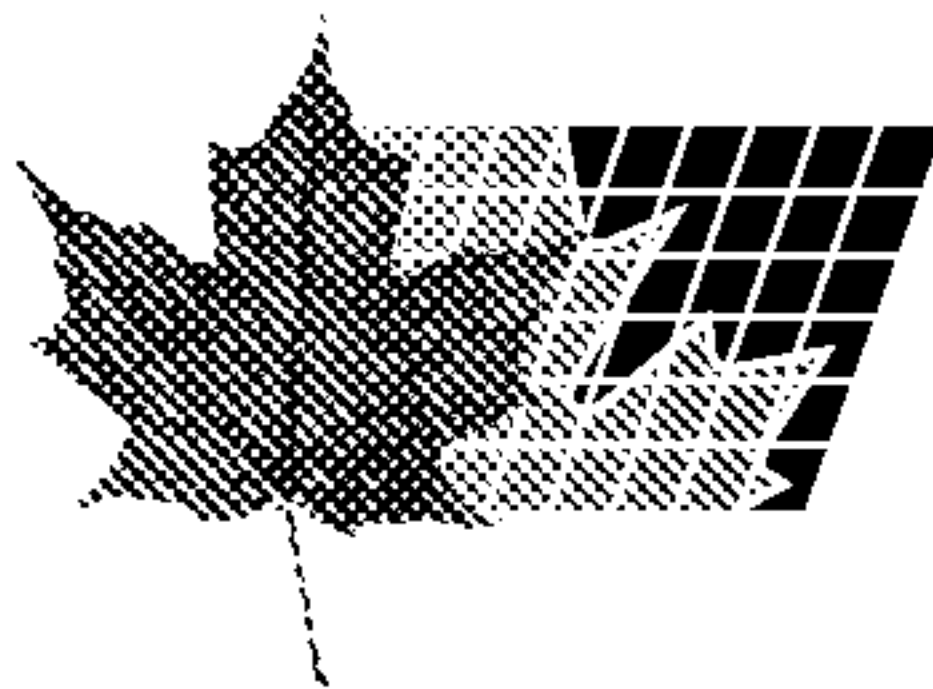




(86) 1990/09/04

(87) 1991/03/21

(45) 2001/05/15

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(51) Int.Cl.⁵ C12N 15/15, G01N 33/574, A61K 39/395, C12N 15/11,
C12N 5/10, C12P 21/02, C07K 16/38, A61K 38/57, C07K 14/81

(30) 1989/09/05 (PJ6179) AU

(54) **PRODUITS DE POLYNUCLEOTIDE RECOMBINANTS
SERVANT A FOURNIR DES SECRETIONS DE PAI-2**

(54) **RECOMBINANT POLYNUCLEOTIDE CONSTRUCTS
SUITABLE FOR PROVIDING SECRETION OF PAI-2**

(57) This invention relates to PAI-2 and its expression as a recombinant molecule in eukaryotic cell lines as a glycosylated secreted molecule, to the constructs expressing it, to host cells expressing it, to compositions comprising it, to methods of treatment, prophylaxis and diagnosis using it and to antibodies raised against it. The invention also provides a 414 amino acid form of PAI-2 wherein the N-terminal methionine residue is deleted, a 60 kD glycosylated secreted recombinant form of PAI-2 and compositions and methods using these molecules. The invention further relates to a novel synthetic signal peptide.

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ABSTRACT

This invention relates to PAI-2 and its expression as a recombinant molecule in eukaryotic cell lines as a glycosylated secreted molecule, to the constructs expressing it, to host cells expressing it, to compositions
5 comprising it, to methods of treatment, prophylaxis and diagnosis using it and to antibodies raised against it. The invention also provides a 414 amino acid form of PAI-2 wherein the N-terminal methionine residue is deleted, a 60 kD glycosylated secreted recombinant form of PAI-2 and compositions and methods using these molecules. The invention further relates to a novel
10 synthetic signal peptide.

Recombinant Product

Technical Field

5 The invention relates to enhanced secretion of PAI-2,
to recombinant polynucleotide constructs suitable for
providing enhanced secretion of PAI-2, to the expression
products of those constructs and to their uses, as well as a
414 amino acid form of PAI-2, a synthetic signal peptide
used in the preparation of glycosylated, secreted PAI-2 and
10 60 kD glycosylated, secreted, recombinant PAI-2.

Deposition of microorganisms

BTA 1445, an E. coli strain harbouring a PAI-2 encoding
construct was deposited with the American Type Culture
15 Collection of 12301 Parklawn Drive, Rockville, MD 20852
U.S.A. under the provisions of the Budapest Treaty on 11
February 1987 under accession number ATCC 53585.

Background Art

20 Plasminogen activator (PA) inhibitor Type 2 (PAI-2)
is one of four immunologically distinct groups of PA
inhibitors. PA inhibitors are members of the serpin gene
family.

PAI-2, also termed minactivin, has been purified from
25 the human monocytic cell line U937 (International Patent
Application PCT/AU85/00191 (filed August 13, 1985 in the name of
Biotechnology Australia Pty. Ltd. and the Australian National
University), published as WO86/01212 and PCT/AU87/00068 (filed
March 13, 1987 in the name of Biotechnology Australia Pty. Ltd.
30 and the Australian National University), published as
WO87/05628). It has also been detected in pregnancy plasma and
in the conditioned medium of several human cells including
peripheral blood monocytes, HT1080 fibrosarcoma cells and HEp3
laryngeal carcinoma cells.

35 Distinct molecular weight species of PAI-2 have been
identified:

A 46kD form is non-glycosylated and primarily cell
associated. This form has been found in lysates of U937
cells and purified from the conditioned medium of phorbol
ester (PMA) stimulated U937 cells (International Patent
Application No. PCT/AU87/00068 (filed March 13, 1987 in the name
of Biotechnology Australia Pty. Ltd. and the Australian National
University).

A 60kD glycosylated form has been found to be secreted by U937 cells (Genton et al., 1987) as well as being present in the maternal plasma of pregnant women (Lecander and Astedt, 1986).

5 The non-glycosylated 46kD intracellular form accounts for 80-90% of PAI-2 synthesized in U937 (Genton et al., 1987) and HT1080 cells (Medcalf et al., 1988)

 Examination of the primary amino acid sequence of PAI-2 reveals that the molecule lacks the characteristic
10 transient signal peptide usually found at the amino (NH₂)-terminus of secreted proteins. Although the NH₂-terminus of the purified 46kD form of PAI-2 from U937 cells has not been determined, apparently because it is blocked (Kruithof et al., 1986), the NH₂-terminus of the
15 60kD glycosylated version has been reported to be the initiator methionine (residue 1, Ye et al., 1988). This implies that secretion occurs without cleavage of a signal peptide from PAI-2. The mode of translocation of PAI-2 through the cell is not understood, however an internal
20 signal has been proposed (Ye et al., 1988).

 As is the case with most potent biologically active proteins, PAI-2 is produced in very small amounts in vivo and as such is difficult to purify and characterise by conventional biochemical approaches. The recent cloning of
25 the gene for PAI-2 and its expression in bacterial cells (International Application No. PCT/AU87/00068 (filed March 13, 1987 in the name of Biotechnology Australia Pty. Ltd. and the Australian National University) now allows the production of significant quantities of purified 46kD PAI-2 which is needed to
30 evaluate its biological efficacy in clinical applications. However this bacterial material is not glycosylated, nor modified post-translationally in a manner analogous to that secreted by human cells. Therefore it is desirable to produce glycosylated forms of PAI-2 using transfected mammalian
35 cells, since the two forms of PAI-2 may differ in their biological activities e.g. binding affinity for urokinase, PA specificity, immunogenicity, in vivo half-life etc.

The native PAI-2 gene has previously been expressed in a number of heterologous mammalian expression systems (International Application No. PCT/AU87/00068 (filed March 13, 1987 in the name of Biotechnology Australia Pty. Ltd. and the Australian National University)). Although PAI-2 is synthesized in these systems, expression levels are low, and the majority of the product (approx. 90%) is non-glycosylated and intracellular. PAI-2 produced in this form is a suitable molecule for prophylactic, therapeutic and/or diagnostic uses but its use is limited by the quantities obtainable and limited glycosylation. It is still desirable to be able to attain efficient secretion, of the 60 kD glycosylated molecule, in order to be able to use the molecule for prophylactic, therapeutic and/or diagnostic purposes on a clinical scale.

In addition to expression of PAI-2 in mammalian cells (International Application No. PCT/AU87/00068 (filed March 13, 1987 in the name of Biotechnology Australia Pty. Ltd. and the Australian National University)), workers have attempted to secrete the 60 kD form of PAI-2. A Swedish group has attempted to secrete PAI-2 from transfected CHO cells but could not detect any secretion of 60 kD PAI-2 (Ny et al, 1988, Ny et al, 1989). A Swiss group attempted the secretion of PAI-2 using transfected WISH cells but likewise found that most of the PAI-2 material was of the intracellular form. Although some high molecular weight material was found in the culture medium, it was not demonstrated that the material was glycosylated (Belin, D., Wohlwend, A., and Vassalli, J.D. (1989); Belin, D., Wohlwend, A., Schleuning, W-D, Kruithof, E.K.O. and Vassalli, J.D. (1989)). The material may have been partially glycosylated because the mobility was different from U937 high molecular weight PAI-2.

Previous studies have shown that the addition of a heterologous hydrophobic signal domain to the NH₂-terminus of a cytoplasmic protein may result in certain circumstances in the translocation of the cytoplasmic protein across the endoplasmic reticulum (Hiebert and Lamb, 1988). Translocation across the ER is the first step along the secretory pathway. The nature of the heterologous signal peptide (e.g. whether or not it is cleaved from the protein) and the inherent properties of the cytoplasmic protein (e.g. the presence of other transport or membrane retention

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signals) then determine whether the chimeric protein is secreted, anchored to a cellular membrane or degraded.

Because of the complexity of the secretion mechanisms it is not possible to convert in a predictable manner a cytoplasmic non-glycosylated molecule into a secreted, glycosylated molecule. Even if by the addition of a signal sequence a cytoplasmic molecule is secreted, the processing (i.e. cleavage) of the added signal sequence may occur at different unpredictable locations.

Transient NH_2 -terminal signal sequences found on most secreted proteins are identified by three distinct regions - a basic N-terminal region which may contain charged residues, a central hydrophobic core and a C-terminal region containing the signal peptidase recognition site.

There is no consensus sequence for signal peptidase recognition although von Heijne (1986) has developed rules that take account of many of the known cleavage sites. The application of these rules, however, does not necessarily allow the prediction of cleavage sites when they are applied to cytoplasmic molecules bearing heterologous signal sequences.

Disclosure of the Invention

In accordance with the present invention, the addition of selected or designed signal sequences to PAI-2 not only facilitates secretion of the glycosylated 60 kD form of PAI-2 but also directs correct processing of the signal. In addition, by utilising these signal sequences, PAI-2 has been secreted either with or without its N-terminal methionine residue.

Definitions

As used throughout the specification the expressions "PAI-2 variant" and "variant of PAI-2" refer to a molecule derived from PAI-2 by alteration to the NH_2 -terminus thereof to provide an efficient signal sequence for secretion of the glycosylated molecule from a host cell.

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The term "parenteral" as used herein includes subcutaneous injections, intravenous, or intramuscular injection, or infusion techniques.

5 The term "synthetic signal peptide" as used throughout the specification refers to a peptide which is made in vitro or in vivo by translating a synthesized oligo-deoxyribonucleotide sequence.

10 According to a first embodiment of this invention there is provided a process for the preparation of recombinant, glycosylated, secreted PAI-2 or recombinant glycosylated secreted PAI-2 without its N-terminal methionine, which process comprises:

15 providing a construct comprising a first polynucleotide molecule encoding PAI-2 or PAI-2 without its N-terminal methionine; and

20 attaching a polynucleotide molecule encoding a transient signal sequence to the 5' end of the first polynucleotide molecule, such that the resulting hybrid protein expressed from the construct will consist of a transient signal sequence attached to the NH₂-terminal of PAI-2 or of PAI-2 without its N-terminal methionine; and expressing the construct in a eukaryotic host cell.

25 The signal peptide may be attached to the N-terminal methionine of PAI-2 or alternatively the N-terminal methionine may be deleted, and the signal peptide attached to the next residue, viz glutamic acid.

30 The signal peptide is designed to be cleaved during translocation of the PAI-2 molecule across the endoplasmic reticulum of the host cell.

The signal peptide may be a synthetic signal peptide.

35 Generally, the synthetic signal sequence is designed so that it has three distinct regions - a basic N-terminal region which may contain charged residues, a central hydrophobic core and a C-terminal region containing the signal peptidase recognition site. Attempts at designing signal peptidase cleavage sites can be assisted using the method of von Heijne (1986).

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A preferred synthetic peptide is:

M K C L L L A L G L L A F V P L V R A

The signal peptide may be naturally occurring.

5 A preferred natural signal peptide is the signal peptide from the protein human α -1-antitrypsin:

M P S S V S W G I L L L A G L C C L V P V S L A.

Other natural signal peptides could function in an analagous manner to the α -1-antitrypsin signal when fused to the NH_2 -terminus of PAI-2.

10 According to a second embodiment of the invention there is provided a recombinant glycosylated PAI-2 encoding construct comprising a polynucleotide molecule encoding a transient signal peptide attached to the NH_2 -terminal methionine or the NH_2 -terminal glutamic acid of PAI-2.

15 Preferably, the construct is a DNA molecule.

According to a third embodiment of the invention there is provided a variant of PAI-2 comprising an altered NH_2 -terminus wherein the NH_2 -terminus of PAI-2 is altered to provide an efficient signal peptide.

20 Typically, the alteration is in the first 22 amino acids of the amino acid sequence of PAI-2.

Preferably, all or some of the asparagine residues at positions 8 and 14, lysine at position 17, and the histidine at position 18 are replaced. Preferably replacement is by
25 in vitro mutagenesis. Preferably the amino acids are replaced by a more hydrophobic amino acid selected from Leu, Phe, Ala, Met, Thr, Ser, Ile and Val.

According to a fourth embodiment of this invention there is provided a polynucleotide molecule encoding a PAI-2
30 variant of the third embodiment.

Preferably, the polynucleotide molecule is a DNA molecule.

According to a fifth embodiment of this invention there is provided the synthetic peptide:

M K C L L L A L G L L A F V P L V R A.

35 Preferably, the signal encoding sequences are prepared by DNA synthesis regardless of whether they are synthetic or naturally occurring signal sequences.

According to a sixth embodiment of the invention there is provided a recombinant DNA molecule comprising a

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polynucleotide molecule of the second embodiment or of the fourth embodiment which polynucleotide molecule is a DNA molecule, and vector DNA.

Typically, the vector DNA is plasmid DNA.

5 Preferred plasmid vectors of the invention include pGEM4Z, pSVL, pBTA613, pBTA830 and baculovirus transfer vectors such as pAC373.

Preferred recombinant DNA molecules of the invention include pBTA822, pBTA823, pBTA825, pBTA826, pBTA827, pBTA828
10 and pBTA839.

According to a seventh embodiment of the invention there is provided a transformed host cell transformed by a recombinant DNA molecule of the sixth embodiment.

Typical host cell lines are derived from eukaryotic
15 organisms and include monkey kidney COS cells, the monkey kidney cell line Vero, Chinese Hamster Ovary (CHO) cells, the human histiocytic lymphoma U937 cell line, derivatives of CHO-K1 cells, for example DG44, the hamster kidney cell line BHK-21, the human kidney cell line 293, the human
20 epithelial cell derived line HeLa S3 and the human monocyte cell line HL60 and cell lines derived from the insects Spodoptera frugiperda and Bombyx mori.

Preferably, the transformation is carried out by the calcium phosphate method or by electroporation.

25 According to an eighth embodiment of this invention there is provided an expression product of a transformed host cell of the seventh embodiment. The expression product is glycosylated PAI-2 or a variant of PAI-2.

The invention also provides the 414 amino acid form
30 of PAI-2 which lacks the N-terminal methionine of native PAI-2 in glycosylated and unglycosylated form. In addition the invention provides for the first time recombinant, glycosylated, secreted 60 kD PAI-2.

According to a ninth embodiment of this invention
35 there is provided a composition comprising an effective amount of an expression product of the eighth embodiment, or of the 60 kD glycosylated, secreted recombinant PAI-2, or of the 414 amino acid form of PAI-2 together with a pharmaceutically acceptable carrier, diluent, excipient,

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and/or adjuvant.

According to a tenth embodiment of this invention there is provided a process for preparing a recombinant DNA molecule of the sixth embodiment which process comprises
5 inserting a DNA molecule of the second embodiment or of the fourth embodiment into vector DNA.

According to an eleventh embodiment of this invention there is provided a process for preparing a transformed host cell of the seventh embodiment which process comprises
10 making a host cell competent for transformation and transforming said competent host cell with a recombinant DNA molecule of the sixth embodiment.

According to a twelfth embodiment of this invention there is provided a process for preparing an expression
15 product of the eighth embodiment which process comprises culturing a transformed host cell of the seventh embodiment and separating the expression product from the culture.

According to a thirteenth embodiment of this invention there is provided a process for preparing a
20 composition of the ninth embodiment which process comprises admixing an expression product of the eighth embodiment or 60 kD recombinant glycosylated secreted PAI-2, or the 414 amino acid form of PAI-2 with a pharmaceutically acceptable carrier, diluent, excipient, and/or adjuvant.

According to a fourteenth embodiment of this invention there is provided an antibody raised against an expression product of the eighth embodiment or against recombinant secreted glycosylated 60 kD PAI-2, or against the 414 amino acid form of PAI-2, or against a composition
25 of the ninth embodiment.
30

According to a fifteenth embodiment of this invention there is provided an antibody composition comprising an effective amount of an antibody of the fourteenth embodiment together with a pharmaceutically acceptable carrier,
35 diluent, excipient, and/or adjuvant.

