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## ABSTRACT

This invention provides genes and their encoded proteins, involved in the biosynthesis of farnesyl dibenzodiazepinones, including ECO-04601. The invention relates to expression vectors comprising the genes and to host cells transformed with these vectors. The invention further relates to methods of producing farnesyl dibenzodiazepinone compounds using the genes and proteins of the invention, for example, involving expression of biosynthetic pathway genes in transformed host cells.

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\%ccs


会 $867 \%$


Figure 1


Figure 2


Figure 3


ECO-04601 ( $20 \mathrm{mg} / \mathrm{kg}$ )
Figure 4

Figure 5

Figure 6

Figure 7

Figure 8


## Figure 9

| Locus A | ------VAELYSTIEESARQLDVPCSRDRVWPILSAYGDAFAHPEAVVAFRVATALRHAG |
| :---: | :---: |
| Locus B | MPGTSEAVELCSTIEESARLLNVACSRDRVWSLLSAYGDAFAHPGAVVAFRVATAMRHVG |
| Locus C | MFATAGAAELHAVVEDSARLLGVTCSPDTVAPILSTYGDTFEHDATVVAFRVATGKRHIG |
|  | * : : ***** *.*.** * * : **:***:* * ${ }^{* * * * * * * * * . ~ * * ~}$ |

Locus A ELDCRFRTHPDDRDPYASALARGLTPRTDHPVGALLSEVHRRCPVESHGIDFGVVGGFKK Locus B ELDCRFTTHPDDRDPYARALSRGLTPETDHPVGTLLSEVQGRCPVESHGIDFGVVGGFKK Locus C ELDCRFTTHPTHRDPYALALSNGLTPKTGHPVGSLLSALQERLPIDSYGIDFGVVGGFKK


| Locus A | IYAAFAPDELQVATSLAGIPAMPRSLAANADFFTRHGLDDRVGVLGFDYPARTVNVYFND |
| :---: | :---: |
| Locus B | IYAFFTPDDLQETSKLAEIPAMPRSLAGNVEFFARHGLDDRVGVFGIDYPSRTVNVYFND |
| Locus C | IYSFFTPDALQEVAALAGIPSMPRSLAG-RDFFERYGCTTGR-VIGIDYPH- |
|  |  |


| Locus A | VPRECFEPETIRSTLRRTGMAEPSEQMLRLGTGAFGLYVTLGWDSPEIERICYAAATTDL |
| :---: | :---: |
| Locus B | VPAESFHSETIRSTLREIGMAEPSERMLKLGEKAFGLYVTLGWDSSRIERICYAAATTDL |
| Locus C |  |
|  | ***.*..********. *******:**:**************..************ |


| Locus A | TTLPVPVEPEIEKFVKSVPYGGGDRKFVYGVALTPKGEYYKLESHYKWKPGAVNFI |
| :---: | :---: |
| Locus B | TTLPVPVEPEIEKFVRSVPYGGEDRKFVYGVALTPHGEYYKLESHYRWKPGAMDFI |
| Locus C |  |
|  | ********: *****************: ************* |

## Figure 10

Locus A VSEPSSSLPRLGQWHGLEDLRRLQEKQLAETFTWAARSPFYRARLASGAPPVTPADLADL Locus B VNDPRPSLPQLGQWHGPEDLQRLQEKQLSQTVTWATRSPFYRDRLDPGALPATAADLADL Locus C VNPTRSSLPRLGQWNGPEDLRLLQEKQLQQTVGWASRSPFYRGRLDTAALPTTIDDLASL *. . . ***:****:* ***: ****** : *. **: ****** ** ..**.* ***.*

Locus A PLTTKQDLRDNYPFGMLAVPRERLATYHESSGTAGKPTPSYYTAEDWTDLAERFARKWIG Locus B PLTTKQDLRDNYPFGMLAVPKERLATYHESSGTAGRPTPSYYTAEDWTDLAERFARKWIG Locus C PLTTKQDLRDNYPFGMLAVPKERLATYHESSGTAGRPTPSYYTADDWIDLAERFARKWIG


Locus A MSADDVFLVRTPYALLLTGHLAHAAARLRGATVVPGDNRSLAMPYARVVRVMHDLDVTLT Locus B MSAEDVFLVRTPYALLLTGHLAHAAGRLRGATVVPGDNRSLAMPYARVVRVMHDLGVTLT Locus C ITAFDVFIVRTPYALTLTGHTAHAAGRLHGATVVPGDNRSLAMPYARVVRVMHDLGVTLT $:: *: * * * * * * * * * * * * * * * * * * * * * 。 * *: * * * * * * * * * * * * * * * * * * * * * * * * * * 。 * * * *$

Locus A WSVPTECLIWAAAAIAAGHRPDIDFPALRALFVGGEPMTDARRRRISRLNGVPVIEEYGS Locus B WSVPTECLIWAAAATAAGHRPDVDFPALRALFVGGEPLTDARRRRISRLWGVPVIEEYGS Locus C WSVPTECLIWAAAATAAGHRPSEDFPALRALFVGGEPLTTARRDRISRLWGVPVIEEYGS


Locus A
TETGSLAGECPEGRLHLWADRALFEVYDPDTGAVRADGDGQLVVTPLFREAMPLLRYNLE
Locus B TETGSLAGECPNGRMHLWADRALFEVYDPRTGTVSADGDGQLVVTPLFREAMPLLRYNLE Locus C TETGSLAGECPHGRMHLWADRALFEVYDPQTGTVRAEGEGQLVVTPLYREAMPLLRYNLE


Locus A
DNVSVSYDDCGCGWKLPTVRVLGRSAFGYRVGGTTITQHQLEELVFSLPEAHRVMFWRAK
Locus B DDVTVSYDDCACGWNLPTVRVLGRAAFGYRVGAATITQHRLEEVVFSLPESHGVVFWRAK Locus C DNVSVAYDDCACGWKLPTVQVLGRAAFGHRVGATTVTQHRLEELVFSLPDAYQVVFWRAR *:*:*:****.***:****:****:***:***.:*:***:***:*****: : *:****:

| Locus A | AEPALLRVEIEVAAAHRVAAEAELTAAIRAAFGVDSEVTGLAPGTLIPLDALTSMPDVVK |
| :--- | :--- |
| Locus B | AEPTVLRIEIEVAEEHRTAAQAELTASVRATFGIDSEVTGLTPGTLVPREALTSMPDVVK |
| Locus C | AEPAALRIEIEVPEEHRAAAEAELVHSVRTAFGVDSTVTGLPPGTLIPHGALTAMPDVVK |
|  |  |
|  |  |
|  |  |

Locus A PRSLFGPDEDWSKALLYY
Locus B PRSLFGPDEDWGKALLYY
Locus C PRSLFGPDEDWGKALLYY

## POLYNUCLEOTIDES FOR PRODUCTION OF FARNESYL DIBENZODLAZEPINONES

## RELATED APPLICATIONS

[0001] This application is a divisional of U.S. patent application Ser. No. 11/330,123, filed Jan. 12, 2006, which is a continuation-in-part of U.S. patent application Ser. No. 10/762,107, filed Jan. 21, 2004, now issued as U.S. Pat. No. $7,101,872$, which claims priority to each of U.S. Provisional Application No. 60/441,126, filed Jan. 21, 2003, U.S. Provisional Application No. 60/492,997, filed Aug. 7, 2003, and U.S. Provisional Application No. 60/518,286, filed Nov. 10, 2003. The entire disclosures of each of these applications are herein incorporated by reference.

## SEQUENCE LISTING ON COMPACT DISK

[0002] The content of the following submissions on compact discs are incorporated herein by reference in its entirety: A compact disc copy of the Sequence Listing (COPY 1) (file name: 3005-5US-50US.ST25.txt, date recorded Jan. 10, 2006, size: 298 KB ) and a duplicate compact disc copy of the Sequence Listing (COPY 2) (file name: 3005-5US-50US. ST25.txt, date recorded Jan. 10, 2006, size: 298 KB).

## FIELD OF THE INVENTION

[0003] The invention relates to novel polynucleotide sequences and their encoded proteins, which are involved in the biosynthesis of a farnesyl dibenzodiazepinone compound and analogs. The invention relates to the use of such polynucleotides and proteins to produce farnesyl dibenzodiazepinone compounds and analogs. One method of obtaining the compound is by cultivation of a novel modified strain of Micromonospora sp., i.e., 046-ECO11 or [S01]046; another method involves expression of biosynthetic pathway genes in transformed host cells. The present invention further relates to cosmids 046 KM and 046 KQ and their methods of use.

## BACKGROUND OF THE INVENTION

[0004] The euactinomycetes are a subset of a large and complex group of Gram-positive bacteria known as actinomycetes. Over the past few decades these organisms, which are abundant in soil, have generated significant commercial and scientific interest as a result of the large number of therapeutically useful compounds produced as secondary metabolites. The intensive search for strains able to produce new secondary metabolites having potential therapeutic applications has led to the identification of hundreds of new species. Many of the euactinomycetes, particularly Streptomyces and the closely related Saccharopolyspora genera, have been extensively studied. Both of these genera produce a notable diversity of biologically active metabolites. Because of the commercial significance of these compounds, much is known about the genetics and physiology of these organisms.
[0005] Microbial genomic information is unique in that, unlike the organization of genomic information in higher life forms, microbial secondary metabolic biosynthetic genes are known to cluster together within the genome. This information allows identification of the gene locus encoding the enzymes responsible for the biosynthesis of a specific molecule. Equally, the identification of the genes present within a cluster allows prediction of the structure of the secondary metabolite. The identification of the genes and proteins
responsible for the production of active molecules allows for example, generation of structural analogs or improvement of the production process.
[0006] U.S. patent application Ser. No. 10/762,107 describes a dibenzodiazepinone secondary metabolite, specifically 10 -farnesyl-4,6,8-trihydroxy-dibenzodiazepin-11one (named ECO-04601) produced by a known euactinomycetes strain, Micromonospora sp. (IDAC 231203-01). Likewise, U.S. Pat. No. 5,541,181 (Ohkuma et al.) also discloses a dibenzodiazepinone secondary metabolite, specifically 5-farnesyl-4,7,9-trihydroxy-dibenzodiazepin-11-one (named "BU-4664L"), produced by a known euactinomycetes strain, Micromonospora sp. M990-6 (ATCC 55378). Both these dibenzodiazepinones have been reported to have anti-tumor activity.
[0007] Although many biologically active compounds have been identified from bacteria, there remains the need to obtain novel naturally occurring compounds with enhanced properties. Current methods of obtaining such compounds include screening of natural isolates and chemical modification of existing compounds, both of which are costly and time consuming. Current screening methods are based on general biological properties of the compound, which require prior knowledge of the structure of the molecules. Methods for chemically modifying known active compounds exist, but still suffer from practical limitations as to the type of compounds obtainable.
[0008] Thus, there exists a considerable need to obtain pharmaceutically active compounds in a cost-effective manner and with high yield. The present invention solves these problems by providing polynucleotides, polypeptides, vectors comprising the polynucleotides and host cells comprising the vectors for production of dibenzodiazepinones, as well as methods to generate farnesyl dibenzodiazepinones by de novo biosynthesis (heterologous or homologous expression of biosynthetic genes) or semi-synthesis rather than by chemical synthesis.

## SUMMARY OF THE INVENTION

[0009] The invention further encompasses an isolated polynucleotide comprising one or more of SEQ ID NOs. 1, 64 and 73, wherein the polynucleotide encodes a polypeptide that participates in a biosynthetic pathway for a farnesyl dibenzodiazepinone.
[0010] The invention further encompasses an isolated polynucleotide comprising SEQ ID NOs. 1, 64 and 73, wherein the polynucleotide encodes a polypeptide that participates in a biosynthetic pathway for a farnesyl dibenzodiazepinone.
[0011] The invention further encompasses an isolated polynucleotide that encodes a polypeptide selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, $24,26,28,30,32,34,36,38,40,42,44,46,48,50,52,54,56$, $58,60,62,65,67,69,71,74,76,78,80,82,84,86,88,90,92$, 94 and 96.
[0012] The invention further provides an isolated nucleic acid comprising a nucleotide sequence identical or complementary to a polynucleotide encoding a polypeptide having at least $70 \%$, at least $75 \%$, at least $80 \%$, at least $85 \%$, at least $90 \%$, at least $95 \%$ or at least $99 \%$ identity to a sequence selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, $10,12,14,16,18,20,22,24,26,28,30,32,34,36,38,40,42$, $44,46,48,50,52,54,56,58,60,62,65,67,69,71,74,76,78$, $80,82,84,86,88,90,92,94$ and 96 said polypeptide having the same biological function as its corresponding protein.
[0013] The invention further provides an isolated nucleic acid comprising a nucleotide sequence hybridizing under low, moderate, high or very high stringency conditions to the complement of a polynucleotide encoding a sequence selected from the group consisting of SEQ ID NOs. 2, 4, 6, 8, $10,12,14,16,18,20,22,24,26,28,30,32,34,36,38,40,42$, $44,46,48,50,52,54,56,58,60,62,65,67,69,71,74,76,78$, $80,82,84,86,88,90,92,94$ and 96 , said polypeptide having the same biological function as its corresponding protein.
[0014] The invention provides an isolated, purified or enriched nucleic acid comprising a polynucleotide, or a nucleotide sequence complementary thereto, said polynucleotide encoding a polypeptide selected from an adenylating amide synthetase (ADSA) having at least $80 \%$, at least $90 \%$, or at least $95 \%$ identity to the adenylating amide synthetase of SEQ ID NO: 48; and an isoprenyl transferase (IPTN) having at least $80 \%$, at least $90 \%$, or at least $95 \%$ identity to the isoprenyl transferase of SEQ ID NO: 22. In one embodiment, the invention provides an expression vector comprising said ADSA or IPTN-encoding nucleic acid. In another embodiment, the invention provides host cells transformed which such vector.
[0015] The invention further provides a polypeptide selected from an adenylating amide synthetase (ADSA) having at least $80 \%$, at least $90 \%$, or at least $95 \%$ identity to the adenylating amide synthetase of SEQ ID NO: 48; and an isoprenyl transferase (IPTN) having at least $80 \%$, at least $90 \%$, or at least $95 \%$ identity to the isoprenyl transferase of SEQ ID NO: 22.
[0016] In one embodiment, the isolated polynucleotide comprising SEQ ID No. 1 encodes a polypeptide selected from the group consisting of SEQ ID Nos. 2, 4, 6, 8, 10, 12, $14,16,18,20,22,24,26,28,30,32,34,36,38,40,42,44,46$, $48,50,52,54,56,58,60$ and 62.
[0017] In another embodiment, the isolated polynucleotide comprising SEQ ID No. 64 encodes a polypeptide selected from the group consisting of SEQ ID NOS: 65, 67, 69 and 71.
[0018] In another embodiment, the isolated polynucleotide comprising SEQ ID No. 73, encodes a polypeptide selected from the group consisting of SEQ ID NOS: 74, 76, 78, 80, 82, 84, 86 and 88.
[0019] The invention further encompasses an isolated polypeptide of SEQ ID NO. 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, $22,24,26,28,30,32,34,36,38,40,42,44,46,48,50,52,54$, $56,58,60,62,65,67,69,71,74,76,78,80,82,84,86,88,90$, 92, 94 and 96.
[0020] The invention further provides an isolated polypeptide having at least $70 \%$, at least $75 \%$, at least $80 \%$, at least $85 \%$, at least $90 \%$, at least $95 \%$ or at least $99 \%$ identity to a sequence selected from the group consisting of SEQ ID NOs. $2,4,6,8,10,12,14,16,18,20,22,24,26,28,30,32,34,36$, $38,40,42,44,46,48,50,52,54,56,58,60,62,65,67,69,71$, $74,76,78,80,82,84,86,88,90,92,94$ and 96 , said polypeptide having the same biological function as its corresponding protein.
[0021] In one embodiment, the polypeptide participates in a biosynthetic pathway for a farnesyl dibenzodiazepinone.
[0022] The invention further encompasses an expression vector comprising one or more of the polynucleotides described herein.
[0023] The invention further encompasses a recombinant prokaryotic organism comprising one or more such expression vectors.
[0024] In one embodiment, the organism is an actinomycete.
[0025] In another embodiment, the organism requires the expression vector to synthesize a farnesyl dibenzodiazepinone. That is, the organism is deficient in the ability to synthesize a farnesyl dibenzodiazepinone before transformation with a polynucleotide as described herein.
[0026] The invention further encompasses a method of making a farnesyl dibenzodiazepinone de novo in a prokaryote, comprising the steps of: (a) providing a prokaryote that is incapable of synthesizing a farnesyl dibenzodiazepinone; (b) transforming the prokaryote with an expression vector as described herein; and (c) culturing the prokaryote under conditions such that a polypeptide of the invention is expressed and catalyses the synthesis of a farnesyl dibenzodiazepinone compound or analog.
[0027] In one embodiment, the prokaryote is an actinomycete.
[0028] In another embodiment, the vector expresses a polypeptide of SEQ ID NO: $2,4,6,8,10,12,14,16,18,20$, $22,24,26,28,30,32,34,36,38,40,42,44,46,48,50,52,54$, $56,58,60,62,65,67,69,71,74,76,78,80,82,84,86,88,90$, 92, 94 and 96.

## BRIEF DESCRIPTION OF THE FIGURES

[0029] FIG. 1: shows inhibition of tumor growth resulting from administration of 10 to $30 \mathrm{mg} / \mathrm{kg}$ of ECO-04601 to glioblastoma-bearing mice beginning one day after tumor cell inoculation.
[0030] FIG. 2: shows inhibition of tumor growth resulting from administration of $20-30 \mathrm{mg} / \mathrm{kg}$ of ECO-04601 to glio-blastoma-bearing mice beginning ten days after tumor cell inoculation.
[0031] FIG. 3: shows micrographs of tumor sections from mice bearing glioblastoma tumors and treated with saline or ECO-04601. The cell density of tumor treated with ECO04601 appears decreased and nuclei from ECO-04601treated tumor cells are larger and pynotic suggesting a cytotoxic effect.
[0032] FIG. 4: shows the biosynthetic locus of ECO-04601, isolated from Micromonospora sp. strain 046-ECO11, including the positions of cosmids 046 KM and 046 KQ .
[0033] FIGS. 5 to 8 : show the different steps involved in the biosynthetic pathway of ECO-04601. Each of FIGS. 5 to 8 shows the three biosynthetic loci A, B and C where ORFs are represented by arrows. Highlighted ORFs are involved in the steps described in the schematic diagram. The biosynthetic enzymes involved in the steps depicted in schematic diagrams are indicated by their family designation and the respective ORF number in each of Loci A, B and C (e.g., 8/7/7).
[0034] FIG. 5: shows a schematic diagram of the biosynthetic pathway for the production of farnesyl-diphosphate, providing the farnesyl group of ECO-04601.
[0035] FIG. 6: shows a schematic diagram of the biosynthetic pathway for the production of 3-hydroxy-anthranilateadenylate precursor of the dibenzodiazepinone group.
[0036] FIG. 7: shows a schematic diagram of the biosynthetic pathway for the production of 2-amino-6-hydroxy-[1, 4]benzoquinone precursor of the core dibenzodiazepinone.
[0037] FIG. 8: shows a schematic diagram of the biosynthetic pathway for the assembly of the ECO-04601 precursors, farnesyl-diphosphate, 3-hydroxy-anthranilate-adenylate and 2 -amino- 6 -hydroxy-[1,4]benzoquinone.
[0038] FIGS. 9 and 10: show clustal alignments respectively of isoprenyl transferase and adenylating amide synthetase enzymes of locus A with the corresponding enzymes present in loci B and C . In each of the clustal alignments: (i) an asterisk "*" indicates positions which have a single, fully conserved residues; (ii) a colon ":" indicates that one of the following strong groups is fully conserved in a specific position: (S, T or A); (N, E, Q or K); (N, H, Q or K); (N, D, E or Q); (Q, H, R or K); (M, I, L or V); (M, I, L or F); (H orY); and (F, Y or W); and (iii) a period "." indicates that one of the following weaker groups is fully conserved: (C, S or A); (A, T or V); (S, A or G); (S, T, N or K); (S, T, P or A); (S, G, N or D); (S, N, D, E, Q or K); (N, D, E, Q, H or K); (N, E, Q, H, R or K); (F, V, L, I or M): and (H, F or Y). The number at the end of each line indicates the position of the last amino acid of the line within the specific domain.
[0039] FIG. 9: shows an amino acid alignment comparing the isoprenyl transferase (IPTN) enzyme of locus A (SEQ ID NO: 22), isolated from Micronospora sp. strain 046-ECO11, with the isoprenyl transferase enzyme of locus B (SEQ IDNO 90) isolated from Micromonospora echinospora challisensis NRRL 12255, and the partial isoprenyl transferase enzyme of locus C (SEQ ID NO: 94) isolated from Streptomyces carzinostaticus neocarzinostaticus ATCC 15944.
[0040] FIG. 10: shows an amino acid alignment comparing the adenylating amide synthetase (ADSA) enzyme of locus A (SEQ ID NO: 48), isolated from Micronospora sp. strain 046-ECO11, with the adenylating amide synthetase of locus B (SEQ ID NO 92) isolated from Micromonospora echinospora challisensis NRRL 12255, and locus C (SEQ ID NO: 96) isolated from Streptomyces carzinostaticus neocarzinostaticus ATCC 15944.

## DETAILED DESCRIPTION OF THE INVENTION

[0041] The present invention provides isolated and purified polynucleotides that encode farnesyl dibenzodiazepinoneproducing enzymes, i.e., polypeptides from farnesyl diben-zodiazepinone-producing microorganisms, fragments thereof, vectors containing those polynucleotides, and host cells transformed with those vectors. These polynucleotides, fragments thereof, and vectors comprising the polynucleotides can be used as reagents in the production of farnesyl dibenzodiazepinones. The invention also relates to a method for producing new farnesyl dibenzodiazepinones, by selectively altering the genetic information of an organism or by feeding the proteins or a host cell transformed with vectors comprising nucleic acids encoding them, with close analogs of the key intermediates. Portions of the polynucleotide sequences disclosed herein are also useful as primers for the amplification of DNA or as probes to identify related domains from other farnesyl dibenzodiazepinone producing microorganisms.

## I. Definitions

[0042] For convenience, the meaning of certain terms and phrases used in the specification, examples, and appended claims, are provided below.
[0043] As used herein, the term "farnesyl dibenzodiazepinone" refers to a class of dibenzodiazepinone compounds containing a farnesyl moiety. The term includes, but is not limited to, the exemplified compound of the present invention, 10 -farnesyl-4,6,8-trihydroxy-dibenzodiazepin-11-one, which is referred to herein as "ECO-04601."
[0044] The terms "farnesyl dibenzodiazepinone-producing microorganism" and "producer of farnesyl dibenzodiazepinone," as used herein, refer to a microorganism that carries genetic information necessary to produce a farnesyl dibenzodiazepinone compound, whether or not the organism naturally produces the compound. The terms apply equally to organisms in which the genetic information to produce the farnesyl dibenzodiazepinone compound is found in the organism as it exists in its natural environment, and to organisms (host cells) in which the genetic information is introduced by recombinant techniques.
[0045] Specific organisms contemplated herein include, without limitation, organisms of the family Micromonosporaceae, of which preferred genera include Micromonospora, Actinoplanes and Dactylosporangium; the family Streptomycetaceae, of which preferred genera include Streptomyces and Kitasatospora; the family Pseudonocardiaceae, of which preferred genera are Amycolatopsis and Saccharopolyspora; and the family Actinosynnemataceae, of which preferred genera include Saccharothrix and Actinosynnema; however the terms are intended to encompass all organisms containing genetic information necessary to produce a farnesyl dibenzodiazepinone compound. A preferred producer of a farnesyl dibenzodiazepinone compound includes microbial strain 046-ECO11, a deposit of which was made on Mar. 7, 2003, with the International Depository Authority of Canada (IDAC), Bureau of Microbiology, Health Canada, 1015 Arlington Street, Winnipeg, Manitoba, Canada R3E 3R2, under Accession No. IDAC 070303-01
[0046] The term "gene" means the segment of DNA involved in producing a polypeptide chain; it includes regions preceding and following the coding region (leader and trailer) as well as, where applicable, intervening regions (introns) between individual coding segments (exons).
[0047] The terms "gene locus, "gene cluster," and "biosynthetic locus" refer to a group of genes or variants thereof involved in the biosynthesis of a farnesyl dibenzodiazepinone compound. For example, the biosynthetic locus in strain 046ECO11 that directs the production of ECO-04601 referred to herein as "046D" or "locus A", the biosynthetic locus in Micromonospora echinospora challisensis NRRL 12255 referred to herein as " 052 E " or "locus B", the biosynthetic locus in Streptomyces carzinostaticus neocarzinostaticus ATCC 15944 referred to herein as " 237 C " or "locus C", or the corresponding biosynthetic locus from a farnesyl dibenzodi-azepinone-producing microorganism. Genetic modification of gene locus, gene cluster or biosynthetic locus refers to any genetic recombinant techniques known in the art including mutagenesis, inactivation, or replacement of nucleic acids that can be applied to generate variants of ECO-04601.
[0048] A DNA or nucleotide "coding sequence" or "sequence encoding" a particular polypeptide or protein, is a DNA sequence which is transcribed and translated into a polypeptide or protein when placed under the control of an appropriate regulatory sequence.
[0049] "Oligonucleotide" refers to a nucleic acid, generally of at least 10 , preferably 15 and more preferably at least 20 nucleotides in length, preferably no more than 100 nucleotides in length, that are hybridizable to a genomic DNA molecule, a cDNA molecule, or an mRNA molecule encoding a gene, mRNA, cDNA or other nucleic acid of interest.
[0050] A promoter sequence is "operably linked to" a coding sequence recognized by RNA polymerase which initiates transcription at the promoter and transcribes the coding sequence into mRNA
[0051] The term "repl icon" as used herein means any genetic element, such as a plasmid, cosmid, chromosome or virus, that behaves as an autonomous unit of polynucleotide replication within a cell. An "expression vector" or "vector" is a replicon in which another polynucleotide fragment is attached, such as to bring about the replication and/or expression of the attached fragment. "Plasmids" are designated herein by a lower case " p " preceded or followed by capital letters and/or numbers. The starting plasmids disclosed herein are commercially available, publicly available on an unrestricted basis, or can be constructed from available plasmids in accordance with published procedures. In addition, equivalent plasmids to those described herein are known in the art and will be apparent to the skilled artisan.
[0052] The terms "express" and "expression" means allowing or causing the information in a gene or DNA sequence to become manifest, for example producing a protein by activating the cellular functions involved in transcription and translation of a corresponding gene or DNA sequence. A DNA sequence is expressed in or by a cell to form an "expression product" such as a protein. The expression product itself, e.g. the resulting protein, may also be said to be "expressed" by the cell. An expression product can be characterized as intracellular, extracellular or secreted.
[0053] "Digestion" of DNA refers to enzymatic cleavage of the DNA with a restriction enzyme that acts only at certain sequences in the DNA. The various restriction enzymes used herein are commercially available and their reaction conditions, cofactors and other requirements were used as would be known to the ordinary skilled artisan. For analytical purposes, typically $1 \mu \mathrm{~g}$ of plasmid or DNA fragment is used with about 2 units of enzyme in about $20 \mu 1$ of buffer solution. For the purpose of isolating DNA fragments for plasmid construction, typically 5 to $50 \mu$ of DNA are digested with 20 to 250 units of enzyme in a larger volume. Appropriate buffers and substrate amounts for particular enzymes are specified by the manufacturer. Incubation times of about 1 hour at $37^{\circ} \mathrm{C}$. are ordinarily used, but may vary in accordance with the supplier's instructions. After digestion the gel electrophoresis may be performed to isolate the desired fragment.
[0054] The term "isolated" as used herein means that the material is removed from its original environment (e.g. the natural environment where the material is naturally occurring). For example, a naturally occurring polynucleotide or polypeptide present in a living animal is not isolated, but the same polynucleotide or polypeptide, which is separated from some or all of the coexisting materials in the natural system, is isolated. Such polynucleotides could be part of a vector and/or such polynucleotides or polypeptides could be part of a composition, and still be isolated in that the vector or composition is not part of the natural environment.
[0055] The term "restriction fragment" as used herein refers to any linear DNA generated by the action of one or more restriction enzymes.
[0056] The term "transformation" means the introduction of a foreign gene, foreign nucleic acid, DNA or RNA sequence to a host cell, so that the host cell will express the introduced gene or sequence to produce a desired substance, typically a protein or enzyme coded by the introduced gene or sequence. The introduced gene or sequence may also be
called a "cloned" or "foreign" gene or sequence, may include regulatory or control sequences, such as start, stop, promoter, signal, secretion, or other sequences used by a cell's genetic machinery. The gene or sequence may include nonfunctional sequences or sequences with no known function. A host cell that receives and expresses introduced DNA or RNA has been "transformed" and is a "transformant" or a "clone" or "recombinant". The DNA or RNA introduced to a host cell can come from any source, including cells of the same genus or species as the host cell, or cells of a different genus or species.
[0057] The terms "recombinant polynucleotide" and "recombinant polypeptide" as used herein mean a polynucleotide or polypeptide which by virtue of its origin or manipulation is not associated with all or a portion of the polynucleotide or polypeptide with which it is associated in nature and/or is linked to a polynucleotide or polypeptide other than that to which it is linked in nature.
[0058] The term "host cell" as used herein, refer to both prokaryotic and eukaryotic cells which are used as recipients of the recombinant polynucleotides and vectors provided herein. In one embodiment, the host cell is a prokaryote.
[0059] The terms "open reading frame" and "ORF" as used herein refers to a region of a polynucleotide sequence which encodes a polypeptide; this region may represent a portion of a coding sequence or a total coding sequence.
[0060] As used herein and as known in the art, the term "identity" is the relationship between two or more polynucleotide sequences, as determined by comparing the sequences. Identity also means the degree of sequence relatedness between polynucleotide sequences, as determined by the match between strings of such sequences. Identity can be readily calculated (see, e.g., Computation Molecular Biology, Lesk, A. M., eds., Oxford University Press, New York (1998), and Biocomputing: Informatics and Genome Projects, Smith, D. W., ed., Academic Press, New York (1993), both of which are incorporated by reference herein). While there exist a number of methods to measure identity between two polynucleotide sequences, the term is well known to skilled artisans (see, e.g., Sequence Analysis in Molecular Biology, von Heinje, G., Academic Press (1987); and Sequence Analysis Primer, Gribskov., M. and Devereux, J., eds., M. Stockton Press, New York (1991)). Methods commonly employed to determine identity between sequences include, for example, those disclosed in Carillo, H., and Lipman, D., SIAM J. Applied Math. (1988) 48:1073. "Substantially identical," as used herein, means there is a very high degree of homology (preferably $100 \%$ sequence identity) between subject polynucleotide sequences. However, polynucleotides having greater than $90 \%$, or $95 \%$ sequence identity may be used in the present invention, and thus sequence variations that might be expected due to genetic mutation, strain polymorphism, or evolutionary divergence can be tolerated.

## II. Method of Making a Farnesyl Dibenzodiazepinone by Fermentation

[0061] The farnesyl dibenzodiazepinone compounds of the present invention may be biosynthesized by various microorganisms. Microorganisms that may synthesize the compounds of the present invention include but are not limited to bacteria of the order Actinomycetales, also referred to as actinomycetes. Non-limiting examples of members belonging to the genera of Actinomycetes include Nocardia, Geo-
dermatophilus, Actinoplanes, Micromonospora, Nocardioides, Saccharothrix, Amycolatopsis, Kutzneria, Saccharomonospora, Saccharopolyspora, Kitasatospora, Streptomyces, Microbispora, Streptosporangium, and Actinomadura. The taxonomy of actinomycetes is complex and reference is made to Goodfellow, Suprageneric Classification of Actinomycetes (1989); Bergey's Manual of Systematic Bacteriology, Vol. 4 (Williams and Wilkins, Baltimore, pp. 2322-2339); and to Embley and Stackebrandt, "The molecular phylogeny and systematics of the actinomycetes," Annu. Rev. Microbiol. (1994) 48:257-289, each of which is hereby incorporated by reference in its entirety, for genera that may synthesize the compounds of the invention.
[0062] Farnesyl dibenzodiazepinone-producing microorganisms are cultivated in culture medium containing known nutritional sources for actinomycetes. Such media having assimilable sources of carbon, nitrogen, plus optional inorganic salts and other known growth factors at a pH of about 6 to about 9. Suitable media include, without limitation, the growth media provided in Table 1. Microorganisms are cultivated at incubation temperatures of about $18^{\circ} \mathrm{C}$. to about $40^{\circ} \mathrm{C}$. for about 3 to about 40 days.
dibenzodiazepinone compounds can be extracted and isolated from the cultivated culture media by techniques known to a skilled person in the art and/or disclosed herein, including for example centrifugation, chromatography, adsorption, filtration. For example, the cultivated culture media can be mixed with a suitable organic solvent such as $n$-butanol, $n$-butyl acetate or 4-methyl-2-pentanone, the organic layer can be separated for example, by centrifugation followed by the removal of the solvent, by evaporation to dryness or by evaporation to dryness under vacuum. The resulting residue can optionally be reconstituted with for example water, ethanol, ethyl acetate, methanol or a mixture thereof, and re-extracted with a suitable organic solvent such as hexane, carbon tetrachloride, methylene chloride or a mixture thereof. Following removal of the solvent, the compounds may be further purified by the use of standard techniques, such as chromatography.
III. Method of Making a Farnesyl Dibenzodiazepinone by Recombinant Technology
[0063] In another embodiment, the present invention relates to nucleic acid molecules that encode proteins useful

TABLE 1

| Component | Examples of Fermentation Media |  |  |  |  | FA | HI | CL |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | QB | MA | KH | RM | JA |  |  |  |
| $\mathrm{pH} * 1$ | 7.2 | 7.5 | 7 | 6.85 | 7.3 | 7.0 | 7.0 | 7.0 |
| Glucose | 12 |  | 10 | 10 |  | 10 |  |  |
| Sucrose |  |  |  | 100 |  |  |  |  |
| Cane molasses |  |  |  |  |  | 15 |  |  |
| Corn starch |  |  |  |  | 30 |  |  |  |
| Soluble starch | 10 | 25 |  |  |  |  |  |  |
| Potato dextrin |  |  | 20 |  |  | 40 | 20 | 20 |
| Corn steep solid | 5 |  |  |  |  |  |  |  |
| Corn steep liquor | 5 |  |  |  | 15 |  |  |  |
| Dried yeast |  | 2 |  |  |  |  |  |  |
| Yeast extract |  |  | 5 |  |  |  | 8.34 |  |
| Malt extract |  |  |  |  | 35 |  |  |  |
| Pharmamedia ${ }^{\text {TM }}$ | 10 |  |  |  | 15 |  |  |  |
| Glycerol |  |  |  |  |  |  | 30 | 20 |
| NZ-Amine A |  |  | 5 |  |  | 10 |  |  |
| Soybean powder |  | 15 |  |  |  |  |  |  |
| Fish meal |  |  |  |  |  |  |  | 10 |
| Bacto-peptone |  |  |  |  |  |  | 2.5 | 5 |
| $\mathrm{MgSO}_{4} \bullet 7 \mathrm{H}_{2} \mathrm{O}$ |  |  |  |  |  | 1 |  |  |
| $\mathrm{CaCO}_{3}$ |  | 4 | 1 |  | 2 | 2 | 3 | 2 |
| NaCl |  | 5 |  |  |  |  |  |  |
| $\left(\mathrm{NH}_{4}\right)_{2} \mathrm{SO}_{4}$ |  | 2 |  |  |  |  |  | 2 |
| $\mathrm{K}_{2} \mathrm{SO}_{4}$ |  |  |  | 0.25 |  |  |  |  |
| $\mathrm{MgCl}_{2} \cdot 6 \mathrm{H}_{2} \mathrm{O}$ |  |  |  | 10 |  |  |  |  |
| $\mathrm{Na}_{2} \mathrm{HPO}_{4}$ |  |  |  |  |  | 3 |  |  |
| Casamino acid |  |  |  | 0.1 |  |  |  |  |
| Proflo oil ${ }^{\text {TM }}$ (mL/L) | 4 |  |  |  |  |  |  |  |
| MOPS |  |  |  | 21 |  |  |  |  |
| Trace element solution ${ }^{* 2} \mathrm{ml} / \mathrm{L}$ |  |  |  | 2 |  |  |  |  |

Unless otherwise indicated all the ingredients are in $\mathrm{g} / \mathrm{L}$.
${ }^{* 1}$ The pH is to adjusted as marked prior to the addition of $\mathrm{CaCO}_{3}$
${ }^{* 2}$ Trace elements solution contains: $\mathrm{ZnCl}_{2} 40 \mathrm{mg}$; $\mathrm{Fe} \mathrm{Cl}_{3} 6 \mathrm{H}_{2} \mathrm{O}(200 \mathrm{mg}) ; \mathrm{CuCl}_{2} 2 \mathrm{H}_{2} \mathrm{O}$ ( 10
mg); $\mathrm{MnCl}_{2} \bullet 4 \mathrm{H}_{2} \mathrm{O} ; \mathrm{Na}_{2} \mathrm{~B}_{4} \mathrm{O}_{7} \bullet 10 \mathrm{H}_{2} \mathrm{O}(10 \mathrm{mg}) ;\left(\mathrm{NH}_{4}\right)_{6} \mathrm{MO}_{7} \mathrm{O}_{24} \bullet 4 \mathrm{H}_{2} \mathrm{O}(10 \mathrm{mg})$ per litre.

