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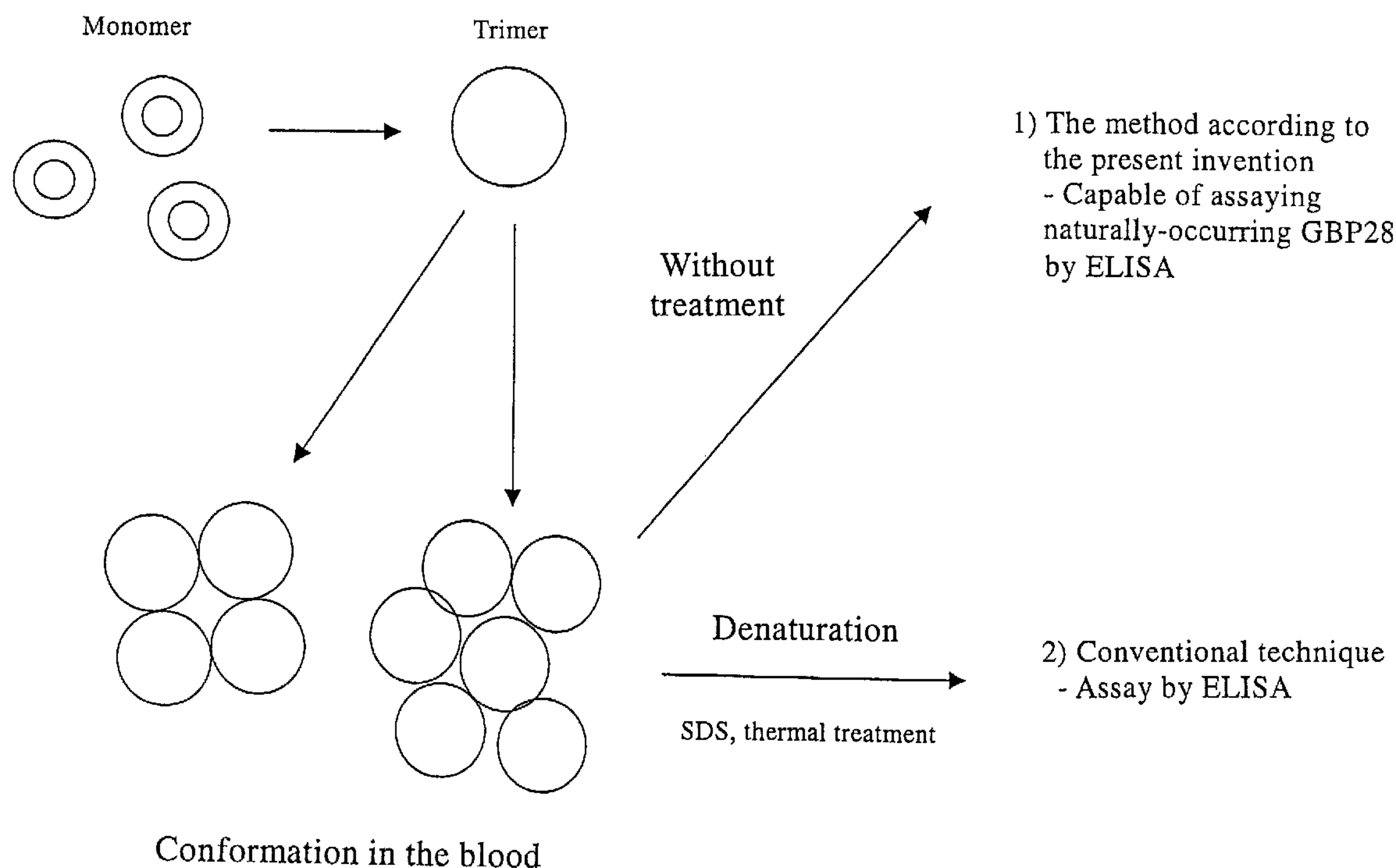
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(54) Titre : METHODE DIAGNOSTIQUE ET DE CONTROLE DES DESORDRES METABOLIQUES GLUCIDIQUES
(54) Title: METHOD FOR DIAGNOSING OR MONITORING CARBOHYDRATE METABOLISM DISORDERS



(57) Abrégé/Abstract:

This invention provides a method for diagnosing geriatric diseases associated with insulin resistance, such as type II diabetes, at an early stage and a method for monitoring the therapeutic effects of agents for type II diabetes by quantitatively assaying GBP28 with the use of a monoclonal antibody that can assay adiponectin (GBP28) in a sample.

ABSTRACT

This invention provides a method for diagnosing geriatric diseases associated with insulin resistance, such as type II diabetes, at an early stage and a method for monitoring the therapeutic effects of agents for type II diabetes by quantitatively assaying GBP28 with the use of a monoclonal antibody that can assay adiponectin (GBP28) in a sample.

DESCRIPTION

METHOD FOR DIAGNOSING OR MONITORING
CARBOHYDRATE METABOLISM DISORDERS

Technical Field

The present invention relates to a method for diagnosing or monitoring carbohydrate metabolism disorders by assaying adiponectin (GBP28) in a sample, a method for monitoring the therapeutic effects of agents for type II diabetes by assaying GBP28 in a sample, a method for assaying GBP28 characterized by assaying naturally-occurring GBP28 in a sample, and a monoclonal antibody that specifically reacts with naturally-occurring GBP28.

Background Art

The number of patients afflicted with diabetes is as many as 100,000,000 or more throughout the world, and 90% or more thereof are afflicted with type II diabetes (Schoonjans, K. and Auwerx, J., *Lancet* 355: 1008-1010, 2000). Type II diabetes is characterized by impaired insulin secretion and/or insulin resistance (DeFronzo, R. A. et al., *Diabetes Care* 15: 318-368, 1992). Thiazolidinediones had been found both in animal experiments and in clinical studies to improve insulin resistance. When such agents were administered to patients with type II diabetes, insulin action was elevated. This adversely resulted in lowered levels of blood glucose, glycohemoglobin, and serum insulin (Schwartz, S. et al., *N. Engl. J. Med.* 338: 861-866, 1998).

Adiponectin, which is a plasma protein derived from the adipocyte (Maeda, K. et al., *Biochem. Biophys. Res. Commun.* 221: 286-289, 1996; Arita, Y. et al., *Biochem. Biophys. Res. Commun.* 257: 79-83, 1999) (also known as a gelatin-binding protein of 28 kDa (GBP28) (Nakano, Y. et al., *J. Biochem.* 120: 803-812, 1996; Saito, K. et al., *Gene* 229: 67-73, 1999; Saito, K. et al., *Biol. Pharm. Bull.* 22: 1158-1162, 1999)), adheres to a damaged vascular wall *in vitro*, blocks endodermal NF- κ B signal

transmission (Ouchi, N. et al., *Circulation* 102: 1296-1301, 2000), and inhibits cell proliferation of smooth muscles induced by HB-EGF and PDGF (Matsuzawa, Y. et al., *Ann. N. Y. Acad. Sci.* 892: 146-154, 1999). In the case of obesity patients, the levels of this protein in serum had been reported to be low (Arita, Y. et al., *Biochem. Biophys. Res. Commun.* 257: 79-83, 1999). In the case of patients with type II diabetes and those of coronary artery diseases, the aforementioned serum levels had been reported to be lowered (Hotta, K. et al., *Arterioscler Thromb. Vasc. Biol.* 20: 1595-1599, 2000).

In the past, blood glucose, HbA_{1C}, serum insulin, HOMA-IR, and the like had been known as factors associated with type II diabetes. These factors were, however insufficient in terms of operability and/or sensitivity as indicators for diagnosing type II diabetes or for monitoring therapeutic effects. As mentioned above, association of GBP28 with type II diabetes had been also suggested. However, it was not sufficiently clarified whether or not GBP28 was actually applicable to the diagnosis of type II diabetes and the monitoring of therapeutic effects in the clinical field.

The ELISA technique of Ohmoto et al. is known as a method for assaying GBP28 (Ohmoto et al., *BIO Clinica* 15(10) 758-761, 2000; Japanese Laid-open Patent Publication (Kokai) No. 2000-304748). Naturally occurring GBP28 in blood is constituted by 3 monomers, and 4 to 6 trimers are aggregated (*J. Biochem.* 120, 803-812, 1996). However, the process of the aforementioned conventional technique was complicated due to the use of a monoclonal antibody to GBP28 having a monomeric structure. This required naturally occurring GBP28 in a sample to be denatured, a specimen to be mixed with an SDS solution at the time of assay, and thermal treatment to be conducted at 100°C. This conventional technique was to assay the naturally occurring GBP28 in a denatured state. Therefore, it could not directly assay the GBP28 in its natural state.