According to a sixteenth embodiment of this invention there is provided a method of passively vaccinating a host in need of such treatment which method comprises administering an effective amount of an antibody of the

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fourteenth embodiment or an antibody composition of the fifteenth embodiment, to the host.

According to a seventeenth embodiment of this invention there is provided a process for producing an antibody of the fourteenth embodiment which process
5 comprises immunising an immunoresponsive host with an expression product of the eighth embodiment, or recombinant glycosylated secreted 60 kD PAI-2, or the 414 amino acid form of PAI-2, or a composition of the ninth embodiment.

10 According to a eighteenth embodiment of the invention there is provided an expression product of the eighth embodiment produced by the process of the twelfth embodiment.

According to a nineteenth embodiment there is provided an antibody of the fourteenth embodiment produced
15 by the process of the seventeenth embodiment.

The antibodies produced may be monoclonal or polyclonal antibodies. For the production of monoclonal antibodies the PAI-2 molecules of the invention are used as immunogens according to standard techniques for the
20 preparation of monoclonal antibodies.

According to a twentieth embodiment of the invention there is provided a diagnostic kit containing an expression product, and/or composition, and/or antibody and/or antibody composition, respectively of the eighth, ninth, fourteenth
25 and fifteenth embodiments, and/or the 414 amino acid form of PAI-2 and/or 60 kD glycosylated secreted recombinant PAI-2, together with positive and negative standards as controls.

According to a twenty-first embodiment of this invention there is provided a reagent for locating and
30 defining the boundaries of tumours in histological specimens or in vivo which reagent comprises a suitably labelled expression product of the eighth embodiment, or suitably labelled 60 kD recombinant secreted glycosylated PAI-2, or suitably labelled 414 amino acid form of PAI-2.

35 According to a twenty-second embodiment of this invention there is provided a method of locating and defining the boundaries of tumours in histological specimens or in vivo which method comprises applying or administering

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a reagent according to the twenty-first embodiment to the specimen or patient respectively and subsequently imaging to determine the site of the concentration of the label.

According to a twenty-third embodiment of this invention there is provided a method of inhibiting tumour invasion or treating tumours comprising administering to a patient requiring such treatment a therapeutically effective amount of an expression product of the eighth embodiment or a composition of the ninth embodiment or 60 kD glycosylated secreted recombinant PAI-2 or the 414 amino acid form of PAI-2 either labelled or unlabelled.

According to a twenty-fourth embodiment of this invention there is provided a method of treatment of chronic inflammation such as rheumatoid arthritis comprising administering to a patient requiring such treatment a therapeutically effective amount of an expression product of the eighth embodiment or a composition of the ninth embodiment or recombinant glycosylated secreted 60 kD PAI-2 or the 414 amino acid form of PAI-2 either labelled or unlabelled.

Labels to be used in conjunction with 60 kD PAI-2, expression products and 414 amino acid forms of PAI-2 in accordance with this invention are those standardly used in the art for labelling for the purposes of diagnosis, in vitro detection, imaging, or therapy.

According to a twenty-fifth embodiment of this invention there is provided a method of monitoring chronic inflammation comprising the detection of PAI-2 in samples of body fluids and tissues using antibodies of the fourteenth embodiment or an antibody composition of the fifteenth embodiment.

According to a twenty-sixth embodiment of the invention there is provided a method of monitoring chronic inflammation comprising using an expression product of the eighth embodiment, recombinant glycosylated secreted 60 kD PAI-2 or the 414 amino acid form of PAI-2.

The PAI-2 variants expression products, 414 amino acid form of PAI-2 and recombinant glycosylated secreted 60 kD PAI-2 produced in accordance with this invention are of

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use in the following areas:

therapy, prophylaxis and diagnosis of inflammatory disease;

therapy, prophylaxis and diagnosis of cancer metastasis and proliferation;

therapy of transplant and placenta rejection crises and graft-versus-host reactions;

therapy and diagnosis of autoimmune diseases and diseases associated with excessive oxygen radical production;

therapy and prophylaxis of wound and bone healing disorders, tissue damage during or after acute reperfusion, and subarachnoidal bleeding disorders;

as topical medicaments for promoting fibrin adhesion, promoting healing of wounds and burns;

treating asthma and treating or preventing diseases associated with high leukocyte activity; and

suppression of the monocyte-macrophage system.

Specific examples from the above classes of disease include:

rheumatoid arthritis

osteoarthritis

colitis ulcerosa

systemic lupus erythematosus

multiple sclerosis

psoriasis

pemphigus

corneal and duodenal ulceration

purpura

periodontitis

muscular dystrophy

The amount of expression product, 60 kD glycosylated secreted recombinant PAI-2 or 414 amino acid form of PAI-2 that may be combined with carrier to produce a single dosage form will vary depending upon the condition being treated, the host to be treated and the particular mode of administration.

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It will be understood, also, that the specific dose level for any particular patient will depend upon a variety of factors including the activity of the specific protein product or antibody employed, the age, body weight, general health, sex, diet, time of administration, route of administration, rate of excretion, drug combination, the particular state being treated and the severity of the particular condition undergoing treatment.

The compositions of the present invention may be administered parenterally or topically or potentially via mucosal routes in dosage unit formulations containing conventional, non-toxic, pharmaceutically acceptable carriers, diluents, adjuvants and/or excipients as desired.

Injectable preparations, for example, sterile injectable aqueous or oleagenous suspensions may be formulated according to the known art using suitable dispersing or wetting agents and suspending agents. The sterile injectable preparation may also be a sterile injectable solution or suspension in a non-toxic parenterally acceptable diluent or solvent, for example, a solution in 1,3-butanediol. Among the acceptable vehicles and solvents that may be employed are water, Ringer's solution, and isotonic sodium chloride solution. In addition, sterile, fixed oils are conventionally employed as a solvent or suspending medium. For this purpose any bland fixed oil may be employed including sythetic mono- or diglycerides, In addition, fatty acids such as oleic acid find use in the preparation of injectables.

The term "pharmaceutically acceptable adjuvant" can mean either the standard compositions which are suitable for human administration or the typical adjuvants employed in animal vaccinations.

At present alum is the only registered adjuvant for human use however, experimental work is being conducted on other adjuvants for human use and it is anticipated that these other adjuvants would be suitable for use in preparing compositions for human vaccination in accordance with this invention.

Suitable adjuvants for the vaccination of animals include but are not limited to oil emulsions such as Freund's complete or incomplete adjuvant (not suitable for livestock use), Marcol 52: Montanide 888 (Marcol is a Trademark of Esso. Montanide is a Trademark of SEPPIC, Paris), squalane or squalene, Adjuvant 65 (containing peanut oil, mannide monooleate and aluminium monostearate), mineral gels such as aluminium hydroxide, aluminium phosphate, calcium phosphate and alum, surfactants such as hexadecylamine, octadecylamine, lysolecithin, dimethyldioctadecylammonium bromide, N,N-dioctadecyl-N',N'-bis(2-hydroxyethyl) propanediamine, methoxyhexadecylglycerol and pluronic polyols, polyanions such as pyran, dextran sulfate, polyacrylic acid and carbopol, peptides and amino acids such as muramyl dipeptide, dimethylglycine, tuftsin and trehalose dimycolate. The expression products of the present invention can also be administered following incorporation into liposomes or other micro-carriers, or after conjugation to polysaccharides, proteins or polymers or in combination with Quil-A to form "Iscoms" (Immunostimulating complexes) (Morein et al., Nature 308, 457-460 [1984]). Other adjuvants suitable for use in the present invention include conjugates comprising the expression product together with an integral membrane protein of prokaryotic origin, such as Trat. (See PCT/AU87/00107, filed April 16, 1987 in the name of Bioenterprises Pty. Ltd.).

Routes of administration, dosages to be administered as well as frequency of injections are all factors which can be optimized using ordinary skill in the art. Typically, the initial vaccination is followed some weeks later by one or more "booster" vaccinations, the net effect of which is the production of high titres of antibodies against the immunogen.

Compositions for topical administration include creams, ointments and pastes. The ingredients that constitute the base of ointments (e.g. petrolatum, waxes) are melted together, powdered drug components are added and the mass stirred with cooling. Generally, the product is then passed through a roller mill to achieve the

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particle-size range desired for the dispersed solid. Pastes are ointments with relatively large, dispersed solid content, and are prepared similarly.

5 Creams are semisolid emulsions, either water-in-oil or oil-in-water. A solid ingredient can be added to the appropriate phase before emulsification or may be dispersed at some point after the emulsification step. Topical dosage forms include disc dosage form systems that have been used for transdermal delivery of therapeutic agents. They
10 provide uniform and prolonged drug release.

It would be desirable in patients suffering from particular disease states to be able to administer the compositions of the invention either orally, rectally or possibly even vaginally. Compositions which could be used
15 in these ways could be prepared as follows:

Suppositories for rectal or vaginal administration of the compositions of the invention can be prepared by mixing the composition with a suitable nonirritating excipient such as cocoa butter, theobroma oil, glycerinated gelatin or
20 polyethylene glycols which are solid at ordinary temperatures but liquid at rectal or vaginal temperature or by contact with fluids present in the appropriate cavity and will therefore melt in the rectum or vagina and release the drug.

25 Solid dosage forms for oral administration may include capsules, tablets, pills, powders, and granules. In such solid dosage forms, expression products, recombinant glycosylated secreted 60 kD PAI-2 or 414 amino acid form of PAI-2 may be admixed with at least one inert diluent such as
30 sucrose, lactose starch or hydrolysed gelatin. Such dosage forms may also comprise, as is normal practice, additional substances other than inert diluents, e.g., lubricating agents such as magnesium stearate. In the case of capsules, tablets, and pills, the dosage forms may also comprise
35 buffering agents. Tablets and pills can additionally be prepared with enteric coatings.

Liquid dosage forms for oral administration may include nanoparticles, microcapsules, LTB conjugates,

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cholera or its B subunit as a conjugate, or vitamin B12 conjugates in pharmaceutically acceptable emulsions, syrups, solutions, suspensions, and elixirs containing inert diluents commonly used in the art, such as water. Such compositions may also comprise adjuvants, such as wetting agents, emulsifying and suspending agents or TraT as a conjugate, and sweetening, flavouring, and perfuming agents including sugars such as sucrose, sorbitol, fructose etc, glycols such as polyethylene glycol, propylene glycol etc, oils such as sesame oil, olive oil, soybean oil etc, antiseptics such as alkylparahydroxybenzoate etc, and flavours such as strawberry flavour, peppermint etc.

The recombinant constructs of the invention provide expression of secreted and glycosylated forms of PAI-2.

Brief Description of Drawings

Figure 1 shows the sequence of oligonucleotides and encoded signal peptides employed in construction of the in vitro and in vivo PAI-2 expression vectors.

Figure 2 shows construction of in vitro expression vectors:

- A: pBTA 821 containing PAI-2 gene without signal
- B: pBTA 822 containing PAI-2 gene with synthetic signal
- C: pBTA 823 containing PAI-2 gene with α -1-antitrypsin signal

Figure 3 shows construction of mammalian cell expression vectors:

- A: pBTA 824 containing PAI-2 gene without signal
- B: pBTA 825 and pBTA 827 containing PAI-2 gene with synthetic signal
- C: pBTA 826 and pBTA 828 containing PAI-2 gene with α -1-antitrypsin signal
- D: pBTA 839 containing PAI-2 gene with synthetic signal

Figure 4 shows the products of the PAI-2 gene and signal variants translated in a cell-free system and electrophoresed through an SDS-PAG.

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Figure 5 shows expression of PAI-2 gene and signal variants in transfected mammalian cells. Aliquots of the medium were immunoprecipitated with an anti-PAI-2 antibody and precipitates run on an SDS-PAG.

5 Figure 6 shows urokinase binding to PAI-2 secreted from transfected COS cells immunoprecipitated with anti-PAI-2 antibodies and electrophoresed through an SDS-PAG.

10 Figure 7 shows DNA and amino acid sequence of preA-PAI2 VIZ PAI-2 plus synthetic signal. Amino acids in capitals indicate the signal sequence.

Figure 8 shows DNA and amino acid sequence of preB-PAI-2 VIZ PAI-2 plus α -1-antitrypsin signal. Amino acids in capitals indicate the signal sequence.

15 Figure 9 shows the digestion of the 60,000 Mr PAI-2 secreted from transfected COS cells with various glycosidases. The products were separated by SDS-PAGE.

Figure 10 shows the peaks of radioactivity released during repeated cycles of Edman degradation on labelled 60,000 Mr PAI-2 secreted from COS cells transfected with:

20 A: pBTA 825 (contains PAI-2 gene with synthetic signal)

 B: pBTA 826 (contains PAI-2 gene with alpha-1 antitrypsin signal)

25 Figure 11 shows the secretion of 60,000 Mr PAI-2 into the medium from a CHO-K1 cell line stably transfected with pBTA 827. The PAI-2 was immunoprecipitated from the medium and then fractionated by SDS-PAGE.

Best Mode of Carrying Out the Invention

30 The recombinant DNA molecules and transformed host cells of the invention are prepared using standard techniques of molecular biology.

35 Expression products of the invention are obtained by culturing the transformed host cells of the invention under standard conditions as appropriate to the particular host cell and separating the expression product from the culture by standard techniques. The expression product may be used

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in impure form or may be purified by standard techniques as appropriate to the expression product being produced.

The compositions of the invention are prepared by mixing, preferably homogeneously mixing, expression product,
5 60 kD recombinant secreted glycosylated PAI-2 or the 414 amino acid form of PAI-2 with a pharmaceutically acceptable carrier, diluent, excipient and/or adjuvant using standard methods of pharmaceutical preparation.

The amount of expression product, recombinant
10 glycosylated secreted 60 kD PAI-2, or the 414 amino acid form of PAI-2 required to produce a single dosage form will vary depending upon the condition to be treated, patient to be treated and the particular mode of administration. The specific dose level for any particular patient will depend
15 upon a variety of factors including the activity of the expression product or PAI-2 molecule employed, the age, body weight, general health, sex, and diet of the patient, time of administration, route of administration, rate of excretion, drug combination and the severity of the
20 condition undergoing treatment.

The composition may be administered parenterally or topically, or possibly by inhalation spray, vaginally, orally or rectally in unit dosage formulations containing conventional, non-toxic, pharmaceutically acceptable
25 carriers, diluents, excipients and/or adjuvants as desired.

Antibodies are raised using standard vaccination regimes in appropriate hosts. The host is vaccinated with an expression product, recombinant glycosylated secreted 60 kD PAI-2 or the 414 amino acid form of PAI-2 or a
30 composition of the invention.

The antibody composition is prepared by mixing, preferably homogeneously mixing, antibody with a pharmaceutically acceptable carrier, diluent, excipient and/or adjuvant using standard methods of pharmaceutical
35 preparation.

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The amount of antibody required to produce a single dosage form will vary depending upon the condition to be treated, patient to be treated and the particular mode of administration. The specific dose level for any particular patient will depend upon a variety of factors including the activity of the antibody employed, the age, body weight, general health, sex, and diet of the patient, time of administration, route of administration, rate of excretion, drug combination, and severity of the condition undergoing treatment.

The antibody composition may be administered parenterally in unit dosage formulations containing conventional, non-toxic, pharmaceutically acceptable carriers, diluents, excipients and/or adjuvants as desired.

Diagnostic kits are prepared by formulating expression product and/or antibodies and/or recombinant glycosylated secreted 60 kD PAI-2 and/or the 414 amino acid form of PAI-2 at appropriate concentration to the substance(s) to be detected with a pharmaceutically acceptable carrier, diluent, excipient and/or adjuvant. A positive control standard of a known concentration of the substance to be detected is prepared similarly. The negative standard comprises carrier, diluent, excipient and/or adjuvant alone. Examples of diagnostic kits include a tumour diagnostic wherein the reagent comprises anti-expression product antibodies and the positive control comprises an expression product standard of known concentration.

We have constructed two variants of PAI-2 that are efficiently secreted from mammalian cells; both contain an NH₂-terminal signal designed to be cleaved during translocation across the endoplasmic reticulum (ER). One of these signals is an artificial signal that we designed according to the above description of signal peptides. It has the sequence:

M K C L L L A L G L L A F V P L V R A

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The second signal is the natural signal peptide from the protein human α -1-antitrypsin:

M P S S V S W G I L L L A G L C C L V P V S L A.

However it is possible that other natural signal peptides could function in an analogous manner to the α -1-antitrypsin signal when fused to the NH_2 -terminus of PAI-2. The examples of signal peptides described are by way of example only and are not intended to be limiting on the scope of the invention.

The first PAI-2 signal-containing variant was constructed such that the artificial signal was fused to the NH_2 -terminal methionine of PAI-2. Correct cellular processing of this signal peptide should result in a mature PAI-2 molecule of 415 amino acids with Met at its NH_2 -terminus. The second variant of PAI-2 was constructed by fusing the human α -1-antitrypsin signal peptide to PAI-2 without its NH_2 -terminal methionine. After correct cellular processing a mature PAI-2 molecule of 414 residues having a glutamic acid residue at its NH_2 -terminus should be generated. Details of the construction of the signal-PAI-2 molecules and their expression in both in vitro and in vivo systems are described below.

An alternative to adding heterologous signal peptides to PAI-2 in order to have PAI-2 secreted efficiently from mammalian cells could be to modify the native PAI-2 sequence. The first 22 amino acids of PAI-2 contain a stretch of hydrophobic residues characteristic of a signal peptide. However experiments below show that this region does not behave as a typical signal i.e. it does not promote translocation of PAI-2 across the ER. However, since some high molecular weight glycosylated PAI-2 has been found in culture media and plasma, residues 1-22 may represent an inefficient signal. The hydrophobic nature of the NH_2 -terminal 22 residues of PAI-2 is compromised by 2 asparagine residues (positions 8 and 14), a lysine at position 17, and a histidine at 18. In vitro mutagenesis of this region to replace one or more of these residues with a

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more hydrophobic amino acid (e.g. Leu, Phe, Ala, Met, Thr, Ser, Ile, Val) could improve the efficiency with which this region acts as a signal peptide. A signal peptidase recognition site is predicted after Ala₂₂. Thus these PAI-2 variants may be secreted without their first 22 residues. Although this region is conserved between members of the serpin gene family its presence may not be required for biological activity of PAI-2.

