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(54) Title: FGF-2 DERIVED PROTEINS FOR THE PREPARATION OF BIOMATERIALS OR MEDICAL DEVICES SUCH AS STENTS

(57) Abstract: The invention relates to the use of basic fibroblast growth factor (FGF-2) derived proteins being chosen among FGF-2 mutants which are unable to interact specifically with the Transloklin, for the preparation of biomaterials or devices chosen among medical prostheses, such as endovascular stems, or bypass grafts, said FGF-2 derived proteins being able to accelerate the endothelialization of said medical prostheses and/or the reendothelialization of injured blood vessels and/or generating an endothelium with macroscopic characteristics similar to a healthy endothelium and in the meantime minimizing restenosis of said medical prostheses or blood vessels.



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FGF-2 DERIVED PROTEINS FOR THE PREPARATION OF BIOMATERIALS OR MEDICAL DEVICES SUCH AS STENTS

5 The present invention relates to the use of basic fibroblast growth factor (FGF-2)
derived proteins for the preparation of biomaterials or devices chosen among medical
prostheses, such as endovascular stents, or bypass grafts, said FGF-2 derived proteins
being able to accelerate the endothelialization of said medical prostheses and/or the
reendothelialization of injured blood vessels and/or generating an endothelium with
10 macroscopic characteristics similar to a healthy endothelium and in the meantime
minimizing restenosis of said medical prostheses or blood vessels.

 Despite therapeutic progress, the cardiovascular diseases remain the first cause of
the mortality in the western countries. This progress has particularly concerned the
surgical cardiology with the angioplasty and the thrombolysis as the current pillars in the
15 treatment of the atherosclerosis. The last twenty years, these therapeutic strategies have
allowed a reduction about 50 % of the mortality of the coronary disease treated in a
hospital environment.

 A percutaneous transluminal angioplasty (PTA) intervention consists in introducing
a balloon catheter along a guidewire via a large artery of the lower or upper limbs to the
20 occluded artery. Once positioned at the level of the occlusion, the balloon is inflated
which presses the atherosclerotic plaque into the arterial wall. This results in an
immediate widening of the arterial lumen and restoration of the arterial caliber. However,
in the months following the intervention a renarrowing (restenosis) occur as a common
($\geq 50\%$) complication. This is most often due to a constrictive remodeling of the arterial
25 wall or the formation of a neointima (Bennett and O'Sullivan, 2001; Van Belle, *et al.*,
1998).

 The narrowing of the vessel lumen due to constrictive remodeling (primarily the
consequence of an inflammatory reaction of the adventitia) can be prevented by the
implantation of an endovascular prosthesis (stent) which mechanically prevents the
30 constrictive reshaping (Fischman, *et al.*, 1994; Serruys, *et al.*, 1994).

 The stent implantation does not however prevent the formation of the neointima,
which is the other important factor provoking restenosis. The neointima is constituted by
proliferating smooth muscle cells originating from the media and extracellular matrix

components secreted from these cells. This hyperplastic response is part of the wound-healing reaction. The angioplasty intervention causes a denudation of the intimal monolayer of endothelial cells. The denuded surface is highly thrombogenic and stimulates the adhesion and aggregation of blood platelets, that upon activation secrete several mitogens and cytokines (George, 2000). Moreover, the inflation of the balloon causes a mechanical stretching of the vessel and rupture of the internal elastic lamina, exposing the media to circulating mitogens, triggering the migration and proliferation of smooth muscle cells. The capacity of these cells to migrate through the struts of the stent is a main cause of the intra-stent restenosis. To prevent this process, the development of stents covered with biocompatible polymers allowing the local liberation of pharmacological agents (Regar, *et al.*, 2001). Among the most promising drugs are two antimitogenic compounds Paclitaxel (Taxol™) and Rapamycin (Sirolimus™) that are able to limit the intrastent restenosis (Degertekin, *et al.*, 2002; Heldman, *et al.*, 2001; Morice, *et al.*, 2002) . However, it was shown that this antimitogenic treatment local inhibit the process of reendothelialization (Farb, *et al.*, 2001), which is important since the intact endothelium represents the best protection towards subsequent thrombosis at the level of the stent. An improvement of a local treatment of the atherosclerotic plaque therefore requires a stimulation of the regeneration of an endothelium with properties as close as possible those of the normal endothelium. The reendothelialization is, indeed, an essential mechanism for the inhibition of the neo-intimal hyperplasia.

The angiogenic factors possess activities which stimulate the reendothelialization: The VEGF (vascular endothelial cell growth factor) is a growth factor produced by a large number of cell types (notably in response to a hypoxia) which stimulates the migration and the proliferation of endothelial cells. However, the VEGF increases the endothelial permeability as well as the deposit of lipids and therefore favors the progression of the atherosclerotic plaque (Celletti, *et al.*, 2001), which limits the potential interest of this factor.

The fibroblast growth factors (FGFs) constitute a family of at least 22 homologous proteins (Nakatake, *et al.*, 2001; Ornitz and Itoh, 2001). FGFs act on a variety of cells by stimulating mitogenesis and/or by inducing morphological changes and differentiation. FGF-2 or basic FGF, one of the first described members of the FGF family, is involved in developmental processes, wound healing, and angiogenesis as well as in tumor progression (Bikfalvi, *et al.*, 1997; Mason, 1994; Yamaguchi and Rossant, 1995). Among these factors, the FGF-1 (or acidic FGF) but especially the FGF-2 (or basic FGF) are

known for their activities on the cells of the vascular wall. The FGF-2 is, with the VEGF, the most powerful angiogenic factor. It stimulates tubulogenesis, that is, the formation of capillaries and stimulates the proliferation and the migration of endothelial but also smooth muscular cells. Five FGF-2 isoforms of 18, 22, 22.5, 24, and 34 kDa are synthesized through an alternative translational initiation process (Arnaud, *et al.*, 1999; Florkiewicz and Sommer, 1989; Prats, *et al.*, 1989). These isoforms differ only in their NH2 extremities, which confer a nuclear localization to the four high molecular mass (HMM) CUG-initiated forms while the smaller AUG-initiated protein of 18 kDa is predominantly cytoplasmic or excreted and stored in the extracellular matrix (Bugler, *et al.*, 1991; Quarto, *et al.*, 1991). These FGF-2s can exert their effects through different pathways. The nuclear HMM forms of FGF-2 act in an intracrine manner that can influence tumor progression and promote cell proliferation under low-serum conditions (Arese, *et al.*, 1999; Bikfalvi, *et al.*, 1995; Couderc, *et al.*, 1991; Okada-Ban, *et al.*, 1999). The AUG-initiated 18 kDa protein is responsible for the auto-/paracrine action of FGF-2 and stimulates cellular proliferation and migration via interaction with (Bikfalvi, *et al.*, 1995) high-affinity transmembrane tyrosine kinase receptors (FGFR) and low-affinity receptors (heparan sulphate-containing proteoglycans) (Johnson and Williams, 1993). Ligand activation of the FGFR causes autophosphorylation of the receptor and the activation of intracellular signaling pathways, in which mitogen-activated protein kinases (MAPK) and/or phospholipase C are important mediators (Cross and Claesson-Welsh, 2001; Friesel and Maciag, 1995). FGF-2 can internalize with both kinds of receptors into the cytoplasm (Gleizes, *et al.*, 1995; Reiland and Rapraeger, 1993; Roghani and Moscatelli, 1992) and translocate into the nucleus during the G1 phase of the cell cycle (Baldin, *et al.*, 1990) by an unknown mechanism distinct from that of HMM FGF-2 (Patry, *et al.*, 1994).

Over the past ten years, accumulating data have suggested that, surprisingly, nuclear targeting and action of growth factors and growth factor receptors constitute a complementary signaling pathway involved in the induction of cell proliferation. In different target cells, nuclear association has been demonstrated for a large number of secreted growth factors or hormones such as FGF, Epidermal Growth Factor (EGF), Platelet Derived Growth Factor (PDGF), Nerve Growth Factor (NGF) and insulin (Jans, 1994; Keresztes and Boonstra, 1999; Mason, 1994; Stachowiak, *et al.*, 1997). Several findings indicate that the nuclear translocation of extracellularly acting FGFs may also be required for the mitogenic effect in different cell types (Imamura, *et al.*, 1990; Joy, *et al.*,

1997; Wiedlocha, *et al.*, 1994). Even though FIBP (Kolpakova, *et al.*, 1998) and Casein Kinase 2 (Bailly, *et al.*, 2000; Skjerpen, *et al.*, 2002) could be mediators of these signals of FGF-1 and/or FGF-2, the details regarding the molecular mechanisms by which the FGFs exert their intracellular effects remain to be elucidated.

5 The three HMM and the 18 kDa intracellular FGFs are found as components of large protein complexes of 320 and 130 kDa, respectively (Patry, *et al.*, 1997).

 In an attempt to identify proteins associated to such complexes, a human placenta cDNA library was screened in the two-hybrid system. A previously unknown protein, named Translokin by the inventors, was characterized which is able to bind 18 kDa
10 FGF-2 in a specific and direct way.

 In the frame of the present invention, the inventors give the demonstration that Translokin co-immunolocalizes with β -tubulin, and by using an FGF-2/FGF-1 chimeric strategy and the inhibition of Translokin expression with a siRNA duplex, that Translokin mediates the translocation of FGF-2 to the nucleus and/or its vicinity.

15 The present invention relies mainly on the demonstration made by the inventors that FGF-2 derived proteins which are unable to interact specifically with Translokin, are defective in stimulating the proliferation of endothelial cells, but their tubulogenesis activity is preserved.

 Thus, the main aim of the present invention is to provide new proteins which can
20 be coated to medical devices, such as endovascular prostheses (stents), in order to promote the endothelialization of said medical devices and the reendothelialization of the vessel in the vicinity of the stent by preserving the tubulogenic and migrating capacity of endothelial cells without stimulating their proliferation.

 The present invention relates to the use of animal or human basic fibroblast
25 growth factor (FGF-2) derived proteins for the preparation of biomaterials or medical devices chosen among endovascular prostheses, such as stents and bypass grafts, or coated endoprotheses, or other kinds of medical prostheses, said FGF-2 derived proteins being chosen among FGF-2 mutants which are unable to interact specifically with the Translokin represented by SEQ ID NO: 2, i.e. having a binding capacity to
30 said Translokin of less than 40%, and more preferably of less than 10%, said FGF-2 mutants being such that they are defective in intracellular trafficking, and in stimulating the proliferation of endothelial cells, and that their tubulogenesis activity is preserved.

The binding capacity to said Translokin mentioned above, and which in the case of the FGF-2 mutants defined above is less than 40%, and more preferably less than 10%, is measured with comparison to bovine FGF-2 in the yeast two-hybrid system as detailed in the material and methods section hereafter.

5 By the expression "defective in intracellular trafficking" mentioned above, it should be understood that FGF-2 mutants according to the invention are unable to translocate from the cell surface to the nuclear compartment. Such property can be measured by subcellular fractionation of fibroblasts (such as NIH 3T3) or endothelial cells (such as ABAE) treated with HA-tagged FGF-2 mutants. Such property can also
10 be assessed by immunocytochemistry of said cells. Detailed examples of these techniques are given in the material and methods section below.

The defective in stimulating the proliferation of endothelial cells property of the FGF-2 mutants mentioned above, can be evaluated by methods measuring cell proliferation, such as cell counting, incorporation of radioactive [H^3]thymidine, MTT
15 assay or other techniques used to assess cell proliferation, of cells treated with FGF-2 mutants. A detailed example of such a technique is given in the material and methods section below.

The tubulogenesis activity of the FGF-2 mutants mentioned above, can be evaluated by methods measuring the capacity of endothelial cells to form capillary
20 structures. One such method is described in the material and methods section below.

FGF-2 mutants according to the invention are able to stimulate the endothelialization of said medical devices, or the reendothelialization of a fully or partially deendothelialized blood vessel, said FGF-2 mutants being able to diminish the frequency of restenosis intra or extrastent.

25 Furthermore, FGF-2 mutants according to the invention are able to regenerate an endothelium with functional or morphological characteristics close to a normal endothelium. Such characteristics could be the capacity to produce NO, as described in (Darblade, *et al.*, 2002). The morphology of the endothelium could be assessed by silver nitrate staining as described in the material and methods section below.

30 The invention relates more particularly to the use as defined above, of FGF-2 mutants corresponding to bovine FGF-2 represented by SEQ ID NO: 4, or human FGF-2 represented by SEQ ID NO: 6, or corresponding to any other animal FGF-2 showing

83 to 99 % of identical amino acids with SEQ ID NO: 4 or SEQ ID NO: 6, and wherein the region delimited by the amino acids located from positions 40 to 130 contains at least one mutation by deletion, or substitution by a heterologous amino acid natural or not, of at least one amino acid of said region, or by insertion of at least one heterologous amino acid natural or not in said region.

The invention concerns more particularly the use as defined above, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6, wherein the region delimited by the amino acids located from positions 40 to 75, and/or the region delimited by the amino acids located from positions 105 to 130, contain at least one mutation as defined above.

The invention also concerns the use as defined above, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6, wherein the region delimited by the amino acids located from positions 44 to 61 contains at least one mutation as defined above.

In this respect, the invention relates to the use as defined above, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6, wherein at least one of the amino acids located in positions 44, 48, 52, 54, 55, 58, 61, 65, 68, 69, 71, and 73, is mutated.

The invention relates more particularly to the use as defined above, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6, and comprising at least one of the following mutations : H44L, R48T, V52T, E54D, K55R, P58Q, K61Q, Q65S, E68S, R69V, V71E, and S73Y, said mutants being represented by SEQ ID NO: 8, or by SEQ ID NO: 20, respectively.

The invention also relates more particularly to the use as defined above, of the following FGF-2 mutants :

- the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations R48T, V52T, and E54D, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 26, or by SEQ ID NO: 28, respectively,

- the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations H44L,

R48T, V52T, and K55R, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 30, or by SEQ ID NO: 32, respectively,

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations H44L, R48T, V52T, and E54D, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 34, or by SEQ ID NO: 36, respectively,

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations Q65S, E68S, R69V, V71E, and S73Y, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 38, or by SEQ ID NO: 40, respectively.

The invention concerns more particularly the use as defined above, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6, wherein at least the amino acids in positions 54 and 55 are mutated, such as FGF-2 mutants comprising the mutations E54D and K55R.

In this respect, the invention relates more particularly to the use as defined above, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6, wherein at least the amino acids in positions 54 and 55 are mutated, and wherein at least one of the amino acids located in positions 44, 48, 52, 58, and 61, is mutated, such as FGF-2 mutants comprising the mutations E54D and K55R, and at least one of the following mutations H44L, R48T, V52T, P58Q, and K61Q, and more particularly :

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations E54D, K55R, and H44L, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 42, or by SEQ ID NO: 44, respectively,

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations E54D, K55R, H44L, and R48T, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 46, or by SEQ ID NO: 48, respectively,

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations E54D,

K55R, H44L, R48T, and V52T, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 50, or by SEQ ID NO: 52, respectively,

the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations E54D, K55R, H44L, R48T, V52T, P58Q, and K61Q, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 54, or by SEQ ID NO: 56, respectively.

The invention more particularly concerns the use as defined above, of FGF-2 mutants corresponding to chimeric FGF-1/FGF-2 proteins.

In this respect, the invention concerns the use as defined above, of the following FGF-2 mutants :

- Nb1a2, represented by SEQ ID NO: 10, corresponding to a chimeric FGF-1/FGF-2 protein wherein the amino acids in positions 1 to 43 and 79 to 155 belong to bovine FGF-2 represented by SEQ ID NO: 4, and the amino acids located in positions 44 to 78 belong to human FGF-1 represented by SEQ ID NO: 14,

- Nb1a1.6, represented by SEQ ID NO: 12, corresponding to a chimeric FGF-1/FGF-2 protein wherein the amino acids in positions 1 to 43 belong to bovine FGF-2 represented by SEQ ID NO: 4, the amino acids located in positions 44 to 61 belong to human FGF-1 represented by SEQ ID NO: 14, and the amino acids located in positions 62 to 155 belong to human FGF-2 represented by SEQ ID NO: 6.

The invention also concerns the use as mentioned above, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6, wherein the region delimited by the amino acids located from positions 109 to 126 contains at least one mutation as defined above.

The invention concerns more particularly the use as defined above, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6, wherein at least one of the amino acids located in positions 109, 111, 116, 118, 120, 121, 122, 124, and 126, is mutated.

In this respect, the invention concerns more particularly the use as defined above, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, and comprising at least one of the following mutations : S109E, N111H, R116T, R118K, Y120H/A/E, S/T121A/E/K, S122E/K/N, Y124F, and A126G, said mutants being represented by SEQ ID NO: 16, or by SEQ ID NO: 22, respectively.

The invention relates more particularly to the use as defined above, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, and comprising the following mutations : S109E, N111H, Y120H/A/E, S/T121A/E/K, S122E/K/N, Y124F, and A126G, said mutants being represented by SEQ ID NO: 58, or by SEQ ID NO: 60, respectively.

The invention concerns more particularly the use as defined above, of the FGF-2 mutant represented by SEQ ID NO: 18, corresponding to a chimeric FGF-1/FGF-2 protein wherein the amino acids in positions 1 to 108 and 127 to 155 belong to human FGF-2 represented by SEQ ID NO: 6, and the amino acids located in positions 109 to 126 belong to human FGF-1 represented by SEQ ID NO: 14.

The invention also concerns the FGF-2 derived proteins chosen among the following FGF-2 mutants :

- FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, and comprising at least one of the following mutations : H44L, R48T, V52T, E54D, K55R, P58Q, K61Q, Q65S, E68S, R69V, V71E, and S73Y, said mutants being represented by SEQ ID NO: 8, or by SEQ ID NO: 20, respectively, and more particularly:

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations R48T, V52T, and E54D, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 26, or by SEQ ID NO: 28, respectively,

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations H44L, R48T, V52T, and K55R, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 30, or by SEQ ID NO: 32, respectively,

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations H44L, R48T, V52T, and E54D, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 34, or by SEQ ID NO: 36, respectively,

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the

mutations Q65S, E68S, R69V, V71E, and S73Y, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 38, or by SEQ ID NO: 40, respectively,

- FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, wherein at least the amino acids in positions 54 and 55 are mutated, such as FGF-2 mutants comprising the mutations E54D and K55R,

- FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, wherein at least the amino acids in positions 54 and 55 are mutated, and wherein at least one of the amino acids located in positions 44, 48, 52, 58, and 61, is mutated, such as FGF-2 mutants comprising the mutations E54D and K55R, and at least one of the following mutations H44L, R48T, V52T, P58Q, and K61Q, and more particularly :

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations E54D, K55R, and H44L, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 42, or by SEQ ID NO: 44, respectively,

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations E54D, K55R, H44L, and R48T, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 46, or by SEQ ID NO: 48, respectively,

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations E54D, K55R, H44L, R48T, and V52T, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 50, or by SEQ ID NO: 52, respectively,

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations E54D, K55R, H44L, R48T, V52T, P58Q, and K61Q, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 54, or by SEQ ID NO: 56, respectively,

- Nb1a2, represented by SEQ ID NO: 10, corresponding to a chimeric FGF-1/FGF-2 protein wherein the amino acids in positions 1 to 43 and 79 to 155 belong to bovine FGF-2 represented by SEQ ID NO: 4, and the amino acids located in positions 44 to 78 belong to human FGF-1 represented by SEQ ID NO: 14,

- Nb1a1.6, represented by SEQ ID NO: 12, corresponding to a chimeric FGF-1/FGF-2 protein wherein the amino acids in positions 1 to 43 belong to bovine FGF-2 represented by SEQ ID NO: 4, the amino acids located in positions 44 to 61 belong to human FGF-1 represented by SEQ ID NO: 14, and the amino acids located in positions
5 62 to 155 belong to human FGF-2 represented by SEQ ID NO: 6,

- FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, and comprising at least one of the following mutations : S109E, N111H, R116T, R118K, Y120H/A/E, S/T121A/E/K, S122E/K/N, Y124F, and A126G, said mutants being represented by SEQ ID NO: 16, or by SEQ ID NO: 22,
10 respectively,

- FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, and comprising the following mutations : S109E, N111H, Y120H/A/E, S/T121A/E/K, S122E/K/N, Y124F, and A126G, said mutants being represented by
15 SEQ ID NO: 58, or by SEQ ID NO: 60, respectively,

- the FGF-2 mutant represented by SEQ ID NO: 18, corresponding to a chimeric FGF-1/FGF-2 protein wherein the amino acids in positions 1 to 108 and 127 to 155 belong to human FGF-2 represented by SEQ ID NO: 6, and the amino acids located in positions 109 to 126 belong to human FGF-1 represented by SEQ ID NO: 14.

The invention also concerns the nucleotide sequences encoding FGF-2 derived
20 proteins as defined above.

The invention also relates to the vectors, such as plasmids, containing the nucleotide sequences as mentioned above.

The invention also concerns the host cells, such as *Escherichia coli* or other unicellular organisms, transformed with vectors as defined above.

The invention also relates to a process for the preparation of FGF-2 derived
25 proteins as defined above, comprising the culturing of host cells in an appropriate medium, and the purification of said FGF-2 derived proteins produced.

The invention also relates to biomaterials or medical devices coated with animal or human basic fibroblast growth factor (FGF-2) derived proteins such as defined
30 above.

The invention relates more particularly to biomaterials or medical devices as mentioned above, coated with basic fibroblast growth factor (FGF-2) derived proteins

as defined above, said biomaterials or medical devices being chosen among endovascular prostheses, such as stents used for inhibiting restenosis or stenosis following angioplasty.

For illustration purpose, such endovascular prostheses and methods for coating proteins thereto, are more particularly described in WO 9749434, or are those currently used in the art. The compounds used for the coating of the prostheses should preferentially permit a controlled release of the FGF-2 derived proteins. Said compounds could be polymers (such as sutures, polycarbonate, Hydron, and Elvax), biopolymers/biomatrices (such as alginate, fucans, collagen-based matrices, heparan sulfate) or synthetic compounds such as synthetic heparan sulfate-like molecules or combinations thereof (Davies, *et al.*, 1997; Desgranges, *et al.*, 2001; Dixit, *et al.*, 2001; Ishihara, *et al.*, 2001; Letourneur, *et al.*, 2002; Tanihara, *et al.*, 2001; Tassiopoulos and Greisler, 2000).

The invention also relates to a process for the screening of FGF-2 mutants according to the invention, said process comprising :

- measurement of the binding capacity of said FGF-2 mutants to Translokin mentioned above, for example in a two-hybrid system as detailed hereafter, and selection of those mutants having less than 40%, and more preferably less than 10%, of binding capacity to Translokin when compared to bovine FGF-2,

- measurement of the property of the FGF-2 mutants selected in the previous step of being unable to translocate from the cell surface to the nuclear compartment ; such property could be measured by subcellular fractionation of fibroblasts (such as NIH 3T3) or endothelial cells (such as ABAE) treated with HA-tagged FGF-2 mutants ; this could also be done using other cell types for which FGF-2 is a mitogen. Such property could also be assessed by immunocytochemistry of said cells. Detailed examples of these techniques are given in the material and methods section below,

- measurement of the property of the FGF-2 mutants selected in the previous steps of being defective in stimulating the proliferation of endothelial cells ; such property could be evaluated by methods measuring cell proliferation (such as cell counting, incorporation of radioactive [H^3]thymidine, MTT assay or other techniques used to assess cell proliferation) of cells treated with FGF-2 derived mutants ; a detailed example of such a technique is given in the material and methods section below,

The tubulogenesis activity of the FGF-2 mutants mentioned above, could be evaluated by methods measuring the capacity of endothelial cells to form capillary structures. One such method is described in the material and methods section below.