Summary of the Invention

In order to overcome the above problems, the present inventors have assayed the blood concentrations of a variety of parameters associated with type II diabetes in the

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patients with this ailment. They have also conducted concentrated studies in order to search for factors that could be clinical indicators for the diagnosis of carbohydrate metabolism disorders, such as type II diabetes, and the monitoring of therapeutic effects of agents for type II diabetes. As a result, they have found that the assay of GBP28 was very effective for the diagnosis of carbohydrate metabolism disorders and for the monitoring of the therapeutic effects of the agents therefore as described in the Examples below. They have also found that GBP28 in a sample can be simply and rapidly assayed in its natural state by assaying the GBP28 in an aggregated form. This has led to the completion of the present invention.

A first aspect of the present invention provides a method for diagnosing type II diabetes or monitoring therapeutic effects of therapeutic agents for type II diabetes, which comprises:

measuring a level of naturally-occurring adiponectin (GBP28) having a trimer structure of GBP28 and/or a structure of aggregated GBP28 trimers in its natural state in a sample,

wherein:

when the method is for diagnosing type II diabetes and the measured level is lower than a normal or boundary value, then that is indicative of type II diabetes, and

when the method is for monitoring the therapeutic effects of the therapeutic agents and the measured level after an initiation of treatment is elevated compared with that before the treatment, then that is indicative of the therapeutic effects of the therapeutic agents.

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A second aspect of the present invention provides a method for providing an indicator of insulin resistance, which comprises:

measuring a level of naturally-occurring
5 adiponectin (GBP28) having a trimer structure of GBP28 and/or a structure of aggregated GBP28 trimers in its natural state in a sample,

wherein when the measured level is lower than a normal or boundary level, then that is indicative of the
10 insulin resistance.

A third aspect of the present invention provides a monoclonal antibody or fragment thereof, which does not react with a monomeric structure of adiponectin (GBP28) but specifically reacts with naturally-occurring GBP28 having a
15 trimer structure of GBP28 and/or a structure of aggregated GBP28 trimers.

A fourth aspect of the present invention provides a use of the above-mentioned monoclonal antibody or segment thereof for the method described above.

20 A fifth aspect of the present invention provides a kit for measuring a level of naturally-occurring adiponectin (GBP28), which comprises:

(A) the monoclonal antibody or fragment thereof as defined above, and

25 (B) a buffer.

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A sixth aspect of the present invention provides a reagent comprising a monoclonal antibody or fragment thereof which does not react with a monomeric structure of adiponectin (GBP28) but specifically reacts with naturally-
5 occurring GBP28 having a trimer structure of GBP28 and/or a structure of aggregated GBP28 trimers.

Brief Description of the Drawings

Fig. 1 schematically shows principles of the method according to the present invention and those of
10 conventional techniques.

Fig. 2 shows changes in serum adiponectin levels caused by three-month administration of pioglitazone (30 mg/day) to 10 patients afflicted with type II diabetes (Japanese males), wherein "O-O" represents diet therapy and
15 administration of pioglitazone (n = 7), and "■-■" represents diet therapy and administration of sulfonylurea and pioglitazone (n = 3).

Fig. 3 shows the calibration curve for GBP28.

Fig. 4 shows the dilution linearity for GBP assay.

Best Modes for Carrying out the Invention

GBP28 in humans is known to have the amino acid sequence as shown in SEQ ID NO: 1 and the nucleotide sequence as shown in SEQ ID NO: 2. GBP28 in the blood is known to form 4 to 6 aggregated trimers, each thereof being constituted by 3 monomers (J. Biochem. 120, 803-812, 1996).

The present inventors have found that the concentration of GBP28 in a sample such as blood obtained from a healthy subject would exhibit a correlation with levels of insulin associated with an indicator of insulin resistance, insulin resistance index (HOMA-IR), triglyceride, HDL-cholesterol, and LDL-cholesterol that are characteristics of type II diabetes. They have also found that the concentration of GBP28 in a sample such as blood obtained from a patient with type II diabetes exhibited more significant changes compared with levels of plasma glucose, HbA_{1C}, serum insulin, and HOMA-IR that have been employed as indicators for type II diabetes when monitoring the therapeutic effects of agents for type II diabetes used therefor. Based on these findings, the concentration of GBP28 in a sample such as blood was found to be very effective as a novel indicator for the diagnosis of carbohydrate metabolism disorders, such as type II diabetes, and the monitoring of the therapeutic effects of agents for type II diabetes.

Accordingly, the present inventors provide a method for diagnosing carbohydrate metabolism disorders such as type II diabetes and a method for monitoring the therapeutic effects of agents for type II diabetes by assaying GBP28, preferably naturally-occurring GBP28, in a sample.

In the present invention, examples of the "sample" include, but are not limited to, serum, plasma, synovial fluid, pleural effusion, tissue extract, tissue, culture supernatant, and urine.

Examples of carbohydrate metabolism disorders include diabetes (type I or II diabetes) and diabetic complications (elevated blood pressure, hyperlipidemia, arteriosclerosis, or glaucoma). The present invention is particularly suitable for the diagnosis of type II diabetes or for the monitoring of the therapeutic effects of the

agents.

A variety of conventional agents can be used as therapeutic agents for carbohydrate metabolism disorders. Examples of therapeutic agents for type II diabetes include: sulfonylurea agents such as glibenclamide, gliclazide, tolbutamide, and glimepiride; immediate-release agents for accelerating insulin secretion such as nateglinide; α -glucosidase inhibitors such as voglibose and acarbose; agents for improving insulin resistance such as pioglitazone; and biguanides such as metformin. These agents can be suitably used in the method according to the present invention. Examples of particularly preferable therapeutic agents for type II diabetes include thiazolidine derivatives such as pioglitazone and troglitazone.

Methods for assaying GBP28 are not particularly limited. Assay can be conducted by any conventional method in the art such as the solid phase method, competitive assay, agglutination assay, turbidimetric assay, or sandwich enzyme immunoassay using a monoclonal antibody that specifically reacts with GBP28, preferably naturally-occurring GBP28 having a trimer structure of GBP28 and/or a structure of aggregated GBP28 trimers. Particularly preferably, assay is conducted by solid-phase enzyme-linked immunosorbent assay (ELISA). Enzyme immunoassay is a common technique in the art, and a person skilled in the art can perform it using an antibody that specifically reacts with the aforementioned GBP28.

When using a monoclonal antibody that specifically reacts with naturally occurring GBP28 in the aforementioned assay, denatured or monomeric GBP28, which does not exist in a natural state, can be excluded from the assay. More specifically, a monoclonal antibody recognizing a site that first appears in a trimer when the GBP28 forms a collagen-like conformation is particularly preferably prepared and used.

Fig. 1 schematically shows principles of the method according to the present invention for assaying GBP28 and those of conventional techniques. In the conventional technique, GBP28 that was expressed by gene recombinant *E. coli* is used as an immunogen in order to obtain a monoclonal antibody to GBP28. Since the recombinant GBP28 is present as an inclusion body in the *E. coli*, however, the protein

must be processed with, for example, guanidine hydrochloride, urea, or a reducing agent when it is subjected to purification. Thus, denatured and monomeric GBP28 is the only type that can be used as an immunogen. Accordingly, the resulting monoclonal antibody reacts only with denatured and monomeric GBP28. When assaying naturally occurring GBP28 in the sample, therefore, it must be previously denatured into monomeric GBP28. In contrast, the method according to the present invention employs a monoclonal antibody that specifically reacts with naturally occurring GBP28. Thus, assay can be conducted without processing naturally occurring GBP28 in the blood.

The monoclonal antibody according to the present invention can be obtained by a technique common in the art using, for example, naturally-occurring GBP28 as an antigen. Naturally occurring GBP28 can be purified by, for example, passing a large amount of human plasma through a gelatin-cellulofine column (J. Biochem 120, 803-812, 1996), utilizing its gelatin-binding properties. For example, an animal is immunized with this naturally occurring GBP28, and a spleen or lymph node is collected from the immunized animal. Antibody-producing cells contained in the spleen or lymph node are allowed to fuse with myeloma cells with the aid of polyethylene glycol or the like to prepare hybridomas. A hybridoma of interest is screened for, this hybridoma is cultured, and a monoclonal antibody can be prepared from the culture supernatant. Culture may be carried out *in vitro* or *in vivo*. For example, a hybridoma is administered to a mouse or the like intraperitoneally, and an antibody of interest can be then obtained from its ascites. A monoclonal antibody can be purified by, for example, fractionation with ammonium sulfate precipitation or anion chromatography or affinity purification using protein A or immobilized antigen. The thus prepared antibody is used for assaying GBP28, and it can be also applied to the antibody treatment or the like of diseases caused by abnormal expression of GBP28.