The culture media inoculated with the farnesyl dibenzodiaz-epinone-producing microorganisms may be aerated by incubating the inoculated culture media with agitation, for example, shaking on a rotary shaker, or a shaking water bath. Aeration may also be achieved by the injection of air, oxygen or an appropriate gaseous mixture to the inoculated culture media during incubation. Following cultivation, the farnesyl
in the production of farnesyl benzodiazepinones. Specifically, the present invention provides recombinant DNA vectors and nucleic acid molecules that encode all or part of the biosynthetic locus in strain 046-ECO11, which directs the production of ECO-04601, and is referred to herein as "046D." The invention further includes genetic modification of 046D using conventional genetic recombinant techniques,
such as mutagenesis, inactivation, or replacement of nucleic acids, to produce chemical variants of ECO-04601.
[0064] The invention thus provides a method for making a farnesyl benzodiazepinone compound using a transformed host cell comprising a recombinant DNA vector that encodes one or more of the polypeptides of the present invention, and culturing the host cell under conditions such that farnesyl benzodiazepinone is produced. In one embodiment, the host cell is a prokaryote. In another embodiment, the host cell is an actinomycete. In another embodiment, the host cell is a Streptomyces host cell. In a further embodiment, the host cell is a non-Streptomyces actinomycete such as a Rhodococcus, a Mycobaterium, or an Amycolatopsis specie.
[0065] The invention provides recombinant nucleic acids that produce a variety of farnesyl dibenzodiazepinone compounds that cannot be readily synthesized by chemical methodology alone. The invention allows direct manipulation of 046D biosynthetic locus via genetic engineering of the enzymes involved in the biosynthesis of a farnesyl dibenzodiazepinone according to the invention. The 046D biosynthetic locus is described in Example 5.
[0066] Farnesyl dibenzodazepinones and analogs are also produced by feeding one or more key intermediates or biosynthetic precursors (as defined in FIGS. 5-8) or close structural analogs, to a host cell comprising a recombinant DNA vector that encodes one or more of the polypeptides of the present invention, and culturing the host cell under conditions such that the farnesyl benzodiazepinone or analog is produced. Key intermediates are contacted directly with an isolated protein of the invention to perform the necessary steps for the production of a farnesyl dibenzodiazepinone (e.g., the farnesyl diphopshate and dibenzodiazepinone precursors can be coupled using an IPTN protein of the invention).
[0067] Key intermediates may be commercially available or may be prepared using standard chemical procedures or using the proteins of this invention. For example, farnesyl diphosphate and 3-hydroxyanthranilic acid are commercially available (e.g., Fluka F6892 and Aldrich 148776). 3-Amino5 -hydroxybenzoic acid, a precursor of the 2 -amino- 6 -hydroxybenzoquinone, is prepared as described in Herlt et al (1981), Aust. J. Chem., vol 34, 1319-1324.
[0068] Recombinant DNA Vectors
[0069] Vectors of the invention typically comprise the DNA of a transmissible agent, into which foreign DNA is inserted. A common way to insert one segment of DNA into another segment of DNA involves the use of specific enzymes called restriction enzymes that cleave DNA at specific sites (specific groups of nucleotides) called restriction sites. A "cassette" refers to a DNA coding sequence or segment of DNA that codes for an expression product that can be inserted into a vector at defined restriction sites. The cassette restriction sites are designed to ensure insertion of the cassette in the proper reading frame. Generally, a nucleic acid molecule that encodes a protein useful in the production of a farnesyl dibenzodiazepinone is inserted at one or more restriction sites of the vector DNA, and then is carried by the vector into a prokaryote e.g. actinomycte, by transformation (see below). A segment or sequence of DNA having inserted or added DNA, such as an expression vector, can also be called a "DNA construct". A common type of vector is a "plasmid" which generally is a self-contained molecule of double-stranded DNA, usually of bacterial origin, that can readily accept additional (foreign) DNA and which can be readily introduced into a suitable host cell. A plasmid vector often contains
coding DNA and promoter DNA and has one or more restriction sites suitable for inserting foreign DNA. Coding DNA is a DNA sequence that encodes a particular amino acid sequence for a particular protein or enzyme. In one embodiment of the invention, the coding DNA encodes for polypeptides of SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, $26,28,30,32,34,36,38,41,42,44,46,48,50,52,54,56,58$, $60,62,65,67,69,70,71,74,76,78,80,82,84,86,88,90,92$, 94,96 or 98 that may be useful for the biosynthesis of a farnesyl dibenzodiazepinone.
[0070] Promoter DNA of a recombinant vector is a DNA sequence that initiates, regulates, or otherwise mediates or controls the expression of the coding DNA. Promoter DNA and coding may be from the same or different organisms. Recombinant cloning vectors will often include one or more replication systems for cloning or expression, one or more markers for selection in the host, e.g. antibiotic resistance, and one or more expression cassettes. Vector constructs may be produced using conventional molecular biology and recombinant DNA techniques within the skill of the art. Such techniques are explained fully in the literature. See, e.g., Sambrook, Fritsch \& Maniatis, Molecular Cloning: A Laboratory Manual, Second Edition (1989) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (herein "Sambrook et al., 1989"); DNA Cloning: A Practical Approach, Volumes I and II (D. N. Glover ed. 1985); F. M. Ausubel et al. (eds.), Current Protocols in Molecular Biology, John Wiley \& Sons, Inc. (1994).
[0071] Examples of promoters that function in actinomycetes, e.g. Streptomyces, are taught in U.S. Pat. Nos. $5,830,695$ and 5,466,590. Another example of a transcription promoter useful in Actinomycetes expression vectors is tipA, a promoter inducible by the antibiotic thiostrepton [c.f. Murakami, T., et al., (1989), J. Bacteriol, 171, 1459].
[0072] Transformation of Actinomycetes
[0073] A suitable transformation method for use with an actinomycete comprises forming the actinomycete culture into spheroplasts using lysozyme. A buffer solution containing recombinant DNA vectors and polyethylene glycol is then added, in order to introduce the vector into the host cells, by using either of the methods of Thompson or Keiser [c. f. Thompson, C. J., et al., (1982), J. Bacteriol., 151, 668-677 or Keiser, T. et al. (2000), "Practical Streptomyces Genetics", The John Innes Foundation, Norwich], for example. A thios-trepton-resistance gene is frequently used as a selective marker in the transformation plasmid [c.f. Hopwood, D.A., et al., (1987), "Methods in Enzymology" 153, 116, Academic Press, New York], but the present invention is not limited thereto. Additional methods for the transformation of actinomycetes are taught in U.S. Pat. No. 5,393,665.
[0074] Assay for Farnesyl Dibenzodiazepinone or Biosynthetic Intermediates
[0075] Actinomycetes defective in farnesyl dibenzodiazepinone biosynthesis are transformed with one or more expression vectors encoding one or more proteins in the farnesyl benzodiazepinone biosynthetic pathway, thus restoring farnesyl benzodiazepinone biosynthesis by genetic complementation of the specific defect.
[0076] The presence or absence of farnesyl dibenzodiazepinone or intermediates in the biosynthetic pathway (see FIGS. 5 to 8 ) in a recombinant actinomycete can be determined using methodologies that are well known to persons of skill in the art. For example, ethyl acetate extracts of fermentation media used for the culture of a recombinant actino-
mycete are processed as described in Example 2 and fractions containing farnesyl dibenzodiazepinone or intermediates detected by TLC on commercial Kieselgel $60 \mathrm{~F}_{254}$ plates. Farnesyl dibenzodiazepinone and intermediate compounds are visualized by inspection of dried plates under UV light or by spraying the plates with a spray containing vanillin ( $0.75 \%$ ) and concentrated sulfuric acid ( $1.5 \%, \mathrm{v} / \mathrm{v}$ ) in ethanol and subsequently heating the plate. The exact identity of the compounds separated by TLC is then determined using gas chromatography-mass spectroscopy. Methods of mass spectroscopy are taught in the published U.S. Patent Application No. US2003/0052268.
[0077] Mutagenesis
[0078] The invention allows direct manipulation of 046D biosynthetic locus via genetic engineering of the enzymes involved in the biosynthesis of a farnesyl benzodiazepinone according to the invention.
[0079] A number of methods are known in the art that permit the random as well as targeted mutation of the DNA sequences of the invention (see for example, Ausubel et. al. Short Protocols in Molecular Biology (1995) 3rd Ed. John Wiley \& Sons, Inc.). In addition, there are a number of commercially available kits for site-directed mutagenesis, including both conventional and PCR-based methods. Examples include the EXSITETM PCR-Based Site-directed Mutagenesis Kit available from Stratagene (Catalog No. 200502) and the QUIKCHANGETM Site-directed mutagenesis Kit from Stratagene (Catalog No. 200518), and the CHAMELEON® double-stranded Site-directed mutagenesis kit, also from Stratagene (Catalog No. 200509).
[0080] In addition the nucleotides of the invention may be generated by insertional mutation or truncation (N-terminal, internal or C-terminal) according to methodology known to a person skilled in the art.
[0081] Older methods of site-directed mutagenesis known in the art rely on sub-cloning of the sequence to be mutated into a vector, such as an M13 bacteriophage vector, that allows the isolation of single-stranded DNA template. In these methods, one anneals a mutagenic primer (i.e., a primer capable of annealing to the site to be mutated but bearing one or more mismatched nucleotides at the site to be mutated) to the single-stranded template and then polymerizes the complement of the template starting from the $3^{\prime}$ end of the mutagenic primer. The resulting duplexes are then transformed into host bacteria and plaques are screened for the desired mutation.
[0082] More recently, site-directed mutagenesis has employed PCR methodologies, which have the advantage of not requiring a single-stranded template. In addition, methods have been developed that do not require sub-cloning. Several issues must be considered when PCR-based site-directed mutagenesis is performed. First, in these methods it is desirable to reduce the number ofPCR cycles to prevent expansion of undesired mutations introduced by the polymerase. Second, a selection must be employed in order to reduce the number of non-mutated parental molecules persisting in the reaction. Third, an extended-length PCR method is preferred in order to allow the use of a single PCR primer set. And fourth, because of the non-template-dependent terminal extension activity of some thermostable polymerases it is often necessary to incorporate an end-polishing step into the procedure prior to blunt-end ligation of the PCR-generated mutant product.
[0083] The protocol described below accommodates these considerations through the following steps. First, the template concentration used is approximately 1000 -fold higher than that used in conventional PCR reactions, allowing a reduction in the number of cycles from 25-30 down to 5-10 without dramatically reducing product yield. Second, the restriction endonuclease Dpn I (recognition target sequence: 5-Gm6ATC-3, where the A residue is methylated) is used to select against parental DNA, since most common strains of $E$. coli Dam methylate their DNA at the sequence 5 -GATC-3. Third, Taq Extender is used in the PCR mix in order to increase the proportion of long (i.e., full plasmid length) PCR products. Finally, Pfu DNA polymerase is used to polish the ends of the PCR product prior to intramolecular ligation using T4 DNA ligase.
[0084] A non-limiting example for the isolation of mutant polynucleotides is described in detail as follows:
[0085] Plasmid template DNA (approximately 0.5 pmole) is added to a PCR cocktail containing: $1 \times$ mutagenesis buffer ( 20 mM Tris $\mathrm{HCl}, \mathrm{pH} 7.5 ; 8 \mathrm{mM} \mathrm{MgCl} 2 ; 40 \mu \mathrm{~g} / \mathrm{ml} \mathrm{BSA}$ ); 12-20 pmole of each primer (one of skill in the art may design a mutagenic primer as necessary, giving consideration to those factors such as base composition, primer length and intended buffer salt concentrations that affect the annealing characteristics of oligonucleotide primers; one primer must contain the desired mutation, and one (the same or the other) must contain a 5 ' phosphate to facilitate later ligation), 250 $\mu$ Meach dNTP, 2.5 U Taq DNA polymerase, and 2.5 U of Taq Extender (Available from Stratagene; See Nielson et al. (1994) Strategies $7: 27$, and U.S. Pat. No. $5,556,772$ ). Primers can be prepared using the triester method of Matteucci et al., 1981, J.Am. Chem. Soc. 103:3185-3191, incorporated herein by reference. Alternatively automated synthesis may be preferred, for example, on a Biosearch 8700 DNA Synthesizer using cyanoethyl phosphoramidite chemistry.
[0086] The PCR cycling is performed as follows: 1 cycle of 4 min at $94^{\circ} \mathrm{C}$., 2 min at $50^{\circ} \mathrm{C}$. and 2 min at $72^{\circ} \mathrm{C}$.; followed by $5-10$ cycles of 1 min at $94^{\circ} \mathrm{C}$., 2 min at $54^{\circ} \mathrm{C}$. and 1 min at $72^{\circ} \mathrm{C}$. The parental template DNA and the linear, PCRgenerated DNA incorporating the mutagenic primer are treated with DpnI (10 U) and Pfu DNA polymerase (2.5 U). This results in the DpnI digestion of the in vivo methylated parental template and hybrid DNA and the removal, by Pfu DNA polymerase, of the non-template-directed Taq DNA polymerase-extended base(s) on the linear PCR product. The reaction is incubated at $37^{\circ} \mathrm{C}$. for 30 min and then transferred to $72^{\circ} \mathrm{C}$. for an additional 30 min . Mutagenesis buffer ( 115 ul of $1 \times$ ) containing 0.5 mMATP is added to the DpnI-digested, Pfu DNA polymerase-polished PCR products. The solution is mixed and 10 ul are removed to a new microfuge tube and T4 DNA ligase ( $2-4 \mathrm{U}$ ) is added. The ligation is incubated for greater than 60 min at $37^{\circ} \mathrm{C}$. Finally, the treated solution is transformed into competent $E$. coli according to standard methods.
[0087] Methods of random mutagenesis, which will result in a panel of mutants bearing one or more randomly situated mutations, exist in the art. Such a panel of mutants may then be screened for those exhibiting reduced uracil detection activity relative to the wild-type polymerase (e.g., by measuring the incorporation of 10 nmoles of dNTPs into polymeric form in 30 minutes in the presence of $200 \mu \mathrm{M}$ dUTP and at the optimal temperature for a given DNA polymerase). An example of a method for random mutagenesis is the socalled "error-prone PCR method". As the name implies, the
method amplifies a given sequence under conditions in which the DNA polymerase does not support high fidelity incorporation. The conditions encouraging error-prone incorporation for different DNA polymerases vary, however one skilled in the art may determine such conditions for a given enzyme. A key variable for many DNA polymerases in the fidelity of amplification is, for example, the type and concentration of divalent metal ion in the buffer. The use of manganese ion and/or variation of the magnesium or manganese ion concentration may therefore be applied to influence the error rate of the polymerase.
[0088] Genes for desired mutant polypeptides generated by mutagenesis may be sequenced to identify the sites and number of mutations. For those mutants comprising more than one mutation, the effect of a given mutation may be evaluated by introduction of the identified mutation to the wild-type gene by site-directed mutagenesis in isolation from the other mutations borne by the particular mutant. Screening assays of the single mutant thus produced will then allow the determination of the effect of that mutation alone.

## IV. Genes and Proteins for the Production of ECO-04601

[0089] As discussed in more detail below, the isolated, purified or enriched nucleic acids of one of SEQ ID NOS: 3, 5, 7, $9,11,13,15,17,19,21,23,25,27,29,31,33,35,37,39,41$, $43,45,47,49,51,53,55,57,59,61,63,66,68,70,72,75,77$, $79,81,83,85,87$ and 89 may be used to prepare one of the polypeptides of SEQ ID NOS: $2,4,6,8,10,12,14,16,18,20$, $22,24,26,28,30,32,34,36,38,41,42,44,46,48,50,52,54$, $56,58,60,62,65,67,69,70,71,74,76,78,80,82,84,86$ and 88 , respectively, or fragments comprising at least $50,75,100$, $200,300,500$ or more consecutive amino acids of one of the polypeptides of SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, $22,24,26,28,30,32,34,36,38,41,42,44,46,48,50,52,54$, $56,58,60,62,65,67,69,70,71,74,76,78,80,82,84,86$ and 88.
[0090] Accordingly, another aspect of the present invention is an isolated, purified or enriched nucleic acid which encodes one of the polypeptides of SEQ ID NOS: $2,4,6,8,10,12,14$, $16,18,20,22,24,26,28,30,32,34,36,38,41,42,44,46,48$, $50,52,54,56,58,60,62,65,67,69,70,71,74,76,78,80,82$, 84,86 and 88 or fragments comprising at least $50,75,100$, $150,200,300$ or more consecutive amino acids of one of the polypeptides of SEQ ID NOS: $2,4,6,8,10,12,14,16,18,20$, $22,24,26,28,30,32,34,36,38,41,42,44,46,48,50,52,54$, $56,58,60,62,65,67,69,70,71,74,76,78,80,82,84,86$ and 88. The coding sequences of these nucleic acids may be identical to one of the coding sequences of one of the nucleic acids of SEQ ID NOS: $3,5,7,9,11,13,15,17,19,21,23,25$, $27,29,31,33,35,37,39,41,43,45,47,49,51,53,55,57,59$, $61,63,66,68,70,72,75,77,79,81,83,85,87$ and 89 or a fragment thereof, or may be different coding sequences which encode one of the polypeptides of SEQ ID NOS: 2, 4, $6,8,10,12,14,16,18,20,22,24,26,28,30,32,34,36,38$, $41,42,44,46,48,50,52,54,56,58,60,62,65,67,69,70,71$, $74,76,78,80,82,84,86$ and 88 or fragments comprising at least $50,75,100,150,200,300$ consecutive amino acids of one of the polypeptides of SEQ ID NOS: $2,4,6,8,10,12,14$, $16,18,20,22,24,26,28,30,32,34,36,38,41,42,44,46,48$, $50,52,54,56,58,60,62,65,67,69,70,71,74,76,78,80,82$, 84, 86 and 88 as a result of the redundancy or degeneracy of the genetic code. The genetic code is well known to those of skill in the art and can be obtained, for example, from Stryer, Biochemistry, $3^{\text {rd }}$ edition, W.H. Freeman \& Co., New York.
[0091] The isolated, purified or enriched nucleic acid which encodes one of the polypeptides of SEQ ID NOS: 2, 4, $6,8,10,12,14,16,18,20,22,24,26,28,30,32,34,36,38$, $41,42,44,46,48,50,52,54,56,58,60,62,65,67,69,70,71$, $74,76,78,80,82,84,86$ and 88 may include, but is not limited to: (1) only the coding sequences of one of SEQ ID NOS: 3 , $5,7,9,11,13,15,17,19,21,23,25,27,29,31,33,35,37,39$, $41,43,45,47,49,51,53,55,57,59,61,63,66,68,70,72,75$, $77,79,81,83,85,87$ and 89 ; (2) the coding sequences of SEQ ID NOS: $3,5,7,9,11,13,15,17,19,21,23,25,27,29,31$, $33,35,37,39,41,43,45,47,49,51,53,55,57,59,61,63,66$, $68,70,72,75,77,79,81,83,85,87$ and 89 and additional coding sequences, such as leader sequences or proprotein; and (3) the coding sequences of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, $47,49,51,53,55,57,59,61,63,66,68,70,72,75,77,79,81$, $83,85,87$ and 89 and non-coding sequences, such as noncoding sequences $5^{\prime}$ and/or $3^{\prime}$ of the coding sequence. Thus, as used herein, the term "polynucleotide encoding a polypeptide" encompasses a polynucleotide that includes only coding sequence for the polypeptide as well as a polynucleotide that includes additional coding and/or non-coding sequence.
[0092] The invention relates to polynucleotides based on SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, $31,33,35,37,39,41,43,45,47,49,51,53,55,57,59,61,63$, $66,68,70,72,75,77,79,81,83,85,87$ and 89 but having polynucleotide changes that are "silent", for example changes which do not alter the amino acid sequence encoded by the polynucleotides of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, $21,23,25,27,29,31,33,35,37,39,41,43,45,47,49,51,53$, $55,57,59,61,63,66,68,70,72,75,77,79,81,83,85,87$ and 89. The invention also relates to polynucleotides which have nucleotide changes which result in amino acid substitutions, additions, deletions, fusions and truncations of the polypeptides of SEQ ID NOS: $2,4,6,8,10,12,14,16,18,20,22,24$, $26,28,30,32,34,36,38,41,42,44,46,48,50,52,54,56,58$, $60,62,65,67,69,70,71,74,76,78,80,82,84,86$ and 88. Such nucleotide changes may be introduced using techniques such as site directed mutagenesis, random chemical mutagenesis, exonuclease III deletion, and other recombinant DNA techniques.
[0093] The isolated, purified or enriched nucleic acids of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, $31,33,35,37,39,41,43,45,47,49,51,53,55,57,59,61,63$, $66,68,70,72,75,77,79,81,83,85,87$ and 89 , the sequences complementary thereto, or a fragment comprising at least 100, 150, 200, 300, 400 or more consecutive bases of one of the sequence of SEQ ID NOS: $3,5,7,9,11,13,15,17,19,21$, $23,25,27,29,31,33,35,37,39,41,43,45,47,49,51,53,55$, $57,59,61,63,66,68,70,72,75,77,79,81,83,85,87$ and 89 , or the sequences complementary thereto may be used as probes to identify and isolate DNAs encoding the polypeptides of SEQ ID NOS: $2,4,6,8,10,12,14,16,18,20,22,24$, $26,28,30,32,34,36,38,41,42,44,46,48,50,52,54,56,58$, $60,62,65,67,69,70,71,74,76,78,80,82,84,86$ and 88 respectively. In such procedures, a genomic DNA library is constructed from a sample microorganism or a sample containing a microorganism capable of producing a farnesyl dibenzodiazepinone. The genomic DNA library is then contacted with a probe comprising a coding sequence or a fragment of the coding sequence, encoding one of the polypeptides of SEQ ID NOS: $2,4,6,8,10,12,14,16,18,20,22,24$, $26,28,30,32,34,36,38,41,42,44,46,48,50,52,54,56,58$, $60,62,65,67,69,70,71,74,76,78,80,82,84,86$ and 88 , or
a fragment thereof under conditions which permit the probe to specifically hybridize to sequences complementary thereto. In a preferred embodiment, the probe is an oligonucleotide of about 10 to about 30 nucleotides in length designed based on a nucleic acid of SEQ ID NOS: 3, 5, 7, 9 , $11,13,15,17,19,21,23,25,27,29,31,33,35,37,39,41,43$, $45,47,49,51,53,55,57,59,61,63,66,68,70,72,75,77,79$, $81,83,85,87$ and 89 . Genomic DNA clones which hybridize to the probe are then detected and isolated. Procedures for preparing and identifying DNA clones of interest are disclosed in Ausubel et al., Current Protocols in Molecular Biology, John Wiley 503 Sons, Inc. 1997; and Sambrook et al., Molecular Cloning: A Laboratory Manual 2d Ed., Cold Spring Harbor Laboratory Press, 1989. In another embodiment, the probe is a restriction fragment or a PCR amplified nucleic acid derived from SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, $17,19,21,23,25,27,29,31,33,35,37,39,41,43,45,47,49$, $51,53,55,57,59,61,63,66,68,70,72,75,77,79,81,83,85$, 87 and 89.
[0094] The isolated, purified or enriched nucleic acids of SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, $31,33,35,37,39,41,43,45,47,49,51,53,55,57,59,61,63$, $66,68,70,72,75,77,79,81,83,85,87$ and 89 , the sequences complementary thereto, or a fragment comprising at least 10 , $15,20,25,30,35,40,50,75,100,150,200,300,400$ or 500 consecutive bases of one of the sequences of SEQ ID NOS: 3 , $5,7,9,11,13,15,17,19,21,23,25,27,29,31,33,35,37,39$, $41,43,45,47,49,51,53,55,57,59,61,63,66,68,70,72,75$, $77,79,81,83,85,87$ and 89 or the sequences complementary thereto may be used as probes to identify and isolate related nucleic acids. In some embodiments, the related nucleic acids may be genomic DNAs (or cDNAs) from potential farnesyl dibenzodiazepinone producers. In such procedures, a nucleic acid sample containing nucleic acids from a potential farnesyl dibenzodiazepinone producer is contacted with the probe under conditions that permit the probe to specifically hybridize to related sequences. The nucleic acid sample may be a genomic DNA (or cDNA) library from the potential farnesyl dibenzodiazepinone-producer. Hybridization of the probe to nucleic acids is then detected using any of the methods described above.
[0095] Hybridization may be carried out under conditions of low stringency, moderate stringency or high stringency. As an example of nucleic acid hybridization, a polymer membrane containing immobilized denatured nucleic acids is first prehybridized for 30 minutes at $45^{\circ} \mathrm{C}$. in a solution consisting of $0.9 \mathrm{M} \mathrm{NaCl}, 50 \mathrm{mMNaH}{ }_{2} \mathrm{PO}_{4}, \mathrm{pH} 7.0,5.0 \mathrm{mMNa}_{2}$ EDTA, $0.5 \%$ SDS, $10 \times$ Denhardt's, and $0.5 \mathrm{mg} / \mathrm{ml}$ polyriboadenylic acid. Approximately $2 \times 10^{7} \mathrm{cpm}$ (specific activity $4-9 \times 10^{8}$ $\mathrm{cpm} / \mathrm{ug}$ ) of ${ }^{32} \mathrm{P}$ end-labeled oligonucleotide probe are then added to the solution. After 12-16 hours of incubation, the membrane is washed for 30 minutes at room temperature in $1 \times$ SET ( $150 \mathrm{mM} \mathrm{NaCl}, 20 \mathrm{mM}$ Tris hydrochloride, $\mathrm{pH} 7.8,1$ $\mathrm{mM} \mathrm{Na} 2_{2}$ EDTA) containing $0.5 \%$ SDS, followed by a 30 minute wash in fresh $1 \times$ SET at $\mathrm{Tm}-10^{\circ} \mathrm{C}$. for the oligonucleotide probe where Tm is the melting temperature. The membrane is then exposed to autoradiographic film for detection of hybridization signals.
[0096] By varying the stringency of the hybridization conditions used to identify nucleic acids, such as genomic DNAs or cDNAs, which hybridize to the detectable probe, nucleic acids having different levels of homology to the probe can be identified and isolated. Stringency may be varied by conducting the hybridization at varying temperatures below the melt-
ing temperatures of the probes. The melting temperature of the probe may be calculated using the following formulas:
[0097] For oligonucleotide probes between 14 and 70 nucleotides in length the melting temperature (Tm) in degrees Celcius may be calculated using the formula: $\mathrm{Tm}=81.5+16$. $6(\log [\mathrm{Na}+])+0.41($ fraction $\mathrm{G}+\mathrm{C})-(600 / \mathrm{N})$ where N is the length of the oligonucleotide.
[0098] If the hybridization is carried out in a solution containing formamide, the melting temperature may be calculated using the equation $\mathrm{Tm}=81.5+16.6(\log [\mathrm{Na}+])+0.41$ (fraction $\mathrm{G}+\mathrm{C})-(0.63 \%$ formamide $)-(600 / \mathrm{N})$ where N is the length of the probe.
[0099] Prehybridization may be carried out in $6 \times$ SSC, $5 \times$ Denhardt's reagent, $0.5 \%$ SDS, $0.1 \mathrm{mg} / \mathrm{ml}$ denatured fragmented salmon sperm DNA or $6 \times \mathrm{SSC}, 5 \times$ Denhardt's reagent, $0.5 \%$ SDS, $0.1 \mathrm{mg} / \mathrm{ml}$ denatured fragmented salmon sperm DNA, $50 \%$ formamide. The composition of the SSC and Denhardt's solutions are listed in Sambrook et al., supra.
[0100] Hybridization is conducted by adding the detectable probe to the hybridization solutions listed above. Where the probe comprises double stranded DNA, it is denatured by incubating at elevated temperatures and quickly cooling before addition to the hybridization solution. It may also be desirable to similarly denature single stranded probes to eliminate or diminish formation of secondary structures or oligomerization. The filter is contacted with the hybridization solution for a sufficient period of time to allow the probe to hybridize to cDNAs or genomic DNAs containing sequences complementary thereto or homologous thereto. For probes over 200 nucleotides in length, the hybridization may be carried out at $15-25^{\circ} \mathrm{C}$. below the Tm . For shorter probes, such as oligonucleotide probes, the hybridization may be conducted at $5-10^{\circ} \mathrm{C}$. below the Tm. Preferably, the hybridization is conducted in $6 \times$ SSC, for shorter probes. Preferably, the hybridization is conducted in $50 \%$ formamide containing solutions, for longer probes. All the foregoing hybridizations would be considered to be examples of hybridization performed under conditions of high stringency.
[0101] Following hybridization, the filter is washed for at least 15 minutes in $2 \times$ SSC, $0.1 \%$ SDS at room temperature or higher, depending on the desired stringency. The filter is then washed with $0.1 \times \mathrm{SSC}, 0.5 \% \mathrm{SDS}$ at room temperature (again) for 30 minutes to 1 hour. Nucleic acids which have hybridized to the probe are identified by conventional autoradiography and non-radioactive detection methods.
[0102] The above procedure may be modified to identify nucleic acids having decreasing levels of homology to the probe sequence. For example, to obtain nucleic acids of decreasing homology to the detectable probe, less stringent conditions may be used. For example, the hybridization temperature may be decreased in increments of $5^{\circ} \mathrm{C}$. from $68^{\circ} \mathrm{C}$. to $42^{\circ} \mathrm{C}$. in a hybridization buffer having a $\mathrm{Na}+$ concentration of approximately 1 M . Following hybridization, the filter may be washed with $2 \times \mathrm{SSC}, 0.5 \% \mathrm{SDS}$ at the temperature of hybridization. These conditions are considered to be "moderate stringency" conditions above $50^{\circ} \mathrm{C}$. and "low stringency" conditions below $50^{\circ} \mathrm{C}$. A specific example of "moderate stringency" hybridization conditions is when the above hybridization is conducted at $55^{\circ} \mathrm{C}$. A specific example of "low stringency" hybridization conditions is when the above hybridization is conducted at $45^{\circ} \mathrm{C}$.
[0103] Alternatively, the hybridization may be carried out in buffers, such as $6 \times \mathrm{SSC}$, containing formamide at a temperature of $42^{\circ} \mathrm{C}$. In this case, the concentration of forma-
mide in the hybridization buffer may be reduced in $5 \%$ increments from $50 \%$ to $0 \%$ to identify clones having decreasing levels of homology to the probe. Following hybridization, the filter may be washed with $6 \times \mathrm{SSC}, 0.5 \% \mathrm{SDS}$ at $50^{\circ} \mathrm{C}$. These conditions are considered to be "moderate stringency" conditions above $25 \%$ formamide and "low stringency" conditions below $25 \%$ formamide. A specific example of "moderate stringency" hybridization conditions is when the above hybridization is conducted at $30 \%$ formamide. A specific example of "low stringency" hybridization conditions is when the above hybridization is conducted at $10 \%$ formamide. Nucleic acids which have hybridized to the probe are identified by conventional autoradiography and non-radioactive detection methods. Examples of conditions of different stringency are also provided in Table 2.
fragments comprising at least $50,75,100,150,200$ or 300 consecutive amino acids thereof. As discussed herein, such polypeptides may be obtained by inserting a nucleic acid encoding the polypeptide into a vector such that the coding sequence is operably linked to a sequence capable of driving the expression of the encoded polypeptide in a suitable host cell. For example, the expression vector may comprise a promoter, a ribosome binding site for translation initiation and a transcription terminator. The vector may also include appropriate sequences for modulating expression levels, an origin of replication and a selectable marker.
[0107] Promoters suitable for expressing the polypeptide or fragment thereof in bacteria include the $E$. coli lac or trp promoters, the lad promoter, the lacZ promoter, the T3 promoter, the T 7 promoter, the gpt promoter, the lambda $\mathrm{P}_{R}$

