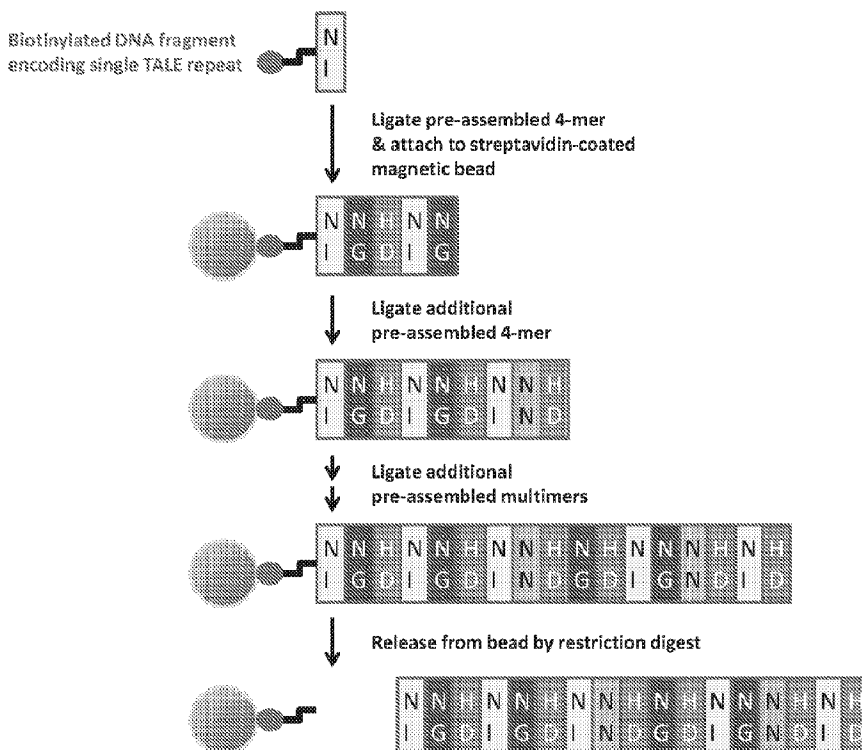




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(57) **Abrégé/Abstract:**

The disclosure describes methods that include providing a first nucleic acid having a sequence encoding a first set comprising one or more transcription activator-like effector (TALE) repeat domains and/or one or more portions of one or more TALE repeat

(57) **Abrégé(suite)/Abstract(continued):**

domains; contacting the first nucleic acid with a first enzyme, wherein the first enzyme creates a first ligatable end; providing a second nucleic acid having a sequence encoding a second set comprising one or more TALE repeat domains and/or one or more portions of one or more TALE repeat domains; contacting the second nucleic acid with a second enzyme, wherein the second enzyme creates a second ligatable end, and wherein the first and second ligatable ends are compatible; and ligating the first and second nucleic acids through the first and second ligatable ends to produce a first ligated nucleic acid, wherein the first ligated nucleic acid is linked to a solid support, and wherein the first ligated nucleic acid encodes a polypeptide comprising said first and second sets.

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[Continued on next page]

(54) Title: METHODS OF TRANSCRIPTION ACTIVATOR LIKE EFFECTOR ASSEMBLY

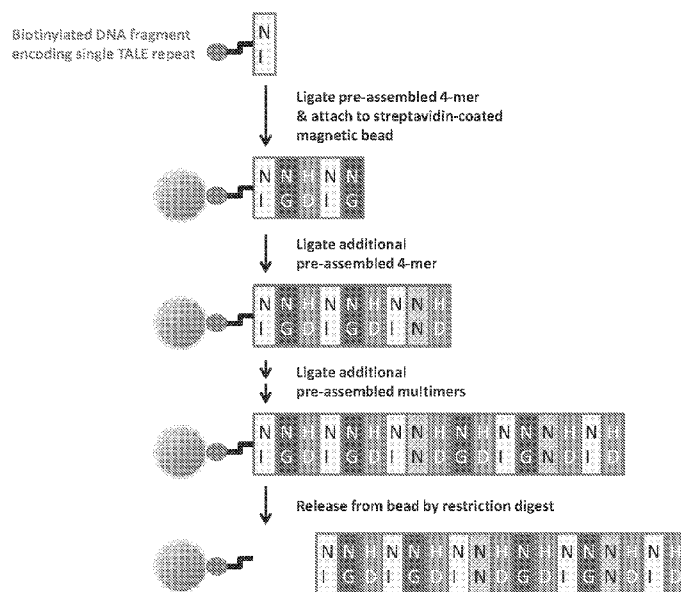


FIG. 1

(57) Abstract: The disclosure describes methods that include providing a first nucleic acid having a sequence encoding a first set comprising one or more transcription activator-like effector (TALE) repeat domains and/or one or more portions of one or more TALE repeat domains; contacting the first nucleic acid with a first enzyme, wherein the first enzyme creates a first ligatable end; providing a second nucleic acid having a sequence encoding a second set comprising one or more TALE repeat domains and/or one or more portions of one or more TALE repeat domains; contacting the second nucleic acid with a second enzyme, wherein the second enzyme creates a second ligatable end, and wherein the first and second ligatable ends are compatible; and ligating the first and second nucleic acids through the first and second ligatable ends to produce a first ligated nucleic acid, wherein the first ligated nucleic acid is linked to a solid support, and wherein the first ligated nucleic acid encodes a polypeptide comprising said first and second sets.

WO 2013/012674 A1



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**METHODS OF TRANSCRIPTION ACTIVATOR LIKE
EFFECTOR ASSEMBLY**

CLAIM OF PRIORITY

STATEMENT AS TO FEDERALLY SPONSORED RESEARCH

5 This invention was made with government support under grant number DP1
OD006862 awarded by the National Institutes of Health. The government has certain
rights in the invention.

TECHNICAL FIELD

10 This invention relates to methods of producing nucleic acids encoding peptides
and polypeptides encoding multiple transcription-like activator effector (TALE) repeat
domains and the proteins themselves.

BACKGROUND

15 TALE proteins of plant pathogenic bacteria in the genus *Xanthomonas* play
important roles in disease, or trigger defense, by binding host DNA and activating
effector-specific host genes (see, e.g., Gu et al., 2005, *Nature* 435:1122; Yang et al., 2006
Proc. Natl. Acad. Sci. USA 103:10503; Kay et al., 2007, *Science* 318:648; Sugio et al.,
2007, *Proc. Natl. Acad. Sci. USA* 104:10720; and Romer et al., 2007, *Science* 318:645).

Specificity for nucleic acid sequences depends on an effector-variable number of imperfect, typically ~33-35 amino acid repeats (Schornack et al., 2006, J. Plant Physiol. 163:256). Each repeat binds to one nucleotide in the target sequence, and the specificity of each repeat for its nucleotide is largely context-independent, allowing for the development of custom sequence-specific TALE proteins (Moscou et al., 2009, Science 326:1501; Boch et al., 2009, Science 326:1509-1512).

SUMMARY

This application is based, at least in part, on the development of rapid, simple, and easily automatable methods for assembling nucleic acids encoding custom TALE repeat array proteins.

Accordingly, this disclosure features a process that includes: (a) providing a first nucleic acid having a sequence encoding a first set comprising one or more (e.g., two or more, three or more, four or more, five or more, six or more, one to six, two to six, three to six, four to six, five or six, one two to five, three to five, four or five, one to four, two to four, three or four, one to three, two or three, one or two, one, two, three, four, five, or six) transcription activator-like effector (TALE) repeat domains and/or one or more portions of one or more TALE repeat domains; (b) contacting the first nucleic acid with a first enzyme, wherein the first enzyme creates a first ligatable end; (c) providing a second nucleic acid having a sequence encoding a second set comprising one or more (e.g., two or more, three or more, four or more, five or more, six or more, one to six, two to six, three to six, four to six, five or six, one two to five, three to five, four or five, one to four, two to four, three or four, one to three, two or three, one or two, one, two, three, four, five, or six) TALE repeat domains and/or one or more portions of one or more TALE repeat domains; (d) contacting the second nucleic acid with a second enzyme, wherein the second enzyme creates a second ligatable end, and wherein the first and second ligatable ends are compatible; and (e) ligating the first and second nucleic acids through the first and second ligatable ends to produce a first ligated nucleic acid, wherein the first ligated nucleic acid is linked to a solid support, and wherein the first ligated nucleic acid encodes a polypeptide comprising said first and second sets.

In some embodiments, the disclosure relates to a process for assembling nucleic acids encoding custom TALE repeat array proteins comprising: (a) providing a first nucleic acid comprising a sequence encoding a first set comprising one or more transcription activator-like effector (TALE) repeat domains; (b) contacting the first nucleic acid with a first enzyme, wherein the first enzyme creates a first ligatable end; (c) providing a second nucleic acid comprising a sequence encoding a second set comprising one or more TALE repeat domains; (d) contacting the second nucleic acid with a second enzyme, wherein the second enzyme creates a second ligatable end, and wherein the first and second ligatable ends are compatible; and (e) ligating the first and second nucleic acids through the first and second ligatable ends to produce a first ligated nucleic acid, wherein the first ligated nucleic acid is linked to a solid support, and wherein the first ligated nucleic acid encodes a polypeptide comprising said first and second sets comprising one or more TALE repeat domains.

In some embodiments, the methods include linking the first nucleic acid to a solid support prior to (b) contacting the first nucleic acid with the first enzyme or prior to (e) ligating the first and second nucleic acids. In some embodiments, the methods include linking the first ligated nucleic acid to a solid support.

In some embodiments, the first set is N-terminal to the second set in the polypeptide. In some embodiments, the second set is N-terminal to the first set in the polypeptide.

In some embodiments, the first and second enzymes are a first and second restriction endonuclease, wherein the first restriction endonuclease cleaves at a site within the first nucleic acid and creates a first cut end, and the second restriction endonuclease cleaves at a site within the second nucleic acid and creates a second cut end, and wherein the first and second ligatable ends are the first and second cut ends. When restriction endonucleases are used, the first ligated nucleic acid cannot include a restriction site recognized by the first restriction endonuclease.

The process can further include: (f) contacting the first ligated nucleic acid with a third enzyme, wherein the third enzyme creates a third ligatable end; (g) providing a third nucleic acid comprising a sequence encoding a third set comprising one or more (e.g., two or more,

three or more, four or more, five or more, six or more, one to six, two to six, three to six, four to six, five or six, one two to five, three to five, four or five, one to four, two to four, three or four, one to three, two or three, one or two, one, two, three, four, five, or six) TALE repeat domains and/or one or more portions of one or more TALE repeat domains; (h) contacting the
5 third nucleic acid with a fourth enzyme, wherein the fourth enzyme creates a fourth ligatable end, and wherein the third and fourth ligatable ends are compatible; and (i) ligating the first ligated and third nucleic acids through the third and fourth ligatable ends to produce a second ligated nucleic acid linked to the solid support, wherein the second ligated nucleic acid encodes a polypeptide comprising said first, second, and third sets.

10 In some embodiments, the third and fourth enzymes are a third and fourth restriction endonuclease, wherein the third restriction endonuclease cleaves at a site within the first ligated nucleic acid and creates a third cut end, and the fourth restriction

endonuclease cleaves at a site within the third nucleic acid and creates a fourth cut end, and wherein the third and fourth ligatable ends are the third and fourth cut ends.

In some embodiments, the ligated nucleic acid does not include a restriction site recognized by the first endonuclease, and the first and third restriction endonucleases are the same. In some embodiments, the second and fourth restriction endonucleases are the same.

The process can further include: (j) contacting the second ligated nucleic acid with a fifth enzyme, wherein the fifth enzyme creates a fifth ligatable end; (k) providing a fourth nucleic acid having a sequence encoding a fourth set comprising one or more (e.g., two or more, three or more, four or more, five or more, six or more, one to six, two to six, three to six, four to six, five or six, one two to five, three to five, four or five, one to four, two to four, three or four, one to three, two or three, one or two, one, two, three, four, five, or six) TALE repeat domains and/or one or more portions of one or more TALE repeat domains; (l) contacting the fourth nucleic acid with a sixth enzyme, wherein the sixth enzyme creates a sixth ligatable end, and wherein the fifth and sixth ligatable ends are compatible; and (m) ligating the second ligated and fourth nucleic acids through the fifth and sixth ligatable ends to produce a third ligated nucleic acid linked to the solid support, wherein the third ligated nucleic acid encodes a polypeptide comprising said first, second, third, and fourth sets. One of ordinary skill would recognize that the process can be repeated with similar additional steps. Such methods are included within this disclosure.

In some embodiments, the fifth and sixth enzymes are a fifth and sixth restriction endonuclease, wherein the fifth restriction endonuclease cleaves at a site within the second ligated nucleic acid and creates a fifth cut end, and the sixth restriction endonuclease cleaves at a site within the fourth nucleic acid and creates a sixth cut end, and wherein the fifth and sixth ligatable ends are the fifth and sixth cut ends.

In some embodiments, the second ligated nucleic acid does not include a restriction site recognized by the first endonuclease, and the first, third, and fifth restriction endonucleases are the same.

In some embodiments, the second, fourth, and sixth restriction endonucleases are the same.

In some embodiments, the solid support and linked nucleic acid are isolated, e.g., following any of the above steps (a) – (m).

5 In some embodiments, the second, third, or fourth set comprises one to four TALE repeat domains.

In some embodiments, the ligatable ends include an overhang of 1-10 nucleotides. In some embodiments, the ligatable ends are blunt ends. In some embodiments, an overhang can be generated using an exonuclease and polymerase in the presence of one or more nucleotides.

In some embodiments, an enzyme or restriction endonuclease used in the above processes is a type IIS restriction endonuclease.

The processes can further comprise unlinking a ligated nucleic acid from the solid support and inserting the ligated nucleic acid (or a processed derivative thereof comprising the TALE repeat array coding sequences) into a vector, e.g., an expression vector. The expression vector can include a sequence encoding an effector domain (e.g., a nuclease domain) configured to create a sequence encoding a fusion protein of the polypeptide and the effector domain. The expression vector can be inserted into a cell to affect the cell directly or for expression of the polypeptide or fusion protein. When the polypeptide or fusion protein is to be expressed, the processes can further include expressing and purifying the polypeptide or fusion protein.

In another aspect, this disclosure features TALE proteins that bind to a target nucleotide sequence (e.g., a “half site”) disclosed herein (e.g., in Table 6 or 7), TALE nucleases that include the TALE proteins, pairs of TALE proteins (e.g., TALENs) that bind to the target sites disclosed herein (e.g., in Table 6 or 7), and nucleic acids that encode any of the above. In some embodiments, the TALE proteins, TALE nucleases, and pairs of TALE proteins (e.g., TALENs) are those disclosed in Example 7. The nucleic acids encoding the TALE proteins, TALE nucleases, and pairs of TALE proteins (e.g., TALENs) can be those disclosed in Example 7 or other sequences that encode the proteins disclosed in Example 7. The disclosure also includes vectors and cells that

include the nucleic acids encoding the TALE proteins, TALE nucleases, or pairs of TALE proteins (e.g., TALENs) disclosed herein and methods of expressing the TALE proteins, TALE nucleases, or pairs of TALE proteins (e.g., TALENs) that include culturing the cells. The methods of expressing the TALE proteins, TALE nucleases, or pairs of TALE proteins (e.g., TALENs) can also include isolating the TALE proteins, TALE nucleases, or pairs of TALE proteins (e.g., TALENs) from the cell culture.

In another aspect, the invention features a set, archive, or library of nucleic acids (e.g., plasmids) that include sequences encoding one or more TALE domains. In some embodiments, the set, archive, or library includes sequences encoding one, two, three, and/or four (or more than four (e.g., five, six, or more)) TALE repeat domains. In some embodiments, the set, library, or archive of nucleic acids includes sequences encoding TALE repeat domains that bind to nucleotide sequences having one, two, three, four (or more than four (e.g., five, six, or more)) nucleotides. In some embodiments, the set, library, or archive includes restriction sites (e.g., sites for type IIS restriction endonucleases) surrounding the sequences encoding the TALE repeat domains.

The methods described herein provide several advantages, including avoiding extensive PCR amplification of the TALE repeats, thereby avoiding the introduction of mutations from PCR errors. Further, TALE repeat arrays of any desired length can be constructed, and the methods can be easily multiplexed and/or automated.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, suitable methods and materials are described below. In case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

The details of one or more embodiments of the invention are set forth in the accompanying drawings and the description below. Other features, objects, and

advantages of the invention will be apparent from the description and drawings, and from the claims.

DESCRIPTION OF DRAWINGS

FIG. 1 is a schematic depiction of an exemplary method of assembling a nucleic acid encoding a TALE protein.

FIG. 2 is a schematic depiction of exemplary archives of nucleic acids encoding single (one-mer), two-mer, three-mer, and four-mer TALE repeat domains.

FIG. 3 depicts the sequence of the pUC57- Δ BsaI plasmid. This plasmid is identical to plasmid pUC57 except for mutation of a single base (in bold, underlined and lowercase) that destroys a BsaI restriction site.

FIG. 4A depicts the polypeptide sequences of exemplary TALE repeats of type α/ϵ , β , γ , and δ . Polymorphic residues characteristic of each type are indicated in bold and italic. The hypervariable triplet SNI for binding to A is indicated in underscore.

FIG. 4B depicts the polynucleotide sequences of the exemplary TALE repeats of FIG. 4A.

FIGs. 5A-5B depict the common sequence of expression plasmids pJDS70, pJDS71, pJDS74, pJDS76, and pJDS78. The region of the variable sequences is depicted as XXXXXXXXXX (underlined and bold).

FIG. 6 is a schematic diagram of the enhanced green fluorescent protein (eGFP) gene and the location of the binding sites for synthetic TALE proteins described herein.

FIG. 7 is a bar graph depicting the % of TALE nuclease-modified, eGFP-negative cells at 2 and 5 days following transfection with plasmids encoding TALE nucleases designed to bind and cleave the eGFP reporter gene.

FIG. 8 is a depiction of the sequences of insertion-deletion mutants of eGFP induced by TALE nucleases. Deleted bases are indicated by dashes and inserted bases indicated by double underlining; the TALEN target half-sites are single underlined. The net number of bases inserted or deleted is shown to the right.

FIG. 9 is a depiction of an electrophoresis gel of assembled DNA fragments encoding 17-mer TALE array preparations.

FIG. 10 is a depiction of an electrophoresis gel of 16-mer TALE array preparations.

FIGs. 11A-11B depict the nucleotide (11A) and polypeptide (11B) sequence of engineered DR-TALE-0003.

5 FIGs. 12A-12B depict the nucleotide (12A) and polypeptide (12B) sequence of engineered DR-TALE-0006.

FIGs. 13A-13B depict the nucleotide (13A) and polypeptide (13B) sequence of engineered DR-TALE-0005.

10 FIGs. 14A-14B depict the nucleotide (14A) and polypeptide (14B) sequence of engineered DR-TALE-0010.

FIGs. 15A-15B depict the nucleotide (15A) and polypeptide (15B) sequence of engineered DR-TALE-0023.

FIGs. 16A-16B depict the nucleotide (16A) and polypeptide (16B) sequence of engineered DR-TALE-0025.

15 FIGs. 17A-17B depict the nucleotide (17A) and polypeptide (17B) sequence of engineered DR-TALE-0020.

FIGs. 18A-18B depict the nucleotide (18A) and polypeptide (18B) sequence of engineered DR-TALE-0022.

20 FIG. 19A is a bar graph depicting activities of 48 TALEN pairs and four ZFN pairs in the EGFP gene-disruption assay. Percentages of EGFP-negative cells as measured 2 and 5 days following transfection of U2OS cells bearing a chromosomally integrated EGFP reporter gene with nuclease-encoding plasmids are shown. Mean percent disruption of EGFP and standard error of the mean from three independent transfections are shown.

25 FIG. 19B is a bar graph depicting mean EGFP-disruption activities from FIG. 19A, grouped by length of the TALENs.

30 FIG. 20A is a graph depicting the ratio of mean percent EGFP disruption values from day 2 to day 5. Ratios were calculated for groups of each length TALEN using the data from FIG. 19B. Values greater than 1 indicate a decrease in the average of EGFP-disrupted cells at day 5 relative to day 2.

FIG. 20B is a graph depicting the ratio of mean tdTomato-positive cells from day 2 to day 5 grouped by various lengths of TALENs. tdTomato-encoding control plasmids were transfected together with nuclease-encoding plasmids on day 0.

FIGs. 21A-E depict DNA sequences and frequencies of assembled TALEN-
5 induced mutations at endogenous human genes. For each endogenous gene target, the wild-type (WT) sequence is shown at the top with the TALEN target half-sites underlined and the translation start codon of the gene (ATG) indicated by a box. Deletions are indicated by dashes and insertions by lowercase letters and double underlining. The sizes of the insertions (+) or deletions (Δ) are indicated to the right of each mutated site. The
10 number of times that each mutant was isolated is shown in parentheses. Mutation frequencies are calculated as the number of mutants identified divided by the total number of sequences analyzed. Note that for several of the genes, we also identified larger deletions that extend beyond the sequences of the TALEN target sites.

FIG. 22 is a schematic depiction of an exemplary method of assembling a nucleic
15 acid encoding a TALE protein containing TALE repeat domains or portions of TALE repeat domains.

DETAILED DESCRIPTION

The methods described herein can be used to assemble engineered proteins containing TALE repeat domains for binding to specific sequences of interest.
20 Assembling long arrays (e.g., 12 or more) of TALE repeat domain repeats can be challenging because the repeats differ only at a small number of amino acids within their highly conserved ~33-35 amino acid consensus sequence. PCR assembly can lead to the introduction of unwanted mutations. Hierarchical assembly methods that involve one or more passages of intermediate plasmid constructs in *E. coli* can also be problematic
25 because the highly repetitive nature of these constructs can make them unstable and prone to recombination and because the need to passage these intermediate constructs makes these approaches difficult to automate.

TAL Effectors

TAL effectors of plant pathogenic bacteria in the genus *Xanthomonas* play important roles in disease, or trigger defense, by binding host DNA and activating effector-specific host genes. Specificity depends on an effector-variable number of imperfect, typically ~33-35 amino acid repeats. Polymorphisms are present primarily at repeat positions 12 and 13, which are referred to herein as the “repeat variable-diresidue” (RVD). The RVDs of TAL effectors correspond to the nucleotides in their target sites in a direct, linear fashion, one RVD to one nucleotide, with some degeneracy and no apparent context dependence. In some embodiments, the polymorphic region that grants nucleotide specificity may be expressed as a triresidue or triplet e.g., encompassing residues 11, 12, and 13.

Each DNA binding repeat can include an RVD that determines recognition of a base pair in the target DNA sequence, wherein each DNA binding repeat is responsible for recognizing one base pair in the target DNA sequence, and wherein the RVD comprises, but is not limited to, one or more of the following: HA for recognizing C; ND for recognizing C; HI for recognizing C; HN for recognizing G; NA for recognizing G; SN for recognizing G or A; YG for recognizing T; and NK for recognizing G, and one or more of: HD for recognizing C; NG for recognizing T; NI for recognizing A; NN for recognizing G or A; NS for recognizing A or C or G or T; N* for recognizing C or T, wherein * represents a gap in the second position of the RVD; HG for recognizing T; H* for recognizing T, wherein * represents a gap in the second position of the RVD; and IG for recognizing T.

TALE proteins are useful in research and biotechnology as targeted chimeric nucleases that can facilitate homologous recombination in genome engineering (e.g., to add or enhance traits useful for biofuels or biorenewables in plants). These proteins also are useful as, for example, transcription factors, and especially for therapeutic applications requiring a very high level of specificity such as therapeutics against pathogens (e.g., viruses) as non-limiting examples.

Assembly Methods

An example of the methods described herein of assembling a TALE repeat domain array is shown in FIG. 1 and includes the following steps: (1) provision a single biotinylated PCR product encoding one single N-terminal TALE repeat domain (a one-mer) with a linker suitable for attachment to a solid support (in the example shown here, a magnetic streptavidin coated bead is used but other solid supports can also be utilized as well as other ways of tethering the initial DNA fragment to the solid support); (2) creation of an overhang at the 3' end of the one-mer DNA (e.g., using a Type IIS restriction enzyme); (3) ligation of a second fragment containing four TALE repeat domain (i.e., a pre-assembled four-mer), creating a five-mer; (4) attachment of the five-mer to the solid support; (5) ligation of additional pre-assembled TALE repeat domains to create a long array, e.g., a piece or pieces of DNA encoding one, two, three, or four TALE repeat domains depending upon the length of the desired final array, and (6) release of the extended DNA encoding the TALE repeats from the solid support (e.g., by using a Type IIS restriction enzyme whose site is built in at the 5' end of the initial biotinylated DNA product). The final fragment can then be prepared for ligation to an appropriate expression plasmid.

Alternatively, the method can proceed as follows: (1) attachment of a single biotinylated PCR product encoding one single N-terminal TALE repeat domains to a solid support (in the example shown here, a magnetic streptavidin coated bead is used but other solid supports such as the streptavidin-coated wells of a multi-well plate can also be utilized as well as other ways of tethering the initial DNA fragment to the solid support), (2) creation of an overhang at the 3' end of the anchored DNA (e.g., using a Type IIS restriction enzyme), (3) ligation of a second fragment containing four TALE repeat domain, (4) additional cycles of steps (2) and (3) to create a long array, (5) in the final cycle performing ligation of a piece of DNA encoding one, two, three, or four TALE repeat domains depending upon the length of the desired final array, and (6) release of the extended DNA encoding the TALE repeats from the solid support (e.g., by using a Type IIS restriction enzyme whose site is built in at the 5' end of the initial biotinylated DNA product).

Another example of a method of assembling a TALE repeat domain array based on the methods described herein is shown in FIG. 22 and includes the following steps: (1) provision a single biotinylated PCR product encoding a portion of one single N-terminal TALE repeat domain (a partial one-mer) with a linker suitable for attachment to a solid support (in the example shown here, a magnetic streptavidin coated bead is used but other solid supports can also be utilized as well as other ways of tethering the initial DNA fragment to the solid support); (2) creation of an overhang at the 3' end of the partial one-mer DNA (e.g., using a Type IIS restriction enzyme); (3) ligation of a second fragment containing consisting of two partial and three full TALE repeats; (4) attachment of the second fragment to the solid support; (5) ligation of additional pre-assembled TALE repeat domains or portions of TALE repeat domains to create a long array, e.g., a piece or pieces of DNA encoding one, two, three, or four TALE repeat domains (or portions of TALE repeat domains) depending upon the length of the desired final array, and (6) release of the extended DNA encoding the TALE repeats from the solid support (e.g., by using a Type IIS restriction enzyme whose site is built in at the 5' end of the initial biotinylated DNA product). The final fragment can then be prepared for ligation to an appropriate expression plasmid.

The initial nucleic acid encoding one or more TALE repeat domains (or portions) is linked to a solid support. The initial nucleic acid can be prepared by any means (e.g., chemical synthesis, PCR, or cleavage from a plasmid). Additionally, the nucleic acid can be linked to the solid support by any means, e.g., covalently or noncovalently.

In some embodiments, the nucleic acid is linked noncovalently by using a nucleic acid modified with one member of a binding pair and incorporating the other member of the binding pair on the solid support. A member of a binding pair is meant to be one of a first and a second moiety, wherein said first and said second moiety have a specific binding affinity for each other. Suitable binding pairs for use in the invention include, but are not limited to, antigens/antibodies (for example, digoxigenin/anti-digoxigenin, dinitrophenyl (DNP)/anti-DNP, dansyl-X/anti-dansyl, Fluorescein/anti-fluorescein, lucifer yellow/anti-lucifer yellow, peptide/anti-peptide, ligand/receptor and rhodamine/anti-rhodamine), biotin/avidin (or biotin/streptavidin) and calmodulin binding

protein (CBP)/calmodulin. Other suitable binding pairs include polypeptides such as the FLAG-peptide (Hopp et al., 1988, *BioTechnology*, 6:1204-10); the KT3 epitope peptide (Martin et al., *Science* 255:192-194 (1992)); tubulin epitope peptide (Skinner et al., *J. Biol. Chem.* 266:15163-66 (1991)); and the T7 gene 10 protein peptide tag (Lutz-Freyer et al., *Proc. Natl. Acad. Sci. USA*, 87:6393-97 (1990)) and the antibodies each thereto.

In some embodiments, the individual nucleic acids encoding one or more TALE repeat domains are present in an archive or library of plasmids (see FIG. 2). Although nucleic acids encoding one to four TALE repeat domains are shown, the library of plasmids can contain nucleic acids encoding more than four (e.g., five, six, or more) TALE repeat domains. Alternatively, as shown Figure 22, the nucleic acids encoding parts or portions of one or more TALE repeat domains can also be joined together to create final DNA fragments encoding the desired full-length arrays of TALE repeat domains. Numerous TALE repeat domain sequences with binding specificity for specific nucleotides or sets of nucleotides are known in the art, and one of ordinary skill can design and prepare a library of plasmids based on these known sequences and the disclosures herein.

As used herein, a solid support refers to any solid or semisolid or insoluble support to which the nucleic acid can be linked. Such materials include any materials that are used as supports for chemical and biological molecule syntheses and analyses, such as, but not limited to: polystyrene, polycarbonate, polypropylene, nylon, glass, dextran, chitin, sand, pumice, agarose, polysaccharides, dendrimers, buckyballs, polyacryl-amide, silicon, rubber, and other materials used as supports for solid phase syntheses, affinity separations and purifications, hybridization reactions, immunoassays and other such applications. The solid support can be particulate or can be in the form of a continuous surface, such as a microtiter dish or well, a glass slide, a silicon chip, a nitrocellulose sheet, nylon mesh, or other such materials. When particulate, typically the particles have at least one dimension in the 5-10 mm range or smaller. Such particles, referred collectively herein as "beads," are often, but not necessarily, spherical. Such reference, however, does not constrain the geometry of the matrix, which can be any

shape, including random shapes, needles, fibers, and elongated. Roughly spherical
“beads,” particularly microspheres that can be used in the liquid phase, also are
contemplated. The “beads” can include additional components, such as magnetic or
paramagnetic particles (see, e.g., Dynabeads (Dynal, Oslo, Norway)) for separation using
5 magnets, as long as the additional components do not interfere with the methods
described herein.

The ligatable ends can be produced by cutting with a restriction endonuclease
(e.g., a type II or type IIS restriction endonuclease) or by “chewing back” the end using
an enzyme (or enzymes) with exonuclease and polymerase activities in the presence of
10 one or more nucleotides (see, Aslanidis et al., 1990, Nucl. Acids Res., 18:6069-74).
Suitable enzymes are known to those of ordinary skill in the art. When restriction
endonucleases are used, the nucleic acids can be designed to include restriction sites for
the enzymes at suitable locations.

Following a ligation reaction, any unligated ends with 5' or 3' overhangs can be
15 “blunted” by use of a polymerase, e.g., a DNA polymerase with both 3'→5' exonuclease
activity and 5'→3' polymerase activity. This blunting step can reduce the appearance of
undesired or partial assembly products. Alternatively, these ends can be capped using
either a “hairpin” oligo bearing a compatible overhang (Briggs et al., 2012, Nucleic Acids
Res, PMID: 22740649) or by short double-stranded DNAs bearing a compatible
20 overhang on one end and a blunt end on the other.

To prepare the ligated nucleic acid for further downstream processing, it can be
useful to select nucleic acids of the expected size, to reduce the presence of minor
products created by incomplete ligations. Methods of selecting nucleic acids by size are
known in the art, and include gel electrophoresis (e.g., slab gel electrophoresis or
25 capillary gel electrophoresis (see, e.g., Caruso et al., 2003, Electrophoresis, 24:1-2:78-
85)), liquid chromatography (e.g., size exclusion chromatography or reverse phase
chromatography (see, e.g., Huber et al., 1995, Anal. Chem., 67:578-585)), and lab-on-a-
chip systems (e.g., LabChip[®] XT system, Caliper Life Sciences, Hopkinton, MA). In
some embodiments, a size exclusion step can be performed using an automated system,

e.g., an automated gel electrophoresis system (e.g., a Pippin Prep™ automated DNA size selection system, Sage Science, Beverly, MA).

Automation

5 The methods disclosed herein can be performed manually or implemented in laboratory automation hardware (e.g., SciClone G3 Liquid Handling Workstation, Caliper Life Sciences, Hopkinton, MA) controlled by a compatible software package (e.g., Maestro™ liquid handling software) programmed according to the new methods described herein or a new software package designed and implemented to carry out the
10 specific method steps described herein. When performed by laboratory automation hardware, the methods can be implemented by computer programs using standard programming techniques following the method steps described herein.

 Examples of automated laboratory system robots include the Sciclone™ G3 liquid handling workstation (Caliper Life Sciences, Hopkinton, MA), Biomek® FX liquid
15 handling system (Beckman-Coulter, Fullerton, California), TekBench™ automated liquid handling platform (TekCel, Hopkinton, Massachusetts), and Freedom EVO® automation platform (Tecan Trading AG, Switzerland).

 The programs can be designed to execute on a programmable computer including at least one processor, at least one data storage system (including volatile and non-
20 volatile memory and/or storage elements, e.g., RAM and ROM), at least one communications port that provides access for devices such as a computer keyboard, telephone, or a wireless, hand-held device, such as a PDA, and optionally at least one output device, such as a monitor, printer, or website. The central computer also includes a clock and a communications port that provides control of the lab automation hardware.
25 These are all implemented using known techniques, software, and devices. The system also includes a database that includes data, e.g., data describing the procedure of one or more method steps described herein.

 Program code is applied to data input by a user (e.g., location of samples to be processed, timing and frequency of manipulations, amounts of liquid dispensed or
30 aspirated, transfer of samples from one location in the system to another) and data in the

database, to perform the functions described herein. The system can also generate inquiries and provide messages to the user. The output information is applied to instruments, e.g., robots, that manipulate, heat, agitate, etc. the vessels that contain the reactants as described herein. In addition, the system can include one or more output
5 devices such as a telephone, printer, or a monitor, or a web page on a computer monitor with access to a website to provide to the user information regarding the synthesis and/or its progress.

Each program embodying the new methods is preferably implemented in a high level procedural or object-oriented programming language to communicate with a
10 computer system. However, the programs can also be implemented in assembly or machine language if desired. In any case, the language can be a compiled or interpreted language.

Each such computer program is preferably stored on a storage medium or device (e.g., RAM, ROM, optical, magnetic) readable by a general or special purpose
15 programmable computer, for configuring and operating the computer when the storage media or device is read by the computer to perform the procedures described herein. The system can also be considered to be implemented as a computer- or machine-readable storage medium (electronic apparatus readable medium), configured with a program, whereby the storage medium so configured causes a computer or machine to operate in a
20 specific and predefined manner to perform the functions described herein.

The new methods can be implemented using various means of data storage. The files can be transferred physically on recordable media or electronically, e.g., by email on a dedicated intranet, or on the Internet. The files can be encrypted using standard
25 encryption software from such companies as RSA Security (Bedford, Massachusetts) and Baltimore[®]. The files can be stored in various formats, e.g., spreadsheets or databases.

As used herein, the term "electronic apparatus" is intended to include any suitable computing or processing apparatus or other device configured or adapted for storing data or information. Examples of electronic apparatus suitable for use with the present
30 invention include stand-alone computing apparatus; communications networks, including local area networks (LAN), wide area networks (WAN), Internet, Intranet, and Extranet;

electronic appliances such as a personal digital assistants (PDAs), cellular telephones, “smartphones,” pagers and the like; and local and distributed processing systems.

As used herein, “stored” refers to a process for encoding information on an electronic apparatus readable medium. Those skilled in the art can readily adopt any of the presently known methods for recording information on known media to generate manufactures comprising the sequence information.

A variety of software programs and formats can be used to store method data on an electronic apparatus readable medium. For example, the data and machine instructions can be incorporated in the system of the software provided with the automated system, represented in a word processing text file, formatted in commercially-available software such as WordPerfect[®] and Microsoft[®] Word[®], or represented in the form of an ASCII file, stored in a database application, such as Microsoft Access[®], Microsoft SQL Server[®], Sybase[®], Oracle[®], or the like, as well as in other forms. Any number of data processor structuring formats (e.g., text file or database) can be employed to obtain or create a medium having recorded thereon the relevant data and machine instructions to implement the methods described herein.

By providing information in electronic apparatus readable form, the programmable computer can communicate with and control the lab automation hardware to perform the methods described herein. One skilled in the art can input data in electronic apparatus readable form (or a form that is converted to electronic apparatus readable form) to describe the completion of various method steps by the lab automation hardware.

Polypeptide Expression Systems

In order to use the engineered proteins of the present invention, it is typically necessary to express the engineered proteins from a nucleic acid that encodes them. This can be performed in a variety of ways. For example, the nucleic acid encoding the engineered TALE repeat protein is typically cloned into an intermediate vector for transformation into prokaryotic or eukaryotic cells for replication and/or expression. Intermediate vectors are typically prokaryote vectors, e.g., plasmids, or shuttle vectors, or

insect vectors, for storage or manipulation of the nucleic acid encoding the engineered TALE protein or production of protein. The nucleic acid encoding the engineered TALE repeat protein is also typically cloned into an expression vector, for administration to a plant cell, animal cell, preferably a mammalian cell or a human cell, fungal cell, bacterial cell, or protozoan cell.

To obtain expression of a cloned gene or nucleic acid, the engineered TALE repeat protein is typically subcloned into an expression vector that contains a promoter to direct transcription. Suitable bacterial and eukaryotic promoters are well known in the art and described, e.g., in Sambrook et al., *Molecular Cloning, A Laboratory Manual* (3d ed. 2001); Kriegler, *Gene Transfer and Expression: A Laboratory Manual* (1990); and *Current Protocols in Molecular Biology* (Ausubel et al., eds., 2010). Bacterial expression systems for expressing the engineered TALE repeat protein are available in, e.g., *E. coli*, *Bacillus* sp., and *Salmonella* (Palva et al., 1983, *Gene* 22:229-235). Kits for such expression systems are commercially available. Eukaryotic expression systems for mammalian cells, yeast, and insect cells are well known in the art and are also commercially available.

The promoter used to direct expression of the engineered TALE repeat protein nucleic acid depends on the particular application. For example, a strong constitutive promoter is typically used for expression and purification of the engineered TALE repeat protein. In contrast, when the engineered TALE repeat protein is to be administered in vivo for gene regulation, either a constitutive or an inducible promoter can be used, depending on the particular use of the engineered TALE repeat protein. In addition, a preferred promoter for administration of the engineered TALE repeat protein can be a weak promoter, such as HSV TK or a promoter having similar activity. The promoter typically can also include elements that are responsive to transactivation, e.g., hypoxia response elements, Gal4 response elements, lac repressor response element, and small molecule control systems such as tet-regulated systems and the RU-486 system (see, e.g., Gossen & Bujard, 1992, *Proc. Natl. Acad. Sci. USA*, 89:5547; Oligino et al., 1998, *Gene Ther.*, 5:491-496; Wang et al., 1997, *Gene Ther.*, 4:432-441; Neering et al., 1996, *Blood*, 88:1147-55; and Rendahl et al., 1998, *Nat. Biotechnol.*, 16:757-761).

In addition to the promoter, the expression vector typically contains a transcription unit or expression cassette that contains all the additional elements required for the expression of the nucleic acid in host cells, either prokaryotic or eukaryotic. A typical expression cassette thus contains a promoter operably linked, e.g., to the nucleic acid sequence encoding the TALE repeat protein signals required, e.g., for efficient polyadenylation of the transcript, transcriptional termination, ribosome binding sites, or translation termination. Additional elements of the cassette can include, e.g., enhancers, and heterologous spliced intronic signals.

The particular expression vector used to transport the genetic information into the cell is selected with regard to the intended use of the engineered TALE repeat protein, e.g., expression in plants, animals, bacteria, fungus, protozoa, etc. Standard bacterial expression vectors include plasmids such as pBR322 based plasmids, pSKF, pET23D, and commercially available fusion expression systems such as GST and LacZ. A preferred fusion protein is the maltose binding protein, "MBP." Such fusion proteins can be used for purification of the engineered TALE repeat protein. Epitope tags can also be added to recombinant proteins to provide convenient methods of isolation, for monitoring expression, and for monitoring cellular and subcellular localization, e.g., c-myc or FLAG.

Expression vectors containing regulatory elements from eukaryotic viruses are often used in eukaryotic expression vectors, e.g., SV40 vectors, papilloma virus vectors, and vectors derived from Epstein-Barr virus. Other exemplary eukaryotic vectors include pMSG, pAV009/A+, pMTO10/A+, pMAMneo-5, baculovirus pDSVE, and any other vector allowing expression of proteins under the direction of the SV40 early promoter, SV40 late promoter, metallothionein promoter, murine mammary tumor virus promoter, Rous sarcoma virus promoter, polyhedrin promoter, or other promoters shown effective for expression in eukaryotic cells.

Some expression systems have markers for selection of stably transfected cell lines such as thymidine kinase, hygromycin B phosphotransferase, and dihydrofolate reductase. High yield expression systems are also suitable, such as using a baculovirus vector in insect cells, with the engineered TALE repeat protein encoding sequence under the direction of the polyhedrin promoter or other strong baculovirus promoters.

The elements that are typically included in expression vectors also include a replicon that functions in *E. coli*, a gene encoding antibiotic resistance to permit selection of bacteria that harbor recombinant plasmids, and unique restriction sites in nonessential regions of the plasmid to allow insertion of recombinant sequences.

5 Standard transfection methods are used to produce bacterial, mammalian, yeast or insect cell lines that express large quantities of protein, which are then purified using standard techniques (see, e.g., Colley et al., 1989, *J. Biol. Chem.*, 264:17619-22; *Guide to Protein Purification*, in *Methods in Enzymology*, vol. 182 (Deutscher, ed., 1990)). Transformation of eukaryotic and prokaryotic cells are performed according to standard
10 techniques (see, e.g., Morrison, 1977, *J. Bacteriol.* 132:349-351; Clark-Curtiss & Curtiss, *Methods in Enzymology* 101:347-362 (Wu et al., eds, 1983)).

Any of the well-known procedures for introducing foreign nucleotide sequences into host cells can be used. These include the use of calcium phosphate transfection, polybrene, protoplast fusion, electroporation, liposomes, microinjection, naked DNA,
15 plasmid vectors, viral vectors, both episomal and integrative, and any of the other well-known methods for introducing cloned genomic DNA, cDNA, synthetic DNA or other foreign genetic material into a host cell (see, e.g., Sambrook et al., *supra*). It is only necessary that the particular genetic engineering procedure used be capable of successfully introducing at least one gene into the host cell capable of expressing the
20 protein of choice.

Characterization of TALE Proteins

Engineered TALE repeat array proteins designed using methods of the present invention can be further characterized to ensure that they have the desired characteristics
25 for their chosen use. For example, TALE repeat array protein can be assayed using a bacterial two-hybrid, bacterial promoter repression, phage-display, or ribosome display system or using an electrophoretic mobility shift assay or "EMSA" (Buratowski & Chodosh, in *Current Protocols in Molecular Biology* pp. 12.2.1-12.2.7). Equally, any other DNA binding assay known in the art could be used to verify the DNA binding
30 properties of the selected protein.

In one embodiment, a bacterial "two-hybrid" system is used to express and test a TALE repeat protein of the present invention. The bacterial two-hybrid system has an additional advantage, in that the protein expression and the DNA binding "assay" occur within the same cells, thus there is no separate DNA binding assay to set up.

5 Methods for the use of the bacterial two-hybrid system to express and assay DNA binding proteins are described in Joung et al., 2000, Proc. Natl. Acad. Sci. USA, 97:7382, Wright et al., 2006, Nat. Protoc, 1:1637-52; Maeder et al., 2008, Mol. Cell, 31:294-301; Maeder et al., 2009, Nat. Protoc., 4:1471-1501; and US Patent Application No. 2002/0119498. Briefly, in a
10 bacterial two-hybrid system, the DNA binding protein is expressed in a bacterial strain bearing the sequence of interest upstream of a weak promoter controlling expression of a reporter gene (e.g., histidine 3 (HIS3), the beta-lactamase antibiotic resistance gene, or the beta-galactosidase (lacZ) gene). Expression of the reporter gene occurs in cells in which the DNA binding protein expressed by the cell binds to the target site sequence.
15 Thus, bacterial cells expressing DNA binding proteins that bind to their target site are identified by detection of an activity related to the reporter gene (e.g., growth on selective media, expression of beta-galactosidase).

 In some embodiments, calculations of binding affinity and specificity are also made. This can be done by a variety of methods. The affinity with which the selected
20 TALE repeat array protein binds to the sequence of interest can be measured and quantified in terms of its K_D . Any assay system can be used, as long as it gives an accurate measurement of the actual K_D of the TALE repeat array protein. In one embodiment, the K_D for the binding of a TALE repeat array protein to its target is measured using an EMSA

25 In one embodiment, EMSA is used to determine the K_D for binding of the selected TALE repeat array protein both to the sequence of interest (i.e., the specific K_D) and to non-specific DNA (i.e., the non-specific K_D). Any suitable non-specific or "competitor" double stranded DNA known in the art can be used. In some embodiments, calf thymus DNA or human placental DNA is used. The ratio of the non-specific K_D to the specific
30 KD is the specificity ratio. TALE repeat array proteins that bind with high specificity

have a high specificity ratio. This measurement is very useful in deciding which of a group of selected TALE should be used for a given purpose. For example, use of TALE repeat array protein in vivo requires not only high affinity binding but also high-specificity binding.

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Construction of Chimeric TALE Proteins

Often, the aim of producing a custom-designed TALE repeat array DNA binding domain is to obtain a TALE repeat array protein that can be used to perform a function. The TALE repeat array DNA binding domain can be used alone, for example to bind to a specific site on a gene and thus block binding of other DNA-binding domains. However, in some embodiments, the TALE repeat array protein will be used in the construction of a chimeric TALE protein containing a TALE repeat array DNA binding domain and an additional domain having some desired specific function (e.g., gene activation) or enzymatic activity i.e., a "functional domain."

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Chimeric TALE repeat array proteins designed and produced using the methods described herein can be used to perform any function where it is desired to target, for example, some specific enzymatic activity to a specific DNA sequence, as well as any of the functions already described for other types of synthetic or engineered DNA binding molecules. Engineered TALE repeat array DNA binding domains, can be used in the construction of chimeric proteins useful for the treatment of disease (see, for example, U.S. patent application 2002/0160940, and U.S. Pat. Nos. 6,511,808, 6,013,453 and 6,007,988, and International patent application WO 02/057308), or for otherwise altering the structure or function of a given gene in vivo. The engineered TALE repeat array proteins of the present invention are also useful as research tools, for example, in performing either in vivo or in vitro functional genomics studies (see, for example, U.S. Pat. No. 6,503,717 and U.S. patent application 2002/0164575).

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To generate a functional recombinant protein, the engineered TALE repeat array DNA binding domain will typically be fused to at least one "functional" domain. Fusing functional domains to synthetic TALE repeat array proteins to form functional transcription factors involves only routine molecular biology techniques which are

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commonly practiced by those of skill in the art, see for example, U.S. Pat. Nos. 6,511,808, 6,013,453, 6,007,988, 6,503,717 and U.S. patent application 2002/0160940).

Functional domains can be associated with the engineered TALE repeat array domain at any suitable position, including the C- or N-terminus of the TALE protein.

5 Suitable "functional" domains for addition to the engineered protein made using the methods of the invention are described in U.S. Pat. Nos. 6,511,808, 6,013,453, 6,007,988, and 6,503,717 and U.S. patent application 2002/0160940.

In one embodiment, the functional domain is a nuclear localization domain which provides for the protein to be translocated to the nucleus. Several nuclear localization
10 sequences (NLS) are known, and any suitable NLS can be used. For example, many NLSs have a plurality of basic amino acids, referred to as a bipartite basic repeats (reviewed in Garcia-Bustos et al, 1991, Biochim. Biophys. Acta, 1071:83-101). An NLS containing bipartite basic repeats can be placed in any portion of chimeric protein and results in the chimeric protein being localized inside the nucleus. It is preferred that a
15 nuclear localization domain is routinely incorporated into the final chimeric protein, as the ultimate functions of the chimeric proteins of the present invention will typically require the proteins to be localized in the nucleus. However, it may not be necessary to add a separate nuclear localization domain in cases where the engineered TALE repeat array domain itself, or another functional domain within the final chimeric protein, has
20 intrinsic nuclear translocation function.

In another embodiment, the functional domain is a transcriptional activation domain such that the chimeric protein can be used to activate transcription of the gene of interest. Any transcriptional activation domain known in the art can be used, such as for
25 example, the VP16 domain from herpes simplex virus (Sadowski et al., 1988, Nature, 335:563-564) or the p65 domain from the cellular transcription factor NF-kappaB (Ruben et al., 1991, Science, 251:1490-93).

In yet another embodiment, the functional domain is a transcriptional repression domain such that the chimeric protein can be used to repress transcription of the gene of interest. Any transcriptional repression domain known in the art can be used, such as for

example, the KRAB (Kruppel-associated box) domain found in many naturally occurring KRAB proteins (Thiesen et al., 1991, *Nucleic Acids Res.*, 19:3996).

In a further embodiment, the functional domain is a DNA modification domain such as a methyltransferase (or methylase) domain, a de-methylation domain, a
5 deaminase domain, a hydroxylase domain, an acetylation domain, or a deacetylation domain. Many such domains are known in the art and any such domain can be used, depending on the desired function of the resultant chimeric protein. For example, it has been shown that a DNA methylation domain can be fused to a TALE repeat array DNA
10 binding protein and used for targeted methylation of a specific DNA sequence (Xu et al., 1997, *Nat. Genet.*, 17:376-378). The state of methylation of a gene affects its expression and regulation, and furthermore, there are several diseases associated with defects in DNA methylation.

In a still further embodiment the functional domain is a chromatin modification domain such as a histone acetylase or histone de-acetylase (or HDAC) domain. Many
15 such domains are known in the art and any such domain can be used, depending on the desired function of the resultant chimeric protein. Histone deacetylases (such as HDAC1 and HDAC2) are involved in gene repression. Therefore, by targeting HDAC activity to a specific gene of interest using an engineered TALE protein, the expression of the gene of interest can be repressed.

In an alternative embodiment, the functional domain is a nuclease domain, such as
20 a restriction endonuclease (or restriction enzyme) domain. The DNA cleavage activity of a nuclease enzyme can be targeted to a specific target sequence by fusing it to an appropriate engineered TALE repeat array DNA binding domain. In this way, sequence specific chimeric restriction enzyme can be produced. Several nuclease domains are
25 known in the art and any suitable nuclease domain can be used. For example, an endonuclease domain of a type IIS restriction endonuclease (e.g., FokI) can be used, as taught by Kim et al., 1996, *Proc. Natl. Acad. Sci. USA*, 6:1156-60). In some embodiments, the endonuclease is an engineered FokI variant as described in US 2008/0131962. Such chimeric endonucleases can be used in any situation where
30 cleavage of a specific DNA sequence is desired, such as in laboratory procedures for the

construction of recombinant DNA molecules, or in producing double-stranded DNA breaks in genomic DNA in order to promote homologous recombination (Kim et al., 1996, Proc. Natl. Acad. Sci. USA, 6:1156-60; Bibikova et al., 2001, Mol. Cell. Biol., 21:289-297; Porteus & Baltimore, 2003, Science, 300:763; Miller et al., 2011, Nat. Biotechnol., 29:143-148; Cermak et al., 2011, Nucl. Acids Res., 39:e82). Repair of
5 TALE nuclease-induced double-strand breaks (DSB) by error-prone non-homologous end-joining leads to efficient introduction of insertion or deletion mutations at the site of the DSB (Miller et al., 2011, Nat. Biotechnol., 29:143-148; Cermak et al., 2011, Nucl. Acids Res., 39:e82). Alternatively, repair of a DSB by homology-directed repair with an
10 exogenously introduced "donor template" can lead to highly efficient introduction of precise base alterations or insertions at the break site (Bibikova et al., 2003, Science, 300:764; Urnov et al., 2005, Nature, 435:646-651; Porteus et al., 2003, Science, 300:763; Miller et al., 2011, Nat. Biotechnol., 29:143-148).

In some embodiments, the functional domain is an integrase domain, such that the
15 chimeric protein can be used to insert exogenous DNA at a specific location in, for example, the human genome.

Other suitable functional domains include silencer domains, nuclear hormone receptors, resolvase domains oncogene transcription factors (e.g., myc, jun, fos, myb, max, mad, rel, ets, bcl, myb, mos family members etc.), kinases, phosphatases, and any
20 other proteins that modify the structure of DNA and/or the expression of genes. Suitable kinase domains, from kinases involved in transcription regulation are reviewed in Davis, 1995, Mol. Reprod. Dev., 42:459-67. Suitable phosphatase domains are reviewed in, for example, Schonthal & Semin, 1995, Cancer Biol. 6:239-48.

Fusions of TALE repeat arrays to functional domains can be performed by
25 standard recombinant DNA techniques well known to those skilled in the art, and as are described in, for example, basic laboratory texts such as Sambrook et al., Molecular Cloning; A Laboratory Manual 3d ed. (2001), and in U.S. Pat. Nos. 6,511,808, 6,013,453, 6,007,988, and 6,503,717 and U.S. patent application 2002/0160940.

In some embodiments, two or more engineered TALE repeat array proteins are
30 linked together to produce the final DNA binding domain. The linkage of two or more

engineered proteins can be performed by covalent or non-covalent means. In the case of covalent linkage, engineered proteins can be covalently linked together using an amino acid linker (see, for example, U.S. patent application 2002/0160940, and International applications WO 02/099084 and WO 01/53480). This linker can be any string of amino acids desired. In one embodiment the linker is a canonical TGEKP linker. Whatever linkers are used, standard recombinant DNA techniques (such as described in, for example, Sambrook et al., *Molecular Cloning: A Laboratory Manual* 3d ed. (2001)) can be used to produce such linked proteins.

In embodiments where the engineered proteins are used in the generation of chimeric endonuclease, the chimeric protein can possess a dimerization domain as such endonucleases are believed to function as dimers. Any suitable dimerization domain can be used. In one embodiment the endonuclease domain itself possesses dimerization activity. For example, the nuclease domain of FokI which has intrinsic dimerization activity can be used (Kim et al., 1996, *Proc. Natl. Acad. Sci.*, 93:1156-60).

Assays for Determining Regulation of Gene Expression by Engineered Proteins

A variety of assays can be used to determine the level of gene expression regulation by the engineered TALE repeat proteins, see for example U.S. Pat. No. 6,453,242. The activity of a particular engineered TALE repeat protein can be assessed using a variety of in vitro and in vivo assays, by measuring, e.g., protein or mRNA levels, product levels, enzyme activity, tumor growth; transcriptional activation or repression of a reporter gene; second messenger levels (e.g., cGMP, cAMP, IP3, DAG, Ca²⁺); cytokine and hormone production levels; and neovascularization, using, e.g., immunoassays (e.g., ELISA and immunohistochemical assays with antibodies), hybridization assays (e.g., RNase protection, northern, in situ hybridization, oligonucleotide array studies), colorimetric assays, amplification assays, enzyme activity assays, tumor growth assays, phenotypic assays, and the like.

TALE proteins can be first tested for activity in vitro using cultured cells, e.g., 293 cells, CHO cells, VERO cells, BHK cells, HeLa cells, COS cells, and the like. In some embodiments, human cells are used. The engineered TALE repeat array protein is

often first tested using a transient expression system with a reporter gene, and then regulation of the target endogenous gene is tested in cells and in animals, both in vivo and ex vivo. The engineered TALE repeat array protein can be recombinantly expressed in a cell, recombinantly expressed in cells transplanted into an animal, or recombinantly expressed in a transgenic animal, as well as administered as a protein to an animal or cell using delivery vehicles described below. The cells can be immobilized, be in solution, be injected into an animal, or be naturally occurring in a transgenic or non-transgenic animal.

Modulation of gene expression is tested using one of the in vitro or in vivo assays described herein. Samples or assays are treated with the engineered TALE repeat array protein and compared to un-treated control samples, to examine the extent of modulation. For regulation of endogenous gene expression, the TALE repeat array protein ideally has a K_D of 200 nM or less, more preferably 100 nM or less, more preferably 50 nM, most preferably 25 nM or less. The effects of the engineered TALE repeat array protein can be measured by examining any of the parameters described above. Any suitable gene expression, phenotypic, or physiological change can be used to assess the influence of the engineered TALE repeat array protein. When the functional consequences are determined using intact cells or animals, one can also measure a variety of effects such as tumor growth, neovascularization, hormone release, transcriptional changes to both known and uncharacterized genetic markers (e.g., northern blots or oligonucleotide array studies), changes in cell metabolism such as cell growth or pH changes, and changes in intracellular second messengers such as cGMP.

Preferred assays for regulation of endogenous gene expression can be performed in vitro. In one in vitro assay format, the engineered TALE repeat array protein regulation of endogenous gene expression in cultured cells is measured by examining protein production using an ELISA assay. The test sample is compared to control cells treated with an empty vector or an unrelated TALE repeat array protein that is targeted to another gene.

In another embodiment, regulation of endogenous gene expression is determined in vitro by measuring the level of target gene mRNA expression. The level of gene

expression is measured using amplification, e.g., using RT-PCR, LCR, or hybridization assays, e.g., northern hybridization, RNase protection, dot blotting. RNase protection is used in one embodiment. The level of protein or mRNA is detected using directly or indirectly labeled detection agents, e.g., fluorescently or radioactively labeled nucleic acids, radioactively or enzymatically labeled antibodies, and the like, as described herein.

Alternatively, a reporter gene system can be devised using the target gene promoter operably linked to a reporter gene such as luciferase, green fluorescent protein, CAT, or beta-galactosidase. The reporter construct is typically co-transfected into a cultured cell. After treatment with the TALE repeat array protein, the amount of reporter gene transcription, translation, or activity is measured according to standard techniques known to those of skill in the art.

Another example of an assay format useful for monitoring regulation of endogenous gene expression is performed in vivo. This assay is particularly useful for examining TALE repeat array proteins that inhibit expression of tumor promoting genes, genes involved in tumor support, such as neovascularization (e.g., VEGF), or that activate tumor suppressor genes such as p53. In this assay, cultured tumor cells expressing the engineered TALE protein are injected subcutaneously into an immune compromised mouse such as an athymic mouse, an irradiated mouse, or a SCID mouse. After a suitable length of time, preferably 4-8 weeks, tumor growth is measured, e.g., by volume or by its two largest dimensions, and compared to the control. Tumors that have statistically significant reduction (using, e.g., Student's T test) are said to have inhibited growth. Alternatively, the extent of tumor neovascularization can also be measured. Immunoassays using endothelial cell specific antibodies are used to stain for vascularization of the tumor and the number of vessels in the tumor. Tumors that have a statistically significant reduction in the number of vessels (using, e.g., Student's T test) are said to have inhibited neovascularization.

Transgenic and non-transgenic animals can also be used for examining regulation of endogenous gene expression in vivo. Transgenic animals can express the engineered TALE repeat array protein. Alternatively, animals that transiently express the engineered TALE repeat array protein, or to which the engineered TALE repeat array protein has

been administered in a delivery vehicle, can be used. Regulation of endogenous gene expression is tested using any one of the assays described herein.

Use of Engineered TALE Repeat-Containing Proteins in Gene Therapy

5 The engineered proteins of the present invention can be used to regulate gene expression or alter gene sequence in gene therapy applications in the same. Similar methods have been described for synthetic zinc finger proteins, see for example U.S. Pat. No. 6,511,808, U.S. Pat. No. 6,013,453, U.S. Pat. No. 6,007,988, U.S. Pat. No. 6,503,717, U.S. patent application 2002/0164575, and U.S. patent application
10 2002/0160940.

 Conventional viral and non-viral based gene transfer methods can be used to introduce nucleic acids encoding the engineered TALE repeat array protein into mammalian cells or target tissues. Such methods can be used to administer nucleic acids encoding engineered TALE repeat array proteins to cells in vitro. Preferably, the nucleic
15 acids encoding the engineered TALE repeat array proteins are administered for in vivo or ex vivo gene therapy uses. Non-viral vector delivery systems include DNA plasmids, naked nucleic acid, and nucleic acid complexed with a delivery vehicle such as a liposome. Viral vector delivery systems include DNA and RNA viruses, which have either episomal or integrated genomes after delivery to the cell. For a review of gene
20 therapy procedures, see Anderson, 1992, Science, 256:808-813; Nabel & Felgner, 1993, TIBTECH, 11:211-217; Mitani & Caskey, 1993, TIBTECH, 11:162-166; Dillon, 1993, TIBTECH, 11:167-175; Miller, 1992, Nature, 357:455-460; Van Brunt, 1988, Biotechnology, 6:1149-54; Vigne, 1995, Restorat. Neurol. Neurosci., 8:35-36; Kremer & Perricaudet, 1995, Br. Med. Bull., 51:31-44; Haddada et al., in Current Topics in
25 Microbiology and Immunology Doerfler and Bohm (eds) (1995); and Yu et al., 1994, Gene Ther., 1:13-26.

 Methods of non-viral delivery of nucleic acids encoding the engineered TALE repeat array proteins include lipofection, microinjection, biolistics, virosomes, liposomes, immunoliposomes, polycation or lipid:nucleic acid conjugates, naked DNA or RNA,
30 artificial virions, and agent-enhanced uptake of DNA or RNA. Lipofection is described

in e.g., U.S. Pat. No. 5,049,386, No. 4,946,787; and No.4,897,355) and lipofection reagents are sold commercially (e.g., Transfectam™ and Lipofectin™). Cationic and neutral lipids that are suitable for efficient receptor-recognition lipofection of polynucleotides include those of Felgner, WO 91/17424, WO 91/16024. Delivery can be to cells (ex vivo administration) or target tissues (in vivo administration).

The preparation of lipid:nucleic acid complexes, including targeted liposomes such as immunolipid complexes, is well known to one of skill in the art (see, e.g., Crystal, 1995, *Science*, 270:404-410; Blaese et al., 1995, *Cancer Gene Ther.*, 2:291-297; Behr et al., 1994, *Bioconjugate Chem.* 5:382-389; Remy et al., 1994, *Bioconjugate Chem.*, 5:647-654; Gao et al., *Gene Ther.*, 2:710-722; Ahmad et al., 1992, *Cancer Res.*, 52:4817-20; U.S. Pat. Nos. 4,186,183, 4,217,344, 4,235,871, 4,261,975, 4,485,054, 4,501,728, 4,774,085, 4,837,028, and 4,946,787).

The use of RNA or DNA viral based systems for the delivery of nucleic acids encoding the engineered TALE repeat array proteins takes advantage of highly evolved processes for targeting a virus to specific cells in the body and trafficking the viral payload to the nucleus. Viral vectors can be administered directly to patients (in vivo) or they can be used to treat cells in vitro and the modified cells are administered to patients (ex vivo). Conventional viral based systems for the delivery of TALE repeat array proteins could include retroviral, lentivirus, adenoviral, adeno-associated, Sendai, and herpes simplex virus vectors for gene transfer. Viral vectors are currently the most efficient and versatile method of gene transfer in target cells and tissues. Integration in the host genome is possible with the retrovirus, lentivirus, and adeno-associated virus gene transfer methods, often resulting in long term expression of the inserted transgene. Additionally, high transduction efficiencies have been observed in many different cell types and target tissues.

The tropism of a retrovirus can be altered by incorporating foreign envelope proteins, expanding the potential target population of target cells. Lentiviral vectors are retroviral vectors that are able to transduce or infect non-dividing cells and typically produce high viral titers. Selection of a retroviral gene transfer system would therefore depend on the target tissue. Retroviral vectors are comprised of cis-acting long terminal

repeats with packaging capacity for up to 6-10 kb of foreign sequence. The minimum cis-acting LTRs are sufficient for replication and packaging of the vectors, which are then used to integrate the therapeutic gene into the target cell to provide permanent transgene expression. Widely used retroviral vectors include those based upon murine leukemia virus (MuLV), gibbon ape leukemia virus (GaLV), Simian Immuno deficiency virus (SIV), human immuno deficiency virus (HIV), and combinations thereof (see, e.g., Buchscher et al., 1992, *J. Virol.*, 66:2731-39; Johann et al., 1992, *J. Virol.*, 66:1635-40; Sommerfelt et al., 1990, *Virology*, 176:58-59; Wilson et al., 1989, *J. Virol.*, 63:2374-78; Miller et al., 1991, *J. Virol.*, 65:2220-24; WO 94/26877).

In applications where transient expression of the engineered TALE repeat array protein is preferred, adenoviral based systems can be used. Adenoviral based vectors are capable of very high transduction efficiency in many cell types and do not require cell division. With such vectors, high titer and levels of expression have been obtained. This vector can be produced in large quantities in a relatively simple system. Adeno-associated virus ("AAV") vectors are also used to transduce cells with target nucleic acids, e.g., in the in vitro production of nucleic acids and peptides, and for in vivo and ex vivo gene therapy procedures (see, e.g., West et al., 1987, *Virology* 160:38-47; U.S. Pat. No. 4,797,368; WO 93/24641; Kotin, 1994, *Hum. Gene Ther.*, 5:793-801; Muzyczka, 1994, *J. Clin. Invest.*, 94:1351). Construction of recombinant AAV vectors are described in a number of publications, including U.S. Pat. No. 5,173,414; Tratschin et al., 1985, *Mol. Cell. Biol.* 5:3251-60; Tratschin et al., 1984, *Mol. Cell. Biol.*, 4:2072-81; Hermonat & Muzyczka, 1984, *Proc. Natl. Acad. Sci. USA*, 81:6466-70; and Samulski et al., 1989, *J. Virol.*, 63:3822-28.