An anti-GBP28 monoclonal antibody that can be particularly preferably used in the present invention is characterized as follows:

the myeloma cell line used: P3U1;

the antigen: a solution of GBP28 (about 50 μ g) isolated and purified from human

serum is micellized with a complete adjuvant, the resultant is administered to a mouse intraperitoneally, two weeks later, the mouse is subjected to booster, the spleen is extirpated 3 days thereafter, and the B-cell is collected;

the number of clones where the monoclonal antibody is prepared: 3 (1H, 5, 6, and 7);

the Ig type: entirely IgG κ -type; and

the epitope: it recognizes only GBP28 trimers that were not thermally denatured, and accordingly, this site is considered to first appear in the trimer when a collagen-like conformation is formed.

Hybridomas IH-6 and IH-7 producing an anti-GBP28 monoclonal antibody that can be suitably used in the present invention have been deposited at the International Patent Organism Depositary of the National Institute of Advanced Industrial Science and Technology (Tsukuba Central 6, 1-1-1 Higashi, Tsukuba, Ibaraki, Japan) as of July 12, 2001, under the accession numbers FERM BP-7660 and FERM BP-7661, respectively.

Specifically, a method for diagnosing carbohydrate metabolism disorders by assaying GBP28 in the sample and a method for monitoring the therapeutic effects of the therapeutic agents can be conducted, for example, in the manner as described below.

1) Assay of the concentration of GBP28

Serums or the like collected from patients or the like are diluted, the diluted serums or the like are added to a 96-well microtiter plate coated with a mouse anti-GBP28 monoclonal antibody, and GBP28 in the serum is allowed to bind to the mouse anti-GBP28 monoclonal antibody in the well. After the well is washed, an enzyme-labeled mouse anti-GBP28 monoclonal antibody is added and then allowed to bind to the mouse anti-GBP28 monoclonal antibody existing in the well. After the well is further washed, a substrate solution of the enzyme is added to develop color. After the termination of color development, the absorbance is assayed. In advance, the absorbance of the GBP28 standard solution at a known concentration level is assayed in the same manner, the calibration curve for the absorbance relative to the concentration of GBP28 is prepared, and the assayed absorbance is converted into the concentration of

GBP28 based on the calibration curve. Thus, the concentration of GBP28 in the serums or the like collected from patients or the like can be assayed.

2) A method for diagnosing or monitoring carbohydrate metabolism disorders

When employing GBP28 as the indicator for diagnosing carbohydrate metabolism disorders such as type II diabetes, for example, the concentration of GBP28 that was assayed by the above method is compared with the normal value or boundary value, which had been previously calculated.

3) A method for monitoring the therapeutic effect of an agent

When employing GBP28 as an indicator for monitoring the therapeutic effects of a therapeutic agent for type II diabetes, for example, blood is sampled from a patient with type II diabetes before and after the initiation of treatment, GBP28 is assayed by the above method, and the rate of changes in the GBP28 content of the blood from before the initiation of treatment to after the initiation of treatment is calculated, thereby evaluating the therapeutic effects. If the blood concentration of GBP28 after the initiation of the treatment is elevated compared with that before the treatment, the therapeutic effects of the agents can be recognized. The interval for blood sampling in this case is preferably between 1 and 3 months.

The present invention also provides a kit for assaying naturally occurring GBP28 in the sample comprising a monoclonal antibody that specifically reacts with naturally-occurring GBP28. The kit according to the present invention may suitably comprise, in addition to the above-mentioned antibody according to the present invention, for example, a buffer, a secondary antibody labeled with a label such as peroxidase, a standard solution, a block solution, an enzyme substrate solution such as peroxidase, and a washing liquid. The monoclonal antibody according to the present invention recognizes the trimer structure of GBP28. Accordingly, sandwich assay can be conducted by immobilizing a non-labeled monoclonal antibody on the plate, adding a GBP28-containing sample, and then using a labeled monoclonal antibody of the same type.

Examples

The present invention is hereafter described in greater detail with reference to the following examples, although the present invention is not limited to these examples.

In the following examples, all the statistical analyses were conducted using the StatView® program for Macintosh (version 4.5-J, Abacus Concepts Inc., Berkeley, California). For the comparison of the baseline and the trace data for 3 months, a Wilcoxon Signed-Rank Test (two-tailed) was conducted. For the comparison of the rate of change in GBP28 and other parameters, significance was evaluated using Pearson's correlation coefficient and the partial correlation coefficient. Serum insulin, TG, and GBP28 levels and HOMA-IR would be normally distributed upon the log conversion in a large-scale study. Thus, logarithmic values were employed for the analyses regarding these parameters. All the data are represented by "mean $\pm$ S.D." When p was smaller than 0.05, it was determined to be statistically significant.

[Example 1] Assay of each parameter

10 male patients with type II diabetes (ages: 40 to 66, 57.7 ± 7.4) either being only under diet therapy (n = 7) or being under diet therapy and on medication of sulfonylurea (n = 3) were subjected to inspection regarding the following parameters before and after a three-month administration of a thiazolidine derivative, pioglitazone. Body weights, blood glucose levels, and blood pressure levels of those patients and treatments for them were at constant levels for at least 3 months before the examination. The sulfonylurea doses for 3 patients were at constant levels during that period. Pioglitazone was administered to all 10 patients after breakfast every day (orally, 30 mg/day). The ages, the body mass indices (BMI), and the clinical profiles of patients at the time of initiation are shown in Table 1.

Table 1
Clinical Profiles of Patients with Type II Diabetes

Parameter		Range
N	10	
Age (years)	57.4 ± 7.4	[40 – 66]
Height (cm)	164.2 ± 5.2	[156.0 – 171.6]
Weight (kg)	71.3 ± 9.8	[57.3 – 85.1]
Body mass index (kg/m^2)	26.4 ± 3.2	[22.4 – 32.5]
Duration of disease (years)	7.0 ± 4.5	[2.0 – 15.0]
Glucose (mmol/l)	8.6 ± 1.4	[6.9 – 11.3]
HbA _{1c} (%)	6.7 ± 0.8	[5.3 – 8.2]

Data are represented by n or mean $\pm$ S.D..

In the table, data are represented by n or mean ($\pm$ S.D.).

The systolic blood pressure (SBP) and the diastolic blood pressure (DBP) were measured twice in a sitting position after an at least 5-minute recess. Body heights, weights, and levels of fasting plasma glucose (FPG), HbA_{1c}, serum insulin, adiponectin, total cholesterol (TC), triglyceride (TG), HDL-cholesterol, LDL-cholesterol, and uric acid were measured in the morning after an overnight fast. Levels of plasma glucose, lipids, and uric acid were measured by a common automated measuring technique. The serum concentration of insulin was measured using a commercially available kit, EIA (Tosoh, Tokyo), as already described (Kawai, T. et al., *Metabolism* 48: 1102-1107, 1999; Hirose, H. et al., *Clin. Sci. (Colch)* 94: 633-636, 1998; Hirose, H. et al., *J. Hypertens.* 16: 2007-2012, 1998; Hirose, H. et al., *Clin. Sci. (Colch)* 100: 145-150, 2001). The insulin resistance index was evaluated by the Homeostasis model assessment (HOMA-IR) (Matthews, D. R. et al., *Diabetologia* 28: 412-419, 1985; Rudenski, A. S. et al., *Metabolism* 40: 908-917, 1991).

The results are shown in Table 2.

