp Ser 275 g Arg Ty Gly Asp 11e Arg Arg Gli Phe Ala Asp Ala Asp Gly Asp 316 g Glu Val Leu Leu Asn Asp Tyr Leu Ala His Gln Arg Lys Leu Leu 330 a Ala Ala Clu Asp Asp 11e Gln Gly Gly Asp Gly Pro Val Asp 315 g Glu Val Leu Leu Asn Asp Tyr Leu Ala His Gln Arg Lys Leu 125 a Arg Gln Arg Glu Leu Glu Asn Arg Thr He Ala Clu 145 330 Thr Thr Tyr Ala 257				100					105					110		
130 135 140 r Asp Val Gln Gln Leu Arg Thr Thr Cys Val Tyr Asp Pro Gln Thr 160 n Thr Phe Asp Leu His Thr Pro His Glu Ala Ala Arg Lys Asp Tyr 175 e Gly Asn Ala Ala Arg Asp Gly Arg Met Ala Val Val Phe Ala Gln 180 u Val Thr Gly Gly Arg Arg His Gly Val His Ala Trp Leu Val Pro 210 e Arg Asp Glu His Gly Lys Pro Met Pro Gly Val Thr Ile Gly Asp 210 a Gly Pro Lys Ala Gly Leu Leu Gly Val Asp Asn Gly Arg Leu Ser 230 c Asp His Val Arg Val Pro Arg Glu Met Leu Leu Asp Gln Tyr Ala 250 n Val Ala Glu Asp Gly Thr Tyr Ser Ser Pro Ile Glu Ann Asp Ser 260 g Arg Phe Phe Thr Met Leu Gly Thr Leu Val Arg Gly Arg Val Ser 285 g Gly Qly Ala Ala Ser Ala Ala Thr Lys Ser Ala Leu Ala Ile Ala 300 130 Tyr Gly Asp Ile Arg Arg Gln Phe Ala Asp Ala Asp Gly Asp 310 130 Glu Val Leu Leu Asp Asp Tyr 1225 g Glu Val Leu Leu Asp Asp Tyr Leu Ala His Gln Arg Lys Leu Leu 220 g Glu Val Leu Leu Asp Asp Tyr Leu Ala His Gln Arg Lys Leu Leu 325 g Glu Val Leu Leu Asp Asp Tyr Ala 255 g Glu Val Leu Leu Asp Asp Tyr Asp 256 g Glu Val Leu Leu Asp Asp Tyr Ala 257 130 Tyr Ala 258 140 Tyr Ala 259 15 Tyr Gly Asp Tir Tyr Ala 256 160 Asp Tyr Gly Asp Tir Arg Arg Arg Gln Pro Arg Asp Ala Asp Ala Asp Gly Asp 300 <td>Ala</td> <td>Leu</td> <td>-</td> <td>Thr</td> <td>ГЛа</td> <td>Arg</td> <td>His</td> <td></td> <td>-</td> <td>Ala</td> <td>Tyr</td> <td>Leu</td> <td>-</td> <td>Asp</td> <td>Ile</td> <td>Val</td>	Ala	Leu	-	Thr	ГЛа	Arg	His		-	Ala	Tyr	Leu	-	Asp	Ile	Val
5 150 155 160 n Thr Phe Asp Leu His Thr Pro His Glu Ala Ala Arg Asp Tyr a Gly Asn Ala Gly Val Asp Ala Gly Ala Gly Ala Gly Ala Gly Ala Gly Ala Ala Sin Ala Ala Sin Ala	Ala	-	Arg	Leu	Phe	Gly	-		Ala	Met	Thr		Thr	Gly	His	Gly
165 170 175 e Gly Asn Ala Asp Gly Gly Asp Gly	Ser 145	-	Val	Gln	Gln		-	Thr	Thr	Суз		Tyr	Asp	Pro	Gln	
180 185 190 u Inr G1y G1y Arg	Gln	Thr	Phe	Asp		His	Thr	Pro	His		Ala	Ala	Arg	Гла		Tyr
195 200 205 e Arg Asp Glu His Gly Lys Pro Met Pro Gly Val Thr Ile Gly Asp 215 Pro Met Pro Gly Val Asp Asn Gly Arg Leu Ser 230 a Gly Pro Lys Ala Gly Leu Leu Gly Val Met Leu Leu Asp Gln Tyr Ala 235 Asp Glu Asp Gly Thr Tyr Ser Ser Pro Ile Glu Asp Asp 275 a Ma Glu Asp Gly Thr Met Leu Gly Thr Leu Val Arg Gly Arg Leu Ser 240 a Ma Glu Asp Gly Thr Tyr Ser Ser Pro Ile Glu Asp Asp Val Ser 270 g Arg Phe Phe Thr Met Leu Gly Thr Leu Val Arg Gly Arg Val Ser 270 g Arg Tyr Gly Asp Ile Arg Arg Gln Phe Ala Asp Ala Asp Gly Asp 310 g Glu Val Leu Leu Asn Asp Tyr Leu Ala His Gln Arg Lys Leu Leu 325 g Ala Leu Ala Thr Thr Tyr Ala 245 a Glu Val Leu Asp Asp Ile Gln Gly Gly Asp Gly Pro Val Asp 310 