In particular, at least six viral vector approaches are currently available for gene transfer in clinical trials, with retroviral vectors by far the most frequently used system. All of these viral vectors utilize approaches that involve complementation of defective vectors by genes inserted into helper cell lines to generate the transducing agent.

pLASN and MFG-S are examples are retroviral vectors that have been used in clinical trials (Dunbar et al., 1995, *Blood*, 85:3048; Kohn et al., 1995, *Nat. Med.*, 1:1017; Malech et al., 1997, *Proc. Natl. Acad. Sci. USA*, 94:12133-38). PA317/pLASN was the

first therapeutic vector used in a gene therapy trial. (Blaese et al., 1995, *Science*, 270:475-480). Transduction efficiencies of 50% or greater have been observed for MFG-S packaged vectors (Ellem et al., 1997, *Immunol Immunother.*, 44:10-20; Dranoff et al., 1997, *Hum. Gene Ther.*, 1:111-112).

5 Recombinant adeno-associated virus vectors (rAAV) are a promising alternative gene delivery systems based on the defective and nonpathogenic parvovirus adeno-associated type 2 virus. Typically, the vectors are derived from a plasmid that retains only the AAV 145 bp inverted terminal repeats flanking the transgene expression cassette. Efficient gene transfer and stable transgene delivery due to integration into the genomes
10 of the transduced cell are key features for this vector system (Wagner et al., 1998, *Lancet*, 351:1702-1703; Kearns et al., 1996, *Gene Ther.*, 9:748-55).

 Replication-deficient recombinant adenoviral vectors (Ad) are predominantly used for colon cancer gene therapy, because they can be produced at high titer and they readily infect a number of different cell types. Most adenovirus vectors are engineered
15 such that a transgene replaces the Ad E1a, E1b, and E3 genes; subsequently the replication defector vector is propagated in human 293 cells that supply deleted gene function in trans. Ad vectors can transduce multiple types of tissues in vivo, including nondividing, differentiated cells such as those found in the liver, kidney and muscle system tissues. Conventional Ad vectors have a large carrying capacity. An example of
20 the use of an Ad vector in a clinical trial involved polynucleotide therapy for antitumor immunization with intramuscular injection (Serman et al., 1998, *Hum. Gene Ther.* 7:1083-89). Additional examples of the use of adenovirus vectors for gene transfer in clinical trials include Rosenecker et al., 1996, *Infection*, 24:15-10; Serman et al., 1998, *Hum. Gene Ther.*, 9:7 1083-89; Welsh et al., 1995, *Hum. Gene Ther.*, 2:205-218; Alvarez et al., 1997, *Hum. Gene Ther.* 5:597-613; Topf et al., 1998, *Gene Ther.*, 5:507-513;
25 Serman et al., 1998, *Hum. Gene Ther.*, 7:1083-89.

 Packaging cells are used to form virus particles that are capable of infecting a host cell. Such cells include 293 cells, which package adenovirus, and Ψ 2 cells or PA317 cells, which package retrovirus. Viral vectors used in gene therapy are usually generated
30 by producer cell line that packages a nucleic acid vector into a viral particle. The vectors

typically contain the minimal viral sequences required for packaging and subsequent integration into a host, other viral sequences being replaced by an expression cassette for the protein to be expressed. The missing viral functions are supplied in trans by the packaging cell line. For example, AAV vectors used in gene therapy typically only
5 possess ITR sequences from the AAV genome which are required for packaging and integration into the host genome. Viral DNA is packaged in a cell line, which contains a helper plasmid encoding the other AAV genes, namely rep and cap, but lacking ITR sequences. The cell line is also infected with adenovirus as a helper. The helper virus promotes replication of the AAV vector and expression of AAV genes from the helper
10 plasmid. The helper plasmid is not packaged in significant amounts due to a lack of ITR sequences. Contamination with adenovirus can be reduced by, e.g., heat treatment to which adenovirus is more sensitive than AAV.

In many gene therapy applications, it is desirable that the gene therapy vector be delivered with a high degree of specificity to a particular tissue type. A viral vector is
15 typically modified to have specificity for a given cell type by expressing a ligand as a fusion protein with a viral coat protein on the viruses outer surface. The ligand is chosen to have affinity for a receptor known to be present on the cell type of interest. For example, Han et al., 1995, Proc. Natl. Acad. Sci. USA, 92:9747-51, reported that Moloney murine leukemia virus can be modified to express human heregulin fused to
20 gp70, and the recombinant virus infects certain human breast cancer cells expressing human epidermal growth factor receptor. This principle can be extended to other pairs of virus expressing a ligand fusion protein and target cell expressing a receptor. For example, filamentous phage can be engineered to display antibody fragments (e.g., Fab or Fv) having specific binding affinity for virtually any chosen cellular receptor. Although
25 the above description applies primarily to viral vectors, the same principles can be applied to nonviral vectors. Such vectors can be engineered to contain specific uptake sequences thought to favor uptake by specific target cells.

Gene therapy vectors can be delivered in vivo by administration to an individual patient, typically by systemic administration (e.g., intravenous, intraperitoneal,
30 intramuscular, subdermal, or intracranial infusion) or topical application, as described

below. Alternatively, vectors can be delivered to cells ex vivo, such as cells explanted from an individual patient (e.g., lymphocytes, bone marrow aspirates, tissue biopsy) or stem cells (e.g., universal donor hematopoietic stem cells, embryonic stem cells (ES), partially differentiated stem cells, non-pluripotent stem cells, pluripotent stem cells, induced pluripotent stem cells (iPS cells) (see e.g., Sipione et al., *Diabetologia*, 47:499-508, 2004)), followed by reimplantation of the cells into a patient, usually after selection for cells which have incorporated the vector.

Ex vivo cell transfection for diagnostics, research, or for gene therapy (e.g., via re-infusion of the transfected cells into the host organism) is well known to those of skill in the art. In a preferred embodiment, cells are isolated from the subject organism, transfected with nucleic acid (gene or cDNA), encoding the engineered TALE repeat array protein, and re-infused back into the subject organism (e.g., patient). Various cell types suitable for ex vivo transfection are well known to those of skill in the art (see, e.g., Freshney et al., *Culture of Animal Cells, A Manual of Basic Technique* (5th ed. 2005)) and the references cited therein for a discussion of how to isolate and culture cells from patients).

In one embodiment, stem cells (e.g., universal donor hematopoietic stem cells, embryonic stem cells (ES), partially differentiated stem cells, non-pluripotent stem cells, pluripotent stem cells, induced pluripotent stem cells (iPS cells) (see e.g., Sipione et al., *Diabetologia*, 47:499-508, 2004)) are used in ex vivo procedures for cell transfection and gene therapy. The advantage to using stem cells is that they can be differentiated into other cell types in vitro, or can be introduced into a mammal (such as the donor of the cells) where they will engraft in the bone marrow. Methods for differentiating CD34+ cells in vitro into clinically important immune cell types using cytokines such as GM-CSF, IFN-gamma and TNF-alpha are known (see Inaba et al., 1992, *J. Exp. Med.*, 176:1693-1702).

Stem cells can be isolated for transduction and differentiation using known methods. For example, stem cells can be isolated from bone marrow cells by panning the bone marrow cells with antibodies which bind unwanted cells, such as CD4+ and CD8+

(T cells), CD45+ (panB cells), GR-1 (granulocytes), and Iad (differentiated antigen presenting cells) (see Inaba et al., 1992, J. Exp. Med., 176:1693-1702).

Vectors (e.g., retroviruses, adenoviruses, liposomes, etc.) containing nucleic acids encoding the engineered TALE repeat array protein can be also administered directly to the organism for transduction of cells in vivo. Alternatively, naked DNA can be administered. Administration is by any of the routes normally used for introducing a molecule into ultimate contact with blood or tissue cells. Suitable methods of administering such nucleic acids are available and well known to those of skill in the art, and, although more than one route can be used to administer a particular composition, a particular route can often provide a more immediate and more effective reaction than another route. Alternatively, stable formulations of the engineered TALE repeat array protein can also be administered.

Pharmaceutically acceptable carriers are determined in part by the particular composition being administered, as well as by the particular method used to administer the composition. Accordingly, there is a wide variety of suitable formulations of pharmaceutical compositions available, as described below (see, e.g., Remington: The Science and Practice of Pharmacy, 21st ed., 2005).

Delivery Vehicles

An important factor in the administration of polypeptide compounds, such as the engineered TALE repeat array proteins of the present invention, is ensuring that the polypeptide has the ability to traverse the plasma membrane of a cell, or the membrane of an intra-cellular compartment such as the nucleus. Cellular membranes are composed of lipid-protein bilayers that are freely permeable to small, nonionic lipophilic compounds and are inherently impermeable to polar compounds, macromolecules, and therapeutic or diagnostic agents. However, proteins and other compounds such as liposomes have been described, which have the ability to translocate polypeptides such as engineered TALE repeat array protein across a cell membrane.

For example, "membrane translocation polypeptides" have amphiphilic or hydrophobic amino acid subsequences that have the ability to act as membrane-

translocating carriers. In one embodiment, homeodomain proteins have the ability to translocate across cell membranes. The shortest internalizable peptide of a homeodomain protein, Antennapedia, was found to be the third helix of the protein, from amino acid position 43 to 58 (see, e.g., Prochiantz, 1996, *Curr. Opin. Neurobiol.*, 6:629-634).

5 Another subsequence, the h (hydrophobic) domain of signal peptides, was found to have similar cell membrane translocation characteristics (see, e.g., Lin et al., 1995, *J. Biol. Chem.*, 270:14255-58).

Examples of peptide sequences that can be linked to a protein, for facilitating uptake of the protein into cells, include, but are not limited to: peptide fragments of the tat protein of HIV (Endoh et al., 2010, *Methods Mol. Biol.*, 623:271-281; Schmidt et al., 2010, *FEBS Lett.*, 584:1806-13; Futaki, 2006, *Biopolymers*, 84:241-249); a 20 residue peptide sequence which corresponds to amino acids 84-103 of the p16 protein (see Fahraeus et al., 1996, *Curr. Biol.*, 6:84); the third helix of the 60-amino acid long homeodomain of Antennapedia (Derossi et al., 1994, *J. Biol. Chem.*, 269:10444); the h region of a signal peptide, such as the Kaposi fibroblast growth factor (K-FGF) h region (Lin et al., supra); or the VP22 translocation domain from HSV (Elliot & O'Hare, 1997, *Cell*, 88:223-233). See also, e.g., Caron et al., 2001, *Mol Ther.*, 3:310-318; Langel, *Cell-Penetrating Peptides: Processes and Applications* (CRC Press, Boca Raton FL 2002); El-Andaloussi et al., 2005, *Curr. Pharm. Des.*, 11:3597-3611; and Deshayes et al., 2005, 15 *Cell. Mol. Life Sci.*, 62:1839-49. Other suitable chemical moieties that provide enhanced cellular uptake can also be chemically linked to TALE repeat array proteins described herein.

Toxin molecules also have the ability to transport polypeptides across cell membranes. Often, such molecules are composed of at least two parts (called "binary toxins"): a translocation or binding domain or polypeptide and a separate toxin domain or polypeptide. Typically, the translocation domain or polypeptide binds to a cellular receptor, and then the toxin is transported into the cell. Several bacterial toxins, including Clostridium perfringens iota toxin, diphtheria toxin (DT), Pseudomonas exotoxin A (PE), pertussis toxin (PT), Bacillus anthracis toxin, and pertussis adenylate cyclase (CYA), 25 have been used in attempts to deliver peptides to the cell cytosol as internal or amino-

terminal fusions (Arora et al., 1993, *J. Biol. Chem.*, 268:3334-41; Perelle et al., 1993, *Infect. Immun.*, 61:5147-56; Stenmark et al., 1991, *J. Cell Biol.*, 113:1025-32; Donnelly et al., 1993, *Proc. Natl. Acad. Sci. USA*, 90:3530-34; Carbonetti et al., 1995, *Abstr. Annu. Meet. Am. Soc. Microbiol.* 95:295; Sebo et al., 1995, *Infect. Immun.*, 63:3851-57; 5 Klimpel et al., 1992, *Proc. Natl. Acad. Sci. USA*, 89:10277-81; and Novak et al., 1992, *J. Biol. Chem.*, 267:17186-93).

Such subsequences can be used to translocate engineered TALE repeat array proteins across a cell membrane. The engineered TALE repeat array proteins can be conveniently fused to or derivatized with such sequences. Typically, the translocation 10 sequence is provided as part of a fusion protein. Optionally, a linker can be used to link the engineered TALE repeat array protein and the translocation sequence. Any suitable linker can be used, e.g., a peptide linker.

The engineered TALE repeat array protein can also be introduced into an animal cell, preferably a mammalian cell, via liposomes and liposome derivatives such as 15 immunoliposomes. The term "liposome" refers to vesicles comprised of one or more concentrically ordered lipid bilayers, which encapsulate an aqueous phase. The aqueous phase typically contains the compound to be delivered to the cell, i.e., the engineered TALE repeat array protein.

The liposome fuses with the plasma membrane, thereby releasing the compound 20 into the cytosol. Alternatively, the liposome is phagocytosed or taken up by the cell in a transport vesicle. Once in the endosome or phagosome, the liposome either degrades or fuses with the membrane of the transport vesicle and releases its contents.

In current methods of drug delivery via liposomes, the liposome ultimately becomes permeable and releases the encapsulated compound (e.g., the engineered TALE 25 repeat array protein or a nucleic acid encoding the same) at the target tissue or cell. For systemic or tissue specific delivery, this can be accomplished, for example, in a passive manner wherein the liposome bilayer degrades over time through the action of various agents in the body. Alternatively, active compound release involves using an agent to induce a permeability change in the liposome vesicle. Liposome membranes can be 30 constructed so that they become destabilized when the environment becomes acidic near

the liposome membrane (see, e.g., Proc. Natl. Acad. Sci. USA, 84:7851 (1987); Biochemistry, 28:908 (1989)). When liposomes are endocytosed by a target cell, for example, they become destabilized and release their contents. This destabilization is termed fusogenesis. Dioleoylphosphatidylethanolamine (DOPE) is the basis of many "fusogenic" systems.

Such liposomes typically comprise the engineered TALE repeat array protein and a lipid component, e.g., a neutral and/or cationic lipid, optionally including a receptor-recognition molecule such as an antibody that binds to a predetermined cell surface receptor or ligand (e.g., an antigen). A variety of methods are available for preparing liposomes as described in, e.g., Szoka et al., 1980, Annu. Rev. Biophys. Bioeng., 9:467, U.S. Pat. Nos. 4,186,183, 4,217,344, 4,235,871, 4,261,975, 4,485,054, 4,501,728, 4,774,085, 4,837,028, 4,235,871, 4,261,975, 4,485,054, 4,501,728, 4,774,085, 4,837,028, 4,946,787, PCT Publication. No. WO 91/17424, Deamer & Bangham, 1976, Biochim. Biophys. Acta, 443:629-634; Fraley, et al., 1979, Proc. Natl. Acad. Sci. USA, 76:3348-52; Hope et al., 1985, Biochim. Biophys. Acta, 812:55-65; Mayer et al., 1986, Biochim. Biophys. Acta, 858:161-168; Williams et al., 1988, Proc. Natl. Acad. Sci. USA, 85:242-246; Liposomes (Ostro (ed.), 1983, Chapter 1); Hope et al., 1986, Chem. Phys. Lip., 40:89; Gregoriadis, Liposome Technology (1984) and Lasic, Liposomes: from Physics to Applications (1993)). Suitable methods include, for example, sonication, extrusion, high pressure/homogenization, microfluidization, detergent dialysis, calcium-induced fusion of small liposome vesicles and ether-fusion methods, all of which are well known in the art.

In certain embodiments, it is desirable to target liposomes using targeting moieties that are specific to a particular cell type, tissue, and the like. Targeting of liposomes using a variety of targeting moieties (e.g., ligands, receptors, and monoclonal antibodies) has been previously described (see, e.g., U.S. Pat. Nos. 4,957,773 and 4,603,044).

Examples of targeting moieties include monoclonal antibodies specific to antigens associated with neoplasms, such as prostate cancer specific antigen and MAGE. Tumors can also be diagnosed by detecting gene products resulting from the activation or over-expression of oncogenes, such as ras or c-erbB2. In addition, many tumors express antigens normally expressed by fetal tissue, such as the alpha-fetoprotein (AFP) and

carcinoembryonic antigen (CEA). Sites of viral infection can be diagnosed using various viral antigens such as hepatitis B core and surface antigens (HBVc, HBVs) hepatitis C antigens, Epstein-Barr virus antigens, human immunodeficiency type-1 virus (HIV1) and papilloma virus antigens. Inflammation can be detected using molecules specifically
5 recognized by surface molecules which are expressed at sites of inflammation such as integrins (e.g., VCAM-1), selectin receptors (e.g., ELAM-1) and the like.

Standard methods for coupling targeting agents to liposomes can be used. These methods generally involve incorporation into liposomes lipid components, e.g., phosphatidylethanolamine, which can be activated for attachment of targeting agents, or
10 derivatized lipophilic compounds, such as lipid derivatized bleomycin. Antibody targeted liposomes can be constructed using, for instance, liposomes which incorporate protein A (see Renneisen et al., 1990, J. Biol. Chem., 265:16337-42 and Leonetti et al., 1990, Proc. Natl. Acad. Sci. USA, 87:2448-51).

15 Dosages

For therapeutic applications, the dose of the engineered TALE repeat array protein to be administered to a patient is calculated in a similar way as has been described for zinc finger proteins, see for example U.S. Pat. No. 6,511,808, U.S. Pat. No. 6,492,117, U.S. Pat. No. 6,453,242, U.S. patent application 2002/0164575, and U.S. patent
20 application 2002/0160940. In the context of the present disclosure, the dose should be sufficient to effect a beneficial therapeutic response in the patient over time. In addition, particular dosage regimens can be useful for determining phenotypic changes in an experimental setting, e.g., in functional genomics studies, and in cell or animal models. The dose will be determined by the efficacy, specificity, and K_D of the particular
25 engineered TALE repeat array protein employed, the nuclear volume of the target cell, and the condition of the patient, as well as the body weight or surface area of the patient to be treated. The size of the dose also will be determined by the existence, nature, and extent of any adverse side-effects that accompany the administration of a particular compound or vector in a particular patient.

30

Pharmaceutical Compositions and Administration

Appropriate pharmaceutical compositions for administration of the engineered TALE repeat array proteins of the present invention can be determined as described for zinc finger proteins, see for example U.S. Pat. No. 6,511,808, U.S. Pat. No. 6,492,117, U.S. Pat. No. 6,453,242, U.S. patent application 2002/0164575, and U.S. patent application 2002/0160940. Engineered TALE repeat array proteins, and expression vectors encoding engineered TALE repeat array proteins, can be administered directly to the patient for modulation of gene expression and for therapeutic or prophylactic applications, for example, cancer, ischemia, diabetic retinopathy, macular degeneration, rheumatoid arthritis, psoriasis, HIV infection, sickle cell anemia, Alzheimer's disease, muscular dystrophy, neurodegenerative diseases, vascular disease, cystic fibrosis, stroke, and the like. Examples of microorganisms that can be inhibited by TALE repeat array protein-mediated gene therapy include pathogenic bacteria, e.g., chlamydia, rickettsial bacteria, mycobacteria, staphylococci, streptococci, pneumococci, meningococci and conococci, klebsiella, proteus, serratia, pseudomonas, legionella, diphtheria, salmonella, bacilli, cholera, tetanus, botulism, anthrax, plague, leptospirosis, and Lyme disease bacteria; infectious fungus, e.g., Aspergillus, Candida species; protozoa such as sporozoa (e.g., Plasmodia), rhizopods (e.g., Entamoeba) and flagellates (Trypanosoma, Leishmania, Trichomonas, Giardia, etc.); viral diseases, e.g., hepatitis (A, B, or C), herpes virus (e.g., VZV, HSV-1, HSV-6, HSV-II, CMV, and EBV), HIV, Ebola, adenovirus, influenza virus, flaviviruses, echovirus, rhinovirus, coxsackie virus, comovirus, respiratory syncytial virus, mumps virus, rotavirus, measles virus, rubella virus, parvovirus, vaccinia virus, HTLV virus, dengue virus, papillomavirus, poliovirus, rabies virus, and arboviral encephalitis virus, etc.

Administration of therapeutically effective amounts is by any of the routes normally used for introducing TALE repeat array proteins into ultimate contact with the tissue to be treated. The TALE repeat array proteins are administered in any suitable manner, preferably with pharmaceutically acceptable carriers. Suitable methods of administering such modulators are available and well known to those of skill in the art, and, although more than one route can be used to administer a particular composition, a

particular route can often provide a more immediate and more effective reaction than another route.

Pharmaceutically acceptable carriers are determined in part by the particular composition being administered, as well as by the particular method used to administer
5 the composition. Accordingly, there is a wide variety of suitable formulations of pharmaceutical compositions that are available (see, e.g., Remington: The Science and Practice of Pharmacy, 21st ed., 2005).

The engineered TALE repeat array proteins, alone or in combination with other suitable components, can be made into aerosol formulations (i.e., they can be
10 "nebulized") to be administered via inhalation. Aerosol formulations can be placed into pressurized acceptable propellants, such as dichlorodifluoromethane, propane, nitrogen, and the like.

Formulations suitable for parenteral administration, such as, for example, by intravenous, intramuscular, intradermal, and subcutaneous routes, include aqueous and
15 non-aqueous, isotonic sterile injection solutions, which can contain antioxidants, buffers, bacteriostats, and solutes that render the formulation isotonic with the blood of the intended recipient, and aqueous and non-aqueous sterile suspensions that can include suspending agents, solubilizers, thickening agents, stabilizers, and preservatives. The disclosed compositions can be administered, for example, by intravenous infusion, orally,
20 topically, intraperitoneally, intravesically or intrathecally. The formulations of compounds can be presented in unit-dose or multi-dose sealed containers, such as ampules and vials. Injection solutions and suspensions can be prepared from sterile powders, granules, and tablets of the kind previously described.

25 Use of TALE Nucleases

TALE nucleases engineered using the methods described herein can be used to induce mutations in a genomic sequence, e.g., by cleaving at two sites and deleting sequences in between, by cleavage at a single site followed by non-homologous end joining, and/or by cleaving at a site so as to remove or replace one or two or a few
30 nucleotides. In some embodiments, the TALE nuclease is used to induce mutation in an

animal, plant, fungal, or bacterial genome. Targeted cleavage can also be used to create gene knock-outs (e.g., for functional genomics or target validation) and to facilitate targeted insertion of a sequence into a genome (i.e., gene knock-in); e.g., for purposes of cell engineering or protein overexpression. Insertion can be by means of replacements of chromosomal sequences through homologous recombination or by targeted integration, in which a new sequence (i.e., a sequence not present in the region of interest), flanked by sequences homologous to the region of interest in the chromosome, is used to insert the new sequence at a predetermined target site via homologous recombination. Exogenous DNA can also be inserted into TALE nuclease-induced double stranded breaks without the need for flanking homology sequences (see, Orlando et al., 2010, Nucl. Acids Res., 1-15, doi:10.1093/nar/gkq512).

As demonstrated in Example 3 below, the TALE nucleases produced by the methods described herein were capable of inducing site-specific mutagenesis in mammalian cells. A skilled practitioner will readily appreciate that TALE nucleases produced by the methods described herein would also function to induce efficient site-specific mutagenesis in other cell types and organisms (see, for example, Cade et al., 2012, Nucleic Acids Res., PMID: 22684503 and Moore et al., 2012, PLoS One, PMID: 22655075).

The same methods can also be used to replace a wild-type sequence with a mutant sequence, or to convert one allele to a different allele.

Targeted cleavage of infecting or integrated viral genomes can be used to treat viral infections in a host. Additionally, targeted cleavage of genes encoding receptors for viruses can be used to block expression of such receptors, thereby preventing viral infection and/or viral spread in a host organism. Targeted mutagenesis of genes encoding viral receptors (e.g., the CCR5 and CXCR4 receptors for HIV) can be used to render the receptors unable to bind to virus, thereby preventing new infection and blocking the spread of existing infections. Non-limiting examples of viruses or viral receptors that can be targeted include herpes simplex virus (HSV), such as HSV-1 and HSV-2, varicella zoster virus (VZV), Epstein-Barr virus (EBV) and cytomegalovirus (CMV), HHV6 and HHV7. The hepatitis family of viruses includes hepatitis A virus (HAV), hepatitis B virus

(HBV), hepatitis C virus (HCV), the delta hepatitis virus (HDV), hepatitis E virus (HEV) and hepatitis G virus (HGV). Other viruses or their receptors can be targeted, including, but not limited to, Picornaviridae (e.g., polioviruses, etc.); Caliciviridae; Togaviridae (e.g., rubella virus, dengue virus, etc.); Flaviviridae; Coronaviridae; Reoviridae; 5 Bimaviridae; Rhabdoviridae (e.g., rabies virus, etc.); Filoviridae; Paramyxoviridae (e.g., mumps virus, measles virus, respiratory syncytial virus, etc.); Orthomyxoviridae (e.g., influenza virus types A, B and C, etc.); Bunyaviridae; Arenaviridae; Retroviridae; lentiviruses (e.g., HTLV-I; HTLV-II; HIV-1 (also known as HTLV-III, LAV, ARV, hTLR, etc.) HIV-II); simian immunodeficiency virus (SIV), human papillomavirus (HPV), 10 influenza virus and the tick-borne encephalitis viruses. See, e.g., Virology, 3rd Edition (W. K. Joklik, ed. 1988); Fundamental Virology, 4th Edition (Knipe and Howley, eds. 2001), for a description of these and other viruses. Receptors for HIV, for example, include CCR-5 and CXCR-4.

In similar fashion, the genome of an infecting bacterium can be mutagenized by 15 targeted DNA cleavage followed by non-homologous end joining, to block or ameliorate bacterial infections.

The disclosed methods for targeted recombination can be used to replace any genomic sequence with a homologous, non-identical sequence. For example, a mutant genomic sequence can be replaced by its wild-type counterpart, thereby providing 20 methods for treatment of e.g., genetic disease, inherited disorders, cancer, and autoimmune disease. In like fashion, one allele of a gene can be replaced by a different allele using the methods of targeted recombination disclosed herein.

Exemplary genetic diseases include, but are not limited to, achondroplasia, achromatopsia, acid maltase deficiency, adenosine deaminase deficiency (OMIM 25 No.102700), adrenoleukodystrophy, aicardi syndrome, alpha-1 antitrypsin deficiency, alpha-thalassemia, androgen insensitivity syndrome, apert syndrome, arrhythmogenic right ventricular, dysplasia, ataxia telangiectasia, barth syndrome, beta-thalassemia, blue rubber bleb nevus syndrome, canavan disease, chronic granulomatous diseases (CGD), cri du chat syndrome, cystic fibrosis, dercum's disease, ectodermal dysplasia, Fanconi 30 anemia, fibrodysplasia ossificans progressive, fragile X syndrome, galactosemia,

Gaucher's disease, generalized gangliosidoses (e.g., GM1), hemochromatosis, the hemoglobin C mutation in the 6th codon of beta-globin (HbC), hemophilia, Huntington's disease, Hurler Syndrome, hypophosphatasia, Klinefelter's syndrome, Krabbes Disease, Langer-Giedion Syndrome, leukocyte adhesion deficiency (LAD, OMIM No. 116920),
5 leukodystrophy, long QT syndrome, Marfan syndrome, Moebius syndrome, mucopolysaccharidosis (MPS), nail patella syndrome, nephrogenic diabetes insipidus, neurofibromatosis, Neimann-Pick disease, osteogenesis imperfecta, porphyria, Prader-Willi syndrome, progeria, Proteus syndrome, retinoblastoma, Rett syndrome, Rubinstein-Taybi syndrome, Sanfilippo syndrome, severe combined immunodeficiency (SCID),
10 Shwachman syndrome, sickle cell disease (sickle cell anemia), Smith-Magenis syndrome, Stickler syndrome, Tay-Sachs disease, Thrombocytopenia Absent Radius (TAR) syndrome, Treacher Collins syndrome, trisomy, tuberous sclerosis, Turner's syndrome, urea cycle disorder, von Hippel-Landau disease, Waardenburg syndrome, Williams syndrome, Wilson's disease, Wiskott-Aldrich syndrome, X-linked lymphoproliferative
15 syndrome (XLP, OMIM No. 308240).

Additional exemplary diseases that can be treated by targeted DNA cleavage and/or homologous recombination include acquired immunodeficiencies, lysosomal storage diseases (e.g., Gaucher's disease, GM1, Fabry disease and Tay-Sachs disease), mucopolysaccharidosis (e.g., Hunter's disease, Hurler's disease), hemoglobinopathies
20 (e.g., sickle cell diseases, HbC, alpha-thalassemia, beta-thalassemia) and hemophilias.

In certain cases, alteration of a genomic sequence in a pluripotent cell (e.g., a hematopoietic stem cell) is desired. Methods for mobilization, enrichment and culture of hematopoietic stem cells are known in the art. See for example, U.S. Pat. Nos. 5,061,620; 5,681,559; 6,335,195; 6,645,489 and 6,667,064. Treated stem cells can be
25 returned to a patient for treatment of various diseases including, but not limited to, SCID and sickle-cell anemia.

In many of these cases, a region of interest comprises a mutation, and the donor polynucleotide comprises the corresponding wild-type sequence. Similarly, a wild-type genomic sequence can be replaced by a mutant sequence, if such is desirable. For
30 example, overexpression of an oncogene can be reversed either by mutating the gene or

by replacing its control sequences with sequences that support a lower, non-pathologic level of expression. As another example, the wild-type allele of the ApoAI gene can be replaced by the ApoAI Milano allele, to treat atherosclerosis. Indeed, any pathology dependent upon a particular genomic sequence, in any fashion, can be corrected or
5 alleviated using the methods and compositions disclosed herein.

Targeted cleavage and targeted recombination can also be used to alter non-coding sequences (e.g., sequences encoding microRNAs and long non-coding RNAs, and regulatory sequences such as promoters, enhancers, initiators, terminators, splice sites) to alter the levels of expression of a gene product. Such methods can be used, for example,
10 for therapeutic purposes, functional genomics and/or target validation studies.

The compositions and methods described herein also allow for novel approaches and systems to address immune reactions of a host to allogeneic grafts. In particular, a major problem faced when allogeneic stem cells (or any type of allogeneic cell) are grafted into a host recipient is the high risk of rejection by the host's immune system,
15 primarily mediated through recognition of the Major Histocompatibility Complex (MHC) on the surface of the engrafted cells. The MHC comprises the HLA class I protein(s) that function as heterodimers that are comprised of a common beta subunit and variable alpha subunits. It has been demonstrated that tissue grafts derived from stem cells that are devoid of HLA escape the host's immune response. See, e.g., Coffman et al., 1993, J.
20 Immunol., 151:425-35; Markmann et al., 1992, Transplantation, 54:1085-89; Koller et al., 1990, Science, 248:1227-30. Using the compositions and methods described herein, genes encoding HLA proteins involved in graft rejection can be cleaved, mutagenized or altered by recombination, in either their coding or regulatory sequences, so that their expression is blocked or they express a non-functional product. For example, by
25 inactivating the gene encoding the common beta subunit gene (beta2 microglobulin) using TALE nuclease fusion proteins as described herein, HLA class I can be removed from the cells to rapidly and reliably generate HLA class I null stem cells from any donor, thereby reducing the need for closely matched donor/recipient MHC haplotypes during stem cell grafting.

Inactivation of any gene (e.g., the beta2 microglobulin gene) can be achieved, for example, by a single cleavage event, by cleavage followed by non-homologous end joining, by cleavage at two sites followed by joining so as to delete the sequence between the two cleavage sites, by targeted recombination of a missense or nonsense codon into the coding region, or by targeted recombination of an irrelevant sequence (i.e., a "stuffer" sequence) into the gene or its regulatory region, so as to disrupt the gene or regulatory region.

Targeted modification of chromatin structure, as disclosed in WO 01/83793, can be used to facilitate the binding of fusion proteins to cellular chromatin.

In additional embodiments, one or more fusions between a TALE binding domain and a recombinase (or functional fragment thereof) can be used, in addition to or instead of the TALE-cleavage domain fusions disclosed herein, to facilitate targeted recombination. See, for example, co-owned U.S. Pat. No. 6,534,261 and Akopian et al. (2003) Proc. Natl. Acad. Sci. USA 100:8688-8691.

In additional embodiments, the disclosed methods and compositions are used to provide fusions of TALE repeat DNA-binding domains with transcriptional activation or repression domains that require dimerization (either homodimerization or heterodimerization) for their activity. In these cases, a fusion polypeptide comprises a TALE repeat DNA-binding domain and a functional domain monomer (e.g., a monomer from a dimeric transcriptional activation or repression domain). Binding of two such fusion polypeptides to properly situated target sites allows dimerization so as to reconstitute a functional transcription activation or repression domain.

Regulation of Gene Expression in Plants

Engineered TALE repeat array proteins can be used to engineer plants for traits such as increased disease resistance, modification of structural and storage polysaccharides, flavors, proteins, and fatty acids, fruit ripening, yield, color, nutritional characteristics, improved storage capability, and the like. In particular, the engineering of crop species for enhanced oil production, e.g., the modification of the fatty acids produced in oilseeds, is of interest.

Seed oils are composed primarily of triacylglycerols (TAGs), which are glycerol esters of fatty acids. Commercial production of these vegetable oils is accounted for primarily by six major oil crops (soybean, oil palm, rapeseed, sunflower, cotton seed, and peanut). Vegetable oils are used predominantly (90%) for human consumption as margarine, shortening, salad oils, and frying oil. The remaining 10% is used for non-food applications such as lubricants, oleochemicals, biofuels, detergents, and other industrial applications.

The desired characteristics of the oil used in each of these applications varies widely, particularly in terms of the chain length and number of double bonds present in the fatty acids making up the TAGs. These properties are manipulated by the plant in order to control membrane fluidity and temperature sensitivity. The same properties can be controlled using TALE repeat array proteins to produce oils with improved characteristics for food and industrial uses.

The primary fatty acids in the TAGs of oilseed crops are 16 to 18 carbons in length and contain 0 to 3 double bonds. Palmitic acid (16:0 [16 carbons: 0 double bonds]), oleic acid (18:1), linoleic acid (18:2), and linolenic acid (18:3) predominate. The number of double bonds, or degree of saturation, determines the melting temperature, reactivity, cooking performance, and health attributes of the resulting oil.

The enzyme responsible for the conversion of oleic acid (18:1) into linoleic acid (18:2) (which is then the precursor for 18:3 formation) is delta-12-oleate desaturase, also referred to as omega-6 desaturase. A block at this step in the fatty acid desaturation pathway should result in the accumulation of oleic acid at the expense of polyunsaturates.

In one embodiment engineered TALE repeat array proteins are used to regulate expression of the FAD2-1 gene in soybeans. Two genes encoding microsomal delta-6 desaturases have been cloned recently from soybean, and are referred to as FAD2-1 and FAD2-2 (Heppard et al., 1996, Plant Physiol. 110:311-319). FAD2-1 (delta-12 desaturase) appears to control the bulk of oleic acid desaturation in the soybean seed. Engineered TALE repeat array proteins can thus be used to modulate gene expression of FAD2-1 in plants. Specifically, engineered TALE repeat array proteins can be used to inhibit expression of the FAD2-1 gene in soybean in order to increase the accumulation

of oleic acid (18:1) in the oil seed. Moreover, engineered TALE proteins can be used to modulate expression of any other plant gene, such as delta-9 desaturase, delta-12 desaturases from other plants, delta-15 desaturase, acetyl-CoA carboxylase, acyl-ACP-thioesterase, ADP-glucose pyrophosphorylase, starch synthase, cellulose synthase, sucrose synthase, senescence-associated genes, heavy metal chelators, fatty acid hydroperoxide lyase, polygalacturonase, EPSP synthase, plant viral genes, plant fungal pathogen genes, and plant bacterial pathogen genes.

Recombinant DNA vectors suitable for transformation of plant cells are also used to deliver protein (e.g., engineered TALE repeat array protein)-encoding nucleic acids to plant cells. Techniques for transforming a wide variety of higher plant species are well known and described in the technical and scientific literature (see, e.g., Weising et al., 1988, *Ann. Rev. Genet.*, 22:421-477). A DNA sequence coding for the desired TALE repeat array protein is combined with transcriptional and translational initiation regulatory sequences which will direct the transcription of the TALE protein in the intended tissues of the transformed plant.

For example, a plant promoter fragment can be employed which will direct expression of the engineered TALE repeat array protein in all tissues of a regenerated plant. Such promoters are referred to herein as "constitutive" promoters and are active under most environmental conditions and states of development or cell differentiation. Examples of constitutive promoters include the cauliflower mosaic virus (CaMV) 35 S transcription initiation region, the 1'- or 2'-promoter derived from T-DNA of *Agrobacterium tumefaciens*, and other transcription initiation regions from various plant genes known to those of skill.

Alternatively, the plant promoter can direct expression of the engineered TALE repeat array protein in a specific tissue or can be otherwise under more precise environmental or developmental control. Such promoters are referred to here as "inducible" promoters. Examples of environmental conditions that can affect transcription by inducible promoters include anaerobic conditions or the presence of light.

Examples of promoters under developmental control include promoters that initiate transcription only in certain tissues, such as fruit, seeds, or flowers. For example, the use of a polygalacturonase promoter can direct expression of the TALE repeat array protein in the fruit, a CHS-A (chalcone synthase A from petunia) promoter can direct
5 expression of the TALE repeat array protein in the flower of a plant.

The vector comprising the TALE repeat array protein sequences will typically comprise a marker gene which confers a selectable phenotype on plant cells. For example, the marker can encode biocide resistance, particularly antibiotic resistance, such as resistance to kanamycin, G418, bleomycin, hygromycin, or herbicide resistance, such
10 as resistance to chlorosulfuron or Basta.

Such DNA constructs can be introduced into the genome of the desired plant host by a variety of conventional techniques. For example, the DNA construct can be introduced directly into the genomic DNA of the plant cell using techniques such as electroporation and microinjection of plant cell protoplasts, or the DNA constructs can be
15 introduced directly to plant tissue using biolistic methods, such as DNA particle bombardment. Alternatively, the DNA constructs can be combined with suitable T-DNA flanking regions and introduced into a conventional *Agrobacterium tumefaciens* host vector. The virulence functions of the *Agrobacterium tumefaciens* host will direct the insertion of the construct and adjacent marker into the plant cell DNA when the cell is
20 infected by the bacteria.

Microinjection techniques are known in the art and well described in the scientific and patent literature. The introduction of DNA constructs using polyethylene glycol precipitation is described in Paszkowski et al., 1984, *EMBO J.*, 3:2717-22. Electroporation techniques are described in Fromm et al. 1985, *Proc. Natl. Acad. Sci. USA*, 82:5824. Biolistic transformation techniques are described in Klein et al., 1987,
25 *Nature*, 327:70-73.

Agrobacterium tumefaciens-mediated transformation techniques are well described in the scientific literature (see, e.g., Horsch et al., 1984, *Science*, 233:496-498; and Fraley et al., 1983, *Proc. Natl. Acad. Sci. USA*, 80:4803).

Transformed plant cells which are derived by any of the above transformation techniques can be cultured to regenerate a whole plant which possesses the transformed genotype and thus the desired TALE repeat array protein-controlled phenotype. Such regeneration techniques rely on manipulation of certain phytohormones in a tissue culture growth medium, typically relying on a biocide and/or herbicide marker which has been introduced together with the TALE repeat array protein nucleotide sequences. Plant regeneration from cultured protoplasts is described in Evans et al., Protoplasts Isolation and Culture, Handbook of Plant Cell Culture, pp. 124-176 (1983); and Binding, Regeneration of Plants, Plant Protoplasts, pp. 21-73 (1985). Regeneration can also be obtained from plant callus, explants, organs, or parts thereof. Such regeneration techniques are described generally in Klee et al., 1987, Ann. Rev. Plant Phys., 38:467-486.

Functional Genomics Assays

Engineered TALE repeat array proteins also have use for assays to determine the phenotypic consequences and function of gene expression. Recent advances in analytical techniques, coupled with focused mass sequencing efforts have created the opportunity to identify and characterize many more molecular targets than were previously available. This new information about genes and their functions will improve basic biological understanding and present many new targets for therapeutic intervention. In some cases analytical tools have not kept pace with the generation of new data. An example is provided by recent advances in the measurement of global differential gene expression. These methods, typified by gene expression microarrays, differential cDNA cloning frequencies, subtractive hybridization and differential display methods, can very rapidly identify genes that are up or down-regulated in different tissues or in response to specific stimuli. Increasingly, such methods are being used to explore biological processes such as, transformation, tumor progression, the inflammatory response, neurological disorders etc. Many differentially expressed genes correlate with a given physiological phenomenon, but demonstrating a causative relationship between an individual differentially expressed gene and the phenomenon is labor intensive. Until now, simple

methods for assigning function to differentially expressed genes have not kept pace with the ability to monitor differential gene expression.

The engineered TALE repeat array proteins described herein can be used to rapidly analyze the function of a differentially expressed gene. Engineered TALE
5 proteins can be readily used to up or down-regulate or knockout any endogenous target gene, or to knock in an endogenous or endogenous gene. Very little sequence information is required to create a gene-specific DNA binding domain. This makes the engineered TALE repeat array technology ideal for analysis of long lists of poorly
10 characterized differentially expressed genes. One can simply build a TALE repeat array protein-based DNA binding domain for each candidate gene, create chimeric up and down-regulating artificial transcription factors and test the consequence of up or down-regulation on the phenotype under study (e.g., transformation or response to a cytokine) by switching the candidate genes on or off one at a time in a model system.

Additionally, greater experimental control can be imparted by engineered TALE
15 repeat array proteins than can be achieved by more conventional methods. This is because the production and/or function of engineered TALE repeat array proteins can be placed under small molecule control. Examples of this approach are provided by the Tet-On system, the ecdysone-regulated system and a system incorporating a chimeric factor including a mutant progesterone receptor. These systems are all capable of indirectly
20 imparting small molecule control on any endogenous gene of interest or any transgene by placing the function and/or expression of a engineered TALE repeat array protein under small molecule control.

Transgenic Animals

25 A further application of engineered TALE repeat array proteins is manipulating gene expression in animal models. As with cell lines, the introduction of a heterologous gene into or knockout of an endogenous in a transgenic animal, such as a transgenic mouse or zebrafish, is a fairly straightforward process. Thus, transgenic or transient expression of an engineered TALE repeat array protein in an animal can be readily
30 performed.

By transgenically or transiently expressing a suitable engineered TALE repeat array protein fused to an activation domain, a target gene of interest can be over-expressed. Similarly, by transgenically or transiently expressing a suitable engineered TALE repeat array protein fused to a repressor or silencer domain, the expression of a target gene of interest can be down-regulated, or even switched off to create "functional knockout". Knock-in or knockout mutations by insertion or deletion of a target gene of interest can be prepared using TALE nucleases.

Two common issues often prevent the successful application of the standard transgenic and knockout technology; embryonic lethality and developmental compensation. Embryonic lethality results when the gene plays an essential role in development. Developmental compensation is the substitution of a related gene product for the gene product being knocked out, and often results in a lack of a phenotype in a knockout mouse when the ablation of that gene's function would otherwise cause a physiological change.

Expression of transgenic engineered TALE repeat array proteins can be temporally controlled, for example using small molecule regulated systems as described in the previous section. Thus, by switching on expression of an engineered TALE repeat array protein at a desired stage in development, a gene can be over-expressed or "functionally knocked-out" in the adult (or at a late stage in development), thus avoiding the problems of embryonic lethality and developmental compensation.

EXAMPLES

Example 1. Assembly of TALE Repeat Arrays using streptavidin coated magnetic beads

An archive of DNA plasmids (~850 different plasmids) encoding one, two, three, or four TALE repeat domains was created for assembly of nucleic acids encoding multiple TALE arrays of any desired length. The plasmids were created by cloning synthetic arrays of one, two, three or four TALE repeat domains into the pUC57- Δ Bsal backbone (FIG. 3). The TALE repeats were of the arrangement α , $\beta\gamma\delta\epsilon$, $\beta\gamma\delta$, $\beta\gamma'$, $\beta\gamma$, $\delta\epsilon'$, and β , and included hypervariable triplet residues at each position to bind to the nucleotides as shown in Table 1. Polypeptide and nucleotide sequences of the TALE

repeat types are shown in FIGs. 4A and 4B, respectively. The polypeptide and polynucleotide sequences were varied slightly among the four types to reduce the possibility of recombination-mediated mutations due to long sequences of exact repeats.

5

Table 1. Nucleotide binding code of TALE triplets

Triplet	Bound Nucleotide
SNI	A
SHD	C
NNN	G
SNK	G
SNG	T

A 16-mer TALE repeat array targeted to the eGFP gene was created by in vitro assembly of 16 TALE repeats designed to bind the target sequence GCAGTGCTTCAGCCGC (SEQ ID NO: 41). In the first step, a plasmid carrying an α -type TALE repeat with an NNN triplet (G) was amplified by PCR using a biotinylated forward primer Biotin-TCTAGAGAAGACAAGAACCTGACC (SEQ ID NO: 42) and a reverse primer GGATCCGGTCTCTTAAGGCCGTGG (SEQ ID NO: 43). The amplified fragment (50 μ l) was purified using a QIA Quick PCR purification kit (QIAGEN), eluted in 40 μ l 0.1X elution buffer (as provided in the QIA Quick PCR purification kit), and digested with BsaI HF (New England Biolabs (NEB)) in NEB Buffer 4 for 15 minutes at 50 °C (40 μ l elution, 5 μ l NEBuffer 4, 5 μ l BsaI HF). The digested fragment was purified using a QIA Quick PCR purification kit and eluted in 0.1X elution buffer (50 μ l).

A plasmid containing a four TALE repeat domain sub-array unit ($\beta\gamma\delta\epsilon$) coding for repeats that each harbor one of the following variable amino acids SHD, SNI, NNN, and SNG (designed to bind the sequence 5'-CAGT-3') was digested with BbsI (NEB) in NEBuffer 2 for 2 hours at 37 °C in 100 μ l (50 μ l plasmid [\sim 200 ng/ μ l], 10 μ l NEBuffer 2, 10 μ l BbsI, 30 μ l water). To the 100 μ l digest was added 25 μ l NEBuffer 4, 2.5 μ l 100X BSA (NEB), 107.5 μ l water, and 5 μ l XbaI (NEB), and the digest was incubated for 5

minutes at 37 °C. To the mixture, 5 µl of BamHI HF was then added for a 5 minute digest at 37 °C, and then 5 µl SalI HF (NEB) was added for an additional 5 minute digest at 37 °C. The resulting fragment was purified using a QIA Quick PCR purification kit (QIAGEN) and eluted in 180 µl 0.1X elution buffer.

5 For the initial ligation, 2 µl of the alpha unit digest was mixed with 2.5 µl of T4 DNA ligase (400 U/µl; NEB) and 27 µl Quick Ligase Buffer (QLB) (NEB). To this 31.5 µl mixture was added 22.5 µl of the first digested subarray, and the mixture was ligated for 15 minutes at room temperature. Magnetic beads were prepared by washing 5 µl of Dynabeads MyOne Streptavidin C1 (Invitrogen) three times with 50 µl 1x B&W
10 Buffer (5.0 mM Tris-HCl [pH 7.5], 0.5 mM EDTA, 1.0 M NaCl, 0.005% Tween 20TM) and resuspending in 54 µl B&W Buffer. The ligated mixture was added to the washed beads and incubated for 15 minutes at room temperature (with mixing every five minutes). The mixture was then placed on a SPRIplate 96-well Ring magnet for 3 minutes. The supernatant was then aspirated, and 100 µl 1x B&W Buffer was added to wash, with
15 mixing by moving the beads 31 times from side to side within the tube using a DynaMag-96 Side magnet (Invitrogen). The B&W Buffer was then aspirated, and 100 µl 1x BSA was added, with mixing, then aspirated. The ligated, bead-bound nucleic acids ($\alpha\beta\gamma\delta\epsilon$) were resuspended in 50 µl BsaI HF mix (5 µl NEBuffer 4, 2 µl BsaI HF, 43 µl water).

The digest was incubated at 50 °C for 10 minutes, and 50 µl 1X B&W buffer was
20 added. The digest was placed on a magnet for 3 minutes, and the supernatant was aspirated. The beads were washed with 100 µl 1X B&W Buffer and 100 µl 1X BSA as above. To the washed beads were added a digested plasmid containing a four TALE repeat domain sub-array unit ($\beta\gamma\delta\epsilon$) coding for repeats that each harbor one of the following variable amino acids NNN, SHD, SNG, and SNG (designed to bind the DNA
25 sequence 5'-GCTT-3') (22.5 µl) and 27.5 µl ligase mix (25 µl Quick Ligase Buffer, 2 µl DNA ligase). The beads were resuspended by pipetting up and down, and the mixture was incubated for 15 minutes at room temperature with mixing every five minutes. To the ligation was added 50 µl 1X B&W Buffer, and the mixture was placed on the magnet for 3 minutes. The supernatant was aspirated, and the beads were washed with 100 µl 1X
30 B&W Buffer and 100 µl 1X BSA as above. The ligated, bead-bound nucleic acids

($\alpha\beta\gamma\delta\epsilon\beta\gamma\delta\epsilon$) were resuspended in 50 μ l BsaI HF mix (5 μ l NEBuffer 4, 2 μ l BsaI HF, 43 μ l water). Two more TALE repeat sub-array units were ligated sequentially as above, the first a four TALE repeat sub-array unit ($\beta\gamma\delta\epsilon$) coding for repeats that each harbor one of the following variable amino acids SHD, SNI, NNN, and SHD (designed to bind the DNA sequence 5'-CAGC-3') and the second a three TALE repeat sub-array unit ($\beta\gamma\delta$) coding for repeats that each harbor one of the following variable amino acids SHD, NNN, and SHD (designed to bind the DNA sequence 5'-CGC-3'). The final TALE repeat array contained subunits of the format $\alpha\beta\gamma\delta\epsilon\beta\gamma\delta\epsilon\beta\gamma\delta\epsilon\beta\gamma\delta$ with individual TALE repeats designed to bind the target DNA sequence 5'-GCAGTGCTTCAGCCGC-3' (SEQ ID NO: 44).

Following the final ligation step, the construct was digested with BsaI HF for eventual cloning into an expression vector and the beads were washed with 1X B&W Buffer and 1X BSA. The washed beads were resuspended in 50 μ l BbsI mix (5 μ l NEBuffer 2, 5 μ l BbsI, 40 μ l water) and incubated at 37 °C for 2 hours with agitation at 1500 rpm to cleave the biotinylated 5' end and release the assembled TALE repeat array from the magnetic beads. The digested mixture was purified by MinElute column purified (QIAGEN) and ligated into a BsmBI-digested TALE expression vector. The ligated mixture was transformed into chemically competent XL1 Blue cells and plated on LB/Carb¹⁰⁰ plates overnight.

The expression vectors each harbor the following elements: a T7 promoter, a nuclear localization signal, a FLAG tag, amino acids 153 to 288 from the TALE13 protein (numbering as defined by Miller et al., 2011, Nat. Biotechnol., 29:143-148), two adjacent BsmBI restriction sites into which a DNA fragment encoding a TALE repeat array can be cloned, a 0.5 TALE repeat, amino acids 715 to 777 from the C-terminal end of the TALE13 protein (numbering as defined by Miller et al., 2011, Nat. Biotechnol., 29:143-148), and the wild-type FokI cleavage domain.

The plasmids differ in the identity of the C-terminal 0.5 TALE repeat. Plasmid pJDS70 encodes a 0.5 TALE repeat with a SNI RVD (for recognition of an A nucleotide), plasmid pJDS71 encodes a 0.5 TALE repeat with a SHD RVD (for recognition of a C nucleotide), plasmid pJDS74 encodes a 0.5 TALE repeat with a NNN RVD (for

recognition of a G nucleotide), plasmid pJDS76 encodes a 0.5 TALE repeat with a SNK RVD (for recognition of a G nucleotide), and plasmid pJDS78 encodes a 0.5 TALE repeat with a NG RVD (for recognition of a T nucleotide). All plasmids share the common sequence shown in FIGs. 5A-5B and differ at just nine nucleotide positions marked as
 5 XXXXXXXXXX (underlined and bold). The sequence of these 9 bps and plasmid names are also shown below in Table 2.

Table 2. DNA sequences of expression vectors

Plasmid name	Sequence of variable 9 bps	SEQ ID NO:	RVD of C-terminal 0.5 TALE repeat
pJDS70	TCTAACATC	45	SNI (for binding to an A nucleotide)
pJDS71	TCCCACGAC	46	SHD (for binding to a C nucleotide)
pJDS74	AATAATAAC	47	NNN (for binding to a G nucleotide)
pJDS76	TCCAATAAA	48	SNK (for binding to a G nucleotide)
pJDS78	TCTAATGGG	49	SNG (for binding to a T nucleotide)

10 This example demonstrates the construction of TALE repeat arrays on an immobilized substrate using preassembled TALE repeat sub-array units. The above method, up to the cloning step, can be performed in one day.

Example 2. Assembly of TALE Repeat Arrays using a streptavidin coated plate

15 TALE repeats are assembled using the archive of DNA plasmids (~850 different plasmids) as described in Example 1. A 16-mer TALE repeat array was created by in vitro assembly of 16 TALE repeats designed to bind a target sequence. In the first step, a plasmid carrying an α -type TALE repeat with an NNN triplet (G) was amplified by PCR using a biotinylated forward primer Biotin-TCTAGAGAAGACAAGAACCTGACC
 20 (SEQ ID NO: 42) and a reverse primer GGATCCGGTCTCTTAAGGCCGTGG (SEQ ID NO: 43). The amplified fragment (50 μ l) was purified using a QIA Quick PCR purification kit (QIAGEN), eluted in 40 μ l 0.1X elution buffer (as provided in the QIA

Quick PCR purification kit), and digested with BsaI HF (New England Biolabs (NEB)) in NEB Buffer 4 for 15 minutes at 50 °C (40 µl elution, 5 µl NEBuffer 4, 5 µl BsaI HF).

The digested fragment was purified using a QIA Quick PCR purification kit and eluted in 0.1X elution buffer (50 µl).

5 A plasmid containing a four TALE repeat domain sub-array unit ($\beta\gamma\delta\epsilon$) coding for repeats that each harbor one of the following variable amino acids SHD, SNI, NNN, and SNG (designed to bind the sequence 5'-CAGT-3') was digested with BbsI (NEB) in NEBuffer 2 for 2 hours at 37 °C in 100 µl (50 µl plasmid [\sim 200 ng/ μ l], 10 µl NEBuffer 2, 10 µl BbsI, 30 µl water). To the 100 µl digest was added 25 µl NEBuffer 4, 2.5 µl 100X
10 BSA (NEB), 107.5 µl water, and 5 µl XbaI (NEB), and the digest was incubated for 5 minutes at 37 °C. To the mixture, 5 µl of BamHI HF was then added for a 5 minute digest at 37 °C, and then 5 µl Sall HF (NEB) was added for an additional 5 minute digest at 37 °C. The resulting fragment was purified using a QIA Quick PCR purification kit (QIAGEN) and eluted in 180 µl 0.1X elution buffer.

15 For the initial ligation, 2 µl of the alpha unit digest was mixed with 2.5 µl of T4 DNA ligase (400 U/ μ l; NEB) and 27 µl Quick Ligase Buffer (QLB) (NEB). To this 31.5 µl mixture was added 22.5 µl of the first digested subarray, and the mixture was ligated for 15 minutes at room temperature. The ligation mixture was then mixed with 2X B&@ buffer (Invitrogen) and added to a well in a 96-well plate coated with
20 streptavidin (Thermo Scientific) and incubated at room temperature for 15min. The supernatant was aspirated. Each well in the 96 well plate was washed with 200ul of 1x Bovine Serum Albumin (BSA) by pipetting up and down 10 times before discarding the 1x BSA. This was repeated for a total of two washes with 1x BSA. Then 50 µl BsaI HF mix (5 µl NEBuffer 4, 2 µl BsaI HF, 43 µl water) was added to the ligated, nucleic acids
25 ($\alpha\beta\gamma\delta\epsilon$) bound to the streptavidin-coated well.

The digest was incubated at 50 °C for 10 minutes and then the supernatant was aspirated. The wells were then washed with 200 µl 1X B&W Buffer and 200 µl 1X BSA twice by pipetting up and down ten times before removal of each supernatant. 22.5 µl of digested plasmid encoding a four TALE repeat domain sub-array unit ($\beta\gamma\delta\epsilon$) coding for
30 repeats that each harbor one of the following variable amino acids NNN, SHD, SNG, and

SNI and 27.5 μ l ligase mix (25 μ l Quick Ligase Buffer, 2 μ l DNA ligase) were added to the well. The supernatant was mixed by pipetting up and down, and the mixture was incubated for 15 minutes at room temperature. The supernatant was removed and the well was washed with 1X B&W and 1x BSA as above. Then 50 μ l BsaI HF mix (5 μ l
5 NEBuffer 4, 2 μ l BsaI HF, 43 μ l water) was added to the ligated nucleic acids ($\alpha\beta\gamma\delta\epsilon\beta\gamma\delta\epsilon$) bound to the well. Two more TALE repeat sub-array units were ligated sequentially as above, the first a four TALE repeat sub-array unit ($\beta\gamma\delta\epsilon$) coding for repeats that each harbor one of the following variable amino acids SHD, SNI, NNN, and SNG and the second a three TALE repeat sub-array unit ($\beta\gamma\delta$) coding for repeats that
10 each harbor one of the following variable amino acids SHD, SNI, NNN, and SHD. The final TALE repeat array contained subunits of the format $\alpha\beta\gamma\delta\epsilon\beta\gamma\delta\epsilon\beta\gamma\delta$ with individual TALE repeats designed to bind a target DNA sequence.

Following the final ligation step, the fragments in the well were digested with BsaI HF for eventual cloning into an expression vector. The well was then washed with
15 1X B&W Buffer and twice with 1X BSA. Then 50 μ l BbsI mix (5 μ l NEBuffer 2, 5 μ l BbsI, 40 μ l water) was added to the well and incubated at 37 °C for 2 hours to cleave the biotinylated 5' end and release the assembled TALE repeat array from the well. The digested mixture was purified, ligated, and transformed as described in Example 1.

20 Example 3. Site-Specific Mutagenesis Using TALE Nucleases

To demonstrate the effectiveness of TALE repeat domains created by the methods described herein, TALE repeat arrays were constructed and cloned into TALE nuclease expression vectors (as described in Example 1) to produce plasmids encoding TALE nuclease monomers targeted to the eGFP coding sequences shown in FIG. 6 and Table 3.
25 Nucleic acid and polypeptide sequences of the TALE nuclease monomers are shown in FIGs. 11A-18B.

Table 3. TALE nuclease monomer target sequences

TALE Fragment	Target Sequence	Length of target sequence	SEQ ID NO:	Site	Position (half-site)	Plasmid name
DR-TALE-0003	TGCAGTGCTTCAGCCGC	17	50	eGFP223	left	SQT70
DR-TALE-0006	TGCAGTGCTTCAGCCGCT	18	51	eGFP223	left	SQT114
DR-TALE-0005	TTGAAGAAGTCGTGCTGC	18	52	eGFP223	right	SQT72
DR-TALE-0010	TGAAGAAGTCGTGCTGCT	18	53	eGFP223	right	SQT56
DR-TALE-0023	TCGAGCTGAAGGGCATC	17	54	eGFP382	left	SQT84
DR-TALE-0025	TCGAGCTGAAGGGCATCG	18	55	eGFP382	left	SQT120
DR-TALE-0020	TTGTGCCCCAGGATGTTG	18	56	eGFP382	right	SQT135
DR-TALE-0022	TGTGCCCCAGGATGTTGC	18	57	eGFP382	right	SQT118

4E5 U2OS-eGFP cells were nucleofected with 400 ng plasmid DNA in solution SE with program DN-100 using Nucleofector™ non-viral transfection (Lonza, Walkersville, MD). The cells were analyzed by flow cytometry at days 2 and 5 (FIG. 7). Non-homologous end joining (NHEJ)-mediated mutagenic repair of TALE nuclease-induced double-stranded breaks led to disruption of eGFP expression (eGFP-negative cells). All eight TALE nuclease pairs tested induced a high percentage of eGFP-negative (eGFP-) cells (y-axis). The percentage of eGFP- cells declined only modestly between day 2 and 5 suggesting that the alterations were stably induced.

A subset of mutated eGFP genes were amplified from cells and sequenced. The resulting mutations are shown in FIG. 8. Sequences targeted by the TALE nucleases encoded by expression plasmids SQT70/SQT56 in human USOS-eGFP cells are

underlined in the wild-type (WT) sequence shown at the top of FIG. 8. Insertion and deletion mutations induced by the TALE nuclease pair are shown below with deleted bases indicated by dashes and inserted bases indicated by double underlining. The net number of bases inserted or deleted is shown to the right. All mutations were isolated once unless otherwise indicated in brackets. The overall frequency of mutagenesis (46%) is also indicated.

Example 4. Automated Assembly of TALE Repeat Arrays

The assembly method described in Example 1 has been automated so as to be performed using a Sciclone™ G3 liquid handling workstation (Caliper Life Sciences, Hopkinton, MA) in 96-well plates. All of the steps were automated except digestion of the nucleic acids prior to ligation and linking to the beads and the steps following release of the assembled TALE repeat array from the magnetic beads. The automated steps were performed essentially as when done manually with minor variations in the number of resuspension and mixing motions. The results of assembly of two 17-mers are shown in FIG. 9. A major product of the expected size can be seen, corresponding to the 17-mer. Additional minor 13-mer, 9-mer, and 5-mer products can also be seen, likely produced by carry forward of incompletely ligated products. A similar result can be seen in FIG. 10, which shows the results of assembly of 16-mers from an N-terminal 1-mer sub-array (1), three 4-mer subarrays (4_A, 4_B, 4_C), and a C-terminal 3-mer subarray (3_D).

This example demonstrates that the methods described herein can be automated for rapid and reproducible synthesis of nucleic acids encoding TALE repeat arrays.

Example 5. Assembly Methods

TALE repeat arrays were created using an architecture in which four distinct TALE repeat backbones that differ slightly in their amino acid and DNA sequences occur in a repeated pattern. The first, amino-terminal TALE repeat in an array was designated as the α unit. This was followed by β , γ , and δ units and then an ϵ unit that is essentially identical to the α unit except for the different positioning of a Type IIS restriction site on the 5' end (required to enable creation of a unique overhang on the α unit needed for

cloning). The ϵ unit was then followed again by repeats of β , γ , δ , and ϵ units. Due to constraints related to creation of a 3' end required for cloning, slightly modified DNA sequences were required for TALE repeat arrays that end with a carboxy-terminal γ or ϵ unit. We designated these variant units as γ^* and ϵ^* .

5 For each type of TALE repeat unit (i.e.— α , β , γ , δ , ϵ , γ^* , and ϵ^*), we commercially synthesized (Genscript) a series of four plasmids, each harboring one of the five repeat variable di-residues (RVDs) that specifies one of the four DNA bases (NI = A; HD = C; NN = G; NG = T, NK = G). Full DNA sequences of these plasmids are provided in Table 4 and FIG. 3. For all 35 of these plasmids, the sequence encoding the TALE repeat domain is flanked on the 5' end by unique XbaI and BbsI restriction sites and on the 3' end by unique BsaI and BamHI restriction sites. Additionally, the overhangs generated by digestion of any plasmids encoding units designed to be adjacent to one another (e.g.— β and γ , or δ and ϵ) with BsaI and BbsI are complementary. Using these 35 different plasmids and serial ligation via the BsaI and BbsI restriction sites, we assembled an archive of all possible combinations of $\beta\gamma$, $\beta\gamma\delta\epsilon$, $\beta\gamma\delta$, $\beta\gamma^*$, and $\delta\epsilon^*$ repeats. In total, this archive consisted of 825 different plasmids encoding 5 α 's, 5 β 's, 25 $\beta\gamma$ combinations, 625 $\beta\gamma\delta\epsilon$ combinations, 125 $\beta\gamma\delta$ combinations, 25 $\beta\gamma^*$ combinations, and 25 $\delta\epsilon^*$ combinations (Table 5). These 825 plasmids plus ten of the original 35 plasmids encoding single TALE repeats (five α and five β plasmids) are required to practice the methods. With this archive of 835 plasmids listed in Table 5, the methods can be used to construct TALE repeat arrays of any desired length and composition.

