



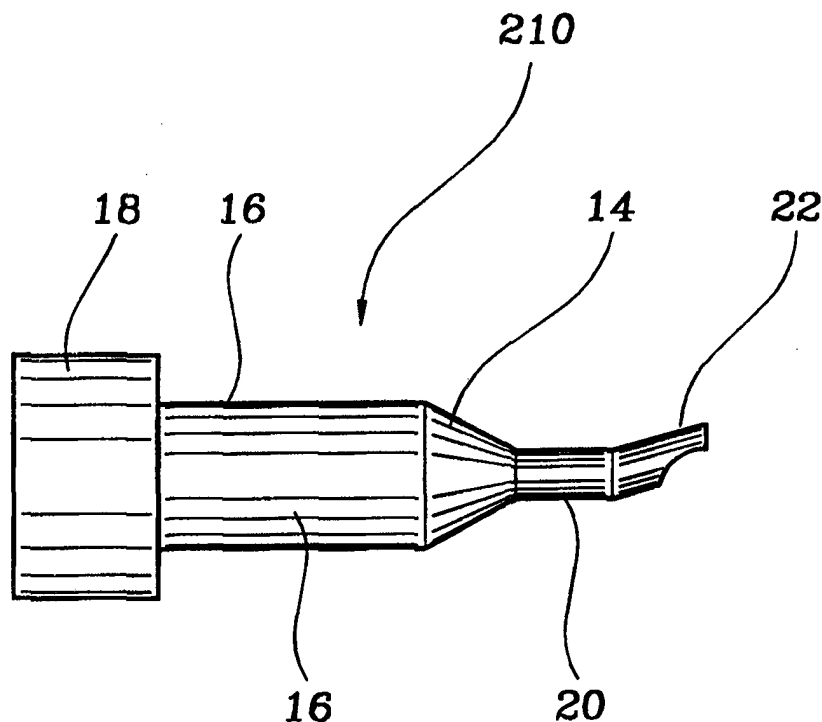
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(54) Title: APPARATUS FOR PREVENTING POSTERIOR CAPSULAR OPACIFICATION

(57) Abstract

An apparatus for destroying residual lens epithelial cells. The apparatus includes a probe (10, 210, 310, 410, 510, 610, 710) configured for insertion into the eye between the iris and the lens capsule. The probe is further configured to deliver energy therefrom to residual lens epithelial cells within the lens capsule in order to destroy them. The distal end (14, 22, 322, 520, 620, 720) of the probe is configured to allow a surgeon using the probe to reach all areas within the lens capsule to ensure that no epithelial cells remain alive after use of the probe.



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**APPARATUS FOR PREVENTING
POSTERIOR CAPSULAR OPACIFICATION**

TECHNICAL FIELD

The present invention relates to a method for
5 preventing the occurrence of posterior capsular
opacification (PCO) or secondary cataract formation
following the extracapsular extraction of a cataractous
lens. More particularly, the present invention is directed
to a method for preventing the occurrence of PCO by
10 destroying residual lens epithelial cells on the interior
surface of the lens capsule of the eye through the
application of energy thereto. In addition, the present
invention is directed to a device configured to deliver
energy to residual lens epithelial cells on the lens capsule
15 of the eye in accordance with the method of the present
invention.

Cataract extraction is among the most commonly
performed operations in the United States and the world.
The cataractous lens is located within a capsular sac or
20 lens capsule which is positioned within the posterior
chamber of the eye. In order to gain access to the
cataractous lens, an incision is made at the limbus of the
eye for the purpose of introducing a surgical instrument
into the anterior chamber of the eye. In the case of
25 extracapsular cataract extraction, a capsularhexis procedure
is performed in which a portion of the anterior membrane of
the lens capsule adjacent to the iris is removed using a
surgical cutting instrument in order to provide direct
access to the cataractous lens from the anterior chamber.
30 The lens is then removed through various known methods,
including phacoemulsification which entails the application
of ultrasonic energy to the lens in order to break it into
small pieces which can be aspirated from the lens capsule.

With the exception of the portion of the anterior membrane of the lens capsule that is removed in order to gain access to the cataractous lens, the lens capsule remains substantially intact throughout an extracapsular cataract extraction. Following removal of the cataractous lens, an artificial intraocular lens typically is implanted within the lens capsule in order to mimic the refractive function of the original lens.

Although cataractous lens removal and intraocular lens implantation provide significant benefits to most cataract patients, it is estimated that up to fifty percent (50%) of all patients who have intraocular lenses implanted within the lens capsule will develop Posterior Capsular Opacification ("PCO") or secondary cataracts within five years after surgery. PCO is caused by the deposit of cells and fibers on the intraocular lens and on the posterior capsular membrane, thereby obstructing light passing through the intraocular lens and obscuring the patient's vision. These cell deposits originate from two sources: (1) the proliferation of residual lens epithelial cells after surgery; and (2) the accumulation of inflammatory cells and protein deposits on the intraocular lens. Of these two sources, the major cause of PCO by far is the proliferation and migration of the residual lens epithelial cells on the capsular membrane.

Ophthalmic surgeons, aware of the problems associated with residual lens epithelial cells, typically take considerable care in trying to remove all residual lens epithelial cells prior to implantation of the artificial intraocular lens. However, despite these efforts, a significant number of lens epithelial cells usually are left on the interior surface of the lens capsule due to the fact that these cells are difficult to identify and are often difficult to reach due to their position on the inside

surface of the lens capsule.

The most common treatment for PCO entails the application of laser energy to the posterior membrane of the lens capsule for the purpose of destroying the lens epithelial cells propagating thereon. However, the laser energy applied to the posterior membrane of the lens capsule is ordinarily directed through the implanted intraocular lens, possibly resulting in damage to the optical and/or structural characteristics of the intraocular lens. The application of laser energy to the posterior membrane of the lens capsule also typically results in the destruction of a portion of the lens capsule as well as the residual lens epithelial cells propagating thereon. The destruction of a portion of the lens capsule creates a risk of exposure to the vitreous, possibly resulting in serious or irreparable damage to the eye. In addition, the destruction of a portion of the lens capsule creates a risk of shrinkage of the lens capsule, possibly resulting in a compromising of the optical characteristics of the intraocular lens. In certain cases, the destroyed posterior capsular tissue may regrow, e.g., as a result of a fibrin clot, thereby creating a renewed possibility of PCO. Accordingly, it is preferable to prevent the occurrence of PCO rather than attempting to treat it.

Various procedures for the prevention of PCO have been suggested in recent years. Many of these procedures have included the application of chemicals to the interior surface of the lens capsule in order to destroy residual lens epithelial cells. However, none of these procedures has proven to be particularly successful in the prevention of PCO due to the fact that it is extremely difficult to destroy residual lens epithelial cells without simultaneously destroying other cells within the eye, including the possible destruction of the corneal

endothelium. Selective destruction of residual lens epithelial cells thus appears to be the key to the prevention of PCO.

DISCLOSURE OF INVENTION

5 The method of the present invention is directed to the application of energy to the interior surface of the lens capsule following extracapsular cataract extraction for the purpose of preventing the occurrence of PCO through the destruction of residual lens epithelial cells. In one
10 embodiment of the method of the present invention, a surgical probe having a capacity to emit energy therefrom in a directionally controlled manner is inserted into the eye following extracapsular cataract extraction such that the distal end portion of the probe is anterior to the anterior
15 membrane of the lens capsule. Energy is then directed to the probe such that energy is emitted therefrom in a predetermined direction through the anterior membrane of the lens capsule in order to destroy residual lens epithelial cells disposed on the interior surface of the lens capsule.
20 The surgical probe can be moved in order to ensure that energy is delivered to substantially all portions of the lens capsule, thereby destroying as many residual lens epithelial cells as possible. The surgical probe is then deactivated and removed from the eye when the surgeon is
25 satisfied that the requisite residual lens epithelial cells have been destroyed through the application of energy from the surgical probe. This embodiment of the present invention can be practiced either before or after extracapsular cataract extraction.
30 The apparatus of the present invention is directed to a surgical probe configured for insertion into the eye such that a distal end portion of the probe can be positioned

between the iris and the lens capsule. The probe includes an electrical conductor configured to deliver energy outwardly therefrom. The probe further includes a non-conductive covering defining a port therethrough whereby
5 energy from the electrical conductor can be emitted outwardly from the probe in a directionally controlled manner. The probe also includes an electrical connector for connecting the electrical conductor to an electrical energy source.

10 Various configurations of the distal end portion of the surgical probe are shown. The surgical probe may emit monopolar or bipolar energy, or any other type of energy sufficient to destroy residual lens epithelial cells. It may also be desirable for the distal end portion of the
15 probe to be angled to either the right or left from the longitudinal axis of the probe handle. Various paddle type configurations of the distal end portion may also be desirable.