Construction of Vectors for Expression of Signal-PAI-2 variants

For details of recombinant DNA techniques used see Maniatis et al (1982). Oligonucleotides encoding the signal peptides (Figure 1) were synthesized on an Applied Biosystems DNA synthesizer (Model 380A), and purified through a polyacrylamide gel. Complementary oligonucleotides (A1 and A2 or B1 and B2 or C1 and C2) were mixed in a 1:1 molar ratio and phosphorylated with 5 units T4 polynucleotide kinase in 65mM Tris-Cl pH 7.5, 10mM MgCl₂, 5mM dithiothreitol, 1mM ATP. The mixture was heated to 100°C for 3 minutes and cooled slowly to room temperature to allow annealing to take place.

The annealed signal peptide oligonucleotides were ligated to the PAI-2 coding sequences and cloned into the vectors pGEM4Z (Promega), pSVL (Pharmacia) pBTA613 and pBTA830. The first vector, pGEM4Z, is useful for expressing gene products in a cell-free transcription/translation system; pSVL is useful for transient expression in monkey kidney COS cells; pBTA613 and pBTA830 are vectors that can be selected and stably integrated in mammalian cell lines such as Chinese Hamster Ovary (CHO) cells, Vero, BHK and U937. The signal-PAI-2 gene constructs could also be cloned into any other vector designed for expression in eukaryotic cells.

The construction of the recombinant plasmids that express the native PAI-2 gene and the signal PAI-2 variants in an in vitro system and in mammalian cells is illustrated in Figures 2 and 3. The various restriction fragments ("inserts" for cloning) were prepared as follows.

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Restriction enzyme digests of purified plasmid DNA were carried out in buffers recommended by the supplier. Required DNA fragments were separated from the plasmid by gel electrophoresis through 0.8-1.5% Sea-Plaque* agarose (FMC Corporation) in Tris-borate buffer (Maniatis et al, 1982). Fragments were visualized by staining with ethidium bromide and UV transillumination. The band of agarose containing the appropriate fragment was sliced out of the gel, melted at 65°C and the DNA was extracted by passing the diluted material through a NACS column (BRL) as recommended by the supplier. The DNA was then precipitated with ethanol in the presence of carrier tRNA (10 µg/ml).

Vectors were typically prepared as follows. Plasmid DNA purified by CsCl density gradient centrifugation was digested with the appropriate restriction enzymes, the digest was extracted with an equal volume of phenol/chloroform (1:1) and the DNA precipitated with 2.5 volumes of ethanol. The digested DNA was resuspended in 50mM Tris-Cl pH 9.0, 1mM MgCl₂, 0.1mM ZnCl₂, 1mM spermidine and incubated with 1-2 units calf intestinal alkaline phosphatase (Boehringer Mannheim) for 30-60 mins at 37°C. The enzyme was heat killed at 70°C for 15 mins then the DNA was extracted with phenol/chloroform and precipitated with ethanol.

Ligations were carried out as follows. Vector and insert DNAs were mixed at a molar ratio of between 1:1 and 1:5 (1:10 if the insert was smaller than 100bp) in 1mM ATP, 10mM MgCl₂, 5mM DTT, 65mM Tris-Cl pH7.5 in a volume of 10-20µl. Ligations were carried out at 16° overnight with 0.5-1 unit T4 DNA ligase (Boehringer Mannheim). Ligation mixes were diluted to 30µl with dH₂O and 10µl removed for transformation into a competent E. coli K12 host (Hanahan, 1985). Transformants were selected by plating onto tryptone-soya agar plates containing 100µg/ml ampicillin.

Plasmid DNA was extracted from individual colonies (Birnboim and Doly, 1979) and the correct recombinant

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plasmids identified by restriction analysis. The 5' ends of the PAI-2 gene in pBTA821, 822 and 823 were sequenced to confirm that the signal peptide and methionine start codons were in frame with the remainder of the PAI-2 sequence.

5 Sequencing was carried out on double-stranded plasmid DNA using the Sequenase* DNA Sequencing Kit (USB) as described in the instruction manual. The primer used was the T7 primer (Promega).

10 The plasmids and strains used in the work are summarized in Table 1.

TABLE 1: Plasmids and Bacterial Strains

BTA					
	<u>Strain#</u>	<u>Plasmid</u>	<u>Host</u>	<u>Description</u>	<u>Signal</u>
15	1702	pBTA613 ^A	HB101 ^B	eukaryotic expression vector	
	1737	pBTA839	"	1.3 kb Hind III-EcoRI from pBTA-822 in pBTA 613	synthetic
20	1740	pBTA830 ^A	"	eukaryotic expression vector	
	1741	pBTA821	"	1.3kb PAI-2 gene in pGEM4Z ^C	none
25	1742	pBTA822	"	1.3kb PAI-2 gene in pGEM4Z	synthetic
	1743	pBTA823	"	1.3kb PAI-2 gene in pGEM4Z	α -1-antitrypsin
	1744	pBTA824	"	1.3kb SalI-SacI from pBTA821 in pSVL ^D	none
30	1745	pBTA825	"	1.3kb SalI-SacI from pBTA822 in pSVL	synthetic
35	1746	pBTA826	"	1.3kb SalI-SacI from pBTA823 in pSVL	α -1-antitrypsin

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Table 1 (continued)

	1747	pBTA827	"	1.3kb HindIII-EcoRI from pBTA822 in pBTA830	synthetic
5	1748	pBTA828	"	1.3kb HindIII-EcoRI from pBTA823 in pBTA830	α -1- antitrypsin
	1613	pBTA446 ^A	C600 λ ^F	PAI-2 gene in pLK58 ^E	none
10	1650	pBTA641 ^A	N4830 ^G	PAI-2 gene in pLK58	none
	1835	pBTA679 ^A	HB101	PAI-2 gene minus 3' untranslated region derived from pBTA438 ^E	none
15	1445 ^H	pBTA438	JM109 ^I	1.6kb cDNA coding PAI-2 cloned pUC18	none

-
- 20 A constructed at Biotechnology Australia Pty Ltd (see text below)
- B See Maniatis et al 1982
- C Purchased from Promega
- D Purchased from Pharmacia LKB Biotechnology
- E Botterman and Zabeau (1985)
- 25 F E. coli K12 C600 λ - thiA1, thr1, leuB6, SupE(gluV)44, lacY1, fhuA21(λ)^T
- G Gottesman et al (1980)
- H Strain deposited with American Type Culture Collection, of 12301 Parklawn Drive, Rockville MD 20852, USA on 11 February 1987 under accession number ATCC 53585 and described in PCT/AU87/00068
- 30 I Yanisch-Perron et al (1985)

Plasmid pBTA641 can be derived from pBTA438 as follows.

35 pBTA438 is partially digested with XhoII plus DraI and a 1550bp fragment isolated and ligated to vector pLK58 cut with BglII and SmaI. The resultant plasmid pBTA446 was linearized with

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BglIII and ligated to a synthetic double stranded 27 mer
oligonucleotide having the sequence GATCT (N)₁₆ ATGGAG
wherein N represents any nucleotide, containing a bacterial
ribosome binding site and the initial nucleotides of the
5 native PAI-2 gene, creating plasmid pBTA641.

pBTA679 contains a 1.3kb fragment of the PAI-2 gene
(including the entire coding region). The 1.3kb fragment
was derived from the 1.6kb EcoRI-DraI fragment from pBTA438
by deleting 300bp of 3' untranslated region between the stop
10 codon and the DraI site (at 1604) using Bal 31 nuclease.
The resultant fragment with linkers ligated to either end
was subcloned into the HindIII-BamHI sites of the multiple
cloning site derived from pUC18 (Yanisch-Perron et al,
1985). The end-point of the deletion was determined by
15 sequencing.

pBTA830 is a mammalian cell expression vector.
Foreign genes are expressed by cloning into the multiple
cloning site (from pUC18) flanked upstream by the SV40 early
promoter and downstream by the SV40 small t intron and early
20 polyadenylation signals. pBTA830 comprises the following
fragments in order: the 345bp PvuII-HindIII fragment from
the SV40 origin (Bethesda Research Laboratories), 51bp
HindIII-EcoRI multiple cloning sites from pUC18, 853bp
XhoI-BamHI fragment from pMSG (a eukaryotic expression
25 vector from Pharmacia) with EcoRI and AatII linkers (GAATTC,
GGACGTCC, New England Biolabs) attached to either end,
2262bp AatII-EagI fragment from pBR327 (Soberon et al,
1980), 345bp PvuII-HindIII fragment from the SV40 origin,
1320bp HindIII-SmaI fragment from Tn5 encoding neomycin
30 phosphotransferase (Pharmacia, Beck et al, 1982), 141bp
Sau3A fragment from the SV40 small t intron region, 293bp
Sau3A fragment from the SV40 early polyadenylation region,
288bp EagI-SalI fragment from pBR327, 345bp PvuII-HindIII
fragment from the SV40 origin, 734bp HindIII-BglIII fragment
35 encoding mouse dihydrofolate reductase from pSV2-DHFR (ATCC
37146, Subramani et al, 1981), 141bp Sau3A fragment from
SV40 small t intron region and 293bp Sau3A from SV40 early

polyadenylation region. Except where indicated incompatible ends were made flush using S1 nuclease or filled in with dNTPs and DNA polymerase I (Klenow).

pBTA613 is a mammalian cell expression vector.

- 5 Foreign genes are expressed by cloning into the multiple cloning site flanked upstream by the SV40 early promoter and downstream by SV40 polyadenylation signals. pBTA613 comprises the following fragments in order. The 345bp PvuII-HindIII fragment from the SV40 origin, 51bp
- 10 HindIII-EcoRI multiple cloning sites from pUC18, 75bp EcoRI-AatII fragment from pBR327, 853bp BamHI-XhoI fragment from pMSG with AatII linkers attached to both ends, 2262bp AatII-EagI fragment from pBR327, 27bp oligonucleotide (GGCCCATATGATATCTCGAGACTAGTC), 288bp EagI-SaII fragment from
- 15 pBR327, 345bp PvuII-HindIII fragment from the SV40 origin, 734bp HindIII-BglII fragment encoding mouse dihydrofolate reductase from pSV2-DHFR, 141bp Sau3A fragment from SV40 small t intron region and 293bp Sau3A fragment from SV40 early polyadenylation region. The HindIII site at the 5'
- 20 end of the dhfr gene was deleted using S1 nuclease, other incompatible ends were made flush using S1 nuclease or filled in with dNTPs and DNA polymerase I (Klenow).

In Vitro Translation

- In pBTA 821, 822 and 823 the PAI-2 gene is placed
- 25 downstream from the T7 RNA polymerase promoter. To generate RNA transcripts from the native PAI-2 gene and the signal variants, plasmid DNA purified on CsCl density gradients was linearized then transcribed using the commercially available Riboprobe*Gene transcription System (Promega). Where
- 30 commercially available kits were used experiments were performed according to the manufacturer's instructions.

- Typically plasmid DNA was linearized by digestion with EcoRI. The EcoRI digested DNA was treated with proteinase K [at 400µg/ml] then extracted with
- 35 phenol/chloroform (1:1), precipitated with 2.5 vols ethanol and resuspended in 10mM Tris-Cl pH 8.0, 1mM EDTA. Approximately 1µg linear DNA was transcribed in a 25µl

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reaction containing 1 x transcription buffer (Promega), 10mM DTT, 0.5mM each ATP, CTP, GTP, UTP, 40 units RNAsin and 7.5 units T7 RNA polymerase (Promega) at 37° for 60 mins.

5 The RNA transcripts were subsequently translated in a rabbit reticulocyte lysate cocktail. Typically, 1µl of the transcription reaction (RNA) was added to a cocktail containing 8µl rabbit reticulocyte lysate (Amersham), 12.5µCi ³⁵S-Methionine (SJ1015 Amersham, 1000Ci/mMole), 20µM amino acid mix minus Met, 20 units RNAsin in a total
10 volume of 12.5µl. In some experiments 1Eq microsomes (Promega) was also included in the translation reaction. Incubation was at 30°C for 1hr.

Following translation, aliquots were removed for further treatment or analysis by SDS-polyacrylamide gel
15 electrophoresis.

To identify whether the synthetic or heterologous natural signal peptides attached to the NH₂-terminal end of PAI-2 were functioning and processed correctly, the reticulocyte lysate translation mix was supplemented with
20 dog pancreatic microsomal vesicles (Promega). Microsomes are generated from endoplasmic reticulum (ER) and allow signal peptide cleavage, protein translocation and core glycosylation events to be studied. Any core carbohydrates added to asparagine residues in PAI-2 molecules on
25 translocation across the ER are removed by digesting the translation products with endoglycosidase H. The extent of translocation of a protein across the membrane is assessed by incubating the processed in vitro translation products with proteinase K. Any of the protein not fully
30 translocated into the lumen of the microsomal vesicle is liable to be digested by proteinase K.

Endoglycosidase H digests were carried out by incubating 2µl translation mix plus 4µl RIPA buffer (1% Triton X100, 1% sodium deoxycholate, 0.1% SDS, 150mM NaCl,
35 50mM Tris-Cl pH7.5, 5mM EDTA) plus 1µg aprotinin with 2mUnits Endo H (Boehringer Mannheim) in a 10µl reaction at 37° for 1hr.

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Proteinase K digests were carried out as follows.

2 μ l translation mix plus 5 μ l phosphate buffered saline plus 0.1 μ g/ μ l proteinase K were incubated on ice for 60 mins. In some reactions 1 μ l 1% Triton X-100 was included.

5 Samples were boiled in 1.5% SDS, 1% 2-mercaptoethanol before electrophoresis on 10% polyacrylamide-SDS gels.

Figures 4A and 4B show the results of typical cell-free translation experiments. The autoradiogram in Figure 4B shows that the primary translation product of the native PAI-2 gene (pBTA821) consists of a 43,000 Mr molecule (Fig 4B lane b). It is not translocated into the microsomes since proteinase K digests the product whether microsomes were present (Fig. 4B lane d) or absent (Fig. 4B lane a). Also the native product is not glycosylated since digestion with endoglycosidase H (Figure 4A lane d) has no effect on the migration of the Mr 43,000 product. This 43,000 Mr product was also immunoprecipitated with anti-PAI-2 monoclonal antibody. In contrast the primary translation products of both signal-PAI-2 variants (pBTA822 and pBTA823, see Figure 4A, lanes g and l) have higher molecular weights (44,000-45,000) than the native PAI-2 (Fig. 4A, lane b) due to the additional signal peptide. When microsomes are included in the reaction, the translation products of pBTA822 and pBTA823 migrate more slowly (Mr 45,000-46,000) due to processing of the signal and addition of carbohydrates. There are two distinct products (44,500 and 46,000 Mr) which are fully protected from digestion with proteinase K (Figure 4B, compare lanes i and j, or o and p). Both these products are reduced in size when digested with Endo H (Figure 4A, compare lanes h and i or m and n) to 43,000 and 41,000 Mr. These results indicate that the addition of either signal peptide to PAI-2 allows PAI-2 to be translocated across the endoplasmic reticulum membrane and to be glycosylated. Furthermore the added signal is cleaved off during translocation.

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Transient Expression In Mammalian Cells

For details of tissue culture techniques used see Freshney (1987). To determine whether the signals attached to PAI-2 enhanced the processing and secretion of PAI-2 from mammalian cells, the native PAI-2 and the two signal variant PAI-2 genes were placed downstream of the SV40 late promoter in the eukaryotic expression vector pSVL (Pharmacia). The construction of these vectors pBTA824, pBTA825 and pBTA826 was described above and is summarized in Figure 3.

Plasmids pBTA824, pBTA825, pBTA826 were transfected into the monkey kidney COS cell line by the calcium phosphate method (Spandidos and Wilkie, 1984). Approximately 6×10^5 cells were seeded into a 100mm culture dish in 10ml Dulbecco's modified Eagle medium supplemented with 10% (v/v) foetal calf serum, 2mM glutamine, 50IU ml penicillin and 50µg/ml streptomycin. Ten micrograms of CsCl purified DNA was precipitated with calcium phosphate and added to the cells. After 4 hrs the cells were treated to a glycerol shock and cultured in the above medium for about 42hrs.

To assay for expression and secretion of PAI-2 from these cells, the medium was removed about 42 hrs after glycerol shock and replaced with Eagles minimal essential medium lacking methionine (Flow) supplemented with 100µCi/ml ^{35} S-methionine (Amersham SJ1015 1000Ci/mmol). After 4 hrs of metabolic labelling the medium was collected for analysis and the cells washed with an ice cold phosphate buffered saline solution, harvested and lysed. Cells were lysed in 1% Triton X100, 0.1% SDS, 150mM NaCl, 50mM Tris-Cl pH7.5, 5mM EDTA. PAI-2 was immunoprecipitated from aliquots of medium and cell lysates by incubating with 25µg/ml anti-PAI-2 mouse monoclonal antibody (MAI-21, Biopool) at 4°C overnight in a solution containing 1% Triton X100 and 10µg/ml aprotinin. Antibody-PAI-2 complexes were extracted with 20µl anti-mouse IgG-agarose beads (Sigma) in 10mM sodium phosphate pH7.2, 0.5M NaCl. The PAI-2 was eluted from the

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beads with 0.15M NaCl/0.1M glycine pH2.4, analysed by SDS-polyacrylamide gel electrophoresis under reducing conditions and visualized by autoradiography as shown in Figure 5. Some cells were incubated with 5µg/ml

5 tunicamycin for 18 hrs prior to and during the labelling, to prevent N-linked glycosylation.