340 a Leu Ala Thr Thr Tyr Ala 245 b Ala Ser 340 a Ala Clu Asp Asp Tile Gln Gly Gly Asp Gly Pro Val Asp 310 a Glu Val Leu Leu Asp Asp Tile Gln Gly Gly Asp Gly Pro Val Asp 310 a Arg Gln Arg Glu Leu Glu Ser Arg Ala Ala Gly Leu Lys Ala Ala 310 a Arg Gln Arg Glu Leu Glu Asp Arg Glu Asp 330 a Thr Trp His Ala Thr Thr Thr Phe Glu Asp Arg Asp 310 a Ma Gly Tyr Leu Ser Glu Asp Arg Asp 310 a Asp 310 a Arg Glu Arg Gly Trp His Ala Thr Arg Thr Ile Gln Ile Cys Arg Glu Ala Cys 345 a Arg Glu Arg Gly Trp His Ala Thr Thr Phe Glu Gly Asp Asp Thr Val Leu Leu 430 <td>Ile</td> <td>Gly</td> <td>Asn</td> <td></td> <td>Ala</td> <td>Arg</td> <td>Asp</td> <td>Gly</td> <td></td> <td>Met</td> <td>Ala</td> <td>Val</td> <td>Val</td> <td></td> <td>Ala</td> <td>Gln</td>	Ile	Gly	Asn		Ala	Arg	Asp	Gly		Met	Ala	Val	Val		Ala	Gln
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5 230 235 240 e Asp His Val Arg Val Pro Arg Glu Met 250 Leu Leu Asp Gln Tyr Ala 255 Ana 255 Ana 255 n Val Ala Glu Asp Gly Thr Tyr Ser Ser Pro Ile Glu Asp Asp Ser 265 Pro Ile Glu Asp Gly Arg Val Ser 285 Ang Gly Arg Val Ser 285 Ang Gly Arg Val Ser 285 g Arg Phe Phe Thr Met Leu Gly Thr Leu Val Arg Gly Arg Val Ser 290 Ana Ala Ser Ala Ala Thr Lys Ser Ala Leu Ala Ile Ala 300 Ang Gly Asp Gly Asp 11e Arg Arg Gln Phe Ala Asp Ala Asp Gly Asp 310 f Arg Tyr Gly Asp Ile Arg Arg Gln Phe Ala His Gln Arg Lys Leu Leu 325 Ser 14 His Gln Arg Lys Leu Leu Asp 345 o Ala Leu Ala Thr Thr Tyr Ala Leu Thr Phe Ala Gln Ala Glu Leu 355 Glu Ala Ala Ser Arg Ala 265 Ang Gly Asp Gly Pro Val Asp Glu 265 s Arg Gln Arg Glu Leu Gly Ser Arg Ala Ala Gly Leu Lys Ala Ala 350 Glu Ala Gly Tyr Leu 366 Ang Gly Asp Glu Ala 265 a Arg Gln Arg Glu Leu 375 Ser Arg Ala Ala Gly Leu Pro Ser Leu Lys Ala 380 Ala 240 g Gly Ala Gly Tyr Leu Ser Glu Asn Arg Leu Pro Ser Leu Lys Ala 415 Ala 445 Ala 445 p Thr Asp Val Phe Thr Thr Phe Glu Asp Arg Ala Cyr 445 Ala Cyr 445 Ala Leu 440 a Gly Ala Gly Tyr Leu Ser Glu Asn Arg Leu Pro Ser Leu Lys Ala 415 Ala 445 p Thr Asp Val Phe Thr Thr Phe Glu Asp Arg Ala Cu Clu Hab 425 Ala 415 a Gly Ala Gly Tyr Gly Arg Ala Ser Phe Val Ala Glu Glu	Ile	-	Asp	Glu	His	Gly	-	Pro	Met	Pro	Gly		Thr	Ile	Gly	Asp
245 250 255 n Val Ala Glu Asp Gly Thr Tyr Ser Ser Pro Ile Glu Asn Asp Ser 265 g Arg Phe Phe Thr Met Leu Gly Thr Leu Val Arg Gly Arg Val Ser 285 1 Gly Gly Ala Ala Ser Ala Ala Thr Lys Ser Ala Leu Ala Ile Ala 290 290 11 Gly Gly Ala Ala Ser Ala Ala Thr Lys Ser Ala Leu Ala Ile Ala 300 1 Arg Tyr Gly Asp Ile Arg Arg Gln Phe Ala Asp Ala Asp Gly Asp 310 290 11 Leu Leu Asp Asp Tyr Leu Ala His Gln Arg Lys Leu Leu 330 310 11 Leu Asp Asp Tyr Leu Ala His Gln Arg Lys Leu Leu 330 311 11 Thr Tyr Ala Leu Ala Thr Tyr Ala 345 312 310 313 310 314 11 Leu Asp Asp 310 320 310 311 11 Leu Asp Asp 310 320 320 311 11 Leu Asp Asp 310 325 11 Leu Asp Asp 310 325 11 Leu Asp Asp 310 326 11 Leu Asp 320 327 11 Leu Asp 310	Ala 225	Gly	Pro	ГЛЗ	Ala	_		Leu	Gly	Val	-	Asn	Gly	Arg	Leu	