BRIEF DESCRIPTION OF DRAWINGS

20 For a more complete understanding of the present invention, reference may be had to the following Detailed Description read in connection with the accompanying drawings in which:

FIGURE 1 is an elevational view of a surgical device
25 constructed in accordance with a first embodiment of the device of the present invention;

FIGURE 2 is an end view of a surgical device constructed in accordance with a first embodiment of the device of the present invention;

30 FIGURE 3 is an elevational view of a second embodiment of a device constructed in accordance with the present invention;

FIGURE 4 is a bottom view of the probe of the second

embodiment of the device of the present invention depicted in FIGURE 3;

FIGURE 5 is a view of an eye undergoing treatment in accordance with the method of the present invention;

5 FIGURE 6 is a partial cross-sectional view of a surgical device constructed in accordance with a third embodiment of the present invention;

FIGURE 7 is a second view of the eye undergoing treatment in accordance with the method of the present invention depicting the surgical device of the present
10 invention in the eye;

FIGURES 8, 9 and 10 are top, side and bottom elevational views, respectively, of a fourth embodiment of a surgical probe constructed in accordance with the present
15 invention;

FIGURES 11, 12 and 13 are perspective, side and bottom elevational views, respectively, of a fifth embodiment of a surgical probe constructed in accordance with the present invention;

20 FIGURES 14 and 15 are side and bottom elevational views, respectively of a sixth embodiment of a surgical probe constructed in accordance with the present invention; and

FIGURES 16 and 17 are side and bottom elevational
25 views, respectively of a seventh embodiment of a surgical probe constructed in accordance with the present invention; and

FIGURE 18 is a third view of the eye undergoing treatment in accordance with the method of the present
30 invention depicting the surgical devices of Figures 11-17 of the present invention in the eye.

MODE FOR CARRYING OUT THE INVENTION

A surgical probe constructed in accordance with the

present invention is generally indicated at 10 of FIG. 1. Surgical probe 10 is constructed to be mounted on handpiece 12 at distal end portion 14 of handpiece 12. Proximal end portion 16 of handpiece 12 is configured to be attached to an energy source 18. Energy supplied by energy source 18 to handpiece 12 is directed from proximal end portion 16 to distal end portion 14. It will be appreciated that the manner in which energy is conducted through handpiece 12 will vary dependent upon the type of energy produced by source 18. For example, when probe 10 is configured to direct electrical energy to residual lens epithelial cells within the lens capsule, electrical energy from source 18 can be directed through handpiece 12 through the use of electrical wiring or through the use of other known electrical conductors. In other words, it is not necessary for the energy source 18 to be actually affixed to the proximal end of handpiece 12. It would be possible for source 1 to be located remote from handpiece 12 such that the energy can be directed to the handpiece 12 through the use of electrical wiring or the like as stated above.

However, it is important that the energy be delivered to distal end portion 14 in a controlled manner in order to prevent the unwanted delivery of energy to the patient or to the surgeon using probe 10 of the present invention.

Probe 10 is dimensioned such that it can be inserted into the anterior chamber of the eye through an incision formed at the limbus in conjunction with the removal of a cataractous lens. Proximal end portion 20 of probe 1 is mounted on distal end portion 14 of handpiece 12. Proximal end portion 20 of probe 10 and distal end portion 14 of handpiece 12 are constructed such that energy directed through handpiece 12 is transmitted to probe 1. Probe can be integrally formed on handpiece 12.

Probe 10 further includes a distal end portion 22. In

one embodiment of the present invention, distal end portion 22 is dimensioned and configured to be inserted into the lens capsule of the eye following extracapsular cataract extraction. The lens capsule is within the posterior chamber of the eye. Probe 10 can have a variety of configurations without departing from the spirit and scope of the present invention. For example, as depicted in FIG. 1, probe 10 can be straight and coaxially mounted on handpiece 12. However, it will be appreciated that it may be preferable to configure probe 10 such that it includes one or more bends along its length in order to enable a surgeon to reach otherwise difficult-to-reach areas of the lens capsule. This is particularly true when the target lens epithelial cells are located on interior surface 100 of anterior membrane 102 of lens capsule 104, as depicted in FIG. 5. The second embodiment of probe 210 depicted in FIGS. 3 and 4 is provided with a single bend in order to provide the surgeon with an enhanced ability to reach these difficult-to-reach areas of lens capsule 104. It will be appreciated that probe 10 can have a variety of other configurations having one or more bends for the purpose of facilitating the application of energy to lens capsule 104 without departing from the spirit and scope of the present invention such as shown in FIGS. 8-10.

Probe 10 is constructed to deliver energy along its length from proximal end portion 20 to distal end portion 22, and then to deliver such energy to interior surface 100 of lens capsule 104 for the purpose of destroying residual lens epithelial cells on interior surface 100. When probe 10 is configured to deliver electrical energy to interior surface 100 of lens capsule 104, it can include a single electrode, in which case electrical energy delivered by the electrode to interior surface 100 of lens capsule 104 travels outwardly from the electrode along the length of the

electrode until it reaches a ground state. In this configuration of the present invention, electrical energy emanating from the single electrode of probe 10 will tend to destroy cells nearer to probe 10 where the electrical energy is at its greatest level.

In the embodiment of the present invention depicted in FIGS. 1 and 2, probe 10 includes first electrode 24 and second electrode 26 which are oriented such that electrical energy will tend to flow from one electrode to the other. Although first electrode 24 and second electrode 26 are depicted as being coaxial in FIGS. 1 and 2, it will be appreciated that the electrodes can be configured in various ways. For example, in the second embodiment of probe 210 depicted in FIGS. 3 and 4, first electrode 224 and second electrode 226 are not coaxially mounted. It is also to be appreciated that more than two electrodes can be used in conjunction with the device and method of the present invention.

In the embodiments of the present invention depicted in FIGS. 1-4, non-conducting zones 28, 228 separate first electrodes 24, 224 and second electrodes 26, 226. Thus, electrical current directed through one of the electrodes will enter the other electrode only after being transmitted through a medium other than non-conducting zones 28, 228. For example, electrical current directed through one electrode can be conducted by residual lens epithelial cells within lens capsule 104 in order to effect the destruction of such residual lens epithelial cells. In the alternative, electrical current can be transmitted through a conductor such as a balanced salt solution that can be introduced into the eye prior to application of power from energy source 18. This aspect of the present invention will be discussed in greater detail below in connection with the method of the present invention.

In a third embodiment of the invention depicted in FIG> 6, distal end portion 322 of probe 310 is configured such that the direction of emission of energy therefrom can be limited by a non-conductive cover 313 positioned about
5 distal end portion 322 of probe 310. Non-conductive cover 313 can be formed from a variety of biocompatible, non-conductive materials, including, but not limited to, silicone. In this third embodiment, one or more portholes 311 are formed through non-conductive cover 313 proximal
10 distal end portion 322 such that electrode 324 is exposed therethrough to an external environment of probe 310. It is believed to be preferable to form portholes 311 such that they are relatively close to tip 315 of probe 310, thereby minimizing the amount of probe 310 that must be inserted
15 into the posterior chamber. Dependent upon the desired direction of emission of energy from probe 310, it may be desirable to insulate tip 315 of probe 310 in order to prevent energy from being emitted therefrom. It has been found in certain cases to be preferable to configure non-
20 conductive cover 313 such that it covers tip 315 of probe 310, thereby preventing the emission of energy outwardly therefrom, when the embodiment of the present invention depicted in FIG. 6 is used to destroy residual lens epithelial cells from a position anterior to the lens
25 capsule, as described in detail below with respect to the method of the present invention.

One of ordinary skill in the art will also recognize that it may be desirable to include an on-conductive cover in the first and second embodiments of the present invention
30 depicted in FIGS. 1-4 in order to control the direction of emission of energy from probe 10, 210.

As above-discussed, non-conductive cover 313 can be constructed of silicone. It is preferable that the non-conductive cover 313 be secured to probe 10, 210, 310 such

that it will not slide during use. It will be appreciated that movement of non-conductive cover 313 may cause damage to eye tissues. In addition, movement of non-conductive cover 313 may result in the unintended application of energy
5 to ocular tissues other than the target tissues, thereby resulting in further ocular damage.

A desirable method for securing a silicone sleeve to a probe includes the step of providing a silicone sleeve having an internal diameter less than an external diameter
10 of the distal end portion of the probe to which it is to be attached. The silicone sleeve is then immersed in ACS grade hexane until the sleeve has expanded sufficiently such that the sleeve can be placed over the distal end portion of the probe. Upon placement of the silicone sleeve over the
15 distal end portion of the probe, the silicone sleeve is permitted to dry under a fume hood. As the silicone sleeve dries, it will tend to return to its original size, thereby securing itself to the probe. In the event that non-conductive covering 313 is to cover tip 316 of probe 310, a
20 drop of silicone can be placed at tip 315 and allowed to dry. Alternatively, the sleeve can be a closed-ended sleeve, thereby obviating the need to apply a drop of silicone to tip 315. One or more portholes 311 can then be formed as desired through the non-conductive covering 313
25 using known cutting tools.

It will be appreciated that energy emitted from electrodes 24, 26 as depicted in FIGS. 1-4 will emanate outwardly from the tip of probe 10 in substantially all directions. In contrast, energy emitted from electrode 324
30 as depicted in FIG. 6 will be directed outwardly in a limited fashion. One of ordinary skill in the art will appreciate that energy will emanate from electrode 324 and through portholes 311 in a substantially conical pattern, such conical pattern having an axis lying substantially

perpendicular to a longitudinal axis of probe 310. The direction of energy emitted from probe 310 can thus be controlled by selectively forming portholes 311 in probe 310. The direction of energy emitted from probe 310 can
5 also be controlled by rotating probe 310 about its axis such that energy is emitted therefrom in the desired direction.