Recombinant PAI-2 is seen in Fig 5, lanes f and h as a broad band of Mr 53,000-60,000. These are samples of medium from cells transfected with the synthetic
10 signal-PAI-2 gene (pBTA825) and α-1-antitrypsin signal-PAI-2 gene (pBTA826) respectively. The medium from mock transfected cells (i.e. no added DNA) and from cells transfected with the vector alone (pSVL) contain no 60,000 Mr material (lanes b and c). However a small amount of
15 60,000 Mr material and a larger amount of 43,000 Mr material is present in the medium from pBTA824 transfected cells (lane d). The 43,000 Mr material appears not to be glycosylated (compare lanes d and e).

Clearly the addition of the signal peptide to the
20 PAI-2 molecule greatly enhances the secretion of the 60,000 Mr glycosylated form from COS cells.

The biological activity of the 58-60,000 Mr forms of PAI-2 were determined by a urokinase (uPA) binding assay. Urokinase (45 IU Low Mol Wt, American Diagnostica) was added
25 to an aliquot of medium containing ³⁵S-labelled PAI-2 from transfected COS cells. Binding was allowed to proceed at room temperature for 90 mins. The uPA-PAI-2 complexes were immunoprecipitated from the reaction using goat anti-PAI-2 antibodies or rabbit anti-uPA serum and a solid phase 2nd
30 antibody. Material was eluted from the immunobeads by boiling in a buffer containing 1.5% SDS and 1% (v/v) 2-mercaptoethanol and then analysed under reducing conditions by SDS-PAGE.

Figure 6 shows the results of such an experiment.
35 The Mr 43,000 nonglycosylated PAI-2 released from COS cells transfected with the native gene (Fig 6, lane c) forms a Mr 76,000 complex when uPA is added

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(Fig 6, lane d). The 58-60,000 glycosylated PAI-2 secreted from COS cells transfected with either of the signal-PAI-2 constructs also binds uPA and a complex of Mr around 89,000 is formed (Fig. 6, lanes f and h). This complex is immunoprecipitable with both anti-PAI-2 and anti-uPA antibodies. This demonstrates that the 58-60kD glycosylated and secreted form of PAI-2 is biochemically active.

Carbohydrate Attachment

To determine the nature of the carbohydrates attached to the 60,000 Mr PAI-2, the material secreted from COS cells was digested with a variety of endoglycosidases (see Glycanalysis Systems Manual from Genzyme Corp. USA). The 60,000 Mr form was immunoprecipitated from the medium of cells transfected with pBTA826 as described above. Aliquots of this material were digested as follows.

- (a) 1% Triton X-100, 1% Na deoxycholate, 0.1% SDS, 150mM NaCl, 50mM Tris-Cl pH 7.5, 5mM EDTA, 2mUnits Endoglycosidase H (Boehringer Manneheim) at 37°C for 1hr.
- (b) 1% Triton X-100, 0.1% SDS, 10mM Na phosphate pH7.2, 400 mUnits glycopeptidase F (Boehringer Mannheim) at 37° for 1hr.
- (c) 1% triton X-100, 0.1% SDS, 10mM D-galactono γ lactone, 10mM Na phosphate pH7.2, 1m Unit neuraminidase and/or 1mU endo- α -N-acetylgalactosaminidase (Boehringer Mannheim) at 37° for 1hr.

After digestion the products were analysed by SDS-PAGE and autoradiography. Digestion with endo H had no effect on the mobility of 60,000 Mr PAI-2 (Figure 9, lane B) indicating that any N-linked carbohydrate contained complex sugars added in the Golgi. However digestion with glycopeptidase F (which removes all N-linked carbohydrate) reduced the 60,000 Mr material to a doublet of Mr 49,000 and 50,000 (Figure 9, lane C). These forms have higher molecular weights than unglycosylated PAI-2 (43,000 Mr). The possibility that some O-linked carbohydrate was also

attached to PAI-2 was confirmed by digesting the 60,000 Mr form with glycopeptidase F, neuraminidase and endo- α -N-acetyl galactosaminidase (which removes O-linked sugars). The combination of these three enzymes reduced the 60,000 Mr form to a 43,000 Mr form (Figure 9, lane E), the size of unglycosylated PAI-2 (Figure 9 lane F).

NH₂ Terminal Determination

To determine the NH₂-terminal residue of the secreted 60,000 Mr form of PAI-2, ³⁵S-methionine or ³H-leucine labelled material secreted from transfected COS cells were subjected to immunoprecipitation using the anti-PAI-2 mouse monoclonal antibody (MAI-21, Biopool) as described above.

An aliquot of the ³⁵S-Met labelled PAI-2 recovered from the medium was analyzed by SDS-polyacrylamide gel electrophoresis and shown to contain 60,000 Mr material. The ³⁵S-Met labelled 60,000 Mr PAI-2 was applied to the Applied Biosystems Protein Sequencer (Model 470A) and subjected to repeated cycles of Edman degradation (see Instruction Manual for Protein Sequencer). Fractions from each cycle were collected and counted in a Liquid Scintillation Counter. The results of this analysis (Figure 10A) indicated that the NH₂ terminal residue of the 60,000 Mr PAI-2 secreted from COS cells transfected with pBTA825 is Met.

The ³H-leu labelled 60,000 Mr forms of PAI-2 were electrophoresed through an SDS-polyacrylamide gel and transferred to Pro Blot* PVDF membrane by electroblotting (Matsudaira, 1987). A fragment of the membrane containing the 60 kDa PAI-2 was excised and then applied to the Protein sequencer and sequenced as above with modifications to the Sequencer program as described by Speicher, 1989.

The results (Figures 8A, 8B) indicated that the 60,000 Mr PAI-2 secreted from COS cells transfected with pBTA825 has Leu residues at positions 4, 10 and 13, i.e. the synthetic signal peptide has been cleaved to leave PAI-2

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with Met at its NH₂-terminus. However, the 60,000 Mr PAI-2 secreted from COS cells transfected with pBTA826 has Leu residues at positions 3, 9 and 12. This indicates that the alpha-1-antitrypsin signal has been cleaved off leaving a glycosylated PAI-2 molecule with Glu at its NH₂-terminus.

These results demonstrate that the signal peptides have been cleaved during processing of PAI-2 through the cell, that cleavage of the synthetic signal peptide occurs between Ala 19 and Met 20 (Figure 7), that cleavage of the alpha-1-antitrypsin signal occurs between Ala 24 and Glu 25 (Figure 8). The 60,000 Mr PAI-2 secreted from COS cells transfected with pBTA825 is 415 amino acid residues. However, the 60,000 Mr PAI-2 secreted from COS cells transfected with pBTA826 contains 414 amino acid residues.

Stable Expression in Mammalian Cells

For details of tissue culture techniques used see Freshney (1987). The stable expression of PAI-2 genes integrated into host cell genomes was investigated in various cell lines. Cell line CHO-K1 (ATCC CCL 61) is derived from a Chinese hamster ovary cell. Cell line U937 is the human monocyte-derived cell which expresses PAI-2 and from which the PAI-2 gene was isolated (Antalis et al, 1988). Other host cells for the expression of PAI-2 include the hamster kidney cell line BHK-21 (ATCC CCL10), the human kidney cell line 293 (ATCC CRL1573), the human epithelial cell-derived line HeLa S3 (ATCC CCL2.2), and the human monocyte cell line HL60 (ATCC CCL 240) and insect cell lines derived from Spodoptera frugiperda (Sf9, ATCC CRL1711) and Bombyx mori (Maeda et al, 1984).

The PAI-2 genes with added signal sequences were inserted into the vector pBTA830 which contains the mouse dhfr gene and the gene conferring resistance to the aminoglycoside antibiotic, G418 (Figures 3B and 3C). The resultant plasmids pBTA827, pBTA828, were transfected into the above cell lines by the calcium phosphate method described, using 20µg plasmid DNA/2 x 10⁵ cells and a total of 3 x 10⁶, or by electroporation. Electroporation

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was carried out using 10µg linearized plasmid DNA/10⁷ cells suspended in phosphate buffered saline/272 mM sucrose, pH 7.4, at 4°C, and pulses of 150-300V at a capacitance of 250µFD (Ausubel et al, 1987). The apparatus used was a Bio-Rad Gene Pulser.

The selection of transfectants of the CHO-K1 cell line was achieved in the nucleoside-free alpha modification of Eagles medium (αMEM) containing 2% (v/v) dialysed foetal bovine serum and 400µg/ml G418. This regime selected for both the dhfr and the G418-resistance genes present in the PAI-2 encoding plasmid. Selection of transfectants of other cell lines was done with αMEM or RPMI 1640 medium (CytoSystems) containing 2-10% foetal bovine serum and 400µg/ml G418. After selection, the PAI-2 genes were amplified along with cotransfected (and host) dhfr genes by exposure of the cultures to increasing levels of methotrexate from 0.1µM to 10µM over three months as described by Zettlemeyers et al. (1987).

The resultant transfected cultures with stably integrated, amplified PAI-2 genes were examined for the secretion of PAI-2 into serum-free culture medium by an ELISA (Biopool, Sweden) performed according to the manufacturers instructions. Different cultures (pools) containing cells originating from many hundreds of transfection events were assayed by ELISA and the pools with the highest PAI-2 expression were chosen. The identity of the molecular weight forms of PAI-2 secreted from these cultures was verified by immunoprecipitation of culture supernatants and SDS-polyacrylamide gel electrophoresis as described above. High molecular weight, glycosylated forms of PAI-2 are produced by representative cultures as shown in Figure 11. The biological activity of the secreted forms of PAI-2 can be determined using a coupled photometric assay for plasminogen activator (Coleman and Green, 1981). The first step of the reaction involves the activation of purified plasminogen to plasmin by uPA. The activity of plasmin is then detected by the formation of a yellow thiophenolate anion. PAI-2 activity is expressed in Plouq units/ml.

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Finally, clones of individual cells from the highest expressing cell pool are isolated by the limiting dilution method (Freshney, 1987). Cell colonies are assayed for PAI-2 by transferring them to nylon filters as described by Raetz et al. (1982) and detecting PAI-2 by Western blot analysis. Colonies with maximal PAI-2 expression are isolated, re-cloned, and the secreted forms of PAI-2 verified as described above.

The stable expression of PAI-2 genes integrated into host genomes was investigated further in various cell lines including CHO, DG44 and Vero. Cell line DG44 is a derivative of the Chinese hamster ovary cell CHO-K1 and contains a deletion of the gene for dihydrofolate reductase (dhfr. Urlaub et al., 1986). Vero is derived from an African green monkey kidney cell (ATCC CCL81).

The PAI-2 gene with added signal sequence was inserted into the vector pBTA613 which contains the mouse dhfr gene (Figure 3D). The resultant plasmid pBTA839 was transfected into the above cell lines by the calcium phosphate method described, using $10\mu\text{g}$ plasmid DNA/ 2×10^5 cells and a total of 2×10^6 cells, or by electroporation. Electroporation was carried out as described by Barsoum (1990) using the Bio-Rad Gene Pulser.

The selection of transfectants of cell line DG44 was achieved in the nucleoside-free medium α MEM (CytoSystems) containing 2% (v/v) dialysed foetal bovine serum and 0.1-0.5 μM methotrexate. The selection of transfectants of Vero cells was done in the same medium. This procedure selects for transfectants with an elevated number of copies of integrated plasmid. After selection, the PAI-2 genes were amplified along with cotransfected (and host) dhfr genes by exposure of the cultures to increasing levels of methotrexate up to 10 μM as described by Zettlemeisl et al (1987). The resultant transfected culture with stably integrated, amplified PAI-2 genes are examined for the secretion of PAI-2 into the serum-free culture medium by an ELISA for PAI-2 (Biopool, Sweden) performed according to the manufacturers instructions. Single colonies arising from

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the transfection and pools containing cells originating from many thousands of transfection events are assayed by ELISA and those with the highest PAI-2 expression are chosen. The identity of the molecular weight forms of PAI-2 secreted from these cultures is verified by immunoprecipitation of culture supernatants and SDS-polyacrylamide gel electrophoresis as described above.

Clones of individual cells are also isolated from the highest expressing cell pool by limiting dilution method (Freshney, 1987). Cell colonies are assayed for PAI-2 by transferring them to nylon filters as described by Raetz et al. (1982) and detecting PAI-2 by Western blot analysis. Colonies with maximal PAI-2 expression are isolated, re-cloned, and the secreted forms of PAI-2 verified as described above.

Absence of Efficient Signal Sequence from Native PAI-2

The in vitro translation experiment with microsomes has for the first time demonstrated that native PAI-2 does not contain a signal sequence that is efficiently recognized by the protein translocation machinery present in some mammalian cell extracts. This may explain the observations that PAI-2 is predominantly an intracellular protein. The little material that is secreted is often unglycosylated and may be released from the cells by an unknown mechanism which does not involve transport of PAI-2 through the normal secretory pathway of the ER and Golgi. However high Mr forms of PAI-2 have also been observed; hence PAI-2 must have the capacity to translocate into the ER and Golgi (where addition of carbohydrate occurs). The signal to allow this has not been identified. Given the results with the in vitro system described above any signal in native PAI-2 must be very inefficiently recognized by the signal recognition protein. To overcome this problem, and to ensure efficient secretion of large amounts of high molecular weight forms of PAI-2 from mammalian cells we have added a signal peptide to the NH₂-terminus of PAI-2. Results described above show that either an artificial

signal peptide or a natural signal peptide can be attached to PAI-2 to create a molecule which is translocated across the endoplasmic reticulum. The signal is cleaved and PAI-2 is processed through the secretory pathway. These high molecular weight forms of PAI-2 are glycosylated and bind to uPA.

Purification

The secreted PAI-2 is purified using established and published procedures (see International Patent Application No. PCT/AU87/00068 (filed March 13, 1987 in the name of Biotechnology Australia Pty. Ltd. and the Australian National University)).

Industrial Applications

The expression products, secreted glycosylated recombinant 60 kD PAI-2 and the 414 amino acid form of PAI-2 of the invention are of use as diagnostic or therapeutic substances in the fields of:

therapy, prophylaxis and diagnosis of inflammatory disease;

therapy, prophylaxis and diagnosis of cancer metastasis and proliferation;

therapy of transplant and placenta rejection crises and graft-versus-host reactions;

therapy and diagnosis of autoimmune diseases and diseases associated with excessive oxygen radical production;

therapy and prophylaxis of wound and bone healing disorders, tissue damage during or after acute reperfusion, and subarachnoidal bleeding disorders;

as topical medicaments for promoting fibrin adhesion, promoting healing of wounds and burns;

treating asthma and treating or preventing diseases associated with high leukocyte activity; and suppression of the monocyte-macrophage system.

Specific examples from the above classes of disease include:

rheumatoid arthritis

osteoarthritis

colitis ulcerosa

systemic lupus erythematosus

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multiple sclerosis

psoriasis

pemphigus

corneal and duodenal ulceration

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purpura

periodontitis

muscular dystrophy

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International Patent Application PCT/AU87/00107.

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THE EMBODIMENTS OF THE INVENTION IN WHICH AN EXCLUSIVE
PROPERTY OR PRIVILEGE IS CLAIMED ARE DEFINED AS FOLLOWS:

1. A process for the preparation of recombinant, glycosylated, secreted PAI-2
or recombinant, glycosylated, secreted PAI-2 without its N-terminal methionine,
5 which process comprises:
 providing a construct comprising a first polynucleotide molecule encoding
PAI-2 or PAI-2 without its N-terminal methionine;
 attaching a polynucleotide molecule encoding the transient signal sequence
M K C L L L A L G L L A F V P L V R A to the 5' end of the first polynucleotide
10 molecule, such that the resulting hybrid protein expressed from the construct will
consist of the transient signal sequence attached directly to the NH₂-terminal of
PAI-2 or PAI-2 without its N-terminal methionine; and expressing the construct in
a eukaryotic host cell.
2. A polynucleotide construct encoding the transient signal peptide
15 M K C L L L A L G L L A F V P L V R A attached to the NH₂-terminal
methionine or the NH₂-terminal glutamic acid of PAI-2.
3. A polynucleotide construct according to claim 2 wherein the construct is a
DNA molecule.
4. A recombinant DNA molecule comprising a DNA molecule according to
20 claim 3 and vector DNA.
5. A recombinant DNA molecule according to claim 4 wherein the vector
DNA is plasmid DNA.
6. A recombinant DNA molecule according to claim 5 wherein the plasmid
DNA is selected from:
25 pGEM4Z, pSVL, pBTA613, pBTA830 and baculovirus transfer vectors.
7. A recombinant DNA molecule selected from pBTA822, pBTA823, pBTA825,
pBTA826, pBTA827, pBTA828 and pBTA839.

8. A eukaryotic host cell transformed by a recombinant DNA molecule according to claim 4.
9. A eukaryotic host cell according to claim 8 wherein the host cell is from a cell line derived from monkey kidney COS cells, the monkey kidney cell line
5 Vero, Chinese hamster ovary cells, the human histiocytic lymphoma U937 cell line, a derivative of CHO-K1 cells, DG-44, the hamster kidney cell line BHK-21, the human kidney cell line 293, the human epithelial cell derived line HeLa S3, the human monocyte cell line HL60 and cell lines derived from the insects *Spodoptera frugiperda* and *Bombyx mori*.
- 10 10. A process for preparing a recombinant DNA molecule which process comprises inserting a DNA molecule according to claim 3 into vector DNA.
11. A process for preparing a host cell which process comprises making a host cell competent for transformation and transforming the host cell with a recombinant DNA molecule according to claim 4.