The invention is further illustrated with the detailed description which follows of mutants according to the present invention, and their biological properties.

RESULTS

Isolation of a novel FGF-2-interacting protein

A yeast two-hybrid method was used to identify intracellular proteins that interact with FGF-2. Using the 18 kDa FGF-2 isoform as bait we screened a human placental complementary DNA library (Clontech). Four positive and distinct clones similar to the KIAA0092 mRNA (Nagase, *et al.*, 1995) were isolated, all encoding the major part of a previously non-characterized protein (Fig. 1). After a more substantial characterization of its function (see below), the protein was named Translokin. The full-length cDNAs encoding human, mouse and bovine Translokin were amplified, cloned and sequenced using either Genebank (human) or EST data (mouse, bovine). The open reading frames encode 500, 500 and 499 amino acids respectively (Fig. 1). The three predicted proteins share an overall sequence identity of 88%, indicating a high degree of evolutionary conservation. To ascertain the specificity of the two-hybrid assay for the observed interaction between Translokin and the 18 kDa FGF-2, Translokin was coexpressed in yeast with a panel of representative FGF-family members (FGF-1, FGF-3, FGF-6, FGF-12), as well as the larger 24 kDa FGF-2. As shown in figure 2a, among the FGFs tested, Translokin was only able to interact with the 18 kDa FGF-2. Surprisingly, Translokin did not interact with the 24 kDa FGF-2, suggesting that the N-terminal extension of this FGF-2 isoform creates a sterical obstacle blocking the interaction at least in its fusion state in yeast. $\alpha\alpha$ homotypic interaction was detected between Translokin molecules indicating its ability to form homodimers. The strength of the Translokin/FGF-2 interaction is noticeable since Translokin interacted with FGF-2 with similar efficiency as the positive control provided by the manufacturer (SV-40 large-T antigen interaction with p53). (Van den Berghe, *et al.*, 2000). To ascertain that the Translokin/FGF-2 interaction was direct and independent of a third partner, the two proteins were expressed in *E.coli* and purified. The interaction between the purified proteins was tested in an *in vitro* ELISA assay. As shown in figure 2b, Translokin and FGF-2 interact strongly in a direct

and dose-dependent manner. This assay was also used to verify *in vitro* the results obtained in the two-hybrid assay concerning the specificity of the interaction between 18 kDa FGF-2 and Translokin. For this, the interaction between Translokin and FGF-1 or 24 kDa FGF-2 was tested, since these two proteins are the most closely related to 18 kDa FGF-2. The lack of interaction between Translokin and FGF-1 was confirmed (figure 2c). However, the interaction obtained between Translokin and 24 kDa FGF-2 was slightly greater than the one obtained in yeast, but remained low compared to the 18 kDa FGF-2/Translokin interaction. To further study the FGF-2/Translokin interaction, co-immunoprecipitation experiments were performed on NIH-3T3 cells, which are known to internalize exogenously added FGF-2 (Bailly, *et al.*, 2000). NIH-3T3 cells, transfected with a plasmid encoding haemagglutinin A (HA)-tagged Translokin, were incubated with or without His-tagged FGF-2 (18 kDa). Immunoprecipitation of Translokin, with an anti-HA antibody, co-precipitated His-FGF-2 as detected by Western-blot analysis of the precipitates with an anti-His antibody (Fig. 2d). The same results were obtained by immunoprecipitation of endogenous Translokin. For this, lysates of NIH-3T3 cells stimulated or not with HA-tagged FGF-2, were incubated with anti-Translokin or with a control antibody. As shown figure 2e, anti-Translokin was able to specifically co-immunoprecipitate the HA-FGF-2. Taken together, these data show that 18 kDa FGF-2 and Translokin are able to interact *in vivo* and *in vitro*.

Expression and subcellular localization of Translokin

The distribution of Translokin mRNA expression in human tissues was next examined by hybridizing a multi-tissue poly(A) Northern blot (Clontech) with a human Translokin cDNA probe. The expression was widespread with highest mRNA levels in heart and skeletal muscle and lowest in lung. A major 3.2 kilobase messenger-RNA species was detected in all human tissues tested as well as two other transcripts of approximately 2.6 and 2.2 kb (Fig. 3a). In accordance with these data, a 1500 bp fragment corresponding to a unique ORF in all human cell lines examined has been detected by RT-PCR (Fig. 3b). A polyclonal antibody against Translokin, developed in the laboratory, detected a protein in all cell lines tested of human, simian, murine and bovine origin, with a molecular mass of 55 kDa corresponding well with the predicted one. Taken together, these results reflect a ubiquitous expression of Translokin. The subcellular localization of Translokin was investigated in COS-7 cells, transfected with a plasmid encoding HA-tagged Translokin. The monoclonal α -HA or the polyclonal α -

Translokin antibodies revealed the same cytoplasmic distribution that entirely overlapped with the staining of microtubules obtained with an anti- β -tubulin antibody (Fig. 3c) and confirmed by the bright color in the merged pictures.

5 **The Translokin-FGF-2 interaction involves two distinct regions in FGF-2**

The inventors subsequently wanted to determine the structural elements in FGF-2 involved in the interaction between FGF-2 and Translokin, in the aim of generating a non-interacting FGF-2 mutant, which would serve as a tool for the evaluation of the physiological role of the Translokin/FGF-2 interaction.

10 The chosen strategy was based on the fact that Translokin is unable to interact with FGF-1, despite the high degree of sequence identity (53%) shared by FGF-1 and 2 and the close resemblance of their three-dimensional structures (Zhu, *et al.*, 1991). This prompted the inventors to choose a cassette-shuffling approach, thereby paying respect to the preservation of the overall structure of FGF. The chimeric FGF-1/FGF-2-proteins were fused with GAL4 DNA-binding domain and the ability of the fusion proteins to interact *in vivo* with Translokin was examined in the yeast two-hybrid system (Fig. 4a). The rationale of the nomenclature of the chimeric proteins is explained in the legend of figure 4. The expression level of the chimeric proteins was assessed by Western blotting using an anti-GAL4DBD monoclonal antibody (Fig. 4b). The results demonstrate the absolute requirement of two separate regions in order to maintain the interaction. The first region is located between β -sheets 2 and 5 (compare Na1 with Na2 or Na1b4 with Na2b4). The replacement in FGF-2 of the equivalent amino acids from FGF-1 (Nb1a2) and the reverse chimera (Na1b2) demonstrated respectively the indispensability but also the insufficiency of this region. Indeed, a second region positioned between β -sheets 8 and 10 is important in order to maintain the interaction (compare Nb2a4 with Nb2a3, Nb3 with Nb4 or Na1b3 with Na1b4). The amino acids comprised within these two regions were therefore replaced in FGF-1, by the corresponding FGF-2 sequences (amino acids 44-78 and 109-126). The fact that this chimera (Na1b2a3b4) exhibited the same capacity to interact with Translokin as FGF-2, demonstrate that the structural motifs involved in the interaction with Translokin are present within these two regions in FGF-2. In the non-interacting FGF-2 mutant, Nb1a2, the FGF-1-derived sequence is restricted to 12 amino acids in the first region. This chimera was chosen to confirm *in vitro* the results obtained in the *in vivo* two-hybrid system. For this, recombinant Nb1a2 was produced in *E. coli* and purified by heparin sepharose chromatography. The ability of the recombinant Nb1a2

protein to interact with Translokin *in vitro* was then tested in an ELISA-based assay. As shown in figure 4c, Nb1a2 has no affinity for Translokin.

Characterization of Nb1a2

5 FGF-2 is a potent mitogen for endothelial cells. Proliferation assays were thus performed with Adult Bovine Aortic Endothelial cells (ABAE), by adding different doses of recombinant FGF-2 or Nb1a2 in culture media. Compared to FGF-2, Nb1a2 exhibited a very low mitogenic activity. The maximal mitogenic effect was obtained for 2 ng/ml of FGF-2 (Fig. 5a), whereas the mitogenic potential of Nb1a2 at this concentration was
10 reduced by 90%. In stark contrast, Nb1a2 was equally efficient as FGF-2 in stimulating tube formation of endothelial cells in Matrigel™ (Fig. 5b) as well as and cell migration.

These results demonstrate that the mitogenic and differentiation activities of FGF-2 can be uncoupled by mutations affecting the FGF-2/Translokin interaction.

15 FGF-mediated activities require the binding of the growth factor to its high-affinity tyrosine kinase receptors. The binding of FGF-2 and Nb1a2 to the FGF receptors was therefore examined by comparing the ability of increasing concentrations of FGF-2 and Nb1a2 to displace [³⁵S]-labeled FGF-2, bound to ABAE cell surface receptors. As shown in figure 5c, both FGF-2 and Nb1a2 displaced the binding of FGF-2 with the same efficiency, indicating equivalent affinities for the high-affinity binding sites on the cell
20 surface. Heparan sulphate-containing proteoglycans (HSPG) are also implicated in FGF-2 mediated signalization, representing low-affinity binding sites. The competition experiments for these sites showed similar results, which is in agreement with the affinity profile towards heparin-sepharose chromatography (Fig. 5d). Moreover, it is well known that phosphorylation and activation of mitogen-activated protein kinases Erk1 and Erk2
25 are required for the mitogenic response induced by FGF-2 (Cross and Claesson-Welsh, 2001; Pages, *et al.*, 1993). By immunoblotting experiments with antibodies raised against phosphorylated forms of Erk1 and 2, it has been observed that FGF-2 and Nb1a2 were both able to activate p42 and p44 in ABAE cells for comparable time lengths (Fig. 5e). Between 4 and 12 hours, the Erk-phosphorylation was low, similar and decreased with
30 the same kinetics in both cases. Activation of p70S6 kinase (a serine/threonine kinase that phosphorylates the ribosomal protein S6 in response to a number of extracellular stimuli) is required for FGF-stimulated cell proliferation but not differentiation of endothelial cells (Kanda, *et al.*, 1997). As for Erk1/2 no difference between FGF-2 and Nb1a2 was found when the phosphorylation status of p70S6K by immunoblotting was assessed.

These results show that Nb1a2 is able to activate the main signaling pathways involved in the mitogenic response induced by FGF-2.

Internalized FGF-2 but not Nb1a2 is associated with the nucleus

5 Exogenously added FGF-2 translocates to the nucleus of proliferating cells (Baldin, *et al.*, 1990; Bouche, *et al.*, 1987; Clarke, *et al.*, 2001; Hawker and Granger, 1992; Reilly and Maher, 2001; Walicke and Baird, 1991) and this nuclear translocation is associated with a proliferative state of the treated cells (Baldin, *et al.*, 1990; Joy, *et al.*, 1997). The inventors therefore tested whether exogenously added Nb1a2 retained the ability of FGF-
10 2 to be internalized from the outside of the cell to the cytoplasm and further translocated to the nucleus. For this purpose, 50% confluent ABAE cells, rendered quiescent after 48h under serum-free conditions, were incubated for 2 h with FGF-2 or Nb1a2. To discriminate from endogenous and exogenous FGF-2, HA-tagged proteins were used. After cell harvesting and fractionation, the respective amounts of HA-FGF-2 or HA-
15 Nb1a2 in the cytoplasmic and nuclear fractions were determined by immunoblotting with an α -HA antibody. As shown in figure 6a, both HA-FGF-2 and HA-Nb1a2 were detected in the cytoplasmic fraction. However, only the HA-FGF-2 was found in the nuclear fraction. These results show that mutations disturbing the interaction between FGF-2 and Translokine lead to a loss of mitogenicity. This could be explained by the inability of the mutant Nb1a2 to be translocated to the nucleus and/or its vicinity. To verify these results,
20 immunofluorescence studies and confocal analysis of the intracellular trafficking of exogenous FGF-2 were performed.

The data, presented in figure 6b (panels B, D), show the distribution of internalized HA-FGF-2 in ABAE cells after two hours incubation at 37°C. The growth factor
25 appeared as intracellular dots, indicating uptake in vesicles, with a prominent perinuclear FGF-2 staining and a relatively sparse FGF-2 staining within the nucleus. The staining obtained with α -HA was specific as no staining was observed when the growth factor was omitted (Fig. 6b panel A). In agreement with the data obtained by cell fractionation (Fig. 6a), no nuclear-associated staining was observed when cells were incubated with
30 HA-Nb1a2 (Fig. 6b panel F). A double-labeling experiment with anti- β -tubulin, resulted in a good overlap of β -tubulin and HA-FGF-2, concentrated in the centrosome area. To verify if the intracellular transport of exogenous FGF-2 was dependent on microtubules, experiments with depolymerizing drugs were carried out. As shown in figure 6b (E),

cytochalasin B, which induces depolymerization of actin microfilaments (compare the actin filament distribution in panels D and E), did not alter the distribution of the growth factor in the nuclear region. On the contrary, treatment with nocodazole, which causes depolymerization of microtubules (compare the microtubule organization in panel B with the one in panel E), prevented the appearance of the growth factor in the juxtannuclear region (panel C). In these cells the punctuated FGF-2 staining was found in the cytoplasm, especially in the vicinity of the plasma membrane. These results strongly suggest that the intracellular trafficking of exogenous 18 kDa FGF-2 is mediated by microtubules in ABAE cells and correlates with the microtubule-associated pattern of Translokine observed by immunocytochemistry (Fig. 3c).

Nuclear association of internalized FGF-2 is essential for its mitogenic activity and is mediated by Translokine

The inventors reasoned that if nuclear translocation was important for mitogenic activity of FGF-2, it may be possible to restore the mitogenic activity of Nb1a2 by the addition of a well characterized nuclear localization sequence (NLS). Therefore, chimeric polypeptides NLS Nb1a2 and NLS FGF as a control, containing the NLS derived from the SV-40 T antigen (PKKKRKRV) at the NH₂-terminus, were constructed. The chimeric proteins were produced in *Escherichia coli* and purified by heparin sepharose chromatography. Both proteins exhibited the same affinity as FGF-2 for immobilized heparin. They were then tested for their mitogenic activity in an *in vitro* proliferation assay using ABAE cells. As shown in figure 7a, the mitogenic activities of FGF-2 and NLS-FGF-2 were comparable. However, the addition of the NLS to Nb1a2 strongly enhanced its mitogenic activity, which reached approximately 60% of the one obtained with WT FGF-2. A fractionation experiment performed on NIH-3T3 cells stimulated or not with NLS-FGF-2 or NLS-Nb1a2, confirmed the ability of these growth factors to be translocated to the nuclear region (Fig. 7b). These results show that the nuclear association of FGF-2 is a critical step for its mitogenic activity.

To further investigate whether Translokine was the mediator for this translocation, NIH-3T3 cells were transfected with small interfering RNA (siRNA) duplexes (Elbashir, *et al.*, 2001) to reduce the expression of endogenous Translokine. The inventors performed a single transfection with a siRNA targeting Translokine or with a scrambled siRNA as control. After 55h of transfection, cells were exposed to 20 ng/ml of HA-tagged FGF-2 for 2h. The cells were then collected and fractionated in order to study the respective

amounts of FGF-2 in the cytoplasmic and nuclear fractions. Since the translocation of FGF-2 is dependent on the cell cycle and on the confluence state of the cells (Baldin, *et al.*, 1990), great care was taken when seeding the cells. In accordance with the putative role of Translokin in FGF-2-induced proliferation, NIH-3T3 cells transfected with Translokin-targeted siRNA grew more slowly than cells transfected with control siRNA. These cells were therefore seeded at a slightly higher density in order to obtain exactly the same number of cells in both conditions at the time the HA-FGF-2 was added. After transfection the protein levels were evaluated by immunoblot analysis. As shown in figure 7c, the specific siRNA led to a decrease of about 78% of Translokin (average obtained from three independent experiments) whereas β -actin remained unaffected. Immunofluorescence studies showed that siRNA treatment did not affect the cytoskeletal organization. Interestingly, the decrease in Translokin expression by siRNA was correlated with a reduction of FGF-2 nuclear accumulation of about 80% (Fig. 7d). A diminution of the cytoplasmic HA-FGF-2 contents in cells transfected with the Translokin siRNA was also observed. However, this decrease, not exceeding 50%, was always less than the one observed for nuclear FGF-2 (Fig. 7d). This could reflect a participation of Translokin complex in the uptake or in the intracellular stability of exogenously added FGF-2. To assess the specificity of the Translokin-siRNA treatment on the FGF-2 translocation, the consequences of the decrease in Translokin expression, on FGF-1 trafficking, were tested. For this purpose, an HA-tagged FGF-1 has been produced and purified. The inventors then proceeded in the same way as for FGF-2. As shown in figure 7e, Translokin-siRNA treatment did not affect the translocation of HA-FGF-1. Taken together these results further demonstrate a direct role of Translokin in the translocation of FGF-2, a step that appears absolutely necessary for the FGF-2 mediated mitogenic activity.

Nb1a2 accelerates the reendothelialisation of injured arteries FGF-2 has been shown to stimulate the regrowth of endothelium after balloon catheter denudation of the rat carotid artery (Lindner, *et al.*, 1990). However, since FGF-2 is able to stimulate the proliferation and migration of smooth vascular cells (Jackson and Reidy, 1993; Lindner, *et al.*, 1991) and thereby also increases and aggravates the neointima after injury (Lindner and Reidy, 1991), it does not constitute a suitable therapeutic tool for arterial healing. Given the characteristics of the biological activity of the Nb1a2 *in vitro*, as described in figure 5, the FGF-2 derived proteins with a strongly reduced binding capacity to

Translokin but with maintained tubulogenic activities, could be more appropriate than wildtype FGF-2 as therapeutic agents to accelerate the arterial healing of injured arteries *in vivo*. To investigate this, an electric injury mouse model was used, which was developed by Carmeliet and coll. (Carmeliet, *et al.*, 1997) and further adapted in the inventors' laboratory (Bouchet, *et al.*, 2001). In this model, an electric pulse is applied to the distal part of the common carotid artery using a bipolar microregulator. Three days following injury, the endothelial regeneration process was evaluated by staining the denuded areas with Evans blue. As expected, 5 μ g of FGF-2 (injected in bolus just after the injury) stimulated the endothelial regrowth (30% above saline-treated controls) (Fig.8). Interestingly, Nb1a2 was as efficient in accelerating the reendothelialisation of the artery.

The formation of the neointima after vascular injury after treatment with FGF-2 or Nb1a2 was investigated in mice, sacrificed ten days after injury. Paraffin embedded sections of fixed carotids, stained with hematoxylin, were examined microscopically. Whereas FGF-2 provoked a thickening of the intima (neointima) at the level of injury, no thickening of the intima was observed in mice treated with Nb1a2. The morphology of the regenerated endothelium was compared by silver nitrate staining of carotids from mice treated with FGF2 or Nb1a2, ten days after injury, as well as from non-injured control animals. As shown in figure 9b and 9c, the shape and the orientation of the endothelial cells of the carotids in the two latter groups were homogenous, following the longitudinal axis of the artery, whereas in the animals treated with FGF-2, the shape and the orientation of the cells were much less regular.

Detailed analysis of the first Translokin-interacting region in FGF-2

As described above, two regions in FGF-2, located between β -strands 2-5 and 8-10, respectively, are important for the interaction with Translokin. As shown in figure 4a, the structural entities within the first region constitute the most critical parameter for maintaining the interaction. In this region, 12 amino acids differ between human FGF-1 and human (or bovine) FGF-2. In order to more precisely define which ones of these amino acids that are implicated in the interaction, a number of more targeted FGF-2 mutants were generated. The capacity of these mutants to interact with Translokin was subsequently assessed in the yeast two-hybrid system. The data, represented in figure 10, show the necessity to mutate several amino acids in order to reduce the affinity to Translokin to a level comparable to the one of FGF-1 or Nb1a2. Indeed, only a hepta-

amino acid mutant: HRVEKPK 44/48/52/54/55/58/61 LTIDRQQ (also referred to as Nb1a1.6) was completely void of any affinity for Translokoin. In analogy with these results, similar studies of the second Translokoin-interacting region in FGF-2, favors an importance of the entire region between β -strands 8 and 10, more than a role of the individual amino acids (data not shown).

Discussion

Over the past decade, it has become increasingly apparent that a number of peptides hormones and growth factors elicit their biological responses in a bi-functional manner, both indirectly with cell surface receptors linked to conventional signal transduction pathways, as well as through direct association with the nuclei of target cells. This growing list of peptides includes fibroblast growth factors and in particular FGF-2 and FGF-1, insulin, EGF, NGF, PDGF, prolactin, angiogenin and parathyroid hormone-related peptide (PTHrP) (Jans, 1994; Keresztes and Boonstra, 1999). Among these examples, FGF-1 and FGF-2 are prototypic for this dual pathway. For both factors the first and indispensable step is the cell surface recognition of the low and high affinity receptors. This step seems to be sufficient to exert the differentiation properties of the factor. Nevertheless, very little is known about intracellular routing of internalized FGFs, even though localization of FGF-2 has been reported in caveolae prior the endosome/lysosome pathway (Gleizes, *et al.*, 1996). Internalization and nuclear translocation of FGF-2 have been abundantly reported and in several studies associated with cell proliferation (Bailly, *et al.*, 2000; Baldin, *et al.*, 1990; Joy, *et al.*, 1997). Similar results have been reported for FGF-1 and different groups have clearly demonstrated that binding and activation of cell surface receptors are not solely sufficient for a full mitogenic response (Grieb and Burgess, 2000; Imamura, *et al.*, 1990; Wiedlocha, *et al.*, 1994), conferring an intracellular activity on internalized FGF. Some intracellular targets of FGF-2 have been previously described including Tax, FIF and CKII, (Bonnet, *et al.*, 1996; Shen, *et al.*, 1998; Van den Berghe, *et al.*, 2000). CKII has also been recently described as an essential target of the intracellular FGF-1 (Skjerpen, *et al.*, 2002). This interaction, as well as the MAPK pathway, are necessary for the proliferative effect of the factor. Furthermore, internalized FGFR1 has been found to be translocated to the nucleus in response to FGF-2 (Reilly and Maher, 2001). This could also constitute a key component of the growth factor response. Nevertheless, the coupling between the different pathways remains to be elucidated.