TABLE 2

|  | Very High Stringency <br> (detects sequences sharing at least 90\% identity) |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
| Hybridization | in | 5 x SCC at | $65^{\circ} \mathrm{C}$. | 16 hours |
| Wash twice | in | 2 x SCC at | room temeprature | 15 mnutes each |
| Wash twice | in | 0.5 x SCC at | $65^{\circ} \mathrm{C}$. | 20 minutes each |
|  |  | High Stringency <br> (detects sequences sharing at least $80 \%$ identity) |  |  |
| Hybridization | in | 5 x SCC at | $65^{\circ} \mathrm{C}$ | 16 hours |
| Wash twice | in | 2 x SCC at | room temeprature | 20 mnutes each |
| Wash once | in | 1x SCC at | $55^{\circ} \mathrm{C}$. | 30 minutes each |
|  |  | Low St sequences shar | ncy <br> t least 50\% identity) |  |
| Hybridization | in | 6 x SCC at | room temperature | 16 hours |
| Wash twice | in | 3 x SCC at | room temeprature | 20 minutes each |

[0104] The preceding methods may be used to isolate nucleic acids having at least $97 \%$, at least $95 \%$, at least $90 \%$, at least $85 \%$, at least $80 \%$, or at least $70 \%$ sequence identity to a nucleic acid sequence selected from the group consisting of the sequences of SEQ ID NOS: $3,5,7,9,11,13,15,17,19$, $21,23,25,27,29,31,33,35,37,39,41,43,45,47,49,51,53$, $55,57,59,61,63,66,68,70,72,75,77,79,81,83,85,87$ and 89. The isolated nucleic acid may have a coding sequence that is a naturally occurring allelic variant of one of the coding sequences described herein. Such allelic variant may have a substitution, deletion or addition of one or more nucleotides when compared to the nucleic acids of SEQ ID NOS: 3, 5, 7, $9,11,13,15,17,19,21,23,25,27,29,31,33,35,37,39,41$, $43,45,47,49,51,53,55,57,59,61,63,66,68,70,72,75,77$, $79,81,83,85,87$ and 89 , or the sequences complementary thereto.
[0105] Additionally, the above procedures may be used to isolate nucleic acids which encode polypeptides having at least $99 \%$, at least $95 \%$, at least $90 \%$, at least $85 \%$, at least $80 \%$, or at least $70 \%$ identity to a polypeptide having the sequence of one of SEQ ID NOS: $2,4,6,8,10,12,14,16,18$, $20,22,24,26,28,30,32,34,36,38,41,42,44,46,48,50,52$, $54,56,58,60,62,65,67,69,70,71,74,76,78,80,82,84,86$ and 88 or fragments comprising at least $50,75,100,150,200$, 300 consecutive amino acids thereof.
[0106] Another aspect of the present invention is an isolated or purified polypeptide comprising the sequence of one of SEQ ID NOS: $2,4,6,8,10,12,14,16,18,20,22,24,26$, $28,30,32,34,36,38,41,42,44,46,48,50,52,54,56,58,60$, $62,65,67,69,70,71,74,76,78,80,82,84,86$ and 88 or
promoter, the lambda $\mathrm{P}_{L}$ promoter, promoters from operons encoding glycolytic enzymes such as 3-phosphoglycerate kinase (PGK), and the acid phosphatase promoter. Fungal promoters include the a factor promoter. Eukaryotic promoters include the CMV immediate early promoter, the HSV thymidine kinase promoter, heat shock promoters, the early and late SV40 promoter, LTRs from retroviruses, and the mouse metallothionein-I promoter. Other promoters known to control expression of genes in prokaryotic or eukaryotic cells or their viruses may also be used.
[0108] Mammalian expression vectors may also comprise an origin of replication, any necessary ribosome binding sites, a polyadenylation site, splice donors and acceptor sites, transcriptional termination sequences, and $5^{\prime}$ flanking nontranscribed sequences. In some embodiments, DNA sequences derived from the SV40 splice and polyadenylation sites may be used to provide the required nontranscribed genetic elements.
[0109] Vectors for expressing the polypeptide or fragment thereof in eukaryotic cells may also contain enhancers to increase expression levels. Enhancers are cis-acting elements of DNA, usually from about 10 to about 300 bp in length that act on a promoter to increase its transcription. Examples include the SV40 enhancer on the late side of the replication origin bp 100 to 270 , the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and the adenovirus enhancers.
[0110] In addition, the expression vectors preferably contain one or more selectable marker genes to permit selection of host cells containing the vector. Examples of selectable
markers that may be used include genes encoding dihydrofolate reductase or genes conferring neomycin resistance for eukaryotic cell culture, genes conferring tetracycline or ampicillin resistance in E. coli, and the S. cerevisiae TRP1 gene.
[0111] The appropriate DNA sequence may be inserted into the vector by a variety of procedures. In general, the DNA sequence is ligated to the desired position in the vector following digestion of the insert and the vector with appropriate restriction endonucleases. Alternatively, appropriate restriction enzyme sites can be engineered into a DNA sequence by PCR. A variety of cloning techniques are disclosed in Ausbel et al. Current Protocols in Molecular Biology, John Wiley 503 Sons, Inc. 1997 and Sambrook et al., Molecular Cloning: A Laboratory Manual 2d Ed., Cold Spring Harbour Laboratory Press, 1989. Such procedures and others are deemed to be within the scope of those skilled in the art.
[0112] The vector may be, for example, in the form of a plasmid, a viral particle, or a phage. Other vectors include derivatives of chromosomal, nonchromosomal and synthetic DNA sequences, viruses, bacterial plasmids, phage DNA, baculovirus, yeast plasmids, vectors derived from combinations of plasmids and phage DNA, viral DNA such as vaccinia, adenovirus, fowl pox virus, and pseudorabies. A variety of cloning and expression vectors for use with prokaryotic and eukaryotic hosts are described by Sambrook et al., Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor, N.Y., (1989), incorporated by reference in its entirety for all purposes.
[0113] Particular bacterial vectors which may be used include the commercially available plasmids comprising genetic elements of the well known cloning vector pBR322 (ATCC 37017), pKK223-3 (Pharmacia Fine Chemicals, Uppsala, Sweden), pGEM1 (Promega Biotec, Madison, Wis., USA) pQE70, pQE60, pQE-9 (Qiagen), pD10, phiX174, pBluescript ${ }^{\text {TM }}$ II KS, $\mathrm{pNH} 8 \mathrm{~A}, \mathrm{pNH} 16 \mathrm{a}, \mathrm{pNH} 18 \mathrm{~A}, \mathrm{pNH} 46 \mathrm{~A}$ (Stratagene), ptrc99a, pKK223-3, pKK233-3, pDR540, pRIT5 (Pharmacia), pKK232-8 and pCM7. Particular eukaryotic vectors include pSV 2 CAT , pOG44, $\mathrm{pXT} 1, \mathrm{pSG}$ (Stratagene), pSVK3, PBPV, PMSG, and PSVL (Pharmacia). However, any other vector may be used as long as it is replicable and stable in the host cell.
[0114] The host cell may be any of the host cells familiar to those skilled in the art, including prokaryotic cells or eukaryotic cells. As representative examples of appropriate hosts, there may be mentioned: bacteria cells, such as E. coli, Streptomyces lividans, Streptomyces griseofuscus, Streptomyces ambofaciens, Rhodococcus, Amycolatopsis, Bacillus subtilis, Salmonella typhimurium and various species within the genera Pseudomonas, Streptomyces, Bacillus, and Staphylococcus, fungal cells, such as yeast, insect cells such as Drosophila S2 and Spodoptera Sf 9 , animal cells such as $\mathrm{CHO}, \mathrm{COS}$ or Bowes melanoma, and adenoviruses. The selection of an appropriate host is within the abilities of those skilled in the art, see for example Manual of Industrial Microbiology and Biotechnology, $2^{\text {nd }}$ Edition, ASM Press, Washington D.C., incorporated by reference in its entirety, and more particularly Sections IV, V and VII.
[0115] The vector may be introduced into the host cells using any of a variety of techniques, including electroporation transformation, transfection, transduction, viral infection, gene guns, or Ti-mediated gene transfer. Where appropriate, the engineered host cells can be cultured in conventional nutrient media modified as appropriate for activating promoters, selecting transformants or amplifying the
genes of the present invention. Following transformation of a suitable host strain and growth of the host strain to an appropriate cell density, the selected promoter may be induced by appropriate means (e.g., temperature shift or chemical induction) and the cells may be cultured for an additional period to allow them to produce the desired polypeptide or fragment thereof.
[0116] Cells are typically harvested by centrifugation, disrupted by physical or chemical means, and the resulting crude extract is retained for further purification. Microbial cells employed for expression of proteins can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing agents. Such methods are well known to those skilled in the art. The expressed polypeptide or fragment thereof can be recovered and purified from recombinant cell cultures by methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography. Protein refolding steps can be used, as necessary, in completing configuration of the polypeptide. If desired, high performance liquid chromatography (HPLC) can be employed for final purification steps.
[0117] Various mammalian cell culture systems can also be employed to express recombinant protein. Examples of mammalian expression systems include the COS-7 lines of monkey kidney fibroblasts (described by Gluzman, Cell, 23:175 (1981)), and other cell lines capable of expressing proteins from a compatible vector, such as the C127,3T3, CHO, HeLa and BHK cell lines. The constructs in host cells can be used in a conventional manner to produce the gene product encoded by the recombinant sequence. Polypeptides of the invention may or may not also include an initial methionine amino acid residue.
[0118] Alternatively, the polypeptides of SEQ ID NOS: 2, $4,6,8,10,12,14,16,18,20,22,24,26,28,30,32,34,36,38$ $41,42,44,46,48,50,52,54,56,58,60,62,65,67,69,70,71$, $74,76,78,80,82,84,86$ and 88 or fragments comprising at least $50,75,100,150,200$ or 300 consecutive amino acids thereof can be synthetically produced by conventional peptide synthesizers. In other embodiments, fragments or portions of the polynucleotides may be employed for producing the corresponding full-length polypeptide by peptide synthesis; therefore, the fragments may be employed as intermediates for producing the full-length polypeptides.
[0119] Cell-free translation systems can also be employed to produce one of the polypeptides of SEQ ID NOS: $2,4,6,8$, $10,12,14,16,18,20,22,24,26,28,30,32,34,36,38,41,42$, $44,46,48,50,52,54,56,58,60,62,65,67,69,70,71,74,76$ $78,80,82,84,86$ and 88 or fragments comprising at least 50 , $75,100,150,200$ or 300 consecutive amino acids thereof using mRNAs transcribed from a DNA construct comprising a promoter operably linked to a nucleic acid encoding the polypeptide or fragment thereof. In some embodiments, the DNA construct may be linearized prior to conducting an in vitro transcription reaction. The transcribed mRNA is then incubated with an appropriate cell-free translation extract, such as a rabbit reticulocyte extract, to produce the desired polypeptide or fragment thereof.
[0120] The present invention also relates to variants of the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, $22,24,26,28,30,32,34,36,38,41,42,44,46,48,50,52,54$
$56,58,60,62,65,67,69,70,71,74,76,78,80,82,84,86$ and 88 or fragments comprising at least $50,75,100,150,200$ or 300 consecutive amino acids thereof. The term "variant" includes derivatives or analogs of these polypeptides. In particular, the variants may differ in amino acid sequence from the polypeptides of SEQ ID NOS: $2,4,6,8,10,12,14,16,18$, $20,22,24,26,28,30,32,34,36,38,41,42,44,46,48,50,52$, $54,56,58,60,62,65,67,69,70,71,74,76,78,80,82,84,86$ and 88 by one or more substitutions, additions, deletions, fusions and truncations, which may be present in any combination.
[0121] The variants may be naturally occurring or created in vitro. In particular, such variants may be created using genetic engineering techniques such as site directed mutagenesis, random chemical mutagenesis, exonuclease III deletion procedures, and standard cloning techniques. Alternatively, such variants, fragments, analogs, or derivatives may be created using chemical synthesis or modification procedures.
[0122] Other methods of making variants are also familiar to those skilled in the art. These include procedures in which nucleic acid sequences obtained from natural isolates are modified to generate nucleic acids that encode polypeptides having characteristics which enhance their value in industrial or laboratory applications. In such procedures, a large number of variant sequences having one or more nucleotide differences with respect to the sequence obtained from the natural isolate are generated and characterized. Preferably, these nucleotide differences result in amino acid changes with respect to the polypeptides encoded by the nucleic acids from the natural isolates.
[0123] The variants of the polypeptides of SEQ ID NOS: 2, $4,6,8,10,12,14,16,18,20,22,24,26,28,30,32,34,36,38$, $41,42,44,46,48,50,52,54,56,58,60,62,65,67,69,70,71$, $74,76,78,80,82,84,86$ and 88 may be variants in which one or more of the amino acid residues of the polypeptides of SEQ ID NOS: $2,4,6,8,10,12,14,16,18,20,22,24,26,28,30$, $32,34,36,38,41,42,44,46,48,50,52,54,56,58,60,62,65$, $67,69,70,71,74,76,78,80,82,84,86$ and 88 are substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue) and such substituted amino acid residue may or may not be one encoded by the genetic code.
[0124] Conservative substitutions are those that substitute a given amino acid in a polypeptide by another amino acid of like characteristics. Typically seen as conservative substitutions are the following replacements: replacements of an aliphatic amino acid such as Ala, Val, Leu and Ile with another aliphatic amino acid; replacement of a Ser with a Thr or vice versa; replacement of an acidic residue such as Asp or Glu with another acidic residue; replacement of a residue bearing an amide group, such as Asn or Gln, with another residue bearing an amide group; exchange of a basic residue such as Lys or Arg with another basic residue; and replacement of an aromatic residue such as Phe or Tyr with another aromatic residue.
[0125] Other variants are those in which one or more of the amino acid residues of the polypeptides of SEQ ID NOS: 2,4 , $6,8,10,12,14,16,18,20,22,24,26,28,30,32,34,36,38$, $41,42,44,46,48,50,52,54,56,58,60,62,65,67,69,70,71$, $74,76,78,80,82,84,86$ and 88 include a substituent group. Still other variants are those in which the polypeptide is associated with another compound, such as a compound to increase the half-life of the polypeptide (for example, polyethylene glycol). Additional variants are those in which addi-
tional amino acids are fused to the polypeptide, such as leader sequence, a secretory sequence, a proprotein sequence or a sequence that facilitates purification, enrichment, or stabilization of the polypeptide.
[0126] In some embodiments, the fragments, derivatives and analogs retain the same biological function or activity as the polypeptides of SEQ ID NOS: $2,4,6,8,10,12,14,16,18$, $20,22,24,26,28,30,32,34,36,38,41,42,44,46,48,50,52$, $54,56,58,60,62,65,67,69,70,71,74,76,78,80,82,84,86$ and 88 . In other embodiments, the fragment, derivative or analogue includes a fused heterologous sequence that facilitates purification, enrichment, detection, stabilization or secretion of the polypeptide that can be enzymatically cleaved, in whole or in part, away from the fragment, derivative or analogue.
[0127] Another aspect of the present invention are polypeptides or fragments thereof which have at least $70 \%$, at least $80 \%$, at least $85 \%$, at least $90 \%$, or more than $95 \%$ identity to one of the polypeptides of SEQ ID NOS: $2,4,6,8,10,12,14$, $16,18,20,22,24,26,28,30,32,34,36,38,41,42,44,46,48$, $50,52,54,56,58,60,62,65,67,69,70,71,74,76,78,80,82$, 84,86 and 88 or a fragment comprising at least $50,75,100$, 150,200 or 300 consecutive amino acids thereof. It will be appreciated that amino acid "substantially identity" includes conservative substitutions such as those described above.
[0128] The polypeptides or fragments having homology to one of the polypeptides of SEQ ID NOS: $2,4,6,8,10,12,14$, $16,18,20,22,24,26,28,30,32,34,36,38,41,42,44,46,48$, $50,52,54,56,58,60,62,65,67,69,70,71,74,76,78,80,82$, 84,86 and 88 or a fragment comprising at least $50,75,100$, 150,200 or 300 consecutive amino acids thereof may be obtained by isolating the nucleic acids encoding them using the techniques described above.
[0129] Alternatively, the homologous polypeptides or fragments may be obtained through biochemical enrichment or purification procedures. The sequence of potentially homologous polypeptides or fragments may be determined by proteolytic digestion, gel electrophoresis and/or microsequencing. The sequence of the prospective homologous polypeptide or fragment can be compared to one of the polypeptides of SEQ ID NOS: $2,4,6,8,10,12,14,16,18,20$, $22,24,26,28,30,32,34,36,38,41,42,44,46,48,50,52,54$, $56,58,60,62,65,67,69,70,71,74,76,78,80,82,84,86$ and 88 or a fragment comprising at least $5,10,15,20,25,30,35$, $40,50,75,100$, or 150 consecutive amino acids thereof.
[0130] The polypeptides of SEQ ID NOS: 2, 4, 6, 8, 10, 12, $14,16,18,20,22,24,26,28,30,32,34,36,38,41,42,44,46$, $48,50,52,54,56,58,60,62,65,67,69,70,71,74,76,78,80$, $82,84,86$ and 88 or fragments, derivatives or analogs thereof comprising at least $40,50,75,100,150,200$ or 300 consecutive amino acids thereof invention may be used in a variety of applications. For example, the polypeptides or fragments, derivatives or analogs thereof may be used to catalyze biochemical reactions as described elsewhere in the specification.

## EXAMPLES

[0131] Unless otherwise indicated, all numbers expressing quantities of ingredients, properties such as molecular weight, reaction conditions, $\mathrm{IC}_{50}$ and so forth used in the specification and claims are to be understood as being modified in all instances by the term "about". Accordingly, unless indicated to the contrary, the numerical parameters set forth in the present specification and attached claims are approxima-
tions. At the very least, and not as an attempt to limit the application of the doctrine of equivalents to the scope of the claims, each numerical parameter should at least be construed in light of the number of significant figures and by applying ordinary rounding techniques. Notwithstanding that the numerical ranges and parameters setting forth the broad scope of the invention are approximations, the numerical values set in the examples, Tables and Figures are reported as precisely as possible. Any numerical values may inherently contain certain errors resulting from variations in experiments, testing measurements, statistical analyses and such.
[0132] Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, suitable methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

## Example 1

## Preparation of Production Culture

[0133] Unless otherwise noted, all reagents were purchased from Sigma Chemical Co. (St. Louis, Mo.), (Aldrich). Micromonospora spp. (deposit accession number IDAC 070303-01) was maintained on agar plates of ISP2 agar (Difco Laboratories, Detroit, Mich.). An inoculum for the production phase was prepared by transferring the surface growth of the Micromonospora spp. from the agar plates to $125-\mathrm{mL}$ flasks containing 25 mL of sterile medium comprised of 24 g potato dextrin, 3 g beef extract, 5 g Bactocasitone, 5 g glucose, 5 g yeast extract, and $4 \mathrm{~g} \mathrm{CaCO}_{3}$ made up to one liter with distilled water ( pH 7.0 ). The culture was incubated at about $28^{\circ} \mathrm{C}$. for approximately 60 hours on a rotary shaker set at 250 rpm . Following incubation, 10 mL of culture was transferred to a 2 L baffled flask containing 500 mL of sterile production medium containing $20 \mathrm{~g} / \mathrm{L}$ potato dextrin, $20 \mathrm{~g} / \mathrm{L}$ glycerol, $10 \mathrm{~g} / \mathrm{L}$ Fish meal, $5 \mathrm{~g} / \mathrm{L}$ Bactopeptone, $2 \mathrm{~g} / \mathrm{L} \mathrm{CaCO}_{3}$, and $2 \mathrm{~g} / \mathrm{L}_{\left(\mathrm{NH}_{4}\right)_{2} \mathrm{SO}_{4}, \mathrm{pH} 7.0 \text {. Fer- }}$ mentation broth was prepared by incubating the production culture at $28^{\circ} \mathrm{C}$. in a rotary shaker set at 250 rpm for one week.

## Example 2

## Isolation

[0134] 500 mL ethyl acetate was added to 500 mL of fermentation broth prepared as described in Example 1 above. The mixture was agitated for 30 minutes on an orbital shaker at 200 rpm to create an emulsion. The phases were separated by centrifugation and decantation. Between 4 and 5 g of anhydrous $\mathrm{MgSO}_{4}$ was added to the organic phase, which was then filtered and the solvents removed in vacuo.
[0135] An ethyl acetate extract from 2 L fermentation was mixed with HP-20 resin ( 100 mL ; Mitsubishi Casei Corp., Tokyo, Japan) in water ( 300 mL ). Ethyl acetate was removed in vacuo, the resin was filtered on a Büchner funnel and the filtrate was discarded. The adsorbed HP-20 resin was then
washed successively with $2 \times 125 \mathrm{~mL}$ of $50 \%$ acetonitrile in water, $2 \times 125 \mathrm{~mL}$ of $75 \%$ acetonitrile in water and $2 \times 125 \mathrm{~mL}$ of acetonitrile.
[0136] Fractions containing ECO-04601 were evaporated to dryness and 100 mg was digested in the 5 mL of the upper phase of a mixture prepared from chloroform, cyclohexane, methanol, and water in the ratios, by volume, of 5:2:10:5. The sample was subjected to centrifugal partition chromatography using a High Speed Countercurrent (HSCC) system (Kromaton Technologies, Angers, France) fitted with a 200 mL cartridge and prepacked with the upper phase of this two-phase system. The HSCC was run with the lower phase mobile and ECO-04601 was eluted at approximately one-half column volume. Fractions were collected and ECO-04601 was detected by TLC of aliquots of the fractions on commercial Kieselgel $60 \mathrm{~F}_{254}$ plates. Compound could be visualized by inspection of dried plates under UV light or by spraying the plates with a spray containing vanillin ( $0.75 \%$ ) and concentrated sulfuric acid ( $1.5 \%, \mathrm{v} / \mathrm{v}$ ) in ethanol and subsequently heating the plate. Fractions contained substantially pure ECO-04601, although highly colored. A buff-colored sample could be obtained by chromatography on HPLC as follows.
[0137] 6 mg of sample was dissolved in acetonitrile and injected onto a preparative HPLC column (XTerra ODS (10 $\mu \mathrm{m}), 19 \times 150 \mathrm{~mm}$, Waters Co., Milford, Mass.), with a 9 $\mathrm{mL} / \mathrm{min}$ flow rate and UV peak detection at 300 nm . The column was eluted with acetonitrile/buffer ( 20 mM of $\mathrm{NH}_{4} \mathrm{HCO}_{3}$ ) according to the following gradient shown in Table 3.

TABLE 3

| Time (min) | Water (\%) | Acetonitrile (\%) |
| :---: | :---: | :---: |
| 0 | 70 | 30 |
| 10 | 5 | 95 |
| 15 | 5 | 95 |
| 20 | 70 | 30 |

[0138] Fractions containing ECO-04061 eluted at approximately 11:0 min and were combined, concentrated and lyophilized to give a yield of 3.8 mg compound.

Example 3
Elucidation of the Structure of ECO-04601
[0139]

$\mathrm{C}_{28} \mathrm{H}_{34} \mathrm{~N}_{2} \mathrm{O}_{4}$
Mol. Wt.: 462.25
[0140] The structure of ECO-04601 above was derived from spectroscopic data, including mass, UV, and NMR spectroscopy. Mass was determined by electrospray mass spectrometry to be 462.6, UV max 230 nm with a shoulder at 290
nm . NMR data were collected dissolved in MeOH-d4 including proton, and multidimensional pulse sequences. Proton and carbon NMR data are detailed in Table 4 below.

TABLE 4

| ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR ( $\delta_{H}, \mathrm{ppm}$ ) of ECO-04601 in MeOH-D ${ }_{4}$ |  |  |  |
| :---: | :---: | :---: | :---: |
| Assignment | ${ }^{1} \mathrm{H}$ | ${ }^{13} \mathrm{C}$ | Group |
| 1 | 7.15 | 122.3 | CH |
| 2 | 6.74 | 121.0 | CH |
| 3 | 6.83 | 116.9 | CH |
| 4 | - | 146.0 | $\mathrm{C}-\mathrm{OH}$ |
| 4 a | - | 142.0 | C |
| 5 a | - | 126.0 | C |
| 6 | - | 148.2 | $\mathrm{C}-\mathrm{OH}$ |
| 7 | 6.20 | 100.0 | CH |
| 8 | - | 153.0 | $\mathrm{C}-\mathrm{OH}$ |
| 9 | 6.25 | 101.0 | CH |
| 9 a | - | 135.0 | C |
| 11 | - | 170.0 | C (O) |
| 11a | - | 125.0 | C |
| $1^{\prime}$ | 4.52 | 48.7 | $\mathrm{CH}_{2}$ |
| $2^{\prime}$ | 5.35 | 121.1 | CH |
| $3^{\prime}$ | - | 138.5 | C |
| $4^{\prime}$ | 2.03 | 39.5 | $\mathrm{CH}_{2}$ |
| $5^{\prime}$ | 2.08 | 26.7 | $\mathrm{CH}_{2}$ |
| $6^{\prime}$ | 5.09 | 124.1 | CH |
| 7 | - | 135.0 | C |
| 8 | 1.95 | 39.6 | $\mathrm{CH}_{2}$ |
| $9^{\prime}$ | 2.02 | 26.3 | $\mathrm{CH}_{2}$ |
| $10^{\prime}$ | 5.06 | 124.4 | CH |
| 11' | - | 130.9 | C |
| 12' | 1.64 | 24.8 | $\mathrm{CH}_{3}$ |
| 1 " | 1.72 | 15.5 | $\mathrm{CH}_{3}$ |
| $2^{\prime \prime}$ | 1.59 | 14.9 | $\mathrm{CH}_{3}$ |
| $3 "$ | 1.55 | 16.5 | $\mathrm{CH}_{3}$ |

[0141] A number of cross peaks in the 2D spectra of ECO04601 are key in the structural determination. For example, the farnesyl chain is placed on the amide nitrogen by a strong cross peak between the proton signal of the terminal methylene of that chain at 4.52 ppm and the amide carbonyl carbon at 170 ppm in the gHMBC experiment. This conclusion is confirmed by a cross peak in the NOESY spectrum between the same methylene signals at 4.52 ppm and the aromatic proton signal at 6.25 ppm from one of the two protons of the tetra substituted benzenoid ring.
[0142] Based on the mass, UV and NMR spectroscopy data, the structure of the compound was determined to be the structure of ECO-04601.

## Example 4

## In Vivo Efficacy in a Glioma Model

[0143] The aim of this study was to test whether ECO04601 when administered by the i.p. route prevents or delays tumor growth in C6 glioblastoma cell-bearing mice, and to determine an effective dosage regimen.
[0144] Animals: A total of 60 six-week-old female mice (Mus musculus nude mice), ranging between 18 to 25 g in weight, were observed for 7 days before treatment. Animal experiments were performed according to ethical guidelines of animal experimentation (Charté du comite d'éthique du CNRS, juillet 2003) and the English guidelines for the welfare of animals in experimental neoplasia (WORKMAN, P., TWENTYMAN, P., BALKWILL, F., et al. (1998). United Kingdom Coordinating Committee on Cancer Research (UKCCCR) Guidelines for the welfare of animals in experi-
mental neoplasia (Second Edition, July 1997; British Journal of Cancer, 77:1-10). Any dead or apparently sick mice were promptly removed and replaced with healthy mice. Sick mice were euthanized upon removal from the cage. Animals were maintained in rooms under controlled conditions of temperature $\left(23+2^{\circ} \mathrm{C}\right.$.), humidity ( $45 \pm 5 \%$ ), photoperiodicity ( 12 hrs light/ 12 hrs dark) and air exchange. Animals were housed in polycarbonate cages ( $5 /$ single cage) that were equipped to provide food and water. Animal bedding consisted of sterile wood shavings that were replaced every other day. Food was provided ad libitum, being placed in the metal lid on the top of the cage. Autoclaved tap water was provided ad libitum. Water bottles were equipped with rubber stoppers and sipper tubes. Water bottles were cleaned, sterilized and replaced once a week. Two different numbers engraved on two earrings identified the animals. Each cage was labeled with a specific code.
[0145] Tumor Cell Line: The C6 cell line was cloned from a rat glial tumor induced by N-nitrosomethyurea (NMU) according to Premont et al. (Premont J, Benda P, Jard S., [3H] norepinephrine binding by rat glial cells in culture. Lack of correlation between binding and adenylate cyclase activation. Biochim Biophys Acta. 1975 Feb. 13; 381(2):368-76.) after series of alternate culture and animal passages. Cells were grown as adherent monolayers at $37^{\circ} \mathrm{C}$. in a humidified atmosphere ( $5 \% \mathrm{CO}_{2}, 95 \%$ air). The culture medium was DMEM supplemented with 2 mML -glutamine and $10 \%$ fetal bovine serum. For experimental use, tumor cells were detached from the culture flask by a 10 min treatment with trypsin-versen. The cells were counted in a hemocytometer and their viability assessed by $0.25 \%$ trypan blue exclusion.
[0146] Preparation of the Test Article: for the Test Article, the Following Procedure was followed for reconstitution (performed immediately preceding injection). The vehicle consisted of a mixture of benzyl alcohol ( $1.5 \%$ ), ethanol ( $8.5 \%$ ), propylene glycol ( $27 \%$ ), PEG 400 ( $27 \%$ ), dimethylacetamide ( $6 \%$ ) and water ( $30 \%$ ). The vehicle solution was first vortexed in order to obtain a homogeneous liquid. 0.6 mL of the vortexed vehicle solution was added to each vial containing the test article (ECO-04601). Vials were mixed thoroughly by vortexing for 1 minute and inverted and shaken vigorously. Vials were mixed again prior to injection into each animal.
[0147] Animal Inoculation with tumor cells: Experiment started at day $0\left(D_{0}\right)$. On $D_{0}$, mice received a superficial intramuscular injection of C 6 tumor cells ( $5 \times 10^{5}$ cells) in 0.1 mL of DMEM complete medium into the upper right posterior leg.
[0148] Treatment Regimen and Results:
[0149] First series of experiments: In a first series of experiments, treatment started 24 hrs following inoculation of C6 cells. On the day of the treatment, each mouse was slowly injected with $100 \mu \mathrm{~L}$ of test or control articles by the i.p. route. For all groups, treatment was performed until the tumor volume of the saline-treated mice (group 1) reached approximately $3 \mathrm{~cm}^{3}$ (around day 16). Mice of group 1 were treated daily with a saline isosmotic solution for 16 days. Mice of group 2 were treated daily with the vehicle solution for 16 days. Mice of group 3 were treated daily with $10 \mathrm{mg} / \mathrm{kg}$ of ECO-04601 for 16 days. Mice of group 4 were treated every two days with $30 \mathrm{mg} / \mathrm{kg}$ of ECO-04601 and received 8 treatments. Mice of group 5 were treated every three days with 30 $\mathrm{mg} / \mathrm{kg}$ of ECO-04601 and received 6 treatments. Measurement of tumor volume started as soon as tumors became
palpable ( $>100 \mathrm{~mm}^{3}$; day 11 post-inoculation) and was evaluated every second day until the end of the treatment using callipers. As shown in Table 5 and FIG. 1, the mean value of the tumor volume of all ECO-4061-treated groups ( 6 mice/ group) was significantly reduced as demonstrated by the oneway analysis of variance (Anova) test followed by the nonparametric Dunnett's multiple comparison test comparing treated groups to the saline group. An asterisk in the P value column of Table 5 indicates a statistically significant value, while "ns" signifies not significant.

TABLE 5

| Treatment | Treatment regimen | $\begin{aligned} & \text { Tumor volume } \\ & \left(\mathrm{mm}^{3}\right) \\ & (\text { mean } \pm \text { SEM }) \end{aligned}$ | \% Inhibition | P <br> value |
| :---: | :---: | :---: | :---: | :---: |
| Saline | Q1 $\times 16$ | $3,004.1 \pm 249.64$ | - | - |
| Vehicle solution | Q1 $\times 16$ | $2,162.0 \pm 350.0$ | 28.0\% | $>0.05 \mathrm{~ns}$ |
| ECO-04601 <br> ( $10 \mathrm{mg} / \mathrm{kg}$ ) | Q1 $\times 16$ | $1,220.4 \pm 283.46$ | 59.4\% | $<0.01$ * |
| $\begin{aligned} & \mathrm{ECO}-04601 \\ & (30 \mathrm{mg} / \mathrm{kg}) \end{aligned}$ | Q2 $\times 8$ | $1,236.9 \pm 233.99$ | 58.8\% | $<0.01 *$ |
| ECO-04601 <br> ( $30 \mathrm{mg} / \mathrm{kg}$ ) | Q $3 \times 6$ | $1,184.1 \pm 221.45$ | 60.6\% | $<0.01 *$ |

[0150] Second series experiments: In a second series of experiments, treatment started at day 10 following inoculation of C6 cells when tumors became palpable (around 100 to $200 \mathrm{~mm}^{3}$ ). Treatment was repeated daily for 5 consecutive days. On the day of the treatment, each mouse was slowly injected with $100 \mu \mathrm{~L}$ of ECO- 04601 by i.p. route. Mice of group 1 were treated daily with saline isosmotic solution. Mice of group 2 were treated daily with the vehicle solution. Mice of group 3 were treated daily with $20 \mathrm{mg} / \mathrm{kg}$ of ECO04601 . Mice of group 4 were treated daily with $30 \mathrm{mg} / \mathrm{kg}$ of ECO-04601. Mice were treated until the tumor volume of the saline-treated control mice (group 1) reached around $4 \mathrm{~cm}^{3}$. Tumor volume was measured every second day until the end of the treatment using callipers. As shown in Table 6 and FIG. 2, the mean value of the tumor volume of all ECO-04601treated groups ( 6 mice/group) was significantly reduced as demonstrated by the one-way analysis of variance (Anova) test followed by the non-parametric Dunnett's multiple comparison test comparing treated groups to the saline group. An asterisk in the P value column of Table 6 indicates a statistically significant value, while "ns" signifies not significant.
[0151] Histological analysis of tumor sections showed pronounced morphological changes between tumors from ECO04601 -treated mice and those from mice in the control groups. In tumors from mice treated with ECO-04601 (20-30 $\mathrm{mg} / \mathrm{kg}$ ), cell density was decreased and the nuclei of remaining tumor cells appeared larger and pycnotic while no such changes were observed for tumors from vehicle-treated mice (FIG. 3).

