(12) STANDARD PATENT

(11) Application No. AU 2002342669 C1

(19) AUSTRALIAN PATENT OFFICE

(54) Title

Peptides and related molecules that bind to TALL-1

(51) International Patent Classification(s)

C07K 7/08 (2006.01) **C12N 15/09** (2006.01) **A61K 38/00** (2006.01) **C07K 14/00** (2006.01) **A61P 17/02** (2006.01) **C07K 14/47** (2006.01) **A61P 35/00** (2006.01) **C07K 16/44** (2006.01) **A61P 35/02** (2006.01) **C07K 19/00** (2006.01) **A61P 37/02** (2006.01) **C12N 1/21** (2006.01) **A61P 37/06** (2006.01) **C12N 15/10** (2006.01) **C12N 15/28** (2006.01) **C07K 1/04** (2006.01) **C07K 5/113** (2006.01) **C40B 40/02** (2006.01) **C07K 7/06** (2006.01)

(21) Application No: **2002342669** (22) Date of Filing: **2002.05.13**

(87) WIPO No: WO02/092620

(30) Priority Data

(31) Number (32) Date (33) Country **60/290,196 2001.05.11 US**

(43) Publication Date: 2002.11.25
 (43) Publication Journal Date: 2003.05.01
 (44) Accepted Journal Date: 2005.09.22
 (44) Amended Journal Date: 2010.10.07

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(56) Related Art

Proc. Natl. Acad. Sci. USA, 28 March 2000, Vol.97, No.7, pp.3370-3375 Proc. Natl. Acad. Sci. USA, 01 August 2000, Vol.97 No.16, pp.9156-9161

(19) World Intellectual Property Organization International Bureau





(43) International Publication Date 21 November 2002 (21.11.2002)

PCT

(10) International Publication Number WO 02/092620 A3

(51) International Patent Classification⁷: C07K 14/52, 14/525, A61K 38/19, C12N 5/10, 15/28

(21) International Application Number: PCT/US02/15273

(22) International Filing Date: 13 May 2002 (13.05.2002)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data: 60/290,196 11 May 2001 (11.05.2001)

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- (81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW.
- (84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

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(54) Title: PEPTIDES AND RELATED MOLECULES THAT BIND TO TALL-1

a¹a²a³CDa⁴La⁵a³a¹¹Ca¹²a¹³a¹⁴

(SEQ. ID. NO: 100),
b¹b²b³Cb⁵b6Db³Lb¹¹b¹¹b¹¹b¹²b¹³b¹⁴Cb¹⁵b¹²b¹³b¹³

(SEQ. ID. NO: 104)
c¹c²c³Cc⁵Dc²Lc³c¹³c¹¹c¹²c¹³c¹⁴Cc¹⁶c¹²c¹²6

(SEQ. ID. NO: 105)
d¹d²d³Cd⁵d6d²WDd¹⁰Ld¹³d¹⁴d¹⁵Cd¹ſd¹³d¹²d¹(SEQ. ID. NO: 106)
e¹e²e³Ce⁵e6e²De³Le¹¹Ke¹³Ce¹⁵e¹6e¹²e¹³6

(SEQ. ID. NO: 107)
f¹f²f³Kf⁵Df″Lf°f¹⁰Qf¹²f¹³f¹⁴

 $(X^1)_a - V^1 - (X^2)_b$ (1)

(SEQ. ID NO: 109)

(57) Abstract: The present invention concerns therapeutic agents that modulate the activity of TALL-1. In accordance with the present invention, modulators of TALL-1 may comprise an amino acid sequence Dz^2Lz^4 wherein z^2 is an amino acid residue and z^4 is threonyl or isoleucyl. Exemplary molecules comprise a sequence of the formulae $a^{1}a^{2}a^{3}CDa^{6}La^{8}a^{9}a^{10}Ca^{12}a^{13}a^{14}$ (SEO.ID.NO:100), $b^1b^2b^3Cb^5b^6Db^8Lb^{10}b^{11}b^{12}b^{13}b^{14}Cb^{16}b^{17}b^{18}\\$ (SEQ.ID.NO:104) $c^{1}c^{2}c^{3}Cc^{5}Dc^{7}Lc^{9}c^{10}c^{11}c^{12}c^{13}c^{14}Cc^{16}c^{17}c^{18}$ (SEQ.ID.NO:105) $d^1d^2d^3Cd^5d^6d^7WDd^{10}Ld^{13}d^{14}d^{15}Cd^{16}d^{17}d^{18}\\$ (SEQ.ID.NO:106) $e^{1}e^{2}e^{3}Ce^{5}e^{6}e^{7}De^{9}Le^{11}Ke^{13}Ce^{15}e^{16}e^{17}e^{18} \\$ (SEQ.ID.NO:107) $f^1f^2f^3Kf^5Df^7Lf^9f^{10}Of^{12}f^{13}f^{14}$ (SEO.ID NO:109) wherein substituents are as defined in the specification. The invention further comprises compositions of matter of the formula $(X^1)_a$ - V^1 - $(X^2)_b$ wherein V1 is a vehicle that is covalently attached to one or more of the above TALL-1 modulating compositions of matter. The vehicle and the TALL-1 modulating composition of matter may be linked through the N- or C-terminus of the TALL-1 modulating portion. The preferred vehicle is an Fc domain, and the preferred Fc domain is an IgG Fc domain.

WO 02/092620 A3 ||||||||||||||



Published:

with international search report

(88) Date of publication of the international search report:

21 August 2003

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

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PEPTIDES AND RELATED MOLECULES THAT BIND TO TALL-1

This application is related to U.S. provisional application no. 60/290,196, filed May 11, 2001, which is hereby incorporated by reference.

Background of the Invention

Any discussion of the prior art throughout the specification should in no way be considered as an admission that such prior art is widely known or forms part of common general knowledge in the field.

After years of study in necrosis of tumors, tumor necrosis factors (TNFs) α and β were finally cloned in 1984. The ensuing years witnessed the emergence of a superfamily of TNF cytokines, including fas ligand (FasL), CD27 ligand (CD27L), CD30 ligand (CD30L), CD40 ligand (CD40L), TNF-related apoptosisinducing ligand (TRAIL, also designated AGP-1), osteoprotegerin binding protein (OPG-BP or OPG ligand), 4-1BB ligand, LIGHT, APRIL, and TALL-1. Smith et al. (1994), Cell 76: 959-962; Lacey et al. (1998), Cell 93: 165-176; Chichepotiche et al. (1997), J. Biol. Chem. 272: 32401-32410; Mauri et al. (1998), <u>Immunity</u> 8: 21-30; Hahne <u>et al</u>. (1998), <u>I. Exp</u>. <u>Med</u>. **188**: 1185-90; Shu <u>et al</u>. (1999), J. Leukocyte Biology 65: 680-3. This family is unified by its structure, particularly at the C-terminus. In addition, most members known to date are expressed in immune compartments, although some members are also expressed in other tissues or organs, as well. Smith et al. (1994), Cell 76: 959-62. All ligand members, with the exception of LT- α , are type II transmembrane proteins, characterized by a conserved 150 amino acid region within the C-terminal extracellular domain. Though restricted to only 20-25% identity, the conserved 150 amino acid domain folds into a characteristic β -pleated sheet sandwich and trimerizes. This conserved region can be proteolytically released, thus generating a soluble functional form. Banner et al. (1993), Cell 73: 431-445.

Many members within this ligand family are expressed in lymphoid enriched tissues and play important roles in the immune system development and modulation. Smith et al. (1994). For example, TNFα is mainly synthesized by macrophages and is an important mediator for inflammatory responses and immune defenses. Tracey & Cerami (1994), Ann. Rev. Med. 45: 491-503. Fas-L, predominantly expressed in activated T cell, modulates TCR-mediated apoptosis of thymocytes. Nagata, S. & Suda, T. (1995) Immunology Today 16: 39-43; Castrim et al. (1996), Immunity 5: 617-27. CD40L, also expressed by activated T cells, provides an essential signal for B cell survival, proliferation and immunoglobulin isotype switching. Noelle (1996), Immunity 4: 415-9.

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The cognate receptors for most of the TNF ligand family members have been identified. These receptors share characteristic multiple cysteine-rich repeats within their extracellular domains, and do not possess catalytic motifs within cytoplasmic regions. Smith et al. (1994). The receptors signal through direct interactions with death domain proteins (e.g. TRADD, FADD, and RIP) or with the TRAF proteins (e.g. TRAF2, TRAF3, TRAF5, and TRAF6), triggering divergent and overlapping signaling pathways, e.g. apoptosis, NF-kB activation, or JNK activation. Wallach et al. (1999), Annual Review of Immunology 17: 331-67. These signaling events lead to cell death, proliferation, activation or differentiation. The expression profile of each receptor member varies. For example, TNFR1 is expressed on a broad spectrum of tissues and cells, whereas the cell surface receptor of OPGL is mainly restricted to the osteoclasts. Hsu et al. (1999) Proc. Natl. Acad. Sci. USA 96: 3540-5.

A number of research groups have recently identified TNF family ligands with the same or substantially similar sequence. The ligand has been variously named neutrokine α (WO 98/18921, published May 7, 1998), 63954 (WO 98/27114, published June 25, 1998), TL5 (EP 869 180, published October 7, 1998), NTN-2 (WO 98/55620 and WO 98/55621,

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published December 10, 1998), TNRL1-alpha (WO 9911791, published March 11, 1999), kay ligand (W099/12964, published March 18, 1999); and TALL-1 (WO 00/68378, published Nov. 16, 2000). Each of these references is hereby incorporated by reference. Hereinafter, the ligands reported therein are collectively referred to as TALL-1.

TALL-1 is a member of the TNF ligand superfamily that is functionally involved in B cell survival and proliferation. Transgenic mice overexpressing TALL-1 had severe B cell hyperplasia and lupus-like autoimmune disease. Khare et al. (2000) PNAS 97 (7): 3370-3375). Both TACI and BCMA serve as cell surface receptors for TALL-1. Gross et al. (2000), Nature 404:995-999; Ware (2000), J. Exp. Med. 192(11):F35-F37; Ware (2000), Nature 404:949-950; Xia et al. (2000), J. Exp. Med. 192(1):137-143; Yu et al. (2000), Nature Immunology 1 (3):252-256; Marsters et al. (2000), <u>Current Biology</u> 10:785-788; Hatzoglou et al. (2000) J. of Immunology 165:1322-1330; Shu et al. (2000) PNAS 97(16):9156-9161; Thompson et al. (2000) J. Exp. Med. 192(1):129-135; Mukhopadhyay et al. (1999) J. Biol. Chem. 274(23):15978-81; Shu et al. (1999) J. Leukocyte Biol. 65:680-683; Gruss et al. (1995) Blood 85(12):3378-3404; Smith et al. (1994), Cell 76:959-962; U. S. Pat. No. 5,969,102, issued October 19,1999; WO 00/67034, published November 9, 2000; WO 00/40716, published July 13, 2000; WO 99/35170, published July 15, 1999. Both receptors are expressed on B cells and signal through interaction with TRAF proteins. In addition, both TACI and BCMA also bind to another TNF ligand family member, APRIL. Yu et al. (2000), Nature Immunology 1(3):252-256. APRIL has also been demonstrated to induce B cell proliferation.

To date, no recombinant or modified proteins employing peptide modulators of TALL-1 have been disclosed. Recombinant and modified

proteins are an emerging class of therapeutic agents. Useful modifications of protein therapeutic agents include combination with the "Fc" domain of an antibody and linkage to polymers such as polyethylene glycol (PEG) and dextran. Such modifications are discussed in detail in a patent application entitled, "Modified Peptides as Therapeutic Agents," publicshed WO 00/24782, which is hereby incorporated by reference in its entirety.

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A much different approach to development of therapeutic agents is peptide library screening. The interaction of a protein ligand with its receptor often takes place at a relatively large interface. However, as demonstrated for human growth hormone and its receptor, only a few key residues at the interface contribute to most of the binding energy. Clackson et al. (1995), Science 267: 383-6. The bulk of the protein ligand merely displays the binding epitopes in the right topology or serves functions unrelated to binding. Thus, molecules of only "peptide" length (2 to 40 amino acids) can bind to the receptor protein of a given large protein ligand. Such peptides may mimic the bioactivity of the large protein ligand ("peptide agonists") or, through competitive binding, inhibit the bioactivity of the large protein ligand ("peptide antagonists").

Phage display peptide libraries have emerged as a powerful method in identifying such peptide agonists and antagonists. See, for example, Scott et al. (1990), Science 249: 386; Devlin et al. (1990), Science 249: 404; U.S. Pat. No. 5,223,409, issued June 29, 1993; U.S. Pat. No. 5,733,731, issued March 31, 1998; U.S. Pat. No. 5,498,530, issued March 12, 1996; U.S. Pat. No. 5,432,018, issued July 11, 1995; U.S. Pat. No. 5,338,665, issued August 16, 1994; U.S. Pat. No. 5,922,545, issued July 13, 1999; WO 96/40987, published December 19, 1996; and WO 98/15833, published April 16, 1998 (each of which is incorporated by reference in its entirety). In such libraries, random peptide sequences are displayed by fusion with

coat proteins of filamentous phage. Typically, the displayed peptides are affinity-eluted against an immobilized target protein. The retained phages may be enriched by successive rounds of affinity purification and repropagation. The best binding peptides may be sequenced to identify key residues within one or more structurally related families of peptides. See, e.g., Cwirla et al. (1997), Science 276: 1696-9, in which two distinct families were identified. The peptide sequences may also suggest which residues may be safely replaced by alanine scanning or by mutagenesis at the DNA level. Mutagenesis libraries may be created and screened to further optimize the sequence of the best binders. Lowman (1997), Ann. Rev. Biophys. Biomol. Struct. 26: 401-24.

Structural analysis of protein-protein interaction may also be used to suggest peptides that mimic the binding activity of large protein ligands. In such an analysis, the crystal structure may suggest the identity and relative orientation of critical residues of the large protein ligand, from which a peptide may be designed. See, e.g., Takasaki et al. (1997), Nature Biotech. 15: 1266-70. These analytical methods may also be used to investigate the interaction between a receptor protein and peptides selected by phage display, which may suggest further modification of the peptides to increase binding affinity.

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Other methods compete with phage display in peptide research. A peptide library can be fused to the carboxyl terminus of the <u>lac</u> repressor and expressed in <u>E</u>. <u>coli</u>. Another <u>E</u>. <u>coli</u>-based method allows display on the cell's outer membrane by fusion with a peptidoglycan-associated lipoprotein (PAL). Hereinafter, these and related methods are collectively referred to as "<u>E</u>. <u>coli</u> display." In another method, translation of random RNA is halted prior to ribosome release, resulting in a library of polypeptides with their associated RNA still attached. Hereinafter, this and related methods are collectively referred to as "ribosome display."

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Other methods employ peptides linked to RNA; for example, PROfusion technology, Phylos, Inc. See, for example, Roberts & Szostak (1997), Proc. Natl. Acad. Sci. USA, 94: 12297-303. Hereinafter, this and related methods are collectively referred to as "RNA-peptide screening." Chemically derived peptide libraries have been developed in which peptides are immobilized on stable, nonbiological materials, such as polyethylene rods or solvent-permeable resins. Another chemically derived peptide library uses photolithography to scan peptides immobilized on glass slides. Hereinafter, these and related methods are collectively referred to as "chemical-peptide screening." Chemical-peptide screening may be advantageous in that it allows use of D-amino acids and other unnatural analogues, as well as non-peptide elements. Both biological and chemical methods are reviewed in Wells & Lowman (1992), Curr. Opin. Biotechnol. 3: 355-62. Conceptually, one may discover peptide mimetics of any protein using phage display, RNA-peptide screening, and the other methods mentioned above. 15

Summary of the Invention

The present invention concerns therapeutic agents that modulate the activity of TALL-1. In accordance with the present invention, modulators of TALL-1 may comprise an amino acid sequence Dz²Lz⁴ (SEQ ID NO: 108) wherein z^2 is an amino acid residue and z^4 is threonyl or isoleucyl. Such modulators of TALL-1 comprise molecules of the following formulae:

 $a^{1}a^{2}a^{3}CDa^{6}La^{8}a^{9}a^{10}Ca^{12}a^{13}a^{14}$ I(a)

(SEQ. ID. NO: 100)

wherein:

a1, a2, a3 are each independently absent or amino acid residues; 25 a6 is an amino acid residue; a9 is a basic or hydrophobic residue; a8 is threonyl or isoleucyl;

a10 is an amino acid residue; a12 is a neutral hydrophobic residue; and a¹³ and a¹⁴ are each independently absent or amino acid residues.

I(b)

 $b^1b^2b^3Cb^5b^6Db^8Lb^{10}b^{11}b^{12}b^{13}b^{14}Cb^{16}b^{17}b^{18}\\$

(SEQ. ID. NO: 104)

wherein:

b1 and b2 are each independently absent or amino acid residues;

b³ is an acidic or amide residue;

b⁵ is an amino acid residue; 10

b6 is an aromatic residue;

b8 is an amino acid residue;

b10 is T or I;

b11 is a basic residue;

b12 and b13 are each independently amino acid residues; 15

b14 is a neutral hydrophobic residue; and

 b^{16} , b^{17} , and b^{18} are each independently absent or amino acid residues.

I(c)

 $c^1c^2c^3Cc^5Dc^7Lc^9c^{10}c^{11}c^{12}c^{13}c^{14}Cc^{16}c^{17}c^{18}$

(SEQ. ID. NO:105)

wherein: 20

c1, c2, and c3 are each independently absent or amino acid residues;

c⁵ is an amino acid residue;

c⁷ is an amino acid residue;

c9 is T or I;

c10 is a basic residue; 25

c11 and c12 are each independently amino acid residues;

c¹³ is a neutral hydrophobic residue;

c14 is an amino acid residue;

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c16 is an amino acid residue;
                c17 is a neutral hydrophobic residue; and
                c18 is an amino acid residue or is absent.
      I(d)
                          d^{1}d^{2}d^{3}Cd^{5}d^{6}d^{7}WDd^{10}Ld^{12}d^{13}d^{14}Cd^{16}d^{17}d^{18}
                                         (SEQ. ID. NO: 106)
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      wherein:
                d<sup>1</sup>, d<sup>2</sup>, and d<sup>3</sup> are each independently absent or amino acid residues;
                d<sup>5</sup>, d<sup>6</sup>, and d<sup>7</sup> are each independently amino acid residues;
                d10 is an amino acid residue;
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                d12 is T or I;
                d13 is an amino acid residue;
                d14 is an amino acid residue; and
                d<sup>16</sup>, d<sup>17</sup>, and d<sup>18</sup> are each independently absent or amino acid residues.
      I(e)
                              e^{1}e^{2}e^{3}Ce^{5}e^{6}e^{7}De^{9}Le^{11}Ke^{13}Ce^{15}e^{16}e^{17}e^{18}
15
                                         (SEQ. ID. NO: 107)
      wherein:
                e1, e2, and e3 are each independently absent or amino acid residues;
                e5, e6, e7, e9, and e13 are each independently amino acid residues;
                e11 is T or I; and
               e<sup>15</sup>, e<sup>16</sup>, e<sup>17</sup>, and e<sup>18</sup> are each independently absent or amino acid residues.
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      I(f)
                                          f^1f^2f^3Kf^5Df^7Lf^9f^{10}Qf^{12}f^{13}f^{14}
                                                  (SEQ. ID NO: 109)
      wherein:
                f<sup>1</sup>, f<sup>2</sup>, and f<sup>3</sup> are absent or are amino acid residues (with one of f<sup>1</sup>, f<sup>2</sup>,
                         and f<sup>3</sup> preferred to be C when one of f<sup>12</sup>, f<sup>13</sup>, and f<sup>14</sup> is C);
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                f<sup>5</sup> is W, Y, or F (W preferred);
                f<sup>7</sup> is an amino acid residue (L preferred);
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f9 is T or I (T preferred);

f¹⁰ is K, R, or H (K preferred);

f¹² is C, a neutral hydrophobic residue, or a basic residue (W, C, or R preferred);

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f¹³ is C, a neutral hydrophobic residue or is absent (V preferred); and f¹⁴ is any amino acid residue or is absent;

provided that only one of f¹, f², and f³ may be C, and only one of f¹², f¹³, and f14 may be C.

Compounds of formulae I(a) through I(f) above incorporate Dz²Lz⁴, as well as SEQ ID NO: 63 hereinafter. The sequence of I(f) was derived as a consensus sequence as described in Example 1 hereinbelow. Of compounds within formula I(f), those within the formula

I(f') f1f2f3KWDf7Lf9KQf12f13f14

(SEQ ID NO: 125)

are preferred. Compounds falling within formula I(f') include SEQ ID NOS: 32, 58, 60, 62, 63, 66, 67, 69, 70, 114, 115, 122, 123, 124, 147-150, 152-177, 179, 180, 187.

In one aspect the present invention provides a TALL-1-binding composition of matter comprising an amino acid sequence Dz²Lz⁴, wherein z² is an amino acid residue and z4 is T or I, and wherein the composition of matter does not comprise the sequence FRKYDLLIHQRV or a fragment of TACI, BCMA, or BAFFR (SEQ ID NOS: 195, 196, and 197).

Further, the present invention provides a composition of matter of the invention, wherein z4 is T.

Also in accordance with the present invention are compounds having the consensus motif: 25

PFPWE

(SEQ ID NO: 110)

which also bind TALL-1.

Further, in accordance with the present invention are compounds of the formulae:

I(g)

 $g^{1}g^{2}g^{3}Cg^{5}PFg^{8}Wg^{10}Cg^{12}g^{13}g^{14}$

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(SEQ. ID. NO. 101)

wherein:

g1, g2 and g3 are each independently absent or amino acid residues;

g⁵ is a neutral polar residue;

g8 is a neutral polar residue;

g10 is an acidic residue; 10

g12 and g13 are each independently amino acid residues; and

g14 is absent or is an amino acid residue.

I(h)

h1h2h3CWh6h7WGh10Ch12h13h14

(SEQ. ID. NO: 102)

15 wherein:

h¹, h², and h³ are each independently absent or amino acid residues;

h⁶ is a hydrophobic residue;

h⁷ is a hydrophobic residue;

h¹⁰ is an acidic or polar hydrophobic residue; and

 h^{12} , h^{13} , and h^{14} are each independently absent or amino acid residues.

I(i)

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 $i^1i^2i^3Ci^5i^6i^7i^8i^9i^{10}Ci^{12}i^{13}i^{14}$

(SEQ. ID. NO: 103)

wherein:

i1 is absent or is an amino acid residue;

25 i² is a neutral hydrophobic residue;

i³ is an amino acid residue:

i⁵, i⁶, i⁷, and i⁸ are each independently amino acid residues;

i9 is an acidic residue;

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i¹⁰ is an amino acid residue;

i¹² and i¹³ are each independently amino acid residues; and i¹⁴ is a neutral hydrophobic residue.

The compounds defined by formulae I(g) through I(i) also bind TALL-1. Further, in accordance with the present invention, modulators of TALL-1 comprise:

- a TALL-1 modulating domain (e.g., an amino acid sequence of a) Formulae I(a) through I(i)), preferably the amino acid sequence Dz²Lz⁴, or sequences derived therefrom by phage display, RNApeptide screening, or the other techniques mentioned above; and
- b) a vehicle, such as a polymer (e.g., PEG or dextran) or an Fc domain, which is preferred;

wherein the vehicle is covalently attached to the TALL-1 modulating domain. The vehicle and the TALL-1 modulating domain may be linked through the Nor C-terminus of the TALL-1 modulating domain, as described further below. The preferred vehicle is an Fc domain, and the preferred Fc domain is an IgG Fc domain. Such Fc-linked peptides are referred to herein as "peptibodies." Preferred TALL-1 modulating domains comprise the amino acid sequences described hereinafter in Tables 1 and 2. Other TALL-1 modulating domains can be generated by phage display, RNA-peptide screening and the other techniques mentioned herein.

Further, in accordance with the present invention is a process for making TALL-1 modulators, which comprises:

- selecting at least one peptide that binds to TALL-1; and a.
- covalently linking said peptide to a vehicle.

The preferred vehicle is an Fc domain. Step (a) is preferably carried out by selection from the peptide sequences in Table 2 hereinafter or from phage display, RNA-peptide screening, or the other techniques mentioned herein.

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Further, the present invention provides a TALL-1 binding composition of matter comprising an amino acid sequence of the formula PFPWE (SEQ ID NO: 110).

Further, the present invention also provides a composition of matter having the formula

$$(X^1)_a - V^1 - (X^2)_b$$

and multimers thereof, wherein:

V¹ is a vehicle:

 X^1 and X^2 are each independently selected from -(L^1)_c- P^1 ,

$$-(L^1)_c - P^1 - (L^2)_d - P^2$$
, $-(L^1)_c - P^1 - (L^2)_d - P^2 - (L^3)_e - P^3$, and

$$\hbox{-(L^1)$}_c\,\hbox{-P}^1\hbox{-($L^2$)$}_d\,\hbox{-P}^2\,\hbox{-(L^3)$}_e\,\hbox{-P}^3\hbox{-($L^4$)$}_f\hbox{-P}^4;$$

L¹, L², L³, and L⁴ are each independently linkers; and

a, b, c, d, e, and f are each independently 0 or 1, provided that at lease one of a and b is 1

15 one or more of P¹, P², P³, and P⁴ each independently comprises a sequence selected from:

a¹a²a³CDa⁶La⁸a⁹a¹⁰Ca¹²a¹³a¹⁴ (SEQ. ID. NO: 100)

b1b2b3Cb5b6Db8Lb10b11b12b13b14Cb16b17b18 (SEO, ID, NO: 104)

 $c^{1}c^{2}c^{3}Cc^{5}Dc^{7}Lc^{9}c^{10}c^{11}c^{12}c^{13}c^{14}Cc^{16}c^{17}c^{18}$ (SEQ. ID. NO: 105)

 $d^{1}d^{2}d^{3}Cd^{5}d^{6}d^{7}WDd^{10}Ld^{12}d^{13}d^{14}Cd^{16}d^{17}d^{18}$ (SEQ. ID. NO: 106)

e¹e²e³Ce⁵e⁶e⁷De⁹Le¹¹Ke¹³Ce¹⁵e¹⁶e¹⁷e¹⁸ (SEQ. ID. NO: 107)

f¹f²f³Kf⁵Df⁷Lf⁹f¹⁰Qf¹²f¹³f¹⁴ (SEQ. ID. NO: 109)

g¹g²g³Cg⁵PFg⁸Wg¹⁰Cg¹²g¹³g¹⁴ (SEQ. ID. NO: 101),

h1h2h3CWh6h7WGh10Ch12h13h14 (SEQ. ID. NO: 102), and

i¹i²i³Ci⁵i6i7i8i9i¹0Ci¹²i¹3i¹4 (SEQ. ID. NO: 103)

wherein:

a¹, a², a³ are each independently absent or amino acid residues; a6 is an amino acid residue:

a9 is a basic or hydrophobic residue; a8 is threonyl or isoleucyl; a¹⁰ is an amino acid residue; a¹² is a neutral hydrophobic residue; a¹³ and a¹⁴ are each independently absent or amino acid residues; 5 b1 and b2 are each independently absent or amino acid residues; b³ is an acidic or amide residue; b⁵ is an amino acid residue: b6 is an aromatic residue; b8 is an amino acid residue; 10 b¹⁰ is T or I; b¹¹ is a basic residue; b¹² and b¹³ are each independently amino acid residues; b14 is a neutral hydrophobic residue; b16, b17, and b18 are each independently absent or amino acid residues; 15 c¹, c², and c³ are each independently absent or amino acid residues; c5 is an amino acid residue; c⁷ is an amino acid residue: c9 is T or I: c10 is a basic residue; 20 c11 and c12 are each independently amino acid residues; c¹³ is a neutral hydrophobic residue; c14 is an amino acid residue; c16 is an amino acid residue; c17 is a neutral hydrophobic residue; and 25 c18 is an amino acid residue or is absent; d¹, d², and d³ are each independently absent or amino acid residues; d⁵, d⁶, and d⁷ are each independently amino acid residues;

d10 is an amino acid residue;

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d12 is T or I;
               d13 is an amino acid residue;
               d14 is an amino acid residue; and
               d<sup>15</sup>, d<sup>16</sup>, and d<sup>17</sup> are each independently absent or amino acid residues;
              e<sup>1</sup>, e<sup>2</sup>, and e<sup>3</sup> are each independently absent or amino acid residues;
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               e<sup>5</sup>, e<sup>6</sup>, e<sup>7</sup>, e<sup>9</sup>, and e<sup>13</sup> are each independently amino acid residues;
               e11 is T or I; and
               e<sup>15</sup>, e<sup>16</sup>, e<sup>17</sup>, and e<sup>18</sup> are each independently absent or amino acid residues;
               f^1, f^2, and f^3 are absent or are amino acid residues;
               f<sup>5</sup> is W or F;
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               f<sup>7</sup> is an amino acid residue;
               f9 is T or I;
               f<sup>10</sup> is K, R, or H;
               f<sup>12</sup> is C, a neutral hydrophobic residue, or a basic residue;
               f<sup>13</sup> is C, a neutral hydrophobic residue or is absent; and
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               f<sup>14</sup> is any amino acid residue or is absent;
               provided that only one of f^1, f^2, and f^3 may be C, and only one of f^{12}, f^{13},
                        and f<sup>14</sup> may be C;
               g1, g2 and g3 are each independently absent or amino acid residues;
               g<sup>5</sup> is a neutral hydrophobic residue;
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               g8 is a neutral hydrophobic residue;
               g10 is an acidic residue;
               g12 and g13 are each independently amino acid residues; and
               g<sup>14</sup> is absent or is an amino acid residue;
               h<sup>1</sup>, h<sup>2</sup>, and h<sup>3</sup> are each independently absent or amino acid residues;
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               h<sup>6</sup> is a hydrophobic residue;
               h<sup>7</sup> is a hydrophobic residue;
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h¹⁰ is an acidic or polar hydrophobic residue; and

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h¹², h¹³, and h¹⁴ are each independently absent or amino acid residues;

i¹ is absent or is an amino acid residue;

i² is a neutral hydrophobic residue;

i³ is an amino acid residue;

i5, i6, i7, and i8 are each independently amino acid residues;

i9 is an acidic residue;

i10 is an amino acid residue;

i¹² and i¹³ are each independently amino acid residues; and

i¹⁴ is a neutral hydrophobic residue.

Further, the present invention also provides DNA encoding a composition of matter of the invention.

Further, the present invention provides an expression vector comprising the DNA of the invention.

Further, the present invention provides a host cell comprising the expression vector of the invention. 15

Further, the present invention provides a method of treating a B-cell mediated autoimmune disease, which comprises administering a composition of matter of the invention.

Further, the present invention provides a method of treating lupus, 20 which comprises administering a composition of matter of the invention.

Further, the present invention provides a method of treating a B-cell mediated cancer, which comprises administering a composition of matter of the invention.

Further, the present invention provides a method of treating B-cell lymphoma, which comprises administering a composition of matter of the invention.

Further, the present invention provides use of a composition of matter to

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the invention for the manufacture of a medicament for the treatment of B-cell mediated autoimmune disease.

Further, the present invention provides use of a composition of matter of the invention in the manufacture of a medicament for the treatment of B-cell mediated autoimmune disease.

Further, the present invention provides use of a composition of matter of the invention in the manufacture of a medicament for the treatment of lupus.

Further, the present invention provides use of a composition of matter of the invention in the manufacture of a medicament for the treatment of lupus.

Further, the present invention provides use of a composition of matter of the invention in the manufacture of a medicament for the treatment of a B-cell mediated cancer.

Further, the present invention provides use of a composition of a matter of the invention in the manufacture of a medicament for the treatment of B-cell lymphoma.

The compounds of this invention may be prepared by standard synthetic methods, recombinant DNA techniques, or any other methods of preparing peptides and fusion proteins. Compounds of this invention that encompass non-peptide portions may be synthesized by standard organic chemistry reactions, in addition to standard peptide chemistry reactions when applicable.

The primary use contemplated for the compounds of this invention is as therapeutic or prophylactic agents. The vehicle-linked peptide may have activity comparable to—or even greater than—the natural ligand mimicked by the peptide.

The compounds of this invention may be used for therapeutic or prophylactic purposes by formulating them with appropriate pharmaceutical carrier materials and administering an effective amount to a patient, such as a

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human (or other mammal) in need thereof. Other related aspects are also included in the instant invention.

Unless the context clearly requires otherwise, throughout the description and the claims, the words 'comprise', 'comprising', and the like are to be construed in an inclusive sense as opposed to an exclusive or exhaustive sense; that is to say, in the sense of "including, but not limited to".

Numerous additional aspects and advantages of the present invention will become apparent upon consideration of the figures and detailed description of the invention.

Brief Description of the Figures

Figure 1 shows exemplary Fc dimers that may be derived from an IgG1 antibody. "Fc" in the figure represents any of the Fc variants within the meaning of "Fc domain" herein. "X1" and "X2" represent peptides or linker-peptide combinations as defined hereinafter. The specific dimers are as follows:

A, D: Single disulfide-bonded dimers. IgG1 antibodies typically have two disulfide bonds at the hinge region of the antibody. The Fc domain in Figures 1A and 1D may be formed by truncation between the two disulfide bond sites or by substitution of a cysteinyl residue with an unreactive residue (e.g., alanyl). In Figure 1A, the Fc domain is linked at the amino terminus of the peptides; in 1D, at the carboxyl terminus.

B, E: Doubly disulfide-bonded dimers. This Fc domain may be formed by truncation of the parent antibody to retain both cysteinyl residues in the Fc domain chains or by expression from a construct including a sequence encoding such an Fc domain. In Figure 1B, the Fc domain is linked at the amino terminus

of the peptides; in 1E, at the carboxyl terminus.

C, F: Noncovalent dimers. This Fc domain may be formed by elimination of the cysteinyl residues by either truncation or substitution. One may desire to eliminate the cysteinyl residues to avoid impurities formed by reaction of the cysteinyl residue with cysteinyl residues of other proteins present in the host cell. The noncovalent bonding of the Fc domains is sufficient to hold together the dimer.

Other dimers may be formed by using Fc domains derived from different types of antibodies (e.g., IgG2, IgM).

Figure 2 shows the structure of preferred compounds of the invention that feature tandem repeats of the pharmacologically active peptide. Figure 2A shows a single chain molecule and may also represent the DNA construct for the molecule. Figure 2B shows a dimer in which the linker-peptide portion is present on only one chain of the dimer. Figure 2C shows a dimer having the peptide portion on both chains. The dimer of Figure 2C will form spontaneously in certain host cells upon expression of a DNA construct encoding the single chain shown in Figure 3A. In other host cells, the cells could be placed in conditions favoring formation of dimers or the dimers can be formed in vitro.

Figure 3 shows exemplary nucleic acid and amino acid sequences (SEQ ID NOS: 1 and 2, respectively) of human IgG1 Fc that may be used in this invention.

Figures 4A through 4F show the nucleotide and amino acid sequences (SEQ ID NOS: 3-27) S of NdeI to SalI fragments encoding peptide and linker.

Figures 5A through 5M show the nucleotide sequence (SEQ ID NO: 28) of pAMG21-RANK-Fc vector, which was used to construct Fc-linked molecules of the present invention. These figures identify a number of features of the nucleic acid, including:

- promoter regions <u>PcopB</u>, <u>PrepA</u>, <u>RNAI</u>, APHII, luxPR, and luxPL;
- mRNA for APHII, luxR;

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 coding sequences and amino acid sequences for the proteins copB protein, copT, repAI, repA4, APHII, luxR, RANK, and Fc;

- binding sites for the proteins copB, CRP;
- hairpins T1, T2, T7, and toop;
- operator site for lux protein;

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enzyme restriction sites for <u>Pflll08I</u>, <u>BglII</u>, <u>ScaI</u>, <u>BmnI</u>, <u>DrdII</u>, <u>DraIII</u>, <u>BstBI</u>,
 <u>AceIII</u>, <u>AflII</u>, <u>PflMI</u>, <u>BglI</u>, <u>SfiI</u>, <u>BstEII</u>, <u>BspLullI</u>, <u>NspV</u>, <u>BplI</u>, <u>EagI</u>, <u>BcgI</u>, <u>NsiI</u>,
 <u>BsaI</u>, <u>Pspl406I</u>, <u>AatII</u>, <u>BsmI</u>, <u>NruI</u>, <u>NdeI</u>, <u>ApaLI</u>, <u>Acc65I</u>, <u>KpnI</u>, <u>SalI</u>, <u>AccI</u>, <u>BspEI</u>,
 <u>AhdI</u>, <u>BspHI</u>, <u>EconI</u>, <u>BsrGI</u>, <u>BmaI</u>, <u>SmaI</u>, <u>SexAI</u>, <u>BamHI</u>, and <u>BlpI</u>.

Figures 6A and 6B show the DNA sequence (SEQ ID NO: 97) inserted into pCFM1656 between the unique <u>Aat</u>II (position #4364 in pCFM1656) and <u>Sac</u>II (position #4585 in pCFM1656) restriction sites to form expression plasmid pAMG21 (ATCC accession no. 98113).

Figure 7 shows that the TALL-1 peptibody (SEQ ID NO: 70) inhibits TALL-1-mediated B cell proliferation. Purified B cells (10^5) from B6 mice were cultured in triplicates in 96-well plated with the indicated amounts of TALL-1 consensus peptibody in the presence of 10 ng/ml TALL-1 plus $2 \mu \text{g/ml}$ anti-IgM antibody. Proliferation was measured by radioactive [3 H]thymidine uptake in the last 18h of pulse. Data shown represent mean \pm SD triplicate wells.

Figure 8 shows that a TALL-1 N-terminal tandem dimer peptibodies (SEQ ID NO: 123, 124 in Table 5B hereinafter) are preferable for inhibition of TALL-1-mediated B cell proliferation. Purified B cells (10^5) from B6 mice were cultured in triplicates in 96-well plated with the indicated amounts of TALL-1 12-3 peptibody and TALL-1 consensus peptibody (SEQ ID NOS: 115 and 122 of Table 5B)or the related dimer peptibodies (SEQ ID NOS: 123, 124) in the presence of 10 ng/ml TALL-1 plus 2 μ g/ml anti-IgM antibody. Proliferation was measured by radioactive [3 H]thymidine uptake in the last 18h of pulse. Data shown represent mean \pm SD triplicate wells.

Figure 9. AGP3 peptibody binds to AGP3 with high affinity.

30 Dissociation equilibrium constant (K_D) was obtained from nonlinear regression

of the competition curves using a dual-curve one-site homogeneous binding model (KinExTM software). K_D is about 4 pM for AGP3 peptibody binding with human AGP3 (SEQ ID NO: 123).

Figures 10A and 10B. AGP3 peptibody blocks both human and murine AGP3 in the Biacore competition assay. Soluble human TACI protein was immobilized to B1 chip. 1 nM of recombinant human AGP3 protein (upper panel) or 5 nM of recombinant murine AGP3 protein (lower panel) was incubated with indicated amount of AGP3 peptibody before injected over the surface of receptor. Relative human AGP3 and murine AGP3 (binding response was shown (SEQ ID NO: 123).

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Figures 11A and 11B. AGP3 peptibody blocked AGP3 binding to all three receptors TACI, BCMA and BAFFR in Biacore competition assay. Recombinant soluble receptor TACI, BCMA and BAFFR proteins were immobilized to CM5 chip. 1 nM of recombinant human AGP3 (upper panel) were incubated with indicated amount of AGP3 peptibody before injected over each receptor surface. Relative binding of AGP3 was measured. Similarly, 1 nM of recombinant APRIL protein was incubated with indicated amount of AGP3 peptibody before injected over each receptor surface. AGP3 peptibody didn't inhibit APRIL binding to all three receptors (SEQ ID NO: 123).