260 265 270 g Arg Phe Phe Phe Thr Met Leu Gly Thr Leu Val Arg Gly Arg Val Ser 275 Phe Phe Thr Met Leu Gly Thr Leu Val Arg Gly Arg Val Ser 1 Gly Gly Ala Ala Ala Ser Ala Ala Thr Lys Ser Ala Leu Ala Ile Ala Ang Gly Arg Gly Asp Gly Arg S Ala Asp Gly Asp 300 1 Arg Tyr Gly Ala Ala Ser Ala Arg Arg Gln Phe Ala Asp Asp 310 Arg Gln Phe Ala Asp Asp Ala Asp Gly Asp 320 Glu Val Leu Leu Asp Asp 11e Arg Arg Gln Phe Ala Asp Asp Ala Asp Gly Asp 320 g Glu Val Leu Lau Asp Asp Tyr Leu Ala His Gln Arg Lys Leu Leu 330 Can Ala Chr Thr Thr Tyr Ala Leu Thr Phe Ala Gln Ala Glu Leu 335 Ang Glu Asp Glu Asp 360 Ang Gly Asp 61u 355 a Arg Gln Arg Glu Leu Asp Asp Ile Gln Gly Gly Asp 365 Gly Fer Val Asp Glu 360 Ang Asp 61u 360 Ang Asp 61u 360 a Arg Gln Arg Glu Leu 375 Glu Ser Arg Ala Ala Gly Leu Lys Ala Ala 380 Ang Glu Ala Cys 390 Ang Glu Ang 375 Ang Glu Ala 280 a Thr Trp His Ala Thr Arg Thr Arg Arg Arg Leu Pro Ser Leu Lys Ala 410 Ang 445 Ang 445 Ang 445 Ang 445 a Thr Asp Val Phe Thr Thr Thr Phe Glu Glu Asp Arg Leu Pro Ser Leu Lys Ala 445 Ang 445 Ang 445 Ang 445 Ang 445 a Leu Val Ala Lys Gly Leu Leu Thr Gly Tyr Arg Asp Glu Phe Gly 445 Ang 445 Ang 445 Ang 445 Ang 445 a Met Val Leu Glu Arg 445 A	Phe	Asp	His	Val	-	Val	Pro	Arg	Glu		Leu	Leu	Asp	Gln	-	Ala
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5 310 315 320 g Glu Val Leu Leu 325 Asn Asp Tyr Leu Ala 330 His Gln Arg Lys Leu Leu 335 Leu 335 o Ala Leu Ala Thr Thr Tyr Ala Leu Thr Phe Ala Gln Ala Glu Leu 345 Glu Sas Glu Sas Glu Sas Glu Sas Glu 355 Glu Asp Glu Sas Glu Sas Glu Sas Glu 365 Asp Glu 365 1 Ala Ala Leu Asp Asp IIe Glu Glu Ser Arg Ala Ala Glu Leu 360 Glu Sas Glu Arg Glu Leu Glu Ser Arg Ala Ala Glu Sas Arg Glu Ala Cys 385 Ala Gly Tyr Leu Ser Glu Asp Asp Glu Sas Arg 410 Glu Pro Ser Leu Lys Ala 415 n Thr Trp His Ala Thr Arg Thr Ihe Glu Glu Asp Asp Glu Ala Cys 400 Glu Ala Cyr 405 Ser Glu Asp Arg 612 Asp Ala 240 g Gly Ala Gly Tyr Leu Ser Glu Asp Arg Leu Pro Ser Leu Lys Ala 415 Ala 425 Asp Asp Asp 140 Asp Asp 415 n Leu Val Ala Lys Gly Leu Leu Thr Gly Tyr Arg Asp Glu Phe Gly 435 Asp 440 Asp 445 Ala Arg 445 n Leu Val Ala Leu Glu Arg Thr Ala Ala Arg Ala Arg Ala Clu Glu Glu Ala Clu Glu Arg 445 Ala Arg 445 Arg 445 n Leu Val Ala Lys Gly Leu Leu Thr Gly Tyr Arg Asp Glu Glu Ala Clu Arg 445 Arg 445 Arg 445 n Leu Val Ala Lys Gly Trp Gly Arg Ala Ser Phe Val Ala Glu Glu Glu Arg 445 Arg 445 Arg 445 n Met Val Leu Glu Arg Thr Arg Asp Asp Glu Val Ala Clu Thr Asp Arg 445 Arg 445 Arg 445 n Met Val Leu Glu Arg 485 Arg Asp Asp Glu Val	Val	-	-	Ala	Ala	Ser		Ala	Thr	Lys	Ser		Leu	Ala	Ile	Ala
325330335o Ala Leu Ala Thr Thr Tyr Ala Leu Thr Phe Ala Gln Ala Glu Leu 340Thr Thr Tyr Ala Leu Thr Phe Ala Gln Ala Glu Leu 345Glu Asp Glu 365Glu Leu 3501 Ala Ala Leu Asp Asp Asp IIe Gln Gly Gly Asp Gly Pro Val Asp Glu 355Arg Gln Arg Glu Leu Glu Ser Arg Ala Ala Gly Leu Lys Ala Ala 370Gln Arg Glu Leu Glu Ser Arg Ala Ala Gly Leu Lys Ala Ala 	Val 305	Arg	Tyr	Gly	Asp		Arg	Arg	Gln	Phe		Asp	Ala	Asp	Gly	