The inventors report the identification of human Translokine, an FGF-2 binding protein involved in the intracellular trafficking of the internalized growth factor. Translokine fails to bind to other members of the FGF family indicating that different intracellular trafficking pathways are used by these growth factors. Surprisingly, 24 kDa FGF-2 does not interact (or very poorly) with Translokine even though the binding sites are present. This could be due to structural changes induced by the presence of an arginine-rich amino-terminal extension in HMM FGF-2 isoforms. Like FGF-2, Translokine is highly conserved among mammals and is also present in *Xenopus*, which is also reflected by the perfect capacity of human Translokine to interact with *Xenopus* FGF-2 in the two-hybrid system (data not shown). Furthermore, Translokine is ubiquitously expressed and localizes with the microtubule network. A two-hybrid screening using Translokine as bait revealed that the protein is also associated with components of microtubule apparatus. These facts, together with the presence of a putative dynactin-related domain in Translokine, reinforce the hypothesis that Translokine is the carrier of internalized exogenously added FGF-2 and belongs to a motor complex. The inventors also describe an FGF-2 mutant, Nb1a2, which is unable to interact with Translokine. Nb1a2 shows uncoupling differentiation and proliferation activities which is due to a lack in its translocation towards the nucleus. This phenotype is consistent with the results reported by Kudla and colleagues (Kudla, *et al.*, 1998) who demonstrated that artificial activation of FGFR1 without FGF-2 leads to differentiation effects but not to proliferation. An hypothesis is that FGF-2 induced proliferation requires, in addition to receptor activation and sustained MAP kinases phosphorylation, an intracellular translocation of the factor to the nucleus or to the perinuclear/centrosomal compartment. Even if it is very difficult to discriminate between "nuclear" and "nucleus-associated" FGF-2, the importance of nuclear FGF-2 as a functional entity is greatly reinforced by the fact that the addition of the Large T antigen NLS to the N-terminus of Nb1a2 leads to a recovery of the mitogenic activity. Moreover, siRNA mediated inhibition of Translokine expression results in a concomitant and similar decrease of the FGF-2 nuclear translocation but has no effect on intracellular FGF-1 trafficking which is consistent with the lack of FGF-1/Translokine interaction and the inability of nocodazole to block the FGF-1 translocation (Malecki, *et al.*, 2002). The inventors also observed, to a lesser extent, a reduced cytoplasmic level of FGF-2. This could suggest that Translokine participates in the uptake of FGF-2, which is the first step of its translocation. Alternatively, Translokine could also protect cytoplasmic FGF-2 from lysosomal

degradation. Taken together these results clearly demonstrate that translocation of FGF-2 to the nucleus and/or its vicinity is an essential step in the mitogenic activity and that Translokin is a major component of this pathway. Bypass of this translocation pathway by the addition of a heterologous NLS to Nb1a2 suggests that the Translokin pathway is highly specific and efficient since this NLS-Nb1a2 is not able to allow more than 60% recovery of the WT FGF-2 mediated mitogenic activity. This specificity does not however exclude the possibility for Translokin to be also involved in the trafficking of other exogenous molecules. The identification of the molecular basis of the FGF-2 translocation machinery will be of great interest and could be a model for the numerous growth factors and peptide hormones that are intracellularly relocated to fully achieve their biological properties.

EXAMPLES

Materials and Methods

Common molecular biology techniques. Unless detailed, the different techniques used for the generation and production of the different constructions included in the invention are basic molecular cloning techniques, known to those skilled in the art. This includes amplification of the ADN by RT-PCR (reverse transcription coupled with PCR (Polymerase Chain Reaction)), PCR, digestion the DNA by restriction enzymes, separation of DNA fragments on agarose gels (1-2 %), ligations with DNA ligase and transformation of competent bacteria, purification of plasmid DNA from transformed bacteria. All constructs were verified by automatic sequencing.

RNAble (Eurobio, Les Ulis, France) was used to extract total RNA from culture cells or mouse organs. SuperScript II (Invitrogen, Groningen, The Netherlands) was used in the reverse transcription (RT) with following the manufacturer's instructions. Automatic nucleotide sequencing was performed using the AmpliTaq FS polymerase and an ABI 373A sequencer (Perkin Elmer Corp, Foster City, CA)

Cloning of FGF-2 for studies in the two-hybrid system.

The following example concerns nucleotide sequences coding for bovine FGF-2, but could concern the FGF-2 from the other mammals or amphibians, since the inventors can provide data demonstrating that human Translokin (SEQ ID NO: 2) interacts equally well with the human or bovine FGF-2 as with the FGF-2 of the amphibian *Xenopus leavis*

(SEQ ID NO: 24). The nucleotide sequence encoding bovine FGF-2 used in the invention was modified with regard to the wild type sequence to better correspond to the codon usage of bacterium *Escherichia coli*, in order to optimize the production of protein in prokaryotic expression systems. This cDNA was cloned in the laboratory in the vector pRF101. For the cloning in the vector aimed for the two-hybrid system (pAS2; Clontech) pRF101 was digested with *NcoI* and *SalI* to release the fragment coding for FGF-2 18 kDa. This fragment was then cloned in the same sites of pAS2, in frame with the DNA binding domain of the yeast transcription factor GAL4.

Cloning of FGF-1 for studies in the two-hybrid system. Human FGF-1 was amplified by RT-PCR of 5 micrograms of total RNA extracted from SK-Hep-1 cells. For the amplification the primers 5'-GGAATTCATATGGCTGAAGGGGAAAT-3' (sense) and 5'-ACGACGTCGACAGATCTCTTTAATCAGAAGAGACT-3' (anti-sense). The product was digested by *NdeI* and *SalI* and cloned in the same sites of the vector pAS2.

Generation of chimeras. For the construction of the first series of chimeras, four junctions were chosen in beta strands 2, 5, 8 and 10 where the sequence is well conserved between both FGF. The nomenclature of the chimeras refer to the origin of N-terminal region ("a" for "acidic FGF" (=FGF-1) and "b" for "basic FGF" (=FGF-2)). The numbers indicate the junctions between FGF-1 and FGF-2.

A first series of eight chimeras, named Nb1-Nb4 and Na1-Na4, was thus conceived. Nb1 was generated by replacement of a 362-base pair *BamHI/SalI* fragment of pAS2/FGF-2 by the corresponding sequence of the vector pAS2/FGF-1. This sequence corresponds in FGF to the sequence downstream the connection 1. In the same manner, the chimera Na1 was generated by the mutual replacement of the *BamHI/SalI* fragment in pAS2/FGF-1 by the one in pAS2/FGF-2. The other chimeras were constructed by a two-step PCR approach. In the first PCR, six anti-sense primers were conceived that were overlapping junctions 2,3 and 4. Taking one of two primers in the junction 2, "2-5a3b": (5'-GCCAGTCTCGGTACTCTTgatagatacaacaccacg-3') as an example. This primer was used to generate Nb2. The sequence in the 5' portion of "2-5a3b" corresponds to the FGF-1 sequence downstream from junction 2 (capital letters in the sequence above) and the sequence in the 3' portion corresponds to the sequence upstream to the junction 2 of FGF-2 (small letters in the sequence). The primer 2-5a3b was used in PCR together with the primer pAS-sense: 5'-AAGATGCCGTCACAGATAGATTG-3' using pAS2/FGF-2

as template. The purified product was then used as a "mega-primer" with a fragment *Bam*HI/*Bgl*II of pAS2 /FGF-1 as a second "mega-primer" in a PCR where the hybridization product of of two "mega-primers" constituted the template. To amplify this product, two primers flanking the FGF sequence (HA: 5 '-CCATACGATGTTCCAGATTACG-3 ' and pAS-term: 5 '-GCGACCTCATGCTATACTGAG-3 ') were used. The final PCR product was then purified, digested by *Bam*HI/*Sal*I, re-purified to finally be cloned in the same sites of pAS2/FGF-2.

Transformation of yeast cells. The method used to transform yeast cells with plasmid DNA is based on the protocol developed Gietz and coll. (Agatep, *et al.*, 1998). A single colony of non-transformed *Saccharomyces cerevisiae* Y190 cells was grown overnight at 30°C (250 rpm) in 20 ml sterile YPD medium [1% w/v yeast extract, 2% w/v peptone, 2% w/v dextrose (glucose)]. The following day the culture was centrifuged for 5 min. at 1000g, 20°C and resuspended in 20 ml YDP medium. The optical density (OD) of the culture was determined by the measurement of an aliquot (2xdilution) in a spectrophotometer at 600nm. A new culture volume (80 ml for 20 independent transformations) with an initial OD^{600nm} of 0.3 was prepared by the addition of an appropriate volume of the overnight culture (5-8 ml for 80 ml fresh YPD). The new culture was incubated at 30°C at 250 rpm until the OD^{600nm} had reached 1.2. The cells were then harvest by centrifugation in sterile 45 ml tubes in a Beckman JS-13 swing-out rotor for 10 min. at 1000g, 20°C. The cells were washed with 25-30 ml H₂O and recentrifuged as above. Each pellet was resuspended in 1 ml 100 mM LiAc and transferred to a 1.5 ml E-tube and recentrifuged at 13000 rpm for 5-7 sec. Finally each pellet was resuspended in 100 mM LiAc to get a final volume of 250µl and the cells were subsequently distributed (25 µl) in 1.5 ml E-tubes (one per transformation). The E-tubes were centrifuged at 13000 rpm for 5-10 sec, the supernatant removed and 178 µl of TRANSFO mix and 0.5-1µg of plasmid DNA were added to each cell pellet. [TRANSFO mix (20 transformations) : 2.4 ml PEG 3350 50% (w/v), 680 µl H₂O, 360 µl 1.0 M LiAc, 100 µl denatured fish-sperm DNA (10mg/ml)]. The tubes were vortexed 1 min in a multivortex and incubated at 30°C for 30 min. The cells were then exposed to a heat shock at 42°C for 30 min. After a final centrifugation at 6500 rpm for 15 sec the supernatant was removed, the pellet resuspended in 250 µl H₂O and the yeast cells plated on selection medium (Yeast Nitrogen Base ; QBIAGEN) lacking appropriate amino acids

to allow the selection of transformed cells. The colonies were allowed to grow at 30°C for 4-5 days.

Yeast two-hybrid screening. The two-hybrid screening was carried out as described by Van den Berghe *et al.* (Van den Berghe, *et al.*, 2000). Briefly, the 18 kDa FGF-2 was used as a bait, and the corresponding cDNA was inserted in the pAS2 vector. A human placenta library (Clontech, Palo Alto, CA) cloned into the pACT2 vector was transformed in the *Saccharomyces cerevisiae* strain Y190 and screened to identify proteins that were able to interact with FGF-2.

Measurement of β -galactosidase activity in yeast cell extract. In order to evaluate the capacity of the chimeras to interact with Translokine, the β -galactosidase activity in yeast extracts was measured from at least 10 pooled colonies. For this the Galacto-Light kit (TROPIX Inc. Bedford, MA) was used. Briefly, the yeast cells, grown over-night in 4 ml selection medium (YNB-Trp/-Leu), were centrifuged (3 min at 13000 rpm, 20°C). The pellet was resuspended in 200 μ l lysis buffer and approximately 30 μ l glass beads (425-600 μ m; Sigma ref. G8772) were added. The cells were lysed mechanically by 2x5 min vigorous vortexing at 4°C, interrupted by a 5 min pause. To recover the cell lysate, the tubes were centrifuged for 5 min at 13000 rpm, 4°C and the supernatants transferred to fresh tubes. Twenty μ l of cell extract were transferred to the well of a microtiter plate to which 80 μ l of Galacton® substrate was added, previously diluted 100x in substrate dilution buffer provided by the manufacturer. In order to standardize the reaction conditions, both the substrate solution and, 60 min later, the light emission accelerator solution were added by automatic injection in a microplate luminometer (EG&G Berthold, Bad Wildbad, Germany). Immediately after injection of the accelerator solution the chemiluminescence was measured during 20 sec per sample. The obtained values were normalized for protein contents, as determined by BCA assay according to the manufacturer's instructions (Pierce Chemical CO., Rockford, IL)

Construction of the plasmid pRF-Nb1a2 for Nb1a2 protein expression. The prokaryotic expression vector pRF-Nb1a2 was generated by a tri-molecular cloning approach. The plasmid pRFC10 (Fig. 11) was digested by the restriction enzymes *Bam*HI and *Eco*RI to obtain a 3.21 kb DNA fragment. In parallel, the digestion of the same plasmid with the restriction enzymes *Eco*RI and *Kpn*I generated a 0.74 kb DNA fragment.

pAS Nb1a2 was cleaved by the restriction enzymes *Bam*HI and *Kpn*I to obtain a 0.1 kb DNA fragment. These three fragments were ligated by using the T4 DNA ligase.

Escherichia coli transformation. *Escherichia coli* BL21 pLys Gold (Invitrogen) were transformed by the plasmid pRF-Nb1a2. For this, 1 µg of pRF-Nb1a2 was incubated on ice with 100 µl of bacteria during 10 minutes. The mix was then subjected to a heat-shock for 2 minutes at 37°C and followed by an incubation on ice for 2 minutes. 500 µl of LB was added to the mix followed by an incubation at 37°C for 45 minutes with shaking at 225-250 rpm. The reaction was then spread on a LB-ampicillin agar plate and the colonies were allowed to grow overnight at 37°C.

Production of Nb1a2 protein. The transformed bacteria were cultured overnight in 250 ml of LB medium containing 2% Glucose and 100 µg/ml of Ampicillin. The overnight culture was mixed with 1 volume of prewarmed (250 ml) said medium. After 20 minutes of shaking at 37°C, the culture was mixed with 500 ml of ice cold said medium containing 4% ethanol. The culture was allowed to grow at 20°C until it obtained an OD^{600nm} of 0.6. Then IPTG was then added to the medium to a final concentration of 0.5 mM and the incubation was continued for 4 hours. The cells were then harvested by centrifugation 5000 rpm at 4°C for 10 min.

Purification of recombinant Nb1a2. The bacterial pellet was resuspended in 30 ml of 20 mM Tris-HCl pH 7.4. and the cells were lysed by three cycles of snap-freezing in liquid nitrogen and sonication 5x 20 seconds. The cell lysate was centrifuged 13000 rpm for 30 min at 4°C. The supernatant fraction containing the recombinant Nb1a2 was adjusted to 0.7 M NaCl and applied directly at 0.5 ml/min to an heparin-sepharose FPLC column (Amersham Pharmacia Biotech, 1 ml of resin), previously equilibrated with 20 ml of 20 mM Tris-HCl pH 7.4, 0.7 M NaCl. The column was then washed with 20 ml of equilibration buffer at 1 ml/min and then with 20 ml of buffer containing 1M NaCl. The recombinant Nb1a2 was eluted with 20 mM Tris-HCl pH 7.4, 2M NaCl and fractions of 1 ml collected. The same protocol was applied to produce other FGF proteins, tagged or not, such as bovine FGF-2, HA-FGF-1, NLS-FGF-2 and NLS-Nb1a2. HA-tagged FGF-2 was kindly provided by G. Bouche (IPBS, Toulouse, France).

Other protein production techniques. His-tagged Translokine was produced using the IPTG-inducible pET expression system (pET-15b, Novagen, Madison, WI) in *E. coli*

BL21 (DE3) pLysS and purified on a nickel-agarose column (Ni-NTA, QIAGEN, Chatsworth, CA). For equilibration and elution, 20 mM and 200 mM imidazole were used, respectively. Protein concentrations were determined by BCA assay (Pierce Chemical CO., Rockford, IL). ³⁵S-labelled FGF-2 was obtained using an *in vitro* translation system. A plasmid encoding FGF-2 was linearized, transcribed *in vitro*, and translated in a rabbit reticulocyte lysate system (RRL; Promega corp., Madison, WI) in the presence of [³⁵S]methionine (Amersham Pharmacia Biotech) as previously described (Huez, *et al.*, 2001). After translation, the lysate containing [³⁵S]-FGF-2 was dialyzed against dialysis buffer (20 mM Hepes pH 7.0, 140 mM NaCl, 2 mM CaCl₂) to remove free [³⁵S]methionine and reducing agents.

Antibodies. Polyclonal anti-Translokin antibodies were generated through the immunization of two rabbits injected three times every 2 weeks and finally one month later with 200 µg of recombinant His-tagged Translokin, electroeluted from a polyacrylamide gel and diluted twice in Freund adjuvant (Sigma-Aldrich). Antiserum was then immunopurified with His-tagged Translokin linked to Affi gel-10 (Bio-Rad Laboratories, Hercules, CA) as previously described (Van den Berghe, *et al.*, 2000). Polyclonal and monoclonal anti-HA (HA.11) were from Babco (Eurogentec, Herstal, Belgium) monoclonal anti-β-actin (AC-15), anti-β-tubulin (TUB 2.1) and peroxidase-labeled anti-His from Sigma, monoclonal anti-FGF2 (Ab3) was from Oncogene Science (Boston, MA), rabbit polyclonal against Erk2 (C14) and mouse monoclonal against the phospho p44/p42 MAPK (E10) antibodies were purchased from Cell Signaling (Beverly, MA). The horse-radish peroxidase-conjugated antibodies were from Amersham, and the FITC or Cy3-conjugated antibodies were from Sigma-Aldrich. The monoclonal anti-GAL4-DBD antibodies were from Clontech. Chromomycin A3 was from Sigma-Aldrich.

***In vitro* interaction.** A 96-well plate was coated overnight at 4°C with 10 µg/ml FGF-2, Nb1a2 or BSA. The plate was washed with PBS-tween 0.2% and 100 µl/well of blocking solution was added for 1h at 37°C. The blocking solution was discarded and the plate washed. Different concentrations of His-tagged Translokin were added for 2 h at 37°C. After three washings, horse-radish-peroxidase-conjugated anti-His antibody (1/1000) was added for 1h at 37°C. Three washings were then performed and the complexes were revealed with 100µl/well of o-phenylenediamine (4 mg/ml) containing

0.03% (v/v) hydrogen peroxide in 0.1 M citrate buffer, pH 5.0. After 20 minutes, the reactions were stopped with 50 μ l of 2M H₂SO₄ and the absorbance was measured at 490 nm. When the experiment was performed in the reversed manner, the plate was coated with Transloklin, and HA-tagged proteins were added. An incubation with a monoclonal anti-HA antibody (1/1000) followed by an incubation with a horse-radish-peroxidase-conjugated anti-mouse IgG antibody (1/1000) were used to reveal the interactions.

Immunoprecipitation. NIH-3T3 cells were transiently transfected with a plasmid encoding the full-length HA-tagged Transloklin. After 48h, the cells were incubated for 2h with 60ng/ml His-tagged FGF-2. The cells were subsequently rinsed in cold PBS, scraped, pelleted, and lysed on ice in 50 mM Tris pH 7.4, 150 mM NaCl, 1% NP-40 and protease inhibitors for 30 min. Cells debris were removed by centrifugation at 10,000 x g for 10 min at 4°C. The extracts were incubated with an monoclonal anti-HA antibody coupled to protein G-Sepharose beads (Sigma) for 2h at 4°C. The beads were washed three times in the lysis buffer and the immobilized proteins were released by boiling in Laemmli buffer and then analyzed by SDS-PAGE.

Northern blot analysis. A human multiple-tissue Northern blot (Clontech) was probed with a 416 nucleotide fragment excised from the human Transloklin cDNA by a *Xba*I digestion and subsequently labelled with [³²P]-ATP using a nick-translation kit (Promega Corp.).

Cell culture. Cells were grown in Dulbecco Modified Eagle Medium (DMEM, Invitrogen) containing antibiotics, 1% glutamin and serum. For COS-7 cells, the medium contained 1 g/l glucose and 5% fetal calf serum (FCS). NIH-3T3 and ABAE cells were grown in DMEM (4 g/l glucose) with 10% FCS or 10% calf serum, respectively. All cell transfections with plasmid DNA were made using Fugene 6 reagent according to the manufacturer's recommendations (Roche Diagnostics, Mannheim, Germany).

Transfection of mammalian cells with siRNA duplexes. The 21-nucleotide siRNA duplexes were synthesized and purified by Dharmacon Research (Boulder, CO). The siRNA sequence, targeting mouse Transloklin, corresponded to the coding region 463-483 relative to the first nucleotide of the start codon. The siRNA control was the

scramble one from Dharmacon. The transfection was performed with 0.1 μ M of siRNA and oligofectamine reagent (Invitrogen) according to the manufacturer's instructions.

***In situ* Immunocytochemistry.** COS-7 cells, grown on coverslips, were transfected and prepared for immunofluorescence microscopy as previously described (Arnaud, *et al.*, 1999). ABAE cells, grown on coverslips, were serum starved for 48h and stimulated with 20 ng/ml HA-FGF or HA-Nb1a2 for 2 hours. In some experiments cells were treated before stimulation with 10 μ M nocodazole or 20 μ g/ml cytochalasin B (Sigma) for 1h. Immunocytochemistry was performed as previously described (Baldin, *et al.*, 1990) using polyclonal anti-HA and monoclonal anti- β -tubulin as primary antibodies and anti-rabbit IgG Cy3 conjugate (1/250) and anti-mouse IgG FITC conjugate (1/50) as secondary antibodies (Sigma). Nuclei were labeled with chromomycin A3 (0.1 mg/ml). Antibody complexes were visualized under a Leica fluorescence microscope or a confocal microscope: Zeiss LSM510 oil objective 63, operture 1.4.

Proliferation of endothelial cells. Five thousand ABAE (adult bovine aortic endothelial) cells were seeded in 35 mm tissue culture dishes in 1 ml of DMEM (Gibco BRL) + 10% Newborn Calf serum (Gibco BRL) + 1% Glutamin (Gibco BRL)+ 1% Amphotericin (Biochrom)+ 0.5 % gentamycin (Gibco BRL) at 37°C 10% CO₂. FGF-2 or Nb1a2, diluted in DMEM + 0.5% BSA, was added to the indicated final concentration on days 1 and 3. The cells were counted on day 5. For this the cells were washed with PBS and trypsinized in 1 ml, and counted with a Coulter Counter (Coulter Electronics Ltd).

Organization of ABAE cells on a Basement membrane *Substratum in vitro.* Twenty-four-well plates were coated with 300 μ l MatriGel™ (B&D, Bedford, MA) per well, which was allowed to polymerize for 1h at 37°C. ABAE cells (2×10^5 cells/ml) suspended in 500 μ l of culture medium were then added to each well. FGF-2 or Nb1a2 were added to the wells at 0.5 ng/ml, and the plates were incubated overnight at 37°C. After removal of the medium, the culture was fixed and the length of the tube network was quantified with the Q Win Leica system.

MAPK activation. ABAE cells growing on 60 mm dishes (at 19,000 cells/cm²) were kept for 1 day in serum-free medium. Subsequently, cells were left untreated or

stimulated with 20 ng/ml FGF-2 or Nb1a2 for the times indicated. The cells were washed twice in PBS and lysed for 15 min on ice in a HEPES-buffer (50 mM, pH 7.4) containing 150 mM NaCl, 100 mM NaF, 10 mM EDTA, 10 mM Na₄P₂O₇, 2 mM sodium orthovanadate, 1 mM PMSF, 2 mg/ml aprotinin, 20 mM leupeptin and 1% Triton. The mixture was gently agitated for 15 min at 4°C and centrifuged at 13,000g for 15 min. Soluble proteins (60 µg) were separated on a 12.5 % SDS-polyacrylamide gel and subjected to immunoblotting with the indicated antibodies.

ABAE fractionation. The fractionation was performed as previously described (Baldin, *et al.*, 1990). Purity and integrity of preparations of nuclei (prior to sonication) were monitored by phase contrast microscopy, and by the quantitative analysis of the cytoplasmic Lactate Dehydrogenase (LDH) content.

Animal studies. All procedures involving experimental animals were performed in accordance with the recommendations of the French Accreditation of Laboratory Animal Care. The mice were housed in stainless-steel cages in groups of 5, kept in a temperature-controlled facility on a 12-h light-dark cycle, and fed normal laboratory mouse chow diet. For all surgical procedures, mice were anesthetized by intraperitoneal injection of 150 mg/kg ketamine and allowed to recover on a thermostated pack. C57BL/6 mice were divided into three groups (n=10 each) for treatment with either FGF-2, Nb1a2 or placebo (saline). FGF-2 and Nb1a2 were administrated by bolus of 5µg IV in the jugular vein, just after electric carotid injury.