Figures 12A and 12B. AGP3 peptibody inhibits mouse serum immunoglobulin level increase induced by human AGP3 challenge. Balb/c mice received 7 daily intraperitoneal injections of 1 mg/Kg human AGP3 protein along with saline, human Fc, or AGP3 peptibody at indicated doses, and were bled on day 8. Serum total IgM and IgA level were measured by ELISA (SEQ ID NO: 123).

Figure 13. AGP3 peptibody treatment reduced arthritis severity in the mouse CIA model. Eight to 12 weeks old DBA/1 male mice were immunized with bovine collagen type II (bCII) emulsified in complete freunds adjuvant intradermally at the base of tail, and were boosted 3 weeks after the initial immunization with bCII emulsified in incomplete freunds adjuvant. Treatment with indicated dosage of AGP3 peptibody was begun from the day of booster

immunization for 4 weeks. As described before (Khare et al., *J. Immunol.*. 155: 3653-9, 1995), all four paws were individually scored from 0-3 for arthritis severity (SEQ ID NO: 123).

Figure 14. AGP3 peptibody treatment inhibited anti-collagen antibody generation in the mouse CIA model. Serum samples were taken one week after final treatment (day 35) as described above. Serum anti-collagen II antibody level was determined by ELISA analysis (SEQ ID NO: 123).

Figures 15A and 15B. AGP3 peptibody treatment delayed proteinuria onset and improved survival in NZB/NZW lupus mice. Five-month-old lupus prone NZBx NZBWF1 mice were treated i.p. 3X/week for 8 weeks with PBS or indicated doses of AGP3 peptibody (SEQ ID NO: 123) or human Fc proteins. Protein in the urine was evaluated monthly throughout the life of the experiment with Albustix reagent strips (Bayer AG).

Figures 16A and 16B show the nucleic acid and amino acid sequences of a preferred TALL-1-binding peptibody (SEQ ID NOS: 189 and 123)

Detailed Description of the Invention

Definition of Terms

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The terms used throughout this specification are defined as follows, unless otherwise limited in specific instances.

General definitions

The term "comprising" means that a compound may include additional amino acids on either or both of the N- or C- termini of the given sequence. Of course, these additional amino acids should not significantly interfere with the activity of the compound.

Additionally, physiologically acceptable salts of the compounds of this invention are also encompassed herein. The term "physiologically acceptable salts" refers to any salts that are known or later discovered to be pharmaceutically acceptable. Some specific examples are: acetate;

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trifluoroacetate; hydrohalides, such as hydrochloride and hydrobromide; sulfate; citrate; tartrate; glycolate; and oxalate.

Amino acids

The term "acidic residue" refers to amino acid residues in D- or L-form having sidechains comprising acidic groups. Exemplary acidic residues include D and E.

The term "amide residue" refers to amino acids in D- or L-form having sidechains comprising amide derivatives of acidic groups. Exemplary residues include N and Q.

The term "aromatic residue" refers to amino acid residues in D- or L-form 10 having sidechains comprising aromatic groups. Exemplary aromatic residues include F, Y, and W.

The term "basic residue" refers to amino acid residues in D- or L-form having sidechains comprising basic groups. Exemplary basic residues include H, K, and R.

The term "hydrophilic residue" refers to amino acid residues in D- or Lform having sidechains comprising polar groups. Exemplary hydrophilic residues include C, S, T, N, and Q.

The term "nonfunctional residue" refers to amino acid residues in D- or Lform having sidechains that lack acidic, basic, or aromatic groups. Exemplary nonfunctional amino acid residues include M, G, A, V, I, L and norleucine (Nle).

The term "neutral hydrophobic residue" refers to amino acid residues in D- or L-form having sidechains that lack basic, acidic, or polar groups. Exemplary neutral hydrophobic amino acid residues include A, V, L, I, P, W, M, and F.

The term "polar hydrophobic residue" refers to amino acid residues in Dor L-form having sidechains comprising polar groups. Exemplary polar hydrophobic amino acid residues include T, G, S, Y, C, Q, and N.

The term "hydrophobic residue" refers to amino acid residues in Dor L-form having sidechains that lack basic or acidic groups. Exemplary hydrophobic amino acid residues include A, V, L, I, P, W, M, F, T, G, S, Y, C, Q, and N.

<u>Peptides</u>

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The term "peptide" refers to molecules of 1 to 40 amino acids, with molecules of 5 to 20 amino acids preferred. Exemplary peptides may comprise the TALL-1 modulating domain of a naturally occurring molecule or comprise randomized sequences.

The term "randomized" as used to refer to peptide sequences refers to fully random sequences (e.g., selected by phage display methods or RNA-peptide screening) and sequences in which one or more residues of a naturally occurring molecule is replaced by an amino acid residue not appearing in that position in the naturally occurring molecule. Exemplary methods for identifying peptide sequences include phage display, <u>E. coli</u> display, ribosome display, RNA-peptide screening, chemical screening, and the like.

The term "TALL-1 modulating domain" refers to any amino acid sequence that binds to the TALL-1 and comprises naturally occurring sequences or randomized sequences. Exemplary TALL-1 modulating domains can be identified or derived by phage display or other methods mentioned herein.

The term "TALL-1 antagonist" refers to a molecule that binds to the TALL-1 and increases or decreases one or more assay parameters opposite from the effect on those parameters by full length native TALL-1. Such activity can be determined, for example, by such assays as described in the subsection entitled "Biological activity of AGP-3" in the Materials & Methods section of the patent application entitled, "TNF-RELATED PROTEINS", WO 00/47740, published August 17, 2000.

Vehicles and peptibodies

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The term "vehicle" refers to a molecule that prevents degradation and/or increases half-life, reduces toxicity, reduces immunogenicity, or increases biological activity of a therapeutic protein. Exemplary vehicles include an Fc domain (which is preferred) as well as a linear polymer (e.g., polyethylene glycol (PEG), polylysine, dextran, etc.); a branched-chain polymer (see, for example, U.S. Patent No. 4,289,872 to Denkenwalter et al., issued September 15, 1981; 5,229,490 to Tam, issued July 20, 1993; WO 93/21259 by Frechet et al., published 28 October 1993); a lipid; a cholesterol group (such as a steroid); a carbohydrate or oligosaccharide (e.g., dextran); any natural or synthetic protein, polypeptide or peptide that binds to a salvage receptor; albumin, including human serum albumin (HSA), leucine zipper domain, and other such proteins and protein fragments. Vehicles are further described hereinafter.

The term "native Fc" refers to molecule or sequence comprising the sequence of a non-antigen-binding fragment resulting from digestion of whole antibody, whether in monomeric or multimeric form. The original immunoglobulin source of the native Fc is preferably of human origin and may be any of the immunoglobulins, although IgG1 and IgG2 are preferred. Native Fc's are made up of monomeric polypeptides that may be linked into dimeric or multimeric forms by covalent (i.e., disulfide bonds) and non-covalent association. The number of intermolecular disulfide bonds between monomeric subunits of native Fc molecules ranges from 1 to 4 depending on class (e.g., IgG, IgA, IgE) or subclass (e.g., IgG1, IgG2, IgG3, IgA1, IgGA2). One example of a native Fc is a disulfide-bonded dimer resulting from papain digestion of an IgG (see Ellison et al.

(1982), <u>Nucleic Acids Res</u>. 10: 4071-9). The term "native Fc" as used herein is generic to the monomeric, dimeric, and multimeric forms.

The term "Fc variant" refers to a molecule or sequence that is modified from a native Fc but still comprises a binding site for the salvage receptor, FcRn. International applications WO 97/34631 (published 25 September 1997) and WO 96/32478 describe exemplary Fc variants, as well as interaction with the salvage receptor, and are hereby incorporated by reference in their entirety. Thus, the term "Fc variant" comprises a molecule or sequence that is humanized from a non-human native Fc. Furthermore, a native Fc comprises sites that may be removed because they provide structural features or biological activity that are not required for the fusion molecules of the present invention. Thus, the term "Fc variant" comprises a molecule or sequence that lacks one or more native Fc sites or residues that affect or are involved in (1) disulfide bond formation, (2) incompatibility with a selected host cell (3) N-terminal heterogeneity upon expression in a selected host cell, (4) glycosylation, (5) interaction with complement, (6) binding to an Fc receptor other than a salvage receptor, or (7) antibody-dependent cellular cytotoxicity (ADCC). Fc variants are described in further detail hereinafter.

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The term "Fc domain" encompasses native Fc and Fc variant molecules and sequences as defined above. As with Fc variants and native Fc's, the term "Fc domain" includes molecules in monomeric or multimeric form, whether digested from whole antibody or produced by other means.

The term "multimer" as applied to Fc domains or molecules comprising Fc domains refers to molecules having two or more polypeptide chains associated covalently, noncovalently, or by both covalent and non-covalent interactions. IgG molecules typically form dimers; IgM, pentamers; IgD, dimers; and IgA, monomers, dimers,

trimers, or tetramers. Multimers may be formed by exploiting the sequence and resulting activity of the native Ig source of the Fc or by derivatizing (as defined below) such a native Fc.

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The term "dimer" as applied to Fc domains or molecules comprising Fc domains refers to molecules having two polypeptide chains associated covalently or non-covalently. Thus, exemplary dimers within the scope of this invention are as shown in Figure 1.

The terms "derivatizing" and "derivative" or "derivatized" comprise processes and resulting compounds respectively in which (1) the compound has a cyclic portion; for example, cross-linking between cysteinyl residues within the compound; (2) the compound is cross-linked or has a cross-linking site; for example, the compound has a cysteinyl residue and thus forms cross-linked dimers in culture or $\underline{\text{in vivo}}$; (3) one or more peptidyl linkage is replaced by a non-peptidyl linkage; (4) the N-terminus is replaced by -NRR¹, NRC(O)R¹, -NRC(O)OR¹, -NRS(O)₂R¹, -NHC(O)NHR, a succinimide group, or substituted or unsubstituted benzyloxycarbonyl-NH-, wherein R and R¹ and the ring substituents are as defined hereinafter; (5) the C-terminus is replaced by -C(O)R² or -NR³R⁴ wherein R², R³ and R⁴ are as defined hereinafter; and (6) compounds in which individual amino acid moieties are modified through treatment with agents capable of reacting with selected side chains or terminal residues. Derivatives are further described hereinafter.

The terms "peptibody" and "peptibodies" refer to molecules comprising an Fc domain and at least one peptide. Such peptibodies may be multimers or dimers or fragments thereof, and they may be derivatized. In the present invention, the molecules of formulae II through VI hereinafter are peptibodies when $V^{\rm I}$ is an Fc domain.

Structure of compounds

In General. The present inventors identified sequences capable of binding to and modulating the biological activity of TALL-1. These sequences can be modified through the techniques mentioned above by which one or more amino acids may be changed while maintaining or even improving the binding affinity of the peptide.

In the compositions of matter prepared in accordance with this invention, the peptide(s) may be attached to the vehicle through the peptide's N-terminus or C-terminus. Any of these peptides may be linked in tandem (i.e., sequentially), with or without linkers. Thus, the vehicle-peptide molecules of this invention may be described by the following formula:

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$$(X^1)_a - V^1 - (X^2)_b$$

15 wherein:

V¹ is a vehicle (preferably an Fc domain);

 X^{1} and X^{2} are each independently selected from -(L¹)_c-P¹, -(L¹)_c-P¹-(L²)_d-P², -(L¹)_c-P¹-(L²)_d-P²-(L³)_e-P³-(L⁴)_f-P⁴

 P^1 , P^2 , P^3 , and P^4 are each independently sequences of TALL-1

20 modulating domains, such as those of Formulae I(a) through I(i);

 L^1 , L^2 , L^3 , and L^4 are each independently linkers; and

a, b, c, d, e, and f are each independently 0 or 1, provided that at least one of a and b is 1.

Thus, compound II comprises preferred compounds of the

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$$X^1-V^1$$

and multimers thereof wherein V^1 is an Fc domain and is attached at the C-terminus of A^1 ;

IV

$$V^1-X^2$$

and multimers thereof wherein V^1 is an Fc domain and is attached at the N-terminus of A^2 ;

5 V

$$V^{1}-(L^{1})_{c}-P^{1}$$

and multimers thereof wherein V^i is an Fc domain and is attached at the N-terminus of -(L^i), - P^i ; and

VI

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$$V^{1}-(L^{1})_{c}-P^{1}-(L^{2})_{d}-P^{2}$$

and multimers thereof wherein V^1 is an Fc domain and is attached at the N-terminus of $-L^1-P^1-L^2-P^2$.

<u>Peptides</u>. The peptides of this invention are useful as TALL-1 modulating peptides or as TALL-1 modulating domains in the molecules of formulae II through VI. Molecules of this invention comprising these peptide sequences may be prepared by methods known in the art.

Preferred peptide sequences are those of the foregoing formulae I(a) having the substituents identified below.

Table 1--Preferred peptide substituents

Formula I(a)	a ⁸ is T;	
	a ⁹ is a basic residue (K most preferred); and	
	a ¹² is a neutral hydrophibic residue (F most preferred).	
Formula I(b)	b ³ is D, Q, or E;	
	b ⁶ is W or Y;	
	b ¹⁰ is T;	
	b ¹¹ is K or R; and	
	b ¹⁴ is V or L.	
Formula I(c)	c ⁹ is T;	
	c ¹⁰ is K or R;	
	c ¹³ is a I, L, or V; and	
	c ¹⁷ is A or L.	
Formula I(d)	d ¹² is T.	
Formula I(e)	e ¹¹ is T.	
Formula I(f)	f ⁹ is T;	
	f ¹⁰ is K; and	
	f ¹³ is V.	
Formula I(g)	g ⁵ is W;	
	g ⁸ is P;	
	g ¹⁰ is E; and	
	g ¹³ is a basic residue.	
Formula I(h)	h ¹ is G;	
	h ⁶ is A;	
	h ⁷ is a neutral hydrophobic residue; and	
	h ¹⁰ is an acidic residue.	
Formula I(i)	i² is W; and	
	i¹⁴ is W.	

Preferred peptide sequences appear in Table 2 below.

Table 2—Preferred TALL-1 modulating domains

Sequence	SEQ ID NO:
PGTCFPFPWECTHA	29
WGACWPFPWECFKE	30
VPFCDLLTKHCFEA	31
GSRCKYKWDVLTKQCFHH	32
LPGCKWDLLIKQWVCDPL	33
SADCYFDILTKSDVCTSS	34
SDDCMYDQLTRMFICSNL	35
DLNCKYDELTYKEWCQFN	36
FHDCKYDLLTRQMVCHGL	37
RNHCFWDHLLKQDICPSP	38
ANQCWWDSLTKKNVCEFF	39
YKGRQMWDILTRSWVVSL	126
QDVGLWWDILTRAWMPNI	127
QNAQRVWDLLIRTWVYPQ	128
GWNEAWWDELTKIWVLEQ	129
RITCDTWDSLIKKCVPQS	130
GAIMQFWDSLTKTWLRQS	131
WLHSGWWDPLTKHWLQKV	132
SEWFFWFDPLTRAQLKFR	133
GVWFWWFDPLTKQWTQAG	134
MQCKGYYDILTKWCVTNG	135
LWSKEVWDILTKSWVSQA	136
KAAGWWFDWLTKVWVPAP	137
AYQTWFWDSLTRLWLSTT	138
SGQHFWWDLLTRSWTPST	139
LGVGQKWDPLTKQWVSRG	140
VGKMCQWDPLIKRTVCVG	141
CRQGAKFDLLTKQCLLGR	142
GQAIRHWDVLTKQWVDSQ	143
RGPCGSWDLLTKHCLDSQ	144
WQWKQQWDLL/TKQMVWVG	145
PITICRKDLLTKQVVCLD	146
KTCNGKWDLLTKQCLQQA	147
KCLKGKWDLLTKQCVTEV	148
RCWNGKWDLLTKQCIHPW	149
NRDMRKWDPLIKQWIVRP	150
QAAAATWDLLTKQWLVPP	151
PEGGPKWDPLTKQFLPPV	152
QTPQKKWDLLTKQWFTRN	153
IGSPCKWDLLTKQMICQT	154
CTAAGKWDLLTKQCIQEK	155
VSQCMKWDLLTKQCLQGW	156
VWGTWKWDLLTKQYLPPQ	157
GWWEMKWDLLTKQWYRPQ	158
TAQVSKWDLLTKQWLPLA	159
QLWGTKWDLLTKQYIQIM	160
WATSQKWDLLTKQWVQNM	161
QRQCAKWDLLTKQCVLFY	162

KTTDCKWDLLTKQRICQV	163
LLCQGKWDLLTKQCLKLR	164
LMWFWKWDLLTKQLVPTF	165
QTWAWKWDLLTKQWIGPM	166
NKELLKWDLLTKQCRGRS	167
GQKDLKWDLLTKQYVRQS	168
PKPCQKWDLLTKQCLGSV	169
GQIGWKWDLLTKQWIQTR	170
VWLDWKWDLLTKQWIHPQ	171
QEWEYKWDLLTKQWGWLR	172
HWDSWKWDLLTKQWVVQA	173
TRPLQKWDLLTKQWLRVG	174
SDQWQKWDLLTKQWFWDV	175
QQTFMKWDLLTKQWIRRH	176
QGECRKWDLLTKQCFPGQ	177
GQMGWRWDPLIKMCLGPS	178
QLDGCKWDLLTKQKVCIP	179
HGYWQKWDLLTKQWVSSE	180
HQGQCGWDLLTRIYLPCH	181
LHKACKWDLLTKQCWPMQ	182
GPPGSVWDLLTKIWIQTG	183
ITQDWRFDTLTRLWLPLR	184
QGGFAAWDVLTKMWITVP	185
GHGTPWWDALTRIWILGV	186
VWPWQKWDLLTKQFVFQD	187
WQWSWKWDLLTRQYISSS	188
NQTLWKWDLLTKQFITYM	60
PVYQGWWDTLTKLYIWDG	61
WLDGGWRDPLIKRSVQLG	62
GHQQFKWDLLTKQWVQSN	63
QRVGQFWDVLTKMFITGS	64
QAQGWSYDALIKTWIRWP	65
GWMHWKWDPLTKQALPWM	66
GHPTYKWDLLTKQWILQM	67
WNNWSLWDPLTKLWLQQN	68
WQWGWKWDLLTKQWVQQQ	69
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It is noted that the known receptors for TALL-1 bear some sequence homology with preferred peptides:

12-3 LPGCKWDLLIKQWVCDPL

BAFFR MRRGPRSLRGRDAPVPTPCVPTECYDLLVRKCVDCRLL

TACI TICNHQSQRTCAAFCRSLSCRKEQGKFYDHLLRDCISCASI

BCMA FVSPSQEIRGRFRRMLQMAGQCSQNEYFDSLLHACIPCQLRC

(SEQ ID NOS: 33, 195, 196, and 197, respectively).

Any peptide containing a cysteinyl residue may be cross-linked with another Cys-containing peptide, either or both of which may be linked to a

vehicle. Any peptide having more than one Cys residue may form an intrapeptide disulfide bond, as well. Any of these peptides may be derivatized as described hereinafter.

Additional useful peptide sequences may result from conservative and/or non-conservative modifications of the amino acid sequences of the sequences in Table 2.

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Conservative modifications will produce peptides having functional and chemical characteristics similar to those of the peptide from which such modifications are made. In contrast, substantial modifications in the functional and/or chemical characteristics of the peptides may be accomplished by selecting substitutions in the amino acid sequence that differ significantly in their effect on maintaining (a) the structure of the molecular backbone in the area of the substitution, for example, as a sheet or helical conformation, (b) the charge or hydrophobicity of the molecule at the target site, or (c) the size of the molecule.

For example, a "conservative amino acid substitution" may involve a substitution of a native amino acid residue with a nonnative residue such that there is little or no effect on the polarity or charge of the amino acid residue at that position. Furthermore, any native residue in the polypeptide may also be substituted with alanine, as has been previously described for "alanine scanning mutagenesis" (see, for example, MacLennan et al., 1998, Acta Physiol. Scand. Suppl. 643:55-67; Sasaki et al., 1998, Adv. Biophys. 35:1-24, which discuss alanine scanning mutagenesis).

Desired amino acid substitutions (whether conservative or non-conservative) can be determined by those skilled in the art at the time such substitutions are desired. For example, amino acid substitutions can be used to identify important residues of the peptide sequence, or to increase or decrease the affinity of the peptide or vehicle-peptide molecules (see preceding formulae) described herein. Exemplary amino acid substitutions are set forth in Table 3.

Table 3—Amino Acid Substitutions

Original Residues	Exemplary Substitutions	Preferred Substitutions
Ala (A)	Val, Leu, Ile	Val
Arg (R)	Lys, Gln, Asn	Lys
Asn (N)	Gln	Gln
Asp (D)	Glu	Glu
Cys (C)	Ser, Ala	Ser
Gln (Q)	Asn	Asn
Glu (E)	Asp	Asp
Gly (G)	Pro, Ala	Ala
His (H)	Asn, Gln, Lys, Arg	Arg
lle (I)	Leu, Val, Met, Ala, Phe, Norleucine	Leu
Leu (L)	Norleucine, Ile, Val, Met, Ala, Phe	lle
Lys (K)	Arg, 1,4 Diamino- butyric Acid, Gln, Asn	Arg
Met (M)	Leu, Phe, Ile	Leu
Phe (F)	Leu, Val, Ile, Ala, Tyr	Leu
Pro (P)	Ala	Gly
Ser (S)	Thr, Ala, Cys	Thr
Thr (T)	Ser	Ser
Trp (W)	Tyr, Phe	Tyr
Tyr (Y)	Trp, Phe, Thr, Ser	Phe
Val (V)	lle, Met, Leu, Phe, Ala, Norleucine	Leu

In certain embodiments, conservative amino acid substitutions also encompass non-naturally occurring amino acid residues which are

typically incorporated by chemical peptide synthesis rather than by synthesis in biological systems.

As noted in the foregoing section "Definition of Terms," naturally occurring residues may be divided into classes based on common sidechain properties that may be useful for modifications of sequence. For example, non-conservative substitutions may involve the exchange of a member of one of these classes for a member from another class. Such substituted residues may be introduced into regions of the peptide that are homologous with non-human orthologs, or into the non-homologous regions of the molecule. In addition, one may also make modifications using P or G for the purpose of influencing chain orientation.

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In making such modifications, the hydropathic index of amino acids may be considered. Each amino acid has been assigned a hydropathic index on the basis of their hydrophobicity and charge characteristics, these are: isoleucine (+4.5); valine (+4.2); leucine (+3.8); phenylalanine (+2.8); cysteine/cystine (+2.5); methionine (+1.9); alanine (+1.8); glycine (-0.4); threonine (-0.7); serine (-0.8); tryptophan (-0.9); tyrosine (-1.3); proline (-1.6); histidine (-3.2); glutamate (-3.5); glutamine (-3.5); aspartate (-3.5); asparagine (-3.5); lysine (-3.9); and arginine (-4.5).

The importance of the hydropathic amino acid index in conferring interactive biological function on a protein is understood in the art. Kyte et al., J. Mol. Biol., 157: 105-131 (1982). It is known that certain amino acids may be substituted for other amino acids having a similar hydropathic index or score and still retain a similar biological activity. In making changes based upon the hydropathic index, the substitution of amino acids whose hydropathic indices are within ± 2 is preferred, those which are within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred.

It is also understood in the art that the substitution of like amino acids can be made effectively on the basis of hydrophilicity. The greatest local average hydrophilicity of a protein, as governed by the hydrophilicity of its adjacent amino acids, correlates with its immunogenicity and antigenicity, <u>i.e.</u>, with a biological property of the protein.

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The following hydrophilicity values have been assigned to amino acid residues: arginine (\pm 3.0); lysine (\pm 3.0); aspartate (\pm 3.0 \pm 1); glutamate (\pm 3.0 \pm 1); serine (\pm 0.3); asparagine (\pm 0.2); glutamine (\pm 0.2); glycine (0); threonine (\pm 0.4); proline (\pm 0.5 \pm 1); alanine (\pm 0.5); histidine (\pm 0.5); cysteine (\pm 1.0); methionine (\pm 1.3); valine (\pm 1.5); leucine (\pm 1.8); isoleucine (\pm 1.8); tyrosine (\pm 2.3); phenylalanine (\pm 2.5); tryptophan (\pm 3.4). In making changes based upon similar hydrophilicity values, the substitution of amino acids whose hydrophilicity values are within \pm 2 is preferred, those which are within \pm 1 are particularly preferred, and those within \pm 0.5 are even more particularly preferred. One may also identify epitopes from primary amino acid sequences on the basis of hydrophilicity. These regions are also referred to as "epitopic core regions."

A skilled artisan will be able to determine suitable variants of the polypeptide as set forth in the foregoing sequences using well known techniques. For identifying suitable areas of the molecule that may be changed without destroying activity, one skilled in the art may target areas not believed to be important for activity. For example, when similar polypeptides with similar activities from the same species or from other species are known, one skilled in the art may compare the amino acid sequence of a peptide to similar peptides. With such a comparison, one can identify residues and portions of the molecules that are conserved among similar polypeptides. It will be appreciated that changes in areas of a peptide that are not conserved relative to such similar peptides would

be less likely to adversely affect the biological activity and/or structure of the peptide. One skilled in the art would also know that, even in relatively conserved regions, one may substitute chemically similar amino acids for the naturally occurring residues while retaining activity (conservative amino acid residue substitutions). Therefore, even areas that may be important for biological activity or for structure may be subject to conservative amino acid substitutions without destroying the biological activity or without adversely affecting the peptide structure.

Additionally, one skilled in the art can review structure-function studies identifying residues in similar peptides that are important for activity or structure. In view of such a comparison, one can predict the importance of amino acid residues in a peptide that correspond to amino acid residues that are important for activity or structure in similar peptides. One skilled in the art may opt for chemically similar amino acid substitutions for such predicted important amino acid residues of the peptides.

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One skilled in the art can also analyze the three-dimensional structure and amino acid sequence in relation to that structure in similar polypeptides. In view of that information, one skilled in the art may predict the alignment of amino acid residues of a peptide with respect to its three dimensional structure. One skilled in the art may choose not to make radical changes to amino acid residues predicted to be on the surface of the protein, since such residues may be involved in important interactions with other molecules. Moreover, one skilled in the art may generate test variants containing a single amino acid substitution at each desired amino acid residue. The variants can then be screened using activity assays know to those skilled in the art. Such data could be used to gather information about suitable variants. For example, if one discovered that a change to a particular amino acid residue resulted in destroyed,

undesirably reduced, or unsuitable activity, variants with such a change would be avoided. In other words, based on information gathered from such routine experiments, one skilled in the art can readily determine the amino acids where further substitutions should be avoided either alone or in combination with other mutations.

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A number of scientific publications have been devoted to the prediction of secondary structure. See Moult J., Curr. Op. in Biotech., 7(4): 422-427 (1996), Chou et al., Biochemistry, 13(2): 222-245 (1974); Chou et al., Biochemistry, 113(2): 211-222 (1974); Chou et al., Adv. Enzymol. Relat. Areas Mol. Biol., 47: 45-148 (1978); Chou et al., Ann. Rev. Biochem., 47: 251-276 and Chou et al., Biophys. J., 26: 367-384 (1979). Moreover, computer programs are currently available to assist with predicting secondary structure. One method of predicting secondary structure is based upon homology modeling. For example, two polypeptides or proteins which have a sequence identity of greater than 30%, or similarity greater than 40% often have similar structural topologies. The recent growth of the protein structural data base (PDB) has provided enhanced predictability of secondary structure, including the potential number of folds within a polypeptide's or protein's structure. See Holm et al., Nucl. Acid. Res., 27(1): 244-247 (1999). It has been suggested (Brenner et al., Curr. Op. Struct. Biol., 7(3): 369-376 (1997)) that there are a limited number of folds in a given polypeptide or protein and that once a critical number of structures have been resolved, structural prediction will gain dramatically in accuracy.

Additional methods of predicting secondary structure include "threading" (Jones, D., <u>Curr. Opin. Struct. Biol.</u>, 7(3): 377-87 (1997); Sippl <u>et al.</u>, <u>Structure</u>, 4(1): 15-9 (1996)), "profile analysis" (Bowie <u>et al., Science</u>, 253: 164-170 (1991); Gribskov et al., Meth. <u>Enzym.</u>, 183: 146-159 (1990);

Gribskov et al., Proc. Nat. Acad. Sci., 84(13): 4355-8 (1987)), and "evolutionary linkage" (See Home, supra, and Brenner, supra).

<u>Vehicles</u>. This invention requires the presence of at least one vehicle (V¹) attached to a peptide through the N-terminus, C-terminus or a sidechain of one of the amino acid residues. Multiple vehicles may also be used; e.g., Fc's at each terminus or an Fc at a terminus and a PEG group at the other terminus or a sidechain. Exemplary vehicles include:

- an Fc domain;
- other proteins, polypeptides, or peptides capable of binding to a salvage receptor;
- human serum albumin (HSA);
- a leucine zipper (LZ) domain;
- polyethylene glycol (PEG), including 5 kD, 20 kD, and 30 kD
 PEG, as well as other polymers;
- dextran;

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and other molecules known in the art to provide extended half-life and/or protection from proteolytic degradation or clearance.

An Fc domain is the preferred vehicle. The Fc domain may be fused to the N or C termini of the peptides or at both the N and C termini.

Fusion to the N terminus is preferred.

As noted above, Fc variants are suitable vehicles within the scope of this invention. A native Fc may be extensively modified to form an Fc variant in accordance with this invention, provided binding to the salvage receptor is maintained; see, for example WO 97/34631 and WO 96/32478.

In such Fc variants, one may remove one or more sites of a native Fc that provide structural features or functional activity not required by the fusion molecules of this invention. One may remove these sites by, for example, substituting or deleting residues, inserting residues into the site, or truncating portions containing the site. The inserted or substituted

residues may also be altered amino acids, such as peptidomimetics or D-amino acids. Fc variants may be desirable for a number of reasons, several of which are described below. Exemplary Fc variants include molecules and sequences in which:

- 1. Sites involved in disulfide bond formation are removed. Such removal may avoid reaction with other cysteine-containing proteins present in the host cell used to produce the molecules of the invention. For this purpose, the cysteine-containing segment at the N-terminus may be truncated or cysteine residues may be deleted or substituted with other amino acids (e.g., alanyl, seryl). In particular, one may truncate the N-terminal 20-amino acid segment of SEQ ID NO: 2 or delete or substitute the cysteine residues at positions 7 and 10 of SEQ ID NO: 2. Even when cysteine residues are removed, the single chain Fc domains can still form a dimeric Fc domain that is held together non-covalently.
- A native Fc is modified to make it more compatible with a selected host cell. For example, one may remove the PA sequence near the N-terminus of a typical native Fc, which may be recognized by a digestive enzyme in <u>E</u>. <u>coli</u> such as proline iminopeptidase. One may also add an N-terminal methionine residue, especially when the molecule is expressed recombinantly in a bacterial cell such as <u>E</u>. <u>coli</u>. The Fc domain of SEQ ID NO: 2 is one such Fc variant.
 - 3. A portion of the N-terminus of a native Fc is removed to prevent N-terminal heterogeneity when expressed in a selected host cell. For this purpose, one may delete any of the first 20 amino acid residues at the N-terminus, particularly those at positions 1, 2, 3, 4 and 5.

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4. One or more glycosylation sites are removed. Residues that are typically glycosylated (e.g., asparagine) may confer cytolytic response. Such residues may be deleted or substituted with unglycosylated residues (e.g., alanine).

5. Sites involved in interaction with complement, such as the C1q binding site, are removed. For example, one may delete or substitute the EKK sequence of human IgG1. Complement recruitment may not be advantageous for the molecules of this invention and so may be avoided with such an Fc variant.

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- 6. Sites are removed that affect binding to Fc receptors other than a salvage receptor. A native Fc may have sites for interaction with certain white blood cells that are not required for the fusion molecules of the present invention and so may be removed.
- 7. The ADCC site is removed. ADCC sites are known in the art; see, for example, Molec. Immunol. 29 (5): 633-9 (1992) with regard to ADCC sites in IgG1. These sites, as well, are not required for the fusion molecules of the present invention and so may be removed.
 - 8. When the native Fc is derived from a non-human antibody, the native Fc may be humanized. Typically, to humanize a native Fc, one will substitute selected residues in the non-human native Fc with residues that are normally found in human native Fc. Techniques for antibody humanization are well known in the art.

Preferred Fc variants include the following. In SEQ ID NO: 2

(Figure 3), the leucine at position 15 may be substituted with glutamate; the glutamate at position 99, with alanine; and the lysines at positions 101 and 103, with alanines. In addition, one or more tyrosine residues can be replaced by phenyalanine residues.

An alternative vehicle would be a protein, polypeptide, peptide, antibody, antibody fragment, or small molecule (e.g., a peptidomimetic compound) capable of binding to a salvage receptor. For example, one could use as a vehicle a polypeptide as described in U.S. Pat. No. 5,739,277, issued April 14, 1998 to Presta et al. Peptides could also be selected by phage display or RNA-peptide screening for binding to the

FcRn salvage receptor. Such salvage receptor-binding compounds are also included within the meaning of "vehicle" and are within the scope of this invention. Such vehicles should be selected for increased half-life (e.g., by avoiding sequences recognized by proteases) and decreased immunogenicity (e.g., by favoring non-immunogenic sequences, as discovered in antibody humanization).

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As noted above, polymer vehicles may also be used for V¹. Various means for attaching chemical moieties useful as vehicles are currently available, see, e.g., Patent Cooperation Treaty ("PCT") International Publication No. WO 96/11953, entitled "N-Terminally Chemically Modified Protein Compositions and Methods," herein incorporated by reference in its entirety. This PCT publication discloses, among other things, the selective attachment of water soluble polymers to the N-terminus of proteins.

A preferred polymer vehicle is polyethylene glycol (PEG). The PEG group may be of any convenient molecular weight and may be linear or branched. The average molecular weight of the PEG will preferably range from about 2 kiloDalton ("kD") to about 100 kD, more preferably from about 5 kD to about 50 kD, most preferably from about 5 kD to about 10 kD. The PEG groups will generally be attached to the compounds of the invention via acylation or reductive alkylation through a reactive group on the PEG moiety (e.g., an aldehyde, amino, thiol, or ester group) to a reactive group on the inventive compound (e.g., an aldehyde, amino, or ester group).

A useful strategy for the PEGylation of synthetic peptides consists of combining, through forming a conjugate linkage in solution, a peptide and a PEG moiety, each bearing a special functionality that is mutually reactive toward the other. The peptides can be easily prepared with conventional solid phase synthesis. The peptides are "preactivated" with

an appropriate functional group at a specific site. The precursors are purified and fully characterized prior to reacting with the PEG moiety. Ligation of the peptide with PEG usually takes place in aqueous phase and can be easily monitored by reverse phase analytical HPLC. The PEGylated peptides can be easily purified by preparative HPLC and characterized by analytical HPLC, amino acid analysis and laser desorption mass spectrometry.

Polysaccharide polymers are another type of water soluble polymer which may be used for protein modification. Dextrans are polysaccharide polymers comprised of individual subunits of glucose predominantly linked by α 1-6 linkages. The dextran itself is available in many molecular weight ranges, and is readily available in molecular weights from about 1 kD to about 70 kD. Dextran is a suitable water soluble polymer for use in the present invention as a vehicle by itself or in combination with another vehicle (e.g., Fc). See, for example, WO 96/11953 and WO 96/05309. The use of dextran conjugated to therapeutic or diagnostic immunoglobulins has been reported; see, for example, European Patent Publication No. 0 315 456, which is hereby incorporated by reference in its entirety. Dextran of about 1 kD to about 20 kD is preferred when dextran is used as a vehicle in accordance with the present invention.

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<u>Linkers</u>. Any "linker" group is optional. When present, its chemical structure is not critical, since it serves primarily as a spacer. The linker is preferably made up of amino acids linked together by peptide bonds. Thus, in preferred embodiments, the linker is made up of from 1 to 30 amino acids linked by peptide bonds, wherein the amino acids are selected from the 20 naturally occurring amino acids. Some of these amino acids may be glycosylated, as is well understood by those in the art. In a more preferred embodiment, the 1 to 20 amino acids are selected from glycine, alanine, proline, asparagine, glutamine, and lysine. Even more preferably,

a linker is made up of a majority of amino acids that are sterically unhindered, such as glycine and alanine. Thus, preferred linkers are polyglycines (particularly (Gly)₄, (Gly)₅), poly(Gly-Ala), and polyalanines. Other specific examples of linkers are:

(Gly)₃Lys(Gly)₄ (SEQ ID NO: 40); (Gly)₃AsnGlySer(Gly)₂ (SEQ ID NO: 41); (Gly)₃Cys(Gly)₄ (SEQ ID NO: 42); and GlyProAsnGlyGly (SEQ ID NO: 43).

To explain the above nomenclature, for example, (Gly)₃Lys(Gly)₄ means Gly-Gly-Gly-Gly-Gly-Gly-Gly (SEQ ID NO: 40). Combinations of Gly and Ala are also preferred. The linkers shown here are exemplary; linkers within the scope of this invention may be much longer and may include other residues.

Preferred linkers are amino acid linkers comprising greater than 5 amino acids, with suitable linkers having up to about 500 amino acids selected from glycine, alanine, proline, asparagine, glutamine, lysine, threonine, serine or aspartate. Linkers of about 20 to 50 amino acids are most preferred. One group of preferred linkers are those of the formulae

GSGSATGGSGSTASSGSGSATx1x2

(SEQ ID NO: 193)

and

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GSGSATGGSGSTASSGSGSATx¹x²GSGSATGGSGSTASSGSGSATx³x⁴
(SEQ ID NO: 194)

wherein x^1 and x^3 are each independently basic or hydrophobic residues and x^2 and x^4 are each independently hydrophobic residues. Specific preferred linkers are:

GSGSATGGSGSTASSGSGSATHM (SEQ ID NO: 59)

GSGSATGGSGSTASSGSGSATGM

(SEQ ID NO: 190)

GSGSATGGSGSTASSGSGSATGS

(SEQ ID NO: 191), and

5 GSGSATGGSGSTASSGSGSATHMGSGSATGGSGSTASSGSGSATHM (SEQ ID NO: 192).

Non-peptide linkers are also possible. For example, alkyl linkers such as -NH-(CH₂)_s-C(O)-, wherein s = 2-20 could be used. These alkyl linkers may further be substituted by any non-sterically hindering group such as lower alkyl (e.g., C_1 - C_6) lower acyl, halogen (e.g., Cl, Br), CN, NH₂, phenyl, etc. An exemplary non-peptide linker is a PEG linker, VII

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wherein n is such that the linker has a molecular weight of 100 to 5000 kD, preferably 100 to 500 kD. The peptide linkers may be altered to form derivatives in the same manner as described above.

<u>Derivatives</u>. The inventors also contemplate derivatizing the peptide and/or vehicle portion of the compounds. Such derivatives may improve the solubility, absorption, biological half life, and the like of the compounds. The moieties may alternatively eliminate or attenuate any undesirable side-effect of the compounds and the like. Exemplary derivatives include compounds in which:

1. The compound or some portion thereof is cyclic. For example, the peptide portion may be modified to contain two or more Cys residues (e.g., in the linker), which could cyclize by disulfide bond formation.

2. The compound is cross-linked or is rendered capable of cross-linking between molecules. For example, the peptide portion may be modified to contain one Cys residue and thereby be able to form an intermolecular disulfide bond with a like molecule. The compound may also be cross-linked through its C-terminus, as in the molecule shown below.

VIII

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$$V^{1}-(X^{1})_{b}-CO-N$$
 $V^{1}-(X^{1})_{b}-CO-N$
 NH_{2}
 NH_{2}

In Formula VIII, each "V" may represent typically one strand of the Fc domain.