Table 2

Effects of three-month administration of pioglitazone on each parameter				
Parameter	Baseline	After the three-month trace	Rate of change in 3 months	
N	10	10		
Body Mass Index (kg/m ²)	26.4 ± 3.2	27.0 ± 3.5	*	2.1 ± 2.2
Body mass index at systolic blood pressure (kg/m ²)	149 ± 18	138 ± 15	**	-7.0 ± 4.9
Body mass index at diastolic blood pressure (kg/m ²)	89 ± 13	83 ± 9	*	-6.3 ± 6.9
Fasting blood glucose (mmol/l)	8.6 ± 1.4	7.0 ± 1.0	**	-18.2 ± 9.3
HbA _{1c} (%)	6.7 ± 0.8	6.1 ± 0.6	*	-9.0 ± 8.8
Serum insulin (pmmol/l) (log)	54 ± 11	30 ± 8	**	-23.7 ± 13.5
HOMA-IR (log)	3.4 ± 1.9	1.6 ± 1.3	*	-62.6 ± 22.6
Total cholesterol (mg/dl)	208 ± 26	218 ± 35		4.8 ± 10.9
Triglyceride (mg/dl) (log)	134 ± 2	127 ± 2		-1.0 ± 6.3
HDL cholesterol (mg/dl)	51.7 ± 9.4	52.1 ± 7.3		2.0 ± 11.2
LDL cholesterol (mg/dl)	124 ± 24	138 ± 24	*	12.3 ± 12.8
Uric acid (mg/dl)	5.9 ± 1.1	5.6 ± 1.3		-5.0 ± 9.5
GBP28 (μg/l) (log)	4.8 ± 1.7	14.4 ± 2.1	**	80.9 ± 58
Data are represented by n or mean ± S.D..		* P < 0.05 (Wilcoxon test)		
		** P < 0.01 (Wilcoxon test)		

In the table, data are represented by n or mean (± S.D.).

As shown in Table 2, a three-month administration of pioglitazone resulted in significantly lowered levels of systolic blood pressure, diastolic blood pressure, fasting plasma glucose, HbA_{1c}, serum insulin, and HOMA-IR. The body mass index (BMI) and the LDL-cholesterol level were elevated, although there was no significant change in levels of triglyceride, HDL-cholesterol, or uric acid.

Effects of controlling blood glucose and blood pressure levels were observed similarly with the case of troglitazone (Kawai, T. et al., *Metabolism*, 48: 1102-1107, 1999), unlike the case of the control group that was subjected to diet therapy. Concerning the effects of thiazolidinedione on blood pressure levels, troglitazone has been reported to lower blood pressure levels in experiments using insulin-resistant animals (Yoshioka, S. et al., *J. Hypertens.* 18: 1857-1864, 2000) and in clinical study (Ogihara, T. et al., *Am. J. Hypertens.* 8: 316-320, 1995). Concerning the effects on LDL-cholesterol, administration of troglitazone (Schwartz, S. et al, *N. Eng. J. Med.* 338: 861-866, 1998) or rosiglitazone (Balfour, J. A. and Plosker, G. L., *Drugs* 57: 921-930, 1999) has been reported to be associated with the elevation of LDL-cholesterol. The

LDL-cholesterol level was slightly increased by the administration of pioglitazone. It was more significant 3 months later.

[Example 2] Assessment of body fat distribution

The patients who had been also employed in Example 1 were subjected to the assessment of body fat distribution before and after the three-month administration of pioglitazone. Distributions of subcutaneous fat and visceral fat were determined by assessing the region of a Hounsfield unit of -150 to -50 by a method modified from the computerized tomography (CT) scan at the level of the navel by Tokunaga et al. (Tokunaga, K. et al., Int. J. Obes. 7: 437-445, 1983). CT images at the baseline and after a three-month treatment were taken in accordance with the same protocols that had been previously reported by the present inventors (Kawai, T. et al., Metabolism, 48: 1102-1107, 1999). The V/S ratio was calculated by dividing the visceral fat area (VFA) by the subcutaneous fat area (SFA). The results are shown in Table 3.

Table 3

Effect of three-month administration of pioglitazone on body fat distribution			
Parameter	Baseline	After the three-month trace	Rate of changes in 3 months
N	10	10	
Visceral Fat Area (cm ²)	165 ± 38	180 ± 46	11.3 ± 21.3
Subcutaneous Fat Area (cm ²)	155 ± 69	179 ± 81	* 16.1 ± 18.7
V/S ratio	1.2 ± 0.3	1.1 ± 0.3	-4.0 ± 11.2
Data are represented by n or mean ± S.D.. * P < 0.05 (Wilcoxon test)			

In the table, data are represented by n or mean (± S.D.).

As shown in Table 3, BMI and the subcutaneous fat area (SFA) significantly increased after the three-month administration of pioglitazone. This result was substantially the same as that obtained by the administration of troglitazone (Kawai, T. et al., Metabolism, 48: 1102-1107, 1999; Kelly, I.E. et al., Diabetes Care 22: 288-293, 1999; Mori, Y. et al., Diabetes Care 22: 908-912, 1999). On the contrary, the visceral fat area (VFA) was not reduced, but it rather slightly increased. This was different from

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the findings obtained based on the administration of troglitazone. Although the reason for such difference has not been elucidated yet, it could be due to structural differences such as the vitamin E structure of troglitazone and/or functions of PPAR α . VFA tended to be larger while the V/S ratio tended to be smaller. However, there was no statistical significance in these differences.

[Example 3] Assay of GBP28

The patients who had been employed in Example 1 were also subjected to an assay of the blood concentration of GBP28 before and after the three-month
10 administration of pioglitazone.

The anti-human GBP28 monoclonal antibody was prepared in the following manner. First, a solution of GBP28 (about 50 μ g) isolated and purified from human serum was micellized with a complete adjuvant, and the resultant was administered to a mouse intraperitoneally. Two weeks later, the mouse was subjected to booster, the spleen was extirpated 3 days thereafter, and the B-cell was collected. The B-cell and the myeloma cell were allowed to fuse with each other to prepare hybridomas, and a hybridoma of interest was administered to a mouse intraperitoneally. Thereafter (20 days later), ascites was processed with DEAE-Sepharose^{*} Fast Flow, and the monoclonal antibody of interest was obtained.

20 GBP28 was assayed by ELISA in the following manner. A standard GBP28 sample, a 441-fold-diluted unknown sample, and a control sample (100 μ l each) were added to a 96-well microtiter plate coated with the mouse anti-GBP28 monoclonal antibody. The plate was incubated for 60 minutes, the well was washed, and incubation was carried out for additional 30 minutes with the horseradish peroxidase-labeled mouse anti-GBP28 monoclonal antibody. The plate was washed again, and incubation was carried out for 30 minutes with a tetramethylbenzidine reagent. Subsequently, 100 μ l of aqueous solution of 0.36N sulfuric acid was added to each well to terminate the reaction, and the absorbance at 450 nm was assayed. Intra- and interassay coefficients were 4.8% to 4.9% and 3.3% to 6.8%, respectively.

*Trade-mark

As a result of the three-month administration of pioglitazone, the elevation in the serum adiponectin level was observed in all 10 patients (1.5 to 6.8 times higher than the baselines and 3 times higher on average) (Table 2 and Fig. 2). Further, the rate of changes in GBP28 and in other parameters shown in Tables 2 and 3 generated over the course of three months were examined. The rate of change in the measured value of each parameter (some were logarithmic values) was compared with each other. As a result, it was found that the concentration of GBP28 in the sample exhibited a larger rate of change (80.9%) than levels of blood glucose (18.2%), HbA_{1c} (9.0%), serum insulin (23.7%), HOMA-IR (62.6%), and the like that have been conventionally known as factors associated with type II diabetes.

[Example 4] Assay results of healthy subjects

In Example 3, GBP28 was found to be applicable to the monitoring of the therapeutic effects of thiazolidine derivatives, i.e., therapeutic agents for type II diabetes, in patients with type II diabetes. Also, the applicability of the concentration of GBP28 as an indicator of insulin resistance could be predicted. Thus, healthy individuals were subjected to inspection for the association between the blood concentration of GBP28 and parameters that have been reported to be associated with insulin resistance.

980 individuals between the ages of 30 and 65 who annually visit the hospital to get medical checkups were subjected to inspections for the parameters that have been reported to be associated with insulin resistance. Specifically, the subjects were 714 males and 266 females, and patients having endocrine diseases or serious kidney or liver diseases were excluded therefrom. Also, patients who had been on insulin, antidiabetic agents, or antihyperlipidemic agents as medications were excluded.

Parameters, i.e., the systolic blood pressure (SBP) and the diastolic blood pressure (DBP), were measured twice in a sitting position after an at least 5-minute recess. Ages, BMI, cardiac rates, levels of fasting plasma glucose (FPG), serum insulin, adiponectin, total cholesterol (TC), triglyceride (TG), HDL-cholesterol, LDL-cholesterol, and uric acid were measured in the morning after an overnight fast. Levels of plasma

glucose and uric acid were measured by a common automated measuring technique. The serum concentration of insulin was measured using a commercially available kit, EIA (Tosoh, Tokyo), as already described. The insulin resistance index was evaluated by the homeostasis model assessment (HOMA-IR).

The results of comparison between each parameter measured in these inspections and GBP28 concerning male subjects are shown in Table 4, and those concerning female subjects are shown in Table 5.