Table 4. DNA sequences encoding individual TALE repeats

TAL ID#	Unit Architecture	RVD	Target Base	DNA Sequence (Cloned between XbaI/BamHI in pUC57- Δ BsaI)	SEQ ID NO:
6	α	NI	A	TCTAGAGAAGACAAGAACCCTGACC CCAGACCAGGTACTCGCAATCGCG TCGAACATTGGGGGAAAGCAAGCC CTGGAAACCGTGCAAAGGTTGTTG CCGGTCCTTTGTCAAGACCACGGC CTTAAGAGACCGGATCC	58

7	α	HD	C	TCTAGAGAAGACAAGAACCTGACC CCAGACCAGGTAGTCGCAATCGCG TCACATGACGGGGGAAAGCAAGCC CTGGAACCGTGCAAAGGTTGTTG CCGGTCCTTTGTCAAGACCACGGC CTTAAGAGACCGGATCC	59
8	α	NK	G	TCTAGAGAAGACAAGAACCTGACC CCAGACCAGGTAGTCGCAATCGCG TCGAACAAGGGGGAAAGCAAGCC CTGGAACCGTGCAAAGGTTGTTG CCGGTCCTTTGTCAAGACCACGGC CTTAAGAGACCGGATCC	60
9	α	NN	G	TCTAGAGAAGACAAGAACCTGACC CCAGACCAGGTAGTCGCAATCGCG ACAATAATGGGGGAAAGCAAGCC CTGGAACCGTGCAAAGGTTGTTG CCGGTCCTTTGTCAAGACCACGGC CTTAAGAGACCGGATCC	61
10	α	NG	T	TCTAGAGAAGACAAGAACCTGACC CCAGACCAGGTAGTCGCAATCGCG TCAAACGGAGGGGGAAAGCAAGCC CTGGAACCGTGCAAAGGTTGTTG CCGGTCCTTTGTCAAGACCACGGC CTTAAGAGACCGGATCC	62
11	β	NI	A	TCTAGAGAAGACAACCTTACACCGG AGCAAGTCGTGGCCATTGCAAGCA ACATCGGTGGCAAACAGGCTCTTG AGACGGTTCAGAGACTTCTCCAG TTCTCTGTCAAGCCCACGGGCTGA AGAGACCGGATCC	63
12	β	HD	C	TCTAGAGAAGACAACCTTACACCGG AGCAAGTCGTGGCCATTGCATCCC ACGACGGTGGCAAACAGGCTCTTG AGACGGTTCAGAGACTTCTCCAG TTCTCTGTCAAGCCCACGGGCTGA AGAGACCGGATCC	64
13	β	NK	G	TCTAGAGAAGACAACCTTACACCGG AGCAAGTCGTGGCCATTGCATCAA ATAAAGTGGCAAACAGGCTCTTG AGACGGTTCAGAGACTTCTCCAG TTCTCTGTCAAGCCCACGGGCTGA AGAGACCGGATCC	65
14	β	NN	G	TCTAGAGAAGACAACCTTACACCGG AGCAAGTCGTGGCCATTGCAAATA ATAACGGTGGCAAACAGGCTCTTG AGACGGTTCAGAGACTTCTCCAG TTCTCTGTCAAGCCCACGGGCTGA AGAGACCGGATCC	66

15	β	NG	T	TCTAGAGAAGACAACCTTACACCGG AGCAAGTCGTGGCCATTGCAAGCA ATGGGGGTGGCAAACAGGCTCTTG AGACGGTTCAGAGACTTCTCCAG TTCCTGTCAAGCCCACGGGCTGA AGAGACCGGATCC	67
16	γ	NI	A	TCTAGAGAAGACAACCTGACTCCCG ATCAAGTTGTAGCGATTGCGTCGA ACATTGGAGGGAAACAAGCATTGG AGACTGTCCAACGGCTCCTTCCCG TGTTGTGTCAAGCCCACGGTTTGA AGAGACCGGATCC	68
17	γ	HD	C	TCTAGAGAAGACAACCTGACTCCCG ATCAAGTTGTAGCGATTGCGTCGC ATGACGGAGGGAAACAAGCATTGG AGACTGTCCAACGGCTCCTTCCCG TGTTGTGTCAAGCCCACGGTTTGA AGAGACCGGATCC	69
18	γ	NK	G	TCTAGAGAAGACAACCTGACTCCCG ATCAAGTTGTAGCGATTGCGTCCA ACAAGGGAGGGAAACAAGCATTGG AGACTGTCCAACGGCTCCTTCCCG TGTTGTGTCAAGCCCACGGTTTGA AGAGACCGGATCC	70
19	γ	NN	G	TCTAGAGAAGACAACCTGACTCCCG ATCAAGTTGTAGCGATTGCGAATA ACAATGGAGGGAAACAAGCATTGG AGACTGTCCAACGGCTCCTTCCCG TGTTGTGTCAAGCCCACGGTTTGA AGAGACCGGATCC	71
20	γ	NG	T	TCTAGAGAAGACAACCTGACTCCCG ATCAAGTTGTAGCGATTGCGTCCA ACGGTGGAGGGAAACAAGCATTGG AGACTGTCCAACGGCTCCTTCCCG TGTTGTGTCAAGCCCACGGTTTGA AGAGACCGGATCC	72
21	δ	NI	A	TCTAGAGAAGACAATTGACGCCTG CACAAGTGGTCGCCATCGCCTCCA ATATTGGCGGTAAGCAGGCGCTGG AAACAGTACAGCGCCTGCTGCCTG TACTGTGCCAGGATCATGGACTGA AGAGACCGGATCC	73
22	δ	HD	C	TCTAGAGAAGACAATTGACGCCTG CACAAGTGGTCGCCATCGCCAGCC ATGATGGCGGTAAGCAGGCGCTGG AAACAGTACAGCGCCTGCTGCCTG TACTGTGCCAGGATCATGGACTGA AGAGACCGGATCC	74

23	δ	NK	G	TCTAGAGAAGACAATTGACGCCTG CACAAGTGGTCGCCATCGCCAGCA ATAAGGGCGGTAAGCAGGCGCTGG AAACAGTACAGCGCCTGCTGCCTG TACTGTGCCAGGATCATGGACTGA AGAGACCGGATCC	75
24	δ	NN	G	TCTAGAGAAGACAATTGACGCCTG CACAAGTGGTCGCCATCGCCAACA ACAACGGCGGTAAGCAGGCGCTGG AAACAGTACAGCGCCTGCTGCCTG TACTGTGCCAGGATCATGGACTGA AGAGACCGGATCC	76
25	δ	NG	T	TCTAGAGAAGACAATTGACGCCTG CACAAGTGGTCGCCATCGCCTCGA ATGGCGGCGGTAAGCAGGCGCTGG AAACAGTACAGCGCCTGCTGCCTG TACTGTGCCAGGATCATGGACTGA AGAGACCGGATCC	77
26	ϵ	NI	A	TCTAGAGAAGACAAGTACCCAG ACCAGGTAGTCGCAATCGCGTCGA ACATTGGGGGAAAGCAAGCCCTGG AAACCGTGCAAAGGTTGTTGCCGG TCCTTTGTCAAGACCACGGCCTTA AGAGACCGGATCC	78
27	ϵ	HD	C	TCTAGAGAAGACAAGTACCCAG ACCAGGTAGTCGCAATCGCGTCAC ATGACGGGGGAAAGCAAGCCCTGG AAACCGTGCAAAGGTTGTTGCCGG TCCTTTGTCAAGACCACGGCCTTA AGAGACCGGATCC	79
28	ϵ	NK	G	TCTAGAGAAGACAAGTACCCAG ACCAGGTAGTCGCAATCGCGTCGA ACAAAGGGGGAAAGCAAGCCCTGG AAACCGTGCAAAGGTTGTTGCCGG TCCTTTGTCAAGACCACGGCCTTA AGAGACCGGATCC	80
29	ϵ	NN	G	TCTAGAGAAGACAAGTACCCAG ACCAGGTAGTCGCAATCGCGAACA ATAATGGGGGAAAGCAAGCCCTGG AAACCGTGCAAAGGTTGTTGCCGG TCCTTTGTCAAGACCACGGCCTTA AGAGACCGGATCC	81
30	ϵ	NG	T	TCTAGAGAAGACAAGTACCCAG ACCAGGTAGTCGCAATCGCGTCAA ACGGAGGGGGAAAGCAAGCCCTGG AAACCGTGCAAAGGTTGTTGCCGG TCCTTTGTCAAGACCACGGCCTTA AGAGACCGGATCC	82

31	γ'	NI	A	TCTAGAGAAGACAACACTGACTCCCG ATCAAGTTGTAGCGATTGCGTCGA ACATTGGAGGGAAACAAGCATTGG AGACTGTCCAACGGCTCCTTCCCG TGTTGTGTCAAGCCCACGGTCTGA AGAGACCGGATCC	83
32	γ'	HD	C	TCTAGAGAAGACAACACTGACTCCCG ATCAAGTTGTAGCGATTGCGTCGC ATGACGGAGGGAAACAAGCATTGG AGACTGTCCAACGGCTCCTTCCCG TGTTGTGTCAAGCCCACGGTCTGA AGAGACCGGATCC	84
33	γ'	NK	G	TCTAGAGAAGACAACACTGACTCCCG ATCAAGTTGTAGCGATTGCGTCCA ACAAGGGAGGGAAACAAGCATTGG AGACTGTCCAACGGCTCCTTCCCG TGTTGTGTCAAGCCCACGGTCTGA AGAGACCGGATCC	85
34	γ'	NN	G	TCTAGAGAAGACAACACTGACTCCCG ATCAAGTTGTAGCGATTGCGAATA ACAATGGAGGGAAACAAGCATTGG AGACTGTCCAACGGCTCCTTCCCG TGTTGTGTCAAGCCCACGGTCTGA AGAGACCGGATCC	86
35	γ'	NG	T	TCTAGAGAAGACAACACTGACTCCCG ATCAAGTTGTAGCGATTGCGTCCA ACGGTGGAGGGAAACAAGCATTGG AGACTGTCCAACGGCTCCTTCCCG TGTTGTGTCAAGCCCACGGTCTGA AGAGACCGGATCC	87
36	ϵ'	NI	A	TCTAGAGAAGACAACACTGACCCAG ACCAGGTAGTCGCAATCGCGTCGA ACATTGGGGAAAGCAAGCCCTGG AAACCGTGCAAAGGTTGTTGCCGG TCCTTTGTCAAGACCACGGCTGA AGAGACCGGATCC	88
37	ϵ'	HD	C	TCTAGAGAAGACAACACTGACCCAG ACCAGGTAGTCGCAATCGCGTCAC ATGACGGGGAAAGCAAGCCCTGG AAACCGTGCAAAGGTTGTTGCCGG TCCTTTGTCAAGACCACGGCTGA AGAGACCGGATCC	89
38	ϵ'	NK	G	TCTAGAGAAGACAACACTGACCCAG ACCAGGTAGTCGCAATCGCGTCGA ACAAAGGGGGAAAGCAAGCCCTGG AAACCGTGCAAAGGTTGTTGCCGG TCCTTTGTCAAGACCACGGCTGA AGAGACCGGATCC	90

39	ϵ'	NN	G	TCTAGAGAAGACAACACTGACCCCAG ACCAGGTAGTCGCAATCGCGAACA ATAATGGGGGAAAGCAAGCCCTGG AAACCGTGCAAAGGTTGTTGCCGG TCCTTTGTCAAGACCACGGCCTGA AGAGACCGGATCC	91
40	ϵ'	NG	T	TCTAGAGAAGACAACACTGACCCCAG ACCAGGTAGTCGCAATCGCGTCAA ACGGAGGGGGAAAGCAAGCCCTGG AAACCGTGCAAAGGTTGTTGCCGG TCCTTTGTCAAGACCACGGCCTGA AGAGACCGGATCC	92

Table 5. Archive of 835 plasmids encoding pre-assembled TALE repeat units

Plasmid ID	DNA Target	RVDs	Unit Architecture
TAL006	A	NI	α
TAL007	C	HD	α
TAL008	G	NK	α
TAL009	G	NN	α
TAL010	T	NG	α
TAL011/016/021/026	AAAA	NI/NI/NI/NI	$\beta\gamma\delta\epsilon$
TAL011/016/021/027	AAAC	NI/NI/NI/HD	$\beta\gamma\delta\epsilon$
TAL011/016/021/028	AAAG	NI/NI/NI/NK	$\beta\gamma\delta\epsilon$
TAL011/016/021/029	AAAG	NI/NI/NI/NN	$\beta\gamma\delta\epsilon$
TAL011/016/021/030	AAAT	NI/NI/NI/NG	$\beta\gamma\delta\epsilon$
TAL011/016/022/026	AACA	NI/NI/HD/NI	$\beta\gamma\delta\epsilon$
TAL011/016/022/027	AACC	NI/NI/HD/HD	$\beta\gamma\delta\epsilon$
TAL011/016/022/028	AACG	NI/NI/HD/NK	$\beta\gamma\delta\epsilon$
TAL011/016/022/029	AACG	NI/NI/HD/NN	$\beta\gamma\delta\epsilon$
TAL011/016/022/030	AACT	NI/NI/HD/NG	$\beta\gamma\delta\epsilon$
TAL011/016/023/026	AAGA	NI/NI/NK/NI	$\beta\gamma\delta\epsilon$
TAL011/016/023/027	AAGC	NI/NI/NK/HD	$\beta\gamma\delta\epsilon$
TAL011/016/023/028	AAGG	NI/NI/NK/NK	$\beta\gamma\delta\epsilon$
TAL011/016/023/029	AAGG	NI/NI/NK/NN	$\beta\gamma\delta\epsilon$
TAL011/016/023/030	AAGT	NI/NI/NK/NG	$\beta\gamma\delta\epsilon$
TAL011/016/024/026	AAGA	NI/NI/NN/NI	$\beta\gamma\delta\epsilon$
TAL011/016/024/027	AAGC	NI/NI/NN/HD	$\beta\gamma\delta\epsilon$
TAL011/016/024/028	AAGG	NI/NI/NN/NK	$\beta\gamma\delta\epsilon$
TAL011/016/024/029	AAGG	NI/NI/NN/NN	$\beta\gamma\delta\epsilon$
TAL011/016/024/030	AAGT	NI/NI/NN/NG	$\beta\gamma\delta\epsilon$
TAL011/016/025/026	AATA	NI/NI/NG/NI	$\beta\gamma\delta\epsilon$

TAL011/016/025/027	AATC	NI/NI/NG/HD	βγδε
TAL011/016/025/028	AATG	NI/NI/NG/NK	βγδε
TAL011/016/025/029	AATG	NI/NI/NG/NN	βγδε
TAL011/016/025/030	AATT	NI/NI/NG/NG	βγδε
TAL011/017/021/026	ACAA	NI/HD/NI/NI	βγδε
TAL011/017/021/027	ACAC	NI/HD/NI/HD	βγδε
TAL011/017/021/028	ACAG	NI/HD/NI/NK	βγδε
TAL011/017/021/029	ACAG	NI/HD/NI/NN	βγδε
TAL011/017/021/030	ACAT	NI/HD/NI/NG	βγδε
TAL011/017/022/026	ACCA	NI/HD/HD/NI	βγδε
TAL011/017/022/027	ACCC	NI/HD/HD/HD	βγδε
TAL011/017/022/028	ACCG	NI/HD/HD/NK	βγδε
TAL011/017/022/029	ACCG	NI/HD/HD/NN	βγδε
TAL011/017/022/030	ACCT	NI/HD/HD/NG	βγδε
TAL011/017/023/026	ACGA	NI/HD/NK/NI	βγδε
TAL011/017/023/027	ACGC	NI/HD/NK/HD	βγδε
TAL011/017/023/028	ACGG	NI/HD/NK/NK	βγδε
TAL011/017/023/029	ACGG	NI/HD/NK/NN	βγδε
TAL011/017/023/030	ACGT	NI/HD/NK/NG	βγδε
TAL011/017/024/026	ACGA	NI/HD/NN/NI	βγδε
TAL011/017/024/027	ACGC	NI/HD/NN/HD	βγδε
TAL011/017/024/028	ACGG	NI/HD/NN/NK	βγδε
TAL011/017/024/029	ACGG	NI/HD/NN/NN	βγδε
TAL011/017/024/030	ACGT	NI/HD/NN/NG	βγδε
TAL011/017/025/026	ACTA	NI/HD/NG/NI	βγδε
TAL011/017/025/027	ACTC	NI/HD/NG/HD	βγδε
TAL011/017/025/028	ACTG	NI/HD/NG/NK	βγδε
TAL011/017/025/029	ACTG	NI/HD/NG/NN	βγδε
TAL011/017/025/030	ACTT	NI/HD/NG/NG	βγδε
TAL011/018/021/026	AGAA	NI/NK/NI/NI	βγδε
TAL011/018/021/027	AGAC	NI/NK/NI/HD	βγδε
TAL011/018/021/028	AGAG	NI/NK/NI/NK	βγδε
TAL011/018/021/029	AGAG	NI/NK/NI/NN	βγδε
TAL011/018/021/030	AGAT	NI/NK/NI/NG	βγδε
TAL011/018/022/026	AGCA	NI/NK/HD/NI	βγδε
TAL011/018/022/027	AGCC	NI/NK/HD/HD	βγδε
TAL011/018/022/028	AGCG	NI/NK/HD/NK	βγδε
TAL011/018/022/029	AGCG	NI/NK/HD/NN	βγδε
TAL011/018/022/030	AGCT	NI/NK/HD/NG	βγδε
TAL011/018/023/026	AGGA	NI/NK/NK/NI	βγδε
TAL011/018/023/027	AGGC	NI/NK/NK/HD	βγδε

TAL011/018/023/028	AGGG	NI/NK/NK/NK	βγδε
TAL011/018/023/029	AGGG	NI/NK/NK/NN	βγδε
TAL011/018/023/030	AGGT	NI/NK/NK/NG	βγδε
TAL011/018/024/026	AGGA	NI/NK/NN/NI	βγδε
TAL011/018/024/027	AGGC	NI/NK/NN/HD	βγδε
TAL011/018/024/028	AGGG	NI/NK/NN/NK	βγδε
TAL011/018/024/029	AGGG	NI/NK/NN/NN	βγδε
TAL011/018/024/030	AGGT	NI/NK/NN/NG	βγδε
TAL011/018/025/026	AGTA	NI/NK/NG/NI	βγδε
TAL011/018/025/027	AGTC	NI/NK/NG/HD	βγδε
TAL011/018/025/028	AGTG	NI/NK/NG/NK	βγδε
TAL011/018/025/029	AGTG	NI/NK/NG/NN	βγδε
TAL011/018/025/030	AGTT	NI/NK/NG/NG	βγδε
TAL011/019/021/026	AGAA	NI/NN/NI/NI	βγδε
TAL011/019/021/027	AGAC	NI/NN/NI/HD	βγδε
TAL011/019/021/028	AGAG	NI/NN/NI/NK	βγδε
TAL011/019/021/029	AGAG	NI/NN/NI/NN	βγδε
TAL011/019/021/030	AGAT	NI/NN/NI/NG	βγδε
TAL011/019/022/026	AGCA	NI/NN/HD/NI	βγδε
TAL011/019/022/027	AGCC	NI/NN/HD/HD	βγδε
TAL011/019/022/028	AGCG	NI/NN/HD/NK	βγδε
TAL011/019/022/029	AGCG	NI/NN/HD/NN	βγδε
TAL011/019/022/030	AGCT	NI/NN/HD/NG	βγδε
TAL011/019/023/026	AGGA	NI/NN/NK/NI	βγδε
TAL011/019/023/027	AGGC	NI/NN/NK/HD	βγδε
TAL011/019/023/028	AGGG	NI/NN/NK/NK	βγδε
TAL011/019/023/029	AGGG	NI/NN/NK/NN	βγδε
TAL011/019/023/030	AGGT	NI/NN/NK/NG	βγδε
TAL011/019/024/026	AGGA	NI/NN/NN/NI	βγδε
TAL011/019/024/027	AGGC	NI/NN/NN/HD	βγδε
TAL011/019/024/028	AGGG	NI/NN/NN/NK	βγδε
TAL011/019/024/029	AGGG	NI/NN/NN/NN	βγδε
TAL011/019/024/030	AGGT	NI/NN/NN/NG	βγδε
TAL011/019/025/026	AGTA	NI/NN/NG/NI	βγδε
TAL011/019/025/027	AGTC	NI/NN/NG/HD	βγδε
TAL011/019/025/028	AGTG	NI/NN/NG/NK	βγδε
TAL011/019/025/029	AGTG	NI/NN/NG/NN	βγδε
TAL011/019/025/030	AGTT	NI/NN/NG/NG	βγδε
TAL011/020/021/026	ATAA	NI/NG/NI/NI	βγδε
TAL011/020/021/027	ATAC	NI/NG/NI/HD	βγδε
TAL011/020/021/028	ATAG	NI/NG/NI/NK	βγδε

TAL011/020/021/029	ATAG	NI/NG/NI/NN	βγδε
TAL011/020/021/030	ATAT	NI/NG/NI/NG	βγδε
TAL011/020/022/026	ATCA	NI/NG/HD/NI	βγδε
TAL011/020/022/027	ATCC	NI/NG/HD/HD	βγδε
TAL011/020/022/028	ATCG	NI/NG/HD/NK	βγδε
TAL011/020/022/029	ATCG	NI/NG/HD/NN	βγδε
TAL011/020/022/030	ATCT	NI/NG/HD/NG	βγδε
TAL011/020/023/026	ATGA	NI/NG/NK/NI	βγδε
TAL011/020/023/027	ATGC	NI/NG/NK/HD	βγδε
TAL011/020/023/028	ATGG	NI/NG/NK/NK	βγδε
TAL011/020/023/029	ATGG	NI/NG/NK/NN	βγδε
TAL011/020/023/030	ATGT	NI/NG/NK/NG	βγδε
TAL011/020/024/026	ATGA	NI/NG/NN/NI	βγδε
TAL011/020/024/027	ATGC	NI/NG/NN/HD	βγδε
TAL011/020/024/028	ATGG	NI/NG/NN/NK	βγδε
TAL011/020/024/029	ATGG	NI/NG/NN/NN	βγδε
TAL011/020/024/030	ATGT	NI/NG/NN/NG	βγδε
TAL011/020/025/026	ATTA	NI/NG/NG/NI	βγδε
TAL011/020/025/027	ATTC	NI/NG/NG/HD	βγδε
TAL011/020/025/028	ATTG	NI/NG/NG/NK	βγδε
TAL011/020/025/029	ATTG	NI/NG/NG/NN	βγδε
TAL011/020/025/030	ATTT	NI/NG/NG/NG	βγδε
TAL012/016/021/026	CAAA	HD/NI/NI/NI	βγδε
TAL012/016/021/027	CAAC	HD/NI/NI/HD	βγδε
TAL012/016/021/028	CAAG	HD/NI/NI/NK	βγδε
TAL012/016/021/029	CAAG	HD/NI/NI/NN	βγδε
TAL012/016/021/030	CAAT	HD/NI/NI/NG	βγδε
TAL012/016/022/026	CACA	HD/NI/HD/NI	βγδε
TAL012/016/022/027	CACC	HD/NI/HD/HD	βγδε
TAL012/016/022/028	CACG	HD/NI/HD/NK	βγδε
TAL012/016/022/029	CACG	HD/NI/HD/NN	βγδε
TAL012/016/022/030	CACT	HD/NI/HD/NG	βγδε
TAL012/016/023/026	CAGA	HD/NI/NK/NI	βγδε
TAL012/016/023/027	CAGC	HD/NI/NK/HD	βγδε
TAL012/016/023/028	CAGG	HD/NI/NK/NK	βγδε
TAL012/016/023/029	CAGG	HD/NI/NK/NN	βγδε
TAL012/016/023/030	CAGT	HD/NI/NK/NG	βγδε
TAL012/016/024/026	CAGA	HD/NI/NN/NI	βγδε
TAL012/016/024/027	CAGC	HD/NI/NN/HD	βγδε
TAL012/016/024/028	CAGG	HD/NI/NN/NK	βγδε
TAL012/016/024/029	CAGG	HD/NI/NN/NN	βγδε

TAL012/016/024/030	CAGT	HD/NI/NN/NG	βγδε
TAL012/016/025/026	CATA	HD/NI/NG/NI	βγδε
TAL012/016/025/027	CATC	HD/NI/NG/HD	βγδε
TAL012/016/025/028	CATG	HD/NI/NG/NK	βγδε
TAL012/016/025/029	CATG	HD/NI/NG/NN	βγδε
TAL012/016/025/030	CATT	HD/NI/NG/NG	βγδε
TAL012/017/021/026	CCAA	HD/HD/NI/NI	βγδε
TAL012/017/021/027	CCAC	HD/HD/NI/HD	βγδε
TAL012/017/021/028	CCAG	HD/HD/NI/NK	βγδε
TAL012/017/021/029	CCAG	HD/HD/NI/NN	βγδε
TAL012/017/021/030	CCAT	HD/HD/NI/NG	βγδε
TAL012/017/022/026	CCCA	HD/HD/HD/NI	βγδε
TAL012/017/022/027	CCCC	HD/HD/HD/HD	βγδε
TAL012/017/022/028	CCCG	HD/HD/HD/NK	βγδε
TAL012/017/022/029	CCCG	HD/HD/HD/NN	βγδε
TAL012/017/022/030	CCCT	HD/HD/HD/NG	βγδε
TAL012/017/023/026	CCGA	HD/HD/NK/NI	βγδε
TAL012/017/023/027	CCGC	HD/HD/NK/HD	βγδε
TAL012/017/023/028	CCGG	HD/HD/NK/NK	βγδε
TAL012/017/023/029	CCGG	HD/HD/NK/NN	βγδε
TAL012/017/023/030	CCGT	HD/HD/NK/NG	βγδε
TAL012/017/024/026	CCGA	HD/HD/NN/NI	βγδε
TAL012/017/024/027	CCGC	HD/HD/NN/HD	βγδε
TAL012/017/024/028	CCGG	HD/HD/NN/NK	βγδε
TAL012/017/024/029	CCGG	HD/HD/NN/NN	βγδε
TAL012/017/024/030	CCGT	HD/HD/NN/NG	βγδε
TAL012/017/025/026	CCTA	HD/HD/NG/NI	βγδε
TAL012/017/025/027	CCTC	HD/HD/NG/HD	βγδε
TAL012/017/025/028	CCTG	HD/HD/NG/NK	βγδε
TAL012/017/025/029	CCTG	HD/HD/NG/NN	βγδε
TAL012/017/025/030	CCTT	HD/HD/NG/NG	βγδε
TAL012/018/021/026	CGAA	HD/NK/NI/NI	βγδε
TAL012/018/021/027	CGAC	HD/NK/NI/HD	βγδε
TAL012/018/021/028	CGAG	HD/NK/NI/NK	βγδε
TAL012/018/021/029	CGAG	HD/NK/NI/NN	βγδε
TAL012/018/021/030	CGAT	HD/NK/NI/NG	βγδε
TAL012/018/022/026	CGCA	HD/NK/HD/NI	βγδε
TAL012/018/022/027	CGCC	HD/NK/HD/HD	βγδε
TAL012/018/022/028	CGCG	HD/NK/HD/NK	βγδε
TAL012/018/022/029	CGCG	HD/NK/HD/NN	βγδε
TAL012/018/022/030	CGCT	HD/NK/HD/NG	βγδε

TAL012/018/023/026	CGGA	HD/NK/NK/NI	βγδε
TAL012/018/023/027	CGGC	HD/NK/NK/HD	βγδε
TAL012/018/023/028	CGGG	HD/NK/NK/NK	βγδε
TAL012/018/023/029	CGGG	HD/NK/NK/NN	βγδε
TAL012/018/023/030	CGGT	HD/NK/NK/NG	βγδε
TAL012/018/024/026	CGGA	HD/NK/NN/NI	βγδε
TAL012/018/024/027	CGGC	HD/NK/NN/HD	βγδε
TAL012/018/024/028	CGGG	HD/NK/NN/NK	βγδε
TAL012/018/024/029	CGGG	HD/NK/NN/NN	βγδε
TAL012/018/024/030	CGGT	HD/NK/NN/NG	βγδε
TAL012/018/025/026	CGTA	HD/NK/NG/NI	βγδε
TAL012/018/025/027	CGTC	HD/NK/NG/HD	βγδε
TAL012/018/025/028	CGTG	HD/NK/NG/NK	βγδε
TAL012/018/025/029	CGTG	HD/NK/NG/NN	βγδε
TAL012/018/025/030	CGTT	HD/NK/NG/NG	βγδε
TAL012/019/021/026	CGAA	HD/NN/NI/NI	βγδε
TAL012/019/021/027	CGAC	HD/NN/NI/HD	βγδε
TAL012/019/021/028	CGAG	HD/NN/NI/NK	βγδε
TAL012/019/021/029	CGAG	HD/NN/NI/NN	βγδε
TAL012/019/021/030	CGAT	HD/NN/NI/NG	βγδε
TAL012/019/022/026	CGCA	HD/NN/HD/NI	βγδε
TAL012/019/022/027	CGCC	HD/NN/HD/HD	βγδε
TAL012/019/022/028	CGCG	HD/NN/HD/NK	βγδε
TAL012/019/022/029	CGCG	HD/NN/HD/NN	βγδε
TAL012/019/022/030	CGCT	HD/NN/HD/NG	βγδε
TAL012/019/023/026	CGGA	HD/NN/NK/NI	βγδε
TAL012/019/023/027	CGGC	HD/NN/NK/HD	βγδε
TAL012/019/023/028	CGGG	HD/NN/NK/NK	βγδε
TAL012/019/023/029	CGGG	HD/NN/NK/NN	βγδε
TAL012/019/023/030	CGGT	HD/NN/NK/NG	βγδε
TAL012/019/024/026	CGGA	HD/NN/NN/NI	βγδε
TAL012/019/024/027	CGGC	HD/NN/NN/HD	βγδε
TAL012/019/024/028	CGGG	HD/NN/NN/NK	βγδε
TAL012/019/024/029	CGGG	HD/NN/NN/NN	βγδε
TAL012/019/024/030	CGGT	HD/NN/NN/NG	βγδε
TAL012/019/025/026	CGTA	HD/NN/NG/NI	βγδε
TAL012/019/025/027	CGTC	HD/NN/NG/HD	βγδε
TAL012/019/025/028	CGTG	HD/NN/NG/NK	βγδε
TAL012/019/025/029	CGTG	HD/NN/NG/NN	βγδε
TAL012/019/025/030	CGTT	HD/NN/NG/NG	βγδε
TAL012/020/021/026	CTAA	HD/NG/NI/NI	βγδε

TAL012/020/021/027	CTAC	HD/NG/NI/HD	βγδε
TAL012/020/021/028	CTAG	HD/NG/NI/NK	βγδε
TAL012/020/021/029	CTAG	HD/NG/NI/NN	βγδε
TAL012/020/021/030	CTAT	HD/NG/NI/NG	βγδε
TAL012/020/022/026	CTCA	HD/NG/HD/NI	βγδε
TAL012/020/022/027	CTCC	HD/NG/HD/HD	βγδε
TAL012/020/022/028	CTCG	HD/NG/HD/NK	βγδε
TAL012/020/022/029	CTCG	HD/NG/HD/NN	βγδε
TAL012/020/022/030	CTCT	HD/NG/HD/NG	βγδε
TAL012/020/023/026	CTGA	HD/NG/NK/NI	βγδε
TAL012/020/023/027	CTGC	HD/NG/NK/HD	βγδε
TAL012/020/023/028	CTGG	HD/NG/NK/NK	βγδε
TAL012/020/023/029	CTGG	HD/NG/NK/NN	βγδε
TAL012/020/023/030	CTGT	HD/NG/NK/NG	βγδε
TAL012/020/024/026	CTGA	HD/NG/NN/NI	βγδε
TAL012/020/024/027	CTGC	HD/NG/NN/HD	βγδε
TAL012/020/024/028	CTGG	HD/NG/NN/NK	βγδε
TAL012/020/024/029	CTGG	HD/NG/NN/NN	βγδε
TAL012/020/024/030	CTGT	HD/NG/NN/NG	βγδε
TAL012/020/025/026	CTTA	HD/NG/NG/NI	βγδε
TAL012/020/025/027	CTTC	HD/NG/NG/HD	βγδε
TAL012/020/025/028	CTTG	HD/NG/NG/NK	βγδε
TAL012/020/025/029	CTTG	HD/NG/NG/NN	βγδε
TAL012/020/025/030	CTTT	HD/NG/NG/NG	βγδε
TAL013/016/021/026	GAAA	NK/NI/NI/NI	βγδε
TAL013/016/021/027	GAAC	NK/NI/NI/HD	βγδε
TAL013/016/021/028	GAAG	NK/NI/NI/NK	βγδε
TAL013/016/021/029	GAAG	NK/NI/NI/NN	βγδε
TAL013/016/021/030	GAAT	NK/NI/NI/NG	βγδε
TAL013/016/022/026	GACA	NK/NI/HD/NI	βγδε
TAL013/016/022/027	GACC	NK/NI/HD/HD	βγδε
TAL013/016/022/028	GACG	NK/NI/HD/NK	βγδε
TAL013/016/022/029	GACG	NK/NI/HD/NN	βγδε
TAL013/016/022/030	GACT	NK/NI/HD/NG	βγδε
TAL013/016/023/026	GAGA	NK/NI/NK/NI	βγδε
TAL013/016/023/027	GAGC	NK/NI/NK/HD	βγδε
TAL013/016/023/028	GAGG	NK/NI/NK/NK	βγδε
TAL013/016/023/029	GAGG	NK/NI/NK/NN	βγδε
TAL013/016/023/030	GAGT	NK/NI/NK/NG	βγδε
TAL013/016/024/026	GAGA	NK/NI/NN/NI	βγδε
TAL013/016/024/027	GAGC	NK/NI/NN/HD	βγδε

TAL013/016/024/028	GAGG	NK/NI/NN/NK	$\beta\gamma\delta\epsilon$
TAL013/016/024/029	GAGG	NK/NI/NN/NN	$\beta\gamma\delta\epsilon$
TAL013/016/024/030	GAGT	NK/NI/NN/NG	$\beta\gamma\delta\epsilon$
TAL013/016/025/026	GATA	NK/NI/NG/NI	$\beta\gamma\delta\epsilon$
TAL013/016/025/027	GATC	NK/NI/NG/HD	$\beta\gamma\delta\epsilon$
TAL013/016/025/028	GATG	NK/NI/NG/NK	$\beta\gamma\delta\epsilon$
TAL013/016/025/029	GATG	NK/NI/NG/NN	$\beta\gamma\delta\epsilon$
TAL013/016/025/030	GATT	NK/NI/NG/NG	$\beta\gamma\delta\epsilon$
TAL013/017/021/026	GCAA	NK/HD/NI/NI	$\beta\gamma\delta\epsilon$
TAL013/017/021/027	GCAC	NK/HD/NI/HD	$\beta\gamma\delta\epsilon$
TAL013/017/021/028	GCAG	NK/HD/NI/NK	$\beta\gamma\delta\epsilon$
TAL013/017/021/029	GCAG	NK/HD/NI/NN	$\beta\gamma\delta\epsilon$
TAL013/017/021/030	GCAT	NK/HD/NI/NG	$\beta\gamma\delta\epsilon$
TAL013/017/022/026	GCCA	NK/HD/HD/NI	$\beta\gamma\delta\epsilon$
TAL013/017/022/027	GCCC	NK/HD/HD/HD	$\beta\gamma\delta\epsilon$
TAL013/017/022/028	GCCG	NK/HD/HD/NK	$\beta\gamma\delta\epsilon$
TAL013/017/022/029	GCCG	NK/HD/HD/NN	$\beta\gamma\delta\epsilon$
TAL013/017/022/030	GCCT	NK/HD/HD/NG	$\beta\gamma\delta\epsilon$
TAL013/017/023/026	GCGA	NK/HD/NK/NI	$\beta\gamma\delta\epsilon$
TAL013/017/023/027	GCGC	NK/HD/NK/HD	$\beta\gamma\delta\epsilon$
TAL013/017/023/028	GCGG	NK/HD/NK/NK	$\beta\gamma\delta\epsilon$
TAL013/017/023/029	GCGG	NK/HD/NK/NN	$\beta\gamma\delta\epsilon$
TAL013/017/023/030	GCGT	NK/HD/NK/NG	$\beta\gamma\delta\epsilon$
TAL013/017/024/026	GCGA	NK/HD/NN/NI	$\beta\gamma\delta\epsilon$
TAL013/017/024/027	GCGC	NK/HD/NN/HD	$\beta\gamma\delta\epsilon$
TAL013/017/024/028	GCGG	NK/HD/NN/NK	$\beta\gamma\delta\epsilon$
TAL013/017/024/029	GCGG	NK/HD/NN/NN	$\beta\gamma\delta\epsilon$
TAL013/017/024/030	GCGT	NK/HD/NN/NG	$\beta\gamma\delta\epsilon$
TAL013/017/025/026	GCTA	NK/HD/NG/NI	$\beta\gamma\delta\epsilon$
TAL013/017/025/027	GCTC	NK/HD/NG/HD	$\beta\gamma\delta\epsilon$
TAL013/017/025/028	GCTG	NK/HD/NG/NK	$\beta\gamma\delta\epsilon$
TAL013/017/025/029	GCTG	NK/HD/NG/NN	$\beta\gamma\delta\epsilon$
TAL013/017/025/030	GCTT	NK/HD/NG/NG	$\beta\gamma\delta\epsilon$
TAL013/018/021/026	GGAA	NK/NK/NI/NI	$\beta\gamma\delta\epsilon$
TAL013/018/021/027	GGAC	NK/NK/NI/HD	$\beta\gamma\delta\epsilon$
TAL013/018/021/028	GGAG	NK/NK/NI/NK	$\beta\gamma\delta\epsilon$
TAL013/018/021/029	GGAG	NK/NK/NI/NN	$\beta\gamma\delta\epsilon$
TAL013/018/021/030	GGAT	NK/NK/NI/NG	$\beta\gamma\delta\epsilon$
TAL013/018/022/026	GGCA	NK/NK/HD/NI	$\beta\gamma\delta\epsilon$
TAL013/018/022/027	GGCC	NK/NK/HD/HD	$\beta\gamma\delta\epsilon$
TAL013/018/022/028	GGCG	NK/NK/HD/NK	$\beta\gamma\delta\epsilon$

TAL013/018/022/029	GGCG	NK/NK/HD/NN	βγδε
TAL013/018/022/030	GGCT	NK/NK/HD/NG	βγδε
TAL013/018/023/026	GGGA	NK/NK/NK/NI	βγδε
TAL013/018/023/027	GGGC	NK/NK/NK/HD	βγδε
TAL013/018/023/028	GGGG	NK/NK/NK/NK	βγδε
TAL013/018/023/029	GGGG	NK/NK/NK/NN	βγδε
TAL013/018/023/030	GGGT	NK/NK/NK/NG	βγδε
TAL013/018/024/026	GGGA	NK/NK/NN/NI	βγδε
TAL013/018/024/027	GGGC	NK/NK/NN/HD	βγδε
TAL013/018/024/028	GGGG	NK/NK/NN/NK	βγδε
TAL013/018/024/029	GGGG	NK/NK/NN/NN	βγδε
TAL013/018/024/030	GGGT	NK/NK/NN/NG	βγδε
TAL013/018/025/026	GGTA	NK/NK/NG/NI	βγδε
TAL013/018/025/027	GGTC	NK/NK/NG/HD	βγδε
TAL013/018/025/028	GGTG	NK/NK/NG/NK	βγδε
TAL013/018/025/029	GGTG	NK/NK/NG/NN	βγδε
TAL013/018/025/030	GGTT	NK/NK/NG/NG	βγδε
TAL013/019/021/026	GGAA	NK/NN/NI/NI	βγδε
TAL013/019/021/027	GGAC	NK/NN/NI/HD	βγδε
TAL013/019/021/028	GGAG	NK/NN/NI/NK	βγδε
TAL013/019/021/029	GGAG	NK/NN/NI/NN	βγδε
TAL013/019/021/030	GGAT	NK/NN/NI/NG	βγδε
TAL013/019/022/026	GGCA	NK/NN/HD/NI	βγδε
TAL013/019/022/027	GGCC	NK/NN/HD/HD	βγδε
TAL013/019/022/028	GGCG	NK/NN/HD/NK	βγδε
TAL013/019/022/029	GGCG	NK/NN/HD/NN	βγδε
TAL013/019/022/030	GGCT	NK/NN/HD/NG	βγδε
TAL013/019/023/026	GGGA	NK/NN/NK/NI	βγδε
TAL013/019/023/027	GGGC	NK/NN/NK/HD	βγδε
TAL013/019/023/028	GGGG	NK/NN/NK/NK	βγδε
TAL013/019/023/029	GGGG	NK/NN/NK/NN	βγδε
TAL013/019/023/030	GGGT	NK/NN/NK/NG	βγδε
TAL013/019/024/026	GGGA	NK/NN/NN/NI	βγδε
TAL013/019/024/027	GGGC	NK/NN/NN/HD	βγδε
TAL013/019/024/028	GGGG	NK/NN/NN/NK	βγδε
TAL013/019/024/029	GGGG	NK/NN/NN/NN	βγδε
TAL013/019/024/030	GGGT	NK/NN/NN/NG	βγδε
TAL013/019/025/026	GGTA	NK/NN/NG/NI	βγδε
TAL013/019/025/027	GGTC	NK/NN/NG/HD	βγδε
TAL013/019/025/028	GGTG	NK/NN/NG/NK	βγδε
TAL013/019/025/029	GGTG	NK/NN/NG/NN	βγδε

TAL013/019/025/030	GGTT	NK/NN/NG/NG	βγδε
TAL013/020/021/026	GTAA	NK/NG/NI/NI	βγδε
TAL013/020/021/027	GTAC	NK/NG/NI/HD	βγδε
TAL013/020/021/028	GTAG	NK/NG/NI/NK	βγδε
TAL013/020/021/029	GTAG	NK/NG/NI/NN	βγδε
TAL013/020/021/030	GTAT	NK/NG/NI/NG	βγδε
TAL013/020/022/026	GTCA	NK/NG/HD/NI	βγδε
TAL013/020/022/027	GTCC	NK/NG/HD/HD	βγδε
TAL013/020/022/028	GTCG	NK/NG/HD/NK	βγδε
TAL013/020/022/029	GTCG	NK/NG/HD/NN	βγδε
TAL013/020/022/030	GTCT	NK/NG/HD/NG	βγδε
TAL013/020/023/026	GTGA	NK/NG/NK/NI	βγδε
TAL013/020/023/027	GTGC	NK/NG/NK/HD	βγδε
TAL013/020/023/028	GTGG	NK/NG/NK/NK	βγδε
TAL013/020/023/029	GTGG	NK/NG/NK/NN	βγδε
TAL013/020/023/030	GTGT	NK/NG/NK/NG	βγδε
TAL013/020/024/026	GTGA	NK/NG/NN/NI	βγδε
TAL013/020/024/027	GTGC	NK/NG/NN/HD	βγδε
TAL013/020/024/028	GTGG	NK/NG/NN/NK	βγδε
TAL013/020/024/029	GTGG	NK/NG/NN/NN	βγδε
TAL013/020/024/030	GTGT	NK/NG/NN/NG	βγδε
TAL013/020/025/026	GTTA	NK/NG/NG/NI	βγδε
TAL013/020/025/027	G TTC	NK/NG/NG/HD	βγδε
TAL013/020/025/028	GTTG	NK/NG/NG/NK	βγδε
TAL013/020/025/029	GTTG	NK/NG/NG/NN	βγδε
TAL013/020/025/030	GTTT	NK/NG/NG/NG	βγδε
TAL014/016/021/026	GAAA	NN/NI/NI/NI	βγδε
TAL014/016/021/027	GAAC	NN/NI/NI/HD	βγδε
TAL014/016/021/028	GAAG	NN/NI/NI/NK	βγδε
TAL014/016/021/029	GAAG	NN/NI/NI/NN	βγδε
TAL014/016/021/030	GAAT	NN/NI/NI/NG	βγδε
TAL014/016/022/026	GACA	NN/NI/HD/NI	βγδε
TAL014/016/022/027	GACC	NN/NI/HD/HD	βγδε
TAL014/016/022/028	GACG	NN/NI/HD/NK	βγδε
TAL014/016/022/029	GACG	NN/NI/HD/NN	βγδε
TAL014/016/022/030	GACT	NN/NI/HD/NG	βγδε
TAL014/016/023/026	GAGA	NN/NI/NK/NI	βγδε
TAL014/016/023/027	GAGC	NN/NI/NK/HD	βγδε
TAL014/016/023/028	GAGG	NN/NI/NK/NK	βγδε
TAL014/016/023/029	GAGG	NN/NI/NK/NN	βγδε
TAL014/016/023/030	GAGT	NN/NI/NK/NG	βγδε

TAL014/016/024/026	GAGA	NN/NI/NN/NI	βγδε
TAL014/016/024/027	GAGC	NN/NI/NN/HD	βγδε
TAL014/016/024/028	GAGG	NN/NI/NN/NK	βγδε
TAL014/016/024/029	GAGG	NN/NI/NN/NN	βγδε
TAL014/016/024/030	GAGT	NN/NI/NN/NG	βγδε
TAL014/016/025/026	GATA	NN/NI/NG/NI	βγδε
TAL014/016/025/027	GATC	NN/NI/NG/HD	βγδε
TAL014/016/025/028	GATG	NN/NI/NG/NK	βγδε
TAL014/016/025/029	GATG	NN/NI/NG/NN	βγδε
TAL014/016/025/030	GATT	NN/NI/NG/NG	βγδε
TAL014/017/021/026	GCAA	NN/HD/NI/NI	βγδε
TAL014/017/021/027	GCAC	NN/HD/NI/HD	βγδε
TAL014/017/021/028	GCAG	NN/HD/NI/NK	βγδε
TAL014/017/021/029	GCAG	NN/HD/NI/NN	βγδε
TAL014/017/021/030	GCAT	NN/HD/NI/NG	βγδε
TAL014/017/022/026	GCCA	NN/HD/HD/NI	βγδε
TAL014/017/022/027	GCCC	NN/HD/HD/HD	βγδε
TAL014/017/022/028	GCCG	NN/HD/HD/NK	βγδε
TAL014/017/022/029	GCCG	NN/HD/HD/NN	βγδε
TAL014/017/022/030	GCCT	NN/HD/HD/NG	βγδε
TAL014/017/023/026	GCGA	NN/HD/NK/NI	βγδε
TAL014/017/023/027	GCGC	NN/HD/NK/HD	βγδε
TAL014/017/023/028	GCGG	NN/HD/NK/NK	βγδε
TAL014/017/023/029	GCGG	NN/HD/NK/NN	βγδε
TAL014/017/023/030	GCGT	NN/HD/NK/NG	βγδε
TAL014/017/024/026	GCGA	NN/HD/NN/NI	βγδε
TAL014/017/024/027	GCGC	NN/HD/NN/HD	βγδε
TAL014/017/024/028	GCGG	NN/HD/NN/NK	βγδε
TAL014/017/024/029	GCGG	NN/HD/NN/NN	βγδε
TAL014/017/024/030	GCGT	NN/HD/NN/NG	βγδε
TAL014/017/025/026	GCTA	NN/HD/NG/NI	βγδε
TAL014/017/025/027	GCTC	NN/HD/NG/HD	βγδε
TAL014/017/025/028	GCTG	NN/HD/NG/NK	βγδε
TAL014/017/025/029	GCTG	NN/HD/NG/NN	βγδε
TAL014/017/025/030	GCTT	NN/HD/NG/NG	βγδε
TAL014/018/021/026	GGAA	NN/NK/NI/NI	βγδε
TAL014/018/021/027	GGAC	NN/NK/NI/HD	βγδε
TAL014/018/021/028	GGAG	NN/NK/NI/NK	βγδε
TAL014/018/021/029	GGAG	NN/NK/NI/NN	βγδε
TAL014/018/021/030	GGAT	NN/NK/NI/NG	βγδε
TAL014/018/022/026	GGCA	NN/NK/HD/NI	βγδε

TAL014/018/022/027	GGCC	NN/NK/HD/HD	βγδε
TAL014/018/022/028	GGCG	NN/NK/HD/NK	βγδε
TAL014/018/022/029	GGCG	NN/NK/HD/NN	βγδε
TAL014/018/022/030	GGCT	NN/NK/HD/NG	βγδε
TAL014/018/023/026	GGGA	NN/NK/NK/NI	βγδε
TAL014/018/023/027	GGGC	NN/NK/NK/HD	βγδε
TAL014/018/023/028	GGGG	NN/NK/NK/NK	βγδε
TAL014/018/023/029	GGGG	NN/NK/NK/NN	βγδε
TAL014/018/023/030	GGGT	NN/NK/NK/NG	βγδε
TAL014/018/024/026	GGGA	NN/NK/NN/NI	βγδε
TAL014/018/024/027	GGGC	NN/NK/NN/HD	βγδε
TAL014/018/024/028	GGGG	NN/NK/NN/NK	βγδε
TAL014/018/024/029	GGGG	NN/NK/NN/NN	βγδε
TAL014/018/024/030	GGGT	NN/NK/NN/NG	βγδε
TAL014/018/025/026	GGTA	NN/NK/NG/NI	βγδε
TAL014/018/025/027	GGTC	NN/NK/NG/HD	βγδε
TAL014/018/025/028	GGTG	NN/NK/NG/NK	βγδε
TAL014/018/025/029	GGTG	NN/NK/NG/NN	βγδε
TAL014/018/025/030	GGTT	NN/NK/NG/NG	βγδε
TAL014/019/021/026	GGAA	NN/NN/NI/NI	βγδε
TAL014/019/021/027	GGAC	NN/NN/NI/HD	βγδε
TAL014/019/021/028	GGAG	NN/NN/NI/NK	βγδε
TAL014/019/021/029	GGAG	NN/NN/NI/NN	βγδε
TAL014/019/021/030	GGAT	NN/NN/NI/NG	βγδε
TAL014/019/022/026	GGCA	NN/NN/HD/NI	βγδε
TAL014/019/022/027	GGCC	NN/NN/HD/HD	βγδε
TAL014/019/022/028	GGCG	NN/NN/HD/NK	βγδε
TAL014/019/022/029	GGCG	NN/NN/HD/NN	βγδε
TAL014/019/022/030	GGCT	NN/NN/HD/NG	βγδε
TAL014/019/023/026	GGGA	NN/NN/NK/NI	βγδε
TAL014/019/023/027	GGGC	NN/NN/NK/HD	βγδε
TAL014/019/023/028	GGGG	NN/NN/NK/NK	βγδε
TAL014/019/023/029	GGGG	NN/NN/NK/NN	βγδε
TAL014/019/023/030	GGGT	NN/NN/NK/NG	βγδε
TAL014/019/024/026	GGGA	NN/NN/NN/NI	βγδε
TAL014/019/024/027	GGGC	NN/NN/NN/HD	βγδε
TAL014/019/024/028	GGGG	NN/NN/NN/NK	βγδε
TAL014/019/024/029	GGGG	NN/NN/NN/NN	βγδε
TAL014/019/024/030	GGGT	NN/NN/NN/NG	βγδε
TAL014/019/025/026	GGTA	NN/NN/NG/NI	βγδε
TAL014/019/025/027	GGTC	NN/NN/NG/HD	βγδε

TAL014/019/025/028	GGTG	NN/NN/NG/NK	βγδε
TAL014/019/025/029	GGTG	NN/NN/NG/NN	βγδε
TAL014/019/025/030	GGTT	NN/NN/NG/NG	βγδε
TAL014/020/021/026	GTA A	NN/NG/NI/NI	βγδε
TAL014/020/021/027	GTAC	NN/NG/NI/HD	βγδε
TAL014/020/021/028	GTAG	NN/NG/NI/NK	βγδε
TAL014/020/021/029	GTAG	NN/NG/NI/NN	βγδε
TAL014/020/021/030	GTAT	NN/NG/NI/NG	βγδε
TAL014/020/022/026	GTCA	NN/NG/HD/NI	βγδε
TAL014/020/022/027	GTCC	NN/NG/HD/HD	βγδε
TAL014/020/022/028	GTCG	NN/NG/HD/NK	βγδε
TAL014/020/022/029	GTCG	NN/NG/HD/NN	βγδε
TAL014/020/022/030	GTCT	NN/NG/HD/NG	βγδε
TAL014/020/023/026	GTGA	NN/NG/NK/NI	βγδε
TAL014/020/023/027	GTGC	NN/NG/NK/HD	βγδε
TAL014/020/023/028	GTGG	NN/NG/NK/NK	βγδε
TAL014/020/023/029	GTGG	NN/NG/NK/NN	βγδε
TAL014/020/023/030	GTGT	NN/NG/NK/NG	βγδε
TAL014/020/024/026	GTGA	NN/NG/NN/NI	βγδε
TAL014/020/024/027	GTGC	NN/NG/NN/HD	βγδε
TAL014/020/024/028	GTGG	NN/NG/NN/NK	βγδε
TAL014/020/024/029	GTGG	NN/NG/NN/NN	βγδε
TAL014/020/024/030	GTGT	NN/NG/NN/NG	βγδε
TAL014/020/025/026	GTTA	NN/NG/NG/NI	βγδε
TAL014/020/025/027	G TTC	NN/NG/NG/HD	βγδε
TAL014/020/025/028	GTTG	NN/NG/NG/NK	βγδε
TAL014/020/025/029	GTTG	NN/NG/NG/NN	βγδε
TAL014/020/025/030	GTTT	NN/NG/NG/NG	βγδε
TAL015/016/021/026	TAAA	NG/NI/NI/NI	βγδε
TAL015/016/021/027	TAAC	NG/NI/NI/HD	βγδε
TAL015/016/021/028	TAAG	NG/NI/NI/NK	βγδε
TAL015/016/021/029	TAAG	NG/NI/NI/NN	βγδε
TAL015/016/021/030	TAAT	NG/NI/NI/NG	βγδε
TAL015/016/022/026	TACA	NG/NI/HD/NI	βγδε
TAL015/016/022/027	TACC	NG/NI/HD/HD	βγδε
TAL015/016/022/028	TACG	NG/NI/HD/NK	βγδε
TAL015/016/022/029	TACG	NG/NI/HD/NN	βγδε
TAL015/016/022/030	TACT	NG/NI/HD/NG	βγδε
TAL015/016/023/026	TAGA	NG/NI/NK/NI	βγδε
TAL015/016/023/027	TAGC	NG/NI/NK/HD	βγδε
TAL015/016/023/028	TAGG	NG/NI/NK/NK	βγδε

TAL015/016/023/029	TAGG	NG/NI/NK/NN	βγδε
TAL015/016/023/030	TAGT	NG/NI/NK/NG	βγδε
TAL015/016/024/026	TAGA	NG/NI/NN/NI	βγδε
TAL015/016/024/027	TAGC	NG/NI/NN/HD	βγδε
TAL015/016/024/028	TAGG	NG/NI/NN/NK	βγδε
TAL015/016/024/029	TAGG	NG/NI/NN/NN	βγδε
TAL015/016/024/030	TAGT	NG/NI/NN/NG	βγδε
TAL015/016/025/026	TATA	NG/NI/NG/NI	βγδε
TAL015/016/025/027	TATC	NG/NI/NG/HD	βγδε
TAL015/016/025/028	TATG	NG/NI/NG/NK	βγδε
TAL015/016/025/029	TATG	NG/NI/NG/NN	βγδε
TAL015/016/025/030	TATT	NG/NI/NG/NG	βγδε
TAL015/017/021/026	TCAA	NG/HD/NI/NI	βγδε
TAL015/017/021/027	TCAC	NG/HD/NI/HD	βγδε
TAL015/017/021/028	TCAG	NG/HD/NI/NK	βγδε
TAL015/017/021/029	TCAG	NG/HD/NI/NN	βγδε
TAL015/017/021/030	TCAT	NG/HD/NI/NG	βγδε
TAL015/017/022/026	TCCA	NG/HD/HD/NI	βγδε
TAL015/017/022/027	TCCC	NG/HD/HD/HD	βγδε
TAL015/017/022/028	TCCG	NG/HD/HD/NK	βγδε
TAL015/017/022/029	TCCG	NG/HD/HD/NN	βγδε
TAL015/017/022/030	TCCT	NG/HD/HD/NG	βγδε
TAL015/017/023/026	TCGA	NG/HD/NK/NI	βγδε
TAL015/017/023/027	TCGC	NG/HD/NK/HD	βγδε
TAL015/017/023/028	TCGG	NG/HD/NK/NK	βγδε
TAL015/017/023/029	TCGG	NG/HD/NK/NN	βγδε
TAL015/017/023/030	TCGT	NG/HD/NK/NG	βγδε
TAL015/017/024/026	TCGA	NG/HD/NN/NI	βγδε
TAL015/017/024/027	TCGC	NG/HD/NN/HD	βγδε
TAL015/017/024/028	TCGG	NG/HD/NN/NK	βγδε
TAL015/017/024/029	TCGG	NG/HD/NN/NN	βγδε
TAL015/017/024/030	TCGT	NG/HD/NN/NG	βγδε
TAL015/017/025/026	TCTA	NG/HD/NG/NI	βγδε
TAL015/017/025/027	TCTC	NG/HD/NG/HD	βγδε
TAL015/017/025/028	TCTG	NG/HD/NG/NK	βγδε
TAL015/017/025/029	TCTG	NG/HD/NG/NN	βγδε
TAL015/017/025/030	TCTT	NG/HD/NG/NG	βγδε
TAL015/018/021/026	TGAA	NG/NK/NI/NI	βγδε
TAL015/018/021/027	TGAC	NG/NK/NI/HD	βγδε
TAL015/018/021/028	TGAG	NG/NK/NI/NK	βγδε
TAL015/018/021/029	TGAG	NG/NK/NI/NN	βγδε

TAL015/018/021/030	TGAT	NG/NK/NI/NG	βγδε
TAL015/018/022/026	TGCA	NG/NK/HD/NI	βγδε
TAL015/018/022/027	TGCC	NG/NK/HD/HD	βγδε
TAL015/018/022/028	TGCG	NG/NK/HD/NK	βγδε
TAL015/018/022/029	TGCG	NG/NK/HD/NN	βγδε
TAL015/018/022/030	TGCT	NG/NK/HD/NG	βγδε
TAL015/018/023/026	TGGA	NG/NK/NK/NI	βγδε
TAL015/018/023/027	TGGC	NG/NK/NK/HD	βγδε
TAL015/018/023/028	TGGG	NG/NK/NK/NK	βγδε
TAL015/018/023/029	TGGG	NG/NK/NK/NN	βγδε
TAL015/018/023/030	TGGT	NG/NK/NK/NG	βγδε
TAL015/018/024/026	TGGA	NG/NK/NN/NI	βγδε
TAL015/018/024/027	TGGC	NG/NK/NN/HD	βγδε
TAL015/018/024/028	TGGG	NG/NK/NN/NK	βγδε
TAL015/018/024/029	TGGG	NG/NK/NN/NN	βγδε
TAL015/018/024/030	TGGT	NG/NK/NN/NG	βγδε
TAL015/018/025/026	TGTA	NG/NK/NG/NI	βγδε
TAL015/018/025/027	TGTC	NG/NK/NG/HD	βγδε
TAL015/018/025/028	TGTG	NG/NK/NG/NK	βγδε
TAL015/018/025/029	TGTG	NG/NK/NG/NN	βγδε
TAL015/018/025/030	TGTT	NG/NK/NG/NG	βγδε
TAL015/019/021/026	TGAA	NG/NN/NI/NI	βγδε
TAL015/019/021/027	TGAC	NG/NN/NI/HD	βγδε
TAL015/019/021/028	TGAG	NG/NN/NI/NK	βγδε
TAL015/019/021/029	TGAG	NG/NN/NI/NN	βγδε
TAL015/019/021/030	TGAT	NG/NN/NI/NG	βγδε
TAL015/019/022/026	TGCA	NG/NN/HD/NI	βγδε
TAL015/019/022/027	TGCC	NG/NN/HD/HD	βγδε
TAL015/019/022/028	TGCG	NG/NN/HD/NK	βγδε
TAL015/019/022/029	TGCG	NG/NN/HD/NN	βγδε
TAL015/019/022/030	TGCT	NG/NN/HD/NG	βγδε
TAL015/019/023/026	TGGA	NG/NN/NK/NI	βγδε
TAL015/019/023/027	TGGC	NG/NN/NK/HD	βγδε
TAL015/019/023/028	TGGG	NG/NN/NK/NK	βγδε
TAL015/019/023/029	TGGG	NG/NN/NK/NN	βγδε
TAL015/019/023/030	TGGT	NG/NN/NK/NG	βγδε
TAL015/019/024/026	TGGA	NG/NN/NN/NI	βγδε
TAL015/019/024/027	TGGC	NG/NN/NN/HD	βγδε
TAL015/019/024/028	TGGG	NG/NN/NN/NK	βγδε
TAL015/019/024/029	TGGG	NG/NN/NN/NN	βγδε
TAL015/019/024/030	TGGT	NG/NN/NN/NG	βγδε

TAL015/019/025/026	TGTA	NG/NN/NG/NI	$\beta\gamma\delta\epsilon$
TAL015/019/025/027	TGTC	NG/NN/NG/HD	$\beta\gamma\delta\epsilon$
TAL015/019/025/028	TGTG	NG/NN/NG/NK	$\beta\gamma\delta\epsilon$
TAL015/019/025/029	TGTG	NG/NN/NG/NN	$\beta\gamma\delta\epsilon$
TAL015/019/025/030	TGTT	NG/NN/NG/NG	$\beta\gamma\delta\epsilon$
TAL015/020/021/026	TTAA	NG/NG/NI/NI	$\beta\gamma\delta\epsilon$
TAL015/020/021/027	TTAC	NG/NG/NI/HD	$\beta\gamma\delta\epsilon$
TAL015/020/021/028	TTAG	NG/NG/NI/NK	$\beta\gamma\delta\epsilon$
TAL015/020/021/029	TTAG	NG/NG/NI/NN	$\beta\gamma\delta\epsilon$
TAL015/020/021/030	TTAT	NG/NG/NI/NG	$\beta\gamma\delta\epsilon$
TAL015/020/022/026	TTCA	NG/NG/HD/NI	$\beta\gamma\delta\epsilon$
TAL015/020/022/027	TTCC	NG/NG/HD/HD	$\beta\gamma\delta\epsilon$
TAL015/020/022/028	TTCG	NG/NG/HD/NK	$\beta\gamma\delta\epsilon$
TAL015/020/022/029	TTCG	NG/NG/HD/NN	$\beta\gamma\delta\epsilon$
TAL015/020/022/030	TTCT	NG/NG/HD/NG	$\beta\gamma\delta\epsilon$
TAL015/020/023/026	TTGA	NG/NG/NK/NI	$\beta\gamma\delta\epsilon$
TAL015/020/023/027	TTGC	NG/NG/NK/HD	$\beta\gamma\delta\epsilon$
TAL015/020/023/028	TTGG	NG/NG/NK/NK	$\beta\gamma\delta\epsilon$
TAL015/020/023/029	TTGG	NG/NG/NK/NN	$\beta\gamma\delta\epsilon$
TAL015/020/023/030	TTGT	NG/NG/NK/NG	$\beta\gamma\delta\epsilon$
TAL015/020/024/026	TTGA	NG/NG/NN/NI	$\beta\gamma\delta\epsilon$
TAL015/020/024/027	TTGC	NG/NG/NN/HD	$\beta\gamma\delta\epsilon$
TAL015/020/024/028	TTGG	NG/NG/NN/NK	$\beta\gamma\delta\epsilon$
TAL015/020/024/029	TTGG	NG/NG/NN/NN	$\beta\gamma\delta\epsilon$
TAL015/020/024/030	TTGT	NG/NG/NN/NG	$\beta\gamma\delta\epsilon$
TAL015/020/025/026	TTTA	NG/NG/NG/NI	$\beta\gamma\delta\epsilon$
TAL015/020/025/027	TTTC	NG/NG/NG/HD	$\beta\gamma\delta\epsilon$
TAL015/020/025/028	TTTG	NG/NG/NG/NK	$\beta\gamma\delta\epsilon$
TAL015/020/025/029	TTTG	NG/NG/NG/NN	$\beta\gamma\delta\epsilon$
TAL015/020/025/030	TTTT	NG/NG/NG/NG	$\beta\gamma\delta\epsilon$
TAL011/016	AA	NI/NI	$\beta\gamma$
TAL011/017	AC	NI/HD	$\beta\gamma$
TAL011/018	AG	NI/NK	$\beta\gamma$
TAL011/019	AG	NI/NN	$\beta\gamma$
TAL011/020	AT	NI/NG	$\beta\gamma$
TAL012/016	CA	HD/NI	$\beta\gamma$
TAL012/017	CC	HD/HD	$\beta\gamma$
TAL012/018	CG	HD/NK	$\beta\gamma$
TAL012/019	CG	HD/NN	$\beta\gamma$
TAL012/020	CT	HD/NG	$\beta\gamma$
TAL013/016	GA	NK/NI	$\beta\gamma$