Energy source 18 can be any of a variety of sources of electrical or thermal energy. It has been found that electrical energy is preferable when used in conjunction
10 with the device and method of the present invention due to the greater on/off capabilities associated with a source of electrical energy and due to the general availability of electrical energy sources in operating rooms. For example, most phacoemulsification systems have the capability of
15 providing the requisite electrical power required by the device and method of the present invention. Energy source 18 can also be provided by a standard operating room system designed for bipolar cautery systems. The voltage and current limitations of such bipolar cautery systems have
20 been shown to be safe and effective when used in conjunction with the device and method of the present invention. In addition, the alternating current produced by power supplies of this type tend to induce the oscillation of charged particles in balanced salt solutions, thereby resulting in a
25 heating of the solution. The importance of this phenomenon will be discussed in greater detail below with respect to the method of the present invention. However, it is to be appreciated that the device and method of the present invention can also be used in conjunction with DC electrical
30 power sources.

Distal end portion 22 of probe 10 is rounded in the embodiment depicted in FIGS. 1 and 2. The rounded configuration of distal end portion 22 facilitates the delivery of energy from electrodes 24, 26 to residual lens

epithelial cells while simultaneously reducing the possibility of damaging lens capsule 104. However, various configurations of distal end portion 22 can be employed in conjunction with the present invention. For example, distal end portion 22 can be configured such that a plurality of electrodes can be extended therefrom when distal end portion 22 is disposed within lens capsule 104. The plurality of electrodes can be positioned relative to each other such that all or substantially all of interior surface 100 of lens capsule 104 can be subjected to energy at the same time using probe 10.

In another possible configuration, distal end portion 22 can include an inflatable tip which can be inflated when distal end portion 22 is in place within lens capsule 104. This embodiment can be used in connection with either electrical or thermal energy. When used in conjunction with electrical energy, the inflatable tip would preferably be constructed of a material having a capacity to conduct electricity such that electrical current could be passed therethrough in order to effect the destruction of residual lens epithelial cells on interior surface 100 of lens capsule 104. When the inflatable tip is used in connection with the application of thermal energy, it is preferably constructed of a heat conducting material such that heat generated within the inflatable tip is delivered to interior surface 100 and to the residual lens epithelial cells disposed thereon. Heating can be effected through a variety of known mechanisms, including the introduction of a heated fluid into the inflatable tip or through the application of energy from an energy source such as a laser to the contents of the inflatable tip.

Probe 10 can include temperature probe 30 disposed at distal end portion 22. Temperature probe 30 has a capacity to measure the temperature at distal end portion 22 and send

a signal to deactivate energy source 18 when the temperature reaches a predetermined level, thereby preventing the possible application of excessive energy levels to the eye. Alternative mechanisms for preventing the application of
5 excessive energy to the eye can also be utilized. For example, energy source 18 can be configured to provide energy pulses of relatively short duration, thereby reducing the likelihood that excessive energy will be delivered to the eye.

10 Probe 10 can also have an irrigation/aspiration capability whereby irrigating fluid can be introduced into the eye and tissue fragments and fluids can be removed from the eye during use of probe 10 in accordance with the method of the present invention.

15 The above-described device of the present invention is constructed for use in conjunction with the extracapsular extraction of a cataractous lens and the subsequent implantation of an artificial intraocular lens for the purpose of destroying residual lens epithelial cells prior
20 to the implantation of the artificial intraocular lens. Extracapsular cataract extraction generally is performed by making an incision through the limbus of the eye in order to provide access to the anterior chamber of the eye. A surgical cutting tool is then inserted through the incision
25 and into the anterior chamber. The surgical cutting tool is used to cut portion 106 from the anterior membrane 102 of lens capsule 104, thereby providing the surgeon direct access to lens 108 within lens capsule 104. Lens 108 is then removed through a known procedure such as
30 phacoemulsification in which ultrasonic energy is imparted to lens 108 in order to break lens 108 into fragments which can then be aspirated from lens capsule 104 through the use of a phacoemulsification system having irrigation/aspiration capabilities.

In a first embodiment of the method of the present invention, a surgeon will employ the device of the present invention in order to remove any residual lens epithelial cells from lens capsule 104 following the removal of lens 5 108 from lens capsule 104. Probe 10 is inserted into the eye such that distal end portion 22 is positioned at a predetermined location within lens capsule 104. Probe 10 can be inserted through a newly formed incision, but preferably is inserted through the incision created in 10 conjunction with the removal of the cataractous lens, thereby minimizing the trauma to the eye. Energy source 18 is then activated in order to provide energy to distal end portion 22 of probe 10. As discussed above, energy source 18 can be either an electrical energy source or a thermal 15 energy source. However, in the preferred embodiment of the method of the present invention, electrical energy is used due to the above-discussed beneficial aspects of using such energy.

It will be appreciated that the energy directed to 20 distal end portion 22 from energy source 18 will be transmitted into the tissues immediately surrounding distal end portion 22. Residual lens epithelial cells thus will tend to be destroyed by the application of energy from probe 10 due to their position on interior surface 100 of lens 25 capsule 104. It has been found that the application of excessive energy to probe 10 will tend to damage lens capsule 104 itself. In particular, it has been discovered that distal end portion 22 of probe 10 will tend to stick to lens capsule 104 in the event that too much energy is 30 directed to a single site on interior surface 100. The further delivery of energy from probe 10 to such a site on interior surface 100 will result in permanent, localized damage to lens capsule 104, including the possible perforation of lens capsule 104. For this reason, it is

imperative that energy be supplied by probe 10 to lens capsule 104 in a controlled manner.

One technique for limiting the amount of energy delivered to a single site on interior surface 100 of lens capsule 104 is to move distal end portion 22 of probe 10 about lens capsule 104 rather than localizing the delivery of energy, thereby minimizing the possibility that too much energy will be delivered to a single site. In the event that this technique is used, it is preferable that probe 10 be moved about lens capsule 104 in a regimented or patterned manner in order to ensure that all areas of interior surface 100 are treated.

It has also been found that the use of balanced salt solutions such as interstitial fluids, osmotically balanced salt solutions, and viscoelastic solutions, can minimize the possibility of probe 10 sticking to interior surface 100 of lens capsule 104 if such solutions are placed in lens capsule 104 prior to the application of power to probe 10. Balanced salt solutions are commonly used in ophthalmic procedures such as extracapsular cataract extraction for the purpose of preventing the collapse of the anterior chamber due to the loss of fluid through the incision. Such balanced salt solutions not only provide a buffer between probe 10 and interior surface 100, but also provide a conducting medium through which electrical energy from probe 10 can pass, thereby facilitating the transfer of energy from probe 10 to the residual lens epithelial cells.

Particularly beneficial results have been achieved through the use of viscoelastic solutions containing 2-hydroxypropylmethyl cellulose, such as the solutions sold by Storz Instrument Company, a wholly-owned subsidiary of the assignee of this invention, under the trademarks "OCCUCOAT" and "OCCUCOAT PF". It has been discovered that probe 10 is less likely to stick to interior surface 100 of lens capsule

104 at a given power setting when "OCCUCOAT" viscoelastic solutions are used as compared to other balanced salt solutions or water. This benefit may be the result of the fact that the application of electrical energy from probe 10
5 to interior surface 100 in the presence of "OCCUCOAT" viscoelastic solution causes the viscoelastic solution to form a precipitate or gel which acts as a barrier between probe 10 and interior surface 100. The resulting precipitate or gel dissipates a few seconds after
10 terminating the application of electrical energy and therefore does not pose any complications in the surgical procedure. In addition, the size and duration of this precipitate or gel has been found to be reproducible and proportional to the intensity of the power and the duration
15 of application of power from probe 10. This predictable change in the physical characteristics and appearance of the "OCCUCOAT" viscoelastic material thus enables a surgeon to identify the areas that have been treated with energy from probe 10 for the purpose of destroying residual lens
20 epithelial cells.

In addition to the above-described benefits, it has also been discovered that the use of a viscoelastic solution containing 2-hydroxypropylmethyl cellulose, such as "OCCUCOAT" viscoelastic material, in conjunction with the
25 method of the present invention results in significantly greater temperature increases when compared to other balanced salt solutions and water. Alternating current produced by energy source 18 causes the oscillation of the charged particles in a viscoelastic solution containing 2-
30 hydroxypropylmethyl cellulose, thereby resulting in the heating of the viscoelastic solution. The maximum temperature achieved using "OCCUCOAT" viscoelastic solution used in conjunction with the method of the present invention is 100°C. Such heat serves to destroy residual lens

epithelial cells within the lens capsule. It is believed that the oscillation of charged particles within the viscoelastic solution caused by the application of AC current thereto, as well as the local osmotic differences
5 resulting from such oscillations, further facilitates the destruction of the lens epithelial cells within the lens capsule.

It has been found that energy emitted from probe 10 in conjunction with the method and device of the present
10 invention may reach the iris, thereby causing tissue damage to the iris. For this reason, it may be desirable to provide an iris shield that can be placed between the lens capsule and the iris prior to directing energy through probe 10. In one embodiment of the method of the present
15 invention, an iris shield formed from a hydrogel material is placed between the iris and the lens capsule in order to prevent energy from probe 10 from adversely affecting the iris. One of ordinary skill in the art will recognize that other materials can be used to form an iris shield in
20 accordance with the teachings of the present invention, so long as such material is biocompatible, is capable of shielding the iris from energy emitted from probe 10, and is not structurally compromised by energy emitted from probe 10.