Electric injury model. The inventors adapted the electric injury model described by Carmeliet *et al.* (Carmeliet, *et al.*, 1997) on the femoral artery to the common carotid artery, the latter being easier to dissect. Electric carotid artery injury was performed in 6-week-old mice. Surgery was carried out using a dissection microscope (Nikon SMZ-2B) in six-week-old female mice weighing 20 g on average. Since the proximal part of the carotid artery is intrathoracic, the injury could not be applied to the whole common carotid artery. The left common carotid artery was exposed via an anterior incision of the neck. The electric injury (mainly thermic) was applied to the distal part of the common carotid artery. The carotid artery was injured by electric current using a bipolar microregulator. In order to standardize the temperature increase in the vessel wall, we used forceps with large tips (1mm), instead of microsurgical forceps (200 µm) and a

bipolar microregulator Force FX (Valleylab®). The "precise" mode of this apparatus allowed delivery of electric energy within a narrow range of resistance, as the generator microprocessor disrupted the electric current when the resistance increased as a consequence of temperature increase. This enabled the increase in tissue temperature to be controlled, and avoided the risk of desiccation and coagulation of the arterial wall. The optimal conditions were determined as follows: electric current of 2 Watts applied for 2 seconds to each mm of carotid artery over a total length of 4mm with the help of a size marker placed parallel to the long axis of the carotid. Despite optimization of the technique, coagulation and thrombosis of the carotid artery occurred in approximately 10 % of the cases, which were then excluded from the study.

Evaluation of Reendothelialization. Three days following the injury, the endothelial regeneration process was evaluated by staining the denuded areas with Evans blue dye. For this, 50µl of solution containing 5% Evans blue diluted in saline was injected in the tail vein using a 30-gauge needle 10 minutes before sacrifice, followed by fixation using a perfusion with 4% phosphate-buffered formalin (pH 7.0) for 5 minutes. Blood, saline and fixative was removed through an incision in the right atrium. The left common carotid artery was dissected with an adjacent portion of the aortic arch and carotid bifurcation. The artery was then opened longitudinally and placed between slides with Fluoprep®. After transparency scanning and numerization, the total and stained carotid artery area were planimeted using an "image analyzer" (VISIOL@b™2000). The ratio between the area stained in blue and the total carotid artery area was calculated. The surface of area remaining deendothelialized was indexed to the total carotid artery area to take into account the changes in vessel area due to both the elasticity of the carotid artery and to the flattening of the vessel between slides. Results are expressed as means ± S.E.M. ANOVA was used for the comparison of the treatments with FGF-2 and Nb1a2 compared to controls.

Morphological studies Ten days following injury, carotids were removed and fixed in 3.7% paraformaldehyde. The samples were dehydrated with ethanol and Microclearing™ (Microstain, Granrolo, Italy), and then embedded in paraffin. Sections (6 µm) were rehydrated and stained with hematoxylin to visualize the nuclei.

Silver nitrate staining was used to examine the morphology of the regenerated endothelium, 10 days after carotid electric injury (n=3). Left ventricle was perfused

immediately after death with solutions of 5% dextrose, 0.25% AgNO₃, 1% NH₄Br, and 3% CoBr each for 1 minute, followed by perfusion of 100 ml buffered formalin. The carotid artery was excised, immersed in formalin for 1 hour, and rinsed in 0.1 M sodium cacodylate buffer. Control and injured carotids were incised longitudinally, dehydrated and examined by light microscopy.

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Figure 1: Characterization of Translokin, an FGF-2 interacting protein

Alignment of the predicted amino acid sequences of human, mouse and bovine Translokin. The cDNAs were cloned from a human placenta two-hybrid library, mice embryos and ABAE (bovine) cells, respectively. The arrows indicate the 5' end of each clone (A,B,C,D) isolated by the two-hybrid system.

Figure 2: Specific and direct interaction between Translokin and FGF-2

a, The specificity of the interaction between Translokin and FGF-2 was first tested in the yeast two-hybrid system. Several FGFs, including the 18 kDa and 24 kDa forms of FGF-2, and Translokin were fused to DNA binding (DB) or activating (AD) domains of Gal4 and subjected to a yeast two hybrid assay to evaluate a potential interaction.

b, Direct interaction between FGF-2 and Translokin by ELISA : FGF-2 and BSA were coated on a 96-well plate and incubated with different concentration of His-tagged recombinant Translokin. The association was revealed using a horse-radish-peroxidase-conjugated antibody against the (His)₆ tag, then read at 490 nm with an ELISA microplate reader. Data are the mean \pm S.D. of triplicate samples.

c, The specificity of the interaction between Translokin and FGF-2 was tested in an ELISA assay. Translokin was coated on a 96-well plate and incubated with HA-tagged 18 kDa FGF-2, FGF-1, or 24 kDa FGF-2. The association was revealed using a monoclonal anti-HA antibody and a horse-radish peroxidase-conjugated anti-mouse IgG antibody. The results were read at 490 nm with an ELISA microplate reader. Data are the mean \pm S.D. of triplicate samples.

d, Co-immunoprecipitation of HA-tagged Translokin and FGF-2. Mouse NIH-3T3 cells were transiently transfected or not (-) with a plasmid encoding the full-length HA-tagged Translokin and incubated during 2h with 60 ng/ml His-tagged FGF-2 (18kDa). Extracts from these cells were immunoprecipitated with monoclonal anti-HA coupled to protein G-Sepharose beads. Immunoprecipitated proteins treated under non-reducing conditions were subjected to Western blotting and revealed with monoclonal anti-HA and anti-His antibodies.

e, Co-immunoprecipitation of Translokin and FGF-2. Extracts from NIH-3T3 cells stimulated (+) or not (-) with 60 ng/ml HA-tagged FGF-2 (18kDa) were subjected to immunoprecipitation with anti-Translokin coupled to protein A-Sepharose beads or with

a control antibody. Immunoprecipitated proteins were subjected to Western blotting and revealed with monoclonal anti-HA to detect HA-tagged FGF-2.

Figure 3: Translokin expression is ubiquitous

5 a, Northern blot (MTN™, Clontech) analysis showing an ubiquitous expression of Translokin in different human tissues.

b, Expression of Translokin mRNA was detected by RT-PCR on mRNA from different cell lines. Ctrl+ is a positive control that corresponded to a PCR done with plasmid containing the Translokin cDNA as the template. Ctrl- is a negative control and
10 corresponds to a PCR done without added template.

c, Immunolocalization of Translokin in transfected COS-7 cells. Transfection was performed with a plasmid encoding the full-length Translokin. Cells were stained using anti-Translokin (top) or polyclonal anti-HA (bottom) followed by the anti-rabbit-Cy3 conjugated antibody and the monoclonal anti- β -tubulin followed by the anti-mouse-FITC
15 conjugated antibody.

Figure 4: Interaction of FGF-2 / FGF-1 chimeras with Translokin

a, Two-hybrid system. Relative intensity of the interactions between FGF-1/2 chimeras and Translokin measured by β -galactosidase activity in yeast extracts. Four
20 junctions were chosen in the well-conserved β -strands 2,5,8 and 10. The regions derived from FGF-1 are represented in white and those from FGF-2 are in black. The nomenclature is based on the origin of the N-terminal extremity; "a" for acidic FGF (FGF-1) and "b" for basic FGF (FGF-2). The numbers indicate the junctions between FGF-1 and FGF-2. The data shown are the mean \pm S.E.M from at least three independent
25 transformations. β -galactosidase assays were performed in duplicates.

b, Analysis of the expression of chimeras/GAL4-DBD fusion proteins. Yeast extracts were subjected to Western blotting and revealed with a monoclonal anti-GAL4-DBD antibody.

c, 10 μ g/ml of FGF-2, Nb1a2 or BSA were immobilized on a 96-well plate and the
30 assay was performed as in figure 2b with 10 μ g/ml of His-tagged Translokin. Data are the mean \pm S.D. of triplicate samples.

Figure 5: Characterization of Nb1a2

a, Proliferation assay. ABAE cells were stimulated by different concentrations of FGF-2 or Nb1a2 added at day 1 and 3. At day 5 cells were trypsinized and counted with a Coulter counter^R ZM. Data are the mean \pm S.D of triplicate samples. The experiment was repeated at least three times with similar results.

b, Organization of ABAE cells on a Basement Membrane Substratum *in vitro*. ABAE cells were plated on MATRIGELTM coated wells in presence of 0.5 ng/ml FGF-2 or Nb1a2 during 12 hours. Tube formation was observed and quantified with a Leica Q Win system.

c, Relative receptors binding affinity : confluent ABAE cells were incubated for 2h at 4°C with [³⁵S]-FGF-2 and increasing concentrations of unlabeled FGF-2 or Nb1a2. Cells were then treated as previously described (Estival, *et al.*, 1996). Values represent the percentage of specific binding of [³⁵S]-FGF-2 on FGF receptors determined at different concentrations of competitors (D: 10 μ g of denatured competitors).

d, Recombinant FGF-2 and Nb1a2 were purified by heparin-sepharose chromatography. Elution was performed using a NaCl gradient. Fractions were analyzed by SDS-PAGE and immunoblotting with an anti-FGF-2 antibody.

e, Effect of FGFs on MAP kinase activation : G0-arrested ABAE cells were stimulated (or not) with 20 ng/ml FGF-2 or Nb1a2 during different periods. Cells were then harvest and MAPK activation was detected by Western blotting using a monoclonal antibody raised against phosphorylated forms of p44 and p42. Immunoblotting with an antibody raised against the non-phosphorylated form of p44 and p42 was performed to verify that equal amounts of cell extracts had been used.

Figure 6: FGF-2 intracellular localization

a, G0-arrested ABAE cells were incubated with or without 20ng/ml HA-FGF-2 or HA-Nb1a2 (20 ng/ml or 100 ng/ml) for 2h. The cells were lysed and nuclear and postnuclear fractions were incubated in the presence of heparin sepharose beads. Proteins bound to heparin sepharose beads were analysed by SDS-PAGE and immunoblotting using monoclonal α -HA antibody.

b, **Confocal microscopy analysis.** ABAE cells were stimulated (B-F) or not (A), by HA-FGF-2 (B-E) or HA-Nb1a2 (F) during 2h at 37°C. In some cases, cells were pretreated, with nocodazole (10 μ M) (C) or cytochalasin B (20 μ g/ml) (E) 1h before HA-

FGF-2 stimulation. After HA-FGF-2 incubation, cells were fixed and stained with polyclonal anti-HA (HA-FGF-2, HA-Nb1a2), monoclonal anti- β -tubulin (A-C, F) or anti- β -actin (D, E) antibodies. Nuclei were labeled with Chromomycin A3.

5 **Figure 7: Recovery of the mitogenic activity of Nb1a2 and inhibition of FGF-2**

10 **a**, Proliferation assay. ABAE cells were stimulated by different concentrations of FGF-2, NLS-FGF-2, Nb1a2 or NLS-Nb1a2 added at day 1 and 3. At day 5 cells were trypsinized and counted with a Coulter counter^R ZM. The experiment was repeated at least three times with similar results, using recombinant proteins from two independent productions.

15 **b**, HA-NLS-FGF-2 and HA-NLS-Nb1a2 are translocated to the nucleus. After stimulation with (+) or without (-) 60 ng/ml NLS-tagged growth factors, NIH-3T3 cells were treated and fractionated as described for figure 6a. Nuclear and postnuclear fractions were analyzed by SDS-PAGE and immunoblotting using monoclonal anti-HA antibody.

20 **c**, Western-blot for Translokin expression in scramble siRNA (lane 1) and Translokin siRNA (lane 2) transfected NIH-3T3 using an anti-Translokin antibody (top panel). β -actin expression was used as a loading control (bottom panel)

25 **d**, Translokin siRNA inhibits FGF-2 translocation. Scramble siRNA (lane 1) and Translokin siRNA (lane 2) transfected NIH-3T3 were stimulated with 20ng/ml HA-FGF-2 for 2h. The cells were then treated and fractionated as described for figure 6a. Nuclear and postnuclear fractions were analyzed by SDS-PAGE and immunoblotting using monoclonal anti-HA antibody. Protein levels were quantified by densitometry of immunoblots, and values normalized to control siRNA transfected cells. Values represent the mean \pm SD of triplicate experiments.

30 **e**, Translokin siRNA does not affect FGF-1 translocation. Scramble siRNA (lane 2) and Translokin siRNA (lane 3) transfected NIH-3T3 were stimulated with (lanes 2 and 3) or without (lane 1) 60ng/ml HA-FGF-1 and 10 u/ml of heparin during 5h. The cells were then treated as described in figure 7d.

35 **Figure 8: FGF-2 and Nb1a2 accelerate the process of reendothelialisation.**

Four millimeters of the distal part of the common carotid artery of C57/Bl6 mice was injured with a bipolar electrosurgical generator (Valleylab Force FX) as described in (Brouchet, *et al.*, 2001). The bars (mean \pm SEM) indicate the reendothelialized surface at

day +3 as percentage of the initially injured surface after one single intravenous administration of 5 μ g of FGF-2 or Nb1a2. (controls (n=7); FGF-2 and Nb1a2 (n=8)).

Figure 9: Morphology of reendothelialized and non-injured carotids.

5 Silver nitrate staining of carotids 10 days after electric injury from (a) FGF-2 treated (5 μ g administrated intravenously (IV) in bolus), (b) Nb1a2 treated (5 μ g IV in bolus) and (c) non-injured animals. After perfusion with silver nitrate solution and buffered formalin, the carotid artery was excised, immersed in formalin, and rinsed in cacodylate buffer. The carotids were incised longitudinally, dehydrated and examined by
10 light microscopy.

Figure 10: Detailed analysis of the first Translokin-interacting region in FGF-2

The capacity of nine targeted mutants in the most N-terminal Translokin-interacting region in FGF-2 to interact with Translokin. The interaction studies were performed in
15 the yeast two-hybrid system as described in the materials and methods section. The results (mean \pm SEM) are represented as the percentage of the beta-galactosidase activity obtained with FGF-2 and Translokin.

Figure 11: Map of the plasmid pRFC10

20 Map of the plasmid pRFC10 used for the construction of pRF-Nb1a2 for the production of recombinant Nb1a2. The cassette BamHI/KpnI in pRFC10 was exchanged by the same fragment in pAS2-Nb1a2.

CLAIMS

5 1. Use of animal or human basic fibroblast growth factor (FGF-2) derived proteins for the preparation of biomaterials or medical devices chosen among medical prostheses, such as endovascular stents, coated endoprotheses or by-pass grafts, said FGF-2 derived proteins being chosen among FGF-2 mutants which are unable to interact specifically with the Translokine represented by SEQ ID NO: 2, i.e. having a
10 binding capacity to said Translokine of less than 40%, and more preferably of less than 10%, said FGF-2 mutants being such that they are defective in intracellular trafficking and in stimulating the proliferation of endothelial cells, and that their tubulogenesis activity is preserved.

15 2. Use according to claim 1, of FGF-2 mutants corresponding to bovine FGF-2 represented by SEQ ID NO: 4, or to human FGF-2 represented by SEQ ID NO: 6, and wherein the region delimited by the amino acids located from positions 40 to 130 contains at least one mutation by deletion, or substitution by a heterologous amino acid natural or not, of at least one amino acid of said region, or by insertion of at least one
20 heterologous amino acid natural or not in said region.

 3. Use according to claim 1 or 2, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, wherein the region delimited by the amino acids located from positions 40 to 75, and/or the region delimited by the
25 amino acids located from positions 105 to 130, contain at least one mutation as defined in claim 2.

 4. Use according to any of claims 1 to 3, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, wherein the region
30 delimited by the amino acids located from positions 44 to 61 contains at least one mutation as defined in claim 2.

5. Use according to claims 1 to 4, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, wherein at least one of the amino acids located in positions 44, 48, 52, 54, 55, 58, 61, 65, 68, 69, 71, and 73, is mutated.

5

6. Use according to any of claims 1 to 5, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, and comprising at least one of the following mutations : H44L, R48T, V52T, E54D, K55R, P58Q, K61Q, Q65S, E68S, R69V, V71E, and S73Y, said mutants being represented by SEQ ID NO: 8, or by SEQ ID NO: 20, respectively.

10

7. Use according to any of claims 1 to 6, of :

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations R48T, V52T, and E54D, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 26, or by SEQ ID NO: 28, respectively,

15

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations H44L, R48T, V52T, and K55R, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 30, or by SEQ ID NO: 32, respectively,

20

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations H44L, R48T, V52T, and E54D, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 34, or by SEQ ID NO: 36, respectively,

25

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations Q65S, E68S, R69V, V71E, and S73Y, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 38, or by SEQ ID NO: 40, respectively.

30

8. Use according to any of claims 1 to 6, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, wherein at least the amino

acids in positions 54 and 55 are mutated, such as FGF-2 mutants comprising the mutations E54D and K55R.

5 9. Use according to claim 8, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, wherein at least the amino acids in positions 54 and 55 are mutated, and wherein at least one of the amino acids located in positions 44, 48, 52, 58, and 61, is mutated, such as FGF-2 mutants comprising the mutations E54D and K55R, and at least one of the following mutations H44L, R48T, V52T, P58Q, and K61Q, and more particularly :

10 · the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations E54D, K55R, and H44L, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 42, or by SEQ ID NO: 44, respectively,

15 · the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations E54D, K55R, H44L, and R48T, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 46, or by SEQ ID NO: 48, respectively,

20 · the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations E54D, K55R, H44L, R48T, and V52T, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 50, or by SEQ ID NO: 52, respectively,

25 · the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations E54D, K55R, H44L, R48T, V52T, P58Q, and K61Q, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 54, or by SEQ ID NO: 56, respectively.

 10. Use according to claims 8 or 9, of FGF-2 mutants corresponding to chimeric FGF-1/FGF-2 proteins.

30 11. Use according to claim 10, of the following FGF-2 mutants :

- Nb1a2, represented by SEQ ID NO: 10, corresponding to a chimeric FGF-1/FGF-2 protein wherein the amino acids in positions 1 to 43 and 79 to 155 belong to

bovine FGF-2 represented by SEQ ID NO: 4, and the amino acids located in positions 44 to 78 belong to human FGF-1 represented by SEQ ID NO: 14,

- Nb1a1.6, also referred to as HRVEKPK 44/48/52/54/55/58/61 LTIDRQQ, represented by SEQ ID NO: 12, corresponding to a chimeric FGF-1/FGF-2 protein wherein the amino acids in positions 1 to 43 belong to bovine FGF-2 represented by SEQ ID NO: 4, the amino acids located in positions 44 to 61 belong to human FGF-1 represented by SEQ ID NO: 14, and the amino acids located in positions 62 to 155 belong to human FGF-2 represented by SEQ ID NO: 6.

12. Use according to any of claims 1 to 3, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, wherein the region delimited by the amino acids located from positions 109 to 126 contains at least one mutation as defined in claim 2.

13. Use according to claim 12, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, wherein at least one of the amino acids located in positions 109, 111, 116, 118, 120, 121, 122, 124, and 126, is mutated.

14. Use according to claim 12 or 13, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, and comprising at least one of the following mutations: S109E, N111H, R116T, R118K, Y120H/A/E, S/T121A/E/K, S122E/K/N, Y124F, and A126G, said mutants being represented by SEQ ID NO: 16, or by SEQ ID NO: 22, respectively.

15. Use according to any of claims 12 to 14, of FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, and comprising the following mutations: S109E, N111H, Y120H/A/E, S/T121A/E/K, S122E/K/N, Y124F, and A126G, said mutants being represented by SEQ ID NO: 58, or by SEQ ID NO: 60, respectively.

16. Use according to any of claims 12 to 15, of the FGF-2 mutant represented by SEQ ID NO: 18, corresponding to a chimeric FGF-1/FGF-2 protein wherein the amino

acids in positions 1 to 108 and 127 to 155 belong to human FGF-2 represented by SEQ ID NO: 6, and the amino acids located in positions 109 to 126 belong to human FGF-1 represented by SEQ ID NO: 14.

5 17. FGF-2 derived proteins chosen among the following FGF-2 mutants :

- FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, and comprising at least one of the following mutations : H44L, R48T, V52T, E54D, K55R, P58Q, K61Q, Q65S, E68S, R69V, V71E, and S73Y, said mutants being represented by SEQ ID NO: 8, or by SEQ ID NO: 20, respectively, and
10 more particularly:

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations R48T, V52T, and E54D, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 26, or by SEQ ID NO: 28, respectively,

15 · the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations H44L, R48T, V52T, and K55R, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 30, or by SEQ ID NO: 32, respectively,

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations H44L, R48T, V52T, and E54D, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 34, or by SEQ ID NO: 36, respectively,

20 · the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations Q65S, E68S, R69V, V71E, and S73Y, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 38, or by SEQ ID NO: 40, respectively,

- FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, wherein at least the amino acids in positions 54 and 55 are mutated, such as FGF-2 mutants comprising the mutations E54D and K55R,

30 - FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, wherein at least the amino acids in positions 54 and 55 are mutated, and wherein at least one of the amino acids located in positions 44, 48, 52, 58, and 61,

is mutated, such as FGF-2 mutants comprising the mutations E54D and K55R, and at least one of the following mutations H44L, R48T, V52T, P58Q, and K61Q, and more particularly :

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations E54D, K55R, and H44L, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 42, or by SEQ ID NO: 44, respectively,

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations E54D, K55R, H44L, and R48T, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 46, or by SEQ ID NO: 48, respectively,

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations E54D, K55R, H44L, R48T, and V52T, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 50, or by SEQ ID NO: 52, respectively,

· the bovine or human FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4 or by SEQ ID NO: 6 respectively, and comprising the mutations E54D, K55R, H44L, R48T, V52T, P58Q, and K61Q, said bovine or human FGF-2 mutants being represented by SEQ ID NO: 54, or by SEQ ID NO: 56, respectively,

- Nb1a2, represented by SEQ ID NO: 10, corresponding to a chimeric FGF-1/FGF-2 protein wherein the amino acids in positions 1 to 43 and 79 to 155 belong to bovine FGF-2 represented by SEQ ID NO: 4, and the amino acids located in positions 44 to 78 belong to human FGF-1 represented by SEQ ID NO: 14,

- Nb1a1.6, represented by SEQ ID NO: 12, corresponding to a chimeric FGF-1/FGF-2 protein wherein the amino acids in positions 1 to 43 belong to bovine FGF-2 represented by SEQ ID NO: 4, the amino acids located in positions 44 to 61 belong to human FGF-1 represented by SEQ ID NO: 14, and the amino acids located in positions 62 to 155 belong to human FGF-2 represented by SEQ ID NO: 6,

- FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, and comprising at least one of the following mutations : S109E, N111H, R116T, R118K, Y120H/A/E, S/T121A/E/K, S122E/K/N, Y124F, and

A126G, said mutants being represented by SEQ ID NO: 16, or by SEQ ID NO: 22, respectively,

- FGF-2 mutants corresponding to FGF-2 represented by SEQ ID NO: 4, or by SEQ ID NO: 6, and comprising the following mutations : S109E, N111H, Y120H/A/E, S/T121A/E/K, S122E/K/N, Y124F, and A126G, said mutants being represented by SEQ ID NO: 58, or by SEQ ID NO: 60, respectively,

- the FGF-2 mutant represented by SEQ ID NO: 18, corresponding to a chimeric FGF-1/FGF-2 protein wherein the amino acids in positions 1 to 108 and 127 to 155 belong to human FGF-2 represented by SEQ ID NO: 6, and the amino acids located in positions 109 to 126 belong to human FGF-1 represented by SEQ ID NO: 14.

18. Nucleotide sequences encoding FGF-2 derived proteins as defined in claim 17.

19. Vectors or plasmids containing nucleotide sequences as defined in claim 18.

20. Host cells transformed with vectors as defined in claim 19.

21. Process for the preparation of FGF-2 derived proteins as defined in claim 17, comprising the culturing of host cells in an appropriate medium, and the purification of said FGF-2 derived proteins produced.