- 3. One or more peptidyl [-C(O)NR-] linkages (bonds) is replaced by a non-peptidyl linkage. Exemplary non-peptidyl linkages are -CH₂-carbamate [-CH₂-OC(O)NR-], phosphonate , -CH₂-sulfonamide [-CH₂-S(O)₂NR-], urea [-NHC(O)NH-], -CH₂-secondary amine, and alkylated peptide [-C(O)NR⁶- wherein R⁶ is lower alkyl].
- acylated or modified to a substituted amine. Exemplary N-terminal derivative groups include -NRR¹ (other than -NH₂), -NRC(O)R¹, -NRC(O)OR¹, -NRS(O)₂R¹, -NHC(O)NHR¹, succinimide, or benzyloxycarbonyl-NH- (CBZ-NH-), wherein R and R¹ are each independently hydrogen or lower alkyl and wherein the phenyl ring may be substituted with 1 to 3 substituents selected from the group consisting of C_1 - C_4 alkyl, C_1 - C_4 alkoxy, chloro, and bromo.

4. The N-terminus is derivatized. Typically, the N-terminus may be

The free C-terminus is derivatized. Typically, the C-terminus is
 esterified or amidated. Exemplary C-terminal derivative groups
 include, for example, -C(O)R² wherein R² is lower alkoxy or -NR³R⁴

wherein R^3 and R^4 are independently hydrogen or C_1 - C_8 alkyl (preferably C_1 - C_4 alkyl).

A disulfide bond is replaced with another, preferably more stable, cross-linking moiety (e.g., an alkylene). See, e.g., Bhatnagar et al. (1996), J. Med. Chem. 39: 3814-9; Alberts et al. (1993) Thirteenth Am. Pep. Symp., 357-9.

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7. One or more individual amino acid residues is modified. Various derivatizing agents are known to react specifically with selected sidechains or terminal residues, as described in detail below.

Lysinyl residues and amino terminal residues may be reacted with succinic or other carboxylic acid anhydrides, which reverse the charge of the lysinyl residues. Other suitable reagents for derivatizing alpha-amino-containing residues include imidoesters such as methyl picolinimidate; pyridoxal phosphate; pyridoxal; chloroborohydride; trinitrobenzenesulfonic acid; O-methylisourea; 2,4 pentanedione; and transaminase-catalyzed reaction with glyoxylate.

Arginyl residues may be modified by reaction with any one or combination of several conventional reagents, including phenylglyoxal, 2,3-butanedione, 1,2-cyclohexanedione, and ninhydrin. Derivatization of arginyl residues requires that the reaction be performed in alkaline conditions because of the high pKa of the guanidine functional group. Furthermore, these reagents may react with the groups of lysine as well as the arginine epsilon-amino group.

Specific modification of tyrosyl residues has been studied extensively, with particular interest in introducing spectral labels into tyrosyl residues by reaction with aromatic diazonium compounds or tetranitromethane. Most commonly, N-acetylimidizole and tetranitromethane are used to form O-acetyl tyrosyl species and 3-nitro derivatives, respectively.

Carboxyl sidechain groups (aspartyl or glutamyl) may be selectively modified by reaction with carbodiimides (R'-N=C=N-R') such as 1-cyclohexyl-3-(2-morpholinyl-(4-ethyl) carbodiimide or 1-ethyl-3-(4-azonia-4,4-dimethylpentyl) carbodiimide. Furthermore, aspartyl and glutamyl residues may be converted to asparaginyl and glutaminyl residues by reaction with ammonium ions.

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Glutaminyl and asparaginyl residues may be deamidated to the corresponding glutamyl and aspartyl residues. Alternatively, these residues are deamidated under mildly acidic conditions. Either form of these residues falls within the scope of this invention.

Cysteinyl residues can be replaced by amino acid residues or other moieties either to eliminate disulfide bonding or, conversely, to stabilize cross-linking. See, e.g., Bhatnagar <u>et al.</u> (1996), J. <u>Med. Chem.</u> 39: 3814-9.

Derivatization with bifunctional agents is useful for cross-linking the peptides or their functional derivatives to a water-insoluble support matrix or to other macromolecular vehicles. Commonly used cross-linking agents include, e.g., 1,1-bis(diazoacetyl)-2-phenylethane, glutaraldehyde, N-hydroxysuccinimide esters, for example, esters with 4-azidosalicylic acid, homobifunctional imidoesters, including disuccinimidyl esters such as 3,3'-dithiobis(succinimidylpropionate), and bifunctional maleimides such as bis-N-maleimido-1,8-octane. Derivatizing agents such as methyl-3-[(p-azidophenyl)dithio]propioimidate yield photoactivatable intermediates that are capable of forming cross-links in the presence of light. Alternatively, reactive water-insoluble matrices such as cyanogen bromide-activated carbohydrates and the reactive substrates described in U.S. Pat. Nos. 3,969,287; 3,691,016; 4,195,128; 4,247,642; 4,229,537; and 4,330,440 are employed for protein immobilization.

Carbohydrate (oligosaccharide) groups may conveniently be attached to sites that are known to be glycosylation sites in proteins.

Generally, O-linked oligosaccharides are attached to serine (Ser) or threonine (Thr) residues while N-linked oligosaccharides are attached to asparagine (Asn) residues when they are part of the sequence Asn-X-Ser/Thr, where X can be any amino acid except proline. X is preferably one of the 19 naturally occurring amino acids other than proline. The structures of N-linked and O-linked oligosaccharides and the sugar residues found in each type are different. One type of sugar that is commonly found on both is N-acetylneuraminic acid (referred to as sialic acid). Sialic acid is usually the terminal residue of both N-linked and Olinked oligosaccharides and, by virtue of its negative charge, may confer acidic properties to the glycosylated compound. Such site(s) may be incorporated in the linker of the compounds of this invention and are preferably glycosylated by a cell during recombinant production of the polypeptide compounds (e.g., in mammalian cells such as CHO, BHK, COS). However, such sites may further be glycosylated by synthetic or semi-synthetic procedures known in the art.

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Other possible modifications include hydroxylation of proline and lysine, phosphorylation of hydroxyl groups of seryl or threonyl residues, oxidation of the sulfur atom in Cys, methylation of the alpha-amino groups of lysine, arginine, and histidine side chains. Creighton, <u>Proteins: Structure and Molecule Properties</u> (W. H. Freeman & Co., San Francisco), pp. 79-86 (1983).

Compounds of the present invention may be changed at the DNA level, as well. The DNA sequence of any portion of the compound may be changed to codons more compatible with the chosen host cell. For \underline{E} . \underline{coli} , which is the preferred host cell, optimized codons are known in the art. Codons may be substituted to eliminate restriction sites or to include silent restriction sites, which may aid in processing of the DNA in the selected

host cell. The vehicle, linker and peptide DNA sequences may be modified to include any of the foregoing sequence changes.

Methods of Making

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The compounds of this invention largely may be made in transformed host cells using recombinant DNA techniques. To do so, a recombinant DNA molecule coding for the peptide is prepared. Methods of preparing such DNA molecules are well known in the art. For instance, sequences coding for the peptides could be excised from DNA using suitable restriction enzymes. Alternatively, the DNA molecule could be synthesized using chemical synthesis techniques, such as the phosphoramidate method. Also, a combination of these techniques could be used.

The invention also includes a vector capable of expressing the peptides in an appropriate host. The vector comprises the DNA molecule that codes for the peptides operatively linked to appropriate expression control sequences. Methods of effecting this operative linking, either before or after the DNA molecule is inserted into the vector, are well known. Expression control sequences include promoters, activators, enhancers, operators, ribosomal binding sites, start signals, stop signals, cap signals, polyadenylation signals, and other signals involved with the control of transcription or translation.

The resulting vector having the DNA molecule thereon is used to transform an appropriate host. This transformation may be performed using methods well known in the art.

Any of a large number of available and well-known host cells may be used in the practice of this invention. The selection of a particular host is dependent upon a number of factors recognized by the art. These include, for example, compatibility with the chosen expression vector, toxicity of the peptides encoded by the DNA molecule, rate of

transformation, ease of recovery of the peptides, expression characteristics, bio-safety and costs. A balance of these factors must be struck with the understanding that not all hosts may be equally effective for the expression of a particular DNA sequence. Within these general guidelines, useful microbial hosts include bacteria (such as <u>E. coli</u> sp.), yeast (such as <u>Saccharomyces</u> sp.) and other fungi, insects, plants, mammalian (including human) cells in culture, or other hosts known in the art.

Next, the transformed host is cultured and purified. Host cells may be cultured under conventional fermentation conditions so that the desired compounds are expressed. Such fermentation conditions are well known in the art. Finally, the peptides are purified from culture by methods well known in the art.

The compounds may also be made by synthetic methods. For example, solid phase synthesis techniques may be used. Suitable techniques are well known in the art, and include those described in Merrifield (1973), Chem. Polypeptides, pp. 335-61 (Katsoyannis and Panayotis eds.); Merrifield (1963), J. Am. Chem. Soc. 85: 2149; Davis et al. (1985), Biochem. Intl. 10: 394-414; Stewart and Young (1969), Solid Phase Peptide Synthesis; U.S. Pat. No. 3,941,763; Finn et al. (1976), The Proteins (3rd ed.) 2: 105-253; and Erickson et al. (1976), The Proteins (3rd ed.) 2: 257-527. Solid phase synthesis is the preferred technique of making individual peptides since it is the most cost-effective method of making small peptides.

Compounds that contain derivatized peptides or which contain non-peptide groups may be synthesized by well-known organic chemistry techniques.

Uses of the Compounds

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Compounds of this invention may be particularly useful in treatment of B-cell mediated autoimmune diseases. In particular, the

compounds of this invention may be useful in treating, preventing, ameliorating, diagnosing or prognosing lupus, including systemic lupus erythematosus (SLE), and lupus-associated diseases and conditions. Other preferred indications include B-cell mediated cancers, including B-cell lymphoma.

The compounds of this invention can also be used to treat inflammatory conditions of the joints. Inflammatory conditions of a joint are chronic joint diseases that afflict and disable, to varying degrees, millions of people worldwide. Rheumatoid arthritis is a disease of articular joints in which the cartilage and bone are slowly eroded away by a proliferative, invasive connective tissue called pannus, which is derived from the synovial membrane. The disease may involve peri-articular structures such as bursae, tendon sheaths and tendons as well as extraarticular tissues such as the subcutis, cardiovascular system, lungs, spleen, lymph nodes, skeletal muscles, nervous system (central and peripheral) and eyes (Silberberg (1985), Anderson's Pathology, Kissane (ed.), II:1828). Osteoarthritis is a common joint disease characterized by degenerative changes in articular cartilage and reactive proliferation of bone and cartilage around the joint. Osteoarthritis is a cell-mediated active process that may result from the inappropriate response of chondrocytes to catabolic and anabolic stimuli. Changes in some matrix molecules of articular cartilage reportedly occur in early osteoarthritis (Thonar et al. (1993), Rheumatic disease clinics of North America, Moskowitz (ed.), 19:635-657 and Shinmei et al. (1992), Arthritis Rheum., 35:1304-1308). TALL-1, TALL-1R and modulators thereof are believed to be useful in the treatment of these and related conditions.

Compounds of this invention may also be useful in treatment of a number of additional diseases and disorders, including:

acute pancreatitis;

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- ALS;
- Alzheimer's disease;
- asthma;
- atherosclerosis;
- autoimmune hemolytic anemia;
 - cancer, particularly cancers related to B cells;
 - cachexia/anorexia;
 - chronic fatigue syndrome;
 - cirrhosis (e.g., primary biliary cirrhosis);
- diabetes (e.g., insulin diabetes);
 - fever;
 - glomerulonephritis, including IgA glomerulonephritis and primary glomerulonephritis;
 - Goodpasture's syndrome;
- Guillain-Barre syndrome;
 - graft versus host disease;
 - Hashimoto's thyroiditis;
 - hemorrhagic shock;
 - hyperalgesia;

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- inflammatory bowel disease;
 - inflammatory conditions of a joint, including osteoarthritis,
 psoriatic arthritis and rheumatoid arthritis;
 - inflammatory conditions resulting from strain, sprain, cartilage damage, trauma, orthopedic surgery, infection or other disease processes;
 - insulin-dependent diabetes mellitus;

• ischemic injury, including cerebral ischemia (e.g., brain injury as a result of trauma, epilepsy, hemorrhage or stroke, each of which may lead to neurodegeneration);

- learning impairment;
- lung diseases (e.g., ARDS);
 - multiple myeloma;
 - multiple sclerosis;
 - Myasthenia gravis;
 - myelogenous (e.g., AML and CML) and other leukemias;
- myopathies (e.g., muscle protein metabolism, esp. in sepsis);
 - neurotoxicity (e.g., as induced by HIV);
 - osteoporosis;
 - pain;
 - Parkinson's disease;
- Pemphigus;
 - polymyositis/dermatomyositis;
 - pulmonary inflammation, including autoimmune pulmonary inflammation;
 - pre-term labor;
- psoriasis;
 - Reiter's disease;
 - reperfusion injury;
 - septic shock;
 - side effects from radiation therapy;
- Sjogren's syndrome;
 - sleep disturbance;
 - temporal mandibular joint disease;

 thrombocytopenia, including idiopathic thrombocytopenia and autoimmune neonatal thrombocytopenia;

- tumor metastasis;
- uveitis; and
- vasculitis.

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Compounds of this invention may be administered alone or in combination with a therapeutically effective amount of other drugs, including analgesic agents, disease-modifying anti-rheumatic drugs (DMARDs), non-steroidal anti-inflammatory drugs (NSAIDs), and any immune and/or inflammatory modulators. Thus, compounds of this invention may be administered with:

- Modulators of other members of the TNF/TNF receptor family, including TNF antagonists, such as etanercept (Enbrel[™]), sTNF-RI, onercept, D2E7, and Remicade[™].
- Nerve growth factor (NGF) modulators.
- IL-1 inhibitors, including IL-1ra molecules such as anakinra and more recently discovered IL-1ra-like molecules such as IL-1Hy1 and IL-1Hy2; IL-1 "trap" molecules as described in U.S. Pat. No. 5,844,099, issued December 1, 1998; IL-1 antibodies; solubilized IL-1 receptor, and the like.
- IL-6 inhibitors (e.g., antibodies to IL-6).
- IL-8 inhibitors (e.g., antibodies to IL-8).
- IL-18 inhibitors (e.g., IL-18 binding protein, solubilized IL-18 receptor, or IL-18 antibodies).
- Interleukin-1 converting enzyme (ICE) modulators.
 - insulin-like growth factors (IGF-1, IGF-2) and modulators thereof.
 - Transforming growth factor- β (TGF- β), TGF- β family members, and TGF- β modulators.

 Fibroblast growth factors FGF-1 to FGF-10, and FGF modulators.

- Osteoprotegerin (OPG), OPG analogues, osteoprotective agents, and antibodies to OPG-ligand (OPG-L).
- bone anabolic agents, such as parathyroid hormone (PTH), PTH fragments, and molecules incorporating PTH fragments (e.g., PTH (1-34)-Fc).
 - PAF antagonists.

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- Keratinocyte growth factor (KGF), KGF-related molecules (e.g.,
 KGF-2), and KGF modulators.
 - COX-2 inhibitors, such as Celebrex[™] and Vioxx[™].
 - Prostaglandin analogs (e.g., E series prostaglandins).
 - Matrix metalloproteinase (MMP) modulators.
 - Nitric oxide synthase (NOS) modulators, including modulators of inducible NOS.
 - Modulators of glucocorticoid receptor.
 - Modulators of glutamate receptor.
 - Modulators of lipopolysaccharide (LPS) levels.
 - Anti-cancer agents, including inhibitors of oncogenes (e.g., fos, jun) and interferons.
 - Noradrenaline and modulators and mimetics thereof.

Pharmaceutical Compositions

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<u>In General</u>. The present invention also provides methods of using pharmaceutical compositions of the inventive compounds. Such pharmaceutical compositions may be for administration for injection, or for oral, pulmonary, nasal, transdermal or other forms of administration. In general, the invention encompasses pharmaceutical compositions comprising effective amounts of a compound of the invention together with pharmaceutically acceptable diluents, preservatives, solubilizers, emulsifiers, adjuvants and/or carriers. Such compositions include diluents of various buffer content (e.g., Tris-HCl, acetate, phosphate), pH and ionic strength; additives such as detergents and solubilizing agents (e.g., Tween 80, Polysorbate 80), anti-oxidants (e.g., ascorbic acid, sodium metabisulfite), preservatives (e.g., Thimersol, benzyl alcohol) and bulking substances (e.g., lactose, mannitol); incorporation of the material into particulate preparations of polymeric compounds such as polylactic acid, polyglycolic acid, etc. or into liposomes. Hyaluronic acid may also be used, and this may have the effect of promoting sustained duration in the circulation. Such compositions may influence the physical state, stability, rate of in vivo release, and rate of in vivo clearance of the present proteins and derivatives. See, e.g., Remington's Pharmaceutical Sciences, 18th Ed. (1990, Mack Publishing Co., Easton, PA 18042) pages 1435-1712 which are herein incorporated by reference in their entirety. The compositions may be prepared in liquid form, or may be in dried powder, such as lyophilized form. Implantable sustained release formulations are also contemplated, as are transdermal formulations.

Oral dosage forms. Contemplated for use herein are oral solid dosage forms, which are described generally in Chapter 89 of Remington's Pharmaceutical Sciences (1990), 18th Ed., Mack Publishing Co. Easton PA 18042, which is herein incorporated by reference in its entirety. Solid dosage forms include tablets, capsules, pills, troches or lozenges, cachets

or pellets. Also, liposomal or proteinoid encapsulation may be used to formulate the present compositions (as, for example, proteinoid microspheres reported in U.S. Patent No. 4,925,673). Liposomal encapsulation may be used and the liposomes may be derivatized with various polymers (e.g., U.S. Patent No. 5,013,556). A description of possible solid dosage forms for the therapeutic is given in Chapter 10 of Marshall, K., Modern Pharmaceutics (1979), edited by G. S. Banker and C. T. Rhodes, herein incorporated by reference in its entirety. In general, the formulation will include the inventive compound, and inert ingredients which allow for protection against the stomach environment, and release of the biologically active material in the intestine.

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Also specifically contemplated are oral dosage forms of the above inventive compounds. If necessary, the compounds may be chemically modified so that oral delivery is efficacious. Generally, the chemical modification contemplated is the attachment of at least one moiety to the compound molecule itself, where said moiety permits (a) inhibition of proteolysis; and (b) uptake into the blood stream from the stomach or intestine. Also desired is the increase in overall stability of the compound and increase in circulation time in the body. Moieties useful as covalently attached vehicles in this invention may also be used for this purpose. Examples of such moieties include: PEG, copolymers of ethylene glycol and propylene glycol, carboxymethyl cellulose, dextran, polyvinyl alcohol, polyvinyl pyrrolidone and polyproline. See, for example, Abuchowski and Davis, Soluble Polymer-Enzyme Adducts, Enzymes as Drugs (1981), Hocenberg and Roberts, eds., Wiley-Interscience, New York, NY,, pp. 367-83; Newmark, et al. (1982), J. Appl. Biochem. 4:185-9. Other polymers that could be used are poly-1,3-dioxolane and poly-1,3,6-tioxocane. Preferred for pharmaceutical usage, as indicated above, are PEG moieties.

For oral delivery dosage forms, it is also possible to use a salt of a modified aliphatic amino acid, such as sodium N-(8-[2-hydroxybenzoyl] amino) caprylate (SNAC), as a carrier to enhance absorption of the therapeutic compounds of this invention. The clinical efficacy of a heparin formulation using SNAC has been demonstrated in a Phase II trial conducted by Emisphere Technologies. See US Patent No. 5,792,451, "Oral drug delivery composition and methods".

The compounds of this invention can be included in the formulation as fine multiparticulates in the form of granules or pellets of particle size about 1 mm. The formulation of the material for capsule administration could also be as a powder, lightly compressed plugs or even as tablets. The therapeutic could be prepared by compression.

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Colorants and flavoring agents may all be included. For example, the protein (or derivative) may be formulated (such as by liposome or microsphere encapsulation) and then further contained within an edible product, such as a refrigerated beverage containing colorants and flavoring agents.

One may dilute or increase the volume of the compound of the invention with an inert material. These diluents could include carbohydrates, especially mannitol, α -lactose, anhydrous lactose, cellulose, sucrose, modified dextrans and starch. Certain inorganic salts may also be used as fillers including calcium triphosphate, magnesium carbonate and sodium chloride. Some commercially available diluents are Fast-Flo, Emdex, STA-Rx 1500, Emcompress and Avicell.

Disintegrants may be included in the formulation of the therapeutic into a solid dosage form. Materials used as disintegrants include but are not limited to starch including the commercial disintegrant based on starch, Explotab. Sodium starch glycolate, Amberlite, sodium carboxymethylcellulose, ultramylopectin, sodium alginate, gelatin, orange

peel, acid carboxymethyl cellulose, natural sponge and bentonite may all be used. Another form of the disintegrants are the insoluble cationic exchange resins. Powdered gums may be used as disintegrants and as binders and these can include powdered gums such as agar, Karaya or tragacanth. Alginic acid and its sodium salt are also useful as disintegrants.

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Binders may be used to hold the therapeutic agent together to form a hard tablet and include materials from natural products such as acacia, tragacanth, starch and gelatin. Others include methyl cellulose (MC), ethyl cellulose (EC) and carboxymethyl cellulose (CMC). Polyvinyl pyrrolidone (PVP) and hydroxypropylmethyl cellulose (HPMC) could both be used in alcoholic solutions to granulate the therapeutic.

An antifrictional agent may be included in the formulation of the therapeutic to prevent sticking during the formulation process. Lubricants may be used as a layer between the therapeutic and the die wall, and these can include but are not limited to; stearic acid including its magnesium and calcium salts, polytetrafluoroethylene (PTFE), liquid paraffin, vegetable oils and waxes. Soluble lubricants may also be used such as sodium lauryl sulfate, magnesium lauryl sulfate, polyethylene glycol of various molecular weights, Carbowax 4000 and 6000.

Glidants that might improve the flow properties of the drug during formulation and to aid rearrangement during compression might be added. The glidants may include starch, talc, pyrogenic silica and hydrated silicoaluminate.

To aid dissolution of the compound of this invention into the aqueous environment a surfactant might be added as a wetting agent. Surfactants may include anionic detergents such as sodium lauryl sulfate, dioctyl sodium sulfosuccinate and dioctyl sodium sulfonate. Cationic detergents might be used and could include benzalkonium chloride or

benzethonium chloride. The list of potential nonionic detergents that could be included in the formulation as surfactants are lauromacrogol 400, polyoxyl 40 stearate, polyoxyethylene hydrogenated castor oil 10, 50 and 60, glycerol monostearate, polysorbate 40, 60, 65 and 80, sucrose fatty acid ester, methyl cellulose and carboxymethyl cellulose. These surfactants could be present in the formulation of the protein or derivative either alone or as a mixture in different ratios.

Additives may also be included in the formulation to enhance uptake of the compound. Additives potentially having this property are for instance the fatty acids oleic acid, linoleic acid and linolenic acid.

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Controlled release formulation may be desirable. The compound of this invention could be incorporated into an inert matrix which permits release by either diffusion or leaching mechanisms; e.g., gums. Slowly degenerating matrices may also be incorporated into the formulation, e.g., alginates, polysaccharides. Another form of a controlled release of the compounds of this invention is by a method based on the Oros therapeutic system (Alza Corp.), i.e., the drug is enclosed in a semipermeable membrane which allows water to enter and push drug out through a single small opening due to osmotic effects. Some enteric coatings also have a delayed release effect.

Other coatings may be used for the formulation. These include a variety of sugars which could be applied in a coating pan. The therapeutic agent could also be given in a film coated tablet and the materials used in this instance are divided into 2 groups. The first are the nonenteric materials and include methyl cellulose, ethyl cellulose, hydroxyethyl cellulose, methylhydroxy-ethyl cellulose, hydroxypropyl cellulose, hydroxypropyl-methyl cellulose, sodium carboxy-methyl cellulose, providone and the polyethylene glycols. The second group consists of the enteric materials that are commonly esters of phthalic acid.

A mix of materials might be used to provide the optimum film coating. Film coating may be carried out in a pan coater or in a fluidized bed or by compression coating.

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Pulmonary delivery forms. Also contemplated herein is pulmonary delivery of the present protein (or derivatives thereof). The protein (or derivative) is delivered to the lungs of a mammal while inhaling and traverses across the lung epithelial lining to the blood stream. (Other reports of this include Adjei et al., Pharma. Res. (1990) 7: 565-9; Adjei et al. (1990), Internatl. J. Pharmaceutics 63: 135-44 (leuprolide acetate); Braquet et al. (1989), J. Cardiovasc. Pharmacol. 13 (suppl.5): s.143-146 (endothelin-1); Hubbard et al. (1989), Annals Int. Med. 3: 206-12 (α1-antitrypsin); Smith et al. (1989), J. Clin. Invest. 84: 1145-6 (α1-proteinase); Oswein et al. (March 1990), "Aerosolization of Proteins", Proc. Symp. Resp. Drug Delivery II, Keystone, Colorado (recombinant human growth hormone); Debs et al. (1988), J. Immunol. 140: 3482-8 (interferon-γ and tumor necrosis factor α) and Platz et al., U.S. Patent No. 5,284,656 (granulocyte colony stimulating factor).

Contemplated for use in the practice of this invention are a wide range of mechanical devices designed for pulmonary delivery of therapeutic products, including but not limited to nebulizers, metered dose inhalers, and powder inhalers, all of which are familiar to those skilled in the art. Some specific examples of commercially available devices suitable for the practice of this invention are the Ultravent nebulizer, manufactured by Mallinckrodt, Inc., St. Louis, Missouri; the Acorn II nebulizer, manufactured by Marquest Medical Products, Englewood, Colorado; the Ventolin metered dose inhaler, manufactured by Glaxo Inc., Research Triangle Park, North Carolina; and the Spinhaler powder inhaler, manufactured by Fisons Corp., Bedford, Massachusetts.

All such devices require the use of formulations suitable for the dispensing of the inventive compound. Typically, each formulation is specific to the type of device employed and may involve the use of an appropriate propellant material, in addition to diluents, adjuvants and/or carriers useful in therapy.

The inventive compound should most advantageously be prepared in particulate form with an average particle size of less than 10 μm (or microns), most preferably 0.5 to 5 μm , for most effective delivery to the distal lung.

Pharmaceutically acceptable carriers include carbohydrates such as trehalose, mannitol, xylitol, sucrose, lactose, and sorbitol. Other ingredients for use in formulations may include DPPC, DOPE, DSPC and DOPC. Natural or synthetic surfactants may be used. PEG may be used (even apart from its use in derivatizing the protein or analog).

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Dextrans, such as cyclodextran, may be used. Bile salts and other related enhancers may be used. Cellulose and cellulose derivatives may be used. Amino acids may be used, such as use in a buffer formulation.

Also, the use of liposomes, microcapsules or microspheres, inclusion complexes, or other types of carriers is contemplated.

Formulations suitable for use with a nebulizer, either jet or ultrasonic, will typically comprise the inventive compound dissolved in water at a concentration of about 0.1 to 25 mg of biologically active protein per mL of solution. The formulation may also include a buffer and a simple sugar (e.g., for protein stabilization and regulation of osmotic pressure). The nebulizer formulation may also contain a surfactant, to reduce or prevent surface induced aggregation of the protein caused by atomization of the solution in forming the aerosol.

Formulations for use with a metered-dose inhaler device will generally comprise a finely divided powder containing the inventive

compound suspended in a propellant with the aid of a surfactant. The propellant may be any conventional material employed for this purpose, such as a chlorofluorocarbon, a hydrochlorofluorocarbon, a hydrofluorocarbon, or a hydrocarbon, including trichlorofluoromethane, dichlorodifluoromethane, dichlorotetrafluoroethanol, and 1,1,1,2-tetrafluoroethane, or combinations thereof. Suitable surfactants include sorbitan trioleate and soya lecithin. Oleic acid may also be useful as a surfactant.

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Formulations for dispensing from a powder inhaler device will comprise a finely divided dry powder containing the inventive compound and may also include a bulking agent, such as lactose, sorbitol, sucrose, mannitol, trehalose, or xylitol in amounts which facilitate dispersal of the powder from the device, e.g., 50 to 90% by weight of the formulation.

<u>Nasal delivery forms</u>. Nasal delivery of the inventive compound is also contemplated. Nasal delivery allows the passage of the protein to the blood stream directly after administering the therapeutic product to the nose, without the necessity for deposition of the product in the lung. Formulations for nasal delivery include those with dextran or cyclodextran. Delivery via transport across other mucous membranes is also contemplated.

<u>Dosages</u>. The dosage regimen involved in a method for treating the above-described conditions will be determined by the attending physician, considering various factors which modify the action of drugs, e.g. the age, condition, body weight, sex and diet of the patient, the severity of any infection, time of administration and other clinical factors. Generally, the daily regimen should be in the range of 0.1-1000 micrograms of the inventive compound per kilogram of body weight, preferably 0.1-150 micrograms per kilogram.

Specific preferred embodiments

The inventors have determined preferred structures for the preferred peptides listed in Table 4 below. The symbol " Λ " may be any of the linkers described herein or may simply represent a normal peptide

5 bond (i.e., so that no linker is present). Tandem repeats and linkers are shown separated by dashes for clarity.

Table 4—Preferred embodiments

Sequence/structure	SEQ ID
-	NO:
LPGCKWDLLIKQWVCDPL-A-V1	44
V¹-∆- LPGCKWDLLIKQWVCDPL	45
LPGCKWDLLIKQWVCDPL -A-	46
LPGCKWDLLIKQWVCDPL -A-V1	
V¹-Λ- LPGCKWDLLIKQWVCDPL -Λ-	47
LPGCKWDLLIKQWVCDPL	
SADCYFDILTKSDVCTSS-A-V ¹	48
V¹-Λ- SADCYFDILTKSDVCTSS	49
SADCYFDILTKSDVTSS-A- SADCYFDILTKSDVTSS	50
$-\Lambda$ - V^1	
V¹-A- SADCYFDILTKSDVTSS -A-	51
SADCYFDILTKSDVTSS	
FHDCKWDLLTKQWVCHGL-A-V ¹	52
V¹-A- FHDCKWDLLTKQWVCHGL	53
FHDCKWDLLTKQWVCHGL -A-	54
FHDCKWDLLTKQWVCHGL -A-V ¹	
V¹-Λ- FHDCKWDLLTKQWVCHGL -Λ-	55
FHDCKWDLLTKQWVCHGL	

"V" is an Fc domain as defined previously herein. In addition to those listed in Table 4, the inventors further contemplate heterodimers in which each strand of an Fc dimer is linked to a different peptide sequence; for example, wherein each Fc is linked to a different sequence selected from Table 2.

All of the compounds of this invention can be prepared by methods described in PCT appl. no. WO 99/25044.

The invention will now be further described by the following working examples, which are illustrative rather than limiting.

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EXAMPLE 1

Peptides

5 Peptide Phage Display

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1. Magnetic bead preparation

A. Fc-TALL-1 immobilization on magnetic beads

The recombinant Fc-TALL-1 protein was immobilized on the Protein A Dynabeads (Dynal) at a concentration of 8 µg of Fc-TALL-1 per 100 µl of the bead stock from the manufacturer. By drawing the beads to one side of a tube using a magnet and pipetting away the liquid, the beads were washed twice with the phosphate buffer saline (PBS) and resuspended in PBS. The Fc-TALL-1 protein was added to the washed beads at the above concentration and incubated with rotation for 1 hour at room temperature. The Fc-TALL-1 coated beads were then blocked by adding bovine serum albumin (BSA) to 1% final concentration and incubating overnight at 4 °C with rotation. The resulting Fc-TALL-1 coated beads were then washed twice with PBST (PBS with 0.05% Tween-20) before being subjected to the selection procedures.

B. Negative selection bead preparation

Additional beads were also prepared for negative selections. For each panning condition, 250 μ l of the bead stock from the manufacturer was subjected to the above procedure (section 1A) except that the incubation step with Fc-TALL-1 was omitted. In the last washing step, the beads were divided into five 50 μ l aliquots.

2. Selection of TALL-1 binding phage

A. Overall strategy

Two filamentous phage libraries, TN8-IX (5X10⁹ independent transformants) and TN12-I (1.4X10⁹ independent transformants) (Dyax Corp.), were used to select for TALL-1 binding phage. Each library was subjected to either pH 2 elution or 'bead elution' (section 2E). Therefore, four different panning conditions were carried out for the TALL-1 project (TN8-IX using the

pH2 elution method, TN8-IX using the bead elution method, TN12-I the using pH2 elution method, and TN12-I using the bead elution method). Three rounds of selection were performed for each condition.

B. Negative selection

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For each panning condition, about 100 random library equivalent ($5X10^{11}$ pfu for TN8-IX and $1.4X10^{11}$ pfu for TN12-I) was aliquoted from the library stock and diluted to 300 μ l with PBST. After the last washing liquid was drawn out from the first 50 μ l aliquot of the beads prepared for negative selections (section 1B), the 300 μ l diluted library stock was added to the beads. The resulting mixture was incubated for 10 minutes at room temperature with rotation. The phage supernatant was drawn out using the magnet and added to the second 50 μ l aliquot for another negative selection step. In this way, five negative selection steps were performed.

C. Selection using the Fc-TALL-1 protein coated beads

The phage supernatant after the last negative selection step (section 1B) was added to the Fc-TALL-1 coated beads after the last washing step (section 1A). This mixture was incubated with rotation for two hours at room temperature, allowing specific phage to bind to the target protein. After the supernatant is discarded, the beads were washed seven times with PBST.

D. pH2 elution of bound phage

After the last washing step (section 2C), the bound phages were eluted from the magnetic beads by adding 200 μ l of CBST (50 mM sodium citrate, 150 mM sodium chloride, 0.05% Tween-20, pH2). After 5 minute incubation at room temperature, the liquid containing the eluted phage were drawn out and transferred to another tube. The elution step was repeated again by adding 200 μ l of CBST and incubating for 5 minutes. The liquids from two elution steps were added together, and 100 μ l of 2 M Tris solution (pH 8) was added to neutralize the pH. 500 μ l of Min A Salts solution (60 mM K₂HPO₄, 33 mM KH₂PO₄, 7.6 mM (NH₄)SO₄, and 1.7 mM sodium citrate) was added to make the final volume to 1 ml.

E. 'bead elution'

After the final washing liquid was drawn out (section 2C), 1 ml of Min A salts solution was added to the beads. This bead mixture was added directly to a concentrated bacteria sample for infection (section 3A and 3B).

3. Amplification

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A. Preparation of plating cells

Fresh <u>E</u>. <u>Coli</u>. (XL-1 Blue MRF') culture was grown to $OD_{600} = 0.5$ in LB media containing 12.5 µg/ml tetracycline. For each panning condition, 20 ml of this culture was chilled on ice and centrifuged. The bacteria pellet was resuspended in 1 ml of the Min A Salts solution.

B. Transduction

Each mixture from different elution methods (section 2D and 2E) was added to a concentrated bacteria sample (section 3A) and incubated at 37 °C for 15 minutes. 2 ml of NZCYM media (2XNZCYM, 50 μ g/ml ampicillin) was added to each mixture and incubated at room temperature for 15 minutes. The resulting 4 ml solution was plated on a large NZCYM agar plate containing 50 μ g/ml ampicillin and incubated overnight at 37 °C.

C. Phage Harvesting

Each of the bacteria/phage mixture that was grown overnight on a large NZCYM agar plate (section 3B) was scraped off in 35 ml of LB media, and the agar plate was further rinsed with additional 35 ml of LB media. The resulting bacteria/phage mixture in LB media was centrifuged to pellet the bacteria away. 50 ml the of the phage supernatant was transferred to a fresh tube, and 12.5 ml of PEG solution (20% PEG8000, 3.5M ammonium acetate) was added and incubated on ice for 2 hours to precipitate phages. Precipitated phages were centrifuged down and resuspended in 6 ml of the phage resuspension buffer (250 mM NaCl, 100 mM Tris pH8, 1 mM EDTA). This phage solution was further purified by centrifuging away the remaining bacteria and precipitating the phage for the second time by adding 1.5 ml of the PEG solution. After a centrifugation step, the phage pellet was resuspended in 400 μl of PBS. This solution was subjected to a final centrifugation to rid of remaining bacteria debris. The resulting phage

preparation was titered by a standard plaque formation assay (Molecular Cloning, Maniatis et al 3rd Edition).

4. Two more rounds of selection and amplification.

In the second round, the amplified phage (10^{10} pfu) from the first round (section 3C) was used as the input phage to perform the selection and amplification steps (sections 2 and 3). The amplified phage (10^{10} pfu) from the 2^{nd} round in turn was used as the input phage to perform 3^{rd} round of selection and amplification (sections 2 and 3). After the elution steps (sections 2D and 2E) of the 3^{rd} round, a small fraction of the eluted phage was plated out as in the plaque formation assay (section 3C). Individual plaques were picked and placed into 96 well microtiter plates containing $100 \, \mu l$ of TE buffer in each well. These master plates were incubated in a 37 °C incubator for 1 hour to allow phages to elute into the TE buffer.

5. Clonal analysis (Phage ELISA and sequencing)

The phage clones were analyzed by phage ELISA and sequencing methods. The sequences were ranked based on the combined results from these two assays.

A. Phage ELISA

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An XL-1 Blue MRF' culture was grown until OD₆₀₀ reaches 0.5. 30 µl of this culture was aliquoted into each well of a 96 well microtiter plate. 10 µl of eluted phage (section 4) was added to each well and allowed to infect bacteria for 15 min at room temperature. 130 µl of LB media containing 12.5 µg/ml of tetracycline and 50 µg/ml of ampicillin was added to each well. The microtiter plate was then incubated overnight at 37 °C. The recombinant TALL-1 protein (1 µg/ml in PBS) was allowed to coat onto the 96-well Maxisorp plates (NUNC) overnight and 4°C. As a control, the recombinant Fc-Trail protein was coated onto a separate Maxisorp plate at the same molar concentration as the TALL-1 protein.

On the following day, liquids in the protein coated Maxisorp plates were

discarded, and each well was blocked with 300 ul of 2% BSA solution at 37 °C

for one hour. The BSA solution was discarded, and the wells were washed three times with the PBST solution. After the last washing step, 50 μ l of PBST was added to each well of the protein coated Maxisorp plates. Each of the 50 μ l overnight cultures in the 96 well microtiter plate was transferred to the corresponding wells of the TALL-1 coated plates as well as the control Fc-Trail coated plates. The 100 μ l mixtures in the two kinds of plates were incubated for 1 hour at room temperature. The liquid was discarded from the Maxisorp plates, and the wells were washed five times with PBST. The HRP-conjugated anti-M13 antibody (Pharmacia) was diluted to 1:7,500, and 100 μ l of the diluted solution was added to each well of the Maxisorp plates for 1 hour incubation at room temperature. The liquid was again discarded and the wells were washed seven times with PBST. 100 μ l of tetramethylbenzidine (TMB) substrate (Sigma) was added to each well for the color reaction to develop, and the reaction was stopped with 50 μ l of the 5 N H₂SO₄ solution. The OD₄₅₀ was read on a plate reader (Molecular Devices).

B. Sequencing of the phage clones.

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For each phage clone, the sequencing template was prepared by a PCR method. The following oligonucleotide pair was used to amplify about 500 nucleotide fragment:

primer #1 (5'-CGGCGCAACTATCGGTATCAAGCTG-3') (SEQ ID NO: 56) and primer #2 (5'-CATGTACCGTAACACTGAGTTTCGTC-3'). (SEQ ID NO: 57) The following mixture was prepared for each clone.

Reagents	volume (μL) / tube
dH ₂ O	26.25
50% glycerol	10
10B PCR Buffer (w/o MgCl ₂)	5
25 mM MgCl ₂	4
10 mM dNTP mix	1
100 μ <u>M</u> primer 1	0.25
100 μ <u>M</u> primer 2	0.25
Taq polymerase	0.25
Phage in TE (section 4)	3
Final reaction volume	50

The thermocycler (GeneAmp PCR System 9700, Applied Biosystems) was used to run the following program: 94°C for 5 min; [94°C for 30 sec, 55°C for 30 sec, 72°C for 45 sec.]x30 cycles; 72°C for 7 min; cool to 4°C. The PCR product was checked by running 5 µl of each PCR reaction on a 1% agarose gel. The PCR product in the remaining 45 µl from each reaction was cleaned up using the QIAquick Multiwell PCR Purification kit (Qiagen), following the manufacturer's protocol. The resulting product was then sequenced using the ABI 377 Sequencer (Perkin-Elmer) following the manufacturer recommended protocol.