25 It will be appreciated that the amount of energy required to perform the method of the present invention will vary dependent upon a number of factors, including the size and configuration of the electrode(s) of probe 10 and the presence or absence of a conducting medium within lens
30 capsule 104. Devices with larger electrode surface areas will have higher power requirements. In addition, the desirable power level will vary dependent upon each surgeon's chosen technique. For example, if probe 10 is used in a relatively quick, sweeping motion within lens

capsule 104, a higher power may be used due to the fact that there will be less power delivered to any single site on interior surface 100 of lens capsule 104. Similarly, greater power levels can be used when a viscoelastic solution containing 2-hydroxycypropylmethyl cellulose is present due to the above-referenced characteristics of such viscoelastic solutions. However, if the surgeon prefers to treat individual sites on a methodical or sequential basis, it may be desirable to utilize lower power levels in order to minimize the possibility of damage to lens capsule 104.

In some cases it may be preferable to utilize two or more different configurations of probe 10 in conjunction with the method of the present invention in order to ensure that all areas of interior surface 100 are subjected to the energy emanating from probe 10. For example, probe 210 may be inserted for use following use of probe 10 in order to ensure that areas of interior surface 100 that may not have been treated using probe 10 are subjected to energy from probe 210. It may also be necessary in certain cases to form a second incision through the limbus in order to provide a different angle of attack for probe 10, thereby ensuring that all areas of interior surface 100 are subjected to the energy emanating from probe 10.

Following the application of energy to lens capsule 104 and the resulting destruction of residual lens epithelial cells, the surgeon deactivates and removes probe 10 from the eye. Upon the removal of particulate matter and any balanced salt solutions from lens capsule 104 using known irrigation/aspiration techniques, the surgeon can proceed with the implantation of an artificial intraocular lens 104 using a variety of known methods.

In a second embodiment of the method of the present invention, a probe 310 capable of emitting energy from its distal end portion in a predetermined direction is provided.

Although probes of various configurations can be used, it has been found to be advantageous to employ a probe such as that depicted in FIG. 6 and disclosed in detail herein in order to control the emission of energy from the probe. It is to be appreciated that the second embodiment of the method of the present invention can be used either before or after the extracapsular extraction of the cataractous lens. In the second embodiment of the method of the present invention, distal end portion 322 is inserted into the eye such that it is positioned within the posterior chamber of the eye between the iris 101 and the anterior membrane 102 of lens capsule 104 and such that porthole 311 is directed posteriorly towards the lens capsule. Energy is then directed to probe 310 such that energy is emitted therefrom at a level sufficient to destroy residual lens epithelial cells on lens capsule 104. It will be appreciated that energy emanating from probe 310 will pass through lens capsule 104 and destroy residual lens epithelial cells disposed on interior surface 100 thereof. Dependent upon the configuration of probe 310, it may be necessary to move probe 310 about in order to ensure that energy is directed to all portions of lens capsule 104, thereby ensuring that as many residual lens epithelial cells as possible are destroyed. The delivery of energy to probe 310 is then ceased and the probe is withdrawn from the eye. It is to be appreciated that a second probe having a different configuration can be inserted as above-discussed in order to reach portions of lens capsule 104 not reachable using probe 310. A second probe also can be inserted through a second incision formed at the limbus as above-discussed in order to reach portions of lens capsule 104 not reachable using probe 310. Furthermore, balanced salt solutions such as interstitial fluids, osmotically balanced salt solutions, and viscoelastic solutions, as above-discussed, can be used

in connection with the second embodiment of the method of the present invention.

A fourth embodiment of the surgical probe is disclosed in FIGS 8-10. The surgical probe 400 is shown having a
5 distal end 410 with longitudinal axis 412. The distal end 410 is provided with a right hand bend 414 as shown in FIG. 8, which is a top view of the probe. It may also be desirable to provide a surgical probe having a left hand bend to further facilitate the application of energy to lens
10 capsule 104 without departing from the scope of the present invention. Furthermore, the probe's distal end 410 is constructed in a similar manner to the probe of FIG. 6. The distal end 410 is provided with a cut away portion 416 as can be seen in FIGS. 9 and 10. The probe 400 is intended to
15 be used with a bipolar energy generator (not shown). As can be seen in FIG. 10, the cut away portion 416 provides a first electrode 418 and a second electrode 420 which are separated by a non-conductive material 422. The non-conductive material 422 surrounds the second electrode 420
20 such that in use the probe 400 will direct energy in a predetermined or downwardly direction when the probe is held in the position shown in FIG. 8. and is emersed in the balanced salt solutions and/or viscoelastic solutions which are discussed above to facilitate the transfer of energy
25 from the probe 400 to the residual lens epithelial cells. The non-conductive material 422 used in probe 400 may be a silicone as discussed above or otherwise be a ceramic material interposed between the electrodes 418 and 420.

Furthermore, some surgeons may wish to perform surgery
30 through only one incision. The difficulties lie in treating and/or destroying the epithelial cells which remain directly below the incision such as shown at 120 in FIG. 7. A single incision surgery can be accomplished by providing a surgical probe such as shown in FIGS. 11-17. FIGS. 11-13 show a

surgical probe 500 which utilizes monopolar energy to provide a predetermined or downwardly directed energy flow to destroy the epithelial cells within the lens capsule as shown in FIG. 18. The probe 500 is provided with a thin
5 ellipsoidal or oval shaped disk 510 which is connected to probe 500 through arm 512. Arm 512 extends axially from probe 500 and bends approximately 90 degrees to connect to a top central portion 514 of the disk 510 as shown in FIG. 11. It may be desirable to bend arm 512 something less than 90
10 degrees so that disk 510 is not parallel to the longitudinal axis of probe 500. However, it is important that arm 512 be attached to the top of disk 510 so that a portion 516 of disk 510 extends behind the connection point 514 between the arm and the disk, and a portion 518 of disk 510 extends
15 forward of the connection point 514 between the arm and disk. FIG. 12 shows more clearly the backward and forward extending portions 516 and 518 of disk 510. Probe 500 can be used in the second embodiment of the preferred method of the present invention, such that probe 500 can be utilized
20 to reach all areas of the lens capsule 104 through a single incision, as shown in FIG. 18. Probe 500 is shown in FIG. 13 having a generally flat bottom which forms a one piece electrode 520. The top and side portions 522 and 524, respectively, are covered with a non-conductive coating 526
25 so that the energy will be directed in a predetermined or downwardly direction when the probe 500 is positioned as shown in FIG. 12. The probe 500 is inserted into the eye such that it is positioned within the posterior chamber of the eye between the iris 101 and the anterior membrane 102
30 of the lens capsule 104 and such that the flat bottom of the disk 500 is directed posteriorly towards the lens capsule such as shown in FIG. 18. Energy is then directed to probe 500 such that energy is emitted therefrom at a level sufficient to destroy residual lens epithelial cells on lens

capsule 104. It will be appreciated that energy emanating from probe disk 510 will pass through lens capsule 104 and destroy residual lens epithelial cells disposed on interior surface 100 thereof. It will also be appreciated upon
5 referral to FIG. 18, that probe 500 can be directed about the whole lens capsule by a circular movement around the iris 101 so that the forward extending portion of the probe will extend under iris 101 to destroy residual lens epithelial cells at 122 as well as at 100. Therefore, the
10 probe 500 can be used through a single incision to destroy any and all such cells existing within the lens capsule.

Referring to FIGS. 14 and 15, a sixth embodiment of surgical probe 600 is shown which utilizes bipolar energy to destroy the residual lens epithelial cells existing within
15 the lens capsule. This probe is similar in design to the probe shown in FIGS. 11-13, except that it is designed to be used with a bipolar energy generator (not shown). Probe 600 is shown having an arm 610 with a "S" type bend 612 to offset itself from the longitudinal axis 614 of the probe
20 600. The arm 610 connects to a thin ellipsoidal or oval shaped disk 616. The outer surface of arm 618 provides a first electrode 620 on the bottom of disk 616, and a second electrode 622 is provided centrally to arm 610 to provide a second ring electrode 624 on the bottom of disk 616. The
25 second electrode 622 is surrounded by a non-conductive material 626 within arm 610 and a non-conductive material 628 on the bottom of disk 616 to separate the first electrode 620 from the second electrode 622 at all times. Energy will flow between the two electrodes when the probe
30 600 is emersed within a fluid solution which exists within the eye as discussed above. The benefit of such a probe 600 design is to provide a bipolar probe device that utilizes bipolar energy only at the periphery of the disk 616 as shown in FIG. 15.

However, as a means to increase the active surface area of a probe device as shown in FIGS. 14 and 15, while reducing the depth to which energy penetrates the lens capsule, it may be desirable to provide a probe 700 as shown in FIGS 16 and 17. Probe 700 is intended to be used with a bipolar energy generator (not shown) and is provided with an arm 710 connected to an ellipsoidal or oval disk 720 as described above. Additionally, the disk 720 is provided with a first electrode 722 and a second electrode 724 which consist of multiple small electrodes in close proximity to each other, but are still insulated from one another by non-conductive material 726, as shown on the bottom of disk 720 in FIG. 17. Reducing the size and distance between the electrodes will reduce the distance which energy radiates away from the bottom of disk 720. It may also be desirable to provide a variety of patterns for the two electrodes on the bottom of disk 720 to allow for an optimal energy pattern to be produced by disk 720 for use in destroying residual lens epithelial cells within the lens capsule of the eye.

Although the device and method of the present invention have been disclosed herein with respect to certain preferred embodiments, it will be apparent to one of ordinary skill in the art that various modifications can be made to the invention without departing from the spirit and scope of the invention disclosed and claimed herein.