TAL013/017	GC	NK/HD	$\beta\gamma$
TAL013/018	GG	NK/NK	$\beta\gamma$
TAL013/019	GG	NK/NN	$\beta\gamma$
TAL013/020	GT	NK/NG	$\beta\gamma$
TAL014/016	GA	NN/NI	$\beta\gamma$
TAL014/017	GC	NN/HD	$\beta\gamma$
TAL014/018	GG	NN/NK	$\beta\gamma$
TAL014/019	GG	NN/NN	$\beta\gamma$
TAL014/020	GT	NN/NG	$\beta\gamma$
TAL015/016	TA	NG/NI	$\beta\gamma$
TAL015/017	TC	NG/HD	$\beta\gamma$
TAL015/018	TG	NG/NK	$\beta\gamma$
TAL015/019	TG	NG/NN	$\beta\gamma$
TAL015/020	TT	NG/NG	$\beta\gamma$
TAL011/016/021	AAA	NI/NI/NI	$\beta\gamma\delta$
TAL011/016/022	AAC	NI/NI/HD	$\beta\gamma\delta$
TAL011/016/023	AAG	NI/NI/NK	$\beta\gamma\delta$
TAL011/016/024	AAG	NI/NI/NN	$\beta\gamma\delta$
TAL011/016/025	AAT	NI/NI/NG	$\beta\gamma\delta$
TAL011/017/021	ACA	NI/HD/NI	$\beta\gamma\delta$
TAL011/017/022	ACC	NI/HD/HD	$\beta\gamma\delta$
TAL011/017/023	ACG	NI/HD/NK	$\beta\gamma\delta$
TAL011/017/024	ACG	NI/HD/NN	$\beta\gamma\delta$
TAL011/017/025	ACT	NI/HD/NG	$\beta\gamma\delta$
TAL011/018/021	AGA	NI/NK/NI	$\beta\gamma\delta$
TAL011/018/022	AGC	NI/NK/HD	$\beta\gamma\delta$
TAL011/018/023	AGG	NI/NK/NK	$\beta\gamma\delta$
TAL011/018/024	AGG	NI/NK/NN	$\beta\gamma\delta$
TAL011/018/025	AGT	NI/NK/NG	$\beta\gamma\delta$
TAL011/019/021	AGA	NI/NN/NI	$\beta\gamma\delta$
TAL011/019/022	AGC	NI/NN/HD	$\beta\gamma\delta$
TAL011/019/023	AGG	NI/NN/NK	$\beta\gamma\delta$
TAL011/019/024	AGG	NI/NN/NN	$\beta\gamma\delta$
TAL011/019/025	AGT	NI/NN/NG	$\beta\gamma\delta$
TAL011/020/021	ATA	NI/NG/NI	$\beta\gamma\delta$
TAL011/020/022	ATC	NI/NG/HD	$\beta\gamma\delta$
TAL011/020/023	ATG	NI/NG/NK	$\beta\gamma\delta$
TAL011/020/024	ATG	NI/NG/NN	$\beta\gamma\delta$
TAL011/020/025	ATT	NI/NG/NG	$\beta\gamma\delta$
TAL012/016/021	CAA	HD/NI/NI	$\beta\gamma\delta$
TAL012/016/022	CAC	HD/NI/HD	$\beta\gamma\delta$

TAL012/016/023	CAG	HD/NI/NK	$\beta\gamma\delta$
TAL012/016/024	CAG	HD/NI/NN	$\beta\gamma\delta$
TAL012/016/025	CAT	HD/NI/NG	$\beta\gamma\delta$
TAL012/017/021	CCA	HD/HD/NI	$\beta\gamma\delta$
TAL012/017/022	CCC	HD/HD/HD	$\beta\gamma\delta$
TAL012/017/023	CCG	HD/HD/NK	$\beta\gamma\delta$
TAL012/017/024	CCG	HD/HD/NN	$\beta\gamma\delta$
TAL012/017/025	CCT	HD/HD/NG	$\beta\gamma\delta$
TAL012/018/021	CGA	HD/NK/NI	$\beta\gamma\delta$
TAL012/018/022	CGC	HD/NK/HD	$\beta\gamma\delta$
TAL012/018/023	CGG	HD/NK/NK	$\beta\gamma\delta$
TAL012/018/024	CGG	HD/NK/NN	$\beta\gamma\delta$
TAL012/018/025	CGT	HD/NK/NG	$\beta\gamma\delta$
TAL012/019/021	CGA	HD/NN/NI	$\beta\gamma\delta$
TAL012/019/022	CGC	HD/NN/HD	$\beta\gamma\delta$
TAL012/019/023	CGG	HD/NN/NK	$\beta\gamma\delta$
TAL012/019/024	CGG	HD/NN/NN	$\beta\gamma\delta$
TAL012/019/025	CGT	HD/NN/NG	$\beta\gamma\delta$
TAL012/020/021	CTA	HD/NG/NI	$\beta\gamma\delta$
TAL012/020/022	CTC	HD/NG/HD	$\beta\gamma\delta$
TAL012/020/023	CTG	HD/NG/NK	$\beta\gamma\delta$
TAL012/020/024	CTG	HD/NG/NN	$\beta\gamma\delta$
TAL012/020/025	CTT	HD/NG/NG	$\beta\gamma\delta$
TAL013/016/021	GAA	NK/NI/NI	$\beta\gamma\delta$
TAL013/016/022	GAC	NK/NI/HD	$\beta\gamma\delta$
TAL013/016/023	GAG	NK/NI/NK	$\beta\gamma\delta$
TAL013/016/024	GAG	NK/NI/NN	$\beta\gamma\delta$
TAL013/016/025	GAT	NK/NI/NG	$\beta\gamma\delta$
TAL013/017/021	GCA	NK/HD/NI	$\beta\gamma\delta$
TAL013/017/022	GCC	NK/HD/HD	$\beta\gamma\delta$
TAL013/017/023	GCG	NK/HD/NK	$\beta\gamma\delta$
TAL013/017/024	GCG	NK/HD/NN	$\beta\gamma\delta$
TAL013/017/025	GCT	NK/HD/NG	$\beta\gamma\delta$
TAL013/018/021	GGA	NK/NK/NI	$\beta\gamma\delta$
TAL013/018/022	GGC	NK/NK/HD	$\beta\gamma\delta$
TAL013/018/023	GGG	NK/NK/NK	$\beta\gamma\delta$
TAL013/018/024	GGG	NK/NK/NN	$\beta\gamma\delta$
TAL013/018/025	GGT	NK/NK/NG	$\beta\gamma\delta$
TAL013/019/021	GGA	NK/NN/NI	$\beta\gamma\delta$
TAL013/019/022	GGC	NK/NN/HD	$\beta\gamma\delta$
TAL013/019/023	GGG	NK/NN/NK	$\beta\gamma\delta$

TAL013/019/024	GGG	NK/NN/NN	$\beta\gamma\delta$
TAL013/019/025	GGT	NK/NN/NG	$\beta\gamma\delta$
TAL013/020/021	GTA	NK/NG/NI	$\beta\gamma\delta$
TAL013/020/022	GTC	NK/NG/HD	$\beta\gamma\delta$
TAL013/020/023	GTG	NK/NG/NK	$\beta\gamma\delta$
TAL013/020/024	GTG	NK/NG/NN	$\beta\gamma\delta$
TAL013/020/025	GTT	NK/NG/NG	$\beta\gamma\delta$
TAL014/016/021	GAA	NN/NI/NI	$\beta\gamma\delta$
TAL014/016/022	GAC	NN/NI/HD	$\beta\gamma\delta$
TAL014/016/023	GAG	NN/NI/NK	$\beta\gamma\delta$
TAL014/016/024	GAG	NN/NI/NN	$\beta\gamma\delta$
TAL014/016/025	GAT	NN/NI/NG	$\beta\gamma\delta$
TAL014/017/021	GCA	NN/HD/NI	$\beta\gamma\delta$
TAL014/017/022	GCC	NN/HD/HD	$\beta\gamma\delta$
TAL014/017/023	GCG	NN/HD/NK	$\beta\gamma\delta$
TAL014/017/024	GCG	NN/HD/NN	$\beta\gamma\delta$
TAL014/017/025	GCT	NN/HD/NG	$\beta\gamma\delta$
TAL014/018/021	GGA	NN/NK/NI	$\beta\gamma\delta$
TAL014/018/022	GGC	NN/NK/HD	$\beta\gamma\delta$
TAL014/018/023	GGG	NN/NK/NK	$\beta\gamma\delta$
TAL014/018/024	GGG	NN/NK/NN	$\beta\gamma\delta$
TAL014/018/025	GGT	NN/NK/NG	$\beta\gamma\delta$
TAL014/019/021	GGA	NN/NN/NI	$\beta\gamma\delta$
TAL014/019/022	GGC	NN/NN/HD	$\beta\gamma\delta$
TAL014/019/023	GGG	NN/NN/NK	$\beta\gamma\delta$
TAL014/019/024	GGG	NN/NN/NN	$\beta\gamma\delta$
TAL014/019/025	GGT	NN/NN/NG	$\beta\gamma\delta$
TAL014/020/021	GTA	NN/NG/NI	$\beta\gamma\delta$
TAL014/020/022	GTC	NN/NG/HD	$\beta\gamma\delta$
TAL014/020/023	GTG	NN/NG/NK	$\beta\gamma\delta$
TAL014/020/024	GTG	NN/NG/NN	$\beta\gamma\delta$
TAL014/020/025	GTT	NN/NG/NG	$\beta\gamma\delta$
TAL015/016/021	TAA	NG/NI/NI	$\beta\gamma\delta$
TAL015/016/022	TAC	NG/NI/HD	$\beta\gamma\delta$
TAL015/016/023	TAG	NG/NI/NK	$\beta\gamma\delta$
TAL015/016/024	TAG	NG/NI/NN	$\beta\gamma\delta$
TAL015/016/025	TAT	NG/NI/NG	$\beta\gamma\delta$
TAL015/017/021	TCA	NG/HD/NI	$\beta\gamma\delta$
TAL015/017/022	TCC	NG/HD/HD	$\beta\gamma\delta$
TAL015/017/023	TCG	NG/HD/NK	$\beta\gamma\delta$
TAL015/017/024	TCG	NG/HD/NN	$\beta\gamma\delta$

TAL015/017/025	TCT	NG/HD/NG	$\beta\gamma\delta$
TAL015/018/021	TGA	NG/NK/NI	$\beta\gamma\delta$
TAL015/018/022	TGC	NG/NK/HD	$\beta\gamma\delta$
TAL015/018/023	TGG	NG/NK/NK	$\beta\gamma\delta$
TAL015/018/024	TGG	NG/NK/NN	$\beta\gamma\delta$
TAL015/018/025	TGT	NG/NK/NG	$\beta\gamma\delta$
TAL015/019/021	TGA	NG/NN/NI	$\beta\gamma\delta$
TAL015/019/022	TGC	NG/NN/HD	$\beta\gamma\delta$
TAL015/019/023	TGG	NG/NN/NK	$\beta\gamma\delta$
TAL015/019/024	TGG	NG/NN/NN	$\beta\gamma\delta$
TAL015/019/025	TGT	NG/NN/NG	$\beta\gamma\delta$
TAL015/020/021	TTA	NG/NG/NI	$\beta\gamma\delta$
TAL015/020/022	TTC	NG/NG/HD	$\beta\gamma\delta$
TAL015/020/023	TTG	NG/NG/NK	$\beta\gamma\delta$
TAL015/020/024	TTG	NG/NG/NN	$\beta\gamma\delta$
TAL015/020/025	TTT	NG/NG/NG	$\beta\gamma\delta$
TAL011/031	AA	NI/NI	$\beta\gamma'$
TAL011/032	AC	NI/HD	$\beta\gamma'$
TAL011/033	AG	NI/NK	$\beta\gamma'$
TAL011/034	AG	NI/NN	$\beta\gamma'$
TAL011/035	AT	NI/NG	$\beta\gamma'$
TAL012/031	CA	HD/NI	$\beta\gamma'$
TAL012/032	CC	HD/HD	$\beta\gamma'$
TAL012/033	CG	HD/NK	$\beta\gamma'$
TAL012/034	CG	HD/NN	$\beta\gamma'$
TAL012/035	CT	HD/NG	$\beta\gamma'$
TAL013/031	GA	NK/NI	$\beta\gamma'$
TAL013/032	GC	NK/HD	$\beta\gamma'$
TAL013/033	GG	NK/NK	$\beta\gamma'$
TAL013/034	GG	NK/NN	$\beta\gamma'$
TAL013/035	GT	NK/NG	$\beta\gamma'$
TAL014/031	GA	NN/NI	$\beta\gamma'$
TAL014/032	GC	NN/HD	$\beta\gamma'$
TAL014/033	GG	NN/NK	$\beta\gamma'$
TAL014/034	GG	NN/NN	$\beta\gamma'$
TAL014/035	GT	NN/NG	$\beta\gamma'$
TAL015/031	TA	NG/NI	$\beta\gamma'$
TAL015/032	TC	NG/HD	$\beta\gamma'$
TAL015/033	TG	NG/NK	$\beta\gamma'$
TAL015/034	TG	NG/NN	$\beta\gamma'$
TAL015/035	TT	NG/NG	$\beta\gamma'$

TAL021/036	AA	NI/NI	$\delta\epsilon'$
TAL021/037	AC	NI/HD	$\delta\epsilon'$
TAL021/038	AG	NI/NK	$\delta\epsilon'$
TAL021/039	AG	NI/NN	$\delta\epsilon'$
TAL021/040	AT	NI/NG	$\delta\epsilon'$
TAL022/036	CA	HD/NI	$\delta\epsilon'$
TAL022/037	CC	HD/HD	$\delta\epsilon'$
TAL022/038	CG	HD/NK	$\delta\epsilon'$
TAL022/039	CG	HD/NN	$\delta\epsilon'$
TAL022/040	CT	HD/NG	$\delta\epsilon'$
TAL023/036	GA	NK/NI	$\delta\epsilon'$
TAL023/037	GC	NK/HD	$\delta\epsilon'$
TAL023/038	GG	NK/NK	$\delta\epsilon'$
TAL023/039	GG	NK/NN	$\delta\epsilon'$
TAL023/040	GT	NK/NG	$\delta\epsilon'$
TAL024/036	GA	NN/NI	$\delta\epsilon'$
TAL024/037	GC	NN/HD	$\delta\epsilon'$
TAL024/038	GG	NN/NK	$\delta\epsilon'$
TAL024/039	GG	NN/NN	$\delta\epsilon'$
TAL024/040	GT	NN/NG	$\delta\epsilon'$
TAL025/036	TA	NG/NI	$\delta\epsilon'$
TAL025/037	TC	NG/HD	$\delta\epsilon'$
TAL025/038	TG	NG/NK	$\delta\epsilon'$
TAL025/039	TG	NG/NN	$\delta\epsilon'$
TAL025/040	TT	NG/NG	$\delta\epsilon'$
TAL011	A	NI	β
TAL012	C	HD	β
TAL013	G	NK	β
TAL014	G	NN	β
TAL015	T	NG	β

To prepare DNA fragments encoding α units for use in assembly, 20 rounds of PCR were performed with each α unit plasmid as a template using primers oJS2581 (5'-Biotin-TCTAGAGAAGACAAGAACCTGACC-3' (SEQ ID NO:237)) and oJS2582 (5'-GGATCCGGTCTCTTAAGGCCGTGG-3' (SEQ ID NO:238)). The resulting PCR products were biotinylated on the 5' end. Each α PCR product was then digested with 40 units of BsaI-HF restriction enzyme to generate 4bp overhangs, purified using the

QIAquick PCR purification kit (QIAGEN) according to manufacturer's instructions except that the final product was eluted in 50 μ l of 0.1X EB.

To prepare DNA fragments encoding β , $\beta\gamma\delta\epsilon$, $\beta\gamma\delta$, $\beta\gamma$, $\beta\gamma^*$, and $\delta\epsilon^*$ repeats, 10 μ g of each of these plasmids was digested with 50 units of BbsI restriction enzyme in
5 NEBuffer 2 for 2 hours at 37 °C followed by serial restriction digests performed in
NEBuffer 4 at 37 °C using 100 units each of XbaI, BamHI-HF, and Sall-HF enzymes that
were added at 5 minute intervals. The latter set of restriction digestions were designed to
cleave the plasmid backbone to ensure that this larger DNA fragment does not interfere
with subsequent ligations performed during the assembly process. These restriction
10 digest reactions were then purified using the QIAquick PCR purification kit (QIAGEN)
according to manufacturer's instructions except that the final product was eluted in 180 μ l
of 0.1X EB.

All assembly steps were performed using a Sciclone G3 liquid handling
workstation (Caliper) in 96-well plates and using a SPRiplate 96-ring magnet (Beckman
15 Coulter Genomics) and a DynaMag-96 Side magnet (Life Technologies). In the first
assembly step, a biotinylated α unit fragment was ligated to the first $\beta\gamma\delta\epsilon$ fragment and
then the resulting $\alpha\beta\gamma\delta\epsilon$ fragments are bound to Dynabeads MyOne C1 streptavidin-
coated magnetic beads (Life Technologies) in 2X B&W Buffer (Life Technologies).
Beads were then drawn to the side of the well by placing the plate on the magnet and then
20 washed with 100 μ l B&W buffer with 0.005% Tween 20 (Sigma) and again with 100 μ l
0.1 mg/ml bovine serum albumin (BSA) (New England Biolabs). Additional $\beta\gamma\delta\epsilon$
fragments were ligated by removing the plate from the magnet, resuspending the beads in
solution in each well, digesting the bead bound fragment with BsaI-HF restriction
enzyme, placing the plate on the magnet, washing with 100 μ l B&W/Tween20 followed
25 by 100 μ l of 0.1 mg/ml BSA, and then ligating the next fragment. This process was
repeated multiple times with additional $\beta\gamma\delta\epsilon$ units to extend the bead-bound fragment.
The last fragment to be ligated was always a β , $\beta\gamma^*$, $\beta\gamma\delta$, or $\delta\epsilon^*$ unit to enable cloning of
the full-length fragment into expression vectors (note that fragments that end with a $\delta\epsilon^*$
unit are always preceded by ligation of a $\beta\gamma$ unit).

The final full-length bead-bound fragment was digested with 40 units of BsaI-HF restriction enzyme followed by 25 units of BbsI restriction enzyme (New England Biolabs). Digestion with BbsI released the fragment from the beads and generated a unique 5' overhang for cloning of the fragment. Digestion with BsaI-HF resulted in
5 creation of a unique 3' overhang for cloning.

DNA fragments encoding the assembled TALE repeat arrays were subcloned into one of four TALEN expression vectors. Each of these vectors included a CMV promoter, a translational start codon optimized for mammalian cell expression, a triple FLAG epitope tag, a nuclear localization signal, amino acids 153 to 288 from the TALE 13
10 protein (Miller et al., 2011, Nat. Biotechnol., 29:143-148), two unique and closely positioned Type IIS BsmBI restriction sites, a 0.5 TALE repeat domain encoding one of four possible RVDs (NI, HD, NN, or NG for recognition of an A, C, G, or T nucleotide, respectively), amino acids 715 to 777 from the TALE 13 protein, and the wild-type FokI cleavage domain. All DNA fragments possessed overhangs that enable directional
15 cloning into any of the four TALEN expression vectors that has been digested with BsmBI.

To prepare a TALEN expression vector for subcloning, 5 µg of plasmid DNA were digested with 50 units of BsmBI restriction enzyme (New England Biolabs) in NEBuffer 3 for 8 hours at 55 degrees C. Digested DNA was purified using 90 µl of
20 AmpureTM XP beads (Agencourt) according to manufacturer's instructions and diluted to a final concentration of 5ng/µl in 1 mM TrisHCl. The assembled TALE repeat arrays were ligated into TALEN expression vectors using 400 U of T4 DNA Ligase (New England Biolabs). Ligation products were transformed into chemically competent XL-1 Blue cells. Six colonies were picked for each ligation and plasmid DNA isolated by an
25 alkaline lysis miniprep procedure. Simultaneously, the same six colonies were screened by PCR using primers oSQT34 (5'-GACGGTGGCTGTCAAATACCAAGATATG-3' (SEQ ID NO:239)) and oSQT35 (5'-TCTCCTCCAGTTCACCTTTTGACTAGTTGGG - 3' (SEQ ID NO:240)). PCR products were analyzed on a QIAxcel capillary electrophoresis system (Qiagen). Miniprep DNA from clones that contained correctly
30 sized PCR products were sent for DNA sequence confirmation with primers oSQT1 (5'-

AGTAACAGCGGTAGAGGCAG-3' (SEQ ID NO:241)), oSQT3
(5'-ATTGGGCTACGATGGACTCC-3' (SEQ ID NO:242)), and oJS2980
(5-TTAATTCAATATATTCATGAGGCAC-3' (SEQ ID NO:243)).

5 Because the final fragment ligated can encode one, two, or three TALE repeats,
the methods disclosed herein can be used to assemble arrays consisting of any desired
number of TALE repeats. Assembled DNA fragments encoding the final full-length
TALE repeat array are released from the beads by restriction enzyme digestion and can
be directly cloned into a desired expression vector of choice.

10 The methods can be efficiently practiced in 96-well format using a robotic liquid
handling workstation. With automation, DNA fragments encoding 96 different TALE
repeat arrays of variable lengths can be assembled in less than one day. Medium-
throughput assembly of fragments can be performed in one to two days using multi-
channel pipets and 96-well plates. Fragments assembled using either approach can then
be cloned into expression vectors (e.g., for expression as a TALEN) to generate sequence-
15 verified plasmids in less than one week. Using the automated assembly approach,
sequence-verified TALE repeat array expression plasmids can be made quickly and
inexpensively.

20 Example 6. Large-scale Testing of Assembled TALENs Using a Human Cell-based
Reporter Assay

To perform a large-scale test of the robustness of TALENs for genome editing in
human cells, the method described in Example 5 was used to construct a series of
plasmids encoding 48 TALEN pairs targeted to different sites scattered throughout the
EGFP reporter gene. Monomers in each of the TALEN pairs contained the same number
25 of repeats (ranging from 8.5 to 19.5 in number), and these pairs were targeted to sites
possessing a fixed length "spacer" sequence (16 bps) between the "half-sites" bound by
each TALEN monomer (Table 6).

Table 6. EGFP reporter gene sequences targeted by 48 pairs of TALENs

TALEN pair #	Position within EGFP of the first nucleotide in the binding site	Target site (half-sites in CAPS, spacer in lowercase)	SEQ ID NO:	# of repeat domains in Left TALEN	# of repeat domains in Right TALEN
1	-8	TCGCCACCATggtgagcaaggg cgagGAGCTGTTCA	93	8.5	8.5
2	35	TGGTGCCCATcctggtcgagct ggacGGCGACGTAA	94	8.5	8.5
3	143	TCTGCACCACcggcaagctgcc cgtgCCCTGGCCCA	95	8.5	8.5
4	425	TGGAGTACAActacaacagcca caacGTCTATATCA	96	8.5	8.5
5	82	TTCAGCGTGTcggcgagggcg agggcGATGCCACCTA	97	9.5	9.5
6	111	TGCCACCTACGgcaagctgacc ctgaaGTCATCTGCA	98	9.5	9.5
7	172	TGGCCACCCTcgtgaccaccc tgaccTACGGCGTGCA	99	9.5	9.5
8	496	TTCAAGATCCGccacaacatcg aggacGGCAGCGTGCA	100	9.5	9.5
9	-23	TAGAGGATCCACcggtcgccac catggtGAGCAAGGGCCA	101	10.5	10.5
10	91	TCCGGCGAGGGCgagggcgatg ccacctACGGCAAGCTGA	102	10.5	10.5
11	194	TGACCTACGGCGtgcaagtgtt cagccgCTACCCGACCA	103	10.5	10.5
12	503	TCCGCCACAACatcgaggacgg cagcgtGCAGCTCGCCGA	104	10.5	10.5
13	44	TCCGTGGTCGAGCTggacggcga cgtaacGGCCACAAGTCA	105	11.5	11.5
14	215	TCAGCCGCTACCCcgaccacat gaagcagCAGACTTCTTCA	106	11.5	11.5
15	251	TCTTCAAGTCCGCcatgccga aggctacTCCAGGAGCGCA	107	11.5	11.5
16	392	TCAAGGAGGACGGcaacatcct ggggcacAAGCTGGAGTACA	108	11.5	11.5
17	485	TCAAGGTGAACTTcaagatccg ccacaacATCGAGGACGGCA	109	11.5	11.5
18	-16	TCCACCGGTCGCCAccatggtg agcaagggCGAGGAGCTGTCA	110	12.5	12.5
19	82	TTCAGCGTGTCCGGcagggcg agggcgatGCCACCTACGCAA	111	12.5	12.5
20	214	TTCAGCCGCTACCCcgaccaca tgaagcagCAGACTTCTTCAA	112	12.5	12.5
21	436	TACAACAGCCACAacgtctata tcatggccGACAAGCAGAAGAA	113	12.5	12.5

22	35	TGGTGCCCATCTGGtegagct ggacggcgaCGTAAACGGCCAC AA	114	13.5	13.5
23	266	TGCCCCAAGGCTACGtccagga gcgaccatCTCTTCAAGGAC GA	115	13.5	13.5
24	362	TGAACCGCATCGAGCtgaagg catcgacttCAAGGAGGACGGC AA	116	13.5	13.5
25	497	TCAAGATCCGCCACAacatcga ggacggcagCGTGCAGCTCGCC GA	117	13.5	13.5
26	23	TGTTACCGGGTGGTgccc cctggtcgagCTGGACGGCGAC GTAA	118	14.5	14.5
27	38	TGCCCATCCTGGTCGagctgga cggcgaagtaAACGGCCACAAG TTCA	119	14.5	14.5
28	89	TGTCCGGCGAGGGCGAgggcga tgccacctacGGCAAGCTGACC CTGA	120	14.5	14.5
29	140	TCATCTGCACCACGGcaagct gcccgtgcccTGGCCACCCCTC GTGA	121	14.5	14.5
30	452	TCTATATCATGGCCGAcaagca gaagaacggcATCAAGGTGAAC TTCA	122	14.5	14.5
31	199	TACGGCGTGCAGTGCTTcagcc gctaccccgacCACATGAAGCA GCACGA	123	15.5	15.5
32	223	TACCCCGACCACATGAAGcagc acgacttcttCAAGTCCGCCAT GCCCCA	124	15.5	15.5
33	259	TCCGCCATGCCCGAAGGctacg tccaggagcgcACCATCTCTT CAAGGA	125	15.5	15.5
34	391	TTCAAGGAGGACGGCAacatcc tggggcacaagCTGGAGTACAA CTACAA	126	15.5	15.5
35	430	TACAAC TACAACAGCCAcaacg tctatatcatgGCCACAAGCA GAAGAA	127	15.5	15.5
36	26	TCACCGGGTGGTGCCAtcct ggtcgagctggaCGGCGACGTA AACGGCCA	128	16.5	16.5
37	68	TAAACGGCCACAAGTTCAgagct gtccggcgaggCGAGGGCGAT GCCACCTA	129	16.5	16.5
38	206	TGCAGTGCTTACGGCGTacc cgaccacatgaaGCAGCAGCAC TTCTCAA	130	16.5	16.5
39	83	TCAGCGTGTCCGGCGAGGGcga gggggatgccaccTACGGCAAG CTGACCCTGA	131	17.5	17.5

40	134	TGAAGTTCATCTGCACCACcgg caagctgccccgtgCCCTGGCCC ACCCTCGTGA	132	17.5	17.5
41	182	TTCGTGACCACCCTGACCTAcgg cgtgcagtgcttcAGCCGCTAC CCCACCACA	133	17.5	17.5
42	458	TCATGGCCGACAAGCAGAAgaa cggcatcaaggtgAACTCAAG ATCCGCCACA	134	17.5	17.5
43	25	TTCACCGGGGTGGTGCCCATcc tggtcgagctggacGGCGACGT AAACGGCCACAA	135	18.5	18.5
44	145	TGCACCACCGCAAGCTGCCcg tgccctggcccaccCTCGTGAC CACCTGACCTA	136	18.5	18.5
45	253	TTCAAGTCCGCCATGCCGAag gctacgtccaggagCGCACCAT CTTCTCAAGGA	137	18.5	18.5
46	454	TATATCATGGCCGACAAGCaga agaacggcatcaagGTGAACCTT CAAGATCCGCCA	138	18.5	18.5
47	139	TTCATCTGCACCACCGCAAGc tgcccgtgcccctggcCCACCCT CGTGACCACCCTGA	139	19.5	19.5
48	338	TGAAGTTCGAGGGCGACACCct ggtgaaccgcatogaGCTGAAG GGCATCGACTTCAA	140	19.5	19.5

Each of the 48 TALEN pairs was tested in human cells for its ability to disrupt the coding sequence of a chromosomally integrated EGFP reporter gene. In this assay, NHEJ-mediated repair of TALEN-induced breaks within the EGFP coding sequence led to loss of EGFP expression, which was quantitatively assessed using flow cytometry 2 and 5 days following transfection. (To ensure that activities of each active TALEN pair could be detected, we only targeted sites located at or upstream of nucleotide position 503 in the gene, a position we had previously shown would disrupt EGFP function when mutated with a zinc finger nuclease (ZFN) (Maeder et al., 2008, Mol. Cell 31:294-301).) Strikingly, all 48 TALEN pairs showed significant EGFP gene-disruption activities in this assay (FIG. 19A). The net percentage of EGFP-disrupted cells induced by TALENs on day 2 post-transfection ranged from 9.4% to 68.0%, levels comparable to the percentage disruption observed with four EGFP-targeted ZFN pairs originally made by the Oligomerized Pool Engineering (OPEN) method (Fig. 19A). These results demonstrate that TALENs containing as few as 8.5 TALE repeats possess significant nuclease

activities and provide a large-scale demonstration of the robustness of TALENs in human cells.

Interestingly, re-quantification of the percentage of EGFP-negative cells at day 5 post-transfection revealed that cells expressing shorter-length TALENs (such as those composed of 8.5 to 10.5 repeats) showed significant reductions in the percentage of EGFP-disrupted cells whereas those expressing longer TALENs did not (FIGs. 19A-B and 20A). One potential explanation for this effect is cellular toxicity associated with expression of shorter-length TALENs. Consistent with this hypothesis, in cells transfected with plasmids encoding shorter-length TALENs, greater reductions in the percentage of tdTomato-positive cells were observed from day 2 to day 5 post-transfection (FIG. 20D) (a tdTomato-encoding plasmid was co-transfected together with the TALEN expression plasmids on day 0). Taken together, our results suggest that although shorter-length TALENs are as active as longer-length TALENs, the former can cause greater cytotoxicity in human cells.

Our EGFP experiments also provided an opportunity to assess four of five computationally-derived design guidelines (Cermak et al., 2011, *Nucleic Acids Res.*, 39:e82). The guidelines proposed by Cermak are as follows:

1. The nucleotide just 5' to the first nucleotide of the half-site should be a thymine.
2. The first nucleotide of the half-site should not be a thymine.
3. The second nucleotide of the half-site should not be an adenosine.
4. The 3' most nucleotide in the target half-site should be a thymine.
5. The composition of each nucleotide within the target half-site should not vary from the observed percentage composition of naturally occurring binding sites by more than 2 standard deviations. The percentage composition of all naturally occurring TALE binding sites is: A = 31±16%, C = 37±13%, G = 9±8%, T = 22±10%. Hence, the nucleotide composition of potential TALE binding sites should be: A = 0% to 63%, C = 11% to 63%, G = 0% to 25% and T = 2% to 42%.

These guidelines have been implemented in the TALE-NT webserver (boglabx.plp.iastate.edu/TALENT/TALENT/) to assist users in identifying potential

TALEN target sites. All 48 of the sequences we targeted in EGFP did not meet one or more of these guidelines (however, note that all of our sites did meet the requirement for a 5' T). The ~100% success rate observed for these 48 sites demonstrates that TALENs can be readily obtained for target sequences that do not follow these guidelines. In addition, for each of the four design guidelines, we did not find any statistically significant correlation between guideline violation and the level of TALEN-induced mutagenesis on either day 2 or day 5 post-transfection. We also failed to find a significant correlation between the total number of guideline violations and the level of mutagenic TALEN activity. Thus, our results show that failure to meet four of the five previously described design guidelines when identifying potential TALEN target sites does not appear to adversely affect success rates or nuclease efficiencies.

Example 7. High-throughput Alteration of Endogenous Human Genes Using Assembled TALENs

Having established the robustness of the TALEN platform with a chromosomally integrated reporter gene, it was next determined whether this high success rate would also be observed with endogenous genes in human cells. To test this, the assembly method described in Example 5 was used to engineer TALEN pairs targeted to 96 different human genes: 78 genes implicated in human cancer (Vogelstein and Kinzler, 2004, Nat. Med., 10:789-799) and 18 genes involved in epigenetic regulation of gene expression (Table 7). For each gene, a TALEN pair was designed to cleave near the amino-terminal end of the protein coding sequence, although in a small number of cases the presence of repetitive sequences led us to target alternate sites in neighboring downstream exons or introns (Table 7). Guided by the results with the EGFP TALENs, TALENs composed of 14.5, 15.5, or 16.5 repeats were constructed that cleaved sites with 16, 17, 18, 19 or 21 bp spacer sequences. All of the target sites had a T at the 5' end of each half-site.

Table 7. Endogenous human gene sequences targeted by 96 pairs of TALENs

Target gene name	% NHEJ	Target site (half-sites in CAPS, spacer in lowercase, ATG underlined)	SEQ ID NO:	Length of LEFT half site (include 5' T)	Length of spacer	Length of RIGHT half site (include 5' T)	Gene Type
ABL1	22.5 ± 7.1	TACCTATTATTACT TTATggggcagcagcctgg aaAAGTACTTGGGG ACCAA	141.	16.5	17	15.5	Cancer
AKT2	14.1 ± 7.3	TGTGTCTTGGGATG AGTGggtcagtggttggtg CTCACAGGATGGCT GGCA	142.	16.5	16	16.5	Cancer
ALK	12.7 ± 2.9	TCCTGTGGCTCCTG CCGCtgccttccacggc AGCTGTGGGCTCCG GGA	143.	16.5	16	15.5	Cancer
APC	48.8 ± 9.8	TATGTACGCCTCCC TGGGctcgggtccggtcgcc CCTTTGCCCGCTTC TGTA	144.	16.5	16	16.5	Cancer
ATM	35.5 ± 15.6	TGAATTGGGATGCT GTTTtaggtattcttcaaa TTTATTTTACTGICT TTA	145.	16.5	18	16.5	Cancer
AXIN2	2.5 ± 0.6	TCCCTCACCATGAG TAGCgctatgttggtgacttG CCTCCCGGACCCCA GCA	146.	16.5	16	16.5	Cancer
BAX	14.7 ± 11.6	TGTGCGATCTCCAA GCACtgagggcagaaact cCCGGATCGGGCGC TGCCA	147.	16.5	16	16.5	Cancer
BCL6	14.9 ± 5.9	TTTTCAAGTGAAGA CAAAatggcctcggcct gACAGCTGTATCCA GTTCA	148.	16.5	16	16.5	Cancer
BMPRI A	50.4 ± 16.4	TACAATTGAACAAT GCCTcagctatacatltacat CAGATTATTGGGAG CCTA	149.	16.5	17	16.5	Cancer
BRCA1	44.5 ± 15.5	TCCGAAGCTGACAG ATGGgtattctttgacgggg GGTAGGGGCGGAA CCTGA	150.	16.5	16	16.5	Cancer
BRCA2	41.6 ± 10.5	TTAGACTTAGGTAA GTAAtgcaataggtagact GGGGAGAACTACA AACTA	151.	16.5	16	16.5	Cancer

CBX3	35.2 ± 22.6	TCTGCAATAAAAAA TGGCctccaacaaaactaca TTGGTAAGTTAATG AAAA	152.	16.5	16	16.5	Epigenetic
CBX8	13.5 ± 3.4	TGGAGCTTTCAGCG GTGGgggagcgggigtctc cgGCCGAAGCCCTC CTGAA	153.	16.5	17	15.5	Epigenetic
CCND1	40.5 ± 2.2	TGGAACACCAGCTC CTGTgctgcgaagtggaaac catCCGCCGCGCGTA CCCCGA	154.	16.5	19	16.5	Cancer
CDC73	36.3 ± 7.7	TGCTTAGCGTCTG CGACagtacaacatccagaa GAAGGAGATTGTG GTGAA	155.	16.5	16	16.5	Cancer
CDHI	none	TGCTGCAGGTACCC CGGAtccccctgacttgcgag GGACGCATTCGGGC CGCA	156.	16.5	16	16.5	Cancer
CDK4	21.5 ± 17.4	TCCCTTGATCTGAG AAtggetacctetegataTG AGCCAGTGGCTGA AA	157.	14.5	16	15.5	Cancer
CHD4	9.6 ± 0.1	TGGCGTCGGGCCTG GGCtccccctccccctctc GGCGGGCAGTGAG GAGGA	158.	15.5	17	16.5	Epigenetic
CHD7	11.4 ± 2.7	TGTGTTGGAAGAAG ATGGcagatccaggaatgat GAGTCTTTTTGGCG AGGA	159.	16.5	16	16.5	Epigenetic
CTNNB1	26.0 ± 8.1	TCCAGCGTGGACAA TGGtactcaaggttggTC ATTAAATCTTTAGT TA	160.	15.5	16	16.5	Cancer
CYLD	24.7 ± 2.3	TAATATCACAATGA GTTcaggcttatggagccaa gaAAAAGTCACTTC ACCCTA	161.	16.5	18	16.5	Cancer
DDB2	15.8 ± 7.2	TCACACGGAGGAC GCGatggctccaagaaac GCCCAGAAACCCA GAAGA	162.	14.5	16	16.5	Cancer
ERCC2	55.8 ± 12.7	TCCGGCCGGCGCCA TGAagtgagaaggggctg GGGGTCGCGCTCGC TA	163.	15.5	16	14.5	Cancer
ERCC5	none	TCCGGGATCGCCAT GGGAactcaatagaaaatcc tcaTCTTCTCACTTG TTCA	164.	16.5	19	16.5	Cancer

EWSR1	14.3 ± 8.2	TGGCGTCCACGGGT GAGTatggtggaactgcggt cGCGCCGGCGGTAG CCGGA	165.	16.5	17	16.5	Cancer
EXT1	9.5 ± 3.0	TGACCCAGGCAGG ACACA ^{tgcaggccaaaaa} cgTATTTTCATCCTG CTCTCA	166.	16.5	17	16.5	Cancer
EXT2	none	TTCCTCCCAGGGGG ATGTcctgcgctcagggtc CGGTGGTGGCCTGC GGCA	167.	16.5	16	16.5	Cancer
EZH2	41.3 ± 2.6	TGCTTTTAGAATAA TCATgggcccagactgggaa gAAATCTGAGAAGG GACCA	168.	16.5	16	16.5	Epigenetic
FANCA	9.7 ± 5.0	TAGGCGCCAAGGC CATGTccgactcgtgggtc ccGAACTCCGCCTC GGCCA	169.	16.5	16	16.5	Cancer
FANCC	23.7 ± 17.8	TGAAGGGACATCA CCTTTtctctttccaagatg GCTCAAGATTCAGT AGA	170.	16.5	17	15.5	Cancer
FANCE	none	TGCCCCGGCATGGC GACAccggacgcggggtc ccTGGGGCTGAGGG CGTGA	171.	16.5	17	16.5	Cancer
FANCF	46.0 ± 7.7	TTGCGCACCTCAT GGaatccctctgcagcaCC TGGATCGCTTTTCC GA	172.	14.5	16	16.5	Cancer
FANCG	26.9 ± 16.2	TCGGCCACCATGTC CCgccagaccacctgtGG GCTCCAGCTGCCTG GA	173.	14.5	16	16.5	Cancer
FES	12.6 ± 10.6	TCCCCAGAACAGCA CTATgggcttcttccgagc tGTGCAGCCCCCAG GGCCA	174.	16.5	18	16.5	Cancer
FGFR1	17.4 ± 6.2	TCTGCTCCCCACCG AGGAcctctgcatcaggca TGAATCCCAGGAGC CTA	175.	16.5	16	15.5	Cancer
FH	20.9 ± 11.8	TGTACCGAGCACTT CGGctctctgcgctcgcg tCCCCTCGTGCGGG CTCCA	176.	16.5	17	16.5	Cancer
FLCN	11.1 ± 4.4	TCTCCAAGGCACCA TGAAtgccatcgtggtctct gCCACTTCTGCGAG CTCCA	177.	16.5	18	16.5	Cancer

FLT3	none	TCCGGAGGCCATGC CGGCgttggcgcgcgacgg cggccaGCTGCCGCTG CTCGGTA	178.	16.5	21	15.5	Cancer
FLT4	9.9 ± 5.0	TGCAGCGGGGCGC CGCGCtgtgcctgcgactgt ggctCTGCCTGGGAC TCCTGGA	179.	16.5	19	16.5	Cancer
FOXO1	8.5 ± 1.1	TCACCATGGCCGAG GCGcctcaggtggtggagaT CGACCCGGACTTCG A	180.	15.5	16	14.5	Cancer
FOXO3	7.3 ± 2.3	TCTCCGCTCGAAGT GGAGctggaccggagtc gagCCCCAGAGCCGT CCGCGA	181.	16.5	18	16.5	Cancer
GLI1	21.5 ± 12.4	TCCTCTGAGACGCC ATGTtcaactegatgacccc ACCACCAATCAGTA GCTA	182.	16.5	16	16.5	Cancer
HDAC1	10.8 ± 3.0	TGGCGCAGACGCA GGGCacccggaggaaagtc tgTTACTACTACGAC GGTGA	183.	15.5	17	16.5	Epigen etic
HDAC2	4.2 ± 0.9	TGCGCTCACTCCC TGCGgectcctgaggtggtt gGTGGCCCCCTCCT CGCGA	184.	16.5	18	16.5	Epigen etic
HDAC6	21.4 ± 2.1	TCCTCAACTATGAC CTCAaccggccaggattcca CCACAACCAGGCA GCGAA	185.	16.5	16	16.5	Epigen etic
HMGA 2	3.0 ± 1.5	TGAGCGCACGCGGT GAGGgcgcggggcagccg tcCACTTCAGCCCAG GGACA	186.	16.5	16	16.5	Cancer
HOXA1 3	7.6 ± 3.1	TCCGTGCTCCTCCA CCCCcgctggatcgagcca cCGTCATGTTCTCT ACGA	187.	16.5	17	16.5	Cancer
HOXA9	6.4 ± 2.7	TGGGCACGGTGATG GCcaccactggggccctgG GCAACTACTACGTG GA	188.	14.5	16	15.5	Cancer
HOXC1 3	10.5 ± 0.3	TCCAGCAGATCATG TCATgacgacttcgctctcc tGCATCCACGCTGG CCGGA	189.	16.5	18	16.5	Cancer
HOXD1 1	none	TTGACGAGTGCGGC CAGagcgcagccagcatgta CCTGCCGGGCTGCG CCTA	190.	15.5	17	16.5	Cancer

HOXD1 3	none	TGCGGGCAGACGG CGGGGgcgccgggtggcgc cccGCCTCTTCCTCC TCCTCA	191.	16.5	17	16.5	Cancer
JAK2	44.9 ± 16.9	TCTGAAAAAGACTC TGCAtggaatggcctgct TACGATGACAGAA ATGGA	192.	16.5	16	16.5	Cancer
KIT	none	TACCGCGATGAGA GGCGCicgcgccctgg gattttCTCTGCGTTCT GCTCCTA	193.	16.5	19	16.5	Cancer
KRAS	9.4 ± 0.9	TGAAAATGACTGA ATATAaactgtgtagttg gaGCTGGTGGCGTA GGCAA	194.	16.5	17	15.5	Cancer
MAP2K 4	11.9 ± 7.1	TAGGGTCCCCGGCG CCAGgccaccggccgtca gCAGCATGCAGGGT AAGGA	195.	16.5	16	16.5	Cancer
MDM2	33.0 ± 20.2	TCCAAGCGCGAAA ACCCCGgatggtgaggag caggTACTGGCCCCGG CAGCGA	196.	16.5	17	15.5	Cancer
MET	40.4 ± 10.7	TTATTATTACATGG CTTTgccttactgaggttcA TCTTGTCTCTGGT CCA	197.	16.5	16	16.5	Cancer
MLH1	44.9 ± 6.3	TCTGGCGCCAAAAT GTCGttcgtggcaggggta TTCGGCGGCTGGAC GAGA	198.	16.5	16	16.5	Cancer
MSH2	27.5 ± 10.4	TGAGGAGGTTTCGA CATGgcggtgcagcgaag gAGACGCTGCAGTT GGAGA	199.	16.5	16	16.5	Cancer
MUTY H	24.9 ± 8.4	TCACTGTCGGCGGC CATGacaccgctcgtccc gcCTGAGTCGTCTGT GGGTA	200.	16.5	18	16.5	Cancer
MYC	13.4 ± 4.0	TGCTTAGACGCTGG ATTTitttcggtagtgaaA ACCAGGTAAGCAC CGAA	201.	16.5	16	16.5	Cancer
MYCL1	17.3 ± 0.6	TCCCGCAGGGAGC GGACAtggactacgacteg taCCAGCACTATTTT TACGA	202.	16.5	16	16.5	Cancer
MYCN	16.3 ± 11.6	TGCCGAGCTGCTCC ACgtccaccatgccggcA TGATCTGCAAGAAC CCA	203.	14.5	16	16.5	Cancer

NBN	46.3 ± 15.5	TGAGGAGCCGGAC CGAtgtggaaactgctgccC GCCGCGGGCCCCGG CA	204.	14.5	16	14.5	Cancer
NCOR1	29.6 ± 13.1	TCTTTACTGATAAT GTCAagttcatgittaccctcC CAACCAAGGAGCA TTCA	205.	16.5	16	16.5	Epigen etic
NCOR2	3.3 ± 0.6	TGGAGGGCCACTG AGCcccgtaccgcccaca CAGCCTTTCCTACC CA	206.	14.5	16	14.5	Epigen etic
NTRK1	none	TCGGCGCATGAAG GAGGTactcctcatttlegtt CTCTCTCTGTGC CCCA	207.	16.5	16	16.5	Cancer
PDGFR A	16.0 ± 4.3	TTGCGCTCGGGGCG GCCAtgtcggecegegagg iCGAGCGCCTAGTG TCGGA	208.	16.5	16	16.5	Cancer
PDGFR B	16.0 ± 3.2	TCTGCAGGACACCA TGCggttccgggtgctgatg CCAGCTCTGGCCCT CAAA	209.	16.5	16	16.5	Cancer
PHF8	22.2 ± 6.1	TGAGTACTCCGCCT CTACcccggctgaagcccg cCCCCGCGCCACC TATTA	210.	16.5	16	16.5	Epigen etic
PMS2	26.9 ± 9.5	TCGGGTGTTGCATC CATGgagcgagctgagagc tcgAGGTGAGCGGG GCTCGCA	211.	16.5	18	16.5	Cancer
PTCH1	27.5 ± 15.9	TGGAAGTCTTAAT AGaaacaggcttgaattGT GAGTCCGCGCTGCA	212.	14.5	16	14.5	Cancer
PTEN	31.5 ± 11.7	TCCCAGACATGACA GCCatcatcaaagatcgT TAGCAGAAACAAA AGGA	213.	15.5	16	16.5	Cancer
RARA	13.4 ± 6.1	TGGCATGGCCAGCA ACAGcagctcctgcccac acCTGGGGGCGGGC ACCTCA	214.	16.5	17	16.5	Cancer
RBBP5	15.7 ± 9.5	TGCTGGGTGAGAA GGGCtgtggtcggttttaga GAAGCGTTGGGTAC TGGA	215.	15.5	17	16.5	Epigen etic
RECQL 4	22.1 ± 16.2	TGCGGGACGTGCG GGAGCggctgcaggcgtg ggaGCGCGCGTCCG ACGGCA	216.	16.5	16	16.5	Cancer

REST	none	TCAGAATACAGTTA TGGCcaccaggtaatggg gCAGTCTTCTGGAG GAGGA	217.	16.5	16	16.5	Epigen etic
RET	5.4 ± 1.8	TGAGTTCTGCCGGC CGCCggctcccaggggc caGGGCGAAGTTGG CGCCGA	218.	16.5	17	16.5	Cancer
RNF2	none	TTCTTTATTTCCAG CAATgtctcaggctgtgcag ACAAACGGAACTC AACCA	219.	16.5	16	16.5	Epigen etic
RUNX1	25.1 ± 6.9	TTCAGGAGGAAGC GATGGcttcagacagcatat tTGAGTCATTTCCCTT CGTA	220.	16.5	16	16.5	Epigen etic
SDHB	36.4 ± 19.2	TCTCCTTGAGGGCGC CGGTtcccggccacaacct TGCCGGAGCCTGCC TGCA	221.	16.5	16	16.5	Cancer
SDHC	13.7 ± 3.4	TGTTGCTGAGGTGA CTTCagtgggactgggagtt ggtGCCTGCGGCCCT CCGGA	222.	16.5	19	15.5	Cancer
SDHD	42.0 ± 7.8	TCAGGAACGAGAT GGCGGttctctggaggctga gtGCCGTTTGCGGTG CCCTA	223.	16.5	17	16.5	Cancer
SETDB 1	33.5 ± 6.1	TGCAGAGGACAAA AGCATgtctccctctctgg gTGCATTGGTTTGG ATGCA	224.	16.5	16	16.5	Epigen etic
SIRT6	43.3 ± 3.1	TTACGCGGGCGGGC TGTCgccgtacgggacaa gggCAAGTGCGGCC TCCCCGA	225.	16.5	18	16.5	Epigen etic
SMAD2	3.9 ± 1.6	TTTGGTAAGAACAT GTCCtccatettgccattcac GCCGCCAGTTGTGA AGA	226.	16.5	17	15.5	Cancer
SS18	31.4 ± 7.9	TGGTGACGGCGGC AACATgtctgtggcttgcg ggCCCCGAGGCAGC GAGGCA	227.	16.5	17	16.5	Cancer
SUZ12	13.1 ± 0.4	TGGCGCTCAGAAG CAcggcgggtggggaggg GGCGGCTCGGGGC CCA	228.	14.5	16	14.5	Epigen etic
TFE3	17.3 ± 2.4	TCATGTCTCATGCG GCCGaccagctcgggatg gCGTAGAGGCCAGC GCGGA	229.	16.5	16	16.5	Cancer

TGFBR 2	none	TCGGGGGCTGCTCA GGGGcctgtggccgctgca caTCGTCTGTGGAC GCGTA	230.	16.5	17	16.5	Cancer
TLX3	none	TTCCGCCCCGCCAG GATGgaggcgcccgcag cgcGCAGACCCCGC ACCCGCA	231.	16.5	17	16.5	Cancer
TP53	19.9 ± 3.6	TTGCCGTCCCAAGC AATGgatgattgatgtg CCCGGACGATATTG AACA	232.	16.5	17	16.5	Cancer
TSC2	30.7 ± 22.7	TCCTGGTCCACCAT GGCcaaaccaacaagcaaa gATTCAGGCTTGAA GGAGA	233.	15.5	17	16.5	Cancer
VHL	19.4 ± 1.1	TCTGGATCGCGGAG GGAAtgccccggagggeg gaGAACTGGGACGA GGCCGA	234.	16.5	16	16.5	Cancer
XPA	12.9 ± 2.2	TGGGCCAGAGATG GCGGCggccgacggggct ttgCCGGAGGCGGCG GCTTA	235.	16.5	16	16.5	Cancer
XPC	31.4 ± 4.2	TGCCCAGACAAGC AACATggctcggaaacgc gggccGGCGGGGAG CCGCGGGGA	236.	16.5	19	16.5	Cancer

The abilities of the 96 TALEN pairs to introduce NHEJ-mediated insertion or deletion (indel) mutations at their intended endogenous gene targets were tested in cultured human cells using a slightly modified version of a previously described T7 Endonuclease I (T7EI) assay (Mussolino et al., 2011, *Nucleic Acids Res.*, 39:9283-93; Kim et al., 2009, *Genome Res.*, 19:1279-88). With this T7EI assay, 83 of the 96 TALEN pairs showed evidence of NHEJ-mediated mutagenesis at their intended endogenous gene target sites, an overall success rate of ~86% (Table 7). The efficiencies of TALEN-induced mutagenesis we observed ranged from 2.5% to 55.8% with a mean of 22.5%. To provide molecular confirmation of the mutations we identified by T7EI assay, we sequenced target loci for 11 different TALEN pairs that induced varying efficiencies of mutagenesis (FIGs. 21A-D). As expected, this sequencing revealed indels at the expected target gene sites with frequencies similar to those determined by the T7EI assays.

The nucleotide and amino acid sequences for 14 of the 96 pairs of TALENs targeted to the endogenous human genes in Table 7 are presented below. Each TALEN monomer is presented as follows:

(1) A header with information presented in the format: Gene target_Left or Right monomer_Target DNA site shown 5' to 3'_TALE repeat monomers and 0.5 repeat plasmid used with code as shown in Table 4.

(2) DNA sequence encoding the N-terminal part of the TALE required for activity, the TALE repeat array, the C-terminal 0.5 TALE repeat domain, and the C-terminal 63 amino acids required for activity from a NheI site to a BamHI site. This sequence is present in the "Vector Sequence" plasmid shown below, taking the place of the underlined X's flanked by NheI and BamHI sites

(3) Amino acid sequences the N-terminal part of the TALE required for activity, the TALE repeat array, the C-terminal 0.5 TALE repeat domain, and the C-terminal 63 amino acids required for activity shown from the start of translation (located just 3' to the NheI site and including an N-terminal FLAG epitope tag) to a Gly-Ser sequence (encoded by the BamHI site) that serves as a linker from the TALE repeat array to the FokI cleavage domain.

VECTOR SEQUENCE

20 GACGGATCGGGAGATCTCCCGATCCCCTATGGTCGACTCTCAGTACAATCTGCTCTGATGCCGCATAGTTA
AGCCAGTATCTGCTCCCTGCTTGTGTGTTGGAGGTCGCTGAGTAGTGCCGCGAGCAAAATTTAAGCTACAAC
AAGGCAAGGCTTGACCGACAATTGCATGAAGAATCTGCTTAGGGTTAGGCGTTTTGCGCTGCTTCGCGATG
TACGGGCCAGATATACGCGTTGACATTGATTATTGACTAGTTATTAATAGTAATCAATTACGGGGTCATTA
25 GTTCATAGCCCATATATGGAGTTCGCGGTTACATAAATTACGGTAAATGGCCCGCCTGGCTGACCGCCCAA
CGACCCCGCCCAATTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGACTTTCATTGAC
GTCAATGGGTGGACTATTTACGGTAAACTGCCCACTTGGCAGTACATCAAGTGTATCATATGCCAAGTACG
CCCCCTATTGACGTCAATGACGGTAAATGGCCCGCCTGGCATTATGCCAGTACATGACCTTATGGGACTT
TCCTACTTGGCAGTACATCTACGTATTAGTCATCGCTATTACCATGGTGATGCGGTTTTGGCAGTACATCA
30 ATGGGCGTGGATAGCGGTTTGACTCACGGGGATTCCAAGTCTCCACCCCAATTGACGTCAATGGGAGTTG
TTTTGGCACCAAAATCAACGGGACTTTCCAAAATGTCGTAAACAACCTCCGCCCAATTGACGCAAAATGGGCGG
TAGGCGTGACGGTGGGAGGTCTATATAAGCAGAGCTCTCTGGCTAACTAGAGAACCCACTGCTTACTGGC
TTATCGAAAATTAACGACTCACTATAGGGAGACCCAAGCTGGCTAGCXXXXXXXXXXGGATCCCAACTAG
TCAAAAGTGAAGTGGAGGAGAAGAAATCTGAACCTCGTCATAAAATTGAAATATGTGCCTCATGAATATATT
35 GAATTAATTGAAATGCCAGAAATCCACTCAGGATAGAATTCTTGAAATGAAGGTAATGGAATTTTTTAT
GAAAGTTTATGGATATAGAGGTAAACATTTGGGTGGATCAAGGAAACCGGACGGAGCAATTTACTGTGCG
GATCTCCTATTGATTACGGTGTGATCGTGGATACTAAAGCTTATAGCGGAGGTTATAATCTGCCAATTTGGC
CAAGCAGATGAAATGCAACGATATGTCGAAGAAAATCAAACACGAAACAAACATATCAACCCTAATGAATG

GTGAAAGTCTATCCATCTTCTGTAACGGAATTTAAGTTTTTATTTGTGAGTGGTCACTTTAAAGGAACT
 ACAAAGCTCAGCTTACACGATTAATCATATCACTAATTGTAATGGAGCTGTTCTTAGTG TAGAAGAGCTT
 TTAATTGGTGGAGAAATGATTAAGCCGGCACATTAACCTTAGAGGAAGTCAGACGGAATTTAATAACGG
 CGAGATAAACTTTAAGGGCCCTTCGAAGGTAAGCCTATCCCTAACCTCTCCTCGGTCTCGATTCTACGC
 5 GTACCGGTCACTATCACCATCACCATTGAGTTTTAAACCCGCTGATCAGCCTCGACTGTGCCTTCTAGTTGC
 CAGCCATCTGTTGTTTGCCTTCCCCGTCCTTCCCTTGACCCTGGAAGGTGCCACTCCCCTGCTTTT
 CTAATAAAATGAGGAAATGTCATCGCATGTCTGAGTAGGTGTCATTCTATTCTGGGGGGTGGGGTGGGGC
 AGGACAGCAAGGGGGAGGATTGGGAAGACAATAGCAGGCATGCTGGGGATGCGGTGGGCTCTATGGCTTCT
 GAGGCGAAAGAACCAGCTGGGGCTTAGGGGTATCCCCACGCGCCCTGTAGCGGCGCATTAAGCGCGGC
 10 GGGTGTGGTGGTTACGCGCAGCGTGACCCTACACTGCCAGCGCCCTAGCGCCGCTCCTTTCGCTTCT
 TCCCTTCCCTTCTCGCCACGTTCCCGGCTTCCCGCTCAAGCTCTAAATCGGGGCATCCCTTAGGGTTC
 CGATTTAGTGCTTACGGCACCTCGACCCAAAAAACTTGATTAGGGTGATGGTTCACGTAGTGGGCCATC
 GCCCTGATAGCGGTTTTTCGCCCTTTCAGCTTGAGTCCACGTTCTTAAATAGTGGACTCTGTCCAAA
 CTGGAACAACACTCAACCCTATCTCGGTCTATTCTTTGATTTATAAGGGATTTGGGGATTTCCGCCAT
 15 TGGTTAAAAATGAGCTGATTTAACA AAAATTTAACGCGAATTAATCTGTGGAATGTGTGCAGTTAGGG
 TGTGGAAGTCCCCAGGCTCCCCAGGCAGGCAGAAGTATGCAAAGCATGCATCTCAATTAGTCAGCAACCA
 GGTGTGGAAGTCCCCAGGCTCCCCAGCAGGCAGAAGTATGCAAAGCATGCATCTCAATTAGTCAGCAACC
 ATAGTCCC GCCCTAACTCCGCCATCCCGCCCTAACTCCGCCAGTTCGCCCATTTCCGCCCATGG
 CTGACTAATTTTTTTATTTATGTCAGAGGCCGAGGCCCTCTGCCTCTGAGCTATTCCAGAAGTAGTGAG
 20 GAGGCTTTTTGGAGGCTTAGGCTTTTGCAAAAAGCTCCCGGAGCTTGATATCCATTTTCGGATCTGAT
 CAGCACGTGTTGACAATTAATCATCGGCATAGTATATCGGCATAGTATAATACGACAAGGTGAGGAACTAA
 ACCATGGCCAAGCCTTGTCTCAAGAAGAATCCACCCTCATGAAAGAGCAACGGCTACAATCAACAGCAT
 CCCCATCTCTGAAGACTACAGCGTCGCCAGCGCAGCTCTCTTAGCGACGGCCGATCTTCACTGGTGTCA
 ATGTATATCATTACTGGGGACCTTGTGCAGAACTCGTGGTGTGGGCACTGCTGCTGCTGCGGCAGCT
 25 GGCAACCTGACTTGTATCGTCGCGATCGGAAATGAGAACAGGGCATCTTGAGCCCTGCGGACGGTGTGCG
 ACAGGTGCTTCTCGATCTGCATCCTGGGATCAAAGCGATAGTGAAGGACAGTGAAGGACAGCCGAGCAG
 TTGGGATTCGTGAATTTGCTGCCCTCTGGTTATGTGTGGGAGGGCTAAGCACTTCGTGGCCGAGGAGCAGGA
 CTGACAGGTGCTACGAGATTTGATTTCCAGTCCAGCCGCTTCTATGAAAGGTTGGGCTTCGGAATCGTTTTCC
 GGGACGCCGCTGGATGATCCTCCAGCGGGGATCTCATGCTGGAGTCTTCGCCACCCCACTTGTTF
 30 ATTGACGCTTATAATGGTTACAAATAAAGCAATAGCATCACAAATTCACAAATAAAGCATTTTTTCTACT
 GCATTTAGTTGTGGTTTGTCCAAACTCATCAATGATCTTATCATGTCTGTATACCGTCGACCTCTAGCT
 AGAGCTTGGCGTAATCATGGTCAATAGCTTTTCCGTGTGAAATTTGTTATCCGCTCACAAATCCACACAAC
 ATACGAGCCGGAAGCATAAAGTGTAAAGCCTGGGGTGCCTAATGAGTGAGCTAACTCACATTAATTGCGTT
 GCGCTCACTGCCGCTTCCAGTCCGGAACCTGCTGTCAGCTGCATTAATGAATCGGCCAACCGCGGG
 35 GGAGAGCGGTTTTGCGTATTGGGCGCTTCCGCTTCTCGCTCACTGACTCGTGCCTCGGTCGTTCCGG
 CTGCGCGGAGCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTATCCACAGAATCAGGGGATAACGCAGG
 AAAGAACATGTGAGCAAAAGGCCAGCAAAAGGCCAGGAACCGTAAAAAGGCCGCTTGTGCGGTTTTTCC
 ATAGGCTCCGCCCTTCCAGTCCGAGCATCAAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGA
 CTATAAAGATAACCAGCGTTTTCCCTTGGAAAGCTCCCTCGTGCCTCTCCTGTTCCGACCTGCCGCTTAC
 40 CGGATACTGTCCGCTTTCTCCCTTCCGGAAGCGTGGCGTTTTCTCAATGCTCAGCTGTAGGTATCTCA
 GTTCGGTGTAGGTCGTTCCGTTCCAAGCTGGGCTGTGTGCACGAACCCCGTTACGCCGACCGCTGCGCC
 TTATCCGTAACATAFCGTCTTGAGTCCAAACCCGTAAGACACGACTTATCGCCACTGGCAGCAGCCACTGG
 TAACAGGATTAGCAGAGCGAGGTATGTAGCGGCTGCTACAGAGTCTTGAAGTGGTGGCCTAACTACGGCT
 ACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCTGAAGCCAGTTACCTTCGGAAAAAGAGTTGGTAGC
 45 TCTTGATCCGGCAAAACAAACCACCGCTGGTAGCGGTGGTTTTTTTGTGTGCAAGCAGCAGATTACGCGCAG
 AAAAAAGGATCTCAAGAAGATCCTTTGATCTTTCTACGGGTCTGACGCTCAGTGGAAACGAAAACCTCAC
 GTTAAGGGATTTGGTATGAGATTATCAAAAAGGATCTTACCTAGATCCTTTTAAATTAATAAAGT
 TTTAAATCAATCTAAAGTATATATGAGTAACTTGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCACC

TATCTCAGCGATCTGTCTATTTTCGTTTCATCCATAGTTGCCTGACTCCCCGTCGTGTAGATAACTACGATAC
GGGAGGGCTTACCACTCTGGCCCCAGTGCTGCAATGATACCGCGAGACCCACGCTCACCGGCTCCAGATTTA
TCAGCAATAAACCAGCCAGCCGGAAGGGCCGAGCGCAGAAGTGGTCCGCAACTTTATCCGCTCCATCCA
5 GTCTATTAATTGTTGCCGGGAAGCTAGAGTAAGTAGTTCGCCAGTTAATAGTTTGGCACAACGTTGTTGCCA
TTGCTACAGGCATCGTGGTGTACGCTCGTTCGTTGGTATGGCTTCATTCAGCTCCGGTTCCCAACGATCA
AGGCGAGTTACATGATCCCCATGTTGTGCAAAAAAGCGGTTAGCTCCTTCGGTCTCCGATCGTTGTCAG
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CCGTAAGATGCTTTTCTGTGACTGGTGAGTACTCAACCAAGTCATTCTGAGAATAGTGTATGCGGCGACCG
10 AGTTGCTCTTGCCCGCGTCAATACGGGATAATACCGGCCACATAGCAGAACTTAAAAGTGCTCATCAT
TGGAAAACGTTCTTCGGGGCGAAAACCTCAAGGATCTTACCGCTGTTGAGATCCAGTTCGATGTAACCCA
CTCGTGCACCCAACCTGATCTTCAGCATCTTTACTTTCACCAGCGTTTCTGGGTGAGCAAAAACAGGAAGG
CAAAATGCCGCAAAAAGGGAATAAGGGCGACACGAAATGTTGAATACTCATACTCTTCTTTTCAATA
TTATTGAAGCATTATCAGGGTTATTGTCTCATGAGCGGATACATATTTGAATGTATTTAGAAAAATAAC
15 AAATAGGGGTTCCGCGCACATTTCCCCGAAAAGTGCCACCTGACGTC SEQ ID NO: 244

TALE REPEAT SEQUENCES

Target	SEQUENCE	SEQ ID NO:	SEQUENCE	SEQ ID NO:
>APC Left TATGTACGCC TCCCTGGG T AL/006/015 /019/025/0 25/012/019 /022/027/0 15/017/022 /027/015/0 19/024/JDS 74/ ('TATGTACG CCTCCCTGGG ' disclosed as SEQ ID NO: 412)	GCTAGCaccATGGACTACAAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGCCCCCAAGAAGAAG AGGAAGGTGGGCATTCACCGGGGTACCTAT GGTGGACTTGAGGACACTCGGTATTTCGCAAC AGCAACAGGAGAAAAATCAAGCCTAAGGTCAGG AGCACCGTCGCGCAACACCACGAGGCGCTTGT GGGGCATGGCTTCACTCATCGCATATTGTCG CGCTTTCACAGCACCCCGCGGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCGAAGCCACGACAGGCAATTGTAG GGTTCGGTAAACAGTGGTCGGGAGCGGAGCA CTTGAGGCGCTGCTGACTGTGGCGGTGAGCT TAGGGGGCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGGGGGAGTA ACAGCGGTAGAGGCAGTGCACGCTGGCCAA TGCGCTCACCGGGCCCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGCGTGAACATT GGGGAAAGCAAGCCCTGGAACCCGTGCAAAG GTTGTTGCCGGTCCCTTGTCAAGACCACGGCC TTACACCGGACCAAGTCGTGCCATTGCAAGC AATGGGGGTGGCAAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGAATAACAATGGAGGGAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCCACGGTTTGACGCTGCACAAGTGGTC GCCATCGCCTCGAATGGCGGGTAAGCAGGC GCTGGAACAGTACAGCGCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG	245.	ASTMDYKDHDGDYKDHDIDYKDDDDKMAPKKRKGVI HRGVPMVDLRILGYSQQQEKIKPKVRSTVAQHHEAL VGHGFTHAHIVALSQHPAALGTVAVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQLDT GQLLKIARGGVTAVEAVHAWRNALTGAPLNLTPDQV VAIASNIGGKQALETVQRLLPVLCDHGLTPEQVVAI ASNGGGKQALETVQRLLPVLCAHGLTPDQVVAIANN NGGKQALETVQRLLPVLCAHGLTPAQVVAIASNGGG KQALETVQRLLPVLCDHGLTPDQVVAIASNIGGKQA LETVQRLLPVLCDHGLTPEQVVAIASHDGGKQALET VQRLLPVLCAHGLTPDQVVAIANNGGKQALETVQR LLPVLCAHGLTPAQVVAIASHDGGKQALETVQRLLP VLCQDHGLTPDQVVAIASHDGGKQALETVQRLLPVL CDHGLTPEQVVAIASNGGGKQALETVQRLLPVLCAH GLTPDQVVAIASHDGGKQALETVQRLLPVLCAHGLT PAQVVAIASHDGGKQALETVQRLLPVLCDHGLTPDQ VVAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVA IASNGGGKQALETVQRLLPVLCAHGLTPDQVVAIAN NNGGKQALETVQRLLPVLCAHGLTPAQVVAIANNG GKQALETVQRLLPVLCDHGLTPEQVVAIANNGGRP ALESIVAQLSRPDALAALNDHLVALACLGGRPALD AVKKGLPHAPALIKRTNRRIPERTSHRVAGS	246.