6. Sequence ranking and consensus sequence determination

A. Sequence ranking

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The peptide sequences that were translated from variable nucleotide sequences (section 5B) were correlated to ELISA data. The clones that showed high OD_{450} in the TALL-1 coated wells and low OD_{450} in the Fc-Trail coated wells were considered more important. The sequences that occur multiple times were also considered important. Candidate sequences were chosen based on these criteria for further analysis as peptides or peptibodies. Five and nine candidate peptide sequences were selected from the TN8-IX and TN12-I libraries, respectively.

B. Consensus sequence determination

The majority of sequences selected from the TN12-I library contained a very conserved DBL motif. This motif was also observed in sequences selected from the TN8-IB library as well. Another motif, PFPWE (SEQ ID NO: 110) was also observed in sequences obtained from the TN8-IB library.

A consensus peptide, FHDCKWDLLTKQWVCHGL (SEQ ID NO: 58), was designed based on the DBL motif. Since peptides derived from the TN12-I library were the most active ones, the top 26 peptide sequences based on the above ranking criteria (section 5A) were aligned by the DBL motif. The underlined "core amino acid sequence" was obtained by determining the amino acid that occur the most in each position. The two cysteines adjacent to the core

sequences were fixed amino acids in the TN12-I library. The rest of the amino acid sequence in the consensus peptide is taken from one of the candidate peptides, TALL-1-12-10 (Table 2, SEQ ID NO: 37). The peptide and peptibody that was derived from this consensus sequence were most active in the B cell proliferation assay.

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EXAMPLE 2

Peptibodies

A set of 12 TALL-1 inhibitory peptibodies (Table 5) was constructed in 10 which a monomer of each peptide was fused in-frame to the Fc region of human IgG1. Each TALL-1 inhibitory peptibody was constructed by annealing the pairs of oligonucleotides shown in Table 6 to generate a duplex encoding the peptide and a linker comprised of 5 glycine residues and one valine residue as an NdeI to SalI fragment. These duplex molecules were ligated into a vector (pAMG21-RANK-Fc, described herein) containing the human Fc gene, also digested with 15 NdeI and SalI. The resulting ligation mixtures were transformed by electroporation into E. coli strain 2596 cells (GM221, described herein). Clones were screened for the ability to produce the recombinant protein product and to possess the gene fusion having the correct nucleotide sequence. A single such clone was selected for each of the peptibodies. The nucleotide and amino acid 20 sequences of the fusion proteins are shown in Figure 4A through 4F.

Table 5. Peptide sequences and oligonucleotides used to generate TALL-1 inhibitory peptibodies.

Peptibody	Peptibody SEQ ID NO	Peptide Sequence	Sense oligo- nucleotide	Antisense oligo-nucleotide
TALL-1-8-1-a	29	PGTCFPFPWECTHA	2517-24	2517-25
TALL-1-8-2-a	30	WGACWPFPWECFKE	2517-26	2517-27
TALL-1-8-4-a	31	VPFCDLLTKHCFEA	2517-28	2517-29
TALL-1-12-4-a	32	GSRCKYKWDVLTKQCFHH	2517-30	2517-31
TALL-1-12-3-a	33	LPGCKWDLLIKQWVCDPL	2517-32	2517-33
TALL-1-12-5-a	34	SADCYFDILTKSDVCTSS	2517-34	2517-35
TALL-1-12-8-a	35	SDDCMYDQLTRMFICSNL	2517-36	2517-37
TALL-1-12-9-a	36	DLNCKYDELTYKEWCQFN	2521-92	2521-93

TALL-1-12-10-a	37	FHDCKYDLLTRQMVCHGL	2521-94	2521-95
TALL-1-12-11-a	38	RNHCFWDHLLKQDICPSP	2521-96	2521-97
TALL-1-12-14-a	39	ANQCWWDSLTKKNVCEFF	2521-98	2521-99
TALL-1-	58	FHDCKWDLLTKQWVCHGL	2551-48	2551-49
consensus				

Table 5B TALL-1 inhibitory peptibodies.

Peptibody	Peptibody SEQ ID NO			Sequence	
TALL-1-8-	111)		VDKTHTCPPC	
1-a		VFLFPPKPKD		TCVVVDVSHE	· ·
		DGVEVHNAKT	~	YRVVSVLTVL	
	'			KGQPREPQVY	
		KNQVSLTCLV		~	
		SDGSFFLYSK SLSLSPGK	LTVDKSRWQQ	GNVFSCSVMH	EALHNHYTQK
TALL-1-8-	112	MWGACWPFPW	ECFKEGGGGG	VDKTHTCPPC	PAPELLGGPS
2-a		VFLFPPKPKD	TLMISRTPEV	TCVVVDVSHE	DPEVKFNWYV
		DGVEVHNAKT	KPREEQYNST	YRVVSVLTVL	HQDWLNGKEY
		KCKVSNKALP	APIEKTISKA	KGQPREPQVY	TLPPSRDELT
		KNQVSLTCLV	KGFYPSDIAV	EWESNGQPEN	NYKTTPPVLD
		SDGSFFLYSK	LTVDKSRWQQ	GNVFSCSVMH	EALHNHYTQK
		SLSLSPGK			
TALL-1-8-	113	MVPFCDLLTK	HCFEAGGGGG	VDKTHTCPPC	PAPELLGGPS
4-a		VFLFPPKPKD	TLMISRTPEV	TCVVVDVSHE	DPEVKFNWYV
-		DGVEVHNAKT	KPREEQYNST	YRVVSVLTVL	HQDWLNGKEY
		KCKVSNKALP	APIEKTISKA	KGQPREPQVY	TLPPSRDELT
		KNQVSLTCLV	KGFYPSDIAV	EWESNGQPEN	NYKTTPPVLD
1		SDGSFFLYSK	LTVDKSRWQQ	GNVFSCSVMH	EALHNHYTQK
		SLSLSPGK			
TALL-1-12-	114	MGSRCKYKWD	VLTKQCFHHG	GGGGVDKTHT	CPPCPAPELL
4-a		GGPSVFLFPP	KPKDTLMISR	TPEVTCVVVD	VSHEDPEVKF
		NWYVDGVEVH	NAKTKPREEQ	YNSTYRVVSV	LTVLHQDWLN
		GKEYKCKVSN	KALPAPIEKT	ISKAKGQPRE	PQVYTLPPSR
		DELTKNQVSL	TCLVKGFYPS	DIAVEWESNG	QPENNYKTTP
		PVLDSDGSFF	LYSKLTVDKS	RWQQGNVFSC	SVMHEALHNH
		YTQKSLSLSP			
TALL-1-12-	115	MLPGCKWDLL	IKQWVCDPLG		CPPCPAPELL
3-a		GGPSVFLFPP	KPKDTLMISR	TPEVTCVVVD	VSHEDPEVKF
		NWYVDGVEVH	NAKTKPREEQ	YNSTYRVVSV	LTVLHQDWLN
		GKEYKCKVSN		ISKAKGQPRE	PQVYTLPPSR
		DELTKNQVSL			QPENNYKTTP
į		1	LYSKLTVDKS	RWQQGNVFSC	SVMHEALHNH
		YTQKSLSLSP			
TALL-1-12-	116	MSADCYFDIL	TKSDVCTSSG		CPPCPAPELL
5-a				TPEVTCVVVD	
		NWYVDGVEVH	NAKTKPREEQ	YNSTYRVVSV	LTVLHQDWLN
		GKEYKCKVSN		ISKAKGQPRE	PQVYTLPPSR
		DELTKNQVSL		DIAVEWESNG	QPENNYKTTP
		PVLDSDGSFF		RWQQGNVFSC	SVMHEALHNH
		YTQKSLSLSP			
TALL-1-12-	117	MSDDCMYDQL			CPPCPAPELL
8-a		GGPSVFLFPP			VSHEDPEVKF
			~	YNSTYRVVSV	~ · · - ·
		GKEYKCKVSN		ISKAKGQPRE	PQVYTLPPSR
		DELTKNQVSL	TCLVKGFYPS	DIAVEWESNG	QPENNYKTTP

	г	DIT DODGOTT	T TOTAL PRINTER	DE TOO CONTINUE	OTRETTO TITLE
			LYSKLTVDKS	RWQQGNVFSC	SVMHEALHNH
		YTQKSLSLSP			
TALL-1-12-	118	ł .	TYKEWCQFNG		
9-a		GGPSVFLFPP	KPKDTLMISR	TPEVTCVVVD	VSHEDPEVKF
		NWYVDGVEVH	NAKTKPREEQ	YNSTYRVVSV	LTVLHQDWLN
		GKEYKCKVSN	KALPAPIEKT	ISKAKGQPRE	PQVYTLPPSR
		DELTKNOVSL	TCLVKGFYPS	DIAVEWESNG	OPENNYKTTP
			LYSKLTVDKS		
		YTOKSLSLSP			
TALL-1-12-	119		TROMVCHGLG	GGGGVDKTHT	CPPCPAPELL
10-a			KPKDTLMISR		
10-a			NAKTKPREEO		
			KALPAPIEKT		~
		1			~
			TCLVKGFYPS		
			LYSKLTVDKS	RWQQGMVFSC	SVMHEALHNH
	100	YTQKSLSLSP		~~~~	
TALL-1-12-	120		LKQDICPSPG		
11-a		l '	KPKDTLMISR		
·			NAKTKPREEQ		
			KALPAPIEKT		
		DELTKNQVSL	TCLVKGFYPS	DIAVEWESNG	QPENNYKTTP
		PVLDSDGSFF	LYSKLTVDKS	RWQQGNVFSC	SVMHEALHNH
		YTQKSLSLSP	GK		
TALL-1-12-	121	MANOCWWDSL	TKKNVCEFFG	GGGGVDKTHT	CPPCPAPELL
14-a			KPKDTLMISR		
14-a			NAKTKPREEQ		
			KALPAPIEKT		
			TCLVKGFYPS		
			LYSKLTVDKS		
		YTQKSLSLSP		TWQQGWVLDC	SVEHILLALIHMI
TALL-1-	122		TKOWVCHGLG	CCCCIDVIII	CDDCDADETT
1	122				
consensus			KPKDTLMISR		
			NAKTKPREEQ		
			KALPAPIEKT		
	į		TCLVKGFYPS		
			LYSKLTVDKS	RWQQGNVFSC	SVMHEALHNH
		YTQKSLSLSP			
TALL-1 12-	123		IKQWVCDPLG		
3 tandem		HMLPGCKWDL	LIKQWVCDPL	GGGGGVDKTH	TCPPCPAPEL
dimer		LGGPSVFLFP	PKPKDTLMIS	RTPEVTCVVV	DVSHEDPEVK
		FNWYVDGVEV	HNAKTKPREE	QYNSTYRVVS	VLTVLHQDWL
		NGKEYKCKVS	NKALPAPIEK	TISKAKGQPR	EPQVYTLPPS
		i e	LTCLVKGFYP		
			FLYSKLTVDK		
		HYTQKSLSLS		~~	
TALL-1	124		TKQWVCHGLG	SGSATGGSGS	TASSGSGSAT
consensus		HMFHDCKWDL	LTKQWVCHGL	GGGGGVDKTH	TCPPCPAPEL
			PKPKDTLMIS		
tandem			HNAKTKPREE		
dimer		ľ			
			NKALPAPIEK	~	~
			LTCLVKGFYP		
			FLYSKLTVDK	SKWQQGNVFS	CSVMHEALHN
	<u> </u>	HYTQKSLSLS	rgk_		

Table 6. Sequences of oligonucleotides used in peptibody construction.

Oligo-	SEQ	Sequence
nucleotide	ID NO	
ID		
number		
number		
2517-24	71	TAT GCC GGG TAC TTG TTT CCC GTT CCC GTG GGA ATG CAC
0515 05		TCA CGC TGG TGG AGG CGG TGG GG
2517-25	72	TCG ACC CCA CCG CCT CCT GGA GCG TGA GTG CAT TCC CAC GGG AAG CCG AAA CAA GTA CCC GGC A
2517-26	73	TAT GTG GGG TGC TTG TTG GCC GTT CCC GTG GGA ATG TTT
2517-20	13	CAA AGA AGG TGG AGG CGG TGG GG
2517-27	74	TCG ACC CCA CCG CCT CCA CCT TCT TTG AAA CAT TCC
2527 27	/=	CACGGG AAC GGC CAA CAAGCA CCC CAC A
2517-28	75	TAT GGT TCC GTT CTG TGA CCT GCT GAC TAA ACA CTG TTT
	, ,	CGA AGC TGG TGG AGG CGG TGG GG
2517-29	76	TCG ACC CCA CCG CCT CCA CCA GCT TCG AAA CAG TGT TTA
		GTC AGC AGG TCA CAGAAC GGA ACC A
2517-30	77	TAT GGG TTC TCG TTG TAA ATA CAA ATG GGA CGT TCT GAC
		TAA ACA GTG TTT CCA CCA CGG TGG AGG CGG TGG GG
2517-31	78	TCG ACC CCA CCG CCT CCA CCG TGG TGG AAA CAC TGT TTA
		GTC AGA ACG TCC CAT TTG TAT TTA CAA CGA GAA CCC A
2517-32	79	TAT GCT GCC GGG TTG TAA ATG GGA CCT GCT GAT CAA ACA
		GTG GGT TTG TGA CCC GCT GGG TGG AGG CGG TGG GG
2517-33	80	TCG ACC CCA CCG CCT CCA CCC AGC GGG TCA CAA ACC CAC
		TGT TTG ATC AGC AGG TCC CAT TTA CAA CCC GGC AGC A
2517-34	81	TAT GTC TGC TGA CTG TTA CTT CGA CAT CCT GAC TAA ATC
		TGA CGT TTG TAC TTC TTG TGG AGG CGG TGG GG
2517-35	82	TCG ACC CCA CCG CCT CCA CCA GAA GAA GTA CAA ACG TCA
		GAT TTA GTC AGG ATG TCG AAG TAA CAG TCA GCA GAC A
2517-36	83	TAT GTC TGA CGA CTG TAT GTA CGA CCA GCT GAC TCG TAT
		GTT CAT CTG TTC TAA CCT GGG TGG AGG CGG TGG GG
2517-37	84	TCG ACC CCA CCG CCT CCA CCC AGG TTA GAA CAG ATG AAC
		ATA CGA GTC AGC TGG TCG TAC ATA CAG TCG TCA GAC A
2521-92	85	TAT GGA CCT GAA CTG TAA ATA CGA CGA ACT GAC TTA CAA
		AGA ATG GTG TCA GTT CAA CGG TGG AGG CGG TGG GG
25221-93	86	TCG ACC CCA CCG CCT CCA CCG TTG AAC TGA CAC CAT TCT
		TTG TAA GTC AGTTCG TCG TAT TTA CAG TTC AGG TCC A
2521-94	87	TAT GTT CCA CGA CTG TAA ATA CGA CCT GCT GAC TCG TCA
		GAT GGT TTG TCA CGG TCT GGG TGG AGG CGG TGG GG
2521-95	88	TCG ACC CCA CCG CCT CCA CCC AGA CCG TGA CAA ACC ATC
		TGA CGA GTC AGC AGG TCG TAT TTA CAG TCG TGG AAC A
2521-96	89	TAT GCG TAA CCA CTG TTT CTG GGA CCA CCT GCT GAA ACA

		GGA	CAT	CTG	TCC	GTC	TCC	GGG	TGG	AGG	CGG	TGG	GG	
2521-97	90	TCG	ACC	CCA	CCG	CCT	CCA	CCC	GGA	GAC	GGA	CAG	ATG	TCC
		TGT	TTC	AGC	AGG	TGG	TCC	CAG	AAA	CAG	TGG	TTA	CGC	A
2521-98	91	TAT	GGC	TAA	CCA	GTG	TTG	GTG	GGA	CTC	TCT	GCT	GAA	AAA
		AAA	CGT	TTG	TGA	ATT	CTT	CGG	TGG	AGG	CGG	TGG	GG	
2521-99	92	TCG	ACC	CCA	CCG	CCT	CCA	CCG	AAG	AAT	TCA	CAA	ACG	TTT
		TTT	TTC	AGC	AGA	GAG	TCC	CAC	CAA	CAC	TGG	TTA	GCC	Ą
2551-48	93	TAT	GTT	CCA	CGA	CTG	CAA	ATG	GGA	CCT	GCT	GAC	CAA	ACA
		GTG	GGT	TTG	CCA	CGG	TCT	GGG	TGG	AGG	CGG	TGG	GG	
2551-49	94	TCG	ACC	CCA	CCG	CCT	CCA	CCC	AGA	CCG	TGG	CAA	ACC	CAC
		TGT	TTG	GTC	AGC	AGG	TCC	CAT	TTG	CAG	TCG	TGG	AAC	A
	1	I.												

pAMG21-RANK-Fc vector

pAMG21. The expression plasmid pAMG21 (ATCC accession no. 98113) can be derived from the Amgen expression vector pCFM1656 (ATCC #69576) which in turn be derived from the Amgen expression vector system described in US Patent No. 4,710,473. The pCFM1656 plasmid can be derived from the described pCFM836 plasmid (U.S. Patent No. 4,710,473) by:

- destroying the two endogenous NdeI restriction sites by end filling with
 T4 polymerase enzyme followed by blunt end ligation;
- replacing the DNA sequence between the unique <u>Aat</u>II and <u>Cla</u>I restriction sites containing the synthetic P_L promoter with a similar fragment obtained from pCFM636 (patent No. 4,710,473) containing the P_L promoter (see SEQ ID NO: 95 below); and
 - substituting the small DNA sequence between the unique <u>ClaI</u> and <u>KpnI</u> restriction sites with the oligonucleotide having the sequence of SEQ ID NO: 96.

SEQ ID NO: 95:

<u>Aat</u>II

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- - $\hbox{-}AAAAAACATACAGATAACCATCTGCGGTGATAAATTATCTCTGGCGGTGTTGACATAAA-TTTTTTGTATGTCTATTGGTAGACCCCACACTATTTAATAGAGACCGCCACAACTGTATTT-$
- 25 -TACCACTGGCGGTGATACTGAGCACAT 3'
 -ATGGTGACCGCCACTATGACTCGTGTAGC 5
 Clai

SEQ ID NO: 96:

 $5' \quad \text{CGATTTGATTCTAGAAGGAGGAATAACATATGGTTAACGCGTTGGAATTCGGTAC} \\ 3' \quad$

3' TAAACTAAGATCTTCCTCCTTATTGTATACCAATTGCGCAACCTTAAGC 5' <u>Cla</u>I <u>Kpn</u>I

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The expression plasmid pAMG21 can then be derived from pCFM1656 by making a series of site-directed base changes by PCR overlapping oligonucleotide mutagenesis and DNA sequence substitutions. Starting with the <u>Bgl</u>II site (plasmid bp # 180) immediately 5' to the plasmid replication promoter $P_{\underline{copB}}$ and proceeding toward the plasmid replication genes, the base pair changes are as shown in Table 7 below.

Table 7—Base pair changes resulting in pAMG21

	pAMG21 bp #	bp in pCFM1656	bp changed to in pAMG21
15			
	# 204	T/A	C/G
	# 428	A/T	G/C
	# 509	G/C	A/T
	# 617		insert two G/C bp
20	# 679	G/C	T/A
	# 980	T/A	C/G
	# 994	G/C	A/T
	# 1004	A/T	C/G
	# 1007	C/G	T/A
25	# 1028	A/T	T/A
	# 1047	C/G	T/A
	# 1178	G/C	T/A
	# 1466	G/C	T/A
	# 2028	G/C	bp deletion
30	# 2187	C/G	T/A
	# 2480	A/T	T/A
	# 2499-2502	<u>AGTG</u>	<u>GTCA</u>
25		TCAC	CAGT
35	# 2642	TCCGAGC AGGCTCG	7 bp deletion
	# 3435	G/C	A/T
40	# 3446	G/C	A/T
	# 3643	A/T	T/A

The DNA sequence between the unique <u>Aat</u>II (position #4364 in pCFM1656) and <u>Sac</u>II (position #4585 in pCFM1656) restriction sites is substituted with the DNA sequence below (SEQ ID NO: 97):.