22. Biomaterials or medical devices coated with basic fibroblast growth factor (FGF-2) derived proteins chosen among FGF-2 mutants which are unable to interact specifically with the translokin represented by SEQ ID NO: 2, i.e. having a binding level to said translokin less than 40%, said FGF-2 mutants being such that they are defective in intracellular trafficking and in stimulation of proliferation of endothelial cells, and that their tubulogenesis activity is preserved, for the preparation of biomaterials or medical devices chosen among medical prostheses, such as endovascular stents, coated endoprotheses or bypass grafts.

23. Biomaterials, or medical devices according to claim 22, coated with basic fibroblast growth factor (FGF-2) derived proteins as defined in claim 17.

5 24. Biomaterials or medical devices according to claim 22 or 23, chosen among endovascular prostheses, such as stents used for inhibiting restenosis or stenosis following angioplasty.

			A →			B →
Human	1	MAAASVSAAS	CSHL	SN	SFAE	PSRSNGSMVRHSSSPYVVVPSDKPFLNSDLRRSPSKPTLA
Bovine	1	MAAASVSEET	SAS	OF	SN	CLAEPSKSNNGSMVRHSSSPYVVVPPDKPFLNSDLRRSPNKPTFA
Mouse	1	MAAASVSAAS	DS	QF	SSVLA	EPSRSNGMVRHSSSPYVVVPPDKPFLNSDLRRSPNKPTFA
				C →	D →	
Human	61	YPESNSRAIF	SALK	NLD	DKIR	RLELERIQABESVKTLSRETIYKVKVLEDEQIOERENSKN
Bovine	61	YPESNSRAIF	SALK	NLD	DKIR	RLELERIQABESVKTLSRETIYKVKVLEDEQIOERENSKN
Mouse	61	YPESNSRAIF	SALK	NLD	DKIR	RLELERIQABESVKTLSRETIYKVKVLEDEQIOERENSKN
Human	121	ESKHNQEL	SS	Q	LLA	ENKCNLEERKLEENRNMKHAEMERTSVLEKQVSLERERQHDQ
Bovine	121	ESKHNQEL	SS	Q	LLA	ENKCNLEERKLEENRNMKHAEMERTSVLEKQVSLERERQHDQ
Mouse	121	ESKHNQEL	SS	Q	LLA	ENKCNLEERKLEENRNMKHAEMERTSVLEKQVSLERERQHDQ
Human	181	HVQSOLER	LD	LL	EG	EVNKLITMORLA
Bovine	181	HVQMLEK	LD	LL	EG	EVNKLITMORLA
Mouse	181	HVQSOLER	LD	LL	EG	EVNKLITMORLA
Human	241	NRLIFED	KAT	EC	VFN	-ARRIKKKSKPPEKKS
Bovine	241	NRLIFED	KAT	EC	VFN	-TRIRIKKKSKPPEKKS
Mouse	241	NRLIFED	KAT	EC	VFN	-TRIRIKKKSKPPEKKS
Human	300	SHAVVAN	VQ	V	HL	MKQHSKALCNDRV
Bovine	300	SHAVVAN	VQ	V	HL	MKQHSKALCNDRV
Mouse	301	SHAVVAN	VQ	V	HL	MKQHSKALCNDRV
Human	360	LSEVLT	Q	L	O	D
Bovine	359	LSEVLT	Q	L	O	D
Mouse	359	LSEVLT	Q	L	O	D
Human	420	KYQALE	K	K	L	E
Bovine	419	KYQALE	K	K	L	E
Mouse	419	KYQALE	K	K	L	E
Human	479	QLLKD	M	Q	S	I
Bovine	478	QLLKD	M	Q	S	I
Mouse	479	QLLKD	M	Q	S	I

Figure 1

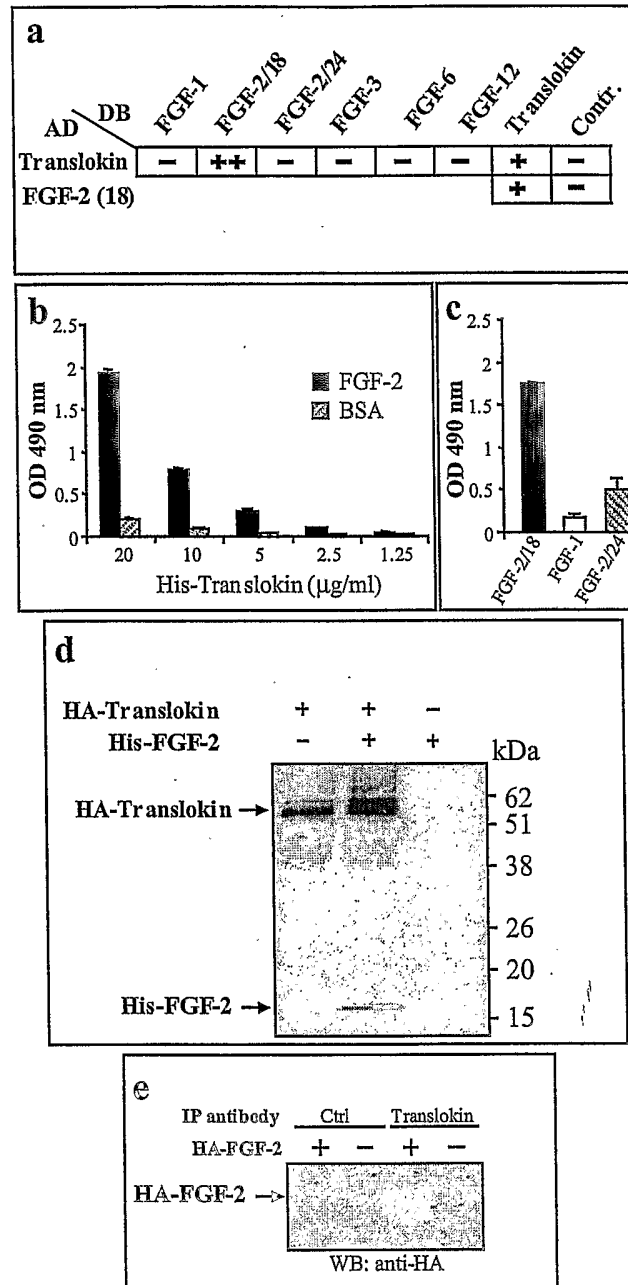


Figure 2

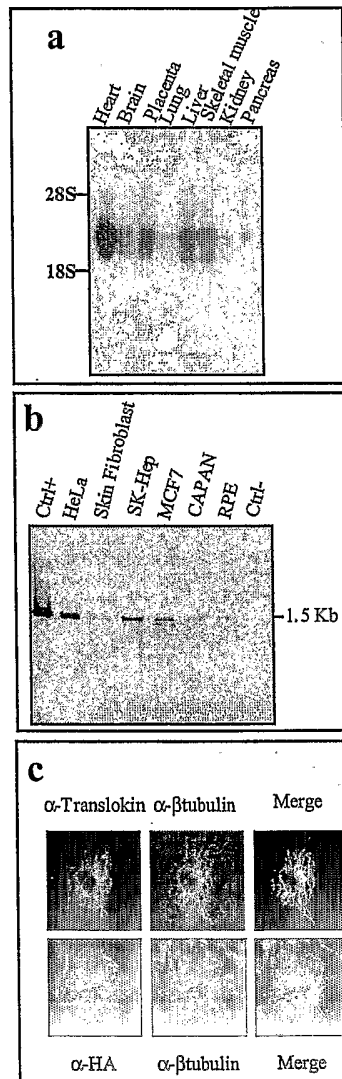


Figure 3

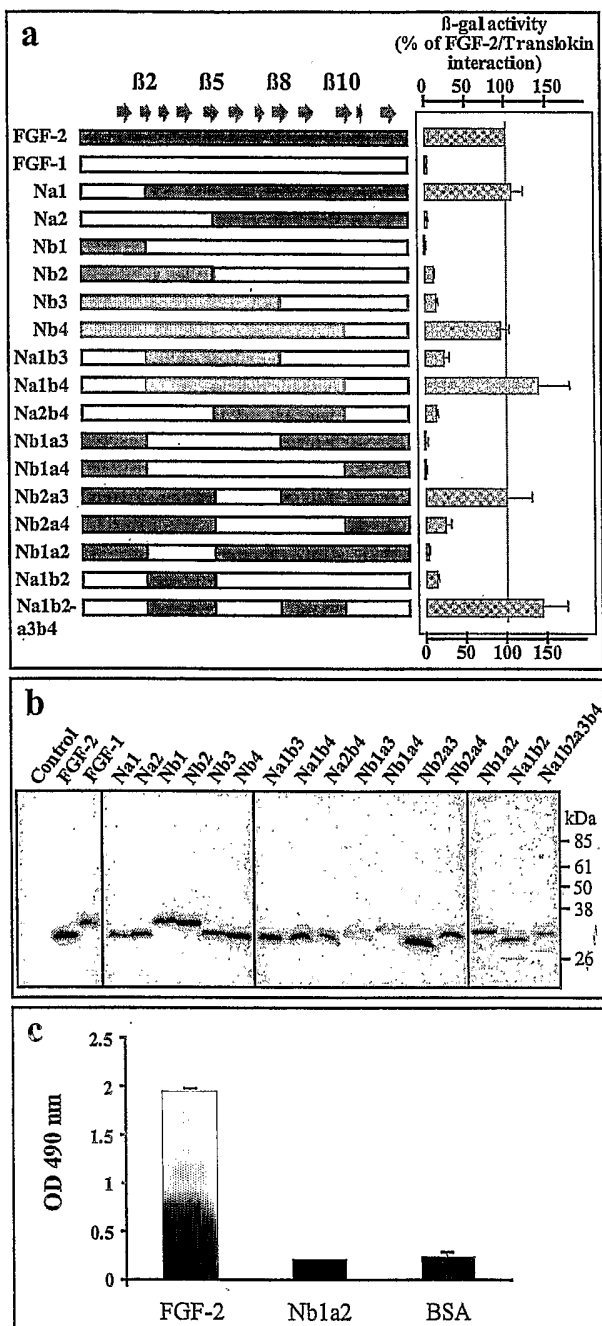


Figure 4

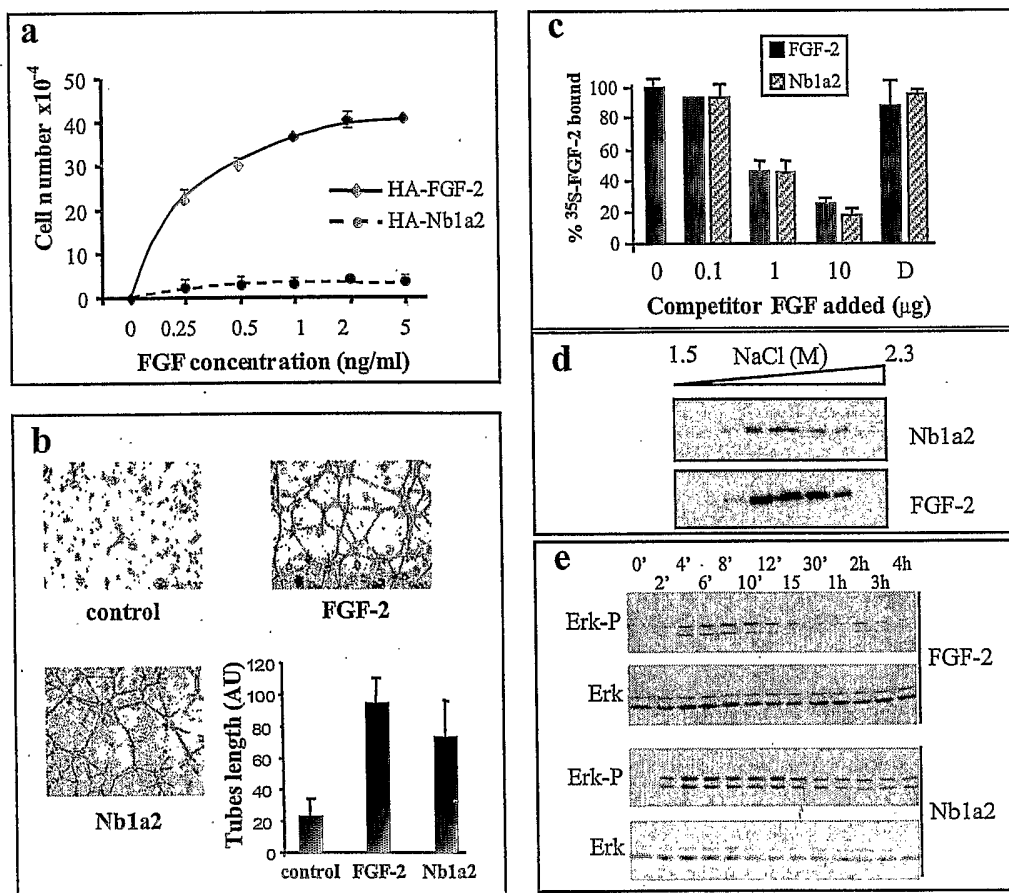
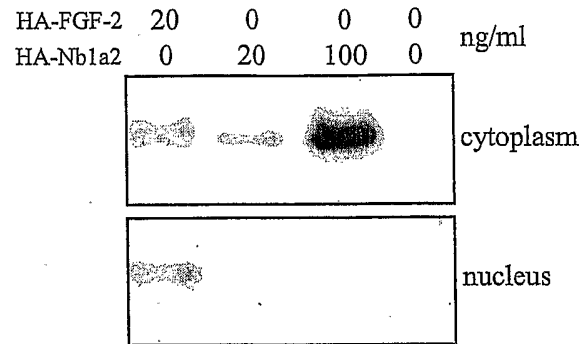


Figure 5

a



b

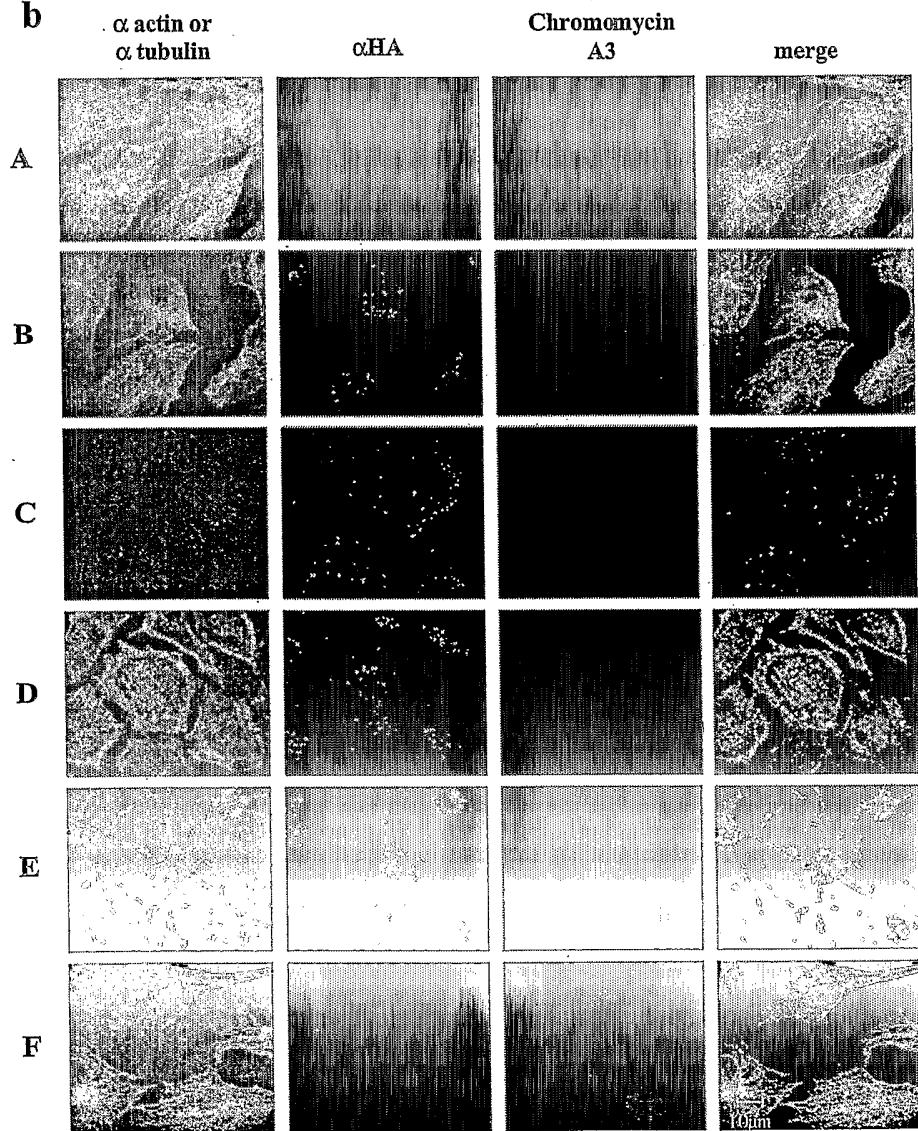


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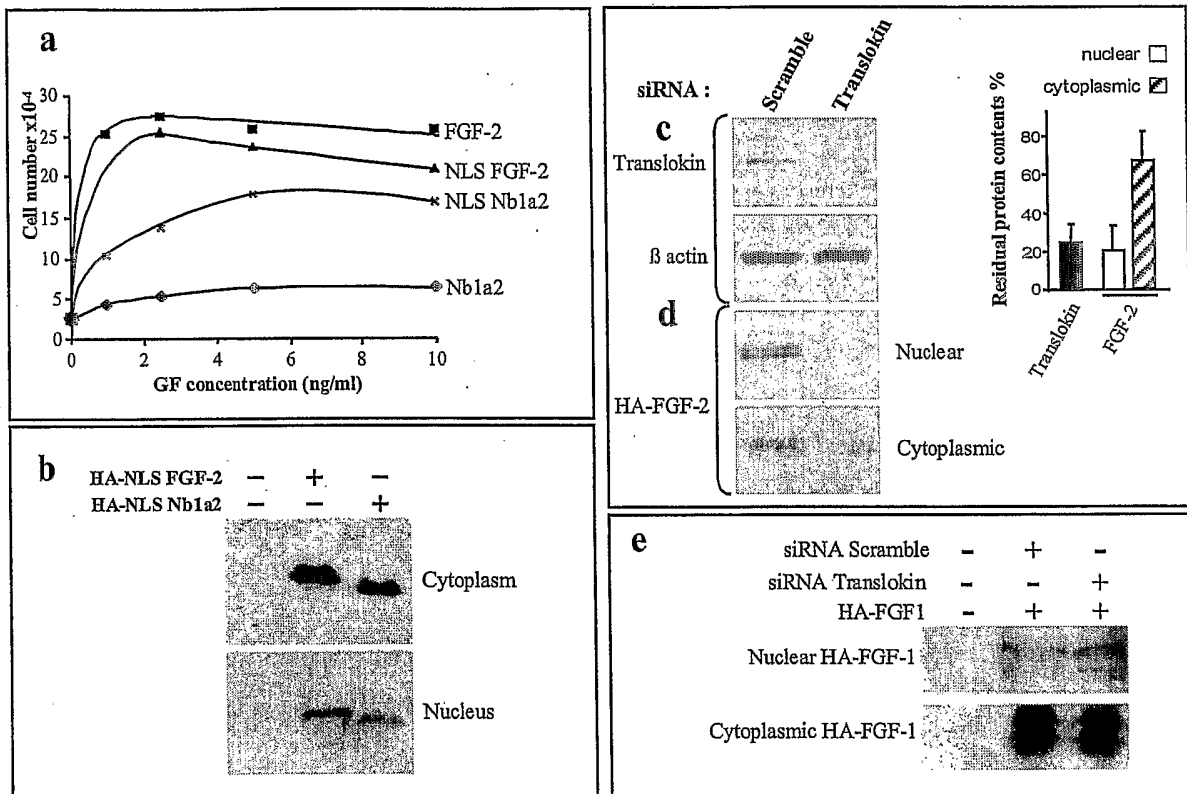


Figure 7

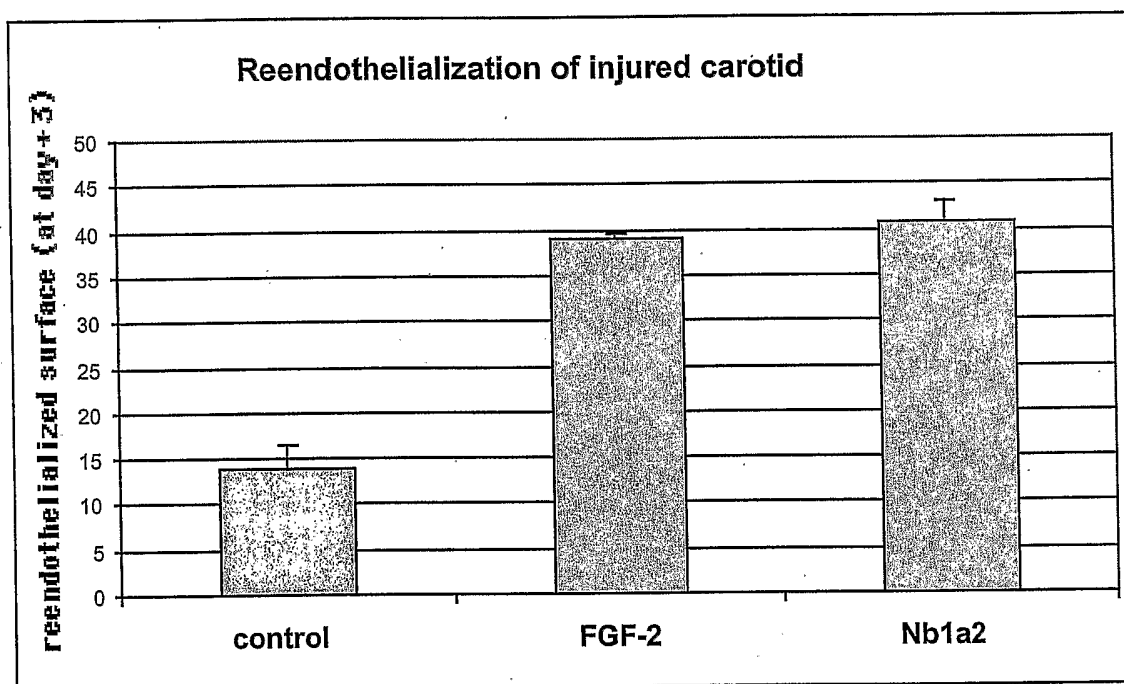


Figure 8

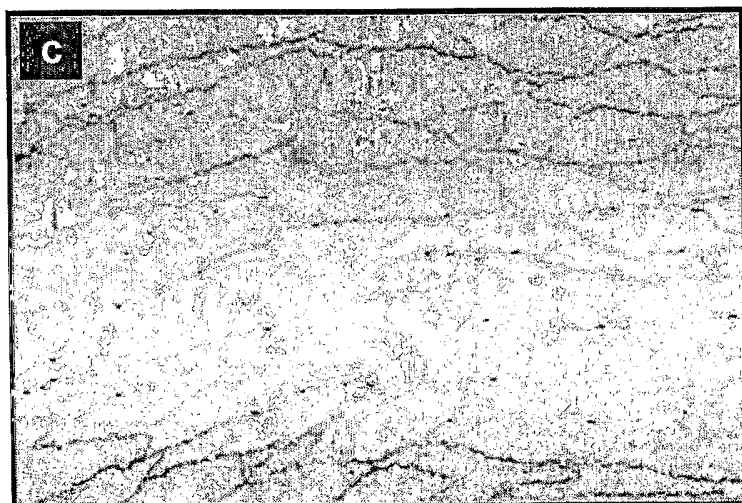
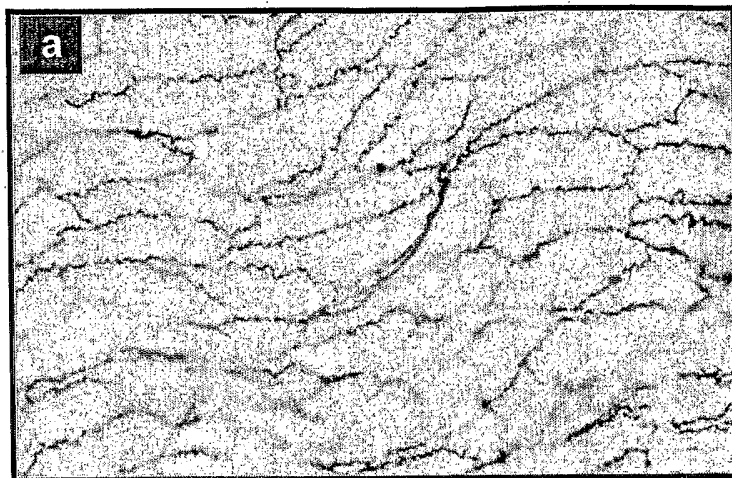