TABLE 6

| Treatment | Treatment regimen | $\begin{aligned} & \text { Tumor volume } \\ & \left(\mathrm{mm}^{3}\right) \\ & (\text { mean } \pm \text { SEM }) \end{aligned}$ | \% Inhibition | P <br> value |
| :---: | :---: | :---: | :---: | :---: |
| Saline | Q1 $\times 5$ | $4,363.1 \pm 614.31$ | - | - |
| Vehicle solution | Q1 $\times 5$ | $3,205.0 \pm 632.37$ | 26.5\% | $>0.05 \mathrm{~ns}$ |

TABLE 6-continued

| Treatment | Treatment regimen | $\begin{aligned} & \text { Tumor volume } \\ & \left(\mathrm{mm}^{3}\right) \\ & (\text { mean } \pm \text { SEM }) \end{aligned}$ | \% Inhibition | P <br> value |
| :---: | :---: | :---: | :---: | :---: |
| ECO-04601 <br> ( $20 \mathrm{mg} / \mathrm{kg}$ ) | Q1 $\times 5$ | $1,721.5 \pm 374.79$ | 60.5\% | $<0.01$ * |
| $\begin{aligned} & \mathrm{ECO}-04601 \\ & (30 \mathrm{mg} / \mathrm{kg}) \end{aligned}$ | Q1 $\times 5$ | $1,131.6 \pm 525.21$ | 74.1\% | $<0.01$ * |

## Example 5

Genes and Proteins for the Production of Farnesyl Dibenzodiazepinones
[0152] Micromonospora sp. strain 046-ECO11 is a representative microorganism useful in the production of the compound of the invention. Strain $046-\mathrm{ECO} 11$ has been deposited with the International Depositary Authority of Canada (IDAC), Bureau of Microbiology, Health Canada, 1015 Arlington Street, Winnipeg, Manitoba, Canada R3E 3R2 on Mar. 7, 2003 and was assigned IDAC accession no. 07030301 . The biosynthetic locus for the production of ECO-04601 was identified in the genome of Micromonospora sp. strain 046-ECO11 using the genome scanning method described in U.S. Ser. No. 10/232,370, CA 2, 352, 451 and Zazopoulos et. al., Nature Biotechnol., 21, 187-190 (2003).
[0153] The biosynthetic locus spans approximately 52,400 base pairs of DNA and encodes 43 proteins. More than 10 kilobases of DNA sequence were analyzed on each side of the locus and these regions were deemed to contain primary genes or genes unrelated to the synthesis of ECO-04601. As illustrated in FIG. 4, the locus is contained within three sequences of contiguous base pairs, namely Contig 1 having the 36,602 contiguous base pairs of SEQ ID NO: 1 and comprising ORFs 1 to 31 (SEQ ID NOS: 3, 5, 7, 9, 11, 13, 15, $17,19,21,23,25,27,29,31,33,35,37,39,41,43,45,47,49$, $51,53,55,57,59,61$ and 63 ), Contig 2 having the 5,960 contiguous base pairs of SEQ ID NO: 64 and comprising ORFs 32 to 35 (SEQ ID NOS: 66, 68, 70 and 72), and Contig 3 having the 9,762 base pairs of SEQ ID NO: 73 and comprising ORFs 36 to 43 (SEQ ID NOS: 75, 77, 79, 81, 83, 85, 87 and 89). The order, relative position and orientation of the 43 open reading frames representing the proteins of the biosynthetic locus are illustrated schematically in FIG. 4. The top line in FIG. 4 provides a scale in base pairs. The gray bars depict the three DNA contigs (SEQ ID NOS: 1, 64 and 73) that cover the locus. The empty arrows represent the 43 open reading frames of this biosynthetic locus. The black arrows represent the two deposited cosmid clones covering the locus.
[0154] The biosynthetic locus will be further understood with reference to the sequence listing which provides contiguous nucleotide sequences and deduced amino acid sequences of the locus from Micromonospora sp. strain 046ECO11. The contiguous nucleotide sequences are arranged such that, as found within the biosynthetic locus, Contig 1 (SEQ ID NO: 1) is adjacent to the $5^{\prime}$ end of Contig 2 (SEQ ID NO: 64), which in turn is adjacent to Contig 3 (SEQ ID NO: 73). The ORFs illustrated in FIG. 4 and provided in the sequence listing represent open reading frames deduced from
the nucleotide sequences of Contigs 1, 2 and 3 (SEQ ID NOS: 1,64 and 73). Referring to the Sequence Listing, ORF 1 (SEQ ID NO: 3) is the polynucleotide drawn from residues 2139 to 424 of SEQ ID NO: 1, and SEQ ID NO: 2 represents that polypeptide deduced from SEQ ID NO: 3. ORF 2 (SEQ ID NO: 5) is the polynucleotide drawn from residues 2890 to 4959 of SEQ ID NO: 1, and SEQ ID NO: 4 represents the polypeptide deduced from SEQ ID NO: 5. ORF 3 (SEQ ID NO: 7) is the polynucleotide drawn from residues 7701 to 5014 of SEQ ID NO: 1, and SEQ ID NO: 6 represents the polypeptide deduced from SEQ ID NO: 7. ORF 4 (SEQ ID NO: 9) is the polynucleotide drawn from residues 8104 to 9192 of SEQ ID NO: 1, and SEQ ID NO: 8 represents the polypeptide deduced from SEQ ID NO: 9. ORF 5 (SEQ ID NO: 11) is the polynucleotide drawn from residues 9192 to 10256 of SEQ ID NO: 1, and SEQ ID NO: 10 represents the polypeptide deduced from SEQ ID NO: 11. ORF 6 (SEQ ID NO: 13) is the polynucleotide drawn from residues 10246 to 11286 of SEQ ID NO: 1, and SEQ ID NO: 12 represents the polypeptide deduced from SEQ ID NO: 13. ORF 7 (SEQ ID NO: 15) is the polynucleotide drawn from residues 11283 to 12392 of SEQ ID NO: 1, and SEQ ID NO: 14 represents the polypeptide deduced from SEQ ID NO: 15. ORF 8 (SEQ ID NO: 17) is the polynucleotide drawn from residues 12389 to 13471 of SEQ ID NO: 1, and SEQ ID NO: 16 represents the polypeptide deduced from SEQ ID NO: 17. ORF 9 (SEQ ID NO: 19) is the polynucleotide drawn from residues 13468 to 14523 of SEQ ID NO: 1, and SEQ ID NO: 18 represents the polypeptide deduced from SEQ ID NO: 19. ORF 10 (SEQ ID NO: 21) is the polynucleotide drawn from residues 14526 to 15701 of SEQ ID NO: 1, and SEQ ID NO: 20 represents the polypeptide deduced from SEQ ID NO: 21. ORF 11 (SEQ ID NO: 23) is the polynucleotide drawn from residues 15770 to 16642 of SEQ ID NO: 1, and SEQ ID NO: 22 represents the polypeptide deduced from SEQ ID NO: 23. ORF 12 (SEQ ID NO: 25) is the polynucleotide drawn from residues 16756 to 17868 of SEQ ID NO: 1, and SEQ ID NO: 24 represents the polypeptide deduced from SEQ ID NO: 25. ORF 13 (SEQ ID NO: 27) is the polynucleotide drawn from residues 17865 to 18527 of SEQ ID NO: 1, and SEQ ID NO: 26 represents the polypeptide deduced from SEQ ID NO: 27. ORF 14 (SEQ ID NO: 29) is the polynucleotide drawn from residues 18724 to 19119 of SEQ ID NO: 1, and SEQ ID NO: 28 represents the polypeptide deduced from SEQ ID NO: 29. ORF 15 (SEQ ID NO: 31) is the polynucleotide drawn from residues 19175 to 19639 of SEQ ID NO: 1, and SEQ ID NO: 30 represents the polypeptide deduced from SEQ ID NO: 31. ORF 16 (SEQ ID NO: 33) is the polynucleotide drawn from residues 19636 to 21621 of SEQ ID NO: 1, and SEQ ID NO: 32 represents the polypeptide deduced from SEQ ID NO: 33. ORF 17 (SEQ ID NO: 35) is the polynucleotide drawn from residues 21632 to 22021 of SEQ ID NO: 1, and SEQ ID NO: 34 represents the polypeptide deduced from SEQ ID NO: 35. ORF 18 (SEQ ID NO: 37 ) is the polynucleotide drawn from residues 22658 to 22122 of SEQ ID NO: 1, and SEQ ID NO: 36 represents the polypeptide deduced from SEQ ID NO: 37. ORF 19 (SEQ ID NO: 39 ) is the polynucleotide drawn from residues 24665 to 22680 of SEQ ID NO: 1, and SEQ ID NO: 38 represents the polypeptide deduced from SEQ ID NO: 39. ORF 20 (SEQ ID $\mathrm{NO}: 41$ ) is the polynucleotide drawn from residues 24880 to 26163 of SEQ ID NO: 1, and SEQ ID NO: 40 represents the polypeptide deduced from SEQ ID NO: 41. ORF 21 (SEQ ID $\mathrm{NO}: 43$ ) is the polynucleotide drawn from residues 26179 to 27003 of SEQ ID NO: 1, and SEQ ID NO: 42 represents the
polypeptide deduced from SEQ ID NO: 43. ORF 22 (SEQ ID NO: 45) is the polynucleotide drawn from residues 27035 to 28138 of SEQ ID NO: 1, and SEQ ID NO: 44 represents the polypeptide deduced from SEQ ID NO: 45 . ORF 23 (SEQ ID NO: 47) is the polynucleotide drawn from residues 28164 to 28925 of SEQ ID NO: 1, and SEQ ID NO: 46 represents the polypeptide deduced from SEQ ID NO: 47. ORF 24 (SEQ ID) NO: 49) is the polynucleotide drawn from residues 28922 to 30238 of SEQ ID NO: 1, and SEQ ID NO: 48 represents the polypeptide deduced from SEQ ID NO: 49. ORF 25 (SEQ ID NO: 51) is the polynucleotide drawn from residues 30249 to 31439 of SEQ ID NO: 1, and SEQ ID NO: 50 represents the polypeptide deduced from SEQ ID NO: 51. ORF 26 (SEQ ID NO: 53) is the polynucleotide drawn from residues 31439 to 32224 of SEQ ID NO: 1, and SEQ ID NO: 52 represents the polypeptide deduced from SEQ ID NO: 53. ORF 27 (SEQ ID NO: 55) is the polynucleotide drawn from residues 32257 to 32931 of SEQ ID NO: 1, and SEQ ID NO: 54 represents the polypeptide deduced from SEQ ID NO: 55. ORF 28 (SEQ ID NO: 57) is the polynucleotide drawn from residues 32943 to 33644 of SEQ ID NO: 1, and SEQ ID NO: 56 represents the polypeptide deduced from SEQ ID NO: 57. ORF 29 (SEQ ID NO: 59) is the polynucleotide drawn from residues 34377 to 33637 of SEQ ID NO: 1, and SEQ ID NO: 58 represents the polypeptide deduced from SEQ ID NO: 59. ORF 30 (SEQ ID NO: 61) is the polynucleotide drawn from residues 34572 to 34907 of SEQ ID NO: 1, and SEQ ID NO: 60 represents the polypeptide deduced from SEQ ID NO: 61. ORF 31 (SEQ ID NO: 63) is the polynucleotide drawn from residues 34904 to 36583 of SEQ ID NO: 1, and SEQ ID NO: 62 represents the polypeptide deduced from SEQ ID NO: 63. ORF 32 (SEQ ID $\mathrm{NO}: 66$ ) is the polynucleotide drawn from residues 23 to 1621 of SEQ ID NO: 64, and SEQ ID NO: 65 represents the polypeptide deduced from SEQ ID NO: 66. ORF 33 (SEQ ID NO: 68) is the polynucleotide drawn from residues 1702 to 2973 of SEQ ID NO: 64, and SEQ ID NO: 67 represents the polypeptide deduced from SEQ ID NO: 68. ORF 34 (SEQ ID NO: 70) is the polynucleotide drawn from residues 3248 to 4270 of SEQ ID NO: 64, and SEQ ID NO: 69 represents the polypeptide deduced from SEQ ID NO: 70. ORF 35 (SEQ ID NO: 72) is the polynucleotide drawn from residues 4452 to 5933 of SEQ ID NO: 64, and SEQ ID NO: 71 represents the polypeptide deduced from SEQ ID NO: 72 . ORF 36 (SEQ ID NO: 75) is the polynucleotide drawn from residues 30 to 398 of SEQ ID NO: 73, and SEQ ID NO: 74 represents the polypeptide deduced from SEQ ID NO: 75. ORF 37 (SEQ ID NO: 77) is the polynucleotide drawn from residues 395 to 1372 of SEQ ID NO: 73, and SEQ ID NO: 76 represents the polypeptide deduced from SEQ ID NO: 77. ORF 38 (SEQ ID NO: 79) is the polynucleotide drawn from residues 3388 to 1397 of SEQ ID NO: 73, and SEQ ID NO: 78 represents the polypeptide deduced from SEQ ID NO: 79. ORF 39 (SEQ ID NO: 81) is the polynucleotide drawn from residues 3565 to 5286 of SEQ ID NO: 73, and SEQ ID NO: 80 represents the polypeptide deduced from SEQ ID NO: 81. ORF 40 (SEQ ID NO: 83) is the polynucleotide drawn from residues 5283 to 7073 of SEQ ID NO: 73, and SEQ ID NO: 82 represents the polypeptide deduced from SEQ ID NO: 83. ORF 41 (SEQ ID NO: 85) is the polynucleotide drawn from residues 7108 to 8631 of SEQ ID NO: 73, and SEQ ID NO: 84 represents the polypeptide deduced from SEQ ID NO: 85. ORF 42 (SEQ ID NO: 87) is the polynucleotide drawn from residues 9371 to 8673 of SEQ ID NO: 73, and SEQ ID NO: 86 represents the polypeptide deduced from SEQ ID NO: 87. ORF 43 (SEQ ID

NO: 89) is the polynucleotide drawn from residues 9762 to 9364 of SEQ ID NO: 73, and SEQ ID NO: 88 represents the polypeptide deduced from SEQ ID NO: 89.
[0155] Some open reading frames provided in the Sequence Listing, namely ORF 2 (SEQ ID NO: 5), ORF 5 (SEQ ID NO: 11), ORF 12 (SEQ ID NO: 25), ORF 13 (SEQ ID NO: 27), ORF 15 (SEQ ID NO:31), ORF 17 (SEQ ID NO: 35), ORF 19 (SEQ ID NO: 39), ORF 20 (SEQ ID NO: 41), ORF 22 (SEQ ID NO: 45), ORF 24 (SEQ ID NO: 49), ORF 26 (SEQ ID NO: 53) and ORF 27 (SEQ ID NO: 55) initiate with non-standard initiation codons (eg. GTG-Valine, or CTGLeucine) rather than standard initiation codon ATG methionine. All ORFs are listed with the appropriate $\mathrm{M}, \mathrm{V}$ or L amino acids at the amino-terminal position to indicate the specificity of the first codon of the ORF. It is expected, however, that in all cases the biosynthesized protein will contain a methionine residue, and more specifically a formylmethionine residue, at the amino terminal position, in keeping with the widely accepted principle that protein synthesis in bacteria initiate with methionine (formylmethionine) even when the encoding gene specifies a non-standard initiation codon (e.g. Stryer BioChemistry $3^{\text {rd }}$ edition, 1998, W.H. Freeman and Co., New York, pp. 752-754).
[0156] ORF 32 (SEQ ID NO: 65) is incomplete and contains a truncation of 10 to 20 amino acids from its carboxy terminus. This is due to incomplete sequence information between Contigs 2 and 3 (SEQ ID NOS: 64 and 73, respectively).
[0157] Deposits of E. coli DH10B vectors, each harbouring a cosmid clone (designated in FIG. 4 as 046 KM and 046 KQ respectively) of a partial biosynthetic locus for the farnesyl dibenzodiazepinone from Micromonospora sp. strain 046ECO11 and together spanning the full biosynthetic locus for production of ECO-04601 have been deposited with the International Depositary Authority of Canada, Bureau of Microbiology, Health Canada, 1015 Arlington Street, Winnipeg, Manitoba, Canada R3E 3R2 on Feb. 25, 2003. The cosmid clone designated 046 KM was assigned deposit accession numbers IDAC 250203-06, and the cosmid clone designated 046KQ was assigned deposit accession numbers IDAC 250203-07. Cosmid 046KM covers residue 1 to residue

32,250 of Contig 1 (SEQ ID NO: 1). Cosmid 046KQ covers residue 21,700 of Contig 1 (SEQ ID NO: 1) to residue 9,762 of Contig 3 (SEQ ID NO: 73). The sequence of the polynucleotides comprised in the deposited strains, as well as the amino acid sequence of any polypeptide encoded thereby are controlling in the event of any conflict with any description of sequences herein.
[0158] The deposit of the deposited strains has been made under the terms of the Budapest Treaty on the International Recognition of the Deposit of Micro-organisms for Purposes of Patent Procedure. The deposited strains will be irrevocably and without restriction or condition released to the public upon the issuance of a patent. The deposited strains are provided merely as convenience to those skilled in the art and are not an admission that a deposit is required for enablement, such as that required under 35 U.S.C. §112. A license may be required to make, use or sell the deposited strains, and compounds derived therefrom, and no such license is hereby granted.
[0159] In order to identify the function of the proteins coded by the genes forming the biosynthetic locus for the production of ECO-04601 the gene products of ORFs 1 to 43, namely SEQ ID NOS: $2,4,6,8,10,12,14,16,18,20,22,24$, $26,28,30,32,34,36,38,40,42,44,46,48,50,52,54,56,58$, $60,62,65,67,69,71,74,76,78,80,82,84,86$ and 88 were compared, using the BLASTP version 2.2.10 algorithm with the default parameters, to sequences in the National Center for Biotechnology Information (NCBI) nonredundant protein database and the DECIPHER® database of microbial genes, pathways and natural products (Ecopia BioSciences Inc. St.Laurent, QC, Canada).
[0160] The accession numbers of the top GenBank ${ }^{\text {TM }}$ hits of this BLAST analysis are presented in Table 7 along with the corresponding E values. The E value relates the expected number of chance alignments with an alignment score at least equal to the observed alignment score. An E value of 0.00 indicates a perfect homolog. The E values are calculated as described in Altschul et al. J. Mol. Biol., 215, 403-410 (1990). The E value assists in the determination of whether two sequences display sufficient similarity to justify an inference of homology.

TABLE 7


TABLE 7-continued

| ORF | $\begin{gathered} \text { SEQ } \\ \text { ID } \end{gathered}$ | Family | \# aa | GenBank <br> homology | Sequence comparison and ORF correlation |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  | Probability | $\begin{gathered} \text { \% Identity } \\ (\% \text { Similarity) } \end{gathered}$ | Proposed function of GenBank match |
| 4 | 8 | IDSA | 362 | $\begin{aligned} & \text { NP_601376.2 } \\ & 371 \mathrm{aa} \end{aligned}$ | 2E-80 | 49\% (65\%) | GGPP synthase [Corynebacterium glutamicum |
|  |  |  |  | $\begin{aligned} & \text { NP_738677.1 } \\ & 366 \mathrm{aa} \end{aligned}$ | 3E-79 | 48\% (62\%) | polyprenyl synthase, Corynebacterium efficiens |
|  |  |  |  | $\begin{aligned} & \text { NP_216689.1 } \\ & 352 \mathrm{aa} \end{aligned}$ | 2E-78 | 46\% (61\%) | idsA2 [Mycobacterium tuberculosis H37Rv] |
| 5 | 10 | MVKA | 354 | $\begin{aligned} & \text { BAB07790.1 } \\ & \text { 345aa } \end{aligned}$ | 2E-71 | 46\% (59\%) | mevalonate kinase [Streptomyces sp. CL190] |
|  |  |  |  | $\begin{aligned} & \text { BAB07817.1 } \\ & 334 \mathrm{aa} \end{aligned}$ | 5E-66 | 45\% (57\%) | mevalonate kinase [Kitasatospora griseola] |
|  |  |  |  | $\mathrm{NP}_{2} 720650.1$ | 3E-36 | 29\% (48\%) | mevalonate kinase [Streptococcus mutans |
| 6 | 12 | DMDA | 346 | $\begin{aligned} & \text { BAB07791.1 } \\ & 350 \mathrm{aa} \end{aligned}$ | 2E-88 | 58\% (65\%) | diphosphomevalonate decarboxylase [Streptomyces sp. |
|  |  |  |  | BAB07818.1 | 2E-69 | 53\% (61\%) | mevalonate diPH decaroboxylase |
|  |  |  |  | 300aa |  |  | [Kitasatospora griseola] |
|  |  |  |  | $\begin{aligned} & \text { NP_785307.1 } \\ & 325 \mathrm{aa} \end{aligned}$ | 3E-44 | 34\% (46\%) | diphosphomevalonate decarboxylase [Lactobacillus plantarum |
| 7 | 14 | MVKP | 369 | $\begin{aligned} & \text { BAB07792.1 } \\ & \text { 374aa } \end{aligned}$ | 4E-93 | 50\% (60\%) | phosphomevalonate kinase [Streptomyces <br> sp. CL190] |
|  |  |  |  | $\begin{aligned} & \text { BAB07819.1 } \\ & 360 \mathrm{aa} \end{aligned}$ | $6 \mathrm{E}-77$ | 48\% (56\%) | phosphomevalonate kinase [Kitasatospora griseola] |
|  |  |  |  | $\begin{aligned} & \text { AAG02442.1 } \\ & 368 \mathrm{aa} \end{aligned}$ | 2E-31 | 29\% (42\%) | 3 phosphomevalonate kinase [Enterococcus faecalis] |
| 8 | 16 | IPPI | 360 | $\begin{aligned} & \text { Q9KWF6 } \\ & \text { 364aa } \end{aligned}$ | 1E-128 | 66\% (74\%) | Isopentenyl-diphosphate delta-isomerase |
|  |  |  |  | $\begin{aligned} & \text { Q9KWG2 } \\ & 363 \mathrm{aa} \end{aligned}$ | 1E-128 | 66\% (77\%) | Isopentenyl-diphosphate delta-isomerase |
|  |  |  |  | $\begin{aligned} & \text { NP_814639.1 } \\ & 347 \mathrm{aa} \end{aligned}$ | 5E-73 | 44\% (61\%) | isopentenyl diphosphate isomerase [Enterococcus faecalis |
| 9 | 18 | HMGA | 351 | $\begin{aligned} & \text { BAA70975.1 } \\ & 353 \mathrm{aa} \end{aligned}$ | 1E-165 | 82\% (91\%) | 3-hydroxy-3-methylglutaryl coenzyme A reductase [Streptomyces sp .] |
|  |  |  |  | $\begin{aligned} & \text { BAA74565.1 } \\ & 353 \mathrm{aa} \end{aligned}$ | 1E-160 | 81\% (89\%) | 3-hydroxy-3-methylglutaryl coenzyme A reductase [Kitasatospora griseola] |
|  |  |  |  | $\begin{aligned} & \text { BAA74566.1 } \\ & 353 \mathrm{aa} \end{aligned}$ | 1E-155 | 80\% (86\%) | 3-hydroxy-3-methylglutaryl coenzyme A reductase [Streptomyces sp .] |
| 10 | 20 | KASH | 391 | $\begin{aligned} & \text { BAB07795.1 } \\ & \text { 389aa } \end{aligned}$ | 1E-148 | 67\% (78\%) | 3-hydroxy-3-methylglutaryl CoA synthase [Streptomyces sp. CL190] |
|  |  |  |  | $\begin{aligned} & \text { BAB07822.1 } \\ & 346 \mathrm{aa} \end{aligned}$ | 1E-136 | 70\% (78\%) | HMG-CoA synthase [Kitasatospora griseola] |
|  |  |  |  | $\begin{aligned} & \text { CAD24420.1 } \\ & \text { 388aa } \end{aligned}$ | $6 \mathrm{E}-79$ | 43\% (54\%) | HMG-CoA synthase [Paracoccus zeaxanthinifaciens] |
| 11 | 22 | IPTN | 290 | $\begin{aligned} & \text { NP_631248.1 } \\ & 295 \text { aa } \end{aligned}$ | 5E-22 | 28\% (44\%) | hypothetical protein [Streptomyces coelicolor A3(2)] |
|  |  |  |  | $\begin{aligned} & \text { AAN65239.1 } \\ & \text { 324aa } \end{aligned}$ | 5E-06 | 25\% (40\%) | cloQ [Streptomyces roseochromogenes subsp. oscitans] |
| 12 | 24 | SPKG | 370 | $\begin{aligned} & \text { AAM78435.1 } \\ & \text { 344aa } \end{aligned}$ | 5E-48 | 54\% (63\%) | two-component sensor [Streptomyces coelicolor A3(2)] |
|  |  |  |  | $\begin{aligned} & \text { NP_630507.1 } \\ & \text { 382aa } \end{aligned}$ | 5E-48 | 54\% (63\%) | sensor kinase [Streptomyces coelicolor $\mathrm{A} 3(2)]$ |
|  |  |  |  | $\begin{aligned} & \text { ZP_00058991.1 } \\ & 407 \mathrm{aa} \end{aligned}$ | 9E-34 | 44\% (58\%) | Signal transduction histidine kinase [Thermobifida fusca] |
| 13 | 26 | RREB | 220 | $\begin{aligned} & \text { NP_630508.1 } \\ & \text { 224aa } \end{aligned}$ | 3E-79 | 67\% (81\%) | regulatory protein [Streptomyces coelicolor $\mathrm{A} 3(2)]$ |
|  |  |  |  | $\begin{aligned} & \text { ZP_00058992.1 } \\ & \text { 221aa } \end{aligned}$ | 4E-67 | 59\% (75\%) | Response regulator [Thermobifida fusca] |
|  |  |  |  | $\begin{aligned} & \text { NP_625364.1 } \\ & 221 \mathrm{aa} \end{aligned}$ | 6E-66 | 60\% (74\%) | response regulator [Streptomyces coelicolor A3(2)] |
| 14 | 28 | UNES | 131 | No hit | - |  |  |
| 15 | 30 | UNEZ | 154 | $\begin{aligned} & \text { NP } 649459.2 \\ & 628 \mathrm{aa} \end{aligned}$ | 7.6E-02 | 38\% (60\%) | CG1090-PB [Drosophila melanogaster] |
|  |  |  |  | $\begin{aligned} & \text { NP_730819.1 } \\ & 473 \mathrm{aa} \end{aligned}$ | 7.6E-02 | 38\% (60\%) | CG1090-PA [Drosophila melanogaster] |
|  |  |  |  | $\begin{aligned} & \text { AAM11079.1 } \\ & 428 \mathrm{aa} \end{aligned}$ | 7.6E-02 | 38\% (60\%) | GH23040p [Drosophila melanogaster] |
| 16 | 32 | OXDS | 661 | $\begin{aligned} & \text { NP_242948.1 } \\ & 500 \mathrm{aa} \end{aligned}$ | 1E-52 | 30\% (46\%) | unknown conserved protein [Bacillus halodurans] |
|  |  |  |  | $\begin{aligned} & \text { ZP_00091617.1 } \\ & 480 \mathrm{aa} \end{aligned}$ | 3E-32 | 29\% (41\%) | Putative multicopper oxidases [Azotobacter vinelandii] |
|  |  |  |  | $\begin{aligned} & \text { NP_252457.1 } \\ & 463 \text { aa } \end{aligned}$ | 1E-31 | 28\% (42\%) | metallo-oxidoreductase [Pseudomonas aeruginosa PA01] |

TABLE 7-continued

| Sequence comparison and ORF correlation |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ORF | $\begin{gathered} \text { SEQ } \\ \text { ID } \end{gathered}$ | Family | \# aa | GenBank homology | Probability | $\begin{gathered} \text { \% Identity } \\ (\% \text { Similarity) } \end{gathered}$ | Proposed function of GenBank match |
| 17 | 34 | UNFD | 129 | $\begin{aligned} & \text { NP_437360.1 } \\ & 127 \mathrm{aa} \end{aligned}$ | 7E-33 | 60\% (72\%) | bleomycin resistance protein family [Sinorhizobium meliloti] |
|  |  |  |  | AAO91879.1 $123 \mathrm{aa}$ | 1E-31 | 58\% (74\%) | unknown [uncultured bacterium] |
|  |  |  |  | $\begin{aligned} & \text { NP_103287.1 } \\ & \text { 131aa } \end{aligned}$ | 1E-23 | 48\% (62\%) | unknown protein [Mesorhizobium loti] |
| 18 | 36 | UNFA | 178 |  |  |  |  |
| 19 | 38 | CSMB | 661 | $\begin{aligned} & \text { ZP_00137697.1 } \\ & \text { 769aa } \end{aligned}$ | 1E-166 | 51\% (66\%) | Anthranilate/para-aminobenzoate synthase [Pseudomonas aemuginosa |
|  |  |  |  | NP_250594.1 | 1E-166 | 51\% (66\%) | phenazine biosynthesis protein PhzE |
|  |  |  |  | 627aa |  |  | [Pseudomonas aeruginosa PA01] |
|  |  |  |  | $\begin{aligned} & \text { ZP_00137701.1 } \\ & \text { 687aa } \end{aligned}$ | 1E-166 | 51\% (66\%) | Anthranilate/para-aminobenzoate synthas [Pseudomonas aeruginosa |
| 20 | 40 | AAKD | 427 | $\begin{aligned} & \text { P41403 } \\ & 421 a a \end{aligned}$ | 1E-64 | 38\% (51\%) | Aspartokinase (Aspartate kinase) |
|  |  |  |  | $\begin{aligned} & \text { ZP_00057166.1 } \\ & 445 \text { aa } \end{aligned}$ | 2E-64 | 37\% (52\%) | Aspartokinases [Thermobifida fusca] |
|  |  |  |  | $\begin{aligned} & \text { AAD49567.1 } \\ & 421 \text { aa } \end{aligned}$ | 6E-64 | 37\% (52\%) | aspartokinase subunit A [Amycolatopsis mediterranei] |
| 21 | 42 | ALDB | 274 | $\begin{aligned} & \text { NP_275722.1 } \\ & 266 \mathrm{aa} \end{aligned}$ | 2E-53 | 45\% (64\%) | conserved protein [Methanothermobacter thermautotrophicus] |
|  |  |  |  | $\begin{aligned} & \text { NP_614692.1 } \\ & 270 \text { aa } \end{aligned}$ | 2E-52 | 43\% (61\%) | Fructose-1,6-bisphosphate aldolase [Methanopyrus kandleri AV19] |
|  |  |  |  | $\begin{aligned} & \mathrm{NP} \text { _615406.1 } \\ & 267 \mathrm{aa} \end{aligned}$ | 2E-50 | 43\% (61\%) | fructose-bisphosphate aldolase <br> [Methanosarcina acetivorans str. C2A] |
| 22 | 44 | UNFC | 367 | $\begin{aligned} & \text { NP_275723.1 } \\ & 378 \mathrm{aa} \end{aligned}$ | 4E-46 | 38\% (56\%) | conserved protein [Methanothermobacter thermautotrophicus] |
|  |  |  |  | $\begin{aligned} & \mathrm{NP} \quad 614691.1 \\ & 402_{3} \end{aligned}$ | 2E-45 | 39\% (55\%) | alternative 3 -dehydroquinate synthase [Methanopyrus kandleri |
|  |  |  |  | NP_248244.1 | 2E-43 | 40\% (59\%) | conserved hypothetical protein <br> [Methanococcus jannaschii |
| 23 | 46 | HYDK | 253 | $\begin{aligned} & \text { NP_577771.1 } \\ & 247 \mathrm{aa} \end{aligned}$ | 4E-14 | $31 \%$ (49\%) | metal-dependent hydrolase [Pyrococcus furiosus DSM 3638] |
|  |  |  |  | $\begin{aligned} & \mathrm{NP} \_142108.1 \\ & 247 \mathrm{aa} \end{aligned}$ | 1E-12 | 33\% (52\%) | hypothetical protein PH0093 [Pyrococcus horikoshii] |
|  |  |  |  | $\begin{aligned} & \text { NP_125791.1 } \\ & 248 \mathrm{aa} \end{aligned}$ | 1E-11 | 28\% (50\%) | hypothetical protein [Pyrococcus abyssi] |
| 24 | 48 | ADSA | 438 | $\begin{aligned} & \text { NP_070499.1 } \\ & 433 \mathrm{aa} \end{aligned}$ | 2E-41 | 35\% (49\%) | coenzyme F390 synthetase [Archaeoglobus fulgidus |
|  |  |  |  | $\begin{aligned} & \text { NP_618724.1 } \\ & \text { 434aa } \end{aligned}$ | 5E-41 | 34\% ( $50 \%$ ) | coenzyme F390 synthetase [Methanosarcina acetivorans |
|  |  |  |  | NP_632700.1 | 7E-41 | 35\% (50\%) | Coenzyme F390 synthetase [Methanosarcina mazei Goe1] |
| 25 | 50 | HOXV | 396 | $\begin{aligned} & \text { ZP_00027430.1 } \\ & 442 \mathrm{aa} \end{aligned}$ | 8E-76 | 42\% (59\%) | 2-polyprenyl-6-methoxyphenol hydroxylase [Burkholderia fungorum] |
|  |  |  |  | $\begin{aligned} & \text { NP_627457.1 } \\ & 420 \mathrm{aa} \end{aligned}$ | 1E-71 | 38\% (51\%) | salicylate hydroxylase [Streptomyces coelicolor A3(2)] |
|  |  |  |  | $\begin{aligned} & \text { ZP_00033877.1 } \\ & 403 a a \end{aligned}$ | 2E-68 | 37\% (51\%) | 2-polyprenyl-6-methoxyphenol hydroxylase [Burkholderia fungorum] |
| 26 | 52 | SDRA | 261 | $\begin{aligned} & \text { NP_391080.1 } \\ & \text { 261aa } \end{aligned}$ | $6 \mathrm{E}-58$ | 46\% (57\%) | 2,3-dihydro-2,3-dihydroxybenzoate dehydrogenase [Bacillus subtilis] |
|  |  |  |  | $\begin{aligned} & \text { ZP_00059512.1 } \\ & 260 \mathrm{aa} \end{aligned}$ | 1E-55 | 45\% (56\%) | Dehydrogenase [Thermobifida fusca] |
|  |  |  |  | $\begin{aligned} & \text { AAG31126.1 } \\ & \text { 257aa } \end{aligned}$ | 9E-55 | 46\% (56\%) | MxcC [Stigmatella aurantiaca] |
| 27 | 54 | DHBS | 224 | $\begin{aligned} & \text { Q51790 } \\ & \text { 207aa } \end{aligned}$ | 7E-60 | 56\% (72\%) | isochorismatase |
|  |  |  |  | $\begin{aligned} & \text { Q51518 } \\ & 207 \mathrm{aa} \end{aligned}$ | 1E-58 | 56\% (71\%) | isochorismatase |
|  |  |  |  | $\begin{aligned} & \text { NP } 391077.1 \\ & 312 \mathrm{aa} \end{aligned}$ | 2E-58 | 52\% (69\%) | isochorismatase [Bacillus subtilis] |
| 28 | 56 | SDRA | 233 | $\begin{aligned} & \text { NP_103491.1 } \\ & 242 \mathrm{aa} \end{aligned}$ | 9E-21 | $32 \%$ (49\%) | acyl-carrier protein reductase [Mesorhizobium loti] |
|  |  |  |  | AAL14912.1 <br> 245aa | 1E-15 | 28\% (44\%) | short-chain dehydrogenase [Rhizobium leguminosarum bv. trifolii] |
|  |  |  |  | $\begin{aligned} & \text { NP } 902480.1 \\ & 235 \text { aa } \end{aligned}$ | 7E-15 | 29\% (44\%) | oxidoreductase [Chromobacterium violaceum |
| 29 | 58 | UNIQ | 246 | $\begin{aligned} & \text { S18541 } \\ & \text { 281aa } \end{aligned}$ | $4.5 \mathrm{E}-02$ | 29\% ( $43 \%$ ) | hypothetical protein 3-Streptomyces coelicolor |
|  |  |  |  | $\begin{aligned} & \text { NP_629228.1 } \\ & 281 \mathrm{aa} \end{aligned}$ | 5.9E-02 | 29\% (43\%) | hypothetical protein [Streptomyces coelicolor A3(2)] |

TABLE 7-continued


TABLE 7-continued

| ORF | $\begin{gathered} \text { SEQ } \\ \text { ID } \end{gathered}$ | Family | \# aa | GenBank homology | Sequence comparison and ORF correlation |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  | Probability | $\begin{gathered} \text { \% Identity } \\ \text { (\% Similarity) } \end{gathered}$ | Proposed function of GenBank match |
| 42 | 86 |  | 232 | NP_830809.1 | 8E-08 | 22\% (35\%) | Transporter, LysE family [Bacillus cereus] |
|  |  |  |  | $\begin{aligned} & \text { NP_844737.1 } \\ & 210 \mathrm{aa} \end{aligned}$ | 2E-07 | 22\% (35\%) | homoserine/threonine efflux protein[Bacillus anthracis |
|  |  |  |  | $\begin{aligned} & \text { NP_655752.1 } \\ & 208 \mathrm{aa} \end{aligned}$ | 1E-06 | 22\% (36\%) | LysE, LysE type translocator [Bacillus anthracis |
| 43 | 88 |  | 132 | $\begin{aligned} & \text { NP_827272.1 } \\ & \text { 127aa } \end{aligned}$ | 4E-09 | 36\% (49\%) | hypothetical protein [Streptomyces avermitilis MA-4680] |
|  |  |  |  | $\begin{aligned} & \text { NP_246491.1, } \\ & \text { 112aa } \end{aligned}$ | 5E-02 | 22\% (47\%) | unknown [Pasteurella multocida] |

[0161] The ORFs encoding proteins involved in the biosynthesis of farnesyl dibenzodiazepinones are assigned a putative function and grouped together in families based on sequence similarity to known proteins. To correlate structure and function, the protein families are given a four-letter designation used throughout the description and figures as indicated in Table 8. The meaning of the four letter designations is as follows: AAKD designates an amino acid kinase; ABCA and ABCC designate ABC transporters; ADSA designates an amide synthetase; ALDB designates an aldolase function; CSMB designates a chorismate transaminase; DAHP designates a 3,4-dideoxy-4-amino-D-arabino-heptulosonic acid 7-phosphate synthase activity; DHBS designates a 2,3-dihy-dro-2,3-dihydroxybenzoate synthase activity; DMDA designates a diphosphomevalonate decarboxylase; EFFT designates an efflux protein; HMGA designates a 3-hydroxy-3-methylglutaryl-CoA reductase; HOXV designates a monooxygenase activity; HOYH designates a hydroxylase/ decarboxylase activity; HYDK designates a hydrolase activity; IDSA designates an isopentenyl diphosphate synthase; IPPI designates an isopentenyl diphosphate isomerase; IPTN designates an isoprenyltransferase; KASH designates 3-hy-droxy-3-methylglutaryl-CoA synthase; MVKA designates a mevalonate kinase; MVPK designates a phosphomevalonate kinase; OXAH designates an acylCoA oxidase; OXDS designates an oxidoreductase; RECH, RECI, REGD, REGG and RREB designate regulators; SDRA designates a dehydrogenase/ketoreductase, SPKG designates a sensory protein kinase; UNES, UNEZ, UNFA, UNFC, UNFD, UNFE, UNFJ and UNIQ designate proteins of unknown function.