	[<u>AatII</u> sticky end] (position #4358 in pAM	G21) 5,	GCGTAACGT TGCACGCATTGCA	'ATGCATGGTCTCC- .TACGTACCAGAGG-
5	-CCATGCGAGAGTAGGGAACTG -GGTACGCTCTCATCCCTTGAC			
	-GGGCCTTTCGTTTTATCTGTT -CCCGGAAAGCAAAATAGACAA			
10	-CGGGAGCGGATTTGAACGTTG -GCCCTCGCCTAAACTTGCAAC			
15	-CATAAACTGCCAGGCATCAAA -GTATTTGACGGTCCGTAGTTT			
13			λа	tII
	-TTCTACAAACTCTTTTGTTTA -AAGATGTTTGAGAAAACAAAT		.CATTCAAATATGGA	.CGTCGTACTTAAC-
20	-TTTTAAAGTATGGGCAATCAA -AAAATTTCATACCCGTTAGTT			
25	-GGTTTGTTGTATTGAGTTTCA -CCAAACAACATAACTCAAAGT			
25	-TACAGCCTAATATTTTTGAAA -ATGTCGGATTATAAAAACTTT			
30	-ATTCTTTTTCTCTTTTTGGTTA -TAAGAAAAAGAGAAAACCAAT			
	-GATAATTATCAACTAGAGAAG -CTATTAATAGTTGATCTCTTC			
35	-AACTATCTATATAGTTGTCTT -TTGATAGATATATCAACAGAA			
	-TAGCAGTATGAATAGGGAAAC -ATCGTCATACTTATCCCTTTG			
40	-TTACATTTGGAGATTTTTTAT -AATGTAAACCTCTAAAAAATA			
45	-AATGATTGGAGTTAGAATAAT -TTACTAACCTCAATCTTATTA			
	-AATATTGCCTCCATTTTTTAG -TTATAACGGAGGTAAAAAATC			
50	-AATGAGGATAAATGATCGCGA -TTACTCCTATTTACTAGCGCT			
	-ATAAGCATTGATTAATATCAT -TATTCGTAACTAATTATAGTA			
55	AAGTGTCGTCGGCATTTATGT TTCACAGCAGCCGTAAATACA			
60	-GCAAGTTTTGCGTGTTATATA -CGTTCAAAACGCACAATATAT			
	-ATTGGATTTTTGTCACACTAT -TAACCTAAAAACAGTGTGATA			

```
-\mathtt{TAGGATCGTACAGGTTTACGCAAGAAAATGGTTTGTTATAGTCGATTAATCGATTTGATT-
     -ATCCTAGCATGTCCAAATGCGTTCTTTTACCAAACAATATCAGCTAATTAGCTAAACTAA-
     -\mathtt{CTAGATTTGTTTTAACTAATTAAAGGAGGAATAACATATGGTTAACGCGTTGGAATTCGA-
5
     -GATCTAAACAAAATTGATTAATTTCCTCCTTATTGTATACCAATTGCGCAACCTTAAGCT-
     -GCTCACTAGTGTCGACCTGCAGGGTACCATGGAAGCTTACTCGAGGATCCGCGGAAAGAA-
     -\texttt{CGAGTGATCACAGCTGGACGTCCCATGGTACCTTCGAATGAGCTCCTAGGCGCCTTTCTT}-
10
     -GAAGAAGAAGAAGCCCGAAAGGAAGCTGAGTTGGCTGCCACCGCTGAGCAATA-
     -\mathtt{CTTCTTCTTCTTCGGGCTTTCCTTCGACCGACGGCGGTGGCGACTCGTTAT} -
     -\texttt{ACTAGCATAACCCCTTGGGGCCTCTAAACGGGTCTTGAGGGGTTTTTTGCTGAAAGGAGG-}
15
     -\mathtt{TGATCGTATTGGGGAACCCCGGAGATTTGCCCAGAACTCCCCAAAAAACGACTTTCCTCC} -
    -AACCGCTCTTCACGCTCTTCACGC 3'
                                            [SacII sticky end]
    -TTGGCGAGAAGTGCGAGAAGTG 5'
                                         (position #5904 in pAMG21)
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During the ligation of the sticky ends of this substitution DNA sequence, the outside <u>AatII</u> and <u>SacII</u> sites are destroyed. There are unique <u>AatII</u> and <u>SacII</u> sites in the substituted DNA.

A gene encoding human RANK fused to the N-terminus of Fc was ligated into pAMG21 as an NdeI to BamHI fragment to generate Amgen Strain #4125. This construct was modified to insert a valine codon at the junction of RANK and Fc. The adjacent valine and aspartate codons create a unique SalI site. This allows for the fusion of peptides at the N-terminus of Fc3 between the unique NdeI and SalI sites. The RANK sequence is deleted upon insertion of a new NdeI-SalI fragment. The sequence of the vector is given in Figure 5A through 5M.

GM221 (Amgen #2596). The Amgen host strain #2596 is an <u>E. coli</u> K-12 strain derived from Amgen strain #393, which is a derivative of <u>E. coli</u> W1485, obtained from the <u>E. coli</u> Genetic Stock Center, Yale University, New Haven, Connecticut (CGSC strain 6159). It has been modified to contain both the temperature sensitive lambda repressor cI857s7 in the early <u>ebg</u> region and the lacI^Q repressor in the late <u>ebg</u> region (68 minutes). The presence of these two repressor genes allows the use of this host with a variety of expression systems, however both of these repressors are irrelevant to the expression from luxP_R. The untransformed host has no antibiotic resistances.

The ribosome binding site of the cI857s7 gene has been modified to include an enhanced RBS. It has been inserted into the ebg operon between

nucleotide position 1170 and 1411 as numbered in Genbank accession number M64441Gb_Ba with deletion of the intervening <u>ebg</u> sequence. The sequence of the insert is shown below with lower case letters representing the <u>ebg</u> sequences flanking the insert shown below (SEQ ID NO: 98):

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The construct was delivered to the chromosome using a recombinant phage called MMebg-cI857s7enhanced RBS #4 into F'tet/393. After recombination and resolution only the chromosomal insert described above remains in the cell. It was renamed F'tet/GM101. F'tet/GM101 was then modified by the delivery of a lacI^Q construct into the ebg operon between nucleotide position 2493 and 2937 as numbered in the Genbank accession number M64441Gb_Ba with the deletion of the intervening ebg sequence. The sequence of the insert is shown below with the lower case letters representing the ebg sequences flanking the insert (SEQ ID NO: 99) shown below:

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ggcggaaaccGACGTCCATCGAATGGTGCAAAACCTTTCGCGGTATGGCATGATAGCGCCCGGAAGA GAGTCAATTCAGGGTGGTGAATGTGAAACCAGTAACGTTATACGATGTCGCAGAGTATGCCGGT AAAAAGTCGAAGCGGCGATGGCGGAGCTGAATTACATTCCCAACCGCGTGGCACAACAACTGG CGGGCAAACAGTCGCTCCTGATTGGCGTTGCCACCTCCAGTCTGGCCCTGCACGCGCCGTCGCA AATTGTCGCGGCGATTAAATCTCGCGCCGATCAACTGGGTGCCAGCGTGGTGGTGCGATGGTA GAACGAAGCGCGTCGAAGCCTGTAAAGCGGCGGTGCACAATCTTCTCGCGCAACGCGTCAGTG TGTTCCGGCGTTATTTCTTGATGTCTCTGACCAGACACCCATCAACAGTATTATTTTCTCCCATGA AGACGGTACGCGACTGGGCGTGGAGCATCTGGTCGCATTGGGTCACCAGCAAATCGCGCTGTTA CAATCAAATTCAGCCGATAGCGGAACGGGAAGGCGACTGGAGTGCCATGTCCGGTTTTCAACAA ACCATGCAAATGCTGAATGAGGGCATCGTTCCCACTGCGATGCTGGTTGCCAACGATCAGATGG CGCTGGGCGCATTGCGGCCATTACCGAGTCCGGGCTGCGCGTTGGTGCGGATATCTCGGTAGT GGGATACGACGATACCGAAGACAGCTCATGTTATATCCCGCCGTTAACCACCATCAAACAGGAT TTTCGCCTGCTGGGGCAAACCAGCGTGGACCGCTTGCTGCAACTCTCTCAGGGCCAGGCGGTGA

 $AGGGCAATCAGCTGTTGCCCGTCTCACTGGTGAAAAGAAAAACCACCCTGGCGCCCAATACGCA\\ AACCGCCTCTCCCCGCGCGTTGGCCGATTCATTAATGCAGCTGGCACGACAGGTTTCCCGACTGG\\ AAAGCGGACAGTAAGGTACCATAGGATCCaggcacagga$

The construct was delivered to the chromosome using a recombinant phage called AGebg-LacIQ#5 into F'tet/GM101. After recombination and resolution only the chromosomal insert described above remains in the cell. It was renamed F'tet/GM221. The F'tet episome was cured from the strain using acridine orange at a concentration of 25 μ g/ml in LB. The cured strain was identified as tetracyline sensitive and was stored as GM221.

Expression in \underline{E} . \underline{coli} . Cultures of each of the pAMG21-Fc-fusion constructs in \underline{E} . \underline{coli} GM221 were grown at 37 °C in Luria Broth medium. Induction of gene product expression from the luxPR promoter was achieved following the addition of the synthetic autoinducer N-(3-oxohexanoyl)-DL-homoserine lactone to the culture media to a final concentration of 20 ng/ml. Cultures were incubated at 37 °C for a further 3 hours. After 3 hours, the bacterial cultures were examined by microscopy for the presence of inclusion bodies and were then collected by centrifugation. Refractile inclusion bodies were observed in induced cultures indicating that the Fc-fusions were most likely produced in the insoluble fraction in \underline{E} . \underline{coli} . Cell pellets were lysed directly by resuspension in Laemmli sample buffer containing 10% β -mercaptoethanol and were analyzed by SDS-PAGE. In each case, an intense Coomassie-stained band of the appropriate molecular weight was observed on an SDS-PAGE gel.

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EXAMPLE 3

TALL-1 peptibody inhibits TALL-1 mediated B cell proliferation

Mouse B lymphocytes were isolated from C57BL/6 spleens by negative selection. (MACS CD43 (Ly-48) Microbeads, Miltenyi Biotech, Auburn, CA). Purified (10⁵) B cells were cultured in MEM, 10% heat inactivated FCS, 5x10⁻⁵M 2-mercaptoethanol, 100 U/ml penicillin, 100 μg/ml streptomycin) in triplicate in 96-well flat bottom tissue culture plates with 10 ng/ml TALL-1 protein and 2 μg/ml of Goat F(ab')₂ anti-mouse IgM (Jackson ImmunoResearch Laboratory,

West Grove, Pennsylvania) with the indicated amount of recombinant TALL-1 peptibody for a period of 4 days at 37 °C, 5%CO2. Proliferation was measured by the uptake of radioactive ³[H] thymidine after an 18-hour incubation period.

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EXAMPLE 4

TALL-1 peptibody blocks TALL-1 binding to its receptors

Reacti-Gel 6x (Pierce) were pre-coated with human AGP3 (also known as TALL-1, Khare et al., Proc. Natl. Acad. Sci. 97:3370-3375, 2000) and blocked with BSA. 100 pM and 40 pM of AGP3 peptibody samples were incubated with indicated various concentrations of human AGP3 at room temperature for 8 hours before run through the human AGP3-coated beads. The amount of the bead-bound peptibody was quantified by fluorescent (Cy5) labeled goat antihuman-Fc antibody (Jackson Immuno Research). The binding signal is proportional to the concentration of free peptibody at binding equilibrium. Dissociation equilibrium constant (KD) was obtained from nonlinear regression of the competition curves using a dual-curve one-site homogeneous binding model (Kin Ex^{TM} software). KD is about 4 pM for AGP3 peptibody (SEQ ID NO: 123) binding with human AGP3 (Figure 9).

To determine if this AGP3 peptibody can neutralize murine AGP3 binding as well as human AGP3, a BIAcore neutralizing assay was utilized. All experiments were performed on a BIAcore 3000 at room temperature. Human TACI-Fc protein (Xia et al, <u>J. Exp. Med</u>. 192: 137-144, 2000) was immobilized to a B1 chip using 10 mM Acetate pH 4.0 to a level of 2900RU. A blank flow cell was used as a background control. Using a running buffer of PBS (without calcium or magnesium) containing 0.005% P20, 1 nM recombinant human AGP3 (in running buffer plus, 0.1 mg/ml BSA) was incubated without and with indicated various amount of AGP3 peptibody (x axis) before injected over the surface of

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was performed with 10 mM HCl pH2, twice for 30 seconds. Relative binding of both human and murine AGP3 at presence vs absence of AGP3 peptibody (SEQ ID NO: 123) was measured (y axis). Relative binding response was determined as (RU-RU blank/ RUo-RU blank). The AGP3 peptibody (SEQ ID NO: 123) inhibited both human and murine AGP3 binding to its receptor TACI (Figures 10A and 10B).

To examine if this AGP3 peptibody blocks AGP3 binding to all three receptors (TACI, BCMA and BAFFR), recombinant soluble receptor TACI, BCMA and BAFFR proteins were immobilized to CM5 chip. Using 10 mM acetate, pH4, human TACI-Fc was immobilized to 6300 RU, human BCMA-Fc to 5000 RU, and BAFFR-Fc to 6000 RU. 1 nM of recombinant human AGP3 (in running buffer containing 0.1 mg/ml BSA and 0.1 mg/ml Heparin) or 1 nM recombinant APRIL protein (Yu, et al., Nat. Immunol., 1:252-256, 2000) were incubated with indicated amount of AGP3 peptibody before injection over each receptor surface. Regeneration for the AGP3 experiment was done with 8 mM glycine, pH 1.5, for 1 minute, followed by 25 mM CAPS, pH 10.5, 1M NaCl for 1 minute. Regeneration for the APRIL experiment was performed with 8 mM glycine, pH 2, for one minute, followed by 25 mM CAPS, pH 10.5, 1 M NaCl for one minute. Relative binding of AGP3 or APRIL was measured. AGP3 peptibody (SEQ ID NO: 123) blocked AGP3 binding to all three receptors

(Figure 11A). AGP3 peptibody didn't affect APRIL binding to the receptors (Figure 11B).

EXAMPLE 5

AGP3 peptibody blocks AGP3 mediated B cell proliferation

Mouse B lymphocytes were isolated from C57BL/6 spleens by negative selection. (MACS CD43 (Ly-48) Microbeads, Miltenyi Biotech, Auburn, CA). Purified (105) B cells were cultured in minimal essential medium (MEM), 10% heat inactivated fetal calf serum (FCS), 5x10⁻⁵ M 2-mercaptoethanol, 100 U/ml penicillin, 100 µg/ml streptomycin) in triplicate in 96-well flat bottom tissue culture plates with 10 ng/ml AGP3 (TALL-1) protein and 2 µg/ml of Goat F(ab')2 anti-mouse IgM (Jackson ImmunoResearch Laboratory, West Grove, Pennsylvania) with the indicated amount of recombinant AGP3 peptibody (SEQ ID NO: 123) for a period of 4 days at 37 °C, 5% CO₂. Proliferation was measured by the uptake of radioactive ³[H] thymidine after an 18-hour incubation period.

EXAMPLE 6

AGP3 peptibody on AGP3-stimulated Ig production in mice

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Mice (Balb/c females of 9-14 weeks of age and 19-21 g of weight) were purchased from Charles River Laboratories, Wilmington, MA. Mice (n = 10) were treated i.p. with 1 mg/Kg of human AGP3 once a day for five consecutive days followed by 5 mg/Kg or 0.5 mg/Kg of AGP3 peptibody (SEQ ID NO: 123) or by saline or by 5 mg/Kg of human Fc. Other mice were left untreated. Mice were sacrificed on the sixth day to measure serum IgM and IgA, which were measured by ELISA. Briefly, plates were coated with capture antibodies specific for IgM or IgA (Southern Biotechnology Associates, Birmingham, AL), blocked,

and added with dilutions of standard (IgM from Calbiochem, San Diego, CA and IgA from Southern Biotechnology Associates) or test samples. Captured Ig were revealed using biotinylated antibodies specific for IgM or IgA (Southern Biotechnology Associates), neutravidin-conjugated peroxidase (Pierce,

Rockford, IL), and tetramethylbenzidine (TMB) microwell peroxidase substrate (KPL, Gaithersburg, MD). Optical densities were quantitated in a Thermomax ELISA reader (Molecular Devices, Menlo Park, CA).

Human AGP3-stimulated increase in serum levels of IgM and IgA was blocked by 5 mg/Kg of the anti-AGP3 peptibody (SEQ ID NO: 123) and not by 0.5 mg/Kg (Figures 12A and 12B).

EXAMPLE 7

AGP3 peptibody reduced spleen B cell number in mice

Mice (as above, n = 7) were treated i.p. for seven consecutive days with 5 mg/Kg or 1.5 mg/Kg or 0.5 mg/Kg of AGP3 peptibody (SEQ ID NO: 123) or with saline or with 5 mg/Kg of human Fc. Mice were sacrificed on the eighth day to count spleen B cell number. Spleens were collected in saline and gently disrupted by manual homogenization to yield a cell suspension. The total cell number was obtained with a H1E counter (Technicon, Tarrytown, NY). Percentages of B cells were derived by immunofluorescence double staining and flow cytometry using fluorescein isothiocyanate (FITC)-conjugated and phycoerythrin (PE)-conjugated Ab against CD3 and B220, respectively (PharMingen, San Diego, CA) and a FACScan analyser (Becton and Dickinson, Mountain View, CA). B cells were identified for being CD3-B220+. At all doses, the AGP3 peptibody (SEQ ID NO: 123) decreased spleen B cell number in a dose-response fashion (Figure 12A and 12B) (SEQ ID NO: 123).

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Table 8

AGP3 Pb Reduces B Cell Number in Normal Mice

n=7	dose (1/day×7)	spleen B cell (1x10e6)	SD	t test
saline		51.3	9.6	
Fc	5mg/Kg	45.5	7.1	
Peptibody	5mg/Kg	20.1	3.8	1.37856E-05
_ <u></u>	1.5mg/Kg	22.6	6.9	5.10194E-05
	0.5mg/Kg	25.8	3.6	0.000111409

EXAMPLE 8

AGP3 peptibody reduced arthritis severity in mouse CIA model

Eight to 12 week old DBA/1 mice (obtained from Jackson Laboratories, Bar Harbor, ME) were immunized with bovine collagen type II (bCII) (purchased from University of Utah), emulsified in complete Freunds adjuvant (Difco) intradermally at the base of tail. Each injection was 100 μl containing 100 μg of bCII. Mice were boosted 3 weeks after the initial immunization with bCII emulsified in incomplete Freunds adjuvant. Treatment was begun from the day of booster immunization for 4 weeks. Mice were examined for the development of arthritis. As described before (Khare et al., J. Immunol. 155: 3653-9, 1995), all four paws were individually scored from 0-3. Therefore arthritis severity could vary from 0 to 12 for each animal. AGP3 (SEQ ID NO: 123) peptibody treatment significantly reduced the severity of arthritic scores (Figure 13).

Serum samples were taken one week after final treatment (day 35) for the analysis of anti-collagen antibody level. High binding ELISA plates (Immulon, Nunc) were coated with 50 μ l of 4 μ g/ml solution of bovine CII in carbonate buffer and plated were kept in cold overnight in the refrigerator. Plates were

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washed three times with cold water. 75 μ l of blocking solution made up of PBS/.05% tween 20/1% BSA was used to block non-specific binding for an hour. Samples were diluted (in blocking buffer) in dilution plates at 1:25, 1:100, 1:400, and 1:1600 and 25 μ l of these samples were added to each well of the ELISA plate for a final dilution of 100, 400, 1600, and 6400 with a final volume of 100 μ l/well. After incubation at room temperature for 3 hours, plates were washed three times again. 100 μ l of secondary antibody diluted in blocking buffer (rat anti-mouse IgM, IgG2a, IgG2b, IgG1, IgG3-HRP) was added to each well and plates were incubated for at least 2 hours. Plates were washed four times. 100 μ l of TMB solution (Sigma) was added to each well and the reaction was stopped using 50 μ l of 25% sulfuric acid. Plates were read using an ELISA plate reader at 450 nm. OD was compared with a standard pool representing units/ml. AGP3 peptibody (SEQ ID NO: 123) treatment reduced serum anti-collagen II IgG1, IgG3, IgG2a, and IgG2b levels compared to PBS or Fc control treatment groups (Figure 14).

EXAMPLE 9

Treatment of AGP3 peptibody in NZB/NZW lupus mice

Five month old lupus prone NZBx NZBWF1 mice were treated i.p. 3X/week for 8 weeks with PBS or indicated doses of AGP3 peptibody or human Fc proteins. Prior to the treatment, animals were pre-screened for protein in the urine with Albustix reagents strips (Bayer AG). Mice having greater than 100 mg/dl of protein in the urine were not included in the study. Protein in the urine was evaluated monthly throughout the life of the experiment. AGP3 peptibody (SEQ ID NO: 123) treatment led to delay of proteinuria onset and improved survival (Figure 15A and 15B).

AGP3 peptibody treatment reduced B cell number in mice. Balb/c mice received 7 daily intraperitoneal injections of indicated amount of AGP3 peptibody (SEQ ID NO: 123) or human Fc protein. On day 8, spleens were collected, and subject to FACS analysis for B220+ B cells as set forth in Table 8.

The invention now being fully described, it will be apparent to one of ordinary skill in the art that many changes and modifications can be made thereto, without departing from the spirit and scope of the invention as set forth herein.

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THE CLAIMS DEFINING THE INVENTION ARE AS FOLLOWS:-

- 1. A TALL-1-binding composition of matter comprising an amino acid sequence Dz²Lz⁴, wherein z² is an amino acid residue and z⁴ is T or I, and wherein the composition of matter does not comprise FRKYDLLIHQRV or a fragment of TACI, BCMA, or BAFFR (SEQ ID NOS: 195, 196, and 197).
- 2. The composition of matter of Claim 1, wherein z^4 is T.
- 3. A TALL-1-binding composition of matter comprising an amino acid sequence Dz²LI, wherein z² is an amino acid residue.
- 4. The composition of matter of Claim 1 comprising an amino acid sequence of 10 the formula

a¹a²a³CDa⁶La⁸a⁹a¹⁰Ca¹²a¹³a¹⁴

(SEQ. ID. NO: 100)

wherein:

a¹, a² and a³ are each independently absent or amino acid residues;

a6 is an amino acid residue;

a⁸ is T or I;

a9 is a basic or hydrophobic residue;

a¹⁰ is an amino acid residue;

a¹² is a neutral hydrophobic residue; and

- a¹³ and a¹⁴ are each independently absent or amino acid residues.
 - 5. The composition of matter of Claim 4 wherein a⁸ is T and a⁹ is a basic residue.
 - 6. The composition of matter of Claim 4 wherein a⁹ is K and a¹² is F.
- 7. The composition of matter of Claim 1 comprising an amino acid sequence of the formula 25

 $b^1b^2b^3Cb^5b^6Db^8Lb^{10}b^{11}b^{12}b^{13}b^{14}Cb^{16}b^{17}b^{18}$

(SEQ. ID. NO: 104)

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wherein:
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b1 and b2 are each independently absent or amino acid residues;

b³ is an acidic or amide residue;

b⁵ is an amino acid residue;

b⁶ is an aromatic residue;

b8 is an amino acid residue;

b10 is T or I;

b11 is a basic residue;

b12 and b13 are each independently amino acid residues;

10 b¹⁴ is a neutral hydrophobic residue; and

 b^{16} , b^{17} , and b^{18} are each independently absent or amino acid residues.

8. The composition of matter of Claim 7 wherein:

b³ is D, Q, or E;

b6 is W or Y;

 b^{10} is T;

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b11 is K or R; and

b14 is V or L.

9. The composition of matter of Claim 1 comprising an amino acid sequence of the formula

 $c^1c^2c^3Cc^5Dc^7L\ c^9c^{10}c^{11}c^{12}c^{13}c^{14}Cc^{16}c^{17}c^{18}$

(SEQ. ID. NO: 105)

wherein:

 c^1 , c^2 , and c^3 are each independently absent or amino acid residues;

c5 is an amino acid residue;

25 c⁷ is an amino acid residue;

c9 is T or I;

c10 is a basic residue;

 c^{11} and c^{12} are each independently amino acid residues;

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c13 is a neutral hydrophobic residue;
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c14 is an amino acid residue;

c16 is an amino acid residue;

c17 is a neutral hydrophobic residue; and

c18 is an amino acid residue or is absent.

10. The composition of matter of Claim 9 wherein:

c9 is T;

c10 is K or R;

c13 is a I, L, or V; and

c¹⁷ is A or L. 10

> 11. The composition of matter of Claim 1 comprising an amino acid sequence of the formula

> > $d^1d^2d^3Cd^5d^6d^7WDd^{10}Ld^{12}d^{13}d^{14}Cd^{16}d^{17}d^{18}$

(SEQ. ID. NO: 106)

15 wherein:

d1, d2, and d3 are each independently absent or amino acid residues;

d⁵, d⁶, and d⁷ are each independently amino acid residues;

d10 is an amino acid residue;

d¹² is T or I:

d¹³ is an amino acid residue; 20

d14 is an amino acid residue; and

 d^{16} , d^{17} and d^{18} are each independently absent or amino acid residues.

12. The composition of matter of Claim 1 comprising an amino acid sequence of the formula

 $e^{1}e^{2}e^{3}Ce^{5}e^{6}e^{7}De^{9}L$ $e^{11}Ke^{13}Ce^{15}e^{16}e^{17}e^{18}$

(SEQ. ID. NO: 107)

wherein:

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e1, e2, and e3 are each independently absent or amino acid residues;

e⁵, e⁶, e⁷, e⁹, and e¹³ are each independently amino acid residues; e11 is T or I; and e15, e16, e17 and e18 are each independently absent or amino acid residues.

13. The composition of matter of Claim 1 comprising an amino acid sequence of the formula

> $f^1f^2f^3Kf^5Df^7Lf^9f^{10}Qf^{12}f^{13}f^{14}$ (SEQ ID NO: 109)

wherein:

 f^1 , f^2 , and f^3 are absent or are amino acid residues; 10

f⁵ is W or F;

f⁷ is an amino acid residue;

f9 is T or I;

f10 is K, R, or H;

f¹² is C, a neutral hydrophobic residue, or a basic residue; 15

f¹³ is C, a neutral hydrophobic residue or is absent; and

f14 is any amino acid residue or is absent;

provided that only one of f1, f2, and f3 may be C, and only one of f12,

 f^{13} , and f^{14} may be C.

- 14. The composition of matter of Claim 13, wherein f⁵ is W.
 - 15. The composition of matter of Claim 13, wherein f' is L.
 - 16. The composition of matter of Claim 13, wherein f9 is T.
 - 17. The composition of matter of Claim 13, wherein f^{10} is K.
- 18. The composition of matter of Claim 13, wherein f12 is C and one of f1, f2, and f³ is C. 25
 - 19. The composition of matter of Claim 13, wherein f^{13} is V.
 - 20. The composition of matter of Claim 13 comprising an amino acid sequence of the formula

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f1f2f3KWDf7Lf9KQf12f13f14

(SEQ ID NO: 125).

- 21. The composition of matter of Claim 20 comprising an amino acid sequence selected from the group consisting of SEQ ID NOS: 32, 33, 58, 60, 63, 66, 67, 69, 114, 115, 122, 123, 124, 147-150, 152-177, 179, 180, and 187.
- 22. The composition of matter of Claim 20 comprising an amino acid sequence of the formula

LPGCKWDLLIKQWVCDPL (SEQ ID NO: 33).

23. A composition of matter comprising an amino acid sequence of the formula 10

$$g^{1}g^{2}g^{3}Cg^{5}PFg^{8}Wg^{10}Cg^{12}g^{13}g^{14}$$

(SEQ. ID. NO: 101)

wherein:

g1, g2 and g3 are each independently absent or amino acid residues;

g⁵ is a neutral hydrophobic residue;

g8 is a neutral hydrophobic residue;

g10 is an acidic residue;

 $g^{\scriptscriptstyle 12}$ and $g^{\scriptscriptstyle 13}$ are each independently amino acid residues; and

g14 is absent or is an amino acid residue.

24. The composition of matter of Claim 23 wherein: 20

g² is G;

g⁵ is W;

g⁸ is P;

g10 is E; and

- 25 g¹³ is a basic residue.
 - 25. A composition of matter comprising an amino acid sequence of the formula

$h^1h^2h^3CWh^6h^7WGh^{10}Ch^{12}h^{13}h^{14}$

(SEQ. ID. NO: 102)

wherein:

h¹, h², and h³ are each independently absent or amino acid residues;

h6 is a hydrophobic residue; 5

h⁷ is a hydrophobic residue;

 h^{10} is an acidic or polar hydrophobic residue; and

 h^{12} , h^{13} , and h^{14} are each independently absent or amino acid residues.

26. The composition of matter of Claim 25 wherein:

h¹ is G; 10

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h6 is A;

h⁷ is a neutral hydrophobic residue; and

h10 is an acidic residue.

27. A composition of matter comprising an amino acid sequence of the formula

 $i^{1}i^{2}i^{3}Ci^{5}i^{6}i^{7}i^{8}i^{9}i^{10}Ci^{12}i^{13}i^{14}$

(SEQ. ID. NO: 103)

wherein:

i1 is absent or is an amino acid residue;

i² is a neutral hydrophobic residue;

i3 is an amino acid residue; 20

 i^5 , i^6 , i^7 , and i^8 are each independently amino acid residues;

i9 is an acidic residue;

i10 is an amino acid residue;

 $i^{12} \mbox{ and } i^{13} \mbox{ are each independently amino acid residues; and }$

i¹⁴ is a neutral hydrophobic residue.

28. The composition of matter of Claim 27 wherein:

i² is W; and

i¹⁴ is W.

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- 29. A TALL-1 binding composition of matter comprising an amino acid sequence of the formula PFPWE (SEQ ID NO: 110). :
- 30. The composition of matter of Claim 1 having the formula

 $(X^1)_a$ - V^1 - $(X^2)_b$ 5

and multimers thereof, wherein:

V¹ is a vehicle;

 X^1 and X^2 are each independently selected from -(L^1)_c- P^1 ,

-(
$$L^1$$
)c- P^1 -(L^2)d - P^2 , -(L^1)c- P^1 -(L^2)d- P^2 -(L^3)e- P^3 , and

 $-(L^1)_{c}-P^1-(L^2)_{d}-P^2-(L^3)_{e}-P^3-(L^4)_{f}-P^4$

one or more of P1, P2, P3, and P4 each independently comprise Dz2Lz4;

- L¹, L², L³, and L⁴ are each independently linkers; and
- a, b, c, d, e, and f are each independently 0 or 1, provided that at least one of a and b is 1;
- and wherein the composition of matter does not comprise the sequence FRKYDLLIHQRV when one of X^1 and X^2 is $-(L^1)_c$ - P^1 and the other is absent.
- 31. The composition of matter of Claim 30 of the formula

 P^1 -(L^1)c- P^2 -(L^2)d.- V^1 . 20

32. The composition of matter of Claim 30 of the formula

$$V^1-(L^1)c-P^1-(L^2)d-P^2$$
.

- 33. The composition of matter of Claim 30, wherein V¹ is an Fc domain.
- 34. The composition of matter of Claim 30 wherein V¹ is an IgG Fc domain. 25
 - 35. The composition of matter of Claim 30 wherein V1 is an IgG1 Fc domain.
 - 36. The composition of matter of Claim 30 wherein V¹ comprises the sequence of SEQ ID NO: 2.

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and multimers thereof, wherein:

V¹ is a vehicle;

 X^1 and X^2 are each independently selected from -(L^1)_c- P^1 ,

$$-(L^1)_c-P^1-(L^2)_d-P^2$$
, $-(L^1)_c-P^1-(L^2)_d-P^2-(L^3)_e-P^3$, and

$$-(L^1)_c-P^1-(L^2)_d-P^2-(L^3)_e-P^3-(L^4)_f-P^4;$$

L¹, L², L³, and L⁴ are each independently linkers; and

a, b, c, d, e, and f are each independently 0 or 1, provided that at least one of

a and b is 1

one or more of P1, P2, P3, and P4 each independently comprises a sequence

selected from:

a¹a²a³CDa6La8a9a¹0Ca¹²a¹³a¹4 (SEQ. ID. NO: 100)

 $b^{1}b^{2}b^{3}Cb^{5}b^{6}Db^{8}Lb^{10}b^{11}b^{12}b^{13}b^{14}Cb^{16}b^{17}b^{18} \ (SEQ. \ ID. \ NO: \ 104)$

 $c^{1}c^{2}c^{3}Cc^{5}Dc^{7}Lc^{9}c^{10}c^{11}c^{12}c^{13}c^{14}Cc^{16}c^{17}c^{18}$ (SEQ. ID. NO: 105)

 $d^1d^2d^3Cd^5d^6d^7WDd^{10}Ld^{12}d^{13}d^{14}Cd^{16}d^{17}d^{18}$ (SEQ. ID. NO: 106)

 $e^{1}e^{2}e^{3}Ce^{5}e^{6}e^{7}De^{9}Le^{11}Ke^{13}Ce^{15}e^{16}e^{17}e^{18} \, (SEQ. \, ID. \, NO: \, 107)$

 $f^1f^2f^3Kf^5Df^7Lf^9f^{10}Qf^{12}f^{13}f^{14}$ (SEQ. ID. NO: 109)

 $g^{1}g^{2}g^{3}Cg^{5}PFg^{8}Wg^{10}Cg^{12}g^{13}g^{14} \ (SEQ \ ID \ NO: \ 101),$

 $h^{\scriptscriptstyle 1}h^{\scriptscriptstyle 2}h^{\scriptscriptstyle 3}CWh^{\scriptscriptstyle 6}h^{\scriptscriptstyle 7}WGh^{\scriptscriptstyle 10}Ch^{\scriptscriptstyle 12}h^{\scriptscriptstyle 13}h^{\scriptscriptstyle 14}$ (SEQ ID NO: 102), and

i¹i²i³Ci⁵i⁶i⁷i⁸i⁹i¹⁰Ci¹²i¹³i¹⁴ (SEQ ID NO: 103)

wherein:

a¹, a², a³ are each independently absent or amino acid residues;

a6 is an amino acid residue;

a9 is a basic or hydrophobic residue;

a8 is threonyl or isoleucyl;

a¹⁰ is an amino acid residue;

a¹² is a neutral hydrophobic residue;

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                                  10
                                  15
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a¹³ and a¹⁴ are each independently absent or amino acid residues; b1 and b2 are each independently absent or amino acid residues; b³ is an acidic or amide residue; b⁵ is an amino acid residue; b6 is an aromatic residue; b8 is an amino acid residue; b^{10} is T or I; b¹¹ is a basic residue; b¹² and b¹³ are each independently amino acid residues; b¹⁴ is a neutral hydrophobic residue; b^{16} , b^{17} , and b^{18} are each independently absent or amino acid residues; c1, c2, and c3 are each independently absent or amino acid residues; c⁵ is an amino acid residue; c⁷ is an amino acid residue; c9 is T or I; c10 is a basic residue; c^{11} and c^{12} are each independently amino acid residues; c13 is a neutral hydrophobic residue; c14 is an amino acid residue;

20 c16 is an amino acid residue;

c17 is a neutral hydrophobic residue; and

c18 is an amino acid residue or is absent;

d¹, d², and d³ are each independently absent or amino acid residues;

d⁵, d⁶, and d⁷ are each independently amino acid residues;

d10 is an amino acid residue;

d12 is T or I;

d13 is an amino acid residue;

d14 is an amino acid residue; and d16, d17, and d18 are each independently absent or amino acid residues; e¹, e², and e³ are each independently absent or amino acid residues; e⁵, e⁶, e⁷, e⁹, and e¹³ are each independently amino acid residues; e11 is T or I; and 5 e¹⁵, e¹⁶, e¹⁷ and e¹⁸ are each independently absent or amino acid residues; f¹, f², and f³ are absent or are amino acid residues; f5 is W or F; f⁷ is an amino acid residue; 10 f⁹ is T or I; f¹⁰ is K, R, or H; f¹² is C, a neutral hydrophobic residue, or a basic residue; f¹³ is C, a neutral hydrophobic residue or is absent; and f¹⁴ is any amino acid residue or is absent; 15 provided that only one of f¹, f², and f³ may be C, and only one of f¹², f¹³, and f¹⁴ may be C; g1, g2 and g3 are each independently absent or amino acid residues; g⁵ is a neutral hydrophobic residue; g8 is a neutral hydrophobic residue; g10 is an acidic residue; 20 g12 and g13 are each independently amino acid residues; and g14 is absent or is an amino acid residue; h¹, h², and h³ are each independently absent or amino acid residues; h⁶ is a hydrophobic residue; 25 h⁷ is a hydrophobic residue; h¹⁰ is an acidic or polar hydrophobic residue; and

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h<sup>12</sup>, h<sup>13</sup>, and h<sup>14</sup> are each independently absent or amino acid residues;
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i1 is absent or is an amino acid residue;

i² is a neutral hydrophobic residue;

i³ is an amino acid residue;

i⁵, i⁶, i⁷, and i⁸ are each independently amino acid residues; 5

i9 is an acidic residue;

i¹⁰ is an amino acid residue;

i¹² and i¹³ are each independently amino acid residues; and

i¹⁴ is a neutral hydrophobic residue.

38. The composition of matter of claim 37, wherein:

a9 is a basic residue;

 b^3 is D, Q, or E;

b6 is W or Y;

b11 is K or R;

b14 is V or L; 15

c10 is K or R;

 c^{13} is a I, L, or V;

c17 is A or L;

f⁵ is W;

f⁷ is L; 20

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f10 is K; and

 f^{13} is V.

39. The composition of matter of Claim 37, wherein one or more of P1, P2, P3, and P⁴ each independently comprises

 $f^1f^2f^3KWDf^7Lf^9KQf^{12}f^{13}f^{14}$

(SEQ ID NO: 125).

40. The composition of matter of Claim 39 of the formula

$$P^1-(L^1)_c-P^2-(L^2)_d-V^1$$
.

41. The composition of matter of Claim 39 of the formula $V^1-(L^1)_c-P^1-(L^2)_d-P^2$.

- 42. The composition of matter of Claim 39 having an amino acid sequence selected from SEQ ID NOS: 122, 123, and 124.
- 43. The composition of matter of Claim 40 wherein L^2 is greater than 5 amino acids.
 - 44. The composition of matter of Claim 43 wherein L² is selected from

GSGSATGGSGSTASSGSGSATx1x2

(SEO ID NO: 193)

and 10

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GSGSATGGSGSTASSGSGSATx1x2GSGSATGGSGSTASSGSGSATx3x4 (SEQ ID NO: 194)

wherein x^1 and x^3 are each independently basic or hydrophobic residues and x^2 and x^4 are each independently hydrophobic residues.

45. The composition of matter of Claim 41 wherein L2 is selected from 15

GSGSATGGSGSTASSGSGSATH

(SEQ ID NO: 59),

GSGSATGGSGSTASSGSGSATGM

(SEQ ID NO: 190)

GSGSATGGSGSTASSGSGSATGS

(SEQ ID NO: 191), and

GSGSATGGSGSTASSGSGSATHMGSGSATGGSGSTASSGSGSATHM (SEQ ID NO: 192).

- 46. The composition of matter of Claim 30 comprising a sequence selected from Table 2 (SEQ ID NOS: 29-39, 60-70, and 126-188).
- 47. The composition of matter of Claim 30 comprising a sequence selected from Table 4 (SEQ ID NOS: 44-55).
- 48. The composition of matter of Claim 46, wherein V¹ is an Fc domain.

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- 49. The composition of matter of Claim 46, wherein V¹ is an IgG Fc domain.
- 50. The composition of matter of Claim 46, wherein V¹ is an IgG1 Fc domain.
- 51. A DNA encoding a composition of matter of Claim 33.
- 52. An expression vector comprising the DNA of Claim 51.
- 53. A host cell comprising the expression vector of Claim 52.
 - 54. The cell of Claim 53, wherein the cell is an \underline{E} . coli cell.
 - 55. A method of treating a B-cell mediated autoimmune disease, which comprises administering a composition of matter of Claim 1.
 - 56. A method of treating a B-cell mediated autoimmune disease, which comprises administering a composition of matter of Claim 13.
 - 57. A method of treating lupus, which comprises administering a composition of matter of Claim 1.
 - 58. A method of treating lupus, which comprises administering a composition of matter of Claim 13.
- 59. A method of treating a B-cell mediated cancer, which comprises administering a composition of matter of Claim 1.
 - 60. A method of treating a B-cell mediated cancer, which comprises administering a composition of matter of Claim 13.
 - 61. A method of treating B-cell lymphoma, which comprises administering a composition of matter of Claim 1.
 - 62. A method of treating B-cell lymphoma, which comprises administering a composition of matter of Claim 13.
 - 63. Use of a composition of matter according to Claim 1 for the manufacture of a medicament for the treatment of B-cell mediated autoimmune disease.
- 64. Use of a composition of matter of Claim 13 in the manufacture of a medicament for the treatment of B-cell mediated autoimmune disease.
 - 65. Use of a composition of matter of Claim 1 in the manufacture of a medicament for the treatment of lupus.

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- 66. Use of a composition of matter of Claim 13 in the manufacture of a medicament for the treatment of lupus.
- 67. Use of a composition of matter of Claim 1 in the manufacture of a medicament for the treatment of a B-cell mediated cancer.
- 68. Use of a composition of matter according to Claim 13 in the manufacture of a medicament for the treatment of B-cell mediated cancer.
 - 69. Use of a composition of a matter of Claim 1 in the manufacture of a medicament for the treatment of B-cell lymphoma.
 - 70. Use of a composition of a matter of Claim 13 in the manufacture of a medicament for the treatment of B-cell lymphoma.
 - 71. A TALL-1 binding composition of matter, substantially as herein described with reference to any one or more of the examples but excluding comparative examples.
 - 72. A composition of matter comprising an amino acid sequence of the formula i¹i²i³Ci⁵i⁶i⁷i⁸i⁹i¹⁰Ci¹²i¹³i¹⁴

(SEQ. ID. NO: 103)

- substantially as herein described with reference to any one or more of the examples but excluding comparative examples.
- 73. A DNA encoding a composition of matter according to Claim 34, 20 substantially as herein described with reference to any one or more of the examples but excluding comparative examples.
 - 74. An expression vector comprising the DNA of Claim 51, substantially as herein described with reference to any one or more of the examples but excluding comparative examples.
 - 75. A host cell comprising the expression vector of Claim 52, substantially as herein described with reference to any one or more of the examples but excluding comparative examples.

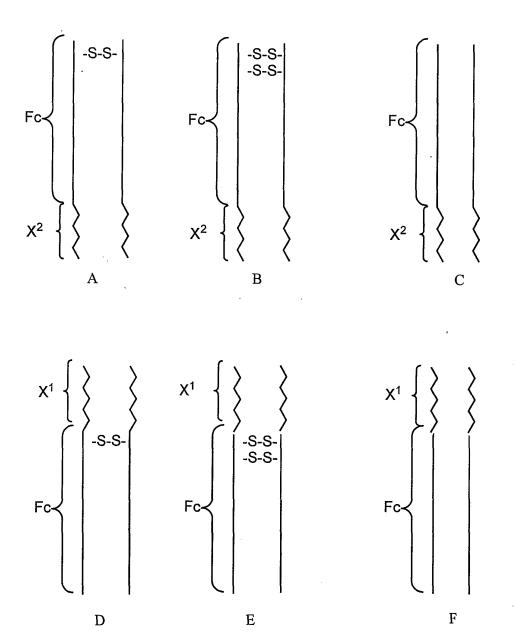
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- 76. A method of treating a B-cell mediated autoimmune disease, substantially as herein described with reference to any one or more of the examples but excluding comparative examples.
- 77. A method of treating lupus, substantially as herein described with reference to any one or more of the examples but excluding comparative examples.
- 78. A method of treating a B-cell mediated cancer, substantially as herein described with reference to any one or more of the examples but excluding comparative examples.
- 79. A method of treating B-cell lymphoma, substantially as herein described with reference to any one or more of the examples but excluding comparative examples.
 - 80. Use of a composition of matter of claim 1, substantially as herein described with reference to any one or more of the examples but excluding comparative examples.
- 81. Use of a composition of matter of claim 13, substantially as herein described 15 with reference to any one or more of the examples but excluding comparative examples.

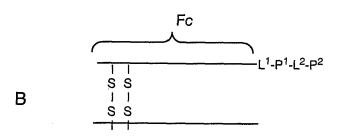
DATED this 31st day of August 2005 Shelston IP

Attorneys for: AMGEN INC. 20

FIG.1



FC FC L1-P1-L2-P2



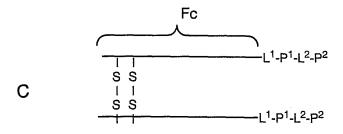


FIG. 3

		ATGG	CAA	AAC'	TCA	.CAC	ATG	TCC	ACC	TTG	TCC	AGC	TCC	GGA	ACT	CCT	GGG	GGG	ACC	GTCA	
	-	TACCI	GTT.	TTG	AGT	GTG	TAC	AGG	TGG	AAC	AGG	TCG	AGG	CCT	TGA	GGA	CCC	CCC	TGG	CAGT	- 60
a		M D	K	Т	H	T	С	P	P	С	P	A	P	E	L	L	G	G	p	s	-
	61	GTCTT	TOOT	CTT(ccc	CCC	AAA	ACC	CAA	GGA	CAC	CCT	CAT	GAT	CTC	CCG	GAC	CCC	TGA	GGTC	120
		CAGA	AGGA(GAA(GGG	GGG	TTT	TGG	GTI	CCT	GTG	GGA	GTA	CTA	GAG	GGC	CTG	GGG	ACT	CCAG	120
a		V F	L	F	P	P	K	P	K	D	T	L	M	I	S	R	T	P.	E	V	-
	121	ACATO	CGT	GGT(GGT	GGA	CGT +	GAG	CCA	CGA	AGA +	.CCC	TGA	GGT -+-	CAA	GTT 	CAA +	CTG	GTA	CGTG	180
		TGTAC																			
a		тС											_	-		F		W			-
	181	GACGG		-+-			+				+			-+-			+			+	240
		CTGCC																	GTC	GTGC	
a		D G	18																S	T	-
	241	TACCO		-+			+				+			-+-			+			+	300
a		ATGGC																			
a		Y R										_									-
	301	TTCAC		-+			+				+	~		-+-		- 	+			+	360
a		K C													K				K		-
									_			_	_	_		_	_	_			
		AAAGG	GCA	3CC(CCG	AGA	ACC.	ACA	GGT	GTA	CAC	CCT	GCC	ccc.	ATC	CCG	GGA'	TGA	GCT	GACC	
	361			-+			+				+		-	-+-			+			+	420
a ·	361	TTTCC		-+	GGC'	TCT	+ TGG	 TGT	CCA	 CAT	+ GTG	GGA	 CGG	-+- GGG	TAG	GGC	+	ACT		+	420
a ·		TTTCC K G AAGAA	CCAC	-+ CGG(P EGT(GGC' R CAG	TCT E CCT	TGG P GAC	TGT Q CTG	CCA V CCT	CAT Y GGT	+ GTG T CAA	GGA L AGG	CGG P	-+- GGG P CTA	TAG S TCC	GGC R CAG	CCT.	ACT E CAT	CGA	T CGTG	~
a ·		TTTCC	Q Q Q	P GGT(GGC' R	TCT E CCT	TGG P GAC +	TGT Q CTG	CCA V CCT	CAT Y GGT	+ GTG T CAA	GGA	CGG	-+- GGG P CTA	TAG S TCC	GGC R CAG	CGA	ACT E	CGA	TCGTG	~
a ·		TTTCC K G AAGAA TTCTT	Q Q Q	P EGTO	GGC' R CAGG	TCT E CCT GGA	TGG P GAC + CTG	TGT Q CTG GAC	CCA V CCT GGA	CAT Y GGT CCA	+ GTG T CAA + GTT	GGA L AGG	CGG P CTT GAA	-+- GGG P CTA -+- GAT	TAG S TCC AGG	GGC R CAG GTC	CGA	ACTO	CGA	T CGTG CGTG + GCAC	~
	421	TTTCC K G AAGAA TTCTT K N GAGTG	CCAC	P EGTO CCAO V	GGC' R CAGG GTCG	TCT E CCT GGA L	TGG P GAC CTG T	TGT Q CTG GAC GCC	CCA V CCT GGA L	CAT Y GGT CCA V GAA	+ GTG T CAA + GTT K	GGA L AGG TCC G	CTTGAA	-+- GGG P CTA -+- GAT Y	TAG S TCC AGG P	GGC R CAG GTC GTC	CGAGGCTG	ACTO E CATO	L CGC CGC A	T CGTG CGTG CGTG CCAC V GGAC	- 480 -
	421	TTTCC K G AAGAA TTCTT K N	CGT(CACCACCACCACCACCACCACCACCACCACCACCACCAC	P EGTO CCAO V EAGO	GGC' R CAGC GTCC	TCT E CCT GGA L	TGG P GAC CTG T GCA+	TGT Q CTG GAC GCC	CCA V CCT GGA L	CAT GGT CCA V GAA	+ GTG T CAA + GTT K CAA	GGA AGG TCC G CTA	CTTGAAG	-+- GGG P CTA GAT GAT	TAG S TCC AGG P CAC	GGC	CGAGGCTG	ACTO	CGA L CGC GCG A	T CGTG CAC V GGAC C+	- 480 -
	421	TTTCC K G AAGAA TTCTT K N GAGTG	CCTC	P EGTO	GGC' R CAGC GTCC S CAA'	TCT E CCT GGA L TGG	TGG P GAC CTG T GCA CGT	TGT Q CTG GAC C GCC GCC	CCA V CCT GGA L GGA CCT	CAT GGT CCA V GAA	T CAA + GTT K CAA CAA CAA GTT	GGA AGG TCC G CTA GAT	CTTGAAG	-+- GGG P CTA -+- GAT Y GAC	TAG S TCC AGG P CAC	GGC	D CGA GCT GCT D TCC AGG	ACTO	CGA	T CGTG CGTG CGTG CAC V GGAC CGGAC CGGAC	- 480 -
a	421	TTTCC K G AAGAA TTCTT K N GAGTG CTCAC	CCTC ECGGC	-+ P GGTC V V GAGC -+ CTCC S	GGC' R CAGG S CAA' S TTO	TCT E CCT GGA L TGG ACC G	TGG PGAC+ CTG TGCA+ CGT Q CCT	TGT Q CTG GAC C GCC CGG P	CCA V CCT GGA L GGA CCT CCT	CAT Y GGT CCA V GAA CTT N CAA	+ GTG T CAA + GTT K CAA N GCT	GGA L AGG GCTA GAT GAT	CGGG P CTT GAA F CAA GTT K CGT	-+- GGG P CTA -+- GAT: Y GAC+- CTG	TAG S TCC AGG P CAC GTG T CAA	GGCC GCCC GCCC GCCC	DCGAGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	E CATO	L CGC A A GCT CGA	T CGTG CAC V GGAC CCTG D GCAG	- 480 - 540 -
a	421	TTTCC K G AAGAA TTCTT K N GAGTG CTCAC	CCTC E	P GGTCC V SAGC	GGC' R CAGG GTCG S CAA' N CTTG	TCT E CCT GGA L TGGG ACC G	TGG PGAC CTG TGCA CGT Q CCT	TGT Q CTG GAC C GCC CGG P	CCA V CCT GGA L GGA CCT E	CAT Y GGT CCA V GAA CTT N CAA	+ GTG T CAA + GTT K CAA + GTT N	GGA L AGG TCC G CTA GAT	CGG P CTT GAA F CAA GTT K	-+- GGG P CTA -+- GAT: Y GAC+- CTG	TAG S TCC AGG P CAC GTG T CAA	GGCCCCGG	D CGAGGGGGCAGGGCAGGGCAGGGCAGGGCAGGGCAGGG	E CATO	CGA L CGCC A GCTC CGA L	T CGTG CGTG CGAC V CGGAC D CGGAC D GGAG	- 480 - 540 -
a	421	TTTCC K G AAGAA TTCTT K N GAGTG CTCAC E W TCCGA	CCTC E	-+ PEGTO V SAGO STOCK STOCK STOCK SAGO SAGO SAGO SAGO SAGO SAGO SAGO SAGO	GGC' R CAGG GTCG S CAA' N CTTG GAAG	E CCTGGACCGAACCGAACCGAACCGAAA	TGG PGAC+ CTG T GCA+ CGT Q CCT+ GGA	TGT Q CTG GAC C GCC CGG P CTA	CCA V CCT GGA L GGA CCT E CAG	CAT Y GGT CCA V GAA CTT N CAA GTT	+ GTG T CAA + GTT K CAA GTT N GCT CGA	GGA L AGG TCC G CTA GAT	CGG P CTT GAA F CAA GTT K CGT GCA	-+- GGGG P CTA -+- GAT: Y GAC+- CTG T GGA	TAG S TCC AGG P CACC GTG T CAAC	GGCCCGGG	D CGAGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	E CATO	CGA L CGC GCG A GCT CGA L CGT	TCTGG TCGTGCAC VGGAC CCTG DGGAG	- 480 - 540 -
a	421 481 541	TTTCC K G AAGAA TTCTT K N GAGTG CTCAC E W TCCGA	CGTC Q CCAC Q GGAC CCCTC E CCGCC G G CCGTC	-+ CGGG P GGTC V SAGC -+ CTCC S CTCC S SCTTCC S CTTCC S	GGC' R CAGG S CAA' STTC N CTTC GAA F	E CCTGGA C CTTGGA G ACC	TGG P GAC+ CTG T GCA+ CGT Q CCT GGA L CTC	TGT Q CTG GAC C GCC CGG P CTA GAT	CCA CCT GGA L GGA CCT E CAG GTC GTC	CAT Y GGT CCA V GAA CTT N CAA GTT K GCA	+ GTG T CAAA + GTT K CAAA + GTT N GCT CGAA L TGAA	GGA L AGGG G CTA GAT CAC GTG T GGGC	CGGG P CTT GAA F CAA GTT K CGT GCA V TCT	-+- GGG P CTA -+- GAT: Y GAC+- CTG T GGA CCT D	TAG S TCC AGG P CAC GTG T CAA GTT K CAA	GGCCCGG	D CGAGGG P CAGGG R CTAG	ACTO	CGA L CGCGC A GCCGC CGA L CGCC	T CTGG T CGTG GCAC V GGAC CCTG D GCAG CCGTC Q	- 480 - 540 - 600
a a	421 481 541	TTTCC K G AAGAA TTCTT K N GAGTG CTCAC E W TCCGA AGGCT S D GGGAA CCCTT	Q CCAC Q Q GGGAC CCCTC E CGGCC G CCGTC	-+ CGGG P GGTCC V SEAGC CTCC S CTTCC S CTTCC S CTTCC S S CTTCC S S CTTCC S S CTTCC S S S S	EGC' R CAGGTC S CAA' N CTTC GAA F CTC GAA GGTT GGAA GGTT GGAA GGGT GGAA GGTT GGAA GGGT GGAA GGGT GGAA GGGT GGAA GGGT GGAA GGGGT GGAA GGGGT GGAA GGGGGGGG	E CCTTGGGAACCCGAACCCGAACCCTTGGAACCCCGAACCCCGAACCCCGAACCCCGAACCCCGAACCCCGAACCCCAACCCCAACCCCAACCCCAACCCCAACCCCAACCCC	TGG P GAC' CTGG T GCA' CGT CGT CGT CGGA L CTC' GAG	TGT Q CTG GAC C GCC P CTA GAT Y CGT	CCA GGA CCTA	CAT Y GGTT CCA CTT K GCA GTT CCAA CTT CCAA CTT CCAA	H GTG T CAA H GTT K CAA GTT N GCT +-CGA L TGA ACT	GGAC GAC GAC GAC GAC GAC GCCC	CGGG P CTT GAA F CAA GTT K CGT GCA V TCT AGA	GAC-CCTC	TAG S TCC AGG P CAC GTG CAA GTT K CAA GTT K CAA GTT	GGCCCGG	D CGAGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	E CATO	CGA A GCTC CGA L CGCTC	T CTGG T CGTGGCAC V GGAC CCTG D GCAG CGCAG Q Q GAAG CCTTC	- 480 - 540 - 600
a	421 481 541	TTTCC K G AAGAA TTCTT K N GAGTG CTCAC E W TCCGA AGGCT S D GGGAA CCCTT	CGTC Q CCAC GGGTC Q GGGAC CCCTC E CGGCC G CCGTC	P GGTCCAC V SAGCCTCC S CTCCC S CTCCC S S CTTCC F F F F F F F F F F F F F F F F F	GGC' R CAGGTC S CAA' T CTTC GAA F CTCC GAA F CTCC S S	E CCTCGGAACCCGAACCC	TGG P GAC+ CTG T GCA+ CGT Q CCT+ GGA L CTC GGA S	TGT Q CTG GAC C GCC GCTA GAT Y CGT GCA V	CCA GGA CCT E CAG CCT E CAG CTC	CAT Y GGTT CCA CTT K GCA GTT CCAA CTT CCAA CTT CCAA	H GTG T CAA H GTT K CAA GTT N GCT +-CGA L TGA ACT	GGAC GAC GAC GAC GAC GAC GCCC GCCC	CGGG P CTT GAA F CAA GTT K CGT GCA V TCT AGA	GAC-CCTC	TAG S TCC AGG P CAC GTG CAA GTT K CAA GTT K CAA GTT	GGCCCGG	D CGAGGG P CAGGC R CTAGCC -++	E CATO	CGA A GCTC CGA L CGCTC	T CTGG T CGTG CGTG CGTG V GGAC CCTG D GCAG Q GAAG	- 480 - 540 - 600
a a	421 481 541	TTTCC K G AAGAA TTCTT K N GAGTG CTCAC E W TCCGA AGGCT G N AGCCT	CGTC Q CCCAC Q GGGAC E CCGCC G CGGCC G CCGTC V CCTCC	P GGTCC S CTTCC S CTTCC F CCTTCC F CCTTCC CTTCC CTTC CTTCC CTTC CT	GGC' R CAGG S CAA' N CTTG GAAG F CTCG S S GTCG S GT	TCT E CCT GGA L TGGG GAA F ATGG CTT TACC	TGG P GAC CTG T GCAA CGT Q CCT GGA L CTC GGA GGA GGGG GGG GGG	TGT Q CTG GAC C GCC P CTA GAT Y CGT GCA V TTAA	CCA GGA CCT E CAG GTC S GTC S GAT CTA	CAT Y GGTT CCA CTT K GCA GTT CCAA CTT CCAA CTT CCAA CTT CCAA	GTG CAA CAA CAA CAA CAA CAA CAA	GGAC GAC GAC GAC GAC GAC GCCC GCCC	CGGG P CTT GAA F CAA GTT K CGT GCA V TCT AGA	GAC-CCTC	TAG S TCC AGG P CAC GTG CAA GTT K CAA GTT K CAA GTT	GGCCCGG	D CGAGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	E CATO	CGA A GCTC CGA L CGCTC	T CTGG T CGTGGCAC V GGAC CCTG D GCAG CGCAG Q Q GAAG CCTTC	- 480 - 540 - 600
a a	421 481 541	TTTCC K G AAGAA TTCTT K N GAGTG CTCAC E W TCCGA AGGCT G D GGGAA CCCTT G N AGCCT TCGGA	CGTC Q CCCAC Q GGGAC E CCGCC G CGGCC G CCGTC V CCTCC	P GGTC V V GAGC CCTC S CTTC S CTTC F GAGC F CCTC CCTC CCTC CCTC CCTC CCTC CCTC	GGC' R CAGG GTCG S CAA' GTTCG GTAGG S GTTCG S	TCT E CCT L TGGG ACC G ACC TACC GAA TACC C TCCC AGGG	TGG P GAC CTG T GCAA CGT Q CCT GGA L CTC GGA GGA GGGG GGG GGG	TGT Q CTGGCCCGG P CTA GAT GAT V TAA ATT	CCA GGA CCT E CAG GTC S GTC S GAT CTA	CAT Y GGT CCA V GAA CTT K GCA GTT K	GTG CAA CAA CAA CAA CAA CAA CAA	GGAC GAC GAC GAC GAC GAC GCCC GCCC	CGGG P CTT GAA F CAA GTT K CGT GCA V TCT AGA	GAC-CCTC	TAG S TCC AGG P CAC GTG CAA GTT K CAA GTT K CAA GTT	GGCCCGG	D CGAGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	E CATO	CGA A GCTC CGA L CGCTC	T CTGG T CGTGGCAC V GGAC CCTG D GCAG CGCAG Q Q GAAG CCTTC	- 480 - 540 - 600

FIG. 4A

1)	AGP3-	8-1-a Nde	eΙ																		
	_		rgcco																		
	1																			+ GCCA	-
a		М	P	G	dr.	C	다	ם	Er.	D	To7	יט	C	m.	7.7	75	C	G	a		
_		SalI	-	J	•	•	r	F	r	r	VV	E.	C	T	п	A	G	G.	G	G	-
		GGGG																•			
	61	CCCCA		69																	
a		G V	D	-																	
2)	AGP3-	8-2-a Nde 	eΙ																		
	1	TAT	GTGG												-	_					
	1		CACC														•			•	60
a		M	I W	G	A	С	W	P	F	P	W	E	С	F	K	E	G	G	G	G	
		SalI GGGG																			
	61	CCCCA		69															-		
a		G V	ח	_																	

FIG. 4B