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FIGURE 1

OLIGO A1 (20mer) - adaptor for cloning native PAI-2 gene into expression vectors

5' AGCTTGTGACACCATGGAA 3'
M E

OLIGO A2 (20 mer) - complementary to A1

5' GATCTTCCATGGTGTGCGACA 3'

OLIGO B1 (77mer) - encodes synthetic signal peptide and signal peptidase recognition site

5' AGCTTGTGACACCATGAAATGTCTGCTCGCTCTCGGCTCTGCTAGCTTTCGTGCCGTTGGTGAGGGCTATGGAA 3'
M K C L L L A L G L L A F V P L V R A M E

OLIGO B2 (77mer) - complementary to B1

5' GATCTTCCATAGCCCTCACCCAACGGCACCAGAAAGCTAGCAGACCCGAGCGAGCAGACATTTTCATGGTGTGCGACA 3'

OLIGO C1 (89mer) - encodes alpha -1- antitrypsin signal and signal peptidase cleavage site

5' AGCTTGTGACACCATGCCCTTCTTCTGCTCTTGGGGCATCCTGCTGCTAGCAGGCCCTGTCTGTCTGCTCCCTGTCTCTGGCTGAA 3'
M P S S V S W G I L L L A G L C C L V P V S L A E

OLIGO C2 (89mer) - complementary to C1

5' GATCTTCCAGCCACAGACAGGACCAACACAGGCCTGCTAGCAGCAGGATGCCCCAAGAGACAGAAGGCATGGTGTGCGACA 3'

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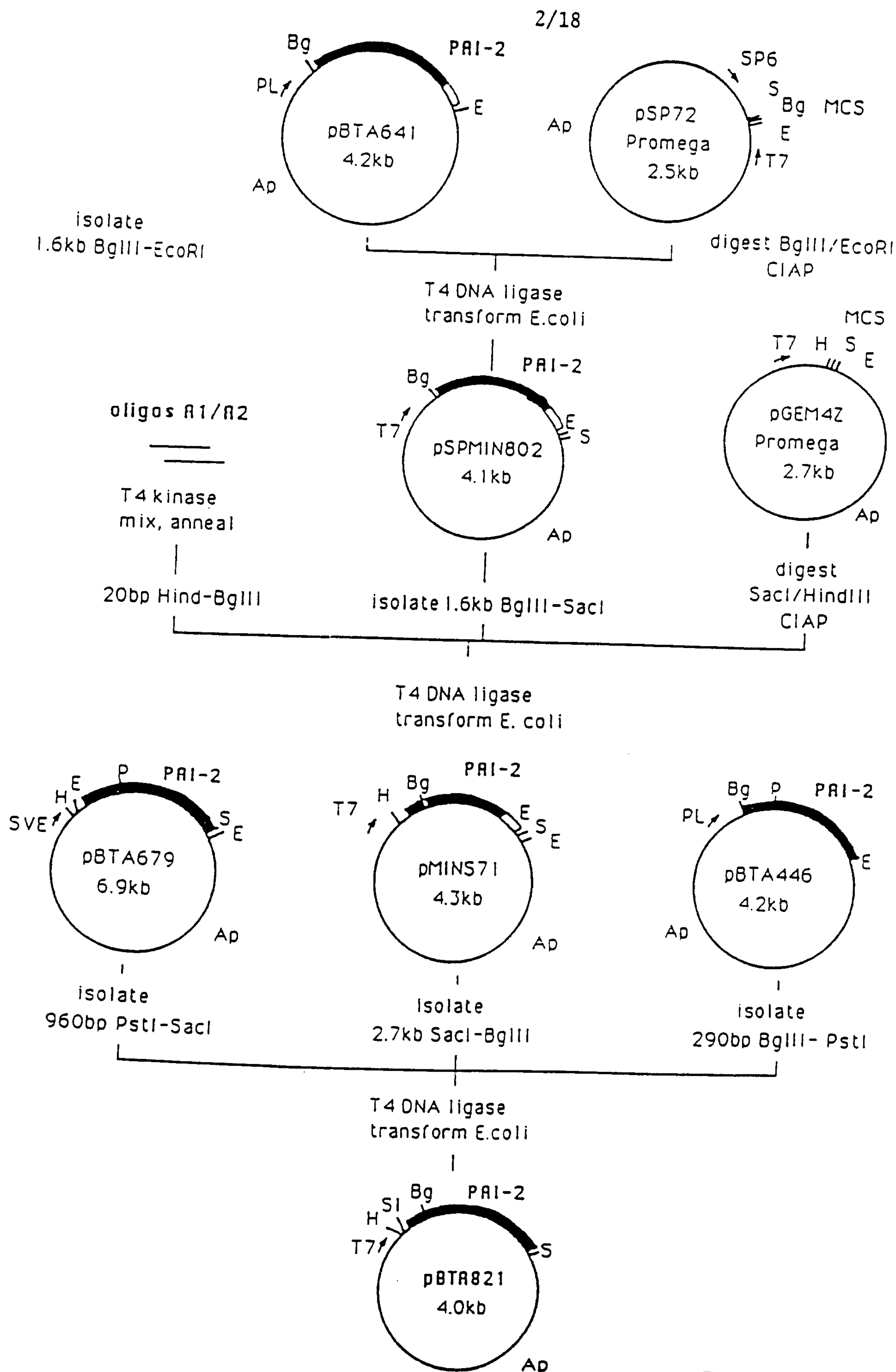


FIGURE 2A

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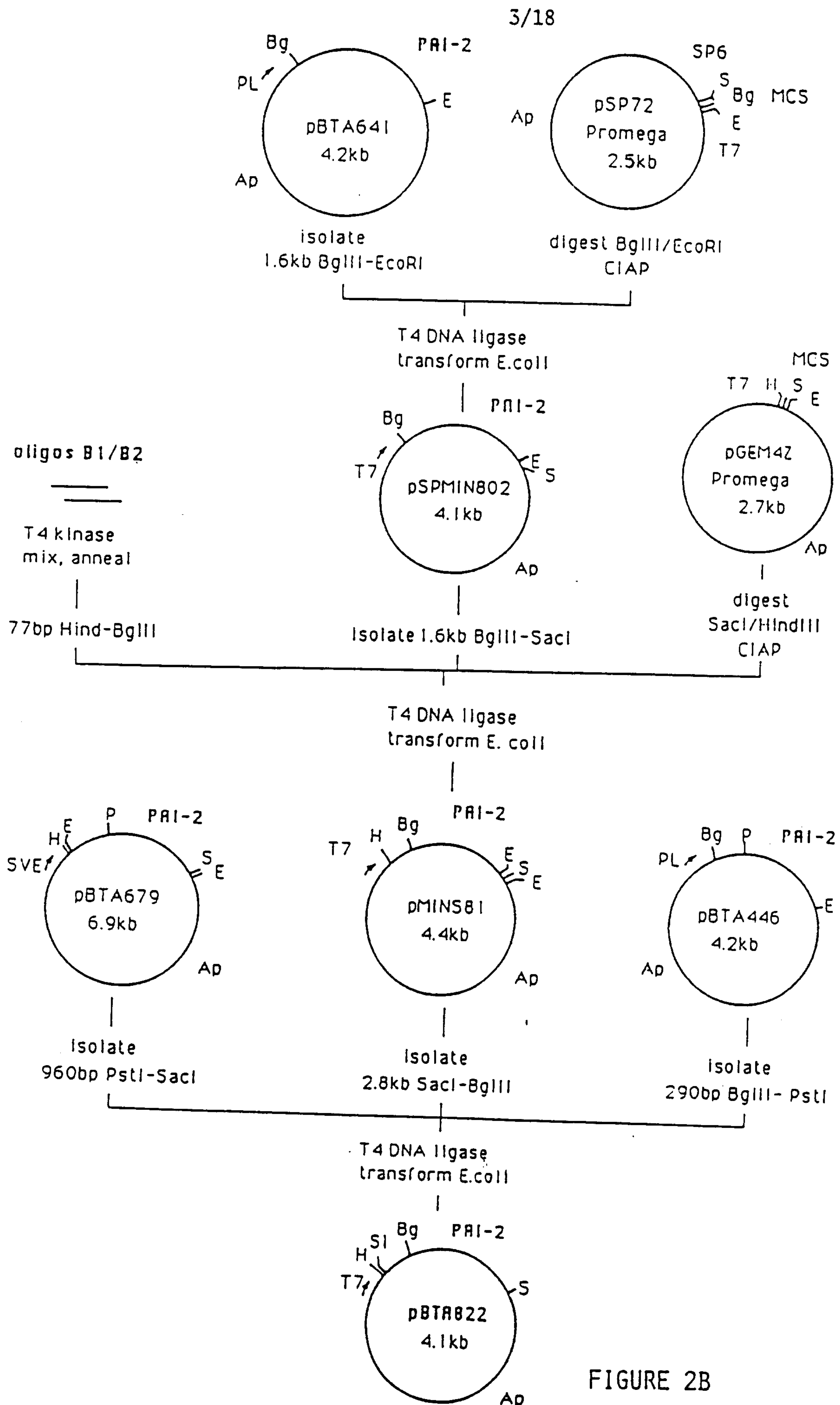


FIGURE 2B

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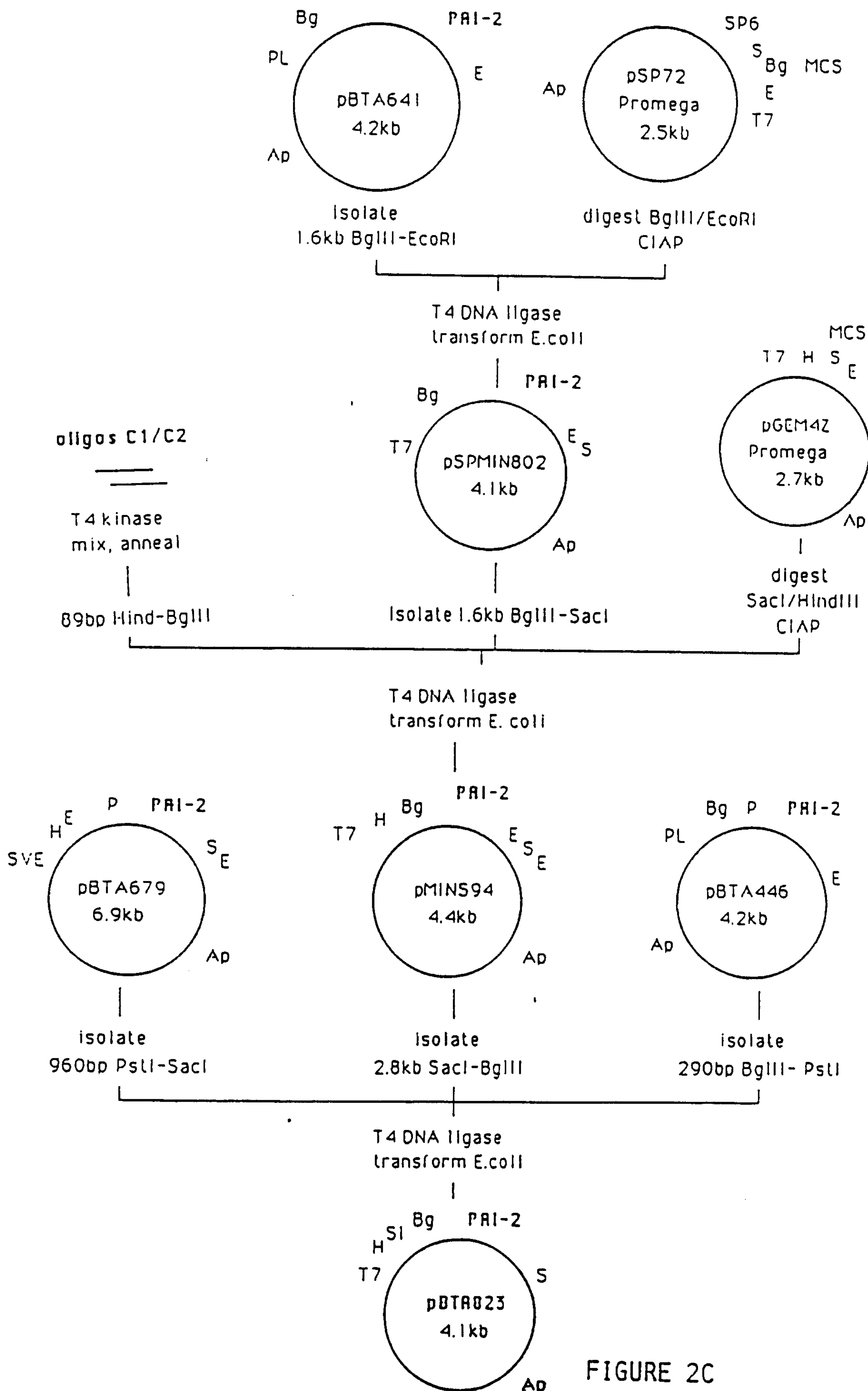
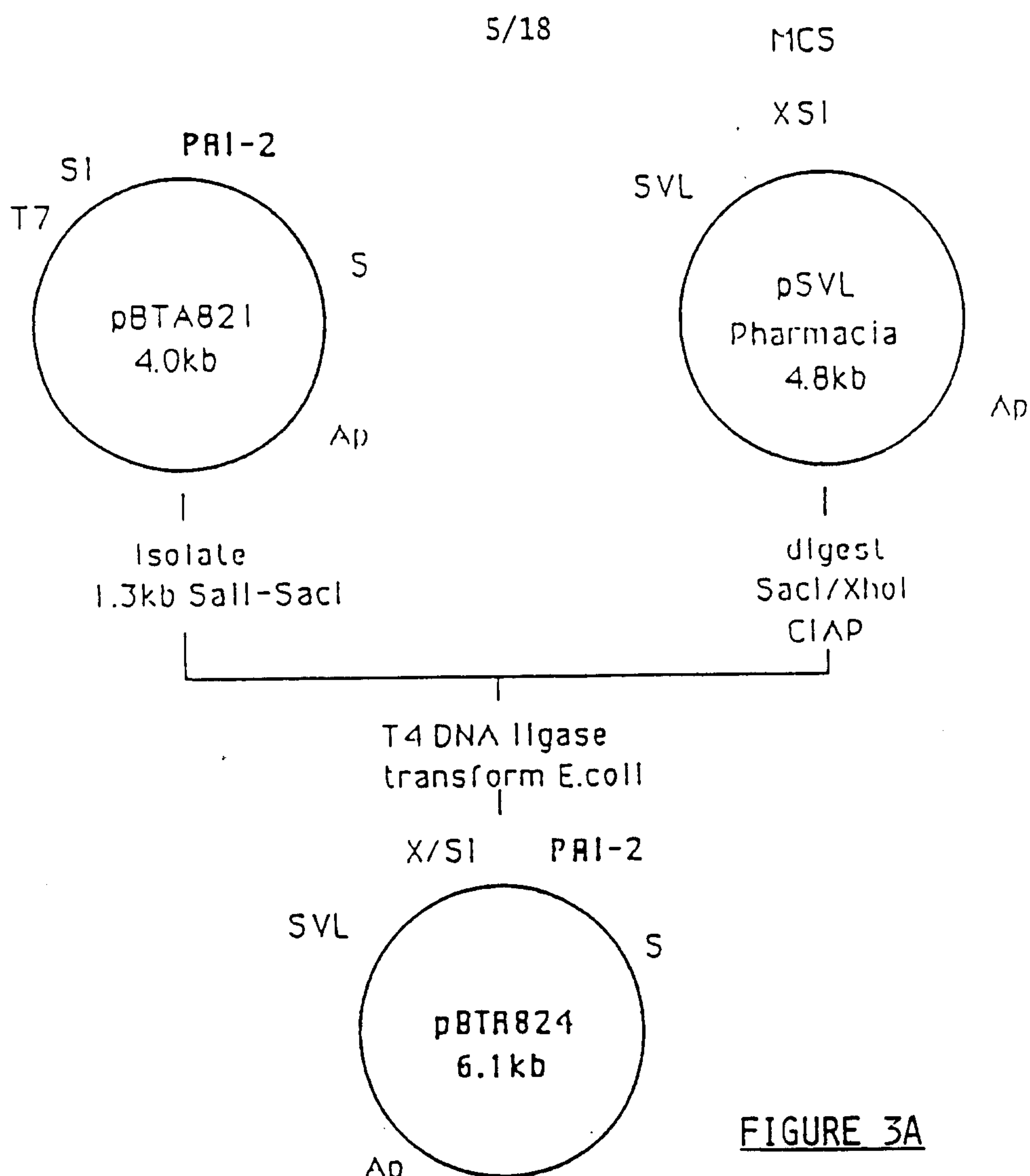


FIGURE 2C

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FIGURE 3AFIGURES 2 AND 3

ABBREVIATIONS :