TABLE 8

| FAMILY | FUNCTION: |
| :--- | :--- |
| AAKD | amino acid kinase; strong homology to <br> primary aspartate kinases, converting L- <br> aspartate to 4-phospho-L-aspartate |
| ABCA | ABC transporter <br> ABCC transporter <br> ADSA |
| ALDB | adenylating amide synthetase <br> aldolase; similarity to fructose-1,6- <br> biphosphate aldolase that generates D- <br> glyceraldehyde-3Ph, precursor of D- <br> erythrose-4Ph involved in the shikimate <br> pathway <br> chorismate transaminase, similarity to <br> anthranilate synthase |
| DSMB | DAHP synthase, class II; involved in <br> formation of aminoDAHP from PEP and <br> erythrose-4-phosphate |
|  | DAHP |

TABLE 8-continued

| FAMILY | FUNCTION: |
| :---: | :---: |
| DHBS | 2,3-dihydro-2,3-dihydroxybenzoate synthase (isochorismatase) |
| DMDA | diphosphomevalonate decarboxylase (mevalonate pyrophosphate decarboxylase) |
| EFFT | efflux protein |
| HMGA | HMG-CoA reductase; converts 3-hydroxy-3-methylglutaryl-CoA to mevalonate plus CoA in isoprenoid biosynthesis |
| HOXV | FAD monooxygenase; shows homology to a variety of monooxygenases including salicylate hydroxylases, zeaxanthin epoxidases |
| HOYH | hydroxylase/decarboxylase; FADdependent monooxygenase |
| HYDK | hydrolase |
| IDSA | isoprenyl diphosphate synthase, catalyzes the addition of 2 molecules of isopentenyl pyrophosphate to dimethylallyl pyrophosphate to generate GGPP |
| IPPI | isopentenyl diphosphate isomerase, catalyzes the isomerization of IPP to produce dimethylallyl diphosphate |
| IPTN | isoprenyltransferase; catalyzes covalent N terminal attachment of isoprenyl units to amide groups of nitrogen-containing heterocycle rings |
| KASH | HMG-CoA synthase; condenses acetyl-CoA with acetoacetyl-CoA to form 3-hydroxy-3-methylglutaryl-CoA |
| MEBI | membrane protein |
| MVKA | mevalonate kinase; converts mevalonate to 5 -phosphomevalonate in the mevalonate pathway of isoprenoid biosynthesis |
| MVKP | phosphomevalonate kinase; converts 5phosphomevalonate to 5-diphosphomevalonate in the mevalonate pathway of isoprenoid biosynyhesis |
| OXAH | acyl CoA oxidase |
| OXDS | oxidoreductase |
| RECH | regulator |
| RECI | regulator; similarity to PadR transcriptional regulators involved in repression of phenolic acid metabolism |
| REGD | transcriptional regulator; relatively large regulators with an N -terminal ATP-binding domain containing Walker A and B motifs and a C-terminal LuxR type DNA-binding domain |
| REGG | regulator |
| RREB | response regulator; similar to response regulators that are known to bind DNA and act as transcriptional activators |

TABLE 8-continued

| FAMILY | FUNCTION: |
| :--- | :--- |
| SDRA | dehydrogenase/ketoreductase, NAD-dependent |
| SPKD | sensory protein kinase, two component system |
| SPKG | sensory protein kinase, two component system |
| UNES | unknown function |
| UNEZ | unknown function |
| UNFA | unknown function |
| UNFC | unknown function |
| UNFD | unknown function |
| UNFE | putative membrane protein |
| UNFJ | unknown function |
| UNIQ | unknown function |

[0162] Biosynthesis of ECO-04601 involves the action of various enzymes that synthesize the three building blocks of the compound, namely the farnesyl-diphosphate component (FIG. 5), the 3-hydroxy-anthranilate-adenylate component (FIG. 6) and the 2-amino-6-hydroxy-benzoquinone component (FIG. 7) that are subsequently condensed to form the final compound (FIG. 8).
[0163] The farnesyl-diphosphate biosynthesis involves the concerted action of seven enzymes (FIG.5). ORF 10 (KASH) (SEQ ID NO: 20) encodes a hydroxymethylglutaryl-CoA synthase that catalyzes an aldol addition of acetyl-CoA onto acetoacyl-CoA to yield 3-hydroxy-3-methylglutaryl-CoA (HMG-CoA). This product is subsequently reduced through the action of ORF 9 (HMGA) (SEQ ID NO: 18) to form mevalonic acid (MVA). ORF 5 (MVKA) (SEQ ID NO: 10) phosphorylates mevalonate to 5'-phosphomevalonate using ATP as the phosphate donor. The next step in the farnesyldiphosphate biosynthesis is the phosphorylation reaction of the 5 '-phosphomevalonate to $5^{\prime}$-pyrophosphomevalonate (DPMVA) that is catalyzed by ORF 7 (MVKP) (SEQ ID NO: 14). Subsequent decarboxylation of $5^{\prime}$-pyrophosphomevalonate catalyzed by ORF 6 (DMDA) (SEQ ID NO: 12) yields isopentenyl diphosphate (IPP) which is then converted to dimethylallyldiphosphate (DMADP) through the action of ORF 8 (IPPI) (SEQ ID NO: 16) that has isomerase enzymatic activity. The final step in the biosynthesis of farnesyl-diphosphate is the condensation of one molecule of dimethylallyldiphosphate with two molecules of isopentenyl diphosphate catalyzed by the isoprenyl diphosphate synthase ORF 4 (IDSA) (SEQ ID NO: 8). The described pathway involved in synthesis of farnesyl-diphosphate is entirely consistent with related mevalonate pathways described in other actinomycete species (Takagi et al., J. Bacteriol. 182, 4153-4157, (2000)).
[0164] Biosynthesis of the 3-hydroxy-anthranilate component involves the use of precursors derived from the shikimate pathway (FIG. 6). Chorismic acid is transaminated through the action of ORF 19 (CSMB) (SEQ ID NO: 38) to form aminodeoxyisochorismic acid. This enzyme resembles anthranilate synthases and is likely to catalyze specifically the transfer of the amino group using glutamine as the amino donor. The next step involves isochorismatase activity and is mediated by ORF 27 (DHBS) (SEQ ID NO: 54). This reaction consists in the removal of the pyruvate side chain from aminodeoxyisochorismic acid to form 6 -amino-5-hydroxy-cyclohexa-1,3-dienecarboxylic acid. This compound is subsequently oxidized through the action of ORF 26 (SDRA) (SEQ ID NO: 52) yielding 3-hydroxy-anthranilic acid. ORF 24 (ADSA) (SEQ ID NO: 48) catalyzes the activation of 3-hydroxy-anthranilic acid through adenylation generating the 3-hydroxy-anthranilate-adenylate component (FIG. 6).
[0165] Biosynthesis of the 2-amino-6-hydroxy-benzoquinone component of the farnesyl dibenzodiazepinone, requires components derived from the aminoshikimate pathway. FIG. 7 depicts the series of enzymatic reactions involved in the biosynthesis of this constituent. ORF 21 (ALDB) (SEQ ID NO: 42) resembles aldolases involved in the generation of precursors of D-erythrose-4-phosphate which is part of the aminoshikimate pathway used for the generation of 2-amino-6-hydroxy-[1,4]-benzoquinone. ORF 33 (DAHP) (SEQ ID NO: 67) catalyzes the initial step in the aminoshikimate pathway that corresponds to the formation of 3,4-dideoxy-4-amino-D-arabino-heptulosonic acid 7-phosphate (amino DAHP) from phosphoenolpyruvate (PEP) and erythrose 4-phosphate (E-4Ph). Subsequent reactions leading to 3 -amino-5-hydroxy-benzoic acid are catalyzed by enzymes provided by primary metabolism biosynthetic pathways present in Micromonospora sp. strain 046-ECO11. ORF 25 (HOXV) (SEQ ID NO: 50) hydroxylates 3-amino-5-hy-droxy-benzoic acid at position 2, generating 3-amino-2,5-dihydroxy-benzoic acid. This intermediate is further modified by ORF 32 (HOYH) (SEQ ID NO: 65) that catalyzes a decarboxylative oxidation reaction yielding 6 -amino-ben-zene-1,2,4-triol. A final oxidation reaction is performed by ORF 16 (OXDS) (SEQ ID NO: 32) yielding 2-amino-6-hy-droxy-[1,4]-benzoquinone (FIG. 7).
[0166] Assembly of the three components resulting in the farnesyl dibenzodiazepinone is catalyzed by ORFs 24 and 11 (FIG. 8). ORF 24 (ADSA) (SEQ ID NO: 48) catalyzes the condensation of the adenylated 3-hydroxy-anthranilate with the 2 -amino-6-hydroxy-[1,4]-benzoquinone component. A spontaneous condensation between the free amino group of the 3-hydroxy-anthranilate and one of the carbonyl groups present on the 2-amino-6-hydroxy-[1,4]-benzoquinone component occurs yielding a dibenzodiazepinone intermediate. This compound is further modified through transfer of the farnesyl group of the farnesyl-diphosphate intermediate onto the nitrogen of the amide of the dibenzodiazepinone catalyzed by ORF 11 (IPTN) (SEQ ID NO: 22) and resulting in the formation of the farnesyl dibenzodiazepinone (FIG. 8).
[0167] Additional ORFs, namely ORF 2 (RECH) (SEQ ID NO: 4), ORF 3 (REGD) (SEQ ID NO: 6), ORF 12 (SPKG) (SEQ ID NO: 24), ORF 13 (RREB) (SEQ ID NO: 26), ORF 34 (REGG) (SEQ ID NO: 69) and ORF 36 (RECI) (SEQ ID NO: 74) are involved in the regulation of the biosynthetic locus encoding the farnesyl dibenzodiazepinone. Other ORFs, namely ORF 1 (ABCC) (SEQ ID NO: 2), ORF 31 (EFFT) (SEQ ID NO: 62), ORFs 39 and 40 (ABCA) (SEQ ID NOS: 80 and 82, respectively) and ORF 42 (SEQ ID NO: 86) are involved in transport. Other ORFs involved in the biosynthesis of the farnesyl dibenzodiazepinone include ORF 20 (AAKD) (SEQ ID NO: 40), ORF 23 (HYDK) (SEQ ID NO: 46), ORF 38 (OXAH) (SEQ ID NO: 78) as well as ORFs 14, $15,17,18,22,29,30,35,37,41$ and 43 (SEQ ID NOS: 28, 30, $34,34,44,58,60,71,76,84$ and 88 , respectively) of unknown function.

## Example 6 <br> Farnesyl Dibenzodiazepinone Loci from Actinomycetes Species

[0168] A. Correlation of Loci A, B and C
[0169] Loci related to the biosynthetic locus present in Micromonospora sp. strain $046 \mathrm{ECO}-11$ as described in Example 5 (referred to herein as locus A) and directing the biosynthesis of farnesyl diabenodiazepinones related to ECO-04601 were detected in the genome of two actinomycetes using the genome scanning method described in U.S.

Ser. No. 10/232,370, CA 2,352,451 and Zazopoulos et. al., Nature Biotechnol., 21, 187-190 (2003).
[0170] Locus B (052E) was detected in Micromonospora echinospora challisensis NRRL 12255. The locus spans approximately 38,000 base pairs of DNA and encodes 33 proteins. Locus C (237C) was detected in Streptomyces carzinostaticus neocarzinostaticus ATCC 15944. This locus spans approximately 37,000 base pairs of DNA and encodes 33 proteins. More than 10 kilobases of DNA sequence were analyzed on each side of the two loci and these regions were deemed to contain primary genes.
[0171] In order to identify the function of the proteins coded by the genes forming the biosynthetic loci $B$ an $C$ the gene products of their ORFs 1 to 33, were compared, using the BLASTP version 2.2.10 algorithm with the default parameters, to sequences in the National Center for Biotechnology Information (NCBI) nonredundant protein database and the DECIPHER $\mathbb{B}$ database of microbial genes, pathways and natural products (Ecopia BioSciences Inc. St.-Laurent, QC, Canada).
[0172] The ORFs encoding proteins present in loci $\mathrm{A}, \mathrm{B}$, and C are assigned a putative function and grouped together in families based on sequence similarity to known proteins. To correlate structure and function, the protein families are given a four-letter designation used throughout the description and figures as indicated in Table 8 of Example 5.
[0173] Comparison of loci A, B and C clearly indicates that all three loci are related and encode similar enzymatic functions. Therefore, the compounds produced by the enzymes encoded by loci B and C are structurally closely related to ECO-04601. Table 9 correlates the protein families of loci B and C to those of locus A. All 33 ORFs found in locus B have counterparts in locus A. Similarly, all 33 ORFs present in locus C have counterpart proteins in locus A, with the exception of ORFs 30,31 , and 32 that encode a sensory protein kinase protein, a response regulator and a membrane protein. These observations suggest that the compounds produced by loci $B$ and $C$ encoded proteins share a high degree of similarity with ECO-04601.

TABLE 9

| Loci A, B and C ORFs function and correlation |  |  |  |
| :--- | :---: | :---: | :---: |
|  | A | B | C |
|  | 1 | - | - |
| ABCC | 2 | 1 | 1 |
| RECH | 3 | 2 | 2 |
| REGD | 4 | 3 | 3 |
| IDSA | 5 | 4 | 4 |
| MVKA | 6 | 5 | 5 |
| DMDA | 7 | 6 | 6 |
| MVKP | 8 | 7 | 7 |
| IPPI | 9 | 8 | 8 |
| HMGA | 10 | - | 9 |
| KASH | 11 | 9 | 10 |
| IPTN | 12 | 15 | 12 |
| SPKG | 13 | 16 | 11 |
| RREB | 14 | 10 | 33 |
| UNES | 15 | 14 | - |
| UNEZ | 16 | 13 | - |
| OXDS | 17 | 12 | - |
| UNFD | 18 | 11 | - |
| UNFA | 19 | 17 | 14 |
| CSMB | 20 | 18 | 15 |
| AAKD | 21 | 19 | 16 |
| ALDB | 22 | 20 | 17 |
| UNFC | 23 | 21 | 18 |
| HYDK | 23 |  |  |

TABLE 9-continued

| Loci A, B and C ORFs function and correlation |  |  |  |
| :--- | :--- | :--- | :--- |
|  | A | B | C |
| ADSA | 24 | 22 | 19 |
| HOXV | 25 | 23 | 20 |
| SDRA | 26 | 24 | 21 |
| DHBS | 27 | 25 | 22 |
| SDRA | 28 | 26 | 23 |
| UNGA | 29 | 27 | 24 |
| UNFE | 30 | 28 | 25 |
| EFFT | 31 | 29 | 26 |
| HOYH | 32 | 30 | 27 |
| DAHP | 33 | 31 | 28 |
| REGG | 34 | 32 | - |
| UNFJ | 35 | 33 | $13 / 29$ |
| RECI | 36 | - | - |
| UNIQ | 37 | - | - |
| OXAH | 38 | - | - |
| ABCA | 39 | - | - |
| ABCA | 40 | - | - |
| UNIQ | 41 | - | 30 |
| SPKD | - | - | 31 |
| RREB | - | - | 32 |
| MEBI | - |  |  |

[0174] FIG. 5 depicts the three biosynthetic loci A, B and C. All ORFs are represented by arrows and their orientation indicate the direction of the transcription of each ORF; highlighted ORFs are involved in the biosynthesis of the farnesyl unit. ORFs 4, 5, 6, 7, 8, 9, and 10 in locus A participate in the synthesis of the farnesyl unit present in the farnesyl dibenzodiazepinone. Counterparts of these ORFs are found in locus $B$ (ORFs 3, 4, 5, 6, 7 and 8 ) as well as in locus C(ORFs 3, 4, 5, $6,7,8$ and 9). As shown in FIG. 5, proteins encoded by these ORFs participate in an orderly fashion in the biosynthesis of farnesyl-diphosphate component starting with acetoacetylCoA and acetyl-CoA. All enzymes necessary for the synthesis of farnesyl-diphosphate are present in all three loci with the exception of a hydroxymethylglutaryl-CoA synthase (KASH) which is absent from locus B. The product of this enzymatic reaction, 3-hydroxy-3-methylglutaryl-CoA is provided by an alternative biosynthetic pathway of the primary metabolism of the microorganism or by a hydroxymethylglu-taryl-CoA synthase located elsewhere in the genome. The described pathway involved in synthesis of farnesyl-diphosphate is entirely consistent with related mevalonate pathways described in other actinomycete species (Takagi et al., J. Bacteriol. 182, 4153-4157, (2000) and FIG. 5).
[0175] FIG. 6 depicts ORFs 19, 21, 24, 26 and 27 in locus A involved in the biosynthesis of the 3-hydroxy-anthranilate component of the farnesyl dibenzodiazepinone. Counterparts of these ORFs are found in locus B (ORFs 17, 19, 22, 24 and 25) as well as in locus C(ORFs 14, 16, 19, 21 and 22). As shown in FIG. 6, proteins encoded by these ORFs participate in an orderly fashion to the biosynthesis of the 3-hydroxy-anthranilate-adenylate component starting with precursors from the pentose phosphate pathway and chorismic acid. In particular, the enzyme responsible for the adenylation of 3-hydroxy-anthranilic acid (ADSA) that corresponds to ORFs 24, 22 and 19 in loci A, B and C respectively is present in all three loci as well as the remaining enzymes that participate in the biosynthesis of 3-hydroxy-anthranilate component present in dibenzodiazepinones.
[0176] FIG. 7 highlights ORFs 16, 24, 25, 32 and 33 in locus A involved in the biosynthesis of the 2 -amino- 6 -hy-
droxy-[1,4]benzoquinone component of the farnesyl dibenzodiazepinone. Counterparts of these ORFs are found in locus B (ORFs 13, 19, 23, 30 and 31) as well as in locus C(ORFs 16, 20, 27 and 28) with the exception of ORF corresponding to the oxidoreductase (OXDS) present in loci A and B. As shown in FIG. 7, proteins encoded by these ORFs participate in an orderly fashion in the biosynthesis of the 2-amino-6-hydroxy-[1,4]benzoquinone component starting with precursors from the pentose phosphate pathway and 3,4-dideoxy-4-amino-D-arabino-heptulosonic acid 7-phosphate (amino DAHP).
[0177] FIG. 8 highlights ORFs 11 (SEQ ID NO: 22) and 24 (SEQ ID NO: 48) in locus A involved in the assembly of all three components, 3-hydroxy-anthranilate, 2-amino-6-hy-droxy-[1,4]benzoquinone and farnesyl-diphosphate to form the farnesyl dibenzodiazepinone. Counterparts of these ORFs are found in locus B (ORFs 9 (SEQ ID NO: 90) and 22 (SEQ ID NO: 92)) as well as in locus C(ORFs 10 (SEQ ID NO: 94) and 19 (SEQ ID NO: 96)). The isoprenyltransferase ORF 10 of locus C (SEQ ID NO: 96) is partial and represents the N-terminal part of the protein. IPTN ORFs 11 (SEQ ID NO: 22), 9 (SEQ ID NO: 90) and 10 (SEQ ID NO: 94) in loci A, B and $C$ respectively catalyze the transfer of the farnesyl unit onto the core element of the farnesyl dibenzodiazepinone and related compounds produced by loci B and C. ADSA ORFs 24 (SEQ ID NO: 48), 22 (SEQ ID NO: 92) and 19 (SEQ ID NO: 96) in loci A, B and C respectively catalyze the condensation of 3-hydroxy-anthranilate and 2-amino-6-hydroxy-[1, 4]benzoquinone and farnesyl-diphosphate to form the dibenzodiazepinone core element of ECO-04601 and related compounds produced by loci B and C .
[0178] B. Clustal ${ }^{\text {TM }}$ Alignments
[0179] Alignements of isoprenyl transferases (IPTN) and adenylating amide synthetases (ADSA) of loci $\mathrm{A}, \mathrm{B}$ and C , respectively presented in FIGS. 9 and 10, were generated by the Clustal ${ }^{\text {TM }}$ alignment method.
[0180] FIG. 9 shows an alignment of ORFs 11 (SEQ ID NO: 22), 9 (SEQ ID NO: 90, which represents the polypeptide deduced from SEQ ID NO:91) and 10 (SEQ ID NO: 94, which represents the polypeptide deduced from SEQ ID NO:95) in loci $\mathrm{A}, \mathrm{B}$ and C respectively, highlighting the phylogenetic relatedness of these three proteins. The amino acid sequence of all three proteins is extremely conserved as shown by the codes on the fourth line, suggesting that these proteins share a well-conserved and related isoprenyltransferase enzymatic function. The following consensus amino acid sequence (also as SEQ ID NO: 98) that represents all three sequences was generated using the hmmemit algorithm (HMMER, Washington University in St-Louis, School of Medicine, MO, USA, http://hmmer.wustl.edu):
[0181] "AaELysviEesARILdvaCsrDrvwpiL-
saYGDaFaHpaavvAFRvAtalRHvGELD CRFttHPddRD-PYAIALsrGLtPktdH-
PvGsLLsevqeRIPvesyGiDFGvvGGFKKiYafFtPDe LqevaaLAgiPamPRsLAgnadFFeR-
yGlddrvGvlGiDYPartvnvyfndvpaesfesetirstlreiGma epsermI kIGekafGlyvtlGwdsseiericyaaattdIttIpvpvepeiekfvksvpyGGedrkfvyGvaltpkGey ykleshykwkpGavdfi"
[0182] FIG. 10 shows an alignment of ORFs 24 (SEQ ID NO: 48), 22 (SEQ ID NO: 92, which represents the polypeptide deduced from SEQ ID NO: 93) and 19 (SEQ ID NO: 96, which represents the polypeptide deduced from SEQ ID NO: 97) in loci A, B and C respectively, highlighting the phyloge-
netic relatedness of these three proteins. The amino acid sequence of all three proteins is extremely conserved as shown by the codes on the fourth line, suggesting that these proteins share a well-conserved and related adenylating amide synthetase enzymatic function. The following consensus amino acid sequence (also as SEQ ID NO: 99) that represents all three sequences was generated using the hmmemit algorithm:

## "VneprssLPrLGqWhGpEDLrrLqEKqLaqtvtWAaRsPFYRdRLds

gAlPvt aaDLAdLPLtt KqDLRDnYPFGmLAvPkERLAt YHEssGtAGr
PtPsYYtAeDWtDLAERFARKWiGmsAeDvFLvRtPYALLLtGHLAH

AAgRLrGAt vvPGDnRsLAmPYARvvRvmHDLgvtLtWsvPtECLiW
AAAAtAAGHRPdvDFPALRALFvGGEPltdARRrRisRLWGvPviEE

YGst EtGsLAGECPeGRIHLWADRALFEvYDPdtGtvrAdGdGqLvv
tPLfREAmPLLRYnLEDnvsvsYDDCaCGWkLPtvrvLGRaAFGyRv

GattitqHrLEElvFsLPeahrvvFwRAkAEPavLRiEiEvaeeHRv
AAeAELtasvRaaFGvDsevtGLaPGtLiPreALtsmPDvvKPRsLF GPDEDWgKALLYY"
[0183] The amino acid shown for the consensus sequences (SEQ ID NOs: 98 and 99) are the highest probability amino acid at that position according to the HMM (hidden Markov model). Highly conserved residues (those with a probability of $>0.5$ ) are shown by capital letters while other residues (lowercase letters) are deduced by the program from the most common amino acid found at the specific position in the aligned proteins (HMMER User's Guide, Sean Eddy, October 2003, Washington University of Medicine, MO, USA, p 23-24).

## Example 7

## Labeled 3-Hydroxyanthranilic Acid Feeding

[0184] This experiment was designed to confirm the farnesyl dibenzodiazepinone biosynthetic pathway involves a 3-hydroxyanthranilate intermediate. First, labeled 4,6-dideu-terio-3-hydroxyanthranilic acid was prepared. Then the labeled intermediate was fed to the Micromonospora sp. strain, the product was purified (see Example 2) and the results were analyzed. The following is an exemplary procedure to accomplish the feeding experiment:
[0185] A. Preparation of 4,6-dideuterio-3-hydroxyanthranilic acid

[0186] 3-Hydroxyanthranilic acid (108 mg, Sigma-Aldrich) was suspended in $\mathrm{D}_{2} \mathrm{O}(2 \mathrm{~mL})$. Potassium t-butoxide ( 154 mg ) was added to give a brown solution. The solution was stirred at $100^{\circ} \mathrm{C}$. under nitrogen for about 6 days. The
reaction mixture was cooled to room temperature. The solution was acidified to pH 6 with 10 N hydrochloric acid and white solid precipitated. The solid was filtered and dried in vacuo ( 93 mg ). The ${ }^{1} \mathrm{H}$ NMR of the isolated product showed about $92-96 \%$ reduction of the proton signals (doublets) at the 4 and 6 positions. The ${ }^{1} \mathrm{H}$ NMR signal of the unchanged proton ( 5 position) also reflected the incorporation of the two deuterium; coupling to the 4 and 6 protons was nearly lost (triplet changed to a singlet having two very small side peaks).
[0187] B. 4,6-dideuterio-3-hydroxyanthranilic Acid Feeding

## B.1. Culture Conditions:

[0188] To prepare a vegetative culture, Micromonospora sp. 046-Eco11 was grown on ISP2 agar (Difco) for 10 to 15 days, and the surface growth from the agar plate was homogenized and transferred to a 125 ml flask containing three glass beads ( 5 mm diameter), and 25 ml of sterile medium KH composed of 10 g glucose, 20 g potato dextrin, 5 g yeast extract, 5 g NZ -Amine A , and 1 g CaCO 3 made up to one liter with tap water and adjusted to pH 7 with 1 M NaOH .). This vegetative culture was incubated at $28^{\circ} \mathrm{C}$. for about 70 hours on a shaker at 250 rpm with a 1 -inch throw.
[0189] Following incubation, 18 ml was used to inoculate 2 L baffled flasks each containing 600 ml of sterile Hi production medium consisting of 20 g potato dextrin, 30 g glycerol, 2.5 g Bacto-peptone, 8.34 g yeast extract, and $3 \mathrm{~g} \mathrm{CaCO}_{3}$ made to one liter with distilled water and adjusted at pH 7.0 with 1 M NaOH . The culture was incubated at $28^{\circ} \mathrm{C}$. for about 96 hours on a shaker at 250 rpm with 1 -inch throw.

## B.2. Feeding Experiment:

[0190] Vegetative cultures of Micromonospora sp. 046Ecol 1 prepared in medium KH as explained above were used to inoculate Hi medium (four $125-\mathrm{mL}$ flasks containing 25 mL ). The medium was fed with $4,6-\mathrm{D}_{2}-3$-hydroxyanthranilic acid at $0.5 \mathrm{mg} / \mathrm{mL}$ before inoculation with the vegetative culture at $2 \%$ level. Control cultures without adding the labeled compound were prepared for each medium in the same way mentioned above. Effect of adding 4,6- $\mathrm{D}_{2}-3$-hydroxyanthranilic acid on the production titre and growth was measured by adding the unlabeled compound to each medium in the same fashion explained above. The purified compound obtained from each experiment was tested by ${ }^{1} \mathrm{H}-\mathrm{NMR}$ for incorporation ratio of the labeled substrate.
[0191] C. Results:
[0192] The purified farnesyl dibenzodiazepinone from the feeding experiment was analyzed both by ${ }^{1} \mathrm{H}$ NMR and mass spectrum. The ${ }^{1} \mathrm{H}$ NMR (in $\mathrm{DMSO}_{-1}$ ) was compared to the unlabelled standard. About $31 \%$ reduction in the intensity of the signals at 6.82 and 7.06 ppm in $\mathrm{DMSO}-\mathrm{d}_{6}$ (correspond to protons signals at 6.83 and 7.14 ppm in $\mathrm{MeOH}-\mathrm{d}_{4}$ ) was observed, which reflected a $31 \%$ incorporation of the deuterium at these positions. Mass spectral analysis gave about $47 \%$ incorporation of the deuterium labeled precursor.

[0193] The result indicated a direct incorporation of 3-hydroxyanthranilate as a precursor in the biosynthesis of ECO04601.