Figure 9

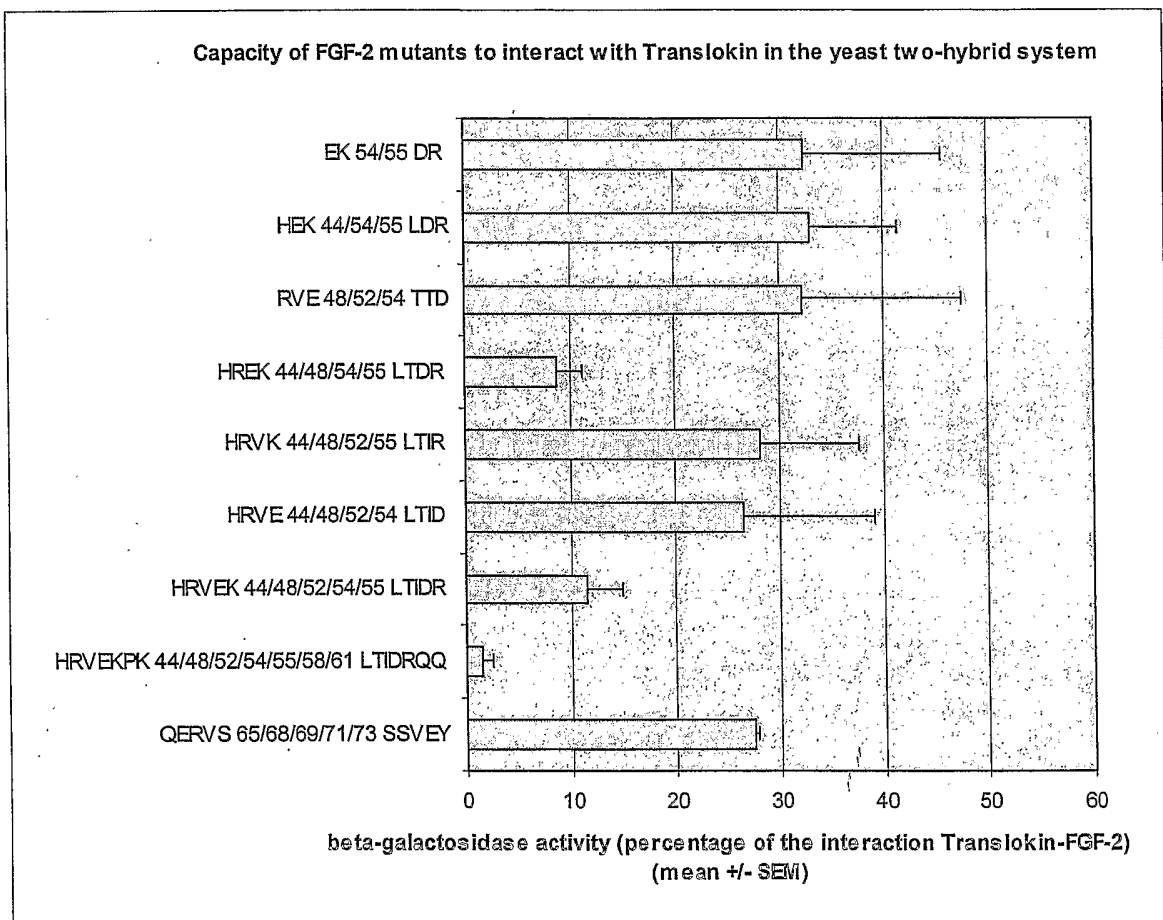
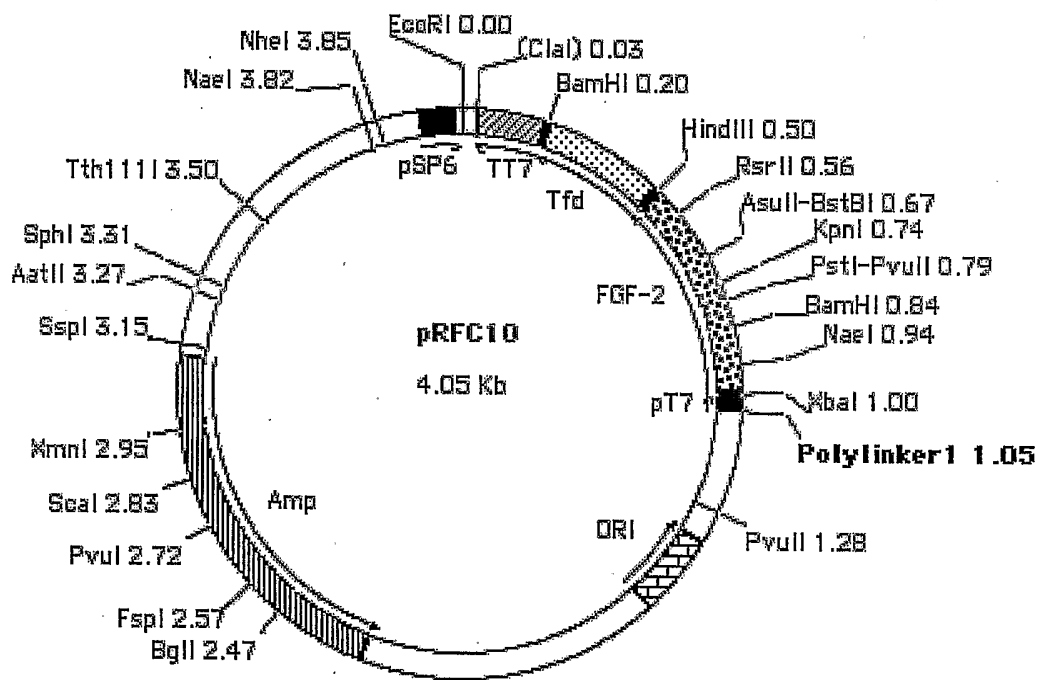


Figure 10



Polylinker1 : 1.05/WbaI.Sall.AccI.HinII.PstI.HindIII.

Figure 11

SEQUENCE LISTING

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Asp Leu Arg Arg Ser Pro Ser Lys Pro Thr Leu Ala Tyr Pro Glu Ser
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Asn Ser Arg Ala Ile Phe Ser Ala Leu Lys Asn Leu Gln Asp Lys Ile
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	Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys					
	130		135		140	
40	gcg atc ctg ttc ctg ccg atg tct gcg aaa tct taa					468
	Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser					
	145		150		155	
45	<210> 4					
	<211> 155					
	<212> PRT					
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	1		5		10	15
55	Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu					
	20		25		30	
60	Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile His Pro Asp Gly Arg					
	35		40		45	
65	Val Asp Gly Val Arg Glu Lys Ser Asp Pro His Ile Lys Leu Gln Leu					
	50		55		60	
70	Gln Ala Glu Glu Arg Gly Val Val Ser Ile Lys Gly Val Cys Ala Asn					
	65		70		75	80

Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

5 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

10 Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
 115 120 125

15 Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys
 130 135 140

Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155

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 25 <213> Homo sapiens

<220>
 <221> CDS
 <222> (1)..(468)
 30 <223>

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 Met Ala Ala Gly Ser Ile Thr Thr Leu Pro Ala Leu Pro Glu Asp Gly
 35 1 5 10 15

ggc agc ggc gcc ttc ccg ccc ggc cac ttc aag gac ccc aag cgg ctg 96
 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30

40 tac tgc aaa aac ggg ggc ttc ttc ctg cgc atc cac ccc gac ggc cga 144
 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile His Pro Asp Gly Arg
 35 40 45

45 gtt gac ggg gtc cgg gag aag agc gac cct cac atc aag cta caa ctt 192
 Val Asp Gly Val Arg Glu Lys Ser Asp Pro His Ile Lys Leu Gln Leu
 50 55 60

caa gca gaa gag aga gga gtt gtg tct atc aaa gga gtg tgt gct aac 240
 Gln Ala Glu Glu Arg Gly Val Val Ser Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

cgt tac ctg gct atg aag gaa gat gga aga tta ctg gct tct aaa tgt 288
 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 55 85 90 95

gtt acg gat gag tgt ttc ttt ttt gaa cga ttg gaa tct aat aac tac 336
 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

60 aat act tac cgg tca agg aaa tac acc agt tgg tat gtg gca ctg aaa 384

Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
 115 120 125

5 cga act ggg cag tat aaa ctt gga tcc aaa aca gga cct ggg cag aaa 432
 Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
 130 135 140

10 gct ata ctt ttt ctt cca atg tct gct aag agc tga 468
 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155

15 <210> 6
 <211> 155
 <212> PRT
 <213> Homo sapiens

<400> 6

20 Met Ala Ala Gly Ser Ile Thr Thr Leu Pro Ala Leu Pro Glu Asp Gly
 1 5 10 15

25 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30

30 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile His Pro Asp Gly Arg
 35 40 45

35 Val Asp Gly Val Arg Glu Lys Ser Asp Pro His Ile Lys Leu Gln Leu
 50 55 60

40 Gln Ala Glu Glu Arg Gly Val Val Ser Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

45 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

50 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

55 Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
 115 120 125

60 Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
 130 135 140

65 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155

60 <210> 7
 <211> 468

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5 <220>
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<220>
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<221> misc_feature
20 <222> (142)..(144)
<223> cgt or act or acc or aca or acg

<220>
25 <221> misc_feature
<222> (154)..(156)
<223> gtt or act or acc or aca or acg

30 <220>
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<223> a or c or g or t

35 <220>
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40 <223> aaa or cgt or cgc or cga or cgg or aga or agg

<220>
<221> misc_feature
<222> (172)..(174)
45 <223> ccg or caa or cag

<220>
50 <221> misc_feature
<222> (181)..(183)
<223> aaa or caa or cag

<220>
55 <221> misc_feature
<222> (193)..(195)
<223> cag or tct or tcc or tca or tcg or agt or agc

60 <220>
<221> misc_feature

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<222> (202)..(204)
<223> gag or tct or tcc or tca or tcg or agt or agc

5 <220>
   <221> misc_feature
   <222> (205)..(207)
   <223> cgt or gtt or gtc or gta or gtg

10 <220>
    <221> misc_feature
    <222> (211)..(213)
    <223> gtt or gaa or gag

15 <220>
    <221> misc_feature
    <222> (218)..(219)
    <223> ct or at or ac

<400> 7
25 atg gct gca ggt tcc atc act acc ctg ccg gct ctg cct gag gac ggt      48
   Met Ala Ala Gly Ser Ile Thr Thr Leu Pro Ala Leu Pro Glu Asp Gly
   1          5          10          15

   ggt agc ggt gct ttt ccg cca ggc cac ttc aaa gac ccg aaa cgt ctg      96
   Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
   20          25          30

   tac tgc aaa aac ggt ggc ttc ttc ctg cgg atc nnn ccg gac ggt nnn      144
   Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Xaa Pro Asp Gly Xaa
   35          40          45

35 gtt gac ggc nnn cgt gan nnn tct gac nnn cac atc nnn ctt cag ctg      192
   Val Asp Gly Xaa Arg Xaa Xaa Ser Asp Xaa His Ile Xaa Leu Gln Leu
   50          55          60

40 nnn gcg gaa nnn nnn ggt nnn gta tnn atc aaa ggt gtt tgc gcg aac      240
   Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
   65          70          75          80

   cgg tac ctg gcg atg aaa gaa gac ggt cgt ctg ctg gcg tct aaa tgc      288
   Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
   85          90          95

   gtt acc gac gaa tgc ttc ttt ttc gaa cgt ctg gag tct aac aat tac      336
   Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
   100          105          110

   aac acc tat cgt tct cgt aaa tac tct tca tgg tat gtt gcg ctc aaa      384
   Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
   115          120          125

55 gcg acg ggt cag tac aaa ctg ggt cca aaa acc ggt ccg ggt cag aaa      432
   Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys
   130          135          140

60 gcg atc ctg ttc ctg ccg atg tct gcg aaa tct taa      468
   Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser

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145

150

155

- 5 <210> 8
<211> 155
<212> PRT
<213> Artificial
- 10 <220>
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<222> (44)..(44)
<223> The 'Xaa' at location 44 stands for His, or Leu.
- 15 <220>
<221> misc_feature
<222> (48)..(48)
<223> The 'Xaa' at location 48 stands for Arg, or Thr.
- 20 <220>
<221> misc_feature
<222> (52)..(52)
<223> The 'Xaa' at location 52 stands for Val, or Thr.
- 25 <220>
<221> misc_feature
<222> (54)..(54)
<223> The 'Xaa' at location 54 stands for Glu, or Asp.
- 30 <220>
<221> misc_feature
<222> (55)..(55)
<223> The 'Xaa' at location 55 stands for Lys, or Arg.
- 35 <220>
<221> misc_feature
<222> (58)..(58)
<223> The 'Xaa' at location 58 stands for Pro, or Gln.
- 40 <220>
<221> misc_feature
<222> (61)..(61)
<223> The 'Xaa' at location 61 stands for Lys, or Gln.
- 45 <220>
<221> misc_feature
<222> (65)..(65)
<223> The 'Xaa' at location 65 stands for Gln, or Ser.
- 50 <220>
<221> misc_feature
<222> (68)..(68)
<223> The 'Xaa' at location 68 stands for Glu, or Ser.
- 55 <220>
<221> misc_feature
<222> (69)..(69)
<223> The 'Xaa' at location 69 stands for Arg, or Val.
- 60 <220>
<221> misc_feature
<222> (71)..(71)

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<223> The 'Xaa' at location 71 stands for Val, or Glu.

<220>

<221> misc_feature

5 <222> (73)..(73)

<223> The 'Xaa' at location 73 stands for Tyr or Ser.

<220>

10 <223> bovine FGF-2 mutant

<400> 8

Met Ala Ala Gly Ser Ile Thr Thr Leu Pro Ala Leu Pro Glu Asp Gly
1 5 10 15

15

Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
20 25 30

20

Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Xaa Pro Asp Gly Xaa
35 40 45

25

Val Asp Gly Xaa Arg Xaa Xaa Ser Asp Xaa His Ile Xaa Leu Gln Leu
50 55 60

30

Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
65 70 75 80

35

Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
85 90 95

40

Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
100 105 110

Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
115 120 125

45

Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys
130 135 140

50

Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
145 150 155

<210> 9

<211> 468

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<212> DNA

<213> Artificial

<220>

60

<223> Chimera FGF-1 FGF-2

<220>

<221> CDS
 <222> (1)..(468)
 <223>

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 1 5 10 15

10 ggt agc ggt gct ttt ccg cca ggc cac ttc aaa gac ccg aaa cgt ctg 96
 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30

15 tac tgc aaa aac ggt ggc ttc ttc ctg cgg atc ctt ccg gat ggc aca 144
 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr
 35 40 45

20 gtg gat ggg aca agg gac agg agc gac cag cac att cag ctg cag ctc 192
 Val Asp Gly Thr Arg Asp Arg Ser Asp Gln His Ile Gln Leu Gln Leu
 50 55 60

25 agt gcg gaa agc gtg ggg gag gtg tat ata aaa ggt gtt tgc gcg aac 240
 Ser Ala Glu Ser Val Gly Glu Val Tyr Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

30 cgg tac ctg gcg atg aaa gaa gac ggt cgt ctg ctg gcg tct aaa tgc 288
 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

35 gtt acc gac gaa tgc ttc ttt ttc gaa cgt ctg gag tct aac aat tac 336
 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

40 aac acc tat cgt tct cgt aaa tac tct tca tgg tat gtt gcg ctc aaa 384
 Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
 115 120 125

45 cgc acg ggt cag tac aaa ctg ggt cca aaa acc ggt ccg ggt cag aaa 432
 Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys
 130 135 140

50 gcg atc ctg ttc ctg ccg atg tct gcg aaa tct taa 468
 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155

<210> 10
 <211> 155
 <212> PRT
 <213> Artificial

<220>
 <223> Chimera FGF-1 FGF-2

55 <400> 10
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 1 5 10 15

60 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu

		20		25		30	
5	Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr	35		40		45	
10	Val Asp Gly Thr Arg Asp Arg Ser Asp Gln His Ile Gln Leu Gln Leu	50		55		60	
15	Ser Ala Glu Ser Val Gly Glu Val Tyr Ile Lys Gly Val Cys Ala Asn	65		70		75	80
20	Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys		85		90		95
25	Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr		100		105		110
30	Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys		115		120		125
35	Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys		130		135		140
40	Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser		145		150		155
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55	ggt agc ggt gct ttt ccg cca ggc cac ttc aaa gac ccg aaa cgt ctg 96 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu 20 25 30						
60	tac tgc aaa aac ggt ggc ttc ttc ctg cgg atc ctt ccg gat ggc aca 144 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr 35 40 45						

gtg gat ggg aca agg gac agg agc gac cag cac att cag ctg cag ctt 192
 Val Asp Gly Thr Arg Asp Arg Ser Asp Gln His Ile Gln Leu Gln Leu
 50 55 60

5 caa gca gaa gag aga gga gtt gtg tct atc aaa gga gtg tgt gct aac 240
 Gln Ala Glu Glu Arg Gly Val Val Ser Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

10 cgt tac ctg gct atg aag gaa gat gga aga tta ctg gct tct aaa tgt 288
 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

15 gtt acg gat gag tgt ttc ttt ttt gaa cga ttg gaa tct aat aac tac 336
 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

20 aat act tac cgg tca agg aaa tac acc agt tgg tat gtg gca ctg aaa 384
 Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
 115 120 125

25 cga act ggg cag tat aaa ctt gga tcc aaa aca gga cct ggg cag aaa 432
 Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
 130 135 140

30 gct ata ctt ttt ctt cca atg tct gct aag agc tga 468
 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155

30 <210> 12
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 <212> PRT
 <213> Artificial

35 <220>
 <223> Chimera FGF-1 FGF-2
 <400> 12

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 1 5 10 15

45 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30

50 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr
 35 40 45

55 Val Asp Gly Thr Arg Asp Arg Ser Asp Gln His Ile Gln Leu Gln Leu
 50 55 60

60 Gln Ala Glu Glu Arg Gly Val Val Ser Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

60 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110
 5
 Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
 115 120 125
 10
 Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
 130 135 140
 15
 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155
 20
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 <212> DNA
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 25
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 1 5 10 15
 35
 aat ctg cct cca ggg aat tac aag aag ccc aaa ctc ctc tac tgt agc 96
 Asn Leu Pro Pro Gly Asn Tyr Lys Lys Pro Lys Leu Leu Tyr Cys Ser
 20 25 30
 40
 aac ggg ggc cac ttc ctg agg atc ctt ccg gat ggc aca gtg gat ggg 144
 Asn Gly Gly His Phe Leu Arg Ile Leu Pro Asp Gly Thr Val Asp Gly
 35 40 45
 45
 aca agg gac agg agc gac cag cac att cag ctg cag ctc agt gcg gaa 192
 Thr Arg Asp Arg Ser Asp Gln His Ile Gln Leu Gln Leu Ser Ala Glu
 50 55 60
 50
 agc gtg ggg gag gtg tat ata aag agt acc gag act ggc cag tac ttg 240
 Ser Val Gly Glu Val Tyr Ile Lys Ser Thr Glu Thr Gly Gln Tyr Leu
 65 70 75 80
 55
 gcc atg gac acc gac ggg ctt tta tac ggc tca cag aca cca aat gag 288
 Ala Met Asp Thr Asp Gly Leu Leu Tyr Gly Ser Gln Thr Pro Asn Glu
 85 90 95
 60
 gaa tgt ttg ttc ctg gaa agg ctg gag gag aac cat tac aac acc tat 336
 Glu Cys Leu Phe Leu Glu Arg Leu Glu Glu Asn His Tyr Asn Thr Tyr
 100 105 110
 60
 ata tcc aag aag cat gca gag aag aat tgg ttt gtt ggc ctc aag aag 384
 Ile Ser Lys Lys His Ala Glu Lys Asn Trp Phe Val Gly Leu Lys Lys
 115 120 125

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aat ggg agc tgc aaa cgc ggt cct cgg act cac tat ggc cag aaa gca 432
 Asn Gly Ser Cys Lys Arg Gly Pro Arg Thr His Tyr Gly Gln Lys Ala
 130 135 140

5 atc ttg ttt ctc ccc ctg cca gtc tct tct gat taa 468
 Ile Leu Phe Leu Pro Leu Pro Val Ser Ser Asp
 145 150 155

10 <210> 14
 <211> 155
 <212> PRT
 <213> Homo sapiens

15 <400> 14
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20 Asn Leu Pro Pro Gly Asn Tyr Lys Lys Pro Lys Leu Leu Tyr Cys Ser
 20 25 30

25 Asn Gly Gly His Phe Leu Arg Ile Leu Pro Asp Gly Thr Val Asp Gly
 35 40 45

30 Thr Arg Asp Arg Ser Asp Gln His Ile Gln Leu Gln Leu Ser Ala Glu
 50 55 60

35 Ser Val Gly Glu Val Tyr Ile Lys Ser Thr Glu Thr Gly Gln Tyr Leu
 65 70 75 80

Ala Met Asp Thr Asp Gly Leu Leu Tyr Gly Ser Gln Thr Pro Asn Glu
 85 90 95

40 Glu Cys Leu Phe Leu Glu Arg Leu Glu Glu Asn His Tyr Asn Thr Tyr
 100 105 110

45 Ile Ser Lys Lys His Ala Glu Lys Asn Trp Phe Val Gly Leu Lys Lys
 115 120 125

50 Asn Gly Ser Cys Lys Arg Gly Pro Arg Thr His Tyr Gly Gln Lys Ala
 130 135 140

55 Ile Leu Phe Leu Pro Leu Pro Val Ser Ser Asp
 145 150 155

60 <210> 15
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<220>
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<223>

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<222> (331)..(333)
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20 <220>
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<222> (346)..(348)
<223> cgt or act or acc or aca or acg

25 <220>
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<222> (352)..(354)
<223> cgt or aaa or aag

30 <220>
<221> misc_feature
<222> (358)..(360)
<223> tac or cat or cac or gaa or gag or gct or gcc or gca or gcg

35 <220>
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<222> (361)..(363)
<223> tct or gct or gcc or gca or gcg or gaa or gag or aaa or aag

40 <220>
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<223> tca or gaa or gag or aaa or aag or aat or aac

45 <220>
<221> misc_feature
<222> (370)..(372)
<223> tat or ttt or ttc

50 <220>
<221> misc_feature
<222> (376)..(378)
<223> gcg or ggt or ggc or gga or ggg

55 <220>
<221> misc_feature
<222> (376)..(378)
<223> gcg or ggt or ggc or gga or ggg

60 <220>
<221> misc_feature
<222> (376)..(378)
<223> gcg or ggt or ggc or gga or ggg

<400> 15
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 Met Ala Ala Gly Ser Ile Thr Thr Leu Pro Ala Leu Pro Glu Asp Gly
 5 1 5 10 15

ggt agc ggt gct ttt ccg cca ggc cac ttc aaa gac ccg aaa cgt ctg 96
 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 10 20 25 30

tac tgc aaa aac ggt ggc ttc ttc ctg ccg atc cac ccg gac ggt cgt 144
 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile His Pro Asp Gly Arg
 35 40 45

gtt gac ggc gtt cgt gaa aaa tct gac ccg cac atc aaa ctt cag ctg 192
 Val Asp Gly Val Arg Glu Lys Ser Asp Pro His Ile Lys Leu Gln Leu
 50 55 60

cag gcg gaa gag cgt ggt gtt gta tct atc aaa ggt gtt tgc gcg aac 240
 Gln Ala Glu Glu Arg Gly Val Val Ser Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

ccg tac ctg gcg atg aaa gaa gac ggt cgt ctg ctg gcg tct aaa tgc 288
 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

gtt acc gac gaa tgc ttc ttt ttc gaa cgt ctg gag nnn aac nnn tac 336
 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Xaa Asn Xaa Tyr
 100 105 110

aac acc tat nnn tct nnn aaa nnn nnn nnn tgg nnn gtt nnn ctc aaa 384
 Asn Thr Tyr Xaa Ser Xaa Lys Xaa Xaa Xaa Trp Xaa Val Xaa Leu Lys
 115 120 125

cgc acg ggt cag tac aaa ctg ggt cca aaa acc ggt ccg ggt cag aaa 432
 Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys
 130 135 140

gcg atc ctg ttc ctg ccg atg tct gcg aaa tct taa 468
 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155

<210> 16
 <211> 155
 <212> PRT
 <213> Artificial

<220>
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 <223> The 'Xaa' at location 109 stands for Ser, or Glu.