**APPARATUS FOR PREVENTING
POSTERIOR CAPSULAR OPACIFICATION**

CLAIMS

1. An instrument for destroying residual lens
5 epithelial cells in a lens capsule of an eye, said
instrument comprising:
 an electrical energy source (18);
 a probe (10, 210, 310) comprising an electrode (24, 26,
224, 226, 322, 418, 420, 620, 720) electrically coupled to
10 said electrical energy source, said probe having a distal
end portion (14, 22, 322) configured for insertion into said
eye between an iris of said eye and said lens capsule; and
 an insulating sleeve (28, 228, 313) surrounding said
distal end portion of said probe, said insulating sleeve
15 defining an aperture therethrough whereby electrical energy
delivered from said electrical energy source to said
electrode is emitted outwardly from said probe through said
aperture defined through said insulating sleeve, and whereby
electrical energy is emitted outwardly from said probe in a
20 directionally controlled manner.

2. An instrument (310) for destroying residual lens
epithelial cells in a lens capsule of an eye in accordance
with claim 1, wherein said insulating sleeve (313) is
constructed of silicone.

25 3. An instrument (310) for destroying residual lens
epithelial cells in a lens capsule of an eye in accordance
with claim 1, wherein said insulating sleeve (313) is
constructed of ceramic.

4. An instrument for destroying residual lens epithelial cells in a lens capsule of an eye in accordance with claim 1, wherein a plurality of apertures (311) are defines through said insulating sleeve (313).

5 5. An instrument for destroying residual lens epithelial cells in a lens capsule of an eye in accordance with claim 1, wherein said distal end portion is straight and coaxially mounted to said probe.

10 6. An instrument (210) for destroying residual lens epithelial cells in a lens capsule of an eye in accordance with claim 1, wherein said distal end portion is bent to offset the distal end portion from the longitudinal axis of said probe.

15 7. An instrument (400,500,600) for destroying residual lens epithelial cells in a lens capsule of an eye in accordance with claim 1, wherein the distal end portion (410) is provided with a generally ellipsoidal shaped disk (516, 616,720) such that said distal end portion is connected to the center (514) of one side of said disk, the
20 other side of said disk defining an electrode (418,420,520) for emitting electrical energy from said probe in a directionally controlled manner.

25 8. An instrument (600) for destroying residual lens epithelial cells in a lens capsule of an eye in accordance with claim 7, wherein the disk (616) has a plane which is offset (612) and generally parallel with the longitudinal axis of the probe.

9. An instrument (400) for destroying residual lens epithelial cells in a lens capsule of an eye in accordance

with claim 1, wherein said electrode comprises:

a first electrode (418) electrically coupled to said electrical energy source;

5 a second electrode (420) coupled to said electrical energy source; and

a non-conductive insulating material (422) interposed between said first and second electrode to provide a bipolar probe, and whereby electrical energy is emitted outwardly from said probe from the first electrode
10 to the second electrode.

10. An instrument (400,500,600) for destroying residual lens epithelial cells in a lens capsule of an eye in accordance with Claim 9, wherein the distal end portion (410) is provided with a generally ellipsoidal shaped disk
15 (510,616) such that said distal end portion (410) is connected to the center (514) of one side of said disk, the other side of said disk defining a generally flat surface, the first and second electrodes (418,420) being provided on said surface and being separated by the non-conductive
20 insulating material (422) for emitting electrical energy from one electrode to the other electrode in a directionally controlled manner.

11. An instrument (600) for destroying residual lens epithelial cells in a lens capsule of an eye in accordance
25 with claim 10, wherein the disk has an axis which is offset (612) and generally parallel with the longitudinal axis of the probe.

12. An instrument (600) for destroying residual lens epithelial cells in a lens capsule of an eye in accordance
30 with Claim 11, wherein the first electrode (618) is provided about the periphery of the disk and the second electrode

(620) is spaced from and generally follows the same shape as the first electrode, the non-conductive insulating material (628) being interposed between the first and second electrodes.

5 13. An instrument (700) for destroying residual lens epithelial cells in a lens capsule of an eye in accordance with Claim 10, wherein said first electrode (722) is provided with a plurality of branches and said second electrode (724) is provided with a plurality of branches
10 covering a majority of the flat surface of said disk such that said first and second electrodes are always separated by the non-conducting insulating material (726) whereby electrical energy is emitted outwardly from said probe in a directionally controlled manner.

15 14. An instrument for destroying residual lens epithelial cells in a lens capsule of an eye, said instrument comprising:
 an electrical energy source (18);
 a probe (10,210,310,410,510,616) comprising an
20 electrode (24,26,224,226,322,418,420,520,620) electrically coupled to said electrical energy source, said probe having a distal end portion configured for insertion into said eye; and

 an insulating sleeve (28,228,313,422,628) surrounding
25 said distal end portion of said probe, said insulating sleeve defining an aperture therethrough whereby electrical energy delivered from said electrical energy source to said electrode is emitted outwardly from said probe through said aperture defined through said insulating sleeve, and whereby
30 electrical energy is emitted outwardly from said probe in a directionally controlled manner to destroy residual lens epithelial cells within the lens capsule.

15. An instrument for destroying residual lens epithelial cells in a lens capsule of an eye in accordance with claim 14, wherein a plurality of apertures (311) are defines through said insulating sleeve (313).

5 16. An instrument for destroying residual lens epithelial cells in a lens capsule of an eye in accordance with claim 14, wherein said distal end portion is straight and coaxially mounted to said probe.

10 17. An instrument (210) for destroying residual lens epithelial cells in a lens capsule of an eye in accordance with claim 14, wherein said distal end portion is bent to offset the distal end portion from the longitudinal axis of said probe.

15 18. An instrument (400,500,600) for destroying residual lens epithelial cells in a lens capsule of an eye in accordance with Claim 14, wherein the distal end (410) of the probe is configured for insertion into said eye between an iris of said eye and said lens capsule, said distal end portion (410) being provided with a generally ellipsoidal
20 shaped disk (510,616,720) such that said distal end portion is connected to the center of one side of said disk, the other side of said disk defining an electrode for emitting electrical energy from said probe in a directionally controlled manner.

25 19. An instrument for destroying residual lens epithelial cells in a lens capsule of an eye in accordance with Claim 18, wherein the disk (616) has a plane which is offset (612) and generally parallel with the longitudinal axis of the probe.

20. An instrument for destroying residual lens epithelial cells in a lens capsule of an eye in accordance with Claim 18, wherein said electrode comprises:

5 a first electrode (418) electrically coupled to said electrical energy source;

a second electrode (420) coupled to said electrical energy source; and

10 a non-conductive insulating material (422) interposed between said first and second electrode to provide a bipolar probe, and whereby electrical energy is emitted outwardly from said probe from the first electrode to the second electrode.

21. An instrument (600) for destroying residual lens epithelial cells in a lens capsule of an eye in accordance
15 with Claim 20, wherein the distal end portion is provided with a generally ellipsoidal shaped disk (616) such that said distal end portion is connected to the center (514) of one side of said disk, the other side of said disk defining a generally flat surface, the first and second electrodes
20 (618, 620) being provided on said surface and being separated by the non-conductive insulating material (628) for emitting electrical energy from one electrode to the other electrode in a directionally controlled manner.

22. An instrument for destroying residual lens
25 epithelial cells in a lens capsule of an eye in accordance with Claim 20, wherein the disk has an axis which is offset (612) and generally parallel with the longitudinal axis of the probe.

23. An instrument for destroying residual lens epithelial cells in a lens capsule of an eye in accordance with Claim 22, wherein the first electrode (618) is provided

about the periphery of the disk and the second electrode (620) is spaced from and generally follows the same shape as the first electrode, the non-conductive insulating material (628) being interposed between the first and second electrodes.

24. An instrument for destroying residual lens epithelial cells in a lens capsule of an eye in accordance with Claim 22, wherein said first electrode is provided with a plurality of branches and said second electrode is provided with a plurality of branches covering a majority of the flat surface of said disk such that said first and second electrodes are always separated by the non-conducting insulating material whereby electrical energy is emitted outwardly from said probe in a directionally controlled manner.