## Example 8

## Methods of Using the Deposited Cosmids

[0194] Two deposits of E. coli DH10B vectors (046KM and 046KQ), having deposit accession numbers IDAC 250203-06 and IDAC 250203-07 respectively, each contain a cosmid clone and together span the whole biosynthetic locus of ECO04601. The coverage of the locus by each deposited cosmid is described in Example 5 and shown on FIG. 4.
[0195] Culture conditions to be employed for growing the deposited cosmid-containing $\mathrm{DH} 10 \mathrm{~B}^{\mathrm{Tm}} E$. coli are understood by a person of skill in the art (Sambrook et al. (1989) Molecular Cloning: A Laboratory Manual, $2^{\text {nd }}$ ed., Cold Spring Harbor Laboratory Press). As a non-limiting example, upon receiving a sample of the deposited strain, either as a frozen glycerol stock or as an agar stab or in a liquid media, a small aliquot of the strain is gathered using a sterile metal loop and thereafter streaked onto a selective media agar on freshly prepared growth plates (e.g. disposable plastic Petri®) plates). The aliquot is streaked so that single bacterial colonies can be isolated. A number of different growth media can be used, provided that the media contain an appropriate amount of a selective agent, for example an antibiotic. Standard growth media are known in the art, such as standard Luria Bertani (LB) media ( 10 grams of $\mathrm{NaCl}, 10$ grams of tryptone, 5 grams of yeast extract, 20 grams of agar, with pH adjusted to 7.0 with 5.0 N NaOH add deionized water to a final volume of 1.0 liters, autoclaved then cooled to $55^{\circ} \mathrm{C}$. followed by addition of 10 mL of $10-\mathrm{mg} / \mathrm{mL}$ filter-sterilized ampicilin or 5 ml of $10-\mathrm{mg} / \mathrm{mL}$ filter-sterilized kanamycin). Plates with streaked bacteria are incubated overnight (approximately 16 hours) at $37^{\circ} \mathrm{C}$. to allow for growth of the bacterial colonies.
[0196] Cosmid DNA containing insert DNA are prepared from the above-noted strains by methods that are known in the art. As a non-limiting example, a single bacterial colony is selected from an agar plate (as referred to above) and restreaked onto a fresh agar plate, containing the appropriate selective agent as noted above, and allowed to grow overnight at $37^{\circ} \mathrm{C}$. From this second agar plate, a single bacterial colony is selected and inoculated into 2.0 to 5.0 ml of liquid broth containing the appropriate amount of a selective agent, for example LB broth (prepared as per LB media, but lacking agar) containing ampicillin or kanamycin in a concentration as noted in the preceding paragraph, in order to generate a liquid starter culture of the single bacterial colony. This starter culture is grown to late logarithmic stage (approximately 8 hours), at which time an aliquot of the starter culture is withdrawn and diluted, by a factor of 500 to 1000 , into a volume
of broth containing the selective agent and grown with vigorous shaking (approximately 300 revolutions per minute) to late logarithmic/stationary phase (approximately 10 to 12 hours) to achieve a cell density of approximately 3 to $4 \times 10^{9}$ cells per ml . Cell density is estimated by taking an aliquot of the liquid culture and obtaining an $\mathrm{OD}_{600}$ reading using a spectrophotometer, or by centrifuging the liquid culture and thereafter measuring the weight of the resulting bacterial pellet. Typically, 1.0 liter volume of an liquid culture of $E$. coli that is grown overnight at $37^{\circ} \mathrm{C}$., 300 rpm with a cell density of approximately 3 to $4 \times 10^{9}$ cells per ml will correspond to a pellet weight of approximately $3 \mathrm{~g} / 1$. Depending on the desired amount of insert-bearing cosmid DNA that is required, a person skilled in the art would understand that either a liquid "mini-culture" of 2.0 to 5.0 ml or a liquid "maxi-culture" of 500 ml may be required to be grown to result in the desired amount of cosmid DNA to be isolated.
[0197] Cosmid DNA, bearing the insert DNA of interest, is isolated from the bacteria grown in liquid cultures, as described in the preceding paragraph, using procedures that are known in the art. Non-limiting examples include the use of commercially available kits, for example the QIAGEN( ${ }^{\text {B }}$ Large-Construct Kit (QIAGEN Inc., Catalogue No. 12462) or Perfectprep $(\mathbb{B})$ BAC 96 Kit (catalogue order number 955150431 ) available from Eppendorf North America (Westbury, N.Y.). Alternatively, the insert-bearing cosmid DNA is isolated by following procedures detailed for a traditional alkaline lysis method as described in Birnboim and Doly (1979) Nucleic Acids Research 7(6): 1513-1523, or in a cosmid-specific manual (e.g. the SuperCos ${ }^{\mathrm{TM}} 1$ Cosmid Vector Kit Instruction Manual published online at www.stratagene.com). As an example of an alkaline lysis procedure, insert-bearing cosmid-containing bacterial cells from a 5.0 ml culture are collected by centrifugation (using an appropriate, sterile centrifuge tube) for 2 minutes followed by aspiration of the supernatant and resuspension of the pellet by vortexing in $200 \mu 1$ of an ice cold solution of 50 mM glucose, 10 mM EDTA, 25 Mm Tris- HCl (pH 8.0). Following resuspension of the bacteria, $400 \mu$ of a freshly prepared solution of $0.2 \mathrm{~N} \mathrm{NaOH}, 1 \% \mathrm{SDS}$ is added and the contents gently mixed by inversion (vortexing must be avoided), followed by incubation on ice for 5 minutes. Following incubation on ice, $300 \mu \mathrm{l}$ of ice-cold potassium acetate (approximate pH 4.8 ) is added, and the tube gently inverted twice and incubated on ice for a further 5 minutes. The tube is then centrifuged for 5 minutes at $4^{\circ} \mathrm{C}$. and $500 \mu \mathrm{l}$ of the supernatant is transferred to a fresh (sterile) tube. The transferred supernatant is deproteinated by extraction with phenol-chloroform, keeping the upper phase to which is then added 1.0 ml of ethanol. The tube is left standing at room temperature for 5 minutes, and thereafter microfuged for 30 minutes, followed by aspiration of the liquid from the tube. The remaining DNA pellet is washed in $70 \%$ ethanol, centrifuged (in a microfuge), and after aspiration of the liquid and drying (avoiding complete dryness) of the pellet, the DNA is resuspended in $50 \mu \mathrm{l}$ of Tris-EDTA (TE). DNA concentration is estimated by taking an $\mathrm{OD}_{600}$ reading on a $1 / 100$ diluted aliquot of the purified insertbearing cosmid DNA. The insert-bearing cosmid DNA is thereafter used in any number of downstream applications that would be appreciated by a person skilled in the art.
[0198] Segments or regions of the insert DNA can be generated by performing a restriction digestion on the insertbearing cosmid DNA using protocols that are known to those of skill in the art. The segments or regions of the insert DNA
may be of interest to the person of skill in the art as the particular nucleotide may be that for a gene(s) that is to be manipulated for a downstream application. As well, the segments or regions of the insert DNA may be of interest to the person of skill in the art as the particular nucleotide may be that for an entire biosynthetic locus, or a portion thereof, that encodes for the production of a natural product. It is possible that the nucleotide sequence of the insert DNA encodes one or more modules, which may be comprised of one or more domains, of a nonribosomal peptide synthetase or a polyketide synthase locus encodes for the production of a bioactive natural product.
[0199] As an example that is not intended to be limiting, if the sequence of the insert DNA is known, the presence of particular restriction enzyme sites within the insert DNA are determined and the region (i.e. the fragment) of DNA situated between two restriction enzyme sites cut or digested from the cosmid DNA. Generally, it is preferred in the art to use a restriction enzyme that recognizes a six base pair (bp) DNA recognition sequence as opposed to a four base pair recognition site, as there will be fewer restriction sites in a given stretch of DNA for six bp restriction enzyme, thereby offering less chance of digesting the cosmid (i.e. the vector) DNA per se. Selection of a given restriction enzyme may also be dependent upon whether the ends of the generated DNA fragment are to be blunt or are to possess overhangs so as to facilitate sub-cloning of the DNA fragment. Restriction digestion conditions are known to those skilled in the art. While not intending to be limiting, a digestion is usually performed using a minimum of $0.2 \mu \mathrm{~g}$ of DNA. If the DNA fragment to be generated is to be used as a probe, for example in Southern blotting, then an amount of DNA of at least $10 \mu \mathrm{~g}$ will be required for digestion. A restriction digestion can usually be performed in a range of reaction volume between $10 \mu \mathrm{l}$ to 50 $\mu 1$, using a requisite number of units of the given restriction endonuclease plus the particular buffer for the restriction enzyme and a necessary amount of sterile water to give the desired reaction volume. One unit of a restriction endonuclease will digest $1 \mu \mathrm{~g}$ of DNA in one hour, and it is common to use a ten-fold excess of the restriction enzyme to ensure complete digestion, provided that the volume of the restriction enzyme used does not exceed $10 \%$ of the final reaction volume. Upon addition of the restriction enzyme as the last component of the reaction mixture, the tube containing the mixture should be gently flicked with a finger to ensure proper mixing of the tube contents, followed by a brief centrifugation and incubation of the tube at $37^{\circ} \mathrm{C}$., or at an elevated temperature $50-65^{\circ} \mathrm{C}$. if the restriction enzyme is one isolated from a thermophilic bacteria, for a time span ranging from one to four hours. The reaction time may be extended beyond for greater lengths of time if it is desired. Reaction and deproteination may be accomplished by heat inactivating the restriction enzyme followed by phenol-chloroform extraction of the reaction (as described above), or by using a commercially available kit such as the MinElute ${ }^{\circledR}$ Reaction Cleanup Kit from QIAGEN.
[0200] Downstream uses of the insert DNA are discussed in Section V11 above and include: Labeling and use of the fragments as probes to detect the presence of the given gene or the expression of the given gene in a different organism; Use of the fragment in hybridization experiments; PCR amplification of the insert DNA or regions of interest of the insert DNA; Mutagenesis of the particular DNA segment of interest in order to produce substitutions, additions, deletions,
fusions or truncations in the expressed polypeptide, which can be accomplished by random chemical mutagenesis, site directed mutagenesis, error-prone PCR, exonuclease II deletion, oligonucleotide mutagenesis for PCR; Generation of variant forms of the peptide of interest with conservative vs. non-conservative changes in the amino acid sequence to result in the production of novel end-product compounds; Cloning and use of the DNA sequence of interest in a heterologous expression system (yeast, mammalian, insect, plant expression vectors) for the production of the peptide of interest, and the creation of tagged (e.g. His, c-myc, Ni-tagged, etc.) fusion proteins; Use of the peptide that is produced to raise polyclonal or monoclonal antibodies (via the production of hybridomas).
[0201] Antibodies (Ab's) are also used as probes to isolate interacting proteins-Ab's are generated against the peptides resulting from the heterologous expression of the DNA sequence of interest. Proteins that may potentially interact with that encoded by the DNA sequence of interest may also be identified by yeast two-hybird screening as described in U.S. Pat. No. 5,283,173.
[0202] All patents, patent applications, and published references cited herein are hereby incorporated by reference in their entirety. While this invention has been particularly shown and described with references to preferred embodiments thereof, it will be understood by those skilled in the art that various changes in form and details may be made therein without departing from the scope of the invention encompassed by the appended claims.

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$<400>$ SEQUENCE: 5atgacaacgg gacggccggg ggagaaccgg gcgacagacg cggcacgaaa tccggggtgg60
gcegccgggg ggceggcgtc ccagccatgg ggcgggggga acgacgagca ggtcetgcge ..... 120
gagatcctcg gggtcgacgt gcaccgcgag ctgattgact tcgcgggtgg tgccggcgga ..... 180
aatccgcacc tggtcgccga actcgcgcgc gggctcgccg aagagggatt gattcgggag ..... 240
acaaacggtc gggcggaatt ggtgtcccgg cgaattcccc ggcgcgtgct gagttttgtc ..... 300
atgcgtcgat tgaatgatgt cagcgccggc tgccagcagt tcttgaaggt tgccgcggca ..... 360
ttgggcagat cettcatgct ggaggacgtt tcgagaatgc tgggccgatc gtcggcggce ..... 420

$<210>$ SEQ ID NO 6
$<211>$ LENGTH: 895
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain $046-$ ECO11
$<400>$ SEQUENCE: 6




-continued

$<210>$ SEQ ID NO 8
$<211>$ LENGTH: 362
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 8



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<210> SEQ ID NO 9
<211> LENGTH: 1089
<212> TYPE: DNA
<213> ORGANISM: Micromonospora sp. strain 046-ECO11
<400> SEQUENCE: 9
```

atgaccgtcg gatatctcgg gacggtcacc gactcggcgc cogtcgacgc cgcgctgcgc
gtcgccgage tggagagcta cgtcetgcgg ggcggcaage gcatccggcc cgecttcgcc ..... 180
tggctgggct ggatcggcgc cggcggcgac ccggaggacc cggtggcgac cgcggtgctg ..... 240
aacgectgcg cegggttcga getgetgcac gegtceggec tcatccacga cgacatcatc ..... 300
gacgcgtcgc agaccegccg cggccatccc gccgcgcacg tcgcgtacgc cgaacggcat ..... 360
cgggcgcggc gcttctccgg tgacccggga acgttcggca ccggcaccgc catcctgatc ..... 420
ggagacctcg tcctgatctg ggccgacgtc ctggtcogcg cetccggcct gccggccgac ..... 480
gcgcacgtgc gggtctcgce ggtgtggtcg gcggtgcgct cegaggtcat gtacggccag ..... 540
ctgctcgatc tgatcagcca ggtgagccgg agcgaggacg tcgacgcggc gctgcgcatc ..... 600
aaccagtaca agaccgcgtc gtacacggtg gagcggccac tgcagttcgg cgcggcgatc ..... 660
gceggcgegg acgacgacct ettcgcggce taccgcgect tcggcgcega egtgggtatt ..... 720
gccttccagc tgcgcgacga cctgctcggc gtgttcggcg acccggtggt gacgggcaag ..... 780
ccgtccggcg acgacctgcg ggagggcaag cggacggtcc tgctcgccac ggcgctcaag ..... 840
cgcgccgacg aacgggaccc ggacgcggcg gcctacctgc gggcgaaggt cggcacggac ..... 900
ctcgeggacg aggagatcgc cegcatccgc gccatcttcc gegacgtcgg cgeggtcgag ..... 960
gagatcgagc ggcagatctc gcagcgcacc gaccgggcgc tggccgcgct ggaggcgagc ..... 1020
agcgccaccg cccecgcgaa gcatcagctc gccgacatgg cgatcaaggc cacccagcgg ..... 1080
gcccagtga


$<210>$ SEQ ID NO 12
$<211>$ LENGTH: 346
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain $046-$ ECO11
$<400>$ SEQUENCE: 12


$<210>$ SEQ ID NO 13
$<211>$ LENGTH: 1041
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE : 13
atgacgactg accaccggge ggagccgtcc gagccggcge tcgaccggce cgcgaccgec ..... 60
gtggcccatc cgaacatcgc gctgatcaag tactggggca agcgcgacga gcagctgatg ..... 120
atcccgtacg cogacagcet gtcgatgacg ctcgacgtct tcecgaccac caccaccgtc ..... 180
cggatcgaca gcggcgcggc ggcegacgag gtcgtcctcg acggctcgcc cgccgacggc ..... 240
gaacggcgac agcgcgtcgt caccttcctg gacctggtac gcaagctggc cgggcgcacg ..... 300
gaacgggcet gegtcgacac cegcaactcc gtgcccaccg gegceggcct ggcgtcetcg ..... 360
gcgagcggat tcgccgccct cgccctcgcc ggcgcegccg cgtacggcct cgacctggac ..... 420
accaccgcge tgtcccgcct ggcecggegg ggatccgtgt eggcctcccg gtcggtcttc ..... 480
ggcggcttcg cgatgtgcca cgcaggcccc ggcgccggga ccgccgcgga cctcggctco ..... 540
tacgecgage cggtgcecgt egcgcecctc gacgtcgcge tggtgatcge gatcgtcgac ..... 600
gccgggccga aggcggtgtc gagccgcgag gggatgcggc gaaccgtccg gacctccccg ..... 660
ctctatcagt cgtgggtcgc ctceggccgc gecgacctgg cegagatgcg ggcegcgetg ..... 720
ctccagggag acctggacgc ggtcggcgag atcgccgaac gcaacgccct cggcatgcac ..... 780
gccaccatgc tggcegccog gceggeggtg cgctacctgg egcoggtcac tgtcgecgtg ..... 840
ctcgacagcg tgctgcgect gegcgccgac ggcgtctccg cetacgccac gatggacgeg ..... 900

$<210>$ SEQ ID NO 14
$<211>$ LENGTH: 369
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 14


| Arg Asp Gly Ala Asp Ala Ser Gly Gly Asp Leu Ala Ala Ser Val Trp |  |
| ---: | ---: |
| 165 | 170 |

Gly Gly Trp Ile Ala Tyr Gln Ala Pro Asp Arg Ala Ala Val Arg Glu
Met Ala Arg Arg Arg Gly val Glu Glu Thr Met Arg Ala Pro Trp Pro

195
200

| Gly Leu Arg Val Arg Arg |  |
| :--- | :--- |
| 210 | Leu Pro Pro Pro Arg Gly Leu Ala Leu Glu <br> 215 |
| 220 |  |


| Val Gly Trp Thr Gly Glu Pro Ala Ser Ser Ser Ser Leu Thr Gly Arg |  |  |  |
| ---: | ---: | ---: | ---: |
| 225 | 230 | 235 | 240 |

Leu Ala Ala Ser Arg Trp Arg Gly Ser Pro Ala Arg Trp Ser Phe Thr

Ser Arg Ser Gln Glu Cys Val Arg Thr Ala Ile Asp Ala Leu Glu Arg \begin{tabular}{rl}
260 <br>
260

$\quad$

270
\end{tabular}

Gly Asp Asp Gln Glu Leu Leu His Gln Val Arg Arg Ala Arg His Val

| Leu Ala Glu Leu Asp Asp Glu Val Arg Leu Gly Ile Phe Thr Pro Arg |  |
| ---: | :--- |
| 290 | 295 |

Leu Thr Ala Leu Cys Asp Ala Ala Glu Thr Val Gly Gly Ala Ala Lys
305
310 $\quad 315 \quad 320$
Pro Ser Gly Ala Gly Gly Gly Asp Cys Gly Ile Ala Leu Leu Asp Ala

$<210>$ SEQ ID NO 16
$<211>$ LENGTH: 360
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 16
Met Ile Ala Asn Arg Lys Asp Asp His Val Arg Leu Ala Ala Glu Gln
$15010 \quad 15$
Gln Gly Arg Leu Gly Gly His His Glu Phe Asp Asp Val Ser Phe Val$20 \quad 25 \quad 30$
His His Ala Leu Ala Gly Ile Asp Arg Ser Asp Val Ser Leu Ala Thr

| Ser Phe Gly Gly Ile Asp |  |
| :---: | :---: |
| 50 | 55 |
| 50 | Trp Pro Val Pro Leu Cys Ile Asn Ala Met |
| 60 |  |


$<210>$ SEQ ID NO 17
$<211>$ LENGTH: 1083
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE : 17
atgatcgcca accgcaagga cgaccacgtc cggctcgccg cegagcagca gggccggctc

| gccaacgtca | acgccaccge ctccgtcgaa cgggcccggc | gggctgtcga cetgatgcgg | 420 |
| :---: | :---: | :---: | :---: |
| gccgacgcge | tgcagatcca cetgaacacc atccaggaga | cggtgatgcc ggagggggac | 480 |
| cggtcgttcg | ccgcctgggg gccgcggatc gaacagatcg | tcgccggcgt cggtgtgccg | 540 |
| gtgatcgtca | aggaggtcgg cttcgggctc agccgcgaaa | cgctgctgcg gctgcgggac | 600 |
| atgggegtcc | gggtggcega cgtcgccggc cgcggcggca | cgaacttcgc gcgeatcgag | 660 |
| aacgaccggc | gggacgecge cgactactcc ttcetcgacg | ggtggggaca gtcgacaccc | 720 |
| gectgcetgc | tggacgccea gggegtggac ctgcecgtgc | tggectcogg cggcatcogc | 780 |
| aaccegctcg | acgtggtccg cgggctggcg ctcggcgccg | gcgcggcegg ggtgtccgga | 840 |
| ctgttcctgc | gcacgctcct ggacggcgge gtgceggcge | tgctgtcget gctgtccacc | 900 |
| tggetcgacc | agatcgaagc cetgatgace gcectgggcg | cgcggaccec ggcegacctg | 960 |
| accogctgcg | acctgctgat ccagggtcgg etgagcgegt | tetgegcgge coggggeatc | 1020 |
| gacacccacc | gcctcgccac cogttccggc gccacccacg | agatgatcgg aggcattcga | 1080 |
| tga |  |  | 1083 |

$<210\rangle$ SEQ ID NO 18
<211> LENGTH: 351
<212> TYPE: PRT
<213> ORGANISM: Micromonospora sp. strain 046-ECO11
<400> SEQUENCE: 18



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<210> SEQ ID NO 19
<211> LENGTH: 1056
<212> TYPE: DNA
<213> ORGANISM: Micromonospora sp. strain 046-ECO11
<400> SEQUENCE: 19
```

atgaacgacg cgatcgccgg tgtgcccatg aatgggtag gtcccgtgcg gatctcggga 60
aacgtggcgc agatcgagac ggaggttccg ctcgccacgt acgagtcgcc gctctggccg 120
tccgtcggcc ggggcgegaa gatctcccgg atggtcgagg cgggcatcgt cgccacgetc 180
gtcgacgagc gcatgacceg ctcggtgttc gtgegcgcca aggacgcgca gaccgcctac 240
ctggcetcgc ttgaggtcga egcgeggttc gacgaactgc gtgacatcgt gcgcacetgc 300
ggcaggttcg tegagctgat cgggttccac cacgagatca cogcgaacct getgttcetg 360
cggttcagtt tcaccaccgg cgacgcgtcc gggcacaaca tggcgacget ggccgccgac 420
gegctgctga agcacatcct ggacaccatt cegggcatct cgtacggctc gatctcgggc 480
aactactgca cogacaagaa ggccaccgcg ataaacggca ttctcggccg gggcaagaac 540
gtggtcaccg agctggtcgt gccgcgggag atcgtccacg acagcctgca cacgacggcg 600
gcggcgatcg cccagctgaa cgtgcacaag aacatgatcg gcacgttgct cgccggcggt 660
atccgctcgg ccaacgccca ctacgcgaac atgctgctcg ggttctacct ggccacgggt 720
caggacgceg cgaacatcgt cgagggctcc cagggegtga eggtcgccga ggaccgcgac 780
ggcgacctct acttctcctg cacgctgcce aacctgatcg tgggcaccgt cggcaacggc 840
aaggggctcg gettcgtcga ggagaacctg gagcggctcg getgcegcgc ctcgcgtgat 900
ccgggcgaga acgcccggcg gctcgcggtc atcgcggccg cgacggtgct ctgcggcgag 960
ctgtcectgc tegcegcgca gaccaacceg ggcgagetga tgegggcgea cgtccggetc 1020
gaacgcccga ccgagaccac gaagatcgga gcctga 1056

```
<210> SEQ ID NO 20
<211> LENGTH: 391
<212> TYPE: PRT
<213> ORGANISM: Micromonospora sp. strain 046-ECO11
<400> SEQUENCE: 20
```



$<210>$ SEQ ID NO 22
$<211>$ LENGTH: 290
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 22


$<210>$ SEQ ID NO 23
$<211>$ LENGTH: 873
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 23
gtggccgagc tctactcgac catcgaggaa tcggcccggc aactggacgt gccgtgttcg
agccacggca tcgacttcgg ggtggtcggc ggcttcaaga agatctacgc ggcettcgce 360
ceggacgagc tgcaggtggc cacgtcgctc gccggcattc eggcgatgcc cegcagcetc ..... 420
gcegcgaacg cegacttctt cacccggcac ggcetcgacg accgggtcgg cgtgctggga ..... 480
ttcgactacc cggcccggac cgtgaacgtc tacttcaacg acgtgccgcg tgagtgcttc ..... 540
gagccggaga ccatccggtc gacgctgcgc cggaccggga tggccgagcc gagcgagcag ..... 600
atgctccggc tcggcaccgg ggcgttcggg ctctacgtca cgctgggetg ggactccceg ..... 660
gagatcgagc ggatctgcta cgccgcggcg accacggacc tgaccacgct tccggtacco ..... 720
gtggaaccgg agatcgagaa gttcgtgaaa agcgttccgt acggcggcgg ggaccggaag ..... 780
ttcgtctacg gcgtggcgct gacccccaag ggggagtact acaaactcga gtcgcactac ..... 840
aatggaagc cgggcgeggt gaacttcatt tga ..... 87.3
$<210>$ SEQ ID NO 24
$<211>$ LENGTH: 370
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain $046-$ ECO11
$<400>$ SEQUENCE: 24


| Ala Ala Ser Arg Arg Ala Pro Ile Ala Val Leu Val Ala Thr Gly Leu |  |
| :---: | :---: |
| 50 | 55 |
| 60 |  |


| Cys Val Val Gly Tyr Asn Ala Ile Gly Phe Gly Val Pro Ala |  |  |  |
| :--- | :---: | :---: | :---: | :---: |
| 65 | 70 | 75 | 80 |




| Val Gln Ala Glu Val Ala Val |  |
| ---: | ---: | ---: |
| 195 | 200 |

Val Pro Glu Ser Leu Leu Ala Ile Gln Glu Ala Gly Arg Ala Ala Thr
210

215
225230235240
Pro Ser His Gly Leu Asp His Leu Pro Glu Leu Leu Ala Gly Ala Glu


Thr Ala Arg Pro Gly Ala Ala Pro Val Pro Gly Val Gly Leu Leu Gly | 330 |
| ---: |
| 325 |

Met His Glu Arg Val Leu Ala Leu Gly Gly Arg Leu Arg Ala Glu Pro
Arg Thr Gly Gly Gly Phe Thr Val Gln Ala Glu Leu Pro Val Val Arg

$<210>$ SEQ ID NO 26
$<211>$ LENGTH: 220
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 26


$<210>$ SEQ ID NO 28
$<211>$ LENGTH: 131
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. Strain 046-ECO11
$<400>$ SEQUENCE: 28


$<210>$ SEQ ID NO 29
$<211>$ LENGTH: 396
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Micromonospora sp. Strain 046-ECO11
$<400>$ SEQUENCE: 29
atgttcatcc gtcgtttgct caccgccgcc gcagccggcg tcctcggtgg gctcgcactc ..... 60
gtcgcaccgg cggccgcgca ggtgacggcc gccgacggtg acggtggttc cggccgcgcc ..... 120
ggatccgtgc tggcgetcgc getcgcgttg ctcggcctcg tcctgggcgg gtgggcgttg ..... 180
cgctccgcgg ggcgeggcgg eggtcgtggc aacgcgatcg cegcgetggt getcgeggtg ..... 240
gceggcetga tcgceggegt ggtcgcectg gccggctcog acggtggtgt cggcagcggc ..... 300
aacggcegtg gtggcgceat cgtggccgtc gtgctggcge tgatcgggat cgccgtcggc ..... 360
ggcetggcat tcaccegctc ceggcgcgec gectga ..... 396

<210> SEQ ID NO 30

<211> LENGTH: 15

<212> TYPE: PRT

$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11

<400> SEQUENCE: 30


| <211> LENGTH: 465 |  |
| :---: | :---: |
| <212> TYPE: DNA |  |
| <213> ORGANISM: Micromonospora sp. strain 046-ECO11 |  |
| <400> SEQUENCE: 31 |  |
| atgcgcaaag tgttcgcegg actggcagcg ttcetgctgc tegtgctcgt ggtgcagttc 60 |  |
| ttcctggceg ccagcggcge gttcagcaac gaggceaacg aggaggcgtt cogccotcac 120 |  |
| cggatcetgg gectggggag catcctcgte gecgtggtge tgacggtggc cgcegcggtg 180 |  |
| atgcggatgc ceggceggat catcggcetg tecggcetgg tegccgggct gggcatcctg 240 |  |
| caggcectga tegcggtcat egceaaggeg ttcggcgact eggceggtga ctcggcegtc 300 |  |
| ggceggtacg tgttcggcet gcacgcggtc aacggactgg tgatggtggc cgtcgcecge 360 |  |
| gtcatcctgc gcagcgtcog ggcggcgecg gacacgacca ceacgccogg egtggacacg 420 |  |
| acggtcaccg gtccggcgge cgactcggcg cgaacggcgt catga | 465 |

$<210>$ SEQ ID NO 32
$<211>$ LENGTH: 661
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 32


Thr Ser His Ala His ..... 660
<210> SEQ ID NO 33 <211> LENGTH: 1986

<212> TYPE: DNA

<213> ORGANISM: Micromonospora sp. strain 046-ECO11

<400> SEQUENCE: 33
atgagcacgc tccaatggat cctcgtggac cacgtcgtgg egctgctcgg tgtcgcgacg 60
tggttcgcaa cgggtgtcac ggcagctctc ggccgccacc ggatcgcgtt ggcgctcctc 120
ggcgccgcgg tgctggtgac agtcgccegc ctgggcaccg tggcgctgct ggccgaccgc 180
ggctggtggt tcgtccagga gaaggttctg ctggggctgc cgatgctcgg cgccgcgggg 240
ctcgtcgcgg tgctcctggc cggcccgcgc ctgctcgcgg cecggcagtc accggcggcg 300
gacctgccgg ceggegcgct ggtcgcggtg ctgaccgccg gcttcgccgc gctggccggc 360
ctggtggtga cgttcaccgc cgggtacccg ctgacgtgga gcaccgcgct gatcgccgtc 420
gccetcgtct gegcegccgc getgetcacc gegcgggtgg teggacgacc egccgccccg 480
gcegeggagg coggctccec ggagcacacg ceggeggegg cogggcccac ggcgetgtcc 540
cgccgceggt tcetcggcgt ggceggggga gtggtcgcgg cgggcgccgg cgccaccggc 600
gtcggcetgc tettcegcga cecggaggeg atggtcaccg gaggeggccc cggacacgcc 660
ggtggcgecc gccceaaggt ctccgtggeg gacctgcgeg gccecggcge tccggcggeg 720
ggcggcacgg cgcgacgcea cgtgctcacc gcceggacgg gcaccgtcac gattccgtcc 780
ggacgtccga tcgacgcctg gagctacgag ggccgcctgc cogggccggc catcaccgeg 840
accgagggcg acctgatcga ggtgacgetc cgcaacgceg acatcgagga cggcgtcacc 900
gtgcactggc acgggtacga egtgcegtgc ggcgaggacg gcgcgccggg cgccacgcag 960
cacgeggtgc agcecggcgg egagttcgtc taccggttcc aggcggacca ggtggggacg 1020
tactggtacc acacccacca ggcgtcgcac cccgccgtgc gcaaagggct gtacgggacg 1080
ctcgtcgtga cgccgcgcga ggaccggccg gaagcggagc gcgggctgga cctgacgctg 1140
coggtgcaca cgttcgacga cgtcacgatc ctcggcgacc aggagggacg cgccgtccac 1200
gacgtccgcc ccggccagce ggtgcgactg cgtctgatca acaccgactc caacccgcac 1260
tggttcgccg tegtcggctc geccttccge gtggtggccg tegacggccg cgacctcaac 1320
cagcegggcg aggtacgega ggtcgggctc cgcetgcceg coggaggceg gtacgacctg 1380
accetggcea tgceggacge caaggtcacg ctgctgctcg acaacgactc cgaccaggge 1440
gtcetgctgc gccegceggg egtcggcggt ggtgaccgce cgetgccgga caccgccgac 1500
tggcecgagt tcgacctgct gggctacggc gagccggcge cegtgccgtt cgacgccgac 1560
gacgecgacc gecacttcac catcgtcctc gaccgggcec tggccatggt cgacggcaag 1620
cecgegtacg cecagaccgt egacggtcge gcacatccct cegtceccga ccagctcgtc 1680
cgggaggggg acgtcgtgcg cttcacggtg gtcaaccgga gcctcgaaac ccaccogtgg 1740
cacctgcacg gccatceggt getgatcctg tccegcgacg gecggecgta ctccggcagc 1800
cegctgtgga tggacacctt egacgtgegg cegggagagg tgtgggaggt ggcgttcegg 1860
gcggacaatc cgggtgtctg gatgaaccac tgccacaacc tgccgcacca ggagcagggc 1920

| atgatgctgc ggctegtcta cgacggtgtc accacgcect tegceagcac gagceacgea | 1980 |
| :---: | :---: |
| cactga | 1986 |
| <210> SEQ ID NO 34 |  |
| <211> LENGTH: 129 |  |
| <212> TYPE: PRT |  |
| <213> ORGANISM: Micromonospora sp. strain 046-ECO11 |  |
| <400> SEQUENCE: 34 |  |
|  |  |
|  |  |
| Val Ser Thr Ala Phe Pro Pro Ala Phe Ala Asp Val Val Arg Gly Pro 35 40 |  |
| Leu Arg Leu Leu Leu Ser Gly Pro Thr Ser Ser Gly Ala Arg Val Thr $50 \quad 5560$ |  |
| Pro Ala Asp Ala Ala Gly Cys Gly Arg Asn Arg Ile His Leu Ile Val 65 70 |  |
| Asp Asp Leu Asp Ala Glu Arg Glu Arg Leu Glu Arg Ala Gly Val Thr 85 |  |
| Leu Arg Ser Asp Val Val Ala Gly Pro Gly Gly Arg Gln Phe Leu Ile 100 105 |  |
| Ala Asp Pro Ala Gly Asn Leu Val Glu Val Phe Glu Pro Ala Ala Arg 115120125 |  |
| Gly |  |
| <210> SEQ ID NO 35 |  |
| <211> LENGTH: 390 |  |
| <212> TYPE: DNA |  |
| <213> ORGANISM: Micromonospora sp. strain 046-ECO11 |  |
| <400> SEQUENCE : 35 |  |
| atgaccgcag acctgcacgg cetggccagc gtccgctaca tcgtcgacga cgtgtcggcg | 60 |
| gcgatcgagt tctacaccac ccacctgggt ttcacggtgt cgaccgcgtt cccgccggce | 120 |
| ttcgcegacg tggtgcgcgg gccgctgcgg ctcctgctgt ccgggccgac cagctcggge | 180 |
| gccegggtca ccceggcgga cgcggccggg tgcgggcgca accgcatcca cetgatcgtc | 240 |
| gacgatctcg acgccgaacg ggagcggctg gagcgcgccg gggtgacgtt gcgcagcgac | 300 |
| gtcgtggccg ggcegggcgg cegtcagttc ctgatcgccg accoggcggg caacctggtc | 360 |
| gaggtgttcg agceggcage cegcggctga | 390 |

$<210>$ SEQ ID NO 36
$<211>$ LENGTH: 178
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain $046-$ ECO11
$<400>$ SEQUENCE : 36


$<210>$ SEQ ID NO 38
$<211>$ LENGTH: 661
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 38



$<210>$ SEQ ID NO 39
$<211>$ LENGTH: 1986
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 39
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ttcgggctcg accgcggaca ggcgctggcg ggcetgcccg aacgcggtgt gccggtgacc 360
gacgccgact tegacctcag egacgaggac tacgccgcga tegtcaagcg ggtggtgggt 420
gacgagatcg ggctgggcgc eggatccaac ttcgtcatcc ggcgcacctt caccgcgegg $\quad 480$
ctggccgact actcgatcge cacggaactg gegctcttcc gecggttgct gaccggcgaa 540
ctgggttcct actggacgtt tetgttccac tecggcgccg gcacgttcat eggcgcgtca 600
ccggaacgac acgtcagcat gatcgacgga accgtctcga tgaatcccat cagcgggacc 660
taccggcacc ceccgaacgg cecggecgtt tccggtctgc tggaattcct gaacgacceg 720
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ccggacggcc cggacggcgg gcggaccacg gccgcgcggg ctcgttcgtc cctggccacc ..... 1260
gacccccggg tacggcgggc gttgcgcgag cgcaacacca cactgtcgag gttctggctc ..... 1320
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ggtcecggce coggegacce gggegacctg accgaccoge gtatgeggac cetgcgeggg ..... 1560
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gtgctcgceg cegaactggg gttccccctc gcceggegcg cggtgcccaa ccagggtgtg ..... 1680
cagaagcgga tcgacctgtt cggccggccg gaactcgtgg ggttctacaa cacctacacc ..... 1740
gcccgctccg cgcacgacgt ggtggccggt ggccggcggg gcccgatcga gatcagccgc ..... 1800
agcecggaca gcggggacgt gcacgcgctg cgcggcccgg gattccgttc cgtccagttc ..... 1860
cacctggagt cegtcctcac ccagcacggc ccacggatcc tgggcgacct gctggtctcc ..... 1920
ctgctcgccg acggcacggc cgccgccgcg gccgaggcgg cgggccggcg cgggaaccgc ..... 1980
cegtga ..... 1986
$<210>$ SEQ ID NO 40
$<211>$ LENGTH: 427
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE : 40