<p>GTAGTCGCAATCGCGTCGAACATTGGGGGAAA GCAAGCCCTGGAAACCGTGCAAAGGTTGTTGC CGGTCCCTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCATCCCACGACGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCCAGTTCTGTCAAGCCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCCAATAA CAATGGAGGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCGTGTGTGTCAAGCCCAC GGTTTGACGCCCTGCACAAGTGGTCGCCATCGC CAGCCATGATGGCGTAAGCAGGCGCTGGAAA CAGTACAGCGCCTGCTGCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGTAGTCGC AATCGCGTCACATGACGGGGAAAGCAAGCCC TGGAACCGTGCAAAGGTTGTTGCCGGTCTT TGTCAAGACCACGGCCTTACACCGGAGCAAGT CGTGGCCATTGCAAGCAATGGGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCCACGGGCTGACTCCGA TCAAGTTGTAGCGATTGCGTCGCATGACGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCCGTGTGTGTCAAGCCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATGCCAGCCATG ATGGCGGTAAGCAGGCGCTGAAACAGTACAG CGCTGCTGCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGT CACATGACGGGGAAAGCAAGCCCTGAAACC GTGCAAAGGTTGTGCCGGTCTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCAAGCAATGGGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTTCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCCAATAACAATGGAGGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAAGCCCACGGTTTGACGCTGCAC AAGTGGTCCCATGCCAACAACAACGGCGGT AAGCAGGCGCTGGAAACAGTACAGCGCCTGCT</p>		
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	<p>GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCGCCATTGCTAATAATAAC GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCGGTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGTCTTGGTGGACGACCCCGCTCGATGCAGT CAAAAGGGTCTGCCTCATGCTCCCGCATTGA TCAAAGAACCAACCGCGGATTCCCGAGAGA ACTTCCCATCGAGTCGCGGGATCC</p>			
<p>>APC_Right _TACAGAAGC GGGCAAAGG TAL/006/01 2/016/024/ 026/011/01 9/022/029/ 014/019/02 2/026/011/ 016/024/JD S74/ ('TACAGAAG CGGGCAAAGG ' disclosed as SEQ ID NO: 413)</p>	<p>GCTAGCaccATGGACTACAAGACCATGACGG TCGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGCCCCCAAGAAGAAG AGGAAGGTGGGCATTACC CGCGGGTACCTAT GGTGGACTTGAGGACACTCGGTTATTCCGAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCGTCCGCGAACACCACGAGGCGCTTGT GGGGCATGGCTTCACTCATGCGCATATTGTCG CGCTTTCACAGCACCCCTGCGGGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCCGAAGCCACGACAGGCAATTGTAG GGGTCGGTAAACAGTGGTGGGAGCGCGAGCA CTTGAGCGCTGTGACTGTGGCGGTTGAGCT TAGGGGGCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCCCGAAGAGAGGGGGAGTA ACAGCGGTAGAGGCAGTGCACGCTGCGCGCAA TGCGCTCACCGGGGCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGCGTGAACATT GGGGAAAGCAAGCCCTGGAAACCGTGCAAAG GTTGTTCGCGGCTCCTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCTGGCCATTGCATCC CACGACGGTGGCAAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGTCGAACATTGGAGGGAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCCACGGTTTGACGCTGCACAAGTGGTC GCCATCGCCAACACACGCGGTAAGCAGGC</p>	<p>247.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDDKMAPKKRKGVI HRGVFVMDLRTLGSQQQEKIKPKVRSVAQHHEAL VGHGFTHAHIVALSQHPAALSTVAVKYQDMIAALPEA THEAIVGVKQWSGARALEALLTVAGELRGPPLQDLD GQLLKIARKGGVTAVEAVHAWRNALTGAPLNLPDQV VAIASNIGGKQALETVQRLLPVLCDHGLTPEQVVAI ASHDGKQALETVQRLLPVLCAHGLTPDQVVAIASN IGGKQALETVQRLLPVLCAHGLTPAQVVAIANNNGG KQALETVQRLLPVLCDHGLTPDQVVAIASNIGGKQ LETVQRLLPVLCDHGLTPEQVVAIASNIGGKQALET VQRLLPVLCAHGLTPDQVVAIANNNGGKQALETVQR LLPVLCAHGLTPAQVVAIASHDGKQALETVQRLLP VLCQDGLTPDQVVAIANNNGGKQALETVQRLLPVL CDHGLTPEQVVAIANNNGGKQALETVQRLLPVLCAH GLTPDQVVAIANNNGGKQALETVQRLLPVLCAHGLT PAQVVAIASHDGKQALETVQRLLPVLCDHGLTPDQ VVAIASNIGGKQALETVQRLLPVLCDHGLTPEQVVA IASNIGGKQALETVQRLLPVLCAHGLTPDQVVAIAS NIGGKQALETVQRLLPVLCAHGLTPAQVVAIANNNG GKQALETVQRLLPVLCDHGLTPEQVVAIANNNGGRP ALESIVAQLSRPDPALAALTNQHLVALACLGGRPALD AVKKGPHAPALIKRTNRRIPERTSHRVAGS</p>	<p>248.</p>

<p>GCTGAAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCGAACATTGGGGAAA GCAAGCCCTGGAAACCGTGCAAAGTTGTTGC CGGTCTTTGTCAAGACCACGGCCTACACCG GAGCAAGTCGTGGCCATTGCAAGCAACATCGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTTCTGTCAAGCCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGAATAA CAATGGAGGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCGTGTGTGTCAAGCCAC GGTTTGACGCCCTGCACAAGTGGTCGCCATCGC CAGCCATGATGGCGTAAGCAGGCGTGGAAA CAGTACAGCGCCTGCTGCCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGAACAATAATGGGGAAAGCAAGCCC TGGAACCGTGCAAAGTTGTTGCCGGTCCTT TGTCAAGACCACGGCCTTACACCGGAGCAAGT CGTGGCCATTGCAATAATAACGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTGTGTCAAGCCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCCAATAACAATGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCCGTGTGTGTCAAGCCCACGGTTGAC GCCTGCACAAGTGGTCGCCATCGCCAGCCATG ATGGCGGTAAGCAGGCGCTGGAAACAGTACAG CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTACTCGCAATCGCGT CGAACATTGGGGAAAGCAAGCCCTGGAACC GTGCAAAGTTGTTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCAAGCAACATCGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTTCTGTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGTCGAACATTGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAAGCCCACGGTTGACGCTGCAC</p>		
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	<p>AAGTGGTCGCCATGCCAACAACAACGGCGGT AAGCAGGCGCTGGAAACAGTACAGCGCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCGCATTGCTAATAATAAC GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCGCGTTGGCTG CGTTAACGAATGACCMCTGGTGGCGTTGGCA TGTCTTGGTGGACGACCCGCGCTCGATGCAGT CAAAAGGGTCTGCCTCATGCTCCCGCATTGA TCAAAGAACAACCGCGGATCCCGAGAGA ACTTCCCATCGAGTCGCGGGATCC</p>			
<p>>BRCA1_Lef t_TCCGAAGC TGACAGATGG TAL/007/0 12/019/021 /026/014/0 17/025/029 /011/017/0 21/029/011 /020/024/J DS74/ ('TCCGAAGC TGACAGATGG disclosed as SEQ ID NO: 414)</p>	<p>GCTAGCaccatGGACTACAAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGCCCCCAAGAAGAAG AGGAAGGTGGCATTACCCGCGGGTACCTAT GGTGACTTGGAGCACTCGGTTATTGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTGAGG AGCACCGTCGCGCAACACCACGAGGCGCTTGT GGGGCATGGCTTCACTCATGCGCATATTGTCG CGCTTTCACAGCACCTTCCGCGCTTGGGACG GTGGCTGTCAATAACCAAGATATGATTGCGGC CCTGCCGAAGCCACGACGAGGCAATTGTAG GGGTCGGTAAACACTGGTGGGAGCGCGAGCA CTTGAGGCGCTGCTGACTGTGGCGGGTGGCT TAGGGGGCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGAGGGGGAGTA ACAGCGGTAGAGGCAGTGCACGCTGGCGCAA TGCGCTCACCAGGGCCCTTGAACCTGACCC CAGACCAGGTAGTCGAATCGCGTCACATGAC GGGGAAAGCAAGCCCTGGAACCGTGCAAAG GTTGTGCCGGTCTTTGTCAAGACCAGGGCC TTACACCGGAGCAAGTCTGTCGATTCGATCC CACGAGGTTGGCAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCACTTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGAATAACAATGGAGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTCCCGTGTGTGTC</p>	<p>249.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDDKMAPKKRKGVI HRGVPMVLDLRTLGSQQQEKIKPKVRSTVAQHHEAL VGHGFTHAHTVALSQHPAALGTVAVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQDLD GQLLKIARGGVTAVEAVHAWRNALTGAPLNLTDPQV VAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVAI ASHDGGKQALETVQRLLPVLCAHGLTPDQVVAIANN NGGKQALETVQRLLPVLCAHGLTPAQVVAIASNIGG KQALETVQRLLPVLCDHGLTPDQVVAIASNIGGKQA LETVQRLLPVLCDHGLTPEQVVAIANNGGKQALET VQRLLPVLCAHGLTPDQVVAIASHDGGKQALETVQR LLPVLCAHGLTPAQVVAIASNGGKQALETVQRLLP VLCQDHLTPDQVVAIANNGGKQALETVQRLLPVL CDHGLTPEQVVAIASNIGGKQALETVQRLLPVLCAH GLTPDQVVAIASHDGGKQALETVQRLLPVLCAHGLT PAQVVAIASNIGGKQALETVQRLLPVLCDHGLTPDQ VVAIANNGGKQALETVQRLLPVLCDHGLTPEQVVA IASNIGGKQALETVQRLLPVLCAHGLTPDQVVAIAS NGGKQALETVQRLLPVLCAHGLTPAQVVAIANNG GKQALETVQRLLPVLCDHGLTPEQVVAIANNGGRP ALESIVAQLSRPDPALAAALNDHLVALACLGGRPALD AVKKGLPHAPALIKRTRRRIPERTSHRVAGS</p>	<p>250.</p>

<p>AAGCCCACGGTTTGACGCCTGCACAAGTGGTC GCCATCGCCTCCAATATTGGCGGTAAAGCAGGC GCTGGAAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCGAACATTGGGGAAA GCAAGCCCTGGAAACCGTGCAAAGGTTGTTGC CGGTCTTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCAAATAATAACGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTTCTCTGTCAAGCCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGTCGCA TGACGGAGGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCGTGTGTGTCAAGCCCAC GGTTTGACGCCTGCACAAGTGGTCGCCATCGC CTCGAATGGCGCGGTAAGCAGGCGTGGAAA CAGTACAGCGCCTGCTGCCGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGGAAACATAATGGGGAAAGCAAGCCC TGGAAACCGTGCAAAGGTTGTTGCCGGTCTTT TGTCAAGACCACGGCCTTACACCGGAGCAAGT CGTGGCCATTGCAAGCAACATCGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCGCATGACGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCCGTGTGTGTCAAGCCCACGGTTGAC GCCTGCACAAGTGGTGCATCGCCTCCAATA TTGGCGGTAAGCAGGCGCTGAAACAGTACAG CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGA ACAATAATGGGGAAAGCAAGCCCTGAAACC GTGCAAAGGTTGTTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCAAGCAACATCGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTTCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGTCCAACGGTGGAGGAAACAA</p>	
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	<p>GCATTGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAAGCCCACGGTTTGACGCCTGCAC AAGTGGTCCCATCGCCAACAACAACGGCGGT AAGCAGGCGCTGGAAACAGTACAGCGCCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCGCCATTGCTAATAATAAC GGAGGACGGCCAGCCTTGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCAGCGTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGTCTTGGTGGACGACCCGCGCTCGATGCAGT CAAAAGGGTCTGCCCTCATGCTCCCGCATTGA TCAAAGAACCAACCGCGGATTCCCAGAGAGA ACTTCCCATCGAGTCGCGGGATCC</p>			
<p>>BRCA1_Rig ht_TCAGGTT CCGCCCCCTAC C_TAL/007/ 011/019/02 4/030/015/ 017/022/02 9/012/017/ 022/027/01 5/016/022/ JDS71/ ('TCAGGTTT CGCCCCCTACC ' disclosed as SEQ ID N: 415)</p>	<p>GCTAGCaccattGGACTACAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGCATTACCCGCGGGTACCTAT GGTGGACTTGAGGACACTCGGTATTTCGCAAC AGCAACAGGAGAAAAATCAAGCCTAAGGTCAGG AGCACCGTCCGGCAACACCACGAGGCGCTTGT GGGGCATGGCTTCACTCATGCGCATATTGTCG CGCTTTCACAGCACCCCTGCGGCGCTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCCGAAGCCACGACGAGGCAATTGTAG GGGTCGGTAACAGTGTGCGGAGCGCGAGCA CTTGAGGCGCTGCTGACTGTGGCGGGTGAGCT TAGGGGGCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGAGGGGAGTA ACAGCGGTAGAGGAGTGCACGCCCTGGCGCAA TCGCCTCACCGGGGCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGGTCACATGAC GGGGAAAGCAGCCCTGGAAACCGTGCAAAG GTTGTGCGCGTCCCTTGTCAAGACCACGGCC TTACACCGGAGCAAGTGTGGCCATTGCAAGC AACATCGGTGCAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT</p>	<p>251.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDDKMAPKKRKRKVI HRGVPMVDLRTLGSQQQEKIKPKVRSTVAQHHEAL VGHGFTHAHIVALSQHPAALGTVAVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQDLD GQLLKIARGGVTAVEAVHAWRNALTGAPLNLPDQV VAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVAI ASNIGGKQALETVQRLLPVLCAHGLTPDQVVAIANN NGGKQALETVQRLLPVLCAHGLTPAQVVAIANNNGG KQALETVQRLLPVLCDHGLTPDQVVAIASNGGKQ LETVQRLLPVLCDHGLTPEQVVAIASNGGKQALET VQRLLPVLCAHGLTPDQVVAIASHDGGKQALETVQR LLPVLCAHGLTPAQVVAIASHDGGKQALETVQRLLP VLCQDHGLTPDQVVAIANNNGGKQALETVQRLLPVL CDHGLTPEQVVAIASHDGGKQALETVQRLLPVLCAH GLTPDQVVAIASHDGGKQALETVQRLLPVLCAHGLT PAQVVAIASHDGGKQALETVQRLLPVLCDHGLTPDQ VVAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVA IASNGGKQALETVQRLLPVLCAHGLTPDQVVAIAS NIGGKQALETVQRLLPVLCAHGLTPAQVVAIASHDG GKQALETVQRLLPVLCDHGLTPEQVVAIASHDGGRP ALESIVAQLSRPDPALAALTNHDLVALACLGGRPALD AVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>	<p>252.</p>

	<p>GCGAATAACAATGGAGGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCCACGGTTTGACGCCTGCACAAGTGGTC GCCATCGCCAACAACAACGGCGGTAAGCAGGC GCTGGAAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCAAACGGAGGGGAAA GCAAGCCCTGGAAACCGTGCAAAGTGTGTGC CGGTCCCTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCAAGCAATGGGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTCTCTGTCAAGCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGTCGCA TGACGGAGGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCGTGTGTGTCAGCCAC GGTTGACGCCTGCACAAGTGGTCGCCATCGC CAGCCATGATGGCGGTAAGCAGGCGCTGAAA CAGTACAGCGCCTGCTGCCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGAACATAATGGGGAAAGCAAGCCC TGGAAACCGTGCAAAGGTTGTTGCCGGTCTT TGTCAAGACCACGGCCTTACACCGGAGCAAGT CGTGCCATTGCATCCACGACGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCCATGACGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCCGTGTGTGTCAAGCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATCGCCAGCCATG ATGGCGGTAAGCAGGCGCTGGAAACAGTACAG CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGT CACATGACGGGGAAAGCAAGCCCTGGAAACC GTGCAAAGGTTGTTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGAGCAAGTCGTGGCCA TTGCAAGCAATGGGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTCTCTG</p>	
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	<p>TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGTTCGAACATTGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAAGCCCACGGTTTGACGCCTGCAC AAGTGGTCGCCATCGCCAGCCATGATGGCGGT AAGCAGGCGCTGGAACAGTACAGCGCCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCGCCATTGCTTCCCACGAC GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCGGTTGGCTG CGTTAACGAATGACCATCTGGTGGCTTGGCA TGCTTGGTGGACGCCCGCTCGATGCAGT CAAAAAGGGTCTGCCTCATGCTCCCGATTGA TCAAAAGAACCAACGGCGGATTCCCGAGAGA ACTTCCCATCGAGTCGCGGGATCC</p>			
<p>>BRCA2_Lef t TTAGACTT AGGTAAGTAA _TAL/010/0 11/019/021 /027/015/0 20/021/029 /014/020/0 21/026/014 /020/021/J DS70/ (TTAGACTT AGGTAAGTAA ' disclosed as SEQ ID NO: 416)</p>	<p>GCTAGCaccATGGACTACAAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGCATTCCACCGGGGTACCTAT GGTGGACTTGAGGACACTCGGTATTTCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTGACG AGCACCGTCGGCAACACCACGAGGCGCTTGT GGGGCATGGCTTCACTCATGCGCATATTGTCG CGCTTTCACAGCACCTGCGGCGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCGAAGCCACGCACGAGGCAATTGTAG GGTCCGGTAAACAGTGGTCGGGAGCGCGAGCA CTTGAGGCGCTGCTGACTGTGGCGGGTGAGCT TAGGGGGCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGAGGGGAGTA ACAGCGGTAGAGGCAAGTGCACCGCTGGCGCAA TGCGCTCACCGGGCCCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGCGTCAAACGGA GGGGAAAGCAAGCCCTGGAACCGTGCAAAG GTTGTTGCCGGTCCCTTGTCAAGACCACGGCC TTACACCGGACCAAGTCGTGGCCATTGCAAGC AACATCGGTGGCAACAGGCTCTTGAGACGGT</p>	<p>253.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDDKMAPKKRKRKVI HRGVPMVDLRTLGYSQQQEKIKPKVRSVVAQHHEAL VGHGFTHAHIVALSQHPAALGTAVVKYQDMIAALPEA THEAIVGVKQWSGARALEALLTVAGELRGPPLQDLD GQLLKIARGGVTAVEAVHAWRNALTGAPLNLTDPQV VAIASNGGKQALETVQRLLPVLCDHGLTPEQVVAI ASNIGGKQALETVQRLLPVLCAHGLTDPQVVAIANN NGGKQALETVQRLLPVLCAHGLTPAQVVAIASNIGG KQALETVQRLLPVLCDHGLTDPQVVAIASHDGGKQA LETVQRLLPVLCDHGLTPEQVVAIASNGGKQALET VQRLLPVLCAHGLTDPQVVAIASNGGKQALETVQR LLPVLCAHGLTPAQVVAIASNIGGKQALETVQRLLP VLCQDHGLTDPQVVAIANNNGGKQALETVQRLLPVL CDHGLTPEQVVAIANNNGGKQALETVQRLLPVLCAH GLTDPQVVAIASNGGKQALETVQRLLPVLCAHGLT PAQVVAIASNIGGKQALETVQRLLPVLCDHGLTDPQ VVAIASNIGGKQALETVQRLLPVLCDHGLTPEQVVA IANNNGGKQALETVQRLLPVLCAHGLTDPQVVAIAS NGGGKQALETVQRLLPVLCAHGLTPAQVVAIASNIG GKQALETVQRLLPVLCDHGLTPEQVVAIASNIGGRP ALESIVAQLSRPDPALAALINDHLVALACLGRPALD AVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>	<p>254.</p>

	<p>TCAGAGACTTCTCCAGTTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGAATAACAATGGAGGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCACGGTTTGACGCCGTCACAAGTGGTC GCCATCGCCTCCAATATTGGGGTAAGCAGGC GCTGGAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCACATGACGGGGAAA GCAAGCCCTGGAAACCGTGCAAGGTTGTTGC CGGTCTTTGTCAAGACCAGGCCTTACACCG GAGCAAGTCGTGGCCATTGCAAGCAATGGGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTTCTCTGTCAAGCCCACGGGCTG ACTCCGATCAAGTTGTAGCGATTGCGTCCAA CGGTGGAGGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCGTGTGTGTCAAGCCCAC GGTTTGACGCCCTGCACAAGTGGTCGCCATCGC CTCCAATATTGGCGGTAAGCAGGCGCTGGAAA CAGTACAGCGCCTGCTGCCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGGAAACAATAATGGGGAAAGCAAGCCC TGGAAACCGTGCAAGGTTGTTGCCGGTCTTT TGTCAAGACCAGGCCTTACACGGAGCAAGT CGTGGCCATTGCAATAATAACGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCCAACGGTGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTCCCGTGTGTGTCAAGCCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATCGCCTCCAATA TTGGCGGTAAGCAGGCGCTGAAACAGTACAG CGCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGT CGAACATTGGGGAAAGCAAGCCCTGAAACC GTGCAAAGTTGTTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA</p>	
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	<p>TTGCAATAATAACGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGTCCAACGGTGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAAGCCCACGGTTTGACGCCTGCAC AAGTGGTCGCCATCGCCTCCAATATTGGCGGT AAGCAGGCGCTGGAAACAGTACAGCGCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCGCCATTGCTTCTAACATC GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCGGTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGTCTTGGTGGACGACCCGCGCTCGATGCAGT CAAAAGGGTCTGCCTCATGCTCCCGCATTGA TCAAAGAACCAACCGGGGATCCCGAGAGA ACTTCCCATCGAGTCGGGGATCC</p>			
<p>>BRCA2_Rig ht_TAGTTG TAGTTCTCCC C TAL/006/ 014/020/02 5/030/014/ 020/021/02 9/015/020/ 022/030/01 2/017/022/ JDS71/ ('TAGTTTGT AGTTCTCCCC ' disclosed as SEQ ID NO: 417)</p>	<p>GCTAGCaccATGGACTACAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACCGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGGCAATCACC CGGGGTACCTAT GGTGGACTTGAGGACACTCGGTTATTCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTGAGG AGCACCCTCGCGCAACACCACGAGGCGTTGT GGGGCATGGCTTCACTCATGCGCATATTGTCG CGCTTTCACAGCACCCCGCGGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCGAAGCCACGCACGAGGCAATTGTAG GGGTCGGTAAACAGTGGTGGGAGCGGAGCA CTTGAGGCGCTGCTGACTGTGGCGGTGAGCT TAGGGGGCTCCGCTCCAGCTCGACACCGGGC ACCTGCTGAAGATCGCGAAGAGAGGGGAGTA ACAGCGGTAGAGGCAGTGCACCGCTGGCGCAA TGCGCTCACCGGGGCCCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGCTCGAACATT GGGGAAAGCAGCCCTGGAAACCGTCAAAG GTTGTGCCCGTCTTGTCAAGACCACGGCC</p>	<p>255.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDDKMAPKKRKRKVI HRCVPMVDLRTLGYSQQQQEKIKPKVRSVAQHHEAL VGHGFTHAHIVALSQHPAALGTVAVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQD GQLLKIARGGVTAVEAVHAWRNALTGAPLNLPDQV VAIASNIGGKQALETVQRLLPVLCDHGLTPEQVVAI ANNNGGKQALETVQRLLPVLCAHGLTDPQVVAIASN GGGKQALETVQRLLPVLCAHGLTPAQVVAIASNNGG KQALETVQRLLPVLCDHGLTDPQVVAIASNNGGKQ LETVQRLLPVLCDHGLTPEQVVAIANNGGKQALET VQRLLPVLCAHGLTDPQVVAIASNNGGKQALETVQR LLPVLCAHGLTPAQVVAIASNIGGKQALETVQRLLP VLCQDHGLTDPQVVAIANNGGKQALETVQRLLPVL CDHGLTPEQVVAIASNNGGKQALETVQRLLPVLCAH GLTDPQVVAIASNNGGKQALETVQRLLPVLCAHGLT PAQVVAIASHDGGKQALETVQRLLPVLCDHGLTDPQ VVAIASNNGGKQALETVQRLLPVLCDHGLTPEQVVA IASHDGGKQALETVQRLLPVLCAHGLTDPQVVAIAS HDGGKQALETVQRLLPVLCAHGLTPAQVVAIASHDG GKQALETVQRLLPVLCDHGLTPEQVVAIASHDGGR</p>	<p>256.</p>

<p>TTACACCGGAGCAAGTCGTGGCCATTGCAAA AATAACGGTGGCAAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCCAGTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGTCCAACGGTGGAGGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCCTGTTGTGTC AAGCCCACGGTTGACGCCGTCACAAGTGGTC GCCATCGCCTCGAATGGCGGCGTAAGCAGGC GGTGGAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCAAACGGAGGGGAAA GCAAGCCCTGGAAACCGTGCAAAGTTGTTGC CGGTCTTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCAATAATAACGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCCAGTCTCTGTCAAGCCCACGGGCTG ACTCCCAGTCAAGTTGTAGCGATTGCGTCAA CGGTGGAGGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCCTGTTGTGTCAAGCCAC GGTGTGACGCTGCACAAGTGGTCCCATCGC CTCCAATATTGGCGTAAGCAGGCGCTGAAA CAGTACAGCGCCTGCTGCCGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGAACAATAATGGGGAAAGCAAGCCC TGGAAACCGTGCAAAGGTTGTTGCCGGTCCTT TGTCAAGACCACGGCCTTACACCGGAGCAAGT CGTGCCATTGCAAGCAATGGGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCCACGGGCTGACTCCCGA TCAAGTTGTAGGATTGCGTCCAACGGTGGAG GAAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCCCTGTTGTGTCAAGCCCACGGTTGAC GCCTGCACAAGTGGTCCCATGCCAGCCATG ATGCCGGTAAGCAGGCGCTGAAACAGTACAG CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGT CAAACGGAGGGGAAAGCAAGCCCTGAAACC</p>	<p>ALESIVAQLSRPDPALAALTNHDLVALACLGGRPALD AVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>
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	<p>GTGCAAAGGTTGTTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCATCCCACGACGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGTCGCATGACGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAGCCCACGGTTTGACGCCTGCAC AAGTGGTCGCCATCGCCAGCCATGATGGCGGT AAGCAGGCGCTGGAAACAGTACAGCGCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCGCCATTGCTTCCCACGAC GGAGGACGGCCAGCCTTGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCGGTTGGCTG CGTTAACGAATGACCATCTGGTGGCCTTGGCA TGTCTTGGTGGACGCCCGGCTCGATGCAGT CAAAAGGGTCTGCCTCATGCTCCCGCATGA TCAAAAGAACCAACCGGGGATCCCGAGAGA ACTTCCCATCGAGTCGGGGATCC</p>			
<p>>ERCC2 Lef t_TCCGGCCG GCGCCATGA_ TAL/007/01 2/019/024/ 027/012/01 9/024/027/ 014/017/02 2/026/015/ 034/JDS70/ ('TCCGGCCG GCGCCATGA' disclosed as SEQ ID NO: 418)</p>	<p>GCTAGCaccATGGACTACAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGGCATTCACCGGGGTACCTAT GGTGGACTTGAGGACACTCGGTTATTCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCGTCGGCACAACACCAGGCGCTTGT GGGGCATGGCTTCACTCATGCGCATATTGTCG CGTTTCACAGCACCTGCGGCGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCGAAGCCACGACGAGGCAATTGTAG GGGTCGGTAAACAGTGGTCGGGAGCGGAGCA CTTGAGGCGCTGCTGACTGTGGCGGTGAGCT TAGGGGGCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGAGGGGGAGTA ACAGCGGTAGAGGCAAGTGCACGCTGGCGCAA TGGCCTCACCGGGGCCCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGCGTCACATGAC</p>	<p>257.</p>	<p>ASTMDYKDHDGDYKDHDIDYKDDDDKMAPKKRKRKVI HRGVPVMDLRTLGYSQQQQEKIKPKVRSTVAQHHEAL VGHGFTHAHIVALSQHPAALGTVAVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQDLD GQLLKI AKRGGVTAVEAVHAWRNALTGAPLNLPDQV VAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVAI ASHDGGKQALETVQRLLPVLCDHGLTPDQVVAIANN NGGKQALETVQRLLPVLCDHGLTPAQVVAIANNNGG KQALETVQRLLPVLCDHGLTPDQVVAIASHDGGKQA LETVQRLLPVLCDHGLTPEQVVAIASHDGGKQALET VQRLLPVLCDHGLTPDQVVAIANNNGGKQALETVQR LLPVLCDHGLTPAQVVAIANNNGGKQALETVQRLLP VLCQDHLTPDQVVAIASHDGGKQALETVQRLLPVLCD QDHLTPDQVVAIANNNGGKQALETVQRLLPVLCDHGL TPAQVVAIASHDGGKQALETVQRLLPVLCDHGLTPDQ VVAIASNIGGKQALETVQRLLPVLCDHGLTPEQVVA IASNGGGKQALETVQRLLPVLCDHGLTPDQVVAIAN</p>	<p>258.</p>

<p>GGGGAAAGCAAGCCCTGGAACCGTGCAAAG GTTGTTGCCGGTCCCTTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCGTGGCCATTGCATCC CACGACGGTGGCAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCCAGTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGAATAACAATGGAGGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCCACGGTTTGACGCTGCACAAGTGGTC GCCATCGCCAACAACAACGGCGTAAGCAGGC GCTGGAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCACATGACGGGGAAA GCAAGCCCTGGAACCGTGCAAAGTTGTTGC CGGTCTTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCATCCCAGACGG TGGCAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCCAGTCTCTGTCAAGCCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGAATAA CAATGGAGGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCGTGTGTGTCAGCCAC GGTTGACGCTGCACAAGTGGTCGCCATCGC CAACAACAACGGCGTAAGCAGCGCTGGAAA CAGTACAGCGCCTGCTGCCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGTCACATGACGGGGAAAGCAAGCCC TGGAAACCGTGCAAAGTTGTTGCCGGTCCCT TGTCAAGACCACGGCCTTACACGGAGCAAGT CGTGCCATTGCAAATAATAACGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCGCATGACGGAG GGAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCCGTGTGTGTCAAGCCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATCGCCAGCCATG ATGGCGGTAAGCAGCGCTGGAACAGTACAG CGCTGCTGCCTGTACTGTGCCAGGATCATGG</p>	<p>NNGGKQALETVQRLPVLQAHGLTPEQVVAIASNIG GRPALESIVAQLSRPDPALAAALNDHLVALACLGGRP ALDAVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>
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	<p>ACTGACCCAGACCAGGTAGTCGCAATCGCGT CGAACATTGGGGAAAGCAAGCCCTGGAACC GTGCAAAGGTTGTTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCAAGCAATGGGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTTCTCTG TCAAGCCCACGGGCTGACTCCCAGTCAAGTTG TAGCGATTGCGAATAACAATGGAGGAAACAA GCATTGGAGACTGTCCAACGGTCTTCCCCT GTTGTGTCAAGCCCAGGTCTGACACCCGAAC AGTGGTCCGATTCGTTCTAACATCGGAGGA CGGCCAGCCTTGGAGTCCATCGTAGCCCAATT GTCCAGGCCCGATCCCCTGTTGGCTGCGTTAA CGAATGACCATCTGGTGGCGTTGGCATGTCTT GGTGACGACCCGCGCTCGATGCAGTCAAAAA GGGTCTGCCCTCATGCTCCCGCATGATCAAAA GAACCAACCGCGGATCCCAGAGAACTTCC CATCGAGTCCGGGATCC</p>			
<p>>ERCC2 Righ ht_TAGCGAG CGCGACCC TAL/006/01 4/017/024/ 026/014/01 7/024/027/ 014/016/02 2/027/012/ JDS71/ ('TAGCGAGC GCGACCCC' disclosed as SEQ ID NO: 419)</p>	<p>GCTAGCaccATGGACTACAAGACCATGACGG TCATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGTGGGCATTCACCGGGGTACCTAT GGTGGACTTGAGGACTCGGTTATTGCGAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCGTCGCGCAACACCAGAGGCGCTTGT GGGGCATGGCTTCACTCATGCGCATATTGTCG CGCTTTCACAGCACCCCTGCCGGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCGAAGCCACGCACGAGCAATTGTAG GGGTCGGTAAACAGTGGTCCGGAGCGCGAGCA CTTGAGGCGCTGCTGACTGTGGCGGTGAGCT TAGGGGGCTCCGCTCCAGTCCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGAGGGGGAGTA ACAGCGGTAGAGGAGTGCACGCCTGGCGCAA TCGCGCTACCGGGGCCCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGCTCGAACATT GGGGAAAGCAAGCCCTGGAACCGTGCAAAG</p>	<p>259.</p>	<p>ASTMDYKDHDGDKDHDIDYKDDDDKMAPKKRKRKVI HRGVPMVDLRTLGSQQQEKIKPKVRSTVAQHHEAL VGHGFTHAHIVALSQHPAALGTAVVKYQDMIAALPEA THEAIVGVKQWSGARALEALLTVAGELRGPPLQDLD GQLLKIARGGVTAVEAVHAWRNALTGAPLNLPDQV VAIASNIGGKQALETVQRLLPVLCDHGLTPEQVVAI ANNNGGKQALETVQRLLPVLCAHGLTPDQVVAIASH DGGKQALETVQRLLPVLCAHGLTPAQVVAIANNGG KQALETVQRLLPVLCDHGLTPDQVVAIASNIGGKQ LETVQRLLPVLCDHGLTPEQVVAIANNGGKQALET VQRLLPVLCAHGLTPDQVVAIASHDGGKQALETVQR LLPVLCAHGLTPAQVVAIANNGGKQALETVQRLLP VLCQDGLTPDQVVAIASHDGGKQALETVQRLLPVL CDHGLTPEQVVAIANNGGKQALETVQRLLPVLCAH GLTPDQVVAIASNIGGKQALETVQRLLPVLCAHGLT PAQVVAIASHDGGKQALETVQRLLPVLCDHGLTPDQ VVAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVA IASHDGGKQALETVQRLLPVLCAHGLTPEQVVAIAS HDGGRPALESIVAQLSRPDPALAALNDHLVALACLG</p>	<p>260.</p>

<p>GTTGTTGCCGGTCCTTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCGTGGCCATTGCAAAAT AATAACGGTGGCAAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCCAGTTCTCTGTCAAGCCC ACGGGCTGACTCCCAGTCAAGTTGTAGCGATT GCGTCGCATGACGGAGGGAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCCTGTTGTGTC AAGCCCACGGTTTGACGCTGCACAAGTGGTC GCCATCGCCAACAACAACGGCGGTAAGCAGGC GCTGGAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCGAACATTGGGGAAA GCAAGCCCTGGAAACCGTGCAAAGTTGTTGC CGGTCTTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCAAATAATAACGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCCAGTTCTCTGTCAAGCCCACGGGCTG ACTCCCAGTCAAGTTGTAGCGATTGCGTCGCA TGACGGAGGGAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCCTGTTGTGTCAAGCCAC GGTTTGACGCTGCACAAGTGGTCGCCATCGC CAACAACAACGGCGTAAGCAGGCGCTGAAA CAGTACAGCGCCTGCTGCCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGTCACATGACGGGGAAAGCAAGCCC TGGAAACCGTGCAAAGTTGTTGCCGGTCTT TGTCAAGACCACGGCCTTACACCGGAGCAAGT CGTGGCCATTGCAAATAATAACGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCGAACATTGGAG GGAACAAGCATTGGAGACTGTCCAACGGCTC CTCCCCTGTTGTGTCAAGCCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATCGCCAGCCATG ATGGCGGTAAGCAGGCGCTGAAAACAGTACAG CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGT</p>	<p>GRPALDAVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>
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	<p>CACATGACGGGGAAAGCAAGCCCTGGAACC GTGCAAAGGTGTGCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCATCCCACGACGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTCTCTG TCAAGCCCACGGGCTGACACCCGAACAGGTGG TCGCCATTGCTTCCCACGACGGAGGACGGCCA GCCTTGGAGTCCATCGTAGCCCAATTGTCCAG GCCCGATCCCGCTGGCTGCGTTAACGAATG ACCATCTGGTGGCTTGGCATGTCTTGGTGA CGACCCGCGCTCGATGCAGTCAAAAAGGTCT GCCTCATGCTCCCGCATTGATCAAAAGAACCA ACCGGCGGATTCCCGAGAGAACTCCCATCGA GTCGCGGATCC</p>			
<p>>FANCA_Lef t_TAGGCGCC AAGGCCATGT _TAL/006/0 14/019/022 /029/012/0 17/021/026 /014/019/0 22/027/011 /020/024/J DS78/ ('TAGGCGCC AAGGCCATGT ' disclosed as SEQ ID NO: 420)</p>	<p>GCTAGCaccATGGACTACAAGACCATGACGG TGATATATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGGCATTACCAGGGGTACCTAT GGTGGACTTGAGGACACTCGGTTATTCCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCGTCCGCAACACCACGAGGCGCTTGT GGGGCATGGCTTCACTCATGCCATATTGTCG CGCTTTCACAGCACCCCTGCGGGCTTGGGACG GTGGCTGTCAATACCAAGATATGATTGCGGC CCTGCCGAAGCCACGACGAGGCAATTGTAG GGGTCGTTAAACAGTGGTCGGGAGCGCAGCA CTTGAGGCGCTGCTGACTGTGGCGGTGAGCT TAGGGGGCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGAGGGGAGTA ACAGCGGTAGAGGCAAGTCCGCTGCAACATT TCGCCTCACCGGGCCCCCTTGAACTGACCC CAGACCAGGTAGTCGCAATCGGCTGCAACATT GGGGAAAGCAAGCCCTGGAACCGTGCAAAG GTTGTGCGGTCTTTGTCAAGACCACGGCC TTACACCGGACCAAGTCTGGCCATTGCAAAT AATAACGGTGGCAAACAGGCTCTTGAGACGGT TCAGAGACTTCCCAGTCTCTGTCAAGCCC</p>	<p>261.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDDKMAPKKRKGVI HRGVPMVDLRTLGSQQQEKIKPKVRSTVAQHHEAL VGHGFTHAHIVALSQHPAALGTVAVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQDLD GQLLKIARKGGVTAVEAVHAWRNALTGAPLNLPDQV VAIASNIGGKQALETVQRLLPVLCDHGLTPEQVVAI ANNNGGKQALETVQRLLPVLCAHGLTPDQVVAIANN NGGKQALETVQRLLPVLCAHGLTPAQVVAIASHDGG KQALETVQRLLPVLCDHGLTPDQVVAIANNNGGKQ ALETVQRLLPVLCDHGLTPEQVVAIASHDGGKQALET VQRLLPVLCAHGLTPDQVVAIASHDGGKQALETVQR LLPVLCAHGLTPAQVVAIASNIGGKQALETVQRLLP VLCQDHGLTPDQVVAIASNIGGKQALETVQRLLPVL CDHGLTPEQVVAIANNNGGKQALETVQRLLPVLCAH GLTPDQVVAIANNNGGKQALETVQRLLPVLCAHGLT PAQVVAIASHDGGKQALETVQRLLPVLCDHGLTPDQ VVAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVA IASNIGGKQALETVQRLLPVLCAHGLTPDQVVAIAS NNGGKQALETVQRLLPVLCAHGLTPAQVVAIANNNG GKQALETVQRLLPVLCDHGLTPEQVVAIASNNGGRP ALESIVAQLSRPDPALAALTNHDLVALACLGGRPALD AVKGLPHAPALIKRTNRRIPERTSHRVAGS</p>	<p>262.</p>

ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGAATAACAATGGAGGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCCACGGTTTGACGCTGCACAAGTGGTC GCCATCGCCAGCCATGATGGCGGTAAGCAGGC GCTGGAAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGGAACAATAATGGGGAAA GCAAGCCCTGGAAACCGTGCAAGGTTGTGC CGGTCTTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCATCCCAGACGG TGGCAAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTTCTCTGTCAAGCCACGGGCTG ACTCCCATCAAGTTGTAGCGATTGCGTCGCA TGACGGAGGGAAACAAGCATTGGAGACTGCC AACGGCTCCTTCCCGTGTGTGTCAGCCAC GGTTTGACGCCTGCACAAGTGGTCGCCATCGC CTCCAATATTGGCGTAAGCAGCGCTGGAAA CAGTACAGCGCCTGCTGCCGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGTCGAACATTGGGGAAAGCAAGCCC TGGAACCGTGCAAGGTTGTTGCCGGTCTT TGTCAAGACCAGGCCTTACACGGAGCAAGT CGTGCCATTGCAATAATAACGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGAATAACAATGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCCGTGTGTGTCAGCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATGCCAGCCATG ATGGCGGTAAGCAGCGCTGGAAACAGTACAG CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGT CACATGACGGGGAAAGCAAGCCCTGGAAACC GTGCAAAGGTTGTGCGGTCCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCAAGCAACATCGGTGGCAAACAGGCTCTT		
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	<p>GAGACGGTTCAGAGACTTCTCCAGTTCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGTCCAACGGTGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAAGCCCACGGTTTACGCCTGCAC AAGTGGTCGCCATCGCCAACAACAACGGCGGT AAGCAGGCGCTGGAAACAGTACAGCGCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCGCCATGCTTCTAATGGG GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCCGATCCCGGTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGTCTTGGTGGACACCCGGCTCGATGCAGT CAAAAAGGGTCTGCCTCATGCTCCCGCATGA TCAAAAGAACCAACCGGGGATTCCCGAGAGA ACTTCCCATCGAGTCGCGGGATCC</p>			
<p>>FANCA Rig ht_TGGCCCG AGGCGGAGTT C_TAL/009/ 014/017/02 2/027/014/ 016/024/02 9/012/019/ 024/026/01 4/020/025/ JDS71/ ('TGGCCCGA GGCGGAGTTC ' disclosed as SEQ ID NO: 421)</p>	<p>GCTAGCaccATGGACTACAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACAGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGCATTACCCGGGGTACCTAT GGTGGACTTGAGGACACTCGGTATTTCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCGTCGGCAACACCACGAGGCGCTTGT GGGCGATGGCTTCACTCATGCGCATATTGTCG CGCTTTCACAGCACCTTCCGGCGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCCGAAGCCACGACGAGGCAATTGTAG GGTTCGGTAACAGTGGTTCGGAGCGCGAGCA CTTGAGGCGCTGCTGACTGTGGCGGTGAGCT TAGGGGGCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGGGGGAGTA ACAGCGGTAGAGGAGTGCACGCTGGCGCAA TGGCTCACCGGGCCCCCTTGAACCTGACCC CAGACCAGGTAGTCGAATCGCGAACAATAAT GGGGAAAGCAAGCCCTGGAACCGTGCAAAG GTTGTTGCCGCTCCTTTGTCAAGACCACGGCC TTACACCGGACCAAGTCTGGCCATTGCAAAAT</p>	<p>263.</p>	<p>ASTMDYKDHDGDKDHDIDYKDDDDKMAPKKRKRKVI HRGVPMVDLRTLGSQQQEKIKPKVRSVVAQHHEAL VGHGFTHAHIVALSQHPAALGTVAVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQDLD GQLLKIARGGVTAVEAVHAWRNALTGAPLNLTDPQV VAIANNGGKQALETVQRLLPVLCDHGLTPEQVVAI ANNNGKQALETVQRLLPVLCAHGLTPDQVVAIASH DGGKQALETVQRLLPVLCAHGLTPAQVVAIASHDGG KQALETVQRLLPVLCDHGLTPDQVVAIASHDGGKQA LETVQRLLPVLCDHGLTPEQVVAIANNGGKQALET VQRLLPVLCAHGLTPDQVVAIASNIGGKQALETVQR LLPVLCAHGLTPAQVVAIANNGGKQALETVQRLLP VLCQDHGLTPDQVVAIANNGGKQALETVQRLLPVL CDHGLTPEQVVAIASHDGGKQALETVQRLLPVLCAH GLTPDQVVAIANNGGKQALETVQRLLPVLCAHGLT PAQVVAIANNGGKQALETVQRLLPVLCDHGLTPDQ VVAIASNIGGKQALETVQRLLPVLCDHGLTPEQVVA IANNGGKQALETVQRLLPVLCAHGLTPDQVVAIAS NNGGKQALETVQRLLPVLCAHGLTPAQVVAIASNGG GKQALETVQRLLPVLCDHGLTPEQVVAIASHDGGRP ALESIVAQLSRPDPALAALNDHLVALACLGRPALD</p>	<p>264.</p>

<p>AATAACGGTGGCAAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCCAGTTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGTCGCATGACGGAGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCCACGGTTTGACGCCTGCACAAGTGGTC GCCATCGCCAGCCATGATGGCGGTAAGCAGGC GCTGAAACAGTACAGCCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCACATGACGGGGAAA GCAAGCCCTGGAACCGTGCAAAGTTGTTGC CGGTCTTTGTCAAGACCAGGCCCTACACCG GAGCAAGTCGTGGCCATTGCAAATAATAACGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCCAGTTCTCTGTCAAGCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGTCGAA CATTGGAGGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCGTGTGTGTCAAGCCAC GGTTTGACGCCTGCACAAGTGGTCGCCATCGC CAACAACAACGGCGGTAAGCAGGCGCTGAAA CAGTACAGCCCTGCTGCCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCCGGAACAATAATGGGGAAAGCAAGCCC TGAAACCGTGCAAAGGTTGTTGCCGGTCTT TGTCAGACCACGGCCTTACCCGGAGCAAGT CGTGGCCATTGCATCCCACGACGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGAATAACAATGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCCGTGTGTGTCAAGCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATCGCCAACAACA ACGGCGGTAAGCAGGCGCTGAAACAGTACAG CGCCTGTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGT CGAACATTGGGGAAAGCAAGCCCTGAAACC GTGCAAAGGTTGTTGCCGGTCTTTGTCAAGA</p>	<p>AVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>
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	<p>CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCAAATAATAACGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGTCCAACGGTGGAGGAAACAA GCATTGGAGACTGICCAACGGCTCCTTCCCGT GTTGTCTCAAGCCCACGGTTTACGCCTGCAC AAGTGGTCGCCATCGCCTCGAATGGCGGCGGT AAGCAGGCGCTGGAAACAGTACAGCGCCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCCGATTCCTTCCCACGAC GGAGGACGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCGCGTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGTCTTGGTGGACACCCCGCTCGATGCAGT CAAAAGGGTCTGCCTCATGCTCCCGCATGA TCAAAGAACCAACCGCGGATCCCGAGAGA ACTTCCCATCGAGTCGCGGGATCC</p>			
<p>>FANCC Lef t TGAAGGGA CATCACCTTT TAL/009/0 11/016/024 /029/014/0 16/022/026 /015/017/0 21/027/012 /020/025/J DS78/ ('TGAAGGGA CATCACCTTT ' disclosed as SEQ ID NO: 422)</p>	<p>GCTAGCaccATGGACTACAAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGGCATTACCCGCGGGTACCTAT GGTGACTTGAGGACACTCGGTATTCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCGTCGCGCAACACCACGAGGCGCTTGT GGGGCATGGCTTCACTCATCGCATATTGTCTG CGCTTTCACAGCACCTTGGCGGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCCGAAGCCACGACGAGGCAATTGTAG GGGTCGGTAAACAGTGGTGGGAGCGGAGCA CTTGAGGCGCTGCTGACTGTGGCGGCTGAGCT TAGGGGCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGGGGGAGTA ACAGCGGTAGAGGAGTGCACCGCTGGCGCAA TGGCCTCACCGGGCCCCCTTGAACCTGACCC CAGACCAGGTAGTCGAATCGCGAACAATAAT GGGGAAAGCAAGCCCTGGAAACCGTGCAAAG</p>	<p>265.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDKMAPKKRKGVI HRGVPMVDLRTLGSQQQEKIKPKVRSVAQHHEAL VGHGFTHAHIVALSQHPAALGTVAVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQDLD GQLLKIAKRGGVTAVEAVHAWRNALTGAPLNLPDQV VAIANNNGKQALETVQRLLPVLCDHGLTPEQVVAI ASNIGGKQALETVQRLLPVLCOAHGLTPDQVVAIASN IGGKQALETVQRLLPVLCOAHGLTPAQVVAIANNNGG KQALETVQRLLPVLCDHGLTPDQVVAIANNNGKQA LETVQRLLPVLCDHGLTPEQVVAIANNNGKQALET VQRLLPVLCOAHGLTPDQVVAIASNIGGKQALETVQR LLPVLCOAHGLTPAQVVAIASHDGGKQALETVQRLLP VLCQDHLTPDQVVAIASNIGGKQALETVQRLLPVL QDHLTPDQVVAIASNIGGKQALETVQRLLPVLCOAH GLTPDQVVAIASHDGGKQALETVQRLLPVLCOAHGLT PAQVVAIASNIGGKQALETVQRLLPVLCDHGLTPDQ VVAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVA IASHDGGKQALETVQRLLPVLCOAHGLTPDQVVAIAS NIGGKQALETVQRLLPVLCOAHGLTPAQVVAIASNIGG</p>	<p>266.</p>

<p>GTTGTTGCCGGTCCTTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCGTGGCCATTGCAAGC AACATCGGTGGCAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGTCGAACATTGGAGGGAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCCACGGTTTGACGCTGCACAAGTGGTC GCCATCGCCAACAACAACGGCGGTAAGCAGGC GCTGGAAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCAACAATAATGGGGAAA GCAAGCCCTGGAAACCGTGCAAAGTTGTGTC CGGTCCTTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCAAATAATAACGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTTCTCTGTCAAGCCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGTCGAA CATTGGAGGGAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCGTGTGTGTCAGCCAC GGTTTGACGCTGCACAAGTGGTCGCCATCGC CAGCCATGATGGCGTAAGCAGGCGCTGAAA CAGTACAGCGCCTGCTGCCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGTCGAACATTGGGGAAAGCAAGCCC TGGAAACCGTGCAAAGGTTGTTGCCGGTCCTT TGTCAAGACCACGGCCTTACACCGGAGCAAGT CGTGGCCATTGCAAGCAATGGGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCGCATGACGGAG GGAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCCGTGTGTGTCAAGCCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATCGCTCCAATA TTGGCGGTAAGCAGGCGCTGAAACAGTACAG CGCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGT</p>	<p>GKQALETVQRLLPVLCCQDHGLTPEQVVAIASNGGGRP ALESIVAQLSRPDPALAAALNDHLVALACLGGRPALD AVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>
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	<p>CACATGACGGGGAAAGCAAGCCCTGGAAACC GTGCAAAGGTTGTTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGCCA TTGCATCCCACGACGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTTCCTG TCAAGCCCACGGGCTGACTCCCAGTCAAGTTG TAGCGATTGCGTCCAACGGTGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAAGCCCACGGTTTGACGCCCTGAC AAGTGGTCGCCATCGCTCGAATGGCGGCGGT AAGCAGGCGCTGGAAACAGTACAGCGCCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGCTGCCATTGCTTCTAATGGG GGAGGACGGCCAGCCTTGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCGGTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGTCTTGGTGGACACCCGCGCTCGATGCAGT CAAAAGGGTCTGCCTCATGCTCCCGCATGA TCAAAGAACCAACCGCGGATTCGCGAGAGA ACTTCCCATCGAGTCGCGGGATCC</p>			
<p>>FANCC Rig ht_TCTACTG AATCTTGAGC TAL/007/0 15/016/022 /030/014/0 16/021/030 /012/020/0 25/029/011 /034/JDS71 / ('TCTACTGA ATCTTGAGC' disclosed as SEQ ID NO: 423)</p>	<p>GCTAGCaccATGGACTACAAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGCATTACCCCGGGGTACCTAT GGTGGACTTGAGGACACTCGGTTATTGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAAG AGCACCGTCGCGCAACACCACGAGGCGCTTGT GGGGCATGGCTTCACTCATGCGCATATTGTCG CGCTTTCACAGCACCCCTGCGGCGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCCGAAGCCACGCACGAGCAATTGTAG GGGTCGGTAAACAGTGGTCGGGAGCGCGAGCA CTTGAGGCGCTGCTGACTGTGGCGGGTGAAGT TAGGGGGCCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGAGGGGGAGTA ACAGCGGTAGAGGAGTGCACCGCTGGCGCAA TGGCTCACCGGGGCCCTTGAACCTGACCC</p>	<p>267.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDDKMAPKKRKRKVI HRGVPMVDLRLTGYQQQEKIKPKVRSTVAQHHEAL VGHGFTHAHIVALSQHPAALGTVAVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQDLD GQLLKIARKGGVTAVEAVHAWRNALTGAPLNLTPDQV VAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVAI ASNGGKQALETVQRLLPVLCDHGLTPEQVVAIASN IGGKQALETVQRLLPVLCDHGLTPEQVVAIASHDGG KQALETVQRLLPVLCDHGLTPEQVVAIASNGGKQA LETVQRLLPVLCDHGLTPEQVVAIANNGGKQALET VQRLLPVLCDHGLTPEQVVAIASNIGGKQALETVQR LLPVLCDHGLTPEQVVAIASNIGGKQALETVQRLLP VLCDHGLTPEQVVAIASNGGKQALETVQRLLPVLCD HDHGLTPEQVVAIASHDGGKQALETVQRLLPVLCDH GLTPEQVVAIASNGGKQALETVQRLLPVLCDHGLT PAQVVAIASNGGKQALETVQRLLPVLCDHGLTPEQ VVAIANNGGKQALETVQRLLPVLCDHGLTPEQVVA</p>	<p>268.</p>

<p>CAGACCAGGTAGTCGCAATCGCGTCACATGAC GGGGAAAGCAAGCCCTGGAACCGTGCAAAG GTTGTTGCCGGTCCCTTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCGTGGCCATTGCAAGC AATGGGGGTGGCAAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCCAGTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGTCGAACATTGGAGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCCACGGTTTGACGCTGCACAAGTGGTC GCCATCGCCAGCCATGATGGCGGTAAGCAGGC GCTGAAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCACAGACCAG GTAGTCGCAATCGCGTCAAACGGAGGGGAAA GCAAGCCCTGGAACCGTGCAAAGTTGTTGC CGGTCTTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCAAATAATAACGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCCAGTCTCTGTCAAGCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGTCGAA CATTGGAGGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCGTGTGTGTCAAGCCAC GGTTGACGCGCTGCACAAGTGGTCGCCATCGC CTCCAATATTGGCGTAAGCAGGCGCTGAAA CAGTACAGCGCCTGCTGCCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGTCAAACGGAGGGGAAAGCAAGCCC TGGAACCGTGCAAAGTTGTGCGGTCCTT TGTCAAGACCAGGCTTACACCGGAGCAAGT CGTGGCCATTGCAITCCACGCGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCACGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCCAACGGTGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCCGTGTGTGTCAAGCCACGGTTGAC GCCTGCACAAGTGGTCGCCATCGCCTCGAATG GCGGCGTAAGCAGGCGCTGGAACAGTACAG</p>	<p>IASNIGGKQALETVQRLLPVLCAHGLTPDQVVAIAN NNGGKQALETVQRLLPVLCAHGLTPEQVVAIASHDG GRPALESIVAQLSRPDPALAAITNDHLVALACLGGRP ALDAVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>
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	<p>CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGA ACAATAATGGGGAAAGCAAGCCCTGGAAACC GTGCAAAGGTTGTTGCCGGTCCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCAAGCAACATCGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTTCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGAATAACAATGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTCCCGT GTTGTGTCAAGCCCACGGTCTGACACCCGAAC AGGTGGTCGCCATTGCTTCCCACGACGGAGGA CGGCCAGCCTTGGAGTCCATCGTAGCCCAATT GTCCAGGCCCGATCCCGGTTGGCTGCGTTAA CGAATGACCATCTGGTGGCGTTGGCATGTCTT GGTGGACGACCCGCGCTCGATGCAGTCAAAA GGGTCTGCCTCATGCTCCCGCATTGATCAAAA GAACCAACCGGGGATTTCCCGAGAGAACTCC CATCGAGTCGCGGGATCC</p>			
<p>>FANCG Lef t TCGGCCAC CATGTCCC T AL/007/014 /019/022/0 27/011/017 /022/026/0 15/019/025 /027/012/J DS71/ ('TCGGCCAC CATGTCCC' disclosed as SEQ ID NO: 424)</p>	<p>GCTAGCaccATGGACTACAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGGCAATCACCAGGGGTACCTAT GGTGACTTGGAGACTCGGTTATTCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCGTCGCGCAACACCAGAGGCGCTTGT GGGGATGGCTTCACTCATGCGCATATTGTCG CCCTTTCACAGCACCTCGCGGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCGAAGCCACGCACGAGGCAATTGTAG GGGTCGGTAAACAGTGGTGGGAGCGCGAGCA CTTGAGGCGCTGCTGACTGTGGCGGTGAGCT TAGGGGCTCCGCTCCAGTCCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGAGGGGAGTA ACAGCGGTAGAGGAGTGCACGCTGGCGCAA TCGGCTCACCGGGCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGGCTCACATGAC</p>	<p>269.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDDKMAPKKRKRKVI HRGVPMVDLRTLGYSQQQEIKPKVRSVAQHHEAL VHGFTHAHIVALSQHPAALGTVAVKYQDMIAALPEA THEAIVGVXQWSGARALEALLTVAGELRGPPLQD TQLLKIARGGVTAVEAVHAWRNALTGAPLNLPDQV VAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVAI ANNNGGKQALETVQRLLPVLCDHGLTPEQVVAIANN NGGKQALETVQRLLPVLCDHGLTPEQVVAIASHDGG KQALETVQRLLPVLCDHGLTPEQVVAIASHDGGKQ ALETVQRLLPVLCDHGLTPEQVVAIASNIGGKQALET VQRLLPVLCDHGLTPEQVVAIASHDGGKQALETVQR LLPVLCDHGLTPEQVVAIASHDGGKQALETVQRLLP VLCQDGLTPEQVVAIASNIGGKQALETVQRLLPVL CDHGLTPEQVVAIASNIGGKQALETVQRLLPVLCDH GLTPEQVVAIASNIGGKQALETVQRLLPVLCDHGLT PAQVVAIASNIGGKQALETVQRLLPVLCDHGLTPEQ VVAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVA IASHDGGKQALETVQRLLPVLCDHGLTPEQVVAIAS</p>	<p>270.</p>

<p>GGGGAAAGCAAGCCCTGGAACCGTGCAAAG GTTGTGGCCGGTCTTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCGTGGCCATTGCAAAT AATAACGGTGGCAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCCAGTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGAATAACAATGGAGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCCTGTTGTGTC AAGCCCACGGTTTGACGCTGCACAAGTGGTC GCCATCGCCAGCCATGATGGCGTAAGCAGGC GCTGGAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCACATGACGGGGAAA GCAAGCCCTGGAACCGTGCAAGGTTGTTGC CGGTCTTTGTCAAGACCAGGCCCTTACACCG GAGCAAGTCGTGGCCATTGCAAGCAACATCGG TGGCAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCCAGTCTCTGTCAAGCCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGTCGCA TGACGGAGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCCTGTTGTCAAGCCAC GGTGACGCTGCACAAGTGGTCCCATCGC CAGCCATGATGGCGTAAGCAGGCGCTGAAA CAGTACAGCGCTGCTGCCGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGTCCAACATTGGGGAAAGCAAGCCC TGGAAACCGTGCAAGGTTGTTGCCGGTCTT TGTCAAGACCAGGCCCTTACACCGGAGCAAGT CGTGGCCATTGCAAGCAATGGGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCCACGGGCTGACTCCGA TCAAGTTGTAGCGATTGCGAATAACAATGGAG GAAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCCCTGTTGTGTCAAGCCCACGGTTGAC GCCTGCACAAGTGGTCCCATCGCTCGAATG GCGGCGGTAAGCAGGCGCTGGAACAGTACAG CGCTGCTGCCTGTACTGTGCCAGGATCATGG</p>	<p>HDGGRPALESIVAQLSRPDPALAALNDHLVALACLG GRPALDAVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>
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	<p>ACTGACCCAGACCAGGTAGTCGCAATCGCGT CACATGACGGGGAAAGCAAGCCCTGAAACC GTGCAAAGGTTGTTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCATCCCACGACGGTGGCAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTTCTCTG TCAAGCCACGGGCTGACACCCGAACAGGTGG TCGCCATTGCTTCCCACGACGGAGGACGGCCA GCCTTGGAGTCCATCGTAGCCCAATTGTCCAG GCCCGATCCCGGCTTGGCTGCGTTAACGAATG ACCATCTGCTGGCGTTGGCATGTCTTGGTGA CGACCCCGCTCGATGCAGTCAAAAAGGTCT GCCTCATGCTCCCGATTGATCAAAAGAACCA ACCGGGGATCCCGAGAGAACTTCCCATCGA GTCCGGGATCC</p>			
<p>>FANCG Righ ht_TCCAGGC AGCTGGAGCC C_TAL/007/ 012/016/02 4/029/012/ 016/024/02 7/015/019/ 024/026/01 4/017/022/ JDS71/ ('TCCAGGCA GCTGGAGCCC disclosed as SEQ ID NO: 425)</p>	<p>GCTAGCaccATGGACTACAAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGCATTACCCGGGGTACCTAT GGTGCATTGAGGACTCGGTTATTGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCGTCGGCAACACCACGAGGCGCTTGT GGGGCATGGCTTCACTCATCGCATATTGTCG CGCTTTCACAGCACCTGCCGGCGTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCCGC CCTGCCCGAAGCCACGCACGAGGCAATTGTAG GGGTCCGTAACAGTGGTCCGGAGCCGAGCA CTTGAGGCGCTGCTGACTGTGGCGGTGAGCT TAGGGGGCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGAGGGGAGTA ACAGCGGTAGAGGCAGTGCACGCTGCGCAA TGCGCTCACCGGGCCCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGCGTCACATGAC GGGGAAAGCAAGCCCTGGAACCGTGCAAAG GTTGTTGCCGGTCTTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCCGTGCCATTGCATCC CACGACGGTGGCAACAGGCTCTTGAGACGGT</p>	<p>271.</p>	<p>ASTMDYKDHDGDYKDHIDYKDDDDKMAPKKRRVGI HRGVPMVLDLRTLGSQQQEKIKPKVRSTVAQHHEAL VGHGFTHAHIVALSQHPAALGTVAVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQDLD GQLLKIARKGGVTAVEAVHAWRNALTGAPLNLPDQV VAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVAI ASHDGGKQALETVQRLLPVLCDHGLTPEQVVAIASN IGGKQALETVQRLLPVLCDHGLTPEQVVAIANNNGG KQALETVQRLLPVLCDHGLTPEQVVAIANNNGGKQA LETVQRLLPVLCDHGLTPEQVVAIASHDGGKQALET VQRLLPVLCDHGLTPEQVVAIASNIGGKQALETVQR LLPVLCDHGLTPEQVVAIANNNGGKQALETVQRLLP VLCQDGLTPEQVVAIASHDGGKQALETVQRLLPVL CDHGLTPEQVVAIASNIGGKQALETVQRLLPVLCDH GLTPEQVVAIANNNGGKQALETVQRLLPVLCDHGLT PEQVVAIANNNGGKQALETVQRLLPVLCDHGLTPEQ VVAIASNIGGKQALETVQRLLPVLCDHGLTPEQVVA IANNNGGKQALETVQRLLPVLCDHGLTPEQVVAIAS HDGGKQALETVQRLLPVLCDHGLTPEQVVAIASHDG GKQALETVQRLLPVLCDHGLTPEQVVAIASHDGGRP ALESIVAQLSRPDPALAALNDHLVALACLGGRPALD AVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>	<p>272.</p>

<p> TCAGAGACTTCTCCCAGTTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGTCGAACATTGGAGGGAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCCACGGTTTGACGCCTGCACAAGTGGTC GCCATCGCCAACAACAACGGCGGTAAGCAGGC GCTGAAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGAACAATAATGGGGGAAA GCAAGCCCTGGAAACCGTGCAAAGTTGTTGC CGGTCTTTGTCAAGACCAGGCCTTACACCG GAGCAAGTCGTGGCCATTGCATCCACGACGG TGGCAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCCAGTTCTGTCAAGCCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGTCGAA CATTGGAGGGAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCGTGTGTCAAGCCCAC GGTTTGACGCCTGCACAAGTGGTCGCCATCGC CAACAACAACGGCGGTAAGCAGGCGCTGGAAA CAGTACAGCGCCTGCTGCCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGTCACATGACGGGGAAAGCAAGCCC TGAAACCGTGCAAAGTTGTTGCCGGTCTT TGTCAGACCACGGCCTTACACCGGAGCAAGT CGTGGCCATTGCAAGCAATGGGGTGGCAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGAATAACAATGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCCGTGTGTGTCAAGCCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATCGCCAACAACA ACGGCGGTAAGCAGGCGCTGGAAACAGTACAG CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGT CGAACATTGGGGGAAAGCAAGCCCTGGAAACC GTGCAAGGTTGTTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA </p>		
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	<p>TTGCAAATAATAACGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTTCCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGTCGCATGACGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAAGCCCACGGTTTGACGCCTGCAC AAGTGGTCGCCATGCCAGCCATGATGGCGGT AAGCAGGCGCTGGAAACAGTACAGCGCCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCGCCATGTCTCCACGAC GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCGCGTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGTCTTGGTGGACGACCCGCGCTCGATGCAGT CAAAAAGGGTCTGCCTCATGCTCCCGCATTGA TCAAAAGAACCAACCGCGGATTCCCGAGAGA ACTTCCCATCGAGTCGCGGGATCC</p>			
<p>>JAK2_Left TCTGAAAA GACTCTGCA TAL/007/01 5/019/021/ 026/011/01 6/021/029/ 011/017/02 5/027/015/ 019/022/JD S70/ ('TCTGAAAA AGACTCTGCA ' disclosed as SEQ ID No: 426)</p>	<p>GCTAGCaccATGGACTACAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGCATTACC CGGGGTACCTAT GGTGGACTTGAGGACACTCGGTTATTCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCCTCGCGCAACACCACGAGGCGCTTGT GGGGCATGGCTTCACTCATGCCATATTGTCCG CGCTTTCACAGCACCCCTGCGCGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTCCGGC CCTGCCCGAAGCCACGCACGAGGCAATTGTAG GGGTCGGTAAACAGTGGTCGGGAGCGGAGCA CTTGAGGCGCTGTGACTGTGGCGGTGAGCT TAGGGGCGCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCCCGAAGAGAGGGGGAGTA ACAGCGGTAGAGGCAGTGCACCGCTGGCGCAA TGCGCTCACCGGGGCCCTTGAACTGACCC CAGACCAGGTAGTCGCAATCGCGTCACATGAC GGGGAAAGCAAGCCCTGGAAACCGTGCAAAG GTTGTGCCGGTCTTGTCAAGACCACGGCC</p>	<p>273.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDDKMAPKKRKRVI HRGVPVMDLRTLGSQQQEKIKPKVRSTVAQHHEAL VGHGFTHAHIVALSQHPAALGTVAVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQD GQLLKIARKGGVTAVEAVHAWRNALTGAPLNLTPDQV VAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVAI ASNGGKQALETVQRLLPVLCAHGLTPDQVVAIANN NNGKQALETVQRLLPVLCAHGLTPAQVVAIASNIGG KQALETVQRLLPVLCDHGLTPDQVVAIASNIGGKQA LETVQRLLPVLCDHGLTPEQVVAIASNIGGKQALET VQRLLPVLCAHGLTPDQVVAIASNIGGKQALETVQR LLPVLCAHGLTPAQVVAIASNIGGKQALETVQRLLP VLCQDHGLTPDQVVAIANNGGKQALETVQRLLPVL CDHGLTPEQVVAIASNIGGKQALETVQRLLPVLCAH GLTPDQVVAIASHDGGKQALETVQRLLPVLCAHGLT PAQVVAIASNIGGKQALETVQRLLPVLCDHGLTPDQ VVAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVA IASNGGKQALETVQRLLPVLCAHGLTPDQVVAIAN NNGKQALETVQRLLPVLCAHGLTPAQVVAIASHDG GKQALETVQRLLPVLCDHGLTPEQVVAIASNIGGRP</p>	<p>274.</p>

<p>TTACACCGGAGCAAGTCGTGGCCATTGCAAGC AATGGGGGTGGCAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGAATAACAATGGAGGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCCACGGTTTGACGCTGCACAAGTGGTC GCCATCGCCTCCAATATTGGCGGTAAGCAGGC GCTGGAAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCAACATTGGGGGAAA GCAAGCCCTGAAACCGTGCAAAGTTGTGTC CGGTCTTTGTCAAGACCAGGCCTTACACCG GAGCAAGTCGTGGCCATTGCAAGCAACATCGG TGGCAAPCAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTTCTCTGTCAAGCCCAGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGTCGAA CATTGGAGGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCGTGTGTGCAAGCCAC GGTTTGACGCTGCACAAGTGGTCCCATCGC CTCCAATATTGGCGGTAAGCAGGCGTGAAA CAGTACAGCGCCTGCTGCCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGAACATAATGGGGAAAGCAAGCCC TGGAACCGTGCAAAGTTGTTGCCGGTCTT TGTCAAGACCAGGCCTTACACCGGAGCAAGT CGTGCCATTGCAAGCAACATCGGTGGCAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCCAGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCGCATGACGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCGTGTTGTGTCAAGCCCAGGTTTGAC GCCTGCACAAGTGGTCCCATCGCCTCGAATG GCGGCGGTAAGCAGGCGCTGAAACAGTACAG CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGT CACATGACGGGGAAAGCAAGCCCTGAAACC</p>	<p>ALESIVAQLSRPDPALAALNDHLVALACLGGRPALD AVKGLPHAPALIKRTRRRIPERTSHRVAGS</p>
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	GTGCAAGGTTGTTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCAAGCAATGGGGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTTCTCTG TCAAGCCACCGGCTGACTCCCGATCAAGTTG TAGCGATTGCGAATAACAATGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAGCCACGGTTTGACGCCTGCAC AAGTGGTCGCCATCGCCAGCCATGATGGCGGT AAGCAGGCGCTGGAACAGTACAGCGCCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCGCCATTGCTTCTAACATC GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCGCGTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGCTTGGTGGACGACCCCGGCTCGATGCAGT CAAAAAGGGTCTGCCTCATGCTCCCGCATTGA TCAAAAGAACCAACCGGGGATCCCGAGAGA ACTTCCCATCGAGTCGCGGGATCC			
>JAK2_Righ t_TCCATTT TGTCATCGTA _TAL/007/0 12/016/025 /030/015/0 17/025/029 /015/017/0 21/030/012 /019/025/J DS70/ (TCCATTT TGTCATCGTA ' disclosed as SEQ ID NO: 427)	GCTAGCaccATGGACTACAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGCATTACCCCGGGTACCTAT GGTGGACTTGAGGACACTCGGTTATTCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCGTCGCGCAACACCAGGAGCGCTTGT GGGCGATGGCTTCACTCATCGCATATTGTCG CGTTTCACAGCACCTCGCGGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCGAAGCCAGCAGGCAATTGTAG GGTTCGGTAAACAGTGGTCGGAGCGCGAGCA CTTGAGCGCTGCTGACTGTGGCGGTCAGCT TAGGGGGCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGAGGGGGAGTA ACAGCGGTAGAGGAGTGCACGCTGGCGCAA TGCGCTCACCGGGCCCCCTTGAACCTGACCC CAGACCAGGTAGTCGAATCGGCTCACATGAC	275.	ASTMDYKDHDGDYKDHDIDYKDDDDKMAPKKRKGVI HRGVPMVDLRTLGSQQQEKIKPKVRSTVAQHHEAL VHGFTHAHIVALSQHPAALSTVAVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQD GQLLKIARGGVTAVEAVHAWRNALTGAPLNLT VVAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVAI ASHDGGKQALETVQRLLPVLCDHGLTPEQVVAIASN IGGKQALETVQRLLPVLCDHGLTPEQVVAIASNGGG KQALETVQRLLPVLCDHGLTPEQVVAIASNGGGKQ LETVQRLLPVLCDHGLTPEQVVAIASNGGGKQALE TVQRLLPVLCDHGLTPEQVVAIASHGGKQALETVQR LLPVLCDHGLTPEQVVAIASNGGGKQALETVQRLLP VLCQDHGLTPEQVVAIANNGGKQALETVQRLLPVL CDHGLTPEQVVAIASNGGGKQALETVQRLLPVLCDH GLTPEQVVAIASHGGKQALETVQRLLPVLCDHGLT PEQVVAIASNIGGKQALETVQRLLPVLCDHGLTPEQ VVAIASNGGGKQALETVQRLLPVLCDHGLTPEQVVA IASHGGKQALETVQRLLPVLCDHGLTPEQVVAIAN	276.