<220>
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 <222> (111)..(111)
 <223> The 'Xaa' at location 111 stands for Asn, or His.

<220>
 <221> misc_feature
 <222> (116)..(116)

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<223> The 'Xaa' at location 116 stands for Arg, or Thr.
 <220>
 <221> misc_feature
 5 <222> (118)..(118)
 <223> The 'Xaa' at location 118 stands for Arg, or Lys.
 <220>
 <221> misc_feature
 10 <222> (120)..(120)
 <223> The 'Xaa' at location 120 stands for Tyr, His, Ala, or Glu.
 <220>
 <221> misc_feature
 15 <222> (121)..(121)
 <223> The 'Xaa' at location 121 stands for Ser, Ala, Glu, or Lys.
 <220>
 <221> misc_feature
 20 <222> (122)..(122)
 <223> The 'Xaa' at location 122 stands for Ser, Glu, Lys, or Asn.
 <220>
 <221> misc_feature
 25 <222> (124)..(124)
 <223> The 'Xaa' at location 124 stands for Tyr, or Phe.
 <220>
 <221> misc_feature
 30 <222> (126)..(126)
 <223> The 'Xaa' at location 126 stands for Ala, or Gly.
 <220>
 <223> bovine FGF-2 mutant
 35 <400> 16
 Met Ala Ala Gly Ser Ile Thr Thr Leu Pro Ala Leu Pro Glu Asp Gly
 1 5 10 15
 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30
 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile His Pro Asp Gly Arg
 35 40 45
 Val Asp Gly Val Arg Glu Lys Ser Asp Pro His Ile Lys Leu Gln Leu
 50 55 60
 Gln Ala Glu Glu Arg Gly Val Val Ser Ile Lys Gly Val Cys Ala Asn
 65 70 75 80
 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95
 60

	Val	Thr	Asp	Glu	Cys	Phe	Phe	Phe	Glu	Arg	Leu	Glu	Xaa	Asn	Xaa	Tyr	
				100					105					110			
5	Asn	Thr	Tyr	Xaa	Ser	Xaa	Lys	Xaa	Xaa	Xaa	Trp	Xaa	Val	Xaa	Leu	Lys	
			115				120						125				
10	Arg	Thr	Gly	Gln	Tyr	Lys	Leu	Gly	Pro	Lys	Thr	Gly	Pro	Gly	Gln	Lys	
			130				135					140					
15	Ala	Ile	Leu	Phe	Leu	Pro	Met	Ser	Ala	Lys	Ser						
	145					150					155						
	<210>	17															
	<211>	468															
	<212>	DNA															
20	<213>	Artificial															
	<220>																
	<223>	Chimera	FGF-1	FGF-2													
25	<220>																
	<221>	CDS															
	<222>	(1)..(468)															
	<223>																
30	<400>	17															
	atg	gca	gcc	ggg	agc	atc	acc	acg	ctg	ccc	gcc	ttg	ccc	gag	gat	ggc	48
	Met	Ala	Ala	Gly	Ser	Ile	Thr	Thr	Leu	Pro	Ala	Leu	Pro	Glu	Asp	Gly	
	1			5					10					15			
35	ggc	agc	ggc	gcc	ttc	ccg	ccc	ggc	cac	ttc	aag	gac	ccc	aag	cgg	ctg	96
	Gly	Ser	Gly	Ala	Phe	Pro	Pro	Gly	His	Phe	Lys	Asp	Pro	Lys	Arg	Leu	
			20					25					30				
40	tac	tgc	aaa	aac	ggg	ggc	ttc	ttc	ctg	cgc	atc	cac	ccc	gac	ggc	cga	144
	Tyr	Cys	Lys	Asn	Gly	Gly	Phe	Phe	Leu	Arg	Ile	His	Pro	Asp	Gly	Arg	
			35				40					45					
45	gtt	gac	ggg	gtc	cgg	gag	aag	agc	gac	cct	cac	atc	aag	cta	caa	ctt	192
	Val	Asp	Gly	Val	Arg	Glu	Lys	Ser	Asp	Pro	His	Ile	Lys	Leu	Gln	Leu	
		50				55			60								
50	caa	gca	gaa	gag	aga	gga	gtt	gtg	tct	atc	aaa	gga	gtg	tgt	gct	aac	240
	Gln	Ala	Glu	Glu	Arg	Gly	Val	Val	Ser	Ile	Lys	Gly	Val	Cys	Ala	Asn	
	65				70				75						80		
55	cgt	tac	ctg	gct	atg	aag	gaa	gat	gga	aga	tta	ctg	gct	tct	aaa	tgt	288
	Arg	Tyr	Leu	Ala	Met	Lys	Glu	Asp	Gly	Arg	Leu	Leu	Ala	Ser	Lys	Cys	
				85					90					95			
60	gtt	acg	gat	gag	tgt	ttc	ttt	ttt	gaa	cga	ttg	gaa	tac	aac	acc	tat	336
	Val	Thr	Asp	Glu	Cys	Phe	Phe	Phe	Glu	Arg	Leu	Glu	Tyr	Asn	Thr	Tyr	
				100					105					110			
65	ata	tcc	aag	aag	cat	gca	gag	aag	aat	tgg	ttt	gtt	ggc	ctc	ctg	aaa	384
	Ile	Ser	Lys	Lys	His	Ala	Glu	Lys	Asn	Trp	Phe	Val	Gly	Leu	Leu	Lys	
			115					120					125				

cga act ggg cag tat aaa ctt gga tcc aaa aca gga cct ggg cag aaa 432
 Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
 130 135 140

5

gct ata ctt ttt ctt cca atg tct gct aag agc tga 468
 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155

10

<210> 18
 <211> 155
 <212> PRT
 <213> Artificial

15

<220>
 <223> Chimera FGF-1 FGF-2

20

Met Ala Ala Gly Ser Ile Thr Thr Leu Pro Ala Leu Pro Glu Asp Gly
 1 5 10 15

25

Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30

30

Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile His Pro Asp Gly Arg
 35 40 45

35

Val Asp Gly Val Arg Glu Lys Ser Asp Pro His Ile Lys Leu Gln Leu
 50 55 60

40

Gln Ala Glu Glu Arg Gly Val Val Ser Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

45

Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

50

Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Tyr Asn Thr Tyr
 100 105 110

55

Ile Ser Lys Lys His Ala Glu Lys Asn Trp Phe Val Gly Leu Leu Lys
 115 120 125

60

Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
 130 135 140

Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155

<210> 19

<211> 468
<212> DNA
<213> Artificial

5 <220>
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<220>
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10 <222> (1)..(468)
<223>

<220>
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15 <222> (130)..(132)
<223> cac or tta or ttg or ctt or ctc or cta or ctg

<220>
<221> misc_feature
20 <222> (142)..(144)
<223> cga or act or acc or aca or acg

<220>
<221> misc_feature
25 <222> (154)..(156)
<223> gtc or act or acc or aca or acg

<220>
<221> misc_feature
30 <222> (162)..(162)
<223> a or c or g or t

35 <220>
<221> misc_feature
<222> (163)..(165)
40 <223> aag or cgt or cgc or cga or cgg or aga or agg

<220>
<221> misc_feature
45 <222> (172)..(174)
<223> cct or caa or cag

<220>
<221> misc_feature
50 <222> (181)..(183)
<223> aag or caa or cag

<220>
<221> misc_feature
55 <222> (193)..(195)
<223> caa or tct or tcc or tca or tcg or agt or agc

60 <220>

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<221> misc_feature
<222> (202)..(204)
<223> gag or tct or tcc or tca or tcg or agt or agc

5
<220>
<221> misc_feature
<222> (205)..(207)
<223> aga or gtt or gtc or gta or gtg

10
<220>
<221> misc_feature
<222> (211)..(213)
<223> gtt or gaa or gag

15
<220>
<221> misc_feature
<222> (218)..(219)
<223> ct or at or ac

20
<400> 19
25 atg gca gcc ggg agc atc acc acg ctg ccc gcc ttg ccc gag gat ggc      48
   Met Ala Ala Gly Ser Ile Thr Thr Leu Pro Ala Leu Pro Glu Asp Gly
   1          5          10          15

   ggc agc ggc gcc ttc ccg ccc ggc cac ttc aag gac ccc aag cgg ctg      96
   Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
   20          25          30

   tac tgc aaa aac ggg ggc ttc ttc ctg cgc atc nnn ccc gac ggc nnn      144
   Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Xaa Pro Asp Gly Xaa
   35          40          45

   gtt gac ggg nnn cgg gan nnn agc gac nnn cac atc nnn cta caa ctt      192
   Val Asp Gly Xaa Arg Xaa Xaa Ser Asp Xaa His Ile Xaa Leu Gln Leu
   50          55          60

   nnn gca gaa nnn nnn gga nnn gtg tnn atc aaa gga gtg tgt gct aac      240
   Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
   65          70          75          80

   cgt tac ctg gct atg aag gaa gat gga aga tta ctg gct tct aaa tgt      288
   Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
   85          90          95

   gtt acg gat gag tgt ttc ttt ttt gaa cga ttg gaa tct aat aac tac      336
   Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
   100         105         110

   aat act tac cgg tca agg aaa tac acc agt tgg tat gtg gca ctg aaa      384
   Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
   115         120         125

   cga act ggg cag tat aaa ctt gga tcc aaa aca gga cct ggg cag aaa      432
   Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
   130         135         140