$<210>$ SEQ ID NO 41
$<211>$ LENGTH: 1284
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 41
gtgaagacga ctgtggacgt getggtccag aaatacgggg gcacctcgct gcagaccetc
ggggcggcgc gggtggaagc ggcgctgggc cgtggcgagg tcgccgtggt caccggattc ..... 420
cagggcatcg accgggccgg tgacgtcgcc acgctgggge gcggcggctc cgacacgaca ..... 480
geggtggcge tegcggcecg gctccgcgeg tcggcgtgeg agatctacac egacgtggac ..... 540

| cocggegtca | tggcggagat | ggcgttcgcc ggcgcgcggg | tectgcacac cogatgcatc | 660 |
| :---: | :---: | :---: | :---: | :---: |
| gagctggceg | ccatggaagg | ggtcgaagtg egcgtgcgca | acgegtcgtc gcaggcgcec | 720 |
| ggaacgatag | tcgtggaccg | gccegacgac eggcegctgg | agacccggcg ggcegtggtg | 780 |
| geggtcaccc | acgacaccga | tgtcgtccgc gtgctggtgc | actgccgcga cggccgccgg | 840 |
| gacatggcac | ccgacgtgtt | cgaggtgctg gccgcecatg | gggcggtggc ggacctggtg | 900 |
| gcecggtceg | ggcectacga | gagcgagtte eggatggggt | tcaccatcog cogcagccag | 960 |
| gcegaagcgg | tgcggaccgc | gctgcacgac ctcaccgcgt | cettcgacgg eggggtccac | 1020 |
| ttcgacgaga | acgtcggcaa | ggtgtccgtg gtcggcatgg | gcetgctcag cegccecgag | 1080 |
| cacacggccc | ggctgatggc | ggcgctggce gcggcgggga | tctcgacgag ctggatctcc | 1140 |
| acctcccaga | tgcggctgtc | ggtgatcgtg tcgcgggacc | gcaccgtcga cgcegtcgaa | 1200 |
| gccetgcacc | gcgcgttccg | cctggaccgg tccgagccgg | cggacgccac gtccctgacc | 1260 |
| tcccgcegtt | ccgccaccgc | ctga |  | 1284 |

$<210>$ SEQ ID NO 42
$<211>$ LENGTH: 274
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 42

Glu Gln Pro Gly Leu Met Ala Ala Ala Val Ala Arg Leu Val His Glu
245
250
$<210>$ SEQ ID NO 43
$<211>$ LENGTH: 825
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 43

$<210>$ SEQ ID NO 44
$<211>$ LENGTH: 367
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain $046-$ ECO11
$<400>$ SEQUENCE: 44



$<210>$ SEQ ID NO 46
$<211>$ LENGTH: 253
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain $046-$ ECO11
$<400>$ SEQUENCE: 46


[^0]
$<210\rangle$ SEQ ID NO 48
<211> LENGTH: 438
<212> TYPE: PRT
<213> ORGANISM: Micromonospora sp. strain 046-ECO11
<400> SEQUENCE: 48



$<210>$ SEQ ID NO 50
$<211>$ LENGTH: 396
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 50


| 245250255 |  |
| :---: | :---: |
| Thr Arg Leu Ile Ala Ala Ala Asp Arg Val Gly Arg Trp Ala Leu His 260 $\quad \begin{array}{r}270\end{array}$ |  |
| Asp Arg Asp 275 |  |
| Gly Asp Ala Ala His Pro Met Leu Pro Phe Gln Ala Gln Gly Ala Asn 290295300 |  |
| Gln Ala Val Glu Asp Ala Val Val Leu Ala Val Cys Leu Ala Gly Val    <br> 305 310 315 320 |  |
| Glu Pro Ala Gly Leu Gly Ala Ala Leu Arg Arg Tyr Glu Arg Ile Arg $\begin{array}{r}325 \\ 330\end{array}$ |  |
| $\begin{array}{rl}\text { Leu Pro Arg Thr Thr Arg Ile Gln Arg Gln Ser Arg Ala Asn Ala Glu } \\ 340 & 345 \\ 350\end{array}$ |  |
| Met Phe His Leu Ala Asp Gly Ala Asp Gln Arg Arg Arg Asp Val Ala $\begin{array}{r}365 \\ 355\end{array}$ |  |
| Ala Gln Ser Ser Ser Gly Leu Asp Arg His Glu Trp Leu Phe Gly Tyr 370375380 |  |
| Asp Ala Glu Lys Ala Thr Thr Thr Ser Gly Ser Ala 385 390 |  |
| <210> SEQ ID NO 51 |  |
| <211> LENGTH: 1191 |  |
| <212> TYPE: DNA |  |
| <213> ORGANISM: Micromonospora sp. strain 046-ECO11 |  |
| <400> SEQUENCE: 51 |  |
| atgccgcaga tgagggtcge cgtggccggc gccggcatcg ccgggctcgc cttcgccgce | 60 |
| gcectgcgec ggaccgggat cgactgccac gtgtacgaac aggccgacca gctcatggag | 120 |
| gtgggegcgg gcgtgcaggt cgcgcegaac gccacccggc tgctgcaccg gctgggcetg | 180 |
| cgtgaccgce tgcgtacggt ggctgtcgeg cogcaggcga togagatgcg cogctgggac | 240 |
| gacggcacge tgctgcaacg cacceagctg ggcagcgtgt gcggacgecg cttcggcgeg | 300 |
| ccgtactacg tggtgcaccg cgcggacctg cacagcagce tgctgtcgct ggtgcegceg | 360 |
| gaccgggtgc acctgggcge cogcetcacc gcegtgacge agaccgecga cgaggegtac | 420 |
| ctgcacctgt ccaacggcac cacggtcgcg gcggatctcg tegtgggcge cgacggcatc | 480 |
| cacteggtcg cgcgggagca gatcgtggcg gaccggcegc gcttctccgg acagtccatc | 540 |
| taccgcggge tggtgceggc egagcgggtg ecgttcctgc teaccgaacc cogggtgcag | 600 |
| ttgtggttcg ggceggacca gcactgcgte tgctacccgg tgtccgcogg coggcaggtg | 660 |
| agcttcggcg egacggtgce cgceaccgac tggeggcagg agtcgtggtc gggcegggge | 720 |
| gacgtgacgc aactcgcggc egcgtacgcg ggctggcace cggacgtcac coggctgate | 780 |
| gccgcggccg accgggtcgg caggtgggcg ctgcacgacc gggacagcat cgaccggctc | 840 |
| agcgegggac gggtgacect gatcggcgac gcegcgcace cgatgctgce gttccaggeg | 900 |
| cagggcgcga accaggccgt cgaggacgcg gtggtgctcg cggtctgcct ggceggcgtg | 960 |
| gaaccggcgg gcetgggcge cgegetgcge cgetacgaac ggatcegcet gceccggace | 1020 |
| acccggatcc agcggcagtc cogggccaac gccgagatgt tccacctggc cgacggcgec | 1080 |
| gaccagcgec gccgggacgt cgcogcacaa tcctcgtcog gcctggaccg ccacgaatgg | 1140 |
| ctettcgggt acgacgecga gaaagceacc acgaccagcg ggagcgectg a | 1191 |


$<210>$ SEQ ID NO 53
$<211>$ LENGTH: 786
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 5360
ggcgccgccg tggccggtgt cctggcgagg gcgggcgcgc aggtggcggc ggtggaccgc ..... 120
aacgccgagg egctgaccac egtcgtgacg aagctcgceg cegagggega ctcggcgegc

$<210>$ SEQ ID NO 54
$<211>$ LENGTH: 224
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 54


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<210> SEQ ID NO 55
<211> LENGTH: 675
<212> TYPE: DNA
<213> ORGANISM: Micromonospora sp. strain 046-ECO11
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$<210>$ SEQ ID NO 56
$<211>$ LENGTH: 233
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 56


| Asn Ile Asp Gly Gly Asp Val Leu Ile 225230 |  |
| :---: | :---: |
| <210> SEQ ID NO 57 |  |
| <211> LENGTH: 702 |  |
| <212> TYPE: DNA |  |
| <213> ORGANISM: Micromonospora sp. strain 046-ECO11 |  |
| <400> SEQUENCE: 57 |  |
| atgtcggatc ggaccegggt egtggtcgtc ggcggaacct cggggatcgg gcggcacttc | 60 |
| gccegattct gegcegaacg eggagacgac gtggtgatca coggcegttc ggcggcecgg | 120 |
| accaagaccg tggcggacga gatcggcggg cggacccgtg ggctcgctct cgacctggce | 180 |
| gagceggaga cgatcgcgga cgcgctcgcc gacgtgcogc acgtcgaccg gctcgtggtc | 240 |
| gcggcgctgg accgcgacta caacaccgtc cgcgcgtacc ggcegggcga cgcggcgegg | 300 |
| ctgctgaccg tcaagctggt cggctacacg gcggtcctgc acgccetcgc cocgcggatg | 360 |
| accgacgaga gegcagtcgt gctgctcggc ggcetggcea gccaccggce gtatcocggc | 420 |
| tccacctccg tcacgaccgc caacggcggg atcagcgcgc tggtgcggac cotggctgtg | 480 |
| gaactctcgc eggtccgggt caacgccctg caccogagca tegtctccga cacgcogttc | 540 |
| tggagcgaca agcecgcege gegggaggce gcegcgacec gegcgetcag cegacggecg | 600 |
| gtcaccatgc aggactgcgc cgaggcgatc gacttcctgc tgacgaaccg ctcgataaac | 660 |
| ggggtcaacc tgaacatcga cggcggggac gtgctcatct ga | 702 |

$<210>$ SEQ ID NO 58
$<211>$ LENGTH: 246
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 58


$<210>$ SEQ ID NO 59
$<211>$ LENGTH: 741
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 59

$<210>$ SEQ ID NO 60
$<211>$ LENGTH: 111
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 60

$<210>$ SEQ ID NO 61
$<211>$ LENGTH: 336
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 61
atggacacgg cagctccggc aacggacgge ggtcgctace tcgccgtcca tcacagcgca 60
gagttcaggg aactacggcg acgatcgagc acgttcacgc tctgggccag cgtcgccttc 120
ttcggctggt ggttcctcgg cagcctgctc gccacctacg egccggactt cttccgggag 180
aaggtggccg gcccggtcaa cgtgggtctg ctcttcgtct tcctgtcgtt cgccttcgtg 240
gtgacgctcg cegcettcta cetgcgttac geccgcacge atctcgatcc gctcagcgag 300
aagatccgtg cogacctgga aggagcgtcc cgatga 336
$<210>$ SEQ ID NO 62
$<211>$ LENGTH: 559
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain $046-$ ECO11
$<400>$ SEQUENCE: 62


$<210>$ SEQ ID NO 63
$<211>$ LENGTH: 1680
$<212>$ TYPE: DNA.
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE : 63
atgagcgtca tcctcgecga cccgccaccc ccggtcgaca acacgtgggc gacgccegcg
accatgctcg cgctgttcct cttcgccggg ccggtgcgca acgtgggcgg ctacacgctc ..... 360
ggtgacctgc tcgcggtccg tacccgggag cggccggcge ggatcgcgtc ggcggtgctc ..... 420
acgctgctga cgtacgtcat gctgacggtg atcatgatgg ccgccatcgc gttcatcttc ..... 480
aaccgctggt tcggcgtcga cgccetcgtc ggcetggtcc tcccggtgtt cgtcgtcggt ..... 540
ctgatcacgg tggggtacgt gtacctcggc gggatgctcg gggtcacccg catcctggtg ..... 600
ttcaagctgg tgctgtcggt ggtcgtcgtg ggcgtgctga ccgcctgggt gctggccogc ..... 660
ttcgacctga acctcttcag cetgctggag cgggcegagg cgaacgcggc gccggtgcce ..... 720
agcggcagcg acctgctggg cccgggccgg ctgttcggcg agggcgcgac cacgctcgtg ..... 780
cacctgtcga agctgttcgc catcgccgtc ggagtggcgg ccattccgtt cetgttcatg ..... 840
cgcaacttcg cggtgaccag cgggcgggac gcgcgccggt cgaccgggtg ggcgtcgatg ..... 900
atcatcgtcg ggttctacct gtgcetgtce gtcgtcggge teggtgecgt egcgatcctc ..... 960
ggccgggaca acatcggcgt catcaaggcc caccgcgaca tcagcttccc caagctcgcc ..... 1020
gacgagctcg gcggtccggt gatggtcggc tccctggcog gcgtcgcggt cctgacgatc ..... 1080
gtcggcgtct tegcgccget getgcacage gecgtgacga cggtgaccaa ggacctgaac ..... 1140
gtgatccgcg gccggcgget ggatccggec gcegagctge gggacatcaa gcgcaacacc ..... 1200
ctgatcatcg gcgtcggctc cgtgctgctg gcggtcgtga tgctgccggt acggacccac ..... 1260
atcttcatcc egacetcgat cgacattgce ggcgeggtgg tcetgccgat cgtcgtctac ..... 1320
gcgttgttct ggcggcgttt caacacccgc ggactgcagt ggacggtcta cggcggcctc ..... 1380
gcgctcaccg cgttcctggt gctgttctcc aacggtgtct cgggcgagcc ggacgccatc ..... 1440
ttcceggacc gcaacttcaa gttegtggac gtcgagcccg cgetgatcac ggtgceggtc ..... 1500
ggettcetgc teggetacct eggetcgatc accagccggg agcgcgacga cgecgegttc ..... 1560
gccgagatge aggtceggtc cetcaccgga getgtegtca egggaccgce geggceggec ..... 1620
gccgtggacg acqaggaccg cqacggccgc caggaccggg cqcccagccc gqtgagctga ..... 1680
<210> SEQ ID NO 64

<211> LENGTH: 5960

<212> TYPE: DNA

$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11

<400> SEQUENCE: 64
ccacacccct egggaggcaa ctgtggatcc ggtaccggtt ctggtcgtgg gcgcgggcce ..... 60
ggtcggcatg gtcaccgcgc tggcgctcgc cegtcacggc gtcgcetgcg tcetcgtcga ..... 120
ccagggettc gagacgtcgg tccatcccaa getggactac gtcaacgccc gcagcatgga ..... 180
gttcctccgc cagttcggcc tcgccgacga cgtccgtgce gccggcgtcg cgcccgagca ..... 240
ccgggccgac gtcatctggt cgaccggcct ggccggtgag ccgatcacca ggtgggggct ..... 300
gccetcggtg acgcaggagt ggcgccgcat cgcegagcac aacgacggca cccagccggc ..... 360
cgagcccggc cagcggatct cccagatcga cctggaaccg gtcctgcggg cccgctgccg ..... 420
gcgggagccc cttgtcgacc tgcgcctcgg cgtacggttc gactcgctga cccaggacga ..... 480
cgcgggggtc accagcgtcc tcgccgacga caccggcggc gaggtccggg tgcggtcgga ..... 540
gtacgtggtc gggtgcgacg gcgcgtcgag ccaggtccge cgggcegtgg gcatcggtga ..... 600
ggaggggttc gacgtgcccg gcctgccggg cgccttcatg gtgcacttca ccagccggga ..... 660




$<210>$ SEQ ID NO 66
$<211>$ LENGTH: 1599
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 66


| accggcetgg | ccggtgagce gatcaccagg tgggggctgc | tcggtgac gcaggagtgg | 300 |
| :---: | :---: | :---: | :---: |
| cgecgcatcg | ccgagcacaa cgacggcacc cagccggceg | agcecggcea geggatctcc | 360 |
| cagatcgacc | tggaaccggt cetgcgggce cgctgccggc | gggagccoct tgtcgacctg | 420 |
| cgcctcggcg | tacggttcga ctcgetgacc caggacgacg | cgggggtcac cagcgtcctc | 480 |
| gccgacgaca | ccggcggcga ggtccgggtg cggtcggagt | acgtggtcgg gtgcgacggc | 540 |
| gcgtcgagce | aggtccgccg ggcegtggge atcggtgagg | aggggttcga cgtgccoggc | 600 |
| ctgcegggeg | cottcatggt gcacttcacc agcogggacc | tggacagcet gcaccggcac | 660 |
| ggceggttct | ggcactactt cgcgttccgg tacgtgatca | tcgeccagga cgaggtcgac | 720 |
| acctggaccg | cgcacgtcaa cggcgtcgac cogaacgagt | togacgagce gceggecgac | 780 |
| ceggaggcgt | tcctgctcga cacgatccgc accgagctgc | ggatcgacaa ggtgctgctc | 840 |
| acctcgeget | ggcgtcccgg cttcatgctc gcegacaggt | accgcgcegg cegggtgctg | 900 |
| ctegccggtg | actcggceca ceggatgttc cecaccggcg | cgtacggcat gaacaccggc | 960 |
| atcggcgacg | ccgtcgacgt ggcetggaag ctggccgctg | tcgtccgggg cttcggcggc | 1020 |
| ccegggctgc | tegacagcta cgacgccgaa egcegccogg | tggggcggcg caacatgcge | 1080 |
| acctcgcacc | ggcacctggg cgtgcacctg cgggcgggcg | agctcctgcg cggcggcgec | 1140 |
| ccgctgcegt | ccgtcgcgge cttcctcgac gccgagcggg | gcgagaacga gtaccggggg | 1200 |
| atcgagctcg | gctaccgcta ctccggcteg ceggtgctct | ggceggaggg cecgggggag | 1260 |
| ccotcggacg | accogcggge gtacgecccg acgacctggc | ccggcgcecg tccgcccagc | 1320 |
| ctcctgctga | gcgacgggca gcagatcttc gaccggttcg | accoggcetc gttcaccotc | 1380 |
| gtggacttca | ccggtgacgg cgcegccggt cegctgctgg | cggcggcgge cgcgcggggg | 1440 |
| ctcecggtca | cccacaccgt ggtgaccgac ceccgggetc | gtgagctgtg ggaacgcgac | 1500 |
| ctegtectgc | tgcggcegga ccaccacgtc gcetggcggg | gaaacaccgt gccgcoggac | 1560 |
| ccogacgecg | tggtccagcg egtgcggggt ggcggatag |  | 1599 |

$<210>$ SEQ ID NO 67
$<211>$ LENGTH: 423
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 67


$<210>$ SEQ ID NO 68
$<211>$ LENGTH: 1272
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 68
atgcagcaat ceggttcaac ggcggaacgc agcccactcg ggcegtggga gggcatgecg 60
geggtccagc aaccggactg gcaggaccac ccggcgtacg eggagacctg tcaggegttg 120
gcgtcggccc egcegctggt cccacccggg gaggtacggg ggttccggca gctgttgtcg 180
gagctggcgt egaccgacgg gctcctgctg cagttgggcg actgcgccga gagcctctac 240

| gagtgcaccc | ccoggcacac ctcggacaag | atcgaggtca | tcgaccgget | gggggaccgg | 300 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ctcagegage | tcaccgggeg caacgtgetg | cgggtgggcc | ggatggcegg | gcagttcgec | 360 |
| aagceccggt | cgcaggcgac ggagtggcac | gacgegctga | gcatcccetc | cttccgcggc | 420 |
| cacatgatca | attcogagct ggcegcgecc | ggtacgcgca | aggccgaccc | tcgccgcatg | 480 |
| tggtgggcgt | acgaggcgag egaccgggtg | cagcgggtcc | tgcgegceca | ccgggagggc | 540 |
| aaccggcgtg | cogcgeggac egaggggecg | tggtcgagce | acgaggcect | ggtcgtegac | 600 |
| tacgagtccc | gectgatcog cegggacecg | gacacgggeg | agcactacct | ggcgtcgacc | 660 |
| cacctgccgt | gggtggggga gcggacccgc | cggtcegceg | aggcgcacgt | ggceatgctg | 720 |
| tccacggtgg | tgaaccoggt cggctgcaag | atcgggccgg | acgccgaccc | ggacgacgtc | 780 |
| ctgcgggtgt | gcgaggcget cgacccgegg | cgcgatccgg | gccgtctcgt | cotgatcceg | 840 |
| cggatgggcc | gggaccggat ccgggagtcc | ctgccgcoga | tcgtccgcgc | ggtggtgaac | 900 |
| gcggggcacc | ccgtgctctg gctgagcgat | cccatgcacg | gcaacaccgt | caaggcctcg | 960 |
| gtcggectga | agacgegcca cctctccgac | gtggtcaccg | aggegctgtg | gttccgcgac | 1020 |
| atcctcgacc | agcagcggca gcacgecgcc | gggctgcaca | tcgaggtcgc | cgecaccgac | 1080 |
| gtgaccgagt | gcgtcggcgg ttcggtggce | ggcgaggagg | acctggcgeg | gcactacacc | 1140 |
| tcgctgtgcg | accegcgget caaccogggt | caggceaccg | agctgatcga | agcgtgggec | 1200 |
| aaggacaccg | cgacggtcgg cecgggaccg | cggcgetccg | gccettcggc | gcggceggag | 1260 |
| gtcgecgcet | ga |  |  |  | 1272 |

$<210>$ SEQ ID NO 69
$<211>$ LENGTH: 340
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 69


<210> SEQ ID NO 70
<211> LENGTH: 1023
<212> TYPE: DNA
<213> ORGANISM: Micromonospora sp. strain 046-ECO11
<400> SEQUENCE: 70
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cccggtgact ccccgcggct ggcgggcgag aacgtcgagc acatccggct gctggccgcg 180
atgcacgacc tcccgccgat cctggtgcaa cgcggcacga tgcgggtgat cgacggcatg 240
caccggctgc gggccgccaa gctgcgcggc gacgagaccg tgcgggtgac gttcttcgac 300
ggggacgacg cegcggcgtt cetgctctcg gtcgacgcca acatcaaaca cgggctgccg 360
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tcggaccgeg cegtcgccge ggcggccggg ctgtcaccga ccacggcgag cggcatccgg 480
cgccgcetge tgcaaccgge ggcgegggag ggcagceggg tgggacggga cgggcgggtg 540
cgccegctgg acggctcgge gggcegacgg cgggceagcg cggtcatcgc gctccggceg 600
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gtgegcgcec ggttgcagge gggcegggac cecgtcetga cetcgcagcg accggeggec 720
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gcggcetggc gegagttcgt egacgccgtg cegcegtact ggcgcaaatc ggtggcegag 960
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$<210>$ SEQ ID NO 72
$<211>$ LENGTH: 1482
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 72
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ttcgacctcg acaacgggaa egccetgacc gacgtcatct acceggccet caacaccgag 180
ccgcgggtcg agtacagcgg coggcecggg tcetgggceg cggaccgcgc catgctcatc 240
gaactgccgt ggttcgacge cetggeggeg taccacccca cogeggtcgg catcttctcc 300
accatcggce gecgtccegc egaggagcac acgacgegca acaagaacat cgccgtcatc 360
tactcggcet acacctcgct cagcaagctc tacccccagc acgaggcgac ctggcagcgg 420
atgatggcea cogcgggcet ggacceggce gtcaccgcgg aggaccggac caccgccagc 480
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cgcgacggcg acgegggcgg cegtcgctac aaccgtgage egtacgccga ccacaccggc 600
taccggccgg tcaacagcce gtacgagctg egcttcccgt egcgctggca gccgaacacc $\quad 660$
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aagcegatca cettcgageg geccgagcag ttceggctca ceccgccgcc gaaccaccac 780
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ggcgagaact tccccgcctc ggtcgcggcc gccgaccagt acgcgccgca gatcggcgac ..... 1440
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<210> SEQ ID NO 73 <211> LENGTH: 9762
<212> TYPE: DNA
<213> ORGANISM: Micromonospora sp. strain 046-ECO11
<400> SEQUENCE: 73
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tgctcgccet gctctcgcgg cgcgacatgt acggcctgga actggccgac tggctcgccg ..... 180
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ccggctccgt gcagacccgg tgggtggcec cegagcaggg gcacgcccgg cggtactacg ..... 300
cgatcaccga ccaggggcgg gcgcacctgc gggtgttcge ggcggtgtgg caggagatce ..... 360
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tgcaggcegg egcgagcetc gcecggetgg teggegtgge cacgetgcce gacaccgcce ..... 6300
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$<210>$ SEQ ID NO 74
$<211>$ LENGTH: 112
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 74

354045
Gly Ser Leu Tyr Pro Leu Leu Ala Arg Met Arg Gln Ala Gly Ser Val
505560

| Gln Thr Arg Trp Val Ala Pro Glu Gln Gly |  |
| :--- | :--- |
| 65 | 70 |
| 65 | 75 |
| 0 |  |

Ala Ile Thr Asp Gln Gly Arg Ala His Leu Arg Val Phe Ala Ala Val
85

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<210> SEQ ID NO 75
<211> LENGTH: 339
<212> TYPE: DNA
<213> ORGANISM: Micromonospora sp. strain 046-ECO11
<400> SEQUENCE: 75
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$<210>$ SEQ ID NO 76
$<211>$ LENGTH: 325
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 76

Gly Asp Ala Ala Ala Thr Pro Gly Pro Pro Ala Arg Pro Glu Pro Ala
305
310 $\quad 320$
$<210>$ SEQ ID NO 77
$<211>$ LENGTH: 978
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 77
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gccgacgccc tcgacagcgg acggagcgcc cacgagatcc tcgccggcct cggcgccgcg 180
cgggacgtgg cecggcagge gegcgaggag ctggggctgc cggcccagga cegcceggec 240
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gtgagcttcc tgetgcegtc egcagtgecg gtggagecga tccaggcegg ceccggegag 360
cagggcgtcc tccgccgget cggccccgga atcgcgctgc tcacgctgct gccggcgetc 420
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gcettctgge tgttcggcgg getgtacctg gcgetcggeg cegctgcgta caccgcctcg 900
cgggecgtcg acggegacge egcegcgacg cecggcecge cggcecggce ggaaccegcg 960
ceggececcg gaggetga 978
$<210>$ SEQ ID NO 78
$<211>$ LENGTH: 663
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 78



$<210>$ SEQ ID NO 79
$<211>$ LENGTH: 1992
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 79
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tacggcgaga coggtgacca ggcecgegag cgcatcacce ggetgetgtc cgaactcccc ..... 180
gtcgagctgg gcatcgcctc cggtttcccc gccgagtacg gcggccgcgg cgacgtgggc ..... 240
gcctcgatcg tcgccaccga gatgctggcc caggtggacc tgtcactgat ggtgaaggcc ..... 300
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<210> SEO ID NO 81
<211> LENGTH: 1722
<212> TYPE: DNA
<213> ORGANISM: Micromonospora sp. strain 046-ECO11
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<400> SEQUENCE: 81
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gaggcgctcg tgccggtggc gatcggcgtc atcatcgacc gggccgtggt gaccggcgac ..... 180
cegtgggcge tegcgtactc egtcgccggc etcgccgcce tgttcaccgt getggcgttc ..... 240
gcctaccgca acggcgcceg ccaggegttc gcggcggtgg aacgggagge gcacctgctg ..... 300
cgggtcgage tggcegageg egcgetcgac ccgcgcggge accgctccgg cetgcgegac ..... 360
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ccgggagaga tcgtcggcgt cetggcgtac gacceggccg acgcggacge getggtggcg ..... 1140
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cccgccgacg acctggacgt cgacgcgetg cgcggcgccg tcctggtcga gccgcacgac ..... 1260
gtgacgetgt tcgagggaac cgtggccgcc aacctcgccg ccgggagcag gaccgaggag ..... 1320
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accaccgccg tggacgcggc caccgaggce caactcgccg acggactggc cggcgcgcgc ..... 1560
cgcgaagcgc cecggggcac getgctggtc accagcagce cegcectgct gcggatcacc ..... 1620
gaccgggtgg tggtgatcgc egacggccgg gtgaccgccg aggggacgca cgagcacctg ..... 1680
ctggccaccg acgcccgcta cegcgaggag acactgcggt ga ..... 1722
$<210>$ SEQ ID NO 82
<211> LENGTH: 596
<212> TYPE: PRT
<213> ORGANISM: Micromonospora sp. strain 046-ECO11
$400>$ SEQUENCE: 82




$<210\rangle$ SEQ ID NO 84
<211> LENGTH: 507
<212> TYPE: PRT
<213> ORGANISM: Micromonospora sp. strain 046-ECO11
<400> SEQUENCE: 84


$<210>$ SEQ ID NO 85
$<211>$ LENGTH: 1524
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE: 85


| tcggtggtga | acgacccgct cgtcggcgac | gagggeggte tcacagtgac cegcgactac | 600 |
| :---: | :---: | :---: | :---: |
| cggacctacg | tegaggcgtt cgccacggce | gcgcagcgca agtgggactc ggtacgccgg | 660 |
| tacgtgcagc | ccggccgcat cgtggacatc | ggctgcggcg cgggcgcegt cetggaactc | 720 |
| gccgaccggg | aggcegcgct gcgtgagagc | gacctgatcg gcgtggaggt cgcccgccac | 780 |
| ctetaccagg | agtgcetgca caagaaggcg | cagggcgtgt tccgcaacge caacgtctac | 840 |
| ttcttccacc | gcaacgtcet cggcggcgeg | gtgttcaagg accgctcggt cgacaccacg | 900 |
| ctcacgttcg | cgetgaccca cgagatctgg | tcgtacggge ggcggcggga gtcgctgctg | 960 |
| cagttcgccc | gcegcatcca cgaccacacg | gtgcecggcg gcgtctggat caacagcgac | 1020 |
| gtgtgcggtc | cggacgacce coggcggcag | gtgctectgc gactgtccac cgacgacgge | 1080 |
| gacaacccgg | cogegcecog cecogacetc | gcegagctga cetcggcgga ggtecggcgt | 1140 |
| tacgtcggcg | ggctgtcgac gcgggcgcgg | ctggaccagt tcgcogtcga cttcgcgttc | 1200 |
| gacttcgact | acgagcegct ceccgacggc | gcggtacgce tgacgctggg cgcegcgatg | 1260 |
| gactacctga | cccgcaagga ctacacggac | aactggctgt cggagacgca ggagcagttc | 1320 |
| tgcggcetga | gettcgccga ctggacggac | ctgctcaccg aggcggggtt cgagatcggc | 1380 |
| ccggcgtcgg | cgceggtgcg caacgagtgg | gtgatcgaca accggatcgc gccagtcgeg | 1440 |
| tccctcaccg | acctcgacgg coggcegctg | gactggccga ccacccacgt cetcaccgtc | 1500 |
| gcecaccgce | cccgcaacca gtga |  | 1524 |

$<210>$ SEQ ID NO 86
$<211>$ LENGTH: 232
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046 -ECO11
$<400>$ SEQUENCE : 86


| $\begin{aligned} & \text { Val Leu Ala Ala Thr Phe Ile Gly Leu Ala Val Leu } \operatorname{Trp} \\ & 180 \\ & 185\end{aligned}$ |  |
| :---: | :---: |
| Tyr Ile Met Leu Ile Asp Arg Leu Gln Pro Trp Leu Thr Arg Pro Ser |  |
| Val Leu Leu Val Ile Glu Arg Leu Thr Gly Leu Ile Leu Ile Val Leu 210215220 |  |
|  |  |
| $<210\rangle$ SEQ ID NO 87 |  |
| <211> LENGTH: 699 |  |
| <212> TYPE: DNA |  |
| <213> ORGANISM: Micromonospora sp. strain 046-ECO11 |  |
| <400> SEQUENCE: 87 |  |
| gtgtctgaca tccagatcat cagtttcgtc gccgccagce tgctcatcat catcgtgccg | 60 |
| ggcgtcgact tcgcgctcgt cacccggcag accgtcaggt acggccggcg ggccgggttc | 120 |
| gtggtgetgg cegggetgtt cgtcgccgcg ctggtgcacg egtegttcgc gaccgccggc | 180 |
| ctgtecgcce tgctggtctc ctcgecgacg ctctacacgg tgctgcgcgt egceggcgeg | 240 |
| ctgtacctgc tctacetggg cggcacgatc ctctgggcga cocggccgcg coggacggtc | 300 |
| coggcggcge agceggtcac tgteggegcg ggcggegceg ggceggacac ggacaccgge | 360 |
| cccgcgcegg tgceggacac cecggcegce gacgagcoge acgtggcecg cegctcgttc | 420 |
| gtcatgggcg tcaccagcea gctgctgaac gtcaaggtgg tegtcttcta cgtctcgttc | 480 |
| gtgcegcagt tcgtcaagce cggcgagggg gcggcggcec gtacggcggt gctcgccgce | 540 |
| acgttcatcg gcctcgcggt gctctggtgg gcctgctaca tcatgctcat cgacaggttg | 600 |
| cagcectggc tgacceggce gtcegtgctg etggtgatcg aacggctgac cgggetcatc | 660 |
| ctgatcgtcc tggcgatccg gatcgcgetg agceggtga | 699 |

$<210>$ SEQ ID NO 88
$<211>$ LENGTH: 132
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora sp. strain 046-ECO11
$<400>$ SEQUENCE : 88


## Arg Arg Arg Val

$$
130
$$

| $<210>$ SEQ ID NO 89 |  |
| :---: | :---: |
| $<211>$$<212>$ TYPE: DNA |  |
|  |  |
| <213> ORGANISM: Micromonospora sp. strain 046-ECO11 |  |
| <400> SEQUENCE: 89 |  |
| gtgggcgtga gcgcgatgac gacattcgac tacgacggce gcgtcttcgt ctcggtggac 6 |  |
| cacgacgceg gtgacggcge cgagcegctg cgggggcact accaccagcg tggcgacetg 120 |  |
| gtctgggcgg agatcaccgg eggcecggtc cggcacggce ggctggcegg cacctgcgac 180 |  |
| gcgcagggcg tegtgcgett egcetacctg gaggtgctca cegacggcac catagtcatc 240 |  |
| ggcgagtgcg agtcecggce cgaacggetg ceggacggce ggatcoggct gegggaacag 300 |  |
| tggcgceggc acggaccacg ccaggacagc ggcgtctccg tcatcgagga ggcagtgceg | 360 |
| gcgctcgceg gaggacagga gagceggcgt egtgtctga | 399 |

$<210>$ SEQ ID NO 90
$<211>$ LENGTH: 296
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora echinospora challisensis
$<400>$ SEQUENCE: 90