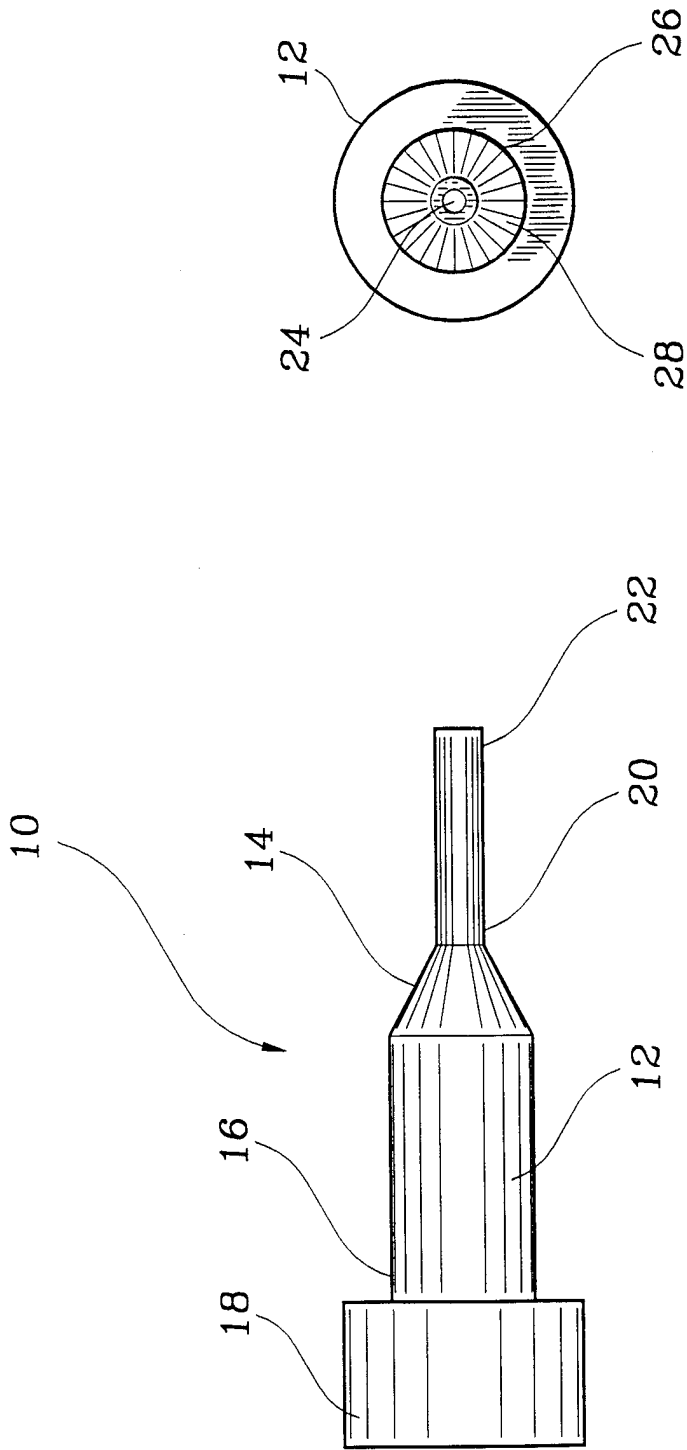


FIGURE 2

FIGURE 1

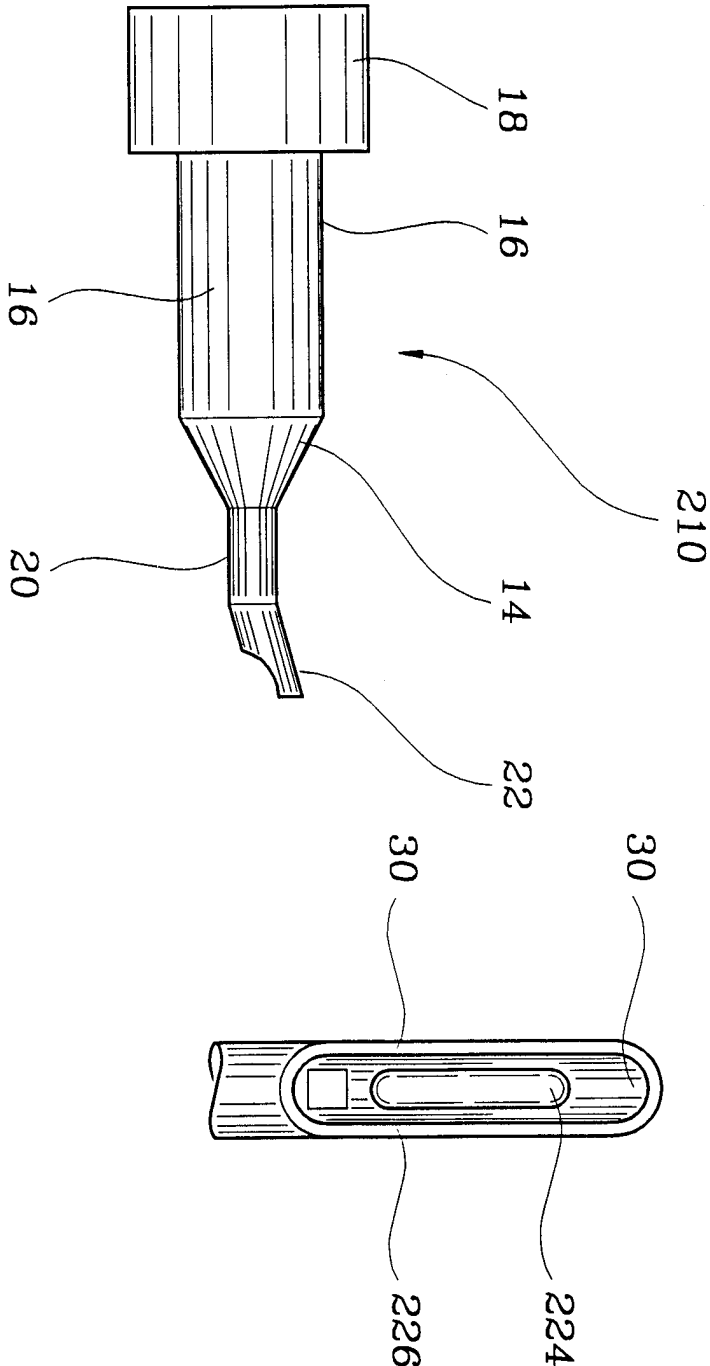


FIGURE 3

FIGURE 4

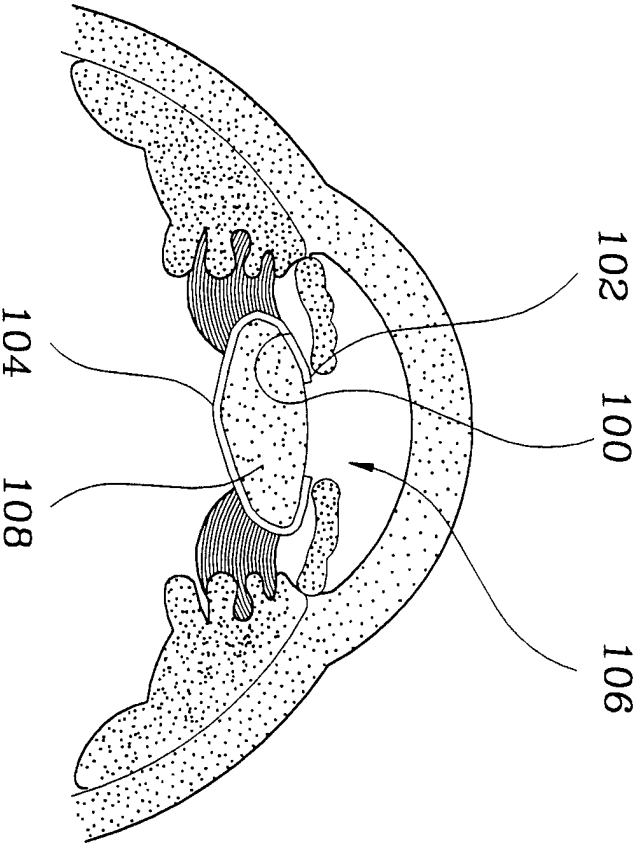


FIGURE 5

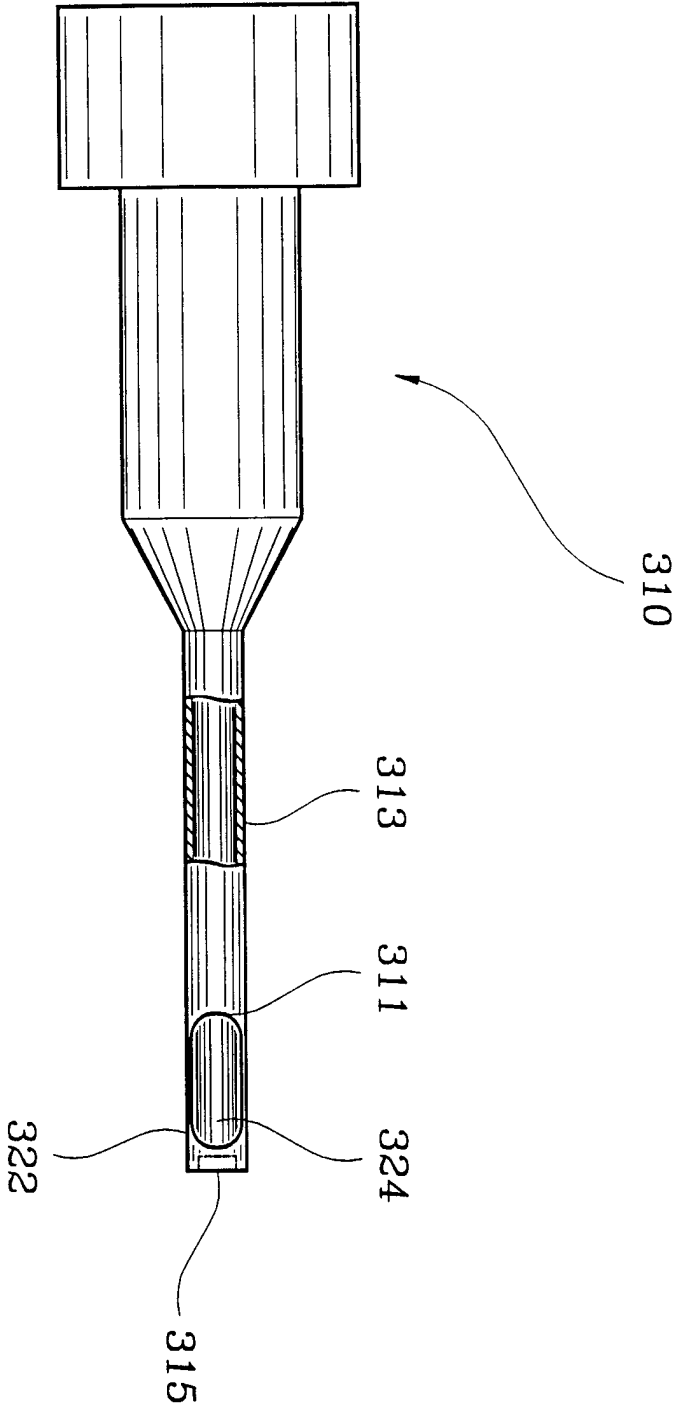


FIGURE 6



FIGURE 7