PROMOTERS

PL - LAMBDA PL
 T7 - T7 RNA POLYMERASE
 SP6 - SP6 RNA POLYMERASE
 SVE - SV40 EARLY
 SVL - SV40 LATE

RESTRICTION SITES

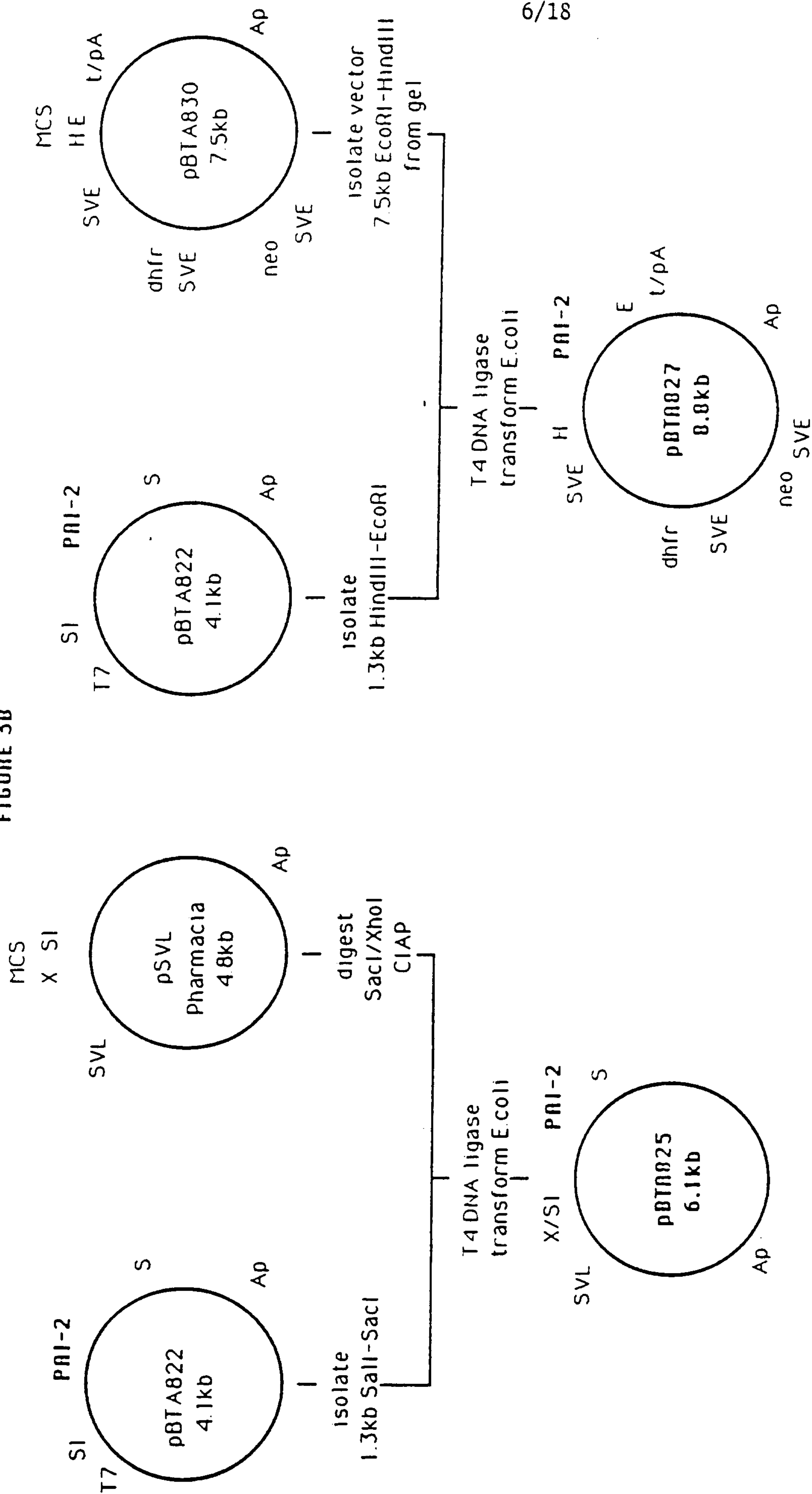
BG - BGLII
 E - ECORI
 H - HINDIII
 P - PSTI
 S - SACI
 SI - SALI
 X - XHOI

AP - AMPICILLIN RESISTANCE GENE
 DHFR - DIHYDROFOLATE REDUCTASE GENE
 NEO - G418/KANAMYCIN RESISTANCE GENE
 MCS - MULTI-CLONING SITE
 T/PA - SV40 T INTRON AND POLYADENYLATION SIGNALS

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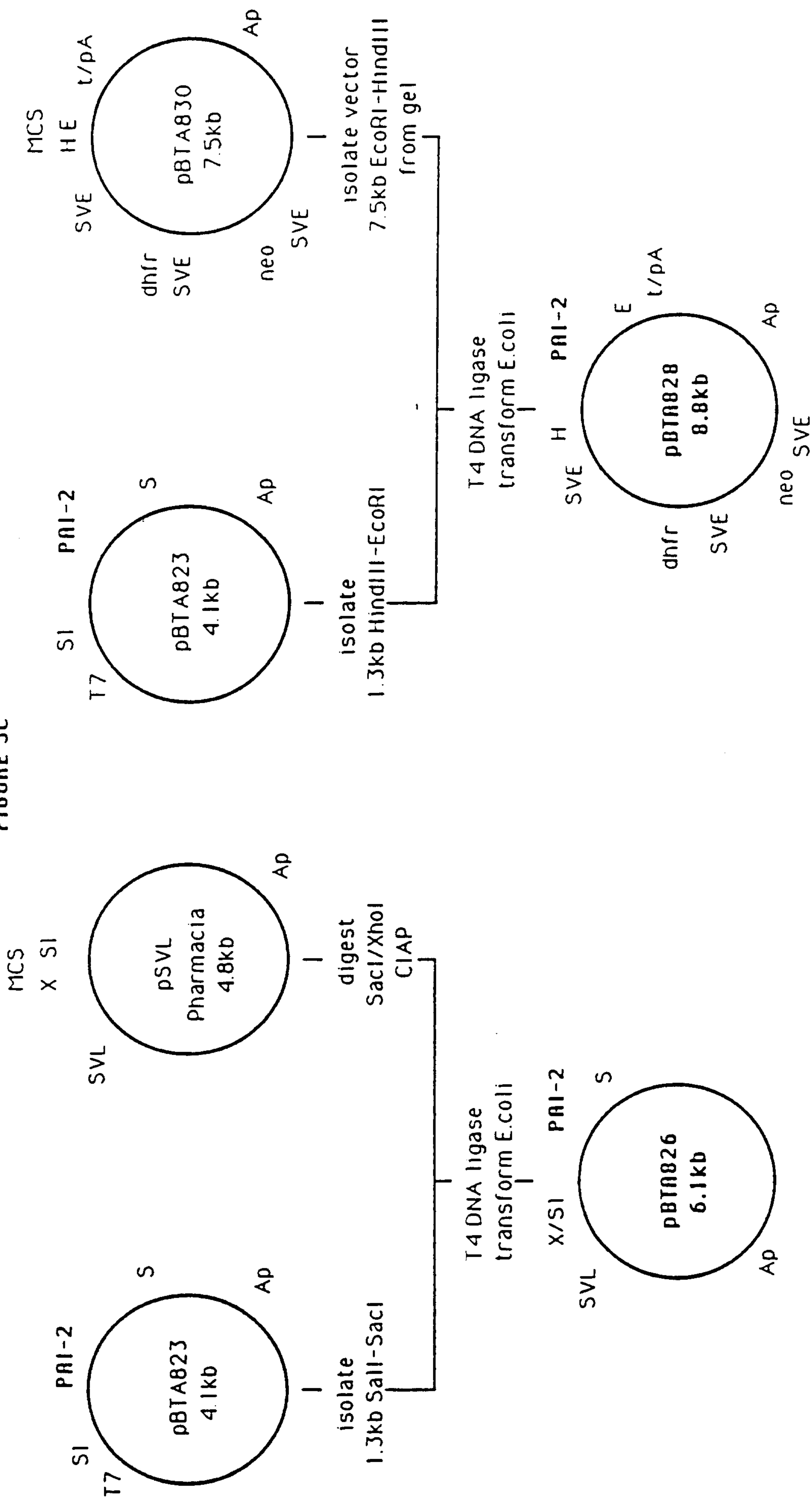
FIGURE 3B



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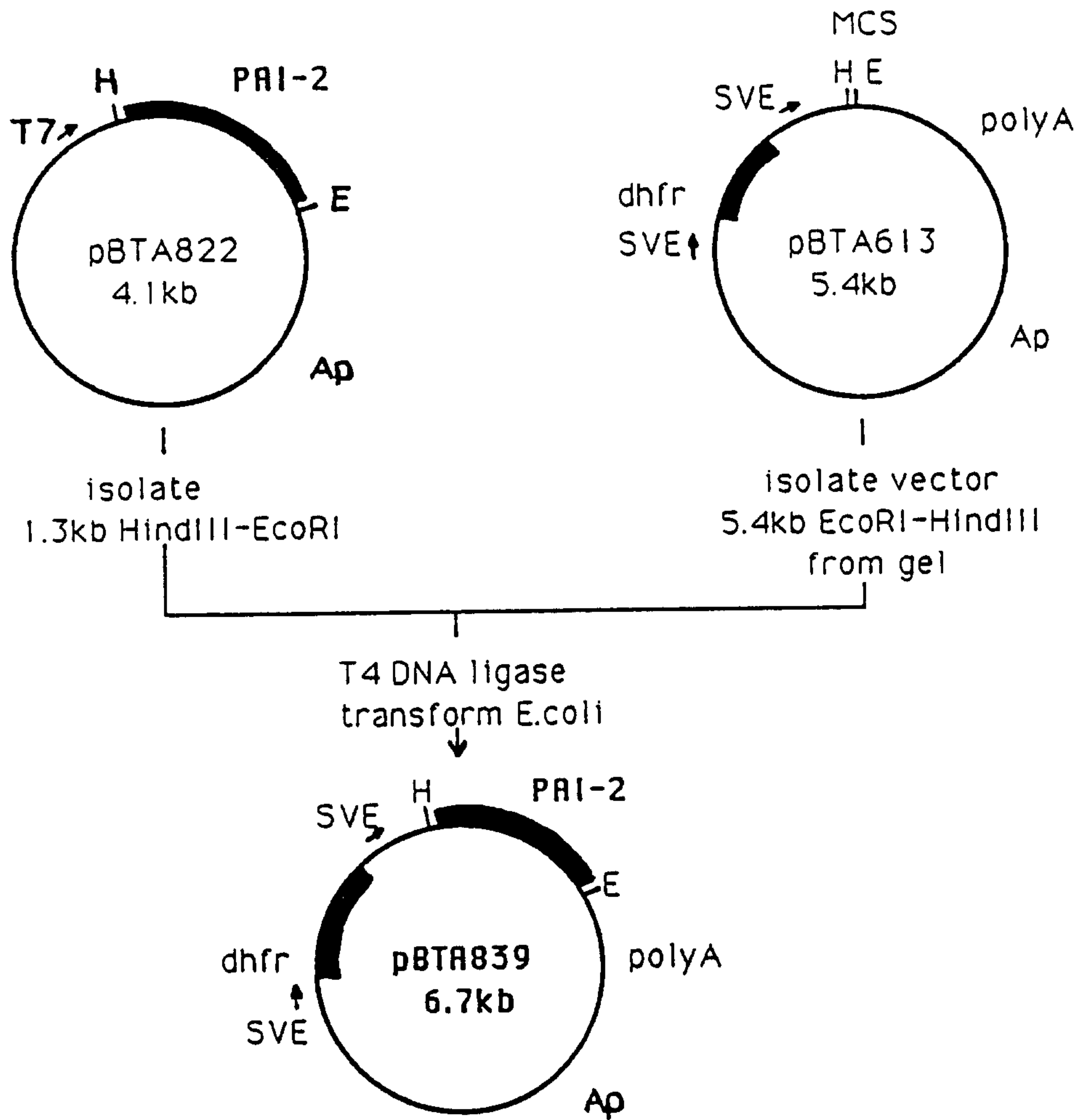
FIGURE 3C



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FIGURE 3D



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FIGURE 4A

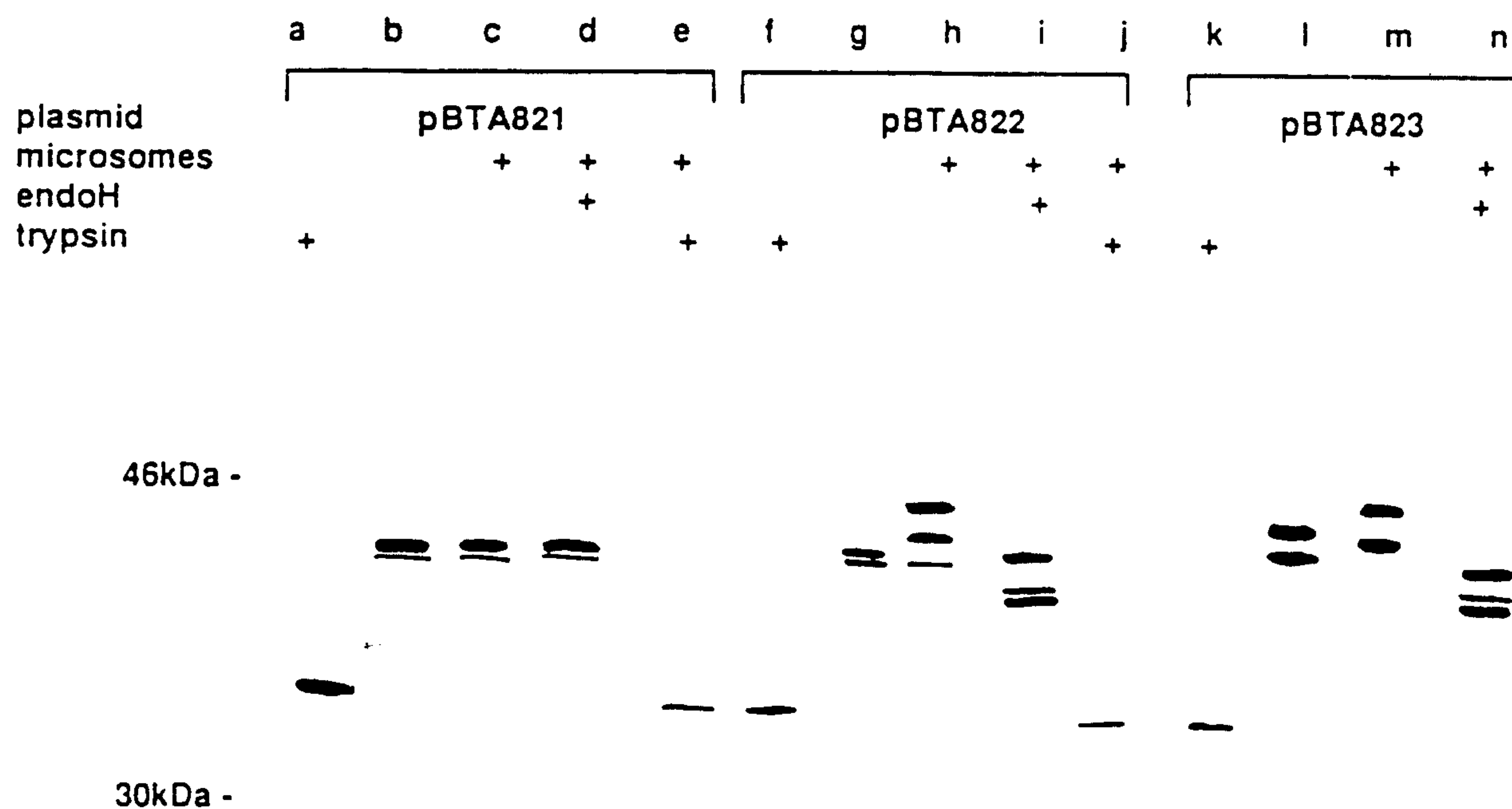
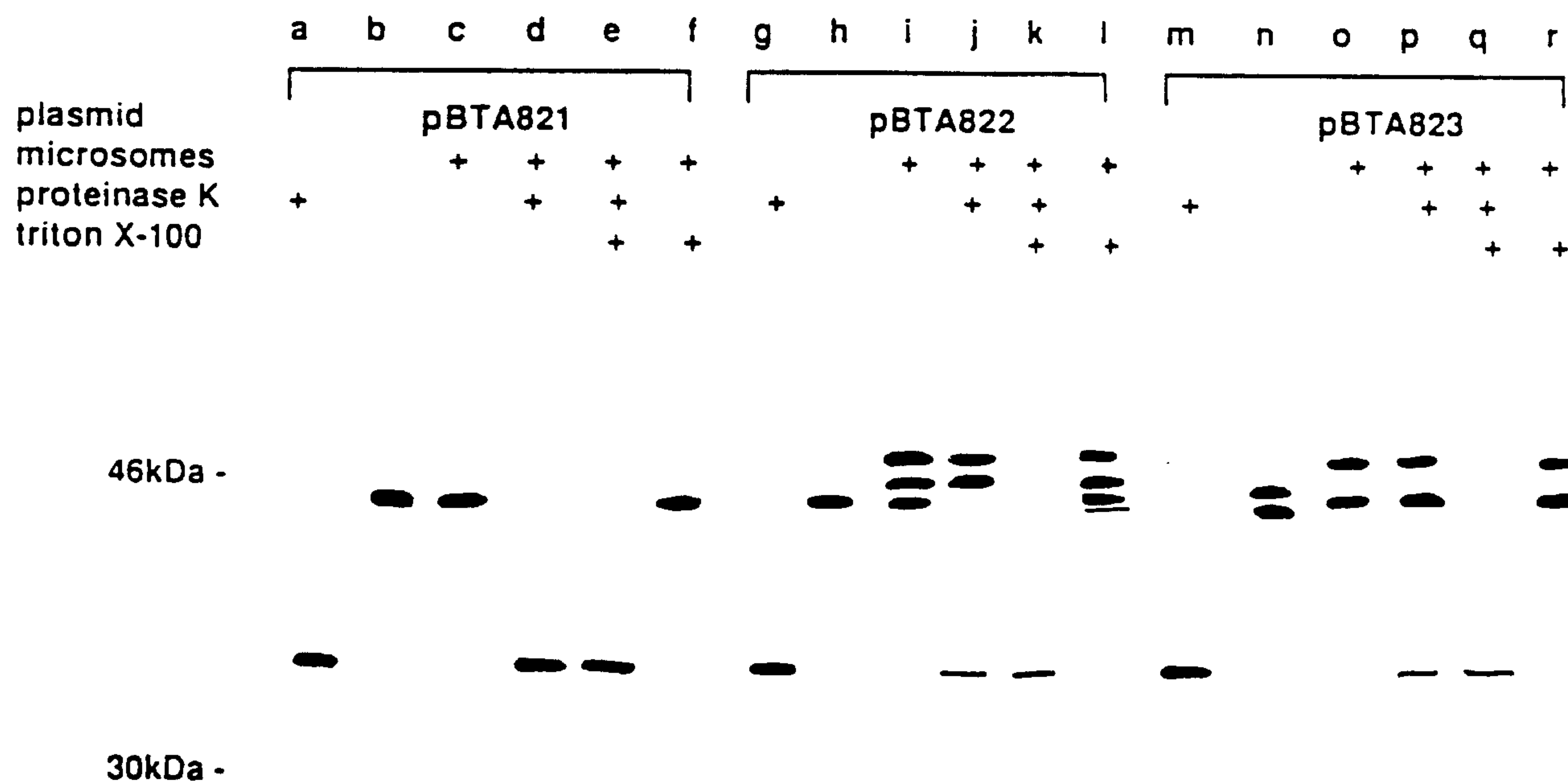