<p>GGGGAAAGCAAGCCCTGAAACCGTGCAAAG GTTGTGCGCGTCCCTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCGTGGCCATTGCATCC CACGACGGTGGCAAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGTCGAACATTGGAGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCCACGGTTTGACGCCGCAACAAGTGGT GCCATCGCCTCGAATGGCGGGGTAAGCAGGC GCTGAAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCAAACGGAGGGGAAA GCAAGCCCTGAAACCGTGCAAAGTTGTTGC CGGTCTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCAAGCAATGGGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTCTCTGTCAAGCCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGTCGCA TGACGGAGGAAACAAGCATTGAGACTGTCC AACGGCTCCTTCCCGTGTGTCAAGCCAC GGTGTGACGCCGTCACAAGTGGTCCCATCGC CTCGAATGGCGGGTAAGCAGCGCTGAAA CAGTACAGCGCCTGCTGCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGGAACAATAATGGGGAAAGCAAGCCC TGGAAACCGTGCAAAGTTGTTGCCGGTCTT TGTCAAGACCACGGCCTTACACCGAGCAAGT CGTGGCCATTGCAAGCAATGGGGTGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTGTCAAGCCCACGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCGCATGACGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCCGTGTGTGTCAAGCCCACGGTTGAC GCCTGCACAAGTGGTCGCCATCGCTCCAATA TTGGCGGTAAGCAGCGCTGAAACAGTACAG CGCTGCTGCCTGTACTGTGCCAGGATCATGG</p>	<p>NNGGKQALETVQRLLPVLCOAHGLTPAQVVAIASNGG GKQALETVQRLLPVLCOHGLTPEQVVAIASNIGGRP ALESIVAQLSRPDPALAALNDHLVALACLGGRPALD AVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>
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	<p>ACTGACCCAGACCAGGTAGTCGCAATCGCGT CAAACGGAGGGGAAAGCAAGCCCTGGAAACC GTGCAAAGGTTGTTGCCGGTCCCTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCATCCCACGACGGTGGCAAACAGGCTCTT GAGACGGTTCAGACACTTCTCCAGTTCTCTG TCAAGCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCCAATAACAATGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAAGCCACGGTTTGACGCCGTGCAC AAGTGGTCGCCATCGCCTCGAATGGCGGCGGT AAGCAGGCGCTGGAAACAGTACAGCGCCTGCT GCCTGTACTGTCCAGGATCATGGACTGACAC CCGAACAGGTGGTCCCATTGCTTCTAACATC GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCCGATCCCGCGTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGTCTTGGTGGACGACCCGCGCTCGATGCAGT CAAAAGGGTCTGCCCTCATGCTCCCGCATTGA TCAAAGAACAACCGCGGATCCCGAGAGA ACTTCCCATCGAGTCGCGGGATCC</p>			
<p>>KRAS_Left TGAAAATGA CTGAATATA TAL/009/01 1/016/021/ 026/015/01 9/021/027/ 015/019/02 1/026/015/ 016/025/JD S70/ ('TGAAAATG ACTGAATATA ' disclosed as SEQ ID</p>	<p>GCTAGCaccATGGACTACAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGGCATTCACCGGGGTACCTAT GGTGGACTTGAGGACACTCGGTATTCCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCGTCGCGCAACACCAGAGCGCTTGT GGGGCATGGCTTCACTCATGCGCATATTGTCG CGCTTTCACAGCACCCCTGCGGCGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCCGAAGCCACGCACGAGGCAATTGTAG GGGTCGGTAAACAGTGGTCGGAGCGCGAGCA CTTGAGGCGCTGCTGACTGTGGCGGGTGAAGT TAGGGGCGCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCCCGAAGAGAGGGGAGTA ACAGCGGTAGAGGAGTGCACGCTGGCGCAA</p>	<p>277.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDDKMAPKKRKGVI HRGVPMVDLRTLGSQQQEKIKPKVRSSTVAQHHEAL VGHGFTHAHIVALSQHPAALGTVAVKYQDMI AALPEA THEAIVGVGRQWSGARALEALLTVAGELRGPPLQDLD GQLLKIARGGVTAVEAVHAWRNALTGAPLNLTDPQV VAIANNNGGKQALETVQRLLPVLCDHGLTPEQVVAI ASNIGGKQALETVQRLLPVLCAHGLTDPQVVAIASN IGGKQALETVQRLLPVLCAHGLTPAQVVAIASNIGG KQALETVQRLLPVLCDHGLTDPQVVAIASNIGGKQA LETVQRLLPVLCDHGLTPEQVVAIASNGGKQALET VQRLLPVLCAHGLTDPQVVAIANNGGKQALETVQR LLPVLCAHGLTPAQVVAIASNIGGKQALETVQRLLP VLCQDHLTPDQVVAIASHDGGKQALETVQRLLPVL CDHGLTPEQVVAIASNGGKQALETVQRLLPVLCAH GLTPDQVVAIANNGGKQALETVQRLLPVLCAHGLT PAQVVAIASNIGGKQALETVQRLLPVLCDHGLTPDQ</p>	<p>278.</p>

<p>NO: 428)</p>	<p>TGCGCTCACCGGGGGCCCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGCGAACAATAAT GGGGGAAAGCAAGCCCTGGAAACCGTGCAAAG GTTGTGCCCGTCCCTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCGTGGCCATTGCAAGC AACATCGGTGGCAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGTCGAACATTGGAGGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTCCCGTGTGTGTGTC AAGCCACCGGTTTGACGCTGCACAAGTGCTC GCCATCGCCTCCAATATTGGCGGTAAGCAGGC GCTGGAAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCGAACATTGGGGAAA GCAAGCCCTGGAAACCGTGCAAAGGTTGTTGC CGGTCCCTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCAAGCAATGGGGG TGGCAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTCTCTGTCAAGCCCACGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGAATAA CAATGGAGGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTCCCGTGTGTGTCAAGCCAC GGTTTGACGCTGCACAAGTGGTCGCCATCGC CTCCAATATTGGCGGTAAGCAGGCGCTGGAAA CAGTACAGCGCCTGCTGCCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGTCACATGACGGGGAAAGCAAGCCC TGGAAACCGTGCAAAGGTTGTGCGGTCCTT TGTCAAGACCACGGCCTTACACCGGAGCAAGT CGTGGCCATTGCAAGCAATGGGGTGGCAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGAATAACAATGGAG GGAACAAGCATTGGAGACTGTCCAACGGCTC CTCCCGTGTGTGTCAAGCCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATCGCCTCCAATA</p>	<p>VVAIASNIGGKQALETVQRLLPVLCDHGLTPEQVVA IASNGGKQALETVQRLLPVLCDHGLTPEQVVAIAS NIGGKQALETVQRLLPVLCDHGLTPEQVVAIASNGG GKQALETVQRLLPVLCDHGLTPEQVVAIASNIGGRP ALESIVAQLSRPDPALAALNDHLVALACLGGRPALD AVKGLPHAPALIKRTRRRIPERTSHRVAGS</p>
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	<p>TTGGCGGTAAGCAGGCGCTGGAACAGTACAG CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCCGAGACCAGGTAGTCGCAATCGCGT CGAACATGGGGGAAAGCAAGCCCTGAAACC GTGCAAGGTTGTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCAAGCAATGGGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTCTCTG TCAAGCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGTCGAACATTGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTCCCGT GTTGTGTCAAGCCACGGTTTGACGCTGCAC AAGTGGTCGCCATCGCCTCGAATGGCGCGGT AAGCAGGCGCTGGAACAGTACAGCGCCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCGCCATTGCTTCTAACATC GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCCGATCCCGCTTGGCTG CGTTAACGAATGACCATCTGGTGGCCTTGCA TGTCTGGTGGACGCCCGGCTCGATGCAGT CAAAAAGGGTCTGCCATGCTCCCGCATGA TCAAAAGAACCAACGGCGGATTCCGAGAGA ACTTCCCATCGAGTCGCGGATCC</p>			
<p>>KRAS_Righ t_TTGCCTAC GCCACCAGC TAL/010/01 4/017/022/ 030/011/01 7/024/027/ 012/016/02 2/027/011/ 034/JDS71/ 'TTGCCTAC GCCACCAGC' disclosed as SEQ ID</p>	<p>GCTAGCaccATGGACTACAAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACCGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGGCATTACCCCGGGTACCTAT GGTGGACTTGAGGACACTCGGTTATTCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCGTCGCGCAACACCAGGCGCTTGT GGGGCATGGCTTCACTCATGCGCATATTGTCG CGCTTTCACAGCACCCCTGCGGCGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCCAAGCCACGACGAGGCAATTGTAG GGTTCGGTAAACACTGCTGGGAGCGCCAGCA CTTGAGGCGCTGACTGTGGCGGGTGAAGCT TAGGGGGCTCCGCTCAGCTCGACACCGGGC</p>	<p>279.</p>	<p>ASTMDYKDHGDYKDHDIDYKDDDKMAPKKRKRKVI HRGVPMVDLRTLGSQQQEKIKPKVRSTVAQHHEAL VGHGFTHAHIVALSQHPAALGTVAVKYQDMIAALPEA THEATVGVGKQWSGARALEALLTVAGELRGPPLQLDT GQLLKIARKGGVTAVEAVHAWRNALTGAPLNLTPDQV VAIASNNGGKQALETVQRLLPVLCDHGLTPEQVVAI ANNNGKQALETVQRLLPVLCAHGLTPDQVVAIASH DGGKQALETVQRLLPVLCAHGLTPAQVVAIASHDGG KQALETVQRLLPVLCDHGLTPDQVVAIASNNGGKQA LETVQRLLPVLCDHGLTPEQVVAIASNIGKQALET VQRLLPVLCAHGLTPDQVVAIASHDGGKQALETVQR LLPVLCAHGLTPAQVVAIANNGGKQALETVQRLLP VLCQDHGLTPDQVVAIASHDGGKQALETVQRLLPVL CDHGLTPEQVVAIASHDGGKQALETVQRLLPVLCAH</p>	<p>280.</p>

<p>NO: 429)</p>	<p>AGCTGCTGAAGATCGCGAAGAGAGGGGGAGTA ACAGCGGTAGAGGCAGTGCACGCCCTGGCGCAA TCGCGTCAACGGGGCCCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGCGTCAAACGGA GGGGAAAAGCAAGCCCTGAAAACCGTGCAAAG GTTGTGGCCGGTCTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCGTGCCATTGCAAAAT AATAACGGTGGCAAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCCAGTTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGTGCATGACGGAGGGAACAAGCATTGGA GACTGTCCAACGGCTCTCCCGTGTGTGTC AAGCCCACGGTTTGACGCTGCACAAGTGTC GCCATCGCCAGCCATGATGGCGGTAAGCAGGC GCTGGAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCAAACGGAGGGGAAA GCAAGCCCTGGAACCGTGCAAAGTTGTTC CGGTCTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCAAGCAACATCGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCCAGTTCTGTCAAGCCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGTCGCA TGACGGAGGGAACAAGCATTGGAGACTGTCC AACGGCTCTTCCCGTGTGTGTCAAGCCAC GGTTTGACGCTGCACAAGTGGTCGCCATCGC CAACAACAACGGCGTAAGCAGGCGCTGAAA CAGTACAGCGCCTGCTGCCGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGTCACATGACGGGGAAAGCAAGCCC TGGAACCGTGCAAAGTTGTTCGCCGTCTT TGTCAAGACCACGGCCTTACACCGGAGCAAGT CGTGGCCATTGCMCCCACGCGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCAACATTGGAG GGAACAAGCATTGGAGACTGTCCAACGGCTC</p>	<p>GLTPDQVVAIASNIGGKQALETVQRLLPVLCAHGLT PAQVVAIASHDGGKQALETVQRLLPVLCDHGLTPDQ VVAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVA IASNIGGKQALETVQRLLPVLCAHGLTPDQVVAIAN NNGGKQALETVQRLLPVLCAHGLTPEQVVAIASHDQ GRPALESIVAQLSRPDPALAALNDHLVALACLGGRP ALDAVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>
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	<p>CTTCCCGTGTGTGTCAGCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATCGCCAGCCATG ATGGCGGTAAGCAGGCGCTGGAACAGTACAG CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGT CACATGACGGGGAAAGCAAGCCCTGGAACC GTGCAAAGGTTGTTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCAAGCAACATCGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTTCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGAATAACAATGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCTTCCCGT GTTGTGTCAAGCCACGGTCTGACACCCGAAC AGGTGGTCGCCATTGCTTCCCACGACGGAGGA CGGCCAGCCTTGGAGTCCATCGTAGCCCAATT GTCCAGGCCGATCCCGGTTGGCTGCGTTAA CGAATGACCATCTGGTGGCGTTGGCATGTCTT GGTGGACGACCCGCGCTCGATGCAGTCAAAA GGGTCTGCCCTCATGCTCCCGCATGATCAAAA GAACCAACCGGGGATCCCGAGAGAACTTCC CATCGAGTCGGGGATCC</p>			
<p>>MYC_Left_ TGCTTAGACG CTGGATTT T AL/009/012 /020/025/0 26/014/016 /022/029/0 12/020/024 /029/011/0 20/025/JDS 78/ ('TGCTTAGA CGCTGGATTT disclosed</p>	<p>GCTAGCaccATGGACTACAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGTGGGCATTACCCGGGGTACCTAT GGTGGACTTGAGGACACTCGGTTATTCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCGTCGCGCAACACCAGGCGCTTGT GGGGATGGCTTCACTCATGCGCATATTGTCG CGCTTTCACAGCACCTGCGGGCCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCCAAGCCACGACGAGGCAATTGTAG GGGTCGGTAAACAGTGGTCGGGAGCGCGAGCA CTGAGGCGCTGCTGACTGTGGCGGTGAGCT TAGGGGGCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGAGGGGGAGTA</p>	<p>281.</p>	<p>ASTMDYKDHDGDKDHDIDYKDDDDKMAPKKRKRKVI HRGVPMVDLRTLGSQQQEKIKPKVRSVAQHHEAL VGHGFTHAHIVALSQHPAALGTAVVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQLDT GQLLKIARKGVTAVEAVHWRNALTGAPLNLTPDQV VAIANNNGKQALETVQRLLPVLCDHGLTPEQVVAI ASHDGKQALETVQRLLPVLCAHGLTPDQVVAIASN GGGKQALETVQRLLPVLCAHGLTPAQVVAIASNNGG KQALETVQRLLPVLCDHGLTPDQVVAIASNIGGKQ LETVQRLLPVLCDHGLTPEQVVAIANNNGKQALET VQRLLPVLCAHGLTPDQVVAIASNIGGKQALETVQR LLPVLCAHGLTPAQVVAIASHDGKQALETVQRLLP VLCQDHGLTPDQVVAIANNNGKQALETVQRLLPVL CDHGLTPEQVVAIASHDGKQALETVQRLLPVLCAH GLTPDQVVAIASNNGGKQALETVQRLLPVLCAHGLT</p>	<p>282.</p>

<p>as SEQ ID NO: 430)</p>	<p>ACAGCGGTAGAGGCAGTGCACGCCTGGCGCAA TGGCTCACCGGGGGCCCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGCGAACAAATAAT GGGGAAAGCAAGCCCTGGAAACCGTGCAAAG GTTGTTGCCGGTCCCTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCGTGGCCATTGCATCC CACGACGGTGGCAAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGTCCAACGGTGGAGGGAAACAAGCATTTGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCACGGTTTGACGCTGCACAAGTGGTC GCCATCGCCTCGAATGGCGGGTAAGCAGGC GCTGGAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCAACATTGGGGAAA GCAAGCCCTGGAAACCGTCAAGGTTGTGTC CGGTCTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCAAATAATAACGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTCTCTGTCAAGCCACGGGCTG ACTCCGATCAAGTTGTAGCGATTGCGTCGAA CATTGGAGGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCGTGTGTCAAGCCAC GGTTTGACGCTGCACAAGTGGTCGCCATCGC CAGCCATGATGGCGTAAGCAGGCGTGGAAA CAGTACAGCGCCTGCTGCCTGTAAGTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCAACAATAATGGGGAAAGCAAGCCC TGGAAACCGTCAAGGTTGTGCGGTCCTT TGTCAAGACCACGGCCTTACACCGGAGCAAGT CGTGGCCATTGCATCCACGACGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCAACGGTGGAG GGAAACAAGCATTGGAGACTGTCAACGGCTC CTTCCCGTGTGTCAAGCCACGGTTGAC</p>	<p>PAQVVAIANNNGGKQALETVQRLLPVLCQDHGLT VVAIANNNGGKQALETVQRLLPVLCQDHGLTPEQVVA IASNIGGKQALETVQRLLPVLCQAHGLTPEQVVAIAS NGGGKQALETVQRLLPVLCQAHGLTPAQVVAIASNGG GKQALETVQRLLPVLCQDHGLTPEQVVAIASNGGGRP ALESIVAQLSRPDALAALNDHLVALACLGGRPALD AVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>
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	<p>GCCTGCACAAGTGGTCGCCATCGCCAACAACA ACGGCGGTAAGCAGGCGCTGGAACAGTACAG CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGA ACAATAATGGGGAAAGCAAGCCCTGGAACC GTGCAAAGGTTGTTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCAAGCAACATCGGTGGCAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTCTCTG TCAAGCCCACGGGTGACTCCCGATCAAGTTG TAGCGATTGCGTCCAACGGTGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTCCCGT GTTGTGTCAAGCCACGGTTTGACGCTGCAC AAGTGGTCGCCATCGCCTCGAATGGCGGCGT AAGCAGGCGCTGGAAACAGTACAGCGCCTGCT GCCTGTACTGTGCCAGGATCATGACTGACAC CCGAACAGGTGGTCGCCATTGCTTCTAATGGG GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCGCTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGTCTTGGTGGACGACCCGCGCTCGATGCAGT CAAAAGGGTCTGCCTCATGCTCCCGCATGA TCAAAGAACCAACCGGGGATTCGCGAGAGA ACTTCCATCGAGTCGCGGGATCC</p>			
<p>>MYC_Right _TTCGGTGCT TACCTGGTT_ TAL/010/01 2/019/024/ 030/014/01 7/025/030/ 011/017/02 2/030/014/ 019/025/JD S78/ ('TTCGGTGC TTACCTGGTT</p>	<p>GCTAGCaccATGGACTACAAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGCATTACCCGCGGGTACCTAT GGTGGACTTGAGGACACTCGGTATTTCGCAAC AGCAACAGGAGAAAAATCAAGCCTAAGGTCAGG AGCACCCTCGCGCAACACCAGGCGCTTGT GGGGCATGGCTTCACTCATGCCATATTGTCG CGCTTTCACAGCACCCCTGCGGCGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCGAAGCCACGACGAGGCAATTGTAG GGGTCGGTAAACACTGTGTCGGGAGCCGAGCA CTTGAGCGCTGCTGACTGTGGCGGTGAGCT</p>	<p>283.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDDKMAPKKRKGVI HRGVPMVDLRTLGSQQQEKIKPKVRSTVAQHHEAL VGHGFTHAHVALSQHPAALGTVAVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQDLD GQLLKIARKGGVTAVEAVHAWRNALTGAPLNLPDQV VAIASNGGKQALETVQRLLPVLCDHGLTPEQVVAI ASHDGKQALETVQRLLPVLCAHGLTPDQVVAIANN NGGKQALETVQRLLPVLCAHGLTPAQVVAIANNNGG KQALETVQRLLPVLCDHGLTPDQVVAIASNGGKQA LETVQRLLPVLCDHGLTPEQVVAIANNNGGKQALET VQRLLPVLCAHGLTPDQVVAIASHDGKQALETVQR LLPVLCAHGLTPAQVVAIASNGGKQALETVQRLLP VLCDHGLTPDQVVAIASNGGKQALETVQRLLPVL</p>	<p>284.</p>

<p>disclosed as SEQ ID NO: 431)</p>	<p>TAGGGGGCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGAGGGGAGTA ACAGCGGTAGAGGCAGTGCACGCCCTGGCGCAA TGCGCTCACCAGGGCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGCGTCAAACGGA GGGGAAAGCAAGCCCTGGAACCCGTGCAAAG GTTGTTGCCGCTCCTTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCGTGCCATTGCATCC CAGCAGGTGGCAAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTTCTGTGTCAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGAATAACAATGGAGGGAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCCACGGTTTGACGCTGCACAAGTGGTC GCCATCGCCAACAACACGGCGTAAGCAGGC GCTGGAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCAAACGGAGGGGAAA GCAAGCCCTGGAAACCGTGCAAAGTTGTGTC CGGCTCTTGTCAAGACCACGGCTTACACCG GAGCAAGTCGTGGCCATTGCAATAATAACGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTTCTGTCAAGCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGTCGCA TGACCGAGGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCGTGTGTGTCAGCCAC GGTTTGACGCTGCACAAGTGGTCGCCATCGC CTCGAATGGCGCGTAAGCAGGCGTGGAAA CAGTACAGCGCTGCTGCTGACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGTCAAACGGAGGGGAAAGCAAGCCC TGGAACCGTCAAAGGTGTGTCGGTCCCTT TGTCAAGACCACGGCTTACACCGGAGCAAGT CGTGGCCATTGCAAGCAACATCGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTGTCAAGCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCCATGACGGAG</p>	<p>QDHGLTPEQVVAIASNIGGKQALETVQRLLPVLCAH GLTPDQVVAIASHDGGKQALETVQRLLPVLCAHGLT PAQVVAIASHDGGKQALETVQRLLPVLCAHGLTPDQ VVAIASNIGGKQALETVQRLLPVLCAHGLTPEQVVA IANNNGGKQALETVQRLLPVLCAHGLTPDQVVAIAN NNGGKQALETVQRLLPVLCAHGLTPAQVVAIASNGG GKQALETVQRLLPVLCAHGLTPEQVVAIASNGGRP ALESIVAQLSRDPALAAALNDHLVALACLGGRPALD AVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>
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	<p>GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTCCCCTGTTGTGTCAAGCCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATCGCCAGCCATG ATGGCGGTAAGCAGGCGCTGGAACAGTACAG CGCTGCTGCCCTGTACTGTGCCAGGATCATGG ACTGACCCCAGACCAGGTAGTCGCAATCGCGT CAAACGGAGGGGAAAGCAAGCCCTGGAACC GTGCAAAGTGTGTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCAAATAATAACGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTTCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGAATAACAATGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCTTCCCCT GTTGTGTCAAGCCCACGGTTTGACGCTGCAC AAGTGGTCGCCATCGCTCGAATGGCGGGGT AAGCAGGCGCTGGAACAGTACAGCGCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCGCCATTGCTTCTAATGGG GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCGCTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGTCTTGGTGGACGACCCGCGCTCGATGCAGT CAAAAGGGTCTGCCATGCTCCCGCATGA TCAAAGAACCAACCGCGGATPCCCGAGAGA ACTTCCCATCGAGTCGCGGGATCC</p>			
<p>>PTEN_Left _TCCCAGACA TGACAGCC_T AL/007/012 /017/021/0 29/011/017 /021/030/0 14/016/022 /026/014/0 32/JDS71/ ('TCCCAGAC</p>	<p>GCTAGCaccATGGACTACAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGCCCCCAAGAAGAAG AGGAAGGTGGCATTACCAGCGGGTACCTAT GGTGGACTTGAGGACACTCGGTTATTGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCCTCGCGCAACACCAGAGGCGCTTGT GGGGCATGGCTTCACTCATGCGCATATTGTCG CGCTTTCACAGCACCCCTGCGGCGCTTGGGACG GTGGCTGTCAANTACCAAGATATGATTGCGGC CCTGCCCAAGCCACGACGAGGCAATTGTAG</p>	<p>285.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDDKMAPKKRRVGI HRGVPMVDLRTLGSQQQEKIKPKVRSVAQHHEAL VGHGFTHAHIVALSQHPAALGTAVVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQLDT GQLLKIAKRGGVTAVEAVHAWRNALTGAPLNLTDPQV VAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVAI ASHDGGKQALETVQRLLPVLQAHGLTPDQVVAIASH DGGKQALETVQRLLPVLQAHGLTPAQVVAIASNIGG KQALETVQRLLPVLCDHGLTDPQVVAIANNNGGKQA LETVQRLLPVLCDHGLTPEQVVAIASNIGGKQALET VQRLLPVLQAHGLTPDQVVAIASHDGGKQALETVQR</p>	<p>286.</p>

<p>ATGACAGCC' disclosed as SEQ ID NO: 432)</p>	<p>GGGTCGGTAAACACTGGTCGGGAGCCGAGCA CTTGAGGCGCTGTGACTGTGGCGGTGAGCT TAGGGGGCCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGAGGGGAGTA ACAGCGGTAGAGGCAGTGCACGCCGTGGCGCAA TGGCCTCACCGGGGCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGCGTCACATGAC GGGGAAAGCAAGCCCTGGAAACCGTGCAAAG GTTGTTGCCGTCTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCGTGGCCATTGCATCC CACGACGGTGGCAAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGTCGCATGACGGAGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTCCCGTGTGTGTC AAGCCCACGGTTTGACCGCTGCACAAGTGGTC GCCATCGCTCCAATATTGGCGTAAGCAGGC GCTGGAAACAGTACAGCGCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGAACAATAATGGGGAAA GCAAGCCCTGGAAACCGTGCAAAGTTGTGC CGGCTCTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCTGGCCATTGCAAGCAACATCGG TGCCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTCTCTGTCAAGCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGTCGCA TGACGGAGGGAACAAGCATTGGAGACTGTCC AACGGCTCCTCCCGTGTGTGTCAAGCCAC GGTTTGACGCCTGCACAAGTGGTCGCCATCGC CTCCAATATTGGCGTAAGCAGGCGCTGGAAA CAGTACAGCGCTGCTGCCTGACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGTCAAACGGAGGGGAAAGCAAGCCC TGGAACCGTGCAAAGGTTGTTGCCGGTCTT TGTCAAGACCACGGCCTTACACCGGAGCAAGT CGTGGCCATTGCAAATAATAACGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA</p>	<p>LLPVLCQAHGLTPAQVVAIASNIGGKQALETVQRLLP VLCQDHGLTPDQVVAIASNNGGKQALETVQRLLPVLC QDHGLTPEQVVAIANNNGGKQALETVQRLLPVLCQAH GLTPDQVVAIASNIGGKQALETVQRLLPVLCQAHGLT PAQVVAIASHDGGKQALETVQRLLPVLCQDHGLTPDQ VVAIASNIGGKQALETVQRLLPVLCQDHGLTPEQVVA IANNNGGKQALETVQRLLPVLCQAHGLTPDQVVAIAS HDGGKQALETVQRLLPVLCQAHGLTPEQVVAIASHDG GRPALSIVAQLSRPDALAALNDHLVALACLGRPR ALDAVKKGLPHAPALIKRNRRIPERTSHRVAGS</p>
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	<p>GTTCTCTGTCAAGCCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCCGAACATTGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCTGGTTGTGTCAAGCCCACGGTTTGAC GCCTGCACAAGTGGTCCCATCGCCAGCCATG ATGGCGGTAAGCAGGCGCTGAAACAGTACAG CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGT CGAACATTGGGGGAAAGCAAGCCCTGAAACC GTGCAAAGTTGTTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCAAATAATAACGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGTCCCATGACGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAAGCCCACGGTCTGACACCCGAAC AGGTGGTCCCATGCTTCCCACGACGGAGGA CGGCCAGCCTTGAGTCCATCGTAGCCCAATT GTCCAGGCCCGATCCCGCGTTGGCTGGTTAA CGAATGACCATCTGGTGGCGTTGGCATGTCTT GGTGGACGACCCGCGCTCGATGCAGTCAAAAA GGGTCTGCCCTCATGCTCCCGCATTGATCAAAA GAACCAACCGCGGATTCCCGAGAGAACTTCC CATCGAGTCCGGGATCC</p>			
<p>>PTEN_Righ t_TCCTTTG TTTCTGCTAA _TAL/007/0 12/020/025 /030/015/0 19/025/030 /015/017/0 25/029/012 /020/021/J DS70/ ('TCCTTTG</p>	<p>GCTAGCaccATGGACTACAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGCATTACCCGGGGGTACCTAT GGTGGACTTGAGGACACTCGGTTATTCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCCTCGCGCAACACCACGAGGCGCTTGT GGGGATGGCTTCACTCATCGGCATATTGTGCG CGCTTTCACAGCACCCCTGCGGGCTTGGGACG GTGGCTGTCAATACCAAGATATGATTGCGGC CCTGCCCGAAGCCACGACGAGGCAATTGTAG GGGTCGGTAAACAGTGGTCCGGAGCGGAGCA</p>	<p>287.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDDKMAPKKRKGVI HRGVPMVDLRTLGSQQQEKIKPKVRSTVAQHHEAL VGHGFTHAHIVALSQHPAALGTAVVKYQDMIAALPEA THEAIVGVGQWSGARALEALLTVAGELRGPPLQLDT GQLLKIARGGVTAVEAVHAWRNALTGAPLNLTDPQV VAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVAI ASHDGGKQALETVQRLLPVLCOAHGLTPDQVVAIASN GGGKQALETVQRLLPVLCOAHGLTPAQVVAIASNGGG KQALETVQRLLPVLCDHGLTPEQVVAIASNGGGKQ LETVQRLLPVLCDHGLTPEQVVAIASNGGGKQALET VQRLLPVLCOAHGLTPDQVVAIANNGGKQALETVQR LLPVLCOAHGLTPAQVVAIASNGGGKQALETVQRLLP</p>	<p>288.</p>

<p>TTTCTGCTAA ' disclosed as SEQ ID NO: 433)</p>	<p>CTTGAGGCGCTGCTGACTGTGGCGGGTGAAGCT TAGGGGGCTCCGCTCCAGCTCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGAGGGGGAGTA ACAGCGGTAGAGGAGTGCACGCCCTGGCGCAA TGGCGTCAACGGGGCCCCCTTGAACCTGACCC CAGACCAGGTAGTGCACATCGCGTCACATGAC GGGGAAAGCAAGCCCTGGAAACCGTGCAAAG GTTGTTGCCGGTCCCTTGTCAAGACCACGGCC TTACACCGGAGCAAGTGGTGGCCATTGCATCC CAGCAGGTGGCAAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGTCCAACGGTGGAGGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCCACGGTTTGACGCTGCACAAGTGGTC GCCATCGCCTCGAATGGCGCGGTAAGCAGGC GCTGGAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCAAACGGAGGGGAAA GCAAGCCCTGAAACCGTGCAAAGTTGTTGC CGGTCCCTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCAAGCAATGGGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTTCTGTCAAGCCACGGGCTG ACTCCGATCAAGTTGTAGCGATTGCGAATAA CAATGGAGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCGTGTGTGTCAAGCCAC GGTTTGACGCTGCACAAGTGGTGCATCGC CTCGAATGGCGCGTAAGCAGGCGTGAAA CAGTACAGCGCCTGCTGCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGTCAAACGGAGGGGAAAGCAAGCCC TGGAAACCGTCAAAGGTTGTTGCCGGTCCCTT TGTCAGACCACGGCCTTACACCGGAGCAAGT CGTGCCATTGCAAGCAATGGGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCACGGGCTGACTCCCGA</p>	<p>VLCQDHGLTPDQVVAIASNGGKQALETVQRLLPVLC QDHGLTPEQVVAIASNGGKQALETVQRLLPVLCQAH GLTPDQVVAIASHDGKQALETVQRLLPVLCQAHGLT PAQVVAIASNGGKQALETVQRLLPVLCQDHGLTPDQ VVAIANNGGKQALETVQRLLPVLCQDHGLTPEQVVA TASHDGGKQALETVQRLLPVLCQAHGLTPDQVVAIAS NGGKQALETVQRLLPVLCQAHGLTPAQVVAIASNIG GKQALETVQRLLPVLCQDHGLTPEQVVAIASNIGGRP ALESIVAQLSRPDPALAALNDHLVALACLGGRPALD AVKGLPHAPALIKRTRRRIPERTSHRVAGS</p>
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	<p>TCAAGTTGTAGCGATTGCGTCGCATGACGGAG GGAAACAAGCATTGGAGACTGTCCAACGGGTC CTGCCCGTGTGTGTCAAGCCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATCGCCTCGAATG GCGGCGGTAAGCAGGCGCTGGAACAGTACAG CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCACAGCAGGTAGTCGCAATCGCGA ACAATAATGGGGAAAGCAAGCCCTGGAACC GTGCAAAGGTTGTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCATCCCACGACGGTGGCAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGTCCAACGGTGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAAGCCCACGGTTGACGCCCTGCAC AAGTGGTCGCCATCGCCTCCAATATGGCGGT AAGCAGGCGCTGGAACAGTACAGCGCCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCCCATGCTTCTAACATC GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCGCTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGTCTTGGTGGACGCCCGCTCGATGCAGT CAAAAGGGTCTGCCTCATGCTCCCGCATTGA TCAAAGAACCAACCGGGGATTCGCGAGAGA ACTTCCCATCGAGTCGCGGGATCC</p>			
<p>>TP53_Left TTGCCGTCC CAAGCAATG TAL/010/01 4/017/022/ 029/015/01 7/022/027/ 011/016/02 4/027/011/ 016/025/JD</p>	<p>GCTAGCaccATGGACTACAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGCATTACCGCGGGGTACCTAT GGTGGACTTGAGGACACTCGGTTATTTCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCGTCGCGCAACACCACGAGGCGCTTGT GGGCATGGCTTCACTCATCGCATATTGTCG CGCTTTCACAGCACCTGCGGCGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC</p>	<p>289.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDDKMAPKKRKRKVI HRGVPMVDLRTLGSQQQEKIKPKVRSTVAQHHEAL VGHGFTHAHIVALSQHPAALGTAVVKYQDMIAALPEA THEAIVGVKQWSGARALEALLTVAGELRGPPLQLDT GQLLKIAKRGGVTAVEAVHAWRNALTGAPLNLTPDQV VAIASNGGKQALETVQRLLPVLCDHGLTPEQVVAI ANNNGKQALETVQRLLPVLCAHGLTPDQVVAIASH DGGKQALETVQRLLPVLCAHGLTPAQVVAIASHDGG KQALETVQRLLPVLCDHGLTPDQVVAIANNNGKQQA LETVQRLLPVLCDHGLTPEQVVAIASNGGKQALET</p>	<p>290.</p>

<p>S74/ (TTGCCGTC CCAAGCAATG disclosed as SEQ ID NO: 434)</p>	<p>CCTGCCGAAGCCACGCACGAGGCAATTGTAG GGGTCCGTAACAGTGGTCGGGAGCGCGAGCA CTTGAGGCGCTGCTGACTGTGGCGGTGAGCT TAGGGGGCCTCCGCTCCAGCTCGACACCGGGC AGTGCTGAAGATCGCGAAGAGAGGGGGAGTA ACAGCGGTAGAGGCAGTGCACGCCTGGCGCAA TCCGCTCACCGGGCCCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGCGTCAAACGGA GGGGAAAGCAAGCCCTGGAACCGTGCAAAG GTTGTTGCCGGTCTTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCGTGGCCATTGCAAA AATAACGGTGGCAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT CGCTCGCATGACGGAGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCACGGTTTGACGCTGCACAAGTGGTC GCCATCGCCAGCCATGATGGCGGTAAGCAGGC GCTGGAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGAACAATAATGGGGAAA GCAAGCCCTGGAACCGTGCAAGGTTGTTGC CGGTCTTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCAAGCAATGGGG TGGCAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTCTCTGTCAAGCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGTCGA TGACGGAGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTCCCGTGTGTGTCAGCCAC GGTTTGACGCTGCACAAGTGGTCGCCATCGC CAGCCATGATGGCGTAAGCAGGCGCTGGAAA CAGTACAGCGCTGCTGCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGTCACATGACGGGGAAAGCAAGCCC TGGAAACCGTGCAAGGTTGTTGCCGCTCCTT TGTCAAGACCAGGCTTACACCGGAGCAAGT CGTGGCCATTGCAAGCAACATCGGTGGCAAC</p>	<p>VQRLLPVLCOAHGLTPDQVVAIASHDGGKQALETVQR LLPVLCOAHGLTPAQVVAIASHDGGKQALETVQRLLP VLCQDHGLTPDQVVAIASHDGGKQALETVQRLLPVL QDHGLTPEQVVAIASNIGGKQALETVQRLLPVLCOAH GLTPDQVVAIASNIGGKQALETVQRLLPVLCOAHGLT PAQVVAIANNGGKQALETVQRLLPVLCOAHGLTPDQ VVAIASHDGGKQALETVQRLLPVLCOAHGLTPEQVVA IASNIGGKQALETVQRLLPVLCOAHGLTPDQVVAIAS NIGGKQALETVQRLLPVLCOAHGLTPAQVVAIASNGG GKQALETVQRLLPVLCOAHGLTPEQVVAIANNGGRP ALESIVAQLSRPDAALNDHLVALACLGRPALD AVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>
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	<p>AGGCTCTTGAGACGGTTCAGAGACTTCTCCCA GTTCTCTGTCAAGCCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCGAACATTGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTCCCGTGTGTGTCAAGCCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATCGCCAACAACA ACGGCGGTAAGCAGGCGCTGAAACAGTACAG CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACAGGTAGTTCGCAATCGCGT CACATGACGGGGAAAGCAAGCCCTGAAACC GTGCAAAGGTTGTGCGGTCCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCAAGCAACATCGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGTCGAACATTGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAAGCCCACGGTTTGACGCGTGCAC AAGTGGTCGCCATCGCCTCGAATGGCGCGGT AAGCAGGCGCTGAAACAGTACAGCGCCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGETCGCCATTGCTAATAATAAC GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCCGATCCCGGTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGTCTTGGTGGACGACCCGCGCTCGATGCAGT CAAAAGGGTCTGCCTCATGCTCCCGCATTGA TCAAAGAACCAACCGCGGATTCCCGAGAGA ACTTCCCATCGAGTCGCGGGATCC</p>			
<p>>TP53_Righ t_TGTTCAAT ATCGTCCGGG _TAL/009/0 15/020/022 /026/011/0 20/021/030 /012/019/0</p>	<p>GCTAGCaccATGGACTACAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGGCATTACCCGCGGGTACCTAT GGTGGACTTGAGGACACTCGGTATTCGCAAC AGCAACAGGAGAAAAATCAAGCCTAAGGTCAGG AGCACCGTCGCGCAACACCACGAGCGCTTGT GGGGCATGGCTTCACTCATGCGCATATTGTCCG</p>	<p>291.</p>	<p>ASTMDYKDHGDYKDHIDYKDDDKMAPKKRKGVI HRGVPMVDLRTLGSQQQEKIKPKVRSTVAQHHEAL VGHGFTHAHIVALSQHPAALGTVAVKYQDMIAALPEA THEATVGVKQWSGARALEALLTVAGELRGPPLQD GQLLKIARGGVTAVEAVHAWRNALTGAPLNLPDQV VAIANNNGGQALETVQRLLPVLCDHGLTPEQVVAI ASNGGKQALETVQRLLPVLCAHGLTPDQVVAIASN GGGKQALETVQRLLPVLCAHGLTPAQVVAIASHDGG</p>	<p>292.</p>

<p>25/027/012 /019/024/J DS74/ (TGTTC AAT ATCGTCCGGG ' disclosed as SEQ ID NO: 435)</p>	<p>CGCTTTCACAGCACCCCTGCGGGCCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCGAAGCCACGCACGAGGCAATTGTAG GGGTCCGTAACAGTGGTCCGGAGCGGAGCA CTTGAGGCGCTGCTGACTGTGGCGGTGAGCT TAGGGGGCCCTCCGCTCCAGCTCGACACCGGGC ACCTGCTGAAGATCGCGAAGAGAGGGGAGTA ACAGCGGTAGAGGCAGTGCACGCCCTGGCGCAA TGCGCTCACCGGGGCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGCGAACAATAAT GGGGAAAGCAAGCCCTGGAACCGTGC AAAG GTTGTGCGCGTCTTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCTGTCGATGCAAGC AATGGGGTGGCAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGTCCAACGGTGGAGGAAACAAGCATTGGA GACTGTCCAACGGTCTCTCCCGTGTGTGTC AAGCCCACGGTTTGACGCCCTGCACAAGTGGTC GCCATCGCCAGCCATGATGGCGGTAAAGCAGC GCTGAAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCAAGCATTGGGGAAA GCAAGCCCTGGAACCGTCAAGGTTGTTGC CGGTCTTTGTCAAGACCACGGCTTACACCG GAGCAAGTCTGGCCATTGCAAGCAACATCGG TGGCAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTCTCTGTCAAGCCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGTCAA CGGTGGAGGAAACAAGCATTGGAGACTGTCC AACGGCTCTTCCCGTGTGTGTCAAGCCAC GGTTTGACGCCCTGCACAAGTGGTCCCATCGC CTCCAATATTGGCGGTAAGCAGGCGCTGGAAA CAGTACAGCGCCTGCTGCCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGTCAAACGGAGGGGAAAGCAAGCCC TGGAAACCGTCAAGGTTGTTGCCGGTCTT</p>	<p>KQALETVQRLLPVLCDHGLTPDQVVAIASNIGGKQA LETVQRLLPVLCDHGLTPEQVVAIASNIGGKQALET VQRLLPVLCDHGLTPEQVVAIASNIGGKQALETVQR LLPVLCDHGLTPEQVVAIASNIGGKQALETVQRLLP VLCDHGLTPEQVVAIASNIGGKQALETVQRLLPVLCD HGLTPEQVVAIASNIGGKQALETVQRLLPVLCDHGL TPEQVVAIANNNNGKQALETVQRLLPVLCDHGLT PEQVVAIASNIGGKQALETVQRLLPVLCDHGLTPEQ VVAIASHGGKQALETVQRLLPVLCDHGLTPEQVVA IASHGGKQALETVQRLLPVLCDHGLTPEQVVAIAN NNNGKQALETVQRLLPVLCDHGLTPEQVVAIANNN GKQALETVQRLLPVLCDHGLTPEQVVAIANNNGRP ALESIVAQLSRPDPALAAALTNHDLVALACLGGRPALD AVKKGLPHAPALIKRTRRRIPERTSHRVAGS</p>
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	<p>TGTC AAGACCACGGCCTTACACCGGAGCAAGT CGTGGCCATTGCATCCCACGACGGTGCCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGAATAACAATGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTCCCCTGTTGTGTCAAGCCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATCGCCTCGAATG GCGGCGGTAAGCAGGCGCTGGAAACAGTACAG CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCACAGACCAGGTAGTCGCAATCGCGT CACATGACGGGGAAAGCAAGCCCTGGAAACC GTGCAAAGGTTGTTGCCGGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCATCCCACGACGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTTCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGAATAACAATGGAGGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCCT GTTGTGTCAAGCCCACGGTTGACGCCTGCAC AAGTGGTCGCCATCGCCAACAACAACGGCGGT AAGCAGGCGCTGGAAACAGTACAGCGCCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCGCCATTGCTAATAATAAC GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCGGTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGTCTTGGTGGACGACCCGGCTCGATGCAGT CAAAAAGGCTCTGCCTCATGCTCCCGCATTGA TCAAAAGAACCAACCGGCGGATTCCCGAGAGA ACTTCCCATCGAGTCGCGGGATCC</p>			
<p>>XPA_Left_ TGGCCAGAG ATGGCGGC_T AL/009/014 /019/022/0 27/011/019</p>	<p>GCTAGCaccATGGACTACAAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGCCCCCAAGAAGAAG AGGAAGGTGGGCATTACCCCGGGTACCTAT GGTGGACTTGAGGACACTCGGTTATTCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG</p>	<p>293.</p>	<p>ASTMDYKDHDGDKHDIDYKDDDDKMAPKKRKRKVI HRGVPMVDLRLGYSQQQEKIKPKVRSVVAQHHEAL VGHGFTHAHIVALSQHPAALGTAVVKYQDMIALPEA THEAIVGVKQWSGARALEALLTVAGELRGPPLQDLD GQLLKIARGGVTAVEAVHAWRNALTGAPLNLTDPQV VAIANNNGGKQALETVQRLLPVLCDHGLTPEQVVAI</p>	<p>294.</p>

<p>/021/029/0 11/020/024 /029/012/0 19/024/JDS 71/ ('TGGGCCAG AGATGGCGGC ' disclosed as SEQ ID NO: 435)</p>	<p>AGCACCGTCGGCGAACACCACGAGGCGCTTGT GGGGCATGGCTTCACTCATGCGCATATTGTCG CGCTTTCACAGCACCCCTGCGGCGCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCGAAGCCACGACGAGGCAATTGTAG GGTTCGGTAAACAGTGGTTCGGGAGCGCGAGCA CTTGAGGCGCTGCTGACTGTGGCGGTGAGCT TAGGGGGCTCCGCTCCAGCTCGACACCGGGC AGTCTGCTGAAGATCGCGAAGAGAGGGGAGTA ACAGCGGTAGAGGCAATGACGCTGGCGCAA TCCGCTCACCGGGCCCCCTTGAACTGACCC CAGACCAGGTAGTCGCAATCGCGAACAAATAT GGGGAAAGCAAGCCCTGGAACCGTGCAAAG GTTGTTGCCGGTCTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCTGGCCATTGCAAAAT AATACCGGTGGCAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTCTCTGTCAAGCCC ACGGGCTGACTCCCAGTCAAGTTGTAGCGATT GCGAATAACAATGGAGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTCCCGTGTGTGTC AAGCCCACGGTTGACGCTGCACAAGTGGTC GCCATCGCCAGCCATGATGGCGGTAAGCAGGC GCTGGAACAGTACAGCCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCACATGACGGGGAAA GCAAGCCCTGGAACCGTGCAAGGTTGTTGC CGGTCTTTGTCAAGACCAGGCTTACACCG GAGCAAGTCTGGCCATTGCAAGCAACATCGG TGGCAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTCTCTGTCAAGCCACGGGCTG ACTCCGATCAAGTTGTAGCGATTGCGAATAA CAATGGAGGAAACAAGCATTGGAGACTGTCC AACGGCTCTTCCCGTGTGTGTCAAGCCAC GGTTTGACGCTGCACAAGTGGTCCGATCGC CTCCAATATTGGCGTAAGCAGGCGCTGAAA CAGTACAGCCCTGCTGCCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCCG</p>	<p>ANNNGGKQALETVQRLLPVLCAHGLTPDQVVAIANN NGGKQALETVQRLLPVLCAHGLTPAQVVAIASHDGG KQALETVQRLLPVLCDHGLTPDQVVAIASHDGGKQA LETVQRLLPVLCDHGLTPEQVVAIASNIGGKQALET VQRLLPVLCAHGLTPDQVVAIANNNGGKQALETVQR LLPVLCAHGLTPAQVVAIASNIGGKQALETVQRLLP VLCQDHGLTPDQVVAIANNNGGKQALETVQRLLPVL CDHGLTPEQVVAIASNIGGKQALETVQRLLPVLCAH GLTPDQVVAIASNIGGKQALETVQRLLPVLCAHGLT PAQVVAIANNNGGKQALETVQRLLPVLCDHGLTPDQ VVAIANNNGGKQALETVQRLLPVLCDHGLTPEQVVA IASHDGGKQALETVQRLLPVLCAHGLTPDQVVAIAN NNGGKQALETVQRLLPVLCAHGLTPAQVVAIANNG GKQALETVQRLLPVLCDHGLTPEQVVAIASHDGGRP ALESIVAQLSRPDPALAAALNDHLVALACLGGRPALD AVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>
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	<p>AATCGGAACAATAATGGGGAAAGCAAGCCC TGGAACCGTGCAAAGGTTGTTGCCGGTCCTT TGTCAAGACCACGGCCTTACACCGGAGCAAGT CGTGGCCATTGCAAGCAACATCGGTGGCAAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCCA GTTCTCTGTCAAGCCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCCAACGGTGGAG GGAAACAAGCATTGGGAGACTGTCCAACGGCTC CTTCCTCGTGTGTGTCAAGCCCACGGTTTGAC GCCTGCACAAGTGGTCCGCATCGCCAACAACA ACGGCGGTAAGCAGGGCCTGAAACAGTACAG CGCCTGCTGCCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCCGAATCGCGA ACAATAATGGGGAAAGCAAGCCCTGGAACC GTGCAAAGGTTGTGCGGGTCCCTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCCGTGGCCA TTGCATCCCACGCGGTGGCAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTTCTCTG TCAAGCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCCAATAACAATGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAAGCCCACGGTTTGACGCTGCAC AAGTGGTCCCATCGCCAACAACAACGGCGGT AAGCAGGCGCTGGAACAGTACAGGCGCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCCCATGCTTCCCACGAC GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCGGTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGTCTTGGTGGACGACCCCGCTCGATGCAGT CAAAAAGGCTCGCCTCATGCTCCCGCATTGA TCAAAGAACCAACGGCGGATTCCCGAGAGA ACTTCCCATCGAGTCGCGGGATCC</p>			
<p>>XPA_Right _TAAAGCCG CGCCTCCGG TAL/006/01</p>	<p>GCTAGCaccATGGACTACAAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGTGGGCATTACCCCGGGGTACCTAT</p>	<p>295.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDKMAPKKRKRKVI HRGVPMVDLRTLGSQQQEKIKPKVRSTVAQHHEAL VGHGFTHAHIVALSQHPAALGTVAVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQLDT</p>	<p>296.</p>

<p>1/016/024/ 027/012/01 9/022/027/ 014/017/02 2/030/012/ 017/024/JD S74/ ('TAAAGCCG CCGCCTCCGG ' disclosed as SEQ ID NO: 437)</p>	<p>GGTGGACTTGAGGACACTCGGTTATTCCGCAAC AGCAACAGGAGAAAAATCAAGCCTAAGGTCAGG AGCACCGTCCGGCAACACCACGAGGCGCTTGT GGGGCATGGCTTCACTCATGCGCATATTGTGCG CGCTTTCACAGCACCCCTGCGGCGCTTGGGACG GTGGCTGTCAATACCAAGATATGATTGCGGC CCTGCCCGAAGCCACGCACGAGGCAATTGTAG GGGTCCGTAACAGTGGTCCGGAGCCGCGAGCA CTTGAGGCGCTGCTGACTGTGGCGGGTGAGCT TAGGGGGCCTCCGCTCCAGCTCGACACCGGGC AGCTCCTGAAGATCCGGAAGAGAGGGGAGTA ACAGCGGTAGAGGAGTGCACGCCCTGGCGCAA TGCGCTCACCGGGGCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGCGTCAACATT GGGGAAAGCAAGCCCTGGAAACCGTGCAAAG GTTGTGCCGGTCTTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCCGTGGCCATTGCAAGC AACATCGGTGGCAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGTCAACATTGGAGGAAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCCACGGTTTGACGCCCTGCACAAGTGGTC GCCATCGCCAACAACAACGGCGTAAGCAGGC GCTGGAACAGTACAGCGCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGGTCACATGACGGGGAAA GCAAGCCCTGGAAACCGTCAAGGTTGTTGC CGGTCTTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCATCCCACGACGG TGGCAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTCTCTGTCAAGCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGAATAA CAATGGAGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTCCCGTGTGTGTCAAGCCAC GGTTTGACGCCCTGCACAAGTGGTCGCCATCGC CAGCCATGATGGCGTAAGCAGGCGTGAAA</p>	<p>GQLLKIARGGVTAVEAVHAWRNALTGAPLNLTDPQV VAIASNIGGKQALETVQRLLPVLCDHGLTPEQVVAI ASNIGGKQALETVQRLLPVLCAHGLTDPQVVAIASN IGGKQALETVQRLLPVLCAHGLTPAQVVAIANNNGG KQALETVQRLLPVLCDHGLTDPQVVAIASHDGKQA LETVQRLLPVLCDHGLTPEQVVAIASHDGKQALET VQRLLPVLCAHGLTDPQVVAIANNNGKQALETVQR LLPVLCAHGLTPAQVVAIASHDGKQALETVQRLLP VLCQDHGLTDPQVVAIASHDGKQALETVQRLLPVL CDHGLTPEQVVAIANNNGKQALETVQRLLPVLCAH GLTDPQVVAIASHDGKQALETVQRLLPVLCAHGLT PAQVVAIASHDGKQALETVQRLLPVLCDHGLTDPQ VVAIASNIGGKQALETVQRLLPVLCDHGLTPEQVVA IASHDGKQALETVQRLLPVLCAHGLTDPQVVAIAS HDGKQALETVQRLLPVLCAHGLTPAQVVAIANNG GKQALETVQRLLPVLCDHGLTPEQVVAIANNGGRP ALESIVAQLSRPDPALAALTDHVALACLGGRPALD AVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>
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	<p>CAGTACAGGCGCTGCTGCCTGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCCCGTCACATGACGGGGAAAGCAAGCCC TGGAAACCGTGCAAAGGTTGTTGCCGGTCCTT TGTCAAGACCACGGCCTTACACCGGAGCAAGT CGTGCCATTGCAAATAATAACGGTGGCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCGCATGACGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCCGTGTGTGTCAAGCCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATCGCCAGCCATG ATGGCGGTAAGCAGGCGCTGAAACAGTACAG CGCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGT CAAACGGAGGGGAAAGCAAGCCCTGAAACC GTGCAAAGGTTGTGCGCGTCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGCCA TTGCATCCCACGACGGTGGCAAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGTCGCATGACGGAGGAAACAA GCATGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAAGCCCACGGTTTGACGCTGCAC AAGTGGTCGCCATCGCCAACAACAACGGCGGT AAGCAGGCGCTGAAACAGTACAGCGCCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGETGGTCCCATGCTAATAATAAC GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCGGTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGTCTTGGTGGACACCCGGCTCGATGCAGT CAAAAAGGGTCTGCCTCATGCTCCCGCATGA TCAAAGAACCAACCGGGGATTCCCGAGAGA ACTTCCCATCGAGTCGCGGGATCC</p>	<p>297.</p>	<p>ASTMDYKDHDGDYKDHDIDYKDDDDKMAPKKRKGVI HRGVPMVDLRTLGSQQQEQEKIKPKVRSTVAQHHEAL</p>	<p>298.</p>
<p>>XPC_Left_ TGCCCAGACA</p>	<p>GCTAGCaccATGGACTACAAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG</p>	<p>297.</p>	<p>ASTMDYKDHDGDYKDHDIDYKDDDDKMAPKKRKGVI HRGVPMVDLRTLGSQQQEQEKIKPKVRSTVAQHHEAL</p>	<p>298.</p>