```
3) AGP3-8-4-a
     NdeI
       {\tt TATGGTTCCGTTGTGACCTGCTGACTAAACACTGTTTCGAAGCTGGTGGAGGCGGT}
    1 ------ 60
        {\tt ACCAAGGCAAGACACTGGACGACTGATTTGTGACAAAGCTTCGACCACCTCCGCCA}
        MVPFCDLLTKHCFEAGGG-
а
      SalI
      GGGG
    61 ---- 69
      CCCCAGCT
     G V D -
4) AGP3-12-4-a
                   November 6, 2000 12:53 ..
     NdeI
       {\tt TATGGGTTCTCGTTGTAAATACAAATGGGACGTTCTGACTAAACAGTGTTTCCACCAC}
    1 ------ 60
        ACCCAAGAGCAACATTTATGTTTACCCTGCAAGACTGATTTGTCACAAAGGTGGTG
       SalI
     GGTGGAGGCGGTGGGG
   61 ------ 81
     CCACCTCCGCCACCCCAGCT
     G G G G V D ~
```

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FIG. 4C

```
5) AGP3-12-3-a
     NdeI
       {\tt TATGCTGCCGGGTTGTAAATGGGACCTGCTGATCAAACAGTGGGTTTGTGACCCGCTG}
     1 ------ 60
        ACGACGGCCCAACATTTACCCTGGACGACTAGTTTGTCACCCAAACACTGGGCGAC
        MLPGCKWDLLIKQWVCDPL -
               SalI
      GGTGGAGGCGGTGGGG
    61 ------ 81
      CCACCTCCGCCACCCCAGCT
      G G G G V D ~
6) AGP3-12-5-a
       NdeI
       {\tt TATGTCTGACTGACTTTCTTCTACTTCGACATCCTGACTAAATCTGACGTTTGTACTTCTTCT}
     1 -----+ 60
         ACAGACGACTGACAATGAAGCTGTAGGACTGATTTAGACTGCAAACATGAAGAAGA
        M S A D C Y F D I L T K S D V C T S S
               SalI
      GGTGGAGGCGGTGGGG
    61 ------ 81
      CCACCTCCGCCACCCCAGCT
     G G G G V D -
```

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FIG. 4D

7)	AGP3-	:	Nde ·	ΕĪ		-																
	1							TAT: +								'GTI	'CAT				CCTG	.
	т.				,											ממים.	CTA				GGAC	60
										-			.010				.0 .11	.0210	21110		GGAC	
a			M	s	D	D	С	М	Y	D	Q	L	T	R	M	F	I	С	s	N	L	-
						Sa	lI 															
		GG'	ľGG	AGG	CGG	TGG	GG															
	61				-+-			;-+	- 8	1												
		CC	ACC	TCC	GCC	ACC	CCA	GCT														
a		G	G	G	G	G ·	V	D	_													
8)	AGP3-	12-	9-a																			
]	Nde	I																		
		,	TAT	GGA	.CCT	'GAA	CTG	TAA	ATA	.CGA	.CGA	ACT	GAC	TTA	CAA	AGA	ATG	GTG	TCA	GTT	CAAC	
	1																				+	60
			A	.CCT	'GGA	CTT	'GAC	TTA:	TAT	GCT	GCT	TGA	.CTG	AAT	GTT	TCT	TAC	CAC	AGT	CAA	GTTG.	
a			M	D	L	N	С	K	Y	D	E	L	T	Y	K	Е	W	С	Q	F	N	-
						Sa	11															
		GG!	rgg.	AGG	CGG	TGG	GG															
	61.				-+-			+	- 8	1												
		CC	ACC	TCC	GCC	ACC	CCA	GCT														
		_	_	,		_	٠	_														
a		Ġ	G	G	G	G	V	D	_													

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FIG. 4E