60 gct ata ctt ttt ctt cca atg tct gct aag agc tga      468

```

Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
145 150 155

- 5 <210> 20
<211> 155
<212> PRT
<213> Artificial
- 10 <220>
<221> misc_feature
<222> (44)..(44)
<223> The 'Xaa' at location 44 stands for His, or Leu.
- 15 <220>
<221> misc_feature
<222> (48)..(48)
<223> The 'Xaa' at location 48 stands for Arg, or Thr.
- 20 <220>
<221> misc_feature
<222> (52)..(52)
<223> The 'Xaa' at location 52 stands for Val, or Thr.
- 25 <220>
<221> misc_feature
<222> (54)..(54)
<223> The 'Xaa' at location 54 stands for Glu, or Asp.
- 30 <220>
<221> misc_feature
<222> (55)..(55)
<223> The 'Xaa' at location 55 stands for Lys, or Arg.
- 35 <220>
<221> misc_feature
<222> (58)..(58)
<223> The 'Xaa' at location 58 stands for Pro, or Gln.
- 40 <220>
<221> misc_feature
<222> (61)..(61)
<223> The 'Xaa' at location 61 stands for Lys, or Gln.
- 45 <220>
<221> misc_feature
<222> (65)..(65)
<223> The 'Xaa' at location 65 stands for Gln, or Ser.
- 50 <220>
<221> misc_feature
<222> (68)..(68)
<223> The 'Xaa' at location 68 stands for Glu, or Ser.
- 55 <220>
<221> misc_feature
<222> (69)..(69)
<223> The 'Xaa' at location 69 stands for Arg, or Val.
- 60 <220>
<221> misc_feature

<222> (71)..(71)

<223> The 'Xaa' at location 71 stands for Val, or Glu.

<220>

5 <221> misc_feature

<222> (73)..(73)

<223> The 'Xaa' at location 73 stands for Tyr or Ser.

<220>

10 <223> human FGF-2 mutant

<400> 20

15 Met Ala Ala Gly Ser Ile Thr Thr Leu Pro Ala Leu Pro Glu Asp Gly
1 5 10 15

Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
20 20 25 30

Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Xaa Pro Asp Gly Xaa
35 40 45

25 Val Asp Gly Xaa Arg Xaa Xaa Ser Asp Xaa His Ile Xaa Leu Gln Leu
50 55 60

30 Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
65 70 75 80

35 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
85 90 95

40 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
100 105 110

Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
115 120 125

45 Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
130 135 140

50 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
145 150 155

<210> 21

55 <211> 468

<212> DNA

<213> Artificial

<220>

60 <223> human FGF-2 mutant

<220>
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<222> (1)..(468)
<223>
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<220>
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<222> (325)..(327)
<223> tct or gaa or gag
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<220>
<221> misc_feature
<222> (331)..(333)
<223> aac or cat or cac
15
<220>
<221> misc_feature
<222> (346)..(348)
<223> cgg or act or acc or aca or acg
20
<220>
<221> misc_feature
<222> (352)..(354)
<223> agg or aaa or aag
25
<220>
<221> misc_feature
<222> (358)..(360)
<223> tac or cat or cac or gaa or gag or gct or gcc or gca or gcg
30
<220>
<221> misc_feature
<222> (361)..(363)
<223> acc or gct or gcc or gca or gcg or gaa or gag or aaa or aag
35
<220>
<221> misc_feature
<222> (364)..(366)
<223> agt or gaa or gag or aaa or aag or aat or aac
40
<220>
<221> misc_feature
<222> (370)..(372)
<223> tat or ttt or ttc
45
<220>
<221> misc_feature
<222> (376)..(378)
<223> gca or ggt or ggc or gga or ggg
50
<400> 21
atg gca gcc ggg agc atc acc acg ctg ccc gcc ttg ccc gag gat ggc
55
60

<221> misc_feature
 <222> (118)..(118)
 <223> The 'Xaa' at location 118 stands for Arg, or Lys.

5 <220>
 <221> misc_feature
 <222> (120)..(120)
 <223> The 'Xaa' at location 120 stands for Tyr, His, Ala, or Glu.

10 <220>
 <221> misc_feature
 <222> (121)..(121)
 <223> The 'Xaa' at location 121 stands for Thr, Ala, Glu, or Lys.

15 <220>
 <221> misc_feature
 <222> (122)..(122)
 <223> The 'Xaa' at location 122 stands for Ser, Glu, Lys, or Asn.

20 <220>
 <221> misc_feature
 <222> (124)..(124)
 <223> The 'Xaa' at location 124 stands for Tyr, or Phe.

25 <220>
 <221> misc_feature
 <222> (126)..(126)
 <223> The 'Xaa' at location 126 stands for Ala, or Gly.

30 <220>
 <223> human FGF-2 mutant

<400> 22

35 Met Ala Ala Gly Ser Ile Thr Thr Leu Pro Ala Leu Pro Glu Asp Gly
 1 5 10 15

Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30

Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile His Pro Asp Gly Arg
 35 40 45

45 Val Asp Gly Val Arg Glu Lys Ser Asp Pro His Ile Lys Leu Gln Leu
 50 55 60

50 Gln Ala Glu Glu Arg Gly Val Val Ser Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

55 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

60 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Xaa Asn Xaa Tyr
 100 105 110

Asn Thr Tyr Xaa Ser Xaa Lys Xaa Xaa Xaa Trp Xaa Val Xaa Leu Lys
 115 120 125

5
 Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
 130 135 140

10
 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155

15
 <210> 23
 <211> 465
 <212> DNA
 <213> *Xenopus laevis*

20
 <220>
 <221> CDS
 <222> (1)..(465)
 <223>

25
 <400> 23
 atg gcg gca ggg agc atc aca act ctg cca act gaa tcc gag gat ggg 48
 Met Ala Ala Gly Ser Ile Thr Thr Leu Pro Thr Glu Ser Glu Asp Gly
 1 5 10 15

30
 gga aac act cct ttt tca cca ggg agt ttt aaa gac ccc aag agg ctc 96
 Gly Asn Thr Pro Phe Ser Pro Gly Ser Phe Lys Asp Pro Lys Arg Leu
 20 25 30

35
 tac tgc aag aac ggg ggc ttc ttc ctc agg ata aac tca gac ggg aga 144
 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Asn Ser Asp Gly Arg
 35 40 45

40
 gtg gac ggg tca agg gac aaa agt gac tcg cac ata aaa tta cag cta 192
 Val Asp Gly Ser Arg Asp Lys Ser Asp Ser His Ile Lys Leu Gln Leu
 50 55 60

45
 caa gct gta gag cgg gga gtg gta tca ata aag gga atc act gca aat 240
 Gln Ala Val Glu Arg Gly Val Val Ser Ile Lys Gly Ile Thr Ala Asn
 65 70 75 80

50
 cgc tac ctt gcc atg aag gaa gat ggg aga tta aca tcg ctg agg tgt 288
 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Thr Ser Leu Arg Cys
 85 90 95

55
 ata aca gat gaa tgc ttc ttt ttt gaa cga ctg gaa gct aat aac tac 336
 Ile Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ala Asn Asn Tyr
 100 105 110

60
 aac act tac cgg tct cgg aaa tac agc agc tgg tat gtg gca cta aag 384
 Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
 115 120 125

65
 cga acc ggg cag tac aaa aat gga tcg agc act gga ccg gga caa aaa 432
 Arg Thr Gly Gln Tyr Lys Asn Gly Ser Ser Thr Gly Pro Gly Gln Lys
 130 135 140

70
 gct att tta ttt ctc cca atg tcc gca aag agc 465

31/94

Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155

- 5 <210> 24
 <211> 155
 <212> PRT
 <213> Xenopus laevis
- 10 <400> 24
- Met Ala Ala Gly Ser Ile Thr Thr Leu Pro Thr Glu Ser Glu Asp Gly
 1 5 10 15
- 15 Gly Asn Thr Pro Phe Ser Pro Gly Ser Phe Lys Asp Pro Lys Arg Leu
 20 25 30
- 20 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Asn Ser Asp Gly Arg
 35 40 45
- 25 Val Asp Gly Ser Arg Asp Lys Ser Asp Ser His Ile Lys Leu Gln Leu
 50 55 60
- 30 Gln Ala Val Glu Arg Gly Val Val Ser Ile Lys Gly Ile Thr Ala Asn
 65 70 75 80
- Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Thr Ser Leu Arg Cys
 85 90 95
- 35 Ile Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ala Asn Asn Tyr
 100 105 110
- 40 Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
 115 120 125
- 45 Arg Thr Gly Gln Tyr Lys Asn Gly Ser Ser Thr Gly Pro Gly Gln Lys
 130 135 140
- 50 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155
- 55 <210> 25
 <211> 468
 <212> DNA
 <213> Artificial
- <220>
 <223> bovine FGF-2 mutant
- 60 <220>
 <221> CDS

<222> (1) .. (468)
<223>

5 <220>
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<222> (130) .. (132)
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10 <220>
<221> misc_feature
<222> (142) .. (144)
<223> act or acc or aca or acg

15 <220>
<221> misc_feature
<222> (154) .. (156)
<223> act or acc or aca or acg

20 <220>
<221> misc_feature
<222> (162) .. (162)
<223> c or t

25 <220>
<221> misc_feature
<222> (163) .. (165)
<223> aaa or cgt or cgc or cga or cgg or aga or agg

30 <220>
<221> misc_feature
<222> (172) .. (174)
<223> ccg or caa or cag

35 <220>
<221> misc_feature
<222> (181) .. (183)
<223> aaa or caa or cag

40 <220>
<221> misc_feature
<222> (193) .. (195)
<223> cag or tct or tcc or tca or tcg or agt or agc

45 <220>
<221> misc_feature
<222> (202) .. (204)
<223> gag or tct or tcc or tca or tcg or agt or agc

50 <220>
<221> misc_feature
<222> (205) .. (207)
<223> cgt or gtt or gtc or gta or gtg

55 <220>
<221> misc_feature
<222> (205) .. (207)
<223> cgt or gtt or gtc or gta or gtg

60 <220>
<221> misc_feature
<222> (205) .. (207)
<223> cgt or gtt or gtc or gta or gtg

<220>
 <221> misc_feature
 5 <222> (211)..(213)
 <223> gtt or gaa or gag

<220>
 10 <221> misc_feature
 <222> (218)..(219)
 <223> ct or at or ac

15 <400> 25
 atg gct gca ggt tcc atc act acc ctg ccg gct ctg cct gag gac ggt 48
 Met Ala Ala Gly Ser Ile Thr Thr Leu Pro Ala Leu Pro Glu Asp Gly
 1 5 10 15

20 ggt agc ggt gct ttt ccg cca ggc cac ttc aaa gac ccg aaa cgt ctg 96
 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30

25 tac tgc aaa aac ggt ggc ttc ttc ctg cgg atc nnn ccg gac ggt nnn 144
 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Xaa Pro Asp Gly Thr
 35 40 45

30 gtt gac ggc nnn cgt gan nnn tct gac nnn cac atc nnn ctt cag ctg 192
 Val Asp Gly Thr Arg Asp Xaa Ser Asp Xaa His Ile Xaa Leu Gln Leu
 50 55 60

nnn gcg gaa nnn nnn ggt nnn gta tnn atc aaa ggt gtt tgc gcg aac 240
 Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

35 cgg tac ctg gcg atg aaa gaa gac ggt cgt ctg ctg gcg tct aaa tgc 288
 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

40 gtt acc gac gaa tgc ttc ttt ttc gaa cgt ctg gag tct aac aat tac 336
 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

45 aac acc tat cgt tct cgt aaa tac tct tca tgg tat gtt gcg ptc aaa 384
 Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
 115 120 125

50 cgc acg ggt cag tac aaa ctg ggt cca aaa acc ggt ccg ggt cag aaa 432
 Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys
 130 135 140

55 gcg atc ctg ttc ctg ccg atg tct gcg aaa tct taa 468
 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155

<210> 26
 <211> 155
 <212> PRT
 60 <213> Artificial

<220>
 <221> misc_feature
 <222> (44)..(44)
 <223> The 'Xaa' at location 44 stands for His, or Leu.

5

<220>
 <221> misc_feature
 <222> (55)..(55)
 <223> The 'Xaa' at location 55 stands for Lys, or Arg.

10

<220>
 <221> misc_feature
 <222> (58)..(58)
 <223> The 'Xaa' at location 58 stands for Pro, or Gln.

15

<220>
 <221> misc_feature
 <222> (61)..(61)
 <223> The 'Xaa' at location 61 stands for Lys, or Gln.

20

<220>
 <221> misc_feature
 <222> (65)..(65)
 <223> The 'Xaa' at location 65 stands for Gln, or Ser.

25

<220>
 <221> misc_feature
 <222> (68)..(68)
 <223> The 'Xaa' at location 68 stands for Glu, or Ser.

30

<220>
 <221> misc_feature
 <222> (69)..(69)
 <223> The 'Xaa' at location 69 stands for Arg, or Val.

35

<220>
 <221> misc_feature
 <222> (71)..(71)
 <223> The 'Xaa' at location 71 stands for Val, or Glu.

40

<220>
 <221> misc_feature
 <222> (73)..(73)
 <223> The 'Xaa' at location 73 stands for Tyr or Ser.

45

<220>
 <223> bovine FGF-2 mutant

50

<400> 26
 Met Ala Ala Gly Ser Ile Thr Thr Leu Pro Ala Leu Pro Glu Asp Gly
 1 5 10 15

55

Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30

60

Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Xaa Pro Asp Gly Thr
 35 40 45

Val Asp Gly Thr Arg Asp Xaa Ser Asp Xaa His Ile Xaa Leu Gln Leu
 50 55 60

5

Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

10

Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

15

Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

20

Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
 115 120 125

25

Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys
 130 135 140

30

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1 5 10 15

ggc agc ggc gcc ttc ccg ccc ggc cac ttc aag gac ccc aag cgg ctg 96
60 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
20 25 30

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	tac tgc aaa aac ggg ggc ttc ttc ctg cgc atc nnn ccc gac ggc nnn	144
	Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Xaa Pro Asp Gly Thr	
	35 40 45	
5		
	gtt gac ggg nnn cgg gan nnn agc gac nnn cac atc nnn cta caa ctt	192
	Val Asp Gly Thr Arg Asp Xaa Ser Asp Xaa His Ile Xaa Leu Gln Leu	
	50 55 60	
10		
	nnn gca gaa nnn nnn gga nnn gtg tnn atc aaa gga gtg tgt gct aac	240
	Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn	
	65 70 75 80	
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	cgt tac ctg gct atg aag gaa gat gga aga tta ctg gct tct aaa tgt	288
	Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys	
	85 90 95	
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	gtt acg gat gag tgt ttc ttt ttt gaa cga ttg gaa tct aat aac tac	336
	Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr	
	100 105 110	
25		
	aat act tac cgg tca agg aaa tac acc agt tgg tat gtg gca ctg aaa	384
	Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys	
	115 120 125	
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	cga act ggg cag tat aaa ctt gga tcc aaa aca gga cct ggg cag aaa	432
	Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys	
	130 135 140	
35		
	gct ata ctt ttt ctt cca atg tct gct aag agc tga	468
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 35 40 45

40 Val Asp Gly Thr Arg Asp Xaa Ser Asp Xaa His Ile Xaa Leu Gln Leu
 50 55 60

45 Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

50 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

55 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
 115 120 125

60 Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys

130

135

140

5 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
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35 1          5          10          15

ggt agc ggt gct ttt ccg cca ggc cac ttc aaa gac ccg aaa cgt ctg      96
Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
40          20          25          30

tac tgc aaa aac ggt ggc ttc ttc ctg cgg atc nnn ccg gac ggt nnn      144
Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr
50          35          40          45

gtt gac ggc nnn cgt gan nnn tct gac nnn cac atc nnn ctt cag ctg      192
Val Asp Gly Thr Arg Xaa Arg Ser Asp Xaa His Ile Xaa Leu Gln Leu
60          50          55          60

nnn gcg gaa nnn nnn ggt nnn gta tnn atc aaa ggt gtt tgc gcg aac      240
Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
70          75          80

cgg tac ctg gcg atg aaa gaa gac ggt cgt ctg ctg gcg tct aaa tgc      288
Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
85          90          95

gtt acc gac gaa tgc ttc ttt ttc gaa cgt ctg gag tct aac aat tac      336
Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
100          105          110

60 aac acc tat cgt tct cgt aaa tac tct tca tgg tat gtt gcg ctc aaa      384

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 5 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
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 10 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr
 35 40 45
 15 Val Asp Gly Thr Arg Xaa Arg Ser Asp Xaa His Ile Xaa Leu Gln Leu
 50 55 60
 20 Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
 65 70 75 80
 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95
 25 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110
 30 Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
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 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30

15 tac tgc aaa aac ggg ggc ttc ttc ctg cgc atc nnn ccc gac ggc nnn 144
 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr
 35 40 45

20 gtt gac ggg nnn cgg gan nnn agc gac nnn cac atc nnn cta caa ctt 192
 Val Asp Gly Thr Arg Xaa Arg Ser Asp Xaa His Ile Xaa Leu Gln Leu
 50 55 60

25 nnn gca gaa nnn nnn gga nnn gtg tnn atc aaa gga gtg tgt gct aac 240
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 65 70 75 80

30 cgt tac ctg gct atg aag gaa gat gga aga tta ctg gct tct aaa tgt 288
 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

35 gtt acg gat gag tgt ttc ttt ttt gaa cga ttg gaa tct aat aac tac 336
 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

40 aat act tac cgg tca agg aaa tac acc agt tgg tat gtg gca ctg aaa 384
 Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
 115 120 125

45 cga act ggg cag tat aaa ctt gga tcc aaa aca gga cct ggg cag aaa 432
 Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
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 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr
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 Val Asp Gly Thr Arg Xaa Arg Ser Asp Xaa His Ile Xaa Leu Gln Leu
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 Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
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 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95
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 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
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Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
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Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
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ggt agc ggt gct ttt ccg cca ggc cac ttc aaa gac ccg aaa cgt ctg 96
45 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
20 25 30

tac tgc aaa aac ggt ggc ttc ttc ctg cgg atc nnn ccg gac ggt nnn 144
50 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr
35 40 45

ggt gac ggc nnn cgt gan nnn tct gac nnn cac atc nnn ctt cag ctg 192
55 Val Asp Gly Thr Arg Asp Xaa Ser Asp Xaa His Ile Xaa Leu Gln Leu
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nnn gcg gaa nnn nnn ggt nnn gta tnn atc aaa ggt gtt tgc gcg aac 240
Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
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Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys

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 100 105 110

10 aac acc tat cgt tct cgt aaa tac tct tca tgg tat gtt gcg ctc aaa 384
 Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
 115 120 125

15 cgc acg ggt cag tac aaa ctg ggt cca aaa acc ggt ccg ggt cag aaa 432
 Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys
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35 40 45
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Val Asp Gly Thr Arg Asp Xaa Ser Asp Xaa His Ile Xaa Leu Gln Leu
50 55 60
25

Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
65 70 75 80
30

Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
85 90 95
35

Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
100 105 110
40

Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
115 120 125
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Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys
130 135 140
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Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
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 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30

tac tgc aaa aac ggg ggc ttc ttc ctg cgc atc nnn ccc gac ggc nnn 144
 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr
 35 40 45

gtt gac ggg nnn cgg gan nnn agc gac nnn cac atc nnn cta caa ctt 192
 Val Asp Gly Thr Arg Asp Xaa Ser Asp Xaa His Ile Xaa Leu Gln Leu
 50 55 60

nnn gca gaa nnn nnn gga nnn gtg tnn atc aaa gga gtg tgt gct aac 240
 Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

cgt tac ctg gct atg aag gaa gat gga aga tta ctg gct tct aaa tgt 288
 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

gtt acg gat gag tgt ttc ttt ttt gaa cga ttg gaa tct aat aac tac 336
 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

aat act tac cgg tca agg aaa tac acc agt tgg tat gtg gca ctg aaa 384
 Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
 115 120 125

cga act ggg cag tat aaa ctt gga tcc aaa aca gga cct ggg cag aaa 432
 Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
 130 135 140

gct ata ctt ttt ctt cca atg tct gct aag agc tga 468
 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155

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 <223> The 'Xaa' at location 61 stands for Lys, or Gln.

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 <223> The 'Xaa' at location 68 stands for Glu, or Ser.

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 <223> The 'Xaa' at location 69 stands for Arg, or Val.

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 <223> The 'Xaa' at location 71 stands for Val, or Glu.

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 35 <222> (73)..(73)
 <223> The 'Xaa' at location 73 stands for Tyr or Ser.

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Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
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Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr
 35 40 45
 55

Val Asp Gly Thr Arg Asp Xaa Ser Asp Xaa His Ile Xaa Leu Gln Leu
 50 55 60
 60

Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
85 90 95

5 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
100 105 110

10 Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
115 120 125

15 Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
130 135 140

Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
145 150 155

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 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
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 35 40 45

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 Val Asp Gly Xaa Arg Xaa Xaa Ser Asp Xaa His Ile Xaa Leu Gln Leu
 50 55 60

nnn gcg gaa nnn nnn ggt nnn gta tnn atc aaa ggt gtt tgc gcg aac 240
 Ser Ala Glu Ser Val Gly Glu Val Tyr Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

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cgg tac ctg gcg atg aaa gaa gac ggt cgt ctg ctg gcg tct aaa tgc 288
 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

10

gtt acc gac gaa tgc ttc ttt ttc gaa cgt ctg gag tct aac aat tac 336
 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

15

aac acc tat cgt tct cgt aaa tac tct tca tgg tat gtt gcg ctc aaa 384
 Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
 115 120 125

20

cgc acg ggt cag tac aaa ctg ggt cca aaa acc ggt ccg ggt cag aaa 432
 Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys
 130 135 140

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gcg atc ctg ttc ctg ccg atg tct gcg aaa tct taa 468
 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
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<220>
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 <223> The 'Xaa' at location 61 stands for Lys, or Gln.

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<220>
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Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30

20

Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Xaa Pro Asp Gly Xaa
 35 40 45

25

Val Asp Gly Xaa Arg Xaa Xaa Ser Asp Xaa His Ile Xaa Leu Gln Leu
 50 55 60

30

Ser Ala Glu Ser Val Gly Glu Val Tyr Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

35

Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

40

Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

45

Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
 115 120 125

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Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys
 130 135 140

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Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155

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ggc agc ggc gcc ttc ccg ccc ggc cac ttc aag gac ccc aag cgg ctg 96
20 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
20 25 30

tac tgc aaa aac ggg ggc ttc ttc ctg cgc atc nnn ccc gac ggc nnn 144
25 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Xaa Pro Asp Gly Xaa
35 40 45

gtt gac ggg nnn cgg gan nnn agc gac nnn cac atc nnn cta caa ctt 192
Val Asp Gly Xaa Arg Xaa Xaa Ser Asp Xaa His Ile Xaa Leu Gln Leu
50 55 60

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Ser Ala Glu Ser Val Gly Glu Val Tyr Ile Lys Gly Val Cys Ala Asn
65 70 75 80

35 cgt tac ctg gct atg aag gaa gat gga aga tta ctg gct tct aaa tgt 288
Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
85 90 95

40 gtt acg gat gag tgt ttc ttt ttt gaa cga ttg gaa tct aat aac tac 336
Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
100 105 110

aat act tac cgg tca agg aaa tac acc agt tgg tat gtg gca ctg aaa 384
45 Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
115 120 125

cga act ggg cag tat aaa ctt gga tcc aaa aca gga cct ggg cag aaa 432
Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
130 135 140

50 gct ata ctt ttt ctt cca atg tct gct aag agc tga 468
Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
145 150 155

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		20					25						30		

50

Tyr	Cys	Lys	Asn	Gly	Gly	Phe	Phe	Leu	Arg	Ile	Xaa	Pro	Asp	Gly	Xaa
		35				40						45			

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Val	Asp	Gly	Xaa	Arg	Xaa	Xaa	Ser	Asp	Xaa	His	Ile	Xaa	Leu	Gln	Leu
	50					55					60				

60

Ser	Ala	Glu	Ser	Val	Gly	Glu	Val	Tyr	Ile	Lys	Gly	Val	Cys	Ala	Asn
65					70					75					80

60

Arg	Tyr	Leu	Ala	Met	Lys	Glu	Asp	Gly	Arg	Leu	Leu	Ala	Ser	Lys	Cys
				85					90					95	

Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110
 5
 Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
 115 120 125
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 Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
 130 135 140
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 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155
 20
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ggt agc ggt gct ttt ccg cca ggc cac ttc aaa gac ccg aaa cgt ctg 96
Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
50 20 25 30

tac tgc aaa aac ggt ggc ttc ttc ctg cgg atc nnn ccg gac ggt nnn 144
Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Xaa
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ggt gac ggc nnn cgt gan nnn tct gac nnn cac atc nnn ctt cag ctg 192
Val Asp Gly Xaa Arg Asp Arg Ser Asp Xaa His Ile Xaa Leu Gln Leu
60 50 55 60

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Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn

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65	70	75	80	
cgg tac ctg gcg atg	aaa gaa gac ggt	cgt ctg ctg gcg tct	aaa tgc	288
Arg Tyr Leu Ala Met	Lys Glu Asp Gly	Arg Leu Leu Ala Ser	Lys Cys	
	85	90	95	
gtt acc gac gaa tgc	ttc ttt ttc gaa	cgt ctg gag tct	aac aat tac	336
Val Thr Asp Glu Cys	Phe Phe Phe Glu	Arg Leu Glu Ser	Asn Asn Tyr	
	100	105	110	
aac acc tat cgt tct	cgt aaa tac tct	tca tgg tat gtt	gcg ctc aaa	384
Asn Thr Tyr Arg Ser	Arg Lys Tyr Ser	Ser Trp Tyr Val	Ala Leu Lys	
	115	120	125	
cgc acg ggt cag tac	aaa ctg ggt cca	aaa acc ggt ccg	ggt cag aaa	432
Arg Thr Gly Gln Tyr	Lys Leu Gly Pro	Lys Thr Gly Pro	Gly Gln Lys	
	130	135	140	
gcg atc ctg ttc ctg	ccg atg tct gcg	aaa tct taa		468
Ala Ile Leu Phe Leu	Pro Met Ser Ala	Lys Ser		
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	<222>	(61)..(61)		
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	<222>	(65)..(65)		
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 25 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Xaa
 35 40 45
 30 Val Asp Gly Xaa Arg Asp Arg Ser Asp Xaa His Ile Xaa Leu Gln Leu
 50 55 60
 35 Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
 65 70 75 80
 40 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95
 45 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110
 50 Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
 115 120 125
 55 Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys
 130 135 140
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 145 150 155

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 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Xaa
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40 nnn gca gaa nnn nnn gga nnn gtg tnn atc aaa gga gtg tgt gct aac 240
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 65 70 75 80

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 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

50 gtt acg gat gag tgt ttc ttt ttt gaa cga ttg gaa tct aat aac tac 336
 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

55 aat act tac cgg tca agg aaa tac acc agt tgg tat gtg gca ctg aaa 384
 Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
 115 120 125

60 cga act ggg cag tat aaa ctt gga tcc aaa aca gga cct ggg cag aaa 432
 Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
 130 135 140

gct ata ctt ttt ctt cca atg tct gct aag agc tga 468
 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
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25

30

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35 40 45

10 Val Asp Gly Xaa Arg Asp Arg Ser Asp Xaa His Ile Xaa Leu Gln Leu
50 55 60

15 Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
65 70 75 80

Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
85 90 95

20 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
100 105 110

25 Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
115 120 125

30 Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
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 <223> The 'Xaa' at location 73 stands for Tyr or Ser.

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35 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30

40 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr
 35 40 45

45 Val Asp Gly Xaa Arg Asp Arg Ser Asp Xaa His Ile Xaa Leu Gln Leu
 50 55 60

50 Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

55 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

60 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
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Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
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ggc agc gcc gcc ttc ccg ccc ggc cac ttc aag gac ccc aag cgg ctg 96
Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
40 20 25 30

tac tgc aaa aac ggg ggc ttc ttc ctg cgc atc nnn ccc gac ggc nnn 144
Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr
45 35 40 45

gtt gac ggg nnn cgg gan nnn agc gac nnn cac atc nnn cta caa ctt 192
Val Asp Gly Xaa Arg Asp Arg Ser Asp Xaa His Ile Xaa Leu Gln Leu
50 50 55 60

nnn gca gaa nnn nnn gga nnn gtg tnn atc aaa gga gtg tgt gct aac 240
Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
65 70 75 80

cgt tac ctg gct atg aag gaa gat gga aga tta ctg gct tct aaa tgt 288
Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
55 85 90 95

gtt acg gat gag tgt ttc ttt ttt gaa cga ttg gaa tct aat aac tac 336
Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
60 100 105 110

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73/94

	aat act tac cgg tca agg aaa tac acc agt tgg tat gtg gca ctg aaa	384
	Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys	
	115 120 125	
5	cga act ggg cag tat aaa ctt gga tcc aaa aca gga cct ggg cag aaa	432
	Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys	
	130 135 140	
10	gct ata ctt ttt ctt cca atg tct gct aag agc tga	468
	Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser	
	145 150 155	
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 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30
 10 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr
 35 40 45
 15 Val Asp Gly Xaa Arg Asp Arg Ser Asp Xaa His Ile Xaa Leu Gln Leu
 50 55 60
 20 Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
 65 70 75 80
 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95
 25 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110
 30 Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
 115 120 125
 35 Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
 130 135 140
 40 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
 145 150 155

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 1 5 10 15

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 ggt agc ggt gct ttt ccg cca ggc cac ttc aaa gac ccg aaa cgt ctg 96
 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30

15
 tac tgc aaa aac ggt ggc ttc ttc ctg cgg atc nnn ccg gac ggt nnn 144
 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr
 35 40 45

20
 gtt gac ggc nnn cgt gan nnn tct gac nnn cac atc nnn ctt cag ctg 192
 Val Asp Gly Thr Arg Asp Arg Ser Asp Xaa His Ile Xaa Leu Gln Leu
 50 55 60

25
 nnn gcg gaa nnn nnn ggt nnn gta tnn atc aaa ggt gtt tgc gcg aac 240
 Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

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 cgg tac ctg gcg atg aaa gaa gac ggt cgt ctg ctg gcg tct aaa tgc 288
 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

35
 gtt acc gac gaa tgc ttc ttt ttc gaa cgt ctg gag tct aac aat tac 336
 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

40
 aac acc tat cgt tct cgt aaa tac tct tca tgg tat gtt gcg ctc aaa 384
 Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
 115 120 125

45
 cgc acg ggt cag tac aaa ctg ggt cca aaa acc ggt ccg ggt cag aaa 432
 Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys
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 20 25 30

40 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr
 35 40 45

45 Val Asp Gly Thr Arg Asp Arg Ser Asp Xaa His Ile Xaa Leu Gln Leu
 50 55 60

50 Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

55 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

60 Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
 115 120 125

Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys
130 135 140

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 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30

45 tac tgc aaa aac ggg ggc ttc ttc ctg cgc atc nnn ccc gac ggc nnn 144
 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr
 35 40 45

50 gtt gac ggg nnn cgg gan nnn agc gac nnn cac atc nnn cta caa ctt 192
 Val Asp Gly Thr Arg Asp Arg Ser Asp Xaa His Ile Xaa Leu Gln Leu
 50 55 60

nnn gca gaa nnn nnn gga nnn gtg tnn atc aaa gga gtg tgt gct aac 240
 Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

55 cgt tac ctg gct atg aag gaa gat gga aga tta ctg gct tct aaa tgt 288
 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

60 gtt acg gat gag tgt ttc ttt ttt gaa cga ttg gaa tct aat aac tac 336
 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr

	100	105	110	
5	aat act tac cgg tca agg aaa tac acc agt tgg tat gtg gca ctg aaa Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys 115 120 125			384
10	cga act ggg cag tat aaa ctt gga tcc aaa aca gga cct ggg cag aaa Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys 130 135 140			432
15	gct ata ctt ttt ctt cca atg tct gct aag agc tga Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser 145 150 155			468
20	<210> 52 <211> 155 <212> PRT <213> Artificial			
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 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
 20 25 30

 15 tac tgc aaa aac ggt ggc ttc ttc ctg cgg atc nnn ccg gac ggt nnn 144
 Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile Leu Pro Asp Gly Thr
 35 40 45

 20 gtt gac ggc nnn cgt gan nnn tct gac nnn cac atc nnn ctt cag ctg 192
 Val Asp Gly Thr Arg Asp Arg Ser Asp Gln His Ile Gln Leu Gln Leu
 50 55 60

 25 nnn gcg gaa nnn nnn ggt nnn gta tnn atc aaa ggt gtt tgc gcg aac 240
 Xaa Ala Glu Xaa Xaa Gly Xaa Val Xaa Ile Lys Gly Val Cys Ala Asn
 65 70 75 80

 30 cgg tac ctg gcg atg aaa gaa gac ggt cgt ctg ctg gcg tct aaa tgc 288
 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
 85 90 95

 35 gtt acc gac gaa tgc ttc ttt ttc gaa cgt ctg gag tct aac aat tac 336
 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
 100 105 110

 40 aac acc tat cgt tct cgt aaa tac tct tca tgg tat gtt gcg ctc aaa 384
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 115 120 125

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40 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
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Asn Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys
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 Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
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gtt acg gat gag tgt ttc ttt ttt gaa cga ttg gaa tct aat aac tac 336
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60 cgc acg ggt cag tac aaa ctg ggt cca aaa acc ggt ccg ggt cag aaa 432
 Arg Thr Gly Gln Tyr Lys Leu Gly Pro Lys Thr Gly Pro Gly Gln Lys
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caa gca gaa gag aga gga gtt gtg tct atc aaa gga gtg tgt gct aac      240
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Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Glu Asn His Tyr
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aat act tac nnn tca nnn aaa nnn nnn nnn tgg nnn gtg nnn ctg aaa      384
Asn Thr Tyr Xaa Ser Xaa Lys Xaa Xaa Xaa Trp Phe Val Gly Leu Lys
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cga act ggg cag tat aaa ctt gga tcc aaa aca gga cct ggg cag aaa      432
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93/94

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- 60 Gln Ala Glu Glu Arg Gly Val Val Ser Ile Lys Gly Val Cys Ala Asn
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- 60 Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
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5 Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Glu Asn His Tyr
100 105 110

Asn Thr Tyr Xaa Ser Xaa Lys Xaa Xaa Xaa Trp Phe Val Gly Leu Lys
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15 Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
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INTERNATIONAL SEARCH REPORT

International Application No
PCT/EP2004/001108A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 A61L31/04 A61L31/10 A61L33/12 C07K14/50

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 C07K A61L

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 99/00071 A (UNIV MICHIGAN) 7 January 1999 (1999-01-07) claims 1-5,15-20 -----	1-24
A	DE 197 13 213 A (RUEBBEN ALEXANDER DR MED) 1 October 1998 (1998-10-01) the whole document -----	1-24

 Further documents are listed in the continuation of box C. Patent family members are listed in annex.

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- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- *Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- *Z* document member of the same patent family

Date of the actual completion of the international search

14 June 2004

Date of mailing of the international search report

30/06/2004

Name and mailing address of the ISA

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Fax: (+31-70) 340-3016

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Heck, G

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/EP2004/001108

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9900071	A	07-01-1999	AU 8267698 A 19-01-1999
			CA 2295040 A1 07-01-1999
			EP 1023005 A1 02-08-2000
			JP 2002507136 T 05-03-2002
			WO 9900071 A1 07-01-1999
DE 19713213	A	01-10-1998	DE 19713213 A1 01-10-1998