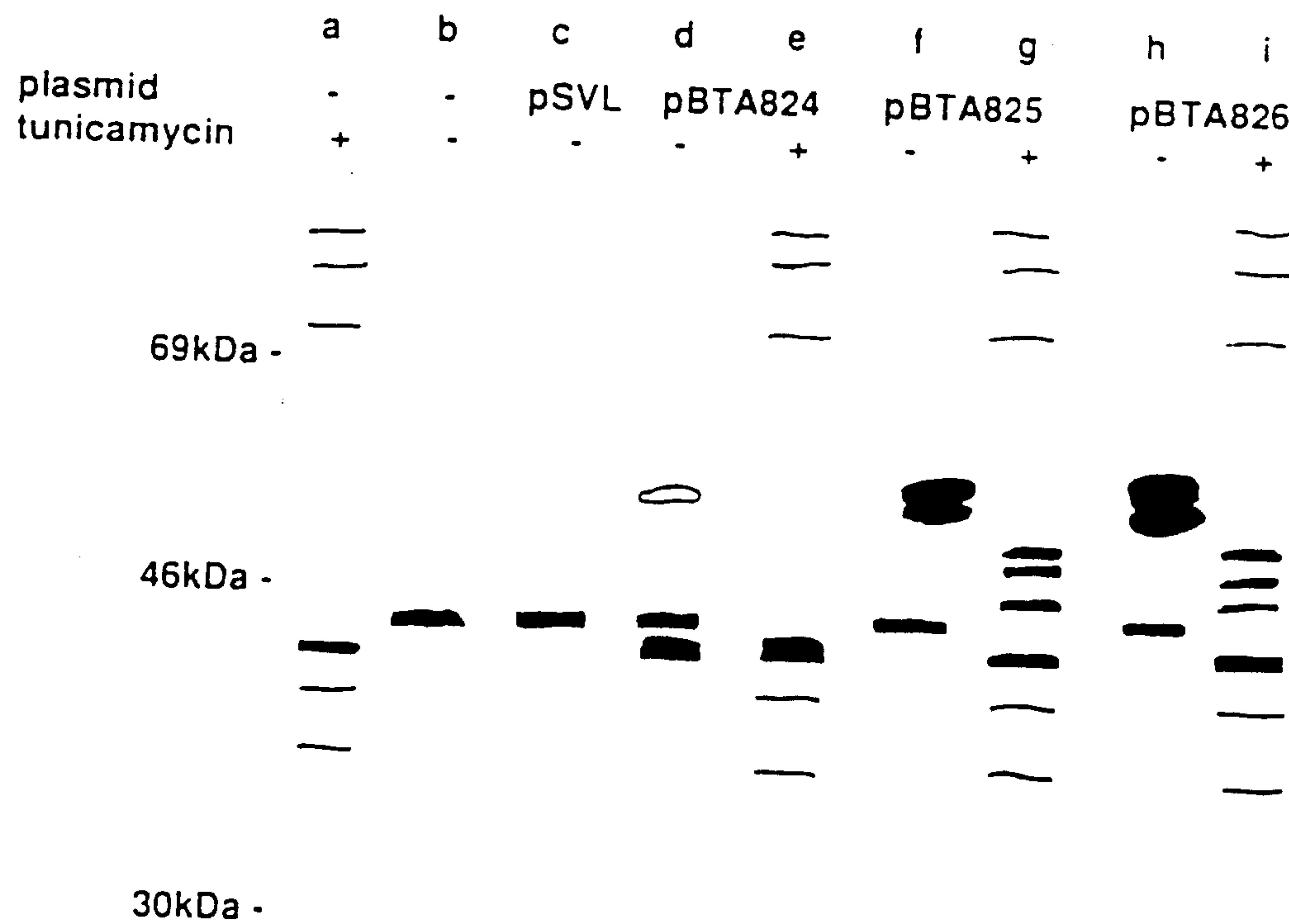
FIGURE 4B



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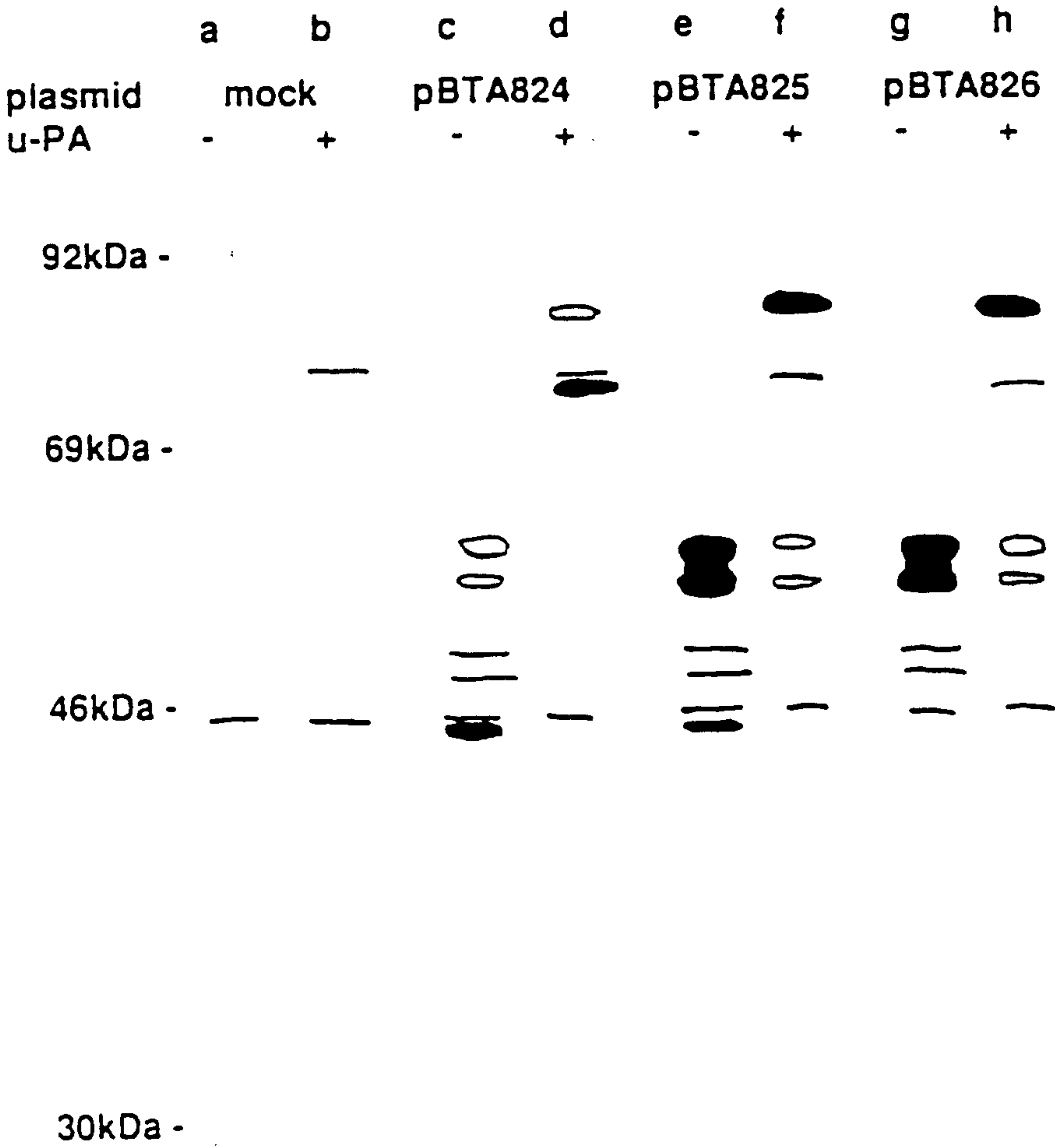
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FIGURE 5 Medium



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FIGURE 6 Urokinase Binding : Medium



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FIGURE 7: Sequence of preA-PAI-2

1										29										83									
AGCTTGTCGACACC										CTG										CTT									
CTC	GCT	CTC	GGT	CTG	CTA	GCT	TTC	GTG	CCG	TTG	GTG	AGG	TGT	CTG	CTG	LEU	LEU	LEU	83										
LEU	ALA	LEU	GLY	LEU	LEU	ALA	PHE	VAL	PRO	LEU	VAL	ARG	ALA	Met	Glu	Asp	Leu	137											
TGT	GTG	GCA	AAC	ACA	CTC	TTT	GCC	CTC	AAT	TTA	TTC	AAG	CAT	CTG	GCA	AAA	GCA	191											
Cys	Val	Ala	Asn	Thr	Leu	Phe	Ala	Leu	Asn	Leu	Phe	Lys	His	Leu	Ala	Lys	Ala	191											
AGC	CCC	ACC	CAG	AAC	CTC	TTC	CTC	TCC	CCA	TGG	AGC	ATC	TCG	TCC	ACC	ATG	GCC	245											
Ser	Pro	Thr	Gln	Asn	Leu	Phe	Leu	Ser	Pro	Trp	Ser	Ile	Ser	Ser	Thr	Met	Ala	245											
ATG	GTC	TAC	ATG	GGC	TCC	AGG	GGC	AGC	ACC	GAA	GAC	CAG	ATG	GGC	AAG	GTG	CTT	299											
Met	Val	Tyr	Met	Gly	Ser	Arg	Gly	Ser	Thr	Glu	Asp	Gln	Met	Ala	Lys	Val	Leu	299											
CAG	TTT	AAT	GAA	GTG	GGA	GCC	AAT	GCA	GTT	ACC	CCC	ATG	ACT	CCA	GAG	AAC	TTT	353											
Gln	Phe	Asn	Glu	Val	Gly	Ala	Asn	Ala	Val	Thr	Pro	Met	Thr	Pro	Glu	Asn	Phe	353											
ACC	AGC	TGT	GGG	TTC	ATG	CAG	CAG	ATC	CAG	AAG	GGT	AGT	TAT	CCT	GAT	GCG	ATT	407											
Thr	Ser	Cys	Gly	Phe	Met	Gln	Gln	Ile	Gln	Lys	Gly	Ser	Tyr	Pro	Asp	Ala	Ile	407											
TTG	CAG	GCA	CAA	GCT	GCA	GAT	AAA	ATC	CAT	TCA	TCC	TTC	CGC	TCT	CTC	AGC	TCT	461											
Leu	Gln	Ala	Gln	Ala	Ala	Asp	Lys	Ile	His	Ser	Ser	Phe	Arg	Ser	Leu	Ser	Ser	461											
GCA	ATC	AAT	GCA	TCC	ACA	GGG	AAT	TAT	TTA	CTG	GAA	AGT	GTC	AAT	AAG	CTG	TTT	515											
Ala	Ile	Asn	Ala	Ser	Thr	Gly	Asn	Tyr	Leu	Leu	Glu	Ser	Val	Asn	Lys	Leu	Phe	515											
GGT	GAG	AAG	TCT	GGC	AGC	TTC	CGG	GAA	GAA	TAT	ATT	CGA	CTC	TGT	CAG	AAA	TAT	569											
Gly	Glu	Lys	Ser	Ala	Ser	Phe	Arg	Glu	Glu	Tyr	Ile	Arg	Leu	Cys	Gln	Lys	Tyr	569											
TAC	TCC	TCA	GAA	CCC	CAG	GCA	GTA	GAC	TTC	CTA	GAA	TGT	GCA	GAA	GAA	GCT	AGA	623											
Tyr	Ser	Ser	Glu	Pro	Gln	Ala	Val	Asp	Phe	Leu	Glu	Cys	Ala	Glu	Glu	Ala	Arg	623											
AAA	AAG	ATT	AAT	TCC	TGG	GTC	AAG	ACT	CAA	ACC	AAA	GGC	AAA	ATC	CCA	AAC	TTG	Leu											
Lys	Lys	Ile	Asn	Ser	Trp	Val	Lys	Thr	Gln	Thr	Lys	Gly	Lys	Ile	Pro	Asn	Leu												

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FIGURE 7 (continued)

TTA Leu	CCT Pro	GAA Glu	GGT Gly	TCT Ser	GTA Val	GAT Asp	GGG Gly	GAT Asp	ACC Thr	AGG Arg	ATG Met	GTC Val	CTG Leu	CTG Val	AAT Asn	GCT Ala	GTC Val	677
TAC Tyr	TTC Phe	AAA Lys	GGA Gly	AAG Lys	TGG Trp	AAA Lys	ACT Thr	CCA Pro	TTT Phe	GAG Glu	AAG Lys	AAA Lys	CTA Leu	AAT Asn	GGG Gly	CTT Leu	TAT Tyr	731
CCT Pro	TTC Phe	CGT Arg	GTA Val	AAC Asn	TCG Ser	GCT Ala	CAG Gln	CGC Arg	ACA Thr	CCT Pro	GTA Val	CAG Gln	ATG Met	ATG Met	TAC Tyr	TTG Leu	CGT Arg	785
GAA Glu	AAG Lys	CTA Leu	AAC Asn	ATT Ile	GGA Gly	TAC Tyr	ATA Ile	GAA Glu	GAC Asp	CTA Leu	AAG Lys	GCT Ala	CAG Gln	ATT Ile	CTA Leu	GAA Glu	CTC Leu	839
CCA Pro	TAT Tyr	GCT Ala	GGA Gly	GAT Asp	GTT Val	AGC Ser	ATG Met	TTC Phe	TTG Leu	TTG Leu	CTT Leu	CCA Pro	GAT Asp	GAA Glu	ATT Ile	CCC Ala	GAT Asp	947
GTG Val	TCC Ser	ACT Thr	GGC Gly	TTG Leu	GAG Glu	CTG Leu	CTG Leu	GAA Glu	AGT Ser	GAA Glu	ATA Ile	ACC Thr	TAT Tyr	GAC Asp	AAA Lys	CTC Leu	AAC Asn	1001
AAG Lys	TGG Trp	ACC Thr	AGC Ser	AAA Lys	GAC Asp	AAA Lys	ATG Met	GCT Ala	GAA Glu	GAT Asp	GAA Glu	GTT Val	GAG Glu	GTA Val	TAC Tyr	ATA Ile	CCC Pro	1055
CAG Gln	TTC Phe	AAA Lys	TTA Leu	GAA Glu	GAG Glu	CAT His	TAT Tyr	GAA Glu	TCC Ser	AGA Arg	ATT Ile	CTG Leu	CTG Leu	AGA Arg	AGC Ser	ATG Met	GGC Gly	1109
ATG Met	GAG Glu	GAC Asp	GCC Ala	TTC Phe	AAC Asn	AAG Lys	GGA Gly	CGG Arg	GCC Ala	AAT Asn	TTC Phe	TCA Ser	GGG Gly	ATG Met	TCG Ser	GAG Glu	AGG Arg	1163
AAT Asn	GAC Asp	CTG Leu	TTT Phe	CTT Leu	TCT Ser	GAA Glu	GTG Val	TTC Phe	CAC His	CAA Gln	GCC Ala	ATG Met	GTG Val	GAT Asp	GTG Val	AAT Asn	GAG Glu	1217
GAG Glu	GGC Gly	ACT Thr	GAA Glu	GCA Ala	GCC Ala	GCT Ala	GGC Gly	ACA Thr	GGG Gly	GGT Gly	GTT Val	ATG Met	ACA Thr	GGG Gly	AGA Arg	ACT Thr	GGA Gly	1271
CAT His	GGA Gly	GGC Gly	CCA Pro	CAG Gln	TTT Phe	GTG Val	GCA Ala	GAT Asp	CAT His	CCT Pro	TTT Phe	CTT Leu	TTT Phe	CTT Leu	ATT Ile	ATG Met	CAT His	1328
AAG Lys	ATA Ile	ACC Thr	AAC Asn	TGC Cys	ATT Ile	TTA Leu	TTT Phe	TTC Phe	GGC Gly	AGA Arg	TTT Phe	TCC Ser	TCA Ser	CCC Pro	TAAAACCTAAGCG			

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FIGURE 8: Sequence of preB-PAI-2

1 AGCTTGTCGACACC										41
CTG	CTG	CTA	GCA	GGC	CTG	TGT	TGT	CTG	CTG	ATC
LEU	LEU	LEU	ALA	GLY	LEU	CYS	CYS	LEU	VAL	ILE
										95
TGT	GTG	GCA	AAC	ACA	CTC	TTT	Phe	CTC	CTG	CTT
Cys	Val	Ala	Asn	Thr	Leu			Leu	ALA	Leu
										149
AGC	CCC	ACC	CAG	AAC	CTC	TTC	CTC	TCC	CTG	GCA
Ser	Pro	Thr	Gln	Asn	Leu	Phe	Leu	Ser	Leu	Ala
										203
ATG	GTC	TAC	ATG	GGC	TCC	AGG	AGG	AGC	ATG	GCC
Met	Val	Tyr	Met	Gly	Ser	Arg	Arg	Met	Thr	Ala
										257
CAG	TTT	AAT	GAA	GTG	GGA	GCC	AGC	AGC	GTG	CTT
Gln	Phe	Asn	Glu	Val	Gly	Ala	Asn	Ala	Val	Leu
										311
ACC	AGC	TGT	GGG	TTC	ATG	CAG	CAG	ATC	AAC	TTT
Thr	Ser	Cys	Gly	Phe	Met	Gln	Gln	Ile	Asn	Phe
										365
TTG	CAG	GCA	CAA	GCT	GCA	GAT	AAA	ATC	AGC	ATT
Leu	Gln	Ala	Gln	Ala	Ala	Asp	Lys	Ile	Ala	Ile
										419
GCA	ATC	AAT	GCA	TCC	ACA	GGG	GGG	TTC	CTG	TCT
Ala	Ile	Asn	Ala	Ser	Thr	Gly	Asn	Arg	Leu	Ser
										473
GGT	GAG	AAG	TCT	GGC	AGC	TTC	Phe	CTC	AAA	TTT
Gly	Glu	Lys	Ser	Ala	Ser	Phe	Arg	Gln	Lys	Phe
										527
TAC	TCC	TCA	GAA	CCC	CAG	GCA	GTA	GAC	GCT	TAT
Tyr	Ser	Ser	Glu	Pro	Gln	Ala	Val	Ala	Ala	AGA
										581
AAA	AAG	ATT	AAT	TCC	TGG	GTC	Val	ATC	AAC	TTG
Lys	Lys	Ile	Asn	Ser	Trp	Val	Lys	Ile	Asn	Leu