$<210>$ SEQ ID NO 91
$<211>$ LENGTH: 891
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora echinospora challisensis
$<400>$ SEQUENCE: 91

| $\begin{aligned} & \text { Ala } \\ & 1 \end{aligned}$ | Thr | Gly | Cys | $\begin{aligned} & \text { Cys } \\ & 5 \end{aligned}$ | Cys | Gly | Gly | $\text { Ala } 7$ | Ala <br> 10 | Cys |  |  | Cys | $\begin{aligned} & \text { Cys } \\ & 15 \end{aligned}$ | Gly |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ala | Gly | Gly | $\begin{aligned} & \text { Cys } \\ & 20 \end{aligned}$ | Gly | $\mathrm{Gly}$ | Thr | Gly | $\begin{aligned} & \text { Gly } \\ & 25 \end{aligned}$ | Ala | Ala | Cys | Thr | $\begin{aligned} & \text { CYs } \\ & 30 \end{aligned}$ |  | Gly |
| Thr | Thr | $\begin{aligned} & \text { Cys } \\ & 35 \end{aligned}$ | Cys A | Ala | Cys | Cys | $\begin{aligned} & \text { Ala } \\ & 40 \end{aligned}$ | Thr | Cys | Gly | Ala | $\begin{aligned} & \text { Gly } \\ & 45 \end{aligned}$ | Gly | Ala | Ala |
| Thr | $\begin{aligned} & \text { Cys } \\ & 50 \end{aligned}$ | Gly | Gly C | Cys | Cys | $\begin{aligned} & \text { Cys } \\ & 55 \end{aligned}$ | Gly | Gly | Cys | Thr | $\begin{aligned} & \text { Gly } \\ & 60 \end{aligned}$ | Cys | Thr | Gly | Ala |
| $\begin{aligned} & \text { Ala } \\ & 65 \end{aligned}$ | Cys | Gly | Thr | $1 Y$ | $\begin{aligned} & \text { Gly } \\ & 70 \end{aligned}$ | Cys | Cys | Thr | Gly | $\begin{aligned} & \text { Cys } \\ & 75 \end{aligned}$ | Thr | Cys | Gly | Cys | $\begin{aligned} & \text { Gly } \\ & 80 \end{aligned}$ |
| Thr | Gly | Ala | Cys | Ala $85$ | Gly | Gly | Gly | Thr | $\begin{aligned} & \text { Cys } \\ & 90 \end{aligned}$ | Thr | Gly | 1 Y | hr | $\begin{aligned} & \text { Cys } \\ & 95 \end{aligned}$ | Cys |
| Cys | Thr | Gly | $\begin{aligned} & \text { Cys T } \\ & 100 \end{aligned}$ | Thr | Cys | Thr | Cys | $\begin{aligned} & \text { Cys } \\ & 105 \end{aligned}$ | Gly | Cys | Gly | Thr | $\begin{aligned} & \text { Ala } \\ & 110 \end{aligned}$ | Cys | Gly |
| Gly | Thr | $\begin{aligned} & \mathrm{Gly} \\ & 115 \end{aligned}$ | Ala | Cys | ly | Cys | $\begin{aligned} & \text { Gly } \\ & 120 \end{aligned}$ | Thr | Thr | Cys | Gly | $\begin{aligned} & \text { Cys } \\ & 125 \end{aligned}$ | Gly | Cys | Ala |
| Cys | $\begin{aligned} & \text { Cys } \\ & 130 \end{aligned}$ | Cys | Cys | Gly | Gly | $\begin{aligned} & \text { Thr } \\ & 135 \end{aligned}$ | Gly | Cys | Cys | Gly | $\begin{aligned} & \text { Thr } \\ & 140 \end{aligned}$ | Gly | Gly | Thr | Cys |
| $\begin{aligned} & \text { Gly } \\ & 145 \end{aligned}$ | Cys | Cys | hr T | hr | $\begin{aligned} & \text { Cys } \\ & 150 \end{aligned}$ | Cys | Gly | Gly | Gly | $\begin{aligned} & \text { Thr } \\ & 155 \end{aligned}$ | Gly | Gly | Cys | Gly | $\begin{aligned} & \text { Ala } \\ & 160 \end{aligned}$ |
| cys | Cys | Gly | Cys | $\begin{aligned} & \mathrm{Gly} \\ & 165 \end{aligned}$ | Ala | Thr | Gly | Cys | $\begin{aligned} & \text { Gly } \\ & 170 \end{aligned}$ | Cys | Cys | Ala | Cys | $\begin{aligned} & \text { Gly } \\ & 175 \end{aligned}$ | Thr |
| Gly | Gly | Gly | $\begin{aligned} & \text { Ala } \\ & 180 \end{aligned}$ | Gly | Ala | Gly | Cys | $\begin{aligned} & \text { Thr } \\ & 185 \end{aligned}$ | Cys | Gly | Ala | Cys | $\begin{aligned} & \text { Thr } \\ & 190 \end{aligned}$ | Gly | Thr |
| Cys | Gly | $\begin{aligned} & \text { Gly } \\ & 195 \end{aligned}$ | Thr T | Thr | cys | Ala | $\begin{aligned} & \text { Cys } \\ & 200 \end{aligned}$ | Gly | Ala | Cys | Gly | $\begin{aligned} & \text { Cys } \\ & 205 \end{aligned}$ | Ala | Cys | Cys |
| Cys | $\begin{aligned} & \text { Gly } \\ & 210 \end{aligned}$ | Gly | la | Cys |  | $\begin{aligned} & \text { Ala } \\ & 215 \end{aligned}$ | Cys | Cys | Gly | Cys | $\begin{aligned} & \text { Gly } \\ & 220 \end{aligned}$ | Ala | CYs | Cys | Cys |
| $\begin{aligned} & \text { Cys } \\ & 225 \end{aligned}$ | Thr | Ala | Cys | Gly | $\begin{aligned} & \text { Cys } \\ & 230 \end{aligned}$ | Cys | Cys | Gly | Thr | $\begin{aligned} & \text { Gly } \\ & 235 \end{aligned}$ | Cys | Gly | Cys | Thr | $\begin{aligned} & \text { Gly } \\ & 240 \end{aligned}$ |
| Thr | Cys | Gly | Cys | $\begin{aligned} & \text { Gly } \\ & 245 \end{aligned}$ | Cys | Gly | Gly | Cys | $\begin{aligned} & \text { Cys } \\ & 250 \end{aligned}$ | Thr | Cys | Ala | Cys | $\begin{aligned} & \text { Cys } \\ & 255 \end{aligned}$ | Cys |
| Cys | Gly | Gly | $\begin{aligned} & \text { Ala } \\ & 260 \end{aligned}$ | Gly | Ala | Cys | Gly | $\begin{aligned} & \text { Gly } \\ & 265 \end{aligned}$ | Ala | Cys | Cys | Ala | $\begin{aligned} & \text { Cys } \\ & 270 \end{aligned}$ | Cys | Cys |
| Gly | Gly | $\begin{aligned} & \text { Thr } \\ & 275 \end{aligned}$ | Cys | Gly | Gly | Cys | $\begin{aligned} & \text { Ala } \\ & 280 \end{aligned}$ | Cys | Cys | Cys | Thr | $\begin{aligned} & \text { Gly } \\ & 285 \end{aligned}$ | Cys | Thr | Cys |


| Thr | $\begin{aligned} & \text { Cys } \\ & 290 \end{aligned}$ | Cys | Gly |  | Gly | $\begin{aligned} & \text { Gly } \\ & 295 \end{aligned}$ |  |  |  | Ala | $\begin{aligned} & \text { Gly } \\ & 300 \end{aligned}$ | Gly |  |  | Cys |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Gly | Thr | Thr | Gly | Thr | Cys | Cys | Gly | Gly | Thr | Gly G | Gly | Ala | Gly | Ala | Gly |
| 305 |  |  |  |  | 310 |  |  |  |  | 315 |  |  |  |  | 320 |
| cys | Cys | Ala | Cys | $\begin{aligned} & \text { Gly } \\ & 325 \end{aligned}$ | Gly | Cys | Ala | Thr | $\begin{aligned} & \text { Cys } \\ & 330 \end{aligned}$ | Gly I | Ala | Cys | Thr | $\begin{aligned} & \text { Thr } \\ & 335 \end{aligned}$ | Cys |
| Gly | Gly | Gly | $\begin{aligned} & \text { Gly } \\ & 340 \end{aligned}$ | Thr | Cys | Gly | Thr | $\begin{aligned} & \text { Cys } \\ & 345 \end{aligned}$ | Gly | Gly | Cys | Gly | $\begin{aligned} & \text { Gly } \\ & 350 \end{aligned}$ | Cys | Thr |
| Thr | Cys | $\begin{aligned} & \text { Ala } \\ & 355 \end{aligned}$ | Ala | Gly | Ala | Ala | $\begin{aligned} & \text { Gly } \\ & 360 \end{aligned}$ | Ala | Thr | Cys | Thr | $\begin{aligned} & \text { Ala } \\ & 365 \end{aligned}$ | Cys | Gl | Cys |
| Gly | $\begin{aligned} & \text { Thr } \\ & 370 \end{aligned}$ | Thr | Cys | hr | Thr | $\begin{aligned} & \text { Cys } \\ & 375 \end{aligned}$ | Ala | Cys | Cys | Cys | $\begin{aligned} & \text { Cys } \\ & 380 \end{aligned}$ | Gly | Gly | Al | Cys |
| $\begin{aligned} & \text { Gly } \\ & 385 \end{aligned}$ | Ala | Cys | Cys | r | $\begin{aligned} & \text { Gly } \\ & 390 \end{aligned}$ | Cys | Ala | Gly | $1 y$ | Ala $395$ | Gly | Ala | Cys | Gly | $\begin{aligned} & \text { Thr } \\ & 400 \end{aligned}$ |
| Cys | Gly | Ala | Ala | $\begin{aligned} & \text { Gly } \\ & 405 \end{aligned}$ | Cys | Thr | Cys | Gly | $\begin{aligned} & \text { Cys } \\ & 410 \end{aligned}$ | Cys | Gly | Ala | Gly | $\begin{aligned} & \text { Ala } \\ & 415 \end{aligned}$ | Thr |
| Cys | Cys | Cys | $\begin{aligned} & \text { Cys } \\ & 420 \end{aligned}$ | Gly | Cys | Cys | Ala | $\begin{aligned} & \text { Thr } \\ & 425 \end{aligned}$ | Gly | Cys | Cys | Gly | $\begin{aligned} & \text { Cys } \\ & 430 \end{aligned}$ | Gl | Cys |
| Ala | Gly | $\begin{aligned} & \text { Cys } \\ & 435 \end{aligned}$ | Cys | Thr | Gly | Gly | $\begin{aligned} & \text { Cys } \\ & 440 \end{aligned}$ | Cys | Gly | Y | ly | $\begin{aligned} & \text { Ala } \\ & 445 \end{aligned}$ | Ala | Cy | Gly |
| Thr | $\begin{aligned} & \text { Cys } \\ & 450 \end{aligned}$ | Gly | Ala | $1 Y$ | $r$ | $\begin{aligned} & \text { Thr } \\ & 455 \end{aligned}$ | Cys | Thr | Thr | ys | $\begin{aligned} & \text { Gly } \\ & 460 \end{aligned}$ | cys | Cys | Cy | Gly |
| $\begin{aligned} & \text { Thr } \\ & 465 \end{aligned}$ | Cys | Ala | Cys | Gly | $\begin{aligned} & \text { Gly } \\ & 470 \end{aligned}$ | Ala | Cys | Thr | Gly | $\begin{aligned} & \text { Gly } \\ & 475 \end{aligned}$ | la | ys | $1 Y$ | Al | Cys 480 |
| Cys | Gly | Gly | Gly | $\begin{aligned} & \text { Thr } \\ & 485 \end{aligned}$ | Cys | Gly | Gly | Gly | $\begin{aligned} & \text { Gly } \\ & 490 \end{aligned}$ | Thr | Gly | hr | Thr | $\begin{aligned} & \text { Cys } \\ & 495 \end{aligned}$ | Gly |
| Gly | Gly | Ala | $\begin{aligned} & \text { Thr } \\ & 500 \end{aligned}$ | Cys | Gly | Ala | Cys | $\begin{aligned} & \text { Thr } \\ & 505 \end{aligned}$ | Ala | Cys | Cys | Cys | $\begin{aligned} & \text { Gly } \\ & 510 \end{aligned}$ | Al | Gly |
| Cys | Cys | $\begin{aligned} & \text { Gly } \\ & 515 \end{aligned}$ | Gly |  | Cys | Gly | $\begin{aligned} & \text { Gly } \\ & 520 \end{aligned}$ | Thr | Gly | $1 a$ | Ala | $\begin{aligned} & \text { Cys } \\ & 525 \end{aligned}$ | Gly | Th | Gly |
| Thr | $\begin{aligned} & \text { Ala } \\ & 530 \end{aligned}$ | Cys | hr | r | ys | $\begin{aligned} & \text { Ala } \\ & 535 \end{aligned}$ | Ala | $y s$ | Gly | a | $\begin{aligned} & \text { Cys } \\ & 540 \end{aligned}$ | Gly | Thr | Al | Cys |
| $\begin{aligned} & \text { Cys } \\ & 545 \end{aligned}$ | Cys | Gly | Cys | Cys | $\begin{aligned} & \text { Gly } \\ & 550 \end{aligned}$ | Ala | Gly | Ala | Gly | $\begin{aligned} & \text { Cys } \\ & 555 \end{aligned}$ | Thr | Thr | Cys | Cys | Ala 560 |
| Cys | Thr | Cys | Gly | $\begin{aligned} & \text { Gly } \\ & 565 \end{aligned}$ | Ala | Gly | la | Cys | $\begin{aligned} & \text { Gly } \\ & 570 \end{aligned}$ | Ala | hr | ys | Cys | $\begin{aligned} & \text { Gly } \\ & 575 \end{aligned}$ | Gly |
| Thr | Cys | Gly | $\begin{aligned} & \text { Ala } \\ & 580 \end{aligned}$ | Cys | Gly | Cys | Thr | $\begin{aligned} & \text { Cys } \\ & 585 \end{aligned}$ | Cys | Gly | Gly | Gly | Ala $590$ | Gly | Ala |
| Thr | Cys | $\begin{aligned} & \text { Gly } \\ & 595 \end{aligned}$ | Gly | Cys | Ala | Thr | $\begin{aligned} & \text { Gly } \\ & 600 \end{aligned}$ | Gly | Cys | Cys | Gly | Ala <br> 605 | Ala | Cy | cys |
| Cys | $\begin{aligned} & \text { Ala } \\ & 610 \end{aligned}$ | Gly | Thr | Gly | Ala | $\begin{aligned} & \text { Gly } \\ & 615 \end{aligned}$ | Cys | Gly | Gly | la | $\begin{aligned} & \text { Thr } \\ & 620 \end{aligned}$ | Gly | Cys | Th | Cys |
| $\begin{aligned} & \text { Ala } \\ & 625 \end{aligned}$ | Ala | Gly | Cys | Thr | $\begin{aligned} & \text { Cys } \\ & 630 \end{aligned}$ | Gly | Gly | Cys | Gly | $\begin{aligned} & \text { Ala } \\ & 635 \end{aligned}$ | Gly | Ala | Ala | Gly | Gly 640 |
| Cys | Gly | Thr | Thr | $\begin{aligned} & \text { Cys } \\ & 645 \end{aligned}$ | Gly | Gly | Ala | Cys | $\begin{aligned} & \text { Thr } \\ & 650 \end{aligned}$ | Gly | Thr | Ala | Thr | $\begin{aligned} & \text { Gly } \\ & 655 \end{aligned}$ | Th |
| Cys | Ala | Cys | $\begin{aligned} & \text { Cys } \\ & 660 \end{aligned}$ | Cys | Thr | Cys | Gly | $\begin{aligned} & \text { Gly } \\ & 665 \end{aligned}$ | Cys | Thr | Gly | Gly | $\begin{aligned} & \text { Gly } \\ & 670 \end{aligned}$ | Al | Th |
| Thr | Cys | $\begin{aligned} & \text { Gly } \\ & 675 \end{aligned}$ | Thr | Cys | Gly | Ala | $\begin{aligned} & \text { Gly } \\ & 680 \end{aligned}$ | Gly | Ala | Thr | Cys | $\begin{aligned} & \text { Gly } \\ & 685 \end{aligned}$ | Ala | Gly | Cys |


|  | 690 |  |  |  |  | 695 |  |  |  |  | 700 |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| $\begin{aligned} & \text { Cys } \\ & 705 \end{aligned}$ | Gly | Cys | Gly | Ala | $\begin{aligned} & \text { Cys } \\ & 710 \end{aligned}$ | Cys | Ala | Cys | Cys | $\begin{aligned} & \text { Gly } \\ & 715 \end{aligned}$ | Ala | Cys | Cys | Thr | $\begin{aligned} & \text { Gly } \\ & 720 \end{aligned}$ |
| Ala | Cys | Gly | Ala | $\begin{aligned} & \text { Cys } \\ & 725 \end{aligned}$ | Cys | Cys | Thr | Gly | $\begin{aligned} & \text { Cys } \\ & 730 \end{aligned}$ | Cys | Cys | Gly | Thr | $\begin{aligned} & \text { Thr } \\ & 735 \end{aligned}$ | Cys |
| Cys | Cys | Gly | $\begin{aligned} & \text { Thr } \\ & 740 \end{aligned}$ | Cys | Gly | Ala | Ala | $\begin{aligned} & \text { Cys } \\ & 745 \end{aligned}$ | Cys | Gly | Gly | Ala | $\begin{aligned} & \text { Gly } \\ & 750 \end{aligned}$ |  | Thr |
| Cys | Gly | $\begin{aligned} & \text { Ala } \\ & 755 \end{aligned}$ | Gly | Ala | Ala | Gly | $\begin{aligned} & \text { Thr } \\ & 760 \end{aligned}$ | Thr | Cys | Gly | Thr | $\begin{aligned} & \text { Ala } \\ & 765 \end{aligned}$ | Cys | Gly | Gly |
| Ala | $\begin{aligned} & \text { Gly } \\ & 770 \end{aligned}$ | Cys | Gly | Thr | Thr | $\begin{aligned} & \text { Cys } \\ & 775 \end{aligned}$ | Cys | Gly | Thr | Ala | $\begin{aligned} & \text { Cys } \\ & 780 \end{aligned}$ | Gly | Gly | Thr | Gly |
| $\begin{aligned} & \text { Gly } \\ & 785 \end{aligned}$ | Gly | Gly | Ala | Ala | $\begin{aligned} & \text { Gly } \\ & 790 \end{aligned}$ | Ala | Cys | Cys | Gly | $\begin{aligned} & \text { Thr } \\ & 795 \end{aligned}$ | Ala | Ala | Gly | Thr | $\begin{aligned} & \text { Thr } \\ & 800 \end{aligned}$ |
| Cys | Gly | Thr | Cys | $\begin{aligned} & \text { Thr } \\ & 805 \end{aligned}$ | Ala | Cys | Gly | Gly | $\begin{aligned} & \text { Cys } \\ & 810 \end{aligned}$ | Gly | Thr | Cys | Gly | $\begin{aligned} & \text { Cys } \\ & 815 \end{aligned}$ | Gly |
| Thr | Thr | Gly | Ala $820$ | Cys | Cys | Cys | Cys | $\begin{aligned} & \text { Gly } \\ & 825 \end{aligned}$ | Cys | Ala | Cys | Gly | $\begin{aligned} & \text { Gly } \\ & 830 \end{aligned}$ | Cys | Gly |
| Ala | Gly | $\begin{aligned} & \text { Thr } \\ & 835 \end{aligned}$ | Ala | Cys | Thr | Ala | $\begin{aligned} & \text { Cys } \\ & 840 \end{aligned}$ | Ala | Ala | Gly | Cys | $\begin{aligned} & \text { Thr } \\ & 845 \end{aligned}$ | Cys | Gly | Ala |
| Gly | $\begin{aligned} & \text { Thr } \\ & 850 \end{aligned}$ | Cys | Gly | Cys | Ala | $\begin{aligned} & \text { Cys } \\ & 855 \end{aligned}$ | Thr | Ala | Cys | Cys | $\begin{aligned} & \text { Gly } \\ & 860 \end{aligned}$ | Gly | Thr |  | Gly |
| $\begin{aligned} & \text { Ala } \\ & 865 \end{aligned}$ | Ala | Gly | Cys | Cys | $\begin{aligned} & \text { Cys } \\ & 870 \end{aligned}$ | Gly | Gly | Gly | Gly | $\begin{aligned} & \text { Cys } \\ & 875 \end{aligned}$ | Gly | Ala | Thr |  | Gly 880 |
| Ala | Cys | Thr | Thr | $\begin{aligned} & \text { Cys } \\ & 885 \end{aligned}$ | Ala | Thr | Cys | Thr | $\begin{aligned} & \text { Gly } \\ & 890 \end{aligned}$ | Ala |  |  |  |  |  |

$<210>$ SEQ ID NO 92
$<211>$ LENGTH: 438
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Micromonospora echinospora challisensis
$<400>$ SEQUENCE: 92


| Leu | Ala | Met | Pro | $\begin{aligned} & \text { TYr } \\ & 165 \end{aligned}$ | Ala | Arg | Val |  | $\begin{aligned} & \text { Arg } \\ & 170 \end{aligned}$ | Val | Met | His | Asp | $\begin{aligned} & \text { Leu } \\ & 175 \end{aligned}$ |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Val | Thr | Leu | $\begin{aligned} & \text { Thr } \\ & 180 \end{aligned}$ | $\operatorname{Trp}$ | Ser | Val | Pro | $\begin{aligned} & \text { Thr } \\ & 185 \end{aligned}$ | Glu | Cys | Leu | Ile | $\operatorname{Trp}$ <br> 190 |  | Ala |
| Ala | Ala | $\begin{aligned} & \text { Thr } \\ & 195 \end{aligned}$ | Ala | Ala | $\mathrm{Gly}$ | His | $\begin{aligned} & \text { Arg } \\ & 200 \end{aligned}$ |  | Asp | Val | Asp | $\begin{aligned} & \text { Phe } \\ & 205 \end{aligned}$ |  |  | Leu |
| Arg | $\begin{aligned} & \text { Ala } \\ & 210 \end{aligned}$ | Leu | Phe | Val | Gly | $\begin{aligned} & \text { Gly } \\ & 215 \end{aligned}$ | Glu | Pro | Leu | Thr | $\begin{aligned} & \text { Asp } \\ & 220 \end{aligned}$ | Ala | Arg | Arg | Arg |
| $\begin{aligned} & \text { Arg } \\ & 225 \end{aligned}$ | Ile | Ser | Arg | Leu | $\begin{aligned} & \operatorname{Trp} \\ & 230 \end{aligned}$ |  | Val | Pro | Val | $\begin{aligned} & \text { Ile } \\ & 235 \end{aligned}$ | Glu | Glu | Tyr | Gly | $\begin{aligned} & \text { Ser } \\ & 240 \end{aligned}$ |
| Thr | Glu | Thr | Gly | $\begin{aligned} & \text { Ser } \\ & 245 \end{aligned}$ | Leu | Ala | Gly |  | $\begin{aligned} & \text { Cys } \\ & 250 \end{aligned}$ | Pro | Asn | $\mathrm{Gly}$ | Arg | $\begin{aligned} & \text { Met } \\ & 255 \end{aligned}$ | His |
| Leu | Trp | Ala | $\begin{aligned} & \text { Asp } \\ & 260 \end{aligned}$ | Arg | Ala |  | Phe | $\begin{aligned} & \text { Glu } \\ & 265 \end{aligned}$ | Val | Tyr | Asp | Pro | $\begin{aligned} & \text { Arg } \\ & 270 \end{aligned}$ | Thr | Gly |
| Thr | Val | $\begin{aligned} & \text { Ser } \\ & 275 \end{aligned}$ | Ala | Asp | Gly | Asp | $\begin{aligned} & \text { Gly } \\ & 280 \end{aligned}$ | $\mathrm{Gln}$ | Leu | Val | Val | $\begin{aligned} & \text { Thr } \\ & 285 \end{aligned}$ | Pro | Leu | Phe |
| Arg | $\begin{aligned} & \text { Glu } \\ & 290 \end{aligned}$ | Ala | Met | Pro | Leu | $\begin{aligned} & \text { Leu } \\ & 295 \end{aligned}$ | Arg | Tyr | Asn | Leu | $\begin{aligned} & \text { Glu } \\ & 300 \end{aligned}$ | Asp | Asp | Val | Thr |
| $\begin{aligned} & \text { Val } \\ & 305 \end{aligned}$ | Ser | Tyr | Asp | Asp | $\begin{aligned} & \text { Cys } \\ & 310 \end{aligned}$ | Ala | Cys | Gly | $\operatorname{Trp}$ | $\begin{aligned} & \text { Asn } \\ & 315 \end{aligned}$ | Leu |  | Thr | Val | $\begin{aligned} & \text { Arg } \\ & 320 \end{aligned}$ |
| Val | Leu | Gly | Arg | $\begin{aligned} & \text { Ala } \\ & 325 \end{aligned}$ | Ala | Phe | Gly | Tyr | $\begin{aligned} & \text { Arg } \\ & 330 \end{aligned}$ | Val | Gly |  |  | $\begin{aligned} & \text { Thr } \\ & 335 \end{aligned}$ | Ile |
| Thr | Gln | His | $\begin{aligned} & \text { Arg } \\ & 340 \end{aligned}$ | Leu | Glu | Glu | Val | $\begin{aligned} & \text { Val } \\ & 345 \end{aligned}$ | Phe |  | Leu | Pro | $\begin{aligned} & \text { Glu } \\ & 350 \end{aligned}$ | Ser | His |
| Gly | Val | $\begin{aligned} & \text { Val } \\ & 355 \end{aligned}$ | Phe | $\operatorname{Trp}$ | Arg | Ala | $\begin{aligned} & \text { Lys } \\ & 360 \end{aligned}$ | Ala | Glu | Pro | Thr | $\begin{aligned} & \text { Val } \\ & 365 \end{aligned}$ | Leu | Arg | Ile |
| Glu | $\begin{aligned} & \text { Ile } \\ & 370 \end{aligned}$ | Glu | Val | Ala | Glu | $\begin{aligned} & \text { Glu } \\ & 375 \end{aligned}$ | His | Arg | Thr | Ala | $\begin{aligned} & \text { Ala } \\ & 380 \end{aligned}$ |  | Ala | Glu | Leu |
| $\begin{aligned} & \text { Thr } \\ & 385 \end{aligned}$ | Ala | Ser | Val | Arg | $\begin{aligned} & \text { Ala } \\ & 390 \end{aligned}$ | Thr |  | Gly | Ile | $\begin{aligned} & \text { Asp } \\ & 395 \end{aligned}$ | Ser |  | Val | Thr | $\begin{aligned} & \text { Gly } \\ & 400 \end{aligned}$ |
| Leu | Thr | Pro | Gly | $\begin{aligned} & \text { Thr } \\ & 405 \end{aligned}$ | Leu |  | Pro | Arg | $\begin{aligned} & \mathrm{Glu} \\ & 410 \end{aligned}$ | Ala | Leu |  |  | $\begin{aligned} & \text { Met } \\ & 415 \end{aligned}$ | Pro |
| Asp | Val | Val | $\begin{aligned} & \text { Lys } \\ & 420 \end{aligned}$ | Pro | Arg | Ser | Leu | Phe <br> 425 | Gly | Pro | Asp | Glu | $\begin{aligned} & \text { Asp } \\ & 430 \end{aligned}$ | Trp | Gly |
| Lys | Ala | Leu $435$ |  | Tyr | Tyr |  |  |  |  |  |  |  |  |  |  |

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taccgcgacc ggctggacce gggggccctg cccgcgaccg cegccgacct cgccgacctg ..... 180
ccgctgacca cgaagcagga cctgcgggac aactacccct tcggcatgct cgccgtcccg ..... 240
aaggagcggc tggccaccta ccacgagtcg agcgggacgg caggccggcc cacgccctcc ..... 300
tactacacgg eggaggactg gaccgacctg gccgagcget tcgcccgcaa gtggatcggg ..... 360
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$<210>$ SEQ ID NO 94
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$<212>$ TYPE: PRT
$<213>$ ORGANISM: Streptomyces carzinostaticus neocarzinostaticus
$<400>$ SEQUENCE: 94

$<210>$ SEQ ID NO 95
$<211>$ LENGTH: 507
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Streptomyces carzinostaticus neocarzinostaticus



$<210>$ SEQ ID NO 98
$<211>$ LENGTH: 290
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Artificial sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: Miscfeature
$<222>$ LOCATION: (1)..(290)
$<223>$ OTHER INFORMATION: HMM consensus seq based on alignment of Fig IO IPTN
$<400>$ SEQUENCE : 98

$3540 \quad 45$
Ala Leu Arg His Val Gly glu Leu Asp Cys Arg Phe Thr Thr His Pro
Asp Asp Arg Asp Pro Tyr Ala Leu Ala Leu Ser Arg Gly Leu Thr Pro

| Lys Thr Asp His Pro Val Gly Ser Leu Leu Ser Glu Val Gln Glu Arg |  |
| :---: | :---: |
| 85 | 90 |
| 95 |  |


| Leu Pro Val Glu Ser Tyr Gly Ile Asp |  |  |
| ---: | ---: | ---: |
| loo | 105 | Ghe Gly Val Val Gly Gly Phe |
| 110 |  |  |

Lys Lys Ile Tyr Ala Phe Phe Thr Pro Asp Glu Leu Gln Glu Val Ala
115
120

| Ala Leu Ala Gly Ile Pro Ala Met Pro Arg Ser Leu Ala Gly Asn Ala |  |
| ---: | :--- |
| 130 | 135 |




Ala Leu Pro Val Thr Ala Ala Asp Leu Ala Asp Leu Pro Leu Thr Thr



| Ala Gly Arg Leu Arg Gly Ala Thr Val Val Pro Gly Asp Asn Arg Ser |  |  |  |
| :--- | ---: | ---: | ---: |
| 145 | 150 | 155 | 160 |

Leu Ala Met Pro Tyr Ala Arg Val Val Arg Val Met His Asp Leu Gly
Val Thr Leu Thr Trp Ser Val Pro Thr Glu Cys Leu Ile Trp Ala Al

Arg Ile Ser Arg Leu Trp Gly Val Pro Val Ile Glu Glu Tyr Gly Ser
Thr Glu Thr Gly Ser Leu Ala Gly Glu Cys Pro Glu Gly Arg Leu His
Leu Trp Ala Asp Arg Ala Leu Phe Glu Val Tyr Asp Pro Asp Thr Gly

Thr Val Arg Ala Asp Gly Asp Gly Gln Leu Val Val Thr Pro Leu Phe | 280 |  |
| ---: | :--- |
| 275 | 280 |

Arg Glu Ala Met Pro Leu Leu Arg Tyr Asn Leu Glu Asp Asn Val Ser$290 \quad 295 \quad 300$
Val Ser Tyr Asp Asp Cys Ala Cys Gly Trp Lys Leu Pro Thr Val Arg


What is claimed is:

1. An isolated polynucleotide comprising a polynucleotide sequence, or a polynucleotide sequence complementary thereto, selected from the group consisting of:
a) a polynucleotide encoding a polypeptide having at least $90 \%$ sequence identity to a polypeptide consisting of amino acids 1-438 of SEQ ID NO: 48 and having adenylating amide synthetase activity;
b) a polynucleotide encoding a polypeptide having at least $90 \%$ sequence identity to a polypeptide consisting of amino acids 1-290 of SEQ ID NO: 22 and having isoprenyl transferase activity;
c) a polynucleotide comprising the nucleic acid sequence of SEQ ID NO:47; and
d) a polynucleotide comprising the nucleic acid sequence of SEQ ID NO:23.
2. An isolated polynucleotide comprising the nucleic acid sequence of SEQ ID NO:47.
3. An isolated polynucleotide comprising the nucleic acid sequence of SEQ ID NO:23.
4. The isolated polynucleotide of claim 1 , wherein said polypeptide of a) has at least $95 \%$ sequence identity to a polypeptide consisting of amino acids 1-438 of SEQ ID NO: 48.
5. The isolated polynucleotide of claim 1, wherein said polypeptide of a) has at least $99 \%$ sequence identity to a polypeptide consisting of amino acids 1-438 of SEQ ID NO: 48.
6. The isolated polynucleotide of claim 1 , wherein said polypeptide of b) has at least $95 \%$ sequence identity to a polypeptide consisting of amino acids 1-290 of SEQ ID NO: 22.
7. The isolated polynucleotide of claim 1, wherein said polypeptide of b) has at least $99 \%$ sequence identity to a polypeptide consisting of amino acids 1-290 of SEQ ID NO: 22.
8. A purified polypeptide selected from the group consisting of:
a) a polypeptide comprising amino acids 1-290 of SEQ ID $\mathrm{NO}: 22$; and
b) a polypeptide having at least $90 \%$ sequence identity to a polypeptide comprising amino acids 1-290 of SEQ ID $\mathrm{NO}: 22$ and having an isoprenyl transferase activity; and
c) a polypeptide encoded by a polynucleotide, the complement of which hybridizes under stringent conditions to a polynucleotide encoding a polypeptide comprising amino acids 1-290 of SEQ ID NO: 22, and having an isoprenyl transferase activity.
9. A purified polypeptide comprising amino acids 1-290 of SEQ ID NO: 22.
10. The purified polypeptide of claim 8 , wherein said polypeptide of $b$ ) has at least $95 \%$ identity to a polypeptide comprising amino acids 1-290 of SEQ ID NO: 22.
11. An expression vector comprising a polynucleotide of claim 1.
12. The expression vector of claim 11, wherein said polynucleotide encodes a polypeptide having at least $90 \%$ sequence identity to a polypeptide comprising amino acids 1-438 of SEQ ID NO: 48 and having adenylating amide synthetase activity.
13. The expression vector of claim 11, wherein said polynucleotide encodes a polypeptide having at least $90 \%$ sequence identity to a polypeptide comprising amino acids 1-290 of SEQ ID NO: 22, and having isoprenyl transferase activity.
14. An isolated host cell transformed with an expression vector of claim 11.
15. The isolated host cell of claim 14 , wherein said host cell is a bacterial host cell.
16. A method for producing a farnesyl dibenzodiazepinone compound, comprising:
a) providing a prokaryote transformed with an expression vector of claim 11; and
b) culturing the prokaryote under conditions such that (i) an adenylating amide synthetase or an isoprenyl transferase is expressed, and (ii) a farnesyl dibenzodiazepinone compound is synthesized.
17. The method of claim 16 , wherein said prokaryote is $E$. coli.
18. The method of claim 16, wherein said prokaryote is an actinomycete.
19. An isolated polynucleotide encoding:
a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOS: 2,4, $6,8,10,12,14,16,18,20,22,24,26,28,30,32,34,36$, $38,41,42,44,46,48,50,52,54,56,58,60,62,65,67$, $69,70,71,74,76,78,80,82,84,86$ and 88 ; or
b) a polypeptide having at least $85 \%$ sequence identity to a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOS: 2,4,
$6,8,10,12,14,16,18,20,22,24,26,28,30,32,34,36$, $38,41,42,44,46,48,50,52,54,56,58,60,62,65,67$, $69,70,71,74,76,78,80,82,84,86$ and 88 , and having the same biological function as the corresponding polypeptide.
20. A cosmid selected from the group consisting of cosmid 046KM deposited under IDAC accession no. 250203-06 and cosmid 046KQ deposited under IDAC accession no. 25020307.
21. The cosmid of claim 20 , wherein said cosmid is inserted into a prokaryotic host for expressing a product.


[^0]:    $<210\rangle$ SEQ ID NO 47
    <211> LENGTH: 762
    <212> TYPE: DNA