340 345 350 1 Ala Ala Ala Asp Asp Ile Gln Gly Gly Asp Gly Pro 365 Val Asp Glu s Arg Gln Arg Glu Leu Glu Ser Arg Ala Ala Gly Leu Leu Ala Ala Ser Arg Ala Gly Leu Leu Ala Ala Ser Arg Ala Gly Leu Leu Ala Ala Ser Arg Ala Gly Leu Leu Ala Ala s Tr Tr His Ala Thr Arg Gln Arg Ala Ala Ser Arg Ala Gly Arg Ala Crs Arg Ala	Arg	Glu	Val	Leu		Asn	Asp	Tyr	Leu		His	Gln	Arg	Lys		Leu
355360365s ArgGlnArgGluLeuGluSerArgAlaAlaGlyLeuLysAlaAlanThrTrpHisAlaThrArgThrIleGlnIleCysArgGluAlaCysyGlyAlaGlyTyrLeuSerGluAsnArgLeuProSerLeuLysAlaCysyGlyAlaGlyTyrLeuSerGluAsnArgLeuProSerLeuLysAlaAlaCysyGlyAlaGlyTyrLeuSerGluAsnArgAsnThrValLeuLeuLeuLeuAnnArgAdoAsnArgAdoAsnArgAdoAsnAnnAsn <td< td=""><td>Pro</td><td>Ala</td><td>Leu</td><td></td><td>Thr</td><td>Thr</td><td>Tyr</td><td>Ala</td><td></td><td>Thr</td><td>Phe</td><td>Ala</td><td>Gln</td><td></td><td>Glu</td><td>Leu</td></td<>	Pro	Ala	Leu		Thr	Thr	Tyr	Ala		Thr	Phe	Ala	Gln		Glu	Leu
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5 390 395 400 y Gly Ala Gly Tyr Leu Ser Glu Asn Arg 410 Leu Pro Ser Leu Lys Ala 415 p Thr Asp Val Phe Thr Thr Phe Glu Gly Asp Asn Thr Val Leu Leu 425 Asn Thr Val Leu Leu 430 n Leu Val Ala Lys Gly Leu Leu Thr Gly Tyr Arg Asp Glu Phe Gly 435 Asp Gly Trp Gly Arg Ala Ser Phe Val Ala Glu Gln Val Arg 445 u Met Val Leu Glu Arg Thr Ala Ala Arg Ala Che Val Leu Clu 430 Arg 450 1 Ser Ala Val Pro Gly Arg Asp Asp Asp Glu Val Ala Val Thr Asp Arg 490 y Trp Gln Leu Lys Leu Phe Glu Asp Arg Glu Glu His Leu Leu Asp	His	-	Gln	Arg	Glu	Leu		Ser	Arg	Ala	Ala	-	Leu	Гла	Ala	Ala
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420 425 430 n Leu Val Ala Lys Oly Clu	Gly	Gly	Ala	Gly	-	Leu	Ser	Glu	Asn	-	Leu	Pro	Ser	Leu	-	Ala
435 440 445 r Leu Asp Gly Trp Gly Arg Ala Ser Phe Val Ala Glu Gln Val Arg 460 Ala Glu Gln Val Arg 460 u Met Val Leu Glu Arg Thr Ala Ala Arg Arg 475 Leu Ala Arg Leu 480 5 Ala Val Pro Gly Arg Asp Asp Asp Glu Val Ala Val Thr Asp Arg 490 9 Trp Gln Leu Lys Leu Phe Glu Asp Arg Glu Asp Arg Glu Glu His Leu Leu Asp	Aap	Thr	Asp		Phe	Thr	Thr	Phe		Gly	Asp	Asn	Thr		Leu	Leu
450 455 460 u Met Val Leu Glu Arg Thr Ala Ala Arg Ala Leu Ile Ala Arg Leu 5 Ser Ala Val Pro Gly Arg Asp Asp Glu Val Ala Val Thr Asp Arg 485 485 490 490 490 490 490 490 490 490 490 490	Gln	Leu		Ala	ГЛа	Gly	Leu		Thr	Gly	Tyr	Arg		Glu	Phe	Gly
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485 490 495 y Trp Gln Leu Lys Leu Phe Glu Asp Arg Glu Glu His Leu Leu Asp	Glu 465	Met	Val	Leu	Glu			Ala	Ala	Arg		Leu	Ile	Ala	Arg	
	Val	Ser	Ala	Val		Gly	Arg	Asp	Asp		Val	Ala	Val	Thr		Arg
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cagetg	gtgg 1	tcacę	geee	ct g	tacco	gcga	g gcé	gatgo	cccc	tgct	cgcgo	cta 🤇	caaco	ctcgag	900	
qacaac	gtgt (cggt	egeet	ta co	gacga	actgo	c gcé	gtgc	ggct	ggaa	agcto	gee (cacgo	gtccag	960	
5																