<p>AGCAACAT T AL/009/012 /017/022/0 26/014/016 /022/026/0 11/019/022 /026/011/0 17/021/JDS 78/ 'TGCCCAGA CAAGCAACAT ' disclosed as SEQ ID NO: 438)</p>	<p>ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGGCATTACCGGGGTACCTAT GGTGGACTTGAGGACACTCGGTATTTCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCGTCGGCAACACCACGAGGCGCTTGT GGGCGATGGCTTCACTCATGCGCATATTGTCG CGCTTTCACAGCACCTGCGGGCCTTGGGACG GTGGCTGTCAAATACCAAGATATGATTGCGGC CCTGCCGAAGCCACGACGAGGCAATTGTAG GGTCCGGTAAACAGTGGTCCGGAGCGCGAGCA CTTGAGGCGCTGCTGACTGTGGCGGTGAGCT TAGGGGGCTCCGCTCCAGTTCGACACCGGGC AGCTGCTGAAGATCGCGAAGAGGGGGAGTA ACAGCGGTAGAGGCGAGTGCACGCTGGCGCAA TCCGCTCACCGGGGGCCCCCTTGAACCTGACCC CAGACCAGGTAGTCCGAATCGCGAACAATAAT GGGGAAAGCAAGCCCTGGAAACCGTGCAAAG GTTGTTGCCGGTCTTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCTGTCGATTCGATCC CAGCAGGTTGCAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGTCCGATGACGGAGGAAACAAGCATTTGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCCACGGTTTGACGCTGCACAAGTGGTC GCCATCGCCAGCCATGATGGCGGTAAGCAGGC GCTGAAAACAGTACAGCGCCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAG GTAGTCGCAATCGCGTCGAACATTTGGGGAAA GCAAGCCCTGAAAACCGTGAAGGTTGTTGC CGGTCTTTGTCAAGACCAGGCTTACACCG GAGCAAGTCGTGGCCATTGCAAATAATAACGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTTCCTGTCAAGCCACGGGCTG ACTCCGATCAAGTTGTAGCGATTGCGTCGAA CATTGGAGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCGTGTGTCAAGCCAC</p>	<p>VHGFTHAHIVALSQHPAALGTAVVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQDLD GQLLKIARKGVTAVEAVHAWRNALTGAPLNLTPDQV VAIANNNGGKQALETVQRLLPVLCDHGLTPEQVVAI ASHDGGKQALETVQRLLPVLCDHGLTPEQVVAIASH DGGKQALETVQRLLPVLCDHGLTPEQVVAIASHDGG KQALETVQRLLPVLCDHGLTPEQVVAIASNIGGKQA LETVQRLLPVLCDHGLTPEQVVAIANNNGGKQALET VQRLLPVLCDHGLTPEQVVAIASNIGGKQALETVQR LLPVLCDHGLTPEQVVAIASHDGGKQALETVQRLLP VLCDHGLTPEQVVAIASNIGGKQALETVQRLLPVLCD HGLTPEQVVAIASNIGGKQALETVQRLLPVLCDHGL TPEQVVAIANNNGGKQALETVQRLLPVLCDHGLT PEQVVAIASHDGGKQALETVQRLLPVLCDHGLTPEQ VVAIASNIGGKQALETVQRLLPVLCDHGLTPEQVVA IASNIGGKQALETVQRLLPVLCDHGLTPEQVVAIAS HDGGKQALETVQRLLPVLCDHGLTPEQVVAIASNIG GKQALETVQRLLPVLCDHGLTPEQVVAIASNNGGRP ALESIVAQLSRPDPALAALTNHDLVALACLGRPALD AVKKGLPHAPALIKRTNRRIPERTSHRVAGS</p>
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<p>GGTTTGACGCCTGCACAAGTGGTCGCCATCGC CAGCCATGATGGCGGTAAGCAGGCGTGGAAA CAGTACAGCGCCTGCTGCCTGACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGTCGAACATTGGGGAAAGCAAGCCC TGGAAACCGTGCAAAGGTTGTTGCCGGTCCTT TGTCAAGACCACGGCCTTACACCGGAGCAAGT CGTGGCCATTGCAAGCAACATCGGTGCCAAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCCTGTCAAGCCCACGGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGAATAACAATGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCCGTGTGTGTCAAGCCCACGGTTTGAC GCCTGCACAAGTGGTCGCCATCGCCAGCCATG ATGGCGGTAAGCAGGCGCTGGAAACAGTACAG CGCCTGCTGCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGT CGAACATTGGGGAAAGCAAGCCCTGGAAACC GTGCAAAGGTTGTTGCCGGTCCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCAAGCAACATCGGTGGCAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTTCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCGTCGCATGACGGAGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCGT GTTGTGTCAAGCCACGGTTTGACGCCCTGCAC AAGTGGTCGCCATCGCCTCCAATATTGGCGGT AAGCAGGCGCTGGAAACAGTACAGCGCCTGCT GCCGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCGCCATTGCTTCTAATGGG GGAGGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCGGTTGGCTG CGTTAACGAATGACCATCTGGTGGCGTTGGCA TGTCTTGGTGGACGACCCCGCTCGATGCAGT CAAAAAGGGTCTGCCTCATGCTCCCGCATTGA TCAAAGAACAACCGGGGATTCCTCCGAGAGA ACTTCCCATCGAGTCGGGGATCC</p>		
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<p>>XPC_Right _TCCCCGCGG CTCCCCGCC TAL/007/01 2/017/022/ 029/012/01 9/024/027/ 015/017/02 2/027/012/ 019/022/JD S71/ ('TCCCCGCG GCTCCCCGCC ' disclosed as SEQ ID NO: 439)</p>	<p>GCTAGCaccATGGACTACAAAGACCATGACGG TGATTATAAAGATCATGACATCGATTACAAGG ATGACGATGACAAGATGGCCCCAAGAAGAAG AGGAAGGTGGGCATTCACCGCGGGTACCTAT GGTGGACTTGAGGACACTCGGTATTTCGCAAC AGCAACAGGAGAAAATCAAGCCTAAGGTCAGG AGCACCGTCGCGCAACACCACGAGGCGCTTGT GGGGCATGGCTTCACTCATGCGCATATTGTCG CGCTTTCACAGCACCCCTGCGGCGCTGGGACG GTGGCTGTCAAAATACCAAGATATGATTGCGGC CCTGCCCGAAGCCACGCACGAGGCAATTGTAG GGGTCGGTAAACAGTGGTCGGGAGCGCGAGCA CTTGAGGCGCTGTGACTGTGGCGGTTGAGCT TAGGGGGCCCTCCGCTCCAGCTCGACACCGGGC ACCTCCTGAAATCGCGAAAGAGGGGGAGTA ACAGCGGTAGAGGCAGTGCACGCTGGCGCAA TGCGCTCACCGGGCCCCCTTGAACCTGACCC CAGACCAGGTAGTCGCAATCGCGTCACATGAC GGGGAAAGCAAGCCCTGGAACCGTGCAAAG GTTGTGGCCGCTCCTTGTCAAGACCACGGCC TTACACCGGAGCAAGTCTGGCCATTGCATCC CACGACGGTGGCAACAGGCTCTTGAGACGGT TCAGAGACTTCTCCAGTCTCTGTCAAGCCC ACGGGCTGACTCCCGATCAAGTTGTAGCGATT GCGTCGCATGACCGAGGGAACAAGCATTGGA GACTGTCCAACGGCTCCTTCCCGTGTGTGTC AAGCCCACGGTTTGACGCCTGCACAAGTGGTC GCCATCGCCAGCCATGATGGCGGTAAGCAGGC GCTGGAACAGTACAGCGCTGCTGCCTGTAC TGTGCCAGGATCATGGACTGACCCAGACCAg GTAGTCGCAATCGCGAACAATAATGGGGAAA GCAAGCCCTGGAACCGTGCAAAGTTGTTGC CGGTCTTTGTCAAGACCACGGCCTTACACCG GAGCAAGTCGTGGCCATTGCATCCCACGACGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGAC TTCTCCAGTCTCTGTCAAGCCACGGGCTG ACTCCCGATCAAGTTGTAGCGATTGCGAATAA</p>	<p>299.</p>	<p>ASTMDYKDHGDYKDHDIYKDDDDKMAPKKRKRKVI HRGVPMVLRITLGYSQQQEKIKPKVRSTVAQHHEAL VGHGETHAHIVALSQHPAALGTVAVKYQDMIAALPEA THEAIVGVGKQWSGARALEALLTVAGELRGPPLQDLD GQLLKIARGGVTAVEAVHAWRNALTGAPLNLTDPQV VAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVAI ASHDGGKQALETVQRLLPVLCAHGLTDPQVVAIASH DGGKQALETVQRLLPVLCAHGLTDPQVVAIASHDGG KQALETVQRLLPVLCDHGLTDPQVVAIANNGGKQ LETVQRLLPVLCDHGLTPEQVVAIASHDGGKQALET VQRLLPVLCAHGLTDPQVVAIANNGGKQALETVQR LLPVLCAHGLTDPQVVAIANNGGKQALETVQRLLP VLCQDHLTDPQVVAIASHDGGKQALETVQRLLPVL CDHGLTPEQVVAIASNGGKQALETVQRLLPVLCAH GLTDPQVVAIASHDGGKQALETVQRLLPVLCAHGLT PAQVVAIASHDGGKQALETVQRLLPVLCDHGLTDPQ VVAIASHDGGKQALETVQRLLPVLCDHGLTPEQVVA IASHDGGKQALETVQRLLPVLCAHGLTDPQVVAIAN NNGGKQALETVQRLLPVLCAHGLTDPQVVAIASHD GKQALETVQRLLPVLCDHGLTPEQVVAIASHDGRP ALESIVAQLSRPDPALAALNDHLVALACLGGRPALD AVKKGLPHAPALIKRTRRRIPERTSHRVAGS</p>	<p>300.</p>
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	<p>CAATGGAGGAAACAAGCATTGGAGACTGTCC AACGGCTCCTTCCCCTGTTGTGTCAAGCCCAC GGTTTGACGCCTGCACAAGTGGTCGCCATCGC CAACAACAACGGCGTAAGCAGGCGCTGAAA CAGTACAGCGCCTGCTGCCGTACTGTGCCAG GATCATGGACTGACCCAGACCAGGTAGTCGC AATCGCGTCACATGACGGGGAAAGCAAGCCC TGGAACCGTGCAAAGGTTGTTGCCGGTCCTT TGTCAGACCACGGCCTTACACCGGAGCAAGT CGTGGCCATTGCAAGCAATGGGGTGGCAAC AGGCTCTTGAGACGGTTCAGAGACTTCTCCA GTTCTCTGTCAAGCCCACGGCTGACTCCCGA TCAAGTTGTAGCGATTGCGTCGCATGACGGAG GGAAACAAGCATTGGAGACTGTCCAACGGCTC CTTCCCCTGTTGTGTCAAGCCCACGGTTGAC GCCTGCACAAGTGGTCGCCATCGCCAGCCATG ATGGCGGTAAGCAGGCGCTGAAACAGTACAG CGCCTGCTGCCCTGTACTGTGCCAGGATCATGG ACTGACCCAGACCAGGTAGTCGCAATCGCGT CACATGACGGGGAAAGCAAGCCCTGAAACC GTGCAAAGGTTGTGCGGTCCTTTGTCAAGA CCACGGCCTTACACCGGAGCAAGTCGTGGCCA TTGCATCCCACGCGGTGGCAACAGGCTCTT GAGACGGTTCAGAGACTTCTCCAGTTCTCTG TCAAGCCCACGGGCTGACTCCCGATCAAGTTG TAGCGATTGCCAATAACAATGGAGGGAAACAA GCATTGGAGACTGTCCAACGGCTCCTTCCCCT GTTGTGTCAAGCCCACGGTTGACGCTGCAC AAGTGGTCGCCATCGCCAGCCATGATGGCGGT AAGCAGGCGCTGAAACAGTACAGCGCCTGCT GCCTGTACTGTGCCAGGATCATGGACTGACAC CCGAACAGGTGGTCGCATTGCTTCCCACGAC GGAGACGGCCAGCCTTGGAGTCCATCGTAGC CCAATTGTCCAGGCCGATCCCCTTGGCTG CGTTAACGAATGACCATCTGGTGGCCTTGGCA TGTCTTGGTGGACACCGCGCTCGATGCAGT CAAAAAGGCTCGCCTCATGCTCCCGCATTGA</p>	
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	TCAAAGAACCAACCGGGGATCCCGAGAGA ACTTCCATCGAGTCGCGGGATCC			
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OTHER EMBODIMENTS

A number of embodiments of the invention have been described. Nevertheless, it will be understood that various modifications may be made without departing from the spirit and scope of the invention. Accordingly, other embodiments are within the scope of the following claims.

SEQUENCE LISTING IN ELECTRONIC FORM

In accordance with Section 111(1) of the Patent Rules, this description contains a sequence listing in electronic form in ASCII text format (file: 60412-4745 Seq 05-FEB-14 v1.txt).

A copy of the sequence listing in electronic form is available from the Canadian Intellectual Property Office.

CLAIMS:

1. A process for assembling nucleic acids encoding custom TALE repeat array proteins comprising:

5 (a) providing a first nucleic acid comprising a sequence encoding a first set comprising one or more transcription activator-like effector (TALE) repeat domains;

(b) contacting the first nucleic acid with a first enzyme, wherein the first enzyme creates a first ligatable end;

(c) providing a second nucleic acid comprising a sequence encoding a second set comprising one or more TALE repeat domains;

10 (d) contacting the second nucleic acid with a second enzyme, wherein the second enzyme creates a second ligatable end, and wherein the first and second ligatable ends are compatible; and

(e) ligating the first and second nucleic acids through the first and second ligatable ends to produce a first ligated nucleic acid, wherein the first ligated nucleic acid is linked to a solid support, and wherein the first ligated nucleic acid encodes a polypeptide comprising said first and second sets comprising one or more TALE repeat domains.

2. The process of claim 1, wherein the first and second enzymes are a first and second restriction endonuclease, wherein the first restriction endonuclease cleaves at a site within the first nucleic acid and creates a first cut end, and the second restriction endonuclease cleaves at a site within the second nucleic acid and creates a second cut end, and wherein the first and second ligatable ends are the first and second cut ends.

3. The process of claim 2, wherein the first ligated nucleic acid does not comprise a restriction site recognized by the first restriction endonuclease.

4. The process of any one of claims 1 to 3, further comprising:

25 (f) contacting the first ligated nucleic acid with a third enzyme, wherein the third enzyme creates a third ligatable end;

(g) providing a third nucleic acid comprising a sequence encoding a third set comprising one or more TALE repeat domains;

(h) contacting the third nucleic acid with a fourth enzyme, wherein the fourth enzyme creates a fourth ligatable end, and wherein the third and fourth ligatable ends are compatible; and

5 (i) ligating the first ligated and third nucleic acids through the third and fourth ligatable ends to produce a second ligated nucleic acid linked to the solid support, wherein the second ligated nucleic acid encodes a polypeptide comprising said first, second, and third sets comprising one or more TALE repeat domains.

10 5. The process of claim 4, wherein the third and fourth enzymes are a third and fourth restriction endonuclease, wherein the third restriction endonuclease cleaves at a site within the first ligated nucleic acid and creates a third cut end, and the fourth restriction endonuclease cleaves at a site within the third nucleic acid and creates a fourth cut end, and wherein the third and fourth ligatable ends are the third and fourth cut ends.

15 6. The process of claim 5, wherein the ligated nucleic acid does not comprise a restriction site recognized by the first endonuclease, and wherein the first and third restriction endonucleases are the same.

7. The process of claim 5 or 6, wherein the second and fourth restriction endonucleases are the same.

8. The process of any one of claims 4 to 7, further comprising:

20 (j) contacting the second ligated nucleic acid with a fifth enzyme, wherein the fifth enzyme creates a fifth ligatable end;

(k) providing a fourth nucleic acid comprising a sequence encoding a fourth set comprising one or more TALE repeat domains;

(l) contacting the fourth nucleic acid with a sixth enzyme, wherein the sixth enzyme creates a sixth ligatable end, and wherein the fifth and sixth ligatable ends are compatible; and

25 (m) ligating the second ligated and fourth nucleic acids through the fifth and sixth ligatable ends to produce a third ligated nucleic acid linked to the solid support, wherein the third ligated nucleic acid encodes a polypeptide comprising said first, second, third, and fourth sets comprising one or more TALE repeat domains.

9. The process of claim 8, wherein the fifth and sixth enzymes are a fifth and sixth restriction endonuclease, wherein the fifth restriction endonuclease cleaves at a site within the second ligated nucleic acid and creates a fifth cut end, and the sixth restriction endonuclease cleaves at a site within the fourth nucleic acid and creates a sixth cut end, and wherein the fifth and sixth ligatable ends are the fifth and sixth cut ends.
10. The process of claim 9, wherein the second ligated nucleic acid does not comprise a restriction site recognized by the first endonuclease, and wherein the first, third, and fifth restriction endonucleases are the same.
11. The process of claim 9 or 10, wherein the second, fourth, and sixth restriction endonucleases are the same.
12. The process of claim 1, wherein the second set comprises one to four TALE repeat domains.
13. The process of any one of claims 1-11, wherein the first and second ligatable ends each comprise an overhang of 1-10 nucleotides; or the first enzyme is a type IIS restriction endonuclease.
14. The process of any one of claims 1-11, further comprising unlinking the first ligated nucleic acid from the solid support and inserting the first ligated nucleic acid into a vector; unlinking the second ligated nucleic acid from the solid support and inserting the second ligated nucleic acid into a vector; unlinking the third ligated nucleic acid from the solid support and inserting the third ligated nucleic acid into a vector.
15. The process of claim 14, wherein the vector is an expression vector.
16. The process of claim 15, wherein the expression vector includes a sequence encoding an effector domain, and wherein the first, second, or third ligated nucleic acid is inserted into the vector such that the vector comprises a sequence encoding a fusion protein of the polypeptide and the effector domain.
17. The process of claim 16, wherein the effector domain is a nuclease domain.

18. The process of claim 16 or 17, further comprising inserting the expression vector into a cell.

19. The process of any one of claims 1 to 18, wherein the first nucleic acid comprises a nucleotide sequence encoding a protein encoded by a sequence selected from the group consisting of SEQ ID NOs: 58-62; and the second nucleic acid comprises a β unit, a γ unit, a δ unit, and a ϵ unit, wherein the β unit comprises a nucleotide sequence encoding a protein encoded by a sequence selected from the group consisting of SEQ ID NOs: 63-67, the γ unit comprises a nucleotide sequence encoding a protein encoded by a sequence selected from the group consisting of SEQ ID NOs: 68-72, the δ unit comprises a nucleotide sequence encoding a protein encoded by a sequence selected from the group consisting of SEQ ID NOs: 73-77, and the ϵ unit comprises a nucleotide sequence encoding a protein encoded by a sequence selected from the group consisting of SEQ ID NOs: 78-82.

20. The process of any one of claims 1 to 18, wherein the first nucleic acid comprises a nucleotide sequence selected from the group consisting of SEQ ID NOs: 58-62; and the second nucleic acid comprises a β unit, a γ unit, a δ unit, and a ϵ unit, wherein the β unit comprises a nucleotide sequence selected from the group consisting of SEQ ID NOs: 63-67, the γ unit comprises a nucleotide sequence selected from the group consisting of SEQ ID NOs: 68-72, the δ unit comprises a nucleotide sequence selected from the group consisting of SEQ ID NOs: 73-77, and the ϵ unit comprises a nucleotide sequence selected from the group consisting of SEQ ID NOs: 78-82.

21. The process of any one of claims 4 to 11 and 13 to 20, wherein the third nucleic acid comprises a β unit, a γ unit, a δ unit, and a ϵ unit, wherein the β unit comprises a nucleotide sequence encoding a protein encoded by a sequence selected from the group consisting of SEQ ID NOs: 63-67, the γ unit comprises a nucleotide sequence encoding a protein encoded by a sequence selected from the group consisting of SEQ ID NOs: 68-72, the δ unit comprises a nucleotide sequence encoding a protein encoded by a sequence selected from the group consisting of SEQ ID NOs: 73-77, and the ϵ unit comprises a nucleotide sequence encoding a protein encoded by a sequence selected from the group consisting of SEQ ID NOs: 78-82.

22. The process of any one of claims 4 to 11 and 13 to 20, wherein the third nucleic acid comprises a β unit, a γ unit, a δ unit, and a ϵ unit, wherein the β unit comprises a nucleotide sequence selected from the group consisting of SEQ ID NOs: 63-67, the γ unit comprises a nucleotide sequence selected from the group consisting of SEQ ID NOs: 68-72, the δ unit
5 comprises a nucleotide sequence selected from the group consisting of SEQ ID NOs: 73-77, and the ϵ unit comprises a nucleotide sequence selected from the group consisting of SEQ ID NOs: 78-82.

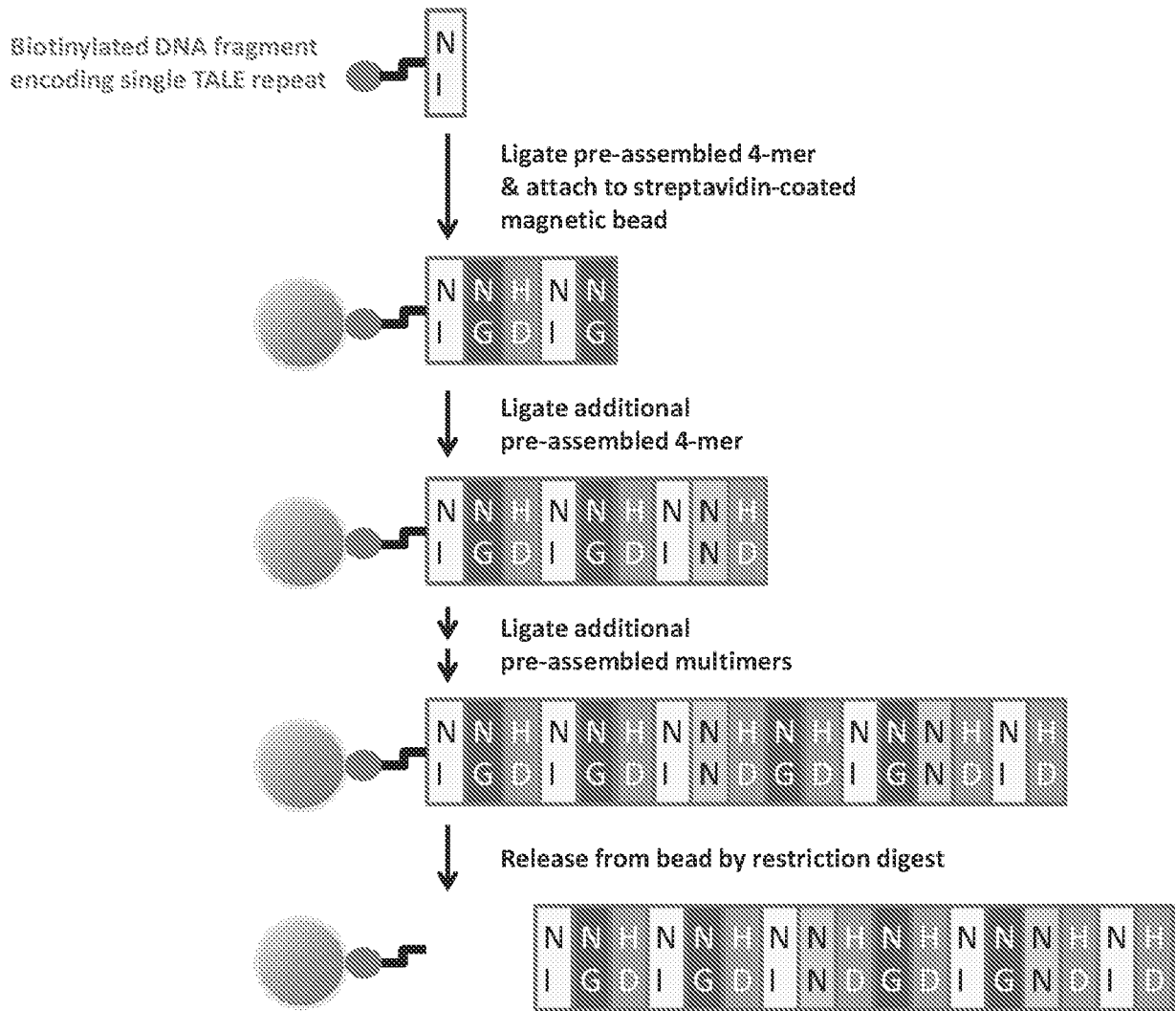


FIG. 1

Archive of 376 pre-assembled TALE repeat units

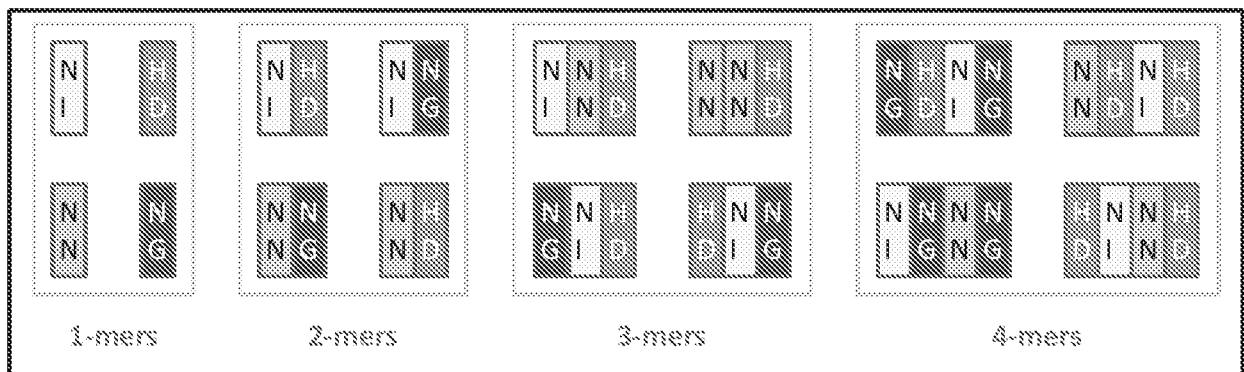


FIG. 2

TCGCGCGTTTCGGTGATGACGGTGAAAACCTCTGACACATGCAGCTCCCGGAGACGGTCCACAGCTTGT
CTGTAAGCGGATGCCGGGAGCAGACAAGCCCCTCAGGGCGCGTCAGCGGGTGTGGCGGGTGTGGGG
CTGGCTTAACATATGCGGCATCAGAGCAGATTGTACTGAGAGTGCACCATATCGGGTGTGAAATACCGC
ACAGATGCGTAAGGAGAAAAATACCGCATCAGGCGCCATTTCGCCATTTCAGGCTGCGCAACTGTTGGGAA
GGCGGATCGGTGCGGGCCTCTTCGCTATTACGCCAGCTGGCGAAAAGGGGGATGTGCTGCAAGGCGATT
AAGTTGGGTAACGCCAGGGTTTTCCAGTCACGACGTTGTA AACGACGGCCAGTGAATTCGAGCTCG
GTACCTCGGAATGCATCTAGATATCGGATCCCGGGCCCCGTCGACTGCAGAGGCTGCATGCAAGCTT
GGCGTAATCATGGTCATAGCTGTTTCCTGTGTGAAATTGTTATCCGCTCACAAATCCACACAACATAC
GAGCCGGAAGCATAAAGTGTAAGCCTGGGGTGCCTAATGAGTGAGCTAACTCACATTAATTGCGTTG
CGTCACTGCCCGCTTTCAGTCGGGAAACCTGTGCTGCCAGCTGCATTAATGAATCGGCCAACGCGC
GGGAGAGGGCGGTTTTGCGTATGGGGCGCTTTCGCTTCCCTCGCTCACTGACTCGCTGCGCTCGGTG
TTCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTATCCACAGAATCAGGGGAT
AACGCAGGAAAGAACATGTGAGCAAAAGGCCAGCAAAAGGCCAGGAACCGTAAAAAGGCCGCGTTGCT
GGCGTTTTTCCATAGGCTCCGCCCCCTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGC
GAAACCCGACAGGACTATAAAGATAACCAGGCGTTTTCCCCCTGGAAGCTCCCTCGTGCCTCTCCTGTT
CCGACCCTGCCGCTTACCGGATACCTGTCCGCTTTCTCCCTTCGGGAAGCGTGGCGCTTCTCATAG
CTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTGCTCCAAGCTGGGCTGTGTGCACGAACCC
CGTTTCAGCCCGACCGCTGCGCCTTATCCGGTAACTATCGTCTTGAGTCCAACCCGTAAGACACGAC
TTATCGCCACTGGCAGCAGCCACTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGA
GTTCTTGAAGTGGTGGCCTAACTACGGCTACACTAGAAGAACAGTATTTGGTATCTGCGCTCTGCTGA
AGCCAGTTACCTTCGGAAAAAGAGTTGGTAGCTCTTGATCCGGCAAACAAACCACCGCTGGTAGCGGT
GGTTTTTTTTGTTTGAAGCAGCAGATTACGCGCAGAAAAAAGGATCTCAAGAAGATCCTTTGATCTT
TTCTACGGGTCTGACGCTCAGTGGAACGAAAACCTCACGTTAAGGGATTTTGGTCATGAGATTATCAA
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TAAACTTGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCACCTATCTCAGCGATCTGTCTATTTG
TTCATCCATAGTTGCCTGACTCCCCGTCGTGTAGATAACTACGATACGGGAGGGCTTACCATCTGGCC
CCAGTGCTGCAATGATAACCGGAGAGCCACGCTCACCGGCTCCAGATTTATCAGCAATAAACCAGCCA
GCCGGAAGGGCCGAGCGCAGAAGTGGTCCTGCAACTTTATCCGCTCCATCCAGTCTATTAATTGTTG
CCGGGAAGCTAGAGTAAGTAGTTCCGCAAGTTAATAGTTTGCGCAACGTTGTTGCCATTGCTACAGGCA
TCGTGGTGTACGCTCGTCGTTTTGGTATGGCTTCATTCAGCTCCGGTCCCAACGATCAAGGCGAGTT
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TAAGATGCTTTTCTGTGACTGGTGAGTACTCAACCAAGTCATTCTGAGAATAGTGTATGCGGCGACCG
AGTTGCTCTTGCCCGGGTCAATAACGGGATAATAACCGGCCACATAGCAGAACTTTAAAAGTGCTCAT
CATGGAAAACGTTCTTCGGGGCGAAAACCTCTCAAGGATCTTACCGCTGTTGAGATCCAGTTCGATGT
AACCCACTCGTGCACCCAACTGATCTTCAGCATCTTTACTTTTACCAGCGTTTCTGGGTGAGCAAAA
ACAGGAAGGCAAAATGCCGCAAAAAAGGGAATAAGGGCGACACGGAAATGTTGAATACTCATACTCTT
CCTTTTTCAATATTATTGAAGCATTATCAGGGTTATTGTCTCATGAGCGGATACATATTTGAATGTA
TTTAGAAAAATAAACAAATAGGGGTTCCGCGCACATTTCCCCGAAAAGTGCCACCTGACGTCTAAGAA
ACCAATTATTATCATGACATTAACCTATAAAAAATAGGCGTATCACGAGGCCCTTTCGTC
(SEQ ID NO:1)

FIG. 3

FIG 4A

Type	SEQUENCE	SEQ ID NO:
α/ϵ	LTPDQVVAIASNIGGKQALETVQRLLPVLCQDHG	2
β	LTPEQVVAIASNIGGKQALETVQRLLPVLCQAHG	3
γ	LTPDQVVAIASNIGGKQALETVQRLLPVLCQAHG	4
δ	LTPAQVVAIASNIGGKQALETVQRLLPVLCQDHG	5

FIG 4B

Type	SEQUENCE	SEQ ID NO:
α/ϵ	CTGACCCAGACCAGGTAGTCGCAATCGCGTCGAACATTGG GGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCG GTCCTTTGTCAAGACCACGGC	6
Type β :	CTTACACCGGAGCAAGTCGTGGCCATTGCAAGCAACATCGG TGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCCAG TTCTCTGTCAAGCCCACGGG	7
Type γ :	CTGACTCCCAGATCAAGTTGTAGCGATTGCGTCGAACATTGGA GGGAAACAAGCATTGGAGACTGTCCAACGGCTCCTTCCCCT GTTGTGTCAAGCCCACGGT	8
Type δ :	TTGACGCCTGCACAAGTGGTCGCCATCGCCTCCAATATTGGCG GTAAGCAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACT GTGCCAGGATCATGGA	9

GACGGATCGGGAGATCTCCCGATCCCCTATGGTCGACTCTCAGTACAATCTGCTCTGATGCCGCATAGTTAAGCCAGTA
TCTGCTCCCTGCTTGTGTGTTGGAGGTCGCTGAGTAGTGCGCGAGCAAAATTTAAGCTACAACAAGGCAAGGCTTGAC
CGACAATTGCATGAAGAATCTGCTTAGGGTTAGGCGTTTTGCGCTGCTTCGCGATGTACGGGCCAGATATACGCGTTG
ACATTGATTATTGACTAGTTATTAATAGTAATCAATTACGGGGTCATTAGTTCATAGCCCATATATGGAGTTCGCGTTA
CATAACTTACGGTAAATGGCCCCGCTGGCTGACCCGCCAACGACCCCCGCCATTGACGTCAATAATGACGTATGTTCC
CATAGTAACGCCAATAGGGACTTTCCATTGACGTCAATGGGTGGACTATTTACGGTAAACTGCCACTTGGCAGTACAT
CAAGTGTATCATATGCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAATGGCCCCGCTGGCATTATGCCAGTACA
TGACCTTATGGGACTTTCTACTTGGCAGTACATCTACGTATTAGTCATCGCTATTACCATGGTGATGCGGTTTTGGCAG
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CGGAAATTTAATAACGGCGAGATAAACTTTTAAGGGCCCTTCGAAGGTAAGCCTATCCCTAACCTCTCCTCGGTCTCG
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GGGCTTAGGGGGTATCCCCACGCGCCCTGTAGCGGCGCATTAAAGCGCGGGCGGGTGTGGTGGTTACGCGCAGCGTG
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GTGGAAGTCCCCAGGCTCCCCAGCAGGCAGAAGTATGCAAAGCATGCATCTCAATTAGTC

(SEQ ID NO:10)

FIG. 5A

FIG. 5B

AGCAACCATAGTCCCGCCCCTAACTCCGCCCATCCCGCCCCTAACTCCGCCCAGTTCGCCCATTCTCCGCCCATGGC
TGACTAATTTTTTTTATTTATGCAGAGGCCGAGGCCGCCTCTGCCTCTGAGCTATCCAGAAGTAGTGAGGAGGCTTT
TTTGGAGGCCTAGGCTTTTGCAAAAAGCTCCCGGGAGCTGTATATCCATTTTCGGATCTGATCAGCACGTGTTGACA
ATTAATCATCGGCATAGTATATCGGCATAGTATAATACGACAAGGTGAGGAACTAAACCATGGCCAAGCCTTTGTCTC
AAGAAGAATCCACCCTCATTGAAAGAGCAACGGCTACAATCAACAGCATCCCCATCTCTGAAGACTACAGCGTCGCC
AGCGCAGCTCTCTAGCGACGGCCGCATCTTCACTGGTGTCAATGTATATCATTTTACTGGGGGACCTTGTGCAGAA
CTCGTGGTGTCTGGGCACTGCTGCTGCTGCGGCAGCTGGCAACCTGACTTGTATCGTCGCGATCGGAAATGAGAACA
GGGGCATCTTGAGCCCCTGCGGACGGTGTGACAGGTGCTTCTCGATCTGCATCCTGGGATCAAAGCGATAGTGAA
GGACAGTGATGGACAGCCGACGGCAGTTGGGATTCTGTAATTGCTGCCCTCTGGTTATGTGTGGGAGGGCTAAGCA
CTTCGTGGCCGAGGAGCAGGACTGACACGTGCTACGAGATTTCCGATTCCACCGCCGCCTTCTATGAAAGGTTGGGCT
TCGGAATCGTTTTCCGGGACGCCGGCTGGATGATCCTCCAGCGCGGGGATCTCATGCTGGAGTCTTCGCCACCCC
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TGCATTCTAGTTGTGGTTTTGTCCAAACTCATCAATGTATCTTATCATGTCTGTATAACCGTCGACCTCTAGCTAGAGCT
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GTCGGGAAACCTGTCGTGCCAGCTGCATTAATGAATCGGCCAACGCGCGGGGAGAGGCGGTTTGCATTTGGGCGC
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AACCGTAAAAAGGCCGCTTGTGGCGTTTTTCCATAGGCTCCGCCCCCTGACGAGCATCACAAAATCGACGCTC
AAGTCAGAGGTGGCGAAACCCGACAGGACTATAAAGATACCAGGCGTTTTCCCCTGGAAGCTCCCTCGTGCCTCTC
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GCAACAACACCCTGCTAGCGGTGGTTTTTTTGTGCAAGCAGCAGATTACGCGCAGAAAAAAGGATCTCAA
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CGGCGTCAATACGGGATAATACCGCGCCACATAGCAGAACTTTAAAAGTGCTCATCATTGGAAAACGTTCTTCGGGG
CGAAAACCTCTCAAGGATCTTACCGCTGTTGAGATCCAGTTCGATGTAACCCACTCGTGCACCCAACTGATCTTCAGCA
TCTTTTACTTTACCAGCGTTTCTGGGTGAGCAAAAACAGGAAGGCAAAATGCCGCAAAAAGGGGAATAAGGGCGA
CACGGAAATGTTGAATACTCATACTCTTCTTTTCAATATTATTGAAGCATTATCAGGGTTATTGTCTCATGAGCGG
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C (SEQ ID NO:11)

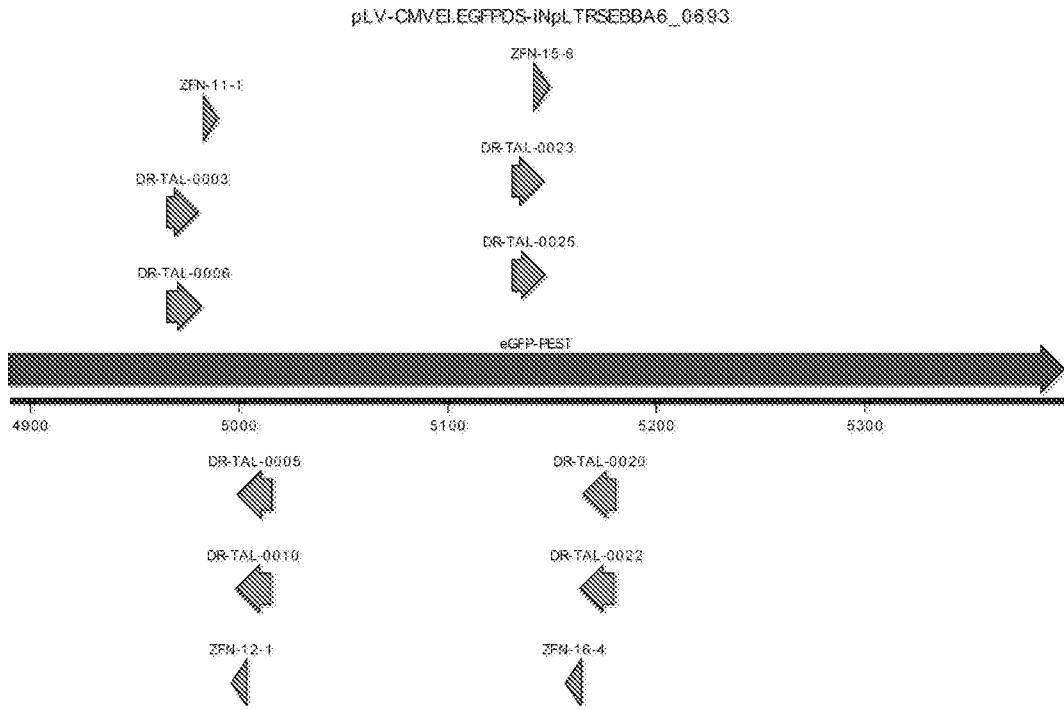


FIG. 6

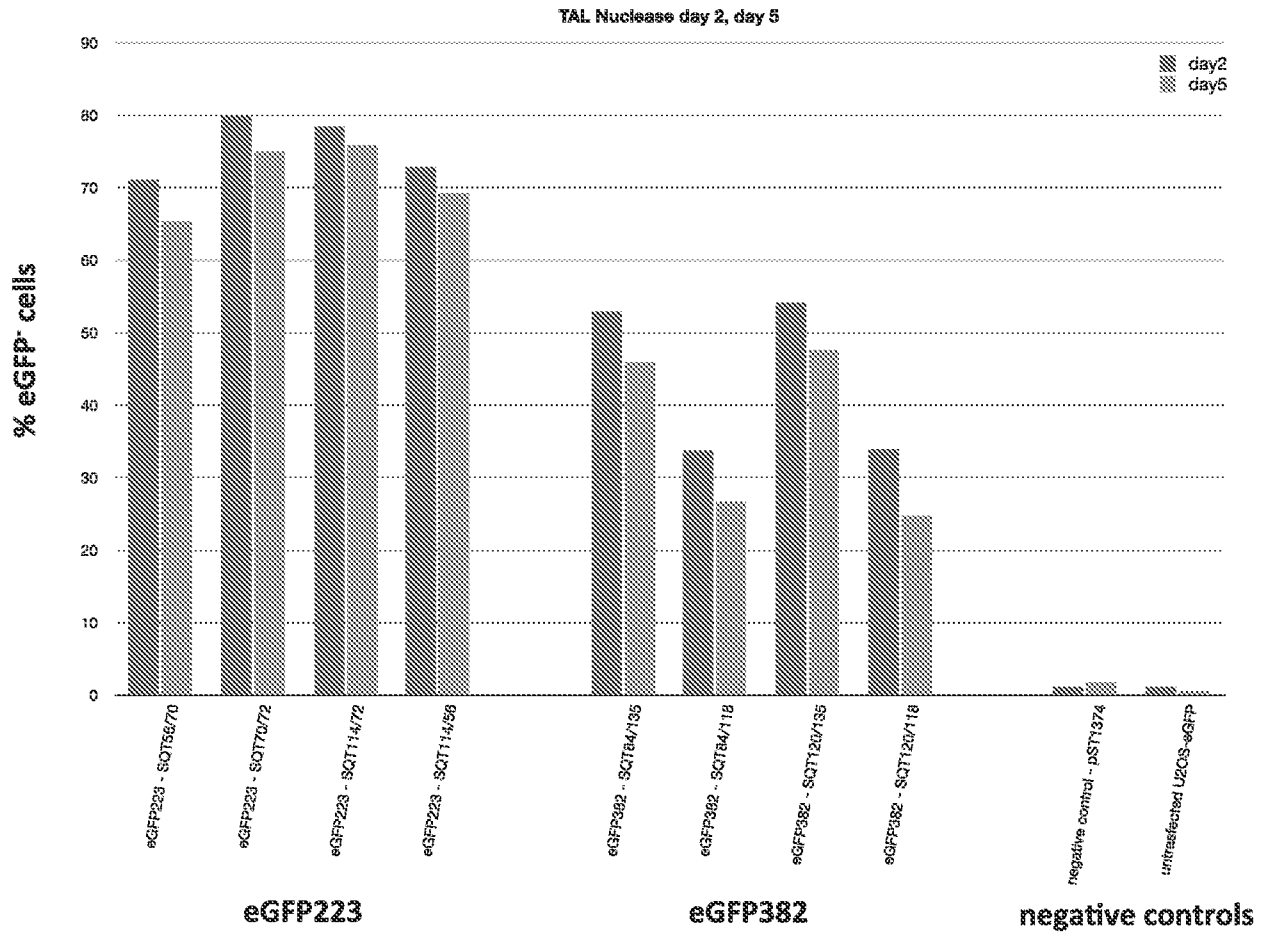


FIG. 7

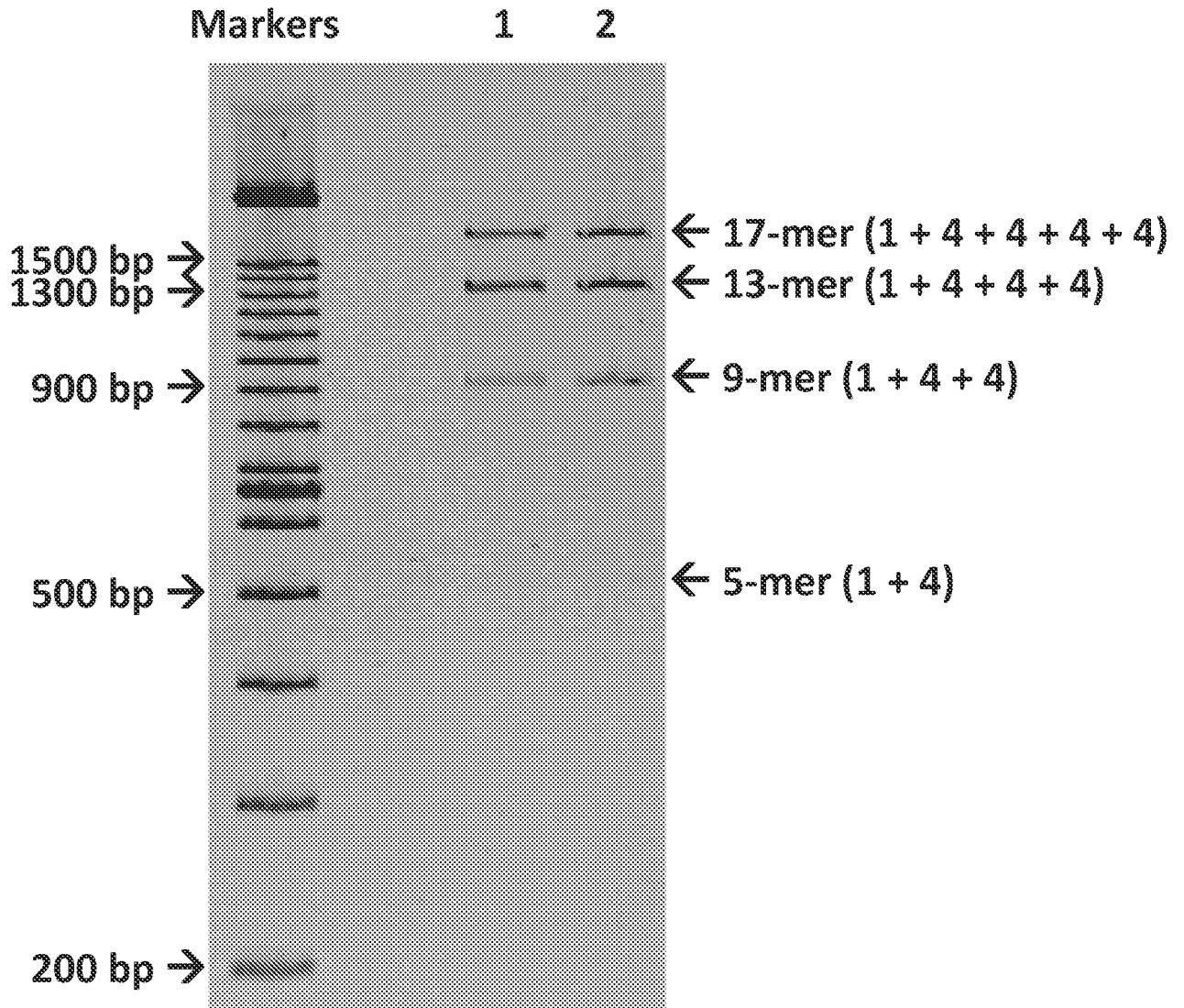
14 of 30 sequences mutated. (46%)

Sequence	Δ SIZE	SEQ ID NO:
CTACGGCGTGCAGTGCCTTACGCCGCTACCCCGACCACTGAAAGCAGCAGCAGTTCAGTCCGCCCATGCCCGAAGGCTACGTCAGGAGCGCACCACTTCTTCAAGGA	WT	12
CTACGGCGTGCAGTGCCTTACGCCGCTACCCCGACCACTGAAAGCAGCAGCAGTTCAGTCCGCCCATGCCCGAAGGCTACGTCAGGAGCGCACCACTTCTTCAAGGA	Δ3 (3X)	13
CTACGGCGTGCAGTGCCTTACGCCGCTACCCCGACCACTGAAAGCAGCAGCAGTTCAGTCCGCCCATGCCCGAAGGCTACGTCAGGAGCGCACCACTTCTTCAAGGA	Δ16	14
CTACGGCGTGCAGTGCCTTACGCCGCTACCCCGACCACTGAAAGCAGCAGCAGTTCAGTCCGCCCATGCCCGAAGGCTACGTCAGGAGCGCACCACTTCTTCAAGGA	Δ21	15
CTACGGCGTGCAGTGCCTTACGCCGCTACCCCGACCACTGAAAGCAGCAGCAGTTCAGTCCGCCCATGCCCGAAGGCTACGTCAGGAGCGCACCACTTCTTCAAGGA	Δ22	16
CTACGGCGTGCAGTGCCTTACGCCGCTACCCCGACCACTGAAAGCAGCAGCAGTTCAGTCCGCCCATGCCCGAAGGCTACGTCAGGAGCGCACCACTTCTTCAAGGA	Δ30 (Δ31 and +1)	17
CTACGGCGTGCAGTGCCTTACGCCGCTACCCCGACCACTGAAAGCAGCAGCAGTTCAGTCCGCCCATGCCCGAAGGCTACGTCAGGAGCGCACCACTTCTTCAAGGA	Δ33	18
CTACGGCGTGCAGTGCCTTACGCCGCTACCCCGACCACTGAAAGCAGCAGCAGTTCAGTCCGCCCATGCCCGAAGGCTACGTCAGGAGCGCACCACTTCTTCAAGGA	Δ33	19
CTACGGCGTGCAGTGCCTTACGCCGCTACCCCGACCACTGAAAGCAGCAGCAGTTCAGTCCGCCCATGCCCGAAGGCTACGTCAGGAGCGCACCACTTCTTCAAGGA	Δ51	20
CTACGGCGTGCAGTGCCTTACGCCGCTACCCCGACCACTGAAAGCAGCAGCAGTTCAGTCCGCCCATGCCCGAAGGCTACGTCAGGAGCGCACCACTTCTTCAAGGA	Δ54	21
CTACGGCGTGCAGTGCCTTACGCCGCTACCCCGACCACTGAAAGCAGCAGCAGTTCAGTCCGCCCATGCCCGAAGGCTACGTCAGGAGCGCACCACTTCTTCAAGGA	Δ60	22
CTACGGCGTGCAGTGCCTTACGCCGCTACCCCGACCACTGAAAGCAGCAGCAGTTCAGTCCGCCCATGCCCGAAGGCTACGTCAGGAGCGCACCACTTCTTCAAGGA	Δ84	23
CTACGGCGTGCAGTGCCTTACGCCGCTACCCCGACCACTGAAAGCAGCAGCAGTTCAGTCCGCCCATGCCCGAAGGCTACGTCAGGAGCGCACCACTTCTTCAAGGA	+2	24

FIG. 8

FIG. 9

Assembled
TALE repeat arrays



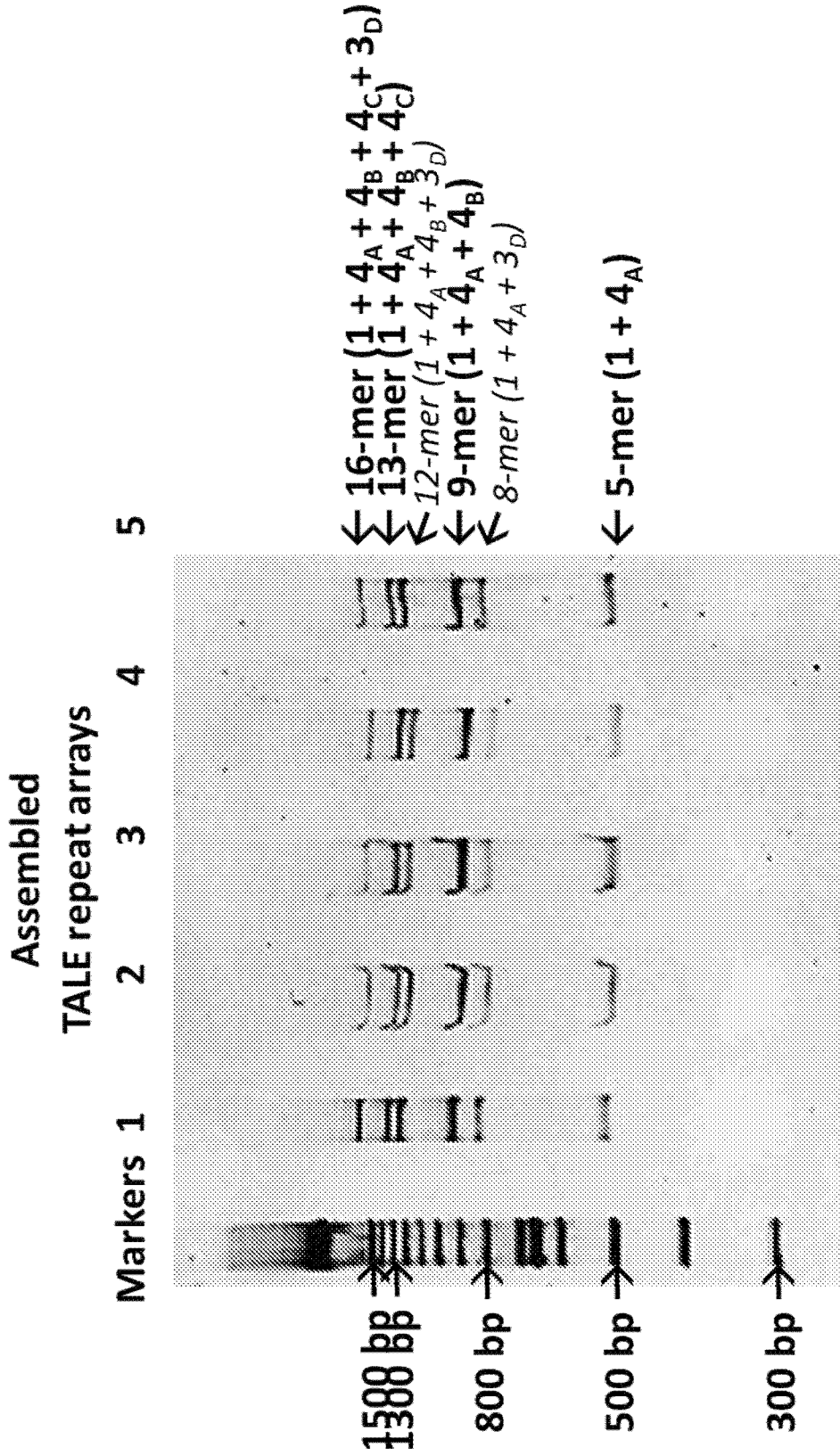


FIG. 10

FIG. 11A

TCTAGAGCTAGCACCATGGACTACAAAGACCATGACGGTGATTATAAAGATCATGACATCGATTACAAGGATGAC
 GATGACAAGATGGCCCCAAGAAGAAGAGGAAGGTGGGCATTACCCGCGGGGTACCTATGGTGGACTTGAGGAC
 ACTCGGTTATTCGCAACAGCAACAGGAGAAAATCAAGCCTAAGGTGAGGAGCACCGTCGCGCAACACCACGAGG
 CGCTTGTGGGGCATGGCTTCACTCATGCGCATATTGTCGCGCTTTACAGCACCTGCGGCGCTTGGGACGGTGGC
 TGTCAAATACCAAGATATGATTGCGGCCCTGCCGAAGCCACGCACGAGGCAATTGTAGGGGTGCGTAAACAGTG
 GTCGGGAGCGCGAGCACTTGAGGCGCTGCTGACTGTGGCGGGTGAGCTTAGGGGGCCTCCGCTCCAGCTCGACA
 CCGGGCAGCTGCTGAAGATCGCGAAGAGAGGGGGAGTAACAGCGGTAGAGGCAGTGCACGCCTGGCGCAATGC
 GCTACCCGGGGCCCCCTTGAACCTGACCCAGACCAGGTAGTCGCAATCGCGAACAATAATGGGGGAAAGCAAG
 CCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCTTTGTCAAGACCACGGCCTTACACCGGAGCAAGTCGTGGCCA
 TTGCATCCACGACGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCAGTTCTCTGTCAAGCCCACGG
 GCTGACTCCCGATCAAGTTGTAGCGATTGCGTGAACATTGGAGGGAAACAAGCATTGGAGACTGTCCAACGGCT
 CCTTCCCGTGTGTGTCAAGCCCACGGTTTGACGCCTGCACAAGTGGTGCCTATCGCCAACAACAACGGCGGTAAG
 CAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGACTGACCCAGACCAGGTAGTC
 GCAATCGCGTCAAACGGAGGGGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCTTTGTCAAGA
 CCACGGCCTTACACCGGAGCAAGTCGTGGCCATTGCAAATAATAACGGTGGCAAACAGGCTCTTGAGACGGTTCA
 GAGACTTCTCCAGTTCTCTGTCAAGCCCACGGGCTGACTCCCGATCAAGTTGTAGCGATTGCGTCGCATGACGGA
 GGGAAACAAGCATTGGAGACTGTCCAACGGCTCCTTCCCGTGTGTGTCAAGCCCACGGTTTGACGCCTGCACAA
 GTGGTGCCTATCGCCTCGAATGGCGGCGGTAAGCAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGC
 CAGGATCATGGACTGACCCAGACCAGGTAGTCGCAATCGCGTCAAACGGAGGGGGAAAGCAAGCCCTGGAAAC
 CGTGCAAAGGTTGTTGCCGGTCTTTGTCAAGACCACGGCCTTACACCGGAGCAAGTCGTGGCCATTGCATCCAC
 GACGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCAGTTCTCTGTCAAGCCCACGGGCTGACTCCCG
 ATCAAGTTGTAGCGATTGCGTGAACATTGGAGGGAAACAAGCATTGGAGACTGTCCAACGGCTCCTTCCCGTGT
 TGTGTCAAGCCCACGGTTTGACGCCTGCACAAGTGGTGCCTATCGCCAACAACAACGGCGGTAAGCAGGCGCTGG
 AAACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGACTGACCCAGACCAGGTAGTCGCAATCGCGT
 CACATGACGGGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCTTTGTCAAGACCACGGCCTTA
 CACCGGAGCAAGTCGTGGCCATTGCATCCACGACGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCC
 CAGTTCTCTGTCAAGCCCACGGGCTGACTCCCGATCAAGTTGTAGCGATTGCGAATAACAATGGAGGGAAACAAG
 CATTGGAGACTGTCCAACGGCTCCTTCCCGTGTGTGTCAAGCCCACGGTCTGACACCCGAACAGGTGGTGCCTAT
 TGCTTCCACGACGGAGGACGGCCAGCCTTGGAGTCCATCGTAGCCCAATTGTCCAGGCCCGATCCCGCCTTGGCT
 GCGTTAACGAATGACCATCTGGTGGCGTTGGCATGTCTTGGTGGACGCCCGCTCGATGCAGTCAAAAAGGGT
 CTGCCTCATGCTCCCGCATTGATCAAAAAGAACCAACCGGCGGATTCCCGAGAGAACTTCCCATCGAGTCGCGGGAT
 CC

(SEQ ID NO:25)

FIG. 11B

MDYKDHDGDYKDHIDYKDDDDKMAPKKRKVGIHRGVPMVDLRTLGYSSQQQEKIKPKVRSTVAQHHEALVGHG
 FTHAHIVALSQHPAALGTVAVKYQDMIAALPEATHEAIVGVGKQWSGARALEALLTVAGELRGPPLQLDGTQLLKIAR
 GGVTAVEAVHAWRNALTGAPLNLTDPDQVVAIANNNGGKQALETVQRLLPVLCQDHGLTPEQVVAIASHDGGKQALE
 TVQRLLPVLCQAHGLTPDQVVAIASNIGGKQALETVQRLLPVLCQAHGLTPAQVVAIANNNGGKQALETVQRLLPVLC
 QDHGLTPDQVVAIASNIGGKQALETVQRLLPVLCQDHGLTPEQVVAIANNNGGKQALETVQRLLPVLCQAHGLTPDQ
 VVAIASHDGGKQALETVQRLLPVLCQAHGLTPAQVVAIASNIGGKQALETVQRLLPVLCQDHGLTPDQVVAIASNGG
 GKQALETVQRLLPVLCQDHGLTPEQVVAIASHDGGKQALETVQRLLPVLCQAHGLTPDQVVAIASNIGGKQALETVQR
 LLPVLCQAHGLTPAQVVAIANNNGGKQALETVQRLLPVLCQDHGLTPDQVVAIASHDGGKQALETVQRLLPVLCQDH
 GLTPEQVVAIASHDGGKQALETVQRLLPVLCQAHGLTPDQVVAIANNNGGKQALETVQRLLPVLCQAHGLTPEQVVAI
 ASHDGGRPALESIVAQLSRPDPALAALNDHLVALACLGGRPALDAVKKGLPHAPALIKRTNRRIPERTSHRVAGS

(SEQ ID NO:26)

FIG. 12A

TCTAGAGCTAGCACCATGGACTACAAAGACCATGACGGTGATTATAAAGATCATGACATCGATTACAAGGATGAC
GATGACAAGATGGCCCCAAGAAGAAGAGGAAGGTGGGCATTACCCGCGGGGTACCTATGGTGGACTTGAGGAC
ACTCGGTTATTCGCAACAGCAACAGGAGAAAATCAAGCCTAAGGTGAGGAGCACCGTCGCGCAACACCACGAGG
CGCTTGTGGGGCATGGCTTCACTCATGCGCATATTGTCGCGCTTTACAGCACCTGCGGCGCTTGGGACGGTGGC
TGTCAAATACCAAGATATGATTGCGGCCCTGCCGAAGCCACGCACGAGGCAATTGTAGGGGTGCGTAAACAGTG
GTCGGGAGCGCGAGCACTTGAGGCGCTGCTGACTGTGGCGGGTGAGCTTAGGGGGCCTCCGCTCCAGCTCGACA
CCGGGCAGCTGCTGAAGATCGCGAAGAGAGGGGGAGTAACAGCGGTAGAGGCAGTGACAGCCTGGCGCAATGC
GCTCACCGGGGCCCTTGAACCTGACCCAGACCAGGTAGTCGCAATCGCGAACAATAATGGGGGAAAGCAAG
CCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCCCTTTGTCAAGACCACGGCCTTACACCGGAGCAAGTCGTGGCCA
TTGCATCCACGACGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCAGTTCTCTGTCAAGCCCACGG
GCTGACTCCCGATCAAGTTGTAGCGATTGCGTCGAACATTGGAGGGAAACAAGCATTGGAGACTGTCCAACGGCT
CCTTCCCGTGTGTGTCAAGCCCACGGTTTGACGCCTGCACAAGTGGTCGCCATCGCCAACAACAACGGCGGTAAG
CAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGACTGACCCAGACCAGGTAGTC
GCAATCGCGTCAAACGGAGGGGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCCCTTTGTCAAGA
CCACGGCCTTACACCGGAGCAAGTCGTGGCCATTGCAAATAATAACGGTGGCAAACAGGCTCTTGAGACGGTTCA
GAGACTTCTCCAGTTCTCTGTCAAGCCCACGGGCTGACTCCCGATCAAGTTGTAGCGATTGCGTCGCATGACGGA
GGGAAACAAGCATTGGAGACTGTCCAACGGCTCCTTCCCGTGTGTGTCAAGCCCACGGTTTGACGCCTGCACAA
GTGGTCGCCATCGCCTCGAATGGCGGCGTAAGCAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGC
CAGGATCATGGACTGACCCAGACCAGGTAGTCGCAATCGCGTCAAACGGAGGGGGAAAGCAAGCCCTGGAAAC
CGTGCAAAGGTTGTTGCCGGTCCCTTTGTCAAGACCACGGCCTTACACCGGAGCAAGTCGTGGCCATTGCATCCCAC
GACGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCAGTTCTCTGTCAAGCCCACGGGCTGACTCCCG
ATCAAGTTGTAGCGATTGCGTCGAACATTGGAGGGAAACAAGCATTGGAGACTGTCCAACGGCTCCTTCCCGTGT
TGTGTCAAGCCCACGTTTGACGCCTGCACAAGTGGTCGCCATCGCCAACAACAACGGCGGTAAGCAGGCGCTGG
AAACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGACTGACCCAGACCAGGTAGTCGCAATCGCGT
CACATGACGGGGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCCCTTTGTCAAGACCACGGCCTTA
CACCGGAGCAAGTCGTGGCCATTGCATCCCACGACGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCC
CAGTTCTCTGTCAAGCCCACGGGCTGACTCCCGATCAAGTTGTAGCGATTGCGAATAACAATGGAGGGAAACAAG
CATTGGAGACTGTCCAACGGCTCCTTCCCGTGTGTGTCAAGCCCACGGTTTGACGCCTGCACAAGTGGTCGCCAT
CGCCAGCCATGATGGCGGTAAGCAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATG
GACTGACACCCGAACAGGTGGTCGCCATTGCTTCTAATGGGGGAGGACGGCCAGCCTTGAGTCCATCGTAGCCC
AATTGTCCAGGCCCGATCCCGCGTTGGCTGCGTTAACGAATGACCATCTGGTGGCGTTGGCATGTCTTGGTGGAC
GACCCGCGCTCGATGCAGTCAAAAAGGGTCTGCCTCATGCTCCCGCATTGATCAAAAAGAACCAACCGGCGGATTC
CCGAGAGAATTCCCATCGAGTCGCGGGATCC

(SEQ ID NO:27)

FIG. 12B

MDYKDHDGDYKDHDIDYKDDDDKMAPKKKRKVGIHRGVPMVDLRTLGYSSQQQEKIKPKVRSVAQHHEALVGHG
FTHAHIVALSQHPAALGTVAVKYQDMIAALPEATHEAIVGVGKQWSGARALEALLTVAGELRGPPLQLDTGQLLKIAGR
GGVTAVEAVHAWRNALTGAPLNLTDPQVVAIANNNGGKQALETVQRLLPVLCQDHGLTPEQVVAIASHDGGKQALE
TVQRLLPVLCQAHGLTPDQVVAIASNIGGKQALETVQRLLPVLCQAHGLTPAQVVAIANNNGGKQALETVQRLLPVLC
QDHGLTPDQVVAIASNIGGKQALETVQRLLPVLCQDHGLTPEQVVAIANNNGGKQALETVQRLLPVLCQAHGLTPDQ
VVAIASHDGGKQALETVQRLLPVLCQAHGLTPAQVVAIASNIGGKQALETVQRLLPVLCQDHGLTPDQVVAIASNGG
GKQALETVQRLLPVLCQDHGLTPEQVVAIASHDGGKQALETVQRLLPVLCQAHGLTPDQVVAIASNIGGKQALETVQR
LLPVLCQAHGLTPAQVVAIANNNGGKQALETVQRLLPVLCQDHGLTPDQVVAIASHDGGKQALETVQRLLPVLCQDH
GLTPEQVVAIASHDGGKQALETVQRLLPVLCQAHGLTPDQVVAIANNNGGKQALETVQRLLPVLCQAHGLTPAQVVAI
ASHDGGKQALETVQRLLPVLCQDHGLTPEQVVAIASNIGGGRPALESIVAQLSRPDPALAALTNDHLVALACLGGRPAL
DAVKKGLPHAPALIKRTNRRIPERTSHRVAGS

(SEQ ID NO:28)