Table 4

A			B		C		D	
vs. log[GBP28]					Correction depending on age and body mass index		Correction depending on age and body mass index	
	Mean	± SD	r	P	r	P	r	P
Age	(46.4)	± 10.1	0.044	NS	-	-	-	-
Body Mass Index	(23.2)	± 2.7	-0.354	<0.0001	-	-	-	-
Body fat	(21.8)	± 4.8	-0.369	<0.0001	-	-	-0.257	0.0069
Systolic blood pressure	(124)	± 18	-0.164	<0.0001	-0.177	0.065	-	-
Diastolic blood pressure	(77)	± 12	-0.140	<0.0002	-0.052	NS	-0.100	NS
Cardiac rate	(75)	± 13	-0.043	NS	-0.047	NS	-0.064	NS
Glucose	(95)	± 16	-0.140	0.0002	-0.033	NS	-0.012	NS
Insulin (log)	(5.58)	± 3.60	-0.338	<0.0001	-0.067	0.068	-0.057	NS
HOMA-IR (log)	(1.33)	± 0.98	-0.348	<0.0001	-0.229	<0.0001	-0.362	<0.0001
Total cholesterol	(200)	± 31	-0.065	0.08	-0.237	<0.0001	-0.362	<0.0001
Triglyceride (log)	(121)	± 84	-0.344	<0.0001	-0.013	NS	0.018	NS
HDL-cholesterol	(54)	± 13	0.414	<0.0001	-0.197	<0.0001	-0.207	0.0005
LDL-cholesterol	(126)	± 28	-0.154	<0.0001	-0.330	<0.0001	0.293	<0.0001
Uric acid	(6.2)	± 1.2	-0.258	<0.0001	-0.083	0.024	-0.030	NS
Data are represented by mean ± S.D..	NS: P > 0.1		n = 330 (relative to body fat)		-0.174	<0.0001	-0.214	0.0003

Table 5

vs. log[GBP28]	A			B		C		D	
	Mean	±	SD	r	P	r	P	r	P
	(	)	(	)	(	)	(	)	(
Age	41.7	±	9.9	0.089	NS	-	-	-	-
Body Mass Index	20.7	±	2.7	-0.226	0.0002	-	-	0.214	NS
Body fat	21.8	±	4.8	-0.360	<0.0001	-0.566	0.0073	-	-
Systolic blood pressure	112	±	16	-0.037	NS	0.050	NS	0.053	NS
Diastolic blood pressure	68	±	11	-0.043	NS	0.016	NS	0.040	NS
Cardiac rate	74	±	10	0.029	NS	0.072	NS	0.067	NS
Glucose	90	±	9	-0.128	0.037	-0.036	NS	-0.072	NS
Insulin (log)	4.82	±	2.50	-0.220	0.0003	-0.137	0.040	-0.127	NS
HOMA-IR (log)	1.08	±	0.61	-0.225	0.0002	-0.134	0.047	-0.126	NS
Total cholesterol	193	±	33	0.010	NS	0.006	NS	0.055	NS
Triglyceride (log)	68	±	37	-0.306	<0.0001	-0.275	<0.0001	-0.134	NS
HDL-cholesterol	69	±	15	0.393	<0.0001	0.345	<0.0001	0.280	0.0007
LDL-cholesterol	112	±	28	-0.175	0.0042	-0.199	0.0041	-0.131	NS
Uric acid	4.3	±	0.94	-0.018	NS	-0.025	NS	0.014	NS
Data are represented by mean ± S.D..				NS: P > 0.1		n = 330 (relative to body fat)			

In these tables, data are represented by "mean $\pm$ S.D.," correlation coefficients, and p values. The blood GBP28 exhibited a wide range of concentration distributions from 0.4 $\mu\text{g/ml}$ to 61.2 $\mu\text{g/ml}$. The blood concentrations of GBP28 of male subjects were $7.2 \pm 4.6 \mu\text{g/ml}$, and those of female subjects were $13.3 \pm 7.3 \mu\text{g/ml}$. That is, female subjects exhibited higher concentration of GBP28 than male subjects.

As shown in Tables 4 and 5, the blood concentration of GBP28 exhibited negative correlations with BMI, the systolic blood pressure, the diastolic blood pressure, fasting plasma glucose, insulin, HOMA-IR, triglyceride, LDL-cholesterol, and uric acid. The blood concentration of GBP28 exhibited positive correlations with HDL-cholesterol. When corrections were made depending on age and BMI and comparisons between GBP28 and each parameter were conducted, statistically significant correlations were attained with a reliability of $p < 0.05$, i.e., regarding insulin (male: $r = -0.229$, female: $r = -0.137$), HOMA-IR (male: $r = -0.237$, female: $r = -0.134$), triglyceride (male: $r = -0.197$, female: $r = -0.275$), HDL-cholesterol (male: $r = -0.330$, female: $r = 0.345$), and LDL-cholesterol (male: $r = -0.083$, female: $r = -0.199$). Therefore, the blood concentration of GBP28 exhibited correlations with HOMA-IR, insulin, triglyceride, HDL-cholesterol, and LDL-cholesterol.

As a result, it was found that the concentration of GBP could also serve as an indicator of insulin resistance for healthy individuals.

[Example 5] Production of a kit for assaying naturally occurring GBP28 in a sample

A. Materials for the kit according to the present invention

(1) A plate coated with an anti-human GBP28 monoclonal antibody

- A solution of anti-human GBP28 monoclonal antibody

The anti-human GBP28 monoclonal antibody obtained by processing mouse ascites with DEAE-Sepharose Fast Flow was adjusted to 10 $\mu\text{g/ml}$ with the aid of 0.1M phosphate buffer (pH 7.0).

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- Composition of washing liquid for preparation of a plate

Table 6

Reagent	Concentration
NaH ₂ PO ₄ ·2H ₂ O	40 mM
Na ₂ HPO ₄ ·12H ₂ O	60 mM
Tween* 20	0.05 %
pH 7.0	-

- Composition of block solution

Table 7

Reagent	Concentration
NaH ₂ PO ₄ ·2H ₂ O	40 mM
Na ₂ HPO ₄ ·12H ₂ O	60 mM
BSA	1.0 %
pH 7.0	-

1. Fractionate an antibody solution at 120 µl/well on the MaxiSorp* F8 plate (Nunc) and then allow the plate to stand at 4°C overnight.

2. Remove the antibody solution and then wash the plate twice with a washing liquid.

3. Fractionate a block solution at 200 µl/well and then allow the plate to stand at 37°C overnight.

4. Remove the block solution.

5. Allow the plate to stand under reduced pressure overnight, dry it thoroughly, seal it hermetically, and then store it at 4°C.

(2) Standard solution

- Solution of GBP28 antigen

GBP28 was purified from the serum pooled from healthy individuals using an affinity column to which the anti-human GBP28 monoclonal antibody had been bound. Thereafter, gel filtration was further carried out. The concentration of GBP28 was determined based on the absorbance at 280 nm (the concentration at Abs. 280 nm = 1,000 was determined to be 1.0 mg/ml).

*Trade-mark

- Dilution buffer

Table 8

Reagent	Concentration
NaH ₂ PO ₄ ·2H ₂ O	4 mM
Na ₂ HPO ₄ ·12H ₂ O	16 mM
BSA	1.0%
pH 7.4	-

The solution of GBP28 antigen was diluted with the aid of the dilution buffer having the above composition. Thus, the concentration of GBP28 was set at 0, 2, 5, 10, 25, and 50 ng/ml.

(3) Solution of peroxidase-labeled anti-human GBP28 monoclonal antibody

- Stock solution of peroxidase-labeled anti-human GBP28 monoclonal antibody

The anti-human GBP28 monoclonal antibody was subjected to pepsin digestion and reduction treatment to prepare the Fab' fragment, and maleimide-activated peroxidase was allowed to bind thereto. The concentration was calculated by determining each concentration of the anti-human GBP28 monoclonal antibody at the Fab' site and at the peroxidase site based on absorbance at 280 nm and 403 nm and adding them up.

- Solution for diluting a labeled antibody

Table 9

Reagent	Concentration
NaH ₂ PO ₄ ·2H ₂ O	4 mM
Na ₂ HPO ₄ ·12H ₂ O	16 mM
BSA	1.0%
pH 6.5	-

The solution of peroxidase-labeled anti-human GBP28 monoclonal antibody was diluted with the aid of a solution for diluting a labeled antibody having the above composition, and the concentration of the labeled antibody was set at 350 ng/ml. Thus, a solution of peroxidase-labeled anti-human GBP28 monoclonal antibody was prepared.