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gtgctcggca gggccgcgtt cggccatcgg gtcggcgcca cgaccgtcac ccagcaccgg ctqqaqqaac tcqtcttctc qctcccqqac qcctaccaqq tqqtqttctq qcqqqqcqcqq geggageegg eegeetgeg categagate gaggtgeeeg aggageaeeg ggeggeegee gaggeggaac tggtgeacte ggtgeggaee gegtteggtg tggaeageae ggteaeegge ctccctccgg gcaccctgat cccccacggc gcgctgaccg ccatgcccga cgtggtcaag ccgcgcagcc tcttcgggcc cgacgaggac tggggcaaag cgctcctcta ctactga <210> SEQ ID NO 98 <211> LENGTH: 290 <212> TYPE: PRT <213> ORGANISM: Artificial sequence <220> FEATURE: <221> NAME/KEY: Miscfeature <222> LOCATION: (1)..(290) <223> OTHER INFORMATION: HMM consensus seq based on alignment of Fig 10 IPTN <400> SEQUENCE: 98 Ala Ala Glu Leu Tyr Ser Val Ile Glu Glu Ser Ala Arg Leu Leu Asp Val Ala Cys Ser Arg Asp Arg Val Trp Pro Ile Leu Ser Ala Tyr Gly Asp Ala Phe Ala His Pro Ala Ala Val Val Ala Phe Arg Val Ala Thr Ala Leu Arg His Val Gly Glu Leu Asp Cys Arg Phe Thr Thr His Pro Asp Asp Arg Asp Pro Tyr Ala Leu Ala Leu Ser Arg Gly Leu Thr Pro 65 70 75 80 Lys Thr Asp His Pro Val Gly Ser Leu Leu Ser Glu Val Gln Glu Arg Leu Pro Val Glu Ser Tyr Gly Ile Asp Phe Gly Val Val Gly Gly Phe Lys Lys Ile Tyr Ala Phe Phe Thr Pro Asp Glu Leu Gl
n Glu Val Ala Ala Leu Ala Gly Ile Pro Ala Met Pro Arg Ser Leu Ala Gly Asn Ala Asp Phe Phe Glu Arg Tyr Gly Leu Asp Asp Arg Val Gly Val Leu Gly Ile Asp Tyr Pro Ala Arg Thr Val Asn Val Tyr Phe Asn Asp Val Pro Ala Glu Ser Phe Glu Ser Glu Thr Ile Arg Ser Thr Leu Arg Glu Ile Gly Met Ala Glu Pro Ser Glu Arg Met Leu Lys Leu Gly Glu Lys Ala Phe Gly Leu Tyr Val Thr Leu Gly Trp Asp Ser Ser Glu Ile Glu Arg Ile Cys Tyr Ala Ala Ala Thr Thr Asp Leu Thr Thr Leu Pro Val Pro Val Glu Pro Glu Ile Glu Lys Phe Val Lys Ser Val Pro Tyr Gly Gly Glu Asp Arg Lys Phe Val Tyr Gly Val Ala Leu Thr Pro Lys Gly Glu