```
9) AGP3-12-10-a
       NdeI
       ] .
       {\tt TATGTTCCACGACTGTAAATACGACCTGCTGACTCGTCAGATGGTTTGTCACGGTCTG}
    ACAAGGTGCTGACATTTATGCTGGACGACTGAGCAGTCTACCAAACAGTGCCAGAC
        M F H D C K Y D L L T R Q M V C H G L -
              SalI
      GGTGGAGGCGGTGGGG
    61 ----- 81
      CCACCTCCGCCACCCCAGCT ~
      G G G G V D -
10) AGP3-12-11-a
       NdeI
       TATGCGTAACCACTGTTTCTGGGACCACCTGCTGAAACAGGACATCTGTCCGTCTCCG
    {\tt ACGCATTGGTGACAAAGACCCTGGTGGACGACTTTGTCCTGTAGACAGGCAGAGGC}
        MRNHCFWDHLLKQDICPSP-
a
              SalI
      GGTGGAGGCGGTGGGG
    61 ------ 81
      CCACCTCCGCCACCCCAGCT
     GGGGGVD -
a
```

FIG. 4F

11) AGP3-12-14-a NdeI 1 ------ 60 MANQCWWDSLLKKNVCEFFа SalI GGTGGAGGCGGTGGGG 61 ------ 81 CCACCTCCGCCACCCCAGCT G G G G V D а AGP3 Consensus 12) NdeI TATGTTCCACGACTGCAAATGGGACCTGCTGACCAAACAGTGGGTTTGCCACGGTCTG 1 ------ 60 $\verb|gtatacaaggtgctgacgtttaccctggacgactggtttgtcacccaaacggtgccagac||$ $\begin{smallmatrix} M \end{smallmatrix} \ F \ H \ D \ C \ K \ W \ D \ L \ L \ T \ K \ Q \ W \ V \ C \ H \ G \ L \ - \\ [4mm]$ а SalI GGTGGAGGCGGTGGGĠ 61 ----- 81 CCACCTCCGCCACCCCAGCT G G G G V D -

FIG. 5A

P f

	1 1 1 0 8	
1	I GATCAGCAGTCCCCGGAACATCGTAGCTGACGCCTTCGCGTTGCTCAGTTGTCCAACCCC	
_	CTAGTCGTCAGGGGCCTTGTAGCATCGACTGCGGAAGCGCAACGAGTCAACAGGTTGGGG	60
61	GGAAACGGGAAAAAGCAAGTTTTCCCCGCTCCCGGCGTTTCAATAACTGAAAACCATACT	120
	CCTTTGCCCTTTTCGTTCAAAAGGGGCCGAGGCCGCAAAGTTATTGACTTTTGGTATGA	120
	B g 1 I	
101	I ATTTCACAGTTTAAATCACATTAAACGACAGTAATCCCCGTTGATTTGTGCGCCAACACA	100
141	TAAAGTGTCAAATTTAGTGTAATTTGCTGTCATTAGGGGCAACTAAACACGCGGTTGTGT	180
	-35 -10	
181	Promoter (PcopB)> GATCTTCGTCACAATTCTCAAGTCGCTGATTTCAAAAAACTGTAGTATCCTCTGCGAAAC	0.40
101	CTAGAAGCAGTGTTAAGAGTTCAGCGACTAAAGTTTTTTGACATCATAGGAGACGCTTTG > mRNA st	>
241	GATCCCTGTTTGAGTATTGAGGAGGCGAGATGTCGCAGACAGA	200
241	CTAGGGACAAACTCATAACTCCTCCGCTCTACAGCGTCTGTCT	300
201	CATTGAGTCAAAAGCGGTTTGTGCGCAGAGGTAAGCCTATGACTGAC	260
201	GTAACTCAGTTTTCGCCAAACACGCGTCTCCATTCGGATACTGAGACTCTTTGTTT L S Q K R F V R R G K P M T D S E K Q M	
361	TGGCCGTTGTTGCAAGAAACGTCTTACACACAAAGAGATAAAAGTTTTTGTCAAAAATC	420
301	ACCGGCAACAACGTTCTTTTGCAGAATGTGTGTTTTCTATTTTCAAAAACAGTTTTTAG A V V A R K R L T H K E I K V F V K N P	
	S C	
	a I	
421	CTCTGAAGGATCTCATGGTTGAGTACTGCGAGAGAGAGGGGATAACACAGGCTCAGTTCG	480
	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	

С

С

С

С

FIG. 5B

	-35
	(PrepA)> OPB binding site AGTAAAGACTTTACT
481+++++++	PCATTTCTGAAATGA
-10 TTGTGGCGTAGCATGCTAGATCGTTTAAGGAATTTTGT0	GCTGGCCACGCCGT
AACACCGCATCGTACGATCTAATGACTAGCAAATTCCTTAAAACA(mRNA> D	++ 600
Br md nI II	
AAGGTGGCAAGGAACTGGTTCTGATGTGGATTTACAGGAGCCAĠAA 601+++++ TTCCACCGTTCCTTGACCAAGACTACACCTAAATGTCCTCGGTCTT C M W I Y R S Q K copt (ORF)>	++ 660
<pre>< copA RNAI ATAATCTTCTTCAACTTTTGCGAGTACGAAAAGATTACCGGGGCCC 661+</pre>	CACTTAAACCGTATA
TATTAGAAGAAGTTGAAAACGCTCATGCTTTTCTAATGGCCCCGGGCC N L L R V R K D Y R G P	
< Promoter (RNAI)	-35
<pre>< GCCAACAATTCAGCTATGCGGGGAGTATAGTTATATGCCCGGAAAA 721++++</pre>	++ 780
C Q Q F S Y A G S I V I C P E K TCTGTGCTCGCTCCTTCTGCGCATTGTAAGTGCAGGATGGTGTGAC	FKTSF-
781++++	ACTAGAAGTGGTTT
D r a	opini procesii
I I I CGTATTACCGCCAGGTAAAGAACCCGAATCCGGTGTTTACACCCCG	TEGA A GETTGGA GGA A
641+	ACTTCCACGTCCTT
CGCTGAAGTTCTGCGAAAAACTGATGGAAAAGGCGGTGGGCTTCAC 901++++	AAGGGCAAAACTAA

FIG. 5C

В S t: В Т ${\tt TCGCCATTCATGTGGCGCACGCCCGTTCGCGTGATCTGCGTCGCCGTATGCCACCAGTGC}$ 961 -----+ 1020 ${\tt AGCGGTAAGTACACCGCGTGCGGGCAAGCGCACTAGACGCAGCGGCATACGGTGGTCACG}$ C AIHVAHARSRDLRRRMPPVL-TGCGTCGTCGGGCTATTGATGCGCTCTTGCAGGGGCTGTGTTTCCACTATGACCCGCTGG 1021 -----+ 1080 ACGCAGCAGCCCGATAACTACGCGAGAACGTCCCCGACACAAAGGTGATACTGGGCGACC C RRRAIDALLQGLCFHYDPLA- $\tt CCAACCGCGTCCAGTGCTCATCACCACGCTGGCCATTGAGTGCGGACTGGCGACGGAGT$ 1081 -----+ 1140 ${\tt GGTTGGCGCAGGTCACGAGGTAGTGGTGCGACCGGTAACTCACGCCTGACCGCTGCCTCA}$ C $\tt CTGCTGCCGGAAAACTCTCCATCACCCGTGCCACCCGTGCCCTGACGTTCCTGTCAGAGC$ 1141 ------ 1200 GACGACGGCCTTTTGAGAGGTAGTGGGCACGGTGGGCACGGGACTGCAAGGACAGTCTCG C AAGKLSITRATRALTFLS.EL-TGGGACTGATTACCTACCAGACGGAATATGACCCGCTTATCGGGTGCTACATTCCGACCG 1201 -----+ 1260 ${\tt ACCCTGACTAATGGATGGTCTGCCTTATACTGGGCGAATAGCCCACGATGTAAGGCTGGC}$ C G L I T Y Q T E Y D P L I G C Y I P T D -ATATCACGTTCACATCTGCACTGTTTGCTGCCCTCGATGTATCAGAGGAGGCAGTGGCCG 1261 -----+----+ 1320 TATAGTGCAAGTGTAGACGTGACAAACGACGGGGAGCTACATAGTCTCCTCCGTCACCGGC C I T F T S A L F A A L D V S E E A V A A- $\verb|CCGCGCGCGCGCGCGTGTGGTATGGGAAAACAAACAACGCAAAAAGCAGGGGCTGGATA|\\$ 1321 -----+ 1380 ${\tt GGCGCGGCGTCGGCACACCATACCCTTTTGTTTGTTGCGTTTTTCGTCCCCGACCTAT}$ C ARRSRVVWENKQRKKQGLDT- $\tt CCCTGGGCATGGATGAACTGATAGCGAAAGCCTGGCGTTTTGTTCGTGAGCGTTTTCGCA$ 1381 -----+ 1440 $\tt GGGACCCGTACCTACTTGACTATCGCTTTCGGACCGCAAAACAAGCACTCGCAAAAGCGT$ С LGMDELIAKAWRFVRERFRS-Α £ 1 Ι Т $\tt GTTATCAGACAGAGCTTAAGTCCCGTGGAATAAAGCGTGCCCGTGCGCGTCGTGATGCGG$ 1441 -----+ 1500 CAATAGTCTGTCTCGAATTCAGGGCACCTTATTTCGCACGGGCACGCGCAGCACTACGCC C Y Q T E L K S R G I K R A R A R R D A D -

FIG. 5D

	1501	ACAGGGAACGTCAGGATATTGTCACCCTGGTGAAACGGCAGCTGACGCGCGAAATCGCGG	
	T20T	TGTCCCTTGCAGTCCTATAACAGTGGGACCACTTTGCCGTCGACTGCGCGCTTTAGCGCC	60
С		R E R Q D I V T L V K R Q L T R E I A E -	
	1561	AAGGGCGCTTCACTGCCAATCGTGAGGCGGTAAAACGCGAAGTTGAGCGTCGTGTGAAGG	
	1201	TTCCCGCGAAGTGACGGTTAGCACTCCGCCATTTTGCGCTTCAACTCGCAGCACACTTCC	520
С		G R F T A N R E A V K R E V E R R V K E -	
		AGCGCATGATTCTGTCACGTAACCGTAATTACAGCCGGCTGGCCACAGCTTCCCCCTGAA	
	1621	TCGCGTACTAAGACAGTGCATTGGCATTAATGTCGGCCGACCGGTGTCGAAGGGGGACTT	680
С		R M I L S R N R N Y S R L A T A S . P *	
		AGTGACCTCCTGAATAATCCGGCCTGCGCCGGAGGCTTCCGCACGTCTGAAGCCCGAC	
	1681	TCACTGGAGGAGACTTATTAGGCCGGACGCGGCCTCCGAAGGCGTGCAGACTTCGGGCTG	40
		P f	
		M I	
	1741	AGCGCACAAAAAATCAGCACCACATACAAAAAACAACCTCATCATCCAGCTTCTGGTGCA	100
		TCGCGTGTTTTTAGTCGTGGTGTATGTTTTTTTTTGTTGGAGTAGTAGGTCGAAGACCACGT	00
	1001	TCCGGCCCCCCTGTTTTCGATACAAAACACGCCTCACAGACGGGGAATTTTGCTTATCC	
	1801	AGGCCGGGGGGACAAAAGCTATGTTTTGTGCGGAGTGTCTGCCCCTTAAAACGAATAGG	60
		ori	
	1061	ACATTAAACTGCAAGGGACTTCCCCATAAGGTTACAACCGTTCATGTCATAAAGCGCCAT	
	1801	TGTAATTTGACGTTCCCTGAAGGGGTATTCCAATGTTGGCAAGTACAGTATTTCGCGGTA	20
		ori	
	1021	CCGCCAGCGTTACAGGGTGCAATGTATCTTTTAAACACCTGTTTATATCTCCTTTAAACT	
	1921	GGCGGTCGCAATGTCCCACGTTACATAGAAAATTTGTGGACAAATATAGAGGAAATTTGA	80
	1001	ACTTAATTACATTCATTTAAAAAGAAAACCTATTCACTGCCTGTCCTTGGACAGACA	
	TOOT	TGAATTAATGTAAGTAAATTTTTCTTTTGGATAAGTGACGGACAGGAACCTGTCTA	40
		ATGCACCTCCCACCGCAAGCGGCCGCCCTACCGGAGCCGCTTTAGTTACAACACTCAG	
	2041	+ 21	00
a		TACGTGGAGGGTGGCGTTCGCCGCCCGGGGATGGCCTCGGCGAAATCAATGTTGTGAGTC M H L P P Q A A G P Y R S R F S Y N T Q -	
		repA4 protein>	->
	2101	ACACAACCACCAGAAAAACCCCGGTCCAGCGCAGAACTGAAACCACAAAGCCCCTCCCT	C 0
	2101	TGTGTTGGTGGTCTTTTTGGGGCCAGGTCGCGTCTTGACTTTGGTGTTTCGGGGAGGGA	60
a		T Q P P E K P R S S A E L K P Q S P S L -	
	2161	ATAACTGAAAAGCGGCCCGGCCCGGTCCGAAGGGCCGGAACAGAGTCGCTTTTAATTAT	20
	2101	TATTGACTTTTCGCCGGGCCGGGCCAGGCTTCCCGGCCTTGTCTCAGCGAAAATTAATA	∠0
a		I T E K R P R P G P K G R N R V A F N Y -	

FIG. 5E

	2221		GAATGTTGTAACTACTTCATCATCGCTGTCAGTCTTCTCGCTGGAAGTTCTCAGTACACG ++++++																		
a	2221		AAC	TTA	GAT		GTA		GCG.	ACA	GTC	AGA	AGA	GCG	ACC	TTC	AAC	GAG'		GTGC	
						B g l I	f i														
	2281	CTCGTA GAGCAT		-+			+				+			+-			+			+	2340
a		L V	S	G	Р	D	G	P	Г	T	R	R	Y	A	P	Т	S	G.	K	P	-
a	2341	TCGTCG AGCAGC S S	CCCI	-+ !GG!	rga(GGC'	+ IGG	CGC	 GTG'	rc'r	rcga	AGA(GAG'	-+- TAC	 CGA	CTI	+ TCG	 :CC	 TATA	CCAG	2400
	2401	TGGCAG		-+			+				+ -			-+-			+			+	2460
a		ACCGTO W Q							CAC'	rtt2	AGA	rag:	(TAC	GTC.	ATG	GCC	:GAA	TG(CGGC	CCGA	
													B s t E I						v		•
	2461	TCGGCG AGCCGC		+			+				<u> </u>			-+-			+		. 	+	2520
													B p L U 1								
	2521	ATGCGG		+			+-			1				-+			+			+	2580
	2581	ACAAAA TGTTTT	ATA	GGA	CAZ	AGTT	raa <i>z</i> +-	AAAT	TT	ACAG	GCG	ATG	CAF	ATG	YTT	CAA	ACA	CGT	TAA	CAAT	2640
	2641	ATCGGG TAGCCC	GGT	GGG	CGZ	AAGZ	AAC:	rcc <i>i</i>	AGC <i>I</i>	ATG#	GAT	ccc	CGC	:GC:	rgg	AGG	ATC	ATC	CAG	CCGG	2700
	2701	CGTCCC GCAGGG		+			-+-			+				-+			+			+	2760

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		N B s p p 1 V I	
	2761	CTCGTGATGGCAGGTTGGCGTCGCTTGGTCGGTCATTTCGAACCCCAGAGTCCCGCTCA	0
		GAGCACTACCGTCCAACCCGCAGCGAACCAGCCAGTAAAGCTTGGGGTCTCAGGGCGAGT	
	2821	GAAGAACTCGTCAAGAAGGCGATAGAAGGCGATGCGCTGCGAATCGGGAGCGGCGATACC++++++	0
f	ķ	* F F E D L L R Y F A I R Q S D P A A I G - < APHII protein [kanamycin resistance gene]	
	2881	GTAAAGCACGAGGAAGCGGTCAGCCCATTCGCCGCCAAGCTCTTCAGCAATATCACGGGT	0
£		CATTTCGTGCTCCTTCGCCAGTCGGGTAAGCGGCGGTTCGAGAAGTCGTTATAGTGCCCA Y L V L F R D A W E G G L E E A I D R T -	
	2941	AGCCAACGCTATGTCCTGATAGCGGTCCGCCACACCCAGCCGGCCACAGTCGATGAATCC	0
f	22	TCGGTTGCGATACAGGACTATCGCCAGGCGGTGTGGGTCGGCCGGTGTCAGCTACTTAGG A L A I D Q Y R D A V G L R G C D I F G -	•
	3001	AGAAAAGCGGCCATTTTCCACCATGATATTCGGCAAGCAGGCATCGCCATGAGTCACGAC	0
f	3001	TCTTTTCGCCGGTAAAAGGTGGTACTATAAGCCGTTCGTCCGTAGCGGTACTCAGTGCTG S F R G N E V M I N P L C A D G H T V V -	•
	2061	GAGATCCTCGCCGTCGGGCATGCGCGCCTTGAGCCTGGCGAACAGTTCGGCTGGCGCGAG	Λ
f	2001	CTCTAGGAGCGGCAGCCGTACGCGCGGAACTCGGACCGCTTGTCAAGCCGACCGCGCTC L D E G D P M R A K L R A F L E A P A L -	J
	2121	CCCCTGATGCTCTTCGTCCAGATCATCCTGATCGACAAGACCGGCTTCCATCCGAGTACG	n
£	J121	GGGGACTACGAGAAGCAGGTCTAGTAGGACTAGCTGTTCTGGCCGAAGGTAGGCTCATGC G Q H E E D L D D Q D V L G A E M R T R -	J
	3181	TGCTCGCTCGATGCGATGTTTCGCTTGGTGGTCGAATGGGCAGGTAGCCGGATCAAGCGT	n
f	3101	ACGAGCGAGCTACGAAAGCGAACCACCAGCTTACCCGTCCATCGGCCTAGTTCGCA A R E I R H K A Q H D F P C T A P D L T -	
	2041	ATGCAGCCGCCGCATTGCATCAGCCATGATGGATACTTTCTCGGCAGGAGCAAGGTGAGA	Λ
£	3241	TACGTCGGCGGCGTAACGTAGTCGGTACCTATGAAAGAGCCGTCCTCGTTCCACTCT H L R R M A D A M I S V K E A P A L H S -	J
_		TGACAGGAGATCCTGCCCGGCACTTCGCCCAATAGCAGCCAGTCCCTTCCCGCTTCAGT	
	3301	ACTGTCCTCTAGGACGGGCCGTGAAGCGGGTTATCGTCGGTCAGGGAAGGCGAAGTCA	0
f		S L L D Q G P V E G L L L W D R G A E T -	
	3361	GACAACGTCGAGCACAGCTGCGCAAGGAACGCCCGTCGTGGCCAGCCA	0
£		V V D L V A A C P V G T T A L W S L R A -	
	3421	TGCCTCGTCCTGCAATTCATTCAGGACACCGGACAGGTCGGTC	0
		ACGGAGCAGGACGTTAAGTAAGTCCTGTGGCCTGTCCAGCCAG	

FIG. 5G

f		A'E DQLENLVGSLDTKVFLVP-	
	2401	GCGCCCTGCGCTGACAGCCGGAACACGGCGGCATCAGAGCAGCCGATTGTCTGTTGTGC	
£	348I	CGCGGGGACGCACTGTCGGCCTTGTGCCGCCGTAGTCTCGTCGGCTAACAGACAACACG R G Q A S L R F V A A D S C G I T Q Q A -	10
		. E	
		a g I	
	3541	CCAGTCATAGCCGAATAGCCTCTCCACCCAAGCGGCCGGAGAACCTGCGTGCAATCCATC	١.0
f		GGTCAGTATCGGCTTATCGGAGAGGTGGGTTCGCCGGCCTCTTGGACGCACGTTAGGTAG W D Y G F L R E V W A A P S G A H . L G D -	, 0
	3601	TTGTTCAATCATGCGAAACGATCCTCATCCTGTCTCTTGATCTGATCTTGATCCCCTGCG	50
£		AACAAGTTAGTACGCTTTGCTAGGAGTAGGACAGAGAACTAGACTAGAACTAGGGGACGC Q E I M	, ,
<	- APHI	II (kanamycin resistance) protein) -10.	
		<pre>< mRNA APHII CCATCAGATCCTTGGCGGCAAGAAAGCCATCCAGTTTACTTTGCAGGGCTTCCCAACCTT</pre>	
	3661	GGTAGTCTAGGAACCGCCGTTCTTTCGGTAGGTCAAATGAAACGTCCCGAAGGGTTGGAA	:0
		−35	
	3721	ACCAGAGGGCGCCCCAGCTGGCAATTCCGGTTCGCTTGCTGTCCATAAAACCGCCCAGTC	10
		TGGTCTCCCGCGGGGTCGACCGTTAAGGCCAAGCGAACGACAGGTATTTTGGCGGGTCAG	
	3781	TAGCTATCGCCATGTAAGCCCACTGCAAGCTACCTGCTTTCTCTTTTGCGCTTGCGTTTTC	.0
		ATCGATAGCGGTACATTCGGGTGACGTTCGATGGACGAAAGAGAAACGCGAACGCAAAAG	
	3841	CCTTGTCCAGATAGCCCAGTAGCTGACATTCATCCGGGGTCAGCACCGTTTCTGCGGACT	0
		GGAACAGGTCTATCGGGTCATCGACTGTAAGTAGGCCCCAGTCGTGGCAAAGACGCCTGA	
	3901	GGCTTTCTACGTGTTCCGCTTCCTTTAGCAGCCCTTGCGCCCTGAGTGCTTGCGGCAGCG	0
		CCGAAAGATGCACAAGGCGAAGGAAATCGTCGGGAACGCGGGACTCACGAACGCCGTCGC	
	3961	TGAAGCTACATATATGTGATCCGGGCAAATCGCTGAATATTCCTTTTGTCTCCGACCATC	. 0
	2901	ACTTCGATGTATATACACTAGGCCCGTTTAGCGACTTATAAGGAAAACAGAGGCTGGTAG	U
		B c	
		g I	
	4021	AGGCACCTGAGTCGCTGTCTTTTCGTGACATTCAGTTCGCTGCGCTCACGGCTCTGGCA	0
		TCCGTGGACTCAGCGACAGAAAAAGCACTGTAAGTCAAGCGACGCGAGTGCCGAGACCGT	
		par locus	

FIG. 5H

4081	GTGAATGGGGGTAAATGGCACTACAGGCGCCTTTTATGGATTCATGCAAGGAAACTACCC	4140
1001	CACTTACCCCCATTTACCGTGATGTCCGCGGAAAATACCTAAGTACGTTCCTTTGATGGG	4140
4141	ATAATACAAGAAAAGCCCGTCACGGGCTTCTCAGGGCGTTTTATGGCGGGTCTGCTATGT	4200
	TATTATGTTCTTTTCGGGCAGTGCCCGAAGAGTCCCGCAAAATACCGCCCAGACGATACA	1200
4201	GGTGCTATCTGACTTTTTGCTGTTCAGCAGTTCCTGCCCTCTGATTTTCCAGTCTGACCA	4260
	CCACGATAGACTGAAAAACGACAAGTCGTCAAGGACGGGAGACTAAAAGGTCAGACTGGT	
4261	CTTCGGATTATCCCGTGACAGGTCATTCAGACTGGCTAATGCACCCAGTAAGGCAGCGGT	4320
	GAAGCCTAATAGGGCACTGTCCAGTAAGTCTGACCGATTACGTGGGTCATTCCGTCGCCA	
	N B s s i a	
4321	ATCATCAACAGGCTTACCCGTCTTACTGTCGAAGACGTGCGTAACGTATGCATGGTCTCC	4380
	TAGTAGTTGTCCGAATGGGCAGAATGACAGCTTCTGCACGCATTGCATACGTACCAGAGG	
	T1 hairpin	
4381	CCATGCGAGAGTAGGGAACTGCCAGGCATCAAATAAAACGAAAGGCTCAGTCGAAAGACT+ GGTACGCTCTCATCCCTTGACGGTCCGTAGTTTATTTTGCTTTCCGAGTCAGCTTTCTGA	4440
4441	GGGCCTTTCGTTTTATCTGTTGTTGTTGTCGGTGAACGCTCTCCTGAGTAGGACAAATCCGC	4500
•	CCCGGAAAGCAAATAGACAACAACAGCCACTTGCGAGAGGACTCATCCTGTTTAGGCG T1 stop>	
	P s	
	p 1	
i	4 0 6	
4501	I CGGGAGCGGATTTGAACGTTGCGAAGCAACGGCCCGGAGGGTGGCGGGCAGGACGCCCGC	4560
#20T	GCCCTCGCCTAAACTTGCAACGCTTCGTTGCCGGGCCTCCCACCGCCCGTCCTGCGGGCG	4500
	T2 hairpin	•
4561	CATAAACTGCCAGGCATCAAATTAAGCAGAAGGCCATCCTGACGGATGGCCTTTTTGCGT	4620
#20T	GTATTTGACGGTCCGTAGTTTAATTCGTCTTCCGGTAGGACTGCCTACCGGAAAAACGCA T2 stop>	-1020

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FIG. 5I

	A a t I TTCTACAAACTCTTTTGTTTATTTTTCTAAATACATTCAAATATGGACGTCGTACTTAAC	
4621	AAGATGTTTGAGAAAACAAATAAAAAGATTTATGTAAGTTTATACCTGCAGCATGAATTG * -	4680
4681	TTTTAAAGTATGGGCAATCAATTGCTCCTGTTAAAATTGCTTTAGAAATACTTTGGCAGC	171C
*	AAAATTTCATACCCGTTAGTTAACGAGGACAATTTTAACGAAATCTTTATGAAACCGTCG S K F Y P C D I A G T L I A K S I S Q C luxR protein	-
4741	GGTTTGTTGTATTGAGTTTCATTTGCGCATTGGTTAAATGGAAAGTGACCGTGCGCTTAC	4900
	CCAAACAACATAACTCAAAGTAAACGCGTAACCAATTTACCTTTCACTGGCACGCGAATG R N T T N L K M Q A N T L H F T V T R K	-
4801	TACAGCCTAATATTTTTGAAATATCCCAAGAGCTTTTTCCTTCGCATGCCCACGCTAAAC	4860
	ATGTCGGATTATAAAAACTTTATAGGGTTCTCGAAAAAGGAAGCGTACGGGTGCGATTTG S C G L I K S I D W S S K G E C A W A L	_
4861		4920
	TAAGAAAAAGAGAAAACCAATTTAGCAACAAACTAAATAAA	-
4921		4980
	CTATTAATAGTTGATCTCTTCCTTGTTAATTACCATACAAGTATGTGCGTACATTTTTAT R Y N D V L S P V I L P I N M C A H L F	_
	B s m	
4981	AACTATCTATATAGTTGTCTTTCTCTGAATGTGCAAAACTAAGCATTCCGAAGCCATTAT	5040
1501	TTGATAGATATCAACAGAAAGAGACTTACACGTTTTGATTCGTAAGGCTTCGGTAATA L S D I Y N D K E S H A F S L M G F G N	-
5041	TAGCAGTATGAATAGGGAAACTAAACCCAGTGATAAGACCTGATGATTTCGCTTCTTTAA	E100
2041	ATCGTCATACTTATCCCTTTGATTTGGGTCACTATTCTGGACTACTAAAGCGAAGAAATT N A T H I P F S F G T I L G S S K A E K	-
5101	TTACATTTGGAGATTTTTTATTTACAGCATTGTTTTCAAATATATTCCAATTAATCGGTG	E160
2101	AATGTAAACCTCTAAAAAATAAATGTCGTAACAAAAGTTTATATAAGGTTAATTAGCCAC I V N P S K K N V A N N E F I N W N I P	- 5T00
F1 64	AATGATTGGAGTTAGAATAATCTACTATAGGATCATATTTTATTAAATTAGCGTCATCAT	
2161	TTACTAACCTCAATCTTATTAGATGATATCCTAGTATAAAATAATTTAATCGCAGTAGTA S H N S N S Y D V I P D Y K I L N A D D	5220 -
	· · · · · · · · · · · · · · · · · · ·	

FIG. 5J

	5221	AATATTGCCTCCATTTTTTAGGGTAATTATCCAGAATTGAAATATCAGATTTAACCATAG	5000
đ	J221	TTATAACGGAGGTAAAAAATCCCATTAATAGGTCTTAACTTTATAGTCTAAATTGGTATC Y Y Q R W K K P Y N D L I S I D S K V M	5280
		N r u T	
	5281	AATGAGGATAAATGATCGCGAGTAAATAATATTCACAATGTACCATTTTAGTCATATCAG	
	0202	TTACTCCTATTTACTAGCGCTCATTTATTATAAGTGTTACATGGTAAAATCAGTATAGTC S H P Y I I A L L Y Y E C H V M K T M D	5340
,	5341	ATAAGCATTGATTAATATCATTATTGCTTCTACAGGCTTTAATTTATTAATŢATTCTGT	F 4 0 0
	3311	TATTCGTAACTAATTATAGTAATAACGAAGATGTCCGAAATTAAAAATAATTAAT	-
	5401	AAGTGTCGTCGGCATTTATGTCTTTCATACCCATCTCTTTATCCTTACCTATTGTTTGT	5460
	2407	TTCACAGCAGCCGTAAATACAGAAAGTATGGGTAGAGAAATAGGAATGGATAACAAACA	5460
		GCAAGTTTTGCGTGTTATATATCATTAAAACGGTAATAGATTGACATTTGATTCTAATAA	
		CGTTCAAAACGCACAATATATAGTAATTTTGCCATTATCTAACTGTAACTAAGATTATT	5520
		< < <	
	lı	1XR mRNA start sites	
		CRP Binding Site	
	5521	ATTGGATTTTTGTCACACTATTATATCGCTTGAAATACAATTGTTTAACATAAGTACCTG	5500
		TAACCTAAAAACAGTGTGATAATATAGCGAACTTTATGTTAACAAATTGTATTCATGGAC	3360
	lux	C B Promoter (luxPR)> 1 b operator site -35 -10 a a	
		I I TAGGATCGTACAGGTTTACGCAAGAAAATGGTTTGTTATAGTCGATTAATCGATTTGATT	
	5581	ATCCTAGCATGTCCAAATGCGTTCTTTTACCAAACAATATCAGCTAATTAGCTAAACTAA	5640 t>
		NdeI	
	E 6 1 1	CTAGATTTGTTTTAACTAATTAAAGGAGGAATAACATATGATCGCTCCACCATGCACCAG	
	2041	GATCTAAACAAAATTGATTAATTTCCTCCTTATTGTATACTAGCGAGGTGGTACGTGGTC	5700
b		M I A P P C T S	-
	5701	TGAGAAGCATTATGAGCATCTGGGACGGTGCTGTAACAAATGTGAACCAGGAAAGTACAT	57 <i>6</i> 0
	2101	ACTCTTCGTAATACTCGTAGACCCTGCCACGACATTGTTTACACTTGGTCCTTTCATGTA	J/0U
b		E K H Y E H L G R C C N K C E P G K Y M	-

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FIG. 5K

	5761	GTC	TTC	TAA	ATO	GCAC	TAC	TAC	CTC	CTGA	CAC	TGT	PTA	TCT	GCC	CTG	TGG	CCC	GGA	TGA	ATA	5820
	3701	CAG	AAG	ATT	'TAC	GTG	ATG	ATG	GAG	ACT	GTC	CACA	TAC	AGA	CGG	GAC	ACC	GGG	CCI	ACI	TAT	5820
b		S	S	K	С	Т	Т	Т	S	D	s	V	С	L	P	С	G	P	D	E	Y	-
	5821	CTT	GGA	TAG	CTC +	GAA	TGA	AGA	AGA	TAA	ATC	CTT	GCI	GCA	TAA	AGI	TTG	TGA	TAC	AGG	CAA	5880
		GAA	CCT	ATC	GAC	CTT	'ACI	TCT	TCT	rra:	TAC	:GAA	CGA	CGI	LTA	TCA	AAC	:ACI	OTA	TCC	GTT	3000
b		L	D	S	W	N	E	E	D	K	С	L	L	H	K	V	С	D	Т	G	K	-
																		paL				
	5881				+			-+-			+				+			-+-			GGC	5940
		CCG	GGA	CCA	.CCG	GCA	.CCA	.GCG	GCC	GTT:	'GTC	ATG	CTG	GGG	GGC	CGC	GAC	:GCG	CAC	GTG	CCG	
b		Α.	L	V	A	V	V	A	G	N	S	Т	Т	P	R	R	С	A	С	T	A	-
	Acc	:65 <u>Į</u>	Kpn	I 																		
	=0.44	TGG	GTA	 CCA	CTG	GAG	CCA	.GG.A	.CTG	CGA	.GTG	CTG	CCG	CCG	CAA	.CAC	CGA	.GTG	CGC	GCC	GGG	
	5941	ACC	CAT	 GGT	+ GAC	CTC	 GGT	CCI	GAC	GCT	CAC	GAC	GGC	 GGC	+ GTT	GTG	GCT	-+- CAC	GCG	CGG	CCC	6000
b		G	Y	Н	W	s	Q	D	С	E	С	С	R	R	N	т	E	С	A	P	G	-
		CCT	ccc	ccc.	CCA	CCA	ccc	cmm	ררא	CCM	<i>~</i> 7 7 7	(1 A A	aa x	a 2 a	7. CIC	ama	~~~	3 C C	mma	aam	таа	
	6001	GGA			+			-+-	~		+				+			-+-			+	6060
b						Н												r GG				_
		AGG							-										_			
	6061				+			-+-			+				+			-+-			+	6120
b			Y				A			S	T			C			W	т			T	_
		CTT	CCT'	rgg	AAA	GAG.	AGT.	AGA	ACA	TCA	TGG	GAC	AGA	GAA.				- GGT	TTG	CAG	_	
	6121	GAA			+			-+-			+				+			-+-			+	6180
b		F	L	G	K	R	V	E	Н	Н	G	т	E	K	s	D	V	V	С	S	s	_
															A	ccI						
															Sa							
	6181	TTC			+			-+-			+				+			-+-			+	6240
_		AAG																				
b		ន	L	P	A	R	K	P	P	N	E	q Q	H	V	Y1	V	D sta:	K	T	Н	${f T}$	-

FIG. 5L

						Ε	3spE	ΞΙ						Ał	ıdI							
	CD 41	ATG	TCC	ACC	ттс	TCC	CAGC	τcc.	CGGZ	AAC	TCC	rggo	GGG	SACC	GTC	AGI	CTI	CCI	CTT	ccc	CCC	
	6241	TAC	CAGO	TGG	AAC	CAGO	TCG	AGC	GCC:	rtg.	AGG	ACCC	ccc	TGG	GAG	TCA	GAA	GGA	GAA	.GGG	+ GGG	6300
b		С	P	P	С	P	A	P	E	L	L	G	G	P	S	V	F	L	F	P	P	-
				•		Bs	IHqa															
	6201	AAA	ACC	CAA	.GGA	CAC	cci	'CAT	'GA'	rct(CCC	GAC	ccc	TGA	GGT	'CAC	ATG	CGT	GGT	GGT	GGA	
	0301	TTT	TGG	GTT	CCI	GTG	GGA	GTA	CTA	AGA	GGG	CCTG	GGG	ACT	CCA	GTC	TAC	-+- GCA	.CCA	.CCA	+ CCT	6360
b		K	P	K	D	T	L	М	I	S	R	Т	P	E	V	Т	С	V	Ÿ	V	D	-
	C2 C1	CGT	GAG	CCA	CGA	AGA	'CCC	TGA	\GG1	ГСА	AGT	CAA	CTG	GTA	CGT	GGA	.CGG	CGT	GGA	GGT	GCA	
	030T	GCA	CTC	GGT	GCI	TCI	'GGG	-+- ACI	CCF	AGT'	rca <i>i</i>	· \GTT	'GAC	CAT	+ GCA	CCT	GCC	-+- GCA	 CCT	CCA	+ CGT	6420
b		V	S	Н	Ε	D	P	E	V	K	F	N	W	Y	V	D	G	V	E	V	Н	-
		TAA	TGC	CAA	.GAC	AAA	.GCC	GCG	GGZ	4GG2	AGC <i>I</i>	AGTA	CAA	CAG	CAC	GTA	CCG	TGT	GGT	CAG	CGT	
	6421				+			-+-			+ rcgi				+			-+-			+	6480
b							P		Е	E		Y		s			R				V	_
					Eco	NI																
	6481	CCT	CAC	CGT	CCT	GĊA	CCA	GGA	CTG	GC.	rga <i>r</i>	TGG	CAA	.GGA	GTA	CAA	GTG	CAA	GGT	CTC	CAA	6540
		GGA																				0340
b		L	T	V	L	H	Q	D	W	L	N	G	K	E	Y	K	С	K	V	S	N	_
	65/1	CAA	AGC	CCT	CCC	AGC	CCC	CAT	'CGA	GAZ	AAAC	CAT	CTC	CAA	AGC	CAA	AGG	GCA	GCC	CCG	AGA	
	6541	GTT	TCG	GGA	GGG	TCG	GGG	GTA	GCI	CTI	rtte	GTA	GAG	GTT	+ TCG	 GTT	TCC	CGT	CGG	GGC'	+ ICT	6600
b		K	A	L	P	A	P	I	E	K	Т	I	S	K	A	K	G	Q	P	R	E	-
			70	O T							naI					~						
		3.00		rGI 		a. a	~~~	aaa		3ma]						~ ~	xAI 					
	6601				+			-+-			+				+			-+-			+	6660
											GGC											
b		P	Q	V	Y	T	L	P	P	S	R	D	E	L	T	K	N	Q	V	S	L	-
	6661	GAC	CTG	CCT	GGT +	CAA 	AGG	CTT -+-	CTA	TCC	CAG	CGA	CAT	CGC	CGT	GGA	GTG(GGA(GAG(CAA	rgg +	6720
		CTG																				0.20
b		T	С	L	V	K	G	F	Y	P	S	D	I	A	V	E	M	E	S	N	G	-
	6721	GCA																				6700
		CGT																				0780

FIG. 5M

b		Q	P	E	N	N Y	K	T	T	P	P	v	L	D	s	D	G	s	F	F	_
	6504	CCT	CTA	CAG	CAAG	CTCA	CCGI	'GGA	CAA	GAG	CAG	GTG	GCA	GCA	GGG	GAA	CGT	CTT(CTC	ATG	
	6781	GGA	GAT	GTC	GTTC	GAGT	GGCA	CCT	GTT	+ CTC	GTC	CAC	CGT	+ CGT	CCC	 CTT	-+- GCA	GAA	GAG	rac +	6840
b		L	Y	S	K	L T	V	D	K	S	R	W	Q	Q	G	N	V	F	S	С	-
	6841	CTC	CGT	GAT(GCAT	GAGG	CTCI	'GCA	CAA	CCA	CTA	CAC	GCA	GAA	GAG	CCT	CTC	CCT	GTC:	rcc	
	0047	GAG	GCA	CTA	CGTA	CTCC	GAGA	CGT	GTT	GGT(GAT	GTG	CGT	CTT	CTC	GGA(GAG(GGA(CAG	+ AGG	6900
b		S	V	M	Н	E A	L	Н	N	Н	Y	т	Q	K	s	L	S	L	S	P	-
				Bai	THn I													•			
	6901	GGG	raa?	ATA	ATĠG	ATCC	GCGG											GGA/	AGC	rga	6060
	0,01					TAGG												CCT:	rcga	ACT	6960
b		G	K	*		Blp:	τ														
						210.	1										т7 ->	hai	irpi	ln	
	6961	GTT	GCT	rgc:	rgcc +	ACCG	CTGA	GCA.	ATA	ACTA	AGC	ATA	ACC	CCT	rgg	GC(CTCT	raaz	ACGO	GT	7020
		CAA	CCGZ	ACG?	ACGG	TGGC	GACT	'CGT'	rat'	rga:	rcg:	TAT'	rgg	GGA	ACC(CCG	GAGA	ATTI	rgco	CA	7020
		<pre>< CTTGAGGGGTTTTTTGCTGAAAGGAGGAACCGCTCTTCACGCTCTTCACGCGGATAAATA</pre>																			
	7021		CTCC	CCC		AACG								+ GCGZ		 \GT(-+ GCGC	CCTA	 \TTT	+ 'AT	7080
		-T7	sto	p -		>															
																to	qoq 	hai	rpi	.n ·->	
	7081			- - - 4		TCCA(+-			+-							+			-+	7140
						AGGT(CATT	ACT(GA(FTC?	rtg <i>i</i>	AGG:	ΓAG	4CC1	'AA'	ACAI	AGTC	TTC	CGA	.GC	
		<			irp		-														
	7141			1		GTTTT	+-			+-							+			-+	7200
		CAAC				CAAA top -		AAC	CACT	CT?	l'AGC	CGT	CGT"	rga <i>i</i>	ACAC	GCGC	CGGI	TAC	CTC	:GG	
		ΆΨСΊ	יריקיזי	יריביחי	ממיי	CGACO	ימפמ	(" አ ጥ	ቦሮ ል ፣	\CZ\ 7	\ ''' ልር	2 (° 2). 7	١٩٥٨	ע כי כי	יחינייר	בא כיז	ארייי	יתותור	יר <i>ד</i> יב <i>ו</i> י	.m.c	
	7201					GCTG(-+-			+-							+			-+	7260
			-502		*O+T		.000	~ 1 tm	1011			-311			. ~~(. 1 ()	. 1 GF	run.		AG.	
	7261	CAGT	CCC	TCT	TCC.	ACCT	CTG.	ACC	3 - 72	85											
						TGGAC															

FIG. 6A

[AatII sticky end] (position #4358 in pAMG21)

- 5' GCGTAACGTATGCATGGTCTCC-3' TGCACGCATTGCATACGTACCAGAGG-
- $-\texttt{CCATGCGAGAGTAGGGAACTGCCAGGCATCAAATAAACGAAAGGCTCAGTCGAAAGACT--GGTACGCTCTCATCCCTTGACGGTCCGTAGTTTATTTTGCTTTTCCGAGTCAGCTTTCTGA--$

- $-{\tt CATAAACTGCCAGGCATCAAATTAAGCAGAAGGCCATCCTGACGGATGGCCTTTTTGCGT-GTATTTGACGGTCCGTAGTTTAATTCGTCTTCCGGTAGGACTGCCTACCGGAAAAACGCA-GTATTTGACGGTCGTAGGACTGCCTACCGGAAAAACGCA-$

<u>Aat</u>II

- -TTCTACAAACTCTTTTGTTTATTTTTCTAAATACATTCAAATATGGACGTCGTACTTAAC--AAGATGTTTGAGAAAACAAATAAAAAGATTTATGTAAGTTTATACCTGCAGCATGAATTG-
- $-TTTTAAAGTATGGGCAATCAATTGCTCCTGTTAAAATTGCTTTAGAAATACTTTGGCAGC-\\-AAAATTTCATACCCGTTAGTTAACGAGGACAATTTTAACGAAATCTTTATGAAACCGTCG-\\$
- $-\mathsf{GGTTTGTATTGAGTTCATTTGCGCATTGGTTAAATGGAAAGTGACCGTGCGCTTAC--CCAAACAACATAACTCAAAGTAAACGCGTAACCAATTTACCTTTCACTGGCACGCGAATG--$
- $-{\tt TACAGCCTAATATTTTTGAAATATCCCAAGAGCTTTTTCCTTCGCATGCCCACGCTAAC-ATGTCGGATTATAAAAACTTTATAGGGTTCTCGAAAAAGGAAGCGTACGGGTGCGATTTG-$
- $-\mathsf{GATAATTATCAACTAGAGAAGGAACAATTAATGGTATGTTCATACACGCATGTAAAAATA-\\-\mathsf{CTATTAATAGTTGATCTCTTTCTTGTTAATTACCATACAAGTATGTGCGTACATTTTTAT-\\$
- $-{\tt TAGCAGTATGAATAGGGAAACTAAACCCAGTGATTAAGACCTGATGATTTCGCTTCTTAA-ATCGTCATACTTATCCCTTTGATTTGGGTCACTATTCTGGACTACTAAAGCGAAGAAATT-$
- $TTACATTTGGAGATTTTTTATTTACAGCATTGTTTTCAAATATATTCCAATTAATCGGTG-\\-AATGTAAACCTCTAAAAAATAAATGTCGTAACAAAAGTTTATATAAGGTTAATTAGCCAC-$
- $-\mathtt{AATATTGCCTCCATTTTTTAGGGTAATTATCCAGAATTGAAATATCAGATTTAACCATAG--\\TTATAACGGAGGTAAAAAATCCCATTAATAGGTCTTAACTTTATAGTCTAAATTGGTATC--$
- $-\mathtt{AATGAGGATAAATGATCGCGAGTAAATAATATTCACAATGTACCATTTTAGTCATATCAG-TTACTCCTATTTACTACTCGCTCATTTATTATAAGTGTTACATGGTAAAATCAGTATAGTC-$

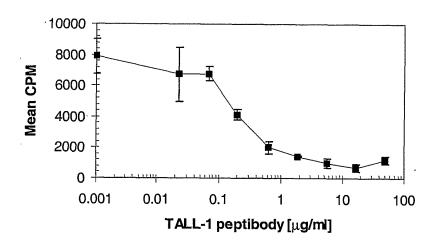
- $-\mathsf{GCAAGTTTTGCGTGTTATATATCATTAAAACGGTAATAGATTGACATTTGACTTCTAATAA-\\-\mathsf{CGTTCAAAACGCACAATATATAGTAATTTTGCCATTATCTAACTGTAAACTAAGATTATT-\\$

FIG. 6B

- $-\mathtt{ATTGGATTTTTGTCACACTATTATATCGCTTGAAATACAATTGTTTAACATAAGTACCTG -\mathtt{TAACCTAAAAACAGTGTGATAATATAGCGAACTTTATGTTAACAAATTGTATTCATGGAC-$
- $-\mathtt{TAGGATCGTACAGGTTTACGCAAGAAAATGGTTTGTTATAGTCGATTAATCGATTTGATT -\mathtt{ATCCTAGCATGTCCAAATGCGTTCTTTTACCAAACAATATCAGCTAATTAGCTAAACTAA-$
- $-\mathtt{CTAGATTTGTTTTAACTAATTAAAGGAGGAATAACATATGGTTAACGCGTTGGAATTCGA-$ -GATCTAAACAAAATTGATTAATTTCCTCCTTATTGTATACCAATTGCGCAACCTTAAGCT-
- SacII $-\texttt{GCTCACTAGTGTCGACCTGCAGGGTACCATGGAAGCTTACTCGAGGATCCGCGGAAAGAA-$ -CGAGTGATCACAGCTGGACGTCCCATGGTACCTTCGAATGAGCTCCTAGGCGCCTTTCTT-
- -GAAGAAGAAGAAAGCCCGAAAGGAAGCTGAGTTGGCTGCCACCGCTGAGCAATA - $-\mathtt{CTTCTTCTTCTTCTGGGCTTTCCTTCGACTCAACCGACGGCGGCGGCGACTCGTTAT} -$
- -ACTAGCATAACCCCTTGGGGGCCTCTAAACGGGTCTTGAGGGGGTTTTTTGCTGAAAGGAGG- $-\mathtt{TGATCGTATTGGGGAACCCCGGAGATTTGCCCAGAACTCCCCAAAAAACGACTTTCCTCC-}$
- -AACCGCTCTTCACGCTCTTCACGC 3' -AACCGCTCTTCACGCTCTTCACGC 3' [SacII sticky end]
 -TTGGCGAGAAGTGCGAGAAGTG 5' (position #5904 in pAMG21)

[SacII sticky end]

FIG. 7





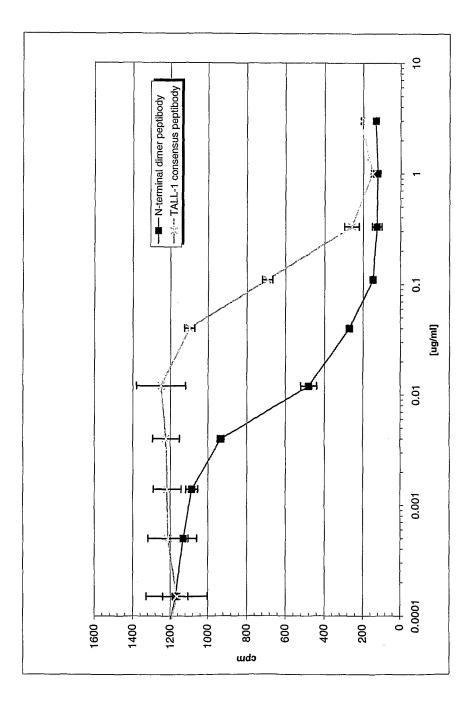
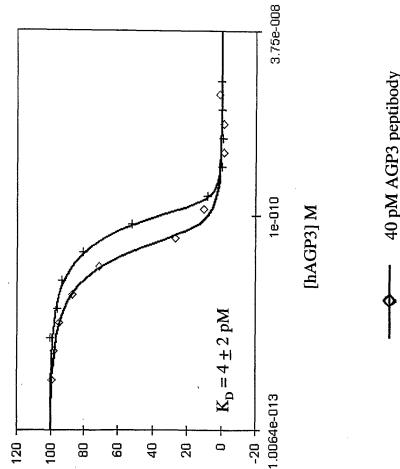


FIG. 9



Percentage of free AGP3 peptibody

◆ 40 pM AGP3 peptibody★ 100 pM AGP3 peptibody

FIG. 10A

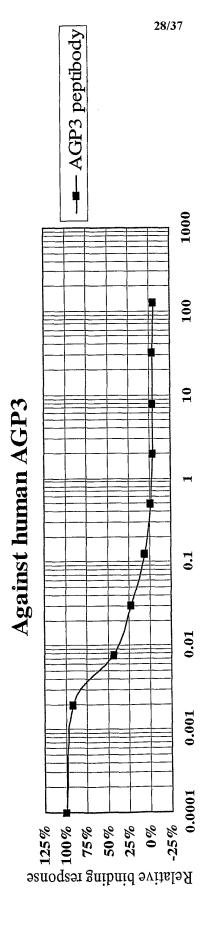


FIG. 10B

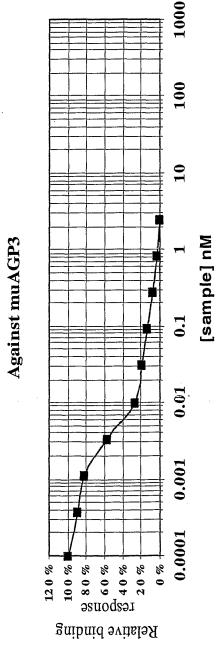
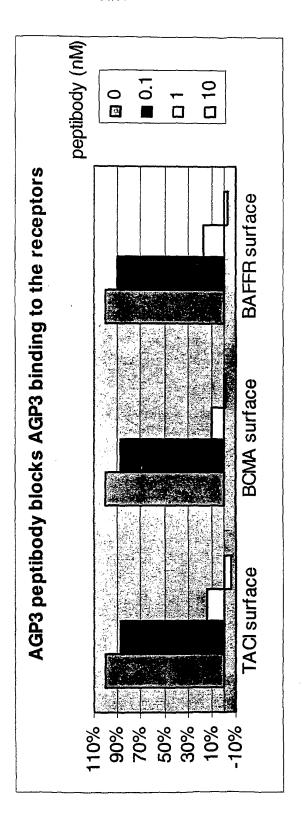


FIG. 11A



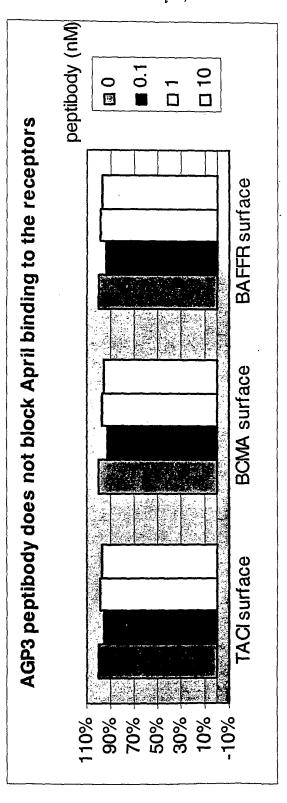
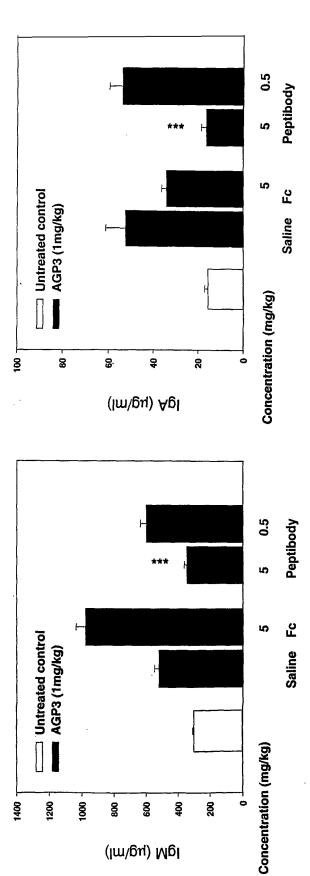
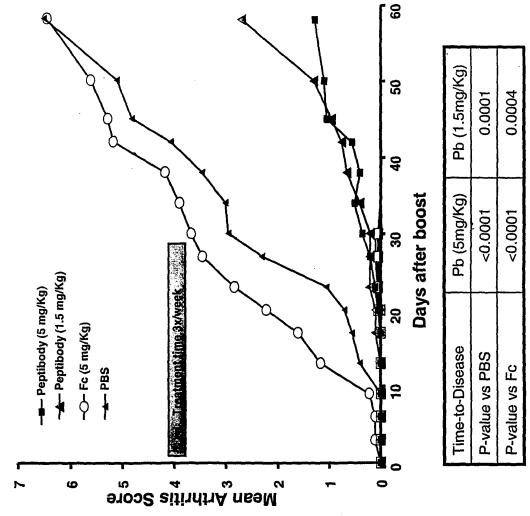


FIG. 12A

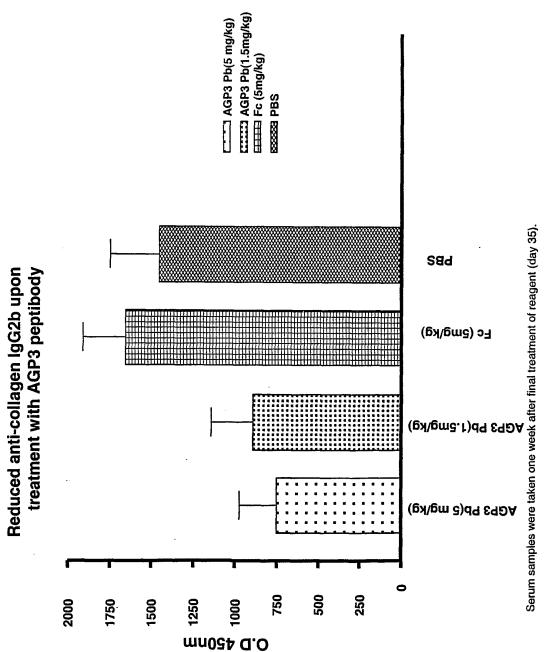






Note: p-value based on log-rank test





The graph above is representative of the IgG1, IgG3, and IgG2a isotypes as well.

0.3685

p-value vs PBS

0.0108

p-value vs PBS

P-vs Fc

0.0573

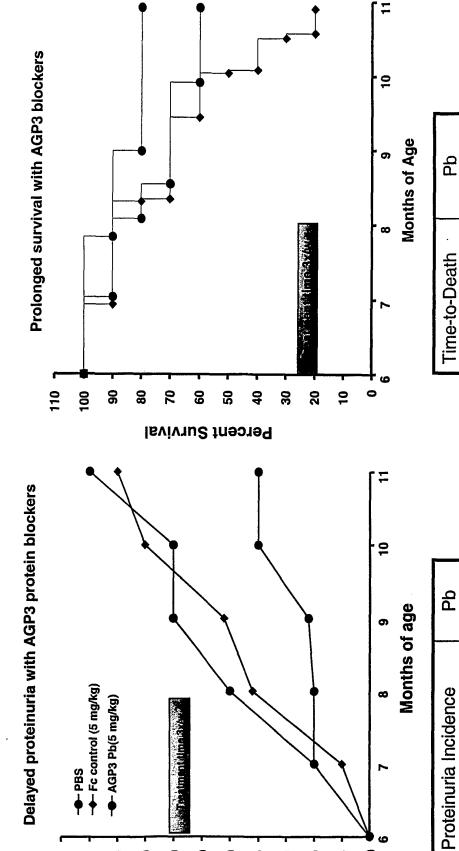
P-value based Fisher's Exact test

0.0159

35/37

Fig. 15A

Fig. 15B



Percent proteinuria (>300mg/dl)

10

P-value based log-rank test

p-value vs Fc

FIG. 16A

ATGCTTCCAGGCTGCAAGTGGGATCTTCTTATTAAGCAATGGGTATGCGATCCACTT TACGAAGGTCCGACGTTCACCCTAGAAGAATAATTCGTTACCCATACGCTAGGTGAA M L P G C K W D L L I K Q W V C D P L TCCGGTTCTGCTACTGGTGGTTCCGGCTCCACCGCAAGCTCTGGTTCAGGCAGTGCC AGGCCAAGACGATGACCACCAAGGCCGAGGTGCGTTCAGGCAGTCCGTCACGC S G S A T G G S G S T A S S G S G S A NdeI CATATGCTGCCGGGTTGTAAATGGGACCTGCTGATCAAACAGTGGGTTTGTGACCCC GTATACGACGGCCCAACATTTĀCCCTGGACGACTAGTTTGTCACCCAAACACTGGGCCGAACACTTGGGCCCAAACACTTGGGCCCAAACACTTGGGCCCCAAACACTTGGGCCCCAAACACTTGGGCCCCAAACACTTGGGCCCCAAACACTTGGGCCCCAAACACTTGGGCCCCAAACACTTGGCCCCAAACACTTGGCCCCAAACACTTGGCCCCAAACACTTGGCCCCAAACACTTGGCCCCAAACACTTGGCCCCAAACACTTGGCCCCAAACACTTGGCCCCAAACACTTGGCCCCAAACACTTGGCCCCAAACACTTGGCCCCAAACACTTGGCCCCAAACACTTGGCCCCAAACACTTGGCCCCAAACACTTGGCCCCAAACACTTGGCCCCAAACACCTGGGCCCCCAAACACCTGGGCCCCAAACACCTGGGCCCCAAACACCTGGGCCCCAAACACCTGGGCCCCAAACACCTGGGCCCCAAACACCTGGGCCCCAAACACCTGGGCCCCAAACACCTGGGCCCCAAACACCTGGGCCCCAAACACCTGGGCCCCAAACACCTGGGCCCCAAACACCTGGGCCCCAAACACCTGGGCCCCAAACACCTGGGCCCCAAACACCTGGGCCCCAAACACCTGGGCCCCAAACACCTGGGCCCCAAACACCTGGCCCCAAACACCTGGCCCCAAACACCTGGCCCCAAACACCTGGCCCCAAACACCTGGCCCCAAACACCTGGCCCCAAACACCTGGCCCCAAACACCTGGCCCCAAACACCTGGCCCCAAACACCTGGCCCCAAACACCTGGCCCCAAACACCTGGCCCCAAACACCTGCCCCAAACACCTGGCCCCAAACACCTGCCCCAAACACCTGCCCCAAACACCTGCCCCAAACACCTGCCCCAAACACCTGCCCCAAACACCTGCCCCAAACACCTGCCCCAAACACCTGCCCCAAACACCTGCCCCCAAACACCTGCCCCAAACACCTGCCCCAAACACCTGCCCCAAACACCTGCCCCAAACACCTGCCCCAAACACCTGCCCCAAACACCTGCCCCAAACACCTGCCCCCAAACACCTCGCCCCAAACACCTCGCCCCAAACACCTCGCCCCAAACACCTCGCCCCAAACACCTCGCCCCCAAACACCTCGCCCCAAACACCTCGCCCCAAACACCTCGCCCCAAACACACCTCGCCCCAAACACCTCGCCCCAAACACACCTCGCCCCAAACACACCTCGCCCCAAACACACCTCGCCCCAAACACACCTCGCCCCAAACACACCTCGCCCCAAACACACCTCGCCCCAAACACACCTCGCCCCAAACACACCTCGCCCCAAACACACCTCGCCCCAAACACACCTCGCCCCAAACACACCTCGCCCCAAACACACCTCCCCCAAACACACCTCCCCCAAACACACCTCGCCCCAAACACACCTCGCCCCAAACACACAC	+ 60 ACCT G - GACT													
TACGAAGGTCCGACGTTCACCCTAGAAGAATAATTCGTTACCCATACGCTAGGTGAA M L P G C K W D L L I K Q W V C D P L TCCGGTTCTGCTACTGGTGGTTCCGGCTCCACCGCAAGCTCTGGTTCAGGCAGTGCC 61	ACCT G - GACT+ 120 CTGA													
TCCGGTTCTGCTACTGGTGGTTCCGGCTCCACCGCAAGCTCTGGTTCAGGCAGTGCC 61++++++	GACT + 120 CTGA													
AGGCCAAGACGATGACCACCAAGGCCGAGGTGGCGTTCGAGACCAAGTCCGTCACGC S G S A T G G S G S T A S S G S G S A NdeI CATATGCTGCCGGGTTGTAAATGGGACCTGCTGATCAAACAGTGGGTTTGTGACCCC	+ 120 CTGA													
AGGCCAAGACGATGACCACCAAGGCCGAGGTGGCGTTCGAGACCAAGTCCGTCACGC S G S A T G G S G S T A S S G S G S A NdeI CATATGCTGCCGGGTTGTAAATGGGACCTGCTGATCAAACAGTGGGTTTGTGACCCCC 121	CTGA													
NdeI CATATGCTGCCGGGTTGTAAATGGGACCTGCTGATCAAACAGTGGGTTTGTGACCCC	т -													
CATATGCTGCCGGGTTGTAAATGGGACCTGCTGATCAAACAGTGGGTTTGTGACCCC														
121+														
GTATACGACGGCCCAACATTTÄCCCTGGACGACGACGTTTTGTCACGGAAAACACTGCACGA	CTG													
	+ 180 CGAC													
H M L P G C K W D L L I K Q W V C D P	L -													
SalI														
 GGTGGAGGCGGTGGGGTCGACAAAACTCACACATGTCCACCTTGTCCAGCTCCGGAA	ACTC													
+++++++														
GGGGVDKTHTCPPCPAPE	L -													
CTGGGGGGACCGTCAGTCTTCCTCTTCCCCCAAAACCCAAGGACACCCTCATGATC														
GACCCCCTGGCAGTCAGAAGGAGAAGGGGGGTTTTGGGTTCCTGTGGGAGTACTAGAGG														
LGGPSVFLFPPKPKDTLMI	S -													
CGGACCCCTGAGGTCACATGCGTGGTGGTGGACGTGAGCCACGAAGACCCTGAGGTC	'AAG													
301														
RTPEVTCVVDVSHEDPEV	к -													
TTCAACTGGTACGTGGACGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAG	GAG													
361+	+ 420													
FNWYVDGVEVHNAKTKPRE	E -													
CAGTACAACAGCACGTACCGTGTGGTCAGCGTCCTCACCGTCCTGCACCAGGACTGG														
421	+ 480													
	L -													

FIG. 16B

181	AA	TGG	CAA	.GGA	GTA	CAA	GTG	CAA	GGT	CTC	CAA	CAA	AGC	CCI	CCC	CAGC	CCC	CAT	'CGA	.GAAA	540									
401	TT	ACC	GTT	CCT	CAT	GTT	CAC	GTI	CCA	GAG	GTT	GTT	TCC	GGA	\GGG	TCG	GGG	+												
	N	G	ĸ	E	Y	ĸ	С	К	v	s	N	K	A	L	P	A	P	I	E	K	-									
541	AC	CAT	CTC	CAA	AGC	CAA	AGG	GCA	.GCC	CCG	AGA	ACC	ACA	GGT	GTA	CAC	CCT	GCC	CCC	ATCC	600									
	TG	GTA	GAG	GTT	TCG	GTT	TCC	CGI	CGG	GGC	TCI	TGG	TGT	'CCA	CAT	GTG	GGA	.CGG	GGG	TAGG	600									
	Т	I	S	K	A	K	G	Q	Р	R	E	P	Q	V	Y	Т	L	P	P	s	-									
601	CG	GGA'	TGA	GCT -+-	GAC	CAA	GAA	CCA	GGT	CAG	CCT	'GAC	CTG	CCT	'GG'I	CAA	AGG	CTT	CTA	TCCC	550									
	GC	CCT.	ACT	CGA	CTG	GTT	CTT	GGT	CCA	.GTC	GGA	CTG	GAC	GGA	CCA	GTT	TCC	GAA	GAT	AGGG	000									
	R	D	Ε	L	T	K	N	Q	V	S	L	T	С	L	V	K	G	F	Y	P	_									
661	AG	CGA	CAT	CGC	CGT	GGA	GTG	GGA	GAG	CAA	TGG	GCA	GCC	GGA	GAA	CAA	CTA	CAA	GAC	CACG	720									
																				GTGC	720									
	s	D	I	A	V	E	W	E	ន	N	G	Q	P	E	N	N	Y	K	Т	T	-									
721	CC	rcc	CGT	GCT	GGA	CTC	CGA	CGG	CTC	CTT	CTT	CCT	CTA	CAG	CAA	GCT	CAC	CGT	GÇA	CAAG	780									
																				GTTC	,00									
	P	P	V	L	D	ġ	D	G	s	F	F	L	Y	s	K	L	T	V	D	K	-									
781	AGO	CAG	GTG(GCA	GCA	GGG	GAA	CGT	СТТ 	CTC	ATG	CTC	CGT	GAT	GCA	TGA	GGC	TCT	GCA(CAAC	840									
																				GTTG	040									
	s	R	W	Q	Q	G	N	V	F	S	С	s	V	M	Н	E	A	L	Н	N	-									
							CCT									82														
							gga								_	~ <u>~</u>														
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atg ato Met Ile	tcc Ser 35	cgg Arg	acc Thr	cct Pro	gag Glu	gtc Val 40	aca Thr	tgc Cys	gtg Val	gtg Val	gtg Val 45	gac Asp	gtg Val	agc Ser	144
cac gaa His Glu 50	gac I Asp	cct Pro	gag Glu	gtc Val	aag Lys 55	ttc Phe	aac Asn	tgg Trp	tac Tyr	gtg Val 60	gac Asp	ggc Gly	gtg Val	gag Glu	192
gtg cat Val His 65	aat Asn	Ala	Lys	aca Thr 70	Lys	Pro	cgg Arg	Glu	G٦u	cag Gln	tac Tyr	aac Asn	Ser	acg Thr 80	240
tac cgt Tyr Arg	gtg Val	gtc Val	agc ser 85	gtc Val	ctc Leu	acc Thr	gtc Val	ctg Leu 90	cac His	cag Gln	gac Asp	tgg Trp	ctg Leu 95	aat Asn	288
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atc gag Ile Glu	aaa Lys 115	acc Thr	atc Ile	tcc ser	aaa Lys	gcc Ala 120	aaa Lys	ggg Gly	cag Gln	ccc Pro	cga Arg 125	gaa Glu	cca Pro	cag Gln	384

gtg val	tac Tyr 130	acc Thr	ctg Leu	ccc Pro	cca	tcc Ser 135	cgg	gat Asp	gag	ctg	acc	aaq	aac	caq	atc	4	32
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gtg Val	gac Asp	aag Lys 195	agc Ser	agg Arg	tgg Trp	cag Gln	cag Gln 200	ggg Gly	aac Asn	gtc Val	ttc Phe	tca Ser 205	tgc Cys	tcc Ser	gtg Val	6	24
atg Met	cat His 210	gag Glu	gct Ala	ctg Leu	cac His	aac Asn 215	cac His	tac Tyr	acg Thr	cag Gln	aag Lys 220	agc Ser	ctc Leu	tcc Ser	ctg Leu	6	72
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Gly	Glу	Pro	Ser 20	Val	Phe	Leu	Phe	Pro 25	Pro	Lys	Pro	Lys	Asp 30	Thr	Leu		
Met	Ile	Ser 35	Arg	Thr	Pro	Glu	Val 40	Thr	Cys	val	۷al	Va1 45	Asp	Val	Ser		
His	Glu 50	Asp	Pro	Glu	۷al	Lys 55	Phe	Asn	Тгр	Tyr	Val 60	Asp	Gly	val	Glu		
Va1 65	His	Asn	Ala	Lys	Thr 70	Lys	Pro	Arg	Glu	Glu 75	Gln	Tyr	Asn	Ser	Thr 80		
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Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val
145 150 155
Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro 165 170 175
Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr 180 \hspace{1cm} 185 \hspace{1cm} 190 \hspace{1cm}
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                                                                                              49
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1 5 10
                                                                                                         49
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1 5 10
                                                                                                         49
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Phe His His Gly Gly Gly Gly Gly
20
                                                                                                         74
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          10
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Page 5

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1 5 10 15
                                                                                                     49
gac ccg ctg ggt gga ggc ggt ggg g
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20
                                                                                                            74
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Thr Ser Ser Gly Gly Gly Gly Gly 20
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1 10 15
                                                                                                            49
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                                                                                                            74
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His Gly Leu Gly Gly Gly Gly Gly
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1 5 10 15
                                                                                                     49
ccg tct ccg ggt gga ggc ggt ggg g
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1 5 10
                                                                                                          49
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Glu Phe Phe Gly Gly Gly Gly Gly 20
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Page 10

20

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Artificial Sequence

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Preferred TALL-1 modulating domain

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Asp Pro Leu
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Thr or Ile
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Thr or Ile
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Amino acid residue

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Absent or amino acid residue
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Amino acid residue
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        MISC_FEATURE
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         (13)..(13)
        Amino acid residue
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         (14)..(14)
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        Absent or amino acid residue
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Absent or amino acid residue
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Xaa Xaa
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        107
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        18
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Xaa Xaa Xaa Cys Xaa Xaa Xaa Asp Xaa Leu Xaa Lys Xaa Cys Xaa Xaa

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1
Xaa Xaa
<210>
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<211>
<212>
           4
           PRT
<213>
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            (2)...(2)
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           (4)..(4)
Thr or Ile
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<400>
           108
Asp Xaa Leu Xaa
<210>
           109
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           14
<212>
           PRT
<213>
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           Modulator of TALL-1
<220>
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           (1)..(1)
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           MISC_FEATURE
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           (3)..(3)
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Corrected sequence listing- final.txt
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        (7)..(7)
        Amino acid residue (Leu preferred)
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        MISC_FEATURE
<222>
        (9)..(9)
<223>
        Thr or Ile (Thr preferred)
<220>
<221>
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        MISC_FEATURE
        (10)..(10)
<223>
        Lys, Arg, or His (Lys preferred)
<220>
<221>
        MISC_FEATURE
<222>
        (12)..(12)
        Cys, neutral hydrophobic residue, or basic residue (Trp, Cys, or Arg preferred, and only one of residues 12, 13, or 14 may be Cys)
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        MISC_FEATURE
        (13)..(13)
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        Cys, neutral hydrophobic residue, or absent (Val preferred, and only one of residues 12, 13, or 14 may be Cys)
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        (14)..(14)
        Absent or amino acid residue (only one of residues 12, 13, or 14
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        may be Cys)
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Xaa Xaa Xaa Lys Xaa Asp Xaa Leu Xaa Xaa Gln Xaa Xaa Xaa
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<211>
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        PRT
        Artificial Sequence
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        Modulator of TALL-1
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        110
Pro Phe Pro Trp Glu
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        111
        248
<211>
<212>
        PRT
<213>
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<220>
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Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro 40 45

Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val 50 60

Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val 65 70 75 80

Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln 85 90 95

Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln 100 105

Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala 115 120 125

Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro 130 140

Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr 145 150 155 160

Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser 165 170 175

Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr 180 185 190

Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr 195 200 205

Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe 210 220

Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys 225 230 235

Ser Leu Ser Leu Ser Pro Gly Lys

<210> 112

<211> <212>

Artificial Sequence

TALL-1 inhibitory peptibody TALL-1-8-2-a

Met Trp Gly Ala Cys Trp Pro Phe Pro Trp Glu Cys Phe Lys Glu Gly
1 10 15

Gly Gly Gly Val Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala 20 25 30

Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro
35 40 45

Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val 50 60

Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val 65 70 75 80

Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln 85 90 95

Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln 100 105

Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala 115 120

Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro 130 135 140

Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr 145 150 155 160

Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser 165 170

Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr 180 185 190

Corrected sequence listing- final.txt
Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr
195 200 205

Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe 210 215 220

Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys 225 230 235 240

Ser Leu Ser Leu Ser Pro Gly Lys 245

113 248

PRT

Artificial Sequence

<220>

TALL-1 inhibitory peptibody TALL-1-8-4-a

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Met Val Pro Phe Cys Asp Leu Leu Thr Lys His Cys Phe Glu Ala Gly
1 10 15

Gly Gly Gly Val Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala 20 25 30

Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro 35 40 45

Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val 50 60

Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val 75 80

Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln 85 90 95

Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln
100 105

Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala 115 120 125

Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro
130 140

Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Page 51

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Corrected sequence listing- final.txt 155 160
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Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser 165 170 175

Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr 180 185 190

Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr 195 200 205

Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe 210 220

Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys 225 230 235 240

Ser Leu Ser Leu Ser Pro Gly Lys 245

<210> 114

<212> PRT

Artificial Sequence

TALL-1 inhibitory peptibody TALL-1-12-4-a

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Met Gly Ser Arg Cys Lys Tyr Lys Trp Asp Val Leu Thr Lys Gln Cys
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Phe His His Gly Gly Gly Gly Val Asp Lys Thr His Thr Cys Pro 20 25 30

Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe 35 40

Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val 50 60

Thr Cys Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe 65 70 75 80

Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro

Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr 100 105 110 Page 52

Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val 115 120 125

Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala 130 135 140

Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg 145 150 155 160

Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly
165 170 175

Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro 180 185 190

Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser 195 200 205

Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln 210 215 220

Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His 225 230 235

Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys 245 250

<210> <211> 115

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PRT

Artificial Sequence

<220>

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Met Leu Pro Gly Cys Lys Trp Asp Leu Leu Ile Lys Gln Trp Val Cys $1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15$

Asp Pro Leu Gly Gly Gly Gly Val Asp Lys Thr His Thr Cys Pro 25 30

Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe 35 40 45

Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val 50 60

Thr Cys Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe 65 70 75 80

Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro 85 90 95

Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr 100 105

Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val 115 120 125

Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala 130 135 140

Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg 145 150 155 160

Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly
165 170 175

Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro 180 185 190

Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser 195 200 205

Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln 210 220

Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His 225 230 235 240

Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys 245

<210> <211> <212> 116 252

PRT

Artificial Sequence

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Corrected sequence listing- final.txt
Thr Ser Ser Gly Gly Gly Gly Val Asp Lys Thr His Thr Cys Pro
20 25 30

Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe 35 40

Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val

Thr Cys Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe 65 70 75 80

Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro

Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr 100 105 110

Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val 115 120 125

Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala 130 135 140

Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg 145 150 155 160

Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly 165 170 175

Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro 180 185 190

Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser 195 200 205

Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln 210 220

Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His 225 235 240

Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys 245

<210> <211> <212> 117 252

	_	Corrected	sequence	11Sting-	rinai.txt
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Met Ser Asp Asp Cys Met Tyr Asp Gln Leu Thr Arg Met Phe Ile Cys $1 \hspace{1cm} 15$

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Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe 35 40 45

Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val 50 60

Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe 65 70 75 80

Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro

Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr $100 \hspace{1cm} 105 \hspace{1cm} 110$

Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val 115 120 125

Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala 130 135 140

Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg 145 150 155 160

Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly
165 170 175

Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro 180 185 190

Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser 195 200 205

Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln 210 215 220

Corrected sequence listing- final.txt
Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His
225 230 235 240

Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys 245

<210> <211> <212>

PRT Artificial Sequence

TALL-1 inhibitory peptibody TALL-1-12-9-a

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Gln Phe Asn Gly Gly Gly Gly Gly Val Asp Lys Thr His Thr Cys Pro 20 25 30

Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe 35 40

Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val 50 60

Thr Cys Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe 65 70 75 80

Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro

Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr 100 105 110

Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val 115 120 125

Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala 130 140

Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg 145 150 155

Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly 165 170 175

Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro

Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser 195 200 205

Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln 210 215 220

Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His 225 230 235 240

Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys 245

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180

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Met Phe His Asp Cys Lys Tyr Asp Leu Leu Thr Arg Gln Met Val Cys 10 15

His Gly Leu Gly Gly Gly Gly Val Asp Lys Thr His Thr Cys Pro

Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe 35 40

Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val 50 60

Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe 65 70 75 80

Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro

Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr 100 105 110

Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val 115 120 125

Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala 130 135 140 Page 58

Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg 145 150 155 160

Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly 165 170

Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro 180 185 190

Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser 195 200 205

Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln 210 215 220

Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His 225 230 235 240

Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys 245

120 252 <210>

Artificial Sequence

<220> <223> TALL-1 inhibitory peptibody TALL-1-12-11-a

<400>

Met Arg Asn His Cys Phe Trp Asp His Leu Leu Lys Gln Asp Ile Cys 1 10 15

Pro Ser Pro Gly Gly Gly Gly Val Asp Lys Thr His Thr Cys Pro 20 25 30

Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe 35 40 45

Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val

Thr Cys Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe 65 70 75 80

Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro

Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr 100 105 110

Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val 115 120 125

Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala 130 135 140

Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg 145 150 155 160

Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly 165 170 175

Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro 180 185 190

Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser 195 200 205

Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln 210 220

Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His 225 230 235

Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys 245

<210> <211> 121

252

Artificial Sequence

<220> <223> TALL-1 inhibitory peptibody TALL-1-12-14-a

<400> 121

Met Ala Asn Gln Cys Trp Trp Asp Ser Leu Thr Lys Lys Asn Val Cys $1 \hspace{1cm} 15$

Glu Phe Phe Gly Gly Gly Gly Val Asp Lys Thr His Thr Cys Pro 20 25 30

Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe 35 40 45

Corrected sequence listing- final.txt
Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val
50 55 60

Thr Cys Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe 65 70 75 80

Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro

Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr $100 \hspace{1cm} 105 \hspace{1cm} 110$

Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val 115 120 125

Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala 130 135 140

Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg 145 150 155 160

Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly
165 170 175

Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro 180 185

Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser 195 200 205

Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln 210 215 220

Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His 225 235 240

Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys 245

122 252 <210>

<211>

PRT

Artificial Sequence <213>

<220>

<223> TALL-1 inhibitory peptibody consensus sequence

<400>

Met Phe His Asp Cys Lys Trp Asp Leu Leu Thr Lys Gln Trp Val Cys

1				5		Corr	ecte	d se	quen 10	ce 1	isti	ng-	fina	1.tx 15	t
His	Gly	Leu	Gly 20	Gly	Gly	Gly	Gly	Val 25	Asp	Lys	Thr	His	Thr 30	Cys	Pro
Pro	Cys	Pro 35	Ala	Pro	Glu	Leu	Leu 40	Glу	Gly	Pro	Ser	Va1 45	Phe	Leu	Phe
Pro	Pro 50	Lys	Pro	Lys	Asp	Thr 55	Leu	Met	Ile	Ser	Arg 60	Thr	Pro	Glu	۷al
Thr 65	Cys	val	val	val	Asp 70	val	Ser	His	Glu	Asp 75	Pro	Glu	Val	Lys	Phe 80
Asn	Trp	Туг	val	Asp 85	Gly	Val	Glu	Val	ніs 90	Asn	Ala	Lys	Thr	Lys 95	Pro
Arg	Glu	Glu	Gln 100	Tyr	Asn	Ser	Thr	Tyr 105	Arg	val	val	Ser	Val 110	Leu	Thr
val	Leu	His 115	Gln	Asp	Trp	Leu	Asn 120	Gly	Lys	Glu	туг	Lys 125	Cys	Lys	val
Ser	Asn 130	Lys	Ala	Leu	Pro	Ala 135	Pro	Ile	Glu	Lys	Thr 140	Ile	Ser	Lys	Ala
Lys 145	Gly	Gln	Pro	Arg	Glu 150	Pro	Gln	٧a٦	Tyr	Thr 155	Leu	Pro	Pro	Ser	Arg 160
Asp	Glu	Leu	Thr	Lys 165	Asn	Gln	Val	Ser	Leu 170	Thr	Cys	Leu	val	Lys 175	Glу
Phe	Tyr	Pro	Ser 180	Asp	Ile	Ala	٧al	Glu 185	Тгр	Glu	Ser	Asn	Gly 190	Gln	Pro
Glu	Asn	Asn 195	Tyr	Lys	Thr	Thr	Pro 200	Pro	val	Leu	Asp	ser 205	Asp	Glу	Ser
Phe	Phe 210	Leu	Туг	ser	Lys	Leu 215	Thr	val	Asp	Lys	Ser 220	Arg	Тгр	Gln	Gln
Gly 225	Asn	val	Phe	Ser	Cys 230	Ser	val	Met	His	G]u 235	Ala	Leu	His	Asn	ніs 240
Tyr	Thr	Gln	Lys	Ser 245	Leu	Ser	Leu	Ser	Pro 250	Glу	Lys				

<210>

Artificial Sequence

<220>

<223> TALL-1 inhibitory peptibody 12-3 tandem dimer

<400>

Met Leu Pro Gly Cys Lys Trp Asp Leu Leu Ile Lys Gln Trp Val Cys $1 \hspace{1cm} 15$

Asp Pro Leu Gly Ser Gly Ser Ala Thr Gly Gly Ser Gly Ser Thr Ala 20 25 30

Ser Ser Gly Ser Gly Ser Ala Thr His Met Leu Pro Gly Cys Lys Trp 35 40 45

Asp Leu Leu Ile Lys Gln Trp Val Cys Asp Pro Leu Gly Gly Gly Gly 50 60

Gly Val Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu 65 70 75 80

Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr 85 90 95

Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val 100 105 110

Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val 115 120 125

Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser 130 140

Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu 145 150 155 160

Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala 165 170 175

Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro 180 185 190

Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln 195 200 205

Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Page 63