FIG. 13A

TCTAGAGCTAGCACCATGGACTACAAAGACCATGACGGTGATTATAAAGATCATGACATCGATTACAAGGATGAC
 GATGACAAGATGGCCCCAAGAAGAAGAGGAAGGTGGGCATTACCCGCGGGGTACCTATGGTGGACTTGAGGAC
 ACTCGGTTATTCGCAACAGCAACAGGAGAAAATCAAGCCTAAGGTCAGGAGCACCGTCGCGCAACACCACGAGG
 CGTTGTGGGGCATGGCTTCACTCATGCGCATATTGTCGCGCTTTCACAGCACCCCTGCGGCGCTTGGGACGGTGGC
 TGTCAAAATACCAAGATATGATTGCGGCCCTGCCGAAGCCACGCACGAGGCAATTGTAGGGGTCGGTAAACAGTG
 GTCGGGAGCGCGAGCACTTGAGGCGCTGCTGACTGTGGCGGGTGAGCTTAGGGGGCCTCCGCTCCAGCTCGACA
 CCGGGCAGCTGCTGAAGATCGCGAAGAGAGGGGGAGTAACAGCGGTAGAGGCACTGCACGCCTGGCGCAATGC
 GCTCACCGGGGCCCCCTTGAACCTGACCCCAGACCAGGTAGTCGCAATCGCGTCAAACGGAGGGGGAAAGCAAG
 CCCTGAAACCGTGCAAAGGTTGTTGCCGTCCTTTGTCAAGACCACGGCCTTACACCGGAGCAAGTCGTGGCCA
 TTGCAAATAATAACGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCCAGTTCTCTGTCAAGCCCACGG
 GCTGACTCCCGATCAAGTTGTAGCGATTGCGTCGAACATTGGAGGGAAACAAGCATTGGAGACTGTCCAACGGCT
 CCTCCCCTGTTGTGTCAAGCCCACGGTTTGACGCCTGCACAAGTGGTCGCCATCGCCTCCAATATTGGCGGTAAG
 CAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGACTGACCCCAGACCAGGTAGTC
 GCAATCGCGAACAATAATGGGGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCTTTGTCAAGAC
 CACGGCCTTACACCGGAGCAAGTCGTGGCCATTGCAAGCAACATCGGTGGCAAACAGGCTCTTGAGACGGTTCAG
 AGACTTCTCCCAGTTCTCTGTCAAGCCCACGGGCTGACTCCCAGTCAAGTTGTAGCGATTGCGTGAACATTGGAG
 GAAACAAGCATTGGAGACTGTCCAACGGCTCCTCCCCTGTTGTGTCAAGCCCACGGTGTGACGCCTGCACAAGT
 GGTGCCATCGCCAACAACAACGGCGGTAAGCAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCA
 GGATCATGGACTGACCCCAGACCAGGTAGTCGCAATCGCGTCAAACGGAGGGGGAAAGCAAGCCCTGGAAACCG
 TGCAAAGGTTGTTGCCGGTCTTTGTCAAGACCACGGCCTTACACCGGAGCAAGTCGTGGCCATTGCATCCCACGA
 CGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCCAGTTCTCTGTCAAGCCCACGGGCTGACTCCCAGT
 CAAGTTGTAGCGATTGCGAATAACAATGGAGGGAAACAAGCATTGGAGACTGTCCAACGGCTCCTCCCCTGTTG
 TGTCAAGCCCACGGTTTGACGCCTGCACAAGTGGTCGCCATCGCCTCGAATGGCGGCGGTAAGCAGGCGCTGGAA
 ACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGACTGACCCCAGACCAGGTAGTCGCAATCGCGAAC
 AATAATGGGGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCTTTGTCAAGACCACGGCCTTACA
 CCGGAGCAAGTCGTGGCCATTGCATCCCACGACGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCA
 GTTCTCTGTCAAGCCCACGGGCTGACTCCCAGTCAAGTTGTAGCGATTGCGTCAAACGGTGGAGGGAAACAAGCA
 TTGGAGACTGTCCAACGGCTCCTCCCCTGTTGTGTCAAGCCCACGGTGTGACGCCTGCACAAGTGGTCGCCATCG
 CCAACAACAACGGCGGTAAGCAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGAC
 TGACACCCGAACAGGTGGTCGCCATTGCTTCCCACGACGGAGGACGGCCAGCCTTGGAGTCCATCGTAGCCCAAT
 TGTCCAGGCCCCGATCCCGCGTTGGCTGCGTTAACGAATGACCATCTGGTGGCGTTGGCATGCTTGGTGGACGAC
 CCGCGCTCGATGCAGTCAAAAAGGGTCTGCCTCATGCTCCCGCATTGATCAAAAAGAACCAACCGGCGGATTCCCG
 AGAGAACTTCCATCGAGTCGCGGGATCC
 (SEQ ID NO:29)

FIG. 13B

MDYKDHDGDYKDHDIDYKDDDDKMAPKKRKRKVGHRGVPVMDLRTLGY5QQQEQEKIKPKVRSTVAQHHEALVGHG
 FTHAHIVALSQHPAALGTVAVKYQDMIAALPEATHEAIVGVGKQWSGARALEALLTVAGELRGPPLQLDGTGQLLKIAR
 GGVTAVEAVHAWRNALTGAPLNLTPDQVVAIASNNGGKQALETVQRLLPVLCQDHGLTPEQVVAIANNNGGKQALE
 TVQRLLPVLCQAHGLTPDQVVAIASNIGGKQALETVQRLLPVLCQAHGLTPAQVVAIASNIGGKQALETVQRLLPVLCQ
 DHGLTPDQVVAIANNNGGKQALETVQRLLPVLCQDHGLTPEQVVAIASNIGGKQALETVQRLLPVLCQAHGLTPDQV
 VAIASNIGGKQALETVQRLLPVLCQAHGLTPAQVVAIANNNGGKQALETVQRLLPVLCQDHGLTPDQVVAIASNNGGK
 QALETVQRLLPVLCQDHGLTPEQVVAIASHDGGKQALETVQRLLPVLCQAHGLTPDQVVAIANNNGGKQALETVQRLL
 PVLCQAHGLTPAQVVAIASNNGGKQALETVQRLLPVLCQDHGLTPDQVVAIANNNGGKQALETVQRLLPVLCQDHGL
 TPEQVVAIASHDGGKQALETVQRLLPVLCQAHGLTPDQVVAIASNNGGKQALETVQRLLPVLCQAHGLTPAQVVAIAN
 NNGGKQALETVQRLLPVLCQDHGLTPEQVVAIASHDGGRPALESIVAQLSRPDPALAALNDHVLVALACLGGRPALDA
 VKKGLPHAPALIKRTNRRIPERTSHRVAGS
 (SEQ ID NO:30)

FIG. 14A

TCTAGAGCTAGCACCATGGACTACAAAGACCATGACGGTGATTATAAAGATCATGACATCGATTACAAGGATGAC
GATGACAAGATGGCCCCAAGAAGAAGAGGAAGGTGGGCATTACCCGCGGGGTACCTATGGTGGACTTGAGGAC
ACTCGGTTATTCGCAACAGCAACAGGAGAAAAATCAAGCCTAAGGTGAGGAGCACCGTCGCGCAACACCACGAGG
CGTTGTGGGGCATGGCTTCACTCATGCGCATATTGTCGCGCTTTACAGCACCTGCGGCGCTTGGGACGGTGGC
TGTCAAATACCAAGATATGATTGCGGCCCTGCCGAAGCCACGCACGAGGCAATTGTAGGGGTGCGTAAACAGTG
GTCGGGAGCGCGAGCACTTGAGGCGCTGCTGACTGTGGCGGGTGAGCTTAGGGGGCCTCCGCTCCAGCTCGACA
CCGGGCAGCTGCTGAAGATCGCGAAGAGAGGGGGAGTAACAGCGGTAGAGGCAGTGCACGCCTGGCGCAATGC
GCTACCGGGGCCCCCTTGAACCTGACCCAGACCAGGTAGTCGCAATCGCGAACAATAATGGGGGAAAGCAAG
CCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCTTTGTCAAGACCACGGCCTTACACCGGAGCAAGTCGTGGCCA
TTGCAAGCAACATCGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCCAGTTCTCTGTCAAGCCCACGG
GCTGACTCCCGATCAAGTTGTAGCGATTGCGTCGAACATTGGAGGGAAACAAGCATTGGAGACTGTCCAACGGCT
CCTTCCCGTGTGTGTCAAGCCCACGGTTTGACGCCTGCACAAGTGGTCGCCATCGCCAACAACAACGGCGGTAAG
CAGGCGCTGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGACTGACCCAGACCAGGTAGTC
GCAATCGCGTCGAACATTGGGGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCTTTGTCAAGAC
CACGGCCTTACACCGGAGCAAGTCGTGGCCATTGCAAGCAACATCGGTGGCAAACAGGCTCTTGAGACGGTTCAG
AGACTTCTCCCAGTTCTCTGTCAAGCCCACGGGCTGACTCCCGATCAAGTTGTAGCGATTGCGAATAACAATGGAG
GGAAACAAGCATTGGAGACTGTCCAACGGCTCCTTCCCGTGTGTGTCAAGCCCACGGTTTGACGCCTGCACAAGT
GGTCGCCATCGCCTCGAATGGCGGCGGTAAGCAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCA
GGATCATGGACTGACCCAGACCAGGTAGTCGCAATCGCGTCACATGACGGGGGAAAGCAAGCCCTGGAAACCG
TGCAAAGGTTGTTGCCGGTCTTTGTCAAGACCACGGCCTTACACCGGAGCAAGTCGTGGCCATTGCAAATAATAA
CGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCCAGTTCTCTGTCAAGCCCACGGGCTGACTCCCGAT
CAAGTTGTAGCGATTGCGTCCAACGGTGGAGGGAAACAAGCATTGGAGACTGTCCAACGGCTCCTTCCCGTGTG
TGTCAGCCCACGGTTTGACGCCTGCACAAGTGGTCGCCATCGCCAACAACAACGGCGGTAAGCAGGCGCTGGAA
ACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGACTGACCCAGACCAGGTAGTCGCAATCGCGTCA
CATGACGGGGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCTTTGTCAAGACCACGGCCTTACA
CCGGAGCAAGTCGTGGCCATTGCAAGCAATGGGGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCCA
GTTCTCTGTCAAGCCCACGGGCTGACTCCCGATCAAGTTGTAGCGATTGCGAATAACAATGGAGGGAAACAAGCA
TTGGAGACTGTCCAACGGCTCCTTCCCGTGTGTGTCAAGCCCACGGTTTGACGCCTGCACAAGTGGTCGCCATCG
CCAGCCATGATGGCGGTAAGCAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGAC
TGACACCCGAACAGGTGGTCGCCATTGCTTCTAATGGGGGAGGACGGCCAGCCTTGGAGTCCATCGTAGCCCAAT
TGTCAGGCCCGATCCCGCGTTGGCTGCGTTAACGAATGACCATCTGGTGGCGTTGGCATGTCTGGTGGACGAC
CCGCGCTCGATGCAGTCAAAAAGGGTCTGCCTCATGCTCCCGCATTGATCAAAAAGAACCAACCGGCGGATTCCCG
AGAGAACTTCCCATCGAGTCGCGGGATCC

(SEQ ID NO:31)

FIG. 14B

MDYKDHDG DYKDHDIDYKDDDDKMAPKKR KVG IHRGVPMVDLRTLGY SQQQEQEKIKPKV RSTVAQHHEALVGHG
FTHAHIVAL SQHPAALGT VAVKYQDMIAALPEATHEAIVGVGKQW SGARALEALLTVAGELRGPPLQLDTGQLLKI AKR
GGVTAVEAVHAWRNALTGAPLNLT PDQVVAIANNNGGKQALETVQRLLPVLCQDHGLTPEQVVAIASNIGGKQALET
VQRLLPVLCQAHGLTPDQVVAIASNIGGKQALETVQRLLPVLCQAHGLTPAQVVAIANNNGGKQALETVQRLLPVLCQ
DHGLTPDQVVAIASNIGGKQALETVQRLLPVLCQDHGLTPEQVVAIASNIGGKQALETVQRLLPVLCQAHGLTPDQV
AIANNNGGKQALETVQRLLPVLCQAHGLTPAQVVAIASNIGGKQALETVQRLLPVLCQDHGLTPDQVVAIASHDGGK
QALETVQRLLPVLCQDHGLTPEQVVAIANNNGGKQALETVQRLLPVLCQAHGLTPDQVVAIASNIGGKQALETVQRLL
PVLCQAHGLTPAQVVAIANNNGGKQALETVQRLLPVLCQDHGLTPDQVVAIASHDGGKQALETVQRLLPVLCQDHG
LTPEQVVAIASNIGGKQALETVQRLLPVLCQAHGLTPDQVVAIANNNGGKQALETVQRLLPVLCQAHGLTPAQVVAIA
SHDGGKQALETVQRLLPVLCQDHGLTPEQVVAIASNIGGGRPALESIVAQLSRPDPALAALTNDHLVALACLGGRPALD
AVKKGLPHAPALIKRTNRRIPERTSHRVAGS

(SEQ ID NO:32)

FIG. 15A

TCTAGAGCTAGCACCATGGACTACAAAGACCATGACGGTGATTATAAAGATCATGACATCGATTACAAGGATGAC
GATGACAAGATGGCCCCAAGAAGAAGAGGAAGGTGGGCATTACCGCGGGGTACCTATGGTGGACTTGAGGAC
ACTCGGTTATTCGCAACAGCAACAGGAGAAAATCAAGCCTAAGGTCAGGAGCACCGTCGCGCAACACCACGAGG
CGCTTGTGGGGCATGGCTTCACTCATGCGCATATTGTCGCGCTTTACAGCACCCCTGCGGCGCTTGGGACGGTGGC
TGTCAAATACCAAGATATGATTGCGGCCCTGCCGAAGCCACGCACGAGGCAATTGTAGGGGTGCGTAAACAGTG
GTCGGGAGCGCGAGCACTTGAGGCGCTGCTGACTGTGGCGGGTGAGCTTAGGGGGCCTCCGCTCCAGCTCGACA
CCGGGACAGCTGCTGAAGATCGCGAAGAGAGGGGGAGTAACAGCGGTAGAGGCAGTGACGCCTGGCGCAATGC
GCTCACCGGGGGCCCCCTGAACCTGACCCAGACCAGGTAGTCGCAATCGCGTCACATGACGGGGGAAAGCAAGC
CCTGGAAACCGTGCAAAGGTTGTTGCCGGTCCTTTGTCAAGACCACGGCCTTACACCGGAGCAAGTCGTGGCCATT
GCAAATAATAACGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCAGTTCTCTGTCAAGCCCACGGG
CTGACTCCCGATCAAGTTGTAGCGATTGCGTCAACATTGGAGGGAAACAAGCATTGGAGACTGTCCAACGGCTC
CTTCCCCTGTTGTGTCAAGCCCACGGTTTGACGCCTGCACAAGTGGTCGCCATCGCCAACAACAACGGCGGTAAGC
AGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGACTGACCCAGACCAGGTAGTCG
CAATCGCGTCACATGACGGGGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCCTTTGTCAAGACC
ACGGCCTTACACCGGAGCAAGTCGTGGCCATTGCAAGCAATGGGGGTGGCAAACAGGCTCTTGAGACGGTTAG
AGACTTCTCCAGTTCTCTGTCAAGCCCACGGGCTGACTCCCGATCAAGTTGTAGCGATTGCGAATAACAATGGAG
GGAAACAAGCATTGGAGACTGTCCAACGGCTCCTTCCCCTGTTGTGTCAAGCCCACGGTTTGACGCCTGCACAAGT
GGTCGCCATCGCCTCCAATATTGGCGGTAAGCAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCA
GGATCATGGACTGACCCAGACCAGGTAGTCGCAATCGCGTCAACATTGGGGGAAAGCAAGCCCTGGAAACCG
TGCAAAGGTTGTTGCCGGTCCTTTGTCAAGACCACGGCCTTACACCGGAGCAAGTCGTGGCCATTGCAAATAATAA
CGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCAGTTCTCTGTCAAGCCCACGGGCTGACTCCCGAT
CAAGTTGTAGCGATTGCGAATAACAATGGAGGGAAACAAGCATTGGAGACTGTCCAACGGCTCCTTCCCCTGTTG
TGTCAAGCCCACGGTTTGACGCCTGCACAAGTGGTCGCCATCGCCAACAACAACGGCGGTAAGCAGGCGCTGGAA
ACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGACTGACCCAGACCAGGTAGTCGCAATCGCGTCA
CATGACGGGGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCCTTTGTCAAGACCACGGCCTTACA
CCGGAGCAAGTCGTGGCCATTGCAAGCAACATCGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCA
GTTCTCTGTCAAGCCCACGGGCTGACTCCCGATCAAGTTGTAGCGATTGCGTCCAACGGTGGAGGGAAACAAGCA
TTGGAGACTGTCCAACGGCTCCTTCCCCTGTTGTGTCAAGCCCACGGTCTGACACCCGAACAGGTGGTCGCCATTG
CTTCCCACGACGGAGGACGGCCAGCCTTGGAGTCCATCGTAGCCCAATTGTCCAGGCCCGATCCCGCGTTGGCTG
CGTTAACGAATGACCATCTGGTGGCGTTGGCATGTCTTGGTGGACGACCCGCGCTCGATGCAGTCAAAAAGGGTC
TGCCCTCATGCTCCCGCATTGATCAAAAGAACCAACCGGCGGATTCCCAGAGAACTTCCCATCGAGTCGCGGGATC
C

(SEQ ID NO: 33)

FIG. 15B

MDYKDHDGDYKDHIDYKDDDDKMAPKKRKRKVGIRGVPMVDLRTLGYSSQQQEKIKPKVRSSTVAQHHEALVGHG
FTHAHIVALSQHPAALGTVAVKYQDMIAALPEATHEAIVGVGKQWSGARALEALLTVAGELRGPPLQLDTGQLLKIAR
GGVTAVEAVHAWRNALTGAPLNLTDPQVVAIASHDGGKQALETVQRLLPVLCQDHGLTPEQVVAIANNNGGKQALE
TVQRLLPVLCQAHGLTPDQVVAIASNIGGKQALETVQRLLPVLCQAHGLTPAQVVAIANNNGGKQALETVQRLLPVLC
QDHGLTPDQVVAIASHDGGKQALETVQRLLPVLCQDHGLTPEQVVAIASNNGGGKQALETVQRLLPVLCQAHGLTPDQ
VVAIANNNGGKQALETVQRLLPVLCQAHGLTPAQVVAIASNIGGKQALETVQRLLPVLCQDHGLTPDQVVAIASNIGG
KQALETVQRLLPVLCQDHGLTPEQVVAIANNNGGKQALETVQRLLPVLCQAHGLTPDQVVAIANNNGGKQALETVQR
LLPVLCQAHGLTPAQVVAIANNNGGKQALETVQRLLPVLCQDHGLTPDQVVAIASHDGGKQALETVQRLLPVLCQDH
GLTPEQVVAIASNIGGKQALETVQRLLPVLCQAHGLTPDQVVAIASNNGGGKQALETVQRLLPVLCQAHGLTPEQVVAIA
SHDGGRPALLESIVAQLSRPDPALAAALTNDHLVAIACLGGRPALDAVKKGLPHAPALIKRTNRRIPERTSHRVAGS
(SEQ ID NO:34)

FIG. 16A

TCTAGAGCTAGCACCATGGACTACAAAGACCATGACGGTGATTATAAAGATCATGACATCGATTACAAGGATGAC
GATGACAAGATGGCCCCAAGAAGAAGAGGAAGGTGGGCATTACCCGCGGGGTACCTATGGTGGACTTGAGGAC
ACTCGGTTATTTCGCAACAGCAACAGGAGAAAATCAAGCCTAAGGTGAGGAGCACCGTCGCGCAACACCACGAGG
CGCTTGTGGGGCATGGCTTCACTCATGCGCATATTGTCGCGCTTTACAGCACCTGCGGCGCTTGGGACGGTGGC
TGTCAAATACCAAGATATGATTGCGGCCCTGCCGAAGCCACGCACGAGGCAATTGTAGGGGTGCGTAAACAGTG
GTCGGGAGCGCGAGCACTTGAGGCGCTGCTGACTGTGGCGGGTGTAGCTTAGGGGGCCTCCGCTCCAGCTCGACA
CCGGGCAGCTGCTGAAGATCGCGAAGAGAGGGGGAGTAACAGCGGTAGAGGCAGTGACAGCCTGGCGCAATGC
GCTACCCGGGGCCCCCTTGAACCTGACCCAGACCAGGTAGTCGCAATCGCGTCACATGACGGGGGAAAGCAAGC
CCTGGAAACCGTGCAAAGGTTGTTGCCGGTCCTTTGTCAAGACCACGGCCTTACACCGGAGCAAGTCGTGGCCATT
GCAAATAATAACGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCAGTTCTCTGTCAAGCCCACGGG
CTGACTCCCGATCAAGTTGTAGCGATTGCGTGAACATTGGAGGGAAACAAGCATTGGAGACTGTCCAACGGCTC
CTTCCCGTGTGTGTCAAGCCCACGGTTTGACGCCTGCACAAGTGGTCGCCATCGCCAACAACAACGGCGGTAAGC
AGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGACTGACCCCAGACCAGGTAGTCG
CAATCGCGTCACATGACGGGGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCCTTTGTCAAGACC
ACGGCCTTACACCGGAGCAAGTCGTGGCCATTGCAAGCAATGGGGGTGGCAAACAGGCTCTTGAGACGGTTCAG
AGACTTCTCCAGTTCTCTGTCAAGCCCACGGGCTGACTCCCGATCAAGTTGTAGCGATTGCGAATAACAATGGAG
GGAAACAAGCATTGGAGACTGTCCAACGGCTCCTTCCCGTGTGTGTCAAGCCCACGGTTTGACGCCTGCACAAGT
GGTCGCCATCGCCTCCAATATTGGCGGTAAGCAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCA
GGATCATGGACTGACCCCAGACCAGGTAGTCGCAATCGCGTGAACATTGGGGGAAAGCAAGCCCTGGAAACCG
TGCAAAGGTTGTTGCCGGTCCTTTGTCAAGACCACGGCCTTACACCGGAGCAAGTCGTGGCCATTGCAAATAATAA
CGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCAGTTCTCTGTCAAGCCCACGGGCTGACTCCCGAT
CAAGTTGTAGCGATTGCGAATAACAATGGAGGGAAACAAGCATTGGAGACTGTCCAACGGCTCCTTCCCGTGTG
TGTCAGCCCACGTTTTGACGCCTGCACAAGTGGTCGCCATCGCCAACAACAACGGCGGTAAGCAGGCGCTGGAA
ACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGACTGACCCCAGACCAGGTAGTCGCAATCGCGTCA
CATGACGGGGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCCTTTGTCAAGACCACGGCCTTACA
CCGGAGCAAGTCGTGGCCATTGCAAGCAACATCGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCA
GTTCTCTGTCAAGCCCACGGGCTGACTCCCGATCAAGTTGTAGCGATTGCGTCCAACGGTGGAGGGAAACAAGCA
TTGGAGACTGTCCAACGGCTCCTTCCCGTGTGTGTCAAGCCCACGGTTTGACGCCTGCACAAGTGGTCGCCATCG
CCAGCCATGATGGCGGTAAGCAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGAC
TGACACCCGAACAGGTGGTCGCCATTGCTAATAATAACGGAGGACGGCCAGCCTTGGAGTCCATCGTAGCCCAAT
TGTCAGGCCCGATCCCGCCTTGGCTGCGTTAACGAATGACCATCTGGTGGCGTTGGCATGTCTTGGTGGACGAC
CCGCGCTCGATGCAGTCAAAAAGGGTCTGCCTCATGCTCCCGCATTGATCAAAAAGAACCAACCGGCGGATTCCCG
AGAGAACTTCCCATCGAGTCGCGGGATCC

(SEQ ID NO:35)

FIG. 16B

MDYKDHDGDYKDHDI DYKDDDDKMAPKKKRKVGIHRGVPMVDLRTLGYSSQQQEKIKPKVRSVAQHHEALVGHG
FTHAHIVALSQHPAALGTAVKYQDMIAALPEATHEAIVGVGKQWSGARALEALLTVAGELRGPPLQLDGTGQLLKIAR
GGVTAVEAVHAWRNALTGAPLNLTDPQVVAIASHDGGKQALETVQRLLPVLCQDHGLTPEQVVAIANNNGGKQALE
TVQRLLPVLCQAHGLTPDQVVAIASNIGGKQALETVQRLLPVLCQAHGLTPAQVVAIANNNGGKQALETVQRLLPVLC
QDHGLTPDQVVAIASHDGGKQALETVQRLLPVLCQDHGLTPEQVVAIASNIGGKQALETVQRLLPVLCQAHGLTPDQ
VVAIANNNGGKQALETVQRLLPVLCQAHGLTPAQVVAIASNIGGKQALETVQRLLPVLCQDHGLTPDQVVAIASNIGG
KQALETVQRLLPVLCQDHGLTPEQVVAIANNNGGKQALETVQRLLPVLCQAHGLTPDQVVAIANNNGGKQALETVQR
LLPVLCQAHGLTPAQVVAIANNNGGKQALETVQRLLPVLCQDHGLTPDQVVAIASHDGGKQALETVQRLLPVLCQDH
GLTPEQVVAIASNIGGKQALETVQRLLPVLCQAHGLTPDQVVAIASNIGGKQALETVQRLLPVLCQAHGLTPAQVVAIA
SHDGGKQALETVQRLLPVLCQDHGLTPEQVVAIANNNGGRPALESIVAQLSRPDPALAALTN DHLVALACLGGRPALD
AVKKGLPHAPALIKRTNRRIPERTSHRVAGS (SEQ ID NO:36)

FIG. 17A

TCTAGAGCTAGCACCATGGACTACAAAGACCATGACGGTATTATAAAGATCATGACATCGATTACAAGGATGAC
 GATGACAAGATGGCCCCAAGAAGAAGAGGAAGGTGGGCATTACCCGCGGGGTACCTATGGTGGACTTGAGGAC
 ACTCGGTTATTCGCAACAGCAACAGGAGAAAATCAAGCCTAAGGTGAGGAGCACCGTCGCGCAACACCACGAGG
 CGCTTGTGGGGCATGGCTTCACTCATGCGCATATTGTCGCGCTTTCACAGCACCTGCGGCGCTTGGGACGGTGGC
 TGTCAAATACCAAGATATGATTGCGGCCCTGCCGAAGCCACGCACGAGGCAATTGTAGGGGTGCGTAAACAGTG
 GTCGGGAGCGCGAGCACTTGAGGCGCTGCTGACTGTGGCGGGTGAGCTTAGGGGGCCTCCGCTCCAGCTCGACA
 CCGGGCAGCTGCTGAAGATCGCGAAGAGAGGGGGAGTAACAGCGGTAGAGGCAGTGCACGCCTGGCGCAATGC
 GCTACCCGGGGCCCCCTTGAACCTGACCCAGACCAGGTAGTCGCAATCGCGTCAAACGGAGGGGGAAAGCAAG
 CCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCTTTGTCAAGACCACGGCCTTACACCGGAGCAAGTCGTGGCCA
 TTGCAAATAATAACGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCAGTTCTGTCAAGCCCACGG
 GCTGACTCCCGATCAAGTTGTAGCGATTGCGTCCAACGGTGGAGGGAAACAAGCATTGGAGACTGTCCAACGGCT
 CCTTCCCGTGTGTGTCAAGCCCACGGTTTGACGCCTGCACAAGTGGTGCCTATCGCCAACAACAACGGCGGTAAG
 CAGGCGCTGGAACAGTACAGCGCCTGCTGCCTGTACTGTCCAGGATCATGGACTGACCCAGACCAGGTAGTC
 GCAATCGCGTCACATGACGGGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCTTTGTCAAGAC
 CACGGCCTTACACCGGAGCAAGTCGTGGCCATTGCATCCCACGACGGTGGCAAACAGGCTCTTGAGACGGTTCAG
 AGACTTCTCCAGTTCTGTCAAGCCCACGGGCTGACTCCCGATCAAGTTGTAGCGATTGCGTCGCATGACGGAG
 GGAAACAAGCATTGGAGACTGTCCAACGGCTCCTTCCCGTGTGTGTCAAGCCCACGGTTTGACGCCTGCACAAGT
 GGTGCCTATCGCCAGCCATGATGGCGGTAAGCAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCA
 GGATCATGGACTGACCCAGACCAGGTAGTCGCAATCGCGTCAACATTGGGGGAAAGCAAGCCCTGGAAACCG
 TGCAAAGGTTGTTGCCGGTCTTTGTCAAGACCACGGCCTTACACCGGAGCAAGTCGTGGCCATTGCAAATAATAA
 CGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCAGTTCTGTCAAGCCCACGGGCTGACTCCCGAT
 CAAGTTGTAGCGATTGCGAATAACAATGGAGGGAAACAAGCATTGGAGACTGTCCAACGGCTCCTTCCCGTGTG
 TGTCAGCCCACGGTTTGACGCCTGCACAAGTGGTGCCTATCGCCTCCAATATTGGCGGTAAGCAGGCGCTGGAA
 ACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGACTGACCCAGACCAGGTAGTCGCAATCGCGTCA
 AACGGAGGGGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCTTTGTCAAGACCACGGCCTTAC
 ACCGGAGCAAGTCGTGGCCATTGCAAATAATAACGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCC
 AGTTCTGTCAAGCCCACGGGCTGACTCCCGATCAAGTTGTAGCGATTGCGTCCAACGGTGGAGGGAAACAAGC
 ATTGGAGACTGTCCAACGGCTCCTTCCCGTGTGTGTCAAGCCCACGGTTTGACGCCTGCACAAGTGGTGCCTATC
 GCCTCGAATGGCGGCGTAAGCAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGG
 ACTGACACCCGAACAGGTGGTGCCTATGCTAATAATAACGGAGGACGGCCAGCCTTGGAGTCCATCGTAGCCCA
 ATTGTCCAGGCCGATCCCGCGTTGGCTGCGTTAACGAATGACCATCTGGTGGCGTTGGCATGTCTGGTGGACG
 ACCCGCGCTCGATGCAGTCAAAAAGGGTCTGCCTCATGCTCCCGCATTGATCAAAAAGAACCAACCGGCGGATTCCC
 GAGAGAACTTCCCATCGAGTCGCGGGATCC (SEQ ID NO:37)

FIG. 17B

MDYKDHGDYKDHDIYKDDDDKMAPKKRKRKVGIRGVPMVDLRTLGYSSQQQEKIKPKVRSTVAQHHEALVGHG
 FTHAHIVALSQHPAALGTAVVKYQDMIAALPEATHEAIVGVGKQWSGARALEALLTVAGELRGPPLQLDGTQLLKIAR
 GGVTAVEAVHAWRNALTGAPLNLTDPQVVAIASNNGGKQALETVQRLLPVLCQDHGLTPEQVVAIANNNNGGKQALE
 TVQRLLPVLCQAHGLTPDQVVAIASNNGGKQALETVQRLLPVLCQAHGLTPAQVVAIANNNNGGKQALETVQRLLPVLC
 QDHGLTPDQVVAIASHDGGKQALETVQRLLPVLCQDHGLTPEQVVAIASHDGGKQALETVQRLLPVLCQAHGLTPDQ
 VVAIASHDGGKQALETVQRLLPVLCQAHGLTPAQVVAIASHDGGKQALETVQRLLPVLCQDHGLTPDQVVAIASNIGG
 KQALETVQRLLPVLCQDHGLTPEQVVAIANNNNGGKQALETVQRLLPVLCQAHGLTPDQVVAIANNNNGGKQALETVQR
 LLPVLCQAHGLTPAQVVAIASNIGGKQALETVQRLLPVLCQDHGLTPDQVVAIASNNGGKQALETVQRLLPVLCQDHG
 LTPEQVVAIANNNNGGKQALETVQRLLPVLCQAHGLTPDQVVAIASNNGGKQALETVQRLLPVLCQAHGLTPAQVVAIA
 SNGGKQALETVQRLLPVLCQDHGLTPEQVVAIANNNNGRPALESIVAQLSRPDPALAALNDHLVALACLGGRPALD
 AVKKGLPHAPALIKRTNRRIPERTSHRVAGS (SEQ ID NO:38)

FIG. 18A

TCTAGAGCTAGCACCATGGACTACAAAGACCATGACGGTGATTATAAAGATCATGACATCGATTACAAGGATGAC
GATGACAAGATGGCCCCAAGAAGAAGAGGAAGGTGGGCATTACCCGCGGGGTACCTATGGTGGACTTGAGGAC
ACTCGGTTATTCGCAACAGCAACAGGAGAAAATCAAGCCTAAGGTGAGGAGCACCGTCGCGCAACACCACGAGG
CGTTGTGGGGCATGGCTTCACTCATGCGCATATTGTCGCGCTTTACAGCACCTGCGGCGCTTGGGACGGTGGC
TGTCAAATACCAAGATATGATTGCGGCCCTGCCGAAGCCACGCACGAGGCAATTGTAGGGGTGCGTAAACAGTG
GTCGGGAGCGCGAGCACTTGAGGCGCTGCTGACTGTGGCGGGTGAGCTTAGGGGGCCTCCGCTCCAGCTCGACA
CCGGGCGAGCTGCTGAAGATCGCGAAGAGAGGGGGAGTAACAGCGGTAGAGGCAGTGCACGCCTGGCGCAATGC
GCTACCCGGGGCCCCCTTGAACCTGACCCAGACCAGGTAGTCGCAATCGCGAACAATAATGGGGGAAAGCAAG
CCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCTTTGTCAAGACCACGGCCTTACACCGGAGCAAGTCGTGGCCA
TTGCAAGCAATGGGGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCAGTTCTCTGTCAAGCCCACG
GGCTGACTCCCGATCAAGTTGTAGCGATTGCGAATAACAATGGAGGGAAACAAGCATTGGAGACTGTCCAACGGC
TCCTTCCCGTGTGTGTAAGCCCACGGTTGACGCCTGCACAAGTGGTCGCCATCGCCAGCCATGATGGCGGTAA
GCAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGACTGACCCAGACCAGGTAGT
CGCAATCGCGTCACATGACGGGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCTTTGTCAAGA
CCACGGCCTTACACCGGAGCAAGTCGTGGCCATTGCATCCCACGACGGTGGCAAACAGGCTCTTGAGACGGTTCA
GAGACTTCTCCAGTTCTCTGTCAAGCCCACGGGCTGACTCCCGATCAAGTTGTAGCGATTGCGTCGCATGACGGA
GGGAAACAAGCATTGGAGACTGTCCAACGGCTCCTTCCCGTGTGTGTAAGCCCACGGTTTGACGCCTGCACAA
GTGGTCGCCATCGCCTCCAATATTGGCGGTAAGCAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGC
CAGGATCATGGACTGACCCAGACCAGGTAGTCGCAATCGCGAACAATAATGGGGGAAAGCAAGCCCTGGAAAC
CGTGCAAAGGTTGTTGCCGGTCTTTGTCAAGACCACGGCCTTACACCGGAGCAAGTCGTGGCCATTGCAAATAAT
AACGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCCAGTTCTCTGTCAAGCCCACGGGCTGACTCCCG
ATCAAGTTGTAGCGATTGCGTCGAACATTGGAGGGAAACAAGCATTGGAGACTGTCCAACGGCTCCTTCCCGTGT
TGTGTCAAGCCCACGGTTTGACGCCTGCACAAGTGGTCGCCATCGCCTCGAATGGCGGGCGGTAAGCAGGCGCTGG
AAACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATGGACTGACCCAGACCAGGTAGTCGCAATCGCGA
ACAATAATGGGGGAAAGCAAGCCCTGGAAACCGTGCAAAGGTTGTTGCCGGTCTTTGTCAAGACCACGGCCTTA
CACCGGAGCAAGTCGTGGCCATTGCAAGCAATGGGGGTGGCAAACAGGCTCTTGAGACGGTTCAGAGACTTCTCC
CAGTTCTCTGTCAAGCCCACGGGCTGACTCCCGATCAAGTTGTAGCGATTGCGTCCAACGGTGGAGGGAAACAAG
CATTGGAGACTGTCCAACGGCTCCTTCCCGTGTGTGTAAGCCCACGGTTTGACGCCTGCACAAGTGGTCGCCAT
CGCCAACAACAACGGCGGTAAGCAGGCGCTGGAAACAGTACAGCGCCTGCTGCCTGTACTGTGCCAGGATCATG
GACTGACACCCGAACAGGTGGTCGCCATTGCTTCCACGACGGAGGACGGCCAGCCTTGGAGTCCATCGTAGCCC
AATTGTCCAGGCCCGATCCCGCGTTGGCTGCGTTAACGAATGACCATCTGGTGGCGTTGGCATGTCTTGGTGGAC
GACCCGCGCTCGATGCAGTCAAAAAGGCTGCTGCCTCATGCTCCCGCATTGATCAAAAAGAACCAACCGGCGGATTC
CCGAGAGAACTTCCATCGAGTCGCGGGATCC (SEQ ID NO:39)

FIG. 18B

MDYKDHDGDYKDHDIDYKDDDDKMAPKKRKRKVGIHRGVPMVDLRTLGYSSQQQEKIKPKVRSTVAQHHEALVGHG
FTHAHIVALSQHPAALGTVAVKYQDMIAALPEATHEAIVGVGKQWSGARALEALLTVAGELRGPPLQLDTGQLLKIAR
GGVTAVEAVHAWRNALTGAPLNLTDPQVVAIANNNGGKQALETVQRLLPVLCQDHGLTPEQVVAIASNGGGKQALE
TVQRLLPVLCQAHGLTPDQVVAIANNNGGKQALETVQRLLPVLCQAHGLTPAQVVAIASHDGGKQALETVQRLLPVLC
QDHGLTPDQVVAIASHDGGKQALETVQRLLPVLCQDHGLTPEQVVAIASHDGGKQALETVQRLLPVLCQAHGLTPDQ
VVAIASHDGGKQALETVQRLLPVLCQAHGLTPAQVVAIASNIGGKQALETVQRLLPVLCQDHGLTPDQVVAIANNNGG
KQALETVQRLLPVLCQDHGLTPEQVVAIANNNGGKQALETVQRLLPVLCQAHGLTPDQVVAIASNIGGKQALETVQR
LPVLCQAHGLTPAQVVAIASNGGGKQALETVQRLLPVLCQDHGLTPDQVVAIANNNGGKQALETVQRLLPVLCQDHG
LTPEQVVAIASNGGGKQALETVQRLLPVLCQAHGLTPDQVVAIASNGGGKQALETVQRLLPVLCQAHGLTPAQVVAIA
NNNGGKQALETVQRLLPVLCQDHGLTPEQVVAIASHDGGRPALASIVAQLSRPDPALAALTNHHLVALACLGGRPALD
AVKKGLPHAPALIKRTNRRIPERTSHRVAGS (SEQ ID NO:40)

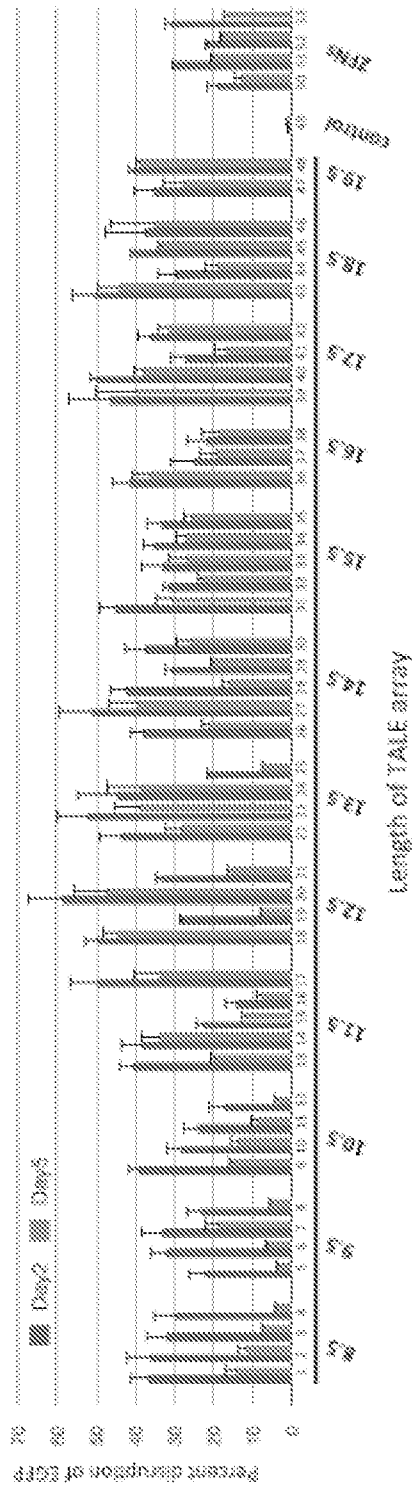


FIG. 19A

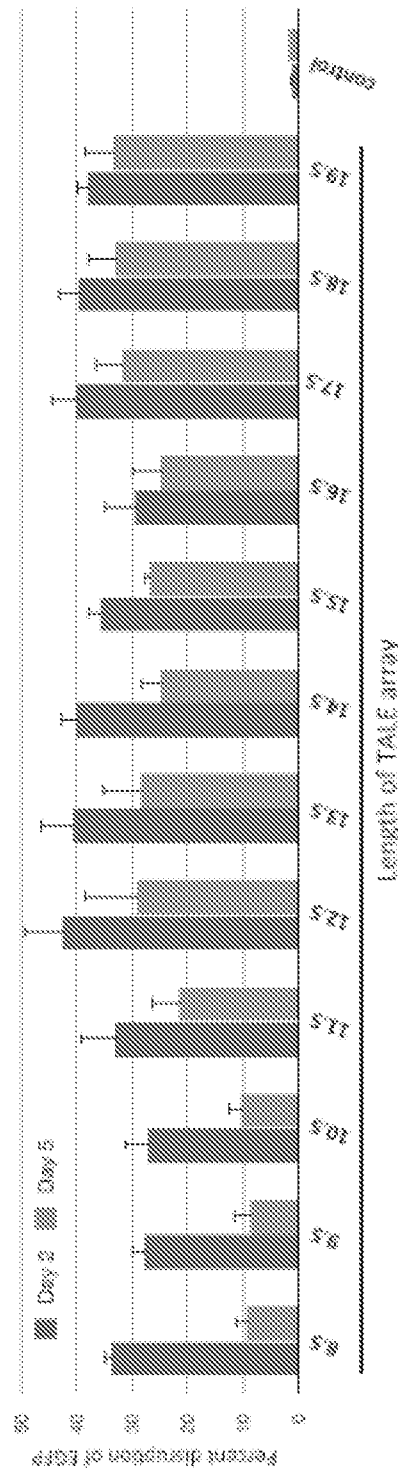


FIG. 19B

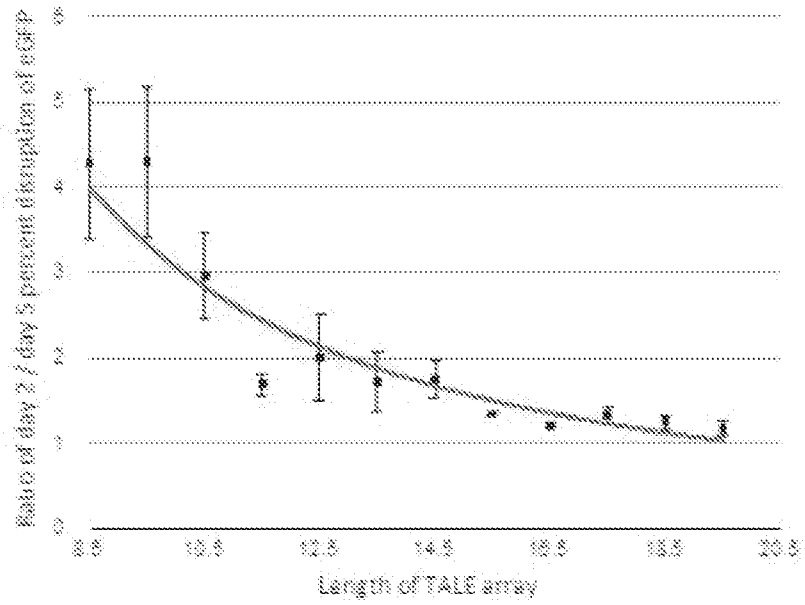


FIG. 20A

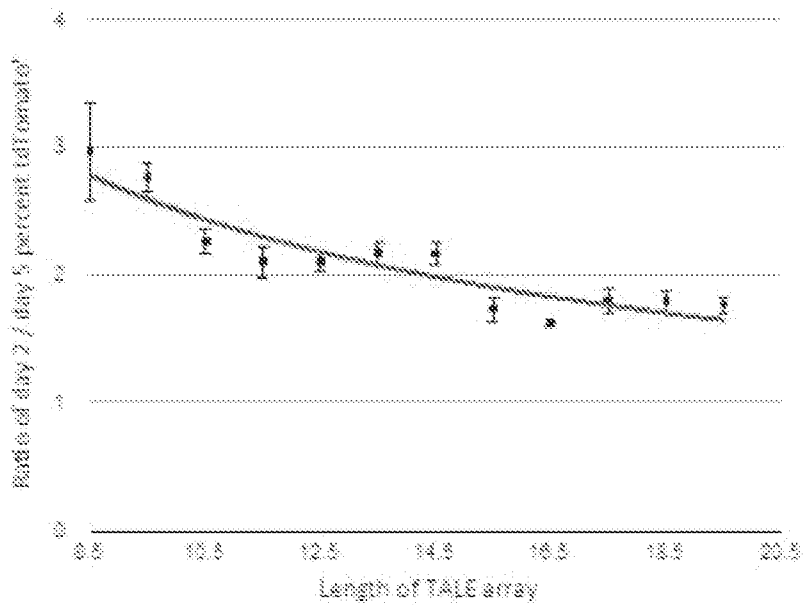


FIG. 20B

SEQUENCE	Δ SIZE	SEQ ID NO:
AXIN2		
TAL2200/TAL2201		
Mutations in 2 of 83 sequences ≈ 2.4%		
TTCCAGACTCAGTGGGAGAGCTCCCTCACCAATGAGTAGCGCTATGTTGGTGACTTGGCTCCCGGACCCAGCAGCAGCTTCCGTGA	WT	301.
TTCCAGACTCAGTGGGAGAGCTCCCTCACCAATGAGTAGCGCTATGTTGGCTCCCGGACCCAGCAGCAGCTTCCGTGA	Δ8	302.
TTCCAGACTCAGTGGGAGAGCTCCCTCACCAATGAGTAGCGCTATGTTGGCTCCCGGACCCAGCAGCAGCTTCCGTGA	Δ31	303.
BRCAL		
TAL2384/TAL2385		
Mutations in 7 of 14 sequences ≈ 50.0%		
GGCGTGGGAGAGTGGATTCCGAAGCTGACAGATGCGTATTCTTTGACGGGGGTAGSGGCGGAACCTGAGAGGGCGTAAGCCGTTGTG	WT	304.
GGCGTGGGAGAGTGGATTCCGAAGCTGACAGATGCGTATTCTTTGACGGGGGTAGSGGCGGAACCTGAGAGGGCGTAAGCCGTTGTG	Δ17	305.
GGCGTGGGAGAGTGGATTCCGAAGCTGACAGATGCGTATTCTTTGACGGGGGTAGSGGCGGAACCTGAGAGGGCGTAAGCCGTTGTG	Δ23	306.
GGCGTGGGAGAGTGGATTCCGAAGCTGACAGATGCGTATTCTTTGACGGGGGTAGSGGCGGAACCTGAGAGGGCGTAAGCCGTTGTG	Δ27	307.
GGCGTGGGAGAGTGGATTCCGAAGCTGACAGATGCGTATTCTTTGACGGGGGTAGSGGCGGAACCTGAGAGGGCGTAAGCCGTTGTG	Δ38	308.
GGCGTGGGAGAGTGGATTCCGAAGCTGACAGATGCGTATTCTTTGACGGGGGTAGSGGCGGAACCTGAGAGGGCGTAAGCCGTTGTG	Δ39	309.
CHD7		
TAL2238/TAL2239		
Mutations in 4 of 81 sequences ≈ 4.9%		
CCTGAGCTGTGGTGGAGGAGCCGTTGGAGAGATGTCAGATCCAGGAATGATGAGTCTTTTGGCCGAGGATGGGAATATTT	WT	312.
CCTGAGCTGTGGTGGAGGAGCCGTTGGAGAGATGTCAGATCCAGGAATGATGAGTCTTTTGGCCGAGGATGGGAATATTT	Δ9 (Δ11 +2)	313.
CCTGAGCTGTGGTGGAGGAGCCGTTGGAGAGATGTCAGATCCAGGAATGATGAGTCTTTTGGCCGAGGATGGGAATATTT	Δ10	314.
CCTGAGCTGTGGTGGAGGAGCCGTTGGAGAGATGTCAGATCCAGGAATGATGAGTCTTTTGGCCGAGGATGGGAATATTT	Δ28	315.
TAAAGAAATATGCAATGACAT	Δ199	316.

FIG. 21A

SEQUENCE	Δ SIZE	SEQ ID NO:
NCOR2		
TAL2284/TAL2285		
Mutations in 7 of 88 ≈ 8.0%		
TCCACACAGCCTGTGGCCACAGACAGTGGAGGCCCACTGAGCCCGCTACCCGCCCCACAGCCCTTCTACCCAGTGCAGATCGCCCGGA	WT	337.
TCCACACAGCCTGTGGCCACAGACAGTGGAGGCCCACTGAGCCCGCTA-CGGCCCCACAGCCCTTCTACCCAGTGCAGATCGCCCGGA	Δ1	338.
TCCACACAGCCTGTGGCCACAGACAGTGGAGGCCCACTGAGCCCGC-----CCCACAGCCTTCTACCCAGTGCAGATCGCCCGGA	Δ7	339.
TCCACACAGCCTGTGGCCACAGACAGTGGAGGCCCACTGAGC-----CCCACAGCCTTCTACCCAGTGCAGATCGCCCGGA	Δ12	340.
TCCACAC-----AGATCGCCCGGA	Δ69	341.
CCCCACCACC-----/-----TTTCTACCCAGTGCAGATCGCCCGGA	Δ70	342.
GCTTATTGGGG-----/-----CTTCTCTGGAGCCAGGCT	Δ213	343.
TCCACACAGCCTGTGGCCACAGACAGTGGAGGCCCACTGAGCCCGCTACAGCCCGCCACAGCCCTTCTACCCAGTGCAGATCGCC	+3	344.
JAK2		
TAL2406/TAL2407		
Mutations in 13 of 21 sequences ≈ 61.9%		
TTTCTCTTACAGGCCAAATGTTCTGAAAAGAACTCTGCATGFGSAATGGCCCTGCTTACGATGACAGAAATGGAGGGAACATCCACCCTCT	WT	345.
TTTCTCTTACAGGCCAAATGTTCTGAAAAGAACTCTGCATG-----GGCCCTGCTTACGATGACAGAAATGGAGGGAACATCCACCCTCT	Δ4	346.
TTTCTCTTACAGGCCAAATGTTCTGAAAAGAACTCTGCATG-----GGCCCTGCTTACGATGACAGAAATGGAGGGAACATCCACCCTCT	Δ5	347.
TTTCTCTTACAGGCCAAATGTTCTGAAAAGAACTCTGCATGGATGG-----TTACGATGACAGAAATGGAGGGAACATCCACCCTCT	Δ6	348.
TTTCTCTTACAGGCCAAATGTTCTGAAAAGAACTCTGCA-----GCCTGCTTACGATGACAGAAATGGAGGGAACATCCACCCTCT	Δ8	349.
TTTCTCTTACAGGCCAAATGTTCTGAAAAGAACTCTGC-----CTTACGATGACAGAAATGGAGGGAACATCCACCCTCT	Δ15 (2x)	350.
TTTCTCTTACAGGCCAAATGTTCTGAAAAGAACTCTCCATGG-----AATGGAGGGAACATCCACCCTCT	Δ24	351.
TTTCTCTTACAGGCCAAATGTTCTGAAAAGAACTCTG-----GATGACAGAAATGGAGGGAACATCCACCCTCT	Δ25	352.
TTTCTCTTACAGGCCAAATGTTCTGAAAAGAACTCTG-----CAGAAATGGAGGGAACATCCACCCTCT	Δ26	353.
TTTCTCTTACAGGCC-----ATGGAGGGAACATCCACCCTCT	Δ53	354.
T-----ACGATGACAGAAATGGAGGGAACATCCACCCTCT	Δ54	355.
TTTCTCTTACAGGCCAAATGTTCTGAAAAGAACTCTGCATGG-----ATLaagtCTTACGATGACAGAAATGGAGGGAACATCCACCCTCT	Δ4 (Δ11 +7)	356.
CGGAGGTTTGTGCAaacagaaatt-----/-----CGATGACAGAAATGGAGGGAACATCCACCCTCT	Δ288 (Δ301 +13)	357.

FIG. 21C

SEQUENCE	Δ SIZE	SEQ ID NO:
MYCN		
TAL2280/TAL2281		
Mutations in 12 of 35 sequences ≈ 34.3%		
CGGAGGCGAGCCCGATGTCGAGCTGCTCCAGTCCACCATGCCGGGCGGATGATCTGCAAGAACCCAGACCTCGAGTTTGACTCGCTACA	WT	358.
CGGAGGCGAGCCCGATGTCGAGCTGCTCCAGTCCACCATGCCGGGCGGATGATCTGCAAGAACCCAGACCTCGAGTTTGACTCGCTACA	Δ6 (Δ8 +2)	359.
CGGAGGCGAGCCCGATGTCGAGCTGCTCCAGTCCACCATGCCGGGCGGATGATCTGCAAGAACCCAGACCTCGAGTTTGACTCGCTACA	Δ8	360.
CGGAGGCGAGCCCGATGTCGAGCTGCTCCAGTCCACCATGCCGGGCGGATGATCTGCAAGAACCCAGACCTCGAGTTTGACTCGCTACA	Δ10	361.
CGGAGGCGAGCCCGATGTCGAGCTGCTCCAGTCCACCATGCCGGGCGGATGATCTGCAAGAACCCAGACCTCGAGTTTGACTCGCTACA	Δ11	362.
CGGAGGCGAGCCCGATGTCGAGCTGCTCCAGTCCACCATGCCGGGCGGATGATCTGCAAGAACCCAGACCTCGAGTTTGACTCGCTACA	Δ12	363.
CGGAGGCGAGCCCGATGTCGAGCTGCTCCAGTCCACCATGCCGGGCGGATGATCTGCAAGAACCCAGACCTCGAGTTTGACTCGCTACA	Δ18	364.
CGGAGGCGAGCCCGATGTCGAGCTGCTCCAGTCCACCATGCCGGGCGGATGATCTGCAAGAACCCAGACCTCGAGTTTGACTCGCTACA	Δ19	365.
AAGGAAGCACCCCGGCTTAA / /		
AGTGTGGAGTCCGGCCCGCCCG / /	Δ100	366.
AAGGAAGCACCCCGGCTTAA / /	Δ122	367.
AAGGAAGCACCCCGGCTTAA / /	Δ141	368.
GAGCGAGCCGATGCCCGTATCTGGGTACGGCTGCTCCAGCTGGAGAGGGGGCTCCCGGGGACCCCTCCFCGGGGGGGGG	+160 (Δ29 +189) (2x)	369.
CCCTGCCATCCCGGACAGGGGCTCAGGCGCTTGTAGTCTCGTATGCTTGGCTGGGAGCATTTGGAGGCGAGTCTAGG		
GGCAGAGGTCCTGTTCCCGCAAGCTTATGATCTGCAAGAACCCCA		
NBN		
TAL2408/TAL2409		
Mutations in 12 of 20 sequences ≈ 60%		
TGCACGTGGGCCCCAGCCCTGAGGAGCCCGGACCGATGGAAACTGCTGCCGCCCGCCGGCAGGAGTTAAGGCGAAGGGAA	WT	370.
TGCACGTGGGCCCCAGCCCTGAGGAGCCCGGACCGATGGG---GCTGCCGCCCGCCGGCAGGAGTTAAGGCGAAGGGAA	Δ5	371.
TGCACGTGGGCCCCAGCCCTGAGGAGCCCGGACCGATGGG---CTGCCGCCCGCCGGCAGGAGTTAAGGCGAAGGGAA	Δ7	372.
TGCACGTGGGCCCCAGCCCTGAGGAGCCCGGACCGATGGG---GCCGCCCGCCGGCAGGAGTTAAGGCGAAGGGAA	Δ13	373.
TGCACGTGGGCCCCAGCCCTGAGGAGCCCGGACCGATGGGACT---CCCCGCCAGGAGTTAAGGCGAAGGGAA	Δ17 (Δ19 and +2)	374.
TGCACGTGGGCCCCAGCCCTGAGGAGCCCGGACCGATGGGA---CCCCGCCAGGAGTTAAGGCGAAGGGAA	Δ19	375.
TGCACGTGGGCCCCAGCCCTGAGGAGCCCGGACCGATGGGACT---CAGGAGTTAAGGCGAAGGGAA	Δ28	376.
TGCACGTGGGCCCCAGCCCTGAGGAGCCCGGACCGATGGGAAA---GTAGGCGAAGGGAA	Δ28	377.
TGCACGTGGGCCCCAGCCCTGAGGAGCCCGGACCGG---GCCAGGTTAAGGCGAAGGGAA	Δ30	378.
TGCACGTGGGCCCCAGCCCTGAGGAGCCCGGACCGG---AGGAGTTAAGGCGAAGGGAA	Δ35	379.
TGCACGTGGGCCCCAGCCCTGAGGAGCCCGGACCGG---CGCGGCCCGGAGGTTAAGGCGAAGGGAA	Δ39	380.
TGCACGTGGGCCCCAGCC---GCCAGGTTAAGGCGAAGGGAA	Δ45	381.
GCTCCGGGAGCGCCAGCTCCCGGAGCCAT / /	Δ183	382.

FIG. 21D

SEQUENCE	Δ SIZE	SEQ ID NO:
XPC - TAL2350/TAL2351		
Mutations in 10 of 36 ≈ 27.8%		
TCCGGAAGTGAATTTGCCAGACAGCAAGCAATGCTCGGAACGCCGGCGGGGGAGCCCGGGGACCCGGAACCTGGCAGCCAG	WT	383.
TCCGGAAGTGAATTTGCCAGACAGCAAGCAATGCTCGGAAG--AACGGCCGGGGAGCCCGGGGACCGGAACCTGGCAGCCAG	Δ 2 (Δ6 and +4)	384.
TCCGGAAGTGAATTTGCCAGACAGCAAGCAATGCTCGGAAG--GGGAGCCCGGGGAGCCCGGGGACCGGAACCTGGCAGCCAG	Δ12	385.
TCCGGAAGTGAATTTGCCAGACAGCAAGCAATGCTCGGAAG--CGCCGGGAGCCCGGGGAGCCCGGGGACCGGAACCTGGCAGCCAG	Δ15	386.
TCCGGAAGTGAATTTGCCAGACAGCAAGCAATGCTCG--GGGAGCCCGGGGAGCCCGGGGACCGGAACCTGGCAGCCAG	Δ17 (x2)	387.
TCCGGAAGTGAATTTGCCAGACAGCAAGCAATGCTGGC--GGGAGCCCGGGGAGCCCGGGGACCGGAACCTGGCAGCCAG	Δ19	388.
TCCGGAAGTGAATTTGCCAGACAGCAAGCAATGTT--GGGAGCCCGGGGAGCCCGGGGACCGGAACCTGGCAGCCAG	Δ35 (Δ37 and +2)	389.
TCCGGAAGTGAATTTGCCAGACAGCAAGCAATGGCTCG--GCGCAAGCCAG	Δ39	390.
TCCGG--CAGCCAG	Δ76	391.
TCCGGAAGTGG-- / /	Δ78	392.
BRCC2 - TAL2306/TAL2307		
Mutations in 19 of 20 sequences ≈ 95.0%		
GTCGACCCCGCTGCACAGTCCGGCCCGGGCTGCTAGTGAAGGGGGCTGGGGTCCCTAGCCGGCCGGGGGCTCTTGAA	WT	393.
GTCGACCCCGCTGCACAGTCCGGCCCGGGCCATGAAGT--GGCTGGGGTCCGCTCGCTAGCCGGCCCGGGGGTCTTGAA	Δ7	394.
GTCGACCCCGCTGCACAGTCCGGCCCGGGCCATGAA--GCTGGGGTCCGCTCGCTAGCCGGCCCGGGGGTCTTGAA	Δ11	395.
GTCGACCCCGCTGCACAGTCCGGCCCGGGCC--TCGCTAGCCGGCCCGGGGGTCTTGAA	Δ31	396.
GTCGACCCCGCTGCACAGTCCGGCCCGGGCCATGAAGT--GGGGTCTTGAA	Δ39	397.
GGCCTCTCGTGAATATTCATG-- / /	Δ99	398.
GTCGACCCCGCTGCACAGTCCGGCCCGGGCCATGAAG-- / /	Δ100	399.
GTCGACCCCGCTGCACAGTCCGGCCCGGGCCATGAAGT-- / /	Δ99 (Δ104 + 5)	400.
GTCGACCCCGCTGCACAGTCCGGCCCGGGCCATGAAGT-- / /	Δ118	401.
ATGGTCTCGTAAATAGTGGAGCGGACCCCTCGAGG-- / /	Δ122	402.
GTCGACCCCGCTG-- / /	Δ200	403.
GGCCTCTCGTGAATATTCATG-- / /	Δ221	404.
GTCGACCCCGCTGCACAGTCCGGCC-- / /	Δ222 (2x)	405.
GTCGACCCCGCTGCACAGTCCGGCC-- / /	Δ238	406.
GTCGACCCCGCTGCACAGTCCGGCCCGGGCC-- / /	Δ242	407.
GTCGACCCCGCTGCACAGTCCGGCC-- / /	Δ249	408.
GTCGACCCCGCTGCACAGTCCGGCC-- / /	Δ313	409.
GTCGACCCCGCTGCACAGTCCGGCC-- / /	Δ319	410.
GGCCTCTCGTGAATATTCATGAG-- / /	Δ340	411.

FIG. 21E

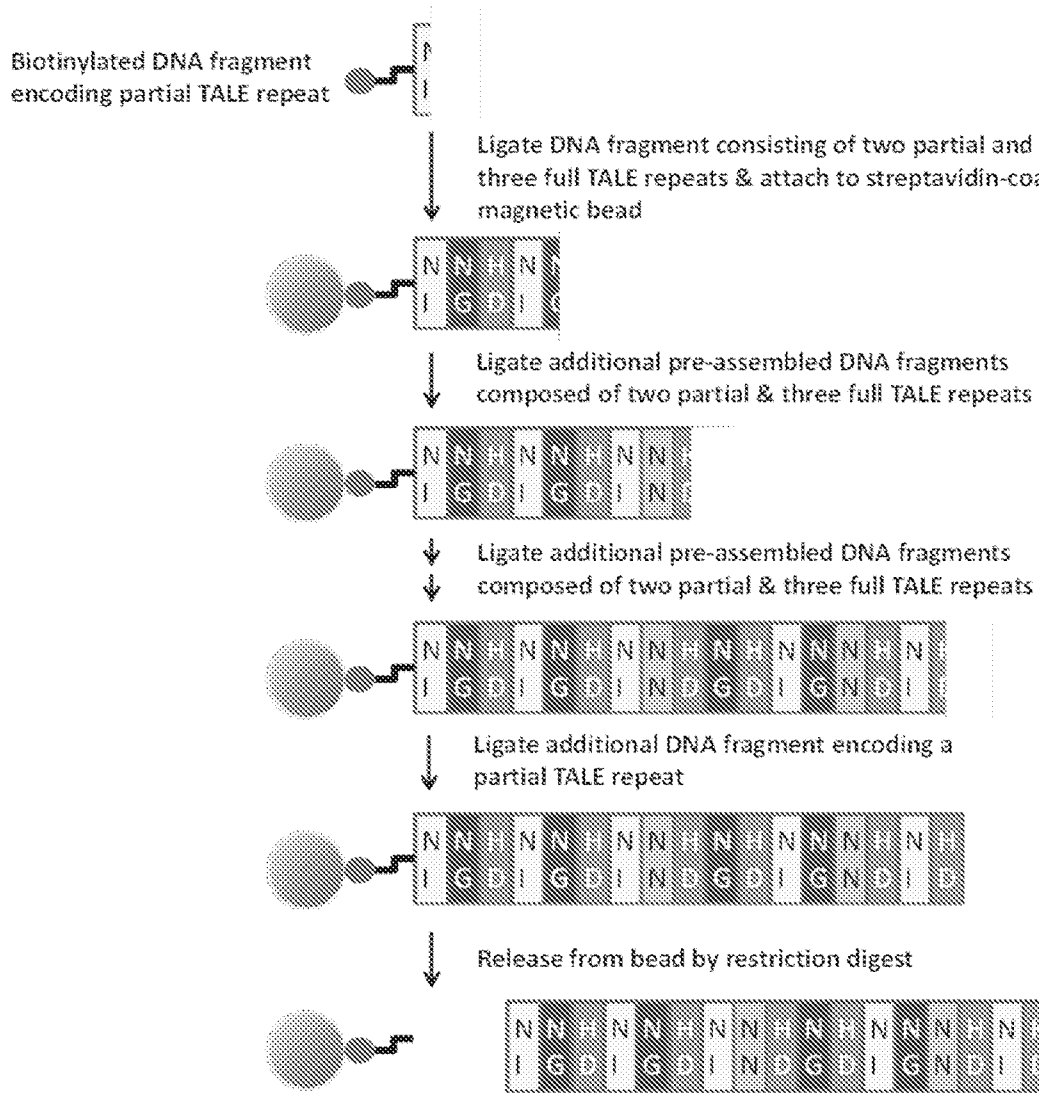


FIG. 22