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Tyr Tyr Lys Leu Glu Ser His Tyr Lys Trp Lys Pro Gly Ala Val Asp Phe Ile <210> SEQ ID NO 99 <211> LENGTH: 438 <212> TYPE: PRT <213> ORGANISM: Artificial sequence <220> FEATURE: <221> NAME/KEY: Miscfeature <222> LOCATION: (1)..(438) <223> OTHER INFORMATION: HMM consensus seq based on alignment of Fig 11 ADSA <400> SEQUENCE: 99 Val Asn Glu Pro Arg Ser Ser Leu Pro Arg Leu Gly Gln Trp His Gly Pro Glu Asp Leu Arg Arg Leu Gln Glu Lys Gln Leu Ala Gln Thr Val Thr Trp Ala Ala Arg Ser Pro Phe Tyr Arg Asp Arg Leu Asp Ser Gly Ala Leu Pro Val Thr Ala Ala Asp Leu Ala Asp Leu Pro Leu Thr Thr Lys Gln Asp Leu Arg Asp Asn Tyr Pro Phe Gly Met Leu Ala Val Pro Lys Glu Arg Leu Ala Thr Tyr His Glu Ser Ser Gly Thr Ala Gly Arg Pro Thr Pro Ser Tyr Tyr Thr Ala Glu Asp Trp Thr Asp Leu Ala Glu Arg Phe Ala Arg Lys Trp Ile Gly Met Ser Ala Glu Asp Val Phe Leu Val Arg Thr Pro Tyr Ala Leu Leu Leu Thr Gly His Leu Ala His Ala Ala Gly Arg Leu Arg Gly Ala Thr Val Val Pro Gly Asp Asn Arg Ser Leu Ala Met Pro Tyr Ala Arg Val Val Arg Val Met His Asp Leu Gly Val Thr Leu Thr Trp Ser Val Pro Thr Glu Cys Leu Ile Trp Ala Ala Ala Ala Thr Ala Ala Gly His Arg Pro Asp Val Asp Phe Pro Ala Leu Arg Ala Leu Phe Val Gly Gly Glu Pro Leu Thr Asp Ala Arg Arg Arg Arg Ile Ser Arg Leu Trp Gly Val Pro Val Ile Glu Glu Tyr Gly Ser Thr Glu Thr Gly Ser Leu Ala Gly Glu Cys Pro Glu Gly Arg Leu His Leu Trp Ala Asp Arg Ala Leu Phe Glu Val Tyr Asp Pro Asp Thr Gly Thr Val Arg Ala Asp Gly Asp Gly Gln Leu Val Val Thr Pro Leu Phe Arg Glu Ala Met Pro Leu Leu Arg Tyr Asn Leu Glu Asp Asn Val Ser Val Ser Tyr Asp Asp Cys Ala Cys Gly Trp Lys Leu Pro Thr Val Arg