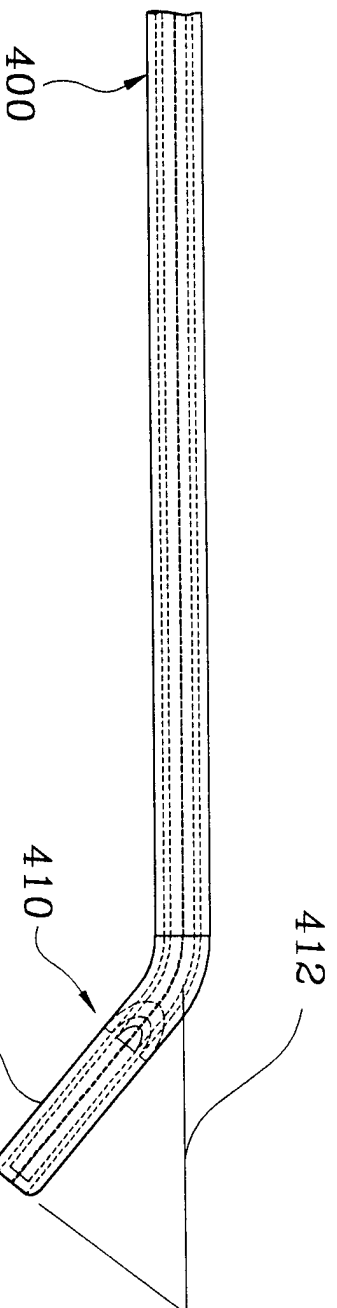


FIGURE 8

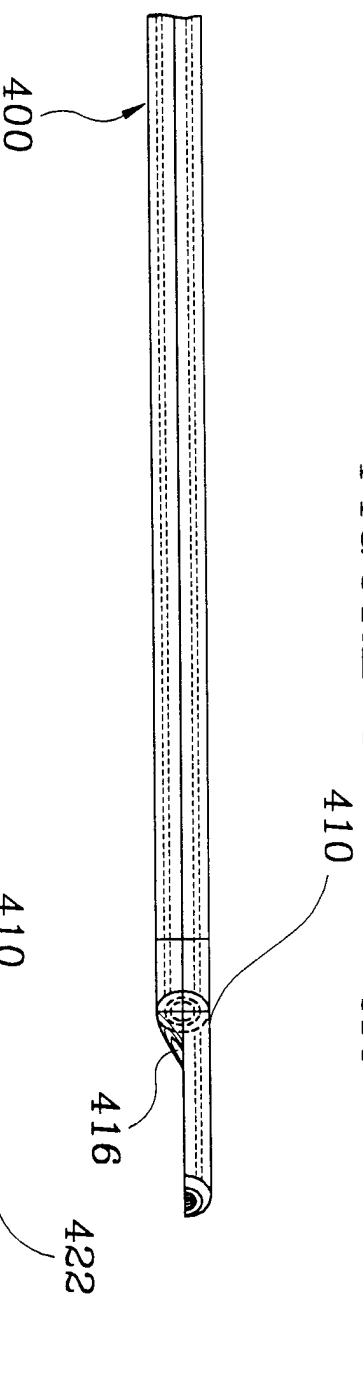


FIGURE 9

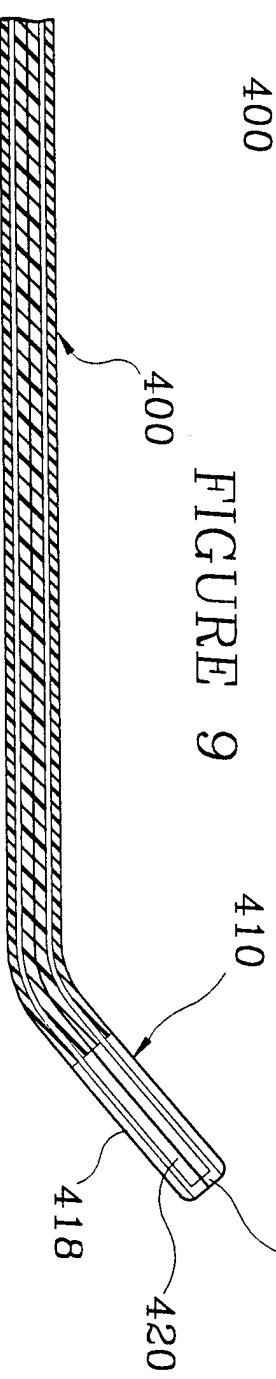
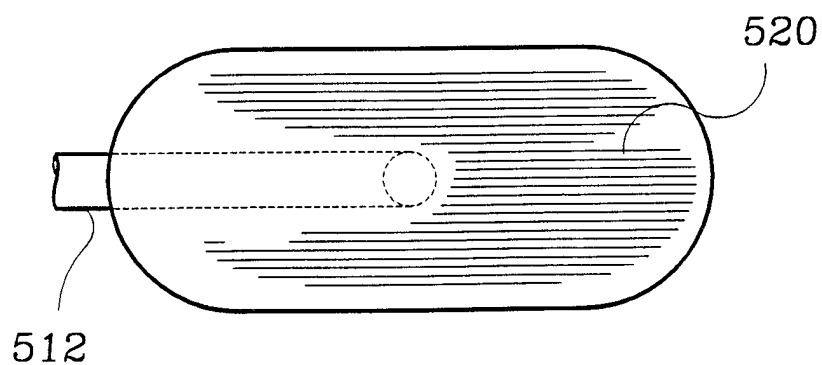
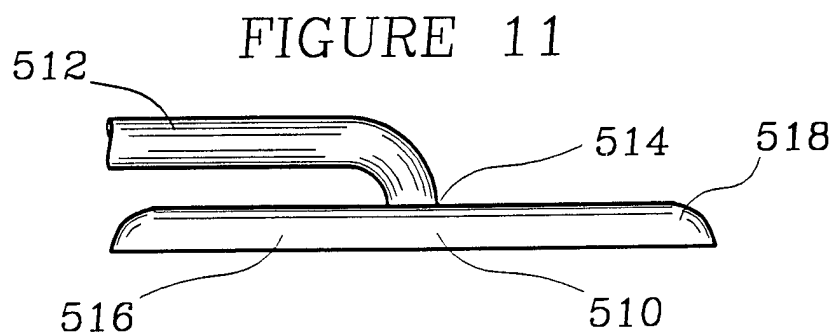
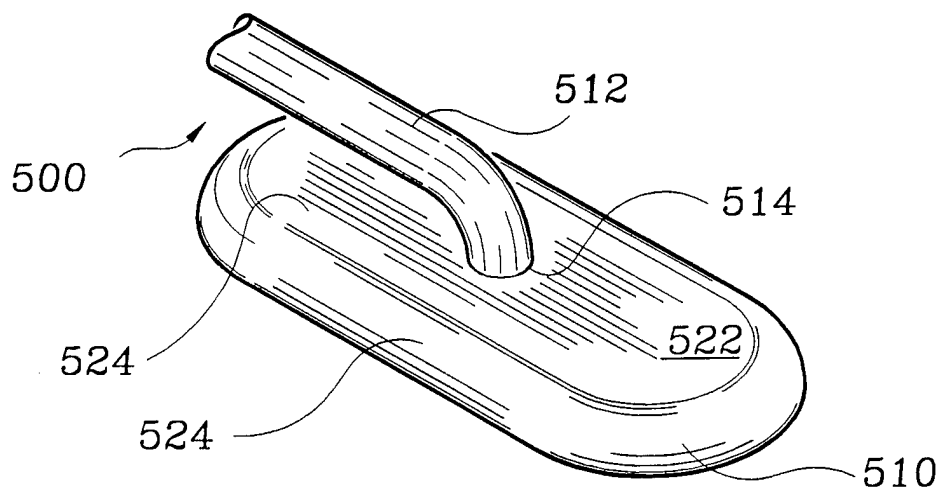