(4) Concentrated washing liquid (diluted 20-fold with purified water at the time of use)

Table 10

Reagent	Concentration
NaH ₂ PO ₄ ·2H ₂ O	40 mM
Na ₂ HPO ₄ ·12H ₂ O	60 mM
Tween 20	1.0%
pH 6.5	-

(5) Substrate solution

A commercially available tetramethylbenzidine (TMB) solution for ELISA was used.

(6) Reaction terminator

0.36N H₂SO₄ was used.

B. Assay protocol

Specimens to be tested are diluted to suitable concentrations (up to 50 ng/ml) with the aid of the dilution buffer which was used for preparation of the standard solution. The dilution ratio of general specimens may be suitably about 400- to 500-fold (e.g., 21-fold dilution is repeated twice to attain 441-fold dilution).

1. Place a standard solution and a specimen to be tested in the wells of a plate (100 µl/well).

2. Stir for about 5 to 10 seconds via a plate stirrer and then allow the plate to stand at room temperature under light-shielded conditions for 60 minutes.

3. Remove the standard solution and the specimen and wash the plate four times with a washing liquid.

4. Place a solution of peroxidase-labeled antibody in the wells of a plate (100 µl/well).

5. Stir for about 5 to 10 seconds via a plate stirrer and then allow the plate to

stand at room temperature under light-shielded conditions for 30 minutes.

6. Remove the solution of peroxidase-labeled antibody and wash the plate four times with a washing liquid.

7. Place a substrate solution in the wells of a plate (100 μ l/well).

8. Stir for about 5 to 10 seconds via a plate stirrer and then allow the plate to stand at room temperature under light-shielded conditions for 30 minutes.

9. Place a reaction terminator in the wells of a plate (100 μ l/well).

10. Stir for about 5 to 10 seconds via a plate stirrer and assay the absorbance of each well (assay wavelength: 450/650 nm (primary/secondary), approximate expression for calibration curve: quadratic).

[Example 6] Preparation of calibration curve

Prepared standard solutions of 0, 2, 5, 10, 25, and 50 ng/ml of GBP28 were assayed as samples, and the calibration curve was inspected. As shown in Fig. 3, the obtained calibration curve was substantially linear. Since the average concentration of GBP28 in the blood was 10 μ g/ml, the specimen was subjected to 21-fold dilution twice to attain 441-fold dilution. Assay was conducted at this concentration (assay range: 0 to 22 μ g/ml).

[Example 7] Test for repeatability

In order to evaluate changes in the assay values obtained by the kit, 3 types of serum samples were used, and each thereof was subjected to assay 24 times. As a result, the CV values were satisfactorily 5% or lower as shown in Table 11. Thus, the assay values obtained by the kit of the present invention could be determined to be reliable.

Table 11

Repeatability of GBP28 assay

GBP28 ($\mu\text{g/ml}$)			
	A	B	C
1	4.95	8.25	15.26
2	5.93	8.59	13.61
3	5.82	6.64	14.82
4	5.63	8.11	12.99
5	5.86	7.93	14.59
6	5.96	7.67	13.34
7	5.55	7.50	14.56
8	5.46	7.78	13.53
9	5.55	7.47	14.70
10	5.62	7.92	13.48
11	5.17	7.97	13.59
12	5.80	7.48	15.00
13	5.94	8.15	14.29
14	5.39	7.99	15.04
15	5.84	7.78	14.22
16	5.46	7.82	14.22
17	5.53	8.09	14.59
18	5.85	7.68	14.01
19	6.01	7.75	14.32
20	5.88	8.18	14.88
21	5.63	7.83	13.13
22	5.80	8.28	15.15
23	6.21	8.41	15.33
24	5.82	8.08	14.80
N	24	24	24
MAX.	6.21	8.59	15.33
MIN.	4.95	6.64	12.99
MEAN	5.694	7.889	14.309
S.D.	0.2762	0.3834	0.6883
C.V.	4.8%	4.9%	4.8%

[Example 8] Test for day-to-day repeatability

In order to evaluate day-to-day variations generated in the assay values obtained by the kit, 5 types of serum samples were used, and each thereof was subjected to the assay five times on five separate days. As a result, variations in the assay values for 5 days were satisfactorily of a c.v. of 3.3% to 6.8% as shown in Table 12. Thus, the assay values obtained by the kit of the present invention could be determined to be reliable.

Table 12

Day-to-day repeatability of GBP28 assay

	GBP28 ($\mu\text{g/ml}$)				
	A	B	C	D	E
1	7.67	5.88	5.38	15.98	11.91
2	8.24	5.61	5.37	16.19	12.12
3	8.08	5.85	6.00	14.58	11.49
4	8.44	6.00	5.72	15.88	11.86
5	7.26	5.32	4.89	14.93	11.04
N	5	5	5	5	5
MAX.	8.44	6.00	6.00	16.19	12.12
MIN.	7.26	5.32	4.89	14.58	11.04
MEAN	7.937	5.730	5.471	15.514	11.684
S.D.	0.4220	0.2432	0.3736	0.6366	0.3835
C.V.	5.3%	4.2%	6.8%	4.1%	3.3%

[Example 9] Dilution test

In order to avoid the influence of the serum component on the reaction system, the serum sample is usually diluted and then used for an assay when the diagnostic agent kit is used. In order to confirm that the dilution ratio is optimal, a sample at a given concentration is suitably diluted and then assayed, and whether or not the obtained values result in a straight line on the graph is determined. A straight line indicates that the properties of the sample are equivalent to those of the standard solution. The assay values are determined not to be affected by the serum component.

Three types of serum samples were diluted 441-fold in accordance with the assay protocol, these samples were further diluted to 1/5 to 5/5, and each thereof was subjected to the assay twice. As a result, all the specimens exhibited good linearity as shown in Table 13 and Fig. 4. This indicated that the dilution ratio of the kit was appropriate.

Table 13

Dilution linearity of GBP28 assay

Serum 1

Dilution (/5)	GBP28 concentration ($\mu\text{g/ml}$)	GBP28 concentration ($\mu\text{g/ml}$)	GBP28 concentration ($\mu\text{g/ml}$)
	Serum 1	Serum 2	Serum 3
1	1.54	1.00	2.88
1	1.59	1.05	3.00
2	2.73	2.28	6.41
2	2.99	2.03	6.05
3	4.36	3.24	9.63
3	4.08	3.21	8.75
4	6.43	4.60	12.34
4	5.88	4.25	11.28
5	7.45	5.20	15.04
5	7.68	5.32	13.88

[Example 10] Addition-recovery test

Unlike isolated proteins, proteins that are present in blood may form a complex with various other proteins or substances existing in blood, or undergo changes in their conformations. Even though good evaluation results were obtained from a standard sample prepared from a isolated protein at a suitable concentration, therefore, an accurate concentration could not always be obtained when serum or the like was employed as a sample (for example, the formation of a complex results in the loss of an epitope to the antibody).

Thus, samples were prepared by adding a GBP28 standard solution adjusted to a suitable concentration to sample serum, and the resulting samples were subjected to the assay. Subsequently, the concentration level of the sample before the addition of the standard sample was subtracted from the obtained value, the resultant was divided by the amount added, and the recovery rate was thus determined. As a result, a good recovery rate of approximately 95% on average was obtained as shown in Table 14. Thus, it was demonstrated that this kit was able to be used for conducting the assay without the influence of the blood component on the added standard sample. This also verified that

the blood concentration of GBP28 could be accurately assayed.

Table 14

Results of addition-recovery test

Serum 1

	None	A	B	C
Mean ($\mu\text{g/ml}$)	10.34	11.85	13.98	19.90
Amount added ($\mu\text{g/ml}$)	-	1.52	3.65	9.57
Recovery rate (%)	-	93.4	95.6	91.9

Serum 2

	None	A	B	C
Mean ($\mu\text{g/ml}$)	13.73	15.25	17.75	23.16
Amount added ($\mu\text{g/ml}$)	-	1.52	4.02	9.43
Recovery rate (%)	-	93.8	105.3	90.6

Serum 3

	None	A	B	C
Mean ($\mu\text{g/ml}$)	26.36	28.13	30.56	36.88
Amount added ($\mu\text{g/ml}$)	-	1.77	4.20	10.52
Recovery rate (%)	-	109.1	110.1	101.1

Industrial Applicability

The present invention enables the simple diagnosis of carbohydrate metabolism disorders, such as type II diabetes, in the clinical field by assaying the concentration of GBP28 in a sample. Also, the present invention enables the monitoring of the therapeutic effects of an agent for type II diabetes, particularly, a thiazolidine derivative. Furthermore, the use of the antibody according to the present invention enables the assay of the blood concentration of GBP28, preferably naturally-occurring GBP28, and GBP28 can be quantitatively assayed without pretreatment, such as thermal treatment, of the GBP28 in the sample. This can lead to simplified assay processes and improved abilities to process specimens.