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FIGURE 8 (continued)

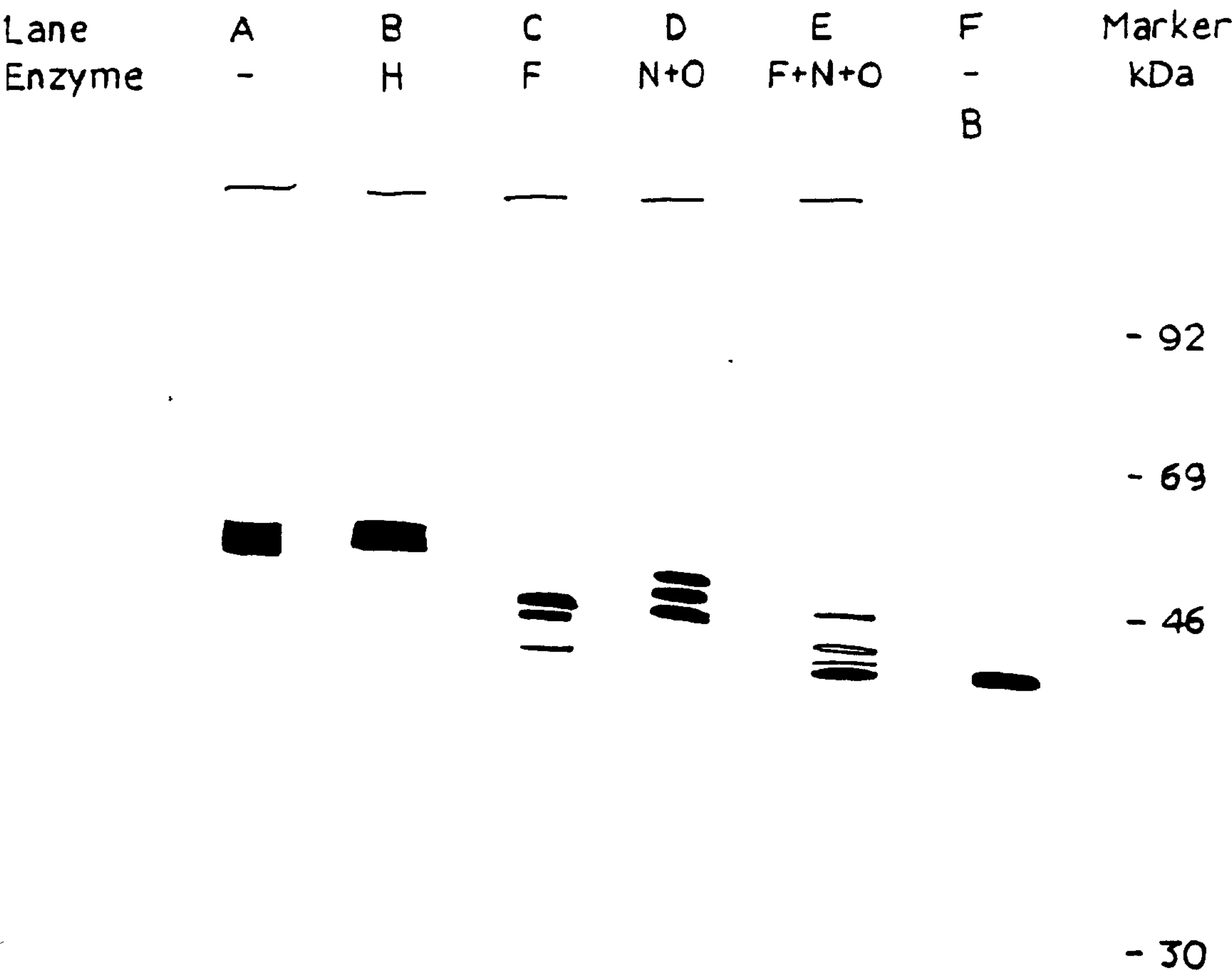
TTA	CCT	GAA	GGT	TCT	GTA	GAT	GGG	GAT	GGG	AGG	ATG	GTC	CTG	CTG	Val	AAT	GCT	GTC	689
Leu	Pro	Glu	Gly	Ser	Val	Asp	Gly	Asp	Gly	Arg	Met	Val	Leu	Leu	Val	Asn	Ala	Val	
TAC	TTC	AAA	GGA	AAG	TGG	AAA	ACT	CCA	ACT	GAG	AAG	AAA	CTA	CTA	TAT	GGG	CTT	TAT	743
Tyr	Phe	Lys	Gly	Lys	Trp	Lys	Thr	Pro	Thr	Glu	Lys	Lys	Leu	Leu	Tyr	Gly	Leu	Tyr	
CCT	TTC	CGT	GTA	AAC	TCG	GCT	CAG	CGC	CAG	CCT	GTA	CAG	ATG	ATG	797	TAC	TTG	CGT	
Pro	Phe	Arg	Val	Asn	Ser	Ala	Gln	Arg	Gln	Pro	Val	Gln	Met	Met	Arg	Tyr	Leu	Arg	
GAA	AAG	CTA	AAC	ATT	GGA	TAC	ATA	GAA	ATA	CTA	AAG	GCT	CAG	ATT	851	CTA	GAA	CTC	
Glu	Lys	Leu	Asn	Ile	Gly	Tyr	Ile	Glu	Ile	Leu	Lys	Ala	Gln	Ile	Leu	Leu	Glu	Leu	
CCA	TAT	GCT	GGA	GAT	GTT	AGC	ATG	TTC	ATG	TTG	CTT	CCA	GAT	CAA	905	ATT	GCC	GAT	
Pro	Tyr	Ala	Gly	Asp	Val	Ser	Met	Phe	Met	Leu	Leu	Pro	Asp	Glu	Asp	Ile	Ala	Asp	
GTC	TCC	ACT	GGC	TTG	GAG	CTG	CTG	GAA	CTG	GAA	ATA	ACC	TAT	GAC	959	AAA	CTC	AAC	
Val	Ser	Thr	Gly	Leu	Glu	Leu	Leu	Glu	Leu	Glu	Ile	Thr	Tyr	Asp	Asn	Lys	Leu	Asn	
AAG	TGG	ACC	AGC	AAA	GAC	AAA	ATG	GCT	ATG	GAT	GAA	GTT	GAG	GTA	1013	TAC	ATA	CCC	
Lys	Trp	Thr	Ser	Lys	Asp	Lys	Met	Ala	Met	Asp	Glu	Val	Glu	Val	Pro	Tyr	Ile	Pro	
CAG	TTC	AAA	TTA	GAA	GAG	CAT	TAT	GAA	TAT	AGA	TCC	ATT	CTG	AGA	1067	AGC	ATG	GCC	
Gln	Phe	Lys	Leu	Glu	Glu	His	Tyr	Glu	Tyr	Arg	Ser	Ile	Leu	Arg	Gly	Ser	Met	Gly	
ATG	GAG	GAC	GCC	TTC	AAC	AAG	GGA	CGG	GGA	AAT	TTC	TCA	GGG	ATG	1121	TCG	GAG	AGG	
Met	Glu	Asp	Ala	Phe	Asn	Lys	Gly	Arg	Gly	Asn	Phe	Ser	Gly	Met	Arg	Ser	Glu	Arg	
AAT	GAC	CTG	TTT	CTT	TCT	GAA	CTG	TTC	CTG	CAA	GCC	ATG	GAT	GAT	1175	GTG	AAT	GAG	
Asn	Asp	Leu	Phe	Leu	Ser	Glu	Val	Phe	Val	Gln	Ala	Met	Val	Asp	Glu	Val	Asn	Glu	
GAG	GGC	ACT	GAA	GCA	GCC	GCT	GGC	ACA	GGC	GGT	GTT	ATG	ACA	GGG	1229	AGA	ACT	GGA	
Glu	Gly	Thr	Glu	Ala	Ala	Ala	Gly	Thr	Gly	Gly	Val	Met	Thr	Gly	Gly	Arg	Thr	Gly	
CAT	GGA	GGC	CCA	CAG	TTT	CTG	GCA	GAT	GCA	CCT	TTT	CTT	TTT	CTT	1283	ATT	ATG	CAT	
His	Gly	Gly	Pro	Gln	Phe	Val	Ala	Asp	Ala	Pro	Phe	Leu	Phe	Leu	His	Ile	Met	His	
AAG	ATA	ACC	AAC	TGC	ATT	TTA	TTT	TTC	TTT	AGA	TTT	TCC	TCA	CCC	1340	ATAA	ACTA	AAGCG	
Lys	Ile	Thr	Asn	Cys	Ile	Leu	Phe	Phe	Phe	Arg	Phe	Ser	Ser	Pro	Pro	Pro	Pro	Pro	

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FIGURE 9 Glycosidase Digestion of 60,000 Mr PAI-2

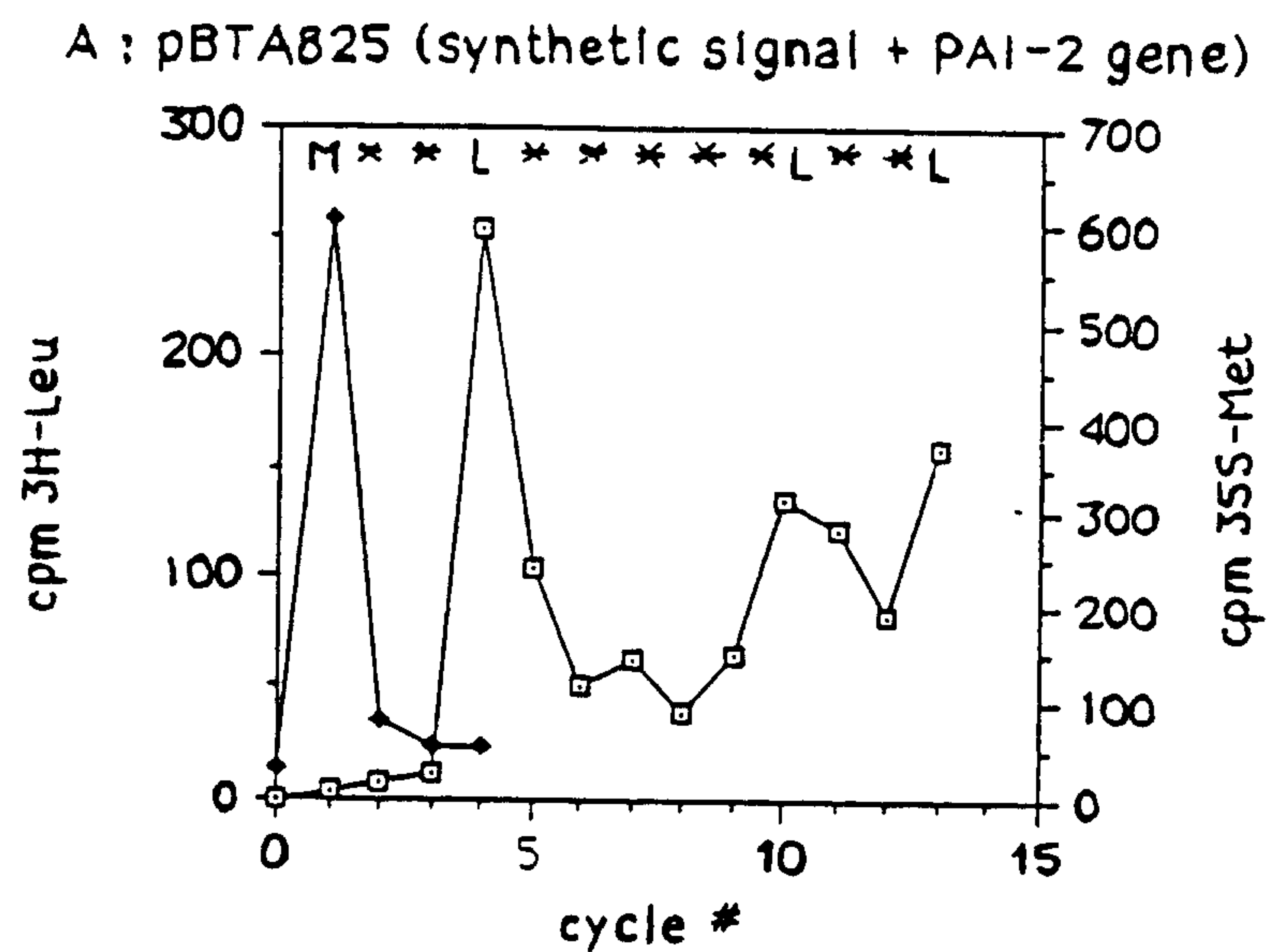


ABBREVIATIONS : H - ENDOGLYCOSIDASE H
F - GLYCOPEPTIDASE F
N - NEURAMINIDASE
O - O-GLYCANASE
B - NON-GLYCOSYLATED PAI-2

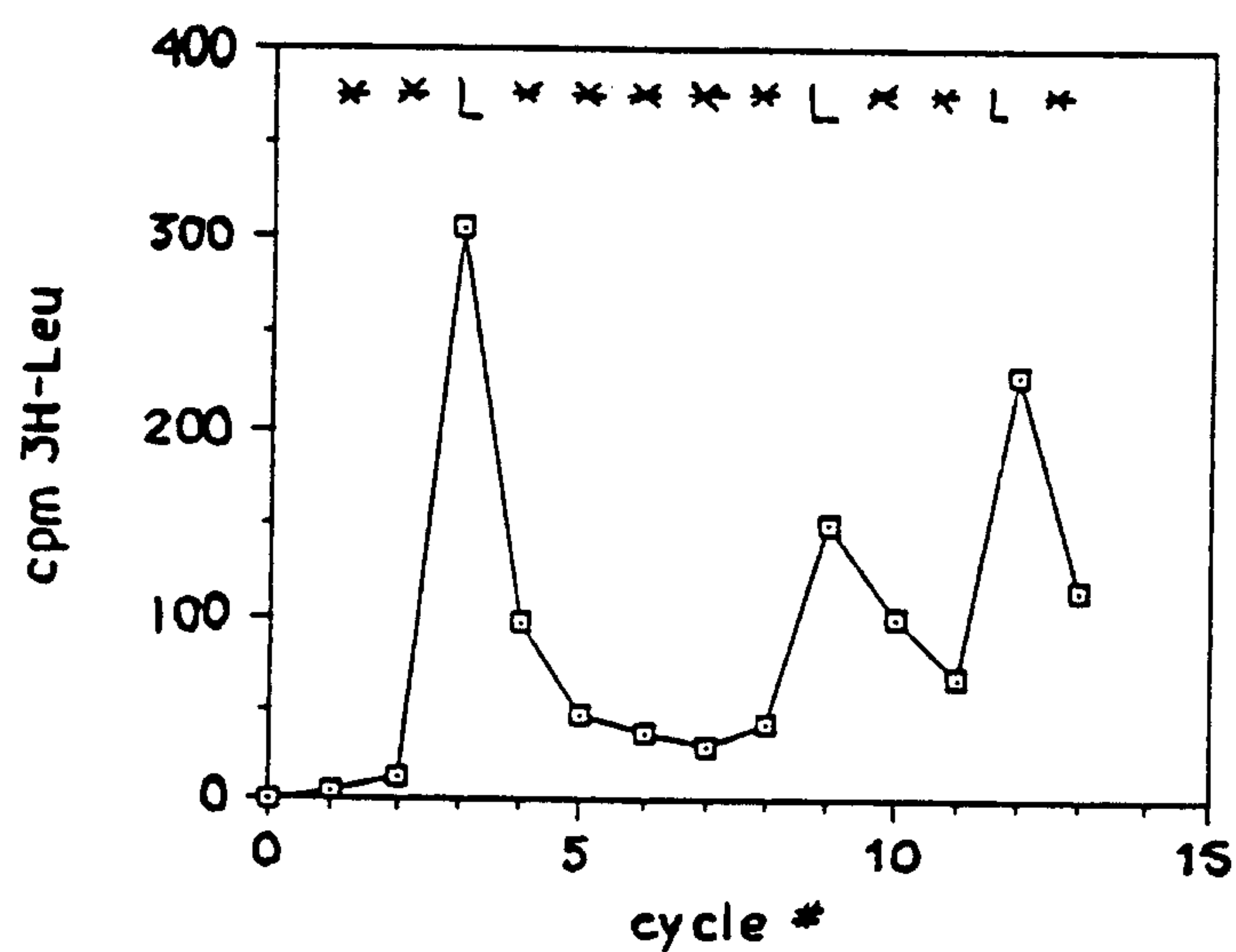
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Figure 10 N-Terminal Analysis of 60,000 Mr PAI-2 Secreted from COS cells transfected with pBTA825 and pBTA826



B : pBTA826 (a-1-antitrypsin signal + PAI-2 gene)

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FIGURE 11 Secretion of PAI-2 into Medium from Transfected CHO-K1 Cells

	transfected plasmid	
Markers (kDa)	pBTA827 vector + PAI-2 gene	pBTA830 vector only
	—	—

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69 -

46 -

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