```
Corrected sequence listing- final.txt 215 220
```

Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr 225 230 235 240

Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu 245 250 255

Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser 260 265 270

Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser 275 280 285

Leu Ser Pro Gly Lys 290

<210> 124

210

<211> 293

PRT <213> Artificial Sequence

<220> TALL-1 inhibitory peptibody consensus tandem dimer <223> <400>

Met Phe His Asp Cys Lys Trp Asp Leu Leu Thr Lys Gln Trp Val Cys 1 5 10 15

His Gly Leu Gly Ser Gly Ser Ala Thr Gly Gly Ser Gly Ser Thr Ala 20 25 30

Ser Ser Gly Ser Gly Ser Ala Thr His Met Phe His Asp Cys Lys Trp $\frac{35}{40}$

Asp Leu Leu Thr Lys Gln Trp Val Cys His Gly Leu Gly Gly Gly 50 60

Gly Val Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu 65 70 75 80

Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr 85 90 95

Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Asp Val 100 105 110

Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val 115 120 125 Page 64

Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser 130 135 140

Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu 145 150 160

Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala 165 170 175

Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro 180 185 190

Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln 195 200 205

Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala 210 220

Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr 225 230 240

Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu 245 250

Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser 260 265 270

Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser 275 280 285

Leu Ser Pro Gly Lys 290

<210> 125

<211>

Artificial Sequence

<220>

Modulator of TALL-1

<220>

<221> MISC_FEATURE

<222> (1)..(1)

Absent or amino acid residue (one of residues 1, 2, or 3 preferably Cys when one of residues 12, 13, or 14 is Cys, and only one of residues 1, 2, or 3 may be Cys)

<220>

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Corrected sequence listing- final.txt
<221>
         MISC_FEATURE
<222>
         (2)..(2)
         Absent or amino acid residue (one of residues 1, 2, or 3
<223>
         preferably Cys when one of residues 12, 13, or 14 is Cys, and only one of residues 1, 2, or 3 may be Cys)
<220>
<221>
         MISC_FEATURE
<222>
         (3)..(3)
         Absent or amino acid residue (one of residues 1, 2, or 3 preferably Cys when one of residues 12, 13, or 14 is Cys, and only one of residues 1, 2, or 3 may be Cys)
<223>
<220>
<221>
<222>
         MISC_FEATURE
         (7)..(7)
<223>
         Amino acid residue (Leu preferred)
<220>
<221>
         MISC_FEATURE
<222>
         (9)..(9)
<223>
         Thr or Ile (Thr preferred)
<220>
<221>
         MISC_FEATURE
<222>
         (12)..(12)
         Cys, neutral hydrophobic residue, or basic residue (Trp, Cys, or Arg preferred, and only one of residues 12, 13, or 14 may be Cys)
<223>
<220>
<221>
<222>
         MISC_FEATURE
         (13)..(13)
         Cys, neutral hydrophobic residue, or absent (val preferred, and only one of residues 12, 13, or 14 may be Cys))
<223>
<220>
<221>
         MISC_FEATURE
<222>
         (14)..(14)
<223>
         Absent or amino acid residue (only one of residues 12, 13, or 14
         may be Cys)
<400>
         125
Xaa Xaa Xaa Lys Trp Asp Xaa Leu Xaa Lys Gln Xaa Xaa Xaa
<210>
         126
<211>
<212>
         18
         PRT
         Artificial Sequence
<213>
<220>
<223>
         Preferred TALL-1 modulating domain
<400>
Tyr Lys Gly Arg Gln Met Trp Asp Ile Leu Thr Arg Ser Trp val val
```

Ser Leu

```
<210>
<211>
        127
18
PRT
        Artificial Sequence
<220>
<223>
       Preferred TALL-1 modulating domain
<400>
Gln Asp Val Gly Leu Trp Trp Asp Ile Leu Thr Arg Ala Trp Met Pro 1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15
Asn Ile
<210>
<211>
<212>
<213>
        128
18
         Artificial Sequence
<220>
<223>
         Preferred TALL-1 modulating domain
<400>
Gln Asn Ala Gln Arg Val Trp Asp Leu Leu Ile Arg Thr Trp Val Tyr 1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15
Pro Gln
<210>
         129
<211>
         18
<212>
        PRT
         Artificial Sequence
<220>
<223>
       Preferred TALL-1 modulating domain
<400>
Gly Trp Asn Glu Ala Trp Trp Asp Glu Leu Thr Lys Ile Trp Val Leu 1 \ 5 \ 10 \ 15
Glu Gln
<210>
         130
<211>
<212>
<213>
         18
         PRT
         Artificial Sequence
<220>
       Preferred TALL-1 modulating domain
                                                   Page 67
```

```
Corrected sequence listing- final.txt
```

<400> 130

Arg Ile Thr Cys Asp Thr Trp Asp Ser Leu Ile Lys Lys Cys Val Pro $1 \hspace{1.5cm} \text{S} \hspace{1.5cm} \text{10} \hspace{1.5cm} \text{15}$

Gln Ser

```
<210>
<211>
<212>
                 131
```

18

<213> Artificial Sequence

<223> Preferred TALL-1 modulating domain

<400> 131

Gly Ala Ile Met Gln Phe Trp Asp Ser Leu Thr Lys Thr Trp Leu Arg $1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15$

Gln Ser

```
<210>
<211>
              132
18
```

<212> Artificial Sequence

<220>

<223> Preferred TALL-1 modulating domain

<400> 132

Trp Leu His Ser Gly Trp Trp Asp Pro Leu Thr Lys His Trp Leu Gln 10 15

Lys Val

<210> 133

18

<212> PRT

<213> Artificial Sequence

<220>

<223> Preferred TALL-1 modulating domain

<400>

Phe Arg

```
<210>
       134
<211>
      18
<212>
      PRT
<213>
       Artificial Sequence
<220>
<223>
      Preferred TALL-1 modulating domain
<400>
      134
Gly Val Trp Phe Trp Trp Phe Asp Pro Leu Thr Lys Gln Trp Thr Gln 10 15
Ala Gly
<210>
<211>
      18
       135
<212>
       PRT
       Artificial Sequence
<220>
<223>
       Preferred TALL-1 modulating domain
<400>
Met Gln Cys Lys Gly Tyr Tyr Asp Ile Leu Thr Lys Trp Cys Val Thr 10 \ 15
Asn Gly
<210>
       136
       18
<212>
<213>
       PRT
      Artificial Sequence
<220>
<223>
       Preferred TALL-1 modulating domain
<400>
Leu Trp Ser Lys Glu Val Trp Asp Ile Leu Thr Lys Ser Trp Val Ser 10 15
Gln Ala
<210>
       137
<211><212><212><213>
       18
       PRT
       Artificial Sequence
<220>
      Preferred TALL-1 modulating domain
                                          Page 69
```

```
Corrected sequence listing- final.txt
```

<400> 137

Lys Ala Ala Gly Trp Trp Phe Asp Trp Leu Thr Lys Val Trp Val Pro $1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15$

Ala Pro

<210> <211> 138

18 <212>

<213> Artificial Sequence

<223> Preferred TALL-1 modulating domain

<400> 138

Ala Tyr Gln Thr Trp Phe Trp Asp Ser Leu Thr Arg Leu Trp Leu Ser $1 \hspace{1cm} 15$

Thr Thr

<210> 139

<211> 18 <212>

PRT Artificial Sequence <213>

<220>

<223> Preferred TALL-1 modulating domain

<400> 139

Ser Gly Gln His Phe Trp Trp Asp Leu Leu Thr Arg Ser Trp Thr Pro $1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15$

Ser Thr

<210> 140

<211> 18

PRT

Artificial Sequence

<220>

<223> Preferred TALL-1 modulating domain

<400> 140

Leu Gly Val Gly Gln Lys Trp Asp Pro Leu Thr Lys Gln Trp Val Ser 10 15

Arg Gly

```
<210>
<211>
        141
        18
<213>
       Artificial Sequence
<220>
<223>
       Preferred TALL-1 modulating domain
<400> 141
Val Gly Lys Met Cys Gln Trp Asp Pro Leu Ile Lys Arg Thr Val Cys 1 \hspace{1cm} 15
val Gly
<210>
        142
<211>
        18
<212>
        PRT
       Artificial Sequence
<220>
<223>
       Preferred TALL-1 modulating domain
<400>
Cys Arg Gln Gly Ala Lys Phe Asp Leu Leu Thr Lys Gln Cys Leu Leu 1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15
Gly Arg
<210>
       143
18
        143
<211>
<212>
       PRT
        Artificial Sequence
<220>
       Preferred TALL-1 modulating domain
<223>
<400> 143
Gly Gln Ala Ile Arg His Trp Asp Val Leu Thr Lys Gln Trp Val Asp 1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15
Ser Gln
<210>
        144
<211>
<212>
        18
        PRT
       Artificial Sequence
<220>
       Preferred TALL-1 modulating domain
                                               Page 71
```

```
Corrected sequence listing- final.txt
```

<400> 144

Arg Gly Pro Cys Gly Ser Trp Asp Leu Leu Thr Lys His Cys Leu Asp $1 \hspace{1.5cm} 15$

Ser Gln

```
<210>
<211>
          145
          18
```

<212>

<213> Artificial Sequence

<223> Preferred TALL-1 modulating domain

<400>

Trp Gln Trp Lys Gln Gln Trp Asp Leu Leu Thr Lys Gln Met Val Trp 1 5 10 15

Val Gly

<210> 146

<211> 18

<212> PRT Artificial Sequence <213>

<220>

<223> Preferred TALL-1 modulating domain

<400> 146

Pro Ile Thr Ile Cys Arg Lys Asp Leu Leu Thr Lys Gln Val Val Cys $1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15$

Leu Asp

147 <210>

<211> 18

PRT

Artificial Sequence

<220>

<223> Preferred TALL-1 modulating domain

<400>

Lys Thr Cys Asn Gly Lys Trp Asp Leu Leu Thr Lys Gln Cys Leu Gln 10 15

Gln Ala

```
<210>
        148
<211>
        18
<213>
       Artificial Sequence
<220>
<223> Preferred TALL-1 modulating domain
<400>
Lys Cys Leu Lys Gly Lys Trp Asp Leu Leu Thr Lys Gln Cys Val Thr 1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15
Glu Val
<210>
        149
<211>
<212>
        18
        PRT
       Artificial Sequence
<220>
<223>
        Preferred TALL-1 modulating domain
<400>
Arg Cys Trp Asn Gly Lys Trp Asp Leu Leu Thr Lys Gln Cys Ile His 1 \phantom{000}5\phantom{000} 15
Pro Trp
<210>
        150
<211>
        18
<212>
        PRT
       Artificial Sequence
<213>
<220>
       Preferred TALL-1 modulating domain
<223>
<400> 150
Asn Arg Asp Met Arg Lys Trp Asp Pro Leu Ile Lys Gln Trp Ile Val 1 	 5 	 10 	 15
Arg Pro
<210>
<211>
<212>
<213>
        151
        18
        PRT
       Artificial Sequence
<220>
<223> Preferred TALL-1 modulating domain
                                               Page 73
```

```
Corrected sequence listing- final.txt
<400> 151
Gln Ala Ala Ala Thr Trp Asp Leu Leu Thr Lys Gln Trp Leu Val 1 	 5 	 10 	 15
Pro Pro
<210>
<211>
<212>
       152
18
       Artificial Sequence
<223>
      Preferred TALL-1 modulating domain
<400>
      152
Pro Glu Gly Gly Pro Lys Trp Asp Pro Leu Thr Lys Gln Phe Leu Pro 1 \ 5 \ 10 \ 15
Pro Val
<210>
<211>
       153
18
<212>
      Artificial Sequence
<220>
<223>
      Preferred TALL-1 modulating domain
<400> 153
Gln Thr Pro Gln Lys Lys Trp Asp Leu Leu Thr Lys Gln Trp Phe Thr 10 15
Arg Asn
        154
<210>
<211>
       18
<212>
       PRT
      Artificial Sequence
<213>
<220>
<223>
       Preferred TALL-1 modulating domain
<400>
Ile Gly Ser Pro Cys Lys Trp Asp Leu Leu Thr Lys Gln Met Ile Cys 1 \hspace{1cm} 15
Gln Thr
```

```
<210>
<211>
       155
18
<213> Artificial Sequence
<220>
<223> Preferred TALL-1 modulating domain
<400>
Cys Thr Ala Ala Gly Lys Trp Asp Leu Leu Thr Lys Gln Cys Ile Gln 10 15
Glu Lys
<210>
<211>
       156
18
<212>
       PRT
      Artificial Sequence
<220>
      Preferred TALL-1 modulating domain
<223>
<400>
Val Ser Gln Cys Met Lys Trp Asp Leu Leu Thr Lys Gln Cys Leu Gln 10 15
Gly Trp
<210>
        157
<211>
        18
<212>
       PRT
      PRT
Artificial Sequence
<220>
<223> Preferred TALL-1 modulating domain
<400> 157
Val Trp Gly Thr Trp Lys Trp Asp Leu Leu Thr Lys Gln Tyr Leu Pro 1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15
Pro Gln
<210>
        158
<211>
        18
<212>
        PRT
        Artificial Sequence
<220> <223> Preferred TALL-1 modulating domain
                                             Page 75
```

Asn Met

```
Corrected sequence listing- final.txt
<400> 158
Gly Trp Trp Glu Met Lys Trp Asp Leu Leu Thr Lys Gln Trp Tyr Arg 1 \hspace{1cm} 5 \hspace{1cm} 15
Pro Gln
<210>
<211>
        159
        18
<212>
<213>
        Artificial Sequence
<223>
        Preferred TALL-1 modulating domain
<400>
       159
Thr Ala Gln Val Ser Lys Trp Asp Leu Leu Thr Lys Gln Trp Leu Pro 1 \hspace{1cm} 10 \hspace{1cm} 15
Leu Ala
<210>
        160
<211>
        18
<212>
        PRT
        Artificial Sequence
<213>
<220>
<223>
        Preferred TALL-1 modulating domain
<400>
        160
Gln Leu Trp Gly Thr Lys Trp Asp Leu Leu Thr Lys Gln Tyr Ile Gln 10 15
Ile Met
<210>
        161
<211>
        18
        PRT
       Artificial Sequence
<220>
<223>
        Preferred TALL-1 modulating domain
<400>
Trp Ala Thr Ser Gln Lys Trp Asp Leu Leu Thr Lys Gln Trp Val Gln 10 15
```

```
<210>
<211>
<212>
        162
        18
        PRT
<213>
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<220>
<223>
       Preferred TALL-1 modulating domain
<400>
       162
Gln Arg Gln Cys Ala Lys Trp Asp Leu Leu Thr Lys Gln Cys Val Leu 1 \hspace{1cm} 15
Phe Tyr
<210>
        163
<211>
<212>
        18
       PRT
       Artificial Sequence
<220>
<223>
       Preferred TALL-1 modulating domain
<400>
Lys Thr Thr Asp Cys Lys Trp Asp Leu Leu Thr Lys Gln Arg Ile Cys 1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15
Gln Val
<210>
        164
<211>
        18
<212>
       PRT
       Artificial Sequence
<213>
<220>
       Preferred TALL-1 modulating domain
<223>
<400> 164
Leu Leu Cys Gln Gly Lys Trp Asp Leu Leu Thr Lys Gln Cys Leu Lys 1 \hspace{1cm} 15
Leu Arg
        165
<210>
<211>
        18
<212>
<213>
        PRT
        Artificial Sequence
<220>
<223> Preferred TALL-1 modulating domain
                                              Page 77
```

```
Corrected sequence listing- final.txt
Leu Met Trp Phe Trp Lys Trp Asp Leu Leu Thr Lys Gln Leu Val Pro 1 \hspace{1cm} 5 \hspace{1cm} 15
```

Thr Phe

<400> 165

```
166
<210>
<211>
<212>
        18
        PRT
<213>
        Artificial Sequence
<220>
```

<223> Preferred TALL-1 modulating domain

<400>

Gln Thr Trp Ala Trp Lys Trp Asp Leu Leu Thr Lys Gln Trp Ile Gly $1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15$

Pro Met

```
<210>
         167
<211>
         18
<212>
        PRT
        Artificial Sequence
<213>
<220>
<223> Preferred TALL-1 modulating domain
<400> 167
Asn Lys Glu Leu Lys Trp Asp Leu Leu Thr Lys Gln Cys Arg Gly 1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15
```

168 <210> 18 PRT Artificial Sequence

<220> Preferred TALL-1 modulating domain <223>

<400>

Arg Ser

Gly Gln Lys Asp Leu Lys Trp Asp Leu Leu Thr Lys Gln Tyr Val Arg $1 \hspace{1.5cm} 5 \hspace{1.5cm} 10 \hspace{1.5cm} 15$

Gln Ser

```
<210>
<211>
       169
       18
<212>
       PRT
<213>
      Artificial Sequence
<220>
<223>
      Preferred TALL-1 modulating domain
<400>
     169
Pro Lys Pro Cys Gln Lys Trp Asp Leu Leu Thr Lys Gln Cys Leu Gly 1 \hspace{1cm} 15
Ser Val
<210>
       170
<211>
<212>
       18
<213>
      Artificial Sequence
<220>
      Preferred TALL-1 modulating domain
<223>
<400>
Gly Gln Ile Gly Trp Lys Trp Asp Leu Leu Thr Lys Gln Trp Ile Gln 10 15
Thr Arg
<210>
      171
<211>
      18
<212>
      PRT
      Artificial Sequence
<213>
<220>
     Preferred TALL-1 modulating domain
<223>
<400> 171
Pro Gln
<210>
      172
<211>
      18
<212>
      PRT
<213>
      Artificial Sequence
<220>
     Preferred TALL-1 modulating domain
                                      Page 79
```

Asp Val

```
Corrected sequence listing- final.txt
<400> 172
Gln Glu Trp Glu Tyr Lys Trp Asp Leu Leu Thr Lys Gln Trp Gly Trp 1 \hspace{1cm} 5 \hspace{1cm} 15
Leu Arg
<210>
<211>
        173
        18
        Artificial Sequence
<223>
        Preferred TALL-1 modulating domain
<400>
        173
His Trp Asp Ser Trp Lys Trp Asp Leu Leu Thr Lys Gln Trp Val Val 1 	 5 	 10 	 15
Gln Ala
<210>
<211>
        174
        18
<212>
<213>
       Artificial Sequence
<220>
<223>
       Preferred TALL-1 modulating domain
<400> 174
Thr Arg Pro Leu Gln Lys Trp Asp Leu Leu Thr Lys Gln Trp Leu Arg 1 \hspace{1cm} 10 \hspace{1cm} 15
Val Gly
        175
<210>
<211>
        18
<212>
        PRT
       Artificial Sequence
<213>
<220>
<223>
        Preferred TALL-1 modulating domain
<400>
```

Ser Asp Gln Trp Gln Lys Trp Asp Leu Leu Thr Lys Gln Trp Phe Trp $1 \hspace{1cm} 15$

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```
<210>
<211>
        176
        18
<212>
<213>
        Artificial Sequence
<220>
<223>
       Preferred TALL-1 modulating domain
<400>
Gln Gln Thr Phe Met Lys Trp Asp Leu Leu Thr Lys Gln Trp Ile Arg 1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15
Arg His
<210>
<211>
<212>
        177
        18
        Artificial Sequence
<220>
<223>
        Preferred TALL-1 modulating domain
<400>
Gln Gly Glu Cys Arg Lys Trp Asp Leu Leu Thr Lys Gln Cys Phe Pro 1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15
Gly Gln
<210>
        178
<211>
        18
<212>
        PRT
        Artificial Sequence
<213>
<220>
<223>
       Preferred TALL-1 modulating domain
<400> 178
Gly Gln Met Gly Trp Arg Trp Asp Pro Leu Ile Lys Met Cys Leu Gly 1 \hspace{1cm} 15
Pro Ser
<210>
        179
<211>
        18
<212><213>
        PRT
        Artificial Sequence
<220>
       Preferred TALL-1 modulating domain
                                                 Page 81
```

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Corrected sequence listing- final.txt
<400> 179
Gln Leu Asp Gly Cys Lys Trp Asp Leu Leu Thr Lys Gln Lys Val Cys 1 \hspace{1cm} 5 \hspace{1cm} 15
Ile Pro
<210>
         180
<211>
<212>
         18
        PRT
<213>
        Artificial Sequence
<220>
<223>
        Preferred TALL-1 modulating domain
<400>
His Gly Tyr Trp Gln Lys Trp Asp Leu Leu Thr Lys Gln Trp Val Ser 1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15
Ser Glu
<210>
        181
<211>
        18
<212>
<213>
        Artificial Sequence
<220>
<223> Preferred TALL-1 modulating domain
<400>
       181
His Gln Gly Gln Cys Gly Trp Asp Leu Leu Thr Arg Ile Tyr Leu Pro 1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15
Cys His
<210>
        182
        18
<212>
<213>
        PRT
       Artificial Sequence
<220>
<223>
        Preferred TALL-1 modulating domain
<400>
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Leu His Lys Ala Cys Lys Trp Asp Leu Leu Thr Lys Gln Cys Trp Pro $1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15$

Met Gln

```
<210>
<211>
        183
        18
<212>
<213>
        Artificial Sequence
<220>
<223>
        Preferred TALL-1 modulating domain
<400>
       183
Gly Pro Pro Gly Ser Val Trp Asp Leu Leu Thr Lys Ile Trp Ile Gln 10 15
Thr Gly
<210>
        184
<211>
<212>
        18
        Artificial Sequence
<213>
<220>
<223>
        Preferred TALL-1 modulating domain
<400>
Ile Thr Gln Asp Trp Arg Phe Asp Thr Leu Thr Arg Leu Trp Leu Pro 1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15
Leu Arg
<210>
        185
<211>
        18
<212>
        PRT
        Artificial Sequence
<213>
<220>
<223>
        Preferred TALL-1 modulating domain
<400>
Gln Gly Gly Phe Ala Ala Trp Asp Val Leu Thr Lys Met Trp Ile Thr 1 10 15
Val Pro
<210>
        186
<211>
        18
        PRT
<212>
<213>
        Artificial Sequence
<220>
       Preferred TALL-1 modulating domain
                                             Page 83
```

<400> 186 Gly His Gly Thr Pro Trp Trp Asp Ala Leu Thr Arg Ile Trp Ile Leu
1 10 15

Gly Val

<210> <211> <212> 187 18 PRT

Artificial Sequence <213>

<220>

<223> Preferred TALL-1 modulating domain

<400>

Val Trp Pro Trp Gln Lys Trp Asp Leu Leu Thr Lys Gln Phe Val Phe 1 5 10 15

Gln Asp

<210> 188 <211> 19

<212> PRT Artificial Sequence

<220>

<223> Preferred TALL-1 modulating domain

<400> 188

Trp Gln Gln Trp Ser Trp Lys Trp Asp Leu Leu Thr Arg Gln Tyr Ile $1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15$

Ser Ser Ser

189 <210>

<211> 882 <212> DNA

<213> Artificial Sequence

<220>

<223> TALL-1 inhibitory peptibody 12-3 tandem dimer

<220>

<221> CDS

<222> (1)..(879)

<400> 189

atg ctt cca ggc tgc aag tgg gat ctt ctt att aag caa tgg gta tgc Met Leu Pro Gly Cys Lys Trp Asp Leu Leu Ile Lys Gln Trp Val Cys

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1	Corr	ected sequ	ence listing-)	final.txt 15
gat cca ctt gga Asp Pro Leu Gly 20	tcc ggt tct Ser Gly Ser	gct act go Ala Thr Gl 25	ot ggt tcc ggc ly Gly Ser Gly	tcc acc gca 96 Ser Thr Ala 30
agc tct ggt tca Ser Ser Gly Ser 35	ggc agt gcg Gly Ser Ala	act cat at Thr His Me 40	g ctg ccg ggt et Leu Pro Gly 45	tgt aaa tgg 144 Cys Lys Trp
gac ctg ctg atc Asp Leu Leu Ile 50	aaa cag tgg Lys Gln Trp 55	gtt tgt ga Val Cys As	c ccg ctg ggt sp Pro Leu Gly 60	gga ggc ggt 192 Gly Gly Gly
ggg gtc gac aaa Gly Val Asp Lys 65	act cac aca Thr His Thr 70	tgt cca co Cys Pro Pr	t tgt cca gct o Cys Pro Ala 75	ccg gaa ctc 240 Pro Glu Leu 80
ctg ggg gga ccg Leu Gly Gly Pro	tca gtc ttc Ser Val Phe 85	ctc ttc cc Leu Phe Pr 90	o Pro Lys Pro	aag gac acc 288 Lys Asp Thr 95
ctc atg atc tcc Leu Met Ile Ser 100	cgg acc cct Arg Thr Pro	gag gtc ac Glu Val Th 105	a tgc gtg gtg or Cys Val Val	gtg gac gtg 336 Val Asp Val 110
agc cac gaa gac Ser His Glu Asp 115	cct gag gtc Pro Glu Val	aag ttc aa Lys Phe As 120	ic tgg tac gtg sn Trp Tyr Val 125	gac ggc gtg 384 Asp Gly Val
gag gtg cat aat Glu Val His Asn 130	gcc aag aca Ala Lys Thr 135	aag ccg cg Lys Pro Ar	gg gag gag cag g Glu Glu Gln 140	tac aac agc 432 Tyr Asn Ser
acg tac cgt gtg Thr Tyr Arg Val 145	gtc agc gtc Val Ser Val 150	ctc acc gt Leu Thr Va	cc ctg cac cag Il Leu His Gln 155	gac tgg ctg 480 Asp Trp Leu 160
aat ggc aag gag Asn Gly Lys Glu	tac aag tgc Tyr Lys Cys 165	aag gtc to Lys Val Se 17	er Asn Lys Ala	ctc cca gcc 528 Leu Pro Ala 175
ccc atc gag aaa Pro Ile Glu Lys 180	acc atc tcc Thr Ile Ser	aaa gcc aa Lys Ala Ly 185	a ggg cag ccc s Gly Gln Pro	cga gaa cca 576 Arg Glu Pro 190
cag gtg tac acc Gln Val Tyr Thr 195	ctg ccc cca Leu Pro Pro	tcc cgg ga Ser Arg As 200	it gag ctg acc p Glu Leu Thr 205	aag aac cag 624 Lys Asn Gln
gtc agc ctg acc Val Ser Leu Thr 210	tgc ctg gtc Cys Leu Val 215	aaa ggc tt Lys Gly Ph	c tat ccc agc le Tyr Pro Ser 220	gac atc gcc 672 Asp Ile Ala
gtg gag tgg gag Val Glu Trp Glu 225	agc aat ggg Ser Asn Gly 230	cag ccg ga Gln Pro Gl	ng aac aac tac u Asn Asn Tyr 235	aag acc acg 720 Lys Thr Thr 240
cct ccc gtg ctg Pro Pro Val Leu	gac tcc gac Asp Ser Asp 245	ggc tcc tt Gly Ser Ph 25	e Phe Leu Tyr	agc aag ctc 768 Ser Lys Leu 255
acc gtg gac aag	agc agg tgg	cag cag gg	g aac gtc ttc Page 85	tca tgc tcc 816

```
Corrected sequence listing- final.txt
Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser
260 265 270
gtg atg cat gag gct ctg cac aac cac tac acg cag aag agc ctc tcc
Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser
275 280 285
                                                                                           864
ctg tct ccg ggt aaa taa
Leu Ser Pro Gly Lys
290
                                                                                           882
         190
<210>
<211>
         23
<212>
         PRT
         Artificial Sequence
<220>
<223>
         Preferred linker sequence
<400>
         190
Gly Ser Gly Ser Ala Thr Gly Gly Ser Gly Ser Thr Ala Ser Ser Gly
1 10 15
Ser Gly Ser Ala Thr Gly Met 20
<210>
         191
<211>
         23
<212>
         PRT
<213>
        Artificial Sequence
<220>
<223>
         Preferred linker sequence
<400>
Gly Ser Gly Ser Ala Thr Gly Gly Ser Gly Ser Thr Ala Ser Ser Gly 1 	 5 	 10
Ser Gly Ser Ala Thr Gly Ser 20
<210>
<211>
<212>
         192
         46
         PRT
<213>
         Artificial Sequence
<220>
         Preferred linker sequence
<223>
<400>
        192
Gly Ser Gly Ser Ala Thr Gly Gly Ser Gly Ser Thr Ala Ser Ser Gly 10 15
Ser Gly Ser Ala Thr His Met Gly Ser Gly Ser Ala Thr Gly Gly Ser
Page 86
```

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Corrected sequence listing- final.txt 25
              20
Gly Ser Thr Ala Ser Ser Gly Ser Gly Ser Ala Thr His Met
<210>
       193
<211>
<212>
       23
       PRT
       Artificial Sequence
<213>
<220>
<223>
       Preferred linker sequence
<220>
<221>
<222>
        MISC_FEATURE
        (22)..(22)
<223>
        Basic or hydrophobic residue
<220>
<221>
<222>
       MISC_FEATURE (23)..(23)
<223>
       Hydrophobic residue
<400>
Gly Ser Gly Ser Ala Thr Gly Gly Ser Gly Ser Thr Ala Ser Ser Gly 10 15
Ser Gly Ser Ala Thr Xaa Xaa 20
<210>
       194
       46
<211>
<212>
       PRT
<213>
       Artificial Sequence
<220>
<223>
       Preferred linker sequence
<220>
<221>
<222>
       MISC_FEATURE
        (22)..(22)
<223>
        Basic or hydrophobic residue
<220>
<221>
       MISC_FEATURE
       (23)..(23)
Hydrophobic residue
<222>
<223>
<220>
<221>
       MISC_FEATURE
<222>
        (45)..(45)
<223>
       Basic or hydrophobic residue
<220>
<221>
<222>
       MISC_FEATURE
       (46)..(46)
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Corrected sequence listing- final.txt
<223>
        Hydrophobic residue
<400>
Gly Ser Gly Ser Ala Thr Gly Gly Ser Gly Ser Thr Ala Ser Ser Gly 1 5 15
Ser Gly Ser Ala Thr Xaa Xaa Gly Ser Gly Ser Ala Thr Gly Gly Ser
20 25 30
Gly Ser Thr Ala Ser Ser Gly Ser Gly Ser Ala Thr Xaa Xaa 35 40
        195
<211>
<212>
        38
        PRT
<213>
        Homo sapiens
<400>
Met Arg Arg Gly Pro Arg Ser Leu Arg Gly Arg Asp Ala Pro Val Pro 1 5 10 15
Thr Pro Cys Val Pro Thr Glu Cys Tyr Asp Leu Leu Val Arg Lys Cys 20 25 30
Val Asp Cys Arg Leu Leu
35
<210>
<211>
        196
        41
<212>
        PRT
<213>
        Homo sapiens
<400>
Thr Ile Cys Asn His Gln Ser Gln Arg Thr Cys Ala Ala Phe Cys Arg 1 	 5 	 10 	 15
Ser Leu Ser Cys Arg Lys Glu Gln Gly Lys Phe Tyr Asp His Leu Leu 20 25 30
Arg Asp Cys Ile Ser Cys Ala Ser Ile
35 40
<210>
<211>
<212>
        197
42
        PRT
       Homo sapiens
<400>
      197
Phe Val Ser Pro Ser Gln Glu Ile Arg Gly Arg Phe Arg Arg Met Leu 1 \hspace{1.5cm} 15
                                              Page 88
```

Gln Met Ala Gly Gln Cys Ser Gln Asn Glu Tyr Phe Asp Ser Leu Leu $20 \hspace{1cm} 25 \hspace{1cm} 30$

His Ala Cys Ile Pro Cys Gln Leu Arg Cys 35