Further, the method according to the present invention can easily process a large amount of specimens. Thus, a very effective assay system is provided, which can be employed in clinical examinations for geriatric diseases associated with insulin resistance, such as diabetes, elevated blood pressure, hyperlipidemia, or arteriosclerosis.

SEQUENCE LISTING

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<120> A METHOD FOR DIAGNOSING OR MONITORING CARBOHYDRATE METABOLISM DISORDERS

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CLAIMS:

1. A method for diagnosing type II diabetes or monitoring therapeutic effects of therapeutic agents for type II diabetes, which comprises:

5 measuring a level of naturally-occurring adiponectin (GBP28) having a trimer structure of GBP28 and/or a structure of aggregated GBP28 trimers in its natural state in a sample,

wherein:

10 when the method is for diagnosing type II diabetes and the measured level is lower than a normal or boundary value, then that is indicative of type II diabetes, and

when the method is for monitoring the therapeutic effects of the therapeutic agents and the measured level
15 after an initiation of treatment is elevated compared with that before the treatment, then that is indicative of the therapeutic effects of the therapeutic agents.

2. A method for providing an indicator of insulin resistance, which comprises:

20 measuring a level of naturally-occurring adiponectin (GBP28) having a trimer structure of GBP28 and/or a structure of aggregated GBP28 trimers in its natural state in a sample,

wherein when the measured level is lower than a
25 normal or boundary level, then that is indicative of the insulin resistance.

3. The method according to claim 1 or 2, wherein the measurement is carried out using a monoclonal antibody or

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fragment thereof which does not react with a monomeric structure of GBP28 but specifically reacts with the naturally-occurring GBP28.

4. The method according to any one of claims 1-3,
5 wherein the structure of aggregated GBP28 trimers consists of from 4 to 6 GBP28 trimers.

5. A monoclonal antibody or fragment thereof, which does not react with a monomeric structure of adiponectin (GBP28) but specifically reacts with naturally-occurring
10 GBP28 having a trimer structure of GBP28 and/or a structure of aggregated GBP28 trimers.

6. The monoclonal antibody or fragment thereof according to claim 5, wherein the structure of aggregated GBP28 trimers consists of from 4 to 6 GBP28 trimers.

15 7. A monoclonal antibody or fragment thereof, which does not react with a monomeric structure of adiponectin (GBP28) but specifically reacts with naturally-occurring GBP28 having a trimer structure of GBP28 and/or a structure of aggregated GBP28 trimers, wherein the monoclonal antibody
20 is produced from hybridoma FERM BP-7660 or FERM BP-7661.

8. A use of the monoclonal antibody or fragment thereof as defined in any one of claims 5-7, for measuring a level of naturally-occurring adiponectin (GBP28) having a trimer structure of GBP28 and/or a structure of aggregated
25 GBP28 trimers.

9. The use according to claim 8, wherein the measurement is carried out for diagnosing type II diabetes, for monitoring therapeutic effects of therapeutic agents for type II diabetes, or for providing an indicator of insulin
30 resistance.

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10. The use according to claim 8 or 9, wherein the structure of aggregated GBP28 trimers consists of from 4 to 6 GBP28 trimers.

11. A method for diagnosing type II diabetes, which
5 comprises:

providing a blood sample of a human patient, the blood sample containing a concentration of naturally-occurring adiponectin (GBP28) having a trimer structure of GBP28 or a structure of aggregated 4 to 6 GBP28 trimers;

10 providing a monoclonal antibody which specifically reacts with the naturally-occurring GBP28 without reacting with monomeric or denatured GBP28;

without processing the blood sample to denature or monomerize the naturally-occurring GBP28, measuring the
15 concentration of the naturally-occurring GBP28 in the blood sample using the monoclonal antibody; and

comparing the measured concentration with a normal value or boundary value previously calculated.

12. The method according to claim 11, wherein the
20 monoclonal antibody is produced by a hybridoma prepared by:

immunizing a non-human animal with naturally-occurring human GBP28 purified from human blood;

collecting spleen or lymph node from the immunized animal; and

25 fusing an antibody-producing cell of the spleen or lymph node with a myeloma cell.

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13. The method according to claim 12, wherein the monoclonal antibody is IgGk-type monoclonal antibody which recognizes only naturally-occurring GBP28 trimer.

14. The method according to claim 11, wherein the
5 monoclonal antibody is produced by hybridoma IH-6 (FERM BP-7660) or hybridoma IH-7 (FERM BP-7661).

15. The method according to any one of claims 11 to 14, wherein the measurement of the concentration of the naturally-occurring GBP28 is conducted by solid-phase
10 enzyme-linked immunosorbent assay (ELISA).

16. A kit for measuring a level of naturally-occurring adiponectin (GBP28), which comprises:

(A) the monoclonal antibody or fragment thereof as defined in any one of claims 5-7, and

15 (B) a buffer.

17. The kit according to claim 16, which further comprises:

a secondary monoclonal antibody or fragment thereof, labeled with a label, the secondary antibody
20 specifically reacting with the GBP28 trimer.

18. The kit according to claim 17, wherein the label is an enzyme and which further comprises a substrate of the enzyme.

19. The kit according to any one of claims 16-18,
25 which still further comprises:

a standard solution containing a known concentration of GBP28;

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a block solution containing bovine serum albumin
(BSA); and

a washing liquid.