FIGURE 10



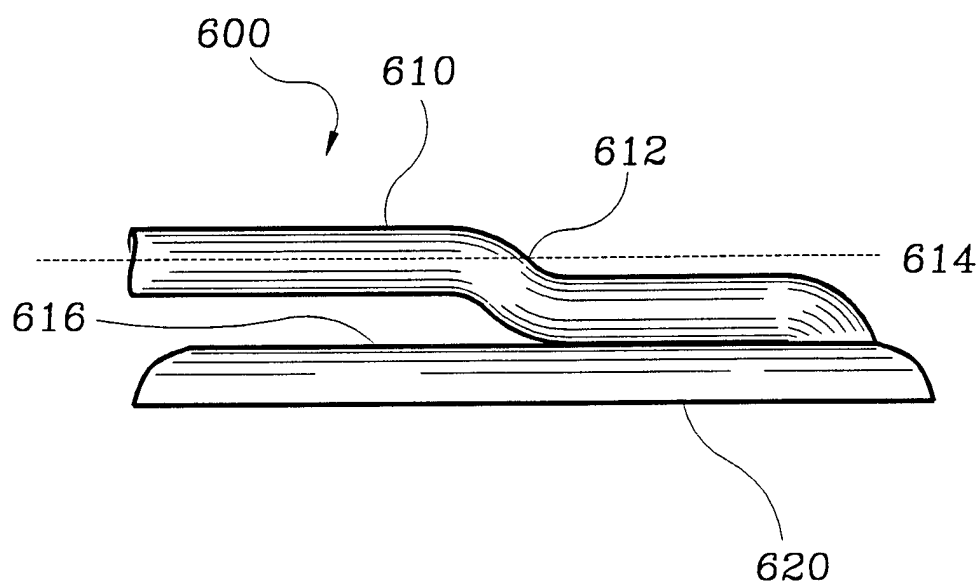


FIGURE 14

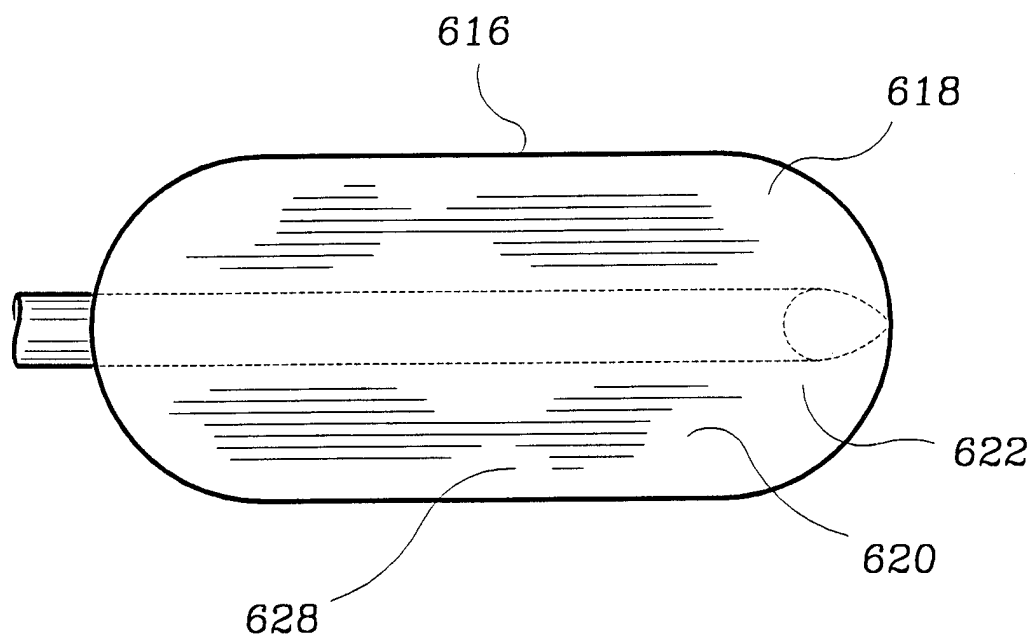


FIGURE 15

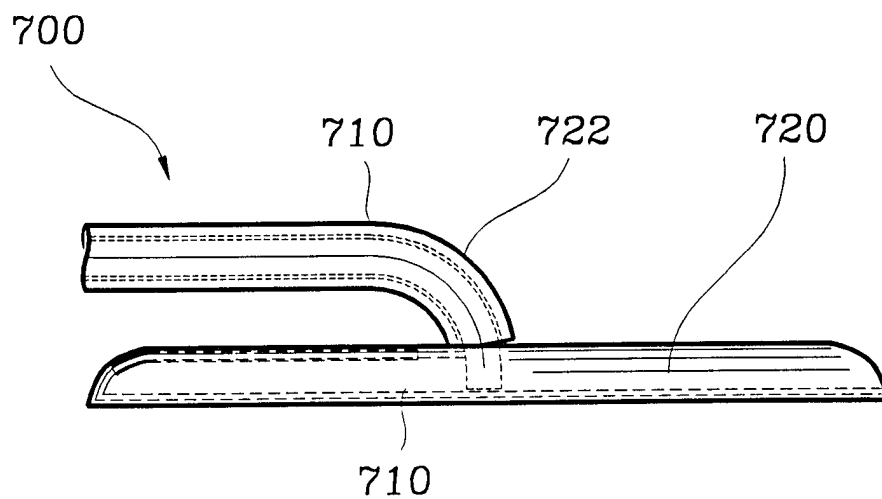


FIGURE 16

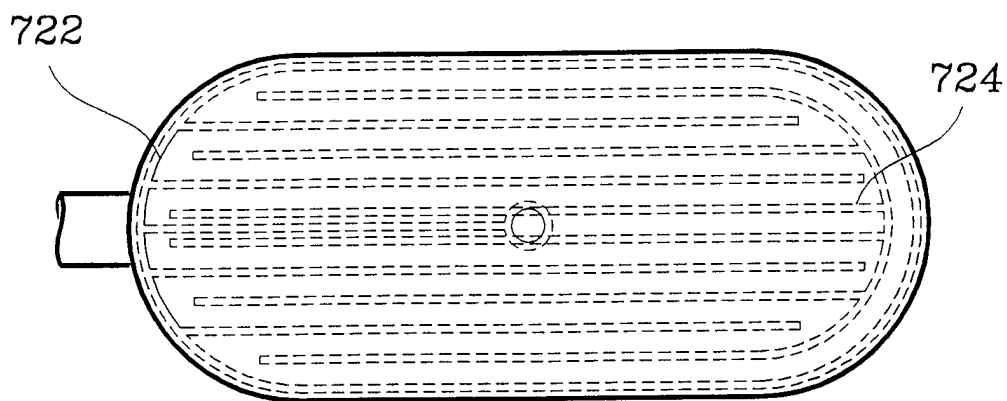


FIGURE 17

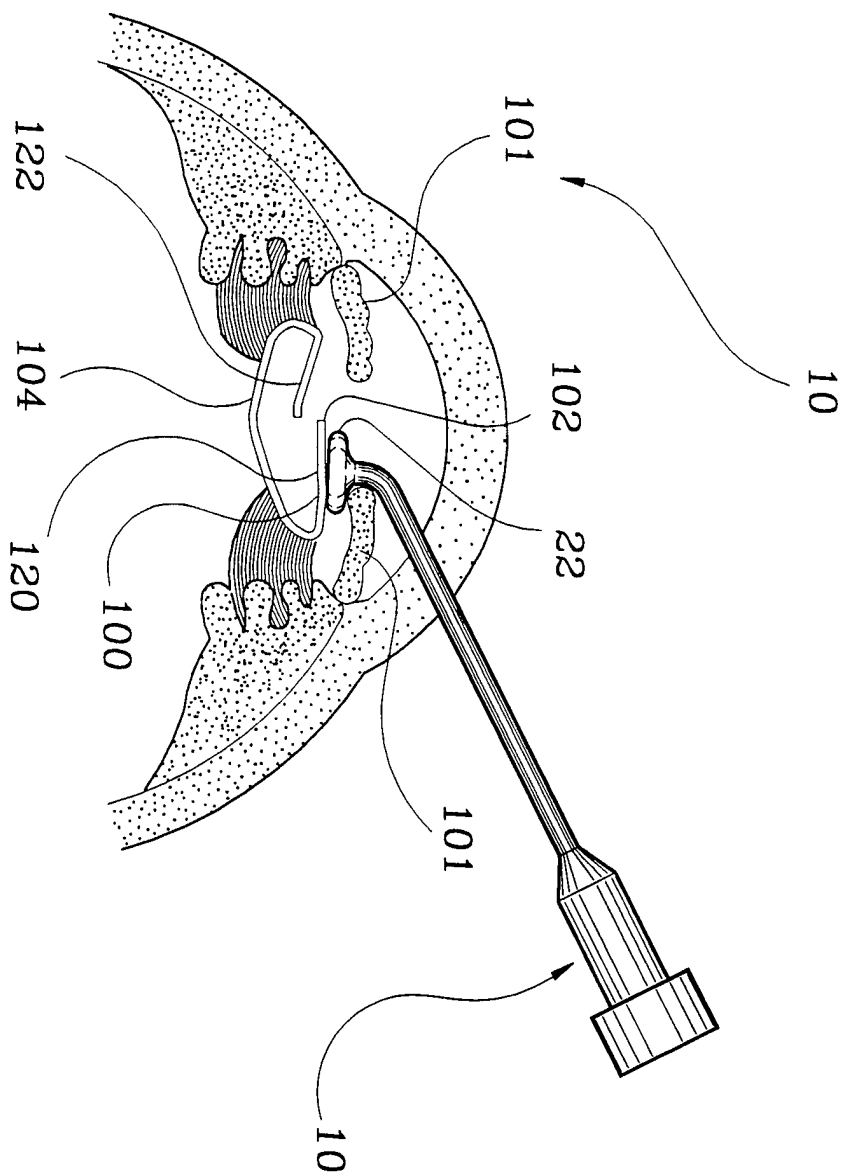


FIGURE 18

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US96/17322

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) :A61B 17/36

US CL :606/004

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 606/4-6, 27-52

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X --- Y	US 5,533,999 A (HOOD et al) 09 July 1996, entire disclosure	1, 5-7, 10-12, 14-19 ----- 2, 3, 8
X --- Y	US 4,381,007 A (DOSS) 26 April 1983, entire document.	1, 5 ----- 2, 3, 7
A, P	US 5,582,608 A (BROWN) 10 December 1996.	1-24



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents:	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier document published on or after the international filing date	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&" document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means	
"P" document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

21 DECEMBER 1996

Date of mailing of the international search report

04 FEB 1997

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