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305				310					315					320
Val Leu	Gly	Arg	Ala 325	Ala	Phe	Gly	Tyr	Arg 330	Val	Gly	Ala	Thr	Thr 335	Ile
Thr Gln	His	Arg 340	Leu	Glu	Glu	Leu	Val 345	Phe	Ser	Leu	Pro	Glu 350	Ala	His
Arg Val	Val 355	Phe	Trp	Arg	Ala	Lys 360	Ala	Glu	Pro	Ala	Val 365	Leu	Arg	Ile
Glu Ile 370	Glu	Val	Ala	Glu	Glu 375	His	Arg	Val	Ala	Ala 380	Glu	Ala	Glu	Leu
Thr Ala 385	Ser	Val	Arg	Ala 390	Ala	Phe	Gly	Val	Aap 395	Ser	Glu	Val	Thr	Gly 400
Leu Ala	Pro	Gly	Thr 405	Leu	Ile	Pro	Arg	Glu 410	Ala	Leu	Thr	Ser	Met 415	Pro
Asp Val	Val	Lys 420	Pro	Arg	Ser	Leu	Phe 425	Gly	Pro	Asp	Glu	Asp 430	Trp	Gly
Lys Ala	Leu 435	Leu	Tyr	Tyr										

What is claimed is:

1. An isolated polynucleotide comprising a polynucleotide sequence, or a polynucleotide sequence complementary thereto, selected from the group consisting of:

- a) a polynucleotide encoding a polypeptide having at least 90% sequence identity to a polypeptide consisting of amino acids 1-438 of SEQ ID NO: 48 and having adenylating amide synthetase activity;
- b) a polynucleotide encoding a polypeptide having at least 90% sequence identity to a polypeptide consisting of amino acids 1-290 of SEQ ID NO: 22 and having isoprenyl transferase activity;
- c) a polynucleotide comprising the nucleic acid sequence of SEQ ID NO:47; and
- d) a polynucleotide comprising the nucleic acid sequence of SEQ ID NO:23.

2. An isolated polynucleotide comprising the nucleic acid sequence of SEQ ID NO:47.

3. An isolated polynucleotide comprising the nucleic acid sequence of SEQ ID NO:23.

4. The isolated polynucleotide of claim **1**, wherein said polypeptide of a) has at least 95% sequence identity to a polypeptide consisting of amino acids 1-438 of SEQ ID NO: 48.

5. The isolated polynucleotide of claim **1**, wherein said polypeptide of a) has at least 99% sequence identity to a polypeptide consisting of amino acids 1-438 of SEQ ID NO: 48.

6. The isolated polynucleotide of claim **1**, wherein said polypeptide of b) has at least 95% sequence identity to a polypeptide consisting of amino acids 1-290 of SEQ ID NO: 22.

7. The isolated polynucleotide of claim 1, wherein said polypeptide of b) has at least 99% sequence identity to a polypeptide consisting of amino acids 1-290 of SEQ ID NO: 22.

8. A purified polypeptide selected from the group consisting of:

- a) a polypeptide comprising amino acids 1-290 of SEQ ID NO: 22; and
- b) a polypeptide having at least 90% sequence identity to a polypeptide comprising amino acids 1-290 of SEQ ID NO: 22 and having an isoprenyl transferase activity; and
- c) a polypeptide encoded by a polynucleotide, the complement of which hybridizes under stringent conditions to a polynucleotide encoding a polypeptide comprising amino acids 1-290 of SEQ ID NO: 22, and having an isoprenyl transferase activity.

9. A purified polypeptide comprising amino acids 1-290 of SEQ ID NO: 22.

10. The purified polypeptide of claim **8**, wherein said polypeptide of b) has at least 95% identity to a polypeptide comprising amino acids 1-290 of SEQ ID NO: 22.

11. An expression vector comprising a polynucleotide of claim **1**.

12. The expression vector of claim **11**, wherein said polynucleotide encodes a polypeptide having at least 90% sequence identity to a polypeptide comprising amino acids 1-438 of SEQ ID NO: 48 and having adenylating amide synthetase activity.

13. The expression vector of claim **11**, wherein said polynucleotide encodes a polypeptide having at least 90% sequence identity to a polypeptide comprising amino acids 1-290 of SEQ ID NO: 22, and having isoprenyl transferase activity.

14. An isolated host cell transformed with an expression vector of claim 11.

15. The isolated host cell of claim **14**, wherein said host cell is a bacterial host cell.

16. A method for producing a farnesyl dibenzodiazepinone compound, comprising:

- a) providing a prokaryote transformed with an expression vector of claim 11; and
- b) culturing the prokaryote under conditions such that (i) an adenylating amide synthetase or an isoprenyl transferase is expressed, and (ii) a farnesyl dibenzodiazepinone compound is synthesized.

17. The method of claim **16**, wherein said prokaryote is *E. coli*.

18. The method of claim **16**, wherein said prokaryote is an actinomycete.

- **19**. An isolated polynucleotide encoding:
- a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88; or
- b) a polypeptide having at least 85% sequence identity to a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOS: 2, 4,

6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 41, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 65, 67, 69, 70, 71, 74, 76, 78, 80, 82, 84, 86 and 88, and having the same biological function as the corresponding polypeptide.

20. A cosmid selected from the group consisting of cosmid 046KM deposited under IDAC accession no. 250203-06 and cosmid 046KQ deposited under IDAC accession no. 250203-07.

21. The cosmid of claim **20**, wherein said cosmid is inserted into a prokaryotic host for expressing a product.

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