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Fig. 1

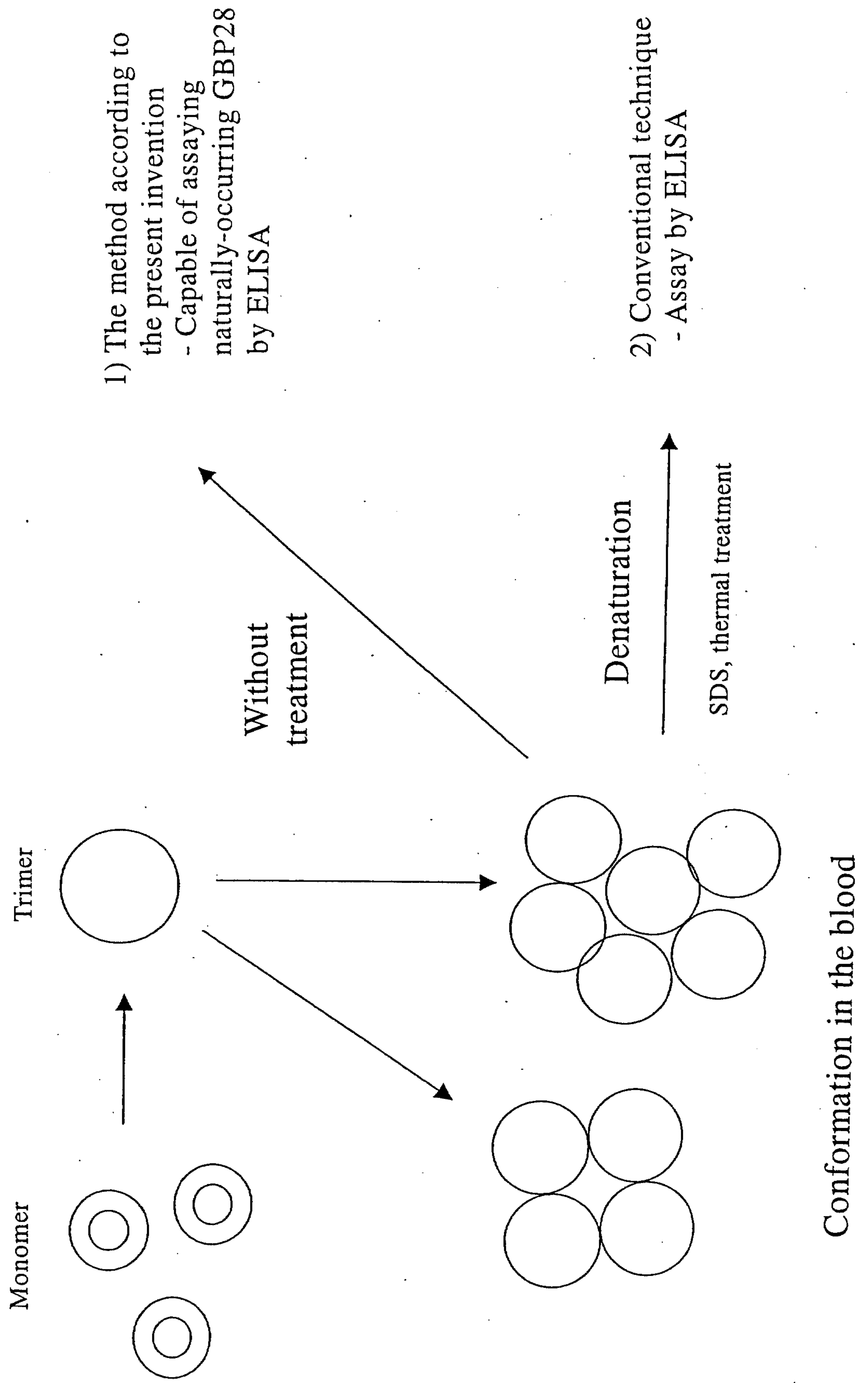


Fig. 2

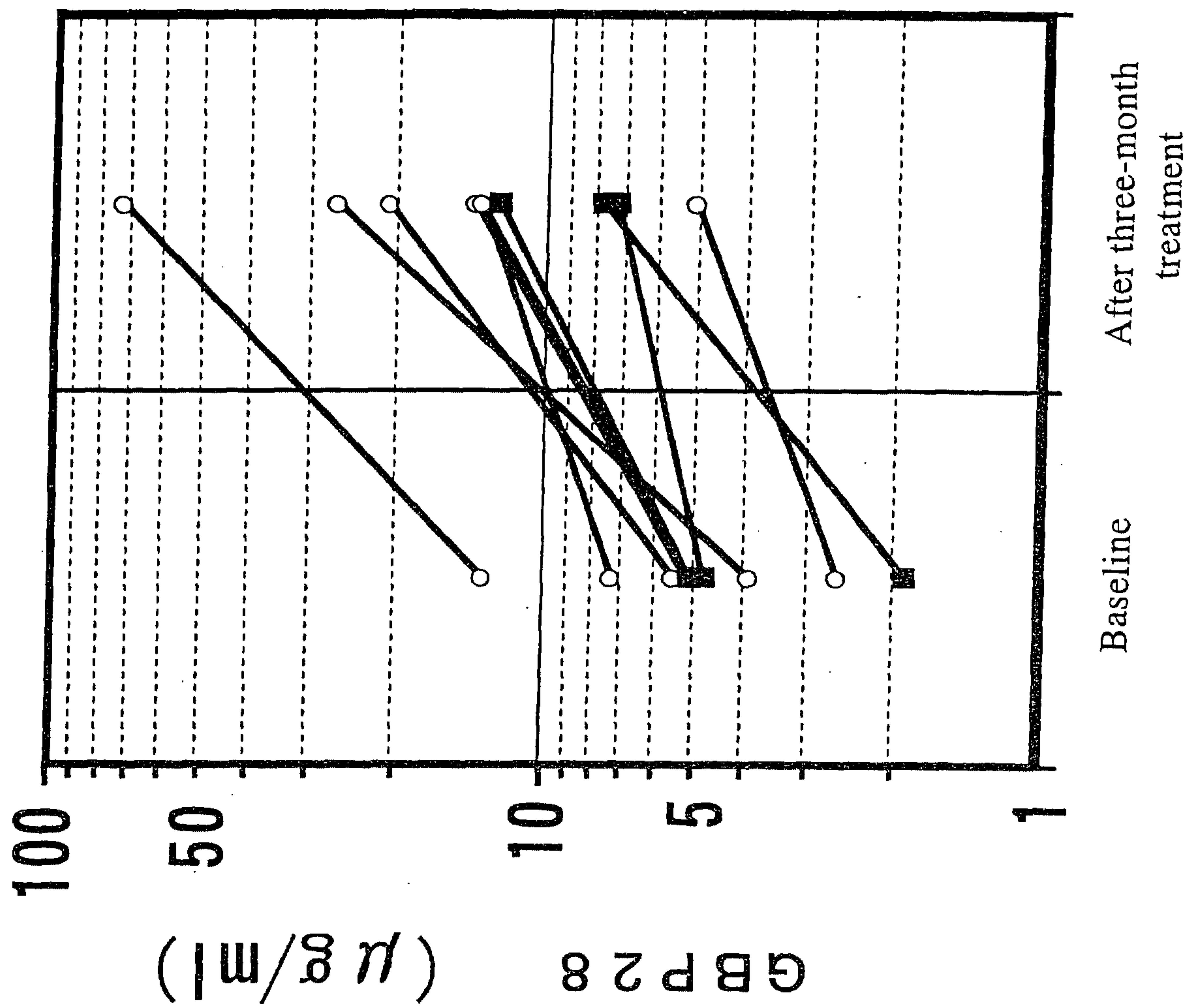


Fig. 3

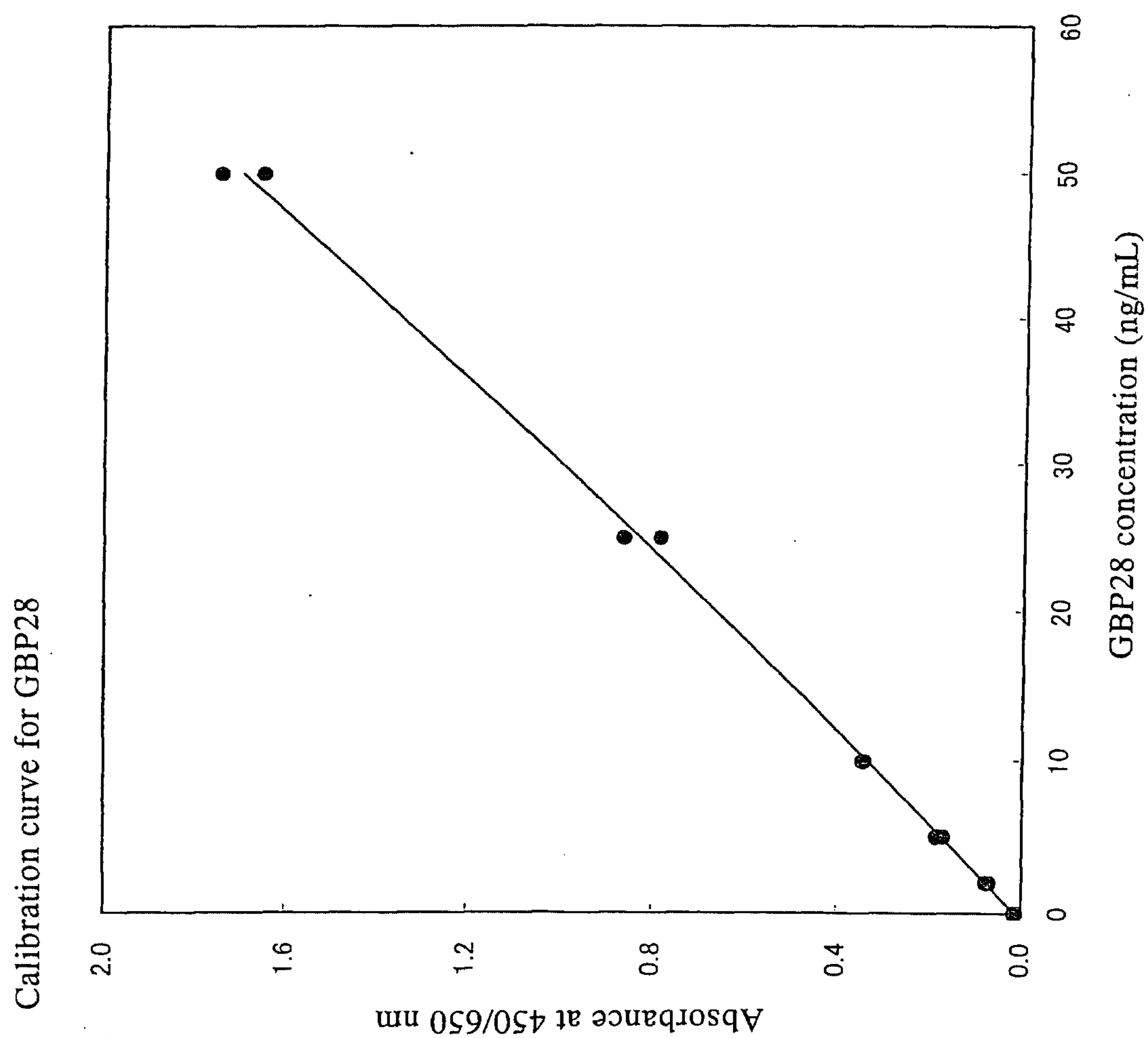


Fig. 4

